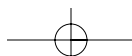
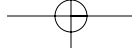


**Late Cretaceous  
and Cenozoic Mammals  
of North America**

---



# **Late Cretaceous and Cenozoic Mammals of North America**

---

Biostratigraphy and Geochronology

---

Edited by Michael O. Woodburne



Columbia University Press

New York



**COLUMBIA UNIVERSITY PRESS**

*Publishers Since 1893*

New York Chichester, West Sussex

Copyright © 2004 Columbia University Press

All rights reserved

Library of Congress Cataloging-in-Publication Data

Late cretaceous and cenozoic mammals of North America : biostratigraphy and geochronology / edited by Michael O. Woodburne.

p. cm.

Includes bibliographical references.

ISBN 0-231-13040-6 (cloth)

1. Mammals, Fossil—North America. 2. Paleontology—Cenozoic. 3. Paleontology—Cretaceous. 4. Paleontology—North America. I. Woodburne, Michael O.

QE881.L26 2004

569.'097—dc21

2003046251

⊗

Columbia University Press books are printed on permanent and durable acid-free paper.

Printed in the United States of America

c 10 9 8 7 6 5 4 3 2 1

---

## Contents

*Preface* M. O. Woodburne *vii*

*List of Contributors* *ix*

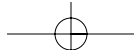
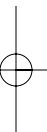
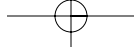
*Definitions* *xi*

*Introduction* M. O. Woodburne *xvii*

- |   |  |
|---|--|
| <p><b>1. Principles and Procedures</b> <span style="float: right;"><b>1</b></span><br/>M. O. Woodburne</p>  | <p><b>6. Mammalian Biochronology of the Arikarean Through Hemphillian Interval (Late Oligocene Through Early Pliocene Epochs)</b> <span style="float: right;"><b>169</b></span><br/>R. H. Tedford,* L. B. Albright III, A. D. Barnosky, I. Ferrusquia-Villafranca, R. M. Hunt Jr., J. E. Storer, C. C. Swisher III, M. R. Voorhies, S. D. Webb, and D. P. Whistler</p> |
| <p><b>2. Mammalian Biochronology of the Latest Cretaceous</b> <span style="float: right;"><b>21</b></span><br/>R. L. Cifelli,* J. J. Eberle, D. L. Lofgren, J. A. Lillegraven, and W. A. Clemens</p>  | <p><b>7. The Blancan, Irvingtonian, and Rancholabrean Mammal Ages</b> <span style="float: right;"><b>232</b></span><br/>E. L. Lundelius Jr. and C. J. Bell,* A. D. Barnosky, R. W. Graham, E. H. Lindsay, D. R. Ruez Jr., H. A. Semken Jr., S. D. Webb, and R. J. Zakrzewski</p>   |
| <p><b>3. Paleocene Biochronology: The Puercan Through Clarkforkian Land Mammal Ages</b> <span style="float: right;"><b>43</b></span><br/>D. L. Lofgren,* J. A. Lillegraven, W. A. Clemens, P. D. Gingerich, and T. E. Williamson</p>                            | <p><b>8. Global Events and the North American Mammalian Biochronology</b> <span style="float: right;"><b>315</b></span><br/>M. O. Woodburne</p>  |
| <p><b>4. Wasatchian Through Duchesnean Biochronology</b> <span style="float: right;"><b>106</b></span><br/>P. Robinson,* G. F. Gunnell, S. L. Walsh, W. C. Clyde, J. E. Storer, R. K. Stucky, D. J. Froehlich, I. Ferrusquia-Villafranca, and M. C. McKenna</p> | <p><i>Systematic Index</i> <i>345</i></p> <p><i>Subject Index</i> <i>361</i></p>   |
| <p><b>5. The Chadronian, Orellan, and Whitneyan North American Land Mammal Ages</b> <span style="float: right;"><b>157</b></span><br/>D. R. Prothero and R. J. Emry</p>   |  |

---

\*Chairman of the committee of contributors



---

## Preface

**T**HIS BOOK UPDATES the information contained in its 1987 progenitor, *Cenozoic Mammals of North America: Geochronology and Biostratigraphy*, to further refine the tempo and mode of mammalian faunal succession in North America, with the major steps being recognized as discrete intervals known as North American land mammal ages. In the present work, the coverage is extended temporally to include the Lancia part of the Late Cretaceous, as precursor to the Cenozoic, and geographically to include information from Mexico, an integral part of the North American fauna, past and present.

This work incorporates new information on the systematic biology of the fossil record inspected herein but also uses the many advances in geochronologic methods and their results obtained since 1987. It is hoped that what follows here can lead to an increasingly high-resolution stratigraphy in which all available temporally significant data and applications are integrated. Fundamental to achieving this goal are using procedures to enable chronologic units to be recognized and their boundaries defined (no gaps or overlaps), establishing the units in actual field settings so that they are both replicable and realistically complete, and using radioisotopic, cyclostratigraphic, and magnetostratigraphic means to assist in developing as highly refined a correlation network as possible. The goal is a robust high-resolution chronology and, potentially, a chronostratigraphy.

As discussed more fully in the *Introduction*, high-resolution chronostratigraphy involves a detailed integration of lithostratigraphic, faunal or (better) biostratigraphic, magnetostratigraphic, cyclostratigraphic,

and radioisotopic data to arrive at the best possible interpretation of the age of a given fossiliferous level.

Whereas radioisotopic data used in 1987 had the advantage of the results of the K--Ar method pioneered by Evernden et al. (1964) unavailable to the original promulgation of the mammal age framework developed by Wood et al. (1941), the present effort benefits from the newly developed  $^{40}\text{Ar}/^{39}\text{Ar}$  laser fusion techniques, unavailable before 1987. Similarly, the 1987 work saw the beginning of the now almost ubiquitous application of paleomagnetic stratigraphy to nonmarine mammal-bearing deposits, and a much richer array of this data set is available for the present book. Isotopic geochemistry provides information on changes in isotopes of oxygen and carbon that are proxies for changes in sea level and climate with implications for the nonmarine record, both as an impetus for faunal change and as tools for correlation. Advances in cyclostratigraphy improve the calibration of the magnetic polarity chronology paradigm, with feedback to the nonmarine correlation framework used here.

Thus the present work differs from the earlier volume in representing improvements in all aspects of the data set designed to promote correlation between fossil mammal-bearing successions in North America and thereby to improve our understanding of the times of faunal change represented by the mammal ages and their chronologic relationship to other important geologic, biological, or climatic events that transpired in the past 80 million years or so and may have shaped the tempo and mode of land mammal faunal succession during that time.

The goal of this book, then, is to place in modern context the information by which North American mammalian paleontologists recognize, divide, calibrate, and discuss intervals of mammalian evolution known as North American land mammal ages.

I dedicate this book to the memory of Donald Elvin Savage and Remmert Daams, two persistent advocates from North America and Europe, respectively, of the efforts and approaches documented herein.

Michael O. Woodburne  
Running Springs, California

---

## REFERENCES

---

- Evernden, J. F., D. E. Savage, G. H. Curtis, and G. T. James. 1964. Potassium–argon dates and the Cenozoic mammal chronology of North America. *American Journal of Science* 262:145–198.
- Wood, H. E. II, R. W. Chaney, J. Clark, E. H. Colbert, G. L. Jepsen, J. B. Reeside Jr., and C. Stock. 1941. Nomenclature and correlation of the North American continental Tertiary. *Bulletin of the Geological Society of America* 52:1–48.
- Woodburne, M. O. (ed.). 1987. *Cenozoic mammals of North America: Geochronology and biostratigraphy*. Berkeley: University of California Press.



---

## List of Contributors

**L. Barry Albright III**

Museum of Northern Arizona, Flagstaff, Arizona 86001

**Anthony D. Barnosky**

Museum of Paleontology, University of California, Berkeley, California

**Christopher J. Bell**

Department of Geological Sciences, University of Texas, Austin, Texas

**Richard L. Cifelli**

Oklahoma Museum of Natural History, University of Oklahoma, Norman, Oklahoma

**William A. Clemens**

Museum of Paleontology, University of California, Berkeley, California

**William C. Clyde**

Department of Geology, University of New Hampshire, Durham, New Hampshire

**Jaelyn J. Eberle**

University of Colorado Museum, Boulder, Colorado

**Robert J. Emry**

Department of Paleobiology, National Museum of Natural History, Smithsonian Institution, Washington, D.C.

**Ismael Ferrusquia-Villafranca**

Instituto de Geologia, UNAM, University of Coyoacan, Mexico

**David J. Froehlich**

Vertebrate Paleontology Laboratory, University of Texas, Austin, Texas

**Philip D. Gingerich**

Museum of Paleontology, The University of Michigan, Ann Arbor, Michigan

**Russell W. Graham**

Denver Museum of Natural History, Denver, Colorado

**Gregg F. Gunnell**

Museum of Paleontology, The University of Michigan, Ann Arbor, Michigan

**Robert M. Hunt Jr.**

University of Nebraska State Museum, Lincoln, Nebraska

**Jason A. Lillegraven**

Department of Geology, The University of Wyoming, Laramie, Wyoming

**Everett H. Lindsay**

Department of Geosciences, University of Arizona, Tucson, Arizona

**Donald L. Lofgren**

Raymond M. Alf Museum of Paleontology, Claremont, California

**Ernest L. Lundelius Jr.**

Department of Geological Sciences, University of Texas, Austin, Texas

**Malcolm C. McKenna**

Department of Vertebrate Paleontology, American Museum of Natural History, New York, New York

**Donald R. Prothero**

Department of Geology, Occidental College, Los Angeles, California

**Peter Robinson**

University of Colorado Museum, Boulder, Colorado

**Dennis R. Ruez Jr.**

Department of Geological Sciences, University of Texas, Austin, Texas

**x** List of Contributors

---

**Holmes A. Semken Jr.**

Department of Geology, University of Iowa, Iowa City, Iowa

**John E. Storer**

Yukon Government Heritage Branch, Whitehorse, Yukon Territory, Canada

**Carl C. Swisher III**

Department of Geological Sciences, Rutgers, The State University of New Jersey, New Brunswick, New Jersey

**Richard K. Stucky**

Denver Museum of Natural History, Denver, Colorado

**Richard H. Tedford**

Department of Vertebrate Paleontology, American Museum of Natural History, New York, New York

**Michael R. Voorhies**

University of Nebraska State Museum, Lincoln, Nebraska

**Stephen L. Walsh**

San Diego Natural History Museum, San Diego, California

**S. David Webb**

Florida Museum of Natural History, University of Florida, Gainesville, Florida

**David P. Whistler**

Division of Earth Sciences, Natural History Museum of Los Angeles County, Los Angeles, California

**Thomas E. Williamson**

New Mexico Museum of Natural History, Albuquerque, New Mexico

**Michael O. Woodburne**

Department of Earth Sciences, University of California, Riverside, California

**Richard J. Zakrzewski**

Department of Earth Sciences and Sternberg Memorial Museum, Fort Hays State University, Hays, Kansas

---

## Definitions

**APTS.** Astronomical polarity time scale. Time scale based on cyclical variations in the stratigraphic record interpreted to reflect astronomical cyclical variations in Earth's orbital progression (Hilgen et al. 1997).

**ASSEMBLAGE CHRON.** This is a new biochronologic unit based on the antecedent assemblage zone and is an interval of time characterized by a distinctive assemblage or association of three or more fossil taxa that, taken together, distinguishes it in biochronologic character from adjacent intervals of time. To the degree that the replication of boundaries is hindered by the number of taxa involved (derived from the antecedent assemblage zone), the utility in precise correlation for the assemblage chron is diminished thereby. Many mammal ages originally were assemblage chrons with little attention given to biostratigraphic data and therefore are not comparable to the assemblage biochron of Walsh (1998) for which the antecedent assemblage zone (Walsh, 1998:160L and figure 5) is based on a biostratigraphic range zone.

**ASSEMBLAGE ZONE.** According to Salvador (1994:62–63), this is “a stratum or body of strata characterized by a distinctive assemblage or association of three or more fossil taxa that, taken together, distinguishes it in biostratigraphic character from adjacent strata.” This is generally similar to Hedberg's (1976:50–52) definition, except for his explicit biofacies connotation. Because of stratigraphic vagaries in ranges of the associated taxa when considered regionally, stratigraphic limits of assemblage zones may be equally variable (Salvador 1994:63). The North American Commission on Stratigraphic Nomenclature (NACSN 1983:863) considers taxon ranges irrelevant and doesn't define boundaries for assemblage zones, apparently because of their ambiguity, whereas boundaries are defined for interval zones and range zones. This treatment differs from the assemblage zone (as-

semblage fossizone or fossilzone) of Walsh (1998, 2000, respectively) in that the latter are based on specified taxon ranges, an approach that effectively synonymizes assemblage and range zones and differs from the language and concept of Hedberg (1976), Salvador (1994), and NACSN (1983).

**BIOCHRON.** According to Salvador (1994), this is “The total time represented by a biozone.” Williams (1901:579) originally defined this term as an interval of geologic time based on the “duration of organic characters.”

**BIOCHRONOLOGY.** “*Geochronology* based on the relative dating of geologic events by biostratigraphic or paleontologic methods or evidence” (Bates and Jackson 1987:69). To the extent that a biochron is based on a biozone, biochronology has a connection to biostratigraphy because the duration of organic characters cannot be demonstrated usefully without recourse to a stratigraphic framework that includes an ordinal paleontologic scale, with or without the addition of numerical data.

**BIOSTRATIGRAPHIC UNIT.** A “body of rock strata that [is] defined or characterized on the basis of [its] contained fossils” (Salvador 1994:53). Kinds of biostratigraphic units include range zone, taxon-range zone, concurrent-range zone, interval zone, lineage zone, assemblage zone, and abundance zone (= acme zone). Fossizone of Walsh (1998) or fossilzone (Walsh 2000) is not used here because it is equivalent in concept to a biozone.

**BIOZONE.** This is a general term for a biostratigraphic zone (Salvador 1994:55).

**CHRON.** *Chron* is the corresponding geochronologic term for a chronozone, the formal lowest-ranking member of the chronostratigraphic hierarchy (Hedberg 1976:69). This means that the chronostratigraphic unit (chronozone) must be established first in order for the chron (geochronologic unit) to be proposed. On this basis, a biozone (bio-

stratigraphic unit) must be developed before an equivalent biologically based chronozone can be identified. In that “the time span of a chronozone is usually defined in terms of the time span of a previously designated stratigraphic unit, such as . . . a biozone” (Hedberg 1976:69), that interval of time is a biochron.

**CHRONOFAUNA.** Following Olson (1952:185), this is a “geographically restricted, natural assemblage of interacting animal populations that has maintained its basic structure over a geologically significant interval of time.” See also Tedford (1970), who stresses that chronofaunas are ecologically interpretive units.

**CHRONOSTRATIGRAPHY.** Chronostratigraphy is “the element of stratigraphy that deals with the *age* of strata and their *time* relations” (Hedberg 1976:66). Salvador (1994:77) replaces *strata* with *rock bodies*, which is not appropriate. According to Aubry et al. (1999:99), chronostratigraphy is “the temporal ordering of geologic strata.” For the purposes of this book, chronostratigraphy deals with strata. Contrary to Walsh (2001), chronostratigraphy is neither solely a method of age determination nor a means of age classification of strata, nor is it a subset of geochronology. In Hedberg (1976) and Salvador (1994), the purpose of a chronostratigraphic classification is “to organize systematically the Earth’s sequence of rock strata into named units (chronostratigraphic units), corresponding to intervals of geologic time (geochronologic units), to serve as a basis for time-correlation and a reference system for recording events of geologic history” (Hedberg 1976:66). Included objectives are to determine local time relations (because this is where the gathering of evidence must begin) and to establish a Standard Global Chronostratigraphic Scale (for global correlation and communication). The determination of the rock record precedes its interpretation (by whatever means) as to the age of that record. The basic chronostratigraphic unit, the stage, therefore precedes the establishment of its geochronologic counterpart, the age, contrary to Walsh (1998, 2001, and references therein).

**CLASSICAL TIME SCALE (CTS; AUBRY 1995).** Time scale based on radioisotopic dating of the stratigraphic record chosen to characterize certain temporal intervals, such as the system, series, and stage.

**CONCURRENT-RANGE CHRON.** Following from the antecedent *concurrent-range zone*, this is a new biochronologic term based on the time of the concurrent, coincident, or overlapping parts of the range chrons of two specified taxa selected from among the total forms contained in a temporal array. This is comparable to the strict overlap biochron of Walsh (1998:161, 2000:771) when two taxa are specified.

**CONCURRENT-RANGE ZONE.** According to Salvador (1994:58) this is “the body of strata including the concurrent, coincident, or overlapping parts of the range zones of two specified taxa selected from among the total forms contained in a sequence of strata.” This is preferred over the

definition of Hedberg (1976:55–57) (“parts of the range-zones of two or more . . . taxons”) because it simplifies boundary definition and recognition. Still, these zones are not as useful in leading to biochronologic correlations as are others. The present definition is comparable to the strict overlap assemblage fossizone (or fossilzone) of Walsh (1998:161, 2000:770) when two taxa are considered.

**CORRELATION.** Stratigraphic correlation shows correspondence in character or stratigraphic position (Salvador 1994:15), but as modified from Aubry (1998:43) as “stratigraphic correlation,” it must mean temporal correlation as based on temporal analysis. Neither diachrony nor synchrony may be accepted on the basis of stratigraphic correlations alone but must be demonstrated on the basis of temporal analysis (Aubry 1995), and a dual terminology for stratigraphic and temporal terms must obtain.

**CYCLOSTRATIGRAPHY.** A discipline of stratigraphy wherein successive repetitions of sedimentary features are considered to be cyclical in nature. Some sedimentary cycles (i.e., varves) are interpreted as being annual features of climatic origin. Others are thought to reflect perturbations in orbital precession and obliquity caused by Earth’s behavior as it orbits the Sun, commonly known as Milankovitch cycles (Hilgen et al. 1997).

**FAD.** First appearance datum. This is a change “in the fossil record with extraordinary geographical limits” (Berggren and Van Couvering 1974:IX). As a chronostratigraphic concept, a FAD expresses an interpretation that the first stratigraphic appearance of a taxon is likely to have been synchronous over a specified geographic region (Woodburne 1996). The origin for a FAD (= appearance) was not constrained by Berggren and Van Couvering (1974, 1978), except that the dispersing taxon would have been newly evolved. For the paleobiotic event to be of “extraordinary geographical limits,” dispersal of an organism at a major scale clearly is the primary consideration, presumably from an indigenous source at some location. Aubry (1995:215) paraphrased this as the FAD being the “first (temporal; evolutionary) appearance datum.” Also, *LO* corresponds to *FAD* if the *LO* is of global significance (Aubry 1997:18, 22).

**FAUNA.** For paleontology, this is an assemblage of vertebrate fossils of similar taxonomic composition obtained from a small number of sites considered to have a limited temporal range. A fauna is commonly composed of a number of local faunas. See Tedford (1970). Depending on historical context and author intent, stratigraphic limits of a fauna may be supplied.

**FAUNULE.** Association of taxa interpreted directly or intentionally for its ecological significance. See Tedford (1970).

**FOD.** First occurrence datum. Aubry (1997:18–19) distinguishes FOD from *LO* and *FAD* as a diachronous *LO* and therefore not an isochronous *FAD*. The word *datum* in the name signifies the temporal connotation rather than the biostratigraphically descriptive *LO*. If a given *LO* can be demonstrated as temporally later than the time of the *FAD* of that taxon,

then it can be segregated from the list of LOs that contribute to the FAD and be designated as a FOD. The FAD is of global significance; the FOD may be regionally important. The FOD is comparable to the dispersal lag of Woodburne and Swisher (1995) if its age can be demonstrated.

**GEOCHRONOLOGY.** According to Hedberg (1976) and Salvador (1994), this is defined as “the science of dating and determining the time sequence of events in the history of the Earth” (Hedberg 1976:15). As expressed by Berggren and Van Couvering (1978:40), geochronology is “geologic time as perceived by the progress in one or another ordinal series of events,” with those events being parts of irreversible systems, such as organic evolution or radioisotopic decay. It is critically important that these ordinal systems “provide a theoretical basis outside of the preserved geologic record by which the nature and relation of the events in the progression can be recognized or predicted, and according to which missing parts of the record can be identified” (Berggren and Van Couvering 1978:40). Other methods useful to geochronology include paleomagnetic stratigraphy, isotope stratigraphy, and Milankovitch cyclostratigraphy. Geochronology is not merely geochronometry, by which numerical ages are applied to rocks or events.

**GEOMAGNETIC POLARITY TIME SCALE (GPTS).** A chronology based on counting reversals of Earth’s magnetic field (Bates and Jackson 1987:272).

**HO.** Highest stratigraphic occurrence (Aubry 1997:18–19). This is effectively similar to HSD. An HO may correspond to a LAD (Aubry 1997:22) if it is of effectively global significance. A series of diachronous HOs can become LODs if of regional significance. An HO also may have no temporal significance because of poor representation, scarcity, and truncation by an unconformity (Aubry 1997:22). See also Walsh (2000).

**HSD.** Highest stratigraphic occurrence of a taxon in a local section (Opdyke et al. 1977). A biostratigraphic term (Lindsay et al. 1987; Woodburne 1996); see *LSD*. Aubry (1997:18–22) prefers to use *HO* for (mostly) the same intent but to reserve the term *datum* for chronologic inference.

**INTEGRATED MAGNETOBIOCHRONOLOGIC SCALE (IMBS; Berggren et al. 1985a, 1985b, 1985c, 1995a).** A time scale consisting of a magnetostratigraphy, a numerical scale, and a magnetobiochronologic framework.

**INTERNATIONAL COMMISSION ON STRATIGRAPHY (ICS),** accepted as such by the International Union of Geological Sciences in 1986. The mandate of the ICS is to develop a standard global stratigraphic scale (Cowie et al. 1986).

**INTERNATIONAL UNION OF GEOLOGICAL SCIENCES.** The IUGS promotes and supports the study of geological problems of worldwide significance and facilitates international and interdisciplinary cooperation in the Earth sciences.

**INTERVAL CHRON.** Following from the terminology of the interval zone (Salvador 1994), this is the interval of time

defined on the earliest age of two successive biohorizons and is comparable to that of Walsh (1998) in representing the span of time between the first or last occurrence of one taxon and the first or last occurrence of another taxon. This is interpreted herein to mean that the boundaries of such a unit would be based on the ages of the LO and HO, respectively, of the taxa in question.

**INTERVAL ZONE.** According to Hedberg (1976:60) this is a biostratigraphic unit defined as the body of fossiliferous strata “between two distinctive biostratigraphic horizons.” Salvador (1994:123) defines this as a “biozone consisting of the body of fossiliferous strata between two specified biostratigraphic horizons (biohorizons).” This is interpreted herein to mean that the boundaries of such a unit would be based on the LOs, respectively, of the taxa in question. Although defining a boundary on an HO is theoretically possible, it generally has a greater potential for stratigraphic inconsistency than a LO (but see Cooper et al. 2001).

**LAD.** Last appearance datum; counterpart to a FAD. A LAD may be identical to the HO if the latter is of global significance (Aubry 1997:22).

**LINEAGE CHRON.** This is a new biochronologic unit. It is based on the corresponding biostratigraphic unit, the lineage zone (Salvador 1994). Thus a lineage chron is the interval of time defined on the earliest age of a taxon or part thereof in a specific evolutionary lineage and on the earliest age of its evolutionary successor. There is no counterpart in Walsh (1998).

**LINEAGE ZONE.** According to Hedberg (1976:58), a lineage zone comprises “the body of strata containing specimens representing a segment of an evolutionary . . . line or trend, defined above and below by changes in features of the line or trend.” In Salvador (1994:125) this is a “body of strata containing specimens representing a specific segment of an evolutionary lineage.” These criteria are interpreted herein to mean that the boundaries of such a unit would be based on the LOs, respectively, of the evolutionary first stratigraphic appearance of the taxon in question and the subsequent evolutionary first stratigraphic appearance of the derivative taxon of the lineage in question (see also NACSN 1983:862). Lineage zones “offers one of the best assurances of reliable time-correlation on a biostratigraphic basis” (Hedberg 1976:59).

**LOCAL FAUNA.** An aggregate of fossil vertebrate species that have a limited distribution in time from a number of closely grouped localities in a limited geographic area. See Tedford (1970). A local fauna could be based on taxa from a single locality.

**LOD.** Last occurrence datum. A series of regionally diachronous highest stratigraphic occurrences can form a number of LODs if they can be documented. See *FOD*.

**LO.** Lowest stratigraphic occurrence (Aubry 1995:17). This may be an LSD. It also may equate to an FAD (Aubry 1997:22) if it is of regional significance. Aubry (1995, 1997) differentiates *LO* as a stratigraphic (descriptive) first occur-

rence and, although *LSD* is equivalent in concept, reserves the term *datum* to signify a temporal connotation. Aubry (1977:18–19) distinguishes a LO from a FOD as well as an FAD. See also Walsh (2000).

**LSD.** Lowest stratigraphic datum (Opdyke et al. 1977:324). This is a biostratigraphic concept of the lowest known occurrence of a taxon in a local stratigraphic sequence (see also Lindsay et al. 1987; Lindsay and Tedford 1990:609; Woodburne 1996). The LO (Aubry 1997:18–22) is in part identical to the LSD.

**MAGNETOSTRATIGRAPHIC POLARITY UNITS.** Throughout the history of its development, workers applied a variety of names to parts of the Geomagnetic Polarity Time Scale, such as *epoch*, *event*, or *interval*. Recent codes or guides have stabilized the nomenclature of magnetic polarity units (e.g., Hedberg 1976; Salvador 1994). The following terminology implies that magnetostratigraphic and chronostratigraphic polarity units are analogous to those based on lithostratigraphy (tables 1.1 and 1.2). In practice, however, the original magnetostratigraphic chrons have no lithostratigraphic or chronostratigraphic base because the magnetic interval is inferred to be present in unseen sea floor lavas as sensed from magnetometers towed through the seas by ships.

**RECOMMENDED TERMINOLOGY FOR  
MAGNETOSTRATIGRAPHIC POLARITY UNITS  
(AFTER SALVADOR 1994:TABLE 2)**

MAGNETO- STRATIGRAPHIC POLARITY UNIT	CHRONOSTRATIGRAPHIC EQUIVALENT	GEOCHRONOLOGIC EQUIVALENT
Polarity superzone	Chronozone (or superchronozone)	Chron (or equivalent)
Polarity zone	Chronozone	Chron
Polarity subzone	Chronozone (or subchronozone)	Chron (or subchron)

**MAMMAL AGES.** Mammal ages make up the basic chronological system used to describe the age and succession of events in mammalian evolution in North America. Mammal ages (commonly known as North American land mammal ages [NALMAs]), are biochronologic units. The interval of time corresponding to each of these is recognized on the basis of mammalian evolution loosely (at least originally) tied to their stratal succession in sedimentary rocks (Wood et al. 1941; Woodburne 1987). In terms of the definitions presented here, mammal ages typically are assemblage chrons, although some have been interval chrons or lineage chrons (Archibald et al. 1987) with varying degrees of biostratigraphic documentation. To the extent that many mammal ages have been defined on the basis of immigrant taxa (Repenning 1967; Woodburne and Swisher 1995), they are effectively interval chrons whose the boundaries are based on first appearance datums. The biostratigraphic

counterpart of most mammal ages is the assemblage zone, “an assemblage zone based on a fossil fauna” (Salvador 1994:63).

**MEGANNUM (MA).** One million years in the radioisotopic time scale. For example, *10 Ma* refers to the 10-million-year level of the radioisotopic scale.

**M.Y. (OR m.y.).** A segment of geologic time 1 million years in duration, or the age of an event (e.g., 10 m.y. ago) without reference to a given point or set of points on the radioisotopic time scale.

**NEOGENE.** This follows Berggren et al. (1995b) to embrace the Miocene through Pleistocene series/epochs.

**NORTH AMERICAN LAND MAMMAL AGE (NALMA);** see *Mammal ages*.

**PALEOGENE.** This follows Berggren et al. (1995b) to embrace the Paleocene through Oligocene series/epochs.

**RANGE CHRON.** This is a biochronologic unit. Following from the language of the antecedent range zone (Salvador 1994), it represents the span of time defined on the age of selected element or elements of a biochronologic sequence. This is interpreted herein to mean that the boundaries of such a unit would be based on the ages of the LO and HO, respectively, of the taxon or taxa in question. The range chron of Walsh (1998) is a subset of the range chron as defined here.

**RANGE ZONE.** According to Salvador (1994:135) this is a biostratigraphic unit comprising the “body of strata representing the known stratigraphic and geographic range of occurrences of any selected element or elements of the assemblage of fossils present in a stratigraphic sequence.” That is interpreted herein to mean that the boundaries of such a unit would be based on the LO and HO, respectively, of the selected element or elements in question.

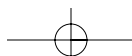
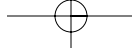
**TAXON-RANGE CHRON.** A taxon-range chron is a new biochronologic unit. Following from the language of the antecedent taxon-range zone (Salvador 1994), a taxon-range chron is defined on the known age range of a specified taxon.

**TAXON-RANGE ZONE.** According to Salvador (1994:140), this is a biostratigraphic unit comprising the “body of strata representing the known range of occurrence (stratigraphic and geographic) of specimens of a certain taxon (species, genus, family, etc.)” That is interpreted herein to mean that the boundaries of such a unit would be based on the LO and HO, respectively, of the taxon in question (see also NACSN 1983:862).

**REFERENCES**

- Archibald, J. D., P. D. Gingerich, E. H. Lindsay, W. A. Clemens, D. W. Krause, and K. D. Rose. 1987. First North American land mammal ages of the Cenozoic Era. In *Cenozoic mammals of North America: Geochronology and biostratigraphy*, ed. M. O. Woodburne. Berkeley: University of California Press, pp. 24–76.

- Aubry, M.-P. 1995. From chronology to stratigraphy: Interpreting the stratigraphic record. In *Geochronology, time scales and global stratigraphic correlation*, ed. W. A. Berggren, D. V. Kent, M.-P. Aubry, and J. Hardenbol. Tulsa: SEPM Special Publication 54, pp. 213–274.
- . 1997. Interpreting the (marine) stratigraphic record. In *Actes du Congrès BiochroM'97*, eds. J.-P. Aguilar, S. Legendre, and J. Michaux. Mémoires et Travaux E.P.H.E., Institut de Montpellier 21:15–32.
- . 1998. Stratigraphic (dis)continuity and temporal resolution of geological events in the Upper Paleocene–Lower Eocene deep sea record. In *Late Paleocene–Early Eocene climatic and biotic events in the marine and terrestrial records*, ed. M.-P. Aubry, S. G. Lucas, and W. A. Berggren. New York: Columbia University Press, pp. 37–66.
- Aubry, M.-P., W. A. Berggren, J. A. Van Couvering, and F. Steininger. 1999. Problems in chronostratigraphy: Stages, series, unit and boundary stratotypes, Global Stratotype Section and Point and tarnished golden spikes. *Earth Science Reviews* 46:99–148.
- Bates, R. L. and J. A. Jackson (eds.). 1987. *Glossary of geology*. Alexandria, VA: American Geological Institute.
- Berggren, W. A., F. J. Hilgen, C. G. Langereis, D. V. Kent, J. D. Obradovich, I. Raffi, M. Raymo, and N. J. Shackleton. 1995a. Late Neogene (Pliocene–Pleistocene) chronology: New perspectives in high resolution stratigraphy. *Geological Society of America Bulletin* 107:1272–1287.
- Berggren, W. A., D. V. Kent, J. J. Flynn, and J. A. Van Couvering. 1985a. Cenozoic geochronology. *Geological Society of America Bulletin* 96:1407–1418.
- . 1985b. Paleogene geochronology and chronostratigraphy. In *The chronology of the geologic record*, ed. N. J. Snelling. *Geological Society of London Memoirs* 10:141–195.
- . 1985c. Neogene geochronology and chronostratigraphy. In *The chronology of the geologic record*, ed. N. J. Snelling. *Geological Society of London Memoirs* 10:211–260.
- Berggren, W. A., Kent, D. V., Swisher, C. C. III, and Aubry, M.-P. 1995b. A revised Cenozoic geochronology and chronostratigraphy. In *Geochronology, time-scales and global stratigraphic correlations: A unified framework for an historical geology*, ed. W. A. Berggren, D. V. Kent, M.-P. Aubry, and J. Hardenbol. Tulsa: SEPM Special Publication 54, pp. 129–213.
- Berggren, W. A. and J. A. Van Couvering. 1974. The late Neogene: Biostratigraphy, geochronology and paleoclimatology of the last 15 million years in marine and continental sequences. *Palaeogeography, Palaeoecology, Palaeoclimatology* 16:1–216.
- . 1978. Biochronology. In *Contributions to the geologic time scale*, ed. G. V. Cohee, M. F. Glaessner, and H. D. Hedberg. Tulsa: American Association of Petroleum Geologists, Studies in Geology 6:39–55.
- Cooper, R. A., J. S. Crampton, J. I. Raine, F. M. Gradstein, H. E. G. Morgans, P. M. Sadler, C. P. Strong, D. Waghorn, and G. J. Wilson. 2001. Quantitative biostratigraphy of the Taranaki Basin, New Zealand: A deterministic and probabilistic approach. *American Association of Petroleum Geologists Bulletin* 85(5):1469–1498.
- Cowie, J. W., W. Ziegler, A. J. Boucot, M. G. Bassett, and J. Remane. 1986. Guidelines and Statutes of the International Commission on Stratigraphy (ICS). *Courier Forschungsinstitut Senckenberg* 83:1–14.
- Hedberg, H. C. (ed.). 1976. *International stratigraphic guide*. New York: Wiley.
- Hilgen, F. J., W. Krijgsman, C. G. Langereis, and L. Lourens. 1997. Breakthrough made in dating of the geologic record. *EOS, Transactions, American Geophysical Union* 78(28):285, 288–289.
- Lindsay, E. H., N. M. Johnson, N. D. Opdyke, and R. F. Butler. 1987. Mammalian chronology and the magnetic polarity time scale. In *Cenozoic mammals of North America; Geochronology and biostratigraphy*, ed. M. O. Woodburne. Berkeley: University of California Press, pp. 269–290.
- Lindsay, E. H. and R. H. Tedford. 1990. Development and application of land mammal ages in North America and Europe, a comparison. In *European Neogene mammal chronology*, ed. E. H. Lindsay, V. Fahlbusch, and P. Mein. New York: Plenum. NATO Advanced Science Institute Series 180:601–624.
- North American Commission on Stratigraphic Nomenclature. 1983. North American stratigraphic code. *American Association of Petroleum Geologists Bulletin* 67(5):841–875.
- Olson, E. C. 1952. The evolution of a Permian vertebrate chronofauna. *Evolution* 6:181–196.
- Opdyke, N. D., E. H. Lindsay, N. M. Johnson, and T. Downs. 1977. The paleomagnetism and magnetic polarity stratigraphy of the mammal-bearing sections of Anza Borrego State Park, California. *Quaternary Research* 7:316–329.
- Repenning, C. A. 1967. Palearctic–Nearctic mammalian dispersal in the late Cenozoic. In *The Bering land bridge*, ed. D. M. Hopkins. Stanford, CA: Stanford University Press, pp. 288–311.
- Salvador, A. (ed.) 1994. *International stratigraphic guide*. Boulder, CO: Geological Society of America.
- Tedford, R. H. 1970. Principles and practices of mammalian geochronology in North America. *Proceedings, North American Paleontological Convention* Pt. F, pp. 666–703.
- Walsh, S. L. 1998. Fossil datum and paleobiological event terms, paleontostratigraphy, chronostratigraphy, and the definition of land mammal “age” boundaries. *Journal of Vertebrate Paleontology* 18(1):150–179.
- . 2000. Eubiostratigraphic units, quasibiostratigraphic units, and “assemblage zones.” *Journal of Vertebrate Paleontology* 20(4):761–775.
- . 2001. Notes on geochronologic and chronostratigraphic units. *Bulletin of the Geological Society of America* 113(6):704–713.
- Williams, H. S. 1901. The discrimination of time-values in geology. *Journal of Geology* 9:570–585.
- Wood, H. E. II, R. W. Chaney, J. Clark, E. H. Colbert, G. L. Jepsen, J. B. Reeside Jr., and C. Stock. 1941. Nomenclature and correlation of the North American continental Tertiary. *Bulletin of the Geological Society of America* 52:1–48.
- Woodburne, M. O. (ed.). 1987. *Cenozoic mammals of North America: Geochronology and biostratigraphy*. Berkeley: University of California Press.
- . 1996. Precision and resolution in mammalian chronostratigraphy: Principles, practices, examples. *Journal of Vertebrate Paleontology* 16(3):531–555.
- Woodburne, M. O. and C. C. Swisher III. 1995. Land mammal high resolution geochronology, intercontinental overland dispersals, sea-level, climate, and vicariance. In *Geochronology, time-scales and global stratigraphic correlations: A unified framework for an historical geology*, ed. W. A. Berggren, D. V. Kent, M.-P. Aubry, and J. Hardenbol. Tulsa: SEPM, Special Publication 54:335–364.





---

## Introduction

Michael O. Woodburne

### **PERSPECTIVE**

---

The chronologic framework of the present book remains the North American mammal age concept articulated by Wood et al. (1941) and Savage (1951) and displayed in a great variety of sources, including Woodburne (1987), hereafter identified as the 1987 volume. It is taken as given that practitioners of stratigraphic paleontology or stratigraphic paleobiology recognize and embrace the principle of paleontological correlation (Smith 1815, 1817) and of Steno's (1669) principles of superposition, original horizontality, and original continuity of strata so that the rock record can be used to order the succession of mammalian (and other) taxa and serve as an empirical basis for recording that succession irrespective of theories of evolution or philosophies of systematic analysis. Even though mammal ages are nominally biochrons (Williams 1901:579; intervals of geologic time based on the "duration of organic characters"), their succession (Wood et al. 1941) was framed by the stratigraphic sequences in which they were found (Tedford 1970; Emry 1973 and references cited therein.). Thus the succession of mammal ages depended on the lithostratigraphic framework for their documentation. Similarly, it follows that refinements in the chronology of mammal ages also depend on increasingly refined documentation of the stratigraphic and chronologic framework in which they occur.

Chapters 1 and 2 of the 1987 volume summarized the variety of biostratigraphic and chronostratigraphic proposals developed to describe mammalian faunal succession and correlation up to that time. Chapter 3 of the 1987 volume nominated a succession of new biostratigraphic

zones for faunas of nominal Paleocene age, and other 1987 chapters evaluated the mammal succession in early and late (or finer-scale) subdivisions of the traditional mammal ages. These frameworks are essentially followed herein. Woodburne and Swisher (1995) gave an update of the mammal age chronology in North America, with emphasis on evidence for the age of the immigrations that define a majority of the mammal ages and the extent to which these corresponded to major episodes of global sea level lowering. Alroy (1992, 1994, 1998a, 1998b) presented a subdivision of the mammalian faunal record in North America based on quantitative analysis and indicated that whereas immigration is a rapid process, the observed sampling-influenced diachroneity is far too great to allow favoring immigrant first occurrences as time indicators. In Alroy's view, only quantitative analyses of entire faunas have any chance of recovering robust biochronological patterns. However, the only way in which quantitative or any other analyses can be improved is by developing new chronologically significant information with which to assess the age of taxa having a taxonomic precision that is underwritten by the experts directly familiar with the fossils they represent.

### **HIGH-RESOLUTION STRATIGRAPHY AND BIOCHRONOLOGY**

---

The present work continues with the integration of stratigraphic and other temporally significant data with the mammal record in its primary physical context so as to

provide an empirical basis on which the tempo and mode of mammalian evolution can be measured. An underlying concern is the degree to which the patterns of mammalian succession are replicable geographically and found to be chronologically consistent, whether these patterns are described as various kinds of biochrons (mammal ages), subdivisions of them, or biostratigraphic or chronostratigraphic zones.

A goal is the development of a high-resolution chronologic network that, to paraphrase Woodburne (1996), involves the development of a detailed stratigraphic framework for the fossil data, whether they are portrayed in a biostratigraphic array or not, determining an approximate age for the fossiliferous levels with respect to radioisotopic calibration or with respect to a magnetozone whose age limits are confidently known. The independent relative chronologic framework of magnetostratigraphy (and assignment of numerical ages to polarity reversal boundaries by various means; Cande and Kent 1992, 1995; Berggren et al. 1995a, 1995b) allows calibration of the fossil level and temporal correlation with any other similarly placed fossil level in another stratigraphic section. See chapter 1 for further consideration of this topic.

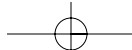
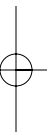
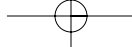
This is not the end of the operation, however. In recent decades, increasing emphasis has been placed on addressing the fidelity of the stratigraphic record through both statistical aspects (Strauss and Sadler 1989; Marshall 1990) and graphic methods (Aubry 1995, 1998; Mann and Lane 1995). Such operations may become increasingly meaningful in recognition of the fact that the  $\pm$  factor as applied for the  $^{40}\text{Ar}/^{39}\text{Ar}$  radioisotopic dating method can produce ancient ages with very small  $\pm$  dimensions (e.g.,  $249.9 \pm 0.1$  Ma; Siberian Traps flood basalts; Renne et al. 1998:130). This can lead to the notion that  $^{40}\text{Ar}/^{39}\text{Ar}$  ages are usually better than those derived from, say, the K–Ar method. But as discussed further in chapter 1, this notion can be somewhat misleading. In any case, there are numerous examples wherein mammalian stratigraphers attempt to use accumulation rate reconstructions (based on extrapolations from or interpolations between radioisotopic or magnetic polarity ages) to estimate the age of biostratigraphic or biochronologic units (Woodburne et al. 1990:474), but almost none use the kinds of procedures outlined in Aubry (1995, 1998) to test rigorously for hidden unconformities or other condensations of stratigraphic section, even though it is a given that any sharp geologic boundary (including a magnetic polarity reversal) may reflect an unconformity in the record (Sadler 1999). In fact, the frequent mismatches in the continental magnetostratigraphic record relative to the Geomagnetic Polarity Time Scale must result at least as much as from

the effect of apparently unappreciated unconformities in the rock record as from imperfections, overprints, or technical errors in the magnetostratigraphic analysis. Before asserting diachrony in the lowest stratigraphic datum of fossil mammals when considered regionally (Alroy 1998), it is necessary to rule out the effect of imperfections in the stratigraphic record. Finally, in order to be precise, boundaries must be defined and the proposed interval characterized (Woodburne, 1977, 1987, 1996), with single-taxon definitions being preferred over those based on multiple taxa because they are less ambiguous.

## REFERENCES

- Alroy, J. 1992. Conjunction among taxonomic distributions and the Miocene mammalian biochronology of the Great Plains. *Paleobiology* 18(3):326–343.
- . 1994. Appearance event ordination: A new biochronologic method. *Paleobiology* 20(2):191–207.
- . 1998a. Diachrony of mammalian appearance events: Implications for biochronology. *Geology* 26(1):23–26.
- . 1998b. Diachrony of mammalian appearance events: Implications for biochronology—Reply. *Geology* 26:956–958.
- Aubry, M.-P. 1995. From chronology to stratigraphy: Interpreting the stratigraphic record. In *Geochronology, time scales and global stratigraphic correlation*, ed. W. A. Berggren, D. V. Kent, M.-P. Aubry, and J. Hardenbol. Tulsa: SEPM Special Publication 54, pp. 213–274.
- . 1998. Stratigraphic (dis)continuity and temporal resolution of geological events in the Upper Paleocene–Lower Eocene deep sea record. In *Late Paleocene–Early Eocene climatic and biotic events in the marine and terrestrial records*, ed. M.-P. Aubry, S. G. Lucas, and W. A. Berggren. New York: Columbia University Press, pp. 37–66.
- Berggren, W. A., F. J. Hilgen, C. G. Langereis, D. V. Kent, J. D. Obradovich, I. Raffi, M. Raymo, and N. J. Shackleton. 1995a. Late Neogene (Pliocene–Pleistocene) chronology: New perspectives in high resolution stratigraphy. *Geological Society of America Bulletin* 107:1272–1287.
- Berggren, W. A., D. V. Kent, C. C. Swisher III, and M.-P. Aubry. 1995b. A revised Cenozoic geochronology and chronostratigraphy. In *Geochronology, time-scales and global stratigraphic correlations: A unified framework for an historical geology*, ed. W. A. Berggren, D. V. Kent, M.-P. Aubry, and J. Hardenbol. Tulsa: SEPM Special Publication 54, pp. 129–213.
- Cande, S. C. and D. V. Kent. 1992. A new geomagnetic polarity time-scale for the Late Cretaceous and Cenozoic. *Journal of Geophysical Research* 97:13,917–13,951.
- . 1995. Revised calibration of the geomagnetic polarity time scale for the Late Cretaceous and Cenozoic. *Journal of Geophysical Research* 100:6093–6095.
- Emry, R. J. 1973. Stratigraphy and preliminary biostratigraphy of the Flagstaff Rim area, Natrona County, Wyoming. *Smithsonian Contributions to Paleobiology* 25:1–20.
- Mann, K. O. and H. R. Lane (eds.). 1995. *Graphic correlation*. Tulsa: SEPM Special Publication 53.

- Marshall, C. R. 1990. Confidence intervals on stratigraphic ranges. *Paleobiology* 16:1–10.
- Renne, P. R., C. C. Swisher III, A. L. Deino, D. B. Karner, T. L. Owens, and D. J. K. DePaolo. 1998. Intercalibration of standards, absolute ages and uncertainties in  $^{40}\text{Ar}/^{39}\text{Ar}$  dating. *Chemical Geology* 145:117–152.
- Sadler, P. M. 1999. The influence of hiatuses on sediment accumulation rates. *GeoResearch Forum* 5:15–40.
- Savage, D. E. 1951. Late Cenozoic vertebrates of the San Francisco Bay region. *University of California Publications in Geological Sciences* 28:215–314.
- Smith, W. 1815. *Memoir to the map and delineation of the strata of England and Wales with a plat of Scotland*. London: Cary.
- Smith, W. 1817. *Stratigraphic system of organized fossils with reference to the specimens of the original collection in the British Museum explaining their state of preservation and their use in identifying the British strata*. London: E. Williams.
- Steno, N. 1669. *De solido intra solidum naturaliter contento dissertationis prodromus*. Florence.
- Strauss, D. and P. M. Sadler. 1989. Classical confidence intervals and Bayesian probability estimates for ends of local taxon ranges. *Mathematical Geology* 21:411–427.
- Tedford, R. H. 1970. Principles and practices of mammalian geochronology in North America. *Proceedings of the North American Paleontological Convention* Pt. F, pp. 666–703.
- Williams, H. S. 1901. The discrimination of time-values in geology. *Journal of Geology* 9:570–585.
- Wood, H. E. II, R. W. Chaney, J. Clark, E. H. Colbert, G. L. Jepsen, J. B. Reeside Jr., and C. Stock. 1941. Nomenclature and correlation of the North American continental Tertiary. *Bulletin of the Geological Society of America* 52:1–48.
- Woodburne, M. O. 1977. Definition and characterization in mammalian chronostratigraphy. *Journal of Paleontology* 51(2):220–234.
- (ed.). 1987. *Cenozoic mammals of North America: Geochronology and biostratigraphy*. Berkeley: University of California Press.
- . 1996. Precision and resolution in mammalian chronostratigraphy: Principles, practices, examples. *Journal of Vertebrate Paleontology* 16(3):531–555.
- Woodburne, M. O. and C. C. Swisher III. 1995. Land mammal high resolution geochronology, intercontinental overland dispersals, sea-level, climate, and vicariance. In *Geochronology, time-scales and global stratigraphic correlations: A unified framework for an historical geology*, ed. W. A. Berggren, D. V. Kent, M.-P. Aubry, and J. Hardenbol. Tulsa: SEPM, Special Publication 54:335–364.
- Woodburne, M. O., R. H. Tedford, and C. C. Swisher III. 1990. Lithostratigraphy, biostratigraphy, and geochronology of the Barstow Formation, Mojave Desert, southern California. *Geological Society of America Bulletin* 102:459–477.



# 1

---

## Principles and Procedures

Michael O. Woodburne

**A** **DISCUSSION OF THE PRINCIPLES** and procedures in methodology and the goal of producing a time scale based on the evolution of fossil mammals that contains neither gaps nor overlaps is as pertinent now as it was in 1987 or, indeed, in 1941 (Wood et al. 1941). Whether or not it is formally identified as biostratigraphy, students of mammalian chronology in North America have continually worked to improve the stratigraphic framework associated with fossil mammals and to integrate it with other chronologic information. Although still biochrons, mammal ages and subdivisions have become stratigraphically assisted (stratigraphically characterized but not defined) to varying degrees since 1941, and this trend continues here. There still are only a few instances in which sufficient stratigraphic information has been added to the fossil mammal biochronologic concept to support the development of a chronostratigraphic stage, and thus a true geochronologic age, hence the common convention of categorizing the biochrons as mammal ages. The main purpose of this chapter is to review the traditional stratigraphic procedures and some innovations designed to improve the development of a correlation network for fossil mammals that is empirically based and noncircular in reasoning. A thesis developed here is that not only is there a distinct progression from biostratigraphy, through biochronology, to chronostratigraphy, and then to geochronology, but that it is appropriate to use a formalized set of biochronologic units as part of that process. In at least one example cited in this chapter, almost the entire process has been achieved for early Paleocene strata of the Hanna Basin, Wyoming, even though the

final procedural documentation of a chronostratigraphic unit (statement of intent, selection of stratotypes, reference sections) has not been completed. In a much larger set of examples (described elsewhere in this book), the mammal age data set is becoming increasingly documented in detail with respect to stratigraphy and with respect to radioisotopic, paleomagnetic, and stable isotope chronology. It is therefore appropriate to review the fundamentals of stratigraphic classification and correlation here.

---

### THE GEOCHRONOLOGIC FRAMEWORK

This is the framework within which the geologist or paleobiologist understands not only the passage of geologic time but also the age and interrelation of past events important for study. In the present case, interest is focused on mammalian evolution and the means by which the results therefrom can be used to develop a framework of data that lead to a chronologic system by which that evolution can be perceived and documented: an increasingly high-resolution stratigraphy in which all available temporally significant data and applications are integrated. The progression toward this goal begins with a consideration of a chronology derived from a physical stratigraphic framework and turns to its calibration, estimation of completeness or fidelity, operations in correlation, the role of biochronology, and the relationship of these factors to mammal ages.

## STRATIGRAPHIC CLASSIFICATION

A major task of a mammalian stratigrapher is to devise and work within a framework of data that leads to the establishment of a succession of temporal intervals that account for all of geologic time, with no overlaps or hiatuses. The following describes the classification of units

**TABLE 1.1** Conventional Hierarchy of Formal Chronostratigraphic and Geochronologic Terms

CHRONOSTRATIGRAPHIC	GEOCHRONOLOGIC
Eonothem	Eon
Erathem	Era
System <sup>a</sup>	Period <sup>a</sup>
Series <sup>a</sup>	Epoch <sup>a</sup>
Stage <sup>b</sup>	Age
Substage	Subage or age
Chronozone	Chron

<sup>a</sup>If additional ranks are needed, the prefixes *sub-* and *super-* may be used with these terms.

<sup>b</sup>Several adjacent stages may be grouped into a superstage.

Hedberg (1976:69–70) considers *chronozone* and *chron* to be members of the formal hierarchy. Salvador (1994:83–84) treats them as formal but nonhierarchical units.

and concepts in the conventional stratigraphic hierarchy. Table 1.1 shows the formal chronostratigraphic and geochronologic terms used in modern stratigraphic guides and codes, and table 1.2 summarizes the classification of units and concepts important to biostratigraphy, chronostratigraphy, and geochronology. The operation of working within this hierarchy to develop a temporal correlation is taken up later in this chapter. With respect to table 1.1, the System–Period pair was the focus of geologists contemporaneous with William Smith, but in subsequent years, increasing attention has been paid to smaller-scale increments of the hierarchy as attempts were made to more finely subdivide (and recognize) intervals of geologic time.

Stratigraphers are increasingly concerned with identifying and using stratigraphically and temporally shorter intervals and use an increasingly sophisticated and refined set of analytical and procedural tools to further those goals. But the first step is to clearly separate physical and tangible units from purely inferential and intangible ones. At one extreme is the lithostratigraphic base on which all other stratigraphic endeavors must be founded. At the other extreme are geochronologic (geologic time) units that are explicitly intangible and inferential. To varying degrees, biostratigraphic units (based

**TABLE 1.2** Independence of Lithostratigraphic Units From and Potential Relationships Between Biostratigraphic, Chronostratigraphic, and Geochronologic Units

LITHOSTRATIGRAPHIC <sup>A</sup>	BIOSTRATIGRAPHIC <sup>B</sup>	CHRONOSTRATIGRAPHIC <sup>C</sup>	GEOCHRONOLOGIC <sup>D</sup>
Formation	Various kinds of biozones	Chronozone	Chron
Member			
Bed			
Horizon			

<sup>A</sup>Physical unit; descriptive, based on lithological characteristics without regard to age of deposition. A formation may be lithologically heterogeneous or homogeneous. A member usually is lithologically more homogeneous and may be interpreted as lithogenesis. A bed commonly is of limited thickness and is at least as homogeneous lithologically as a member. A horizon is of very limited (conceptually zero) thickness but is a traceable marker.

<sup>B</sup>Fundamentally a physical unit, descriptive of the occurrence of fossils in their stratigraphic context. Procedurally independent of other stratigraphic units, biostratigraphic units can be developed for their biochronological significance and ultimately transformed into the paleontological basis for chronostratigraphic units (see text). The biozones most useful in chronostratigraphy are the taxon-range zones and lineage zones that describe the stratigraphic range of a single taxon without regard to sampling factors (e.g., abundance). The interpretive aspect is the subjective identification that the specimens on which the zone is based pertain to a given paleospecies. Thus if Lineage zone  $\alpha$  is followed stratigraphically by Lineage zone  $\theta$ , the lower boundary of  $\theta$  may depend on an arbitrary decision on the part of the stratigrapher. Thus to some extent the stratigraphic range of each biozone has an interpretive aspect.

<sup>C</sup>This is a physical unit in that it is the rock deposited during an interval of geologic time. It is conceptual in that the means by which the unit is recognized (most commonly fossils) are presumed to have a temporal component that is unique. Once a chronostratigraphic (time–rock) unit is created, the corresponding geochronologic (geologic time) unit of equal rank is thereby defined (Table 1.1). The chronostratigraphic unit (e.g., Ypresian Stage) is defined in a given type section or reference sections, and stratigraphic sequences in other areas are referred to this stage based on having sufficient defining or characteristic criteria (usually fossils) to warrant such a correlation. Chronostratigraphic units are the fundamental means for building a time–rock record that accounts for all of geologic time that has neither overlaps nor hiatuses. In contrast to geochronologic units, chronostratigraphic units are limited by the rock record. The chronozone is the basal element of the hierarchy (Table 1.1).

<sup>D</sup>This is a conceptual and intangible unit that stands for an interval of geologic time. It is not a stratigraphic unit, even though it may correspond to the time span of a stratigraphic unit. Thus one may speak of events that transpired during the Ypresian age without reference to a specific section of strata. The chron is the basal element of the hierarchy (Table 1.1).

on the physical disposition of fossils in the rock) and chronostratigraphic units (sections of rock that document intervals of geologic time) are both physical (tangible) and inferential (intangible), as summarized in table 1.2 and as discussed more fully later in this chapter. In brief, lithostratigraphic units provide the objective physical framework for geologic data; biostratigraphic units provide the objective physical framework for paleontologic data; chronostratigraphic units give a physical, stratigraphic record of the passage of time, drawn in large part from biostratigraphic information; and geochronologic units are intangible representations of the intervals of time contained in chronostratigraphic units.

**Lithostratigraphic Units** With regard to lithostratigraphic units, present North American and international codes and guides are consistent in separating the concepts and operations of lithostratigraphy as distinct from those dealing directly, or potentially, with biostratigraphy, chronostratigraphy, or geochronology. This stems from the conviction that the basic physical and descriptive framework for historical geology should be separate from any interpretive concepts or operations. Schenk and Muller (1941) clearly articulated this principle.

“Lithostratigraphic units are bodies of rock, bedded or unbedded, that are defined and characterized on the basis of their observable lithologic properties” (Salvador 1994:31). The objectively observed lithologic criteria are paramount in establishing a lithostratigraphic unit, regardless of age. Thus whereas fossils can be recognized as an identifying component (e.g., a *coquina*), they are treated as lithologic properties similar to kinds of rocks, minerals, and the like.

A primary purpose of lithostratigraphic units is to demonstrate a physical framework at a level pertinent to the study at hand, not always necessitating the construction of a geologic map. The typically mappable unit is the formation (table 1.2), but other and generally thinner but not necessarily areally less extensive units may be used. Examples of the latter include air-fall or ash-flow tuffs, debris flows, or other (usually thin, measured in meters or less) beds of distinctive lithology relative to those above or below. Whereas formations or other units may be homogenous lithologically, others may be differentiated by being lithologically heterogeneous in contrast to those above and below. Also, it is convenient if the boundaries of the lithologic unit are sharp and unambiguously detected, but in other cases boundaries may be gradational. As Schenk and Muller (1941:1424) point out, boundaries of lithologic units commonly are chosen at unconformities, across which trenchant changes in lithology may be

observed. These authors further assert that this is in distinct contrast to the goals of time stratigraphy, in which it is desirable to have a setting in which deposition was effectively continuous, especially at the boundaries between the units. For lithostratigraphic units, the basic issue is developing a physical stratigraphic framework that is empirically constructed and reliably replicable in the district under study. Here, and for other units, base defines boundary. Hedberg (1976) and Salvador (1994) summarize the need to specify stratotypes or type localities of lithostratigraphic units.

**Biostratigraphic Units** As given in Salvador (1994:53), “biostratigraphic units (biozones) are bodies of rock strata that are defined or characterized on the basis of their contained fossils.” As summarized in tables 1.2 and 1.3, biostratigraphic units are material, physical units (also Walsh 1998:163). Determining the base of the unit in places other than the stratotype is only as valid as its definition. As discussed later in this chapter (see “Definition and Characterization”), the best definition is based on the lowest stratigraphic occurrence of a single taxon. Whereas some biozones illustrate variations in abundance of paleospecies, others clearly are intended for use in correlation, including the development of chronostratigraphic units. Some biozones are based on the record of single taxa, others on the occurrence of several.

The reason for establishing biostratigraphic units is to develop an empirical record of taxonomic occurrence in the rock record. Whether codified as a given category (table 1.3) or not, the pattern of biostratigraphic information that may be constructed forms an empirical framework parallel in concept to the development of lithostratigraphic information. Both frameworks are conceptually independent of other kinds of considerations, such as time or ecology, and on this basis can form the legitimate foundation from which those other considerations may be developed. Regarding biostratigraphy, the species from which biozones are described have a distinct, limited sojourn in geologic time. Once described and found to be replicated geographically and consistently with respect to geologically isochronous markers, biostratigraphic data can be interpreted for their temporal significance and form the basis for defining and characterizing chronostratigraphic units (table 1.2). Thus biozones are basically descriptive units but also have the potential for temporal interpretation. This aspect of biostratigraphy is taken up later in this chapter (“The Role of Biochronology”).

Salvador (1994:57–64) summarizes the various kinds of biozones. They are categorized as range zone, interval

TABLE 1.3 Biostratigraphic and Biochronologic Categories

AFTER SALVADOR (1994)		AFTER WALSH (1998, 2000)		THIS BOOK
BIOSTRATIGRAPHIC CATEGORIES	(EU)BIOSTRATIGRAPHIC CATEGORIES	BIOCHRONOLOGIC CATEGORIES		
Biostratigraphic units are empirical units based on the stratigraphic disposition of fossils. The basic unit is the biozone. <sup>a</sup>	As in Biostratigraphic categories.	Biochronologic units are interpretive units; intervals of time as represented by fossils. The basic unit is the biochron (Williams 1991).	Biostratigraphic categories follow Salvador (1994). Biochronologic categories are interpretive units based on the age of the equivalent biostratigraphic unit of Salvador (1994).	
Range zone: The body of strata containing the total known stratigraphic and geographic range of any selected elements of the assemblage of fossils in a stratigraphic sequence.	No comparably general unit in Walsh (1998, 2000).		The basic unit is the biochron. General correspondence to certain units of Walsh (1998, 2000) is indicated where appropriate.	
a. Taxon-range zone: The body of strata containing the total known stratigraphic and geographic range of specimens of a particular taxon (species, genus, family, etc.).	Range zone: The set of strata between the LSD and HSD of a single taxon. <sup>b</sup>	Range chron: The span of time between the FHA and LHA of a single taxon. <sup>c</sup>	Range chron: Interval of time defined on the age of selected element or elements of a biochronologic sequence, in part inclusive of range chron of Walsh (1998).	Taxon-range chron: Interval of time defined on the known age range of a specified taxon.
b. Concurrent-range zone: The body of strata including coincident, or overlapping, parts of the range zones of two taxa selected from those contained in a sequence of strata.	Strict overlap assemblage fossilzone: Where $N = 2$ . The more general case is the thickness of strata between the lowest co-occurrence and highest co-occurrence of every one of a specified set of two or more taxa.	Strict overlap assemblage biochron: The span of time between the FHAs and LHAs of (in the restricted context) two specified taxa.	Concurrent-range chron: Interval of time defined by the concurrent, coincident, or overlapping parts of the range of two specified taxa selected from the total forms contained in a temporal array.	
Interval zone: The body of fossiliferous strata between two biohorizons. The base is set by the lower biohorizon, the top by the upper.	Interval zone: The set of strata between the LSD or HSD of one taxon and the LSD or HSD of another taxon. <sup>d</sup>	Interval chron: The span of time between the FHA or LHA of one taxon and the FHA or LHA of another taxon.	Interval chron: Interval of time defined on the earliest age of two successive biohorizons.	
Lineage zone: The body of strata containing specimens representing a specific segment of an evolutionary lineage. <sup>e</sup>	No specific category in Walsh (1998, 2000). <sup>f</sup>		Lineage chron: Interval of time defined on the earliest age of a taxon or part thereof in a specific evolutionary lineage and on the earliest age of its evolutionary successor.	



**Assemblage zone:** The body of strata containing a distinctive assemblage of three or more fossil taxa that, taken together, distinguishes it in biostratigraphic character from adjacent strata.

**Assemblage fossil zone:** In a given superposed section, a thickness of strata based on the partial or combined range fossil zones of one, some, or all the taxa present in a set of two or more taxa.

**Assemblage biochron:** In a given geographic area, a span of time based on the partial or combined range biochrons of one, some, or all the taxa present in a single set of two or more taxa.

**Disjunctive assemblage fossil zone (the thickness of strata between the lowest LSD and the highest HSD of whichever taxa are present out of a specified set of two or more taxa) and Minimal overlap assemblage fossil zone (the thickness of strata between the lowest and highest co-occurrence of any combination of  $n$  taxa in a specified set of at least  $N + 1$  taxa where  $N \geq 2$ ) and**

**Disjunctive biochron:** The span of time between whichever of the FHAs of a specified set of two or more taxa is the oldest and whichever of the LHAs is the youngest.

**Minimal overlap biochron:** The span of time between whichever of the FHAs of a specified set of at least  $N + 1$  taxa is the  $n$ th oldest and whichever of the LHAs is the  $n$ th youngest (where  $N \leq 2$ ).

**Strict overlap assemblage fossil zone (the thickness of strata between the lowest and highest co-occurrence of every one of a specified set of two or more taxa.**

**Strict overlap assemblage biochron:** The span of time between whichever of the FHAs of a specified set of two or more taxa is the youngest and whichever of the LHAs is the oldest.

**Abundance zone:** A body of strata in which the abundance of a particular taxon is significantly greater than in the adjacent parts of the section regardless of either association or range.

No specific category in Walsh (1998, 2000).  
No closely applicable chronologic significance.

No specific category. No closely applicable chronologic significance.

FHA; HSD, highest stratigraphic datum; LHA; LSD, lowest stratigraphic datum.

<sup>a</sup>Walsh (1998) uses the term *fossil zone* for an equivalent concept, modified to *fossil zone* in Walsh (2000).

<sup>b</sup>Range zones and interval zones are each divided into a range epizone, range entozone, lineage epizone, and lineage entozone. Epizones are based on LSDk and HSDk (empirical zones based on the LSD and HSD as originally defined by Opdyke et al. (1977). Entozones are based on LSDa and HSDa, the "actual" occurrence in the rock record, but this is a theoretical concept in that the fossils involved have not yet been found. Even though they are used in a strat context, LSDa and HSDa are comparable to interpretive, biochronologic units and not recognized here as a separate biostratigraphic category. *Range zone* of Walsh (1998) is most similar to *taxon-range zone* of Salvador (1994).

<sup>c</sup>Range chron are based on FHA and LHA (first and last historical occurrence as a temporal interpretation). If a taxon is thought to have been present in a given area (FHA), it presumably would leave a potential rock record (LSDa) that, when demonstrated stratigraphically, would become an LSDk. Regardless of being oriented toward a potential rock record rather than a potential temporal record, an LSDk is interpretive to the same extent as is an FHA. Both are interpretive units. Thus entozones are not proper biostratigraphic units and are not used here. *Range chron* of Walsh (1998) appears most comparable to *taxon-range chron* as used here.

<sup>d</sup>Walsh (1998) also nominates assemblage interval fossil zones and biochrons (renamed in 2000 as multiple-taxon interval fossil zones and biochrons). These are not discussed further here.

<sup>e</sup>A lineage zone has strong temporal significance and approaches a chronozone (basic unit of chronostratigraphy). Once defined, however, a chronozone contains all strata that can be shown to correspond to the specified interval of time, regardless of fossil content.

<sup>f</sup>This apparently reflects Walsh's (1988) view that procedures leading to the development of chronostratigraphic units via the usual methods of performing a biochronologic correlation based on biostratigraphic data are irrelevant to finding geochronologic events that can be formalized as a Global Boundary Stratotype Section and Point and then used to define the boundaries of chronostratigraphic units.

zone, lineage zone, assemblage zone, and abundance zone (table 1.3). Table 1.3 also compares biostratigraphic categories of Salvador (1994) with those of Walsh (1998) and concepts used herein. Either taxon-range zones and concurrent-range zones represent the total stratigraphic and geographic range of taxa. The taxon-range zone (range zone of Walsh 1998) is based on the range of a single taxon, whether of specific rank or greater. Its boundaries therefore are defined on the presence of the taxon in question. Thus the zone begins and ends stratigraphically with the known range of the specified taxon. The concurrent-range zone (strict overlap assemblage fossilzone of Walsh 1998) is similar, except that its extent is defined on the shared ranges of two taxa, although other taxa can help characterize the zone. As Salvador (1994:58) notes, a succession of concurrent-range zones can have gaps or overlaps between them.

An interval zone is a unit of fossiliferous rock having boundaries specified by two bounding biohorizons, although the fossil content of the zone within the interval itself is not specified. The interval zone of Walsh (1998) is comparable. This kind of zone appears to be most useful in analyzing cores of subsurface drilling but also has been used in mammalian biochronology (Archibald et al. 1987). This exemplifies the trend reflected in this book whereby mammal paleontologists strive to increase the role of biostratigraphy as applied to rock sequences bearing fossil mammals.

Lineage zones may be chronostratigraphic in character. These comprise a body of strata having specimens of a specific part of an evolutionary lineage, which could include the entire range of a given species. In that evolutionary innovations are temporally unique, this type of zone approaches a chronozone, the basic hierarchical unit of chronostratigraphy (table 1.1). To the extent that limits between species or other parts of the lineage must be interpreted by a paleontologist, there may be an intangible aspect to these zones that could have a temporal ramification (Woodburne 1987b, 1996a). A lineage zone still is empirical in that its presence is recognized solely on the stratigraphic occurrence of the taxon in question and has the additional provision that the base of the lineage defines the base of the zone, and its top is defined by the base of the descendant taxon. A chronozone based on a lineage zone theoretically would have boundaries with neither gaps nor overlaps. Walsh (1998) has no counterpart to the lineage zone.

An assemblage zone is characterized by the co-occurrence of three or more taxa that together distinguish the stratigraphic interval in which they occur from those above or below. In addition to usually being limited to

specific areas or regions, the boundaries of assemblage zones are imprecise. In that the zone is based on the ranges of three or more taxa, it is possible that none of them will be necessarily exclusive to the zone. If two or more taxa share the same lowest stratigraphic occurrence (LO), it is likely that this results from extrinsic factors (such as an underlying unconformity) and should be viewed with caution.

In part to address the vagaries of boundary definition for assemblage zones (or assemblage fossilzones), Walsh (1998:160L, 2000:770) proposes three different kinds—disjunctive, minimal overlap, or strict overlap assemblage fossilzones—and proposes to restrict the concept to a the lowest and highest occurrences of a single specified set of two or more taxa. This initiative moves well beyond the apparent consensus (at least among Hedberg 1976; Salvador 1994; NACSN 1983) that assemblage zones are generalized concepts for which boundary precision is unwarranted or at least not relevant. These three works also state or imply that range zones and interval zones are capable of precise boundary definition and also by implication or statement are most relevant as a basis for chronostratigraphic zonation. The formulations of Walsh (1998, 2000) move assemblage zones into the realm of range zone and interval zone, for which definitions already obtain (table 1.3).

Finally, abundance (or acme) zones are self-explanatory, with the abundance of a taxon or group of taxa relative to that of other taxa being distinctive of a certain stratal succession. The subjective nature of the sampling aspect in determining taxon abundance speaks strongly against such zones being precisely replicable stratigraphically; therefore, they are of limited use as a basis for chronostratigraphy. Walsh (1998) has no comparable category to abundance zone.

**Chronostratigraphic Units** According to Hedberg (1976:67), a chronostratigraphic unit is “a body of rock strata that is unified by being the rocks formed during a specific interval of geologic time, [and] . . . represents all rocks formed during a certain time span of Earth history and only those rocks formed during that time span. Chronostratigraphic units are bounded by isochronous surfaces.” Salvador (1994:88) states, “The essential part of the definition of a chronostratigraphic unit is the time span during which the unit described was formed.” Contrary to Walsh (2001 and references cited therein) this does not mean that the span of time (and its limits) have been identified before the definition of the chronostratigraphic unit (table 1.4). Salvador (1994:88) continues, “Since the only record of geologic time and of the events of geologic history lies in the rocks themselves, the

**TABLE 1.4** Chronostratigraphic and Geochronologic Categories

AFTER SALVADOR (1994)	AFTER WALSH (1998, 2001)
<b>CHRONOSTRATIGRAPHIC CATEGORIES</b>	<b>CHRONOSTRATIGRAPHIC CATEGORIES</b>
<p>Bodies of rock formed during a specified interval of geologic time.<sup>a</sup> The basic nonhierarchical unit is the chronozone.</p> <p>Formalized hierarchic units are the System, Series, and Stage. Intervals are based on boundary stratotypes and unit stratotypes; golden spikes (GSSPs) may be used. The boundary stratotype is the same concept as the golden spike. GSSPs are chronostratigraphic entities (also as per the International Commission on Stratigraphy, even if correlation precedes definition).</p>	<p>A set of rocks formed during a chronologic unit</p> <p>Geochronostratigraphic unit: The set of all existing rocks formed during a specified geochronologic unit.<sup>1</sup> The basic nonhierarchical unit is the geochronozone.<sup>b</sup> Formalized hierarchic units are the System, Series, and Stage.<sup>c</sup></p> <p>Biochronostratigraphic unit: The set of all existing rocks formed during a specified biochronologic unit. The basic nonhierarchical unit is the biochronozone.<sup>d</sup> Informal hierarchic units are the “System,” “Series,” and “Stage.”<sup>e</sup></p>
<b>GEOCHRONOLOGIC CATEGORIES</b>	<b>CHRONOLOGIC CATEGORIES</b>
<p>The units of geologic time during which chronostratigraphic units were formed.</p> <p>Limits are established by equivalent and precursor chronostratigraphic units.</p> <p>The basic nonhierarchical unit is the chron.</p> <p>Formalized hierarchic units are the Period, Epoch, and Age.</p>	<p>A span of time defined by two historical events.</p> <p>Geochronologic unit: A span of time defined by two geologic events, that is, the deposition of two exactly specified sedimentary layers, marked by golden spikes, in specified stratigraphic sections (boundary stratotypes). GSSPs are geochronologic entities.<sup>f</sup> The basic non hierarchic unit is the geochron.<sup>g</sup> Formalized hierarchic units are the period, epoch, and age.<sup>h</sup></p> <p>Biochronologic unit: A span of time defined by two paleobiologic events (e.g., evolution, extinction, immigration). The basic nonhierarchical unit is the biochron.<sup>i</sup> Informal hierarchic units are the Period, Epoch, and Age.</p>

GSSP, Global Boundary Stratotype Section and Point.

<sup>a</sup>In Salvador (1994) the interval of time is based on evidence intrinsic to the unit in question, as presented in its original formulation and definition. Stage precedes Age. In Walsh (1998) the interval of time is based on evidence extrinsic to the unit in question, on a precedent geochronologic unit. Age precedes Stage.

<sup>b</sup>This is the same concept as chronozone in Salvador (1994). Both are stratigraphically based units representing an interval of time.

<sup>c</sup>These are the same hierarchic units as in chronostratigraphic unit hierarchies of Salvador (1994).

<sup>d</sup>This is effectively the same concept as the biozone in Salvador (1994), a stratigraphic demonstration of paleobiologic phenomena. In that the boundaries of such units in Walsh (1998) are placed on only paleobiologic evidence (biochronologic unit), they are relegated to informal status. This is contrary to Salvador (1994), wherein biozones commonly contain temporal information ultimately taken to define the boundaries of chronostratigraphic units, which then lead to formalized Stages and higher hierarchic units.

<sup>e</sup>This subordinates paleontologically defined events (biochronologic units) relative to geologically defined events (geochronologic units; boundaries based on GSSPs).

<sup>f</sup>Golden spikes are used in fundamentally different ways: for chronostratigraphic units in Salvador (1994) and for geochronologic units in Walsh (1998).<sup>h</sup>

<sup>g</sup>Walsh (1998) uses the geochron as a time interval based on the deposition of a body of rock (Williams 1901); it is distinct from the chron of Salvador (1994), a strictly temporal term (Williams 1901). This is consistent with Salvador (1994).

<sup>h</sup>Implicit here is that only “geologic” events are appropriate to boundary definition for the Period, Epoch, and Age, with GSSPs being documented in a lithostratigraphic context, rather than these being based on chronostratigraphic data. What formerly were chronostratigraphic procedures now have become those of geochronology.

<sup>i</sup>This again implies that paleontologic data are subordinate in importance to GSSPs or other “geologic” events in determining and defining ages of stratigraphic sequences.

best standard for a chronostratigraphic unit is a body of rocks formed between two designated instants of geologic time.” Implicit in discussions of both Hedberg (1976) and Salvador (1994) are the fundamental presumptions that rocks are tangible records of the passage of time, that a limit in the rock record implies a limit in time, that chronostratigraphic units are based on marine strata, that the disposition and delimitation of that record is based

on criteria meaningful to that situation, and that the defining boundaries are normally based on paleontological data. Addressing the requirement of boundary isochroneity leads to examination of all possibly time-significant information that, following the examples provided in Aubry (1991, 1995, 1997) and Berggren and Aubry (1996), can lead to consideration of fidelity of the record and identification of phenomena useful in correlation

that have no intrinsic numerical age connotation, including paleomagnetic pattern. Therefore neither chronostratigraphy or geochronology is hostage to numerical analysis, for which direct evidence is difficult to achieve in marine contexts in any case.

Walsh (1998, 2001) cites some general concerns as to the establishment of chronostratigraphic units (the need for type and reference sections) and their global role in correlation. Actually, there is no confusion between chronostratigraphic units and unit stratotypes because the stratigrapher must proceed from the local to the more regional context for any chronostratigraphic unit, and the location of golden spikes or other formalized important referents must be determined at the end of the process, not at its beginning. To do otherwise presupposes the stratigrapher having prescient knowledge of all globally significant information at the beginning rather than at the end of the operation. The current chronostratigraphic scale has its own historical precedents, and the present-day focus on global geologic events may differ from those considered important when some or most of that scale was first devised. Still, the fact that the stratigraphic community now can provide numerical statements for an increasingly diverse range of geologic situations doesn't mean that the basic principles for establishing and dealing with chronostratigraphic and geochronologic time scales must be abandoned. Stage still must come before Age. By definition, chronostratigraphic units are bounded by isochronous horizons that are everywhere the same age, but demonstration of this goal is a continuing process and ultimately is achievable only within the abilities of a geochronologist to identify the limits of accuracy of a given time indicator (the  $\pm$  factor).

Although the basic hierarchical chronostratigraphic unit is the chronozone (table 1.1), the lower boundary of the stage is formally recognized as also setting the basal boundary of the Series and System (Hedberg 1976:71–74), and because the issue of temporal accuracy and repeatability is so important, current codes and guides specify that the stage be defined by its boundary stratotype. This may be based on a Global Boundary Stratotype Section and Point (GSSP; Cowie et al. 1986) because the intent of unit definition is that it is recognizable on a global scale. Therefore the means of identifying the boundary of the unit also should be globally relevant. Here is where the operation of high-resolution chronostratigraphy is paramount because all available means must be organized so as to describe and define an isochronous horizon. Until recently, fossils have been the primary means for age evaluation so that the considerations addressed earlier in this

chapter applied exclusively. The hierarchy of units above and below a stage are given in table 1.1.

In that regard, the chronozone is the most useful chronostratigraphic unit for fine-scale correlation. For all chronostratigraphic units, the means for identifying the span of time they represent must be specified. Critically, base defines boundary, so it is important to specify the basis on which the original (stratotypic) unit was founded and considered to be identifiable in other places around the world. Therefore not only must boundary stratotypes be established for the lower boundary of the stage and the one that succeeds it, but also their locale must be chosen in places where unique instants of geologic time may be recognized stratigraphically in places having a record of effectively continuous (Sadler 1981, 1999) sedimentary accumulation (Salvador 1994:90). Once established, a chronozone, a stage, or any other chronostratigraphic unit is theoretically global in extent. If time-significant criteria can identify the boundaries of the unit in places other than its type area, the unit may be extended laterally regardless of whether the original (e.g., paleontological) criteria are present.

**Geochronologic Units** In Salvador (1994:16) a geochronologic unit is “a unit of geologic time (time determined by geochronologic methods). It is not a body of rocks and therefore not a stratigraphic unit, although it may correspond to the time span of a stratigraphic unit.” Still, as indicated in table 1.1, geochronologic units are the lingua franca of the geologic time scale for general communication and have a formal relationship to chronostratigraphic units. The fact that North American land mammal ages are so designated results from the fact that there is no antecedent chronostratigraphic stage in most cases (Wood et al. 1941; Woodburne 1987a, 1996a). Walsh (1998, 2001) argues in favor of transferring to geochronology many of the operations herein maintained for chronostratigraphy, as indicated in table 1.4. Based on the preceding discussions, these suggestions are not followed here.

#### THE $^{40}\text{Ar}/^{39}\text{Ar}$ DATING SYSTEM IN COMPARISON WITH OTHER GEOCHRONOMETRIC METHODS

The laser total-fusion single-crystal  $^{40}\text{Ar}/^{39}\text{Ar}$  radioisotopic dating method is one of the most widely used and precise methods of geochronologic analysis (Renne et al. 1998), and commonly considered, at least informally, as superior to the K–Ar method, which had such an important role to play in the initial calibration of the mammal age chronology (Evernden et al. 1964). For example, Swisher

et al. (1993) report a mean age of  $65.16 \pm 0.04$  Ma for a volcanic ash in the IRZ coal of Montana, coincident with the iridium anomaly that identified the Cretaceous–Tertiary (K–T) boundary. This can be compared with an  $^{40}\text{Ar}/^{39}\text{Ar}$  age of  $15.85 \pm 0.04$  Ma for the Oreodont Tuff of the upper Miocene Barstow Formation of California. These dates, of strongly different parts of the Classical Time Scale (CTS), appear to have similarly small  $\pm$  factors, but those actually reflect analytical precision alone. Comparisons of these ages are valid as long as only the  $^{40}\text{Ar}/^{39}\text{Ar}$  or K–Ar system is used. Renne et al. (1998:131) indicate that when dates derived from  $^{40}\text{Ar}/^{39}\text{Ar}$  analyses are compared with those derived from other systems, such as the U/Pb method, additional corrections to the  $\pm$  factor should be applied to reflect uncertainties in the decay constants and the absolute ages of standards used in the analyses. In such a situation, recalculation of the IRZ coal date would result in an age of  $65.46 \pm 0.63$  Ma (Renne et al. 1998) and would be more appropriate for comparison with ages derived from other chronometric systems.

Regarding the accuracy of  $^{40}\text{Ar}/^{39}\text{Ar}$  dates and those derived from  $^{40}\text{K}$ – $^{40}\text{Ar}$  analysis, Woodburne et al. (1990) and MacFadden et al. (1990) report on reanalyzed  $^{40}\text{K}$ – $^{40}\text{Ar}$  ages for some of the tuffs of the Barstow Formation. For example, the  $^{40}\text{K}$ – $^{40}\text{Ar}$  age given for sanidine crystals in a sample of the Oreodont Tuff in Coon Canyon is  $15.8 \pm 0.2$  Ma (MacFadden et al. 1990, table 2). This is very close to the  $^{40}\text{Ar}/^{39}\text{Ar}$  date of  $15.9 \pm 0.06$  Ma from sanidine of the same sample in Coon Canyon and to a  $^{40}\text{Ar}/^{39}\text{Ar}$  date of  $15.85 \pm 0.04$  Ma from a site in Rainbow Basin, about 1 mile east (MacFadden et al. 1990:489). A similarly close correspondence in age when comparing the  $^{40}\text{Ar}/^{39}\text{Ar}$  and  $^{40}\text{K}$ – $^{40}\text{Ar}$  systems is provided by the Dated Tuff of the Barstow Formation. Biotite from this tuff dated by the  $^{40}\text{Ar}/^{39}\text{Ar}$  method yielded an age of  $14.8 \pm 0.06$  Ma. Analysis of the same biotite by the  $^{40}\text{K}$ – $^{40}\text{Ar}$  method yielded an age of  $14.8 \pm 0.15$  Ma (MacFadden et al. 1990:490). These examples show not only that the  $^{40}\text{K}$ – $^{40}\text{Ar}$  and  $^{40}\text{Ar}/^{39}\text{Ar}$  methods are capable of similar results but also that the correspondence in age corroborates the age derived from the  $^{40}\text{K}$ – $^{40}\text{Ar}$  system.

#### THE GEOMAGNETIC POLARITY TIME SCALE (GPTS), THE INTEGRATED MAGNETOBIOSTRATIGRAPHIC SCALE (IMBS), AND ESTIMATING THE AGE OF GEOBIOLOGICAL EVENTS

**GPTS and IMBS** It is now well known that Earth's magnetic field reverses periodically and that such reversals are recorded in magnetically susceptible minerals of molten

igneous or fluid sedimentary rocks. It also has become obvious that fossil mammals can be found in sequences of nonmarine strata that record patterns of geomagnetic reversals that can be correlated with the global pattern of the GPTS. Extension of the global pattern to mammal-bearing sequences provides a major basis by which events important to mammalian paleontologists can be compared with those of other disciplines worldwide, despite the fact that nonmarine sequences generally are considered to be less complete than those deposited in marine environments. Still, Aubry (1995) has shown that even the deep marine record may be significantly incomplete, and a thesis of this book is that in the best examples, stratigraphic and chronologic analysis of nonmarine sequences can approach that embodied by the IMBS of the marine realm. For both the marine and nonmarine realms, correlating an indigenous magnetic reversal pattern to part of the global template is one operation. It is another matter to assign numerical ages to that pattern in any realm.

As illustrated by Aubry (1995), the close link between the stratigraphic record and geologic time is manifested in a number of ways important to our purposes. Most mammalian paleontologists work with elements of the CTS, wherein the age of events in the stratigraphic record is supplied by radioisotopic data. Increasingly, nonmarine stratigraphers also have become accustomed to taking advantage of the GPTS as applied to continental strata. This chronologic scale is derived from a linear distance relationship of the age of magnetic polarity reversals recorded in oceanic basalts preserved on the floors of ocean basins derived from spreading rates calibrated by interpolations between, or extrapolations from, radioisotopic dating of selected magnetic reversals (Cande and Kent 1992, 1995; Berggren et al. 1995a). The radioisotopic information transforms the relative magnetic reversal pattern into a numerical chronology. In that it is almost impossible to directly date the oceanic basalts, the chronologic scale is developed by use of marine or nonmarine stratigraphic sections wherein the polarity pattern can be directly calibrated radioisotopically (see Vine and Matthews 1963; Heirtzler et al. 1968; Berggren et al. 1985; Lindsay et al. 1987; Aubry 1995 for summaries). Whereas initial correlation of a given nonmarine magnetostratigraphic succession typically is based on radioisotopic data, Albright (1999, 2000) gives a recent example in which biostratigraphic and biochronologic data were used for both original and refined age control of a lengthy magnetic polarity sequence in the absence of radioisotopic dates.

Berggren et al. (1995a, 1995b) introduced the concept of the IMBS as consisting of three elements: a magne-

tochronology, a numerical scale (combined as the GPTS), and a magnetobiochronologic framework. In that the GPTS is independent of biological evolution, the relative magnetic reversal pattern can be applied to the relative biochronologic framework without circularity in reasoning, and the calibration applied to the reversal pattern also can be carried into the biochronologic data set, providing its numerical calibration. Among other operations, the numerical scale allows quantification of geologic (and paleontologic) processes.

**Estimating the Age of Biochronologic Events** Despite advances in radioisotopic dating, biostratigraphy (augmented by magnetostratigraphy, chemostratigraphy, and cyclostratigraphy) still is the most important and widespread means of obtaining numerical ages from the stratigraphic record. The primary importance of biostratigraphy is that it is noniterative; that is, it is an ordinal scale. This is in contrast to magnetic polarity zones, chemostratigraphic signatures, and so on, which require either biostratigraphy or radioisotopic dating to order the other events in a temporal context.

Because not all biochronologic boundaries are directly calibrated, and because many of them fall within rather than at magnetic polarity reversal boundaries, it is necessary to estimate the ages of the biochronologic framework, either in the deep sea or on land. This operation also can be significant in refining, via feedback, the points at which the GPTS is calibrated and can effect revisions therein (e.g., Cande and Kent 1992, 1995). In this context, the term *estimation* has taken on a formal meaning in chronostratigraphy. Rather than connoting a guess at a reasonable calibration of a biochronologic boundary, *estimation* (sometimes called temporal analysis; Aubry 1995) means addressing this goal with deliberate attention given to rational means by which it may be achieved. The many examples from Berggren and Van Couvering (1974, 1978), Berggren et al. (1995b), and Aubry (1995, 1998) illustrate that evaluation of accumulation rate (its steadiness or erraticism) in sequences under discussion comprises a primary objective as addressed from any pertinent perspective, including magneto-, bio-, cyclo-, chemo-, and lithostratigraphy. Rather than guesswork, this is a rigorous and time-consuming process. Once stratigraphic completeness has been appraised it is possible (where warranted) to derive numerical ages for paleobiological or other events based on interpolations between, or extrapolations from, parts of the succession that are directly calibrated by radioisotopic means.

In theory, two magnetozones (for instance) may be collapsed into one, or be cut out entirely, by an unconfor-

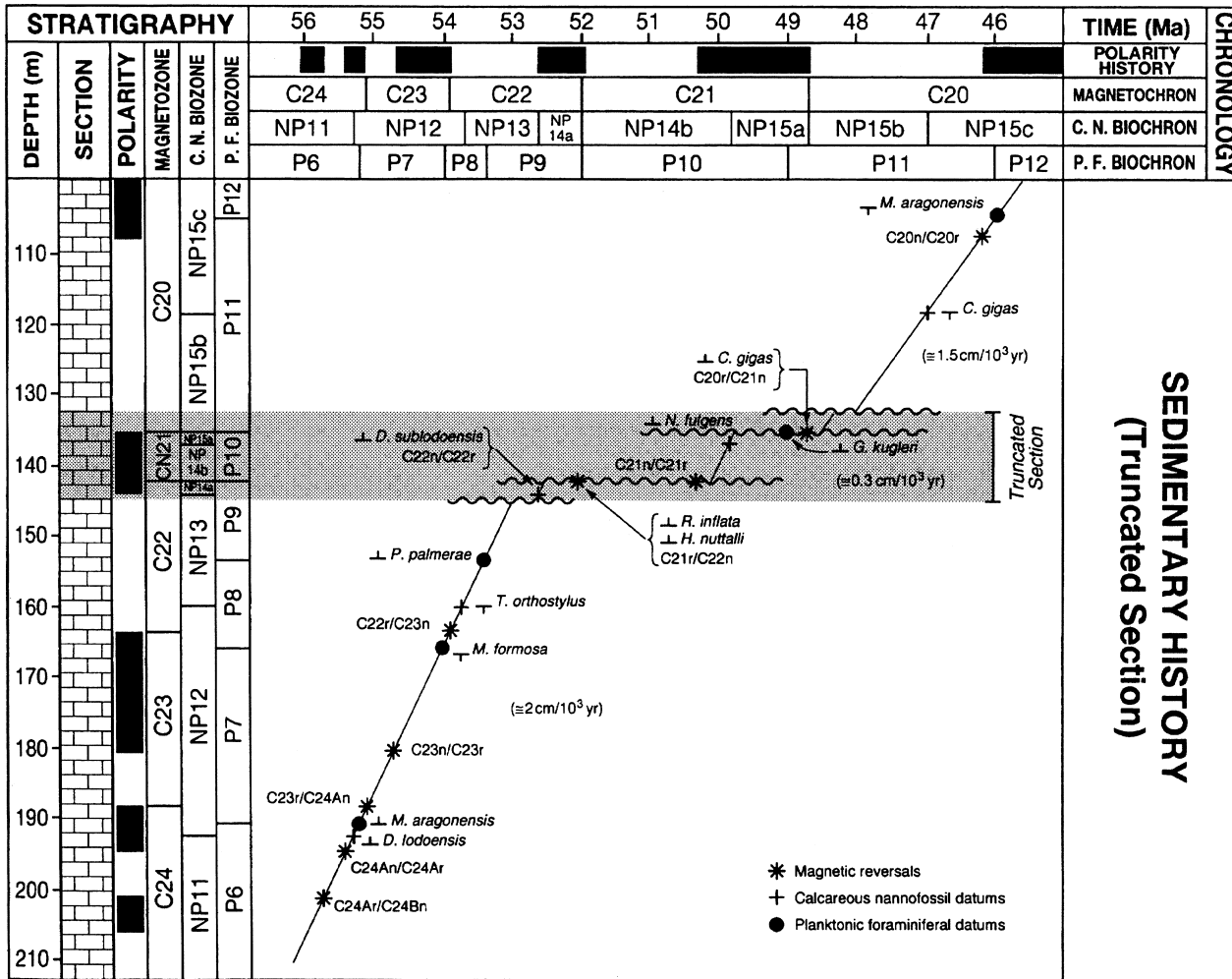
mity (Aubry 1991, 1998; Aubry et al. 1996). Similarly, geochemical profiles or any other pattern may be interrupted and condensed by unconformities, giving spurious results (Aubry 1998). Truncation of taxon ranges may appear to have been diachronous on a regional basis, but this appearance may reflect only an incomplete stratigraphic record. Also, an apparent range truncation might result from parts of a taxon record being eliminated within the stratigraphic extent of the range (preserving both top and bottom). A visually thin biozone thus might translate into spurious estimations of accumulation rate.

To test for hidden unconformities, Aubry (1991, 1995) proposes a method as excerpted here. If the stratigraphic section is continuous, the thickness of the magnetozones and biozones in the stratigraphic record should be proportional to their respective durations as magnetostratigraphic and biostratigraphic units of the IMBS. The accumulation curve calculated on this basis should be a straight line. If the record reflects a condensed section, the apparent succession of events still will be recorded, but the accumulation rate constructed from them will be of a shallower slope than that previously or subsequently (figure 1.1). If the record reflects a truncated section (unconformity), the accumulation rate slope will be disjunct within the zone of unconformity (figure 1.2). Whereas it probably is not possible to determine the amount of sediment removed (or not deposited) by the processes that result in an unconformity, it is possible to determine the amount of time represented by the hiatus.

In this exercise, the two surfaces that bound an unconformity are of different genetic significance. The upper surface is the bounding surface that reflects renewed deposition after the hiatus. The lower surface reflects all the events that conspired to form the hiatus in the first place. Dating the surfaces of an unconformity involves three steps (Aubry 1991, 1995).

1. Recognizing the presence of an unconformity as in severe truncation of biozones, magnetostratigraphic units, and so on (e.g., figure 1.2) or sharp contacts between lithologic units or subunits.
2. Estimating the duration of the hiatus by estimating the duration of the magnetostratigraphic units or biozones that are not represented in the various stratigraphic sections
3. Dating the age of the boundaries using any information or approach (e.g., accumulation rate) that narrows the level of imprecision as to the likely age of the unconformable surfaces

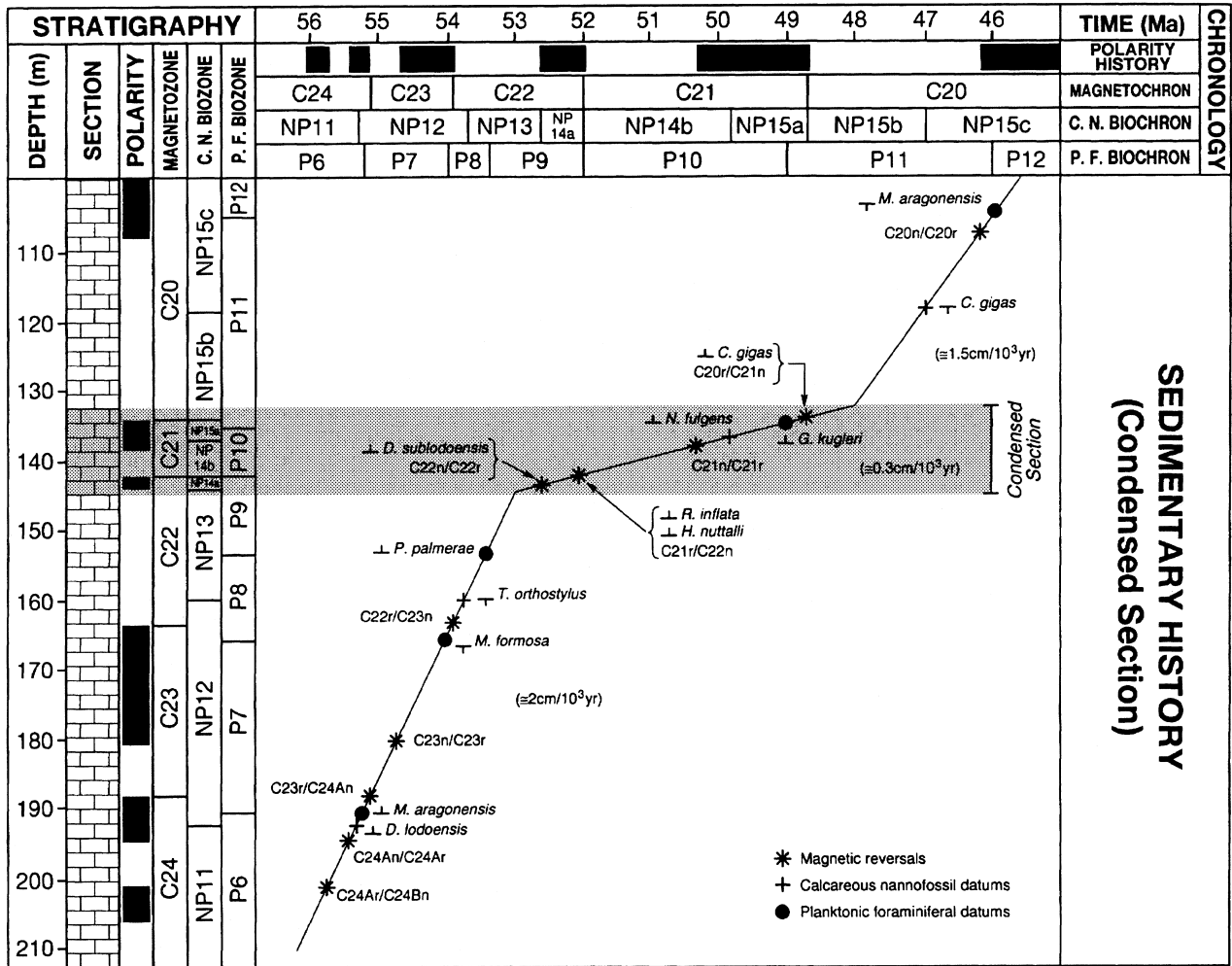
In an important illustration of the significance of these considerations, Aubry et al. (1996) developed a detailed examination of the fidelity of the core retrieved from Deep Sea Drilling Project (DSDP) site 550. This core contains



**FIGURE 1.1** Sedimentary history of a section containing successive truncations (stratigraphic gaps), inferred from the sedimentation rate curve. The truncated section is characterized by the anomalous stratigraphic juxtaposition of paleontologic events and magnetic reversals that are temporally well separated. Note the disjunct nature of the sedimentation rate curve in the truncated interval. After Aubry (1995). C.N., calcareous nannofossil; P.F., planktonic foraminiferal.

the data from which a date of 55 m.y. was estimated for the age of the Paleocene–Eocene Series boundary in the GPTS (Cande and Kent 1992, 1995) on the basis of the nannoplankton (NP) 9/10 zonal boundary. This chronozone boundary occurs 13 m and 7 m, respectively, below ashes dated by the laser total fusion  $^{40}\text{Ar}/^{39}\text{Ar}$  method at  $54.0 \pm 0.53$  Ma and  $54.51 \pm 0.05$  Ma, from which Swisher and Knox (1991) derived an accumulation rate for this part of the DSDP core. This accumulation rate was extrapolated to the part of the section containing the NP9/10 zonal boundary, resulting in its estimated age of 55 Ma. Cande and Kent (1992, 1995) used this age for the NP9/10 chronozone boundary as one of the nine calibration points from which their GPTS was constructed. But Aubry et al. (1996) showed that there is an unconformity at level 408 m in the core at site 550 and that part of zone NP10 is missing and suggested

that the age of the NP9/10 zonal boundary probably is 0.3–0.4 m.y. older than previously thought. At the same time, one cannot arbitrarily change the age of the NP9/10 chronozone boundary because that has not yet been demonstrated in a section that is complete across that boundary and because such a change carries implications for the age of all other magnetostratigraphic units in that part of the GPTS and thus for all biostratigraphic events in the relevant part of the IMBS (Aubry et al. 1996). Wing et al. (2000) explore this situation and also that for the best developed data set for the age of the  $\delta^{13}\text{C}$  excursion in nonmarine strata of the Fort Union and Willwood formations of the Big Horn Basin, Wyoming. The age of this excursion in the marine realm (55.5 Ma; Aubry et al. 1996) depends on the same data set as that from which an age of 55.0 Ma was derived for the Paleocene–Eocene boundary. Although dif-



**FIGURE 1.2** Sedimentary history of a section containing a condensed interval, inferred from the sedimentation rate curve. The condensed section yields the sequential occurrences of paleontologic events and magnetic reversals that characterize the temporal interval during which it was deposited. After Aubry (1995). C.N., calcareous nannofossil; P.F., planktonic foraminiferal.

ferent in detail, the analysis of sequence and evidence offered by Wing et al. (2000) for the Big Horn Basin succession is comparable in pitfalls and uncertainty to the marine situation under discussion (although for different reasons, including possible variations in local tectonism that might affect accumulation rates used in extrapolations; Bao et al. 1999). Neither the marine nor the nonmarine realm offers a single stratigraphic section that is complete across the interval involved (inclusive of the relevant calibration points), so the proposal that the NP9/10 chrono-zonal boundary is 55.0 Ma (but might be 55.3 Ma; Berggren and Aubry 1996) cannot be reconciled with the proposal (Wing et al. 2000, table 7.5) that the comparable level in the nonmarine record of the GPTS (two-thirds of the way down in C24r [= C24r.0.66]) is 55.315 Ma. Although the numbers appear similar, either or both might be inaccurate because of uncertainties in each case.

As of this writing, this situation had not been resolved. On one hand, the ages of reversal boundaries for the GPTS are commonly reported to three decimal places (e.g., Berggren et al. 1995b:132–133; chron C29r: 64.745–65.578 Ma). On the other hand, users of the numerical estimates for the age of reversal boundaries of the GPTS should be aware of the methods and the facts by which they are determined and treat them with a level of caution appropriate to the scale of the study for which they are being used.

### OPERATIONS IN CORRELATION

The purpose of this section is to outline some further operational aspects basic to developing temporal correlations of paleontological data before turning to a discussion of biochronology.



**Base Defines Boundary** This concept (George et al. 1969) is fundamental to all unit stratigraphic boundaries intended for potential or actual use in correlation. As indicated earlier, boundaries of lithostratigraphic units typically are chosen on sharp or at least replicable changes in lithology. Although not universal, it is common that lithostratigraphic boundaries are based on breaks in the rock record (i.e., at unconformities). As Schenk and Muller (1941) point out, this may be desirable for rock units, but it is definitely undesirable for chronostratigraphic units or related concepts (e.g., table 1.2). When the intent is to develop a succession important for correlation and replication of the passage of time, units and the boundaries between them should be chosen in locations where gaps (unconformities) do not exist or at least are made as minimal as possible.

It is certainly necessary to have a stratotype section for a formalized unit and its boundaries so that subsequent observers can replicate the criteria on which the unit was based originally (Salvador 1994:88). It also is intellectually appealing to have the bottom and top of a particular unit displayed in a single stratigraphic section and to have the type section that forms the basis of the unit to be a single, continuous stratigraphic transect. However, this goal may not be attainable realistically, and rather than having the one-section notion result in loss of important data because of imperfections in the rock record, modern codes and guides (e.g., Hedberg 1976; Salvador 1994) provide for a variety of typical sections to address the situations in which not only the base of the unit (boundary stratotype) but also characteristically typical representations of the unit (composite or component stratotypes) may be portrayed. Thus a given stratigraphic unit could have a boundary stratotype for its base, another for its top (the base of the next overlying unit), and one or more unit stratotypes where the main characterizing features (rocks or taxa) of the unit are well displayed and replicable by subsequent workers (Aubry et al. 1999).

**Definition and Characterization** The utility of a sequence of chronostratigraphic units for correlation is directly proportional to the degree to which its boundaries may be unambiguously defined and to the interval being characterized for the purpose of general recognition and correlation. In order to have unambiguous utility, the base of a stratigraphic unit must be defined as unambiguously as possible (and therefore must be repeatable and replicable precisely by other workers). A unit is characterized by the joint occurrence of a number of taxa (in this case), so that finding a representative number of them in a rock sequence is sufficient to propose a correlation

to somewhere in the unit under discussion. It commonly is convenient to use immigrant taxa as allochthonous novelties (various kinds of datums) in stratigraphic sections as a means for boundary definition (Repenning 1967; Woodburne and Swisher 1995), although this is not always the case, so that taxa resulting from endemic evolution must be considered as well (Murphy 1977; Woodburne 1977, 1987b, 1996a).

Walsh (1998) presents a convenient summary of concerns commonly offered regarding the choice between single-taxon boundary definitions and multiple-taxon characterizations of biostratigraphic or biochronologic units. In short, if the boundary can't be found because the single definition is absent, it is better to increase the chance of finding the boundary with a number of ways (taxa) in which to do so.

If the boundary is to be used in correlation, one logically would seek the least ambiguous (= best) means to identify that point in time. When originally addressing boundaries for fossil mammals, Woodburne (1977) was proposing the best means for that purpose and clearly recognized that a single-taxon definition would be elusive. After all, the best often is a rare entity. Ideally, a single taxon would be widespread, geologically instantaneous in distribution, and abundantly found. The *Hippotherium* datum is one such candidate (Woodburne 1996a). It also is a rare example of such a phenomenon (Walsh 1998). The *Hippotherium* datum illustrates a fairly common situation regarding such phenomena, at least for the fossil mammal record: Although this datum is of nearly global significance, its source still has not been recognized. Although the North American genus *Cormohipparion* appears to be the best candidate for the source of the dispersal event known as the *Hippotherium* datum, neither the species of *Cormohipparion* that probably was the source for the datum nor its phyletic relationship to sister taxa within *Cormohipparion* has been securely identified (Woodburne 1996b). Whereas the time of origin in North America of the Old World *Hippotherium* datum may be unresolved, the age of the regional first appearance datum (FAD) is closely approached at 11.1 Ma (Woodburne 1996a; Garcés et al. 1996; Rögl and Daxner-Höck 1996).

Practical realities suggest that most correlations, as demonstrated by the presence of one or more taxa characteristic of them, are to some point within a chronologic unit or interval. Most correlations are thus centrist in tendency. Still, it is rational to propose the best means by which a biostratigraphic or chronostratigraphic boundary may be recognized, even if having found it means that the boundary already has been crossed to some extent

(Walsh 1998). The extent to which such crossing is temporally significant is open to further testing depending on operational goals, and such testing is futile absent an unambiguous, temporally distinctive boundary definition.

## THE ROLE OF BIOCHRONOLOGY

Berggren and Van Couvering (1978:40) indicate that “long-distance correlations are geochronologic in substance,” and because of the prevalence of fossils over other kinds of chronologically significant data, such long-distance correlations are effectively biochronologic.

Berggren and Van Couvering (1978:40) portray the evidentiary and interpretive progression from biostratigraphy to biochronology and ultimately to chronostratigraphy. In that report, biochronologic correlations are based on recognizing the most widespread and distinctive events in biologic history (mostly FAD and LAD), identifying (replicating) those events in local biostratigraphies (lowest stratigraphic datums [LSDs] or LOs of current terminology) and evaluating their age with respect to as many other criteria as possible, and stratigraphically relating the events to evidence for other biochronologic datum events and to radiometrically dated or calibrated levels such as a volcanic layer or a paleomagnetic boundary in order to justify the interpretation that the event is isochronous throughout its area of extent.

Both Salvador (1994) and Hedberg (1976) state that biostratigraphic units are material and descriptive, rather than interpretive, whereas chronostratigraphic units are interpretive, representing an interval of time as recorded in the strata of the unit. Still, biostratigraphic correlation may approach chronostratigraphic correlation, and biostratigraphic (paleontologic) data give the basic material on which time correlations are based because they are nearly ubiquitous and they reflect the irreversibility of organic evolution.

Implicit in both Salvador (1994) and Hedberg (1976) are the commonly used procedures in developing correlations of rock units based on fossils: Biostratigraphic procedures allow the documentation in a physical framework of the presence and stratigraphic occurrence of a distinctive paleontological event, such as a new species or genus, developed either by in situ evolution or as reflecting dispersal from a possibly unknown source. This situation is conveniently and accurately described as an LSD or LO.

Finding the same paleontological stratigraphic pattern in other geographically distributed sites (other LSDs or

LOs) permits the inference that the pattern may be of temporal significance and that the various LSDs or LOs may be correlated on the basis of their biostratigraphic similarity. From the paleontological (including mammal age) standpoint, these correlations are biocorrelations of Salvador (1994:15), which, when demonstrated to be temporally valid, become chronocorrelations. From the perspective of this book, mammalian biostratigraphic information (LO) is proposed to have biochronologic significance. The biochronologic significance is rooted in the irreversibility of organic evolution and documented as to its place in time and space by the stratigraphic framework in which it is manifested. Once frozen in stratigraphy, the biotic properties can be inspected for their relationship to any other kind of potentially significant temporal information and integrated with any iterative or ordinal data so as to eventually arrive at a level of documentation that enables the original biostratigraphic LOs to be awarded a temporal connotation. This operation begins with biostratigraphy, proceeds via biochronology, and leads to chronostratigraphy. In that mammal ages are biochrons, their role in correlation is clear, and a purpose of this book is to identify the quantity and quality of temporally significant information that can be related to mammal ages so that their chronologic utility is increasingly refined and illuminated.

Placing disparate sections (a number of LOs) into a chronologic framework based on stratigraphically adjacent phenomena considered to have their own temporal significance (i.e., other fossil zones, radioisotopic or magnetostratigraphic data) allows the further inference that the boundaries under discussion are isochronous (and at what scale). The resulting FAD is one example of that operation and would define the lower boundary of a given chronostratigraphic unit. Finding the next highest datum (and successfully validating its relevance to correlation; Aubry et al. 1999:113) would permit the designation of a chronozone with both boundaries defined. When this is accomplished, the entire package of strata can be combined into a chronostratigraphic unit of regional scale, with its implication for correlation, which is the purpose of the exercise in the first place.

Thus biostratigraphically based biochronologic correlation is an integral part of assessing age relationships of fossils and the rocks in which they occur, but is not formalized in current codes and guides. The important relationship between biochronology, biostratigraphy, and chronostratigraphy justifies the formalization of biochronologic units, as pioneered by Walsh (1998:158). This is followed here in principle but not in all details of nomenclature (table 1.3). In all cases an attempt has been

made to preserve much of the pertinent language of the antecedent biostratigraphic unit, following Salvador (1994). Although the categories nominated by Walsh (1998, 2000) are similar in name and content, they are put to very different uses. In the proposal favored here, biochronologic correlations are of primary importance in establishing formal chronostratigraphic units, which are the necessary precursors to formulating geochronologic units. In the proposals represented by Walsh (1998, 2000, 2001, and references therein), geochronologic units are constructed in advance of chronostratigraphic units, biochronology plays only a subsidiary role, and chronostratigraphic units are “merely abstract sets of material strata” (Walsh 2001:708). One of the most useful aspects of Walsh (1998) is the emphasis on separating empirical and descriptive operations and concepts as distinct from theoretical and interpretive concepts. As Aubry (1997:22) stresses, the goal is to discuss rock and time with a simple but dual nomenclature.

The following biochronologic units are suggested (table 1.3): assemblage chron (reflective of many original mammal ages being effectively assemblage zones: “an assemblage zone based on a fossil fauna” [Salvador 1994:63]), range chron, taxon-range chron, concurrent-range chron, interval chron, and lineage chron. The assemblage chron stems from the assemblage zone of Salvador (1994) rather than that of Walsh (1998; assemblage fossizone) because the latter is directed toward range zones, which are a different category as used here (table 1.3). The range chron stems from the range zone of Salvador (1994). The range zone of Walsh (1998) is effectively the same as the taxon-range zone of Salvador (1994), which here is taken as antecedent to the taxon-range chron. The concurrent-range chron stems from the concurrent-range zone of Salvador (1994), which is the same as the strict overlap assemblage fossizone where the number of taxa is two (Walsh 1998:161). The interval chron stems from the interval zone of Salvador (1994) and is the interval of time defined on the earliest age of two successive biohorizons. This is comparable to that of Walsh (1998), which is a restatement of Salvador (1994). The lineage chron stems from the lineage zone of Salvador (1994) and has no counterpart in Walsh (1998). In any case, the purpose of identifying these biochronologic units is to emphasize their application to interpretive correlation as biochrons subsequent to empirical biostratigraphic documentation.

In this context, *datum* carries a temporal (time = interpretive) connotation and should be kept separate from descriptive and empirical considerations (rock), also as stressed by Walsh (1998). Thus *FAD* is appropriate for a

temporal analysis, but *LSD* is a misnomer because it is a biostratigraphic (empirical, rock) concept (see *Definitions*). For that reason, *LO* (Aubry 1995, 1997) is preferred here (also Walsh 2000). Substituting *LO* for *LSD* preserves the intent promulgated by Opdyke et al. (1977), Lindsay et al. (1987), and Lindsay and Tedford (1990) but reserves the term *datum* for temporal interpretations rather than empirical stratigraphy. To paraphrase and substitute as to the proposal of Woodburne (1996a), a *FAD* is composed of a number of *LOs*. Each *LO* is a biostratigraphic record. If the *LOs* are associated with independent temporally significant information that shows them to be closely time correlative, they collectively support the interpretation that a *FAD* may be established (and within what temporal limits). If the proposal can be made that the *FAD* is isochronous over a large geographic area, then that concept (*FAD*) is appropriate to this interpretation. This is effectively similar to the proposals of Aubry (1995, 1997) except that the *FOD* (see *Definitions*) is not addressed here.

Hedberg (1976:86–92) and Salvador (1994:92–97) consider means by which established chronostratigraphic units may be extended away from their boundary or other stratotypes. Except for paleontological or radioisotopic criteria, the remainder<sup>1</sup> fall into various kinds of lithologic or other (paleomagnetic reversal) iterative, rather than ordinal, phenomena that depend on the former two kinds of data for demonstrating their temporal significance. Although mammal ages are not chronostratigraphic units, the means by which they may be correlated are the same because even though they were loosely constrained stratigraphically as originally formulated (Wood et al. 1941), mammal ages still had a stratigraphic component, and this has become increasingly refined ever since (although not necessarily uniformly).

## RELATIONSHIP TO MAMMAL AGES

North American land mammal ages (Wood et al. 1941; Savage 1951) typically are identified as biochronologic units constructed to recognize discrete intervals of time based on the evolution of fossil mammals. Historically these biochrons were only loosely tied to a stratigraphic framework. To a real but perhaps originally somewhat limited extent, mammal ages were basically assemblage zones (i.e., there was a biostratigraphic base of sorts), and their correlation made use of the derivative assemblage chron (although they were not labeled as such). After the work of Savage (1977), various kinds of chronostratigraphic and biostratigraphic units have been proposed for some of these intervals. Savage proposed the

Wasatchian Stage/Age and its correlation to the European Sparnacian Stage (nominally lower Eocene or upper Paleocene depending on interpretations of the Paleocene–Eocene Series boundary), and although it was provided only in summary form, it is clear that the Wasatchian Stage is represented in Colorado, New Mexico, and Wyoming, as well as in Europe. Although study of the Wasatchian Stage still is not complete, it can be considered as under current discussion (Gingerich and Clyde 2001; Gingerich 2001; Bowen et al. 2001; Clyde 2001; Strait 2001).

Rose (1980, 1981) proposed the Clarkforkian Stage/Age (nominally upper Paleocene, but pre-Wasatchian). Walsh (1998) calls the Wasatchian and Clarkforkian proposals into question on the basis of their having been no boundary stratotype identified for either of them. Still, attention to stratigraphic detail was implicit (Savage 1977) or explicit (Rose 1981) and contributed to the interpretation that the biostratigraphic patterns were chronologically significant. Rose (1981, figures 2 and 3) shows the distribution of measured sections and localities that demonstrate the lower and upper limits of the Clarkforkian Stage, even though type and reference sections are not designated by those names. Thus formalization of the unit remains only a matter of procedural assertion. Rose (1981:26–27) provides additional faunal characterization of the Clarkforkian to facilitate its recognition elsewhere and points out that stratigraphic demonstration of the unit in other areas still is under development. Archibald et al. (1987) indicate that Clarkforkian mammals are rarely found beyond the Big Horn and Clark's Fork basins of Wyoming. The best stratigraphic documentation of elements of the Clarkforkian Stage is provided by Clyde (2001) for the McCullough Peaks area situated between the Clark's Fork and Big Horn basins (figure 3.1, this volume). The current status of the Clarkforkian Stage is one of further development, documentation, and demonstration.

Archibald et al. (1987) offer a succession of biostratigraphic zones within the four mammal ages that equate about with the Paleocene Epoch. Lacking formal biostratigraphic stratotypes and detailed stratigraphic documentation, these interval zones, lineage zones, and acme zones remain biochronologic units, although their temporal succession still is viable. Williamson (1996) proposes a biostratigraphic zonation for rocks of the same age in the San Juan Basin, New Mexico, that in part validates the biochronology proposed by Archibald et al. (1987) through its explicit use of a physical stratigraphic framework. Although the biostratigraphic data were generalized into intervals, each of which was on the order of 5–10 m thick (or more in some cases) and separated by

apparently unfossiliferous intervals 10 or more meters thick, the stratigraphic order is clear. The suites of locality intervals were physically correlated between the various stratigraphic sections in the southern San Juan Basin and grouped into eight informal biostratigraphic zones, labeled A–H (Williamson 1996:27, figure 18), with many extending laterally up to about 40 km. The physical versus biological correlation of the localities leads these lettered units to be dubbed as paleontologically distinct lithozones by Walsh (2000). The physical correlation also was consistent with its magnetostratigraphic characterization. The physical framework thus constructed was transformed into eight formal biostratigraphic zones as defined paleontologically. The type stratigraphic section was explicitly given for each zone (Williamson 1996:49–53), and the stratigraphic ranges for the taxa on which the zones are based is clearly shown at the indicated level of precision mentioned earlier (Williamson 1996, figure 19). As one example, the stratigraphically lowest biozone was nominated as the *Hemithlaeus kowalevskianus*–*Taeniolabis taoensis* Zone. Although not so designated by Williamson (1996), this zone and all of the others are interval zones, save the stratigraphically highest *Mixodectes pungens* Taxon-range Zone. The biozones are defined on the basis of single taxa and are provided with characterizations based on index taxa and first and last occurrences. As indicated (Williamson 1996, figure 24), these biozones contain taxa correlative with Puercan (but not earliest Puercan) and Torrejonian mammal ages, with the *Hemithlaeus*–*Taeniolabis* Zone being about correlative with the *Ectoconus*–*Taeniolabis* (Pu2) biochron of Archibald et al. (1987) and the *Taeniolabis taoensis*–*Periptychus carinidens* Interval Zone about correlative with the *Taeniolabis*–*Periptychus* (Pu3) biochron. Williamson (1996) effectively validates the Pu2 and Pu3 tentative proposals of Archibald et al. (1987; see also chapter 3, this volume), but fossiliferous strata equivalent to Pu1, typified by faunas in Wyoming, Montana, and Colorado (Archibald et al. 1987), are not present in the San Juan Basin.

Superposition of the Puercan interval zones is demonstrated by Eberle and Lillegraven (1998a, 1998b) and Lillegraven and Eberle (1999) in their seminal studies of the Ferris Formation in the Hanna Basin of Wyoming. The Ferris Formation contains a sedimentary succession about 1200 m thick with fossil mammals of Late Cretaceous (Lancian) to early Paleocene (Puercan) age. Puercan fossils occur in the upper 600 m of section, and a detailed biostratigraphic array allows the superpositional demonstration of biochrons Pu1–3 of Archibald et al. (1987), with the content of Pu1 being modified somewhat

to take into account proposals for (Archibald and Lofgren 1990) and against (Lofgren 1995) the recognition of an earliest Puo zone (see also chapter 3, this volume). The Hanna Basin record is sufficient to validate as biostratigraphic interval zones the Pu1–3 biochrons originally proposed by Archibald et al. (1987) and to stabilize the allocation of mammal-bearing sites in other parts of North America to this chronologic scheme. Neither Eberle and Lillegraven (1998a), Lillegraven and Eberle (1999), nor the authors of chapter 3 take the additional steps needed to formalize these Puercan biostratigraphic units.

In a similar fashion, Williamson (1996) validates the Torrejonian biochrons of Archibald et al. (1987). Williamson's (1996) *P. carinidens*–*Protoselene opisthacus* Interval Zone is about correlative with the *Periptychus*–*Tetraclaenodon* (To1) biochron. Collectively the Williamson (1996) *P. opisthacus*–*Ellipsodon grangeri* Interval Zone, the *E. grangeri*–*Arctocyon ferox* Interval Zone, the *A. ferox*–*Pantolambda cavirictum* Interval Zone, and the *P. cavirictum*–*Mixodectes pungens* Interval Zone are about correlative with the *Tetraclaenodon*–*Pantolambda* (To2) biochron (chapter 3). The *Mixodectes pungens* Taxon-range Zone of Williamson (1996) is about correlative with the *Pantolambda*–*Plesiadapis praecursor* (To3) biochron. Biostratigraphic validation of the Archibald et al. (1987) biochrons of the Tiffanian mammal age remains to be accomplished.

The biostratigraphic zones of the Fort Union and Willwood formations of the Big Horn Basin, Wyoming (Gingerich 1976, 1980, 1983, 1991), recently have been formalized (Gingerich 2001). Up to this point, these were biochronologic units. As summarized in Bown et al. (1994), P. D. Gingerich and colleagues have embarked on a detailed stratigraphic documentation of fossil-bearing sites of nominally Paleocene and early Eocene age in this region over a period of nearly 30 years. Implicit as well as explicit in these studies has been the correlation of the fossil sites to a number of master stratigraphic sections, with stratigraphic assignments commonly given in meters with respect to a given datum. Although the methods used were those that can lead to biostratigraphic zonations, and the density of biostratigraphic data has greatly increased over time, the proposed zonation has been formally described only recently (Gingerich 2001). Thus the succession of lineage zones, interval zones, and acme zones previously used now is provided with a clear, if terse, definition (lowest range datum [LRD]), stratigraphic documentation, and nominal abbreviation (Cf3 = *Phenacodus*–*Ectocion* Acme Zone).

Previously, the only procedurally valid (in part) biostratigraphic zonation for part of this succession was pro-

posed by Schankler (1980) for the Willwood Formation of nominally early Eocene age (Bown et al. 1994:35). Following recommendations in Hedberg (1976), a threefold zonation was developed in reference to a measured section approximately 730 m thick, with taxon ranges displayed with respect to the fossil localities that were tied or referred to that master section (grouped into 10-m intervals). The boundaries of the zones were chosen at places showing a marked change in faunal composition. Schankler (1980) chose the boundary between the first two zones, the *Haplomylus*–*Ectocion* Range Zone and the *Bunophorus* Interval Zone at a point labeled biohorizon B, located at a level (380 m) intermediate between a major extinction (370 m) and immigration (390 m) event. In comparable fashion, the boundary between the *Bunophorus* Interval Zone and the next higher *Heptodon* Range Zone was placed at a level (530 m) at which another immigration event is recorded. Although Schankler (1980) follows the “base defines boundary” principle, it would have been preferable to begin the *Bunophorus* Interval Zone at the immigration level (390 m) where data actually are present rather than in the unfossiliferous gap at 380 m. It also would have been preferable had the localities not been lumped into 10-m intervals, an unavoidable procedure at the time. Moreover, Bown et al. (1994:35–36) point out that the stratigraphic section that provided Schankler's zonation was never located on a map; his biostratigraphic zones thus were not provided with a type section and so are of limited utility. The promise of completely demonstrating a viable biostratigraphy was not realized by Schankler (1980), and these are not incorporated in the compilation of Gingerich (2001). Clyde (2001) used the part of the zonations in Gingerich (2001) in the McCullough Peaks Formation, found between the Clark's Fork and southern Big Horn basins. The McCullough Peaks Formation contains fossil mammals that range from Tiffanian (Ti3) through Wasatchian (Wa7) and contributes to the paleomagnetic characterization of these zones in comparison to those from sequences in Polecat Bench. Taking into account the interpolation of zonal boundaries in unfossiliferous parts of the stratigraphic sections, Clyde (2001) proposes two models for age assignment for the biozones of the two regions (also see Wing et al. 2000), based on interpolating zonal boundaries with respect to the magnetostratigraphic zonation. In model 1, the Clarkforkian begins at 56.17 Ma, the Wasatchian zone Wao at 54.96 Ma. In model 2, these ages are 56.47 Ma and 55.23 Ma, respectively.

D. R. Prothero and colleagues have been long involved in a faunal succession of the nominally Oligocene strata in North America. The interval zone framework used in

chapter 5 for fossil mammals exemplifies the detailed use of magnetostratigraphic information to assist in temporal discrimination of this zonation.

The small number of these and possible other examples reflects, in part, the generally discontinuous record of mammal-bearing deposits in North America but also probably results to some extent from the fact that, overall, mammal age correlations provide results that are satisfactory to their users. In fact it can be argued that valuable refinements can be made to the framework in which mammalian biochronology resides without resorting to the kind of effort needed for their detailed formalization into biostratigraphic or chronostratigraphic units (Lindsay and Tedford 1990). But if for no other reason than to be able to accurately relate events in the fossil mammal time scale to those of global extent (Zachos et al. 2001), close attention still must be given to the chronology of mammal-bearing sequences.

Examples in chapters 3–7 illustrate ongoing refinements in biostratigraphically assisted biochronology, including increased scrutiny of boundaries of mammal ages and their subdivisions, and the principles and practices that can lead to the development of soundly based biostratigraphic and chronostratigraphic units (and the disciplinary rigor needed for this purpose) are exactly those needed to increase the documentation and integration of information with which to assess the quality, validity, and scale of resolution of correlations based on fossil mammals. One of the features of mammalian biochronologies in the 1987 volume and in the current volume is the use of immigration events as defining the boundaries of mammal ages or other units. Whether reflecting endemic genesis or intercontinental correlation, the question of diachroneity is either cited and recognized (Woodburne 1977, 1996a) or emphasized (Alroy 1998). Still, Aubry (1995, 1997) has implied that at basinal (transpose continental here) scale, diachrony is less likely to be an operational problem than is the impact of stratigraphic imperfection on local sections. A major charge to the mammalian biostratigraphic and biochronologic community is to increase the number of pertinent stratigraphic sections and to improve the estimates of age, correlation, and completeness for all of them.

#### NOTE

1. These include physical interrelations of strata (e.g., superposition); lithic correlations, especially volcanogenic units; geomagnetic reversals; climatic change (the  $\delta^{13}\text{C}$  excursion, mentioned earlier, could be one such example); eustatic sea level changes or other processes that create unconformities;

and subsequent reflooding or other kinds of resumed deposition.

#### REFERENCES

- Albright, L. B. 1999. Magnetostratigraphy and biochronology of the San Timoteo Badlands, southern California, with implications for local Pliocene–Pleistocene tectonic and depositional patterns. *Geological Society of America Bulletin* 111(9):1265–1293.
- . 2000. Biostratigraphy and vertebrate paleontology of the San Timoteo badlands, southern California. *University of California Publications in Geological Sciences* 144.
- Alroy, J. 1998. Diachrony of mammalian appearance events: Implications for biochronology. *Geology* 26:91–207.
- Archibald, J. D., P. D. Gingerich, E. H. Lindsay, W. A. Clemens, D. W. Krause, and K. D. Rose. 1987. First North American land mammal ages of the Cenozoic era. In *Cenozoic Mammals of North America: Geochronology and Biostratigraphy*, ed. M. O. Woodburne. Berkeley: University of California Press, pp. 24–76.
- Archibald, J. D. and D. L. Lofgren. 1990. Mammalian zonation near the Cretaceous–Tertiary boundary. In *Dawn of the age of mammals in the northern part of the Rocky Mountain Interior, North America*, ed. T. Bown and K. D. Rose. Boulder, CO: Geological Society of America Special Paper 243:31–50.
- Aubry, M.-P. 1991. Sequence stratigraphy: Eustacy or tectonic imprint? *Journal of Geophysical Research* 96:6641–6679.
- . 1995. From chronology to stratigraphy: Interpreting the stratigraphic record. In *Geochronology, time scales and global stratigraphic correlation*, ed. W. A. Berggren, D. V. Kent, M.-P. Aubry, and J. Hardenbol. Tulsa: SEPM Special Publication 54, pp. 213–274.
- . 1997. Interpreting the (marine) stratigraphic record. In *Actes du Congrès BiochroM '97*, ed. J.-P. Aguilar, S. Legendre, and J. Michaux. *Mémoires et Travaux E.P.H.E., Institut de Montpellier* 21:15–32.
- . 1998. Stratigraphic (dis) continuity and temporal resolution of geological events in the upper Paleocene–lower Eocene deep sea record. In *Late Paleocene–early Eocene climatic and biotic events in the marine and terrestrial records*, ed. M.-P. Aubry, S. G. Lucas, and W. A. Berggren. New York: Columbia University Press, pp. 37–66.
- Aubry, M.-P., W. A. Berggren, L. Stott, and A. Sihna. 1996. The upper Paleocene–lower Eocene stratigraphic record and the Paleocene–Eocene boundary carbon isotope excursion: Implications for geochronology. In *Correlation of the early Paleogene in northwest Europe*, eds. R. W. O. Knox, R. M. Corfield, and R. E. Dunay. Geological Society Special Publication 101:353–380.
- Aubry, M.-P., W. A. Berggren, J. A. Van Couvering, and F. Steininger. 1999. Problems in chronostratigraphy: Stages, series, unit and boundary stratotypes, global stratotype section and point and tarnished golden spikes. *Earth Science Reviews* 46:99–148.
- Bao, H., P. L. Koch, and D. Rumble III. 1999. Paleocene–Eocene climatic variations in western North America: Evidence from the  $\delta^{18}\text{O}$  of pedogenic hematite. *Geological Society of America Bulletin* 111(9):1405–1415.
- Berggren, W. A. and M.-P. Aubry. 1996. A late Paleocene–early Eocene NW European and North Sea magnetobiochronological correlation network. In *Correlation of the early Paleogene in*

- northwest Europe, ed. R. W. O. Knox, R. M. Corfield, and R. E. Dunay. Geological Society Special Publication 101: 309–352.
- Berggren, W. A., F. J. Hilgen, C. C. Langereis, D. Kent, J. D. Obradovich, I. Raffi, M. Raymo, and N. J. Shackleton. 1995a. Late Neogene (Pliocene–Pleistocene) chronology: New perspectives in high resolution stratigraphy. *Geological Society of America Bulletin* 107(11):1271–1287.
- Berggren, W. A., D. V. Kent, J. J. Flynn, and J. A. Van Couvering. 1985. Cenozoic geochronology. *Geological Society of America Bulletin* 96:1407–1418.
- Berggren, W. A., D. V. Kent, C. C. Swisher III, and M.-P. Aubry. 1995b. A revised Cenozoic geochronology and chronostratigraphy. In *Geochronology, time-scales and global stratigraphic correlations: A unified framework for an historical geology*, ed. W. A. Berggren, D. V. Kent, M.-P. Aubry, and J. Hardenbol. Tulsa: SEPM Special Publication 54:129–213.
- Berggren, W. A. and J. A. Van Couvering. 1974. The late Neogene: Biostratigraphy, geochronology and paleoclimatology of the last 15 million years in marine and continental sequences. *Palaeogeography, Palaeoecology, Palaeoclimatology* 16:1–216.
- . 1978. Biochronology. In *Contributions to the geologic time scale*, ed. G. V. Cohee, M. F. Glaessner, and H. D. Hedberg. Tulsa: American Association of Petroleum Geologists, Studies in Geology 6:39–55.
- Bowen, G. J., P. L. Koch, P. D. Gingerich, R. D. Norris, S. Bains, and R. M. Corfield. 2001. Refined isotope stratigraphy across the continental Paleocene–Eocene boundary on Polecat Bench in the northern Bighorn Basin. In *Paleocene–Eocene stratigraphy and biotic change in the Bighorn and Clark's Fork basins, Wyoming*, ed. P. D. Gingerich. *University of Michigan Papers on Paleontology* 33:73–88.
- Bown, T. M., K. D. Rose, E. L. Simons, and S. L. Wing. 1994. Distribution and stratigraphic correlation of upper Paleocene and lower Eocene fossil mammal and plant localities of the Fort Union, Willwood, and Tatman formations, southern Bighorn Basin, Wyoming. *U.S. Geological Survey Professional Paper* 1540.
- Cande, S. C. and D. V. Kent. 1992. A new geomagnetic polarity time-scale for the Late Cretaceous and Cenozoic. *Journal of Geophysical Research* 97:13,917–13,951.
- . 1995. Revised calibration of the geomagnetic polarity time scale for the Late Cretaceous and Cenozoic. *Journal of Geophysical Research* 100:6093–6095.
- Clyde, W. C. 2001. Mammalian biostratigraphy of the McCullough Peaks area in the northern Bighorn Basin. In *Paleocene–Eocene stratigraphy and biotic change in the Bighorn and Clark's Fork basins, Wyoming*, ed. P. D. Gingerich. *University of Michigan Papers on Paleontology* 33:109–126.
- Cowie, J. W., W. Ziegler, A. J. Boucot, M. G. Bassett, and J. Remane. 1986. Guidelines and statutes of the International Commission on Stratigraphy (ICS). *Courier Forschungsinstitut Senckenberg* 83:1–14.
- Eberle, J. J. and J. A. Lillegraven. 1998a. A new important record of earliest Cenozoic mammalian history: Eutheria and paleogeographic/biostratigraphic summaries. *Rocky Mountain Geology* 33(1):49–117.
- . 1998b. A new important record of earliest Cenozoic mammalian history: Geologic setting, *Multituberculata*, and *Paradectia*. *Rocky Mountain Geology* 33(1):3–47.
- Evernden, J. F., D. E. Savage, G. H. Curtis, and G. T. James. 1964. Potassium–Argon dates and the Cenozoic mammalian geochronology of North America. *American Journal of Science* 262:145–198.
- Garcés, M., J. Agustí, L. Cabrera, and J. M. Parés. 1996. Magnetostratigraphy of the Vallesian (late Miocene) in the Vallès-Penedès Basin (northeast Spain). *Earth and Planetary Science Letters* 142:381–396.
- George, T. N., W. B. Harland, D. V. Ager, H. W. Ball, W. H. Blow, R. Casey, C. H. Holland, N. F. Hughes, G. A. Kellaway, P. E. Kent, W. H. C. Ramsbottom, J. Stubblefield, and A. W. Woodland. 1969. Recommendations on stratigraphical usage. *Proceedings of the Geological Society of London* 1969:139–166.
- Gingerich, P. D. 1976. Cranial anatomy and evolution of early Tertiary Plesiadapidae (Mammalia, Primates). *University of Michigan Papers on Paleontology* 15:1–141.
- . 1980. Evolutionary patterns in early Tertiary mammals. *Annual Review of Earth and Planetary Science* 8:407–424.
- . 1983. Paleocene–Eocene faunal zones and a preliminary analysis of Laramide structural deformation in the Clark Fork's Basin, Wyoming. *Wyoming Geological Association Guidebook, 34th Annual Field Conference* 1983:185–195.
- . 1991. Systematics and evolution of the early Eocene Perisodactyla (Mammalia) in the Clark's Fork Basin, Wyoming. *University of Michigan Museum of Paleontology Contributions* 28(8):181–123.
- . 2001. Biostratigraphy of the continental Paleocene–Eocene boundary interval on Polecat Bench in the northern Bighorn Basin. In *Paleocene–Eocene stratigraphy and biotic change in the Bighorn and Clark's Fork basins, Wyoming*, ed. P. D. Gingerich. *University of Michigan Papers on Paleontology* 33:37–72.
- Gingerich, P. D. and W. C. Clyde. 2001. Overview of mammalian biostratigraphy in the Paleocene–Eocene Fort Union and Willwood formations of the Bighorn and Clark's Fork basins. In *Paleocene–Eocene stratigraphy and biotic change in the Bighorn and Clark's Fork basins, Wyoming*, ed. P. D. Gingerich. *University of Michigan Papers on Paleontology* 33:1–14.
- Hedberg, H. C. (ed.). 1976. *International stratigraphic guide*. New York: Wiley.
- Heirtzler, J. R., D. O. Dickson, E. M. Herron, W. C. Pitman III, and X. Le Pichon. 1968. Marine magnetic anomalies, geomagnetic field reversals, and motions of the ocean floor and continents. *Journal of Geophysical Research* 73:2119–2136.
- Lillegraven, J. A. and J. J. Eberle. 1999. Vertebrate faunal changes through Lancian and Puercan time in southern Wyoming. *Journal of Paleontology* 73(4):691–710.
- Lindsay, E. H., N. D. Opdyke, N. M. Johnson, and R. F. Butler. 1987. Mammalian chronology and the magnetic polarity time scale. In *Cenozoic mammals of North America: Geochronology and biostratigraphy*, ed. M. O. Woodburne. Berkeley: University of California Press, pp. 269–284.
- Lindsay, E. H. and R. H. Tedford. 1990. Development and application of land mammal ages in North America and Europe, a comparison. In *European Neogene mammal chronology*, eds. E. H. Lindsay, V. Fahlbusch, and P. Mein. New York: Plenum Press. NATO Advanced Science Institute Series 180:601–624.
- Lofgren, D. L. 1995. The Bug Creek problem and the Cretaceous–Tertiary transition at McGuire Creek, Montana. *University of California Publications in Geological Sciences* 140:1–185.
- MacFadden, B. J., C. C. Swisher III, N. D. Opdyke, and M. O. Woodburne. 1990. Paleomagnetism, geochronology, and possible tectonic rotation of the middle Miocene Barstow Formation,

- Mojave Desert, California. *Geological Society of America Bulletin* 102:478–493.
- Murphy, M. A. 1977. On chronostratigraphic units. *Journal of Paleontology* 51:213–219.
- North American Commission on Stratigraphic Nomenclature. 1983. North American stratigraphic code. *American Association of Petroleum Geologists Bulletin* 67(5):841–875.
- Opdyke, N. D., E. H. Lindsay, N. M. Johnson, and T. Downs. 1977. The paleomagnetism and magnetic polarity stratigraphy of the mammal-bearing section of Anza-Borrego State Park, California. *Quaternary Research* 7:316–329.
- Renne, P. R., C. C. Swisher III, A. L. Deino, D. B. Karner, T. L. Owens, and D. J. K. DePaolo. 1998. Intercalibration of standards, absolute ages and uncertainties in  $^{40}\text{Ar}/^{39}\text{Ar}$  dating. *Chemical Geology* 145:117–152.
- Repenning, C. A. 1967. Palearctic–Nearctic mammalian dispersal in the late Cenozoic. In *The Bering land bridge*, ed. D. M. Hopkins. Stanford, CA: Stanford University Press, pp. 288–311.
- Rögl, R. and G. Daxner-Höck. 1996. Late Miocene paratethys correlations. In *The evolution of western Eurasian Neogene mammal faunas: The 1992 Schloss-Reisenburg workshop concept*, ed. R. L. Bernor, V. Fahlbusch, and W.-W. Mittmann. New York: Columbia University Press, pp. 47–55.
- Rose, K. D. 1980. Clarkforkian land mammal age: Revised definition, zonation and tentative intercontinental correlation. *Science* 208:744–746.
- . 1981. The Clarkforkian land-mammal age and mammalian faunal composition across the Paleocene–Eocene boundary. *University of Michigan Papers on Paleontology* 26:1–197.
- Sadler, P. M. 1981. Sediment accumulation rates and the completeness of stratigraphic sections. *Journal of Geology* 89:589–584.
- . 1999. The influence of hiatuses on sediment accumulation rates. *GeoResearch Forum* 5:15–40.
- Salvador, A. (ed.) 1994. *International stratigraphic guide*. Boulder, CO: The Geological Society of America.
- Savage, D. E. 1951. Late Cenozoic vertebrates of the San Francisco Bay region. *University of California Publications in Geological Sciences* 28:215–314.
- . 1977. Aspects of vertebrate paleontological stratigraphy and geochronology. In *Concepts and methods of biostratigraphy*, ed. E. G. Kauffmann and J. E. Hazel. Stroudsburg, PA: Dowden, Hutchinson, and Ross, pp. 427–442.
- Schankler, D. M. 1980. Faunal zonation of the Willwood Formation in the central Bighorn Basin, Wyoming. In *Early Cenozoic paleontology and stratigraphy of the Bighorn Basin Wyoming*, ed. P. D. Gingerich. *University of Michigan Papers on Paleontology* 24:99–114.
- Schenk, H. G. and S. W. Muller. 1941. Stratigraphic terminology. *Geological Society of America Bulletin* 52:1414–1426.
- Strait, S. G. 2001. New Wa-o mammalian fauna from Castle Gardens in the southeastern Bighorn Basin. In *Paleocene–Eocene stratigraphy and biotic change in the Bighorn and Clark's Fork basins, Wyoming*, ed. P. D. Gingerich. *University of Michigan Papers on Paleontology* 33:127–144.
- Swisher, C. C. III, L. Dingus, and R. F. Butler. 1993.  $^{40}\text{Ar}/^{39}\text{Ar}$  dating and magnetostratigraphic correlation of the terrestrial Cretaceous–Paleogene boundary and Puercan mammal age, Hell Creek–Tullock formations, eastern Montana. *Canadian Journal of Earth Sciences* 30:1981–1996.
- Swisher, C. C. III and R. O. Knox. 1991. The age of the Paleocene/Eocene boundary:  $^{40}\text{Ar}/^{39}\text{Ar}$  dating of the lower part of NP10, North Sea Basin and Denmark. In *IGCP 308 (Paleocene/Eocene Boundary Events) International Meeting and Field Conference 2–6 December 1991*. Brussels: Abstracts with Programs, p. 16.
- Vine, F. J. and D. H. Matthews. 1963. Magnetic anomalies over oceanic ridges. *Nature* 199:947–949.
- Walsh, S. L. 1998. Fossil datum and paleobiological event terms, paleontostratigraphy, chronostratigraphy, and the definition of land mammal “age” boundaries. *Journal of Vertebrate Paleontology* 18(1):150–179.
- . 2000. Eubiostratigraphic units, quasibiostratigraphic units, and “assemblage zones.” *Journal of Vertebrate Paleontology* 20(4):761–775.
- . 2001. Notes on geochronologic and chronostratigraphic units. *Bulletin of the Geological Society of America* 113(6):704–713.
- Williams, H. S. 1901. The discrimination of time-values in geology. *Journal of Geology* 9:570–585.
- Williamson, T. E. 1996. The beginning of the age of mammals in the San Juan Basin, New Mexico: Biostratigraphy and evolution of Paleocene mammals of the Nacimiento Formation. *New Mexico Museum of Natural History Science Bulletin*.
- Wing, S. L., H. Bao, and P. L. Koch. 2000. An early Eocene cool period? Evidence for continental cooling during the warmest part of the Cenozoic. In *Warm climates in Earth history*, ed. B. T. Huber, K. G. MacLeod, and S. L. Wing. New York: Cambridge University Press, pp. 197–236.
- Wood, H. E. II, R. W. Chaney, J. Clark, E. H. Colbert, G. L. Jepsen, J. B. Reeside Jr., and C. Stock. 1941. Nomenclature and correlation of the North American continental Tertiary. *Bulletin of the Geological Society of America* 52:1–48.
- Woodburne, M. O. 1977. Definition and characterization in mammalian chronostratigraphy. *Journal of Paleontology* 51(2):220–234.
- . 1987a. Mammal ages, stages, and zones. In *Cenozoic mammals of North America: Geochronology and biostratigraphy*, ed. M. O. Woodburne. Berkeley: University of California Press, pp. 18–23.
- . 1987b. Principles, classification, and recommendations. In *Cenozoic mammals of North America: Geochronology and biostratigraphy*, ed. M. O. Woodburne. Berkeley: University of California Press, pp. 9–17.
- . 1996a. Precision and resolution in mammalian chronostratigraphy: Principles, practices, examples. *Journal of Vertebrate Paleontology* 16(3):531–555.
- . 1996b. Reappraisal of the *Cormohipparion* from the Valentine Formation, Nebraska. *American Museum of Natural History Novitates* 3163.
- Woodburne, M. O. and C. C. Swisher III. 1995. Land mammal high-resolution geochronology, intercontinental overland dispersals, sea level, climate, and vicariance. In *Geochronology, time scales and global stratigraphic correlation*, ed. W. A. Berggren, D. V. Kent, M.-P. Aubry, and J. Hardenbol. Tulsa: SEPM Special Publication 54:335–364.
- Woodburne, M. O., R. H. Tedford, and C. C. Swisher III. 1990. Lithostratigraphy, biostratigraphy and geochronology of the Barstow Formation Mojave Desert, southern California. *Geological Society of America Bulletin* 102:459–477.
- Zachos, J., M. Pagani, L. Sloan, E. Thomas, and K. Billups. 2001. Trends, rhythms, and aberrations in global climate 65 Ma to present. *Science* 292:686–693.



# 2

## Mammalian Biochronology of the Latest Cretaceous

Richard L. Cifelli, Jaelyn J. Eberle,  
Donald L. Lofgren, Jason A. Lillegraven,  
and William A. Clemens

**M**AMMALIAN DIVERSIFICATION, in both the ecologic and taxonomic senses, sharply increased in the early Tertiary. Consideration of Late Cretaceous assemblages therefore provides essential background to interpretation of the great evolutionary radiations that followed. Most relevant in this connection is the Lancian land mammal age, which preceded the Puercan and is the focus of this summary. However, the Lancian itself must be viewed in the context of preceding land mammal ages and assemblages that have not yet been assigned to a land mammal age. Particularly significant problems remain in identifying the beginning of the Lancian, biochronologically and chronostratigraphically. For this reason, we begin with a brief general review of the record of North American, Late Cretaceous mammals, with particular emphasis on the widely used land mammal ages correlative with the Campanian and Maastrichtian marine stages/ages of Europe. A summary of the age relationships of better-known mammal faunas and ages for the Late Cretaceous of North America is shown in figure 2.1.

Land mammal ages have been established in North America only for the latter part of the Cretaceous period, and for this reason we must make frequent reference to European marine stages in reviewing temporal relationships, especially among the older faunas. The Aquilan, Judithian, “Edmontonian” (not formally recognized at present), and Lancian North American land mammal ages (NALMAs) were proposed initially as true chronostratigraphic ages or stages (Dorf 1942; Russell 1964, 1975). That is, they were based on the physical limits of specific rock bodies and their contained fossils (NACSN 1983), and therefore they differ conceptually from North Amer-

ican land mammal ages (Wood et al. 1941), which are intended to serve as time units. The original definitions of the Cretaceous ages also included other fossils, including plants, molluscs, and lower vertebrates such as dinosaurs. With great increases in knowledge of Late Cretaceous mammals, however, attention later became focused largely on mammalian assemblages in characterizing these ages (Fox 1978). Lillegraven and McKenna (1986) formally proposed Aquilan, Judithian, and Lancian land mammal ages. We follow their definitions and usage herein, modifying the characterization of the Lancian and its principal correlates based on new information.

The beginning of the Puercan land mammal age is defined by the first appearance of *Protungulatum donnae* (Archibald and Lofgren 1990; chapter 3, this volume). A commonly used marker of the Cretaceous–Tertiary boundary in more global terms is at the base of a clay containing anomalously high levels of iridium in the marine boundary stratotype at El Kef, Tunisia (Keller et al. 1995). The age of the boundary currently is placed at  $65.5 \pm 0.1$  Ma (Obradovich and Hicks 1999), which, because of the use of different standard monitor ages in their calculations, is older than the widely cited age of  $65.16 \pm 0.04$  (Swisher et al. 1993). The Lancian–Puercan and Cretaceous–Tertiary boundaries correspond closely in time, and the two are commonly considered to have been synchronous (see discussion and references in Eberle and Lillegraven 1998b). However, definitions of the Lancian–Puercan and Cretaceous–Tertiary are based on differing criteria, studied in different depositional settings on distant continents. As discussed later in this chapter, at least one transitional mammalian assemblage appears to be a Puercan fauna of Late Creta-

Late Cretaceous Mammal Ages and Faunas										
Ma	Polarity Chrono-Zones	Polarity	Period	Epoch	Age	NALMA	Pre-Aquilian Faunas	Notable First Occurrences	Geographic Range	
60	C26		TERTIARY PALEOGENE	PALEOCENE	SELANDIAN	Tiffanian				
	C27				DANIAN	Torrejonian				
	C28					Puercan				
	C29					65.0 Ma				
	C30		CRETACEOUS	LATE		LANCIAN		<i>Cimexomys minor</i> , <i>Parectypodus</i> , <i>Glasbius</i> , <i>Alphadon jasoni</i> , <i>Gypsonictops illuminatus</i>	AK, AB, SK, MT, ND, SD, WY, CO, UT, NM, TX*, NJ*	
	C31				MAASTRICHTIAN					
70	C32				71.3 Ma	"Edmontonian"			<i>Meniscoessus collomensis</i> , <i>Didelphodon</i>	AB, MT, CO
					CAMPANIAN	JUDITHIAN			<i>Meniscoessus major</i> , <i>M. intermedius</i> , <i>Mesodma primaeva</i> , <i>Cimolomys clarki</i> , <i>Turgidodon</i> , <i>Gypsonictops</i>	AB, SK, MT, WY, UT, NM, TX
	C33									
									<i>Mesodma</i> , <i>Eodelphis</i> , <i>Paranyctoidea</i>	AB, UT
					83.5 Ma	SANTONIAN			John Henry Mbr. fauna (Coniacian–Santonian)	UT
					85.8 Ma	CONIACIAN			Iron Springs Fm. fauna (Turonian–Santonian)	UT
90					89.0 Ma	TURONIAN			Smoky Hollow Mbr. fauna (Turonian)	UT
			93.5 Ma							
				CENOMANIAN			Dakota Fm. fauna (late Cenomanian)	UT		
			98.0 Ma				Mussentuchit L. F. (98.5 Ma)	UT		
100				ALBIAN						

\* = Isolated occurrences of single specimens that may be of Lancian age

**FIGURE 2.1** Late Cretaceous mammal ages and pre-Aquilian faunas. Polarity and polarity chronozones are from Gradstein et al. (1995); normal polarity chrons are shaded black. Numeric ages for stage boundaries are from Palmer and Geissman (1999). North American land mammal age (NALMA), pre-Aquilian fauna, notable first occurrence, and geographic range information is found in the main text and is based on references cited therein. Shaded boundaries between NALMAs indicate that they are not well defined (see text for discussion).

ceous age. Radioisotopic ages for chrons of the Geomagnetic Polarity Time Scale (GPTS) as used in our summary follow Berggren et al. (1995). The term *faunule* is used herein as a diminutive form of *fauna*, referring to a fauna known by only a few taxa rather than to an association of taxa interpreted intentionally for its ecological significance (see *Definitions*).

### PRE-LANCIAN FAUNAS

#### ALBIAN-CENOMANIAN INTO SANTONIAN

Until recently, the North American record of Late Cretaceous mammals older than approximately the early Campanian was almost nonexistent, consisting of a few, largely

indeterminate specimens (McNulty and Slaughter 1968; Clemens et al. 1979; Krause and Baird 1979; Emry et al. 1981). Several reasonably well-represented mammalian faunas, all from Utah, are now known for the approximately 15-m.y. time span separating the beginning of Cenomanian from the beginning of Campanian time. Oldest of these is the Mussentuchit local fauna, collected from a restricted stratigraphic interval in upper parts of the Cedar Mountain Formation. <sup>40</sup>Ar/<sup>39</sup>Ar age determinations indicate the fauna of about 28 mammalian varieties to be about 98.5 Ma, placing it near the Albian–Cenomanian (Early–Late Cretaceous) boundary (Cifelli et al. 1997, 1999b).

A somewhat younger fauna is known from the Dakota Formation, southern Utah. Mammalian fossils come from the middle member of the unit and are considered late Cenomanian in age, based on invertebrate fossils from

marine facies in parts of the Dakota Formation itself (Eaton 1991). The next youngest, reasonably well-known mammalian fauna comes from the Smoky Hollow Member of the Straight Cliffs Formation, also in southern Utah. The age of this assemblage is Turonian, based on molluscs from sub- and superjacent marine units (Eaton 1987, 1991). Late Cretaceous, pre-Campanian faunules, as yet not well known, have been reported from the John Henry Member (Coniacian–Santonian) of the Straight Cliffs Formation, which overlies the Smoky Hollow Member (Eaton et al. 1999a); an unspecified unit of ?Coniacian–Santonian age (Eaton et al. 1999b); and various horizons in the Iron Springs Formation, southwestern Utah, possibly including mammals of Turonian and Coniacian–Santonian age (Eaton 1999a).

The discontinuous record, limited geographic distribution, and in most cases scant sampling of these faunas, together with the preliminary nature of much of the taxonomic information available about them, severely limit interpretation of mammalian biochronology of these older strata. Accordingly, we limit ourselves in this summary to general observations on pre-Campanian mammals of North America. Members of Triconodontidae, represented by an endemic North American clade (Alti-conodontinae), are reasonably abundant in the Albian–Cenomanian (Cifelli and Madsen 1998). However, they are lacking from all later faunas except an Aquilan assemblage from Canada (Fox 1976a). Spalacotheriid symmetrodonts are present in all pre-Campanian faunas (Cifelli and Gordon 1999). Multituberculate assemblages are dominated by species of the “*Paracimexomys* group.” Relictual “plagiaulacidans” may be present in the Albian–Cenomanian, when Neoplagiaulacidae first appear (Eaton and Cifelli, unpublished data). Cimolodontidae are possibly present as early as the Cenomanian and, more securely, by the Coniacian–Santonian, when Cimolomyidae first appear (Eaton 1995, 1999a, 1999b; Eaton et al. 1999a, 1999b). Marsupials are present from the Albian–Cenomanian onward (Cifelli et al. 1999b), with Stagodontidae appearing by the Cenomanian (Cifelli and Eaton 1987) and the paraphyletic “Pediomyidae” by the Santonian (Eaton et al. 1999b). Notable by their absence throughout are Eutheria, which may have been present in the Aptian–Albian of North America but do not reappear until the early Campanian (Cifelli 1999).

## AQUILAN

The Aquilan, the oldest of North America’s formally defined Late Cretaceous land mammal ages, was characterized on the basis of a mammalian fauna from Verdigris

Coulee, in upper parts of the Milk River Formation, Alberta (Lillegraven and McKenna 1986). The age of the fauna is conventionally regarded as early Campanian, although placement of the Santonian–Campanian boundary has been disputed (Lillegraven 1991). An alternative interpretation would place the upper Milk River Formation in late Santonian time (Leahy and Lerbekmo 1995). The mammalian fauna, described in a series of papers by Fox (1971a, 1971b, 1976a, 1980b, 1982, 1984a, 1984b, 1987), is highly distinctive. In large part, this is almost surely a result of artifacts caused by hiatuses in the fossil record. The Aquilan is not immediately preceded by any well-known mammalian fauna and, because it is probably at least 80 Ma in age, it antedates Judithian assemblages by several million years. In addition to the fauna of some 31 mammalian varieties from Verdigris Coulee, the Aquilan is represented by several smaller assemblages from Utah. Of these, the most diverse is from the Wahweap Formation of the Kaiparowits Plateau (Cifelli and Madsen 1986; Eaton 1987; Cifelli 1990b, 1990c); faunules also are known from the ?Wahweap Formation, Paunsaugunt Plateau (Eaton et al. 1998), and the Masuk Formation, Henry Mountains (Eaton 1990).

The Aquilan includes representations of a number of archaic mammalian lineages; most notable among last appearances are the spalacotheriid symmetrodonts and Triconodontidae. Nonetheless, the Aquilan differs significantly from older Late Cretaceous assemblages, particularly in the greater diversity of more advanced forms, including cimolodontid, cimolomyid, and neoplagiaulacid multituberculates (Fox 1971a; Eaton 1987), along with pediomyid marsupials (Fox 1971b). Notable first occurrences include the multituberculate *Mesodma*, the stagodontid marsupial *Eodelphis*, and the lipotyphlan insectivore *Paranyctoides* (see Fox 1984a; Cifelli 1990c). A species referred to *Paranyctoides* also has been described from Coniacian strata of Uzbekistan (Nessov 1993). This suggests the possibility that *Paranyctoides* arrived via dispersal from Asia rather than having evolved in situ in North America. Given that it is widely distributed among Aquilan (and later) faunas, *Paranyctoides* may eventually prove useful as a first appearance datum (FAD) for the Aquilan.

## JUDITHIAN

The basis of the Judithian land mammal age (Lillegraven and McKenna 1986) is a fauna from three localities situated near the top of the Judith River Formation, Choteau and Blaine counties, north-central Montana (Sahni 1972). Stratigraphic correlation of the type Judithian (and pre-

sumed correlatives mentioned later in this chapter) with adjacent marine units has yielded mixed interpretations for the age of the Judithian because of conflicting zonations based on foraminiferans and ammonites (see references and discussion in Lillegraven and McKenna 1986; Lillegraven and Ostresh 1990). Radioisotopic dates for a correlative assemblage (the Hill County Local Fauna), together with correlations based on ammonites, suggest that the Judithian had a duration of at least 5 m.y. and extended back to at least 78 Ma, placing it within standard chronology of the European Campanian (Goodwin and Deino 1989). Weil (1999) summarizes available biostratigraphic and geochronologic data and concludes that known Judithian faunas span the interval from approximately 79 to 74 Ma.

Geographically, the Judithian is the most widely represented of North America's Late Cretaceous land mammal ages. Correlative faunas from the Judith River Formation in Montana include a well-sampled assemblage from the northern part of the state, the Hill County Local Fauna (Montellano 1992), as well as a faunule from near the Mussellshell River, Wheatland and Golden counties (Fiorillo 1989; Fiorillo and Currie 1994), in south-central Montana. The Egg Mountain locality, in the Two Medicine Formation, northern Montana, has yielded rare but more complete specimens (Montellano 1988; Montellano et al. 2000). To the north, in Alberta, correlative faunas (arbitrarily grouped, for present purposes, as the Oldman assemblage) have been recovered from what is now called the Judith River Group (Eberth and Hamblin 1993). Best represented of these are fossils from the Dinosaur Park (upper Oldman) Formation (Fox 1979a, 1979b, 1979c, 1980a, 1981); a few specimens have been collected from the underlying Oldman and Foremost formations (lower Oldman of Fox 1976b). A few mammalian specimens also have been collected from the Judith River Group in Saskatchewan (Storer 1993). Principal correlatives to the south of the type Judithian include faunas from the "Mesaverde" Formation, Wind River and Bighorn basins, Wyoming (Lillegraven and McKenna 1986); Kaiparowits Formation, southern Utah (Cifelli 1990a, 1990c; Eaton 1993; Eaton et al. 1999a); upper part of the Fruitland Formation and transition zone between Fruitland and Kirtland formations, New Mexico (Clemens 1973; Flynn 1986; Rigby and Wolberg 1987); and Aguja Formation, southern Texas (Rowe et al. 1992; Weil 1992; Cifelli 1994; Sankey 1998).

The Judithian is highly distinctive, sharing few species with other land mammal ages (Lillegraven and McKenna 1986; see table 2.2), and it appears to have been stable

through its duration (Goodwin and Deino 1989). Perhaps the biochronologically most useful first appearances are those of the multituberculates *Meniscoessus major*, *M. intermedius*, *Mesodma primaeva*, and *Cimolomys clarki*; marsupial *Turgidodon*; and leptictoid eutherian *Gypsonictops*. On the basis of dinosaur assemblages, Lehman (1997) defines northern and southern faunal provinces for the Judithian, corresponding to the *Aquilapollenites* and *Normapolles* palynofloras, respectively. Heterogeneity in composition of Judithian mammalian assemblages also appears to be related, at least in part, to latitudinal provinciality. PEDIOMYID and STAGODONTID marsupials, for example, are rare or lacking in southern assemblages. Statistical analyses of the Judithian mammalian assemblages show that available data do not permit formal recognition of one or more southern faunal provinces (Weil 1999), but mammalian local faunas of the San Juan Basin, New Mexico, await full description and modern systematic analyses of all their members.

#### "EDMONTONIAN"

It is generally recognized that a significant temporal hiatus separates typical Judithian from Lancian faunas (Lillegraven and McKenna 1986). Indeed, given the age range for the Judithian recognized here (ca. 79–74 Ma) and the fact that known Lancian local faunas correlate with the late Maastrichtian (ca. 67.5–65.5 Ma), it seems probable that the two land mammal ages are separated by as much as 7 m.y. Russell (1964, 1975) proposes an intervening terrestrial vertebrate stage, the "Edmontonian." Formal recognition of the "Edmontonian" as a land mammal age is not yet possible, however, and the term is used in quotation marks here, following Lillegraven and McKenna (1986). Fossiliferous nonmarine strata of probable "Edmontonian" age are not common in the Western Interior because of a late Campanian–early Maastrichtian marine transgression. In Montana, for example, this interval is represented by the Bearpaw Shale, which in most areas of outcrop lies stratigraphically between the largely terrestrial Judith River and Hell Creek formations (Lillegraven 1987). A second problem in recognizing an "Edmontonian" land mammal age is that a high proportion of the few mammals represented is either inadequately known or conspecific with Lancian taxa, so the species are not temporally diagnostic (Lillegraven and McKenna 1986). Fortunately, discoveries in recent years have greatly improved knowledge of this poorly understood interval, and we suggest that a defensible basis for definition of the "Edmontonian" soon may be possible.

The first mammalian assemblage to be referred to the "Edmontonian" was the Scabby Butte Local Fauna from the St. Mary River Formation, Alberta (Sloan and Russell 1974; Russell 1975); a second occurrence in the same formation is near Lundbreck, Alberta (Russell 1975). Two sites of probable "Edmontonian" age, also in Alberta, are known from the Horseshoe Canyon Formation (Fox and Naylor 1986). A small mammalian assemblage from the St. Mary River Formation in Montana remains under study (Heinrich et al. 1998). Most recently, a diverse mammalian assemblage has been reported from the Williams Fork Formation, northwestern Colorado. Correlation based on cephalopods indicates the fauna to be intermediate in age between the Judithian and Lancian (Lillegraven 1987). Although study of the fauna is still in progress (Archibald 1987a; Diem 1999), the mammals of the Williams Fork Formation show great promise for resolving problems inherent in defining the "Edmontonian" as a land mammal age. At present, the most diagnostic "Edmontonian" mammals are species of the multituberculate *Meniscoessus* and the stagodontid marsupial *Didelphodon* (see Fox and Naylor 1986; Lillegraven 1987).

