THERMODYNAMICS of Pharmaceutical Systems

An Introduction to Theory and Applications

KENNETH A. CONNORS SANDRO MECOZZI



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Second Edition

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PREFACE

The preparation of this second edition is motivated by the effectiveness of the first edition in classroom instruction, so the original structure and content have been retained. Our changes have been directed to the needs of students and other readers. Most obviously, we have moved the review of mathematics, formerly an appendix, to the front of the book (and, by labeling it Chapter 0, have conveniently preserved the structure and numbering throughout the following body of the text). We have added new textual material and figures in several places, most notably in Chapter 14, the chapter treating noncovalent chemical interactions; it is this chapter that most clearly distinguishes the content of this book from that of other thermodynamic texts. Two new appendices provide ancillary material that expands certain matters bordering the subject of classical thermodynamics.

Obviously, we have taken this opportunity to correct errors in the first edition, and we hope that we have not introduced many new ones.

We thank our colleague Professor Lian Yu for valuable discussions.

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Madison, Wisconsin

PREFACE TO THE FIRST EDITION

Classical thermodynamics, which was largely a nineteenth-century development, is a powerful descriptive treatment of the equilibrium macroscopic properties of matter. It is powerful because it is general, and it is general because it makes no assumptions about the fundamental structure of matter. There are no atoms or molecules in classical thermodynamics, so if our ideas about the atomic structure of matter should prove to be wrong (a very possible outcome to many nineteenth-century scientists), thermodynamics will stand unaltered. What thermodynamics does is to start with a few very general experimental observations expressed in mathematical form and then develop logical relationships among macroscopic observables such as temperature, pressure, and volume. These relationships turn out to have great practical value.

Of course, we now have firm experimental and theoretical reasons to accept the existence of atoms and molecules, so the behavior of these entities has been absorbed into the content of thermodynamics, which thereby becomes even more useful to us. In the following we will encounter the most fundamental ideas of this important subject, as well as some applications of particular value in pharmacy. In keeping with our needs, the treatment will in places be less rigorous than that in many textbooks, and we may omit descriptions of detailed experimental conditions, subtleties in the arguments, or limits on the conclusions when such omissions do not concern our practical applications. But despite such shortcuts, the thermodynamics is sound, so if you later study thermodynamics at a deeper level, you will not have to "unlearn" anything. Thermodynamics is a subject that benefits from, or may require, repeated study, and the treatment here is intended to be the introductory exposition.

Here are a few more specific matters that may interest readers. Throughout the text there will be citations to the Bibliography at the end of the book and to the Notes sections that appear at the end of most chapters. Students will probably not find it necessary to consult the cited entries in the Bibliography, but I encourage you to glance at the Notes, which you may find to be interesting and helpful. Two of my practices in the text may be regarded by modern readers as somewhat old-fashioned, and perhaps they are, but here are my reasons. I make considerable use of certain units, such as the kilocalorie and the dyne, that are formally obsolete; not only is the older literature expressed in terms of these units, but they remain in active use, so the student must learn to use them. Appendix B treats the conversion of units. My second peculiar practice, which may seem quaint to students who have never used a table of logarithms, is often to express logarithmic relationships in terms of Briggsian (base 10) logarithms rather than natural logarithms. There are two reasons for the continued use of base 10 logarithms; one is that certain functions, such as pH and p*K*, are defined by base 10 logs, and these definitions can be taken as invariant components of chemical description; and the second reason, related to the first, is that order-of-magnitude comparisons are simple with base 10 logarithms, since we commonly operate with a base 10 arithmetic.

Obviously, there is no new thermodynamics here, and I have drawn freely from several of the standard references, which are cited. Perhaps the only unusual feature of the text is my treatment of entropy. The usual development of the entropy concept follows historical lines, invoking heat engines and Carnot cycles. I agree with Guggenheim (1957, p. 7), however, that the idea of a Carnot cycle is at least as difficult as is that of entropy. Guggenheim then adopts a postulational attitude toward entropy [a method of approach given very systematic form in a well-known book by Callen (1960)], whereas I have developed a treatment aimed at establishing a stronger intuitive sense in my student readers [Nash (1974, p. 35) uses a similar strategy]. My approach consists of these three stages: (1) The basic postulates of statistical mechanics are introduced, along with Boltzmann's definition of entropy, and the concept is developed that spontaneous processes take place in the direction of greater probability and therefore of increased entropy; (2) with the statistical definition in hand, the entropy change is calculated for the isothermal expansion of an ideal gas; and (3) finally, we apply classical thermodynamic arguments to analyze the isothermal expansion of an ideal gas. By comparing the results of the statistical and the classical treatments of the same process, we find the classical definition of entropy, dS = dq/T, that will provide consistency between the two treatments.

Lectures based on this text might reasonably omit certain passages, only incidentally to save time; more importantly, the flow of ideas may be better served by making use of analogy or chemical intuition, rather than rigorous mathematics, to establish a result. For a good example of this practice, see Eq. (4.1) and the subsequent discussion; it seems to me to be more fruitful educationally to pass from Eq. (4.1), which says that, for a pure substance, the molar free energies in two phases at equilibrium are equal, to the conclusion for mixtures, by analogy, that the chemical potentials are equal, without indulging in the proof, embodied in Eqs. (4.2)–(4.6). But different instructors will doubtless have different views on this matter.

I thank my colleague George Zografi for providing the initial stimulus that led to the writing of this book. The manuscript was accurately typed by Tina Rundle. Any errors (there are always errors) are my responsibility.

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0

REVIEW OF MATHEMATICS

0.1. INTRODUCTION

The extent of mathematics required in order to master the material in the professional pharmacy curriculum is really quite modest, comprising the basic operations of arithmetic and algebra; some fundamentals of plane geometry, analytic geometry, and calculus; and ideas from mathematical statistics. Nearly everything mathematical that you will need to know as a pharmacy student you have presumably already mastered in high school or college courses, except perhaps for the mathematical statistics content and the concept of partial differentiation (which will be treated in the following pages). You already know, or you once knew, about 95% of the material in this review (exclusive of the statistics section), which therefore should not present a difficult intellectual challenge. Although our immediate concern is to provide a basis for the mathematical needs of thermodynamics, this review goes beyond thermodynamics to include the mathematical methods useful in other parts of the curriculum.

Mathematics is treated by scientists as a tool or a language, and it is this attitude that you should adopt. Your goal in reviewing this material is to develop such a familiarity with the mathematical operations that you need not worry about them or even give much conscious thought to them. Such a capability will allow you to concentrate your attention on the new ideas being presented in your courses, whether they are physical chemical, pharmacokinetic, or biological. Not every bit of mathematics that you may encounter in your future studies will be treated here, but nearly all of it will be found here.

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Symbol	Meaning	
=	Equals; is equal to	
=	Is equivalent to	
\sim or \approx or \cong	Is approximately equal to	
≠	Is not equal to	
α	Is proportional to	
<	Is less than	
< < > 2	Is less than or equal to	
>	Is greater than	
\geq	Is greater than or equal to	
+	Plus	
_	Minus	
\times or \cdot	Times	
÷ or /	Divided by	
±	Plus or minus	
	The square root of	
v 	Therefore	
II	The absolute value of	

Table 0.1. Common mathematical symbols

As a first step in developing this review of essential mathematics as the language of science, Table 0.1 lists some standard mathematical symbols.

As you read the following pages, keep in mind that this is a review, not a logical development of the subject. Results are usually presented without derivation; you have already seen the derivations in your mathematics courses. Examples will draw on chemical concepts that, in many instances, will already be familiar to you, but a few ideas may appear that are new.

0.2. DIMENSIONS AND UNITS

Base and Derived Units. The value of a physical quantity is expressed as the product of a numerical value and a unit; thus

Physical quantity = numerical value \times unit

For example,

Distance
$$= 125 \text{ km}$$

Many systems of units have been used, and several systems are in use today, but in scientific work the standard system of units is the Système International (SI), occasionally varied with a few units from the older cgs system (IUPAC, 1993). There are seven SI *base units*, each of these being independent of the others. Table 0.2 lists the SI base units. No punctuation is used with symbols, nor are symbols pluralized.

Physical Quantity	Symbol for Quantity (Dimension)	Name of SI Unit	Symbol for SI Unit
Length	l	meter	m
Mass	m	kilogram	kg
Time	t	second	S
Electric current	Ι	ampere	А
Temperature	Т	kelvin	Κ
Amount of substance	n	mole	mol
Luminous intensity	$I_{ m v}$	candela	cd

Table 0.2. SI base units

Table 0.3. So	ome SI deriv	ved units
---------------	--------------	-----------

Physical Quantity	Defining Relationship	Derived SI Unit	
Area	$l \times l$	m ²	
Volume	$l \times l \times 1$	m ³	
Velocity	Distance per second	${ m ms^{-1}}$	
Density	Mass per volume	$kg m^{-3}$ mol m ⁻³	
Concentration	Amount per volume	$mol m^{-3}$	

All other units are called *derived units*; these can be obtained by appropriate algebraic combination of the base units. Table 0.3 gives a few derived units to illustrate the manner in which they are formed. Of course, in common practice we make wide use of traditional alternatives. For example, we may express area in square centimeters (cm^2) if this is convenient to the problem. Density is usually expressed in grams per milliliter $(g m L^{-1})$; and concentration is expressed in moles per liter $(mol L^{-1})$, which is sometimes written mol dm⁻³.

A few important physical quantities have special names and symbols for their units; a selection of these is shown in Table 0.4. The expression of a derived unit in terms of base units, as illustrated in Table 0.3, is obtained from the appropriate physical law or definition. For example, from Newton's law of motion, we have

 $Force = Mass \times Acceleration$

Physical Quantity	SI Unit	SI Symbol	Base Unit Expression
Frequency	hertz	Hz	s^{-1}
Force	Newton	Ν	$m kg s^{-2}$
Pressure	pascal	Ра	$Nm^{-2} \equiv m^{-1}kgs^{-2}$
Energy, heat, work	joule	J	$N m \equiv m^2 kg s^{-2}$
Electric charge	coulomb	С	A s
Electric potential	volt	V	$JC^{-1} \equiv m^2 kg s^{-1} A^{-1}$
Electric resistance	ohm	Ω	$VA^{-1} \equiv m^2 kg^{-1} s^3 A^2$

Table 0.4. Some named derived units

and

Acceleration
$$= \frac{\text{Velocity}}{\text{Time}} = \frac{\text{Distance}}{\text{Time}^2}$$

so, in terms of SI units we have

$$N \equiv m \, kg \, s^{-2}$$

Similarly,

$$Pressure = \frac{Force}{Area}$$

so

$$Pascal \equiv N m^{-2}$$

Also

Work = Force \times Distance

so

$Joule\equiv N\,m$

Not all scientists restrict themselves to SI units, and a few quantities are occasionally written in units from the cgs system. Moreover, the published scientific literature obviously makes use of older units, and so it is essential to be able to interconvert units of the several systems. Table 0.5 gives the relationships between the most important of these.

Table 0.5. Some cgs units

 Unit	SI Equivalent
	Si Equivalent
1 g (gram)	10^{-3}kg
1 mL (milliliter)	10^{-3} L
1 L (liter)	$10^{-3} \mathrm{m}^3$
1μ (micron)	$10^{-6} \mathrm{m}$
1 Å (angstrom)	$10^{-10} \mathrm{m} (10^{-8} \mathrm{cm})$
1 atm (atmosphere)	101,325 Pa
1 dyn (dyne)	10^{-5} N
1 erg	$10^{-7} \mathrm{J}$
1 cal (calorie)	4.184 J
1 D (debye)	$3.336 \times 10^{-30} \text{ C m}$

Submultiple	Prefix	Symbol	Multiple	Prefix	Symbol
10^{-1}	deci	d	10	deca	da
10^{-2}	centi	с	10 ²	hecto	h
10^{-3}	milli	m	10 ³	kilo	k
10^{-6}	micro	μ	10^{6}	mega	М
10^{-9}	nano	n	10 ⁹	giga	G
10^{-12}	pico	р	10 ¹²	tera	Т
10^{-15}	femto	f	10^{15}	peta	Р
10^{-18}	atto	а	10^{18}	exa	Е
10^{-21}	zepto	Z	10^{21}	zetta	Ζ
10^{-24}	yocto	У	10^{24}	yotta	Y

Table 0.6. Submultiple and multiple prefixes

It is often convenient to represent quantities in terms of multiples or submultiples of SI units. Table 0.6 gives the prefixes and symbols for these multiplicative operations.

Quantity Algebra. We earlier wrote this equation:

Physical quantity = Numerical value
$$\times$$
 Unit (0.1)

We now assert that each of the three quantities in this equation can be treated as an algebraic quantity and manipulated by the rules of algebra. Thus Eq. (0.1) could be written as

$$\frac{\text{Physical quantity}}{\text{Unit}} = \text{Numerical value}$$
(0.2)

This equation is particularly convenient because, since the right-hand side is a pure number (it has no dimensions), so is the left-hand side. To illustrate this with an example, let us write

$$T = 273.15 \,\mathrm{K}$$

in the form of Eq. (0.1). Transforming to the form of Eq. (0.2) gives

$$\frac{\mathrm{T}}{\mathrm{K}} = 273.15$$

We will call this method of manipulating units *quantity algebra*. It is particularly convenient for designing headings of table columns and for labeling the axes of figures. We will make use of the quantity algebra when converting from one unit to another.

Conversion of Units. We often find it convenient or necessary to convert from one system of units to another. Setting up a proportion is one method for converting units.

Example 0.1. Convert an energy change of 125 kJ mol^{-1} to kcal mol⁻¹. From Table 0.5 we have the essential relationship

$$1 \text{ cal} = 4.184 \text{ J}$$

Multiplying each side by 10^3 gives

$$1 \text{ kcal} = 4.184 \text{ kJ}$$

Now we form the proportion from statements:

Since 1 kcal = 4.184 kJx kcal = 125 kJ

or

$$\frac{1}{x} = \frac{4.184}{125}$$

 $x = 29.9$

Therefore 125 kJ mo1^{-1} is equal to $29.9 \text{ kcal mol}^{-1}$.

Quantity algebra offers a general procedure for the interconversion of units. This is the method: Multiply the quantity whose unit must be converted by one or more quotients, each being equal to the pure number one, but having the units needed to get the job done.

Example 0.2. Convert a wavelength of 560 nm to angstroms.

We know (Table 0.6) that $1 \text{ nm} = 10^{-9} \text{ m}$, so it follows that $1 \text{ nm}/10^{-9} \text{ m} = 1$. Similarly, from Table 0.5, $1 \text{ Å} = 10^{-10} \text{ m}$, so $1 \text{ Å}/10^{-10} \text{ m} = 1$. Therefore we can write

$$560 \,\mathrm{nm} \left(\frac{10^{-9} \mathrm{m}}{1 \,\mathrm{nm}}\right) \left(\frac{1 \,\mathrm{\mathring{A}}}{10^{-10} \mathrm{m}}\right) = 5600 \,\mathrm{\mathring{A}}$$

The method works because each quantity in parentheses is equal to 1, and we know that we can multiply fearlessly by 1. These quantities in parentheses are chosen so that the units that we do not want will cancel, and the units that we want will remain.

Example 0.3. The surface tension of water at 25° C is 71.8 erg cm^{-2} . Convert this to SI units:

$$71.8 \frac{\text{erg}}{\text{cm}^2} \left(\frac{1 \text{ J}}{10^7 \text{ erg}}\right) \left(\frac{1 \text{ N m}}{1 \text{ J}}\right) \left(\frac{10^2 \text{ cm}}{1 \text{ m}}\right) = 71.8 \times 10^{-3} \text{ N m}^{-1}$$

This is usually written 71.8 mN m^{-1} . Note in this multiplication how the unitary multipliers are oriented so as to achieve the desired cancellations. Incidentally, 71.8 erg cm^{-2} also equals 71.8 dyn cm^{-1} , because 1 erg = 1 dyn cm.

Extensive and Intensive Properties. A physical property whose magnitude is additive is called an *extensive property*; its magnitude depends on the extent (size) of the system. Mass, volume, and energy are examples of extensive properties.

A property whose magnitude is independent of the size of the system is called an *intensive property*. Temperature, pressure, and concentration are intensive properties.

An extensive property can be converted to an intensive property by dividing it by a mass or an amount of substance, thus placing it on a per unit basis. If an extensive property is divided by mass, the adjective *specific* is often used to describe it. Usually the mass unit gram (g) is used, so the specific property refers to the quantity per gram. For example, the volume V is an extensive property, but if V is divided by the mass, we get the *specific volume* (the volume per gram), which is intensive.

If we divide V by the amount of substance in moles, we have V/n, which is called the *molar volume*, and is interpreted as the volume per mole.

Example 0.4. The density of acetone is 0.788 at 25°C. Calculate the specific volume and the molar volume of acetone.

The units of density, which are $g m L^{-1}$ (or $g cm^{-3}$), are not always written out. The specific volume is simply the reciprocal of the density, as we can deduce from its units.

Specific volume =
$$\left(\frac{1 \text{ mL}}{0.788 \text{ g}}\right) = 1.269 \text{ mL g}^{-1}$$

Molar volume = $\left(\frac{1.269 \text{ mL}}{\text{g}}\right) \left(\frac{58.08 \text{ g}}{1 \text{ mol}}\right) = 73.7 \text{ mL mol}^{-1}$

These two statements have the same meaning:

- 1. The heat of solution of succinyl sulfathiazole is $12.0 \text{ kcal mol}^{-1}$.
- 2. The molar heat of solution of succinyl sulfathiazole is 12.0 kcal.

Dimensional Consistency. We have seen that units can be treated algebraically in that they undergo division and multiplication just as do numerical values. Units have two additional characteristics of great importance when carrying out calculations:

- 1. In what may appear to be a disagreement with the assertion that units can be treated algebraically, we note that when we add or subtract physical quantities, they must possess the same units, but the units themselves do not add or subtract.
- 2. When using or deriving physicochemical equations, the left-hand and righthand sides of the equation must have the same units.

Example 0.5. In Chapter 14 we will encounter this equation:

$$S_t = s_0 + \frac{K_{11}s_0L_t}{1 + K_{11}s_0}$$

We do not need to consider the detailed meaning of the equation at this point, because our present interest is only in its dimensional nature. (Indeed, this example will show how we can even bypass the detail and yet learn something useful). With the information that the quantities S_t , s_0 , and L_t are all molar concentrations, and that the equation is dimensionally consistent (point 2 above), let us determine the units of K_{11} .

Evidently, S_t (in mol L⁻¹, or M) is given by the sum of s_0 (also in M) and the final term, whose units must, by point 1 above, be M. Consider the denominator of this term. Since 1 is dimensionless, the product $K_{11}s_0$ must be dimensionless, which requires that the units of K_{11} be M⁻¹. Does this work out in the numerator? Yes, it does, because it yields (M⁻¹)(M)(M) = M, as required by dimensional consistency.

Dimensional consistency in equations describing physical systems is a necessary condition for their validity, but it is not a sufficient condition. Incidentally, if approximations are introduced into equations, the approximations must not alter the dimensional consistency.

0.3. LOGARITHMS AND EXPONENTS

Definition and Properties. Suppose we have the power function of Eq. (0.3):

$$a = b^x \tag{0.3}$$

We define¹ the logarithm of a to the base b by

$$\log_b a = x \tag{0.4}$$

There are only two logarithmic bases (values of *b*) in common use, namely, b = 10 (giving *Briggsian logarithms*) and b = e (giving *natural logarithms*). Briggsian logarithms are denoted *log*, whereas natural logarithms are denoted *ln*. An important property² of all logarithms is stated by Eq. (0.5):

$$\log_b b = 1 \tag{0.5}$$

It follows that

$$\log 10 = 1$$
 (0.6)

$$\ln e = 1 \tag{0.7}$$

¹ Equation (0.4) is not the standard definition, which requires calculus; rather, it is a consequence of the standard definition.

² From Eq. (0.3), if $b = b^x$, x = 1, so $\log_b b = 1$.

Another property³ is given by Eqs. (0.8) and (0.9):

$$\log 1 = 0 \tag{0.8}$$

$$\ln 1 = 0$$
 (0.9)

The number e is extremely important in mathematics and science. It has the value 2.718...

At one time, Briggsian logarithms were widely used to carry out arithmetic operations, but with electronic calculators and computers this use is obsolete. However, the base 10 log function continues to be indispensable in the sciences, for two reasons: (1) Some important physicochemical concepts (such as pH) are defined in terms of Briggsian logarithms and (2) logarithms to the base 10 have the convenient property of revealing order-of-magnitude changes or differences at a glance, For example, $\log 10^2 = 2.00$, $\log 10^3 = 3.00$, and so on.

Let us obtain a relationship by means of which we can interconvert Briggsian and natural logarithms of the same number. Let a be the number, and write

$$\ln a = c \log a \tag{0.10}$$

where we want to find the conversion factor c. Suppose for convenience (any number would do) that we set a = 10. From Eq. (0.10), we obtain

$$c = \frac{\ln 10}{\log 10} = \frac{2.303\dots}{1} \tag{0.11}$$

where the numerator is found by means of an electronic calculator, and the denominator is from Eq. (0.6). Therefore we can interconvert ln and log values by Eq. (0.12):

$$\ln a = 2.303 \log a$$
 (0.12)

Operations with Logarithms. Suppose we have both Eqs. (0.13) and (0.14):

$$a = b^x \tag{0.13}$$

$$c = b^{\mathsf{y}} \tag{0.14}$$

We know from Eq. (0.4) that

 $\log_b a = x$ $\log_b c = y$

But from Eqs. (0.13) and (0.14) we can also write

$$ac = b^x \cdot b^y = b^{x+y}$$

³ From Eq. (0.3), if a = 1 and b = 10, then x = 0, so log 1 = 0.

It follows that

$$\log_h ac = \log_h a + \log_h c \tag{0.15}$$

Thus, the logarithm of a product of two numbers is equal to the sum of the logarithms of the individual numbers. This very valuable result permits several other relationships to be derived; these are stated as Eqs. (0.16)–(0.19). Although these are written in terms of log, exactly analogous equations can be given for ln:

$$\log pq = \log p + \log q \tag{0.16}$$

$$\log \frac{p}{q} = \log p - \log q \tag{0.17}$$

$$\log \frac{1}{q} = -\log q \tag{0.18}$$

$$\log p^n = n \log p \tag{0.19}$$

Example 0.6. It is convenient to define, as a measure of the acidity of an aqueous solution, the pH by

$$pH = -log [H^+]$$

where $[H^+]$ is the hydrogen ion concentration in mol L⁻¹. This means that⁴

$$\mathrm{pH} = \log \frac{1}{[H^+]}$$

(a) Calculate the pH if $[H^+] = 5.00 \times 10^{-3} M$.

$$pH = -\log 5.00 \times 10^{-3}$$

= -(log 5.00 + log 10^{-3})
= -(0.70-3.00)
= 2.30

The pH is a positive number provided that $[H^+] < 1$ M. Seldom is it justifiable to express pH values beyond the second decimal place (i.e., 0.01 pH unit).

⁴ We can only take the logarithm of a pure number, thus this definition of pH appears to be invalid (and so shortly after we have considered dimensional consistency!). One way out of this dilemma is to say that we are really defining the pH as

$$pH = -\log([H^+]/M)$$

because the quantity in parentheses is dimensionless.

(b) What is the hydrogen ion concentration if pH = 8.67?

$$pH = 8.67$$

-log [H⁺] = 8.67
log [H⁺] = -8.67 = -9.00 + 0.33
[H⁺] = 2.14 × 10⁻⁹ M

This last step is called "taking the antilogarithm." It is the reverse of taking the logarithm, and it is easily accomplished on the electronic calculator. A useful check on the calculation is to convert the $[H^+]$ back to pH. Another check is to be sure that the order of magnitude is reasonable; this means that the answer is as expected at least to within a factor of 10.

Operations with Exponents. The following identities follow from the properties of logarithms [actually we used Eq. (0.22) in order to derive Eq. (0.15)]:

$$a^0 = 1$$
 (0.20)

$$a^1 = a \tag{0.21}$$

$$a^u a^v = a^{u+v} \tag{0.22}$$

$$a^{-u} = \frac{1}{a^u} \tag{0.23}$$

$$\left(a^{u}\right)^{v} = a^{uv} \tag{0.24}$$

$$(ab)^u = a^u b^u \tag{0.25}$$

Scientific Notation. Many quantities in theoretical and experimental science are either extremely large or extremely small, so an exponential form of expressing them is convenient. Nearly always these are written as an integral power of 10, as shown in these examples:

$$51,000 \equiv 5.1 \times 10^4 \equiv 51 \times 10^3$$
$$0.00000417 \equiv 4.17 \times 10^{-6}$$

As seen in Example 0.6, this is a very convenient way in which to express a number whose logarithm is to be taken, because [as seen in Eq. (0.15)] the logarithm of a product is equal to the sum of the logs and because $\log 10^n = n \log 10 = n$. Thus

$$\log 4.17 \times 10^{-6} = (\log 4.17) - 6 = -5.38$$

0.4. ALGEBRAIC AND GRAPHICAL ANALYSIS

Setting Up Proportions. It often happens that we know three related items of information and seek a fourth member of the set. In order to solve for this fourth, unknown item, we write an equation. One way to do this is to express the relationships in terms of sentences, by means of which we can more readily detect analogous patterns. Then we replace the English words with mathematical symbols.

Example 0.7. How many moles of sodium chloride are contained in 5.00 g of NaCl?

The first of the following statements gives two pieces of known information; the second analogous statement contains the unknown.

58.45 g of NaCl corresponds to 1 mol

so

5.00 g of NaCl corresponds to how many moles?

Now we replace the words with symbols (and notice how the units cancel):

$$\frac{58.45}{5.00} = \frac{1}{x}$$
$$x = \frac{5.00}{58.45} = 0.0855$$

Generalizing this equation gives these important formulas:

$$mol = \frac{W(g)}{MW}$$
(0.26)

$$mmol = \frac{W(mg)}{MW}$$
(0.27)

Equation (0.27) is obtained from Eq. (0.26) by multiplying both sides by 10^3 . A millimole (mmol) is one-thousandth of a mole.

Example 0.8. A sample of aspirin weighing 305 mg was found by analysis to contain 294 mg of aspirin. What is its percent purity?

If the sample contained 305 mg, it would be 100% pure. Actually it contains 294 mg, so it is x% pure. Therefore

$$\frac{305}{294} = \frac{100}{x}$$

or 96.4% pure. Generalizing this result, we obtain

$$\% \text{ purity} = \frac{W_{\text{found}}}{W_{\text{taken}}} \times 100$$
 (0.28)

Example 0.9. Convert 75 min to hours.

If 60 min is equivalent to 1 h, then 75 min is equivalent to x h?

$$\frac{60}{75} = \frac{1}{x}$$
$$x = 1.25 \text{ h}$$

Ratios and Fractions. We are going to consider this subject in the context of chemistry rather than of pure mathematics. To clarify the distinction between a ratio and a fraction, consider the dissociation of a weak acid (like a carboxylic acid) in water:

$$HA \rightleftharpoons H^+ + A^-$$

Here we let HA represent any neutral weak acid, where A^- is the anion resulting from the dissociation. We define the *ratio* of anion to neutral acid by

$$\frac{[A^-]}{[HA]} = \text{ratio of anion to acid} = R_{A^-}$$

where square brackets signify molar concentrations. Similarly we have

$$\frac{[\text{HA}]}{[\text{A}^-]} = \text{ratio of acid to anion} = R_{\text{HA}}$$

These ratios play an important role in describing chemical equilibria.

There can also be situations in which we wish to define the *fraction* of anion in the mixture of A⁻ and HA. This fraction, which we will label F_{A^-} , is defined as

$$\frac{[\mathbf{A}^-]}{[\mathbf{H}\mathbf{A}] + [\mathbf{A}^-]} = F_{\mathbf{A}^-} = \text{fraction of anion}$$

Similarly the fraction present as the neutral acid is given by

$$\frac{[\text{HA}]}{[\text{HA}] + [\text{A}^-]} = F_{\text{HA}} = \text{fraction of acid}$$

By substitution it is easily found that

$$F_{\mathrm{A}^-} + F_{\mathrm{HA}} = 1$$

This is a general property of fractions as defined in this way; they sum to unity. As a consequence, if the whole is divided into n fractions, only n-1 of these are independent (i.e., capable of independent variation).

Example 0.10. A solution made up to contain a *total* concentration (also called an *analytical concentration*) of weak acid 3.50×10^{-3} M is found to have an anion concentration of 8.95×10^{-4} M. Calculate the fractions of solute in the anion and neutral forms.

We are told that the total concentration is 3.50×10^{-3} M. This is the sum [HA] + [A⁻], since the solute can exist in either or both forms, and no other. Hence

$$F_{\rm A^-} = \frac{[{\rm A}^-]}{[{\rm HA}] + [{\rm A}^-]} = \frac{0.895 \times 10^{-3}}{3.50 \times 10^{-3}} = 0.256$$

From the identity $F_{A^-} + F_{HA} = 1$, we immediately obtain $F_{HA} = 1 - 0.256 = 0.744$.

A fraction can be converted to a percentage through multiplication by 100, Eq. (0.29); see also Eq. (0.28).

$$Percent = 100 \times Fraction \tag{0.29}$$

Thus in this example 25.6% is in the form of the anion and 74.4% is in the form of the neutral acid.

Example 0.11. For the system in Example 0.10, calculate the ratios $[A^-]/[HA]$ and $[HA]/[A^-]$.

We know that $[A^-] = 0.895 \times 10^{-3} \text{ M}$ and $[HA] + [A^-] = 3.50 \times 10^{-3} \text{ M}$, so we find that $[HA] = 2.605 \times 10^{-3} \text{ M}$. Therefore

$$R_{\rm A^-} = \frac{[{\rm A}^-]}{[{\rm HA}]} = \frac{0.895}{2.605} = 0.344$$

The reciprocal gives us the other ratio:

$$R_{\rm HA^-} = \frac{[\rm HA]}{[\rm A^-]} = \frac{1}{0.344} = 2.91$$

Fractions must be less than (or equal to) unity, but a ratio can be greater than unity. Ratios and fractions (in this context) are dimensionless.

Students sometimes wonder why, in calculations like this (the dissociation of HA according to $HA \rightarrow H^+ + A^-$) we were able to ignore the H^+ produced in the reaction. The answer is that we are counting molecules or moles, not grams; each molecule of HA that dissociates yields one H^+ ion and one A^- ion. If we counted both ions, we would be double-counting, with the result that we would appear to be creating matter, which we know is impossible.

Solving Simultaneous Equations. From algebra we know that if we have n simultaneous independent equations in n unknowns, we can solve the equations to find all n unknowns. Usually, of course, we encounter the simplest case in which we

have one equation and one unknown. It is more interesting when we have two or more unknowns.

Let us continue to discuss our earlier example of the dissociation of weak acid HA to yield anion A^- . Often it happens that we know the total concentration and can measure the ratio of concentrations. From this information we seek the individual concentrations [HA] and [A⁻].

We have two unknowns, so we hope that we can write two independent equations. We begin by setting in equation form that which we know. We write

$$\frac{[A^-]}{[HA]} = R_{A^-} \tag{0.30}$$

$$[HA] + [A^{-}] = c_t \tag{0.31}$$

where we know (have numerical values for) R_{A^-} and c_t . Equations (0.30) and (0.31) are two independent equations in two unknowns.

There are several ways to solve simultaneous equations; the simplest is by algebraic substitution. Solve Eq. (0.30) for [A⁻] and substitute this into Eq. (0.31). From Eq. (0.30), [A⁻] = R_{A^-} [HA], so from Eq. (0.31) we get

$$[\mathrm{HA}] + R_{\mathrm{A}^{-}}[\mathrm{HA}] = c_t$$

We have reduced two equations with two unknowns to one equation with one unknown, for which we solve

$$[\mathrm{HA}] = \frac{c_t}{1 + R_{\mathrm{A}^-}}$$

Having found [HA], we can use Eq. (0.30) or (0.31) to find [A⁻].

The requirement that the equations be independent means that there is no possible way to obtain (to derive) one of them from the others; that is, each equation must contribute some additional information to the problem. For example, in the preceding case suppose that we only know c_t . There is no way from this knowledge alone that we could deduce the value of R_{A^-} , so we could not calculate individual values of [HA] and [A⁻]; we would have a single equation [Eq. (0.31)] and two unknowns.

The solution of simultaneous equations is usually easy. The difficult tasks are to analyze the problem so as to identify and write down the equations and to collect the necessary information so they can be solved numerically.

Solution of Quadratic Equations. Everyone learns in high school algebra how to solve a quadratic equation, that is, an equation in which the unknown quantity appears to the second power. In its "standard" form a quadratic equation can be written

$$ax^2 + bx + c = 0 (0.32)$$

where x is the unknown quantity and a, b, c are known quantities; a, b, and c can be viewed as constants in the particular situation. The solution of Eq. (0.32) is given by Eq. (0.33).

$$x = \frac{-b \pm \sqrt{b^2 - 4ac}}{2a} \tag{0.33}$$

This equation should be memorized. It shows that there are two solutions to Eq. (0.32), corresponding to the use of either the plus or the minus sign in Eq. (0.33). Usually only one of these solutions has physical significance; the other solution can be ignored. By "physical significance" we mean that it makes sense in the context of the physical situation. For example, if *x* is a concentration, obviously a negative value has no physical significance.

Example 0.12. We will continue to use our example of the dissociation of weak acid HA according to

$$HA \rightleftharpoons H^+ + A^-$$

Let us define an equilibrium constant by

$$K_a = \frac{[\mathrm{H}^+][\mathrm{A}^-]}{[\mathrm{HA}]} \tag{0.34}$$

We also have the equation giving the total solute concentration:

$$c_t = [HA] + [A^-]$$
 (0.35)

We now observe that each molecule of HA that dissociates yields one H^+ and one A^- , suggesting (this is not quite true, but is nearly always a very good approximation) that we can write

$$[A^{-}] = [H^{+}]$$

Making this substitution in Eq. (0.34) gives

$$K_a = \frac{[\mathrm{H}^+]^2}{c_t - [\mathrm{H}^+]} \tag{0.36}$$

This is a quadratic equation. First we rearrange it to place it in the standard form

$$[\mathrm{H}^+]^2 + K_a[\mathrm{H}^+] - K_a c_t = 0 \tag{0.37}$$

Comparing Eqs. (0.32) and (0.37) gives these identities:

$$a = 1$$

$$b = K_a$$

$$c = K_a c_t$$

$$x = [H^+]$$

The solution is accordingly

$$[H^+] = \frac{-K_a \pm \sqrt{K_a^2 - 4K_a c_t}}{2} \tag{0.38}$$

Example 0.13. Calculate the pH in a solution that is 0.010 M with respect to a weak acid if $K_a = 1.00 \times 10^{-5}$.

We use Eq. (0.38):

$$[H^+] = \frac{-1.0 \times 10^{-5} \pm \sqrt{1.0 \times 10^{-10} + (4 \times 10^{-5})(1 \times 10^{-2})}}{2}$$
$$= \frac{-1.0 \times 10^{-5} \pm \sqrt{1.0 \times 10^{-10} + 4 \times 10^{-7}}}{2}$$
$$= \frac{-1.0 \times 10^{-5} \pm \sqrt{0.01 \times 10^{-8} + 40 \times 10^{-8}}}{2}$$
$$= \frac{-1.0 \times 10^{-5} \pm \sqrt{40.01 \times 10^{-8}}}{2}$$
$$= \frac{-1.0 \times 10^{-5} \pm 6.325 \times 10^{-4}}{2}$$
$$= \frac{-1.0 \times 10^{-5} \pm 63.25 \times 10^{-4}}{2}$$
$$= 31.125 \times 10^{-5} = 3.11 \times 10^{-4}$$

So $pH = -log[H^+] = 3.51$. Note that the solution employing the negative sign has been rejected.

Approximations. Scientists, unlike mathematicians, are often satisfied with approximate rather than exact solutions. The level of approximation that is acceptable is determined by practical criteria and is not completely arbitrary. If we cannot experimentally detect or measure an effect, we may feel justified in neglecting it. Or how we use the result may determine the acceptable level of approximation; for example, if we are to measure the pH of Lake Mendota water, we may be satisfied

with a result accurate to ± 0.1 pH unit (for various practical reasons), whereas if we are measuring the equilibrium constant of a weak acid, we will surely hope for measurements accurate to ± 0.01 pH unit. This kind of thinking is then extended to the equations that enable us to describe and interpret the measurements. If we make an approximation in an equation, it is no longer exact in a mathematical sense,⁵ but it may be acceptably accurate in a scientific sense.

Example 0.14. Calculate the pH in a solution that is 0.010 M with respect to a weak acid if $K_a = 1.00 \times 10^{-5}$.

This is the same problem for which we obtained an accurate solution in Example 0.13. In that calculation we used the equation

$$K_a = \frac{[\mathrm{H}^+]^2}{c_t - [\mathrm{H}^+]} \tag{0.39}$$

Now suppose that the hydrogen ion concentration is much smaller than is the total concentration c_t . Then we can reasonably make this approximation:

$$c_t - [\mathrm{H}^+] \approx c_t$$

Making this substitution in Eq. (0.39) gives

$$K_a \approx \frac{\left[\mathrm{H}^+\right]^2}{c_t}$$

or

$$[\mathrm{H}^+] \approx \sqrt{K_a c_t} \tag{0.40}$$

Applying Eq. (0.40) to our problem, we get

$$[\mathrm{H^+}] = \sqrt{(1 \times 10^{-5})(1 \times 10^{-2})}$$
$$= \sqrt{1 \times 10^{-7}}$$
$$= 3.16 \times 10^{-4} \,\mathrm{M}$$

or pH = 3.50. Our approximation has led to a very simple solution, and comparison with Example 0.13 shows the level of error that has been introduced.

When is it acceptable to use such an approximation? This depends on those factors already mentioned. In the present example we might examine the quantity $c_t - [H^+]$ to

⁵ The equation probably never was truly exact in a physicochemical sense anyway because of the complexity of chemical systems.

see if $[H^+]$ is indeed much smaller than c_t (because that is the assumption on which the approximation is based). In Example 0.14 we can compare these numbers:

Exact:
$$c_t = 1.00 \times 10^{-2} - 3 \times 10^{-4} = 0.97 \times 10^{-2}$$

Approximate: $c_t - [\text{H}^+] \sim c_t = 1.00 \times 10^{-2}$

Therefore the approximation introduces about a 3% error into the denominator of Eq. (0.39). The error in the final answer is even smaller than this, and for most purposes the approximate result would be acceptable. This is the practical point of view that is adopted, and it can be seen that decisions of this sort depend greatly on experience with making calculations and with interpreting experimental results.

Graphical Properties of Linear Functions. Let us begin this treatment by writing this linear equation in a standard form:

$$y = mx + b \tag{0.41}$$

We call this "linear" because y depends on x to the first power. The following terminology is used:

x, *y* are *variables m*, *b* are *parameters* (i.e., constants of the system)

More specifically, x is often called the *independent variable* and y the *dependent variable*. This terminology signifies that the value of x is under our control and that the value of y then depends on x.

If paired *x*, *y* data are related by Eq. (0.41), when plotted on graph paper they yield a straight line (which is another reason why we call Eq. (0.41) "linear"). Figure 0.1 shows how the plot is conventionally made. It is traditional to plot the *x* values on the horizontal axis (the abscissa) and the *y* values on the vertical axis (the ordinate).

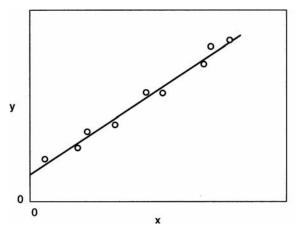


Figure 0.1. A typical straight-line plot of data points.

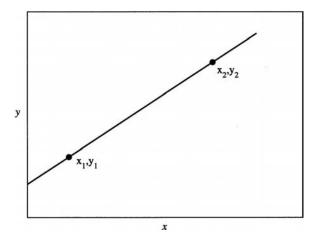


Figure 0.2. Illustrating the calculation of the slope of a straight line; see text.

Let us set x = 0 in Eq. (0.41); we get y = b; that is, b is the value of y when x = 0. We call this the *intercept on the y axis*. The parameter m describes the steepness with which y increases (or decreases) as x increases; m is called the *slope*. Evidently, if we know the values of m and b, we know everything there is to know about Eq. (0.41).

Now let us review the calculation of m, the slope. In Fig. 0.2, two points, on the line, having coordinates (x_1, y_i) and (x_2, y_2) , are identified. We write the equation of the line for each of these points (for the equation is satisfied at every point on the line):

$$y_1 = mx_1 + b$$
$$y_2 = mx_2 + b$$

Now subtract y_1 from y_2 and solve for *m*:

$$m = \frac{y_2 - y_1}{x_2 - x_1} \tag{0.42}$$

This equation allows us to calculate the slope of any straight line. The slope *m* can be positive (*y* increases as *x* increases), negative (*y* decreases as *x* increases), or zero (*y* does not depend on *x*). We can read *b* from the graph as the value of *y* when x = 0; or more accurately and sometimes more conveniently (because sometimes the plot doesn't include the x = 0 region), *b* can be calculated from Eq. (0.41), since we now know *m*. Simply choose any point on the line and substitute the corresponding numerical values into b = y - mx.

Here are two interesting special cases of Eq. (0.41).

1. m = 0. Then the line is horizontal, parallel to the *x* axis, since substituting this value of *m* into Eq. (0.41) gives y = b.

Sodium Salicylate Concentration/(M)	Theophylline Concentration/(M)
0	0.0387
0.0252	0.0517
0.0504	0.0622
0.1009	0.0860
0.1261	0.0946
0.1514	0.1094
0.2018	0.1288

Table 0.7. Solubility of the ophylline in water at $25.0^\circ C$ as a function of added sodium salicy late

2. b = 0. Then Eq. (0.41) yields

$$y = mx \tag{0.43}$$

showing that the line passes through the origin at y = 0, x = 0.

Example 0.15. Table 0.7 lists experimental data (Cohen and Connors, 1967) giving the solubility of the drug theophylline (in water at 25° C) as a function of the concentration of added sodium salicylate. Analyze the data.

The first thing to do is to plot the data. Use graph paper having fine enough divisions to preserve the accuracy of the data; 1-mm divisions usually work well, whereas 1/4'' divisions are too coarse. Figure 0.3 shows the plot of the data. Since the sodium salicylate concentration was set by the experimenter, we consider it to be the independent variable *x*.

The data points appear to describe a straight line. There is a theoretical reason for this, but we can view it purely experimentally as a pleasingly simple result. Note that

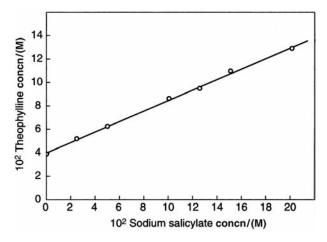


Figure 0.3. Plot of the data in Example 0.10.

the data points show some small but significant "scatter" about the line that has been drawn. This scatter introduces ambiguity into the question of how best to draw the line. Later, in Section 0.6, we will deal with this issue of the "best straight line," but for the present it is sufficient to note that we draw a line that appears to approximately balance the deviations of the points above and below the line.

Next we calculate the slope of the line. Because of the scatter seen in the experimental points, our two points for the slope calculation will be taken from the line itself, because it is the slope of the line that we want.⁶ Any two points will do, but they should be far enough apart to yield an accurate result. Points corresponding to $x_1 = 0.060$ and $x_2 = 0.160$ were chosen. Reading directly from Fig. 0.3 gives

At $x_1 = 0.060$ M, $y_1 = 0.0669$ M At $x_2 = 0.160$ M, $y_2 = 0.1122$ M

These numbers are used in Eq. (0.42):

$$m = \frac{0.1122 - 0.0669}{0.160 - 0.060} = 0.453$$

Now we calculate *b* using (for convenience, because we already have the values) point x_1, y_1 :

$$b = y - mx$$

= 0.0669 - (0.453)(0.060)
= 0.0397 M

Note that *b* has the units of *y* whereas *m* has the units of y/x. Also observe that *b* as calculated from the line is slightly different from the experimental value of 0.0387 M when x = 0. Such discrepancies are common in scientific work. They add interest to the interpretation of experimental data.

We can now state that the set of data given in Example 0.15 can be described by the equation

$$y = 0.453x + 0.0397$$

where *x* is the sodium salicylate concentration in M, *y* is the theophylline concentration in M, and the equation is valid over the range x = 0 - 0.20 M. The physical interpretation is that the presence of sodium salicylate increases the theophylline solubility in a linear manner. The chemical interpretation of this phenomenon has to do with the mutual binding interaction between theophylline and the salicylate ion (Cohen and Connors, 1967). A more detailed explanation of this interaction, along with a rigorous treatment of the binding between two or more drug molecules, can be found in Chapter 14.

⁶ Here is the attitude we have adopted. The experimental points have led us to the line as the best interpretation of the dependence of the *y* values on the *x* values, so henceforth we base our interpretation on the line and not on the data points.

Linearization of Nonlinear Functions. The experimental data of Example 0.15 consisted of molar concentrations, and we found empirically that a plot of theophylline solubility as a function of sodium salicylate concentration gave a straight line. We could conclude that the concentrations were linearly related, and we obtained the parameters of the straight-line relationship.

It is not always this simple. Often, perhaps usually, the data are smoothly but nonlinearly related. In such circumstances we may be able to carry out a mathematical transformation that converts a nonlinear function to a linear function. This is often very desirable, because of the simplicity with which we can describe straight lines. We may have theoretical reasons to expect certain transformations to work in this way, or we may just try different plotting forms empirically, hoping that we will generate a straight line. Here are some of the most common transformations.

1. Exponential Functions. These have the general form

$$y = ae^{bx} \tag{0.44}$$

where, as before, x and y are variables, a and b are parameters, and e is the base of natural logarithms. We take the natural logarithm of both sides:

$$\ln y = \ln a + bx \tag{0.45}$$

This operation has converted the nonlinear function Eq. (0.44) to the linear function Eq. (0.45), which has the form of Eq. (0.41) for a straight line. In other words, if Eq. (0.44) describes the data, a plot of $\ln y$ against *x* should be linear with slope value *b* and intercept (on the vertical axis when x = 0) equal to $\ln a$. This is often called a *semilog plot*. It is widely used in kinetic studies.

2. Power Functions

$$y = ax^b \tag{0.46}$$

Again we take the logarithm:

$$\ln y = \ln a + b \ln x \tag{0.47}$$

A plot of $\ln y$ against $\ln x$ will be linear if Eq. (0.46) describes the data. This is called a *log–log plot*.

3. Polynomial Functions

$$y = a + bx + cx^2 \tag{0.48}$$

This equation is a quadratic function, which can be linearized as in

$$\frac{y-a}{x} = b + cx \tag{0.49}$$

Evidently a plot of (y - a)/x against x will give a straight line if Eq. (0.48) describes the data. Of course, the parameter a must be known in order to construct this plot.

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4. *Rectangular Hyperbola* Any equation having the following form is called a *rectangular hyperbola*:

$$y = \frac{ax}{1+bx} \tag{0.50}$$

This is nonlinear when plotted as *y* against *x*. We can linearize it in several ways. Taking the reciprocals of both sides gives

$$\frac{1}{y} = \frac{1}{ax} + \frac{b}{a} \tag{0.51}$$

so a plot of 1/y against 1/x (this is called a *double-reciprocal plot*) will be linear. You may encounter this plot in your study of enzyme kinetics, where it is called the *Lineweaver–Burk* plot. Algebraic manipulation also leads to the following equation, another linear plotting form:

$$\frac{x}{y} = \frac{b}{a}x + \frac{1}{a} \tag{0.52}$$

In this version, x/y is plotted against x. A third linear transformation can be obtained; this is shown as

$$\frac{y}{x} = a - by \tag{0.53}$$

A plot of y/x against y is linear. Notice that the slope is negative. In protein binding studies a plot according to Eq. (0.53) is called a *Scatchard plot*. The rectangular hyperbola arises naturally in mathematical descriptions of chemical equilibria.

It will be evident that a facility for recognizing the standard linear form y = mx + b (where the definitions of the variables and parameters depend on the particular system) is invaluable in seeking linear forms from nonlinear functions.

Example 0.16. Table 0.8 gives kinetic data for the decomposition of nitrogen pentoxide in carbon tetrachloride solution at 45° C.⁷ Analyze the data.

The experimental data consist of the concentration of $N_2O_5(c)$ as a function of time (*t*). Figure 0.4 is a direct plot of *c* (as the dependent variable) against *t*. The plot is obviously nonlinear. As it happens, we have some theoretical guidance for this reaction (and for many other processes that follow the same functional dependence

 7 See Daniels and Alberty (1955), p. 323). The reaction yields oxygen and $N_2O_4,$ which exists in equilibrium with $NO_2.$

$$N_2O_5 \rightarrow N_2O_4 + \frac{1}{2}O_2$$
$$N_2O_4 \rightleftharpoons 2NO_2$$

t/(s)	c/(M)	$\log \left[c/(\mathrm{M}) \right]$
0	2.33	0.367
184	2.08	0.318
319	1.91	0.281
526	1.67	0.223
867	1.36	0.134
1198	1.11	0.045
1877	0.72	-0.143
2315	0.55	-0.260
3144	0.34	-0.469

Table 0.8. Kinetics of N₂O₅ decomposition at 45°C in CCl₄

on time). We anticipate that the progress of the reaction is described by Eq. (0.54), where c_0 is the reactant concentration when t = 0 (the beginning of the reaction) and *c* is the concentration at any time *t*:

$$c = c_0 e^{-kt} \tag{0.54}$$

The parameter *k* is called the *rate constant*.

We recognize Eq. (0.54) as an exponential function, which we can linearize by taking the logarithm

$$\ln c = \ln c_0 - kt \tag{0.55}$$

or

 $\log c = \log c_0 - \frac{kt}{2.303} \tag{0.56}$

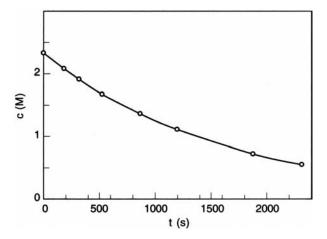


Figure 0.4. Direct plot of data in Example 0.11.

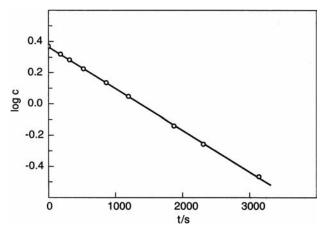


Figure 0.5. Semilog plot of data in Example 0.16.

We can use either Eq. (0.55) or Eq. (0.56). The table of data shows values of log c, and Fig. 0.5 is the plot of log c against t as suggested by Eq. (0.56). Evidently we have succeeded in linearizing the data, which we interpret to mean that Eq. (0.54) describes the reaction.

From Fig. 0.5 we calculate the slope as follows:

Slope =
$$\frac{-0.167 - (+0.100)}{2000 - 1000}$$

= $-2.67 \times 10^{-4} \text{ s}^{-1}$

Observe the sign and units of the slope. Now, from Eq. (0.56) we can write

$$\text{Slope} = -\frac{k}{2.303}$$

so

$$k = 6.15 \times 10^{-4} \, \mathrm{s}^{-1}$$

0.5. DEALING WITH CHANGE

Change in Physical and Chemical Processes. We are often interested in some sort of process that has happened, is happening, or is expected to happen; in other words, something changes. We need to be able to describe and analyze such changes. Table 0.9 shows the kinds of processes that may concern us.

Suppose that we want to specify quantitatively the value of some property P that changes as the physical or chemical system passes from its initial state A to its

Solid	melting (fusion) crystallization	Liquid
Liquid	condensation	Gas (vapor)
Solid	sublimation	Gas
Solvent + solid	dissolution	Solution
Solute in solvent A	partitioning	Solute in solvent B
Gas at volume 1	expansion	Gas at volume 2
System at temperature 1	heating	System at temperature 2
Reactants	chemical reaction	Products

Table 0.9. Types of physicochemical processes

final state Z. The process itself is symbolized $A \rightarrow Z$, and the change (or increment) in P is defined as

Change in P = (Value of P in final state) - (Value of P in initial state)

or

$$\Delta P = P_Z - P_A \tag{0.57}$$

If *P* increases during the process, then $P_Z > P_A$ and ΔP is positive; if *P* decreases, then ΔP is negative.

Incremental and Differential Change. We have defined an increment in a quantity by Eq. (0.57). Let us now revert to our study of a straight-line function

$$y = mx + b \tag{0.58}$$

and recall how we calculated the value of the slope m. We chose two points on the line (see Fig. 0.2) and found that m is equal to the function in Eq. (0.59):

$$m = \frac{y_2 - y_1}{x_2 - x_1} \tag{0.59}$$

If we think of larger positive values of x (values increasing toward the right on the abscissa) as "later," then we can identify the point (x_1, y_1) as an initial state and point (x_2, y_2) as a final state. Then we see that m can be written in our delta symbolism as

$$m = \frac{\Delta y}{\Delta x} \tag{0.60}$$

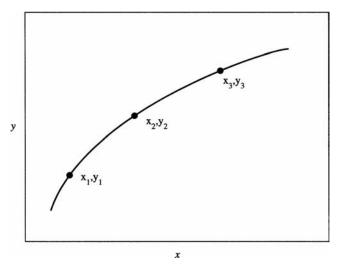


Figure 0.6. A curvilinear function y = f(x)

Carrying out the division on the right-hand side with numerical quantities yields a number equal to *m*. Since this result can be viewed as the number divided by 1, we interpret $m = \Delta y / \Delta x$ as the change in y per unit change in x. In Example 0.15 we found m = 0.453, which we interpret to mean that as x increases by one unit, y increases by 0.453 unit. Recall also that *m* can be negative; this would mean that y decreases as x increases.

Now the essence of a straight line is that its slope m is a constant, so that we would obtain the same value for *m* no matter which two points, wherever they may be on the line, we choose for its calculation. But suppose the plot of *y* against *x* yields a curved (nonlinear) line, as in Fig. 0.6? Obviously if we calculate the slope value using points (x_1, y_1) and (x_2, y_2) , we will get a different value from that using points (x_2, y_2) and (x_3, y_3) . In fact, we have to ask if the concept of "slope" has meaning in this

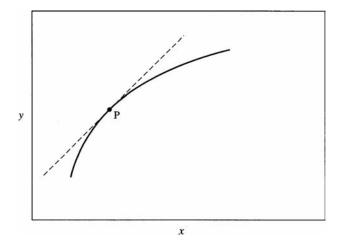


Figure 0.7. The slope at point *p* is the value of the tangent to the curve at *p*.

circumstance. The answer is that we can expand our definition of the slope to include such circumstances. We will now define the slope of any line *at a point p* as the value of the tangent to the line at *p*, as shown in Fig. 0.7.

How can we evaluate the numerical value of the slope at point p? We simply return to our original definition

$$m = \frac{\Delta y}{\Delta x}$$

and require that Δx (in the immediate vicinity of point *p*) be made smaller and smaller until it is so small that *m* approaches an essentially constant value; that is, making Δx still smaller doesn't sensibly alter the value of *m*. Now the ratio $\Delta y/\Delta x$ gives the slope (tangent) of the line at point *p*. We write this

$$m = \lim_{\Delta x \to 0} \frac{\Delta y}{\Delta x} \tag{0.61}$$

which is read "*m* is the limiting value of $\Delta y/\Delta x$ as Δx approaches zero." Of course, it would be cumbersome to write Eq. (0.61) repeatedly, so a new terminology is introduced, namely

$$m = \lim_{\Delta x \to 0} \frac{\Delta y}{\Delta x} = \frac{dy}{dx}$$
(0.62)

where dy and dx are individually referred to as *differentials* and the ratio dy/dx is called *the derivative of y with respect to x*.

We interpret the derivative dy/dx of any function y = f(x) as the slope of the function (when plotted in the usual manner). If the function is a straight line, dy/dx is a constant (its value does not depend on *x*), whereas for curved functions the slope dy/dx depends on (varies with) *x*. Thus dy/dx, for both straight and curved lines, is a measure of change.

Formulas for Derivatives. One of the interesting properties of derivatives is that the individual differentials can be treated as algebraic quantities. Thus we can write

$$\frac{dy}{dx}dx = dy \tag{0.63}$$

and

$$\frac{dy}{dx} = \frac{dy}{du}\frac{du}{dx} \tag{0.64}$$

Equation (0.64) is known as the "chain rule." Evidently we also have dx/dx = 1.

Table 0.10 collects some formulas [Eqs. (0.65)-(0.72)] for derivatives and differentials that will be found useful in scientific settings. In these formulas *u* and *v* are functions of *x*, *c* and *n* are constants, and *e* is the base of natural logarithms. The differential form is obtained from the derivative through multiplication by *dx*. Table 0.10 is a compact way of summarizing these formulas, and a more expanded

Derivative	Differential	Eq.
$\frac{dc}{dx} = 0$	dc = 0	(0.65)
$\frac{d(cu)}{dx} = c\frac{du}{dx}$	d(cu) = c du	(0.66)
$\frac{d(u+v)}{dx} = \frac{du}{dx} + \frac{dv}{dx}$	d(u+v) = du + dv	(0.67)
$\frac{d(uv)}{dx} = u\frac{dv}{dx} + v\frac{du}{dx}$	d(uv) = udv + vdu	(0.68)
$\frac{d(u/v)}{dx} = \frac{v(du/dx) - u(dv/dx)}{v^2}$	$d\frac{u}{v} = \frac{vdu - udv}{v^2}$	(0.69)
$\frac{du^n}{dx} = nu^{n-1}\frac{du}{dx}$	$d(u^n) = nu^{n-1}du$	(0.70)
$\frac{d\ln u}{dx} = \frac{1}{u}\frac{du}{dx}$	$d\ln u = \frac{du}{u}$	(0.71)
$\frac{de^u}{dx} = e^u \frac{du}{dx}$	$de^u = e^u du$	(0.72)

	Table 0.10.	Some	derivatives	and	differentials
--	-------------	------	-------------	-----	---------------

 ^{a}u , v, x are variables; c, n are constants.

form might express the results as follows, where we take Eq. (0.67) as an example:

Function:	y = u + v
Derivative :	$\frac{dy}{dx} = \frac{du}{dx} + \frac{dv}{dx}$
Differential:	dy = du + dv

Observe also that some entries can be derived from others; for example, Eq. (0.66) is a special case of Eq. (0.68), and Eq. (0.69) can be obtained from Eq. (0.68) by writing the quotient u/v as the product uv^{-1} .

All the derivatives in Table 0.10 are first derivatives. We can also define higher derivatives. Suppose that we define u = dy/dx; then

$$\frac{du}{dx} = \frac{d(dy/dx)}{dx} = \frac{d^2y}{dx^2}$$

is called a *second derivative*. A second derivative is a measure of change of the first derivative.

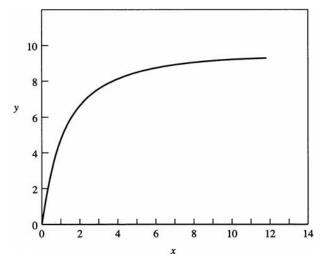


Figure 0.8. The rectangular hyperbula, Eq. 0.73, for the case a = 10, b = 1.

Example 0.17. Find the slope of the hyperbolic function

$$y = \frac{ax}{1+bx} \tag{0.73}$$

We want the slope dy/dx. Equation (0.73) is a quotient, so we use Eq. (0.69) combined with Eq. (0.66):

$$\frac{dy}{dx} = \frac{(1+bx)a - (ax)b}{(1+bx)^2} = \frac{a}{(1+bx)^2}$$
(0.74)

Thus the slope depends on x, as can be seen graphically in Fig. 0.8. As $x \to 0$, $dy/dx \to a$, a constant; this is the "limiting slope" or "initial slope," that is, the tangent to the curve at the point x = 0. On the other hand, when $x \to \infty$ [x becomes so large that the denominator of Eq. (0.73) becomes much greater than the numerator], $dy/dx \to 0$; in this circumstance the value of y becomes essentially independent of x.

Maxima, Minima, and Inflection Points. Figure 0.9 shows a function y = f(x) that passes through a maximum.

Picture how the slope dy/dx changes as x increases from left to right. This is indicated by the tangent lines. In the vicinity of the maximum, whose location is labeled x_{max} , the slope decreases as x approaches x_{max} from the left. At $x = x_{\text{max}}$, dy/dx = 0; then as x leaves x_{max} , moving to the right, dy/dx becomes an increasingly large negative number.

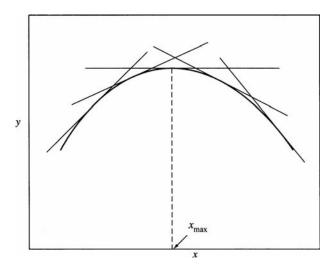


Figure 0.9. The tangent lines show how dy/dx changes as y passes through a maximum.

This behavior provides a criterion for the location of a maximum in a function. At a maximum, dy/dx = 0, dy/dx is positive at $x < x_{max}$, and dy/dx is negative at $x > x_{max}$. (In addition, the second derivative d^2y/dx^2 is negative at a maximum.) Analogous conditions locate a minimum: dy/dx = 0, dy/dx is negative for $x < x_{min}$, dy/dx is positive for $x > x_{min}$ and the second derivative is positive.

An inflection point in a plot of y against x is a point where the first derivative passes through a maximum; hence it is also detectable as the point where the second derivative is equal to zero. This idea is clarified with some experimental data in Example 0.18 (Connors, 1967, Chapter 6).

Example 0.18. Given: 0.3070 g of a weak base dissolved in glacial acetic acid was titrated with 0.1138 M acetous perchloric acid; the data in Table 0.11 are the titrant volume V and the electrochemical cell potential E in the vicinity of the endpoint. Estimate the endpoint volume by the second derivative method.

The third column in Table 0.11 shows $\Delta E/\Delta V$ (an approximation to the first derivative) as calculated from the V, *E* data; note how the values of $\Delta E/\Delta V$ are centered on the *V*, *E* intervals. In column 4 the "second derivative" $\Delta^2 E/\Delta V^2$ is similarly calculated from

$$\frac{\Delta^2 E}{\Delta V^2} = \frac{(\Delta E / \Delta V)_2 - (\Delta E / \Delta V)_1}{V_2 - V_1}$$

Figure 0.10 shows plots of *E* against *V*, of $\Delta E/\Delta V$ against *V*, and of $\Delta^2 E/\Delta V^2$ against *V*.

The endpoint could be estimated directly from Fig. 0.10a as the volume corresponding to the steepest point of the curve, but Fig. 0.10b shows how this point can also be found through extrapolation from the $\Delta E/\Delta V$ values on either side of the maximum. Figure 0.10c uses an interpolation from the $\Delta^2 E/\Delta V^2$ calculation to find the endpoint;

	· ·	,
	$\Delta E / \Delta V$	$\Delta^2 E / \Delta V^2$
E (mV)	(mV/mL)	(mV/mL^2)
490		
	44	
501		112
	72	
519		192
	120	
549		48
	132	
582		-176
	88	
604		-112
	60	
619		
	501 519 549 582 604	$\begin{array}{c c} & \Delta E / \Delta V \\ E (mV) & (mV/mL) \\ \hline 490 & & \\ & 44 \\ 501 & & \\ & 72 \\ 519 & & \\ & 120 \\ 549 & & \\ & 132 \\ 582 & & \\ 582 & & \\ & 88 \\ 604 & & \\ & 60 \\ \hline \end{array}$

Table 0.11. Potentiometric titration data (Example 0.18)

the calculation, from the table, is

$$V_{\text{endpoint}} = 8.25 + 0.25(\frac{48}{48 + 176}) = 8.30 \text{ mL}$$

Thus, the point at which $\Delta^2 E/\Delta V^2 = 0$ is found by linear interpolation between the two data points on either side of zero. (We now have enough data to calculate the equivalent weight of the weak base).

Integration. Integration is the opposite of differentiation; starting from the derivative, we seek to recover the original function. An equation of the form

$$\frac{dy}{dx} = f(x) \tag{0.75}$$

is called a *differential equation*; we have seen many examples. We wish to learn how y is related to x; our only clue is Eq. (0.75). We rearrange to the form

$$dy = f(x) \, dx \tag{0.76}$$

and integrate both sides as directed by the integral signs:

$$\int dy = \int f(x) \, dx \tag{0.77}$$

(An integral sign must always be accompanied by a differential.) On the left-hand side of Eq. (0.77) the integral sign reverses the action of the differential sign, since y is the

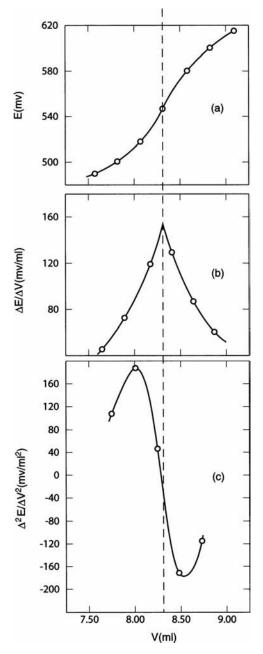


Figure 0.10. Plots of the data in Example 0.18. The dashed line shows the endpoint of the titration. [Reproduced by permission from Connors (1967).]

function whose differential is dy. Hence

$$y = \int f(x) \, dx \tag{0.78}$$

To proceed, we will take some special cases.

Example 0.19.

(a) Integrate dy/dx = a, where a is a constant. In the form of Eq. (0.78) we have

$$y = \int a \, dx \tag{0.79}$$

One rule of integration is that we can take constants outside the integral sign, so

$$y = a \int dx \tag{0.80}$$

This suggests that y = ax. But consider the following. Suppose that the original function was y = ax + 2, or $y = ax + 10^6$, or y = ax - 15, or the like. All of these functions (and there are an infinite number of them) have the derivative dy/dx = a, because the derivative of a constant is zero. So when we integrate Eq. (0.80) we must restore the constant that *might* have been there. Our final result is therefore

$$y = ax + C \tag{0.81}$$

where C is called the constant of integration.

(b) Integrate $dy/dx = 24x^2$. Experience with derivatives leads us to expect [see Eq. (0.70) in Table 0.10] that if $dy/dx = 24x^2$, then y is some function of x^3 . A bit of guessing shows us that if $y = 8x^3$, then $dy/dx = 24x^2$. We therefore have

$$y = 24 \int x^2 dx = (24) \left(\frac{x^3}{3}\right) + C = 8x^3 + C$$

Integrals of the type discussed above are called *indefinite integrals*, at least in part because the value of the constant *C* is unspecified. A short tabulation of indefinite integrals is given in Table 0.12 [which contains Eqs. (0.82)–(0.90)]. The *CRC Handbook of Chemistry and Physics* gives a table with hundreds of integrals.

A *definite integral* has specified initial and final points over which the integral is to be evaluated. We will develop this concept with an example of great importance in many types of scientific work.

Table 0.12. Some indefinite integrals

$$\int du = u + C \tag{0.82}$$

$$\int a \, du = a \int du \tag{0.83}$$

$$\int (du + dv) = \int du + \int dv \tag{0.84}$$

$$\int u^n du = \frac{u^{n+1}}{n+1} \ (n \neq -1) \tag{0.85}$$

$$\int \frac{du}{u} = \ln u + C \tag{0.86}$$

$$\int e^{au} du = \frac{e^{au}}{a} + C \tag{0.87}$$

$$\int a^u du = \frac{a^u}{\ln a} + C \tag{0.88}$$

$$\int (a+bu)^n du = \frac{(a+bu)^{n+1}}{(n+1)b} + C(n \neq -1)$$
(0.89)

$$\int \frac{du}{a+bu} = \frac{1}{b} \ln\left(a+bu\right) \tag{0.90}$$

Example 0.20. Let us study a chemical reaction described by the scheme $A \rightarrow Z$, where *A* is the reactant and *Z* is the product. It is often observed experimentally, for reactions fitting this scheme, that the rate of loss of reactant is directly proportional to the concentration of *A* at that moment. Expressed mathematically, this observation becomes

$$-\frac{dc_A}{dt} = kc_A \tag{0.91}$$

Here dc_A/dt is the *reaction rate*; the negative sign arises because c_A , the concentration of A, is decreasing as time t increases. The quantity k is the *rate constant*. Because c_A

appears on the right-hand side to the first power, Eq. (0.91) is called the *first-order* differential rate equation, and k is a first-order rate constant. [In Problem 0.22, Eq. (0.91) is obtained from a set of experimental kinetic data.]

Equation (0.91) expresses a rate as a function of a concentration, but it is more convenient experimentally to measure a concentration as a function of time. Integration will convert Eq. (0.91) into such a form. Let us make these definitions:

- c_A^0 is the concentration of A when t = 0, which means "at the beginning of the reaction."
- c_A is the concentration of A at any time t.

Now, to place Eq. (0.91) into an integrable form, we collect like variables; in this case we want c_A and dc_A on the same side of the equation. Algebraic rearrangement gives

$$\frac{dc_A}{c_A} = -k \, dt \tag{0.92}$$

We are going to carry out a definite integration between the limits $c_A = c_A^0$ when t = 0and $c_A = c_A$ when t = t. This is indicated as in

$$\int_{C_{A}}^{C_{A}} \frac{dc_{A}}{c_{A}} = -k \int_{0}^{t} dt$$
 (0.93)

Note that the initial state is at the lower end of the integral sign and the final state at the upper end. Next we integrate both sides, making use of Eqs. (0.82) and (0.86) (from Table 0.12):

$$\ln c_A]_{c_A^0}^{c_A} = -kt]_0^t \tag{0.94}$$

This symbolism shows that we have carried out the integration but have not yet applied the integration limits, so next we do this, writing the final state first and subtracting off the initial state, exactly as in our earlier definitions of changes:

$$\ln c_A - \ln c_A^0 = -k(t-0)$$

This is obviously equivalent to

$$\ln \frac{c_A}{c_A^0} = -kt \tag{0.95}$$

or, in Briggsian logarithms,

$$\log \frac{c_A}{c_A^0} = \frac{-kT}{2.303} \tag{0.96}$$

Yet another rearranged form is shown as

$$c_A = c_A^0 e^{-kt} \tag{0.97}$$

Equations (0.95)–(0.97) are equivalent forms of the *integrated first-order rate equation* (this is sometimes called the *first-order rate law*, *first-order decay*, or the *exponential rate law*). We have seen here that it appears naturally in the field of chemical kinetics. It is also important for describing radioactive decay, pharmacokinetic processes, and other applications. Mastery of the material in Example 0.20 is essential. Note also how Example 0.16 and Problem 0.22 are related to this treatment.

More complicated rate equations will sometimes be encountered, but a thorough treatment is not appropriate here. Sometimes the kinetic scheme and the differential rate equation can suggest the qualitative nature of the integrated equation. Consider this scheme of two first-order reactions in series:

$$A \xrightarrow{k_1} B \xrightarrow{k_2} C$$

The differential equations are

$$-\frac{dc_A}{dt} = k_1 c_A$$
$$\frac{dc_B}{dt} = k_1 c_A - k_2 c_B$$
$$\frac{dc_c}{dt} = k_2 c_B$$

We can therefore anticipate that the dependence of c_A on t will be given by Eq. (0.97), since the rate of loss of A depends only on rate constant k_1 . Intermediate B, however, involves both k_1 and k_2 , so we expect that c_B will be a biexponential function. The integrated result for c_B [which we will not derive here (Connors, 1990, Chapter 3)], is

$$c_b = \frac{c_A^0 k_1}{k_2 - k_1} \left[e^{-k_1 t} - e^{-k_2 t} \right]$$
(0.98)

except for the special case $k_1 = k_2$. The dependence of c_C on time can be found from the conservation equation $c_A^0 = c_A + c_B + c_C$ combined with Eqs. (0.97) and (0.98) for c_A and c_B .

Partial Differentiation and the Total Differential. Up to this point in our treatment of change, we have dealt with functions of a single variable. For example, if y is a function of the variable x alone, we write y = f(x), and we can evaluate the derivative dy/dx. Very often, however, we encounter functions of two or more variables, and here we

consider how to describe changes in such quantities. This material may not have been covered in an introductory calculus course.

Suppose that we have a function z = f(x, y), meaning that the value of z depends on both of the variables x and y. We wish to describe how z changes when we alter x or y, or both x and y. We do this as follows. First take the derivative of z with respect to x while holding y constant. In order to make transparently clear what is going on, we use a slightly different symbolism for this derivative, writing

$$\left(\frac{\partial z}{\partial x}\right)_y$$

Note the subscript y; this tells us that y is held constant while x is changing. In like manner we can take the derivative of z with respect to y while x is held constant, writing the result as

$$\left(\frac{\partial z}{\partial y}\right)_x$$

Observe the use of the symbol \hat{o} in these expressions. The operation just described is called partial differentiation and the quantities written above are *partial derivatives*.

Next we make use of a chain rule to define the *total differential* dz of the function z = f(x, y):

$$dz = \left(\frac{\partial z}{\partial x}\right)_{y} dx + \left(\frac{\partial z}{\partial y}\right)_{x} dy \qquad (0.99)$$

These considerations may seem somewhat abstract, but it happens that many quantities of scientific interest are functions of more than one variable. Incidentally, Eq. (0.99) can be generalized to functions of more than two variables; the pattern will be evident from Eq. (0.99) (see also Problem 0.31).

Example 0.21. The volume V of a fixed amount of a homogeneous gas depends only on its temperature T and its pressure P. Write the total differential dV.

By analogy with the foregoing, we write

$$V = f(T, P)$$

so

$$dV = \left(\frac{\partial V}{\partial T}\right)_P dT + \left(\frac{\partial V}{\partial P}\right)_T dP \tag{0.100}$$

Experimental measurements can provide numerical quantities for these partial derivatives, so Eq. (0.100) has a definite physical meaning.

Recall from our earlier definition of change in a chemical process that we defined an increment as the difference between values in final and initial states. Taking volume *V* as an example of a property undergoing change, using the language of calculus, we have,

$$\Delta V = \int_{V_{\text{initial}}}^{V_{\text{final}}} dV = V_{\text{final}} - V_{\text{initial}}$$
(0.101)

Now suppose that we carry out the change in two steps, first $A \to B$ and then $B \to C$. What is the change ΔV for the overall process $A \to C$? We apply Eq. (0.101):

$$\Delta V = \int_{V_A}^{V_B} dV + \int_{V_B}^{V_C} dV = (V_B - V_A) + (V_C - V_B)$$
$$\Delta V = V_C - V_A$$

We have found that state *B* plays no role in determining ΔV . State *B* is an intermediate state on the path from *A* to *C*, and since *B* could have been any intermediate state, evidently the change ΔV is independent of the path or mechanism of the process.

If the change in a property or function between two *states* of a system is independent of the path taken between the states, the total differential is called an *exact differential* and the property or function is called a *state function*. Whether or not a function is a state function (path-independent) is ultimately based on experimental observations, but extensive laboratory studies have clarified the situation. The subject of thermo-dynamics (which describes systems at equilibrium) deals largely with state functions, including the temperature, volume, pressure, and energy.⁸ On the other hand, chemical kinetics, which describes systems changing in time, largely treats path-dependent quantities; in this context, the path taken by a reaction is the reaction mechanism, and a major role of chemical kinetics studies is to investigate reaction mechanisms.

0.6. STATISTICAL TREATMENT OF DATA

The results of a quantitative experimental study often consist of a set of numbers obtained from replicate determinations made under essentially identical conditions, or a set of numbers corresponding to one quantity as a function of another quantity. These numbers may represent concentrations, weights, equilibrium constants, rate constants, pH values, and so on. The interpretation of these data by the experimentalist answers two general questions:

- 1. What is the *best estimate* of the quantity or function being investigated?
- 2. *How reliable* is this estimate as a measure of the true value?

 $^{\rm 8}$ Heat and work, however, which are important thermodynamic quantities, are not, in general, state functions.

The branch of mathematics called *statistics* deals with such issues. Our present treatment will be very brief, intended to provide an immediately usable tool for the interpretation of data obtained in laboratory coursework.

Random Errors and the Normal Distribution. Every measurement has some error associated with it. These errors may be of two types. *Systematic errors* are errors introduced by some inadequacy in the experimental technique (such as an improperly calibrated balance or an interference caused by impurities in reagents) or poor judgment or unconscious bias on the part of the experimentalist (as may happen through parallax error when reading the meniscus in a burette). Systematic errors can be tracked down and substantially eliminated (or the method may have to be abandoned); we will not consider them further.

Even after all systematic errors have been eliminated, it is a familiar observation that repeated determinations of a quantity almost never result in the same number. This variability in experimental data is a manifestation of *random error*. We usually observe that replicate observations are grouped about a most frequently observed value and that large deviations from this value are rarer than small deviations. Random errors are the consequence of limitations inherent in the observational method. They can perhaps be reduced in magnitude by careful work, but they cannot be eliminated. Statistics helps the experimentalist to interpret the data, given this inevitable presence of random error.

The statistician adopts a point of view concerning experimental data that may seem peculiar to the experimentalist. The statistician considers an experimental observation to be a single member that has been randomly selected from an infinite population of individual observations that are characteristic of the system being investigated. Replicate observations will exhibit variation owing to the operation of random errors. If we draw a graph of the value of each experimental observation on the horizontal (x) axis against the number of times each value is observed (its frequency) on the vertical (y) axis, a symmetric figure usually is obtained, with a maximum corresponding to the most frequently observed value. As the number of observations increases toward the (unattainable) limit of infinity, the graph will assume the form of a smooth curve. This curve is called a *frequency distribution*.

In scientific practice, experimental frequency distributions can usually be closely fitted to a theoretical frequency distribution that is called the *normal distribution*, the *Gaussian error curve*, or the *normal error curve*. The equation of the normal distribution is

$$f(x) = \frac{1}{\sigma\sqrt{2\pi}} e^{-(1/2)[(x-\mu)/\sigma]^2}$$
(0.102)

where f(x) is the frequency of occurrence of the value x of the "random variable," which is the experimental observable, and σ and μ are parameters of the population. The symbol μ signifies the *population mean*, which is the value of x corresponding to the maximum in the distribution; and σ , the *population standard deviation*,

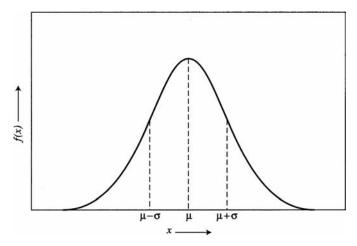


Figure 0.11. The normal error curve with mean μ and standard deviation σ .

determines the "spread" or width of the bell-shaped curve. Figure 0.11 shows a plot of the normal distribution. Graphically, the quantity σ is the horizontal distance from the mean to either inflection point of the curve, as shown in Fig. 0.11. Approximately 68% of the area under the normal curve, and therefore 68% of the members of the population, lie within one standard deviation of the mean, $\mu \pm \sigma$. About 95% of the population are found in the range $\mu \pm 2\sigma$, and over 99% in the range $\mu \pm 3\sigma$. Scientists usually assume that their data can be described by the normal distribution, and we will adopt this assumption in the following treatment.

Estimation of Statistical Parameters. The population parameters σ and μ are not accessible to us because we would have to sample an infinity of random variables in order to define the population frequency distribution. Nevertheless, we desire estimates of μ and σ . Because of the symmetry of the normal distribution, the best estimate of the population mean is the arithmetic average, or *mean*, defined by

$$\bar{x} = \frac{\sum x_i}{n} \tag{0.103}$$

where x is the value of the ith observation (i = 1, 2, 3, ..., n) and n is the total number of observations.

The *accuracy* of an experimental result is the closeness with which the experimental mean \bar{x} approaches the population mean μ . Since we do not know μ , we cannot in general assess the accuracy. Sometimes, however, an experiment can be designed so that the experimental \bar{x} can be compared with a known value that has been set by the experimentalist with a standard sample. There also exist a few quantities that have been measured by so many workers, using different methods in different laboratories, that we have developed great confidence in their accuracy. For example, the K_a value of benzoic acid at 25° C is 6.26×10^{-5} , an average of many experimental results; and so much care and effort has gone into this number that we may reasonably take it as very accurate.

The *median* is sometimes used as an estimate of the population mean, especially if the set of experimental results includes one or two widely divergent results. The median is that value that exceeds as many values as it itself is exceeded by, for n odd. If *n* is even, the median is the average of the two values satisfying the same criterion. The median is easily picked out if the numbers are arranged in order of increasing magnitude.

It is a valuable result of statistical theory that, even if the experimental variable is not normally distributed, the means of small sets of the variable are normally distributed. Since the mean \bar{x} is our best estimate of the true value, we are assured that these \bar{x} estimates follow a normal distribution.

We have next to obtain an estimate of σ , the population standard deviation, which we see from Fig. 0.11 to be a measure of the width of the normal distribution. This width reflects the reproducibility of the measurements; the more reproducible, the narrower the curve, and the smaller the value of σ . *Precision* is the term usually used to describe reproducibility. Our estimate of σ is labeled *s* and is called the *experimental standard deviation; s* is our measure of the precision of \bar{x} , and sometimes *s* is said to measure the "uncertainty" of \bar{x} . Its calculation begins with the definition of the following equation, where s^2 is called the *variance*:

$$s^{2} = \frac{\sum (x_{i} - \bar{x})^{2}}{n - 1} \tag{0.104}$$

The standard deviation is then found as the square root of the variance. Note the denominator in this equation; the quantity n-1 is called the *degrees of freedom*, because it specifies the number of independently assignable quantities needed to completely determine the system. This is n-1 because we already have calculated one parameter, namely, \bar{x} . The units of *s* are the same as those of \bar{x} .

At one time (before electronic calculators and computers made statistical calculations easy), the *average deviation*, expressed as follows, was often used as a measure of precision:

Average deviation
$$= \frac{\sum |x_i - \bar{x}|}{n-1}$$
 (0.105)

The *range* w is the difference between the largest and the smallest results in a set; w can be a useful indicator of the spread of results when n is small.

The precision can be expressed in relative terms as the quotient s/\bar{x} , or more frequently on a percent basis by $100s/\bar{x}$, which is called the *coefficient of variation* or the *relative standard deviation* (RSD). Similarly, we might report

Precision in parts per thousand
$$=$$
 $\frac{10^3 s}{\bar{x}}$
Precision in parts per million $=$ $\frac{10^6 s}{\bar{x}}$

It is important to appreciate that the accuracy and precision of an experimental result are very different concepts. In the best of circumstances we may have good accuracy (\bar{x})

closely approximates μ) and high precision (*s* is small relative to \bar{x}), but it is also possible to encounter poor accuracy with high precision.

A more complete description of *s* is that it is the standard deviation of a single observation. However, it is the mean \bar{x} that we take as our best estimate, so we really would like a measure of the precision of the mean. This is provided by the *standard deviation of the mean*, s_m :

$$s_m = \frac{s}{\sqrt{n}} \tag{0.106}$$

Some authors call s_m the standard error.

Example 0.22. The following numbers are the percent recoveries in seven identical nonaqueous titrations of a urea sample: 98.4, 100.2, 99.3, 101.7, 97.4, 98.2, and 100.8. Calculate the mean, median, range, variance, standard deviation, relative standard deviation, and standard deviation of the mean.

It is best to arrange the work in tabular form. Notice that the variance is calculated according to Eq. (0.104) with the arithmetic operations carried out in the following order: (1) differences are taken, (2) differences are squared, and (3) squares are summed:

Titration, <i>i</i>	Percent Recovery (x_i)	$(x_i - \bar{x})$	$(x_i - \bar{x})^2$
1	98.41	-1.01	1.00
2	100.2	0.8	0.64
3	99.3	-0.1	0.10
4	101.7	2.3	5.29
5	97.4	-2.0	4.00
6	98.2	1.2	1.44
7	100.8	1.4	1.96
	696.0		14.34

TITRATION DATA

Mean: \bar{x} = 696.0%/7 = 99.4% Median: 99.3% Range: 101.7% - 97.4% = 4.3% Variance: $s^2 = [14.34(\%^2)]/6 = 2.39(\%^2)$ Standard deviation: $s = \sqrt{2.39} = 1.55\%$ RSD: 100(1.55/99.4) = 1.56% Standard deviation of the mean: $s_m = 1.55/2.65 = 0.58\%$

Equation (0.104) for the variance can be placed in other forms that are more convenient for electronic calculation. You will undoubtedly use your electronic calculator or computer (which should be capable of the routine statistical calculations described in this section), but make sure that the program uses n - 1 rather than n in the

Degrees of Freedom	P = 0.10	0.05	0.01
1	6.314	12.706	63.657
2	2.920	4.303	9.925
3	2.353	3.182	5.841
4	2.132	2.776	4.604
5	2.015	2.571	4.032
6	1.943	2.447	3.707
7	1.895	2.365	3.499
8	1.860	2.306	3.555
9	1.833	2.262	3.250
10	1.812	2.228	3.169

Table 0.13. Some values of student's t distribution

denominator of Eq. (0.104). Of course, if *n* is very large (an unusual circumstance in much experimental work), the difference between *n* and n - 1 becomes negligible.

Confidence Limits of the Mean. We have seen that the mean \bar{x} can be treated as a normally distributed random variable. \bar{x} is our best estimate of the quantity we seek, and now we would like some measure of the reliability of \bar{x} . It might seem that we could achieve this by taking note of the properties of the normal distribution. For example, we saw that 95% of the members of a normally distributed population fall within two standard deviations of the mean, so perhaps we could state that the interval $\bar{x} \pm 2s$ should include 95% of any future estimates of \bar{x} for the same quantity.

This sounds plausible, but as it happens, if *n* is fairly small, the actual distribution of \bar{x} is somewhat wider than is specified by the normal distribution. The actual distribution for small *n* is given by a different function called Student's *t* distribution,⁹ which approaches the normal distribution as *n* becomes large. Table 0.13 lists some values of Student's *t*. In this table the column headed "Degrees of Freedom" is to be interpreted as n - 1. The headings of the other columns give the value of *P*, which is the probability that the limits to be calculated may be exceeded by chance. The values in the table are the Student's *t*. Observe how *t* seems to be approaching 2 for the column P = 0.05 and recall that for the normal distribution 95% of the population lies in the interval $\bar{x} \pm 2s$; this comparison gives some meaning to the *t* values.

We now wish to measure the reliability of our \bar{x} estimate. This measure of reliability is called the *confidence limits of the mean*, and it is calculated with the aid of Student's *t* distribution. We suppose that *n* replicate observations have been made, and \bar{x} , *s*, and *s_m* have been calculated. We then calculate the confidence limits of \bar{x} by

 $\bar{x} \pm ts_m$

⁹ "Student" was the pen name of W. S. Gosset, a British statistician and chemist who worked at Guinness Breweries.

where *t* is read from the appropriate column in Table 0.13. For example, if we wish to know the limits within which 95% of further values would fall (if we undertook to measure them), we would choose the column P = 0.05, where the quantity *P* is the fraction of results that are expected to fall *outside* the limits. Similarly, the P = 0.01 column gives the 99% confidence limits.

Example 0.23. Find 95% and 99% confidence limits of the mean calculated in Example 0.22.

95% Confidence Limits. For P = 0.05 and 6 degrees of freedom, Table 0.13 gives t = 2.447. The confidence limits (P = 0.05) are therefore $\bar{x} = \pm ts_m$, or 99.4 \pm 2.447 (0.58)% = 99.4 \pm 1.4%. The meaning of this result is that, if many additional sets of seven observations were made, about 95% of the means of these sets would be expected to fall within the range 98.0–100.8% (the confidence interval).