The age of the Lower Hunter Wash fauna (from the upper part of the Fruitland Formation and lower part of the Kirtland Formation), New Mexico, has been disputed (see Butler et al. 1977; Butler and Lindsay 1985; Flynn 1986; Rigby and Wolberg 1987). As noted earlier, the range of ages of Judithian local faunas is estimated at 79–74 Ma, but paleontological criteria for recognition of a boundary between the Judithian and "Edmontonian" have yet to be established. The ages of the Lower Hunter Wash local faunas appear to approximate this undefined boundary. In addition to revisions of biostratigraphic correlations by Rowe et al. (1992), Fassett and Obradovich (1996) and Fassett and Steiner (1997) report  $^{40}\text{Ar}/^{39}\text{Ar}$  determinations that constrain the age of at least some of the localities yielding the Lower Hunter Wash fauna to the interval 74.56–74.11 Ma and show that they were deposited during chron C33n. Contrary to arguments presented by Lillegraven (1987) and Lillegraven and Ostresh (1990), this fauna is herein tentatively considered to be Judithian in age. A second biogeographically highly significant Judithian–"Edmontonian" boundary local fauna comes from three localities in the "El Gallo Formation" in Baja California del Norte (Lillegraven 1972). All the localities are above a tuff dated at 73 Ma, and ammonites from within and above the El Gallo Formation suggest a Campanian age. Finally, a possible Judithian (or ?"Edmontonian") correlative,

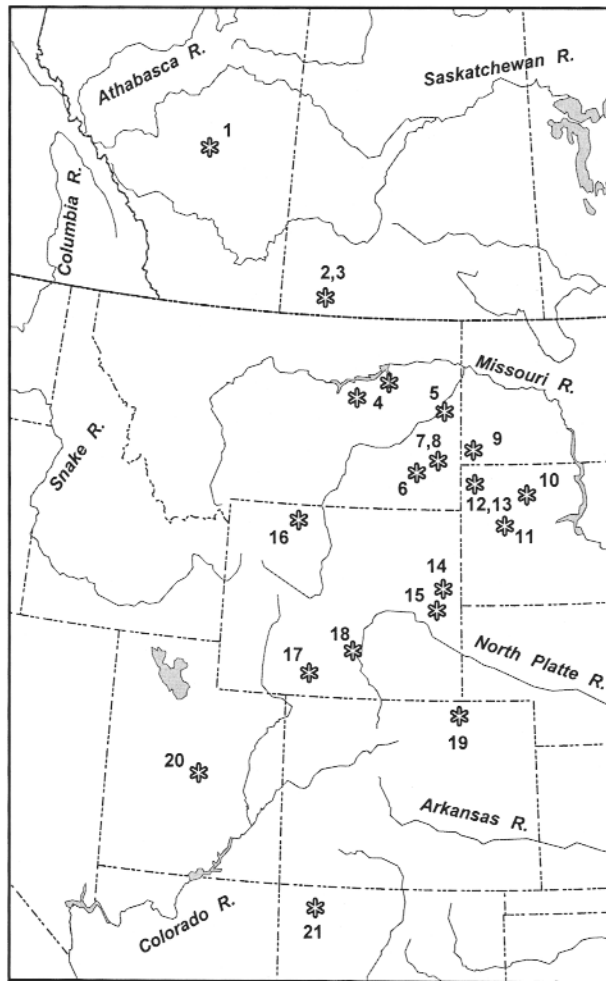
dated at 71.6 Ma, is known from the Marshalltown Formation, New Jersey (Grandstaff et al. 1992).

## LANCIAN LAND MAMMAL AGE

"The Lancian 'age' is a new (or modified) provincial time term, based upon mammalian fauna from type Lance Formation of east-central Wyoming in Niobrara County, faunal type areas in valleys of Lance Creek and its tributaries north of town of Lance Creek" (Lillegraven and McKenna 1986:51). The first discovered Cretaceous mammals are Lancian in age (Van Valen 1967), and occurrences of Lancian mammals account for nearly a quarter of the entire record of Mesozoic mammals worldwide. As is true for most Cretaceous mammals from North America, Lancian mammals have been collected principally through use of underwater screenwashing and associated techniques (e.g., Lillegraven 1969). Thus, despite the enormous samples now known, most fossils are fragmentary, consisting largely of dentulous jaw fragments and isolated teeth. Identification and interpretation of such material must be approached with caution.

Many Lancian localities yielding individual specimens, faunules, or well-represented local faunas are known (see Webb 1998). Herein (figure 2.2, table 2.1), we treat only principal correlates or those that are of special interest for other reasons (i.e., geographic or stratigraphic distribution, availability of geochronologic data). Mammalian fossils are known from many localities in the type Lance Formation, but the majority of specimens comes from three main sites in upper parts of the unit (Clemens 1963, 1966, 1973). Additional specimens and faunules from the Lance Formation are known from several nearby areas not far to the north of the type area (Clemens et al. 1979; Whitmore 1985; Whitmore and Martin 1986) and in southern Wyoming (Breithaupt 1982), the Bighorn Basin area (Webb 1998, 2001), and south-central Montana (Clemens et al. 1979).

The Hell Creek Formation of Montana and adjacent parts of the Dakotas is distinguished from the Lance Formation mainly on the basis of discontinuity in exposure between the two (Clemens 1963; Clemens et al. 1979). Many Lancian localities in the upper part of the Hell Creek Formation are known from vicinity of the Fort Peck Reservoir, Montana; the Flat Creek Local Fauna (principally from one of five included sites) is the best-sampled assemblage (Archibald 1982). Currently, G. P. Wilson (Museum of Paleontology, University of Califor-



**FIGURE 2.2** Principal Lancian localities, local faunas, and areal assemblages. Numbered areas are the same as those used in tables 2.1 and 2.2.

nia Berkeley) is having success in discovering and sampling mammalian local faunas from the middle and lower parts of the formation. Another cluster of sites in the upper Hell Creek Formation occurs to the southeast, in Carter and Fallon counties. Some, such as the Claw Butte and Blacktail local faunas, are well represented, including 25–30 mammalian species (Hunter and Archibald 2002). Only individual specimens or faunules have been reported from other localities (Clemens 1973; Clemens et al. 1979; Archibald 1982). The Muddy Tork Local Fauna, from the upper Hell Creek Formation, Williston Basin, eastern Montana, includes only nine mammalian taxa but is well documented through study of palynomorphs, non-marine molluscs, and paleomagnetic stratigraphy (Hunter et al. 1997).

Additional occurrences of Lancian mammals from the Hell Creek Formation are known from North Dakota

(Hoganson et al. 1994; Murphy et al. 1995; Hunter and Archibald 2002) and South Dakota (Wilson 1965, 1983). Mammals also are known from at least two localities in the Fox Hills Formation, South Dakota (Waage 1968; Wilson 1983). The Red Owl Local Fauna, Meade County, is in the lower part of the unit and therefore probably is equivalent, stratigraphically, to a part of the Fox Hills Formation in the area of the type Lance Formation, Wyoming (Wilson 1987). The Red Owl Local Fauna therefore is noteworthy because it is almost surely older than typical Lancian assemblages from upper parts of the Lance and Hell Creek formations.

Three principal correlative faunas are known from the prairies of Canada. The Trochu Local Fauna, Red Deer River Valley, Alberta, is in lower parts of the Scollard Formation (see review by Clemens et al. 1979). The fauna includes 22 mammalian varieties, several of which are represented by complete jaws (Lillegraven 1969). The other two faunas are from closely spaced sites on opposite sides of the Frenchman River, Cypress Hills region, Saskatchewan. The Gryde and Wounded Knee local faunas are diverse, well-represented assemblages from slightly different horizons in the upper Frenchman Formation (Fox 1989; Storer 1991). Two other localities in Saskatchewan, Fr-1 and Long Fall, have yielded mammals from the Ravenscrag or Frenchman formation (Johnston and Fox 1984; Fox 1989). The age and correlation of Fr-1 and Long Fall have been contentious; as discussed later in this chapter, they have been interpreted as Cretaceous by some workers and Paleocene by others. We refer both localities to the Puercan land mammal age on the basis of key mammalian fossils.

The most northerly known occurrence of Late Cretaceous mammals is from a site in the Prince Creek Formation, northern Alaska. Although only three mammalian species are yet known, the occurrence is notable for its high paleolatitude, which during the Cretaceous was greater than the present position near 70°N (Clemens 1995). Precise correlation of this northern local fauna with those to the south is bedeviled by faunal and floral provincialism and the lack of a formal definition of a boundary between the “Edmontonian” and Lancian. The site that has yielded mammals is estimated but not clearly demonstrated to lie stratigraphically below the major dinosaur-bearing sites near Ocean Point. Interbedded strata of tephra at the major dinosaur bearing sites yielded K–Ar and <sup>40</sup>Ar/<sup>39</sup>Ar age determinations of 69.1 ± 0.3 Ma (weighted mean of all analyses, Conrad et al. 1992). This radiometric age determination indicates that the Alaskan mammalian fauna is older than most Lancian faunas in the northern Western Interior. To the southwest of the

**TABLE 2.1** Major Areal Assemblages, Local Faunas, and Localities of the Lancian Land Mammal Age

Areas are numbered generally north to south then east to west and agree with the numbers used in figure 2.2 and table 2.2.

**Prince Creek Formation, Alaska**

0. Colville River (not shown on map)

Reference: Clemens 1995

**Scollard Formation, Alberta**

1. Trochu local fauna

Reference: Lillegraven 1969

**Frenchman Formation, Saskatchewan**

2. Wounded Knee local fauna

3. Gryde local fauna

References: Fox 1989; Storer 1991

**Hell Creek Formation, Montana and North Dakota**

4. Vicinity of Fort Peck Reservoir, Montana

5. Muddy Tork local fauna, Williston Basin, Montana

6. Powderville, Montana

7. Claw Butte local fauna, Montana

8. Blacktail local fauna, Montana

9. Localities in the Little Missouri badlands, Montana and North Dakota

References: Clemens et al. 1979; Archibald 1982; Lofgren 1995; Hunter and Pearson 1996; Hunter et al. 1997; Hunter and Archibald 2002

**Fox Hills Formation, South Dakota**

10. Iron Lightning

11. Red Owl local fauna

References: Waage 1968; Clemens et al. 1979; Wilson 1983, 1987

**Hell Creek Formation, South Dakota**

12. Joe Painter Quarry

13. Eureka Quarry

Reference: Wilson 1983

**Lance Formation, Wyoming**

14. Localities near Mule Creek Junction

15. Localities in Lance Creek drainage, type Lance Formation

16. Hewitt's Foresight

17. Black Butte Station

References: Clemens 1963, 1966, 1973; Breithaupt 1982; Whitmore 1985; Webb 1998, 2001

**Ferris Formation, Wyoming**

18. Localities in Hanna Basin

References: Eberle and Lillegraven 1998a, 1998b; Lillegraven and Eberle 1999

**Laramie Formation, Colorado**

19. Site in Weld County

Reference: Carpenter 1979

**North Horn Formation, Utah**

20. Localities on North Horn Mountain and in South Dragon Canyon

References: Clemens 1961; Cifelli and Muizon 1998; Cifelli et al. 1999a

**Kirtland Formation, New Mexico**

21. Alamo Wash local fauna, San Juan Basin

References: Lehman 1981; Flynn 1986

type Lance Formation, an assemblage of Lancian mammals from the Ferris Formation of the Hanna Basin, Wyoming, is particularly notable because the sequence also includes mammals of early Puercan age in unequivocal superposition (Eberle and Lillegraven 1998a, 1998b;

Lillegraven and Eberle 1999). Smaller Lancian faunules are known from the Laramie Formation, Colorado (Carpenter 1979), and North Horn Formation, Utah (Cifelli et al. 1999a). For many years only a few isolated teeth documented the mammalian fauna of the

Naashoibito Member of the Kirtland Formation, New Mexico (Flynn 1986). Isolated occurrences of single specimens that may be of Lancian age are known from the Kemp Clay Formation, Texas (Tokaryk 1987), and the Mount Laurel Formation, New Jersey (Krause and Baird 1979).

Despite this broad geographic spread, currently well-represented and published Lancian assemblages are confined to north-central parts of the Western Interior, so that biogeographic variation among the mammals remains difficult to assess. Dinosaurian and floral distributions suggest some degree of latitudinal provinciality, as was true for mammals in the older Judithian (Lehman 1987; Clemens 2002) and younger Puercan (Eberle and Lillegraven 1998a) ages. Some geographic variation of mammals is seen among the well-sampled local faunas of central and northern parts of the Western Interior. Eutherians, for example, are generally more abundant and diverse in northerly assemblages, such as Trochu and Wounded Knee (see Hunter and Pearson 1996; Clemens 2002). The Flat Creek Local Fauna of Montana is geographically intermediate between Trochu (to the north) and the type Lance (to the south), and this is reflected in distribution of mammalian species among the three faunas (Archibald 1982).

## DEFINITION AND CHARACTERIZATION

Definition of the beginning of the Lancian age through designation of an FAD is problematic. The principal problem is questionable occurrences in older faunas, especially inadequate knowledge and characterization of the preceding "Edmontonian." Local ancestry for most first-appearing Lancian taxa appears probable. However, Weil and Clemens (1998; see also Clemens 2002) identify the eutherian *Batodon* and the marsupial *Glasbius* as Lancian "aliens" that lack records in preceding faunas of the northern Western Interior. To this list we add the multituberculate *Essonodon*, which appears to be closely related to known North American taxa (Weil 1999) but nonetheless is highly distinctive and has no known close morphologic antecedent in preceding faunas. We do not formally propose an FAD for the Lancian, but *Batodon*, *Glasbius*, and *Essonodon* appear to be the best candidates based on evidence in hand. Onset of the succeeding Puercan age is defined by first appearance of the archaic ungulatormorph *Protungulatum donnae* (see Archibald and Lofgren 1990; Eberle and Lillegraven 1998a, 1998b; Clemens 2002; and chapter 3). We provisionally characterize the Lancian here (also see tables 2.2 and 2.3). Note in what follows that taxa listed as possible appearances or occurrences may appear in more than one category.

First appearances: *Cimexomys minor*, *Parectypodus*, *Alphadon jasoni*, and *Gypsonictops illuminatus*; possible other first appearances: *Essonodon*, *Mesodma formosa*, *M. hensleighi*, *Neoplagiulax*, *Kimbetohia campi*, *Pedionomys elegans*, "P." *krejci*, *Cimolestes cerberoides*, and *C. stirtoni*

Last appearances: *Paracimexomys*, *P. priscus*, *Meniscoessus collomensis*, *M. conquistus*, cf. *Deltatheridium* sp., *Alphadon wilsoni*, *Protalphadon*, *P. lulli*, *Turgidodon*, *T. rhaister*, "P." *Pedionomys* *cooki*, "P." *hatcheri*, "P." *krejci*, Stagodontidae, and *Didelphodon*; possible other last appearances: Cimolomyidae, *Cimolomys*, *C. gracilis*, *Meniscoessus robustus*, *Cimolestes cerberoides*, *C. stirtoni*, and *Paranyctoides*

Unique occurrences: *Bubodens magnus*, *Cimolomys trochuus*, *Essonodon browni*, *Meniscoessus seminoensis*, *Clemensodon*, *C. megaloba*, ?*Neoplagiulax burgessi*, *Parectypodus foxi*, *Alphadon eatoni*, *Glasbius*, *G. intricatus*, *G. twitchelli*, *Protalphadon foxi*, *Turgidodon petaminis*, "P." *Pedionomys* *florenceae*, *Didelphodon padanicus*, *D. vorax*, *Cimolestes incisus*, *C. magnus*, *C. propalaeoryctes*, *Telacodon*, *T. laevis*, *Gypsonictops hypoconus*, *Batodon*, *B. tenuis*, *Alostera*, and *A. saskatchewanensis*; possible other unique occurrences: *Essonodon*, "P." *Pedionomys* *hatcheri*, "P." *krejci*, *Cimolestes cerberoides*, and *C. stirtoni*

## AGE RELATIONSHIPS BETWEEN LANCIAN LOCAL FAUNAS

With a few exceptions noted earlier, Lancian local faunas are highly similar in composition across northwestern North America. Some of the few known differences may reflect age variation between sites rather than biogeographic provinciality or geographic distance (Hunter and Archibald 2002), but it is not yet possible to propose a defensible zonation to subdivide the Lancian land mammal age. Detailed, multidisciplinary studies of age relationships have been conducted for Lancian and Puercan faunas in the Hell Creek and Tullock formations near Ft. Peck Reservoir, Montana (see Clemens 2002 and references therein). Stratigraphic positions commonly are determined by thickness below the local Hell Creek–Tullock contact, with most Lancian sites being in upper parts of the Hell Creek Formation, 3.5 to 45 m below the contact.

Reference to the presumed K–T iridium anomaly has proven of limited utility in the area because its presence depends on local depositional settings. Iridium anomalies are preserved in lignites near the base of the Z coal complex (which defines the formational contact) at sev-



TABLE 2.2 Temporal Ranges for Judithian Through Puercan Mammals

TAXON	JU	"ED"	LA	PU		LOCALITIES
				PU1	PU2+	
Multituberculata						
Family <i>incertae sedis</i>						
<i>Bubodens</i>			X			11
<i>B. magnus</i>			X			11
? <i>Bryceomys</i>	X					
<i>Cimexomys</i>	X	X	X	X	X	4, 11, 15, 17
<i>C. gregoryi</i>	X					
<i>C. judithae</i>	X	X				
<i>C. minor</i>			X	X		4, 15, 16, 17
<i>Paracimexomys</i>	X	X	X			1, 2, 3, 4, 20
<i>P. priscus</i>	X	O	X			1, 2, 3, 4
<i>P. magnus</i>	X					
Cimolodontidae						
<i>Cimolodon</i>	X	X	X	X	X	0, 1, 2, 3, 4, 7, 8, 9, 11, 13, 14, 15, 18
<i>C. electus</i>	?	X				
<i>C. nitidus</i>	?	X	X	T		0, 1, 2, 3, 4, 7, 8, 9, 11, ?13, 14, 15, 16, 18
<i>C. similis</i>	?					
Cimolomyidae						
<i>Cimolomys</i>	X	X	X	X		1, 3, 4, 8, 13, 15, 19, 20
<i>C. clarki</i>	X					
<i>C. gracilis</i>		X	X	?T		1, 3, 4, 8, 13, 15, 16
<i>C. milliensis</i>	X					
<i>C. trochuus</i>			X			1
<i>Essonodon</i>		?	X			2, 4, 7, 15, 17, 21
<i>E. browni</i>			X			4, 7, 15, 21
<i>Meniscoessus</i>	X	X	X	T		2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 17, 18, 19, 21
<i>M. collomensis</i>		X	X			19
<i>M. conquistus</i>		X	X			
<i>M. intermedius</i>	X	X				
<i>M. major</i>	X	X				
<i>M. robustus</i>		X	X	?T		2, ?3, 4, 5, 6, 7, 8, 9, 10, 11, 12, ?13, 14, 15, 16, 17
<i>M. seminoensis</i>			X			18
Eucosmodontidae						
<i>Clemensodon</i>			X			15
<i>C. megaloba</i>			X			15
<i>Stygimys</i>		X	O	X	X	
<i>S. cupressus</i>				X		
Neoplagiaulacidae						
<i>Mesodma</i>	X	X	X	X	X	1, 2, 3, 4, 5, 6, 7, 8, 9, 11, 12, 13, 15, 17, 20, 21
<i>M. formosa</i>	?	?	X	X	X	1, 2, 3, ?4, 7, 8, 11, 12, 13, 15, 16, 17, 18, 21
<i>M. hensleighi</i>	?	O	X	X		1, 3, 7, 8, 11, 12, ?13, 15, 16, 17, 18, ?20
<i>M. primaeva</i>	X					
<i>M. senecta</i>	?	?				
<i>M. thompsoni</i>		X	X	X	X	1, ?2, ?3, 4, ?5, 6, ?7, 8, 9, ?11, 12, 13, 15, 16, 17

TABLE 2.2 (continued)

TAXON	JU	"ED"	LA	PU		LOCALITIES
				PU1	PU2+	
<i>Neoplagiaulax</i>			?	O	X	?4
? <i>N. burgessi</i>			X			4, 16
<i>Parectypodus</i>			X	O	X	3
<i>P. foxi</i>			X			3
Ptilodontidae						
<i>Kimbetohia</i>	?		X	O	X	15
<i>K. campi</i>			X	O	X	15
Taeniolabididae						
<i>Catopsalis</i>				X	X	
<i>C. joyneri</i>				X	X	
<i>C. johnstoni</i>				X		
"Eupantotheria"						
Dryolestidae	X					
Boreosphenida, <i>incertae sedis</i>						
Deltatheridiidae						
cf. <i>Deltatheridium</i>	X	O	X			1, 15
Family <i>incertae sedis</i>						
<i>Falepetrus</i>	X					
<i>F. barwini</i>	X					
<i>Bistius</i>		X				
<i>B. bondi</i>		X				
<i>Palaeomolops</i>	X					
<i>P. langstoni</i>	X					
Marsupialia						
Alphadontidae						
<i>Aenigmadelphys</i>	X	X				
<i>A. archeri</i>	X					
<i>Alphadon</i>	X	X	X	T		1, 2, 3, 4, 7, 8, 9, 11, 12, 13, 17, 20, 21
<i>A. attaragos</i>	X					
<i>A. eatoni</i>			X			20
<i>A. halleyi</i>	X	X				
<i>A. jasoni</i>			X	T		1, 2, 3, 4, 15, 16
<i>A. marshi</i>	X		X	T		1, 4, 7, 8, 9, 11, ?12, 15, 16, 17, 21
<i>A. perexiguus</i>	X					
<i>A. sahnii</i>	X					
<i>A. wilsoni</i>	?	X	X			1, 4, ?7, 8, 15
<i>Glasbius</i>			X			1, 4, 7, 15
<i>G. intricatus</i>			X			15, 16
<i>G. twitchelli</i>			X			1, 4, 7
<i>Protalphadon</i>	X	O	X			4, 7, 8, 11, 15, 18
<i>P. foxi</i>			X			4
<i>P. lulli</i>	X	O	X			7, 8, ?11, 15, 16, 18
<i>Turgidodon</i>	X	X	X			1, 3, 4, 5, 7, 8, 15
<i>T. lillegraveni</i>	X					
<i>T. madseni</i>	X					
<i>T. parapraesagus</i>		X				
<i>T. petaminis</i>			X			3
<i>T. praesagus</i>	X					
<i>T. rhaister</i>		X	X			1, 4, 5, 7, 8, 15, 16
<i>T. russelli</i>	X	X				

TABLE 2.2 (continued)

TAXON	JU	"ED"	LA	PU		LOCALITIES
				PU1	PU2+	
<i>Varalphadon</i>	X					
<i>V. wahweapensis</i>	X					
"Pediomyidae"						
<i>Aquiladelphis</i>	O	X				
<i>A. incus</i>	O	X				
<i>A. paraminor</i>		X				
"Pediomys"	X	X	X	T		1, 2, 3, 4, 5, 6, 7, 8, 9, ?10, 11, 12, 13, 14, 15, 17, 20
"P." <i>clemensi</i>	X					
"P." <i>cooki</i>	X	X	X			4, 7, 8, 11, 15, 16
"P." <i>elegans</i>	?	O	X	T		1, 2, 3, 4, 7, 8, 12, 13, 15, 16, 17
"P." <i>fasseti</i>		X				
"P." <i>florencae</i>			X			4, 5, ?6, 7, 8, 9, 13, 15, 16
"P." <i>hatcheri</i>	?	O	X			1, ?3, 4, 5, ?6, 7, 8, 11, 12, 13, 14, 15, 20
"P." <i>krejci</i>		?	X			1, 3, 4, ?5, 7, 8, ?11, 12, 15
"P." <i>prokrejci</i>	X					
Stagodontidae						
<i>Didelphodon</i>		X	X			1, 2, 3, 4, 5, 6, 8, 9, 11, ?12, 13, 14, 15
<i>D. coyi</i>		X				
<i>D. padanicus</i>			X			
<i>D. vorax</i>			X			1, 2, 3, 4, 5, 6, 8, 9, ?11, ?13, 14, 15, 16
<i>Eodelphis</i>	X	X				
<i>E. browni</i>	X					
<i>E. cutleri</i>	X					
Eutheria						
Arctocyonidae						
<i>Baioconodon</i>				X	X	
<i>Oxyprimus</i>				X		
<i>O. erikseni</i>				X		
<i>Protungulatum</i>				X	X	
<i>P. donnae</i>				X	X	
Cimolestidae						
<i>Cimolestes</i>	?	X	X	X	X	1, 2, 3, 4, 8, 9, ?14, 15, 18
<i>C. cerberoides</i>			X	?T		1, ?4
<i>C. incisus</i>			X			?2, 3, 4, 8, 15, 16
<i>C. magnus</i>			X			1, 3, 4, 9, 15, 16
<i>C. propalaeoryctes</i>			X			1, ?2, 4, 8, 16
<i>C. stirtoni</i>			X	?T		2, 4, 8, 15
<i>Procerberus</i>				X	X	
<i>P. formicarum</i>				X		
<i>Telacodon</i>			X			15
<i>T. laevis</i>			X			15
Gypsonictopidae						
<i>Gypsonictops</i>	X	X	X	T		0, 1, 2, 3, 4, 6, 7, 8, 9, 11, 12, 13, 15, 17, 18, 19
<i>G. clemensi</i>		X				
<i>G. hypoconus</i>			X			1, 4, 7, 11, 12, 13, 15, 16, 17
<i>G. illuminatus</i>			X	T		1, ?2, 3, 4, ?6, 9, 16
<i>G. lewisi</i>	X	?				

TABLE 2.2 (continued)

TAXON	JU	"ED"	LA	PU		LOCALITIES
				PU1	PU2+	
Periptychidae						
<i>Mimatuta</i>				X	X	
Soricomorpha, family <i>incertae sedis</i>						
<i>Batodon</i>			X			1, 3, 4, 7, 8, 15
<i>B. tenuis</i>			X			1, 3, 4, 7, 8, 15
<i>Paranyctooides</i>	X	X	?			?8
<i>P. maleficus</i>	X					
<i>P. megakeros</i>	X					
<i>P. sternbergi</i>	X	?				
Ungulatomorpha, family <i>incertae sedis</i>						
<i>Alostera</i>			X			1, 2, 3, 4
<i>A. saskatchewanensis</i>			X			1, 2, 3, 4
<i>Avitotherium</i>	X					
<i>A. utahensis</i>	X					
<i>Gallolestes</i>	X	X				
<i>G. agujaensis</i>	X					
<i>G. pachymandibularis</i>		X				

Puercan taxa include only those also known from older faunas or those appearing in the earliest Puercan. See table 2.1 for references and suites of localities.

"Ed," "Edmontonian"; Ju, Judithian; La, Lancian; Pu (and first two subdivisions), Puercan.

X, presence; O, absence; ?, questionable occurrence; T, otherwise Lancian taxon with Puercan occurrence only in one of the transitional faunas of Saskatchewan (Long Fall, Fri).

eral sites near Hell Creek; a weighted mean age determination for the horizon is  $65.14 \pm 0.04$  Ma (Swisher et al. 1993). However, the base of the Z coal complex is not synchronous from place to place; it is about 200 k.y. younger to the east, in vicinity of McGuire Creek, where it overlies Puercan faunas recovered from the uppermost Hell Creek Formation. In absence of the iridium anomaly (which often has been assumed to be synchronous on a hemispheric basis), palynological criteria have been used to recognize the local Cretaceous–Tertiary boundary (Lofgren 1995). As far as is known, Lancian assemblages in vicinity of the Ft. Peck Reservoir lie within the youngest Late Cretaceous palynological zone of the Western Interior, the *Wodehousia spinata* Assemblage Zone, and in sediments deposited during magnetic polarity chron 29r. A chemostratigraphic method of correlation, based on carbon isotope excursions before and associated with the K–T boundary, gives great promise of providing precise correlations of terrestrial sections in the Western Interior (Arens and Jahren 2000). In the Dakotas, Montana, and adjacent areas the Hell Creek Formation probably spans about 1.75 m.y. (Hicks et al. 1999a), but the time represented by the better sampled Lancian faunas in the area is far less, insofar as these known localities are concentrated in upper parts of the unit.

Some Lancian faunas from the Hell Creek Formation further to the east (in easternmost Montana and southwestern North Dakota) appear to be somewhat older, representing younger parts of magnetic polarity chron 30n (Hunter et al. 1997; Hicks et al. 1999a). Where correlation to the GPTS is known or can be reasonably inferred, Lancian faunas from most other units also appear to lie within strata representing chron 29r or the younger parts of chron 30n (table 2.4). All known Lancian faunas are older than the generally accepted Cretaceous–Tertiary boundary, placed at 65.5 Ma by Obradovich and Hicks (1999). Given the age of chron 30n (65.58–67.61 Ma; Berggren et al. 1995), a maximum duration for the Lancian, based on these limited data from the Hell Creek Formation, would be about 2 m.y. To our knowledge, the only radioisotopic date that in part constrains the age of the Lancian is a determination of  $66.8 \pm 1.1$  Ma from the Kneehills Tuff, considered to represent the base of the *Triceratops* zone in the lower part of the Scollard Formation (Obradovich 1993). Again it must be stressed that establishment of a boundary between the Lancian and the older "Edmontonian" remains a challenge for future research.

The two Lancian mammal localities in the Fox Hills Formation of South Dakota—Red Owl Quarry (Wilson 1983, 1987) and the Iron Lightning locality (Waage 1968)

**TABLE 2.3** Taxonomic List of Lancian Mammals and Their Biochronologic Significance

Multituberculata family <i>incertae sedis</i>	<i>A.marshi</i>
<i>Bubodens</i>	<i>A. wilsoni</i> (*L)
<i>B.magnus</i> (U)	<i>Glasbius</i>
<i>Cimexomys</i>	<i>G. intricatus</i> (U)
<i>C. minor</i> (*F)	<i>G. twitchelli</i> (*U)
<i>Paracimexomys</i> (*L)	<i>Protalphadon</i> (*L)
<i>P. priscus</i> (*L)	<i>P. foxi</i> (U)
Cimolodontidae	<i>P. lulli</i> (*L)
<i>Cimolodon</i>	<i>Turgidodon</i> (*L)
<i>C.nitidus</i>	<i>T. petaminis</i> (U)
Cimolomyidae	<i>T. rhaister</i> (*L)
<i>Cimolomys</i>	“Pediomyidae”
<i>C. gracilis</i> (*?L)	“Pediomys”
<i>C.trochuus</i> (U)	“P.” <i>cooki</i> (*L)
<i>Essonodon</i> (?U)	<i>P. elegans</i> (*?F)
<i>E. browni</i> (*U)	“P.” <i>florenceae</i> (*U)
<i>Meniscoessus</i>	“P.” <i>hatcheri</i> (?U)
<i>M. collomensis</i> (L)	“P.” <i>krejicii</i> (?U)
<i>M. conquistus</i> (L)	Stagodontidae (*L)
<i>M. robustus</i> (*?L)	<i>Didelphodon</i> (*L)
<i>M. seminoensis</i> (U)	<i>D. padanicus</i> (U)
Eucosmodontidae	<i>D. vorax</i> (*U)
<i>Clemensodon</i>	Eutheria
<i>C. megaloba</i> (U)	Cimolestidae
Neoplagiulacidae	<i>Cimolestes</i>
<i>Mesodma</i>	<i>C. cerberoides</i> (?U)
<i>M. formosa</i> (*?F)	<i>C. incisus</i> (*U)
<i>M. hensleighi</i> (*?F)	<i>C. magnus</i> (*U)
<i>M.thompsoni</i>	<i>C. propalaeoryctes</i> (*U)
<i>Neoplagiulax</i> (?F)	<i>C. stirtoni</i> (?U)
? <i>N. burgessi</i> (U)	<i>Telacodon</i>
<i>Parectypodus</i> (F)	<i>T. laevis</i> (U)
<i>P. foxi</i> (U)	Gypsonictopidae
Ptilodontidae	<i>Gypsonictops</i>
<i>Kimbetohia</i>	<i>G. hypoconus</i> (*U)
<i>K. campi</i> (?F)	<i>G. illuminatus</i> (*F)
Boreosphenida, <i>incertae sedis</i>	Soricomorpha, family <i>incertae sedis</i>
Deltatheridiidae	<i>Batodon</i>
cf. <i>Deltatheridium</i> sp. (L)	<i>B. tenuis</i> (*U)
Marsupialia	cf. <i>Paranyctooides</i> (?L)
Alphadontidae	Ungulatomorpha, family <i>incertae sedis</i>
<i>Alphadon</i>	<i>Alostera</i>
<i>A. eatoni</i> (U)	<i>A. saskatchewanensis</i> (*U)
<i>A. jasoni</i> (*F)	

Taxa not surely identified are excluded unless their occurrences are significant. First (F), last (L), and unique (U) occurrences in the Lancian are designated for supraspecific taxa only when their occurrences differ from those of included species. F, L, and U occurrences are queried if the respective taxon is tentatively recorded from another land mammal age; asterisked taxa are known from more than one Lancian assemblage and therefore are more useful biostratigraphically.

**TABLE 2.4** Correlation of Lancian Faunas to the Geomagnetic Polarity Time Scale

LITHOLOGIC UNIT AND LOCATION	TEMPORAL CORRELATION AND REFERENCES
Scollard Formation, Alberta	According to Lerbekmo et al. (1979) and Lerbekmo (1985), Cretaceous strata of Scollard Formation in Red Deer Valley, Alberta, are in younger half of chron 30n and 29r.
Hell Creek Formation, vicinity of Fort Peck Reservoir, Montana	Chron 29r; see Clemens (2002) and references therein.
Hell Creek Formation, Muddy Tork local fauna, Montana	Younger part of chron 30n (Hunter et al. 1997).
Hell Creek Formation, SW North Dakota (loc. PTRM-V92067 of Hunter and Pearson 1996)	Youngest chron 30n (K. R. Johnson, pers. comm., 2000; Hicks et al. 1999a).
Lance Formation, Wyoming	According to Keating and Helsey (1983), strata of the Lance Formation in east-central Wyoming are of reversed polarity, but no mention is made of correlation to the Geomagnetic Polarity Time Scale or to chron 29r.
Upper part of Laramie Formation, Weld County, Colorado	No study was done in Weld County, but paleomagnetic data from elsewhere in the Denver Basin (Castle Pines core and Jimmy Camp Creek Section) suggest normal polarity rocks, probably representing C30n or possibly C31n (K. R. Johnson, pers. comm. 2000).
Fox Hills Formation, South Dakota	Indirect correlation to section at Red Bird, Wyoming (this chapter), suggests placement of Red Owl and Iron Lightning faunas in chron 31N (see Wilson 1987; Hicks et al. 1999b).
Alamo Wash local fauna, Naashoibito Member of Kirtland Formation, San Juan Basin, New Mexico	Chron 29r (see Butler and Lindsay 1985; Flynn 1986).

Locality suites listed in table 2.1 but not appearing here lack available paleomagnetic information.

warrant further discussion. Indirect evidence suggests that both sites may be older than other Lancian local faunas in the Western Interior and consequently may be useful for constraining the earlier age limit of the Lancian NALMA. The Iron Lightning locality is in the type section for the Iron Lightning Member (Waage 1968), and Red Owl Quarry is in the Fairpoint Member (Wilson 1987), which “is equivalent to at least part of the Colgate and Bullhead Members and perhaps to the Trail City and Timber Lake Members farther east” (Pettyjohn 1967:1367). Red Owl Quarry is lower in the local section than the Iron Lightning locality (Wilson 1983). Wilson (1987) surmised that strata containing Red Owl Quarry probably are equivalent to some part of the Fox Hills Formation in the Lance Creek area and that the mammalian fauna from Red Owl therefore is older than that of the type Lance Formation.

Correlation of the ammonite zonation preserved in members of the Fox Hills Formation to the GPTS (Gradstein et al. 1995) appears to support Wilson’s (1987) interpretation. The *Jeletzkytes nebrascensis* Range Zone, the youngest ammonite zone in the Western Interior, begins with the first appearance of *J. nebrascensis* in the uppermost Trail City Member and extends through the Iron Lightning Member of the upper Fox Hills Formation (Cobban and Reeside 1952; Landman and Waage 1993). Landman and Waage report that the *J. nebrascensis* Range

Zone locally extends into the basal Hell Creek Formation. According to Gradstein et al. (1995, figure 8), the *J. nebrascensis* Range Zone occurs within C31. At Red Bird, approximately 40 km northeast of Lance Creek, Hicks et al. (1999b) place the C31r–C31n boundary in the upper part of the Fox Hills Formation, within the *Hoploscaphites birkelundi* Range Zone (see Landman and Waage 1993, called the *H. aff. nicoletti* Range Zone by Gradstein et al. 1995), and estimate the age of the boundary to be  $69.01 \pm 0.5$  Ma. The *J. nebrascensis* Range Zone overlies the *H. birkelundi* Range Zone and thus presumably lies within C31n.

As discussed earlier, other Lancian localities for which correlation to the GPTS has been proposed are placed in the younger part of C30n or in C29r, thus suggesting that the mammalian assemblages of the Iron Lightning Formation are somewhat older. Given an estimated 1.02-m.y. duration for C31n (Berggren et al. 1995) and an estimated age of 69.01 Ma for the C31r–C31n boundary (Hicks et al. 1999b), the mammals of the Fox Hills Formation would fall within the range 69.01–67.99 Ma. Thus if the ammonite-based correlation of the Fox Hills strata were to be confirmed (a possibility reflected in figure 2.1), the 2-m.y. duration of the Lancian would be nearly doubled. The mammalian fauna of the Iron Lightning locality is poorly known, including four varieties, only one of which (the com-

mon Lancian multituberculate *Meniscoessus robustus*) is identified at the species level (Clemens et al. 1979). Fourteen kinds of mammals have been reported from Red Owl Quarry (Wilson 1983, 1987). None of these suggests a temporal difference from other Lancian faunas. One species, the large multituberculate *Bubodens magnus*, is endemic to Red Owl Quarry, however. Specimens referred to another multituberculate, *Meniscoessus robustus*, have somewhat smaller average dimensions than those from other Lancian localities (Eberle and Lillegraven 1998b). Wilson (1983) also noted the absence at Red Owl of the marsupial *Pediomys elegans* as unusual.

### PROBLEMATIC LANCIAN-PUERCAN FAUNAS

Several mammalian faunas or assemblages include a mixture of species that are typical of the Lancian on one hand and Puercan on the other. Most celebrated are the Bug Creek assemblages, from several localities in northeastern Montana. When first described, they were interpreted as a temporally ordered series (from oldest to youngest: Bug Creek Anthills, Bug Creek West, Harbicht Hill), apparently documenting appearance of mammalian taxa typical of the Paleocene during latest Cretaceous time (Sloan and Van Valen 1965). Subsequently, the issue of ecological versus temporal differences between Cretaceous and Tertiary faunas (see Matthew 1937) was reopened, and the Bug Creek assemblages were then suggested to represent a faunal facies that was contemporaneous with "typical" Lancian faunas (see also Clemens et al. 1979; Archibald 1982). Still later, the Bug Creek assemblages were assigned to their own, slightly younger land mammal age, the "Bugcreekian" (Archibald 1987b; Sloan 1987). Subsequently, the "Bugcreekian" mammal age was abandoned, with the Bug Creek assemblages being placed within the Puercan NALMA, and *Protungulatum donnae* was recognized as the FAD for advent of the Puercan's initial interval zone (Archibald and Lofgren 1990). The original Bug Creek "faunas" and their correlatives (see Sloan et al. 1986, Rigby 1989, and Lofgren 1995 for correlatives) are now interpreted to represent time-averaged assemblages that include both Lancian and Puercan elements mixed together through reworking caused by the incision of large Paleocene channels into fossiliferous Cretaceous strata (Lofgren 1995). The result of this reworking is that older Lancian fossils are present as sedimentary particles in younger

channel fills, deposited during the Puercan, that also yield *Protungulatum* (see Lofgren 1995). Thus Lancian taxa in the original Bug Creek assemblages, and their correlatives probably are represented by reworked fossils. As a practical issue, these fossils must be ignored for purposes of biochronology because it cannot be determined to what extent reworking might have extended their apparent geologic range.

The other two problematic faunas, both from the Cypress Hills of southwestern Saskatchewan, also include a mixture of Lancian species with those that are more typical of the Puercan. The sites in question, Long Fall (Medicine Hat Brick and Tile Quarry) and Fr-1, have been interpreted as late Lancian in age (Johnston 1980; Johnston and Fox 1984; Fox 1989, 1997) and therefore were considered to have been faunally transitional between the Lancian and Puercan NALMAs. Long Fall lies about 3 m stratigraphically below another site, Rav W-1 (Lerbekmo 1985), the latter of which yielded an assemblage of Puercan mammals.

Sloan (1987) interprets Long Fall as earliest Paleocene in age. Similarly, Lerbekmo (1985) considers both Long Fall and Rav W-1 to be in the same point bar deposit in the Frenchman Formation and suggests that the Lancian fossils were reworked during Puercan time (see also Lofgren 1995). Lerbekmo's (1985) interpretation was vigorously challenged by Fox (1989, 1997), however, who argued that the sites are separated by a disconformity and that both lie in the Ravenscrag Formation. The regional, traditionally used marker bed for recognizing the Cretaceous-Tertiary boundary, the Ferris (= No. 1) Coal Seam (Lerbekmo and Coulter 1985), unfortunately is not found in the section that included Long Fall and Rav w-1, and attempts to recover palynomorphs were unsuccessful (Fox 1997). Magnetostratigraphic studies by Lerbekmo (1985) place the Long Fall site within chron 29r, which spans both the Lancian-Puercan and globally recognized Cretaceous-Tertiary boundaries. Unfortunately, the two localities have since been destroyed through mining operations (Fox 1997), so questions regarding the age of Long Fall remain unanswered.

The Fr-1 site lies unequivocally in the Frenchman Formation. The Ferris Coal Seam is absent from vicinity of this site as well, and no sampling for iridium concentrations has been done. However, palynomorphs that are typical of strata in the Western Interior considered to be of Maastrichtian age have been recovered from Fr-1 (Fox 1997). In addition, articulated dinosaur remains have been reported from what appears to represent the same horizon several hundred meters east of Fr-1 (Fox 1989). Also, the relative abundance of mammals typical of the

Lancian at Fr-1 is high (Fox 1989), much higher than in the mixed assemblages from the Bug Creek area of Montana. These relationships make it difficult to dismiss the mixture of Lancian and Puercan mammals at Fr-1 as having resulted from reworking (see criteria and discussion by Lofgren 1995).

We assign both the Long Fall and Fr-1 localities to the Puercan land mammal age on the basis of presence of *Protungulatum* cf. *P. donnae* (see discussion by Eberle and Lillegraven 1998a); at least two other taxa characteristic of the Puercan are known from Fr-1, and many more, including a moderate diversity of archaic ungulates, are known from the Long Fall site (Fox 1989). But both sites also include several varieties of vertebrates that are otherwise known only from the Lancian (see table 2.2). Taken at face value, available data suggest the working hypotheses that (1) one or both of these transitional assemblages are of Cretaceous age and are therefore older than Puercan fossils from elsewhere (notably the Tullock Formation and the uppermost strata of the Hell Creek Formation in some areas of Montana); (2) the beginning of Puercan time is diachronous between the prairie provinces and northern Montana, at the level of resolution currently available; and (3) *Protungulatum* and a few other mammalian genera thought to be exclusively Paleocene in age apparently occur in the Late Cretaceous of southwestern Saskatchewan.

### PROSPECTS FOR FUTURE RESEARCH ON THE LANCIAN AGE

Definition of the onset of Lancian time continues to be problematic because of inadequate knowledge and characterization of the preceding, informally recognized “Edmontonian.” This is particularly unfortunate because it has direct bearing on the duration of the Lancian and therefore of the magnitude of faunal change through that age. Such information would provide an important baseline for comparison to apparently much greater evolutionary rates for mammals during the Puercan. Field research focused on lower parts of the Lance, Ferris, Hell Creek, and contemporaneous formations and older units is a priority.

Problems of correlation of the Lancian–Puercan boundary (see chapter 3) with the Cretaceous–Tertiary boundary have resulted from a wealth of data and application of different correlation techniques. In the northern Western Interior, Brown’s (1952) widely applied “formula” placed the Cretaceous–Tertiary boundary at the base of the first regionally continuous lignite found strati-

graphically above the last record of dinosaurs. Subsequently, biostratigraphic criteria for recognition of the Cretaceous–Tertiary boundary (e.g., marked changes in palynofloras) have been used. On a global basis, mineralogic and chemostratigraphic evidence of the impact of an extraterrestrial body, as well as radiometric age determinations, have been used in attempts to correlate the Cretaceous–Tertiary boundary (as defined at the stratotype in Tunisia) with particular levels in sections in the Western Interior. Changes in carbon isotope ratios are providing another tool for long-range correlations (Arens and Jahren 2000).

Given the increased precision of correlation methods, it is not surprising that formation boundaries long held to mark the Cretaceous–Tertiary boundary (e.g., the Hell Creek–Tullock boundary) can now be shown to be time transgressive (Lofgren 1995). An example of Puercan local faunas of possible Late Cretaceous age was discussed in a preceding section of this chapter. As Lillegraven and Eberle (1999) emphasize, although the Lancian–Puercan boundary appears to approximate the Cretaceous–Tertiary boundary, greater care must be exercised in defining units and their boundaries and applying different methods of correlation with the stratotype of the Cretaceous–Tertiary boundary.

One of the most serious shortcomings of the Lancian as a provincial time term has been restriction of its definition to faunas in north-central parts of the Western Interior. Only a few minor faunules were known outside this region and, as a result, little was known of biogeographic variation. Given the clearly recognized latitudinal and biogeographic differences that characterize Judithian and Puercan faunas (Weil 1999; Clemens 2002), the hypothesis that such provinciality also characterized Lancian mammals—as apparently was the case with dinosaurs (Lehman 1987)—was open to testing.

Recent discoveries by Weil and Williamson (2000) provide the first extensive sample of a mammalian fauna in the uppermost Kirtland Formation (Naashoibito Member) of the San Juan Basin, New Mexico. In their initial report on this collection they document the common occurrence of the multituberculate *Essonodon*, whose first occurrence has been suggested as a FAD for the Lancian; and as yet unidentified therian mammals. The composition of the mammalian fauna is reported to be distinctly different from that of Lancian faunas in the northern Western Interior. Additional collecting in the San Juan Basin, along with a geographically expanded program of sampling of Lancian and older Late Cretaceous formations, will provide additional insights into biogeographic deployment of highly distinctive taxa such as *Essonodon*



and *Glasbius*. These genera appear in the fossil record without obvious antecedents, and it will be important to determine the extent to which appearance of such taxa in different parts of the continent was synchronous.

Renewed field investigations in known fossiliferous Lancian units in southerly realms such as Colorado, Utah, New Mexico, and Texas offer promise for more complete documentation of Lancian biogeography in western North America. Extending still farther south, Mexico's highly diverse Late Cretaceous dinosaurian faunas show marked similarities to correlatives to the north (Ferrusquia-Villafranca 1998). To what extent do Lancian mammalian assemblages show faunal continuity between Canada and Mexico?

Compositions of North American Late Cretaceous faunas are providing pivotal data for analyses of macroevolutionary patterns of terrestrial vertebrates before and across the Cretaceous–Tertiary boundary. Debate swirls around conflicts between analyses of the available fossil record (suggesting that most of the major ordinal clades of mammals did not differentiate until after the Cretaceous–Tertiary boundary; Foote et al. 1999) and comparative molecular studies (suggesting much more ancient times of divergence; Kumar and Hedges 1998). Recent studies of the patterns of mammalian extinction at the Cretaceous–Tertiary boundary and restoration of their taxonomic diversity during the early Paleocene (Alroy 1999; Lillegraven and Eberle 1999; Clemens 2002) are based primarily, if not exclusively, on the fossil record available from the northern Western Interior.

How adequately does the Lancian and Puercan fossil record from the Western Interior represent the global pattern of evolution of mammalian faunas across the Cretaceous–Tertiary boundary? On a worldwide basis, a survey of known latest Cretaceous (Maastrichtian) mammalian faunas highlights significant additions since they were surveyed by Clemens et al. (1979). Various localities have been added to the record from the Western Interior, and new discoveries provide glimpses of contemporaneous faunas in eastern North America. Discoveries in Europe (e.g., Gheerbrant and Astibia 1999) are beginning to document Maastrichtian local faunas of distinctly different compositions. Increased exploration of Late Cretaceous and Paleocene faunas of South America (see Bonaparte 1996; Rougier et al. 2000) reveals endemism.

Unfortunately, we continue to lack detailed records of mammalian evolution across the Cretaceous–Tertiary boundary from continents other than North America. Knowledge of mammalian evolution during this interval is limited almost exclusively to a major window encompassing only the North American Western Interior and

much more limited data from South America and Europe. All of these records indicate that immigration from unsampled areas played major roles in establishing earliest Paleocene faunas on other continents. Despite contrary suggestions (e.g., Fara and Benton 2000), the incompleteness and geographic bias of the available fossil record must be considered in studies of patterns of mammalian evolution.

Studies of the mammalian fossil record in the Western Interior highlight the rapid rate of increase in taxonomic diversity and appearance of major groups of multituberculates and eutherian mammals in the Puercan. The heightened diversity becomes particularly clear during Pu<sub>2</sub> and Pu<sub>3</sub> times (see chapter 3 and Alroy 1999). Does this rapid increase in taxonomic diversity represent a high rate of evolutionary radiation of locally surviving lineages of multituberculates and eutherians after they were freed from dinosaurian tyranny? Or are we seeing effects of the immigration of new taxa from outside the Western Interior?

As noted earlier, the vertebrate fossil record of the northern Western Interior for approximately the last 2 million years of the Cretaceous suggests that its mammalian fauna remained stable in composition, even though the flora of the area changed markedly (Johnson and Hickey 1990). Most Lancian mammals were closely related to those known from Judithian strata; only a few genera appear to qualify as FADs for the Lancian land mammal age.

Analyses of phylogenetic relationships both of multituberculates and eutherians in earliest Puercan (Pu<sub>1</sub>, see chapter 3) local faunas show that most species are not closely related to known Lancian mammals (Weil and Clemens 1998; Clemens 2002). Unexpectedly, among Puercan representatives are various early ungulates that appear to be most closely related to Cretaceous forms known from Campanian local faunas in Texas and Baja California as well as members of older assemblages in Asia. Among the FADs for Pu<sub>1</sub>, the multituberculate *Stygimys kuszmauli* is not closely related to any known species in Lancian or Judithian local faunas of the northern Western Interior. However, that genus (if not the same species) is represented in the Campanian local fauna known from Baja California. Pre-Lancian records of primitive ungulatomorphs (such as *Gallolestes*) and the lipotyphlan soricomorph *Paranyctoides* suggest that the evolutionary radiation of eutherians had roots well within the Late Cretaceous and therefore was not solely the product of post-Cretaceous diversification.

In summary, both direct and indirect evidence (including analyses of phylogenetic relationships of immi-

grant taxa) points to extensive biogeographic diversification throughout Late Cretaceous mammalian faunas of the world. Although the specific groups of mammals differ, patterns of mammalian evolution across the Cretaceous–Tertiary boundary in other parts of the world show some basic similarities to those documented in the Western Interior. In the few areas currently sampled, Cretaceous extinctions appear to have terminated many lineages. Recovery of taxonomic diversity in the earliest Paleocene appears to have been the combined product of evolutionary radiations of locally surviving stocks and immigration of new groups. Although we may be approaching comprehensive knowledge of Lanciaan mammalian diversification in northern parts of the Western Interior, much remains to be discovered there and in other areas outside this small window of current knowledge.

#### ACKNOWLEDGMENTS

We are grateful to Donna Naughton, Canadian Museum of Nature, for help in constructing figure 2.1. This work was supported or benefited from research made possible by the following grants from the National Science Foundation: DEB-9870173 (to RLC) and EAR-8205211, EAR-9506462, and EAR-9909354 (to JAL).

#### REFERENCES

- Alroy, J. 1999. The fossil record of North American mammals: Evidence for a Paleocene evolutionary radiation. *Systematic Biology* 48:107–118.
- Archibald, J. D. 1982. A study of Mammalia and geology across the Cretaceous–Tertiary boundary in Garfield County, Montana. *University of California Publications in Geological Sciences* 122:1–286.
- . 1987a. Late Cretaceous (Judithian and Edmontonian) vertebrates and geology of the Williams Fork Formation, N.W. Colorado. In *Fourth symposium on Mesozoic terrestrial ecosystems*, ed. P. J. Currie and E. H. Koster. Drumheller, Alberta, Canada: Tyrrell Museum of Palaeontology, pp. 7–11.
- . 1987b. Latest Cretaceous and early Tertiary mammalian biochronology/biostratigraphy in the Western Interior. *Geological Society of America, Abstracts with Programs* 19(5):258.
- Archibald, J. D. and D. L. Lofgren. 1990. Mammalian zonation near the Cretaceous–Tertiary boundary. *Geological Society of America, Special Paper* 243:31–50.
- Arens, N. C. and A. H. Jahren. 2000. Carbon isotope excursion in atmospheric CO<sub>2</sub> at the Cretaceous–Tertiary boundary: Evidence from terrestrial sediments. *Palaeos* 15:314–322.
- Berggren, W. A., D. V. Kent, C. C. Swisher, and M.-P. Aubry. 1995. A revised Cenozoic geochronology and chronostratigraphy. *Society for Sedimentary Geology, Special Publication* 54:129–212.
- Bonaparte, J. F. 1996. Cretaceous tetrapods of Argentina. *Münchener Geowissenschaftliche Abhandlungen, Reihe A* 30:73–130.
- Breithaupt, B. H. 1982. Paleontology and paleoecology of the Lance Formation (Maastrichtian), east flank of Rock Springs uplift, Sweetwater County, Wyoming. *Contributions to Geology, University of Wyoming* 21:123–151.
- Brown, R. 1952. Tertiary strata in eastern Montana and western North and South Dakota. *Billings Geological Society Guidebook* 3:89–92.
- Butler, R. F. and E. H. Lindsay. 1985. Mineralogy of magnetic minerals and revised magnetic polarity stratigraphy of continental sediments, San Juan Basin, New Mexico. *Journal of Geology* 93:535–554.
- Butler, R. F., E. H. Lindsay, L. L. Jacobs, and N. M. Johnson. 1977. Magnetostratigraphy of the Cretaceous–Tertiary boundary in the San Juan Basin, New Mexico. *Nature* 267:318–323.
- Carpenter, K. 1979. Vertebrate fauna of the Laramie Formation (Maastrichtian), Weld County, Colorado. *Contributions to Geology, University of Wyoming* 17:37–49.
- Cifelli, R. L. 1990a. Cretaceous mammals of southern Utah. I. Marsupial mammals from the Kaiparowits Formation (Judithian). *Journal of Vertebrate Paleontology* 10:295–319.
- . 1990b. Cretaceous mammals of southern Utah. II. Marsupials and marsupial-like mammals from the Wahweap Formation (early Campanian). *Journal of Vertebrate Paleontology* 10:320–331.
- . 1990c. Cretaceous mammals of southern Utah. IV. Eutherian mammals from the Wahweap (Aquilan) and Kaiparowits (Judithian) formations. *Journal of Vertebrate Paleontology* 10:346–360.
- . 1994. Therian mammals of the Terlingua Local Fauna (Judithian), Aguja Formation, Big Bend of the Río Grande, Texas. *Contributions to Geology, University of Wyoming* 30:117–136.
- . 1999. Tribosphenic mammal from the North American Early Cretaceous. *Nature* 401:363–366.
- Cifelli, R. L. and J. G. Eaton. 1987. Marsupial from the earliest Late Cretaceous of western US. *Nature* 325:520–522.
- Cifelli, R. L. and C. L. Gordon. 1999. Symmetrodonts from the Late Cretaceous of southern Utah, and comments on the distribution of archaic mammalian lineages persisting into the Cretaceous of North America. *Geology Studies, Brigham Young University* 44:1–16.
- Cifelli, R. L., J. I. Kirkland, A. Weil, A. L. Deino, and B. J. Kowallis. 1997. High-precision <sup>40</sup>Ar/<sup>39</sup>Ar geochronology and the advent of North America's Late Cretaceous terrestrial fauna. *Proceedings of the National Academy of Sciences USA* 94:11163–11167.
- Cifelli, R. L. and S. K. Madsen. 1986. An Upper Cretaceous symmetrodont (Mammalia) from southern Utah. *Journal of Vertebrate Paleontology* 6:258–263.
- . 1998. Triconodont mammals from the medial Cretaceous of Utah. *Journal of Vertebrate Paleontology* 18:403–411.
- Cifelli, R. L. and C. de Muizon. 1998. Marsupial mammal from the Upper Cretaceous North Horn Formation, central Utah. *Journal of Paleontology* 72:532–537.
- Cifelli, R. L., R. L. Nydam, J. G. Eaton, J. D. Gardner, and J. I. Kirkland. 1999a. Vertebrate faunas of the North Horn Formation (Upper Cretaceous–lower Paleocene), Emery and Sanpete counties. In *Vertebrate paleontology in Utah*, ed. D. D. Gillette. Salt Lake City: Utah Geological Survey, Special Publication 99-1, pp. 377–388.
- Cifelli, R. L., R. L. Nydam, A. Weil, J. D. Gardner, J. I. Kirkland, J. G. Eaton, and S. K. Madsen. 1999b. Vertebrate fauna of the upper Cedar Mountain Formation (Cretaceous; Albanian–Cenomanian),

- Emery County: The Mussentuchit local fauna. In *Vertebrate fossils of Utah*, ed. D. D. Gillette. Salt Lake City: Utah Geological Survey, pp. 219–242.
- Clemens, W. A. 1961. A Late Cretaceous mammal from Dragon Canyon, Utah. *Journal of Paleontology* 35:578–579.
- . 1963. Fossil mammals of the type Lance Formation, Wyoming. Part I. Introduction and Multituberculata. *University of California Publications in Geological Sciences* 48:1–105.
- . 1966. Fossil mammals from the type Lance Formation Wyoming. Part II. Marsupialia. *University of California Publications in Geological Sciences* 62:1–122.
- . 1973. Fossil mammals of the type Lance Formation, Wyoming. Part III. Eutheria and summary. *University of California Publications in Geological Sciences* 94:1–102.
- . 1995. A latest Cretaceous, high paleolatitude mammalian fauna from the north slope of Alaska. In *Fifth symposium on Mesozoic terrestrial ecosystems and biota*, ed. Z. Kielan-Jaworowska, N. Heintz, and H. A. Nakrem. Oslo: Contributions from the Paleontological Museum, University of Oslo, no. 364, pp. 15–16.
- . 2002. Evolution of the mammalian fauna across the Cretaceous–Tertiary boundary in northeastern Montana and other areas of the Western Interior. In *The Hell Creek Formation and the Cretaceous–Tertiary boundary in the northern Great Plains: An integrated continental record of the end of the Cretaceous*, ed. J. H. Hartman, K. R. Johnson, and D. J. Nichols. Denver, CO: Geological Society of America Special Paper 361:217–245.
- Clemens, W. A., J. A. Lillegraven, E. H. Lindsay, and G. G. Simpson. 1979. Where, when, and what: A survey of known Mesozoic mammal distribution. In *Mesozoic mammals: The first two-thirds of mammalian history*, ed. J. A. Lillegraven, Z. Kielan-Jaworowska, and W. A. Clemens. Berkeley: University of California Press, pp. 7–58.
- Cobban, W. A. and J. B. Reeside Jr. 1952. Correlation of Cretaceous formations of the Western Interior of the United States. *Geological Society of America Bulletin* 63:1011–1044.
- Conrad, J. E., E. H. McKee, and B. D. Turrin. 1992. Age of tephra beds at the Ocean Point dinosaur locality, north slope, Alaska, based on K–Ar and  $^{40}\text{Ar}/^{39}\text{Ar}$  analyses. *U.S. Geological Survey Bulletin* 1990C:C1–C12.
- Diem, S. 1999. *Vertebrate faunal analysis of the Upper Cretaceous Williams Fork Formation, Rio Blanco County, Colorado*. M.S. thesis, San Diego State University, San Diego.
- Dorf, E. 1942. Upper Cretaceous floras of the Rocky Mountain region. II: Flora of the Lance Formation at its type locality, Niobrara County, Wyoming. *Carnegie Institute of Washington* 508:79–159.
- Eaton, J. G. 1987. *Stratigraphy, depositional environments, and age of Cretaceous mammal-bearing rocks in Utah, and systematics of the Multituberculata*. Ph.D. dissertation, University of Colorado, Boulder.
- . 1990. Stratigraphic revision of Campanian (Upper Cretaceous) rocks in the Henry Basin, Utah. *Mountain Geologist* 27:27–38.
- . 1991. Biostratigraphic framework for the Upper Cretaceous rocks of the Kaiparowits Plateau, southern Utah. *Geological Society of America Special Paper* 260:47–63.
- . 1993. Mammalian paleontology and correlation of the uppermost Cretaceous rocks of the Paunsaugunt Plateau, Utah. *Museum of Northern Arizona Bulletin* 59:163–180.
- . 1995. Cenomanian and Turonian (early Late Cretaceous) multituberculate mammals from southwestern Utah. *Journal of Vertebrate Paleontology* 15:761–784.
- . 1999a. Vertebrate paleontology of the Iron Springs Formation, Upper Cretaceous, southwestern Utah. In *Vertebrate paleontology in Utah*, ed. D. D. Gillette. Salt Lake City: Utah Geological Survey, Miscellaneous Publication 99-1, pp. 339–343.
- . 1999b. Vertebrate paleontology of the Paunsaugunt Plateau, Upper Cretaceous, southwestern Utah. In *Vertebrate paleontology in Utah*, ed. D. D. Gillette. Salt Lake City: Utah Geological Survey, Miscellaneous Publication 99-1, pp. 335–338.
- Eaton, J. G., R. L. Cifelli, J. H. Hutchison, J. I. Kirkland, and J. M. Parrish. 1999a. Cretaceous vertebrate faunas from the Kaiparowits Plateau. In *Vertebrate fossils of Utah*, ed. D. D. Gillette. Salt Lake City: Utah Geological Survey, pp. 345–354.
- Eaton, J. G., S. Diem, J. D. Archibald, C. Schierup, and H. Munk. 1999b. Vertebrate paleontology of the Upper Cretaceous rocks of the Markagunt Plateau, southwestern Utah. In *Vertebrate paleontology in Utah*, ed. D. D. Gillette. Salt Lake City: Utah Geological Survey, Miscellaneous Publication 99-1, pp. 323–333.
- Eaton, J. G., H. Munk, and M. A. Hardman. 1998. A new vertebrate fossil locality within the Wahweap Formation (Upper Cretaceous) of Bryce Canyon National Park and its bearing on the presence of the Kaiparowits Formation on the Paunsaugunt Plateau. *Technical Reports, National Park Service* 98/1:36–40.
- Eberle, J. J. and J. A. Lillegraven. 1998a. A new important record of earliest Cenozoic mammalian history: Eutheria and paleogeographic/biostratigraphic summaries. *Rocky Mountain Geology* 33:49–117.
- . 1998b. A new important record of earliest Cenozoic mammalian history: Geologic setting, Multituberculata, and Peradectia. *Rocky Mountain Geology* 33:3–47.
- Eberth, D. A. and A. P. Hamblin. 1993. Tectonic, stratigraphic, and sedimentologic significance of a regional discontinuity in the upper Judith River Group (Belly River wedge) of southern Alberta, Saskatchewan, and northern Montana. *Canadian Journal of Earth Sciences* 30:174–200.
- Emry, R. J., J. D. Archibald, and C. C. Smith. 1981. A mammalian molar from the Late Cretaceous of northern Mississippi. *Journal of Paleontology* 55:953–956.
- Fara, E. and M. J. Benton. 2000. The fossil record of Cretaceous tetrapods. *Palaio* 15:161–165.
- Fassett, J. E. and J. D. Obradovich. 1996. Subsidence/sedimentation rates as Campanian Pictured Cliffs shoreline regressed across San Juan Basin, New Mexico and Colorado based on five new  $^{40}\text{Ar}/^{39}\text{Ar}$  ages spanning 2.5 MY. *Geological Society of America, Abstracts with Programs* 28(1):66.
- Fassett, J. E. and M. B. Steiner. 1997. Precise age of C33n–C32r magnetic polarity reversal, San Juan Basin, New Mexico and Colorado. *New Mexico Geological Society Guidebook, 48th Field Conference, Mesozoic Geology and Paleontology of the Four Corners Region* 1997:239–247.
- Ferrusquia-Villafranca, L. 1998. Mexico's dinosaur record: Its paleogeographic significance. *Geological Society of America, Abstracts with Programs* A170.
- Fiorillo, A. R. 1989. The vertebrate fauna from the Judith River Formation (Late Cretaceous) of Wheatland and Golden Valley counties, Montana. *The Mosasaur* 4:127–142.

- Fiorillo, A. R. and P. J. Currie. 1994. Theropod teeth from the Judith River Formation (Upper Cretaceous) of south-central Montana. *Journal of Vertebrate Paleontology* 14:74–80.
- Flynn, L. J. 1986. Late Cretaceous mammal horizons from the San Juan Basin, New Mexico. *American Museum Novitates* 2845:1–30.
- Foote, M., J. P. Hunter, C. M. Janis, and J. J. Sepkoski. 1999. Evolutionary and preservational constraints on origins of biologic groups: Divergence times of eutherian mammals. *Science* 283:1310–1314.
- Fox, R. C. 1971a. Early Campanian multituberculates (Mammalia: Allotheria) from the upper Milk River Formation, Alberta. *Canadian Journal of Earth Sciences* 8:916–938.
- . 1971b. Marsupial mammals from the early Campanian Milk River Formation, Alberta, Canada. In *Early mammals*, ed. D. M. Kermack and K. A. Kermack. *Zoological Journal of the Linnean Society, London*, pp. 145–164.
- . 1976a. Additions to the mammalian local fauna from the upper Milk River Formation (Upper Cretaceous), Alberta. *Canadian Journal of Earth Sciences* 13:1105–1118.
- . 1976b. Cretaceous mammals (*Meniscoessusintermedius*, new species, and *Alphadon* sp.) from the lowermost Oldman Formation, Alberta. *Canadian Journal of Earth Sciences* 13:1216–1222.
- . 1978. Upper Cretaceous terrestrial vertebrate stratigraphy of the Gobi Desert (Mongolian People's Republic) and western North America. In *Western and Arctic Canadian biostratigraphy*, ed. C. R. Stelck and D. E. Chatterton. Geological Association of Canada, Special Paper 18, pp. 577–594.
- . 1979a. Mammals from the Upper Cretaceous Oldman Formation, Alberta. I. *Alphadon* Simpson (Marsupialia). *Canadian Journal of Earth Sciences* 16:91–102.
- . 1979b. Mammals from the Upper Cretaceous Oldman Formation, Alberta. II. *Pediomys* Marsh (Marsupialia). *Canadian Journal of Earth Sciences* 16:103–113.
- . 1979c. Mammals from the Upper Cretaceous Oldman Formation, Alberta. III. Eutheria. *Canadian Journal of Earth Sciences* 16:114–125.
- . 1980a. Mammals from the Upper Cretaceous Oldman Formation, Alberta. IV. *Meniscoessus* Cope. *Canadian Journal of Earth Sciences* 17:1480–1488.
- . 1980b. *Picopsispattersoni*, n. gen. and sp., an unusual therian from the Upper Cretaceous of Alberta, and the classification of primitive tribosphenic mammals. *Canadian Journal of Earth Sciences* 17:1489–1498.
- . 1981. Mammals from the Upper Cretaceous Oldman Formation, Alberta. V. *Eodelphis* Matthew, and the evolution of the Stagodontidae. *Canadian Journal of Earth Sciences* 18:350–365.
- . 1982. Evidence of new lineage of tribosphenic therians (Mammalia) from the Upper Cretaceous of Alberta, Canada. *Géobios, Mémoire Spécial* 6:169–175.
- . 1984a. *Paranyctoidesmaleficus* (new species), an early eutherian mammal from the Cretaceous of Alberta. *Special Publication, Carnegie Museum of Natural History* 9:9–20.
- . 1984b. A primitive, “obtuse-angled” symmetrodont (Mammalia) from the Upper Cretaceous of Alberta, Canada. *Canadian Journal of Earth Sciences* 21:1204–1207.
- . 1987. An ancestral marsupial and its implications for early marsupial evolution. In *Fourth symposium on Mesozoic terrestrial ecosystems*, ed. P. J. Currie and E. H. Koster. Drumheller, Alberta, Canada: Tyrrell Museum, pp. 101–105.
- . 1989. The Wounded Knee local fauna and mammalian evolution near the Cretaceous–Tertiary boundary, Saskatchewan, Canada. *Palaeontographica, Abt. A* 208:11–59.
- . 1997. Upper Cretaceous and Tertiary stratigraphy and paleontology of southern Saskatchewan. In *Canadian paleontology conference, field trip guidebook no. 6*, ed. L. McKenzie-McAnally. St. John's, Newfoundland: Geological Association of Canada, pp. 70–85.
- Fox, R. C. and B. G. Naylor. 1986. A new species of *Didelphodon* Marsh (Marsupialia) from the Upper Cretaceous of Alberta, Canada: Paleobiology and phylogeny. *Neues Jahrbuch für Geologie und Paläontologie Abhandlungen* 172:357–380.
- Gheerbrant, E. and H. Astibia. 1999. The Upper Cretaceous mammals from Laño (Spanish Basque country). *Estudios del Museo de Ciencias Naturales de Alava* 14:295–323.
- Goodwin, M. B. and A. L. Deino. 1989. The first radiometric ages from the Judith River Formation (Upper Cretaceous), Hill County, Montana. *Canadian Journal of Earth Sciences* 26:1384–1391.
- Gradstein, F. M., F. P. Agterberg, J. G. Ogg, J. Hardenbol, P. Van Veen, J. Thierry, and Z. Huang. 1995. A Triassic, Jurassic and Cretaceous time scale. In *Geochronology, time scales and global stratigraphic correlation*, ed. W. A. Berggren, D. V. Kent, M.-P. Aubry, and J. Hardenbol. Tulsa: SEPM (Society for Sedimentary Geology) Special Publication, pp. 95–126.
- Grandstaff, B. S., D. C. Parris, and R. K. Denton. 1992. *Alphadon* (Marsupialia) and Multituberculata (Allotheria) in the Cretaceous of eastern North America. *Journal of Vertebrate Paleontology* 12:217–222.
- Heinrich, R. E., J. P. Hunter, and D. B. Weishampel. 1998. Mammals from the St. Mary River Formation (Upper Cretaceous), Montana: New insights on an “Edmontonian” land mammal age. *Journal of Vertebrate Paleontology* 18:50A.
- Hicks, J. F., K. R. Johnson, L. Tauxe, D. Clark, and J. D. Obradovich. 1999a. Geochronology of the Hell Creek Formation of southwestern North Dakota: A multidisciplinary approach using biostratigraphy, isotopic dating, geochemistry, and magnetic stratigraphy. *Geological Society of America, Abstracts with Programs* 30:A-71.
- Hicks, J. F., J. D. Obradovich, and L. Tauxe. 1999b. Magnetostratigraphy, isotope age calibration and intercontinental correlation of the Red Bird section of the Pierre Shale, Niobrara County, Wyoming, USA. *Cretaceous Research* 20:1–27.
- Hoganson, J. W., J. M. Campbell, and E. C. Murphy. 1994. Stratigraphy and paleontology of the Cretaceous Hell Creek Formation, Stumpf site, Morton County, North Dakota. *Proceedings of the North Dakota Academy of Science* 48:95.
- Hunter, J. P. and J. D. Archibald. 2002. Mammals from the end of the age of dinosaurs in North Dakota and southeastern Montana, with a reappraisal of geographic differentiation among Lancian mammals. In *The Hell Creek Formation and the Cretaceous–Tertiary boundary in the northern Great Plains: An integrated continental record of the end of the Cretaceous*, ed. J. H. Hartman, K. R. Johnson, and D. J. Nichols. Denver, CO: Geological Society of America, Special Paper 361:191–216.
- Hunter, J. P., J. H. Hartman, and D. W. Krause. 1997. Mammals and molluscs across the Cretaceous–Tertiary boundary from Makoshika State Park and vicinity (Williston Basin), Montana. *Contributions to Geology, University of Wyoming* 32:61–114.

- Hunter, J. P. and D. A. Pearson. 1996. First record of Lancian (Late Cretaceous) mammals from the Hell Creek Formation of southwestern North Dakota. *Cretaceous Research* 17:633–643.
- Johnson, K. R. and L. J. Hickey. 1990. Megafloal change across the Cretaceous/Tertiary boundary in the northern Great Plains and Rocky Mountains, U.S.A. *Geological Society of America, Special Paper* 247:433–444.
- Johnston, P. A. 1980. First record of Mesozoic mammals from Saskatchewan. *Canadian Journal of Earth Sciences* 17:512–519.
- Johnston, P. A. and R. C. Fox. 1984. Paleocene and Late Cretaceous mammals from Saskatchewan, Canada. *Palaeontographica, Abteilung A* 186:163–222.
- Keating, B. H. and C. E. Helsey. 1983. The magnetostratigraphy of the Cretaceous–Tertiary boundary in the continental Lance Formation and five marine sequences. *Eos, Transactions, American Geophysical Union* 64:219.
- Keller, G., L. Li, and N. MacLeod. 1995. The Cretaceous/Tertiary boundary stratotype section at El Kef, Tunisia: How catastrophic was the mass extinction? *Palaeogeography, Palaeoclimatology, Palaeoecology* 119:221–254.
- Krause, D. W. and D. Baird. 1979. Late Cretaceous mammals east of the North American Western Interior Seaway. *Journal of Paleontology* 53:562–565.
- Kumar, S. and S. B. Hedges. 1998. A molecular timescale for vertebrate evolution. *Nature* 392: 917–920.
- Landman, N. H. and K. M. Waage. 1993. Scaphitid ammonites of the Upper Cretaceous (Maastrichtian) Fox Hills Formation in South Dakota and Wyoming. *Bulletin of the American Museum of Natural History* 215:1–257.
- Leahy, G. D. and J. F. Lerbekmo. 1995. Macrofossil magnetostratigraphy for the upper Santonian–lower Campanian interval in the Western Interior of North America: Comparisons with European stage boundaries and planktonic foraminiferal zonal boundaries. *Canadian Journal of Earth Sciences* 32:247–260.
- Lehman, T. M. 1981. The Alamo Wash local fauna: A new look at the old Ojo Alamo fauna. In *Advances in San Juan Basin paleontology*, ed. S. G. Lucas, J. K. Rigby Jr., and D. Wolberg. Albuquerque: University of New Mexico Press, pp. 189–221.
- . 1987. Late Maastrichtian paleoenvironments and dinosaur biogeography in the Western Interior of North America. *Palaeogeography, Palaeoclimatology, Palaeoecology* 60:189–217.
- . 1997. Late Campanian dinosaur biogeography in the Western Interior of North America. *Dinofest International Proceedings 1997*:223–240.
- Lerbekmo, J. F. 1985. Magnetostratigraphic and biostratigraphic correlations of Maastrichtian to early Paleocene strata between south-central Alberta and southwestern Saskatchewan. *Bulletin of Canadian Petroleum Geology* 33:213–226.
- Lerbekmo, J. F. and K. C. Coulter. 1985. Late Cretaceous to early Tertiary magnetostratigraphy of a continental sequence: Red Deer Valley, Alberta, Canada. *Canadian Journal of Earth Sciences* 22:567–583.
- Lerbekmo, J. F., M. E. Evans, and H. Baadsgaard. 1979. Magnetostratigraphy, biostratigraphy, and geochronology of the Cretaceous–Tertiary boundary sediments, Red Deer Valley. *Nature* 279:26–30.
- Lillegraven, J. A. 1969. Latest Cretaceous mammals of upper part of Edmonton Formation of Alberta, Canada, and review of marsupial–placental dichotomy in mammalian evolution. *University of Kansas Paleontological Contributions* 50:1–122.
- . 1972. Preliminary report on Late Cretaceous mammals from the El Gallo Formation, Baja California del Norte, Mexico. *Natural History Museum of Los Angeles County, Contributions in Science* 232:1–11.
- . 1987. Stratigraphic and evolutionary implications of a new species of *Mensicoessus* (Multituberculata, Mammalia) from the Upper Cretaceous Williams Fork Formation, Moffat County, Colorado. In *Papers in vertebrate paleontology in honor of Morton Green*, ed. J. E. Martin and G. E. Ostrander. Rapid City: South Dakota School of Mines and Technology, *Dakoterra*, Special Paper 3, pp. 46–56.
- . 1991. Stratigraphic placement of the Santonian–Campanian boundary (Upper Cretaceous) in the North American Gulf Coastal Plain and Western Interior, with implications to global geochronology. *Cretaceous Research* 12:115–136.
- Lillegraven, J. A. and J. J. Eberle. 1999. Vertebrate faunal changes through Lancian and Puercan time in southern Wyoming. *Journal of Paleontology* 73:691–710.
- Lillegraven, J. A. and M. C. McKenna. 1986. Fossil mammals from the “Mesaverde” Formation (Late Cretaceous, Judithian) of the Bighorn and Wind River basins, Wyoming, with definitions of Late Cretaceous North American land-mammal “ages.” *American Museum Novitates* 2840:1–68.
- Lillegraven, J. A. and L. M. Ostresh Jr. 1990. Late Cretaceous (earliest Campanian/Maastrichtian) evolution of western shorelines of the North American Western Interior seaway in relation to known mammalian faunas. *Geological Society of America, Special Paper* 243:1–30.
- Lofgren, D. L. 1995. The Bug Creek problem and the Cretaceous–Tertiary transition at McGuire Creek, Montana. *University of California Publications in Geological Sciences* 140:1–185.
- Matthew, W. D. 1937. Paleocene fauna of the San Juan Basin, New Mexico. *Transactions of the American Philosophical Society*, new series 30:1–510.
- McNulty, C. J. and B. Slaughter. 1968. Stratigraphy of the Woodbine Formation, Tarrant County, Texas (locality 3). In *Field trip guidebook, Second Annual Meeting, South-Central Section of the Geological Society of America*, pp. 68–72.
- Montellano, M. 1988. *Alphadonhalleyi* (Didelphidae, Marsupialia) from the Two Medicine Formation (Late Cretaceous, Judithian) of Montana. *Journal of Vertebrate Paleontology* 8:378–382.
- . 1992. Mammalian fauna of the Judith River Formation (Late Cretaceous, Judithian), north-central Montana. *University of California Publications in Geological Sciences* 136:1–115.
- Montellano, M., A. Weil, and W. A. Clemens. 2000. An exceptional specimen of *Cimexomys judithae* (Mammalia: Multituberculata) from the Campanian Two Medicine Formation of Montana, and the phylogenetic status of *Cimexomys*. *Journal of Vertebrate Paleontology* 20:333–340.
- Murphy, E. C., D. J. Nichols, J. W. Hoganson, and N. F. Forsman. 1995. The Cretaceous/Tertiary boundary in south-central North Dakota. *North Dakota Geological Survey, Report of Investigation* 98:1–74.
- NACSN (North American Commission on Stratigraphic Nomenclature). 1983. North American stratigraphic code. *Bulletin of the American Association of Petroleum Geologists* 67:841–875.

- Nessov, L. A. 1993. New Mesozoic mammals of middle Asia and Kazakhstan, and comments about evolution of theriofaunas of Cretaceous coastal plains of ancient Asia (in Russian). *Trudy Zoologicheskogo Instituta RAN* 249:105–133.
- Obradovich, J. D. 1993. A Cretaceous time scale. In *Evolution of the Western Interior Basin*, ed. W. G. E. Caldwell and E. G. Kauffman. St. John's, Newfoundland: Geological Association of Canada, Special Paper 39:379–396.
- Obradovich, J. D. and J. F. Hicks. 1999. A review of the isotopic calibration points for the geomagnetic polarity time scale in the interval 83 to 33 Ma (C<sub>34n</sub> to C<sub>13n</sub>). *Geological Society of America, Abstracts with Programs* 30(7):A-71.
- Palmer, A. R. and J. Geissman. 1999. *1999 geological time scale*. Boulder, CO: The Geological Society of America.
- Pettyjohn, W. A. 1967. New members of Upper Cretaceous Fox Hills Formation in South Dakota, representing delta deposits. *Bulletin of the American Association of Petroleum Geologists* 51:1361–1367.
- Rigby, J. K. Jr. 1989. The Cretaceous–Tertiary boundary of the Bug Creek drainage: Hell Creek and Tullock formations: McCone and Garfield counties, Montana. In *Mesozoic/Cenozoic vertebrate paleontology: Classic localities, contemporary approaches. Field trip guidebook T322*, ed. J. J. Flynn and M. C. McKenna. Washington, DC: American Geophysical Union, pp. 67–73.
- Rigby, J. K. Jr. and D. L. Wolberg. 1987. The therian mammalian fauna (Campanian) of Quarry 1, Fossil Forest study area, San Juan Basin, New Mexico. *Geological Society of America, Special Paper* 209:51–79.
- Rougier, G. W., M. J. Novacek, R. Pascual, and J. N. Gelfo. 2000. New Late Cretaceous mammals from Argentina and the survival of Mesozoic lineages into the Patagonian early Tertiary. *Journal of Vertebrate Paleontology* 20:65A.
- Rowe, T. B., R. L. Cifelli, T. M. Lehman, and A. Weil. 1992. The Campanian age Terlingua local fauna, with a summary of other vertebrates from the Aguja Formation, Trans-Pecos Texas. *Journal of Vertebrate Paleontology* 12:472–493.
- Russell, L. S. 1964. Cretaceous non-marine faunas of western North America. *Royal Ontario Museum, University of Toronto, Life Sciences Contribution* 61:1–24.
- . 1975. Mammalian faunal succession in the Cretaceous System of western North America. In *The Cretaceous system in the Western Interior of North America*, ed. W. G. E. Caldwell. Geological Association of Canada, Special Paper 13, pp. 137–161.
- Sahni, A. 1972. The vertebrate fauna of the Judith River Formation, Montana. *Bulletin of the American Museum of Natural History* 147:321–412.
- Sankey, J. 1998. *Vertebrate paleontology and magnetostratigraphy of the upper Aguja Formation (late Campanian), Talley Mountain area, Big Bend National Park, Texas*. Ph.D. dissertation, Louisiana State University, Baton Rouge.
- Sloan, R. E. 1987. Paleocene and latest Cretaceous mammal ages, biozones, magnetozones, rates of sedimentation, and evolution. *Geological Society of America, Special Paper* 209:165–200.
- Sloan, R. E., J. K. Rigby Jr., L. Van Valen, and D. Gabriel. 1986. Gradual dinosaur extinction and simultaneous ungulate radiation in the Hell Creek Formation. *Science* 232:629–633.
- Sloan, R. E. and L. S. Russell. 1974. Mammals from the St. Mary River Formation (Cretaceous) of southwestern Alberta. *Royal Ontario Museum, Contributions in Life Sciences* 95:1–21.
- Sloan, R. E. and L. Van Valen. 1965. Cretaceous mammals from Montana. *Science* 148:220–227.
- Storer, J. E. 1991. The mammals of the Gryde local fauna, Frenchman Formation (Maastrichtian: Lancian), Saskatchewan. *Journal of Vertebrate Paleontology* 11:350–369.
- . 1993. Additions to the mammalian paleofauna of Saskatchewan. *Modern Geology* 18:475–487.
- Swisher, C. C. III, L. Dingus, and R. F. Butler. 1993. <sup>40</sup>Ar/<sup>39</sup>Ar dating and magnetostratigraphic correlation of the terrestrial Cretaceous–Paleogene boundary and Puercan mammal age. *Canadian Journal of Earth Sciences* 30:1981–1996.
- Tokaryk, T. T. 1987. An eutherian from the Late Cretaceous (Maestrichtian, Kemp Clay Formation) of Texas. *Occasional Short Notes, Saskatchewan Museum of Natural History* 2:1–6.
- Van Valen, L. 1967. The first discovery of a Cretaceous mammal. *American Museum Novitates* 2285:1–4.
- Waage, K. M. 1968. The type Fox Hills Formation, Cretaceous (Maestrichtian), South Dakota. *Bulletin of the Peabody Museum of Natural History* 27:1–175.
- Webb, M. W. 1998. A revised summary of Lancian (latest Cretaceous) mammal localities with introduction to a new Lancian locality (Lance Formation) in the southwestern Bighorn Basin. In *Cretaceous and lower Tertiary rocks of the Bighorn Basin, Wyoming and Montana*, ed. W. R. Keefer and J. E. Goolsby. Casper: Wyoming Geological Association, Forty-ninth Guidebook, pp. 131–136.
- . 2001. *Fluvial architecture and Late Cretaceous mammals of the Lance Formation, southwestern Bighorn Basin, Wyoming*. Ph.D. dissertation, University of Wyoming, Laramie.
- Weil, A. 1992. *The Terlingua local fauna: Stratigraphy, paleontology, and multituberculate systematics*. M.S. thesis, University of Texas, Austin.
- . 1999. *Multituberculate phylogeny and mammalian biogeography in the Late Cretaceous and earliest Paleocene Western Interior of North America*. Ph.D. dissertation, University of California, Berkeley.
- Weil, A. and W. A. Clemens. 1998. Aliens in Montana: Phylogenetically and biogeographically diverse lineages contributed to an earliest Cenozoic community. *Geological Society of America, Abstracts with Programs* 30(5):69–70.
- Weil, A. and T. E. Williamson. 2000. Diverse Maastrichtian terrestrial vertebrate fauna of the Naashoibito Member, Kirtland Formation (San Juan Basin, New Mexico) confirms “Lancian” faunal heterogeneity in western North America. *Geological Society of America, Abstracts with Programs* 32:498.
- Whitmore, J. L. 1985. Fossil mammals from two sites in the Late Cretaceous Lance Formation in northern Niobrara County, Wyoming. *Dakoterra* 2:353–367.
- Whitmore, J. L. and J. E. Martin. 1986. Vertebrate fossils from the Greasewood Creek locality in the Late Cretaceous Lance Formation of Niobrara County, Wyoming. *Proceedings of the South Dakota Academy of Sciences* 65:33–50.
- Wilson, R. W. 1965. Type localities of Cope's Cretaceous mammals. *Proceedings of the South Dakota Academy of Sciences, University of South Dakota Bulletin* 44:88–90.
- . 1983. Late Cretaceous mammals of western South Dakota, 1974. *National Geographic Society Research Reports* 15:749–752.
- . 1987. Late Cretaceous (Fox Hills) multituberculates from the Red Owl local fauna of western South Dakota. *Dakoterra* 3:118–122.
- Wood, H. E., R. W. Chaney, J. Clark, E. H. Colbert, G. L. Jepsen, J. B. Reeside Jr., and C. Stock. 1941. Nomenclature and correlation of the North American continental Tertiary. *Geological Society of America Bulletin* 52:1–48.

# 3

## Paleocene Biochronology: The Puercan Through Clarkforkian Land Mammal Ages

Donald L. Lofgren, Jason A. Lillegraven,  
William A. Clemens, Philip D. Gingerich,  
and Thomas E. Williamson

**P**ALEOCENE CONTINENTAL STRATA of the Western Interior of North America preserve the world's most complete and most thoroughly studied record of early Cenozoic mammalian evolution. We examine this record. Our examination updates and amplifies earlier ones, specifically Wood et al. (1941) and Archibald et al. (1987) on the first four North American land mammal ages (NALMAs) of the Cenozoic era: the Puercan, Torrejonian, Tiffanian, and Clarkforkian. For brevity, we refer to these as NALMAs or individually as mammal ages. Wood et al. (1941) recognized a fifth mammal age, the Dragonian, between the Puercan and Torrejonian mammal ages. Van Valen (1978) later proposed another, the Mantuan, preceding the Puercan mammal age. Also, Sloan (1987) and Archibald (1987a, 1987b) both tentatively proposed a Bugcreekian mammal age preceding the Puercan (and Mantuan). Archibald et al. (1987) considered the Dragonian mammal age to be part of the Torrejonian and the Mantuan mammal age to be part of the Puercan, and we follow that usage. Also, we follow Archibald and Lofgren (1990) and consider the Bugcreekian to be part of the Puercan.

Wood et al. (1941) correlated the advent of the Paleocene Epoch with the beginning of the Puercan mammal age and the end of the Paleocene Epoch with the end of the Clarkforkian mammal age. Archibald et al. (1987) also correlated the beginning of the Puercan with that of the Paleocene but placed the Clarkforkian mammal age as straddling the Paleocene–Eocene boundary based on correlations by Gingerich (1976) and Rose (1980, 1981a). More recent developments in geochronology based on presumed unique geochemical phenomena indicate that

the Puercan may have begun in the latest Cretaceous, and the beginning of the Wasatchian mammal age (which followed the Clarkforkian) correlates closely to the advent of the Eocene. These geochemical phenomena are an elevated concentration of iridium at the Cretaceous–Paleocene boundary (see Clemens 2002 and references therein) and carbon isotope excursions at both the Cretaceous–Tertiary boundary (Arens and Jahren 1999; Arens et al. 2000) and the Paleocene–Eocene boundary (see Gingerich 2000, 2001, 2003 and references therein).

In this chapter, we are concerned with two types of geochronologic units: the mammal age and the mammal zone or biochron (treated as a subdivision of a mammal age). Mammal ages and mammal zones or biochrons are types of biochronologic units, units that are not used by the North American Stratigraphic Code (NACSN 1983) and are only briefly considered in the *International Stratigraphic Guide* (ISG; Hedberg 1976). Mammal ages and mammal zones are biochronologic units characterized by faunal content and thus are not chronostratigraphic units because they are not tied to a body of rock established to serve as the material reference for all strata deposited during the same span of time. For this reason, Savage (1962) advocated that the word *ages* in *North American land mammal ages* be placed in quotes to reflect the fundamental difference between North American land mammal “ages” and ages as later defined by the NACSN (1983). We agree with this reasoning but omit the quotes as unnecessarily repetitive in this chapter.

The Puercan, Torrejonian, Tiffanian, and Clarkforkian NALMAs differ from one another in their historical development, but their defining and characterizing criteria

are procedurally similar. However, three different kinds of zones are used to subdivide them. The first kind is illustrated by subdivisions of the Puercan and Torrejonian mammal ages and the oldest zone of the Clarkforkian mammal age. Both the Puercan and Torrejonian are subdivided into three zones, all of which are defined and limited by successive first appearances of unrelated taxa (first appearance datum [FAD]). The oldest zone of the Clarkforkian, for example, is defined by the first appearance of Rodentia. Zones such as these are recognized by the NACSN and ISG as one type of interval zone.

The second kind of zone is recognized for all six of the Tiffanian zones and for the second oldest Clarkforkian zone. These seven zones conform to the concept of a range zone according to the ISG and to a type of interval zone according to the NACSN. Both codes are in agreement on lineage zones because the successive earliest appearances defining the zones form a presumed phylogenetic lineage. The final type of zone is represented by the youngest of the three zones in the Clarkforkian mammal age and is called an abundance zone by the NACSN and an acme zone by the ISG.

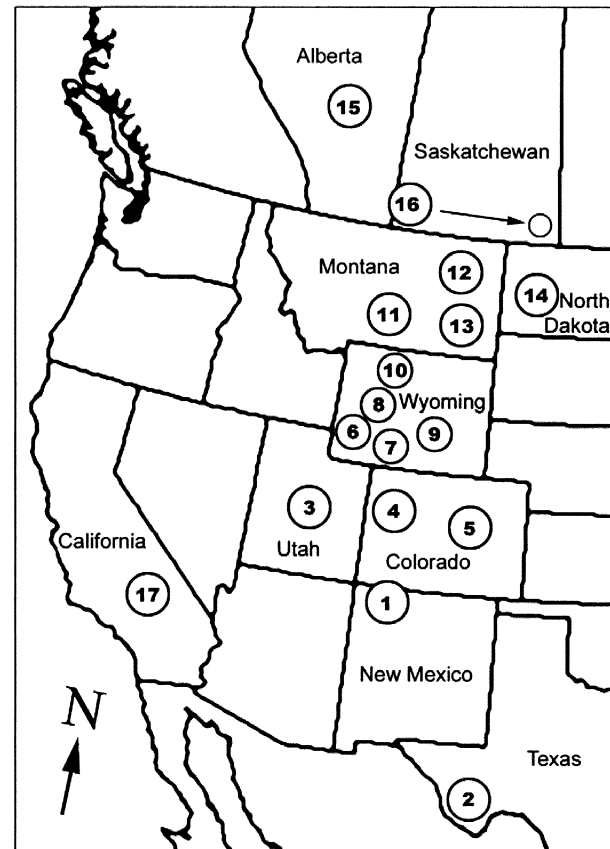
We follow Archibald et al. (1987) in their use of ISG terminology and refer to the three types of zones as interval zones, lineage zones, and acme zones. We also follow Archibald et al. (1987) in naming and subsequent usage of zonal names by including the name of the commencing taxon and the closing taxon for both the interval and lineage zones. The oldest zone in the Puercan mammal age, for example, is the *Protungulatum/Ectoconus* Interval Zone, and the oldest in the Tiffanian mammal age is the *Plesiadapis praecursor/Plesiadapis anceps* Lineage Zone. Also, each zone can be referred to by an abbreviation of the mammal age in which it occurs plus a number indicating its temporal sequence within the given mammal age. For example, the two zones just mentioned are known informally as the Pui interval zone and the Tii lineage zone, or simply Pui and Tii, respectively.

For each of the four mammal ages and fifteen zones discussed in this chapter, we provide a standardized definition and characterization. A formal definition is not given for the fifteenth zone, the *Phenacodus-Ectocion* Acme Zone. The definitions for all four mammal ages and for the remaining fourteen zones are based on the first appearance of a single taxon. Characterization for each age and zone consists of five parts: “first appearances” for taxa that appear for the first time within a mammal age or zone (but not necessarily in the oldest faunas of that mammal age or zone), “last appearances” for taxa that appear for the last time within a mammal age or zone (but not necessarily in the youngest faunas of that mammal age or zone), “index fossils” for taxa limited to a

mammal age or zone, “characteristic fossils” for all taxa (not just common taxa) that occur within a mammal age or zone (but do not belong to one of the three preceding categories), and taxa that are recognized before and after a given mammal age or zone but not within it.

We follow the recently published classification of mammals by McKenna and Bell (1997). Also, we limit our review to Paleocene mammalian biochronology of North America, associated magnetostratigraphic and radioisotopic data, and intercontinental correlations. Thus for discussion of community structure of Mammalia during the Paleocene, rates of origination and turnover, diversity histories, and effects of climate change on mammalian evolution in North America, we recommend use of Alroy (1999, 2000), Alroy et al. (2000), Wilf (2000), Maas and Krause (1994), Maas et al. (1995), and Wing et al. (1995) as starting points.

We present two figures and two tables to augment the text. In general, they are updates of the two figures and two tables provided by Archibald et al. (1987). Figure 3.1



**FIGURE 3.1** Map showing approximate locations of Puercan through Clarkforkian mammal-bearing regions in western North America. Each number refers to a region in which known mammal localities are present (not shown are Louisiana and South Carolina) and relates to a stratigraphic column presented in figure 3.2.



is a map of western North America showing the approximate location of areas from which Puercan through Clarkforkian mammalian faunas have been recovered (equivalent to figure 3.1 of Archibald et al. 1987). Figure 3.2 is a biochronologic correlation of Puercan through Clarkforkian localities clustered according to geographic and geologic features (equivalent to figure 3.2 of Archibald et al. 1987). Table 3.1 is a list of localities and references for each locality (equivalent to table 3.1 of Archibald et al. 1987). Table 3.2 is a list of taxa and their temporal ranges (similar to table 3.2 of Archibald et al. 1987 but lacking the locality component).

We would be remiss if we did not formally acknowledge that the organizational content of this chapter closely follows that of Archibald et al. (1987). We owe them a tremendous debt for compiling and producing the first update of Paleocene NALMAs since Wood et al. (1941). We were able to build on their framework. As we present our version of the current state of knowledge concerning the Puercan through Clarkforkian NALMAs, we hope our work is also a worthy framework for a future update. All interpretations, as well as errors or inaccuracies in the text or figures, are our own and not attributable to Archibald et al. (1987).

## PUERCAN MAMMAL AGE

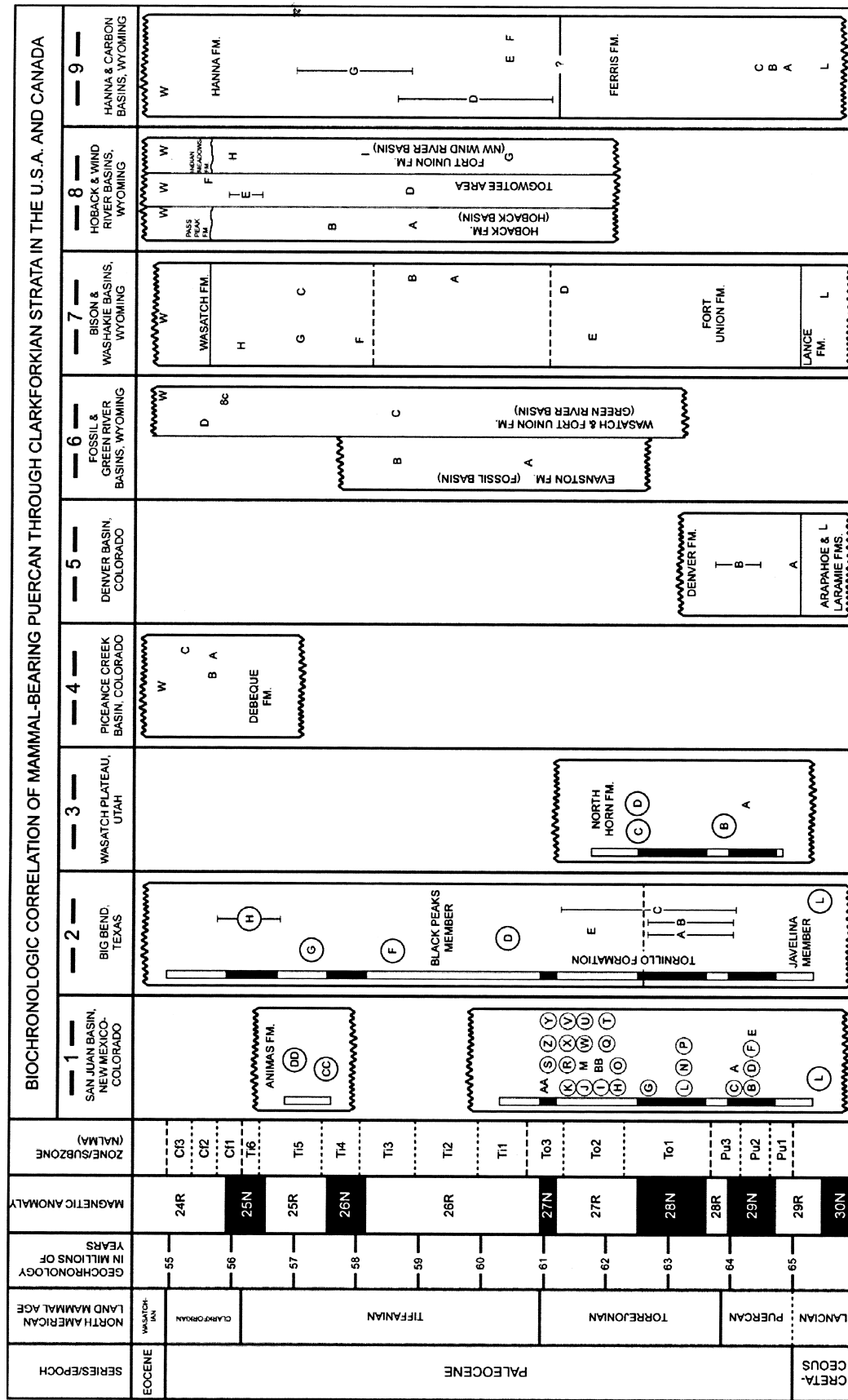
The concepts of the Puercan and Torrejonian NALMAs grew out of work done in the San Juan Basin, New Mexico, in the late nineteenth century with fossils supplied to E. D. Cope from the “Puerco Marls” (Cope 1875) by professional collector David Baldwin (Cope 1884, 1888). Cope’s “Puerco Marls” were subsequently subdivided into two formations, the Puerco and overlying Torrejon (Matthew 1897). When naming the Puercan mammal age, Wood et al. (1941) based it on the Puerco Formation and named the type locality the Rio Puerco area (near Cuba, New Mexico). They also noted that the “most typical and only fossiliferous exposures” of the Puercan are “the escarpment running from northwest of Ojo Alamo about 25 miles to Arroyo Eduardo, east of Kimbetoh” (1941:8). Similarly, Wood et al. (1941:9) based the Torrejonian mammal age on the Torrejon Formation, described the type locality as situated at “the heads of Arroyo Torrejon,” and stated that the “typical area runs from there northwest to Ojo Alamo, with additional, poorer localities scattered to the north.” After much discussion concerning the difficulty of differentiating the Puerco and Torrejon formations based on lithologic criteria, it was discovered that the type locality of the Puercan mammal

age in the Rio Puerco area, as recognized by Wood et al. (1941), yielded fossils that were Torrejonian in age (Simpson 1959).

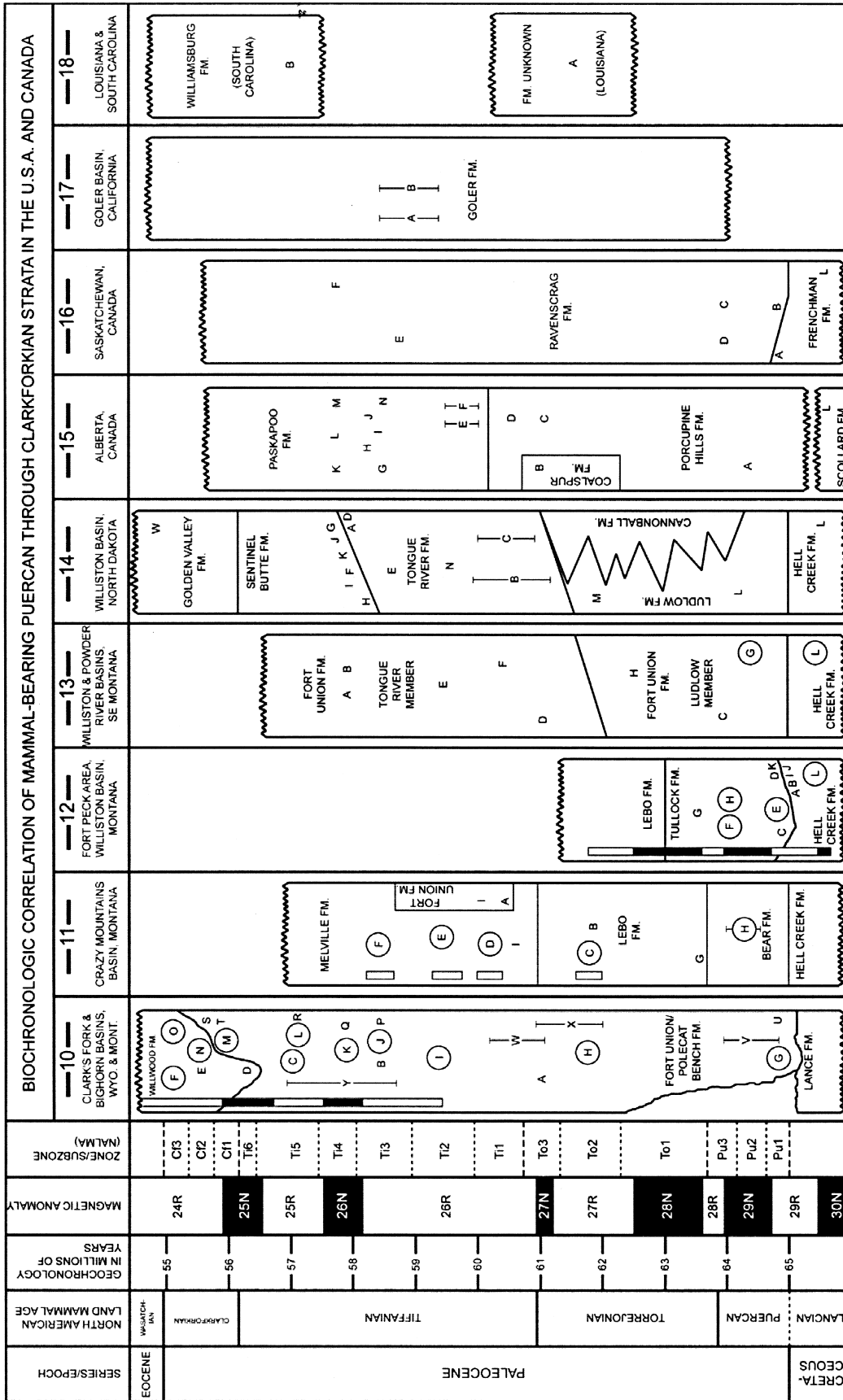
Simpson (1959) further noted that the “Puerco” and “Torrejon” formations could not be differentiated based on lithologic criteria and suggested that the Nacimiento Formation of Gardner (1910) be used in a restricted sense to replace them. This usage was adopted by subsequent workers, although the Nacimiento Formation was later subdivided into three members, the Arroyo Chijuillita, Ojo Encino, and Escavada (Williamson and Lucas 1992). Thus since 1959 only the exposures on the escarpment running from northwest of Ojo Alamo about 25 miles to Arroyo Eduardo, east of Kimbetoh have been recognized as yielding the type Puercan fauna (Simpson 1959; Archibald et al. 1987; Williamson 1996). These Puercan sites are all restricted to the Arroyo Chijuillita Member of the Nacimiento Formation (Williamson and Lucas 1992; Williamson 1996). For purposes of discussion, we divide these exposures geographically into the De-na-zin (= Barrel Spring Arroyo), Alamo, Kimbetoh, and Bettonie-Tsotie washes and the West Fork of Gallegos Canyon.

The only correlative of the Puercan assemblages recognized by Wood et al. (1941) is what Archibald et al. (1987) called the Mantua Lentil Local Fauna from the Fort Union Formation (= Polecat Bench Formation of Jepsen 1940), Wyoming. Archibald et al. (1987) retained the Mantua Lentil Local Fauna in the Puercan mammal age, although they demonstrated that it was older than the type Puercan assemblage (*sensu* Simpson 1959) of the San Juan Basin, basing their correlation on the stage of evolution of the mammals, the demonstrable superposition in northeastern Montana of type Puercan-like local faunas over Mantua-like local faunas, and faunal and magnetostratigraphic correlation between type Puercan-like local faunas of Montana and the type Puercan fauna of New Mexico. Thus, when dividing the Puercan into three interval zones, Archibald et al. (1987) referred the Mantua Lentil Local Fauna to the oldest (Pu1) and the type Puercan to the youngest (Pu2–Pu3) rather than accepting the argument by Van Valen (1978; further supported by Williamson 1996) that the Mantua Lentil Local Fauna should be the basis for a new Mammal Age, the “Mantuan.” Thus Archibald et al. (1987) defined the beginning of Pu1 interval zone, the initial interval zone of the Puercan mammal age, as occurring with the first appearance of the marsupial *Peradectes*.

However, in 1987, when Archibald et al. was published, there was much uncertainty about the age of a series of localities in eastern Montana and western Canada that contained unusual mammalian assemblages with taxa indicative of both the Lancian and Puercan NALMAs. These



**FIGURE 3.2** Correlation of Puercan through Clarkforkian faunas and mammal-bearing strata in United States and Canada, with localities clustered according to geographic or geologic features (often sedimentary basins). Each column is denoted by a number matching those used in figure 3.1. Number 18, in Louisiana and South Carolina, is not shown in figure 3.1. Mammal-bearing localities or suites of localities in each region are designated by letters. Letters L and W indicate Lancian and Wasatchian sites, respectively. Letters that are circled indicate that strata containing the localities or suite of localities have undergone paleomagnetic analysis and have been correlated to a magnetic anomaly. Within regions, correlation between faunas is based on biochronology or biostratigraphy. Between regions, correlation is biochronologic except in areas where magnetostratigraphic studies have been conducted. The approximate geochronologic ages and epochal and magnetic chrons boundaries are after Berggren et al. (1995) and Cande and Kent (1995). Note that for some sections (e.g., Clark's (continued on next page)



(continued from previous page) Fork and Bighorn basins) biochronologically recognized mammal ages and zones can be placed in a geochronologic context using magnetostratigraphy. Certain areas (e.g., Fort Peck area of Williston Basin) provide both magnetostratigraphy and radioisotopic dating as means of placement of biochronologically recognized mammal ages and zones into geochronologic context. Such correlations provide some degree of confidence to chronostratigraphic and geochronologic assignments for certain strata. However, this is not possible for most sections. Thus a given segment of a formation should not be assumed to correlate across the chart to a given mammal age or zone (e.g., in the absence of relevant data, the lowermost Hoback Formation should not be assumed to be latest Torrejonian in age at all places of occurrence).

**TABLE 3.1** List of Regions in Which Mammal-Bearing Strata Occur and Known Localities in Each Region**I. SAN JUAN BASIN, NEW MEXICO-COLORADO****PUERCAN**

- A. West Fork of Gallegos Canyon, AMNH locality 4 (Pu3)
- B. De-na-zin Wash AMNH locality 2, lower fossil level (Pu2)
- C. De-na-zin Wash AMNH locality 2, upper fossil level (Pu3)
- D. West Flank Kimbeto Wash AMNH locality 5 (Pu2)
- E. East Flank Kimbeto Wash AMNH localities 6 and 7 (Pu2)
- F. Betonnie-Tsosie Wash including Mammalon Hill NMMNH L-317 (Pu2)

**TORREJONIAN**

- G. Kutz Canyon NMMNH localities 2659, 2660 (To1)
- H. Kutz Canyon AMNH locality "Sec. 3, T27N, R11W," KU locality 14 (To2)
- I. KU Locality 13, "Big Pocket" (To2)
- J. AMNH localities "1 or 2 miles west of Angel Peak" (To2)
- K. NMMNH localities 1482, 2658 (To2)
- L. De-na-zin Wash AMNH locality 3 (To1)
- M. Gallegos Canyon AMNH locality 1 (To2)
- N. Lowest Torrejonian of Kimbeto Wash, NMMNH 692 (= UALP 77113) (To1)
- O. Head of Kimbeto Wash AMNH locality 8 in part, KU 9 "Little Pocket" (To2)
- P. Betonnie-Tsosie Wash ("lowest Torrejonian") (To1)
- Q. 44 store localities low and high, AMNH locality 9 in part? (To2)
- R. Escavada Wash AMNH locality 14 in part, lower horizon (To2)
- S. Escavada Wash AMNH locality 14 (head of Escavada Wash) (To3)
- T. Torrejon Wash NMMNH localities 2693, 2714 (To2)
- U. Torrejon Wash NMMNH locality 2723 (To2)
- V. Torrejon Wash NMMNH locality 2709 (To2)
- W. East Branch of Torrejon Wash NMMNH 2724 (AMNH locality 11 in part?) (To2)
- X. East Branch of Torrejon Wash AMNH locality 11 lower horizon (To2)
- Y. West Branch of Torrejon Wash AMNH locality 10 upper horizon, including Tsentas Microvertebrate locality NMMNH L-312 (To3)
- Z. East Branch of Torrejon Wash AMNH locality 10 upper horizon (To3)
- AA. Mesa Chijuilla AMNH locality 17 (To3)
- BB. Mesa de Cuba and Mesa Portales AMNH localities 222, 226, 229, 230 (To2)

**TIFFANIAN**

- CC. Mason Pocket near Tiffany (Ti4)
- DD. Bayfield and others (Ti5)

References: Simmons 1987; Thewissen 1990 (DD); Williamson and Lucas 1993; Gunnell 1994 (CC); Van Valen 1994; Williamson 1996; Lucas et al. 1997; Rose and Lucas 2000

**II. BIG BEND, TEXAS**

- A. Dogie LSUMG VL-108 (Pu3-To1?)
- B. Tom's Top LSUMG VL-111 (Pu3-To1?)
- C. Glenn Eleven LSUMG VL-107 (Pu3-To3)
- D. C-Con and Schiebout-Reeves Quarry (TMM 41274, 41377) (Ti1)
- E. The Middle Peak and Alligator Alley (TMM 40147) (To2?)
- F. Ray's Bonebed (TMM 40536, 40537) (Ti3)
- G. Joe's Bonebed (TMM 41365, 41366) (Ti5)
- H. New Taeniodont Site (TMM 41364) (Ti or Cf)

References: Schiebout 1974; Schoch 1986 (H); Schiebout et al. 1987; Thewissen 1990 (D, F); Standhardt 1986; Williamson 1996

**TABLE 3.1** (continued)**III. WASATCH PLATEAU, UTAH**

- A. Gas Tank Hill local fauna (Pu2?)
- B. Wagonroad; lower part of Gazin's locality 4 (Pu3)
- C. Dragon; upper part of Gazin's locality 4 (To1)
- D. Dragon; locality 2 of Gazin (To1)

References: Robison 1986; Cifelli et al. 1995, 1999; Lucas et al. 1997; Williamson 1996

**IV. PICEANCE CREEK BASIN, COLORADO**

- A. Plateau Valley local fauna (Cf2)
- B. Big Rock Ranch local fauna (Cf2?)
- C. Later Clarkforkian local fauna (including Flynn Hill, Oval Hill) (Cf3)

References: Kihm 1984; Thewissen and Gingerich 1987 (A)

**V. DENVER BASIN, COLORADO**

- A. Littleton local fauna (Alexander and South Table Mountain) (Pu1)
- B. Coral Bluffs, Jimmy Camp Creek, West Bijou Creek (Pu2–3)

References: Middleton 1982, 1983; Williamson 1996

**VI. FOSSIL AND GREEN RIVER BASINS, WYOMING**

- A. Little Muddy Creek (Ti1)
- B. Twin Creek (Ti3)
- C. Chappo Type Locality (Chappo 17) (Ti3)
- D. Buckman Hollow Locality (Chappo 1, Chappo 12) (Cf2)

References: Dorr and Gingerich 1980 (C, D); Gunnell 1989 (B), 1994 (B, C); Krause 1987b (D); Williamson 1996 (A); Gingerich 1976 (A, B)

**VII. BISON AND WASHAKIE BASINS, WYOMING****BISON BASIN**

- A. Saddle Locality (Ti2)
- B. Ledge Locality, Saddle Annex, West End (Ti3)
- C. *Titanoides* Locality (Ti5)

**WASHAKIE BASIN**

- D. University of Wyoming Localities V77009–10, 12, 14, V78055 (To3)
- E. Swain Quarry (To2?)
- F. University of Wyoming Localities V77005–8, 13, 15–16, V77061 (Ti4)
- G. University of Wyoming Localities V76008, V77059–60, V78052–54 (Ti5)
- H. Big Multi Quarry (Cf1)

References: Gunnell 1994 (A); Gingerich 1976, 1983 (A, B, C); Rigby 1980 (E); Winterfeld 1982 (D, F, G); Rose 1981a (H); Thewissen 1990 (E); Williamson 1996 (E); Dawson and Beard 1996 (H); Wilf et al. 1998 (H)

**VIII. HOBACK AND WIND RIVER BASINS, WYOMING**

- A. Battle Mountain (Ti3)
- B. Dell Creek Quarry (Ti5)
- C. UM-Sub-Wy localities 7, 10, 20 (Cf2) (locality plotted in #6 column of figure 3.2)
- D. Love Locality (Ti3)
- E. Low Locality, Rohrer Locality (Ti6–Cf1)

**TABLE 3.1** (continued)

- 
- F. Purdy Basin (Red Creek Localities) (Cf2)
  - G. Keefer Hill (Twin Buttes) (Ti1)
  - H. West Side of Shotgun Butte (Cf2?)
  - I. "Malcolm's Locality" (Badwater Locality) (Ti4)

References: Gunnell 1989 (G), 1994 (A, D, J); Gingerich 1976; 1983 (A, B); Thewissen and Gingerich 1987 (C); Rose and Krause 1982 (E); Cifelli et al. 1989 (F); Rose 1981a (F)

## **IX. HANNA AND CARBON BASINS, WYOMING**

### **HANNA BASIN**

- A. Ferris Formation Pu1 Level (Pu1)
- B. Ferris Formation Pu2 Level (Pu2)
- C. Ferris Formation Pu3 Level (Pu3)
- D. The Breaks local fauna (To3–Ti3)

### **CARBON BASIN**

- E. Grayson Ridge fauna (Ti1?)
- F. Halfway Hill fauna (Ti1?)
- G. Sand Creek fauna (Ti3–Ti5)

References: Eberle 1999 (B); Eberle and Lillegraven 1998a (A–C), 1998b (A–C); Higgins 2000 (D–G); Lillegraven and Eberle 1999 (A–C); Secord 1998 (D–G)

## **X. CLARK'S FORK AND BIGHORN BASINS, WYOMING AND MONTANA**

### **WEST AND NORTHWEST OF POLECAT BENCH**

- A. Cub Creek (To3)
- B. Seaboard Well (Ti3)
- C. Princeton Quarry, Schaff Quarry, Fossil Hollow, Brice Canyon, Fritz Quarry, Jepsen Valley (Ti5)
- D. Little Sand Coulee (Cf1), various UM localities (including 52a) (Cf1), Bear Creek (Cf1), various UM localities (Ti6)
- E. *Franimys* Hill, Phil's Hill, Paint Creek, Krause Quarry, Holly's Microsite, various other localities (Cf2)
- F. Granger Mountain, Rainbow Valley, various UM localities (Cf3)

### **VICINITY OF POLECAT BENCH AND POWELL, WYOMING**

- G. Mantua Lentil (Pu1)
- H. Rock Bench Quarry (To2)
- I. UM Locality 263 (Ti2)
- J. Various UM localities (Ti3)
- K. Airport Locality, Long Draw Quarry (Ti4)
- L. Various UM localities (Ti5)
- M. Various UM localities (Cf1)
- N. Various UM localities (Cf2)
- O. Various UM localities (Cf3)

### **SOUTH AND EAST OF POWELL, WYOMING**

- P. Cedar Point Quarry, Jepsen Quarry (Ti3)
- Q. Lower Sand Draw, Witter Quarry (Croc Tooth), Divide Quarry, Sand Draw Anthill (Ti4)
- R. Middle Sand Draw, Sunday Locality (Ti5)
- S. Foster Gulch (Cleopatra Reservoir Quarry), Rough Gulch, Upper Sand Draw (Cf2)
- T. Ries Locality, Foster Gulch Oil Well #1 (Cf, but zone uncertain)

### **SOUTHERN BIGHORN BASIN**

- U. Leidy Quarry (Pu1)
- V. Cedar Mountain Sites, low in Fort Union Formation (Pu, but zone uncertain)

**TABLE 3.1** (*continued*)

---

W. Cedar Mountain Sites, high in Fort Union Formation (To3 or Ti1)

X. Grass Creek–Blue Mesa #1 (To2–To3)

Y. Grass Creek Blue Mesa #2 (Ti3–Ti5)

---

References: Thewissen 1990; Gunnell 1988, 1989, 1994; Krause 1987a, 1987b; Gingerich 1976, 1987, 1989; Cifelli et al. 1989; J. E. Hartman 1986 (V–W); Leite 1992 (X–Y); Gunnell and Gingerich 1991; Rose 1981a; Van Valen 1978; Williamson 1996 (H); Bloch et al. 2001 (Q); Bloch et al. 2002

#### **XI. CRAZY MOUNTAINS BASIN, MONTANA (BANGTAIL LOCALITY LOCATED IN BRIDGER RANGE)**

A. Bangtail Locality (Ti1)

B. Gidley Quarry (To2)

C. Silberling Quarry (To2)

D. Douglass Quarry (Ti1)

E. Scarritt Quarry (Ti2)

F. Simpson Locality 13 (= Melville Locality) (Ti3)

G. Simpson Locality 65 (To1)

H. Simpson Quarry (Pu2 or Pu3)

I. Glennie and Bingo localities (Ti1)

---

References: Hartman and Krause 1993; Hartman et al. 1989 (H); Buckley 1994 (H), 1995 (H), 1997 (H); Hartman 1999; Butler et al. 1987; Krause and Maas 1990; Gunnell 1994; Thewissen 1990; Wall and Krause 1992 (C); Williamson 1996 (B, H)

#### **XII. FORT PECK AREA, WILLISTON BASIN, MONTANA**

A. Bug Creek Anthills (Pu1)

B. Bug Creek West, Harbicht Hill, Ferguson Ranch, Chris's Bonebed (Pu1)

C. Morales 1 and Herpijunk Promontory (Pu1)

D. McKeever Ranch localities (Pu1)

E. Hell's Hollow local fauna (including Worm Coulee #1) (Pu1)

F. Garbani Channel localities (including Garbani Quarry) (Pu3?)

G. Mosquito Gulch localities (including Farrand Channel) (To1)

H. Purgatory Hill (Pu3?)

I. McGuire Creek local faunas (Little Roundtop, Black Spring Coulee, Shiprock, Brown Grey, Second Level, Up Up the Creek) (Pu1)

J. Z-Line Channel local fauna (Pu1)

K. Jacks Channel local fauna (Pu1)

---

References: Lofgren 1995; Archibald 1982; Archibald and Lofgren 1990; Clemens 2002; Van Valen 1978, 1994; Swisher et al. 1993; Rigby 1987, 1989; Rigby et al. 1987; Sloan et al. 1986; Lupton et al. 1980 (B); Simmons 1987 (F, H); Luo 1991 (A); Fox 1989 (A)

#### **XIII. WILLISTON AND POWDER RIVER BASINS, SOUTHEAST MONTANA**

A. Olive (Ti4)

B. Circle (Ti4)

C. Bechtold Site (Pu3?)

D. Medicine Rocks 1, Mehling Site (To3?)

E. White Site, 7-Up Butte, Highway Blowout (Ti2)

F. Newell's Nook (USGS D-2003) (Ti1)

G. Hiatt local fauna (Pu2)

H. School Well local fauna (To, but zone uncertain)

---

References: Hunter et al. 1997 (G–H); Robinson and Honey 1987 (F); Simmons 1987 (C); Thewissen 1990; Gunnell 1989, 1994; Krause 1987a (D); Gingerich 1976; Rose 1975

**TABLE 3.1** (continued)**XIV. WILLISTON BASIN, NORTH DAKOTA**

- A. Wannagan Creek Quarry (Ti4)
- B. Donnybrook (To or Ti)
- C. Lloyd and Hares Site (= Heart Butte) (To3 or Ti1)
- D. Judson (Ti4)
- E. Brisbane (Ti3)
- F. Riverdale (Ti4)
- G. Cross Locality (L5377 a and b) (Ti4)
- H. White's River Basin Survey Site 3 (Ti3?)
- I. Red Spring Locality (Ti4)
- J. Locality L5500b (Ti4)
- K. Witter Locality (Ti4)
- L. Pita Flats (Pu2–Pu3)
- M. Brown Ranch localities (To2)
- N. X-X Locality (Ti2)

References: Hartman and Kihm 1991, 1992, 1995, 1999; J. H. Hartman 1999; Erickson 1991 (A), 1999 (A); Gunnell 1994 (D–F); Thewissen 1990 (D, F); Krause 1987b (D); Hunter 1999 (L–N)

**XV. ALBERTA, CANADA**

- A. R.C.A. Corehole 66-1 (Balzac) (Pu2?)
- B. Diss (To3?)
- C. Calgary 2E (inc. Calgary 7E) (To3?)
- D. Cochrane 1, Cochrane 2 (Ti1)
- E. Aaron's Locality (Ti1 or Ti2)
- F. Hand Hills West lower level (Ti1 or Ti2)
- G. Hand Hills West upper level (Ti3)
- H. Blindman River localities (DW-1, DW-2, DW-3, Mel's Place) (Ti3)
- I. Burbank (Ti3)
- J. Joffe Bridge localities (including Erickson's Landing) (Ti3)
- K. Crestomere School (Ti4)
- L. Canyon Ski Quarry (Ti4)
- M. Swan Hills Site (Ti4)
- N. Birchwood Locality (Ti3)

References: Fox 1990a, 1990b, 1990c, 1997; Webb 1995 (N); MacDonald 1995 (F, G); Gunnell 1994 (M); Thewissen 1990 (D, J); Williamson 1996 (B)

**XVI. SASKATCHEWAN, CANADA (POLICE POINT LOCATED IN SOUTHEAST ALBERTA)**

- A. Frenchman 1 (Pu1)
- B. MHBT Quarry, Long Fall (Pu1)
- C. MHBT Quarry, Rav W-1 (Pu3?)
- D. Croc Pot (Pu3)
- E. Police Point (Ti3)
- F. Roche Percée (Ti4)

References: Fox 1990c, 1997; Lofgren 1995 (A, B); Gunnell 1989 (C), 1994 (F, G); Krause 1987a (C), 1987b (G); Simmons 1987 (C); Williamson 1996 (C)



TABLE 3.1 (continued)

**XVII. GOLER BASIN, CALIFORNIA**

A. Laudate local fauna (Ti2–Ti3)

B. Edentulous Jaw Site (Ti2–Ti3)

References: McKenna et al. 1987 (A); Lofgren et al. 1999 (A, B); McKenna and Lofgren in press (A)

**XVIII. LOUISIANA AND SOUTH CAROLINA (NOT SHOWN IN FIGURE 2)**

A. Junior Oil Co. Beard #1 Well (Louisiana) (To2?)

B. Santee River Rediversion Project (South Carolina) (Ti5)

References: Russell 1967 (A); Simpson 1932 (A); Schoch 1985, 1998 (B)

The same convention of numbering (region) and lettering (locality) used in figures 3.1 and 3.2 is used here. The abbreviation in parentheses after each locality is the zone to which the fauna from that locality is referred. References follow each numbered section; those listed are seminal works that refer to most of the relevant localities or are references that appeared after publication of Archibald et al. (1987). References followed by a letter or letters in parentheses denote the specific locality or localities to which each reference relates. For a more extensive list of cited literature, see table 3.1 in Archibald et al. (1987).

were the Bug Creek Faunas (Bug Creek Anthills, Bug Creek West, Harbicht Hill) from the upper Hell Creek Formation of eastern Montana, first described by Sloan and Van Valen (1965), and the Frenchman 1 (Frenchman Formation) and Long Fall (Ravenscrag Formation) sites in southern Saskatchewan, described by Johnston (1980) and Johnston and Fox (1984). Archibald et al. (1987) tentatively recognized the Bug Creek Faunas and correlatives as Lancian in age because their stratigraphic position was thought to be laterally equivalent to nearby Lancian sites (Archibald 1982). But they also suspected that they “may be younger than known typical Lancian localities” (Archibald et al. 1987:41).

Subsequently, Archibald (1987a, 1987b) and Sloan (1987) both proposed a Bugcreekian mammal age preceding the Puercan, based on the Bug Creek Faunas and presumed correlatives (i.e., Frenchman 1 and Long Fall). As defined, the Bugcreekian began with the first appearance of the ungulate *Protungulatum* (Archibald 1987a, 1987b). It should be noted that when the Bugcreekian mammal age was proposed, the first appearance of *Protungulatum* was in channel fillings in the upper Hell Creek Formation that were thought to be of Cretaceous age. The stratigraphically lowest record of *Peradectes* was in the lowest part of the overlying Tullock Formation and was recognized to be of Puercan age. Shortly thereafter, Lofgren (1995) reported the co-occurrence of *Protungulatum* and *Peradectes* at a number of sites in the upper Hell Creek Formation in the McGuire Creek area, which is 3 miles south of Bug Creek, where two of the three original

Bug Creek sites are located. Based primarily on the co-occurrence of *Protungulatum* and *Peradectes* in the upper Hell Creek Formation at McGuire Creek, the Bugcreekian mammal age was abandoned and the Puo interval zone of the Puercan NALMA was proposed to replace it (Archibald and Lofgren 1990). Thus the first occurrence of *Protungulatum* defined the beginning of both the Puo interval zone and the Puercan mammal age (Archibald and Lofgren 1990). Later, Lofgren (1995) discussed the practical difficulties of paleontologically distinguishing Puo from Pui, noting that Puo involves no genera of unique occurrence and must be differentiated from Pui by the absence of *Peradectes*, a typically rare genus. Also, as noted, Puo and Pui faunas are found in the same stratigraphic interval in Montana, the upper Hell Creek Formation, and further complicating the biostratigraphy is that Puo and Pui faunas in the Bug Creek and McGuire Creek areas contain reworked Lancian fossils (Lofgren 1995). Thus in both a biostratigraphic and biochronologic sense the Puo interval zone, although demonstrably older in two cases than any Pui site (based on faunal content at Bug Creek Anthills [Lofgren 1995] and palynology at Frenchman 1 [Fox 1995, 1997]), is still too similar to the Pui interval zone to be of practical use. Therefore we merge Puo and Pui and recognize the advent of the Puercan mammal age as beginning with the first occurrence of the ungulate *Protungulatum*.

A recent test of the utility of the Puo interval zone that supports our decision to merge the Puo and Pui interval zones comes from the type Ferris Formation, roughly 2



TABLE 3.2 (continued)

TAXON	L+	PU1	PU2	PU3	T01	T02	T03	T11	T12	T13	T14	T15	T16	CF1	CF2	CF3	W+
<i>Glasbiidae</i>																	
<i>Glasbius</i>	←	?															
<i>Pediomyidae</i>																	
<i>Pediomys</i>	←	?															
<i>Stagodontidae</i>																	
<i>Didelphodon</i>	←	?															
LEPTICTIDA																	
<i>Gypsonictopidae</i>																	
<i>Gypsonictops</i>	←	?															
<i>Stilpnodon</i>						X	O	O	O	?							
<i>Leptictidae</i>																	
<i>Leptonysson</i>							X										
<i>Prodiacodon</i>			X	O	X	X	X	X	X	X	O	X	O	O	?	?	→
<i>Palaeictops</i>			X	O	X	?	O	O	O	X	X	O	O	O	O	O	→
<i>Myrmecoboides</i>					X	X	X	O	X	O	X						
<i>Xenacodon</i>											X						
ANAGALIDA																	
<i>Pseudictopidae</i>																	
<i>Mingotherium</i>													X				
RODENTIA																	
<i>Alagomyidae</i>																	
<i>Alagomys</i>														X			
<i>Ischyromyidae</i>																	
<i>Acritoparamys</i>														X	O	O	→
<i>Microparamys</i>															X	X	→
<i>Paramys</i>														X	X	X	→
<i>Franimys</i>															X	O	→
<i>Apatosciuravus</i>														X	X	X	→
<i>Reithroparamyidae</i>																	
<i>Reithroparamys</i>															X	O	→
CIMOLESTA																	
<i>Family indeterminate</i>																	
<i>Alostera</i>	←	?															
<i>Ravenictis</i>			X														
<i>Pararyctes</i>						X	X	X	X	X							
<i>Palaeoryctidae</i>																	
<i>Palaeoryctes</i>					X	X	X	O	X	X	X	O	X	X	X		
<i>Aptoryctes</i>												X					



TABLE 3.2 (continued)

TAXON	L+	PU1	PU2	PU3	T01	T02	T03	T11	T12	T13	T14	T15	T16	CF1	CF2	CF3	W+
<b>TILLODONTIA</b>																	
<i>Tillotheriidae</i>																	
<i>Esthonyx</i>													?	X	X	X	→
<i>Deltatherium</i>						X											
<b>PANTODONTA</b>																	
<i>Titanoideidae</i>																	
<i>Titanoides</i>								X	X	X	X	X	O	?			
<i>Pantolambidae</i>																	
<i>Pantolambda</i>						X	X	X									
<i>Caenolambda</i>									X	X							
<i>Barylambdidae</i>																	
<i>Barylamba</i>										X	X	X	O	X	X	?	→
<i>Haplolambda</i>											X	X	O	X			
<i>Ignatiolambda</i>											X	X					
<i>Cyriacotheriidae</i>																	
<i>Cyriacotherium</i>										X	X	X	X	X	X	X	
<i>Coryphodontidae</i>																	
<i>Coryphodon</i>														X	X	X	→
<b>CREODONTA</b>																	
<i>Oxyaenidae</i>																	
<i>Oxyaena</i>												X	X	X	X	X	→
<i>Dipsalidictides</i>														X	X	X	→
<i>Tytthaena</i>									X								
<i>Dipsalodon</i>												X	?	O	X	?	
<i>Palaeonictis</i>																X	
<b>CARNIVORA</b>																	
<i>Viverravidae</i>																	
<i>Pristinictis</i>								X									
<i>Simpsonictis</i>						X	O	X	?								
<i>Viverravus</i>												X	O	X	X	X	→
<i>Ictidopappus</i>			?	O	O	X											
<i>Didymictis</i>												X	X	X	X	X	→
<i>Protictis</i>					X	X	X	X	X	X	X	X					
<i>Raphictis</i>										X	X						
<i>Miacidae</i>																	
<i>Uintacyon</i>															X	X	→



TABLE 3.2 (continued)

TAXON	L+	PU1	PU2	PU3	T01	T02	T03	T11	T12	T13	T14	T15	T16	CF1	CF2	CF3	W+
<i>Nannodectes</i>								X	X	X	X	X					
<i>Plesiadapis</i>								X	X	X	X	X	X	X	X	X	→
<b>Palaechthonidae</b>																	
<i>Palaechthon</i>					X	X	X	X	O	X							
<i>Premnoides</i>										X							
<i>Anasazia</i>										X							
<i>Palenochtha</i>						X	X	X									
<i>Plesiolestes</i>					X	X	X	X									
<b>Picrodontidae</b>																	
<i>Draconodus</i>					X												
<i>Picrodus</i>						X	X	X	X	X	O	X					
<i>Zanycteris</i>										X	X						
<b>DERMOPTERA</b>																	
<b>Paromomyidae</b>																	
<i>Paromomys</i>					X	X	X	X	?	?							
<i>Ignacius</i>						X	X	X	X	X	X	O	X	X	X	X	→
<i>Dillerlemur</i>												X	X	X	X	X	→
<i>Phenacolemur</i>										?	O	O	X	X	X	X	→
<i>Acidomomys</i>															X		
<b>Plagiomenidae</b>																	
<i>Elpidophorus</i>							X	X	X	X	X						
<i>Eudaemonema</i>					X	X	X	O	X								
<i>Planetetherium</i>													X	?			
<i>Worlandia</i>															X	X	→
<i>Plagiomene</i>															X	X	→
<b>Mixodectidae</b>																	
<i>Mixodectes</i>						X	X										
<i>Dracontolestes</i>					X												
<b>Family indeterminate</b>																	
<i>Thylacaelurus</i>								?	O	O	O	O	O	O	O	O	→
<b>EUPRIMATES</b>																	
<b>Carpolestidae</b>																	
<i>Elphidotarsius</i>					X	X	X	O	X								
<i>Carpodaptes</i>							X	X	X	X							
<i>Carpolestes</i>												X	X	X	X	X	
<i>Carpocristes</i>								X	X	X							
<i>Carpomegodon</i>										X							
<b>UNGULATA</b>																	
<i>Protungulatum</i>		X	X	X													
<b>DINOCERATA</b>																	
<b>Uintatheriidae</b>																	
<i>Prodinoceras</i>												X	X	X	X	X	→





TABLE 3.2 (continued)

TAXON	L+	PU1	PU2	PU3	T01	T02	T03	T11	T12	T13	T14	T15	T16	CF1	CF2	CF3	W+
<i>Ectocion</i>								X	X	X	X	X	X	X	X	X	→
<i>Meniscotherium</i>																X	→
<i>Copecion</i>										X							
<b>Periptychidae</b>																	
<i>Mimatuta</i>		X	X														
<i>Anisonchus</i>			X	X	X	X	X	X									
<i>Haploconus</i>			X	X	X	X	X										
<i>Mithrandir</i>			X	X													
<i>Hemithlaeus</i>		?	X														
<i>Ampliconus</i>		X	X														
<i>Auraria</i>		X															
<i>Ectoconus</i>			X	X													
<i>Alticonus</i>			X														
<i>Maiorana</i>		X															
<i>Periptychus</i>			X	X	X	X	X	X	O	X	X						
<i>Tinuviel</i>		?	X	X													
<i>Oxyacodon</i>		X	X	X													
<i>Conacodon</i>		X	X	X	O	O	O	O	?	?							
<b>ARCTOSTYLOPIDA</b>																	
<b>Arctostylopidae</b>																	
<i>Arctostylops</i>													X	O	O	X	X
<b>CETE</b>																	
<b>Family indeterminate</b>																	
<i>Microclaenodon</i>							X	X									
<b>Triisodontidae</b>																	
<i>Goniacodon</i>				X	X	X	X										
<i>Eoconodon</i>		X	X	X													
<i>Triisodon</i>					X	X											
<i>Stelocyon</i>								X									
<b>Mesonychidae</b>																	
<i>Ankalagon</i>						X	X										
<i>Dissacus</i>						X	X	X	X	X	X	X	X	X	X	X	→

Arrows indicate documented ranges earlier than Puercan or later than Clarkforkian. Genera listed have been reported in published literature or are in press (except *Auraria* from Middleton 1983). Unpublished faunal data from dissertations are included in a few cases. Symbols: X, known from zone; ?, questionably present; O, not known. Taxa are listed following McKenna and Bell (1997). It is beyond the scope of this work to provide a locality list for each genus listed. For a partial list of localities for each genus, see tables 3.1 and 3.2 in Archibald et al. (1987) and the more recently published references listed in table 3.1.

km thick, exposed in the Hanna Basin of Wyoming (Eberle and Lillegraven 1998a, 1998b). Eberle and Lillegraven report a series of 39 localities referable to the Lancian and interval zones Pu1, Pu2, and Pu3 of the Puercan. A mammalian assemblage referable to the Pu0 in-

terval zone was not found in the Ferris Formation, even in this unusually thick package of sedimentary rock.

Finally, in contrast to our decision to refer faunas with *Protungulatum* to the Puercan mammal age, Fox (1990c, 1997) argued that the mammalian assemblages contain-

ing *Protungulatum* from the Frenchman 1 and Long Fall sites in western Canada are Cretaceous in age and referable to the Lancian mammal age. Their Cretaceous age assignment was based on stratigraphic position (Johnston 1980; Johnston and Fox 1984; Fox 1990c, 1997) and palynology (Fox 1995, 1997). Thus, as we recognize it, the Puercan mammal age may span the Cretaceous–Tertiary boundary (i.e., the first occurrence of *Protungulatum* may have occurred in the Cretaceous in Canada or elsewhere in North America). In any case, the first appearance of ungulates (i.e., the advent of the Puercan mammal age) signals the beginning of a dramatic shift in composition of mammalian assemblages in North America (Alroy 1999). We see no conceptual reason why the Lancian–Puercan and Cretaceous–Tertiary boundaries should necessarily coincide. Also, we see no reason why the first appearance of *Protungulatum* could not have occurred in the Late Cretaceous.

The number of known Puercan localities has increased greatly since publication of Wood et al. (1941) and Archibald et al. (1987). New Puercan faunas or additional taxa reported from previously known sites have been reported from the Javelina Member of the Tornillo Formation of Texas (Standhardt 1986; Schiebout et al. 1987), the North Horn Formation of central Utah (Robison 1986; Cifelli et al. 1995, 1999), the Ferris Formation of south-central Wyoming (Eberle 1999; Eberle and Lillegraven 1998a, 1998b; Lillegraven and Eberle 1999), the Fort Union Formation near Cedar Mountain, Wyoming (Hartman 1986), the Bear Formation in south central Montana (Buckley 1994, 1995, 1997), the upper Hell Creek and lower Tullock formations of eastern Montana (Lofgren 1995; Rigby 1989; Sloan et al. 1986), the Ludlow Member of the Fort Union Formation of southeastern Montana (Hunter et al. 1997), the Ludlow Formation of North Dakota (Hunter 1999), the Ravenscrag Formation of Saskatchewan (Fox 1990c, 1997), and the Nacimiento Formation of New Mexico (Williamson 1996).

## DEFINITION AND CHARACTERIZATION

We define the Puercan mammal age to include faunas that occurred between first appearance of the arctocyonid condylarth *Protungulatum* and first appearance of the periptychid condylarth *Periptychus carinidens*.

Wood et al. (1941) noted that the following taxa first appeared in the Puercan mammal age: *Anisonchus*, Condylarthra, Creodonta, *Eucosmodon*, *Oxyclaenus*, Taeniodonta, and Taligrada. The taxa in this list are still thought to have appeared during Puercan time, although not necessarily at the earliest sites referable to this age.

Of the four orders listed as first appearances, only the Taeniodonta retains a taxonomic usage comparable to that applied by Wood et al. in 1941 (Archibald et al. 1987). This order is still considered to have appeared in the Puercan mammal age but not at the earliest sites (Eberle 1999). Assuming that Wood et al. (1941) followed Matthew's (1937) concept of the Condylarthra, Creodonta, and Taligrada, then, following the classification by McKenna and Bell (1997), Puercan representatives of Matthew's Condylarthra are now included in the condylarthran families Mioclaenidae (*Protoselene*, *Ellipsodon*) and Periptychidae (*Oxyacodon*). According to McKenna and Bell (1997), Matthew's Creodonta are now included in the orders Procreodi (Oxyclaenidae: *Carcinodon*, *Oxyclaenus*, *Chriacus* including *Tricentes*, and Arctocyonidae: *Loxolophus*) and Cete (Triisodontidae: *Eoconodon*, *Goniacodon*), and Matthew's Taligrada are now included in the condylarthran family Periptychidae (*Haploconus*, *Anisonchus*, *Hemithlaeus*, *Periptychus*, *Ectoconus*, *Conacodon*). Taligrada is no longer used in standard mammalian taxonomy (see McKenna and Bell 1997).

In addition to first appearances, Wood et al. (1941) identified the following as index fossils thought to have been limited to the Puercan mammal age: *Carsiptychus*, *Conacodon*, *Ectoconus*, *Eoconodon*, *Loxolophus*, *Onychodectes*, *Oxyacodon*, *Taeniolabis*, and *Wortmania*. Of these genera, three are now known to occur in the Torrejonian or Torrejonian and Tiffanian mammal ages. *Loxolophus* has been reported from the Dragon Local Fauna, North Horn Formation of Utah (Gazin 1941; Robison 1986) and Swain Quarry of Wyoming (Rigby 1980), both of Torrejonian age (Archibald et al. 1987). *Conacodon* has been identified from the Laudate Local Fauna in the Tiffanian part of the Goler Formation of California (McKenna 1955; McKenna and Lofgren in press). McKenna and Bell (1997) consider *Carsiptychus* a junior synonym of *Periptychus*, which extended into the Tiffanian mammal age (Archibald et al. 1987).

Archibald et al. (1987) presented a list of first and last appearances, index fossils, and characteristic fossils for the Puercan mammal age. Our updated list reflects new knowledge of temporal ranges, new genera described since 1987, and the following taxonomic and biostratigraphic considerations. First, in addition to the synonymy of *Periptychus* and *Carsiptychus*, McKenna and Bell (1997) recognized the following generic synonyms: *Mithrandir* (*Gillisonchus*), *Oxyclaenus* (*Thangorodrim*), *Oxyacodon* (*Escatepos*), *Mimatuta* (*Earendil*), and *Ellipsodon* (*Bomburia*). Second, we recognize *Ragnarok* as a junior synonym of *Baioconodon*, an idea first proposed by Middleton (1983), formalized by Hunter et al. (1997),

and accepted by McKenna and Bell (1997). Third, the occurrence of *Gypsonictops*, *Alostera*, *Batodon*, *Didelphodon*, *Glasbius*, *Pedimys*, *Alphadon*, *Turgidodon*, *Essonodon*, *Meniscoessus*, and *Cimolomys* in strata representing the Puercan mammal age probably is the result of reworking because their Puercan records are limited to channels that were incised into Lancian strata (Lofgren 1995). Finally, as a result of changing the definition of the beginning of the Puercan mammal age to the first appearance of *Protungulatum*, the following genera represent Puercan index fossils rather than taxa that first appeared in the Lancian and last appeared in the Puercan mammal ages: *Baioconodon* (including *Ragnarok*), *Mimatuta*, *Oxyprimus*, *Procerberus*, *Protungulatum*, and *Purgatorius*.

A combined biochronologic–magnetostratigraphic correlation of Puercan local faunas from the Nacimiento Formation, San Juan Basin, New Mexico (Butler and Lindsay 1985; Williamson and Lucas 1992; Williamson 1996); North Horn Formation, central Utah (Tomida and Butler 1980); Tullock Formation, northeastern Montana (Archibald et al. 1982; Swisher et al. 1993); Fort Union Formation, northern Wyoming (Butler et al. 1987); and Ludlow Formation, southeastern Montana (Hunter et al. 1997) indicates that Puercan mammals first occur and last occur in strata of reversed polarity (Tomida and Butler 1980; Swisher et al. 1993). We interpret these data to signify that the Puercan mammal age (Pu1 interval zone) began during magnetic polarity chron C29r and ended (To1 interval zone) in chron C28r.

Puercan local faunas in the upper Hell Creek and Tullock formations in eastern Montana have been correlated to stratigraphic units (in this case lignites) that contain bentonitic and volcanic crystal rich partings suitable for argon–argon dating (Swisher et al. 1993). These data indicate that the entire Puercan mammal age was approximately 1 million years in duration (Swisher et al. 1993).

Further discussion of biochronologic–paleomagnetic correlations is provided in the appropriate sections of this chapter dealing with interval zones Pu1, Pu2, and Pu3 of the Puercan mammal age.

First and last appearances, index fossils, and fossils characteristic of the Puercan mammal age are as follows.

First appearances: *Anisonchus*, *Catopsalis*, *Chriacus*, *Conacodon*, *Desmatoclaenus*, *Ectypodus*, *Ellipsodon*, *Eucosmodon*, *Goniacodon*, *Haplaletes*, *Haploconus*, *Ictidopappus?*, *Leptacodon?*, *Litaletes?*, *Litomyilus*, *Loxolophus*, *Microcosmodon*, *Mimotricentes*, *Oxyclaenus*, *Palaeictops*, *Peradectes*, *Periptychus*, *Prodiacodon*, *Promioclauenus*, *Protoselene?*, *Ptilodus*, and *Xyronomys*

Last appearances: *Alostera?*, *Alphadon?*, *Batodon?*, *Cimexomys*, *Cimolestes*, *Cimolodon?*, *Cimolomys?*, *Didelphodon?*, *Essonodon?*, *Glasbius?*, *Gypsonictops?*, *Kimbetohia*, *Meniscoessus?*, *Pedimys?*, *Turgidodon?*, and *Viridomys*

Index fossils: *Acheronodon*, *Alticonus*, *Alveugena*, *Ampliconus*, *Auraria*, *Baioconodon*, *Bubogonia*, *Carcinodon*, *Choeroclaenus*, *Ectoconus*, *Eoconodon*, *Hemithlaeus*, *Maiorana*, *Mimatuta*, *Mithrandir*, *Onychodectes*, *Oxyacodon*, *Oxyprimus*, *Pandemonium*, *Platymastus*, *Procerberus*, *Protungulatum*, *Purgatorius*, *Ravenictis*, *Schochia*, *Taeniolabis*, *Tinuviel*, *Tiznatzinia*, and *Wortmania*

Characteristic fossils: *Mesodma*, *Neoplagiaulax?*, *Parectypodus*, and *Styгимys*

## ZONATION

We recognize three interval zones in the Puercan mammal age. From oldest to youngest, they are the *Protungulatum/Ectoconus* Interval Zone (Pu1), the *Ectoconus/Taeniolabis taoensis* Interval Zone (Pu2), and the *Taeniolabis taoensis/Periptychus carinidens* Interval Zone (Pu3).

***Protungulatum/Ectoconus* Interval Zone (Pu1)** We define the *Protungulatum/Ectoconus* Interval Zone to include faunas that occurred between the first appearance of *Protungulatum* and the first appearance of *Ectoconus*. The Pu1 interval zone is represented by the following sets of localities and local faunas: Mantua Lentil Local Fauna, Leidy Quarry, and sites low in the Fort Union Formation near Cedar Mountain, Bighorn Basin, northern Wyoming; Hell's Hollow Local Fauna, McKeever Ranch localities, Bug Creek Anthills and correlatives (Bug Creek West, Harbicht Hill, Ferguson Ranch, Chris's Bonebed), Morales 1 and Herpijunk Promontory, McGuire Creek local faunas, Z-line Channel Local Fauna, Jacks Channel Local Fauna, northeastern Montana; Littleton Local Fauna, Denver Basin, central Colorado; Long Fall and Frenchman 1 sites in western Canada; and a series of localities in the Hanna Basin, south-central Wyoming.

The Mantua Lentil Local Fauna from the Fort Union (= Polecat Bench) Formation was first described by Jepsen (1930, 1940). Van Valen (1978) later provided abbreviated descriptions of a number of new species of ungulates and a list of ungulates present in the local fauna (*Oxyprimus*, *Baioconodon*, *Eoconodon*, *Maiorana*, *Mimatuta*, and *Oxyacodon*). Van Valen (1978) also briefly described the mammalian fauna from the Leidy Quarry in the southern Bighorn Basin, a locality also referable to the Pu1 interval zone. Leidy Quarry yielded specimens

that are referable to *Baioconodon* and *Mimatuta*. More recently, J. E. Hartman (1986) described a sparse fauna containing *Mesodma*, *Baiotomeus*, *Peradectes*, *Oxyclaenus*, and *Oxyprimus* from four sites low in the Fort Union Formation near Cedar Mountain, also in the southern Bighorn Basin. Presence of the ungulate *Oxyprimus* suggests an early Puercan age because this is an index fossil for the Pu1 interval zone based on current knowledge. However, *Baiotomeus* is reported from the same locality (Hartman 1986), and this genus is known only from the early Tiffanian elsewhere. Thus these sites low in the Fort Union Formation from Cedar Mountain probably are Puercan in age (Hartman 1986), but there are not enough data available to refer them to a specific interval zone.

Beginning with a brief description of the mammalian fauna from Bug Creek Anthills and correlatives by Sloan and Van Valen in 1965, the uppermost Hell Creek and the lower Tullock formations in northeastern Montana have yielded many sites referable to the Pu1 interval zone. From the lower Tullock Formation in Garfield County, Archibald (1982) described the Hell's Hollow Local Fauna and a sparse fauna from the McKeever Ranch localities, and he noted the presence of *Baioconodon* from Morales 1. From McCone County further to the east, Sloan and Van Valen (1965), Lupton et al. (1980), Sloan et al. (1986), Rigby (1989), Fox (1989), Luo (1991), and Lofgren (1995) provided lists or descriptions of taxa from the Bug Creek Anthills locality and nearby correlatives (Bug Creek West, Harbicht Hill, Ferguson Ranch, Chris's Bonebed) from the upper Hell Creek Formation. Also, Lofgren (1995) described a series of mammalian faunas from the upper Hell Creek Formation from the McGuire Creek area just south of Bug Creek, which are referred to here as the McGuire Creek and Z-line local faunas. Lofgren (1995) also provided a list of the small mammalian assemblage from the Jacks Channel Local Fauna, which consists of two sites in the lower Tullock Formation near McGuire Creek. All these sites or local faunas are referred to the Pu1 interval zone based on presence of the multituberculates *Stygimys*, *Catopsalis*, *Mesodma*, *Cimexomys*, and *Acheronodon* (Hells Hollow only), the cimolestid *Procerberus*, the marsupial *Peradectes*, the ungulates *Protungulatum*, *Baioconodon*, *Oxyprimus*, and *Mimatuta*, and the primate *Purgatorius* (occurrence at Harbicht Hill is questionable). The presence of Lancian taxa at some of these sites (i.e., *Meniscoessus*, *Essonodon*, *Cimolodon*, *Cimolomys*, *Alphadon*, *Turgidodon*, *Pedionomys*, *Glasbius*, *Didelphodon*, *Gypsonictops*, *Alostera*, and *Batodon*) probably is the result of reworking (Lofgren 1995).

The Littleton Local Fauna from the Denver Formation in central Colorado was described by Middleton (1983) based on specimens collected from South Table Mountain and the Alexander Locality. In this mostly unpublished dissertation, Middleton (1983) described 13 new mammalian species and two new genera (*Auraria* and *Ampliconus*). He also argued based on phyletic relationships that the Littleton Local Fauna is younger than the Mantua Lentil Local Fauna but older than the type Puercan faunas of the San Juan Basin or the faunas that define the Pu2 and Pu3 interval zones. Therefore Archibald et al. (1987) assigned the Littleton Local Fauna to the Pu1 interval zone.

Two sites from southern Saskatchewan are of special interest to discussions of the Cretaceous–Tertiary and Lancian–Puercan boundaries. These localities are Frenchman 1, Frenchman Formation, and Long Fall, Ravenscrag Formation. The sample of mammals from Frenchman 1 is small and contains *Protungulatum* and mammals previously restricted to Lancian-aged strata (*Meniscoessus* and *Cimolodon*) along with the bones and teeth of dinosaurs (Johnston 1980; Fox 1988, 1990c, 1997). The mammalian sample from Long Fall is much larger and yields the ungulates *Protungulatum*, *Oxyprimus*, *Baioconodon*, and *Mimatuta*, Lancian mammals (*Alphadon*, *Pedionomys*, *Meniscoessus*, *Cimolomys*, *Cimolodon*, and *Gypsonictops*), and dinosaurs (Johnston and Fox 1984; Fox 1988, 1990c, 1997). We assign these sites to the Pu1 interval zone based on the presence of *Protungulatum* (and in the case of Long Fall, the other ungulates as well). Fox (1988, 1990c, 1997) argued that these sites are both Cretaceous and Lancian in age. A Cretaceous age for Frenchman 1 is further supported by a preliminary report based on palynology (Fox 1995). The Frenchman 1 and Long Fall sites may indeed be Cretaceous in age. However, we choose to define the boundaries of biochronologic units based on new appearances of mammals (see Woodburne 1977, 1987 for further discussion). Thus age assignment of assemblages such as Frenchman 1, Long Fall, and Bug Creek Anthills from eastern Montana to a particular interval zone does not require one to first determine whether the Lancian mammal component of the fauna may have been reworked (see Lofgren 1995 for discussion). Also, the first appearance of ungulates in North America signals the advent of a dramatic change in the taxonomic composition of its mammalian faunas. We use that event to define the beginning of the Puercan mammal age (i.e., the first appearance of *Protungulatum*) whether or not it occurs in the late Cretaceous or early Paleocene. This line of reasoning was first proposed by Archibald and Lofgren (1990) and further advocated by Lofgren (1995), Eberle

and Lillegraven (1998a), and Lillegraven and Eberle (1999).

Recently, Eberle and Lillegraven (1998a, 1998b) documented a series of stratigraphically superposed localities containing mammals typical of the Puercan mammal age from the type Ferris Formation in the Hanna Basin of south-central Wyoming. These mammalian fossils allowed a detailed biostratigraphic zonation of the Puercan section of the formation, which is approximately 540 m thick, an order of magnitude thicker than any other known of that age (Eberle and Lillegraven 1998a). Preserved in this thick section are mammalian assemblages that represent all three Puercan interval zones (Pu1 as defined here and Pu2 and Pu3 as defined by Archibald et al. 1987). In the Ferris Formation, Eberle and Lillegraven (1998a, 1998b) used the lowest stratigraphic occurrences of Puercan interval zone FADs *Protungulatum* (Pu1), *Ectoconus* (Pu2), and *Taeniolabis taoensis* (Pu3) to determine the bases of their three respective Puercan interval zones. As Eberle and Lillegraven (1998a, 1998b) noted, the lowest stratigraphic occurrence of *Ectoconus* is 77 m above that of *Protungulatum*, whereas the lowest stratigraphic occurrence of *Taeniolabis taoensis* is 240 m above that of *Ectoconus*. No other section in North America has mammalian faunas that represent Puercan interval zones Pu1, Pu2, and Pu3 in direct superposition. Documented differences in mammalian assemblages combined with unequivocal superposition of fossil-bearing localities from the Ferris Formation in the Hanna Basin help to confirm the distinctiveness of all three Puercan interval zones (Eberle and Lillegraven 1998a). This issue was a matter of debate concerning the Pu2 and Pu3 interval zones because it was questioned whether one could distinguish these interval zones outside their type areas in the San Juan Basin (Archibald et al. 1987). In addition to *Protungulatum*, Pu1 interval zone localities of the Ferris Formation have yielded specimens referred to *Mesodma*, *Peradectes*, *Cimolestes*, *Oxyprimus*, *Mimatuta*, *Maiorana*, and *Eoconodon*.

Of the three states (Montana, Wyoming, Colorado) and one province (Saskatchewan) in which the Pu1 interval zone can be recognized, only local faunas in northern Wyoming (Mantua Lentil) and eastern Montana have been correlated to the Geomagnetic Polarity Time Scale (GPTS). In these areas, Pu1 local faunas lie in a reversed magnetozone that has been correlated to magnetic polarity chron C29r (Archibald et al. 1982; Butler et al. 1987; Swisher et al. 1993). The entire Pu1 interval zone may lie in magnetic polarity chron C29r because the oldest part of the *Ectoconus/Taeniolabis taoensis* Interval Zone (Pu2) in the San Juan Basin is contained in strata of normal po-

larity that are correlated to magnetic anomaly chron C29n (Butler and Lindsay 1985; Williamson and Lucas 1992). Also, Hunter et al. (1997) suggested that the mammalian assemblage from the Hiatt local fauna (Ludlow Formation, southeastern Montana), which they refer to the Pu2 interval zone, may occur in strata of reversed polarity, which they correlate with magnetic polarity chron C29r. If they are correct, then the entire Pu1 interval zone would indeed lie in chron C29r.

In eastern Montana, Pu1 local faunas have been correlated to lignites that contain volcanic units suitable for argon-argon dating (Swisher et al. 1993). Data presented by Swisher et al. (1993) indicate that the Pu1 interval zone was approximately 400,000 years in duration.

First appearances: *Ampliconus*, *Baiococonodon*, *Catopsalis*, *Conacodon*, *Eoconodon*, *Hemithlaeus?*, *Mimatuta*, *Oxyacodon*, *Oxyclaenus*, *Peradectes*, *Procerberus*, *Protungulatum*, *Purgatorius?*, and *Tinuviel?*

Last appearances: *Alostera?*, *Alphadon?*, *Batodon?*, *Cimolomys?*, *Didelphodon?*, *Essonodon?*, *Glasbius?*, *Gypsonictops?*, *Meniscoessus?*, *Pedionomys?*, and *Turgidodon?*

Index fossils: *Acheronodon*, *Auraria*, *Maiorana*, and *Oxyprimus*

Characteristic fossils: *Cimexomys*, *Cimolestes*, *Mesodma*, and *Stygmimys*

Taxa absent but known before and after Pu1: *Cimolodon?*, *Kimbetohia*, *Neoplagiaulax?*, *Parectypodus*, and *Viridomys?*

***Ectoconus/Taeniolabis taoensis* Interval Zone (Pu2)** We define the *Ectoconus/T. taoensis* Interval Zone to include faunas that occurred between the first appearance of *Ectoconus* and the first appearance of *T. taoensis*.

In 1892, Wortman recognized two faunal zones for Puercan assemblages in what are now called the De-na-zin and Alamo washes (see Osborn and Earle 1895; Sinclair and Granger 1914). These are the lower “*Ectoconus* zone” (= “*Hemithlaeus* zone” of Van Valen 1978) and the upper “*Taeniolabis* zone” (Sinclair and Granger 1914; Matthew 1937). Archibald et al. (1987) used these zones as the basis for the naming and characterization of the *Ectoconus/Taeniolabis taoensis* (Pu2) and the *Taeniolabis taoensis/Periptychus* (Pu3) interval zones. However, they did so with caution because faunal differences between the zones were slight and there was concern that the difference between the zones may have resulted more from ecological variation than from significant differences in age (Lindsay et al. 1981; Archibald et al. 1987). There was little doubt that the interval zones were superposed in De-na-zin and Alamo washes and

represented some difference in time, but the absence of the multituberculate *Taeniolabis* in the other two major collecting areas for the type Puercan fauna, the Kimbetoh and Betonnie–Tsosie arroyos, supported the view that the differences may have been ecological. Also, in strata located along the West Fork of Gallegos Canyon (another collecting area for Puercan mammals northwest of De-na-zin and Alamo washes; locality A in column 1 of figure 3.2), Lucas (1984) described a small assemblage of mammals that included *Taeniolabis*. However, stratigraphically below the fauna that included *Taeniolabis*, Lucas (1984) was unable to locate a mammalian assemblage referable to the *Ectoconus*/*Taeniolabis taoensis* Interval Zone (Pu2). These data presented Archibald et al. (1987) with a problem regarding the Puercan mammal age: the inability to clearly identify zones within the type Puercan mammal age of the San Juan Basin and then to extend this zonation beyond the limits of that basin. Also, paleomagnetic analysis adds little to differentiation of Pu2 and Pu3 interval zones in the San Juan Basin because both interval zones occur in strata of normal polarity that is correlated with magnetic chron C29n (Lindsay et al. 1981; Butler and Lindsay 1985; Williamson and Lucas 1992). Accordingly, Archibald et al. (1987) defined the Pu2 and Pu3 interval zones with the caveat that revision may be needed and that they should be used provisionally in other parts of the San Juan Basin and western North America.

Reanalysis of the Puercan sites and establishment of a local biostratigraphic zonation in the San Juan Basin (Williamson 1996) and the recent discovery of new Puercan local faunas from the Ferris Formation of Wyoming (Eberle and Lillegraven 1998a, 1998b) provide support for subdividing the middle and late parts of the Puercan mammal age into the Pu2 and Pu3 interval zones. Concerning the possibility that the “*Ectoconus* zone” or Pu2 interval zone (H–T zone of Williamson 1996) and the “*Taeniolabis* zone” or Pu3 interval zone (T–P zone of Williamson 1996) represented ecological differences rather than differences in time, Williamson (1996:54) argued that “there is no doubt that the two faunas are superposed and separated by a barren interval of approximately 25 meters in the De-na-zin Wash section” and that 25 m of sedimentation suggests that a significant interval of time elapsed between the deposition of the two faunas. Williamson (1996:54) also stated that “there are no significant facies changes between the two fossiliferous zones and therefore no direct evidence to indicate different environments,” data that further support the biochronologic utility of the Pu2 and Pu3 interval zones.

The first record of superposed mammalian faunas referable to the Pu2 and Pu3 interval zones in an area outside the San Juan Basin comes from the type Ferris Formation in the Hanna Basin of Wyoming (Eberle and Lillegraven 1998a, 1998b). Preserved in a 540-m-thick section of the formation are mammalian assemblages indicating that the lowest stratigraphic occurrence of *Taeniolabis taoensis* is 240 m above the lowest stratigraphic occurrence of *Ectoconus* (similarly, the lowest stratigraphic occurrence of *Ectoconus* is 77 m above that of *Protungulatum*; Eberle and Lillegraven 1998a, 1998b). The Hanna Basin is the only area in North America in which all three Puercan interval zones are found in superposition. Thus mammalian faunas from the type Ferris Formation help to confirm the distinctiveness of the Pu2 and Pu3 interval zones outside the San Juan Basin and provide a unique reference section for the entire Puercan mammal age.

In the San Juan Basin, mammalian faunas that form the basis for the definition of the Pu2 interval zone are found in the Arroyo Chijuillita Member of the Nacimiento Formation, located in De-na-zin, Alamo, Kimbetoh, and Betonnie–Tsosie washes (Williamson 1996). Based on a list of therian taxa and their biostratigraphic ranges provided by Williamson (1996), it should be noted that the Pu2 mammalian fauna is distinguished from the Pu3 mammalian fauna primarily by species-level differences in ungulate genera. Three genera (the tillodont *Schochia* and the ungulates *Platymastus* and *Hemithlaeus*) have unique occurrences in the Pu2 interval zone, but *Schochia* and *Platymastus* are known only from their holotypes (Williamson 1996) and represent taxa with limited biochronologic utility. Thus detailed taxonomic comparisons of ungulate species are needed before biochronologic correlations to the Pu2 interval zone of the San Juan Basin from elsewhere in North America can be made with confidence.

In the Hanna Basin of Wyoming, 240 m of the Ferris Formation represent strata referred to the Pu2 interval zone by Eberle and Lillegraven (1998a, 1998b). The multituberculates *Ptilodus* and *Ectypodus* and the ungulates *Loxolophus*, *Mithrandir*, *Oxyacodon*, *Conacodon*, *Ectoconus*, *Periptychus*, and *Promioclænus* are genera that appear in Pu2 strata from the Ferris Formation that are typical elsewhere of the Pu2 interval zone. A net result of these introductions is that, compared with the Pu1 interval zone, the Pu2 interval zone is characterized by more than two times more diversity of ungulate genera and by greater body size and dental specialization (Eberle and Lillegraven 1998a).

From the Denver Formation in the Denver Basin of eastern Colorado, Middleton (1983) described the mam-

malian assemblages found at the Corral Bluffs and West Bijou Creek-1 localities. These sparsely fossiliferous sites share some ungulate species in common with the Pu2 and Pu3 interval zones of the San Juan Basin, but the multi-tuberculate *Taeniolabis* has not been recovered from either locality. Therefore Corral Bluffs or West Bijou Creek-1 cannot be assigned to the Pu2 or Pu3 interval zone with confidence.

The North Horn Formation contains the Gas Tank Local Fauna, named by Robison (1986) after the Gas Tank Hill locality (Van Valen 1978; = Flagstaff Peak Locality in Spieker 1960). The Gas Tank Local Fauna is composed of a series of widely separated localities grouped based on general stratigraphic position (distance in meters below the base of the overlying Flagstaff Peak Formation) and faunal similarity (Robison 1986). We suggest caution in grouping these localities and analyzing them as one local fauna because most sites referred to the Gas Tank Hill Local Fauna occur in areas where the strata have been slumped (see Robison 1986). Archibald et al. (1987) tentatively referred the Gas Tank Local Fauna to the Pu2 interval zone because it is lower stratigraphically than the Wagonroad Local Fauna, and the latter fauna contains *Taeniolabis* (see Gazin 1941; Tomida and Butler 1980). However, Williamson (1996) noted that of the five ungulate species the Gas Tank Local Fauna shares with Puercan faunas in the San Juan Basin, only *Loxolophus pentacus* shows a range limited to either the Pu2 or Pu3 interval zone, in this case the Pu2 interval zone. Also, Robison (1986) tentatively referred an incisor fragment from the Ferron Mountain Locality of the Gas Tank Local Fauna to cf. *Taeniolabis taoensis*. If this identification is correct, then at least one locality (Ferron Mountain) of the Gas Tank Local Fauna would be referable to the Pu3 interval zone. Therefore we tentatively follow Archibald et al. (1987) and refer the Gas Tank Local Fauna to the Pu2 interval zone, recognizing that much more work must be done to confirm this correlation.

From the lower part of the Ludlow Formation in southwestern North Dakota, Hunter (1999) reported a sparse mammalian assemblage from the Pita Flats locality. The presence of *Oxyacodon priscilla* at Pita Flats indicates a middle to late Puercan age or referral to the Pu2 or Pu3 interval zone (Hunter 1999).

In southeastern Montana near Glendive, Hunter et al. (1997) described the mammalian assemblage from the Hiatt Local Fauna located in the lower part of the Ludlow Member of the Fort Union Formation. Hunter et al. (1997) argued that the diverse assemblage of archaic ungulates in the Hiatt Local Fauna in conjunction with the absence of *Taeniolabis* suggests that the Hiatt Local Fauna

is referable to the Pu2 interval zone. They also suggested that the Hiatt Local Fauna may occur in strata of reversed polarity that they correlated with magnetic polarity chron C29r. Because Pu2 faunas elsewhere are known only from strata of normal polarity correlated with magnetic chron C29n, if the Hiatt Local Fauna is indeed in strata of reversed polarity representing chron C29r, it could not be younger than Pu2.

In addition to the Hiatt Local Fauna, the only other place at which local faunas referred to the Pu2 interval zone have been correlated to the magnetic anomaly correlation scale occur in the San Juan Basin. These local faunas lie in strata of normal polarity correlated with magnetic anomaly chron C29n (Lindsay et al. 1981; Butler and Lindsay 1985; Williamson and Lucas 1992).

First appearances: *Anisonchus*, *Bubogonia*, *Carcinodon*, *Choeroclaenus*, *Chriacus*, *Desmatoclaenus*, *Ectoconus*, *Ectypodus*, *Ellipsodon*, *Eucosmodon*, *Haploconus*, *Ictidodopappus?*, *Litomylus*, *Loxolophus*, *Microcosmodon*, *Mimotricentes*, *Mithrandir*, *Onychodectes*, *Pandemonium*, *Periptychus*, *Promioclænus*, *Ptilodus*, *Tiznatzinia*, *Wortmania*, and *Xyromys*

Last appearances: *Alticonus*, *Ampliconus*, *Hemithlaeus*, and *Mimatuta*

Index fossils: *Alveugena*, *Platymastus*, and *Schochia*

Characteristic fossils: *Baiocodon*, *Catopsalis*, *Cimexomys*, *Cimolestes*, *Conacodon*, *Eoconodon*, *Kimbetohia*, *Mesodma*, *Neoplagiaulax*, *Oxyacodon*, *Oxyclaenus*, *Parectypodus*, *Peradectes*, *Procerberus?*, *Protungulatum*, *Purgatorius*, *Stygimys*, and *Tinuviel*

Taxa absent but known before and after Pu2: *Cimolodon?* and *Viridomys?*

***Taeniolabis taoensis*/*Periptychus carinidens* Interval Zone (Pu3)** We define the *T. taoensis*/*P. carinidens* Interval Zone to include faunas that occurred between the first appearance of *T. taoensis* and the first appearance of *P. carinidens*.

In the San Juan Basin, mammalian assemblages that comprise the type fauna of the Pu3 interval zone (= T-P interval zone of Williamson 1996) are located in the Arroyo Chijuillita Member of the Nacimiento Formation in De-na-zin and Alamo washes and in the West Fork of Gallegos Canyon (Williamson 1996). Based on faunal data presented by Williamson (1996), the Pu2 and Pu3 interval zones in the San Juan Basin differ in the following ways. First, the tillodont *Schochia* and the ungulates *Hemithlaeus* and *Platymastus* are not present in Pu3 faunas; *Hemithlaeus* is common in Pu2 faunas, but *Platymastus* and *Schochia* are rare. Second, a number of un-

gulate species that occur in Pu2 did not persist into the Pu3 interval zone. Finally, the ungulates *Loxolophus pentacus*, *L. priscus*, and *Oxyclaenus antiquus* and the multi-tuberculate *Taeniolabis taoensis* are index taxa for the Pu3 interval zone. Because the Pu2 and Pu3 interval zones are faunally similar, the fact that *Taeniolabis taoensis* is limited to the Pu3 interval zone in the San Juan Basin becomes, by default, a critical factor in assigning middle to late Puercan faunas elsewhere in North America to the Pu2 or Pu3 interval zone. Paleomagnetic correlations in the San Juan Basin offer little assistance in this regard because both the Pu2 and Pu3 interval zones are located in strata of normal polarity correlated to magnetic anomaly chron C29n (Lindsay et al. 1981; Butler and Lindsay 1985; Williamson and Lucas 1992).

The Wagonroad Local Fauna from the North Horn Formation in Utah was first described by Gazin (1941), with additions from Tomida and Butler (1980), Robison (1986), and Cifelli et al. (1995). The Wagonroad Local Fauna is small and shares only three species with Puercan faunas from the San Juan Basin (Williamson 1996), making a firm biochronologic correlation difficult. However, Gazin (1941:8) described a broken first lower molar from the Wagonroad Local Fauna and referred it only to the generic level (*Taeniolabis*) even though "in size and appearance the specimen closely resembles this portion of the m1 in *Taeniolabis taoensis* from the Puerco of New Mexico." Therefore, although the Wagonroad Local Fauna may contain *Taeniolabis taoensis*, better material is needed for confirmation. Paleomagnetic evidence indicates that strata containing the Wagonroad Local Fauna straddle a transition from normal to reversed polarity that is correlated with the transition from magnetic polarity chron C29n to C28r (Tomida and Butler 1980). Based on both biochronologic (presence of *Taeniolabis*) and paleomagnetic correlations (Pu3 in San Juan Basin is in C29n), we refer the Wagonroad Local Fauna to the Pu3 interval zone.

In the section of the Ferris Formation assigned to the Pu3 interval zone (more than 200 m thick) based on the lowest stratigraphic occurrence of *Taeniolabis taoensis*, there is not as dramatic a difference in composition of mammalian assemblages between Pu2 and Pu3 as there is between Pu1 and Pu2. Also, the number of ungulate genera remains nearly constant between Pu2 and Pu3 (Eberle and Lillegraven 1998a). It is interesting to note that the only known occurrence of *Protoselene* in the Puercan mammal age is based on a dentary fragment referred to cf. *Protoselene* from the Pu3 interval zone in the Ferris Formation (Eberle and Lillegraven 1998a). As to the importance of the Ferris Formation to Puercan biochronology, we again stress that superposition of localities in the Fer-

ris Formation yielding Pu2 and Pu3 mammalian assemblages strongly supports the reality of discrete Pu2 and Pu3 interval zones. This northward extension of the Pu2 and Pu3 interval zones from the San Juan Basin into southern Wyoming suggests that we may be able to recognize discrete Pu2 and Pu3 interval zones even further north (Canada and Montana), although the significant biogeographic provinciality evident in the late Puercan makes correlations into Canada and Montana very tentative.

Based on a well-preserved taeniolabidid dentary from the Bechtold Site, Ludlow Formation, southeastern Montana, Simmons (1987) described a second species of *Taeniolabis*, *T. lamberti*. The presence of at least one species of *Taeniolabis* distinct from *T. taoensis* outside the San Juan Basin does not necessarily indicate that this or other sites with this species (Garbani Local Fauna, discussed later in this chapter) are correlatives of the Pu3 interval zone. However, in the absence of well-documented evidence showing that *Taeniolabis* occurs in a fauna referable to an interval zone other than Pu3, we tentatively refer the Bechtold Site and others sites yielding *Taeniolabis* (see further discussion later in this chapter) to the Pu3 interval zone.

The Purgatory Hill and Garbani local faunas from the Tullock Formation in northeastern Montana appear to be correlatives of the Pu3 interval zone. The Purgatory Hill Local Fauna was the first Puercan fauna in Montana to be discovered and was briefly described by Van Valen and Sloan (1965) and Van Valen (1978). Sloan (1970) assigned the Purgatory Hill Local Fauna a late Puercan age based in part on the presence of *Taeniolabis* (fragment of an M2). This specimen was described and referred to *Taeniolabis* by Simmons (1987). The Garbani channel complex is more fossiliferous than the Purgatory Hill site, and Simmons (1987) described seven isolated teeth of *Taeniolabis* from the Garbani channel complex, two of which she referred to *Taeniolabis lamberti* and the other five to *Taeniolabis* species. Although Clemens (2002) presented an updated list of the mammalian assemblages for both the Garbani and Purgatory Hill local faunas, in reality few taxa in either fauna have been thoroughly described (see Clemens 1974; Novacek 1977; Novacek and Clemens 1977; Archibald et al. 1983; Simmons 1987; Weil 1998), making comparisons to other faunas uncertain. This problem is exacerbated by the reality that a considerable level of biogeographic provinciality exists in the Puercan mammal age, especially between northern and southern faunas (Buckley 1994; Williamson 1996; Eberle and Lillegraven 1998a). Therefore we tentatively assign the Garbani and Purgatory Hill local faunas to the Pu3 interval zone, recognizing that our correlation is based primarily on the



occurrence of *Taeniolabis* in both faunas. In the absence of evidence to the contrary, we speculate that *Taeniolabis* has a biochronologic range limited to that of the Pu3 interval zone of the San Juan Basin. Both the Purgatory Hill and Garbani local faunas occur in strata of normal polarity correlated to magnetic anomaly chron C29n (Archibald et al. 1982; Swisher et al. 1993), which is consistent with a correlation to the Pu2 or Pu3 interval zone of the San Juan Basin.

The Rav W-1 and Croc Pot sites from the Ravenscrag Formation in southern Saskatchewan yield mammalian assemblages similar in composition to the Garbani and Purgatory Hill local faunas of eastern Montana and the Simpson Quarry from south central Montana. The fauna from the Rav W-1 site was described by Johnston (1980) and Johnston and Fox (1984), with updated lists provided by Fox (1990c, 1997). Johnston and Fox (1984) suggested that Rav W-1 is older than or occurs very early in the *Hemithlaeus* zone (Pu2 interval zone of Archibald et al. 1987), even though a species of *Taeniolabis* is present. They argued that the species of *Taeniolabis* from Rav W-1 (based on tooth fragments) and the Garbani Local Fauna (based on isolated teeth) are the same and that this species is more primitive than *Taeniolabis taoensis* from the type Puercan assemblage of the San Juan Basin. From her detailed analysis of *Taeniolabis*, Simmons (1987) noted that the ambiguous nature of dental characters and the limited material available for study make it impossible to determine whether *Taeniolabis lamberti* (Garbani and Bechtold sites), *Taeniolabis* sp. (Garbani, Purgatory Hill, and Rav W-1), or *T. taoensis* is more or less derived. Also, based on their interpretation of the stage of evolution of ungulate species, Johnston and Fox (1984) argued that some of the ungulate species at Rav W-1 are less derived than their generic counterparts from the *Hemithlaeus* zone (Pu2 interval zone) of the San Juan Basin. Later, Fox (1997:77) stated that the Rav W-1 mammals "correlate most closely with mid-Puercan (Pu2) faunas in the US, although species differences make comparisons with standard Puercan sequences of the San Juan Basin, New Mexico difficult." As with the Garbani and Purgatory Hill local faunas, correlation is limited by faunal provinciality, but we tentatively assign the Rav W-1 fauna to the Pu3 interval zone based on the presence of *Taeniolabis*. The mammalian fauna from the Croc Pot site is small and undescribed, but faunal lists provided by Fox (1990c, 1997) indicate that the site yields *Taeniolabis taoensis*, and its overall mammalian assemblage is similar to that of Rav W-1. Therefore we assign Croc Pot to the Pu3 interval zone. Magnetostratigraphic data are not available for the Rav W-1 and Croc Pot sites.

In a mostly unpublished dissertation, Buckley (1994) described a large mammalian assemblage from Simpson Quarry, located in the Bear Formation in the Crazy Mountains Basin of south-central Montana. The fauna from Simpson Quarry is most similar to that from Rav W-1 in Canada, but Simpson Quarry has not yielded *Taeniolabis* (see Buckley 1994). Taxonomic composition of the mammalian assemblage from Simpson Quarry also has close affinities to local faunas from Purgatory Hill and Garbani Quarry in Montana and the Croc Pot site in Canada. Taken together, these four sites or local faunas form a biogeographic province that differs from southern sites in the San Juan Basin and Utah (Buckley 1994). Because of the absence of *Taeniolabis* and the level of biogeographic provincialism between northern and southern middle-late Puercan faunas, we assign Simpson Quarry to an undifferentiated Pu2–Pu3 interval zone. Magnetostratigraphic analysis indicates that Simpson Quarry lies in strata of normal polarity correlated to magnetic anomaly chron C29n (Buckley 1994), which is consistent with a Pu2 or Pu3 interval zone assignment.

The Big Bend area of Texas may yield mammalian assemblages that are Puercan in age, but correlations are uncertain. In a mostly unpublished dissertation, Standhardt (1986) described small faunas from the Dogie, Glenn Eleven, and Tom's Top sites, located in the Javelina Member of the Tornillo Formation, and assigned them to the Puercan mammal age. The Javelina and Blacks Peak formations were reduced to member status in the Tornillo Formation by Schiebout et al. (1987). The taxa listed for each site by Standhardt (1986) show affinities to both the Puercan and Torrejonian mammal ages (Williamson 1996). Also, small sample sizes and fragmentary material make some identifications tenuous (see Williamson 1996). The three sites may be Puercan or Torrejonian. Tom's Top occurs in strata of reversed polarity that are tentatively correlated to magnetic polarity chron C28r (Standhardt 1986; Schiebout et al. 1987), which would support a late Puercan or early Torrejonian age for the site.

Magnetostratigraphic analysis of the Pu3 interval zone in the San Juan Basin indicates that the Pu3 interval zone occurs in strata of normal polarity interpreted as magnetic polarity chron C29n. Because the Pu2 interval zone also occurs in rocks of normal polarity correlated to magnetic polarity chron C29n, magnetostratigraphic analysis offers no support for differentiating the Pu2 and Pu3 interval zones.

First appearances: *Goniacodon*, *Haplaletes*, *Leptacodon*?, *Litaletes*, *Palaeictops*, *Prodiacodon*, and *Protoselene*?

Last appearances: *Baiiconodon*, *Bubogonia*, *Carcinodon*, *Choeroclaenus*, *Cimexomys*, *Cimolestes*, *Cimolodon*?, *Ec-*

*toconus*, *Eoconodon*, *Kimbetohia*, *Mithrandir*, *Onychodectes*, *Oxyacodon*, *Pandemonium*, *Procerberus*, *Protungulatum*, *Purgatorius*, *Tinuviel*, *Tiznatzinia*, and *Wortmania*

Index fossils: *Ravenictis* and *Taeniolabis*

Characteristic fossils: *Anisonchus*, *Catopsalis*, *Chriacus*, *Conacodon*, *Desmatoclaenus*, *Ellipsodon*, *Eucosmodon*, *Haploconus*, *Loxolophus*, *Mesodma*, *Microcosmodon*, *Mimotricentes*, *Neoplagiaulax*, *Oxyclaenus*, *Parectypodus*, *Peradectes*, *Periptychus*, *Promioclauenus*, and *Stygimys*

Taxa absent but known before and after Pu3: *Ectypodus*, *Ictidopappus?*, *Litomylus*, *Ptilodus*, *Viridomys?*, and *Xyronomys*

### TORREJONIAN MAMMAL AGE

Initially, the Torrejonian mammal age included only the *Pantolambda* zone of Osborn and Matthew (1909). Later, however, Sinclair and Granger (1914) recognized two faunal zones, a lower *Deltatherium* and an upper *Pantolambda* zone. The zones were questioned by Matthew (1937) and others, mainly because neither *Deltatherium* nor *Pantolambda* was abundant enough to warrant much confidence in their stratigraphic range.

Wood et al. (1941) defined the Puercan and Torrejonian NALMAs based on fossils from the "Puerco" and "Torrejon" formations in the San Juan Basin. Later, Simpson (1959) reported the presence of fossils of Torrejonian age from the type section of the Puerco Formation. Because the formations apparently were defined on flawed paleontologic and not lithologic criteria, the "Puerco" and "Torrejon" formations were abandoned and replaced by a redefined Nacimiento Formation (see Gardner 1910; Simpson 1959). But the concept of using the Puercan and Torrejonian faunas (within the redefined Nacimiento Formation) as reference assemblages for biochronologic units for part of the Puercan and Torrejonian mammal ages was retained (Archibald et al. 1987).

The concept of the Torrejonian mammal age has been substantially modified since Wood et al. (1941) with the inclusion of the Dragonian mammal age by Archibald et al. (1987). Wood et al. (1941) defined a Dragonian mammal age between the Puercan and Torrejonian mammal ages, based on the Dragon Local Fauna from the North Horn Formation of Utah. Because it was convincingly demonstrated that a Dragonian faunal correlative is present in the San Juan Basin (Tomida and Butler 1980; Tomida 1981), Archibald et al. (1987) recognized the tem-

poral equivalent to the Dragonian mammal age as the first interval zone in a redefined Torrejonian mammal age. Recent work in the Nacimiento and North Horn formations lends further support for inclusion of the Dragonian in the Torrejonian mammal age (see Williamson 1996; Lucas et al. 1997). Also, Williamson (1996:61) contends "that Gazin (1938, 1939, 1941), convinced of the intermediate age of the Dragonian fauna between known Puercan and Torrejonian faunas, was led to give taxonomic significance to any small difference in morphology." Therefore detailed reanalysis of the Dragon Local Fauna may result in synonymy of many species described by Gazin (1938, 1939, 1941) with those from other Puercan and Torrejonian sites (Williamson 1996). As a first step, a revised faunal list for the Dragon Local Fauna was provisionally provided by Williamson (1996, table 12), pending thorough revision.

Wood et al. (1941) recognized two correlatives of the Torrejonian mammal age: the "Lebo" and "Rock Bench." No correlatives were listed for the Dragonian mammal age. "Lebo" refers to the Lebo Member of the Fort Union Formation in Simpson's (1935a, 1937b) Crazy Mountain Field, Montana. (In present usage in the Crazy Mountains area, Lebo is elevated to formation status and Fort Union to group status.) Simpson's (1937b) "Lebo" is now considered to include both Torrejonian and Tiffanian local faunas. "Rock Bench" refers to the Rock Bench Quarry from the Fort Union Formation in the Bighorn Basin of Wyoming, a large quarry sample of Torrejonian age.

Archibald et al. (1987) listed the following as correlatives of the Torrejonian mammal age: Farrand Channel Local Fauna, Tullock Formation, northeastern Montana (Archibald 1982); Goler Formation, California (McKenna 1960; West 1976), now considered Tiffanian (McKenna and Lofgren in press); Porcupine Hills Formation, Alberta (Russell 1958; Krause 1978); and Fort Union Formation of Washakie Basin, Wyoming (Rigby 1980). Since publication of Archibald et al. (1987), a significant number of new Torrejonian localities or additions to existing localities have been reported from the Nacimiento Formation of New Mexico (Williamson 1996), Tornillo Formation of Texas (Standhardt 1986), North Horn Formation of Utah (Robison 1986), Hanna Formation of Wyoming (Secord 1998; Higgins 2000), Fort Union Formation of Wyoming (Hartman 1986; Leite 1992), Fort Union Group of the Crazy Mountains area of south-central Montana (Hartman and Krause 1993; Buckley 1994), Fort Union Formation of the Clark's Fork area of south-central Montana (Butler et al. 1987), Fort Union Formation of southeast Montana (Hunter et al. 1997),

Ludlow Formation of North Dakota (Hunter 1999), and Coalspur Formation of Alberta, Canada (Fox 1990c).

### PUERCAN-TORREJONIAN BOUNDARY

The beginning of the Torrejonian mammal age is recognized by appearance of the periptychid *Periptychus carinidens*. *P. carinidens* is a distinctive member of Torrejonian faunas, including the Dragon Local Fauna of Utah (see Williamson 1996 for discussion of synonymy of *P. carinidens* and *Periptychus gilmorei*) on which the Dragonian mammal age was based (Wood et al. 1941). Tomida and Butler (1980) established a magnetic polarity sequence in the North Horn Formation that indicated that the Dragon Local Fauna occurred in strata that span magnetic chron C28n and C27r. Also, Tomida (1981) described a small fauna from the San Juan Basin correlated to the upper part of magnetic chron C28n and base of chron C27r that yielded typical Torrejonian mammals. Because it was well documented that the Dragonian interval occurred in the San Juan Basin and yielded a Torrejonian mammalian assemblage (Tomida and Butler 1980; Tomida 1981), the Dragonian mammal age was assigned to the Torrejonian mammal age as the To1 interval zone (Archibald et al. 1987).

De-na-zin Wash is the only known area in the San Juan Basin in which the Pu2, Pu3, and To1 interval zones are in direct superposition (Williamson 1996). In De-na-zin Wash, *P. carinidens* is present 52.5 m above the Pu3 interval zone (see Section J in Williamson 1996). Sites correlated to the To1 interval zone also overlie Pu2 interval zone sites (by more than 70 m) in both Kimbeto and Betonnie-Tsosis washes (Lindsay et al. 1981; Williamson 1996). The highest stratigraphic records of Puercan mammals from the San Juan Basin occur in strata with normal polarity, interpreted as magnetic polarity chron C29n. The lowest records of *P. carinidens* in the San Juan Basin occur in the next higher normal polarity magnetozone, interpreted as magnetic polarity chron C28n. Therefore, a stratigraphic interval of more than 50 m, which entirely spans the duration of magnetic polarity chron C28r, separates Puercan and Torrejonian assemblages in the San Juan Basin.

### DEFINITION AND CHARACTERIZATION

We follow Archibald et al. (1987) by defining the Torrejonian mammal age to include faunas that occur during the time between the first appearance of the periptychid condylarth *P. carinidens* and the first appearance of the plesiadapid primate *Plesiadapis*.

As mentioned previously, Tomida and Butler (1980) demonstrated that strata of the North Horn Formation yielding the Dragon Local Fauna, type fauna for Dragonian mammal age, correlate with a stratigraphic level in the San Juan Basin containing typical Torrejonian mammals (Tomida 1981). Tomida (1981) also showed that the Dragonian level in the San Juan Basin is faunally distinct and designated this interval as the *Periptychus-Loxolophus* chronozone. As discussed later in this chapter, our *Periptychus carinidens/Protoselene opisthacus* Interval Zone (To1) is equivalent to the Dragonian mammal age, Tomida's *Periptychus-Loxolophus* chronozone, and the To1 interval zone of Archibald et al. (1987), except for tentative inclusion of American Museum of Natural History (AMNH) locality 230 of Mesa de Cuba in our To1 interval zone.

There have been changes in the taxonomic characterization of the Torrejonian mammal age (in addition to inclusion of Dragonian mammal age) since publication of Wood et al. (1941). Of the taxa listed by Wood et al. (1941) as first appearances for the Torrejonian mammal age (including Dragonian), five are now known from the Puercan (*Periptychus*, *Catopsalis*, *Haploconus*, *Ptilodus*, and *Chriacus*), and *Didymictis* is now known to first occur in the Tiffanian mammal age. Similarly, in last appearances the only change is the probable occurrence of *Eucosmodon* in the Tiffanian mammal age. The index and characteristic fossils for the Torrejonian mammal age of Wood et al. (1941) have remained unchanged.

Based especially on the Kutz Canyon section in the San Juan Basin, Torrejonian faunas appear in rocks of normal polarity (interpreted as magnetic polarity chron C28n), continue upward in strata of reversed polarity, and are last known in superjacent layers of normal polarity (interpreted as magnetic polarity chron C27n by Lindsay et al. 1981; Butler and Lindsay 1985; Williamson and Lucas 1992; Williamson 1996). This is supported by new paleomagnetic data from the Tornillo Formation of Texas (Standhardt 1986; Schiebout et al. 1987); Fort Union Group, Crazy Mountains, and Fort Union Formation, Clark's Fork Basin, both south-central Montana (Butler et al. 1987; Buckley 1994); and Tullock Formation, eastern Montana (Swisher et al. 1993).

The faunal characterization of the Torrejonian mammal age is as follows.

First appearances: *Acmeodon*, *Adunator*, *Anconodon*, *Aphronorus*, *Baiotomeus*, *Claenodon*, *Colpoclaenus*, *Coryphagus*, *Diacodon?*, *Dissacus*, *Elphidotarsius*, *Elpidophorus*, *Eudaemonema*, *Gelastops*, *Ignacius*, *Jepsenella*, *Krauseia*, *Litocherus*, *Mimetodon*, *Myrmecoboides*, *Neo-*

*claenodon*, *Palaechthon*, *Palaeoryctes*, *Palenochtha*, *Paletomus*, *Pantolambda*, *Pararyctes*, *Paromomys*, *Pentacodon*, *Phenacodus*, *Picrodus*, *Plesiolestes*, *Pronothodectes*, *Propalaeosinopa*, *Prothryptacodon*, *Protictis*, *Psittacotherium*, *Simpsonictis*, *Stilpnodon*, *Tetraclaenodon*, and *Unuchinia*

Last appearances: *Ellipsodon*, *Goniacodon*, *Haploconus*, *Ictidopappus*, *Loxolophus*, *Oxyclaenus*, *Stygmimys*, *Viridomys?*, and *Xyromomys*

Index fossils: *Anasazia*, *Ankalagon*, *Avunculus*, *Conoryctella*, *Conoryctes*, *Deltatherium*, *Deuterogonodon*, *Draconodus*, *Dracontolestes*, *Escavadodon*, *Huerfanodon*, *Leptonysson*, *Microclaenodon*, *Mioclauenus*, *Mixodectes*, *Oxytomodon*, *Premnoides*, *Stelocyon*, *Swaindelphys*, *Trisodon*, and *Xanoclomys*

Characteristic fossils: *Anisonchus*, *Catopsalis*, *Chriacus*, *Desmatoclaenus*, *Ectypodus*, *Eucosmodon*, *Haplaletes*, *Leptacodon*, *Litaletes*, *Litomyilus*, *Mesodma*, *Mimotricentes*, *Neoplagiaulax*, *Palaeictops*, *Parectypodus*, *Peradectes*, *Periptychus*, *Prodiacodon*, *Promioclauenus*, *Protoselene*, and *Ptilodus*

Taxa absent but known before and after the Torrejonian: *Conacodon?* and *Microcosmodon*

## ZONATION

Archibald et al. (1987) proposed three Torrejonian interval zones: the *Periptychus carinidens*/*Tetraclaenodon* Interval Zone (To1), the *Tetraclaenodon*/*Pantolambda* Interval Zone (To2), and the *Pantolambda*/*Plesiadapis praecursor* Interval Zone (To3). Their To1 interval zone was equivalent to both the *Periptychus*–*Loxolophus* chronozone of Tomida (1981) and the Dragonian mammal age of Wood et al. (1941). Their To2 interval zone was approximately equivalent to the *Deltatherium* zone of Osborn (1929) and the *Deltatherium* chronozone of Tomida (1981). Finally, their To3 interval zone was approximately equivalent to the *Pantolambda* zone of Osborn (1929) and the *Pantolambda* chronozone of Tomida (1981).

Recent collecting efforts in the San Juan Basin have resulted in downward extension of the stratigraphic ranges of *Tetraclaenodon* and *Pantolambda* (see Williamson 1996). *Tetraclaenodon* is now known from low in the section at Kutz Canyon from a horizon correlated to the Dragon Canyon Local Fauna, the type fauna for the To1 interval zone (Williamson 1996). A specimen of *Pantolambda* has been found low in the Kutz Canyon section approximately equivalent to a horizon that previously recorded the lowest stratigraphic occurrence of *Tetraclaenodon* (see Williamson 1996). Therefore, if *Tetraclaenodon* and *Pantolambda* were retained as defining taxa

for the To2 and To3 interval zones in the San Juan Basin, the To3 interval zone would include all localities formerly in To2, and the To2 interval zone would include a significant part of To1.

Rather than expand the concepts of the *Tetraclaenodon*/*Pantolambda* Interval Zone (To2) and the *Pantolambda*/*P. praecursor* Interval Zone (To3), we redefine the interval zones for the Torrejonian mammal age. The *Periptychus carinidens*/*Tetraclaenodon* Interval Zone is renamed the *P. carinidens*/*Protoselene opisthacus* Interval Zone (To1), the *Tetraclaenodon*/*Pantolambda* Interval Zone is redefined as the *Protoselene opisthacus*/*Mixodectes pungens* Interval Zone (To2), and the *Pantolambda*/*P. praecursor* Interval Zone is redefined as the *M. pungens*/*P. praecursor* Interval Zone (To3). In relation to the local zonation proposed for the San Juan Basin by Williamson (1996, figures 18 and 19), the To1, To2, and To3 interval zones correlate as follows: To1 is equal to Williamson's P–P zone (= Dragonian mammal age); To2 is equal to his combined P–E, E–A, A–P, and P–M zones (approximately equivalent to *Deltatherium* zone); and To3 is equal to the M zone (= *Pantolambda* zone).

It is important to point out that *Protoselene opisthacus* and *M. pungens*, although common in the San Juan Basin at their defining horizons, are not ideal species to use for defining the To2 and To3 interval zones because their known geographic ranges are limited. However, we could not identify a more appropriate choice because taxa that are simultaneously common and geographically widespread are not available. Therefore the To2 and To3 interval zones may be difficult to distinguish outside the San Juan Basin based only on faunal correlations. Magnetostratigraphy appears to be helpful in correlating to the To2 and To3 interval zones because the To2 interval zone is almost entirely within magnetic polarity chron C27r, and the To3 interval zone is almost entirely within magnetic polarity chron C27n (Williamson 1996).

In the San Juan Basin, the To1 interval zone is present in De-na-zin, Betonnie–Tsosie, and Kimbeto washes, Kutz Canyon (as “main body” of Nacimiento Formation) and perhaps Mesa de Cuba, from the Arroyo Chijullita Member of the Nacimiento Formation (Williamson 1996). Similarly, the To2 interval zone is present in Kutz and Gallegos canyons, the Betonnie–Tsosie, Kimbeto, Escavada, and Torrejon washes, and Mesa de Cuba–Mesa Portales from the Ojo Encino Member of the Nacimiento Formation (except for Kutz Canyon and Mesa de Cuba). The To3 interval zone is present in Torrejon and Escavada washes and Mesa Chijuilla, all from the Ojo Encino Member of the Nacimiento Formation (Williamson 1996).

The appearance of *Plesiadapis praecursor* establishes the upper limit of the To3 interval zone and the beginning of the Tiffanian mammal age. Gingerich et al. (1983) noted the appearance of *Microcosmodon*, *Carpodaptes*, *Nannodectes*, *Ectocion*, and *Phenacodus* as well as *Plesiadapis* at the beginning of the Tiffanian. *Microcosmodon* has since been reported from the Puercan mammal age (Johnston and Fox 1984; Weil 1998), and *Phenacodus* is present in the Hanna Basin in strata interpreted to represent late phases of the Torrejonian mammal age (Higgins 2000). It is important to note that neither *Plesiadapis* nor a mammalian assemblage of early Tiffanian age is known from the Nacimiento Formation in the San Juan Basin. Consequently, the top of the To3 interval zone cannot be identified in the San Juan Basin.

***Periptychus carinidens*/*Protoselene opisthacus* Interval Zone (To1)** We define this interval zone to include faunas that occurred between the first appearance of *P. carinidens* and the first appearance of *P. opisthacus*. The Dragon Local Fauna of Utah has the best faunal representation of the To1 interval zone. It was described by Gazin (1938, 1939, 1941), with notable additions and revisions by Wilson (1956), MacIntyre (1966), Szalay (1969), West (1976), Tomida and Butler (1980), Robison (1986), Cifelli et al. (1995), and Williamson (1996). A revised faunal list and discussion were presented by Williamson (1996).

In addition to the Dragon Local Fauna, the To1 interval zone is documented in strata of the Nacimiento Formation of the San Juan Basin (Tomida 1981; Williamson 1996; Lucas et al. 1997), the Tullock Formation of eastern Montana (Archibald 1982; Clemens, pers. obs., 2000), and the Lebo Formation of south-central Montana (Hartman and Krause 1993, table 2).

A small and undescribed mammalian assemblage from the Farrand Channel (Mosquito Gulch Local Fauna) in the Tullock Formation of Montana contains *Paromomys* (Clemens, pers. obs., 2000), a distinctive Torrejonian to early Tiffanian taxon. Separated by approximately 25 m of strata, the Farrand Channel overlies the Garbani Channel, which yielded the Garbani Local Fauna, the mammalian assemblage we tentatively correlate to the Pu3 interval zone. Strata that contain the Farrand Channel are of reversed polarity, correlated with magnetic anomaly chron C28r (Swisher et al. 1993). The Farrand Channel is also bracketed by lignitic beds that bear volcanic crystal-rich partings suitable for dating. Two dated units (64.11 Ma below the channel, 63.90 Ma above) constrain the age of the channel filling (Swisher et al. 1993). These dates are consistent with correlation to magnetic anomaly chron C28r. Thus the Farrand Channel may be the

oldest known Torrejonian fauna in North America; To1 interval zone faunas in the Nacimiento and North Horn formations occur in strata correlated to magnetic anomaly chron C28n or the base of C27r (Tomida and Butler 1980; Lindsay et al. 1981; Butler and Lindsay 1985; Williamson and Lucas 1992; Williamson 1996).

From the lower part of the Lebo Formation in the Crazy Mountains Basin of south-central Montana are Simpson's (1937b) localities 9, 65, and 78. These sites have not been collected extensively, but they could be temporal equivalents of the Dragon Local Fauna (Sloan 1987).

From the Lebo Member of the Fort Union Formation in southeastern Montana is a sparse mammalian assemblage called the School Well Local Fauna that contains *Ptilodus*, *Litaletes*, *Periptychus*, and *Paromomys* (Hunter et al. 1997). The School Well Local Fauna probably is Torrejonian in age, but not enough information is available to firmly place it in a particular interval zone.

The To1 interval zone is not very fossiliferous in the San Juan Basin (Williamson 1996). As noted earlier, mammals assigned to the To1 interval zone have been found at sites superposed over localities yielding Puercan faunas in the De-na-zin, Kimbeto, and Betonnie-Tsosie washes (Lindsay et al. 1981; Tomida 1981; Williamson 1996). However, those Puercan and Torrejonian assemblages are separated by a wide stratigraphic interval. In Kutz Canyon and Mesa de Cuba, faunas of the To1 interval zone also are present, but they are not underlain by known Puercan sites. However, they are overlain by faunas referable to the To2 interval zone (Williamson 1996). Where sampling has been done, sites representing the To1 interval zone in the San Juan Basin occur in strata with normal polarity (Tomida 1981), interpreted as magnetic polarity chron C28n (Williamson and Lucas 1992; Williamson 1996). Combined with paleomagnetic correlations described previously from Montana and Utah, the To1 interval zone is interpreted to correlate with part of magnetic polarity chron C28r, the entirety of magnetic polarity chron C28n, and the lowermost part of magnetic polarity chron C27r (Tomida and Butler 1980; Butler and Lindsay 1985; Williamson and Lucas 1992; Swisher et al. 1993; Williamson 1996).

First appearances: *Acmeodon*, *Aphronorus*, *Conoryctella*, *Mioclaenus*, *Palaechthon*, *Paromomys*, *Plesiolestes*, *Proctictis*, *Tetraclaenodon*, and *Triisodon*

Last appearances: *Oxyclaenus* and *Viridomys*?

Index fossils: *Draconodus*, *Dracontolestes*, and *Oxytomodon*

Characteristic fossils: *Anisonchus*, *Catopsalis*, *Chriacus*, *Desmatoclaenus*, *Eucosmodon*, *Goniacodon*, *Haplo-*

*conus*, *Litaletes*, *Litomylus*, *Loxolophus*, *Mimotricentes*, *Parectypodus*, *Periptychus*, *Promioclauenus*, *Protoselene*, and *Ptilodus*

Taxa absent but known before and after To1: *Conacodon?*, *Ectypodus*, *Ellipsodon*, *Haplaletes*, *Ictidopappus?*, *Leptacodon?*, *Mesodma*, *Microcosmodon*, *Neoplagiaulax*, *Palaeictops*, *Peradectes*, *Prodiacodon*, *Styginmys*, and *Xyronomys*

***Protoselene opisthacus*/*Mixodectes pungens* Interval Zone (To2)** We define this interval zone to include faunas that occurred between the first appearance of *P. opisthacus* and the first appearance of *M. pungens*. In the San Juan Basin, the To2 interval zone is better represented in abundance of fossils and localities than the To1 interval zone. The To2 interval zone as defined here includes, in ascending stratigraphic order, the P–E, E–A, A–P, and P–M biostratigraphic zones of Williamson (1996). Inclusion of the uppermost of these local San Juan Basin zones (i.e., the P–M zone) in the To2 interval zone is a departure from Archibald et al. (1987). They included this local zone in the To3 interval zone. As a result of this change, sites from the head of Kimbeto Wash (including AMNH locality 8 and Kansas University [KU] locality 9, “Little Pocket”) and the south end of Kutz Canyon (including AMNH 1482 and 2658 and University of Arizona Laboratory of Paleontology [UALP] 7650) are now assigned to the To2 interval zone. The To3 interval zone thus becomes limited to the stratigraphic interval equivalent to the *Pantolambda* zone of Osborn (1929), including the richly fossiliferous Tsentas Microsite (New Mexico Museum of Natural History [NMMNH] locality L-312). Also, some sites previously assigned to the To3 interval zone are now correlated with the redefined To2 interval zone (discussed later in this chapter).

The To2 interval zone is well represented in Kutz Canyon, San Juan Basin, New Mexico (Williamson 1996). In Kutz Canyon this interval zone is superposed on the To1 interval zone. The To2 interval zone is overlain by the To3 interval zone in Torrejon and Escavada washes (Williamson 1996). Based on paleomagnetic correlations (Williamson 1996, figure 9), the To2 interval zone correlates closely with nearly the entire span of magnetic polarity chron C27r. This is important to note because the redefined To3 interval zone in the San Juan Basin correlates with nearly the entire span of magnetic polarity chron C27n.

Other diverse and well-known local faunas in the To2 interval zone are those from Gidley and Silberling quarries (Simpson 1937b; Rose 1981a) in the Crazy Mountains Basin, Montana; Rock Bench Quarry (Jepsen 1930, 1940;

Rose 1981a) in the Bighorn Basin, Wyoming; and Swain Quarry (Rigby 1980) in the Washakie Basin, Wyoming. Williamson (1996) tentatively correlated the mammalian assemblages from the Swain, Gidley, and Rock Bench quarries to his P–M zone in the San Juan Basin. As indicated previously, we recognize Williamson’s (1996) P–M zone as the highest stratigraphic interval in the San Juan Basin that is within the redefined To2 interval zone. Thus we tentatively assign the Gidley, Silberling, Rock Bench, and Swain quarries to the To2 interval zone (faunas from Gidley and Silberling quarries are similar; see Simpson 1937b and Rose 1981a). Rock Bench and Silberling quarries both occur in strata of reversed polarity correlated to magnetic polarity chron C27r (Butler et al. 1987). This is consistent with paleomagnetic results from the San Juan Basin in that faunas assigned to the To2 interval zone there occur in rocks of reversed polarity, also correlated to magnetic anomaly chron C27r (Williamson and Lucas 1992; Williamson 1996).

Rigby (1980) indicated the presence, based on isolated teeth, of the hyaenodontid *Prolimnocyon* in the mammalian sample from the Swain Quarry. This occurrence would greatly extend the first record of North American hyaenodontids from the Wasatchian into the Torrejonian mammal age. Pending further documentation, we omit this questionable record from our To2 fauna list.

Hunter (1999) reported the presence of two identifiable taxa (the taeniodont *Conoryctella* and the mesonychid *Dissacus*) from the Brown Ranch localities in the upper part of the Ludlow Formation of North Dakota. The co-occurrence of these taxa indicates that the Brown Ranch localities probably are referable to the To2 interval zone.

The Black Peaks Member of the Tornillo Formation of Texas yielded a small mammalian fauna at locality Texas Memorial Museum (TMM) 40147 (The Middle Peak–Alligator Alley) that shows strong Torrejonian affinities (Standhardt 1986; Williamson 1996). Paleomagnetic analysis of strata containing TMM 40147 indicates that the locality occurs in rocks that probably are of reversed polarity (Rapp et al. 1983; Standhardt 1986). If so, this might indicate a correlation with magnetic polarity chron C27r. Therefore we tentatively correlate TMM 40147 to the To2 interval zone.

A summary of magnetostratigraphic data for the To2 interval zone based mainly on analysis of the Nacimiento Formation in the San Juan Basin (see Williamson 1996 for summary), with additions by Butler et al. (1987) from Wyoming and Montana, indicates that the To2 interval zone occurs in reversely magnetized strata correlated with magnetic polarity chron C27r.

First appearances: *Adunator*, *Anconodon*, *Ankalagon*, *Avunculus*, *Claenodon*, *Colpoclaenus?*, *Conoryctes*, *Corphagus*, *Deuteronodon*, *Diacodon?*, *Dissacus*, *Elphidotarsius*, *Eudaemonema*, *Gelastops*, *Ignacius*, *Jepsenella*, *Microclaenodon*, *Mixodectes*, *Myrmecoboides*, *Palaeoryctes*, *Palenochtha*, *Paleotomus*, *Pantolambda*, *Pentacodon*, *Picrodus*, *Pronothodectes*, *Propalaeosinopa*, *Prothryptacodon*, *Psittacotherium*, *Simpsonictis*, *Stilpnodon*, and *Unuchinia*

Last appearances: *Conoryctella*, *Ellipsodon*, *Ictidopappus*, *Stygimys*, *Triisodon*, and *Xyronomys*

Index fossils: *Deltatherium*, *Huerfanodon*, *Leptonysson*, and *Xanoclomys*

Characteristic fossils: *Acmeodon*, *Anisonchus*, *Aphronorus*, *Catopsalis*, *Chriacus*, *Ectypodus*, *Eucosmodon*, *Goniacodon*, *Haplaletes*, *Haploconus*, *Leptacodon?*, *Litaletes*, *Litomylus*, *Loxolophus*, *Mesodma?*, *Mimotricentes*, *Mioclaenus*, *Neoplagiaulax*, *Palaechthon*, *Palaeictops*, *Parectypodus*, *Paromomys*, *Peradectes*, *Periptychus*, *Plesiolestes*, *Prodiacodon*, *Promioclaenus*, *Protictis*, *Protoselene*, *Ptilodus*, and *Tetraclaenodon*

Taxa absent but known before and after To<sub>2</sub>: *Conacodon?*, *Desmatoclaenus* and *Microcosmodon*

***Mixodectes pungens*/*Plesiadapis praecursor* Interval Zone (To<sub>3</sub>)** We define this interval zone to include faunas that occurred between the first appearance of *M. pungens* and the first appearance of *P. praecursor*.

The To<sub>3</sub> interval zone is found only in the eastern part of the San Juan Basin from the Ojo Encino Member of the Nacimiento Formation in Escavada and Torrejon washes and at Mesa Chijuilla, but these areas are richly fossiliferous. The best representation of To<sub>3</sub> faunas actually is from the type Torrejonian at the head of Torrejon Wash. This area was collected in the 1890s by J. Wortman and W. Granger for the American Museum of Natural History (= AMNH locality 10). Many other institutions have collected from these strata over the last hundred years. Tsentas (1981) reported a productive screenwashing site from the To<sub>3</sub> interval zone in the type area that subsequently yielded small mammals not previously recorded from this interval zone (Williamson 1996, table 2).

Two faunas from Alberta that might belong in the To<sub>3</sub> interval zone are the Calgary 2E Local Fauna (L. Russell 1958; D. Russell 1967; Krause 1978; Fox 1990c) from the Porcupine Hills Formation and the Diss Local Fauna from the Coalspur Formation (Fox 1990c). The small mammal fauna from the Calgary 2E Local Fauna includes *Tetraclaenodon*, *Claenodon*, *Protictis*, *Pronothodectes*, and the multituberculates *Baiotomeus*, *Neoplagiaulax*,

*Anconodon*, *Eucosmodon*, and *Catopsalis* (see Fox 1990c). The Diss Local Fauna is also small and includes *Pararyctes*, *Propalaeosinopa*, *Aphronorus*, *Colpoclaenus*, *Promioclaenus*, *Simpsonictis*, *Palaechthon*, and the multituberculates *Ptilodus*, *Baiotomeus*, *Mimetodon*, *Parectypodus*, and *Neoplagiaulax* (see Fox 1990c). The presence of *Baiotomeus* in both local faunas and *Pararyctes* in the Diss Local Fauna suggests an early Tiffanian age. However, most taxa in these two local faunas have strong Torrejonian affinities. We tentatively suggest an assignment of the Diss and Calgary 2E local faunas to the To<sub>3</sub> interval zone.

A series of localities near Cub Creek in the Clark's Fork Basin of south-central Montana shows superposition of early Tiffanian (Ti<sub>1</sub>) over late Torrejonian (To<sub>3</sub>) sites in a measured section that has undergone magnetostratigraphic analysis (Butler et al. 1987). Here, a site (Cub Creek 2) referable to the To<sub>3</sub> interval zone occurs in strata of normal polarity and is overlain by strata containing sites (Cub Creek 1, Cub Creek 3, Eagle Quarry) in rocks of reversed polarity that yield early Tiffanian mammals. Thus Cub Creek 2 is correlated with magnetic polarity chron C27n and the other three sites with magnetic polarity chron C26r (Butler et al. 1987). Until recently, this was the only section in North America known to have early Tiffanian faunas in superposition above a late Torrejonian site. However, biostratigraphic analysis of sections spanning the Torrejonian–Tiffanian boundary in the Bighorn and Hanna basins of Wyoming by Hartman (1986) and Higgins (2000), respectively, allow further refinement of this important biochronologic boundary (see “Torrejonian–Tiffanian Boundary” later in this chapter for further discussion).

Other sites containing faunas that might be referable to the To<sub>3</sub> interval zone are the Medicine Rocks 1–Mehling Site in southeastern Montana, Donnybrook and the Lloyd and Hares sites in North Dakota, and a series of localities in the Washakie Basin, Wyoming (Winterfeld 1982).

In general, the To<sub>3</sub> interval zone is dominated by *Ptilodus*, *Mimotricentes*, and *Promioclaenus*. Primates, especially *Palaechthon* and *Paromomys*, may be dominant in northern faunas, whereas *Tetraclaenodon*, *Periptychus*, and *Mixodectes* may be dominant in southern faunas. Plesiadapid and carpolestid primates show strong affinity for northern latitudes. In contrast, the mixodectids are more common in southern latitudes in the time represented by the To<sub>3</sub> interval zone (Archibald et al. 1987).

It has not been possible to identify a biochronologic basis or magnetostratigraphic limit for the Torrejon-

ian–Tiffanian boundary in the San Juan Basin; definitive Tiffanian taxa such as *Plesiadapis* have not been recovered from the Nacimiento Formation. Therefore the top of the To3 interval zone in the San Juan Basin remains biochronologically unconstrained.

A summary of magnetostratigraphic data for the To3 interval zone, based largely on work in the San Juan Basin (see Williamson 1996), shows that the To3 interval zone occurs in normally magnetized strata correlated with magnetic polarity chron C27n.

First appearances: *Baiotomeus*, *Elpidophorus*, *Krauseia*, *Litocherus*, *Mimetodon*, *Neoclaenodon*, *Pararyctes*, and *Phenacodus*

Last appearances: *Ankalagon*, *Avunculus*, *Conoryctes*, *Deuteronogodon*, *Goniacodon*, *Haploconus*, *Loxolophus*, *Microclaenodon*, *Mioclaenus*, and *Mixodectes*

Index fossils: *Anasazia*, *Escavadodon*, *Premnoides*, *Stelocyon*, and *Swaindelphys*

Characteristic fossils: *Acmeodon*, *Adunator*, *Anconodon*, *Anisonchus*, *Aphronorus*, *Catopsalis*, *Chriacus*, *Claenodon*, *Colpoclaenus*, *Coriphagus*, *Diacodon?*, *Dissacus*, *Ectypodus*, *Elphidotarsius*, *Eucosmodon*, *Eudaemonema*, *Gelastops*, *Haplaletes*, *Ignacius*, *Jepsenella*, *Leptacodon*, *Litaletes*, *Litomylus*, *Mesodma*, *Mimotricentes*, *Myrmecoboides*, *Neoplagiaulax*, *Palaechthon*, *Palaeictops?*, *Palaeoryctes*, *Palenochtha*, *Paleotomus*, *Pantolambda*, *Paractypodus*, *Paromomys*, *Pentacodon*, *Peradectes*, *Periptychus*, *Picrodus*, *Plesiolestes*, *Prodiacodon*, *Promioclaenus*, *Pronothodectes*, *Propalaeosinopa*, *Prothryptacodon*, *Protictis*, *Protoselene*, *Psittacotherium*, *Ptilodus*, and *Tetraclaenodon*

Taxa absent but known before and after To3: *Conacodon?*, *Desmatoclaenus*, *Microcosmodon*, *Simpsonictis*, *Stilpnodon?*, and *Unuchinia*

## TIFFANIAN MAMMAL AGE

*Tiffany* refers to a small settlement in southern Colorado; Walter Granger (1917) first used the term to refer to strata and their contained faunas in the northern San Juan Basin. The “Tiffany beds” are now assigned to the Animas Formation, which intertongues with the Nacimiento and San Jose formations to the south. The Tiffany beds probably are equivalent to part of the Cuba Mesa Sandstone member, the unfossiliferous basal unit of the San Jose Formation. However, definite stratigraphic placement of these beds laterally between the underlying Nacimiento Formation and overlying San

Jose Formation has not been demonstrated (Archibald et al. 1987).

The first collection of fossil mammals reported from the Tiffany beds was made by Wegemann (1917). Later that same year Granger (1917) presented a more detailed treatment of the Tiffany beds and their mammalian fauna and named the principal locality the Mason Quarry or Mason Pocket. Granger (1917) also suggested that the Tiffany mammalian assemblage was intermediate in evolutionary grade between “Torrejon” and “Wasatch” faunas and that it might be correlative with the Clark’s Fork fauna of Wyoming, which he tentatively interpreted as early Eocene in age. The Tiffany Local Fauna from Mason Pocket and specimens from sites nearby were described by Simpson (1935c, 1935d, 1935e), and few additions or modifications have appeared since then.

In the 1920s and 1930s, additional mammalian assemblages were discovered that resembled the Tiffany fauna. These included Erickson’s Landing and Red Deer from the Paskapoo Formation and Cochrane II from the Porcupine Hills Formation of Alberta (Simpson 1927; Russell 1929), a series of localities from the Fort Union Formation (= Polecat Bench Formation) in Wyoming (Jepsen 1930, 1940), and another series of localities from the Crazy Mountain field in Montana (Simpson 1936, 1937a, 1937b). These sites demonstrated that Tiffany faunas were late Paleocene in age and were younger than Torrejon faunas and older than Clark’s Fork faunas (Simpson 1933).

Wood et al. (1941) named the Tiffanian land mammal age based on the fauna from the Tiffany beds of Colorado. Its two principal correlatives were Bear Creek and Silver Coulee. These refer to Bear Creek Local Fauna, Fort Union Formation, Montana (Simpson 1928, 1929a, 1929c; Jepsen 1937), and Silver Coulee Local Fauna, Fort Union Formation, Wyoming (Jepsen 1930, 1940). Although the faunas discovered in the Silver Coulee beds confirm the Tiffanian mammal age assigned to them by Wood et al. (1941), the Bear Creek Local Fauna has since been determined to be of Clarkforkian age (Rose 1981a).

Archibald et al. (1987) provided an updated characterization and zonation of the Tiffanian mammal age and discussed the many Tiffanian faunas discovered and described since the publication by Wood et al. (1941). These faunas were from the Paskapoo and Ravenscrag formations, Alberta and Saskatchewan (Russell 1967; Krishtalka 1973; Krause 1977, 1978); Fort Union Formation, central Montana (Gingerich et al. 1983); Tongue River Formation, eastern Montana (Wolberg 1979); Tongue River and Sentinel Butte formations, North Dakota (Holtzman 1978); Fort Union Formation, Clark’s Fork and Bighorn



basins, northern Wyoming (Gingerich 1976; Rose 1981a); Hoback Formation, Hoback Basin, Wyoming (Dorr 1952, 1958, 1978); the "sandstone and shale sequence," Togwotee Pass area, Wyoming (McKenna 1980); Shotgun Member, Fort Union Formation, Wind River Basin, Wyoming (Patterson and McGrew 1962; Gazin 1971; Krishtalka et al. 1975); Fort Union Group, Bison Basin, Wyoming (Gazin 1956a); Chappo Member, Wasatch Formation, Wyoming (Dorr and Gingerich 1980); Evanston Formation, Fossil Basin, Wyoming (Gazin 1956b, 1969); and Black Peaks Formation, Big Bend area, Texas (Schiebout 1974).

Since the publication by Archibald et al. (1987), additional Tiffanian faunas have been reported or previous ones have been described more completely. This applies to faunas from the Hanna Formation, Carbon and Hanna basins, south-central Wyoming (Secord 1998; Higgins 2000); Fort Union Formation, southern Bighorn Basin, central Wyoming (Hartman 1986; Leite 1992); Wasatch Formation, Wyoming (Gunnell 1994), Fort Union Formation, Crazy Mountains Basin, Montana (Hartman and Krause 1993); Fort Union Formation, Powder River Basin, Montana (Robinson and Honey 1987); Sentinel Butte and Tongue River formations, Williston Basin, North Dakota (Erickson 1991, 1999; Hartman and Kihm 1991, 1995, 1999; Hunter 1999); Paskapoo Formation, Alberta (Fox 1990c); Goler Formation, California (Lofgren et al. 1999; McKenna and Lofgren in press); and Williamsburg Formation, South Carolina (Schoch 1998).

#### TORREJONIAN-TIFFANIAN BOUNDARY

When Archibald et al. (1987) was published, the only continuously exposed sections of strata that documented superposition of Tiffanian over Torrejonian faunal assemblages were in the Clark's Fork Basin (at Cub Creek) in south-central Montana and the northern Bighorn Basin (at Polecat Bench) in northern Wyoming. As mentioned previously, strata that yield Eagle Quarry, Cub Creek 1, and Cub Creek 3 (all T<sub>11</sub>) overlie strata that yield the Cub Creek 2 locality (T<sub>03</sub>), and this sequence of localities begins in strata of normal polarity and continues up into those of reversed polarity correlated with magnetic polarity chron C<sub>27n</sub> and C<sub>26r</sub>, respectively (Butler et al. 1987).

This situation has changed: Since 1987 a series of mammalian assemblages from the southern Bighorn and Hanna basins in Wyoming have been reported that span the Torrejonian–Tiffanian boundary (Hartman 1986; Higgins 2000). Also, sites in the Carbon Basin adjacent to the Hanna Basin contain mammalian faunas that are

either latest Torrejonian or earliest Tiffanian in age (Secord 1998).

In the southern Bighorn Basin near Cedar Mountain, 185–300 m above the base of the Fort Union Formation is a series of localities that yield late Torrejonian mammals. In direct superposition are strata 300–338 m above the base of the formation that yield early Tiffanian mammals, including *Plesiadapis praecursor* and *Nannodectes intermedius*. Hartman (1986:51) noted that although *Plesiadapis* is the FAD of the Tiffanian mammal age, early Tiffanian faunas at Cedar Mountain that contain *Plesiadapis* have a "strong Torrejonian element, with the transition to the Tiffanian marked only by appearance of the two plesiadapids indicative of the earliest Tiffanian."

A similar situation exists in the Carbon Basin, where the Grayson Ridge and Halfway Hill faunas yield mammalian assemblages with affinities to both the Torrejonian and Tiffanian mammal ages (Secord 1998). Because of the small sample sizes from Grayson Ridge and Halfway Hill, Secord (1998:135) argued that although "these faunas are close in age to the Torrejonian–Tiffanian boundary," a confident assignment to the latest Torrejonian or earliest Tiffanian mammal age is not warranted based on the available data. However, the presence of *Ectocion* in the Grayson Ridge Fauna and *Thryptacodon* in the Halfway Hill Fauna, taxa both limited to the Tiffanian elsewhere, would suggest an early Tiffanian (T<sub>11</sub>) age.

In the Hanna Basin, an excellent faunal record of the transition from Torrejonian to Tiffanian faunas is available. Unique structural and depositional conditions in south-central Wyoming during Paleocene time resulted in deposition of the Hanna Formation, a rock unit more than 2 miles thick. Age of the formation ranges from Torrejonian at its base to early Wasatchian at its erosional top (Lillegraven and Snoke 1996). Lower reaches of the Hanna Formation in the Hanna Basin's northeastern corner exhibit a richly fossiliferous, 550-m-thick interval yielding latest Torrejonian (T<sub>03</sub>) through middle Tiffanian (T<sub>13</sub>) mammalian assemblages. The fossils represent The Breaks Local Fauna, described from 136 recorded localities by Higgins (2000). Minimally, the local fauna is composed of 72 mammalian species, identified at least to generic levels using data from 57 sites. A 55-m-thick interval, placed centrally in stratigraphic limits of the local fauna, has yielded an assemblage of species that ordinarily would be considered characteristic of latest Torrejonian (T<sub>03</sub>) or earliest Tiffanian (T<sub>11</sub>) age. Because of the grand thickness and richly fossiliferous nature of this section, it shows, for the first time, a clear picture of the nature of faunal change at the Torrejonian–Tiffanian boundary.

The zone of overlap in The Breaks Local Fauna documents first appearances of two species of plesiadapids (*Nannodectes intermedius* and *Plesiadapis praecursor*), traditional index taxa for recognizing advent of the Tiffanian mammal age. Additionally, the zone of overlap documents persistence, presumably as evolutionary relicts, of 14 species considered typical of Torrejonian time (*Krauseia clemensi*, *Ectypodus szalayi*, *Gelastops joni*, *Paleotomus milleri*, *Aphronorou ratatoski*, *Litaletes disjunctus*, *Anisonchus sectorius*, *Tetraclaenodon puercensis*, *Prodiacodon puercensis*, *Mimotricentes subtrigonus*, *Palaechthon alticuspis*, *Palaechthon nacimienti*, *Palenochtha minor*, and *Paramomys depressidens*). Indeed, were it not for presence of the plesiadapids, age of the zone of overlap probably would be identified as Torrejonian. This raises concern about accuracy of age control in stratigraphic sections elsewhere, supposedly representing latest Torrejonian time, in which plesiadapids may have been rare or absent. With this caveat in mind, we continue to use the first appearance of *Plesiadapis* in defining the advent of the Tiffanian mammal age. We believe that *Plesiadapis* is an appropriate taxon for this purpose because *Plesiadapis* has traditionally served as a taxon marking the beginning of the Tiffanian mammal age (Wood et al. 1941; Archibald et al. 1987); *Plesiadapis* is common and widespread, at least in northern faunas; and it has not been confidently documented that *Plesiadapis* occurs in strata of Torrejonian age (although this is somewhat circular because the presence of *Plesiadapis* in a transitional fauna of uncertain age, by definition, indicates a Tiffanian age for the fauna). Discovery of *Plesiadapis* in an unquestionably Torrejonian fauna (such as one in the Torrejonian interval of the Nacimiento Formation from the San Juan Basin) or in strata of normal polarity correlated to magnetic polarity chron C27n would be powerful evidence of Torrejonian occurrence of this genus.

As interpreted from Paleocene mammalian faunas in the Hanna Basin, important paleogeographic modifications occurred in western North America between Puercan and Tiffanian time. For example, Eberle and Lillegraven (1998b) reported that Puercan assemblages of the Hanna Basin had much greater taxonomic affinities with contemporaneous mammalian faunas to the south than to the north. Just the opposite became the case during late Torrejonian and early Tiffanian time (Higgins 2000). Such major alterations of distributional ranges may well reflect continental climatic change in the earlier half of the Paleocene.

Archibald et al. (1987) noted that all genera first appearing at the beginning of the Tiffanian mammal age

were plausibly derived from genera present in North America during the Torrejonian mammal age. Therefore, the boundary between the Torrejonian and the Tiffanian mammal ages probably was a product of intracontinental evolution and local dispersal. This is in contrast to the situation between the Tiffanian and the Clarkforkian mammal ages, which was marked by immigrations of new genera from Asia (Archibald et al. 1987). The immigrants represented new families, and even orders, on the North American continent.

## DEFINITION AND CHARACTERIZATION

We define the Tiffanian mammal age to include faunas that occurred between the first appearance of the plesiadapid primate *Plesiadapis* and the first appearance of Rodentia.

Wood et al. (1941) listed the following taxa as making their first appearance in the Tiffanian mammal age: *Palaeosinopa*, *Phenacodus*, *Plesiadapis*, "*Probathyopsis*," Rodentia, and *Thryptacodon*. Based on present knowledge, *Palaeosinopa*, *Plesiadapis*, "*Probathyopsis*" (included in *Prodinoceras* by McKenna and Bell 1997), and *Thryptacodon* still occur first in the Tiffanian, whereas *Phenacodus* is now known from late Torrejonian rocks in the Hanna Basin of Wyoming (Higgins 2000); the Rodentia did not appear until the Clarkforkian mammal age. Four of the five genera noted as last appearances for the Tiffanian mammal age (*Tetraclaenodon*, *Anisonchus*, *Pantolambda*, and *Claenodon*) retain this distinction, whereas *Leptacodon* is questionably reported from the Clarkforkian mammal age. Wood et al. (1941) listed *Barylambda*, "*Bathyopsoides*," "*Labidolemur*," *Phenacodus grangeri*, "*Sparactolambda*," and *Titanoides* as index fossils for the Tiffanian mammal age. "*Sparactolambda*" is now considered a synonym of *Titanoides*, "*Bathyopsoides*" a synonym of *Prodinoceras*, and "*Labidolemur*" a synonym of *Apatemys* (see McKenna and Bell 1997). Of the other index fossils listed by Wood et al. (1941), only *Titanoides* and *Phenacodus grangeri* are limited to the Tiffanian; *Barylambda* is known from the Clarkforkian mammal age. *Ectypodus* was listed as the only characteristic fossil for the Tiffanian, and this designation remains valid.

Paleomagnetic sections have been developed in Clark's Fork and Bighorn basins, northern Wyoming (Butler et al. 1980, 1981) in conjunction with fossil localities ranging from the Ti2 lineage zone through Cf3 acme zone. Therefore these paleomagnetic sections make up one of the most comprehensive for Paleocene terrestrial strata in North America. Although not all of the fossiliferous

parts of these strata were amenable to magnetostratigraphic analysis, the available part extended from within magnetic polarity chron C26r through C24r and included the Paleocene–Eocene boundary. In these basins Tiffanian faunas extend from magnetic polarity chron C26r into C25n (Archibald et al. 1987).

Rapp et al. (1983) presented a paleomagnetic sequence for the Black Peaks Member of the Tornillo Formation. The lower part of the Black Peaks Member, which includes Ray's Bonebed (TMM 40536–37, a locality correlated to the Ti3 lineage zone), occurs in strata of reversed polarity. Joe's Bone Bed (TMM 41365–66), a locality correlated with the Ti5 lineage zone, occurred in the overlying, reversed magnetozone. Rapp et al. (1983) and Schiebout et al. (1987) correlated these reversed magnetozones containing Joe's Bonebed (Ti5) with magnetic polarity chron C25r and Ray's Bonebed (Ti3) with magnetic polarity chron C26r, both of which are consistent with the paleomagnetic correlation of Tiffanian lineage zones in the Clark's Fork Basin (Butler et al. 1981).

To summarize, we place the Torrejonian–Tiffanian boundary at or near the boundary of magnetic polarity chrons C27n and C26r. The To3 interval zone is correlated with magnetic polarity chron C27n, the younger part of magnetic polarity chron C27r, and the older part of C26r. The Ti1 and Ti2 lineage zones are correlated with most of magnetic polarity chron C26r. The Ti3 lineage zone is correlated with younger parts of magnetic polarity chron C26r and part of C26n. The Ti4 lineage zone is correlated with most of magnetic polarity chron C26n and the beginning of chron C25r. The Ti5 lineage zone is correlated with most of magnetic polarity chron C25r and the earliest part of chron C25n. The Ti6 interval subzone of the Ti6–Cf1 lineage zone is correlated with part of magnetic polarity chron C25n. These data indicate that the Tiffanian mammal age began during magnetic polarity chron C26r and ended in magnetic polarity chron C25n.

Paleomagnetic samples also were analyzed from strata that yielded the type Tiffanian mammal age in southern Colorado, including Mason Pocket (Butler et al. 1981). The strata sampled were of reversed polarity, correlated to magnetic polarity chron C25r based on faunal correlation to mammalian assemblages in the Clark's Fork and Bighorn basins (Butler et al. 1981).

Characterization of the Tiffanian mammal age is as follows.

First appearances: *Aletodon*, *Anacodon*, *Apatemys*, *Arcostylops*, *Barylambda*, *Carpolestes*, *Chiromyoides*, *Cyriacotherium*, *Didymictis*, *Dillerlemur*, *Dipsalodon*, *Ecto-*

*cion*, *Ectoganus*, *Entomolestes*, *Esthonyx?*, *Haplolambda*, *Lambertocyon*, *Limaconyssus*, *Micromomys*, *Neoliotomus*, *Oxyaena*, *Palaeosinopa*, *Phenacodaptes*, *Phenacolemur?*, *Plesiadapis*, *Princetonia*, *Prochetodon*, *Prodinoceras*, *Thryptacodon*, *Thylacaelurus?*, *Titanoides*, and *Viverravus*

Last appearances: *Acmeodon*, *Anconodon*, *Anisonchus*, *Aphronorus*, *Baiotomeus*, *Catopsalis*, *Claenodon*, *Colpoclaenus*, *Conacodon?*, *Coriphagus*, *Desmatoclaenus*, *Elphidotarsius*, *Elpidophorus*, *Eucosmodon?*, *Eudaeomonema*, *Gelastops*, *Haplaletes*, *Jepsenella*, *Krauseia*, *Litaletes*, *Litocherus*, *Litomylus*, *Mesodma*, *Mimetodon*, *Mimotricentes*, *Myrmecoboides*, *Neoclaenodon*, *Neoplagiaulax*, *Palaechthon*, *Palenochtha*, *Paleotomus*, *Pantolambda*, *Pararyctes*, *Paromomys*, *Pentacodon*, *Periptychus*, *Picrodus*, *Plesiolestes*, *Promioclaenus*, *Pronothodectes*, *Propalaeosinopa*, *Prothryptacodon*, *Protictis*, *Protoselene*, *Psittacotherium*, *Ptilodus*, *Simpsonictis*, *Stilpnodon?*, *Tetraclaenodon*, and *Unuchinia*

Index fossils: *Aptoryctes*, *Amelotabes*, *Bisonalveus*, *Caenolambda*, *Carpocristes*, *Carpodaptes*, *Carpomogodon*, *Cedrocherus*, *Copecion*, *Dorraletes*, *Fractinus*, *Ignatiolambda*, *Liotomus*, *Litolestes*, *Melaniella*, *Mentoclaenodon*, *Mingotherium*, *Nannodectes*, *Navajovius*, *Pentacosmodon*, *Pristinictis*, *Propalaeonodon*, *Raphictis*, *Saxonella*, *Tytthaena*, *Utemylus*, *Xenacodon*, and *Zanycteris*

Characteristic fossils: *Adunator*, *Chriacus*, *Diacodon*, *Disacus*, *Ectypodus*, *Ignacius*, *Leptacodon*, *Microcosmodon*, *Palaeictops*, *Palaeoryctes*, *Parectypodus*, *Peradectes*, *Phenacodus*, and *Prodiacodon*

Taxa absent but known before and after the Tiffanian: none

## ZONATION

The large number of mammalian faunas belonging to the Tiffanian mammal age and the fact that many of them are located in separate depositional basins has made it difficult to determine the relative ages of localities and their contained faunas. Archibald et al. (1987) formalized the methods used to place Tiffanian localities in successive biostratigraphic zones or faunally equivalent biochronologic zones. We follow Archibald et al. (1987) and subdivide the Tiffanian mammal age into five lineage zones (Ti1–Ti5) and one interval subzone (Ti6, part of a sixth lineage zone, Ti6–Cf1) based on apparently nonoverlapping species of the primate *Plesiadapis* that have been argued to represent a single evolving lineage (Gingerich 1976). *Plesiadapis* is one of the most abundant and most widely distributed genera of late Paleocene

mammals, at least in northerly realms of the Western Interior, and is a suitable taxon for lineage-based zonation. The five lineage zones and one interval subzone of the Tiffanian that we recognize are *Plesiadapis praecursor*/*P. anceps* Lineage Zone (Ti1), *P. anceps*/*P. rex* Lineage Zone (Ti2), *P. rex*/*P. churchilli* Lineage Zone (Ti3), *P. churchilli*/*P. simonsi* Lineage Zone (Ti4), *P. simonsi*/*P. gingerichi* Lineage Zone (Ti5), and *P. gingerichi*/Rodentia Interval Subzone (Ti6) of the *P. gingerichi*/*P. cookei* Lineage Zone (Ti6–Cfi). The first three lineage zones are represented by faunas from the Crazy Mountains Basin, Montana (Simpson 1937a; Gingerich 1976). The second through fifth lineage zones and the sixth interval subzone are represented by faunas from the Clark's Fork and Bighorn basins, Wyoming (Gingerich 1976, 2000; Gingerich et al. 1980).

#### ***Plesiadapis praecursor*/*P. anceps* Lineage Zone (Ti1)**

We define the *P. praecursor*/*P. anceps* Lineage Zone to include faunas that occurred between the first appearance of *P. praecursor* and the first appearance of *P. anceps*.

As noted by Archibald et al. (1987), earliest Tiffanian faunas were known from central Alberta to probably southern Texas, but the zone was one of the least well known for the Tiffanian mammal age; samples from many Ti1 localities were either small or only partially described. This statement is still valid concerning Cochrane I in central Alberta (Russell 1958; Fox 1990c), Little Muddy Creek in the Fossil Basin of southwestern Wyoming (Gazin 1969), Bangtail Locality in the western Crazy Mountains Basin of south-central Montana (Gingerich et al. 1983), and Schiebout-Reeves Quarry in the Big Bend area of Texas (Schiebout 1974; Rapp et al. 1983; Schiebout et al. 1987).

Two of the largest and potentially most important collections from the Ti1 lineage zone, those from Keefer Hill in the Wind River Basin of Wyoming and Cochrane II from central Alberta, remain largely undescribed. Only preliminary faunal lists have been published for the Cochrane II (Fox 1990c) and Keefer Hill local faunas (Keefer 1961; D. Russell 1967), and some taxa cited in these preliminary lists are new records for the Ti1 lineage zone. We include most in table 3.2, which lists temporal ranges of North American genera of Paleocene mammals. Pending descriptions of specimens, we exclude *Oxyprimus*? and *Acheronodon* reported from Cochrane II because these undocumented records would drastically alter the temporal ranges of these genera, now both known elsewhere from the Pu1 interval zone only.

In the eastern Crazy Mountains Basin are the Douglass, Glennie, and Bingo localities. The mammalian assemblage from the Douglass Quarry was described by Krause

and Gingerich (1983), but extensive collections have been made since then. These collections, as well as those made from the Glennie and recently discovered Bingo locality, remain undescribed (Hartman and Krause 1993). These three quarries are especially important because they are located low in the Melville Formation. Stratigraphically, they occur above the Gidley and Silberling quarries (To3 interval zone) in the upper Lebo Formation and stratigraphically below Scarritt Quarry (Ti2 lineage zone) in the Melville Formation (Simpson 1937b; Krause and Gingerich 1983; Hartman and Krause 1993). Although no faunal list was presented, Hartman and Krause (1993) suggested that the mammalian assemblage from the Bingo Quarry appears to be correlative to the Bangtail Locality (Ti1 lineage zone) in the western Crazy Mountains Basin, which was described by Gingerich et al. (1983).

Robinson and Honey (1987) described a diverse new vertebrate fauna, including 21 mammalian species, from Newell's Nook (U.S. Geological Survey D-2003), Tongue River Member of the Fort Union Formation in the northern Powder River Basin of Montana. No specimens referable to *Plesiadapis* were reported. However, the mammalian assemblage does contain *Anconodon*, *Acmeodon*, *Nannodectes*, *Ectocion*, and *Anisonchus*, all taxa for which known temporal ranges overlap in the Ti1 lineage zone.

As mentioned earlier in the discussion of the Torrejonian–Tiffanian boundary, new earliest Tiffanian (Ti1) localities from Wyoming have been reported from the southern Bighorn Basin (Hartman 1986), the Hanna Basin (Higgins 2000), and probably the Carbon Basin (Secord 1998). Also, Cub Creek 1, Cub Creek 3, and Eagle Quarry from the Clark's Fork Basin in southern Montana yield sparse faunas referred to the Ti1 lineage zone.

The Douglass Quarry, Cub Creek 1, Cub Creek 3, Eagle Quarry, and probably Schiebout–Reeves Quarry all occur in strata of reversed polarity correlated with magnetic polarity chron C26r (Rapp et al. 1983; Schiebout et al. 1987; Butler et al. 1987). Thus, based on present knowledge, the Ti1 lineage zone is constrained within the older part of magnetic polarity chron C26r.

First appearances: *Bisonalveus*, *Carpodaptes*, *Ectocion*, *Limaconyssus*, *Nannodectes*, *Navajovius*, *Plesiadapis*, *Thryptacodon*, *Thylacaelurus*?, and *Titanoides*

Last appearances: *Anisonchus*, *Coriphagus*, *Eucosmodon*?, *Jepsenella*, *Krauseia*, *Litaletes*, *Palenochtha*, *Pantolambda*, *Pentacodon*, *Plesiolestes*, *Pronothodectes*, *Prothryptacodon*, and *Tetraclaenodon*

Index fossils: *Fractinus*, *Liotomus*, *Nannodectes intermedius*, *Plesiadapis praecursor*, and *Pristinictis*

Characteristic fossils: *Acmeodon*, *Adunator*, *Anconodon*, *Aphronorus*, *Baiotomeus*, *Catopsalis*, *Chriacus*, *Claenodon*, *Colpoclaenus*, *Desmatoclaenus*, *Diacodon?*, *Dissacus*, *Ectypodus*, *Elphidotarsius*, *Elpidophorus*, *Eudaemonema*, *Gelastops*, *Haplaletes*, *Ignacius*, *Leptacodon*, *Litocherus*, *Litomylylus*, *Mesodma*, *Microcosmodon*, *Mimetodon*, *Mimotricentes*, *Myrmecoboides*, *Neoclaenodon*, *Neoplagiaulax*, *Palaechthon*, *Palaeoryctes*, *Paleotomus*, *Pararyctes*, *Parectypodus*, *Paromomys*, *Peradectes*, *Periptychus*, *Phenacodus*, *Picrodus*, *Prodiacodon?*, *Promioclauenus*, *Propalaeosinopa*, *Protictis*, *Protoselene*, *Psittacotherium*, *Ptilodus*, and *Simpsonictis*

Taxa absent but known before and after T1: *Conacodon?*, *Palaeictops*, *Stilpnodon?*, and *Unuchinia*

#### ***Plesiadapis anceps*/*Plesiadapis rex* Lineage Zone (T12)**

We define the *P. anceps*/*P. rex* Lineage Zone to include faunas that occurred between the first appearance of *P. anceps* and the first appearance of *P. rex*.

Localities referred to the T12 lineage zone are known only from Montana and Wyoming and possibly Canada and California. Like the T11 lineage zone, the T12 lineage zone is not well known. Faunal studies on Scarritt Quarry from the Crazy Mountains Basin of south-central Montana (Simpson 1936, 1937a) and Saddle Locality from the Bison Basin of south-central Wyoming (Gazin 1956b) are the only published descriptions available for taxa occurring in the T12 lineage zone, and both local faunas are small. An updated faunal list for Scarritt Quarry was presented by Rose (1981a), and the sample appears to be biased in favor of small mammals (Rose 1981a, 1981b).

Much smaller collections of mammals from the T12 lineage zone are known from University of Michigan (UM) locality 263 in the Polecat Bench section, Bighorn Basin, Wyoming; the Tongue River Formation of North Dakota (Hunter 1999); and several localities (White Site, 7-Up Butte, Highway Blowout) in the Medicine Rocks area of southeastern Montana.

Fox (1990c) provided lists of taxa from two sites, Aaron's Locality and Hand Hills West lower level, from the Paskapoo Formation of Alberta that may be referable to the T12 lineage zone. Aaron's Locality contains *Bisonalveus*, *Ignacius*, *Elphidotarsius*, and *Ptilodus*, and the fauna at Hand Hills West lower level includes *Neoplagiaulax*, *Elphidotarsius*, *Picrodus*, and *Pararyctes*. Not enough faunal data are available to determine whether these sites are T11 or T12 in age (Fox 1990c).

The Goler Formation of California contains two mammalian assemblages, the Laudate Local Fauna and that from the Edentulous Jaw Site. Both are apparently Tiffanian

in age because both sites yield *Plesiadapis* (see Lofgren et al. 1999; McKenna and Lofgren in press). The Laudate Local Fauna had yielded *Microcosmodon?*, *Neoliotomus?*, *Neoplagiaulax*, *Mesodma?*, *Ptilodus*, *Paromomys*, *Conacodon*, and *Dissacus* and was tentatively assigned a late Torrejonian or early Tiffanian age (McKenna 1955, 1960; McKenna et al. 1987; Lofgren et al. 1999). Recently, a specimen of a smaller species of *Plesiadapis* was recovered, which indicates that the site probably is early Tiffanian in age (McKenna and Lofgren in press). The small fauna from the Edentulous Jaw Site may be a correlative of the T12 lineage zone. A small to medium-sized species of *Plesiadapis* occurs at the Edentulous Jaw Site (Lofgren et al. 1999) that shows many similarities in size and morphology (Lofgren unpubl. data) to *Plesiadapis anceps*. Thus correlation to the T12 lineage zone is plausible, pending the results of further analysis of screen-washed samples from this site.

The Hanna Basin of Wyoming is the only other area that has yielded sites referable to the T12 lineage zone. These sites are part of a series of localities that make up The Breaks Local Fauna of Higgins (2000), which is late Torrejonian (T03) through mid-Tiffanian (T13) in age.

T12 lineage zone sites that have undergone paleomagnetic analysis are Scarritt Quarry in the Crazy Mountains Basin of Montana and localities in the Clark's Fork Basin of Wyoming. The strata that contain all these sites are of reversed polarity and are correlated to magnetic polarity chron C26r (Butler et al. 1981, 1987).

First appearances: *Caenolambda*, *Mentoclaenodon*, *Neoliotomus?*, and *Palaeosinopa*

Last appearances: *Anconodon*, *Desmatoclaenus*, *Neoclaenodon*, and *Simpsonictis?*

Index fossils: *Plesiadapis anceps* and *Nannodectes gazini*

Characteristic fossils: *Bisonalveus*, *Carpodaptus*, *Chriacus*, *Claenodon*, *Conacodon?*, *Dissacus*, *Ectocion*, *Ectypodus*, *Elpidophorus*, *Haplaletes*, *Ignacius*, *Leptacodon*, *Litocherus*, *Litomylylus*, *Mesodma*, *Microcosmodon*, *Mimotricentes*, *Nannodectes*, *Neoplagiaulax*, *Paleotomus*, *Pararyctes*, *Paromomys?*, *Peradectes*, *Phenacodus*, *Picrodus*, *Plesiadapis*, *Prodiacodon*, *Promioclauenus*, *Propalaeosinopa*, *Protictis*, *Protoselene*, *Ptilodus*, *Thryptacodon*, *Titanoides*, and *Unuchinia*

Taxa absent but known before and after T12: *Acmeodon*, *Adunator*, *Aphronorus*, *Baiotomeus*, *Catopsalis*, *Colpoclaenus*, *Diacodon?*, *Elphidotarsius*, *Eudaemonema*, *Gelastops*, *Limaconyssus*, *Mimetodon*, *Myrmecoboides*, *Navajovius*, *Palaechthon*, *Palaeictops*, *Palaeoryctes*, *Parectypodus*, *Periptychus*, *Psittacotherium*, *Stilpnodon?*, and *Thylacaelurus?*

***Plesiadapis rex/Plesiadapis churchilli* Lineage Zone (Ti3)** We define the *P. rex/P. churchilli* Lineage Zone to include faunas that occurred between the first appearance of *P. rex* and the first appearance of *P. churchilli*.

The Ti3 lineage zone is much better represented than the Ti1 and Ti2 lineage zones. Fifty percent more genera and localities are known from the Ti3 lineage zone than from Ti1 or Ti2 (Archibald et al. 1987). Cedar Point Quarry, in the Bighorn Basin of northwestern Wyoming, contains the largest sample, numbering almost 2000 specimens. Available papers describe several taxonomic groups such as adapisoricids (Krishtalka 1976a; Gingerich 1983), nyctitheriids (Krishtalka 1976b), apatemyids (West 1973), plesiadapids (Gingerich 1976), carpolestids (Rose 1975), arctocyonids (Van Valen 1978), phenacodontids (West 1971, 1976; Thewissen 1990), carnivores (Gingerich and Winkler 1985), creodonts (Gingerich 1980b), pantodonts (Simons 1960), and microsypoidea (Gunnell 1989). Rose (1981a) presented a preliminary list of the Cedar Point Quarry fauna and the relative abundances of taxa in the sample, which was dominated by *P. rex* and *Ptilodus*.

Smaller samples that have received descriptive treatment are Ray's Bonebed from the Big Bend area of Texas (Schiebout 1974; Standhardt 1986; Schiebout et al. 1987); the Ledge, Saddle Annex, and West End sites in the Bison Basin of Wyoming (Gazin 1956b); the Brisbane and White's River Basin Survey Site from the Williston Basin of North Dakota (Holtzman 1978; Hartman and Kihm 1991, 1995); the Chappo Type Locality from the Green River Basin of southwest Wyoming (Dorr and Gingerich 1980; Gunnell 1994); the Battle Mountain locality from the Hoback Basin of southwest Wyoming (Dorr 1958); and the Police Point Local Fauna from southeast Alberta (Krishtalka 1973).

Other important sites representing the Ti3 lineage zone include Simpson's Locality 13 (Simpson 1937b) in the Crazy Mountains Basin of south-central Montana, a site that occurs in strata of reversed polarity correlated with magnetic polarity chron C26r (Butler et al. 1987); The Breaks Local Fauna from the Hanna Basin (Higgins 2000) and perhaps the Sand Creek fauna (two taxa) from the Carbon Basin (Secord 1998), both of south-central Wyoming; and a series of sites from the Paskapoo Formation in Alberta (Fox 1983, 1984a–1984d, 1990c; Webb 1995). Preliminary faunal lists provided by Fox (1990c) from these sites (Hand Hills West, upper level; Blindman River localities DW-1, DW-2, DW-3, Mel's Place; Burbank; Joffre Bridge localities) indicate that a diverse middle Tiffanian fauna is present in this area of Alberta. Specimens from the DW-2 site are especially noteworthy because of their exceptional preservation and completeness (Fox 1990c).

Of the many Ti3 sites, only Ray's Bonebed, Simpson Locality 13, and localities in the Clark's Fork Basin have undergone paleomagnetic analysis. Results indicate that these sites occur in strata of both reversed and normal polarity correlated magnetic polarity chron C26r and C26n (Butler et al. 1981; Rapp et al. 1983; Schiebout et al. 1987; Butler et al. 1987).

First appearances: *Aletodon*, *Apatemys*, *Barylambda*, *Carpocristes*, *Chiromyoides*, *Cyriacotherium*, *Dorraletes*, *Lambertocyon*, *Litolestes*, *Micromomys*, *Prochetodon*, *Raphictis*, and *Zanycteris*

Last appearances: *Acmeodon*, *Aphronorus*, *Baiotomeus*, *Bisonalveus*, *Caenolambda*, *Colpoclaenus*, *Conacodon?*, *Elphidotarsius*, *Eudaemonema*, *Gelastops*, *Mentoclaenodon*, *Palaechthon*, *Paromomys?*, *Promioclaenus*, *Protoselene*, and *Stilpnodon?*

Index fossils: *Cedrocherus*, *Chiromyoides minor*, *Copacion*, *Melaniella*, *Nannodectes simpsoni*, *Plesiadapis rex*, *Saxonella*, and *Tytthaena*

Characteristic fossils: *Adunator*, *Carpodaptes*, *Chriacus*, *Claenodon*, *Dissacus*, *Ectocion*, *Ectypodus*, *Elpidophorus*, *Haplaletes*, *Ignacius*, *Leptacodon*, *Litocherus*, *Litomylus*, *Mesodma*, *Microcosmodon*, *Mimetodon*, *Mimotricentes*, *Myrmecoboides*, *Nannodectes*, *Navajovius*, *Neoliotomus?*, *Neoplagiaulax*, *Palaeoryctes*, *Palaeosinopa*, *Paleotomus*, *Pararyctes*, *Parectypodus*, *Peradectes*, *Periptychus*, *Phenacodus*, *Picrodus*, *Plesiadapis*, *Prodiacodon*, *Propalaeosinopa*, *Protictis*, *Psittacotherium*, *Ptilodus*, *Thryptacodon*, *Titanoides*, and *Unuchinia*

Taxa absent but known before and after Ti3: *Catopsalis*, *Diacodon?*, *Limaconyssus*, *Palaeictops*, and *Thylacaelurus?*

***Plesiadapis churchilli/Plesiadapis simonsi* Lineage Zone (Ti4)** We define the *P. churchilli/P. simonsi* Lineage Zone to include faunas that occurred between the first appearance of *P. churchilli* and the first appearance of *P. simonsi*.

The type Tiffanian fauna from the Mason Pocket site of southwestern Colorado lies in the Ti4 lineage zone and was described in detail by Simpson (1935c, 1935d, 1935e). Paleomagnetic analysis of strata containing Mason Pocket exhibit reversed polarity, and the rocks are correlated with magnetic polarity chron C25r based on faunal correlation to mammalian assemblages in the Clark's Fork and Bighorn basins (Butler et al. 1981).

The largest sample (exceeding 5000 specimens) of Ti4 lineage zone mammals is from the Roche Percée localities in southeastern Saskatchewan. Thus far, only multituberculates (Krause 1977), primates (Krause 1978), and

a pantodont (Rose and Krause 1982) have been described from the Roche Percée Local Fauna.

Also from Canada is a series of sites referred to the Ti4 lineage zone from the Paskapoo Formation. The Crestomere School and Canyon Ski Quarry sites have yet to be fully described, although a preliminary faunal list was provided by Fox (1990c). Swan Hills Site 1 is the most northerly Tiffanian and North American Paleocene fossil mammal locality known (Archibald et al. 1987), and its fauna has been described in some detail (L. Russell 1967; Krishtalka 1973; Krause 1978; Gingerich 1986; Fox 1990c).

Wolberg (1979) presented preliminary faunal lists for the Olive and Circle local faunas of eastern Montana. Included in Wolberg's list from the Olive site are several taxa that otherwise are unknown from the Ti4 lineage zone (e.g., *Nyctitherium*, *Protentomodon*, cf. *Purgatorius*). They have not been included as first or last appearances in the lists in this chapter pending publication of full descriptions and analyses.

In the Bighorn Basin are several Ti4 lineage zone localities of interest (Airport, Witter Quarry, Divide Quarry) because they can be placed in stratigraphic relationship to sites that yield mammalian assemblages representative of the Ti3 or Ti5 lineage zones (Archibald et al. 1987).

Mammalian assemblages from other Ti4 lineage zone localities include a series of sites in the Rock Springs uplift of southwestern Wyoming (Winterfeld 1982), Malcolm's Locality in the Wind River Basin of central Wyoming (Krishtalka et al. 1975), and a series of localities in the Williston Basin of central North Dakota (Holtzman 1978; Erickson 1991, 1999; Hartman and Kihm 1991, 1995, 1999). The Wannagan Creek Local Fauna is of special interest because the preliminary faunal list includes cf. *Phenacolemur*, *Entomolestes* sp., and *Leptictis* sp. (see Erickson 1991, 1999). All three taxa represent significant range extensions. Pending description of specimens, we tentatively include *Phenacolemur* and *Entomolestes* in our faunal characterization for the Ti4 lineage zone.

Paleomagnetic analyses of strata yielding mammals representing the Ti4 lineage zone from southern Colorado and the Clark's Fork Basin of Wyoming indicate that these rocks are of reversed and normal polarity correlated with magnetic polarity chrons C26n and C25r (Butler et al. 1981).

First appearances: *Anacodon*, *Entomolestes*, *Haplolambda*, *Ignatiolambda*, *Phenacodaptes*, and *Phenacolemur*?

Last appearances: *Catopsalis*, *Elpidophorus*, *Litomylus*, *Mesodma*, *Pararyctes*, *Periptychus*, *Raphictis*, and *Zanycteris*

Index fossils: *Amelotabes*, *Carpomegodon*, *Chiromyoides caesor*, *Nannodectes gidleyi*, *Plesiadapis churchilli*, *Utemylus*, and *Xenacodon*

Characteristic fossils: *Adunator*, *Aletodon*, *Apatemys*, *Barylambda*, *Carpocristes*, *Carpodaptes*, *Chiromyoides*, *Chriacus*, *Claenodon*, *Cyriacotherium*, *Dissacus*, *Ectocion*, *Ectypodus*, *Haplaletes*, *Ignacius*, *Lambertocyon*, *Leptacodon*, *Litocherus*, *Litolestes*, *Microcosmodon*, *Micromomys*, *Mimetodon*, *Mimotricentes*, *Nannodectes*, *Navajovius*, *Neoplagiaulax*, *Palaeictops*, *Palaeoryctes*, *Palaeosinopa*, *Peradectes*, *Phenacodus*, *Plesiadapis*, *Prochetodon*, *Propalaeosinopa*, *Protictis*, *Ptilodus*, *Thryptacodon*, *Titanoides*, and *Unuchinia*

Taxa absent but known before and after Ti4: *Diacodon*?, *Dorraletes*, *Limaconyssus*, *Myrmecoboides*, *Neoliotomus*?, *Paleotomus*, *Parectypodus*, *Picrodus*, *Prodiacodon*, *Psittacotherium*?, and *Thylacaelurus*?

***Plesiadapis simonsi*/*Plesiadapis gingerichi* Lineage Zone (Ti5)** We define the *P. simonsi*/*P. gingerichi* Lineage Zone to include faunas that occurred between the first appearance of *P. simonsi* and the first appearance of *P. gingerichi*.

Most Ti5 lineage zone localities are from the Clark's Fork and Bighorn basins. Of these, the mammalian fauna from Princeton Quarry is the best known because the fauna was examined in detail by Rose (1981a, 1981b). Rose (1981a) noted that there may be a significant size bias in the sample because the fauna is dominated by smaller taxa (*Phenacodaptes*, *Plesiadapis*), whereas larger forms are rare or absent.

Small collections of Ti5 lineage zone mammals have been described from the Bayfield area, northern San Juan Basin, Colorado (Simpson 1935c, 1935d, 1935e); various localities at the Rock Springs uplift, Wyoming (Winterfeld 1982); the *Titanoides* Locality, Bison Basin, Wyoming (Gazin 1956b); the Dell Creek Quarry, Hoback Basin, Wyoming (Dorr 1952, 1958, 1978); and Joe's Bonebed, Big Bend area, Texas (Schiebout 1974; Standhardt 1986; Schiebout et al. 1987).

Recently, Schoch (1985, 1998) reported a sparse but unique occurrence of Paleocene land mammals from the East Coast of North America (Williamsburg Formation, South Carolina) consisting of three identifiable specimens referred to *Mingotherium*, *Ectoganus*, and *Phenacodus grangeri*. *Mingotherium* is a genus of uncertain affinity for which the temporal range is unknown. However, *Ectoganus* occurs in strata elsewhere from the Ti5 lineage zone through the Clarkforkian Mammal Age (Archibald et al. 1987), and *P. grangeri* is known from the Ti1 to Ti5 lineage zones (Thewissen

1990). Therefore we tentatively assign this fauna to the Ti5 lineage zone.

Strata that contain Joe's Bonebed and Ti5 sites in the Clark's Fork Basin are of reversed and normal polarity correlated with magnetic polarity chron C25r and C25n (Butler et al. 1981; Rapp et al. 1983; Schiebout et al. 1987).

First appearances: *Arctostylops*, *Carpolestes*, *Didymictis*, *Dillerlemur*, *Dipsalodon*, *Ectoganus*, *Oxyaena*, *Princetonia*, *Prodinoceras*, and *Viverravus*

Last appearances: *Carpocristes*, *Carpodaptes*, *Claenodon*, *Dorraletes*, *Haplaletes*, *Ignatiolambda*, *Litocherus*, *Litolestes*, *Mimetodon*, *Myrmecoboides*, *Nannodectes*, *Navajovius*, *Neoplagiaulax*, *Paleotomus*, *Picrodus*, *Propalaeosinopa*, *Protictis*, *Psittacotherium?*, *Ptilodus*, and *Unuchinia*

Index fossils: *Aaptoryctes*, *Chiromyoides potior*, *Mingotherium*, *Pentacosmodon*, *Plesiadapis fodinatus*, *Plesiadapis simonsi*, and *Propalaeonodon*

Characteristic fossils: *Adunator*, *Aletodon*, *Anacodon*, *Barylambda*, *Chiromyoides*, *Chriacus*, *Cyriacotherium*, *Diacodon*, *Dissacus*, *Ectocion*, *Ectypodus*, *Haplolambda*, *Ignacius*, *Lambertocyon*, *Leptacodon*, *Microcosmodon*, *Micromomys*, *Mimotricentes*, *Neoliotomus*, *Palaeictops*, *Palaeoryctes*, *Palaeosinopa*, *Parectypodus*, *Peradectes*, *Phenacodaptes*, *Phenacodus*, *Plesiadapis*, *Prochetodon*, *Prodiacodon*, *Thryptacodon*, and *Titanoides*

Taxa absent but known before and after Ti5: *Apatemys*, *Entomolestes*, *Limaconyssus*, *Phenacolemur?*, and *Thylacaelurus?*

#### ***Plesiadapis gingerichi*/Rodentia Interval Subzone (Ti6) of the *P. gingerichi*/*P. cookei* Lineage Zone (Ti6–Cf1)**

*P. gingerichi* was described by Rose (1981a), who interpreted it to be phylogenetically and temporally intermediate between *P. simonsi* (latest Tiffanian, Ti5) and *P. cookei* (middle Clarkforkian, Cf2). Following Rose (1981a) and Archibald et al. (1987), we define the *P. gingerichi*/*P. cookei* Lineage Zone to include faunas that occurred between the first appearance of *P. gingerichi* and the first appearance of *P. cookei*.

The Ti6–Cf1 lineage zone as first defined by Rose (1980, 1981a) was interpreted to straddle the Tiffanian–Clarkforkian boundary (Archibald et al. 1987). To specify this relationship, Archibald et al. (1987) subdivided the lineage zone into two interval subzones. The end of the Ti6 interval subzone and commencement of the Cf1 interval subzone equaled the Tiffanian–Clarkforkian boundary (as well as first appearance of the Rodentia). Thus Archibald et al. (1987) defined the first interval subzone, the *P. gingerichi*/Rodentia Interval Subzone (Ti6),

to include faunas that occurred between the first appearance of *P. gingerichi* and first appearance of the Rodentia. The purpose in recognizing two interval subzones in the Ti6–Cf1 lineage zone was to highlight that the lineage zone straddles the Tiffanian–Clarkforkian boundary (Archibald et al. 1987). We accept and follow this reasoning.

The Ti6 interval subzone has been recognized with certainty only in the Clark's Fork Basin (and perhaps the Wind River Basin) of Wyoming, and the subzone is poorly known (Archibald et al. 1987). The long list of genera given here for taxa absent but known before and after Ti6 is evidence of this fact. Several other faunas that may be referable to the Ti6 interval subzone are discussed later in this chapter under the Cf1 interval subzone.

Sites referred to the Ti6 interval subzone from the Clark's Fork Basin occur in strata of normal polarity correlated to magnetic polarity chron C25n (Butler et al. 1981).

First appearances: *Esthonyx?*, *P. gingerichi*, and *Plesiadapis dubius*

Last appearances: *Mimotricentes*

Index fossils: none

Characteristic fossils: *Adunator*, *Carpolestes*, *Chiromyoides*, *Cyriacotherium*, *Didymictis*, *Dillerlemur*, *Dipsalodon?*, *Dissacus*, *Ectocion*, *Ectoganus*, *Leptacodon*, *Neoliotomus*, *Oxyaena*, *Palaeosinopa*, *Phenacodus*, *Plesiadapis*, *Prodinoceras*, and *Prochetodon*

Taxa absent but known before and after Ti6: *Aletodon*, *Anacodon*, *Apatemys*, *Arctostylops*, *Barylambda*, *Chriacus*, *Diacodon?*, *Ectypodus*, *Entomolestes*, *Haplolambda*, *Ignacius*, *Lambertocyon*, *Limaconyssus*, *Microcosmodon*, *Micromomys*, *Palaeictops*, *Palaeoryctes*, *Parectypodus*, *Peradectes*, *Phenacodaptes*, *Phenacolemur?*, *Princetonia*, *Prodiacodon*, *Thryptacodon*, *Thylacaelurus?*, *Titanoides?*, and *Viverravus*

## **CLARKFORKIAN MAMMAL AGE**

Granger (1914) applied the term *Clark Fork* to a stratigraphic interval at the southwestern end of Polecat Bench in the Clark's Fork Basin of northwestern Wyoming. Fossils in this "Clark Fork fauna" were described by Matthew (1915a, 1915b, 1915c), Granger (1915), Jepsen (1930, 1940), and Simpson (1929b, 1937c). Wood et al. (1941:9) formally proposed the Clarkforkian as a North American provincial age, "based on the Clark Fork member (and faunal zone) of the Polecat Bench Formation"; they selected Granger's locality near Polecat Bench as the type locality.



Also, Wood et al. (1941) considered the Clarkforkian as the youngest of the Paleocene provincial ages.

Because of the small collections, inadequate stratigraphic documentation, and other factors, there was initial skepticism about legitimacy of the Clarkforkian mammal age. Eventually, that skepticism culminated in rejection by R. Wood (1967) of the Clarkforkian as a valid mammal age. Nevertheless, collecting in the Clark's Fork Basin in the 1970s and 1980s yielded thousands of specimens that clearly reaffirmed the recognizability of a distinctive Clarkforkian fauna (Gingerich and Rose 1977; Rose 1978, 1980, 1981a). In addition, study of these collections indicated that Clarkforkian assemblages in the type area are not limited to the Fort Union Formation (= Polecat Bench Formation) but occur primarily in lower parts of the overlying Willwood Formation (Archibald et al. 1987).

Wood et al. (1941) did not recognize any faunal correlatives elsewhere of the Clarkforkian mammal age. Later, however, correlatives of this age were recognized and discussed by Rose (1981a). These faunas occur in the following formations: Fort Union and Willwood formations, Clark's Fork and Bighorn basins, northern Wyoming and southern Montana (Sinclair and Granger 1912; Simpson 1928, 1929a, 1929c; Jepsen 1937; Van Houten 1944; Van Valen and Sloan 1966; Gingerich and Rose 1979); "lower variegated sequence," Togwotee Pass area, northwestern Wyoming (McKenna 1972, 1980); Chappo Member, Wasatch Formation, Hoback Basin, western Wyoming (Dorr 1952, 1958, 1978; Dorr and Steidtmann 1970; Dorr et al. 1977) and Green River Basin, southwestern Wyoming (Gazin 1942, 1956a; Dorr and Gingerich 1980); Fort Union Formation, Washakie Basin, south-central Wyoming (Rose 1981a); Debeque Formation, Piceance Creek Basin, northwestern Colorado (Patterson 1933, 1936, 1937, 1939, 1949; Patterson and Simons 1958; Patterson and West 1973; Kihm 1984); and possibly localities in the Tornillo Formation, Big Bend area, southwestern Texas (Schiebout 1974; Schiebout et al. 1987). Rose (1981a) provided stratigraphic sections, descriptions, and ranges of Clarkforkian mammals from the type area and a discussion of most of the Clarkforkian assemblages listed here.

#### TIFFANIAN-CLARKFORKIAN BOUNDARY

In the Clark's Fork Basin, the beginning of the Clarkforkian mammal age can be recognized by the first occurrence of the orders Rodentia and Tillodontia (*Esthonyx*) and the genera *Haplomyilus* (Condylarthra) and *Coryphodon* (Pantodonta). It was appearance of the Rodentia, however, that was used to define the beginning of

this mammal age (Gingerich and Gunnell 1979; Rose 1980, 1981a; Archibald et al. 1987). All of these first appearances seem to represent immigrants, and discoveries in Paleocene strata of China suggest that rodents and tillodonts may have originated in Asia (Wang 1975; Zhou et al. 1977; Gingerich 1980a; Zhang 1980; Dawson et al. 1984). The first occurrence of any of these four immigrants is a good indication of the beginning of the Clarkforkian mammal age, but none is common in early parts of this mammal age (Rose 1981a; Archibald et al. 1987).

Wood et al. (1941) listed the Tiffanian as the oldest record of Rodentia. However, this was based on the occurrence of rodents at Bear Creek, Montana (Jepsen 1937), a locality that is now considered to be of Clarkforkian age (Van Valen and Sloan 1966; Sloan 1970; Rose 1975, 1977, 1981a; Gingerich 1976).

Exposures yielding the type Clarkforkian fauna in vicinity of the Polecat Bench–Clark's Fork Basin occur in an interval about 470 m thick in the upper Fort Union and lower Willwood formations. The earliest Clarkforkian faunas are found about 350 m above the level of the late Tiffanian Princeton Quarry. The only Clarkforkian faunas known outside the Clark's Fork Basin that may overlie Tiffanian faunas and thus include the Tiffanian–Clarkforkian boundary are from strata preserved in the Togwotee Pass area and the Hoback Basin, both in western Wyoming. However, collections from these strata do not permit precise location of the Tiffanian–Clarkforkian boundary (Archibald et al. 1987).

#### DEFINITION AND CHARACTERIZATION

Archibald et al. (1987), using Rose's (1980, 1981a) study of the mammalian fauna from the area of the type Clarkforkian, defined the Clarkforkian mammal age to include faunas that occurred between the first appearance of the Rodentia and the first appearance of the Artiodactyla.

Wood et al. (1941) listed cf. *Coryphodon*, *Ectocion*, *Esthonyx*, and *Oxyaena* as first appearing in the Clarkforkian mammal age. Indeed, *Coryphodon* and perhaps *Esthonyx* (there is a questionable occurrence in the Ti6 interval subzone) are now known to appear for the first time in the Clarkforkian. However, *Ectocion* and *Oxyaena* have been recorded subsequently from Tiffanian faunas. Wood et al. (1941) also listed *Carpolestes* and *Plesiadapis* as having last appearances in the Clarkforkian mammal age. The final appearance of *Carpolestes* is still known from Clarkforkian strata, but a single specimen of *Plesiadapis dubius* was reported from an early Wasatchian fauna (Rose and Bown 1982). Wood et al. (1941) listed *Plesiadapis cookei* as the only index fossil for the Clarkforkian

mammal age, and this assignment remains valid. Characteristic fossils listed by Wood et al. (1941) for this mammal age (i.e., *Didymictis*, *Ectypodus*, *Phenacodus*, “*Probatyopsis*,” and *Thryptacodon*) remain valid in that sense, although McKenna and Bell (1997) synonymized “*Probatyopsis*” with *Prodinoceras*.

In their update of Wood et al. (1941), Archibald et al. (1987) commented on the following aspects of Clarkforkian faunas. First, common index fossils of the Clarkforkian include *Plesiadapis cookei*, *Carpolestes nigridentis*, *Aletodon gunnelli*, *Apheliscus nitidus*, *Haplomyilus simpsoni*, *Dissacus praenuntius*, *Esthonyx xenicus*, *Esthonyx ancylion*, and *Acritoparamys atavus*, and all occur in at least one Clarkforkian fauna outside the Clark’s Fork Basin in addition to the type Clarkforkian fauna. Second, the most common taxa of the Clarkforkian mammal age are phenacodontid condylarths (*Ectocion osbornianus*, *Phenacodus primaevus*, and *Phenacodus vortmani*), which together constitute about 50 percent of individuals at most levels in the Clark’s Fork Basin (Rose 1981a, 1981b). Third, *Ectocion* is the most common taxon at most localities. Finally, phenacodontids persist into Wasatchian time but exhibit a dramatic decline in abundance at the Clarkforkian–Wasatchian boundary.

Magnetostratigraphic sections have been developed in the Clark’s Fork and Bighorn basins, northern Wyoming (Butler et al. 1980, 1981), and these sections encompass all three zones recognized by Rose (1981a) for the type Clarkforkian fauna and part of the next younger, Wasatchian fauna. These data indicate that Clarkforkian faunas extend from magnetic polarity chron C25n into C24r.

The characterization of the Clarkforkian mammal age is as follows.

First appearances: *Acritoparamys*, *Apatosciuravus*, *Apheliscus*, *Arctodontomys*, *Chalicomomys*, *Coryphodon*, *Dipsalidictides*, *Franimys*, *Haplomyilus*, *Hyopsodus*, *Icaronycteris?*, *Leipsanolestes*, *Meniscotherium*, *Microparamys*, *Mimoperadectes?*, *Niptomomys*, *Palaeonodon*, *Paramys*, *Plagiocetenodon*, *Plagiomene*, *Pontifactor?*, *Reithroparamys*, *Tinimomys*, *Uintacyon*, and *Worlandia*

Last appearances: *Adunator*, *Aletodon*, *Anacodon*, *Arctostylops*, *Carpolestes*, *Chiromyoides*, *Cyriacotherium*, *Dipsalodon*, *Haplolambda*, *Lambertocyon*, *Limaconysus*, *Microcosmodon*, *Palaeoryctes*, *Phenacodaptes*, *Prochetodon*, and *Titanoides?*

Index fossils: *Acidomomys*, *Acritoparamys atavus*, *Alagomys*, *Aletodon gunnelli*, *Apheliscus nitidus*, *Carpolestes nigridentis*, *Ceutholestes*, *Dissacus praenuntius*, *Esthonyx ancylion*, *Esthonyx xenicus*, *Haplomyilus simp-*

*soni*, *Palaeonictis*, *Planetetherium*, *Plesiadapis cookei*, *Protentomodon*, *Thelysia*, and *Wyonycteris*

Characteristic fossils: *Apatemys*, *Barylambda*, *Chriacus*, *Diacodon?*, *Didymictis*, *Dillerlemur*, *Dissacus*, *Ectocion*, *Ectoganus*, *Ectypodus*, *Esthonyx*, *Ignacius*, *Leptacodon*, *Neoliotomus*, *Oxyaena*, *Palaeosinopa*, *Parectypodus*, *Peradectes*, *Phenacodus*, *Phenacolemur*, *Plesiadapis*, *Princetonia*, *Prodiacodon*, *Prodinoceras*, *Thryptacodon*, and *Viverravus*

Taxa absent but known before and after the Clarkforkian: *Entomolestes*, *Micromomys*, *Palaeictops*, and *Thylacaelurus?*

## ZONATION

Archibald et al. (1987) subdivided the Clarkforkian mammal age into one subzone and two zones based on work by Rose (1980, 1981a) in the Clark’s Fork Basin. The first subzone in the Clarkforkian mammal age, the Rodentia/*Plesiadapis cookei* Interval Subzone (Cf1), is the second of two interval subzones in the *P. gingerichi*/*P. cookei* Lineage Zone (Ti6–Cf1). As mentioned previously, the Ti6–Cf1 lineage zone, as defined by Rose (1980, 1981a), straddles the Tiffanian–Clarkforkian boundary, and Archibald et al. (1987) recognized two interval subzones within it. The Ti6 interval subzone corresponds to the Tiffanian part of the Ti6–Cf1 lineage zone, and the Cf1 interval subzone corresponds to its Clarkforkian part.

The second Clarkforkian zone, the *Plesiadapis cookei* Lineage Zone (Cf2), bears only the name of the species defining the beginning of the zone; the third Clarkforkian zone is not based on the first appearance of a single taxon and therefore is neither a lineage zone nor an interval zone (Archibald et al. 1987). This third zone, the *Phenacodus*/*Ectocion* Acme Zone (Cf3), is based on the simultaneous abundance of the phenacodontid condylarths *Phenacodus* and *Ectocion* (Archibald et al. 1987).

Recognition of this zonation in strata of the Clark’s Fork Basin was augmented by use of species of *Phenacolemur* and *Esthonyx* (see Rose 1981a; Archibald et al. 1987). *Esthonyx xenicus* occurs in the Ti6–Cf1 lineage zone and through the lowest 30 m of strata bearing faunas referable to the Cf2 lineage zone. *Esthonyx ancylion* continues above the 30-m level in strata bearing faunas of the Cf2 lineage zone into about the lowest 20 m of strata with faunas assigned to the Cf3 acme zone. *Esthonyx grangeri* continues through the remainder of the Cf3 acme zone into the Wasatchian mammal age. *Phenacolemur pagei* first appears in the later part of the Tiffanian mammal age and continues into the Clarkforkian mammal age, coexisting with *E. xenicus* and *E. ancylion*. *Phenacolemur praecox* fol-

lows *P. pagei* and, as does *E. grangeri*, continues into the Wasatchian mammal age. Archibald et al. (1987) indicated that although species of *Esthonyx* and *Phenacolemur* may be useful in recognition of Clarkforkian zones, these two genera constitute evolving lineages that are somewhat arbitrarily divided into species.

**Rodentia/*Plesiadapis cookei* Interval Subzone (Cf1) of the *P. gingerichi*/*P. cookei* Lineage Zone (Ti6–Cf1)** We follow Archibald et al. (1987) and define the Rodentia/*P. cookei* Interval Subzone to include faunas that occurred between the first appearance of the Rodentia and the first appearance of *P. cookei*.

The Cf1 interval subzone can be recognized with certainty only in the Clark's Fork Basin (Archibald et al. 1987). The only other local faunas (in addition to those from the type area in the Clark's Fork Basin) that are referred to this interval subzone are Bear Creek, some sites in the Bighorn Basin, perhaps a site in the Togwotee Pass area west of the Wind River Basin (Archibald et al. 1987), and probably the Big Multi Quarry in the Washakie Basin of southwestern Wyoming (Wilf et al. 1998). The Big Multi Quarry is especially important because it has yielded a large and diverse assemblage (Rose 1981a; Wilf et al. 1998, table 3) and it records the only known Clarkforkian occurrence of the rodent *Alagomys* (Dawson and Beard 1996). The Big Multi Quarry was previously assigned to the Cf2 lineage zone (Archibald et al. 1987). However, a correlation with the Cf1 interval subzone is now advocated by Wilf et al. (1998) based on additional samples collected from the site beginning in 1992.

Based on paleomagnetic correlations provided by Butler et al. (1981) from the Clark's Fork Basin, the Cf1 interval subzone begins in strata of normal polarity and continues up into those of reversed polarity correlated to magnetic polarity chrons C25n and C24r.

First appearances: *Acritoparamys*, *Apatosciuravus*, *Apheliscus*, *Arctodontomys*, *Ceutholestes*, *Chalicomomys*, *Coryphodon*, *Dipsalidictides*, *Haplomylus*, *Leipsanolestes*, *Palaeonodon*, *Paramys*, *Plagioctenodon*, *Planetetherium*, *Tinimomys*, and *Wyonycteris*

Last appearances: *Haplolambda*, *Lambertocyon*, *Phenacodaptes*, *Plesiadapis gingerichi*, and *Titanoides*?

Index fossils: *Alagomys* and *Protentomodon*

Characteristic fossils: *Adunator*, *Aletodon*, *Apatemys*, *Barylambda*, *Carpolestes*, *Chiromyoides*, *Chriacus*, *Cyriacotherium*, *Didymictis*, *Dillerlemur*, *Dissacus*, *Ectocion*, *Ectogonus*, *Ectypodus*, *Esthonyx*, *Ignacius*, *Leptacodon*, *Limaconyssus*, *Microcosmodon*, *Neoliotomus*, *Oxyaena*, *Palaeoryctes*, *Palaeosinopa*, *Parectypodus*, *Per-*

*aedectes*, *Phenacodus*, *Phenacolemur*, *Plesiadapis*, *Prochetodon*, *Prodinoceras*, *Thryptacodon*, and *Viverravus*  
Taxa absent but known before and after Cf1: *Anacodon*, *Arctostylops*, *Diacodon?*, *Dipsalodon*, *Entomolestes*, *Micromomys*, *Palaeictops*, *Princetonia*, *Prodiacodon*, and *Thylacaelurus?*

***Plesiadapis cookei* Lineage Zone (Cf2)** *P. cookei* is limited to the middle of the Clarkforkian mammal age. Its appearance marks the beginning of the lineage zone bearing its name, and it is common in this interval. The species disappears abruptly in the Clark's Fork Basin section (and apparently elsewhere), and its absence is characteristic of the succeeding Cf3 acme zone (Rose 1981a). Based on these data, Archibald et al. (1987) defined this lineage zone to include faunas that occurred between the first and last appearances of *P. cookei*. We follow their usage.

Of the three Clarkforkian zones, the Cf2 lineage zone can be recognized over the widest geographic area. In the Clark's Fork Basin section, local faunas referable to the Cf2 lineage zone occur in the middle 200 m of strata containing Clarkforkian faunas (Archibald et al. 1987). Another series of major Cf2 interval zone sites is located in the Bighorn Basin, and these are the Rough Gulch, Foster Gulch, and Ries localities. Rough Gulch and Foster Gulch have yielded *P. cookei*, the index taxon for the Cf2 lineage zone (Archibald et al. 1987). One of the largest and most diverse Clarkforkian assemblages outside the Clark's Fork or Bighorn basins occurs in the Togwotee Pass area in northwestern Wyoming (Rose 1981a), where *P. cookei* also has been recovered. The local fauna from Buckman Hollow of the Green River Basin, Wyoming, is another site that yields a mammalian assemblage referable to the Cf2 lineage zone based on many diagnostic taxa including *P. cookei* (Gazin 1942, 1956; Archibald et al. 1987). Other areas that might contain Clarkforkian local faunas that might be referable to the Cf2 lineage zone are the Hoback and Wind River basins of Wyoming and the Piceance Creek Basin of Colorado (Kihm 1984).

Paleomagnetic correlations indicate that the Cf2 lineage zone occurs entirely in magnetic polarity chron C24r (Butler et al. 1981).

First appearances: *Aletodon gunnelli*, *Esthonyx ancylion*, *Franimys*, *Microparamys*, *Niptomomys*, *Plagiomene*, *Pontifactor?*, *Reithroparamys*, *Uintacyon*, and *Worlandia*

Last appearances: *Anacodon*, *Chiromyoides*, *Esthonyx xenicus*, *Microcosmodon*, and *Planetetherium?*

Index fossils: *Acidomomys*, *Chiromyoides major*, *Plesiadapis cookei*, and *Thelysia*

Characteristic fossils: *Adunator*, *Aletodon*, *Apatemys*, *Apatosciuravus*, *Apheliscus*, *Arctodontomys*, *Arctostylops*, *Barylambda*, *Carpolestes*, *Chriacus?*, *Coryphodon*, *Cyriacotherium*, *Diacodon?*, *Didymictis*, *Dillerlemur*, *Dipsalidictides*, *Dipsalodon*, *Dissacus*, *Ectocion*, *Ectogonus*, *Ectypodus*, *Esthonyx*, *Haplomylus*, *Ignacius*, *Leipsanolestes*, *Leptacodon*, *Neoliotomus*, *Oxyaena*, *Palaeonodon*, *Palaeoryctes*, *Palaeosinopa*, *Paramys*, *Parectypodus*, *Peradectes*, *Phenacodus*, *Phenacolemur*, *Plagiocetenodon?*, *Plesiadapis*, *Princetonia*, *Prochetodon*, *Prodiacodon?*, *Prodinoceras*, *Thryptacodon*, *Tinimomys*, and *Viverravus*

Taxa absent but known before and after Cf<sub>2</sub>: *Acritoparamys*, *Ceutholestes*, *Chalicomomys*, *Entomolestes*, *Limaconyssus*, *Micromomys*, *Palaeictops*, *Thylacaelurus?*, and *Wyonycteris*

**Phenacodus–Ectocion Acme Zone (Cf<sub>3</sub>)** The beginning of the Cf<sub>3</sub> acme zone occurs immediately after the last appearance of *Plesiadapis cookei* (Rose 1981a; Archibald et al. 1987). It can be recognized further by the evolutionary first occurrence of *Esthonyx grangeri* and *Phenacolemur praecox*, which make their appearances during but not at the beginning of the zone (Rose 1981a). Archibald et al. (1987) did not offer a formal definition of this zone but noted that the end of the zone is marked by the appearance of Artiodactyla. Other taxa common in the Cf<sub>3</sub> acme zone are *Ectocion*, *Phenacodus*, *Prodinoceras*, and *Didymictis* (Archibald et al. 1987).

Outside the Clark's Fork and Bighorn basins, no assemblages can be referred with certainty to the Cf<sub>3</sub> acme zone (Archibald et al. 1987). The "cf. Late Clarkforkian Local Fauna" from the Piceance Creek Basin of Colorado described by Kihm (1984) may be referable to the Cf<sub>3</sub> acme zone because it includes *Phenacolemur* sp. near *P. praecox*.

Paleomagnetic correlations indicate that the Cf<sub>3</sub> acme zone occurs entirely within magnetic polarity chron C24r (Butler et al. 1981).

First appearances: *Esthonyx grangeri*, *Hyopsodus*, *Icaronycteris?*, *Meniscotherium*, *Mimoperadectes?*, and *Phenacolemur praecox*

Last appearances: *Adunator*, *Aletodon*, *Apheliscus nitidus*, *Arctostylops*, *Carpolestes*, *Ceutholestes*, *Cyriacotherium*, *Dipsalodon?*, *Dissacus praenuntius*, *Esthonyx ancylion*, *Haplomylus simpsoni*, *Limaconyssus*, *Palaeoryctes*, *Prochetodon*, and *Wyonycteris*

Index fossils: *Palaeonictis*

Characteristic fossils: *Apatemys*, *Apatosciuravus*, *Apheliscus*, *Arctodontomys*, *Barylambda?*, *Chriacus?*, *Coryphodon*, *Didymictis*, *Dillerlemur*, *Dipsalidictides*, *Dissacus*, *Ectocion*, *Ectogonus*, *Ectypodus*, *Esthonyx*, *Haplomylus*,

*Ignacius*, *Leipsanolestes*, *Leptacodon*, *Microparamys*, *Nipptomomys*, *Oxyaena*, *Palaeonodon*, *Palaeosinopa*, *Paramys*, *Peradectes*, *Phenacodus*, *Phenacolemur*, *Plagiomene*, *Plesiadapis*, *Prodiacodon?*, *Prodinoceras*, *Thryptacodon*, *Uintacyon*, *Viverravus*, and *Worlandia*

Taxa absent but known before and after Cf<sub>3</sub>: *Acritoparamys*, *Chalicomomys*, *Diacodon?*, *Entomolestes*, *Franimys*, *Micromomys*, *Neoliotomus*, *Palaeictops*, *Parectypodus*, *Plagiocetenodon*, *Pontifactor?*, *Princetonia*, *Reithroparamys*, *Thylacaelurus?*, and *Tinimomys*

## INTERCONTINENTAL CORRELATIONS

Intercontinental correlations are important in assessing global distribution patterns of mammals, centers of origin for major taxonomic groups, and dispersal events. These sorts of information also provide useful global contexts to questions of detailed correlation of biological and geologic happenings in North America itself. Therefore we briefly discuss Paleocene mammalian assemblages from other continents. We review their proposed correlations to the succession of mammal ages from North America.

## EUROPEAN PALEOCENE FAUNAS

Mammalian faunas of earliest Paleocene age are not known from Europe. The sparse mammalian assemblage from Hainin in Belgium appears to be Danian in age and is the oldest known European Paleocene fauna (Russell et al. 1982). The Hainin Local Fauna includes endemic elements mixed with taxa of North American Torrejonian affinity (Vianey-Liaud 1979). There may have been some mammalian dispersal between Europe and North America in the early Paleocene (Savage and Russell 1983). If so, however, it was minor in terms of number of taxa (Woodburne and Swisher 1995).

The next younger Paleocene assemblages in Europe appear to be those from a fissure-fill at the Walbeck site in Germany and the Campo locality in Spain. Based on stage of evolution, the Walbeck assemblage may be a temporal equivalent of the early Tiffanian (Savage and Russell 1983). The Campo locality yields a sparse assemblage of Paleocene mammals (Tambareau et al. 1992) associated with marine fossils that indicate a Thanetian age (Gheerbrant et al. 1997). The Campo assemblage is interpreted to be slightly older than the Walbeck assemblage (Gheerbrant et al. 1997). However, magnetostratigraphic analysis of rocks containing the Campo locality indicate that the site lies close to the boundary between magnetic polarity chrons C26n and

C25r, a paleomagnetic interval that correlates to the T14 lineage zone in North America (Gheerbrant et al. 1997).

The Cernaysian mammal age was proposed by Savage and Russell (1983) for a series of mammalian assemblages that occurs in sites located primarily in the Paris Basin of France. There is consensus on a late Paleocene (Thanetian) age assignment for the Cernay faunas (Berggren and Aubry 1998; Lucas 1998), younger than Campo and Walbeck. Cernaysian mammal faunas lack the large herbivores (e.g., pantodonts and uinatheres) present in presumably temporally equivalent assemblages from North America. Cernaysian assemblages show some taxonomic resemblance to late Paleocene North American faunas, but this may principally reflect inherited vestiges from preceding faunas rather than solid evidence of dispersals (Savage and Russell 1983).

The Cernay Conglomerate (type Cernaysian) is overlain by the Argile Plastique, which yields a dramatically different assemblage of mammals of Sparnacian age (Savage and Russell 1983). European Sparnacian faunas are similar to North American Wasatchian faunas, and therefore the Cernaysian is certainly older than Wasatchian. Younger Cernaysian faunas probably are a temporal equivalent of the T16–Cf1 lineage zone based on comparisons of lineages of plesiadapids (Rose 1981a; Gingerich 1976; Gingerich and Rose 1977). However, Wing (1984) suggested that Cernaysian faunas correlate with later Tiffanian rather than early Clarkforkian faunas. Lucas (1998), in further contrast, considers the Cernaysian to be correlative with the Clarkforkian.

Recently, a sparse mammalian assemblage was reported from the “Calcaires de Rona,” Transylvania, Romania (Gheerbrant et al. 1999). The fauna, which includes an incisor fragment referred to Rodentia, is the oldest mammalian assemblage known from eastern Europe. Based on preliminary analysis, the fauna may be a Clarkforkian equivalent (Gheerbrant et al. 1999). Another small assemblage of mammals recently reported from Spain may also be a Clarkforkian equivalent (Lopez-Martinez and Pelaez-Campomanes 1998).

## THE PALEOCENE-EOCENE BOUNDARY

The Paleocene–Eocene boundary has long been controversial because of the paucity of biostratigraphic sections spanning the boundary and ambiguity in original definition of the nonmarine Paleocene in conjunction with the marine Eocene sections. The Paleocene Epoch was named by Schimper (1874) for a distinctive terrestrial flora that held plants then considered to have both a Cretaceous and an Eocene aspect. Schimper’s Paleocene floras came from rocks in the Paris Basin, different strata of which

now are assigned to the Thanetian, Sparnacian, and Cuisian ages. At about the same time, Gervais (1873) reported a distinctive assemblage of terrestrial vertebrates, also then thought to exhibit Cretaceous and Eocene aspects, from the Cernay Conglomerate in the Paris Basin. Later, Gervais (1877) described more specimens from the Paris Basin including *Plesiadapis tricuspidens* and proposed a new faunal interval between those characteristic of the Cretaceous and Eocene.

Paleocene and early Eocene mammals have been studied intensively in Europe since the time of Gervais. The Cernay fauna (and Cernaysian mammal age) is accepted as late Paleocene, whereas Sparnacian faunas are generally regarded as early Eocene. Sparnacian faunas differ from those of the Paleocene in having the modern, cosmopolitan orders Perissodactyla, Artiodactyla, and Primates, along with representatives of the Hyaenodontidae. There are several problems with study of Paleocene–Eocene vertebrate faunas in Europe. Many samples come from isolated exposures, there are few thick stratigraphic sections, and deposition near sea level means that substantial hiatuses are common (Pomerol 1989; Dashzeveg 1988; Hooker 1998). These uncertainties are compounded by the fact that the Paleocene–Eocene boundary is currently placed at two different stratigraphic levels separated by approximately 1 m.y. (Berggren and Aubry 1998; Aubry et al. 1999). One level, based on micropaleontology, is the base of the Ypresian Stage (narrowly interpreted to exclude Sparnacian strata), which corresponds approximately to the NP9/NP10 (calcareous nannofossil) and P5/P6 (planktonic foraminiferal) zonal boundaries. The second is the base of the Sparnacian Stage, which corresponds to major turnover in mammalian assemblages. Although the latter is based mainly on fossil mammals, it may correlate approximately with the P4/P5 zonal boundary (Aubry et al. 1999).

The zone of uncertainty between the two stratigraphic levels apparently occurs in magnetic polarity chron C24r (see figure 1 in Berggren et al. 1995 and figure 2.2 in Berggren and Aubry 1998). A highly significant negative carbon isotope excursion is associated with “type” (“lower”) Sparnacian mammals in the Paris Basin and with global extinction or turnover of benthic foraminiferans in marine sections (Hooker 1998; Berggren and Aubry 1998; Steurbaut et al. 1999 and references therein). Documentation of a carbon isotope excursion associated with latest Clarkforkian and Wasatchian mammals (Koch et al. 1992; Bowen et al. 2001; Bains et al. 2003) is seen more clearly in Wyoming than in association with Sparnacian mammals of Europe. A carbon isotope excursion associated with earliest Bumbanian mammals has recently been identified in Asia (Bowen et al. 2002; Ting et al. 2003).

Assuming that these various carbon isotope excursions represent the same event recognizable on a global scale, they provide a geochemical means of correlating nonmarine and marine rocks between continents and marine basins.

The North American record of vertebrate change across the Paleocene–Eocene boundary is more continuous and more complete than in Europe. Indeed, the former may eventually hold a reference nonmarine global stratotype section and point (GSSP) for the series boundary (Lucas 1998). In the Clark's Fork and northern Bighorn basins of Wyoming, a stratigraphic section about 2300 m thick spans much of late Paleocene (Tiffanian and Clarkforkian mammal ages) and early Eocene (Wasatchian) time. Recent work in that area has concentrated on Wasatchian biostratigraphy and faunal change (Gingerich 1989, 1991; Clyde and Gingerich 1998). As in Europe, *Plesiadapis* is present in North American Paleocene faunas. Rodents, tillodonts, *Haplomytus*, and *Coryphodon* appear at or near the beginning of the Clarkforkian, and perissodactyls, artiodactyls, primates (including *Cantius*), and hyaenodontids appear at the beginning of the Wasatchian. Late Paleocene mammals tend to be endemic on the northern continents. In contrast, Wasatchian mammals in North America are part of a Holarctic assemblage that appears at or near the beginning of Eocene time in Europe (Hooker 1998) and central Asia (Meng and McKenna 1998).

Therefore, the presumed boundary between Paleocene and Eocene time is sharply distinct on all three northern continents. In North America, the age of the Clarkforkian–Wasatchian faunal turnover is constrained in two ways. Paleomagnetic stratigraphy has been studied in two nearby areas by Butler et al. (1981, 1987) and Clyde et al. (1994). The early Wasatchian falls in chron C24r. Interpolation using the numerical calibration of Cande and Kent (1995) places the base of the Wasatchian near or slightly before 55.0 Ma. Carbon isotope stratigraphy has been studied by Koch et al. (1992, 1995), and the negative carbon isotope excursion interpreted as correlative with extinction of benthic foraminifera falls in the earliest parts of the Wasatchian. The beginning of the carbon isotope excursion occurs at approximately 54.95 Ma, with the event lasting 120–220 k.y. (Norris and Rohl 1999; Rohl et al. 2000). Thus major mammalian turnovers at the Cernaysian–Sparnacian and Clarkforkian–Wasatchian boundaries appear to coincide with two other major events, a benthic foraminiferal extinction and a carbon isotope excursion. Although agreement on final placement of the Paleocene–Eocene boundary remains unresolved, the

International Geological Correlation Programme (IGCP) Paleocene–Eocene boundary working group formally voted recently that the boundary will be linked to the carbon isotope excursion (Luterbacher et al. 2000).

In any event, the North American record of mammalian evolution across the Paleocene–Eocene boundary is exceptionally complete, and it corroborates the sharp distinction of Paleocene and Eocene mammalian faunas first discovered in Europe. A major dispersal of mammals occurred between Europe and North America in the latest Paleocene or earliest Eocene. Wasatchian and Sparnacian mammalian assemblages of the two continents are very similar, more so than during any other time of the Cenozoic (Savage and Russell 1983). Dispersal was aided by sea level lowstands in a time of high sea level (Haq et al. 1987; Woodburne and Swisher 1995). The route probably was via a corridor between Greenland, Spitzbergen, and adjacent land masses (McKenna 1975, 1983).

#### ASIAN PALEOCENE FAUNAS

In recent years, the number of depositional basins reported from China and Mongolia that yield Paleocene mammals has increased significantly. The total now stands at 12 in China and 2 in Mongolia (Wang et al. 1998; Ting 1998). In tandem with the increased knowledge of Paleocene mammalian assemblages from China and Mongolia, three Asian land mammal ages—the Shanghuan, Nongshanian, and Gashatan—have now been formally named and defined (Ting 1998). The names *Shanghuan* and *Nongshanian* were first proposed by Li and Ting (1983), but they were not defined. Romer (1966) proposed the term *Gashatan*, which was later defined by Szalay and McKenna (1971). In defining the Shanghuan, Nongshanian, and Gashatan, Ting (1998) uses a system similar to that used for NALMAs, in which biochronologic units are defined by successive appearances of unrelated taxa.

In contrast to those from North America, Paleocene mammalian assemblages from Asia were dominated by anagalids, pantodonts, and, to a lesser extent, mesonychids (Wang et al. 1998; see tables 1–3 and figures 4–6 in Ting 1998). Also, no species and only a few genera (*Disacus*, *Prodinoceras*, *Coryphodon*, and perhaps *Oxyaena*) are shared between North America and Asia in strata of undoubted Paleocene age (Wang et al. 1998; Ting 1998). Thus Asian Paleocene mammalian assemblages display endemism, especially before the late Paleocene. As a result, proposed correlations between Asian and North American mammal ages remain tentative.

The Shanghuan Asian land mammal age was defined by Ting (1998) to include the time between appearance of the Pantodonta, represented by *Bemalambda*, and the first appearance of the Arctostylopida, represented by *Asiostylops*. Ting (1998) also subdivided the Shanghuan into two interval zones, the *Bemalambda* and *Archaeolambda* interval zones.

Correlation between the Shanghuan mammal age and those of the North American Paleocene have been based on comparisons of stage of evolution using taxa that are largely endemic. Thus Shanghuan assemblages (*sensu* Li and Ting 1983) and the Shanghuan land mammal age as defined by Ting (1998) have been regarded variously as correlative to the Tiffanian (Savage and Russell 1983), Torrejonian (Ting 1998), Torrejonian and latter part of the Puercan (Zhou et al. 1977; Li and Ting 1983; Sloan 1987; Mateer and Chen 1992; Russell et al. 1993), Puercan and all but the latter part of the Torrejonian (Wang et al. 1998), and Puercan (Lucas and Williamson 1995).

Paleomagnetic correlations and radiometric dating techniques are helpful in assessing correlations between Asia and North America. Paleomagnetic data from the Nanxiong Basin suggest that the Shanghuan spans the latter part of polarity chron C29r through C27r (Zhao et al. 1991). Also, paleomagnetic data from the Shanyang Basin indicate that *Bemalambda* occurs in rocks of reversed polarity correlated with chron C27r (Xue et al. 1994, 1996). Magnetic polarity correlations in North America indicate that the interval from the latter part of C29r through C27r spans the Puercan and most of the Torrejonian. Also, magnetic polarity chron C27r would be approximately correlative to the T02 interval zone (see figure 3.2). The only radiometric date associated with rocks yielding Paleocene mammals in China comes from the Xinzhou Basin. There, intrusive basaltic rocks thought to be younger than beds bearing Paleocene mammals yield a date of  $61.63 \pm 0.92$  (Wang et al. 1998), which is consistent with correlation to chron C27r or the Torrejonian mammal age (figure 3.2; see also Berggren et al. 1995; Cande and Kent 1995).

Lucas and Williamson (1995) argue that the sudden appearance in North America, the lack of older North American close relatives, and the presence of older, more primitive Shanghuan relatives in China supports an Asian origin and immigration of Paleocene mammalian groups. Specifically, they suggest dispersal to North America of Carnivora, Mesonychia, Pantodonta, and Tillodontia at about the beginning of Torrejonian time. However, the presence of an earlier and possibly more plesiomorphic carnivoran (*Ravenictis krausei*) from the Puercan of Saskatchewan (Fox and Youzwshyn 1994) might suggest a North American origin for the Carnivora.

The Nongshanian Asian land mammal age was defined by Ting (1998) to include the time between the first appearance of the Arctostylopida, represented by *Asiostylops*, and the first appearance of Rodentia, represented by *Tribosphenomys*. Ting (1998) subdivided the Nongshanian into the *Asiostylops* and *Sinostylops* interval zones. Dinocerata and the families Phenacolphidae and Ernanodontidae (Edentata?) made their first appearance in Asia during the Nongshanian (Ting 1998).

Paleomagnetic and radiometric data are not available from strata that yield mammalian assemblages assigned to the Nongshanian mammal age. Correlations between Asia and North America for the Nongshanian are very uncertain, therefore, because they are based on comparisons between faunas that are largely endemic. First occurrences of the orders Arctostylopida and Dinocerata in both the Nongshanian and the Tiffanian (T<sub>15</sub> lineage zone) nevertheless suggest correlation between the Tiffanian and the Nongshanian (Ting 1998). A strong sea level lowstand in the medial Tiffanian (Woodburne and Swisher 1995) may have contributed to the presumed exchange of these taxa between Asia and North America. Beard and Dawson (1999) correlated the Nongshanian with the late Torrejonian and the first half of the Tiffanian. Similarly, Wang et al. (1998) correlated the Nongshanian with the late Torrejonian through middle Tiffanian (T<sub>03</sub>–T<sub>14</sub>). They based their correlation on the conclusion by Cifelli et al. (1989) that the Nongshanian arctostylopids (*Bothriostylops* and *Sinostylops*) are more primitive than the Tiffanian (T<sub>15</sub>) *Arctostylops*. Also, *Ernanodon*, the supposed edentate from China, known from both the Nongshanian and Gashatan mammal ages, could suggest a faunal tie to South America, presumably via North America during the middle to late Paleocene.

The Gashatan Asian land mammal age was defined by Ting (1998) to include the time between the appearance of Rodentia, represented by *Tribosphenomys*, and the first appearance of Perissodactyla, represented by *Orientolophus*. In contrast to the Shanghuan and Nongshanian mammal ages, subdivision of the Gashatan mammal age into interval zones was not proposed.

Archibald et al. (1987) used the first occurrence of Rodentia in North America to define the beginning of the Clarkforkian mammal age, a definition that we follow in this update. Tillodontia (minus *Deltatherium*) and *Coryphodon* also apparently occurred synchronously with Rodentia in the earliest Clarkforkian (Archibald et al. 1987), and these first occurrences in North America may reflect dispersals from Asia across Beringia or the Euroamerican corridor (Krause and Maas 1990; Woodburne and Swisher 1995).

Ting (1998) correlated the Gashatan with the Clarkforkian mammal age in North America based mainly on the first occurrences of Rodentia and the pantodont family Coryphodontidae in both the Gashatan and Clarkforkian ages. In contrast, Wang et al. (1998) tentatively correlated the Gashatan with the late Tiffanian through Clarkforkian based on similarities between Asian and North American arctostylopids and dinoceratans. Beard (1998) and Beard and Dawson (1999) argued that dispersal of mammals rarely coincides precisely with their phylogenetic origin. They reasoned further that the beginnings of the Gashatan and Bumbanian mammal ages probably are older than the Clarkforkian and Wasatchian, respectively. Recognition of a carbon isotope excursion in the earliest Bumbanian confirms that the Gashatan–Bumbanian boundary coincides closely with both the Clarkforkian–Wasatchian and Paleocene–Eocene boundaries (Bowen et al. 2002; Ting et al. 2003).

#### AFRICAN PALEOCENE FAUNAS

The only known mammalian faunas of Paleocene age from all of Africa are from the Ouarzazate and Ouled Abdoun basins of Morocco. Mammals from the Ouarzazate Basin were first reported by Cappetta et al. (1978). The assemblage from the Ouarzazate Basin is diverse, including palaeoryctids, adapisoriculids, ?creodonts, carnivores, condylarths, todralestids, and an omomyid primate (Sige et al. 1990; Sudre et al. 1993; Gheerbrant 1991, 1992, 1994, 1995). Fossils of these taxa were recovered primarily from two localities in the Jbel Guersif Formation: Adrar Mgorn 1 and Ihadjamene (Gheerbrant et al. 1993). The Jbel Guersif Formation is thought to be Thanetian based on chondrichthyans (Cappetta et al. 1987). Thanetian is considered late Paleocene in age (Berggren et al. 1995). Paleomagnetic study of the Jbel Guersif Formation in the vicinity of Adrar Mgorn 1 indicates that the site occurs in rocks of reversed polarity that reasonably could be correlated either to magnetic polarity chron C25r or C24r (Gheerbrant et al. 1998b). Compared with correlations of magnetic polarity chrons with mammal ages in North America, Adrar Mgorn 1 would correlate with late phases of the Tiffanian if C25r is correct or Clarkforkian if C24r is correct.

Before the late 1990s, the phosphates of the Ouled Abdoun Basin yielded two mammalian specimens, both of which were identified as *Phosphatherium*, interpreted to be the oldest known proboscidean (Gheerbrant et al. 1996, 1998a). The locality from which the specimens were derived was unknown, but analysis of matrix from the specimens confirmed its phosphatic nature. Foraminifera and

tiny chondrichthyian teeth from the matrix also indicated a Thanetian age (Gheerbrant et al. 1996, 1998a). However, additional specimens of *Phosphatherium* were recently collected from strata in the Ouled Abdoun Basin that have been reported as Ypresian or early Eocene in age (Gheerbrant et al. 2001).

Paleocene faunal comparisons between North America and Africa are limited by endemism, but there is some relationship at the familial level (palaeoryctids), and the genera *Cimolestes* and *Palaeoryctes* are found on both continents (Gheerbrant et al. 1998b). In North America, *Cimolestes* is known from the Lancian and Puercan, and *Palaeoryctes* is known from the mid-Torreonian to late Clarkforkian. The antiquity of cimolestids in North America suggests a dispersal event between North America and Africa, probably via Europe, in the early Paleocene (Gheerbrant 1990).

#### SOUTH AMERICAN PALEOCENE FAUNAS

Paleocene mammalian assemblages from South America are large, diverse, and dominated by endemic types of marsupials, leptictids, notoungulates, litopterns, and condylarths, along with lesser numbers of trigonostylopoids, astrapotheres, xenungulates, and xenarthrans. One species each of an enigmatic multituberculate-like gondwanathere (Krause et al. 1997; Pascual et al. 1999), a pantodont (de Muizon and Marshall 1992), and a monotreme (Pascual et al. 1992) also are present. The South American Paleocene fauna is distinctly different from those of probable Campanian–Maastrichtian age (the Los Alamos and La Colonia local faunas of Argentina; see Bonaparte 1990; Pascual et al. 2000). These Late Cretaceous mammalian faunas are composed of a variety of nontherian mammals, a faunal assemblage dubbed the “Gondwanan Stage” by Pascual (see Pascual et al. 2000 and references therein). During an interval after the last records of the “Gondwanan Stage,” which has been estimated to be approximately 10 m.y. (Pascual 1998), most of these Gondwanan nontherian lineages became extinct. For the most part, South American Paleocene mammalian faunas are made up of immigrants or descendants of immigrants from Laurasian continents (Luo et al. 2001).

Paleocene mammalian assemblages are known from four main areas of South America: Laguna Umayo, southeastern Peru; the Andean Basin of northwest Argentina and southern Bolivia (includes Tiupampa and Tres Cruces); Itaborai, southeastern Brazil; and the San Jorge Basin of southern Argentina (includes the Rio Chico and Punta Peligro faunas). Marshall and Sempere (1993) re-



viewed the Cenozoic land mammal record of South America and recognized three South American land mammal ages: the Tiupampian (early but not earliest Paleocene), including the faunas from Laguna Umayo and Tiupampa; the Itaboraian (middle? Paleocene to early Eocene?), including the fissure fill faunas from Itaborai; and the Riochican (late Paleocene), including faunas from the Rio Chico Formation. Marshall and Sempere (1993) also noted the lack of geochronologic control for the Itaborai faunas, which Savage and Russell (1983) included in the Riochican age.

More recently, Marshall et al. (1997) proposed a refined zonation for Paleocene land mammal faunas of South America, suggested that all these faunas are of late Paleocene age, and identified two mid- to late Paleocene interchange events between North and South America. Their refined zonation and new correlation were based on revised calibration of the Paleocene mammalian local faunas from the southern Andean Basin using regional stratigraphy and magnetostratigraphy (Sempere et al. 1997) along with reassessment of the ages of local faunas from the San Jorge Basin and Itaborai. Marshall et al. (1997) recognized a single Riochican South America land mammal age (spanning 60.0–55.5 Ma) with four subages. Listed chronologically, they are the Peligran (based on sparse faunas from Tres Cruces in the Andean Basin and Punta Peligro in the San Jorge Basin), Tiupampian, Itaboraian, and Riochican (*sensu stricto*). Marshall et al. (1997) did not propose a definition or characterization of their Riochican age or any of its four subages. This reassessment by Marshall et al. (1997), which suggests that no known mammalian assemblage from South America predates 60 Ma, is a radical departure. Earlier interpretations of faunal data from Tiupampa suggested an age of approximately 64–63 Ma, if correlations to North American faunas are accurate (Van Valen 1988; Williamson 1996).

To provide a structure for evaluating these conflicting interpretations, we will discuss South American Paleocene mammal faunas and their correlation to NALMAs in the context of their four main geographic areas (Laguna Umayo, Andean Basin, Itaborai, and San Jorge Basin). The new calibration of Paleocene mammalian faunas from the Andean Basin, reassessment of the age of other local faunas, and the refined zonation for Paleocene land mammal faunas of South America proposed by Sempere et al. (1997) and Marshall et al. (1997) will be addressed where appropriate.

The Laguna Umayo Local Fauna described by Sige (1972) is from a red bed succession (Umayo Formation) exposed near Lake Titicaca in southeastern Peru. More

recently, Crochet and Sige (1993) reported another mammalian assemblage from the Umayo Formation that was recovered about 200 m stratigraphically higher than the Laguna Umayo Local Fauna. The magnetostratigraphic section of the Umayo Formation is entirely of reversed polarity (Sige et al. in prep., cited in Marshall et al. 1997), so correlation to the GPTS is difficult. Various correlations to magnetic polarity chrons (C29r, C28r, C27r, or C26r) are possible, and egg shells of dinosaurs or large ground birds are associated with the mammals (see Marshall et al. 1997 for discussion). Therefore reasonable age assignments for the mammalian assemblages from the Umayo Formation based on available data vary from Late Cretaceous to late Paleocene.

The Andean Basin of northwestern Argentina and southern Bolivia yields the Tiupampa Local Fauna, one of the best-known Paleocene mammalian assemblages in South America. The Tiupampa Local Fauna from the Santa Lucia Formation of south-central Bolivia includes a large and diverse assemblage of mammals. It includes several groups of marsupials and members of the placental orders Pantodonta, Leptictida, Condylarthra, and Nontoungulata (de Muizon and Marshall 1992; Marshall et al. 1997). This fauna was originally assigned a Late Cretaceous age, but later it was considered to represent the early Paleocene (de Muizon and Marshall 1992; Marshall and Sempere 1993 and references therein).

The Tiupampa Local Fauna includes two mioclaenids, *Tiuclaenus* and *Molinodus*, along with the only known South American pantodont, *Alcidedorbignya*. The mioclaenids represent a distinct stock (Williamson 1996) and have been assigned to an endemic subfamily, Kollpaniinae (de Muizon and Cifelli 2000). Currently, the oldest records of mioclaenids in North America (five genera) are from the Pu2 interval zone. The contemporaneous appearances of such diverse genera strongly suggests an earlier radiation of the group in an as yet unsampled area outside the Western Interior of North America.

The South American pantodont *Alcidedorbignya* may be more primitive than the earliest known North American pantodonts. The presence of primitive molar characters in several Asian taxa and their absence in North and South American pantodonts points to an Asiatic origin for the group (de Muizon and Marshall 1992). Also, de Muizon and Marshall (1992) suggested that presence of *Alcidedorbignya* in South America in the early Paleocene requires a Maastrichtian (Late Cretaceous) or older, Asia to South America, via North America, dispersal route for pantodonts. However, the earliest North American pantodont is *Pantolambda intermedium* from the To2 interval zone in the San Juan Basin (Williamson 1996).

Therefore presence of *Alcidedorbignya*, as well as *Tiupampius* and *Molinodus*, in the Tiupampa Local Fauna suggests a pre-Torrejonian age for dispersal of primitive ungulates from North America to South America and a dispersal of Pantodonta to South America from Asia predating the San Juan Basin occurrence (To2) in North America. Similarly, based on comparison of the fauna of placental mammals from Tiupampa with those from the Paleocene of North America, Van Valen (1988) correlated the Tiupampa Local Fauna to the late Puercan or early Torrejonian (approximately 64–63 Ma).

Recent stratigraphic and magnetostratigraphic correlations proposed by Sempere et al. (1997) indicate that the section of the Santa Lucia Formation containing the Tiupampa Local Fauna is of reversed polarity, which they correlate to magnetic polarity chron C26r or about 59 Ma. Based on this interpretation, the Tiupampa Local Fauna would be about mid-Tiffanian in age and thus much younger than previously thought. However, the magnetostratigraphic correlation proposed by Sempere et al. (1997) for the Tiupampa Local Fauna depends heavily on the accuracy of their correlation of the magnetostratigraphic sequence at La Palca (which includes the El Molino Formation and the overlying Santa Lucia Formation) to the GPTS. It is important to note that the La Palca magnetostratigraphic section is complicated and records many geomagnetic polarity reversals (see figure 10 in Sempere et al. 1997). A single radioisotopic date ( $72.1 \pm 0.5$  Ma) from the lower part of the El Molino Formation is used to correlate the lower part of the paleomagnetic section to the GPTS (Sempere et al. 1997). Radioisotopic data are not available for the Santa Lucia Formation. With this degree of uncertainty in correlation to the GPTS, more than one interpretation is plausible. Thus the reversed polarity section of the Santa Lucia Formation at Tiupampa might correlate with magnetic polarity chron C28r or perhaps C27r. Either chron would be more in agreement with the faunal correlations proposed by Van Valen (1988) and Williamson (1996).

At Sao Jose de Itaborai, inland from Rio De Janeiro, Brazil, a large and diverse assemblage of mammals was collected from marl fills in karst cavities that developed in limestones of the Sao Jose de Itaborai Formation (Savage and Russell 1983 and references therein). Based on the Itaborai fauna, an Itaboraian mammal age was proposed by Paula Couto (1952), but it was later placed in the Riochican mammal age (Savage and Russell 1983). Recently, Marshall et al. (1997) proposed that karst formation and infilling at Itaborai coincided with the large sea level lowstand between 58.5 and 56.5 Ma recognized by Haq et al. (1987). This sea level lowstand would cor-

relate approximately with the late Paleocene (late Tiffanian) interval of dispersal advocated by Gingerich (1985), in which representatives of the Dinocerata, Notoungulata (represented by Arctostylopidae), and Edentata dispersed from South America to North America. Because of its fissure fill provenance, the mammalian assemblage from Itaborai remains unconstrained geochronologically.

The last area of South America known to yield Paleocene mammals is the San Jorge Basin, located in southern Argentina. There, a succession of mudstone and sandstone beds, which yields a diverse and endemic assemblage of mammals, is referred to the Rio Chico Formation. The Rio Chico Formation probably is no older than late Paleocene. It is superposed on the marine Salamanca Formation, which has been correlated with the Danu-Montian Stage of Europe based on foraminiferans (Loeblich and Tappan 1957; Savage and Russell 1983). The age of the Itaborai fauna was approximately correlated with mammalian assemblages from the Rio Chico Formation. Together, they were considered to represent the Riochican mammal age of late Paleocene age (Simpson 1940; Savage and Russell 1983).

The mammalian assemblages from the Rio Chico Formation occur near the Atlantic coast, and Marshall et al. (1997) attempted to correlate lithologic changes with transgressive–regressive cycles proposed by Haq et al. (1987). Marshall et al. (1997) identified several guide levels in the detailed sections provided by Simpson (1935b) and Feruglio (1949). Combining these guide levels and lithologic changes with paleomagnetic data from Cerro Redondo (Marshall et al. 1981) and other stratigraphic sections, Marshall et al. (1997) developed a tentative correlation of mammal-bearing sections of the Rio Chico Formation (see figure 6 and table 2 in Marshall et al. 1997). They were able to distinguish at least four distinct ages of mammalian assemblages in the Rio Chico Formation. The oldest assemblage, containing South America's only known occurrence of Monotremata, *Monotrematum* (Pascual et al. 1992), comes from the Banco Negro Inferior at Punta Peligro, which Marshall et al. (1997) dated at approximately 60 Ma. The fauna from the Banco Negro Inferior and that from Tres Cruces in the Andean Basin form the Peligran subage (Marshall et al. 1997). The lower mammal-bearing levels at Bajo de la Palangana and Cerro Redondo are interpreted to have been deposited about 59 Ma. They are considered approximate equivalents of the Tiupampa fauna from the Andean Basin. These three faunas form the Tiupampian subage (Marshall et al. 1997). The remainder of the mammalian assemblages from the Rio Chico Formation are referred to the Itaboraian or Ri-

ochican subages, interpreted to range from 58.0 to 56.0 Ma (Marshall et al. 1997).

In summary, based on similarity of faunas, there appear to have been two distinct intervals of dispersal between North and South America. The older occurred in the early Paleocene, perhaps during the late Puercan or early Torrejonian. Faunal data from Tiupampa and Puercan and early Torrejonian sites in North America suggest a connection between the continents in the early Paleocene (de Muizon and Marshall 1992; Van Valen 1988; Williamson 1996). However, recent biostratigraphic and geochronologic correlations proposed by Sempere et al. (1997) and Marshall et al. (1997) suggest that the Tiupampa Local Fauna is significantly younger than the early Paleocene. The second interval of dispersal was during the late Paleocene, probably in the late Tiffanian (Gingerich 1985; Marshall et al. 1997). Unlike the earlier interval, proposed geochronologic correlations between the Americas indicate that the late Tiffanian faunas of North America are approximate time equivalents of the Itaboraian and Riochican subages of the Riochican mammal age of Marshall et al. (1997) or the Riochican mammal age of Savage and Russell (1983).

### SUGGESTIONS FOR FUTURE RESEARCH

With the publications by Wood et al. (1941) and Archibald et al. (1987), North American land mammal ages have become widely accepted biochronologic units. However, this does not mean that these mammal ages could ever replace a detailed chronostratigraphic framework for continental strata deposited in North America during the Paleocene; definitions and characterizations of mammal ages and zones still rely heavily on faunal data. Is a truly chronostratigraphic framework of stages based on tighter stratigraphic control of mammalian faunas possible for the North American Paleocene? Yes, but this will entail continued emphasis on developing detailed biostratigraphic and stratigraphic fieldwork coupled with reanalysis of previously published biostratigraphic data from a number of well-studied basins in western North America. That is, we must continue to gather additional faunal data from local sections with determinable superpositional relationships, interpret the data in terms of biogeographic provinces, and then correlate to other faunal provinces based on comparison of detailed faunal successions. Research in each area must be followed by refinement of correlation within and be-

tween provinces through use of magnetostratigraphy. Finally, temporal calibration using radioisotopes must be applied if possible. Further refinement of the Puercan through Clarkforkian mammal ages and development of greater solidity of the chronostratigraphic framework is desirable and should be a high priority for future research.

To aid in developing a firmer chronostratigraphic framework, in concert with additional biostratigraphic data, sampling rocks associated with mammalian faunas for magnetostratigraphic and radioisotopic analysis should be emphasized. Only one Lancian to early Torrejonian sequence of rocks (encompassing the upper Hell Creek and Tullock formations in eastern Montana) has undergone all three (biostratigraphic, magnetostratigraphic, and radioisotopic) primary types of analyses (Swisher et al. 1993; Clemens 2002). In fact, the radioisotopic dates from Montana are among the few associated with Paleocene mammalian faunas in North America. Granted, the paucity of available data is partially a factor of the rarity in many Paleocene basins of rocks suitable for isotopic analysis. But some areas known to have Paleocene mammals and volcanic units suitable for dating remain unsampled. For the time being, magnetostratigraphy remains a primary method of providing independent checks on faunal correlation for mammal-bearing sections throughout western North America. Wherever possible, additional paleomagnetic data should be obtained from mammal-bearing rocks.

Finally, the available record of Paleocene mammalian faunas is geographically concentrated in northern and mid-continental Rocky Mountain states, from the San Juan Basin of New Mexico north to Alberta, Canada. Significant faunal provinciality is evident in comparisons between mammalian assemblages from New Mexico and Montana to southern Canada (for example, see the discussion of Pu2 and Pu3 interval zones). Wyoming shows varying intermediacies, with greater affinities to the south during the Puercan and greater northward affinities during the late Torrejonian and early Tiffanian. Therefore additional faunal data are needed throughout all latitudes of the North American continent to fully assess temporally shifting patterns of faunal distribution. Although Paleocene nonmarine rocks that yield mammals are known from southwest Texas and southern California, faunal data available from these areas are sparse. Also, Mexico has yet to yield Paleocene mammals. But because Cretaceous and Eocene mammals are known from that country, it is only a matter of time until Paleocene mammals are discovered. The same can be said for Alaska because nonmarine strata of proba-

ble Paleocene age are exposed along the banks of some of the major rivers on its North Slope.

## ACKNOWLEDGMENTS

We thank E. McGee for sharing her computer database on Puercan through Early Arikareean mammalian faunas, which aided in getting our update started; M. Woodburne for comments on the manuscript and assistance in gathering references; M. Lauria for constructing figures 3.1 and 3.2; A. Raser for help with table 3.2; and the following grants from the National Science Foundation (EAR-9506462, EAR-9909354, EAR-0125502, and DGE-9253953) and the National Geographic Society (6736-00).