99% Confidence Limits. For P = 0.01, t = 3.707. The limits (P = 0.01) are 99.4 \pm 2.2%. The limits are wider at P = 0.01 than at P = 0.05 because we have used the same information but have required that a larger percentage of additional results will fall within specified limits; that is, we have asked for a greater level of confidence, so we must pay for this with a wider confidence interval.

Comparison of Two Means. It is often required that two experimental results are to be compared to determine whether they are different. For example, we might wish to know if a newly developed chromatographic method yields the same analytic result as a well-tested spectrophotometric method. Such problems are dealt with by *tests of significance*, meaning that a decision is sought concerning whether the difference between the two results is significant or negligible.

In statistical terms, one tentatively assumes that there is no difference between the two results—this is called the *null hypothesis—and* then tests this assumption. Usually two means are to be compared. Let us symbolize these means as \bar{x} and \bar{y} . According to the null hypothesis, \bar{x} and \bar{y} describe the same population. To show the basis of the significance test, suppose that \bar{x} and \bar{y} have the same value s_m for the standard deviation of the mean. As we have seen, confidence intervals can be defined by

Confidence interval for $\bar{x} = \bar{x} \pm ts_m$ Confidence interval for $\bar{y} = \bar{y} \pm ts_m$

We can therefore say that \bar{x} and \bar{y} are not significantly different (at the *P* level of confidence expressed by *t*) if \bar{x} lies within the confidence interval of \bar{y} (and vice versa), as indicated graphically in Fig. 0.12.

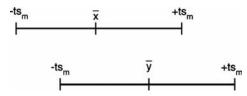


Figure 0.12. The means \bar{x} and \bar{y} are not significantly different if their confidence intervals overlap.

Therefore $\bar{x} = \bar{y} \pm ts_m$ expresses the null hypothesis; by rearranging, we obtain

$$t = \frac{\bar{x} - \bar{y}}{s_m} \tag{0.107}$$

The approach is to calculate t by Eq. (0.107). If the calculated t exceeds the tabulated t value at the chosen level of significance, the null hypothesis may be rejected; that is, \bar{x} and \bar{y} are significantly different.

This demonstrates the principle. Some subtleties enter when, as may happen, the numbers of observations contributing to \bar{x} and \bar{y} are different and when s_m for \bar{x} is different from s_m for \bar{y} . We will not pursue these matters here, except as seen in the following example.

Example 0.24. These are analytical results for the analysis of two lots of aspirin tablets, given in milligrams of aspirin per tablet. Are the aspirin contents of the two lots different at the 95% significance level?

	x_i	y_i
	295.4	300.5
	301.1	310.9
	297.8	307.1
	305.0	302.6
	297.5	305.9
For lot x we find $\bar{x} = 299.4$, s	= 3.755	$s, s_m^x = 1.679.$
For lot y we find $\bar{y} = 305.4$, s	= 4.039	$s_m^y = 1.806.$

We now introduce this modification. Somewhat later in this section we will discover that variances of both sums and differences are additive. Since the numerator of Eq. (0.107) is a difference, we really want the standard deviation of the mean of this difference in the denominator. We therefore take the square root (to get the standard deviation) of the sum of the variances:

$$s_m = \sqrt{\left(s_m^x\right)^2 + \left(s_m^y\right)^2} = 2.466$$

Now we calculate t with Eq. (0.107):

$$t = \frac{305.4 - 299.4}{2.466} = 2.433$$

Consulting Table 0.13, we find that the tabulated $t[P = 0.05; (n_x - 1) + (n_y - 1) = 8$ degrees of freedom] is 2.306. The calculated *t* exceeds the tabulated *t*, so the null hypothesis may be rejected, and we conclude that the two batches are different at the 95% significance level.

It is interesting that at the 99% significance level the tabulated t = 3.555, so the null hypothesis may not be rejected; the two batches are not different at this significance level. This result shows that statistics alone cannot make decisions; the experimentalist must exercise judgment. When reporting statistical results of this type, it is essential to specify the chosen level of significance.

Linear Correlation. In Section 0.4 we discussed linear relationships in detail. Our general formula for a *linear correlation* is

$$y = mx + b \tag{0.108}$$

where *m* is the slope and *b* is the intercept on the *y* axis. When confronted with a set of *x*, *y* data, we constructed a plot of *y* against *x*, determined by inspection whether it was linear, and drew our best straight line using a straightedge (ruler) and our intuition. We now wish to learn how statistics can assist us in such an analysis.

Our first task is to decide whether the correlation between *x* and *y* is linear. Visual inspection is usually adequate, but many scientists like to calculate a quantity *r* called the *correlation coefficient*:

$$r = \frac{s_{xy}}{s_x s_y} \tag{0.109}$$

where s_x is the standard deviation of all the *x* values, calculated in the usual way, and similarly s_y is the standard deviation of the *y* values. The quantity s_{xy} is called the *covariance* and is given by

$$s_{xy} = \frac{\sum (x_i - \bar{x})(y_i - \bar{y})}{n - 1} \tag{0.110}$$

Electronic calculators and computers yield r directly without requiring the user to proceed through Eqs. (0.109) and (0.110).

The numerator of Eq. (0.109) is a measure of how strongly correlated the *x* and *y* values are, and the denominator "normalizes" *r* so that it must lie in the interval 0 to 1 (for positive slope) or 0 to -1 (for negative slope). The closer *r* is to 1 (or -1), the more linear the correlation. That, at least, is the conventional interpretation, but it must be treated with caution. This is because *r* depends not only on the linearity of the relationship but also on the slope of the line (a steeper slope giving a larger *r* value) and on the scatter of the points (more scatter giving a smaller *r* value).

Figures 0.13 and 0.14 illustrate these points. These figures have been constructed so that the slopes of the lines are very similar. Figure 0.13 has more scatter than does Fig. 0.14, and this is reflected in their *r* values: r = 0.990 for Fig. 0.13; r = 0.999 for Fig. 0.14. But close observation will reveal that although the points in Fig. 0.13 show considerable scatter, they very reasonably follow a straight line. The points in Fig. 0.14, however, clearly describe a curve, although a straight line has been drawn through them. The lesson is that one should not rely on the correlation coefficient alone, but should also plot the data and examine the graph critically. A good way to do

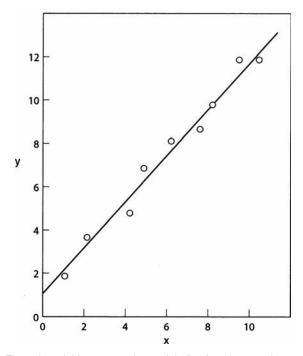


Figure 0.13. The points yield r = 0.990; the straight line has the equation y = 1.16x + 1.15.

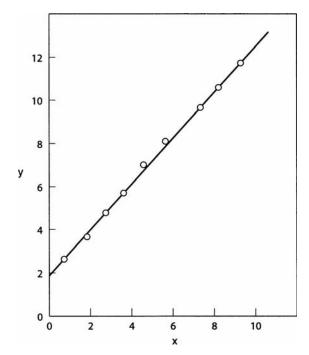


Figure 0.14. The points yield r = 0.999; the straight line has the equation y = 1.07x + 2.49.

this is by "sighting" along the data points as a carpenter sights along the edge of a board to determine how straight it is.

If we conclude that the data are linearly correlated, the next step is to draw the "best" straight line. This is nearly universally done now by a statistical technique called the *method of least squares* or *linear regression analysis*. The line is drawn such that the sum of the squares of the vertical distances of the points from the line is a minimum.¹⁰

Let the least-squares line be written

$$\hat{y}_i = mx_i + b \tag{0.111}$$

where *m* and *b* are the slope and intercept of the line (not yet known) and \hat{y}_i is the value of *y* on the least squares line corresponding to x_i . Then we write

$$G = \sum (y_i - \hat{y}_i)^2$$
 (0.112)

where y_i is the experimental value of y for observation i and \hat{y}_i is the corresponding value from the line. Thus the right-hand side expresses the sum of squares of differences of the points from the line. To minimize this sum, we use the methods of calculus, taking the partial derivatives $\partial G/\partial m$ and $\partial G/\partial b$, setting these equal to zero, and solving for the parameters. The results are

$$b = \bar{y} - m\bar{x} \tag{0.113}$$

$$m = \frac{\sum xy - n\bar{x}\bar{y}}{\sum x^2 - n\bar{x}^2} \tag{0.114}$$

In these equations, \bar{x} and \bar{y} are the mean values of these variables, and *n* is the number of data points. Inasmuch as scientific electronic calculators and computers are capable of generating the parameters *m* and *b* of the least-squares regression line, it is unlikely that you will make direct use of Eqs. (0.113) and (0.114). This is how the calculator or computer does it, however.

Example 0.25. In Example 0.15, the data in Table 0.7 were analyzed as a linear correlation. Repeat the analysis using the method of least squares.

Using an electronic calculator, we obtain the following for the least-squares regression line:

$$y = 0.447x + 0.0397$$

In Example 0.15 the line was drawn "by eye," and its equation turned out to be y = 0.453x + 0.0397. These two results compare quite favorably. The reason for the general preference for the least-squares treatment is that it is objective; any number of scientists applying the least squares method to the same set of data will obtain the same result.

¹⁰ It might seem simpler to minimize the sum of the vertical distances themselves, but this sum is zero, because some points are above the line and some are below the line. By squaring the distances, all quantities are converted to positive numbers. It will be seen that this procedure has much in common with the calculation of the variance, Eq. (0.104).

Since the slope and intercept values of the least-squares regression line are themselves estimates of statistical parameters, they possess uncertainties that can be expressed as their standard deviations. We will not go into the evaluation of the standard deviations of the slope and intercept values.

Propagation of Errors. We nearly always subject our raw experimental data to arithmetic operations in order to arrive at a number that is useful to us, as when we convert a titrant volume to a concentration or a weight of sample or when we operate on a parameter of an equation to give us a rate constant. When we make such calculations, we are ultimately interested in the uncertainty (expressed as a standard deviation) of the final calculated result, whereas what we may know are the standard deviations of the quantities that are used in the calculation. We therefore wish to learn how the errors (uncertainties) in the primary quantities are propagated through the calculation into the final result.

Suppose that we let Q represent our final result, which is obtained by carrying out calculations on the two primary quantities x and y; that is, we have

$$Q = f(x, y)$$

Then the equation for the propagation of the uncertainties in x and y into Q, which we will not derive here, is

$$s_{Q}^{2} = \left(\frac{\partial Q}{\partial x}\right)^{2} s_{x}^{2} + \left(\frac{\partial Q}{\partial y}\right)^{2} s_{y}^{2}$$
(0.115)

where the quantities in parentheses are partial derivatives, and s_Q^2 , s_x^2 , s_y^2 are the variances of the subscripted quantities. We will apply Eq. (0.115) to the basic arithmetic operations.

Sum

$$Q = x + y$$
$$\frac{\partial Q}{\partial x} = 1; \qquad \frac{\partial Q}{\partial y} = 1$$

 $\Omega = r \eta$

so, from eq. (0.115), we have

$$s_Q^2 = s_x^2 + s_y^2 \tag{0.116}$$

Difference

$$Q = x - y$$

$$\frac{\partial Q}{\partial x} = 1; \qquad \frac{\partial Q}{\partial y} = -1$$

$$s_Q^2 = s_x^2 + s_y^2 \qquad (0.117)$$

Thus the errors (variances) add for both sums and differences. (We used this result in Example 0.24.) This is why the relative error of a difference of two numbers similar in magnitude may be very large.

 \sim

Product

$$Q = xy$$

$$\frac{\partial Q}{\partial x} = y; \qquad \frac{\partial Q}{\partial y} = x$$

$$s_Q^2 = y^2 s_x^2 + x^2 s_y^2$$

which is equivalent to

$$\frac{s_Q^2}{Q^2} = \frac{s_x^2}{x^2} + \frac{s_y^2}{y^2} \tag{0.118}$$

Quotient

$$Q = \frac{y}{x}$$

Equation (0.118) is again obtained.

The essential result of these demonstrations is that *variances are additive*. For sums and differences, the absolute variances add [Eqs. (0.116) and (0.117)]; for products and quotients, relative variances add [Eq. (0.118))]. To find the standard deviation of Q, we first calculate s_Q^2 as in the examples above and then take its square root.

Example 0.26. Suppose that we titrate a solution of acetic acid with 15.00 mL of 0.1000 N NaOH. What is the uncertainty in the weight of acetic acid found if $s_N = 0.0003$ and $s_V = 0.02$ mL?

Since mmol of HOAc = VN = w/M, where V is volume of titrant, N is titrant normality (molarity), w is weight of acetic acid in milligrams, and M is the molecular weight of acetic acid, our function is w = MVN. Thus

$$s_w^2 = \left(\frac{\partial w}{\partial V}\right) s_V^2 + \left(\frac{\partial w}{\partial N}\right) s_N^2$$

We have $(\partial w/\partial V) = MN$ and $(\partial w\partial N) = MV$, so

$$s_w^2 = M^2 N^2 s_V^2 + M^2 V^2 s_N^2$$

= (60)²(0.1)²(4 × 10⁻⁴) + (60)²(15)²(9 × 10⁻⁸)

$$= 14.4 \times 10^{-3} + 8.1 \times 10^{-3} = 2.25 \times 10^{-2} \text{ mg}^2$$

 $s_w = 0.15 \text{ mg}$

Since w = (60)(15)(0.1) = 90 mg, the relative standard deviation (RSD, or coefficient of variation) is (100)(0.15)/90 = 0.17%.

A practical consequence of the propagation of error treatment is that if one of the uncertainties (say, σ_x^2) is very much larger than the other, then the uncertainty σ_Q^2 will receive its largest contribution from σ_x^2 . Any experimental attempts to reduce the error in Q should obviously focus on reducing the error in x, because the error in y makes a negligible contribution to the error in Q.

Significant Figures. In reporting a final numerical result, it is necessary to decide how many digits will be retained. This is especially pertinent in modern science because electronic calculators and computers routinely generate numbers having 10 or more digits. There is a temptation to record the entire display, but this is seldom justified. Instead we should retain and report only those digits that have physical significance.

Here is a simple criterion for deciding on how many digits to retain. The final digit should possess some uncertainty, whereas the digit preceding this one should be essentially certain. Some flexibility (within one digit) is acceptable. The number of significant figures in a result is then the total number of digits (exclusive of zeros that are needed solely to establish the position of the decimal point).

In order to use this criterion, we need a means for establishing whether a digit is essentially certain or is uncertain. We have this means at hand in the standard deviation *s* (or the standard deviation of the mean s_m if it is a mean that we are reporting). Consider, for example, the analytical results reported in Example 0.24. For the *x* series we calculated x = 299.4, $s_m^x = 1.679$, and for the *y* series we calculated, $\bar{y} = 305.4$, $s_m^y = 1.806$. For both series, therefore, the s_m values indicate variability in the unit place, suggesting that the means are uncertain in the unit place. Accordingly, we would report these results as $\bar{x} = 299$, $s_m^x = 1.7$ and $\bar{y} = 305$, $s_m^y = 1.8$. Incidentally, it is good practice to retain more digits than otherwise may be justified throughout the calculation in order that "rounding errors" not be inadvertently introduced. Then the significant-figure criterion is applied to the final result.

Another guide may be available in experimental practice and experience. For example, much laboratory experience has shown us that very precise titrimetric analysis can routinely be accomplished with precision corresponding to standard deviations in the fourth decimal place of molar concentrations; this is the justification for expressing solution concentrations to the fourth place. Another example occurs in pH measurements, where experience shows that, in careful but nevertheless routine work, the tenth unit is certain and the hundredth unit is uncertain, so we usually write pH values with two decimal places. The experimental procedure itself may determine the significance. For example, on a typical buret the finest graduations are in tenths of a milliliter, and we estimate to the hundredths place; thus we write the volume to two decimal places.

PROBLEMS

- **0.1.** Convert these pH values to hydrogen ion concentrations:
 - (a) pH = 4.75
 - **(b)** pH = 11.13
 - (c) pH = 7.19
 - (d) pH = 2.00
- **0.2.** Convert these hydrogen ion concentrations to pH values:
 - (a) $[H^+] = 3.85 \times 10^{-3} M$
 - **(b)** $[H^+] = 1.15 \times 10^{-9} M$
 - (c) $[H^+] = 6.46 \times 10^{-8} M$
 - (d) $[H^+] = 1.15 M$
- **0.3.** Convert ln *x* to the corresponding Briggsian logarithm.
- **0.4.** How much more acidic is a solution of pH 3.00 compared with a solution of pH 9.00?
- **0.5.** Solve for *q* in the equation

$$13.52^q = 5.62 \times 10^4$$

- **0.6.** Calculate the product 6, 942, 821×0.0057384 by using logarithms. Check the result by direct multiplication on an electronic calculator.
- **0.7.** How many grams of benzoic acid are required to prepare 500 mL of a 0.0025 M solution?
- **0.8.** Ethyl acetate hydrolyzes according to the reaction

$$CH_3COOC_2H_5 + H_2O \rightarrow C_2H_5OH + CH_3COOH$$

How much acetic acid is produced by the hydrolysis of 10.00 g of ethyl acetate?

- **0.9.** Let $R_{A^-} = [A^-]/[HA]$ and $F_{A^-} = [A^-]/([HA] + [A^-])$. Then derive an equation giving R_{A^-} as a function of F_{A^-} (or the reverse).
- **0.10.** Given [HA] = [A⁻], calculate the ratios R_{HA} , R_{A^-} and the fractions F_{HA} , F_{A^-} .
- **0.11.** A solution of weak acid is prepared to have a total concentration of 7.50×10^{-4} M, and analysis shows that the ratio [A⁻]/[HA] is 0.15. Calculate the individual concentrations [HA] and [A⁻].
- **0.12.** Vinegar contains ~5% acetic acid (i.e., 5 g of acetic acid per 100 mL). The K_a of acetic acid is 1.78×10^{-5} . Calculate the pH of vinegar.
- **0.13.** Repeat Problem 0.12 using the approximate solution [Eq. (0.40)].

- **0.14.** Find the value of the intercept on the x axis for the straight line of Eq. (0.41).
- **0.15.** Carry out the algebraic manipulations that give Eqs. (0.51)–(0.53) from Eq. (0.50).
- **0.16.** In this set of data, *c* is the molar concentration of *trans*-cinnamic acid (C₆H₅CH=CHCOOH) in acidic solution, and *A* is a dimensionless measure of light-absorbing ability of the solution called the absorbance. It is anticipated that the data are described by the equation $A = \varepsilon bc$, which is called Beer's law, where *b* is the "pathlength" of light through the solution (*b* = 1 cm in this experiment) and ε is called the *molar absorptivity*. Analyze the data:

$10^5 c (M)$	A
0	0
1.083	0.224
2.165	0.450
3.248	0.679
4.330	0.901

0.17. The noncovalent interaction of cinnamic acid anion with theophylline was studied optically with a pathlength b of 1 cm. These are the data, where we let c be the theophylline concentration.

$c(\mathbf{M})$	Α
0	$0.98(A_0)$
0.0111	1.375
0.0125	1.418
0.0143	1.472
0.0167	1.544
0.0200	1.638
0.0250	1.767

We expect the system to be described by

$$\frac{\Delta A}{b} = \frac{BKc}{1+Kc}$$

where $\Delta A = A - A_0$. Find the equilibrium constant *K*.

0.18. The equilibrium solubility of a solute usually varies with temperature according to

$$\ln s = -\frac{\Delta H}{RT} + C$$

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where *s* is the solubility corresponding to absolute temperature *T*, *R* is the gas constant, *C* is a constant, and ΔH is the molar heat of solution. These are solubility data for γ -cyclodextrin in water, where *s* is the mole fraction solubility:

t (°C)	S
25	0.003539
30	0.004456
35	0.005476
40	0.006239
42	0.007583

Find the heat of solution.

0.19. The total solubility S_t of 4,4'-dihydroxybiphenyl varies with the concentration [L] of α -cyclodextrin according to

$$S_t = s_0 + K_{11}s_0[L] + K_{11}K_{12}s_0[L]^2$$

where s_0 is the solubility when [L] = 0, and K_{11} and K_{12} are equilibrium constants for the formation of 1:1 (SL) (substrate:ligand) and 1:2 (SL₂) complexes. Here are data for 25°C:

$10^{2}\left[L\right]\left(M\right)$	$10^4 S_t (\mathbf{M})$
0	1.98
0.311	2.53
0.412	2.74
0.512	3.13
0.611	3.51
0.709	4.02
0.806	4.39
0.810	4.62
0.902	4.94
0.998	5.74
1.19	6.86

Find K_{11} and K_{12} .

0.20. The rate of hydrolysis of 4-nitrophenyl glutarate at 25°C and pH7 follows a rate law like Eq. (0.54). When the absorbance *A* is measured as a function of time,

the equation is written

$$A_{\infty} - A_t = (A_{\infty} - A_0)e^{-kt}$$

where A_t is the absorbance at time t, A_0 at t = 0, and A_∞ at $t = \infty$ ("infinity" time, when the reaction is essentially complete). Here are the data; find the rate constant k:

t(s)	A _t	t(s)	At
10	0.129	120	0.453
20	0.168	140	0.492
30	0.200	160	0.531
40	0.237	180	0.565
50	0.270	200	0.598
60	0.300	240	0.650
70	0.330	280	0.692
80	0.357	320	0.729
90	0.381	360	0.758
100	0.407	400	0.783
		8	0.900

0.21. Rearrange this equation to give a linear plotting form

$$y = \frac{1 + ax}{1 + bx}$$

- **0.22.** Consult the set of data in Example 0.16. For each pair of adjacent time points, calculate the increment in time (Δt) and the increment in concentration (Δc) . Take their quotient $\Delta c/\Delta t$ as a crude estimate of dc/dt. Now plot each $\Delta c/\Delta t$ value against the mean value of c (i.e., $\bar{c} = (c_1 + c_2)/2$] corresponding to the time interval. Interpret the result.
- **0.23.** If $y = a^u$, find dy/dx, where *a* is a constant and *u* is a function of *x*. (*Hint:* Start by taking the logarithm of *y*.)
- **0.24.** Equation (0.46), a power function, has the form $y = ax^{b}$. (a) Find dy/dx.
 - (b) For the special case b = 2, evaluate both y and dy/dx when x = 0.5, 1, and 2. Compare the x dependencies of y and dy/dx.

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- **0.25.** Equation (0.48) is a polynomial, $y = a + bx + cx^2$. Find the first derivative dy/dx and the second derivative d^2y/dx^2 .
- **0.26.** Assuming that 48.2 mg of an acid of unknown structure was titrated with 0.0988 M NaOH, the following data (pH as a function of titrant volume *V*) were recorded. Find the endpoint volume by the second-derivative technique and calculate the equivalent weight of the acid (for a monoprotic acid, the equivalent weight equals the molecular weight):

$V(\mathrm{mL})$	pН
1.40	2.74
1.50	2.86
1.60	3.12
1.70	3.60
1.80	6.15
1.90	9.74
2.00	10.40
2.10	10.62
2.20	10.75

0.27. This equation arises in the study of the effect of pH on the rate of hydrolysis of many drugs, where k is an observed rate constant and k_1 , k_2 , k_3 are constants of the system:

$$k = k_1[\mathrm{H}^+] + k_2 + k_3[\mathrm{OH}^-]$$

Find the value of pH at which the rate of hydrolysis is a minimum.

0.28. Evaluate this definite integral.

$$y = 24 \int_{1}^{4} x^2 dx$$

- **0.29.** Derive an expression giving the time elapsed for the concentration in a first-order reaction to decrease to one-half its initial value. (This time is called the *half-life* of the reaction; it is symbolized $t_{1/2}$).
- **0.30.** Why is the constant of integration omitted when evaluating a definite integral?
- **0.31.** Write the total differential dw for the function w = f(x, y, z).
- **0.32.** The equilibrium constant K_{11} for the complexation of 4-nitrophenol anion with α -cyclodextrin (in water at 25°C) has been measured by many laboratories;

these are the reported values (units are M^{-1}): 2290, 2200, 2500, 2700, 1590, 2439, 1890, 2720, 2143, 2408, 2270, 3550. Calculate the mean, standard deviation, standard deviation of the mean, and the relative standard deviation.

0.33. The acetylation of isopropyl alcohol by acetic anhydride is a second-order reaction:

$$Ac_2O + (CH_3)_2CHOH \rightarrow HOAc + CH_3COOCH(CH_3)_2$$

It is described by this integrated rate equation

$$\log \frac{c_B}{c_A} = \frac{(c_B^0 - c_A^0)kt}{2.303} + \log \frac{c_B^0}{c_A^0}$$

where *A* is isopropyl alcohol and *B* is acetic anhydride. These are experimental kinetic data for this reaction:

t (min)	$c_A(M)$	$c_{\rm B}({\rm M})$
0	0.456	0.876
1.50	0.248	0.668
3.13	0.138	0.558
4.50	0.088	0.508
6.10	0.062	0.482
8.00	0.040	0.460
12.10	0.012	0.432

Analyze the data by the method of least squares, and report the value of the second-order rate constant k.

- **0.34.** These values have been reported in the literature for the dipole moment of phenol at 25°C: 1.45, 1.53, 1.54, 1.65, 1.72, 1.86, 1.53, 1.43, 1.64. Calculate the usual statistical parameters, and give 95% confidence limits for the mean.
- **0.35.** Return to Problem 0.32 and express the mean with an appropriate number of significant figures.
- **0.36.** The activation energy for the hydrolysis of aspirin in acid solution is $16.7 \text{ kcal} \text{ mol}^{-1}$. Convert this to kJ mol⁻¹.
- **0.37.** A rate constant for the uncatalyzed hydrolysis of succinylcholine chloride has been reported to be $5.0 \times 10^{-6} \text{ h}^{-1}$. Convert this to s⁻¹ (reciprocal seconds).
- **0.38.** Convert a density of 1.86 g mL^{-1} to SI units.
- **0.39.** The tetrahedral covalent radius of carbon is 0.77 A. Convert this to nanometers.

- **0.40.** R (the gas constant) is equal to $8.314 \,\mathrm{J}\,\mathrm{K}^{-1}\,\mathrm{mol}^{-1}$. Convert this to cal $\mathrm{K}^{-1}\,\mathrm{mol}^{-1}$.
- **0.41.** Physicochemical equations often call for taking the logarithm of a quantity having units. It is difficult to conceive of the logarithm of a unit. Describe a way out of this quandary.
- **0.42.** Beer's law for the absorption of light is written $A = \varepsilon bc$, where *b* is pathlength in centimeters, *c* is molar concentration, and *A* is absorbance, defined by $A = \log(I_0/I)$, where the *I* quantities are light intensities. Deduce the units of ε , which is called the *molar absorptivity*.
- **0.43.** Both of these equations have been published in the scientific literature (by coauthor KAC of the present book). One of them is incorrect. The quantities k_s , k_L , and k_{11} all have the same units. K_{11} is defined $K_{11} = [SL]/[S][L]$. The *F* quantities are fluorescent intensities. Use dimensional analysis to detect the incorrect equation.

(a)
$$\frac{F}{F_0} = \frac{1 + (k_{11}/k_s)K_{11}[L]}{1 + K_{11}[L]} + \frac{k_L}{K_s}[L]$$

(b)
$$\frac{F}{F_0} = \frac{1 + (k_{11}/k_s)K_{11}[L]}{1 + K_{11}[L]} + \frac{(k_L/k_s)[L]}{[S](1 + K_{11}[L])}$$

BASIC THERMODYNAMICS

1

ENERGY AND THE FIRST LAW OF THERMODYNAMICS

1.1. FUNDAMENTAL CONCEPTS

Temperature and the Zeroth Law. The concept of temperature is so familiar to us that we may not comprehend why scientists two centuries ago tended to confuse temperature with heat. We will start with the notion that temperature corresponds to "degree of hotness" experienced as a sensation. Next we assign a number to the temperature based on the observation that material objects (gases and liquids in particular) respond to "degree of hotness" through variations in their volumes. Thus we should be able to associate a number (its temperature) with the volume of a specified amount of material. We call the instrument designed for this purpose a thermometer.

The first requirement in setting up a scale of temperatures is to choose a zero point. In the common *Celsius* or *centigrade* scale we set the freezing point of water (which is also the melting point of ice) at 0 °C [more precisely, 0 °C corresponds to the freezing point of water (called the "ice point") in the presence of air at a pressure of 1 atmosphere (atm)]. The second requirement is that we must define the size of the degree, which is done for this scale by setting the boiling point of water (the "steam point") at 100 °C. The intervening portion of the scale is then divided linearly into 100 segments. We will let *t* signify temperature on the Celsius scale.

Experience shows that different substances may give different temperature readings under identical conditions even though they agree perfectly at 0 °C and 100 °C. For example, a mercury thermometer and an alcohol thermometer will not give precisely the same readings at (say) room temperature. In very careful work it would be

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advantageous to have available an "absolute" temperature scale that does not depend on the identity of the thermometer substance. Again we appeal to laboratory experience, which has shown that the dependence of the volume of a fixed amount of a gas on temperature, at very low pressures of the gas, is independent of the chemical nature of the gas. Later we will study the behavior of gases at low pressures in more detail; for the present we can call such gases "ideal gases" and use them to define an *absolute ideal-gas temperature scale.* We define the absolute temperature as directly proportional to the volume of a given mass of ideal gas at constant pressure (i.e., letting *T* be the absolute temperature and *V* the gas volume):

$$T \propto V$$

For convenience we define the size of the absolute temperature degree to be identical to the Celsius degree. If V_0 and V_{100} are the volumes of the ideal gas at the ice and steam points of water, respectively, the size of the degree is given by

$$\frac{V_{100} - V_0}{100}$$

Then our absolute temperature scale is defined by

$$T = \frac{V}{(V_{100} - V)/100} \tag{1.1}$$

Now suppose that we apply our ideal-gas thermometer to water at the ice point. In this special case, Eq. (1.1) becomes

$$T_0 = \frac{V_0}{(V_{100} - V_0)/100} \tag{1.2}$$

Careful experimental work with numerous gases has revealed that $T_0 = 273.15$ K. Thus the Celsius and absolute scales are related by

$$T = t + 273.15$$

The absolute temperature scale is also called the *thermodynamic scale* or the *Kelvin scale*, and temperatures on this scale are denoted K (pronounced Kelvin, with no degree symbol or word).

According to Eq. (1.1), when T = 0 K, V = 0; the volume of the ideal gas goes to zero at the *absolute zero*. Modern experimental techniques have achieved temperatures within microdegrees of the absolute zero, but T = 0 K appears to be an unattainable condition.

The concept and practical use of temperature scales and thermometers is based on the experimental fact that if two bodies are each in thermal equilibrium with a third body, they are in thermal equilibrium with each other. This is the *zeroth law of thermodynamics*.

Work and Energy. Let us begin with the mechanical concept of *work* as the product of a force and a displacement:

$$Work = Force \times Displacement$$
(1.3)

The units of work are consequently those of force and length. Now from Newton's laws of motion,

$$Force = Mass \times Acceleration$$
(1.4)

In SI units, force therefore has the units kg m s⁻², which is also called a newton, N. Hence the units of work are either kg m² s⁻² or N m.

Energy is defined as any property that can be produced from or converted into work (including work itself). Therefore work and energy have the same dimensions, although different units may be used to describe different manifestations of energy and work. For example, 1 Nm = 1 J (joule), and energy is often given in joules or kilojoules. Here are relationships to earlier energy (cgs) units:

$$1 \text{ J} = 10^7 \text{ erg}$$

4.184 J = 1 cal (calorie)

Note from the definition (1.3) that work is a product of an intensive property (force) and an extensive property (displacement). In general, work or energy can be expressed as this product:

Work (energy) = intensity factor
$$\times$$
 capacity factor (1.5)

Here are several examples of Eq. (1.5):

Mechanical work = Mechanical force × Distance Work of expansion = Pressure × Volume change Electrical work = Electric potential × Charge Surface work = Surface tension × Area change

All forms of work are, at least in principle, completely interconvertible. For example, one could use the electrical energy provided by a battery to drive a (frictionless) piston that converts the electrical work to an equivalent amount of work of expansion.

Heat and Energy. Heat has been described as *energy in transit* (Glasstone 1947, p. 7) or as a *mode of energy transfer* (Denbigh 1966, p. 18). Heat is that form of energy that is transferred from one place to another as a consequence of a difference in temperature between the two places. Numerically, heat is expressed in joules (J) or calories (cal). Heat is not "degree of hotness," which, as we have seen, is measured by temperature.¹

Since both work and heat are forms of energy, they are closely connected. Work can be completely converted into an equivalent amount of heat (e.g., through friction).

¹ Note that temperature is an intensive property, whereas heat is an extensive property. Two hot potatoes differing in size may have the same temperature, but the larger potato possesses more heat than the smaller one.

	Total energy of a b	ody	
Thermodynam	ic energy (U)	Mechanic	al energy
Kinetic energy (translational energy)	Internal energy (vibrational, rotational, and electronic energy)	Kinetic energy as a result of the body's motion as a whole	Potential energy as a result of the body's position

Table 1.1. The energy of a thermodynamic system

The converse is not possible, however; it is found experimentally that heat cannot be completely converted into an equivalent amount of work (without producing changes elsewhere in the surroundings). This point will be developed later; for the present we observe that this finding is the basis for the impossibility of a "perpetual-motion machine."

We find it convenient to divide energy into categories. This is arbitrary, but there is nothing wrong with it provided that we are careful to leave nothing out. Now, we have seen that thermodynamics is not built on the atomic theory; nevertheless, we can very usefully invoke the atomic and molecular structure of matter in our interpretation of energy. In this manner we view heat as thermal energy, equivalent to, or manifesting itself as, motions of atoms and molecules. The scheme shown in Table 1.1 clarifies the several "kinds" of energy that a body (the "system") can possess.²

Chemical thermodynamics is concerned with the energy U. This energy is a consequence of both (a) the *electronic* distribution within the material and (b) three types of atomic or molecular motion: (1) *translation*, the movement of individual molecules in space; (2) *vibration*, the movement of atoms or groups of atoms with respect to each other within a molecule; and (3) *rotation*, the revolution of molecules about an axis. If a material object is subjected to an external source of heat, so that the object absorbs heat and its temperature rises, the atoms and molecules increase their translational, vibrational, and rotational modes of motion. Energy is not a "thing" but is, instead, one way of describing and measuring these molecular and atomic distributions and motions, as well as the electronic distribution within atoms and molecules.

Systems and States. In order to carry out experimental studies and to interpret the results, we must focus on some part of the universe that interests us. In thermodynamics, this portion of the universe is called a *system*. The system typically consists of a specified amount of chemical substance or substances, such as a given mass of a gas,

² This scheme is consistent with the usage of most authors, but some variation is found in the literature. The thermodynamic energy U may also be symbolized E, and some authors label the thermodynamic energy the internal energy. The internal energy shown Table 1.1 may be identified with the potential energy of the molecules (to be distinguished from the potential energy of the body as a whole).

liquid, or solid. Whatever exists outside of the system is called the *surroundings*. Certain conditions give rise to several types of systems:

- *Isolated Systems.* These systems are completely uninfluenced by their surroundings. This means that neither matter nor energy can flow into or out of the system.³
- *Closed Systems.* Energy may be exchanged with the surroundings, but there can be no transfer of matter across the boundaries of the system.

Open Systems. Both energy and matter can enter or leave the system.

We can also speak of a *homogeneous* system, which is completely uniform in composition, or speak of a *heterogeneous* system, which consists of two or more phases.

Experiment has shown that the *state* of a system can be completely defined by specifying four observable thermodynamic variables: the composition, temperature, pressure, and volume. If the system is homogeneous and consists of a single chemical substance, only three variables suffice. Moreover, it is known that these three variables are not all independent; if any two are known, the third is thereby fixed. Thus the thermodynamic state of a pure homogeneous system is completely defined by specifying any two of the variables pressure (P), volume (V), and temperature (T). The quantitative relationship, for a given system, among P, V, and T is called an *equation of state*. Generally the equation of state of a system must be established experimentally.

The fact that the state of a system can be completely defined by specifying so few (two or three) variables constitutes a vast simplification in the program of describing physicochemical systems, because this means that all the other macroscopic physical properties (density, viscosity, compressibility, etc.) are fixed. We don't know their values, but we know that they depend only on the thermodynamic variables and therefore are not themselves independent. With this terminology we can now say that thermodynamics deals with changes in the energy U of a system as the system passes from one state to another state.

Thermodynamic Processes and Equilibrium. A system whose observable properties are not undergoing any changes with time is said to be in *thermodynamic equilibrium*. Thermodynamic equilibrium implies that three different kinds of equilibrium are established: (1) *thermal equilibrium* (all parts of the system are at the same temperature), (2) *chemical equilibrium* (the composition of the system is not changing), and (3) *mechanical equilibrium* (there are no macroscopic movements of material within the system).

Many kinds of processes can be carried out on thermodynamic systems, and some of these are of special theoretical or practical significance. *Isothermal processes* are

³ A truly isolated system is an idealization, but a very close approximation can be achieved inside a closed *thermos* (derived from the original trade name *Thermos* in 1907) bottle. (The laboratory version is called a *Dewar flask*.)

those in which the system is maintained at a constant temperature. (This is easy to do with a constant-temperature bath or oven.) Since it is conceivable that heat is given off or taken up by the system during the process, maintaining a constant temperature requires that the heat loss or gain be offset by heat absorbed from or given up to the surroundings. Thus an isothermal process requires either a closed or an open system, both of these allowing energy to be exchanged with the surroundings. An *adiabatic process* is one in which no heat enters or leaves the system. An adiabatic process requires an isolated system. Obviously, if the process is adiabatic, the temperature of the system may change.

A *spontaneous process* is one that occurs "naturally"; it takes place without intervention. For example, if a filled balloon is punctured, much of the contained gas spontaneously expands into the surrounding atmosphere. In an equilibrium chemical reaction, which we may write as

$$A + B \rightleftharpoons M + N$$

it is conventional to consider the reaction as occurring from left to right as written. Thus if the position of equilibrium favors M + N (the products), the reaction is said to be spontaneous. If the reactants (A + B) are favored, the reaction is nonspontaneous as written. (Obviously we can change these designations simply by writing the reaction in the reverse direction.)

It is the business of thermodynamics to tell us whether a given process is spontaneous or nonspontaneous. However, thermodynamics, which deals solely with systems at equilibrium, cannot tell us how fast the process will be. For example, according to thermodynamic results, a mixture of hydrogen and oxygen gases will spontaneously react to yield water. This is undoubtedly correct—but it happens that (in the absence of a suitable catalyst) the process will take millions of years.

There is one more important type of thermodynamic process: the *reversible process*. Suppose we have a thermodynamic system at equilibrium. Now let an infinitesimal alteration be made in one of the thermodynamic variables (say, T or P). This will cause an infinitesimal change in the state of the system. If the alteration in the variable is reversed, the process will reverse itself exactly, and the original equilibrium will be restored. This situation is called *thermodynamic reversibility*. Reversibility in this sense requires that the system always be at, or infinitesimally close to, equilibrium and that the infinitesimally small alterations in variables be carried out infinitesimally slowly. Because of this last factor, thermodynamically reversible processes constitute an idealization of real processes, but the concept is theoretically valuable. One feature of a reversible process is that it can yield the maximum amount of work; any other (irreversible) process would generate less work, because some energy would be irretrievably dissipated (e.g., by friction).

Now suppose that a system undergoes a process that takes it from state A to state B:

$$A \longrightarrow B$$

We define a change in some property Q of the system by

$$\Delta Q = Q_{\rm B} - Q_{\rm A} \tag{1.6}$$

In other words, the incremental change in the property is equal to its value in the final state minus its value in the initial state.

Next consider this series of processes, which constitutes a thermodynamic cycle:

$$A \xrightarrow{1} B \qquad \Delta Q_1 = Q_B - Q_A$$

$$4 \xrightarrow{1} Q_2 = Q_C - Q_B$$

$$\Delta Q_2 = Q_C - Q_B$$

$$\Delta Q_3 = Q_D - Q_C$$

$$\Delta Q_4 = Q_A - Q_D$$
Sum: $\Delta Q = 0$

In any cycle in which the system is restored exactly to its original state, the total incremental change is zero.

1.2. THE FIRST LAW OF THERMODYNAMICS

Statement of the First Law. To this point we have been establishing a vocabulary and some basic concepts, and now we are ready for the first powerful thermodynamic result. This result is solidly based on extensive experimentation, which tells us that although energy can be converted from one form to another, it cannot be created or destroyed [this statement is completely general in the energy regime characteristic of chemical processes; relativistic effects (i.e., the famous equation $E = mc^2$) do not intrude here]. This is the great conservation of energy principle, which is expressed mathematically as Eq. (1.7), the first law of thermodynamics.

$$\Delta U = q - w \tag{1.7}$$

Here ΔU is the change in thermodynamic energy of the system, q is the amount of energy gained by the system as heat, and w is the amount of energy lost by the system by doing work on its surroundings. These are the sign conventions that we will use:

- q is positive if the heat is taken up by the system (i.e., energy is gained by the system).
- w is positive if work is done by the system (i.e., energy is lost by the system).⁴

Equation (1.7) is the incremental form of the first law. The differential form is

$$dU = dq - dw \tag{1.8}$$

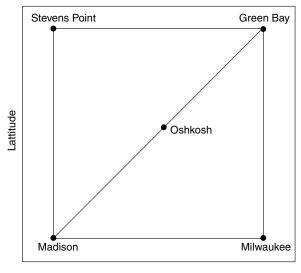
⁴ This is the sign convention used by most authors, but the International Union of Pure and Applied Chemistry (IUPAC) reverses the convention for w, giving as the first law $\Delta U = q + w$.

But now we must make a very clear distinction between the quantity dU and the quantities dq and dw. U is a *state function* and dU is an *exact differential*. This terminology means that the value of ΔU , which is obtained by integrating dU over the limits from the initial state to the final state, is independent of the path (i.e., the process or mechanism) by which the system gets from the initial state to the final state. A state function depends only on the values of the quantity in the initial and final states.

It is otherwise with q and w, for these quantities may be path-dependent. For example, the amount of work done depends on the path taken (e.g., whether the process is reversible or irreversible). Therefore dq and dw are not exact differentials, and some authors use different symbols to indicate this. Nevertheless, although q and w individually may be path-dependent, the combination q-w is independent of path, because it is equal to ΔU .⁵

The Ideal Gas. Experimental measurements on gases have shown that as the pressure is decreased, the volume of a definite amount of gas is proportional to the

⁵ This analogy will clarify the difference between path-dependent and path-independent quantities. Suppose we wish to drive from Madison (WI) to Green Bay. Obviously there are numerous routes we might take. We could drive via Milwaukee, or via Oshkosh, or via Stevens Point, and so on. Graphically, the possibilities can be represented on a map, as shown in the accompanying figure. Now, no matter which path we take, the changes in latitude, Δ Lat, and in longitude, Δ Lon, will be exactly the same for each route; for example, Δ Lat = Lat(GB) – Lat(MAD), and this quantity is independent of the route. Thus latitude and longitude are state functions. But the amount of gasoline consumed, the time spent driving, and the number of miles driven all depend on the path taken; these are not state functions. This analogy is taken from Smith (1977).



Longitude

reciprocal of the pressure:

$$V \propto \frac{1}{P}$$

As P is decreased toward zero, all gases (at constant temperature) tend to behave in the same way, such that Eq. (1.9) is satisfied:

$$PV = \text{constant}$$
 (1.9)

This result can be generalized as Eq. (1.10), which is called the *ideal-gas equation* (or the ideal-gas law):

$$PV = nRT \tag{1.10}$$

where *P*, *V*, and *T* have their usual meanings; *n* is the number of moles of gas; and *R* is a proportionality constant called the *gas constant*. Equation (1.10) is the equation of state for an ideal gas (sometimes called the "perfect gas"), and it constitutes a description of real-gas behavior in the limit of vanishingly low pressure. The macroscopic properties of an ideal gas such as pressure, volume, and temperature are related to the continuous motion of the gas molecules. A theory called the kinetic theory of gases has been developed to derive these properties and their relationship (ideal gas law) from first principles. Details on the kinetic theory of gases can be found in Appendix B.

Example 1.1. Experiment has shown that 1 mol of an ideal gas occupies a volume of 22.414 L at 1 atm pressure when T = 273.15 K. Calculate R:

$$R = \frac{PV}{nT} = \frac{(1 \text{ atm})(22.414 \text{ L})}{(1 \text{ mol})(273.15 \text{ K})}$$
$$= 0.082057 \text{ L atm mol}^{-1} \text{ K}^{-1}$$

We can use a dimensional analysis treatment to convert to other energy units, as described in Chapter 0:

$$R = \left(\frac{0.082057 \text{ L atm}}{\text{mol K}}\right) \left(\frac{101, 325 \text{ Pa}}{1 \text{ atm}}\right) \left(\frac{1 \text{ N m}^{-2}}{1 \text{ Pa}}\right) \left(\frac{10^3 \text{ cm}^3}{1 \text{ L}}\right) \left(\frac{1 \text{ J}}{1 \text{ Nm}}\right) \left(\frac{1 \text{ m}}{10^2 \text{ cm}}\right)^3$$
$$= 8.3144 \text{ J mol}^{-1} \text{K}^{-1}$$

and since 1 cal = 4.184 *J*, R = 1.987 cal mol⁻¹ K⁻¹. Notice that in this calculation of *R*, its units are energy per mol per K. That is, since R = PV/nT, the units of the product *PV* are energy, which we expressed in the particular units L atm, J, or cal. These several values of *R* are widely tabulated, and they can serve as readily accessible conversion factors among these energy units.

We earlier mentioned a type of work called *work of expansion*. This is the work done by a gas when it expands against a resisting pressure, as happens when a piston moves in a cylinder. We can obtain a simple expression for work of expansion. Suppose that a piston of cross-sectional area A moves against a constant pressure P. We know that mechanical work is the product of force (F) and distance, or

$$w = F(L_2 - L_1)$$

where L_1 is the initial position of the piston and L_2 is its final position. Pressure is force per unit area (A), so F = PA, giving

$$w = PA(L_2 - L_1)$$

But $A(L_2 - L_1) = V_2 - V_1$, where V_1 and V_2 are volumes, so

$$w = P(V_2 - V_1) = P \Delta V$$
 (1.11)

where ΔV is the volume displaced. Thus work of expansion is the product of the (constant) pressure and the volume change; in fact, we often refer to work of expansion as $P\Delta V$ work.

Now, if the process is carried out reversibly, so that the pressure differs only infinitesimally from the equilibrium pressure, the volume change will be infinitesimal, and Eq. (1.11) can be written

$$dw = P \, dV \tag{1.12}$$

We can integrate this between limits:

$$w = \int_{V_1}^{V_2} P \, dV \tag{1.13}$$

(In the case of an isothermal, reversible expansion, *w* does not depend upon the path, but this is a special case.) Now suppose that the gas is ideal and that the process is carried out isothermally. From the ideal gas law, P = nRT/V, so

$$w = nRT \int_{V_1}^{V_2} \frac{dV}{V}$$
(1.14)

$$w = nRT \ln \frac{V_2}{V_1} \tag{1.15}$$

If $V_2 > V_1$, the system does work on the surroundings, and *w* is positive. If $V_1 > V_2$, the surroundings do work on the system, and *w* is negative.

In developing Eq. (1.15) we saw an example of thermodynamic reasoning, and we obtained a usable equation from very sparse premises. Here is another example, again based on the ideal gas. Suppose that such a gas expands into a vacuum. Since the resisting pressure is zero, Eq. (1.11) shows that w = 0; that is, no work is done. Careful experimental measurements by Joule and Kelvin in the nineteenth century showed that there is no heat exchange in this process, so q = 0. The first law therefore tells us that $\Delta U = 0$. Since the energy depends on just two variables, say, volume and temperature,

we can express the result as

$$\left(\frac{\partial U}{\partial V}\right)_T = 0 \tag{1.16}$$

which says that the energy of an ideal gas is independent of its volume at constant temperature. We can interpret this thermodynamic result in molecular terms as follows. A gas behaves ideally when the intermolecular forces of attraction and repulsion are negligible. (This is why real gases approach ideality at very low pressures, because then the molecules are so far apart that they do not experience each others' force fields.) If there are no forces between the molecules, no energy is required to change the intermolecular distances, and so expansion (or compression) results in no energy change.

1.3. THE ENTHALPY

Definition of Enthalpy. In most chemical studies we work at constant pressure. (The reaction vessel is open to the atmosphere, and P = 1 atm, approximately.) Consequently the system is capable of doing work of expansion on the surroundings. From the first law we can write $q = \Delta U + w$; and since $w = P \Delta V$, we have

$$q = \Delta U + P \Delta V$$

at constant P. Writing out the increments, we obtain

$$q = (U_2 - U_1) + P(V_2 - V_1)$$

Rearranging, we have

$$q = (U_2 + PV_2) - (U_1 + PV_1)$$
(1.17)

where U, P, and V are all state functions. We define a new state function H, the *enthalpy*, by

$$H = U + PV \tag{1.18}$$

giving, from Eq. (1.17), the following:

$$q = \Delta H \tag{1.19}$$

Although Eq. (1.18) defines the enthalpy, it is usually interpreted according to Eq. (1.19), because we can only measure *changes* in enthalpy (as with all energy quantities). The enthalpy change is equal to the heat gained or lost in the process, at constant pressure (there is another restriction, namely, that work of expansion is the only work involved in the process). Since enthalpy is an energy, it is measured in the usual energy units.

From Eq. (1.18) we can write

$$\Delta H = \Delta U + P \Delta V \tag{1.20}$$

For chemical processes involving only solids and liquids, ΔV is usually quite small, so $\Delta H \approx \Delta U$; but for gases, where ΔV may be substantial, ΔH and ΔU are different. We can obtain an estimate of the difference by supposing that 1 mol of an ideal-gas is evolved in the process. From the ideal gas law we write

$$P\Delta V = (\Delta n)RT$$

For 1 mol we obtain $\Delta n = 1$, so from Eq. (1.20) we have

$$\Delta H = \Delta U + RT$$

At 25 °C, this gives

$$\Delta H = \Delta U + (1.987 \text{ cal mol}^{-1} \text{ K}^{-1})(298.15 \text{ K})$$

= $\Delta U + 592 \text{ cal mol}^{-1}$

which is a very appreciable difference.

When a chemical process is carried out at constant pressure, the heat evolved or absorbed, per mole, can be identified as ΔH . Specific symbols and names have been devised to identify ΔH with particular processes. For example, the heat absorbed by a solid on melting is called the *heat of fusion* and is labeled ΔH_m , or ΔH_f . The *heat* of solution is the enthalpy change per mole when a solute dissolves in a solvent. For a chemical reaction, ΔH is called a *heat of reaction*. The heat of reaction may be positive (heat is absorbed) or negative (heat is evolved). By writing a reaction on paper in reverse direction its ΔH changes sign. For example, this reaction absorbs heat:

$$6C(s) + 3H_2(g) \longrightarrow C_6H_6(l)$$
 $\Delta H = +11.7 \text{ k cal mol}^{-1}$

This reaction, its reverse, therefore evolves heat:

$$C_6H_6(l) \longrightarrow 6C(s) + 3H_2(g) \quad \Delta H = -11.7 \text{ k cal mol}^{-1}$$

We will later see how enthalpy changes for chemical processes can be measured.

Heat Capacity. A quantity C, called the heat capacity, is defined as

$$C = \frac{dq}{dT} \tag{1.21}$$

C is a measure of the temperature change in a body produced by an increment of heat. The concept of the heat capacity is essential in appreciating the distinction between heat and temperature.

Chemical processes can be carried out at either constant volume or constant pressure. First consider constant volume. If only work of expansion is possible, at constant volume we obtain $\Delta V = 0$, so w = 0, and from the first law we have dq = dU. We therefore define the heat capacity at constant volume by

$$C_V = \left(\frac{\partial U}{\partial T}\right)_V \tag{1.22}$$

At constant pressure, on the other hand, we have, from Eq. (1.19), dq = dH, and we define the heat capacity at constant pressure by

$$C_P = \left(\frac{\partial H}{\partial T}\right)_P \tag{1.23}$$

In the preceding section we had obtained, for one mole of an ideal gas, Eq. (1.24).

$$\Delta H = \Delta U + RT \tag{1.24}$$

Let us differentiate this with respect to temperature. Using Eqs. (1.22) and (1.23), we get

$$C_P = C_V + R \tag{1.25}$$

For argon, at room temperature, $C_P = 20.8 \text{ J K}^{-1} \text{ mol}^{-1}$ and $C_v = 12.5 \text{ J K}^{-1} \text{ mol}^{-1}$; hence $C_P - C_V = 8.3 \text{ J K}^{-1} \text{ mol}^{-1}$, which is *R*.

For most compounds only C_P has been measured. Values of C_P for typical organic compounds lie in the range 15–50 cal K⁻¹ mol⁻¹. As seen here, heat capacity is expressed on a per mole basis, and is sometimes called the *molar heat capacity*. When the heat capacity is expressed on a per gram basis it is called the *specific heat*.

Taking the constant-pressure condition of Eq. (1.23) as understood, we can write $C_P = dH/dT$, or $dH = C_P dT$. If we suppose that C_P is essentially constant over the temperature range T_1 to T_2 , integration gives

$$\Delta H = C_P \,\Delta T \tag{1.26}$$

Example 1.2. The mean specific heat of water is $1.00 \text{ cal g}^{-1} \text{ K}^{-1}$. Calculate the heat required to increase the temperature of 1.5 L of water from 25 °C to the boiling point.⁶

As a close approximation we may take the density of water as 1.00 g mL^{-1} and the boiling point as $100 \,^{\circ}$ C, so from Eq. (1.26) we obtain

$$\Delta H = \left(\frac{1.00 \text{ cal}}{\text{g K}}\right) (1500 \text{ g})(75 \text{ K}) = 112,500 \text{ cal}$$

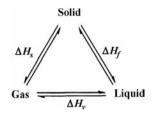
or 112.5 kcal.

⁶ It is not a coincidence that the specific heat of water is 1.00 cal $g^{-1} K^{-1}$, because this is how the calorie was originally defined: One calorie was the amount of heat required to raise the temperature of one gram of water by 1 °C. Actually the specific heat of water varies slightly with the temperature.

PROBLEMS

- **1.1.** A piston 3.0 in. in diameter expands into a cylinder for a distance of 5.0 in. against a constant pressure of 1 atm. Calculate the work done in joules.
- **1.2.** What is the work of expansion when the pressure on 0.5 mol of ideal gas is changed reversibly from 1 atm to 4 atm at 25 °C? (*Hint:* For an ideal gas $P_1V_1 = P_2V_2$.)
- **1.3.** Derive an equation giving the heat change in the isothermal reversible expansion of an ideal gas against an appreciable pressure. [*Hint:* Make use of Eq. (1.16) and the first law.]
- **1.4.** What is the molar heat capacity of water? (See Example 1.2 for the specific heat.)
- **1.5.** The molar heat capacity of liquid benzene is $136.1 \,\mathrm{J}\,\mathrm{mol}^{-1}\,\mathrm{K}^{-1}$. What is its specific heat?
- **1.6.** The specific heat of solid aluminum is $0.215 \text{ cal g}^{-1} \text{ K}^{-1}$. If a 100-g block of aluminum, initially at 25 °C, absorbs 1.72 kcal of heat, what will be its final temperature?
- **1.7.** A 500-g piece of iron, initially at 25 °C, is plunged into 0.5 L of water at 75 °C in a Dewar flask. When thermal equilibrium has been reached, what will the temperature be? The specific heat of iron is $0.106 \text{ cal g}^{-1} \text{ K}^{-1}$.
- **1.8.** A thermally unstable drug needs to be mailed to a patient. The drug is put into a 100-mL bottle, and the bottle is put in a 2-L Styrofoam box. Dry ice (solid carbon dioxide) is added to the box in the amount of 352 g to cool the drug and maintain it cold until it reaches its destination. Dry ice has the property of sublimating upon heating. No liquid carbon dioxide is formed. The box is sealed and it represents a thermodynamic closed system. Assume that the initial temperature of dry ice is its sublimation temperature, -78 °C. The package takes two days to be delivered. Every day, the package absorbs 600 J of heat from the environment. Calculate the pressure inside the box when the package reaches its destination (neglect the amount of air initially present in the container) and the temperature of the drug when the package is opened. Dry ice heat of sublimation = 416 J/mol, molecular weight of dry ice = 44 g/mol, density of dry ice = 1.56 g/mL.
- **1.9.** A sample of an unknown gas at atmospheric pressure weighs 2.3 g/L at the temperature of 25 °C. Calculate the molecular weight of the gas. Using the information in Appendix B, calculate the square root velocity of the gas molecules at 25 °C. Express the velocity in cm/s.
- **1.10.** Consider 3 mol of an ideal gas having a molecular weight of 32 g/mol. The gas is at the temperature of 300 °C and at the pressure of 6.3 atm. The gas is then expanded until it reaches a pressure of 0.001 atm. What is the gas density in g/mL in the initial and final states?

- **1.11.** An experimenter wishes to determine the partial pressure of chloroform required to anesthetize a 28.0-g mouse in a 2.37-L container at $20 \,^{\circ}$ C. If 2.00 mL of liquid chloroform is introduced into the closed vessel through a valve, what is the partial pressure of chloroform in the container? Assume complete evaporation of the chloroform. Calculate the partial pressure using the ideal gas equation. Assume the density of the mouse to be 1 g/mL. The density of liquid chloroform at $20 \,^{\circ}$ C is 1.484 g/mL.
- **1.12.** Nitrous oxide (N₂O) is used for the rapid induction of anesthesia. Using the ideal gas equation, calculate the molecular weight of this gas, given that 1 L at the temperature of 0 °C and at the pressure of 760 mmHg weighs 1.97 g. In addition, calculate the root mean square velocity, μ , of nitrous oxide and from μ calculate the density of gaseous N₂O at 1 atm and 0 °C. (See Appendix B).
- **1.13.** In the following thermodynamic cycle, ΔH_f , ΔH_v and ΔH_s , are, respectively, molar heats of fusion, vaporization, and sublimation for a pure substance. Obtain an equation connecting these three quantities. (*Hint:* Pay careful attention to the directions of the arrows.)



2

THE ENTROPY CONCEPT

2.1. THE ENTROPY DEFINED

Why Energy Alone Is Not a Sufficient Criterion for Equilibrium. Let us try to develop an analogy, based on what we know from classical mechanics, between a mechanical system and a chemical (thermodynamic) system. The position of equilibrium in a mechanical system is controlled by potential energy. Consider a rock poised near the top of a hill. It possesses potential (gravitational) energy as a consequence of its position. If it is released, its potential energy will be converted to heat (through friction) and to kinetic energy as it rolls down the hill. It will come to rest, having zero potential energy, at the foot of the hill (since we can measure only *changes* in energy, we mean that the potential energy is zero relative to some arbitrary reference value, which we are free to take as the value at the foot of the hill). It is now at mechanical equilibrium. Thus the criterion for a spontaneous mechanical process is that the change in potential energy be negative (it gets smaller), and the criterion for mechanical equilibrium is that the change in potential energy be zero.

Why don't we simply apply an analogous criterion to chemical systems? We might argue that ΔU (for a system at constant volume) or ΔH (for a system at constant pressure) plays the role of potential energy in the mechanical system. But we find experimentally that this suggestion is inadequate to account for the observations. Consider first the following experiment (Smith 1977, p. 6):

1. Dissolve some solid NaOH in water. The solution becomes warm; that is, heat is liberated in the process. This means that ΔH is negative in the spontaneous

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process of NaOH dissolving in water. (The reaction is said to be *exothermic.*) This is entirely in accord with the proposal we are examining.

2. Dissolve some solid NaNO₃ in water. The solution becomes cool; that is, heat is absorbed as the dissolution occurs, and this cools the solution. Therefore ΔH is positive in this spontaneous process. (It is an *endothermic* reaction.) This behavior is in conflict with the proposal.

Here is another pertinent experiment. Suppose that we have two identical chambers connected by a stopcock. With the stopcock closed, we let one chamber contain a gas and the other chamber be evacuated (i.e., it "contains" a vacuum). Now we open the stopcock. We know what will happen—the gas will spontaneously distribute itself uniformly throughout the two chambers. If the gas is ideal (and most gases behave nearly ideally at low pressures), we know [see Eq. (1.16)] that $\Delta U = 0$ for this spontaneous process. Thus, with *no energy change at all*, the system spontaneously underwent a change to an equilibrium position.

This inability to predict the direction of chemical change based on energy considerations alone was one of the great nineteenth-century scientific problems. Since energy minimization alone is not an adequate criterion for chemical equilibrium, *something else* must be involved. This is our next concern, and we are going to use an approach somewhat different from that taken in many textbooks, which adopt an argument based on the historical development of the ideas. We are going to sidestep classical thermodynamic history by turning to a description based on the particulate (i.e., atomic) nature of matter.

The Statistical Mechanical Entropy. We have seen that classical thermodynamics is based on macroscopic observations and makes no assumptions about the ultimate structure of matter. An alternative viewpoint, called *statistical mechanics* (or *statistical thermodynamics* when applied to thermodynamic problems), adopts the assumption that matter is composed of vast numbers of very small particles (which we now identify as electrons, atoms, molecules, etc.). In many circumstances this point of view provides physical insight not available from classical thermodynamics, and we will turn to it to illuminate our present problem.

Let us reconsider the example of the apparatus with two chambers, in one of which a gas was initially confined. Suppose that only a single molecule of gas had been present. After the stopcock is opened (and presuming that both chambers have equal volumes), evidently the probability that the molecule will be in one specified chamber (say, the left chamber) is $\frac{1}{2}$. Next suppose we were to start with two molecules—say, a and b—and ask for the probability that both will be found, at equilibrium, in the left chamber. These are the only possible distributions:

Left	Right
a	b
b	а
a,b	
	a,b

Thus, of four possible distributions, only one places both a and b in the left chamber, so the probability¹ of this distribution is $\frac{1}{4} = (\frac{1}{2})^2$. Generalizing to *N* molecules, we get $(\frac{1}{2})^n$ for the probability that at equilibrium all *N* molecules will be found in the left chamber. Since for chemical systems the number of atoms or molecules, *N*, can be very large indeed, we see that the probability is extremely small that all of the molecules will end up in one chamber. On the other hand, the probability is extremely high that the molecules will be distributed equally between the two chambers.

This simple example (Glasstone 1947, p. 184) suggests a general statement, which in fact constitutes a basic premise of statistical mechanics, namely, that *all spontaneous processes represent changes from a less probable to a more probable state.* This postulate leads us to the next stage of our inquiry, which consists essentially of counting all possible distributions that are accessible to a system, because this is how the probability of a state is to be established.

In this next example the system is more complicated, although still artificially simple. We imagine that two crystals of different elements, A and B, are placed in contact, so that atoms of A may diffuse into the B crystal and vice versa [this example is given by Denbigh (1966, p. 49)]. In this simple example we suppose that crystal A contains four A atoms (4A), and likewise crystal B contains four B atoms (4B). We can distinguish between A and B atoms, but all A atoms are indistinguishable among themselves, and similarly for B. The sites that the atoms occupy in the crystals are distinguishable. Initially, let all A atoms be in the left-hand crystal and let all B atoms be in the right-hand crystal.

We are going to count all possible configurations (called *microstates*) of our system. There are 4A and 4B to be distributed among eight sites. (We assume that the energies of interaction are identical no matter which type of atom is on which site.) Clearly, there is only one microstate having 4A in the left crystal and 4B in the right crystal:

A	А	В	В
Α	А	В	В

Similarly, there is only one microstate with 4B in the left and 4A in the right crystal.

But now consider the number of ways we can have 3A + 1B on the left and 3B + 1A on the right. We could argue in this way: The A atom on the right has any one of four right-hand sites available to it, and likewise the B atom on the left has four sites available, making $4 \times 4 = 16$ configurations. These 16 microstates are explicitly shown in Fig. 2.1. Obviously the symmetrical arrangement of 1A + 3B on the left and 3A + 1B on the right will also have 16 microstates.

The remaining arrangement of 2A + 2B (left) and 2A + 2B (right) is slightly more difficult. First consider the left crystal. The first B atom has four sites available, whereas the second B atom has only three accessible sites. Hence there appear to be 4×3 possible configurations. However, the two B atoms are indistinguishable, so we

¹ We are making the unstated assumption that the molecules behave independently, so that each has a probability of $\frac{1}{2}$ of being in the left chamber. The probability that both will be in the left chamber is the product of the individual probabilities.

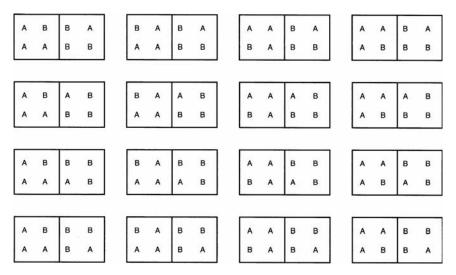


Figure 2.1. The 16 microstates possessing 3A + 1B on the left and 3B + 1A on the right.

have double-counted, and must compensate, giving $(4 \times 3)/2$ as the number of microstates. But an equal number is contributed by the right-hand side, making in all $(4 \times 3)/2 \times (4 \times 3)/2 = 36$ microstates. Here are the results summarized.²

Atoms to Left	Atoms to Right	Number of Microstates
4A	4B	1
3A + 1B	1A + 3B	16
2A + 2B	2A + 2B	36
1A + 3B	3A + 1B	16
4B	4A	1
		70

In modern terminology, the microstates are called quantum states.

² Probability theory gives a simple expression for calculating the number of ways N objects can be distributed into n_1 of type 1, n_2 of type 2, and so on; this is the expression:

$$\frac{N!}{n_1!n_2!\dots}$$

where N! is read N factorial and is the product $1 \times 2 \times 3 \times 4 \times ... (N-1) \times N$. For our system this gives 8!/4! 4! = 70.

Now, another key premise of statistical mechanics is that the system is as likely to be in any one microstate as in another. That is, *all microstates are equally probable*. In our example above, there is a probability of $\frac{1}{35}$ that all of the A atoms will be found in a single crystal (either left or right); but there is a probability of $\frac{36}{70}$ that the atoms will be uniformly distributed. All the microstates are accessible, and the system is simply more likely to be found (at equilibrium) in the state possessing the largest number of microstates. (It can also be stated that the system spends an equal amount of time in each microstate, so it spends the most time in the system with the most microstates.)

For chemical systems the number of particles is extremely large (recall that Avogadro's number is about 6×10^{23}), so the number of microstates is vast, and the consequence is that the most probable state of the system is so probable that all other states (although possible in principle) may be disregarded in practice. The total number of microstates accessible to a system (which, we have just noted, is essentially equal to the number of microstates in the most probable state) we label *W*. (Some authors use Ω .) We now define a quantity *S*, called *entropy*, by Eq. (2.1), which is due to Boltzmann:

$$S = k \ln W \tag{2.1}$$

This is a definition. We will later establish the significance of the proportionality constant k. The equation says that the entropy S of a system increases logarithmically as W, the number of accessible microstates, increases.

We have noticed in our crystal diffusion example how W is composed of contributions from various configurations, and within each configuration the contributions are multiplicative; for example, for the 3A + 1B (left) and 1A + 3B (right) state we had $4 \times 4 = 16$ microstates. If, more generally, we write $W = W_L \times W_R$, then Eq. (2.1) gives us

$$S = k \ln W_{\rm R} = k \ln W_{\rm L} + k \ln W_{\rm R} = S_{\rm L} + S_{\rm R}$$

Thus, entropy is additive. (This is one reason why Boltzmann used a logarithmic function in his definition.)

Another point is to be made here. Our crystal diffusion example involved microstates all having the same energy, and we calculated all possible configurations. The resulting entropy is known as the *configurational entropy*. More generally, in chemical systems we must also consider a very large number of quantum states, most of which occupy different energy levels. The total entropy receives contributions from both sources: the microstates counting all configurations and those counting all energies.

Summarizing to this point, we conclude that spontaneous processes occur in a direction of increasing probability and that entropy as calculated by the statistical mechanical definition is a quantitative measure of this probability. Therefore *spontaneous processes occur with an increase in entropy.*³

³ In making this statement we are neglecting concurrent energy changes; specifically, we are assuming $\Delta U = 0$ (for the present). Note also that we measure *changes* in entropy, ΔS , so the statement says that if ΔS is positive, the process is spontaneous.

Before leaving the statistical mechanical treatment, let us apply our results to the calculation of the entropy change accompanying the isothermal expansion of an ideal gas from volume V_1 to volume V_2 (Glasstone 1947, p. 186; Denbigh 1966, p. 55; Rossini 1950, p. 73). (We will shortly see the point of this particular calculation.) Recall that $\Delta U = 0$ in this process, so the only driving force for the expansion is the increase in probability of the system.

If W_1 and W_2 are the numbers of microstates associated with volumes V_1 and V_2 , then $S_1 = k \ln W_1$ and $S_2 = k \ln W_2$, so

$$\Delta S = S_2 - S_1 = k \ln \frac{W_2}{W_1} \tag{2.2}$$

The probability that a single molecule will be found in any volume V is proportional to that volume, and the number of microstates accessible to a molecule is proportional to V (Hill 1960, Chapter 4). For a single molecule we therefore can write, $W_2/W_1 = V_2/V_1$, and for N_A (one mole of) molecules we have

$$\frac{W_2}{W_1} = \left(\frac{V_2}{V_1}\right)^{N_A} \tag{2.3}$$

Combining Eqs. (2.2) and (2.3) gives as the statistical mechanical result

$$\Delta S = k N_A \ln \frac{V_2}{V_1} \tag{2.4}$$

Entropy in Classical Thermodynamics. Now we are going to treat the isothermal reversible expansion of an ideal gas classically. Our goal is to establish the classical thermodynamic equivalent of the statistical mechanical entropy. We begin with the first law:

$$dU = dq - dw \tag{2.5}$$

On expanding from volume V_1 to volume V_2 against pressure P, the gas is capable of doing work of expansion dw = P dV. Moreover, we know from our earlier discussion that dU = 0 for this process, so we have dq = P dV. For one mole of an ideal gas we have P = RT/V, giving dq = RT(dV/V), or

$$\frac{dq}{T} = R \frac{dV}{V} \tag{2.6}$$

We will integrate Eq. (2.6) between our expansion limits of V_1 and V_2 , giving

$$\int_{\text{state 1}}^{\text{state 2}} \frac{dq}{T} = R \ln \frac{V_2}{V_1}$$
(2.7)

Now let us compare Eq. (2.7), derived classically, with Eq. (2.4), derived statistically. These equations describe the same process, and they reveal that consistency between

the classical and statistical treatments can be achieved by writing the identities

$$kN_A = R \tag{2.8}$$

$$\int_{\text{state1}}^{\text{state2}} \frac{dq}{T} = \Delta S \tag{2.9}$$

and Eq. (2.9) implies

$$\frac{dq}{T} = dS \tag{2.10}$$

These are powerful results. From Eq. (2.8) we achieve a physical interpretation of the proportionality constant *k* in Eq. (2.1), Boltzmann's definition of entropy, as

$$k = \frac{R}{N_A} \tag{2.11}$$

where k is the gas constant per molecule; this quantity is known as the *Boltzmann* constant. It has the value $k = 1.38 \times 10^{-23}$ J K⁻¹. Any equation containing R is on a per mole basis; replace the R with k and the equation is on a per molecule basis.

According to Eq. (2.10), the differential entropy change is equal to the differential heat change divided by the absolute temperature. Moreover, from Eq. (2.7), since *V* is a state function, the entropy *S* is a state function. With a combination of statistical and classical arguments we can make some general statements about entropy changes. From statistical mechanics we had seen that ΔS increases during a spontaneous process, so we infer that $\Delta S = 0$ at equilibrium. Reverting to a differential symbolism, these results give us

$$dS > 0$$
 for a spontaneous (irreversible) process
 $dS = 0$ for a system at equilibrium

Recall that in a reversible process the system is always virtually at equilibrium, and the system is then capable of performing the maximum work (because irreversible losses, such as to friction, are minimized). In a spontaneous (irreversible) process, the amount of work that can be done is less than the maximum. From the first law, since dU is a state function and is the same no matter what path is taken, we have

$$dU = dq_{\rm rev} - dw_{\rm rev} = dq_{\rm irr} - dw_{\rm irr}$$

so

$$dq_{\rm rev} - dq_{\rm irr} = dw_{\rm rev} - dw_{\rm irr}$$

Since $dw_{\rm rev} > dw_{\rm irr}$, it follows that $dq_{\rm rev} > dq_{\rm irr}$. We therefore can write

$$\frac{dq_{\rm rev}}{T} > \frac{dq_{\rm irr}}{T}$$

The entropy is a state function, independent of path, so the differential dS has a definite value for a given process regardless of whether that process is carried out reversibly. Equation (2.10) for the classical definition of entropy can be more explicitly written

$$dS = \frac{dq_{\rm rev}}{T}$$

Classically, then, the entropy increase is equal to the heat change in an isothermal reversible process divided by the absolute temperature at which the heat change occurs. All spontaneous (i.e., natural) processes occur with a gain of entropy by the system and the surroundings. Note that it is conceivable for the system to experience an entropy decrease (dS < 0), but this will inevitably be accompanied by a more-than-compensating entropy increase in the surroundings.

2.2. THE SECOND LAW OF THERMODYNAMICS

Statement of the Second Law. Entropy plays a critical role in thermodynamic analysis, because it is the missing factor that we were seeking to allow us to predict the direction of change in atomic or molecular systems. The essential result constitutes the *second law* of thermodynamics, which can be stated in several ways, not all of them obviously equivalent, but in fact all of them providing the same message. Here are some of them:

- 1. Heat does not spontaneously flow from a cold body to a hot body.
- 2. Spontaneous processes are not thermodynamically reversible.
- 3. The complete conversion of heat into work is impossible without leaving some effect elsewhere.
- 4. It is impossible to convert heat into work by means of a constant temperature cycle.
- 5. All natural processes are accompanied by a net gain in entropy of the system and its surroundings.

This last statement is most useful to us. Let us write

$$dS_{\rm net} = dS_{\rm system} + dS_{\rm surroundings}$$

Then the second law says

 $dS_{\text{net}} > 0$ (spontaneous process) $dS_{\text{net}} = 0$ (reversible process)

Interpretations of Entropy. Entropy is an abstract concept of thermodynamics and statistical mechanics that plays a practical role in providing a criterion for equilibrium.

Despite its technical and abstract nature, it has passed into popular culture and language, where its use is sometimes casual and inexact. Let us consider some interpretations that have been given to entropy. The statistical mechanical picture is clearest. We found that the entropy increases logarithmically with the number of microstates accessible to the system, and we concluded that entropy is correlated with the increase in "mixed-up-ness" of the system [Denbigh (1966, p. 55) attributes this term to Gibbs]. Entropy is widely interpreted as a measure of randomness or of *disorder*, an increase in entropy being associated with an increase in these properties. This is because spontaneous processes occur with an increase in entropy and lead to more extensive mixing of the units in a system. This interpretation directly concerns the configurational entropy, which measures the spatial disposition of units; in addition, there is the thermal entropy, which measures the distribution of quantum states having different energies. (But note that an increase in configurational entropy might conceivably be accompanied by a decrease in thermal entropy; it is the net entropy change that is decisive.) E. A. Guggenheim [cited by Denbigh (1966, p. 56)] refers to entropy as a measure of spread-that is, dispersion over a larger number of quantum states, either configurational or thermal.

A fundamental basis of the second law is closely connected to these interpretive notions. As we have seen, it is possible to convert work completely into heat, but we cannot completely convert heat into work. The reason for this dissymmetry lies in the atomic structure of matter. Doing work means making use of the *directed* motion of an assemblage of particles (as by rubbing a metal block on a surface, or drilling a hole in a solid with a drill bit). This work is converted (through friction) to heat, which raises the temperature of the contacting bodies. The temperature increase reflects the increased kinetic energy of the atoms in the bodies, and (this is the essential point) the motions of these atoms are undirected, because they are largely chaotic. Without adding energy from the surroundings, there is no possible way to transform completely this *undirected* motion (heat) back into work. The basis of this irreversibility is the increased randomness on the atomic scale. There is a modern tendency to describe this phenomenon (increased spread or randomness, therefore increased entropy) as reflecting a loss of information about the system, but this usage is not helpful and may be misleading. Let us consider this further.

Some authors give as examples of processes that involve an increase in entropy (and a corresponding loss of information) the change of a new deck of cards to a randomly shuffled deck, or the change from the orderly arrangement of books and papers in a student's room at the beginning of the semester to a disorderly distribution of these objects about the room by the time of final exams. But we can easily be led astray by such examples, which as analogies to the atomic scale are erroneous. There are two factors to be kept in mind: (1) We earlier reached the conclusion that *spontaneous processes occur with an increase in entropy*. But a deck of cards does not spontaneously shuffle itself into randomness, nor do the books in a student's room spontaneously adopt new locations. So the examples were incomplete; they omitted to consider the second factor: (2) The *system and its surroundings* must both be taken unto account. In the case of the playing cards, the cards themselves constitute the sytem; the shuffler of the cards constitutes the effective surroundings, and the

Law	State Function	Characteristic
0	Т	Determines thermal equilibrium
1	U	Conservation of energy (The energy of the universe is constant) (You can't get something for nothing)
2	S	Determines direction of spontaneous change (The entropy of the universe is increasing) (You can't break even)

Table 2.1. The laws of thermodynamics

surroundings did work on the system; this was not a spontaneous process. Nor was there any change in the number of microstates available to the cards before and after shuffling, so there was no entropy increase in the cards.

What is the essential difference between a collection of playing cards and a collection of atoms and molecules? Fundamentally, it is a matter of size. Atoms and molecules are in incessant motion because of their thermal energy; through such motion they undergo collisions resulting in energy exchange and distribution, and they spontaneously adopt configurations with larger number of microstates. Cards, however, are macroscopic objects; their movement in space is not spontaneous, but rather requires action by an external agent (which *does* experience an entropy increase in the process because of the accompanying dispersal of energy). Lambert (1999, 2002) has given interesting discussions of these issues.

Summary of Fundamental Thermodynamics. Our development of the first and second laws of thermodynamics has provided the entire basis of this subject. Everything else (and there is a great deal more) follows from this by introducing definitions of new quantities or functions and manipulating them mathematically. Before we proceed, we summarize our results⁴ in Table 2.1.

2.3. APPLICATIONS OF THE ENTROPY CONCEPT

Entropy Relationships A few simple manipulations will demonstrate the involvement of entropy in thermodynamic relationships. The first law is dU = dq - dw. If the only work done in a process is work of expansion, then dw = P dV. Moreover, we have seen that dS = dq/T, so dq = T dS, and we get

$$dU = T \, dS - P \, dV \tag{2.12}$$

Die Energie der Welt ist constant.

⁴ The concept of entropy was introduced by Clausius in 1854, and he introduced the word *entropy* in 1865. This is how Clausius expressed the first and second laws:

Die Entropie der Welt strebt einem Maximum zu.

as another statement of the first law. The product TdS (or $T\Delta S$) is pervasive in thermodynamics, and this is its source; observe that this product is an energy.

Now rearrange Eq. (2.12) to

$$dS = \frac{dU + P \, dV}{T} \tag{2.13}$$

and consider processes at constant pressure. From the definition of enthalpy applied to Eq. (2.12) we find

$$dS = \frac{dH}{T} \tag{2.14}$$

We recall that the heat capacity at constant pressure is defined $C_P = dH/dT$, so $dH = C_P dT$. Using this in Eq. (2.14) gives

$$dS = C_P \frac{dT}{T} \tag{2.15}$$

where the constant pressure condition is understood and is not explicitly indicated. We can integrate Eq. (2.15) between the limits T_1 and T_2 :

$$\Delta S = S_2 - S_1 = \int_{T_1}^{T_2} C_P \frac{dT}{T} = \int_{T_1}^{T_2} C_P d \ln T$$
(2.16)

If C_P is substantially independent of temperature over the integration range, Eq. (2.16) becomes

$$\Delta S = C_P \ln \frac{T_2}{T_1} \tag{2.17}$$

at constant pressure [a corresponding equation, $\Delta S = C_v \ln (T_2/T_1)$, applies at constant volume].

An interesting case of Eq. (2.16) arises when we set $T_1 = 0$ K, giving

$$S = S_0 + \int_0^T C_P d \ln T$$
 (2.18)

The quantity S_0 is to be interpreted as the value of the entropy at the absolute zero. Planck in 1912 proposed that S_0 may assume the value zero at 0 K for a perfect crystal, which possesses no disorder. This proposal is known as the *third law of thermodynamics*. By means of the third law combined with Eq. (2.18), it is possible to evaluate the entropy S of substances from measurements of C_P as a function of temperature. The procedure is to plot experimental values of C_P against ln T for the entire range of experimental temperatures. Since T = 0 K is unattainable, the curve thus generated is extrapolated to 0 K with the aid of a theoretical function. The area under the curve, from 0 K up to any specified temperature, is then equal to the entropy of the substance at that temperature.

Entropy Changes. Despite the possibility afforded by the third law to evaluate absolute entropies of substances, in nearly all practical applications of the entropy concept we evaluate *changes* in entropy. Here we will see some examples of such determinations. Later in the book we will consider the estimation of entropy changes for additional types of processes.

From the definition dS = dq/T it is evident that the units of entropy are energy per Kelvin, and it is expressed either in J K⁻¹ or cal K⁻¹. Since entropy is an extensive property, we convert it to an intensive property by expressing it on a per mole basis. Consequently, ΔS values will always be encountered in the units J K⁻¹ mol⁻¹ or cal K⁻¹ mol⁻¹ (the combination cal K⁻¹ mol⁻¹ is sometimes referred to as the *entropy unit*, abbreviated e.u.).

We will calculate the entropy changes accompanying phase changes, as when a solid melts (fusion) or a liquid evaporates (vaporization). These processes can be carried out reversibly at constant temperature (the temperature being called the melting point, T_m , for fusion, or the boiling point, T_b , for vaporization.)⁵ The system is not isolated, because heat must be supplied in order that the process take place. The heat supplied in the fusion process is ΔH_f , the heat of fusion; whereas ΔH_v , the heat of vaporization, is furnished in the vaporization process. These enthalpy changes are expressed on a per mole basis. Many experimental ΔH_f and ΔH_v values are available in the common reference handbooks.

From Eq. (2.14) applied to our present concern we can write

$$\Delta S_f = \frac{\Delta H_f}{T_m} \tag{2.19}$$

$$\Delta S_{\nu} = \frac{\Delta H_{\nu}}{T_b} \tag{2.20}$$

Table 2.2 shows enthalpy data for a few phase changes.

Substance	mp^a (°C)	ΔH_f (kcal mol ⁻¹)	bp^b (°C)	$\Delta H_{\nu} (\mathrm{kcal} \mathrm{mol}^{-1})$
Benzoic acid	122.1	4.32		_
Phenol	40.9	2.75		_
Acetone		_	56.2	6.95
Water	0.0	1.436	100.0	9.717

Table 2.2. Heats of fusion and vaporization for some solids and liquids

^{*a*}Melting Point.

^bBoiling Point.

 5 The boiling point is commonly considered to be the temperature at which the liquid and vapor are in equilibrium at atmospheric pressure. However, Eq. (2.20) can also be applied to data at other pressures, with the appropriate temperature inserted.

Substance	ΔS_f (cal K ⁻¹ mol ⁻¹)	$\Delta S_{\nu} (\operatorname{cal} \mathrm{K}^{-1} \mathrm{mol}^{-1})$
Benzoic acid	+ 10.93	_
Phenol	+8.76	
Acetone		+21.11
Water	+5.26	+26.04

Table 2.3. Entropies of fusion and vaporization

Example 2.1. Calculate the entropy of fusion of benzoic acid.

$$\Delta S_f = \frac{\Delta H_f}{T_m} = \frac{4320 \text{ cal mol}^{-1}}{395.25 \text{ K}} = +10.93 \text{ cal K}^{-1} \text{ mol}^{-1}$$

Table 2.3 gives ΔS_f and ΔS_v results for the processes described in Table 2.2.

It has been known since 1884 that for very many nonassociated liquids (i.e., liquids whose molecules do not interact strongly with each other), $\Delta S_{\nu} \approx 21 \text{ cal } \text{K}^{-1} \text{mo1}^{-1}$. This empirical observation is known as *Trouton's rule*, and it provides a simple though approximate estimate of ΔH_{ν} , by means of Eq. (2.20), since the boiling point is easily measured. Such a convenient generalization cannot be made for ΔS_f values, although some definite patterns have been observed [see Yalkowski and Valvani (1980); in Chapter 10 we make use of these observations].

Notice that all ΔS_f and ΔS_v values are positive, because the system in each case is proceeding from a state of relative order to a state of relative disorder. Molecules in the liquid state possess a larger number of accessible quantum states (both configurational and thermal) than in the more restricted solid state, and similarly for the vaporization process.

We will subsequently learn how to calculate ΔS for chemical reactions, where we will find that ΔS can be either positive or negative, just as with ΔH values, depending on the direction in which the reaction is written. Very generally we anticipate that if the product state (the right-hand side of the equation) possesses more particles (molecules or ions) than the reactant state, ΔS will be positive, reflecting the availability to the system of more microstates.

PROBLEMS

- **2.1.** Predict the sign of ΔS for these processes.
 - (a) Crystallization of benzoic acid from its melt.
 - (b) Evaporation of spilled gasoline.
 - (c) This chemical reaction

 $Me_2C = CH_2 + Cl_2 \rightarrow Me_2CClCH_2Cl$

- **2.2.** Look up the boiling point of benzene, and estimate its molar heat of vaporization.
- **2.3.** The heat of fusion of 4-nitroaniline is $5.04 \text{ kcal mol}^{-1}$. Look up its melting point, and calculate its entropy of fusion.
- **2.4.** Sublimation is the process in which a solid is transformed directly to the vapor state. The heat of sublimation of naphthalene is $17.6 \text{ kcal mol}^{-1}$ at $25 \degree \text{C}$. Calculate its entropy of sublimation.
- **2.5.** Calculate the entropy change during the isothermal expansion of 0.5 mol of an ideal gas from 100 mL to 1 L.
- **2.6.** The heat capacity of chloroform in the vicinity of 600 K is $20.4 \text{ cal } \text{K}^{-1} \text{ mol}^{-1}$. Calculate the entropy change per mole when chloroform is brought from 550 K to 625 K.
- **2.7.** Derive an equation for the molar entropy change when the pressure on an ideal gas is isothermally changed from P_1 to P_2 atm. [*Hint:* Start with Eq. (2.4).]
- **2.8.** Inhalation of a low-boiling-point anesthetic is a common way of inducing general anesthesia. A commonly used gaseous anesthetic is isoflurane (1-chloro-2,2,2-trifluoroethyl difluoromethyl ether). Its boiling point is 48.5 °C and its molecular weight is 184.50 g mol⁻¹. In a certain anesthetic procedure, 92.25 g of isoflurane is mixed with 90% pure oxygen by blowing a flow of oxygen at 25 °C on the liquid isoflurane. The final mixture, at the pressure of 1 atm, contains all the isoflurane and 144 g of oxygen. Calculate the change in entropy of isoflurane in going from a liquid state at 25 °C to a vapor state at 25 °C mixed with oxygen. Assume that all heat needed for the vaporization of isoflurane is given from the surrounding environment (open system) and that the isoflurane heat of vaporization is 275 cal mol⁻¹.
- **2.9.** A thermos jug contains 108 mL of water at the temperature of 13 °C. Ice (90 g) at the temperature of -3 °C is added Assume no exchange of energy and mass with the environment. Calculate the change in entropy of the system if the process proceeds at constant pressure. Heat capacity of liquid water = 18 cal mol⁻¹, density of liquid water = 1 g mL⁻¹, heat of fusion of ice = 1437 cal mol⁻¹, heat capacity of ice = 9 cal mol⁻¹, and molecular weight of water = 18 g mol⁻¹.
- **2.10.** A 500-g piece of iron at the temperature of 978 °C is plunged into 200 mL of water at the temperature of 25 °C in a Dewar flask. Knowing that the specific heat of iron is $0.106 \text{ cal g}^{-1} \text{ K}^{-1}$, atomic weight of iron = 56 g mol⁻¹, heat of vaporization of liquid water = 10,400 cal mol⁻¹, and molecular weight of water = 18 g mol⁻¹, specific heat of water = 1 cal g⁻¹ K⁻¹ (assume this is the specific heat for both liquid and gaseous (C_P) water, density of water = 1 g mL⁻¹, boiling point of water = 100 °C, calculate the final temperature of the iron and the change in entropy of the system.