## REFERENCES

- Alroy, J. 1999. The fossil record of North American mammals: Evidence for a Paleocene evolutionary radiation. *Systematic Biology* 48:107–118.
- . 2000. New methods for quantifying macroevolutionary patterns and processes. *Paleobiology* 26:707–734.
- Alroy, J., P. L. Koch, and J. C. Zachos. 2000. Global climate change and North American mammalian evolution. *Paleobiology* 26 (suppl. 4):259–288.
- Archibald, J. D. 1982. A study of Mammalia and geology across the Cretaceous–Tertiary boundary in Garfield County, Montana. *University of California Publications in Geological Sciences* 122:1–286.
- . 1987a. The Bugcreekian land mammal age: A reassessment. *Journal of Vertebrate Paleontology* 7(suppl. 3):10A.
- . 1987b. Latest Cretaceous and Early Tertiary mammalian biochronology/biostratigraphy in the Western Interior. *Geological Society of America Abstracts with Programs* 19:258.
- Archibald, J. D., R. F. Butler, E. H. Lindsay, W. A. Clemens, and L. Dingus. 1982. Upper Cretaceous–Paleocene biostratigraphy and magnetostratigraphy, Hell Creek and Tullock formations, north-eastern Montana. *Geology* 10:153–159.
- Archibald, J. D., W. A. Clemens, P. D. Gingerich, D. W. Krause, E. H. Lindsay, and K. D. Rose. 1987. First North American land mammal ages of the Cenozoic era. In *Cenozoic mammals of North America*, ed. M. O. Woodburne. Berkeley: University of California Press. pp. 24–76.
- Archibald, J. D., and D. L. Lofgren. 1990. Mammalian zonation near the Cretaceous–Tertiary boundary. In *Dawn of the age of mammals in the northern part of the Rocky Mountain Interior, North America*, ed. T. M. Bown and K. D. Rose. Geological Society of America Special Paper 243:31–50.
- Archibald, J. D., J. K. Rigby Jr., and S. F. Robison. 1983. Systematic revision of *Oxyacodon* (Condylarthra, Periptychidae) and a description of *O. ferronensis* n. sp. *Journal of Paleontology* 57:53–72.
- Arens, N. C. and A. H. Jahren. 1999. Chemostratigraphic recognition of the Cretaceous–Tertiary boundary in North Dakota: Precise stratigraphic correlation of fossil-bearing sections. *Geological Society of America Abstracts with Programs* 30:71–72.
- Arens, N. C., A. H. Jahren, and R. Amundson. 2000. Can C<sub>3</sub> plants faithfully record the carbon isotopic composition of atmospheric carbon dioxide? *Paleobiology* 26:137–164.
- Aubry, J. P., W. A. Berggren, J. A. Van Couvering, and F. Steininger. 1999. Problems in chronostratigraphy: Stages, series, unit and boundary stratotypes, global stratotype section and point and tarnished golden spikes. *Earth Science Reviews* 46:99–148.
- Bains, S., R. D. Norris, R. M. Corfield, G. J. Bowen, P. D. Gingerich, and P. L. Koch. 2003. Marine–terrestrial linkages at the Paleocene–Eocene boundary. In *Causes and consequences of globally warm climates in the early Paleogene*, ed. S. L. Wing, P. D. Gingerich, E. Thomas, and B. Schmitz. Geological Society of America Special Paper 369:1–9.
- Beard, K. C. 1998. East of Eden: Asia as an important center of taxonomic origination in mammalian evolution. In *Dawn of the age of mammals in Asia*, ed. K. C. Beard and M. R. Dawson. *Bulletin of the Carnegie Museum of Natural History* 34:5–39.
- Beard, K. C. and M. R. Dawson. 1999. Intercontinental dispersal of Holarctic land mammals near the Paleocene/Eocene boundary: Paleogeographic, paleoclimatic and biostratigraphic implications. *Bulletin de la Société Géologique de France* 170:697–706.
- Berggren, W. A. and M.-P. Aubry. 1998. The Paleocene/Eocene epoch/series boundary: Chronostratigraphic framework and estimated geochronology. In *Late Paleocene–Early Eocene climatic and biotic evolution*, ed. M.-P. Aubry, S. Lucas, and W. A. Berggren. New York: Columbia University Press, pp. 18–36.
- Berggren, W. A., D. V. Kent, C. C. Swisher III, and M.-P. Aubry. 1995. A revised Cenozoic geochronology and chronostratigraphy. In *Geochronology, time scales and global stratigraphic correlation*, ed. W. A. Berggren, D. V. Kent, M.-P. Aubry, and J. Hardenbol. SEPM Special Publication 54:129–212.
- Bloch, J. I., D. M. Boyer, P. D. Gingerich, and G. F. Gunnell. 2002. New primitive paromomyid from the Clarkforkian of Wyoming and dental eruption in Plesiadapiformes. *Journal of Vertebrate Paleontology* 22:366–379.
- Bloch, J. I., D. C. Fisher, K. D. Rose, and P. D. Gingerich. 2001. Stratocladistic analysis of Paleocene Carpolestidae (Mammalia, Plesiadapiformes) with a description of a new Late Tiffanian genus. *Journal of Vertebrate Paleontology* 21:119–131.
- Bonaparte, J. F. 1990. New Late Cretaceous mammals from the Los Alamos Formation, northern Patagonia. *National Geographic Research* 6:63–93.
- Bowen, G. J., W. C. Clyde, P. L. Koch, S. Ting, J. Alroy, T. Tsubamoto, Y. Wang, and Y. Wang. 2002. Mammalian dispersal at the Paleocene/Eocene boundary. *Science* 295:2062–2065.
- Bowen, G. J., P. L. Koch, P. D. Gingerich, R. D. Norris, S. Bains, and R. M. Corfield. 2001. Refined isotope stratigraphy across the continental Paleocene–Eocene boundary on Pocat Bench in the northern Bighorn Basin. In *Paleocene–Eocene stratigraphy and biotic change in the Bighorn and Clark's Fork basins, Wyoming*, ed. P. D. Gingerich. *University of Michigan Papers on Paleontology* 33:73–88.
- Buckley, G. A. 1994. *Paleontology, geology, and chronostratigraphy of Simpson Quarry (early Paleocene), Bear Formation, Crazy Mountains Basin, south central Montana*. Ph.D. dissertation, Rutgers University, New Brunswick, NJ.
- . 1995. The multituberculate *Catopsalis* from the early Paleocene of the Crazy Mountain Basin in Montana. *Acta Palaeontologica Polonica* 40:389–398.
- . 1997. A new species of *Purgatorius* (Mammalia, Primatomorpha) from the Lower Paleocene Bear Formation, Crazy Mountains Basin, south central Montana. *Journal of Paleontology* 71:149–155.
- Butler, R. F., P. D. Gingerich, and E. H. Lindsay. 1981. Magnetic polarity stratigraphy and biostratigraphy of Paleocene and lower

- Eocene continental deposits, Clark's Fork Basin, Wyoming. *Journal of Geology* 89:299–316.
- Butler, R. F., D. W. Krause, and P. D. Gingerich. 1987. Magnetic polarity stratigraphy and biostratigraphy of middle-late Paleocene continental deposits of south-central Montana. *Journal of Geology* 95:647–657.
- Butler, R. F. and E. H. Lindsay. 1985. Mineralogy of magnetic minerals and revised magnetic polarity stratigraphy of continental sediments, San Juan Basin, New Mexico. *Journal of Geology* 94:535–554.
- Butler, R. F., E. H. Lindsay, and P. D. Gingerich. 1980. Magnetic polarity stratigraphy and Paleocene–Eocene biostratigraphy of Polecat Bench, northwestern Wyoming. In *Early Cenozoic paleontology and stratigraphy of the Bighorn Basin, Wyoming, 1880–1980*, ed. P. D. Gingerich. *University of Michigan Papers on Paleontology* 24:95–98.
- Cande, S. C. and D. V. Kent. 1995. Revised calibration of the geomagnetic polarity timescale for the Late Cretaceous and Cenozoic. *Journal of Geophysical Research* 100:6093–6095.
- Cappetta, H., J.-J. Jaeger, M. Sabatier, J. Sudre, and M. Vianey-Liaud. 1978. Découverte dans le Paléocène du Maroc des plus anciens mammifères eutheriens d'Afrique. *Géobios* 11:257–263.
- Cappetta, H., J.-J. Jaeger, B. Sige, J. Sudre, and M. Vianey-Liaud. 1987. Compléments et précisions biostratigraphiques sur la faune Paléocène à Mammifères et Sélaciens du bassin d'Ouarzazate (Maroc). *Tertiary Research* 8:147–157.
- Cifelli, R. L., N. J. Czaplewski, and K. D. Rose. 1995. Additions to knowledge of Paleocene mammals from the North Horn Formation, central Utah. *Great Basin Naturalist* 55:304–314.
- Cifelli, R. L., R. L. Nydam, J. G. Eaton, J. D. Gardner, and J. I. Kirkland. 1999. Vertebrate faunas of the North Horn Formation (Upper Cretaceous–Lower Paleocene), Emery and San Pete counties, Utah. In *Vertebrate paleontology in Utah*, ed. D. D. Gillette. Miscellaneous Publication 99-1 Utah Geological Survey, pp. 377–388.
- Cifelli, R. L., C. R. Schaff, and M. C. McKenna. 1989. The relationships of Arctostylopidae (Mammalia): New data and interpretation. *Bulletin of the Museum of Comparative Biology* 152:1–44.
- Clemens, W. A. 1974. *Purgatorius*, an early paromomyid primate (Mammalia). *Science* 184:903–905.
- . 2002. Evolution of the mammalian fauna across the Cretaceous–Tertiary boundary in northeastern Montana and other areas of the Western Interior. In *Hell Creek Formation and the Cretaceous–Tertiary boundary in the northern Great Plains: An integrated continental record of the end of the Cretaceous*, ed. J. H. Hartman, K. R. Johnson, and D. J. Nichols. Geological Society of America Special Paper 361:217–245.
- Clyde, W. C. and P. D. Gingerich. 1998. Mammalian community response to the latest Paleocene thermal maximum: An isotaphonomic study in the northern Bighorn Basin, Wyoming. *Geology* 26:1011–1014.
- Clyde, W. C., J. Stamatakos, and P. D. Gingerich. 1994. Chronology of the Wasatchian land-mammal age (early Eocene): Magnetostratigraphic results from the McCullough Peaks section, northern Bighorn Basin, Wyoming. *Journal of Geology* 102:367–377.
- Cope, E. D. 1875. Report on the geology of that part of northwestern New Mexico examined during the field season of 1874. In *Geographical Surveys West of the 100th Meridian* and Appendix LL, *Annual Report of the Chief of Engineers for 1875*, pp. 61–97 of separate issue; pp. 981–1017 of full report.
- . 1884. The Vertebrata of the Tertiary formations of the West. Book I. *Report of the U.S. Geological Survey of the Territories*, F. V. Hayden in charge, 3:1–1009.
- . 1888. Synopsis of the vertebrate fauna of the Puerco series. *Transactions of the American Philosophical Society* 16:298–361.
- Crochet, J.-Y. and B. Sige. 1993. Les mammifères de Chulpas (Formation Umayo, transition Crétacé–Tertiaire, Pérou), données préliminaires. *Documents du Laboratoire de Géologie de Lyon* 125:97–107.
- Dashzeveg, D. 1988. Holarctic correlation of non-marine Palaeocene–Eocene boundary strata using mammals. *Journal of the Geological Society of London* 145:473–478.
- Dawson, M. R. and K. C. Beard. 1996. New late Paleocene rodents (Mammalia) from the Big Multi Quarry, Washakie Basin, Wyoming. *Palaeovertebrata* 25:301–321.
- Dawson, M. R., C.-K. Li, and T. Qi. 1984. Eocene ctenodactyloid rodents (Mammalia) of eastern and central Asia. *Carnegie Museum of Natural History Special Publication* 9:138–150.
- Dorr, J. A. Jr. 1952. Early Cenozoic stratigraphy and vertebrate paleontology of the Hoback Basin, Wyoming. *Bulletin of the Geological Society of America* 63:59–94.
- . 1958. Early Cenozoic vertebrate paleontology, sedimentation and orogeny in central western Wyoming. *Bulletin of the Geological Society of America* 69:1217–1244.
- . 1978. Revised and amended fossil vertebrate faunal lists, early Tertiary, Hoback Basin, Wyoming. *Contributions to Geology, University of Wyoming* 16:79–84.
- Dorr, J. A. Jr. and P. D. Gingerich. 1980. Early Cenozoic mammalian paleontology, geologic structure, and tectonic history in the overthrust belt near LaBarge, western Wyoming. *Contributions to Geology, University of Wyoming* 18:101–115.
- Dorr, J. A. Jr., D. R. Spearing, and J. R. Steidtmann. 1977. Deformation and deposition between a foreland uplift and an impinging thrust belt, Hoback Basin, Wyoming. *Geological Society of America Special Paper* 177:1–82.
- Dorr, J. A. Jr. and J. R. Steidtmann. 1970. Stratigraphic–tectonic implications of a new, earliest Eocene, mammalian faunule from central western Wyoming. *Michigan Academician* 3:25–41.
- Eberle, J. J. 1999. Bridging the transition between didelphodonts and taeniodonts. *Journal of Paleontology* 73:936–944.
- Eberle, J. J. and J. A. Lillegraven. 1998a. A new important record of earliest Cenozoic mammalian history: Eutheria, and paleogeographic/biostratigraphic summaries. *Rocky Mountain Geology* 33:49–117.
- . 1998b. A new important record of earliest Cenozoic mammalian history: Geologic setting, Multituberculata, and Peradectia. *Rocky Mountain Geology* 33:3–47.
- Erickson, B. R. 1991. Flora and fauna of the Wannagan Creek Quarry: Late Paleocene of North America. *Science Publications of the Science Museum of Minnesota* 7:1–19.
- . 1999. Fossil Lake Wannagan (Paleocene: Tiffanian) Billings County, North Dakota. *North Dakota Geological Survey Miscellaneous Publication* 87:1–9.
- Feruglio, E. 1949. Descripción geológica de la Patagonia. *Buenos Aires, Yacimientos Petrolíferos Fiscales* 2:1–349.
- Fox, R. C. 1983. Notes on the North American Tertiary marsupials *Herpotherium* and *Peradectes*. *Canadian Journal of Earth Sciences* 20:1565–1578.
- . 1984a. The definition and relationships of the Paleocene primate *Micromomys* Szalay, with description of a new species. *Canadian Journal of Earth Sciences* 21:1262–1267.
- . 1984b. First North American record of the Paleocene primate *Saxonella*. *Journal of Paleontology* 58:892–894.

- . 1984c. *Melaniella timosa* n. gen. and n. sp.: An unusual mammal from the Paleocene of Alberta, Canada. *Canadian Journal of Earth Sciences* 21:1335–1338.
- . 1984d. A new species of the Paleocene primate *Elphidotarsius* Gidley: Its stratigraphic position and evolutionary relationships. *Canadian Journal of Earth Sciences* 21:1268–1277.
- . 1988. Late Cretaceous and Paleocene mammal localities of southern Alberta. *Occasional Paper of the Tyrrell Museum of Paleontology* 6:1–38.
- . 1989. The Wounded Knee Local Fauna and mammalian evolution near the Cretaceous–Tertiary boundary, Saskatchewan, Canada. *Palaeontographica Abt. A* 208:11–59.
- . 1990a. *Pronothodectes gaoi* n. sp. from the Late Paleocene of Alberta, Canada, and the early evolution of the Plesiadapidae (Mammalia, Primates). *Journal of Paleontology* 64:637–647.
- . 1990b. *Saxonella* (Plesiadapiformes: ?Primates) in North America: *S. naylori*, sp. nov., from the Late Paleocene of Alberta, Canada. *Journal of Vertebrate Paleontology* 11:334–349.
- . 1990c. The succession of Paleocene mammals in western Canada. In *Dawn of the age of mammals in the northern part of the Rocky Mountain Interior of North America*, ed. T. M. Bown and K. D. Rose. *Geological Society of America Special Paper* 243:51–70.
- . 1995. “Paleocene aspect” mammals from the Late Cretaceous of Alberta and Saskatchewan. *Geological Society of America Abstracts with Programs* 27:11.
- . 1997. Late Cretaceous and Paleocene mammals, Cypress Hills region, Saskatchewan, and mammalian evolution across the Cretaceous–Tertiary boundary. In *Upper Cretaceous and Tertiary stratigraphy and paleontology of southern Saskatchewan*, ed. L. McKenzie-McAnally. *Canadian Paleontology Conference Field Trip Guidebook* 6:70–85.
- Fox, R. C. and G. P. Youzwyshyn. 1994. New primitive carnivorans (Mammalia) from the Paleocene of western Canada, and their bearing on relationships of the order. *Journal of Vertebrate Paleontology* 14:382–404.
- Gardner, J. H. 1910. The Puerco and Torrejon formations of the Nacimiento Group. *Journal of Geology* 18:702–741.
- Gazin, C. L. 1938. A Paleocene mammalian fauna from central Utah. *Journal of the Washington Academy of Sciences* 28:271–277.
- . 1939. A further contribution to the Dragon Paleocene fauna of central Utah. *Journal of the Washington Academy of Sciences* 29:273–286.
- . 1941. The mammalian faunas of the Paleocene of central Utah, with notes on the geology. *Proceedings of the U.S. National Museum* 91:1–53.
- . 1942. Fossil Mammalia from the Almy Formation in western Wyoming. *Journal of the Washington Academy of Sciences* 32:217–220.
- . 1956a. The occurrence of fossil mammalian remains in the Fossil Basin of southwestern Wyoming. *Journal of Paleontology* 30:707–711.
- . 1956b. Paleocene mammalian faunas of the Bison Basin in south-central Wyoming. *Smithsonian Miscellaneous Collections* 131(6):1–57.
- . 1956c. The upper Paleocene Mammalia from the Almy Formation in western Wyoming. *Smithsonian Miscellaneous Collections* 131(7):1–18.
- . 1969. A new occurrence of Paleocene mammals in the Evanston Formation of southwest Wyoming. *Smithsonian Contributions to Paleobiology* 2:1–17.
- . 1971. Paleocene primates from the Shotgun member of the Fort Union Formation in Wind River Basin, Wyoming. *Proceedings of the Biological Society of Washington* 84:13–38.
- Gervais, P. 1873. Enumeration de quelques ossements d’animaux vertebres recueillis aux environs de Reims par M. Lemoine. *Journal de Zoologie Paris* 2:351–355.
- . 1877. Enumeration de quelques ossements d’animaux vertebres recueillis aux environs de Reims par M. Lemoine. *Journal de Zoologie Paris* 6:74–79.
- Gheerbrant, E., 1990. On the early biogeographical history of African placentals. *Historical Biology* 4:107–116.
- . 1991. *Todralestes variabilis* n. g., n. sp., nouveau Proteutherian (Eutheria, Todralestidae fam. nov.) du Paleocene du Maroc. *Comptes Rendus Académie des Sciences Paris* 312:1249–1255.
- . 1992. Les mammiferes paleocenes du Bassin d’Ouarzazate (Maroc). I. Introduction generale et Palaeoryctidae. *Palaeontographica Abt. A* 224:67–132.
- . 1994. Les Mammiferes paleocenes du Bassin d’Ouarzazate (Maroc). II. Todralestidae (Proteutheria, Eutheria). *Palaeontographica Abt. A* 231:133–188.
- . 1995. Les Mammiferes Paleocenes du Bassin D’Ouarzazate (Maroc). III. Adapisoriculidae et autres mammiferes (Carnivora, ?Creodonta, Condylarthra, ?Ungulata et incertae sedis). *Palaeontographica Abt. A* 237:39–132.
- Gheerbrant, E. H., C. Abrial, and G. Cuenca. 1997. Nouveau mammifere condylarthre du Paleocene superieur de campo (Pyrenees aragonaises, Espagne). *Comptes Rendus Académie des Sciences Paris* 243 series 11a:599–606.
- Gheerbrant, E., H. Cappetta, M. Feist, J.-J. Jaeger, J. Sudre, M. Vianey-Liaud, and B. Sige. 1993. La succession des faunes de vertebres d’age paleocene superieur et eocene inferieur dans la Bassin d’Ouarzazate. Porte biostratigraphique et paleogeographique. *Newsletters on Stratigraphy* 28:33–58.
- Gheerbrant, E. H., V. Codrea, A. Hosu, S. Sen, C. Guernet, F. de Lapparent de Broin, and J. Rivelinee. 1999. Decouverte de vertebres dans les Calcaires de Rona (Thanetian ou Sparnacien), Transylvanie, Roumanie: Les plus anciens mammiferes cenozoiques d’Europe Orientale. *Eclogae Geologicae Helvetiae* 92:517–535.
- Gheerbrant, E., J. Sudre, and H. Cappetta. 1996. A Palaeocene proboscidean from Morocco. *Nature* 383: 68–70.
- Gheerbrant, E., J. Sudre, H. Cappetta, and G. Bignot. 1998a. *Phosphatherium escuilliei* du Thanetian du Bassin des Ouled Abdoun (Maroc), plus ancien proboscidean (Mammalia) d’Afrique. *Geobios* 30:247–269.
- Gheerbrant, E., J. Sudre, M. Laroche, and A. Moumni. 2001. First ascertained African “condylarth” mammals (primitive ungulates: cf. Bulbulodentata and cf. Phenacodonta) from the earliest Ypresian of the Ouled Abdoun Basin, Morocco. *Journal of Vertebrate Paleontology* 21:107–118.
- Gheerbrant, E., J. Sudre, S. Sen, C. Abrial, B. Marandat, B. Sige, and M. Vianey-Liaud. 1998b. Nouvelles donnees sur les mammiferes du Thanetian et de l’Ypresien du Bassin d’Ouarzazate (Maroc) et leur contexte stratigraphique. *Palaeovertebrata* 27:155–202.
- Gingerich, P. D. 1976. Cranial anatomy and evolution of early Tertiary Plesiadapidae (Mammalia, Primates). *University of Michigan Papers on Paleontology* 15:1–141.
- . 1980a. Evolutionary patterns in early Cenozoic mammals. *Annual Review of Earth and Planetary Sciences* 8:407–424.
- . 1980b. *Tytthaena parrisi*, oldest known oxyaenid (Mammalia, Creodonta) from the later Paleocene of western North America. *Journal of Paleontology* 54:570–576.

- . 1983. New Adapisoricidae, Pentacodontidae, and Hypsodontidae (Mammalia, Insectivora, and Condylarthra) from the late Paleocene of Wyoming and Colorado. *Contributions from the Museum of Paleontology of the University of Michigan* 26:227–255.
- . 1985. South American mammals in the Paleocene of North America. In *The great American biotic interchange*, ed. F. G. Stehli and S. D. Webb. New York: Plenum, pp. 123–137.
- . 1986. Systematic position of *Litomylus* (?) *alphamon* Van Valen (Mammalia: Insectivora): Further evidence for the late Paleocene age of Swan Hills Site-1 in the Paskapoo Formation of Alberta. *Journal of Paleontology* 60:1153–1137.
- . 1987. Early Eocene bats (Mammalia, Chiroptera) and other vertebrates in freshwater limestones of the Willwood Formation, Clarks Fork Basin, Wyoming. *Contributions from the Museum of Paleontology of the University of Michigan* 27:275–320.
- . 1989. New earliest Wasatchian mammalian fauna from the Eocene of northwestern Wyoming: Composition and diversity in a rarely sampled high-floodplain assemblage. *University of Michigan Papers on Paleontology* 28:1–97.
- . 1991. Systematics and evolution of early Eocene Perissodactyla (Mammalia) in the Clark's Fork Basin, Wyoming. *Contributions from the Museum of Paleontology of the University of Michigan* 28:181–213.
- . 2000. Paleocene–Eocene boundary and continental vertebrate faunas of Europe and North America. In *Early Paleogene warm climates and biosphere dynamics*, ed. B. Schmitz, B. Sundquist, and F. P. Andreasson. Uppsala: Geological Society of Sweden, pp. 57–59.
- . 2001. Biostratigraphy of the continental Paleocene–Eocene boundary interval on Polecat Bench in the northern Bighorn Basin. In *Paleocene–Eocene stratigraphy and biotic change in the Bighorn and Clark's Fork basins, Wyoming*, ed. P. D. Gingerich. *University of Michigan Papers on Paleontology* 33: 37–71.
- . 2003. Mammalian responses to climate change at the Paleocene–Eocene boundary: Polecat Bench record in the northern Bighorn Basin, Wyoming. In *Causes and consequences of globally warm climates in the early Paleogene*, ed. S. L. Wing, P. D. Gingerich, E. Thomas, and B. Schmitz. Geological Society of America Special Paper 369:463–478.
- Gingerich, P. D. and G. F. Gunnell. 1979. Systematics and evolution of the genus *Esthonyx* (Mammalia, Tillodontia) in the early Eocene of North America. *Contributions from the Museum of Paleontology of the University of Michigan* 25:125–153.
- Gingerich, P. D., P. Houde, and D. W. Krause. 1983. A new earliest Tiffanian (late Paleocene) mammalian fauna from Bangtail Plateau, western Crazy Mountain Basin, Montana. *Journal of Paleontology* 57:957–970.
- Gingerich, P. D. and K. D. Rose. 1977. Preliminary report on the American Clark Fork mammal fauna, and its correlation with similar faunas in Europe and Asia. *Géobios, Mémoire Spécial* 1:39–45.
- . 1979. Anterior dentition of the Eocene condylarth *Thryptacodon*: Convergence with the tooth comb of lemurs. *Journal of Mammalogy* 50:16–22.
- Gingerich, P. D., K. D. Rose, and D. W. Krause. 1980. Early Cenozoic mammalian faunas of the Clark's Fork Basin–Polecat Bench area, northwestern Wyoming. In *Early Cenozoic paleontology and stratigraphy of the Bighorn Basin, Wyoming*, ed. P. D. Gingerich. *University of Michigan Papers on Paleontology* 24:51–64.
- Gingerich, P. D. and D. A. Winkler. 1985. Systematics of Paleocene Viverravidae (Mammalia, Carnivora) in the Bighorn Basin and Clark's Fork Basin, Wyoming. *Contributions from the Museum of Paleontology of the University of Michigan* 27:87–128.
- Granger, W. 1914. On the names of lower Eocene faunal horizons of Wyoming and New Mexico. *Bulletin of the American Museum of Natural History* 33:201–207.
- . 1915. A revision of the lower Wasatch and Wind River faunas. Pt. III. Order Condylarthra, families Phenacodontidae and Meniscotheriidae. *Bulletin of the American Museum of Natural History* 34:329–361.
- . 1917. Notes on Paleocene and lower Eocene mammal horizons of northern New Mexico and southern Colorado. *Bulletin of the American Museum of Natural History* 37:821–830.
- Gunnell, G. F. 1988. New species of *Unuchinia* (Mammalia, Insectivora) from the middle Paleocene of North America. *Journal of Paleontology* 62:139–141.
- . 1989. Evolutionary history of Microsypoidea (Mammalia, ?Primates) and the relationship between Plesiadapiformes and Primates. *University of Michigan Papers on Paleontology* 27:1–157.
- . 1994. Paleocene mammals and faunal analysis of the Chappo type locality (Tiffanian), Green River Basin, Wyoming. *Journal of Vertebrate Paleontology* 14:81–104.
- Gunnell, G. F. and P. D. Gingerich. 1991. Systematics and evolution of late Paleocene and early Eocene Oxyaenidae (Mammalia, Creodonta) in the Clark's Fork Basin, Wyoming. *Contributions from the Museum of Paleontology of the University of Michigan* 28:141–180.
- Haq, B. U., J. Hardenbol, and P. R. Vail. 1987. Chronology of fluctuating sea-levels since the Triassic. *Science* 35:1156–1166.
- Hartman, J. E. 1986. Paleontology and biostratigraphy of lower part of Polecat Bench Formation, southern Bighorn Basin, Wyoming. *Contributions to Geology University of Wyoming* 24:11–63.
- Hartman, J. H. 1999. Western exploration along the Missouri River and the first paleontological studies in the Williston Basin, North Dakota and Montana. *Proceedings of the North Dakota Academy of Science* 53:158–165.
- Hartman, J. H., G. A. Buckley, D. W. Krause, and T. J. Kroeger. 1989. Paleontology, stratigraphy, and sedimentology of Simpson Quarry (early Paleocene), Crazy Mountains Basin, South-central Montana. *Montana Geological Society Guidebook, Geological Resources of Montana* 1:173–185.
- Hartman, J. H. and A. J. Kihm. 1991. Stratigraphic distribution of *Titanooides* (Mammalia: Pantodonta) in the Fort Union Group (Paleocene) of North Dakota. In *Sixth International Williston Basin Symposium*, ed. J. E. Christopher and F. M. Haidl. *Special Publication Saskatchewan Geological Society* 11:207–215.
- . 1992. Chronostratigraphy of Paleocene strata in the Williston Basin. In *Geology and utilization of lignites*, ed. R. B. Finkelman, D. J. Daly, and S. J. Tewalt. Reston, VA: Environmental and Coal Associates, pp. 52–75.
- . 1995. Age of Meek and Hayden's Fort Union Group (Paleocene), upper Missouri River, North Dakota–Montana. In *Seventh International Williston Basin Symposium*, ed. L. D. V. Hunter and R. A. Schalla. *Montana Geological Society Guidebook*, pp. 417–428.
- . 1999. The discovery and preliminary record of North Dakota Paleocene mammals from the Lloyd and Hares localities. *Proceedings of the North Dakota Academy of Science* 53:149–154.
- Hartman, J. H. and D. W. Krause. 1993. Cretaceous and Paleocene stratigraphy and paleontology of the Shawmut anticline and the Crazy Mountains Basin, Montana: Road log and overview of re-

- cent investigations. *Montana Geological Society Guidebook, Energy and Mineral Resources of Central Montana*, pp. 71–84.
- Hedberg, H. D., ed. 1976. *International stratigraphic guide*. New York: Wiley.
- Higgins, P. 2000. *Re-evaluation of the boundary between the Torrejonian and Tiffanian North American land mammal "ages" with description of a new fauna from the Hanna Basin, south-central Wyoming*. Ph.D. dissertation, University of Wyoming, Laramie.
- Holtzman, R. C. 1978. Late Paleocene mammals of the Tongue River Formation, western North Dakota. *North Dakota Geological Survey Report of Investigations* 65:1–88.
- Hooker, J. J. 1998. Mammalian faunal change across the Paleocene–Eocene transition in Europe. In *Late Paleocene–Early Eocene climatic and biotic evolution*, ed. M. P. Aubry, S. Lucas, and W. A. Berggren. New York: Columbia University Press, pp. 428–450.
- Hunter, J. P. 1999. The radiation of Paleocene mammals with the demise of the dinosaurs: Evidence from southwestern North Dakota. *Proceedings of the North Dakota Academy of Science* 53:141–144.
- Hunter, J. P., J. H. Hartman, and D. W. Krause. 1997. Mammals and mollusks across the Cretaceous–Tertiary boundary from Makoshika State Park and vicinity (Williston Basin), Montana. *Contributions to Geology, University of Wyoming* 32:61–114.
- Jepsen, G. L. 1930. Stratigraphy and paleontology of the Paleocene of northeastern Park County, Wyoming. *Proceedings of the American Philosophical Society* 69:463–528.
- . 1937. A Paleocene rodent, *Paramys atavus*. *Proceedings of the American Philosophical Society* 78:291–301.
- . 1940. Paleocene faunas of the Polecat Bench Formation, Park County, Wyoming. Pt. I. *Proceedings of the American Philosophical Society* 83:217–341.
- Johnston, P. A. 1980. First record of Mesozoic mammals from Saskatchewan. *Canadian Journal of Earth Sciences* 17:512–519.
- Johnston, P. A. and R. C. Fox. 1984. Paleocene and late Cretaceous mammals from Saskatchewan, Canada. *Palaeontographica Abt. A* 186:166–222.
- Keefer, W. R. 1961. Waltman Shale and Shotgun members of Fort Union Formation (Paleocene) in Wind River Basin, Wyoming. *American Association of Petroleum Geologists Bulletin* 45:1310–1323.
- Kihm, A. J. 1984. *Early Eocene mammalian faunas of the Piceance Creek Basin, northwestern Colorado*. Ph.D. dissertation, University of Colorado, Boulder.
- Koch, P. L., J. C. Zachos, and D. L. Dettman. 1995. Stable isotope stratigraphy and paleoclimatology of the Paleogene Bighorn Basin (Wyoming, USA). *Palaeogeography, Palaeoclimatology, Palaeoecology* 115:61–89.
- Koch, P. L., J. C. Zachos, and P. D. Gingerich. 1992. Correlation between isotope records in marine and continental carbon reservoirs near the Palaeocene–Eocene boundary. *Nature* 358:319–322.
- Krause, D. W. 1977. Paleocene multituberculates (Mammalia) of the Roche Percée Local Fauna, Ravenscrag Formation, Saskatchewan, Canada. *Palaeontographica Abt. A* 159:1–36.
- . 1978. Paleocene primates from western Canada. *Canadian Journal of Earth Sciences* 15:1250–1271.
- . 1987a. *Baiotomeus*, a new ptilodontid multituberculata (Mammalia) from the middle Paleocene of western North America. *Journal of Paleontology* 61:595–603.
- . 1987b. Systematic revision of the genus *Prochetodon* (Ptilodontidae, Multituberculata) from the late Paleocene and early Eocene of western North America. *Contributions from the Museum of Paleontology of the University of Michigan* 27:221–236.
- Krause, D. W. and P. D. Gingerich. 1983. Mammalian fauna from Douglass Quarry, earliest Tiffanian (late Paleocene) of the eastern Crazy Mountain Basin, Montana. *Contributions from the Museum of Paleontology of the University of Michigan* 26:157–196.
- Krause, D. W. and M. C. Maas. 1990. The biogeographic origins of late Paleocene–early Eocene mammalian immigrants to the Western Interior of North America. In *Dawn of the age of mammals in the northern part of the Rocky Mountain Interior*, ed. T. M. Bown and K. D. Rose. *Geological Society of America Special Paper* 243:71–106.
- Krause, D. W., G. V. R. Prasad, W. von Koenigswald, A. Sahni, and F. E. Grine. 1997. Cosmopolitanism among Gondwanan Late Cretaceous mammals. *Nature* 390:504–507.
- Krishtalka, L. 1973. Late Paleocene mammals from the Cypress Hills, Alberta. *Special Publications, The Museum of Texas Tech University* 2:1–77.
- . 1976a. Early Tertiary Adapisoricidae and Erinaceidae (Mammalia, Insectivora) of North America. *Bulletin of the Carnegie Museum of Natural History* 1:1–40.
- . 1976b. North American Nyctitheriidae (Mammalia, Insectivora). *Annals of the Carnegie Museum of Natural History* 46:7–28.
- Krishtalka, L., C. C. Black, and D. W. Riedel. 1975. Paleontology and geology of the Badwater Creek area, central Wyoming. *Annals of the Carnegie Museum of Natural History* 45:179–212.
- Leite, M. B., 1992. *Vertebrate biostratigraphy and taphonomy of the Fort Union Formation (Paleocene) east of Grass Creek Basin, southwestern Bighorn Basin, Wyoming*. Ph.D. dissertation, University of Wyoming, Laramie.
- Li, C.-K. and S.-Y. Ting. 1983. The Paleocene mammals of China. *Bulletin of the Carnegie Museum of Natural History* 21:1–93.
- Lillegraven, J. A. and J. J. Eberle. 1999. Vertebrate faunal changes through Lancian and Puercan time in southern Wyoming. *Journal of Paleontology* 73:689–708.
- Lillegraven, J. A. and A. W. Snoke. 1996. A new look at the Laramide orogeny in the Seminoe and Shirley mountains, Freezout Hills, and Hanna Basin, south-central Wyoming. *Geological Survey of Wyoming Public Information Circular* 36:1–56.
- Lindsay, E. H., R. F. Butler, and N. M. Johnson. 1981. Magnetic polarity zonation and biostratigraphy of late Cretaceous and Paleocene continental deposits, San Juan Basin, New Mexico. *American Journal of Science* 281:390–435.
- Loeblich, A. R. and H. Tappan. 1957. Correlation of the Gulf and Atlantic coastal plain Paleocene and lower Eocene formations by means of planktonic Foraminifera. *Journal of Paleontology* 31:1109–1137.
- Lofgren, D. L. 1995. The Bug Creek problem and the Cretaceous–Tertiary boundary at McGuire Creek, Montana. *University of California Publications in Geological Sciences* 140:1–185.
- Lofgren, D. L., M. C. McKenna, and S. L. Walsh. 1999. New records of Torrejonian–Tiffanian mammals from the Paleocene–Eocene Goler Formation, California. *Journal of Vertebrate Paleontology* 19(3):60A.
- Lopez-Martinez, N. and P. Pelaez-Campomanes. 1998. Late Paleocene mammals from the Tremp Formation (southern Pyrenees, Lleida, Spain). *Strata* 1(9):79–82.
- Lucas, S. G. 1984. Early Paleocene vertebrates, stratigraphy, and biostratigraphy, West Fork of Gallegos Canyon, San Juan Basin, New Mexico. *New Mexico Geology* 6:56–60.



- . 1998. Fossil mammals and the Paleocene/Eocene series boundary in Europe, North America, and Asia. In *Late Paleocene–Early Eocene climatic and biotic evolution*, ed. M. P. Aubry, S. Lucas, and W. A. Berggren. New York: Columbia University Press, pp. 451–500.
- Lucas, S. G. and T. E. Williamson. 1995. Systematic position and biochronologic significance of *Yuodon* and *Palasiodon*, supposed Paleocene “condylarths” from China. *Neues Jahrbuch für Geologie und Paläontologie Abhandlungen* 196:93–107.
- Lucas, S. G., T. E. Williamson, and M. D. Middleton. 1997. *Catopsalis* (Mammalia: Multituberculata) from the Paleocene of New Mexico and Utah. *Journal of Paleontology* 71:484–493.
- Luo, Z. 1991. Variability of dental morphology and relationships of the earliest arctocyonid species. *Journal of Vertebrate Paleontology* 11:452–471.
- Luo, Z., R. L. Cifelli, and Z. Kielan-Jaworowska. 2001. Dual origin of tribosphenic mammals. *Nature* 409:53–57.
- Lupton, C. D., D. Gabriel, and R. W. West. 1980. Paleobiology and depositional setting of a Late Cretaceous vertebrate locality, Hell Creek Formation, McCone County, Montana. *Contributions to Geology University of Wyoming* 18:117–126.
- Luterbacher, H., J. A. Hardenbol, and B. Schmitz. 2000. Decision of the voting members on the International Subcommittee on Paleogene stratigraphy on the criterion for the recognition of the Paleocene–Eocene boundary. *Newsletter of the International Subcommittee on Paleogene Stratigraphy* 9:13.
- Maas, M. C., M. R. L. Anthony, P. D. Gingerich, G. F. Gunnell, and D. W. Krause. 1995. Mammalian generic diversity and turnover in the late Paleocene and early Eocene of the Bighorn and Crazy Mountains basins, Wyoming and Montana. *Palaeogeography, Palaeoclimatology, Palaeoecology* 115:181–207.
- Maas, M. C. and D. W. Krause. 1994. Mammalian turnover and community structure in the Paleocene of North America. *Historical Biology* 8:91–128.
- MacDonald, T. E. 1995. Late Paleocene (Tiffanian) mammalian local faunas in superposition from near Drumheller, Alberta. *Journal of Vertebrate Paleontology* 15(3):41A.
- MacIntyre, G. T. 1966. The Miacidae (Mammalia, Carnivora) Pt. 1: The systematics of *Ictidopappus* and *Protictis*. *Bulletin of the American Museum of Natural History* 131:115–210.
- Marshall, L. G., R. F. Butler, R. E. Drake, and G. H. Curtis. 1981. Calibration of the beginning of the age of mammals in Patagonia. *Science* 212:43–45.
- Marshall, L. G. and T. Sempere. 1993. Evolution of the Neotropical Cenozoic land mammal fauna in its geochronologic, stratigraphic, and tectonic context. In *Biological relationships between Africa and South America*, ed. P. Goldblatt. New Haven, CT: Yale University Press, pp. 329–392.
- Marshall, L. G., T. Sempere, and R. F. Butler. 1997. Chronostratigraphy of the mammal-bearing Paleocene of South America. *Journal of the South American Earth Sciences* 10:49–70.
- Mateer, N. J. and P. Chen. 1992. A review of the nonmarine Cretaceous–Tertiary transition in China. *Cretaceous Research* 13:81–90.
- Matthew, W. D. 1897. A revision of the Puerco fauna. *Bulletin of the American Museum of Natural History* 9:259–323.
- . 1915a. A revision of the lower Eocene Wasatch and Wind River faunas. Pt. I: Order Ferae (Carnivora), suborder Creodonta. *Bulletin of the American Museum of Natural History* 34:4–103.
- . 1915b. A revision of the lower Eocene Wasatch and Wind River faunas. Pt. II: Order Condylarthra, family Hypsodontidae. *Bulletin of the American Museum of Natural History* 34:311–328.
- . 1915c. A revision of the lower Eocene Wasatch and Wind River faunas. Pt. IV: Entelonychia, Primates, Insectivora (part). *Bulletin of the American Museum of Natural History* 34:429–483.
- . 1937. Paleocene faunas of the San Juan Basin, New Mexico. *Transactions of the American Philosophical Society* 30:1–510.
- McKenna, M. C. 1955. Paleocene mammal, Goler Formation, Mojave Desert, California. *American Association of Petroleum Geologists Bulletin* 39:512–515.
- . 1960. A continental Paleocene vertebrate fauna from California. *American Museum Novitates* 2024:1–20.
- . 1972. Vertebrate paleontology of the Togwotee Pass area, northwestern Wyoming. In *Guidebook, field conference on tertiary biostratigraphy of southern and western Wyoming*, ed. R. M. West. Privately printed, pp. 80–101.
- . 1975. Fossil mammals and the early Eocene North Atlantic land continuity. *Annals of the Missouri Botanical Garden* 62:335–353.
- . 1980. Late Cretaceous and early Tertiary vertebrate paleontological reconnaissance, Togwotee Pass area, northwestern Wyoming. In *Aspects of vertebrate history: Essays in honor of Edwin Harris Colbert*, ed. L. L. Jacobs. Flagstaff: Museum of Northern Arizona Press, pp. 321–343.
- . 1983. Holarctic landmass rearrangement, cosmic events, and Cenozoic terrestrial organisms. *Annals of the Missouri Botanical Garden* 70:459–489.
- McKenna, M. C. and S. K. Bell. 1997. *Classification of mammals above the species level*. New York: Columbia University Press.
- McKenna, M. C., J. H. Hutchison, and J. H. Hartman. 1987. Paleocene vertebrates and nonmarine Mollusca from the Goler Formation, California. In *Basin analysis and paleontology of the Paleocene and Eocene Goler Formation, El Paso Mountains, California*, ed. B. C. Cox. SEPM Pacific Section Publication 57:31–41.
- McKenna, M. C. and D. L. Lofgren. In press. *Mimotricentes tedfordi*, a new arctocyonid from the Paleocene of California. *American Museum Novitates*.
- Meng, J. and M. C. McKenna. 1998. Faunal turnover of Palaeogene mammals from the Mongolian Plateau. *Nature* 394:364–367.
- Middleton, M. D. 1982. A new species and additional material of *Catopsalis* (Mammalia, Multituberculata) from the Western Interior of North America. *Journal of Paleontology* 56:1197–1206.
- . 1983. *Early Paleocene vertebrates of the Denver Basin, Colorado*. Ph.D. dissertation, University of Colorado, Boulder.
- Muizon, C. de and R. L. Cifelli. 2000. The “condylarths” (archaic Ungulata, Mammalia) from the early Palaeocene of Tiupampa (Bolivia): implications on the origin of South American ungulates. *Geodiversitas* 22:47–150.
- Muizon, C. de and L. G. Marshall. 1992. *Alcidedorbignya inopinata* (Mammalia, Pantodonta) from the Early Paleocene of Bolivia: Phylogenetic and paleobiogeographic implications. *Journal of Paleontology* 66:499–520.
- NACSN. 1983. North American Commission on Stratigraphic Nomenclature. *American Association of Petroleum Geologists Bulletin* 67:841–875.
- Norris, R. D. and U. Rohl. 1999. Carbon cycling and chronology of climate warming during the Palaeocene–Eocene transition. *Nature* 401:775–778.
- Novacek, M. J. 1977. A review of Paleocene and Eocene Leptictidae (Eutheria: Mammalia) from North America. *Paleobios* 24:1–42.

- Novacek, M. J. and W. A. Clemens. 1977. Aspects of intra-generic variation and evolution of *Mesodma* (Multituberculata, Mammalia). *Journal of Paleontology* 51:701–717.
- Osborn, H. F. 1929. The titanotheres of ancient Wyoming, Dakota, and Nebraska. *U.S. Geological Survey Monographs* 55:1–701.
- Osborn, H. F. and C. Earle. 1895. Fossil mammals of the Puerco beds: Collection of 1892. *Bulletin of the American Museum of Natural History* 7:1–70.
- Osborn, H. F. and W. D. Matthew. 1909. Cenozoic mammal horizons of western North America. *Bulletin of the U.S. Geological Survey* 361:1–138.
- Pascual, R. 1998. The history of South American land mammals: The seminal Cretaceous–Tertiary transition. *Asociacion Paleontologia Argentina, Publication Especial*, pp. 5–18.
- Pascual, R., M. Archer, E. O. Juareguizar, J. L. Prado, H. Godthelp, and S. J. Hand. 1992. First discovery of monotremes in South America. *Nature* 356:704–706.
- Pascual, R., F. J. Goin, P. Gonzales, A. Ardolino, and P. F. Puerta. 2000. A highly derived docodont from the Patagonian Late Cretaceous: Evolutionary implications for Gondwanan mammals. *Geodiversitas* 22:395–414.
- Pascual, R. F. J. Goin, D. W. Krause, E. Ortiz-Jaureguizar, and A. A. Carlini. 1999. The first gnathic remains of *Sudamerica*: Implications for gondwanan relationships. *Journal of Vertebrate Paleontology* 19:373–382.
- Patterson, B. 1933. A new species of the ambylopod *Titanoides* from western Colorado. *American Journal of Science* 25:415–425.
- . 1936. Mounted skeleton of *Titanoides* with notes on the associated fauna. *Proceedings of the Geological Society of America* 1935:397–398.
- . 1937. A new genus, *Barylambda*, for *Titanoides faberi*, Paleocene ambylopod. *Geological Series of the Field Museum of Natural History* 6:229–231.
- . 1939. New Pantodontia and Dinocerata from the upper Paleocene of western Colorado. *Geological Series of the Field Museum of Natural History* 6:351–384.
- . 1949. A new genus of taeniodonts from the late Paleocene. *Fieldiana: Geology* 10:41–42.
- Patterson, B. and P. O. McGrew. 1962. A new arctocyonid from the Paleocene of Wyoming. *Breviora* 174:1–10.
- Patterson, B. and E. L. Simons. 1958. A new barylambdid pantodont from the late Paleocene. *Breviora* 93:1–8.
- Patterson, B. and R. M. West. 1973. A new late Paleocene phenacodont (Mammalia, Condylarthra) from western Colorado. *Breviora* 403:1–7.
- Paula Couto, C. de. 1952. Fossil mammals from the beginning of the Cenozoic in Brazil. *Bulletin of the American Museum of Natural History* 99:359–394.
- Pomerol, C. 1989. Stratigraphy of the Palaeogene: Hiatus and transitions. *Proceedings of the Geologists' Association* 100:313–324.
- Rapp, S. D., B. J. MacFadden, and J. A. Schiebout. 1983. Magnetic polarity stratigraphy of the early Tertiary Black Peaks Formation: Big Bend National Park, Texas. *Journal of Geology* 91:555–572.
- Rigby, J. K. Jr. 1980. Swain Quarry of the Fort Union Formation, middle Paleocene (Torrejonian), Carbon County, Wyoming: Geologic setting and mammalian fauna. *Evolutionary Monographs* 3:1–179.
- . 1987. The last of the North American dinosaurs. In *Dinosaurs past and present*, Vol. II, ed. S. Czerkas and E. Olsen. Seattle: University of Washington Press, pp. 119–153.
- . 1989. The Cretaceous–Tertiary boundary of the Bug Creek drainage: Hell Creek and Tullock formations, McCone and Garfield counties, Montana. In *Mesozoic/Cenozoic vertebrate paleontology: Classic localities, contemporary approaches*, ed. J. J. Flynn. *American Geophysical Union Field Trip Guidebook* T322:67–73.
- Rigby, J. K. Jr., K. R. Newmann, J. Smit, S. Van der Kaars, R. E. Sloan, and J. K. Rigby. 1987. Dinosaurs from the Paleocene part of the Hell Creek Formation, McCone County, Montana. *Palaaios* 2:296–302.
- Robinson, L. N. and J. G. Honey. 1987. Geologic setting of a new Paleocene mammal locality in the northern Powder River Basin, Montana. *Palaaios* 2:87–90.
- Robison, S. F. 1986. Paleocene (Puercan–Torrejonian) mammalian faunas of the North Horn Formation, central Utah. *Brigham Young University Geology Studies* 33:87–133.
- Rohl, U., T. J. Bralower, R. D. Norris, and G. Wefer. 2000. New chronology for the late Paleocene thermal maximum and its environmental implications. *Geology* 28:927–930.
- Romer, A. S. 1966. *Vertebrate paleontology*. Chicago: University of Chicago Press.
- Rose, K. D. 1975. The Carpolestidae: Early Tertiary primates from North America. *Bulletin of the Museum of Comparative Zoology* 147:1–74.
- . 1977. Evolution of carpolestid primates and chronology of the North American middle and late Paleocene. *Journal of Paleontology* 51:536–542.
- . 1978. A new Paleocene epiocotheriid (Mammalia), with comments on the Palaeodonta. *Journal of Paleontology* 52:658–674.
- . 1980. Clarkforkian land-mammal age: Revised definition, zonation, and tentative intercontinental correlation. *Science* 208:744–746.
- . 1981a. The Clarkforkian land-mammal age and mammalian faunal composition across the Paleocene–Eocene boundary. *University of Michigan Papers on Paleontology* 26:1–197.
- . 1981b. Composition and species diversity in Paleocene and Eocene mammal assemblages: An empirical study. *Journal of Vertebrate Paleontology* 1:367–388.
- Rose, K. D. and T. M. Bown. 1982. New plesiadapiform primates from the Eocene of Wyoming and Montana. *Journal of Vertebrate Paleontology* 2:63–69.
- Rose, K. D. and D. W. Krause. 1982. Cyriacotheriidae: A new family of early Tertiary pantodonts from western North America. *Proceedings of the American Philosophical Society* 126:26–50.
- Rose, K. D. and S. G. Lucas. 2000. An early Paleocene palaeodont (Mammalia, ?Philodota) from New Mexico, and the origin of the Palaeodonta. *Journal of Vertebrate Paleontology* 20:139–156.
- Russell, D. A., D. E. Russell, and A. R. Sweet. 1993. The end of the dinosaurian era in the Nanxiong Basin. *Vertebrata Palasiatica* 31:139–145.
- Russell, D. E. 1967. Le Paléocène continental d'Amerique du Nord. *Mémoires du Musée National d'Histoire Naturelle, Série C* 16:1–99.
- Russell, D. E., J.-L. Hartenberger, C. Pomerol, S. Sen, N. Schmidt-Kittler, and M. Vianey-Liaud. 1982. Mammals and stratigraphy: The Paleocene of Europe. *Palaeoverm. Mem. Ext.* pp. 1–77.
- Russell, L. S. 1929. Paleocene vertebrates from Alberta. *American Journal of Science* 217:162–178.
- . 1958. Paleocene mammal teeth from Alberta. *Bulletin of the National Museum of Canada* 147:96–103.

- . 1967. Palaeontology of the Swan Hills area, north-central Alberta. *Royal Ontario Museum, Life Sciences Contribution* 71:1–31.
- Savage, D. E. 1962. Cenozoic geochronology of the fossil mammals of the Western Hemisphere. *Museo Argentino de Ciencias Naturales, Ciencias Zoológicas* 8:51–67.
- Savage, D. E. and D. E. Russell. 1983. *Mammalian paleofaunas of the world*. Reading, MA: Addison-Wesley.
- Schiebout, J. A. 1974. Vertebrate paleontology and paleoecology of Paleocene Black Peaks Formation, Big Bend National Park, Texas. *Texas Memorial Museum Bulletin* 24:1–88.
- Schiebout, J. A., C. A. Rigsby, S. D. Rapp, J. A. Hartnell, and B. R. Stanhardt. 1987. Stratigraphy of the Cretaceous–Tertiary and Paleocene–Eocene transition rocks of Big Bend National Park, Texas. *Journal of Geology* 95:359–371.
- Schimper, W. P. 1874. *Traité de paléontologie végétale*, Vol. 3. Paris: J. B. Bailliere, pp. 1–896.
- Schoch, R. M. 1985. Preliminary description of a new late Paleocene land-mammal fauna from South Carolina, USA. *Postilla* 196:1–13.
- . 1986. Systematics, functional morphology, and macroevolution of the extinct mammalian order Taeniodonta. *Peabody Museum of Natural History Bulletin* 42:1–307.
- . 1998. Late Paleocene land-mammals from the Williamsburg Formation (Black Mingo Group) of South Carolina. *Transactions of the American Philosophical Society* 88:229–245.
- Secord, R. 1998. Paleocene mammalian biostratigraphy of the Carbon Basin, southeastern Wyoming, and age constraints on local phases of tectonism. *Rocky Mountain Geology* 33:119–154.
- Sempere, T., R. F. Butler, D. R. Richards, L. G. Marshall, W. Sharp, and C. C. Swisher III. 1997. Stratigraphy and chronology of Upper Cretaceous–lower Paleogene strata in Bolivia and northwest Argentina. *Geological Society of America Bulletin* 109:709–727.
- Sige, B. 1972. La faunule de mammifères du Cretace supérieur de Laguna Umayo (Andes péruviennes). *Bulletin du Musée National d'Histoire Naturelle Paris* 99, *Sciences de la Terre* 19:375–409.
- Sige, B., J.-J. Jaeger, J. Sudre, and M. Vianey-Liaud. 1990. *Altitlasius koulchii* n. gen. et sp., primate Omomyidae du Paleocene supérieur du Maroc, et les origines des Euprimates. *Palaeontographica Abt. A* 214:31–56.
- Simmons, N. B. 1987. A revision of *Taeniolabis* (Mammalia, Multituberculata) with a new species from the Puercan of eastern Montana. *Journal of Paleontology* 61:794–808.
- Simons, E. L. 1960. The Paleocene Pantodonta. *Transactions of the American Philosophical Society* 50:1–99.
- Simpson, G. G. 1927. Mammalian fauna and correlation of the Paskapoo Formation of Alberta. *American Museum Novitates* 268:1–10.
- . 1928. A new mammalian fauna from the Fort Union Formation of southern Montana. *American Museum Novitates* 297:1–15.
- . 1929a. A collection of Paleocene mammals from Bear Creek, Montana. *Annals of the Carnegie Museum of Natural History* 19:115–122.
- . 1929b. A new Paleocene uinthere and molar evolution in the Amblypoda. *American Museum Novitates* 387:1–9.
- . 1929c. Third contribution to the Fort Union fauna at Bear Creek, Montana. *American Museum Novitates* 345:1–12.
- . 1932. A new Paleocene mammal from a deep well in Louisiana. *Proceedings of the U.S. National Museum* 82:1–4.
- . 1933. Glossary and correlation charts of North American Tertiary mammal-bearing formations. *Bulletin of the American Museum of Natural History* 67:79–121.
- . 1935a. New mammals from the Fort Union of Montana. *Proceedings of the U.S. National Museum* 83:221–244.
- . 1935b. Occurrence and relationships of the Rio Chico fauna of Patagonia. *American Museum Novitates* 818:1–21.
- . 1935c. The Tiffany fauna, upper Paleocene. I. Multituberculata, Marsupialia, Insectivora, and ?Chiroptera. *American Museum Novitates* 795:1–19.
- . 1935d. The Tiffany fauna, upper Paleocene. II. Structure and relationships of *Plesiadapis*. *American Museum Novitates* 816:1–30.
- . 1935e. The Tiffany fauna, upper Paleocene. III. Primates, Carnivora, Condylarthra, and Amblypoda. *American Museum Novitates* 817:1–28.
- . 1936. A new fauna from the Fort Union of Montana. *American Museum Novitates* 873:1–27.
- . 1937a. Additions to the upper Paleocene fauna of the Crazy Mountain Field. *American Museum Novitates* 940:1–15.
- . 1937b. The Fort Union of the Crazy Mountain Field, Montana, and its mammalian faunas. *U.S. National Museum Bulletin* 169:1–287.
- . 1937c. Notes on the Clark Fork, upper Paleocene, fauna. *American Museum Novitates* 954:1–24.
- . 1940. Review of the mammal-bearing Tertiary of South America. *Proceedings of the American Philosophical Society* 83:649–709.
- . 1959. Fossil mammals from the type area of the Puerco and Nacimiento strata, Paleocene of New Mexico. *American Museum Novitates* 1957:1–22.
- Sinclair, W. J. and W. Granger. 1912. Notes on the Tertiary deposits of the Bighorn Basin. *Bulletin of the American Museum of Natural History* 31:57–67.
- . 1914. Paleocene deposits of the San Juan Basin, New Mexico. *Bulletin of the American Museum of Natural History* 33:297–316.
- Sloan, R. E. 1970. Cretaceous and Paleocene terrestrial communities of western North America. *Proceedings of the North American Paleontology Convention* 1:427–453.
- . 1987. Paleocene and latest Cretaceous mammal ages, biozones, magnetozones, rates of sedimentation, and evolution. In *The Cretaceous–Tertiary boundary in the San Juan and Raton basins, New Mexico and Colorado*, ed. J. E. Fassett and J. K. Rigby Jr. Geological Society of America Special Paper 209:165–200.
- Sloan, R. E., J. K. Rigby Jr., L. Van Valen, and D. Gabriel. 1986. Gradual dinosaur extinction and simultaneous ungulate radiation in the Hell Creek Formation. *Science* 232:629–633.
- Sloan, R. E. and L. Van Valen. 1965. Late Cretaceous mammals from Montana. *Science* 148:220–227.
- Spieker, E. M. 1960. The Cretaceous–Tertiary boundary in Utah. *Proceedings of the 21st International Geological Congress* 5(5):14–24.
- Standhardt, B. R. 1986. *Vertebrate paleontology of the Cretaceous/Tertiary transition of Big Bend National Park, Texas*. Ph.D. dissertation, Louisiana State University, Baton Rouge.
- Staubert, E., J. D. Connick, E. Roche, and T. Smith. 1999. The Dormaal sands and the Paleocene–Eocene boundary in Belgium. *Bulletin de la Société Géologique de France* 170: 217–227.
- Sudre, J., J.-J. Jaeger, B. Sige, and M. Vianey-Liaud. 1993. Nouvelles données sur les condylarthres du Thanetian et de l'Ypresien du bassin d'Ouarzazate (Maroc). *Geobios* 26:609–615.
- Swisher, C. C. III, L. Dingus, and R. F. Butler. 1993.  $^{40}\text{Ar}/^{39}\text{Ar}$  dating and magnetostratigraphic correlation of the terrestrial Cre-

- taceous–Paleogene boundary and Puercan Mammal Age, Hell Creek–Tullock formations, eastern Montana. *Canadian Journal of Earth Sciences* 30:1981–1996.
- Szalay, F. S. 1969. Mixodectidae, Microsypidae, and the insectivore–primate transition. *Bulletin of the American Museum of Natural History* 140:195–330.
- Szalay, F. S. and M. C. McKenna. 1971. Beginning of the age of mammals in Asia: The late Paleocene Gashato fauna, Mongolia. *Bulletin of the American Museum of Natural History* 144:273–314.
- Tambareau, Y., D. E. Russell, D. Sigogneau-Russell, and J. Villatte. 1992. Découverte de Vertébrés dans le Paleocene de Campo (Pyrenees aragonaises). *Bulletin de la Société d'Histoire Naturelle de Toulouse* 128:73–76.
- Thewissen, J. G. M. 1990. Evolution of Paleocene and Eocene Phenacodontidae (Mammalia, Condylarthra). *University of Michigan Papers on Paleontology* 29:1–107.
- Thewissen, J. G. M. and P. D. Gingerich. 1987. Systematics and evolution of *Probatyopsis* (Mammalia, Dinocerata) from the late Paleocene and early Eocene of western North America. *Contributions from the Museum of Paleontology of the University of Michigan* 27:195–219.
- Ting, S. 1998. Paleocene and early Eocene land mammal ages of Asia. In *Dawn of the age of mammals in Asia*, ed. K. C. Beard and M. R. Dawson. *Bulletin of the Carnegie Museum of Natural History* 34:124–147.
- Ting, S., G. J. Bowen, P. L. Koch, W. C. Clyde, Y. Wang, Y. Wang, and M. C. McKenna. 2003. Biostratigraphic, chemostratigraphic, and magnetostratigraphic study across the Paleocene/Eocene boundary in the Hengyang Basin, Hunan, China. In *Causes and consequences of globally warm climates in the early Paleogene*, ed. S. L. Wing, P. D. Gingerich, E. Thomas, and B. Schmitz. *Geological Society of America Special Paper* 369:521–535.
- Tomida, Y. 1981. “Dragonian” fossils from the San Juan Basin and status of the “Dragonian” land mammal “age.” In *Advances in San Juan Basin paleontology*, ed. S. G. Lucas, J. K. Rigby Jr., and B. S. Kues. Albuquerque: University of New Mexico Press, pp. 222–241.
- Tomida, Y. and R. F. Butler. 1980. Dragonian mammals and Paleocene magnetic polarity stratigraphy, North Horn Formation, central Utah. *American Journal of Science* 280:787–811.
- Tsentas, C. 1981. Mammalian biostratigraphy of the middle Paleocene (Torrejonian) strata of the San Juan Basin: Notes on Torreon Wash and the status of the *Pantolambda* and *Deltatherium* faunal “zones.” In *Advances in San Juan Basin paleontology*, ed. S. G. Lucas, J. K. Rigby Jr., and B. S. Kues. Albuquerque: University of New Mexico Press, pp. 262–292.
- Van Houten, F. B. 1944. Stratigraphy of the Willwood and Tatman formations in northwestern Wyoming. *Bulletin of the Geological Society of America* 55:165–210.
- Van Valen, L. 1978. The beginning of the age of mammals. *Evolutionary Theory* 4:45–80.
- . 1988. Paleocene dinosaurs or Cretaceous ungulates in South America. *Evolutionary Monographs* 10:1–79.
- . 1994. The origin of the plesiadapid primates and the nature of *Purgatorius*. *Evolutionary Monographs* 15:1–79.
- Van Valen, L. and R. E. Sloan. 1965. The earliest primates. *Science* 150:743–745.
- . 1966. The extinction of the multituberculates. *Systematic Zoology* 15:261–278.
- Vianey-Liaud, M. 1979. Les mammifères montiens de Hainin (Paléocène moyen de Belgique), part I: Multituberculés. *Palaeovertebrata* 9:117–139.
- Wall, C. E. and D. W. Krause. 1992. A biomechanical analysis of the masticatory apparatus of *Ptilodus* (Multituberculata). *Journal of Paleontology* 12:172–187.
- Wang, B. 1975. Paleocene mammals of Chaling Basin, Hunan. *Vertebrata Palasiatica* 13:154–162.
- Wang, Y., Y. Hu, M. Chow, and C. Li. 1998. Chinese Paleocene mammal faunas and their correlation. In *Dawn of the age of mammals in Asia*, ed. K. C. Beard and M. R. Dawson. *Bulletin of the Carnegie Museum of Natural History* 34:89–123.
- Webb, M. W. 1995. A new Paleocene (Tiffanian) mammalian local fauna from near Brayton Valley, central Alberta, Canada. *Journal of Vertebrate Paleontology* 15(3):59A.
- Wegemann, C. H. 1917. Wasatch fossils in so-called Fort Union beds of the Powder River Basin, Wyoming. *U.S. Geological Survey Professional Paper* 108-D:57–60.
- Weil, A. 1998. A new species of *Microcosmodon* (Mammalia: Multituberculata) from the Paleocene Tullock Formation of Montana, and an argument for the Microcosmodontinae. *PaleoBios* 18:1–15.
- West, R. M. 1971. Deciduous dentition of the early Tertiary Phenacodontidae (Condylarthra, Mammalia). *American Museum Novitates* 2461:1–37.
- . 1973. Antemolar dentitions of the Paleocene apatemyid insectivorans *Jepsenella* and *Labidolemur*. *Journal of Mammalogy* 54:33–40.
- . 1976. The North American Phenacodontidae (Mammalia, Condylarthra). *Contributions in Biology and Geology, Milwaukee Public Museum* 6:1–78.
- Wilf, P. 2000. Late Paleocene–early Eocene climate changes in southwestern Wyoming: Paleobotanical analysis. *Bulletin of the Geological Society of America* 112:292–307.
- Wilf, P., K. C. Beard, K. S. Davies-Vollum, and J. W. Norejko. 1998. Portrait of a late Paleocene (early Clarkforkian) terrestrial ecosystem: Big Multi quarry and associated strata, Washakie Basin, southwestern Wyoming. *Palaaios* 13:514–532.
- Williamson, T. E. 1996. The beginning of the age of mammals in the San Juan Basin: Biostratigraphy and evolution of Paleocene mammals of the Nacimiento Formation. *New Mexico Museum of Natural History Bulletin* 8:1–141.
- Williamson, T. E. and S. G. Lucas. 1992. Stratigraphy and mammalian biostratigraphy of the Paleocene Nacimiento Formation, southern San Juan Basin, New Mexico. *New Mexico Geological Society Guidebook* 43:265–296.
- . 1993. Paleocene vertebrate paleontology of the San Juan Basin, New Mexico. In *Vertebrate paleontology in New Mexico*, ed. S. G. Lucas and J. Zidek. *New Mexico Museum of Natural History and Science Bulletin* 2:105–135.
- Wilson, R. W. 1956. The condylarth genus *Ellipsodon*. *University of Kansas, Museum of Natural History Publications* 9:105–116.
- Wing, S. L. 1984. A new basis for recognizing the Paleocene–Eocene boundary in the Western Interior North America. *Science* 226:439–441.
- Wing, S. L., J. Alroy, and L. J. Hickey. 1995. Plant and mammal diversity in the Paleocene and early Eocene of the Bighorn Basin. *Palaeogeography, Palaeoclimatology, Palaeoecology* 115:117–155.

- Winterfeld, G. F. 1982. Mammalian paleontology of the Fort Union Formation (Paleocene), eastern Rock Springs uplift, Sweetwater County, Wyoming. *Contributions to Geology, University of Wyoming* 21:73–112.
- Wolberg, D. L. 1979. Late Paleocene (Tiffanian) mammalian fauna of two localities in eastern Montana. *Northwest Geology* 8:83–93.
- Wood, H. E. II, R. W. Chaney, J. Clark, E. H. Colbert, G. L. Jepsen, J. B. Reeside Jr., and C. Stock. 1941. Nomenclature and correlation of the North American continental Tertiary. *Bulletin of the Geological Society of America* 52:1–48.
- Wood, R. C. 1967. A review of the Clark Fork vertebrate fauna. *Breviora* 257:1–30.
- Woodburne, M. O. 1977. Definition and characterization in mammalian chronostratigraphy. *Journal of Paleontology* 51:220–234.
- . 1987. Mammal ages, stages, and zones. In *Cenozoic mammals of North America*, ed. M. O. Woodburne. Berkeley: University of California Press, pp. 18–23.
- Woodburne, M. O. and C. C. Swisher III. 1995. Land mammal high-resolution geochronology, intercontinental overland dispersals, sea level, climate, and vicariance. In *Geochronology time scales and global stratigraphic correlation*, ed. W. A. Berggren, D. V. Kent, M.-P. Aubry, and J. Hardenbol. SEPM Special Publication 54: 335–364.
- Xue, X., L. Yue, and Y. Zhang. 1994. Magnetostratigraphical, biostratigraphical, and lithostratigraphical correlation of the red beds of Shanyang Basin, Shaanxi Province. *Science in China (ser. B)* 24:413–417.
- Xue, X., Y. Zhang, Y. B. L. Yue, and D. Chen. 1996. *The development and environmental changes of the intermontane basins in the eastern part of Qinling Mountains*. Beijing: Geological Publishing House.
- Zhang, Y.-P. 1980. A new tillodont-like mammal from the Paleocene of Nanxiong Basin, Guangdong. *Vertebrata Palasiatica* 18:126–130.
- Zhao, Z., J. Ye, H. Li, Z. Zhao, and Z. Zan. 1991. Extinction of the dinosaurs across the Cretaceous–Tertiary boundary in Nanxiong Basin, Guangdong Province. *Vertebrata Palasiatica* 29:1–20.
- Zhou, M., Y. Zhang, B. Wang, and S. Ding. 1977. Mammalian fauna from the Paleocene of Nanxiong Basin, Guangdong. *Palaeontologia Sinica, New Series C*, 20:1–100.

# 4

## Wasatchian Through Duchesnean Biochronology

Peter Robinson, Gregg F. Gunnell,  
Stephen L. Walsh, William C. Clyde,  
John E. Storer, Richard K. Stucky,  
David J. Froehlich,  
Ismael Ferrusquia-Villafranca,  
and Malcolm C. McKenna

**T**HE PREVIOUS EDITION of this chapter (Krishtalka et al. 1987) assembled much information about the areal distribution and biochronology of the continental rocks and faunas representing the Wasatchian through Duchesnean North American land mammal ages (NALMAs). That edition detailed the history of the classification of terrestrial rocks, particularly of western North America, and included some information on radioisotopic and paleomagnetic determinations. This revision concentrates on the inclusion of new data and includes expanded information concerning Mexico and Canada. We introduce new information about radioisotopic ages and paleomagnetic correlations. Wherever possible, we include data from unpublished sources and do not repeat data already cited in detail. The bibliography is as inclusive as possible. We stress field information.

Recently the definition of the Paleocene–Eocene boundary has been reviewed and is still under discussion (Aubry 1996, 1998, 2000; Berggren and Aubry 1998; Gingerich 1989, 2000; Gunnell 1998b; Koch et al. 1992). A recommendation has been put forward to locate the lower boundary of the Eocene at the negative carbon isotope excursion (CIE) in chron C24r (Koch et al. 1992; Gingerich 2001), at the beginning of the Wasatchian land mammal age (LMA) in North America. The base of the Wasatchian is marked by a major vertebrate faunal immigration event including the widespread dispersal in the Northern Hemisphere of the orders Artiodactyla, Perisodactyla, and Primates, as well as hyaenodontid Creodonta. The ultimate location of the Paleocene–Eocene boundary is in some sense irrelevant to the goals of this chapter (but see “Geochronology” later in this chapter),

which is refinement of Wasatchian through Duchesnean biochronology. This biochronologic framework will remain unaffected by the ultimate placement of the Paleocene–Eocene boundary, which may be placed at the Dababiya section in Egypt (Aubry 2001). It is clear that most of the Wasatchian and all of the Bridgerian through Duchesnean will be recognized as representing the Eocene and contingent on the final position of the Global Stratotype Section and Point (GSSP), all of the Wasatchian may ultimately be included in the Eocene. The Chadronian LMA is considered latest Eocene (Swisher and Prothero 1990) but is addressed in the following chapter.

The prior edition of this chapter pointed out that the terminology for the NALMA discussed was based on formational names and that the resulting confusion of the differences between what is contained in a rock unit and what is contained in a temporal unit with essentially the same name is regrettable, even though the temporal boundaries are flexible (Wilson 1975). This situation has not changed significantly for the LMAs in the intervening years, although there has been much more precision in the methods of collecting and recording data, and subdivisional terminology has helped. That the inconsistencies of the Wood committee (Wood et al. 1941) remain is obvious, but they are recognized as such. Many assemblages have been studied since the publication of the Wood committee report. Faunal sequences in several of these show that the age boundaries are not, need not be, and probably should not be synchronous with lithostratigraphic boundaries. For example, in several subunits of the Greater Green River Basin, Bridgerian mammals have

been recovered from rocks referred to the upper part of the Cathedral Bluffs Tongue of the Wasatch Formation, and Uintan mammals have been recovered from beds high in the Bridger Formation (Evanoff et al. 1994). Most of the lower part of the Duchesne River Formation is Uintan in age, whereas some of the lower part and the upper part are Duchesnean (Gazin 1955; Clark et al. 1967; Tedford 1970). Table 4.1 lists the more significant Wasatchian through Duchesnean fossil mammal localities relevant to this chapter.

The four NALMAs discussed here are based on mammalian fossil assemblages (figure 4.1) from two Rocky Mountain basins: the western Green River Basin (Wasatchian and Bridgerian) and the Uinta Basin (Uintan and Duchesnean). This terminology has been correlated outward from these two basins and is used throughout North America. The term *Wasatchian* is derived from the Wasatch Formation and its faunas in the Greater Green River and Fossil basins; all of the Wasatchian subages are actually derived from the faunal content of the Willwood and Wind River Formations (Sandcouleean, Graybullian, Lysitean, and Lostcabinian). The term *Bridgerian* is taken from the Bridger Formation of the Greater Green River Basin, and two of its accepted sub-age names are based on lithologic members of that formation, a situation that has to change, as will be shown later in this chapter.

Gingerich (1989, 2001) and his coworkers have developed a terminology for the subdivision of the Wasatchian (biochrons Wao to Wa7) that has been modified for other NALMAs, such as biochrons Bro–Br3 (Gunnell 1998a; Gunnell and Yarborough 2000). This terminology will be used and expanded here to include a revised subdivision of the Uintan; the Duchesnean is not subdivided here.

## HISTORY OF TERMINOLOGY

We present here an abridged version of historical terminology. See Krishtalka et al. (1987) and references therein for more detailed accounts.

### WASATCH FORMATION AND WASATCHIAN LMA

Hayden (1869) gave the name *Wasatch Group* to a reddish fluvial sequence and was derived from Wasatch Station in Weber Canyon, Summit County, Utah. This type locality (as cited by Veatch 1907) includes sediments from two separate basins, the southwestern part of the Greater Green

River Basin (Carter, Wyoming, located in the Bridger Basin of authors) and the southern part of the Fossil Basin (Wasatch Station). Cope (1877, 1882) extended the usage of Wasatch beds to the San Juan Basin in New Mexico and the Bighorn Basin in northwestern Wyoming, identifying the rocks as the *Coryphodon* zone. Hayden (1878) similarly referred sedimentary rocks in the Wind River Basin of central Wyoming to the Wasatch Formation.

The informal name *Gray Bull beds* was proposed (Granger 1914) for the lower part of the Bighorn Basin sequence. Granger (1914) extended the Lost Cabin beds (or *Lambdotherium* level) into the Bighorn Basin and referred rocks between the *Lambdotherium*-bearing deposits and the Gray Bull beds to the Lysite beds. The Lysite and Lost Cabin beds had initially been recognized as distinct units by Granger (1910), and the two names were proposed as formations by Sinclair and Granger (1911). A geologically older stratum, the Sand Coulee beds, was recognized in the Bighorn Basin by Granger (1914).

Lithostratigraphic terminology has been standardized in Wyoming basins. Wasatchian rocks of the Wind River Basin are represented in part by the Wind River Formation and include the Lysite and Lostcabin members. Temporally, the Wind River Formation spans the late Wasatchian to early Bridgerian time, with three faunal levels: Lysitean, Lostcabinian, and Gardnerbuttean. The “Big Horn Wasatch” was formalized by Van Houten (1944) as the Willwood Formation. The Willwood Formation is conformable in most places on rocks variously referred to the Fort Union Formation and is subdivided lithologically, in the southern part of the basin, into the Elk Creek and Sand Creek facies (Bown 1979b). The term *Wasatch Formation* unfortunately is still used for the upper Paleocene and lower Eocene sediments of the Powder River Basin, although this basin is physically separated from the Greater Green River Basin by both the Wind River and Hanna basins.

Veatch (1907) subdivided Hayden’s Wasatch into three formations: Almy at the base, Fowkes disconformably overlying it, and Knight at the top. The type localities of these formations are all in the Fossil Basin. In 1960, the U.S. Geological Survey standardized usage of Wasatch Formation across the entire Green River Basin area, including the Washakie Basin and Great Divide Basin (*sensu* Love 1961; now all included in the Greater Green River Basin). Oriel (1962) advocated elimination of Veatch’s formations altogether in favor of a broad usage of the Wasatch Formation. Veatch’s Fowkes Formation is now known to be Bridgerian in age (Nelson 1973, 1979).

Roehler (1992a–1992c) points out that the eastern (Great Divide, Washakie, and Sand Wash) and western parts

**TABLE 4.1** Wasatchian Through Duchesnean Fossil Localities in North America

1. Sand Coulee beds, Willwood Formation, Bighorn Basin (Wa0–Wa2)
2. “Wasatch” Formation, Powder River Basin (Wa0–Wa3)
3. Lower variegated beds, Togwotee Pass (Wa1)
4. Red Hot L.F., Tuscahoma Formation, Lauderdale County, Mississippi (Wa1)
5. Four Mile area, Wasatch Formation, Washakie–Sand Wash Basin divide, eastern Greater Green River Basin (Wa1–Wa2)
6. Wasatch and Pass Peak formations, Hoback Basin (Wa1–Wa5)
7. DeBeque Formation, Shire Member, Piceance Creek Basin (Wa2–Wa7)
8. Indian Meadows Formation, northwest Wind River Basin (Wa2–Wa3)
9. Wasatch Formation, Northern Washakie Basin (Wa3–Wa7)
10. Raven Ridge, Colton Formation, eastern Uinta Basin (Wa2–Wa7)
11. Golden Valley Formation, North Dakota (Wa3)
12. Bashi Formation, Lauderdale County, Mississippi (Wa3)
13. Las Lomas de Tetas de Cabra Formation, Baja California (Wa3? and later?)
14. “Cuchara” Formation, Raton Basin (Wa3?)
15. Gray Bull beds, Willwood Formation, Bighorn Basin (Wa3–Wa5)
16. Cooper Creek area, “Wind River” Formation, Laramie Basin (Wa3–Wa5)
17. Fisher/Sullivan Site, Nanjemoy Formation, Virginia (Wa3–Wa5)
18. San Jose Formation, San Juan Basin (Wa5–Wa6)
19. Main Body, Wasatch Formation, eastern Greater Green River Basin including Dad L.F. (Wa5–Wa7)
20. Lower Huerfano Formation, Raton Basin (Wa5–Br0)
21. Lysite Member, Wind River Formation, Wind River Basin (Wa6)
22. Lysite beds, Willwood Formation, Bighorn Basin (Wa6)
23. Morena Boulevard L.F., unnamed formation, southern California (Wa5–Wa6)
24. Cerrillos L.F., Galisteo Formation, New Mexico (Wa5?–Wa6?)
25. Book Cliffs area, Colton Formation, southern Uinta Basin (Wa6–Wa7)
26. Wasatch Formation, Fossil Basin (Wa6–Wa7)
27. LaBarge area, Wasatch Formation, western Greater Green River Basin (Wa6–Wa7)
28. North and South Fork localities, Willwood and Aycross formations, Bighorn Basin (Wa6–Br2)
29. Hannold Hill Formation, Big Bend National Park, Texas (Wa7)
30. Lost Cabin beds, Willwood Formation, Bighorn Basin (Wa7)
31. Hatchetigbee Bluff L.F., Hatchetigbee Formation, Wilcox Group, Washington County, Alabama (Wa7–Br0)
32. Niland Tongue, Wasatch Formation, northeastern Greater Green River Basin (Wa7)
33. Lost Cabin Member, Wind River Formation, northern Wind River Basin (Wa7–Br1a)
34. Farisita Formation, Raton Basin (Wa7–Br1a)
35. Locality L-41, Aycross Formation, Togwotee Pass (Wa7–Br1a)
36. Raven Ridge, Green River Formation, eastern Uinta Basin (Wa7–Br2)
37. Ellesmere and Axel Heiberg Islands, Eureka Sound Group, Nunavut Territory, Canada (Wa–Br)
38. Sage Creek Formation, Sage Creek Basin, Montana (Br0–Br3)
39. Aycross Formation, northwestern Wind River Basin (Br1b–Br2)
40. Upper Huerfano Formation, Raton Basin (Br1a)
41. Cathedral Bluffs Tongue, Wasatch Formation, Greater Green River Basin (Wa7–Br1a)
42. Lower Blacks Fork “Member” (Bridger A), Bridger Formation, southwestern Greater Green River Basin (Br1b)
43. Powder Wash locality, Green River Formation, Raven Ridge, Uinta Basin (Br1b?)
44. Middle Bridgerian beds, northwest Green River Basin (Br2)
45. Upper Blacks Fork “Member” (Bridger B), Bridger Formation, southwestern Greater Green River Basin (Br2)
46. Aycross Formation, southern Absaroka Range, Bighorn Basin (Br2)
47. Localities 17 and 18, Wagon Bed Formation, northern Wind River Basin (Br2)
48. Elderberry Canyon L.F., Sheep Pass Formation, Nevada (Br2)
49. Hart Mine Formation, New Mexico (Br3)
50. Princeton Locality, British Columbia (Br2?)
51. Togwotee Summit, Aycross Formation, Togwotee Pass (Br2?)
52. Swami’s Point L.F., Delmar Formation, southern California (Br2)
53. Tatman Formation, Bighorn Basin (Wa7–Br2)
54. Kinney Rim Member, Washakie Formation, eastern Greater Green River Basin (Br3)
55. Twin Buttes “Member,” Bridger Formation, western Greater Green River Basin (Br2–Br3)
56. Tabernacle Butte, Upper Bridger Formation, western Greater Green River Basin (Br3)
57. Fowkes Formation, Fossil Basin, Wyoming and Utah (Br3)
58. Nut Bed, Clarno Formation, Oregon (Br3)
59. Lower Adobe Town Member, Washakie Formation, eastern Green River Basin (Br3)
60. Black’s Beach L.F., Scripps Formation, southern California (Br3–Ui1)
61. Marfil, Conglomerado Rojo de Guanajuato, Guanajuato, Mexico (Br–Ui)
62. Little Stave Creek L.F., Gosport Sand, Claiborne Group, Clarke County, Alabama (Wa?, Br–Ui)
63. Washakie Formation, Sand Wash Basin, eastern Greater Green River Basin (Br3–Ui2)
64. Jackson Group, St. Francis County, Arkansas (Br–Ch?)
65. Junction localities, Agua Fria area, Devil’s Graveyard Formation, west Texas (Ui1)
66. Bridger E, Turtle Bluffs Member, Bridger Formation, southwestern Greater Green River Basin (Ui1)
67. Canoe assemblage A, Canoe Formation and Big Bend lateral equivalent, Devil’s Graveyard Formation, west Texas (Ui1?)
68. Bone Bed A, Tepee Trail Formation, East Fork River, Wind River Basin (Ui1–Ui2)
69. Middle Adobe Town Member, Washakie Formation, eastern Greater Green River Basin (Ui1–Ui2)
70. Beaver Divide, Wagon Bed Formation, southern Wind River Basin (Ui2)



TABLE 4.1 (continued)

71. Owl Creek area, Wiggins Formation, Absaroka Mountains (Ui2)	95. Lower Member C, Santiago Formation, southern California (Ui3)
72. Uinta B1 and B2, Wagonhound Member, Uinta Formation, Uinta Basin (Ui2)	96. Laguna Riviera and Camp San Onofre L.F., upper Member C of the Santiago Formation, southern California (Ui3–Du)
73. Friars L.F., Friars Formation, southern California (Ui1–Ui2)	97. Strathern/Hartman Ranch L.F., Sespe Formation, southern California (Ui3–Du)
74. Member B, Santiago Formation, southern California (Ui1–Ui2)	98. Geodetic Hills, Axel Heiberg Island, Eureka Sound Group, Nunavut Territory, Canada (Ui–Du)
75. Lower Member, Stadium Conglomerate, southern California (Ui2)	99. Skyline Channels L.F., Devil's Graveyard Formation, west Texas (Du)
76. Locality 1, Wagon Bed Formation, northern Wind River Basin (Ui2)	100. Porvenir L.F., Chambers Tuff, west Texas (Du)
77. Upper Adobe Town Member, Washakie Formation, greater Green River Basin (Ui2–Ui3)	101. Cotter Channel, Devil's Graveyard Formation, west Texas (Du)
78. Whistler Squat Quarry, Devil's Graveyard Formation, west Texas (Ui1–Ui2)	102. Green River Formation, central Utah (Du)
79. Myton Member, Uinta Formation, Uinta Basin (Ui3)	103. Dry Gulch Creek Member, Duchesne River Formation, Uinta Basin (Du)
80. Unnamed L.F., Sespe Formation, southern California (Ui3)	104. LaPoint Member, Duchesne River Formation, Uinta Basin, Utah (Du)
81. Tapo Canyon and Brea Canyon localities, Sespe Formation, southern California (Ui3)	105. Pearson Ranch/Simi Valley Landfill L.F., Sespe Formation, southern California (Du)
82. Canoe assemblage B, Canoe Formation, West Texas (Ui3)	106. Bonita L.F., "Sweetwater" Formation, southern California (Ui3–Du)
83. Chisos Formation, west Texas (Ui3)	107. Shoddy Springs L.F., Climbing Arrow Formation, Montana (Du)
84. Stonecrest L.F., Upper Member, Stadium Conglomerate, southern California (Ui3)	108. Localities Wood, Rodent, and 20, Hendry Ranch Member, Wagon Bed Formation, northern Wind River Basin (Du)
85. Serendipity L.F., Devil's Graveyard Formation, west Texas (Ui3)	109. Antelope Creek L.F., Slim Buttes Formation, South Dakota (Du)
86. Lake Casa Blanca L.F., Laredo Formation, Webb County, Texas (Ui3)	110. Hancock Quarry, Clarno Formation, Oregon (Du)
87. Candelaria L.F., Colmena Tuff, west Texas (Ui3)	111. Tonque L.F., Galisteo Formation, New Mexico (Du)
88. Swift Current Creek beds, Cypress Hills Formation, Saskatchewan (Ui3)	112. Baca and Cub Mountain formations, New Mexico (Du)
89. Mission Valley L.F., Mission Valley Formation, southern California (Ui3)	113. Carthage Coal Field, Baca Formation, New Mexico (Du)
90. Eastview L.F., Lower and Miramar Sandstone members, Pomerado Conglomerate, southern California (Ui3)	114. Windmill Hill locality, New Mexico (Du)
91. "Randlett Fauna," Brennan Basin Member, Duchesne River Formation, Uinta Basin (Ui3)	115. Turtle Basin L.F., "Brian Head Formation," Sevier Plateau, Utah (Du)
92. Localities 5, 6, and 7, Hendry Ranch Member, Wagon Bed Formation, northern Wind River Basin (Ui3)	116. Flathead Valley, Kishenehn Formation, British Columbia, Canada (Du–Ch)
93. Douglass Draw and Hough Draw L.F., "Dell Beds," Montana (Ui3)	117. Lac Pelletier, lower and upper faunas, Saskatchewan, Canada (Du–Ch)
94. Upper Assemblage, Pomerado Conglomerate, southern California (Du–Ch)	118. Diamond O L.F., Climbing Arrow Formation, Montana (?Du–?Ch)

Biochronologic zone range in parentheses: Br, Bridgerian; Ch, Chadronian; Du, Duchesnean; Ui, Uintan; Wa, Wasatchian. L.F., Local Fauna. The locality numbers appear in figures 4.1–4.7.

(Green River Basin) of the major southwestern Wyoming basin are really parts of a Greater Green River Basin in which much of the middle part of the basin is divided by the structurally younger Rock Springs Uplift. The uplift does not cut the basin fully in half, and the units can be traced around the uplift on the northern side. The two sides of the basin have similar depositional histories. Physical correlation of the rock units between the Greater Green River Basin and other basins such as the Uinta and Piceance

Creek basins to the south is much less easily accomplished, despite the superficial similarity of the rock units.

Granger (1914) divided Cope's (1877) "Wahsatch" in the San Juan Basin of New Mexico into lower Almagre beds and upper Largo beds, basing the difference on the presence of *Meniscotherium* in the Largo (Lucas 1977 has since reported it from the Almagre). Simpson (1948) concluded that the Almagre and Largo should be regarded as facies of a single formation, the San Jose Formation. Re-

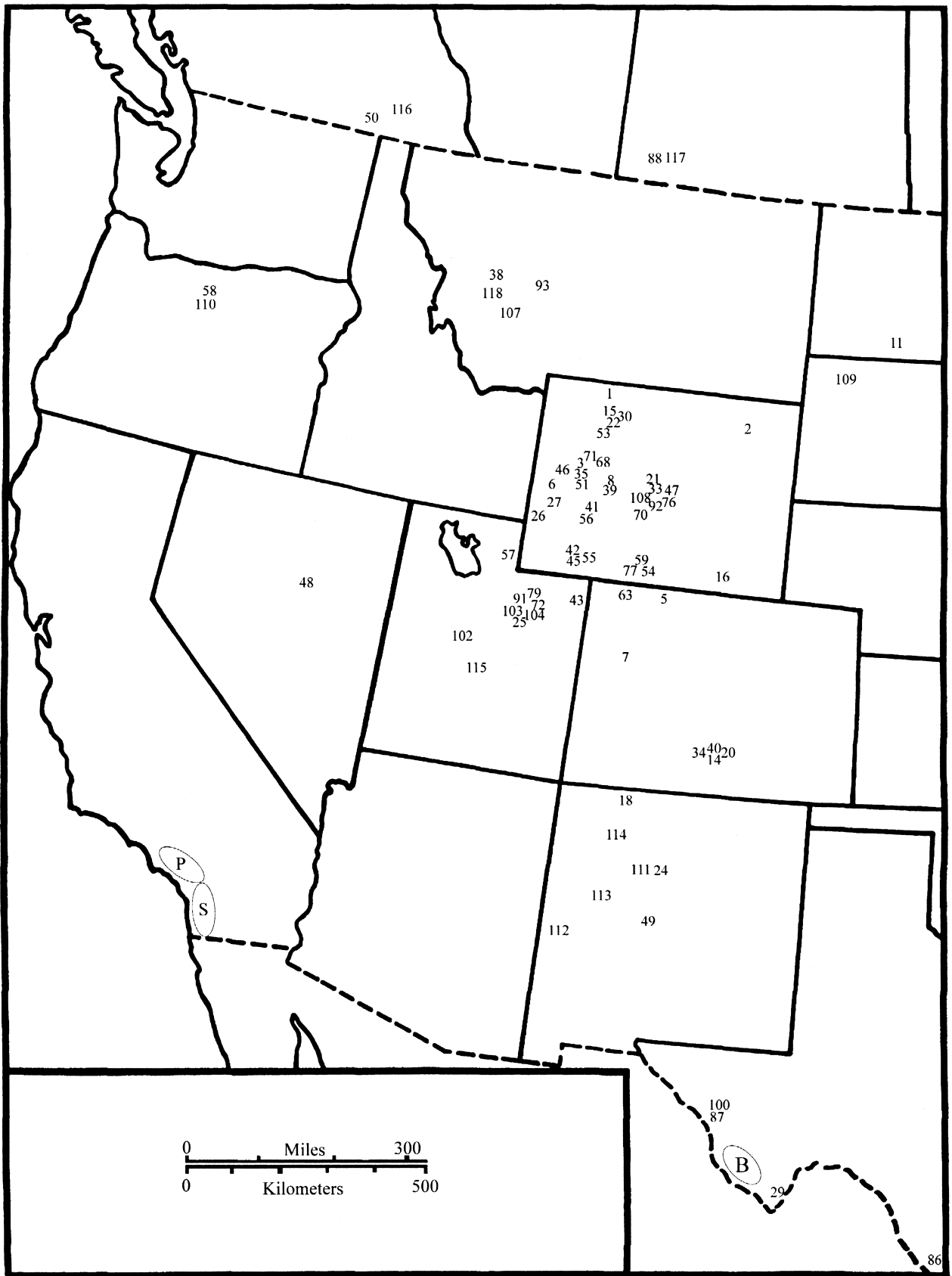


FIGURE 4.1 Map of the main Wasatchian through Duchesnean fossil mammal sites in western North America; localities are listed in table 4.1.

cent work indicates that most of the faunas of the San Jose Formation are Lysitean (Wa6) in age, based primarily on the distribution of the temporally limited taxon *Xenicohippus*. The presence of a large paromomyid in the fauna (Smith and Lucas 1991, locality 20) indicates either that rocks as old as Wa2 may be in the San Jose Formation (Robinson and Ivy 1994) or that large paromomyids survived much later in the Wasatchian as stated by Smith and Lucas (1991:35). Some of the San Jose Formation is Wa5 (upper Graybullian) in age.

There has been a general recognition that although the Wasatchian age sediments of each of the intermontane basins have certain similarities, the history of each basin is distinct and the geologic formations are separate sedimentologic entities. Other areas in which Wasatchian time is represented include the Hoback Basin of central western Wyoming; the Togwotee Pass area southeast of Yellowstone National Park; the Powder River Basin; the Williston Basin, Golden Valley Formation, in south-central North Dakota; the Laramie Basin of southeastern Wyoming; the Piceance Creek Basin of western Colorado; the Raton Basin of south-central Colorado; the Tusahoma and Bashi formations in Mississippi; Hannold Hill Formation of west Texas; the Lomas las Tetas de Cabra Formation of Baja California; Axel Heiberg and Ellesmere Islands in Canada; and the Talkeetna Mountains in Alaska.

#### BRIDGER FORMATION AND BRIDGERIAN LMA

The Bridgerian age of Wood et al. (1941) is based on the time of deposition and the faunas of Bridger A–D. The Bridger Formation was named by Hayden (1869) as the Bridger Group, with the type area at Church Buttes Station, Wyoming. Matthew (1909) published a comprehensive Bridger stratigraphy, dividing the formation into five alphabetically designated zones (A–E) demarcated by laterally extensive white marker beds and further subdivided ( $B_1$ ,  $B_2$ , etc.) on the basis of persistent benches held up by resistant marl beds (Matthew's Bridger E is now known to be fossiliferous [West and Hutchison 1981; Evanoff et al. 1994] and should not be included in the definition of the Bridgerian). Wood (1934) recognized the apparent faunal similarity between Bridger A and B and between Bridger C and D and combined these zones into two members, the lower Blacks Fork Member and the higher Twin Buttes Member. West and Hutchison (1981) designated the uppermost part of the Bridger Formation (Matthew's Bridger E) as the Cedar Mountain Member. This member name is precluded because of prior usage for the Cedar Mountain Formation, Cretaceous, of Utah

and has been replaced by the term *Turtle Bluffs Member* (Evanoff et al. 1998). McGrew and Sullivan (1970) examined the exposures of Matthew's Bridger A near Opal, Wyoming, and argued for the basic faunal similarity of Bridger A and B. However, Gingerich (1979) recognized different species of adapiform euprimates in Bridger A and Bridger B, and this appears to be the case for much of the fauna (Gunnell 1998a; Zonneveld et al. 2000).

The Bridger Formation has never been called by any other name, and there are only the lithologic subdivisions just mentioned (West 1976b). Other areas where Bridgerian time is represented include elsewhere in the Greater Green River Basin, Wind River Basin, and Bighorn Basin all in Wyoming; the Uinta Basin of Utah and Colorado; the Piceance Creek Basin of Colorado; the northern Raton Basin of Colorado; west Texas, Oregon, New Jersey, Ellesmere Island, and perhaps Axel Heiberg Island, Canada.

#### UINTA FORMATION AND UINTAN LMA

Comstock (1875) first formally recognized the Uinta Formation of northeastern Utah. Peterson (in Osborn 1895) divided the Uinta Formation into Horizons A, B, and C, a division that was followed by Riggs (1912) and Douglass (1914). As noted by Cashion and Donnell (1974) and Prothero (1996b), Osborn (1929) restricted Uinta A to only the lowermost, barren part of the formation, named Uinta B<sub>1</sub> for the sparsely fossiliferous upper part of Peterson's original Horizon A, and named Uinta B<sub>2</sub> for Peterson's original Horizon B. As noted by Walsh (1996a), Osborn (1929) also lowered the Uinta B–C boundary to the top of the *Amynodon* sandstone, which resulted in the grayish Devil's Playground beds being transferred from the upper part of Uinta B to the lower part of Uinta C. Osborn's redefinition of the Uinta B–C boundary is problematic because the *Amynodon* sandstone is not a widely mappable unit (Cashion 1986). Most recently, Crawford et al. (2002) advocated returning the Uinta B–C lithostratigraphic contact to the striking gray-to-red color change that marked Peterson's original boundary between these units.

Wood (1934) proposed the Wagonhound Member for the lower part of the Uinta Formation (A and B) and Myton Member for the upper part (C). The final alteration in Uinta Formation usage was the separation of the beds above Uinta C as the Duchesne Formation (Peterson 1932), modified to Duchesne River Formation (Kay 1934).

As discussed in Krishtalka et al. (1987), the Uintan LMA originally was based on the fauna and time of deposition of Uinta A–C, but the age of Uinta A is essentially unknown, and this lithostratigraphic-based characterization is replaced by a purely biochronologic definition later in

this chapter. Other areas containing Uintan rocks and faunas include southwestern Saskatchewan; southwestern Montana; northwestern South Dakota; southwestern, central, and northwestern Wyoming; Trans-Pecos and southern Texas; northern New Mexico; northwestern Colorado; southern California; and Mississippi.

### DUCHESNE RIVER FORMATION AND DUCHESNEAN LMA

As with the underlying Uinta Formation, the upper part of the Eocene section in the Uinta Basin has had a complex nomenclatorial history. Peterson (1932) named the Duchesne Formation for the sparsely fossiliferous upper part of the original Horizon C of the Uinta Formation. This was soon changed by Kay (1934) to the Duchesne River Formation because the original name was already in use. The Duchesne River Formation overlies and intertongues with the Uinta Formation, although there is a substantial change in lithofacies (Kay 1934; Scott 1945; Anderson and Picard 1972). Three lithostratigraphic and biostratigraphic horizons were recognized by Kay (1934) in the Duchesne River Formation, but they have since been abandoned in favor of the formal lithostratigraphic members of Anderson and Picard (1972). From oldest to youngest, Kay's subdivisions were the Randlett, Halfway, and LaPoint horizons. The Randlett corresponds roughly to the lower two-thirds of the Brennan Basin Member of Anderson and Picard (1972). The Halfway horizon is equivalent to the upper third of the Brennan Basin Member plus the Dry Gulch Creek Member, and the LaPoint horizon is essentially equivalent to the LaPoint Member. The uppermost unit (the Starr Flat Member of Anderson and Picard 1972) has yet to yield fossils.

The entire thickness of the Duchesne River Formation was the original basis for the Duchesnean of Wood et al. (1941), but the Duchesnean LMA has now been restricted to faunas that occur in the Dry Gulch Creek and lower LaPoint members (Gazin 1955; Clark et al. 1967; Tedford 1970; Wilson 1978, 1986; Emry 1981; Lucas 1992; Rasmussen, Hamblin, and Tabrum 1999). In addition to the Uinta Basin, Duchesnean faunas have been found in Saskatchewan, Montana, Oregon, Wyoming, southwestern Texas, New Mexico, and southern California.

### BIOCHRONOLOGY OF WASATCHIAN-DUCHESNEAN LMAS

The Wood et al. (1941) committee definitions of NALMA faunal elements have been greatly modified by nomen-

clatural revisions and refinements and by more detailed geologic work. Despite these changes, the basic structure of the Wood et al. framework has held up very well. We present updated biochronologic range information in summary form here. For a more detailed historical review of biochronologic ranges see Krishtalka et al. (1987).

### WASATCHIAN

Wasatchian first appearances include *Perissodactyla*, *Artiodactyla*, *Primates*, *Hyaenodontidae*, *Sciuravidae*, *Palaeictops*, *Didelphodus*, *Macrocranion*, *Pachyaena*, *Miacis*, *Vulpavus*, and *Megalestonyx* (Schankler 1980; Rose 1981). Of these, the simultaneous appearance of *Perissodactyla* (*Hyracotherium*), *Artiodactyla* (*Diacodexis*), omomyid euprimates (*Teilhardina*), adapiform euprimates (*Cantius*), and *Hyaenodontidae* mark the onset of the Wasatchian and the end of the Clarkforkian.

Important last occurrences in the Wasatchian (a criterion of lesser biostratigraphic utility) include *Meniscotherium*, *Homogalax*, *Xenicohippus*, *Lambdotherium*, *Anacodon*, *Niptomomys*, *Tetonius*, *Pelycodus* (*sensu strictu* [s.s.]), and *Pachyaena* (Schankler 1980; Stucky 1984b; Bown and Kihm 1981).

Wasatchian index taxa (restricted to the Wasatchian) include *Ambloctonus*, *Diacodon*, *Homogalax*, *Lambdotherium*, *Meniscotherium*, *Pachyaena*, *Tetonius*, *Pelycodus* (s.s.), *Ectoganus*, *Wasatchia* (s.s.), *Xenicohippus*, *Copelemur*, *Anemorhysis*, *Loveina*, and *Notoparamys* (Korth 1982). *Cantius*, once considered a Wasatchian index taxon, is now known from Bridgerian beds in the Piceance Creek Basin (University of Colorado Museum [UCM] loc. 96197), from the Cathedral Bluffs Tongue of the Wasatch Formation, and probably the Huerfano formation as well (UCM loc. 77039).

Taxa typical of the Wasatchian (not all of which are restricted to Wasatchian time) include *Hyracotherium* (which is probably the wrong name for the North American taxon), *Coryphodon*, *Phenacodus*, *Ectocion*, *Hyopsodus*, *Paramys*, *Microsyops*, *Cantius*, *Absarokius*, *Prolimnocyon*, *Prototomus*, *Arfia*, *Oxyaena*, *Didymictis*, *Viverravus*, *Diacodexis*, *Lambdotherium*, *Homogalax*, *Esthonyx*, and *Ectoganus*. The presence of abundant *Hyopsodus*, *Cantius*, *Hyracotherium*, and *Diacodexis* differentiates the Wasatchian from the earlier Clarkforkian. The end of the Wasatchian (= the beginning of the Bridgerian) can now be defined by the penecontemporaneous appearance of *Hyrachyus*, *Palaeosyops*, *Eotitanops* (this genus may occur in latest Wasatchian rocks; Gunnell and Yarborough 2000; Smith and Holroyd 2001), *Trogosus*, *Omomys* (the presumed Wasatchian *Omomys* are now placed in other genera; Stucky 1984a; Bown and Rose 1984; Honey 1990),

*Washakius*, *Anaptomorphus*, *Smilodectes*, *Megadelphus*, *Pantolestes*, and *Microsus*.

Work by Gingerich and coworkers has established numerical biochrons Wao–Wa7 for the Wasatchian LMA. The Sandcouleean (early Wasatchian) includes Wao–Wa2, the Graybullian (middle Wasatchian) spans Wa3–Wa5, and the Lysitean (Wa6) and Lostcabinian (Wa7) together represent the late Wasatchian.

The Sandcouleean is characterized by the occurrence of the following genera (not all limited to the Sandcouleean): *Phenacodus*, *Coryphodon*, *Hyracotherium*, *Hyopsodus*, *Haplomytus*, *Apheliscus*, *Ectocion*, *Arctodontomys*, *Phenacolemur*, *Cantius*, *Teilhardina*, and *Diacodexis*. *Plesiadapis* makes its last appearances, and multituberculates are widespread.

The Graybullian is characterized by many of these same genera, but *Arctodontomys* is replaced by *Microsypops*, *Homogalax protapirinus* appears and is both common and widespread, and rodents become more diverse. The Graybullian is also characterized by the last appearances of *Neoliotomus* and *Haplomytus* (Wa5). *Apheliscus* was thought to be included here, but recent work by Denver Museum parties has documented early Wa6 *Apheliscus* in association with *Heptodon* at Denver Museum of Nature and Science locality 66 in the Wind River Formation.

Lysitean (Wa6) first appearances include *Heptodon*, *Loveina*, and *Hexacodus* (Guthrie 1967; Krishtalka and Stucky 1985). The Lostcabinian (Wa7) is marked by the first appearance of *Lambdaotherium*, a genus that is widespread, common, and apparently limited to Wa7. Wa7 as a whole is characterized by the first occurrence of *Shoshonius*, *Antiacodon*, *Orohippus*, *Megalestonyx*, *Pauromys*, *Sciuravus*, and *Armintodelphys* and derived species of *Estonyx*, *Diacodexis*, *Absarokius*, *Hyopsodus*, *Cantius/Notharctus*, and *Hyracotherium*. Last appearances in Wa7 are *Lambdaotherium* and *Meniscotherium*.

## WASATCHIAN-BRIDGERIAN BOUNDARY

Transitional Wasatchian–Bridgerian faunal assemblages are not widely represented. The best of these are known from the upper part of the Wind River Formation in the Wind River Basin (Stucky 1984a; Stucky and Krishtalka 1983; Krishtalka and Stucky 1983), Huerfano Park in Colorado (P. Robinson 1966), and at South Pass along the northeastern margin of the Greater Green River Basin (Gunnell and Yarborough 2000; Gunnell and Bartels 2001). P. Robinson (1966) originally assigned the fauna from the upper Huerfano Formation to a new subage, the Gardnerbuttean, which he included as the last subage of the Wasatchian. The upper Huerfano and the *Palaeosyops borealis* Assemblage Zone (Bro) faunas

(Stucky 1984a) are now considered earliest Bridgerian, an action endorsed by Robinson and other workers. Gunnell (1998a) and Zonneveld et al. (2000) have shown that much of the Gardnerbuttean occurs above Bro in biochron Br1a. Corollaries of this conclusion are that the end of the Lostcabinian (and the *Lambdaotherium* Range Zone) marks the end of the Wasatchian, and the Lost Cabin Member in the Wind River Basin straddles the Wasatchian–Bridgerian boundary. The Wasatchian–Bridgerian boundary is now placed near a date of  $50.56 \pm 0.13$  Ma from the Grey Tuff of the Wilkins Peak Member of the Green River Formation (M. E. Smith et al. 2003).

Another area that preserves faunal elements of the Wasatchian–Bridgerian transition is in the northern Greater Green River Basin north of Tabernacle Butte. In this area, a referred section of the Cathedral Bluffs Tongue was thought to overlie the New Fork Tongue of the Wasatch Formation (West 1973a). Upon reexamination by Roehler (1991), the New Fork Tongue is now considered to be the westward extension of the Cathedral Bluffs Tongue, and what had been considered Cathedral Bluffs Tongue is now considered to be the Laney Shale Member of the Green River Formation. The New Fork Tongue was thought to have produced a mixed late Wasatchian–early Bridgerian fauna from more than 250 feet of strata (*Hyracotherium*, *Meniscotherium*, *Coryphodon*, *Palaeosyops*, *Lambdaotherium*, *Phenacodus*, and *Oxyaena*) (West 1973a). More recent collections from this area by L. Ivy (UCM) indicate that the Wasatchian and Bridgerian faunas are not mixed; these collections indicate that the characteristic Lostcabinian genera of *Lambdaotherium* (UCM loc. 84211, 84228) and *Meniscotherium* (UCM loc. 84228, 84220) occur in the lower parts of the outcrop area and that the typical Bridgerian taxa of *Palaeosyops* (UCM loc. 84219) and *Hyrachyus* (UCM loc. 84213) occur in the upper. None of the stratigraphically higher UCM localities have Wasatchian fossils in them, and none of the lower have Bridgerian. The overlying “Cathedral Bluffs Tongue” (West and Dawson 1973) is miscorrelated (Roehler 1991) and is a fluvial facies of the Green River Formation and contains a typically Bridgerian faunal sample including *Orohippus*, *Helaletes*, *Antiacodon*, *Pauromys*, *Tillomys*, and *Mysops*. There is a lithologic difference between the “New Fork Tongue” (= Cathedral Bluffs Tongue) and the “Cathedral Bluffs Tongue” (= Laney Shale Member of the Green River Formation) in the northern Green River Basin, and according to Roehler (1991) there is an erosional unconformity between the top of the Cathedral Bluffs Tongue and the overlying Laney Shale member of the Green River Formation in part of this area. Based on this new infor-

mation (Roehler 1991), the Wasatchian–Bridgerian boundary occurs in the Cathedral Bluffs Tongue of the Wasatch Formation on the western side of the Greater Green River Basin and on the eastern (Honey 1988; Gunnell and Yarborough 2000; Clyde et al. 2001).

The absolute dating of the Wasatchian–Bridgerian boundary is still being evaluated. Recent absolute dates (Clyde et al. 2001; M. E. Smith et al. 2003) are significantly at variance by approximately 2 Ma. As described in the preceding two paragraphs, the physical location of the boundary can be demonstrated in several basins by faunal turnover. The only date that is well placed in regard to the physical position of the boundary is that of the Grey Tuff cited by Smith et al. from the Wilkins Peak Member of the Green River Formation, a lateral correlative of the Cathedral Bluffs Member of the Wasatch Formation. If this date is accurate, then it is the best date available at this time.

The onset of the Bridgerian is defined by the first appearance of *Palaeosyops*, *Trogosus*, *Hyrachyus*, *Homacodon*, *Helohyus*, *Megadelphus*, *Smilodectes*, *Omomys*, *Washakius*, and *Pantolestes*. However, these occurrences are not necessarily simultaneous. *Trogosus* occurs first in such localities as Huerfano VII (Bro). *Hyrachyus* and *Palaeosyops fontinalis* occur slightly higher in the section in the Huerfano Basin (locs. II, III, V; P. Robinson 1966) and apparently in the Green River and Wind River basins as well (Gunnell 1998a; Zonneveld et al. 2000). *Trogosus*, *Hyrachyus*, and *Palaeosyops* are all widespread, and *Palaeosyops* and *Hyrachyus* are particularly common.

Other areas that record the Wasatchian–Bridgerian transition include the Green River Formation–Colton Formation intertonguing area of the eastern Uinta Basin (Utah and Colorado; Doi 1990), the Piceance Creek Basin of Colorado (Kihm 1984; Honey 1990), and the Willwood and Aycross formations (Bown 1979a, 1982; Gunnell et al. 1992) in the Bighorn Basin.

## BRIDGERIAN

The earliest Bridgerian (middle and upper Gardnerbuttean, Br1a) is marked by the first appearance of *Palaeosyops*, *Utahia*, *Trogosus*, *Uintanius*, *Hyrachyus*, *Homacodon*, *Helohyus*, *Megadelphus*, *Patriofelis*, *Sinopa*, *Microsus*, *Thinocyon*, *Smilodectes*, *Omomys*, *Washakius*, and *Pantolestes*. In addition, there are several holdover Wasatchian taxa typical of the earliest Bridgerian, including *Esthonyx*, *Coryphodon*, *Bunophorus*, *Diacodexis*, *Didymictis*, *Hyracotherium*, *Shoshonius*, *Bathyopsis*, *Absarokius*, *Thryptacodon*, *Palaeosinopa*, and *Knightomys*.

Other Bridgerian first appearances (Br1b and later) include *Pantolestes*, *Anaptomorphus*, *Taxymys*, *Mysops*, *Mesatirhinus*, *Harpagolestes*, *Limnocyon*, *Mesonyx*, *Telmatherium*, *Hemiacodon*, *Dilophodon*, *Tillomys*, *Manteoceras*, *Parisectolophus*, *Uintatherium*, and leptocherids.

Bridgerian last occurrences include *Vulpavus*, *Patriofelis*, *Phenacodus*, *Hyracotherium*, *Antiacodon*, *Diacodexis*, *Bunophorus*, *Coryphodon*, *Bathyopsis*, *Absarokius*, *Shoshonius*, *Hapalodectes*, *Knightomys*, *Palaeosinopa*, *Thryptacodon*, *Esthonyx*, *Heptodon*, *Helaletes*, *Ectocion*, *Prolimnocyon*, and *Didymictis*.

Characteristic Bridgerian taxa include *Hyopsodus*, *Miacis*, *Sciuravus*, *Paramys*, *Pauromys*, *Peratherium*, *Antiacodon*, *Microsyops*, *Notharctus*, *Orohippus*, *Helaletes*, *Apatemys*, *Scenopagus*, *Pontifactor*, *Nyctitherium*, *Centetodon*, *Entomolestes*, and *Uintasorex*.

Gunnell (1998a) has proposed a subdivision of the Bridgerian that is similar in concept to that of the Wasatchian. He recognizes Bro, Br1a, Br1b, Br2, and Br3 biochrons. Bro is the *Eotitanops borealis* Range Zone and is the lowest part of the Gardnerbuttean. This biochron is recognized in the Wind River Formation (Upper Lost Cabin Member; Stucky 1984a), in the Aycross Formation (Flynn 1986), in the Cathedral Bluffs Tongue of the Wasatch Formation near Boulder, Wyoming (West 1973a), and in the Huerfano Formation of Colorado (American Museum of Natural History [AMNH] loc. VII). Bro forms the lower part of the Gardnerbuttean Subage of the Bridgerian LMA. *Eotitanops borealis* is restricted to this range zone; *Trogosus*, a Bridgerian immigrant, occurs with it at Huerfano VII.

Br1a comprises the middle and later parts of the Gardnerbuttean. It is present in the Wind River Formation (several Carnegie Museum [CM]/UCM Locs), in the Cathedral Bluffs Tongue of the Wasatch Formation and Laney Shale Member of the Green River Formation of the Greater Green River Basin (several localities), in tongues of the Colton and Green River formations at Raven Ridge in the Uinta Basin, in the Green River Formation of the northern Piceance Basin (Honey 1990), and in the Huerfano Formation (AMNH locs. I, II, III, and V). *Hyrachyus*, *Helaletes*, *Patriofelis*, *Bathyopsis fissidens*, *Trogosus*, *Mesonyx*, *Notharctus robinsoni*, *Omomys carteri*, *Smilodectes*, *Scenopagus*, and *Palaeosyops* are present and widespread. First appearances are *Uintanius*, *Utahia*, and *Megadelphus*. *Eotitanops minimus*, *Megadelphus lundeliusi*, and *Didymictis vanceaveae* are restricted to Br1a. Last appearances in Br1a are *Hyracotherium*, *Didymictis*, *Coryphodon*, and *Cantius* (Green River Formation of the Piceance Creek Basin; Huerfano Formation). We disagree with the assignment by Rose et al.

(1999) of the Uintan *Hesperolemur* to *Cantius* (Gunnell and Rasmussen in prep.).

The Blacksforkian Subage comprises biochrons Br1b and Br2. Br1b is the biochron for the fauna from Matthew's Bridger A rocks. It is characterized by a typical Bridgerian fauna with several species restricted to it. Among these are *Bathyopsis middleswarti*, *Anaptomorphus westi*, and *Smilodectes mcgrewi*. It also has several species characteristic of the Br2 or Br3 biochrons such as *Washakius insignis*, *Paramys delicatus*, *Paramys delicatior*, *Sinopa rapax*, and taxa of *Orohippus*, *Antiacodon*, and *Microsycop*s that are very close to Br2 forms. The last appearance of *Bathyopsis* is recorded in the Br1b biochron. Br1b faunas are rare outside the type area in southwestern Wyoming, but a notable exception is the rich micro-mammal locality in the Green River Formation at Powder Wash on Raven Ridge in the Uinta Basin (Gunnell and Bartels 1999). Recent fieldwork by Doi (1990) has significantly augmented the Raven Ridge collections, and several Br1b localities are now known there.

The Br2 fauna (upper Blacksforkian) is characterized by *Notharctus pugnax* and *N. tenebrosus*, *Smilodectes gracilis*, *Orohippus major*, *Tillodon*, and *Microsycop*s *elegans*. Uintatheres are lacking in this faunal unit. The last appearances of *Trogosus* and *Tillodon* are recorded; *Trogosus* therefore is a very good indicator of Br0 to Br2 age rocks. The boundary between the Br2 and Br3 biochrons is not clear-cut. Matthew (1909), Wood (1934), and Wood et al. (1941) assumed that the faunal break occurred at the boundary between Matthew's Bridger B and C, the Sage Creek White Layer (SCWL). Recent collecting by UCM parties has shown that the lower 70 m of Bridger C are sparsely fossiliferous. Br2 faunas are not widespread in North American intermontane basins. Other areas where these faunas are known include Raven Ridge (Uinta Basin) and possibly the Green River Formation of the Washakie Basin.

The Twinbuttean Subage fauna (Br3) contains abundant uintatheres (*Uintatherium*), and its brontotheriid diversity (*Telmatherium*, *Manteoceras*, *Mesatirhinus*) is greater than that of earlier faunas. Two species of large *Notharctus* (Covert et al. 1998), *Homacodon vagans*, *Orohippus sylvaticus*, *Hyopsodus lepidus*, and *Microsycop*s *annectens* are also unique to the late Bridgerian. Br3 age rocks are found in the Washakie Formation (Kinney Rim Member and lower Adobe Town Member; McC Carroll et al. 1996a), the Wind River Basin (Wagon Bed Formation at Beaver Divide and Badwater Creek), and the Piceance Creek Basin (*Tethyopsis*, UCM loc. 84115).

It is now evident that the Gardnerbuttean (Br0, Br1a; Upper Cathedral Bluffs, Laney Shale Member of the Green River Formation) is the oldest part of the Bridger-

ian LMA, followed by the Blacksforkian (Br1b, Br2; Bridger A and Bridger B; Bridger C *pro parte*) and the Twinbuttean (Br3, middle and upper Bridger C and Bridger D) (see Wood 1934; Matthew 1909; P. Robinson 1966; Gazin 1976; Gunnell 1998a). The faunal boundary between the Blacksforkian (Br2) fauna and the Twinbuttean (Br3) fauna apparently does not occur at the Bridger B–C boundary (Sage Creek White Layer, the lithologic boundary between the Black Fork Member and the Twin Buttes Member) but some 70 m above it, between the Hickey Mountain Limestone and the Burnt Fork Limestone (Murphey 2001). This probably explains why so many typical Bridger B *Notharctus* and *Smilodectes* are recorded from the Twin Buttes member (P. Robinson 1957b; Gingerich 1979) and further illustrates the bad practice of naming faunal and stratigraphic units with the same term. One UCM specimen of *Notharctus pugnax* (a Br2 species) comes from 69 m above the base of the Sage Creek Limestone near Sage Creek Mountain, and the fauna from the Hickey Mountain Limestone, a thin lacustrine bed some 69 m above the base of the Sage Creek White Layer, appears to be Br2 in age. In general, the lower 70–80 m of Bridger C (lower Tertiary Bridger C [TBC] of Evanoff et al. 1998; Murphey 2001) are sparsely fossiliferous and poorly represented in collections, so most Bridger C specimens present in collections probably have come from the upper two thirds of the unit. Only the UCM collections in the Twin Buttes member made since 1991 have adequate stratigraphic data.

## BRIDGERIAN-UINTAN BOUNDARY

There are few sequences that preserve continuous, fossiliferous sequences across the Bridgerian–Uintan boundary. Sediments in the East Fork Basin, Wyoming (McKenna 1980b); the Washakie Basin, Wyoming (Roehler 1973); the Uinta Basin, Utah (Kay 1957); Beaver Divide, Wyoming (Emry 1975); the Baca and Galisteo formations in New Mexico (Lucas et al. 1981; Lucas and Williamson 1993); the Badwater Creek area, Wyoming (Black 1969); the Sand Wash Basin, Colorado (West and Dawson 1975; Stucky et al. 1996); the Trans-Pecos area, Texas (West 1982); the Green River Basin, Wyoming (Evanoff et al. 1994); and the Scripps, Friars, and Santiago formations in southern California (Walsh 1996a) may preserve all or portions of this transitional interval.

One difficulty surrounding the definition of the Bridgerian–Uintan boundary concerns the essentially unknown fauna of Uinta A (as restricted by Osborn 1929 and discussed by Cashion and Donnell 1974). Prothero

(1996b) points out that several Uintan taxa assumed by Krishtalka et al. (1987) to have been collected from Uinta A were almost certainly obtained from Uinta B1 of Osborn (1929). Support for this conclusion comes from the fact that Peterson's (1924) "*Dolichorhinus* Quarry" (rediscovered by Bilbey et al. 2002) is listed from Uinta A in Carnegie Museum records (A. Tabrum, pers. comm., 2000) but actually occurs about 122 m stratigraphically above the Uinta A–B contact as mapped by Cashion (1974).

Two areas that may be particularly relevant to the Bridgerian–Uintan boundary issue are the Agua Fria area in Trans-Pecos, Texas, and the newly discovered Bridger E faunal sample from the Green River Basin. In the Agua Fria area, the Devil's Graveyard Formation unconformably overlies the Cretaceous. The basal unit, the basal Tertiary conglomerate, contains two localities, Junction and 0.6 miles east of Junction. These localities are not part of the Whistler Squat Quarry assemblage. The faunal assemblage from these two localities includes a mix of Bridgerian (*Herpetotherium*, *Scenopagus*, *Omomys*, *Notharctus*, *Microsypops annectens*, *Thisbemyx*, *Mysops*, *Stylinodon*, *Hyrachyus*, and *Helohyus*) and Uintan (*Triplopus*, *Ourayia*, *Macrotarsius*, and *Leptoreodon*) elements as well as some rangethrough taxa (*Centetodon*, *Microparamys*, and *Uintatherium*).

The Bridger E faunal assemblage (Evanoff et al. 1994; UCM loc. 92189) occurs approximately 7 m above the Bridger D–E lithologic boundary and contains faunal elements similar to those found at the Junction localities, including many Bridgerian taxa along with *Triplopus*, Eomyidae, and a very derived *Hemicacodon* that resembles *Macrotarsius*. The base of the Bridger E sequence is dated at  $46.16 \pm 0.44$  Ma (Murphey et al. 1999).

The Agua Fria and Bridger E assemblages indicate that there is a transitional faunal interval between Bridgerian biochron Br3 and the early Uintan faunal assemblages typified by Uinta B in Utah and the early Uintan samples from southern California, neither of which contain so many holdover Bridgerian taxa. This interval may represent the earliest Uintan and is assignable to biochron Ui1. The typical fauna of Uinta B beds therefore would comprise biochron Ui2 and that of Uinta C and the lower Duchesne River Formation comprise biochron Ui3. The early Uintan appears to fall entirely within chron C2or, which has a duration of 2 Ma (Berggren et al. 1995).

Flynn (1986) proposes a subage of the Uintan, the Shoshonian, as the basal Uintan temporal unit. This subage was originally proposed based on faunal samples from Bone Bed A of the Tepee Trail Formation (McKenna 1980b) and from the greater San Diego area, with faunal

samples from the upper part of the lower Adobe Town Member of the Washakie Formation being included with less certainty (Flynn 1986). In two more recent articles, McCarroll et al. (1996a, 1996b) argue for the presence of a Shoshonian interval in the middle unit of the Adobe Town Member in the Washakie Basin. Stucky et al. (1996) also recognize an earliest Uintan or "Shoshonian" (their quotations) interval from above the Robin's Egg Blue Tuff in the Washakie Formation, Sand Wash Basin in Colorado (West and Dawson 1975). McKenna (1990) uses the term *Shoshonian* for the Tepee Trail Bone Bed A Quarry Fauna, most of which remains undescribed.

Flynn (1986) characterizes Shoshonian assemblages as those containing the first appearances of typical Uintan taxa such as *Amyrnodon*, *Leptoreodon*, *Protoreodon*, *Protylopus*, *Macrotarsius*, *Oligoryctes*, *Achaenodon*, and possibly *Epihippus* and co-occurring Bridgerian taxa such as *Notharctus*, *Microsypops annectens*, *Sciuravus*, *Hemicacodon*, *Washakius*, *Omomys*, *Herpetotherium*, *Apatemys*, and *Uintasorex*. Additional Shoshonian first appearances cited include *Triplopus*, *Uintaceras*, *Ourayia*, *Oromeryx* from the Sand Wash Basin (Stucky et al. 1996; D. T. Rasmussen, pers. comm., 2000), and *Dolichorhinus*, *Eobasileus*, *Metarhinus*, and some of the Sand Wash taxa from the middle Adobe Town Member, Washakie Basin (McCarroll et al. 1996a, 1996b).

Recent taxonomic revision and stratigraphic study have called some of these associations into question. Many of the San Diego primate taxa cited by Flynn as Bridgerian co-occurrences have been assigned to different genera or species from those known in the Bridgerian (Mason 1990; Gunnell 1995; Walsh 1996a; Walsh and Rasmussen in prep.). Some of the co-occurring taxa (*Herpetotherium*, *Nyctitherium*, *Apatemys*, and *Sciuravus*) are rangethrough taxa with temporal ranges that span much of the Wasatchian through Uintan and therefore are of limited biostratigraphic use. Most of the taxa cited by Flynn (1986) as Uintan first occurrences are not known from all or even most possible Shoshonian localities but are unevenly distributed among the various faunal samples. For these reasons we believe that, although there is clear evidence that a Bridgerian–Uintan transitional interval exists, adoption of a Shoshonian subage may be premature because of the lack of compelling samples from areas other than Agua Fria and especially Bridger E. When such samples do become available and if they corroborate the findings at Agua Fria and Bridger E, then *Shoshonian* would be the appropriate subage name for this interval. As of now, we prefer to recognize an earliest Uintan biochron (Ui1) best represented by the faunal assemblages at the basal Tertiary conglomerate and



Bridger E (Turtle Bluffs Member) while leaving the question of subage status for the Shoshonian open for further testing and corroboration. It may be that more than one faunal unit is located between Br3 and Ui2. The length of time involved in chron C2or indicates that this may be possible.

The beginning of Uintan biochron, Ui1, can be defined by the first appearances of the taxa listed earlier that characterize the beginning of the Uintan. Its ending is defined by several taxa listed later in this chapter that tentatively characterize the beginning of biochron Ui2. Taxa known only from Ui1, and probably the latter part of it, include *Patriolestes*, *Hesperolemur*, *Stockia*, and *Merycobunodon* (all endemic to southern California). Ui1 is further characterized by the presence of certain Bridgerian holdover taxa that apparently do not persist into Ui2, including *Antiacodon*, *Hemiacodon*, *Notharctus*, *Washakius*, and *Hyrachyus*. Strata and faunas assigned to Ui1 include the lowermost Tepee Trail Formation in the East Fork Basin, Wyoming; Bridger E; the upper Sand Wash Fauna from Colorado; and the lower part of the lower member of the Devil's Graveyard Formation in Texas. The fauna from the upper Friars assemblage in California may be late Ui1. Further collecting from Bridger E, Uinta A and B1 (Utah), and the lower and middle units of the Adobe Town Member of the Washakie Formation (Wyoming) and its correlative in the Sand Wash Basin of Colorado is needed to determine whether Ui1 is represented in these strata.

## UINTAN

There is significant disagreement among members of the committee revising this chapter on the definition of the Uintan and its subunits. In part this is caused by the lack of adequate transitional sections and partly by the paucity of good basal Uintan faunal samples. In the Rocky Mountain basins, at least, most of the Bridgerian–Uintan potential or actual boundary sections are in areas of steep exposures where erosion rapidly removes, breaks up, and forces downhill exposed bones. Quarriable sites (Bone Bed A, East Fork Basin; UCM loc. 92189, Bridger E) are limited and not found in sequence above other quarriable sites and often are difficult to access.

Uintan index taxa tentatively include *Achaenodon*, *Amyndodon*, *Diplacodon*, *Eobasileus*, *Leptotragulus*, *Oxyaenodon*, *Oromeryx*, *Prodaphænus*, *Protitanotherium*, *Protoptychus*, *Dolichorhinus* (= *Sphenocoelus*), *Ourayia*, *Procyonictis*, *Metarhinus*, *Mesomeryx*, *Bunomeryx*, *Hylomeryx*, and *Mytonomeryx*; the southern Californian endemic taxa *Patriolestes*, *Dyseolemur*, *Eohaplomys*, *Tapoc-*

*hoerus*, *Merycobunodon*, and a new tapiroid genus (Colbert and Schoch 1998); and the Texas endemic taxon *Prolapsus*.

Uintan first appearances include agriochoerids, *Amyndodon*, lagomorphs, soricids, *Sespedectes*, *Proterixoides*, *Wallia*, *Ankyledon*, *Thylacaelurus*, *Procaprolagus*, *Rapamys*, *Janimus*, *Pareumys*, *Pseudocylindrodon*, *Oligoryctes*, *Ourayia*, *Metanoiamys*, *Protadjidaumo*, *Simimys*, *Griphomys*, *Tapocyon*, *Uintaceras*, *Epitriplopus*, *Colodon*, *Ibarus*, *Tapochoerus*, *Texodon*, *Malaquiferus*, *Diplobunops*, *Simimeryx*, *Mytonomeryx*, *Toromeryx*, *Stockia*, *Craseops*, *Tapomys*, *Eomoropus*, *Grangeria*, *Apriculus*, *Pentacemylus*, *Macrotarsius*, *Chipetaia* (Rasmussen 1996), and Camelidae (Poebrodon).

Uintan last occurrences include Dinocerata, *Hyrachyus*, *Limnocyon*, oxyaenids, *Notharctus*, *Paramys*, *Sciuravus*, taeniodonts, *Heloxyus*, *Scenopagus*, *Macrocranion*, *Entomolestes*, *Ourayia*, *Pantolestes*, *Washakius*, *Hemiacodon*, *Aethomylos*, *Crypholestes*, *Thisbemys*, *Reithroparamys*, *Mysops*, *Pauromys*, *Viverravus*, *Dilophodon*, *Antiacodon*, *Mesonyx*, and *Microsnyops*.

Characteristic Uintan taxa are *Epihippus*, *Protoreodon*, *Amyndodon*, *Eobasileus*, *Metarhinus*, *Dolichorhinus*, *Triplopus*, *Protylopus*, *Mytonolagus*, and *Ischyrotomus*.

The Uintan appears to be subdivisible faunally into early (Ui1), middle (Ui2), and late (Ui3) segments. Ui1 was defined earlier, Ui2 is based on the fauna of Uinta B from the Wagonhound Member, and Ui3 is based on the fauna of Uinta C, from the Myton Member of the Uinta Formation, and now also includes faunas from the lower part of the Brennan Basin Member of the Duchesne River Formation. Based on the magnetic polarity scale, in terms of elapsed time, not faunal development, almost half of the Uintan is represented by Ui1 (Berggren et al. 1995).

The beginning of Ui2 can be tentatively defined by the first appearances of *Bunomeryx*, *Mesomeryx*, and *Protoptychus*, and its ending is defined by the appearance of taxa listed later in this section that characterize the beginning of biochron Ui3. Taxa known only from Ui2 include *Chipetaia*, *Eomoropus*, *Mesomeryx*, and possibly *Protoptychus*, although only the latter two genera are common, and both are known only from the Rocky Mountain region. *Crypholestes* and *Pauromys* have their last occurrences in Ui2. Strata assigned to Ui2 include most of Uinta B2 in Utah and the upper part of the middle Adobe Town Member of the Washakie Formation in Wyoming. Strata potentially assignable to Ui2 include Uinta B1 in Utah, the upper part of the Tepee Trail Formation and the Wiggins Formation in Wyoming, the upper part of the lower member of the Devil's Graveyard Formation of Texas, and

the lower member of the Stadium Conglomerate in California.

Some additional comments on U<sub>11</sub> and U<sub>12</sub> are warranted here. U<sub>11</sub> and U<sub>12</sub> faunas have yet to be demonstrated in a single, superposed section. No distinction between U<sub>11</sub> and U<sub>12</sub> faunas has yet been made in the thick Uintan deposits of the Tepee Trail and Wiggins formations (Eaton 1985). However, the Uintan deposits in Texas may show distinct U<sub>11</sub> (Junction Localities) and overlying U<sub>11</sub> or U<sub>12</sub> (Whistler Squat Quarry) faunal assemblages. Unfortunately, micromammal faunas from Uinta B<sub>1</sub> and the lower part of Uinta B<sub>2</sub> in the Uinta Basin are poorly known, making characterizations of U<sub>11</sub> and U<sub>12</sub> biochrons necessarily tentative.

Krishtalka et al. (1987) list camelids (*Poebrodon*), eomyids, and possibly canids as making first appearances in the late Uintan (U<sub>13</sub>) and also note that limnocyonids decreased in diversity. Other first appearances listed by Krishtalka et al. include *Domnina*, *Thylacaelurus*, *Colodon*, *Prodaphænus*, *Simidectes*, *Procynodictis*, and *Epitriplopus*. Of these, *Colodon* (Eaton 1985), *Poebrodon* (McCarroll et al. 1996a; although Walsh questions this identification), and eomyids (Chiment and Korth 1996; Walsh 1997) are now known from the early Uintan, and canids are known for certain only from the Duchesnean (Bryant 1992). The systematics of *Prodaphænus* are in question, and a new, undescribed species of *Simidectes* is now known from the early Uintan Friars Formation. Other first appearances in U<sub>13</sub> are *Sespedectes*, *Proterixoides*, *Dyseolemur*, *Mytonolagus*, *Rapamys*, *Janimus*, *Protadjidaumo*, *Simimys*, *Griphomys*, *Diplacodon*, *Protitanotherium*, *Epitriplopus*, *Ibarus*, *Tapochoerus*, *Diplobunops*, *Mytonomeryx*, *Malaquiferus*, *Simimeryx*, *Pentacemylus*, and *Toromeryx*. Of these, *Sespedectes*, *Proterixoides*, *Dyseolemur*, *Rapamys*, *Simimys*, and *Tapochoerus* may be very useful in characterizing the U<sub>12</sub>–U<sub>13</sub> boundary.

Taxa making last appearances in U<sub>13</sub> include *Aethomylos*, *Batodontoides*, *Macrocranium*, *Ourayia*, *Microsyops*, *Eohaplomys*, *Sciuravus*, *Mesonyx*, *Oxyaenodon*, “*Proviverra*,” *Limnocyon*, *Tapocyon*, *Epihippus* (Rasmussen, Conroy, et al. 1999), *Hylomeryx*, *Bunomeryx*, and possibly *Auxontodon*.

Other taxa only known from U<sub>13</sub> include *Dyseolemur*, *Eotitanotherium*, *Protitanotherium*, *Epitriplopus*, *Ibarus*, *Tapochoerus*, *Mytonomeryx*, *Laredochoerus*, *Microeutypomys*, *Laredomys*, *Toromeryx*, *Procaprolagus*, *Tapomys*, and *Craseops*.

Strata and faunas assigned to U<sub>13</sub> include the Myton Member of the Uinta Formation and the lower part of the Brennan Basin Member of the Duchesne River Formation in Utah; the upper part of the Wagon Bed For-

mation, Wind River Basin in Wyoming; the Swift Current Creek Local Fauna (L.F.), Cypress Hills Formation in Saskatchewan; the Serendipity L.F. (Devil’s Graveyard Formation), Candelaria (Colmena Formation), and Lake Casa Blanca (Laredo Formation) local faunas in Texas; the lower part of member C of the Santiago Formation, the upper member of the Stadium Conglomerate, the Mission Valley Formation, and the lower two members of the Pomerado Conglomerate, San Diego County, California; and the lower part of the middle member of the Sespe Formation, Ventura County, California.

### UINTAN-DUCHESNEAN BOUNDARY

The nature of the Uintan–Duchesnean transition in the Uinta Basin is unclear, mostly because, according to Kay (1934), there are approximately 320 m of barren strata between the base of the Brennan Basin Member (which yielded the late Uintan assemblage from Randlett Point) and the base of the LaPoint Member (which has yielded the type Duchesnean assemblage). Krishtalka et al. (1987) define the Uintan–Duchesnean boundary on the first appearances of *Duchesneodus*, *Brachyhyops*, *Hyaenodon*, *Simimeryx*, *Poabromylus*, *Hyracodon*, and *Agriochoerus*. However, *Poabromylus* is known from the late Uintan Badwater locality 7 (Black 1978), and a new species of *Simimeryx* is present in the late Uintan Tapo Canyon Local Fauna (Mason 1988). In his review of the Duchesnean problem, Lucas (1992) lists numerous taxa with Duchesnean first appearances but does not explicitly define the Uintan–Duchesnean boundary. We tentatively use the first appearances of *Hyaenodon*, *Duchesneodus*, *Duchesnehippus intermedius*, *Amynodontopsis*, and *Eotylolopus* for this purpose. We also accept the proposals of Lucas (1992) and Rasmussen, Hamblin, and Tabrum (1999) to include the “Halfway Fauna” (now apparently consisting only of *Duchesnehippus intermedius*) in the Duchesnean.

The numerical age of the Uintan–Duchesnean boundary is unclear, again mainly because we do not know where this boundary falls in the Duchesne River Formation. The LaPoint Ash (which forms the base of the LaPoint Member) has been dated at 39.74 Ma ± 0.07 Ma (Prothero and Swisher 1992). The Carnegie Museum *Duchesneodus* Quarry (from which most of the LaPoint Fauna is derived) is located about 37 m above the LaPoint Ash (Kay 1934). However, the fauna of the underlying Dry Gulch Creek Member is largely unknown, and the potential Duchesnean index taxon *Duchesneodus* has been recorded from the base of the Brennan Basin Member (Black and Dawson 1966b; Rasmussen, Hamblin,

and Tabrum 1999). The faunal assemblage from the La-Point Member has generally been suggested to be of middle to later Duchesnean age given the presence of several Chadronian-aspect taxa (Emry 1981; Wilson 1986; Kelly 1990; Prothero and Emry 1996). Therefore the Uintan–Duchesnean boundary could be significantly older than the 39.74 Ma date obtained on the La-Point Ash.

## DUCHESNEAN

Though still far from complete, our knowledge of Duchesnean faunas has increased greatly since the very preliminary characterization of this LMA by Wood et al. (1941). Lucas (1992) reviewed several controversies surrounding the Duchesnean and upheld its validity as a distinct LMA. The Duchesnean is at the heart of a profound middle Eocene faunal replacement in North America (Black and Dawson 1966b) and features an unusual number of first and last appearances but few genera that are restricted to it. Thus the faunal signature of the Duchesnean, instead of being based on the occurrence of typical taxa, is based on the co-occurrence of archaic groups and more advanced genera that came to dominate Chadronian faunas.

The Duchesnean assemblage is all the more difficult to characterize because of the already high and apparently increasing provinciality of the North American fauna at that time (Lillegraven 1979a; Storer 1989, 1996). Typical Duchesnean assemblages and taxa have very different attributes in different faunal provinces (e.g., southern California, west Texas, Gulf Coast Plain, intermontane basins, Great Plains), and it appears that the centers of evolution for most groups were well separated, probably by major physiographic barriers. This combination of provinciality and local evolution makes it exceptionally difficult to correlate assemblages across long distances. However, by the end of the Duchesnean, much of this provinciality disappeared, making Chadronian faunas easier to characterize if not necessarily to correlate.

Lucas (1992) notes that several genera are known only from the Duchesnean, including *Duchesneodus*, *Amyndontopsis*, *Rooneyia*, *Mahgarita*, *Presbymys*, *Viejadjidaumo*, and *Haplohippus*. However, he also notes that with the exception of *Duchesneodus* and possibly *Amyndontopsis*, these taxa are either too rare or too limited in geographic distribution to be regarded as Duchesnean index taxa. Lucas also includes *Protictops*, *Ischnognathus*, and *Hidrosotherium* as taxa unique to the Duchesnean, but *Protictops* is now regarded as a junior synonym of

*Centetodon* (Rasmussen, Hamblin, and Tabrum 1999), *Ischnognathus* is of doubtful taxonomic placement (Gustafson 1986; Lucas 1992), and *Hidrosotherium* is now regarded as a junior synonym of “*Leptomeryx*” (?*Hendryomeryx*) *defordi* (Prothero 1996c).

Other, mostly species-level taxa known only from the Duchesnean include *Simidectes merriami*, *Chumashius balchi*, *Metanoiamys korthi*, *Simimys landeri*, *Simiacritomys whistleri*, *Protoreodon pacificus*, *Protylopus pearsonensis*, and *Simimeryx hudsoni*, all from the Pearson Ranch and Simi Valley Landfill local faunas of the Sespe Formation (Kelly 1990, 1992; Kelly and Whistler 1998), and *Trogolemur leonardi*, *Microparamys nimius*, *Anonymus baroni*, *Pseudotomus timmys*, *Microeutypomys tilliei*, *Eutypomys acares*, *Eutypomys obliquidens*, *Metanoiamys lacus*, *Protadjidaumo pauli*, *Adjidaumo craigi*, *Tachylagus gawneae*, and *Heptacodon pellionis*, all from the Lac Pelletier lower and upper faunas of the Cypress Hills Formation (Storer 1995, 1996).

Excluding the aforementioned species, genera with Duchesnean first appearances include *Sinclairella*, *Apternodus*, *Ischyromys*, *Ardynomys*, *Jaywilsonomys*, *Yoderimys*, *Aulolithomys*, *Heliscomys*, *Hemipsalodon*, *Hesperocyon*, *Daphoenus*, *Mesohippus*, *Trigonias?*, *Hyracodon*, *Subhyracodon?*, *Menops*, *Toxotherium*, *Brachyhyops*, *Heptacodon*, *Agriochoerus*, *Aclistomycter*, *Heteromeryx*, *Pseudoprotoceras*, *Trigenicus*, and *Leptomeryx*.

Excluding the aforementioned species, genera with Duchesnean last appearances include *Apatemys*, *Palaeictops*, *Didelphodus*, *Nyctitherium*, *Talpavus*, *Sespedectes*, *Proterixoides*, *Simidectes*, *Janimus*, *Trogolemur*, *Chumashius*, *Omomys*, *Rapamys*, *Pareumys*, *Simimys*, *Griphomys*, *Mytonomys*, *Hessolestes*, *Harpagolestes*, *Uintacyon*, *Uintasorex (sensu lato)*, *Miocyon*, *Amynodon?*, *Triplopus*, *Protoreodon*, *Leptoreodon*, *Leptotragulus*, *Protylopus*, and *Simimeryx*.

Wilson (1984) proposed an informal subdivision of the Duchesnean into early and late parts. He characterized the early Duchesnean by the presence of certain Uintan holdover taxa (e.g., *Simidectes*, *Harpagolestes*, and *Diplobunops*) and the absence of certain Chadronian-aspect taxa (e.g., *Hemipsalodon*, *Mesohippus*, *Toxotherium*, *Hyracodon primus*, *Brachyops*, and *Merycoidodon*) known from later Duchesnean faunas. Wilson’s general proposal was accepted by Kelly (1990), but Lucas (1992) and Storer (1996) maintained that the Duchesnean record was still inadequate to support a formal subdivision. Nevertheless, we agree with these authors that the relative ages of several Duchesnean assemblages are readily discernible. For example, we agree with Wilson (1986) and Kelly (1990) that the Pearson Ranch

L.F., Badwater Locality 20, and the Skyline Channels are early Duchesnean and that the Porvenir L.F. is late Duchesnean. Prothero and Emry (1996) also propose that the Simi Valley Landfill L.F., the LaPoint Fauna, and the Galisteo Formation assemblage are roughly middle Duchesnean. These suggestions appear reasonable and await corroboration by new fossil discoveries and additional radioisotopic and paleomagnetic work.

The end of the Duchesnean and the beginning of the Chadronian can be characterized by the first appearances of Sciuridae, Nimravidae, Tayassuidae, *Palaeolagus*, *Daphænictis*, *Daphænocyon*, *Penetrigonias*, *Stibarus*, *Bathhygenys*, *Merycoidodon*, *Poebrotherium*, *Montanatylopus*, and *Hypisodus* (Lucas 1992; note that *Heptacodon* and *Pseudoprotoceras* have now been reported from the Duchesnean by Storer 1996 and Eaton et al. 1999). The numerical age of the Duchesnean–Chadronian boundary is estimated at about 37 Ma by Prothero (1996a) and Prothero and Emry (1996) based on dates obtained from the Buckshot Ignimbrite (underlying the late Duchesnean Porvenir L.F.) and the Bracks Rhyolite (overlying the early Chadronian Little Egypt L.F.) in Texas.

## GEOGRAPHIC DISTRIBUTION OF WASATCHIAN-DUCHESNEAN FAUNAL SAMPLES

### WIND RIVER BASIN

**Northwestern Wind River Basin (Figure 4.2)** The oldest fauna in the northwestern Wind River Basin is derived from the upper part of the “lower variegated sequence” (loc. 3, table 4.1) of Love (1947) south of Togwotee Pass in the Fish Creek drainage. Vertebrate assemblages derived from this sequence (McKenna 1980b; Rose 1981) range from Clarkforkian (Red Creek) through Wasatchian (Hardscrabble Creek) as higher levels in the unit are sampled. The presence of *Cantius*, *Hyopsodus*, and *Diacodexis* firmly places the upper part of the “lower variegated sequence” in the early Wasatchian, but the lower part is clearly Clarkforkian, with *Coryphodon*, *Plesiadapis dubius*, *P. cookei*, *Azygonyx ancylion*, and *Arctostylops steini*.

Stratigraphically above the lower variegated sequence is Love’s (1947) “upper variegated unit.” Its fauna, although not well known, is clearly still Wasatchian in age (McKenna 1980b). Overlying the upper variegated sequence is a volcanoclastic sequence (Aycross Formation) that contains the Coyote Creek flora (MacGinitie et al. 1974), fossil land mollusks, and a few mammalian and

other vertebrate remains. The vertebrate fossils are representative of the late Wasatchian or early Bridgerian (McKenna 1980b).

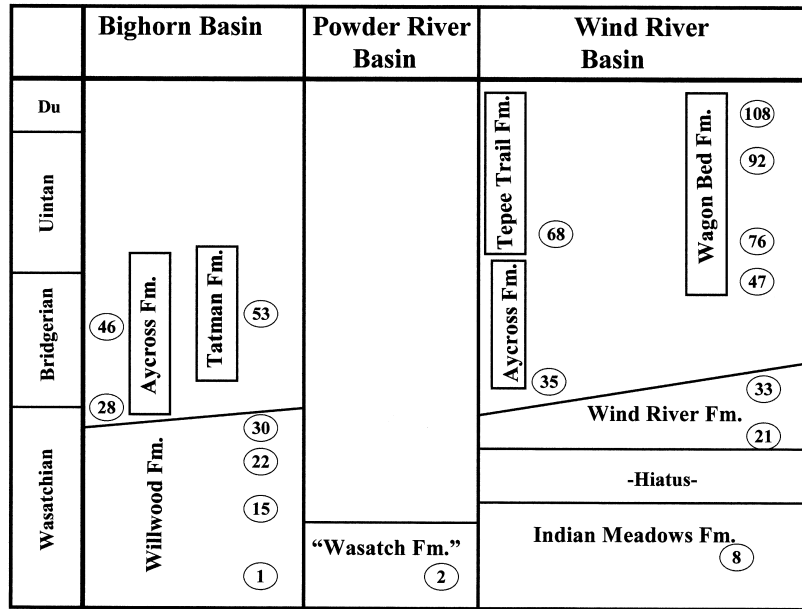
A highly tuffaceous third variegated unit is also present in the northwestern Wind River Basin. This unit and the volcanoclastic sequence beneath it are referred to the Aycross Formation (Love 1939). Locality L-41 (loc. 35, table 4.1) is in these Aycross rocks southwest of the summit of Togwotee Pass, and its small faunal assemblage suggests a latest Wasatchian or early Bridgerian age (McKenna 1980b). Overlying locality L-41 is a greenish volcanoclastic unit, exposed in a small badlands area at the summit of Togwotee Pass (loc. 51, table 4.1), referred to the Aycross Formation. The faunal sample from Togwotee Summit (including *Hyrachyus*, *Palaeosyops*, cf. *Trogosus*, *Microsyops*, *Washakius*, *Tillomys*, *Sciuravus*, *Hyopsodus*, and cf. *Orohippus*) is early Bridgerian (Br2).

Another Wasatchian assemblage is derived from the lower part of the Indian Meadows Formation (Love 1939). This Indian Meadows Fauna (loc. 8, table 4.1) may be Waz to Wa3 in age, based on the presence of *Haplomyilus speirianus* and an *Absarokius*- or *Absarokius*-like omomyid (Winterfeld 1986).

Approximately 64 km east of Togwotee Pass in the East Fork Basin, a distinctive vertebrate assemblage has been quarried from unit 24 (loc. 68, table 4.1; Bone Bed A) of the type section of the Tepee Trail Formation (Love 1939). It contains several apparently endemic taxa (the “dermopteran” *Tarka* and *Hyopsodus lovei*), a new erinaceid also known from the late Uintan of the Badwater area (Krishtalka and Setoguchi 1977), and many characteristic Uintan taxa including eomyid rodents, *Epihippus*, *Dilophodon*, *Amyndodon*, cf. *Tapocyon*, *Achaenodon*, *Uintaceras*, and primitive selenodont and bunodont artiodactyls (McKenna 1980b, 1990; MacFadden 1980; Flynn 1991). This assemblage is tentatively assigned to U<sub>11</sub>, although paleomagnetic evidence obtained by Sundell et al. (1984), Flynn (1986), and Prothero (1996c) suggests that Bone Bed A may correlate with some part of Uinta B<sub>1</sub> (early chron C<sub>20r</sub>). The Tepee Trail Formation below Bone Bed A probably is Br<sub>3</sub> or U<sub>11</sub> in age.

At the southeastern end of the Absaroka Mountains, in the drainage of Owl Creek, Eaton (1980, 1982) and Bown (1982) have collected definitive Bridgerian faunas from the Aycross Formation (loc. 39, table 4.1) and other units. The occurrence of *Scenopagus*, *Microsyops*, *Uintasorex*, *Omomyis*, *Anaptomorphus*, *Washakius*, *Viverravus*, *Mesonyx*, *Hyopsodus paulus*, *Trogosus*, *Orohippus*, *Helohipus*, *Helaletes*, and *Hyrachyus* confirm a Bridgerian age.

Above the Aycross Formation in the drainage of the North Fork of Owl Creek, Eaton (1980, 1985) has docu-



**FIGURE 4.2** Correlation of rock units containing fossil mammals of Wasatchian through Duchesnean (Du) age in northwestern, central, and eastern Wind River Basin, Bighorn Basin, and Powder River Basin. Numerals in ovals refer to localities listed in table 4.1. Fm., Formation.

mented several superposed vertebrate localities (loc. 71, table 4.1) in the Tepee Trail and Wiggins formations. Eaton’s “Holy City beds” and “Foggy Day beds” are of early Uintan age (*Amyrnodon*, *Uintaceras*, *Epihippus*, and *Achaenodon*) and possibly assignable to Uii. The precise ages of Eaton’s “upper Tepee Trail” and “lower Wiggins” are uncertain, but the “upper Wiggins” may be of late Uintan age based on taxa such as *Macrotarsius* sp., cf. *M. siegerti*, cf. *Procyonodictis*, *Colodon*, and *Protoreodon* sp., cf. *P. pumilus* (Eaton 1985). Fossils from referred Wiggins exposures at the southern margin of the Wind River Basin also are Uintan in age (Emry 1975).

**Central and Eastern Wind River Basin (Figure 4.2)** An early Wasatchian fauna has been collected from Shotgun Butte in the Indian Meadows Formation and includes *Hyracotherium*, *Hyopsodus*, *Haplomyilus*, *Coryphodon*, *Diacodexis*, and *Cantius* (Keefer 1965).

The Wind River Formation produces several discrete faunas including a Wa6 assemblage from the Lysite Member (loc. 21, table 4.1), a Wa7 assemblage from the lower part of the Lost Cabin Member (loc. 33, table 4.1), and two assemblages (loc. 33, table 4.1; Bro and Bria) from the upper part of the Lost Cabin Member (Stucky and Krishtalka 1983; Stucky 1984a; Krishtalka and Stucky 1983; Gunnell and Yarborough 2000). The Lostcabinian is

based on the *Lambdaotherium* Range Zone; the Gardnerbuttean, originally defined by the Huerfano B fauna (Bria, late Gardnerbuttean; see P. Robinson 1966), is now also characterized by the *Palaeosyops* Assemblage Zone as defined by Stucky (1984a; Bro, early Gardnerbuttean; see Gunnell and Yarborough 2000).

Faunas from the Badwater area (Black and Dawson 1966a) suggest that *Phenacodus*-bearing locality 17 (loc. 47, table 4.1; West and Atkins 1970) is early Bridgerian in age, as is nearby locality 18 (see also Wood et al. 1936). Localities 5, 5A, 6, and 7 (loc. 92, table 4.1) and associated sites are Ui3 equivalents. Still higher are the Wood and Rodent localities and locality 20 (loc. 108, table 4.1). The last of these is particularly important because it probably represents an earliest Duchesnean assemblage (see Maas 1985 for faunal list).

The nomenclatural treatment of rocks of Bridgerian, Uintan, and Duchesnean ages in the central and eastern Wind River Basin remains unclear. *Wagon Bed Formation* may be the appropriate name for units that contain Bridgerian to Duchesnean faunas, but the upper unit, the Hendry Ranch Member, may be a distinct mappable unit, thus deserving formation rank, or it may be a distal (and later) fine-grained expression of the Wiggins Formation (Emry 1975). The Wagon Bed Formation typically crops out at Beaver Divide in the southern Wind

River Basin where it contains a Uintan fauna (Emry 1975).

### BIGHORN BASIN AND SOUTHWESTERN MONTANA (FIGURE 4.2)

The 770-m Willwood Formation of north-central Wyoming has produced a large series of fossil mammal assemblages (locs. 1, 15, 22, and 30, table 4.1), apparently continuous through Clarkforkian and Wasatchian time. Local disconformities do occur in the sequence. Clarkforkian time is defined on the fauna that appears in both the upper Fort Union Formation and in the lowermost Willwood Formation. Early and middle Wasatchian time (Sandcouleean and Graybullian) is based on assemblages from the Willwood Formation (Van Houten 1945; Gingerich 1980a). Higher beds in the Willwood Formation produce assemblages that correlate with the faunas of the Lysite and Lost Cabin (late Wasatchian) members of the Wind River Formation.

The onset of Wasatchian time is marked by the first appearances of artiodactyls, perissodactyls, euprimates, and hyaenodontid creodonts in the earliest Sandcouleean (Wao). The Graybullian is marked by the first appearance of *Homogalax* (Wa3), and the first appearance of *Heptodon* marks the beginning of the late Wasatchian (= Lysitean, Wa6). The first appearance of *Lambdaotherium* indicates the onset of the Lostcabinian (Wa7). It first occurs at about 650 m above the base of the Willwood Formation and is found up to within 8 m of the Willwood–Tatman formational contact. Local biozonations based on perissodactyls and euprimates (Gingerich 1980b, 1983, 1989, 2001) and *Haplomylus*, *Ectocion*, and *Bunophorus* (Schankler 1980) now define Wasatchian biochrons Wao through Wa7.

The Tatman Formation (loc. 53, table 4.1), which conformably overlies and intertongues with the Willwood Formation in the southwestern part of the Bighorn Basin, was presumed to be Bridgerian in age by Van Houten (1944). D. Parris (Bown 1982) has found late Wasatchian mammals from low in the formation. So-called Tatman equivalents are overlain by and intertongue with the Aycross Formation to the south of Carter Mountain (loc. 46, table 4.1) in the southeastern Absarokas (Bown 1982); the Aycross is of early Bridgerian age, as indicated by the work of Jepsen (1939) and Bown (1979a, 1982).

North of Carter Mountain, the Willwood is overlain unconformably by the Wapiti Formation, a lateral equivalent of the Aycross (loc. 46, table 4.1) and Tepee Trail formations (Bown 1982; Eaton 1982). The Wapiti contains a middle Eocene fauna (Bown 1979a) and is overlain un-

conformably by the Trout Peak Trachyandesite and the Wiggins Formation. A Bridgerian fauna (including *Stylinodon*, *Notharctus*, *Washakius*, *Palaeictops*, *Hyopsodus*, *Phenacodus*, *Orohippus*, cf. *Palaeosyops*, *Hyrachyus*, and *Helohyus*) is known from rocks referred to the Wiggins Formation at Carter Mountain (Eaton 1980, 1982).

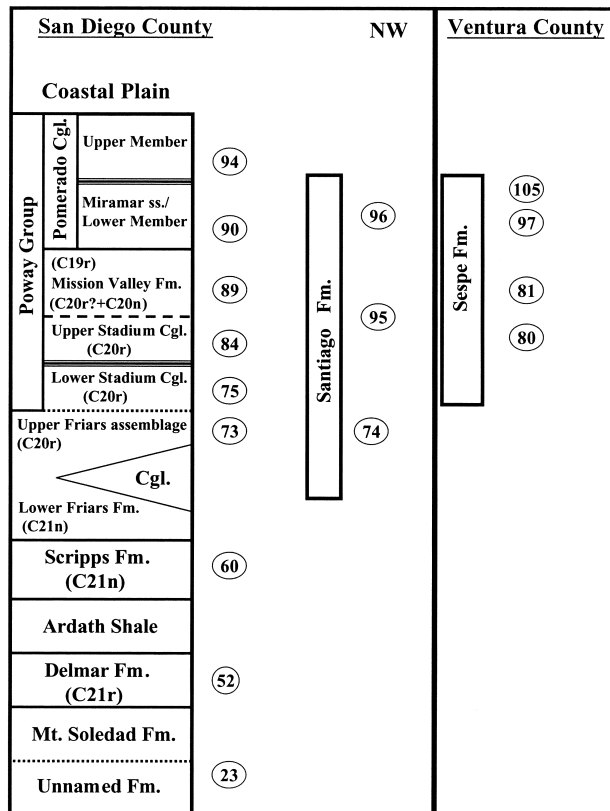
To the west of Cody, Wyoming, along the north and south forks of the Shoshone River, a small vertebrate fauna has been collected from the Willwood and overlying Aycross formations (loc. 28, table 4.1) in Wapiti Valley (Gunnell et al. 1992). This fauna ranges from Lysitean (Wa6) through middle Bridgerian (Br2) and demonstrates that Willwood deposition continued into the Bridgerian along the northwestern margin of the Bighorn Basin.

### SOUTHERN CALIFORNIA (FIGURE 4.3)

Southern Californian Eocene mammal-bearing strata crop out in San Diego and Ventura counties and range from the Wasatchian through at least the late Duchesnean, although most deposits are Uintan. Interdigitations of Uintan strata with fossiliferous marine units permits many southern California sites to be correlated with standard marine reference biochronologies (Bukry and Kennedy 1969; Berggren et al. 1978; Flynn 1986; Bukry 1991; Walsh 1996a). A detailed summary of known occurrences of land vertebrate taxa from deposits in southern California can be found in Golz and Lillegraven (1977), Kelly (1990), Kelly et al. (1991), Walsh (1991a, 1991b, 1996a), and Walsh and Gutzler (1999).

A number of areas in San Diego County have yielded early Cenozoic faunal samples, and revisions of stratigraphy (Walsh 1996a; Walsh et al. 1996) have clarified relationships between rock units and allow documentation of a more complete biostratigraphic framework for this area. The oldest known Wasatchian fossil vertebrates in San Diego County are from an unnamed formation situated between the upper Cretaceous Cabrillo Formation and the Mount Soledad Formation (Kennedy and Moore 1971) and disconformably separated from both. The Morena Boulevard Local Fauna (loc. 23, table 4.1; Walsh 1991a, 1996a) contains typical Wasatchian taxa that probably correlate with the Lysitean or Lostcabinian (Wa6–Wa7) of the western interior (Williamson and Lucas 1992).

The only known Bridgerian aged mammals from California come from the Delmar Formation (Swami's Point L.F., loc. 52, table 4.1; Walsh 1996a). Included in this local fauna are *Hyrachyus* and *Trogosus*, which, along with Flynn's (1986) assignment of the Delmar Formation re-



**FIGURE 4.3** Correlation of rock units containing fossil mammals of Wasatchian through Duchesnean age in southern California. Numerals in ovals refer to localities listed in table 4.1. Cgl., Conglomerate; Fm., Formation; SS, Sandstone.

versed polarity interval to paleomagnetic chron C21r, point to a Br2 correlation.

Overlying the Delmar Formation in turn are the shallow marine Torrey Sandstone, deep marine Ardath Shale, and the shelfal marine Scripps Formation (Kennedy and Moore 1971). The Ardath Shale and Scripps Formation normal polarity interval was correlated with magnetostratigraphic chron C21n by Flynn (1986). Contained in the conglomeratic sandstones in the basal part of the Scripps Formation is the Black's Beach Local Fauna (loc. 60, table 4.1; Walsh 1991a, 1996a). This sample contains cf. *Uin-tatherium* and can be assigned to a late Bridgerian or early Uintan age.

Overlying the Scripps Formation is the Friars Formation (loc. 73, table 4.1), which was informally divided by Walsh et al. (1996) into a lower tongue, a middle conglomerate tongue, and an upper tongue, all apparently of U1i age. The conglomerate tongue of the Friars Formation was previously mapped as the Stadium Conglomerate (Kennedy and Peterson 1975; Kennedy and Peterson 1975), and the upper tongue was mapped by the same au-

thors as Mission Valley Formation. Clarification of these lithostratigraphic relationships resolves the anomaly observed by Golz and Lillegraven (1977) and Novacek and Lillegraven (1979) concerning the very different mammal assemblages obtained from the southern and northern outcrops of the "Mission Valley Formation" (Walsh 1996a; Walsh et al. 1996). The Friars Formation has yielded the well-known early Uintan fauna from San Diego, whose taxa have been described in publications by Lillegraven (1976, 1979b, 1980), Novacek (1976, 1985), Gunnell (1995), and Walsh (1996b, 1997, 1998, 2000) and references therein. The magnetostratigraphic pattern of several Friars Formation sections is difficult to interpret, but lower normal and upper reversed magnetozones in the type area are correlated with C21n and C20r (Flynn 1986; Walsh et al. 1996).

Disconformably overlying the Friars Formation is the lower member of the Stadium Conglomerate, which contains a distinct assemblage (loc. 75, table 4.1; possibly U12) including the oldest known occurrence of *Eohaplomys*, a new species of *Crypholestes*, common specimens of *Pauromyys lillegraveni*, and several taxa found in the Friars Formation (Walsh 1996a, 1997). The reversed polarity interval is correlated with C20r by Walsh et al. (1996).

The upper member of the Stadium Conglomerate disconformably overlies the lower member and contains the late Uintan Stonecrest L.F. (loc. 84, table 4.1; early U13; Walsh 1996a), similar to the late Uintan faunal assemblage from the gradationally overlying Mission Valley Formation (loc. 89; table 4.1). Both faunal assemblages are very different from the well-documented assemblages from the Friars Formation and lower Stadium Conglomerate in that they contain many taxa characteristic of the late Uintan part of the Sespe Formation such as *Sespedectes*, *Proterixoides*, and *Simimys*. The lower normal and upper reversed magnetozones in the type section of the Mission Valley Formation are correlated with paleomagnetic chrons C20n and C19r by Walsh et al. (1996). A <sup>40</sup>Ar/<sup>39</sup>Ar date of 42.83 ± 0.24 Ma was obtained by J. D. Obradovich on a bentonite from the Mission Valley Formation (Walsh 1996a).

Overlying the Mission Valley Formation in the Miramar Reservoir area is the Pomerado Conglomerate, which is divisible into a lower conglomeratic member, the Miramar Sandstone Member (loc. 90, table 4.1), and an upper conglomeratic member (Peterson and Kennedy 1974). The lower two units have yielded late Uintan vertebrate assemblages indistinguishable from those of the Mission Valley Formation. The upper unit (loc. 94, table 4.1) contains a late Duchesnean–early Chadronian assemblage (Walsh and Gutzler 1999), indicating that there is an un-

conformity between the Miramar Sandstone Member and the upper conglomeratic member of the Pomerado Conglomerate that may represent as much as 5 m.y. In the extreme southwestern part of San Diego County, the Pomerado Conglomerate is absent, and the Mission Valley Formation is overlain by reddish mudstones of the "Sweetwater" Formation. The "Sweetwater" contains sparse mammal fossils of latest Uintan to Duchesnean age (Walsh 1991b, 1996a).