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2.11. A block of ice weighing 360 g at the temperature $-20 \,^{\circ}$ C is poured in a bucket containing 4L of water at 31 $^{\circ}$ C. Consider the system to be in adiabatic conditions. What is the change in entropy of the system? Molecular weight of water = 18 g mol⁻¹, density of water = 1 g mL⁻¹, heat capacity of ice = 9 cal mol⁻¹, heat capacity of liquid water = 18 cal mol⁻¹, and heat of fusion of water = 1437 cal mol⁻¹.

3

THE FREE ENERGY

3.1. PROPERTIES OF THE FREE ENERGY

The Gibbs Free Energy. We have seen that for a mechanical system (which consists of relatively few bodies or "particles") the condition for a spontaneous process is that the potential energy change be negative, whereas for a chemical system (which consists of an almost unimaginably large number of particles) we learned that, even when no energy change occurs, spontaneous processes can take place, and we concluded that spontaneous processes occur with an increase in entropy. Now we are going to bring this together, recognizing that there are two factors involved in determining the direction of chemical change: The system seeks to minimize its energy *and* to maximize its entropy, and the position of equilibrium depends upon a combination of (and perhaps a compromise between) these factors. Several thermodynamic functions have been proposed to describe the situation, but we will make use of only one of these, which is particularly useful for our purposes because it invokes the commonly controlled experimental conditions of temperature and pressure. This function, termed *Gibbs free energy G*,¹ is defined as follows:

$$G = H - TS \tag{3.1}$$

¹ After J. Willard Gibbs, a physicist at Yale University, who provided much of the theoretical development of thermodynamics and statistical mechanics in the second half of the nineteenth century. The *Helmholtz free energy A*, defined A = U - TS, is more useful than *G* under conditions of constant volume. We will not make use of *A* (but see also note 2).

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This equation is actually a definition. Since H, T, and S are state functions, G is also a state function. As seen from its definition, the Gibbs free energy (which is often referred to simply as the "free energy" for convenience) is an energy quantity. We are, for the present, restricting attention to a closed system, which is one across whose boundaries no matter is exchanged with the surroundings.

Since by definition H = U + PV, Eq. (3.1) can be written

$$G = U + PV - TS \tag{3.2}$$

and its complete differential is

$$dG = dU + P \, dV + V \, dP - T \, dS - S \, dT \tag{3.3}$$

We saw earlier [Eq. (2.12)] that if the only work done in a reversible process is work of expansion, the first law can be written

$$dU = T \, dS - P \, dV \tag{3.4}$$

which, combined with Eq. (3.3), gives

$$dG = V \, dP - S \, dT \tag{3.5}$$

Equation (3.5) shows how the free-energy change depends on changes in the pressure and the temperature for a reversible process in a closed system.² If the temperature is constant, dT = 0, and from Eq. (3.5) we obtain

$$\left(\frac{\partial G}{\partial P}\right)_T = V \tag{3.6}$$

 2 Each of the functions *U*, *H*, *G*, and *A* can be written in parallel form as a function of two variables, namely (for closed systems)

$$dU = T dS - P dV$$
$$dH = T dS + V dP$$
$$dA = -S dT - P dV$$
$$dG = V dP - S dT$$

These all contain the same information, but it is because *G* is expressible as a function of the variables *P* and *T* that we find it especially useful. If a system is at equilibrium, any infinitesimal change is reversible. The preceding four relationships, which are called *characteristic functions*, provide equivalent criteria for equilibrium. From the first one, at constant entropy and volume (i.e., dS = 0, dV = 0), the condition for equilibrium is dU = 0. From the second, at constant entropy and pressure, dH = 0 defines equilibrium; from the third, at constant temperature and volume, dA = 0 at equilibrium; finally, at constant pressure and temperature, the condition for equilibrium is dG = 0.

If the pressure is constant, dP = 0 and from Eq. (3.5) we obtain

$$\left(\frac{\partial G}{\partial T}\right)_P = -S \tag{3.7}$$

From the definition Eq. (3.1), if the temperature is constant, we obtain

$$dG = dH - T \, dS \tag{3.8}$$

or, in incremental form,

$$\Delta G = \Delta H - T \,\Delta S \tag{3.9}$$

This last equation is an especially useful relationship because experimentally we measure these incremental quantities. Observe in this equation how ΔG , the change in free energy, is composed of an energy component, ΔH , and an entropic term, $-T\Delta S$.

We can obtain some insight into the meaning of free energy from the following development. We can write the work done by or on the system as

$$dw = dw_{\text{expansion}} + dw_{\text{additional}}$$

where $dw_{\text{expansion}} = P \, dV$ and $dw_{\text{additional}}$ represents work other than $P \, dV$ work (such as electrical work). The first law is dU = dq - dw, and for a reversible process we have $dq = T \, dS$. Combining these relationships gives

 $-dw_{\text{additional}} = dU + P \, dV - T \, dS$

But dU + P dV = dH, so finally, by comparison with Eq. (3.8), we have $-dG = dw_{additional}$.

It is for this reason that the free energy change is said to be a measure of the maximum work available from a process (exclusive of work of expansion). That is, $-dG = dw - P \, dV$. When the system can do no useful work, dG = 0; a spontaneous process has a negative value of dG (or of ΔG). In a chemical reaction the approach to the position of equilibrium may be from either direction, depending on the initial conditions (i.e., the concentration of the reacting species). Figure 3.1 shows this schematically.

The essential characteristic of the Gibbs free-energy function is its combination of both the energy and entropy components in a form that reveals how these two thermodynamic concepts compete to generate a compromise that determines the position of equilibrium in a chemical process.³ A more negative ΔH favors

³ It is not too fanciful to draw an analogy with a political science setting, in which each society must choose its own compromise position between the extremes of maximum security (the energy component) and maximum liberty (the entropy component).

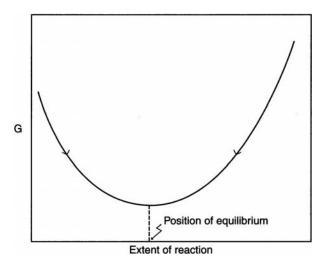


Figure 3.1. Free energy of a reacting chemical system, showing how the direction of the reaction depends on the initial state of the system.

spontaneous reaction, and a more positive ΔS favors spontaneous reaction, in both instances by making ΔG more negative.

We are now in a position to better understand our earlier calculations of entropies of fusion and vaporization. These systems were at equilibrium, so $\Delta G = 0$, and, from Eq. (3.9), $\Delta S = \Delta H/T$.

Pressure Dependence of the Free Energy. From either Eq. (3.5) or Eq. (3.6), at constant temperature, we obtain

$$dG = -V \, dP \tag{3.10}$$

Now let us consider the special case of one mole of an ideal gas, so PV = RT and V = RT/P, giving

$$dG = RT \frac{dP}{P} = RT d \ln P \tag{3.11}$$

Integrating between the limits of P_1 and P_2 , we obtain

$$\Delta G = RT \ln \frac{P_2}{P_1} \tag{3.12}$$

which could alternatively have been integrated to the form

$$G = G^* + RT \ln P \tag{3.13}$$

where G^* is the constant of integration.

Although Eqs. (3.12) and (3.13) apply only to ideal gases, the mathematical form of these equations turns out to be ubiquitous, and we will subsequently encounter the form of Eq. (3.13) in several contexts.

Temperature Dependence of the Free Energy. If the definition G = H - TS is combined with Eq. (3.7), we obtain

$$G = H + T \left(\frac{\partial G}{\partial T}\right)_P \tag{3.14}$$

Next divide through by T^2 and rearrange to the form of Eq. (3.15):

$$-\frac{G}{T^2} + \frac{1}{T} \left(\frac{\partial G}{\partial T}\right)_p = -\frac{H}{T^2}$$
(3.15)

Now we call attention to the nonobvious fact that the left-hand side of Eq. (3.15) is equal to the derivative d(G/T)/dT

$$\frac{d(G/T)}{dt} = -\frac{G}{T^2} + \frac{1}{T} \left(\frac{\partial G}{\partial T}\right)_P$$
(3.16)

Combining Eqs. (3.15) and (3.16) therefore yields

$$\left[\frac{\partial(G/T)}{\partial T}\right]_{P} = -\frac{H}{T^{2}}$$
(3.17)

The incremental form of Eq. (3.17) is

$$\left[\frac{\partial(\Delta G/T)}{\partial T}\right]_{P} = -\frac{\Delta H}{T^{2}}$$
(3.18)

Equations (3.14), (3.17), and (3.18) are equivalent forms of the *Gibbs–Helmholtz* equation. We will later make use of Eq. (3.18).

3.2. THE CHEMICAL POTENTIAL

Definition of the Chemical Potential. All of the relationships that we have seen to this point deal with closed systems (no matter can enter or leave the system) in complete internal equilibrium (no chemical reactions are occurring in the system). But of course we are very interested in chemical reactions, and we would also like to be able to describe open systems, in which matter may be exchanged between the system and its surroundings. In order to do this, we expand our concept of the free energy to include the amounts (numbers of moles) of the chemical constituents of

the system, writing

$$G = f(T, P, n_1, n_2, \ldots)$$
 (3.19)

where n_1 is the number of moles of chemical substance 1, and so on. From Eq. (3.19) we write out the total differential

$$dG = \left(\frac{\partial G}{\partial T}\right)_{P,n_1,n_2,\dots} dT + \left(\frac{\partial G}{\partial P}\right)_{T,n_1,n_2,\dots} dP + \left(\frac{\partial G}{\partial n_1}\right)_{T,P,n_2,\dots} dn_1 + \left(\frac{\partial G}{\partial n_2}\right)_{T,P,n_1,\dots} dn_2 + \cdots$$
(3.20)

where the subscripts indicate the quantities that are held constant during the evaluation of the partial derivatives. We have already dealt with the partial derivatives $(\partial G/\partial T)$ and $(\partial G/\partial P)$, and now we turn our attention to the new quantities appearing in Eq. (3.20). These partial derivatives are called *partial molar free energies*. They have this significance: They represent the change in the total free energy of the system when one mole of constituent i (i = 1, 2, ...) is added while T, P, and all other constituent amounts are held constant. This quantity is so important that it has been given the special name *chemical potential* and its own symbol μ . Thus we define⁴

$$\left(\frac{\partial G}{\partial n_i}\right)_{T,P,n_j \neq n_i} = \mu_i \tag{3.21}$$

Now let us rewrite Eq. (3.20), making use of Eqs. (3.6), (3.7) and (3.21):

$$dG = -S dT + V dP + \mu_1 dn_1 + \mu_2 dn_2 + \cdots$$
 (3.22)

which can be written more succinctly as

$$dG = -S dT + V dP + \sum \mu_i dn_i \qquad (3.23)$$

The chemical potential μ is an intensive property, its units being energy per mole, as can be seen from its definition, Eq. (3.21).

Now let us consider Eq. (3.22) at constant temperature and pressure:

$$dG_{T,P} = \mu_1 \, dn_1 + \mu_2 \, dn_2 + \cdots \tag{3.24}$$

On integration this gives

$$G_{T,P} = n_1 \mu_1 + n_2 \mu_2 + \cdots$$
 (3.25)

⁴ Partial molar quantities are sometimes indicated with the conventional letter symbol and a bar above it, so the chemical potential may also be written \overline{G}_i .

which can be generally differentiated to give

$$dG_{T,P} = (n_1 d\mu_1 + \mu_1 dn_1) + (n_2 d\mu_2 + \mu_2 dn_2) + \cdots$$
 (3.26)

which is rearranged to

$$dG_{T,P} = (n_1 d\mu_1 + n_2 d\mu_2 + \dots) + (\mu_1 dn_1 + \mu_2 dn_2 + \dots)$$
(3.27)

Comparison of Eqs. (3.24) and (3.27) leads to

$$n_1 d\mu_1 + n_2 d\mu_2 + \dots = 0 \tag{3.28}$$

This last equation is called the *Gibbs–Duhem equation*.

Dependence of Chemical Potential on Pressure. We had earlier applied Eq. (3.6), reproduced here, to establish the dependence of the free energy of a closed system on pressure [Eq. (3.13)]:

$$\left(\frac{\partial G}{\partial P}\right)_T = V \tag{3.29}$$

We are now interested in mixtures, that is, systems of more than one substance, so we must make use of chemical potentials (partial molar free energies). We state without derivation the analog to Eq. (3.9), which is intuitively evident⁴:

$$\left(\frac{\partial \mu_i}{\partial P}\right)_T = \overline{V_i} \tag{3.30}$$

where $\overline{V_i}$ is the *partial molar volume* of substance *i*. The physical interpretation of this quantity is that it is the volume per mole of *i* at the composition specified. (In general, $\overline{V_i}$ is not equal to V_i , the molar volume of pure *i*, because of intermolecular interactions in the mixture.)

Now let us consider a mixture of ideal gases. From Eq. (3.30), the variation in chemical potential for constituent *i* can be written $d\mu_i = \overline{V_i} dP$, where *P* is the total pressure. Since the total number of moles *n* in the ideal-gas equation PV = nRT is just the sum $(n_1 + n_2 + \cdots)$, we obtain

$$V = (n_1 + n_2 + \cdots) \frac{RT}{P}$$

so the partial molar volume of *i* is

$$\overline{V_i} = \left(\frac{\partial V}{\partial n_i}\right)_{T,P,n_j \neq n_i} = \frac{RT}{P}$$
(3.31)

which is nicely simple (because we are dealing with ideal gases). Therefore we can write

$$d\mu_i = \overline{V_i} \, dP = RT \frac{dP}{P} = RT \, d\ln P \tag{3.32}$$

For a mixture of ideal gases, we have

$$p_i = x_i P \tag{3.33}$$

where p_i is the partial pressure of gas *i* and x_i is its mole fraction. At constant x_i , therefore, $d \ln p_i = d \ln P$, giving from Eq. (3.32)

$$d\mu_i = RT \, d \ln p_i \tag{3.34}$$

which is integrated to

$$\boldsymbol{\mu}_i = \boldsymbol{\mu}_i^* + RT \ln p_i \tag{3.35}$$

where μ_i^* is the constant of integration. According to Eq. (3.35), the chemical potential of *i* is logarithmically related to its partial pressure. The value of μ_i^* can be evaluated by setting $p_i = 1$ atm; then we see that μ_i^* is the chemical potential of gas *i* when $p_i = 1$ atm.

Although these ideas seem rather remote from our main interests, they are leading to an important result. In particular, Eq. (3.35) and its predecessor Eq. (3.13) possess the general form

$$\mu_i = \text{constant} + RT \ln(\text{composition variable})$$

which will recur in important contexts. We will also have to pay some attention to the constant term.

The Fugacity. In the development leading to Eq. (3.35), we supposed that we were dealing with a mixture of ideal gases, and the resulting expression for the chemical potential was very simple. In real circumstances, gases are not ideal (although at low pressures their behavior may closely approach ideality). We therefore must accept that Eq. (3.35) will not be an exact description for real-gas mixtures. The simplicity of the equation is so attractive, however, that standard practice is to preserve the form of the equation by replacing the partial pressure p_i with a quantity symbolized f_i and called the *fugacity*. Thus Eq. (3.35) applied to real gases becomes

$$\mu_i = \mu_i^* + RT \ln f_i \tag{3.36}$$

The fugacity may be thought of as a measure of the "escaping tendency" of the gaseous constituent (consider Latin *fugio*, to flee; Italian *fuggire*, to flee; French *fugace*,

fleeting; English *fugitive*). Again μ_i^* is the value of μ_i when the logarithmic term vanishes, that is, when $f_i = 1$. Of course, when $f_i = 1$, p_i probably does not equal 1 for a real gas, but as the pressure becomes smaller, p_i , and f_i approach each other, and ultimately as $p_i \rightarrow 0$ we obtain $f_i/p_i \rightarrow 1$. Thus the ratio f_i/p_i is a measure of the extent of nonideal behavior of gas *i* in the mixture.

Experimental methods are available for the measurement of fugacities. We will not pursue this aspect of the problem, except to note that fugacity has the units of pressure.

Activity and Activity Coefficient. We now turn to liquid mixtures, which are of great importance in pharmaceutical, chemical, and biological systems. Equation (3.36) applies to each constituent in a liquid solution because (as we will prove in Chapter 4) at equilibrium the chemical potential of each constituent is equal in the liquid phase and in the vapor phase in contact with it; therefore the fugacity f_i of component *i* is the same in the liquid and the vapor phases. But it is more convenient to express the chemical potential in a liquid solution in terms of a quantity having units more familiar than those of pressure. We therefore build on the foregoing developments, anticipating that the form of Eq. (3.36) is applicable, to write, for constituent *i* in a liquid mixture,

$$\mu_i = \mu_i^0 + RT \ln a_i \tag{3.37}$$

This equation is of great importance to us. The quantity a_i , is called the *activity* of constituent *i*, μ_i is its chemical potential, and μ_i^0 is the *standard chemical potential* of *i*. Evidently, $\mu_i^0 = \mu_i$ when $a_i = 1$. Of course, the activity a_i , and the chemical potential potential μ_i depend on the conditions of temperature, pressure, and composition of the system.

At this stage in our treatment the activity is still a concept without a context. Let us relate this concept to an experimental observable by focusing attention on a solution of solute *i* in a liquid solvent. Let *c* be the concentration (in mol L^{-1}) of the solute. Then we write the activity of *i* as

$$a_i = \gamma_i c_i \tag{3.38}$$

where γ_i , a proportionality constant, is called the *activity coefficient*. Combining Eqs. (3.37) and (3.38), we obtain

$$\mu_i = \mu_i^0 + RT \ln \gamma_i c_i \tag{3.39}$$

Why do we need the activity and the activity coefficient at all? Why not just write

$$\mu_i = \mu_i^0 + RT \ln c_i \tag{3.40}$$

The answer to these questions is that real solutions do not behave ideally (just as real gases do not behave ideally). When treating gases we replaced the pressure p_i with the fugacity f_i and saw that the ratio f_i/p_i is a measure of nonideal behavior.

Now in treating liquids we replace the concentration c_i , with the activity a_i , and use the ratio $a_i/c_i = \gamma_i$, as a measure of nonideal behavior. The source of the nonideal behavior is the noncovalent forces of interaction between molecules and ions. These interactions perturb the chemical and physical properties of the molecules characteristic of their isolated states, when they are sufficiently far apart that they are not sensibly affected by other particles. We therefore expect that deviations from ideal behavior will become greater as the molecules are forced closer together, which will happen as the pressure increases (for gases) or as the concentration increases (for liquids).

Let us now return to our consideration of Eqs. (3.37) and (3.39). At the moment we cannot use these equations, because the only quantity that we presumably know is the concentration c_i . It is necessary to introduce some definitions and to adopt some conventions. First, here are the definitions:

- The *standard state* (with respect to constituent *i*) is that state of the system in which $a_i = 1$; then $\mu_i = \mu_i^0$.
- The *reference state* (with respect to constituent *i*) is that state of the system in which $\gamma_i = 1$; then $a_i = c_i$.

From this point on, the rigor with which applications are made depends on the level of accuracy required in the results. Here are conventions that provide realistic approximations for practical calculations that do not require the highest accuracy.

- 1. The fugacity of a *gas* may be taken equal to its pressure (or partial pressure) in atm, at low to moderate pressures.
- 2. The activity of a liquid *solvent* in a solution is equal to its mole fraction. It follows that the activity of a pure liquid is 1.00.
- 3. The activity of a pure *solid* is 1.00. (This is consistent with convention 2.)
- 4. The activity of a *solute* in an infinitely dilute liquid solution (concentrations of $\sim \le 10^{-4}$ M may be considered infinitely dilute for this purpose) may be taken equal to its molar concentration. Thus, $\gamma_i = 1.00$; the solute is in its reference state.
- 5. The activity coefficient of an uncharged solute may be taken as 1.00 at any concentration (because uncharged molecules experience much weaker forces of interaction than do ions).

The reference state of a solute is usually taken to be the infinitely dilute solution, so that $\gamma_i \rightarrow 1$ as $c_i \rightarrow 0$. (Activity coefficients of ions are more complex than has been implied by our treatment, as a consequence of the impossibility of experimentally studying an ionic solution of either a cation or an anion by itself.) Table 3.1 (Harned and Owen 1958, pp. 484, 488) lists some experimental values of activity coefficients to give a sense of the extent to which solution behavior may depart from ideality. It is

	γ	
m^a	HCl	NaCl
0.001	0.965	
0.002	0.952	_
0.005	0.928	0.928
0.007	_	0.917
0.01	0.905	0.903
0.02	0.875	0.873
0.03	_	0.851
0.04	_	0.835
0.05	0.831	0.822
0.06	_	0.812
0.08	_	0.794
0.10	0.797	0.780

Table 3.1. Activity coefficients of hydrochloric acid and sodium chloride in aqueous solution at 25 $^\circ\text{C}$

^aMolality.

Source: Harned and Owen (1958, pp. 484, 488).

also possible to calculate theoretically the activity coefficients of ions by means of the Debye–Huckel theory. Chapter 8 treats ionic activity coefficients in more detail.

In most of the practical situations of interest to us we deal with mixtures, so the proper notation and terminology consists of μ , the chemical potential, and $\Delta\mu$, the change in chemical potential. For pure substances, there is no difference between μ and G, the molar free energy, or between $\Delta\mu$ and ΔG . In common practice we tend to be careless and to use G and ΔG where we really should be using μ and $\Delta\mu$, but this should cause no confusion. Note, however, that when we are talking about the free energy or free-energy change of the *system*, G or ΔG is appropriate even for mixtures. The chemical potential rightly applies to specific constituents of the mixture.

It may be helpful to collect the several equations having the form characteristic of, for example, Eq. (3.37). Table 3.2 lists these equations.

System	Equation	Equation Number
Pure ideal gas	$G = G^* + RT \ln P$	(3.13)
Mixture of ideal gases	$\mu_i = \mu_i^* + RT \ln p_i$	(3.35)
Mixture of real gases	$\mu_i = \mu_i^* + RT \ln f_i$	(3.36)
Ideal liquid mixture	$\mu_i = \mu_i^{\rm o} + RT \ln c_i$	(3.40)
Real liquid mixture	$\mu_i = \mu_i^{\rm o} + RT \ln a_i$	(3.37)
Real liquid mixture	$\mu_i = \mu_i^{\rm o} + RT \ln \gamma_i c_i$	(3.39)

Table 3.2. Equations relating chemical potentials to composition variables

PROBLEMS

- **3.1.** Calculate the activity of 0.02 m HC1 at $25 \circ$ C.
- 3.2. Calculate the free-energy change accompanying the process

NaCl $(0.005 m) \rightarrow$ NaCl (0.05 m)

- **3.3.** Estimate the free-energy difference $\Delta \mu = \mu \mu^0$ for a solution 0.25 M in sucrose.
- **3.4.** On the basis of the results of Problems 3.2 and 3.3, comment on the spontaneity or nonspontaneity of making a solution more concentrated.
- **3.5.** A certain substance undergoes a phase transition at the temperature T_c . What is the change of free energy of that compound at the temperature T_c ?

4

EQUILIBRIUM

4.1. CONDITIONS FOR EQUILIBRIUM

We have concluded that a spontaneous process, at constant temperature and pressure, possesses a negative value of dG (or of ΔG) and that the condition for equilibrium is that dG = 0 (or $\Delta G = 0$). We are now going to examine some specific systems to uncover an important consequence of the preceding statements.

First, suppose that the system consists of a single pure substance at constant temperature and pressure, the substance existing (at this temperature and pressure) in two phases at equilibrium. A solid and its melt at the melting point, or a liquid and its vapor at the boiling point, are the most common examples of such a system. We will use the solid (s)–liquid (l) equilibrium in what follows. Since the system is at equilibrium, we know that $\Delta G = 0$, where ΔG is the free energy change per mole associated with the process. Writing the process as

Solid = Liquid

we have, from our general definition of incremental change in a process,

$$\Delta G = G_l - G_s$$

where G_l and G_s are the molar free energies of the substance in the liquid and solid phases, respectively. But since $\Delta G = 0$, we conclude

$$G_s = G_l \tag{4.1}$$

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at equilibrium. Thus, for a pure substance, whenever two (or three) phases are in equilibrium, at fixed temperature and pressure, the molar free energy of the substance has the same value in each phase.

Now we will extend this argument to a closed system at constant temperature and pressure, where the system is at equilibrium and contains *P* phases (*P* = *a*, *b*, *c*,...) and *C* components (*C* = 1, 2, 3,...).¹ Since this is a mixture, we use chemical potentials (partial molar free energies) rather than molar free energies. Imagine infinitesimally small amounts *dn* of components being transferred from one phase to another. Since the system remains at equilibrium during this reversible process, dG = 0, and we write out Eq. (3.24) for the system, obtaining (Glasstone 1947, p. 238)

$$\mu_{1(a)}dn_{1(a)} + \mu_{1(b)}dn_{1(b)} + \dots + \mu_{1(P)}dn_{1(P)} + \mu_{2(a)}dn_{2(a)} + \mu_{2(b)}dn_{2(b)} + \dots + \mu_{2(P)}dn_{2(P)}$$

$$\vdots \qquad (4.2)$$

$$+\mu_{C(a)}dn_{C(a)}+\mu_{C(b)}dn_{C(b)}+\cdots+\mu_{C(P)}dn_{C(P)}=0$$

which is succinctly written

$$\sum \mu_{C(P)} dn_{C(P)} = 0 \tag{4.3}$$

Since the system is closed, the total amount of each component is constant, giving

$$dn_{1(a)} + dn_{1(b)} + \dots + dn_{1(P)} = 0$$

$$dn_{2(a)} + dn_{2(b)} + \dots + dn_{2(P)} = 0$$

$$\vdots$$

$$dn_{C(a)} + dn_{C(b)} + \dots + dn_{C(P)} = 0$$
(4.4)

or

$$\sum dn_{C(P)} = 0 \tag{4.5}$$

The only way in which Eqs. (4.2) and (4.4) [or Eqs. (4.3) and (4.5)] can simultaneously be satisfied is if

$$\mu_{1(a)} = \mu_{1(b)} = \cdots \mu_{1(P)}$$

$$\mu_{2(a)} = \mu_{2(b)} = \cdots \mu_{2(P)}$$

$$\vdots$$

$$\mu_{C(a)} = \mu_{C(b)} = \cdots \mu_{C(P)}$$
(4.6)

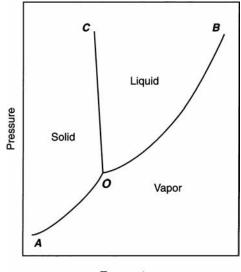
¹ A *component* is defined in this way: *C*, the number of components, is the minimum number of substances needed to make up the equilibrium mixture. For example, pure water contains H_2O , H_3O^+ , OH^- , and a mixture of hydrogen-bonded water multimers, but these are all connected by (established by) equilibria, so C = 1; you need take only one substance, water, to create this system.

Thus, under these conditions (consisting of *P* phases, *C* components, where the closed system is at equilibrium at fixed temperature and pressure), the *chemical potential of each individual component has the same value in all phases*. This is an obvious generalization of our earlier result for a pure substance. Since, from Eq. (4.6), we have $\mu_{i(a)} = \mu_{i(b)} = \dots = \mu_{i(P)}$ for component *i*, it follows that, at equilibrium, $d\mu_i = 0$ for the transfer of an infinitesimal amount of component *i* from one phase to another.²

4.2. PHYSICAL PROCESSES

Phase Transitions (Single Component). We now return to a system consisting of a single component in a closed system at fixed temperature and pressure. This substance is capable of existing in three states of matter: the solid, the liquid, and the gas (vapor). The manner in which these states are controlled by the values of temperature and pressure is readily displayed on a pressure–temperature *phase diagram.* Figure 4.1 shows a schematic phase diagram. For each chemical substance the phase diagram must be experimentally determined.

Any selected pair of coordinates P, T determine the state of the system. Of special interest are those coordinates describing the lines in the phase diagram, because along these lines two phases coexist in equilibrium. Thus line OC describes the melting



Temperature

Figure 4.1. Pressure-temperature phase diagram of a pure substance.

² The equilibrium condition $d\mu_i = 0$ also applies to the case P = 1, as in a liquid solution, and it determines the direction in which solute diffusion takes place. If a nonequilibrium distribution of solute exists, the solute particles will diffuse in the direction so as to achieve the condition $d\mu_i = 0$ throughout the solution. This means that the direction of diffusion is from regions of higher chemical potential to regions of lower chemical potential. transition; at any point on this line the solid and liquid phases are in equilibrium, and the value of T corresponding to any given value of P is the *melting point* at that pressure. The line is very steep because the melting point is not very sensitive to pressure changes (i.e., the melting point does not change much when the pressure is changed).

Along the line *OB* the liquid and its vapor are in equilibrium, so the value of *T* on this line is the *boiling point* corresponding to the selected *P* value. Observe that the boiling point is quite sensitive to pressure.³ Along the line *OA* the solid and the vapor are in equilibrium. The direct conversion of solid to vapor is called *sublimation*.⁴ At point O all three phases coexist in equilibrium. This is obviously a unique set of circumstances; it is called the *triple point*. The triple point of water is 273.16 K (i.e., 0.01 °C) and 4.58 mmHg pressure.

Now we treat phase transitions thermodynamically. Consider a pure substance at temperature and pressure such that two phases, 1 and 2 (which may be gas, liquid, or solid), are in equilibrium. Thus $G_1 = G_2$, and so $dG_i = dG_2$, which means that if the temperature or pressure is changed infinitesimally, the changes in free energy of the two phases will be identical and the phases will remain in equilibrium. From Eq. (3.5) we write dG = V dP - S dT, or

$$V_1 dP - S_1 dT = V_2 dP - S_2 dT$$

Rearranging, we obtain

$$\Delta S dT = \Delta V dP$$

where $\Delta S = S_2 - S_1$ and $\Delta V = V_2 - V_1$. Therefore

$$\frac{dP}{dT} = \frac{\Delta S}{\Delta V} \tag{4.7}$$

But the system is at equilibrium, so $\Delta G = \Delta H - T \Delta S = 0$, giving $\Delta S = \Delta H/T$, or, from Eq. (4.7)

$$\frac{dP}{dT} = \frac{\Delta H}{T \,\Delta V} \tag{4.8}$$

This is the *Clapeyron equation*. It describes the slope of the line in the phase diagram for a pure substance.

The Clapeyron equation is especially useful when applied to the liquid-vapor transition (boiling), and in this application we usually can employ an approximate

³ Conventionally, the boiling point is considered to be the temperature at which the liquid and vapor are in equilibrium at atmospheric pressure. This definition can be assumed when the pressure is not stated.

⁴ Sublimation is less familiar than melting or boiling, but it can be important. Salicylic acid readily sublimes. An old bottle of aspirin tablets may contain partially hydrolyzed aspirin (acetylsalicylic acid). The products are acetic acid (a liquid whose vapor smells like vinegar) and salicylic acid, which may sublime and then condense in white crystals on the wall of the bottle.

version. Writing Eq. (4.8) specifically for this transition, we obtain

$$\frac{dP}{dT} = \frac{\Delta H_{\rm vap}}{T\Delta V_{\rm vap}} \tag{4.9}$$

where *T* is the boiling temperature and $\Delta V_{\text{vap}} = V_{\text{vapor}} - V_{\text{liquid}}$. We neglect the molar volume of the liquid as being very small relative to the molar volume of the vapor. We will also assume that the vapor phase behaves ideally, so $V_{\text{vap}} = RT/P$. Combining these approximations with Eq. (4.9) yields

$$\frac{1}{P}\frac{dP}{dT} = \frac{\Delta H_{\text{vap}}}{RT^2} \tag{4.10}$$

or

$$\frac{d\ln P}{dT} = \frac{\Delta H_{\rm vap}}{RT^2} \tag{4.11}$$

These two equations are versions of the *Clausius–Clapeyron equation*, which relates the boiling point *T* to the vapor pressure *P*.

If ΔH_{vap} should happen to be essentially constant (independent of temperature), we can integrate the Clausius–Clapeyron equation. First integrating generally gives

$$\ln P = -\frac{\Delta H_{\rm vap}}{RT} + \text{constant}$$
(4.12)

or

$$\log P = -\frac{\Delta H_{\rm vap}}{2.3RT} + C \tag{4.13}$$

where *C* is a constant. Equation (4.12) or (4.13) provides a means for evaluating the molar heat of vaporization ΔH_{vap} from vapor pressure-temperature data.

Example 4.1. Table 4.1 gives vapor pressure–temperature data for *n*-octane. Calculate ΔH_{vap} .

$t(^{\circ}C)$	$T(\mathbf{K})$	1/T	P(mmHg)	log[P(mmHg)]
-14.0	259.15	0.00386	1	0.000
+19.2	292.35	0.00342	10	1.000
45.1	318.25	0.00314	40	1.602
65.7	338.85	0.00295	100	2.000
104.0	377.15	0.00265	400	2.602
125.6	398.75	0.00251	760	2.881

Table 4.1. Vapor pressure-temperature data for n-octane

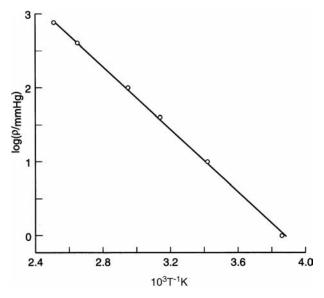


Figure 4.2. Plot of Eq. (4.13) for *n*-octane. Data from Table 4.1.

Figure 4.2 is a plot of the data according to Eq. (4.13). From the plot we evaluate the slope as -2091 K. Equation (4.13) shows this identity:

$$\text{Slope} = -\frac{\Delta H_{\text{vap}}}{2.3R}$$

Thus we calculate

$$\Delta H_{\text{vap}} = -2.3(1.987 \text{ cal mol}^{-1} \text{ K}^{-1})(-2091 \text{ K})$$

= 9658 cal mol⁻¹
= 9.57 kcal mol⁻¹
= 40.0 kJ mol⁻¹

The heat of vaporization is positive because heat is absorbed by the system during the vaporization process.

We can also integrate the Clausius–Clapeyron equation between the limits T_1 and T_2 , presuming that ΔH_{vap} is essentially constant in this temperature range. From Eq. (4.11), we obtain

$$\int_{P_{1}}^{P_{2}} d\ln P = \frac{\Delta H_{\text{vap}}}{R} \int_{T_{1}}^{T_{2}} \frac{dT}{T^{2}}$$

which gives

$$\ln \frac{P_2}{P_1} = -\frac{\Delta H_{\text{vap}}}{R} \left(\frac{1}{T_2} - \frac{1}{T_1} \right) = \frac{\Delta H_{\text{vap}}}{R} \left(\frac{T_2 - T_1}{T_1 T_2} \right)$$
(4.14)

or

$$\log \frac{P_2}{P_1} = \frac{\Delta H_{\text{vap}}}{2.3R} \left(\frac{T_2 - T_1}{T_1 T_2} \right)$$
(4.15)

Example 4.2. The vapor pressure of water is 17.535 mmHg at $20.0 \degree \text{C}$ and 31.824 mmHg at $30.0 \degree \text{C}$. Calculate the heat of vaporization of water in this temperature interval.

Using Eq. (4.15), we have

$$\log \frac{31.824}{17.535} = \frac{\Delta H_{\text{vap}}}{(2.3)(1.987)} \left(\frac{10}{293.15 \times 303.15}\right)$$
$$\Delta H_{\text{vap}} = 10,530 \text{ cal mol}^{-1}$$
$$= 10.53 \text{ kcal mol}^{-1}$$

In Examples 4.1 and 4.2 we see how heats of transition are determined. With such ΔH values at hand, we can calculate entropies of transition as shown in Example 2.1.

The Phase Rule (Multiple Components). We have already developed the condition for equilibrium in a system at equilibrium containing multiple phases and components; the condition is that the chemical potential of each component be the same in all phases [Eq. (4.6)]. This will lead us to a general rule connecting the number of phases, the number of components, and the number of variables (this last factor is called the *degrees of freedom F*) that must be specified in order to define the system completely (in a thermodynamic sense). Let *P* be the number of phases and *C* the number of components (see note 1). Here are the steps in the reasoning:

- 1. The composition of a phase containing C components can be specified by giving C-1 concentrations. This is because the final concentration can be obtained by difference.
- 2. If there are *P* phases, in order to completely define the compositions of all phases, P(C-1) concentrations must be specified. This is the total number of concentration variables in the system.
- 3. The temperature and pressure variables must be added to the P(C-1) concentration variables. This gives us

Total number of variables = P(C-1) + 2

- 4. Since for component 1 we know that $\mu_{1(\text{phase }a)} = \mu_{1(\text{phase }b)} = \dots$, and similarly for all *C* components, this condition gives C(P-1) independent equations, which fix C(P-1) variables.⁵
- 5. The number of variables left undetermined (i.e., the number of degrees of freedom) is equal to the total number of variables minus the number of variables that are determined by the equilibrium condition. Thus

Number of degrees of freedom = P(C-1) + 2 - C(P-1)

or

$$F = C - P + 2 \tag{4.16}$$

This last equation is the *phase rule* of Gibbs.

We will apply the phase rule to the phase diagram of Fig. 4.1 in order to learn how it works. First consider the liquid–gas equilibrium. There are two phases and one component, so F = 1 - 2 + 2 = 1. This means that specifying one degree of freedom suffices to completely define the system. If we choose to specify the temperature (as our one degree of freedom), the condition of equilibrium uniquely guarantees that the pressure will be fixed at the value given by the line *OB* corresponding to the specified temperature. Alternatively, we might have specified the pressure; then the temperature would be defined by the system. In this system as described, we cannot independently choose both the temperature and the pressure.

Next consider the triple point *O*. Here P = 3 and C = 1, so F = 0. There is no freedom to alter the system variables while maintaining the system at the triple point.

A very practical kind of system is that of a pure solid substance placed in contact with a second pure substance, a liquid, which we call the *solvent*. To be specific, let us add solid benzoic acid to water. Presuming that sufficient benzoic acid has been added so that at equilibrium some solid is present, how many degrees of freedom does the system possess?

The process that occurs is dissolution of benzoic acid in water. Evidently P = 2, for two phases, solid benzoic acid and liquid solution, are present. Moreover, C = 2, because the system can be prepared from benzoic acid and water. The phase rule gives us F = 2 - 2 + 2 = 2 degrees of freedom. These are the temperature and the

$$\mu_{1(a)} = \mu_{1(b)}, \qquad \mu_{2(a)} = \mu_{2(b)}$$

 $\mu_{1(b)} = \mu_{1(c)}, \qquad \mu_{2(b)} = \mu_{2(c)}$

The equations $\mu_{1(a)} = \mu_{1(c)}$ and $\mu_{2(a)} = \mu_{2(c)}$ are not independent, but follow from the preceding four equations.

⁵ The multiplier is P - 1 rather than P because if we know C(P - 1) relationships, we have exhausted the independent equations. For instance, suppose C = 2 (components numbered 1,2) and P = 3 (phases labeled a, b, c). Then the $C(P - 1) = 2 \times 2 = 4$ independent equations are

pressure, both of which must be fixed in order to completely define the position of equilibrium of the system. (In practice this system is not very sensitive to pressure, which is commonly the ambient atmospheric pressure, but it is very sensitive to the temperature.) An equivalent description of this system is that at fixed temperature and pressure, the concentration of dissolved benzoic acid is invariant; no further degrees of freedom remain. We call this invariant dissolved concentration the *equilibrium solubility* of benzoic acid at the experimental temperature.

4.3. CHEMICAL EQUILIBRIUM

The Equilibrium Constant. We now turn to a treatment of chemical reactions, namely, processes in which chemical bonds (covalent bonds) or noncovalent interactions are formed or broken, or both. Because liquid systems are of special interest to us, suppose that the process occurs in a homogeneous (single phase) liquid system. Let the generalized balanced chemical reaction be written

$$a\mathbf{A} + b\mathbf{B} \Longrightarrow m\mathbf{M} + n\mathbf{N}$$
 (4.17)

where A and B represent the reactant chemical species; M and N are the product chemical species; and *a*, *b*, *m*, *n* are stoichiometric coefficients in the balanced reaction.⁶ For the moment we do not require that the system be at equilibrium, but the temperature and pressure are fixed.

As is our usual practice, the incremental change in free energy is defined (on a per mole basis) as the difference between the final state and the initial state:

$$\Delta G = \sum G_{\text{products}} - \sum G_{\text{reactants}}$$
(4.18)

Recognizing that our reaction system is a mixture, we know that we should express the free energies of reactants and products in terms of chemical potentials; thus Eq. (4.18) becomes

$$\Delta G = m\mu_{\rm M} + n\mu_{\rm N} - a\mu_{\rm A} - b\mu_{\rm B} \tag{4.19}$$

Notice that each term on the right is the product of an intensity factor (e.g., μ_M , chemical potential per mole) and a capacity factor (*m*, number of moles).

Next we call on our fundamental relationship for the chemical potential in terms of activity [Eq. (3.37)]:

$$\boldsymbol{\mu} = \boldsymbol{\mu}_i^0 + RT \ln a_i \tag{4.20}$$

⁶ Our modern interpretation of a balanced chemical reaction views the species symbols as representing atoms, molecules, or ions, but the balanced reaction does not necessarily imply an atomic viewpoint. The reaction simply describes an experimental observation and is a classical thermodynamic concept.

We simply substitute from Eq. (4.20) into Eq. (4.19):

$$\Delta G = m\mu_{\rm M}^0 + mRT \ln a_{\rm M} + n\mu_{\rm N}^0 + nRT \ln a_{\rm N}$$
$$- a\mu_{\rm A}^0 - aRT \ln a_{\rm A} - b\mu_{\rm B}^0 - bRT \ln a_{\rm B}$$

Collecting terms and making use of one of the properties of logarithms gives

$$\Delta G = (m\mu_{\rm M}^0 + n\mu_{\rm N}^0 - a\mu_{\rm A}^0 - b\mu_{\rm B}^0)$$
$$+ RT \ln a_{\rm M}^m + RT \ln a_{\rm N}^n$$
$$- RT \ln a_{\rm A}^a - RT \ln a_{\rm B}^b$$

which can be written

$$\Delta G = \Delta G^0 + RT \ln \frac{a_{\rm M}^m a_{\rm N}^n}{a_{\rm A}^a a_{\rm B}^b} \tag{4.21}$$

where $\Delta G^0 = m\mu_M^0 + n\mu_N^0 - a\mu_A^0 - b\mu_B^{0.7}$ Equation (4.21) is called the *reaction isotherm* (the term *isotherm* merely signifies that the equation or phenomenon takes place at, or applies to, a constant temperature).

Recall that we have not yet required that the system be at equilibrium. Equation (4.21) gives the free energy change as a function of the activities of reactants and products of the reaction. But it is the equilibrium condition that specifically interests us. Let the activities now be the activities at equilibrium, and define

$$K = \frac{a_{\rm M}^m a_{\rm N}^n}{a_{\rm A}^a a_{\rm B}^b} \tag{4.22}$$

Moreover, we recall that the condition for equilibrium is that $\Delta G = 0$. Putting this condition and the definition of Eq. (4.22) into Eq. (4.21) gives the simple but very important result:

$$\Delta G^0 = -RT \ln K \tag{4.23}$$

The quantity K is called the *equilibrium constant* for the reaction; its general form can be inferred from Eq. (4.22). It is conventional to write the products in the numerator and the reactants in the denominator. Notice, therefore, that the reciprocal of K as thus defined is the equilibrium constant for the reaction when written in the reverse direction.

Equilibrium constants can be measured experimentally; in effect, one needs to determine each activity in the definition and then to calculate K according to

⁷ It would be perfectly correct to write $\Delta \mu^0$ instead of ΔG^0 . The latter symbolism is used in order to be consistent with conventional practice.

Eq. (4.22). Then, with Eq. (4.23), the quantity ΔG^0 (which is pronounced "delta *G* naught") is calculated. ΔG^0 is called the *standard free-energy change* for the reaction, and it is interpreted as the change in free energy per mole when the reactants in their standard states are transformed into the products in their standard states. This concept is difficult to visualize in physical terms, and it may be better to note the obvious, namely, that [see Eq. (4.23)] ΔG^0 and *K* contain the same information about the system. The logarithmic relationship between ΔG^0 and *K*, as well as the form of the equilibrium constant definition, is a direct consequence of the form of Eq. (4.20) for the chemical potential.

In the chemical literature many equilibrium constants are described by adjectives that provide information on the chemical process and on the definition of the constant. For example, a weak acid HA dissociates according to

$$HA \stackrel{K_a}{=} H^+ + A^-$$

Placing the equilibrium constant symbol over the arrows tells the reader how the constant is to be defined; in this case

$$K_a = \frac{a_{\rm H^+} a_{\rm A^-}}{a_{\rm HA}} \tag{4.24}$$

This particular equilibrium constant is called an *acid dissociation constant* (or *acid ionization constant*). For a reaction, especially a reaction involving noncovalent interactions, having the form

$$A + B \Longrightarrow M$$

the equilibrium constant

$$K = \frac{a_{\rm M}}{a_{\rm A}a_{\rm B}}$$

may be called an *association constant, formation constant, stability constant*, or *binding constant*. Turn the reaction around, and its equilibrium constant (which will be the reciprocal of *K*) becomes a dissociation constant or instability constant.

Example 4.3. For the acid dissociation of acetic acid in water at 25 °C, the experimental value of K_a is 1.75×10^{-5} . Calculate ΔG^0 for this process.

We can use Eq. (4.23) directly or in the form

$$\Delta G^{0} = -2.303 RT \log K_{a}$$

$$= -(2.303)(1.987 \text{ cal mol}^{-1} \text{ K}^{-1})(298.15 \text{ K})(-4.757)$$

$$= +6490 \text{ cal mol}^{-1}$$

$$= +6.49 \text{ kcal mol}^{-1}$$
(4.25)

From either Eq. (4.23) or (4.25) we obtain these correspondences:

If K < 1, then $\Delta G^0 > 0$. If K = 1, then $\Delta G^0 = 0$. If K > 1, then $\Delta G^0 < 0$.

In Example 4.3, ΔG^0 has a positive sign because *K* is smaller than unity. This tells us that, at equilibrium, the reactant state is "favored" in this process.

The units of the equilibrium constant require comment. From its definition in terms of activities, it is clear that we need to know the units of activities. Our earlier conventions concerning standard states and reference states provide guidance. Evidently the activities of solvents and solids, which are taken equal to their mole fractions in practical work, are dimensionless, because the mole fraction is dimensionless. The activities of uncharged molecules are taken equal to the concentrations (usually in mol L^{-1}) of these molecules, so the activities are reasonably given the same units. To maintain consistency, it is advisable to assign the units of concentration to the activities of other solution species.

According to these recommendations, the unit of the acid dissociation constant K_a is [from Eq. (4.24)] M (i.e., mol L⁻¹). However, units are conventionally not stated for K_a values.⁸

The standard free-energy change ΔG^0 can be related to a standard enthalpy change ΔH^0 and a standard entropy change ΔS^0 by the usual form:

$$\Delta G^0 = \Delta H^0 - T \Delta S^0 \tag{4.26}$$

 ΔH^0 and ΔS^0 are interpreted analogously to ΔG^0 , that is, in terms of the process in which reactants are transformed into products, and all species are in their standard states.

Temperature Dependence of the Equilibrium Constant. The Gibbs–Helmholtz equation [Eq. (3.18), repeated here as Eq. (4.27)] has a useful form for our present purpose:

$$\frac{d(\Delta G/T)}{dT} = -\frac{\Delta H}{T^2} \tag{4.27}$$

From Eq. (4.23), $\Delta G^0 = -RT \ln K$, rearrangement gives

$$\frac{\Delta G^0}{T} = -R \ln K \tag{4.28}$$

⁸ An alternative viewpoint is that activities are dimensionless, thus requiring the activity coefficient to have units. Each point of view is acceptable, provided that it is consistently applied throughout a calculation.

Combination of Eqs. (4.27) and (4.28) yields

$$\frac{d\ln K}{dT} = \frac{\Delta H^0}{RT^2} \tag{4.29}$$

Equation (4.29), the *van't Hoff equation*, describes how the equilibrium constant varies with the temperature. The quantity ΔH^0 is the standard enthalpy change, sometimes called the *heat of reaction*. If ΔH^0 is essentially independent of temperature, general integration of Eq. (4.29) gives

$$\ln K = -\frac{\Delta H^0}{RT} + \text{constant}$$
(4.30)

or

$$\log K = -\frac{\Delta H^0}{2.3RT} + C \tag{4.31}$$

Alternatively, integration between the temperature limits T_1 and T_2 gives

$$\ln \frac{K_2}{K_1} = \frac{\Delta H^0}{R} \left(\frac{T_2 - T_1}{T_1 T_2} \right)$$
(4.32)

$$\log \frac{K_2}{K_1} = \frac{\Delta H^0}{2.3R} \left(\frac{T_2 - T_1}{T_1 T_2} \right)$$
(4.33)

These integrated equations may seem familiar; they have the same form as the vapor pressure equations (4.12)–(4.15), and they are used similarly.

Free-Energy, Enthalpy, and Entropy Changes in Chemical Reactions. We now have at hand all the thermodynamic theory needed to calculate (from the appropriate experimental data) these standard thermodynamic quantities for a chemical reaction: ΔG^0 , ΔH^0 , and ΔS^0 . These are the steps:

1. From measurement of the equilibrium constant *K* at a given temperature, calculate ΔG^0 from

$$\Delta G^0 = -RT \ln K \tag{4.34}$$

- 2. From measurements of *K* at several temperatures, calculate ΔH^0 by means of one of Eqs. (4.30)–(4.33).
- 3. From $\Delta G^0 = \Delta H^0 T \Delta S^0$, calculate ΔS^0 .

Since K in general varies with temperature, evidently ΔG^0 does as well. When integrating the van't Hoff equation, we assumed that ΔH^0 is a constant,

T(°C)	$K(M^{-1})$	<i>T</i> (K)	1/T	log K
15.5	11.6	288.65	0.00346	1.064
25.0	8.7	298.15	0.00335	0.940
40.0	5.9	313.15	0.00319	0.771

 Table 4.2.
 Dependence of equilibrium constant on temperature for the binding of methyl *trans*-cinnamate and 8-chlorotheophylline anion in water

independent of temperature, and although this may constitute an acceptable approximation for many reactions, it is not generally true, and it may lead to poor estimates of ΔH^0 and ΔS^0 . The data themselves, if carefully interpreted, will reveal whether ΔH^0 is reasonably constant over the temperature range that was investigated experimentally.

Example 4.4. These are data (Table 4.2) for the equilibrium constant (a stability constant) describing the noncovalent association between methyl *trans*-cinnamate and 8-chlorotheophyllinate in aqueous solution. Find ΔG^0 , ΔH^0 , and ΔS^0 at 25 °C.

1. From Eq. (4.34), or its equivalent, we have

$$\Delta G^{0} = -(2.303)(1.987 \text{ cal mol}^{-1}\text{K}^{-1})\log 8.7$$

= -1282 cal mol⁻¹
= -1.28 kcal mol⁻¹

2. ΔH^0 will be obtained from a plot according to Eq. (4.31); this is called a *van't Hoff plot:*

$$\log K = -\frac{\Delta H^0}{2.3RT} + C$$
$$\text{Slope} = -\frac{\Delta H^0}{2.3R}$$

Figure 4.3 shows the plot, which is acceptably straight over the temperature range given in the table; this linearity is consistent with the constancy of ΔH^0 over this temperature range. The slope of the line is 1083 K, giving for ΔH^0

$$\Delta H^0 = -(2.3)(1.987 \text{ cal mol}^{-1}\text{K}^{-1})(1083 \text{ K})$$

= -4996 cal mol^{-1}
= -5.00 kcal mol^{-1}

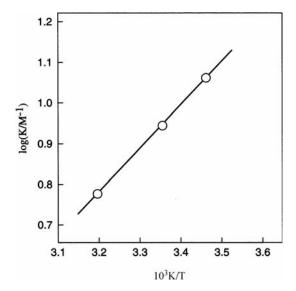


Figure 4.3. van't Hoff plot of the data in Table 4.2.

3.

$$\Delta S^{0} = \frac{-(\Delta G^{0} - \Delta H^{0})}{T}$$

= $\frac{-1.28 - (5.00) \text{ kcal mol}^{-1}}{-298.15 \text{ K}}$
= $-0.0125 \text{ kcal mol}^{-1} \text{K}^{-1}$
= $-12.5 \text{ cal mol}^{-1} \text{K}^{-1}$

We can interpret these results chemically. The reaction has the form

$$A + B \rightleftharpoons C$$

where A is methyl cinnamate and B is 8-chlorotheophyllinate; C is the complex formed from these. Since K > 1, we have $\Delta G^0 < 0$; the product C is favored over the reactants. But this result is seen to be a consequence of a competition between ΔH^0 and ΔS^0 . ΔH^0 is negative, so the enthalpy change makes a favorable contribution to ΔG^0 (it makes ΔG^0 more negative). The negative value of ΔH^0 suggests that fairly strong noncovalent binding is occurring between A and B, because ΔH^0 is an energy value. On the other hand, the negative value of ΔS^0 opposes product formation by making a positive contribution to ΔG^0 (because of the negative sign in $\Delta G^0 = \Delta H^0 - T \Delta S^0$). The negative entropy change may arise because two particles (A and B) are being transformed into a single C particle, with a resultant decrease in number of configurational and thermal microstates of the system. If we were to write this reaction as

$$C \stackrel{K'}{=} A + B$$

we would find that K' = 1/K and that ΔG^0 , ΔH^0 , and ΔS^0 each possesses the same numerical value as found in this example, but with the opposite sign.

A subtlety of such calculations, often overlooked, is that the numerical values of ΔG^0 and ΔS^0 (but not of ΔH^0) depend on the concentration scale in which the equilibrium constant is expressed. In thermodynamic terms, this is stated as follows: The numerical values of ΔG^0 and ΔS^0 depend on the choice of standard state.

PROBLEMS

- **4.1.** The vapor pressure of heptane, C_7H_{16} , is 100 mmHg at 41.8 °C and 760 mmHg (i.e., 1 atm) at 98.4 °C. Calculate its molar heat of vaporization over this temperature range.
- **4.2.** The equilibrium constant for this reaction in aqueous solution at $25 \,^{\circ}\text{C}$ is $21.5 \,\text{M}^{-1}$:

Theophylline + Salicylate anion $\stackrel{K}{\longrightarrow}$ Complex

Calculate the standard free energy change for this reaction.

4.3. These are literature data for the vapor pressure of ethyl acetate, $CH_3COOC_2H_5$, as a function of temperature. Calculate the heat of vaporization.

T(°C)	P(mmHg)
-43.4	1
-13.5	10
9.1	40
27.0	100
59.3	400
77.1	760

4.4. A quantity pK_a is defined by the relationship

$$pK_a = -\log K_a$$

where K_a is the acid dissociation constant of a weak acid in water. Obtain an equation by which the standard free-energy change can be calculated directly from the p K_a .

4.5. The p K_a value of phenol is 10.0 at 25 °C. Write the chemical reaction, define K_a , and calculate ΔG^0 for the process.

- **4.6.** The p K_a value of chloroacetic acid, ClCH₂COOH, is 2.87 at 25 °C, and its heat of ionization (ΔH^0) has been measured to be -1.12 kcal mol⁻¹. Calculate its standard entropy of ionization.
- **4.7.** The ionization (acid dissociation) of chloroacetic acid takes place in aqueous solution according to

$$CICH_2COOH \stackrel{K_a}{\longrightarrow} H^+ + CICH_2COO^-$$

In view of this, attempt to rationalize the sign of ΔS^0 obtained in Problem 4.6.

4.8. The equilibrium constant for the addition of hydrogen cyanide to acetaldehyde is 7100 M^{-1} at 25 °C. Calculate ΔG^0 .

$$H_3C-CHO + HCN \longrightarrow H_3C-CHO + HCN$$

- **4.9.** A pharmacy has received 0.5 kg of a certain drug with a molecular weight of 250 g mol^{-1} . The drug undergoes a degradation reaction with a heat of reaction of 750 cal mol⁻¹ and an equilibrium constant equal to 1.2×10^{-3} . The drug is kept in a thermostated stockroom at 25 °C. Calculate the entropy of the degradation reaction.
- **4.10.** The pH of a 0.01 M solution of chloroacetic acid is 2.44 at 25 °C. The entropy change ΔS^0 of the acid dissociation is -16.9 cal mol⁻¹ K⁻¹. Calculate the heat of dissociation ΔH^0 of chloroacetic acid. Based on the value of the calculated ΔH^0 , predict if the equilibrium constant of chloroacetic acid at 50 °C is smaller or larger than the equilibrium constant at 25 °C.
- **4.11.** The vapor pressure of *n*-octane is 100 mmHg at 65.7 °C and 400 mmHg at 104.0 °C. Knowing that the boiling point of *n*-octane is 126.5 °C and assuming that the heat of vaporization is constant in the range of temperatures $40 \,^{\circ}\text{C}$ -126.5 °C, calculate the entropy of vaporization of *n*-octane.
- **4.12.** Calculate the number of degrees of freedom *F* of the following systems:
 - (a) Carbonated water.
 - (b) A suspension of erythromycin stearate (a powerful antibiotic) in water.
- **4.13.** The dissociation of aqueous ammonia NH₄OH yields the ions NH₄⁺ and OH⁻. Knowing that the dissociation constant of NH₄OH is 1.652×10^{-5} at $15 \,^{\circ}$ C and 1.892×10^{-5} at $50 \,^{\circ}$ C, calculate the standard ΔG^0 (ΔG at $25 \,^{\circ}$ C) knowing that ΔS^0 is -19.3 cal K⁻¹.

THERMODYNAMICS OF PHYSICAL PROCESSES

5

INTRODUCTION TO PHYSICAL PROCESSES

5.1. SCOPE

The separation of properties or processes into physical and chemical categories is arbitrary but useful, and in most instances it is not notably ambiguous. Here are the criteria adopted in making the present separation:

- 1. All covalent bond changes (which necessarily result in alterations in primary molecular structure) are *chemical*. This category includes most of the reactions of interest in organic chemistry, inorganic chemistry, and biochemistry. Part III deals with such processes.
- 2. All changes in physical state or phase that do not involve covalent bond changes are *physical*. Such processes include melting, vaporization, sublimation, dissolution, partitioning, and adsorption. These are of concern in Part II.
- 3. There is an exception: electrolyte dissociation and behavior are treated generally, in Part II, as a physical phenomenon, but the special case of acid–base equilibrium is discussed as a chemical phenomenon in Part III.
- 4. An ambiguous area remains, consisting of changes in noncovalent interactions roughly in the ΔG^0 range of 0–10 kcal mol⁻¹. These processes include molecular complex formation (binding phenomena of many types) and conformational changes. We classify these as *chemical*, and treat them in Part III.

Some slight repetition of material from Part I will be encountered in Part II, where it is inserted for convenience.

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5.2. CONCENTRATION SCALES

Solution composition can be expressed on a physical (empirical) basis in terms of the quantities measured in the laboratory. These are the common concentration scales of this type:

- *Percent by Weight.* The number of grams of solute contained in 100 g of solution. The concentration of the strong mineral acids, as available commercially, are expressed as percent by weight.
- *Percent Weight/Volume (% w/v).* The number of grams of solute contained in 100 mL of solution. This scale is often used to describe the composition of solutions of solids in liquids.
- *Percent by Volume*. The number of milliliters of solute contained in 100 mL of solution. Solutions of liquids in liquids are commonly specified in this way. It is important to note a possible ambiguity in this designation. Consider these two operations: (1) 80.0 mL of ethanol is dissolved in water to make a final total volume of 100 mL; (2) 80.0 mL of ethanol is mixed with 20.0 mL of water. These solutions have different compositions, because preparation 2 does not yield a final volume of 100 mL. In thermodynamic terms, the partial molar volume of ethanol is not equal to its molar volume. In molecular terms, the spatial and energetic character of the ethanol–water interaction is different from those of ethanol–ethanol or water–water interactions. Solution 1 has a composition of 80.0% by volume of ethanol. The composition of solution 2 is most easily specified in terms of its *volume fraction* φ , where the volume fraction φ_i of component *i* is defined

$$\varphi_i = \frac{V_i}{\sum\limits_{i=1}^n V_i}$$

Thus solution 2 has volume fraction $\varphi_1 = 0.20$ of water and $\varphi_2 = 0.80$ of ethanol. In order to communicate without possible confusion, statements of solution composition should specify clearly, as, for example, by describing the manner of preparing the solution, which concentration scale is meant.

Observe that these physical concentration scales constitute the three combinations of mass/mass, mass/volume, and volume/volume. Other units may, however, often be encountered. For example, milligram percent is the number of milligrams of solute contained in 100 mL of solution. Another common unit is milligrams per milliliter (mg/mL), which is numerically equal to grams per liter (g/L). Very dilute solutions may be expressed in parts per million (ppm), which is the number of grams of solute contained in 10^6 g of solution. (If the solvent is water, this is effectively the number of grams of solute in 10^6 mL of solution.) Similarly, ppb means parts per billion. The chemical concentration scales are based on the concept of the amount of substance as expressed in number of moles:

- *Molarity* (c)—the number of moles of solute contained in 1000 mL of solution. Molarity has the units mol L^{-1} , which is often designated M; it is expressed as mol dm⁻³ in SI units. Some investigators use the unit millimolar (mM), which denotes the number of millimoles contained in 1000 mL of solution; for example, 0.030 M and 30 mM have the same meaning. Molarity is a very practical concentration scale, but it has the disadvantage that the molarity of a solution depends on the temperature, because the volume is temperature-dependent.
- *Molality* (m)—the number of moles of solute per 1000 g of solvent. The molality is temperature-independent, and for this reason it is often preferred in precise physical chemical experimental work.
- *Mole fraction*(x)—the number of moles of solute divided by the total number of moles in the solution. The mole fraction is temperature-independent. A convenient attribute of the mole fraction (as of all fractional quantities) is that the sum of the mole fractions of all constituents in a solution is unity.

In general the molarity, molality, and mole fraction scales are not directly proportional to each other, but in very dilute solutions of a solute *i* the relationships are

$$x_i = \frac{c_i M_1}{1000\rho_1} \qquad (x_i \ll 1) \tag{5.1}$$

$$x_i = \frac{m_i M_1}{1000} \qquad (x_i \ll 1) \tag{5.2}$$

where subscript 1 refers to the solvent and *i* to the solute. In these dilute solutions, Eqs. (5.1) and (5.2) show that the various concentration scales are proportional to each other; M_1 is the molecular weight of the solvent, and ρ_1 is its density.

5.3. STANDARD STATES

In Chapter 3 we encountered the concept of the standard state, and here we will extend the treatment by explicitly invoking the concentration scales used in laboratory work. We begin with Eq. (3.37), repeated here:

$$\mu_i = \mu_i^0 + RT \ln a_i \tag{5.3}$$

In this equation, μ_i is the chemical potential (partial molar free energy) of constituent *i* in a liquid mixture, μ_i^0 is its standard chemical potential, and a_1 is its activity. The activity, which can be thought of as an "effective concentration," is related to the actual

concentration by

Activity = Activity coefficient
$$\times$$
 Concentration

where the activity coefficient is a number that accounts for deviations from ideal behavior. Let us recall these definitions:

- The *standard state* (with respect to constituent *i*) is that state of the system in which the activity of *i* is unity; then, from Eq. (5.3), $\mu_i = \mu_i^0$.
- The *reference state* (with respect to constituent i) is that state of the system in which the activity coefficient of i is unity; then its activity equals its concentration.

For the present let us suppose that the system is in its reference state (for constituent i), so the activity coefficient is unity, and the activity is equal to the concentration. But we have seen that concentrations can be expressed in molarity, molality, or mole fraction. For constituent i in a given system, Eq. (5.3) may be written in terms of each of these concentration units:

$$\mu_i = \mu_i^0 + RT \ln c_i \tag{5.4}$$

$$\mu_i = \mu_m^0 + RT \ln m_i \tag{5.5}$$

$$\mu_i = \mu_x^0 + RT \ln x_i \tag{5.6}$$

The situation represented by Eqs. (5.4)–(5.6) is analogous to the specification of mechanical potential energy, say, of a rock on top of a hill. We can only speak numerically of energy *differences*, and we calculate the potential energy of the rock as the product *mgh*, where *m* is its mass, *g* is the gravitational acceleration, and *h* is its height. But before we can make the calculation, we must define a state at which *h* = 0, and this is arbitrary, meaning that we can choose any state we wish. We might, in the case of the rock on the hill, choose to measure *h* from the valley floor, but we could just as well choose sea level as this state. Equations (5.4)–(5.6) present the same kinds of choices. In each case, μ_i is the same definite quantity (corresponding to the potential energy of the rock on the hill). Each equation can be rearranged to the form

$$\mu_i - \mu_c^0 = RT \ln c_i$$

using Eq. (5.4) as an example. Since, for a given solution, c_i , m_i , and x_i have different numerical values, so, too, do the quantities $(\mu_i - \mu_c^0)$, $(\mu_i - \mu_m^0)$, and $(\mu_i - \mu_x^0)$. Each of these quantities gives the value of the chemical potential μ_i , relative to its standard state value. Each expression is thermodynamically acceptable; they differ only in the values they assign to the standard potential. We can easily find the relationship between these different standard chemical potentials. Let us compare Eqs. (5.4) and (5.6). Recalling that we have assumed that the system is in its reference state, which usually is the infinitely dilute solution, we can use Eq. (5.1), substituting it into (5.6) to eliminate x_i , and setting Eqs. (5.4) and (5.6) equal to yield

$$\mu_c^0 = \mu_x^0 + RT \ln \frac{M_1}{1000\rho_1}$$

For example, if the solvent is water, then $M_1 = 18$ and $\rho_1 = 1.0$, and we find $\mu_c^0 = \mu_x^0 - RT$ ln 55.5. Concluding this exposition, we see that the selection of a standard state is arbitrary and that we select a standard state when we choose a concentration scale. [Some authors write of "adopting the 1 M standard state," for example, which merely means that the molar scale was used, with Eq. (5.4) showing that when $c_i = 1$, then $\mu_i = \mu_c^0$.]

In the more general case where activity coefficients may deviate from unity, Eqs. (5.4)–(5.6) become

$$\mu_i = \mu_c^0 + RT \ln \gamma_c c_i \tag{5.7}$$

$$\mu_i = \mu_m^0 + RT \ln \gamma_m m_i \tag{5.8}$$

$$\mu_i = \mu_x^0 + RT \ln \gamma_x x_i \tag{5.9}$$

where the subscript *i* has been omitted from the μ^0 and γ terms to reduce typographical clutter. In general γ_c , γ_m , and γ_x differ for the same system, although in dilute solutions they have nearly the same value (Glasstone 1947, p. 355). Certain conventions allow us to carry out calculations to levels of accuracy appropriate to many practical situations where extreme accuracy is not required. We will adopt these conventions, which are repeated from Chapter 3 for convenience:

- 1. The fugacity of a *gas* is taken equal to its pressure (or partial pressure) in atm, at low to moderate pressures.
- 2. The activity of a liquid *solvent* in a solution is equal to its mole fraction. It follows that the activity of a pure liquid is 1.00 and that this is its standard state. (This convention also applies to liquid solutes if desired.)
- 3. The activity of a pure *solid* is 1.00; this is the same as convention 2.
- 4. The activity of a *solute* in an infinitely dilute solution will be taken equal to its molar concentration; that is, $\gamma_c = 1.00$; the solute is in its reference state. The standard state is rather peculiar; it approximates to the 1 M solution, but at this concentration the solution probably does not behave ideally, so although its activity is unity by definition, some of its properties are those of the reference state. (We further expand on this in Section 7.2.)
- 5. The activity of an *uncharged solute* will be taken as 1.00 at any concentration. This is an approximation justified by recognizing that uncharged molecules experience much weaker forces of interaction than do ions. The approximation improves as the solution is made more dilute.
- 6. The activity coefficient of an *ion* can be drawn from experimentally determined results (Harned and Owen 1958) or calculated from theory, as will be described in Section 8.3.

It is instructive to write Eq. (5.7) in this expanded form

$$\mu_i = \mu_i^0 + RT \ln c_i + RT \ln \gamma_i \tag{5.10}$$

This equation shows how the numerical value of the chemical potential μ_i receives a contribution from the arbitrarily assigned standard state (μ_i^0) , a contribution from $RT \ln c_i$, which describes the composition dependence of the ideally behaved constituent, and a contribution from $RT \ln \gamma_i$, which describes the nonideal behavior of the constituent. Although not explicitly indicated, γ_i is also composition-dependent, and as $c_i \rightarrow 0$, $\gamma_i \rightarrow 1$.

PROBLEMS

- **5.1.** Calculate the mole fraction x_2 of benzoic acid in an aqueous solution 1.5 mM in benzoic acid. Also calculate the mole fraction x_i of water in this solution.
- **5.2.** Concentrated hydrochloric acid is labeled to contain about 38.0% by weight of HCl, and its density is about $1.19 \,\mathrm{g}\,\mathrm{mL}^{-1}$. Calculate the approximate molar concentration of HCl in this solution.
- 5.3. What is the molar concentration of pure water?
- **5.4.** Calculate the difference between standard chemical potentials in dilute aqueous solution based on the molar and the mole fraction standard states, at 25 °C.
- **5.5.** Obtain an equation with which molar and molal concentrations may be interconverted in dilute solution.
- **5.6.** Consider a liquid solution of solvent 1 and solute 2. Let n_1 and n_2 , respectively, be the numbers of moles of solvent and solute in a given mass of solution, whose density is *p*. Then derive an *exact* equation relating x_2 and c_2 . [*Hint:* The correct result must reduce to Eq. (5.1) in very dilute solution.]
- **5.7.** A 0.1 M solution of glucose (MW = 180 g/mol) has a density of 1.04 g/mL. What is the molality of the solution?
- **5.8.** Consider a 0.28 m solution of glucose. Calculate the mole fraction and the %w/w of glucose in this solution.

6

PHASE TRANSFORMATIONS

6.1. PURE SUBSTANCES

Phase Diagrams. In this section we treat equilibria in heterogeneous systems, that is, systems consisting of more than one phase. A phase of matter is uniform in chemical composition and physical state. It may be subdivided, but it remains a single phase. For example, a system consisting of ordinary crushed ice dispersed in water possesses two phases: ice and water. These types of systems were treated briefly in Chapter 4. Figure 6.1, which appeared earlier as Fig. 4.1, is a pressure–temperature phase diagram for the simplest case, a pure substance possessing only one form of each of the three phases solid, liquid, and vapor (gas). (A pure substance can have only a single vapor phase, and most pure substances have only a single liquid phase,¹ but many solid phases may exist, as will be described below.)

The line *OC* in Fig. 6.1 describes all systems in which the solid and liquid phases coexist in equilibrium; that is, any pair of *P*, *T* coordinates on this line describe the *melting temperature* of the solid at that pressure. Similarly, line *OB* gives the *boiling temperature* as a function of pressure, and line *OA* gives the *sublimation temperature*. Point *O* is called the *triple point*, a unique pair of *P*, *T* values at which the solid, liquid, and vapor are in mutual equilibrium. The slopes of these lines are given by Eq. (6.1), the Clapeyron equation, from Chapter 4:

¹ Some substances reveal the existence of a second liquid phase called the *liquid crystalline phase*. It is recognized by its optical properties.

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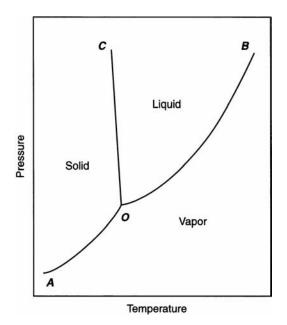


Figure 6.1. Pressure-temperature phase diagram of a pure substance.

$$\frac{dP}{dT} = \frac{\Delta H}{T\Delta V} \tag{6.1}$$

where ΔH is the enthalpy change for the process and ΔV is the volume change. For the special case of the liquid–vapor transition (boiling), this equation is usually used in the approximate version called the *Clausius–Clapeyron equation*:

$$\frac{d\ln P}{dT} = \frac{\Delta H_{\rm vap}}{RT^2} \tag{6.2}$$

In its integrated forms the Clausius-Clapeyron equation becomes

$$\log P = \frac{\Delta H_{\text{vap}}}{2.3RT} + C \tag{6.3}$$

or

$$\log \frac{P_2}{P_1} = -\frac{\Delta H_{\text{vap}}}{2.3R} \left(\frac{T_2 - T_1}{T_1 T_2} \right)$$
(6.4)

where *C* is a constant; ΔH_{vap} , the molar heat of vaporization, is assumed to be a constant throughout the temperature range of interest; and the equations relate boiling temperature to pressure.

The Gibbs phase rule is

$$F = C - P + 2 \tag{6.5}$$

where *P* is now the number of phases in the system at equilibrium, *C* is the number of components, and *F* is the number of degrees of freedom.²*F* is the number of variables that must be fixed in order to completely define the system. For example, along line *OC* in Fig. 6.1 there are two phases (solid and liquid) and one component (since this is the diagram for a pure substance), so F = 1 - 2 + 2 = 1. According to this result, the system along line *OC* possesses one degree of freedom. This means that we can select *either* the pressure *or* the temperature at will (provided the value lies within the *OC* range); the other variable is then established by the equilibrium.

At the triple point O, F = 1 - 3 + 2 = 0; the system has no degrees of freedom. The three phases solid, liquid, and vapor are all in equilibrium, and if this condition is to be maintained, neither the temperature nor the pressure may be altered.

Figure 6.1 and the Clausius–Clapeyron equation show that the pressure may be a fairly sensitive function of temperature along the vaporization line *OB*. By definition, the *normal boiling point* T_b is the boiling temperature when P = 1 atm; this is the boiling temperature usually measured in the laboratory (or the kitchen). The melting point (also called the *freezing point*) is not very sensitive to pressure.

Polymorphism. A pure substance may be capable of existing in more than one crystalline solid form. Each such crystalline solid is a separate phase, and these forms are called *polymorphs*. The phenomenon of polymorphism (also known as *allotropy*) is widespread, and it has pharmaceutical ramifications. Polymorphs have different arrangements of the molecules in their crystal structures, but chemically they are identical. The two or more polymorphs of a substance possess different free energies, and the polymorph that has the lowest free energy is the thermodynamically most stable form. The other forms are thermodynamically unstable relative to the stable form, but it may happen that the rate of transformation from the unstable to the stable forms is so slow as to be negligible or practically unimportant, in which case the unstable polymorph is said to be *metastable*. For example, the element carbon can exist in two polymorphic forms called graphite and diamond. Graphite is the thermodynamically stable form, and diamond is metastable with respect to it, but although diamond is thermodynamically unstable, the timescale on which it transforms to the more stable form is of no human concern. Some polymorphic transformations may be quite fast, however.

A given substance may possess numerous polymorphs—phenobarbital has at least 8 and may have 11 of them—but according to the phase rule, the maximum number of phases, including solid phases, that can coexist in equilibrium is P = C + 2 (i.e., P is maximized when F is set to 0); for a pure substance this is three phases. Figure 6.2

 $^{^2}$ The number of components may differ from the number of constituents. Here is a simple way to determine *C*, the number of components: *C* is equal to the minimum number of bottles of pure substances required to prepare the system in the laboratory.

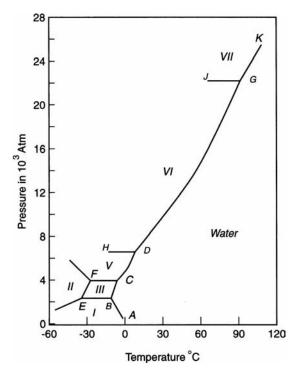


Figure 6.2. Pressure–temperature phase diagram of water at high pressures, showing the six ice polymorphs. [Reproduced by permission from Findlay et al. (1951).]

is the phase diagram of water at extremely high pressures (Findlay et al. 1951). This phase diagram (which is based on the experimental work of P. W. Bridgman) shows six ice polymorphs; these are labeled ice I (this is ordinary ice), ice II, ice III, ice V, ice VI, and ice VII (the reported discovery of ice IV was erroneous). Observe that a maximum of three phases may exist at any fixed combination of temperature and pressure.³

The pharmaceutical significance of polymorphism lies in two features: (1) The different crystal forms have different physical properties and (2) polymorphs may interconvert on a pharmaceutically pertinent timescale. These features have led to much pharmaceutical research in this area (Carstensen 1973, pp. 113–124; Haleblian and McCrone 1969; Haleblian 1975; Florence and Attwood 1981, Chapter 2; Yu 2007; Chen et al. 2005). It has been found that the solubilities of polymorphic forms of a drug are different. If the solubility of the less stable form is greater than that of the more stable form, its solution will be unstable with respect to the more stable solid form because it is supersaturated with respect to this form. Precipitation may occur unexpectedly in such a situation unless some form of stabilization can be devised. The bioavailability of a drug may depend on the drug's

³ For a fictional use of the concept of polymorphism see the note on ice-nine in Wikipedia (http://en. wikipedia.org/wiki/Ice-9).

polymorphic form. Chemical stability of drugs, as well as the physical stability of pharmaceutical dosage forms, may be dependent on the polymorphic form of the drug and its propensity for transformation to a more stable polymorph. The presence of more stable inactive polymorphs of a drug can lead to very serious problems. The case of the AIDS drug Ritonavir is exemplary. This very effective drug could not be used after the sudden appearance of a new, more stable but less soluble polymorph. This new inactive polymorph completely overcame the original drug. Any attempt at making the original polymorph only resulted in the inactive form of the drug. Eventually, the production of Ritonavir was abandoned (Bauer et al. 2001). It is also possible that a single solid compound may form a number of different polymorphs. ROY, 5-methyl-2-[(2-nitrophenyl)amino]-3-thiophenecarbonitrile, a synthetic drug-substance intermediate, dramatically illustrates this latter concept. ROY takes its name from the colors (red, orange, and yellow) of three of its polymorphs. Further studies identified up to seven more polymorphs, making ROY the most polymorphic organic compound in the Cambridge Structural Database (Chen 2005). The recognition of the possible existence of inactive, more stable polymorphs of a certain drug makes polymorph screening an important component of modern drug development. One subtle aspect of polymorphism is that polymorphs can form not only upon crystallization from solutions, but also from cooling of pure melted compounds and as the result of crystal conversion directly in the solid state.

Note that the formation of a crystalline hydrate (or other solvate), in which the compound crystallizes with one or more molecules of solvent in its crystal structure, is not true polymorphism; a crystal hydrate is not chemically the same substance as the unhydrated substance.

The Amorphous State. We have seen that a pure substance may assume any one of several crystalline solid forms called *polymorphs*. There exists yet another possibility called the *amorphous* or *glassy* state, in which the substance appears to be solid in its consistency, yet X-ray diffraction data show the absence of the periodic array of molecules characteristic of the crystalline state. The amorphous (i.e., formless) state is really the supercooled liquid, which, although below its normal freezing point, has not adopted the orderly arrangement of molecules characteristic of the crystall. Although it appears to be a solid, it is really a highly viscous liquid. Presumably some kinetic barrier to crystallization permits supercooling to take place. This pathway to the amorphous state is not the only one, however, and it has been found possible to generate amorphous samples by subjecting crystalline solids to high-energy processes such as grinding, milling, and freeze drying. The amorphous state is best detected by means of X-ray powder spectra.

The amorphous state is unstable with respect to (it is of higher energy than) the crystalline solid, to which it may revert on a generally unpredictable timescale. Its pharmaceutical advantages and disadvantages follow from these properties. Higher solubility and bioavailability may be achieved with amorphous solids, but transformation to the crystalline state is a possibility. Experimental study of each substance is required to establish its characteristic behavior.

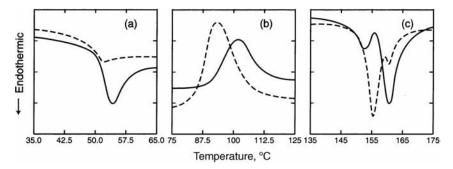


Figure 6.3. DSC traces of two amorphous samples of indomethacin. See text for explanation. [Reproduced by permission from Yoshioka et al. (1994).]

The amorphous state is often studied by means of differential scanning calorimetry (DSC), in which the temperature of the sample is raised while the heat absorbed or released by the sample is monitored. Figure 6.3 shows DSC curves for two amorphous samples of the drug indomethacin (Yoshioka et al. 1994). One sample (dashed lines) had been prepared by rapidly cooling the melted drug; the other sample (solid lines) had been more slowly cooled. Besides these amorphous samples, indomethacin also forms two crystalline polymorphs: the α form with melting point 155 °C and the γ form with melting point 161 °C.

As the temperature sweeps through the range 35–65 °C, in Fig. 6.3a, both amorphous samples show endothermic peaks (they are absorbing heat) as they undergo a transition. The onset of this transition, at about 50 °C, is called the *glass transition temperature*, T_g . With increased temperatures, the samples undergo crystallization in Fig. 6.3b, with the release of heat (the heat of crystallization, which can be measured from the areas under the crystallization peaks). Finally, in Fig. 6.3c, both samples melt. Observe that the two melting crystalline samples are actually mixtures of the α and γ polymorphs (each curve has two components), but they differ in terms of which polymorph is present in major fraction.

The properties of the amorphous state are a consequence of its high energy content, which is its dominant characteristic. It is a relatively unstudied state of matter, for which increasing pharmaceutical applications may be expected.

6.2. MULTICOMPONENT SYSTEMS

This is an abbreviated treatment of this topic, limited in these two ways: (1) We consider only binary (i.e., two-component) systems, and (2) we omit certain topics as not particularly pertinent to our interests (fractional distillation is an example). Fuller treatments are available (Rossini 1950, Chapter 32; Atkins 1994, Chapter 8).

Liquid–Liquid Systems. We will omit consideration of the vapor phase, in principle by postulating that it is excluded from the system, in practice by working (usually) under the ambient fixed atmospheric pressure. We begin by considering a system of two liquids. Of course, whether a substance is a liquid or a solid depends (at fixed pressure) on the temperature, but common usage denotes as liquids those substances that exist in this state at or near room temperature. Pairs of liquids often are classified as essentially completely immiscible (such as mercury and water), as completely miscible in all proportions (e.g., ethanol and water), or as partially miscible (e.g., diethyl ether and water). The completely immiscible case need not concern us, since it effectively consists of two separate pure substances. Completely miscible systems are dealt with in Chapter 7. We are left to consider those pairs of liquids that are miscible in some proportions but are immiscible in other proportions.

Inasmuch as we have fixed the pressure, the two experimental variables by means of which the system may be manipulated are the temperature and the composition of the system, and phase diagrams are commonly constructed with these variables as the coordinates. Usually the composition is expressed as mole fraction or as percent by weight. Figure 6.4 shows a schematic temperature–composition phase diagram for a partially miscible pair of liquids, 1 and 2. Any combination of temperature and composition giving a point outside the phase boundary line describes a homogeneous system; in this region, 1 and 2 are mutually miscible. Note that small concentrations of 1 will dissolve in 2, and vice versa; moreover, as the temperature increases, the extent

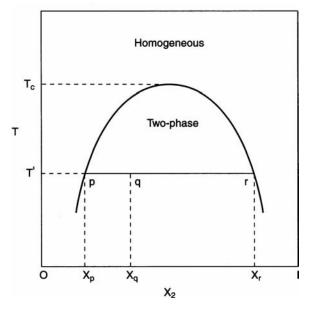


Figure 6.4. Schematic temperature-composition phase diagram for two partially immiscible liquids 1 and 2; x_2 is the mole fraction of 2 and T_c is the upper critical temperature.

of mutual solubility increases. At any temperature above T_c (which is called the upper critical temperature), the two liquids are miscible in all proportions.

But if the temperature–composition combination places the system under (within) the phase boundary line, two phases form. One phase is predominantly 1 saturated with 2, the other is largely 2 saturated with 1. At any given temperature, say, T' in Fig. 6.4, the horizontal *tieline pr* connects the arms of the phase diagram, and the compositions of the two phases are given by x_p and x_r . Moreover, if x_q is the overall composition of the system, the amounts of the two phases are in the ratio of the distances pq/qr. Figure 6.5 shows the experimental phase diagram for the phenol–water system [see Findlay et al. (1951, p. 95); the melting point of phenol is 41°C, and phenol is being treated as a liquid in this context]. This diagram is helpful in determining the ranges of compositions that will yield homogeneous solutions of phenol in water at room temperature (25°C). Liquefied Phenol U.S.P. contains 89% by weight of phenol, placing it in the single-phase region of the diagram.

Example 6.1. 50.0 g of Liquefied Phenol U.S.P. is diluted with 50.0 mL of water at room temperature. Analyze the outcome of this procedure.

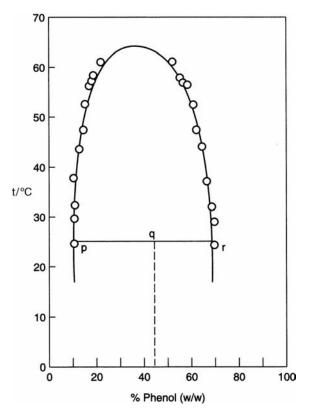


Figure 6.5. Phase diagram for the phenol-water system. See discussion in Example 6.1.

Since Liquefied Phenol contains 89% w/w of phenol and the density of water is 1.0 g mL^{-1} , the system as prepared contains 44.5% w/w phenol with a total weight of 100 g. Figure 6.5 shows the 25°C tieline with point *q* given by the 44.5% system composition. This point lies within the two-phase boundary, so the system will separate into two layers. Reading the compositions of the layers at points *p* and *r* tells us that one phase will contain 8% phenol and the other phase will contain 71% phenol. The ratio pq/qr = (44.5 - 8)/(71 - 44.5) = 1.38.