The Santiago Formation in northwestern San Diego County has yielded several faunal assemblages. The local member B (Wilson 1972) assemblage (loc. 74, table 4.1) is early Uintan in age and very similar to that obtained from the Friars Formation, although not as diverse. The local member C assemblage (loc. 95, table 4.1), including the Jeff's Discovery and Rancho del Oro L.F., is late Uintan and very similar to that from the Mission Valley Formation, being dominated by *Sespedectes*, *Proterixoides*, *Dyseolemur*, *Simimys*, *Eohaplomys*, *Microparamys*, and several species of *Leptoreodon* and *Protylopus* not present in earlier Uintan assemblages from southern California.

Other Santiago Formation faunas from San Diego County include the Laguna Riviera L.F. (loc. 106, table 4.1; Golz 1976), the Camp San Onofre Locality assemblage (loc. 96, table 4.1; University of California Museum of Paleontology [UCMP] loc. V-72088; Golz and Lillegraven 1977), the San Diego Society of Natural History (SDSNH) loc. 3495 sample, and the Mission del Oro L.F. (Walsh 1996a). All of these faunal samples are of probable late Uintan or earliest Duchesnean age and may correlate best with the Brea Canyon and Strathern local faunas of the Sespe Formation in Ventura County (Kelly 1990; Kelly et al. 1991). The Santiago Formation localities are U13 to early Duchesnean in age.

The Sespe Formation (Kew 1924; Dibblee 1966a, 1966b) is a continental clastic unit of great temporal range, extending from the late Uintan at its base to the early Arikareean and, in some areas, the early Hemingfordian at its top (Donohoo and Prothero 1999). The Sespe crops out in numerous places, but most Eocene vertebrates have been found in the well-studied areas north of Simi Valley in Ventura County. As traditionally understood, the Sespe Formation is a large clastic wedge that is thickest to the east and laterally correlative with several marine formations to the west (Van de Kamp et al. 1974). The formation generally has been viewed as representing a gradual, prolonged, and major westward regression of the sea from the southern California landscape.

In the Simi Valley area, the Sespe Formation rests unconformably on the marine Llajas Formation, which may

be in part coeval with the Delmar Formation and Ardath Shale in San Diego County (Givens and Kennedy 1979), suggesting that the Llajas Formation is temporally equivalent to the late Bridgerian or early Uintan. The Sespe Formation was divided into three members by Taylor (1983): a conglomeratic lower member, an interbedded sandstone and claystone middle member containing vertebrate fossils, and a thick upper member of heterogeneous lithology.

The oldest known vertebrate fossils (loc. 80, table 4.1) so far recovered from the Sespe Formation come from the top of the lower member (Kelly et al. 1991) and appear to be late Uintan in age based on the presence of *Sespedectes singularis* and *Simimys* sp. The well known Tapo Canyon and Brea Canyon local faunas (loc. 81, table 4.1) occur in the basal part of the middle member and are also of late Uintan age (Kelly 1990). The Strathern L.F. (loc. 97, table 4.1) is transitional between the latest Uintan and earliest Duchesnean and was obtained from a stratigraphic interval immediately below the interval that yields the early Duchesnean Pearson Ranch L.F. (loc. 105, table 4.1; Kelly 1990; Kelly et al. 1991). The youngest Eocene assemblage known from the Sespe is the Duchesnean Simi Valley L.F. (loc. 105, table 4.1) from the upper part of the middle member (Kelly et al. 1991).

Northwest of Simi Valley in the Sespe Creek and Pine Mountain areas (Golz and Lillegraven 1977), the Sespe Formation gradationally overlies the Coldwater Formation (Kew 1924; Dibblee 1966a, 1966b). The Coldwater Formation is a mostly marine sandstone unit up to 750 m thick (Prothero and Vance 1996). The Coldwater Formation overlies the Cozy Dell Shale, which in turn overlies the Matilija Sandstone. Based on molluscan faunal samples (Squires 1994) and coccoliths and planktonic foraminifera (Berman 1979) obtained from these formations, the base of the overlying Sespe Formation cannot be older than late Uintan.

The Hartman Ranch L.F. (loc. 97, table 4.1; UCMP loc. V-5814) from upper Sespe Creek is of late Uintan or early Duchesnean age (Lindsay 1968; Kelly 1990; Lander 1994) and was originally reported to occur in the transitional zone between the Sespe and the Coldwater formations. Later, Lander (1994) placed this locality (UCMP V-5814) about 100 feet above the base of the Sespe. Stock (1938) described a brontotheriid from Sespe Creek (loc. Los Angeles County Museum-California Institute of Technology [LACM-CIT] 292) that was later identified as *Duchesneodus* sp., cf. *D. uintensis* by Kelly (1990). LACM-CIT 292 occurs stratigraphically above UCMP loc. V-5814. A specimen of *Amynodontopsis* was discovered by Kelly (1990) from a locality just above LACM-CIT 292. To-

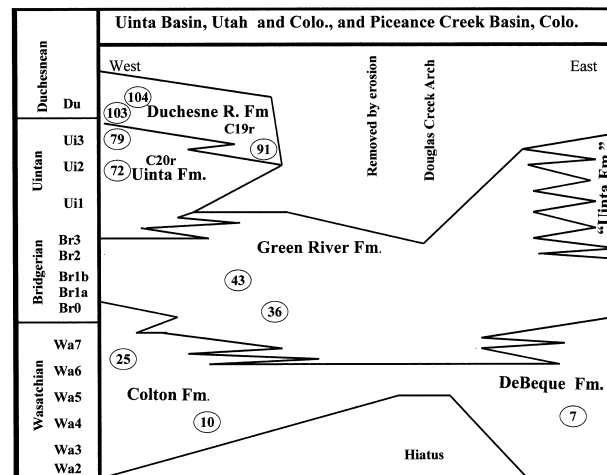


gether these constitute Kelly's Sespe Creek Local Fauna and suggest a Duchesnean age for this faunal level. If the Hartman Ranch L.F. is of Duchesnean age, then the base of the Sespe Formation in the Sespe Creek and Pine Mountain areas is somewhat younger than the base of the Sespe Formation in Simi Valley.

Paleomagnetic correlations for several sections of the Sespe Formation have been proposed by Prothero and Vance (1996) and Prothero et al. (1996). The latter authors correlated a reversed interval yielding the late Uintan Tapo Canyon and Brea Canyon local faunas (loc. 81, table 4.1) to chron C18r and an overlying normal interval yielding the early Duchesnean Pearson Ranch L.F. (loc. 105, table 4.1) to chron C18n. However, the Tapo Canyon and Brea Canyon faunal assemblages are extremely similar to the late Uintan assemblages from San Diego County (Walsh 1996a), suggesting a close temporal correlation. Walsh et al. (1996) correlated the late Uintan faunal assemblages from San Diego County with chron C20n–C19r. These correlations indicate that if both the Walsh et al. (1996) and Prothero et al. (1996) magnetochron assignments are correct, then a difference of about 1.5 m.y. exists between the San Diego and Simi Valley late Uintan assemblages according to the Berggren et al. (1995) time scale. Because a correlation of the Tapo Canyon and Brea Canyon faunas with chron C18r makes them younger than the *Duchesneodus*-bearing basal part of the Duchesne River Formation in Utah (correlated with chron C19n by Prothero 1996b), it seems likely that the correlation of these Ventura County localities with chron C18r may be incorrect.

#### UINTA BASIN AND PICEANCE CREEK BASIN (FIGURE 4.4)

The Wasatchian faunas in these areas are best represented on the southeastern and northern sides of the Piceance Basin (loc. 7, table 4.1) of Colorado and Raven Ridge (loc. 10, table 4.1) on the eastern margin of the Uinta Basin. In a review of the geology and mammalian paleontology of the Debeque Formation (Wasatch Formation of authors) in the Piceance Basin, Kihm (1984) has recorded 124 mammalian species. They can be assigned to faunas of middle Clarkforkian through late Wasatchian age. Additionally, the presence of *Tethyopsis* in the Piceance Creek Basin in the intertongues of the "Uinta" and Green River formations provides evidence of Br3 rocks in that basin that cannot be documented in the Uinta Basin, further illustrating the detailed differences in the basinal histories that are a standard feature of intermountain basins. The Plateau Valley Local Fauna, previously assigned to the Tiffanian, is re-



**FIGURE 4.4** Correlation of rock units containing fossil mammals of Wasatchian through Duchesnean age in Uinta and Piceance Creek basins, Utah and Colorado. Numerals in ovals refer to localities listed in table 4.1. Fm., Formation.

garded by Kihm as mid-Clarkforkian. However, J. Honey (pers. comm. in Kihm 1984) also reported a Tiffanian fauna from the extreme northwestern part of the Debeque Formation. Apparently, much—if not all—of the Wasatchian is represented in the Piceance Basin, although Wao and Wa1 time may be represented in the less fossiliferous Molina Member. Kihm's work corroborates the Wasatchian faunal sequences established in the Bighorn and Wind River basins, although there are some differences in faunal composition. However, certain Wasatchian biochrons, such as Wa5, are much better represented than others. Indeed, Wa5 may be the best represented biochron in the entire region; extensive faunal samples are present in the San Juan, Raton, Piceance Creek, Uinta, Greater Green River, and Bighorn basins, indicating a regional episode of sedimentation. Beds mapped as tongues of the Uinta Formation intertongue with the Green River Formation in the central Piceance Plateau.

Two fossiliferous early Eocene units are present on the southwest side of the Uinta Basin. The Colton Formation, a clastic fluvial floodplain deposit, is considered to be Wasatchian in age because of its intertonguing relationship with the Wasatchian Flagstaff Limestone (Marcantel and Weiss 1968). Fragments of mammals have been recovered by McKenna from the Colton, but no identifiable material is yet available from the southwestern part of the basin. The Flagstaff Limestone has produced one dentary of *Vulpavus australis* (Rich and Collinson 1973).

A series of localities in the Colton (Wasatch Formation of authors) and Green River formations from Raven

Ridge (loc. 10, table 4.1) in the northeastern Uinta Basin, collectively known as the Powder Wash sites (Burke 1935; Kay 1957; Doi 1990), have many Wasatchian assemblages and one rich Bridgerian fauna. The Bridgerian locality (loc. 43, table 4.1) is located 82 m below the Mahogany Oil Shale bed, in the Douglas Creek Member of the Green River Formation (Cashion, written comm. in Dawson 1968) and places the Powder Wash site low in the general Uinta Basin Green River sequence; however, at Raven Ridge the Green River Formation is thinner than elsewhere and it is nearer to the top of the formation as preserved there. Detailed faunal studies for most groups have not been completed; only the marsupials (Krishtalka and Stucky 1984), primates (Gazin 1958; Szalay 1976), rodents (Dawson 1968), an artiodactyl (Burke 1969), and adapisoricid, nyctitheriid, and geolabidid insectivores (Krishtalka 1975, 1976a, 1976b; Lillegraven et al. 1981) have been described. Krishtalka and Stucky (1984) and Gunnell and Bartels (1999) have presented revised faunal lists concluding that this fauna is early Bridgerian and post-Gardnerbuttean in age. Work by Doi (1990) indicates that the lower Green River Formation at Raven Ridge is thick and that the upper (Bridgerian) Powder Wash locality is stratigraphically above the middle of the formation at that locality. The Wa7 and Bro parts of the sequence (loc. 36, table 4.1) are quite thick in comparison with the Wa5 and Wa6 units; this parallels the situation in the eastern part of the Greater Green River Basin (Roehler 1992a). Doi has also found many additional localities in the Raven Ridge area spanning the time between Wa2 and Br2.

The section in the eastern Uinta Basin is not uniform. Near Bonanza, there is a thick sequence of Uinta A that thins dramatically to the north and northeast. At the northwestern end of Raven Ridge on the eastern margin of the Uinta Basin, some 45 km to the north, the underlying Green River Formation is significantly thinner than in the Bonanza area. If the Uinta Formation is present there, it is very thin. Many of the classic Uinta B localities are 5 to 8 km north of Bonanza. North of these localities, the Uinta Formation is much thinner and is overlain by the Duchesne River Formation or covered.

Lacustrine conditions persisted until near the end of the middle Eocene in the Uinta Basin, as recovery of the Duchesnean brontotheriid *Duchesneodus uintensis* from the Green River Formation in Sanpete County, Utah, indicates (Nelson et al. 1980). The younger fluvial formations of the Uinta Basin, the Uinta Formation, and the Duchesne River Formation are the type areas for the two youngest ages (Uintan and Duchesnean) considered in this chapter.

The consensus here is that the faunas from the Wagonhound Member (loc. 72, table 4.1) are primarily Uinta B

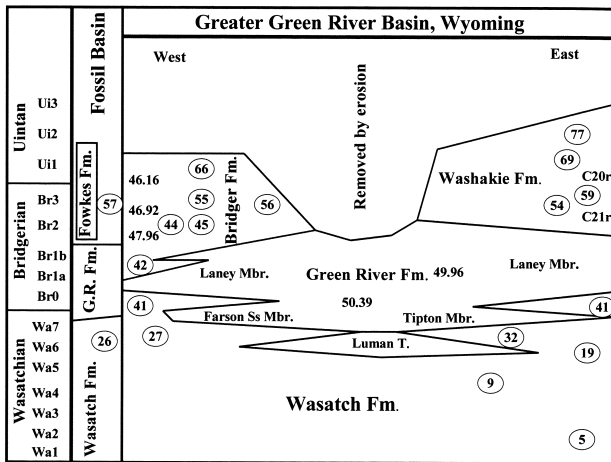
(= Ui2), and those from the Myton Member (loc. 79, table 4.1) are Uinta C (= Ui3). Much more fieldwork and collecting are necessary, especially in the areas of intertonguing of the Green River and Uinta formations (Cashion 1957; Dane 1954) in the region of the Uinta A deposits. This intertonguing occurs southwest of Raven Ridge in areas where faunal remains are very scarce. The intertonguing indicates that rocks present in that area are missing in the hiatus above the Green River Formation further north at Raven Ridge.

The Brennan Basin Member (loc. 91, table 4.1) of the Duchesne River Formation contains the Randlett Fauna; the Halfway Fauna comes from the lower two-thirds of the Dry Gulch Creek Member (loc. 103, table 4.1) of the Duchesne River Formation. These two are considered Ui3 in age, and the faunal names have been abandoned. The fauna from the LaPoint Member (loc. 104, table 4.1) is Duchesnean.

#### EASTERN GREATER GREEN RIVER BASIN (GREAT DIVIDE, WASHAKIE, AND SAND WASH BASINS; FIGURE 4.5)

The Wasatch Formation exposed around the flanks of the Washakie Basin and extensively in the Great Divide Basin (Bradley 1964) has yielded numerous local faunas. These demonstrate the apparent presence of most of Clarkforkian and Wasatchian time as the oldest assemblages appear to be roughly equivalent in age to those from the lowest part of the Willwood Formation, and the youngest are transitional into the Bridgerian. The distribution of these faunas is not uniform, however, and local sedimentologic hiatuses may be present. Two major assemblages come from the lower part of the Wasatch Formation. At the southeastern edge of the basin near the divide with the Sand Wash Basin, in Moffat County, Colorado, are the numerous localities that produce the Four Mile Fauna (loc. 5, table 4.1; McKenna 1960). These faunal samples are equivalent to those from the lower Willwood Formation (McKenna 1960). Recent work in the Four Mile Creek area indicates the presence of a slightly younger Wasatchian fauna above the Four Mile Fauna localities cited earlier (Hill et al. 2000).

In the vicinity of Bitter Creek (loc. 9, table 4.1), along the northern edge of the Washakie Basin, C. L. Gazin (1950s and 1960s), D. Savage and associates (1970s), H. H. Covert and students (1980s and 1990s) and K. C. Beard (1990s) made collections from a series of localities through a continuous sedimentary section from the upper part of the Fort Union Formation to the Cathedral Bluffs Tongue of the Wasatch Formation. Faunal sam-



**FIGURE 4.5** Correlation of rock units containing fossil mammals of Wasatchian through Duchesnean age in the Greater Green River (G.R.) Basin. Numerals in ovals refer to localities listed in table 4.1. Fm., Formation; Mbr., Member; T, Tongue.

ples from these localities span the entire Clarkforkian and the Wasatchian from Wa3 to Wa7 and either Bro or Br1a (P. Holroyd and H. H. Covert, pers. comm., 2000).

Other Washakie Basin and Great Divide Basin Wasatchian localities were cited by Gazin (1962) and have been recollected by recent work of Anemone et al. (2000). Although these are generally small assemblages, they further confirm the age of the Wasatch Formation in the Washakie Basin. Recent work by H. H. Covert in the Bitter Creek area and by P. Robinson (UCM loc. 83120) and J. Honey (1988) on the eastern part of the Washakie Basin confirms the evidence that the Wasatchian–Bridgerian boundary in this area is also in the Cathedral Bluffs member.

The Niland Tongue (= upper Main Body) of the Wasatch Formation along the east side of the Washakie Basin has produced the Dad Local Fauna (loc. 32, table 4.1), which is Wa7 in age. It contains the typical Wa7 perissodactyl *Lambdotherium* and other characteristic late Wasatchian mammals. The Niland Tongue is overlain by the Tipton Tongue and Wilkins Peak members of the lacustrine Green River Formation, and they are overlain by or interdigitate with the uppermost Wasatch Formation subunit, the Cathedral Bluffs Tongue.

The Washakie Formation contains the youngest record of early Tertiary fossil mammals in the Washakie Basin. Biostratigraphic work by Turnbull (1972, 1978), Roehler (1973), and McCarroll et al. (1996a, 1996b) has established the presence of considerable geologic time in the Washakie Formation. Roehler (1973) presented detailed sections for the Washakie Formation and designated two

members, the lower Kinney Rim Member and the upper Adobe Town Member, the latter being divided into lower, middle, and upper parts.

The Kinney Rim Member (loc. 54, table 4.1) of the Washakie Formation contains Bridgerian taxa present in both Br2 and Br3; however, one taxon, *Hyrachyus eximius*, which is found in Br3 and Ui1, occurs near the base of the Kinney Rim Member and indicates that the age of that member is Br3 (McCarroll et al. 1996a). The lowest part of the Adobe Town Member (loc. 59, table 4.1) is Granger’s (1909) Washakie A and is equivalent to Br3. Such taxa as *Notharctus robustior*, *Hemiacodon*, *Stylinodon*, *Tethyopsis*, *Uintatherium*, and *Mesatirhinus* secure this age assignment. The middle Adobe Town Member (loc. 69, table 4.1), Washakie B, of Granger (1909) is either Ui1 or Ui2 in age given the occurrence of *Protoptychus*, *Eobasileus*, *Dolichorhinus*, *Eomoropus*, *Triplopus*, *Amyrnodon*, *Achaenodon*, and *Protylepus*. The highest part of the Washakie Formation (upper Adobe Town Member, loc. 77, table 4.1) may represent another zone not recognized by Granger, who assumed that the summit of Haystack Mountain was the uppermost part of the Washakie Basin section (McCarroll et al. 1996a, 1996b).

The Great Divide Basin has produced a few localities of significance ranging in age from early Wasatchian to early Bridgerian. Recent fieldwork in the area has been carried out by R. Anemone in the southern part of the basin and by G. F. Gunnell and W. S. Bartels in the northwestern part.

The Sand Wash Basin (loc. 63, table 4.1), the southeastern extension of the Greater Green River Basin, contains Washakie Formation rocks (Stucky et al. 1996) equivalent to, and initially continuous with, the lower and middle parts of the Adobe Town Member of the Washakie Formation. The Robin’s Egg Blue Tuff marker is present and aids in correlation with the Washakie Basin. The small upper fauna (West and Dawson 1975; Stucky et al. 1996; D. T. Rasmussen, pers. comm., 2000) is early Uintan (Ui1 or Ui2) (*Triplopus*, *Eobasileus*, *Protoreodon*, *Ourayia*) and includes several Bridgerian holdover taxa such as *Notharctus*.

**WESTERN GREATER GREEN RIVER BASIN (INCLUDING HOBACK AND FOSSIL BASINS) (FIGURE 4.5)**

The Wasatch Formation is exposed along the western and northern margins of the Western Green River Basin (Bradley 1964). In the type area, near Evanston (Fossil Basin) in southwestern Wyoming, there is a small Wa6 mammalian assemblage (loc. 26, table 4.1) that includes

*Heptodon*. Clarkforkian and early Wasatchian mammals have been reported from near LaBarge (loc. 27, table 4.1), on the western side of the Green River Basin (Dorr and Gingerich 1980). Younger Wa7 equivalents have been found at several localities along the western and northern margins of the basin (Gazin 1952, 1962; West 1970, 1973a). The age is confirmed by the presence of *Lambdotherium* in the LaBarge and Cathedral Bluffs (formerly New Fork) faunas (loc. 41, table 4.1) and by the positions of the fossiliferous fluvial rocks relative to the tongues of the Green River Formation.

The Bridger Formation crops out in the southern part of the western Greater Green River Basin. This formation has been famous since its discovery in the 1860s for the abundance and quality of its vertebrate fauna and became the basis for the Bridgerian LMA. Matthew (1909) divided the Bridger Formation into five ascending lithologic units, A to E, and separated them on the basis of lithologic marker beds ("white layers"), many of which are lacustrine. At the time of Matthew's monograph, Bridger A was considered sparsely fossiliferous, Bridger B, C, and D were considered rich, and Bridger E was thought to be barren. Bridger A (loc. 42, table 4.1) is found north of highway Interstate 80 near the town of Opal, Wyoming; Bridger B (loc. 45, table 4.1) crops out extensively between the towns of Lyman and Green River; and Bridger C–D (loc. 55, table 4.1) and E (loc. 66, table 4.1) are found in a more restricted southern area at Sage Creek, Cedar and Hickey Mountains. Outliers of the upper Bridger formation occur north of Farson (Tabernacle Butte). Work by McGrew and Sullivan (1970) and more recently by Gunnell and Bartels (1994) and Gunnell (1998a) has demonstrated that the Bridger A beds (loc. 42, table 4.1) intertongue with the upper units of the Green River Formation and that they have a diverse fauna. Mapping of the Bridger B, C, and D has developed a significant number of stratigraphic markers, allowing more detailed stratigraphic positioning of fossil collections (Evanoff et al. 1998). The Bridger E faunal assemblage is now considered earliest Uintan. Following suggestions in Matthew (1909), UCM parties have accumulated significant microvertebrate samples from lacustrine units in Bridger C and D.

Clyde et al. (2001) place the onset of Bridgerian time at greater than 52 Ma; however, more recent work by M. E. Smith et al. (2003) suggests a date of 50.39 Ma based on the Grey Tuff of the Wilkins Peak member of the Green River Formation. More recent dates on three tuffs from the Bridger Formation (Murphey et al. 1999) indicate that middle Bridgerian was at about 48 Ma and that the Bridgerian ended near 46 Ma. The Bridger Formation

proper may represent only about 60 percent of Bridgerian time (approximately 3.2 m.y.) from Bridger A through Bridger D (Murphey 2001). On the basis of the various referred areas in the South Pass and Bridger Basin, we regard the Bridgerian as possibly lasting as much as 5 million years, from greater than 51 Ma to about 46 Ma.

Isolated areas of Bridgerian rocks crop out in the northern Green River Basin, in downdropped blocks related to the western end of the Continental Fault system (McKenna et al. 1962; West and Atkins 1970; West 1973a; West and Dawson 1973). The oldest Bridgerian assemblage in this region comes from arkosic rocks of the Cathedral Bluffs Tongue of the Wasatch Formation (loc. 41, table 4.1) and seems best placed in Br1a. It is overlain by more typical Bridger Formation rocks that produce a Br1b fauna (loc. 44, table 4.1). Conformably overlying this early Bridgerian sequence are the tuffaceous rocks of Tabernacle Butte (loc. 56, table 4.1) with a late Bridgerian assemblage (McGrew 1959). This particular assemblage is of interest because it contains a late Bridgerian *Phenacodus* and the only multituberculate known from the Bridgerian.

Fossil Basin (loc. 26, table 4.1) is a separate depositional basin a few miles west of the western edge of the Green River Basin proper from near Evanston north to near Kemmerer, Wyoming. Mammalian assemblages of Wasatchian age have been found at several localities (Gazin 1962; Oriel et al. 1962), and the upper assemblages (Wa6 and Wa7) are physically coincident with fish-bearing beds of the Green River Formation. Recently, exposures of the Wasatch and Green River Formations at Fossil Butte National Monument have produced Wa6 and Wa7 vertebrates (Ambrose et al. 1997; Froehlich and Breithaupt 1998).

Nelson (1973, 1974, 1977) collected a suite of Bridgerian mammals from the Fowkes Formation (loc. 57, table 4.1), northwest of Evanston, Wyoming. The Fowkes Formation was initially recognized by Veatch (1907) as the middle part of the Wasatch Group. Oriel and Tracey (1970) have shown that this formation is younger than the Wasatch Formation. Nelson's faunal sample from the Fowkes Formation includes *Notharctus*, *Hemiacodon*, *Omomys*, *Uintasorex*, *Hyopsodus lepidus*, *Orohippus*, and several late Bridgerian ischyromyid rodents.

North of the western Green River Basin is a small physiographic depression, the Hoback Basin. J. A. Dorr and students from the University of Michigan demonstrated virtually continuous sedimentation from Paleocene into medial Wasatchian time (Dorr 1952, 1958, 1969; Dorr and Steidtmann 1971; Dorr et al. 1977). Although the paleontological record from the Wasatch and Pass Peak forma-

tions (loc. 6, table 4.1) is not particularly good, samples (Dorr 1978) are adequate to demonstrate the presence of Clarkforkian and early and middle Wasatchian sites (Dorr and Gingerich 1980).

**WEST TEXAS (FIGURE 4.6)**

Rocks producing fossils of late Wasatchian, Uintan, and Duchesnean ages are present in three areas of West Texas: Big Bend National Park, the Agua Fria area of Brewster County, and the Vieja-Ojinaga area of Presidio County.

The oldest Eocene faunas have been collected from the Hannold Hill Formation (loc. 29, table 4.1) in Big Bend National Park (Schiebout 1974; Rapp et al. 1983; Rose 1981). These faunas include *Phenacodus*, *Phenacolemur*, and two species of *Hyracotherium*, a possible *Lambdotherium*, and several disarticulated skeletons of *Coryphodon* from the Fossil Bone Exhibit locality. These Hannold Hill assemblages are clearly Wasatchian and probably late Wasatchian (Wa7), especially if the record of *Lambdotherium* is valid (but there are several small assemblages from differing levels that have been combined into one “local fauna” for the Hannold Formation, so it is possible that other biochrons of the Wasatchian are also represented).

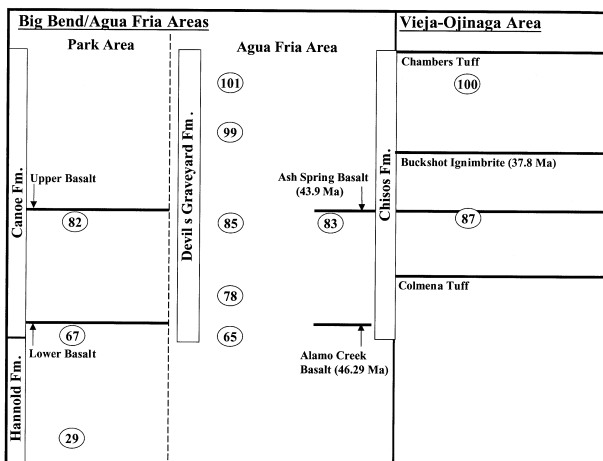
Unconformably overlying the Hannold Hill Formation is the Canoe Formation, which contains two distinct faunal assemblages, Canoe assemblage A and Canoe assemblage B. Canoe assemblage A (loc. 67, table 4.1) is from just below the lower basalt. Canoe assemblage A and an equivalent sample from inside Big Bend National Park (from the Devil’s Graveyard Formation) have a mixed

Bridgerian–Uintan fauna including *Hyrachyus*, *Helohyus*, *Hyopsodus*, *Prolapsus*, *Peratherium*, *Scenopagus*, *Microsycop*, *Uintasorex*, *Omomys*, *Triplopus*, *Amynodon*, *Leptoreodon*, *Microparamys*, *Leptotomus*, and *Pauromys*. Both of these faunal samples can best be interpreted as early Uintan (Ui1), possibly equivalent with the Junction localities (J. A. Wilson 1967 1977; Runkel 1988). Canoe assemblage B (loc. 82, table 4.1) is located beneath the upper basalt in the Canoe Formation. The Canoe assemblage B, along with an equivalent sample from the Chisos Formation, includes *Metamynodon*, *Protoreodon*, *Leptoreodon major*, *Leptoreodon pusillus*, *Leptoreodon edwardsi*, *Uintacyon scotti*, *Epihippus gracilis*, and *Triplopus*. These faunas are equivalent to the Serendipity L.F. from Agua Fria and are late Uintan (Ui3).

North of Big Bend National Park is the Agua Fria area, where a series of superposed localities are found in the Devil’s Graveyard Formation. The earliest occurring faunal assemblage comes from the Junction and 0.6 miles east of Junction localities (loc. 65, table 4.1; basal Tertiary conglomerate; Wilson 1986). These samples include *Herpetotherium*, *Scenopagus*, *Centetodon*, *Nyctitherium*, *Omomys*, *Ourayia*, *Notharctus*, *Microsycop*, *Thisbemys*, *Microparamys*, *Mysops*, *Prolapsus*, *Hyopsodus*, *Uintatherium*, *Stylinodon*, *Hyrachyus*, *Triplopus*, *Helohyus*, *Parahyus*, and *Leptoreodon pusillus*. This assemblage is very similar to that of Bridger E (Evanoff et al. 1994), and we interpret it to represent the earliest Uintan (Ui1).

The Whistler Squat Quarry (loc. 78, table 4.1) occurs above the Junction Locality and has a fauna containing *Amynodon*, “*Sthenodectes*,” *Protoreodon*, *Malaquiferus*, and *Leptoreodon* and is interpreted as Ui1 or Ui2. Above Whistler Squat is the Serendipity L.F. (loc. 85, table 4.1), which appears equivalent to Canoe B and the Chisos Formation sample (Ui3). Higher in the Devil’s Graveyard Formation, above the Ash Spring Basalt, are the Skyline Channels localities (loc. 99, table 4.1). These localities have produced a typical Duchesnean assemblage including *Simidectes*, *Leptotomus*, *Mahgarita*, *Hyaenodon*, *Harpagolestes*, *Amynodontopsis*, *Toxotherium*, *Protoreodon pumilus*, *Agriochoerus*, *Hendryomeryx*, and an unidentified brontotheriid. The Cotter Channel assemblage (loc. 101, table 4.1), found above the Skyline Channels, probably is early Duchesnean and includes *Mytonomys*, *Duchesneodus?*, *Amynodontopsis*, *Protoreodon pumilus*, *Protoreodon petersoni*, *Agriochoerus*, *Hyperragulus*, *Aclistomycter*, and *Eotylopus*. Above these localities are several localities of the Chadronian Coffee Cup L.F.

Approximately 100 km northwest of the park in the Vieja-Ojinaga area is another fossiliferous sequence



**FIGURE 4.6** Correlation of rock units containing fossil mammals of Wasatchian through Duchesnean age in western Texas. Numerals in ovals refer to localities listed in table 4.1. Fm., Formation.

(Wilson 1978; Walton 1992). The lower part of the Vieja Group, which unconformably overlies Cretaceous rocks, includes the fossiliferous Colmena Formation. The Colmena produces the Candelaria L.F. (loc. 87, table 4.1; *Manitsha*, *Pseudotomus*, *Ephippus*, *Sihenodectes*, *Protoreodon*, *Leptoreodon*, and *Toromeryx*), which correlates with the Randlett Fauna (loc. 91, table 4.1) of Utah (Ui3) and with other Texas local faunas including Canoe assemblage B, Chisos Formation, and Serendipity L.F. The Porvenir L.F. (loc. 100, table 4.1) comes from the Chambers Formation, above the Buckshot Ignimbrite (37.8 Ma; Swisher and Prothero 1990). It correlates biostratigraphically with the Montgomery bone bed and is considered late Duchesnean. Above the Porvenir L.F. are found the Chadronian Little Egypt, Airstrip, and Ash Spring localities. Across the Rio Grande in Mexico, the Rancho Gaitan sample appears to correlate with the Chadronian Little Egypt locality.

#### SAN JOSE, HUERFANO, RIO GRANDE RIFT (FIGURE 4.7)

Several separate occurrences of fossil mammals are present in southern Colorado and New Mexico. The most extensive assemblage occurs in the San Jose Formation (loc. 18, table 4.1) of the northern San Juan Basin of northwestern New Mexico. Granger (1914) and Simpson (1948) recognized two main fossiliferous units of the San Jose Formation: the lower Almagre and upper Largo beds or facies. Baltz (1967) divided the San Jose into four members: the Regina Member, including the Almagre beds;

the Tapicitos Member, including the Largo beds; the Llaves Member; and the Cuba Mesa Member. The two main faunal assemblages, one from the Almagre and the other from the Largo, are Wasatchian. Van Houten (1945) and Simpson (1948) regarded them as spanning the Graybullian to early Lysitean, a conclusion corroborated by Lucas et al. (1981), Stucky and Krishtalka (1983), Smith and Lucas (1991), and Lucas and Williamson (1993). These two faunas are not as different as previously thought, and Lucas et al. (1981) suggested abandonment of the Largo–Almagre faunal distinction. For the most part, the San Jose faunas may be Wa6 in age.

The Galisteo Formation of north-central New Mexico (Stearns 1943) is represented by as much as 1300 m of fluvial mudstone, sandstone, and conglomerate (Lucas and Williamson 1993). A small Wasatchian sample (the Cerillos Local Fauna, loc. 24, table 4.1) has been found in the lower part of the Galisteo Formation and includes *Ectoganus*, *Coryphodon*, *Microsypops*, *Hyopsodus powellianus*, *Homogalax*, *Hyracotherium*, and paramyid rodents. The age is indeterminate, but Wa5 or Wa6 is probable. The upper part of the Galisteo Formation has yielded the Tonque L.F. (loc. 111, table 4.1) and includes *Duchesneodus*, *Hemipsalodon grandis*, *Amyrnodon*, *Protoreodon*, *Poabromylus*, brontotheriids, hyracodontids, and protoceratids (Lucas and Williamson 1993), indicating a Duchesnean age. Other scattered fossils from the Galisteo (Galusha 1966; Galusha and Blick 1971) are Eocene in aspect but not definitive of any particular subage.

The Baca Formation may be as much as 1100 m thick and is well exposed in western New Mexico. It has produced small suites of fossil mammals from localities west of the Rio Grande that indicate an age span of Uintan to Chadronian (Schiebout and Schrodtt 1981; Lucas et al. 1981; Lucas and Williamson 1993). The Baca Formation faunal assemblage (locs. 112–113, table 4.1) includes *Hyaenodon*, *Diplacodon*, *Brachyops*, *Protoreodon*, an agriocherid, a protoceratid, and a camelid. The fauna seems to be a mix of Uintan and Duchesnean taxa, but no stratigraphic organization of Baca Formation localities exists, so it is difficult to sort out the precise distribution of these various taxa. Lucas (1990) and Lucas and Williamson (1993) noted the presence of a small brontotheriid metacarpal from the base of the Baca Formation in Cibola County that may indicate that Baca deposition began in the late Bridgerian. The Baca Formation preserves three mammalian trackways in Socorro County (Lucas 1983; Lucas and Williamson 1993) that are interpreted as representing those of artiodactyls.

Lucas and Williamson (1993) recognized a new formation, the Hart Mine Formation, for rocks in the

	E. Colorado	New Mexico	Great Plains
Duchesnean			Saskatchewan (117) South Dakota (109)
Uintan	Northern Raton Basin	Baca Fm. (111, 112, 113)	Saskatchewan (88)
Bridgerian	Huerfano/Farísita Fms. (34, 40, 20)	Hart Mine Fm. (49)	
Wasatchian	Denver Basin (14) Dawson Fm.	Galisteo Fm. (24) San Jose Fm. (18)	North Dakota (11)

**FIGURE 4.7** Correlation of rock units containing fossil mammals of Wasatchian through Duchesnean age in New Mexico and eastern Colorado. Numerals in ovals refer to localities listed in table 4.1. Fm., Formation.

Carthage–La Joya Basin, east of the Neogene Rio Grande rift in Socorro and Valencia counties in central New Mexico. These rocks originally were assigned to the Baca Formation but are lithologically distinct from the Baca Formation west of the rift. A small assemblage of vertebrates from the Hart Mine Formation (loc. 49, table 4.1) includes *Glyptosaurus*, *Telmatherium*, and *Notharctus*. These indicate a Bridgerian age, probably Br<sub>3</sub> (Lucas and Williamson assign the *Notharctus* specimen to *N. tenebrosus* based on tooth size but point out that it could represent a later species of *Notharctus* as well; the presence of *Telmatherium* suggests a later rather than earlier Bridgerian interval).

Other Eocene sediments represented in New Mexico include the Cub Mountain Formation (loc. 112, table 4.1), the Rubio Peak Formation, the Palm Park Formation, and the Love Ranch Formation. There are small vertebrate samples from these formations that indicate ages ranging from the Wasatchian through the Chadronian (Lucas and Williamson 1993).

Well to the north, in the northern Raton Basin of southern Colorado, five superposed assemblages have been collected from the Huerfano Formation (P. Robinson 1966; Stucky 1984b). They correspond to biochrons Wa<sub>5</sub> (loc. 20, table 4.1; Huerfano Locality XIII or UCM loc. 77041; P. Robinson 1963), Wa<sub>6</sub> (loc. 20, table 4.1; Huerfano locs. VIII, IX, and XII; lower part of Huerfano A), Wa<sub>7</sub> (locs. IV, VI, and XI; upper part of Huerfano A), and Bro–Bria (locs. 20 and 40, table 4.1; loc. VII, uppermost part Huerfano A; University of Michigan locs. I, II, III, and V; Huerfano B). As in the Wind River Basin, the Wa<sub>7</sub> biochron is defined by the presence of *Lambdaotherium*. The Gardnerbuttean, first defined by P. Robinson (1966) on the basis of the fauna from Huerfano B (loc. 40, table 4.1), is now also recognized in the Wind River, Green River, and Uinta basins. The Huerfano Formation is at least 1525 m thick in the vicinity of the Spanish Peaks (central Raton Basin), where it unconformably overlies the Paleocene Poison Canyon Formation; scant faunal remains from that area may indicate that pre-Wa<sub>5</sub> sediments occur there as well (P. Robinson 1960). The Farisita Formation (loc. 34, table 4.1; Johnson and Wood 1956) contains sediments that in part intertongue with the Huerfano Formation and in part overlie it (Briggs and Goddard 1956; Berner and Briggs 1958). The only known faunas from the Farisita Formation come from beds of the intertonguing sequence, and these contain both Wa<sub>7</sub> and Bria taxa (P. Robinson 1966). The term *Cuchara Formation* (loc. 14, table 4.1) has been applied to beds that supposedly underlie the Huerfano Formation (Johnson and Wood 1956; Johnson et al. 1958) but that in part are in fact contem-

poraneous with the lower part of the Huerfano Formation (P. Robinson 1966) and from localities that cannot be distinguished from the Huerfano Formation lithologically (P. Robinson 1960, 1963, 1966).

## MISCELLANEOUS LOCALITIES

A number of miscellaneous North American sites produce fossil mammals that are Wasatchian through Duchesnean in age. Most are geologically and geographically isolated from the aforementioned areas and usually are single fossiliferous levels correlative with other areas only biochronologically. Continued discovery of these sites indicates that much is yet to be found in North America.

**Mexico: Baja California** A small collection from the Lomas las Tetas de Cabra (loc. 13, table 4.1) Formation, 30 km south of the village of Punta Prieta, Baja California del Norte, correlates with the Early Eocene Bateque formation (marine). The fauna includes *Hyracotherium*, an endemic marsupial *Esteslestes ensis*, *Esthonyx*, an endemic pantodont, *Meniscotherium*, *Hyopsodus*, and a creodont (Morris 1966; Flynn and Novacek 1984; Novacek et al. 1991). The age of the fauna, originally suggested as Clarkforkian (Morris 1966; Rose 1981), is now thought to be early Wasatchian (Novacek et al. 1991), based primarily on the occurrence of *Wyolestes*. Other elements of the fauna include typical Wasatchian genera such as *Meniscotherium* (Wao–Wa<sub>7</sub>), *Ectocion* (Tiffanian–Wa<sub>7</sub>), *Prolimnocyon* (Wa<sub>3</sub>–Bria), *Diacodexis* (Wao–Wa<sub>7</sub>), *Oxyaena* (Wa<sub>1</sub>–Wa<sub>7</sub>), *Phenacodus* (Tiffanian to Bridgerian), *Hyopsodus* (Clarkforkian–Ui<sub>3</sub>), *Dissacus* (Torrejonian–Wa<sub>4</sub>), and *Esthonyx* (Wao–Bria). There is also a new pantolambdid pantodont, a group usually Paleocene in distribution. The presence of the brontotheriid *Eotitanops* (usually considered early Bridgerian) in the fauna raises some questions because even its late Wasatchian occurrence in Wyoming (Smith and Holroyd 2001) indicates a younger age for the fauna than the other taxa present do. This specimen was not collected at the same time as most of the material cited by Novacek et al., and its provenience is not as certain. Preliminary paleomagnetic study suggests that this sequence can be correlated with either chron C23r or C24n.

**Mexico: Guanajuato** Paleogene vertebrates from Guanajuato, Mexico, at the Marfil locality (loc. 61, table 4.1) have been described by Fries et al. (1955), Black and Stephens (1973), Ferrusquia-Villafranca (1984, 1989). The mammals so far recovered from Marfil include a sciuravid rodent, *Floresomys guanajuatoensis*, two rodents of

uncertain taxonomic placement (*Guanajuatomys hibbardi* and *Marfilomys aewoodi*), fragments of a tapiroid forelimb, a small carnivoran possibly representing *Viverravus*, a hyopsodontid condylarth of uncertain affiliation, and a small palaeonodont. These taxa do not permit a precise age assignment, but a probable Bridgerian–Uintan interval is most likely. A K–Ar date from an andesitic lava flow near the Marfil site produced a date of 49.3 Ma, but the precise stratigraphic relationship between this basalt and the Marfil sediments is not yet known.

**Alaska** A recent discovery represents the only known Paleogene mammalian record from Alaska. In the Matanuska Valley east of Palmer, individual footprints and trackways of mammals and birds have been discovered. Two of the mammalian ichnomorphs represent a perissodactyl and a possible amblypod (K. May, pers. comm., 2001). The trackways were found in the Chickaloon Formation, which spans the Paleocene–Eocene boundary (Triplehorn et al. 1984).

**Golden Valley** The fluvial Golden Valley Formation (loc. 11, table 4.1) conformably overlies the Sentinel Butte Shale Member of the Fort Union Formation in southwestern North Dakota. A collection made by Jepsen (1963) and supplemented by West (1973b) is early Wasatchian in age. The Golden Valley Formation extends well below the mammal-producing level and, if fossiliferous, could yield a series of faunas spanning the Tiffanian through Wasatchian in southwestern North Dakota.

**Powder River Basin** Delson (1971) reported on collections from the Powder River Basin (loc. 2, table 4.1) of northeastern Wyoming. The rocks there are regarded by the U.S. Geological Survey as belonging to the Wasatch Formation, although they are separated from the typical Wasatch Formation of the Greater Green River Basin by the Wind River Basin with its Wasatchian Indian Meadows and Wasatchian/Bridgerian Wind River Formations. The assemblage discussed by Delson is early Wasatchian in age, mostly Wa<sub>2</sub>, with one locality, Monument Blowout (UCM loc. 88052) of Wa<sub>3</sub> age based on the presence of *Homogalax protapirinus*.

Recent work by University of Colorado parties has found that several early Wasatchian faunas are represented in the Powder River Basin (Robinson and Ivy 1994; Robinson and Williams 1998). These faunas are correlative with the Sandcouleean (Wao–Wa<sub>2</sub>) and early Graybullian (Wa<sub>3</sub>) of the Willwood Formation. Reexamination of a deciduous perissodactyl premolar (USNM 187546) from Pumpkin Buttes ascribed to *Lambdotherium*

(Whitmore in Soister 1968; Sharp et al. 1964; Robinson and Ivy 1994) indicates that it is not that taxon and that no Wasatchian sediments younger than Wa<sub>3</sub> definitely occur in the Powder River Basin. The *Homogalax protapirinus* from North Pumpkin Butte cited by Sharp et al. (1964) is the highest fossil occurrence yet documented from the Wasatchian rocks of the Powder River Basin and is a species usually found in Wa<sub>3</sub>–Wa<sub>5</sub> rocks. *Coryphodon* and *Hyacotherium*, also present in the Powder River Basin “Wasatch” Formation (Wegemann 1917; Sharp et al. 1964; Soister 1968), generally indicate only a Wasatchian age. The Powder River Basin sediments are in sharp contrast to the coeval Willwood Formation of the Bighorn Basin because of the large number of lignites present in the former and by the lack of beds definitely younger than Wa<sub>3</sub>. The Powder River sequence may well represent a much more paludal environment.

**Laramie and Shirley Basins** A small mammalian fauna from rocks in the Cooper Creek area (loc. 16, table 4.1) referred to the Wind River Formation 40 km northwest of Laramie, Wyoming, was studied by Prichinello (1971). The fauna includes *Cantius*, *Tetonius*, *Haplomytus*, *Hyopsodus*, *Phenacodus*, *Coryphodon*, and *Hyacotherium* and is early Wasatchian in age.

In the Shirley Basin, north of the Laramie Basin, Harshman (1972) mapped both the Wind River and Wagon Bed formations. Fossils found in the Wind River Formation are characteristic of the Wasatchian (Harshman 1972) but cannot be assigned more precisely. The Wagon Bed conformably overlies the Wind River in the Shirley Basin; a specimen of *Notharctus tenebrosus* (Harshman 1972) suggests an early Bridgerian age for the Wagon Bed Formation there.

**New Jersey** An isolated lower molar of the tillodont *Anchippodus* (?*Trogosus*) was collected from the marine Shark River Marls of northeastern New Jersey. The tooth is at the same stage of evolutionary development as those of western Bridgerian genera (Gazin 1953).

**Canada: British Columbia** Two teeth referable to the Bridgerian tillodont *Trogosus* have been collected at a coal mine near the town of Princeton (Russell 1935; Gazin 1953).

An assemblage from the Kishenehn Formation (loc. 116, table 4.1) in the Flathead Valley of southeastern British Columbia includes the type material of *Thylacaelurus* and also citations of *Peratherium*, *Pseudocylindrodon*, *Protadjidaumo*, *Paradjidaumo*, *Desmatolagus*, and *Leptotragulus* (Russell 1954). The Kishenehn Fauna was



originally thought to be Duchesnean (Russell 1954), but Lucas (1992) considered that the assemblage was more likely to be Chadronian. It is difficult to be certain considering the number of taxa now known.

A brontotheriid lower molar from the Australian Creek beds along the Fraser River at Quesnel was interpreted by L. S. Russell (figured in McAnally 1996) as Chadronian. It has been used to correlate a critical palynofloral sequence with the Eocene–Oligocene boundary (Ridgway et al. 1995). This area should be restudied because the modest size of the specimen in question raises the distinct possibility that it might be Uintan or Duchesnean.

**Canada: Saskatchewan** In the Cypress Hills Formation, Swift Current Plateau (loc. 88, table 4.1) of southwestern Saskatchewan, a diverse mammalian fauna has been collected by the Canadian Museum of Nature (Russell and Wickenden 1933), the Royal Ontario Museum (Russell 1965), and the Royal Saskatchewan Museum (Storer 1978, 1984; Krishtalka 1979). The presence of *Procaprolagus*, *Miocyon*, *Auxontodon*, *Colodon*, *Epihippus*, *Protoreodon*, and *Leptoreodon* confirms a late Uintan (U<sub>13</sub>) age.

Two superposed Duchesnean local faunas (loc. 117, table 4.1; Lac Pelletier Lower Fauna, Lac Pelletier Upper Fauna) occur in the Cypress Hills Formation, also on the Swift Current Plateau (Storer 1996). Both would be interpreted by most authors as late Duchesnean because of a group of first appearances of typically Chadronian genera including *Sinclairiella*, *Hesperocyon*, *Heptacodon*, *Pseudoprotoceras*, and *Trigenicus*. Lucas (1992) has suggested that this assemblage actually is Chadronian. On the other hand, *Palaeictops*, *Didelphodus*, *Apatemys*, *Talpavus*, *Sespedectes*, *Nyctitherium*, *Uintasorex*, three omomyid primates, *Miocyon*, *Janimus*, and *Microeutypomys* are also present in these samples and are not known from any Chadronian faunas. Storer (1996) hesitated to assign these assemblages to either early or late Duchesnean, noting only that they appear older than the latest Duchesnean Porvenir Local Fauna of Texas (Wilson 1986).

No Duchesnean assemblages are documented from the Cypress Hills Plateau, although the Duchesnean index taxon, *Duchesneodus primitivus* (Lambe 1908; Lucas and Schoch 1989), comes from an undetermined locality there.

**Canadian Arctic** Vertebrates from several localities at 79°–80° north latitude on Ellesmere Island (Dawson et al. 1976; West and Dawson 1978; McKenna 1980a) and Axel Heiberg Island (Dawson et al. 1993), Nunavut Territory, Canada, suggest a Wasatchian and possible early Bridger-

ian age. These fossils occur in the Eureka Sound Group (loc. 37, table 4.1), the formations within which are differently named and divided by Miall (1986) and Ricketts (1986). As currently known, the assemblages do not correlate readily with any particular southern fauna, and the high diversity of plagiomenids is their most striking peculiarity.

Slightly higher in the Eureka Sound Group of Axel Heiberg Island, the beds preserving the Geodetic Hills (loc. 98, table 4.1) mummified forest appear to be Uintan or Duchesnean, based on fragments of brontotheriid teeth found there (Eberle and Storer 1999).

**Oregon** The Clarno Formation has two mammal-producing levels. The lower level, the Nut Bed (loc. 58, table 4.1), has produced a small assemblage (*Patriofelis*, *Orohippus*, *Telmatherium*, and *Hyrachyus*) indicative of the Bridgerian. A stratigraphically higher locality, Hancock Quarry (loc. 110, table 4.1), has produced *Epihippus* and *Diplobunops*, generally regarded as Uintan, but other taxa (*Hemipsalodon*, *Haplohippus*, *Protapirus*, cf. *Procadurcodon*, and *Caenopus*) are more usually considered Duchesnean or Chadronian.

**South Dakota** The Slim Buttes Formation (loc. 109, table 4.1) of northwestern South Dakota contains *Duchesnehippus intermedius*, *Duchesneodus*, *Colodon*, *Amyndontopsis*, an indeterminate agriochoerid, and an indeterminate leptotraguline (Bjork 1967). A Duchesnean age is assignable to this small assemblage (Lucas 1992), which is similar to the LaPoint Fauna of Utah.

**Virginia** Rose (1999) has recently reported on a small fauna from the Fisher/Sullivan site, Nanjemoy Formation (loc. 17, table 4.1), which may well be middle Wasatchian (Wa<sub>3</sub>–Wa<sub>5</sub>) in age. Included in this fauna are isolated teeth of a probable pantolestid, a miacid carnivoran, an esthonychid tillodont, a hypsodontid condylarth, an isctolophid perissodactyl, an undetermined rodent, and a single dentary containing three teeth of a possible nyctitheriid insectivoran.

**Mississippi** The Tuscahoma Formation of Lauderdale County (loc. 4, table 4.1) has yielded a fauna of 25 species of land mammals and other vertebrates (Red Hot L.F. of Beard et al. 1995 and Beard and Dawson 2001). This assemblage probably is Wa<sub>1</sub> in age, based on the co-occurrence of *Ectocion parvus* and *Haplomyilus speirianus* (Thewissen 1990; Robinson and Williams 1998). On the basis of associated marine fossils, Beard et al. (1995) correlated the Red Hot L.F. with nannoplankton zone NP<sub>9</sub>, which straddles

the Paleocene–Eocene boundary (Berggren et al. 1995). The Bashi Formation (loc. 12, table 4.1) overlies the Tuscahoma Formation and has produced an omomyid primate of probably Wasatchian age (Beard and Tabrum 1990).

A single brontotheriid skull, the holotype of *Notiotitanops mississippiensis*, was found in Clark County, Mississippi, in strata previously referred to the Lisbon Formation (Gazin and Sullivan 1942). The locality is now referred to the Archusa Marl Member of the middle Eocene Cook Mountain Formation (Westgate 1986). The skull is morphologically closest to several Uintan brontotheriids of the Utah–Wyoming region (Mader 1989), and the locality is best considered late Uintan in age (Westgate, pers. comm., 2002).

**Alabama** Westgate (2001) cites a single specimen of *Coryphodon* from the Hatchetigbee Bluff L.F. (loc. 31, table 4.1) in the Hatchetigbee Formation, Wilcox Group, in Washington County. *Coryphodon* occurs from the Clarkforkian to earliest Bridgerian (Br1a); the location of this specimen argues for a Wasatchian age.

Another Alabama locality with a single mammalian specimen is the Little Stave Creek L.F. (loc. 62, table 4.1) in the Gosport Sand, Claiborne Group, in Clarke County (Westgate 2001). This locality has produced the miacid carnivore *Vulpavus*, indicating a Wasatchian or Bridgerian age.

**Arkansas** Westgate (2001) notes the presence of *Diplobunops* and *Pantolestes* in the Jackson Group (loc. 64, table 4.1) of St. Francis County. These two genera are not usually found together. *Pantolestes* normally is restricted to the Bridgerian and early Uintan (although probable sister taxa survive into the Chadronian in North America) and *Diplobunops* to the late Uintan (Ui3).

**Texas Coastal Plain** A diverse late Uintan vertebrate assemblage (loc. 86, table 4.1; Casa Blanca L.F.) has been collected from strata of the Laredo Formation in Webb County, Texas (Westgate 1988, 1990, 1994c, 1999; Wilson and Westgate 1991; Walton 1993a, 1993b). Marine invertebrates in the Laredo Formation permit a correlation with the Cook Mountain Formation, which contains nannoplankton assigned to zone NP16 (late medial Eocene; Berggren et al. 1995). A bentonite in the Cook Mountain Formation has been dated at  $42.0 \pm 0.8$  Ma (Berggren et al. 1992).

Westgate (1988, 1994a–1994c, 2001) summarizes the Lake Casa Blanca L.F. (loc. 86, table 4.1) from near Laredo in Webb County. This diverse vertebrate assemblage con-

tains many Uintan mammals and is correlated with Ui3 and the Serendipity L.F. of the Big Bend area. The age assignment is derived from the artiodactyls present at Lake Casa Blanca and the presence of the rodent *Mytonomys*.

**Montana** There are several small faunal samples from isolated areas in Montana representing the Bridgerian through Duchesnean. The earliest of these is from the type area (loc. 38, table 4.1) of the Sage Creek Formation, Beaverhead County, Montana, and includes *Helalestes*, *Hyrachyus*, *Eotitanops*, and *Palaeosyops*, indicating a Bridgerian age, perhaps even early Bridgerian (Bro–Br1a; Wallace 1980; Tabrum et al. 1996). Two successive late Uintan local faunas, the Douglass Draw L.F. and the Hough Draw L.F. (loc. 93, table 4.1), are located in the “Dell beds” stratigraphically above the Sage Creek Formation (Tabrum et al. 1996).

Two local faunas are known from the Climbing Arrow Formation near Three Forks, in the Beaverhead Basin. Shoddy Springs (G. E. Robinson 1963; Black 1967; G. E. Robinson et al. 1957; Krishtalka 1979; Krishtalka and Black 1975; Lillegraven and Tabrum 1983) is a Duchesnean locality (loc. 107, table 4.1) tentatively correlated with Locality 20 (loc. 108, table 4.1) in the Wagon Bed Formation, Wind River Basin. An additional latest Duchesnean or earliest Chadronian sample from the Diamond O L.F. (loc. 118, table 4.1) was reported by Tabrum et al. (1996).

## CORRELATION AND CALIBRATION

### GEOCHRONOLOGY

During the last two decades of magnetostratigraphic and radioisotopic research, significant progress has been made toward placing the NALMA record into a chronostratigraphic framework that can be readily correlated to the global time scale. Table 4.2 gives a summary of the most recent reliable geochronologic constraints on the Wasatchian through Duchesnean NALMA record. Unfortunately, several intervals remain poorly sampled and controversial. The following is a brief discussion of the key information that constrains the NALMA record in time, with particular attention to intervals that are still characterized by uncertainty.

No global stratotype section and point (GSSP) has yet been determined for the Paleocene–Eocene boundary, so the position of the Clarkforkian–Wasatchian NALMA boundary with respect to the Paleocene–Eocene boundary remains ambiguous (Aubry 2000). However, it is clear

**TABLE 4.2** Geochronology of North American Land Mammal Ages (NALMAs)

CALIBRATION POINT	AGE	BIOCHRON	NALMA SUBAGE	NALMA	LOCATION	REFERENCE	COMMENTS
Paleocene-Eocene isotope excursion	54-96	Wao	Sandcouleean	Wasatchian	Bighorn Basin	Wing et al. 2000	Age corroborated by Norris and Rohl 1999
Chron C24r-C24n.3n	53-35	Wa4-Wa5	Graybullian	Wasatchian	Bighorn Basin	Clyde et al. 1994 Tauxe et al. 1994	
Chron C24n.3n-C24n.2r	52-90	Wa6	Lysitean	Wasatchian	Bighorn Basin	Clyde et al. 1994 Tauxe et al. 1994	
Chron C24n.2r-C24n.2n	52-80	Wa6	Lysitean	Wasatchian	Bighorn Basin	Clyde et al. 1994 Tauxe et al. 1994	
Chron C24n.2n-C24n.1r	52-76	Wa6	Lysitean	Wasatchian	Bighorn Basin	Clyde et al. 1994 Tauxe et al. 1994	
<sup>40</sup> Ar/ <sup>39</sup> Ar-sandine	52.8 ± 0.3 (WM)	Wa7	Lostcabinian	Wasatchian	Bighorn Basin	Wing et al. 1991	Upper Willwood Formation
<sup>40</sup> Ar/ <sup>39</sup> Ar-sandine	52.6 (TG)	Wa7	Lostcabinian	Wasatchian	Bighorn Basin	Wing et al. 1991	Upper Willwood Formation
Chron C24n.1r-C24n.1n	52-66	Wa7	Lostcabinian	Wasatchian	Bighorn Basin	Tauxe et al. 1994	
Chron C24n.1n-C23r	52-36	Wa7	Lostcabinian	Wasatchian	Green River Basin	Clyde et al. 1997, 2001	
Chron 23r-C23n.2n	51-74	Bria	Gardnerbuttean	Bridgerian	Green River Basin	Clyde et al. 1997, 2001	
Chron C23n.2n-C23n.1r	51-05	Bria	Gardnerbuttean	Bridgerian	Green River Basin	Clyde et al. 1997, 2001	
Chron C23n.1r-C23n.1n	50-95	Bria	Gardnerbuttean	Bridgerian	Green River Basin	Clyde et al. 1997, 2001	
Chron 23n.1n-C22r	50-78	Bria	Gardnerbuttean	Bridgerian	Green River Basin	Clyde et al. 1997, 2001	
Chron C22r-C22n	49-71	Brib	Bridger A	Bridgerian	Green River Basin	Clyde et al. 1997, 2001	
Chron C22n-C21r	49-04	Br2	Bridger B	Bridgerian	East Fork Basin	Flynn 1986	
<sup>40</sup> Ar/ <sup>39</sup> Ar-sandine/ biotite/plagioclase/ hornblende	47.96 ± 0.13 (WM)	Br2	Bridger B	Bridgerian	Green River Basin	Murphey et al. 1999	Bridger Formation
Chron C21r-C21n	47-91	Br2	Bridger B	Bridgerian	Washakie Basin East Point Basin	Flynn 1986	
<sup>40</sup> Ar/ <sup>39</sup> Ar-sandine/ biotite/plagioclase/ hornblende	46.92 ± 0.17 (WM)	Br3	Bridger C	Bridgerian	Green River Basin	Murphey et al. 1999	Bridger Formation

TABLE 4.2 (continued)

CALIBRATION POINT	AGE	BIOCHRON	NALMA SUBAGE	NALMA	LOCATION	REFERENCE	COMMENTS
<sup>40</sup> Ar/ <sup>39</sup> Ar-sanidine	46.29 ± 0.05	Uii?	"Shoshonian"	Uintan	Trans-Pecos Texas	Henry et al. 1986	Below Alamo Creek Basalt Junction L.F.
Chron C21n-C20r	46.26	Uii?	"Shoshonian"	Uintan	Washakie Basin Uinta Basin Southern California	Flynn 1986 Prothero 1996b Walsh 1996b	Correlation A of McCarrroll et al. 1996a, Washakie Formation
<sup>40</sup> Ar/ <sup>39</sup> Ar-sanidine/ biotite/plagioclase/ hornblende	46.16 ± 0.44 (WM)	Uii?	Bridger E	Uintan	Green River Basin	Murphey et al. 1999	Bridger Formation
Chron C20r-C20n	43.79	Uj3	Uinta A/B	Uintan	Uinta Basin	Prothero and Swisher 1992	
<sup>40</sup> Ar/ <sup>39</sup> Ar-sanidine	42.83 ± 0.24 Uj2		Uinta C	Uintan	Southern California	Obradovich in Walsh 1996b	Mission Valley Formation
Chron C20n-C19r	42.54	Uj3	Uinta C	Uintan	Uinta Basin	Prothero and Swisher 1992	
Chron C19r-C19n	41.52	Uj3	Uinta C	Uintan	Uinta Basin	Prothero 1996b	Brennan Basin Member, Duchesne River Formation
Chron C19n-C18r	41.26	Uj3	Uinta C	Uintan	Uinta Basin	Prothero 1996b	Brennan Basin Member, Duchesne River Formation
Chron C18r-C18n.2n	40.13	Uj3/Du	Uinta C/Duchesnean	Uintan/Duchesnean	Texas Southern California	Prothero 1996c Prothero et al. 1996	
Chron C18n.2n-C18n.1r	39.63	Uj3/Du	Uinta C/Duchesnean	Uintan/Duchesnean	Texas Southern California	Prothero 1996c Prothero et al. 1996	
<sup>40</sup> Ar/ <sup>39</sup> Ar-biotite	39.74 ± 0.07	Du		Duchesnean	Uinta Basin	Prothero and Swisher 1992	LaPoint Tuff
Chron C18n.1r-C18n.1n	39.55	Du		Duchesnean	Texas	Prothero 1996c	
Chron C18n.1n-C17r	38.43	Du		Duchesnean	Texas New Mexico	Prothero 1996c Prothero and Lucas 1996	
Chron C17r-C17n.3n	38.11	Du		Duchesnean	Texas New Mexico	Prothero 1996c Prothero and Lucas 1996	
Chron C17n.3n-C17n.2r	37.92	Du		Duchesnean	Texas New Mexico	Prothero 1996c Prothero and Lucas 1996	

TABLE 4.2 (continued)

CALIBRATION POINT	AGE	BIOCHRON	NALMA SUBAGE	NALMA	LOCATION	REFERENCE	COMMENTS
Chron C17n.2r-C17n.2n	37.85	Du		Duchesnean	Texas New Mexico	Prothero 1996c Prothero and Lucas 1996	
<sup>40</sup> Ar/ <sup>39</sup> Ar-sanidine	37.8 ± 0.35	Du		Duchesnean	Texas	Prothero and Swisher 1992	Buckshot Ignimbrite
Chron C17n.2n-C17n.1r	37.60	Du		Duchesnean	Texas New Mexico	Prothero 1996c Prothero and Lucas 1996	
Chron C17n.1r-C17n.1n	37.47	Ch		Chadronian	Texas	Prothero 1996c	
<sup>40</sup> Ar/ <sup>39</sup> Ar-sanidine	36.7 ± 0.07	Ch		Chadronian	Texas	Prothero and Swisher 1992	Bracks Rhyolite, confirmed by Henry et al. 1994

Ages of magnetic reversals are from the most recent Geomagnetic Polarity Time Scale (Cande and Kent 1995). L.F., Local Fauna; TG, total gas; WM, weighted mean.

that the Clarkforkian–Wasatchian boundary correlates with a global carbon isotope excursion (CIE) that occurs within chron C24r and has been dated at approximately 55.5 Ma (Koch et al. 1992; Wing et al. 2000; Rohl et al. 2000). Although the 55.5 Ma age for the CIE conflicts with one of the calibration points for the most recent GPTS (Cande and Kent 1995; see discussion in Berggren and Aubry 1998) and therefore is not included in table 4.2, the age of the CIE changes only slightly when it is estimated independently of the GPTS (Wing et al. 2000, model 2). This CIE represents a profound global biogeochemical event that lasted only 200,000 years (or less) and therefore represents a potentially powerful point for intracontinental and intercontinental correlations of mammalian biostratigraphies.

Studies from the Bighorn Basin (Butler et al. 1981; Clyde et al. 1994; Tauxe et al. 1994), Baja California (Flynn et al. 1989), and West Texas (Rapp et al. 1983) confirm that the Wasatchian LMA extends from late chron C24r to at least early chron C23r. Although there is some uncertainty as to the placement of the C24r–C24n.3n boundary with respect to the Wa4–Wa5 faunal zone boundary (Clyde et al. 1994; Tauxe et al. 1994), research in progress should resolve this soon. Reanalysis of the magnetostratigraphy from the Eureka Sound Group in Ellesmere Island provided no support for previous suggestions of large-scale faunal and floral latitudinal diachroneity during Wasatchian time (Tauxe and Clark 1987; see also Hickey et al. 1983; Kent et al. 1984).

The age of the Wasatchian–Bridgerian NALMA boundary is less clear. It was traditionally placed at ~50 Ma in chron C22r (Krishtalka et al. 1987:91–95; Prothero 1995; Clyde et al. 1997, Correlation 2), but new studies in the Green River Basin suggest that the boundary may be significantly older, in chron C23r at ~52 Ma (Clyde et al. 1999, 2001; Murphey et al. 1999). More recent work by M. E. Smith et al. (2003) indicates that the boundary is near the Grey Tuff, dated at  $50.39 \pm 0.13$  Ma.

The geochronology of the Bridgerian–Uintan NALMA boundary is also poorly constrained at present because of contradictory evidence from well-sampled sections in the Washakie Basin, Bighorn Basin, Uinta Basin, Bridger Basin, southern California, and Texas that span this interval (see discussions in Flynn 1986; Prothero 1996b; McCarroll et al. 1996a; Stucky et al. 1996; Walsh 1996b, 2001). Despite various magnetobiostratigraphic inconsistencies with these sections, it is generally agreed that rocks characterized by Bridgerian mammalian assemblages extend into chron C21n; rocks characterized by Uintan mammalian assemblages correlate to chron C20r and C20n and above and may occur in C21n (Flynn 1986; Walsh et al.

1996; Prothero 1996b). Resolving the detailed chronostratigraphy of the intervening interval that includes the Bridgerian–Uintan NALMA boundary (and the proposed Shoshonian subage) will entail further fieldwork.

Magnetostratigraphic and radioisotopic data from the Uinta Formation in Utah, mixed volcanic and sedimentary strata from Trans-Pecos Texas, and the Coldwater Sandstone/Sespe Formation in California all suggest that the Uintan NALMA extends into chron C18r (Prothero 1996b, 1996c; Prothero and Vance 1996; Prothero et al. 1996). An earliest Duchesnean fauna from California (Pearson Ranch Local Fauna) is known from a normal interval correlated to chron C18n, indicating that the Uintan–Duchesnean boundary lies somewhere in chron C18 (Prothero et al. 1996). This correlation is further supported by radioisotopic dates and magnetostratigraphy from Trans-Pecos Texas (Prothero 1996c). The Texas sections also provide the best evidence for constraining the timing of the Duchesnean–Chadronian boundary. A late Duchesnean local fauna (Porvenir Local Fauna) lies just above the Buckshot Ignimbrite, dated at  $37.8 \pm 0.15$  Ma ( $Ar^{39}/Ar^{40}$ ; Prothero and Swisher 1992). An early Chadronian fauna (Little Egypt Local Fauna) from the Vieja area of Trans-Pecos Texas immediately underlies the Bracks Rhyolite, dated at  $36.7 \pm 0.07$  Ma ( $Ar^{39}/Ar^{40}$ ; Prothero and Swisher 1992; Henry et al. 1994). The intervening interval is characterized by predominantly normal polarity and was correlated to chron C17n by Prothero (1996c). This suggests that the Duchesnean–Chadronian NALMA boundary lies in chron C17n at about 37 Ma.

## POSTSCRIPT

After having reflected on what has been presented here, the two principal authors of this chapter (G.G. and P.R.) feel it is appropriate to look back on what has been accomplished since the last edition of this volume and what lies ahead. It is clear that a great deal has been learned since 1987 about the NALMAs that make up the subject of this chapter. Yet there is also a lingering feeling that much more must be learned before a true understanding of the intricacies of this important time period will be realized. The problems that remain, though intellectual in nature, can be addressed only by additional field exploration using the most advanced technology available to fieldworkers.

It is clear that we have a much better understanding of the biochronologic history of the Wasatchian and Bridgerian than we did in 1987. However, we still don't know just how applicable the biochronologies developed

in the Bighorn and Green River basins will be outside these type areas. The biochronologic frameworks of the Uintan and Duchesnean are still in their infancy and await much better records not only from the type areas of these land mammal ages but also from other areas that will affect our understanding of each of these time periods. It is still not clear that similar biochronologic approaches will be equally applicable to each of these land mammal ages, and we almost certainly will have to adapt as our knowledge becomes greater. We are getting a much better understanding of intercontinental relationships but are also raising many questions that need to be addressed. Additional fieldwork in areas that can add to our knowledge of these events must be undertaken. Much of our considerations of this are east–west focused. Perhaps we should also consider more north–south possibilities.

We believe that the issues surrounding the recognition and placement of the Paleocene–Eocene boundary will be resolved in the near future, perhaps even before this volume appears in print. Other issues will take more time and effort. As befits transitional periods, the Wasatchian–Bridgerian, Bridgerian–Uintan, and Uintan–Duchesnean will remain contentious for the foreseeable future. We believe that the Wasatchian–Bridgerian transition has come into focus recently, yet we still do not understand the mechanisms of this change particularly well. There is tantalizing evidence of the possible existence of an additional recognizable subage at the Bridgerian–Uintan boundary, yet the evidence is far from compelling. The Uintan–Duchesnean transition remains shrouded in mystery, as does the very nature of these two land mammal ages.

Much has been accomplished since 1987, yet not all questions have been answered and not all problems solved. With new information come new questions. Renewed efforts are needed to address the issues raised by what has been presented in this chapter. We look forward to seeing what the next 15 years will bring.

#### ACKNOWLEDGMENTS

*The original chapter by Krishtalka et al. (1987) benefited from the assistance of C. Castelli, C. Costello, G. Curtis, J. A. Dorr Jr., J. Eaton, P. Gingerich, B. Hanson, E. A. Hill, J. H. Hutchison, A. Kihm, C. A. Knox, J. D. Love, S. Lucas, E. Manning, M. Mason, R. Mauger, J. Obradovich, N. Perkins, P. Robinson, W. Rohrer, K. Rose, D. E. Savage, J. Schiebout, S. Speerbrecher, J. Wahlert, and J. A. Wilson.*

*This second version was aided significantly by information and advice from R. L. Anemone, W. S. Bartels, S. A. Bilbey, J. I. Bloch, R. J. Burnham, H. H. Covert, E. Evanoff, J. J. Flynn, P. D. Gingerich, P.*

*Holroyd, L. Ivany, L. D. Ivy, S. G. Lucas, K. May, K. M. Muldoon, P. C. Murphey, D. T. Rasmussen, K. D. Rose, A. Tabrum, M. D. Uhen, and J.-P. Zonneveld.*

#### REFERENCES

- Ambrose, P., W. S. Bartels, G. F. Gunnell, and E. M. Williams. 1997. Stratigraphy and vertebrate paleontology of the Wasatch Formation, Fossil Butte National Monument, Wyoming. *Journal of Vertebrate Paleontology* 17(suppl. 3):29A.
- Andersen, D. W. and M. D. Picard. 1972. Stratigraphy of the Duchesne River Formation (Eocene–Oligocene), northern Uinta Basin, northeastern Utah. *Bulletin of the Utah Geological Mineralogical Survey* 97:1–29.
- Anemone, R. L., D. J. Over, B. A. Nachman, and J. Harris. 2000. A new late Wasatchian mammalian fauna from the Great Divide Basin, Sweetwater County, Wyoming. *Journal of Vertebrate Paleontology* 20(suppl. 3):26A.
- Aubry, M.-P. 1996. Towards an upper Paleocene–lower Eocene high resolution stratigraphy. *Israel Journal of Earth Sciences* 44:239–253.
- . 1998. Stratigraphic (dis)continuity and temporal resolution in the upper Paleocene–lower Eocene deep sea record. In *Late Paleocene–early Eocene climatic and biotic events in the marine and terrestrial records*, ed. M.-P. Aubry, S. G. Lucas, and W. A. Berggren. New York: Columbia University Press, pp. 37–66.
- . 2000. Where should the global stratotype section and point (GSSP) for the Paleocene/Eocene boundary be located? *Bulletin de la Société Géologique de France* 17:461–476.
- . 2001. *Annual report 2001 of the Paleocene/Eocene Boundary stratotype working group*. Unpublished report of the International Commission on Paleogene Stratigraphy.
- Baltz, E. H. 1967. *Stratigraphy and regional tectonic implications of part of Upper Cretaceous and Tertiary rocks, east-central San Juan Basin, New Mexico*. U.S. Geological Survey Professional Paper 552.
- Beard, K. C. and M. R. Dawson. 2001. Early Wasatchian mammals from the Gulf Coast Plain of Mississippi: Biostratigraphic and paleobiogeographic implications. In *Eocene biodiversity: Unusual occurrences and rarely sampled habitats*, ed. G. F. Gunnell. New York: Kluwer/Plenum, Topics in Geobiology Series, pp. 75–94.
- Beard, K. C., M. R. Dawson, and A. R. Tabrum. 1995. First diverse land mammal fauna from the Early Cenozoic of the southeastern United States: The early Wasatchian Red Hot local fauna, Lauderdale County, Mississippi. *Journal of Vertebrate Paleontology* 15(suppl. 3):18A.
- Beard, K. C. and A. R. Tabrum. 1990. The first Early Eocene mammal from eastern North America: An omomyid primate from the Bashi Formation, Lauderdale County, Mississippi. *Mississippi Geology* 11:1–6.
- Berggren, W. A. and M.-P. Aubry. 1998. The Paleocene/Eocene epoch/series boundary: Chronostratigraphic framework and estimated geochronology. In *Late Paleocene–early Eocene climatic and biotic events in the marine and terrestrial records*, ed. M.-P. Aubry, S. G. Lucas, and W. A. Berggren. New York: Columbia University Press, pp. 18–36.

- Berggren, W. A., D. V. Kent, J. D. Obradovich, and C. C. Swisher. 1992. Toward a revised Paleogene geochronology. In *Eocene–Oligocene climatic evolution*, ed. D. R. Prothero and W. A. Berggren. Princeton, NJ: Princeton University Press, pp. 29–45.
- Berggren, W. A., D. V. Kent, C. C. Swisher, and M.-P. Aubry. 1995. A revised Cenozoic chronology and chronostratigraphy. In *Geochronology, time scales and global stratigraphic correlations: A unified temporal framework for an historical geology*, ed. W. A. Berggren, D. V. Kent, and J. Hardenbol. Society of Economic Paleontologists and Mineralogists Special Publication 54.
- Berggren, W. A., M. C. McKenna, J. Hardenbol, and W. Obradovich. 1978. Revised Paleogene polarity time scale. *Journal of Geology* 86:67–81.
- Berman, B. H. 1979. *Biostratigraphy of the Cozy Bell Formation in Ventura and Santa Barbara counties*. M.S. thesis, California State University, Long Beach.
- Berner, R. and L. I. Briggs. 1958. Continental Eocene sedimentation in Huerfano Park, Colorado. *Bulletin of the Geological Society of America* 69:1533.
- Billbey, S. A., E. Hall, and D. Hall. 2002. Old and new vertebrate sites in the Uinta Basin, Utah. *Journal of Vertebrate Paleontology* 22(suppl. 3):36A.
- Bjork, P. R. 1967. Latest Eocene vertebrates from northwestern South Dakota. *Journal of Paleontology* 41:227–236.
- Black, C. C. 1967. Middle and late Eocene mammal communities: A major discrepancy. *Science* 156:62–64.
- . 1969. Fossil vertebrates from the late Eocene and Oligocene Badwater Creek area, Wyoming, and some regional correlations. *Wyoming Geological Association Guidebook, 21st Annual Field Conference*, pp. 43–48.
- . 1978. Geology and paleontology of the Badwater Creek area, central Wyoming. Pt. 14: The artiodactyls. *Annals of the Carnegie Museum* 47:223–259.
- Black, C. C. and M. R. Dawson. 1966a. Paleontology and geology of the Badwater Creek area, central Wyoming. Part 1. History of fieldwork and geological setting. *Annals of the Carnegie Museum* 38:297–307.
- Black, C. C. and M. R. Dawson. 1966b. A review of late Eocene mammalian faunas from North America. *American Journal of Science* 264:321–349.
- Black, C. C. and J. J. Stephens III. 1973. Rodents from the Paleogene of Guanajuato, Mexico. *Occasional Papers of the Museum of Texas Tech University* 14.
- Bown, T. M. 1979a. Correlation of Eocene volcanoclastic rocks, southeastern Absaroka Range in northwestern Wyoming. *U.S. Geological Survey Professional Paper* 1150:68–69.
- . 1979b. Geology and mammalian paleontology of the Sand Creek facies, lower Willwood Formation (lower Eocene), Washakie County, Wyoming. *Geological Survey of Wyoming Memoir* 2.
- . 1982. Geology, paleontology, and correlation of Eocene volcanoclastic rocks, southeast Absaroka Range, Hot Springs County, Wyoming. *U.S. Geological Survey Professional Paper* 1201-A.
- Bown, T. M. and A. J. Kihm. 1981. *Xenicohippus*, an unusual new hyracotheriine (Mammalia, Perissodactyla) from lower Eocene rocks of Wyoming, Colorado, and New Mexico. *Journal of Paleontology* 55(1):257–270.
- Bown, T. M. and K. D. Rose. 1984. Reassessment of some early Eocene Omomyidae, with a description of a new genus and three new species. *Folia Primatologica* 43:97–112.
- Bradley, W. H. 1964. Geology of Green River Formation and associated Eocene rocks in southwestern Wyoming and adjacent parts of Colorado and Utah. *U.S. Geological Survey Professional Paper* 496-A.
- Briggs, L. I. and E. N. Goddard. 1956. Geology of Huerfano Park Colorado. *Rocky Mountain Association Geology Guidebook, Geology of the Raton Basin, Colorado*, pp. 40–45.
- Bryant, H. N. 1992. The Carnivora of the Lac Pelletier lower fauna (Eocene, Duchesnean), Cypress Hills Formation, Saskatchewan. *Journal of Paleontology* 66:847–855.
- Bukry, D. 1991. Transoceanic correlation of middle Eocene coccolith subzone CP14a at Bataquitos Lagoon, San Diego County. In *Eocene geologic history, San Diego region*, ed. P. L. Abbott and J. A. May (eds.). *Pacific Section SEPM* 68:189–193.
- Bukry, D. and M. P. Kennedy. 1969. Cretaceous and Eocene coccoliths at San Diego, California. *California Division of Mines and Geology Special Report* 100:33–43.
- Burke, J. J. 1935. Preliminary report of fossil mammals from the Green River Formation in Utah. *Annals of the Carnegie Museum* 25:13–14.
- . 1969. An antiacodont from the Green River Eocene of Utah. *Kirtlandia* 5:1–7.
- Butler, R. F., P. D. Gingerich, and E. H. Lindsay. 1981. Magnetic polarity stratigraphy and biostratigraphy of Paleocene and lower Eocene continental deposits, Clark's Fork Basin, Wyoming. *Journal of Geology* 89:299–316.
- Cande, S. C. and D. V. Kent. 1995. Revised calibration of the geomagnetic polarity time scale for the Late Cretaceous and Cenozoic. *Journal of Geophysical Research* 100:6093–6095.
- Cashion, W. B. 1957. Stratigraphic relations and oil shale of the Green River Formation in the eastern Uinta Basin. *Intermountain Association of Petroleum Geologists Guidebook, 8th Annual Field Conference*, pp. 131–135.
- . 1974. *Geologic map of the Southam Canyon Quadrangle, Uintah County, Utah*. U.S. Geological Survey Miscellaneous Field Studies Map MF-579.
- . 1986. *Geologic map of the Bonanza Quadrangle, Uintah County, Utah*. U.S. Geological Survey Miscellaneous Field Studies Map MF-1865.
- Cashion, W. B. and J. R. Donnell. 1974. Revision of nomenclature of the upper part of the Green River Formation, Piceance Creek Basin, Colorado, and eastern Uinta Basin, Utah. *Bulletin of the U.S. Geological Survey* 1394-G.
- Chiment, J. J. and W. W. Korth. 1996. A new genus of eomyid rodent (Mammalia) from the Eocene (Uintan–Duchesnean) of southern California. *Journal of Vertebrate Paleontology* 16:116–124.
- Clark, J., J. R. Beerbower, and K. E. Kietzke. 1967. Oligocene sedimentation, stratigraphy, paleoecology and paleoclimatology in the Big Badlands of South Dakota. *Fieldiana: Geology Memoir* 5.
- Clyde, W. C., N. D. Sheldon, and P. L. Koch. 1999. New paleomagnetic and stable-isotope results from the Green River Basin (South Pass, Wyoming): Geological, climatic and biotic interpretations. *Geological Society of America Abstracts with Programs* 31:A416.
- Clyde, W. C., N. D. Sheldon, P. L. Koch, G. F. Gunnell, and W. S. Bartels. 2001. Linking the Wasatchian/Bridgerian boundary to the Cenozoic global climate optimum: New magnetostratigraphic and isotopic results from South Pass, Wyoming. *Palaeogeography, Palaeoclimatology, Palaeoecology* 167:175–199.



- Clyde, W. C., J. Stamatikos, and P. D. Gingerich. 1994. Chronology of the Wasatchian land-mammal age (early Eocene): Magnetostratigraphic results from the McCullough Peaks section, northern Bighorn Basin, Wyoming. *Journal of Geology* 102:367–377.
- Clyde, W. C., J.-P. Zonneveld, J. Stamatikos, G. F. Gunnell, and W. S. Bartels. 1997. Magnetostratigraphy across the Wasatchian–Bridgerian (early to middle Eocene) in the western Green River Basin, Wyoming. *Journal of Geology* 105:657–669.
- Colbert, M. W. and R. M. Schoch. 1998. Tapiroidea and other morphomorphs. In *Evolution of Tertiary mammals of North America*. Vol. 1, *Terrestrial carnivores, ungulates and ungulate-like mammals*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, pp. 569–582.
- Comstock, T. B. 1875. *Report upon the reconnaissance of northwestern Wyoming including Yellowstone National Park, for 1873, by Wm. A. Jones*. House Report Executive Documents no. 285, 43d Congress, 1st session, January 1875.
- Cope, E. D. 1877. Report upon the extinct Vertebrata obtained in New Mexico by parties of the expedition of 1874. *Geographical Survey West of the 100th Meridian*, pt. 2.
- . 1882. Contributions to the history of the Vertebrata of the lower Eocene of Wyoming and New Mexico made during 1881. *Proceedings of the American Philosophical Society* 20:139–197.
- Covert, H. H., P. Robinson, and J. R. Harris. 1998. Evidence for two lineages of *Notharctus* during Bridger C and D. *Journal of Vertebrate Paleontology* 18(suppl. 3):36A.
- Crawford, T., D. T. Rasmussen, and G. Conroy. 2002. Forelimb morphology and limb proportions of middle Eocene *Oxyaenodon* (Mammalia, Creodonta). *Journal of Vertebrate Paleontology* 22(suppl. 3):46A.
- Dane, C. H. 1954. Stratigraphic and facies relationships of upper part of Green River Formation and lower part of Uinta Formation in Duchesne, Uintah, and Wasatch counties, Utah. *American Association of Petroleum Geologists Bulletin* 38:405–425.
- Dawson, M. R. 1968. Middle Eocene rodents (Mammalia) from northeastern Utah. *Annals of the Carnegie Museum* 39:327–370.
- Dawson, M. R., M. C. McKenna, K. C. Beard, and J. H. Hutchison. 1993. An early Eocene plagiomenid mammal from Ellesmere and Axel Heiberg Islands, Arctic Canada. *Kaupia-Darmstädter Beiträge zur Naturgeschichte* 3:179–192.
- Dawson, M. R., R. M. West, W. Langston Jr., and J. H. Hutchison. 1976. Paleogene terrestrial vertebrates: Northernmost occurrence, Ellesmere Island, Canada. *Science* 192:781–782.
- Delson, E. 1971. Fossil mammals of the early Wasatchian Powder River local fauna, Eocene of northeast Wyoming. *Bulletin of the American Museum of Natural History* 146:305–364.
- Dibblee, T. W. Jr. 1966a. *Geologic map of the Wheeler Springs Quadrangle, Ventura County, California*. Dibblee Geological Foundation Map no. DF-01.
- . 1966b. Geology of the central Santa Ynez Mountains, Santa Barbara County, California. *California Division of Mines and Geology Bulletin* 186.
- Doi, K. 1990. *Geology, and paleontology of two primate families of the Raven Ridge, northwestern Colorado and northeastern Utah*. Unpublished M.S. thesis, University of Colorado, Boulder, pp. 1–215.
- Donohoo, L. and D. R. Prothero. 1999. Magnetic stratigraphy of the lower Miocene Sespe–Vaqueros formations, Orange County, California. *Journal of Vertebrate Paleontology* 19(suppl. 3):42A (abstract).
- Dorr, J. A. Jr. 1952. Early Cenozoic stratigraphy and vertebrate paleontology of the Hoback Basin, Wyoming. *Bulletin of the Geological Society of America* 63:59–94.
- . 1958. Early Cenozoic vertebrate paleontology, sedimentation, and orogeny in central western Wyoming. *Bulletin of the Geological Society of America* 69:1217–1244.
- . 1969. Mammalian and other fossils, early Eocene Pass Peak Formation, central western Wyoming. *University of Michigan, Contributions from the Museum of Paleontology* 22:207–219.
- . 1978. Revised and amended fossil vertebrate faunal lists, early Tertiary, Hoback Basin, Wyoming. *University of Wyoming, Contributions to Geology* 16:79–84.
- Dorr, J. A. Jr. and P. D. Gingerich. 1980. Early Cenozoic mammalian paleontology, geologic structure and tectonic history in the overthrust belt near LaBarge, western Wyoming. *University of Wyoming, Contributions to Geology* 18(2):101–115.
- Dorr, J. A. Jr., D. R. Spearing, and J. R. Steidtmann. 1977. Deformation and deposition between a foreland uplift and an impinging thrust belt: Hoback Basin, Wyoming. *Geological Society of America Special Paper* 177.
- Dorr, J. A. Jr. and J. R. Steidtmann. 1971. Stratigraphic–tectonic implications of a new, earliest Eocene, mammalian faunule from central western Wyoming. *Michigan Academician* 3:25–41.
- Douglass, E. 1914. Geology of the Uinta Formation. *Bulletin of the Geological Society of America* 25:417–420.
- Eaton, J. G. 1980. Preliminary report on paleontological exploration of the southeastern Absaroka Range, Wyoming. In *Early Cenozoic paleontology and stratigraphy of the Bighorn Basin, Wyoming*, ed. P. D. Gingerich. *University of Michigan Papers on Paleontology* 24:139–142.
- . 1982. Paleontology and correlation of Eocene volcanic rocks in the Carter Mountain area, Park County, southeastern Absaroka Range, Wyoming. *University of Wyoming, Contributions to Geology* 21(2):153–194.
- . 1985. Paleontology and correlation of the Eocene Tepee Trail and Wiggins formations in the North Fork of Owl Creek area southeastern Absaroka Range, Hot Springs County, Wyoming. *Journal of Vertebrate Paleontology* 5:345–370.
- Eaton, J. G., J. H. Hutchison, P. A. Holroyd, W. W. Korth, and P. M. Goldstrand. 1999. Vertebrates of the Turtle Basin local fauna, middle Eocene, Sevier Plateau, south-central Utah. In *Vertebrate paleontology in Utah*, ed. D. D. Gillette. Utah Geology Survey Miscellaneous Publications 99-1:463–468.
- Eberle, J. J. and J. E. Storer. 1999. Northernmost record of bronotheres, Axel Heiberg Island, Canada. *Journal of Paleontology* 73:979–983.
- Emry, R. J. 1975. Revised Tertiary stratigraphy of the western Beaver Divide, Fremont County, Wyoming. *Smithsonian Contributions to Paleobiology* 25.
- . 1981. Additions to the mammalian fauna of the type Duchesnean, with comments on the status of the Duchesnean “Age.” *Journal of Paleontology* 55:563–570.
- Evanoff, E., L. Brand, and P. C. Murphey. 1998. Bridger Formation (middle Eocene) of southwestern Wyoming: Widespread marker units and subdivisions of Bridger B through D. *Dakoterra* 5:115–121.

- Evanoff, E., P. Robinson, P. C. Murphey, D. G. Kron, D. Engard, and P. Monaco. 1994. An early Uintan fauna from Bridger E. *Journal of Vertebrate Paleontology* 14(suppl. 3):24A.
- Ferrusquia-Villafranca, I. 1984. Review of the early and middle Tertiary mammal faunas of Mexico. *Journal of Vertebrate Paleontology* 4:187–198.
- . 1989. A new rodent genus from central Mexico and its bearing on the origin of the Caviomorpha. In *Papers on fossil rodents in honor of Albert Elmer Wood*, ed. C. C. Black and M. R. Dawson. *Los Angeles County, Museum of Natural History, Series in Science* 33:91–117.
- Flynn, J. J. 1986. Correlation and geochronology of Middle Eocene strata from the western United States. *Palaeogeography, Palaeoclimatology, Palaeoecology* 55:335–406.
- . 1991. *Hyopsodus* (Mammalia) from the Tepee Trail Formation (Eocene), northwestern Wyoming. *American Museum Novitates* 3007:1–19.
- Flynn, J. J., R. M. Cipolletti, and M. J. Novacek. 1989. Chronology of early Eocene marine and terrestrial strata, Baja California, Mexico. *Geological Society of America Bulletin* 101:1182–1196.
- Flynn, J. J. and M. J. Novacek. 1984. Early Eocene vertebrates from Baja California: Evidence for intracontinental age correlations. *Science* 224:151–153.
- Fries, C. Jr., C. W. Hibbard, and D. H. Dunkle. 1955. Early Cenozoic vertebrates in the red conglomerate at Guanajuato, Mexico. *Smithsonian Miscellaneous Collections* 123(7).
- Froehlich, D. J. and B. H. Breithaupt. 1998. Mammals from the Eocene Epoch Fossil Butte Member of the Green River Formation, Fossil Basin, Wyoming. *Journal of Vertebrate Paleontology* 18(suppl. 3):43A.
- Galusha, T. 1966. The Zia Sand Formation, new early to medial Miocene beds in New Mexico. *American Museum Novitates* 2271.
- Galusha, T. and J. C. Blick. 1971. Stratigraphy of the Santa Fe group, New Mexico. *Bulletin of the American Museum of Natural History* 144:1–128.
- Gardner, J. H. 1910. The Carthage Coal Field, New Mexico. *Bulletin of the U.S. Geological Survey* 381:452–460.
- Gazin, C. L. 1952. The lower Eocene Knight Formation of western Wyoming and its mammalian faunas. *Smithsonian Miscellaneous Collections* 117(8).
- . 1953. The Tillodontia: An early Tertiary order of mammals. *Smithsonian Miscellaneous Collections* 121(10).
- . 1955. A review of the upper Eocene Artiodactyla of North America. *Smithsonian Miscellaneous Collections* 128(8).
- . 1958. A review of the middle and upper Eocene primates of North America. *Smithsonian Miscellaneous Collections* 136(1).
- . 1962. A further study of the lower Eocene mammalian faunas of southwestern Wyoming. *Smithsonian Miscellaneous Collections* 141(1).
- . 1976. Mammalian faunal zones of the Bridger middle Eocene. *Smithsonian Contributions to Paleobiology* 26.
- Gazin, C. L. and J. M. Sullivan. 1942. A new titanotherium from the Eocene of Mississippi, with notes on the correlation between the marine Eocene of the Gulf coastal plain and continental Eocene of the Rocky Mountain region. *Smithsonian Miscellaneous Collections* 101(13).
- Gingerich, P. D. 1979. Phylogeny of middle Eocene Adapidae (Mammalia, Primates) in North America: *Smilodectes* and *Notharctus*. *Journal of Paleontology* 53:153–163.
- , ed. 1980a. Early Cenozoic paleontology and stratigraphy of the Bighorn Basin, Wyoming. *University of Michigan Papers on Paleontology* 24.
- . 1980b. Evolutionary patterns in early Cenozoic mammals. *Annual Review of Earth and Planetary Science* 8:407–424.
- . 1983. Paleocene–Eocene faunal zones and a preliminary analysis of Laramide structural deformation in the Clark's Fork Basin, Wyoming. *Wyoming Geological Association Guidebook, 34th Annual Field Conference*, pp. 185–195.
- . 1989. New earliest Wasatchian mammalian fauna from the Eocene of northwestern Wyoming: Composition and diversity in a rarely sampled high-floodplain assemblage. *University of Michigan Papers on Paleontology* 28:1–97.
- . 2000. Paleocene–Eocene boundary and the continental vertebrate faunas of Europe and North America. In *Early Paleogene warm climates and biosphere dynamics*, ed. B. Schmitz, B. Sundquist, and F. P. Andreasson. *Geologiska Föreningen Förhandlingar, Uppsala* 122:57–59.
- . 2001. Biostratigraphy of the continental Paleocene–Eocene boundary interval on Polecat bench in the northern Bighorn Basin. In *Paleocene–Eocene stratigraphy and biotic change in the Bighorn and Clark's Fork basins, Wyoming*, ed. P. D. Gingerich. *University of Michigan Papers on Paleontology* 33:37–71.
- Givens, C. R. and M. P. Kennedy. 1979. Eocene molluscan stages and their correlation, San Diego area California. In *Eocene depositional systems, San Diego, California*, ed. P. L. Abbott. Pacific Section, SEPM Guidebook, pp. 81–95.
- Golz, D. J. 1976. Eocene Artiodactyla of southern California. *Natural History Museum of Los Angeles County Science Bulletin* 26.
- Golz, D. J. and J. A. Lillegraven. 1977. Summary of known occurrences of terrestrial vertebrates from Eocene strata of southern California. *University of Wyoming, Contributions to Geology* 15:43–65.
- Granger, W. 1909. Faunal horizons of the Washakie Formation of southern Wyoming. *Bulletin of the American Museum of Natural History* 26:13–23.
- . 1910. Tertiary faunal horizons in the Wind River Basin, Wyoming, with descriptions of new Eocene mammals. *Bulletin of the American Museum of Natural History* 28:235–251.
- . 1914. On the names of lower Eocene faunal horizons of Wyoming and New Mexico. *Bulletin of the American Museum of Natural History* 33:201–207.
- Gunnell, G. F. 1995. New notharctine (Primates, Adapidae) skull from the Uintan (middle Eocene) of San Diego County, California. *American Journal of Physical Anthropology* 98:447–470.
- . 1998a. Mammalian fauna from the lower Bridger Formation (Bridger A, early Middle Eocene) southern Green River Basin, Wyoming. *University of Michigan, Contributions from the Museum of Paleontology* 30:83–130.
- . 1998b. Mammalian faunal composition and the Paleocene/Eocene epoch/series boundary: Evidence from the Northern Bighorn Basin, Wyoming. In *Late Paleocene–early Eocene climatic and biotic events in the marine and terrestrial records*, ed. M.-P. Aubry, S. G. Lucas, and W. A. Berggren. New York: Columbia University Press, pp. 409–427.
- Gunnell, G. F. and W. S. Bartels. 1994. Early Bridgerian (middle Eocene) vertebrates, paleontology and paleoecology of the southern Green River Basin, Wyoming. *University of Wyoming, Contributions to Geology* 30:57–70.

- . 1999. Middle Eocene vertebrates from the Uinta Basin, Utah and their relationship with faunas from the southern Green River Basin, Wyoming. In *Vertebrate paleontology in Utah*, ed. D. D. Gillette. *Utah Geological Survey Miscellaneous Publication* 99:429–422.
- . 2001. Basin margins, biodiversity, evolutionary innovation, and the origin of new taxa. In *Eocene biodiversity: Unusual occurrences and rarely sampled habitats*, ed. G. F. Gunnell. New York: Kluwer/Plenum, Topics in Geobiology Series, pp. 403–440.
- Gunnell, G. F., W. S. Bartels, P. D. Gingerich, and V. Torres. 1992. Wapiti Valley faunas: Early and middle Eocene fossil vertebrates from the North Fork of the Shoshone River, Park County, Wyoming. *University of Michigan, Contributions from the Museum of Paleontology* 28:247–287.
- Gunnell, G. F. and V. L. Yarbrough. 2000. Brontotheriidae (Perissodactyla) from the late Early and Middle Eocene (Bridgerian) Wasatch and Bridger Formations, Southern Green River Basin, southwestern Wyoming. *Journal of Vertebrate Paleontology* 20:349–368.
- Gustafson, E. P. 1986. Carnivorous mammals of the late Eocene and early Oligocene of Trans-Pecos, Texas. *Texas Memorial Museum Bulletin* 33:1–66.
- Guthrie, D. A. 1967. The mammalian fauna of the Lysite Member, Wind River Formation (early Eocene) of Wyoming. *Memoir, Southern California Academy of Science* 5.
- Harshman, E. N. 1972. Geology and uranium deposits, Shirley Basin area, Wyoming. *U.S. Geological Survey Professional Paper* 745.
- Hayden, F. V. 1869. First, second, and third annual reports of the United States Geological Survey of the territories for the years 1867, 1868, and 1869. *U.S. Geological Survey of the Territories, 3d Annual Report*, pp. 109–199.
- . 1878. Wasatch group. In General notes, geology, and paleontology. *American Naturalist* 12:831.
- Henry, C. D., M. J. Kunk, and W. C. McIntosh. 1994.  $^{40}\text{Ar}/^{39}\text{Ar}$  chronology and volcanology of silicic volcanism in the Davis Mountains, Trans-Pecos, Texas. *Geological Society of America Bulletin* 106:1359–1376.
- Henry, C. D., F. W. McDowell, J. G. Price, and R. C. Smyth. 1986. Compilation of potassium–argon ages of Tertiary igneous rock, Trans-Pecos Texas. *Bureau of Economic Geology of the University of Texas Circular* 86-2:1–34.
- Hickey, L. J., R. M. West, M. R. Dawson, and D. K. Choi. 1983. Arctic terrestrial biota: Paleomagnetic evidence of age disparity with mid-northern latitudes during the late Cretaceous and early Tertiary. *Science* 221:1153–1156.
- Hill, R. V., J. G. Honey, and M. A. O'Leary. 2000. New fossils from the early Eocene Four Mile Area and improved relative dating of vertebrate localities. *Journal of Vertebrate Paleontology* 20(suppl. 3):48A.
- Honey, J. G. 1988. A mammalian fauna from the base of the Eocene Cathedral Bluffs Tongue of the Wasatch Formation, Cottonwood Creek Area, Southeast Washakie Basin, Wyoming. *U.S. Geological Survey Bulletin* 1669:C1–C14, pl. 1–2.
- . 1990. New washakiin primates (Omomyidae) from the Eocene of Wyoming and Colorado, and comments on the evolution of the Washakiini. *Journal of Vertebrate Paleontology* 10:206–221.
- Jepsen, G. L. 1939. Dating Absaroka rocks by vertebrate fossils (abstract). *Bulletin of the Geological Society of America* 50:1914.
- . 1963. Eocene vertebrates, coprolites, and plants in the Golden Valley Formation, western North Dakota. *Bulletin of the Geological Society of America* 74: 673–684.
- Johnson, R. B. and G. H. Wood Jr. 1956. Stratigraphy of the Upper Cretaceous and Tertiary Rocks of the Raton Basin, Colorado and New Mexico. *Bulletin of the American Association of Petroleum Geologists* 40:707–721.
- Johnson, R. B., G. H. Wood Jr., and R. L. Harbour. 1958. *Preliminary geologic map of the northern part of the Raton Mesa region and Huerfano Park in parts of Las Animas, Huerfano and Custer Counties, Colorado*. U.S. Geological Survey, Oil and Gas Investigations, Map OM 183.
- Kay, J. L. 1934. The Tertiary formations of the Uinta Basin, Utah. *Annals of the Carnegie Museum* 23:357–371.
- . 1957. The Eocene vertebrates of the Uinta Basin, Utah. *Intermountain Association of Petroleum Geologists Guidebook, 8th Annual Field Conference* 110–114.
- Keefer, W. R. 1965. Stratigraphy and geologic history of the uppermost Cretaceous, Paleocene, and lower Eocene rocks in the Wind River Basin, Wyoming. *U.S. Geological Survey Professional Paper* 495-A.
- Kelly, T. S. 1990. Biostratigraphy of the Uintan and Duchesnean land mammal assemblages from the middle member of the Sespe Formation, Simi Valley, California. *Los Angeles County Natural History Museum Contributions in Science* 419:1–42.
- . 1992. New Uintan and Duchesnean (middle and late Eocene) rodents from the Sespe Formation, Simi Valley, California. *Bulletin of the Southern California Academy of Science* 91:97–120.
- Kelly, T. S., E. B. Lander, D. P. Whistler, M. A. Roeder, and R. E. Reynolds. 1991. Preliminary report on a paleontologic investigation of the lower and middle members of the Sespe Formation, Simi Valley landfill, Ventura County, California. *Paleobios* 13(50):1–13.
- Kelly, T. S. and D. P. Whistler. 1998. A new eomyid rodent from the Sespe Formation of southern California. *Journal of Vertebrate Paleontology* 18:440–443.
- Kennedy, M. P. and G. W. Moore. 1971. Stratigraphic relations of Upper Cretaceous and Eocene formations, San Diego coastal area, California. *American Association of Petroleum Geologists Bulletin* 55:709–722.
- Kennedy, M. P. and G. L. Peterson. 1975. Geology of the eastern San Diego Metropolitan area, California. *Bulletin of the California Division of Mines and Geology* 200-B:43–56.
- Kent, D. V., M. C. McKenna, N. D. Oldie, J. J. Flynn, and B. J. MacFadden. 1984. Arctic biostratigraphic heterochrony. *Science* 224:173–174.
- Kew, W. S. W. 1924. Geology and oil resources of a part of Los Angeles and Ventura counties, California. *Bulletin of the U.S. Geological Survey* 753.
- Kihm, A. J. 1984. *Early Eocene mammalian faunas of the Piceance Creek Basin, northwestern Colorado*. Ph.D. dissertation, University of Colorado, Boulder.
- Koch, P. L., J. C. Zachos, and P. D. Gingerich. 1992. Correlation between isotope records in marine and continental carbon reservoirs near the Paleocene/Eocene boundary. *Nature* 358:319–322.
- Korth, W. W. 1982. Revision of the Wind River faunas, early Eocene of central Wyoming. Pt. 2: Geologic setting. *Annals of the Carnegie Museum* 51:57–78.

- Krishtalka, L. 1975. *Systematics and relationships of early Tertiary Lipotyphla (Mammalia, Insectivora) of North America*. Ph.D. dissertation, Texas Tech University, Lubbock.
- . 1976a. Early Tertiary Adapisoricidae and Erinaceidae (Mammalia, Insectivora) of North America. *Bulletin of the Carnegie Museum of Natural History* 1:1–40.
- . 1976b. North American Nyctitheriidae (Mammalia, Insectivora). *Annals of the Carnegie Museum* 46:7–28.
- . 1979. Paleontology and geology of the Badwater Creek area, central Wyoming. Pt. 18: Revision of late Eocene *Hyopsodus*. *Annals of the Carnegie Museum* 48:377–389.
- Krishtalka, L. and C. C. Black. 1975. Paleontology and geology of the Badwater Creek area, central Wyoming. Pt. 12: Description and review of late Eocene Multituberculata from Wyoming and Montana. *Annals of the Carnegie Museum* 45:287–297.
- Krishtalka, L. and T. Setoguchi. 1977. Paleontology and geology of the Badwater Creek area, central Wyoming. Pt. 13: The late Eocene Insectivora and Dermoptera. *Annals of the Carnegie Museum* 46:71–99.
- Krishtalka, L. and R. K. Stucky. 1983. Revision of the Wind River faunas, early Eocene of central Wyoming. Pt. 3: Marsupialia. *Annals of the Carnegie Museum* 52:205–227.
- . 1984. Middle Eocene marsupials (Mammalia) from northeastern Utah, and the mammalian fauna from Powder Wash. *Annals of the Carnegie Museum* 53:31–45.
- . 1985. Revision of the Wind River faunas, early Eocene of central Wyoming. Pt. 7: Revision of *Diacodexis* (Mammalia, Artiodactyla). *Annals of the Carnegie Museum* 54:413–486.
- Krishtalka, L., R. M. West, C. C. Black, M. R. Dawson, J. J. Flynn, W. D. Turnbull, R. K. Stucky, M. C. McKenna, T. M. Bown, D. J. Golz, and J. A. Lillegraven. 1987. Eocene (Wasatchian through Duchesnean) biochronology of North America. In *Cenozoic mammals of North America: Geochronology and biostratigraphy*, ed. M. O. Woodburne. Berkeley: University of California Press, pp. 77–117.
- Lambe, L. M. 1908. The Vertebrata of the Oligocene of the Cypress Hills, Saskatchewan. *Contributions to Canadian Paleontology* 3:1–64.
- Lander, B. 1994. Recalibration and causes of marine regressive–transgressive cycle recorded by middle Eocene to lower Miocene nonmarine Sespe Formation, southern California continental plate margin. In *Sedimentology and paleontology of Eocene rocks in the Sespe Creek area, Ventura County, California*, ed. A. E. Fritsche. *Pacific Section SEPM (Society for Sedimentary Geology) Book* 74:79–88.
- Lillegraven, J. A. 1976. Didelphids (Marsupialia) and *Uintasorex* (?Primates) from the later Eocene sediments of San Diego County, California. *San Diego Society of Natural History Transactions* 18:85–112.
- . 1979a. A biogeographical problem involving comparisons of later Eocene terrestrial vertebrate faunas of western North America. In *Historical biogeography, plate tectonics and the changing environment*, ed. J. Gray and A. J. Boucot. Corvallis: Oregon State University Press, pp. 333–347.
- . 1979b. Small rodents (Mammalia) from the Eocene deposits of San Diego County, California. *Bulletin of the American Museum of Natural History* 158:221–262.
- . 1980. Primates from later Eocene rocks of southern California. *Journal of Mammalogy* 61:181–204.
- Lillegraven, J. A., M. C. McKenna, and L. Krishtalka. 1981. Evolutionary relationships of middle Eocene and younger species of *Centetodon* (Mammalia, Insectivora, Geolabididae) with a description of the dentition of *Ankylodon* (Adapisoricidae). *University of Wyoming Publications* 45:1–115.
- Lillegraven, J. A. and A. Tabrum. 1983. A new species of *Centetodon* (Mammalia, Insectivora, Geolabididae) from southwestern Montana and its biogeographical implications. *University of Wyoming, Contributions to Geology* 22:57–73.
- Lindsay, E. H. 1968. Rodents from the Hartman Ranch local fauna, California. *Paleobios* 6: 1–20.
- Love, J. D. 1939. Geology along the southern margin of the Absaroka Range, Wyoming. *Geological Society of America Special Paper* 20.
- . 1947. Tertiary stratigraphy of the Jackson Hole area, northwestern Wyoming. *U.S. Geological Survey Oil and Gas Investigations Preliminary Chart* 27.
- . 1961. Definition of Green River, Great Divide, and Washakie basins, southwestern Wyoming. *American Association of Petroleum Geologists Bulletin* 45:1749–1755.
- Lucas, S. G. 1977. Vertebrate paleontology of the San Jose Formation, east-central San Juan Basin, New Mexico. *New Mexico Geological Society Guidebook, 28th Field Conference*, pp. 221–225.
- . 1983. The Baca Formation and the Eocene–Oligocene boundary in New Mexico. *New Mexico Geological Society Guidebook* 34:187–192.
- . 1990. Middle Eocene mammal from the base of the Baca Formation, west-central New Mexico. *New Mexico Journal of Science* 30:35–39.
- . 1992. Redefinition of the Duchesnean land mammal “age,” Late Eocene of western North America. In *Eocene–Oligocene climatic and biotic evolution*, ed. D. R. Prothero and W. A. Berggren. Princeton, NJ: Princeton University Press, pp. 88–105.
- Lucas, S. G. and R. M. Schoch. 1989. Taxonomy of *Duchesneodus* (Brontotheriidae) from the late Eocene of North America. In *The evolution of the perissodactyls*, ed. D. R. Prothero and R. M. Schoch. Oxford, UK: Oxford University Press, pp. 490–503.
- Lucas, S. G., R. M. Schoch, E. Manning, and C. Tsentas. 1981. The Eocene biostratigraphy of New Mexico. *Bulletin of the Geological Society of America*, 92(1):951–967.
- Lucas, S. G. and T. E. Williamson. 1993. Eocene vertebrates and late Laramide stratigraphy of New Mexico. In *Vertebrate paleontology in New Mexico*, ed. S. G. Lucas and J. Zidek. *Bulletin of the New Mexico Museum of Natural History and Science* 2: 145–158.
- Maas, M. C. 1985. Taphonomy of a late Eocene microvertebrate locality, Wind River Basin, Wyoming (U.S.A.). *Palaeogeography, Palaeoclimatology, Palaeoecology* 52:123–142.
- MacFadden, B. J. 1980. Eocene perissodactyls from the type section of the Tepee Trail Formation of northwestern Wyoming. *University of Wyoming, Contributions to Geology* 18:135–143.
- MacGinitie, H. D., E. B. Leopold, and W. L. Rohrer. 1974. An early middle Eocene flora from the Yellowstone–Absaroka volcanic province, northwestern Wind River Basin, Wyoming. *University of California Publications in Geological Sciences* 108.
- Mader, B. J. 1989. The Brontotheriidae: A systematic revision and preliminary phylogeny of North American genera. In *The evolution of the perissodactyls*, ed. D. R. Prothero and R. M. Schoch. Oxford, UK: Oxford University Press, pp. 458–484.

- Marcantel, E. L. and M. P. Weiss. 1968. Colton Formation (Eocene) fluviatile and associated lacustrine beds, Gunnison Plateau, central Utah. *Ohio Journal of Science* 68:40–49.
- Mason, M. A. 1988. *Mammalian paleontology and stratigraphy of the early to middle Tertiary Sespe and Titus canyon formations, southern California*. Unpublished Ph.D. dissertation, University of California, Berkeley, pp. 1–257.
- . 1990. New fossil Primates from the Uintan (Eocene) of southern California. *Paleobios* 13(49):1–7.
- Matthew, W. D. 1909. The Carnivora and Insectivora of the Bridger Basin, middle Eocene. *Memoir of the American Museum of Natural History* 9:291–567.
- McAnally, L. M. 1996. Paleogene mammals on land and sea. In *Life in stone. A natural history of British Columbia's fossils*, ed. R. Ludvigsen. Vancouver: University of British Columbia, pp. 202–211.
- McCarroll, S. M., J. J. Flynn, and W. D. Turnbull. 1996a. Biostratigraphy and magnetostratigraphy of the Bridgerian–Uintan Washakie Formation, Washakie Basin, Wyoming. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 25–39.
- McCarroll, S. M., J. J. Flynn, and W. D. Turnbull. 1996b. The mammalian faunas of the Washakie Formation, Eocene age, of southern Wyoming. Part III. The perissodactyls. *Fieldiana: Geology* 33:1–38.
- McGrew, P. O., ed. 1959. The geology and paleontology of the Elk Mountain and Tabernacle Butte area, Wyoming. *Bulletin of the American Museum of Natural History* 117:117–176.
- McGrew, P. O. and R. Sullivan. 1970. The stratigraphy and paleontology of Bridger A. *University of Wyoming, Contributions to Geology* 9:66–85.
- McKenna, M. C. 1960. Fossil Mammalia from the early Wasatchian Four Mile fauna, Eocene of northwest Colorado. *University of California Publications in Geological Sciences* 37:1–130.
- . 1980a. Eocene paleolatitude, climate, and mammals of Ellesmere Island. *Palaeogeography, Palaeoclimatology, Palaeoecology* 30:349–362.
- . 1980b. Late Cretaceous and early Tertiary vertebrate paleontological reconnaissance, Togwotee Pass area, northwestern Wyoming. In *Aspects of vertebrate history*, ed. L. L. Jacobs. Flagstaff: Museum of Northern Arizona Press, pp. 321–343.
- . 1990. Plagiomenids (Mammalia, ?Dermoptera) from the Oligocene of Oregon, Montana and South Dakota and the middle Eocene of northwestern Wyoming. In *Dawn of the age of mammals in the northern part of the Rocky Mountain Interior, North America*, ed. T. M. Bown and K. D. Rose. *Geological Society of America Special Paper* 243:211–234.
- McKenna, M. C., P. Robinson, and D. W. Taylor. 1962. Notes on Eocene Mammalia and Mollusca from Tabernacle Butte, Wyoming. *American Museum Novitates* 2102.
- Miall, A. D. 1986. The Eureka Sound Group (Upper Cretaceous–Oligocene) Canadian Arctic Islands. *Bulletin of Canadian Petroleum Geology* 34:240–270.
- Morris, W. J. 1966. Fossil mammals from Baja California: New evidence on early Tertiary migrations. *Science* 153:1376–1378.
- Murphey, P. C. 2001. *Stratigraphy, fossil distribution, and depositional environments of the upper Bridger Formation (Middle Eocene) of southwestern Wyoming, and taphonomy of an unusual Bridger microfossil assemblage*. Unpublished Ph.D. dissertation, University of Colorado, Boulder.
- Murphey, P. C., A. Lester, B. Bohor, P. Robinson, E. Evanoff, and E. Larson. 1999.  $^{40}\text{Ar}/^{39}\text{Ar}$  dating of volcanic ash deposits in the Bridger Formation (middle Eocene), southwestern Wyoming. *Geological Society of America Abstracts with Programs* 33:A-233.
- Nelson, M. E. 1973. Age and stratigraphic relations of the Fowkes Formation, Eocene, of southwestern Wyoming and northeastern Utah. *University of Wyoming, Contributions to Geology* 12:27–31.
- . 1974. Middle Eocene rodents from southwestern Wyoming. *University of Wyoming, Contributions to Geology* 15:1–10.
- . 1977. Middle Eocene primates (Mammalia) from southwestern Wyoming. *Southwestern Naturalist* 22:487–493.
- . 1979. K–Ar age for the Fowkes Formation (middle Eocene) of southwestern Wyoming. *University of Wyoming, Contributions to Geology* 17:51–52.
- Nelson, M. E., J. H. Madsen Jr., and W. L. Stokes. 1980. A titanothere from the Green River Formation, central Utah: *Teleodius uintensis* (Perissodactyla: Brontotheriidae). *University of Wyoming, Contributions to Geology* 18:127–134.
- Norris, R. D. and U. Rohl. 1999. Carbon cycling and chronology of the climate during the Paleocene/Eocene transition. *Nature* 401:775–778.
- Novacek, M. J. 1976. Insectivora and Proteutheria of the later Eocene (Uintan) of San Diego County, California. *Los Angeles County Natural History Museum Contributions to Science* 283:1–52.
- . 1985. The Sespedectinae, a new subfamily of hedge-hog-like insectivores. *American Museum of Natural History Novitates* 2822.
- Novacek, M. J., I. Ferrusquia-Villafranca, J. J. Flynn, A. R. Wyss, and M. Norell. 1991. Wasatchian (early Eocene) mammals and other vertebrates from Baja California, Mexico: The Lomas las Tetas de Cabra Fauna. *Bulletin of the American Museum of Natural History* 208:1–88.
- Novacek, M. J. and J. A. Lillegraven. 1979. Terrestrial vertebrates from the later Eocene of San Diego County, California: A conspectus. In *Eocene depositional systems, San Diego, California*, ed. P. L. Abbott. Los Angeles: Pacific Section, SEPM, pp. 69–79.
- Oriel, S. S. 1962. Main body of Wasatch Formation near LaBarge, Wyoming. *American Association of Petroleum Geologists Bulletin* 46:2161–2173.
- Oriel, S. S., C. L. Gazin, and J. I. Tracey Jr. 1962. Eocene age of Almy Formation, Wyoming, in its type area. *American Association of Petroleum Geologists Bulletin* 48:1936–1943.
- Oriel, S. S. and J. I. Tracey Jr. 1970. Uppermost Cretaceous and Tertiary stratigraphy of Fossil Basin, southwestern Wyoming. *U.S. Geological Survey Professional Paper* 635.
- Osborn, H. F. 1895. Fossil mammals of the Uinta Basin: Expedition of 1894. *Bulletin of the American Museum of Natural History* 7:71–105.
- . 1929. The titanotheres of ancient Wyoming, Dakota, and Nebraska. *U.S. Geological Survey Monograph* 55.
- Peterson, G. L. and M. P. Kennedy. 1974. Lithostratigraphic variations in the Poway Group near San Diego, California. *San Diego Society of Natural History Transactions* 17:251–258.
- Peterson, O. A. 1924. Osteology of *Dolichorhinus longiceps* Douglass, with a review of the species of *Dolichorhinus* in the order of their publication. *Carnegie Museum Memoirs* 9:405–492.
- . 1932. New species from the Oligocene of the Uinta. *Annals of the Carnegie Museum* 21:61–78.
- Prichinello, K. A. 1971. Earliest Eocene mammalian fossils from the Laramie Basin of southeast Wyoming. *University of Wyoming, Contributions to Geology* 10:73–88.

- Prothero, D. R. 1995. Geochronology and magnetostratigraphy of Paleogene North American land mammal "ages." In *Geochronology, time scales and global stratigraphic correlation*, ed. W. A. Berggren, D. V. Kent, M.-P. Aubry, and J. Hardenbol. Tulsa, OK: SEPM (Soc. Sed. Geol.), pp. 295–315.
- . 1996a. Camelidae. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 609–651.
- . 1996b. Magnetic stratigraphy and biostratigraphy of the middle Eocene Uinta Formation, Uinta Basin, Utah. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 1–24.
- . 1996c. Magnetostratigraphy of the Eocene–Oligocene transition in Trans-Pecos Texas. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 189–198.
- Prothero, D. R. and C. R. Denham. 1981. Magnetostratigraphy of the White River Group and its implications for Oligocene geochronology. *Geological Society of America Abstracts with Programs* 13:534.
- Prothero, D. R. and R. J. Emry. 1996. Summary. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 664–683.
- Prothero, D. R., J. L. Howard, and T. H. H. Dozier. 1996. Stratigraphy and paleomagnetism of the upper middle Eocene to lower Miocene (Uintan–Arikarean) Sespe Formation, Ventura County, California. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 171–188.
- Prothero, D. R. and S. G. Lucas. 1996. Magnetic stratigraphy of the Duchesnean part of the Galisteo Formation, New Mexico. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 199–205.
- Prothero, D. R. and C. C. I. Swisher. 1992. Magnetostratigraphy and geochronology of the terrestrial Eocene–Oligocene transition in North America. In *Eocene–Oligocene climatic and biotic evolution*, ed. D. R. Prothero and W. A. Berggren. Princeton, NJ: Princeton University Press, pp. 74–87.
- Prothero, D. R. and E. H. Vance Jr. 1996. Magnetostratigraphy of the upper middle Eocene Coldwater Sandstone, central Ventura County, California. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 155–170.
- Rapp, S. D., B. J. MacFadden, and J. A. Schiebout. 1983. Magnetic polarity stratigraphy of the early Tertiary Black Peaks Formation, Big Bend National Park, Texas. *Journal of Geology* 91:555–572.
- Rasmussen, D. T. 1996. A new middle Eocene omomyine primate from the Uinta Basin, Utah. *Journal of Human Evolution* 31:75–87.
- Rasmussen, D. T., G. C. Conroy, A. R. Friscia, K. E. Townsend, and M. D. Kinkel. 1999. Mammals of the middle Eocene Uinta Formation. In *Vertebrate paleontology in Utah*, ed. D. D. Gillette. *Utah Geological Survey Miscellaneous Publications* 99-1:401–420.
- Rasmussen, D. T., A. H. Hamblin, and A. R. Tabrum. 1999. The mammals of the Eocene Duchesne River Formation. In *Vertebrate paleontology in Utah*, ed. D. D. Gillette. *Utah Geological Survey Miscellaneous Publications* 99-1:421–427.
- Rich, T. H. V. and J. W. Collinson. 1973. First mammalian fossil from the Flagstaff limestone, central Utah: *Vulpavus australis* (Carnivora: Miacidae). *Journal of Paleontology* 47:854–860.
- Ricketts, B. D. 1986. New formations of the Eureka Sound Group, Canadian Arctic Islands. Current Research, Part B. *Geological Survey of Canada Paper* 86-1 B:363–374.
- Ridgway, K. D., A. R. Sweet, and A. R. Cameron. 1995. Climatically induced floristic changes across the Eocene–Oligocene transition in the northern high latitudes, Yukon Territory, Canada. *Bulletin of the Geological Society of America* 107: 676–696.
- Riggs, E. S. 1912. New or little-known titanotheres from the lower Uintah formations. *Field Museum of Natural History Geological Series* 4(2):17–41.
- Robinson, G. E. 1963. Geology of the Three Forks Quadrangle, Montana. *U.S. Geological Survey Professional Paper* 370.
- Robinson, G. E., E. Lewis, and D. W. Taylor. 1957. Eocene continental deposits in Three Forks Basin, Montana (abstract). *Bulletin of the Geological Society of America* 68:1786.
- Robinson, P. 1957a. Age of Galisteo Formation, Santa Fe County, New Mexico. *American Association of Petroleum Geologists Bulletin* 41:757.
- Robinson, P. 1957b. The species of *Notharctus* from the middle Eocene. *Yale Peabody Museum Postilla* 28:1–27.
- . 1960. *Sinopa* from the Cuchara Formation of Colorado. *Yale Peabody Museum Postilla* 44:1–4.
- . 1963. Fossil vertebrates and age of the Cuchara Formation of Colorado. *University of Colorado Studies, Series in Geology* 1:1–5.
- . 1966. Fossil mammals of the Huerfano Formation, Eocene, of Colorado. *Peabody Museum of Natural History Yale University Bulletin* 21.
- Robinson, P. and L. D. Ivy. 1994. Paromomyidae (Dermoptera?) from the Powder River Basin, Wyoming, and a discussion of microevolution in closely related species. *University of Wyoming, Contributions to Geology* 30:91–116.
- Robinson, P. and B. A. Williams. 1998. Species diversity, tooth size, and shape of *Haplomylys* (Condylarthra, Hyopsodontidae) from the Powder River Basin, northeastern Wyoming. *University of Wyoming, Contributions to Geology* 31: 59–78.
- Roehler, H. W. 1973. Stratigraphy of the Washakie Formation in the Washakie Basin, Wyoming. *U.S. Geological Survey Bulletin* 1369.
- . 1991. Revised stratigraphic nomenclature for the Wasatch and Green River Formations of Eocene Age, Wyoming, Utah and Colorado. *U.S. Geological Survey Professional Paper* 1506-B:1–38.
- . 1992a. Correlation, composition, areal distribution and thickness of Eocene stratigraphic units, Greater Green River Basin, Wyoming, Utah, and Colorado. *U.S. Geological Survey Professional Paper* 1506-E:1–49.
- . 1992b. Description and correlation of Eocene rocks in the stratigraphic reference sections for the Green River and Washakie Basins, southwest Wyoming. *U.S. Geological Survey Professional Paper* 1506-D:1–83.
- . 1992c. Introduction to Greater Green River Basin geology, physiography and history of investigations. *U.S. Geological Survey Professional Paper* 1506-A:1–14.
- Rohl, U., T. J. Bralower, R. D. Norris, and G. Welfer. 2000. New chronology for the late Paleocene thermal maximum and its environmental implications. *Geology* 28:927–930.
- Rose, K. D. 1981. The Clarkforkian land mammal age and mammalian faunal composition across the Paleocene–Eocene boundary. *University of Michigan Papers on Paleontology* 26.

- . 1999. Fossil mammals from the Early Eocene Fisher/Sullivan site. In *Early Eocene vertebrates from the Fisher/Sullivan site (Nanjemoy Formation) Stafford County, Virginia*, ed. R. E. Weems and G. J. Grimsley. *Virginia Division of Mineral Resources Publication* 152:133–137.
- Rose, K. D., R. D. E. MacPhee, and J. P. Alexander. 1999. The skull of Early Eocene *Cantius abditus* (Primates, Adapiformes) and its phylogenetic implications, with a reevaluation of “*Hesperolemur*” *actius*. *American Journal of Physical Anthropology* 109:523–539.
- Runkel, A. C. 1988. *Stratigraphy, Sedimentology and vertebrate paleontology of Eocene rocks, Big Bend region, Texas*. Unpublished Ph.D. dissertation, University of Texas, Austin, pp. 1–310.
- Russell, L. S. 1935. A middle Eocene mammal fauna from British Columbia. *American Journal of Science* 29:54–55.
- . 1954. Mammalian fauna of the Kishenehn Formation, southeastern British Columbia. *Annual Report of the National Museum, Fiscal Year 1952–53, Bulletin* 132:92–111.
- . 1965. Tertiary mammals of Saskatchewan. Pt. I: The Eocene fauna. *Royal Ontario Museum, University of Toronto, Life Sciences Contribution* 67.
- Russell, L. S. and R. T. D. Wickenden. 1933. An upper Eocene Vertebrate fauna from Saskatchewan. *Transactions of the Royal Society of Canada Series 3*, 27(4):53–65.
- Schankler, D. M. 1980. Faunal zonation of the Willwood Formation in the central Bighorn Basin, Wyoming. In *Early Cenozoic paleontology and stratigraphy of the Bighorn Basin, Wyoming*, ed. P. D. Gingerich. *University of Michigan Papers on Paleontology* 24:99–114.
- Schiebout, J. A. 1974. Vertebrate paleontology and paleoecology of Paleocene Black Peaks Formation, Big Bend National Park, Texas. *Texas Memorial Museum Bulletin* 24.
- Schiebout, J. A. and A. K. Schrod. 1981. Vertebrate paleontology of the lower Tertiary Baca Formation of western New Mexico. *American Association of Petroleum Geologists Bulletin* 65:568.
- Scott, W. B. 1945. The Mammalia of the Duchesne River Oligocene. *Transactions of the American Philosophical Society* 34:209–233.
- Sharp, W. N., E. J. McKay, F. A. McKeown, and A. M. White. 1964. Geology and uranium deposits of the Pumpkin Buttes area of the Powder River Basin, Wyoming. *Bulletin of the U.S. Geological Survey* 1107-H:541–638.
- Simpson, G. G. 1948. The Eocene of the San Juan Basin, New Mexico. *American Journal of Science* 246:257–282, 363–385.
- Sinclair, W. J. and W. Granger. 1911. Eocene and Oligocene of the Wind River and Bighorn basins. *Bulletin of the American Museum of Natural History* 30:83–117.
- Smith, K. T. and P. A. Holroyd. 2001. “Gardnerbuttean” mammals in a classic Lostcabinian locality, northern Green River Basin, Wyoming, with implications for the nature of high-latitude corridor routes. In *Climate and biota of the early Paleogene*, ed. A. W. Ash and S. L. Wing. Powell, WY: Smithsonian Institution, July 3–8, 2001, [http://www.paleogene.net/abstract\\_volume.html](http://www.paleogene.net/abstract_volume.html):83.
- Smith, L. N. and S. G. Lucas. 1991. Stratigraphy, sedimentology, and paleontology of the lower Eocene San Jose Formation in the central portion of the San Juan Basin, northwestern New Mexico. *New Mexico Bureau of Mines and Mineral Resources Bulletin* 126:1–44.
- Smith, M. E., B. Singer, and A. R. Carroll. 2003.  $^{40}\text{Ar}/^{39}\text{Ar}$  geochronology of the Eocene Green River Formation, Wyoming. *Bulletin of the Geological Society of America* 115:549–565.
- Soister, P. E. 1968. Stratigraphy of the Wind River Formation in south-central Wind River Basin, Wyoming. *U.S. Geological Survey Professional Paper* 594-A.
- Squires, R. L. 1994. Macropaleontology of Eocene marine rocks, upper Sespe Creek area, Ventura County, southern California. In *Sedimentology and paleontology of Eocene rocks in the Sespe Creek area, Ventura County, California*, ed. A. E. Fritsche. Pacific Section SEPM (Soc. for Sed. Geol.) Book 74:39–56.
- Stearns, C. E. 1943. The Galisteo Formation of north-central New Mexico. *Journal of Geology* 51:301–319.
- Stock, C. 1938. A titanothera from the type Sespe of California. *Proceedings of the National Academy of Science* 24:507–512.
- Storer, J. E. 1978. Rodents of the Swift Current Creek local fauna (Eocene: Uintan) of Saskatchewan. *Canadian Journal of Earth Sciences* 15:1673–1674.
- . 1984. Mammals of the Swift Current Creek local fauna (Eocene: Uintan, Saskatchewan). *Contributions of the Saskatchewan Museum of Natural History* 7:1–158.
- . 1989. Rodent faunal provinces, Paleocene–Miocene of North America. In *Papers on fossil rodents in honor of Albert Elmer Wood*, ed. C. C. Black and M. R. Dawson. *Natural History Museum of Los Angeles County, Science Series* 33:17–29.
- . 1995. Small mammals of the Lac Pelletier lower fauna, Duchesnean, of Saskatchewan, Canada: Insectivores and insectivore-like groups, a plagiomenid, a microsypid and Chiroptera. In *Vertebrate fossils and evolution of scientific concepts*, ed. W. A. S. Sarjeant. Amsterdam: Gordon and Breach, pp. 595–615.
- . 1996. Eocene–Oligocene faunas of the Cypress Hills Formation, Saskatchewan. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 240–261.
- Stucky, R. K. 1984a. Revision of the Wind River faunas, early Eocene of central Wyoming. Pt. 5: Geology and mammalian biostratigraphy of the upper part of the Wind River Formation. *Annals of the Carnegie Museum* 53:231–294.
- . 1984b. The Wasatchian–Bridgerian land mammal age boundary (early to middle Eocene) in western North America. *Annals of the Carnegie Museum* 53:347–382.
- Stucky, R. K. and L. Krishtalka. 1983. Revision of the Wind River faunas, early Eocene of central Wyoming. Pt. 4: The Tillodontia. *Annals of the Carnegie Museum* 52:375–391.
- Stucky, R. K., D. R. Prothero, W. G. Lohr, and J. R. Snyder. 1996. Magnetic stratigraphy, sedimentology, and mammalian faunas of the early Uintan Washakie Formation, Sand Wash Basin, northwestern Colorado. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 40–51.
- Sundell, K. A., P. N. Shive, and J. G. Eaton. 1984. Measured sections, magnetic polarity and biostratigraphy of the Eocene Wiggins, Tepee Trail and Aycross formations within the southeastern Absaroka Range, Wyoming. *Earth Science Bulletin, Wyoming Geological Association* 17:1–48.
- Swisher, C. C. III and D. R. Prothero. 1990. Single crystal  $^{40}\text{Ar}/^{39}\text{Ar}$  dating of the Eocene–Oligocene transition in North America. *Science* 249:760–762.
- Szalay, F. S. 1976. Systematics of the Omomyidae (Tarsiiformes, Primates): Taxonomy, phylogeny, and systematics. *Bulletin of the American Museum of Natural History* 156:157–450.

- Tabrum, A., D. R. Prothero, and D. Garcia. 1996. Magnetostratigraphy and biostratigraphy of the Eocene–Oligocene transition, southwestern Montana. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 278–311.
- Tauxe, L. and D. R. Clark. 1987. New paleomagnetic results from the Eureka Sound Group: Implications for the age of the Early Tertiary arctic biota. *Geological Society of America Bulletin* 99:739–747.
- Tauxe, L., J. Gee, Y. Gellert, T. Pick, and T. Bown. 1994. Magnetostratigraphy of the Willwood Formation, Bighorn Basin, Wyoming: New constraints on the location of the Paleocene/Eocene boundary. *Earth and Planetary Science Letters* 125:159–172.
- Taylor, G. E. 1983. Braided-river and flood-related deposits of the Sespe Formation, northern Simi Valley, California. In *Cenozoic geology of the Simi Valley area, southern California*, ed. R. L. Squires and M. V. Filewicz. *Pacific Section, Society of Economic Paleontologists and Mineralogists Volume and Guidebook*, pp. 129–140.
- Tedford, R. H. 1970. Principles and practices of mammalian geochronology in North America. *Proceedings of the North American Paleontological Convention*, Pt. F, pp. 666–703.
- Thewissen, J. G. M. 1990. Evolution of Paleocene and Eocene Phenacodontidae (Mammalia, Condylarthra). *University of Michigan, Papers on Paleontology* 29:1–107.
- Triplehorn, D. M., D. L. Turner, and C. W. Naeser. 1984. Radiometric age of the Chickaloon Formation of south-central Alaska: Location of the Paleocene–Eocene boundary. *Bulletin of the Geological Society of America* 95:740–742.
- Turnbull, W. D. 1972. The Washakie Formation of Bridgerian–Uintan ages, and the related fauna. *Guidebook, Field Conference on Tertiary Biostratigraphy of Southern and Western Wyoming*, pp. 20–31.
- . 1978. The mammalian faunas of the Washakie Formation, Eocene age, of southern Wyoming. Pt. 1: Introduction: The geology, history, and setting. *Fieldiana: Geology* 33:569–601.
- Van de Kamp, P. C., J. D. Harper, J. J. Conniff, and D. A. Morris. 1974. Facies relations in the Eocene–Oligocene Santa Ynez Mountains, California. *Journal of the Geological Society of London* 130:545–565.
- Van Houten, F. B. 1944. Stratigraphy of the Willwood and Tatman formations in northwestern Wyoming. *Bulletin of the Geological Society of America* 65:165–210.
- . 1945. Review of latest Paleocene and early Eocene mammalian faunas. *Journal of Paleontology* 19:421–461.
- Veatch, A. C. 1907. Geography and geology of a portion of southwestern Wyoming. *U.S. Geological Survey Professional Paper* 56.
- Wallace, S. M. 1980. *A revision of North American early Eocene Bronthotheriidae (Mammalia, Perissodactyla)*. M.S. thesis, University of Colorado, Boulder.
- Walsh, S. L. 1991a. Eocene mammal faunas of San Diego County. In *Eocene geologic history San Diego region*, ed. P. L. Abbott and J. A. May. *Pacific Section, Society of Economic Paleontologists and Mineralogists* 68:161–178.
- . 1991b. Late Eocene mammals from the Sweetwater Formation, San Diego County, California. In *Eocene geologic history, San Diego region*, ed. P. L. Abbott and J. A. May. *Pacific Section, Society of Economic Paleontologists and Mineralogists* 68:149–159.
- . 1996a. Middle Eocene Mammalian faunas of San Diego County, California. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 75–119.
- . 1996b. Theoretical biochronology, the Bridgerian–Uintan boundary and the “Shoshonian Subage” of the Uintan. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 52–74.
- . 1997. New specimens of *Metanoiamys*, *Pauromys* and *Simimys* (Rodentia: Myomorpha) from the Uintan (middle Eocene) of San Diego County, California, and comments on the relationships of selected Paleogene Myomorpha. *Proceedings, San Diego Society of Natural History* 32:1–20.
- . 1998. Fossil datum terms, paleobiological event terms, paleontostratigraphy, chronostratigraphy and the definition of land mammal “age” boundaries. *Journal of Vertebrate Paleontology* 18:150–179.
- . 2000. Bunodont artiodactyls (Mammalia) from the Uintan (middle Eocene) of San Diego County, California. *Proceedings, San Diego Society of Natural History* 37:1–27.
- . 2001. Eubiostratigraphic units, quasibiostratigraphic units, and “assemblage zones.” *Journal of Vertebrate Paleontology* 20:761–775.
- Walsh, S. L. and R. Q. Gutzler. 1999. Late Duchesnean–early Chadronian mammals from the upper member of the Pomerado Conglomerate, San Diego County, California. *Journal of Vertebrate Paleontology* 19(suppl. 3):82A.
- Walsh, S. L., D. R. Prothero, and D. J. Lundquist. 1996. Stratigraphy and paleomagnetism of the middle Eocene Friars Formation and Poway Group, southwestern San Diego County, California. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 120–151.
- Walton, A. H. 1992. Magnetostratigraphy and the ages of the Bridgerian and Uintan faunas of the lower and middle members of the Devil’s Graveyard Formation, Trans-Pecos Texas. In *Eocene–Oligocene climatic and biologic evolution*, ed. D. R. Prothero and W. A. Berggren. Princeton, NJ: Princeton University Press, pp. 74–87.
- . 1993a. A new genus of eutypomyid (Mammalia: Rodentia) from the middle Eocene of Texas. *Journal of Vertebrate Paleontology* 13:262–266.
- . 1993b. *Pauromys* and other small Sciuravidae (Mammalia: Rodentia) from the Middle Eocene of Texas. *Journal of Vertebrate Paleontology* 23:243–261.
- Wegemann, C. H. 1917. Wasatch fossils in the so-called Fort Union beds of the Powder River Basin, Wyoming. *U.S. Geological Survey Professional Paper* 108-D:55–60.
- West, R. M. 1970. Sequence of mammalian faunas of Eocene age in the northern Green River Basin, Wyoming. *Journal of Paleontology* 14:142–147.
- . 1973a. Geology and mammalian paleontology of the New Fork–Big Sandy area, Sublette County, Wyoming. *Fieldiana: Geology* 29.
- . 1973b. New records of fossil mammals from the early Eocene Golden Valley Formation, North Dakota. *Journal of Mammalogy* 54:749–750.
- . 1973c. Review of the North American Eocene and Oligocene Apatemyidae (Mammalia: Insectivora). *Special Publications, Museum of Texas Tech University* 3.
- . 1976a. The North American Phenacodontidae (Mammalia, Condylarthra). *Milwaukee Public Museum Contributions in Biology and Geology* 6.



- . 1976b. Paleontology and geology of the Bridger Formation, southern Green River Basin, southwestern Wyoming. Pt. 1: History of fieldwork and geological setting. *Milwaukee Public Museum Contributions in Biology and Geology* 7.
- . 1982. Fossil mammals from the lower Buck Hill Group, Eocene of southwest Texas: Marsupicarnivora, Primates, Taeniodonta, Condylarthra, bunodont Artiodactyla, and Dinocerata. *Pearce-Sellards Series* 35:1–20.
- West, R. M. and E. G. Atkins. 1970. Additional middle Eocene (Bridgerian) mammals from Tabernacle Butte, Sublette County, Wyoming. *American Museum Novitates* 2404.
- West, R. M. and M. R. Dawson. 1973. Fossil mammals from the upper part of the Cathedral Bluffs Tongue of the Wasatch Formation (early Bridgerian), northern Green River Basin, Wyoming. *University of Wyoming, Contributions to Geology* 12:33–41.
- . 1975. Eocene fossil Mammalia from the Sand Wash Basin, northwestern Moffat County, Colorado. *Annals of the Carnegie Museum* 45:231–253.
- . 1978. Vertebrate paleontology and the Cenozoic history of the North Atlantic region. *Polarforschung* 48:103–119.
- West, R. M. and J. H. Hutchison. 1981. Geology and paleontology of the Bridger Formation, southern Green River Basin, southwestern Wyoming. Pt. 6: The fauna and correlation of Bridger E. *Milwaukee Public Museum Contributions in Biology and Geology* 46.
- Westgate, J. W. 1986. Late Eocene mammals from the Gulf Coastal Plain. In *Montgomery Landing Site, marine Eocene (Jackson) of central Louisiana*, ed. J. A. Schiebout W. van den Bold. Proceedings of a symposium, 1986 Annual Meeting, Gulf Coast Geological Society, pp. 223–239.
- . 1988. Biostratigraphic implications of the first Eocene land-mammal fauna from the North American coastal plain. *Geology* 16:995–998.
- . 1990. Uintan land mammals (excluding rodents) from the estuarine facies of the Laredo Formation (middle Eocene, Claiborne Group), Webb County, Texas. *Journal of Paleontology* 64:454–468.
- . 1994a. Eocene forest-swamp. *National Geographic Research and Exploration* 10:80–91.
- . 1994b. A new leptocherid from middle Eocene (Uintan) deposits of the Texas Coastal Plain. *Journal of Vertebrate Paleontology* 14:296–299.
- . 1994c. Uintan land mammals (excluding rodents) from an estuarine facies of the Laredo Formation (Middle Eocene, Claiborne Group) of Webb County, Texas. *Journal of Paleontology* 64:454–468.
- . 1999. *After the dinosaurs: A Texas tropical paradise recovered at Lake Casa Blanca*. Austin: Texas Parks and Wildlife Press, pp. 1–69.
- . 2001. Paleogeology and biostratigraphy of marginal marine Gulf Coast Eocene vertebrate localities. In *Eocene biodiversity: Unusual occurrences and rarely sampled habitats*, ed. G. F. Gunnell. New York: Kluwer/Plenum, Topics in Geobiology Series, pp. 263–297.
- Williamson, T. E. and S. G. Lucas. 1992. Fossil mammals and the early Eocene age of the San Jose Formation. *New Mexico Geological Society Guidebook* 43:311–316.
- Wilson, J. A. 1967. Early Tertiary mammals. In *Geology of Big Bend National Park*, ed. R. A. Maxwell, J. T. Lonsdale, R. T. Hazzard, and J. A. Wilson. University of Texas Publication 6711:157–169.
- . 1975. Geochronology, stratigraphy, and typology. *Fieldiana: Geology* 33:193–204.
- . 1977. Early Tertiary faunas, Big Bend area, Trans-Pecos Texas: Brontotheriidae. *Pearce-Sellards Series* 25.
- . 1978. Stratigraphic occurrence and correlation of early Tertiary vertebrate faunas, Trans-Pecos Texas. Pt. 1: Vieja area. *Texas Memorial Museum Bulletin* 25.
- . 1984. Vertebrate faunas forty-nine to thirty-six million years ago and additions to the species of *Leptoreodon* (Mammalia: Artiodactyla) found in Texas. *Journal of Vertebrate Paleontology* 4:199–207.
- . 1986. Stratigraphic occurrence and correlation of early Tertiary vertebrate faunas, Trans-Pecos, Texas. Agua Fria–Green Valley areas. *Journal of Vertebrate Paleontology* 6:350–373.
- Wilson, J. A. and J. W. Westgate. 1991. A lophodont rodent from the middle Eocene of the Gulf Coastal Plain, Texas. *Journal of Vertebrate Paleontology* 11:257–260.
- Wilson, K. L. 1972. *Eocene and related geology of a portion of San Luis Rey and Encinitas quadrangles, San Diego County, California*. Unpublished M.S. thesis, University of California, Riverside, pp. 1–135.
- Wilson, R. W. 1967. Fossil mammals in Tertiary correlations. In *Essays in paleontology and stratigraphy*, ed. C. Teichert and E. L. Yochelson. *University of Kansas Department of Geology Special Publication* 2:590–606.
- Wing, S. L., H. Bao, and P. L. Koch. 2000. An early Eocene cool period? Evidence for continental cooling during the warmest part of the Cenozoic. In *Warm climates in Earth history*, ed. B. T. Huber, K. G. Macleod, and S. L. Wing. New York: Cambridge University Press, pp. 197–237.
- Wing, S. L., T. M. Bown, and J. D. Obradovich. 1991. Early Eocene biotic and climatic change in interior western North America. *Geology* 19:1189–1192.
- Winterfeld, G. F. 1986. *Laramide tectonism, deposition, and Early Cenozoic stratigraphy of the northwestern Wind River Basin and Washakie Range, Wyoming*. Unpublished Ph.D. dissertation, University of Wyoming, Laramie, pp. 1–248.
- Wood, H. E. 1934. Revision of the Hyrachyidae. *Bulletin of the American Museum of Natural History* 67:182–295.
- Wood, H. E., R. W. Chaney, J. Clark, E. H. Colbert, G. L. Jepsen, J. B. Reeside Jr., and C. Stock. 1941. Nomenclature and correlation of the North American continental Tertiary. *Bulletin of the Geological Society of America* 52:1–48.
- Wood, H. E., H. Seton, and C. J. Hares. 1936. New data on the Eocene of the Wind River Basin, Wyoming. *Proceedings of the Geological Society of America* 1935:394–395.
- Zonneveld, J.-P., G. F. Gunnell, and W. S. Bartels. 2000. Early Eocene fossil vertebrates from the southwestern Green River Basin, Lincoln and Uinta Counties, Wyoming. *Journal of Vertebrate Paleontology* 20:369–386.

#### ADDITIONAL READINGS

- Bandy, O. L. and R. L. Kolpack. 1963. Foraminiferal and sedimentological trends in the Tertiary section of Tecolote Tunnel, California. *Micropaleontology* 9:117–170.
- Beard, K. C. 1987. *Jemezius*, a new omomyid primate from the early Eocene of northwestern New Mexico. *Journal of Human Evolution* 16:457–468.

- . 1988. New notharctine primate fossils from the early Eocene of New Mexico and southern Wyoming and the phylogeny of the Notharctinae. *American Journal of Physical Anthropology* 75:439–469.
- Beard, K. C. and M. R. Dawson. 1999. Intercontinental dispersal of holarctic land mammals near the Paleocene/Eocene boundary: Paleogeographic, paleoclimatic and biostratigraphic considerations. *Bulletin de la Société Géologique de France* 170:697–706.
- Berggren, W. A., D. V. Kent, J. J. Flynn, and J. A. Van Couvering. 1985. Cenozoic geochronology. *Bulletin of the Geological Society of America* 96:1407–1418.
- Bown, T. M. 1975. Paleocene and lower Eocene rocks in the Sand Creek–No Water Creek area, Washakie County, Wyoming. *Wyoming Geological Association Guidebook, 27th Annual Field Conference*, pp. 55–61.
- . 1980. The Willwood Formation (lower Eocene) of the southern Bighorn Basin, Wyoming, and its mammalian fauna. In *Early Cenozoic paleontology and stratigraphy of the Bighorn Basin, Wyoming, 1880–1980*, ed. P. D. Gingerich. *University of Michigan Papers on Paleontology* 24:127–138.
- Bown, T. M. and K. D. Rose. 1976. New early Tertiary primates and a reappraisal of some Plesiadapiformes. *Folia Primatologica* 26:109–138.
- Bown, T. M. and D. M. Schankler. 1982. A review of the Proteutheria and Insectivora of the Willwood Formation (lower Eocene), Bighorn Basin, Wyoming. *U.S. Geological Survey Professional Paper* 1201.
- Bradley, W. H. 1931. Origin and microfossils of the oil shale of the Green River Formation of Colorado and Utah. *U.S. Geological Survey Professional Paper* 168:1–58.
- Burbank, W. S., T. S. Lovering, E. N. Goddard, and E. B. Eckel. 1935. Reprinted 1967. *Geologic map of Colorado*. Boulder, CO: U.S. Geological Survey.
- Butler, R. F., E. H. Lindsay, and P. D. Gingerich. 1980. Magnetic polarity stratigraphy and Paleocene–Eocene biostratigraphy of Polecat Bench, northwestern Wyoming. In *Early Cenozoic paleontology and stratigraphy of the Bighorn Basin, Wyoming*, ed. P. D. Gingerich. *University of Michigan Papers on Paleontology* 24:95–98.
- Cashion, W. B. 1994. *Geologic map of the Nutter's Hole Quadrangle, Uintah and Carbon Counties, Utah*. U.S. Geological Survey Miscellaneous Field Studies Map MF-2250.
- Clark, B. L. and H. E. Vokes. 1936. Summary of marine Eocene sequence of western North America. *Bulletin of the Geological Society of America* 47:851–877.
- Coombs, M. C. 1971. Status of *Simidectes* (Insectivora, Pantolestoidea) of the late Eocene of North America. *American Museum of Natural History Novitates* 2455:1–41.
- Crochet, J.-Y. 1977. Les Didelphidae (Marsupicarnivora, Marsupialia) holarctiques tertiaires. *Comptes Rendus Académie de Sciences Paris* 284(D):357–360.
- Culbertson, W. C. 1961. Stratigraphy of the Wilkins Peak Member of the Green River Formation, Firehole Basin Quadrangle, Wyoming. *U.S. Geological Survey Professional Paper* 424-D:170–173.
- Dalrymple, G. B. 1979. Critical tables for conversion of K–Ar ages from old to new constants. *Geology* 7:558–559.
- Dawson, M. R. 1970. Paleontology and geology of the Badwater Creek area, central Wyoming. Part 6. The leporid *Mytonolagus* (Mammalia, Lagomorpha). *Annals of the Carnegie Museum* 41:215–230.
- . 1980. Geology and paleontology of the Badwater Creek area, central Wyoming. Pt. 20: The late Eocene Creodonta and Carnivora. *Annals of the Carnegie Museum* 49:79–91.
- Denison, R. H. 1937. Early lower Eocene mammals from the Wind River Basin, Wyoming. *Proceedings of the New England Zoological Club* 16:11–14.
- Dibblee, T. W. Jr. 1950. Geology of southwestern Santa Barbara County, California. *California Division of Mines Bulletin* 150.
- Dockery, D. T. III, K. C. Beard, A. R. Tabrum, and G. R. Case. 1991. New Early Eocene land mammal faunas from the Tusahoma and Bashi formations in Mississippi. *Journal of the Mississippi Academy of Science* 36:41.
- Donnell, J. R. 1969. Paleocene and lower Eocene units in the southern part of the Piceance Creek Basin, Colorado. *Bulletin of the U.S. Geological Survey* 1274-M.
- Dreyer, E. E. 1935. *Geology of a portion of Mt. Pinos Quadrangle, Ventura County, California*. M.A. thesis, University of California, Los Angeles.
- Durham, J. W., R. H. Johns, and D. E. Savage. 1954. Marine–non-marine relationships in the Cenozoic section of California. *California Division of Mines Bulletin* 170:59–71.
- Emry, R. J. 1973. Stratigraphy and preliminary biostratigraphy of the Flagstaff Rim area, Natrona County, Wyoming. *Smithsonian Contributions to Paleobiology* 18:1–43.
- . 1990. Mammals of the Bridgerian (middle Eocene) Elderberry Canyon Local Fauna of eastern Nevada. In *Dawn of the age of mammals in the northern part of the Rocky Mountain Interior*, ed. T. M. Bown and K. D. Rose. *Geological Society of America Special Paper* 243:187–210.
- Emry, R. J. and W. W. Korth. 1989. Rodents of the Bridgerian (middle Eocene) Elderberry Canyon Local Fauna of eastern Nevada. *Smithsonian Contributions to Paleobiology* 67:1–14.
- Evernden, J. R., D. E. Savage, G. H. Curtis, and G. T. James. 1964. Potassium–argon dates and the Cenozoic mammalian chronology of North America. *American Journal of Science* 262:145–198.
- Ferrusquia-Villafranca, I., E. Jimenez-Hidalgo, J. A. Ortiz-Mendieta, and V. M. Bravo-Cuevos. In prep. El registro paleogénico de mamíferos en México y su significación geológico-paleontológica. In *Avances en la paleomastozoología mexicana*, ed. J. Arroyo-Cabrales and M. Montellano-Ballesteros. Lubbock: Texas Tech University.
- Flynn, J. J. 1981. Magnetic polarity stratigraphy and correlation of Eocene strata from Wyoming and southern California (abstract). *Eos, Transactions, American Geophysical Union* 62:264.
- . 1998. Early Cenozoic Carnivora (“Miacoidea”). In *Evolution of Tertiary mammals in North America*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press 1:110–123.
- Gazin, C. L. 1956. The upper Paleocene Mammalia from the Almy Formation in western Wyoming. *Smithsonian Miscellaneous Collections* 131(7).
- . 1965. Early Eocene mammalian faunas and their environment in the vicinity of the Rock Springs Uplift, Wyoming. *Wyoming Geological Association Guidebook, 19th Annual Field Conference*, pp. 171–180.
- Gibson, J. M. 1971. Benthonic foraminifera of the Ardath Shale and Stadium Conglomerate (Eocene), San Diego Basin, California. *Bulletin of the Southern California Academy of Science* 70:125–130.
- Gingerich, P. D. 1978. New Condylarthra (Mammalia) from the Paleocene and early Eocene of North America. *University of Michigan, Contributions from the Museum of Paleontology* 25:1–9.

- Gingerich, P. D. and G. F. Gunnell. 1979. Systematics and evolution of the genus *Esthonyx* (Mammalia, Tillodontia) in the early Eocene of North America. *University of Michigan, Contributions from the Museum of Paleontology* 25:125–153.
- Gingerich, P. D. and R. A. Haskin. 1981. Dentition of early Eocene *Pelycodus jarrovii* (Mammalia, Primates) and the generic attribution of species formerly referred to *Pelycodus*. *University of Michigan, Contributions from the Museum of Paleontology* 25:327–337.
- Gingerich, P. D., K. D. Rose, and D. W. Krause. 1980. Early Cenozoic mammalian faunas of the Clark's Fork Basin–Polecat Bench area, northwestern Wyoming. In *Early Cenozoic paleontology and stratigraphy of the Bighorn Basin, Wyoming*, ed. P. D. Gingerich. *University of Michigan Papers on Paleontology* 24:51–68.
- Gingerich, P. D. and E. L. Simons. 1977. Systematics, phylogeny and evolution of early Eocene Adapidae (Mammalia, Primates) in North America. *University of Michigan, Contributions from the Museum of Paleontology* 24:245–279.
- Givens, C. R. 1974. Eocene molluscan biostratigraphy of the Pine Mountain area, Ventura County, California. *University of California Publications in Geological Sciences* 109.
- Givens, C. R. and M. P. Kennedy. 1976. Middle Eocene mollusks from northern San Diego County, California. *Journal of Paleontology* 50:954–975.
- Gunnell, G. F. 1985. Systematics of early Eocene Microsypinae (Mammalia, Primates) in the Clark's Fork Basin, Wyoming. *University of Michigan, Contributions from the Museum of Paleontology* 27:51–71.
- . 1989. Evolutionary history of the Microsypoidea (Mammalia, ?Primates) and the relationship between Plesiadapiformes and Primates. *University of Michigan Papers on Paleontology* 27:1–157.
- Guthrie, D. A. 1971a. The mammalian fauna of the Lost Cabin Member, Wind River Formation (lower Eocene) of Wyoming. *Annals of the Carnegie Museum* 43:47–113.
- . 1971b. A titanothere (Mammalia, Perissodactyla) from the early Eocene of Wyoming. *Journal of Mammalogy* 52:474–475.
- Hamblin, A. H. 1987. Paleogeography and paleoecology of Myton Pocket, Uinta Basin, Utah (Uinta Formation–Upper Eocene). *Brigham Young University Geological Studies* 34:33–60.
- Hanna, M. A. 1926. Geology of the La Jolla Quadrangle, California. *University of California Publications in Geological Sciences* 16:187–246.
- Hanson, C. B. 1996. Stratigraphy and vertebrate faunas of the Bridgerian–Duchesnean Clarno Formation, north-central Oregon. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 206–239.
- Haq, B. Q., J. Hardenbol, and P. R. Vail. 1987. Chronology of fluctuating sea levels since the Triassic. *Science* 235:1156–1167.
- Harland, W. B., R. L. Armstrong, A. V. Cox, L. E. Craig, A. G. Smith, and D. G. Smith. 1990. *A geologic time scale*. New York: Cambridge University Press.
- Hatcher, J. B. 1895. On a new species of *Diplacodon*, with a discussion of the relations of that genus to *Telmatherium*. *American Naturalist* 29:1084–1090.
- Hickey, L. J., R. M. West, and M. R. Dawson. 1984. Arctic biostratigraphic heterochrony. *Science* 224:175–176.
- Holbrook, L. T. and S. G. Lucas. 1997. A new genus of rhinocerotoid from the Eocene of Utah and the status of North American "*Forstercooperia*." *Journal of Vertebrate Paleontology* 17:384–396.
- Howard, J. L. 1989. Conglomerate clast populations of the upper Paleogene Sespe Formation, southern California. In *Conglomerates in basin analysis. A symposium dedicated to A. O. Woodford*, ed. I. P. Colburn, P. L. Abbott, and J. A. Minch. *Pacific Section SEPM* 62:269–280.
- Hutchison, J. H. 1971. Cf. *Uintatherium* (Dinocerata, Mammalia) from the Uintan (middle to late Eocene) of southern California. *Paleobios* 12:1–8.
- Jepsen, G. L. 1930. New vertebrate fossils from the lower Eocene of the Big Horn Basin, Wyoming. *Proceedings of the American Philosophical Society* 69:117–131.
- . 1940. Paleocene faunas of the Polecat Bench Formation, Park County, Wyoming. *Proceedings of the American Philosophical Society* 83:217–341.
- . 1966. Early Eocene bat from Wyoming. *Science* 154:1333–1339.
- Kay, J. L. and V. Garwood. 1953. *Guidebook, sixth annual field conference of the Society of Vertebrate Paleontology in northeastern Utah*. Pittsburgh: Utah Field House of Natural History and Carnegie Museum, pp. 1–34.
- Keefer, W. R. 1956. Tertiary rocks in the northwestern part of the Wind River Basin, Wyoming. *Wyoming Geological Association Field Conference Guidebook*, pp. 109–116.
- Kelly, T. S. and D. P. Whistler. 1994. Additional Uintan and Duchesnean (middle and late Eocene) rodents from the Sespe Formation, Simi Valley, California. *Los Angeles County Natural History Museum Contributions in Science* 439:1–29.
- Kitts, D. B. 1956. American *Hyracotherium* (Perissodactyla, Equidae). *Bulletin of the American Museum of Natural History* 110:1–60.
- Korth, W. W. 1984. Earliest Tertiary evolution and radiation of rodents in North America. *Bulletin of the Carnegie Museum of Natural History* 24:1–71.
- . 1985. The rodents *Pseudotomus* and *Quadratimus* and the content of the tribe Manitshini (Paramyinae, Ischyromyidae). *Journal of Vertebrate Paleontology* 5:139–152.
- . 1994. *The Tertiary record of rodents in North America*. New York: Plenum Press, pp. 1–319.
- Korth, W. W. and R. L. Evander. 1982. A new species of *Orohippus* (Perissodactyla, Equidae) from the early Eocene of Wyoming. *Journal of Vertebrate Paleontology* 2:167–171.
- Krishtalka, L. 1978. Paleontology and geology of the Badwater Creek area, central Wyoming. Pt. 15: A review of the late Eocene primates of Wyoming and Utah, and the Plesitarsiiformes. *Annals of the Carnegie Museum* 47:335–360.
- Krishtalka, L., C. C. Black, and D. W. Riedel. 1975. Paleontology and geology of the Badwater Creek area, central Wyoming. Pt. 10: A late Paleocene mammal fauna from the Shotgun Member of the Fort Union Formation. *Annals of the Carnegie Museum* 45:179–212.
- Krishtalka, L., R. J. Emry, J. E. Storer, and J. F. Sutton. 1982. Oligocene Multituberculates (Mammalia, Allotheria): Youngest known records. *Journal of Paleontology* 56:791–794.
- Krishtalka, L. and R. K. Stucky. 1986. Early Eocene artiodactyls from the San Juan Basin, New Mexico, and the Piceance Basin, Colorado. *University of Wyoming, Contributions to Geology Special Paper* 3.
- Lander, B. 1997. Geology and vertebrate paleontology of Cenozoic nonmarine rock units in Simi Valley. In *Simi Valley, a journey*

- through time, ed. P. Havens and B. Appleton. Simi Valley, CA: Simi Valley Historical Society and Museum, pp. 303–319.
- . 1998. Oreodontoida. In *Evolution of Tertiary mammals in North America*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, 1:402–425.
- Lillegraven, J. A. and R. W. Wilson. 1975. Analysis of *Simimys simplex*, an Eocene rodent (?Zapodidae). *Journal of Paleontology* 49:856–874.
- Link, M. H. and J. E. Welton. 1982. Sedimentology and reservoir potential of Matalija Sandstone: An Eocene sand-rich deep-sea fan and shallow-marine complex, California. *Bulletin of the American Association of Petroleum Geologists* 66: 1514–1534.
- Loomis, F. R. 1907. Origin of the Wasatch deposits. *American Journal of Science* 4th ser. 23:356–364.
- Love, J. D. 1970. Cenozoic geology of the Granite Mountains area, central Wyoming. *U.S. Geological Survey Professional Paper* 495-C.
- . 1978. Cenozoic thrust and normal faulting, and tectonic history of the Badwater area, northeastern margin of Wind River Basin, Wyoming. *Wyoming Geological Association Guidebook, 30th Annual Field Conference*, pp. 235–238.
- Love, J. D., A. C. Christiansen, J. L. Earle, and R. W. Jones. 1978. Preliminary geologic map of the Arminto 1' \_ 2' Quadrangle, central Wyoming. *U.S. Geological Survey Open-File Report* 78-1089.
- Love, J. D. and W. R. Keefer. 1975. Geology of sedimentary rocks in southern Yellowstone National Park, Wyoming. *U.S. Geological Survey Professional Paper* 729-D.
- Love, J. D., M. C. McKenna, and M. R. Dawson. 1976. Eocene, Oligocene, and Miocene rocks and vertebrate fossils at the Emerald Lake locality, three miles south of Yellowstone National Park, Wyoming. *U.S. Geological Survey Professional Paper* 932-A.
- Love, L. L., A. M. Kudo, and D. W. Love. 1976. Dacites of Bunsen Peak, the Birch Hills, and the Washakie Needles, northwest Wyoming, and the relationship to the Absaroka volcanic fields, Wyoming–Montana. *Bulletin of the Geological Society of America* 87:1455–1462.
- Lucas, S. G. 1982. Vertebrate paleontology, stratigraphy, and biostratigraphy of the Eocene Galisteo Formation, north-central New Mexico. *New Mexico Bureau of Mines and Mineral Resources Circular* 186:1–3.
- Lucas, S. G. and B. S. Kues. 1979. Vertebrate biostratigraphy of the Eocene Galisteo Formation, north-central New Mexico. *New Mexico Geological Society Guidebook, 30th Field Conference*, pp. 225–229.
- Lucas, S. G. and R. M. Schoch. 1982. *Duchesneodus*, a new name for some titanotheres (Perissodactyla, Brontotheriidae) from the late Eocene of western North America. *Journal of Paleontology* 56:1018–1023.
- Mader, B. J. 1998. Brontotheriidae. In *Evolution of Tertiary mammals of North America*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press 1:525–536.
- Marsh, O. C. 1870. Professor Marsh's Rocky Mountain expedition: Discovery of the Mauvaises Terres Formation in Colorado. *American Journal of Science* 2d ser. 50:292.
- . 1871. On the geology of the eastern Uinta Mountains. *American Journal of Science* 3rd ser. 2:191–198.
- . 1877. Introduction and succession of vertebrate life in America. *American Journal of Science* 3rd ser. 14:337–378.
- . 1885. Dinocerata. *U.S. Geological Survey Monograph* 10.
- Mason, M. A. and C. C. Swisher. 1989. New Evidence for the age of the South Mountain Local Fauna, Ventura County, California. *Los Angeles County Natural History Museum Contributions in Science* 410:1–9.
- Mauger, R. L. 1977. K–Ar ages of biotites from tuffs in Eocene rocks of the Green River, Washakie, and Uinta basins, Utah, Wyoming, and Colorado. *University of Wyoming, Contributions to Geology* 15:17–41.
- Maxwell, R. A., J. T. Lonsdale, R. T. Hazzard, and J. A. Wilson. 1967. Geology of Big Bend National Park. *University of Texas Publications* 6711.
- McDowell, F. W. 1979. Potassium–argon dating in the Trans-Pecos Texas volcanic field. *Bureau of Economic Geology, University of Texas Guidebook* 19:10–18.
- McDowell, F. W., J. A. Wilson, and J. Clark. 1973. K–Ar dates for biotite from two paleontologically significant localities: Duchesne River Formation, Utah, and Chadron Formation, South Dakota. *Isochron/West* 7:11–12.
- McGrew, P. O. and H. W. Roehler. 1960. Correlations of Tertiary units in southwestern Wyoming. *Wyoming Geological Association Guidebook, 15th Annual Field Conference*, pp. 156–158.
- McKenna, M. C. 1972. Vertebrate paleontology of the Togwotee Pass area, northwest Wyoming. In *Guidebook, Field Conference, Tertiary Biostratigraphy of Southern and Western Wyoming*, pp. 80–101.
- . 1976. *Esthonyx* in the upper faunal assemblage, Huerfano Formation, Eocene of Colorado. *Journal of Paleontology* 50:354–355.
- McKenna, M. C. and J. D. Love. 1970. Local stratigraphic and tectonic significance of *Leptoceratops*, a Cretaceous dinosaur in the Pinyon Conglomerate, northwestern Wyoming. *U.S. Geological Survey Professional Paper* 700-D:D55–D61.
- Mellet, J. S. 1977. Paleobiology of North American *Hyaenodon* (Mammalia, Creodonta). *Contributions to Vertebrate Evolution*, no. 1. Basel: S. Karger.
- Morris, W. J. 1954. An Eocene fauna from the Cathedral Bluffs Tongue of the Washakie Basin, Wyoming. *Journal of Paleontology* 28:195–203.
- Neasham, J. W. and C. F. Vondra. 1972. Stratigraphy and petrology of the lower Eocene Willwood Formation, Big Horn Basin, Wyoming. *Bulletin of the Geological Society of America* 83:2167–2180.
- Nightingale, W. T. 1930. Geology of Vermilion Creek gas area in southwest Wyoming and northwest Colorado. *American Association of Petroleum Geologists Bulletin* 14:1013–1040.
- Norris, G. and A. D. Miall. 1984. Arctic biostratigraphic heterochroneity. *Science* 224:174–175.
- Okada, H. and D. Bukry. 1980. Supplementary modification and introduction of code numbers to the low-latitude coccolith biostratigraphic zonation (Bukry 1973, 1975). *Marine Micropaleontology* 5:321–324.
- Osborn, H. F. and W. D. Matthew. 1909. Cenozoic mammal horizons of western North America. *Bulletin of the U.S. Geological Survey* 361:1–138.
- Ostrander, G. E. 1980a. An early Oligocene vertebrate fauna from northwestern Nebraska. *Geological Society of America Abstracts with Programs* 12(6):300.
- . 1980b. *Mammalia of the Early Oligocene (Chadronian) Raben Ranch local fauna of northwestern Nebraska*. Unpublished M.S. thesis, South Dakota School of Mines Technology, pp. 1–28.

- Ostrander, G., C. A. Jones, and R. Cape. 1979. The occurrence of a multituberculate in the lower Oligocene Chadron Formation of northwest Nebraska. *Geological Society of America Abstracts with Programs* 11:299.
- Patterson, B. 1934. A contribution to the osteology of *Titanoides* and the relationships of the Amblypoda. *Proceedings of the American Philosophical Society* 73:71–101.
- . 1935. Second contribution to the osteology and affinities of the Paleocene amblypod *Titanoides*. *Proceedings of the American Philosophical Society* 75:143–162.
- . 1937. A new genus, *Barylambda*, for *Titanoides faberi*, Paleocene amblypod. *Field Museum of Natural History Geology Series* 6:229–231.
- . 1939a. New Pantodonta and Dinocerata from the upper Paleocene of western Colorado. *Field Museum of Natural History Geology Series* 6:351–383.
- . 1939b. A skeleton of *Coryphodon*. *Proceedings of the New England Zoological Club* 17:97–110.
- . 1949. A new genus of taeniodonts from the late Paleocene. *Fieldiana: Geology* 10:41–42.
- Patterson, B. and E. L. Simons. 1958. A new barylambdid pantodont from the late Paleocene. *Breviora* 93.
- Patterson, B. and R. M. West. 1973. A new late Paleocene phenacodont (Mammalia, Condylarthra) from western Colorado. *Breviora* 403.
- Pekarek, A., R. F. Maruin, and H. H. Mehnert. 1974. K–Ar ages of the volcanics in the Rattlesnake Hills, central Wyoming. *Geology* 2:282–285.
- Peterson, O. A. 1919. Report upon the material discovered in the upper Eocene of the Uinta Basin by Earl Douglass in the years 1908–1909, and by O. A. Peterson in 1912. *Annals of the Carnegie Museum* 12:40–169.
- . 1934. List of species and description of new material from the Duchesne River Oligocene, Uinta Basin, Utah. *Annals of the Carnegie Museum* 23:373–389.
- Premoli Silva, I. and D. G. Jenkins. 1993. Decision on the Eocene–Oligocene boundary stratotype. *Episodes* 16:379–382.
- Radinsky, L. 1963. Origin and early evolution of North American Tapiroidea. *Peabody Museum of Natural History Yale University Bulletin* 17:1–106.
- . 1967a. *Hyrachyus*, *Chasmothereium*, and the early evolution of helaletid tapiroids. *American Museum Novitates* 2313.
- . 1967b. Review of the rhinocerotoid Family Hyracodontidae (Perissodactyla). *Bulletin of the American Museum of Natural History* 136:1–45.
- Rasmussen, D. T., M. Shekelle, S. L. Walsh, and B. O. Riney. 1995. The dentition of *Dyseolemur*, and comments on the use of anterior teeth in primate systematics. *Journal of Human Evolution* 29:301–320.
- Robinson, P. 1968. The paleontology and geology of the Badwater Creek area, central Wyoming. Pt. 4: Late Eocene Primates from Badwater, Wyoming, with a discussion of material from Utah. *Annals of the Carnegie Museum* 39:307–326.
- Rohrer, W. L. and J. D. Obradovich. 1969. Age and stratigraphic relations of the Tepee Trail and Wiggins formations, northwestern Wyoming. *U.S. Geological Survey Professional Paper* 650-B:57–62.
- Rowley, P. D., W. R. Hansen, O. Tweto, and P. E. Carrara. 1985. Geologic map of the Vernal 1° 2' Quadrangle, Colorado, Utah and Wyoming. *U.S. Geological Survey Misc. Investigations Series* Map I-1526.
- St. John, O. H. 1883. Report on the geology of the Wind River district. *U.S. Geological Survey Territories 12th Annual Report*, pt. 1, pp. 173–269.
- Savage, D. E. 1955. Nonmarine lower Pliocene sediments in California. *University of California Publications in Geological Sciences* 31:1–26.
- . 1977. Aspects of vertebrate paleontological stratigraphy and geochronology. In *Concepts and methods of biostratigraphy*, ed. E. G. Kauffmann and J. E. Hazel. Stroudsburg, PA: Dowden, Hutchinson, and Ross, pp. 427–442.
- Savage, D. E. and D. E. Russell. 1983. *Mammalian paleofaunas of the world*. Reading, MA: Addison-Wesley.
- Savage, D. E. and B. T. Waters. 1978. A new omomyid primate from the Wasatch Formation of southern Wyoming. *Folia Primatologica* 30:1–29.
- Shankler, D. M. 1981. Local extinction and ecological re-entry of early Eocene mammals. *Nature* 293:135–138.
- Schassberger, H. T. 1972. A K–Ar age of a quartz monzonite dike in the Kerwin [sic] mining district, Park County, Wyoming. *Ischron/West* 4:31.
- Schiebout, J. A. 1977. Eocene Perissodactyla from the La Jolla and Poway Groups, San Diego County, California. *San Diego Society of Natural History Transactions* 18:217–227.
- Scott, W. B. and H. F. Osborn. 1890. The Mammalia of the Uinta Formation. *Transactions of the American Philosophical Society* 16:461–572.
- Shive, P. N., K. A. Sundell, and J. Rutledge. 1980. Magnetic polarity stratigraphy of Eocene volcanoclastic rocks from the Absaroka Mountains of Wyoming (abstract). *Eos, Transactions, American Geophysical Union* 61:945.
- Simpson, G. G. 1933. Glossary and correlation charts of North American Tertiary mammal-bearing formations. *Bulletin of the American Museum of Natural History* 67:79–121.
- . 1946. Discussion of the Duchesnean fauna and the Eocene–Oligocene boundary. *American Journal of Science* 244:52–57.
- Sinclair, W. J. and W. Granger. 1912. Note on the Tertiary deposits of the Big Horn Basin. *Bulletin of the American Museum of Natural History* 31:57–67.
- Smedes, H. W. and H. J. Prostka. 1972. Stratigraphic framework of the Absaroka volcanic supergroup in the Yellowstone National Park region. *U.S. Geological Survey Professional Paper* 729-C.
- Snyder, D. O. 1970. Fossil evidence of Eocene age of Baca Formation, New Mexico. *New Mexico Geological Society Guidebook* 21:65–67.
- Squires, R. L. 1984. Megapaleontology of the Eocene Lajas Formation, Simi Valley, California. *Los Angeles County Museum of Natural History Contributions in Science* 350:1–76.
- . 1988. Geologic age refinements of West Coast Eocene marine mollusks. In *Paleogene stratigraphy, West Coast of North America*, ed. M. V. Filewicz and R. L. Squires. *Pacific Section SEPM* 58:107–112.
- Stagner, W. L. 1941. The paleogeography of the eastern part of the Uinta Basin during Uinta B time. *Annals of the Carnegie Museum* 28:273–308.
- Steiger, R. H. and E. Jager. 1977. Subcommission on geochronology: Conventions on use of decay constants in geo- and cosmochronology. *Earth and Planetary Science Letters* 36:359–362.
- Steineck, P. L. and J. M. Gibson. 1971. Age and correlation of the Eocene Ulatisian and Narizian stages, California. *Bulletin of the Geological Society of America* 82:447–480.

- Steineck, P. L., J. M. Gibson, and R. W. Morin. 1972. Foraminifera from the middle Eocene Rose Canyon and Poway formations, San Diego, California. *Journal of Foraminiferal Research* 2:137–144.
- Stevens, J. B., M. S. Stevens, and J. A. Wilson. 1984. Devil's Graveyard Formation (new), Eocene and Oligocene age, Trans-Pecos Texas. *Texas Memorial Museum Bulletin* 32:1–21.
- Stock, C. 1932. Eocene land mammals on the Pacific Coast. *Proceedings of the National Academy of Sciences* 18:518–523.
- . 1936. Perissodactyla of the Sespe Eocene, California. *Proceedings of the National Academy of Sciences* 22:260–265.
- . 1948. Pushing back the history of land mammals in western North America. *Bulletin of the Geological Society of America* 59:327–332.
- Stokes, W. L. and J. H. Madsen Jr. 1961. *Geologic maps of Utah (northeast quarter)*. Salt Lake City: College of Mines and Mining Industry, University of Utah.
- Storer, J. E. 1987. Dental evolution and radiation of Eocene and early Oligocene Eomyidae (Mammalia, Rodentia) of North America with new material from the Duchesnean of Saskatchewan. *Dakoterra* 3:108–117.
- . 1993. Multituberculates of the Lac Pelletier lower fauna, Late Eocene (Duchesnean), of Saskatchewan. *Canadian Journal of Earth Sciences* 30:1613–1617.
- Stucky, R. K. 1982. *Mammalian fauna and biostratigraphy of the upper part of the Wind River Formation (early to middle Eocene), Natrona County, Wyoming, and the Wasatchian–Bridgerian boundary*. Ph.D. dissertation, University of Colorado, Boulder.
- . 1996. The mammalian faunas of North America of Bridgerian to early Arikareean “ages” (Eocene to Oligocene). In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 464–493.
- . 1998. Eocene bunodont and bunoselenodont Artiodactyla (“dichobunids”). In *Evolution of Tertiary mammals of North America*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, 1:358–374.
- Stucky, R. K. and L. Krishtalka. 1982. Revision of the Wind River faunas, early Eocene of central Wyoming. Pt. 1: Introduction and Multituberculata. *Annals of the Carnegie Museum* 51:39–56.
- Stucky, R. K., L. Krishtalka, and M. R. Dawson. 1989. Paleontology, geology and remote sensing of Paleogene rocks in the northeastern Wind River Basin, Wyoming, USA. In *Mesozoic/Cenozoic vertebrate paleontology: Classic localities, contemporary approaches*, ed. J. J. Flynn. 28th International Geological Congress Field Trip Guide T322:34–44.
- Sullivan, R. 1980. A stratigraphic evaluation of Eocene rocks of southwestern Wyoming. *Geological Survey of Wyoming Report of Investigations* 20:1–50.
- Sutton, J. F. and C. C. Black. 1972. Oligocene and Miocene deposits of Jackson Hole, Wyoming. *Guidebook, Field Conference on Tertiary Biostratigraphy, Southern and Western Wyoming*, pp. 73–79.
- Szalay, F. S. 1969. Mixodectidae, Microsypidae, and the insectivore-primate transition. *Bulletin of the American Museum of Natural History* 140:193–330.
- Tedford, R. H., M. F. Skinner, R. W. Fields, J. M. Rensberger, D. P. Whistler, T. Galusha, B. E. Taylor, J. R. McDonald, and S. D. Webb. 1987. Faunal succession and biochronology of the Arikarean through Hemphillian interval (late Oligocene through earliest Pliocene) in North America. In *Cenozoic mammals of North America, geochronology and biostratigraphy*, ed. M. O. Woodburne. Berkeley: University of California Press, pp. 153–210.
- Testarmata, M. M. and W. A. Gose. 1979. Magnetostratigraphy of the Eocene–Oligocene Vieja Group, Trans-Pecos Texas. In *Cenozoic geology of the Trans-Pecos volcanic field of Texas*, ed. A. W. Walton and C. D. Henry. *Bureau of Economic Geology, University of Texas, Guidebook* 19:55–66.
- . 1980. Magnetostratigraphy in the Trans-Pecos volcanic field: Preliminary results from the Eocene–Oligocene Vieja Group. *New Mexico Geological Society Guidebook, 31st Annual Field Conference*, pp. 101–103.
- Tourtletot, H. A. 1948. Tertiary rocks in the northeastern part of the Wind River Basin, Wyoming. *Guidebook, 3d Annual Field Conference, Society of Vertebrate Paleontology in Southeastern Wyoming*, pp. 53–67.
- . 1957. The geology and vertebrate paleontology of upper Eocene strata in the northeastern part of the Wind River Basin, Wyoming. Pt. 1: Geology. *Smithsonian Miscellaneous Collections* 134(4).
- Truex, J. N. 1976. Santa Monica and Santa Ana mountains: Relation to Oligocene Santa Barbara Basin. *American Association of Petroleum Geologists Bulletin* 60:65–86.
- Turnbull, W. D. 1991. *Protoptychus hatcheri* Scott, 1985. The mammalian faunas of the Washakie Formation, Eocene Age, of southern Wyoming. Part II. The Adobetown Member, middle division (= Washakie B), Twka/2 (in part). *Fieldiana: Geology* new series 21:1–33.
- Tweto, O. 1979. *Geologic map of Colorado*. Boulder: U.S. Geological Survey.
- Untermann, G. E., B. R. Untermann, and D. M. Kinney. 1964. Geologic map of Uintah County, Utah (south half). *Bulletin of the Utah Geological Mineralogical Survey* 72 (suppl.).
- Vedder, J. G. 1972. Revision of stratigraphic names for some Eocene formations in Santa Barbara and Ventura counties, California. *Bulletin of the U.S. Geological Survey* 1354-C.
- Walton, A. H. 1992. Magnetostratigraphy and the ages of Bridgerian and Uintan faunas in the lower and middle members of the Devil's Graveyard Formation, Trans-Pecos, Texas. In *Eocene–Oligocene climatic and biotic evolution*, ed. D. R. Prothero and W. A. Berggren. Princeton, NJ: Princeton University Press, pp. 74–87.
- Weaver, C. E., S. Beck, M. N. Bramlette, S. Carlson, B. L. Clark, T. W. Dibblee Jr., W. Durham, G. C. Ferguson, L. C. Forest, U. S. Grant IV, M. Hill, F. R. Kelley, R. M. Kleinpell, W. D. Kleinpell, J. Marks, W. C. Putnam, H. G. Schenk, N. T. Taliaferro, R. R. Thorup, E. Watson, and R. T. White. 1944. Correlation of the marine Cenozoic formations of western North America. *Bulletin of the Geological Society of America* 55:569–598.
- West, R. M. 1969. Biostratigraphy of fluvial sediments of the upper Wasatch Formation in the northern Green River Basin, Wyoming. *University of Wyoming, Contributions to Geology* 8:184–196.
- Westgate, J. W. and R. J. Emry. 1985. Land mammals of the Crow Creek local fauna, late Eocene, Jackson Group, St. Francis County, Arkansas. *Journal of Paleontology* 59:242–248.
- Wheeler, W. H. 1961. Revision of the uintatheres. *Peabody Museum of Natural History Yale University Bulletin* 14.

- Wilson, J. A. 1971. Early Tertiary vertebrate fauna, Vieja Group, Trans-Pecos Texas: Agriochoeridae and Merycoidodontidae. *Texas Memorial Museum Bulletin* 18.
- . 1980. Geochronology of the Trans-Pecos Texas volcanic field. *New Mexico Geological Society Guidebook, 31st Annual Field Conference*, pp. 205–211.
- Wilson, J. A., P. C. Twiss, R. K. DeFord, and S. E. Clabaugh. 1968. Stratigraphic succession, potassium–argon dates, and vertebrate faunas, Vieja Group, Rim Rock County, Trans-Pecos Texas. *American Journal of Science* 266:590–604.
- Wing, S. L. 1998. Late Paleocene–early Eocene floral and climatic change in the Bighorn Basin, Wyoming. In *Late Paleocene–early Eocene climatic and biotic events in the marine and terrestrial records*, ed. M.-P. Aubry, S. G. Lucas, and W. A. Berggren. New York: Columbia University Press, pp. 308–400.
- Wood, A. E. 1962. The early Tertiary rodents of the Family Paramyidae. *Transactions of the American Philosophical Society* 52.
- . 1973. Eocene rodents, Pruett Formation, southwest Texas: Their pertinence to the origin of the South American Caviomorpha. *Pearce-Sellards Series* 20.
- . 1974. Early Tertiary vertebrate faunas, Vieja Group, Trans-Pecos Texas: Rodentia. *Texas Memorial Museum Bulletin* 21.
- Woodburne, M. O. 1977. Definition and characterization in mammalian chronostratigraphy. *Journal of Paleontology* 51:220–234.
- Woodring, W. P. 1931. Age of the orbitoid-bearing limestone and *Turritella varinta* zone of the western Santa Ynez Range, California. *Transactions of the San Diego Natural History Society* 6:371–388.
- Woodring, W. P. and W. P. Popenoe. 1945. *U.S. Geological Survey Oil and Gas Investigations Preliminary Chart* 12.
- Wortman, J. L. 1901. Studies of Eocene Mammalia in the Marsh Collection, Peabody Museum. Pt. 1: Carnivora. *American Journal of Science*, 4th ser. 11:333–348.

# 5

---

## The Chadronian, Orellan, and Whitneyan North American Land Mammal Ages

Donald R. Prothero and Robert J. Emry

**I**N THE 16 YEARS SINCE EMRY, BJORK, and Russell (1987) reviewed the Chadronian, Orellan, and Whitneyan land mammal ages for the original version of this volume, an enormous amount of information has been published concerning this interval of time. Many of the crucial sections have been studied by means of magnetic stratigraphy, and many new  $^{40}\text{Ar}/^{39}\text{Ar}$  dates have been analyzed, which have radically changed our concept of the correlation of these beds (Swisher and Prothero 1990; Prothero and Swisher 1992; Prothero 1996b; Prothero and Whittlesey 1998). Systematic reviews of most of the biostratigraphically informative taxa have been completed (see chapters in Prothero and Emry 1996b), allowing a formal range zone biostratigraphy to be erected for most of this interval (Prothero and Emry 1996a; Prothero and Whittlesey 1998). Calibrated by means of magnetostratigraphy, these biostratigraphic zones now offer very high-resolution dating of most of this time interval. In Emry et al. (1987), it was not possible to resolve many events to the nearest 2–3 million years, but the current high-resolution chronostratigraphic framework now allows many events in better-studied sections to be dated to the nearest 100,000 years or less.

In addition to the improvements in chronostratigraphy, the new dates have radically changed our correlation of the Chadronian, Orellan, and Whitneyan land mammal ages to the global time scale. When these terms were originally coined by the Wood committee (1941), they were thought to correlate with the early, middle, and late Oligocene, respectively. But the new  $^{40}\text{Ar}/^{39}\text{Ar}$  dates, combined with the magnetostratigraphy (Swisher and Prothero 1990; Prothero and Swisher 1992),

and changes in the dating of the global time scale (Berggren et al. 1992, 1995) showed that the Chadronian correlates with the late Eocene, the Orellan and Whitneyan with the early Oligocene, and the Arikareean (long considered early Miocene) with the late Oligocene and earliest Miocene (as first shown by Evernden et al. 1964 and Tedford et al. 1987). Thus the original concept of the Chadronian, Orellan, and Whitneyan land mammal ages as the “Oligocene” chapter in the original book has lost its meaning. This chapter now covers the late Eocene to early Oligocene, and the rest of the Eocene and Oligocene is covered in other chapters. Nevertheless, the Chadronian, Orellan, and Whitneyan land mammal ages are important because they represent the best-studied terrestrial record of the Eocene–Oligocene transition in the world and are therefore an important data set for studying the climatic changes that occurred when the global climate shifted from greenhouse to icehouse states (Prothero 1994; Prothero and Emry 1996b; Prothero and Heaton 1996; Prothero 1999).

### LITHOSTRATIGRAPHY OF CHADRONIAN, ORELLAN, AND WHITNEYAN STRATA

---

In this chapter, we will not repeat the historical introduction to the White River Group (figure 5.1), more fully discussed in Emry et al. (1987), or much of the stratigraphic background discussed in the latter part of that chapter. Instead, we will summarize the recent developments in



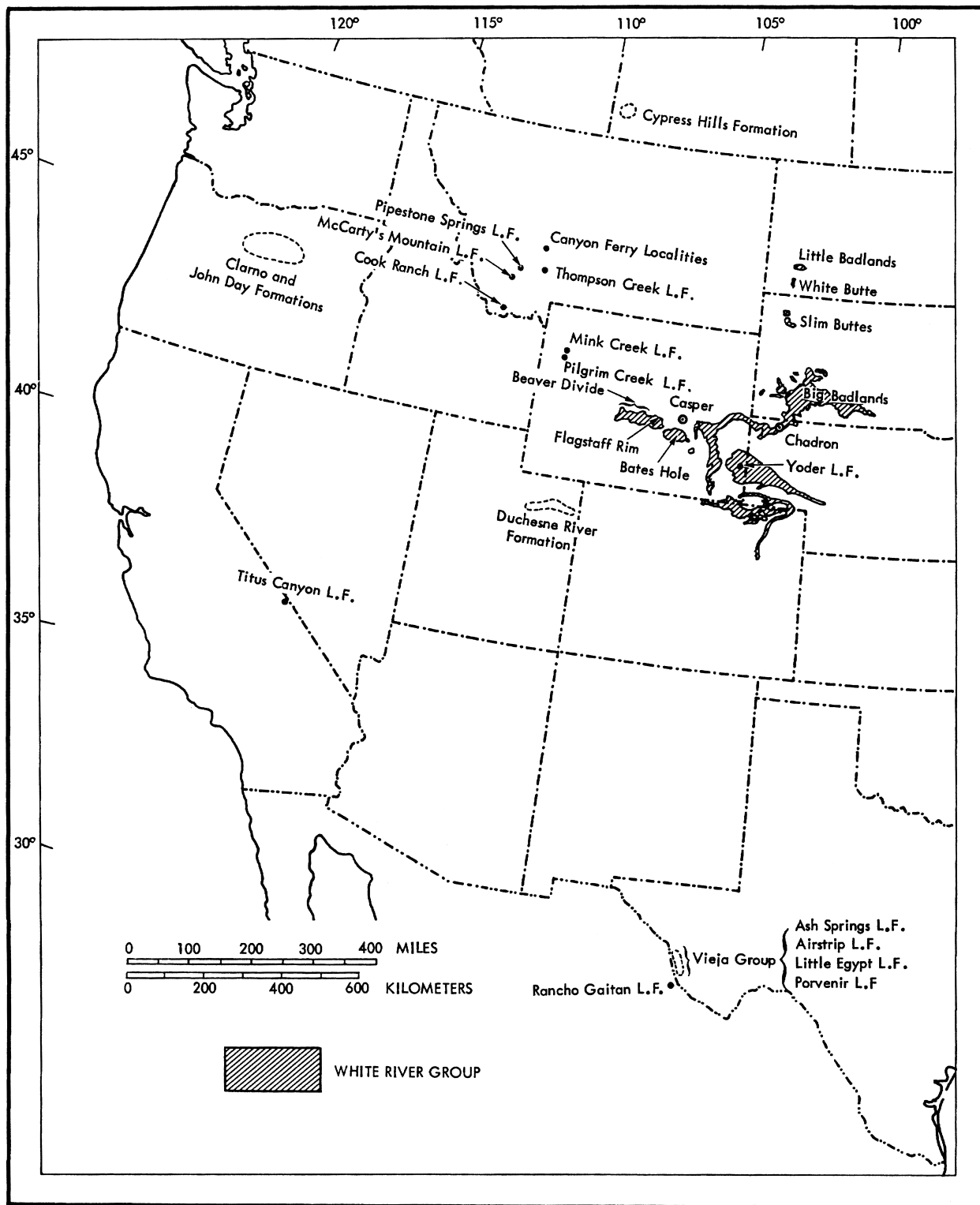


FIGURE 5.1 Index map showing location of rock units mentioned in text (after Emry et al. 1987). L.F., Local Fauna.

biostratigraphy, magnetostratigraphy, and  $^{40}\text{Ar}/^{39}\text{Ar}$  dating that were published since 1987 (Swisher and Prothero 1990; Prothero and Swisher 1992; various chapters in Prothero and Emry 1996b, including Prothero and Emry 1996a; Prothero and Whittlesey 1998).

Some important changes in the lithostratigraphic framework have taken place, however (figure 5.2). Terry (1998) proposed a new lithostratigraphic framework for the lower part of the White River Group in South Dakota and Nebraska. The lowest part of the former Chadron Formation in South Dakota (containing the Interior and Weta paleosols) was named the Chamberlain Pass Formation. Of the three members of the Chadron Formation recognized in South Dakota by Clark (1937), only the Peanut Peak Member is recognized in Nebraska. Terry (1998) rejects the old correlation of the Ahearn and Crazy Johnson Members with Chadron A and B in Nebraska. Chadron B and C and Orella A in Nebraska (*sensu* Schultz and Stout 1955, 1961) are now placed in the new Big Cottonwood Creek Member of the Chadron Formation by Terry (1998) and Terry and LaGarry (1998).

LaGarry (1998) also proposed revisions of the Brule Formation in Nebraska. As just mentioned, "Orella A" in Nebraska has been reassigned to the Big Cottonwood Creek Member of the Chadron Formation. With this change, the Orellan North American land mammal age (NALMA) now coincides even better with the Orella Member as redefined. LaGarry (1998) redefined the boundary between the Orella and Whitney members to a position slightly lower in the section.

Finally, Swinehart et al. (1985) and Tedford et al. (1996) recognized a unit above the Whitney Member of the Brule Formation that they informally called the "brown siltstone" member. Many of the rocks that were once assigned to the upper Whitney Member or lower Gering Formation by Schultz and Stout (1955, 1961) were reassigned to this unit.

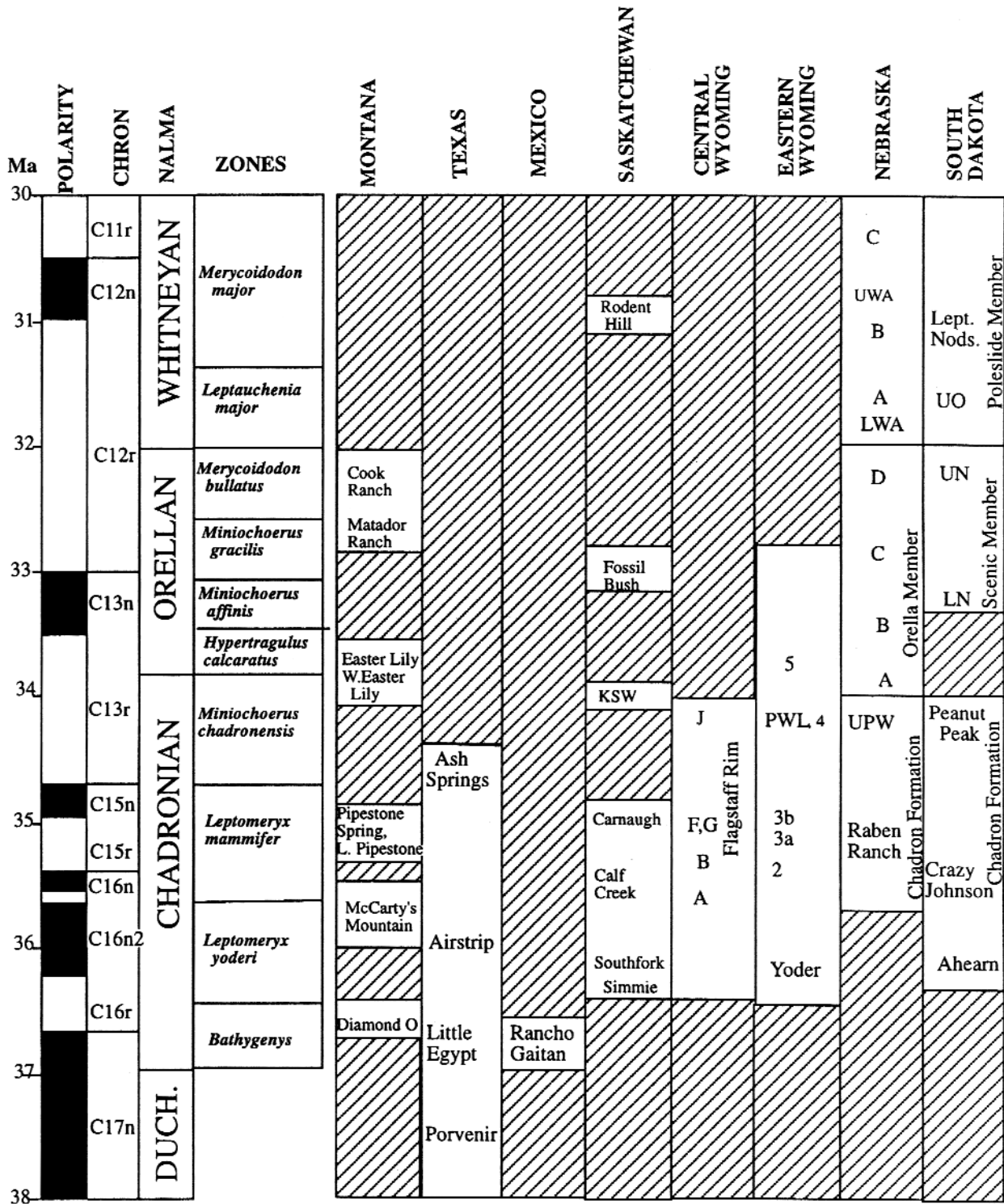
#### GEOGRAPHY OF THE CHADRONIAN, ORELLAN, AND WHITNEYAN NALMAS

All three of these NALMAs were originally based of parts of the White River Group and its contained faunas in northwestern Nebraska and southwestern South Dakota (Wood et al. 1941). With the exception of the earliest Chadronian, the subdivisions of these units that we now recognize are still based on parts of the White River Group and its faunas, although several of the subdivisions are based on White River sequences that are well outside the original type areas of the whole NALMAs. This primarily reflects the fact that faunas of Chadronian age are

much better known in these other areas. Therefore it seems worthwhile to generalize briefly about the geographic distribution of the White River Group and the non-White River faunal correlates.

The White River Group is exposed widely in southwestern South Dakota and northwestern Nebraska and north of the Pine Ridge escarpment in adjacent Niobrara County, Wyoming (figure 5.1). From there it can be traced, nearly continuously, westward to the vicinity of Douglas, Wyoming, and from there southward east of the Laramie Range nearly to the Colorado border and westward north of the Laramie Range to the vicinity of Glenrock. The White River Group crops out extensively in the valley of the North Platte River and its tributaries in western Nebraska and eastern Wyoming. In northeastern Colorado, extensive outcrops are found along the south-facing escarpment where the High Plains break off into the Colorado Piedmont. In northwestern South Dakota, southeastern Montana, and southwestern North Dakota, isolated remnants of White River are preserved, mainly in the upper parts of buttes and higher elevations. Figure 5.1 indicates the approximate areal extent, at the surface, of the White River Group.

Intermontane basins of central Wyoming have extensive White River deposits that are not continuous with the main body of the White River Group farther east. In the Bates Hole Area, at Flagstaff Rim at the southeastern end of the Wind River Basin, and at Beaver Divide along the southern margin of the Wind River Basin, the White River deposits have the general lithologic characteristics that justify the White River terminology but are generally so uniform that they are not subdivided lithologically and are locally treated as a formation rather than group. At their westernmost limit, deposits that can be recognized lithologically as White River lap onto the eastern end of the Wind River Range and extend from there southeast into the northwestern edge of the Great Divide Basin in Wyoming and northeast along Beaver Divide. These areas of White River Formation undoubtedly are remnants of a blanketing deposit that once extended north across the Wind River Basin and continued northeast across the Powder River Basin (where remnants remain at the top of Pumpkin Buttes) and east into the classic areas of White River in eastern Wyoming, western Nebraska, and southwestern South Dakota. The White River deposits retain their general lithologic character over such a broad area (across what would have been separate depositional basins with respect to fluvial components) because the bulk of the formation consists of eolian volcanoclastics, transported and deposited across this broad area irrespective of the fluvial systems.



**FIGURE 5.2** Correlation chart of deposits of Chadronian, Orellan, and Whitneyan age in North America. Time scale after Berggren et al. (1995), Prothero and Emry (1996a), and Prothero and Whittlesey (1998). Lithostratigraphic and magnetostratigraphic correlations after Emry et al. (1987), Swisher and Prothero (1990), Prothero and Swisher (1992), Prothero (1996a, 1996b), Tabrum et al. (1996), Tedford et al. (1996), Prothero and Whittlesey (1998), Terry (1998), and various other chapters in Prothero and Emry (1996b). KSW, Keasey Springs West; Lept. Nods., *Leptauchenia* Nodules; LN, Lower Nodules; LWA, Lower Whitney Ash; PWL, Persistent (= Purplish) White Layer; UN, Upper Nodules; UO, Upper Oreadon Beds; UPW, Upper Purplish White Layer; UWA, Upper Whitney Ash.

Chadronian faunas occur in several areas in rocks that are beyond the geographic limits of the White River Group. Among the more important of these are the Chambers Tuff (with the Porvenir and Little Egypt local faunas) and the Capote Mountain Tuff (with the Airstrip Local Fauna) of the Vieja Group in Trans-Pecos Texas. Other important Chadronian faunas occur in the Cypress Hills Formation in southwestern Saskatchewan and in the Climbing Arrow, Dunbar Creek, and Renova formations (the latter with the important Pipestone Springs Local Fauna), all in southwestern Montana (Tabrum et al. 1996). Mammalian assemblages from these units can be correlated, with varying degrees of precision, with the White River faunal sequence. The Vieja Group of Trans-Pecos Texas is especially important for the recognition and definition of earlier Chadronian subdivisions.

Through the Chadronian–Whitneyan interval, the known geographic range of faunas becomes progressively limited. Orellan faunas occur in southwestern Montana in the Dunbar Creek Formation of the Three Forks, Clarkston, and South Townsend basins, in the Dunbar Creek Member of the Renova Formation in the Jefferson Basin, and in the Cook Ranch Local Fauna east of Lima. Otherwise, Orellan faunas are largely limited to the Brule Formation in its typical areas: isolated localities in southwestern North Dakota, northwestern South Dakota, the Scenic Member in the Big Badlands of South Dakota, the Orella Member in western Nebraska and eastern Wyoming, and the equivalent Cedar Creek Member in northeastern Colorado.

Whitneyan faunas are even more limited geographically, essentially to the Brule Formation: the Whitney Member in western Nebraska and eastern Wyoming, the Vista Member in northeastern Colorado, and the Poleslide Member in South Dakota. Possible outliers are the Cedar Ridge Local Fauna in the Badwater Creek area in the northeastern part of the Wind River Basin of Wyoming (Black 1968; Setoguchi 1978) and the I-95 Local Fauna (Patton 1969) in northern Florida.

To our knowledge, no assemblages of Chadronian or Whitneyan land mammals occur in deposits that can be related unambiguously to any marine sequence. Orellan mammals have been reported from the lower Oligocene Byram Formation, Mississippi (Manning et al., 1986; Manning, 1997). Correlation with other chronologies is accomplished principally through radiometric dates and magnetic polarity stratigraphy. It does appear that brontotheres, whose latest occurrences caused confusion in defining and recognizing the Chadronian–Orellan boundary, can be rehabilitated as a useful biochronologic taxon; the extinction of bron-

totheres appears now to have been a global event at the end of the Eocene and thus provides supporting evidence for correlating the Chadronian–Orellan boundary with the Eocene–Oligocene boundary.

## CHADRONIAN

The original definition of the Chadronian by Wood et al. (1941) has caused considerable confusion about recognizing the Chadronian–Orellan boundary. Wood et al. (1941:11) wrote that “the Chadronian is based on the Chadron formation, type locality near Chadron, Nebraska, type area, northwestern Nebraska and southwestern South Dakota, includes the old term ‘*Titanotherium* beds,’ used in its most extended sense. It may also be defined, faunally, as the time during which *Mesohippus* and titanotheres coexisted.” As several authors have pointed out (Emry et al. 1987; Prothero and Whittlesey 1998), this definition combines lithostratigraphy (the geochron of the Chadron Formation) and biochronology (the overlapping ranges of *Mesohippus* and brontotheres, or titanotheres). As long as *Mesohippus* was not known in strata older than the Chadron Formation and brontotheres were not known above the Chadron Formation, the dual definition was not internally inconsistent and remained workable.

Confusion regarding the Chadronian–Orellan boundary results ultimately from the uncritical definition of the Chadron Formation–Orella Member boundary by Schultz and Stout (1938). In the Toadstool Park area of northwestern Nebraska (the type localities for the Orella and Whitney members of the Brule Formation), the uppermost of several “purplish white layers” (volcanic ash beds) was defined by Schultz and Stout (1938, 1955) as marking the top of the Chadron Formation. What is apparently the same purplish white ash occurs widely in the Seaman Hills area to the west in Niobrara County, Wyoming, and can be traced eastward into Sioux County, Nebraska, where it occurs in a sequence similar to that at the Orella type locality. In the Seaman Hills area, the lithology is the same above and below this ash bed, so there is no logical reason to place a formational, lithologic boundary at this ash bed. Moreover, as mentioned by Emry et al. (1987) and Prothero and Whittlesey (1998), brontothere (titanothere) bones have been found up to 25 feet above the ash bed in the Seaman Hills area. On September 6, 1953, Morris Skinner (unpublished section book, vol. 5, pp. 9–11, in Department of Vertebrate Paleontology Archives, American Museum of Natural History) discovered titanothere bones in a channel-fill de-

posit that cut down from about 6.5 m (20 feet) above the "Persistent White Layer" (PWL), Skinner's term for the "purplish white layer" of Schultz and Stout. In his unpublished 1960 summary of Seaman Hills stratigraphy, Skinner indicated "Chadronian age at least to here" at the level 6.5 m (20 feet) above the PWL. There are other occurrences of brontotheres (titanotheres) that would be above the top of the Chadron Formation as it was recognized by Schultz and Stout (1938, 1955). Emry et al. (1987) and Prothero and Whittlesey (1998) mention another occurrence in the White River southeast of Douglas, Wyoming, about 25 feet above a white ash bed that Skinner called the "100' correlator white zone" and Evanoff et al. (1992) call the "5 tuff," which is presumed to be the same ash bed as the upper PWL of Schultz and Stout. Even Schultz and Stout (1955, figure 10) indicate a titanotherid metapodial in a channel sandstone incised from about 7 feet above the upper PWL at Scottsbluff National Monument in the Platte Valley of western Nebraska, and in a footnote (1955:27) they comment that "all parts of the Chadron and the lowest parts of the Brule (basal Orella or Orella A) should be expected to yield titanotherid remains."

It is important to note that Skinner did not consider these brontotherid occurrences to be in the Brule Formation. On the contrary, in his section books and the unpublished summary mentioned earlier and in his daily diaries that he maintained, Skinner referred to the beds above the PWL (in northwestern Nebraska and Niobrara County, Wyoming) as the "Trunk Butte Member of the Chadron Formation." In the Seaman Hills area in Niobrara County, Wyoming, Skinner recognized about 30 m (100 feet) of this unit above the PWL and lesser thicknesses to the eastward into Nebraska as far as the vicinity of Chadron (Trunk Butte is southwest of Chadron). Skinner's name for this unit was never published, but Terry and LaGarry (1998) applied the name *Big Cottonwood Creek Member* of the Chadron Formation to this body of rock.

Thus the apparent inconsistency in the definition of the Chadronian by Wood et al. (1941) really resulted from the uncritical definition of the top of the Chadron Formation by Schultz and Stout (1938, 1955). In the Toadstool Park area, the base of the Toadstool Park channel sequence is disconformable, and at some places the channel sequence has cut nearly to the level of the PWL, but as Terry and LaGarry (1998) clearly show, and Skinner had recognized long before, lateral to the channel sequence the Chadron Formation extends well above the PWL. The Chadronian–Orellan boundary as it was determined by Prothero and Whittlesey (1998) in the Dou-

glas, Wyoming, sequence and in the Seaman Hills sequence includes these latest known brontotherid occurrences.

Emry et al. (1987) reviewed the problems with the original Wood committee definition of the Chadronian and made some suggestions for an improved definition of this time interval. They recommended that the Chadronian be defined in faunal or biostratigraphic terms and argued that lithostratigraphic units had no role in the definition of a biochronologic unit. One of the major problems at that time was that there was much debate and confusion over how to distinguish the Chadronian from the preceding Duchesnean age and whether the Duchesnean could be distinguished at all. Some authors (Emry 1981; Wilson 1984, 1986) even recommended that the early Duchesnean be considered a subage of the Uintan and the late Duchesnean a subage of the Chadronian. But subsequent authors (Kelly 1990; Lucas 1992) pointed out the distinctiveness of the Duchesnean, and since then its biostratigraphy and chronostratigraphy have become much better understood (several chapters in Prothero and Emry 1996b).

Prothero and Emry (1996a) reviewed the latest biostratigraphic, magnetostratigraphic, and geochronologic data of the Chadronian and proposed a revised definition. The Chadron Formation in South Dakota and Nebraska proved to be too poorly fossiliferous and its biostratigraphy too poorly resolved to serve as the basis for the chronostratigraphy of the Chadronian. Instead, Prothero and Emry (1996a) recommended that the faunas of much better sections in Trans-Pecos Texas and at Flagstaff Rim and Douglas, Wyoming, serve as the chronostratigraphic standards for the Chadronian.

Prothero and Emry (1996a) designated the first appearance of *Bathygenys*, *Merycoiodon dunagani*, *Brachyrhynchocyon* (formerly *Daphoenocyon*) *dodgei*, and *Archaeotherium* as the best indicators of the beginning of the Chadronian, with *Bathygenys* as the defining taxon and the rest as characterizing taxa. In Trans-Pecos Texas, these distinctive earliest Chadronian taxa all occur in the Little Egypt Local Fauna, which lies immediately above the late Duchesnean Porvenir Local Fauna. Both faunas are bracketed by  $^{40}\text{Ar}/^{39}\text{Ar}$  dates of  $37.8 \pm 0.15$  Ma and  $36.7 \pm 0.07$  Ma, placing the Duchesnean–Chadronian boundary at approximately 37.0 Ma (and thus coincident with the middle–late Eocene boundary on the time scale of Berggren et al. 1995). If a stratotype were to be designated for this boundary, then the Trans-Pecos region of Texas is the only suitable place.

Because of the problem with the highly variable and infrequent occurrence of the latest surviving brontotheres with respect to the top of the Chadron Formation, the

original Wood committee (1941) definition of the end of the Chadronian is insufficient by itself, and although it can be part of the definition, it is impractical as a means of recognizing the end of Chadronian time. Prothero and Whittlesey (1998) instead recommended that the Chadronian–Orellan boundary be placed at the first appearance of the distinctive taxon *Hypertragulus calcareatus*, along with the first appearance of a number of additional reference taxa. This boundary is discussed further later in this chapter.

### SUBDIVISIONS OF THE CHADRONIAN

As reviewed by Emry et al. (1987), early concepts of the Chadronian essentially used lithostratigraphic units as de facto subdivisions of the time interval. Clark (1937, 1954) proposed a threefold division of the Chadron Formation in the Big Badlands of South Dakota into Ahearn, Crazy Johnson, and Peanut Peak members and suggested that these could also serve as time divisions of the Chadronian. Schultz and Stout (1955, 1961) designated informal members A, B, and C of the Chadron Formation in the Toadstool Park area of Nebraska. These rocks were also subdivided by the oreodont faunal zones of Schultz and Falkenbach (1968). The stratigraphic schemes of Schultz and Stout (1955, 1961) have not stood the test of time, with revised definitions (Terry 1998) placing Orella A in the Chadron Formation and changing the correlation with South Dakota radically from earlier concepts. Unfortunately, both the Clark (1954) and Schultz and Stout (1955, 1961) schemes used lithostratigraphic units for essentially biochronologic and biostratigraphic concepts, so they were fundamentally unsound. In addition, the actual stratigraphic range of the key fossils in these units was poorly documented, so they were unsuitable for a detailed range zone biostratigraphy. A much better, more complete section with excellent biostratigraphic data representing most of the Chadronian occurs at Flagstaff Rim, Wyoming (Emry 1973, 1992; Emry et al. 1987), and this serves as the basis for the much of the present zonation of the Chadronian.

Prothero and Emry (1996a) suggested a subdivision of the Chadronian into four biostratigraphically defined intervals, based on the best available lithostratigraphic sections known and calibrated by magnetostratigraphy and  $^{40}\text{Ar}/^{39}\text{Ar}$  dating. These four intervals were known informally as earliest, late early, middle, and late Chadronian. Each of these four intervals is about 1 million years in duration, except for the earliest Chadronian, which is only about 500,000 years in duration. Although possible biostratigraphic index taxa were suggested, no formal bio-

stratigraphic zones were erected in that publication. In this chapter, we will finish this procedure by formally designating these zones.

**Earliest Chadronian (36.5–37.0 Ma)** Prothero and Emry (1996a) suggested that the biochronologic interval represented by the Little Egypt Local Fauna of Texas and Rancho Gaitan Local Fauna in Chihuahua, Mexico, be considered earliest Chadronian. This interval might be represented by the very base of the section at Flagstaff Rim, Wyoming, but the faunal evidence is very limited. Based on the local magnetostratigraphy (Prothero 1996a), this interval falls entirely in magnetic chron C16r, so it apparently spans the time interval from 36.5 to 37.0 Ma. As discussed earlier, the first appearances of *Bathygenys* and several other index taxa characterize this interval, so if a formal biostratigraphic zonation were needed, it could be known as the *Bathygenys* Interval Zone, and its stratotype is here designated as the Reeves Bonebed section (Wilson 1978, figure 9; Prothero 1996a) in Trans-Pecos Texas.

**Late Early Chadronian (35.7–36.5 Ma)** This interval is much more completely represented and geographically widespread than the earliest Chadronian. The main reference section is at Flagstaff Rim, Wyoming (from just above the base of the section to 50 feet below Ash B), and the principal correlatives are the McCarty's Mountain Local Fauna of Montana, the faunas of the Ahearn Member in South Dakota, and the Yoder Local Fauna of Wyoming. Based on the magnetostratigraphy, this interval correlates with magnetic chron C15n2 (35.7–36.2 Ma). Ash B has been  $^{40}\text{Ar}/^{39}\text{Ar}$  dated at  $35.9 \pm 0.2$  Ma (Prothero and Swisher 1992) or  $35.41 \pm 0.14$  Ma (Obradovich et al. 1995), with the more recent Obradovich date taking into account some laboratory problems with Swisher's date and better fitting all the other age constraints. This places an upper age constraint on the late early Chadronian.

Prothero and Emry (1996a) gave a list of taxa that first appear in and are limited to this interval, including *Palaeolagus primus*, *Merycoidodon presidioensis*, *Pseudoproteroceras semicinctus*, *Litoyoderimys lustrorum*, *Yoderimys stewarti*, and *Leptomeryx yoderi*. In addition, a number of taxa (including *Daphoenictis tedfordi*, *Hyaenodon montanus*, *Ischyromys veterior*, *Centetodon chadronensis*, *Parvitrugulus priscus*, *Patriomanis americanus*, *Trigenicus profectus*, and *Sinclairiella dakotensis*) first occur at this time but range into the middle Chadronian. Prothero and Emry (1996a) suggested that this interval be named the *Leptomeryx yoderi* Interval Zone, with its type section in the main Flagstaff Rim section along Little Lone Tree Gulch (Emry 1973).

**Middle Chadronian (34.7–35.7 Ma)** The middle Chadronian is much better known than the two earlier Chadronian time intervals. It is typified by the large fossil collections recovered from the interval from 50 feet below Ash B to 50 feet above Ash G at Flagstaff Rim, Wyoming (Emry 1973, 1992). There are several principal correlatives, including the Pipestone Springs and Little Pipestone Creek local faunas of Montana (Tabrum et al. 1996), the Raben Ranch Local Fauna of Nebraska (Ostrander 1985), and the fauna of the Crazy Johnson Member of South Dakota. These beds all appear to correlate with magnetic chrons C15n–C15r (34.7–35.7 Ma). This interval has been dated not only by the  $^{40}\text{Ar}/^{39}\text{Ar}$  date on Ash B cited earlier but also by dates of  $35.7 \pm 0.1$  Ma (biotite) and  $35.8 \pm 0.04$  Ma (anorthoclase) on Ash F and  $35.6 \pm 0.06$  Ma (biotite) and  $35.7 \pm 0.03$  Ma (anorthoclase) on Ash G (Swisher and Prothero 1992).

Prothero and Emry (1996a) noted a number of taxa that are limited to this interval, including *Leptomeryx mammifer*, *Zemiodontomys burkei*, *Miniochoerus forsythae*, *Prosciurus vetustus*, *Hyaenodon microdon*, *Hyaenodon megalodon*, *Chadrolagus emryi*, and *Frictops emryi*. A number of taxa first appear in the middle Chadronian and range into younger strata, including *Palaeolagus temnodon*, *Dinictis felina*, *Meliakrounomys*, *Eutyromys magnus*, *Mesohippus westoni*, *Leptomeryx speciosus*, *Pseudoprotoceras longinarius*, large *Ischyromys*, *Hoplophoneus mentalis*, *Miohippus grandis*, and *Megalagus brachyodon*. Emry et al. (1987) and Prothero and Emry (1996a) recommended that *Leptomeryx mammifer* be designated the index fossil of this zone (which could be called the *Leptomeryx mammifer* Interval Zone), with a type section in the Flagstaff Rim section from 50 feet below Ash B to 50 feet above Ash G. We so designate it in this chapter.

**Late Chadronian (34.7–33.7 Ma)** The Flagstaff Rim section is sparsely fossiliferous from about 50 feet above Ash G, so Prothero and Emry (1996a) suggested the section below the PWL (= Ash J at Flagstaff Rim, the 4 tuff at Douglas, Wyoming, according to Larson and Evanoff 1998) in the Seaman Hills section, north of Lusk, Wyoming, better typifies this interval. Additional referred sections include the Douglas section below the 4 tuff (Evanoff et al. 1992), the late Chadronian in western Nebraska (Ostrander 1985; Terry 1998), the Peanut Peak Member in South Dakota, and possibly the Ash Spring Local Fauna in Texas. All of these strata appear to correlate with late chron C13r–C15n (33.7–34.7 Ma). Ash J at Flagstaff Rim has been  $^{40}\text{Ar}/^{39}\text{Ar}$  dated at  $34.7 \pm 0.04$  Ma (Prothero and Swisher 1992) or  $34.36 \pm 0.11$  Ma

(Obradovich et al. 1995). As discussed earlier, the Obradovich date probably is more reliable.

Only a few taxa, such as *Pseudoprotoceras taylori*, are limited to the late Chadronian at Flagstaff Rim (Emry 1992). In other late Chadronian sections, the interval is marked by the first occurrences of *Merycoiodon culbertsoni*, *Poebrotherium franki*, *Miniochoerus chadronensis*, *Ischyromys typus*, *Mesohippus exoletus*, *Mesohippus bairdi*, and *Scottimus viduus*. A number of taxa last appear in this interval and terminate near the Chadronian–Orellan boundary, including Brontotheriidae, *Xenocranium*, *Pseudocylindrodon*, *Toxotherium*, *Palaeolagus temnodon*, *Hoplophoneus mentalis*, *Miohippus grandis*, *Mesohippus westoni*, *Eotylopus reedi*, *Poebrotherium eximium*, and *Archaeotherium coarctatum*. Prothero and Emry (1996a) recommended that *Miniochoerus chadronensis* is the best candidate for index fossil in this interval because it is abundant, distinctive, and limited to the late Chadronian. Thus the late Chadronian could be designated as the *Miniochoerus chadronensis* Interval Zone and typified by the Seaman Hills section in Niobrara County, Wyoming. We formally designate the type section of the *Miniochoerus chadronensis* Interval Zone as the interval from 25 m below to 7 m above the PWL in the Boner Ranch section (SW sec. 9, T35N R61W, South Oat Creek 7.5' Quadrangle, Niobrara County, Wyoming; see Prothero and Whittlesey 1998, figure 5).

## ORELLAN

The Wood committee (1941:11) based the Orellan on “the Orella member of the Brule Formation, type locality, Orella, northwestern Nebraska, southwestern South Dakota and eastern Wyoming; includes the old term, ‘*Oreodon* beds,’ used in the most extended sense.” Once again, the Wood committee based a biochronologic concept on a lithostratigraphic unit, which is no longer acceptable in modern stratigraphic practice (Emry et al. 1987). In addition, Terry (1998) transferred Schultz and Stout’s (1955, 1961) “Orella A” to the Chadron Formation, so that the Chadron–Orella lithostratigraphic boundary now closely corresponds to the Chadronian–Orellan biochronologic and biostratigraphic boundary. Emry et al. (1987:139) pointed out that the “old term ‘*Oreodon* beds’ used in the broadest sense” applies to many rocks not considered Orellan. As discussed by Prothero and Whittlesey (1998), the Wood committee’s (1941) original criteria for recognizing the Chadronian–Orellan boundary (the last occurrence of brontotheres

and the top of the Chadron Formation) is also unsuitable, but with Terry's revision, this definition is no longer in conflict because at least four documented occurrences of brontotheres are now known above the Chadron–Brule contact of Schultz and Stout (1955, 1961) but within the Chadron Formation as redefined by Terry (1998).

For these reasons, Prothero and Whittlesey (1998) recommended redefining the Chadronian–Orellan boundary as the first appearance of *Hypertragulus calcaratus*, with the first appearances of *Leptomeryx evansi*, *Palaeolagus intermedius*, and small *Miniochoerus chadronensis* (= "*M. douglasensis*" of Stevens 1977) serving as characterizing biostratigraphic datums. The last appearance of *Poebrotherium eximium* and *Miohippus grandis* (with brontotheres being used only in a secondary role) can also be used to mark this boundary. The Chadronian–Orellan boundary falls in the upper part of magnetic chron C13r, 7 m (20 feet) above the 5 tuff at Douglas, Wyoming, and the same distance above the PWL in the Seaman Hills, near Lusk, Wyoming. Swisher and Prothero (1990) and Prothero and Swisher (1992) reported a  $^{40}\text{Ar}/^{39}\text{Ar}$  date of  $33.91 \pm 0.058$  Ma on the 5 tuff, and, based on the Berggren et al. (1995) time scale, the Chadronian–Orellan boundary falls very close to 33.7 Ma, or the Eocene–Oligocene boundary.

## SUBDIVISIONS OF THE ORELLAN

Emry et al. (1987) reviewed the earlier attempts to subdivide the Orellan, based on the lithostratigraphy of the Orella Member or on the "oreodont faunal zones" of Schultz and Falkenbach (1968), and showed that they were inadequate as then constituted. Korth (1989) proposed a biostratigraphy of the Orellan, again based on the Orella Member in Nebraska, but as Prothero and Whittlesey (1998) pointed out, there were several problems with this zonation. First of all, it was based on the University of Nebraska collections, most of which do not offer stratigraphic resolution of taxa in the alphabetically labeled units in the Orella Member, so the "biostratigraphic zones" of Korth (1989) are essentially equivalent to lithostratigraphic units. In addition, Prothero and Whittlesey (1998) found problems with several of the zonal indicator taxa proposed by Korth (1989). Finally, Korth's (1989) biostratigraphic zones lacked several key criteria (such as type sections) required by the North American Code of Stratigraphic Nomenclature (North American Commission on Stratigraphic Nomenclature 1983).

For these reasons, Prothero and Whittlesey (1998) proposed a division of the Orellan into four successive biostratigraphic zones, complete with defining and charac-

terizing taxa based on the fine-scale biostratigraphic data available from the Frick Collection at numerous localities and including all information required by the code, including type sections. Thus the Orellan is now based on formal biostratigraphic zones as required by the code and can now be considered a stage and age in the sense of the code rather than an informal biochronologic unit.

**Earliest Orellan *Hypertragulus calcaratus* Interval Zone (33.7–33.4 Ma)** As discussed by Prothero and Whittlesey (1998), the earliest Orellan is now marked by the first appearance of *Hypertragulus calcaratus* and characterized by the first appearances of *Leptomeryx evansi* and *Palaeolagus intermedius*. Its upper limit is marked by the first appearance of the indicator of the next zone, *Miniochoerus affinis*. The type section for this interval zone was designated as the strata from 7 to 17 m (20–50 feet) above the 5 tuff in the Douglas area (full details given in Prothero and Whittlesey 1998:55). This zone is also known from the Seaman Hills section near Lusk, Wyoming. It is correlated with the latest part of magnetic chron C13r and the earliest part of chron C13n (33.7–33.4 Ma).

**Late Early Orellan *Miniochoerus affinis* Interval Zone (33.4–33.1 Ma)** Prothero and Whittlesey (1998:55) recognized an interval defined by the first appearance of the oreodont *Miniochoerus affinis* and characterized by the first appearances of *Eumys elegans*, *Pelycomys brulanus*, *Adjidaumo minutus*, *Cedromus wardi*, and *Hoplophoneus occidentalis*. This end of this interval is also marked by the last appearance of *Ischyromys parvidens* and the first appearance of *Miniochoerus gracilis*. They designated the type section of the *Miniochoerus affinis* Interval Zone as the strata between 17 and 27 m (63–80 feet) above the PWL in the Boner Ranch section in the Seaman Hills, near Lusk, Wyoming. Principal referred sections include the interval between 17 and 25 m (50–75 feet) above the 5 tuff in Douglas, Wyoming (Evanoff et al. 1992), portions of Orella B in Toadstool Park, Nebraska, and the Lower Nodular Zone in the Big Badlands of South Dakota. These strata correlate with the later part of magnetic chron C13n (33.4–33.1 Ma).

**Early Late Orellan *Miniochoerus gracilis* Interval Zone (33.1–32.5 Ma)** Prothero and Whittlesey (1998:56) recognized a zone defined by the first appearance of the dwarfed oreodont *Miniochoerus gracilis* and characterized by the first appearance of *Mesohippus barbouri*, *Agnotocaster readingi*, *Paradjidaumo validus*, *Eutypomys thomsoni*, and *Eumys parvidens*. Its end is marked by the last



appearance of *Ischyromys parvidens* and the first appearance of the next zonal indicator, *Merycoiodon bullatus*. They designated the type section as the strata 27–50 m (80–150 feet) above the PWL at Boner Ranch in the Seaman Hills, near Lusk, Wyoming. Principal reference sections included Orella C in Toadstool Park, Nebraska, and the strata just above the Lower *Oreodon* Beds but below the Upper *Oreodon* Beds in the Big Badlands of South Dakota. This zone correlates with the latest part of chron C13n and the early part of chron C12r (33.1–32.5 Ma).

**Latest Orellan *Merycoiodon bullatus* Interval Zone (32.5–32.0 Ma)** The latest Orellan is marked by a large number of distinctive taxa. Prothero and Whittlesey (1998:56) recognized a biostratigraphic zone based on the first appearance of the large-bulla oreodont *Merycoiodon bullatus* and characterized by the first appearance of *Miniochoerus starkensis*, *Palaeolagus burkei*, *Prosciurus magnus*, *Ecclesimus tenuiceps*, *Tenudomys basilaris*, *Pelycomys placidus*, *Heliscomys vetus*, *Heliscomys mcgrewi*, *Wilsonemys planidens*, and *Campestralomys annectens*. A large number of characteristically Orellan taxa last appear in this interval, including *Hyaenodon crucians*, *Ischyromys typus*, *Paratylopus labiatus*, *Archaeotherium mortoni*, *Thinohyus lentus*, *Stibarus quadricuspis*, *Leptochoerus emilyae*, *Subhyracodon occidentalis*, “*Hesperocyon*” *coloradensis*, *Prosciurus*, *Pelycomys*, *Protosciurus*, *Oligospermophilus*, *Eutyromys*, *Adjidaumo*, *Paradjidaumo*, *Heliscomys*, *Wilsonemys*, *Eoemys*, *Tenudomys*, *Pipestoneomys*, *Megalagus*, *Palaeolagus intermedius*, *Centetodon marginalis*, *Leptictis haydeni*, *Herpetotherium fugax*, *Copedelphys stevensoni*, *Nanodelphys hunti*, and all surviving species of *Mesohippus* (*M. bairdi*, *M. exoletus*, *M. westoni*, and *M. barbouri*). This concentration of last appearances (especially of micromammals) may be partially an artifact of poor collecting and lack of screenwashing from strata of the early Whitneyan and may be modified by range extensions once adequate sampling is done. However, for most of the larger mammals, this is not the case, and these taxa almost certainly last occur in this zone.

Prothero and Whittlesey (1998) designated the type section of this zone as the Upper Nodular Zone on the east side of Sheep Mountain Table in the Big Badlands (full details given in Prothero and Whittlesey 1998:56). Principal reference sections include the upper part of Orella C and Orella D in the Toadstool Park area, Nebraska, and the upper part of the Cedar Creek Member in Colorado. These beds correlate with the early part of magnetic chron C12r (32.0–32.5 Ma).

## WHITNEYAN

Wood et al. (1941:11) based the Whitneyan “on the Whitney Member of the Brule Formation, type locality, Whitney, northwestern Nebraska, type area, northwestern Nebraska, southwestern South Dakota and eastern Wyoming, including the old term, ‘*Protoceras–Leptauchenia* beds,’ used in the most extended sense.” As Emry et al. (1987) pointed out, there were numerous problems with this definition, not the least of which is the fact that it bases a biochronologic interval of time on a lithostratigraphic unit. They recommended that the Whitneyan be defined on biostratigraphic criteria and suggested some possibilities for an appropriate set of criteria.

The multiple sections and high-resolution biostratigraphic data for the Orellan allowed Prothero and Whittlesey (1998) to finely subdivide it into four zones, each less than a million years in duration. Unfortunately, such data are not yet available for the Whitneyan. Most of the best biostratigraphic data come from the Poleslide Member of the Brule Formation in the Big Badlands of South Dakota, which was typically subdivided into the Upper *Oreodon*, *Protoceras*, and *Leptauchenia* beds of Osborn (1907), Osborn and Matthew (1909), Wanless (1923), Skinner (unpublished field notes), and many other workers. A smaller biostratigraphic database comes from the Whitney Member of the Brule Formation in northwestern Nebraska, which was subdivided into units A, B, and C by Schultz and Stout (1955, 1961). Because fossils usually were zoned only to one of the three subdivisions of the Whitneyan in these regions, the Whitneyan cannot be more finely subdivided than the original data resolution allows. In addition, Whitneyan fossils are much scarcer and more sparsely distributed than they are in the Orellan, further hampering fine-scale biostratigraphic zonation. Finally, the Whitneyan is known well only from two regions (Nebraska and South Dakota), with lesser collections from Colorado, North Dakota, and Wyoming, so many of the key index taxa only occur in one place.

## SUBDIVISIONS OF THE WHITNEYAN

Prothero and Whittlesey (1998) proposed a formal division of the Whitneyan into two biostratigraphic zones, based on the excellent biostratigraphic data now available from the Frick Collection. These zones include the following.

**Early Whitneyan *Leptauchenia major* Interval Zone (32.0–31.4 Ma)** A number of taxa are typical of the early Whitneyan, including abundant *Leptauchenia decora* (the

traditional indicator of this zone but known to occur rarely in older rocks) and the first occurrences of *Lep- tauchenia major*, *Hyracodon leidymanus*, *Paratylopus pri- maevus*, *Paralabis cedrensis*, *Diceratherium tridactylum*, *Protapirus obliquidens*, *Ectopocynus antiquus*, *Oxetocyon cuspidatus*, *Cynodesmus thoooides*, *Agnotocastor praeterea- dens*, and *Oropyctis pediasius*. Last occurrences in this zone include *Miniochoerus starkensis*, *Stibarus obtusilobus*, *Hyaenodon horridus*, *Cedromus wilsoni*, *Metadjidaumo hendryi*, *Agnotocastor praetereadens*, and *Oropyctis pedia- sius*. Prothero and Whittlesey (1998) designated a type section for this zone as the Upper *Oreodon* Beds (0–30 m, or 0–90 feet, above the Scenic-Poleslide contact) on the south side of Sheep Mountain Table in the Big Badlands of South Dakota (see Prothero and Whittlesey 1998:56 for details). Its principal reference section is Whitney A in the Toadstool Park area, Nebraska, and the lower part of the Vista Member in Colorado. This zone correlates with the middle of magnetic chron C12r (32.0–31.4 Ma). It is calibrated by the Lower Whitney Ash, which was  $^{40}\text{Ar}/^{39}\text{Ar}$  dated at  $31.8 \pm 0.023$  Ma (Swisher and Prothero 1990; Prothero and Swisher 1992).

**Late Whitneyan *Merycooidodon major* Interval Zone (31.4–30.0 Ma)** The traditional “*Protoceras* beds” and “*Lep- tauchenia* beds” of the Big Badlands yield an essentially homogeneous fauna, as does Whitney B–C in Nebraska. For that reason, Prothero and Whittlesey (1998) were not able to maintain this distinction based on lithostratigraphy rather than faunal change. Instead, they recognized a single late Whitneyan zone, defined by the first appearance of the distinctive large oreodont *Merycooidodon major* and characterized by the first appearances of *Protoceras celer* (the traditional zonal indicator but known only from the channel sandstones in the Big Badlands); the camel *Pseudolabis dakotensis*; the horses *Miohippus intermedius*, *Miohippus annectens*, *Miohippus equinanus*, and *Miohippus gidleyi*; and *Hoplo- phoneus dakotensis*, *Eusmilus cerebrealis*, *Nimravus brachy- ops*, *Hyaenodon brevirostrus*, *Eumys brachyodus*, and *Scot- timus lophatus*. Last occurrences include *Palaeolagus burkei*. Prothero and Whittlesey (1998) designated this interval as the *Merycooidodon major* Interval Zone, with its type section as the strata between 30 and 103 m (90–310 feet) above the Scenic-Poleslide contact on the south side of Sheep Mountain Table in the Big Badlands (see Prothero and Whittlesey 1998:57 for full details). This interval correlates with late chron C12r to early chron C11r (31.4–30.0 Ma). It is calibrated by the Upper Whitney Ash, which was  $^{40}\text{Ar}/^{39}\text{Ar}$  dated at  $30.58 \pm 0.61$  Ma, and by the overlying Nonpareil Ash in the Arikareean, which

was dated at  $30.05 \pm 0.19$  Ma (Swisher and Prothero 1990; Prothero and Swisher 1992).

## WHITNEYAN-ARIKAREEAN BOUNDARY

Tedford et al. (1996) reviewed the abundant new data on strata spanning the Whitneyan–Arikareean boundary, including the previously undescribed “brown siltstone member” of the White River Group above the Whitney Member in Nebraska. After reviewing all the available biostratigraphic data, they placed the end of the Whitneyan and beginning of the Arikareean at the first appearance of *Nanotragulus loomsi*, *Palaeolagus hypsodus*, *Palaeocastor nebrascensis*, *Leidymys blacki*, and *Mesore- odon minor*. These taxa first occur low in the Sharps For- mation in South Dakota (near the Rockyford Ash) and near the second Nonpareil Ash Zone of the “brown silt- stone” in Nebraska. This boundary occurs near the base of chron C11n (30.0 Ma). The end of the Whitneyan is also marked by the last occurrences of a number of taxa typical of the White River Chronofauna, including *Lep- tomeryx*, *Merycooidodon*, *Paratylopus*, *Paralabis*, *Per- choerus*, *Heptacodon*, *Leptochoerus*, *Colodon*, *Protapirus*, *Hesperocyon*, *Osbornodon*, *Dinictis*, *Paradjidaumo*, *Eumys*, and *Scottimus*.

## CONCLUSION

Although the fossils of the White River Group have been known for more than 150 years, only recently has it been possible to use these collections for classic range zone bio- stratigraphy. The enormous, stratigraphically zoned col- lections made by the Frick Laboratory and the strati- graphic research of Frick workers such as Morris Skinner finally made that achievement possible. The additional breakthroughs of magnetic stratigraphy and  $^{40}\text{Ar}/^{39}\text{Ar}$  dating allowed numerical dating, detailed correlation of individual sections, and calibration of these bio- stratigraphic zones to the global time scale. Consequently, some the great potential of the White River Group, with its excellent sections, abundant ashes, and enormous fos- sil collections, can now be realized.

We now have four biostratigraphically distinctive inter- vals in the Chadronian, four formally proposed range zones in the Orellan, and two formal zones in the Whitneyan. For the Orellan and Whitneyan, at least, this means that these land mammal ages are actually based on bio-

stratigraphic zones and stages, and so they are true stage/age units in the sense of the North American Code of Stratigraphic Nomenclature (North American Commission on Stratigraphic Nomenclature 1983). The Chadronian subdivisions have not yet been formally proposed, but Prothero and Emry (1996a) and this chapter have indicated the best candidates for the zonal index fossils and for type sections. Each of the four Chadronian, four Orellan, and two Whitneyan intervals are quite short, most less than a million years in duration, which offers the first high-resolution, well-calibrated chronostratigraphy for the White River Group. Such high-resolution data have already proven useful for a number of evolutionary and paleoclimatic studies (Prothero and Heaton 1996; Prothero 1999), where precise dating of faunal events is necessary.

## ACKNOWLEDGMENTS

We thank Mike Woodburne for inviting us to contribute this chapter and for his helpful comments and suggestions.

## REFERENCES

- Berggren, W. A., D. V. Kent, M.-P. Aubry, C. C. Swisher III, and K. G. Miller. 1995. A revised Paleogene geochronology and chronostratigraphy. *SEPM Special Publication* 54:129–212.
- Berggren, W. A., D. V. Kent, J. D. Obradovich, and C. C. Swisher III. 1992. Toward a revised Paleogene geochronology. In *Eocene–Oligocene climatic and biotic evolution*, ed. D. R. Prothero and W. A. Berggren. Princeton, NJ: Princeton University Press, pp. 29–45.
- Black, C. C. 1968. Late Oligocene vertebrates from the northeastern Wind River Basin. In *University of Colorado Museum, field conference guidebook for the high altitude and mountain basin deposits of Miocene age in Wyoming and Colorado*. Boulder: University of Colorado Museum, pp. 50–64.
- Clark, J. 1937. The stratigraphy and paleontology of the Chadron Formation in the Big Badlands of South Dakota. *Annals of the Carnegie Museum of Natural History* 25:261–350.
- . 1954. Geographic designation of the members of the Chadron Formation in South Dakota. *Annals of the Carnegie Museum of Natural History* 33:197–198.
- Emry, R. J. 1973. Stratigraphy and preliminary biostratigraphy of the Flagstaff Rim area, Natrona County, Wyoming. *Smithsonian Contributions to Paleobiology* 18.
- . 1981. Additions to the mammalian fauna of the type Duchesnean, with comments on the status of the Duchesnean. *Journal of Paleontology* 55:563–570.
- . 1992. Mammalian range zones in the Chadronian White River Formation at Flagstaff Rim, Wyoming. In *Eocene–Oligocene climatic and biotic evolution*, ed. D. R. Prothero and W. A. Berggren. Princeton, NJ: Princeton University Press, pp. 106–115.
- Emry, R. J., P. R. Bjork, and L. S. Russell. 1987. The Chadronian, Orellan, and Whitneyan land mammal ages. In *Cenozoic mammals of North America: Geochronology and biostratigraphy*, ed. M. O. Woodburne. Berkeley: University of California Press, pp. 118–152.
- Evanoff, E., D. R. Prothero, and R. H. Lander. 1992. Eocene–Oligocene climatic change in North America: The White River Formation near Douglas, east-central Wyoming. In *Eocene–Oligocene climatic and biotic evolution*, ed. D. R. Prothero and W. A. Berggren. Princeton, NJ: Princeton University Press, pp. 116–130.
- Evernden, J. F., D. E. Savage, G. H. Curtis, and G. T. James. 1964. Potassium–argon dates and the Cenozoic mammalian chronology of North America. *American Journal of Science* 262:145–198.
- Kelly, T. S. 1990. Biostratigraphy of Uintan and Duchesnean land mammal assemblages from the middle member of the Sespe Formation, Simi Valley, California. *Contributions to Science of the Natural History Museum of Los Angeles County* 419:1–42.
- Korth, W. W. 1989. Stratigraphic occurrence of rodents and lagomorphs in the Orella Member, Brule Formation (Oligocene), northwestern Nebraska. *Contributions to Geology, University of Wyoming* 27(1):15–20.
- LaGarry, H. E. 1998. Lithostratigraphic revision and redescription of the Brule Formation (White River Group) of northwestern Nebraska. *Geological Society of America Special Paper* 325:63–91.
- Larson, E. E. and E. Evanoff. 1998. Tephrostratigraphy and source of the tuffs of the White River sequence. *Geological Society of America Special Paper* 325:1–14.
- Lucas, S. G. 1992. Redefinition of the Duchesnean land mammal “age,” late Eocene of western North America. In *Eocene–Oligocene climatic and biotic evolution*, ed. D. R. Prothero and W. A. Berggren. Princeton, NJ: Princeton University Press, pp. 88–105.
- Manning, E. M. 1997. An early Oligocene rhinoceros jaw from the marine Byram Formation of Mississippi. *Mississippi Geology* 18:14–31.
- Manning, E. M., D. T. Dockery III, and J. A. Schiebout. 1986. Preliminary report of a Metamynodon skull from the Byram Formation (lower Oligocene) in Mississippi. *Mississippi Geology* 6:1–16.
- North American Commission on Stratigraphic Nomenclature. 1983. North American Code. *American Association of Petroleum Geologists Bulletin* 67:841–875.
- Obradovich, J. D., E. Evanoff, and E. E. Larson. 1995. Revised single-crystal laser-fusion  $^{40}\text{Ar}/^{39}\text{Ar}$  ages of Chadronian tuffs in the White River Formation of Wyoming. *Geological Society of America, Abstracts with Programs* 27(3):77–78.
- Osborn, H. F. 1907. Tertiary mammal horizons of North America. *Bulletin of the American Museum of Natural History* 23:237–254.
- Osborn, H. F. and W. D. Matthew. 1909. Cenozoic mammal horizons of western North America. *U.S. Geological Survey Bulletin* 361:1–138.
- Ostrander, G. E. 1985. Correlation of the early Oligocene (Chadronian) in northwestern Nebraska. In *Fossiliferous Cenozoic deposits of western South Dakota and northwestern Nebraska*, ed. J. E. Martin. *Dakoterra* 2:205–231.
- Patton, T. H. 1969. An Oligocene land vertebrate fauna from Florida. *Journal of Paleontology* 43:543–546.
- Prothero, D. R. 1994. *The Eocene–Oligocene transition: Paradise lost*. New York: Columbia University Press.
- . 1996a. Magnetostratigraphy of the Eocene–Oligocene transition in Trans-Pecos Texas. In *The terrestrial Eocene–Oligocene*

- transition in North America*, ed. D. R. Prothero and R. J. Emry. Cambridge, UK: Cambridge University Press, pp. 189–198.
- . 1996b. Magnetostratigraphy of the White River Group in the High Plains. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. Cambridge, UK: Cambridge University Press, pp. 247–262.
- . 1999. Does climatic change drive mammalian evolution? *GSA Today* 9(9):1–5.
- Prothero, D. R. and R. J. Emry. 1996a. Summary. In *The terrestrial Eocene–Oligocene transition in North America*, in D. R. Prothero and R. J. Emry. Cambridge, UK: Cambridge University Press, pp. 646–664.
- (eds.). 1996b. *The terrestrial Eocene–Oligocene transition in North America*. Cambridge, UK: Cambridge University Press.
- Prothero, D. R. and T. H. Heaton. 1996. Faunal stability during the early Oligocene climatic crash. *Palaeogeography, Palaeoclimatology, Palaeoecology* 127:239–256.
- Prothero, D. R. and C. C. Swisher III. 1992. Magnetostratigraphy and geochronology of the terrestrial Eocene–Oligocene transition in North America. In *Eocene–Oligocene climatic and biotic evolution*, ed. D. R. Prothero and W. A. Berggren. Princeton, NJ: Princeton University Press, pp. 46–74.
- Prothero, D. R. and K. E. Whittlesey. 1998. Magnetostratigraphy and biostratigraphy of the Orellan and Whitneyan land mammal “ages” in the White River Group. *Geological Society of America Special Paper* 325:39–61.
- Schultz, C. B. and C. H. Falkenbach. 1968. The phylogeny of the oreodonts, Parts 1 and 2. *Bulletin of the American Museum of Natural History* 139:1–498.
- Schultz, C. B. and T. M. Stout. 1938. Preliminary remarks on the Oligocene of Nebraska (abstract). *Geological Society of America Bulletin* 49:1921.
- . 1955. Classification of the Oligocene sediments in Nebraska. *Bulletin of the University of Nebraska State Museum* 4:17–52.
- . 1961. Field conference on the Tertiary and Pleistocene of western Nebraska. *Special Publication of the University of Nebraska State Museum* 2:1–54.
- Setoguchi, T. 1978. Paleontology and geology of the Badwater Creek area, central Wyoming. Part 16: The Cedar Ridge Local Fauna (late Oligocene). *Carnegie Museum of Natural History Bulletin* 9.
- Stevens, M. S. 1977. *Re-evaluation of taxonomy and phylogeny of some oreodonts (Artiodactyla)*. Unpublished manuscript in the Osborn Library of the American Museum of Natural History.
- Swinehart, J. B., V. L. Souders, H. M. DeGraw, and R. F. Diffendal. 1985. Cenozoic paleogeography of western Nebraska. In *Cenozoic paleogeography of the west-central United States*, ed. R. M. Flores and S. S. Kaplan. Denver: Rocky Mountain Section SEPM, pp. 209–229.
- Swisher, C. C. III and D. R. Prothero. 1990. Single-crystal  $^{40}\text{Ar}/^{39}\text{Ar}$  dating of the Eocene–Oligocene transition in North America. *Science* 249:760–762.
- Tabrum, A. R., D. R. Prothero, and D. Garcia. 1996. Magnetostratigraphy and biostratigraphy of the Eocene–Oligocene transition in western Montana. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 278–312.
- Tedford, R. H., T. Galusha, M. F. Skinner, B. E. Taylor, R. W. Fields, J. R. MacDonald, J. M. Rensberger, S. D. Webb, and D. P. Whistler. 1987. Faunal succession and biochronology of the Arikareean through Hemphillian interval (late Oligocene through earliest Pliocene epochs) in North America. In *Cenozoic mammals of North America: Geochronology and biostratigraphy*, ed. M. O. Woodburne. Berkeley: University of California Press, pp. 152–210.
- Tedford, R. H., J. Swinehart, C. C. Swisher III, D. R. Prothero, S. A. King, and T. E. Tierney. 1996. The Whitneyan–Arikareean transition in the High Plains. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 312–334.
- Terry, D. O. Jr. 1998. Lithostratigraphic revision and correlation of the lower part of the White River Group: South Dakota to Nebraska. *Geological Society of America Special Paper* 325:15–38.
- Terry, D. O. Jr. and H. E. LaGarry. 1998. The Big Cottonwood Creek member: A new member of the Chadron Formation in northwestern Nebraska. *Geological Society of America Special Paper* 325:117–141.
- Wanless, H. R. 1923. The stratigraphy of the White River beds of South Dakota. *Proceedings of the American Philosophical Society* 62:190–269.
- Wilson, J. A. 1978. Stratigraphic occurrence and correlation of early Tertiary vertebrate faunas, Trans-Pecos Texas, Part 1: Vieja area. *Texas Memorial Museum Bulletin* 25:1–42.
- Wilson, J. A. 1984. Vertebrate fossil faunas 49 to 36 million years ago and additions to the species of *Leptoreodon* found in Texas. *Journal of Vertebrate Paleontology* 4:199–207.
- . 1986. Stratigraphic occurrence and correlation of early Tertiary vertebrate faunas, Trans-Pecos Texas: Agua Fria–Green Valley areas. *Journal of Vertebrate Paleontology* 6:350–373.
- Wood, H. E. II, R. W. Chaney Jr., J. Clark, E. H. Colbert, G. L. Jepsen, J. B. Reeside, and C. Stock. 1941. Nomenclature and correlation of the North American continental Tertiary. *Geological Society of America Bulletin* 52:1–48.

# 6

## Mammalian Biochronology of the Arikareean Through Hemphillian Interval (Late Oligocene Through Early Pliocene Epochs)

Richard H. Tedford, L. Barry Albright III,  
Anthony D. Barnosky, Ismael Ferrusquia-  
Villafranca, Robert M. Hunt Jr., John E. Storer,  
Carl C. Swisher III, Michael R. Voorhies,  
S. David Webb, and David P. Whistler

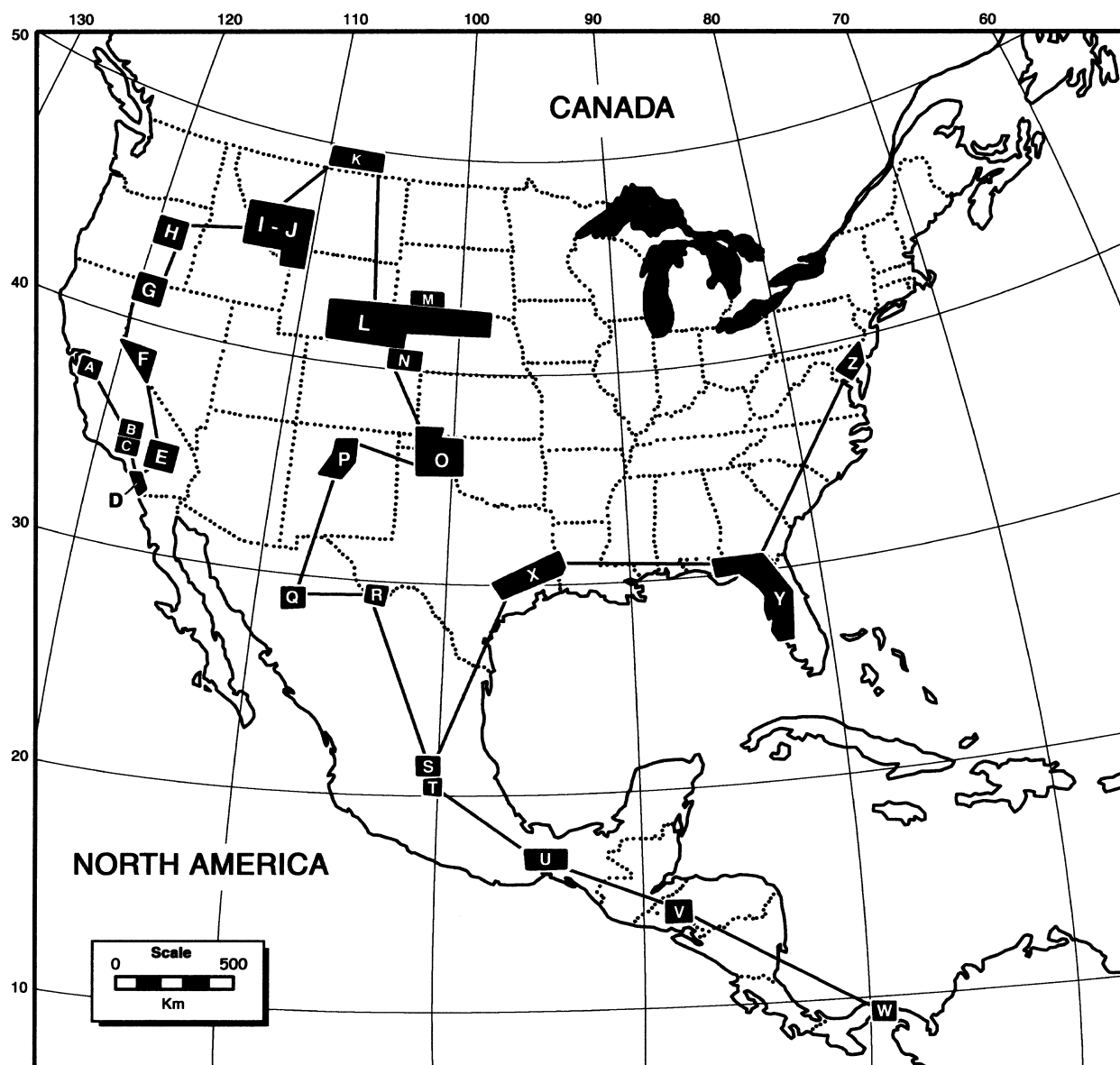
**I**N THIS CHAPTER, as in its predecessor (Tedford et al. 1987), we review the most important evidence bearing on the chronologic succession of the North American mammal faunas as revealed by stratigraphic superposition and biological correlation. The assembled record is calibrated by reference to the radioisotope ages of volcanic rocks interbedded with fossil mammal-bearing deposits (see “Appendix”) or by reference of magnetostratigraphies containing fossil mammals to the Geomagnetic Polarity Time Scale (GPTS). The wealth of new geochronologic data gives this treatment further rigor and guides the biochronology across zoogeographic discontinuities. This is easily seen in comparing the correlation charts in the 1987 work (figure 6.2) with that herein (figure 6.2). We have included nearly all the data shown in the previous correlation chart to present a comprehensive summary of knowledge of the temporal relationships of all assemblages discussed in Tedford et al. 1987 and in this revision. We follow the calibration of the GPTS given by Berggren et al. (1995) throughout this chapter.

The focus of the present work is to emphasize information gathered since 1987, particularly that important in assessing the relationship of faunal data to independent chronologic systems. Localities or districts where no new information has been obtained are not reconsidered here, nor is their significance called into question.

The geographic scope of this compilation has been extended to the southern Great Basin of New Mexico, Texas, and Mexico; the northern Great Plains of Saskatchewan, Canada; the Atlantic coast of the northeastern United States; and southern Mexico and Central America (figure 6.1). This extended geographic coverage will again

serve as a test of the application of the North American land mammal ages (NALMAs) to the limits of the evidence available from the middle and southern latitudes of the continent.

Over the past years there have been a number of reviews and restatements of the principles discussed in the 1987 work (Lindsay and Tedford 1990; Woodburne and Swisher 1995; Woodburne 1996). Walsh (1998, 2000) presents a more detailed analysis of biochronology in the context of paleobiological event terms in general. Woodburne (chapter 1, this volume) reviews a number of Walsh’s recommendations, but some are worth brief additional remarks as they impinge on the principles used in this chapter. Fundamental to our data analysis is the use of the local sample of paleontological materials derived from a single site or compiled from separate sites at a single stratigraphic level in a limited geographic area. These samples are called a local fauna (note that this term was given greater stratigraphic and, necessarily, geographic scope in Tedford 1970 than is recommended in Tedford et al. 1987 or here). As Walsh (2000:769) emphasizes, these are not biostratigraphic units (i.e., they are not bodies of rock that contain fossils) but the co-occurrences of specific taxa at a geologic instant in time. Local faunas may occur in stratigraphic relationship to each other, thus acquiring a relative temporal position, and such data are used to establish the biological content of temporal units derived from them. Thus local faunas are operational units (*sensu* Walsh 1998) in that they are empirical entities whose components are observed to co-exist in nature. “Faunas” are theoretical units of greater temporal and geographic scope. Their components are



**FIGURE 6.1** Map of part of North and Central America showing (lettered black quadrangles) geographic regions from which data for the correlation charts (figure 6.2) were derived. Lines linking these regions indicate the sequence of regions discussed in the text.

united on taxonomic (usually species-level) content rather than empirical occurrence. They resemble neontological “faunas” as they characterize zoogeographic regions but obviously differ from the former in that a substantial temporal span is necessarily implied. They have been confused with assemblage zones, but they are purely biological associations, not bodies of rock. Faunas can be composed of many local faunas that share the same taxonomy. Commonly in continental strata, fossil remains are scattered geographically and stratigraphically within a given lithostratigraphic unit. In cases in which these data have not been gathered biostratigraphically but there is an

overall taxonomic uniformity of the composite assemblage, it may be called the “fauna of the X-Formation, Member etc.,” or simply the “X-Fauna.” In this case the composite assemblage or fauna is assumed to represent a coeval association present throughout the time span implied by the rock unit from which it was obtained.

With the help of the timely supraspecific classification of mammals of McKenna and Bell (1997), we have continued to upgrade the taxonomy of the assemblages discussed in this volume. However, we do not review all the taxa mentioned in the 1987 work, so this version will not suffice as a completely up-to-date taxonomy. We con-

tinue the practice of denoting original taxonomy, which we still question by placing quotation marks around the original usage.

In making this compilation we have often referred to the monumental compilation of various authors under the editorship of Janis, Scott, and Jacobs (1998) on matters of taxonomy, phylogeny, and geologic and geographic ranges, but we are not altogether constrained by this source. Likewise we have turned to Korth's (1994) review of the record of rodents in the Tertiary of North America. Thanks to the generosity of M. S. Stevens, we have had the benefit of her comprehensive review of the oreodonts, most of which remains unpublished (see Stevens and Stevens 1996 for the methods used in that work). Our oreodont references are drawn from her work. Ongoing revision of the Canidae (Wang 1994; Wang et al. 1999) have resulted in many nomenclatural changes cited here.

## THE FAUNAL SUCCESSION

### CALIFORNIA COAST RANGES

**San Francisco Bay Area (A)** A significant addition to our knowledge of the chronology of this region was provided by Baskin's (1981) discussion of the Black Hawk Ranch Local Fauna in the course of his contribution to the study of the temporally related fauna of the Love Bone Bed in Florida. This work (missed in the compilation of Tedford et al. 1987) provides a revision of the Carnivora of the California site, especially the recognition of the biochronologically important *Barbourofelis lovei* ("machairoidont" of Macdonald 1948). Wang et al. (1999) confirmed the presence of *Aelurodon taxoides* ("*Aelurodon aphobus*," Macdonald 1948) and attributed Macdonald's (1948) "*Osteoborus diabloensis*" to *Borophagus littoralis*. Baskin (1981) pointed out the close similarity of *Nimravides thinobates* to *Nimravides galiani* from the Love Bone Bed. Kelly's (1998) description of the horses strongly suggests that the Black Hawk *Pliohippus leardi* does belong to that genus, although he continued to refer this taxon to *Dinohippus*. Savage (1955) based most of the biological typification of his Montediablan Stage on the Black Hawk Ranch Local Fauna. In an effort to test the correlation of the local faunas used to characterize this stage and those held typical of the underlying Cerrotejonian Stage, Prothero (in Prothero and Tedford 2000) ran a long magnetostratigraphic section from the upper part of the shallow marine San Pablo Group at the level of the

ostensibly Cerrotejonian Sycamore Creek Local Fauna through the Black Hawk Ranch Quarry and upward for 350 m of the enclosing continental Sycamore Canyon Formation (Green Valley Formation in Tedford et al. 1987). The entire sequence was reversely magnetized and correlated with part of chron C4A (8–9 Ma).

**Southern San Joaquin Valley (B)** The composite suite of taxa present at the Tecuya Formation sites are best regarded as representing an early Arikareean fauna, limited by the presence of *Nanotragulus* and *Desmatochoerus-megalodon* (= *Promerycochoerus erythroceps* Stock 1932 *vide* M. S. Stevens, pers. comm., 2002) and the limit of the range of the White River relict *Protosciurus*. This limits the temporal range of these sites shown in Tedford et al. (1987:156 and figure 6.2).

In 1987 Tedford et al. (p. 156) mentioned in passing the terrestrial mammal faunas from the predominantly marine Temblor Formation outcrops on the western and eastern margins of the southern San Joaquin Valley. These assemblages can be directly related to marine megafaunal and microfaunal biostratigraphies, which in turn have been calibrated radiometrically. The most important assemblage is the North Coalinga Local Fauna ("The fauna of the *Merychippus* zone," Bode 1935a, 1935b), which lies near the top of the correlated Temblor strata in deposits that represent either the upper part of the Relizian benthonic foraminiferal stage or the boundary between the Relizian and superadjacent Luisian stages (Durham et al. 1954). In 1970 Turner calibrated the boundary between these stages as 13.7–14.5 Ma but admitted that large analytic uncertainties and tenuous correlations to the benthonic time scale limit the accuracy of this determination. These results were referred to by Tedford et al. (1987) in determining the age of the North Coalinga Local Fauna. The presence of Proboscidea in that fauna was said to correlate the assemblage with the Barstow Fauna despite previous workers' (Downs 1956, 1961; Bode 1935a) placement of this fauna in the proximity of the Mascall and Virgin Valley faunas of the Columbia Plateau and definitely older than the "Barstow Fauna" (having the same connotation to those authors as in this volume, i.e., the youngest faunal unit in the Barstow Formation). A review of the North Coalinga Local Fauna confirms their opinion: Stirton (1935) referred the *Monosaulax* sp. of Bode (1935a) to *M. pansus* in the sense of his sample from Stewart Springs, Nevada, which has in turn been allocated to *M. skinneri* Evander (1999) by Korth (1999c). The North Coalinga borophagines include *Paracynarctus kelloggi*, *Protomarctus* cf. *optatus*, and *Aelurodon asthenostylus*; the amphicyonid is *Amphicyon sinapius*; the horses

include *Hypohippus* cf. *osborni*, *Archaeohippus mourningi*, *Desmatippus avus*, *Merychippus brevidontus*, and “*Merychippus*” *californicus*; a rhino near *Aphelops* is present; the camels include *Aepycamelus* and *Miolabis*; and the cranioceratine dromomerycid seems to be *Bouromeryx*. The composition of this fauna has a Columbia Plateau–like character and also includes taxa found in the early Barstovian of the Great Plains. There are some range extensions (higher for *Prototomarcus*, lower for *Aelurodon*) for an early Barstovian assemblage. This conclusion is now confirmed by the latest calibration of the Relizian–Luisian boundary at 15.7 Ma by Barron and Isaacs (2001). This occurrence lends additional support for the early Barstovian appearance of Proboscidea, especially mammutids (*Zygodolophodon*, “*Miomastodon* sp.,” of Bode 1935a, figure 5a, c, and d), and possibly gomphotheriids as well (Bode 1935a, figure 5b) and extends the geographic range to the Pacific coastal sites in California. Curiously, the North Coalinga Local Fauna shares few taxa with early Barstovian assemblages of the nearby Transverse Ranges and Mojave Desert reviewed in this chapter.

Along the southeastern outcrop of the Temblor Formation, faunas of like composition include the late Hemingfordian Barker’s Ranch Local Fauna and early Barstovian Sharktooth Hill Local Fauna (also with *Zygodolophodon*) mentioned in Tedford et al. (1987:156 and footnotes 3 and 4), the latter attributed to the late Barstovian.

Savage (1955) typified his Cerrotejonian Stage with the South Tejon Hills Fauna (composite of Los Angeles County Museum [LACM] California Institute of Technology [CIT] sites 303, 304, and 307) from the Tejon Hills outcrops in the southeastern margin of the San Joaquin Valley. At the time the containing strata were referred to the marine Santa Margarita Formation, but later Bartow and McDougall (1984) revised the lithostratigraphy of the Tejon Hills in light of the regional relationships of the rock units in the southeastern San Joaquin Valley. They now attribute these mammal-bearing strata to the non-marine Bena Formation, which underlies a thin representative of the Santa Margarita with local angular unconformity. The South Tejon Hills Fauna includes *Hipparion tehonense*, “*Pliohippus*” *tejonensis*, *Megahippus* sp., *Borophagus littoralis* (Wang et al. 1999), *Ustatochoerus* sp., *Cranioceras* sp., and *Paracosoryx* cf. *furlongi*.

In the northern part of the hills, the thin Santa Margarita equivalent contains the Comanche Point Local Fauna with *Cormohipparion* cf. *occidentale* and “*Pliohippus*” *tehonensis*. These shallow water marine beds grade laterally and upward into the nonmarine Chanac Formation. This younger interval contains the North Tejon Hills

Fauna (LACM [CIT] localities 104, 302, and 305): *Hipparion forcei*, *Pliohippus leardi*, *Barbourofelis* cf. *lovei*, *Borophagus littoralis*, *Epicyon*, *Nimravides* cf. *thinobates*, *Ustatochoerus* cf. *californicus*, *Cranioceras* sp., and *Plioceros* sp. This fauna closely resembles that from the Black Hawk Ranch Quarry and was Savage’s (1955) principal reference assemblage for his Montediablan Stage. Thus the Tejon Hills contained not only the typifying fauna for the Cerrotejonian but also the superposed Montediablan reference. In an effort to calibrate this sequence, Wilson and Prothero (1997) constructed a nearly 200-m magnetostratigraphic section in the northern Tejon Hills extending from below the level of the Comanche Point Local Fauna through the exposed Chanac Formation. This sequence begins in a mixed-polarity interval containing the Comanche Point Local Fauna overlain by a long reversed section containing the North Tejon Hills Fauna. Unfortunately a magnetostratigraphy could not be constructed in the South Tejon Hills, but like Savage, Wilson and Prothero (1997) assumed that the northern hills section included correlative strata (“Santa Margarita”) at the base. These results were interpreted to indicate that the Cerrotejonian and Montediablan stages pertained to successive magnetic chrons assumed to be chron C5An to C5r (11–12 Ma). Later work (Prothero and Tedford 2000), informed by studies in the Cuyama Valley, shifted the correlation of the reversed interval to chron C4Ar.

**Transverse Ranges (C)** Continued study of the faunal sequence in the Transverse Ranges south of the San Joaquin Valley has improved our knowledge of the taxa that occur there and the chronologic relationships of the fossil assemblages.

Recent work on the lithostratigraphy, magnetostratigraphy, and radioisotopic age determination of faunal occurrences in the upper part of the Sespe Formation (Mason and Swisher 1989; Prothero et al. 1996) has confirmed the relationships of these scattered assemblages. Prothero et al. (1996) constructed a composite Sespe section on the north side of Simi Valley, eastern Ventura County, including the Alamos Canyon Local Fauna in the upper part of a 370-m segment of the upper member of the Sespe Formation above the intraformational hiatus separating the medial Eocene from late Oligocene rocks. The Alamos Canyon Local Fauna (R. W. Wilson 1949; Lander 1983) lies within a reversed interval that Prothero et al. (1996) correlate with chron C9r (27.9–28.3 Ma). It contains ?*Archaeolagus*, *Leidymys nematodon*, and *Hypertragulus* cf. *hesperus*, all taxa that occur in the John Day Formation in the interval between the Deep Creek and



Tin Roof tuffs (27.9–25.9 Ma). This suggests either a somewhat younger correlation with the GPTS than indicated by Prothero et al. (1996) or an upward extension of the local range zones for these taxa.

The Kew Quarry Local Fauna has been a consistent enigma as to its age, largely because of inadequate identification of constituent taxa and knowledge of their biochrons. Revision of some of the Kew Quarry canid, nimravid, and camelid taxa (in Prothero and Emry 1996) yields a revised mammal faunal list: *Palaeolagus*, *Sespeymys thurstoni*, *Paciculus* sp., *Temnocyon altigenis*, *Mesocyon brachyops* (Wang 1994), *Archaeocyon pavidus* (Wang et al. 1999), *Nimravus brachyops* (Bryant 1996), *Eusmilus cerebralis* (Bryant 1996), *Miohippus*, *Subhyracodon occidentalis*, *Elomeryx armatus*, *Miotylopus gibbi* (Prothero 1996), and *Hypertragulus calcaratus*. Judging from the ranges of these taxa in the older part of the John Day Formation and younger part of the Brule Formation, this assemblage is of early Arikareean age, ca. 28–29 Ma, and shows the characteristic persistence of elements of the White River Chronofauna coupled with new elements (*Mesocyon*, *Miotylopus*, possibly *Paciculus* if truly present, and *Temnocyon*; Hunt 1998). A 500-m section to determine the polarity of the structurally isolated Kew Quarry Local Fauna was found to be reversed except for the lowest site. The reversed strata in which it occurs probably lies in chron C10r.

In the folded Sespe outcrops at South Mountain, north of the Simi Valley, elements of the South Mountain Fauna occur through 640 m of section (Mason and Swisher 1989). The tiny leptachenine *Sespia* occurs throughout the entire sequence, with isolated occurrences of other taxa: *Eporeodon thurstoni* Stock (1934; *Eporeodon thurstoni* of Stevens, pers. comm., 2002) in the lower part of the section and *Promerycochoerus hesperus* Stock (1930; *Desmatochoerus megalodon* of Stevens, pers. comm., 2002), *Diceratherium*, and *Gregorymys* (or *Grangerimus*) near the top. The composition of this fauna resembles those of the volcanoclastic aeolian deposits of the lower Arikaree Group of the Great Plains. In California they occur in an interval dominated by variegated sandstones and red mudstones, carrying the same implications of well-oxidized sediments above groundwater. A datable ash bed (Willard Canyon Tuff) occurs near the top of the fossiliferous interval and gives a K–Ar date of  $28.20 \pm 0.2$  Ma. It lies in a normal polarity interval (Prothero et al. 1996) that appears to be chron C10n. The stratal range of the South Mountain Fauna probably encompasses most of chrons C10 and C9r, approximately 29.4–27.9 Ma. The fauna thus is partly an ecofacies of the Kew Quarry Local Fauna and partly succeeds it.

A significant faunal occurrence, the Vedder Local Fauna, in a nonmarine tongue in the otherwise marine Branch Canyon Formation of the eastern Caliente Range was briefly mentioned by Tedford et al. (1987:158). This late Hemingfordian assemblage provides a larger view than other West Coast sites of the diversity of mammals during that span. The local fauna was described by Hutchinson and Lindsay (1974), Lindsay (1974), and Munthe (1979a) and is particularly notable in containing the only North American occurrence of the Eurasian petauristine squirrel *Blackia*. The microfauna has broad resemblances to late Hemingfordian assemblages of the Great Plains and late Hemingfordian to early Barstovian faunas elsewhere in North America in the occurrence of *Hypolagus*, *Miospermophilus*, *Pseudotheridomys*, *Lepidotomys* (= “*Eomys*,” Korth and Bailey 1992), *Mookomys*, *Proheteromys sulcatus*, and *Proheteromys magnus*. The ochotonid *Cuyamalagus* is present, as are such mammals as *Protomarctus* cf. *optatus*, *Archaeohippus* cf. *penultimus*, *Parapliohippus carrizoensis*, *Ticholeptus zygomatus*, *Protolabis*, and a dromomerycid. Many of these taxa are also present at CIT loc. 315 (Dougherty 1940) and the Hidden Treasure Springs Site in the Caliente Formation to the east and southeast into the Cuyama Badlands. Like CIT loc. 315, the Vedder site lies below the projection of the lowest Triple Basalt K–Ar dated at  $16.5 \pm 1.3$  Ma (Turner 1970).

In recent years Kelly (Kelly and Lander 1988) has reevaluated the biostratigraphy of the Caliente Formation in the Cuyama badlands first monographed by James (1963). Kelly reviewed the horses (1995, 1998) and camels (1992), and Lander (Kelly and Lander 1988) reevaluated the oreodonts. The biochronology was later reaffirmed (Kelly and Lander 1992). A redating of the upper biotite tuff, or “Dated Tuff,” in the section by Swisher (1992) using  $^{40}\text{Ar}/^{39}\text{Ar}$  has shown it to be markedly younger ( $13.4 \pm 0.14$  Ma) than previously believed (15.6 Ma). This important sequence is revised here: The lowest assemblage, the Hidden Treasure Spring Fauna, lies in the lower part of the Caliente Formation as exposed locally. It contains a limited number of large mammal taxa, including “*Parahippus*,” “*Parapliohippus*,” *Acritohippus* cf. *tertius*, a chalicotherid, *Ticholeptus zygomatus*, *Aepycamelus*, and “*Merycodus*.” These taxa indicate correlation with the Vedder site to the northwest in the Caliente Range, as suggested earlier. A newly available magnetostratigraphy that includes the Hidden Treasure Springs level places this faunal association in the earlier part of chron C5Cn or in C5Cr, roughly equivalent to the position of the Red Division Quarry Local Fauna of the Barstow Formation. These sites share *Parapliohippus carrizoensis* but no other taxa.

Higher in the section, the West Dry Canyon and overlying Lower Dome Spring faunas (*sensu* Kelly and Lander 1988) are biologically correlated with the Green Hills Fauna of the Barstow Formation and contain *Parapliohippus*, *Acritohippus*, *Cynorca occidentale*, *Brachycrus*, *Miolabis fricki*, *Aepycamelus*, and *Rakomeryx raki* among the taxa in common. In addition, these faunas contain the cranio-ceratin dromomerycid *Bouromeryx cf. milleri*.

A larger assemblage is represented by the Upper Dome Spring Fauna, whose elements occur above the Lower Dome Spring Fauna and extend nearly to the base of the "Dated Tuff." This fauna includes the lowest local occurrences of *Copemys cf. dentalis*, *Perognathus furlongi*, *Proheteromys maximus*, and *Petauristodon uphami*. *Perognathus* and *Proheteromys* overlap only in the early Barstovian. The presence of the immigrant *Lanthanותרium* may be its earliest North American occurrence. Species of this genus have their first local occurrence in the younger part of the span of the Barstow Fauna. The large mammal fauna, which includes *Tomarctus hippophaga*, *Acritohippus quinni*, *Merychippus brevidontus*, *Archaeohippus mourningi*, *Cynorca occidentale*, *Miolabis fricki*, *Paramiolabis taylori*, *Hesperocamelus cf. alexandrae*, and *Bouromeryx milleri*, shows a close resemblance to the Lower Dome Spring Fauna except that it lacks the limited early Barstovian *Brachycrus* and *Rakomeryx* and adds *Paramiolabis* and *Hesperocamelus*. *Tomarctus hippophaga* seems limited to the early Barstovian (Wang et al. 1999). This and the rodent evidence suggest a late early Barstovian assemblage rather like that of the Second Division Fauna of the Barstow Formation.

The faunal evidence suggests that despite the close proximity of the "Dated Tuff" to the highest stratigraphic occurrence of the Upper Dome Spring Fauna, there must be a hiatus greater than 1 m.y. separating the fauna and ash.

Only scattered fossils occur above the Dome Spring faunas. The Doe Spring Canyon Fauna of Quatal Canyon in the western part of the Cuyama badlands lies above the "Dated Tuff." It contains *Gomphotherium*, *Acritohippus quinni* (highest local occurrence), *Merycodus cf. cerroensis*, *Ramoceros*, and *Petauristodon uphami*.

Although the sites containing the Doe Spring Canyon Fauna lack direct superposition with the interval in the Caliente Formation containing the Mathews Ranch and Nettle Spring faunas to the east in Apache Canyon, there seems little doubt of its relative stratigraphic position (Kelly and Lander 1988). These younger faunas contrast in their horse taxa but are largely conspecific with regard to their rodent assemblages. The Mathews Ranch faunal span lies mostly in a mixed-polarity interval correlative with chron C5An and the older part of C5r, whereas the immediately

overlying Nettle Spring Fauna lies in the younger part of chron C5r (Prothero and Tedford 2000). Savage (1955) regarded these faunas as part of the reference sequence for the Cerrotejonian and Montediablan stages, but in fact the supposed Montediablan Nettle Spring Fauna lacks the horse and other taxa characteristic of that stage. Nevertheless, the faunal correlation of Mathews Ranch and South Tejon Hills faunas seems valid, and the magnetics thus suggest a chron C5An–C5r age for the Cerrotejonian, or about 11.5–12.5 Ma. Thus in the Cuyama badlands, the superposed faunal sequence in the Caliente Formation suggests that older Clarendonian faunas extend to 12.4 Ma. The meager evidence hints at a faunal turnover in this coastal site that approximates the striking change in the Mojave Desert at the end of the Barstovian.

**Peninsular Ranges (D)** Important new information is now available regarding Arikareean faunas from the western flanks of the Peninsular Ranges in southern Los Angeles, Orange, and San Diego counties of southwestern California.

The large collection from the Otay Formation, south of San Diego (Démere 1988), the East Lake Local Fauna, closely resembles early Arikareean faunas of western Nebraska and adjacent Wyoming. Notable genera in common are *Leidyomys*, *Mesocyon*, *Hyaenodon*, *Subhyracodon*, *Mesoreodon*, *Sespia*, *Miotylopus*, ?*Dyseotylopus*, and *Nanotragulus*. The fossiliferous beds lie in a reversed interval correlated with chron C10r (incorrectly correlated in the "emended" figure 3 of Prothero 1991), as do coeval faunas of similar character in the Great Plains. Except for the occurrence of *Meniscomys* and *Mesocyon coryphaeus*, which are limited to rocks of comparable age in the John Day Formation of Oregon, the Otay Fauna closely resembles those of the earliest Arikareean of northern Great Plains, indicating very broad zoogeographic regions within North America in the late Oligocene, as already suggested by the Sespe Formation assemblages of the Transverse Ranges.

Newly discovered local faunas in the undifferentiated Sespe–Vaqueros formation strata of the Santa Ana Mountains (northern Peninsular Ranges) of Orange County include the Upper Oso Dam and Bee Canyon Landfill faunas (Raschke 1984), the Bolero Lookout Local Fauna (Lucas et al. 1997), and sites encountered during development of the Eastern Transportation Corridor (Prothero and Donohoo 2001). Many of these sites were discovered during recent road and housing development. They indicate the presence of early Hemingfordian assemblages resembling those of the Vaqueros and basal Caliente formations in the Transverse Ranges ("A" sites

in Repenning and Vedder 1961) and more widely into the midcontinent.

One of the most diverse of such assemblages is that from the Santiago Canyon Landfill, which contains such widely distributed early Hemingfordian taxa as *Trogomys*, *Proheteromys*, *Schizodontomys*, *Cupidinimus*, *Pseudotheridomys*, *Leidymys*, *Yatkolamys*, *Metatomarctus*, *Menoceras barbouri*, *Parahippus pawniensis*, *Anchitherium clarencei*, *Merychyus arenarum*, *Pseudoblastomeryx advena*, *Machaeromeryx*, *Michenia agatense*, and *Tanymycter brevidontus* (Lander 1994). The correlative Bolero Lookout Local fauna adds *Dinohyus hollandi* and contains *Parahippus pawniensis* and *Michenia agatense* (Lucas et al. 1997). Magnetostratigraphic work by Prothero and Donohoo (2001) indicates that the Sespe red beds containing such faunas lie below marine sandstones with Vaquerosian mollusks and reversed polarity in an interval of mixed polarity that can be correlated with chron C5Dr (18.3–17.6 Ma), slightly younger than the calibration of the lower part of the Runningwater Formation of Nebraska, which begins in chron C5Er (at 18.8 Ma) and extends to 18.0 Ma in chron C5Dr (MacFadden and Hunt 1998).

A thick sedimentary wedge on the northeastern flank of the Peninsular Ranges outlined by the San Andreas (north) and San Jacinto (south) faults includes the late Hemphillian Mount Eden Formation and Local Fauna near the base and succeeding Blancan assemblages in the conformably overlying San Timoteo Formation (Frick 1921; Albright 1999a). Frick (1921) described the Mount Eden Local Fauna, later adding an updated faunal list in his study of the Proboscideans (Frick 1933). In 1937 he added the presence of the dromomerycid *Pediomeryx* (mistaken for a cervid and named *Procoelius*; Frick 1937). A few other taxa were added by later authors: *Repomys* (May and Repenning 1982) and *Megacamelus* (Harrison 1985). Biochronologically important taxa include megalonychid sloths, the cricetine *Repomys maxumi*, the ursid *Agriotherium gregoryi*, the horse *Dinohippus osborni*, the rhino *Teleoceras*, the camel *Megacamelus*, and the dromomerycid *Pediomeryx*. The magnetostratigraphy of Albright (1999a) supports a correlation with chron C3r and an interpolated age of 5.6 Ma for the Mount Eden Local Fauna, comparable to the latest Hemphillian sites in the Panhandle of Texas and adjacent Oklahoma.

## NORTHERN GREAT BASIN

**Mojave Desert, California (E)** Major advances have been made in the knowledge of the chronology of the faunal sequence in the Barstow Formation of the Mud Hills, central Mojave Desert (MacFadden et al. 1990;

Woodburne et al. 1990), and the Ricardo faunal sequence of the Dove Spring Formation of the Ricardo Group on the northwestern edge of the desert (Whistler and Burbank 1992). In addition, other early Hemingfordian faunas have been discovered in the eastern part of the province (Reynolds et al. 1995) that help to define, and partially calibrate, such assemblages in the southwestern United States.

Woodburne et al. (1990) have provided a more detailed faunal sequence in the type area of the Barstow Formation (also principal reference sequence for the Barstovian land mammal age) than presented in Tedford et al. (1987). The oldest local fauna was obtained at a single site, Red Division Quarry, which contains only *Parapliohippus carizoensis*, *Paramiolabis tenuis*, and *Merychyus relictus fletcheri* (included in *Merychyus elegans* by Lander in Kelly and Lander 1988), clearly equating it with the Upper Cady Mountains Local Fauna farther east in the Mojave (see also Woodburne 1998:208) and the Phillips Ranch Local Fauna of the Tehachapi Mountains on the western desert margin. The limited occurrence of *P. tenuis* in the Sheep Creek Formation of western Nebraska indicates correlation with the principal reference fauna of the Hemingfordian land mammal age. At Barstow the local fauna from the Red Division Quarry lies in chron C5Cr (MacFadden et al. 1990) at about 16.7 Ma (Woodburne 1998).

The fauna of the Rak Division of the Barstow sequence lies stratigraphically between the late Hemingfordian Red Division Quarry and the diverse Green Hills Fauna of early Barstovian age. Chronologically the Rak Division encompasses chron C5Cn3–C5Cn1 (MacFadden et al. 1990; Woodburne, pers. comm., 1999). The Rak Division contains *Protomarctus optatus* (Wang et al. 1999), *Amphicyon* cf. *sinapius*, *Acritohippus* cf. *tertius*, *Aphelops*, *Paramiolabis tenuis*, *P. cf. singularis*, *Protolabis*, *Michenia*, *Aepycamelus*, and *Meryceros*. The top of this interval also contains the lowest observed occurrence of *Copemys* in the Barstow Formation (Lindsay 1995; at the base of chron C5Cn1, 16.2 Ma; Woodburne pers. comm., 1999). It was used by Tedford et al. (1987) as a defining taxon, along with *Pliothocyon*, for the beginning of Barstovian time (see discussion in Lindsay 1995). The Rak Division assemblage contains a mixture of taxa found only in the late Hemingfordian Sheep Creek Fauna or early Barstovian Lower Snake Creek Fauna of the classic sequence of western Nebraska, that is, *Protomarctus optatus*, *Acritohippus* cf. *tertius*, and *Paramiolabis tenuis* of the Sheep Creek coupled with *Amphicyon* cf. *sinapius* and *Paramiolabis* cf. *singularis* of the Lower Snake Creek, although we cannot demonstrate that the ranges of all these taxa are strictly coeval in the Rak Division.

The Green Hills Fauna is based on material from many local concentrations of remains and scattered finds. It maintains taxic uniformity through chron C5Br (15.3–16.0 Ma), and its containing strata are conformable with the Rak Division deposits below and with those containing the younger Barstow assemblages above. The Green Hills Fauna included the oldest known occurrence of *Hemicyon* (*Plithocyon*) in North America (at Steepside Quarry at the base of the stratal span containing the fauna) at approximately 16.0 Ma. The remaining fauna contains genera and species limited to the early Barstovian: *Tomarctus hippophaga*, *Amphicyon ingens*, *Acritohippus stylodontus*, *Brachycrus buwaldi*, *Rakomeryx*, and *Merriamoceros*. *Microtomarctus conferta* (Wang et al. 1999) first appears in the Green Hills and some correlative assemblages. This interval contains the last occurrence of *Euoplocyon*.

Woodburne et al. (1990) distinguish a Second Division Fauna that extends from Valley View Quarry upward to just below New Year Quarry. This interval is marked by an increase in diversity among the borophagine canids (Wang et al. 1999), especially taxa such as *Protepicyon raki* and *Aelurodon asthenostylus* that presage the late Barstovian and Clarendonian ascension of species in these clades. Species of *Cynarctus* and *Paratomarctus* also appear at these levels, and *Cynarctoides* has its last appearance there. The interval has a brief co-occurrence of the two common Barstow horse species, *Acritohippus stylodontus* and “*Merychippus*” *intermontanus* at the level of the Skyline Quarries. *Archaeohippus* (*A. mourningi*) is limited to just below the Skyline Tuff in the Second Division. The Second Division spans chron C5Bn (15.3–14.8 Ma).

The first local appearance of Proboscidea (here nearly synchronous for both mammutids and gomphotheriids) occurs near the base of the First Division (at about the level of New Year Quarry and just below the Dated Tuff, 14.8 Ma; Woodburne et al. 1990) at the initiation of the interval typified by the limited Barstow Fauna. Important taxa characterizing this interval (Barstow Fauna s.s.) include *Ischyrocyon* (Hunt 1998), *Hemicyon* (*Plithocyon*) *barstowensis*, “*Merychippus*” *intermontanus* “*Merychippus*” *sumani*, rare rhinos, *Meryceros joraki*, *Paramoceros*, and *Procamelus*. The anchitherine horse *Megahippus* (*M. mckennai*) has its first occurrence. *Parapliosacomys* has its first, and the oreodont *Mediochoerus* (*M. mohavensis*) has its only Great Basin appearance and the latest for the genus. Radioisotopic dating of ash beds and magnetostratigraphy calibrate the span of the First Division as 14.8–13.4 Ma in chrons C5AD–C5ACn. In the eastern Mojave Desert the Cronese Local Fauna, a correlative of the Barstow Fauna, is associated with an ash date of 12.6 Ma,

suggesting a somewhat longer span for the biochron of the Barstow Fauna. If accurate, this date and that for the basal part of the Ricardo faunal sequence nearly overlap, pointing to a phase of rapid turnover in western North America in the medial Miocene.

In 1998 Woodburne reviewed the lithostratigraphy and biostratigraphy of the Hector Formation in the Cady Mountains and clarified the correlation of the faunal successions in the northern and southern parts of the outcrop belt. Critical in this correlation was the recognition that the Logan Mine Local Fauna in the southern part of the range was contained in sands and gravels whose source was the Peach Springs Tuff, whereas the Lower Cady Mountains Local Fauna in the northern part of the outcrop was obtained from rocks underlying the tuff itself. Thus the Logan Mine is slightly younger than the Lower Cady Mountain assemblage, both about 18–19 Ma, in agreement with their early Hemingfordian faunal content. The Hector Formation contains faunas of late Arikarean to early Hemingfordian age and supports the calibration of that boundary near 19 Ma. Their strong biological relationships with correlative faunas of the midcontinent suggest that ecological similarity across the western half of the continent seen in the Late Oligocene continued into early Miocene time.

In recent years a number of mammal faunas have been collected in eastern San Bernardino County, California, by R. E. Reynolds and parties from the San Bernardino County Museum. One of the more informative of these (Reynolds et al. 1995) was obtained in Hackberry Wash from lacustrine deposits laid down on the margin of the Woods Mountain volcanic center and intercalated with the regionally traceable Peach Spring Tuff (18.5 Ma) and the Wild Horse Mesa Tuff (17.75–17.73 Ma). Fossil mammals were obtained at several sites in these deposits and are called the Hackberry Fauna, which seems to represent a coeval assemblage including (list revised from that of Reynolds et al. 1995 using casts generously donated by Reynolds): ochotonid, cf. *Trogomys*, *Metatomarctus* cf. *canavus*, *Menoceras barbouri*, *Protolabis* sp., cf. *Hesperocamelus*, and *Aletomeryx occidentalis*. This fauna is of early Hemingfordian age, correlative with the Boron Local Fauna of the western Mojave Desert and also the Peninsular Range sites in Orange County, California, discussed earlier. Chronologic information accompanying all these sites places them between 17 and 19 Ma, similar to the calibration of early Hemingfordian faunas in the northern Great Plains (MacFadden and Hunt 1998).

Continued work by one of us (D.P.W.) on the biostratigraphy of the Dove Spring Formation (Ricardo Group) has resulted in a biostratigraphy for the mammals

(Whistler in Whistler and Burbank 1992) and a chemical characterization of ash beds (Perkins et al. 1998) to provide radioisotopic ages by correlation of the ash beds with their sources. This work has shown that there is a significant hiatus in the lower part of the section where chron C5r is almost entirely removed at roughly the transition between the Iron Canyon and Ricardo faunas of our previous discussion (Tedford et al. 1987:159). Using the "base defines boundary" principle in chronostratigraphy, the *Cupidinimus avawatzensis/Paracosoryx furlongi* Assemblage Zone is still defined by the biostratigraphy even though the upper part of the zone is lost in the unconformity. Further difficulties with the composite magnetostratigraphy below this hiatus have led to some uncertainty in the calibration of the Iron Canyon interval, but a credible ash identification in this interval (Cougar Point Tuff V,  $12.07 \pm 0.04$  Ma, of Perkins et al. 1998, about 100 m above the base) suggests that the age of the base of the formation is close to 12.5 Ma. A concordance of all evidence indicates that the top of the Dove Spring Formation extends to 8.0 Ma and beyond into unfossiliferous rocks.

The new work has been fortunate in finding a local fauna in the basal gravels of the Dove Spring Formation that clearly links the base of the unit faunally to the Iron Canyon Fauna above. This local fauna contains *Copemys russelli*, *Cupidinimus tertius*, sciurids, *Pliohippus* cf. *tantalus*, and *Paracosoryx*.

Whistler (in Whistler and Burbank 1992) subdivide the Dove Spring biostratigraphy into four assemblage zones, characterized by the coexistence in each zone by two taxa: the *Ustatochoerus profectus/Copemys russelli* Assemblage Zone and the *Cupidinimus avawatzensis/Paracosoryx furlongi* Assemblage Zone (together roughly equivalent to the Iron Canyon Fauna of Tedford et al. 1987), the *Epi cyon aphobus* (= *haydeni*)/*Hipparion forcei* Assemblage Zone roughly equivalent to the Ricardo Fauna s.s., and the *Paronychomys/Osteoborus diabloensis* (= *Borophagus littoralis*) Assemblage Zone (roughly equivalent to the Dove Spring Fauna).

One of the important conclusions that emerges from this biostratigraphy is the length and overlap of the local range zones of the horse taxa used by Savage to define his Cerrotejonian and Montediablan stages in coastal California (Prothero and Tedford 2000). All the taxa have longer ranges in the Dove Spring Formation, with broad overlap between *Hipparion tehonense* and *Pliohippus tehonensis* (Cerrotejonian) and the supposedly replacing *Hipparion forcei* and *Pliohippus leardi* (Montediablan). The only replication of a like biostratigraphic character lies in the *E. haydeni/H. forcei* Assemblage Zone, in late chron C5n and C4Ar, where the local range

zones of the Cerrotejonian pair terminate before the Montediablan pair. Such evidence would confirm the young position for the Montediablan but leaves the exclusive overlap of the Cerrotejonian taxa in an interval attributed to chron C5n by Prothero and Tedford (2000). This conclusion is compatible with the biostratigraphic relationships of other component taxa of these stages.

The succession of taxa in the Dove Spring Formation is similar to that seen in the upper Valentine and Ash Hollow formations of north-central Nebraska. Of particular interest is the correlation of the oldest Dove Spring levels (Iron Canyon Fauna). As advocated by Tedford et al. (1987), this assemblage shares *Cormohipparion occidentale*, *Megahippus* cf. *matthewi*, *Ischyrocyon*, *Ustatochoerus medius*, and large merycodonts (*Paracosoryx* rather than the Great Plains *Cosoryx*) with the Burge Fauna. Judging from the Dove Spring evidence, this association of taxa would have an age of about 12–12.5 Ma. In great contrast to western North America, however, there is no faunal turnover in the Great Plains at the close of the Barstovian but rather chronofaunal continuity between the faunas of the Burge and older parts of the Valentine, a situation strikingly unlike the relationships between the faunas of the Barstow and Dove Spring formations. The effect of this turnover in the West is the introduction of faunas more like those of the Great Plains to western assemblages of Clarendonian and Hemphillian ages. This evidence also highlights the strong zoogeographic contrast between the Barstow Fauna and its temporal equivalents in the lower part of the Valentine Formation of north-central Nebraska.

**Western Nevada (F)** New  $^{40}\text{Ar}/^{39}\text{Ar}$  dates now available for the Barstovian Tonopah Local Fauna and Stewart Springs Fauna of western Nevada (Swisher 1992) were obtained from ash beds more intimately associated with these assemblages and help clarify their temporal positions. Biotite and plagioclase separated from ash falls in the Siebert Tuff immediately below the main fossil quarry of Tonopah yielded dates of  $15.16 \pm 0.063$  Ma (mean of three determinations), and a biotite tuff overlying the Stewart Springs Tedford Pocket locality yielded  $14.96 \pm 0.24$  Ma on biotite and  $14.89 \pm 0.53$  Ma on plagioclase. These dates confirm the near contemporaneity of these faunas and give a 14.9 Ma date for the occurrence of mammutid Proboscidea at the local assemblages (specifically the Savage Canyon site) grouped as the Stewart Springs Fauna.

The Tonopah and Stewart Springs faunas share such elements as the borophagines *Paratomarctus temerarius* and *Aelurodon asthenostylus*. In addition, Tonopah has *Micro-*

*tomarctus conferta* and *Paracynarctus kelloggi*; Stewart Springs has *Carpocyon compressus*. Both have “*Merychippus*” cf. *intermontanus* (“*M. calamarius*”), *Peraceras supercilliosum*, and *Paracosoryx loxoceras*. These assemblages correlate temporally and faunally with the Second Division and early Barstow faunas of the Barstow Formation.

New  $^{40}\text{Ar}/^{39}\text{Ar}$  dates (Swisher 1992) refine the calibration of the mammal-bearing deposits in the Fish Lake Valley and Stewart Valley sites that were combined as the Fish Lake Valley Fauna in Tedford et al. (1987). These new dates confirm the contemporaneity of these sites within a range of 11.6–11.7 Ma. The composition of the Fish Lake Valley Fauna closely resembles those of early Clarendonian age from the northern Great Plains and stands in strong compositional contrast to late Barstovian sites in the Great Basin region, emphasizing once again the remarkable turnover at the close of the Barstovian in western North America.

Recently discovered localities in thick unnamed basin fill cropping out on the western flank of the Pine Nut Mountains in Douglas County, Nevada, near the California border, have yielded an unbroken Hemphillian–Blancan succession (Kelly 1994, 1997). A small assemblage of latest Hemphillian mammals including the leporids *Hypolagus gidleyi* and *Lepoides lepoides*, the rhino *Teleoceras*, the horse *Dinohippus*, the llama *Hemiauchenia*, and a cervid (Kelly 1997) make up the Washoe Local Fauna of late Hemphillian age. An ash bed near the lowest occurrence of Blancan mammals (*Equus* sp. and *Ursus abstrusus*, taxa included in the Buckeye Creek Local Fauna) gave a 4.96 Ma radiometric date (Lindsay et al. 2002). This is a slightly older date for the Hemphillian–Blancan transition than reported from the Yepómera or Rancho El Ocote faunal sequences that also include this superposition.

## COLUMBIA PLATEAU

**Northwestern Nevada (G)** Fossil mammals occur in volcanoclastic sediments in the rhyolitic volcanic province of northern Nevada. Ash flow tuffs there facilitate the correlation of isolated sites over long distances and provide radioisotope calibration of the sequence. Especially noteworthy is the volcanic field of northwestern Nevada (Washoe and Humboldt counties) and adjacent Oregon, which contains the superposed Massacre Lake Local Fauna (late Hemingfordian), the Virgin Valley Fauna (early Barstovian), and the High Rock Lake sites (early late Barstovian). These sites document the early appearance of zygodont mammutids in the northwestern United States and reinforce the evidence from Oregon of a distinguishable zoogeographic province in that region.

The Massacre Lake Local Fauna lies below an ash flow tuff thought to be an outlier of the Summit Lake Tuff (Noble et al. 1970, 1973). This unit was dated by Evernden et al. (1964, who first announced discovery of the fossil mammal site) at 15.6 Ma but was later revised to 16.5 Ma by Swisher (1992), who named this local unit the Tuff of Big Basin. The Massacre Lake Local fauna has a unique mixture of Columbia Plateau Barstovian forms such as *Liodontia*, *Paracynarctus kelloggi*, *Desmatippus*, *Dromomeryx*, and *Ticholeptus*, with Great Plains Hemingfordian forms including *Alphagaulus*, *Protomarctus*, *Anchitherium*, *Parahippus*, and *Bouromeryx* and western Hemingfordian–Barstovian forms such as *Parapliohippus*. A fragment of a mammutid tooth was found at this site (Morea 1981; Woodburne and Swisher 1995).

The Virgin Valley Fauna (Merriam 1911) was obtained from the “lower member of the Virgin Valley beds” of Merriam (1910). Subsequent work recognized that the “upper member” was equivalent to his “Thousand Creek beds” of Hemphillian age, and the Virgin Valley beds were accordingly amended. The rocks containing the Virgin Valley Fauna, the Virgin Valley Formation (*sensu stricto* [s.s.]), interfinger with and overlie the Cañon Rhyolite, which has given an  $^{40}\text{Ar}/^{39}\text{Ar}$  date of  $16.28 \pm 0.07$  Ma (Swisher 1992) beneath the fauna, perhaps a maximum age. The fauna is more closely bracketed by ash beds in the Virgin Valley Formation that range from  $15.85 \pm 0.05$  to  $15.18 \pm 0.03$  Ma. Unlike the Massacre Lake Fauna, the Virgin Valley Fauna lacks most Great Plains elements and instead resembles Columbia Plateau equivalents. It is the type locality for such taxa as *Liodontia alexandrae*, *Oreolagus nevadensis*, *Paracynarctus kelloggi*, *Zygodont merriami*, *Moropus? merriami*, and *Parablastomeryx mollis* and also includes *Protomarctus*, *Hypohippus*, *Desmatippus*, *Merychippus brevidontus*, *Acritohippus isonesus*, and *Dromomeryx*.

This endemism continues into the fauna at the High Rock Lake sites (Merriam 1911; Stirton 1939), which are contained in volcanoclastic sediments bracketed below by the Soldier Meadow Tuff, best dated at  $16.12 \pm 0.03$  Ma (Swisher 1992) and above by a local ash dated approximately 14.5 Ma (Swisher 1992). Most of the fauna listed for the Virgin Valley continues in this assemblage, including *Zygodont*, but the equine horse “*Merychippus californicus*” (Downs 1961), a large oreodont, and *Merycodus nevadensis* (type locality) join the fauna.

**Eastern Oregon (H)** The historic John Day region’s rich fossil deposits have been repeatedly exploited since the 1860s and made famous by collectors for Marsh (Schuchert and LeVene 1940) and Cope (1884). They

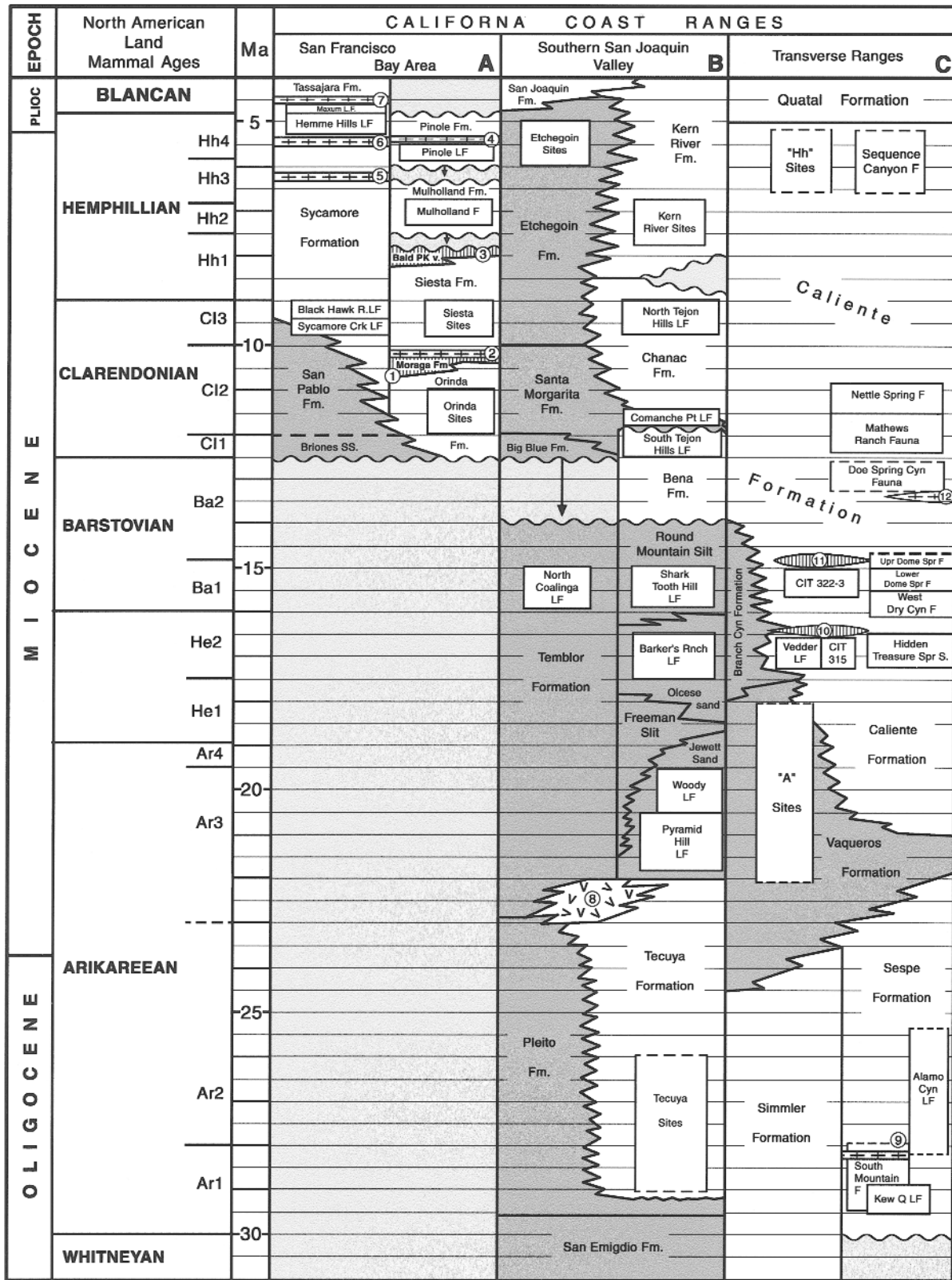
were first studied stratigraphically by Merriam and Sinclair (1907), later revised by Fisher and Rensberger (1972), and are now subject to further revision since the most important sites were incorporated into the John Day National Monument in 1980. This sequestering of fossiliferous terrain has initiated new field studies by T. Fremd and three of us (L.B.A., R.M.H., C.C.S.), in company with M. O. Woodburne. This new work is still ongoing, but a progress report (Fremd et al. 1994) compiled the fauna of intervals to produce the first biostratigraphy for major elements of the fauna, including the placement of many historic specimens. Stepleton and Hunt (1994; Hunt and Stepleton 2001) have focused on the Upper John Day Formation lithostratigraphy and biostratigraphy. Albright, Woodburne, and Swisher have initiated a magnetostratigraphic study to revise Prothero and Rensberger's (1985) pioneer effort. Swisher continues to provide  $^{40}\text{Ar}/^{39}\text{Ar}$  single-crystal dates for the many ash beds that underpin the correlation of the magnetostratigraphy with the Geomagnetic Polarity Time Scale and provide timing for significant volcanic events. Rensberger's (1971, 1973, 1983) systematic studies of the geomorph and aplodontid rodents are important modern contributions, but their chronology will necessarily be modified by the studies enumerated earlier. Comparable systematic work on canids (Wang 1994; Wang et al. 1999), amphicyonids (Hunt 2001 and pers. comm., 2003) and other groups are under way. Such studies are particularly important because faunas of the John Day Formation, along with comparable assemblages in the northern Rocky Mountains west of the Continental Divide, have a distinct taxonomic composition that delineates a distinctive zoogeographic region that existed there during early Arikarean time.

The middle part of the John Day Formation, above the basal Big Basin Member, is composed of the Turtle Cove Member and its partial facies, the Kimberly Member overlain disconformably by the Haystack Valley Member (*sensu lato* [s.l.]; Fisher and Rensberger 1972). The faunal succession is now keyed to a succession of widely traceable tuffs, most of which have been dated by  $^{40}\text{Ar}/^{39}\text{Ar}$  on single crystals using the laser fusion method (Swisher, pers. comm., 2002). Fremd et al. (1994) have given faunal lists for the interash intervals (lithostratigraphic units A–L; see figure 6.2H) of five linked biostratigraphic sections extending along a 10-km segment of the Main Fork of the John Day River from Turtle Cove to Bone Creek in western Grant and eastern Wheeler counties, Oregon. Based on recent U.S. National Parks collecting and knowledge of the location of historical specimens, many taxa have been allocated to these intervals, yielding a bio-

stratigraphy that contains much of the John Day "Fauna" (figure 6.2H) in a total section about 500 m thick.

Radioisotopic dates and magnetostratigraphy suggest that the sequence begins at about 30 Ma and extends to about 18 Ma. Prothero and Rensberger's (1985) magnetostratigraphy is called into question by this extended temporal range and their necessary reliance on a single date that was much too young (the Picture Gorge Ignimbrite at 26 Ma). However, their magnetostratigraphy for the Picture Gorge–Deep Creek Tuff interval can be correlated with the chron C<sub>10n</sub>–C<sub>9n</sub> interval (27–29 Ma, Berggren et al. 1995) using the new data.

In the northern Great Plains the disconformity between the White River and Arikaree Groups falls in chron C<sub>10n</sub>, but many elements of the White River Chronofauna survive into the base of the Arikaree; their extinction occurs just above the basal sands (Gering and upper Sharps formations) of the Arikaree Group. Although we see loss of some of these elements in the John Day (*Eusmilus*, *Nimravus*, *Dinictis*, and *Perchoerus*) below the Deep Creek Tuff (27.89 ± 0.57 Ma), others survive nearly to the top of the middle John Day (*Agriochœrus*, *Eporeodon*, *Hypetragulius*, and *Palaeolagus*) and beyond (*Miohippus*). Some of the taxa that appear in the northern Great Plains after the extinction of White River elements, including *Archaeolagus*, *Alwoodia*, *Pleurolicus*, *Desmatochoerus megalodon*, and *Oreodontoides oregonensis*, are also present at the Picture Gorge Ignimbrite (28.7 ± 0.07 Ma) or below, a little earlier in the John Day Formation than in the Plains (Tedford et al. 1996). Other taxa that first appear at the Deep Creek Tuff include *Entoptychus*, *Paroreodon*, *Merycoides*, *Hypsiops*, *Promerycochoerus*, and *Gentilicamelus*. Most of these genera extend to the top of the middle John Day and some beyond into the Haystack Valley Member. Rensberger's (1971, 1973, 1983) biostratigraphy is compromised by recent work that shows that although the local range zones of *Meniscomys* and *Entoptychus* are successional, they overlap in the Deep Creek Tuff–Tin Roof Tuff interval, and the local range for *Pleurolicus* occurs with *Meniscomys* only at the base of the latter's range zone, below the Deep Creek Tuff (see local range zones in figure 6.2H). *Allomys* traverses the entire middle John Day. *Diceratherium* and *Meniscomys* do not extend beyond the Tin Roof Tuff (25.9 ± 0.31 Ma), but some new elements are added in the succeeding interval ("L" in figure 6.2H: *Schizodontomys*, *Leidymys nematodon*, *Phenacocoelus*, *Moropus*, and *Nexuotapirus robustus*). The occurrence and succession of taxa in the John Day Formation above the Deep Creek Tuff represent a span of time that has not been clearly delineated biochronologically in the Great Plains.



**FIGURE 6.2** Correlation charts showing the chronologic sequence of rock units and contained fossil assemblages discussed in this chapter and in Tedford et al. (1987). The time scale follows figure 6.3. AZ, assemblage zone; B., Basalt; CIT, California Institute of Technology; F., Fauna; Fm., Formation; LF, Local Fauna; M. or Mbr., Member; Q, Quarry; Riv., River; S., site; SS, sandstone; V, volcanic.



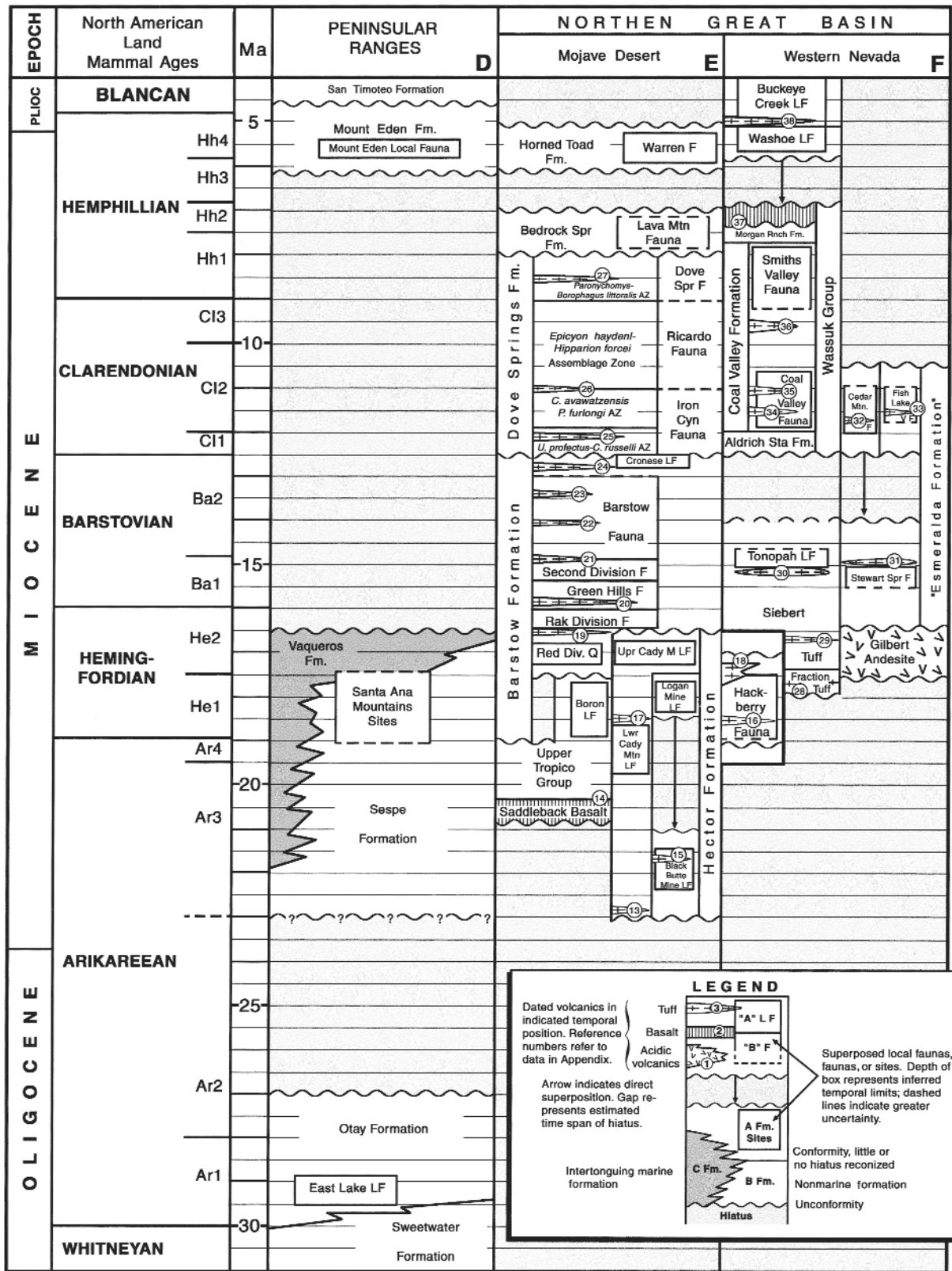


FIGURE 6.2 (continued)

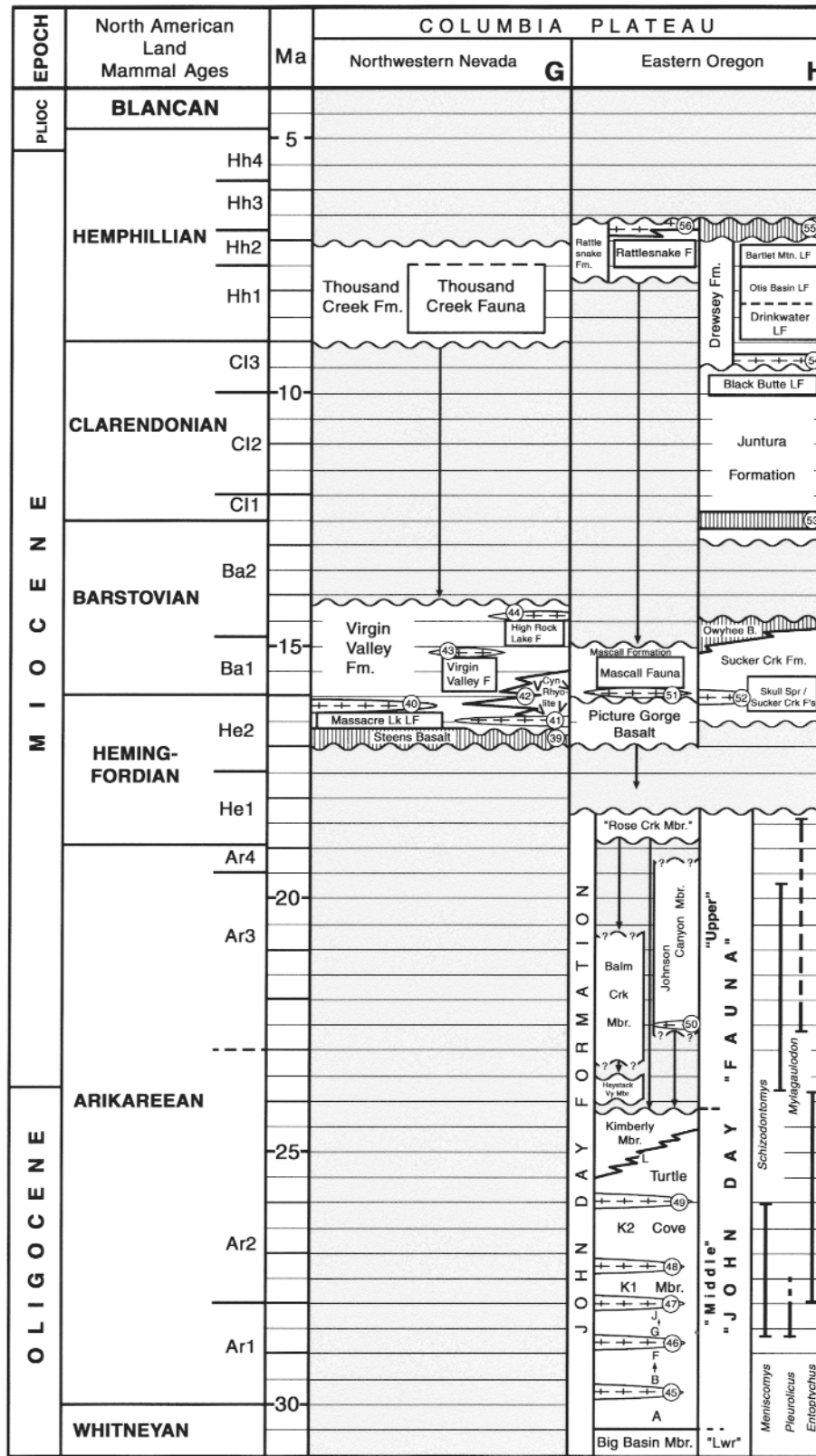


FIGURE 6.2 (continued)

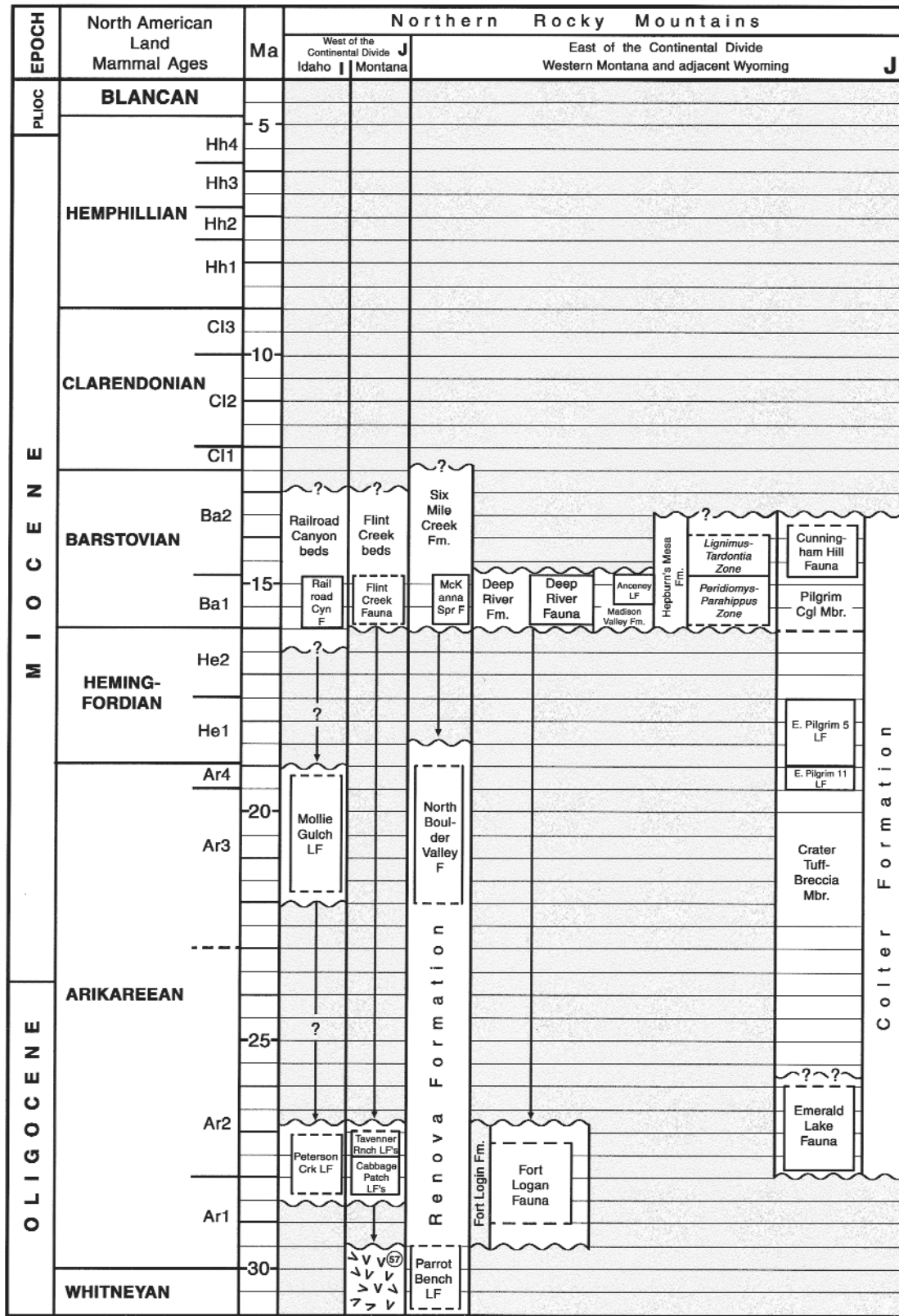


FIGURE 6.2 (continued)

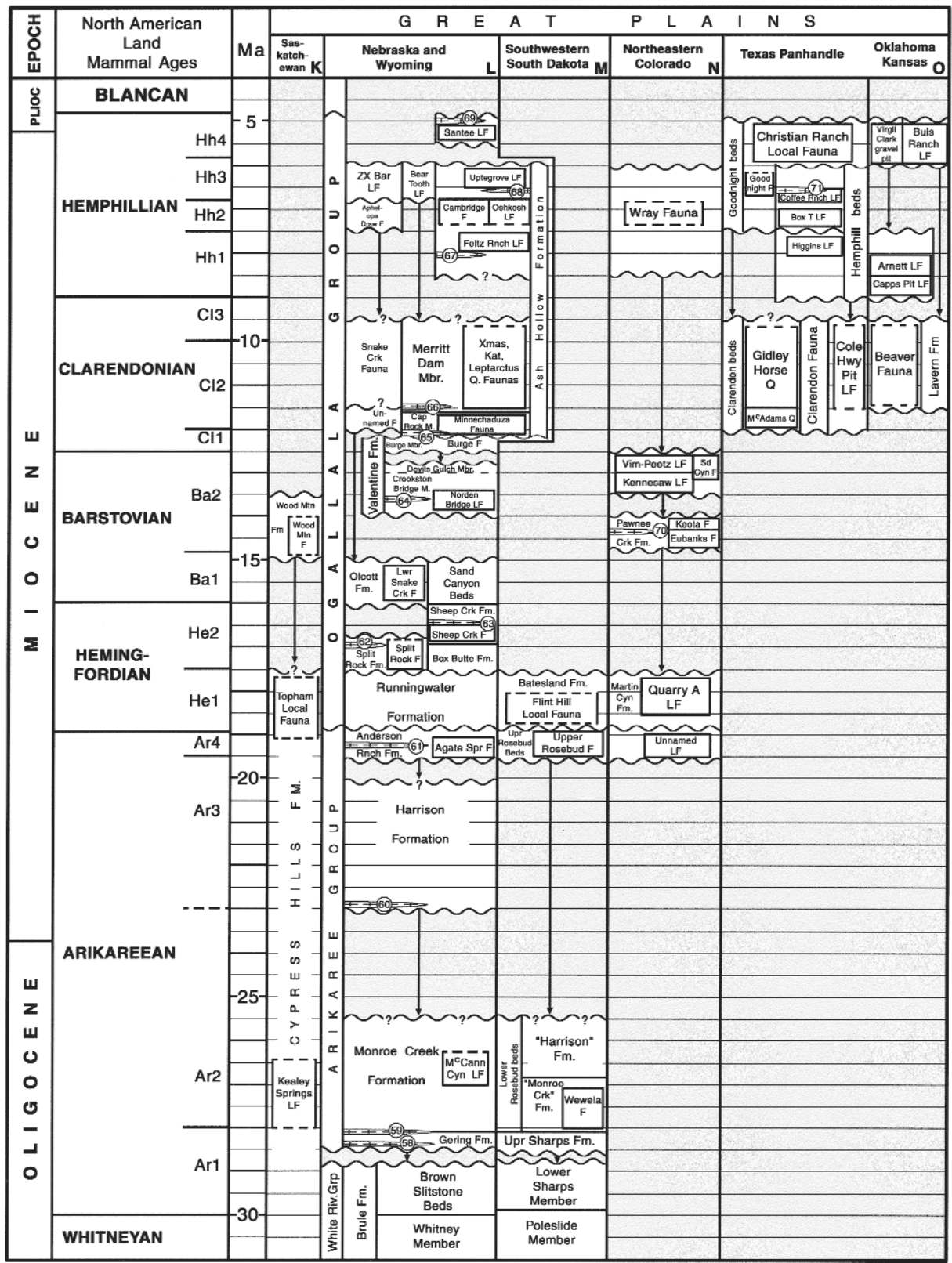


FIGURE 6.2 (continued)

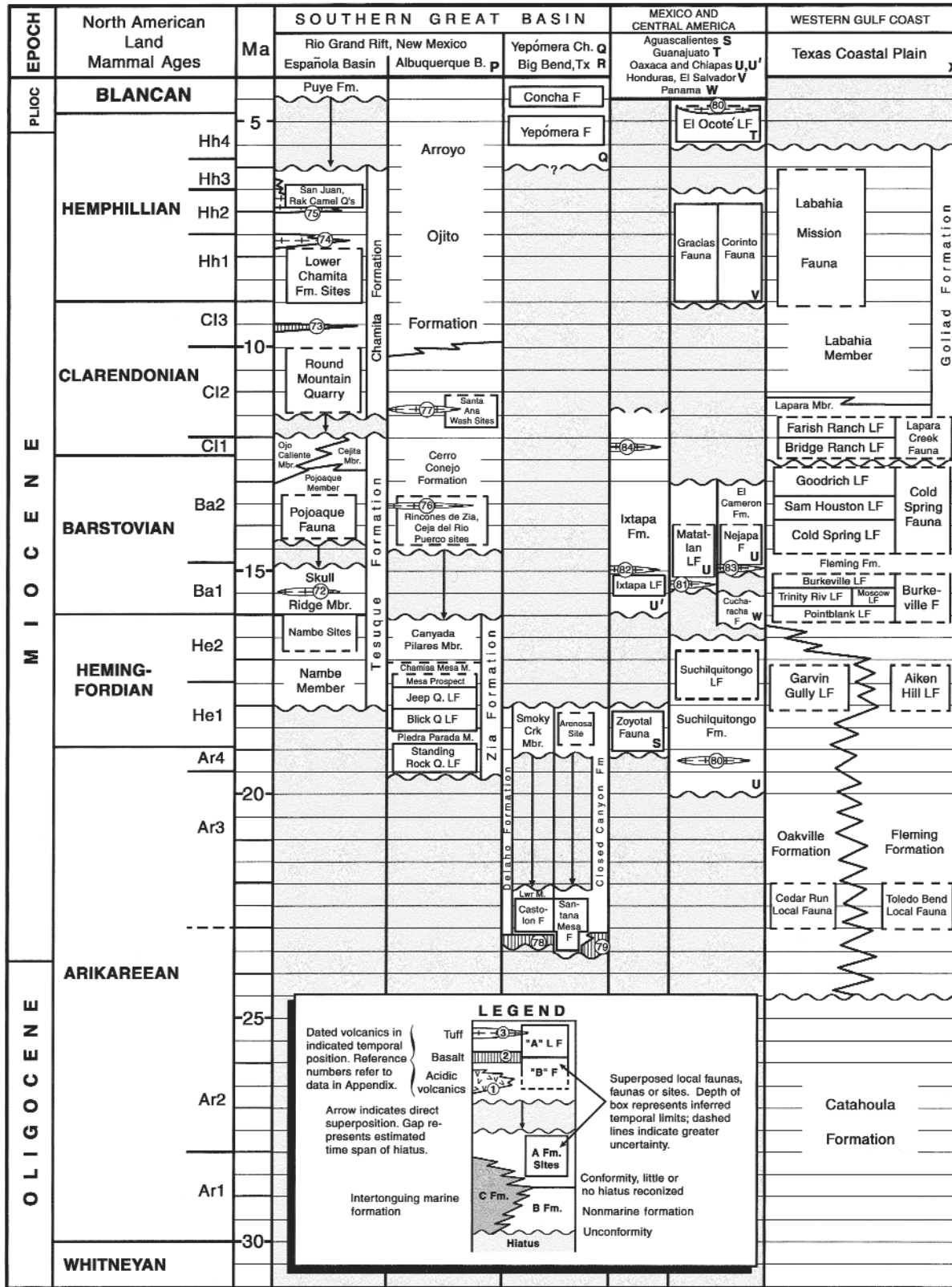


FIGURE 6.2 (continued)

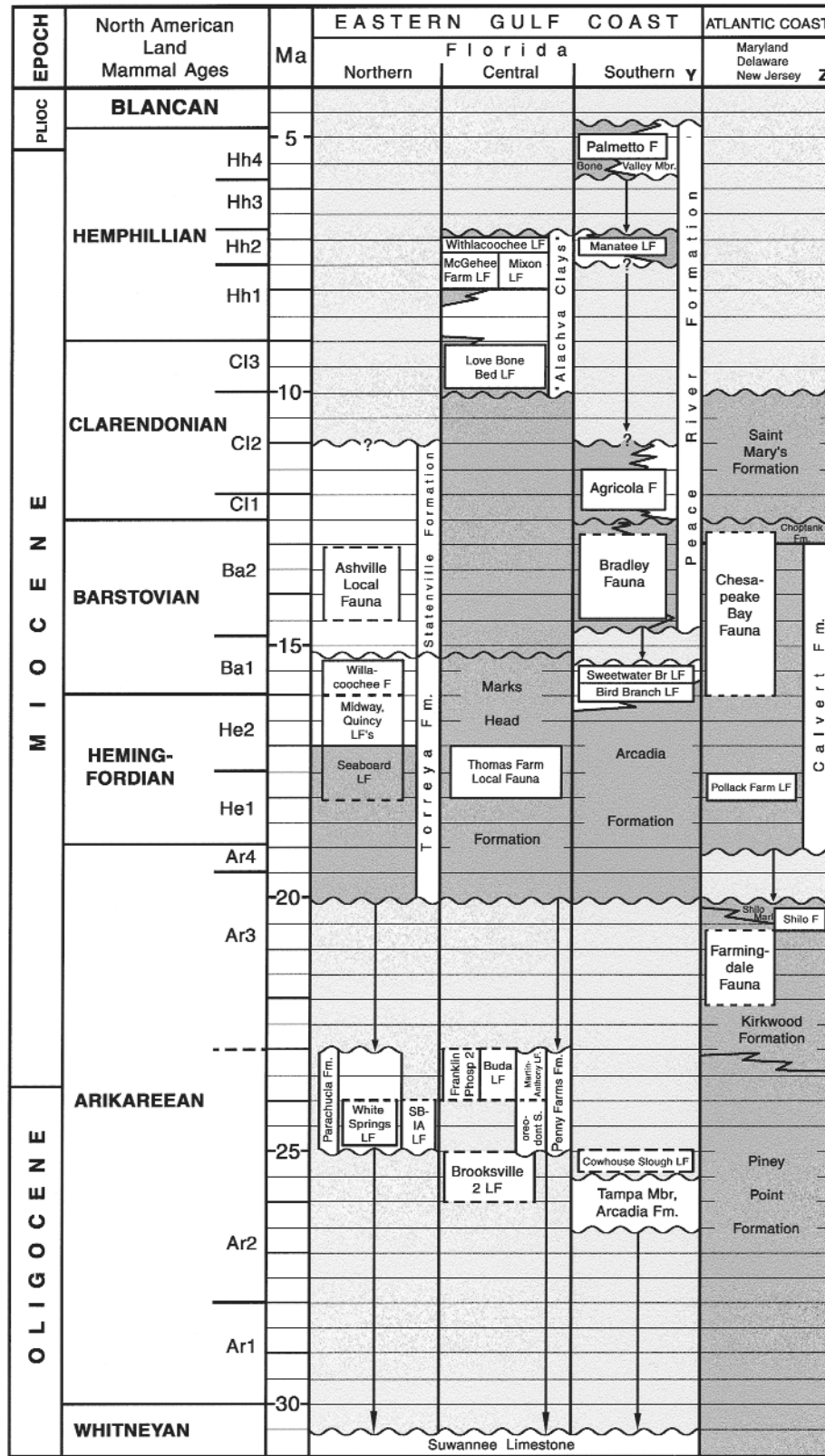


FIGURE 6.2 (continued)

Recent work by Hunt and Stepleton (2001) has established that the fauna of the Haystack Valley Member of Fisher and Rensberger (1972) is a composite assemblage derived from at least three formation rank rock units that extend in age from the early late Arikareean to the early Hemingfordian. *Merycochoerus*, *Moropus*, *Parahippus pawniensis*, *Mylagaulodon*, and a *Barbouromeryx*-like dromomerycid occur in the uppermost unconformity-bounded unit of the lithostratigraphic succession south of Kimberly post office (Rose Creek Member of Hunt and Stepleton in review). Several of these taxa indicate a correlation with the lower part of the Runningwater Formation of western Nebraska (Northeast of Agate Local Fauna; MacFadden and Hunt 1998, ~18.8–18.2 Ma) and suggest a greater affinity to Great Plains faunas of this age than previously believed. A fauna from the western facies of the John Day Formation on the eastern flank of the Cascade Range (Warm Springs Local Fauna of Woodburne and Robinson 1977; Dingus 1990) is of similar age.