We can go further than this. Since 100 g of total system contains 44.5 g of phenol, we can write

Weight of phenol in aqueous layer + weight of phenol in phenolic layer = 44.5 g Letting *x* be the weight of the aqueous layer in 100 g of sample gives

$$0.08x + 0.71(100 - x) = 44.5$$

resulting in x = 42.1 g as the weight of the aqueous layer and therefore 57.9 g as the weight of the phenolic layer. The aqueous layer contains (0.08) (42.1) = 3.4 g of phenol, and the phenolic layer contains (0.71)(57.9) = 41.1 g of phenol. Note, incidentally, that pq/qr = 1.38 = 57.9/42.1.

In these two-component systems each phase is a solution, which can be defined as a phase of variable composition. Notice that we have not identified one of the components as the solute and the other as the solvent; such a designation has no thermodynamic significance and is done solely for our convenience.

Liquid–Solid Systems. Imagine a two-component system consisting of two solids A and B brought to a temperature above the melting points of both. Then in the simplest instance a one-phase system will form consisting of a liquid solution of A and B (Findlay et al. 1951, p. 135). Referring to Fig. 6.6, the area labeled L (for liquid phase) will include the system as described, with its precise location in the diagram depending on the temperature and the composition.

The points A and B in Fig. 6.6 represent the melting points of solids A and B. Now let the temperature be lowered (always allowing the system to remain at equilibrium). Suppose the system initially is represented by point E. When the temperature reaches point G, pure solid A will begin to form, and as the temperature continues to fall (as heat is withdrawn from the system), more solid is formed. Throughout the area ADC the system consists of pure solid A dispersed in a solution of A and B. Its composition is given by tielines, such as HK in the figure.

When the system temperature reaches level C, the temperature ceases to fall, even though heat continues to be withdrawn from the system; point C has no degrees of freedom. (Recall that we have fixed the pressure.) At point C, solid A, solid B, and solution phase are in mutual equilibrium. The solid phase at this point is a finely divided two-phase dispersion of crystalline A and B called a *eutectic*, and C is the *eutectic point*. Microscopic examination reveals that the eutectic is a mixture and not a single phase. The composition of the eutectic mixture is fixed for a given pair of substances. Observe that the eutectic melts at a lower temperature than does either of

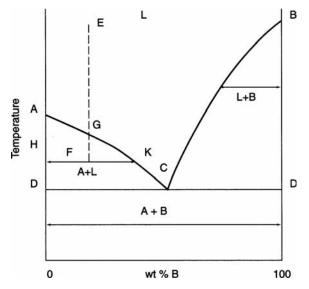


Figure 6.6. The simplest solid–liquid phase diagram for a two-component system of A and B, where *L* represents the liquid (solution) phase; *C* is the eutectic point. [Reproduced by permission from Findlay et al. (1951).]

its pure components. Eutectic formation is observed widely in geologic deposits and metal alloys, and the phenomenon is of pharmaceutical importance. Numerous drugs form eutectic mixtures, with the consequence that they may liquefy at ambient temperature owing to the melting point decrease characteristic at the eutectic point. Acetaminophen, aspirin, menthol, phenacetin, phenol, and thymol are some of these substances that are prone to eutectic formation. Special care in formulating or compounding these compounds is necessary (Thompson 1998, pp. 34–35).

We tend to think of curves such as AC and BC in Fig. 6.1 as freezing point (or melting point) curves, but from the thermodynamic point of view they can just as well be viewed as solubility curves. Suppose, for example, that A is a liquid at room temperature but that B is a solid. Then the curve BC can be interpreted as the solubility of B in A. We will not pursue this line of interpretation because Chapter 10 is entirely concerned with solubility.

A traditional laboratory technique for the confirmation of identity of a solid substance is to mix some of the sample with an authentic specimen and to measure the melting point. If this *mixed melting point* is the same as that of the melting point of the authentic specimen, the sample is very likely the same compound. If, on the other hand, the melting point of the mixture is decreased, the two substances are different. This is a consequence of the mutual depression of melting points seen in Fig. 6.6 when two components are mixed.⁴

⁴ The melting of ice on winter roads by spreading salt is another manifestation of the phenomenon. NaCl and H₂O form a eutectic of composition 23.3% NaCl at a eutectic temperature of -21.1° C.

The region in Fig. 6.6 labeled A + B and lying entirely below point *C* may consist merely of the two crystalline phases of A and B (leaving aside the phenomenon of polymorphism). But another possibility is that A and B may form a *solid solution*, which is a homogeneous single-phase state of matter, no different in principle from a liquid solution. Some drugs are known to form solid solutions (Carstensen 1977, pp. 23–26).

PROBLEMS

- **6.1.** Suppose that a solution is prepared at 70°C to contain 65% by weight of phenol in water. The solution is slowly cooled. At what temperature will it separate into two phases?
- 6.2. Calculate the degrees of freedom at the eutectic point C in Fig. 6.6.

7

SOLUTIONS OF NONELECTROLYTES

7.1. IDEAL SOLUTIONS

A nonelectrolyte is a substance that is uncharged and that does not sensibly give rise to ions. Our analysis will be sufficiently general if we consider solutions of two nonelectrolytes, labeled 1 and 2; the results can be extended to more components if necessary. For the present we limit discussion to single-phase systems.

A convenient starting place is with the experimental observation known as *Raoult's law*, which describes a particularly simple type of solution behavior in the form of

$$p_i = x_i P_i^* \tag{7.1}$$

Raoult's law states that the partial pressure p_i of constituent *i* over its solution is directly proportional to its mole fraction in the solution, where the proportionality constant P_i^* is the vapor pressure of the pure liquid (i.e., when $x_i = 1$). An ideal liquid solution is then one in which Raoult's law is obeyed over the entire range of composition, at all temperatures and pressures. As may be imagined, Raoult's law represents a limit of simple behavior toward which certain systems tend, rather than an exact description; but if the solution components are chemically very similar and are nonpolar molecules, behavior very close to the ideal may be observed. A solution of benzene and toluene illustrates such behavior.

It can be proved (Glasstone 1947, p. 320) that if Raoult's law applies to one of the constituents of a solution, then it must also apply to the other. Figure 7.1 shows Raoult's

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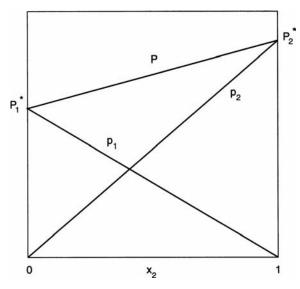


Figure 7.1. Raoult's law behavior of both components of an ideal solution; p_1 and p_2 are the partial pressures; *P* is the total pressure.

law behavior for an ideal solution. Since the total pressure is the sum of the partial pressures, then

$$P = x_1 P_1^* + x_2 P_2^* \tag{7.2}$$

This is the equation of the topmost line in Fig. 7.1.

We know from Chapter 4 that at equilibrium the chemical potentials of constituent *i* are equal in the vapor and liquid phases, or $\mu_i(g) = \mu_i(l)$. For a mixture of ideal gases, we also can write [see Eq. (3.35)], that

$$\mu_i(g) = \mu_i^*(g) + RT \ln P_i$$

If we combine this relationship with the foregoing equality and with Raoult's law, we obtain

$$\mu_i(l) = \mu_i^{\circ}(l) + RT \ln x_i \tag{7.3}$$

where $\mu_i^{\circ}(l) = \mu_i^*(g) + RT \ln P_i^*$. Equation (7.3) may be taken as an alternative description of an ideal-liquid solution (Smith 1977, p. 78). The standard chemical potential $\mu_i^{\circ}(l)$ is the chemical potential of pure component i (i.e., when $x_i = 1$).

We can develop the thermodynamic properties of the ideal solution as follows. The total free energy of the solution is given by

$$G = \mu_1 x_1 + \mu_2 x_2 \tag{7.4}$$

where x_1 and x_2 are mole fractions. The free-energy change on mixing 1 and 2 is equal to the free energy of the solution after mixing minus the free energy of the pure components before mixing, or

$$\Delta G_{\rm mix} = \sum \mu_i x_i - \sum \mu_i^o x_i \tag{7.5}$$

Substituting from Eq. (7.3) into Eq. (7.5) leads to

$$\Delta G_{\text{mix}}^{\text{ideal}} = RTx_1 \ln x_1 + RTx_2 \ln x_2 \tag{7.6}$$

as the ideal free energy of mixing. Since the mole fractions are less than one, the free energy of mixing is negative and the process is spontaneous.

The entropy of mixing is easily obtained by applying the relationship [Eq. (3.7)]

$$\left(\frac{\partial\Delta G}{\partial t}\right)_P = -\Delta S \tag{7.7}$$

to Eq. (7.6). The result is

$$\Delta S_{\text{mix}}^{\text{ideal}} = -Rx_1 \ln x_1 - Rx_2 \ln x_2 \tag{7.8}$$

Therefore the entropy of mixing is positive, as we would expect. From the identity $\Delta G = \Delta H - T \Delta S$ we obtain, making use of Eqs. (7.6) and (7.8),

$$\Delta H_{\rm mix}^{\rm ideal} = 0 \tag{7.9}$$

Finally, from Eq. (3.6) we obtain

$$\left(\frac{\partial\Delta G}{\partial P}\right)_T = \Delta V \tag{7.10}$$

Applying this to Eq. (7.6), we get

$$\Delta V_{\rm mix}^{\rm ideal} = 0 \tag{7.11}$$

Equations (7.6), (7.8), (7.9) and (7.11) give the essential thermodynamic properties of the ideal solution. We can make some molecular interpretations of these results. In an ideal solution, the three pairwise interactions between 1–1 molecules, 2–2 molecules, and 1–2 molecules are all energetically and spatially identical, so replacement of a 1 molecule by a 2 molecule anywhere in the solution leads to no energy or volume changes; hence $\Delta H_{\text{mix}}^{\text{ideal}} = 0$ and $\Delta V_{\text{mix}}^{\text{ideal}} = 0$. (It is these stringent constraints that account for the rarity of experimental examples of ideal solutions, because if two real

molecules have different identities, their energies and space-filling requirements will differ to at least some degree.) The ideal entropy of mixing is positive because the mixed system is more disordered than is the initial system of separated species and the number of configurational microstates is greater. Since $\Delta H_{\text{mix}}^{\text{ideal}} = 0$ and $\Delta S_{\text{mix}}^{\text{ideal}} > 0$, the negative value of $\Delta G_{\text{mix}}^{\text{ideal}}$ is entirely entropy-driven.

7.2. NONIDEAL SOLUTIONS

It will be no surprise to learn that few real solutions behave ideally. Nevertheless, fairly simple behavior is widely observed in solutions that are very dilute with respect to one component. It will now be convenient to designate the component (to be labeled component 1) that is present in great excess as the *solvent* and designate component 2, present in low concentration, as the *solute*. The solvent is obviously a liquid, but the solute may be either a liquid or a solid.

First consider Fig. 7.2, which shows vapor pressure–composition curves for both solution components when derivations from ideality occur. In this figure the dashed lines show ideal Raoult's law behavior (compare with Fig. 7.1), whereas the solid lines show positive deviations from Raoult's law (Fig. 7.2a) and negative deviations (Fig. 7.2b).¹ The two components may exchange roles as solvent and solute, depending on which is in excess.

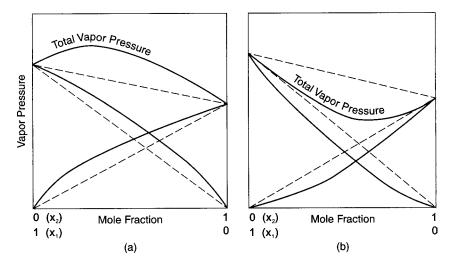


Figure 7.2. Nonideal solution behavior showing positive deviations (a) and negative deviations (b) from Raoult's law. The mole fraction scale runs from 0 to 1 for one of the components and from 1 to 0 for the other.

¹ A solution having the composition corresponding to either a maximum (Fig. 7.2a) or a minimum (Fig. 7.2b) in the vapor pressure curve will distill as a constant boiling mixture of constant composition, called an *azeotrope*. For instance, 95% alcohol is an azeotrope containing 95.57% by weight (94.9% by volume) of C_2H_5OH .

Now, in a dilute solution (i.e., dilute with respect to component 2, the solute), the solvent, component 1, approaches a mole fraction of unity, and its vapor pressure approaches that expected from Raoult's law; this behavior can be seen in Fig. 7.2, where the dashed and solid lines approach asymptotically as *x* approaches unity. This is reasonable behavior, since in this circumstance (the very dilute solution) the solvent molecules are surrounded essentially only by other solvent molecules, and hence are practically unperturbed by solute molecules. But it is otherwise for the solute molecules in dilute solution, because then each solute molecule finds itself in an environment of essentially only solvent molecules, which is clearly not typical of the purely solute environment. Consequently the solute does not follow Raoult's law in dilute solution.

Despite this result, a certain simplicity of behavior by the solute can be discerned. Experiment shows that in the very dilute solution the vapor pressure of the solute is, in the limit of zero concentration, a linear function of its mole fraction, as in

$$p_2 = x_2 k_2^x \tag{7.12}$$

which should be compared with Raoult's law, Eq. (7.1). Equation (7.12) is called *Henry's law*, and the constant of proportionality k_2^x is the Henry's law constant. The distinction between Raoult's law and Henry's law is easily seen graphically in Fig. 7.3.

The thermodynamic description of solute behavior in very dilute solutions is based on Henry's law, and it leads, by the same kind of argument used for ideal solutions,

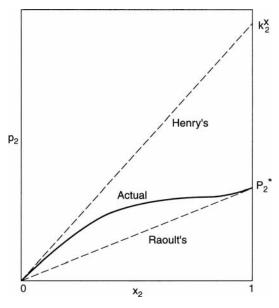


Figure 7.3. P_2^* is the vapor pressure of the pure solute ($x_2 = 1$), and the actual vapor pressure curve tends to this value. The Henry's law constant k_2^x is a hypothetical value obtained by linear extrapolation to $x_2 = 1$ of the tangent to the actual curve at $x_2 = 0$.

to Eq. (7.13), which is very similar to Eq. (7.3) for the ideal solution, except that the standard chemical potential incorporates the Henry's law constant:

$$\mu_2 = \mu_2^0 + RT \ln x_2 \tag{7.13}$$

We saw in Chapter 5 that we may base our standard state definitions on either the mole fraction, the molal, or the molar concentration scales. Equations (7.12) and (7.13) make use of the mole fraction convention, and this is shown in Fig. 7.3 and again in Fig. 7.4a. Figure 7.4b shows the significance of the standard state convention on the molar scale. Note how the superscript *x* or *c* is used with the Henry's law constant to clarify the definition. Of course, k_2^x and k_2^c are different; their relationship can be worked out as we did in Chapter 5 [see also Grant and Higuchi (1990, p. 93)].

Finally we must consider nonideal solution behavior outside the very dilute solution range. We have seen how to cope with this behavior (Section 5.3) by defining activity coefficients, so that the actual behavior is quantitatively expressed by an activity coefficient that measures the deviation between real and ideal behavior. Now we can see that the adoption of a criterion of ideal behavior is critical to expressing the extent of deviation from this ideal. We are here approaching from a different direction an issue already faced in Section 5.3. At that point we had defined the activity of the solvent to be equal to its mole fraction. We now see that this is equivalent to assuming that Raoult's law is obeyed by the solvent, which is a reasonable assumption in the dilute solution range. The activity of the solute, on the other hand, we took as equal to its molar concentration in the very dilute range. This is a Henry's law reference state; the standard state is as shown in Fig. 7.4b. In Chapter 8 we will learn how to estimate activity coefficients for ionic species, which are notorious for their nonideal behavior; except in very concentrated solutions,

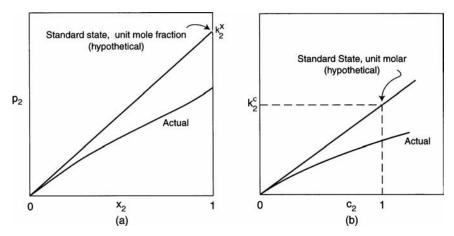


Figure 7.4. Henry's law constant k_2^x on (a) the mole fraction scale and (b) the molar scale. These standard states are different. They are both hypothetical, because real behavior (solid curves) deviates from these linear extrapolations based on very dilute behavior.

however, the activity coefficients of nonelectrolytes can be taken as unity for most practical work.

The thermodynamic properties of real solutions are sometimes expressed in terms of *excess functions*, which are defined as the difference between the actual value of the function and the ideal value. For example, the excess entropy of mixing is

$$S^E = \Delta S^{\text{real}}_{\text{mix}} - \Delta S^{\text{ideal}}_{\text{mix}}$$

The excess functions can be positive or negative.²

7.3. PARTITIONING BETWEEN LIQUID PHASES

The Partition Coefficient. Suppose that we bring two immiscible liquids in contact and then incorporate a nonelectrolyte solute such that its concentration is in the dilute solution range. The solute will distribute itself between the two phases, each of which constitutes a solution.³ Since the phases will arrange themselves according to their densities, let us identify them as the upper (U) and lower (L) phases. The distribution of solute between the phases is called *partitioning*. The typical separatory funnel operation exemplifies this system. We take the pressure and temperature as fixed.

At equilibrium the chemical potentials of the solute in the upper and lower phases are equal:

$$\boldsymbol{\mu}_2^U = \boldsymbol{\mu}_2^L \tag{7.14}$$

These chemical potentials will be written out for the Henry's law molar standard state definition, giving

$$\mu_U^* + RT \ln c_U = \mu_L^* + RT \ln c_L \tag{7.15}$$

where for convenience the subscript 2 is omitted, assuming that the solute is meant.

Rearrangement of Eq. (7.15) gives

$$\Delta \mu^0 = -RT \ln \frac{c_U}{c_L} \tag{7.16}$$

where $\Delta \mu^0 = \mu_U^* - \mu_L^*$. Comparison of Eq. (7.16) with the important equation [Eq. (4.23)]

$$\Delta G^0 = -RT \ln K \tag{7.17}$$

shows that the ratio c_U/c_L has the character of an equilibrium constant. In fact, it is the equilibrium constant of this "reaction":

² A class of solutions called *regular solutions* is defined to have $S^E = 0$ and $H^E \neq 0$, so that entropically such solutions behave ideally, but they undergo nonideal energy changes. Regular solutions are commonly formed from nonpolar components [see Hildebrand et al. (1970); see also Chapter 10 (below)].

³ Besides the phenomenon in which the solute distributes between the two phases, the upper phase will be saturated with respect to the lower phase solvent, and vice versa. This mutual saturation alters the solvent properties of the two phases, but it does not affect the thermodynamic argument.

Solute in phase $L \rightleftharpoons$ Solute in phase U

This quantity is labeled P and is called the partition coefficient

$$P = \frac{c_U}{c_L} \tag{7.18}$$

Partition coefficients are usually expressed in terms of their base 10 logarithms, $\log P$. This makes the numerical values directly proportional to the standard free-energy change, according to Eq. (7.17), and it provides convenient magnitudes, since P itself can be much smaller or larger than unity.

Log P values have great utility in drug discovery and drug delivery research programs. For these purposes the solvent of the upper phase is usually selected to be 1-octanol, with water serving as the lower phase solvent. The partition coefficient is then defined as

$$P = \frac{c_{\text{octanol}}}{c_{\text{water}}} \tag{7.19}$$

Since water is more polar than is octanol, very polar solutes tend to have greater affinity for the aqueous phase and therefore to have P values smaller than unity, whereas nonpolar solutes have P values greater than unity. Consequently, log P is often used as a quantitative measure of a compound's polarity.⁴

Log P values can be measured experimentally by the separatory funnel technique, or modifications of it. Sometimes it is useful to be able to predict a log P value, as for example if a compound of interest is not available or has not yet been synthesized. Empirical methods, making use of a large body of experimental log P values, have been developed that allow log P to be estimated solely on the basis of knowledge of the solute's molecular structure (Leo et al. 1971; Nys and Rekker 1974).

Table 7.1 lists a few $\log P$ values. Notice, for the series of normal alcohols, how the trend of $\log P$ values appears to accord with our qualitative notions of the polarities in this series. The $\log P$ of the aromatics is also consistent with expectations.

Solute	Log P	Solute	Log P
Methanol	-0.74	Benzene	2.13
Acetic acid	-0.24	Phenol	1.46
Ethanol	-0.32	Aniline	0.94
1-Propanol	0.34	Toluene	2.69
1-Butanol	0.88	Naphthalene	3.37
1-Pentanol	1.40	Aspirin	1.21

Table 7.1. Log P (octanol/water) for some solutes

 4 *P* (and therefore also log *P*) is a perfectly well-defined thermodynamic quantity. The concept that log *P* is a measure of polarity is not a part of thermodynamics, however, and since this concept, and others like it, lie outside of thermodynamics, it is said to be *extrathermodynamic*. Appendix C is a brief introduction to extrathermodynamic arguments and relationships.

Example 7.1. 170.0 mg of benzylpenicillin (MW 334.4) was shaken with 10.0 mL of 1-octanol and 25.0 mL of water. After the phases separated, the aqueous phase was analyzed and found to contain 7.20×10^{-4} M benzylpenicillin. Calculate the partition coefficient of benzylpenicillin in this system.

The total number of moles of benzylpenicillin is $n_{\text{total}} = w/M$, where w is the weight in grams and M is the molecular weight. Obviously, n_{total} is the sum of the amounts in the octanol and aqueous phases, or

$$n_{\rm oct} + n_{\rm aq} = n_{\rm total}$$

We also have the partition coefficient definition,

$$P = \frac{c_{\rm oct}}{c_{\rm aq}}$$

and the concentrations (in $mol L^{-1}$) are given by

$$c_{
m oct} = rac{n_{
m oct}}{V_{
m oct}}, \quad c_{
m aq} = rac{n_{
m aq}}{V_{
m aq}}$$

where the volumes are in liters. These equations suffice to solve the problem. We find $n_{\text{total}} = 0.170/334.4 = 5.08 \times 10^{-4}$ mol. Then, from the definition of c_{aq} , we obtain

$$n_{aq} = c_{aq} V_{aq}$$

= (7.20 × 10⁻⁴ mol L⁻¹)(0.025 L)
= 0.18 × 10⁻⁴ mol

It follows that $n_{oct} = n_{total} - n_{aq}$, or $n_{oct} = 5.08 \times 10^{-4} - 0.18 \times 10^{-4} = 4.90 \times 0^{-4}$ mol, and therefore that

$$c_{\text{oct}} = \frac{4.90 \times 10^{-4} \text{ mol}}{0.01 \text{ L}}$$
$$= 4.90 \times 10^{-2} \text{ mol } \text{L}^{-1}$$

Finally

$$P = \frac{c_{\text{oct}}}{c_{\text{aq}}} = \frac{4.90 \times 10^{-2}}{7.20 \times 10^{-4}}$$
$$= 68.1$$

or $\log P = 1.83$.

Example 7.2. Log *P* (octanol/water) of caffeine is -0.07 at 25°C. Calculate the standard free-energy change for the partitioning process.

From Eq. (7.17), we have

$$\Delta \mu^{0} = -2.303RT \log P$$

= (-2.303)(1.987 cal mol⁻¹ K⁻¹)(298.15 K)(-0.07)
= 96 cal mol⁻¹ = 400 J mol⁻¹

The interpretation of $\Delta \mu^0$ is that it is the free-energy change when one mole of caffeine in its standard state in water is transferred to its standard state in octanol. (This quantity is sometimes called the *transfer free energy*.)

From the log *P* value we find P = 0.85. Caffeine partitions nearly equally between the octanol and water phases, with a very slight preference for the water.

Solvent Extraction. Partitioning of a solute between immiscible phases is a valuable analytical technique, and it forms the basis of some chromatographic separation methods. In the simplest case we have the type of system described in the preceding discussion. Let p be the fraction of solute present in the upper phase and q the fraction in the lower phase, so p + q = 1. This quantity p is defined as

$$p = \frac{\text{amount of solute in upper phase}}{\text{total amount of solute}}$$
(7.20)

If $c_{\rm U}$ and $c_{\rm L}$ are the concentrations and $V_{\rm U}$ and $V_{\rm L}$ are the volumes of the upper and lower phases, then

$$p = \frac{c_{\rm U} V_{\rm U}}{c_{\rm U} V_{\rm U} + c_{\rm L} V_{\rm L}} \tag{7.21}$$

Let us define the ratio of phase volumes as

$$R = \frac{V_{\rm U}}{V_{\rm L}} \tag{7.22}$$

and of course $P = c_U/c_L$ from Eq. (7.18). Combining these relationships gives

$$p = \frac{PR}{PR+1} \tag{7.23}$$

and so

$$q = \frac{1}{PR+1} \tag{7.24}$$

n	Fraction of Total Extracted in <i>n</i> th Extraction	Total Fraction Extracted	Fraction Remaining
1	р	р	1 - p = q
2	pq	p + pq	$1 - (p + pq) = q^2$
3	pq^2	$p + pq \ p + pq + pq^2$	$1 - (p + pq) = q^{2}$ 1 - (p + pq + pq^{2}) = q^{3}
n	$pq^{(n-1)}$	$\sum_{n=l}^{n} pq^{(n-1)}$	q^n

Table 7.2. Calculation of the progress of extraction

Note that the product *PR* is equal to the ratio (amount in upper phase)/(amount in lower phase); this quantity is called the *capacity factor*. Equation (7.23) gives the fraction of solute extracted into the upper phase, and 100*p* is the percent extracted.

Example 7.3. Log P (octanol/water) = 0.70 for ethyl acetate. If 10.0 mL of an aqueous solution of ethyl acetate is extracted with one 25.0 mL portion of octanol, what percentage of the ethyl acetate will be extracted into the octanol layer?

Since log P = 0.70, P = 5.0. We also have R = 2.50. Applying Eq. (7.23) gives p = 0.926, so 92.6% will be found in the octanol.

Unless *P* is very large or very small, a significant fraction of solute will be found in both phases after a single extraction, as seen in Example 7.3. If the experimental goal is to remove essentially all the solute from one phase into the other, common practice is to reextract with fresh portions of the extracting solvent, pooling the extracts, until the solute has been quantitatively removed. We can calculate the number of extractions required to extract any specified fraction of solute.

As earlier, p is the fraction of solute extracted into the upper phase in a single extraction, and q is the fraction in the lower phase. The first line in Table 7.2. shows the state of the extraction after the first extraction.

If we accept the assumption that P is a true equilibrium constant, so that P has the same value irrespective of the absolute concentrations,⁵ then the same fraction p of solute remaining in the lower phase will be extracted into the upper phase each time. (We assume that identical volumes of fresh upper phase are used in each extraction.) Then the fraction of *total* solute removed in the *n*th extraction is equal to the product of the fraction remaining and the fraction extracted in a single extraction:

Fraction of total extracted in *n*th extraction = fraction of total left after

$$(n-1)$$
th extraction $\times p$ (7.25)

⁵ From Eq. (7.18), $c_U = Pc_L$, which states that a plot of c_U versus c_L should be linear if *P* is a constant independent of concentration. This plot is called a *partition or distribution isotherm*. A linear partition isotherm shows that *P* is independent of concentration.

Applying this equation to the second extraction gives

Fraction of total extracted in 2nd extraction = pq

This is entered on the second line of Table 7.2. The total fraction extracted is now equal to the sum of the fractions extracted in the first and second extractions, which is p + pq, and the fraction remaining is 1 - (total fraction extracted), which is equal to q^2 as seen in Table 7.2. In this way Table 7.2 is completed.

A final convenient expression is obtained by noting that

Total fraction extracted = 1 - Fraction remaining

or, from the final entry in Table 7.2, after n extractions

Total fraction extracted = $1 - q^n$ (7.26)

Example 7.4. For the system described in Example 7.3, calculate the total fraction extracted after 1, 2, 3, 4, and 5 extractions, if R = 1.0.

Since P = 5.0 and R = 1.0, we find with Eq. (7.24) that $q = \frac{1}{6} = 0.167$. Applying Eq. (7.26) gives the results in Table 7.3.

Observe the asymptotic approach to complete extraction, which in principle can never be achieved because, in the terms of classical thermodynamics, at equilibrium (which is reached at each stage of the extraction process) the chemical potential of the solute must be identical in both phases, so the solute cannot be absent from one phase and present in the other. In practice, of course, we can often carry out the extraction to an extent that is practically indistinguishable from completion.

Example 7.5. Using the same system of Examples 7.3 and 7.4, for which P = 5.0, compare the efficiency of extraction of a 15-mL aqueous solution of ethyl acetate with (a) one 60-mL portion of octanol and (b) four 15-mL portions of octanol.

- (a) With Eq. (7.23) and the quantities P = 5.0, R = 4.0, we find p = 0.952, or 95.2% extracted in this experiment.
- (b) With Eq. (7.26) and the quantities p = 5.0, R = 1.0, n = 4 we find q = 0.0476 and total fraction extracted = 0.9992, or 99.92% extracted in this experimental design.

Number of Extractions, <i>n</i>	Total Extracted (%)
1	83.33
2	97.21
3	99.53
4	99.92
5	99.99

Table 7.3. Multiple extractions of a solute with P = 5 and R = 1

Example 7.4 demonstrates an important result of extraction theory. A more efficient extraction is achieved with several extractions than with a single extraction, even when the same volume of extracting solvent is employed in the different operations.

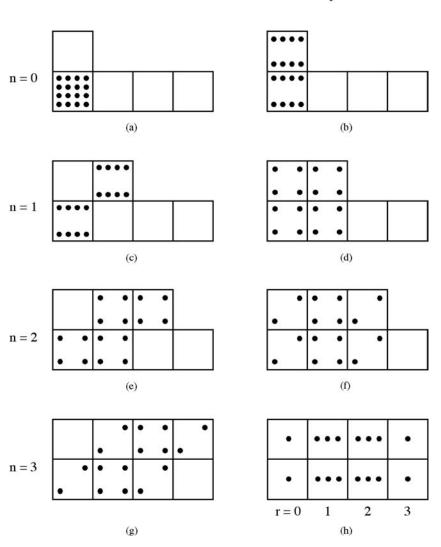
Countercurrent Distribution. Although a single solute can be exhaustively extracted from solution by means of multiple extractions, it is not possible to separate two solutes (leaving one in each phase) by this technique unless the partition coefficient of one of them is effectively zero or infinite. An alternative experimental design, called countercurrent distribution (CCD), has been invented to allow solutes having similar (yet quantitatively different) partition coefficients to be separated. The term "countercurrent" means that the two phases move in opposite directions, although actually one phase is held motionless and the other moves, so the phases are in relative motion. Although CCD as a separation technique has been superseded by chromatography, a description is worthwhile for two reasons: (1) Since thermodynamic equilibrium can be achieved at each stage of the process, an exact mathematical analysis is possible, and the mathematics turn out to be of a much wider applicability; and (2) CCD constitutes an excellent introduction to the technique of partition chromatography, which in fact was initially developed as a modification of CCD [the present treatment of CCD draws heavily on earlier work (Connors 1982, pp. 357-364)].

The countercurrent distribution experiment uses a train of tubes within which the individual equilibrations occur. At the beginning of the experiment each tube is charged with an identical volume of the lower phase (e.g., water or an aqueous buffer). These tubes are numbered 0, 1, 2, ..., r. Into tube 0 a suitable volume of the upper-phase solvent (e.g., ether) is introduced. The solute is added to tube 0; it is immaterial whether the solute is added in the upper or the lower phase. Figure 7.5 is a schematic rendering of a countercurrent distribution of a single solute; it is assumed, in this case, that p = q = 0.5. Figure 7.5a represents the train of tubes as it has been described above, with 16 parts of solute added to the lower phase of tube 0. Now the tube is shaken to allow distribution to occur; in Fig. 7.5b the resulting partitioning of the solute is shown as 8 parts in each phase, since p = q for this particular solute.

Next the upper phase of tube 0 is transferred to tube 1 (this is called the *first transfer*) and fresh solvent is added to tube 0 (Fig. 7.5c). The tubes are equilibrated to give the distribution shown in Fig. 7.5d. This sequence is repeated until three transfers have been effected (n = 3), as shown in Fig. 7.5h.

The result of these operations has been to transfer the solute in the direction of motion of the upper phase. This process may be repeated many times. Since only the upper phase is transferred, clearly the solute can progress along the train of tubes only by being extracted into the upper phase. Therefore the greater the value of p, the further along the tube train the solute will progress in a given number of transfers. Actually the solute is distributed over many tubes, as can be seen by the sample shown in Fig. 7.5. If the original sample contains two solutes with different partition coefficients, they will progress along the tubes at different "rates," the substance with the larger

Equilibration



Transfer

Figure 7.5. Schematic representation of countercurrent distribution with three transfers of a solute with p = 0.5. [Reproduced by permission from Connors (1987).]

partition coefficient traveling faster. In order to separate the solutes, it is necessary only to perform enough transfers.

It is possible to predict quantitatively the countercurrent distribution behavior of a solute if its partition coefficient is known for the liquid–liquid system. Since P and R are known quantities, p and q may be calculated.

Suppose that one unit of a single solute is placed in the lower phase of tube 0; the situation may be represented as in the first row of Table 7.4, where, as in the

Transfer Number, <i>n</i>		Tube Number, r				
		0	1	2	3	4
0	\mathbf{B}^{a} \mathbf{A}^{b}	0/1 p/q				
1	В	$\frac{p}{q}$	<i>p</i> /0			
	А	pq/q^2	p^2/pq			
2	В	$0/q^{2}$	pq/pq	$p^{2}/0$		
	А	pq^{2}/q^{3}	$2p^2q/2pq^2$	$p^{3\prime}p^2q$		
3	В	$0/q^{3}$	$pq^{2\prime}2pq^2$	$2p^2q/p^2q$	$p^{3}/0$	
	А	pq^{3}/q^{4}	$3p^2q^{2/}3pq^3$	$3p^3q/3p^2q^2$	$p4/p^3q$	
4	В	$0/q^4$	$pq^{3/}3pq^{3}$	$3p^2q^2/3p^2q^2$	$3p^3q/p^3q$	p ⁴ /0
	ls after four ansfers	q^4	$4pq^3$	$6p^2q^2$	$4p^3q$	p^4

Table 7.4. Calculation of the distribution through four transfers^a

^aBefore equilibration.

^bAfter equilibration.

ⁱSource: Reproduced by permission from Connors (1987).

earlier discussion, the tubes are numbered 0, 1, 2, ..., r and transfers are numbered 0, 1, 2, ..., n. Before equilibration all the solute is in the lower phase, and after equilibration a fraction p of the solute is in the upper phase and q is in the lower phase.

Next the upper phase of tube 0 is transferred to tube 1 (which contains fresh lower phase) and fresh upper phase is placed in tube 0. The phases are equilibrated. The fraction of total solute extracted into the upper phase of tube 0 will be p times the fraction of solute in the tube, or pq. Similarly, the fraction of solute in the lower phase is q times the fraction of solute in the tube, or q^2 . In this way the distribution has been calculated through four transfers, as seen in Table 7.4.

In the last row of the table the total fraction of original solute in each tube is listed. The distribution exhibits a marked symmetry in p and q. Obviously, the calculation of such a distribution for many transfers would be extremely laborious, but it is fortunately not necessary to proceed as in the previous example. It has been observed that the total fraction of original solute in each tube is given by the corresponding term in the binomial expansion, $(q + p)^n$. Two implications of this result are as follows: (1) For n transfers there are n + 1 terms and therefore n + 1 tubes; and (2) the sum of all the terms is 1, since p + q = 1, and 1 to any power is 1.

The expansion of the function $(q + p)^n$ is laborious for large *n*, and an easier calculation is available. The binomial expansion may be written

$$(q+p)^n = q^n + nq^{n-1}p + \frac{n(n-1)}{2}q^{n-2}p^2 + \cdots + p^n$$

which can be expressed

$$(q+p)^{n} = \sum_{r=0}^{n} \frac{n!}{r!(n-r)!} p^{r} q^{(n-r)}$$

where *r* is the number of the corresponding term in the expansion (the quantity *n*! is called "*n* factorial" and means $n! = 1 \times 2 \times 3 \times 4 \times ... n$; the relationship 0! = 1 is a definition). Interpreting this in the context of CCD, we write Eq. (7.27) for the *r*th term in the binomial expansion

$$T_{nr} = \frac{n!}{r!(n-r)!} p^r q^{(n-r)}$$
(7.27)

where the quantity T_{nr} is read "the fraction of total solute contained in both layers of the *r*th tube after *n* transfers." A calculated countercurrent distribution is usually exhibited as a plot of T_{nr} versus *r*. Equation (7.27) is called the *binomial distribution*.

Calculation of the CCD curve may be further simplified. According to statistical theory, the mean of the binomial distribution is equal to np. The mean corresponds to the maximum; therefore the tube number of the maximum in the curve, r_{max} , is given by

$$r_{\max} = np \tag{7.28}$$

This simple expression permits one to calculate the maximum in the CCD curve if p is known. Although n must be an integral number, r_{max} need not be. Note that r_{max} is directly proportional to p. If Eq. (7.27) is written for T_{nr} and for $T_{n(r-1)}$, these expressions can be combined to give

$$\frac{T_{nr}}{T_{n(r-1)}} = \frac{p(n-r+1)}{qr}$$
(7.29)

with which the fraction of solute in any tube can be calculated if the fraction in an adjacent tube is known.

The easiest way to calculate an entire distribution curve with these equations (assuming that *p* is known) is to first find r_{max} with Eq. (7.28). Next calculate T_{nr} with Eq. (7.27) for one tube in the vicinity of r_{max} . Finally, calculate the fractions of solute in all surrounding tubes by means of Eq. (7.29). Figure 7.6 shows the results of such a calculation for a typical separation of two solutes; it was assumed that $P_1 = 0.5$, $P_2 = 2.0$, and R = 1.00 for this system. In Fig. 7.6a the distribution of each solute is shown after four transfers. In an actual experiment the tube contents would be analyzed for total solutes present, and the experimental curve would therefore represent the sum of the fractions of the individual solutes; this curve is shown as the solid line in Fig. 7.6a. Separation is not yet apparent in this curve. The individual distribution curves, however, show that a partial resolution has occurred, with tubes 0 and 1 enriched in solute 1, tubes 3 and 4 enriched in solute 2, and tube 2 containing equal fractions of solutes 1 and 2.

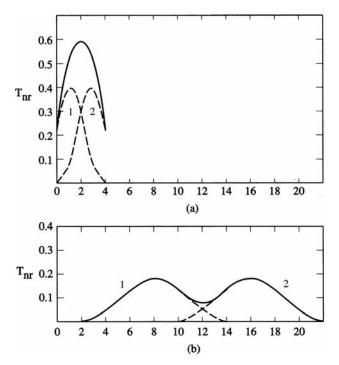


Figure 7.6. Countercurrent distribution of two solutes in a system where $P_1 = 0.5$, $P_2 = 2.0$, R = 1.0: (a) Distribution after 4 transfers; (b) distribution after 24 transfers. The calculated points are connected by smooth curves, although in fact the distribution is discontinuous. [Reproduced by permission from Connors (1987).]

Figure 7.6b shows the same system after 24 transfers. Separation of the solutes is now apparent. Tubes 0–9 contain essentially only solute 1, whereas tubes 15–24 contain only solute 2. Portions of both solutes will be found in tubes 10–14. If the experiment were extended to a larger number of transfers, a complete separation could eventually be achieved. Note, however, that the width of the "zones," or distribution curves, increases as the number of transfers increases.

In a real experimental situation, the quantity plotted on the vertical axis would usually be an analytical quantity, such as weight of solute per tube, rather than the fraction T_{nr} . It may be noted that from such an experimental distribution curve the quantity r_{max} may be read and, by utilizing Eqs. (7.28) and (7.23), the partition coefficient may be estimated.

The countercurrent distribution curve is not symmetric (unless p = q), but as *n* becomes larger, the curve approaches very closely a symmetric distribution.

The binomial distribution is a mathematical function that yields the probability of "success" in what are known as *Bernoulli trials*. These are events, such as coin tosses, in which there are only two possible outcomes (heads or tails). The analogy to CCD is that a molecule of solute has only two possible choices: It must take up residence in either the upper phase or the lower phase. As the number of transfers becomes very

large, the discontinuous (i.e., stepwise) binomial distribution approaches closely a continuous function, the normal distribution, which provides for a faster means of calculating the CCD curve (Connors 1982, pp. 357–364).

PROBLEMS

- **7.1.** Calculate the ideal entropy of mixing and free energy of mixing when 10.0 g of benzene and 15.0 g of toluene are mixed at 25° C.
- **7.2.** For alanine, $\log P$ (octanol/water) = -2.94. For phenothiazine, $\log P = 4.15$. Calculate the standard free energy changes for these phase transfer processes at 25°C.
- **7.3.** These are experimental partial pressures of benzene (B) and toluene (T) over their solutions at 20°C:

$x_{\rm B}$	$P_{\rm B} ({\rm mmHg})$	P_T (mmHg)
0.00	0	22
0.27	18	17
0.44	34	12
0.55	41	11
0.67	49	8
1.00	75	0

Confirm the validity of Raoult's law for this system by plotting the data. By calculation, determine the solution composition at which the partial pressures of benzene and toluene are equal, and check your result on the graph you have plotted. What is the total vapor pressure over the solution at this composition?

7.4. These are partial pressures of chloroform over chloroform–acetone solutions at $35 \,^{\circ}\text{C}$:

<i>x</i> (CHC1 ₃)	P (mmHg)	
0.0	0	
0.2	34	
0.4	82	
0.6	148	
0.8	225	
1.0	293	

Plot the data, confirming the asymptotic approach to Raoult's law in the nearly pure chloroform, and the nonideal behavior in dilute solutions of chloroform. Estimate the Henry's law constant.

- **7.5.** How many extractions are necessary to remove 99.9% of a drug from 30 mL of an aqueous solution if it is extracted with 20 mL portions of ether and log *P* (ether/water) = 0.54?
- **7.6.** (a) Consider the distribution of neutral weak acid HA between an organic phase and an aqueous phase. Define the true partition coefficient as $P = [HA]_{org}/[HA]_{aq}$. Presuming that the anion does not detectably partition into the organic phase, derive this relationship between the true partition coefficient *P* and the apparent partition coefficient *P*_{app}:

$$P_{\rm app} = \frac{P[H^+]}{[H^+] + K_a}$$

where K_a is the acid dissociation constant of HA, [H⁺] is the aqueous phase concentration of hydrogen ion, and P_{app} is the ratio of total concentrations of solute in the organic and aqueous phases.

- (b) Show how P_{app} is related to *P* at the limits of very low and very high hydrogen ion concentration. How are *P* and P_{app} related when $pH = pK_a$?
- (c) Calculate P_{app} as a function of pH for an acid having $pK_a = 4.0$ and for which log P = 1.00. Plot P_{app} against pH.
- **7.7.** Consider a system consisting of a single solute partitioned among three mutually immiscible phases A, B, and C, the system being at equilibrium.
 - (a) Define the three partition coefficients.
 - (b) Derive an equation relating one of the partition coefficients to the other two.
 - (c) Derive an equation relating the fraction of solute in phase A to the partition coefficients and the volumes of the two phases.
- **7.8.** A mixture of three compounds was subjected to countercurrent distribution. After 150 transfers, with each tube containing 5 mL of water and 5mL of ether, the maxima in the CCD curve appeared at tubes 30, 75, and 120. Calculate the partition coefficients of the three compounds.
- **7.9.** Ordinary water contains small amounts of dissolved oxygen. What is the unit of the ratio between the oxygen Henry's constant and the partial pressure of the water vapor in equilibrium with the solution?
- **7.10.** Calculate the ideal entropy and free energy of mixing when 36 g of water (MW = 18 g/mol) are mixed with 230 g of ethanol (MW = 46 g/mol) at 25 °C. Express the results in cal/mol K (ΔS) and cal/mol ((ΔG).

8

SOLUTIONS OF ELECTROLYTES

8.1. COULOMBIC INTERACTION AND IONIC DISSOCIATION

An electrolyte is a substance that produces ions. Since the ions are charged species, the force of interaction between them is a convenient starting point for our discussion. The force of interaction between two particles having charges Q_1 and Q_2 , separated by distance *r*, is given by *Coulomb's law*:

$$F = \frac{Q_1 Q_2}{4\pi \varepsilon r^2} \tag{8.1}$$

where ε is a property of the medium, to be dealt with shortly. The potential energy of interaction, *V*, is equal to the product of force and distance. The Coulombic potential energy is therefore

$$V = \frac{Q_1 Q_2}{4\pi\varepsilon r} \tag{8.2}$$

Since the charge on an ion can be written as the product of its valence z (including its sign) and the electronic charge e ($e = 1.602 \times 10^{-19}$ C), Eq. (8.2), for our purposes, is equivalent to

$$V = \frac{z_1 z_2 e^2}{4\pi\varepsilon r} \tag{8.3}$$

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For two ions of like charge, V and F are both positive and the force is repulsive, whereas if the ions are of unlike charge, V and F are negative and are attractive. The zero of potential energy is taken to be when the ions are separated to infinity $(r = \infty)$.

The quantity ε is called the permittivity, and it is best introduced through the expression

$$\varepsilon_r = \frac{\varepsilon}{\varepsilon_0} \tag{8.4}$$

where ε_0 is the *permittivity of the vacuum* and ε_r is called the *relative permittivity*. Chemists, however, have traditionally referred to ε_r as the *dielectric constant*. The relative permittivity or dielectric constant is measured as the electrical capacitance of the medium (solvent) relative to the capacitance of the vacuum. It follows that ε_r is a dimensionless number greater than one. Equation (8.3) is often written in the form

$$V = \frac{z_1 z_2 e^2}{4\pi\varepsilon_0 \varepsilon_r r} \tag{8.5}$$

The permittivity of the vacuum ε_0 has the value $8.854 \times 10^{-12} \text{ C}^2 \text{J}^{-1} \text{ m}^{-1}$. Table 8.1 gives some dielectric constant values.

Example 8.1. Calculate the energy of the Coulombic interaction between a sodium ion and a chloride ion, at contact distance, in vacuum and in water.

The ionic radii of Na⁺ and of Cl⁻ (available in reference handbooks) are 0.95 and 1.81 Å, respectively, equivalent to an internuclear distance of $r = 2.76 \times 10^{-10}$ m. In vacuum

$$V(\text{vacuum}) = -\frac{(1.602 \times 10^{-19} \text{ C})^2}{4\pi (8.854 \times 10^{-12} \text{ C}^2 \text{ J}^{-1} \text{ m}^{-1})(1)(2.76 \times 10^{-10} \text{ m})}$$

= -8.36 × 10⁻¹⁹ J

Solvent	ε _r	Solvent	ε_r
<i>n</i> -Hexane	1.89	Methanol	32.6
Cyclohexane	2.02	Nitrobenzene	35
1,4-Dioxane	2.21	Acetonitrile	36.2
Benzene	2.28	N,N-Dimethylformamide	36.7
Diethyl ether	4.34	Ethylene glycol	37.7
Ethyl acetate	6.02	N,N-Dimethylacetamide	37.8
Acetic acid	6.19	Glycerol	42.5
n-Butyl alcohol	17.1	Dimethyl sulfoxide	49
<i>i</i> -Propyl alcohol	17.7	Formic acid	58
Acetone	20.7	Water	78.5
Ethanol	24.3	Formamide	110

Table 8.1. Dielectric constants of some solvents

This is the energy of interaction between one Na^+ and one $C1^-$. If we multiply by Avogadro's number to find the energy per mole of sodium chloride, we get

$$V(\text{vacuum}) = -504 \text{ kJ mol}^{-1}$$
$$= -120 \text{ kcal mol}^{-1}$$

a very strong interaction. In water $\varepsilon_r = 78.5$, and the calculation gives

$$-0.106 \times 10^{-19}$$
 J per ion pair = -6.42 kJ mol⁻¹
V(water) = -1.53 kcal mol⁻¹

Example 8.1 shows that the dielectric constant of the medium markedly influences the strength of the interionic interaction energy. The dielectric constant is a measure of the ability of the medium to separate charges of unlike sign. (Not coincidentally, the dielectric constant roughly parallels our chemical notion of solvent polarity and is often taken as a quantitative measure of polarity.) The larger the dielectric constant, the more easily two unlike charges can be separated. The high dielectric constant of water is a manifestation of the very unusual nature of water as a solvent. In fact, the classification of electrolytes into the categories of strong electrolytes (i.e., essentially completely dissociated into ions in solution) and weak electrolytes (incompletely dissociated) is based on the use of water as the solvent. Substances that are strong electrolytes in water act as weak electrolytes in low dielectric constant solvents.¹ Let us pursue this issue by writing Eq. (8.6) for an electrolyte, schematically denoted AB, when dissolved in a solvent:

$$AB \stackrel{\text{ionization}}{=\!\!=\!\!=\!\!=\!\!=\!\!=\!\!=\!\!=\!\!=\!\!=\!\!A^+ B^- \stackrel{\text{dissociation}}{=\!\!=\!\!=\!\!=\!\!=\!\!=\!\!A^+ + B^-$$
(8.6)

Ionization is the production of ions,² and dissociation is the separation of species (whether ionic or uncharged). The extent of ionic dissociation is reasonably described by Coulomb's law. This is why we do not distinguish between ionization and dissociation for aqueous solutions; because water's dielectric constant is quite large, the force between ions is relatively small, and as soon as ions form, they dissociate. Ion

¹ The dielectric constant is a bulk property of matter, and its incorporation into Coulomb's law means that we are treating the solvent as a continuum; that is, the molecular (particulate) nature of the solvent is ignored in this treatment.

² The molecular interpretation of the ionization process may be complex and will depend on the molecular identity. One possibility is that two kinds of ion pairs may form. One of these, represented A^+B^- , is an *intimate ion pair*; the other, shown as A^+SB^- , where S is a molecule of solvent, is a *solvent-separated ion pair*.

pairs (the species $A^+ B^-$) are seldom detectable in water. But in solvents of low dielectric constant (typically with ε_r values less than ~25), the extent of dissociation is reduced, as may be demonstrated by repeating Example 8.1 with some different ε_r values. Glacial acetic acid (the term "glacial" simply means essentially pure in this context) is an important analytical solvent that has been carefully studied. Because of its low dielectric constant, ionpairs can be detected in acetic acid solutions. Letting HOAc represent acetic acid (since acetic acid is CH₃COOH, the symbol Ac represents the acetyl group CH₃CO), a solute acid HX reacts according to

$$HX + HOAc \rightleftharpoons H_2OAc^+X^- \rightleftharpoons H_2OAc^+ + X^-$$

In this scheme, HOAc is acting to solvate the hydrogen ion, and H_2OAc^+ in acetic acid is analogous to H_3O^+ in water. For convenience we usually omit the solvent, writing simply

$$HX \rightleftharpoons H^+ X^- \rightleftharpoons H^+ + X^- \tag{8.7}$$

Now, in the conventional manner we define an ionization constant K_i , and a dissociation constant K_d as follows, using Eq. (8.7) as the defining reaction.

$$K_i = \frac{[\mathrm{H}^+\mathrm{X}^-]}{[\mathrm{H}\mathrm{X}]} \tag{8.8}$$

$$K_d = \frac{[\mathrm{H}^+][\mathrm{X}^-]}{[\mathrm{H}^+\mathrm{X}^-]}$$
(8.9)

Next we define an overall dissociation constant K_{HX} ; we place all dissociated species in the numerator and all undissociated species in the denominator:

$$K_{\rm HX} = \frac{[{\rm H}^+][{\rm X}^-]}{c_{\rm HX}}$$
(8.10)

where $c_{\text{HX}} = [\text{HX}] + [\text{H}^+\text{X}^-]$. In these equations brackets signify molar concentrations. Combining Eqs. (8.8)–(8.10) gives

$$K_{\rm HX} = \frac{K_i K_d}{1 + K_i} \tag{8.11}$$

Similar equations can be written for bases and for salts. One of the consequences is that the pH, which in water is the controlling factor in acid–base equilibria, does not play a comparable role in glacial acetic acid. This is because very little of the acidic species is present as dissociated H^+ ; most of the acid is in the undissociated form c_{HX} .

8.2. MEAN IONIC ACTIVITY AND ACTIVITY COEFFICIENT

Let us now consider a strong electrolyte, such as a salt in aqueous solution. The solute is completely dissociated into its constituent ions according to

$$\mathbf{M}_{p}\mathbf{X}_{q} \rightleftharpoons p\mathbf{M}^{q+} + q\mathbf{X}^{p-} \tag{8.12}$$

where p and q denote the number of positive and negative ions, respectively, generated by one molecule of the salt. The following development is motivated by the impossibility of separately varying and studying the cations and the anions; electroneutrality dictates that only their combination in the ratio p/q can be manipulated.

We will adopt the infinite dilution Henry's law reference state in the molar concentration scale for all species. Then we can write for the cation and the anion

$$\mu_{+} = \mu_{+}^{0} + RT \ln a_{+} \tag{8.13a}$$

$$\mu_{-} = \mu_{-}^{0} + RT \ln a_{-} \tag{8.13a}$$

and for the solute as a whole

$$\mu_2 = \mu_2^0 + RT \ln a_2 \tag{8.14}$$

Now we postulate (assuming complete dissociation)

$$\mu_2 = p\mu_+ + q\mu_- \tag{8.15}$$

and analogously

$$\mu_2^0 = p\mu_+^0 + q\mu_-^0 \tag{8.16}$$

Simple algebraic combination of Eqs. (8.13)-(8.16) yields

$$a_2 = a_+^p a_-^q \tag{8.17}$$

We define v as the number of ions generated by one molecule of solute, so

$$v = p + q \tag{8.18}$$

It is now conventional to define the *mean ionic activity* a_{\pm} by $a_{\pm}^{\nu} = a_2$, giving the following, from Eq. (8.17):

$$a_{\pm}^{\nu} = a_{\pm}^{p} a_{-}^{q} \tag{8.19}$$

A self-consistent set of relations is obtained by making these further definitions; the *mean ionic activity coefficient* is

$$\gamma^{\nu}_{\pm} = \gamma^{p}_{+} \gamma^{q}_{-} \tag{8.20}$$

and the mean ionic molarity is

$$c_{\pm}^{\nu} = c_{+}^{p} c_{-}^{q} \tag{8.21}$$

so that we can write

$$a_{\pm} = \gamma_{\pm} c_{\pm} \tag{8.22}$$

The significance of these relationships is easiest to comprehend for the simplest case of a 1:1 electrolyte such as NaCl. For this case p = 1, q = 1, v = 2, and we write from the foregoing

$$a_{\pm}^2 = a_{\pm}a_{-} \tag{8.23a}$$

$$\gamma_{\pm}^2 = \gamma_+ \gamma_- \tag{8.23b}$$

$$c_{\pm}^2 = c_{+}c_{-} \tag{8.23c}$$

Although we separately know c_+ and c_- from c_2 , the solute concentration, we cannot separately determine y_+, y_-, a_+ , and a_- . The effect of the definitions given above is to assign the extent of nonideality equally (when p = q) to the cation and the anion.

Example 8.2

- (a) What is the mean ionic molarity of an aqueous solution 0.15 M in sodium chloride? Since sodium chloride is completely dissociated, $c_{+} = 0.15$ M and $c_{-} = 0.15$ M, giving, from Eq. (8.23c), $c_{\pm} = 0.15$ M.
- (b) What is the mean ionic molarity of an aqueous solution 0.25 M in K₂SO₄? For this system p = 2, q = 1, v = 3. The concentration of potassium ions, c_+ , is 0.50 M and c_- , the concentration of sulfate ions, is 0.25 M. From Eq. (8.21), we have

$$c_{\pm}^{3} = (0.50)^{2}(0.25)$$

 $c_{\pm} = 0.397M$

8.3. THE DEBYE-HÜCKEL THEORY

In an infinitely dilute solution each solute ion is resident in an environment that consists effectively only of the solvent (which we continue to treat as a continuum).

In such a situation the ion is free to exert whatever effects are characteristic of its identity, unperturbed by other solute species; it is in its Henry's law reference state, and it behaves ideally.

If the ionic concentration of the solution is raised, either by increasing the concentration of the solute of interest or by adding ions of a different electrolyte solute, the environment of our ion changes. As the ionic concentrations increase, the distance between ions decreases, and the Coulombic interaction energies come into play. Ions of like charge tend to repel each other, and ions of unlike charge attract each other. The consequence of these interactions is that instead of a random distribution of ions throughout the solution, an *ionic atmosphere* develops such that the volume centered on a cation possesses a net negative charge, whereas the volume centered on an anion possesses a net positive charge (of course, the solution as a whole is electrically neutral). These charge distributions, constituting perturbations of the infinite dilution environment, are manifested in solute behavior that we interpret as nonideal and that we measure in terms of a mean ionic activity coefficient.

In 1923 Debye and Hückel developed a quantitative theory of this ionic atmosphere effect. Although the Debye–Hückel theory is not itself part of thermodynamics, its final result has been absorbed into thermodynamics, and it is routinely used to interpret and to predict nonideal behavior in electrolyte solutions. The *Debye–Hückel equation* is written

$$\operatorname{Log} \gamma_{\pm} = -\frac{A|z_{\pm}z_{-}|\sqrt{I}}{1+aB\sqrt{I}}$$
(8.24)

where *A* and *B* are constants whose values depend on the dielectric constant and the temperature, and *a* is closely related to an ionic radius. The quantity *I* is the *ionic strength* and is defined by Eq. (8.25), where c_i is the molar concentration of ion *i* and *z* is its charge.

$$I = \frac{1}{2} \sum c_i z_i^2 \tag{8.25}$$

In Eq. (8.24), z_+ and z_- are the (absolute values of the) charges on the electrolyte of interest; in Eq. (8.25), the c_i and z_i include *all* the ions in the solution.

For aqueous solutions at 25°C, Eq. (8.24) takes the specific form

$$\log \gamma_{\pm} = -\frac{0.509|z_{\pm}z_{-}|\sqrt{I}}{1+\sqrt{I}}$$
(8.26)

and at very low ionic strengths Eq. (8.26) approaches Eq. (8.27), which is known as the *Debye–Hückel limiting law:*

$$\log \gamma_{\pm} = -0.509 |z_{+}z_{-}| \sqrt{I}$$
(8.27)

Example 8.3. What is the ionic strength of (a) a solution 0.10 M in NaCl and 0.05 M in HCl; (b) a solution 0.25 M in K₂SO₄?

(a) $c_{\text{Na}^+} = 0.10 \text{ M}$, $c_{\text{H}^+} = 0.05 \text{ M}$, $c_{\text{Cl}^-} = 0.15 \text{ M}$; $z_i^2 = 1$ for all ions. From Eq. (8.25), we have

$$I = \frac{1}{2}(0.10 + 0.05 + 0.15) = 0.15 \,\mathrm{M}$$

The ionic strength of a solution of 1:1 electrolytes is equal to the total solute concentration.

(b)
$$c_{K^+} = 0.50 \text{ M}, \qquad z_{K^+} = +1, \qquad c_{SO_4^{2^-}} = 0.25 \text{ M}, \qquad z_{SO_4^{2^-}} = -2$$

$$I = \frac{1}{2} (0.50 \times 1 + 0.25 \times 4) = 0.75 \text{ M}$$

The ionic strength of a solution containing polyvalent ions reflects the dominant effect of the square of the charge on the ionic atmosphere. Notice that the concentrations of H^+ and OH^- arising from the dissociation of water are not included in the calculation because they make a negligible contribution to the ionic strength.

Example 8.4. Calculate the mean ionic activity coefficient of a 1:1 electrolyte at concentrations of 0.001, 0.010, and 0.10 mol L^{-1} , in water at 25°C. Use the Debye–Hückel equation in the form of Eq. (8.26), and also use the limiting law, Eq. (8.27).

From the given data, $z_+ = +1$, $z_- = -1$, so $|z_+z_-| = 1$, and I = c, the molar concentration. These results are found:

	γ	±
<i>c</i> (M)	Eq. (8.26)	Eq. (8.27)
0.001	0.965	0.964
0.010	0.899	0.889
0.100	0.756	0.690

The results in Example 8.4 show that the limiting law and the full Debye–Hückel equation agree closely in extremely dilute solution, but they begin to differ significantly in the concentration region of ~0.01 M (i.e., when $\sqrt{I} = 0.1$). Above this ionic strength Eq. (8.26) is necessary, but even this equation fails to agree closely with experimental results at ionic strengths above about 0.05 M, where effects specific to each electrolyte are observed. Table 8.2 lists some experimentally determined mean ionic activity coefficients.³ We expect, from the appearance of the product $|z_+z_-|$ in the Debye–Hückel equation, that different charge types of electrolytes will behave

³ See Glasstone (1947, p. 402). These activity coefficients can be measured in various ways. One approach is to measure the deviation from ideality of the *solvent* and to relate this to the nonideality of the solute.

		·	γ_{\pm}	
т	HCl	NaCl	CaCl ₂	ZnSO ₄
0.001	0.966	0.966	0.888	0.734
0.005	0.928	0.929	0.789	0.477
0.01	0.905	0.904	0.732	0.387
0.02	0.875	0.875	0.669	0.298
0.05	0.830	0.823	0.584	0.202
0.1	0.796	0.778	0.531	0.148
0.2	0.767	0.732	0.482	0.104
0.5	0.757	0.679	0.457	0.063
1.0	0.809	0.656	0.509	0.044
2.0	1.009	0.670	0.807	0.35
3.0	1.316	0.719	1.55	0.041

Table 8.2. Mean ionic activity coefficients in water at 25°C

Source: Data from Glasstone (1947, p. 402).

differently, and this is seen. However, it is also observed that electrolytes of the same charge type display behavior characteristic of the individual electrolyte; compare HCl and NaCl in Table 8.2. Empirical extensions of the Debye–Hückel equation have been proposed of the form (at 25° C in water)

$$\log \gamma_{\pm} = -\frac{0.509|z_{\pm}z_{-}|\sqrt{I}}{1 + aB\sqrt{I}} + CI$$
(8.28)

where the parameters aB and C are chosen to best fit the experimental data.

Figure 8.1 is a plot of the data from Table 8.2 in a format consistent with the manner in which the Debye–Hückel equation is written, that is, as a plot of $\log \gamma_+$

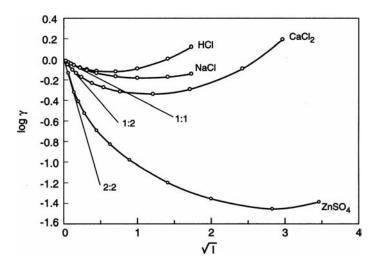


Figure 8.1. Plot of data in Table 8.2. Limiting law slopes are drawn for 1:1, 1:2, and 2:2 electrolytes.

against \sqrt{I} . Figure 8.1 shows several interesting features. The individual character of the nonideal behavior is clearly evident in the curves for HCl and NaCl. The minima observed in these curves are not predicted by the Debye–Hückel equation, and in some instances the mean ionic activity coefficients rise to values greater than unity. From the limiting law, Eq. (8.27), we can predict the slope of the plot for each charge type of electrolyte at infinite dilution, and these slopes are drawn in Fig. 8.1. The Debye–Hückel limiting law gives a satisfactory account of nonideal electrolyte behavior in very dilute solutions.

The Debye–Hückel theory finds very practical application in obtaining thermodynamic acid dissociation constants for weak acids and bases. An apparent constant is measured experimentally at, necessarily, finite ionic strength, and the theory is used to correct the value to zero ionic strength. This calculation is described in Chapter 13. One powerful consequence of the Debye–Hückel theory is that it provides a firm theoretical basis for the extrapolation of electrolyte experimental data to infinite dilution; the appropriate independent variable is the square root of the ionic strength.

PROBLEMS

- **8.1.** Write the reactions for ionization and dissociation of a base B in glacial acetic acid.
- **8.2.** Calculate the ionic strengths of these three solutions (from Table 8.2): 3.0 m NaCl; 3.0 m CaCl₂; 3.0 m ZnSO₄.
- **8.3.** Calculate the mean ionic activity coefficient of 0.05 m NaCl in water at 25°C, and compare your result with the experimental value in Table 8.2.
- **8.4.** Estimate the mean ionic activity of 0.001 M HCl in an aqueous solution containing 0.025 M KCl at 25°C.
- **8.5.** Calculate the ionic strength of a solution containing $0.10 \text{ M} \text{ Na}_3 \text{PO}_4$ and 0.05 M KBr.
- **8.6.** Obtain an estimate of the parameter C in Eq. (8.28) for CaCl₂ by use of the data in Fig. 8.1. [*Hint:* find the derivative $d \log \gamma_{\pm}/d\sqrt{I}$ of Eq. (8.28), set equal to zero (at the minimum), and solve for C.]

9

COLLIGATIVE PROPERTIES

Several properties of solutions depend (mainly) only on the *number* of solute particles (molecules or ions) and not on their identity. These are called the *colligative properties*. They are pharmaceutically relevant.

9.1. BOILING POINT ELEVATION

The boiling point of a solution of a nonvolatile solute is higher than the boiling point of the pure solvent. This observation is readily explicable on the following basis. The normal boiling point T_b of the solvent is the temperature at which its vapor pressure is equal to 1 atm. When a solute is incorporated into the solvent, according to Raoult's law the vapor pressure over the solution is $p_1 = x_1 P_1^*$ [Eq. (7.1)]. (We will use subscript 1 to designate the solvent and 2 for the solute.) Since $x_1 + x_2 = 1$, an increase in x_2 results in a decrease in x_1 and therefore a decrease in p_1 , at a given temperature. In order to cause the solvent to boil, it is now necessary to raise the temperature until p_1 becomes 1 atm. This phenomenon is known as the *boiling point elevation*, and it is seen to be a consequence of the vapor pressure lowering by the presence of solute particles. As ordinarily discussed, the boiling point elevation is treated as a phenomenon of nonelectrolyte solutions, but solutions of electrolytes show the same effect. It is necessary to keep in mind that the number of solute particles (i.e., their concentration) is the controlling factor, and if the solute is an electrolyte, the number of particles depends on the charge type and the extent of dissociation. For example, in a 0.10 M aqueous solution of NaCl the effective concentration, as concerns the colligative properties, is 0.20 M.

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A thermodynamic description of the boiling point elevation effect can be achieved by the application of concepts that we have already developed. At equilibrium (i.e., at the boiling point), the chemical potential of the solvent is equal in the vapor and liquid phases:

$$\boldsymbol{\mu}_1(\mathbf{g}) = \boldsymbol{\mu}_1(\mathbf{l}) \tag{9.1}$$

By restricting attention to dilute solutions, we can treat the solution as an ideal solution, writing

$$\mu_1(1) = \mu_1^0(1) + RT \ln x_1 \tag{9.2}$$

$$\mu_1(g) = \mu_1^0(g) + RT \ln p_1 \tag{9.3}$$

Setting these equal, noting that $p_1 = 1$ atm at the boiling point, and writing $\Delta G_{\text{vap}}^0 = \mu_1^0(g) - \mu_1^0(l)$ gives

$$\Delta G_{\rm vap}^0 = RT \ln x_1 \tag{9.4}$$

Putting Eq. (9.4) into the form $\Delta G_{\text{vap}}^0/T = R \ln x_1$ and applying the Gibbs–Helmholtz equation [Eq. (3.18)] leads to

$$\frac{d\ln x_1}{dT} = -\frac{\Delta H_{\rm vap}^0}{RT^2} \tag{9.5}$$

where constant pressure is understood.¹

When $x_1 = 1$, we obtain $T = T_b$ (the normal boiling point). Equation (9.5) is integrated between the limits shown:

$$\int_{1}^{x_1} d\ln x_1 = -\frac{\Delta H_{\mathrm{vap}}^0}{R} \int_{T_b}^T \frac{dT}{T^2}$$

The result is

$$\ln x_1 = -\frac{\Delta H_{\rm vap}^0}{R} \left(\frac{1}{T_b} - \frac{1}{T}\right)$$

¹ $\Delta G_{\text{vap}} = \mu_1(g) - \mu_1(l) = 0$, since the system is at equilibrium, but $\Delta G_{\text{vap}}^0 = \mu_1^0(g) - \mu_1^0(l)$ is not zero; it is the standard free-energy change. It is therefore correct to label the enthalpy change ΔH_{vap}^0 . However, ΔH_{vap}^0 is numerically identical to ΔH_{vap} , which appears in the Clausius–Clapeyron equation [Eq. (4.11)]. The reason for this equality is that we have postulated ideal solution behavior, and for the ideal solution, the enthalpy of mixing is zero (Section 7.1). Consequently there is no enthalpy change on bringing the solution from $x_1 = 1$ to $x_1 < 1$.

which can be written

$$\ln x_1 = -\frac{\Delta H_{\text{vap}}^0}{R} \left(\frac{T - T_b}{T T_b}\right)$$
(9.6)

Now define the boiling point elevation as $\Delta T_b = T - T_b$, and, since T and T_b are quite close together, approximate TT_b by T_b^2 . Making these substitutions in Eq. (9.6) gives

$$\ln x_1 = -\frac{\Delta H_{\rm vap}^0 \Delta T_b}{RT_b^2}$$

Since x_2 is small (the solution is dilute), we write² ln $x_i = \ln (1 - x_2) \approx -x_2$. We also convert x_2 , the mole fraction of solute, to m_2 , the molality of solute, with $x_2 = m_2 M_1/1000$, where M_1 is the molecular weight of solvent [Eq. (5.2)]. The final result of these substitutions is

$$\Delta T_b = \frac{RT_b^2 M_1 m_2}{1000\Delta H_{\rm vap}^0} \tag{9.7}$$

which can be written $\Delta T_b = K_b m_2$, where

$$K_b = \frac{RT_b^2 M_1}{1000 H_{\rm vap}^0} \tag{9.8}$$

The proportionality constant K_b is called the boiling point elevation constant or the *ebullioscopic constant*. Note that K_b can be calculated solely from properties of the solvent and that ΔT_b depends only on the identity of the solvent and the concentration (not the identity) of the solute.

Example 9.1. The heat of vaporization of water is $9.717 \text{ kcal mol}^{-1}$ at its boiling point. Calculate the ebullioscopic constant of water:

$$K_b = \frac{(1.987 \text{ cal mol}^{-1} \text{ K}^{-1})(373.15K)^2 (18.02 \text{ g mol}^{-1})}{(1000)(9717 \text{ cal mol}^{-1})}$$

= 0.513 K g mol⁻¹

The result in Example 9.1 may appear to say that the boiling point of a 1 m solution will be raised 0.513 K, but of course a 1 m solution lies outside the dilute solution range where this equation is valid.³

² We are using the series expansion $\ln(1-x) = -x + x^2/2 - x^3/3 + \cdots$

³ Some discrepancies in the form of Eq. (9.8) for K_b will be noted in the literature. Some authors (Williamson 1967, p. 102; Atkins 1994, p. 229) omit the factor 1000 in the denominator. Others (Smith 1977, p. 90; Gupta 2000) include the 1000 and assign K_b the unit K. The distinction lies in the units given to the molality m_2 , that is, whether molality has the units mol kg⁻¹ or mol (1000) g⁻¹ or is considered to be a dimensionless number.

9.2. FREEZING POINT DEPRESSION

We have seen in Chapter 6 that the freezing point (melting point) of a two-component system is lowered (relative to a pure substance). A simple quantitative treatment can be based on the assumption that the solute does not dissolve in solid solvent; then the solid that forms is pure solvent. The method is identical in form with that used for the analysis of the boiling point elevation; we replace the chemical potential of the gaseous solvent with that of the solid solvent. The result can be written

$$\Delta T_f = K_f m_2 \tag{9.9}$$

where $\Delta T_f = T_f - T$ and

$$K_f = \frac{RT_f^2 M_1}{1000\Delta H_f^0} \tag{9.10}$$

where ΔH_f^0 is the heat of fusion and K_f is the *cryoscopic constant* or *freezing point* depression constant.

Example 9.2. The heat of fusion of water is 6.01 kJ mol^{-1} . Calculate the freezing point depression constant of water:

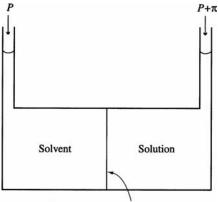
$$K_f = \frac{(8.314 \text{ J mol}^{-1} \text{ K}^{-1})(273.15 \text{ K})^2 (18.02 \text{ g mol}^{-1})}{(1000)(6010 \text{ J mol}^{-1})}$$

= 1.86 K g mol⁻¹

9.3. OSMOTIC PRESSURE

Consider the experimental arrangement in Fig. 9.1, which shows a solvent compartment (left) and a compartment containing a dilute solution in the same solvent (right), the two compartments separated by a *semipermeable membrane*, which permits the passage of solvent molecules but prevents the passage of solute molecules. The presence of solute in the right-hand compartment reduces the mole fraction of solvent in that compartment and thereby reduces its activity and chemical potential below their values in the pure solvent in the left-hand compartment.

In order to achieve equilibrium, the chemical potential of the solvent must be equal on both sides of the membrane. There is thus a driving force for the passage of solvent molecules from left to right. (Although it is actually the chemical potential difference that is responsible for this effect, it can also be rationalized as a simple concentration effect, because the solvent concentration is higher on the left.) The flow of solvent from left to right continues until it is opposed by the backpressure generated by the increased height of solution in the right-hand column. (Or alternatively, the experiment can be arranged so as to apply an excess pressure to the right until the flow of solvent is exactly



Semipermeable membrane

Figure 9.1. Principle of osmosis and the osmotic pressure π .

balanced.) This phenomenon of the passage of solvent through a semipermeable membrane under the influence of a difference in chemical potentials is called *osmosis*, and the excess pressure (π in Fig. 9.1) that equalizes the chemical potentials is the *osmotic pressure*.

Our thermodynamic analysis of osmosis begins with the statement of osmotic equilibrium

$$\mu_1^0(P, T) = \mu_1(P + \pi, T, x_1) \tag{9.11}$$

where the left side of the equation refers to pure solvent (the left-hand compartment in Fig. 9.1) and the right side, to the solution. The parentheses contain the variables controlling the particular quantities, in order to make explicit how the two sides differ. We assume that the solution is dilute and behaves ideally. Then, on expanding μ_1 in the usual manner, Eq. (9.11) becomes

$$\mu_1^0(P, T) = \mu_1^0(P + \pi, T) + RT \ln x_1 \tag{9.12}$$

where now $\mu_1^0(P + \pi, T)$ designates the chemical potential of pure solvent at pressure $P + \pi$. We now develop this quantity as in

$$\mu_1^0(P+\pi, T) = \mu_1^0(P, T) + \int_P^{P+\pi} V_1 \, dP \tag{9.13}$$

where V_1 is the molar volume of solvent [Eq. (3.6)]. Putting Eq. (9.13) into Eq. (9.12) results in

$$\int_{P}^{P+\pi} V_1 \, dP = -RT \ln x_1 \tag{9.14}$$

Treating V_1 as independent of pressure and integrating yields

$$\pi V_1 = -RT \ln x_1 \tag{9.15}$$

Using once again (see note 2) the approximation (for dilute solution) that $\ln x_1 = -x_2$, Eq. (9.15) becomes $\pi V_1 = x_2 RT$. Now, $x_2 = n_2/(n_1 + n_2)$, which in dilute solution is nearly equal to the mole ratio n_2/n_1 . This gives

$$\pi n_1 V_1 = n_2 R T \tag{9.16}$$

The product n_1V_1 equals V, the total volume of the solution, or

$$\pi V = n_2 R T \tag{9.17}$$

The formal resemblance of Eq. (9.17) to the ideal-gas law is obvious. We can take one further step by noting that the ratio n_2/V is the molar concentration c_2 of the solute:

$$\pi = c_2 R T \tag{9.18}$$

Example 9.3. Calculate the osmotic pressure of an 0.01 M solution at 25°C. Since we find it convenient to express pressure in atmospheres, we use *R* as 0.08206 L atm mol⁻¹ K⁻¹. From Eq. (9.18), we have

$$\pi = (0.01 \text{ mol } \text{L}^{-1})(0.08206 \text{ L atm } \text{mol}^{-1} \text{ K}^{-1})(298.15 \text{ K})$$

= 0.245 atm

Example 9.3 shows that osmosis is a very sensitive effect, much more so than are the other colligative properties. This same solution would exhibit a boiling point elevation of 0.0051 K (Example 9.1) and a freezing point depression of 0.0186 K (Example 9.2). This sensitivity forms the basis of an experimental method, called *osmometry*, for measuring molecular weights of solutes. Solutions are prepared with known concentrations in grams per liter (g L⁻¹) and their osmotic pressures are measured. From Eq. (9.18) the corresponding molar concentration c_2 is calculated. Since c_2 has the units mol L⁻¹ and mol = g/M₂ (where M_2 is the molecular weight of the solute), the quantity M₂ can be obtained. In practice, deviations from ideality must be taken into account (Atkins 1994, p. 229).

Osmotic Pumps. The phenomenon of osmosis has found important applications in the delivery of drugs by both oral tablets and implantable systems. The concept in all cases is very simple: A membrane semipermeable to water but not to the drug to be delivered is used to encapsulate the drug, which occupies one chamber in the tablet (one layer in the implant). Additives such as osmotic agents (salts like magnesium)

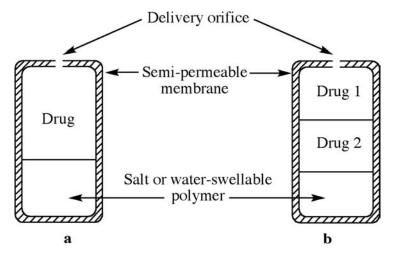


Figure 9.2. (a) An osmotic pump can be used to release drug at a controlled rate from an oral tablet. (b) The same concept applied to the delivery of two drugs through one tablet.

sulfate) or water-swellable polymers occupy a second chamber (different layer in an implant). Figure 9.2a shows the simplest osmotic pump. The tablet (or the implant) is equipped with one or more micro-orifices drilled in the system with a laser. Upon exposure to an aqueous environment, the semipermeable membrane will allow water to enter the tablet. The chamber where the salt or the polymer is contained will increase in volume, and this increase will generate enough pressure to push the drug out of the orifice as an aqueous suspension. The rate of release will be determined by the rate of water-diffusion and the orifice diameter. This conceptually remarkably simple concept can be extended to tablets containing more than one drug as shown in Fig. 9.2b.

The use of a number of different additives as well as complexing agents for enhancing the water-solubility of hydrophobic drugs (cyclodextrins, an important class of drug-complexing agents are described in Chapter 14) has led to a number of modifications of the original osmotic pump design. Osmotic pumps present the advantage of leading to zero-order kinetics in drug release, that is, constant drug release over time. This consistent, stable drug release helps to achieve better therapeutic control.

9.4. ISOTONICITY CALCULATIONS

Body membranes, including cell membranes, are semipermeable membranes to some degree. They are generally permeable to water and are impermeable, or nearly so, to many (but obviously not all) solutes. Thus we anticipate the existence of osmotic pressure differences across these membranes. It is known that irritation caused by foreign solutions is in part related to their osmotic pressure; the closer the osmotic pressure of an administered solution is to that of the physiological solution on the other side of the membrane, the less the discomfort that is experienced.

Two solutions are said to be *isoosmotic* if they have the same osmotic pressure. Isoosmoticity is therefore a physical property, based on the thermodynamic concept of Section 9.3. A solution is said to be *isotonic* if it has the same osmotic pressure as a reference body fluid, measured with respect to the appropriate body membrane. Isotonicity is therefore a physiological concept. It is possible for a pair of solutions to be both isoosmotic and isotonic, but they need not be, as when the biological membrane is not perfectly impermeable to the solute. For example, 1.9% boric acid solution (in water, the solvent for all solutions in this section) is both isoosmotic and isotonic with respect to the eye, because the corneal membrane is impermeable to boric acid. On the other hand, 1.9% boric acid is isoosmotic with the red blood cell contents, but it is not isotonic toward this biological medium, because the red blood cell membrane is permeable to boric acid. (Boric acid is often used to render ophthalmic solutions isotonic.)

A solution having an osmotic pressure greater than that of physiological fluids is *hypertonic*; if its osmotic pressure is less than that of physiological fluids, it is *hypotonic*. Consider a red blood cell surrounded by a hypertonic solution. Since the osmotic pressure of the surrounding solution is greater than that inside the cell (i.e., the water activity is less outside than inside the cell), water will flow out of the cell, which shrinks and shrivels. If the red blood cell should be immersed in a hypotonic solution, water will flow into the cell, which swells and may burst. The goal of rendering pharmaceutical solutions isotonic is directed toward preventing or minimizing such physiological consequences.

Experimental work has shown that the freezing point of human blood is -0.52° C. We consider that all other physiological fluids are effectively in equilibrium with blood, and so they are isotonic with blood. Although we saw that osmotic pressure is the most sensitive of the colligative properties, the freezing point depression is much the easiest to measure, and so it forms the basis of all isotonicity calculations. Here we will describe the simplest of these, called straightforwardly the *freezing point depression method*. For other methods the literature may be consulted (Thompson 1998, Chapter 10; Windholz 1983, pp. MISC-47–MISC-69; Reich et al. 2000). The basis of the method is the assumption that contributions to the freezing point depression from multiple solutes are additive, and so the goal of the calculation is to achieve a freezing point depression of -0.52° C.

Example 9.4. Estimate the concentration of sodium chloride required to produce an isotonic aqueous solution.

The calculation is based on Eq. (9.9), $\Delta T_f = K_f m_2$, with $K_f = 1.86^{\circ}$ C (see Example 9.2). We seek $\Delta T_f = 0.52^{\circ}$ C; hence we need $m_2 = 0.52/1.86 = 0.28$. But NaC1 is a 1 : 1 strong electrolyte, so each molecule yields two particles on dissociation; therefore we actually need to prepare a solution 0.28/2 = 0.14 m. Commonly this molal concentration is expressed as a molar concentration, neglecting the difference; to obtain 0.14 M NaCl, whose molecular weight is 58.5, we take (0.14) (58.5) = 8.19 g L⁻¹, or 0.82 g/100 mL, which is 0.82%.

Experimentally it is observed that a solution 0.5% in NaCl exhibits a freezing point depression of 0.289°C. Setting up the proportion

$$\frac{0.289^{\circ}C}{0.5\%} = \frac{0.52^{\circ}C}{x}$$

we find x = 0.9% NaCl. The slight discrepancy with the result in Example 9.4 probably is a consequence of nonideal solution behavior. An aqueous solution containing 0.9% (w/v) sodium chloride is isotonic. This solution is called *normal saline* or *physiological saline*.

Because of nonideal solution behavior or incomplete electrolyte dissociation, it is preferable to make use of published freezing point depression data. However, if such data are not available, a very reasonable calculation can be based on the known K_f value of 1.86 together with chemical knowledge of the nature of the solute.

Example 9.5. Give directions for the preparation of 100 mL of isotonic 1% hexamethonium tartrate. The following data are available (Windholz 1983, pp. MISC-47–MISC-69; Reich et al. 2000):

Concentration	ΔT_f
0.5	0.045
1	0.089
2	0.181
3	0.271
5	0.456

We set up the problem in tabular form:

Desired ΔT_f value	0.52°C
ΔT_f of 1% of drug	0.089°C
Difference to be made up	0.431°C

We will use NaCl to make the solution isotonic. From the proportion

$$\frac{0.9\%}{0.52^{\circ}\text{C}} = \frac{x}{0.431^{\circ}\text{C}}$$

we find x = 0.75%. Therefore we proceed by dissolving 1.0 g of hexamethonium tartrate and 0.75 g of NaCl in enough water to make 100 mL.

If data are not available for the concentration we require, interpolation or extrapolation may give the information.

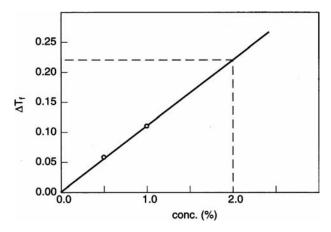


Figure 9.3. Freezing point depression of hexamethonium tartrate. See Example .

Example 9.6. Prepare 2 oz of 2% isotonic imipramine hydrochloride.

Sources give $\Delta T_f = 0.058^{\circ}$ C at 0.5% and 0.110°C at 1%. We can plot these data as in Fig. 9.3 and obtain the estimate $\Delta T_f = 0.225^{\circ}$ C at 2%. (Extrapolation is risky, but it may be our best recourse.) We proceed as before:

Desired ΔT_f value	0.52°C
ΔT_f of 2% of drug	0.225°C
Difference to be made up	0.295°C

Again using NaCl, we obtain

$$\frac{0.9\%}{0.52^{\circ}C} = \frac{x}{0.295^{\circ}C}$$
$$x = 0.51\%$$

If we needed to prepare 100 mL of the solution we would take 2.0 g of drug and 0.51 g of NaCl. However, only 2 oz (60 mL) of solution is required, so we take (60/100) (2) = 1.2 g of drug and (60/100) (0.51) = 0.31 g of NaC1 in enough water to make 60 mL.

If the prescription calls for more than one drug, the ΔT_f contributions of the several drugs are summed. In ophthalmic solutions, boric acid may be used to make the solution isotonic; 1.9% boric acid solution is isotonic.

Although the several alternative methods of isotonicity calculation are all derived from the freezing point depression method, one of these approaches merits comment because of its simplicity and utility. The principle is as follows. If two isotonic solutions are mixed, the resulting solution is isotonic. For example, if the prescribed amount of drug is dissolved in just enough water to make an isotonic solution, this solution could be diluted to the desired volume with normal saline and the result will be isotonic. The *United States Pharmacopeia* gives formulas for isotonic phosphate buffer solutions that can be used in this way, particularly for ophthalmic solutions.

PROBLEMS

- **9.1.** Starting with Raoult's law, show that the extent of vapor pressure lowering is directly proportional to solute concentration.
- **9.2.** Calculate the boiling point elevation constant of ethanol, whose heat of vaporization is 38.6 kJ mol^{-1} .
- **9.3.** Calculate the freezing point depression constant of glacial acetic acid, whose heat of fusion is 11.7 kJ mo1^{-1} .
- **9.4.** Give directions for the preparation of 1 oz of this ophthalmic solution; ΔT_f for 2% pilocarpine nitrate is 0.247: Pilocarpine nitrate 2% Make isotonic with boric acid
- **9.5.** Estimate ΔT_f of 5% dextrose solution. Dextrose (glucose) is available as the monohydrate, MW 198.2. What concentration of dextrose will yield an isotonic solution?
- **9.6.** A and B are two pure liquids of similar freezing points and molecular weights. Liquid A freezing-point depression constant is twice that of liquid B. What is the relationship between the heats of melting of A and B?
- **9.7.** A solution is made at 25° C by mixing 0.50 g of liquid C (molecular weight = 250 g/mol) and 1.60 g of liquid D (molecular weight = 200 g/mol). The vapor pressure of liquid C at 25° C is 268 mmHg. The vapor pressure of liquid D at 25° C is 236 mmHg. It is found experimentally that the partial vapor pressure of C and D above this solution are 45 mmHg and 177 mmHg, respectively. What are the activity coefficients of C and D in this solution?
- **9.8.** What are the freezing point and the osmotic pressure of an aqueous 0.2 m solution of sucrose at 25° C? Consider the solution as dilute and assume density = 1.02 g cm^{-3} .