Hunt and Stepleton (2001) limit the Haystack Valley Member to lithically homogeneous tuffs with fluvial welded tuff-bearing gravels at Haystack Valley, best exposed along Balm Creek in the type area of Fisher and Rensberger (1972). The Balm Creek outcrops of the revised member produced nearly the entire fauna attributed to the Haystack Valley Member of Fisher and Rensberger in Haystack Valley. This fauna includes the first occurrence of the peccary *Hesperhys*, the pleurolicine rodent *Schizodontomys*, and the last occurrence of *Entoptychus*, *Allomys*, and *Miohippus*. Hunt and Stepleton regard this fauna as of early late Arikareean age, supported by an  $^{40}\text{Ar}/^{39}\text{Ar}$  age determination of 23.5–23.8 Ma.

It has long been known that one of the striking contrasts between the John Day faunas and approximate contemporaries from the northern Great Plains and the northern Rocky Mountains east of the Continental Divide was the absence of the leptachenine oreodonts and protoceratids at John Day and the abundant presence of *Entoptychus* species there. Other compositional differences mark the John Day faunas, especially the presence of diverse aplodontid rodents, a largely endemic fauna of small hypocarnivorous borophagines (Wang et al. 1999), a different oreodont fauna except for the shared occurrence of *Promerycochoerus*, the presence of *Gentilicamelus* and *Paratylopus* camels, and particularly the survival of species of White River genera beyond 25 Ma.

Recently available  $^{40}\text{Ar}/^{39}\text{Ar}$  dates (Swisher 1992) refine the chronologic position of the Mascall Formation sites in the John Day region and also provide better calibration of the Skull Springs Fauna of eastern Oregon. At the Mascall Formation type section, a tuff in unit 2 of

Downs's (1956) measured section, 25 feet below the lower mammal-bearing unit 5, yielded plagioclase that gave a date (average of four determinations) of  $15.77 \pm 0.07$  Ma. The Owyhee Basalt drawn by Tedford et al. (1987) beneath the Sucker Creek faunal sites actually overlies them. Its whole rock K–Ar dates, averaged to  $14.7 \pm 0.3$  Ma (Bottomley and York 1976), give a minimum age for the fauna. The Leslie Gulch Tuff in the Sucker Creek Formation, of unknown stratigraphic relationship to the faunal occurrence, was K–Ar dated (sanidine; Ekren et al. 1984) at  $15.8 \pm 0.6$  Ma, in agreement with the dates for the correlative Mascall and Skull Springs faunas. Recently a suite of tuffs in the Sucker Creek Formation were dated by  $^{40}\text{Ar}/^{39}\text{Ar}$  single-crystal laser fusion methods (Downing and Swisher 1993), bracketing the fauna between 15 and 15.5 Ma.

## NORTHERN ROCKY MOUNTAINS

**Eastern Idaho (I)** A zoogeographically important sequence of faunas is contained in strata of early Arikareean and early Hemingfordian age that crop out on the western flanks of the Beaverhead Mountains west of the Continental Divide in Lemhi County, eastern Idaho. The oldest faunal sequence occurs in outcrops along Peterson Creek, a tributary of the Lemhi River, about 10 miles northwest of Leadore. Nichols (1976, 1979) described the biostratigraphy and the contained fossils from collections made by him (at the University of Montana) and parties from the Idaho State Museum. Lagomorphs are represented by the archaeolagine *Palaeolagus*. Aplodontid rodents are diverse: *Allomys*, *Niglarodon*, and two stratigraphically successive *Meniscomys* species. The promylagauline *Trilaccogaulus* is present, as is the beaver *Palaeocastor*, the cricetid *Pacculus*, two stratigraphically successive *Entoptychus* species; the horse *Miohippus*, and the oreodonts *Megoreodon* and *Mesoreodon*. In the main this fauna resembles those of the Cabbage Patch sequence, the Fort Logan Formation, and Great Plains faunas of the early Arikareean, especially those from the upper Sharps and lowest Arikaree ("Monroe Creek") of South Dakota and the Gering of Nebraska. Of interest in an assemblage from west of the Continental Divide are the high-crowned *Entoptychus* species typical of the John Day; however, these species (*E. fieldsi* and the younger *E. sheppardi*) are, in Nichols's (1976) estimation, more primitive than *E. basilaris*, the earliest occurring form in the John Day Formation. The latter taxon first occurs above the Deep Creek Tuff of the John Day Formation, which is dated close to 28 Ma, in agreement with the calibration of the Great Plains early Arikareean (Tedford et al. 1996).

Charles Falkenbach of the Frick Laboratory, American Museum of Natural History (AMNH), working in the same area in 1942, discovered a younger fauna in Mollie Gulch about 5 miles southeast of Peterson Creek. These beds, also sandy clays with interbedded ash, are more than 45 m thick but seem to contain a unified fauna, mostly of large ungulates. The Frick collection includes the horse *Parahippus*; the camels *Oxydactylus* cf. *longipes*, *Michenia* cf. *agatense*, and *Protolabis*; the oreodont *Merychys arenarum*; and the moschid *Blastomeryx*. All these taxa are typical of latest Arikareean assemblages of the Great Plains. Superposition of the rocks in Mollie Gulch with those in Peterson Creek has not been observed.

A younger fauna comes from strata exposed in Railroad Canyon on the high terrain at Bannock Pass. These deposits lie on pre-Tertiary basement rocks and dip eastward into the Horse Prairie Basin east of the Continental Divide in Beaverhead County, Montana. Some 300 m of clastic strata are exposed in the pass above the mid-Tertiary unconformity with sparsely fossiliferous deposits of early Miocene age. Fossil mammal remains were obtained throughout these exposures, but the largest collections made by Falkenbach, Nichols, and Barnosky come from the upper half of the sequence. Magnetostratigraphic studies (Zheng 1996) indicate that this part of the section lies in an interval bracketed by 16–13 Ma. Curiously, there is no detectable biostratigraphic change in the character of the fauna, which is an early Barstovian assemblage similar in composition to the Deep River, Madison Valley, and the lower zone at Hepburn's Mesa, Montana, which have been correlated with the early Barstovian 16.2–14.8 Ma interval. A few micromammals are known, and they significantly include *Peridimys* and *Cupidinimus*, which range through the sequence, the former genus being unknown below early late Barstovian rocks. Horses are diverse, including *Archaeohippus* cf. *ultimus*, *Hypohippus* cf. *osborni*, *Merychippus* cf. *insignis*, and *Acritohippus isonesus*. The oreodonts include *Merychys* (*Metoreodon*), *Ticholeptus zygomatus*, and *Brachycrus*. The camels are dominated by species of *Aepycamelus*, but *Paramiolabis* cf. *singularis* also occurs. Ruminants include *Blastomeryx*, *Rakomeryx* cf. *kinseyi*, and *Merycodus*. Such a fauna is typical of the Great Plains and northern Rocky Mountains early Barstovian assemblages from rocks lying directly on the mid-Tertiary unconformity (base of the fourth sequence of Hanneman and Wideman 1991).

**Western Montana and Adjacent Wyoming (J)** In later years workers (especially Kuenzi and Fields 1971) recognized that a regional unconformity separates the Tertiary basin fill of Montana into two sequences: the older Re-

nova Formation, volcanoclastic deposits of medial Tertiary age that range upward into the early Hemingfordian (ca. 18 Ma), and the unconformably overlying, predominantly epiclastic Six Mile Creek Formation of early Barstovian (ca. 16 Ma) and younger age. The seismic stratigraphic studies of Hanneman and Wideman (1991) reveal that most of the basins of southwestern Montana contain three sequences of unconformity-bounded later Tertiary units comparable with the White River, Arikaree, and Ogallala groups of the northern Great Plains. The Renova Formation subsumes two sequences; the Six Mile Creek Formation contains a single late Tertiary sequence.

The studies under way on the John Day faunal succession and those reviewed for the early Arikareean of the northern Great Plains again focus attention on the biostratigraphy of the similar span of time recorded in the Cabbage Patch beds in the Flint Creek and adjacent Deer Lodge basins of western Montana studied by Rasmussen (1969, 1977). This largely unpublished work is again reviewed in light of later studies in correlative regions discussed in this work. Rasmussen reconstructed the biostratigraphic succession from scattered fossiliferous outcrops largely on the basis of similarity of faunal composition. He recognized three faunal associations that could be attributed to the lower, middle, and upper part of the Cabbage Patch beds. The lowest fauna, largely from the well-known outcrops 4 km east of Drummond, contains *Ocajila*, *Pseudotrimylus*, *?Palaeolagus*, *Megalagus*, *Niglarodon*, *Agnotocastor*, *Eutypomys*, *Leidymys*, *Plesiosminthus*, *Cynodesmus* s.s., *?Perchoerus*, *Megoreodon*, *Desmatochoerus*, and *Pronodens* among the taxa that also occur in early Arikareean faunas of the John Day and Montana sites east of the Continental Divide (Fort Logan Formation) and those in the lower Arikaree Group of Nebraska. As Rasmussen (1977) concluded, this fauna has its closest similarity to that from the Gering Formation of Nebraska and the "Sharps Formation" (i.e., the "upper Sharps Formation" of Tedford et al. 1996). Differences are the diverse aplodontid assemblage (*Downsimus*, three *Niglarodon* species, and the endemic *Fossodontia*), an earlier occurrence of the heterosoricine *Pseudotrimylus*, and the endemic leptomerycid *Pronodens* in the lower Cabbage Patch beds.

The fauna of the middle part of the Cabbage Patch beds includes the first local occurrence of *Parvericius*, *Ampechinus*, *Archaeolagus*, *Alwoodia*, *Pleurolicus*, *Pacculus*, and *Diceratherium*. This fauna contains taxa in common with the lower Arikaree strata in South Dakota and western Nebraska. *Archaeolagus*, *Alwoodia*, and *Pleurolicus* also have their first appearances in strata just above the Sharps and Gering formations, there correlated with



chron C9n (27–28 Ma). At approximately the same temporal position in the middle John Day, *Alwoodia* and *Pleurolicus* appear just above the Picture Gorge Ignimbrite and *Archaeolagus* somewhat below.

The upper Cabbage Patch fauna contains the first local occurrences of *Stenoechinus*, ?*Gripholagomys*, *Gregorymys*, *Mookomys*, and ?*Euhapsis*, taxa largely limited to the northern Rockies and Great Plains, hence affording little basis for correlation with the John Day sequence. In South Dakota *Gregorymys* has its first local occurrence in the correlated “Monroe Creek Formation” above the Sharps Formation (L. MacDonald 1972), as does *Gripholagomys*. The genotypic species of *Euhapsis* occurs in the upper part of the type Monroe Creek Formation of Nebraska (Martin 1987). Thus the faunal sequence in the Cabbage Patch beds seems to represent a short span during early Arikareean time from about 28.3 to perhaps 27.5 Ma, although the sparse mammal fauna of the upper Cabbage Patch beds does not constrain the sequence precisely, and it could range into significantly younger intervals.

The Blacktail Deer Creek Fauna, an isolated occurrence in Beaverhead County, east of the Continental Divide in the southwest corner of Montana, is important to mention because it contains the holotypes of two widely recognized early Arikareean genera: the beaver *Neotocastor* (*N. hesperus* Douglass 1901) and the anthracothere *Arretotherium* (*A. acridens* Douglass 1901). The outcrop from which these fossils were obtained is about 122 m thick; the anthracothere and a rhino (“*Caenopus*,” Douglass 1901) were found near the base of the section, and the beaver was found near the top in the alternating succession of light tan sandstones and clays. Subsequently Hibbard and Keenmon (1950) revisited the locality and added a new species, *Gregorymys montanensis*, and the oreodont *Desmatochoerus megalodon* (“*Promerycochoerus (Parapromerycochoerus) barbouri*,” Hibbard and Keenmon 1950) from near the lower fossil occurrence. *Gregorymys* has its first record in the Great Plains in the “Monroe Creek” of South Dakota, and species of the genus extend only through the Arikareean. *Desmatochoerus* occurs in the Gering and equivalent strata but is limited to the early Arikareean. The earliest record of *Arretotherium* (as *A. leptodus* closely allied or conspecific with *A. acridens*) is in the “Monroe Creek” of South Dakota. This taxon is replaced in the early Hemingfordian by *A. fricki*. The genus is unknown in the intervening strata. Outside Montana *Neotocastor hesperus* curiously occurs only in faunas of early or medial Arikareean age in the Gulf Coast of Texas and Florida. These data sum to an early Arikareean age for the Blacktail Deer Creek assemblage.

The Fort Logan Formation (*sensu* Koerner 1940) from the Smith (“Deep”) River basin, east of the Continental Divide in western Montana, has a microfauna (Black 1961; Rensberger 1979, 1981) that includes taxa (*Palaeolagus hypsodus*, *Megalagus*, *Agnotocastor*, and *Eumys*) that do not occur above the top of the Sharps and Gering formations of the Great Plains. Likewise, the occurrence of *Pacliculus* has its earliest Great Plains occurrence in the upper Sharps Formation, and *Cynodesmus* does not occur above the basal Arikaree (Wang 1994). The Fort Logan oreodont fauna includes the leptachenines *Sespia* and *Leptauchenia* and the genera *Promerycochoerus*, *Mesoreodon*, *Desmatochoerus*, and *Merycoides*. The latter four oreodonts co-occur in the lower Arikaree Group and in the John Day Formation around the level of the Deep Creek Tuff. Again the inference is that the Fort Logan Formation contains faunas basically of early Arikareean age. As in the case of the Cabbage Patch sequence, the presence of younger Arikareean assemblages is not evident.

Imprecise mapping of the contact between the Fort Logan and Deep River formations by Koerner (1940) failed to remove the confusion regarding the faunal content of these units, especially the assignment of clearly Arikareean oreodonts (the leptachenines, *Desmatochoerus*, *Merycoides*, and *Promerycochoerus*) to the Deep River, which otherwise bears a Barstovian fauna. Because the fauna of the Deep River Formation was one of the earliest early Barstovian assemblages (along with Mascall) to be described (by Cope and Scott in the nineteenth century), much of the faunal list represents specific and generic holotypes. Therefore it stood as a singular geochronologic reference fauna into the early twentieth century. Included are the erinaceid insectivores *Brachyterix* and a late occurrence of *Parvericius* and the last occurrence of the Proscalopidae (*Mesoscalops*). The last occurrences of the mylagaulid *Mesogaulus*, the heteromyine *Harrymys*, and the marmotine *Protospermophilus* are in this and correlative faunas. The Mammutidae are present (*Zygodon brevidens*; Lambert and Shoshani 1998), as are the horses *Desmatippus crenidens* (Scott 1893), *Hypohippus equinus* (Scott 1893), and *Acritohippus isonesus*; the oreodont *Ticholeptus zygomatus* (Cope 1875); the camel *Aepycamelus*; and the dromomerycids *Subdromomeryx antilopinus* (Scott 1893), *Dromomeryx borealis* (Cope 1878), and *Rakomeryx kinseyi* (Frick 1937). The presence of a mammutid proboscidean is noteworthy in this early Barstovian site, adding credence to other records of the group in deposits of similar age elsewhere in North America.

Although fossil mammal remains had been noted by early geologists (Hayden and especially Peale 1896) in the

bluffs of the lower Madison River where it crosses Tertiary deposits of the Three Forks Basin (Robinson 1961, 1963; Dorr 1956), Earl Douglass was the first to make collections in this region beginning in 1894 and reported in his University of Montana thesis of 1899. The greater part of the fauna was described by him in that and subsequent works, later revisions of some taxa have been made, but few additions to the faunal list have resulted from fieldwork in the twentieth century.

Douglass (1899) gives a stratigraphic section of the Madison River bluffs that stretch along the east side of the river south of Three Forks and Logan in Gallatin County, Montana. Initially, he referred to the fossil mammal-bearing rocks as the "Loup Fork beds" or the "Loup Fork Epoch," further noting, "The beds overlie the White River and occupy the top of the triangular bench between the Madison and Gallatin rivers from the vicinity of Logan on the north, nearly to Elk Creek on the south; also the tops of the high bench west of the Madison River" (1899:155). Dorr (1956) proposed a type section for the Madison Valley Formation in the northern part of the outcrop belt described by Douglass but included in its base rocks referred to the White River beds by Douglass. It is clear from Dorr's figures and description that the "Madison Valley beds" of Douglass include only the conglomerates containing fossil wood and intercalated sands and clays equivalent to the "fossil wood conglomerate" at the top of Dorr's section.

Douglass (1899, 1903) and Robinson (1963) report the occurrence of "mastodon" remains in high gravels in the Three Forks basin, but only one of these, a juvenile ramus, ascribed to the "Madison Valley beds" by Douglass (1903), has been described. Without a figure it is difficult to make comparisons, but the remains seem to be compatible with those of *Zygodolophodon*, perhaps like *Z. brevidens* of Cope. In any event, the remarks definitely put such Proboscidea in the fauna of the "Madison Valley beds." The remainder of the assemblage resembles that of the Deep River with some exceptions that may denote either environmental or temporal differences. Douglass reviewed the content of the fauna in 1903, and we follow this list and the work of later authors for the taxa contained in the Madison Valley beds. The rodent fauna is dominated by sciurids: *Spermophilus* (*Otospermophilus*) *primitivus* (Bryant 1945), the marmotines *Arctomyoides* and *Palaeoarctomys*, and the mylagaulid *Alphagaulus pristinus* are present. The carnivores *Aelurodon* cf. *asthenostylus*, *Miomustela madisonae*, and *Pliocyon ossifragus*; the horses *Archaeohippus minimus*, *Acritohippus isonesus*, and "Protohippus"; the rhino *Peraceras superciliosum*; the oreodonts *Brachycrus rusticus*

and *Ticholeptus zygomatiscus*; the camels *Miolabis montanus*, *Procamelus lacustris*, *Aepycamelus elrodi*, and *Aepycamelus madisonensis*; the moschid *Blastomeryx gemmifer*; the dromomerycids *Dromomeryx borealis*, *Bouromeryx americanus*, and *Rakomeryx kinseyi*; and the antilocaprids *Merycodus ?agilis* and *Paracosoryx furcatus* complete the known fauna.

Like the fauna of the Deep River Formation, the Madison Valley also seems to be of early Barstovian age principally on the biochrons of its wider-ranging horses, oreodonts, and camels as seen in the Great Plains. A strong local element is present in the rodents and dromomerycids.

Dorr (1956) and Sutton and Korth (1995) described the Anceney Local Fauna collected topographically high in the southernmost part of the Madison Valley Formation outcrop. They suggest that this site lies stratigraphically above the part of the section containing the classic fauna. This sample was obtained from a road cut and represents a coeval assemblage. It is strongly biased toward the microfauna, which includes the hedgehog *Brachyerix*; the soricids *Limnoecus* and *Angustidens* (its last occurrence); the talpid *Domninoidea*; the lagomorphs *Oreolagus nevadensis* and *Hypolagus*; the last promylagauline, *Gabreathia*; the mylagauline *Alphagaulus*; the castorid *Euroxenomys*; the sciurids *Tamias*, *Spermophilus* (*Otospermophilus*), and *Cynomysoides* (Korth 1996b); diverse geomyids *Mookomys*, *Peridiomys*, *Perognathus*, *Cupidinimus*, and *Phelosacomys*; the cricetid *Copemys nebrascensis*; the borophagine canids *Paratomarctus temerarius* and *Aelurodon* cf. *asthenostylus*; the mustelids *Leptarctus primus*, *Plionictis*, and *Martes*; the horses *Hypohippus* cf. *osborni* and *Acritohippus styodontus* (Evander 1996; "Merychippus cf. *M. intermontanus*" of Dorr 1956); the rhino *Peraceras superciliosum*; the moschid *Blastomeryx* cf. *elegans*; the camel *Aepycamelus elrodi*; and the antilocaprid *Merycodus* cf. *M. necatus*.

Even considering the differences in manner of collection, there are many comparable taxa between the Anceney Local Fauna and the classic fauna of the Madison Valley beds. Like the latter, the assemblage agrees best with assemblages of early Barstovian age, especially the co-occurrence of the mylagaulids *Gabreathia* and *Alphagaulus*, the heteromyids *Mookomys* and *Peridiomys*, the *Cupidinimus* species, and the ochotonid *Oreolagus*. Zoogeographically the relationship of this assemblage lies largely with the Great Plains.

South of the Three Forks Basin in the Yellowstone River valley, just north of the National Park, outcrops on the eastern side of the river called the Chalk Cliffs have yielded an important Barstovian faunal sequence from the Hepburn's Mesa Formation (Barnosky and Labar

1989). A magnetostratigraphy (Burbank and Barnosky 1990) for this sequence places it more securely in time than the assemblages discussed earlier. The fossiliferous sequence extends from chron C5Br into chron C5ADn. The biostratigraphy has been generalized to two zones, and the taxa (at the generic level) have been assigned to a *Peridiomys/Parahippus* zone older than 14.8 Ma and a *Lignimus/Tardontia* zone extending from 14.8 to about 14 Ma (the top of the exposed section). The lower zone contains *Alphagaulus* (*A. douglassi*; McKenna 1955), *Peridiomys*, *Oreolagus*, *Hypolagus*, “*Merychippus*,” “*Parahippus*” (including *Desmatippus*), *Blastomeryx*, *Dromomeryx*, and *Merycodus*, all of which also occur in the Madison Valley and Deep River faunas. The limited occurrence of the rodents and *Desmatippus* indicates an early Barstovian age, as does the calibration of this part of the sequence (older than 14.8 Ma).

The upper zone is defined by micromammals from collections at the top and bottom of the zone. Of particular significance is the occurrence of the aplodontid rodent *Tardontia*, the geomyids *Diprionomys* and *Lignimus*, the zapodid *Pseudadjidaumo*, and the sciurid *Spermophilus*. Some of these taxa characterize an assemblage zone (Lindsay 1972) low in the younger part of the Barstow Formation that dates at about 14.8 Ma, as at Hepburn’s Mesa. The survival of other taxa, namely *Mesoscalops* and *Alphagaulus*, into the lower part of the upper zone indicates that some otherwise early Barstovian forms lingered into the earliest late Barstovian in the northern Rocky Mountains. Although these microfaunas mainly have zoogeographic affinities with the Great Plains (*Lignimius*, *Schaubeumys*), there are some western taxa (*Tardontia*, *Pseudadjidaumo*, *Pseudotheridiomys*, and *Mojavemys*) as well (see discussion in Barnosky 1986a), in contrast with the Ancney Local Fauna, which lacks late Barstovian rodent taxa.

The Colter Formation crops out in Jackson Hole, Teton County, Wyoming, just south of Yellowstone Park and within 130 km of the Chalk Cliffs (Hepburn’s Mesa) north of the park. This unit is 1500 m thick and contains an intermittent faunal succession (Barnosky 1986a) that helps fill some gaps in our knowledge of mammalian history in the northern Rocky Mountains. Near the base of the formation tuffaceous sediments produce a small collection of taxa, the Emerald Lake Fauna, that includes the aplodontids *Niglarodon* cf. *blacki* and *Alwoodia cristabrevis*, the leporid *Archaeolagus emeraldensis*, and the oreodont *Promerycochoerus superbus* (“*Desmatochoerus leidyi*,” Barnosky 1986a:45; M. Stevens, pers. comm., 2002). Such an assemblage finds equivalents at the generic level in the early Arikareean (Ar2) of Montana (Fort Logan Fauna) and in the “Monroe Creek–Harrison” interval in

southwestern South Dakota. More than 50 m higher in the Crater Tuff–Breccia Member, two local faunas separated by about 250 m were grouped by Barnosky (1986a) as the East Pilgrim Assemblage. The lowest local fauna, East Pilgrim 11, is represented by a single taxon, the oreodont *Merychys arenarum*, whose range is confined to Upper Harrison strata (latest Arikareean) in the Great Plains. The upper local fauna, East Pilgrim 5, contains the equid *Parahippus tyleri* and oreodont *Merycochoerus magnus*, both of which occur in the Runningwater Formation of Nebraska (early Hemingfordian).

Nearly 400 m higher in the Colter Formation, in the Pilgrim Conglomerate Member, Barstovian mammals, the Cunningham Hill Fauna, appear and are represented by a more diverse array of taxa, especially microfauna. The greatest resemblance of the rodent fauna is with early late Barstovian faunas such as the *Lignimus/Tardontia* zone assemblage in the upper part of the Hepburn’s Mesa Formation (Burbank and Barnosky 1990) and specifically with the Norden Bridge and Railway Quarries (Crookston Bridge Member, Valentine Formation) of north-central Nebraska in the joint occurrence of *Oregonomys agrarius* (*sensu* Voorhies 1990a), *Copemys kelloggae*, *Cupidinimus* (as *C. whitlocki* Barnosky 1986b close to *C. nebrascensis*), and *Lignimus* (*L. transversus* Barnosky 1986a, closely related to Norden Bridge *L. cf. montis*, Voorhies 1990b). The few larger mammals do not materially contribute to the age determination other than the little worn hipparionine p4, which resembles *Cormohipparion quinni* (“*Neohipparion republicanus*” of Voorhies 1990a).

## NORTH GREAT PLAINS

**Saskatchewan (K)** Scattered outcrops are all that remain of the once continuous Tertiary blanket of the northern Great Plains in Canada. Profound preglacial erosion, occurring in the late Miocene and Pliocene, has removed most of this record. The Hand Hills of southeastern Alberta preserve limited ?Barstovian (Storer 1978) and late Hemingfordian (Burns and Young 1988) assemblages, and the former record is the youngest in the uplands of the region. Some upland surfaces in the Cypress Hills Plateau have been dated as late Miocene (Barendregt et al. 1997), and in that part of the region up to 150 m of erosion may have occurred since that time. The Wellsch Valley Local Fauna of Saskatchewan (Stalker and Churcher 1972; Harrington 1978) appears to correlate most reasonably as early Pleistocene (?early Irvingtonian), and as the earliest low-elevation assemblage in the area, it may provide a constraint for the end of downcutting.

Informative faunas occur in the southern plateaus of Saskatchewan, just north of the U.S.–Canada border in two areas: The Cypress Hills Plateau has a notable post–White River record (Skwara 1988; Storer 1993, 1996; Storer and Bryant 1993), and the Wood Mountain Plateau has an important late Barstovian fauna, the Wood Mountain Fauna (Storer 1975).

Fossil mammal remains have been collected from the sands and gravels of the Cypress Hills Formation outcrops for more than 100 years, but only recently have faunas markedly younger than Eocene (Chadronian) been acknowledged. Now the formation is known to contain faunas spanning the Arikareean to Hemingfordian interval from scattered localities in the southeastern part of the Cypress Hills Plateau (Storer 1993, 1996). The Kealey Springs Local Fauna, northwest of Anxiety Butte, has White River taxa such as *Centetodon*, *Eutypomys*, *Scottimus*, *Leptomeryx*, and *Elomeryx* associated with *Parvericius*, *Crucimys*, *Parallomys*, *Downsimus*, *Pseudotheridomys*, *Schizodontomys*, *Geringia*, *Archaeolagus*, *Promerycochoerus*, and *Sespia* (Storer 2002), suggesting correlation with lower Arikareean sites in western Nebraska. Recently Williams and Storer (1998) reviewed the cricetid rodents and *Pseudotheridomys* and correlated the Kealey Springs Local Fauna with assemblages from the Monroe Creek of the central Great Plains.

Some fragmentary remains from Anxiety Butte suggest that late Arikareean faunas may be present (Storer 1993), but the definitive presence of early Hemingfordian assemblages at that locality was demonstrated by Storer and Bryant (1993). Two sites on the southern edge of the butte, and at approximately equivalent elevations, yielded *Archaeohippus stenolophus* (Lambe 1905, including “*Mesohippus*” *planidens* Lambe 1905) associated with *Parahippus*, *Diceratherium*, *Arretotherium* cf. *fricki*, *Merycochoerus* cf. *proprius*, *Michenia*, *?Pseudoparablastomeryx*, and *Blastomeryx*. This suite of taxa is comparable with forms from the basal Ogallala strata (Runningwater Formation) of Nebraska. A larger Hemingfordian fauna, the Topham Local Fauna (Skwara 1988), is the youngest fauna obtained from the Cypress Hills Formation. It was found west of Anxiety Butte and includes *Oreolagus*, *Megalagus*, *Mylagaulodon*, *Protospermophilus*, *Miospermophilus*, *Leidyomys*, *Schaubeumys*, *Pseudotheridomys*, *Leptodontomys*, *Heliscomys*, *Proheteromys* (three species), *Parahippus*, *Hypohippus*, entelodonts, *Blastomeryx*, *Parablastomeryx*, dromomerycids, *Michenia*, and *Merychys*, all of which compare best with Great Plains early Hemingfordian faunas, especially those rich in micromammals (e.g., Quarry A; Wilson 1960).

The Wood Mountain gravels and sands cap the plateau in southernmost Saskatchewan near the international

boundary, 200 km southeast of the Cypress Hills Plateau. More than 30 m of these deposits yield a number of local faunas that are sufficiently alike compositionally to be grouped as the Wood Mountain Fauna (Storer 1975, 1978, 1993). This assemblage of more than 55 species includes the following taxa important in correlation: *Brachyerix*, *Megasmithus*, *Leptodontomys*, *Lignimus*, *Copemys*, *Anchitheriomys*, *Monosaulax*, *Hesperolagomys*, *Russellagus*, *Paratomarctus* (not *Tomarctus* of Storer 1975), *Aelurodon* cf. *ferox* (not “*A. cf. saevus*” of Storer 1975), *Amphicyon*, *Ursavus*, *Hemicyon*, *Zygodon proavus* (in Madden and Storer 1985), *Hypohippus osborni*, *Archaeohippus*, *Acritohippus* cf. *isonesus*, *Hipparion*, *Calippus* (“Equidae gen et sp. indet.,” Storer 1975), *Ticholeptus*, *Blastomeryx*, *Procranioceras* cf. *skinneri*, *Merycodus necatus*, *M. sabulonus*, and *Paracosoryx* cf. *alticornis*. As Storer (1975:126–127) pointed out, this fauna, though retaining early Barstovian elements, includes a number of taxa common to the earliest Valentine Formation assemblages (later or medial Barstovian) of Nebraska such as *Russellagus*, *Megasmithus*, *Lignimus*, *Paratomarctus*, *Aelurodon* cf. *ferox*, *Ursavus*, *Hemicyon*, *Hipparion*, *Calippus*, and *Procranioceras*. This suite of taxa includes genera that also characterize the Keota Fauna in the Pawnee Creek Formation of northeastern Colorado, where geochronologic evidence supports a pre-Valentine position for a comparable assemblage.

The only Canadian Miocene fauna outside the Great Plains is preserved in sediment filling the Houghton Astrobleme on Devon Island in the Canadian Arctic Islands above the Arctic Circle at 75°22'N latitude (Omar et al. 1987; Hickey et al. 1988; Whitlock and Dawson 1990). Fission-track dating of apatite grains (Omar et al. 1987) from the gneiss shocked by impact gives a  $22.4 \pm 1.4$  Ma date for that event and a maximum age for the fauna contained in the sediments infilling the crater. This undescribed assemblage is of great zoogeographic interest. It contains a heterosoricine shrew, cf. *Domnina*; an ochotonid, cf. *Desmatolagus*; a rhino; and an artiodactyl of “uncertain affinities,” none of which are exclusively North American groups.

**Nebraska and Wyoming (L)** Since the previous review (Tedford et al. 1987) a number of new studies have focused on the biochronology of parts of the Nebraskan record, particularly the early and late Arikareean, early Hemingfordian, and early late Barstovian, all of which provide new insights and information on the mammal ages classically typified by evidence from Nebraska. These studies have met the challenge of our previous review, which called for improvement of biostratigraphic knowl-

edge above that of just the composite faunal content of whole lithostratigraphic units.

Continued interest in the White River–Arikaree Group contact has focused on the lithostratigraphy of the bracketing units, their biostratigraphy and magnetostratigraphy, coupled with radioisotopic dating of the contained ash beds. These studies have led to a closer look at an important interval on the northern Great Plains, one broken by faunal turnover at and after the demise of the White River Chronofauna. This subject was explored in a preliminary way by Tedford et al. (1985), and the results were incorporated in Tedford et al. (1987). A more comprehensive treatment resulted in Tedford et al. (1996), which provides the substance for the following summary.

Tedford et al. (1996) abandoned the lithostratigraphy of Vondra (1963), with its downward extension of Darton's (1899) Gering Formation into White River Group strata. The latter rocks were subsequently recognized as the "Brown Siltstone beds" by Swinehart et al. (1985), who considered the upper member of the Brule Formation to represent an upward and gradational coarsening of the White River Group conformably above the Whitney Member. The Brown Siltstone beds contain many new taxa coexisting with forms typical of the White River Chronofauna, and these new taxa define and characterize the beginning of Arikareean time in the Great Plains. In a closer look at the stratigraphy of this interval, Tedford et al. (1996, figure 9) show that, of the immigrant taxa previously used to define the beginning of the Arikareean (*Ocajila*, *Talpinae*, *Plesiosminthus*, and "Allomys" = *Alwoodia*), only the zapodid *Plesiosminthus* occurs near the base of the Brown Siltstone. The other immigrant taxa have first stratigraphic occurrences as high as the basal Arikaree strata, where the local extinction of White River chronofaunal elements has been completed and a new fauna appears. This turnover episode is an important feature of the Great Plains faunal succession. We noted earlier that the other classic biostratigraphic record of this interval, that in the John Day Formation of Oregon, also shows a turnover, beginning slightly earlier, but the new fauna is characterized by persistent co-occurrence of some White River taxa into much younger deposits.

The new fauna that occurs in the lower Arikaree strata is biased by the dominance of aeolian facies, so the record is composed largely of oreodonts and a few horses, camels, and rhinos. Burrowing rodents were not common, and the beavers so conspicuous in younger Arikaree rocks were just beginning their diversification. Local fluvial settings provide most of our knowledge of mammalian diversity of this span. For these reasons it has been difficult to acquire a biostratigraphy for the Monroe

Creek and undifferentiated lower Arikaree rocks in Nebraska. The classic work by Peterson (1907) remains largely unchallenged. Hunt (1985) reinvestigated the stratigraphic allocation of Peterson's collection from the type area of the Monroe Creek Formation, north of Harrison, Nebraska, and reaffirmed the assignment of *Lepotauchenia*, *Desmatochoerus megalodon*, and *Diceratherium* to the lower part of the unit.

In an effort to provide a magnetostratigraphy for the lower part of the Arikaree Group, including Hatcher's (1902) Monroe Creek Formation, MacFadden and Hunt (1998) chose a section at Pants Butte, 8 km east of Monroe Creek Canyon, to avoid faulting. Only the lowermost 98 m of the Arikaree Group is exposed at Pants Butte (compared with 200–215 m at Monroe Creek Canyon). The Pants Butte section was correlated with chron C9r–C9n, probably continuing into chron C8r–C8n, and possibly to C7Ar or C7r, suggesting that the entire section at Pants Butte is older than 25 Ma (Berggren et al. 1995). The lack of faunal control at Pants Butte makes it uncertain where the "lower Monroe Creek Fauna" would occur in this section. Biochronologic data from the Platte Valley do not constrain the upper part of the ranges of the three lower Monroe Creek taxa.

An important insight into the nature of the small mammals of the lower Monroe Creek interval is the collections described by Korth (1992) from exposures along the north side of the Niobrara River near the mouth of McCann Canyon in eastern Cherry County, Nebraska. The McCann Canyon Local Fauna was taken from strata referred to the Harrison Formation that are unconformably overlain by the Valentine Formation (Skinner and Johnson 1984:229). However, as Korth concluded, the systematic relationships of this assemblage lie most closely with those described by J. R. Macdonald (1963, 1970) and L. J. Macdonald (1972) from strata referred to the Monroe Creek Formation in the Pine Ridge Reservation just north of the border in South Dakota. Taxa in common include *Alwoodia*, *Parallomys*, *Trilaccogaulus*, *Gregorymys*, *Pseudotheridomys*, *Plesiosminthus*, and *Archaeolagus*, all taxa that characterize the lower Arikaree Group deposits above the basal strata (Gering, Sharps formations) and thus younger than 28 Ma. (Tedford et al. 1996). In addition, the McCann Canyon Local Fauna contains such White River holdovers as *Domnina*, *Centetodon*, and the yoderomyine *Arikareomys* but lacks *Palaeolagus*. The presence of a low-crowned species of *Entoptychus* (*E. grandiplanus*) is the only well-represented occurrence of the genus in the Great Plains (see Korth 1992 for discussion).

Hunt (1985) also affirmed that the upper part of the Monroe Creek Formation of Hatcher (1902) contained

the type specimens of Peterson's (1907) taxa: *Promerycochoerus carrikeri*, *Phenacocoelus typus*, and *Euhapsis platyceps*. These taxa also occur in the overlying Harrison Formation, where they are joined by a diverse and clearly new fauna, indicating that a turnover event took place between the Monroe Creek and Harrison formations. MacFadden and Hunt (1998, figures 15 and 16) were aware that their magnetostratigraphic study of the Arikaree Group at Pine Ridge in northwest Nebraska sampled only the lower part of the Arikaree Group at Pants Butte and the uppermost part at Eagle Crag. Therefore their magnetostratigraphy contained a hiatus in the middle part of the Arikaree section. This lacuna can be filled, at least in part, by rocks of the Monroe Creek Canyon section. A magnetostratigraphic study of that section by Glynn Hayes and R. M. Hunt is in progress. However, one or more hiatuses appear to be widespread in the Arikaree Group of the northern Great Plains, so we lack biostratigraphic characterization of the important turnover event or events that occur somewhere in this span.

The Harrison is primarily an aeolian deposit with local fluvial facies near the base of the 50-m-thick unit. The latter facies produce a sufficiently varied mammal fauna to typify this interval biochronologically as previously described (Tedford et al. 1987). MacFadden and Hunt (1998) correlated the upper part of the Harrison Formation at Monroe Creek Canyon with the long chron C6n, although the lower part of the formation in the Niobrara River valley at Agate National Monument, limited by a 22.9-Ma K–Ar date on the Agate Ash (Izett and Obradovich 2001), ranges downward into earlier chrons. The top of the Harrison Formation is limited by the zircon fission-track date on the Eagle Crag Ash (Hunt et al. 1983), which occurs at the base of the disconformably overlying Upper Harrison beds. Few details of the biostratigraphy in the Harrison Formation have been recognized, but exposures in the Niobrara Valley near Agate and in the type area of the formation along the Pine Ridge in Sioux County, Nebraska, and adjacent Niobrara County, Wyoming, contain most of the fauna listed.

MacFadden and Hunt (1998) postulated a very short span for the volcanoclastic loess of the Upper Harrison beds (Peterson 1907, 1909), which lies on the Harrison in a deeply dissected surface. The Eagle Crag Ash broadly constrains the age of its base ( $19.2 \pm 0.5$  Ma). The 60-m-thick unit lies mostly in a reversed interval that was correlated with part of chron C5Er, its base at the top of chron C6n, a 400-k.y. interval spanning 19.2–18.8 Ma. Again, fluvial lithofacies such as that containing the Agate Bone Bed at the base of the dominantly aeolian unit contain most of the faunal typification of the lower part

of the interval. The upper part yields a fauna from aeolian volcanoclastic loess of more advanced character. This composite fauna shows a strong resemblance to that of the preceding Harrison assemblages but includes new elements that are phylogenetically exotic to the North American fauna and are therefore regarded as immigrants at the middle latitudes. Hunt (2002) recently proposed a replacement name, *Anderson Ranch Formation*, for the Upper Harrison beds, based on the type exposures in the Niobrara Canyon, Sioux County, Nebraska, where O. A. Peterson (1909) first defined the formation.

Rocks correlated with the Runningwater Formation fill a shallow paleovalley cut into the Upper Harrison in the region northeast of the Agate Post Office in Sioux County. These are epiclastic and volcanoclastic fluvial sandstones and loessic deposits about 30 m thick. The lower 20 m were found to lie in a mixed-polarity interval (MacFadden and Hunt 1998, figure 14) correlated with chron C5E and the base of C5Dr, the base at 18.8 Ma and the local top at 18.2 Ma, possibly extending to 18.0 Ma. The fauna from these deposits occurs as scattered specimens and has not been fully described. It represents a continuation of the chronofauna first seen in the Harrison Formation, but it differs from the Runningwater Local Fauna from the type section to the east of the Agate area in that *Syndyoceras* continues from the Harrison but not into the type Runningwater; *Merycochoerus magnus* replaces *M. matthewi* and is replaced by *M. proprius* in the younger part of the Runningwater.

In the Niobrara River area of northwestern Nebraska, the Runningwater Formation represents the basal unit of the principally epiclastic Ogallala Group. These fluvial strata cut across and may remove deposits of the underlying volcanoclastic Arikaree and White River groups. This episode marks a profound change in depositional style from broad sheetlike bodies with prominent aeolian facies to local valley filling, principally epiclastic fluvial units, lying in a dissected terrain. This regional shift in depositional style is not immediately reflected in biological change, but local stream channel deposits referred to the Box Butte and Sheep Creek formations in western Nebraska contain faunas that indicate that a significant event involving extinction and evolution ensued during early Ogallala time. This turnover initiated an early phase of one of the more enduring chronofaunas of the midcontinent late Cenozoic. This event cannot be precisely pinpointed in time but must have occurred rapidly between 17 and 17.5 Ma. Prominent among the many changes are the origin and initial diversification of equine horses (MacFadden and Hulbert 1988; Hulbert and MacFadden 1991); the diversification of mylagauline rodents (Korth

2000) and dromomerycids; antilocaprids, advanced camelids, and the rise of advanced borophagine canids (early cynarctines, aelurodontines, and borophagines; Wang et al. 1999); the first occurrence of immigrant rhinos (*Aphelops* and *Teleoceras*) and felids; and the loss of entelodonts, many oreodonts, early amphicyonids, most hesperocyonine canids (Wang 1994), and *Menoceras* (Tedford et al. 1987). Because of the manner in which the Hemingfordian was initially proposed, this natural biochronologic event is used only to divide the Hemingfordian into two parts, with the turnover defining the late Hemingfordian.

In recent years there has been increased interest in faunas that lie at the base of the Valentine Formation and equivalent units in Nebraska. They record a significant step in the evolution of the chronofauna that leads to assemblages of maximum diversity by the close of the medial Miocene. Voorhies (1990a, 1990b) and associates have described the local faunas of the closely associated Norden Bridge, Egelhoff, and Carrot Top quarries (Voorhies 1990b) from the basal part of the Valentine Formation (Cornell Dam Member of Skinner and Johnson 1984) in north-central Nebraska. The presence of gomphotheriid proboscideans in these sites records a significant expansion of range of these animals into the midcontinent; this appearance is used to define the beginning of the late (or medial, Voorhies 1990a) Barstovian time. However, in the case of the Nebraska sequence, the profound disconformity beneath the Valentine strata truncates the actual first occurrence of these proboscideans in that area. We must turn to evidence from the more complete stratigraphic sequence in the Pawnee Creek Formation, northeastern Colorado, for the first local appearance of the gomphotheriids. The Hurlbut Ash near the base of the Valentine Formation has been fission-track dated  $13.6 \pm 1.3$  Ma (Boellstorff and Skinner 1977), and the date is now confirmed at  $13.55 \pm 0.09$  Ma by  $^{40}\text{K}$ - $^{40}\text{Ar}$  dating of glass shards (Swisher 1992) and at  $13.5 \pm 0.1$  Ma by identification of the element composition of the Hurlbut Ash with its source in an eruptive center in northern Nevada (Perkins and Nash 2002, table DR2).

Voorhies's review (1990b) shows that the faunas of the Cornell Dam Member record an early phase of organization of the Miocene Chronofauna of the Great Plains. Cornell Dam representatives of long-ranging genera typical of this chronofauna in addition to Proboscidea are the beaver *Eucastor*; such borophagine dogs as *Paratomarctus*, *Aelurodon*, *Carpocyon*, and *Cynarctus* (Wang et al. 1999); the canine *Leptocyon*; the mustelid *Leptarctus*; such horses as *Cormohipparion* ("*Neohipparion*"), *Protohippus*, and *Calippus*; the rhinos *Aphelops*,

*Teleoceras*, and *Peraceras*; the oreodont *Ustatochoerus*; camels such as *Aepyamelus*, *Procamelus*, and *Protolabis*; and diverse merycodontine antilocaprids. This early fauna contains many taxa such as the beavers *Anchitheriomys* and *Monosaulax*, the aplodontid *Allomys*, the horses *Anchitherium* and *Archaeohippus*, the protoceratid *Prosynthetoceras*, the dromomerycid *Dromomeryx*, the merycodonts *Ramoceros* and *Submeryceros*, and the oreodont *Ticholeptus* that reach the limit of their geologic ranges at this time, giving rise to an extinction event that continues into the faunas of the overlying Crookston Bridge and Devil's Gulch members.

At the family level such venerable groups as aplodontids, chalicotheres, and leptomerycids make their final appearances in the Great Plains before the Burge Member. This extinction, along with continued evolution (much of it anagenetic) in surviving groups, but little immigration, shapes the faunas of the succeeding Burge and Ash Hollow strata. Unfortunately we have little evidence for the timing of these events beyond the knowledge of the approximate initiation of Valentine sedimentation (13.6 Ma) and an  $^{40}\text{Ar}/^{39}\text{Ar}$  age for vitric ash in the lower part of the overlying Ash Hollow Formation ( $12.18 \pm 0.12$  Ma, Swallow Ash; Swisher 1992). Because there are regional disconformities at the base of the Burge Member of the Valentine and at the base of the Ash Hollow, these data imply that the depositional span of the Valentine must be less than 1 m.y.

New  $^{40}\text{Ar}/^{39}\text{Ar}$  dates obtained from glass shards in the Swallow and Davis ashes allow calibration of the lower part of the Ash Hollow Formation of north-central Nebraska (Swisher 1992) to 11.5–12.2 Ma. Faunas from the upper part of this unit indicate that it must reach nearly to the Clarendonian–Hemphillian boundary. Younger Ash Hollow faunas (including the Feltz Ranch, Oshkosh, and Uptegrove faunas in the type area of that unit) are compositionally medial and later Hemphillian assemblages, as supported by bracketing dates. The Santee Local Fauna, the youngest Nebraska Hemphillian fauna, occurs just below a vitric tuff (Santee Ash) fission-track dated to  $5.0 \pm 0.2$  Ma (Boellstorff 1978).

An important record of later Hemingfordian faunas is contained in the Split Rock Formation (as restricted by Munthe 1979b) of central Wyoming. These rocks were deposited in the Granite Mountains Basin formed by mid-Tertiary subsidence and folding (Love 1970) along the Split Rock syncline. Older faunas occur along the northern flank of the basin that yield scattered remains including *Oxydactylus* cf. *longipes*, *Paracynarctus kelloggi*, and *Merychyus* (Munthe 1979b) of early Hemingfordian age. A concentration of remains scattered through 130 m

of the section exposed in Fremont and adjoining Natrona counties near their common junction with Carbon County seems to represent a taxonomically unified assemblage called the Split Rock Fauna here ("local fauna" in Munthe 1979b, 1988). An ash in the faunal sequence produced a radiometric date of  $17.4 \pm 0.08$  Ma (Izett and Obradovich 2001). The fauna includes a diverse lower vertebrate assemblage and 45 mammalian taxa, among which are the erinaceid insectivores *Brachyerix* and *Parvericus*; the mole *Mesoscalops*; the ochotonids *Desmatolagus schizopetrus* and *Oreolagus nebrascensis*; the mylagaulids *Galbreathia* and *Alphagaulus*; the squirrels *Protospermophilus* and *Miospermophilus*; the geomyoids *Harrymys*, *Schizodontomys*, *Proheteromys*, and *Peridiomys*; the zopodid *Plesiosminthus*; the borophagine canids *Protomarctus*, *Metatomarctus*, *Paracynarctus*, and *Cynarctoides*; the equids *Hypohippus* and "*Merychippus*" *primus*; the dome-headed chalicotheres *Tylocephalonyx*; the oreodont *Brachycrus*; the camels *Protolabis* and *Blickomylylus*; the dromomerycids *Bouromeryx* and possibly *Barbouromeryx*; and the antilocaprids *Merycodus* and *Paracosoryx*. This assemblage clearly is a late Hemingfordian fauna with strong affinities to assemblages in adjacent Nebraska. Several genera do not range into the Barstovian (*Protomarctus*, *Metatomarctus*, *Tylocephalonyx*, and *Blickomylylus*), and others do not occur in earlier Hemingfordian assemblages (*Galbreathia*, *Alphagaulus*, *Hypohippus*, "*Merychippus*" *primus*, and *Brachycrus*). In the northwestern Albuquerque Basin *Blickomylylus* occurs in the red mudstones of Canyada Pilares Member of the Zia Formation in strata as young as the base of chron C5Cn (Tedford and Barghoorn 1999), calibrated at 16.7 Ma, a date comparable with that determined from the ash in the Split Rock faunal span.

**Southwestern South Dakota (M)** A recent review (Tedford et al. 1996) of the Whitneyan–Arikareean transition in southwestern South Dakota gives further substance to the reconstruction proposed in Tedford et al. (1987) for the Sharps Formation. Lithologically this unit lies astride the boundary between the White River and Arikaree groups. The lower part is lithologically identical to the Brown Siltstone beds of the Brule Formation in adjacent Nebraska, including the correlative Rockyford and Nonpareil ash beds. Like the Arikaree regionally, the disconformably overlying upper part of the unit has a basal channel filling sandstone that grades upward into massive silts and fine sands of typical Arikaree lithologies. The Wounded Knee–Sharps Fauna of Macdonald (1963, 1970), from the upper part of the Sharps Formation, contains a mixture of relictual White River

taxa and new records, including the introduction of taxa typical of the Arikaree.

Our lack of knowledge of the biostratigraphy of the younger levels in the Arikaree Group of the Great Plains has already been discussed in connection with the work of Hunt (1985) and MacFadden and Hunt (1998) in the Monroe Creek–Harrison region of northwestern Nebraska. When traced northeastward toward South Dakota, the Monroe Creek and Harrison formations are truncated by the disconformity at the base of the Anderson Ranch Formation (former Upper Harrison beds; Hunt 2002) so that around Chadron, Nebraska, the Pine Ridge Escarpment shows the Anderson Ranch resting directly on lower Arikaree rocks that contains a Wounded Knee–Sharps like fauna (Tedford et al. 1996). Northeast at the Beaver Wall in Nebraska and the adjacent Slim Buttes in South Dakota the intervening strata reemerge. At Slim Buttes, just east of the White Clay Fault Zone, the lithic sequence seems similar to that in the classic "Rosebud" outcrops along Porcupine Creek, further east in South Dakota. In the latter area, the nodular, massive to thick-bedded, silty sandstones of the upper part of the Sharps Formation are gradationally overlain by nodular, massive, pink sandy siltstones that form vertical outcrops (usually called "Lower Rosebud" or "Monroe Creek"; Harksen 1969; Macdonald 1963). At Bear Creek Bluff, at the entrance of Porcupine Creek Canyon, these rocks contain an upward extension of the Sharps oreodonts *Mesoreodon* *minor*, *Desmatochoerus megalodon*, *Megoreodon grandis*, and *Leptauchenia* (*Sespia* is absent). New taxa appearing in the "Monroe Creek" Formation in southwestern South Dakota include *Amphechinus*, *Archaeolagus*, *Gripholagomys*, *Desmatolagus*, *Fossorcaptor*, *Alwoodia*, *Pleurolicus*, *Gregorymys*, *Pseudotheridomys*, *Mammacyon*, *Nexuotapirus*, and *Oreodontoides* along with immigrant carnivores *Promartes* and *Plesictis* (J. R. Macdonald 1963, 1970; L. J. Macdonald 1972; Tedford et al. 1996). These strata record the last Great Plains occurrence of some White River taxa: *Proscalops*, *Geolabis*, *Palaeolagus*, *Palaeocastor*, *Eutypomys*, *Heliscomys*, and possibly *Nimravus* and the leptauchinine oreodonts.

The Turtle Butte Formation, which crops out 163 km southeast of the classic Pine Ridge Reservation region, contains a fauna of large mammals, the Wewela Fauna of Skinner et al. (1968), similar in character to those found in the lower Arikaree (faunas of the "Lower Rosebud" or "Monroe Creek" formations) of western South Dakota and adjacent Nebraska. The following forms (taxonomy revised) are present: the canine *Leptocyon*; the hesperocyonine *Enhydrocyon pahinsintewakpa* (Wang 1994); a large nimravid carnivore, possibly *Hoplophoneus occiden-*



*talis*; the horse *Miohippus equinanus*; the giant entelodont *Archaeotherium trippensis* (cf. “*Dinohyus*” *minimus*; Schlaikjer 1935); the camel *Miotylopus*; and the oreodonts *Megoreodon grandis* and *Paramerychius* cf. *harrisonensis* (Stevens, pers. comm., 2002).

In the Porcupine Creek outcrops, massive buff to gray sandstones with slabby cemented zones (often referred to the “Harrison” Formation; Harksen 1969; Macdonald 1963, 1970) conformably overlie the “Monroe Creek” siltstones and introduce a different oreodont fauna with *Promerycochoerus superbus*, *Promerycochoerus* cf. *carrickeri*, and ?*Hypsiops latidens*; *Oreodontoides oregonensis* and *Desmatochoerus megalodon* continue, but the leptachenines appear to be absent; neither *Paramerychius* nor *Merychius* is recorded. Additional first appearances at these levels include *Pseudopalaecastor*, *Parahydrocyon*, *Parahippus*, and *Arretotherium*, but in general the “Monroe Creek” and “Harrison” formation mammal faunas retain several genera (and even species) in common, such as *Fossorcastor*, *Gregorymys*, *Enhydrocyon*, *Miohippus*, *Diceratherium*, *Desmatochoerus*, *Oreodontoides*, and *Nanotragus*. At Slim Buttes, strata with such a fauna are strongly disconformable on the lower Arikaree. In turn these rocks are disconformably overlain by the “Upper Rosebud” (= Anderson Ranch Formation) strata in the upper part of the Porcupine Creek drainage. This is a frustratingly limited view of a critical faunal transition, biased by faunal facies and the imposition of disconformities with ostensibly large hiatuses. The post-Sharps faunal succession resembles that in the type Monroe Creek described by Peterson (1907) and reviewed by Hunt (1985).

**Northeastern Colorado (N)** The lithostratigraphy and biostratigraphy of the Pawnee Creek Formation and younger Ogallala rocks were presented in Tedford et al. (1987). They accepted the lectotype of the Pawnee Creek Formation proposed by Galbreath (1953), removing only the capping gravels of the type section as undivided upper Ogallala beds. The Pawnee Creek Formation, as emended, fills the course of a single river valley that follows a broadly sinuous path through the Pawnee Buttes area, 7–20 km east of Grover, Weld County, Colorado (Tedford 1999, figures 16 and 19). Dated ash beds (Swisher in Tedford 1999) in this unit show that the Pawnee Creek Formation spans the interval between 14 and 14.5 Ma. Temporally and biochronologically it lies between the older Olcott (15–16 Ma) and younger (less than 14 Ma) Valentine formations of adjacent Nebraska and their abundant faunas and thus fills an important hiatus in the classic Nebraska sequence. Significantly, the capping

gravels of the Pawnee Creek type section are part of the overlying upper Ogallala rocks containing local faunas (Kennesaw, Vim-Peetz, and Sand Canyon; Galbreath 1953) that readily correlate with those from the lower Valentine of Nebraska. The deposition of the Pawnee Creek Formation, as now restricted, took sufficient time so that there are some differences in the local range zones of certain taxa. Galbreath (1953) used the term *Eubanks Fauna* for the assemblage from the lower part of the section. This fauna is a mixture of survivors from the early Barstovian (e.g., *Microtomarctus*, *Leptarctus primus*, *Desmatippus*, *Merychippus insignis*, *Ticholeptus*, and *Brachycrus*), derived members of early Barstovian clades (e.g., *Hypohippus osborni* vs. *Hypohippus pertinax*), and new genera that assume an important role in the gathering Miocene Chronofauna (i.e., *Aelurodon*, *Calippus*, *Protohippus*, *Megahippus*, *Ustatochoerus*, and *Cranioceras*). The fauna of the upper part of the Pawnee Creek Formation, the Keota Fauna of Tedford (1999), is typified by the local fauna of the Horse and Mastodon Quarry. This assemblage is marked by the first appearance of Proboscidea, both mammutids (*Zygodolophodon*) and gomphotheriids (*Gomphotherium*) just above an ash whose glass  $^{40}\text{Ar}/^{39}\text{Ar}$  age is  $14.4 \pm 0.02$  Ma (Swisher in Tedford 1999). The accompanying fauna differs little from that of the Eubanks Fauna except for the appearance of *Carpocyon compressus* and *Cormohipparion paniense* and the loss of the holdover taxa listed earlier. The continued coexistence of *Ticholeptus* with *Ustatochoerus medius*, *Dromomeryx pawniensis* with *Procranioceras pawniensis*, and *Paracosoryx* with *Cosoryx* gives the Keota Fauna a significant biostratigraphic signature compared with assemblages from the lower part of the Valentine Formation. A hiatus of nearly 1 m.y. may separate the Keota and Norden Bridge faunas judging from ash dates now available.

#### **Texas Panhandle, Adjacent Oklahoma and Kansas (O)**

The importance of the southern Great Plains region in establishing a biostratigraphic sequence of faunas was emphasized by Tedford et al. (1987:175) and by Schultz’s (1990) excellent review. This sequence has gained in value in recent years with the addition of microfaunal elements to the better-known large mammal assemblages through the work of the late Walter Dalquest and his colleagues.

A case in point are the Beaver County, Oklahoma, Clarendonian sites, known since the nineteenth century. Hesse (1936) gave the first review of the Beaver County sites, adding new information from the collections of the universities of Kansas and California. Three sites in the Laverne Formation produce locally abundant fossil mammals. These sites are grouped as the Beaver Fauna

because they are identical taxonomically. They are spread through many meters stratigraphically and distributed from near the town of Beaver east almost to Laverne (Harper County), a distance of some 30 km. The remains occur in locally deformed fine sands and interbedded diatomites that lie unconformably beneath younger Ogallala strata (containing the *Biorbia* fossil seed assemblage). The Whisenhunt Quarry Local Fauna (Dalquest et al. 1996) supplies most of the microfaunal material; the classic Cragin Quarry and Beaver Quarry local faunas (Hesse 1936) supply the macromammals. Biochronologically important elements of this fauna include the hedgehog *Untermannerix*, the mole *Domninoidea*, the endemic ochotonid *Oklahomalagus*, an early leporine *Pronotolagus*, a late occurrence of the marmotine *Miospermophilus*, the beaver *Eucastor planus*, the zapodid *Macrognothomys*, and the cricetids *Tregomys* and *Copemys lindsayi*. Large mammals include the borophagine canid *Aelurodon taxoides* and the horses *Calippus* (*Grammohippus*) *martini*, *Pseudhipparion gratum*, *Hipparion* cf. *tehonense*, and *Cormohipparion occidentale*, whose overlapping range zones indicate a medial Clarendonian age. The Wakeeny Local Fauna from western Kansas (R. L. Wilson 1968) represents a closely similar fauna and ecology with beavers, cricetids, and rare geomyoids.

Thirteen kilometers north of the Beaver local faunal site on the south bank of the Cimarron River, the upper part of the Ogallala Formation lies in a solution collapse basin developed in the Permian gypsum. These sediments are capped by a caliche and contain a late Hemphillian fauna, the Buis Ranch Local Fauna (Hibbard 1954, 1963; Stevens 1966). This assemblage includes the following taxa: the talpid *Hesperoscalops*; the ground squirrel *Spermophilus* (*Buiscitellus*) *dotti*; the geomyoids *Perognathus*, *Prodipodomys*, and *Pliogeomys*; the skunk *Buisnictis* and fox *Vulpes*; the horse *Nannippus aztecus*; the rhino *Teleoceras*; and the llama *Hemiauchenia*.

Microfaunas associated with the well-known early and late Hemphillian sites in the Texas Panhandle fill the gap between the Oklahoma assemblages discussed earlier. Dalquest and Patrick (1989) described a microfauna from Sebitts Ranch Site 24B, from which most of the early Hemphillian Higgins Local Fauna was collected. It curiously lacks cricetid rodents but yields the eomyid *Kansasimys dubius*, the geomyoids *Perognathus* and *Pliosacomys*, a ground squirrel *Spermophilus*, and the archaeolagine rabbit *Hypolagus vetus*. In contrast, the Coffee Ranch Quarry site produced a larger fauna of late Hemphillian age (Dalquest 1983; Dalquest and Patrick 1989) with *Hypolagus* cf. *vetus*; the eomyid *Comancheomys*; the ground squirrel *Spermophilus*; diverse ge-

omyoids *Progeomys*, *Perognathus*, *Cupidinimus*, and *Prodipodomys*; and the cricetids *Calomys* (*Bensonomys*), *Prosigmodon*, *Peromyscus*, and *Neotoma* (*Paraneotoma*). This fauna resembles the Buis Ranch and Saw Rock Canyon local faunas at the generic level but not at the specific. It also lacks the primitive arvicoline present at Saw Rock Canyon.

## SOUTHERN GREAT BASIN

**Rio Grande Rift, New Mexico (P)** The Española and adjacent Albuquerque basins contain important superposed sequences of faunas of late Oligocene through Miocene age. Cope discovered the faunas of the "Santa Fe Marls," but the decades of collecting and stratigraphic study by the Frick Laboratory have proven the great value of these areas to biochronology. Galusha and Blick (1971) and Galusha (1966) laid out the lithostratigraphy for both basins. Barghoorn (1981) provided a magnetostratigraphy for both basins (Tedford and Barghoorn 1993, 1999), which was constrained by ash dating. The fauna was initially described by Cope (1874, 1875), and the newer collections were informally reviewed by Tedford (1981) and Tedford and Barghoorn (1997). The faunal sequence in both basins is partially overlapping; that in the Española Basin is the longest.

The oldest assemblages are best represented in the northern Albuquerque Basin at Standing Rock Quarry in the base of the Piedra Parada Member of the Zia Formation. This local fauna includes the leporid *Archaeolagus*; the heteromyids *Ziamys* and *Proheteromys*; the carnivores *Cynarctoides*, *Promartes*, *Daphoenodon*, and *Cephalogale*; and the camel *Stenomylus*. These taxa are comparable with assemblages from the Anderson Ranch Formation (formerly Upper Harrison beds; Hunt 2002) of Nebraska. The Piedra Parada Member is a volcanoclastic dune sand, including interdune pond and fluvial facies in its lower part (Gawne 1981). These deposits yield additional taxa from the northwestern part of the basin that are approximately contemporaneous with those from the Standing Rock Quarry (*Diceratherium*, *Merychys* cf. *arenarum*, *Oxydactylus*, and *Michenia* cf. *agatense*) that confirm the correlation of these faunas with those from the Upper Harrison beds of Nebraska.

The Chamisa Mesa Member of the Zia Formation lies conformably above the Piedra Parada dunefield and represents a transitional environment from aeolian dominated to mixed aeolian-fluvial deposits (Gawne 1981). In its type area in the northernmost Albuquerque Basin there is a sequence of superposed fossil concentrations ("quarries") that appear to span most of the Hemingfor-

dian. These sites are dominated by the stenomyline camel *Blickomylus*. In the lowest site (Blick Quarry) it occurs with *Stenomylus*, *Archaeolagus*, *Oreolagus*, *Pleurolicus*, *Cynarctoides*, and *Protomarctus optatus*. At an intermediate stratigraphic level (Jeep Quarry) *Blickomylus* occurs with *Alphagaulus*, *Archaeolagus*, *Cynarctoides*, *Desmocyon thomsoni*, *Metatomarctus canavus*, *Promartes*, *Amphicyon*, *Menoceras*, *Michenia*, *Protolabis*, *Nothotylopus*, and *Merycodus*. Five meters higher in the conformable section at Mesa Prospect, *Parahippus* cf. *tyleri* and “*Merychippus*” sp. (cement bearing, hypsodont cheek teeth) join the artiodactyl fauna seen at Jeep Quarry. The endemic rodents, very rare oreodonts, diverse camelids, and the peculiarly limited distribution of a very derived stenomyline places a distinctive zoogeographic stamp on a faunal sequence that otherwise resembles that of the Runningwater to Sheep Creek interval in Nebraska. Magnetostratigraphic work in correlative rocks along the northwestern side of the Albuquerque Basin (Tedford and Barghoorn 1999) indicates that the Zia Formation (including the local Canyada Pilares Member of Gawne 1981 at the top) extends to chron C5Cn close to 16.0 Ma. Unfortunately there are few fossils other than the camels *Michenia* and *Protolabis* cf. *barstowensis* at the top of the section in that area.

A regional disconformity occupies the span of the early Barstovian in the Albuquerque Basin, but this interval and the late Hemingfordian are represented in the Española Basin. In the latter basin, the upper part of the Nambé Member of the Tesuque Formation has a more diverse fauna than the Zia Formation, including the horses *Archaeohippus*, *Acritohippus isonesus*, and *Protohippus*; the rhino *Aphelops*; the camels *Aepyamelus*, *Nothotylopus*, *Michenia*, and *Paramiolabis* (but no stenomylines); the borophagines *Cynarctoides*, *Paracynarctus* (also present in the Canyada Pilares Member, Zia Formation), and *Microtomarctus conferta*; and the immigrant felid *Pseudaelurus*. This fauna comes from rocks that lie in the normal interval at the top of chron C5C and seems more derived than the youngest well-represented assemblage from the Zia Formation.

The rich fauna of the overlying Skull Ridge Member of the Tesuque Formation was unknown to Cope but was discovered and well sampled by the Frick Laboratory. Surprisingly, a very derived stenomyline, *Rakomylus*, reappears but is nevertheless a very rare taxon. The camel fauna of this unit is diverse and includes species of *Miolabis*, *Paramiolabis*, *Nothotylopus*, *Australocamelus*, *Aepyamelus*, *Protolabis*, and *Michenia*. The horses are rare but include “*Merychippus*” cf. *intermontanus*, *Acritohippus isonesus*, *Anchitherium*, and *Hypohippus*; *Megahippus*

makes its earliest occurrence. The rhinos are *Teleoceras* and *Peraceras*. The arctoids *Hemicyon* and *Amphicyon* cf. *ingens* are present, and the borophagine canid fauna closely resembles that of the Lower Snake Creek Fauna of western Nebraska, with *Cynarctoides*, *Paracynarctus*, *Psalidocyon*, *Microtomarctus*, *Tomarctus hippophaga*, and *Tomarctus brevirostris* (Wang et al. 1999) and the last hesperocyonine, *Osbornodon*. The oreodonts *Merychys* and *Brachycrus*, the antilocaprid *Merycodus*, and the dromomerycid *Rakomeryx* complete the known fauna. Magnetostratigraphic work (Barghoorn 1981; Tedford and Barghoorn 1993) and ash dating show that the Skull Ridge Member spans nearly all of chron C5B (16.0–14.9 Ma; Berggren et al. 1995). Sanidine  $^{40}\text{Ar}/^{39}\text{Ar}$  dates of  $15.42 \pm 0.06$  Ma (McIntosh and Quade 1995) and  $15.3 \pm 0.05$  Ma (Izett and Obradovich 2001) were obtained on the White Ash No. 4 near the top of the Skull Ridge Member, in agreement with the magnetostratigraphic results.

Compared with the approximately contemporaneous assemblages from the northern Great Plains (Sheep Creek through Lower Snake Creek interval), the Nambe and Skull Ridge faunas have greater numbers and diversity of camels and fewer horses, antilocaprids, moschids, and dromomerycids. The canid and amphicyonid fauna is largely the same, but oreodonts are rare in New Mexico during this span. These compositional differences probably reflect ecological contrasts, resulting in some endemism (*Rakomylus*), but fundamentally the New Mexican faunas represent zoogeographic outliers of those of the Great Plains. Farther west in the Great Basin faunal diversity declines markedly during this interval.

In the Española Basin, as in the Albuquerque Basin, a regional hiatus breaks stratigraphic continuity beneath the Pojoaque Member of the Tesuque Formation. This hiatus removes the record of most of chron C5AD so that the Pojoaque Member deposition begins in late C5ADn at about 14.3 Ma and extends into chron C5An2 at approximately 12 Ma. This was the unit that yielded nearly all of Cope’s collection of 1874. The most fossiliferous interval is the lower 120 m of the unit, which produced the following taxa important in correlation: *Monosaulax pantsus* (Cope 1874, now clearly differentiated from *Eucastor tortus*; Korth 1999c), *Copemys loxodon* (Cope 1874), “*Mylagaulus*,” *Hesperolagomys*, *Russellagus*, *Panolax santae-fidei* (Cope 1874), and *Hypolagus*; the borophagine canids (Wang et al. 1999) *Microtomarctus*, *Aelurodon* (*Strobodon*) *stirtoni*, *A. ferox* (= *A. wheelerianus*; Cope 1877), *Paratomarctus*, *Carpocyon*, and the canine *Leptocyon*; *Hemicyon ursinus* (Cope 1875); the horses *Hipparion sanfondensis* (Frick 1933; *Hippotherium speciosum* of Cope 1877), *Neohipparion coloradense* (MacFadden 1984), *Pro-*

*tohippus*, *Dinohippus*, and *Pliohippus*, associated with the large and primitive *Merychippus calamarius* (Cope 1875) and the anchitherines *Hypohippus* and *Megahippus* cf. *mckennai*; the rhinos *Teleoceras*, *Aphelops*, and *Peraceras*. The camel *Procamelus* makes its first appearance, and the genera *Aepycamelus*, *Paramiolabis*, *Miolabis*, *Nothotylopus*, *Protolabis*, *Michenia*, and *Australocamelus* continue from the Skull Ridge. The merycodonts are more diverse, including species of *Meryceros*, *Cosoryx*, and *Ramoceros*. The moschids, tragulids, a surviving leptomerycid, and *Cranioceras teres* (Cope 1874) are present; *Ustatochoerus medius* is the only oreodont. Significantly, gomphotheriid proboscideans are common, with all the morphological diversity grouped into a single taxon, *Gomphotherium productum* (Cope 1874), by Tobien (1972, 1973). This diverse fauna has a strong systematic relationship with faunas of the Valentine Formation of Nebraska and other early late Barstovian faunas of the northern Great Plains. Most of the Pojoaque taxa co-occur in the chron C5AC and C5AB interval (Barghoorn 1981; Tedford and Barghoorn 1993), 14.3 to 13.3 Ma, and biotite from a thin white ash about 170 m above the base of the Pojoaque Member gave an  $^{40}\text{Ar}/^{39}\text{Ar}$  date of  $13.7 \pm 0.18$  Ma (Izett and Obradovich 2001, mean of four determinations). The fauna of this interval is thus temporally correlative with the basal Valentine Norden Bridge to Railway Quarry local faunas in agreement with the age of the Hurlbut Ash,  $13.6 \pm 0.2$  Ma (Swisher 1992).

Poorly fossiliferous, structurally disturbed, and unfavorable lithofacies (dune sands, Ojo Caliente Member, Tesuque Formation) limit the biostratigraphy of the Pojoaque to its lower levels. The record reemerges at the base of the overlying Chamita Formation, where Round Mountain Quarry yields an early Clarendonian assemblage: *Epicyon saevus* has its first local occurrence with *Aelurodon taxoides* (Wang et al. 1999); the horses include *Megahippus* cf. *matthewi*, *Pliohippus* cf. *pernix*, and *Pseudhipparion gratum*; the merycodonts *Cosoryx* and *Paramoceros*, the gelocid *Pseudoceras*, and camel *Hemiauchenia* are represented at the quarry; and correlative sediments in the vicinity add *Epicyon haydeni*, *Dinohippus*, *Cormohipparion occidentale* (MacFadden 1984), *Aphelops jemezianus* (Cope 1875), *Teleoceras*, *Peraceros*, *Gomphotherium productum*, *Megatylopus*, *Ustatochoerus major*, *Longirostromeryx*, *Blastomeryx*, and *Plioceros*. These taxa also occur together in the classic Clarendonian sites in the Great Plains. A basalt flow higher in the local section gives a whole rock date of  $9.6 \pm 0.2$  Ma (Aldrich and Dethier 1990) as minimum age for the Round Mountain and stratigraphically associated fauna.

In the type section of the Chamita Formation, MacFadden's (1977) magnetostratigraphy for the incomplete 500-m-thick section extends from the top of chron C5n to the base of chron C3r, 9.7–5.8 Ma. Scattered mammal remains occur in the lower part of the section including the sloth *Pliometanastes*, the canid *Epicyon* cf. *haydeni*, the camel *Megatylopus*, and the antilocaprine *Plioceros* sp.; the first two taxa co-occur in the Great Plains only in strata of early Hemphillian age, 8–7 Ma. Thus the exposed base of the Chamita type section probably overlaps the top of the referred exposures in the vicinity of Round Mountain Quarry. A late Hemphillian fauna occurs in the upper tuffaceous zone of the type Chamita at the approximately contemporaneous San Juan and Rak Camel quarries. The composite assemblage would come from "chron 6N" (Lindsay et al. 1987, now chron C3Bn, about 7 Ma, Berggren et al. 1995), early in the late Hemphillian as calibrated here. Sanidine from the upper tuffaceous zone at the level of the quarries yielded a  $^{40}\text{Ar}/^{39}\text{Ar}$  date of  $6.9 \pm 0.03$  Ma (Izett and Obradovich 2001). The fauna includes *Dipoides williamsi*, *Eucyon davisii*, *Plesiogulo*, *Astrohippus ansae*, *Dinohippus interpolatus*, *Hemiauchenia*, and *Megatylopus matthewi* and has a clear taxic relationship to broadly comparable assemblages from the Hemphill beds of the Texas Panhandle.

**Yepómera, Northern Chihuahua, Mexico (Q)** Flat-lying, fine-grained sediments exposed in the valley of the Rio Papagochic, 200 km west of Chihuahua City, yield a very late Hemphillian (Yepómera Fauna) through earliest Blancan (Concha Fauna) succession calibrated magnetostratigraphically (Lundelius et al. 1987). The composition of the Yepómera Fauna is similar to that of latest Hemphillian faunas of the Texas Panhandle and adjacent Oklahoma (Axtel and Buis Ranch local faunas) that are contained in rocks superposed on the Hemphill beds carrying the well-known Coffee Ranch Local Fauna. The Yepómera Fauna (composite of several sites; Lindsay and Jacobs 1985) includes *Notolagus*<sup>\*</sup>, *Paenemarmota*<sup>\*</sup>, *Prodipodomys*, *Pliogeomys*<sup>\*</sup>, *Copemys*<sup>†</sup>, *Calomys* (*Bensonomys*), *Baiomys*, *Prosigmodon*<sup>\*</sup>, *Eucyon davisii*<sup>†</sup>, *Vulpes stenognathus*<sup>†</sup>, *Agriotherium*, *Taxidea*<sup>\*</sup>, *Machairodus*<sup>†</sup>, *Astrohippus stocki*<sup>†</sup>, *Dinohippus mexicanus*<sup>†</sup>, *Neohipparion eurystyle*<sup>†</sup> (MacFadden 1984), *Nannipus aztecus*<sup>†</sup>, *Teleoceras*<sup>†</sup>, *Prosthennops*, *Megatylopus*, *Hemiauchenia*, and *Hexobelomeryx*<sup>†</sup> (asterisks denote earliest appearance of taxon, daggers indicate last appearance). The notations indicate the nature of this turnover fauna, which is placed magnetically just straddling the Sidufjall subchron of the Gilbert chron with a terminal date for the fauna, and the

Hemphillian mammal age, at about 4.8 Ma in earliest Pliocene time.

This fauna is one of a series of sites of Hemphillian age stretching south from the Southern Great Plains of western Texas into the Chihuahua–Coahuila Plateaus and Range (Yepómera), the Trans-Mexican Volcanic Belt (Rancho El Ocote), and into Central America (e.g., the Gracias and Corinto local faunas). All have taxonomically similar ungulate components implying broad environmental similarity deep into the modern Neotropics in the late Miocene.

**Big Bend, Texas (R)** Basin-forming extensional tectonics of the Big Bend, Presidio County, Texas, roughly corresponds in time with those of the rest of the Chihuahua–Coahuila Plateaus and Ranges Province and gave rise to a series of small basins, among which are the Delaho and, to the west, the Santana graben. Both are filled with coarse clastic debris and contain mammal faunas of similar composition. An interbedded basalt in the base of the lower member of the Delaho Formation underlying the Castolon Fauna yields a whole rock K–Ar date of 23 Ma (Stevens et al. 1969; Stevens and Stevens 1989). Rocks (Closed Canyon Formation [Henry et al. 1998], formerly member 9 of the Rawls Formation) containing the Santana Mesa Fauna are bracketed by flows and dikes dated at 23.3–22.5 Ma. The Castolon Fauna (Stevens et al. 1969) contains (taxa denoted with an asterisk are unique to the Big Bend) *Archaeolagus*, *Gregorymys*, *Similosciurus*\*, *Phlaocyon annectens* (Wang et al. 1999), *Parenhydrocyon wallovianus* (Wang 1994), *Moschoedestes delahoensis*\*, *Merychys* cf. *calaminthus*, *Ustatochoerus leptoscelos*\*, *Priscocamelus wilsoni*\*, *Michenia*, *Stenomylus*, *Aguascalientia*, *Delahomeryx*\*, and *Nanotragulus ordinatus*. The Santana Mesa Fauna does not extend this list, and *Aguascalientia* is shared with the Zoyotal Fauna of central Mexico. The level of endemicity of this assemblage poses problems in correlation, but the basalt dates agree with an early late Arikareean age that is close to the age of the base of the Harrison Formation of Nebraska.

The Closed Canyon Formation also has been mapped in a small graben that is not continuous with the Santana Bolson. This site contains the Arenosa (formerly Hodo) site with *Merychys elegans* and indeterminate camelid material (Stevens and Stevens 1989:77) indicating an early Hemingfordian age for that rock body. The unfossiliferous Smokey Creek Member of the Delaho Formation probably is of similar age because the histories of these basins seem roughly concurrent.

## CENTRAL MEXICO

**Aguascalientes (S)** In Mexico, 800 km south of the Big Bend, near the city of Aguascalientes, rhyolitic tuff and interbedded tuffaceous sandstones (Zoyotal Tuff of Hernandez 1981) quarried for building material contain the Zoyotal Fauna (Dalquest and Mooser 1974). Only four taxa represent this assemblage: the oreodont *Merychys* cf. *elegans*, the floridatragulid camel *Aguascalientia wilsoni*, a peccary referred to *Dyseohyus*, and the rhino *Menoceras* (Ferrusquia-Villafranca 1990). The oreodont is a common early Hemingfordian taxon, and the floridatraguline is more derived than the *Aguascalientia* sp. from the Castolon Fauna (Stevens 1977). Although younger than the Castolon and Santana Mesa faunas of Texas, the Zoyotal is dominated by similar elements, suggesting that northern Mexico and adjacent Texas shared environmental features (Stevens 1977), in the early Miocene.

**Guanajuato (T)** In the Trans-Mexican Volcanic Belt several sites occur in fine-grained sediments deposited in a local fault-bounded basin just north of the city of San Miguel de Allende and 1100 km southeast of the temporally and biologically closely allied Yepómera Fauna of Chihuahua. Late Hemphillian fossils occur in the lower 10 m of the exposed basin fill, and at the better-known Rancho El Ocote site they have been divided into two levels. However, the faunas differ mainly in the distribution of large (lower) and small (upper level) taxa, and they are treated as essentially coeval in this work (Carranza-Castañeda and Walton 1992, table 1). The carnivore and ungulate faunas are similar to those at Yepómera at the species level despite the efforts of earlier authors (Dalquest and Mooser 1980) to see the horses, in particular, of the “Ocoté Local Fauna” as “slightly advanced” over those at Yepómera and hence needing taxonomic separation (see MacFadden 1984 for the different opinion followed here). Significant differences with regard to Yepómera are the presence of a sloth identified as *Glossotherium* that, if verified as to genus, would be an earlier occurrence than previously documented in the United States (late Blancan); the slightly greater species diversity of sigmodontine rodents; and the lack of the gomomyids and heteromyids present at Yepómera. These differences probably reflect ecological contrasts between the widely separated sites, but they do not mask the close similarity of these faunas. A fission-track date on an ash in the upper part of the beds of Hemphillian age (Kowallis et al. 1986) gave an age of 4.6 Ma, remarkably close to that

inferred from magnetostratigraphy for comparable strata at Yepómera.

### SOUTHEASTERN MEXICO

**Oaxaca (U)** In Oaxaca the continental Tertiary deposits are largely preserved in narrow northwest–southwest trending grabens such as the Valle de Oaxaca, in the center of the state. Surrounded by horsts of pre-Tertiary rocks, the Cenozoic graben fill includes Paleogene conglomerates and andesitic flows unconformably overlain in the northern part of the graben by the Miocene Suchilquitongo Formation of Wilson and Clabaugh (1970), unconformably overlain by Plio–Pleistocene deposits. The Suchilquitongo Formation is a fluviolacustrine sequence with an interbedded rhyolitic tuff, the Etna Member, that yielded biotite and plagioclase K–Ar ages of  $19.3 \pm 0.3$  and  $20.6 \pm 0.3$  Ma, respectively (Ferrusquia-Villafranca 1992). The Suchilquitongo Local Fauna was obtained 80 m above the Etna Tuff and includes the oreodont *Merychius minimus*, *Merychippus* sp., and a new kyptoceratine protoceratid larger and more hypsodont than *Syndyoceras cooki* (Ferrusquia-Villafranca 1990). This local fauna is best regarded as an early late Hemingfordian assemblage. However, the occurrence of hypsodont equines with a limiting date established for such taxa at 17.5 Ma, that is, younger than the Etna Tuff, may indicate a younger age for the fauna or the need to extend the range of hypsodont equines beyond this limiting date.

In the southeastern part of the Valle de Oaxaca Graben the Tertiary deposits include an extensive pyroclastic sheet partly intertongued and overlain by a tuffaceous fluviolacustrine unit that crops out in the vicinity of Matatlán, 40 km southeast of Oaxaca City. The pyroclastics were K–Ar dated (Ferrusquia-Villafranca 1992) at  $15.3 \pm 0.8$  (biotite) and  $16.0 \pm 0.8$  Ma (plagioclase). The Matatlán Local Fauna (Ferrusquia-Villafranca 1990) was obtained from epiclastic strata overlying the dated pyroclastics, and although represented by fragmentary remains it appears to be Barstovian in age. The horses are diverse and suggest an early late Barstovian age. Continued study of the site by Ferrusquia-Villafranca and associates produced remains of *Leptarctus*, a felid (cf. *Pseudaelurus*), “*Merychippus*” s.l. (both hipparionine and equine forms), a rhino, the camel *Protolabis*, and a leptomerycid (cf. *Pseudoparablastomeryx*).

The first site discovered in Oaxaca was at El Gramal, along the Pan-American Highway in the Nejapa Valley near the Rio Tehuantepec, about 95 km southeast of Oaxaca City. Stirton (1954) reported the discovery, in gently dipping volcanoclastic sediments, of part of a palate and

skeletal fragments of “*Merychippus*” associated in the same concretion with part of an unfused cannonbone of a small camelid identified as “*Oxydactylus*.” The fauna was named El Gramal. Subsequent work in the vicinity of the type locality by J. A. Wilson (1967) and colleagues yielded a broken M3 of “*Merychippus*” sp., a fragment of a gomphotherium tusk (with enamel band), and an upper tooth fragment of a protoceratid. Later collecting by Ferrusquia-Villafranca (1975, 1990) and associates yields the following composite list for the El Gramal Fauna: *Gomphotherium* sp., “*Merychippus*” sp., cf. *Miolabis*, cf. *Protolabis*, and a protoceratid. Two additional horses are represented in the collection, one a small hipparionine similar in size and morphology to *Hipparion shirleyi* and the other a small protohippine, perhaps *Calippus*.

Continued geologic study in the Nejapa area by Ferrusquia-Villafranca and associates has shown that the fossiliferous deposits, the tuffaceous fluviolacustrine El Camaron Formation, overlies and partly intertongues with a sheet of felsic tuff, the Yautepec Tuff, for which K–Ar determinations (Ferrusquia-Villafranca 1992) indicate an age between  $15.0 \pm 0.8$  (biotite) and  $16.7 \pm 0.71$  Ma (plagioclase). The Yautepec Tuff is younger than the Etna Tuff, underlying the Suchilquitongo Formation, and its contained fauna. The El Camaron Formation, contained in a small graben, can be divided into two informal units: a fine-grained lower member and a coarse-grained upper member. The lower member has yielded fossil mammals at a number of localities in the vicinity of El Gramal.

At El Camaron, 7 km south of El Gramal, correlative rocks (Ferrusquia-Villafranca 1990) yield *Plionictis oaxacaensis*, a gomphotheriid, a small protohippine, and a merycodont. Further exploration (Ferrusquia-Villafranca 1990:109) 7 km north-northeast of El Gramal, near the village of La Mancornada, produced *Gomphotherium*, two equids (one referred to the El Gramal hipparionine, the other like the Camaron protohippine), a small and medium-sized camel, and carnivores referred to Canidae and Felidae.

Despite the fragmentary nature of the evidence, the taxonomic similarity in all these Nejapa sites suggests a fauna of broadly late early Barstovian age. The composite fauna is similar to contemporary assemblages of the Great Plains and contains no recognizable Gulf Coast endemics beyond perhaps the protoceratid, depending on its precise identity.

**Central-West Chiapas (U)** This site has yielded the southeasternmost Tertiary mammal fauna of Mexico; it comes from the thick marine and terrestrial sequence preserved in the Ixtapa Graben, located 25 km east-

northeast of Tuxtla Gutierrez, the state capital. The fossil-bearing unit is the Middle Miocene Ixtapa Formation (redefined, Ferrusquia-Villafranca 1996), which is a 3000- to 3500-m-thick calcilitic conglomerate, tuffaceous sandstone, and siltstone sequence interbedded with felsic tuffs, which have yielded biotite and plagioclase K–Ar ages of  $15.02 \pm 0.35$  and  $16.02 \pm 0.53$  Ma in the lower part and  $12.12 \pm 0.19$  and  $12.24 \pm 0.19$  Ma in the middle part (Ferrusquia-Villafranca 1992, 1996).

The small but significant mammal assemblage that forms the Ixtapa Local Fauna (Ferrusquia-Villafranca 1990) was collected from epiclastic beds in the lower part of the formation, located some 200 m below the older dated tuff. The fauna includes the equid *Cormohipparion*, the rhino cf. *Teleoceras*, and the proboscidean *Gomphotherium*.

The equid is close in degree of hypsodonty and occlusal pattern to *Cormohipparion quinni*, known from the late Barstovian (MacFadden 1998 and references therein). *Teleoceras* and *Gomphotherium* are long-lived and widespread genera in temperate North America; however, the latter is represented in Chiapas by a species reminiscent in molar size and occlusal pattern to the Barstovian *G. obscurum*, the most primitive North American species of this genus (Lambert and Shoshani 1998). On this basis, the age of the Ixtapa Local Fauna probably would fall in the late early Barstovian. The stratigraphic position of the fossiliferous beds (about 200 m below the older dated tuff strata, 15–16 Ma), constrains the age to the early Barstovian. This datum is significant on two counts. First, the Ixtapa *Cormohipparion* record antedates by 1 Ma that of *C. quinni*, probably its closest North American sister species. Second, the Ixtapa gomphotheriid is practically coeval with the possible earliest record of *Gomphotherium* in the United States (North Coalinga Local Fauna from the California Coast Ranges; Bode 1935a; Tedford et al. 1987:156) and somewhat older than the earliest widespread appearance of undisputed gomphotheriid remains across the United States, an event that occurred *circa* 14.5–14.8 Ma (Tedford et al. 1987).

## CENTRAL AMERICA

**Honduras and Adjacent El Salvador (V)** Small basins faulted into the ignimbrite plateau of Pacific coastal Honduras and adjacent El Salvador have produced late Miocene faunas resembling those of midlatitude North America. The best known assemblage is the Gracias Fauna of central southern Honduras (reviewed in Webb and Perrigo 1984) and the more recently obtained Corinto Fauna of northeastern El Salvador (Webb and Perrigo

1984). These sites are about 75 km apart and have similar geologic settings: volcanoclastic fine to coarse debris with interbedded ash beds resting on the widespread ignimbrites. The composite fauna includes *Borophagus secundus*, *Rhynchotherium blicki* (type locality), *Hipparion plicatile*, *Cormohipparion cf. occidentale*, *Calippus hondurensis* (type locality), *Prosthennops cf. serus*, *Protolabis cf. heterodontus*, *Procamelus cf. grandis*, and *Pseudoceras skinneri*. The distribution of these taxa in northern faunas implies an early Hemphillian age, marked in Central America by the earliest record of *Rhynchotherium* and a *Calippus* species limited to Florida sites of similar age. The other taxa have wider distributions into the Great Plains. In any event, this is a fauna of midlatitude character at 14°N latitude, 1700 km south of the Gulf Coast of the United States.

**Panama (W)** In 1962 fossil mammals were found in the Cucaracha Formation, exposed in the Gaillard Cut of the Panama Canal by R. H. Stewart, a Canal Zone geologist. The fragmentary remains occurred in bentonitic clay and siltstone in several horizons 1.2–1.5 m thick through a stratigraphic thickness of 45 m of the 90-m-thick unit. The Cucaracha Formation is interbedded with fossiliferous marine beds that are usually assigned an early Miocene age. Whitmore and Stewart (1965) gave a preliminary report on the fauna but never fully described the assemblage. In 1977 Bob Slaughter washed a large volume of matrix from this unit and recovered some rodent remains that represented four taxa, one assigned to a new species of the geomyoid *Texomys*, *T. stewarti*; the other three include two genera that “belong to known North American forms and a third [that] is new to science” (Slaughter 1981:711), which have not been described. The faunal list known so far is *Texomys stewarti*, *Archaeohippus*, *Anchitherium*, *?Diceratherium*, *Brachycrus* (Schultz and Falkenbach 1949 and M. S. Stevens, pers. comm., 2002 all concur with this identification made from the specimens and casts), and *Paratoceras*. Such a collection of taxa is familiar in early Barstovian faunas (e.g., Trinity River Local Fauna) of the Gulf Coast of North America. *Texomys* and *Paratoceras* are geographically limited to Panama and the Gulf Coast. The presence of the peculiar oreodont *Brachycrus* represents a major extension (4000 km) in geographic range from its southwestern United States records. Thus this early Miocene fauna has a composition, like those of the late Miocene faunas of Central America (Gracias, Corinto), consisting of wide-ranging (20° of latitude) forms and Gulf Coast endemics demonstrating the breadth of Miocene faunal provinces and, by inference, comparable ecological regions.

## GULF COAST, NORTH AMERICA

**Texas Coastal Plain (X)** A substantial addition to our knowledge of the earliest known faunas of the Gulf Coastal Plain of Texas comes from Albright's description and analysis (1994, 1996, 1998a, 1998b, 1999b) of the Toledo Bend Local Fauna of easternmost Texas and his review of Arikareean assemblages of Texas and Florida (Albright 1998a). The composition of the Toledo Bend mammal fauna, particularly the lower vertebrate component (Albright 1994), clearly indicates a forested subtropical to tropical coastal lowland paleoenvironment. This affords a rare view of an Arikareean environment that is almost the antithesis of the dry upland ecologies represented by the Great Plains assemblages historically used to typify the Arikareean. Similar settings are indicated for the Florida Arikareean, which may have been subjected to additional endemism by isolation from the mainland by the Gulf Trough. An additional difficulty in placing these assemblages chronologically is the lack of biostratigraphic typification of the medial part of the Arikareean in the Great Plains, a topic we have raised elsewhere that is also discussed later in this chapter. For these reasons correlation of Gulf Coast assemblages with typical Arikareean faunas has been uncertain. However, some taxa in these faunas have broader ecological tolerances and hence larger geographic ranges that provide points of comparison with the midcontinent.

The Cedar Run Local Fauna, previously discussed by Tedford et al. (1987:176) and revised by Albright (1998a), contains *Palaeolagus*, *Palaeocastor*, *Pseudopalaeocastor*, *Daphoenodon notionastes*, ?*Anchippus*, ?*Miohippus* ("Archaeohippus"), *Nexuotapirus*, *Diceratherium*, and two species assigned to *Prosynthetoceras*, as in the Toledo Bend Local Fauna. In addition, Wang et al. (1999:66) found *Phlaocyon minor* at Cedar Run. In the Great Plains comparable genera occur in the correlated "Monroe Creek" and "Harrison" formation sites in southwestern South Dakota, with first occurrences of *Pseudopalaeocastor* (Martin 1987, for *P. barbouri* Peterson 1905) in the "Harrison" and *Nexuotapirus* in the "Monroe Creek." The White River holdovers, *Palaeolagus* and *Palaeocastor*, last occur together in the "Monroe Creek." *Diceratherium* (as *D. annectens*) has a long range in the Arikareean, and the species was described from the Middle John Day.

Albright (1998a) correlated the Cedar Run Local Fauna with the larger Toledo Bend Local Fauna obtained near the Sabine River, Newton County, Texas, just west of the Louisiana border. They share *Daphoenodon notionastes*, possibly *Anchippus* and *Archaeohippus*, *Nexuotapirus*,

*Diceratherium*, *Prosynthetoceras orthrionanus*, and *Prosynthetoceras texanus*. Significant additions in the Toledo Bend Local Fauna are *Neotocastor*, *Protospermophilus*, *Proheteromys* (two endemic species), *Texomys*, *Moropus*, *Dinohyus*, "Cynorca," *Arretotherium acridens*, *Nothokemas* (Albright 1999b), and *Nanotragulus*. There are no identified White River relicts. Albright (1999b) pointed out that the occurrence of *Daphoenodon*, *Moropus*, *Diceratherium*, and *Nanotragulus* limited the fauna to the Arikareean. The presence of Gulf Coast endemic genera *Texomys*, *Anchippus*, *Nothokemas*, and *Prosynthetoceras* has little biochronologic significance in this context, so the weight of evidence for the age of this assemblage rests with zoogeographically more widespread taxa such as *Daphoenodon* (although *D. notionastes* is more primitive than *D. superbus* and has not been recognized outside of the Gulf Coast); *Diceratherium* (as *D. annectens* and *D. armatum*); *Moropus* (a species much smaller than *M. elatus* of the Great Plains late Arikareean, approximating the John Day *M. oregonensis*; see Coombs et al. 2001), very rare in the early Arikareean of the Great Plains (*M. elatus*-sized phalanges are known from Muddy Creek, Wyoming; Coombs 1978:9); *Nexuotapirus marslandensis*, which ranges through the Arikareean into the earliest Hemingfordian (it is only distantly related to prior and contemporary North American tapirs [e.g., *Miotapirus*] and may be an immigrant); *Arretotherium* (compared with *A. acridens* but not sufficiently distinguished from early Arikareean *A. leptodus*); and *Nanotragulus* (possibly close to *N. ordinatus*, which has a late Arikareean range in the northern Great Plains, succeeding *N. loomisi* of the early Arikareean). These taxa imply that the Toledo Bend Local Fauna, like that from Cedar Run, could lie in the medial Arikareean position advocated by Albright (1999b).

**Florida (Y)** Substantial advances have been made in the discovery and description of Arikareean faunas from central Florida, clarification of the age and composition of Barstovian assemblages of northern Florida, and further analysis of the faunal succession in the Bone Valley and correlative faunas of southern Florida.

In a recent review of Arikareean mammal faunas of Florida from the perspective of the Toledo Bend Local Fauna of easternmost Texas, Albright (1998a) assembled a relative Arikareean sequence based on the mammals with reference to the Great Plains. Albright (1999b) reaffirmed this sequence. One of the oldest Florida Arikareean assemblages is the Cow House Slough Local Fauna, from deposits filling the karst developed on the Tampa Limestone Member of the Arcadia Formation. This local



fauna was first discussed by Morgan (1993) and later revised by Hayes (2000). It includes *Centetodon*, *Megalagus*, *Agnotocastor*, *Arikareeomys*, *Proheteromys* (three species), an entoptychine, *Miohippus*, “*Cynorca*,” *Phenacocoelus*, and cf. *Nothokemas*. As Albright (1998a) points out, the joint association of *Centetodon*, *Megalagus*, and *Agnotocastor* implies an early Arikareean age for these holdover taxa from the White River Chronofauna. To this may be added the species of *Proheteromys* that represent post-White River taxa and the yoderomyine *Arikareeomys* (Morgan 1993; the “new large eomyid” of Albright 1998a, not listed by Hayes 2000) typified by material from the McCann Canyon Local Fauna of north-central Nebraska, held by Korth (1992) to be of great “homotaxial similarity to the Monroe Creek Fauna of South Dakota.” Isotopic ages using  $^{87}\text{Sr}/^{86}\text{Sr}$  from mollusks in the fossiliferous fissure containing the Cow House Slough Local Fauna range between 25 and 26 Ma, thus yielding a maximum age for the fauna (Jones et al. 1993).

Hayes (2000) recently described another early Arikareean fauna from the central Florida karst that is more diverse than the Cow House Slough Local Fauna. It is a composite of five sites, closely associated geographically. This assemblage is known as the Brooksville 2 Local Fauna, and like Cow House Slough it contains *Centetodon*, *Megalagus*, and *Agnotocastor*. These White River holdovers are joined by *Parvericius* (or *Amphechinus*); *Proheteromys*; an entoptychine; *Palaeogale*; two endemic musteloids; the canids *Phlaocyon*, *Enhydrocyon* cf. *pahinsintewakpa*, and *Osbornodon*; the horse *Miohippus*; and the artiodactyls *Phenacocoelus*, *Nothokemas*, and *Nanotragulus loomisi*. The canids, oreodont, and *Nanotragulus* again point to an early Arikareean age for the Brooksville 2 Local Fauna. Hayes opted for a “medial” Arikareean age for Brooksville 2 and its correlate Cow House Slough but admitted that they lie in an “undefined biochronologic interval” that lacks typification in the Great Plains reference section. Note that the “Brooksville Fauna” of Tedford et al. (1987) is a younger assemblage tentatively correlated with the Buda Local Fauna.

The White Springs Local Fauna (Morgan 1989 and pers. comm., 2002) of northernmost Florida is important in being a near-shore assemblage with a diverse and abundant shark, ray, and dugongid fauna. The associated land mammals include *Megalagus*, cf. *Neotocastor*, *Arikareeomys*, *Protosciurus*, *Heliscomys*, *Leidymys*, *Anchippus*, *Mesoreodon*, cf. *Oxydactylus*, cf. *Gentilicamelus*, and *Nothokemas* cf. *waldropi*. This fauna, like that of Cow House Slough, contains White River relicts (*Megalagus*, *Heliscomys*) but differs significantly in some taxa (*An-*

*chippus texanus* and *Neotocastor*; Korth 1996a), whose previous records have been considered later Arikareean. Jones et al. (1993) obtained a  $^{87}\text{Sr}/^{86}\text{Sr}$  date of 24.4 Ma on mollusk shells at the White Springs Local Faunal level. Both Albright (1998a, 1999b) and Hayes (2000) show the White Springs Local Fauna as younger than Cow House Slough and Brooksville 2.

In Albright’s (1998a, 1999) and Hayes’s (2000) estimation, the Franklin Phosphate Pit No. 2 and Buda local faunas are correlatives. Of these the Buda Local Fauna, principally described by Frailey (1979), is the larger assemblage. This fauna has been revised by Albright (1998a), and other taxonomic modifications have been provided by systematic studies. It includes *Centetodon*, *Parvericius*, *Texomys*, *Arikareeomys*, *Proheteromys* (two species, including *P.* cf. *magnus* as at White Springs and SB-1A), *Daphoenodon notionastes*, *Phlaocyon achoros* (Wang et al. 1999), *Cynarctoides lemur* (Wang et al. 1999), *Cormocyon* cf. *copei* (Wang et al. 1999), a nimravid, *Moropus* cf. *oregonensis*, *Cynorca*, *Phenacocoelus*, *Nothokemas*, and *Nanotragulus loomisi*. Many of these taxa are endemic to the Gulf Coast, some representing White River relicts (e.g., *Centetodon* and the nimravid). *Nanotragulus loomisi* has a Great Plains range confined to latest White River through Monroe Creek, where it is succeeded by *Nanotragulus ordinatus* in the Harrison. The Buda Local Fauna is compositionally much like the Toledo Bend and Cedar Run faunas of coastal Texas in the continued presence of White River relicts (*Centetodon*) and taxa antecedent to later Arikareean forms (*Texomys*, *Daphoenodon*, and *Moropus*). The Franklin Phosphate Pit No. 2 of Simpson (1930) in northern Florida previously mentioned (Tedford et al. 1987) is regarded as equivalent (Albright 1999b).

The fauna of the SB-1A site (also known as the Live Oak site) is derived from conglomeratic sands, resting on the marine early Oligocene Suwanee Limestone of northern Florida. The local fauna lacks marine or aquatic vertebrates but contains taxa similar to those of the Buda Local Fauna, including *Protosciurus*, *Arikareeomys*, *Proheteromys*, *Mammacyon* cf. *obtusidens*, *Phlaocyon leucosteus* (Wang et al. 1999), “*Paroligobunis*” *frazieri*, *Palaeogale*, and *Nothokemas waldropi*. The rodent fauna is basically like the Florida faunas previously discussed, and the carnivores also suggest an earlier Arikareean age. *Phlaocyon* occurs in the Great Plains before the Harrison Formation, “*Paroligobunis*” *frazieri* is an earlier form preceding the late Arikareean species of *Megalictis* from the Great Plains, *Palaeogale* is similar to the Arikareean species of the genus, and *Mammacyon obtusidens* does not occur in the late Arikareean. Locality SB-1A also has

*Daphoenodon notionastes* (Hunt, pers. comm., 2001), as at the Buda and Franklin Phosphate Pit No. 2 sites. Thus the SB-1A Local Fauna appears similar in age to the Buda Local Fauna and Gulf Coast correlatives.

MacFadden (1980) described the Martin–Anthony oreodont site and overlying strata containing a few terrestrial mammal remains. The terrestrial elements accompanied dugongs and turtles in an estuarine sequence. Marine invertebrates accompanying the oreodont *Phenacocoelus typus* (“*P. luskensis*” of MacFadden 1980) have been  $^{87}\text{Sr}/^{86}\text{Sr}$  dated to 24.6 Ma (Jones et al. 1993), remarkably close to the date for the White Springs Local Fauna. The holotype of this oreodont taxon occurs in the upper part of the type Monroe Creek Formation of Nebraska (Hunt 1985), and the taxon ranges into the Harrison Formation in the northern Great Plains. A similar oreodont is also present in the Buda Local Fauna, here deemed slightly younger than White Springs.

The overlying siliciclastics (“Hawthorne Formation” of MacFadden 1980, now Penny Farms Formation of the Hawthorn Group) contain the Martin–Anthony Local Fauna, whose land mammals include (Albright 1998a) *Mammacyon* (“*Temnocyon*,” Albright 1998a; reidentified by Hunt 1998), *Menoceras arikareense*, and a “small new genus and species” of camelid. The latter unnamed taxon was described by Frailey (1979:151–154) from the Buda Local Fauna and has also been recognized (Albright 1998a) in the White Springs, Franklin Phosphate Pit 2, and Cow House Slough faunas.

The Gulf Coast Arikareean sites have been difficult to place precisely in time mainly because of the lack of correlative faunas in the Great Plains and the zoogeographic and ecologic peculiarities of the John Day sequence, whose radioisotopic calibration indicates that it covers the interval in question. As a first step we have tried to deduce the relative sequence of the Gulf Coast assemblages. Second, we have accepted the 24 Ma strontium age for the White Springs Local Fauna as a calibration point.

If this is credible, the sequence is marked by three features: upward extension of range zones of members of the White River Chronofauna, in dwindling numbers through the reconstructed sequence; presence of several Great Plains Arikareean taxa whose records begin in the late early Arikareean (Ar2) but do not occur in the late Arikareean (Ar3) of the plains; and downward extension of species in genera whose oldest Great Plains record is late Arikareean (Ar3). These include two immigrants, *Menoceras* and *Moropus*; an important member of the Ar3 characterizing fauna, *Daphoenodon*; and such Gulf Coast endemics as *Texomys* and *Prosynthetoceras* better known in younger deposits.

Bryant et al. (1992) obtained a  $^{87}\text{Sr}/^{86}\text{Sr}$  date on mollusks just below the site producing the Seaboard Local Fauna in the lower Torreya Formation of northern Florida (Olsen 1964; Tedford et al. 1987). The date,  $18.4 \pm 1.0$  Ma, provides the only isotopic indication of the age of the Seaboard Local Fauna and its correlative Thomas Farm Local Fauna. The limits of this date overlap those provided by the same method at the Pollack Farm site in Delaware and suggest equivalence with the Runningwater faunas of the Great Plains.

Important new data are now available for faunas from north Florida that come from the upper or Dogtown Member of the Torreya Formation. The marine Torreya Formation provided a correlation with planktonic foraminiferal zones in our previous work (Tedford et al. 1987:179). This evidence has been reviewed and extended by  $^{87}\text{Sr}/^{86}\text{Sr}$  ages and magnetostratigraphy by Bryant et al. (1992) and the fauna revised by Bryant (1991). The Midway and Quincy faunas of the northern Florida fuller’s earth mines were treated together by Tedford et al. (1987). The Quincy Local Fauna contains only “*Merychippus gunteri*” as a land mammal. Bryant (1991) has shown that this taxon has a longer stratigraphic range in the Torreya, so the age of that local fauna is not constrained on that basis. The Midway Fauna (s.s.) has a larger suite of nonmarine mammals that appear to represent a younger fauna than the medial Hemingfordian Thomas Farm Local Fauna of central Florida.

Bryant (1991) was able to lithologically correlate local quarry exposures in northern Gadsden County over several kilometers and thereby relate fossiliferous levels into a coeval fauna, the Willacoochee Creek Fauna. This assemblage contains the following nonmarine mammals: *Lanthanotherium*, *Mylagaulus*, ?*Protospermophilus*, *Perognathus*, *Proheteromys*, *Copemys*, *Cynorca*, *Ticholeptus*, *Bouromeryx*, *Rakomeryx*, ?*Blastomeryx*, *Anchitherium clarencei*, “*Merychippus gunteri*,” “*Merychippus primus*,” *Acritohippus* cf. *isonesus*, ?*Aphelops*, and a possible proboscidean (not in situ or from the better-sampled sites). This fauna has several wide-ranging taxa whose biochrons are well known from the midcontinent to the West Coast, supporting an early Barstovian age with range extensions needed only for *A. clarencei* and “*M. primus*.” If verified as a part of this fauna, the mastodont would constitute the earliest Florida occurrence of proboscideans (early Barstovian).

Strontium isotope age estimates between  $14.7 \pm 1.5$  and  $16.6 \pm 1.0$  Ma were obtained from mollusk shells in the fossil vertebrate-containing strata within the Dogtown Member of the Torreya Formation. A magnetostratigraphic study, including the fossiliferous interval produc-

ing the Willacoochee Creek Fauna, showed that the interval is entirely reversed and probably correlates with the distinctive long reversed portion of chron C5B (16.2–15.3 Ma; Berggren et al. 1995). All these data are compatible with calibrations of the early Barstovian interval elsewhere. The strong relationship of the Willacoochee Creek Fauna with correlatives in the midcontinent is echoed by the coeval Trinity River Local Fauna of the Texas Gulf Coast and represents a feature of the zoogeographic relationships of these and younger Miocene faunas of the Gulf Coast.

Barstovian and Clarendonian assemblages lately have been found in the top of the Arcadia and base of the overlying Peace River Formation of the phosphate mining district of southern Florida (Morgan 1993). These assemblages help to fill a previous gap in the Florida sequence between the Hawthorn Group and “Alachua Clays” of the older literature. All these assemblages have a strong affinity with correlatives in midcontinental North America.

The stratigraphically oldest is the Bird Branch Local Fauna, for which only *Copemys* and teeth of *Acritohippus* cf. *isonesus* (Hulbert and MacFadden 1991, figure 9A–B) are known, both compatible with early Barstovian faunas elsewhere. Above this level, in the top of the Arcadia or base of the Peace Valley formations, lies the Sweetwater Branch Local Fauna (the fauna of Unit 1 of Webb and Crissinger 1983), which includes a small mustelid, “*Merychippus*” *goorisi*; *Merychippus* cf. *brevidontus*, a “parahippine”; a rhino, cf. *Peraceras hessei*; a camelid; and a dromomerycid, cf. *Bouromeryx* (Hulbert and MacFadden 1991:37–39, figure 9B–I). This assemblage can be correlated with the Trinity River Local Fauna of the Texas coastal plain and with early Barstovian assemblages in the midcontinent.

The lower part of the Peace River Formation yields the Bradley Fauna (the fauna of units 2–3 of Webb and Crissinger 1983) of early late Barstovian age, which contains the horses *Megahippus*, *Protohippus perditus*, *Pliohippus mirabilis*, and *Calippus proplacidus*; the dromomerycid *Procranioceras* cf. *skinneri*; *Zygodon*; and a gomphothere near *Gomphotherium calvertensis* of the Atlantic coastal plain. The Agricola Fauna of early Clarendonian age (the fauna of unit 5 of Webb and Crissinger 1983) contains the borophagine *Epicyon*; the horses *Hypohippus affinis*, *Protohippus supremus*, *Calippus martini*, *Cormohipparion occidentale*, and *Pseudhipparion curtivalum* (Hulbert 1988a, 1988b); the rhino *Teleoceras*; and the protoceratid *Synthetoceras*, all taxa with geographic ranges that include the Great Plains.

Recent additions to the Palmetto Fauna (“Upper Bone Valley Fauna” in Tedford et al. 1987) have elucidated some

elements of this late late Hemphillian fauna in the Gulf Coastal Plain. Description of *Eocoileus gentryorum*, one of the earliest cervids in North America, clarifies its relationships to other odocoileines and indicates that more than one such genus immigrated to North America (Webb 2000). The presence of *Miopetaurista*, previously known from the late Blancan of Florida, indicates this large flying squirrel as a latest Hemphillian immigrant from Eurasia. The Palmetto Fauna is noted for holding over several lineages that evidently survived in the Gulf Coastal Plain after they had disappeared from the High Plains. Examples include *Cormohipparion emsleyi*, *Pseudhipparion simpsoni*, *Kyptoceras amatorum*, the last of the protoceratids, and *Pseudoceras*, the last North American gelocid.

**Atlantic Coast (Z)** Tedford and Hunter (1984) tried to use the association of terrestrial mammals in near-shore marine deposits as a tool to compare marine–nonmarine correlations of the eastern seaboard. For this purpose they reviewed the available data known at that time, particularly those of the historic sites in the Kirkwood Formation of New Jersey and the Calvert and overlying Choptank formations of Maryland and Virginia. These results were discussed only briefly by Tedford et al. (1987). Since that time important new analyses of the mammalian biostratigraphy of the Calvert and Choptank strata have been advanced, and a very important new site in Delaware has greatly improved our knowledge of the early to middle Miocene of the Atlantic coastal region.

Peccaries are the most common fossil mammals in the Calvert Formation faunas. Wright and Eshelman (1987) reviewed this record and on that basis proposed a more detailed biostratigraphy for the containing rocks. Tedford and Hunter (1984) had combined the mammal remains from the upper (Plum Point Marl Member) of the Calvert Formation (Shattuck 1904, Beds 10, 13–15) and the basal strata of the overlying Choptank Formation as a coeval Chesapeake Bay Fauna. Wright and Eshelman (1987) have shown that there is a biostratigraphic sequence of tayassuid taxa through this interval. Following their lead, we present the following bed-by-bed analysis in ascending order:

- Bed 10 (Calvert Formation): ?*Cynarctus marylandica* (Wang et al. 1999), and *Cynorca proterva*
- Bed 12: *Cynorca proterva*
- Bed 13 or 14: *Gomphotherium calvertensis* (Gazin and Collins 1950), holotype
- Bed 14: “*Prosthennops*” *xiphodonticus*
- Bed 14 or 15: ?*Aphelops* sp.

Bed 17 (Choptank Formation): cf. *Gomphotherium calvertensis*, “*Prosthennops*” *niobrarensis* (“*Hesperhys*” of Gazin and Collins 1950)

The remainder of the Chesapeake Bay Fauna, including taxa identified by C. D. Frailey from collections made since Gazin and Collins (1950), cannot be stratigraphically allocated. These include *Amphicyon*, *Tapiravus* cf. *validus* (holotype from the Shiloh Local Fauna, Kirkwood Formation of southern New Jersey, probably of early Miocene age; Emry and Eshelman 1998), *Archaeohippus*, “*Merychippus*,” a dromomerycid, and ?*Aepycamelus*. Wright and Eshelman (1987) compared the Calvert and Choptank peccary taxa with the midcontinent biostratigraphy of these forms to indicate that Bed 10 was early Barstovian, Bed 14 early late Barstovian, and Bed 17 late late Barstovian. Thus the earliest evidence of gomphotheres on the east coast lies in the early part of the late Barstovian.

Overlying the Choptank Formation in Maryland is the St. Mary’s Formation, whose basal sands, the Little Cove Point Unit (Blackwelder and Ward 1976), produced a jaw of *Cormohipparion* and a glauconite K–Ar date of  $12.3 \pm 0.5$  Ma (Blackwelder 1981) that limits the underlying Choptank to the late Barstovian.

The Pollack Farm site in Kent County, central Delaware, was discovered in 1991, while the enclosing rocks were being quarried for building materials. The land mammal remains, grouped as the Pollack Farm Local Fauna (Emry and Eshelman 1998), were obtained from spoil piles and in situ so that the fauna can be confidently assigned to the lower shell bed of the deltaic Cheswold Sand unit of the lower Calvert Formation. The terrestrial mammals include *Anchitheriomys*, “*Monosaulax*” (cf. *Neotocastor*), *Proheteromys magnus*, *Plesiosminthus*, “?Hemicyon” (cf. *Phoberocyon*), *Amphicyon*, *Cynelos*, *Paracynarctus kelloggi* (Wang et al. 1999), *Metatomarctus canavus* (Wang et al. 1999), *Archaeohippus* cf. *blackbergi*, *Anchitherium*, *Parahippus leonensis*, cf. *Tylocephalonyx*, *Cynorca sociale*, and *Blastomeryx galushai*. This assemblage has a strong resemblance to midcontinent faunas of early Hemingfordian age; the only Gulf Coast taxa are the horses, but that may reflect the lack of knowledge of the taxonomic status of the midcontinent equid fauna of this age. Strontium isotopic age estimates from shells in the lower shell bed range from 17.8 to 18.2 Ma, with a mean of  $17.9 \pm 0.5$  Ma (Jones et al. 1998). This is slightly younger than the paleomagnetically established local upper limit of the Runningwater Formation of western Nebraska (18.0 Ma; MacFadden and Hunt 1998) but in agreement with the estimated age of

the upper Runningwater Formation, which probably extends from 17.5 to 18.0 Ma (Hunt, pers. comm., 2001).

Emry and Eshelman (1998) reevaluated the ages of the Shiloh and Farmingdale faunas of New Jersey in light of new stratigraphic and geochronologic information (see also Benson 1998). Subsurface tracing of the Shiloh Marl from its outcrops to the Pollack Farm site shows that it stratigraphically underlies the Pollack Farm Local Fauna (separated by a disconformity fide Benson 1998), so that the holotype of *Tapiravus validus* (Marsh 1871) is not correlative with the Pollack Farm or Chesapeake Bay assemblages (the latter *contra* Tedford and Hunter 1984). Sugarman et al. (1993) give strontium isotope estimates of 20.0 to 20.3 Ma for the Shiloh Marl, clearly late Arikarean dates (MacFadden and Hunt 1998).

The Farmingdale Fauna from the basal Kirkwood Formation of central New Jersey, placed in the early Hemingfordian by Tedford and Hunter (1984), is more properly dated to the late Arikarean on the joint occurrence of *Diceratherium* and *Menoceras*, as pointed out by Emry and Eshelman (1998). The fauna, assembled from sites at and near Farmingdale, Monmouth County, New Jersey, was described by Marsh in the nineteenth century: *Diceratherium matutinum* (Marsh 1870), *Daeodon leidymanum* (Marsh 1893), and *Hesperhys antiquus* (Marsh 1870), to which have been subsequently added (AMNH collection) *Menoceras* cf. *arikareense*, *Anchitherium*, and *Prosynthetoceras*. Strontium isotopic estimation of age from nearby boreholes penetrating the base of the Kirkwood give values of 19.2–22.6 Ma (Sugarman et al. 1993), in good agreement with dated ash beds and magnetostratigraphy for the type Harrison Formation of western Nebraska (MacFadden and Hunt 1998).

## BIOCHRONOLOGY

In the 1987 report the nature and methods of typification of the North American mammal ages were discussed in their historical context and in light of the refinements that were possible at that time. The present review has marshaled more evidence for the faunal succession in various parts of North America, which leads in some instances to better biostratigraphic documentation of the faunal sequence, augmented by improved taxonomic resolution. In addition, a rapidly growing body of geochronologic data is available from ash dating and element comparison accompanied by comprehensive magnetostratigraphies relating fossil mammal occurrences to the radioisotopic and Geomagnetic Polarity Time Scale

(GPTS). It is possible to correlate distant biostratigraphies on these physical bases to test biological correlations and to better place events in geologic time.

The accumulated evidence, once again, allows a rough delineation of zoogeographic provinces characterized by contrasting but contemporaneous faunal compositions. This is particularly true of the Hemingfordian through Barstovian interval, where magnetic data from the Great Plains to West Coast allow such precise correlation.

This review has also reinforced the existence and faunal distinction of later Cenozoic chronofaunas and provided data on their assembly, acme, and decline. Webb (1983) has traced the history of the Miocene Chronofauna and, in a more general treatment (Webb and Opdyke 1995), has further delineated the chronology and succession of such events in the Oligocene and Miocene and tried to show that aspects of their history seem to relate to global climatic change. In a related treatment, Woodburne and Swisher (1995) have examined faunal change, particularly the episodes marked by immigration, in terms of sea level history. In this work we refine the chronology and nature of these events in terms of the revisions now possible and show that chronofaunal turnover occupies a span of time during which the previous assemblage is replaced by extinction and the elevated evolution of new autochthones as new allochthonous taxa ("immigrants") are added.

Dawson's (1999) recent analysis of dispersal of Miocene land mammals between Eurasia and North America provides a critique of the taxa chosen in 1987 to define the mammal ages and their subdivisions. We have accepted this timely advice for the most part but depart when we believe the evidence is secure. In the recognition of allochthones we place particular weight on their phyletic isolation from comparable taxa in the contemporary and preceding autochthonous faunas. A penetrating analysis of the relationship of New and Old World Miocene insectivores and rodents by Engesser (1979) rejected most previous claims of allochthonous status where he detected morphologic differences between the presumed vicars. He attributed the similarity of the putative Old and New World vicars to parallel evolution involving similar adaptive trends and conservation of primitive features. These hypotheses deserve testing by more detailed phyletic analysis and serve as a warning regarding facile assumptions of close affinity demanded by the hypothesis of dispersal. A summary of the defining taxa proposed here for each mammal age and their subdivisions is presented in figure 6.3.

Typology still plays a controlling role in determining the temporal span of the NALMAs such that the limits

set by Wood et al. (1941) are not significantly compromised. Evidence discussed in 1987 and in this revision certainly suggests striking turnover events more suitable for biochronologic boundaries, but these must be subordinated to the traditional limits of each age to avoid wholesale revision of the content of every NALMA. Following Woodburne and Swisher (1995) we designate subdivisions of each NALMA by numbered intervals preceded by an abbreviation for the mammal age.

## ARIKAREEAN

This NALMA was equated with the limits of the Arikaree Group as "redefined by Schultz (1938), but including the Rosebud" (Wood et al. 1941:11). In these terms the Arikaree (and Arikareean) was originally confined to the Gering, Monroe Creek, and Harrison formations of northwestern Nebraska and their postulated equivalent in adjacent South Dakota, the Rosebud Formation of Matthew and Gidley (1904).

In faunal terms the Arikareean included the total faunal content of the Arikaree Group so defined but with "Agate [i.e. Agate Springs Local Fauna] being [the] most typical locality" and the "John Day (in part)" as a "principal correlative" (Wood et al. 1941:11). However, the faunal characterization ("Index fossils" and "First appearances") listed mainly taxa drawn from the Harrison Formation, and the "Last appearances" were taxa characteristic of the assemblages of the Gering, Monroe Creek, and Lower Rosebud strata.

Modification of the limits of the stratal span of the Arikareean has since been proposed. At the base, the age now includes the fauna of the uppermost member of the Brule Formation ("Brown siltstone" unit of Swinehart et al. 1985) of the White River Group, a unit included in the Gering Formation by Vondra et al. (1969), whose lithostratigraphy was the basis of Martin's (1973) pioneering study of the "Gering Fauna." The detailed biostratigraphic and geochronologic study of the Brown Siltstone and overlying Gering and of correlative Sharps Formation strata discussed earlier now provides a more secure basis for a faunal definition and characterization of the early Arikareean.

Faunas representing the earliest phase of the Arikareean (Ar1) combine elements of both the White River Chronofauna and new taxa arising by evolution and immigration that increase faunal diversity. Study of this early phase by Tedford et al. (1996) has modified the definition and characterization of the early Arikareean. The Arikareean is now defined by the earliest appearance of a single immigrant, the zapodid rodent *Plesiosminthus*.

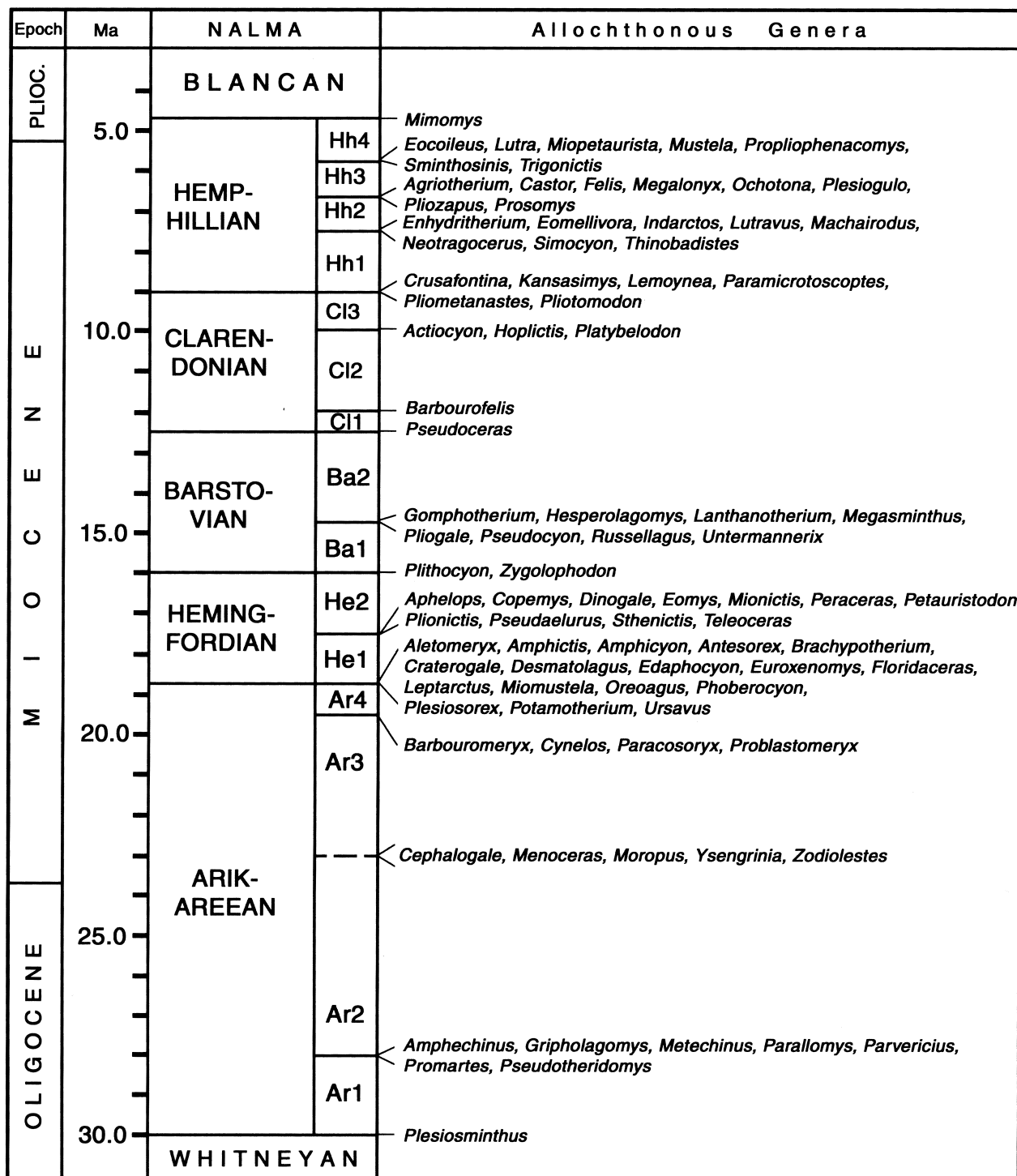


FIGURE 6.3 Chronology used in this chapter. Epoch boundaries and chronologic scale follow Berggren et al. (1995). North American land mammal age (NALMA) boundaries as proposed in this work are denoted by solid lines where well calibrated, dashed where approximately located. Allochthonous genera represent taxa exotic to the North American fauna. They are regarded as immigrants that have their oldest records as indicated. This figure supersedes figure 6.3 of Tedford et al. (1987).

The age is characterized by the earliest appearance in the Great Plains of autochthonous taxa including the beavers *Capacikala* (*C. gradatus*) and *Capatanka* (*C. cankpeopi*); the oreodonts *Sespia* (*S. nitida*) and ?*Mesoreodon* (?*M. minor*); the hypertragulid *Nanotragulus* (*N. loomisi*) and within the interval, but not necessarily in its oldest phase, the appearance of the primate *Ekgmowechasala*; the erinaceid *Ocajila*; the talpid *Scalopoides*; the murid *Paciculus* (*P. woodi*); the rodent families Mylagaulidae (Promylagaulinae) and Aplodontidae (*Niglarodon*); the hesperocyonine canid *Enhydrocyon* (*E. pahinsintewakpa*); the entelodont *Dinohyus*; the oreodont *Megoreodon* (*M. grandis*); and the stenomyline camel *Miotylopus*.

Confined to the early early Arikareean (Ar<sub>1</sub>) are the limited occurrences of the rabbit *Palaeolagus hypsodus*, the beaver *Palaeocastor nebrascensis*, the florentiamyid *Kirkomys* (*K. schlaikjeri*), and the hesperocyonine canids *Cynodesmus* (*C. martini*) and *Shunkahetanka* (*S. gerinensis*). Correlative faunas of the John Day Formation are bracketed by the AB Tuff (29.75 ± 0.02 Ma) and Deep Creek Tuff (27.89 ± 0.57 Ma). These John Day strata indicate first appearances of the following additional autochthonous genera: the rabbit *Archaeolagus* (*A. ennisianus*); the aplodontids *Allomys*, *Meniscomys*, and *Alwoodia* (*A. magna*); the entoptychine *Pleurolicus* (*P. sulcifrons*); the oreodont *Oreodontoides* (*O. oregonensis*); and the *Hypertragulus* subgenus *Allomeryx* (*A. planiceps*).

Last occurrences in the early early Arikareean (Ar<sub>1</sub>) of the Great Plains include such White River genera as *Agnotocastor*, *Palaeocastor*, *Eumys*, *Hyaenodon*, *Eusmilus*, *Hesperocyon*, *Cynodesmus*, *Protapirus*, *Hyracodon*, *Elomeryx*, *Leptochoerus*, *Perchoerus*, *Chaenohyus*, *Hypertragulus*, *Leptomeryx*, *Hypisodus*, and *Agriochcerus*. In the John Day Formation *Agriochcerus*, *Leptomeryx*, and *Hypertragulus* persist into younger strata, and in the Gulf Coast *Agnotocastor* and *Palaeocastor* also persist.

Additional taxa appear in lower Arikaree strata above the basal Gering and correlative strata in the Great Plains (Tedford et al. 1996). These can be used to define and characterize a second phase in the early Arikareean (Ar<sub>2</sub>): the late early Arikareean. This phase is defined by the first appearance of allochthonous taxa: the erinaceids *Amphichinus* and *Parvericius* and the brachyericine *Metechinus*, the Ochotonidae (*Gripholagomys*; Green 1972), the aplodontid *Parallomys*, the eomyid *Pseudotheridomys*, and the mustelid *Promartes*. Accompanying these in the Great Plains are the first appearance of such autochthones as the leporid *Archaeolagus* (*A. cf. ennisianus*); the beaver *Fossorcastor* (*F. fossor* and *F. brachyiceps*); the promylagaulines *Trilaccogaulus* and *Promylagaulus*; the aplodontid *Alwoodia* (*A. harkseni*); the entoptychines *Entopty-*

*chus*, *Pleurolicus*, and *Gregorymys*; the florentiamyids *Fanimus* and *Florentiamys*; the amphicyonid *Mammocyon*; the tapir *Nexuotapirus*; the peccary *Cynorca*; the anthracothere *Arretotherium*; the oreodonts *Oreodontoides* (*O. oregonensis*) and *Promerycochoerus*; and the camel *Stenomylus*. In the Gulf Coast such taxa as *Daphoenodon* and *Prosynthetoceras* have their first appearances in the later part of the Ar<sub>2</sub> interval but achieve larger ranges in late Arikareean and Hemingfordian time.

Last occurrences in the late early Arikareean (Ar<sub>2</sub>) of the Great Plains include forms that do not occur in the Harrison Formation (ca. 23 Ma) after a hiatus in the local record of nearly 3 m.y. These include the remaining White River taxa: the Nimravidae (*Nimravus*), the Proscalopidae (*Proscalops*), the Geolabididae (*Centetodon*), the Palaeolaginae (*Palaeolagus*), the Eutypomyidae (*Eutypomys*), and the Yoderomyinae (*Arikareeomys*), as well as the florentiamyid rodents *Sanctimus* and *Hitonkala*, the cricetine *Geringia*, the promylagauline *Crucimys*, the equid *Mesohippus*, the entelodont *Archaeotherium*, and the leptachenine oreodonts. In addition, many of the taxa listed as first appearances in the late early Arikareean also do not survive the turnover concealed by the mid-Arikareean hiatus.

In the John Day Formation, the interval from the Deep Creek Tuff (27.89 ± 0.57 Ma) to the Tin Roof Tuff (25.9 ± 0.31 Ma) covers the early part of the Great Plains hiatus. Local first appearances during this interval include the entoptychine *Entoptychus*; the oreodonts *Merycooides*, *Hypslops* (*H. brachymelis*), and *Paroreodon*; and the nothokematine camelid *Gentilicamelus* (*G. sternbergi*).

Our reconstruction and calibration of the early Arikareean faunal sequence in the Gulf Coast suggest the continued presence of such White River taxa as *Centetodon*, *Palaeolagus*, and *Palaeocastor* and the presence of such late Arikareean taxa as *Moropus* and *Daphoenodon*, indicating that a phase of turnover to faunas typical of the late Arikareean of the Great Plains was under way by 24 Ma in that region. The Castolon and to a greater extent the Santana Mesa faunas of the Big Bend in Texas are associated with 23 Ma basalts and contain *Merychys*, early *Ustatochoerus*, *Michenia*, and *Nanotragulus ordinatus*, showing that faunas of late Arikareean (Ar<sub>3</sub>) were already in place in the continental interior by the earliest Miocene.

We believe that the Florida "medial" Arikareean lies in the Great Plains Arikaree hiatus and therefore carries elements that later typify the late Arikareean at least to 23 Ma, especially the immigrant rhinocerotid *Menoceras* and chalicothere *Moropus*. We use these two taxa to define

the earliest late Arikareean (Ar<sub>3</sub>). Other immigrants have their earliest appearance in the Harrison Formation maximally calibrated to 23 Ma. The following taxa also define the Ar<sub>3</sub>: the amphicyonid *Ysengrinia*, the ursid *Cephalogale* (Hunt, pers. comm., 2001), and the mustelid *Zodiolestes*.

The late Arikareean as a unit (i.e., Ar<sub>3</sub> and Ar<sub>4</sub>) is characterized by the earliest occurrence of such autochthones as the mylagaulid *Mylagaulodon*; the amphicyonid *Daphoenodon*; the borophagine canid *Desmocyon*; the tapir *Miotapirus*; the horses *Kalobatippus*, *Archaeohippus*, *Desmatippus*, and “*Parahippus*”; the tayassuid *Hesperhys*; the oreodonts *Merycochoerus*, *Ustatochoerus*, and *Merychys*; the kryptocerine protoceratid *Syndyceras*; the protolabine camels *Michenia*, *Protolabis*, and *Tanymyktekter*; and the “oxydactylid” camels *Priscocamelus* and *Oxydactylus*.

The late Arikareean is also characterized by the last occurrence of *Gripholagomys*; the flat-incisor, burrowing beavers *Euhapsis*, *Pseudopalaeocastor* (*P. barbouri*, incorrectly assigned to a new genus *Nannasfiber* by Xu 1996), and *Fossorcastor* (*F. fossor*, *sensu* Xu 1996, referred to *Palaeocastor* by Martin 1987, the *Daemonelix* burrow maker); most entoptychines and florentiamyid rodents; the canid *Cormocyon*; the amphicyonids *Temnocyon* and *Mammacyon*; the mustelids *Zodiolestes* and *Megalictis*; the horse *Miohippus*; the rhino *Diceratherium* (may survive into the late Hemingfordian in Panama); the oreodonts *Eporeodon*, *Phenacocoelus*, *Promerycochoerus*, *Pseudodesmatochoerus*, *Desmatochoerus*, *Hypsiops*, *Oreodontoides*, *Paroreodon*, *Paramerychys*, and *Submerycochoerus*; the camels *Pseudolabis* and *Miotylopus*; the protoceratid *Syndyceras*; the hypertragulids *Hypertragulus* and *Nanotragulus*; and the leptomerycid *Pronodens*.

A further subdivision, Ar<sub>4</sub>, the late late Arikareean (fauna of the Upper Harrison beds, now Anderson Ranch Formation; Hunt 2002), is defined by the first appearance of the immigrant amphicyonid *Cynelos*, the first Moschidae (*Problastomeryx*), the earliest representatives of the Dromomerycidae (*Barbouromeryx*), and Antilocapridae (*Paracosoryx*).

Autochthonous taxa appearing first in the late late Arikareean, Ar<sub>4</sub>, as shown principally by the fauna of the Upper Harrison and Rosebud (*sensu* Macdonald 1963) beds of the Great Plains include the mylagaulid *Mylagaulodon*, the mustelid *Megalictis*, the equid *Desmatippus*, the oreodonts *Merycochoerus* and *Merychys arenarum*, the protolabine camels *Tanymyktekter* and *Protolabis*, the miolabine camel *Miolabis*, and the moschids *Blastomeryx*, *Parablastomeryx*, and *Machaeromeryx*.

The loss of burrowing beavers and representatives of many oreodont clades that rapidly diversified in the early

Arikareean, as well as members of other earlier autochthonous lineages, indicates that a major turnover was in progress before the time when younger Arikaree Group rocks began to accumulate in the Great Plains. Limited evidence from the Gulf Coast and Columbia Plateau (John Day) indicates that this change was under way by 24 Ma. This extinction is accompanied by renewed immigration, the true dimensions of which become more evident in the succeeding Hemingfordian, when small mammal faunas also become available. By the close of the Arikareean the remaining autochthonous fauna and the evolving allochthonous clades (Moschidae, Dromomerycidae, and Antilocapridae) formed the core assemblage for the Hemingfordian.

## HEMINGFORDIAN

The Hemingfordian of Wood et al. (1941) was “based on the Hemingford Group including the Marsland and, especially, the limited or lower Sheep Creek fauna (Cook and Cook 1933, pp. 38–40), and not on the formation limits as extended upward (Lugn 1939b).” These qualifications demonstrate that the Hemingfordian is not to be equated with the total Hemingford Group of Lugn (1939), but it was still ambiguous with regard to its lower limits because of the confusion regarding Schultz’s (1938) original definition of the Marsland Formation. As discussed in more detail in Tedford et al. (1987:185–186), the view we have accepted equates the Marsland Formation as used by Schultz (1938) as a composite made up of two formation rank rock units: the Upper Harrison beds of Peterson (1907, 1909), a name recently replaced by the term *Anderson Ranch Formation* (Hunt 2002), and the superjacent Runningwater Formation of Cook (1965). Schultz considered Peterson’s Upper Harrison beds as roughly equivalent to his “lower Marsland” and Cook’s Runningwater Formation as his “upper Marsland.” It is clear from the characterizing assemblage of the Hemingfordian given by Wood et al. (1941) that the base of the Hemingfordian was marked by taxa of the Runningwater Formation, not those of the Upper Harrison (containing Agate Springs Quarries, the “typical locality” for the Arikareean; Hunt 1985). These considerations remove the potential typologic overlap between the Arikareean and Hemingfordian.

As noted in Tedford et al. (1987), the typification of the early Hemingfordian (and thus the total NALMA) is based mainly on evidence from the Great Plains, supplemented by Gulf Coast assemblages. Scattered local faunas from the Great Basin and West Coast can be identified as representing the early Hemingfordian. Although



they do not contribute much to the biological characterization of the Hemingfordian, they do contribute to its chronology and zoogeography.

An invasion of immigrants defines the beginning of Hemingfordian and hence early Hemingfordian (He1) time. Small mammals become more visible, and the immigrant fauna is more balanced in size range and breadth of adaptation. This event is accompanied by an overall change in sedimentary environment as volcanoclastic aeolian deposits of great lateral extent representing the Arikaree Group give way to coarser epiclastic fluvial sands and gravels of the Ogallala Group that locally fill deeply incised river valleys containing evidence of more varied ecologies. The Hemingfordian can be defined by the following immigrants: the Soricinae (*Antesorex*), the Plesiosoricidae (*Plesiosorex*), the ochotonids *Oreolagus* and ?*Desmatolagus*, the trogontheriine beaver *Euroxenomys*, the amphicyonid *Amphicyon*, the Hemicyonidae (*Phoberocyon*), the Ursidae (*Ursavus* earliest certain record), the Procyonidae (*Amphictis* and *Edaphocyon*), the Leptarctinae (*Craterogale* and *Leptarctus*), the Mustelinae (*Miomustela*), the Semantorinae (*Potamothenium*), the Teleoceratinae (*Brachypotherium*), the Aceratherinae (*Floridaceras*), and the dromomerycid *Aletomeryx*.

The early Hemingfordian (He1) is characterized by the earliest appearance of the mylagaulid *Mesogaulus*; the heteromyine *Cupidinimus*; the borophagine canids *Paracynarctus*, *Metatomarctus*, *Euoplocyon*, *Microtomarctus*, and *Prototomarctus* (Wang et al. 1999); the mustelid *Brachypsalis*; the equids *Hypohippus* and "*Merychippus*" ("*M. gunteri*"); the peccary *Floridachoerus*; the oreodonts *Merychys elegans*, *Ticholeptus*, and *Mediochoerus*; the protoceratid *Lambdoceras*; and the camels *Floridatragulus*, *Blickomylus*, and *Australocamelus*.

The early Hemingfordian (He1) contains the limited occurrence of *Amphictis*, *Craterogale*, *Floridaceras*, *Lambdoceras*, *Aletomeryx*, and *Probarbouromeryx* and the latest occurrence of florentiamyine rodents (Korth et al. 1990), *Desmocyon*, *Menoceras*, *Nexuotapirus*, *Kalobatippus*, Entelodontidae, Anthracotheriidae, *Merycochoerus*, *Tanymykter*, and *Stenomylus*.

A striking faunal change takes place between the early and later Hemingfordian, which demonstrates the folly of tying a biochronology to the depositional record. This turnover event is not clearly recorded in any continuously fossiliferous section in North America, perhaps exaggerating the perception of its rate of change. It is particularly well documented in groups that have received phylogenetic study such as canids and equids but seems evident in most well-represented clades and is marked by

accelerated cladogenesis and extinction and some immigration. In the borophagine canids (Wang et al. 1999) the event signals the extinction of nearly all Phlaocyonini and primitive Borophagini, accompanied by the origin and fundamental cladogenesis of all later borophagine clades in Cynarctina, Aelurodontina, and Borophagina. The horses (Hulbert and MacFadden 1991) show the early stages of cladogenesis of the Equinae, with both the Equini and Hipparionini in evidence in the late Hemingfordian. Similar events are seen in the Camelidae, Dromomerycidae, and merycodontine Antilocapridae. Chronologically this turnover takes place between the youngest dated early Hemingfordian deposits (about 17.5 Ma for the type area; MacFadden and Hunt 1998; Hunt, pers. comm., 2002) and the oldest dated late Hemingfordian (about 17 Ma for the Barstow sequence; MacFadden et al. 1990), or during a span of about 0.5 m.y.

The late Hemingfordian (He2) is defined by the earliest appearance of the immigrant petauristine squirrel (*Petauristodon*); the cricetine *Copemys* (toward the end of the span); the eomyid *Eomys*; the mustelines *Plionictis*, *Dinogale*, *Sthenictis*, and *Mionictis*; the Felidae (*Pseudaelurus*); and the rhinos *Peraceras*, *Teleoceras*, and *Aphelops*. It is characterized by the earliest appearance of the rabbit *Hypolagus*; the beaver *Anchitheriomys*; the amphicyonid *Pliocyon*; the Equinae (both Equini and Hipparionini clades, including *Merychippus*, *Protohippus*, *Parapliohippus*, and *Acritohippus*); the oreodont *Brachycrus*; the camelids *Paramiolabis*, *Nothotylopus*, and *Aepycamelus*; the dromomerycids *Sinclairiomeryx*, *Rakomeryx*, *Subdromomeryx*, *Dromomeryx*, and *Bouromeryx*; and the antilocaprids *Merriamoceros*, *Merycodus*, *Meryceros*, and *Submeryceros* and by the last appearance of *Phlaocyon*, *Metatomarctus*, *Prototomarctus*, *Barbouromeryx*, *Blickomylus*, and *Oxydactylus*.

## BARSTOVIAN

Originally the Barstovian had a very local rock and faunal basis ("Barstow Formation, San Bernardino County, California, and specifically on the fossiliferous tuff member in the Barstow syncline and its fauna"; Wood et al. 1941:12), but as discussed previously (Tedford et al. 1987:187–188) this left in limbo a large part of the biostratigraphic section between those containing late Hemingfordian faunas and those designated Barstovian. Tedford et al. (1987) recommended extending the Barstovian to the intervening strata to include the presence of taxa found in the principal correlative faunas (Pawnee Creek, Deep River, Virgin Valley, and Mascall, all reviewed in this and the previous work).

The Barstovian faunas identified across North America indicate the presence of zoogeographic provinces, especially the northwest, far west, midcontinent, and southeast and, with our extended coverage, into Central America as well. Radioisotopic and paleomagnetic data for faunal sequences scattered across these provinces have linked the Barstovian faunas of the Caliente Formation (California Coast Ranges), Barstow Formation (Northern Great Basin, California), Tesuque Formation (Southern Great Basin, New Mexico), and Pawnee Creek Formation (Northern Great Plains, Colorado) so that their respective biostratigraphies can be compared chronologically with some precision. In addition to some endemism, diversity changes in space can be documented as previously noted and as briefly explored by Tedford and Barghoorn (1997, figure 10). To a certain extent this partitioning of the mid-Miocene biota of North America presents problems in defining and characterizing the NALMAs and their subdivisions. To this is added the difficulty that there are remarkably few defining taxa noted for this interval.

In addition to these problems, the classic "standard" succession of Miocene faunas from the Great Plains of Nebraska is contained in a rock sequence that is notoriously incomplete, being broken by disconformities inherent in the style of sedimentation in valley-filling epiclastic deposits. There, much of the record is represented by lateral rather than the vertical accretion characteristic of the tectonically active basins of the western part of the continent. Thus the Hemingfordian–Barstovian transition in Nebraska is broken at the contact between the Sheep Creek and Olcott formations, which sharpens the perception of faunal change but does not show its transitional steps. Fortunately the sequences in the Tesuque Formation, New Mexico, and the Barstow Formation, California, contain these spans of time and show local biostratigraphic details.

A general problem in defining these medial Miocene NALMAs is the paucity of immigrant taxa in comparison with those of the early and late Miocene. Reliance on one or two such taxa, often with limited geographic ranges, limits the application of this method of recognition so useful for most NALMAs. This problem is further compounded by the largely anagenetic sequences of autochthonous taxa in this chrono-faunal interval so that characterizing faunas are best delineated at the specific rather than at the generic level.

Prominent among the allochthonous taxa suggested to define parts of the Barstovian are the earliest appearances of mammutid and gomphotheriid proboscideans in North America. These events need to be reevaluated ac-

ording to the data summarized earlier. Woodburne and Swisher (1995:347) indicated the presence of the Proboscidea (a tooth fragment, thought to be a mammutid) in the late Hemingfordian Massacre Lake Local Fauna of northwestern Nevada. Recently Reynolds and Woodburne (2001) identified proboscidean trackways in the Barstow Formation of the Calico Mountains, adjacent to the Mud Hills, Mojave Desert, California, that date to about 16.2 Ma, as in northwestern Nevada. These are the earliest known occurrences, but we have noted several other early Barstovian occurrences as well: California Coast Range (North Coalinga Local Fauna) and Sharktooth Hill Local Fauna, *Zygodolophodon* and possibly *Gomphotherium*, near the Relizian–Luisian boundary, 15.7 Ma; Columbia Plateau (Virgin Valley Fauna and High Rock Lake site, northwestern Nevada, Sucker Creek and Skull Springs faunas of eastern Oregon), *Zygodolophodon*, 15.7–14.7 Ma (note that the Sucker Creek occurrence lies below the Owyhee Basalt, 14.7 Ma *contra* Tedford et al. 1987); Northern Rocky Mountains (Deep River and Madison Valley faunas), *Zygodolophodon*, 16.2–14.6 Ma by local correlation; and Gulf Coastal Texas and Florida (Burkeville and Willacoochee Creek faunas), undetermined Proboscidea, chron C5Br, 16.2–15.3 Ma. Where the fossil material is well preserved, these early Barstovian remains seem to be largely *Zygodolophodon* sp., or *Z. merriami*. Thus the record indicates the presence of Proboscidea at the earliest in the late Hemingfordian, and thereafter mammutids are certainly known in the early Barstovian in the northwestern and coastal parts of North America. Confirmed records of gomphotheriids in the continental interior are limited to post–14.8 Ma strata. However, the record of *Gomphotherium* in Ixtapa, Chiapas, collected from strata located 200 m below a tuff dated as 15–16 Ma, and the North Coalinga specimen would extend the occurrence of such records into the early Barstovian or even late Hemingfordian. At their first occurrence, the Proboscidea are too rare to be used as a defining taxon for the beginning of the late Hemingfordian, and their actual first appearance probably is late in the span, but it may be useful to include *Zygodolophodon* as part of the characterizing fauna of the early Barstovian and to regard the midcontinent appearance of gomphotheriids as an event defining the beginning of the late Barstovian (Ba2) at about 14.5–14.8 Ma.

With all these problems in mind we propose the following revision of the Barstovian. It previously relied on two immigrant taxa for its definition: the hemicyonine *Plithocyon* and the cricetid *Copemys*. Both taxa were found together in the Barstow Formation at the base of the stratal span characterized by the Green Hills Fauna

(Steepside Quarry; Woodburne et al. 1990). Subsequent collecting of small mammals from the underlying Rak Division by Lindsay (1995) has carried *Copemys* (and most of the Green Hills rodent fauna) down into the top of the Rak Division and into the base of chron C5Cn2 (16.2 Ma; Berggren et al. 1995), only 164 m below and 0.2 m.y. older than the Steepside Quarry occurrence. This does not substantially change the chronologic position of this defining taxon but does serve as a warning (if one is needed) of the stratigraphic behavior of defining taxa, especially those that are rare or needing special collecting techniques for their detection.

The beginning of Barstovian time, and hence the early Barstovian (Ba1), can be defined by the first appearance of the hemicyonine *Plithocyon* and the definite occurrence of *Zygodolopodon*. It can be characterized by the first appearance of such autochthones as the mylagaulines *Umbogaulus*, *Pterogaulus*, and *Ceratogaulus* in the Great Plains and *Hesperogaulus* in the Columbia Plateau (Korth 1999a, 2000); the beaver *Monosaulax* (*sensu* Korth 1999c); the heteromyid rodents *Perognathus*, *Peridiomys*, and *Mojavemyx*; the borophagine canids *Cynarctus*, *Paratomarctus*, and in the west *Aelurodon*, *Paratomarctus*, *Carpocyon*, and *Protepicyon*; the procyonids *Probassariscus* and *Arctonasua*; the mustelid *Martes*; the equids *Megahippus*, *Calippus*, and *Hipparion*; the peccaries *Dyseohyus* and *Prosthennops*; the camelids *Procamelus* and *Rakomyx*; and the merycodontine antilocaprids *Ramoceros* and *Cosoryx*.

The early Barstovian (Ba1) contains the last occurrence of the Promylagaulinae (*Galbreathia*; Korth 1999b), the amphicyonids *Pliocyon* and *Amphicyon*, the Hesperocyoninae (*Osbornodon*), the borophagine canids *Cynarctoides* and *Euoplocyon*, the equids *Desmatippus* and *Parahippus*, the peccary *Cynorca*, the oreodonts *Brachycrus* and *Merychys*, the camelid *Paramiolabis*, the moschid *Problastomeryx*, the dromomerycids *Subdromomeryx* and *Rakomeryx*, the Stenomylinae (*Rakomyx*), and the antilocaprid *Merriamoceros*. The early Barstovian contains the limited occurrence of *Tomarctus*, *Psolidocyon*, *Tephrocyon*, *Edaphocyon*, *Rakomyx*, *Drepanomeryx*, and *Matthomeryx*.

The beginning of the late Barstovian (Ba2, medial Barstovian *sensu* Voorhies 1990a) is defined by the first appearance of gomphotheriid Proboscidea. Taxonomically identifiable gomphotheriid remains are largely confined to late Barstovian assemblages in North America, and many from California to Maryland, where datable, first occur around 14.8–14.5 Ma. This proboscidean datum is reasonably isochronous in the majority of instances, and we will continue to regard it as the most useful way

to define late Barstovian time, bearing in mind that this definition is also backed up by the characterizing fauna of that span.

Fortunately the Barstovian was based fundamentally on the faunal sequence in an unbroken sedimentary section in the Great Basin, thus avoiding the hiatuses that break the more fossiliferous midcontinent sections that have been so important in determining the faunal diversity of this phase of the Miocene of North America. There is a trade-off here: The Great Basin Barstovian faunas do not display a chrono-faunal relationship with those of the Great Plains; rather, they remain at lower diversity and have an endemic character that must reflect important environmental gradients across North America.

There is a gap between the early and late Barstovian faunas of the Great Plains, only partially filled by the faunal sequence in the Pawnee Creek Formation of northeastern Colorado. In Nebraska these phases of the Barstovian are not superposed and are separated from one another by 300 km. The time value of the hiatus between them is difficult to judge, but the late Barstovian sequence in the Valentine Formation of north-central Nebraska must begin shortly before the dated Hurlbut Ash (13.6 Ma) near the base of the unit. This equates on geochronologic grounds with the upper part of the Barstow faunal span in California (about the level of the Hemicyon Tuff, chron C5An, 14.0 Ma). Thus the true biochrons of the taxa first appearing at the base of the Valentine Formation (e.g., the gomphotheriid Proboscidea) may extend into the hiatus, as these proboscideans do elsewhere (14.8 Ma at Barstow, older than 14.6 Ma in New Mexico, 14.4 Ma in Colorado).

Because of the zoogeographic diversity of the Barstovian assemblages involved in defining and characterizing the late Barstovian (Ba2), we indicate with an asterisk the taxa limited to the Great Plains and Gulf Coast provinces. The late Barstovian is defined by the earliest appearance of the Gomphotheriidae, the hedgehogs *Lanthanotherium* and *Untermannerix*\*, the ochotonids *Hesperolagomys* and *Russellagus*\*, the limited occurrence of the zapodid *Megasmithus*\*, the mephitine *Pliogale*, and the amphicyonid *Pseudocyon*\*. The characterizing fauna of Ba2 includes the earliest occurrences of the Leporinae *Alilepus* and *Pronotolagus* (Voorhies and Timperley 1997); the mylagauline *Mylagaulus* (Korth 2000); the beaver *Eucastor* (*E. tortus*); the geomyoid *Lignimus*; the eomyid *Leptodontomys*; the cricetid *Tregomys*\*; the borophagine canids *Cynarctus saxatilis*\*, *Aelurodon* (*Strobodon*)\*, and *A. ferox*\*; the ursid *Ursavus pawniensis*; the amphicyonid *Ischyrocyon*; the horses *Pliohippus* (*P. mirabilis*), *Pseudhipparion*\*, *Neohipparion* (*N. coloradense*), and *Cormohipparion*\*

ion; the oreodont *Ustatochoerus medius*; the moschid *Longirostromeryx*\*; the dromomerycid *Procranioceras*; and the merycodontine *Ramoceros*. The late Barstovian contains the last occurrence of the erinaceid *Parvericius*, the mylagauline *Umbogaulus* (Korth 2000), the beavers *Monosaulax* and *Anchitheriomys*, the ochotonid *Oreolagus*, the aplodontid *Allomys*, the zapodid *Megasmithus*, the Chalicotheriidae (*Moropus*), the equids *Archaeohippus* and *Anchitherium*, the oreodont *Ticholeptus*, the protoceratid *Prosynthetoceras*, the Leptomerycidae (*Pseudoparablastomeryx*), and the dromomerycids *Dromomeryx* and *Bouromeryx*.

### CLARENDONIAN

The Clarendonian (Wood et al. 1941:12) originally was based on the “Clarendon local fauna (and member?) near Clarendon, Donley County, Panhandle of Texas” and the age limits suggested by the list of principal correlatives, “Burge, Big Spring Canyon, Fish Lake Valley, Ricardo,” and also by the correlation chart and glossary entries that constrain the upper limit of the age to the “Minnechaduzza local fauna, late Clarendonian or, possibly, early Hemphillian” (Wood et al. 1941:26). Superposed younger faunas from the “Xmas and Kat Quarries, local channel fauna or faunas in the Ash Hollow Formation, later than the Minnechaduzza, northeastern Cherry County, Nebraska” (Wood et al. 1941:30), were shown as basal Hemphillian on the correlation chart. Thus in terms of the superposed sequence in north-central Nebraska, the Burge Fauna from the uppermost part of the Valentine Formation delimited the oldest and the Minnechaduzza Local Fauna from the lower part of the Ash Hollow Formation was considered the youngest part of the Clarendonian. In his review of the Burge and Minnechaduzza faunas, Webb (1969) followed Wood et al. (1941) in including the Burge Fauna in the Clarendonian. He also included the “Leptarctus B” site in his concept of the Minnechaduzza Fauna, but in his discussion of the antilocaprine, *Plioceros flobairi*, from that site, he indicated that “the exact age of material from this quarry is in doubt. Several species indicative of a late Clarendonian age occur, but there are other unusual forms that either indicate a different ecological situation or a younger age. A transitional Clarendonian–Hemphillian age is suggested” (Webb 1969:172–173). Much later, Skinner and Johnson (1984), based on a regional stratigraphic synthesis, separated the rocks above the Cap Rock Member (containing the Minnechaduzza Fauna) as the Merritt Dam Member of the Ash Hollow Formation and listed many sites in this unit, including the *Leptarctus* Quarry,

and the Xmas and Kat quarries as containing a post-Minnechaduzza fauna. The fauna of these younger deposits has a strong chronofaunal relationship with the Burge and Minnechaduzza assemblages. Tedford et al. (1987) included the post–Cap Rock faunas in their concept of the Clarendonian without specific discussion, but their intent was clear from their correlation chart and is followed here.