10

SOLUBILITY

10.1. SOLUBILITY AS AN EQUILIBRIUM CONSTANT

The topic of solubility merits special attention because of its great importance in pharmaceutical systems. We can generally anticipate that a drug must be in solution if it is to exert its effect. Typically, the type of system we encounter is a pure solid substance (the solute) in contact with a pure liquid (the solvent). We allow equilibrium to be achieved at fixed temperature and pressure, such that at equilibrium the system consists of (excess) pure solid phase and liquid solution of solute dissolved in solvent. According to Gibbs' phase rule, P = 2 and C = 2, so F = C - P + 2 = 2 degrees of freedom. These are the temperature and pressure, which we have specified as fixed. Thus there remain no degrees of freedom; the system is invariant. This means that at fixed temperature and pressure, the concentration of dissolved solute is fixed. We call this invariant dissolved concentration the *equilibrium solubility* of the solute at this pressure and temperature. (We say that the solution is *saturated*.) Our present concern is with how the equilibrium solubility depends on the temperature and on the chemical natures of the solute and the solvent.

Expressed as a reaction, the dissolution process is

Pure solvent + Pure solute \rightleftharpoons Solute in solution

At equilibrium the chemical potentials of the solute in the two phases are equal; or, letting component 1 be the solvent and component 2 the solute, we obtain

 $\mu_2(\text{solid}) = \mu_2(\text{solution})$

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Writing out the chemical potentials gives

$$\mu_2^0(\text{solid}) + RT \ln a_2(\text{solid}) = \mu_2^0(\text{solution}) + RT \ln a_2(\text{solution})$$
(10.1)

where the standard state of the solid is the pure solid, and we will adopt as the standard state of the solute in solution the Henry's law definition on the molar concentration scale. Rearranging Eq. (10.1) leads to

$$\Delta \mu^0 = -RT \ln \frac{a_2(\text{solution})}{a_2(\text{solid})}$$
(10.2)

where $\Delta \mu^0 = \mu_2^0$ (solution) $-\mu_2^0$ (solid). But the solid is in its standard state, so $a_2(\text{solid}) = 1.0$ by definition and we obtain

$$\Delta \mu^0 = -RT \ln a_2(\text{solution}) \tag{10.3}$$

We have seen that a_2 (solution) is invalant—it is the activity corresponding to the equilibrium solubility—so comparison of Eq. (10.3) with the fundamental thermo-dynamic result

$$\Delta G^0 = -RT \ln K \tag{10.4}$$

leads to the conclusion that a_2 (solution), the activity of the solute in a saturated solution, must have the character of an equilibrium constant. As a consequence, we can evaluate standard free energy, enthalpy, and entropy changes for the soution process in the usual manner (Chapter 4). These quantities are respectively called the *free energy*, *heat*, and *entropy of solution*.

For nonelectrolyte solutes, particularly those of limited solubility, so that the saturated solution is fairly dilute, it will be acceptable to approximate the activity a_2 (soln) by the equilibrium solubility concentration. This is usually in molar concentration units and is often symbolized *s*.

10.2. THE IDEAL SOLUBILITY

A thermodynamic argument can predict the equilibrium solubility of a nonelectrolyte, provided that it dissolves to form an ideal solution. Ideal behavior does not mean that intermolecular interactions are absent. On the contrary, solids and liquids would not exist without the intermolecular forces of interaction. In the present context, ideal behavior means that the energy of interaction between two solvent molecules is identical to that between one solvent and one solute molecule, so that a solvent molecule may be replaced with a solute molecule without altering the intermolecular energies. (This requires that the solvent and solute molecules have the same size, shape, and chemical nature, a demanding set of limitations.)

Quantitatively, an ideal solution can be defined as one having the following properties (Chapter 7):

$$\Delta H_{\rm mix} = 0 \tag{10.5}$$

$$\Delta V_{\rm mix} = 0 \tag{10.6}$$

$$\Delta S_{\rm mix} = -R(x_1 \ln x_1 + x_2 \ln x_2) \tag{10.7}$$

According to Eqs. (10.5) and (10.6), there is no heat or volume change on mixing the solute and solvent in an ideal solution, and the entropy change is given by Eq. (10.7). Since $x_1 + x_2 = 1$, the logarithmic terms are necessarily negative, so ΔS_{mix} is positive, and this constitutes the "driving force" for dissolution, because of the relationship $\Delta G = \Delta H - T \Delta S$.

If the entropy of mixing is the driving force for dissolution, what is the "resistance"? It is the solute–solute interaction forces, which, for solids, lead to the "crystal lattice energy." These must be overcome for the solute to dissolve. Now, the free-energy change for the dissolution process is the same no matter what reversible mechanism (path) is taken to pass from the initial state (pure solute) to the final state (saturated solution), so we can divide the process as follows (for a solid solute):

Sum : Crystalline solute + Pure liquid solvent \rightleftharpoons Saturated solution

Since in an ideal solution the solvent–solvent interactions match the solvent–solute interactions, the energy required to create molecule-sized cavities in the solvent is offset by the energy recovered when the solute molecules are inserted into these cavities. The energetic cost of the dissolution process then appears in the first step, the melting of the solid. An equivalent viewpoint (Grant and Higuchi, 1990, p. 16) is that the enthalpy of solution is given by

$$\Delta H_{\rm soln} = \Delta H_{\rm fusion} + \Delta H_{\rm mix}$$

But $\Delta H_{\text{mix}} = 0$ for an ideal solution, so $\Delta H_{\text{soln}} = \Delta H_{\text{fusion}}$.

The saturation solubility, we have seen, is an equilibrium constant, so the van't Hoff equation [Eq. (4.29)] is applicable:

$$\frac{d\ln x_2}{dT} = \frac{\Delta H_f}{RT^2} \tag{10.8}$$

where the solubility is expressed as the mole fraction simply to maintain consistency with Eq. (10.7) and where ΔH_f is the heat of fusion and *T* is the absolute temperature. We have seen above why the heat of fusion appears in a solubility expression. (Incidentally, a dissolved solid should be viewed as possessing some of the properties of the liquid state, consistent with the above view that fusion is the first step in the dissolution process.) Now suppose that ΔH_f is independent of temperature, which is equivalent to writing for the solute, from Eq. (1.23),

$$\Delta C_p = C_p^{\text{liq}} - C_p^{\text{solid}} = 0 \tag{10.9}$$

Then integrating Eq. (10.8) from T_m to T gives

$$\ln x_2 = -\frac{\Delta H_f}{R} \left(\frac{T_m - T}{T T_m} \right) \tag{10.10}$$

where T_m is the melting temperature and T is the experimental temperature. Equation (10.10) allows us to calculate the ideal solubility.

Example 10.1. The melting point of naphthalene is $80.2 \degree \text{C}$, and its heat of fusion at the melting point is $4.54 \text{ kcal mo1}^{-1}$. What is the ideal solubility of naphthalene at $20 \degree \text{C}$?

$$Log x_{2} = \frac{-4540 \text{ cal mol}^{-1}}{(2.303)(1.987 \text{ cal mol}^{-1}\text{K}^{-1})} \left(\frac{60.2 \text{ K}}{353.35 \text{ K} \times 293.15 \text{ K}}\right)$$
$$= -0.577$$
$$x_{2} = 0.265$$

Deviations from ideality will be manifested by discrepancies from the ideal solubility as calculated with Eq. (10.10). Table 10.1 lists equilibrium solubilities for naphthalene in many solvents. Observe that those solvents most chemically like

Solvent	<i>x</i> ₂
(Ideal)	0.265
Chlorobenzene	0.256
Benzene	0.241
Toluene	0.224
Carbon tetrachloride	0.205
Hexane	0.090
Aniline	0.130
Nitrobenzene	0.243
Acetone	0.183
<i>n</i> -Butanol	0.0495
Methanol	0.0180
Acetic acid	0.0456
Water (25 °C)	0.0000039

Table 10.1. Naphthalene solubility at 20 °C

naphthalene—that is, aromatic and nonpolar solvents—show behavior most closely approximating ideal behavior.

At the melting temperature T_m the solid and liquid forms of the solute are in equilibrium, so $\Delta G_f = 0$ and we get $\Delta H_f = T_m \Delta S_f$, giving Eq. (10.11) as an alternative form of Eq. (10.10):

$$\ln x_2 = -\frac{\Delta S_f(T_m - T)}{RT}$$
(10.11)

10.3. TEMPERATURE DEPENDENCE OF THE SOLUBILITY

Since ΔH_f is always a positive quantity, Eq. (10.10) predicts that the solubility of a solid will increase with temperature. Moreover, Eq. (10.10) shows that if two solid substances have the same heat of fusion, the one with the higher melting point will have the lower solubility. Conversely, if they have the same melting point, the one with the lower heat of fusion will have the higher solubility. All of these inferences from Eq. (10.10) refer to systems forming ideal solutions, so deviations from the predictions can occur for real systems. Nevertheless, the increase of solubility with temperature is very widely observed for solids. Even the relationship of solubility to melting point can be a useful guide, though confounding phenomena can introduce complications; for example, hydrogen-bonding or other polar interactions may raise both the melting point and the aqueous solubility. The comparison of the temperature dependence of solubility of solids and gases is instructive; see Table 10.2.

Equation (10.10) can be rearranged to Eq. (10.12):

$$\ln x_2 = -\frac{\Delta H_f}{RT} + \frac{\Delta H_f}{RT_m}$$
(10.12)

If ΔH_f is essentially constant over the experimental temperature range, Eq. (10.12) predicts that a plot of $\ln x_2$ against 1/T will be linear with a slope equal to $-\Delta H_f/R$. The line should terminate at the melting point, where $1/T = 1/T_m$. Often such lines are straight, probably because the usual range of temperatures is small. The slope gives ΔH_f in principle, but in actuality the quantity evaluated from the slope is not precisely

Table 10.2. The contrary effects of temperature on the solubilities of solids and gases

Solid
$$\stackrel{\Delta H_{\rm f}}{\rightleftharpoons}$$
 Liquid $\stackrel{\Delta H_{\rm v}}{\underset{\Delta H_c}{\rightleftharpoons}}$ Gas

Solids	Gases
Solution is the process of passing from solid to liquid (fusion, ΔH_f)	Solution is the process of passing from gas to liquid (condensation, ΔH_c), which is
ΔH_f is positive, so x_2 increases as	the reverse of vaporization (ΔH_{ν}) ΔH_{ν} is positive, so ΔH_c is <i>negative</i> ;
T increases	thus x_2 decreases as T increases

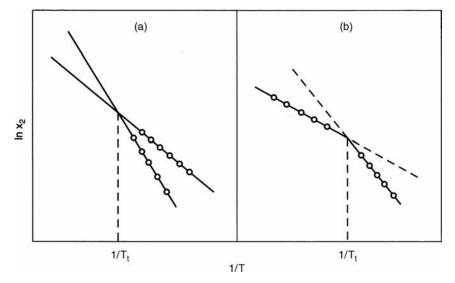


Figure 10.1. Hypothetical solubility van't Hoff plots for polymorphs.

 ΔH_f because the solution is seldom ideal, and instead the quantity found in this way is termed the *heat of solution*.

Throughout this discussion we have been assuming that the solid phase consists of the pure solid and not a solid solution. Another possible complication arises if the solid substance can exist in two crystalline forms (polymorphs; Chapter 6), which interconvert at transition temperature T_i . The van't Hoff plot can resemble Fig. 10.1a or Fig. 10.1b, depending primarily on the kinetics of the transformation. In Fig. 10.1a, the two forms are sufficiently stable that their solubilities can be separately measured at the same temperatures, which are below the transition temperature. Nevertheless, the crystal form having the higher solubility (at a given temperature) is thermodynamically unstable (it is said to be metastable, since its kinetics of transformation permit it to exist for some period during which it acts as if it were stable) and will ultimately be converted to the stable form. Extrapolation of the lines to the transition temperature may be possible. Sulfathiazole in 95% ethanol shows the Fig. 10.1a behavior (Milosovich 1964; Carstensen 1977, p. 7).

In Fig. 10.1b, one form exists in one temperature range, while the other form exists in a temperature range on the other side of T_t . The melting point observed will be that of the higher-melting polymorph. Carbon tetrabromide exemplifies this behavior (Hildebrand et al., 1970, p. 23).

Let us return to the assumption that the change in heat capacities, ΔC_p , is zero, for all the subsequent discussion was based on this assumption. If ΔH_f in fact is a function of temperature, then ΔC_p is not zero. Suppose we make the more reasonable assumption that ΔC_p is a nonzero constant, and write ΔH_f as

$$\Delta H_f = \Delta H_f^m - \Delta C_p (T_m - T) \tag{10.13}$$

where ΔH_f^m is the heat of fusion at T_m . Equation (10.13) is inserted into Eq. (10.8), which can be rearranged and integrated to give Eq. (10.14):

$$\ln x_2 = -\frac{\Delta H_f^m}{R} \left(\frac{T_m - T}{TT_m} \right) + \frac{\Delta C_p}{R} \left(\frac{T_m - T}{T} \right) - \frac{\Delta C_p}{R} \ln \frac{T_m}{T}$$
(10.14)

This equation is useful for assessing the error that may be introduced by making the simple assumption $\Delta C_p = 0$. Suppose, for example, that the experimental temperature is 25 °C and the melting point is 100 °C. Then the last two terms in Eq. (10.14) become equal to $0.25\Delta C_p/R - 0.22\Delta C_p/R = 0.03\Delta C_p/R$. Thus considerable compensation can take place, making the approximation $\Delta C_p = 0$ more acceptable than it might have seemed.

Example 10.2. These are solubility data for nitrofurantoin in water (Chen et al. 1976). Analyze the data to obtain the heat of solution.

$T(^{\circ}C)$	$10^6 x_2$
24	6.01
30	8.57
37	13.16
45	18.99

The data are manipulated as required to make the van't Hoff plot according to Eq. (10.12):

$10^{3} K/T$	$\log x_2$
3.37	-5.22
3.30	-5.06
3.23	-4.88
3.14	-4.72

The plot is shown in Fig. 10.2. It is possible that the points describe a curve, but this is uncertain with the data as given, because conceivably the scatter is a consequence of experimental random error. A straight line has therefore been drawn. Its slope is -2300 K, so we calculate

$$\Delta H_{\text{soln}} = (2300 \text{ K})(1.987 \text{ cal mol}^{-1} K^{-1})$$

= 4570 cal mol⁻¹
= 4.57 kcal mol⁻¹
= 19.1 kJ mol⁻¹

Note that the enthalpy change is labeled ΔH_{soln} to indicate explicitly that this is a heat of solution.

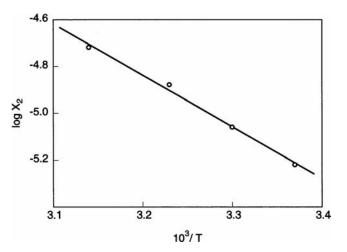


Figure 10.2. van't Hoff plot for nitrofurantoin solubility.

10.4. SOLUBILITY OF SLIGHTLY SOLUBLE SALTS

Many salts exhibit very low solubilities in water. Silver chloride is an example; If aqueous solutions of silver nitrate and sodium chloride are mixed, solid silver chloride precipitates. It is conventional to describe this process as the reverse of the precipitation reaction, namely, as the dissolution of the salt. Let us begin with the simplest case of a 1 : 1 sparingly soluble salt MX. The solid crystalline form is ionic. When it dissolves in water the ions dissociate, and no ion pairs are detectable. We therefore write the equilibrium as

$$MX(s) \rightleftharpoons M^+ + X^- \tag{10.15}$$

Proceeding as we have done for several earlier processes, we equate the chemical potentials of the solid and the dissolved solute at equilibrium:

$$\mu(\mathbf{s}) = \mu(\mathbf{soln})$$

Expanding these gives

$$\mu^{0}(s) + RT \ln a(s) = \mu^{0}_{+} + RT \ln a_{+} + \mu^{0}_{-} + RT \ln a_{-}$$

and collecting terms (and noting that a(s) = 1 by our standard state definition),

$$\Delta \mu^0 = -RT \ln a_+ a_- \tag{10.16}$$

where $\Delta \mu^0 = \mu^0_+ + \mu^0_- - \mu^0(s)$. Evidently then [compare with Eq. (10.4)1, the product a_+a_- is an equilibrium constant. By Eq. (8.23a) we see that $a_+a_- = a_{\pm}^2$,

where a_{\pm} is the mean ionic activity, and since $a_{\pm}^2 = \gamma_{\pm}^2 c_{\pm}^2$, Eq. (10.16) can be written

$$\Delta \mu^0 = -RT \ln \gamma_+^2 c_+^2 \tag{10.17}$$

If no extraneous ions are present, so that the ionic strength is due solely to the ions from the sparingly soluble salt (and hence is very low), the activity coefficient term is essentially unity. Moreover, the molar concentrations of the cation M^+ and the anion X^- are equal, and each is numerically equal to the equilibrium molar solubility of the salt, which is commonly denoted *s*. Thus Eq. (10.17) becomes

$$\Delta \mu^0 = -RT \ln s^2 \tag{10.18}$$

Equation (10.17) is exact; Eq. (10.18) is usually a reasonable approximation, and both implicitly define the equilibrium constant for Eq. (10.15). This constant is symbolized K_{sp} and is called the *solubility product*. Since solubility products are very small numbers, it is common to state them as pK_{sp} , where $pK_{sp} = -\log K_{sp}$. Table 10.3 lists some pK_{sp} values.

Example 10.3. What is the solubility of silver chloride in water? From Table 10.3, $pK_{sp} = 9.75$ for AgCl, so $K_{sp} = 1.78 \times 10^{-10}$. From Eq. (10.18), $K_{sp} = s^2$, so $s = \sqrt{K_{sp}} = 1.33 \times 10^{-5}$ M.

In the general case of the salt whose formula is M_pX_q the solubility product is defined, in accordance with the usual formulation of equilibrium constants:

$$K_{\rm sp} = c_M^p c_X^q \tag{10.19}$$

The quantity that we label *s* then depends on the stoichiometry.

Example 10.4. What is the molar solubility of ferrous hydroxide?

Salt	pK _{sp}	Salt	pK _{sp}
BaSO ₄	9.96	PbCO ₃	13.13
CaCO ₃	8.54	PbS	27.9
Ca(OH) ₂	5.26	MgCO ₃	7.46
$Ca_3(PO_4)_2$	28.7	Hg_2S	47.0
CuI	11.96	HgS (red)	52.4
AuCl	12.7	HgS (black)	51.8
AuCl ₃	24.5	AgBr	12.30
Fe(OH) ₂	15.1	AgCl	9.75
Fe(OH) ₃	37.4	AgI	16.08

Table 10.3. Solubility products for slightly soluble salt^a

^aIn the temperature range 18–25 °C; water is the solvent.

From Table 10.3, $pK_{sp} = 15.1$, or $K_{sp} = 7.9 \times 10^{-16}$. The dissolution reaction is

$$Fe(OH)_2 \rightleftharpoons Fe^{2+} + 2OH^{-}$$

so $K_{\rm sp} = c_{\rm Fe}c_{\rm OH}^2$. (The charges on the subscripts are omitted for clarity.) Since each molecule of Fe(OH)₂ that dissolves yields one Fe²⁺ ion, we define the solubility as the concentration of ferrous ion, or $c_{\rm Fe}$, = s. The stoichiometry yields $c_{\rm OH} = 2c_{\rm Fe}$, so the result is ¹

$$K_{\rm sp} = {\rm s} \times (2{\rm s})^2 = 4s^3$$

Therefore $s = 5.8 \times 10^{-6}$ M.

Example 10.5. What is the solubility of silver chloride in 0.02 M KCl? Assume that activity coefficients are unity.

Again we set $c_{Ag} = s$, the solubility. The solubility product is defined $K_{sp} = c_{Ag}c_{Cl}$; however, the chloride concentration has been augmented by the addition of potassium chloride, so we write $c_{Cl} = 0.02 + s$; that is, the chloride concentration is the sum from two sources, the KCl and the AgCl. We therefore have $K_{sp} = s(0.02 + s)$, which is a quadratic equation that can be solved for *s*. Before doing that, however, it is worth trying the approximation $c_{Cl} = 0.02$, which involves neglecting the relatively small contribution from dissolution of the AgCl. Thus

$$K_{\rm sp} = 0.02s = 1.78 \times 10^{-10}$$

 $s = 8.9 \times 10^{-9} M$

First note that the approximation seems well justified. More interestingly, observe that the solubility of silver chloride has been reduced from about 1×10^{-5} M in water (Example) to about 1×10^{-8} M in 0.02 M KCl. This is an example of the *common ion effect*. The solubility of any slightly soluble salt can be reduced by adding an excess of one of its constituent ions.

The accuracy of such calculations can be improved by making use of the Debye–Hückel equation to estimate the values of mean ionic activity coefficients.

10.5. SOLUBILITIES OF NONELECTROLYTES: FURTHER ISSUES

Salt Effects. In Example 10.5 we encountered one type of salt effect. There is another type of salt effect that is observed when the solubility of a nonelectrolyte is studied as a function of ionic strength (or of the concentration of an added electrolyte).

¹ Equations like this one in Example 10.4 are easily solved by a logarithmic technique. We have $7.9 \times 10^{-16} = 4 \text{ s}^3$, or $1.975 \times 10^{-16} = \text{s}^3$. Take logarithms of both sides, obtaining $-15.70 = 3 \log s$, or $-5.235 = \log s$. The antilogarithm gives *s*.

Compare the nonelectrolyte solubility in the absence and presence of added salt. Since the solid solute is present in both cases, we obtain

$$\mu(\text{solid}) = \mu(c_s = 0) = \mu(c_s)$$

where c_s , is the concentration of added salt. Therefore $a(c_s = 0) = a(c_s)$, or

$$s_0 \gamma_0 = s \gamma \tag{10.20}$$

where s_0 and s are the solubilities in the two cases. Thus $\gamma/\gamma_0 = s_0/s$; and since $\gamma_0 = 1$ is a reasonable assumption, $\gamma = s_0/s$, and we have a method for measuring nonelectrolyte activity coefficients. Moreover, it is found experimentally that the quantity log (s_0/s) often varies linearly with c_s , or

$$\operatorname{Log}\frac{s_0}{s} = k_s c_s \tag{10.21}$$

If $s_0/s > 1$, then k_s is positive, and the nonelectrolyte is said to be "salted out"; if $s_0/s < 1$, then k_s is negative, and the solute is "salted in." These are called the "salting-out and salting-in effects," and the constant k_s , is known as the Setschenow constant.

Regular Solution Theory. We have seen that an ideal solution has thermodynamic mixing quantities $\Delta H_{\text{mix}} = 0$ and $\Delta S_{\text{mix}} = -R(x_1 \ln x_1 + x_2 \ln x_2)$. A *regular solution* is defined to be one having an ideal entropy of mixing but a nonideal enthalpy of mixing. Recall also that the ideal solubility of a nonelectrolyte (i.e., the solubility when a nonelectrolyte forms an ideal solution) is given by

$$\ln x_2 = \frac{-\Delta H_f}{R} \left(\frac{T_m - T}{TT_m} \right) \tag{10.22}$$

where ΔC_p is assumed to be zero or negligible. The molecular interpretation of an ideal solution is that the energy of interaction of a solute molecule with a solvent molecule is identical with the energy of interaction of two solvent molecules.

The molecular interpretation of regular solution theory is quite different; in regular solution theory the energy of 1-2 interactions (where 1 is the solvent, 2 is the solute) is approximated as the geometric mean of 1-1 and 2-2 interaction energies, or ²

$$U_{12} = \left(U_{11}U_{22}\right)^{1/2} \tag{10.23}$$

This approximation results in regular solution theory being applicable mainly to fairly nonpolar systems, that is, nonpolar nonelectrolytes dissolved in nonpolar solvents. For our present interest, the essential result (Hildebrand and Scott, 1964, p. 271) of regular solution theory is embodied in Eq. (10.24), which may be compared

² The arithmetic mean of two numbers is (a + b)/2; their geometric mean is $(ab)^{1/2}$.

with Eq. (10.22):

$$\ln x_2 = \frac{-\Delta H_f}{R} \left(\frac{T_m - T}{TT_m} \right) - \frac{V_2 \varphi_1^2}{RT} (\delta_1 - \delta_2)^2$$
(10.24)

where V_2 is the molar volume of solute and φ_1 is the volume fraction concentration of solvent in the solution. The quantities δ_1 and δ_2 are the *solubility parameters* of the solvent and solute. These are physical properties with the following significance.

The term ΔH_{vap} , the molar heat of vaporization, is the enthalpy required to effect the transformation of one mole of liquid to its vapor state. During this process all the solvent–solvent interactions (which are responsible for the existence of the liquid phase) are overcome. A quantity called the *cohesive energy density* (CED) is defined

$$CED = \frac{\Delta H_{vap} - RT}{V}$$
(10.25)

where *V* is the molar volume of the liquid. We anticipate, and we find, that liquids with strong intermolecular interactions (especially polar "associated" liquids having the potential for strong dipole–dipole and hydrogen-bonding interactions) have larger CED values than do nonpolar liquids. Table 10.4 lists some CED values.

Because of the manner in which CED appears in regular solution theory equations, Hildebrand (Hildebrand et al. 1970; Hildebrand and Scott 1964, p. 271) defined the solubility parameter δ by Eq. (10.26). Table 10.4 also gives δ values.

$$\delta = (\text{CED})^{1/2} \tag{10.26}$$

Solvent	CED (cal cm $^{-3}$)	$\delta ({\rm cal}^{1/2}{\rm cm}^{-3/2})$
<i>n</i> -Pentane	50.2	7.0
Cyclohexane	67.2	8.2
1,4-Dioxane	96	10.0
Benzene	84.6	9.2
Diethyl ether	59.9	7.4
Ethyl acetate	83.0	9.1
Acetic acid	102	10.1
<i>n</i> -Butyl alcohol	130.0	11.4
<i>n</i> Propyl alcohol	141.6	11.9
Acetone	95	9.9
Ethanol	168	12.7
Methanol	212	14.5
Acetonitrile	141.6	11.9
Dimethylformamide	146.4	12.1
Ethylene glycol	212	14.6
Glycerol	272	16.5
Dimethylsulfoxide	144	12.0
Water	547.6	23.4

Table 10.4. Cohesive energy densities and solubility parameters

Referring now to Eq. (10.24), note that if $\delta_1 = \delta_2$, we recover Eq. (10.22) for the ideal solution; in other words, the condition $\delta_1 = \delta_2$ is equivalent to the condition $\Delta H_{\text{mix}} = 0$. The greater the difference $\delta_1 - \delta_2$ (or $\delta_2 - \delta_1$, because the difference is squared), the greater the deviation from ideality, and, as Eq. (10.24) shows, the lower the solubility that is predicted. This provides a guide for experimental design; to achieve maximal solubility according to regular solution theory, strive to equate the solubility parameters of solvent and solute. Since the solute identity is usually established by the nature of the problem, the experimental variable is the solvent identity. Sometimes mixed solvent systems function better than pure solvents for this reason. For example, a mixture of ether ($\delta = 7.4$) and ethanol ($\delta = 12.7$) dissolves nitrocellulose ($\delta = 11.2$), although neither pure liquid serves as a good solvent for this solute.³

Although the cohesive energy density, and therefore the solubility parameter, is a well-defined physical property for any solvent, regular solution theory is limited (e.g.,by the geometric mean approximation) to solutions of nonpolar substances. It should therefore not be expected to apply quantitatively to polar systems such as aqueous solutions.

Example 10.6. Predict the solubility of naphthalene in *n*-hexane at 20 °C. The solubility parameters are $\delta_1 = 7.3$ and $\delta_2 = 9.9$ (both in cal^{1/2} cm^{-3/2}), and the molar volumes are $V_1 = 132$ cm³ mol⁻¹ and $V_2 = 123$ cm³ mol⁻¹. See Example 10.1 for additional data.

We use Eq. (10.24), which in Example 10.1 was expressed in terms of log x_2 . In that form the first term on the right had the value -0.577, which we need not recalculate. Now we consider the second term. We lack only the quantity φ_1 , the volume fraction of solvent. This appears to be a dilemma, because we cannot estimate φ_1 until we know x_2 , which is what we want to calculate.

If we anticipate that the solute has a low solubility, it may be acceptable to make the approximation $\varphi_1 = 1$. An alternative is to take the result for an ideal solution (Example 10.1, which gave $x_2 = 0.265$) as a basis for estimating φ_1 . We will do the problem in both ways.

(a) Let $\varphi_1 = 1$. Then from Eq. (10.24), we obtain

$$\log x_2 = -0.577 - \frac{(123 \text{ cm}^3)(1)^2 (7.3 - 9.9 \text{ cal}^{1/2} \text{ cm}^{-3/2})^2}{(2.303)(1.987 \text{ cal mol}^{-1} \text{K}^{-1})(293.15 \text{ K})}$$

= -0.577 - 0.620 = -1.197
$$x_2 = 0.064$$

³ As a strategy for optimizing solvent selection, evidently this approach requires an estimate of the solubility parameter of the solute. There are several ways to obtain this. One method is suggested by the example; presumably the solubility parameter of the solvent mixture that maximizes solubility is also the solubility parameter of the solute.

(b) The volume fraction is defined as follows:

$$\varphi_1 = \frac{n_1 V_1}{n_1 V_1 + n_2 V_2} \tag{10.27}$$

Suppose $n_1 + n_2 = 1$; from Example 10.1, $x_2 = 0.265$, or $n_2 = 0.265$ and $n_1 = 0.735$. Using these numbers in Eq. (10.27) gives $\varphi_1 = 0.748$. (Note how close φ_1 is to x_1 , because V_1 and V_2 are similar.) Repeating the calculation gives

$$\log x_2 = -0.577 - 0.335 = -0.912$$
$$x_2 = 0.122$$

We therefore predict that x_2 is between 0.064 and 0.122, and we might take the average as our best estimate. The experimental result (Table 10.1) is $x_2 = 0.090$.

Prediction of Aqueous Solubilities. Water is the preferred solvent for liquid dosage forms because of its biological compatibility, but unfortunately many drugs are poorly soluble in water. To be able to predict the aqueous solubility of compounds, even if only approximately, is a valuable capability because it can guide or reduce experimental effort. Water is a highly polar and structured medium in which nonideal behavior is commonly observed, so we must abandon hope that the ideal solubility prediction of Eq. (10.10) will be useful, and even the regular solution theory [Eq. (10.24)] is ineffectual in solving this problem. Effective approaches may be guided by thermodynamic concepts, but they incorporate much empirical (i.e., experimental) content.

Although the ideal solubility equation will not suffice to predict nonelectrolyte solubility in water, the solute–solute interactions responsible for maintaining the crystal lattice must nevertheless be overcome, so Eq. (10.10) will still be applicable as a means of estimating the solute-solute interaction. What must be done in addition is to take account of the solvent–solvent and solvent–solute interactions, because these will in general not offset each other. In a paper that includes a valuable collection of solubility data, Yalkowsky and Valvani (1980) have developed a very useful method based on this approach. They start with Eq. (10.10), which they transform to Eq. (10.11), repeated here:

$$\ln x_2 = -\frac{\Delta S_f(T_m - T)}{RT}$$
(10.28)

They then carry out an analysis of experimental entropies of fusion, reaching these conclusions:

For spherical (or nearly so) molecules: $\Delta S_f = 3.5 \text{ cal mol}^{-1} \text{ K}^{-1}$

For rigid molecules: $\Delta S_f = 13.5 \text{ cal mol}^{-1} \text{ K}^{-1}$

For molecules having n > 5 flexible chain atoms: $\Delta S_f = 13.5 + 2.5 (n-5)$ cal mol⁻¹ K⁻¹

In the following we will use only the result for rigid molecules.

Yalkowsky and Valvani then take the log *P* value of the solute (where *P* is the 1-octanol/water partition coefficient) as an empirical measure of the solution phase nonidealities. They combine this with Eq. (10.28), convert to molar concentration, and apply a small statistical adjustment, finally getting Eq. (10.29) for the calculation of rigid nonelectrolyte molar solubility in water at 25 °C:

$$\log c_2 = -0.0011(t_m - 25) - \log P + 0.54 \tag{10.29}$$

where t_m is the solute melting point in centigrade degrees. For liquid nonelectrolytes, t_m is set to 25, so the first term vanishes. Log *P* may be available from experimental studies, but it may have to be estimated by methods cited in Chapter 7.

Yalkowsky and Valvani applied Eq. (10.29) to solubility data on 167 compounds whose solubilities ranged over nine orders of magnitude, finding that the estimated solubilities agreed with the observed solubilities to within 0.5 log unit for all but eight compounds, and in no case was the error greater than a factor of 10. Equation (10.29) is a very practical solution to the problem of predicting aqueous solubilities.

Amidon and Williams (1982) refined the approach of Yalkowsky and Valvani, achieving better accuracy but at the cost of increased complexity in the equation. Grant and Higuchi (1990) describe alternative methods of calculation that are based on different pathways from the initial to the final state.

Equation (10.10) and equations derived from it, such as Eqs. (10.28) and (10.29), contain the difference $(T_m - T)$, showing that a higher melting temperature is reflecting stronger solute–solute interactions in the solid state. As a general but not precise rule, we may anticipate that very polar molecules (or functional groups) will conduce to strong intermolecular interactions by means of electrostatic forces, which for certain groups may include hydrogen bonding. Thus high molecular polarity tends to be associated with high melting temperature, and higher melting temperatures lead to lower solubilities, at least as they are described by Eq. (10.10).

Now consider the special case of water as a solvent. Water is a very polar solvent and is capable of functioning as a hydrogen bond donor and acceptor. Very polar solute molecules will tend to interact strongly with the solvent water; these are the solvent–solute or solvation interactions that increase solubility. But we have seen that highly polar substances tend to have high melting temperatures, so we are led to the tentative conclusion that melting temperature may be an approximate indicator of the extent of solvent–solute interaction. It follows (still arguing in this approximate mode) that the opposing factors of solute–solute (crystal lattice) and solvent–solute (solvation) interactions are both measured by, or at least indicated by, the same quantity, namely, the melting temperature.⁴ Thus in some degree we may anticipate that these two factors will compensate each other, with the consequence that the solubility will become essentially independent of the melting temperature. But then the first term in Eq. (10.29) will (approximately) vanish, leading to a dependence solely on log *P*.

⁴ Polarity is just one factor controlling the melting temperature. Symmetry is another; the more symmetric the molecule, the higher the melting temperature (when comparing "similar" molecules).

Correlations of log c with log P are well known (Yalkowsky and Valvani 1980; Grant and Higuchi, 1990, Chapter 8). Equations (10.30a) and (10.30b) are such correlations, based on solubility data for compounds having a considerable range of structural features. These equations are to be judged solely by their success in reproducing or predicting solubilities; they are purely empirical.

$$\log c_2 = -\log P - 1.00 \quad \text{(for solids)} \tag{10.30a}$$

$$\log c_2 = -\log P + 0.27$$
 (for liquids) (10.30b)

A comparison of the performance of Eq. (10.29) with Eqs. (10.30) indicates that Eq. (10.29) is slightly superior, but there are some reversals. If the solute melting point is not available, Eq. (10.30a) offers an alternative method of estimation.

Example 10.7. Estimate the aqueous solubility at 25 °C of isophthalic acid, for which log P = 1.73 and whose melting temperature is 346 °C.

With Eq. (10.29), $\log c_2 = -4.72$; with Eq. (10.30a), $\log c_2 = -2.73$. The experimental result is $\log c_2 = -3.40$. Evidently neither Eq. (10.29) nor Eq. (10.30a) yields a fully satisfactory answer in this case. (Although interestingly their average is 3.73, in error by only about a factor of 2 in the solubility c_2 .) Obviously there is scope for improved methods of estimation.

Solubility in Mixed Solvents. If the equilibrium solubility of a solute in water is too low to achieve the desired "target" concentration, a preferred approach in many instances is to incorporate an organic solvent in the aqueous solution, in this way increasing the solubility of the solute. This organic solvent (often called the *cosolvent*) must be miscible with water, at least in the proportions used, and if the solution is to be a dosage form, the cosolvent must be physiologically acceptable. These requirements severely limit the cosolvent selection. But beyond this issue is the matter of the optimal cosolvent concentration in the mixed solvent system of water and cosolvent. As in our treatment of aqueous solubility, we seek methods that are rapid and easy to apply, even though approximate in their accuracy, because the calculation will always be followed by laboratory studies to confirm or refine the numerical estimate.

If the solute and solvent molecules in a solution differ greatly in size, plots of various experimental quantities against solvent composition tend to be more symmetrical when solvent composition is given in volume fraction than in mole fraction (Williamson 1967, p. 44). This observation forms the basis of a model proposed by Yalkowsky and Rubino (1985). For these three-component systems, let water be component 1, the cosolvent component 2, and the solute component 3. The molar solubility of solute in water is written $(c_3)_1$ and its molar solubility in pure cosolvent as $(c_3)_2$. In solvent of any composition the solute solubility is written c_3 . Then the Yalkowsky–Rubino model becomes

$$\log c_3 = \varphi_1 \log(c_3)_1 + \varphi_2 \log(c_3)_2 \tag{10.31}$$

where φ_1 and φ_2 are the volume fractions of water and cosolvent, respectively. Since $\varphi_1 + \varphi_2 = 1$, an equivalent form of Eq. (10.31) is

$$\log c_3 = \varphi_2 [\log(c_3)_2 - \log(c_3)_1] + \log(c_3)_1$$
(10.32)

This equation predicts that log c_3 will be a linear function of φ_2 .

Equation (10.31) is a postulate. It can be described as a linear combination model, or as a weighted average; that is, $\log c_3$ is postulated to be an average of $\log (c_3)_1$ and $\log (c_3)_2$, each of these making a contribution according to (weighted by) its volume fraction.

The procedure for testing and using this model is simple. On a graphical scale of φ_2 , one plots log $(c_3)_1$ at $\varphi_2 = 0.0$ and log $(c_3)_2$ at $\varphi_2 = 1.0$. These points are connected by a straight line, which is the graphical representation of Eq. (10.32). A test of the model consists of plotting experimental solubilities at intermediate values of φ_2 to learn how well they agree with the straight-line prediction. Alternatively, if (as is usually the case) such data are not available, the model is assumed to be (approximately) valid, and that value of φ_2 is read off the line that will achieve a desired target solubility. It is not necessary to carry this operation out graphically, because by rearrangement of Eq. (10.32) we obtain

$$\varphi_2 = \frac{\log c_3 - \log(c_3)_1}{\log(c_3)_2 - \log(c_3)_1} \tag{10.33}$$

With this equation the required volume fraction of cosolvent can be calculated, according to this model.

Figure 10.3 shows solubility data for the system water (1)–ethanol (2)–naphthalene (3) (LePree et al., 1994). The straight line connecting the extreme points constitutes the linear combination model, Eq. (10.32); the points are experimental. Obviously the points do not describe a straight line, so in this sense, and for this system, the model does not appear to be valid. On the other hand, as an approximate guide to the dependence of solubility on solvent composition it may be helpful to the experimentalist, and it is in this sense that the model should be judged. It is not a precise description of physicochemical behavior, but rather is a useful tool in formulation development.

Example 10.8. Propose a water/ethanol mixed solvent composition that will dissolve 2.5 mg mL⁻¹ of naphthalene. The solubility of naphthalene in water is 2.14×10^{-4} M, and in ethanol it is 0.675 M.

The target concentration of 2.5 mg mL⁻¹ is equivalent to 2.5 g L⁻¹. The molecular weight of naphthalene is 128.2, so the molar target concentration c_3 is 0.0195 M, or $\log c_3 = -1.71$. From the given data we have $\log (c_3)_1 = -3.67$ and $\log (c_3)_2 = -0.17$. Applying Eq. (10.33), we obtain

$$\begin{split} \varphi_2 = \frac{-1.71 - 3.67}{-0.17 - (-3.67)} \\ = 0.56 \end{split}$$

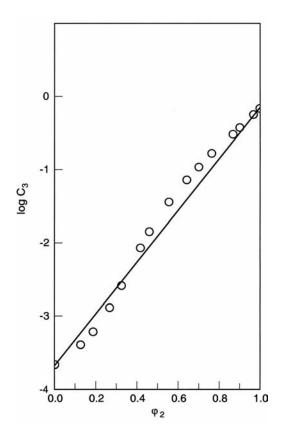


Figure 10.3. The linear combination model for naphthalene solubility in aqueous ethanol solutions.

Thus we predict that a volume fraction of 0.56 ethanol will dissolve the target concentration. This result could also have been obtained by reading from the straight line of Fig. 10.3. It is interesting to note, from the experimental points in Fig. 10.3, that a volume fraction $\varphi_2 = 0.51$ will actually dissolve the target concentration.

More accurate models of solvent effects are available, but these require much experimental effort and are computationally more elaborate (see Khossravi and Connors, 1992).

PROBLEMS

- **10.1.** The melting point of benzoic acid is $122.4 \,^{\circ}$ C, and its heat of fusion is $4.44 \,\text{kcal mol}^{-1}$. Calculate its ideal solubility at $25 \,^{\circ}$ C.
- **10.2.** From the data in Example 10.2, convert the mole fraction solubilities to molar solubilities, construct the van't Hoff plot, and evaluate the heat of solution.

- **10.3.** Derive an equation relating the molar solubility of calcium phosphate to its solubility product, and calculate its molar solubility.
- **10.4.** A solution containing NaBr, NaCl, and NaI is titrated with silver nitrate solution. Predict the order in which the silver halides will precipitate.
- **10.5.** Predict the solubility of iodine in carbon tetrachloride at 25 °C. The melting point of iodine is 113.6 °C, its heat of fusion is 3.71 kcal mol⁻¹, its molar volume is 59 cm³, and its solubility parameter is 14.1. The solubility parameter of carbon tetrachloride is 8.6.
- **10.6.** Predict the molar solubility of progesterone in water at 25 °C. The melting point of progesterone is 131 °C and its log *P* value is 3.87.
- **10.7.** The solubility of naphthalene in water at $25 \,^{\circ}$ C is 2.14×10^{-4} M, and its solubility in dimethylsulfoxide (DMSO) is 1.920 M. Estimate the mixed solvent composition required to dissolve $4 \,\text{mg mL}^{-1}$ of the solute.
- **10.8.** The salt form of warfarin has a solubility in water of 2×10^{-3} g/mL at 25 °C. Calculate the standard free energy of solubilization of the warfarin salt. Assume activity = concentration. Warfarin salt molecular weight = 330 g/mol.

11

SURFACES AND INTERFACES

Up to this point in our study of thermodynamics we have dealt with bulk phases only; that is, we have ignored possible influences of the surfaces of these phases. This attitude is justified when the surface constitutes a very small fraction of the system. In some circumstances, however, the surface:volume ratio of the system becomes relatively large, and then the properties of the surface may dominate the behavior of the system. In the pharmaceutical field, the dosage forms called *emulsions, suspensions,* and *foams* exemplify such circumstances; collectively these are known as *disperse systems*; emulsions are dispersions of liquid droplets in an immiscible liquid, suspensions are dispersions of solid particles in a liquid, and foams are dispersions of gases in liquids.

Let us begin with a clarification of terminology. Strictly speaking, the boundary between any two phases constitutes an interface, but it is conventional to call this interface a "surface" when one of the phases is a vapor or gas (especially, and usually, when it is air). We therefore have these identifications:

Surfaces	Interfaces
Solid–gas	Solid-solid
Liquid–gas	Solid-liquid
	Liquid–liquid

Despite these definitions, it is common to use the word "surface" in a generic sense to embrace all such phase boundaries.

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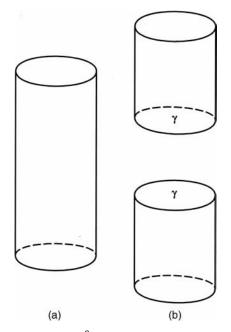


Figure 11.1. (a) A column of liquid 1 cm² in cross section; (b) the column separated to create two surfaces, each with area 1 cm² and surface tension γ .

11.1. THERMODYNAMIC PROPERTIES

Surface Tension and Interfacial Tension. Let us carry out this thought experiment. Figure 11.1a illustrates a column of a liquid, constituting a single phase and having a cross-sectional area of 1 cm^2 . Now imagine this column to be pulled apart cleanly into two parts, as shown in Fig. 11.1b. The result of this imagined experiment has been to create two surfaces of the liquid, each of area 1 cm^2 .

Energy was required to create these surfaces, because molecules had to be pulled apart and the forces of intermolecular interaction had to be overcome. The work of carrying out this process is called the *work of cohesion*, w_c , and it is set equal to the energy of the surfaces that were created. (Because we can neither create nor destroy energy, the work w_c done on the system is now possessed by the system in the form of surface energy.) We write

$$w_c = 2\gamma \tag{11.1}$$

where γ is the surface energy per square centimeter and is called the *surface tension*. (The factor 2 appears because 2 cm² of surface was created in Fig. 11.1.)

We can now connect this concept of surface energy to other kinds of energy by recalling (Chapter 1) that work (or energy) can be expressed as the product of an

Solvent	$\gamma(dyn \ cm^{-1})$	Solvent	$\gamma(\mathrm{dyn}\ \mathrm{cm}^{-1})$
<i>n</i> -Hexane	17.9	Acetone	22.9
Cyclohexane	19.8	Ethanol	21.8
Benzene	28.2	Methanol	22.4
Diethyl ether	16.5	Acetonitrile	28.5
Chloroform	26.5	Glycerol	62.5
Ethyl acetate	23.2	Dimethylsulfoxide	42.8
<i>n</i> -Butyl alcohol	24.2	Water	71.8
n-Propyl alcohol	23.4	Mercury	485.5

Table 11.1. Surface tensions at 25 °C

intensity factor and a capacity factor. For surface work or energy we have

Surface energy = Surface tension \times Change in surface area

$$\Delta G = \gamma \Delta A \tag{11.2}$$

If ΔA is positive (surface area is created), ΔG is positive and the process is nonspontaneous; work must be done to create new surface area. If ΔA is negative, ΔG is negative and the process is spontaneous. The surface energy is identified with the change in Gibbs free energy if the temperature and pressure are constant.

Table 11.1 lists the surface tensions of some liquids. First consider the units of γ , namely energy per unit area. Thus in SI units, J m⁻² is a correct designation for the units of γ . Moreover, since work (energy) is the product of force and length and 1 J = 1 N m, the units N m⁻¹ are also acceptable. In the older cgs system, in which most of the literature values of γ are recorded, the corresponding units are erg cm⁻² and dyn cm⁻¹. However, in order that the numerical values of γ be identical in the cgs and SI systems, the SI units are multiplied by 10³. Thus we can state the surface tension of water in these equivalent units:

$$\gamma = 71.8 \frac{\text{erg}}{\text{cm}^2} = 71.8 \frac{\text{dyn}}{\text{cm}} = 71.8 \frac{\text{mJ}}{\text{m}^2} = 71.8 \frac{\text{mN}}{\text{m}}$$

Inasmuch as the surface tension is a measure of the energy required to create unit area of surface, we might expect γ to be larger for solvents having stronger intermolecular forces of interaction, and generally we see that this expectation is borne out. Very polar molecules and those capable of hydrogen bonding tend to have higher values of γ than do nonpolar substances.

As the temperature of a liquid is increased, the liquid acquires more thermal energy, and so less additional energy needs to be supplied to create new surface. As a consequence, the surface tension is smaller at higher temperatures.

Another point of view may be helpful in visualizing the physical nature of the surface tension. The properties of a system consisting of a liquid in contact with its vapor do not change discontinuously at the surface, but rather change in a smooth continuous fashion. Another way to say this is to point out that the surface is not a mathematical boundary, but is a region having a thickness of several molecular diameters. Consider the density as a property that varies from a relatively high value in the bulk liquid (expressed as g mL⁻¹ or as number of molecules per unit volume) to a

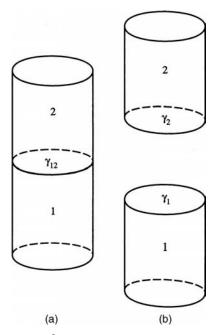


Figure 11.2. (a) A column of 1-cm² cross-sectional area of phases 1 and 2 in contact; (b) the phases are separated to create two surfaces; one of liquid 1, the other of liquid 2.

very low value in the vapor. Evidently the density will have an intermediate value in the surface region. Taking a slice coplanar with the surface, then, the number of molecules per unit area in the surface is smaller than in the bulk. In other words, the surface is analogous to an extended spring; energy is required to create the surface (or to stretch the spring), and this energy is manifested as a tension in the plane of the surface; this is the surface tension (Fowkes 1964).

Now we will carry out another thought experiment in the style of Fig. 11.1, this one as shown in Fig. 11.2. Here we have a column of 1-cm² cross-sectional area, but consisting of two immiscible liquid phases, 1 and 2, in contact. We now imagine the phases separated at their boundary.

According to our earlier analysis, we can expect an amount of work to be required equal to the sum $\gamma_1 + \gamma_2$, because 1 cm² of surface of each 1 and 2 is created in this process. But there is a further factor to consider, because in the initial state of the system there existed an interface between 1 and 2, and this interface itself possesses surface (or interface) energy, labeled γ_{12} . In the final state of the system this interface no longer exists, so neither does its energy, which has, in effect, been applied to defray the energetic cost of creating the two new surfaces. The work involved in passing from the initial state in Fig. 11.2a to the final state in Fig. 11.2b is called the *work of adhesion*, w_a , and is given by

$$w_a = \gamma_1 + \gamma_2 - \gamma_{12} \tag{11.3}$$

Liquids	$\gamma(dyn \ cm^{-1})$	Liquids	$\gamma(\mathrm{dyn}\ \mathrm{cm}^{-1})$
Water/mercury Water/n-octane Water/benzene Mercury/benzene	375 50.8 35.0 375	Water/n-hexane Water/CCl ₄ Water/ether	51.1 45 10.7

Table 11.2. Interfacial tensions at 20 °C

The quantity γ_{12} is the *interfacial tension*. It has the same units and the same physical significance as the surface tension. Table 11.2 gives some interfacial tensions. It must be realized that in such systems the two liquids are mutually saturated.

The interfacial tension can be measured, or it can be calculated with reasonable accuracy by means of

$$\gamma_{12} = \gamma_1 + \gamma_2 - 2(\gamma_1^d \gamma_2^d)^{1/2}$$
(11.4)

where γ_1^d and γ_2^d are the London dispersion force contributions to γ_1 and γ_2 (Fowkes 1964). The dispersion force is an attractive force (sometimes called the van der Waals force) between all molecules. For nonpolar molecules such as saturated hydrocarbons, it is the only attractive force, so $\gamma^d = \gamma$ for such liquids. For mercury γ^d is 200 dyn cm⁻¹ and for water $\gamma^d = 21.8$ dyn cm⁻¹. Equation (11.4) incorporates the geometric mean approximation of regular solution theory (Section 10.5); see Eq. (10.23) in particular.

Example 11.1. Estimate the interfacial tension at the water/*n*-hexane interface. We use Eq. (11.4):

$$\gamma_{12} = 71.8 + 17.9 - 2(21.8 \times 17.9)^{1/2} = 50.2 \text{ dyn cm}^{-1}$$

The experimental value (Table 11.2) is 51.1 dyn cm^{-1} .

Because a surface is of relatively high energy compared to the bulk, there is a thermodynamic driving force for minimization of surface area. A sphere is the geometric form having the minimum surface:volume ratio; thus under the influence of surface tension alone, units of matter will tend to assume a spherical shape, as with droplets of liquids. This tendency may be opposed by other forces, such as the gravitational force. The trend to surface minimization also accounts for the tendency of a dispersion of liquid drops in a liquid (an emulsion) to coalesce into two bulk phases.

Spreading of Liquids on Liquids. Picture a small volume of liquid 2 placed on the planar surface of liquid 1. What will happen? Will it just sit there as a globule (flattened by gravity into a "lens" shape), or will it spread out into a very thin film? This question can be answered by considering the work of adhesion (which measures the attraction between 1 and 2) and the work of cohesion (which measures the attraction within 2) (Adamson 1960, p. 107; Bummer 2000). The *spreading coefficient* $S_{2/1}$ for the spreading of 2 on 1 is defined as

$$S_{2/1} = w_a - w_{c(2)} \tag{11.5}$$

where $w_{c(2)}$ is the work of cohesion of liquid 2. Similarly, the spreading coefficient $S_{1/2}$ for the spreading of 1 on 2 is

$$S_{1/2} = w_a - w_{c(1)} \tag{11.6}$$

Incorporating Eqs. (11.1) and (11.3) into these definitions gives

$$S_{2/1} = \gamma_1 - (\gamma_2 + \gamma_{12}) \tag{11.7}$$

$$S_{1/2} = \gamma_2 - (\gamma_1 + \gamma_{12}) \tag{11.8}$$

Let us try to anticipate the possible outcomes of numerical calculations of spreading coefficients. If $w_a \gg w_{c(2)}$, evidently the attraction between 1 and 2 is greater than the self-attraction of liquid 2, so the spreading coefficient will be positive and we may expect 2 to spread on 1. Compare the rather extreme cases in Example 11.2.

Example 11.2. Calculate the spreading coefficients of water on mercury and of mercury on water and interpret the results. (Ignore the density difference!)

Let mercury be liquid 1 and water liquid 2. From Eqs. (11.7) and (11.8) and data in Tables 11.1 and 11.2, we have

$$S_{2/1} = 485.5 - (71.8 + 375) = +38.7$$

 $S_{1/2} = 71.8 - (485.5 + 375) = -788.7$

Thus water will spread on mercury, but mercury will not spread on water.

In general a liquid of lower surface tension will spread on a liquid of higher surface tension. However, spreading does not continue indefinitely, because the phenomenon in which liquid 2 spreads on liquid 1 creates a new surface tension, leading to a situation in which the spreading coefficient is negative and addition of further 2 leads to formation of a lens rather than continued spreading.

Wetting of Solids by Liquids. Consider the system of a drop of liquid L on a planar surface of a solid S, where the system is at equilibrium with the vapor V of the liquid. The shape of the drop can be specified in terms of the angle θ that a tangent makes to the surface of the liquid at its point of contact with the solid. This angle is called the *contact angle*. It can be measured experimentally.

A simple relationship can be obtained connecting θ with the system surface and interfacial tensions. Figure 11.3 shows the construction, the lengths of the arrows (vectors) being proportional to the indicated tensions. At equilibrium, the force associated with tension γ_{SV} is exactly balanced by the sum of the forces due to γ_{SL} , and the component of γ_{LV} lying in the same plane and direction as γ_{SL} . This component has the magnitude *ab* in the figure, and is seen to be equal to $\gamma_{LV} \cos \theta$. Thus Eq. (11.9) can be written.

$$\gamma_{\rm SV} = \gamma_{\rm SL} + \gamma_{\rm LV} \cos\theta \tag{11.9}$$

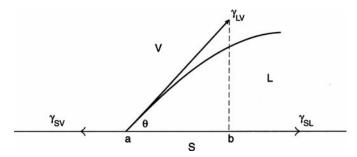


Figure 11.3. Contact angle θ of a solid on a solid.

The tension γ_{LV} is, of course, simply the surface tension of the liquid. γ_{SL} is the liquid/ solid interfacial tension, and γ_{SV} is the surface tension of the solid.

If the liquid wets the solid, the liquid spreads out on the surface, reducing the angle θ ; commonly a liquid that wets a solid is considered to have a contact angle of 0°. If $\theta > 90^\circ$, the liquid does not wet the solid. Examination of Fig. 11.3 shows that θ will be reduced under the influence of these factors: a larger value of γ_{SV} , or smaller values of γ_{LV} and of γ_{SL} . For a given solid, wetting will be favored by liquids with lower surface tensions. For a given liquid, wetting will be favored by solids with higher surface tensions.¹

Pressure Difference across Spherical Surfaces. A *bubble* is a region of vapor (often including air) surrounded by a thin film of liquid. A *cavity* is a hole (containing vapor) in a liquid. A *droplet* is a small volume of liquid. Bubbles, cavities, and droplets are approximately spherical; they would be spheres if surface forces only were active, but other forces distort their shape. We will assume that they are spheres. Bubbles actually have double walls, an inner film and an outer film, so they have twice the surface area of a sphere their size. Cavities and droplets have single surfaces.

An interesting property of these systems has been known for a long time. Consider a spherical system (we may think of a bubble, while neglecting the double wall), of radius r. There is a driving force for reducing r as a consequence of two factors. One of these is the external pressure on the bubble, and the other is the surface energy. At equilibrium this combined force is just balanced by that due to the internal pressure. We seek the condition of equilibrium balance. First we write the external and internal work terms, then differentiate these with respect to r (giving forces), and then equate the forces.

The external work is a sum of a work of expansion contribution and a surface work term:

$$dw_{\text{ext}} = P_{\text{ext}} \, dV + \gamma \, dA$$

¹ Solids with higher surface tensions are said to be higher energy solids. Such solids tend to possess polar functional groups. A solid such as paraffin, which is a saturated hydrocarbon, is a low-energy solid. Silica (glass) is of higher energy.

The force is $dw_{\rm ext}/dr$, or

$$\frac{dw_{\text{ext}}}{dr} = P_{\text{ext}}\frac{dV}{dr} + \gamma \frac{dA}{dr}$$

For a sphere $V = (4/3)\pi r^3$ and $A = 4\pi r^2$, so we get

$$\frac{dw_{\text{ext}}}{dr} = 4\pi r^2 P_{\text{ext}} + 8\pi r\gamma \qquad (11.10)$$

The internal work is $dw_{int} = P_{int}dV$, giving in the same way

$$\frac{dw_{\rm int}}{dr} = 4\pi r^2 P_{\rm int} \tag{11.11}$$

Equating Eqs. (11.10) and (11.11) yields

$$P_{\rm int} = P_{\rm ext} + \frac{2\gamma}{r} \tag{11.12}$$

According to this surprising result, the pressure inside the bubble or cavity is greater than that outside by the amount $2\gamma/r$.

Example 11.3. Calculate the pressure difference inside and outside a cavity in water whose radius is 0.01 mm.

We use eq. (11.12) with $r = 10^{-5}$ m and $\gamma = 71.8$ mN m⁻¹, finding

$$P_{\rm int} - P_{\rm ext} = 1.436 \times 10^7 {\rm mN} {\rm m}^{-2}$$

This quantity can be converted to more familiar units by using the identities

$$1 \text{ Pa} = 1 \text{ Nm}^{-2}$$

1 atm = 101325 Pa

The result is 0.142 atm.

For a planar surface, r is infinite, so the pressure difference vanishes.

Two practical consequences of Eq. (11.12), which we will not develop here, are as follows: (1) The vapor pressure of very small droplets is greater than that of large droplets, so the small droplets evaporate; and (2) the solubility of very small particles is greater than that of larger particles (Glasstone 1947, p. 247).

11.2. ADSORPTION

The Surface Phase. To the eye a surface or interface at equilibrium appears to be a quiescent, two-dimensional element; but since matter is atomic or molecular in

structure, we have to consider the nature of the surface in molecular terms. We know that molecules possess thermal energy; in particular, they possess translational energy and are incessantly in motion. Thus at the boundary between a liquid and its vapor, molecules are passing from the liquid state to the vapor state, and vice versa. If the system is at thermodynamic equilibrium, these two rates are equal, so the net transport between the phases is zero, but the extent of molecular traffic is prodigious. The molecules within the liquid phase are also in thermal motion. We noted earlier that the surface region extends several molecular diameters in thickness into the liquid phase. This surface region or phase cannot be sharply demarcated at either of its boundaries (with the bulk liquid or with the vapor) because of the molecular motion. Far from being a quiescent two-dimensional construction, the surface region is a turbulent three-dimensional "interphase."

Molecules in the surface region experience a force field different from that experienced by those in the bulk. A molecule within a bulk liquid is enveloped in a homogeneous force field (on average), because its molecular environment is the same in every direction. Consequently, the forces on the molecule are everywhere balanced out, so it experiences no net force. At the surface, on the other hand, the molecule's environment is disymmetric; it experiences more intermolecular forces from the bulk liquid side than from the vapor side. (This disymmetric nature of the forces is treated by some authors as the source of the surface tension.)

Notwithstanding our recognition that the surface region is ill-defined geometrically on the molecular scale, we find it convenient, and also justifiable, as we will see, to treat the surface as a two-dimensional mathematical abstraction. First, treating the surface region realistically as a phase, we may conclude from standard thermodynamics that the chemical potentials of all constituents in a system at equilibrium are identical in all phases, including the surface phase. Next, turning to the surface region treated abstractly as a mathematical surface dividing two phases, refer to Fig. 11.4a, which depicts two phases, α and β , divided by surface *s*, whose location is arbitrarily chosen to separate chemically bulk α from chemically bulk β ; that is, *s* is located *somewhere* within the surface region. Suppose the system consists of two components, 1 and 2. The element shown in the figure has cross-sectional area *A*. If n_1 and n_2 are the total numbers of moles of 1 and 2 contained between the limits *aa* and *bb*, respectively, we can write these mass balances:

$$n_1^{\alpha} + n_1^{\beta} + n_1^s = n_1 \tag{11.13a}$$

$$n_2^{\alpha} + n_2^{\beta} + n_2^s = n_2 \tag{11.13b}$$

Now since the molar concentration of 1 in phase α is $c_1^{\alpha} = n_1^{\alpha}/V^{\alpha}$, where $V^{\alpha} = xA$, and so on, we expand Eq. (11.13) to

$$xAc_1^{\alpha} + yAc_1^{\beta} + n_1^s = n_1 \tag{11.14a}$$

$$xAc_2^{\alpha} + yAc_2^{\beta} + n_2^s = n_2 \tag{11.14b}$$

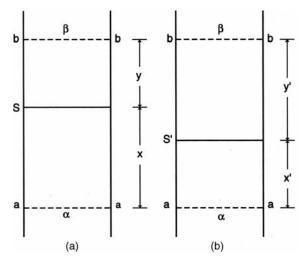


Figure 11.4. (a) Two-phase system, with *aa* and *bb* located within bulk phases α and β and *s* the dividing boundary within the surface phase; (b) the same as (a) but with the dividing boundary moved to *s'*.

We divide by *A* and define $\Gamma = n^s/A$, obtaining

$$xc_1^{\alpha} + yc_1^{\beta} + \Gamma_1 = \frac{n_1}{A}$$
(11.15a)

$$xc_2^{\alpha} + yc_2^{\beta} + \Gamma_2 = \frac{n_2}{A} \tag{11.15b}$$

This quantity Γ is called the *surface excess* of the specified constituent. Referring to Eq. (11.13a), we can write

$$\Gamma_1 = \frac{n_1}{A} - \frac{n_1^{\alpha} + n_1^{\beta}}{A}$$

which shows that Γ_1 can be zero, positive, or negative. If Γ_1 , is positive, an excess of constituent 1 (relative to the bulk) is located at the surface, whereas a negative value of Γ_1 , indicates a deficiency at the surface (although it is still called the surface excess).

Returning to our mathematical treatment, let us suppose that phase β is a vapor phase and that c^{β} is negligible. We then have

$$xc_1^{\alpha} + \Gamma_1 = \frac{n_1}{A} \tag{11.16a}$$

$$xc_2^{\alpha} + \Gamma_2 = \frac{n_2}{A} \tag{11.16b}$$

Next we construct Fig. 11.4b for the same system, altering the construction only by displacing boundary s to position s' (but still within the surface region).

Repeating the treatment gives the following, by analogy with Eq. (11.16):

$$x'c_1^{\alpha} + \Gamma'_1 = \frac{n_1}{A}$$
(11.17a)

$$x'c_2^{\alpha} + \Gamma'_2 = \frac{n_2}{A}$$
(11.17b)

Since n_1 and n_2 are fixed and independent of the choice of *s* or *s'*, we write from Eqs. (11.16) and (11.17)

$$xc_1^{\alpha} + \Gamma_1 = x'c_1^{\alpha} + {\Gamma'}_1 \tag{11.18a}$$

$$xc_2^{\alpha} + \Gamma_2 = x'c_2^{\alpha} + \Gamma'_2$$
 (11.18b)

These two equations combine to give

$$\frac{\Gamma_1' - \Gamma_1}{c_1^{\alpha}} = \frac{\Gamma_2' - \Gamma_2}{c_2^{\alpha}}$$

which further rearranges to

$$\Gamma'_1 c_2^{\alpha} - \Gamma'_2 c_1^{\alpha} = \Gamma_1 c_2^{\alpha} - \Gamma_2 c_1^{\alpha} = \text{constant}$$
(11.19)

That is, the left-hand portion of Eq. (11.19) refers solely to dividing line *s'*, whereas the right-hand portion refers solely to dividing line *s*, and these are equal. If we created yet more dividing surfaces (within the surface region), say, *s''*, *s'''*, and so on, the corresponding terms would all be the same. The conclusion is that the choice of dividing surface is arbitrary and irrelevant, provided that it lies within the surface region. Of course, the numerical values of the surface excesses depend on the location of the dividing line, so practical issues may lead to a preferred location, but there is no fundamental issue involved (Adamson 1960, pp. 73–79).

This consideration leads to a very convenient simplification in our next development, because, if we can place the dividing surface as we wish, why not place it so that one of the surface excesses is equal to zero?

The Gibbs Adsorption Equation. We are going to develop a famous equation of Gibbs by following a path used earlier in analyzing an open system without consideration of a surface phase. Where we earlier wrote the free energy as the general function [Eq. (3.19)]

$$G = f(T, P, n_1, n_2 \ldots)$$

we now expand the description, writing

$$G = f(T, P, \gamma n_1, n_2, \ldots)$$
 (11.20)

In order to simplify the treatment, we will consider that the temperature and pressure are constant and that only two components are present. Carrying through the earlier development gives the surface phase analog to the Gibbs–Duhem equation [Eq. (3.28)]

$$Ad\gamma + n_1^s d\mu_1 + n_2^s d\mu_2 = 0 \tag{11.21}$$

where μ_1 and μ_2 are the chemical potentials of the two components in the surface phase (but we do not need to distinguish them as surface potentials because at equilibrium the potential is the same everywhere).

Now dividing through by A and recalling the definition of surface excess gives

$$d\gamma + \Gamma_1 d\mu_1 + \Gamma_2 d\mu_2 = 0 \tag{11.22}$$

Next we make use of the demonstration of the preceding analysis, where we saw that the dividing surface can be arbitrarily placed, and we choose to place it such that Γ_1 , the surface excess of component 1 (which we can take as the solvent), is zero. Thus

$$d\gamma = -\Gamma_2 d\mu_2 \tag{11.23}$$

At low solute concentrations, where the solute activity coefficient may be taken as essentially unity, we have

$$\mu_2 = \mu_0 + RT \ln c_2 \tag{11.24}$$

Combining Eqs. (11.23) and (11.24) gives the *Gibbs adsorption equation*²:

$$\Gamma_2 = -\frac{c_2}{RT} \left(\frac{d\gamma}{dc_2} \right) \tag{11.25}$$

The great value of the adsorption equation is that it connects the readily measurable quantities on the right-hand side with the somewhat abstract concept of the surface excess. Qualitatively, we can see that if the surface tension of a solution of solute 2 in solvent 1 decreases as the concentration of 2 increases, then Γ_2 is positive, meaning that solute 2 is more concentrated at the surface than in the bulk of the solution. Substances that exhibit positive surface excesses, and therefore produce surface tension decreases, are called *surface active agents*, or *surfactants*. Soaps and detergents are surfactants. Such agents tend to be fairly large molecules, having one end

² See Gibbs (1876, 1878). Equations (11.23) and (11.25) are obtained on pp. 232 and 235, respectively, of the Dover (1961) edition. Much of Gibbs' massive work on thermodynamics is difficult for the modern reader to follow, in part because his symbolism differs from our usage, but two of the symbols Gibbs introduced, μ for chemical potential and Γ for surface excess, are still used, so his surface equations are more accessible. Incidentally, Gibbs would write Eqs. (11.23) and (11.25) with the symbol $\Gamma_{2(1)}$, thus explicitly noting that this is the value of Γ_2 given that Γ_1 is set to zero.

of the molecule polar (and therefore attracted to a polar solvent like water) and the other end nonpolar (and therefore preferring to reside in a nonpolar phase). Even quite small molecules with these molecular attributes exhibit positive surface excesses. Organic solvents such as alcohols, acetone, acetonitrile, and dimethylsulfoxide decrease the surface tension of water.³ Inorganic salts, on the other hand, increase the surface tension of water.

Equation (11.25) can be written in the equivalent form

$$\Gamma_2 = -\frac{1}{RT} \left(\frac{d\gamma}{d \ln c_2} \right) \tag{11.26}$$

or

$$d\gamma = -RT \,\Gamma_2 d \ln c_2 \tag{11.27}$$

Of course, Γ_2 is a function of concentration, so simple integration is unwarranted, but Eqs. (11.26) and (11.27) suggest that plots of γ against $\ln c_2$ may be fruitful forms of data analysis. Such plots are curved, but their slopes (tangents to the curve) yield estimates of Γ_2 as a function of concentration (Bummer 2000).

Adsorption lsotherms. Adsorption is that process in which a substance develops a positive surface excess at a surface or interface. In our present discussion we will restrict attention to those systems in which a component of a gas or vapor phase is adsorbed to a solid surface, or in which a component of a liquid solution is adsorbed to a solid surface. The substance that is adsorbed is called the *adsorbate*, ⁴ and the solid is called the *adsorbent*. It might seem that the Gibbs surface excess Γ is the quantity that should be sought experimentally in a study of adsorption, but usually some more accessible measure of the extent of adsorption is determined. A graph of this measure of extent of adsorption against the partial pressure of the adsorbate in the vapor (for vapor–solid systems) or against its solution concentration (for liquid–solid systems) is called an *adsorption isotherm*. (The term *isotherm* simply means that the temperature is held constant.)

Many shapes of adsorption isotherms have been found experimentally, but we will consider only one of these in detail. This function, called the *Langmuir adsorption isotherm*, is widely observed, its physical basis is simple, and its mathematics turn out also to be applicable to other types of systems involving noncovalent interactions, such as enzyme–substrate complexing and molecular complex formation. To focus attention, suppose that we have a two-component solution of solvent 1 and solute

³ Equations have been described with which the surface tensions of binary aqueous–organic solvent mixtures can be modeled over the entire composition range; see Connors and Wright (1989) and Khossravi and Connors (1993).

⁴ In *adsorption* the adsorbate adheres to the surface of the solid. In *absorption* the solute is taken up throughout the volume of the solid. When the nature of the process is unknown, the generic term *sorption* may be used.

(adsorbate) 2 in contact, and at equilibrium, with solid. The process can be portrayed as

Adsorbate in solution
$$\stackrel{adsorption}{\underbrace{\qquad}}$$
 Adsorbate on surface

Our approach, oddly, is a kinetic one. At equilibrium the rate at which molecules of adsorbate adsorb to the surface is just equal to the rate at which they desorb from the surface. We write these expressions for these rates:

Rate of adsorption
$$= k_a c_2 A_f$$
 (11.28)

Rate of desorption
$$= k_d A_b$$
 (11.29)

In these equations, c_2 is the solution concentration of adsorbate; k_a and k_d are rate constants for adsorption and desorption, respectively; A_f is the solid surface area per unit mass (usually per gram) that is "free," namely, unoccupied by adsorbate molecules; and A_b is the solid surface area per unit mass that is "bound," or occupied by adsorbate molecules. Equation (11.28) postulates that the rate of adsorption is directly proportional to the concentration (actually activity) of adsorbate and to the amount of space available for adsorption on the surface. Equation (11.29) says that the rate of desorption is directly proportional to the extent of surface already covered by adsorbate molecules. It is important to recognize that, for any given system, Eqs. (11.28) and (11.29) a priori may or may not be valid; they constitute hypotheses, to be justified by subsequent testing against experimental data.

At equilibrium these two rates are equal. Let us work out the ramifications of this equality:

$$k_a c_2 A_f = k_d A_b \tag{11.30}$$

Define the total surface area per unit mass as A_t , so

$$A_t = A_f + A_b \tag{11.31}$$

Eliminating A_f , defining $K = k_a/k_d$ (K is an equilibrium constant), and rearranging give

$$Kc_2 = \frac{A_b}{A_t - A_b} \tag{11.32}$$

We now define θ , the *degree of saturation*, by

$$\theta = \frac{A_b}{A_t} \tag{11.33}$$

Combining Eqs. (11.32) and (11.33) gives

$$Kc_2 = \frac{\theta}{1 - \theta} \tag{11.34}$$

which is one form of the Langmuir adsorption isotherm. More commonly, it is encountered in this rearranged form:

$$\theta = \frac{Kc_2}{1 + Kc_2} \tag{11.35}$$

Equation (11.35) is equivalent to

$$A_b = \frac{KA_t c_2}{1 + Kc_2} \tag{11.36}$$

Now let σ be the area occupied per molecule on the surface, so A_b/σ is the number of molecules adsorbed per gram of solid, and $n_b = A_b/\sigma N_A$ (where N_A is Avogadro's number) is the number of moles of adsorbate bound per gram of solid. Similarly, $A_t/\sigma N_A n_{\text{max}}$ is the maximum number of moles that can be bound per gram of solid. Making substitutions in Eq. (11.36) gives finally

$$n_b = \frac{Kn_{\max}c_2}{1+Kc_2} \tag{11.37}$$

This is a very practical form of the Langmuir adsorption isotherm. Experimentally, we know c_2 , the solution concentration of adsorbate, and we measure n_b , the number of moles of adsorbate per gram of solid.

Two practical problems remain. We wish to test the hypothesis against the data (i.e., we wish to establish whether the system is described by the Langmuir isotherm), and, if Eq. (11.37) is in fact descriptive of the system, we wish to evaluate the model parameters K and n_{max} . Both problems have traditionally been solved by rearranging Eq. (11.37) to this form:

$$\frac{c_2}{n_b} = \frac{c_2}{n_{\max}} + \frac{1}{Kn_{\max}}$$
(11.38)

This is the equation of a straight line. If the Langmuir isotherm is obeyed, a plot of c_2/n_b against c_2 should be linear, with slope = $1/n_{max}$, and intercept = $1/Kn_{max}$. Thus $n_{max} = 1$ /slope, and K = slope/intercept.

From Eq. (11.37) we can analyze Langmuirian behavior. If c_2 is very low, so that $Kc_2 \ll 1$, then $n_b \approx Kn_{\max}c_2$; the isotherm is nearly linear at very low adsorbate concentrations. At relatively high c_2 values, where $Kc_2 \gg 1$, then $n_b \approx n_{\max}$; the extent of adsorption reaches a maximum value and becomes independent of the solution concentration. The physical interpretation of this result is that the surface is completely covered with adsorbate molecules and that no further adsorption can occur because no more solid surface is accessible. This is interpreted to mean that the surface is covered with a layer of adsorbate one molecule thick (a monomolecular layer). This is one of the physical implications of the Langmuir isotherm. Another implication is that adsorption "sites" on the solid surface are independent in the sense that the energy of adsorbate molecule to a surface site is independent of whether an adjacent site is already occupied.

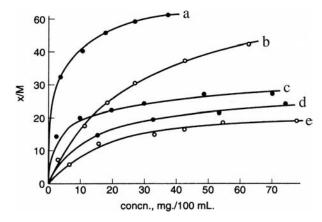


Figure 11.5. Plots of milligrams of dye absorbed per gram of cornstarch [x(M)], versus milligrams of dye per 100 mL of solution at equilibrium for FD&C Red No. 3 (a), FD&C Blue No. 2 (b), Ext. D&C Red No. 15 (c), FD&C Yellow No. 5 (d), and FD&C Green No. 1 (e). [Reproduced with permission from Zografi and Mattocks (1963).]

These physical restrictions are rather severe, so it may seem unlikely that the Langmuir isotherm will be followed by real systems, yet many adsorption systems fit the equation well or at least reasonably so as a good approximation. Figure 11.5 shows some isotherms for the adsorption of dyes on cornstarch (Zografi and Mattocks 1963). The shapes of these curves are typical of Langmuirian adsorption, and the plots according to Eq. (11.38), in Fig. 11.6, confirm the validity of Eq. (11.37) as a description of the phenomenon.

Example 11.4. Figure 11.7 is the isotherm for the adsorption of indolinospiropyran from cyclohexane solution onto alumina of specific area $155 \text{ m}^2 \text{ g}^{-1}$ (Connors and Jozwiakowski 1987; Jozwiakowski 1987). The quantity *y* is the amount adsorbed in

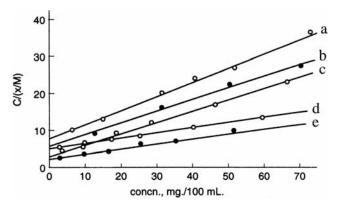


Figure 11.6. Plots according to Eq. (11.38) for the isotherms shown in Fig. 11.5. [Reproduced with permission from Zografi and Mattocks (1963).]

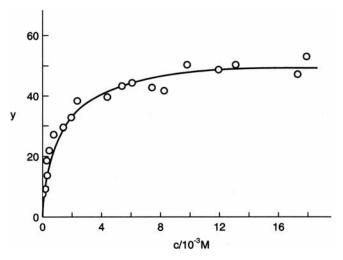


Figure 11.7. Adsorption isotherm at 25 °C for indolinospiropyran from cyclohexane onto alumina. The smooth line was drawn with Eq. (11.37) and the parameters given in Example 11.4. [Reproduced with permission from Connors and Jozwiakowski (1987).]

mg g⁻¹. From the plot according to Eq. (11.38), these parameters were found: $K = 810 \text{ M}^{-1}$ and $y_{\text{max}} = 53.9 \text{ mg g}^{-1}$. The molecular weight of the adsorbate is 327. Calculate the area occupied per molecule of adsorbate on the surface of the solid.

The area we want is σ , which appears in the relationship $A_t/\sigma N_A = n_{\text{max}}$. We therefore first convert y_{max} to n_{max} ; $y_{\text{max}} = 53.9 \text{ mg g}^{-1} = 0.0539 \text{ g g}^{-1}$; dividing by the molecular weight gives us $n_{\text{max}} = 1.65 \times 10^{-4} \text{ mol.}$

We solve for σ in $\sigma = A_t/N_A n_{max}$, where $A_t = 155 \text{ m}^2 \text{g}^{-1}$, finding $\sigma = 1.56 \times 10^{-18} \text{ m}^2 \text{molecule}^{-1}$, which is equivalent to 156 A^2 molecule⁻¹. Incidentally, the specific area A_t is itself determined by a gas adsorption method.

Solids as adsorbents have limited but important application as pharmaceutical agents; they serve mainly to adsorb toxic agents in the gastrointestinal tract. Activated charcoal is used for this purpose, being administered as soon as possible after ingestion of a toxic dose of a drug or poison, which on adsorption to the solid is effectively inactivated and is excreted. Besides this emergency application, adsorbents are widely used in the purification of chemicals, because they are effective in adsorbing colored impurities (colored molecules tend to have large surface areas and numerous polar groups) and in separation processes such as chromatography.

PROBLEMS

- **11.1.** Calculate the work of adhesion at the water-diethyl ether interface.
- **11.2.** Calculate the spreading coefficients of water on ether and of ether on water, and interpret the results.

- **11.3.** Calculate the surface work required to enlarge a cavity in water from a diameter of 0.1 mm to a diameter of 1 mm.
- **11.4.** Equation (11.38) is a linearized version of Eq. (11.37). There exist two additional linearized forms of Eq. (11.37). Find these linear equations, and show how the model parameters can be obtained from the slope and intercept of the plots.
- **11.5.** These are data for the adsorption of 6-methoxybenzoindolinopyran from cyclohexane solution onto silica gel of specific area $300 \text{ m}^2 \text{ g}^{-1}$. The quantity y is the amount of adsorbate adsorbed in mg g⁻¹. The molecular weight of the adsorbate is 307. Analyze the data; that is, determine if the data fit the Langmuir isotherm and, if so, evaluate the model parameters (Connors and Jozwiakowski 1987; Jozwiakowski 1987).

$10^{3}c_{2}(M)$	$y(\text{mg g}^{-1})$
1.18	58.2
2.08	60.6
4.26	94.7
5.75	103.5
8.65	105.8
10.62	107.4
12.27	119.7
14.13	123.8
16.06	124.8
16.98	127.6
19.81	134.3

THERMODYNAMICS OF CHEMICAL PROCESSES

Parts I and II of this book provide most of the concepts and quantitative relationships that we will need in our treatment of chemical processes. Occasional reference will be made to passages or equations in earlier chapters so as to minimize repetition of material. For brevity we state here that, except when indicated otherwise, constant temperature and pressure may be assumed; thus minimization of the Gibbs free energy is the criterion for equilibrium. Commonly, the fixed pressure is the ambient (atmospheric) pressure and the fixed temperature is 25° C or "room temperature." Usually, solute concentrations are given in moles per liter (molarity), symbolized either c_A or [A]; these represent concentrations at equilibrium unless noted otherwise. The solute reference state is the infinitely dilute solution, and very often we will suppose that the solute is in its reference state so that its activity coefficient is unity and its activity is equal to its concentration; this condition will allow us to focus on the essential chemistry of the process without being needlessly distracted by considering corrections for nonideal behavior. Such corrections can always be brought into the description as they are found to be required.

12

ACID-BASE EQUILIBRIA

Most drug molecules and biomolecules include one or more acidic or basic functional groups, so acid-base chemistry is pervasive in pharmaceutical systems. Acid-base equilibria therefore deserve detailed attention.

12.1. ACID-BASE THEORY

Definitions. Acid–base phenomena were observed very early in the development of chemical science, but their systematic understanding is a twentieth-century accomplishment. The Arrhenius theory of acids and bases, dating from the close of the nineteenth century, postulated that an acid is a substance that in water gives rise to hydronium ions, and that a base is a substance that in water gives rise to hydroxide ions. Thus HCl is an acid and NaOH is a base. The admittedly basic properties of compounds like amines, which do not contain the elements of the hydroxide ion, were proposed to result from reaction with water (hydrolysis) to generate hydroxide ions, as in

 $RNH_2 + H_2O \rightleftharpoons RNH_3^+ + OH^-$

The Arrhenius theory provided a satisfactory conceptual basis for understanding acid–base behavior in aqueous solution, but it was limited by its dependence on water as a solvent.

Acid-base behavior is observable in many solvents other than water, and such systems became comprehensible with the introduction, in 1923, of an acid-base theory

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by Bronsted (and independently by Lowry) based on these definitions; an *acid* is a species that can donate a proton, and a *base* is a species that can accept a proton. In reaction form, this is

$$\underset{\text{acid}}{\text{HA}} \rightleftharpoons \text{H}^+ + \text{A}^- \tag{12.1}$$

$$\mathbf{H}^{+} + \underset{\text{base}}{\mathbf{B}} \rightleftharpoons \mathbf{B}\mathbf{H}^{+} \tag{12.2}$$

Observe that the Bronsted definitions are built on the proton, and not the hydronium ion, so the new definition is independent of the solvent. We now proceed to explore the ramifications of this powerful definition.¹

The Conjugate Pair Concept. Observe in Eq. (12.1), on reading it from right to left, that the species A^- is accepting a proton, and so it must, by definition, be a base. Similarly, in Eq. (12.2) read from right to left, BH⁺ is donating a proton, so it must be an acid. The Bronsted acid–base definitions can be generally represented by

$$Acid \rightleftharpoons H^+ + base \tag{12.3}$$

An acid-base pair related by Eq. (12.3) is called a *conjugate acid-base pair*. Thus, referring to Eq. (12.1), we speak of A^- as the conjugate base of acid HA; from Eq. (12.2), BH⁺ is the conjugate acid of base B.

Equation (12.3) and the associated definitions make no mention of the charge types of the acid or base; the only requirement is that an acid be one positive charge greater than its conjugate base. Here are examples of equilibria that fit Eq. (12.3), showing acids and bases of various charge types:

$$\frac{\text{Acid}}{\text{CH}_3\text{CH}_2\text{COOH}} \rightleftharpoons \text{H}^+ + \text{CH}_3\text{CH}_2\text{COO}^-$$
$$\text{HCO}_3^- \rightleftharpoons \text{H}^+ + \text{CO}_3^{2-}$$
$$\text{CH}_3\text{CH}_2\text{NH}_3^+ \rightleftharpoons \text{H}^+ + \text{CH}_3\text{CH}_2\text{NH}_2$$
$$^+ \text{H}_3\text{N} - \text{C}_6\text{H}_4 - \text{NH}_3^+ \rightleftharpoons \text{H}^+ + \text{H}_2\text{N} - \text{C}_6\text{H}_4 - \text{NH}_3^+$$

¹ G. N. Lewis, also in 1923, proposed an even more general acid-base theory. Just as Bronsted improved on the Arrhenius theory by eliminating the hydroxide ion as a defining feature, so Lewis generalized the Bronsted theory by eliminating the proton. Lewis defined an *acid as an electron pair acceptor* and defined a *base as an electron pair donor*. Thus every Bronsted base is also a Lewis base, but the Lewis acid concept greatly expands our ideas of acid character. In the reaction

$$BF_3 + NH_3 \rightleftharpoons F_3B : NH_3$$

boron trifluoride is a Lewis acid.

The proton of Eq. (12.3), which is a naked nucleus, is a species of extremely high reactivity, a reactivity so high that in ordinary chemical systems the proton will not be detectable (because it will certainly combine with some other species in its vicinity). Consequently, we never actually observe Eq. (12.3) by itself. But if we combine two conjugate pairs, the proton donated by the acid of one pair can be accepted by the base of the other pair, and we observe an overall *proton transfer reaction*. Writing the conjugate pair reactions separately and adding them to give the net reaction:

Pair 1:
$$HA_{acid 1} \rightleftharpoons H^+ + A_{base1}^-$$

Pair 2: $B_{base 2} + H^+ \rightleftharpoons BH^+_{acid 2}$
Overall: $HA + B \rightleftharpoons A^- + BH^+$

The net result is that a proton has been transferred from the first pair to the second pair (reading from left to right), or vice versa when reading from right to left.

Next suppose that one of the pairs is the solvent. In particular, let it be water, our most important solvent. Water reacts in the pattern of Eq. (12.3):

$$\underset{\text{acid}}{\text{H}_2\text{O}} \rightleftharpoons \text{H}^+ + \underset{\text{base}}{\text{OH}^-}$$
 (12.4)

Thus H_2O is an acid, and OH^- is its conjugate base. But water is also capable of functioning in this version of Eq. (12.3):

$$\underset{\text{acid}}{\text{H}_3\text{O}^+} \rightleftharpoons \underset{\text{base}}{\text{H}^+} + \underset{\text{base}}{\text{H}_2\text{O}}$$
(12.5)

Thus H_2O is the conjugate base of H_3O^+ . A substance that (like water) can be either an acid or a base is said to be *amphoteric*. The amphoteric nature of water allows it to play the role of the second conjugate pair for either acids or bases. If the solute is an acid (say, HA), then H_2O functions as a base:

$$HA + H_2O \rightleftharpoons A^- + H_3O^+$$
(12.6)

whereas if the solute is a base (say, B), then H₂O functions as an acid:

$$\mathbf{B} + \mathbf{H}_2\mathbf{O} \rightleftharpoons \mathbf{B}\mathbf{H}^+ + \mathbf{O}\mathbf{H}^- \tag{12.7}$$

Throughout most of Chapter 12 we will be concerned with reactions like those in Eqs. (12.6) and (12.7).

Dissociation Constants. Consider the system described by Eq. (12.6) at equilibrium.² We can apply the thermodynamic concept of the equilibrium constant (Section 4.3) to define the quantity K_a :

$$K_a = \frac{[H_3O^+][A^-]}{[HA]}$$
(12.8)

where we assume that activity coefficients are unity. (The activity of water is equal to its mole fraction, essentially unity in dilute solution.) Very commonly, when water is understood to be the solvent, Eq. (12.6) is abbreviated to

$$HA \rightleftharpoons H^+ + A^-$$

and Eq. (12.8) becomes

$$K_a = \frac{[\mathrm{H}^+][\mathrm{A}^-]}{[\mathrm{HA}]}$$
(12.9)

The equilibrium constant K_a is known as the *acid dissociation constant* (also called the *acid ionization constant* or the *acidity constant*). In like manner, from Eq. (12.7) we define the *base dissociation constant* K_b :

$$K_b = \frac{[BH^+][OH^-]}{[B]}$$
(12.10)

We can apply the same formalism to water. From Eq. (12.4), we have

$$K_w = [\mathrm{H}^+][\mathrm{OH}^-] \tag{12.11}$$

where again the activity of water in the denominator is unity. K_w is called the *ion* product or autoprotolysis constant of water.

We are now prepared to develop the most powerful quantitative result of the Bronsted acid–base theory. Consider the equilibria of HA as an acid and of A^- as its conjugate base:

$$HA \rightleftharpoons H^{+} + A^{-} \qquad K_{a} = \frac{[H^{+}][A^{-}]}{[HA]} \qquad (12.12)$$

$$\mathbf{A}^{-} + \mathbf{H}_{2}\mathbf{O} \rightleftharpoons \mathbf{H}\mathbf{A} + \mathbf{O}\mathbf{H}^{-} \qquad K_{b} = \frac{|\mathbf{H}\mathbf{A}|[\mathbf{O}\mathbf{H}^{-}]}{[\mathbf{A}^{-}]} \qquad (12.13)$$

Now multiply together the K_a of Eq. (12.12) and the K_b of Eq. (12.13). The result is $[H^+][OH^-]$, or

$$K_w = K_a K_b \tag{12.14}$$

 $^{^2}$ Proton transfer reactions are extremely fast, so as soon as the solution has been made macroscopically homogeneous by mixing, the system is at equilibrium.

where the K_a and K_b of Eq. (12.14) refer to a conjugate acid–base pair. Since K_w is a constant (at a given temperature), Eq. (12.14) says that K_a and K_b are reciprocally related; that is, $K_b = K_w/K_a$. Values of K_w are known over the entire practical temperature range, so a result of Eq. (12.14) is that if we know either K_a or K_b , we can calculate the other. It is not necessary to measure both quantities.

We will subsequently see that K_a is a measure of acid strength and that K_b is a measure of base strength. Thus Eq. (12.14) shows that the strengths of an acid and its conjugate base are not independent; on the contrary, the stronger the acid, the weaker the base, and vice versa. Because of this relationship, we commonly specify acid strength in terms of K_a , but we do not use K_b to describe the strength of a base. Instead we use the K_a of the conjugate acid of the base. This may seem illogical, but it is traditional, and we will return to the issue in Section 12.7.

As it happens, K_a and K_b values are usually very small numbers, so for convenience (as one motive) we define

$$pK = -\log K \tag{12.15}$$

where *K* may be K_a , K_b , or K_w . The symbol *p* is a mathematical operator that turns a quantity into its negative logarithm. Applying Eq. (12.15) to Eq. (12.14) gives

$$\mathbf{p}K_w = \mathbf{p}K_a + \mathbf{p}K_b \tag{12.16}$$

which is a very convenient form.

Conventionally, we divide acids and bases into the classes of strong acids and bases and of weak acids and bases. Strong acids and bases are strong electrolytes, essentially completely dissociated in water; HCl, H_2SO_4 , HNO₃, NaOH, and KOH are examples. Weak acids and bases are incompletely dissociated in water; carboxylic acids, phenols, and amines are examples. The concept of the equilibrium constant is usefully applied only to the weak acids and bases.

pH. Let us apply the *p* operator to the hydrogen ion concentration. We write

$$pH = -\log \left[H^+\right] \tag{12.17}$$

Extending this formalism to Eq. (12.11) gives

$$pK_w = pH + pOH \tag{12.18}$$

from which we see that the acidity of a solution (measured by pH) and its alkalinity (measured by pOH) are coupled, so we do not need to measure both quantities. In the laboratory it is much easier to measure pH, so this is the quantity that we use to describe solution acidity or alkalinity. Table 12.1 lists pK_w values at several temperatures (Harned and Owen 1958, p. 638). Note that $pK_w = 14.00$ at 25°C.

A solution in which pH = pOH is said to be *neutral*; hence a neutral solution has pH = 7.00 (but only at 25°C). The practical pH range in water is essentially defined by

t(°C)	pK _w
0	14.94
10	14.54
20	14.17
25	14.00
30	13.83
40	13.54
50	13.26
60	13.02

Table 12.1. Ion product of water as a function of temperature

the value of pK_w , or 14 pH units. The lower the pH, the more acidic the solution; the higher the pH, the more alkaline the solution.³

Example 12.1 (a) Convert $K_a = 2 \times 10^{-4}$ to p K_a .

$$pK_a = -\log K_a$$

= -(log 2 + log 10⁻⁴)
= -(0.30 - 4)
= -(-3.70)
= 3.70

If $pK_a = 5.30$, what is K_a ?

$$-\log K_a = 5.30$$

$$\log K_a = -5.30$$

$$= -6.00 + 0.70$$

$$K_a = 5.0 \times 10^{-6}$$

Example 12.2. $pK_a = 4.75$ for acetic acid at 25°C. What is K_b of acetate ion? From $pK_a = 4.75$ we find $K_a = 1.78 \times 10^{-5}$. Using either Eq. (12.14) or Eq. (12.16), we find $pK_b = 9.25$ or $K_b = 5.62 \times 10^{-10}$.

Example 12.3. What is the hydroxide ion concentration of a solution having pH = 6.50 at $25^{\circ}C$?

From Eq. (12.18), pOH = 14.00 - 6.50 = 7.50, so $[OH^{-}] = 3.16 \times 10^{-8} M$.

³ It is important to appreciate a critical difference in the meanings of Eq. (12.16), $pK_w = pK_a + pK_b$; and of Eq. (12.18), $pK_w = pH + pOH$. Equation (12.16) refers to a *conjugate acid–base pair*; pK_a and pK_b , are constants, although mutually dependent. Equation (12.18) refers to a solution; pH and pOH are variables, although mutually dependent.

Example 12.4. The pK_a of the conjugate acid of methylamine is 10.64 at 25°C. Calculate the standard free energy change for the acid dissociation process.

From the basic thermodynamic result

$$\Delta G^0 = -RT \ln K$$

and the definition $pK_a = -\log K_a$ we derive

$$\Delta G^{0} = 2.303 RT pK_{a}$$

= (2.303)(1.987 cal mol⁻¹ K⁻¹)(298.15 K)(10.64)
= 14517 cal mol⁻¹
= 14.5 kcal mol⁻¹
= 60.7 kJ mol⁻¹

 ΔG^0 is positive because K_a is smaller (much smaller) than one. pK_a is seen to be directly proportional to ΔG^0 .

12.2. pH DEPENDENCE OF ACID-BASE EQUILIBRIA

Fractional Distribution of Acid–Base Species. Some of the treatment described here follows Connors (1982). Picture a very dilute solution of a given weak acid or base in an aqueous medium whose pH can be controlled, independently of the solute of interest, by the experimenter. Such pH control is easy to accomplish. We now assert, and will shortly demonstrate, that the fractions of solute present in the conjugate acid and base forms depend only on the pH of the solution and the pK_a of the acid. Thus the pH is a "master variable" that controls all acid–base equilibria in the solution.

Consider acid HA (whose charge type is irrelevant in what follows), having acid dissociation constant K_a . Let *c* be its total molar concentration, so that the mass balance expression Eq. (12.19) can be written.

$$c = [HA] + [A^{-}] \tag{12.19}$$

We introduce the fractions of solute in the conjugate acid and base forms with these definitions:

$$F_{\rm HA} = \frac{[\rm HA]}{c} \tag{12.20}$$

$$F_{\rm A} = \frac{\left[{\rm A}^{-}\right]}{c} \tag{12.21}$$

Algebraic combination of Eqs. (12.9) and (12.19)-(12.21) gives

$$F_{\rm HA} = \frac{[{\rm H}^+]}{[{\rm H}^+] + K_a} \tag{12.22}$$

$$F_{\rm A} = \frac{K_a}{[{\rm H}^+] + K_a}$$
(12.23)

which relate the fractions of solute in the conjugate acid and base forms to the acid dissociation constant and the hydrogen ion concentration of the medium. Equations (12.22) and (12.23) confirm the earlier assertion that the solution pH (which we recall can be established independently of the solute acid) is the only variable controlling the position of the acid–base equilibrium. Figure 12.1 is a plot of $F_{\rm HA}$ and $F_{\rm A}$ against pH, calculated with Eqs. (12.22) and (12.23) for a hypothetical acid having p $K_a = 4.0$. This curve is called a *sigmoid curve* because of its shape.

From Eqs. (12.22) and (12.23) we can deduce these general properties of such distribution curves. At any given pH, $F_{HA} + F_A = 1$ [a conclusion implicit in Eq. (12.19)]. At the point where the two curves cross, $F_{HA} = F_A = 0.5$, and at this point $[H^+] = K_a$, or pH = p K_a . At pH values much less than p K_a , F_{HA} approaches unity and F_A approaches zero; at pH values much greater than p K_a , F_A approaches unity and F_{HA} approaches zero. The curves in Fig. 12.1 apply to any monoprotic

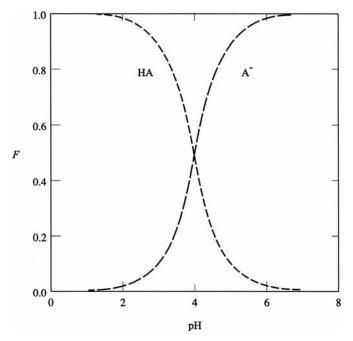


Figure 12.1. Variation with pH of the fraction F_{HA} (conjugate acid) and F_A (conjugate base) for an acid with $pK_a = 4.0$. [Reproduced by permission from Connors (1982).]

acid–base pair (i.e., one having only a single acid–base group and therefore only a single pK_a value) merely by sliding the curves along the pH axis until their pH of intersection matches the solute pK_a . From Fig. 12.1 we also can see that most of the interesting acid–base chemistry of this system takes place in the approximate pH range of $pK_a \pm 2$ units; outside this range the solute exists largely either as HA (at low pH) or as A⁻ (at high pH).

Rearrangement of Eqs. (12.20) and (12.21) gives $[HA] = cF_{HA}$ and $[A^-] = cF_A$; if, therefore, the total concentration *c* is known (*c* is sometimes called the *analytical concentration*), and if the pH and p K_a are known, the individual conjugate acid and base species concentrations are easily calculated.

Example 12.5. The pK_a of benzoic acid is 4.20. Calculate the concentrations of benzoic acid and benzoate ion in a solution whose pH = 5.20 and which was prepared to contain 0.005 M benzoic acid.

The preceding wording may seem confusing, but it is a fair example of the terminology that might be used in a laboratory. Its meaning is that c = 0.005 M.

From the pK_a we find $K_a = 6.31 \times 10^{-5}$ and from the pH we find $[H^+] = 6.31 \times 10^{-6}$. Equations (12.22) and (12.23) then give $F_{HA} = 0.0909$ and $F_A = 0.909$. Thus $[HA] = 0.005 \times 0.0909 = 4.55 \times 10^{-4}$ M and $[A^-] = 0.005 \times 0.909 = 4.55 \times 10^{-3}$ M. The slight discrepancy between the given value of *c* and the value obtained by summing [HA] and $[A^-]$ results from rounding errors (and is probably experimentally negligible). Notice that $pH = pK_a + 1$ and that at this condition $[A^-] = 10$ [HA].

A diprotic acid (sometimes called a dibasic acid) possesses two acidic groups and two pK_a values. We can symbolize such an acid as H_2A and write the acid–base equilibria as follows:

$$H_{2}A \stackrel{K_{1}}{\rightleftharpoons} H^{+} + HA^{-}$$
$$HA^{-} \stackrel{K_{2}}{\rightleftharpoons} H^{+} + A^{2-}$$

The placement of K_1 and K_2 over the arrows effectively defines these constants. We proceed to define fractions as before, except that now we have three solute species:

$$F_{\rm H_2A} = \frac{[{\rm H_2A}]}{c}$$
$$F_{\rm HA} = \frac{[{\rm HA}^-]}{c}$$
$$F_{\rm A} = \frac{[A^{2-}]}{c}$$

where $c = [H_2A] + [HA^-] + [A^{2-}]$. Combining these relations with the two dissociation constants gives

$$F_{\rm H_2A} = \frac{[{\rm H^+}]^2}{[{\rm H^+}]^2 + K_1[{\rm H^+}] + K_1K_2}$$
(12.24)

$$F_{\rm HA} = \frac{K_1[{\rm H}^+]}{\left[{\rm H}^+\right]^2 + K_1[{\rm H}^+] + K_1K_2}$$
(12.25)

$$F_{\rm A} = \frac{K_1 K_2}{\left[{\rm H}^+\right]^2 + K_1 \left[{\rm H}^+\right] + K_1 K_2}$$
(12.26)

Obviously $F_{H_2A} + F_{HA} + F_A = 1$. Observe how the three terms that make up the denominator constitute in turn the numerators of Eqs. (12.24)–(12.26). At high values of the hydrogen ion concentration the term $[H^+]^2$ dominates; as $[H^+]$ decreases, the middle term, $K_1[H^+]$, takes over as the largest contributor, and finally at small values of $[H^+]$ the last term, K_1K_2 , becomes the largest. Again some general relationships can be derived from Eqs. (12.24)–(12.26). Where the curves for F_{H_2A} and F_{HA} cross, equating Eqs. (12.24) and (12.25) gives $pH = pK_1$. Similarly, where F_{HA} and F_A are equal, $pH = pK_2$. The pH at which the maximum in F_{HA} appears is found by differentiating Eq. (12.25) with respect to $[H^+]$ and setting the derivative equal to zero; the result is

$$pH_{max} = \frac{pK_1 + pK_2}{2}$$
(12.27)

Figure 12.2 is a plot of Eqs. (12.24)–(12.26) for a hypothetical diprotic acid having $pK_1 = 5.0$ and $pK_2 = 10.0$. Note that the fraction F_{HA} rises essentially to unity at a pH given by Eq. (12.27).

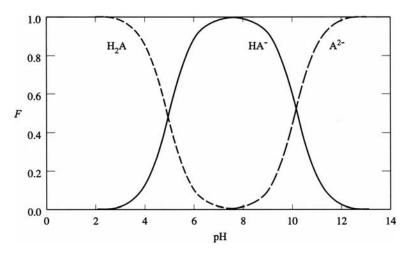


Figure 12.2. Species distribution diagram for a dibasic acid H_2A with $pK_1 = 5.0$ and $pK_2 = 10.0$. [Reproduced by permission from Connors (1982).]

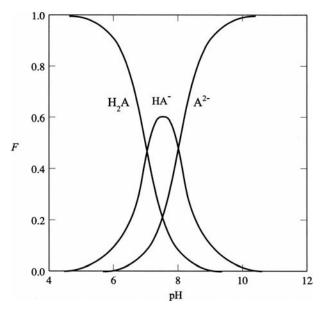


Figure 12.3. Species distribution diagram for a dibasic acid H_2A with $pK_1 = 7.0$ and $pK_2 = 8.0$. [Reproduced by permission from Connors (1982).]

Now compare Fig. 12.3 with Fig. 12.2. Again the curves are calculated with Eqs. (12.24)–(12.26), but now $pK_1 = 7.0$ and $pK_2 = 8.0$. Equation (12.27) is still obeyed, but now the fraction F_{HA} fails to rise to anywhere near unity. The behavior seen here is closely connected with the observation, in Fig. 12.1, that the essential acid-base chemistry occurs largely in the pH region $pK_a \pm 2$ units. In Fig. 12.2 the pK_1 and pK_2 values differ by 5 units, so the two acid-base equilibria described by K_1 and K_2 act essentially independently. In Fig. 12.3, however, $\Delta pK_a = pK_2 - pK_1$ is only 1 unit. As, in imagination, the pH is raised, swept from left to right across Fig. 12.3, H₂A is converted to HA⁻; this transformation commences at about $pK_1 - 2$ units, or pH 5. But before it can be carried to completion (which would not occur until $pK_1 + 2$, or pH 9), the system has been brought to within the range $pK_2 - 2$, or pH 6, so the second transformation, of HA⁻ to A^{2-} , must take place. The consequence is that F_{HA} cannot rise as high as it did in Fig. 12.2, where $\Delta p K_a$ was 5 units. Generally, one may expect successive p K_a values to control essentially independent equilibria if $\Delta p K_a > 4$. The consequence in Fig. 12.2 is that the system contains essentially only two acid-base species $(H_2A + HA^{-} or HA^{-} + A^{2-})$ at any given pH. In Fig. 12.3, however, the three species coexist in a wide pH range.⁴

⁴ Of course, these are equilibria, and all species are, strictly speaking, present in all solutions. From the practical point of view, however, we are often justified in neglecting the presence of a species if it is experimentally undetectable or exhibits no detectable influence.

These principles can be extended to acids and bases with any number of acid–base groups. The form of the expressions for the fractions is seen to take on a predictable pattern, and it is possible to write these expressions down without derivation. For the tribasic acid H_3A , these expressions become

$$F_{\rm H_3A} = \frac{[{\rm H^+}]^3}{[{\rm H^+}]^3 + K_1 [{\rm H^+}]^2 + K_1 K_2 [{\rm H^+}] + K_1 K_2 K_3}$$
$$F_{\rm H_2A} = \frac{K_1 [{\rm H^+}]^2}{[{\rm H^+}]^3 + K_1 [{\rm H^+}]^2 + K_1 K_2 [{\rm H^+}] + K_1 K_2 K_3}$$

and so on.

Example 12.6. Calculate the concentration of monoanion in an aqueous solution 0.01 M in phthalic acid at pH 5.00. $pK_1 = 2.95$; $pK_2 = 5.41$.

Phthalic acid is



1

The two ionizable groups are obviously equivalent. We calculate $K_1 = 1.12 \times 10^{-3}$ and $K_2 = 3.89 \times 10^{-6}$. From Eq. (12.25), we obtain

$$F_{\text{HA}} = \frac{(1.12 \times 10^{-3})(1 \times 10^{-5})}{(1 \times 10^{-5})^2 + (1.12 \times 10^{-3})(1 \times 10^{-5}) + (1.12 \times 10^{-3})(3.89 \times 10^{-6})}$$

= 0.713

Then from $[HA^-] = cF_{HA}$, we have

$$[HA^{-}] = 0.01 \times 0.713$$
$$= 7.13 \times 10^{-3} M$$

Buffer Solutions. Suppose that an aqueous solution is prepared to contain $a \mod L^{-1}$ of a weak acid HA and $b \mod L^{-1}$ of its conjugate base A⁻. (Of course A⁻ will be accompanied by $b \mod L^{-1}$ of its counterion, a cation.) As we have done earlier, the K_a for this system is defined

$$K_a = \frac{[\mathrm{H}^+][\mathrm{A}^-]}{[\mathrm{HA}]}$$
(12.28)

and we now apply the p operator to Eq. (12.28) to obtain the convenient form

$$pK_a = pH - \log \frac{[A^-]}{[HA]}$$
(12.29)

The quantities a and b are the *formal* concentrations of HA and A⁻; the actual equilibrium concentrations are somewhat different from these values because *both* of these equilibria must be mutually satisfied:

$$HA \rightleftharpoons H^{+} + A^{-}$$
$$H_{2}O \rightleftharpoons H^{+} + OH^{-}$$

We can find exact expressions for these concentrations by making use of mass balance and electroneutrality relationships. To be specific, suppose that the cationic counterion is labeled M^+ . Then these two mass balance expressions can be written as follows:

$$b = [\mathsf{M}^+] \tag{12.30}$$

$$a+b = [HA] + [A^-]$$
 (12.31)

The electroneutrality principle asserts that any macroscopic volume of solution is electrically neutral. This means that the sum of positive charges equals the sum of negative charges. For the aqueous solution under discussion we have

$$[\mathrm{H}^+] + [\mathrm{M}^+] = [\mathrm{OH}^-] + [\mathrm{A}^-]$$
(12.32)

Algebraic combination of Eqs. (12.30)–(12.32) gives the desired relationships:

$$[HA] = a - [H^+] + [OH^-]$$
(12.33)

$$[A^{-}] = b + [H^{+}] - [OH^{-}]$$
(12.34)

Combining Eqs. 12.29), (12.33) and (12.34), we obtain

$$pK_a = pH - \log \frac{b + [H^+] - [OH^-]}{a - [H^+] + [OH^-]}$$
(12.35)

If the solution pH is in the approximate range 4–10, the contributions of $[H^+]$ and $[OH^-]$ in Eq. (12.35) are usually negligible, and Eq. (12.35) becomes

$$pK_a = pH - \log\frac{b}{a} \tag{12.36}$$

In this form, Eq. (12.36) is known as the *Henderson–Hasselbalch equation*. This is a very convenient form for carrying out calculations.

A solution that contains comparable and appreciable concentrations of a conjugate weak acid–base pair is called a *buffer solution* because it resists a change in pH on the addition of a small amount of acid or base. This phenomenon can be demonstrated with an example. Suppose a solution is prepared to be 0.1 M in acetic acid (pK_a 4.76) and 0.1 M in sodium acetate. Then b/a = 1.00 and, according to Eq. (12.36), $pH = pK_a$, or pH = 4.76. Now let 1.0 mL of 0.1 M NaOH be added to 100 mL of this solution; what will be the new value of pH? It may be assumed that the sodium hydroxide converts an equivalent amount of acetic acid to sodium acetate. The solution therefore contains, after addition of the sodium hydroxide, (100)(0.1) - (1)(0.1) = 9.9 mmol of acetic acid, and (100)(0.1) + (1)(0.1) = 10.1 mmol of acetate, all in 101 mL of solution. The ratio b/a is now 1.02, its logarithm is 0.01, and Eq. (12.36) shows that the new pH is 4.77. Addition of the alkali has resulted in a pH change of only 0.01 unit. If the same volume of the sodium hydroxide solution had been added to 100 mL of pH 4.76 strong acid, the pH would have changed to about 11.

Equation (12.36) will sometimes be encountered in different guises, because some authors consider that a weak acid, on treatment with a strong base, is converted to its salt; thus the equation could be written $pH = pK_a + \log$ (salt/acid); when a weak base is treated with a strong acid to form its salt, the equivalent form is $pH = pK_a + \log$ (base/salt). Because of the possible confusion resulting from this terminology, we will use the Bronsted terminology by speaking of conjugate acid and base species. Thus Eq. (12.36) and equivalent versions always can be written in the form

$$pK_a = pH - \log \frac{[\text{conjugate base}]}{[\text{conjugate acid}]}$$
(12.37)

This equation relates the three quantities pH, pK_a , and the ratio b/a. Often two of these are known and the third may then be calculated.

Example 12.7. Calculate the pH of a buffer solution prepared by dissolving 242.2 mg of tris(hydroxymethyl)aminomethane in 10.0 mL of 0.170 M HC1 and diluting to 100 mL with water. The molecular weight of the solute is 121.1. It is a primary amine of structure (HOCH₂)₃CNH₂, with $pK_a = 8.08$ for the conjugate acid.

A total of 2.00 mmol of solute was weighed out, and 1.70 mmol of HCl was added. Since the HCl reacts with the amine to convert an equivalent amount to its conjugate acid (protonated) form, this means that a = 1.7/100 M and b = (2.0 - 1.7)/100 M. Using these figures in the Henderson–Hasselbalch equation, we obtain

$$pH = pK_a + \log \frac{0.003}{0.017} = 8.08 - \log \frac{0.017}{0.003}$$
$$= 8.08 - \log 5.67 = 7.25$$

In this calculation the ratio was inverted merely to give a value greater than unity, for ease in taking the logarithm.

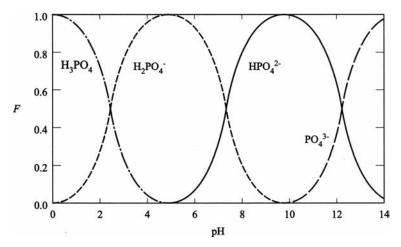


Figure 12.4. Species distribution diagram for phosphoric acid: $pK_1 = 2.23$, $pK_2 = 7.21$, $pK_3 = 12.32$. [Reproduced by permission from Connors (1982).]

Example 12.8. Calculate the pH of a buffer prepared to contain $0.09 \text{ M NaH}_2\text{PO}_4$ and $0.01 \text{ M K}_2\text{HPO}_4$. p $K_1 = 2.23$, p $K_2 = 7.21$, p $K_3 = 12.32$ for phosphoric acid.

In general a buffer of a polyprotic acid may be a very complex mixture, and a species distribution diagram is helpful in clarifying the problem. Figure 12.4 shows this diagram for phosphoric acid. The pK values of this acid are widely spaced, and phosphoric acid behaves essentially as if it were an equimolar mixture of three monobasic acids of the given pK values. From the experimental values a = 0.09 M and b = 0.01 M, Fig. 12.4 clearly shows that the pH will be approximately 6.3 and that the solution contains practically no H₃PO₄ or PO₃⁴⁻ at this pH. We have now simplified the problem to that of a monobasic acid (H₂PO₄⁻) and its conjugate base (HPO₄²⁻), with the dissociation constant $pK_2 = 7.21$. Applying the Henderson–Hasselbalch equation gives pH = $7.21 - \log 9 = 6.26$.

Table 12.2 was constructed by means of Eq. (12.36). Table 12.2 illustrates the symmetry and simplicity provided by the logarithmic form of the dissociation constant expression. The table also confirms the suggestion derived from Fig. 12.1 that most of the acid–base "action" takes place in the approximate pH range $pK_a \pm 2$.

Inasmuch as the function of a buffer is to minimize changes in pH, it is useful to have a measure of the "buffering capacity" of a buffer solution. This is provided by the *buffer index* β . Let *b* be the concentration of strong base added to a solution containing total concentration c of a weak acid. Then β is defined by

$$\beta = \frac{db}{d\mathrm{pH}} \tag{12.38}$$

Thus β is the concentration of strong base required to change the pH by a given amount; the larger the value of β , the greater is the buffer capacity of the solution. Since the

b/a	pH
$\overline{0.001(10^{-3})}$	$pK_a - 3$
$0.01(10^{-2})$	$pK_a - 2$
$0.1(10^{-1})$	$pK_a - 1$
$1(10^{0})$	pK _a
$10(10^1)$	$pK_a + 1$
$100(10^2)$	$pK_a + 2$
$1000(10^3)$	$pK_a + 3$

Table 12.2. Relationship of pH, pK_a , and the conjugate base/acid ratio

strong base converts the weak acid to its conjugate base, *b* has the meaning given to it earlier, and from Eqs. (12.21) and (12.23), we write $F_A = b/c = K_a/([H^+] + K_a)$, or

$$b = \frac{cK_a}{[\mathrm{H^+}] + K_a}$$

Therefore

$$\frac{db}{d[\mathrm{H^+}]} = \frac{-cK_a}{\left([\mathrm{H^+}] + K_a\right)^2}$$

and

$$\frac{db}{dpH} = -2.3[H^+]\frac{db}{d[H^+]}$$

$$\beta = 2.3cF_{HA}F_A \qquad (12.39)$$

Equation (12.39) shows that β is directly proportional to total buffer concentration *c*, as well as to the product $F_{\text{HA}}F_{\text{A}}$. It is easy to show (e.g., by inserting numbers for the fractions) that this product is maximal when $F_{\text{HA}} = F_{\text{A}} = 0.5$, which, we have seen, occurs when $pH = pK_a$. This result, together with extensive laboratory experience, leads to the guideline that buffer capacity is maximal when $pH = pK_a$ and is acceptable in the approximate range of $pH = pK_a \pm 1$. This is the information needed to design effective buffer solutions.⁵

12.3. CALCULATION OF SOLUTION pH

We have already seen some calculations of solution pH, but here the treatment will be more systematic. There are two kinds of aqueous solutions to consider: (1) a solution

⁵ Buffers are commonly described in an abbreviated terminology that must be understood. For example, a 0.10 M pH 5.00 acetate buffer means that the total buffer concentration *c* is 0.10 M and the pH is 5.00; obviously the solution contains both acetate and acetic acid, in concentrations that can be worked out from the Henderson–Hasselbalch equation. Similarly, a 0.05 M pH 7.0 phosphate buffer contains both dihydrogen phosphate (HPQ₄^{2–}), usually taken as their salts.

prepared with a single solute, whether a strong acid, strong base, weak acid, or weak base; and (2) a solution prepared to contain two conjugate species, namely a weak acid and its conjugate base. Obviously, all acid–base systems contain both species; the distinction being made is that in (1) one of these arises solely from the operation of the solution equilibria, whereas in (2) the experimenter ensures by manipulation that appreciable concentrations of both are present.

Any aqueous acid–base system can be completely described by carrying through the following procedure:

- 1. Write the electroneutrality equation for the solution.
- 2. Write the mass balance expressions for each solute.
- 3. Define all pertinent K_a values.
- 4. Define K_w .
- 5. Algebraically combine the preceding expressions.

Seldom is it necessary to carry through the system in its full generality, and we will see that shortened versions, often employing chemically reasonable approximations, will usually suffice. The level of accuracy sought is determined by the typical accuracy in an experimental measurement, which is, at best, about 0.01 pH unit.

Strong Acid or Base. A strong acid or base is essentially completely dissociated in dilute aqueous solution. The common strong acids are hydrochloric (HCl), sulfuric (H₂SO₄), nitric (HNO₃), and perchloric (HClO₄); the common strong bases are sodium and potassium hydroxides (NaOH, KOH).

Let c be the total (analytical) molar concentration of strong acid HX. According to the electroneutrality principle applied to this solution, we obtain

$$[H^+] = [OH^-] + [X^-]$$
(12.40)

The source of the hydroxide ion is the dissociation of water. The mass balance expression for this solution is

$$c = [X^-]$$
 (12.41)

Equations (12.40) and (12.41) can be combined with the definition of K_w to give a quadratic equation, which can be solved for $[H^+]$. However, if the concentration *c* is greater than $\sim 10^{-6}$ M, then $[X^-]$ will be much greater than $[OH^-]$, and we can write the acceptable approximation $[H^+] = [X^-]$, or

$$[{\rm H}^+] = c \tag{12.42}$$

which states that the hydrogen ion concentration is numerically equal to the total concentration of strong acid. Similarly for a strong base MOH the electroneutrality equation is

$$[\mathrm{H}^+] + [\mathrm{M}^+] = [\mathrm{OH}^-] \tag{12.43}$$

which simplifies to $c = [OH^-]$, where *c* is the analytical concentration of the strong base.

These calculations have ignored nonideality effects, which, we have seen, set in at fairly low ionic strengths for ionic species (Chapter 8). For example, the calculated pH of 0.10 M HCl is 1.00, whereas the measured pH is 1.10.

Example 12.9

- (a) What is the pH of 0.005 M H₂SO₄? Sulfuric acid dissociates according to $H_2SO_4 \rightarrow 2H^+ + SO_4^{2-}$. Thus in this solution $[H^+] = 2c = 0.010$ M, and pH = 2.00.
- (b) What is the pH of 0.025 M NaOH? We have $[OH^-] = 0.025$ M, so pOH = 1.60 and pH = 14.00 - 1.60 = 12.40.

Weak Acid. Let HA be a monoprotic weak acid at total concentration c. The dissociation reaction is

$$\mathrm{HA} \stackrel{K_a}{\rightleftharpoons} \mathrm{H}^+ + \mathrm{A}^-$$

From the electroneutrality principle we write

$$[H^+] = [OH^-] + [A^-]$$
(12.44)

and the mass balance expression is

$$c = [HA] + [A^{-}] \tag{12.45}$$

An exact solution combines Eqs. (12.44) and (12.45) with the definitions of K_a and K_w . Usually, however, it is reasonable to approximate Eq. (12.44) by $[H^+] = [A^-]$. Using this equality in the definition of K_a yields

$$K_a = \frac{[\mathrm{H}^+]^2}{c - [\mathrm{H}^+]} \tag{12.46}$$

which can be rearranged to the quadratic form

$$[\mathbf{H}^+]^2 + K_a [\mathbf{H}^+] - K_a c = 0$$
(12.47)

Application of the quadratic formula gives

$$[\mathrm{H}^+] = \frac{-K_a \pm \sqrt{K_a^2 + 4K_a c}}{2} \tag{12.48}$$

One uses the physically meaningful solution.

Suppose that $[H^+] \ll c$; then the denominator in Eq. (12.46) can be approximated by $c - [H^+] = c$. If this is acceptable, we obtain $K_a = [H^+]^2/c$, or

$$[\mathrm{H}^+] = \sqrt{K_a c} \tag{12.49}$$

This equation offers an extremely simple solution to the problem. Whether this approximation is reasonable can be assessed by comparing the results calculated by Eqs. (12.48) and (12.49).

The preceding derivations are applicable to aqueous solutions of monoprotic acids regardless of charge type, so Eqs. (12.48) and (12.49) apply to neutral acids (such as RCOOH), to positively charged acids (like RNH_3^+), and to negatively charged acids (like $\text{HPO}_4^{2^-}$).

Example 12.10

(a) Calculate the pH of 0.02 M *trans*-cinnamic acid (p $K_a = 4.30$). We can use Eq. (12.49) with c = 0.02 and $K_a = 5.0 \times 10^{-5}$; the result is $[H^+] = 1.0 \times 10^{-3}$ M, or pH = 3.00. Assessing the validity of the approximation leading to Eq. (12.49), we see that c = 0.020 M and $[H^+] = 0.001$ M (as calculated), so $[H^+]$ is about 5% of *c*. Let us repeat the calculation with Eq. (12.48):

$$[H^+] = \frac{-5 \times 10^{-5} \pm \sqrt{(5 \times 10^{-5})^2 + (4)(0.02)(5 \times 10^{-5})}}{2}$$
$$= \frac{-5 \times 10^{-5} \pm \sqrt{25 \times 10^{-10} + 0.4 \times 10^{-5}}}{2}$$
$$= \frac{-5 \times 10^{-5} \pm \sqrt{4 \times 10^{-6}}}{2}$$
$$= \frac{0.002 - 0.00005}{2}$$
$$= 0.975 \times 10^{-3}$$

so pH = 2.99. The approximate solution, Eq. (12.49), gave pH = 3.00, which usually would be considered acceptable, because pH seldom can be measured to better than 0.01 unit.

(b) Calculate the pH of 0.05 M ammonium chloride ($pK_a = 9.25$). Ammonium chloride is a salt. It dissociates completely according to

$$NH_4Cl \rightarrow NH_4^+ + Cl^-$$

The ammonium ion NH_4^+ is a weak acid:

$$NH_4^+ \rightleftharpoons H^+ + NH_3$$

Therefore we anticipate that the solution will be acidic.⁶ The given pK_a is for the acid NH₄⁺. Using Eq. (12.49) with c = 0.05 and $K_a = 5.62 \times 10^{-10}$ gives $[H^+] = 5.30 \times 10^{-6}$ M, or pH = 5.28. Our experience with Example 12.10(a) convinces us that this is an acceptably accurate solution.

Weak Base. The base dissociation reaction is

$$B + H_2O \rightleftharpoons^{K_b} BH^+ + OH^-$$

and by reasoning identical with that applied to the weak acid case, we derive

$$K_b = \frac{[\text{OH}^-]^2}{c - [\text{OH}^-]}$$
(12.50)

When $[OH^-] \ll c$, Eq. (12.50) becomes

$$[OH^{-}] = \sqrt{K_b c} \tag{12.51}$$

where *c* is the total base concentration. Equations (12.50) and (12.51) apply to neutral bases (like RNH₂), to positively charged bases (like $H_2NCH_2CH_2NH_3^+$), and to negatively charged bases (like RCOO⁻).

Example 12.11. What is the pH of 0.10 M potassium acetate? ($pK_a = 4.76$).

We will use Eq. (12.51) with c = 0.10. The p K_a that is given is for the conjugate acid, namely, acetic acid. The chemistry of this system mimics what we saw in Example 12.10(b). First the salt potassium acetate (symbolized KOAc) completely dissociates:

$$KOAc \rightarrow K^+ + OAc^-$$

The potassium ion is neutral, but OAc^- is a weak base; it is the conjugate base of acetic acid, and it makes the solution basic because of the equilibrium

$$OAc^- + H_2O \rightleftharpoons HOAc + OH^-$$

 $^{^{6}}$ The chloride ion is neutral, as may be deduced by noting that HCl is a strong acid; that is, its "conjugate base" Cl⁻ is so weak that it is completely ineffectual at capturing the proton.

From $pK_a 4.76$ we find $pK_b = 14.00 - 4.76 = 9.24$, or $K_b = 5.75 \times 10^{-10}$. Applying Eq. (12.51), we obtain

$$[OH^{-}] = \sqrt{(5.75 \times 10^{-10})(0.1)}$$
$$= \sqrt{5.75 \times 10^{-11}}$$
$$= \sqrt{57.5 \times 10^{-12}}$$
$$= 7.6 \times 10^{-6}$$

so pOH = 5.12 and pH = 8.88. It is helpful to check that the calculated pH is on the correct side of neutrality.

Mixture of Weak Acid and Its Conjugate Base. We have already treated this case under *Buffer Solutions*. The appropriate relationship is

$$pK_a = pH - \log \frac{[A^-]}{[HA]}$$
(12.52)

and we have seen how this equation can be applied to calculate the pH of buffers. Of course, if we wish to prepare a buffer of given pH, we use the equation in an alternative manner. First a buffer substance is selected according to the criterion that the desired pH be in the range $pK_a \pm 1$. Then we use Eq. (12.52) to find the ratio $[A^-]/[HA]$ that is needed to deliver this pH. Finally we decide on a total buffer concentration *c*. Then, since we have the sum $c = [HA] + [A^-]$ and the ratio $[A^-]/[HA]$, we solve for the individual concentrations [HA] and $[A^-]$ and prepare the solution to contain these concentrations.

Some mixtures of acids and bases can be quite complex, consisting of polyprotic acids or bases, or of a weak acid and a weak base (not its conjugate). A complete analytical description can always be obtained by means of the general scheme outlined at the beginning of this section, but the solution of the final equation will usually require approximations of the type we have made use of above.⁷

12.4. ACID–BASE TITRATIONS

A titration is an experimental operation in which a solution of one reactant (the *titrant*), this solution having an accurately known concentration, is added to a solution of a substance (the sample or *analyte*) with which it will stoichiometrically and quantitatively react, until chemically equivalent amounts of titrant and sample have been mixed. From the stoichiometry of the known reaction between the titrant and sample substances, along with the known concentration of titrant, the amount or concentration

⁷ The exact equation obtained by applying this general scheme will be a polynomial in [H⁺], whose highest power will be equal to the number of dissociation constants (including K_w) plus one.

of the sample substance can be calculated. The theoretical point at which the amounts of titrant and sample are equivalent is called the *equivalence point*, and its experimental estimate is the *endpoint* of the titration. If the titrant is an acid and the sample a base, or the reverse, the operation is called *acid–base titration*. The analytical calculations, which are simple, lie outside our present concern. Here we are interested in how the pH of the sample solution varies, throughout the course of the titration, as increasing volumes of titrant solution are added to it. A plot of this pH [on the vertical axis (ordinate; *y* axis)] against the volume of titrant added [on the horizontal axis (abscissa; *x* axis)] is called an *acid–base titration curve*. We will see how it is possible to calculate such a titration curve, along with the information to be derived from it.

Strong Acid–Strong Base Titration. A strong acid is completely dissociated into H^+ and its anion, which is neutral; a strong base is completely dissociated into OH^- and its cation, which is neutral. Consequently, the reaction that occurs in the sample solution is

$$H^+ + OH^- \rightleftharpoons H_2O$$

This is the reverse of the autoprotolysis of water, so its equilibrium constant is $1/K_{w}$, or 1×10^{14} . The reaction is obviously quantitative; in casual terms, "it goes completely to the right." At each stage in the titration the sample solution consists of a solution of a strong acid or a strong base, so the calculation of the titration curve involves no new concepts.

Example 12.12. Calculate the titration curve for the titration of 25.0 mL of 0.05 M HCl with 0.10 M NaOH.

In this titration HCl is the sample and NaOH is the titrant. Before any titrant has been added, the solution consists of 0.05 M HCl, so $[H^+] = 0.05$ M and pH = 1.30. Now suppose we add 1.0 mL of titrant. We can arrange the work in tabular form:

Initially the sample had	(25)(0.05)	=	1.25 mmol H^+
We have added	(1)(0.10)	=	0.10 mmol OH^-
Remaining in the solution are			1.15 mmol H^+

This 1.15 mmol of H⁺ is contained in 26 mL of solution, so the new concentration is $[H^+] = 1.15 \text{ mmol}/26 \text{ mL} = 0.04423 \text{ M}$, and the new pH is 1.35. Obviously the pH has risen because we have added a strong base to the solution.

This calculation is repeated with increasing volumes of titrant. Here are some results: at 5 mL of titrant, pH = 1.60; at 8 mL, pH = 1.87; at 10 mL, pH = 2.15; at 12 mL, pH = 2.87; at 12.2 mL, pH = 3.09; at 12.4 mL, pH = 3.57.

The equivalence point occurs at 12.5 mL of titrant, because at this volume the number of millimoles (mmol) of NaOH added exactly matches the number of mmol of HCl initially present; and at this point the foregoing method of calculation gives an embarrassing result, because it leads to the conclusion that the number of mmol of H⁺ remaining is zero. But of course there always will be hydrogen ions in water. At the

equivalence point we simply have a solution of sodium and chloride ions, and (neglecting impurities from the atmosphere) the solution is neutral, so pH = 7.00.

Let us continue to add titrant; suppose that 13 mL has been added. All that is happening is that the titrant is being diluted. Since the first 12.5 mL of titrant was consumed in the titration reaction, we have added 0.5 mL in excess, or (0.5) (0.1) = 0.05 mmol of OH⁻. This is contained in 38 mL of solution, so $[OH^-] = 0.05/38 = 0.001316$ M, pOH = 2.88, and pH = 11.12.

It is left as an exercise to plot the titration curve and to locate the equivalence point. The calculation of a few more points in the titration, especially after the equivalence point, may be helpful in defining the shape of the curve.

Weak Acid–Strong Base Titration. We could apply the systematic treatment outlined at the beginning of Section 12.3 to obtain a general equation applicable throughout the course of the titration, but it is simpler to recognize that at any stage in the titration the solution consists of an example of a type that we have already considered. For the titration of a weak acid HA with a strong base MOH, here are the four such stages into which we divide the titration:

- *Stage 1: Before the Titration Begins.* The sample solution is simply a solution of a weak acid, and Eq. (12.48) or (12.49) is applicable.
- Stage 2: During Titration. Since some strong base has been added and has converted an equivalent amount of weak acid to its conjugate base according to

$$HA + OH^{-} \rightleftharpoons A^{-} + H_2O$$

the solution contains appreciable quantities of both HA and A^- . It is a buffer solution throughout much of this stage, and Eq. (12.36) applies.

- *Stage 3: At the Equivalence Point.* Now the weak acid HA has been quantitatively converted to A⁻. The sample solution consists of this weak base in water, and Eq. (12.50) or Eq. (12.51) may be used.
- *Stage 4: After the Equivalence Point.* The solution contains excess strong base and the weak base A⁻, whose dissociation is repressed by the common ion effect of the hydroxide from the strong base. We therefore ignore the contribution of A⁻, and we calculate the concentration of hydroxide exactly as in Example 12.12.

Example 12.13. Calculate the titration curve for the titration of 10.0 mL of 0.2 M weak acid ($pK_a = 5.0$) with 0.2 M sodium hydroxide.

Stage 1. We have $K_a = 1 \times 10^{-5}$ and c = 0.2, so, from Eq. (12.49),

$$[\mathrm{H^{+}}] = \sqrt{(1 \times 10^{-5})(0.2)}$$
$$= 1.41 \times 10^{-3}$$

or pH = 2.85.

Stage 2. Let 2.0 mL of titrant be added. This is the situation:

Initially in the sample :	(10.0)(0.2)	=	2.00 mmol HA
Strong base added :	(2.0)(0.2)	=	$0.40 \mathrm{~mmol~OH^{-}}$
Weak acid remaining			1.60 mmol HA

Using Eq. (12.36), we have

$$pH = 5.00 + \log \frac{0.40}{1.60} = 4.40$$

Of course, the logarithmic term is strictly a ratio of *concentrations*, so we should have written

$$pH = 5.00 + \log \frac{0.40/12}{1.60/12}$$

where 12 mL is the total volume, but these volumes cancel, so we effectively calculate a ratio of *amounts*. Using the same calculational method, we obtain these further results: at 5.0 mL, pH 5.00; at 7.0 mL, pH = 5.37.

Stage 3. The equivalence point obviously corresponds to 10.0 mL of titrant added. Initially we had (10.0) (0.2) = 2.00 mmol of HA, and this has now been converted to 2.00 mmol of A⁻, which is contained in 20.0 mL of solution. Using Eq. (12.51), with $K_b = 1 \times 10^{-9}$ and c = 0.10 M, we obtain

$$[OH^{-}] = \sqrt{(1 \times 10^{-9})(0.1)}$$
$$= 1 \times 10^{-5} M$$

so pOH = 5.00, and pH = 9.00. Obviously the solution is basic at the equivalence point, since it contains only a weak base.

Stage 4. Let 12.0 mL of titrant be added. This constitutes an excess of 2.0 mL of 0.2 M strong base in a total volume of 22.0 mL, so $[OH^-] = (2.0)(0.2)/22 = 0.0182$ M, giving pOH = 1.74 and pH = 12.26.

Figure 12.5 is a plot of the full titration curve. Three important lessons are to be learned from this graphical display:

Detection of the Endpoint. Observe the relatively sharp "break" in the curve corresponding to the equivalence point at 10.0 mL of titrant. The point at which the slope has its maximum value gives us our experimental estimate of the endpoint. In the laboratory we make use of this information in either of two ways: (1) If we know the pH at the endpoint by either calculation or experience, we can titrate to that pH

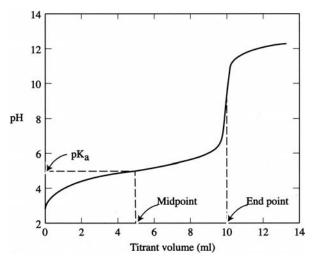


Figure 12.5. Calculated curve for the titration of 10.0 mL of 0.2 N weak acid ($pK_a = 5.00$) with 0.2 N strong base. [Reproduced by permission from Connors (1982).]

(usually as detected with a visual indicator); or (2) we can experimentally measure the pH, plot the curve, and establish the endpoint graphically.

Buffer Properties. We have seen that the sample solution throughout much of the titration is a buffer, and this property is manifested in the slope of the titration curve, which is seen to be the reciprocal of β , the buffer index [Eq. (12.38)]. Thus the shallower the slope, the greater the buffer capacity. Recall our earlier claim that buffer capacity is acceptable in the range $pH = pK_a \pm 1$, and observe how this range is reflected on the titration curve.⁸

Determination of pK_a. We will subsequently learn how to measure the pH experimentally, so the titration curve can be determined. Writing Eq. (12.37) again

 $pH = pK_a + \log \frac{[\text{conjugate base}]}{[\text{conjugate acid}]}$

we see that if we know the ratio appearing in the logarithmic term and can measure the pH of the solution, we can calculate pK_a . In particular, consider the point in the titration corresponding to one-half way to the endpoint, as measured in titrant volume. At this point (call it the midpoint), one-half of the weak acid has been converted to its conjugate base, so the ratio [conjugate base]/[conjugate acid] = 1.00, and pH = pK_a . This relationship is shown on Fig. 12.5. (Of course, the pK_a thus determined is not a "thermodynamic pK_a " because we have not yet applied activity coefficient corrections; these are discussed in Chapter 13.) An interesting capability of this technique is

⁸ Figure 12.5 also shows that fairly high concentrations of a strong base (seen well beyond the endpoint in Fig. 12.5) constitute good buffers. The same is true of strong acids.

that it allows us to determine the pK_a of an acid whose identity is unknown; we merely need to determine the titration curve, read off the endpoint volume, and then, by interpolation, establish the pH corresponding to one-half the endpoint volume.

This equality $pH = pK_a$ at the titration midpoint is yet another manifestation of a condition we have encountered before, notably in discussing Eqs. (12.22) and (12.23), Eqs. (12.24)–(12.26), Table 12.2, and Eq. (12.39).

Calculation of the titration curve for the titration of a weak base with a strong acid is analogous to the preceding treatment. For such a titration the pH will initially be on the alkaline side of neutrality and will decrease throughout the titration. At the endpoint the solution contains the conjugate acid of the sample base, so the solution is acidic.

The titration curves of polyfunctional acids and bases, if their successive pK_a values differ by about 4 or more units, show a "break" at each endpoint for the successive titration of the groups (in the order strongest to weakest). If, however, the successive pK_a values are not widely spaced, the successive breaks are less distinct because the phenomenon seen in Fig. 12.3, in which more than two solute species coexist at some pH values, intrudes. Such systems can be algebraically described, and in this manner the experimental data can be fitted to the equation to extract the pK_a values. We will not pursue this analysis.

Acid–Base Indicators. An acid–base indicator is a compound whose conjugate acid and base forms exhibit different colors. There is no limitation on the charge type of the indicator. Indicators are used to detect the endpoint in a titration; the selection of an indicator is based on the simple principles to be discussed here.

Consider the indicator acid HI. This acid will undergo dissociation in aqueous solution:

$$HI \rightleftharpoons H^+ + I^-$$

The acid dissociation constant has the form of K_a , but it is often symbolized K_I :

$$K_{\rm I} = \frac{[{\rm H}^+][{\rm I}^-]}{[{\rm H}{\rm I}]} \tag{12.53}$$

The acid form HI is responsible for the acid color of the indicator solution, and I^- shows the base color. The color that our eyes see is related to the relative concentrations of these two forms of the indicator. Rearranging Eq. (12.53) gives

$$\frac{[\mathrm{I}^{-}]}{[\mathrm{HI}]} = \frac{K_{\mathrm{I}}}{[\mathrm{H}^{+}]} \tag{12.54}$$

Two important conclusions follow from Eq. (12.54). The color is controlled by the pH of the solution; and the color change during a titration is not abrupt but occurs in a continuous manner, since the pH changes continuously, as we saw earlier.

It is characteristic of the typical human eye that in order to detect the first deviation from the pure acid color in a solution of the indicator, the ratio $[I^-]/[HI]$ must be at least

1/10; that is, about 10% of the indicator must be in the base form. Similarly, about 10% of the indicator must be in the acid form to detect any acid color. (These statements apply to two-color indicators.) In between these limits the eye recognizes that a mixture of colors is present and that the indicator color change is taking place if a titration is being carried out. These limits for $[I^-]/[HI]$ of 0.1 to 10 have no theoretical chemical significance but are related to the sensitivity of the observer's eyes and to the particular indicator used; some colors are more readily detected than others.

The pH values at which these limits of observable color change occur are easily calculated. From Eq. (12.54):

$$pH = pK_I + \log \frac{[I^-]}{[HI]}$$
(12.55)

For the limit on the acid side, $[I^-]/[HI] = 0.1$, or $pH = pK_I - 1$. For the limit on the base side, $[I^-]/[HI] = 10$, or $pH = pK_I + 1$. The pH range within which the indicator can be observed to be changing color is thus given approximately by $pH = pK_I \pm 1$. This is called the *transition interval* of the indicator, and it clearly depends on the pK_I of the indicator. This is why indicators of different structure change color at different pH values.

Table 12.3 gives the colors and transition intervals of some useful acid–base indicators. Many of the intervals are less than 2 pH units, suggesting that the limits $pK_I \pm 1$ are rather conservative. One-color indicators, in which only one of the conjugate forms possesses a visible color, will not behave visually in accordance with the above-mentioned treatment, although of course their equilibria will be

Indicator	Transition Interval	Acid Color	Base Color
Methyl violet	0.15-3.2	Yellow	Violet
Thymol blue	1.2–2.8	Red	Yellow
Quinaldine red	1.4–3.2	Colorless	Red
2,4-Dinitrophenol	2.4-4.0	Colorless	Yellow
Methyl yellow	2.9-4.0	Red	Yellow
Bromcresol blue	3.0-4.6	Yellow	Blue
Methyl orange	3.1-4.4	Red	Yellow
Bromcresol green	3.8–5.4	Yellow	Blue
Methyl red	4.4-6.2	Red	Yellow
Bromcresol purple	5.2-6.8	Yellow	Purple
4-Nitrophenol	5.6-7.6	Colorless	Yellow
Bromothymol blue	6.0–7.6	Yellow	Blue
Phenol red	6.4-8.2	Yellow	Red
Cresol red	7.2-8.8	Yellow	Red
Thymol blue	8.0-9.6	Yellow	Blue
Phenolphthalein	8.2-10	Colorless	Red
Thymolpthalein	9.3-10.5	Colorless	Blue
α -Naphtholbenzein	9.8-11.0	Yellow	Blue

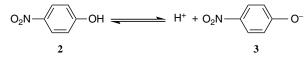
Table 12.3. Acid-base indicators

described by the same equations. The pK_I of an indicator, and therefore its transition interval, can be affected by the salt concentration of the solution and by organic solvents incorporated into the aqueous medium.

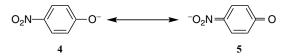
In order to achieve an accurate visual detection of the endpoint in an acid–base titration, evidently the pH of the solution must change by about 2 units in the immediate vicinity (say, $\pm 0.2\%$) of the endpoint. Whether this condition is satisfied in any given circumstance can be determined by calculating the titration curve. Figure 12.5 shows the results of such a calculation, indicating that this would be a feasible titration with visual endpoint detection because of the sharp break in the region of the theoretical equivalence point. Calculations show that the more concentrated the solution and the stronger the acid (for titrations with base), the greater this break.

An indicator should now be chosen such that the pH at the titration equivalence point falls within the transition interval of the indicator. In the titration of a weak acid with a strong base, at the endpoint the solution contains the conjugate base of the acid, so its pH is in the alkaline range, as shown in Fig. 12.5. Weak acids therefore are usually titrated using thymol blue, phenolphthalein, or thymolphthalein as indicators. In titrations of weak bases with strong acids the endpoint pH will be in the acidic range, and methyl red, methyl orange, and bromcresol green are commonly used indicators.

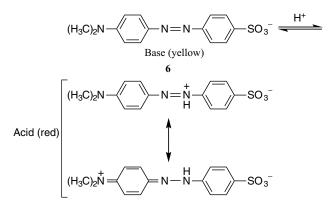
Without some understanding of the relationship of molecular structure to optical absorption spectra, a full accounting for the color changes of indicators is not possible, but an approximate treatment is feasible. The essential fact about acid–base indicators is that the acid and base forms have different colors. All acid–base indicators in common use are organic compounds. Apparently the reason for the different colors must be sought in the different structures of the acid and base forms of the indicator. It is possible to account for the fact of color differences on this basis: if two forms of the indicator differ markedly in their electronic distribution, and particularly in their extents of resonance delocalization, two colors will be observed. Color is associated with the capability of the compound to absorb visible light, and this capability can be related to the electronic structure. In the resonance hybrid several factors may contribute, but we can simplify and say that a change in the length of conjugation path or in extent of electronic delocalization will result in absorption of a different color component of white light, with a resultant color change. For a simple example we take 4-nitrophenol, one of the indicators in Table 12.3. The acid–base dissociation is



The acid form is colorless, but the base form is yellow. This yellow color can be correlated with electron delocalization in the base form as indicated in this conventional depiction of a resonance hybrid:



Possibilities for such electron delocalization increase with the size of the molecule, and most indicators are quite large molecules. Here is the kind of electron distribution responsible for the color change in methyl orange:



12.5. AQUEOUS SOLUBILITY OF WEAK ACIDS AND BASES

Many acidic and basic drugs possess a limited solubility in water, and we have a practical interest in being able to increase their solubility. We can often accomplish this by means of pH control. The pH of an aqueous solution can usually be adjusted independently of the acid–base equilibrium of the solute drug by means of a buffer solution.

This is the general principle that we apply—the total solubility is limited by the intrinsic solubility of the uncharged (nonionic) form of the drug. We can assume that the ionic form has unlimited solubility; this is not strictly true, but the assumption carries no practical drawbacks. Notice that for the first time in our consideration of acid–base chemistry we are directing our attention to the charge types of the species.

The experimental approach is to place enough of the solute in its ionic (charged) form to achieve the desired total concentration. This is accomplished either by raising the pH if the drug is an acid (thus deprotonating it) or lowering the pH if the drug is a base (thus protonating it). We therefore must recognize whether the drug is an acid or a base, and we require the pK_a in order to calculate the needed pH of the solution.

As noted in the preceding paragraph, we have two cases to consider: (1) a neutral weak acid, such as a carboxylic acid, to be symbolized HA; and (2) a neutral weak base, such as an amine, symbolized B. For the present, concentrations will be in molar units. We make these definitions:

Let s_0 be the equilibrium solubility of the neutral (uncharged) form of the drug. Let S_t be the total (apparent) solubility of the drug at any given pH of the solution.

We take it that the pH of the solution is under our control, for example by adding a buffer. Also note that we are restricting attention to compounds having a single

ionizable group. Then we can write in general

Total concentration = Solubility of uncharged form + Concentration of charged form

We will develop the two cases in parallel:

Neutral Acid, HA
 Neutral Base, B

$$S_t = [HA] + [A^-] = s_0 + [A^-]$$
 $S_t = [B] + [BH^+] = s_0 + [BH^+]$
 $K_a = \frac{[H^+][A^-]}{[HA]} = \frac{[H^+](S_t - s_0)}{s_0}$
 $K_a = \frac{[H^+][B]}{[BH^+]} = \frac{[H^+]s_0}{S_t - s_0}$
 $pK_a = pH - \log \frac{S_t - s_0}{s_0}$
 (12.56a)
 $pK_a = pH - \log \frac{s_0}{S_t - s_0}$
 (12.56b)

Observe that Eqs. (12.56a) and (12.56b) are simply forms of the familiar Henderson–Hasselbalch equation. The physical interpretation of these equations is that S_t is the maximum concentration of drug that can be achieved *at the given pH*. An alternative view is that the pH given by the equation is the limit beyond which precipitation of the uncharged drug will occur *at the given* S_t ; the pH directional change that will produce such precipitation depends on the solute; acids will precipitate as the pH is lowered, and bases will precipitate as the pH is raised.

Equations (12.56) contain two parameters (pK_a and s_0) and two variables (pH and S_t). In the most desirable situation, pK_a and s_0 will be available as experimentally measured quantities; otherwise they must be estimated. Methods for estimating the solubilities of nonelectrolytes are available (Chapter 10).

Let us examine Eqs. (12.56) more closely. As noted above, these equations contain the four quantities pK_a , pH, S_t , and s_0 . In most applications we will know pK_a and s_0 , and will either set a "target" S_t value and calculate pH, or set a target pH and calculate S_t . The concentrations appear as the ratio $(S_t - s_0)/s_0$ or its reciprocal, so it makes no difference whether molar units or physical units (such as mg mL⁻¹) are used, so long as S_t and s_0 are expressed in the same units.

The nonlogarithmic form of Eq. (12.56a) can be arranged to Eq. (12.57):

$$S_t = s_0 + \frac{K_a}{[\mathrm{H}^+]} s_0 \tag{12.57}$$

When $[H^+] \gg \gg K_a$ (i.e., when pH $\ll pK_a$), for this neutral acid, Eq. (12.57) becomes $S_t = s_0$. Essentially all of the solute is in the conjugate acid form. When $[H^+] \ll K_a$ (pH $\gg \gg pK_a$), Eq. (12.57) approaches

$$S_t = \frac{K_a}{[\mathrm{H}^+]} s_0$$

which can be written

$$\log S_t = \log s_0 + pH - pK_a \tag{12.58}$$

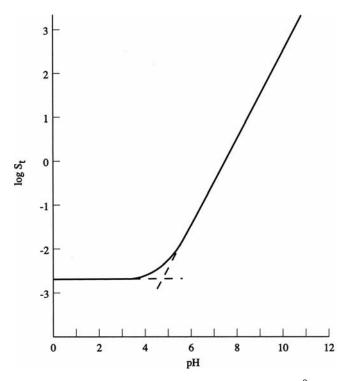


Figure 12.6. Plot of Eq. (12.56a); $pK_a = 5.0$, $s_0 = 2 \times 10^{-3}$ M.

Under these conditions the slope of a plot of log S_t versus pH is unity; that is, for each unit increase in pH the concentration increases tenfold. At the point where pH = p K_a , log $S_t = \log s_0$ [but this is an extrapolated condition, because Eq. (12.58) does not hold when pH = p K_a]. The reverse pH dependence (slope of the plot of log S_t vs. pH is -1.00) will be seen for a neutral base. Figure 12.6 shows this behavior for a hypothetical weak acid having p $K_a = 5.00$ and $s_0 = 2 \times 10^{-3}$ M.

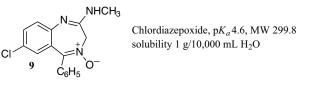
In applying either Eq. (12.56a) or Eq. (12.56b), we must first establish whether the drug is a neutral acid or a neutral base. Consider sodium benzoate, useful as a preservative. We recognize this as the sodium salt of benzoic acid, prepared by reacting benzoic acid with sodium hydroxide. The solute in this case is a neutral weak acid, even though we weigh it out and dissolve it as the salt (the charged conjugate base). Benzoic acid has a limited water solubility, whereas sodium benzoate is quite soluble. But we realize that sodium benzoate, in solution, is in equilibrium with benzoic acid, as the position of equilibrium is determined solely by the pK_a of benzoic acid and the pH of the solution. Despite the high solubility of sodium benzoate, if at the experimental pH the concentration placed in solution exceeds the S_t value given by Eq. (12.56a), free benzoic acid will precipitate until the equation is satisfied. We reiterate: The original form of the solute is irrelevant; this is an equilibrium situation entirely controlled by pK_a , pH, and s_0 .

We mentioned earlier that the pH is under our control, but this may not always be so, and in some cases the pH may be established by a complex mixture of buffers or other formulation components. Calculation of the pH may not be feasible in such circumstances; instead the pH should be measured. Even pH indicator paper may be adequate to this purpose.

Example 12.14

- (a) Suppose a drug is available as the sodium salt of a carboxylic acid whose molecular weight is 150, $pK_a = 5.00$, and solubility of the unionized acid form is 2×10^{-3} M. What is the maximum concentration of drug that can be dissolved at pH 5.00? Applying Eq. (12.56a) with $s_0 = 2 \times 10^{-3}$ M gives $S_t = 4 \times 10^{-3}$ M. Compare this with Fig. 12.6, which was calculated with these same parameters. Note that in this problem pH = pK_a , so equal concentrations of the ionized and unionized forms are present.
- (b) Can 0.5% of the sodium salt described in Example 12.14(a) be dissolved at pH 4.00? Using Eq. (12.56a) gives $S_t = 2.2 \times 10^{-3}$ M. The desired concentration is 0.5 g/100 mL = 5 g/L = 3.3×10^{-2} M, which far exceeds the calculated S_t . All drug exceeding 2.2×10^{-3} M will precipitate as the free acid, at pH 4.00.
- (c) For the drug of Examples 12.14(a) and 12.14(b), what pH range will permit 0.5% to be dissolved? Now we let $S_t = 3.3 \times 10^{-2}$ M, $s_0 = 2 \times 10^{-3}$ M, and $pK_a = 5.00$, applying Eq. (12.56a) to get pH = 6.19. Thus any pH equal to 6.19 *or higher* will allow 0.5% to be dissolved. (See Fig. 12.6; log 0.033 = -1.48.)

Example 12.15. In what pH range is it possible to prepare an aqueous solution of chlordiazepoxide (9) at a concentration of 10 mg/5 mL?



Chlordiazepoxide is obviously a neutral weak base. It is not obvious to which nitrogen atom the pK_a should be assigned, but its value is reasonable for an aromatic amine. The reported aqueous solubility corresponds to 0.01%. (The drug is also available as the hydrochloride salt, which is quite soluble, but as noted in the earlier discussion, this is entirely irrelevant to the problem.)

The desired concentration of 10 mg/5 mL is equivalent to 200 mg/100 mL or 0.2%. We can use Eq. (12.56b) with $s_0 = 0.01$ and $S_t = 0.2$. The result is pH = 3.3; that is, at any pH of 3.3 or below (more acidic), this concentration of drug can be dissolved.

We have emphasized that the form (neutral molecule or its salt) used experimentally is irrelevant, with the total achievable concentration depending solely on the parameters pK_a and s_0 and on the assigned pH. This is true when the pH is fixed by the experimentalist, by means of a buffer. But if control of pH is not important, then the simplest procedure is merely to dissolve the salt form, which is usually very soluble, to the desired concentration. The pH will shift to a value determined by pK_a and concentration. [And we know how to calculate this pH by using either Eq. (12.49) or Eq. (12.51).] But this procedure may be unacceptable if the resulting pH is outside the desired range. Such a solution, moreover, is unbuffered and is susceptible to perturbation of its pH by other ingredients.

12.6. NONAQUEOUS ACID-BASE BEHAVIOR

Although water is our most important solvent, in some applications we often make use of nonaqueous solvents. Acid–base behavior (reaction of acids with bases, attainment of acid–base equilibrium, acid–base indicator color changes) is widely observable in nonaqueous media, and here we survey this very large class of substances. It would be possible to develop quantitative acid–base theories for these solvents, but the results would be much more complicated than the acid–base theory of aqueous solutions owing to the lower dielectric constants of organic solvents. This circumstance leads to the formation of ion pairs (and even of ion triplets and higher aggregates), whose existence complicates the description, so we will be satisfied with a qualitative treatment.

Dissociating Solvents. Let us take water as our model of a dissociating solvent. Omitting the molecule of water that hydrates the hydrogen ion:

$$H_2O \rightleftharpoons H^+ + OH^-$$

A large number of *dissociating solvents* fit this pattern. Methanol and other alcohols give alkoxide ions, analogous to hydroxide in water:

$$MeOH \rightleftharpoons H^+ + MeO^+$$

It is important to realize that the symbol H^+ represents a different species in these two equations; in water it means H_3O^+ and in methanol it means $MeOH_2^+$.

Liquid carboxylic acids, of which glacial acetic acid is the most important, also are dissociating solvents. Letting Ac represent CH₃CO, the acetyl group, we obtain

$$AcOH \rightleftharpoons H^+ + AcO^-$$

(glacial acetic acid simply denotes pure acetic acid). In this equation, H^+ represents $AcOH_2^+$. The proton is not a necessary product of solvent dissociation. Here is how acetic anhydride dissociates:

$$Ac_2 O \rightleftharpoons Ac^+ + AcO^-$$

The symbol Ac⁺ represents the acetylium ion, CH_3CO^+ .

Solvent	pK _s	Dielectric Constant
Water	14.00	78.5
Methanol	16.70	32.6
Ethanol	19.10	24.3
Acetic acid	14.45	6.19
Formic acid	6.20	58
Acetic anhydride	14.5	21
Acetonitrile	26.5	36.2

Table 12.4. Properties of some solvents

There are also many nondissociating solvents, such as hydrocarbons, ethers, and carbon tetrachloride. Actually the distinction between dissociating and nondissociating solvents is a matter of degree, because with sufficiently sensitive techniques we might detect some level of dissociation by nearly any solvent. We adopt a practical viewpoint based on ordinary laboratory experience.

Let us generally represent a dissociating solvent by the symbol CA, where C is the cationic component and A is the anionic component. Then the solvent dissociation is

$$CA \rightleftharpoons C^+ + A^-$$

and, just as we did for water, we define an ion product, K_s :

$$K_s = [C^+][A^-] \tag{12.59}$$

Table 12.4 lists some ion products as $pK_s = -\log K_s$.

Here is an interesting consequence of the pK_s value. We will compare water and ethanol. In water, in order to pass from an acidic solution in which pC (i.e., pH) = 1 to an alkaline solution in which pA (i.e., pOH) = 1, a range of acidity corresponding to 12 pH units must be traversed, since pC + pA = 14 for water. For ethanol, however, pC + pA = 19.1, so to go from pC = 1 to pA = 1 requires that 17.1 orders of magnitude be covered. Very roughly, we may expect that the smaller the value of K_s for a solvent, the greater the range of the acidity scale available for studying or titrating sample solutes.

Acid–Base Properties. Solvents may be discussed as acids or bases just as are any other substances. The Bronsted theory forms the basis of the discussion, and the terms used are given in Table 12.5. Very generally we recognize that protogenic and amphiprotic solvents are dissociating solvents, whereas protophilic and aprotic solvents are nondissociating solvents.

A useful analogy may now be made between water, whose acid-base properties we understand, and several nonaqueous solvents. The dissociating solvent CA yields

Class	Characteristics	Examples
Protogenic	Acidic; donate a proton	Glacial HOAc; H ₂ SO ₄
Protophilic	Basic; accept a proton	Amines; ethers; esters
Amphiprotic	Both acidic and basic; can donate or accept a proton	Water; alcohols
Aprotic	Neither acidic nor basic.	Hydrocarbons, CCl ₄ ,CH ₃ CN, dioxane

Table 12.5. Solvent classifications

the cation C^+ , which is called the *lyonium ion*, and the anion A^- , called the *lyate* ion:

Solvent	\Rightarrow	Lyonium ion + Lyate
$2H_2O$	1	$H_3O^+ + OH^-$
2MeOH	\$	$MeOH_2^+ + MeO^-$
2AcOH	\$	$AcOH_2^+ + AcO^-$
Ac ₂ O	\$	$Ac^+ + AcO^-$
2NH ₃ (liq)	2	$NH_4^+ + NH_2^-$

With water as the solvent, we are accustomed to regarding its lyonium ion, H_3O^+ , as the strongest possible acid, and OH^- , the lyate ion, as the strongest possible base. Let us extend this concept to the other solvents. In glacial acetic acid as a solvent, we would conclude that the acetate ion AcO^- is the strongest possible base. In other words, sodium acetate (which we recall is a weak base in water) should be a strong base in glacial acetic acid, just as sodium hydroxide is a strong base in water. This expectation is borne out by experiment. Similarly, we predict that ammonium chloride should be a strong acid in liquid ammonia. This analogy is a powerful concept for the design of experiments.

The Leveling and Differentiating Effects. Since solvents can be acids or bases, an acidic or basic solute reacts with such a solvent to a degree determined by their relative strengths. We can distinguish two possibilities. Let S represent a basic solvent and HX a strongly acidic solute. Then one possibility is that the reaction

$$HX + S \rightleftharpoons SH^+ + X^-$$

goes essentially completely to the right. Thus, the solute is quantitatively transformed into the lyonium ion of the solvent, which is the strongest acid that can exist in this solvent. (Any acid stronger than SH⁺, such as HX, will be transformed to SH⁺; this lowers the free energy of the system.) We say that the solvent S is a *leveling solvent* for HX, or that HX is leveled by S. If we also had a second very strong acid HY that was leveled by S, the acids HX and HY would appear to be of the same strength, because they both have been converted to SH⁺. This is what happens when the familiar strong acids (HCl, HNO₃, H₂SO₄, HClO₄) are dissolved in water; they are leveled to H₃O⁺ and all appear to be of the same acid strength. Glacial acetic acid as a solvent, which is fairly acidic, levels many bases, such as amines, by quantitatively transforming them to its lyate ion:

$$RNH_2 + AcOH \rightleftharpoons RNH_3^+ + AcO^-$$

The other possibility, of course, is that the reaction between solvent and solute does not go to completion. Imagine the acid HA reacting with solvent S:

$$HA + S \rightleftharpoons SH^+ + A^-$$

If the reaction does not go completely to the right, we can measure an equilibrium constant for it, and this constant is a quantitative measure of the extent of reaction. Now if we take a second comparable acid HB and measure its equilibrium constant, we can compare the acid strengths of HA and HB with respect to the reference base S. We call S a *differentiating solvent* for HA and HB. This is just what we do in water when we measure K_a values for weak acids and bases.

Figure 12.7 is a schematic representation of these ideas. Here we have supposed that there exists for every solute an innate absolute acidity or basicity (which is not true, but is a reasonable practical approximation), and as solutes we have taken, in order of decreasing acidity and increasing basicity, $HClO_4$, AcOH, ArOH (a phenol), H_2O , $ArNH_2$ (an aromatic amine), RNH_2 (an aliphatic amine), NaOH. These are spread out on scales, which may be taken as proportional to pH or pK_a .

The key idea is that the pure solvent is taken as the neutral point of the scale. Thus in the upper scale, showing H₂O as the solvent, H₂O is the neutral point. Any solute, acid or base, lying very far from this point, will be leveled by water, because the difference in their acid–base properties is great. Those solutes not falling very distant from the neutral point are differentiated by water, as we know that AcOH, ArOH, ArNH₂, and RNH₂ are differentiated, because we can measure their K_a values.

Now turn to the scale for glacial acetic acid as solvent. Acetic acid now becomes our neutral point, and we may question whether even such a strong acid as perchloric acid $(HClO_4)$ is leveled by acetic acid. On the other hand, we readily accept that aliphatic amines are leveled, because they lie distant from the neutral point.

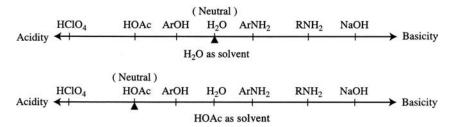


Figure 12.7. Illustrating acidity/basicity relative to the solvent, and the leveling and differentiating effects.

These considerations of acid–base strength are somewhat complicated by the concomitant phenomenon of limited dissociation as a result of the low dielectric constant of some nonaqueous solvents, but as a qualitative guide they are useful. The role of dielectric constant and the distinction between ionization and dissociation was treated earlier, in Chapter 8.

12.7. ACID–BASE STRUCTURE AND STRENGTH

Principles. We should be able to look at a molecular structure and to make reasonable, even though approximate, estimates of the acid or base strengths of functional groups. The ability to do this from fundamental theoretical principles is almost nonexistent at present and need not be considered. Quite sophisticated yet practical empirical techniques are available, but they are beyond our present requirements (Perrin et al., 1981). Our treatment will be very brief.

We begin by repeating Eq. (12.16),

$$\mathbf{p}K_w = \mathbf{p}K_a + \mathbf{p}K_b$$

along with the insight provided by the Bronsted theory that acid and base strength, for a conjugate pair, are reciprocally related, that is, $K_a = K_w/K_b$ or $K_b = K_w/K_a$. Now, it is the essence of the acid–base definitions that we can make these statements:

- 1. As K_a increases, pK_a decreases, and acid strength increases.
- 2. As K_b increases, pK_b decreases, and base strength increases.

For a conjugate pair, a smaller pK_a (stronger acid) must be accompanied by a larger pK_b (weaker base); this is the reciprocal effect. Observe in Table 12.6, these pairs of pK_a and pK_b values for (hypothetical) conjugate acid–base pairs.

It is obvious that pK_a is a reasonable quantitative measure of acid strength. What is not so obvious is that it has become conventional to use the pK_a of the conjugate acid to specify base strength! And Table 12.6 shows that a stronger base is associated with a larger pK_a (of its conjugate acid—but this parenthetical addition is seldom stated). So

	Conjugate Acid, pK_a	Conjugate Base, pK_b	
	3	11	
	4	10	
Î	5	9	S
cid	6	8	troj
r a	7	7	nge
Stronger acid	8	6	Stronger base
troi	9	5	ase
S	10	4	Ĵ
	11	3	

Table 12.6. Measures of acid and base strength for
hypothetical conjugate pairs

Conjugate Acid	pK _a	Conjugate Base	pK _b
$\begin{array}{c} CH_{3}COOH\\ H_{2}PO_{4}^{-}\\ NH_{4}^{+} \end{array}$	4.76	CH ₃ COO ⁻	9.24
	7.21	HPO ₄ ²⁻	6.79
	9.25	NH ₃	4.75

Table 12.7. pK_a and pK_b of some acids and bases

our chemical problem is twofold: (1) We must be able to recognize, by examination of the molecular structure, whether a functional group is acidic or basic and (2) we must estimate its pK_a value. Alternatively, if the pK_a is known, we must examine the structure to determine whether the pK_a describes an acid or a base.

Table 12.7 gives data for some important acid–base pairs. Thus we see that acetic acid is a stronger acid than is dihydrogen phosphate, which is stronger than ammonium ion. It follows inevitably that ammonia is a stronger base than monohydrogen phosphate, which is stronger than acetate ion. To find this set of data in the literature, one looks for the pK_a of acetic acid; the *second* $pK_a(pK_2)$ of phosphoric acid, H₃PO₄; and the pK_a of *ammonia*.

A very simplified view of pK_a prediction is often adequate. First we note that these commonly seen functional groups can be considered to have essentially no acidic or basic character in aqueous solution:

Alcohols and sugars, ROH Amides, RCONH₂ Ethers, ROR' Esters, RCOOR' Carbonyls, RCOR'; RCHO

Table 12.8 gives pK_a ranges that will include many of the commonly encountered acidic and basic functional groups. Recall that an aromatic amine has the nitrogeneither as part of the ring system (as in pyridine) or directly attached to an aromatic ring (aniline).

It is valuable to memorize (or otherwise keep readily available) a few typical pK_a values to serve as reference points. Here are some examples:

Acetic acid, CH ₃ COOH	р <i>К_а</i> 4.76
Benzoic acid, C ₆ H ₅ COOH	p <i>K_a</i> 4.20
Phenol, C ₆ H ₅ OH	p <i>K_a</i> 10.00
4-Nitrophenol, O ₂ N-C ₆ H ₄ OH	р <i>К_а</i> 7.14
Triethylamine, Et ₃ N	р <i>К_a</i> 10.78
Aniline, C ₆ H ₅ NH ₂	p <i>K_a</i> 4.69

Туре	pK _a
Acids	
Carboxylic acids, RCOOH	2-6 (3-5 typical)
Sulfonic acids, RSO ₃ H	-1 to 1
Phenols, ArOH	7–11
Thiols, RSH	7–10
Imides-CONHCO-	8-11
Bases	
Aliphatic amines	8-11
Aromatic amines	4–7
Guanidines, (RNH) ₂ C=NH	11–14

Table 12.8. pKa ranges of acids and bases

Bearing in mind the definitions of acid (proton donor) and base (proton acceptor), it is clear that a structural change that increases the electron density at a functional group will weaken an acid (raise its pK_a) by making it more difficult for the proton to leave; and it will strengthen a base (raise its pK_a) by more avidly attracting the proton, and vice versa for electron withdrawal from the functional group. This is why 4-nitrophenol is a much stronger acid than is phenol. With the few pK_a values given here and the ranges of Table 12.8, quite useful estimates can be made by analogy.

Structural Effects. We have seen that electron-withdrawing structural features are acid-strengthening and base-weakening, whereas electron-donating structural features are acid-weakening and base-strengthening. Discrete charges display these effects very clearly. Compare these data:

⁺ H_3 N-CH ₂ COOH \rightleftharpoons ⁺ H_3 N-CH ₂ COO ⁻ + H ⁺	$pK_a = 2.31$
$H_3C-CH_2COOH \rightleftharpoons H_3C-CH_2COO^- + H^+$	$pK_a = 4.88$
$^{-}O_{2}C$ -CH ₂ COOH $\rightleftharpoons ^{-}O_{2}C$ -CH ₂ COO ⁻ + H ⁺	$pK_a = 5.69$

Taking propionic acid as our reference, the pK_a of the positively substituted acid lies as expected, because a simple electrostatic argument states that like charge repulsion will facilitate the departure of the proton, thus enhancing acid strength. Just the opposite effect is seen with the negative substituent, which through unlike charge attraction inhibits the dissociation of the proton, weakening the acid. This is why pK_2 of a diprotic acid is always larger than pK_1 .

More subtle effects are seen with substituents capable of exerting electron release or electron withdrawal by inductive or resonance mechanisms. The aromatic ring provides good examples of such effects, and in fact much of our information on the electronic effects of substituents has come from pK_a measurements. Table 12.9 lists pK_a values for monosubstituted benzoic acids.

Some of the effects are easy to rationalize. For example, the nitro group is electronwithdrawing from any position. (The ortho substituent often is atypical because steric

Substituent	Position		
	Ortho	Meta	Para
-Н	4.20	4.20	4.20
-NO ₂	2.17	3.45	3.44
-Cl	2.94	3.83	3.99
-OCH ₃	4.09	4.09	4.47
-CH ₃	3.91	4.24	4.34
-C(CH ₃) ₃	3.46	4.28	4.40
-COOH	2.95	3.54	3.51
-COO ⁻	5.41	4.60	4.82
-OH	2.98	4.08	4.58
-NH ₂	4.98	4.79	4.92

Table 12.9. pK_a values of benzoic acids at 25°C

as well as electronic effects operate.) Similarly, the amino group is electron-releasing from every position. But some of the results may seem anomalous. Thus methoxy is acid-weakening in the para position but acid-strengthening in the meta position. How can this be explained?

Recall that both the inductive and resonance effects are present. The inductive effect (a through-bond displacement of electron density) is governed mainly by the electronegativity difference of the bonded atoms. The resonance effect is an electron delocalization resulting from molecular orbital overlap. These two effects may operate in the same directions, thus largely adding their effects; or they may oppose each other. In the methoxy case such opposition occurs; the methoxy group is electron-releasing by the resonance effect but electron-withdrawing by the inductive effect. In the para position the resonance effect dominates, but in the meta position resonance is largely ineffective, and the inductive effect dominates.

Table 12.10 gives pK_a values for phenols. The substituent effects on phenolic pK_a values are more marked than those on the benzoic acid series because the phenolic group can enter into direct conjugation with the substituent (as we saw in describing 4-nitrophenol as an acid–base indicator).

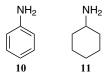
Substituent	Position		
	Ortho	Meta	Para
-H	10.00	10.00	10.00
$-NO_2$	7.23	8.35	7.14
-Cl	8.48	9.02	9.35
-OCH ₃	9.93	9.65	10.20
-CH ₃	10.28	10.08	10.19
-NH ₂	9.71	9.87	10.30

Table 12.10. pK_a values for monosubstituted phenols in water

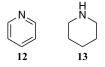
Amine	pK _a	
Ammonia, NH ₃	9.25	
Methylamine	10.64	
Ethylamine	10.67	
Dimethylamine	10.73	
Ethanolamine, HOCH ₂ CH ₂ NH ₂	9.50	
Hydroxylamine, HONH ₂	5.96	
Hydrazine, H ₂ NNH ₂	8.12	
Aniline	4.58	$\int AnK = 6.06$
Cyclohexylamine	10.64	$\int \Delta p R_a = 0.00$
Pyridine	5.17	$\Big\} \Delta p K_a = 6.06 \\ \Big\} \Delta p K_a = 5.96 $
Piperidine	11.13	$\int \Delta p \Lambda_a = 5.50$

Table 12.11. Amine pK_a values

Table 12.11 lists a few p K_a values of amines. These display the range of p K_a values typically seen with this class of compound, although even more dramatic substituent effects may be encountered; for example, p $K_a = 1.11$ for 4-nitroaniline. Comparison of the aromatic amine aniline **10** with its saturated analog cyclohexylamine **11** shows that the aromatic ring decreases base strength by a millionfold ($\Delta p K_a = 6.06$):

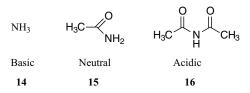


The same effect is seen with the pair pyridine and piperidine:



Although it may seem counterintuitive, the only conclusion that can be drawn is that the aromatic ring is responsible for reducing the electron density on the nitrogen and therefore must be functioning as an electron-withdrawing substituent (Brown et al., 1955). This is a resonance delocalization effect.

The appearance of the imide structure as an acid in Table 12.8 may be surprising, so let us view an imide as a product of successive acylations of ammonia:



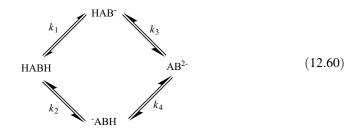
It follows that the acyl group is electron-withdrawing, reducing the electron density on nitrogen to such an extent that the basic ammonia molecule is converted to the neutral acetamide, with a second acetyl group producing an imide, which has an ionizable hydrogen. Many drug molecules, including the barbituric acid derivatives and the hydantoins, include the imide group. The sulfonamide drugs contain the functional group



which bears some relationship to the imide group, and is likewise acidic.

Assignment of pK_a Values.. To this point we have been considering the problem of predicting, to a semiquantitative level, the pK_a of a molecule from knowledge of its molecular structure, the method being based on analogy with pK_a values of model compounds. Now we face a related but distinctly different problem. Suppose a compound of known structure is studied experimentally and its pK_a value or values are measured. The problem is to associate these pK_a values with the functional groups responsible for them. This is called assigning the pK_a values.

If only a single pK_a is measurable, the problem is usually trivial, because there will be only a single reasonable choice of functional group to associate with the experimental value.⁹ We therefore turn to the more interesting case of a diprotic acid, which we symbolize HABH. In general the two ionizable hydrogens are associated with chemically different functional groups, so we must expand our description of the acid–base equilibria to accommodate two possible pathways:



Here HAB⁻ and ⁻ABH represent the two possible monoprotic species. The constants are called *microscopic dissociation constants* and are obviously defined as follows:

$$k_1 = \frac{[\mathrm{H}^+][\mathrm{HAB}^-]}{[\mathrm{HABH}]}$$

⁹ The choice is not always obvious, however, as is illustrated by the data in Example 12.15 for chlordiazepoxide.

$$k_{2} = \frac{[\mathrm{H}^{+}][-\mathrm{ABH}]}{[\mathrm{HABH}]}$$

$$k_{3} = \frac{[\mathrm{H}^{+}][\mathrm{AB}^{2-}]}{[\mathrm{HAB}^{-}]}$$

$$k_{4} = \frac{[\mathrm{H}^{+}][\mathrm{AB}^{2-}]}{[-\mathrm{ABH}]}$$
(12.61)

The observed stepwise dissociation constants are given by

$$K_{1} = \frac{[\mathrm{H}^{+}]([\mathrm{HAB}^{-}] + [^{-}\mathrm{ABH}])}{[\mathrm{HABH}]}$$
(12.62)

$$K_{2} = \frac{[\mathrm{H}^{+}][\mathrm{AB}^{2-}]}{[\mathrm{HAB}^{-}] + [^{-}\mathrm{ABH}]}$$
(12.63)

Algebraic combination of Eqs. (12.61)-(12.63) gives

$$K_1 = k_1 + k_2 \tag{12.64}$$

$$\frac{1}{K_2} = \frac{1}{k_3} + \frac{1}{k_4} \tag{12.65}$$

From Eqs. (12.61) we find

$$k_1k_3 = k_2k_4 \tag{12.66}$$

showing that only three of the four microscopic constants are independent. We might even define a fifth microscopic constant according to

$$HAB^- \rightleftharpoons^{K_{iso}} - ABH$$

and further manipulation results in

$$k_{\rm iso} = \frac{k_2}{k_1} = \frac{k_3}{k_4} \tag{12.67}$$

Before continuing with this general case, let us pause to analyze the special case in which the two ionizable groups are chemically identical and are independent of each other. The fact that they are identical means that $k_1 = k_2$ and that $k_3 = k_4$. The fact that they are independent means that $k_1 = k_3$ and $k_2 = k_4$. Inserting these special conditions

into Eqs. (12.64) and (12.65) gives $K_1 = 2k$ and $1/K_2 = 2/k$, or by combining these we obtain

$$\frac{K_1}{K_2} = 4 \tag{12.68}$$

This result is usually described as a statistical effect. Equation (12.68) is not closely obeyed by long-chain dicarboxylic acids because of the superimposed electrostatic effects of the anionic charges (Brown et al., 1955); in other words, the two groups, although identical, are not independent.

We now return to the general case in which the two functional groups are different. From Eqs. (12.64),(12.65) and (12.67) we get

$$K_1 = k_1 (1 + k_{\rm iso}) \tag{12.69}$$

$$K_2 = \frac{k_3}{1 + k_{\rm iso}} \tag{12.70}$$

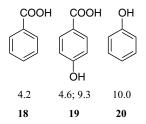
Now suppose that k_{iso} is very much smaller than one. From Eq. (12.67), this means that $k_2 \ll k_1$ and $k_3 \ll k_4$, or from Eqs. (12.69)–(12.70)

$$K_1 = k_1$$
 (12.71)

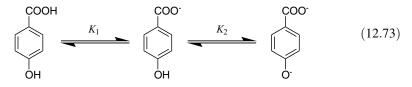
$$K_2 = k_3$$
 (12.72)

In this case the observed K_a values can be equated to microscopic constants. Chemically, this means that essentially only the uppermost pathway in Eq. (12.60) is followed, and the only monoprotic species is HAB⁻. The condition $k_{iso} \ll 1$ (or the reverse, $k_{iso} \gg 1$), which leads to a single ionization pathway, is satisfied if the observed p K_a values are widely spaced.

We now turn to specific chemical compounds. In making pK_a assignments, begin by writing the compound in its fully protonated form, whether this is uncharged or cationic. First consider 4-hydroxybenzoic acid. We place this in the center, with its measured pK_1 and pK_2 values, and flank it with very obvious model compounds and their pK_a values:

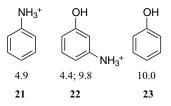


It is reasonable and even obvious to assign pK_1 to the carboxylic acid and pK_2 to the phenolic group. Moreover, since $K_1 \gg K_2$, we can expect that only a single pathway is followed and that the only monoprotic species present in significant concentration is the carboxylate. Thus for this compound the ionization pathway is

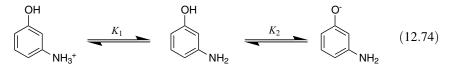


Incidentally, the differences between 4.6 and 4.2 and between 9.3 and 10.0 are to be ascribed to electronic substituent effects.

Next consider 3-aminophenol:



By analogy we assign the pK_1 to the amine and pK_2 to the phenol. Again the two dissociation constants are widely spaced, and this is the sequence:



Finally consider glycine, 25:

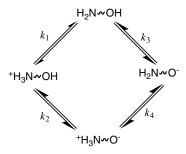
CH ₃ NH ₃ ⁺	⁺ H ₃ NCH ₂ COOH	CH ₃ COOH
10.7	2.4; 9.8	4.8
24	25	26

Once again by using model compounds as guides we make our assignment, this time of pK_1 to ionization of the carboxylic acid and pK_2 to the amine function. The ionization sequence is therefore

$${}^{+}\mathrm{H}_{3}\mathrm{NCH}_{2}\mathrm{COOH} \stackrel{K_{1}}{\rightleftharpoons} {}^{+}\mathrm{H}_{3}\mathrm{NCH}_{2}\mathrm{COO}^{-} \stackrel{K_{2}}{\rightleftharpoons} \mathrm{H}_{2}\mathrm{NCH}_{2}\mathrm{COO}^{-}$$
(12.75)

The intermediate species, carrying both a positive charge and a negative charge, is called a *zwitterion*.

What is the difference between 3-aminophenol (which does not yield a zwitterion) and glycine? Representing both molecules in the scheme



we see that 3-aminophenol follows the $k_1 \rightarrow k_3$ pathway, because the protonated amine, which is the conjugate acid of an aromatic amine, is a stronger acid than is the phenol. For glycine, on the other hand, the carboxylic acid is stronger than is the protonated amine (which in this compound is the conjugate acid of an aliphatic amine), so the $k_2 \rightarrow k_4$ route predominates.

If K_1 and K_2 are not widely separated, then both ionization routes are followed, and both monoprotic species may be present in the solution in significant concentration. Unique assignment of the pK_1 and pK_2 values then is not possible, and the interesting problem, which we will not pursue here, is to determine the values of the microscopic constants.

During our discussion of the relationship of acid–base strength to chemical structure, we have strayed from the path of classical equilibrium thermodynamics, because we have invoked nonthermodynamic concepts or "models" (such as the inductive and resonance effects of electron displacement) in aid of molecular-level interpretations. The equilibrium constants that we have called upon are thermodynamic, and the treatment embodied in Eqs. (12.60)–(12.71) is thermodynamic, but the assignment of p K_a values to functional groups lies outside of thermodynamics. Such interpretations are therefore referred to as *extrathermodynamic*. The distinction is worth keeping in mind because, valuable as the extrathermodynamic insights may be, they are provisional, subject to modification as new data or ideas become available, unlike the thermodynamic results, which do not rely on molecular models. Appendix C provides an introduction to the extrathermodynamic approach.

PROBLEMS

- 12.1. Calculate the pH of each of these aqueous solutions at 25°C.
 - (a) 2.50×10^{-4} M HCl
 - **(b)** $2.50 \times 10^{-4} \text{ M H}_2 \text{SO}_4$

- (c) 0.04 M ammonia
- (d) 0.04 M ammonium chloride
- (e) 3.0×10^{-3} M KOH
- **12.2.** A buffer was prepared by mixing 25.0 mL of 0.10 M acetic acid and 15.0 mL of 0.075 M KOH and then diluting to 50.0 mL with water. Calculate its pH $(pK_a = 4.76)$.
- **12.3.** Give directions for the preparation of 500 mL of 0.10 M pH 8.35 tris buffer, starting with pure tris and 0.10 M HCl. (See Example 12.7 for needed information.)
- **12.4.** Sorensen buffer solutions are prepared by mixing appropriate volumes of these stock solutions:

Stock solution A. 9.91 g of NaH_2PO_4 H_2O (MW 138.0) is dissolved in water to make 1 L.

Stock solution B. 9.47 g of Na_2HPO_4 (MW 142.0) is dissolved in water to make 1 L.

- (a) Calculate the molar concentrations of stock solutions A and B.
- (b) Calculate the pH of a Sorensen's buffer prepared by mixing 40 mL of A and 60 mL of B. Use $pK_2 = 6.80$ as the effective pK_2 of phosphoric acid.
- **12.5.** Calculate the titration curve for the titration of 24.0 mL of 0.20 M *n*-butylamine ($pK_a = 10.60$) with 0.30 M HCl when the following volumes of titrant have been added: 0 mL, 2 mL, 5 mL, 8 mL, 12 mL, 16 mL, 18 mL. Suggest a suitable indicator for the titration.
- 12.6. Calculate and plot the species distribution curves for these solutes.
 - (a) Phenol, $pK_a = 10.00$.
 - (**b**) Hydroxylamine, $pK_a = 5.96$.
 - (c) Phthalic acid, $pK_1 = 2.95$, $pK_2 = 5.41$.
 - (d) Citric acid, $pK_i = 3.06$, $pK_2 = 4.74$, $pK_3 = 5.40$.
 - (e) Is there any pH range within which significant concentrations of the uncharged forms of phenol and hydroxylamine can coexist in solution?
- 12.7. Give numerical values for the equilibrium constants of these reactions.
 - (a) $C_6H_5COOH + CH_3NH_2 \rightleftharpoons C_6H_5COO^- + CH_3NH_3^+$
 - (b) $C_6H_5OH + OH^- \rightleftharpoons C_6H_5O^- + H_2O$
- **12.8.** Assign the pK_a values of these compounds. (Look up the structures as necessary.)
 - (a) Salicylic acid; $pK_1 = 2.98$, $pK_2 = 13.00$.
 - **(b)** Arginine; $pK_1 = 2.17$, $pK_2 = 9.04$, $pK_3 = 12.48$.

- (c) Quinine; $pK_1 = 6.66$, $pK_2 = 9.48$.
- (d) Theophylline; $pK_a = 8.77$.
- **12.9.** For the titration of weak acid HA with strong base MOH, let $b = [M^+]$ and $c = [HA] + [A^-]$. Derive an *exact* equation relating $[H^+]$, K_a , K_w , b, and c throughout the entire titration. (*Hint:* Apply the systematic procedure outlined at the beginning of Section 12.3.)
- **12.10.** Write the electroneutrality equation for aqueous solutions of each of these solutes.
 - (a) Ammonium chloride.
 - (b) The triprotic neutral acid H_3A .
 - (c) A pH 7 phosphate buffer (assume sodium ion is the counterion).
- **12.11.** What is the pH of a 2.00% solution of ephedrine hydrochloride (MW 211.7, pK_a 9.60)?
- **12.12.** Given: 5.444 g of KH₂PO₄ (MW 136.1) was dissolved in 100.0 mL of 0.300 M KOH and the solution was diluted to 1000 mL. Calculate the pH ($pK_1 = 2.23$, $pK_2 = 7.21$, $pK_3 = 12.32$).
- **12.13.** Given: 50.0 mL of an aqueous solution of ammonia (pK_a 9.25) titrated with 0.100 M HCl, 9.50 mL of titrant being required.
 - (a) What was the pH of the solution when 4.75 mL of titrant had been added?
 - (b) What was the pH of the solution at the endpoint?
- **12.14.** Calculate the standard free-energy change at 25°C for the dissociation of phenol in water ($pK_a = 10.00$).
- **12.15.** What weight of anhydrous sodium acetate (MW 82.0) must be added to 1.00 L of pH 3.75 acetate buffer containing a total buffer concentration of 0.055 M in order to change the pH to 3.90 (p $K_a = 4.75$)?
- **12.16.** Show how a fraction and a ratio are related; in particular, how are the fraction F = [HA]/c and the ratio $R = [HA]/[A^-]$ related (*c* is the total concentration)?
- **12.17.** What is the concentration of benzoate ion in a solution prepared to be 0.10 M in acetic acid, 0.10 M in potassium acetate, and 5×10^{-4} M in (total) benzoic acid (p K_a = 4.75 for acetic acid; p K_a = 4.20 for benzoic acid)?
- **12.18.** Suppose that the pH of a solution of a diprotic weak acid H₂A is adjusted to be equal to $(pK_1 + pK_2)/2$.
 - (a) Will the concentration of the monoanion HA⁻ increase or decrease when some HCl is added?
 - (b) Will the concentration of the monoanion HA⁻ increase or decrease when some KOH is added?
- **12.19.** What is the pH at the midpoint of the titration of 15.0 mL of 0.050 M HCl with 0.10 M NaOH?

- **12.20.** You wish to prepare 1.00 L of pH 7.00 phosphate buffer containing a total phosphate concentration of 0.100 M. You have available crystalline NaH₂PO₄·H₂O (MW 138.0) and Na₂HPO₄ (MW 142.0). For phosphoric acid, $pK_1 = 2.23$, $pK_2 = 7.21$, $pK_3 = 12.32$. Neglecting activity coefficient effects, what weights of the two solutes must be taken?
- **12.21.** What is the standard free energy change of this reaction at 25°C?

$$H^+ + OH^- \rightleftharpoons H_2O$$

13

ELECTRICAL WORK

13.1. INTRODUCTION

In Chapter 1 we saw that work can be expressed as the product of an intensive property and an extensive property

 $Work(energy) = Intensity factor \times Capacity factor$

and these four examples were given:

Mechanical work = Mechanical force × Distance Work of expansion = Pressure × Volume change Surface work = Surface tension × Area change Electrical work = Electric potential × Charge

Mechanical work is dealt with in classical mechanics. In earlier chapters we treated expansion work and surface work. The present section develops the idea of electrical work. Recall from Chapter 3 that the Gibbs free-energy change in a reversible process when carried out reversibly is equal to the maximum work obtainable from the system (exclusive of work of expansion). One kind of useful work measured by the free-energy change is electrical work. We routinely exploit this application of thermodynamics when we use batteries.

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The essential phenomenon that we will study consists of a transfer of charge between an electrolyte solution and another phase (usually a solid). There are two ways in which this charge can be transferred: by electron transfer (the subject of Sections 13.2–13.4) and by ion transfer (Section 13.5), commonly called *ion exchange*.

13.2. OXIDATION-REDUCTION REACTIONS

Inorganic Redox Reactions. We saw in Chapter 12 that acid–base reactions are manifested as proton transfers from one conjugate acid–base pair to another. Now we encounter a formal analogy in the phenomenon of electron transfer. First we define an oxidation–reduction (*redox*) half-reaction:

$$\operatorname{Red} \underbrace{\xrightarrow{\text{oxidation}}}_{\text{reduction}} \operatorname{Ox} + ne \tag{13.1}$$

where "Red" is the reduced form of the reacting species (also known as the *reductant* or *reducing agent*), "Ox" is the oxidized form (the *oxidant* or *oxidizing agent*), *e* is the electron, and *n* is the number of electrons in the balanced half-reaction. Of course, in ordinary chemical systems we do not observe the half-reaction; instead two half-reactions are coupled, and the net process consists of one or more electrons being transferred from one redox pair to another:

$$\operatorname{Red}(1) = \operatorname{Ox}(1) + ne$$
$$ne + \operatorname{Ox}(2) \rightleftharpoons \operatorname{Red}(2)$$
$$\operatorname{Net:} \operatorname{Red}(1) + \operatorname{Ox}(2) \rightleftharpoons \operatorname{Ox}(1) + \operatorname{Red}(2)$$

Equation (13.1) shows that oxidation is the process in which a substance loses electrons and reduction is the process in which a substance gains electrons. Here are some simple examples of redox half-reactions.

$$H_{2} \rightleftharpoons 2H^{+} + 2e$$

$$Na \rightleftharpoons Na^{+} + e$$

$$Fe^{2+} \rightleftharpoons Fe^{3+} + e$$

$$2Cl^{-} \rightleftharpoons Cl_{2} + 2e$$

As in other types of chemical processes, redox reactions must be written in balanced form in order to express the experimental stoichiometry and to define equilibrium constants. For the simple examples shown above, it is easy to combine half-reactions into balanced net reactions, such as

$$H_2 + 2Na^+ \rightleftharpoons 2H^+ + 2Na$$

 $2Fe^{2+} + Cl_2 \rightleftharpoons 2Fe^{3+} + 2Cl^-$

Observe that a balanced redox reaction (whether a half-reaction or a net reaction) is balanced both chemically and electrically.

Some inorganic redox species take part in more complicated processes, and the balancing of their reactions is not intuitively obvious. For example, $Cr_2O_7^{2-}$ is reduced to Cr^{3+} . There is a systematic balancing procedure that serves to generate balanced redox reactions in all instances.

- *Step I.* Balance each half-reaction both chemically and electrically, using water and hydrogen ions where necessary. (We assume that the solvent is water.)
- *Step 2.* Equate the electron yield in the oxidation to the electron consumption in the reduction.
- Step 3. Add the balanced half-reactions, thus canceling electrons.
- *Step 4.* If necessary, reduce stoichiometric coefficients to whole numbers, and, if desired, express the balanced equation in molecular rather than ionic form.

Example 13.1. Balance the reaction in which potassium dichromate oxidizes ferrous ion. The unbalanced dichromate half-reaction is $Cr_2O_7^{2-} \rightleftharpoons Cr^{3+}$.

$$6e + Cr_2O_7^{2-} + 14H^+ \rightleftharpoons 2Cr^{3+} + 7H_2O$$

$$Fe^{2+} \rightleftharpoons Fe^{3+} + e$$
(Step 1)

In balancing the dichromate half-reaction, two Cr^{3+} are placed on the right side. Then 7H₂O are added to make up the oxygens in dichromate, and 14H⁺ are added to the left side to balance the water. Finally the 6 electrons balance the half-reaction electronically.

$$6e + Cr_2O_7^{2-} + 14H^+ \rightleftharpoons 2Cr^{3+} + 7H_2O$$

$$6Fe^{2+} \rightleftharpoons 6Fe^{3+} + 6e$$
(Step 2)

$$Cr_2O_7^{2-} + 6Fe^{2+} + 14H^+ \rightleftharpoons 2Cr^{3+} + 6Fe^{3+} + 7H_2O$$
 (Step 3)

$$K_2Cr_2O_7 + 6FeCl_2 + 14HCl \rightleftharpoons 2CrCl_3 + 6FeCl_3 + 7H_2O + 2KCl \qquad (Step 4)$$

Very effective analytical methods have been based on redox reactions. In redox titrations a reductant is titrated with an oxidant (or vice versa). The endpoint can be detected with a redox indicator, which is a substance whose oxidized and reduced forms exhibit different colors. Alternatively, an instrumental method of detection, to be described in Section 13.3, may be applied.

Organic Redox Reactions. The oxidation of organic compounds can be extremely complicated, and many of these reactions are not effectively reversible. Nevertheless,

this kind of reaction is important pharmaceutically because many drug molecules undergo degradative oxidation (Connors et al. 1986), and a brief discussion is appropriate. The essential concept is that the oxidation state of a carbon atom is a result of the number of bonds from carbon to oxygen; the greater the number of carbon–oxygen bonds, the more highly oxidized the carbon atom is.¹ The simplest example of this concept is provided by these one-carbon compounds:

Now, if oxidation is the addition of oxygen, then the reverse reaction must be reduction. This leads to the identification of reduction with the addition of hydrogen. Let us broaden this idea to consider the addition of hydrogen to an olefin:

$$H_2 + RCH = CHR \xrightarrow[oxidation]{reduction} RCH_2CH_2R$$

Since the hydrogenation reaction is a reduction, the reverse of this, a dehydrogenation, must be an oxidation. And now we see that we have an organic oxidation process that does not involve oxygen. We can apply the same idea to other reactions, as in the formal conversion of a sulfhydryl (mercaptan) group to a disulfide:

$$2RSH \xrightarrow[reduction]{\text{oxidation}} RSSR + H_2$$

In the presence of oxygen (as the oxidizing agent) this reaction proceeds according to

$$2RSH + \frac{1}{2}O_2 \xrightarrow[reduction]{\text{oxidation}} RSSR + H_2O$$

The mechanisms of organic redox reactions are seldom simple. Mechanisms (the detailed pathways from the initial to the final states) are investigated by the methods of kinetics and do not form part of the field of classical thermodynamics. (Despite this rather flat statement, a connection between the fields of thermodynamics and chemical kinetics can be made, and this connection is discussed in Appendix C.)

¹ This concept is really the genesis of the term *oxidation* as it is also applied to inorganic reactions. Compare FeO (ferrous oxide) and Fe_2O_3 (ferric oxide). The conceptual transformation from oxygen gain to electron loss is very broadening, but it obscures the historical basis. The term *reduction* originally arose in the processing of ores, when it can be said that an ore is reduced to the pure metal, the term referring to both a typical process (also historically called *revivification*) and a diminution in volume.

We have now seen oxidation described as an electron loss, as an addition of oxygen, or as a loss of hydrogen, with reduction as the reverse process. Here is a yet broader context. We noted that a reducing agent is a substance that yields electrons and, from Chapter 12, that a base is a substance that donates an electron pair. More generally we label as a *nucleophile* ("nucleus lover") a substance that furnishes electrons; thus reducing agents and bases are special cases of nucleophiles. Similarly, the class of *electrophiles* ("electron lovers") includes oxidizing agents and Lewis acids.

13.3. ELECTROCHEMICAL CELLS

Electrodes. In the present context, an electrode is a conductor of electricity immersed in an electrolyte solution. A transfer of charge may take place at the interface between the electrode surface and the solution. This charge transfer may result from the transfer either of electrons or of ions across the interface. Our present concern is with electron transfer, which arises from the occurrence of oxidation—reduction reactions.

Suppose that we assemble a system consisting of two electrodes, one each of two different redox half-reactions. One of these might be a piece of zinc metal partly immersed in a solution of zinc sulfate, with the other a piece of copper metal partly immersed in a solution of cupric sulfate. Assume that the solutions are in electrical contact but prevented from mixing and the metal electrodes are connected externally by a conductor, such as a length of copper wire. This assembly is an example of an *electrochemical cell*, this particular example being known as the *Daniell cell*. Figure 13.1 shows the Daniell cell.

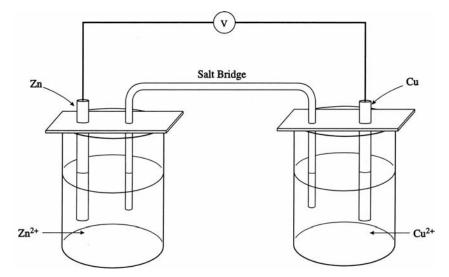


Figure 13.1. The Daniell cell. The salt bridge is a gel containing an electrolyte; it permits the passage of current, but prevents mixing of the solutions. [Reproduced by permission from Connors (1982).]

If we prepare the solutions each to be about 1 M, the redox half-reactions that take place in this cell will be

$$Zn \rightleftharpoons Zn^{2+} + 2e$$
$$2e + Cu^{2+} \rightleftharpoons Cu$$

Thus, oxidation occurs at the zinc electrode (removing metal from the solid zinc) and reduction occurs at the copper electrode (plating copper metal onto the solid copper). We will shortly learn how the direction of reaction can be predicted. On atomic and subatomic scales, electrons are being released at the zinc electrode, they flow through the external conductor, constituting an electric current, and they are consumed at the copper electrode by reduction of cupric ions. If a voltmeter is introduced to the external circuit, it will indicate a voltage difference between the electrodes. The *electric potential* between the electrodes is the source of this measured voltage. [Electric potential is also called *electromotive force* (emf).] Electric potential is a consequence of a difference in charge between two points; it is the electrical analog of the chemical potential.

There are two ways to operate an electrochemical cell:

- 1. If we simply connect the external leads, thus creating a closed circuit, the redox reactions at the electrodes occur as we have described, and the cell generates an emf, which we can use to do work. In fact, the cell is a battery. In this mode of operation the cell half-reactions are occurring spontaneously. The cell is called a *galvanic* or *voltaic cell* when it is operating spontaneously. The essential thermodynamic phenomenon taking place is that chemical energy is being transformed to electrical energy.
- 2. The second mode of operation of a cell opposes the cell potential with a greater external potential, thus reversing the cell reactions. Now we are converting electrical energy to chemical energy. This is how we recharge a battery. In this type of operation the cell is called an *electrolytic cell*. The cell reactions are occurring nonspontaneously, provided with energy from an external source to drive them in reverse.

Whether an electrochemical cell is operating as a galvanic cell (spontaneously) or as an electrolytic cell (nonspontaneously), the electrode at which oxidation occurs is called the *anode* and the electrode at which reduction occurs is called the *cathode*. In the Daniell cell of Fig. 13.1, for example, when operating spontaneously the zinc electrode is the anode and the copper electrode is the cathode, whereas in the nonspontaneous mode the copper electrode is the anode and the zinc electrode is the cathode.

All electrochemistry takes place at interfaces between phases. This is explicitly indicated in a shorthand notation for describing electrochemical cells. The physical state (gas, liquid, solid) may be shown if not obvious, and solution activities or

concentrations or gas pressure may be indicated. Here is the designation of the cell in Fig. 13.1:

$$Zn|ZnSO_4(1M)||CuSO_4(1M)|Cu$$

A single vertical line represents an interface across which a potential difference exists. This potential difference is called an *electrode potential*. The potential of the cell as a whole, which is what we measure, is the sum of the two electrode potentials (we cannot measure a single electrode potential because a single electrode does not constitute a closed electric circuit). The double vertical line signifies a *salt bridge*, which is an electrolyte solution (usually in a viscous medium like agar or gelatin) that establishes electrical contact between the electrode solutions while keeping them physically separated. The existence of the salt bridge introduces a complication, because at each end of the salt bridge there is another interface, and across these interfaces there are compositional differences. This means that charge differences must exist across each solution/salt bridge interface, and therefore electric potentials must exist. This liquid junction potential is small but is measurable, and it is difficult to account for theoretically in a rigorous and accurate way, so an experimental approach is taken to render it negligible. Since the source of the liquid junction potential is known to be the different mobilities of cations and anions, salt bridges are constructed with high concentrations of KCl as the electrolyte, because K⁺ and Cl⁻ have nearly the same mobility; thus the bulk of the current is carried by these ions, and the liquid junction potential is minimized. In extremely accurate work cells without liquid junctions may be used, but in usual laboratory situations our cells possess liquid junctions. We will not need to take the liquid junction potential into account.

The Nernst Equation. Consider the generalized chemical reaction

$$aA + bB \rightleftharpoons mM + nN$$

In Chapter 4, Section 4.3 we derived Eq. (13.2) for this system:

$$\Delta G = \Delta G^0 + RT \ln \frac{a_M^m a_N^n}{a_A^a a_B^b}$$
(13.2)

where a_i represents the activity of substance $i, \Delta G$ is the free-energy change (per mole), and ΔG^0 is the standard free-energy change. It is important to realize that the activities in Eq. (13.2) do not necessarily represent the values at equilibrium. If, however, we now impose the equilibrium condition, meaning that $\Delta G = 0$, we obtain the important relationship (which we have used frequently in earlier developments)

$$\Delta G^0 = -RT \ln K \tag{13.3}$$

where *K* is the equilibrium constant:

$$K = \frac{(a_M^m)_{eq}(a_N^n)_{eq}}{(a_A^a)_{ea}(a_B^b)_{eq}}$$
(13.4)

All the foregoing is familiar. We now make a connection to the electrochemical cell, in which the reaction is an oxidation–reduction reaction, and the chemical process results in electron transfer across the electrode–solution interfaces. The key idea is that if the cell operates reversibly (in the thermodynamic meaning of this term), the electrical work produced will be the free energy change for the process, which is the maximum work (not counting work of expansion) that the system can perform. The cell will operate reversibly if an infinitesimal amount of reaction occurs, and this condition requires that zero or an infinitesimal amount of current flow through the external circuit. This is accomplished by opposing the cell potential with an external potential such that zero current flows. The measured potential is then the cell potential at zero current.

From Section 13.1, electrical work is the product of potential and charge. Let *E* represent the cell potential [measured in volts (V)] and let *F* be the charge of one mole of electrons [measured in coulombs (C)], so that $F = eN_A = 96,485 \text{ C mol}^{-1}$. The quantity *F* is called the *Faraday*. Then for a redox reaction in which *n* electrons per molecule are transferred, we have

$$\Delta G = -nFE \tag{13.5}$$

where the negative sign is inserted so as to achieve agreement between the sign conventions applying to free-energy changes and cell potentials, as we will later see. By analogy with Eq. (13.5), we also write

$$\Delta G^0 = -nFE^0 \tag{13.6}$$

where E^0 , called the standard cell potential, is the value of the cell potential when reactants and products are in their standard states. For convenience let us define

$$L = \frac{a_M^m a_N^n}{a_A^a a_B^b} \tag{13.7}$$

where the activities need not be equilibrium activities. Now combining Eqs. (13.2) and (13.5)–(13.7) we get

$$E = E^0 - \frac{RT}{nF} \ln L \tag{13.8}$$

This is the *Nernst equation*, which relates the cell potential at zero current, E, to the activities (or concentrations, when activity coefficients are essentially unity) of the reactant and product species of the electrochemical cell reaction.

Equations (13.3) and (13.6) combined give

$$E^0 = \frac{RT}{nF} \ln K \tag{13.9}$$

where K is the equilibrium constant of the cell reaction.

At 25°C the Nernst equation takes the form

$$E = E^0 - \frac{0.059}{n} \log L \tag{13.10}$$

where the final term is now expressed in base 10 logarithms.² According to Eq. (13.10), a change in *L* by a factor of 10 results in a change in the cell potential of 59/n mV. This result is said to be a "Nernstian response" and is evidence that the electrochemical system is well-defined and is operating reversibly.

The foregoing equations allow us to make these associations:

- 1. If ΔG is negative, *E* is positive. The cell reaction is spontaneous; that is, with the reactant and product concentrations as specified, the reaction spontaneously proceeds from left to right.
- 2. If ΔG is positive, *E* is negative, and the reaction is nonspontaneous; it proceeds from right to left.
- 3. If the system is at equilibrium, then L = K, $\Delta G = 0$, E = 0. The cell can do no work.
- 4. If K > 1, E^0 is positive; if K < 1, E^0 is negative.

It is important to keep in mind that we have the experimental ability to (a) arrange the reactant and product concentrations at levels far from equilibrium and (b) maintain them in such a state by means of the salt bridge separation shown in Fig. 13.1, or an equivalent device; moreover, by holding the system in the zero-current condition, we achieve thermodynamic reversibility. However, if we allow current to flow, the available electrical work is less than the maximum work as measured or as calculated under conditions of reversibility.

Standard Potentials and Sign Conventions. We have seen that a functioning electrochemical cell requires a complete circuit, which means that it must possess two electrodes. We measure the cell potential. Although we cannot measure individual electrode potentials (because we would then have an open circuit), the concept of an electrode potential is so attractive that we adopt the view that a cell potential can be expressed as the sum of its electrode potentials. Consider again the Daniell cell:

² One joule = 1 volt coulomb, so $R = 8.314 \text{ J K}^{-1} \text{ mol}^{-1} = 8.314 \text{ V C K}^{-1} \text{ mol}^{-1}$. This provides *E* with the unit V.

According to this concept we write

$$E_{\rm cell} = E_{\rm Zn, Zn^{2+}} + E_{\rm Cu^{2+}, Cu}$$
(13.11)

Observe how the order of the subscripts tells which half-reaction is an oxidation and which is a reduction.³ An analogous equation relates the standard cell potential to the standard electrode potentials:

$$E_{\text{cell}}^{0} = E_{\text{Zn},\text{Zn}^{2+}}^{0} + E_{\text{Cu}^{2+},\text{Cu}}^{0}$$
(13.12)

Strictly speaking, the term *standard electrode potential* should be used only for the quantity describing the half-reaction as a reduction (IUPAC, 1993, p. 61), but we will relax this nomenclature, using the direction of the subscripts to make fully explicit whether the half-reaction is an oxidation or a reduction.

We must now consider the sign conventions relating to potentials. Two different concepts are involved, and some confusion is possible because historically we have used the words *positive* and *negative* to denote both a mathematical sign and a physical characteristic (Anson 1959).

Concept 1: The Potential of the Physical Electrode. We have seen that it would be convenient to have available values of individual electrode potentials, but we have also seen that we can only measure cell potentials. This difficulty is overcome by means of the universal agreement that the standard hydrogen electrode, diagrammed as

$$Pt, H_2(1 atm)|H^+(a = 1)$$

is to be assigned an electrode potential of zero volts at all temperatures. The electrode reaction is

$$\mathrm{H}_{2} \rightleftharpoons 2\mathrm{H}^{+} + 2e \qquad (E_{\mathrm{H}_{2} \mathrm{H}^{+}}^{0} = 0.00 \mathrm{V})$$

Now, if we form a cell of the standard hydrogen electrode with any other electrode, the measured cell potential is to be assigned to the second electrode. But besides its magnitude, this potential has a sign, which is determined by whether the second electrode has an excess or a deficiency of electrons relative to the hydrogen electrode. Suppose that we construct the following cell:

Pt, H₂(1 atm)|H⁺ (
$$a = 1$$
)||Zn²⁺ ($a = 1$)Zn

³ Some authors represent all electrode potentials as reductions, and then Eq. (13.11) will appear as

$$E_{\text{cell}} = E_{\text{Cu}^{2+},\text{Cu}} - E_{\text{Zn}^{2+},\text{Zn}}$$

This sign change is discussed later in the text.

We write the cell reaction with the left-hand electrode expressed as an oxidation and the right-hand electrode as a reduction:

$$H_2 + Zn^{2+} \rightleftharpoons 2H^+ + Zn$$

Of course, it is irrelevant to the physical system how we choose to write the reaction, and in the laboratory it is observed that, in this cell, the zinc electrode is negative relative to the hydrogen electrode; that is, oxidation occurs at the zinc electrode, releasing electrons. This is the physical characteristic alluded to earlier; it is a convention that we assign a negative charge to the electron, but it is a physical fact that oxidation in this cell occurs at the zinc electrode. The measured potential (at 25°C) is -0.76 V, and since we can write

$$E_{\text{cell}}^0 = E_{\text{H}_2,\text{H}^+}^0 + E_{\text{Zn}^{2+},\text{Zn}}^0 = -0.76 V$$

we can state that $E_{Zn^{2+},Zn}^0 = -0.76$ V. This is how standard electrode potentials are measured. Table 13.1 lists some standard electrode potentials. From the Nernst equation, Eq. (13.10), which can be written for either an electrode or a cell, the standard potential is equal to the potential when L = 1, and this condition is met when

Reduction Half-Reaction	$E^0(\mathbf{V})$
$\overline{\mathrm{Na}^+ + e} = \mathrm{Na}$	-2.71
$Zn^{2+} + 2e = Zn$	-0.76
$Fe^{2+} + 2e = Fe$	-0.44
$Ni^{2+} + 2e = Ni$	-0.25
$AgI(s) + e = Ag + I^{-}$	-0.15
$\mathrm{Sn}^{2+} + 2e = \mathrm{Sn}$	-0.14
$Pb^{2+} + 2e = Pb$	-0.13
$2\mathrm{H}^+ + 2e = \mathrm{H}_2$	(0.00)
$AgBr(s) + e = Ag + Br^{-}$	0.07
$S_4 O_6^{2-} + 2e = 2S_2 O_3^{2-}$	0.08
$\mathrm{Sn}^{4+} + 2e = \mathrm{Sn}^{2+}$	0.15
$\operatorname{AgCl}(s) + e = \operatorname{Ag} + \operatorname{Cl}^{-}$	0.22
$\mathrm{Hg}_{2}\mathrm{Cl}_{2}(\mathrm{s}) + 2e = 2\mathrm{Hg} + 2\mathrm{Cl}^{-}$	0.27
$Cu^{2+} + 2e = Cu$	0.34
$Cu^+ + e = Cu$	0.52
$I_3^- + 2e = 3I^-$	0.55
$\mathrm{Fe}^{3+} + e = \mathrm{Fe}^{2+}$	0.77
$Ag^+ + e = Ag$	0.80
$Cu^{2+} + I^- + e = CuI$	0.86
$\mathrm{Cl}_2 + 2e = 2\mathrm{Cl}^-$	1.36
$MnO_4^- + 8H^+ + 5e = Mn^{2+} + 4H_2O$	1.51
$H_2O_2 + 2H^+ + 2e = 2H_2O$	1.77

Table 13.1. Standard electrode potentials at 25°C

all reactant and product species are in their standard states. When determining standard potentials, the hydrogen electrode is always written on the left (as an oxidation) (IUPAC, 1993, p. 61).

If a cell, or indeed a reaction mixture, is constituted of any two half-reactions listed in Table 13.1, all species being in their standard states of unit activity, then the more negative E^0 value will represent the oxidation. Another way to say this is that the more positive the electrode potential, the greater its oxidizing power. Thus we can predict that permanganate (MnO₄⁻) will oxidize Fe²⁺ to Fe³⁺ and that iodine (as the triiodide ion I₃⁻) will oxidize (i.e., will be reduced by) thiosulfate, S₂O₃²⁻. This argument is made more systematic in Example 13.5 to follow.

Concept 2: The Potential of a Half-Reaction. From Eq. (13.5), $\Delta G = -nFE$, and from the thermodynamic result that a spontaneous reaction has a negative free energy change, we conclude that a spontaneous reaction has a positive potential. But we know that the reverse reaction is nonspontaneous, so its free-energy change is positive; hence its potential is negative. Therefore, in complete harmony with our practice in altering the sign of a thermodynamic quantity when we reverse the direction of the reaction, we also alter the sign of a potential when we reverse the direction of the reaction. Thus since $E_{Zn^{2+},Zn}^o = -0.76$ V for the half-reaction $Zn^{2+} + 2e = Zn$, we can write $E_{Zn,Zn^{2+}}^0 = +0.76$ V for the half-reaction $Zn = Zn^{2+} + 2e$.

From Eq. (13.9), this sign change convention is equivalent to converting an equilibrium constant to its reciprocal when a reaction is written in the reverse direction.

Applications of Electrode Potentials. Electrochemical calculations are not routinely necessary in the solution of pharmaceutical problems, so our treatment will be brief; but there are two reasons to include some level of discussion. The first is that these calculations offer a perfect example of the power of thermodynamics to predict the direction of chemical change. The second reason is to provide a sound basis for the discussions in Sections 13.4 and 13.5.

We begin with the Nernst equation

$$E = E^0 - \frac{0.059}{n} \log L \tag{13.13}$$

where

$$L = \frac{\prod a_p^{\nu}}{\prod a_r^{\nu}} \tag{13.14}$$

In Equation (13.14), Π signifies "the product of," a_p represents activity of product(s), a_r is the activity of reactant(s), and ν specifies the power to which each activity is raised; ν is the appropriate stoichiometric coefficient in the balanced cell reaction. The Nernst equation bears a formal resemblance to the Henderson–Hasselbalch equation, Eq. (12.36), and it is used in much the same way. The equation relates the three

quantities E, E^0 , and L, and if we know two of these, we can calculate the third. The methods will be illustrated with numerical examples.

Example 13.2. Calculate the potential of this cell at 25°C and predict the direction of the reaction. That is, will ferrous ion be oxidized or will ferric ion be reduced under these conditions?

$$Pt|Fe^{2+}(1 \times 10^{-6} M), Fe^{3+}(0.05 M)||KI(0.001 M)|Agl, Ag$$

The Pt is inert and serves only to make electrical contact between the solution and the external conductor (as in the hydrogen electrode).

Always begin by writing the balanced cell reaction, treating the left-hand electrode as the anode (where oxidation takes place). For this cell

$$Agl + Fe^{2+} \rightleftharpoons Fe^{3+} + Ag + I^{-}$$

We now set up the Nernst equation:

$$E_{\text{cell}} = E_{\text{cell}}^0 - 0.059 \log \frac{a_{\text{Fe}^{3+}} a_{\text{I}} - a_{\text{Ag}}}{a_{\text{Agl}} a_{\text{Fe}^{2+}}}$$

The standard electrode potentials are found in Table 13.1:

$$E_{\text{cell}}^{0} = E_{\text{Fe}^{2+},\text{Fe}^{3+}}^{0} + E_{\text{Agl},\text{Ag}}^{0}$$

= -0.77 - 0.15 = -0.92 V

Since Ag and AgI are solids, they are in their standard states of unit activity, and activities of solutes will be approximated by concentrations:

$$E_{\text{cell}} = -0.92 - 0.059 \log \frac{(0.05)(1 \times 10^{-3})}{(1 \times 10^{-6})}$$
$$= -0.92 - 0.059 \log 50 = -0.82 \text{ V}$$

Since the calculated cell potential is negative, the reaction as it has been written is nonspontaneous. Therefore, at these concentrations, ferric ion will be reduced to ferrous ion and silver will be oxidized to Ag^+ (which precipitates as AgI).

Example 13.3. What is the solubility product of silver bromide?

This may seem a peculiar question in the present context, because the solubility product (Chapter 10) does not describe a redox reaction.

$$AgBr(s) \rightleftharpoons Ag^+ + Br$$

 $K_{sp} = a_{Ag^+}a_{Br^-}$

However, we can "compose" this reaction from two redox half-reactions as follows:

$$AgBr(s) + e \rightleftharpoons Ag + Br^{-}$$

$$Ag \rightleftharpoons Ag^{+} + e$$

$$AgBr(s) \rightleftharpoons Ag^{+} + Br^{-}$$

From Table 13.1 we find

$$E_{\text{cell}}^{0} = E_{\text{AgBr,Ag}}^{0} + E_{\text{Ag,Ag}^{+}}^{0}$$

= 0.07 - 0.80
= -0.73 V

We now apply Eq. (13.9) at 25°C, with n = 1, or

$$\log K_{\rm sp} = \frac{E_{\rm cell}^0}{0.059}$$
$$= -12.37$$

or $K_{\rm sp} = 4.3 \times 10^{-13}$. It is often more accurate to determine such equilibrium constants from potential measurements than by direct chemical analysis.

Example 13.4. Calculate the potential of this Daniell cell:

$$Zn|Zn^{2+}(0.35 \text{ M})||Cu^{2+}(0.001 \text{ M})|Cu$$

The cell reaction is

$$Zn + Cu^{2+} \rightleftharpoons Zn^{2+} + Cu$$

The Nernst equation for this cell is

$$E_{\text{cell}} = E_{\text{cell}}^0 - 0.059 \log \frac{a_{\text{Zn}^{2+}} a_{\text{Cu}}}{a_{\text{Cu}^{2+}} a_{\text{Zn}}}$$

where n = 2. We find the standard electrode potentials in Table 13.1:

$$E_{\text{cell}}^{0} = E_{\text{Zn},\text{Zn}^{2+}}^{0} + E_{\text{Cu}^{+2},\text{Cu}}^{0}$$

= 0.76 + 0.34
= +1.10 V

Potentials are intensive quantities, so they are independent of the amount of reaction. From the equation $\Delta G = -nFE$ we see that $E = -\Delta G/nF$; a larger *n* value gives a correspondingly larger ΔG , but *E* is unaffected. This holds because the amount of oxidation is exactly balanced by the amount of reduction. Continuing with the calculation, we have

$$E_{\text{cell}} = +1.10 - \frac{0.059}{2} \log \frac{0.35}{0.001}$$

= +1.02 V

The cell reaction is spontaneous as written.

Example 13.5. Predict what will happen in this cell:

Ag, AgCl
$$|Cl^-(1 M)||Cl_2(p = 1 atm)|Pt$$

The cell reaction is balanced as follows:

$$2Ag + 2Cl^{-} \rightleftharpoons 2AgCl + 2e$$

$$Cl_2 + 2e \rightleftharpoons 2Cl^{-}$$

$$2Ag + Cl_2 \rightleftharpoons 2AgCl$$

We recognize that all species are in their standard states, so L = 1 and $E_{cell} = E_{cell}^0$. From Table 13.1

$$E_{\text{cell}}^{0} = E_{\text{Ag,AgCl}}^{0} + E_{\text{Cl}_{2}\text{Cl}}^{0} - \frac{1}{2} = -0.22 + 1.36$$
$$= +1.14 \text{ V}$$

The reaction is spontaneous as written; that is, Cl_2 oxidizes Ag. Note that in the silver–silver chloride electrode the chloride ion does not undergo a redox process, whereas in the chlorine electrode Cl^- is a product of a reduction.

Example 13.5 places on a formal basis the predictions made earlier when discussing standard potentials; these are seen to constitute a special case (the case L = 1) of the Nernst equation.

Example 13.6. What is the potential of this cell at 25° C?

$$Cu|Cu^{2+}(0.25 M)||Cu^{2+}(0.01 M)|Cu$$

Obviously the two electrodes are identical except for the ion concentrations. We proceed in the usual manner:

Left electrode :
$$Cu \rightleftharpoons Cu^{2+}(0.25 \text{ M}) + 2e$$
Right electrode : $2e + Cu^{2+}(0.01 \text{ M}) \rightleftharpoons Cu$ Overall reaction : $Cu^{2+}(0.01 \text{ M}) \rightleftharpoons Cu^{2+}(0.25 \text{ M})$

This is called a *concentration cell*. The Nernst equation applied to this cell is

$$E_{\text{cell}} = E_{\text{cell}}^0 - \frac{0.059}{2} \log \frac{0.25}{0.01}$$

where $E_{cell}^0 = E_{Cu,Cu^{2+}}^0 + E_{Cu^{2+},Cu}^0 = 0$. Thus we find $E_{cell} = -0.041$ V. The cell reaction is nonspontaneous, which accords with expectation; the spontaneous direction of diffusion is from higher to lower concentration (actually chemical potential).

The experimental measurement of electric potential is called *potentiometry*. When a redox titration is carried out with measurement of the potential as a function of titrant volume, the plot of potential E against titrant volume has the appearance of the acid–base titration curves that we studied in Chapter 12, and the endpoint is determined from the break in this potentiometric titration curve. By applying the Nernst equation at successive stages in the titration, it is possible to calculate such a curve.

13.4. pH MEASUREMENT

pH and the Hydrogen Electrode. The potential of certain electrochemical cells depends on the pH of the cell solution, and this dependence offers a means for the potentiometric measurement of pH. The hydrogen electrode is the classic type of a pH responsive electrode. Let us consider the following electrochemical cell

Pt,
$$H_2(p = 1 \text{ atm})|H^+(a_{H^+})|$$
 reference electrode

where the reference electrode need not be specified at present except to require that its potential not be affected by the pH of the cell solution. The hydrogen electrode consists of hydrogen gas bubbled through the aqueous solution; electrical contact is made through platinum metal, which is unreactive. Proceeding to describe this cell quantitatively, we write

$$E_{\text{cell}} = H_{\text{H}_2,\text{H}^+} + E_{\text{ref}}$$

The potential of the hydrogen electrode is expanded by means of the Nernst equation; the electrode reaction is

$$H_2 \rightleftharpoons 2H^+ + 2e$$

The result is (at 25° C)

$$E_{\text{cell}} = E_{\text{H}_2,\text{H}^+}^0 - \frac{0.059}{2} \log \frac{a_{\text{H}^+}^2}{p_{\text{H}_2}} + E_{\text{ref}}$$

We combine this equation with the convention $E_{\text{H}_2,\text{H}^+}^0 = 0$ and the definition $\text{pH} = -\log a_{\text{H}^+}$, obtaining

$$E_{\rm cell} = 0.059 \,\mathrm{pH} + E_{\rm ref}$$
 (13.15)

This last equation says that the cell potential is linearly related to the pH of the cell solution. The slope of the line is $dE_{cell}/dpH = 0.059$ V or 59 mV per pH unit. This is the theoretical response of a pH responsive electrode at 25°C. (In general, the response is 2.303 *RT/F* Vper pH unit.) This means that in order to measure pH to within 0.01 pH unit, we must be able to measure the cell emf to within 0.00059 V.

Practical Electrodes for pH Measurement. First we consider the reference electrode whose potential appears in Eq. (13.15). The most widely used reference electrode for practical pH measurements is the *saturated calomel electrode* (SCE). The SCE is composed of a saturated solution of potassium chloride that is also saturated with mercurous chloride (calomel). The solution is in contact with mercury metal, through which electrical contact is made with the external circuit. The SCE half-reaction is

$$Hg_2Cl_2 + 2e \rightleftharpoons 2Hg + 2Cl^-$$

and its potential, which is unaffected by pH, is + 0.244 V. A salt bridge must be placed between the SCE and the "test solution" whose pH is to be measured so that the SCE saturated solution is not diluted. Then a complete cell consisting of a hydrogen electrode and the SCE looks like this:

$$Pt, H_2(p = 1 \text{ atm})|H^+(a_{H^+})||KCl(sat), Hg_2Cl_2(sat)|Hg$$

Its cell potential is given as follows, by an argument identical to the preceding one:

$$E_{\text{cell}} = 0.059 \,\text{pH} - (E_{\text{SCE}} + E_{\text{lj}}) \tag{13.16}$$

where E_{lj} is the liquid junction potential.

Since E_{lj} is unknown, we are unable to apply Eq. (13.16) directly to calculate pH from a measured value of E_{cell} . We therefore proceed by, in effect, measuring differences in pH between two solutions. Let *E* represent the cell potential when the cell solution has a certain pH, and similarly let E_s be the cell potential when the cell solution has the value pH_s. Then writing Eq. (13.16) for both solutions and subtracting one equation from the other gives

$$pH = pH_s + \frac{E - E_s}{0.059}$$
(13.17)

at 25°C or in general

$$pH = pH_{s} + \frac{E - E_{s}}{2.303 \, RT/F}$$
(13.18)

The success of this procedure depends on the liquid junction potential remaining essentially constant as the pH is changed from pH to pH_S. This assumption is reasonable if pH and pH_s are not greatly different. Although Eq. (13.18) is not thermodynamically exact because of this assumption of the constancy of E_{lj} , it constitutes the operational definition of pH for practical laboratory work. If we know the value pH_S of a standard solution and measure its corresponding E_S, then replacement of the standard solution by the unknown test solution and measurement of *E* permits us to calculate the unknown pH by Eq. (13.18). Actually the calculation is automatically carried out by the electronic potentiometer (called a *pH meter*) used to measure the cell potential. The cell is "calibrated" by setting the pH meter to the known pH_S value with the standard solution in the cell. Then the standard solution is replaced with the test solution and the meter gives the unknown pH value. An adjustment on the meter calculates the value of 2.303 *RT/F* for the experimental temperature.

Obviously the accuracy of this pH measurement procedure depends on the accuracy with which pH_s values are known. For all pH measurement in aqueous solutions the pH_s values given in Table 13.2 will suffice. These pH_s values were determined on cells without liquid junctions by Bates and co-workers (Bates 1962; Staples and Bates 1969). (Ordinary laboratory work yields pH measurements

$t(^{\circ}C)$	Tartrate	Citrate	Phthalate	Phosphate (1:1)	Phosphate (1:3.5)	Borax	Carbonate
0		3.864	4.003	6.984	7.534	9.464	10.321
5	_	3.839	3.999	6.951	7.500	9.395	10.243
10	_	3.819	3.998	6.923	7.472	9.332	10.178
15	_	3.802	3.999	6.900	7.448	9.276	10.116
20	_	3.788	4.002	6.881	7.429	9.225	10.060
25	3.557	3.776	4.008	6.865	7.413	9.180	10.012
30	3.552	3.767	4.015	6.853	7.400	9.139	9.968
35	3.549	3.759	4.024	6.844	7.389	9.102	9.928
38	3.548	_	4.030	6.840	7.384	9.081	
40	3.547	3.754	4.035	6.838	7.380	9.068	9.892
45	3.547	3.750	4.047	6.834	7.363	9.038	9.856
50	3.549	3.749	4.060	6.833	7.367	9.011	9.825
55	3.554	_	4.075	6.834		8.985	
60	3.560	_	4.091	6.836		8.962	
70	3.580		4.126	6.845		8.921	
80	3.609	_	4.164	6.859		8.885	
90	3.650	_	4.205	6.877		8.850	
95	3.674	—	4.227	6.886	—	8.833	

Table 13.2. Standard pH_s values

Solution	т	Substance	Weight $(g)^b$	pH _s at 25°C
Tartrate	~0.034	KHC ₄ H ₄ O ₆	Saturated at 25°C	3.557
Citrate	0.05	KH ₂ C ₆ H ₅ O ₇	11.41	3.776
Phthalate	0.05	KHC ₈ H ₄ O ₄	10.12	4.008
Phosphate (1:1)	$\left\{ \begin{array}{l} 0.025 \\ 0.025 \end{array} \right.$	KH ₂ PO ₄ Na ₂ HPO ₄	$3.39 \\ 3.53 $	6.865
Phosphate (1:3.5)	$\left\{\begin{array}{c} 0.008695\\ 0.03043 \end{array}\right.$	KH ₂ PO ₄ Na ₂ HPO ₄	$\left. \begin{array}{c} 1.179\\ 4.30 \end{array} \right\}$	7.413
Borax	0.01	$Na_2B_4O_7 \cdot 10H_2O$	3.80	9.180
Carbonate	${0.025 \\ 0.025}$	NaHCO ₃ NaCO ₃	$2.092 \\ 2.640 $	10.012

Table 13.3. Compositions of standard buffer solutions^a

^aBates 1962; Staples and Bates 1969.

^bWeight of substance (in air near sea level) per liter of solution, prepared with carbonate-free distilled water.

accurate at best to ± 0.01 pH unit.) Table 13.3 gives the compositions of these standard pH buffer solutions.

We have seen that the cell should be standardized with a standard buffer whose pH_s is close to the anticipated pH of the test solution in order to minimize variation in the liquid junction potential. It is also good experimental practice to measure the pH of a second standard buffer after standardization against the first one; this step checks the correct operation of the electrode system, the pH meter, and the standardization of the cell.

To this point we have treated the hydrogen electrode as the pH-responsive electrode. The hydrogen electrode, though thermodynamically well-defined, is not very practical for routine use, so alternatives have been sought, and in all practical laboratory and field measurements the pH-responsive electrode is a device known as the *glass electrode*. This electrode possesses a very thin glass membrane separating the test solution from a solution of fixed pH. A potential is developed across this glass membrane that is related to the pH difference on the two sides of the glass. (This is an ion-transfer effect, not a redox phenomenon; it is treated in Section 13.5.) A practical cell for pH measurement then has this form:

Ag|AgCl(sat), HCl|glass|test solution||SCE

The silver–silver chloride electrode serves to make electrical connection between the glass membrane and the external circuit. The pH response of this electrode is Nernstian, so the measurement of pH with a glass–SCE cell follows Eq. (13.16).

 pK_a Determination. Accurate values of pK_a can be measured with a glass–SCE cell if appropriate attention is paid to temperature control and standardization of the electrodes. For weak acid HA the thermodynamic constant K_a is defined as

$$K_a = \frac{a_{\rm H^+} a_{\rm A^-}}{a_{\rm HA}}$$
(13.19)

The quantity we actually measure in the laboratory, however, is called the apparent constant, K'_a , defined by

$$K'_{a} = \frac{a_{\mathrm{H}^{+}}[\mathrm{A}^{-}]}{[\mathrm{HA}]}$$
(13.20)

This is because the glass-SCE cell gives a pH value defined by

$$pH = -\log a_{H^+}$$
(13.21)

and because we prepare solutions to have known *concentrations* of A^- and HA.⁴ Our first problem is to measure pK'_a ; next we must correct pK'_a to pK_a .

From Eq. (13.20) we obtain

$$pK'_{a} = pH - \log \frac{[A^{-}]}{[HA]}$$
(13.22)

and from Eqs. (12.33) and (12.34) we have

$$pK'_{a} = pH - \log \frac{b + [H^{+}] - [OH^{-}]}{a - [H^{+}] + [OH^{-}]}$$
(13.23)

where *b* is the concentration of the counterion to A^- and a + b is the total solute concentration: $a + b = [HA] + [A^-]$. Since we know *a* and *b* from the manner in which the solution was prepared, and we measure pH, we can calculate pK'_a .⁵ A series of solutions can efficiently be prepared, each solution having varied values of *b* such that the pH of all solutions will be within about 1 unit of pK_a (so as to ensure good buffer capacity). The pH values of the solutions are measured, and pK_a is calculated for each solution. If pH is in the approximate range of 4–10, the $[H^+]$ and $[OH^-]$ quantities in Eq. (13.23) will usually be negligible.

It will be seen that this procedure is essentially a titration of the weak acid HA with the strong base MOH, where $b = [M^+]$. If the pK_a of a weak base B is to be measured, the usual procedure is to titrate B with strong acid HX in an analogous manner. Now $a + b = [BH^+] + [B]$ and $a = [X^-]$. Equation (13.23) is again applicable.

The same value for pK'_a should be obtained whether we start with the conjugate acid (e.g., acetic acid) and titrate with strong base, or start with its conjugate base (e.g., sodium acetate) and titrate with strong acid, provided that the ionic strength is substantially identical in the two titrations.

⁴ For any practical purpose Eq. (13.21) is an adequate interpretation, but we must recall that this cell possesses a liquid junction. Moreover, Eq. (13.21) implies that we can measure the activity of a single ionic species, whereas we have seen in Chapter 8 that we can measure only a mean ionic activity.

⁵ There is a subtlety here. From the pH measurement we know the *activity* of H^+ but Eq. (13.23) calls for the *concentrations* of H^+ and OH^- . Activity coefficient corrections can be applied, but this will seldom be necessary.

With the apparent constant pK'_a at hand, we turn to the problem of finding the thermodynamic constant pK_a . For reasons discussed in Chapter 8, pK'_a varies with the ionic strength of the solution, so it is not a true constant. On the other hand, pK_a is defined to be a constant (at fixed temperature and pressure); in other words, pK_a is that unique value of pK'_a in the reference state of the solute where activity coefficients are unity. We can correct a pK'_a value for nonideal behavior by applying the Debye–Hückel theory (Section 8.3).

Consider the uncharged acid HA, whose apparent constant pK'_a is defined in Eq. (13.20). We may take it that at low to moderate values of ionic strength the activity of the uncharged form HA is equal to its concentration. The activity of the ion A⁻, however, is given by

$$a_{\mathrm{A}} = \gamma_{\mathrm{A}^{-}} [\mathrm{A}^{-}]$$

where γ_{A^-} is the mean ionic activity coefficient. With these identities, Eqs. (13.19) and (13.20) become

$$K_a = \frac{a_{\mathrm{H}^+} \gamma_{\mathrm{A}} [\mathrm{A}^-]}{a_{\mathrm{HA}}}$$

and

$$K'_a = \frac{a_{\mathrm{H}^+} \left[\mathrm{A}^-\right]}{a_{\mathrm{HA}}}$$

or by combining these equations we obtain

$$K_a = \gamma_A - K'_a$$

which in logarithmic form becomes

$$\mathbf{p}K_a = \mathbf{p}K'_a - \log \gamma_{A^-} \tag{13.24}$$

Now we use the Debye–Hückel equation, Eq. (8.26), in Eq. (13.24):

$$pK_a = pK'_a + \frac{0.509\sqrt{I}}{1+\sqrt{I}}$$
(13.25)

The ionic strength I is calculated with

$$I = \frac{1}{2} \sum c_i z_i^2$$
(13.26)

where *c* represents the concentration of species *i* and z_i is its charge; the summation includes all the ions in the solution. Equation (13.25) corrects pK'_a to pK_a for uncharged acids.

For a positively charged acid such as BH⁺, the ionic activity coefficient appears in the denominator of K_a , with the result that Eq. (13.24) takes the form

$$pK_a = pK'_a + \log \gamma_{BH^+} \tag{13.27}$$

leading to

$$pK_a = pK'_a - \frac{0.509\sqrt{I}}{1 + \sqrt{I}}$$
(13.28)

Equations (13.25) and (13.28) apply at 25° C.

Example 13.7. Suppose that 10.0 mL of a solution 0.10 M with respect to both acetic acid and sodium chloride was mixed with 10.0 mL of 0.05 M sodium hydroxide at 25°C. The pH of this solution was 4.64. What is the thermodynamic pK_a of acetic acid?

The total solute concentration (i.e., acetic acid plus acetate ion) is 0.05 M; this is equal to a + b in the symbolism of Eq. (13.23). The quantity b is given by (10.0)(0.05)/20.0 = 0.025 M. From Eq. (13.23), we obtain $pK'_a = 4.64$, because the quantity [H⁺] is negligible.

The ionic strength receives contributions from both the sodium chloride (0.05 M) and the sodium acetate (0.025 M); from Eq. (13.26), I = 0.075 M. Finally, Eq. (13.25) gives

$$pK_a = 4.64 + \frac{(0.509)(0.274)}{1.274} = 4.75$$

Example 13.8. Given: 0.0541 g of 4-cresol (MW 108.1) dissolved in 47.5 mL of water at 20°C. After 1.0 mL of 0.10 M KOH was added, the pH was 9.55; after the addition of 3.0 mL of 0.10 M KOH, the pH was 10.29. Calculate the apparent constant and the thermodynamic constant.

We calculate that $0.0541/108.1 = 5.00 \times 10^{-4}$ mol of solute, or 0.5 mmol, was taken. When 1.0 mL of strong base titrant had been added, the volume was 48.5 mL, so the total solute concentration was 0.5/48.5 = 0.01031 M; this is the quantity a + b. The value of b is (1.0) (0.10)/48.5 = 0.002062. Since the pH is quite high, let us apply the correction for [OH⁻]. At 20°C, pK_w = 14.17 (Table 12.1), so pOH = 4.62 and [OH⁻] = 2.4×10^{-5} . Equation (13.23) gives

$$pK'_{a} = 9.55 - \log \frac{0.002062 - 0.000024}{0.00825 + 0.000024}$$
$$= 10.16$$

The ionic strength of this solution is (neglecting the hydroxide) equal to b, or 0.00206 M. Applying Eq. (13.25) gives $pK_a = 10.16 + 0.02 = 10.18$.

After the addition of 3.0 mL of titrant, we calculate these quantities: $a + b = 0.00990 \text{ M}; b = 0.00594 \text{ M}; [OH⁻] = 1.3 \times 10^{-4}$, and Eq. (13.23) gives

$$pK'_{a} = 10.29 - \log \frac{0.00594 - 0.00013}{0.00396 + 0.00013}$$
$$= 10.14$$

The ionic strength is now 0.00607 M, and Eq. (13.25) gives $pK_a = 10.14 + 0.04 = 10.18$.

The data in Example 13.8 are from Albert and Serjeant (1984), who give much useful information on pK_a determination. These numerical examples are helpful in showing typical magnitudes of the activity coefficient corrections, which in Example 13.8 appear to be only slightly greater than the usual experimental uncertainty in the pH measurement.

An alternative approach to estimating pK_a from measurements of pK'_a is to note that Eq. (13.25) can be written in the form

$$pK'_{a} = pK_{a} - \frac{0.509\sqrt{I}}{1 + \sqrt{I}}$$
(13.29)

which suggests that if pK'_a is measured as a function of ionic strength, a plot of pK'_a against $\sqrt{I}/(1 + \sqrt{I})$ will be a straight line, which can be extrapolated to yield pK_a as the intercept. Figure 13.2 is a plot of Eq. (13.29) for β -naphthol (Albert and Serjeant 1984). The line extrapolates to $pK_a = 9.63$; its slope is -0.505.

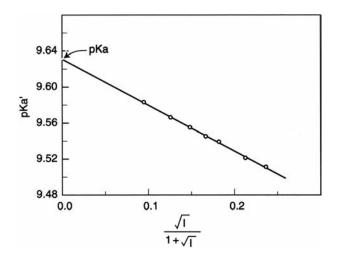


Figure 13.2. Plot according to Eq. (13.29) for β -naphthol at 20°C (Albert and Serjeant 1984).

13.5. ION-SELECTIVE MEMBRANE ELECTRODES

Except for our brief discussion of the pH-sensitive glass electrode, all the preceding treatment of electrical phenomena has been based on electron transfer across the electrode–solution interface. We now turn to the ion transfer mechanism as the basis of an electrochemical cell.

Theory of the Electrode Response. Before we examine the physical nature of the electrode substance, we will simply designate it as a "membrane"; it is the site of ion exchange with the electrolyte solution. A schematic representation of an electrochemical cell incorporating such a membrane electrode is

Internal reference electrode $|a_{int}|$ Membrane $|a_{ext}|$ External reference electrode

The activities refer to the ion for which the membrane serves as an ion exchanger; a_{int} is the internal (fixed) ion activity and a_{ext} is the sample solution ion activity. The reference electrodes are connected to the external circuit. The potential of this cell is given by

$$E_{\text{cell}} = E_{\text{int ref}} + E_{\text{ext ref}} + E_{\text{lj}} + E_{\text{membrane}}$$
(13.30)

Although we anticipate that the membrane response is a consequence of ion exchange between the membrane and the solution, we can model this formally as the result of two redox half-reactions. Suppose the ion in question is the cation M^{n+} . Then

Left(oxidation) :
$$M_{int} \rightleftharpoons M_{int^-}^{n+} + n(e)$$
Right(reduction) : $n(e) + M_{ext^-}^{n+} \rightleftharpoons M_{ext}$ Overall reaction : $M_{ext}^{n+} \rightleftharpoons M_{int}^{n+}$

Note that, in the usual manner, we write the oxidation at the left electrode. (We do not include M_{int} and M_{ext} in the overall reaction because, as pure metal, they are in the standard state of unit activity.) Comparison of the development to this point with Example 13.6 reveals that the membrane electrode is formally part of a concentration cell. The Nernst equation gives

$$E_{\text{membrane}} = -\frac{RT}{nF} \ln \frac{a_{\text{int}}}{a_{\text{ext}}}$$

which can be combined with Eq. (13.30) to yield

$$E_{\text{cell}} = \text{constant} + \frac{RT}{nF} \ln a_{\text{ext}}$$

or at 25°C we obtain

$$E_{\text{cell}} = \text{constant} + \frac{0.059}{n} \ln a_{\text{ext}}$$
(13.31)

Thus we predict Nernstian behavior of this cell.

Suppose that the exchangeable ion is an anion. Then the development is based on these redox reactions:

Left (oxidation) :
$$2X_{int}^{-} \rightleftharpoons (X_2)_{int} + 2e$$
Right (reduction) : $2e + (X_2)_{ext} \rightleftharpoons 2X_{ext}^{-}$ Overall reaction : $X_{int}^{-} \rightleftharpoons X_{ext}^{-}$

The membrane potential is given by

$$E_{\text{membrane}} = -\frac{RT}{nF} \ln \frac{a_{\text{int}}}{a_{\text{ext}}}$$

and the analog to Eq. (13.31) becomes

$$E_{\text{cell}} = \text{constant} - \frac{0.059}{n} \ln a_{\text{ext}}$$
(13.32)

Summing this up, we anticipate an electrode response following the form

$$E_{\text{cell}} = \text{constant} \pm \frac{0.059}{n} \ln a_{\text{ext}}$$
(13.33)

with the + sign applying to cations and the - sign to anions.

We next turn to the mechanisms for ion-exchange between the membrane and the solution.

Glass Electrodes. A typical glass is composed of Na_2O and SiO_2 , and its surface layers possess exchangeable cations. An ion exchange equilibrium is set up between the glass surface and an electrolyte solution according to

$$-SiO^{-}Na^{+} + \underset{(solution)}{H^{+}} \rightleftharpoons -SiO^{-}H^{+} + \underset{(solution)}{Na^{+}}$$

When a pH-responsive glass electrode is first placed in water this process occurs, and when equilibrium has been reached the electrode is said to be hydrated and is ready for use. The full thickness of a glass membrane is typically 0.03-0.1 mm, and the thickness of the hydrated layers (internal and external) is only about 10^{-4} mm (Evans 1987).

Now, when this hydrated glass electrode is placed in a sample solution, in general the hydrogen ion activities in the external hydrated glass layer and in the sample solution will be different. This difference gives rise to the potential described by Eq. (13.31). In practice there can be some complicating factors. The hydrogen ion activities of the internal and external hydrated layers are seldom exactly equal, and this difference gives rise to an *asymmetry potential*, which is compensated for in the calibration process. Another complication occurs when the pH is being measured in solutions containing high sodium ion concentrations. In this case a potential difference may develop based on the different activities of sodium ion in the glass and in the solution; in effect, the electrode (or the experimentalist) is fooled into thinking that the sample solution hydrogen ion activity is higher than it really is. Since the usual source of high sodium ion concentrations is NaOH, this experimental artifact is called the *glass electrode alkaline error*. Newer glasses replace some of the sodium content with lithium and so are less susceptible to the alkaline error.

The foregoing description of the sodium ion error of the glass electrode is suggestive of an opportunity to modify the glass composition and in this way to develop glass electrodes with enhanced selectivity for ions other than H. This approach has been successful. Glasses are described in a shorthand manner by the symbol NAS X-Y, meaning that this glass contains X% of Na₂O, Y% of Al₂O₃, and 100 - (X + Y)% of SiO₂. The conventional pH glass electrode has little Al₂O₃, and its sensitivity pattern is

$$H^+ \gg Na^+ >, Rb^+, Cs^+, \gg Ca^{2+}$$

NAS 11-18 glass has this sensitivity sequence:

$$Ag^+ > H^+ > Na^+ \gg K^+, Li^+, > Ca^{2+}$$

This glass has been used as a sodium ion-selective electrode. Obviously Ag^+ must be absent and H^+ must be fixed, but its advantage is its high sensitivity to Na^+ relative to K^+ or Li⁺. Many of the newer glasses, especially the lithium glasses, possess analytically useful selectivity profiles. Glass membrane electrodes are particularly valuable because, although they are physically delicate, they are chemically quite robust.

Solid Membrane Electrodes. A sparingly soluble crystalline solid can serve as an ion-exchange electrode. Usually, crystals of the substance are dispersed in a polymer matrix. Suppose we wish to design a halide ion (X^-) -sensitive electrode. We can form such an electrode with the slightly soluble AgX salt. The membrane sets up this solubility equilibrium with the solution:

AgX_{membrane}
$$\xrightarrow{K_{sp}}$$
 Ag⁺_{soln} + X⁻_{soln}

where K_{sp} is the solubility product (Chapter 10).

Presuming that the sample solution (into which the electrode is introduced) initially contains no Ag^+ but a relatively high concentration of X^- , the common ion effect of X^- will depress the solubility of AgX, and the concentration (activity) of the silver ion will be determined by that of X^- :

$$a_{\rm Ag^+} = \frac{K_{\rm sp}}{a_{X^-}} \tag{13.34}$$

From Eq. (13.31), therefore

$$E_{\text{cell}} = \text{constant} + 0.059 \ln a_{\text{Ag}^+} \tag{13.35}$$

or by making use of Eq. (13.34) we obtain

$$E_{\text{cell}} = (\text{constant})' - 0.059 \ln a_{X^-}$$
 (13.36)

where a_{X^-} refers to the total activity of X⁻ in the solution. This has an interesting consequence. Reverting to concentrations for convenience, we can write

$$[\mathbf{X}^-]_{total} = [\mathbf{X}^-]_{initial} + [\mathbf{X}^-]_{dissolved}$$

where $[X^-]_{initial}$ is the quantity sought from the measurement and $[X^-]_{dissolved}$ is the concentration contributed by dissolution of the electrode substance. Because of the stoichiometry of the dissolution process, $[X^-]_{dissolved} = [Ag^+]$, so we find

$$[\mathrm{Ag}^+] = [\mathrm{X}^-]_{\mathrm{total}} - [\mathrm{X}^-]_{\mathrm{initial}}$$

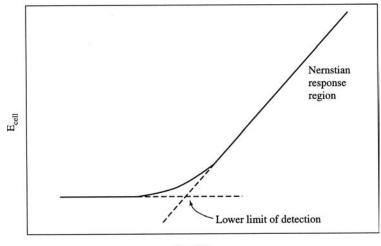
Restating Eq. (13.34), we also have

$$a_{\mathrm{Ag}^+} = \frac{K_{\mathrm{sp}}}{\left[\mathrm{X}^-\right]_{\mathrm{total}}}$$

Combining these gives a quadratic in $[X^-]_{total}$ whose solution is

$$[\mathbf{X}^{-}]_{\text{total}} = \frac{[\mathbf{X}^{-}]_{\text{initial}} \pm ([\mathbf{X}^{-}]_{\text{initial}}^{2} + 4K_{\text{sp}})^{1/2}}{2}$$
(13.37)

There are two extreme cases to consider. If $[X^-]_{initial}^2 \gg 4K_{sp}$, then we find that $[X^-]_{total} = [X^-]_{initial}$. This means that the dissolved anion makes a negligible contribution to the total, and that in this region of behavior a tenfold increase in $[X^-]_{initial}$ results in a 59-mV change in cell potential [Eq. (13.36)]. At the other extreme of behavior, where $[X^-]_{initial}^2 \ll 4K_{sp}$, Eq. (13.37) yields $[X^-]_{total} = \sqrt{K_{sp}} = [X^-]_{dissolved}$; here the initial concentration of anion is so low that it makes no sensible contribution, and from Eq. (13.36) we find that the potential is independent of $[X^-]_{initial}$. At intermediate conditions the dependence is more



log [X-] initial

Figure 13.3. Electrode response curve of a solid membrane electrode.

complicated, as shown in Fig. 13.3. The sensitivity of the system may be defined as the slope of the response curve in Fig. 13.3; thus the electrode is considered to lose useful sensitivity below the lower limit of detection.

Solid-state electrodes are quite rugged. Their sensitivity limits are, as we have seen, controlled by the K_{sp} value of the solid. Mixed solids also have been used. For example, an electrode composed of CuS and Ag₂S can respond to Ag⁺, S²⁻, or Cu²⁺; the solubility products of both salts must be simultaneously satisfied. Some lower limits of detection of solid-state electrodes are, for Cl⁻, 10⁻⁵ M, for I⁻, 10⁻⁸ M, for Cu²⁺, 10⁻⁹ M, and for Bi³⁺, 10⁻¹¹ M. These are very high sensitivities.

Liquid Membrane Electrodes. We are not accustomed to picturing liquid membranes, but the term simply implies a thin liquid phase. As the sample solution is aqueous, the liquid membrane phase will be an organic liquid that is immiscible with water. Hence the liquid membrane is very hydrophobic. By making the membrane phase quite thin, a faster response is achieved. Figure 13.4 shows some simple designs of liquid membrane electrodes. The electrode design of Fig. 13.4a will produce a slow response because of the considerable distance between the inner and outer interfaces, whereas Fig. 13.4b shows a faster responding electrode.

The basic idea is that an equilibrium is established within the membrane phase between the sample ion of interest and some hydrophobic molecule with which the ion forms a noncovalently bound complex. The sample ion thus is capable of exchanging between the membrane phase and the external aqueous sample solution. This ion exchange process is responsible for developing the potential at the interface as we have described earlier.

In one general type of electrode, a hydrophobic anion is dissolved in a hydrophobic solvent. The anion is selected so that it will form a strong complex with the

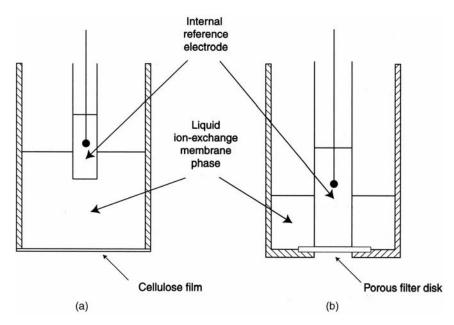


Figure 13.4. Schematic diagrams of liquid ion-selective membrane electrodes. [Reproduced by permission from Connors (1982).]

exchangeable cation of interest. Since the anion is hydrophobic, it is constrained to remain in the membrane phase, but the cation can exchange between the membrane and the aqueous sample solution. A calcium-selective electrode is based on this principle. The complexing agent is bis(di-*n*-decyl)phosphate, and this is the equilibrium within the membrane phase

$$Ca^{2+} + 2R_2P \bigvee_{OH}^{O} \xrightarrow{(R_2PO_2)_2Ca + 2H^+}$$

where R is $CH_3(CH_2)_9$. Clearly this electrode will suffer interference from hydrogen ions, and it is found to be usable in the pH range 5.5–11. (Above pH 11 calcium hydroxide precipitates.)

The alternative version of this electrode design makes use of a hydrophobic cation, chosen to form a complex with an exchangeable anion. Thus a nitrate ion-selective electrode is based on the ion pair formation of NO_3^- with the tridodecylhexadecyl-ammonium ion; we can represent this equilibrium as

$$NO_3^- + R_4N^+ \rightleftharpoons R_4N^+ NO_3^-$$

The second general type of liquid membrane electrode is based on an uncharged complexing agent. A potassium ion electrode uses the antibiotic valinomycin as a selective complexing agent. Much synthetic organic research is leading to the availability of macrocyclic compounds capable of complex formation with a wide variety of smaller species, and many of these compounds may be suitable as complexes in liquid membrane electrodes. An especially attractive possibility is to develop electrodes specific for drug molecules. Since most drugs are acids or bases, they are easily transformed into ions by pH control, and then their electrochemical detection becomes possible if a complexing agent can be identified that will selectively bind the ionic drug. Many such electrodes have been developed.

PROBLEMS

- **13.1.** Balance these equations:
 - (a) $ClO_3^- + Sn^{2+} = Cl^- + Sn^{4+}$ (b) $PbO_2 + I^- = I_2 + Pb^{2+}$ (c) $OCl^- + NH_3 = Cl^- + N_2$ (d) $MnO_4^- + H_2O_2 = Mn^{2+} + O_2$ (e) $MnO_4^- + CuI = MnO_2 + Cu^{2+}$ (f) $S_2O_8^{2-} + 2Fe^{2+} = 2SO_4^{2-} + 2Fe^{3+}$ (g) $NH_2OH + Ce^{4+} = N_2O + Ce^{3+}$ (h) $RSH + I_2 = RSSR + I^-$
- **13.2.** Calculate the ionic strength of a phosphate buffer prepared to be 0.025 M in NaH₂PO₄, 0.025 M in K₂HPO₄, and 0.050 M in NaCl.
- **13.3.** Calculate the change in cell potential when a glass electrode–SCE pair is moved from 0.010 M HCI to 0.010 M KOH. (Assume that activities are equal to concentrations.)
- **13.4.** Given this electrochemical cell, where "tris" is tris(hydroxymethyl)aminomethane and trisH⁺ is protonated tris:

Pt, $H_2(p = 1 \text{ atm}) |tris(0.05M), trisH^+(0.05M)||SCE$

The potential of the SCE is +0.244 V and the measured cell potential was +0.717 V at 25°C. What is the p K'_a of trisH⁺?

13.5. A cell consisting of a calcium ion-sensitive membrane electrode and a reference electrode gave the following cell potentials at 25° C with solutions having the listed Ca²⁺ activities:

Ca ²⁺ Activity	$E_{\text{cell}}(\mathbf{V})$
0.000135	+0.136
0.00541	+0.183
0.0382	+0.208

- (a) Determine whether the cell response is Nernstian.
- (b) Predict the cell potential if the Ca²⁺ activity is 1.00×10^{-3} .
- **13.6.** (a) What is the ionic strength of a solution 0.01 M in KCl and 0.01 M in Na_2SO_4 ?
 - (b) Calculate the mean ionic activity coefficient of sulfate ion in this solution.
- **13.7.** (a) Calculate the cell potential at 25°C of this electrochemical cell:

- (b) Will copper metal be plated out or will it be dissolved in this cell?
- **13.8.** (a) Calculate the potential of this cell at 25° C:

Zn|Zn(NO₃)₂(0.01 M), AgNO₃(0.001 M)|Ag

(b) Will this cell produce zinc-plated silver or silver-plated zinc?

13.9. What is the equilibrium constant of this reaction at 25° C?

$$AgCl + Br^{-} \rightleftharpoons AgBr + Cl^{-}$$

13.10. Standard potentials are given in parentheses for the following reactions:

$$\begin{split} \mathrm{Mg(OH)}_2 + 2e \ \rightleftharpoons \ \mathrm{Mg} + 2\mathrm{OH}^- \ (E^0_{\mathrm{Mg(OH)}_2\mathrm{Mg}} = \ -2.69 \ \mathrm{V}) \\ \mathrm{Mg}^{2+} + 2e \ \rightleftharpoons \ \mathrm{Mg} \qquad (E^0_{\mathrm{Mg}^{2+},\mathrm{Mg}} = \ -2.37 \ \mathrm{V}) \end{split}$$

Calculate the solubility product of magnesium hydroxide.

14

NONCOVALENT BINDING EQUILIBRIA

14.1. INTRODUCTION

The principles of thermodynamics apply to all physical and chemical equilibria, and the purpose of the present section is to concentrate attention on yet another class of chemical equilibria, namely, those processes in which the noncovalent forces of interaction are operative. The distinction between covalent ("chemical") bonds and noncovalent ("physical") interactions is not clearcut, and it is based in part on theory and in part on an empirical criterion. Theoretically, using the language of molecular orbital theory, a covalent bond is a consequence of the formation of molecular orbitals from atomic orbitals; the bond usually consists of one or more electron pairs shared between the bound atoms, and is highly directional in character. Noncovalent interactions, on the other hand, are electrostatic in nature and are not highly directional. The empirical criterion is energetic; covalent bonds tend to be strong whereas noncovalent interactions are weak. Obviously this is not an absolute criterion. The single, double, and triple bonds familiar from organic chemistry are perfect examples of covalent bonds. The interactions that are responsible for the existence of condensed phases (solids and liquids) are noncovalent. Notice that we refrain from speaking of noncovalent "bonds"; this restraint is a reflection of the weakness of these interactions as well as their nondirectional nature. Most noncovalent interactions exhibit energies of $\sim 10 \text{ kcal mol}^{-1}$ or less (ΔH^0 values), whereas covalent bond energies largely lie in the range 20-200 kcal mol^{-1.1}

¹ The classic treatment of covalent bonding is Linus Pauling's *The Nature of the Chemical Bond* (1960). Of course, more recent sources must be consulted for later theoretical and experimental findings.

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The noncovalent interactions occur between two or more molecules or ions. The manner in which we designate these interactants is arbitrary and may be determined for our convenience. We choose to call one of the interactants the *substrate* S and the other interactant the *ligand* L. Usually some chemical or physical property of the substrate, a property that is altered on interaction with the ligand, is observed by the experimenter. The product of the interaction between the substrate and the ligand is generally called a *complex*. Complexes may form between two (or more) small molecules or ions; or small molecules may bind to a macromolecule, as when a drug–protein complex is formed. Noncovalent complex formation is an essential step in many biological processes, such as antigen–antibody interaction, transport of ions across membranes, control of metabolic pathways, signal transduction, oxygen transport, and the regulation of gene expression.

14.2. THE NONCOVALENT INTERACTIONS

Potential Energy Functions. The detailed nature of the noncovalent interaction forces is not the concern of thermodynamics, but some understanding of these phenomena will be helpful in picturing the molecular processes being studied. Although we commonly speak of the forces of interaction, it is actually the energies of interaction that are of primary interest. The force F is related to the potential energy V by

$$F = -\frac{dV}{dr}$$

where r is the distance between two interacting particles. Physical theory has elucidated the potential energy functions of the noncovalent interactions (Hirschfelder et al. 1954; Israelachvili, 1985). The interacting species may be ions or molecules, and the molecules may be nonpolar or may possess permanent dipole moments.

Table 14.1 gives the noncovalent potential energy functions, which are seen to fall into three classes. The Table 14.1 functions are written for interactions between species S and L in vacuo, that is, in the absence of a solvent. These are the three classes of noncovalent interactions:

- 1. *Electrostatic Interactions*. These take place between ions (*C*) or dipoles (μ). The magnitude of a charge is to be accompanied by its sign, and a negative value of energy is attractive, a positive value repulsive. (The charge–charge interaction term is Coulomb's law, which we encountered in Chapter 8.) Observe that charges and dipole moments appear as squared quantities.
- 2. Induction (Polarization) Interactions. These are a result of an ion or a dipole inducing a temporary dipole in an adjacent molecule. The interaction then consists of the electrostatic interaction between the temporary dipole and the permanent dipole or the ion. The quantity α in the expressions is the polarizability of the molecule, a measure of the ease with which the molecule's electron cloud can be deformed in the presence of an electric field.

Type of Interaction		Potential Energy Function
	Electrostatic	
Charge-charge		$+\frac{C_{\rm S}C_{\rm L}}{r}$
Charge-dipole		$+ \frac{C_{\rm s}C_{\rm L}}{r} \\ - \frac{1}{3k{\rm T}} \cdot \frac{C_{\rm s}^2 \cdot \mu_{\rm L}^2}{r^4} \\ - \frac{2}{3k{\rm T}} \cdot \frac{\mu_{\rm s}^2 \cdot \mu_{\rm L}^2}{r^6}$
Dipole-dipole		$-\frac{2}{3k\mathrm{T}}\cdot\frac{\mu_{\mathrm{S}}^{2}\cdot\mu_{\mathrm{L}}^{2}}{r^{6}}$
	Induction	
Charge-induced dipole		$-\frac{C_{\rm S}^2 \cdot \alpha_{\rm L}}{2r^4} \\ -\frac{\mu_{\rm S}^2 \cdot \alpha_{\rm L}}{r^6}$
Dipole-induced dipole		$-rac{\mu_{ m S}^2\cdotlpha_{ m L}}{r^6}$
	Dispersion	
Induced dipole-induced dipole		$-\frac{3}{4} \left[\frac{\in_{S} \cdot \in_{L}}{\in_{S} + \in_{L}} \right] \frac{\alpha_{S} \cdot \alpha_{L}}{r^6}$

Table 14.1. Potential energy functions for noncovalent interactions^a

^{*a*}C is the charge on an ion, μ is permanent dipole moment, α is polarizability, *r* is intermolecular distance, \in is a specific energy term, *T* is absolute temperature, and *k* is Boltzmann's constant, where $k = R/N_A$.

3. *Dispersion (London) Interaction.* This is a quantum-mechanical phenomenon, although we can give a reasonable classical interpretation. Any molecule, including a nonpolar molecule, may develop a momentary dipole as a result of transitory electron density displacements. This momentary dipole can induce a dipole in a neighboring molecule, and these two temporary dipoles then interact. The dispersion interaction takes place between all species; its effectiveness is dependent on their polarizabilities.

Note that, for uncharged molecules, each class of interaction includes an r^{-6} dependence on the internuclear separation distance *r*. For such interacting species the combined effect of these r^{-6} terms constitutes the noncovalent interactions.² Under the attractive influence of these terms, two molecules approach until they experience an opposing repulsive force, which initially is a result of electron–electron repulsion and, at even closer distances, of nucleus-nucleus repulsion. The net effect of these forces of attraction and repulsion is reasonably expressed in Eq. (14.1), an empirical function called the Lennard-Jones 6–12 potential.

$$V = 4V_{\min}\left[\left(\frac{r_0}{r}\right)^{12} - \left(\frac{r_0}{r}\right)^6\right]$$
(14.1)

² Some authors refer collectively to these r^{-6} terms (r^{-7} in the forces) as *van der Waals* interactions, but there is some ambiguity in this term, because certain authors seem to mean only the London force by the designation van der Waals.

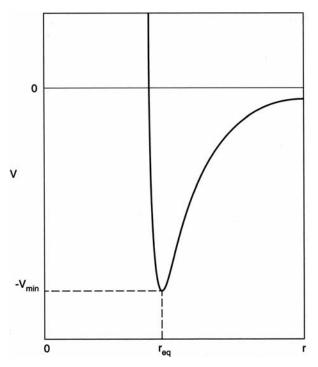


Figure 14.1. Potential energy dependence on intermolecular distance according to the Lennard-Jones potential function.

In Eq. (14.1), which is shown graphically as Fig. 14.1, V is the potential energy of interaction between two uncharged particles separated by distance r. V_{\min} is the value of V at the equilibrium separation r_{eq} , which is the value of r when the attractive and repulsive forces exactly balance each other; r_0 is the value of r when V=0. The r^{-12} term empirically expresses the repulsive energy. Observe the r^{-6} term, which is attractive and embodies the r^{-6} terms in Table 14.1.

The potential energy functions in Table 14.1 are written for interactions in vacuo, that is, without a solvent present. But we are interested in condensed phase systems, most commonly consisting of substrate S and ligand L dissolved in a solvent. In such real systems the noncovalent interactions will be moderated to a degree expressed by the dielectric constant of the solvent, in the manner made explicit in our treatment of the Coulomb interaction (Chapter 8).

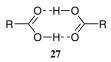
Chemical Interpretations. The noncovalent interactions of Table 14.1 constitute all of the forces that we need to consider,³ yet we are accustomed to invoke phenomena that may appear to be additional forces. But these phenomena are really just the

³ This statement is not strictly correct, because certain molecules possessing quadrupolar moments can undergo additional forces, but these can usually be neglected.

noncovalent forces (perhaps with some covalent bonding mixed in) masquerading in chemically useful forms. One of these forms is *hydrogen-bonding*. A hydrogen bond (H bond) is represented by the dots in the reaction

$$A-H+B \rightleftharpoons A-H \cdots B$$

The A-H bond in the H-bond donor AH is largely covalent; the H \cdots B bond to the Hbond acceptor B is largely a dipole-dipole interaction. Though exceptions to the following are known, H bonding is most effective when the A-H \cdots B angle is 180° and when the atoms A and B are nitrogen or oxygen. The protein α -helix and the folded conformations of proteins are largely maintained by H-bonding, as is the DNA double helix. Carboxylic acids in the vapor state and in nonpolar solvents form H-bonded dimers as in



The rather surprising pK_a values of salicylic acid ($pK_1 = 2.98$, pK_2 13.00; for 4-hydroxybenzoic acid $pK_1 = 4.58$, $pK_2 = 9.39$) are ascribed to stabilization of the monoanion by H bonding:



It is interesting that intramolecular H bonding can be effective in aqueous solution, whereas intermolecular H bonding is relatively ineffective because of the overwhelming competition from the solvent, itself a good H bond donor and acceptor. The structure of water as a medium is a result of its H-bonding properties.

Another chemical phenomenon is *charge transfer* (CT) complexing, also known as electron donor–acceptor (EDA) complexing. This seems to be a combination of covalent and noncovalent effects. Apparently an electron is partly or wholly transferred from an orbital on the donor molecule to one on the acceptor molecule, so the interaction has some covalent character and it results in definite and detectable changes in electron configuration; however, the noncovalent forces also are involved. Although some covalent electron-sharing takes place, CT complexes often are very weak.

Finally we must take notice of the solvent. We can do this by treating the solvent as a structureless continuum, as when we introduce the dielectric constant as a measure of the ability of the solvent to separate charges; or more realistically we can recognize that the solvent is itself composed of molecules, which undergo the same kinds of intermolecular interactions as do the solute species. We are then led to the view that the formation of a complex SL through the reaction of S with L in the presence of solvent (medium) M is a competition among three pairwise types of interactions: solute–solute (SL), solute–solvent (SM, LM, and SL-M), and solvent–solvent (MM). Thus we can write

$$\Delta G_{\text{net}}^0 = \Delta G_{\text{solute-solute}}^0 + \Delta G_{\text{solvation}}^0 + \Delta G_{\text{solvent-solvent}}^0$$

Each term on the right is the result of noncovalent interactions. The solute–solute interaction term is complex-stabilizing. The solvation term could be complex-stabilizing if the SL-M interaction (solvation of the complex) is stronger than the sum of the substrate–solvent plus the ligand–solvent terms, but this is unlikely, so solvation usually destabilizes the complex. The role of the solvent–solvent interaction term depends greatly on the nature of the solvent and the solute species.

Suppose, for example, that we wish to study H bonding between phenol (H-bond donor) and pyridine (H-bond acceptor). If we attempt this in water as the solvent, we are unlikely to be successful because the solvation term (arising from the phenol–water and pyridine–water solvation interaction) will be complex-destabilizing. On the other hand, if carbon tetrachloride, with no H-bonding capability of its own, is the solvent, we will be able to detect the phenol–pyridine interaction.

There is a particularly important kind of solvent effect observed with nonpolar solutes in aqueous solution. Since water is both an H-bond donor and an H-bond acceptor, the solvent-solvent interaction term ΔG^0 is appreciable and may even be overwhelming. Water molecules form an H-bonded network having definite structural features. Now, when a nonpolar solute molecule (such as a saturated hydrocarbon) dissolves in water, no H bonds from the solute to the surrounding water molecules will form, because the solute is neither a donor nor an acceptor. The water structure in the immediate vicinity of the solute re-forms to compensate for the broken water-water H bonds. The result is that the number of possible orientations of water molecules is reduced by the presence of the nonpolar solute; this means that the configurational entropy of the system is reduced. From this point of view, the very low aqueous solubility of nonpolar substances is caused by this unfavorable entropy change in the dissolution process. This is called the *hydrophobic effect*.

Now if two such dissolved nonpolar solute molecules come into contact, some of the "structured water" in their vicinity is released, thereby increasing the system entropy and resulting in a negative free-energy change. This is the driving force for association of nonpolar species in aqueous solution. This phenomenon is termed *hydrophobic interaction*. Observe that complex formation driven by this hydrophobic phenomenon owes little or nothing to solute–solute interaction (which is simply a result of the ever-present dispersion forces); rather, it is essentially completely driven by solvent–solvent interaction. In a sense the solute is "squeezed out" of the water. This water-structure interpretation of the hydrophobic interaction applies strictly only to nonpolar solutes. Many solutes, however, including most drug molecules, possess both nonpolar and polar groups (we can call such molecules "semipolar"), and although the hydrophobic interaction may play a role in their complex formation, the driving force may appear as either a favorable entropy change or a favorable enthalpy change.

An alternative description of the hydrophobic effect and hydrophobic interaction adopts a thermodynamic viewpoint. The solvent is pictured as a structureless continuum. In order to dissolve a molecule in this medium, a molecule-sized cavity must be created; then the solute molecule is inserted into the cavity, and finally solute–solvent interaction takes place. Creation of a cavity having surface area A requires an expenditure of free energy equal to $A\gamma$, where γ is the solvent surface tension. We have seen (Chapter 11) that the surface tension of water is unusually high (this itself is a result of the strong water–water interaction).

On this basis alone, all solutes might be expected to show low aqueous solubilities. However, recall the step in the dissolution process in which solute–solvent interactions may occur. In this step the energy expended in creating the cavity may be recovered, provided that the solute structure is such as to permit strong solute–solvent interactions. Semipolar solutes do lead to such interactions, but nonpolar solutes do not. Hence this thermodynamic approach accounts for the hydrophobic effect.

When two dissolved solute molecules come into contact, the total cavity surface area is reduced as the two separate cavities coalesce into a single cavity. The free energy change for this process is therefore $\gamma \Delta A$, where ΔA , a negative quantity, is the change in cavity surface area. This is the driving force for the hydrophobic interaction. Quantitative theories of this phenomenon have been developed. (See Connors and Khossravi, 1993; Mulski and Connors, 1995).

If the hydrophobic interaction is substantially responsible for the strength of a given complex in aqueous solution, we can predict that incorporation of an organic solvent into the aqueous medium will result in a weakening of the complex (observed as a decrease in the equilibrium constant for its formation). We often turn the argument around and use the complex-weakening effect of organic solvents as a diagnostic criterion for the operation of the hydrophobic interaction.

14.3. BINDING MODELS

General. We can write any complex formation equilibrium as

$$mS + nL \rightleftharpoons S_mL_n$$

and define the overall binding constant

$$\boldsymbol{\beta}_{mn} = \frac{[\mathbf{S}_m \mathbf{L}_n]}{[\mathbf{S}]^m [\mathbf{L}]^n} \tag{14.2}$$

where we assume that activities are equal to concentrations. The simultaneous collision of *m* molecules of S and *n* molecules of L to produce the complex S_mL_n is so improbable (unless m = 1 and n = 1) that it can be neglected, and so a more

realistic view of what happens is that complex S_mL_n is built up by successive bimolecular collisions. For example, the complex SL_2 forms in these successive steps:

$$S + L \rightleftharpoons SL$$

 $SL + L \rightleftharpoons SL_2$

For each such step we define a *stepwise binding constant*:

$$K_{11} = \frac{[SL]}{[S][L]}$$
(14.3)

$$K_{12} = \frac{[SL_2]}{[SL][L]}$$
(14.4)

Algebraic substitution shows that $\beta_{12} = K_{11}K_{12}$. Observe how the subscripts denote the stoichiometry of the complex.

Usually, reactant concentrations are expressed in mol L^{-1} (molarity, M); consequently, all stepwise binding constants have the unit M^{-1} . Binding constants are also known as stability constants, association constants, or formation constants. In some fields of study it is traditional to write the reaction in the opposite direction, and then the corresponding equilibrium constant is the reciprocal of the binding constant and is called a dissociation constant.

The experimental recognition of the existence of a complex requires that some property of the complex be quantitatively different from that property as possessed by the two interactants giving rise to the complex; in the following section we will see examples of such properties. Our first experimental goal is to establish the stoichiometry of the complex or complexes; our second goal is to evaluate the stepwise binding constants. The most general way to accomplish both goals is to postulate a reasonable stoichiometry, to derive a quantitative description of the expected experimental outcomes based on this stoichiometry, and then to test this model (hypothesis) with experimental data. The usual criterion is that the binding constant evaluated in such a test should be constant over a wide range of system concentrations. This somewhat abstract account will be made more explicit by considering several important stoichiometric models. More extensive treatments are available (Connors 1987; Wyman and Gill 1990).

The 1:1 Stoichiometric Model. This, the simplest of stoichiometric models, also happens to be applicable to many real systems, and its description is fundamental for understanding chemical equilibria. The complex formation reaction is

$$S + L \stackrel{K_{11}}{\Longrightarrow} SL$$

with K_{11} defined by Eq. (14.3). Let us define f_{11} as the fraction of substrate present in the complexed (bound) form, or

$$f_{11} = \frac{[\mathrm{SL}]}{S_t} \tag{14.5}$$

where S_t is the total substrate concentration:

$$S_t = [\mathbf{S}] + [\mathbf{SL}] \tag{14.6}$$

Algebraic substitution of Eqs. (14.3) and (14.5) into Eq. (14.6) gives the quantitative 1:1 stoichiometric model:

$$f_{11} = \frac{K_{11}[L]}{1 + K_{11}[L]}$$
(14.7)

This equation is the 1:1 *binding isotherm.* Its functional form is known as a rectangular hyperbola. We can easily gain a sense of how f_{11} depends on [L] by making a calculation for a hypothetical system; this has been done in Table 14.2 by assuming that $K_{11} = 50 \text{ M}^{-1}$ and using Eq. (14.7) to calculate f_{11} for assigned values of [L]. The results are plotted in Fig. 14.2. The shape of this curve is very characteristic; at low values of [L], where K_{11} [L] $\ll 1, f_{11}$ is nearly a linear function of [L], as can be seen from Eq. (14.7), whereas at high values of [L], where K_{11} [L] $\gg 1, f_{11}$ becomes nearly independent of [L]. This region in which f_{11} shows little dependence on [L] is called the *saturation effect*; the physical interpretation is that nearly all the S molecules are already bound to L, so the addition of more L has little

Table 14.2. Hypothetical 1 : 1 binding isotherm calculated with Eq. (14.7) and the value $K_{11} = 50 \text{ M}^{-1}$

[L] (M)	f_{11}
0	0
0.005	0.200
0.01	0.333
0.02	0.500
0.04	0.667
0.06	0.750
0.08	0.800
0.10	0.833
0.15	0.882
0.20	0.909
0.30	0.938
0.40	0.952

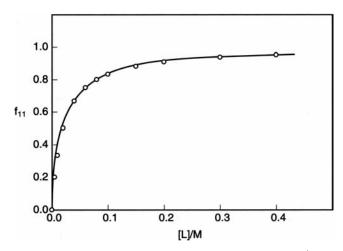


Figure 14.2. Plot of Eq. (14.7), the 1:1 binding isotherm, for $K_{11} = 50 \text{ M}^{-1}$ (see Table 14.2).

effect on increasing f_{11} , which increases asymptotically to a value of unity. Notice that $K_{11} = 1/[L]$ when $f_{11} = 0.5$.⁴

The chemical interpretation of 1:1 binding is that the substrate S possesses a single "binding site," as does the ligand L; and when the complex SL forms, no further sites are available for the binding of additional species.

A quantitative test of the 1:1 model may be made by rearranging Eq. (14.7) into a linear plotting form. Several linear plots have been used. Since f_{11} is the fraction complexed (also known as the *fraction bound*), then $1 - f_{11}$ is the fraction uncomplexed (fraction free), or $1 - f_{11} = 1/(1 + K_{11} [L])$. Thus $f_{11}/(1 - f_{11}) = K_{11}[L]$, or

$$\log\left(\frac{f_{11}}{1-f_{11}}\right) = \log\left[L\right] - \log K_{11}$$
(14.8)

A log-log plot should be linear with a slope of unity if the stoichiometry is 1:1. This is called a *Hill plot*. More simply, Eq. (14.7) can be rearranged to three nonlogarithmic linear plotting forms. Equation (14.9) is used by plotting $1/f_{11}$ against 1/[L], so it is called the *double-reciprocal plot*:

$$\frac{1}{f_{11}} = \frac{1}{K_{11}[L]} + 1 \tag{14.9}$$

When spectrophotometry is the experimental method of study, the double-reciprocal plot is called the *Benesi–Hildebrand plot*.

⁴ We have seen earlier manifestations of Eq. (14.7). The Langmuir adsorption isotherm (Chapter 11) and Eq. (12.22) have this form. The identity K = 1/[L] when $f_{11} = 0.500$ is the same as the condition $pK_a = pH$ at the midpoint of an acid–base titration.

Equation (14.10) is another plotting form of Eq. (14.7); a plot of $[L]/f_{11}$ against [L] is expected to be linear:

$$\frac{[\mathbf{L}]}{f_{11}} = [\mathbf{L}] + \frac{1}{K_{11}} \tag{14.10}$$

Finally, Eq. (14.11) is a third plotting form:

$$\frac{f_{11}}{[L]} = -K_{11}f_{11} + K_{11} \tag{14.11}$$

This plot of $f_{11}/[L]$ against f_{11} is sometimes called a *Scatchard plot*.

Linearity in these plots is a necessary condition if the 1:1 model is valid; and from the parameters of the equation, K_{11} can be evaluated. Actually we seldom measure f_{11} directly, but rather some experimental quantity that is related to f_{11} , so the interpretation of the plots depends on the particular experimental method, as will be shown in Section 14.4.

A very interesting version of the 1:1 model arises in the study of many small molecule–protein binding systems. In these systems we may consider that n small molecules, which will play the role of ligand L, may bind to a single protein molecule, which we will call the substrate S. As a measure of the extent of binding, we define the quantity

$$\bar{i} = \frac{L_t - [\mathrm{L}]}{S_t} \tag{14.12}$$

where L_t and S_t refer respectively to the total ligand and substrate concentrations; i can be interpreted as the average number of ligand molecules bound per molecule of substrate.

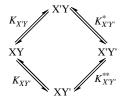
Now, if we impose the rather stringent conditions that all n binding sites on the protein are identical and independent, it is possible to derive the following isotherm:

$$\bar{i} = \frac{nk[\mathrm{L}]}{1+k[\mathrm{L}]} \tag{14.13}$$

where k is the constant for binding to a single site. According to this equation, this rather complicated system follows the hyperbolic function characteristic of simple 1:1 binding. Usually n and k are evaluated from a Scatchard plot.

The 1:1 + 1:2 Model. This stoichiometric model will serve as an example of the treatment of multiple complexes. The complex stoichiometries are SL and SL₂, and the stepwise binding constants are defined in Eqs. (14.3) and (14.4). Obviously, both of these equations must be simultaneously satisfied.

The existence of the SL_2 complex means that there must be two binding sites on S. If these two sites are chemically different, there may exist two different 1:1 complexes. All of these possibilities can be related by the scheme



where XY represents the uncomplexed substrate, and X and Y are the two different binding sites. A superscript prime indicates that the site is complexed to an L molecule, so X'Y and XY' are the two possible isomeric 1:1 complexes and X'Y' is the 1:2 complex. The equilibrium constants shown are microscopic binding constants.

We experimentally measure K_{11} and K_{12} . These are related (Connors 1987) to the microscopic constants by

$$K_{11} = K_{X'Y} + K_{XY'} \tag{14.14}$$

$$K_{11}K_{12} = a_{XY}K_{X'Y}K_{XY'} \tag{14.15}$$

where $a_{XY} = K_{X'Y'}^*/K_{XY'} = K_{X'Y'}^{**}/K_{X'Y'}$. Since these two equations constitute two independent equations with three unknowns, we are in general unable to evaluate the microscopic constants. Two special cases are of interest, however. First, suppose that the two binding sites are identical. This means that $K_{X'Y} = K_{XY}$, so $K_{11} = 2K_{X'Y}$ and Eq. (14.15) becomes $K_{11} = 4K_{12}/a_{XY}$. The second special case occurs if the two sites are identical and are also independent. This last condition means that $a_{XY} = 1$, and we then find that $K_{11} = 4K_{12}$.

Perhaps this treatment of the 1:1 + 1:2 model has seemed somewhat familiar. In fact, it is merely a rephrasing of the arguments used in Section 12.7 when discussing the assignment of pK_a values of diprotic acids. The only difference in the treatments is that for acids we defined K_a values as dissociation constants whereas for complex formation our equilibrium constants are association constants. In the acid–base system the proton plays the role of the ligand.

14.4. MEASUREMENT OF BINDING CONSTANTS

In this section we describe a few experimental techniques for the determination of complex binding constants; many more techniques are available (Connors 1987). It is obviously essential that the temperature be controlled and that the system be at equilibrium; this latter condition is easy to achieve because most noncovalent interactions take place very rapidly.

Spectrophotometry. A basic knowledge of absorption spectroscopy is assumed. We will treat the 1:1 stoichiometric model, assuming that Beer's law is obeyed by all species. The method requires that the free substrate S and the complex SL possess significantly different absorption spectra. A wavelength is chosen at which ε_S and ε_{11} are different. (Subscripts refer to the species; 11 indicates the complex SL.) S_t and L_t represent total substrate and ligand concentrations.

In the absence of ligand we can write

$$A_0 = \varepsilon_S b S_t \tag{14.16}$$

In the presence of ligand at fixed S_t value, the solution absorbance is given by

$$A_{\rm L} = \varepsilon_{\rm S} b[{\rm S}] + \varepsilon_{\rm L} b[{\rm L}] + \varepsilon_{11} b[{\rm SL}]$$

which is combined with the mass balances $L_t = [L] + [SL]$ and $S_t = [S] + [SL]$ to give

$$A_{\rm L} = \varepsilon_S b S_t + \varepsilon_{\rm L} b L_t + \Delta \varepsilon_{11} b [\rm{SL}]$$

where $\Delta \varepsilon_{11} = \varepsilon_{11} - \varepsilon_S - \varepsilon_L$. If the solution absorbance is measured against a reference containing L_t mol L⁻¹ of ligand, the absorbance actually measured is

$$A = \varepsilon_S b S_t + \Delta \varepsilon_{11} b [SL] \tag{14.17}$$

Now subtract Eq. (14.16) from Eq. (14.17) and incorporate the definition of K_{11} [Eq. (14.3)], obtaining

$$\Delta A = K_{11} \Delta \varepsilon_{11} b[\mathbf{S}][\mathbf{L}] \tag{14.18}$$

where $\Delta A = A - A_0$. From the mass balance $S_t = [S] + [SL]$ and the expression for K_{11} we find $S_t = [S](1 + K_{11}[L])$, which is used in Eq. (14.18):

$$\frac{\Delta A}{b} = \frac{S_t K_{11} \Delta \varepsilon_{11} [\mathbf{L}]}{1 + K_{11} [\mathbf{L}]} \tag{14.19}$$

This equation is the 1:1 binding isotherm expressed for spectrophotometric observation. Note that the functional dependence of the dependent variable $\Delta A/b$ on [L] is identical with that in Eq. (14.7).

There is a subtlety in the application of Eq. (14.19), because in this equation [L] is the free (uncomplexed) concentration of ligand, which we do not (yet) know. What we know is L_t , the total ligand concentration, because we prepare the solutions with a known constant value of S_t and varying but known values of L_t ; then we measure $\Delta A/b$ in these solutions. The answer to this problem is to start with the mass balance

 $L_t = [L] + [SL]$, incorporating the expression for K_{11} to yield

$$L_t = [L] + \frac{S_t K_{11}[L]}{1 + K_{11}[L]}$$
(14.20)

Equations (14.19) and (14.20) together provide a complete description of the system. Often it is possible to design the experiment such that $L_t \gg S_t$; then Eq. (14.20) shows that $L_t \approx [L]$, and Eq. (14.19) can be analyzed with this approximation. More generally an iterative method is required; with a preliminary estimate of K_{11} , Eq. (14.20) is solved to give the [L] value corresponding to each L_t . These [L] values are used in Eq. (14.19) to obtain an improved estimate of K_{11} , and this process is repeated until the K_{11} estimates reach a constant value. The solution of Eq. (14.19) can be carried out graphically as described earlier; for example, the double-reciprocal form (the Benesi–Hildebrand plot) is

$$\frac{b}{\Delta A} = \frac{1}{S_t K_{11} \Delta \varepsilon_{11}[L]} + \frac{1}{S_t \Delta \varepsilon_{11}}$$
(14.21)

According to this equation, a plot of $b/\Delta A$ against 1/[L] will be linear if the stoichiometry is 1:1. The value of K_{11} is found from

$$K_{11} = \frac{\text{intercept on vertical axis}}{\text{slope}}$$
(14.22)

The parameter $\Delta \varepsilon_{11}$ may be evaluated from the intercept value.

Table 14.3 lists experimental data from a study of the complex between cinnamic acid anion (the substrate S) and theophylline (the ligand L) (Kramer and Connors 1969). Figure 14.3 is the plot of these data according to Eq. (14.21). From this plot the value $K_{11} = 10.5 \text{ M}^{-1}$ was evaluated.

This absorption spectrophotometric method can be applied throughout the ultraviolet, visible, and infrared regions of the spectrum. Moreover, the same approach is applicable to fluorescence data and to nuclear magnetic resonance chemical shifts.

$\overline{10^2 L_t (\mathrm{M})}$	A ^a
0.00	0.530
1.11	0.791
1.25	0.826
1.43	0.858
1.67	0.906
2.00	0.965
2.50	1.053

Table 14.3. Complexing of sodium cinnamate and the ophylline at pH 6.5 and 25 $^\circ\text{C}$

^{*a*}Pathlength b = 1 cm; wavelength = 315 nm; S_t = 0.001 M.

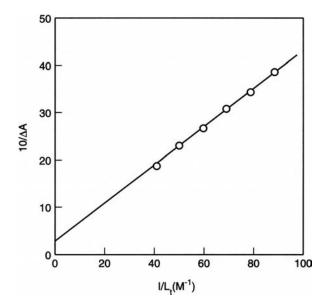


Figure 14.3. Plot of spectral data for the sodium cinnamate (S)–theophylline (L) system; pH 6.5; at 25 °C; $K_{11} = 10.5 \text{ M}^{-1}$. Values of ordinates are an arithmetic average of five determinations. [Reproduced by permission from Kramer and Connors (1969).]

Solubility. When a complex is formed, the total (apparent) solubility of an interactant is increased, and this change in apparent solubility can be analyzed to derive the binding constant. The experiment consists of the addition of an identical weight (in considerable excess of its intrinsic solubility) of substrate S to each of numerous vials. Increasing amounts of soluble ligand L are added to successive vials, and solvent is added to make an identical volume of solution in each vial. Now the vials are agitated at constant temperature until solubility equilibrium is achieved (typically about 24 h). Then the solution phase of each vial is analyzed for its total dissolved concentration of substrate, S_t . A solubility phase diagram is constructed by plotting S_t (vertical axis) against L_t .

We will carry out the interpretation of the phase diagram for the case of 1:1 stoichiometry. The mass balance expressions are

$$S_t = [S] + [SL]$$
$$L_t = [L] + [SL]$$

as we have often written. The manner in which the experiment was carried out ensured that pure solid S remained in each vial at equilibrium, which means that the chemical potential, and therefore the activity, of uncomplexed substrate S was the same in each vial; we can therefore reasonably conclude that the concentration of free S, namely [S], was the same in each vial, including the vial containing no ligand

$\overline{10^2 L_t (\mathrm{M})}$	$10^2 S_t (\mathrm{M})$
0	3.87
2.52	5.17
5.04	6.22
10.09	8.60
12.61	9.46
15.14	10.94
20.18	12.88

 Table 14.4.
 Solubility data for the theophylline

 (S)-sodium salicylate (L) system^a

^{*a*}At 25 °C in pH 6.5 phosphate buffer.

at all. This quantity is therefore the intrinsic solubility s_0 of substrate. The mass balance on substrate can therefore be written

$$S_t = s_0 + [SL] \tag{14.23}$$

Combining Eq. (14.23) with the mass balance on ligand and the definition of K_{11} gives

$$S_t = s_0 + \frac{K_{11}s_0L_t}{1 + K_{11}s_0} \tag{14.24}$$

as the solubility binding isotherm. Equation (14.24) predicts that S_t will be a linear function of L_t ; the intercept will be s_0 , and the binding constant can be calculated from

$$K_{11} = \frac{\text{slope}}{S_0(1-\text{slope})} \tag{14.25}$$

Table 14.4 gives solubility data for a study of the complexing between theophylline (the substrate) and sodium salicylate (the ligand) (Cohen and Connors 1967). Figure 14.4 is the plot according to Eq. (14.25). From this phase diagram the value $K_{11} = 21.5 \text{ M}^{-1}$ was evaluated.⁵

The solubility method can be extended to the study of multiple complexes.

⁵ Earlier 1:1 binding isotherms [Eqs. (14.7) and (14.19)] have been hyperbolic functions of [L], whereas Eq. (14.25) is a linear function of L_t . This distinction arises because of the unusual feature of the solubility experiment in which [S] is maintained constant while S_t varies; in the other methods, S_t is held constant and [S] varies.

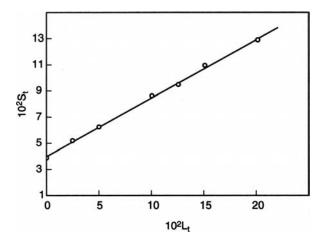


Figure 14.4. Phase solubility diagram for the theophylline (S)–sodium salicylate (L) system at $25 \degree C$. [Reproduced by permission from Cohen and Connors (1967).]

Potentiometry. We saw in Chapter 13 that it may be possible to design an electrochemical cell such that an ion activity is related to cell potential by the Nernst equation:

$$E = \text{constant} \pm \frac{RT}{nF} \ln a$$

If the activity of this ion changes in the process of complex formation, then potentiometry offers a means for studying the complex. This is what we do when we measure pK_a values of weak acids. The principle can be extended to study the formation of complexes between metal ions and bases. These complexes, which can be exceedingly strong, are the result of covalent bonding and lie somewhat outside our present concern, but they can be studied in the same way (Connors 2000).

We will develop the potentiometric method for the formation of complexes between a weak acid–base conjugate pair (the substrate) and a neutral ligand. The hydrogen ion activity changes as the ligand concentration is altered, so that the ion we monitor, H^+ , is a measure of the extent of complex formation but is not itself the substrate or ligand. In general, both the acid form of the substrate, HA, and its conjugate base A^- , can complex with the ligand L, so we write

$$HA + L \stackrel{K_{11a}}{\longrightarrow} HAL$$
$$A^{-} + L \stackrel{K_{11b}}{\longrightarrow} AL^{-}$$

We also have the usual acid-base equilibrium

$$\mathrm{HA} \stackrel{K_a}{=} \mathrm{H}^+ + \mathrm{A}^-$$

and there also exists this acid-base equilibrium:

$$HAL \stackrel{K_{a11}}{\longleftarrow} H^+ + AL^-$$

Defining and manipulating these four equilibrium constants gives

$$\frac{K_a}{K_{11}} = \frac{K_{11a}}{K_{11b}} \tag{14.26}$$

Thus only three of these four quantities are independent; that is, we need measure and discuss only three of them to give a full description of this 1:1 stoichiometric system. We will choose K_a , K_{11a} , and K_{11b} .

The experiment consists of measuring the apparent pK'_a of HA as a function of added ligand. We start the analysis by defining the apparent acid dissociation constant by

$$K'_{a} = \frac{[\text{HA}]([\text{A}^{-}] + [\text{AL}^{-}])}{[\text{HA}] + [\text{HAL}]}$$
(14.27)

Then the expressions for K_a , K_{11a} , and K_{11b} are combined with Eq. (14.27), yielding

$$K'_{a} = K_{a} \frac{(1 + K_{11b}[L])}{(1 + K_{11a}[L])}$$
(14.28)

Defining $\Delta pK'_a = pK'_a - pK_a$ allows us to write Eq. (14.28) as

$$\Delta p K'_{a} = \log \frac{(1 + K_{11a}[L])}{(1 + K_{11b}[L])}$$
(14.29)

Further defining the quantity C by

$$C = \frac{1 + K_{11a}[L]}{1 + K_{11b}[L]}$$
(14.30)

we have $\Delta pK'_a = \log C$. Equation (14.30) is the 1:1 binding isotherm. We measure $\Delta pK'_a$ as a function of L_t , where pK_a is the value of $\Delta pK'_a$ when $L_t = 0$. Then C is found from $\Delta pK'_a$. A separate mass balance relates [L] to L_t . Equation (14.30) is a rectangular hyperbola, and the usual three linear plotting forms can be used to extract values of K_{11a} and K_{11b} . Equation (14.31) shows the form corresponding to Eq. (14.11), the Scatchard plot:

$$\frac{C-1}{[L]} = K_{11a} - K_{11b}C \tag{14.31}$$

	$\Delta \mathrm{p} K'{}_a$		
L_{t} (M)	Benzoic Acid	4-Cyanophenol	
0.0017	0.24		
0.0020		-0.16	
0.0030		-0.21	
0.0045		0.28	
0.0057	0.61	_	
0.0075		-0.38	
0.01	0.84	-0.42	
0.02	1.10	-0.52	
0.03	1.25	-0.56	
0.04	1.34	-0.58	
0.05		-0.58	
0.06	1.45	-0.59	
0.07	1.48	-0.58	
0.08	1.51	-0.59	
0.09	1.54	-0.59	
0.10	1.55	-0.58	

Table 14.5. $\Delta p K'_a$ values at 25 °C for the complexing of benzoic acid and of 4-cyanophenol with α -cyclodextrin

Table 14.5 gives $\Delta pK'_a$ values for two complexing systems; in both of these α -cyclodextrin, a macrocyclic natural product formed of six residues of glucose is the ligand. The substrates are benzoic acid and 4-cyanophenol (Connors et al. 1982; Lin and Connors 1983). The first interesting feature of these data is the very substantial values of $\Delta pK'_a$ that may occur. The second point is that $\Delta pK'_a$ may be either positive or negative. [If $\Delta pK'_a$ is negative, it can be redefined as $\Delta pK'_a = pK_a - pK'_a$ in order to generate a positive number; alternatively, define C' = 1/C and use C' instead of C in Eqs. (14.30) and (14.31).] Referring to Eq. (14.30) shows that a positive $\Delta pK'_a$, which signifies that C > 1, means that $K_{11a} > K_{11b}$; the conjugate acid binds to the ligand more strongly than does the conjugate base. Similarly, a negative $\Delta pK'_a$ means that $K_{11b} > K_{11a}$. If $K_{11a} = K_{11b}$, then $\Delta pK'_a = pK_a$ so $\Delta pK'_a = 0$, and the method is inapplicable; this circumstance is rare, however, because the conjugate acid and base differ electronically very profoundly, so their noncovalent interaction possibilities differ as a consequence.

From plots of Eq. (14.31), the values $K_{11a} = 722 \text{ M}^{-1}$ and $K_{11b} = 11.2 \text{ M}^{-1}$ were found for the benzoic acid- α -cyclodextrin complexes. The 4-cyanophenol complexes gave $K_{11a} = 158 \text{ M}^{-1}$ and $K_{11b} = 662 \text{ M}^{-1}$.

Dialysis. This interesting method is applicable to the study of the binding of small molecules to macromolecules, and it has been widely applied to drug–protein binding. The experimental method requires that two solution compartments be separated by a semipermeable membrane, that is, a membrane through which the small molecule can freely pass but the macromolecule cannot. Let us call the macromolecule

the substrate S and call the small molecule the ligand L. We label the compartments 1 and 2.

The substrate, at known total concentration S_t , is placed in compartment 1, where it is constrained to remain because of its size. The small ligand is placed in compartment 2. The system is allowed to come to equilibrium at constant temperature.

At equilibrium the chemical potentials of free L, the diffusible ligand, in the two compartments are equal. We may therefore (usually) infer that the concentrations of L are equal. Using subscripts to identify the compartments, we write this equality as

$$[L]_1 = [L]_2 \tag{14.32}$$

The solutions in the two compartments are analyzed to determine their total ligand concentrations, $(L_t)_1$ and $(L_t)_2$. For compartment 1 we can now state

$$(L_t)_1 = [L]_1 + [bound L]_1$$
(14.33)

and for compartment 2 we have

$$(\mathbf{L}_t)_2 = [\mathbf{L}]_2 \tag{14.34}$$

We therefore have all the information required to calculate the quantity i [Eq. (14.12)], rewritten here explicitly for compartment 1:

$$\bar{i} = \frac{(L_t)_1 - [L]_1}{(S_t)_1} \tag{14.35}$$

This experiment is repeated over a range of ligand concentrations, and then binding isotherm (14.13) is applied to the data to determine the parameters *n* and *k*.

Strengths of Complexes. We might express the strength of a complex in terms of either its binding constant or (what is equivalent) its standard free energy change ΔG^0 , or as its standard enthalpy change ΔH^0 . These may parallel each other for comparable systems, but they may diverge markedly because of the thermodynamic relationship $\Delta G^0 = \Delta H^0 - T \Delta S^0$. Let us make some simple calculations to develop a sense of the magnitudes to be expected.

We recall from an earlier discussion that the net free-energy change may be divided into contributions from solute–solute (i.e., substrate–ligand) interactions, from solute–solvent (solvation) interactions, and from solvent–solvent interactions, or

$$\Delta G^0 = \Delta G^0_{\rm SL} + \Delta G^0_{\rm solvation} + \Delta G^0_{\rm MM} \tag{14.36}$$

Let us first consider an H-bond complex formation reaction in vacuo or in a very nonpolar solvent, so that essentially only the solute–solute interaction makes a contribution; thus $\Delta G^0 = \Delta G_{SL}^0$. Suppose that ΔH^0 for the reaction is -3 kcal mol⁻¹,

a typical value for a noncovalent interaction. We also need an estimate for ΔS^0 , which we will obtain with the following (oversimplified) argument. In the process of the two species S and L interacting to form the complex C, some modes of molecular motion are lost. Even supposing that all internal modes (vibrational and rotational) of S and L are preserved in C, three translational modes will be lost. Applying the statistical mechanical definition of entropy [Chapter 2, Eqs. (2.1) and (2.2)] to the reaction $S + L \rightleftharpoons C$, we obtain $\Delta S^0 = -R \ln 3 = -9.1 \text{ J K}^{-1} \mod^{-1}$ or $-2.2 \arctan K^{-1} \mod^{-1}$. For our hypothetical system we therefore calculate $\Delta G^0 = -3000 + (298)$ (2.2) = $-2350 \arctan^{-1}$. With the basic equation $\Delta G^0 = -RT \ln K_{11}$ we calculate $K_{11} = 53 \text{ M}^{-1}$ at 25 °C.

Now let us consider the quite different system of two nonpolar molecules in aqueous solution. The solute–solvent and solute–solute interactions will be negligible or will largely offset each other, so in this case we can write $\Delta G^0 = \Delta G_{MM}^0$. To estimate ΔG^0 , we draw on the cavity theory of the hydrophobic interaction, according to which the change in free energy is equal to the product $\gamma \Delta A$, where γ is the solvent surface tension and ΔA is the change in nonpolar surface area in contact with the solvent. Suppose $\Delta A = -50 \text{ A}^2$; this is about the cross-sectional area of an aromatic ring. For water $\gamma = -71.8 \text{ erg cm}^{-2}$. Thus

$$\Delta G^{0} = \left(71.8 \frac{\text{erg}}{\text{cm}^{2}}\right) \left(\frac{1J}{10^{7} \text{erg}}\right) - \left(50 \text{ Å}^{2}\right) \left(\frac{10^{-8} \text{cm}}{1 \text{ Å}}\right) = -3.6 \times 10^{-20} \text{J}$$

This is the free-energy change per complex; we multiply by N_A to put it on a molar basis, finding $\Delta G^0 = -21.7 \text{ kJ mol}^{-1}$ or $-5.2 \text{ kcal mol}^{-1}$. Converting to K_{11} , gives $K_{11} = 6.5 \times 10^3 \text{ M}^{-1}$.

These calculations illustrate the magnitudes of binding constants that may be expected, although the range is even larger than indicated by these numbers. Values of K_{11} smaller than 1 M⁻¹ have been reported, although such results should be interpreted with caution. Very large values are not unusual. Consider the p K_a of phenol, which is 10.00 at 25 °C. K_a is a dissociation constant. Its reciprocal, $I/K_a = K_{11}$, is a binding constant; this is obviously $1 \times 10^{10} \text{ M}^{-1}$.

As was pointed out for acids and bases in Table 12.8, values of equilibrium constants for closely related reactants tend to fall into characteristic ranges. This behavior provides a coarse level of predictability. There is some evidence that for each identifiable "population" or class of complex equilibria the log K_{11} values are normally distributed (Connors 1995; Burnette and Connors 2000). Figure 14.5 is a plot of log K_{11} for the complexation of 663 substrates with the ligand α -cyclodextrin. The mean value of log K_{11} is 2.11, and its standard deviation is 0.90. With these parameters the normal distribution was plotted as the smooth curve in Figure 14.5. [The equation of this curve is given by Eq. (0.102) and the properties of the normal distribution are discussed in Section 0.6].

This kind of systematic behavior is seen also with other classes of noncovalent interactions, and Table 14.6 collects some examples. Observe that the mean value covers a range of more than seven orders of magnitude in K_{11} while the standard

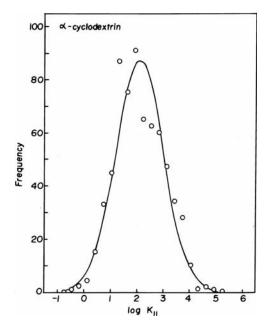


Figure 14.5. Frequency distribution (points) and normal distribution (curve) calculated with the parameters n = 663, $\mu = 2.11$, and $\sigma = 0.90$ for α -cyclodextrin complex stabilities with both uncharged and ionic substrates.

deviations differ by no more than a factor of two. On this basis, the following postulate has been suggested (Connors, 1997):

Any chemically well-defined class of noncovalent association equilibria will be normally distributed in the variable log K_{11} , the standard deviation of all such populations being about one log K_{11} unit.

Table 14.6. Statistical parameters for noncovalent binding equilibria in aqueous solution at 25 $^\circ\text{C}$

Equilibrium (Reactants Shown)	n	\bar{x}^{a}	s ^a
$\overline{\alpha$ -CyD + substrate ^b }	663	2.11	0.90
β -CyD + substrate ^b	721	2.69	0.89
γ -CyD + substrate ^b	166	2.55	0.93
$R-COO^- + H^+$	458	4.07	0.67
c X-COO ⁻ + H ⁺	161	2.32	0.62
$^{-}$ OOC-COO $^{-}$ + H $^{+}$	174	5.55	1.27
$ArO^{-} + H^{+}$	116	9.42	1.21

 ${}^{a}\bar{x}$ and s in units of log K_{11} .

 ${}^{b}CyD = cyclodextrin.$

^cX represents strongly electron-withdrawing group(s).

14.5. APPLICATIONS

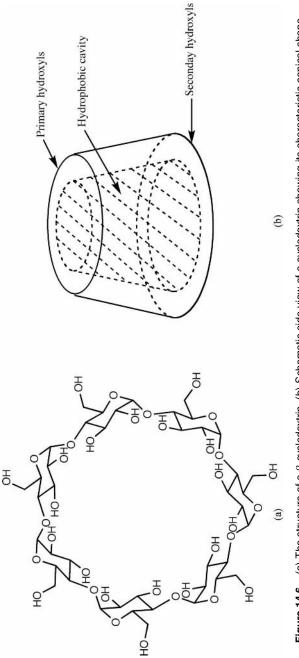
General. The interaction between two or more molecules through noncovalent interactions is referred to as molecular recognition. Molecular recognition is at the basis of the inner working of biological molecules, including proteins and nucleic acids. Antigene–antibody, DNA transcription, RNA translation, and receptor–ligand binding are all examples of biological molecular recognition. Many drugs exert their action by simply interfering with noncovalent interactions between specific molecules in the cell, so the field of molecular recognition is of special importance to pharmaceutical systems. The formation of complexes between molecules of various size has been extensively exploited for the purpose of drug delivery. A particularly significant example is given by the use of cyclodextrins.

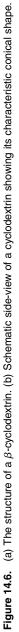
Cyclodextrins. Cyclodextrins (Connors 1997, Challa 2005). are cyclic oligomers of α -D-glucose having a truncated-cone shape (Fig. 14.6). Three kinds of cyclodextrins are readily available: α -, β -, and γ -cyclodextrins, composed of six, seven, and eight units of α -D-glucose, respectively. The utility of cyclodextrins stems from their lipophilic inner cavities, which can complex hydrophobic molecules, and their hydrophilic external surface, which imparts considerable water-solubility to these molecules and to their complexes. (The strength of cyclodextrin complexes were discussed in Section 14.4.) Thus, if a hydrophobic drug can be complexed inside a cyclodextrin, it can be readily made water-soluble, while at the same being protected from the aqueous environment. Release of the drug is achieved by equilibrium dissociation. It should be noted that the inner cavity of a cyclodextrin is quite small and can accommodate only small molecules such as phenyl derivatives. Nevertheless, much larger molecules can be solubilized in water by using cyclodextrins. It is often sufficient for only one part of a hydrophobic molecule to be complexed inside a cyclodextrin to dramatically increase the water solubility of that molecule. Figure 14.7 shows the binding mode of a cytoprotective agent with various β-cyclodextrins at different pHs (Nagase 2001). Note how most of the drug molecule is outside of the cyclodextrin hydrophobic cavity.

Complexation of a drug with a cyclodextrin may bring the following advantages: (a) improved water-solubility and dissolution, (b) enhanced bioavailability, (c) reduced drug toxicity, and (d) improved drug stability. Cyclodextrins have been used in oral, parenteral, ocular, nasal, mucosal, dermal and transdermal, and brain targetting drug delivery. Hundreds of drugs have been studied as complexes with cyclodextrins. These include sulfomethiazole, naproxen, ketoprofen, fluasterone, doxorubibin, teststerone, danazol, tolbutamide, cyclosporine, and dexamethasone.

A useful property of cyclodextrins is their relative resistance to hydrolysis. This ability makes cyclodextrins especially useful for targeting the delivery of drugs to the colon. In addition, cyclodextrins have been used for the delivery of peptides, proteins, genes, and oligonucleotides.

RNA Binding and Recognition. The use of cyclodextrins in drug delivery shows very clearly the importance of noncovalent complexes in pharmaceutical research.





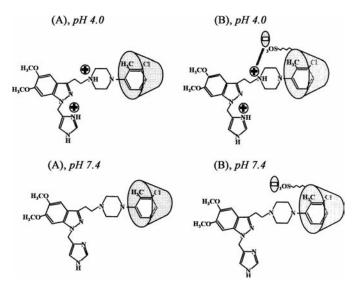


Figure 14.7. Proposed modes for the inclusion complexes of DY-9760e with HP- β -Cyclodextrin (A) and SBE- β -Cyclodextrin (B) at pH 4.0 (upper) and 7.4 (lower). [Reproduced with permission from Nagase et al. (2001).]

A second large field of investigation in molecular recognition involves the study of the features that allow certain molecules to recognize only specific compounds. The careful appreciation of how a network of intermolecular interactions leads to highly selective binding and recognition is the key for designing novel noncovalent complexes and for tuning their binding affinity. Among all molecules that have been studied in the molecular recognition field, biopolymers hold a special place. Proteins and nucleic acids are among the molecules that allow life as we know it. While a large body of research exists on proteins, the binding ability of nucleic acids for small molecules—and, more specifically, the ability of oligoribonucleotides to be able to recognize and bind small molecules with high affinity-has recently emerged as a crucially important field. The discovery of small-molecule binding RNAs such as those found in riboswitches and various forms of MicroRNAs has led to work on understanding the binding ability of these biopolymers. RNAs can fold in complex three-dimensional structures resembling those of proteins and enzymes. Some RNAs have been found to be able to bind small molecules with high affinity, and others can catalyze certain reactions. The former are called RNA aptamers, whereas the latter are referred to as ribozymes. The ability of RNAs to selectively bind small molecules and to catalyze reactions hints at the possibility that RNAs possess many different functions inside a cell, besides being involved in transcription and translation processes. An important aspect of RNA recognition studies is the minimum nucleotide sequence able to recognize and bind selectively a small molecule. A combination of computational and experimental studies has allowed the identification of the miminum sequence requirements for the binding of a number of small molecules (Anderson and Mecozzi 2005a, 2005b, 2006, 2007). For instance, starting from the

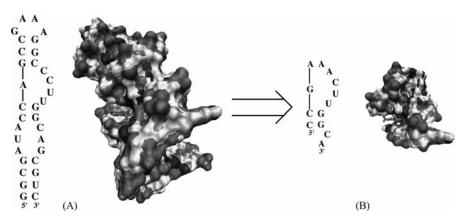
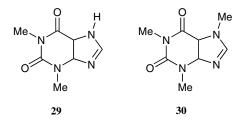


Figure 14.8. Using computational methodologies based on molecular mechanics and molecular dynamics simulations, an RNA sequence binding the drug theophylline with high selectivity and affinity has been reduced from 33 nucleotides (A) to only 13 nucleotides (B).

crystal structure of the anti-asthma drug theophylline **29** and its RNA aptamer, it has been possible to reduce RNA size from 33 to just 13 nucleotides while retaining binding ability (Fig. 14.8) (Anderson 2005a).



Moreover, while the identified 13-mer RNA is the smallest RNA ever shown to have a function (in this case the binding of theophylline), it should also be noted that this RNA was even able to selectively bind theophylline in presence of caffeine **30**, a closely related structural analog of theophylline. The selectivity achieved by this small RNA indicates how recognition and binding affinity of biopolymers are related not only to their complexity but also to their preorganized structure and to the possibility of forming a rather large number of noncovalent interactions. Similar studies have allowed the identification of the RNA minimum sequence requirements for the binding of the antibiotic paromomycin to the ribosome decoding site A (Anderson 2007) and of flavin mononucleotide (Anderson 2005b).

PROBLEMS

14.1. Define overall and stepwise binding constants for the formation of complex S_2L_3 , and show how these constants are related.

- **14.2.** Construct plots according to Eqs. (14.9)–(14.11) of the simulated data in Table 14.2.
- **14.3.** Equation (14.19) describes the binding curve for the spectrophotometric study of 1:1 complex formation. What is the value of the initial slope (i.e., the slope when [L] = 0) of the plot of $\Delta A/b$ versus [L]?
- **14.4.** (a) From Eq. (14.19) derive the Scatchard linear plotting form (corresponding to Eq. (14.11)).
 - (**b**) Plot the data in Table 14.3 to evaluate K_{11} and $\Delta \varepsilon_{11}$. Assume $L_t = [L]$.
- **14.5.** What is the value of the initial slope of a plot of *C* against [L] according to Eq. (14.30)?
- **14.6.** The anti-asthma drug theophylline is often sold as a complex with salicylic acid. In solution the compounds form a 1:1 complex. Calculate the concentration of free theophylline assuming that the binding constant is 21.5 M^{-1} and the fraction of complexed salicylic acid (substrate) is 0.99.
- 14.7. Tests on a newly isolated protein show that warfarin is a major binder to the protein. A Scatchard analysis of the binding between the protein and warfarin shows that the binding constant K_{11} is equal to 10^5 L mol^{-1} and that the number *n* of molecules of warfarin bound on average to one molecule of protein is equal to 6. Knowing that the equation used for the Scatchard plot is

$$\frac{f_{11}}{[L]} = nK_{11} - f_{11}K_{11}$$

Calculate:

- (a) The equation defining the binding isotherm.
- (b) The fraction of uncomplexed protein when the concentration of the free ligand [L] is 5×10^{-7} M.
- (c) The concentration of free ligand that corresponds to a fraction of complexed protein equal to 0.5.
- **14.8.** In Table 14.6, means and standard deviations are given in units of $\log K_{11}$ for populations (classes) of cyclodextrin complex formation and weak acid equilibria in aqueous solution. How is the p K_a of a weak acid related to $\log K_{11}$?

APPENDIXES

$_{\text{APPENDIX}}A$

PHYSICAL CONSTANTS

Quantity	Symbol	Value
Avogadro's number	N _A	$6.022 \times 10^{23} \mathrm{mo1^{-1}}$
Gas constant	R	$8.314 \mathrm{J}\mathrm{K}^{-1}\mathrm{mo1}^{-1}$
		$1.987 \mathrm{cal} \mathrm{K}^{-1} \mathrm{mol}^{-1}$
		$0.08206 \mathrm{L}\mathrm{atm}\mathrm{K}^{-1}$
Boltzmann's constant	k	$1.381 \times 10^{-23}\mathrm{JK^{-1}}$
Faraday	F	$9.6485 \times 10^4 \mathrm{C mol^{-1}}$
Elementary charge	е	$1.602 \times 10^{-19} \mathrm{C}$
Planck's constant	h	$6.626 imes 10^{-34} \mathrm{Js}$
Speed of light in vacuum	с	$2.9979 imes 10^8 { m m s^{-1}}$

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APPENDIX B

KINETIC THEORY OF GASES

The kinetic theory of gases is a classical approach to explaining the macroscopic properties of gases such as temperature, pressure, and volume and the relationship among these properties. This is done by considering the incessant motion of the particles composing the gas and their collisions with the walls of the container holding them. The theory is based on these assumptions:

- 1. A gas consists of separate particles, either molecules or atoms, characterized by a certain mass per particle.
- 2. The gas particles can be considered as points; that is, their volume is negligible compared with the volume of the container holding them.
- 3. Collisions between gas particles and the walls of the container in which they are held are perfectly elastic.
- 4. The gas particles do not exert any attractive or repulsive interactions on each other.
- 5. The average kinetic energy of the particles depends only on the gas temperature.

Let's consider N gas particles, each of mass m, held inside a cubic box of side b. Let c be the average molecular velocity. This velocity can be resolved along the three x, y, and z axes to give the c_x , c_y , and c_z components of the velocity c. Consider a particle moving along the x axis. Upon collision with the box wall perpendicular to the x axis, the particle will reverse its motion while the velocity component c_x will remain the same. Thus, the change in momentum of the particle can be written as

$$\Delta(\mathrm{mc}_x) = \mathrm{final\ momentum} - \mathrm{initial\ momentum} = -\mathrm{mc}_x - \mathrm{mc}_x = -2\mathrm{mc}_x \quad (\mathrm{B.1})$$

The pressure exerted by the gas will be given by the average force the particles exert on the wall per unit area. This average force will be given by the change in momentum p per unit time. The particle will, on average, hit the wall perpendicular to its motion c/2b

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times per second. Then, the force acting on the wall will be, per particle,

$$F_x = \frac{\Delta p}{\Delta t} = \frac{2mc_x}{\frac{2b}{c_x}} = \frac{mc_x^2}{b}$$
(B.2)

The total force acting on the wall in the x direction will be given by considering all the N molecules:

$$F = \frac{m \sum_{i=1}^{N} c_{ix}^2}{b}$$
(B.3)

The total force acting on all six walls of the cubic container will be simply

$$F_{\text{tot}} = 2 \frac{m \sum_{i=1}^{N} c_i^2}{b}$$
(B.4)

where the factor 2 considers both walls in a certain direction. The total force acting on just one of the six walls can be obtained by dividing Eq. (B.4) by 6:

$$F = \frac{m\sum_{i=1}^{N} c_i^2}{3b}$$
(B.5)

The quantity $\sum_{i=1}^{N} c_i^2$ can be written as $N\overline{c^2}$. The average $\sqrt{c^2}$ is called the rootmean-square velocity of the particles, and it is often symbolized μ . The pressure *P* can be obtained by noting that the area *A* of the wall on which the force *F* is exerted is given by b^2 . Then

$$P = \frac{F}{A} = \frac{Nm\overline{c^2}}{3b^3} = \frac{Nm\overline{c^2}}{3V}$$
(B.6)

which can be rewritten as

$$P = \frac{1}{3} \frac{M}{V} \overline{c^2} = \frac{1}{3} \delta \overline{c^2}$$
(B.7)

where M = Nm is the total mass of the gas and δ is its density. We can then calculate the root-mean-square velocity μ from the density of the gas:

$$\mu = \sqrt{\overline{c^2}} = \sqrt{\frac{3PV}{Nm}} = \sqrt{\frac{3PV}{M}} = \sqrt{\frac{3P}{\delta}}$$
(B.8)

The ideal gas law can easily be derived from Eq. B.6. The kinetic energy associated with just one gas molecule will be given by $\frac{1}{2}mc^2$. This kinetic energy is linearly

proportional to the temperature through a constant α : $\frac{1}{2}mc^2 = \alpha T$. In addition, the number *N* of molecules will be given by the product of the Avogadro number N_A and the number of moles of gas *n*. This leads us to

$$PV = \frac{Nm\overline{c^2}}{3} = \frac{2}{3}\alpha N_A nT = nRT$$
(B.9)

where we have indicated the constant $\frac{2}{3}\alpha N_A$ with R.

APPENDIX C EXTRATHERMODYNAMIC RELATIONSHIPS

Several passages in this book have mentioned interpretations of relationships that do not constitute part of classical equilibrium thermodynamics but share some of its concepts and methods. These *extrathermodynamic* ideas make use of atomic and molecular models such as modes of intramolecular transmission of electronic effects or steric effects; and these ideas, while having great value in guiding interpretations of chemical and physical phenomena, are in some degree conjectural and subject to modification. Here we describe some features of this extrathermodynamic approach in order to demonstrate the attitude that is adopted. The great simplifying generalization of (mainly) early twentieth-century chemical research was that *like structural changes produce like chemical effects*. Extrathermodynamic methods lend quantitative precision to this observation.

Linear Free Energy Relationships. We define as a *reaction series* X a set of compounds having the same reaction site X but containing varying substituents elsewhere in the molecule; thus let R_1X , R_2X , R_3X , ... constitute reactions series X. Similarly, consider reaction series Y consisting of compounds R_1Y , R_2Y , R_3Y , Suppose we subject both series to reactions appropriate to the functional groups X and Y and measure the equilibrium constants of all these reactions.

According to thermodynamics, each equilibrium constant is related to a standard free energy change by Eq. (C.1) (see Chapter 4, Section 4.3):

$$\Delta G^0 = -RT \ln K \tag{C.1}$$

In attempting to interpret our set of experimental data, therefore, we might be led by this equation to make a plot of the logarithms of the equilibrium constants of series X against those of series Y; that is, $\log K_1^X$ (for compound R₁X) is plotted against $\log K_1^Y$ (for compound R₁X), and so on. If the two reaction series are not greatly different (and

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there is much chemical meaning embodied in those few words), we often observe that this plot yields a straight line. Let us work out the consequences of such a result.

The empirically observed linear plot of log K^X against log K^Y means that

$$\log K^{\rm X} = m \log K^{\rm Y} + b \tag{C.2}$$

where *m* is the slope of the plot and *b* is the intercept. Combining Eqs. (C.1) and (C.2) gives

$$\Delta G_{\rm X}^0 = m \Delta G_{\rm Y}^0 + b' \tag{C.3}$$

where b' = -2.3RTb. Because of the form of Eq. (C.3), a linear plot according to Eq. (C.2) is called a *linear free energy relationship* (LFER). Such a relationship clearly has practical uses in succinctly expressing a large mass of experimental data, and it offers a means for predicting equilibrium constants for members of a reaction series that have not been experimentally studied. Beyond this, the LFER suggests the existence of an underlying simplicity of chemical behavior, and thus it may stimulate further experimentation or theoretical investigation.

LFER sometimes appear in a slightly different format. Suppose that, in each reaction series, a particular substituent is designated as a "reference" substituent; very often the unsubstituted member of the series plays this role. Let K_0^X and K_0^Y be the equilibrium constants for the reference compounds. Equation (C.4) can be written

$$\log K_0^{\mathrm{X}} = m \log K_0^{\mathrm{Y}} + b \tag{C.4}$$

because the point $(\log K_0^X, \log K_0^Y)$ satisfies Eq. (C.2). Subtracting Eq. (C.4) from Eq. (C.2) gives

$$\log \frac{K^{\mathrm{X}}}{K_0^{\mathrm{X}}} = m \log \frac{K^{\mathrm{Y}}}{K_0^{\mathrm{Y}}} \tag{C.5}$$

as an alternative plotting form.

This treatment can be carried somewhat further. Let us suppose that reaction series Y is exceptionally "well-behaved" in the sense that the equilibrium is simple, the equilibrium constant can be accurately measured, and for good chemical reasons we think that K^{Y} is an uncomplicated measure of some molecular-level phenomenon such as electron distribution.

Then there is an advantage in defining series Y as a reference series, and using it to attempt to discover LFER with many other processes. This has been done. The reference reaction that is widely useful is the aqueous acid dissociation at 25°C of *meta*- and *para*-substituted benzoic acids; Table 12.9 lists pK_a values for many of these acids. A quantity σ is then defined

$$\sigma = \log \frac{K_a}{K_a^0} \tag{C.6}$$

where K_a^0 is the acid dissociation constant for unsubstituted benzoic acid, and K_a is the corresponding constant for a substituted benzoic acid. σ is called the *Hammett substituent constant*, and with its use we can rewrite Eq. (C.5) as

$$\log \frac{K^{\rm X}}{K_0^{\rm X}} = \rho \sigma \tag{C.7}$$

where ρ (called the reaction constant) is the slope of a plot of the left-hand side, obtained from data on a reaction series X, against σ . The values of σ (the *meta* and *para* series have different σ values) are interpreted as measures of the substituent's ability to attract electrons from or to release electrons to the reaction site. (The student may easily calculate σ_m and σ_p values from the data in Table 12.9.)

Note that σ , defined by Eq. (C.6), is a perfectly valid thermodynamic quantity, because the equilibrium constants that enter its calculation are thermodynamically defined. However, the interpretation of σ is a measure of electron-release or electron-withdrawal is extrathermodynamic in nature, as is the inference that the LFER in Eq. (C.7) has fundamental chemical significance.

The Connection Between Rates and Equilibria. In Chapter 1 a distinction was made between path-independent functions, called state functions, and path-dependent functions. Many of the important quantities in thermodynamics, such as changes in free energy, enthalpy, and entropy, are state functions: their values depend only upon the values of the quantity in the initial and final states. In consequence, we can learn nothing from such quantities about the path between the initial and final states. In reacting chemical systems, the path taken by the system in passing from the initial to the final state is called the reaction mechanism. Thus thermodynamics does not in detail inform us about reaction mechanisms. In order to acquire mechanistic information, we must study the rate of a chemical reaction, because the rate does depend upon the path. The study of reaction rates is called chemical kinetics.

Not surprisingly, the field of chemical kinetics has its own theories, and for our purpose the most useful of these is called the *transition state theory*. Let us begin by writing this generalized chemical reaction:

I⇒F

where I represents the initial state [i.e., reactant(s)] and F denotes the final state [product(s)]. We suppose that we can measure the equilibrium constant *K* and thus, from Eq. (C.1), calculate ΔG^0 for the reaction.

Now let us ask, Why does not all of I instantaneously transform itself into F? After all, if some of I reacts to give F in a given time interval, what is holding back the rest of it?

The answer is that there is an energy barrier on the reaction path between I and F, and at any given temperature only a certain fraction of I possesses sufficient thermal energy to surmount this barrier. The essence of the transition state theory is that at or near the top of this energy barrier the reacting system exists as a *transition state*, and the

initial and transition states are in virtual equilibrium. (That "virtual" is necessary because the transition state is a very special kind of molecule, possessing a vibrational mode of motion different from an initial state molecule.) If we assume equilibrium between the initial state and the transition state, then all of the equilibrium concepts of thermodynamics can be applied to this process. So let us expand our hypothetical reaction to include its transition state:

$$I \rightleftharpoons [M^{\ddagger}] \rightleftharpoons F$$

(The "double dagger" symbol always refers to the transition state or a quantity associated with it.) The transition state theory then gives Eq. (C.8) as the key result (Connors, 1990, Chapter 5).

$$k = \frac{kT}{h}K^{\ddagger} \tag{C.8}$$

In Eq. (C.8), *k* is the rate constant of the reaction (an introduction to rate constants is given in Chapter 0), $\tilde{k} = R/N_A$ is the Boltzmann constant, *T* is the absolute temperature, *h* is Planck's constant (see Appendix A), and K^{\ddagger} is the equilibrium constant for the initial state-to-transition state process.

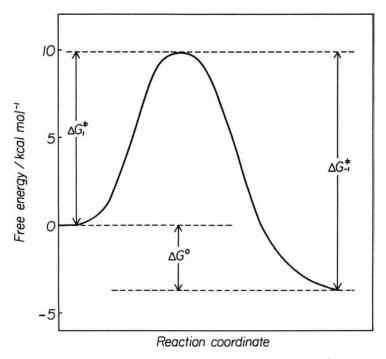


Figure C.1. Free energy reaction coordinate diagram for the reaction $I \rightleftharpoons [M^{\ddagger}] \rightleftharpoons F$ [Reproduced by permission from Connors (1990).]

Since, through this transition state concept, we have imported the results of thermodynamics into chemical kinetics, we go further and define ΔG^{\ddagger} , ΔH^{\ddagger} , and ΔS^{\ddagger} , which are respectively called the free energy, enthalpy, and entropy of activation. These can be evaluated from measurements of the rate constant *k* by means of these familiar equations:

$$\Delta G^{\ddagger} = -RT \ln K^{\ddagger} \tag{C.9}$$

$$\Delta \mathbf{G}^{\ddagger} = \Delta H^{\ddagger} - T \Delta S^{\ddagger} \tag{C.10}$$

$$\frac{d\ln K^{\ddagger}}{d(1/T)} = -\frac{\Delta H^{\ddagger}}{R} \tag{C.11}$$

We can make these ideas more concrete with Fig. C.1, which is a *reaction* coordinate diagram. The vertical axis is measured in units of free energy per mole. The horizontal axis, labeled reaction coordinate, is not a well-defined quantity, but can be interpreted as a measure of the progress (reading left to right) of the transformation of initial state to final state. In the figure, the thermodynamic quantity ΔG^0 is seen to be determined solely by the free energy difference between the initial and final states.

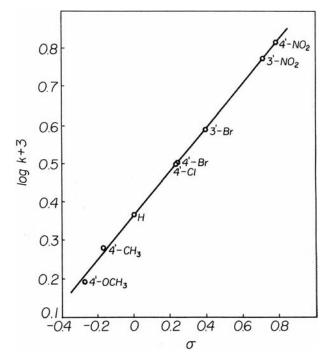


Figure C.2. Linear free energy relationship for the kinetics of alkaline hydrolysis of substituted ethyl 4-biphenylcarboxylates at 40°C; σ is the Hammett substituent constant. [Reproduced by permission from Connors (1990).]

Now consider the quantity labeled ΔG_1^{\ddagger} . This is the free energy of activation for the forward reaction, and it is given by the difference in free energies of the transition and initial states. (The quantity ΔG_{-1}^{\ddagger} is the corresponding change for the reverse reaction.)

The point we have reached allows us to consider the question, Is there a relationship between ΔG^0 (i.e., the equilibrium) and ΔG_1^{\ddagger} (the rate) of the reaction? From Fig. C.1 we evidently must answer that there is no *necessary* relationship. On the other hand, Fig. C.1 does bring these two quantities together in a theoretical context that perhaps leads us to suppose that there *may* be such a relationship. And indeed such relationships have been observed. Even more interesting, perhaps, are examples of LFER between rates and equilibria of *different* reactions. Suppose the rate constants of reaction series B are linearly correlated with the equilibrium constants of reaction series A:

$$\log k_B = m \log K_A + b \tag{C.12}$$

From Eqs. (C.1) and (C.9), combined with Eq. (C.12), we find

$$\Delta G_B^{\ddagger} = m \Delta G_A^0 + b' \tag{C.13}$$

Figure C.2 is such an LFER in which the Hammett substituent constant provides the equilibrium quantity and the rate constants are for an alkaline ester hydrolysis.

ANSWERS TO PROBLEMS

Chapter 0

- 0.1. (a) $[H^+] = 1.78 \times 10^{-5} M$ (b) $[H^+] = 7.41 \times 10^{-12} M$ (c) $[H^+] = 6.46 \times 10^{-8} M$ (d) $[H^+] = 1.00 \times 10^{-2} M$
- **0.2.** (a) pH = 2.41
 - **(b)** pH = 8.94
 - (c) pH = 7.19
 - (**d**) pH = -0.06
- **0.3.** $\log x = (\ln x)/2.303 = 0.434 \ln x$
- **0.4.** One million times more acidic (i.e., 10^6).
- **0.5.** *q* = 4.20
- **0.7.** 0.1526 g benzoic acid
- **0.8.** 6.82 g acetic acid
- **0.9.** $R_{A^-} = F_{A^-} / (1 F_{A^-}) \text{ or } F_{A^-} = R_{A^-} / (1 + R_{A^-})$
- **0.10.** $R_{\text{HA}} = R_{\text{A}^-} = 1.0; F_{\text{HA}} = F_{\text{A}^-} = 0.5$

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0.11. [HA] = 6.52×10^{-4} M; [A⁻] = 0.98×10^{-4} M **0.12.** pH = 2.41 **0.13.** pH = 2.41 **0.14.** -b/m **0.16.** $\varepsilon = 2.08 \times 10^{4}$ M⁻¹ cm⁻¹ **0.17.** K = 10.8 M⁻¹ **0.18.** $\Delta H = 7.73$ kcal mol⁻¹ **0.19.** $K_{11} = 41$ M⁻¹; $K_{12} = 345$ M⁻¹

- **0.20.** $k = 4.90 \times 10^{-3} \, \mathrm{s}^{-1}$
- **0.21.** (y-1)/x = a bx (there are other possible linear forms also).
- **0.22.** The plot of $\Delta c/\Delta t$ against \bar{c} is reasonably linear, so it has the equation $-\Delta c/\Delta t = k\bar{c}$, with $k = 6.1 \times 10^{-4} \,\mathrm{s}^{-1}$. You have obtained an experimental rate equation, where $\Delta c/\Delta t$ is the rate of reaction and k the rate constant. Compare your value of k with the value calculated in Example 0.11.
- $0.23. \ dy/dx = a^u \ln a(du/dx)$
- **0.24.** (a) $dy/dx = abx^{b-1}$ (b) At x = 0.5, y = a/4, dy/dx = a; at x = 1, y = a, dy/dx = 2a; at x = 2, y = 4a, dy/dx = 4a

0.25.
$$dy/dx = b + 2cx$$
; $d^2y/dx^2 = 2c$

- **0.26.** $V_{\text{endpoint}} = 1.83 \text{ mL}$; equivalent weight = 266.6
- **0.27.** $pH_{\min} = \frac{1}{2}pK_w + \frac{1}{2}\log(k_1/k_3)$, where $pK_w = -\log K_w$ and $K_w = [H^+][OH^-]$
- **0.28.** *y* = 504
- **0.29.** $t_{1/2} = \ln 2/k = 0.693/k$
- **0.30.** Because the constant of integration will disappear anyway in the subtraction process.

0.31.
$$dw = \left(\frac{\partial w}{\partial x}\right)_{y,z} dx + \left(\frac{\partial w}{\partial y}\right)_{x,z} dy + \left(\frac{\partial w}{\partial z}\right)_{x,y} dz$$

0.32. $\bar{x} = 2392 \text{ M}^{-1}$; $s = 484 \text{ M}^{-1}$; $s_m = 140 \text{ M}^{-1}$; RSD = 20.2%

- **0.33.** The plot of log (c_B/c_A) against *t* has r = 0.981, intercept = 0.262, slope = 0.107 min⁻¹, from which is obtained $k = 0.585 \text{ M}^{-1} \text{ min}^{-1} = 9.75 \times 10^{-3} \text{ M}^{-1} \text{s}^{-1}$.
- **0.34.** $\bar{x} = 1.59$, s = 0.139, $s_m = 0.046$; confidence limits $(P = 0.05) = 1.59 \pm 0.11$

- **0.35.** $\bar{x} = 2.4 \times 10^3 \text{ M}^{-1}$; $s_m = 1.4 \times 10^2 \text{ M}^{-1}$ (note that each of these quantities possesses two significant figures; the numbers 2.4×10^3 and 2400 are subtly different in this regard)
- **0.36.** 69.9 kJ mo1^{-1}
- **0.37.** $1.39 \times 10^{-9} \, \mathrm{s}^{-1}$

0.38.
$$\left(\frac{1.86 \text{ g}}{\text{mL}}\right) \left(\frac{10^3 \text{mL}}{1 \text{ L}}\right) \left(\frac{1 \text{ L}}{10^{-3} \text{m}^3}\right) \left(\frac{1 \text{ kg}}{10^3 \text{g}}\right) = 1.86 \times 10^3 \text{kgm}^{-3}$$

0.39.
$$(0.77\text{ Å})\left(\frac{10^{-10}\text{m}}{1\text{ Å}}\right)\left(\frac{10^9\text{ nm}}{1\text{ m}}\right) = 0.077\text{ nm}$$

0.40.
$$\left(\frac{8.314 \text{ J}}{\text{K mol}}\right) \left(\frac{1 \text{ cal}}{4.184 \text{ J}}\right) = 1.987 \text{ cal } \text{K}^{-1} \text{mol}^{-1}$$

- **0.41.** Use quantity algebra to generate a pure number, and then take the logarithm of this pure number. For example, the expression log K_a really should be interpreted as log (K_a/M) . We use the former symbolism in equations for convenience.
- **0.42.** Since A is dimensionless and $\varepsilon = A/bc$, ε has the units $L \mod^{-1} \operatorname{cm}^{-1}$, or $\operatorname{M}^{-1}\operatorname{cm}^{-1}$.
- 0.43. Equation (a) is incorrect.

Chapter 1

- **1.1.** $\Delta V = 35.343 \text{ in.}^3 = 0.579 \text{ L}$ $w = P\Delta V = 0.579 \text{ L} \text{ atm} = 58.7 \text{ J}$
- **1.2.** $w = -1718 \,\mathrm{J}$
- **1.3.** $\Delta U = 0$ and $w = P\Delta V$, so $q = w = P\Delta V$, leading to Eq. (1.15) for q.
- **1.4.** $C_P = 18.02 \text{ cal mol}^{-1} \text{ K}^{-1}$
- **1.5.** Specific heat = $1.74 \text{ Jg}^{-1} \text{ K}^{-1}$
- **1.6.** T (final) = 25°C + ΔT = 105 °C
- **1.7.** Write Eq. (1.26) as $\Delta H = C_P(T_2 T_1)$, applying it both to water and to iron. Since ΔH (water) = $-\Delta H$ (iron), and T_2 is the same for both, we find $T_2 = 343.36$ K or 70.2 °C.
- **1.8.** P = 26.3 atm; T = -78 °C (not all the dry ice has sublimated).
- **1.9.** MW = 56.3 g/mol; $\mu = 3.6 \times 10^4$ cm/s.
- **1.10.** $\delta_{\text{Initial}} = 4.28 \times 10^{-3} \text{ g cm}^{-3}; \ \delta_{\text{Final}} = 6.8 \times 10^{-7} \text{ g cm}^{-3}.$

- **1.11.** $P_{CHCl_3(ideal)} = 0.255 \text{ atm.}$
- **1.12.** MW_{nitrous oxide} = 44.15 g/mol; $\mu = 3.93 \times 10^4$ cm s⁻¹; $\delta = 1.97 \times 10^{-3}$ g cm⁻³.
- **1.13.** $\Delta H_f + \Delta H_v \Delta H_s = 0$

- 2.1. (a) (-) System becomes more ordered as it crystallizes.
 - (**b**) (+) System becomes more random as it vaporizes.
 - (c) (-) Two particles combine to yield one particle, with decrease in number of microstates.
- **2.2.** Using Trouton's rule, $\Delta H_v = (21)(353.25) 7.42 \text{ kcal mol}^{-1}$. (The experimental value is 7.35 kcal mol⁻¹.)
- **2.3.** $\Delta S_f = 5040/419.15 = 12.02 \text{ cal } \text{K}^{-1} \text{ mol}^{-1}$
- **2.4.** $\Delta S_s = 17,600/298.15 = 59.0 \text{ cal } \text{K}^{-1} \text{ mol}^{-1}$
- **2.5.** $\Delta S = 9.57 \,\mathrm{J}\,\mathrm{K}^{-1}$ (i.e., 19.14 $\mathrm{J}\,\mathrm{K}^{-1}\,\mathrm{mo1}^{-1}$)
- **2.6.** $\Delta S = 2.61 \text{ cal K}^{-1} \text{ mol}^{-1}$
- **2.7.** $\Delta S = R \ln (P_1/P_2)$
- **2.8.** $\Delta S = 2.75 \text{ cal } \text{K}^{-1}.$
- **2.9.** $\Delta S = 0.11 \text{ cal } \text{K}^{-1}$.
- **2.10.** $T = 100 \,^{\circ}\text{C}$; $\Delta S = 68 \,\text{cal }\text{K}^{-1}$.
- **2.11.** $\Delta S = 12 \text{ cal } \text{K}^{-1}$.

- **3.1.** a = 0.0175 m (from data in Table 3.1)
- **3.2.** $\Delta \mu = \mu_{(0.05m)} \mu_{(0.005m)} = 5.41 \text{ kJ mol}^{-1}$
- **3.3.** $\Delta \mu = -3.44 \text{ kJ mol}^{-1}$
- **3.4.** Making a solution more concentrated yields a positive free-energy change; the process is nonspontaneous. (See Problem 3.2.) In Problem 3.3 the process takes the solution from 1 M (the standard state) to 0.25 M, and this is spontaneous. It is possible for nonideal behavior (activity coefficient effects) to reverse this conclusion, however.
- **3.5.** $\Delta G = 0.$

- **4.1.** From the integrated Clausius–Clapeyron equation, $\Delta H_{\text{vap}} = 9.45 \text{ kcal mol}^{-1} = 39.5 \text{ kJ mol}^{-1}$
- **4.2.** $\Delta G^0 = -1.82 \text{ kcal mol}^{-1} = -7.61 \text{ kJ mol}^{-1}$
- **4.3.** $\Delta H_{\rm vap} = +8.79 \,\rm kcal \, mol^{-1}$
- **4.4.** $\Delta G^0 = 2.303 \, \text{RT} \, \text{p}K_a$
- **4.5.** $C_6H_5OH \rightleftharpoons H^+ + C_6H_5O^ K_a = a_{H^+}a_{PhO^-}/a_{PhoH}(Ph \equiv C_6H_5)$ $\Delta G^0 = +1.36 \text{ kcal mol}^{-1}$
- **4.6.** $\Delta S^0 = -16.9 \text{ cal mol}^{-1} \text{ K}^{-1}$
- 4.7. We might expect ΔS^0 to be positive, because one particle is being transformed into two particles, with (presumably) an increase in number of accessible microstates. Since ΔS^0 is negative, however, we may infer that something else, something not apparent in the reaction as written, must be occurring. The most likely possibility is that the ions on the right-hand side of the equation are limiting the motions of solvent (water) molecules via strong ion–dipole interactions. The implication is that, taking into account the solvent, there are more particles, and therefore more accessible microstates, on the left-hand side of the equation.
- **4.8.** $\Delta G^0 = -5.25 \text{ kcal mol}^{-1}$.
- **4.9.** $\Delta S = -10.84 \text{ cal mo1}^{-1}$.
- **4.10.** $\Delta H^0 = -1.358 \text{ kcal mo1}^{-1}$; The van't Hoff equation shows that if the heat of dissociation is negative, then the equilibrium constant at a certain temperature will be smaller than the equilibrium constant at a lower temperature.
- **4.11.** $\Delta S_v = 23.0 \text{ cal } \text{K}^{-1} \text{ mol}^{-1}$.
- **4.12.** (a) 2; (b) 2.
- **4.13.** $\Delta G^0 = 6.47 \, \text{kcal mol}^{-1}$.

- **5.1.** $x_2 = 0.0000270$, so $x_1 = 1 x_2 = 0.999973$
- 5.2. About 12.4 M
- **5.3.** 55.5 M
- **5.4.** $\mu_c^0 \mu_x^0 = -RT \ln 55.5 = -2.38 \text{ kcal mol}^{-1} = -9.96 \text{ kJ mol}^{-1}$

5.5. $c_i = M_i \rho_1$

5.6.
$$x_2 = \frac{c_2(n_1M_1 + n_2M_2)}{1000(n_1 + n_2)}$$

where M_1 and M_2 are molecular weights. [Suggestion: Start by writing the mass of solution as $n_1M_1 + n_2M_2$; then use ρ to write the volume of solution as $(n_1M_1 + n_2M_2)/1000 \rho$ in liters.]

- 5.7. 0.098 m.
- **5.8.** X = 0.005; % w/w = 4.8%

Chapter 6

- **6.1.** 45 °C
- 6.2. C=2 and P=3, so F=1; however, the pressure is fixed by experimental design, so there remain no degrees of freedom.

Chapter 7

- 7.1. $\Delta S_{\text{mix}}^{\text{ideal}} = + 1.36 \text{ cal mol}^{-1} \text{ K}^{-1} = + 5.70 \text{ J mol}^{-1} \text{ K}^{-1}$ $\Delta G_{\text{mix}}^{\text{ideal}} = -0.41 \text{ kcal mol}^{-1} = -1.72 \text{ mol}^{-1}$
- 7.2. Alanine: $\Delta \mu^0 = +4.01 \text{ kcal mol}^{-1} = 16.8 \text{ kJ mol}^{-1}$ Phenothiazine: $\Delta \mu^0 = -5.66 \text{ kcal mol}^{-1} = -23.7 \text{ kJ mol}^{-1}$
- **7.3.** $X_B = 0.23$; total pressure = 34 mmHg
- **7.4.** k^{x} (CHC1₃) \approx 157 mmHg
- 7.5. Six extractions
- 7.7. (a) $P_{AB} = c_A/c_B$; $P_{BC} = c_B/c_C$; $P_{AC} = c_A/c_C$ (b) $P_{AB} = P_{AC}/P_{BC}$ (c) $p_A = P_{AB}V_A/(P_{AB}V_A + V_B + V_C/P_{BC})$

7.8. P = 0.25; 1.00; 4.00

7.9. $k_{\rm H}$ has the dimensions of a pressure, therefore $k_{\rm H}/P$ is a dimensionless quantity.

7.10. $\Delta S_{\text{mix}} = 1.19 \text{ cal mol}^{-1} \text{ K}^{-1}; \Delta G_{\text{mix}} = 354.6 \text{ cal mol}^{-1}.$

- 8.1. $B + AcOH \rightleftharpoons AcO^{-}BH^{+} + AcO^{-}$
- 8.2. NaCl, I = 3.0 m; CaCl₂, I = 9.0 m; ZnSO₄, I = 12.0 m

- 8.3. $\gamma_{\pm} = 0.807$
- **8.4.** $\gamma_+ = 0.850; a_{\pm} = 0.0085M$
- 8.5. I = 0.65 M
- 8.6. $C = 0.21|z_+z_-|$

- **9.1.** From $p_1 = x_1 P_1^*$, derive $P_1^* p_1 = x_2 P_1^*$.
- **9.2.** $K_b = 1.23 \,^{\circ}\text{C}$
- **9.3.** $K_f = 3.58 \,^{\circ}\text{C}$
- **9.4.** Take 0.6 g pilocarpine nitrate, 0.3 g boric acid, and dilute to 30 mL ("q.s. ad" is a prescription abbreviation meaning "a quantity sufficient to make"; sterile water is intended in this case).
- **9.5.** $\Delta T_f = 0.469 \,^{\circ}\text{C}$ (the experimental value is 0.470 $^{\circ}\text{C}$); 5.54% dextrose will be isotonic.
- 9.6. $2\Delta H_{f_A} = \Delta H_{f_B}$
- **9.7.** $\gamma_{\rm C} = 0.84; \gamma_{\rm D} = 0.94.$
- **9.8.** $\Delta T_f = K_f \text{ m} = 0.372 \text{ K}; \ \pi = cRT = 4.99 \text{ atm}$

Chapter 10

10.1. $x_2 = 0.158$

- **10.2.** At 24 °C (e.g.), $c_2 = 3.34 \times 10^{-4}$ M. The heat of solution is identical with that obtained in Example 10.2. (Enthalpy changes do not change value when the concentration scale is altered, unlike free-energy and entropy changes.)
- **10.3.** $K_{\rm sp} = 108 {\rm s}^5$; $s = 7.14 \times 10^{-7} {\rm M}$
- 10.4. AgI, then AgBr, then AgCl
- **10.5.** $x_2 = 0.0117$ (calculated taking $\phi_1 = 1$) (the experimental value is $x_2 = 0.0142$)
- **10.6.** $\log c_2 = -4.50$ [Eq. (10.29)] or $\log c_2 = -4.87$ [Eq. (10.30a)] (the experimental value is $\log c_2 = -4.42$)
- **10.7.** $\phi_2 = 0.55$.
- **10.8.** $\Delta G^0 = 3.03 \text{ kcal mol}^{-1}$.

11.1. $67.6 \,\mathrm{dyn} \,\mathrm{cm}^{-1}$

11.2. Water on ether: S = -66Ether on water: S = +44.6Ether will spread on water readily; water will not spread on ether.

11.3.
$$\Delta G = \gamma \Delta A = 4\pi \gamma (r_{final}^2 - r_{initial}^2)$$
$$\Delta G = 2.23 \times 10^{-4} \,\mathrm{mJ}$$

11.4. (a) $l/n_b = 1/Kn_{max}c_2 + l/n_{max}$ (b) $n_b/c_2 = -Kn_b + Kn_{max}$

11.5. $K = 386 \text{ M}^{-1}$; $y_{\text{max}} = 146 \text{ mg g}^{-1}$ (we also calculate $\sigma = 105 \text{ Å}^2 \text{ molecule}^{-1}$)

Chapter 12

- **12.1.** (a) pH = 3.60(b) pH = 3.30(c) pH = 10.93(d) pH = 5.32(e) pH = 11.48
- **12.2.** pH = 4.67
- **12.3.** Dissolve 6.055 g tris in 325.3 mL 0.10 M HCl and dilute to 500 mL.
- **12.4.** (a) Both are $\frac{1}{15}$ M = 0.0667 M (b) pH = 6.98
- **12.5.** At 0 mL, pH = 11.95; at 2 mL, pH = 11.45; at 5 mL, pH = 10.94; at 8 mL, pH = 10.60; at 12 mL, pH = 10.12; at 16 mL, 5.76; at 18 mL, pH = 1.84; methyl red or bromcresol purple
- **12.7.** (a) $K_{eq} = K_a$ (benzoic acid)/ K_a (methylamine) = 2.76 × 10⁶ (b) $K_{eq} = K_a$ (phenol)/ $K_w = 1.00 \times 10^4$
- **12.8.** pK_1 (COOH); pK_2 (OH) pK_1 (COOH); pK_2 (NH₂); pK_3 (guanidine) pK_1 (aromatic amine); pK_2 (aliphatic amine) This is an acidic group, the 7-NH is structurally similar to an imide.

12.9.
$$[\mathrm{H}^+]^3 + (K_a + b)[\mathrm{H}^+]^2 - (K_a c - K_a b + K_w)[\mathrm{H}^+] - K_a K_w = 0$$

12.10. (a) $[NH_4^+] + [H^+] = [OH^-] + [Cl^-]$ (b) $[H^+] = [OH^-] + [H_2A^-] + 2[HA^{2-}] + 3[A^{3-}]$ (c) $[Na^+] = [H^+] + [OH^-] + [H_2PO_4^-] + 2[HPO_4^{2-}]$ **12.11.** pH = 5.31

- 12.12. pH = 7.69
- **12.13.** (a) $pH = pK_a = 9.25$ (b) pH = 5.52
- **12.14.** $\Delta G^0 = +13.64 \text{ kcal mol}^{-1}$
- **12.15.** 0.1689 g of sodium acetate
- **12.16.** F = R/R + 1)
- **12.17.** $3.91 \times 10^{-4} \,\mathrm{M}$
- **12.18.** (a) Decrease (b) Decrease

12.19. pH = 1.70

- **12.20.** 8.528 g of NaH_2PO_4 ·H₂O and 5.424 g of Na_2HPO_4
- **12.21.** $\Delta G^0 = -19.1 \text{ kcal mol}^{-1} = -79.8 \text{ kJ mol}^{-1}$

13.1. (a)
$$ClO_3^- + 3Sn^{2+} + 6H^+ = Cl^- + 3Sn^{4+} + 3H_2O$$

(b) $PbO_2 + 2I^- + 4H^+ = I_2 + Pb^{2+} + 2H_2O$
(c) $3OCl^- + 2NH_3 = 3Cl^- + N_2 + 3H_2O$
(d) $2MnO_4^- + 5H_2O_2 + 6H^+ = 2Mn^{2+} + 5O_2 + 8H_2O$
(e) $MnO_4^- + 3CuI + 4H^+ = MnO_2 + 3Cu^{2+} + 3I^- + 2H_2O$
(f) $S_2O_8^{2-} + 2Fe^{2+} = 2SO_4^{2-} + 2Fe^{3+}$
(g) $2NH_2OH + 4Ce^{4+} = N_2O + 4Ce^{3+} + H_2O + 4H^+$
(h) $2RSH + I_2 = RSSR + 2HI$

- **13.2.** *I* = 0.150 M
- **13.3.** $\Delta E = -0.59 \, \mathrm{V}$
- **13.4.** p*K*′_{*a*} = 8.02
- **13.5.** (a) Slope of plot of E_{cell} versus log (Ca²⁺ activity) = 0.029, consistent with 0.059/2
 - **(b)** $E_{\text{cell}} = + 0.162 \text{ V}$
- **13.6.** (a) I = 0.04 M(b) $\gamma_{\pm} = 0.458$
- **13.7.** (a) $E_{\text{cell}} = -0.548 \text{ V}$
 - (b) E_{cell} is negative, so reaction is nonspontaneous (proceeds from right to left); copper dissolves. $2Fe^{2+} + Cu^{2+} \rightleftharpoons Cu + 2Fe^{3+}$

- **13.8.** (a) $E_{cell} = +1.44 \text{ V}$ (b) Silver-plated zinc
- **13.9.** $E^0 = +0.15$ V; K = 349**13.10.** $pK_{sp} = -10.85$

14.1. $\beta_{23} = K_{11}K_{21}K_{22}K_{23}$ **14.3.** Initial slope $= S_tK_{11}\Delta\varepsilon_{11}$ **14.4.** (a) $\frac{\Delta A/b}{[L]} = -K_{11}(\Delta A/b) + S_tK_{11}\Delta\varepsilon_{11}$

(b)
$$K_{11} = 10.2 \,\mathrm{M}^{-1}; \Delta \varepsilon_{11} = 257 \,\mathrm{M}^{-1} \,\mathrm{cm}^{-1}$$

- **14.5.** Initial slope $= K_{11a} K_{11b}$
- **14.6.** [theophylline]_{free} = 5×10^{-5} M.

14.7. (a)
$$f_{11} = \frac{[L]nK_{11}}{[L]K_{11}+1}$$
; (b) $1 - f_{11} = 0.71$; (c) $[L] = 9 \times 10^{-7}$ M.

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