On the other hand, Tedford et al. (1987:189) excluded the fauna of the Burge Member of the Valentine Formation from their concept of the Clarendonian on the grounds that “the hiatus between the Valentine and Ash Hollow formations is accompanied by perceivable faunal change, and this provides the basis for an objective characterization of the Clarendonian.” The Burge Member is separated from adjacent units by disconformities that contain hiatuses that break chronofaunal continuity. In their view the faunal relationships of assemblages from the Burge and older Valentine deposits were more closely related than they were with the Ash Hollow faunas. Also on the matter of definition, always difficult in a span of low immigration, the first appearance of the nimravid *Barbourofelis* seemed more recognizable (it has a unique skeleton) despite the inherent rarity of carnivores in any fossil assemblage. This genus has been recognized at several sites scattered across the North American midcontinent. On this basis *Barbourofelis* seemed a plausible defining taxon for the Clarendonian.

Subsequently Voorhies (1990a:142) raised objections to this truncation of the Clarendonian on several grounds. First, the Burge Fauna was originally included in the Clarendonian as a “principal correlative” (Wood et al. 1941). Second, the degree of faunal change between the Burge and Ash Hollow “does not appear to be more remarkable than that which separate the Burge faunas from those collected at lower stratigraphic levels in the Valentine Formation.” Third, the presence of *Barbourofelis* at the base of the Ash Hollow did not seem an intrinsically more useful marker than the presence of the gelocid *Pseudoceras* at the base of the Burge. Finally, although not strongly advocated, the short Clarendonian of Tedford et al. (1987) is “severely curtailed in stature.” Of these arguments the most telling is the typology, which we have otherwise tried to preserve, and the arbitrariness of any change in boundary position within a chronofaunal sequence. The choice of a definitive taxon in this type of faunal sequence usually is subordinate to the characterizing fauna in age determination.

Recent studies of the geochronology of faunal sequences provide means for determining the tempo of faunal change and for estimating the time value of hiatuses

that break faunal sequences. In the Mojave Desert region of California, the principal reference sequence for the Barstovian extends upward into deposits lying in chron C5ABn at about 13.4 Ma (in agreement with ash dates at that level). The Ricardo Fauna, “principal correlative” of the Clarendonian, extends downward (as the Burge equivalent Iron Canyon Fauna) into deposits older than 12 Ma and probably 12.5 Ma (magnetostratigraphy and ash dates; Whistler and Burbank 1992; Perkins et al. 1998). The striking turnover implied by this faunal and geochronologic data suggests a major event in western North America contemporaneous with the little-perturbed chronofaunal progression characteristic of the midcontinent. The fauna that emerges from this event in the Great Basin is compositionally like those of the Great Plains Clarendonian. The fact that this break takes place below the Burge-equivalent local faunas at the base of the Ricardo succession is another reason to place the Barstovian–Clarendonian boundary in its typologic position.

We thus return to the typologic boundary for definition of the beginning of Clarendonian time, and for lack of other defining taxa we use the first appearance of the gelocid *Pseudoceras* for this purpose. It is morphologically distinct in dentition and extends from the northern Great Plains around the Gulf of Mexico and into Honduras but is geographically limited to east of the Rocky Mountains. It is usually rare but has a well-defined biochron—early Clarendonian (Cl1) to late early Hemphillian (Hh2)—and occurs only as a relict in the latest Hemphillian (Hh4) of Florida. The characterizing fauna of the early Clarendonian (Cl1) includes the first appearances of the beavers *Eucastor planus* and *E. dividerus*; the gomphotheriids *Eubelodon* and *Megabelodon*; the borophagine canids *Cynarctus voorhiesi*, *Aelurodon* (*Strobodon*) *stirtoni*, and *Paratomarctus euthos*; the equids *Plihippus pernix*, *Cormohipparion occidentale*, *Pseudhipparion retrusum*, *Protohippus supremus*, and *Megahippus matthewi*; the oreodont *Ustatochoerus major* (including *U. profectus* *fide* Stevens, pers. comm., 2002); and the dromomerycid *Cranioceras*.

The medial Clarendonian (Cl2) is defined by the reappearance of the Nimravidae (*Barbourofelis whitfordi* and *Barbourofelis osborni*) in North America. It is characterized by the following first appearances of autochthonous clades: the borophagine canids *Borophagus* (*B. littoralis*) in western North America and *Epicyon* (*E. saevus*), *Carpocyon robustus*, *Aelurodon taxoides*, and *Cynarctus crucidens* in the midcontinent; the equids *Pseudhipparion gratum*, *Hipparion tehonense*, *Neohipparion affine*, and *Dinohippus*; the oreodont *Ustatochoerus californicus*; the protoceratid *Synthetoceras*; and the limited occurrence of

*Proantilocapra*. A few taxa show last appearances in the medial Clarendonian: the mustelid *Brachypsalis*, the ursid *Hemicyon*, the equids *Merychippus* (s.s.) and *Megahippus*, the rhino *Peraceras*, and the protoceratine *Paratoceras*.

The late Clarendonian (Cl3) is defined by the first appearance of the shovel-tusk gomphotheriid *Platybelodon*, the ischyricine mustelid *Hoplictis* (= *Beckia*), and the simocyonine ailurid *Actiocyon*. The characterizing fauna includes the first appearances of the beaver *Hystricops*; the sigmodontine cricetid *Antecalomys*; the marmot *Marmota*; the procyonids *Protoprocyon* and *Paranasua*; the Canini (?*Eucyon*); the nimravids *Barbourofelis morrisoni* and *Barbourofelis lovei*; the felids *Nimravides thinobates* and *Nimravides galiani*; the equids *Pseudhipparion skinneri*, *Hipparion forcei*, *Neohipparion trampasense*, *Protohippus gidleyi*, and *Calippus cerasinus*; and the antilocaprine *Plioceros*.

The close of the Clarendonian is marked by an extinction event removing a number of taxa characteristic of the medial Miocene. At the same time, new autochthones appear and faunal diversity begins to decline toward the comparatively depauperate assemblages of the late Miocene. The following taxa have their last appearance in the late Clarendonian: the Erinaceidae (*Untermannerix* and *Metechinus*); the beaver *Eucastor*; the sciurids *Protospermophilus* and *Petauristodon*; the geomyids *Phelosacomys* and *Mioheteromys*; the eomyid *Pseudotheridomys*; the cricetine *Tregomys*; the ochotonids *Hesperolagomys* and *Russellagus*; the mustelids *Hoplictis* and *Mionictis*; the Amphicyonidae (*Ischyrocyon* and *Pseudocyon*); the borophagine canids *Aelurodon*, *Paratomarctus*, and *Cynarctus*; the Anchitheriinae (*Hypohippus*); the Merycoidodontidae (*Ustatochoerus*); the miolabine camels *Miolabis* and *Nothotylopus*; the Moschidae (*Blastomeryx*, *Parablastomeryx*, and *Longirostromeryx*); the dromomerycid *Cranioceras*; and the Merycodontinae (*Cosoryx*).

## HEMPHILLIAN

The principal characterization of the Hemphillian shifts to the southern Great Plains, where a sequence was chosen in the “Hemphill Member of the Ogallala, which includes both the Hemphill Local Fauna from the Coffee Ranch Quarry and the Higgins Local Fauna, Hemphill [and Lipscomb] County, Panhandle of Texas” (Wood et al. 1941:12). This was a particularly fortunate choice because superposition of the Hemphill Member on deposits containing Clarendonian faunas is present, as is a fairly detailed biostratigraphic sequence in the Hemphill. Schultz (1990) gives a very useful review of our understanding of this sequence, and with Voorhies’s (1990a) discussion of the Nebraska Hemphillian, the biochronol-

ogy of this interval in the Great Plains has been conveniently reviewed.

The Hemphillian, approximately the late Miocene and a little more than 4 m.y. in duration, is marked by continual loss of autochthonous taxa and by a rising immigration rate marked by the first waifs from South America; diverse rodents, insectivores, and carnivores; and some ungulates from Asia. Most of the remaining elements of the "Miocene" chronofauna are extinct by 6–7 Ma, so the Hemphillian can be conveniently divided by that turnover event, as previously noted (Tedford et al. 1987:191–192). Hemphillian faunas seem more homogeneous in composition across North America than earlier NALMAs, although the Great Basin and West Coast show particularly reduced horse diversity, as in older parts of the Miocene. The Gulf Coast retains some relictual Clarendonian taxa, and the Great Plains (including Mexico and Central America) show the greatest ungulate diversity.

Immigrant taxa whose first appearances in North America have been established in the Great Plains can be used (as in Voorhies 1990a) to distinguish four phases of the Hemphillian, with important turnover events marking the end of the late early (Hh2) and the close of the Hemphillian.

The beginning of Hemphillian time (Hh1) is defined by the first appearance of the Edentata (Megalonychidae, *Pliometanastes*), the prometheomyine murid *Paramicroscoptes*, the eomyid *Kansasimys*, the cricetodontine murid *Pliotomodon*, the desmanine mole *Lemoynea*, and the nectogaline shrew *Crusafontina* (including *Anoureneomys*). First appearances in the autochthonous characterizing fauna include the archaeolagine *Hypolagus vetus*; the beaver *Dipoides*; the geomyid *Pliosacomys*; the sigmodontine murid *Paronychomys*; the badger *Pliotaxidea*; the borophagine canid *Borophagus pugnator* and its coexistence with *Epicyon haydeni*; the Vulpini (*Vulpes stenognathus*) and Canini (*Eucyon davisii*); the gomphothere *Rhynchotherium*; the rhino *Teleoceras fossiger*; the equids *Neohipparion leptode*, *Nannippus*, *Hippidion*, and *Pliohippus nobilis*; the llamine *Alforjas* and cameline *Megatylopus gigas*; and the antilocaprine *Osbornoceras*.

Late early (or medial; Voorhies 1990a) Hemphillian (Hh2) is defined by the first appearance of the sloth *Thiobadistes* (Mylodontidae), the bear *Indarctos*, the ailurid *Simocyon*, the galictine mustelid *Lutravus*, the otter *Enhydritherium*, the mellivorine *Eomellivora*, the felid *Machairodus*, and the Bovidae (*Neotragoceros*). The characterizing fauna includes the first appearances of the beaver *Dipoides stirtoni*; the murid *Goniodontomys*; the tremarctine bear *Plionarctos*; the borophagine canid *Borophagus secundus*; the felid *Nimravides catocopsis*; the

shovel-tusked gomphothere *Amebelodon*; the rhino *Aphelops mutilus*; the horses *Dinohippus leidymanus*, *Astrohippus ansae*, *Pliohippus spectans*, and *Neohipparion eurystyle*; the peccaries *Prosthennops serus* and *Platygonus*; and the antilocaprines *Sphenophalos*, *Ilingoceros*, and *Texoceros*.

During the late early Hemphillian (Hh2) a number of taxa reached the limit of their chronologic ranges, including some characteristic of the Clarendonian. The list of last occurrences includes the beaver *Hystricops*; the eomyid *Leptodontomys*; the mustelids *Leptarctus* and *Sthenictis*; the felid *Nimravides*; the Nimravidae (*Barbourofelis*); the horses *Protohippus*, *Hipparion*, *Calippus*, and *Pliohippus*; the Protolabinae (*Protolabis* and *Micheinia*); the llamine *Aepyamelus*; the cameline *Procamelus*; the dromomerycid *Yumaceras*; and the antilocaprines *Osbornoceros* and *Plioceros*. Most of the early Hemphillian immigrants also do not survive except *Machairodus*, *Neotragoceros*, and the sloths.

The late Hemphillian (Hh3) is defined by several immigrants: the sloth *Megalonyx*, the pika *Ochotona*, the arvicoline murid *Prosomys*, the zapodid *Pliozapus*, the beaver *Castor*, the ursid *Agriotherium*, the wolverine *Plesiogulo*, and the cat *Felis*. The characterizing fauna of the early late Hemphillian (Hh3) includes first occurrences of the mole *Scalopus* (*Hesperoscalops*); the horned mylagauline *Ceratogaulus hatcheri*; the large marmot *Paenemarmota*; the sigmodontine cricetids *Calomys* and *Prosigmodon*; the cricetines *Peromyscus*, *Galushamys*, *Onychomys*, and *Paraneotoma*; the geomyid *Pliogeomys*; the heteromyid *Prodipodomys*; the raccoon *Procyon*; the tremarctine bear *Plionarctos edensis*; the borophagine canid *Borophagus parvus*; the equids *Dinohippus interpolatus*, *Nannippus aztecus*, *Nannippus lenticularis*, and *Onohippidion* (*O. galushai*); and the dromomerycid *Pediomyerx*. Taxon extinctions during this interval include the immigrants *Platybelodon*, *Prosomys*, *Pliozapus*, *Eomellivora*, *Lutravus*, and *Pliogale* and the autochthones *Amebelodon*, *Alforjas*, *Sphenophalos*, *Ilingoceros*, and *Texoceros*.

Chronologically the latest (or late late) Hemphillian (Hh4) just extends across the Miocene–Pliocene boundary (5.3 Ma) to 4.6–4.9 Ma, as shown by the ash date at Rancho El Ocote, Guanajuato, the magnetostratigraphy at the Yepómera site in Chihuahua, Mexico, and the early Blancan ash date from Nevada (data presented earlier). This final phase of the Hemphillian begins at the chron C3r–C3An boundary at about 5.9 Ma (Berggren et al. 1995). It closes with the extinction of most of the clades characteristic of the late Hemphillian: the families Plesiosoricidae, Mylagaulidae, Rhinocerotidae, Protoceratidae, and Dromomerycidae; the wolverine *Plesiogulo*; the badger *Pliotaxidea* (survives as *Taxidea*); the borophagine

canids *Carpocyon* and *Borophagus secundus*; the sabre-cat *Machairodus*; the proboscidean *Gomphotherium*; the equids *Dinohippus* (survives as *Equus*), *Astrohippus*, and *Neohipparion*; and the antilocaprine *Hexameryx*.

The defining taxa of this final phase of the Hemphillian (Hh4) include the flying squirrel *Miopetaurista*, the arvicoline *Propliophenacomys*, the weasel *Mustela*, the mustelids *Trigonictis* and *Sminthosinis*, the otter *Lutra*, the cat *Megantereon*, and the Odocoileini (*Eocoileus*; Webb 2000). The characterizing fauna includes the leporine *Lepoides*; the marmots *Marmota* (*M. vetus*) and *Paenemarmota sawrockensis*; the gopher *Thomomys* (*Plesiothomomys*); the cricetines *Baiomys* and *Repomys* (*R. gustelyi*); the canids *Borophagus hilli* and *Borophagus dudleyi*; the skunk *Buisnictis*; the horses *Dinohippus mexicanus*, *Neohipparion* cf. *eurystyle*, and *Astrohippus stocki*; the tayassuid *Mylohyus*; the camel *Megacamelus*; the antilocaprine *Subantilocapra*; and the last protoceratid, *Kyptoceras*. Most of the ungulates on this list have biochrons limited to the late late Hemphillian. A number of lineages extended their ranges into the late Hemphillian but limited their geographic ranges to the southern Great Plains, Gulf Coastal Plain, and Mexico, including a small species of *Gomphotherium*, the Protoceratidae (*Kyptoceras*), the Gelocidae (*Pseudoceras*), and the hipparionine equids *Cormohipparion emsleyi*, *Nannippusa ztecus*, and *Pseudhipparion simpsoni*, the last reaching as far north as Buis Ranch, Oklahoma, and MacPherson, Kansas.

## ACKNOWLEDGMENTS

A summary such as this that builds on a previous version owes much to our former collaborators, some of whom (M. F. Skinner, R. W. Fields, and T. Galusha) have since passed away. We dedicate this work to their memory.

The present compilation was put together in much the same way as the previous, with the senior author distilling the combined efforts at his colleagues into a synthesis that represents a largely consensus effort. We have been assisted in important ways by the following colleagues who contributed data and ideas to this work: T. Fremd, R. Hulbert, W. W. Korth, E. B. Lander, E. L. Lundelius, G. S. Morgan, M. S. Stevens, X.-M. Wang, and M. O. Woodburne.

Alejandra Lora was responsible for bringing the text into order, and Edward Heck translated pencil sketches into clear graphics.

## APPENDIX

Selected radiometric and fission-track dates used to calibrate the correlation charts (reference numbers on figure 6.2). Dates published before 1979 were recalculated using

International Union of Geological Sciences constants following Dalrymple (1979). This list includes most material previously published by Tedford et al. (1987), augmented by relevant dates obtained since that writing.

NO.	DATE	UNIT	REMARKS
	(MA, ± SD)	DATED	
1	10.3	Moraga Fm. <sup>a</sup>	Revised date of basal andesitic basalt flow (see KA 993, 1001) <sup>b</sup>
2	10.2	Siesta Fm. <sup>b</sup>	Plagioclase from tuff, base of unit, KA 829
3	7.9	Bald Peak Volcanics <sup>b</sup>	Basalt, minimum date, KA 1003
4	5.3 ± 0.1	Pinole Tuff <sup>f</sup>	Feldspar from tuff, a few feet above fauna, KA 1005
5	5.9 ± 0.6; 6.3 ± 0.1	Sycamore Fm. <sup>c,d</sup>	Tuff near top of unit
6	5.5 ± 0.2	Pinole Tuff <sup>f</sup>	Tuff at top of Sycamore Fm., immediately beneath Hemme Hills LF
7	4.1 ± 1.0; 4.6 ± 0.5	Lawlor Tuff <sup>c</sup>	Tuff in Tassajara Fm.
8	22.1–22.9	Dacite, dacite agglomerate <sup>e</sup>	Intertongues with top of Tecuya Fm., minimum date for faunas, KA 2114–5, 2166, 2175
9	28.20 ± 0.2	Willard Canyon Tuff <sup>f</sup>	Top of fossiliferous interval containing South Mountain Fauna
10	16.5 ± 1.3	Caliente Fm. <sup>e</sup>	Lowest Triple Basalt, KA 2127
11	14.6 ± 0.6; 14.8 ± 0.8	Caliente Fm. <sup>e</sup>	Uppermost Triple Basalt, KA 2116, 2125
12	13.4 ± 0.14	Dry Canyon Tuff <sup>g</sup>	Ar–Ar on biotite, revises age of “Dated Tuff,” unit 10 in Tedford et al. (1987)
13	22.9 ± 0.4	Hector Fm. <sup>h</sup>	K–Ar on biotite from tuff near base Hector Fm., North Cady Mountains; maximum date for overlying faunas
14	20.3 ± 0.7	Saddleback Basalt <sup>i</sup>	Boron LF 146 m above top of basalt, B-4
15	21.6	Hector Fm. <sup>j</sup>	Tuff near top of stratigraphic occurrence of Black Butte Mine LF, KA 2223
16	18.5 ± 0.2	Peach Spring Tuff <sup>k</sup>	Interbedded with rocks containing Hackberry Fauna
17	18.5 ± 0.2	Peach Springs Tuff <sup>h</sup>	K–Ar on sanidine from tuff stratigraphically between local faunas, northeastern Cady Mountains
18	17.8	Wildhorse Mesa Tuff <sup>l</sup>	Interbedded with top of unit yielding Hackberry Fauna
19	16.3 ± 0.3	Rak Tuff <sup>m</sup>	K–Ar date on biotite from tuff near base of Rak Division, Barstow Formation
20	15.8 ± 0.02	Oreodont Tuff <sup>m</sup>	K–Ar date on biotite from tuff, 40 m above base of Green Hills Division, Barstow Fauna
21	14.8 ± 0.06	Dated Tuff <sup>m</sup>	Ar–Ar date on biotite from tuff near base of First Division of Barstow Fm.
22	14.0 ± 0.09	Hemicyon Tuff <sup>m</sup>	Ar–Ar date on biotite from tuff about 80 m above base of First Division, Barstow Fm.

NO.	DATE (MA, ± SD)	UNIT DATED	REMARKS	NO.	DATE (MA, ± SD)	UNIT DATED	REMARKS
23	13.4 ± 0.2	Lapilli Tuff <sup>m</sup>	K-Ar date on biotite from tuff about 30 m below top Barstow Fm.	46	28.7 ± 0.07	Picture Gorge Ignimbrite <sup>s</sup>	Ar-Ar date, John Day Fm.
24	12.6 ± 0.1	Cronese Tuff <sup>g</sup>	Pumice, just above fossil site	47	27.89 ± 0.57	Deep Creek Tuff <sup>g</sup>	Ar-Ar date, John Day Fm.
25	12.07 ± 0.04	Cougar Point Tuff V <sup>n</sup>	Ar-Ar date, type locality northeast Nevada, identified by element analysis; about 100 m above base of Dove Spring Fm.	48	27.18 ± 0.13	Biotite Tuff <sup>g</sup>	Ar-Ar date, John Day Fm.
26	10.94 ± 0.03	Cougar Point Tuff XIII <sup>n</sup>	Ar-Ar date type locality northeastern Nevada, identified by element analysis; just above upper basalt, Dove Spring Fm.	49	25.9 ± 0.31	Tin Roof Tuff <sup>g</sup>	Ar-Ar date, John Day Fm.
27	8.50 ± 0.13	Unnamed tuff <sup>n</sup>	K-Ar date (86CS-R4), 1400 m above base of Dove Spring Fm.	50	22.6 ± 0.13	Across the River Tuff <sup>g</sup>	Ar-Ar date, John Day Fm.
28	17.4 ± 0.3; 17.6 ± 0.3	Fraction Tuff <sup>o</sup>	K-Ar date, sanidine and biotite, respectively, U.S. Geological Survey (M) 11549-1	51	15.77 ± 0.04	Mascall Fm. <sup>g</sup>	Ar-Ar date on plagioclase from tuff 25 feet below base of Unit 5 of Downs (1956) and below fauna
29	16.6 ± 0.4	Brougher Dacite <sup>p</sup>	K-Ar date, sanidine and biotite, 659-66 <sup>q</sup>	52	15.79 ± 0.07	Unnamed tuff <sup>g</sup>	Ar-Ar date on sanidine, from tuffaceous fossil horizon, Skull Springs Fauna; Red Basin
30	15.16 ± 0.063	Tonopah Tuff <sup>g</sup>	K-Ar date on plagioclase from just below fossil quarry	53	12.4	Juntura Fm. <sup>l</sup>	Basalt at top of lower member, Juntura Fm., above Stinking Water Flora, KA 1240
31	14.89 ± 0.53	Unnamed ash <sup>g</sup>	K-Ar date on plagioclase; ash overlies Stewart Springs localities	54	9.4 ± 0.6	Drewsey Fm. <sup>u</sup>	Sanidine from Welded Tuff of Devine Canyon, lowest unit in Drewsey Fm., beneath local faunas
32	11.74 ± 0.03	Esmeralda Fm. <sup>g</sup>	Ar-Ar date on biotite, tuff from middle of fossiliferous section <sup>b</sup>	55	7.1 ± 1.09	Drinkwater Basalt <sup>v</sup>	Whole rock date; minimum age for Bartlett Mountain and other local faunas in the Drewsey Fm.
33	11.58 ± 0.05	Esmeralda Fm. <sup>g</sup>	Ar-Ar date on biotite, tuff just beneath small mammal site <sup>b</sup>	56	6.6 ± 0.1; 6.8 ± 0.2	Rattlesnake Fm. <sup>q</sup>	Sanidine from ignimbrite member of Rattlesnake Fm., above fauna
34	11.5	Coal Valley Fm. <sup>b</sup>	K-Ar date on biotite, tuff near base of fossiliferous section, KA 414	57	29.5 ± 2.8	Unnamed ignimbrite <sup>w</sup>	Fission-track from zircon in welded tuff unconformably beneath Cabbage Patch Fm.
35	11.1	Coal Valley Fm. <sup>b</sup>	K-Ar date on biotite, tuff near middle of fossiliferous section	58	28.26 ± 0.05	Chimney Rock Perrierite Ash <sup>x</sup>	Ar-Ar date on sanidine, near base of Gering Fm.
36	9.6	Coal Valley Fm. <sup>b</sup>	K-Ar on biotite from tuff below beds containing Smiths Valley Fauna, KA 485	59	28.11 ± 0.18	Roundhouse Rock Pisolithic Ash <sup>x</sup>	Ar-Ar date on sanidine and plagioclase, near top of Gering Fm.
37	6.76 ± 0.06; 7.20 ± 0.07	Unnamed basalt <sup>q</sup>	Whole rock K-Ar dates from basalts unconformably overlying Wassuk Group, KA 2365 and 2369, respectively	60	22.9 ± 0.08	Agate Ash <sup>y</sup>	K-Ar biotite from tuff near base of Harrison Fm.
38	4.96	Unnamed tuff <sup>f</sup>	Tuff in the base of section yielding Blancan mammals	61	19.2 ± 0.5	Eagle Crag Ash <sup>z</sup>	Fission-track on zircon from ash just above Agate Springs LF
39	16.58 ± 0.12	Steens Basalt <sup>g</sup>	Ar-Ar whole rock date on upper flow in type area southeastern Oregon; minimum age for genetically related basalt at site of Massacre Lake LF	62	17.4 ± 0.08	Split Rock Ash <sup>y</sup>	K-Ar date on sanidine from an ash (unit 2 of Love 1961:19) in stratal span of Split Rock Fauna
40	16.47 ± 0.04	Tuff of Big Basin <sup>g</sup>	Ar-Ar date on anorthoclase; ash flow immediately above rocks with Massacre Lake LF	63	16.36 ± 0.07; 16.4 ± 0.07	Sheep Creek Tuff <sup>g,y</sup>	Ar-Ar date on anorthoclase from the third ash in the Sheep Creek Fm. type section above fauna
41	16.13 ± 0.03	Canyon Rhyolite <sup>g</sup>	Ar-Ar on anorthoclase from rhyolite flow that underlies Virgin Valley Fauna	64	13.55 ± 0.09	Hurlbut Ash <sup>g</sup>	Ar-Ar date on glass, ash above Norden Bridge LF, Cornell Dam Mbr., Valentine Fm.
42	15.84 ± 0.13	Virgin Valley Pumice <sup>g</sup>	Ar-Ar date on sanidine, lower part of Virgin Valley Fm. below fossil sites	65	12.18 ± 0.12	Swallow Ash <sup>g</sup>	K-Ar date on glass, ash near local base, Cap Rock Mbr., Ash Hollow Fm.
43	15.18 ± 0.76	Virgin Valley Tuff <sup>g</sup>	Ar-Ar date on anorthoclase, overlies main fossil horizon	66	11.55 ± 0.12	Davis Ash <sup>g</sup>	K-Ar date on glass, ash near base Merritt Dam Mbr., Ash Hollow Fm.
44	14.49 ± 1.14	Fly Tuff <sup>g</sup>	K-Ar date on plagioclase, 8 m above main fossil horizon	67	8.0 ± 0.7	Unnamed ash <sup>aa</sup>	Fission-track from glass in Ogallala lectotype section of Elias, <sup>bb</sup> beneath Feltz Ranch LF
45	29.75 ± 0.02	AB Tuff <sup>g</sup>	Ar-Ar date, John Day Fm.	68	6.6 ± 0.3	Ash Hollow Fm. <sup>g</sup>	Fission-track from glass near top of type section, Garden County, Nebraska
				69	5.0 ± 0.2	Santee Ash <sup>cc</sup>	Fission-track from glass, ash overlies Santee LF



NO.	DATE (MA, + SD)	UNIT DATED	REMARKS
70	14.36 ± 0.18	Pawnee Creek Fm. <sup>g</sup>	Ar–Ar date on glass, ash near middle of section underlies first local appearance of Proboscidea
71	6.8 ± 0.2; 6.5; 6.8 + 0.03	Coffee Ranch Ash <sup>y,dd,ee</sup>	Fission-track date on zircon from ash just above Coffee Ranch Quarry; age revised using magnetostratigraphy, <sup>ee</sup> maximum date; Ar–Ar date on sanidine <sup>y</sup>
72	15.3 ± 0.05; 15.45 ± 0.06	No. 4 White Ash <sup>y,ff</sup>	Ar–Ar date on sanidine; ash in upper part of Skull Ridge Mbr., Tesuque Fm., 3 m above White Operation Quarry <sup>gg</sup>
73	9.6 ± 0.2	Lobato Basalt <sup>hh</sup>	Whole rock K–Ar age from a flow assigned to the Lobato field; minimum date for lower part of Chamita Fm.
74	7.7 ± 0.3	Lower Tuffaceous Zone <sup>ff</sup>	Ar–Ar on hornblende, Lower Tuffaceous Zone, Chamita Fm., maximum date for fauna
75	6.93 ± 0.05; 6.78 ± 0.03;	Upper Tuffaceous Zone <sup>ff</sup>	Ar–Ar on sanidine, samples from lower and upper part of Upper Tuffaceous Zone, Chamita Fm., bracketing fauna in ash
76	13.64 ± 0.09	Cerro Conejo Fm. <sup>ii</sup>	K–Ar on biotite from ash near top of fossiliferous interval, Ceja del Rio Puerco
77	11.3 ± 0.10	Cerro Conejo Fm. <sup>jj</sup>	Ash tentatively identified with Cougar Point Tuff XI, <sup>ii</sup> Ar–Ar dated at type locality, northeastern Nevada
78	23	Basalt <sup>kk</sup>	Whole rock K–Ar date on basalt in base of the Delaho Fm., underlies fauna
79	23.3	Basalt <sup>kk</sup>	Whole rock K–Ar date on basalts at base of Closed Canyon Fm., brackets fauna
80	19.3 ± 0.3; 20.3 ± 0.3	Etla Tuff <sup>ll</sup>	K–Ar dates on biotite and plagioclase, respectively
81	15.3 ± 0.8; 16.0 ± 0.8	Unnamed <sup>ll</sup>	K–Ar dates on biotite and plagioclase, respectively; pyroclastics underlying fauna
82	15.02 ± 0.35; 16.02 ± 0.53	Unnamed lower tuff <sup>ll</sup>	K–Ar dates on biotite and plagioclase, respectively; tuff overlies fauna
83	15.0 ± 0.8; 16.47 ± 0.71	Yautepec Tuff <sup>ll</sup>	K–Ar dates on biotite and plagioclase, respectively; tuff underlies faunal sites
84	12.12 ± 0.19; 12.24 ± 0.19	Unnamed upper tuff <sup>ll</sup>	K–Ar dates on biotite and plagioclase, respectively; tuff overlies fauna
85	4.6	Unnamed <sup>mmm</sup>	Fission-track age of tuff, overlying fauna

Fm., Formation; LF, Local Fauna; Mbr., Member.

#### References:

<sup>a</sup>D. E. Savage, pers. comm., 1973.

<sup>b</sup>Evernden et al. 1964.

<sup>c</sup>Sarna-Wojcicki 1976.

<sup>d</sup>Bartow et al. 1973.

<sup>e</sup>Turner 1970.

<sup>f</sup>Mason and Swisher 1989.

<sup>g</sup>Swisher 1992.

<sup>h</sup>Woodburne 1998.

<sup>i</sup>Armstrong and Higgins 1973.

<sup>j</sup>Woodburne et al. 1974.

<sup>k</sup>Nielson et al. 1990.

<sup>l</sup>Reynolds et al. 1995.

<sup>m</sup>MacFadden et al. 1990.

<sup>n</sup>Perkins et al. 1995, 1998.

<sup>o</sup>Silberman and McKee 1972.

<sup>p</sup>Albers and Stewart 1972.

<sup>q</sup>Parker and Armstrong 1972.

<sup>r</sup>Lindsay et al. 2002.

<sup>s</sup>Swisher 1992.

<sup>t</sup>Evernden and James 1964.

<sup>u</sup>Greene 1973.

<sup>v</sup>Greene et al. 1972.

<sup>w</sup>C. W. Naeser, pers. comm., 1973.

<sup>x</sup>Tedford et al. 1996.

<sup>y</sup>Izett and Obradovich 2001.

<sup>z</sup>Hunt et al. 1983:366.

<sup>aa</sup>Boellstorff 1976.

<sup>bb</sup>Stirton 1936.

<sup>cc</sup>Boellstorff 1978.

<sup>dd</sup>Naeser et al. 1980.

<sup>ee</sup>Lindsay et al. 1984.

<sup>ff</sup>McIntosh and Quade 1995.

<sup>gg</sup>Barghoorn and Tedford 1993.

<sup>hh</sup>Tedford and Barghoorn 1993.

<sup>ii</sup>Tedford and Barghoorn 1999.

<sup>jj</sup>S. Connell, pers. comm., 2002.

<sup>kk</sup>Stevens and Stevens 1989; Henry et al. 1998.

<sup>ll</sup>Ferrusquia-Villafranca 1992, 1996.

<sup>mmm</sup>Kowallis et al. 1986.

## REFERENCES

- Albers, J. P. and J. H. Stewart. 1972. Geology and mineral resources of Esmeralda County, Nevada. *Nevada Bureau of Mines and Geology Bulletin* 78.
- Albright, L. B. III. 1994. Lower vertebrates from an Arikareean (earliest Miocene) fauna near the Toledo Bend Dam, Newton County, Texas. *Journal of Paleontology* 68:1131–1145.
- . 1996. Insectivores, rodents and carnivores of the Toledo Bend Local Fauna: An Arikareean (earliest Miocene) assemblage from the Texas Coastal Plain. *Journal of Vertebrate Paleontology* 16:458–473.
- . 1998a. The Arikareean land mammal age in Texas and Florida: Southern extension of Great Plains faunas and Gulf Coastal Plain endemism. In *Depositional environments, lithostratigraphy and biostratigraphy of the White River and Arikaree*

- groups (*Late Eocene to Early Miocene, North America*), ed. D. O. Terry, H. E. LaGarry, and R. M. Hunt Jr. *Geological Society of America Special Paper* 325:167–196.
- . 1998b. New genus of tapir (Mammalia: Tapiridae) from the Arikareean (earliest Miocene) of the Texas Coastal Plain. *Journal of Vertebrate Paleontology* 18:200–217.
- . 1999a. Biostratigraphy and vertebrate paleontology of the San Timoteo Badlands, southern California. *University of California Publications, Geological Sciences* 144.
- . 1999b. Ungulates of the Toledo Bend Local Fauna (late Arikareean, early Miocene), Texas Coastal Plain. *Florida Museum of Natural History Bulletin* 42:1–80.
- Aldrich, M. P. Jr. and D. P. Dethier. 1990. Stratigraphy and tectonic evolution of the northern Espanola Basin, Rio Grande Rift, New Mexico. *Geological Society of America Bulletin* 102:1695–1705.
- Armstrong, R. L. and R. E. Higgins. 1973. K–Ar dating of the beginning of Tertiary volcanism in the Mojave Desert, California. *Geological Society of America Bulletin* 84:1095–1100.
- Barendregt, R. W., W. J. Vreeken, E. Irving, and J. Baker. 1997. Stratigraphy and paleomagnetism of the late Miocene Davis Creek Silt, East Block of the Cypress Hills, Saskatchewan. *Canadian Journal of Earth Sciences* 34:1325–1332.
- Barghoorn, S. F. 1981. Magnetic-polarity stratigraphy of the Miocene type Tesuque Formation, Santa Fe Group, in the Española Valley, New Mexico. *Geological Society of America Bulletin* 92:1027–1041.
- Barghoorn, S. F. and R. H. Tedford. 1993. Road Log. Neogene geology of the Española Basin, New Mexico. *New Mexico Museum of Natural History and Science Bulletin* 2:169–178.
- Barnosky, A. D. 1986a. Arikareean, Hemingfordian, and Barstovian mammals from the Miocene Colter Formation, Jackson Hole, Teton County, Wyoming. *Carnegie Museum of Natural History Bulletin* 26.
- . 1986b. New species of the Miocene rodent *Cupidinimus* (Heteromyidae) and some evolutionary relationships within the genus. *Journal of Vertebrate Paleontology* 6:46–64.
- Barnosky, A. D. and W. J. Labar. 1989. Mid-Miocene (Barstovian) environmental and tectonic setting near Yellowstone Park, Wyoming and Montana. *Geological Society of America Bulletin* 101:1448–1456.
- Barron, J. A. and C. M. Isaacs. 2001. Updated chronostratigraphic framework for the California Miocene. In *The Monterey Formation from rocks to molecules*, ed. C. M. Isaacs and J. Rullkötter. New York: Columbia University Press, pp. 393–395.
- Bartow, J. A. and K. McDougall. 1984. Tertiary stratigraphy of the southwestern San Joaquin Valley, California. *U.S. Geological Survey Bulletin* 1529J.
- Bartow, J. A., A. Sarna-Wojcicki, and W. O. Addicott. 1973. Correlation of marine and continental Pliocene deposits in northern California by tephrochronology (abstract). *American Association of Petroleum Geologists Bulletin* 57(4):769.
- Baskin, J. A. 1981. *Barbourofelis* (Nimravidae) and *Nimravides* (Felidae), with a description of two new species from the late Miocene of Florida. *Journal of Mammalogy* 62:122–139.
- Benson, R. N. 1998. Radiolarians and diatoms from the Pollack Farm Site, Delaware: Marine–terrestrial correlation of Miocene vertebrate assemblages of the Atlantic Coastal Plain. In *Geology and paleontology of the lower Miocene Pollack Farm Site, Delaware*, ed. R. N. Benson. *Delaware Geological Survey Special Publication* 21:5–19.
- Berggren, W. A., D. V. Kent, C. C. Swisher III, and M.-P. Aubry. 1995. A revised Cenozoic geochronology and chronostratigraphy. In *Geochronology, time scales and global stratigraphic correlation*, ed. W. A. Berggren, D. V. Kent, M.-P. Aubry, and J. Hardenbol. *Society for Sedimentary Geology Special Publication* 54:129–212.
- Black, C. C. 1961. Rodents and lagomorphs from the Miocene Fort Logan and Deep River formations of Montana. *Postilla* 48.
- Blackwelder, B. W. 1981. Late Cenozoic marine deposition in the United States Atlantic Coastal Plain related to tectonism and global climate. *Palaeogeography, Palaeoclimatology, Palaeoecology* 34:87–114.
- Blackwelder, B. W. and L. W. Ward. 1976. *Stratigraphy of the Chesapeake Group of Maryland and Virginia*. Arlington, VA: Geological Society of America Guidebook for Field Trip 7b, joint meeting of NE and SE sections.
- Bode, F. C. 1935a. The fauna of the Merychippus Zone, North Coalinga district, California. *Carnegie Institution of Washington Publication* 453:66–96.
- . 1935b. Tooth characters of protohippine horses with special reference to species from the Merychippus Zone, California. *Carnegie Institution of Washington Publication* 453:39–63.
- Boellstorff, J. D. 1976. The succession of late Cenozoic volcanic ashes in the Great Plains. A progress report. Guidebook, 24th Annual Meeting of the Midwest Friends of the Pleistocene. *Kansas Geological Survey Guidebook Series* 1:37–71.
- . 1978. Chronology of some late Cenozoic deposits from the central United States and the Ice Ages. *Nebraska Academy of Sciences Transactions* 6:35–49.
- Boellstorff, J. D. and M. F. Skinner. 1977. A fission-track date from post-Rosebud, early Valentine rocks. *Nebraska Academy of Sciences Proceedings* 1977:39.
- Bottomley, R. J. and D. York. 1976. <sup>40</sup>Ar–<sup>39</sup>Ar age determinations on the Owyhee Basalt of the Columbia Plateau. *Earth and Planetary Science Letters* 31:75–84.
- Bryant, H. N. 1996. Nimravidae. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 453–475.
- Bryant, J. D. 1991. New early Barstovian (middle Miocene) vertebrates from the upper Torreya Formation, eastern Florida Panhandle. *Journal of Vertebrate Paleontology* 11:472–489.
- Bryant, J. D., B. J. MacFadden, and P. A. Mueller. 1992. Improved chronologic resolution of the Hawthorn and the Alum Bluff Groups in northern Florida: Implications for Miocene chronostratigraphy. *Geological Society of America Bulletin* 104:208–218.
- Bryant, M. D. 1945. Phylogeny of Nearctic Sciuiridae. *The American Midland Naturalist* 33:257–390.
- Burbank, D. W. and A. D. Barnosky. 1990. The magnetochronology of Barstovian mammals in southwestern Montana and implications for the initiation of Neogene crustal extension in the northern Rocky Mountains. *Geological Society of America Bulletin* 102:1093–1104.
- Burns, J. A. and R. R. Young. 1988. Stratigraphy and paleontology of the Hand Hills region. *Tyrell Museum of Paleontology Occasional Paper* 9.
- Carranza-Castañeda, O. and A. H. Walton. 1992. Cricetid rodents from the Rancho El Ocote Fauna, late Hemphillian (Pliocene), state of Guanajuato. Universidad Nacional Autónoma de México. *Instituto de Geología, Revista* 10:71–93.
- Cook, H. J. 1965. Runningwater Formation, middle Miocene of Nebraska. *American Museum Novitates* 2227.

- Cook, H. J. and M. C. Cook. 1933. Faunal lists of the Tertiary Vertebrata of Nebraska and adjacent areas. *Nebraska Geological Survey Paper* 5.
- Coombs, M. C. 1978. A reevaluation of Early Miocene North American *Moropus* (Perissodactyla, Chalicotheriidae, Schizotheriinae). *Carnegie Museum of Natural History Bulletin* 4:1–62.
- Coombs, M. C., R. M. Hunt Jr., E. Stepleton, L. B. Albright III, and T. J. Fremd. 2001. Stratigraphy, chronology, biogeography, and taxonomy of early small chalicotheres in North America. *Journal of Vertebrate Paleontology* 21:607–620.
- Cope, E. D. 1874. Notes on the Eocene and Pliocene lacustrine formations of New Mexico, including descriptions of certain new species of vertebrates. In *Annual report upon the geographical explorations and surveys west of the one hundredth meridian, in California, Nevada, Utah, Arizona, Colorado, New Mexico, Wyoming and Montana*, ed. G. M. Wheeler. Washington, DC: U.S. GPO, Appendix FF, Annual Report Chief of Engineers for 1874, Appendix FF3, pp. 115–130.
- . 1875. Report on the geology of that part of northwestern New Mexico examined during the field season of 1874. In *Annual report upon the geographical explorations and surveys west of the hundredth meridian in California, Nevada, Nebraska, Utah, Arizona, Colorado, New Mexico, Wyoming and Montana*, ed. G. M. Wheeler. Washington, DC: U.S. GPO, Appendix LL, Annual Report, Chief of Engineers for 1875, Appendix GI, pp. 61–97.
- . 1877. Report upon the extinct vertebrata obtained in New Mexico by parties of the expedition of 1874. In *Report upon United States geographical surveys west of the one hundredth meridian*, ed. G. M. Wheeler. Washington, DC: U.S. GPO, Vol. 4:1–370.
- . 1878. Descriptions of new Vertebrata from Upper Tertiary formations of the west. *American Philosophical Society Proceedings* 17:219–231.
- . 1884. The Vertebrata of the Tertiary formations of the West. Book I. In *Report of the U.S. Geological Survey of the Territories*, ed. F. V. Hayden. Washington, DC: U.S. GPO, pp. i–xxxv, I-1009.
- . 1889. The Proboscidea. *American Naturalist* 23:191–211.
- Dalquest, W. W. 1983. Mammals of the Coffee Ranch Local Fauna, Hemphillian of Texas. *Texas Memorial Museum, Pearce–Sellards Series* 38:1–41.
- Dalquest, W. W., J. A. Baskin, and G. E. Schultz. 1996. Fossil mammals from a late Miocene (Clarendonian) site in Beaver County, Oklahoma. *Museum of Texas Tech University, contributions in mammalogy: A memorial volume honoring Dr. J. Knox-Jones Jr.* Lubbock: Texas Tech University, pp. 107–137.
- Dalquest, W. W. and O. Mooser. 1974. Miocene vertebrates from Aguascalientes, Mexico. *Texas Memorial Museum, Pearce–Sellards Series* 21.
- . 1980. Late Hemphillian mammals of the Ocote Local Fauna, Guanajuato, Mexico. *Texas Memorial Museum, Pearce–Sellards Series* 32.
- Dalquest, W. W. and D. B. Patrick. 1989. Small mammals from the early and medial Hemphillian of Texas, with descriptions of a new bat and gopher. *Journal of Vertebrate Paleontology* 9:78–88.
- Dalrymple, G. B. 1979. Critical tables for conversion of K–Ar ages from old to new constants. *Geology* 7:558–560.
- Darton, N. H. 1899. Preliminary report on the geology and water resources of Nebraska west of the one hundred and third meridian. *U.S. Geological Survey, 19th annual report*, part 4 (for 1897–1898), pp. 719–814.
- Dawson, M. R. 1999. Bering down: Miocene dispersals of land mammals between North America and Europe. In *The Miocene land mammals of Europe*, ed. G. E. Rössner and K. Heissig. Munich: Dr. F. Pfeil, pp. 473–483.
- Démere, T. A. 1988. Early Arikareean (late Oligocene) vertebrate fossils and biostratigraphic correlations of the Otay Formation at Eastlake, San Diego County, California. In *Paleogene stratigraphy, West Coast of North America, Pacific Section, Society of Economic Paleontologists and Mineralogists, West Coast Paleocene Symposium*, ed. M. V. Filewicz and R. L. Squires, 58:35–45.
- Dingus, L. 1990. Systematics, stratigraphy and chronology for mammalian fossils (late Arikareean to Hemingfordian) from the uppermost John Day Formation, Warm Springs, Oregon. *PaleoBios* 12:1–24.
- Dorr, J. A. Jr. 1956. Anceney local mammalian fauna, latest Miocene, Madison Valley formation, Montana. *Journal of Paleontology* 30:62–74.
- Dougherty, J. F. 1940. A new mammalian fauna from Caliente Mountain, California. *Carnegie Institution of Washington Publication* 514:111–143.
- Douglass, E. 1899. *The Neocene lake beds of western Montana*. M.S. thesis, University of Montana, Missoula.
- . 1901. Fossil Mammalia of the White River beds of Montana. *Transactions of the American Philosophical Society* 20:1–42.
- . 1903. New vertebrates from the Montana Tertiary. *Annals of the Carnegie Museum* 2:145–200.
- Downing, K. F. and C. C. Swisher III. 1993. New <sup>40</sup>Ar/<sup>39</sup>Ar dates and refined geochronology of the Sucker Creek Formation, Oregon. *Journal of Vertebrate Paleontology* 13(suppl. 3):33A.
- Downs, T. 1956. The Mascall fauna from the Miocene of Oregon. *University of California Publications in Geological Sciences* 31:199–354.
- . 1961. A study of variation and evolution of Miocene *Merychippus*. *Los Angeles County Museum Contributions in Science* 45:1–75.
- Durham, J. W., R. H. Johns, and D. E. Savage. 1954. Marine–non-marine relationships in the Cenozoic section of California. In *Geology of southern California*, chapter III, Historical geology. San Francisco: Division of Mines, pp. 59–71.
- Ekren, E. B., D. H. McIntyre, and E. H. Bennett. 1984. High temperature, large volume, lava-like ash flow tuffs without calderas in southwestern Idaho. *U.S. Geological Survey Professional Paper* 1272.
- Emry, R. E. and R. E. Eshelman. 1998. The early Hemingfordian (early Miocene) Pollack Farm Local Fauna: First Tertiary land mammals described from Delaware. In *Geology and paleontology of the lower Miocene Pollack Farm fossil site, Delaware*, ed. R. N. Benson. *Delaware Geological Survey Special Publication* 21:153–173.
- Engesser, B. 1979. Relationships of some insectivores and rodents from the Miocene of North America and Europe. *Carnegie Museum of Natural History Bulletin* 14.
- Evander, R. L. 1996. Horses (Mammalia, Equidae) from the Barstovian (Miocene) Anceney Local Fauna, Montana. *Paludicola* 1:1–4.
- . 1999. Rodents and lagomorphs (Mammalia) of the Railway Quarries Local Fauna (Miocene, Barstovian) of Nebraska. *Paludicola* 2:248–257.
- Evernden, J. F. and G. T. James. 1964. Potassium–argon dates and the Tertiary floras of North America. *American Journal of Science* 262:945–974.

- Evernden, J. F., D. E. Savage, G. H. Curtis, and G. T. James. 1964. Potassium-argon dates and the Cenozoic mammalian chronology of North America. *American Journal of Science* 262:145-198.
- Ferrusquia-Villafranca, I. 1975. Mamíferos miocénicos de México: Contribución al conocimiento de la paleozoogeografía del continente. *Universidad Nacional Autónoma de México, Instituto Geología, Revista* 1:12-18.
- . 1990. Biostratigraphy of the Mexican continental Miocene: Part I, Introduction and the northwestern and central faunas; Part II, the southeastern (Oaxacan) faunas; Part III, the southeasternmost (Chiapas) fauna and concluding remarks on the discussed vertebrate record. *Universidad Nacional Autónoma de México, Instituto Geología, Paleontología Mexicana* 56.
- . 1992. Contribución al conocimiento del Cenozoico en el Sureste de México y de su relevancia en el entendimiento de la evolución geológica regional. VII Congreso Geológico Latinoamericano, Salamanca, Spain. *Actas Sesiones Técnicas y Simposia* 4:40-44.
- . 1996. Contribución al conocimiento geológico de Chiapas: El Área Ixtapa-Soyaló. *Universidad Nacional Autónoma de México, Instituto Geología, Boletín* 109:1-130.
- Fisher, R. V. and J. M. Rensberger. 1972. Physical stratigraphy of the John Day Formation, central Oregon. *University of California Publication in Geological Sciences* 101:1-33.
- Frailey, C. D. 1979. The large mammals of the Buda Local Fauna (Arikarean: Alachua County, Florida). *Florida State Museum Bulletin* 24:123-173.
- Fremd, T., E. A. Bestland, and G. J. Retallack. 1994. *John Day Basin paleontology field trip guide and road log*. Prepared for 54th Annual Meeting, Society of Vertebrate Paleontology by John Day Fossil Beds National Monument, John Day, Oregon, unpagued.
- Frick, C. 1921. Extinct vertebrate faunas of Bautista Creek and San Timoteo Canyon, southern California. *University of California Publications, Bulletin of the Department of Geology* 12:122-176.
- . 1933. New remains of trilophodont-tetrabelodon mastodons. *American Museum of Natural History Bulletin* 59:505-652.
- . 1937. Horned ruminants of North America. *American Museum of Natural History Bulletin* 69:i-xxviii, 1-669.
- Galbreath, E. C. 1953. A contribution to the Tertiary geology and paleontology of northeastern Colorado. *University of Kansas Paleontological Contributions, Vertebrata*, Article 4.
- Galusha, T. 1966. The Zia Sand Formation, new early to medial Miocene beds in New Mexico. *American Museum Novitates* 2271.
- Galusha, T. and J. C. Blick. 1971. Stratigraphy of the Santa Fe Group, New Mexico. *American Museum of Natural History Bulletin* 144:1-128.
- Gawne, C. E. 1981. Sedimentology and stratigraphy of the Miocene Zia Sand of New Mexico: Summary. *Geological Society of America Bulletin*, Part I, 92:999-1007.
- Gazin, C. L. and R. L. Collins. 1950. Remains of land mammals from the Miocene of the Chesapeake Bay region. *Smithsonian Miscellaneous Contributions* 116:1-21.
- Gilbert, C. M. and M. W. Reynolds. 1973. Character and chronology of basin development, western margin of the Basin and Range Province. *Geological Society of America Bulletin* 84:2489-2510.
- Green, M. 1972. Lagomorpha from the Rosebud Formation, South Dakota. *Journal of Paleontology* 46:377-385.
- Greene, R. C. 1973. Petrology of the Welded Tuff of Devine Canyon, southwestern Oregon. *U.S. Geological Survey Professional Paper* 797.
- Greene, R. C., G. W. Walker, and R. E. Corcoran. 1972. *Geologic map of the Burns Quadrangle, Oregon*. U.S. Geological Survey, Miscellaneous. Geological Investigations Map I-680.
- Hanneman, D. L. and O. J. Wideman. 1991. Sequence stratigraphy of Cenozoic continental rocks, southwestern Montana. *Geological Society of America Bulletin* 103:1335-1345.
- Harksen, J. C. 1969. The Cenozoic history of southwestern South Dakota. In *Guidebook to the major Cenozoic deposit of southwestern South Dakota*, ed. J. C. Harksen and J. R. Macdonald. *South Dakota Geological Survey Guidebook* 2:11-28.
- Harrington, C. R. 1978. Quaternary vertebrate faunas of Canada and Alaska and their suggested chronological sequence. *National Museums of Canada Syllogus* 15:1-105.
- Harrison, J. A. 1985. Giant camels from the Cenozoic of North America. *Smithsonian Contributions to Paleobiology* 57:1-29.
- Hatcher, J. B. 1902. Origin of the Oligocene and Miocene deposits of the Great Plains. *Proceedings of the American Philosophical Society* 41:113-131.
- Hayes, F. G. 2000. The Brooksville 2 Local Fauna (Arikarean, latest Oligocene): Hernando County, Florida. *Florida Museum of Natural History Bulletin* 43:1-47.
- Henry, C. D., L. L. Davis, M. J. Kunk, and W. C. McIntosh. 1998. Tertiary volcanism of the Bofecillas Mountains and Big Bend Ranch State Park Texas: Revised stratigraphy and <sup>40</sup>Ar/<sup>39</sup>Ar geochronology. *Bulletin of Economic Geology, University of Texas*, Report of Investigations no. 253.
- Hernandez, L. D. 1981. Estratigrafía de la región central de Aguascalientes, Ags, México. *Society Geología Mexicana Gaceta Geológica* 31:17-40.
- Hesse, C. J. 1936. Lower Pliocene vertebrate fossils from the Ogalala Formation (Lavern Zone) of Beaver County, Oklahoma. Carnegie Institution of Washington, Publication 476:47-71.
- Hibbard, C. W. 1954. A new Pliocene vertebrate fauna from Oklahoma. *Michigan Academy of Science, Arts, and Letters Papers* 39:339-359.
- . 1963. *Tanupoloma vera* (Matthew) from the late Hemphillian of Beaver County, Oklahoma. *Kansas Academy of Science Transactions* 66:267-269.
- Hibbard, C. W. and K. A. Keenmon. 1950. New evidence of the lower Miocene age of the Blacktail Deer Creek Formation in Montana. *University of Michigan Contributions from the Museum of Paleontology* 8:193-204.
- Hickey, L. J., K. R. Johnson, and M. R. Dawson. 1988. The stratigraphy, sedimentology and fossils of the Haughton Formation: A post-impact crater fill, Devon Island, N.W.T., Canada. *Meteoritics* 23:221-231.
- Hulbert, R. C. Jr. 1988a. *Calippus* and *Protohippus* (Mammalia, Perissodactyla, Equidae) from the Miocene (Barstovian-early Hemphillian) of the Gulf Coastal Plain. *Florida State Museum Biological Sciences Bulletin* 32:221-340.
- . 1988b. *Cormohipparion* and *Hipparion* (Mammalia, Perissodactyla, Equidae) from the late Neogene of Florida. *Florida State Museum Biological Sciences Bulletin* 33:229-338.
- Hulbert, R. C. Jr. and B. J. MacFadden. 1991. Morphological transformation and cladogenesis at the base of the adaptive radiation of Miocene hypsodont horses. *American Museum Novitates* 3000.

- Hunt, R. M. Jr. 1985. Faunal succession, lithofacies, and depositional environments in Arikaree rocks (Lower Miocene) of the Hartville table, Nebraska and Wyoming. *Dakoterra* 2:155–204.
- . 1998. Amphicyonidae. In *Evolution of Tertiary mammals of North America*, Vol. 1: *Terrestrial carnivores, ungulates and ungulate-like mammals*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, pp. 196–227.
- . 2001. Small Oligocene amphicyonids from North America (*Paradaphoenus*, Mammalia, Carnivora). *American Museum Novitates* 3331.
- . 2002. New amphicyonid carnivorans (Mammalia, Daphoeninae) from the early Miocene of Southeastern Wyoming. *American Museum Novitates* 3385.
- Hunt, R. M. Jr. and E. Stepleton. 2001. Geology and paleontology of the Upper John Day, lithostratigraphic and biochronologic revision of the beds, John Day River valley, Oregon: Haystack Valley and Kimberly areas. *PaleoBios* 21(suppl. 2):69–70.
- Hunt, R. M. Jr., X.-X. Xue, and J. Kaufman. 1983. Miocene burrows of extinct bear dogs: Indication of early denning behavior of large mammalian carnivores. *Science* 221:364–366.
- Hutchinson, J. H. and E. H. Lindsay. 1974. The Hemingfordian mammal fauna of the Vedder Locality, Branch Canyon Formation, Santa Barbara County, California. Part I: Insectivora, Chiroptera, Lagomorpha and Rodentia (Sciuridae). *PaleoBios* 15.
- Izett, G. A. and J. D. Obradovich. 2001.  $^{40}\text{Ar}/^{39}\text{Ar}$  ages of Miocene tuffs in basin fill deposits (Santa Fe Group, New Mexico and Troublesome Formation, Colorado) of the Rio Grande system. *The Mountain Geologist* 38:77–86.
- James, G. T. 1963. Paleontology and nonmarine stratigraphy of the Cuyama Valley badlands, California. Part 1. Geology, faunal interpretations, and systematic descriptions of Chiroptera, Insectivora, and Rodentia. *University of California Publications in Geological Sciences* 45:i–iv, 1–154.
- Janis, C. M., K. M. Scott, and L. L. Jacobs. 1998. *Evolution of Tertiary mammals of North America*, Vol. 1: *Terrestrial carnivores, ungulates and ungulate-like mammals*. New York: Cambridge University Press, pp. i–xi, 1–691.
- Jones, D. S., P. A. Mueller, D. A. Hodell, and L. A. Stanley. 1993.  $^{87}\text{Sr}/^{86}\text{Sr}$  geochronology of Oligocene and Miocene strata in Florida. In *The Neogene of Florida and adjacent areas*, ed. V. A. Zullo, W. B. Harris, T. M. Scott, and R. W. Portell. *Florida Geological Survey Special Publication* 37:55–66.
- Jones, D. S., L. W. Ward, P. A. Mueller, and D. A. Hodell. 1998. Age of marine molluscs from the lower Miocene Pollack Farm site, Delaware, determined by  $^{87}\text{Sr}/^{86}\text{Sr}$  geochronology. In *Geology and paleontology of the lower Miocene Pollack Farm Site, Delaware*, ed. R. N. Benson. *Delaware Geological Survey Special Publication* 21:21–25.
- Kelly, T. S. 1992. New middle Miocene camels from the Caliente Formation, Cuyama Valley badlands, California. *PaleoBios* 13:1–22.
- . 1994. Two Pliocene (Blancan) vertebrate faunas from Douglas County, Nevada. *PaleoBios* 16:1–23.
- . 1995. New Miocene horses from the Caliente Formation, Cuyama Valley badlands, California. *Natural History Museum of Los Angeles County Contributions in Science* 455.
- . 1997. Additional late Cenozoic (latest Hemphillian to earliest Irvingtonian) mammals from Douglas County, Nevada. *PaleoBios* 18:1–31.
- . 1998. New middle Miocene equid crania from California and their implications for the phylogeny of the Equini. *Natural History Museum of Los Angeles County Contributions in Science* 473.
- Kelly, T. S. and E. B. Lander. 1988. Biostratigraphy and correlation of Hemingfordian and Barstovian land mammal assemblages, Caliente Formation, Cuyama Valley area, California. In *Tertiary tectonics and sedimentation in the Cuyama Basin, San Luis Obispo, Santa Barbara and Ventura counties, California*, ed. W. J. M. Bazeky. *Society of Economic Paleontologists and Mineralogists, Pacific Section* 59:1–19.
- . 1992. Miocene land mammal faunas from the Caliente Formation, Cuyama Valley badlands, California. *PaleoBios* 14:3–8.
- Koerner, H. E. 1940. The geology and vertebrate paleontology of the Fort Logan and Deep River formations of Montana, Part 1: New vertebrates. *American Journal of Science* 238:837–862.
- Korth, W. W. 1992. Fossil small mammals from the Harrison Formation (late Arikareean: earliest Miocene), Cherry County, Nebraska. *Annals of the Carnegie Museum* 61:69–131.
- . 1994. *The Tertiary record of rodents in North America*. New York: Plenum Press.
- . 1996a. A new genus of beaver (Mammalia: Castoridae: Rodentia) from the Arikareean (Oligocene) of Montana and its bearing on castorid phylogeny. *Annals of the Carnegie Museum* 65:167–179.
- . 1996b. A new genus of prairie dog (Sciuridae, Rodentia) from the Miocene (Barstovian of Montana and Clarendonian of Nebraska) and the classification of Nearctic ground squirrels (Marmotini). *Transactions of the Nebraska Academy of Sciences* 23:109–113.
- . 1999a. *Hesperogaulus*, a new genus of mylagaulid rodent (Mammalia) from the Miocene (Barstovian to Hemphillian) of the Great Basin. *Journal of Paleontology* 73:938–944.
- . 1999b. A new genus of derived promylagauline rodent (Mylagaulidae) from the Miocene (late Hemingfordian–early Barstovian). *Journal of Vertebrate Paleontology* 19:752–756.
- . 1999c. A new species of beaver (Rodentia, Castoridae) from the earliest Barstovian (Miocene) of Nebraska and the phylogeny of *Monosaulax* Stirton. *Paludicola* 2:258–264.
- . 2000. Review of the Miocene (Hemingfordian to Clarendonian) mylagaulid rodents (Mammalia) from Nebraska. *Annals of the Carnegie Museum* 69:227–280.
- Korth, W. W. and B. E. Bailey. 1992. Additional specimens of *Leptodontomys douglassi* (Eomyidae, Rodentia) from the Arikareean (late Oligocene) of Nebraska. *Journal of Mammalogy* 73:651–662.
- Korth, W. W., B. E. Bailey, and R. M. Hunt Jr. 1990. Geomyid rodents from the early Hemingfordian (Miocene) of Nebraska. *Annals of the Carnegie Museum* 59:25–47.
- Kowallis, B. J., J. S. Heaton, and K. Bringhurst. 1986. Fission-track dating of volcanically derived sedimentary rocks. *Geology* 14:19–22.
- Kuenzi, W. D. and R. W. Fields. 1971. Tertiary stratigraphy, structure, and geological history, Jefferson Basin, Montana. *Geological Society of America Bulletin* 82:3373–3394.
- Lambe, L. M. 1905. Fossil horses of the Oligocene of the Cypress Hills, Assiniboia. *Royal Society of Canada Transactions*, series 2, 11(4):43–52.
- Lambert, W. D. and J. Shoshani. 1998. Proboscidea. In *Evolution of Tertiary mammals of North America*, Vol. 1: *Terrestrial carnivores*,

- ungulates and ungulate-like mammals, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, pp. 606–621.
- Lander, E. B. 1983. Continental vertebrate faunas from the upper member of the Sespe Formation, Simi Valley, California, and the Terminal Eocene Event. In *Cenozoic geology of the Simi Valley area, southern California, Pacific Section, Society of Economic Paleontologists and Mineralogists, Fall Field Trip Guidebook*, ed. R. R. Squires and M. V. Filewicz, pp. 142–144.
- Lander, E. G. 1994. *Paleontologic Resource Impact Mitigation Program, final report, Santiago Canyon Land Fill, southeast and southwest borrows, Orange County, California*. Irvine, CA: Chambers Group Inc.
- Lindsay, E. H. 1972. Small mammal fossils from the Barstow Formation, California. *University of California Publications in Geological Sciences* 93:1–104.
- . 1974. The Hemingfordian mammal fauna of the Vedder Locality, Branch Canyon Formation, Santa Barbara County, California. Part II: Rodentia (Eomyidae and Heteromyidae). *Paleo-Bios* 16.
- . 1995. *Copemys* and the Barstovian/Hemingfordian boundary. *Journal of Vertebrate Paleontology* 15:357–365.
- Lindsay, E. H. and L. L. Jacobs. 1985. Pliocene small mammal fossils from Chihuahua, Mexico. *Universidad Nacional Autonoma de Mexico, Instituto de Geologia, Paleontologia Mexicana* 51.
- Lindsay, E. H., N. M. Johnson, N. D. Opdyke, and R. F. Butler. 1987. Mammalian chronology and the magnetic polarity time scale. In *Cenozoic mammals of North America: Geochronology and biostratigraphy*, ed. M. O. Woodburne. Berkeley: University of California Press, pp. 269–284.
- Lindsay, E., Y. Mou, W. Downs, J. Pedersson, T. Kelly, C. Henry, and J. Trexler. 2002. Recognition of the Hemphillian/Blancan boundary in Nevada. *Journal of Vertebrate Paleontology* 22:429–442.
- Lindsay, E. H., N. D. Opdyke, and N. M. Johnson. 1984. Blancan–Hemphillian land mammal ages and Late Cenozoic mammal dispersal events. *Annual Review of Earth and Planetary Sciences* 12:445–488.
- Lindsay, E. H. and R. H. Tedford. 1990. Development and application of land mammal ages in North America and Europe, a comparison. In *European Neogene mammal chronology*, ed. E. H. Lindsay, V. Fahlbusch, and P. Mein. New York: Plenum, pp. 601–624.
- Love, J. D. 1961. Split Rock Formation (Miocene) and Moonstone Formation (Pliocene) in Central Wyoming. *U.S. Geological Survey Bulletin* 1121-I.
- . 1970. Cenozoic geology of the Granite Mountains area, central Wyoming. *U.S. Geological Survey Professional Paper* 495-C.
- Lucas, S. G., D. P. Whistler, and H. M. Wagner. 1997. Giant entelodont (Mammalia, Artiodactyla) from the early Miocene of Southern California. *Natural History Museum of Los Angeles County Contributions in Science* 466.
- Lugn, A. L. 1939. Classification of the Tertiary system in Nebraska. *Geological Society of America Bulletin* 50:1245–1276.
- Lundelius, E. L., C. S. Churcher, T. Downs, C. R. Harington, E. H. Lindsay, G. E. Schultz, H. A. Semken, S. D. Webb, and R. J. Zakrzewski. 1987. The North American Quaternary sequence. In *Cenozoic mammals of North America: Geochronology and biostratigraphy*, ed. M. O. Woodburne. Berkeley: University of California Press, pp. 211–235.
- Macdonald, J. R. 1963. The Miocene faunas from the Wounded Knee area of western South Dakota. *American Museum of Natural History Bulletin* 125:139–238.
- . 1970. Review of the Wounded Knee faunas of southwestern South Dakota. *Los Angeles County Museum of Natural History and Science Bulletin* 9.
- Macdonald, L. J. 1948. The Pliocene carnivores of the Black Hawk Ranch fauna. *University of California Publications, Bulletin of the Department of Geological Sciences* 28:53–80.
- . 1972. Monroe Creek (early Miocene) microfossils from the Wounded Knee area, South Dakota. *South Dakota Geological Survey Report of Investigation* 105:i–iii, 1–43.
- MacFadden, B. J. 1977. Magnetic-polarity stratigraphy of the Chamita Formation stratotype (Mio-Pliocene) of north-central New Mexico. *American Journal of Science* 277:769–800.
- . 1980. An early Miocene land mammal (Oreodonta) from a marine limestone in northern Florida. *Journal of Paleontology* 54:93–101.
- . 1984. Systematics and phylogeny of *Hipparion*, *Neohipparion*, *Nannippus*, and *Cormohipparion* (Mammalia, Equidae) from the Miocene and Pliocene of the New World. *American Museum of Natural History Bulletin* 179:1–196.
- MacFadden, B. J. and R. C. Hulbert Jr. 1988. Explosive speciation at the base of the adaptive radiation of Miocene grazing horses. *Nature* 336:466–468.
- MacFadden, B. J. and R. M. Hunt. 1998. Magnetic polarity stratigraphy and correlation of the Arikaree Group, Arikarean (late Oligocene–early Miocene) of northwestern Nebraska. In *Depositional environments, lithostratigraphy, and biostratigraphy of the White River and Arikaree groups (Late Eocene to Early Miocene, North America)*, Boulder, Colorado, ed. D. O. Terry Jr., H. E. LaGarry, and R. M. Hunt Jr. *Geological Society of America Special Paper* 325:143–165.
- MacFadden, B. J., C. C. Swisher III, N. D. Opdyke, and M. O. Woodburne. 1990. Paleomagnetism, geochronology, and possible tectonic rotation of the middle Miocene Barstow Formation, Mojave Desert, southern California. *Geological Society of America Bulletin* 102:478–493.
- Madden, C. T. and J. E. Storer. 1985. The Proboscidea from the middle Miocene Wood Mountain Formation, Saskatchewan. *Canadian Journal of Earth Sciences* 22:1345–1350.
- Marsh, O. C. 1870. Remarks on *Hadrosaurus minor*, *Mosasaurus crassidens*, *Liodon laticaudus*, *Baptosaurus* and *Rhinoceras matutinus*. *Proceedings of the Academy of Natural Sciences Philadelphia* 1870:2–3.
- . 1871. Description of *Lophiodon validus*. *Proceedings of the Academy of Natural Sciences Philadelphia* 1871:10.
- . 1893. Description of Miocene Mammalia. *American Journal of Science* 46:407–412.
- Martin, L. D. 1973. *The mammalian fauna of the lower Miocene Gering Formation of western Nebraska and the early evolution of the North American Cricetidae*. Ph.D. dissertation, University of Kansas, Lawrence.
- . 1987. Beavers from the Harrison Formation (early Miocene), with a revision of *Euhapsis*. *Dakoterra* 3:73–91.
- Mason, M. A. and C. C. Swisher III. 1989. New evidence for the age of the South Mountain Local Fauna, Ventura County, California. *Natural History Museum of Los Angeles County Contributions in Science* 410.

- Matthew, W. D. and J. W. Gidley. 1904. New or little known mammals from the Miocene of South Dakota. American Museum expedition of 1903. *American Museum of Natural History Bulletin* 20:241–268.
- May, S. R. and C. A. Reppenng. 1982. New evidence for the age of the Mount Eden Fauna, southern California. *Journal of Vertebrate Paleontology* 2:109–113.
- McIntosh, W. C. and J. Quade. 1995.  $^{40}\text{Ar}/^{39}\text{Ar}$  geochronology of tephra layers in the Santa Fe Group, Española Basin, New Mexico. *New Mexico Geological Society Guidebook, 46th Field Conference, Geology of Santa Fe Region*, pp. 279–287.
- McKenna, M. C. 1955. A new species of mylagaulid from the Chalk Cliffs Local Fauna, Montana. *Journal of the Washington Academy of Sciences* 45:107–110.
- McKenna, M. C. and S. K. Bell. 1997. *Classification of mammals above the species level*. New York: Columbia University Press, pp. i–xii, 1–631.
- Merriam, J. C. 1910. Tertiary mammal beds of Virgin Valley and Thousand Creek in northwestern Nevada. Part I, geologic history. *University of California Publications, Bulletin of the Department of Geology* 6:21–53.
- . 1911. Tertiary mammal beds of Virgin Valley and Thousand Creek in northwestern Nevada. Part II, vertebrate faunas. *University of California Publications, Bulletin of the Department of Geology* 6:199–304.
- Merriam, J. C. and W. J. Sinclair. 1907. Tertiary faunas of the John Day region. *University of California Publications, Bulletin of the Department of Geological Sciences* 5:171–205.
- Morea, M. F. 1981. *The Massacre Lake local fauna (Mammalia, Hemingfordian) from northwestern Washoe County, Nevada*. Ph.D. dissertation, University of California, Riverside.
- Morgan, G. S. 1989. Miocene vertebrate faunas from the Suwanee River basin of north Florida and south Georgia. *Southeastern Geological Society Guidebook* 30:26–53.
- . 1993. Mammalian biochronology and marine–nonmarine correlations in the Neogene of Florida. *Florida Geological Survey Special Publication* 37:55–66.
- Munthe, J. 1979a. The Hemingfordian mammal fauna of the Vedder Locality, Branch Canyon Formation, Santa Barbara County, California, Part III: Carnivora, Perissodactyla, Artiodactyla and summary. *PaleoBios* 29.
- . 1979b. Summary of Miocene vertebrate fossils of the Granite Mountains Basin, central Wyoming. *University of Wyoming Contributions to Geology* 18:33–46.
- . 1988. Miocene mammals of the Split Rock area, Granite Mountains Basin, central Wyoming. *University of California, Publications of the Department of Geological Sciences* 126:1–136.
- Naeser, C. W., G. A. Izett, and J. D. Obradovich. 1980. Fission-track and K–Ar ages of natural glasses. *U.S. Geological Survey Bulletin* 1489.
- Nichols, R. 1976. Early Miocene mammals from the Lemhi Valley of Idaho. *Tebiwa* 18:9–47.
- . 1979. Additional early Miocene mammals from the Lemhi Valley of Idaho. *Tebiwa* 17.
- Nielson, J. E., D. R. Lux, G. B. Dalrymple, and A. F. Glazner. 1990. The age of the Peach Springs Tuff, southeastern California and western Arizona. *Journal of Geophysical Research* 95:571–580.
- Noble, D. C., C. E. Hedge, E. H. McKee, and M. K. Korrington. 1973. Reconnaissance study of the strontium isotopic composition of Cenozoic volcanic rocks in the northwestern Great Basin. *Geological Society of America Bulletin* 84:1393–1406.
- Noble, D. C., E. H. McKee, J. G. Smith, and M. K. Korrington. 1970. Stratigraphy and geochronology of Miocene volcanic rocks in northwestern Nevada. *U.S. Geological Survey Professional Paper* 700-D:D23–D32.
- Olsen, S. J. 1964. The stratigraphic importance of a lower Miocene vertebrate fauna from north Florida. *Journal of Paleontology* 38:477–482.
- Omar, G., K. R. Johnson, L. J. Hickey, P. B. Robertson, M. R. Dawson, and C. W. Barnosky. 1987. Fission-track dating of Haughton Astrobleme and included biota, Devon Island, Canada. *Science* 237:1603–1605.
- Parker, D. and R. C. Armstrong. 1972. K–Ar dates and Sr isotope initial ratios for volcanic rocks in the Harney Basin, Oregon. *Isochron/West* 5:7–12.
- Peale, A. C. 1896. Description of the Three Forks Sheet, Montana. *U.S. Geological Survey Geological Atlas*, Folio 24.
- Perkins, M. E., F. H. Brown, and W. Nash. 1998. Sequence, age, and source of silicic fallout tuffs in the middle to late Miocene basins of the northern Basin and Range Province. *Geological Society of America Bulletin* 110:344–360.
- Perkins, M. E. and B. P. Nash. 2002. Explosive silicic volcanism of the Yellowstone hotspot: The ash fall tuff record. *Geological Society of America Bulletin* 114:367–381.
- Perkins, M. E., W. P. Nash, F. H. Brown, and R. J. Fleck. 1995. Fall-out tuffs of Trapper Creek, Idaho: A record of Miocene explosive volcanism in the Snake River Plain volcanic Province. *Geological Society of America Bulletin* 107:1484–1506.
- Peterson, O. A. 1905. Description of new rodents and discussion of the origin of *Daemonelex*. *Memoirs of the Carnegie Museum* 2:139–200.
- . 1907. The Miocene beds of western Nebraska and eastern Wyoming and their vertebrate faunas. *Annals of the Carnegie Museum* 4:21–72.
- . 1909. A revision of the Entelodontidae. *Memoirs of the Carnegie Museum* 4:41–158.
- Prothero, D. R. 1991. Magnetic stratigraphy of Eocene–Oligocene mammal localities in southern San Diego County. In *Eocene geologic history, San Diego region*, ed. P. L. Abbott and J. A. May. Society for Sedimentary Geology, Pacific Section, Publication 68:125–130.
- . 1996. Camelidae. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 609–651.
- Prothero, D. R. and L. L. Donohoo. 2001. Magnetic stratigraphy of the lower Miocene (early Hemingfordian) Sespe–Vaqueros Formations, Orange County, California. In *Magnetic stratigraphy of the Pacific Coast Cenozoic*, ed. D. R. Prothero. Society for Sedimentary Geology, Pacific Section, Publication 91:242–253.
- Prothero, D. R. and R. J. Emry. 1996. *The terrestrial Eocene–Oligocene transition in North America*. New York: Cambridge University Press, pp. i–xii, 1–688.
- Prothero, D. R. and J. M. Rensberger. 1985. Preliminary magnetostratigraphy of the John Day Formation, Oregon, and the North American Oligocene–Miocene boundary. *Newsletter in Stratigraphy* 15:59–70.
- Prothero, D. R., J. L. Howard, and T. H. H. Dozier. 1996. Stratigraphy and paleomagnetism of the upper middle Eocene to lower

- Miocene (Uintan to Arikarean) Sespe Formation, Ventura County, California. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 171–188.
- Prothero, D. R. and R. H. Tedford. 2000. Magnetic stratigraphy of the type Montediablan Stage (Late Miocene), Black Hawk Ranch, Contra Costa County, California: Implications for regional correlations. *PaleoBios* 20:1–10.
- Raschke, R. E. 1984. Early and middle Miocene vertebrates from the Santa Ana Mountains, California. In *The natural sciences of Orange County*, ed. B. Butler, J. Gant, and C. J. Stadum. *Memoirs of the Natural History Foundation of Orange County* 1:61–67.
- Rasmussen, D. L. 1969. *Late Cenozoic geology of the Cabbage Patch area, Granite and Powell counties, Montana*. M.A. thesis, University of Montana, Missoula.
- . 1977. *Geology and mammalian paleontology of the Oligocene–Miocene Cabbage Patch Formation, central-western Montana*. Ph.D. dissertation, University of Kansas, Lawrence.
- Rensberger, J. M. 1971. Entoptychine pocket gophers (Mammalia, Geomyoidea) of the early Miocene John Day Formation, Oregon. *University of California Publications in Geological Sciences* 90:1–163.
- . 1973. Pleurolicine rodents (Geomyoidea) of the John Day Formation, Oregon and their relationships to taxa from the early and middle Miocene. South Dakota. *University of California Publications in Geological Sciences* 102:1–95.
- . 1979. *Promylagaulus*, progressive aplodontoid rodents of the early Miocene. *Natural History Museum of Los Angeles County Contributions in Science* 312.
- . 1981. Evolution in a late Oligocene–early Miocene succession of meniscomyine rodents in the Deep River Formation, Montana. *Journal of Vertebrate Paleontology* 1:185–209.
- . 1983. Successions of meniscomyine and allomyine rodents (Aplodontidae) in the Oligo–Miocene John Day Formation, Oregon. *University of California Publications, Geological Sciences* 124.
- Repenning, C. A. and J. G. Vedder. 1961. Continental vertebrates and their stratigraphic correlation with marine mollusks, eastern Caliente Range, California. *U.S. Geological Survey Professional Paper* 424-C:C235–C239.
- Reynolds, R. E., R. M. Hunt Jr., and B. Albright III. 1995. Rhinoceros in Landfair Valley. *San Bernardino County Museum Association Quarterly* 42:107–110.
- Reynolds, R. E. and M. O. Woodburne. 2001. Review of the proboscidean datum within the Barstow Formation, Mojave Desert, California. *Journal of Vertebrate Paleontology Abstracts* 21:93A.
- Robinson, G. D. 1961. Origin and development of the Three Forks Basin, Montana. *Geological Society of America Bulletin* 72:1003–1014.
- . 1963. Geology of the Three Forks Quadrangle, Montana. *U.S. Geological Survey Professional Paper* 370.
- Sarna-Wojcicki, A. M. 1976. Correlation of late Cenozoic tuffs in the central coast ranges of California by means of trace and minor element chemistry. *U.S. Geological Survey Professional Paper* 972.
- Savage, D. E. 1955. Nonmarine lower Pliocene sediments in California: A geochronologic–stratigraphic classification. *University of California Publications in Geological Sciences* 31:1–26.
- Schlaikjer, E. M. 1935. Contributions to the stratigraphy and palaeontology of the Goshen Hole area, Wyoming IV. New vertebrates and the stratigraphy of the Oligocene and early Miocene. *Bulletin of the Museum of Comparative Zoology, Harvard College* 76:99–189.
- Schuchert, C. and C. M. Levene. 1940. *O. C. Marsh: Pioneer in paleontology*. New Haven, CT: Yale University Press.
- Schultz, C. B. 1938. The Miocene of western Nebraska. *American Journal of Science* 35:441–444.
- Schultz, C. B. and C. H. Falkenbach. 1949. Promerycochoerinae, a new subfamily of oreodonts. *American Museum of Natural History Bulletin* 93:69–198.
- . 1968. The phylogeny of the oreodonts. Parts 1 and 2. *American Museum of Natural History Bulletin* 139:1–498.
- Schultz, G. E. 1990. Clarendonian and Hemphillian vertebrate faunas from the Ogallala Formation (late Miocene–early Pliocene) of the Texas Panhandle and adjacent Oklahoma. In *Geologic framework and regional hydrology: Upper Cenozoic Blackwater Draw and Ogallala Formations, Great Plains*, ed. T. C. Gustavson. Austin: Bureau of Economic Geology, the University of Texas, pp. 56–97.
- Scott, W. B. 1893. The mammals of the Deep River beds. *American Naturalist* 27:659–662.
- Shattuck, G. B. 1904. Geologic and paleontological relations with a review of earlier investigations. In *The Miocene deposits of Maryland*, ed. W. B. Clark, G. B. Shattuck, and W. H. Dall. Baltimore: Maryland Geological Survey, Miocene volume, pp. 73–94.
- Silberman, M. L. and E. H. McKee. 1972. A summary of radiometric determinations on Tertiary volcanic rocks from Nevada and eastern California: Part II, western Nevada. *Ischron/West* 4:7–28.
- Simpson, G. G. 1930. Tertiary land mammals of Florida. *American Museum of Natural History Bulletin* 59:149–211.
- Skinner, M. F. and F. W. Johnson. 1984. Tertiary stratigraphy and the Frick Collection of fossil vertebrates from north-central Nebraska. *American Museum of Natural History Bulletin* 178:215–368.
- Skinner, M. F., S. M. Skinner, and R. J. Gooris. 1968. Cenozoic rocks and faunas of Turtle Buttes, south-central South Dakota. *American Museum of Natural History Bulletin* 138:379–436.
- Skwara, T. 1988. Mammals of the Topham Local Fauna: Early Miocene (Hemingfordian). Cypress Hills Formation, Saskatchewan. *Saskatchewan Museum of Natural History, Natural History Contributions* 9:1–169.
- Slaughter, B. H. 1981. A new genus of geomyoid rodent from the Miocene of Texas and Panama. *Journal of Vertebrate Paleontology* 1:111–115.
- Stalker, A. M. and C. S. Churcher. 1972. Glacial stratigraphy of the southwestern Canadian prairies: The Laurentide record. *24th International Geological Congress*, section 12, pp. 110–119.
- Stepleton, E. and R. M. Hunt Jr. 1994. Geology and paleontology of the Upper John Day beds. *Society of Vertebrate Paleontology Field Conference Guidebook*.
- Stevens, M. S. 1966. The osteology and relationships of the Pliocene ground squirrel, *Citellus dotti* Hibbard from the Ogallala Formation of Beaver County, Oklahoma. *Texas Memorial Museum, Pearce–Sellards Series* 4.
- . 1977. Further study of Castolon Local Fauna (early Miocene), Big Bend National Park, Texas. *Texas Memorial Museum, Pearce–Sellards Series* 28.
- Stevens, M. S. and J. B. Stevens. 1989. Neogene–Quaternary deposits and vertebrate faunas Trans-Pecos Texas. In *Vertebrate paleontology, biostratigraphy and depositional environments, latest Creta-*



- ceous and Tertiary, Big Bend Area, Texas*, ed. A. B. Busbey II and T. M. Lehman. Austin, TX: Society of Vertebrate Paleontology Guidebook, Field Trips 1A, B, C. 49th Annual Meeting, pp. 67–90.
- . 1996. Merycoidodontinae and Miniochoerinae. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 498–573.
- Stevens, M. S., J. B. Stevens, and M. R. Dawson. 1969. New Early Miocene formation and vertebrate local fauna, Big Bend National Park, Brewster County, Texas. *Texas Memorial Museum, Pearce–Sellards Series* 15.
- Stirton, R. A. 1935. A review of the Tertiary beavers. *University of California Publications, Bulletin of the Department of Geological Sciences* 23:391–458.
- . 1936. Succession of North American continental Pliocene mammalian faunas. *American Journal of Science* 32:161–206.
- . 1939. The Nevada Miocene and Pliocene mammalian faunas as faunal units. *Sixth Pacific Science Congress Proceedings*, pp. 627–638.
- . 1954. Late Miocene mammals from Oaxaca, Mexico. *American Journal of Science* 252:634–638.
- Stock, C. 1930. Oreodonts from the Sespe deposits of South Mountain, Ventura County, California. *Carnegie Institution of Washington Contributions to Paleontology* 404:27–42.
- . 1932. Additions to the mammalian fauna of the Tecuya beds, California. *Carnegie Institution of Washington Publication* 418:89–92.
- . 1934. On the occurrence of an oreodont skeleton in the Sespe of South Mountain, California. *Proceedings of the National Academy of Sciences* 20:518–523.
- Storer, J. E. 1975. Tertiary mammals of Saskatchewan. Part III: The Miocene fauna. *Royal Ontario Museum Life Sciences Contribution* 103.
- . 1978. Tertiary sands and gravels in Saskatchewan and Alberta: Correlation of mammalian faunas. In *Western and Arctic Canadian biostratigraphy*, ed. C. R. Stelck and B. D. E. Challerton. *Geological Association of Canada Special Paper* 18:595–602.
- . 1993. Additions to the mammalian palaeofauna of Saskatchewan. *Modern Geology* 18:475–487.
- . 1996. Eocene–Oligocene faunas of the Cypress Hills Formation, Saskatchewan. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry Jr. New York: Cambridge University Press, pp. 240–261.
- . 2002. Small mammals of the Kealey Springs Local Fauna (Early Arikareean, Late Oligocene) of Saskatchewan. *Paludicola* 3:105–133.
- Storer, J. E. and H. N. Bryant. 1993. Biostratigraphy of the Cypress Hills Formation (Eocene to Miocene), Saskatchewan: Equid types (Mammalia: Perissodactyla) and associated faunal assemblages. *Journal of Paleontology* 67:660–669.
- Sugarman, P. J., K. G. Miller, J. P. Owens, and M. D. Feigenson. 1993. Strontium-isotope and sequence stratigraphy of the Miocene Kirkwood Formation, southern New Jersey. *Geological Society of America Bulletin* 105:423–436.
- Sutton, J. F. and W. W. Korth. 1995. Rodents (Mammalia) from the Barstovian (Miocene) Anceney Local Fauna, Montana. *Annals of the Carnegie Museum* 64:267–314.
- Swinehart, J. B., V. L. Souders, H. M. DeGraw, and R. F. Diffendal Jr. 1985. Cenozoic paleogeography of western Nebraska. In *Cenozoic paleogeography of west-central United States*. *Rocky Mountain Section, Society of Economic Paleontologists and Mineralogists*, ed. R. M. Flores and S. S. Kaplan. Denver, CO: Rocky Mountain Paleogeography Symposium 3, pp. 204–229.
- Swisher, C. C. III. 1992.  $^{40}\text{Ar}/^{39}\text{Ar}$  dating and its application to the calibration of the North American land-mammal ages. Ph.D. dissertation, University of California, Berkeley.
- Tedford, R. H. 1970. Principles and practices of mammalian geochronology in North America. Chicago: North American Paleontological Convention, 1969, Proceedings F, pp. 666–703.
- . 1981. Mammalian biochronology of the late Cenozoic basins of New Mexico. *Geological Society of America Bulletin*, Part I, 92:1008–1022.
- . 1999. Rocks and faunas, Ogallala Group, Pawnee Buttes area, Weld County Colorado. In *The Tertiary record of Weld County, northeastern Colorado*, ed. E. Evanoff, R. W. Graham, and R. H. Tedford. *Denver Museum of Natural History and Science Field Guide*, pp. 31–47.
- Tedford, R. H. and S. Barghoorn. 1993. Neogene stratigraphy and mammalian biochronology of the Española Basin, northern New Mexico. *New Mexico Museum of Natural History and Science Bulletin* 2:159–168.
- . 1997. Miocene mammals of the Española and Albuquerque basins, north-central New Mexico. In *New Mexico's fossil record 1*, ed. S. G. Lucas, J. W. Estep, T. E. Williamson, and G. S. Morgan. *New Mexico Museum of Natural History and Science Bulletin* 11:77–95.
- . 1999. Santa Fe Group (Neogene), Ceja del Rio Puerco, Northwestern Albuquerque Basin, Sandoval County, New Mexico. *New Mexico Geological Society Guidebook, 50th Field Conference, Albuquerque Geology*, pp. 327–335.
- Tedford, R. H., T. Galusha, M. F. Skinner, B. E. Taylor, R. W. Fields, J. R. Macdonald, J. M. Rensberger, S. D. Webb, and D. P. Whistler. 1987. Faunal succession and biochronology of the Arikareean through Hemphillian interval (late Oligocene through earliest Pliocene epochs) in North America. In *Cenozoic mammals of North America: Geochronology and biostratigraphy*, ed. M. O. Woodburne. Berkeley: University of California Press, pp. 153–210.
- Tedford, R. H. and M. E. Hunter. 1984. Miocene marine–nonmarine correlations, Atlantic and Gulf Coastal Plains, North America. *Palaeogeography, Palaeoclimatology, Palaeoecology* 47:129–151.
- Tedford, R. H., J. B. Swinehart, R. M. Hunt Jr., and M. R. Voorhies. 1985. Uppermost White River and lowermost Arikaree rocks and faunas, White River Valley, northwestern Nebraska, and their correlation with South Dakota. *Dakoterra* 2:335–352.
- Tedford, R. H., J. B. Swinehart, C. C. Swisher III, D. R. Prothero, S. A. King, and T. E. Tierney. 1996. The Whitneyan–Arikareean transition in the High Plains. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 312–334.
- Tobien, H. 1972. Status of the genus *Serridentinus* Osborn 1923 (Proboscidea, Mammalia) and related forms. *Mainzer Geowissenschaften Mitteilungen* 1:143–191.
- . 1973. On the evolution of mastodonts (Probosciden, Mammalia) Part 1: The bunodont trilophodont groups. *Notizblatt des Hessischen Landesamtes für Bodenforschung zu Weisbaden* 101:202–276.
- Turner, D. L. 1970. Potassium–argon dating of Pacific Coast Miocene foraminiferal stages. *Geological Society of America Special Paper* 124:91–129.

- Vondra, C. F. 1963. *The stratigraphy of the Gering Formation in the Wildcat Ridge in western Nebraska*. Ph.D. dissertation, University of Nebraska, Lincoln.
- Vondra, C. F., C. B. Schultz, and T. M. Stout. 1969. New members of the Gering formation (Miocene) in Western Nebraska, including a geologic map of Wilcat Ridge and related outliers. *Nebraska Geological Survey Paper* 18.
- Voorhies, M. R. 1990a. Vertebrate biostratigraphy of the Ogallala Group in Nebraska. In *Geologic framework and regional hydrology: Upper Cenozoic Blackwater Draw and Ogallala Formations, Great Plains*, ed. T. C. Gustavson. Austin: Bureau of Economic Geology, University of Texas, pp. 115–151.
- . 1990b. Vertebrate paleontology of the proposed Norden Reservoir area, Brown, Cherry and Keyapaha counties, Nebraska. U.S. Bureau of Reclamation, Denver, Technical Report 82-09, Division of Archaeological Research, University of Nebraska, Lincoln, pp. 1–138, A1–A593.
- Voorhies, M. R. and C. L. Timperley. 1997. A new *Pronotolagus* (Lagomorpha: Leporidae) and other leporids from the Valentine Railway quarries (Barstovian, Nebraska), and the archaeologine–leporine transition. *Journal of Vertebrate Paleontology* 17:725–737.
- Walsh, S. L. 1998. Fossil datum terms, paleobiological event terms, paleostratigraphy, chronostratigraphy and the definition of land-mammal “age” boundaries. *Journal of Vertebrate Paleontology* 18:150–179.
- . 2000. Eubiostratigraphic units, quasibiostratigraphic units and “assemblage zones.” *Journal of Vertebrate Paleontology* 20:761–775.
- Wang, X-M. 1994. Phylogenetic systematics of the Hesperocyoninae (Carnivora: Canidae). *American Museum of Natural History Bulletin* 221.
- Wang, X-M., R. H. Tedford, and B. E. Taylor. 1999. Phylogenetic systematics of the Borophaginae (Carnivora: Canidae). *American Museum of Natural History Bulletin* 243.
- Webb, S. D. 1969. The Burge and Minnechaduzza Clarendonian mammalian faunas of north-central Nebraska. *University of California Publications in Geological Sciences* 78:1–191.
- . 1983. The rise and fall of the late Miocene ungulate fauna in North America. In *Coevolution*, ed. M. H. Nitecki. Chicago: University of Chicago Press, pp. 267–306.
- . 2000. Evolutionary history of New World Cervidae. In *Antelopes, deer and relatives: Fossil record, behavioral ecology, systematics, and conservation*, ed. E. S. Vrba and G. B. Schaller. New Haven, CT: Yale University Press, pp. 38–64.
- Webb, S. D. and D. B. Crissinger. 1983. Stratigraphy and vertebrate paleontology of the central and southern phosphate districts of Florida. *Geological Society of America, Southeastern Section, Field Trip Guidebook, Central Florida Phosphate District*, pp. 28–72.
- Webb, S. D. and N. D. Opdyke. 1995. Global climatic influence on Cenozoic land mammal faunas. In *Effects of past global change on life. Studies in geophysics*, ed. J. P. Kennen and S. M. Stanley. Washington, DC: National Academy of Sciences, pp. 184–208.
- Webb, S. D. and S. C. Perrigo. 1984. Late Cenozoic vertebrates from Honduras and El Salvador. *Journal of Vertebrate Paleontology* 4:237–254.
- Whistler, D. P. and D. W. Burbank. 1992. Miocene biostratigraphy and biochronology of the Dove Spring Formation, Mojave Desert, California, and characterization of the Clarendonian mammal age (late Miocene) in California. *Geological Society of America Bulletin* 104:644–658.
- Whitlock, C. and M. R. Dawson. 1990. Pollen and vertebrates of the early Neogene Houghton Formation, Devon Island, Arctic Canada. *Arctic* 43:324–330.
- Whitmore, F. C. Jr. and R. H. Stewart. 1965. Miocene mammals and Central American seaways. *Science* 148:180–185.
- Williams, M. R. and J. E. Storer. 1998. Cricetid rodents of the Kealey Springs Local Fauna (early Arikareean, late Oligocene) of Saskatchewan. *Paludicola* 1:143–149.
- Wilson, J. A. 1967. Additions to El Gramal Fauna Nejapa, Oaxaca, Mexico. *Sociedad Geológica Mexicana Boletín* 30:1–4.
- Wilson, J. A. and S. E. Clabaugh. 1970. A new Miocene formation and a description of volcanic rocks, northern Valley of Oaxaca, State of Oaxaca. *Sociedad Geológica Mexicana, Guidebook, Excursion Mexico–Oaxaca*, pp. 120–128.
- Wilson, L. L. and D. R. Prothero. 1997. Magnetic stratigraphy and tectonic rotations of the middle–upper Miocene “Santa Margarita” and Chanac formations, north-central Transverse Ranges, California. In *Geology of the western cordillera: Perspectives from undergraduate research*, ed. G. H. Girty, R. E. Hanson, and J. D. Cooper. *Pacific Section, Society of Economic Paleontologists and Mineralogists* 82:35–48.
- Wilson, R. L. 1968. Systematics and faunal analysis of a lower Pliocene vertebrate assemblage from Trego County, Kansas. *The University of Michigan Contributions from the Museum of Paleontology* 22:75–126.
- Wilson, R. W. 1949. Rodents and lagomorphs of the Upper Sespe. *Carnegie Institution of Washington Publication* 584:53–65.
- . 1960. Early Miocene rodents and insectivores from north-western Colorado. *University of Kansas Paleontological Contributions, Vertebrata*, Article 7, pp. 1–92.
- Wood, H. E., R. W. Chaney, J. Clark, E. H. Colert, G. L. Jepsen, J. B. Reeside Jr., and C. Stock. 1941. Nomenclature and correlation of the North American continental Tertiary. *Geological Society of America Bulletin* 52:1–48.
- Woodburne, M. O. 1996. Precision and resolution in mammalian chronostratigraphy: Principles, practices, examples. *Journal of Vertebrate Paleontology* 16:531–555.
- . 1998. Arikareean and Hemingfordian faunas of the Cady Mountains, Mojave Desert Province, California. In *Depositional environments, lithostratigraphy and biostratigraphy of the White River and Arikaree groups (late Eocene to early Miocene, North America)*, ed. D. O. Terry Jr., H. E. LaGarry, and R. M. Hunt Jr. *Geological Society of America Special Paper* 325:197–210.
- Woodburne, M. O. and P. T. Robinson. 1977. A new late Hemingfordian mammal fauna from the John Day Formation, Oregon, and its stratigraphic implications. *Journal of Paleontology* 51:750–757.
- Woodburne, M. O. and C. C. Swisher III. 1995. Land mammal high-resolution geochronology, intercontinental overland dispersals, sea level, climate and vicariance. In *Geochronology, time scales, and global stratigraphic correlation*, ed. W. A. Berggren, D. V. Kent, M.-P. Aubry, and J. Hardenbol. *Society for Sedimentary Geology Special Publication* 54:335–364.
- Woodburne, M. O., R. H. Tedford, M. S. Stevens, and B. E. Taylor. 1974. Early Miocene mammalian faunas, Mojave Desert, California. *Journal of Paleontology* 48:6–26.

- Woodburne, M. O., R. H. Tedford, and C. C. Swisher III. 1990. Lithostratigraphy, biostratigraphy and geochronology of the Barstow Formation, Mojave Desert, southern California. *Geological Society of America Bulletin* 102:459–477.
- Wright, D. B. and R. E. Eshelman. 1987. Miocene Tayassuidae (Mammalia) from the Chesapeake Group of the Mid-Atlantic Coast and their bearing on marine–nonmarine correlation. *Journal of Paleontology* 61:604–618.
- Xu, X.-F. 1996. Castoridae. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 417–432.
- Zheng, J.-Y. 1996. *Magnetostratigraphy of a Miocene sedimentary sequence in Railroad Canyon, Idaho*. Ph.D. dissertation, University of Pittsburgh.

# 7

## The Blancan, Irvingtonian, and Rancholabrean Mammal Ages

Christopher J. Bell and Ernest L. Lundelius Jr.  
(co-chairmen), Anthony D. Barnosky,  
Russell W. Graham, Everett H. Lindsay,  
Dennis R. Ruez Jr., Holmes A. Semken Jr.,  
S. David Webb, and Richard J. Zakrzewski

**T**HIS CHAPTER EXAMINES the last three North American land mammal ages of the Cenozoic: the Blancan, Irvingtonian, and Rancholabrean. It also incorporates the arvicoline rodent biochronology that was a separate chapter in the first edition of this volume (Repenning 1987). These mammal ages encompass approximately the last 5 million years and span most of the Pliocene and all of the Pleistocene (the latest Hemphillian is part Pliocene and is covered by Tedford et al., chapter 6, this volume). Vertebrate faunas from these epochs are known from several thousand localities in North America and are distributed over a wider geographic area than those of any other epoch or equivalent interval of time. The short duration of the Pliocene and Pleistocene and the availability of numerous and diverse dating methods (Noller et al. 2000) permit a refined temporal resolution and critical evaluation of faunal change on much shorter time scales than is possible for most of the Cenozoic. In addition, many of the taxa represented in Pliocene and Pleistocene faunas are still extant or have close living relatives (this is particularly true for the latter part of the Pleistocene). This temporal proximity between fossil and living forms creates a unique bridge between neontology and paleontology and provides a valuable perspective on the biological components that add to the complexities of biostratigraphy and biochronology. Although there is extensive evidence of evolutionary change in many Pleistocene mammal lineages, the relative paucity of cladogenic speciation probably is a result of the short time involved. Detailed excavations of fossil deposits that accumulated over sometimes quite short periods of time reveal that complex faunal changes took place over short

time intervals. These deposits thus permit a greater understanding of the interaction between biotic and abiotic factors in shaping mammalian faunal dynamics and provide a broader perspective on the implications of time-averaging and provinciality for biochronology. These implications are relevant not only for the Pliocene–Pleistocene but also for all the Cenozoic.

The problems involved in the use of vertebrate fossils for latest Tertiary and Quaternary geochronology are similar to those of older time periods. Additional complications are caused by strong environmental zonation that resulted from climatic changes due to repeated pulses of glacial advance and retreat. The boundaries of these environmental zones shifted over short time intervals and were often associated with dramatic faunal changes.

Long, richly fossiliferous sequences with well-dated superpositional control provide the ideal basis for studying faunal succession in the Pliocene and Pleistocene of North America. We are fortunate to have several complementary sections in the West and Midwest that provide an adequate geochronologic framework for the study of late Cenozoic mammalian biostratigraphy. These include deposits in Meade County in Kansas, the western Snake River Plain of Idaho, the San Pedro Valley of Arizona, the Gila River Valley in southeastern Arizona and western New Mexico, the Medicine Hat sequence in Alberta, the Anza-Borrego Desert in the Salton Sea Basin of southern California, the San Timoteo Badlands of California, and the Ringold Formation of southeastern Washington. Geochronologic data from these and other isolated localities make the North American Pliocene and

Pleistocene mammal ages, despite their limitations, among the best dated in the world.

External age control of Pliocene and Pleistocene faunas comes primarily from radioisotopic dating techniques and paleomagnetism. Volcanic ash, pumice, and rhyolite units significant for correlation of these faunas are summarized in table 7.1. The dates of the geomagnetic polarity chrons are subject to revision as improved methods and procedures are developed and as new stratigraphic sections are discovered and studied. To retain some measure of consistency in this volume we follow Berggren et al. (1995) for the dates of geomagnetic polarity event boundaries. In a few cases, radiometric dates of a particular sequence conflict with the global Geomagnetic Polarity Time Scale (GPTS) proposed by Berggren et al. (1995); these cases are discussed individually. When citing prior publications in which time designations for faunas were based on older geomagnetic polarity stratigraphy calibrations, we altered the published designation to conform to that of Berggren et al. (1995); in those cases our ages do not necessarily match those given in the earlier publications.

Four magnetic polarity chrons were initially established for the Pliocene and Pleistocene (Cox et al. 1963); from oldest to youngest these are the Gilbert, Gauss, Matuyama, and Brunhes. These names, and the names of associated subchrons (e.g., Jaramillo, Olduvai), are now firmly established in the literature and are used in this chapter. The numeric designations proposed for chrons from the Miocene through Mesozoic also are standardized (Opdyke and Channell 1996) and are extended through to today (e.g., in that system the Brunhes chron is redesignated chron C<sub>1n</sub>). For completeness and clarity, we provide the numeric chron and subchron designations in addition to the traditional named chrons.

The correlation of the North American mammal ages with the epochs of the geologic time scale was difficult to establish previously, especially for the Pliocene through Holocene. Resolution of debates over the epochal boundaries and subsequent designation of stratotypes for these boundaries in the last two decades provide new opportunities to evaluate temporal correlation between defined European epochal stratotypes and the North American mammal ages. In this chapter, we provide a brief summary of epochal boundary stratotypes and current interpretations of their ages. Subsequent sections of this chapter address the Blancan, Irvingtonian, and Rancholabrean mammal ages in terms of their history, definition, characterization, and temporal extent and the geographic distribution of relevant faunas for each age. Summary characterizations are shown in table 7.2.

## LATE CENOZOIC EPOCHAL BOUNDARIES

### THE MIOCENE-PLIOCENE BOUNDARY

The complicated early history of the use of the term *Pliocene*, from its original definition by Lyell (1833) through the early twentieth century, was reviewed by Wilmarth (1925). In recent usage, the base of the Pliocene was placed at the base of the Zanclean Stage at Capo Rossello, Sicily, marked by an acknowledged unconformity between the Trubi Formation (Trubi marls) and the underlying alluvial/lacustrine Arenazzolo Formation of the Messinian Stage (Cita 1975). This boundary was interpreted to mark the return of marine conditions to the Mediterranean after the Messinian salinity crisis. The lack of good magnetostratigraphy in the proposed type section was pointed out by Rio et al. (1991), who also indicated that the Miocene strata at Capo Rossello are non-marine (making biostratigraphic correlation to the marine section difficult) and suggested the need for a boundary stratotype outside the Mediterranean to avoid the problem of endemism in the marine faunas. The Capo Rossello section does not satisfy the operational requirements of a global boundary and stratotype section because of the unconformity at the base of the Trubi Formation at that locality (Hilgen and Langereis 1993). An alternative boundary in the Eraclea Minoa (Sicily) section shows evidence of a transitional interval rather than an unconformity at the base of the Trubi Formation (Hilgen and Langereis 1993). The evaporite cycles in the Trubi Formation are tuned to the astronomical precession cycles and thereby provide a presumably precise determination of the date. There are five precession cycles between the base of the Thvera subchron (chron C<sub>3n.4n</sub>) and the basal contact of the Zanclean. With an average cycle length of 21,000 years, the base of the Zanclean therefore is approximately 105,000 years older than the base of the Thvera (given as 5.23 Ma by Berggren et al. 1995). The age of the base of the Pliocene therefore would be considered 5.335 Ma using the Berggren et al. (1995) GPTS. This boundary was ratified by the International Commission on Stratigraphy in January 2000 and now is established as the base of the Pliocene series (see Van Couvering et al. 2000 for discussion).

### THE PLIOCENE-PLEISTOCENE BOUNDARY

**International Stratotype** The term *Pleistocene* had a complicated early history (reviewed by Wilmarth 1925), but modern usage corresponds roughly to the definition

**TABLE 7.1** Late Cenozoic Volcanic Units Useful in Correlating North American Mammal Faunas

<b>NAME</b>	<b>PUBLISHED AGES</b>	<b>REFERENCES</b>
Lava Creek B Ash (Pearlette type O)	0.602 ± 0.004 Ma Between 0.66 ± 0.01 and 0.67 ± 0.01 Ma	Gansecki et al. 1998 Izett and Honey 1995; Izett et al. 1992
Hartford Ash	Approximately 0.74 Ma	Boellstorff 1973, 1978
Bishop Ash	0.7589 ± 0.0018 Ma Approximately 0.70 Ma	Sarna-Wojcicki et al. 2000 Izett et al. 1970
Tsankawi Pumice Bed (lower bed of the Tshirege Member of the Bandelier Tuff)	1.22 ± 0.018 Ma 1.12 ± 0.03 Ma	Izett and Obradovich 1994 Izett et al. 1981
Cerro Toledo Rhyolite (upper)	1.23 ± 0.02 Ma	Izett et al. 1981
Cerro Toledo "X" Ash (suspected to be derived from Cerro Toledo Rhyolite)	1.2–1.3 Ma (by tentative correlation)	Izett 1977; G. Schultz 1990b
Mesa Falls Ash (Pearlette type S, Coleridge Ash, "Sappa ash")	1.293 ± 0.012 Ma 1.27 Ma (average) 1.21 Ma 1.21 ± 0.05 Ma 1.20 ± 0.04 Ma About 1.2 Ma	Gansecki et al. 1998 Izett 1981 Boellstorff 1973 Boellstorff 1976 Naeser et al. 1973 Naeser et al. 1971
Cerro Toledo "B" Ash (from eruptions associated with Cerro Toledo Rhyolite)	Between 1.2 and 1.4 Ma Probably close to 1.4 Ma (by correlation)	Izett et al. 1981 Izett and Honey 1995
Cerro Toledo Rhyolite (lower)	1.47 ± 0.04 Ma	Izett et al. 1981
Cerro Toledo Rhyolites (as a whole)	Pulse eruptions between approximately 1.54 and 1.22 Ma	Spell et al. 1996
Guaje Pumice Bed (lower bed of the Otowi Member of the Bandelier Tuff)	1.608 ± 0.01 Ma 1.613 ± 0.011 Ma 1.40 ± 0.04 Ma	Spell et al. 1996 Izett and Obradovich 1994 Izett et al. 1981
"Guaje ash" (informal) near Mt. Blanco in Texas (chemical and petrographic resemblance to Guaje Pumice Bed in New Mexico)	1.77 ± 0.44 Ma 1.4 ± 0.2 Ma	Boellstorff 1976 Izett et al. 1972
Huckleberry Ridge Ash (Pearlette type B, "Borchers ash")	2.003 ± 0.014 Ma 2.10 ± 0.02 Ma 2.09 ± 0.01 Ma 2.09 ± 0.01 Ma 2.01 Ma 1.96 ± 0.2 Ma	Gansecki et al. 1998 Honey et al. 1998 Izett and Honey 1995 Obradovich and Izett 1991 Izett 1981 Boellstorff 1976
Blanco Ash	2.8 ± 0.3 Ma	Boellstorff 1976
Peters Gulch Ash	3.75 ± 0.36 Ma	Izett 1981
Lawlor Tuff	4.1 ± 0.2 Ma	Sarna-Wojcicki et al. 1991
Healdsburg tephra	4.69 Ma or 4.64 ± 0.03 Ma	Lindsay et al. 2002
"Tracer pumice bed," Nevada	4.96 ± 0.02 Ma	Lindsay et al. 2002
Pinole Tuff	5.2 ± 0.1 Ma	Evernden et al. 1964

The names used in this chapter are provided with alternative names in parentheses. Ages and references for the dates are provided.

**TABLE 7.2** Characteristic Taxa of the Blancan, Irvingtonian, and Rancholabrean Mammal Ages

A, begins in mammal age but also persists after it; B, begins before mammal age but persists into it; C, confined to mammal age; 5T, species with five closed triangles on lower first molar. Asterisk (\*) indicates taxa that first appear in the latest Blancan faunas that are elsewhere often considered to be early Irvingtonian.

	BLANCAN	IRVINGTONIAN	RANCHOLABREAN
<i>Megalonyx leptostomus</i>	C		
<i>Glossotherium chapadmalense</i>	C		
<i>Aluralagus</i>	C		
<i>Pewelagus dawsonae</i>	C		
<i>Pratilepus kansasensis</i>	C		
<i>Dipoides rexroadensis</i>	C		
<i>Sigmodon curtisi</i>	C		
<i>Sigmodon minor</i>	C		
<i>Mictomys vetus</i>	C		
<i>Ogmodontomys</i>	C		
<i>Ophiomys</i>	C		
<i>Pliopotamys</i>	C		
<i>Ondatra idahoensis</i>	C		
<i>Pliolemmus</i>	C		
<i>Borophagus diversidens</i>	C		
<i>Canis lepophagus</i>	C		
<i>Chasmaporthetes</i>	C		
<i>Ursus abstrusus</i>	C		
<i>Platygonus bicalcaratus</i>	C		
<i>Platygonus pearcei</i>	C		
<i>Nannippus peninsulatus</i>	C		
<i>Alilepus</i>	B		
<i>Hypolagus</i>	B		
<i>Nekrolagus</i>	B		
<i>Paenemarmota</i>	B		
<i>Prodipodomys</i>	B		
<i>Satherium</i>	B		
<i>Borophagus hilli</i>	B		
<i>Trigonicictis</i>	B		
<i>Megantereon</i>	B		
<i>Rhynchotherium</i>	B		
<i>Nothrotheriops*</i>	A	B	B
<i>Paramylodon harlani*</i>	A	B	B
<i>Glyptotherium</i>	A	B	B
<i>Holmesina</i>	A	B	B
<i>Blarina</i>	A	B	B
<i>Sylvilagus</i>	A	B	B
<i>Lepus*</i>	A	B	B
<i>Allophaiomys pliocaenicus*</i>	A	B	
<i>Microtus (5T)*</i>	A	B	B
<i>Mictomys kansasensis/meltoni*</i>	A	B	
<i>Neofiber*</i>	A	B	B
<i>Ondatra annectens*</i>	A	B	
<i>Phenacomys*</i>	A	B	B
<i>Synaptomys*</i>	A	B	B
<i>Sciurus*</i>	A	B	B
<i>Lutra canadensis*</i>	A	B	B
<i>Homotherium*</i>	A	B	
<i>Miracinonyx inexpectatus</i>	A	B	

TABLE 7.2 (continued)

	BLANCAN	IRVINGTONIAN	RANCHOLABREAN
<i>Smilodon gracilis</i>	A	B	
<i>Canis edwardii</i> *	A	B	
<i>Platygonus vetus</i> *	A	B	
<i>Stegomastodon</i>	A	B	
<i>Mammut americanum</i>	A	B	B
<i>Microtus llanensis</i>		C	
<i>Microtus meadensis</i>		C	
<i>Microtus paroperarius</i>		C	
<i>Canis armbrusteri</i>		C	
<i>Tetrameryx irvingtonensis</i>		C	
<i>Didelphis</i>		A	B
<i>Brachylagus idahoensis</i>		A	B
<i>Sylvilagus palustris</i>		A	B
<i>Clethrionomys</i>		A	B
<i>Lemmiscus curtatus</i>		A	B
<i>Ondatra zibethicus</i>		A	B
<i>Marmota flaviventris</i>		A	B
<i>Marmota monax</i>		A	B
<i>Cynomys gunnisoni</i>		A	B
<i>Cynomys ludovicianus</i>		A	B
<i>Panthera onca</i>		A	B
<i>Smilodon populator</i>		A	B
<i>Mustela erminea</i>		A	B
<i>Brachyprotoma</i>		A	B
<i>Conepatus</i>		A	B
<i>Canis latrans</i>		A	B
<i>Canis lupus</i>		A	B
<i>Arctodus simus</i>		A	B
<i>Euceratherium</i>		A	B
<i>Oreamnos</i>		A	B
<i>Mammuthus</i>		A	B
<i>Megalonyx jeffersonii</i>			C
<i>Platygonus compressus</i>			C
<i>Canis dirus</i>			C
<i>Panthera atrox</i>			C
<i>Miracinonyx trumani</i>			C
<i>Aplodontia rufa</i>			A
<i>Ovis canadensis</i>			A
<i>Bison</i>			A
<i>Rangifer tarandus</i>			A
<i>Felis concolor</i>			A
<i>Vulpes velox</i>			A
<i>Homo sapiens</i>			A

proposed by Forbes (1846:386), who equated the Pleistocene with the glacial deposits of England. The concept of worldwide Pleistocene glaciation followed soon thereafter, and the recognition of multiple glaciations on a worldwide scale was seen as a means to establish temporal divisions within the Pleistocene.

A more precise and traditional placement for the base of the Pleistocene was given in 1948 by the 18th International Geological Congress. By their edict, the Lower Pleistocene Series included the marine Calabrian Stage of Gignoux (1910, 1916), with its recognized terrestrial equivalent, the Villafranchian Stage, as its basal member. The



Pliocene–Pleistocene Commission of that congress noted that this boundary coincided with evidence of climatic cooling in the Italian Neogene succession. Some significant changes in the planktonic foraminiferal record ultimately were driven by climatic change, and Berggren and Van Couvering (1974) correlated these changes with the GPTS. Their efforts provided an important means of simultaneously including paleontological, paleomagnetic, and climate change data in a discussion of the boundary.

Disagreement over the relative importance of climatic events in the determination of the Pliocene–Pleistocene boundary stimulated an extensive debate over the placement of the boundary. Recent arguments presented by Morrison (1991) pertaining to the boundary were based in part on a desire to identify the boundary with a significant climatic event. However, a strict reliance on climatic criteria for establishing boundaries is not advisable because of the time-transgressive nature and wide geographic variance of climatic conditions.

The placement of the Pliocene–Pleistocene boundary at Vrica, Italy, at the base of the claystone conformably overlying sapropelic marker bed “e” was adopted by the International Geological Correlation Project 41 and the International Union for Quaternary Research (INQUA) Sub-Commission 1-d at the 27th International Geological Congress at Moscow in 1984. The boundary is situated just above the top of the Olduvai normal polarity subchron (chron 2n; now dated at 1.77 Ma; Cande and Kent 1995; Berggren et al. 1995). The age of the boundary therefore is slightly younger than 1.77 Ma. An age of 1.8 Ma, derived by rounding off the upper age of the Olduvai event to one decimal place, was given by Pasini and Colalongo (1997). This date is close to several microplankton events and seems to coincide with the entrance of the mollusk *Arctica islandica* into the Mediterranean (the historical basis for recognizing the base of the Pleistocene in Italy). This boundary was ratified and adopted as a Global Stratotype Section and Point (GSSP) by the International Union of Geological Sciences in 1985 (Bassett 1985). Because at present it is not possible to resolve time intervals as short as 30,000 years in 2-m.y.-old sediments, an age of 1.8 Ma can be accepted as an approximation for the age of the Pliocene–Pleistocene boundary, but we emphasize that the actual age is slightly younger than 1.77 Ma. A more detailed history of the dispute over the placement of the Pliocene–Pleistocene boundary was provided by Berggren and Van Couvering (1974) and Van Couvering (1997).

**North American Boundary Recognition** Recognition of the Pliocene–Pleistocene boundary in North America ultimately must be based on the boundary as defined in

a marine section in Italy. Early efforts to recognize the Pliocene–Pleistocene boundary in North America were based on the traditional concept that this boundary was marked by the initiation of widespread glaciation. Consequently, North American stratigraphers turned their attention to the extensive glacial deposits in the northern plains and midwestern states, where the most complete glacial sequence is preserved.

Temporal relationships among Pleistocene terrestrial deposits in North America traditionally were based on the recognition of four major glacial pulses, designated (from oldest to youngest) the Nebraskan, Kansan, Illinoian, and Wisconsinan. Interglacial units between these major glacial pulses were called (from oldest to youngest) Aftonian, Yarmouthian, and Sangamonian. Study of deep sea stratigraphic cores, application of new radiometric dating techniques, and detailed stratigraphic reconstructions in the Great Plains reveal that this traditional classification is oversimplified; as many as 20 discrete glacial pulses over the last 2 million years are now recognized (e.g., Richmond and Fullerton 1986).

Fission-track dates of volcanic ash deposits in stratigraphic association with classic glacial till sequences demonstrate that continental glaciation in the New World predates 2.2 Ma (Boellstorff 1978). Especially significant in this respect is Nebraska Geological Survey core 5-A-75, recovered from a well near Afton, Iowa (Boellstorff 1978). This core revealed at least two tills underlying the classic Nebraskan till exposed in the region. The lowermost till in this core is overlain by a “Pearlette-type” ash dated to 2.2 Ma by Hallberg and Boellstorff (1978). Thus the “Nebraskan” till certainly does not represent the earliest North American continental glaciation, and the differentiation of at least seven tills in the type “Nebraskan–Kansan” sequence demonstrates that these terms are meaningless where they were defined originally (Boellstorff 1978). Ashes dated at approximately 0.60–0.67 Ma (“Type O” or “Lava Creek B” of Izett et al. 1981; Izett and Wilcox 1982; “Pearlette restricted” of Boellstorff 1973) and 0.74 Ma (“Hartford Ash” of Boellstorff 1973, 1978) lie stratigraphically above classic Nebraskan till and below classic Kansan till (Hallberg 1980). The Cudahy Fauna in Kansas also lies immediately below the 0.60–0.67 Ma Lava Creek B Ash. Although this fauna is approximately the age of classic “Aftonian” sediments, it traditionally was considered to be “late Kansan” in age (Hibbard 1970; Paulson 1961).

The age and stratigraphic relations of these ashes clearly indicate that traditional glacial age designations of High Plains faunas are misaligned with respect to the till sequences and that the terms *Nebraskan*, *Aftonian*, and

*Kansan* have lost meaning as chronologic units (see also Richmond and Fullerton 1986). Similar problems are associated with the use of *Yarmouthian*, *Illinoian*, and *Sangamonian*. It is questionable whether climatic criteria that lack independent chronologic calibration are a sound basis for any chronostratigraphic or geochronologic unit. Thus, use of this nomenclature with Pleistocene faunas should be abandoned. A possible exception to this general statement is the Wisconsinan glacial interval, which is unique among the traditional terms because it is reliably and accurately dated by radiocarbon dating and is still in wide use.

Correlations for the base of the Pleistocene in North America must be established on evidence independent of local climate and ultimately must be correlated with the stratotype base in Italy. Although paleontological data relevant to the establishment of the boundary stratotype are derived from marine organisms, land mammal faunas do contribute significantly to correlations. European terrestrial mammalian faunas were successfully correlated with the classic Italian marine sequences as early as the late 1960s (Hürzeler 1967; Savage and Curtis 1970; Azzaroli and Ambrosetti 1970; Azzaroli and Berzi 1970). Terrestrial mammalian faunas in North America and Italy can be correlated, at least loosely, with roughly contemporaneous mammalian faunas elsewhere in Europe and Asia. In addition to faunal correlations, direct radiometric dates and paleomagnetic stratigraphy are used to correlate mammalian faunas within and between North America and Europe (Repenning and Fejfar 1977; Repenning et al. 1990; Azzaroli et al. 1997; Fejfar et al. 1998). Well-dated faunas in turn form the foundation for correlating North American faunas that lack independent age control.

#### PLEISTOCENE-HOLOCENE BOUNDARY

Lyell (1833) defined the Recent based on the appearance of humans, but the term *Holocene* was first used by Gervais (1869:32), who gave no definition other than that it follows the “diluvial or Pleistocene deposits.” Modern authors equate Lyell’s Recent with Gervais’s Holocene, but there is no agreement on the placement of a boundary between the Pleistocene and Holocene. Holocene sediments are entirely within the Brunhes chron (chron C<sub>1n</sub>), and although there is polar wander and minor fluctuations in geomagnetic paleointensity during this interval, these data are not yet appropriate for deriving potential boundary markers (Opdyke and Channell 1996; Stoner et al. 1998, 2002). Four alternatives for the place-

ment of the boundary were noted by the 6th INQUA subcommittee for the Holocene (de Jong 1965): the beginning of the Bølling (13 ka), the beginning of the Allerød (11.8 ka), between the Younger Dryas and Preboreal (between zones III and IV of the pollen chronologic sequence, or at approximately 10 ka), and at the disappearance of large, extinct mammals in Europe (we followed Mangerud et al. 1974 for approximate boundary ages of Bølling and Allerød). The terms *Bølling*, *Allerød*, *Younger Dryas*, and *Preboreal* are derived from the pollen and plant sequence in Scandinavia (see Flint 1971 and Mangerud et al. 1974 for summaries). The beginning of the Allerød marks the beginning of deposition of autochthonous continental organic remains after the recession of the last ice sheet in northwestern Europe (a biotic event tied to a climatic event in a localized area) and was proposed by Neustadt (1967) as an appropriate boundary. The term *Younger Dryas* is widely used outside Scandinavia for a cool interval corresponding in age to the Younger Dryas in the type area (Rodbell 2000). Its recognition is not on the basis of mammalian faunas but on pollen or geochemical data. The applicability of these terms in North America and elsewhere outside of Scandinavia is questionable (Bennett et al. 2000; but see Mayle and Cwynar 1995). A similar European nomenclature (the Blytt–Sernander sequence) was reported to be applicable to the Holocene worldwide (Bryson et al. 1970), and it is used in North America, especially in archaeological literature (e.g., Semken and Falk 1987).

Based on data from the deep sea record, Morrison (1991) suggested that the boundary should be placed between oxygen isotope stages 2 and 1. This transition usually is radiocarbon dated to between 11 and 12 ka, but on a worldwide basis this boundary is time transgressive between 9 and 13 ka. There is currently no agreed-upon stratotype for the Pleistocene–Holocene boundary. Operationally, most North American workers appear to be using either an arbitrary age of 10 ka or the disappearance of large-bodied extinct mammals to mark the boundary. The latter event is widely recognized to be time-transgressive between about 23 and 9.5 ka (Grayson 1989, 1991).

Interest in a nontraditional and arbitrary boundary, expressed in terms of thousands of years, has been in the literature for decades (de Jong 1965). The Holocene Commission of INQUA in 1973 confirmed the Paris decision of 1969 to place the base of the Holocene at 10,000 years B.P. and recommended that the international boundary stratotype be located in southern Sweden (Fairbridge 1974). Hopkins (1975) pointed out that eight contributors

to the *Quaternary of the United States* volume (Wright and Frey 1965) placed the boundary in seven positions over a span of 6000 years and that a date within every millennium from 18,000 to 4000 has been proposed. He suggested adoption of an arbitrary boundary at 10 ka as a compromise "because it is a nice round number" (Hopkins 1975:10). That compromise was adopted as a provisional date by Richmond and Fullerton (1986) with the notation that it is a chronometric boundary without a stratigraphic basis. That proposal does not meet the requirement of the International Stratigraphic Code that a chronostratigraphic boundary of this rank be based on an internationally accepted stratotype, but we accept it here as a working definition that is in keeping with general operational practice.

### FAUNAL CORRELATIONS

The problem of faunal contemporaneity and the duration of dispersal events and transitions can best be resolved by dating methods that are independent of the faunas themselves. Figure 7.1 places the major Blancan, Irvingtonian, and Rancholabrean mammal faunas of North America into a chronologic framework based on faunal correlations, volcanic ash beds, radiometric dates, and correlations with the GPTS. We recognize that many other faunas are known from other locations on the continent (e.g., Alaska, the Yukon, Mexico, Greenland), but because most of them lack reliable stratigraphic, radiometric, or paleomagnetic data on which to base their ages and because of the probability of diachrony across wide geographic and ecological boundaries, many were omitted from figure 7.1. Recent summaries of the Pliocene and Pleistocene mammalian faunas from other parts of North America can be found in the following sources: Greenland (Bennike 1997), Alaska (Guthrie and Matthews 1971; Repenning et al. 1987; Repenning and Brouwers 1992), Canada (Churcher 1984a; Harington 1978, 1989, 1990a, 1990b; Harington and Clulow 1973; Morlan 1984, 1996; Stalker 1996), Mexico (Ferrusquía-Villafranca 1978; Carranza-Castañeda and Ferrusquía-Villafranca 1978; Miller 1980; Lindsay 1984; Carranza-Castañeda and Miller 1996, 1998, 2000; Miller and Carranza-Castañeda 1984, 1998a, 1998b; Kowallis et al. 1998). Some of the faunas shown in figure 7.1 are tentatively placed because of inadequate dating or faunal correlation but are included because they are of historical significance or are persistently referenced in the litera-

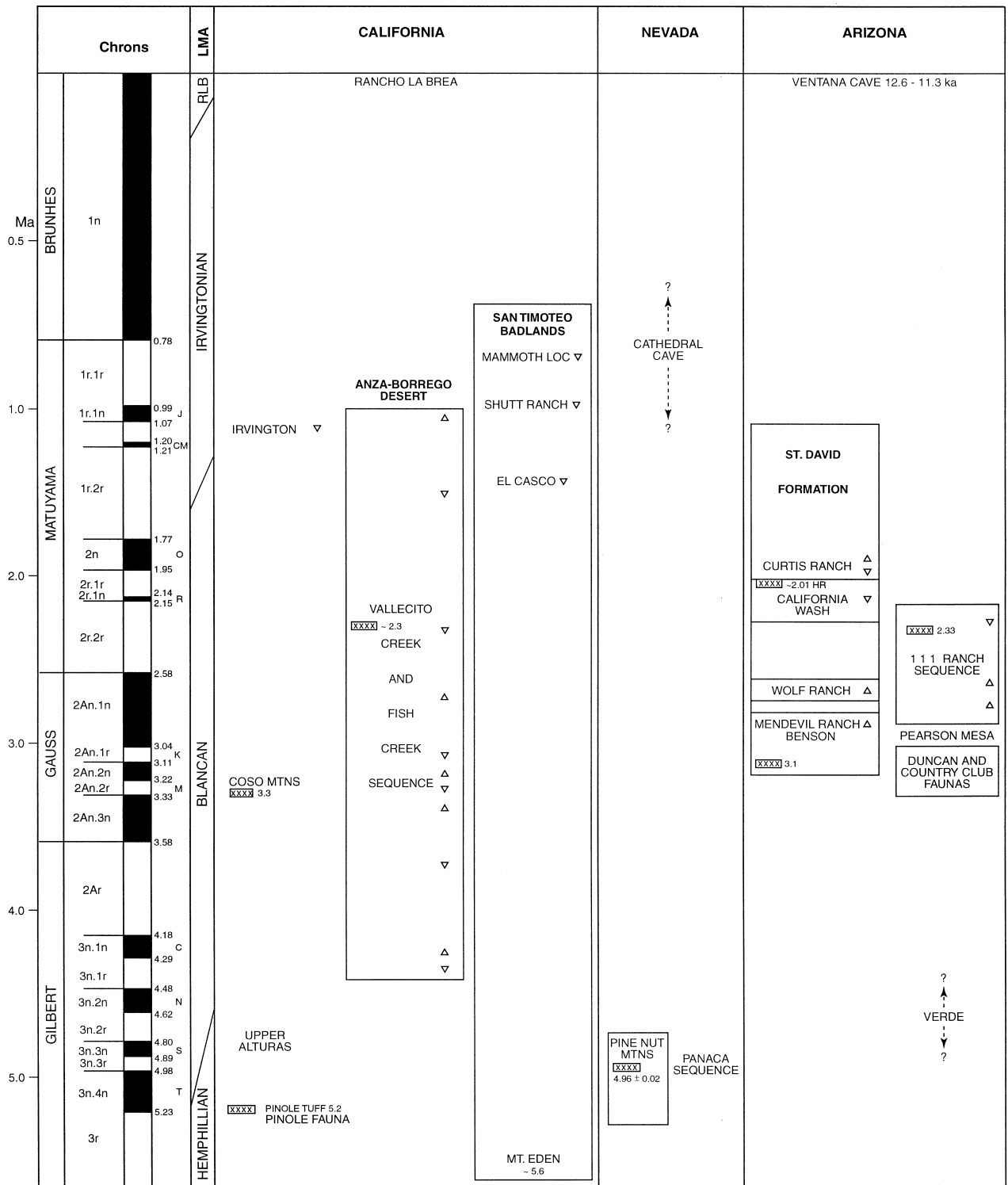
ture. A range chart for selected taxa discussed in this chapter is provided in figure 7.2.

### TAXONOMIC CONSIDERATIONS

Like all healthy sciences, mammalian biostratigraphy and biochronology develop with continued efforts in the field and laboratory. Although all biostratigraphers deal with fossils preserved in the rock record, alternative perspectives on the systematic relationships of taxa and their evolutionary histories can lead to alternative, sometimes confusing taxonomic treatments of the same specimens. New philosophical and methodological approaches to the recognition and naming of fossil taxa are now available (Rowe 1987; Rowe and Gauthier 1992; de Queiroz 1992) and are in wide use, but these methods are rarely applied in research on Pliocene and Pleistocene mammals.

The differing perspectives on the evolutionary history and taxonomy of the voles, lemmings, and muskrats, collectively called microtines or arvicoline, are of special significance for this chapter because of their historical importance in North American Pliocene and Pleistocene biochronology. Controversy over the taxonomy and systematic relationships of arvicoline rodents complicates attempts to summarize their evolutionary history in North America and to establish unambiguous geochronologic boundaries based on various members of the group. Unfortunately, these complications sometimes overshadow the potential importance of these rodents in biostratigraphic correlations and biochronologic interpretations of Pliocene and Pleistocene North American faunas.

The higher-level taxonomy (e.g., family, subfamily, tribe, genus) of the group is especially complicated. Most neontologists now classify these animals in the family Muridae, subfamily Arvicolinae (following Carleton and Musser 1984; Musser and Carleton 1993), and some paleontologists adopted this classification in their work (McKenna and Bell 1997). Earlier systems placed these rodents either in the family Muridae, subfamily Microtinae (Hinton 1926; Hall 1981); family Cricetidae, subfamily Microtinae (Simpson 1945; Gromov and Polyakov 1992; Corbet 1978); or family Cricetidae, with several subfamilies (Repenning et al. 1990). Some paleontologists often classify the group in the family Arvicolidae (Chaline 1987; Fejfar and Heinrich 1990; R. Martin et al. 2000). Korth (1994) recognized a family Microtidae but classified it in the family Cricetidae. The informal terms *microtine* and



**FIGURE 7.1** Correlation of selected Pliocene and Pleistocene faunas discussed in the text. Positions and radiometric ages of key ash beds are indicated by “xxxx.” Faunas with known paleomagnetic orientations are indicated by triangles to the right of faunal name, upright for normal polarity and inverted for reversed polarity. The dates of the geomagnetic polarity boundaries follow Berggren et al. (1995). LMA, land mammal age; RLB, Rancholabrean. Geomagnetic polarity subchrons: C, Cochiti; CM, Cobb Mountain; J, Jaramillo; K, Kaena; M, Mammoth; N, Nunivak; O, Olduvai; R, Reunion; S, Sidufjall; T, Thvera. Key ash and lava beds: Ba, Blanco Ash; CT-B, Cerro Toledo B Ash; CT-X, Cerro Toledo X Ash; DGL, Deer Gulch Lava; Ga, Guaje Ash; HR, Huckleberry Ridge Ash; HA, Hall Ash; HT, Hartford Ash; LC-B, Lava Creek B Ash; PG, Peters Gulch Ash.

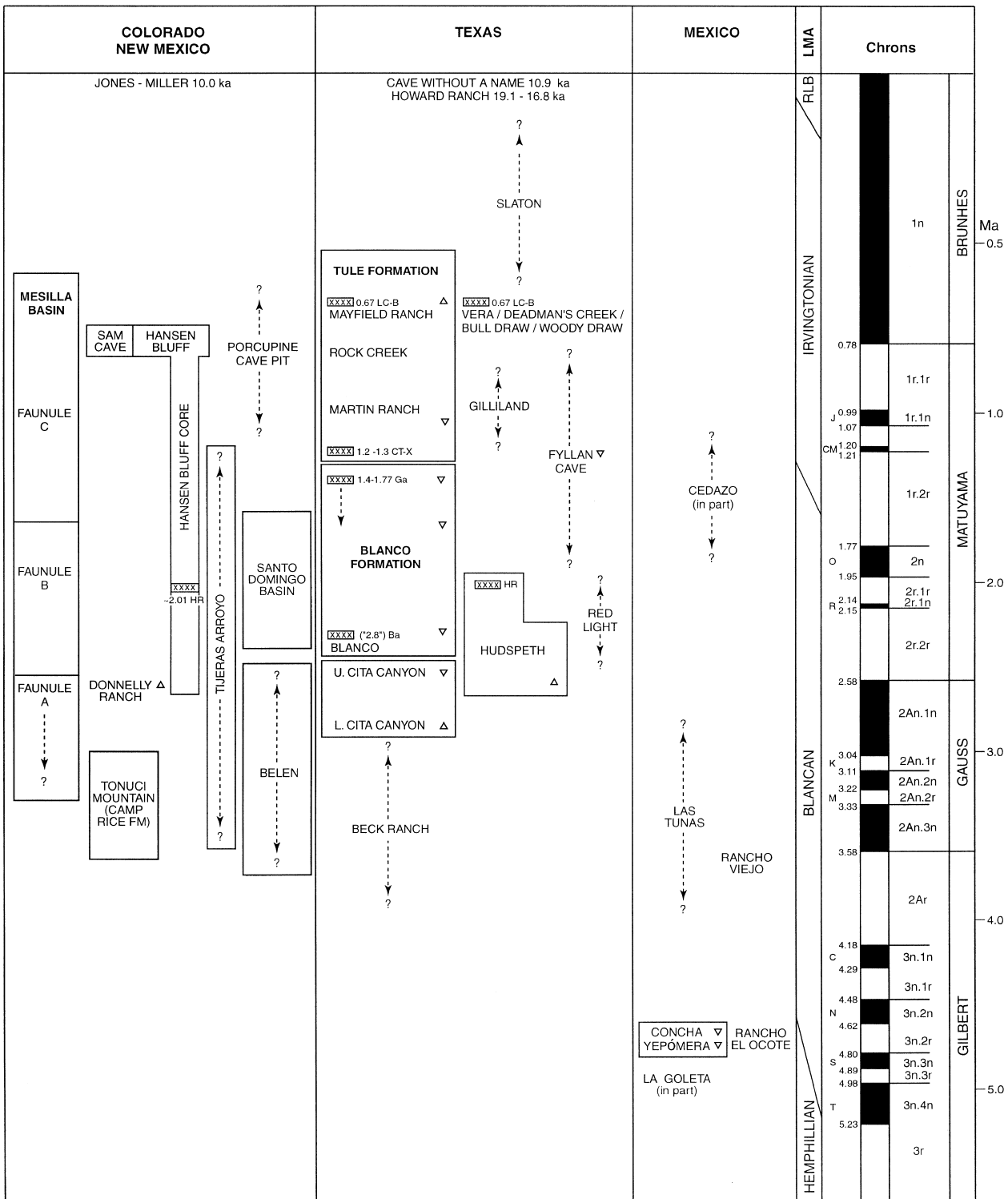


FIGURE 7.1 (continued)

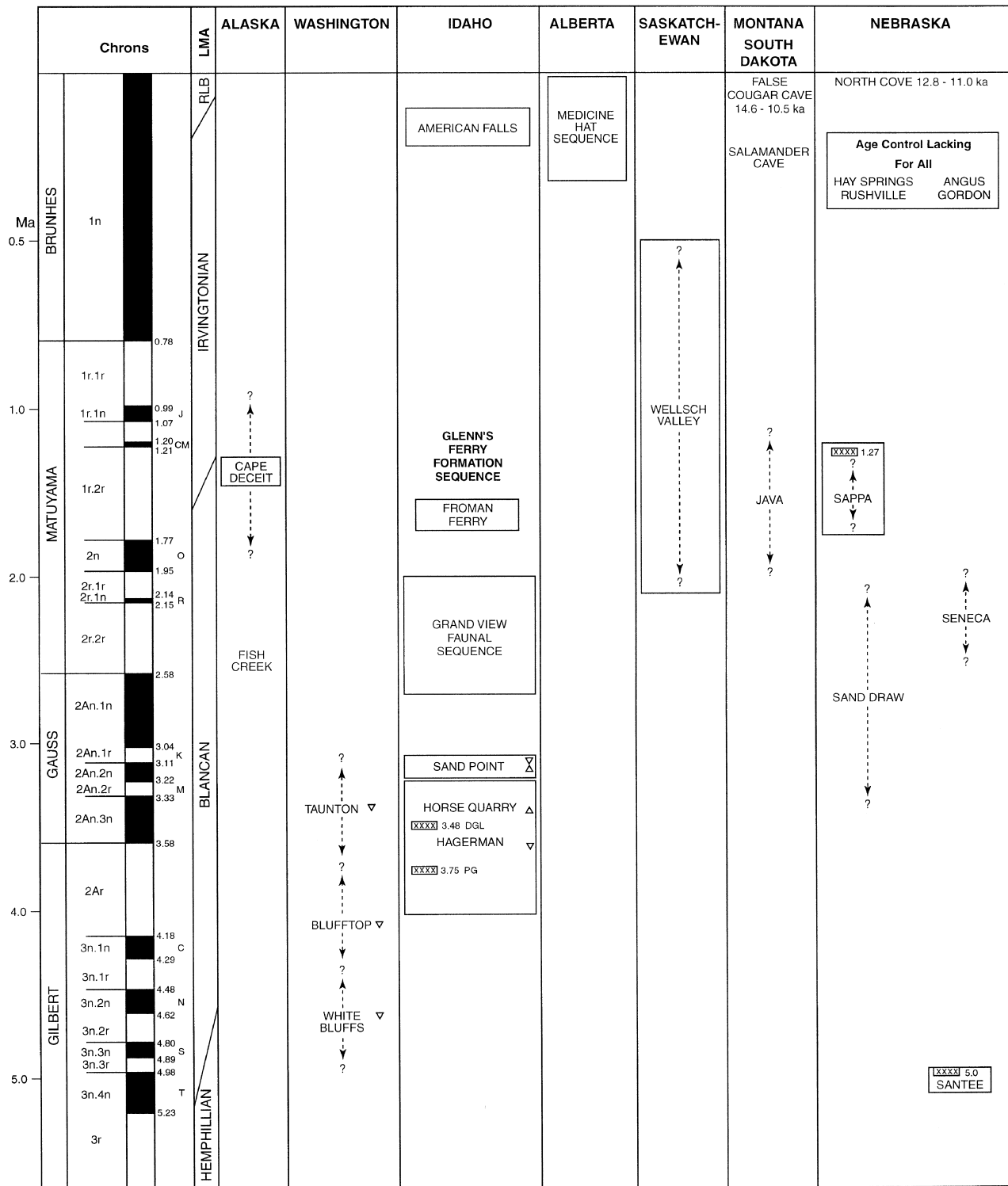


FIGURE 7.1 (continued)

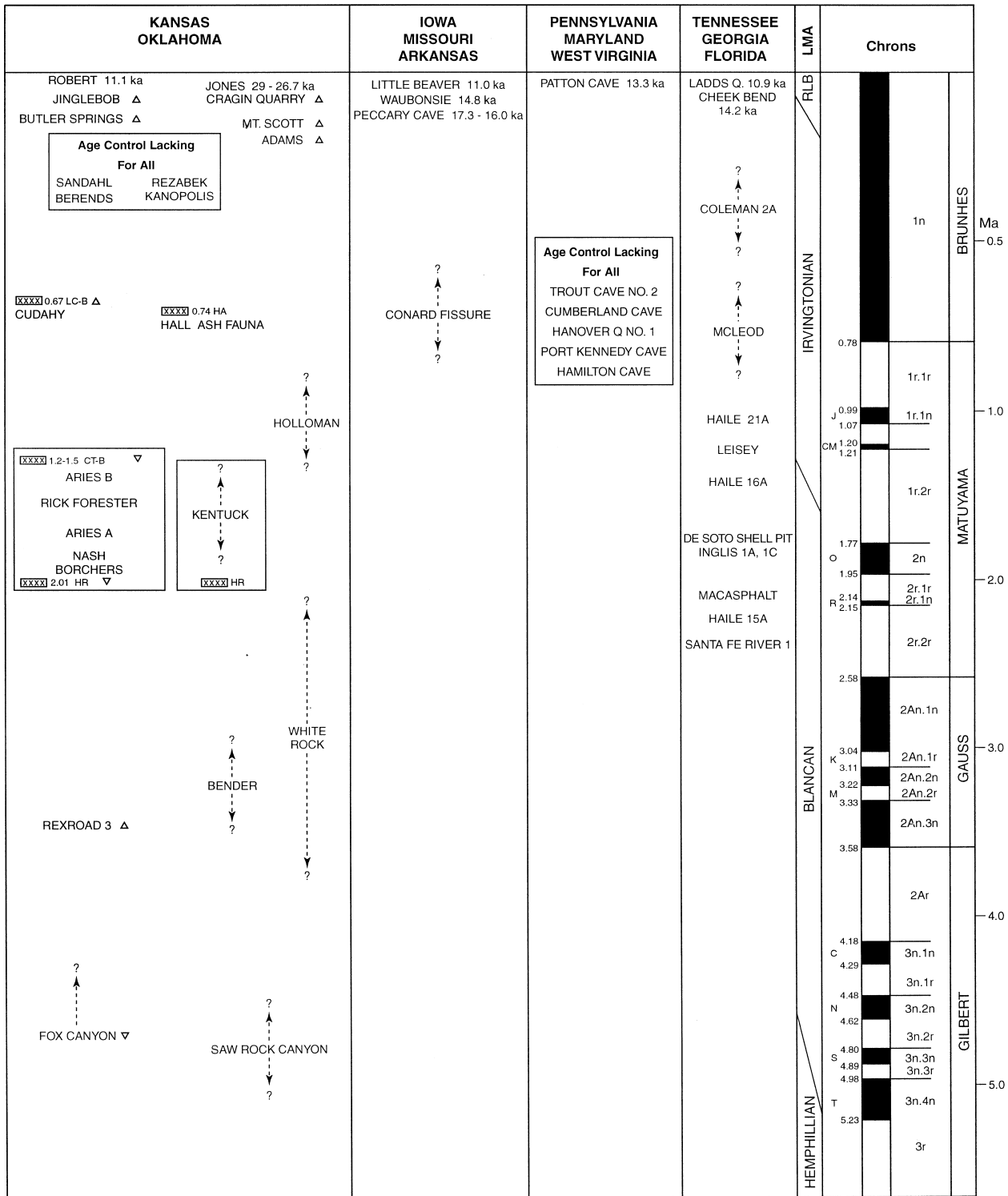
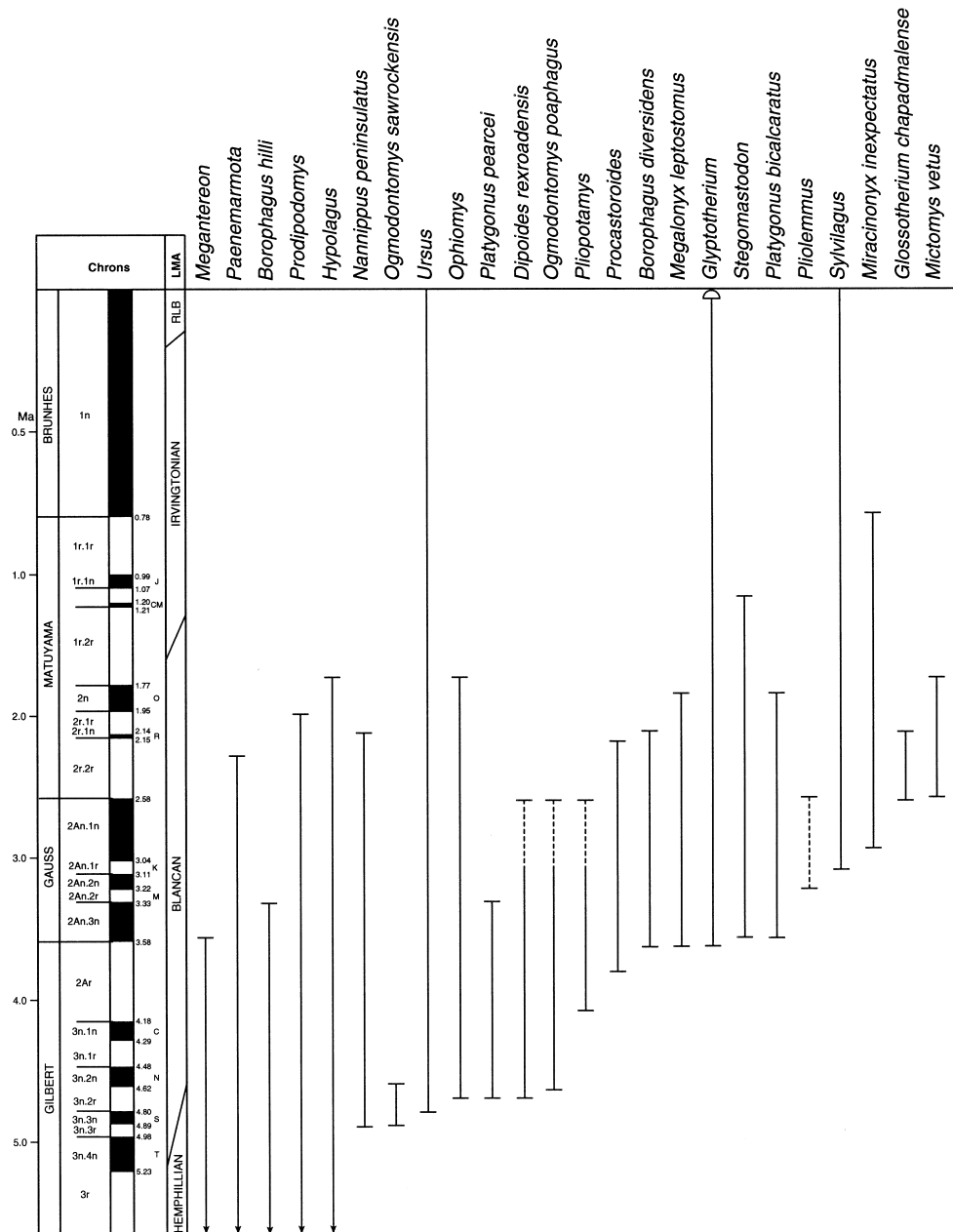
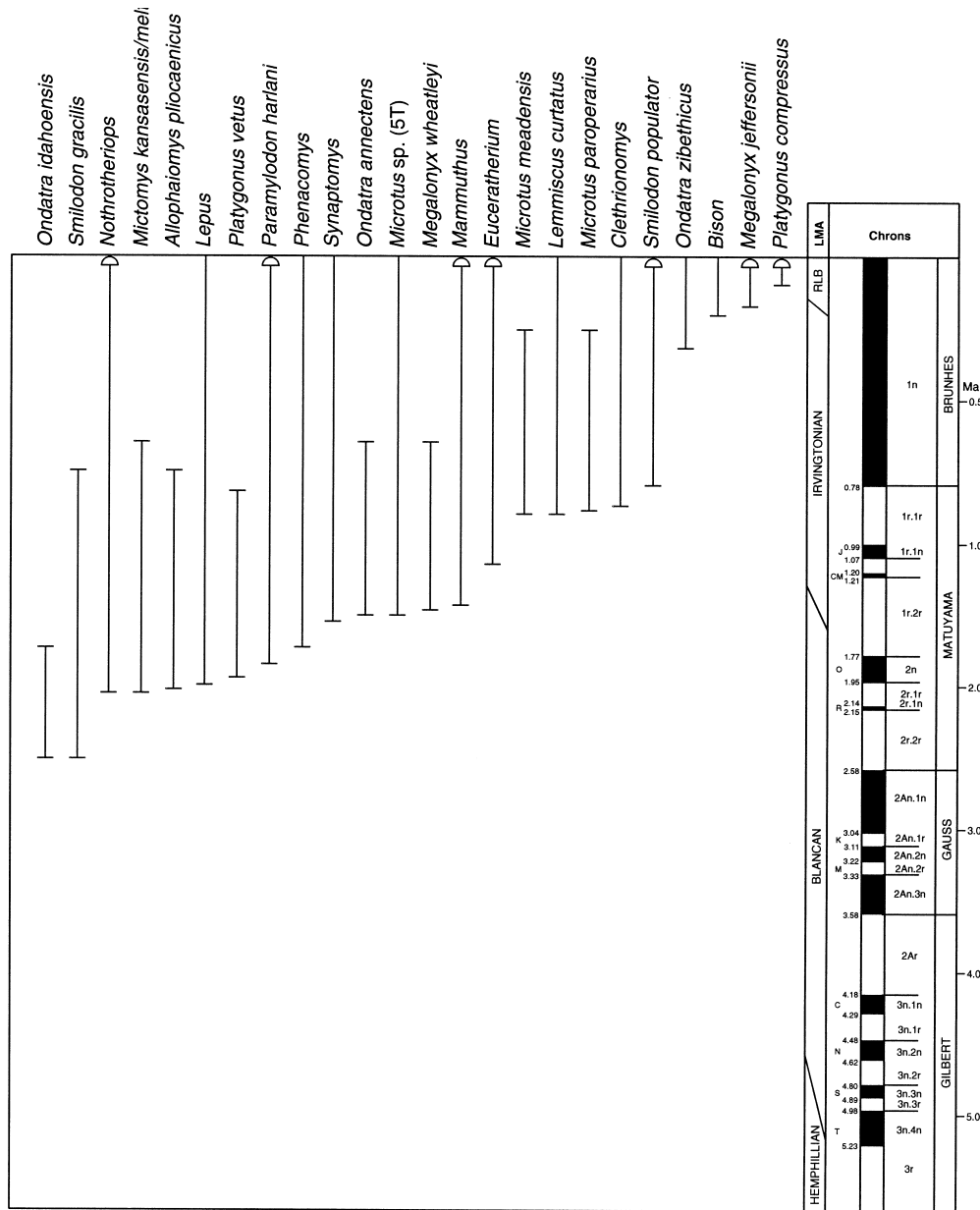


FIGURE 7.1 (continued)



**FIGURE 7.2** Temporal ranges of selected North American Pliocene and Pleistocene mammals. Many ranges are based on faunas with external age control; possible older or younger occurrences in faunas that are poorly constrained temporally are discussed in the text. Dashed lines indicate a high degree of uncertainty. E, earliest known occurrence; Hemp, Hemphillian; L, latest known occurrence; LMA, land mammal age; RLB, taxon became extinct near the end of the Rancholabrean, between approximately 24 ka and 9.5 ka; 5T, 5 closed triangles on lower first molar. *Megantereon*, E Hemp, L Rexroad 3; *Paenemarmota*, E Hemp, L Blanco; *Borophagus hilli*, E Hemp, L Hagerman; *Prodipodomys*, E Hemp, L Borchers; *Hypolagus*, E Hemp, L Froman Ferry; *Nannippus peninsulatus*, E La Goleta, L Macaspalt; *Ogmodontomys sawrockensis*, from three approximately contemporaneous localities (Upper Alturas, Maxum, and Saw Rock Canyon); *Ursus*, E above ash in Pine Nut Mountains, L Recent; *Ophiomys*, E White Bluffs, L Froman Ferry; *Platygonus pearcei*, E White Bluffs, L Hagerman; *Dipoides rexroadensis*, E White Bluffs, L Sand Draw; *Ogmodontomys poaphagus*, E Verde, L Sand Draw; *Pliopotamys*, E Kettleman Hills, L Sand Draw; *Procastoroides*, E Hagerman, L Grand View (Jack Ass Butte); *Borophagus diversidens*, E Rancho Viejo, L Vallecito Creek–Fish Creek sequence (at approximately 2.1 Ma); *Megalonyx leptostomus*, E Hagerman, L De Soto Shell Pit; *Glyptotherium*, E Rancho Viejo, L RLB; *Stegomastodon*, E Rexroad 3, L Martin Ranch; *Platygonus bicalcaratus*, E Rexroad 3, L Inglis 1C; *Pliolemmus*, E Bender, L Sanders or Cita Canyon (all known records lack external age control); *Sylvilagus*, E Vallecito Creek–Fish Creek sequence (ca. 3.15 Ma), L Recent; *Miracinonyx inexpectatus*, E Cita Canyon, L Conard Fissure, Port Kennedy Cave, or Hamilton Cave; *Glossotherium chapadmalense*, E 111 Ranch, L Macaspalt; *Mictomys vetus*, E 111 Ranch, L Froman Ferry; *Ondatra idahoensis*, E Seneca or White Rock, L Froman Ferry; *Smilodon gracilis*, E Santa Fe River 1A, L McLeod or Port Kennedy Cave; *Nothrotheriops*, E Vallecito Creek–Fish Creek sequence, L RLB; *Mictomys kansasensis/meltoni*, E Nash, L Cudahy; *Allophaiomys pliacaenicus*, E Nash or Hansen Bluff Core, L Porcupine





**FIGURE 7.2** (continued) Cave Pit; *Lepus*, E Borchers, L Recent; *Platygonus vetus*, E Inglis 1A, L Cumberland Cave; *Paramylodon harlani*, E Inglis 1A, L RLB; *Phenacomys*, E Froman Ferry, L Recent; *Synaptomys*, E Haile 16A, L Recent; *Ondatra annectens*, E Kentuck or Java, L Cudahy; *Microtus* sp. (5T), E Vallecito Creek–Fish Creek sequence, L Recent; *Megalonyx wheatleyi*, E Haile 16A, L Cudahy; *Mammuthus*, E Bruneau Idaho, L RLB; *Euceratherium*, E Vallecito Creek–Fish Creek sequence, L RLB; *Microtus meadensis*, E Hansen Bluff or Porcupine Cave Pit, L Salamander Cave; *Lemmiscus curtatus*, E Sam Cave or base of Porcupine Cave Pit, L Recent; *Microtus paroperarius*, E Hamilton Cave, L Salamander Cave; *Clethrionomys*, E Sam Cave, L Recent; *Smilodon populator*, E Conard Fissure, L RLB; *Ondatra zibethicus*, E Kanopolis or Rezabek, L Recent; *Bison*, E American Falls, L Recent; *Megalonyx jeffersonii*, E multiple Rancholabrean sites (questionable record from Holloman not plotted here), L RLB; *Platygonus compressus*, E Papago Springs Cave, L RLB.

*arvicoline* persist in the mammalogical and paleontological literature, in some cases reflecting an author’s formal taxonomic preference or conceptualization of higher-level systematics, in other cases representing only a convenient vernacular. In this chapter we follow Musser and Carleton (1993) in the higher-level taxonomy and use *arvicoline* for informal designations.

Variable generic classifications of the Arvicolinae introduce additional nomenclatural complications for a review such as ours. Many nomenclatural controversies result from the fact that many paleontologists want their taxonomic allocations to reflect their conceptualization of the systematic relationships of the various taxa, yet there is no agreement on the systematic relationships within the

TABLE 7.3 Taxonomic Synonymies for Arvicoline Rodents

THIS CHAPTER	ALTERNATIVE NAMES
<i>Allophaiomys pliocaenicus</i>	<i>Microtus pliocaenicus</i>
<i>Cosomys primus</i>	<i>Mimomys (Cosomys) primus</i>
<i>Lasiopodomys deceitensis</i>	<i>Microtus deceitensis</i>
<i>Lemmiscus curtatus</i>	<i>Lagurus curtatus</i>
<i>Loupomys monahani</i>	<i>Mimomys monahani</i>
<i>Microtus aratai</i>	<i>Pitymys aratai</i>
<i>Microtus guildayi</i>	<i>Allophaiomys, Pedomys, or Pitymys guildayi</i>
<i>Microtus llanensis</i>	<i>Pedomys llanensis</i>
<i>Microtus meadensis</i>	<i>Pitymys meadensis, Terricola meadensis</i>
<i>Mictomys kansasensis</i>	<i>Synaptomys kansasensis</i>
<i>Mictomys meltoni</i>	<i>Synaptomys meltoni</i>
<i>Mimomys virginianus</i>	<i>Mimomys (Cromeromys) virginianus</i>
<i>Mimomys dakotaensis</i>	<i>Mimomys (Cromeromys) dakotaensis</i>
<i>Ogmodontomys poaphagus</i>	<i>Mimomys (Ogmodontomys) poaphagus</i>
<i>Ogmodontomys sawrockensis</i>	<i>Mimomys (Cosomys) sawrockensis</i>
<i>Ondatra annectens</i>	<i>Ondatra zibethicus/annectens</i> (chronomorph)
<i>Ondatra idahoensis</i>	<i>Ondatra zibethicus/idahoensis</i> (chronomorph)
<i>Ondatra zibethicus</i> (in part)	<i>Ondatra nebracensis, Ondatra nebrascensis, Ondatra zibethicus/zibethicus</i> (chronomorph)
<i>Ophiomys</i>	<i>Mimomys (Ophiomys)</i>
<i>Pliopotamys meadensis</i>	<i>Ondatra zibethicus/meadensis</i> (chronomorph)
<i>Propliophenacomys parkeri</i>	<i>Pliophenacomys parkeri, ?Cseria parkeri</i>

group as a whole or of species within certain groups. Perhaps the most controversial generic classifications concern species variously placed in the genera *Cosomys*, *Mimomys*, *Ogmodontomys*, and *Ophiomys* (representing predominantly Blancan radiations) and those placed in *Allophaiomys*, *Arvicola*, *Lasiopodomys*, *Microtus*, *Pedomys*, *Phaiomys*, *Pitymys*, and *Terricola* (representing predominantly Irvingtonian and Rancholabrean radiations). A recent molecular analysis of the extant members of the latter group resulted in the recognition of a monophyletic clade of endemic North American species that includes the species placed in the “pitymyine” genera *Pedomys* and *Pitymys* and those placed in the genus *Microtus* (Conroy and Cook 2000). “Pitymyines” were found to be paraphyletic (supporting previous statements to that effect by Moore and Janecek 1990 and Repenning 1992). In light of these multiple studies indicating an independent derivation of “pitymyine” species, we place these species in the genus *Microtus*. To provide a guide to the literature, we list common alternative nomenclature in table 7.3 and indicate alternative generic assignments parenthetically in the text where confusion is most likely to occur.

A similar problem, perhaps even more intractable, surrounds the nomenclature and relationships of Pliocene and Pleistocene horses. Recent taxonomic and systematic treat-

ments (Winans 1989; Azzaroli and Voorhies 1993; Downs and Miller 1994; Azzaroli 1995; Repenning et al. 1995) differ significantly in conclusions and recommendations. We made no effort to resolve these issues, particularly those surrounding the propriety of using *Plesippus* or *Equus* for the nominal species *francescana*, *idahoensis*, and *simplicidens* (see also Albright 1999). For these latter taxa, we retain the most common (or most recent) usage in our discussion of faunas but indicate the taxonomic uncertainty with the use of quotation marks around the name.

The fossil equids assigned to *Nannippus phlegon* are more properly referred to *Nannippus peninsulatus*. *Equus phlegon* was transferred to *Nannippus* by Hibbard (1937) and Stirton (1940). The type specimen of *Hippotherium peninsulatum* was subsequently determined to be indistinguishable from *N. phlegon* (MacFadden 1984). *Nannippus peninsulatus* is used throughout our text for this species.

The details of the taxonomic problems with North American mylodont sloths were reviewed by McDonald (1995), who recommended using *Paramylodon harlani* for the Irvingtonian and Rancholabrean form and *Glossotherium chapadmalense* for Blancan forms. Because the recognized temporal ranges of land mammal ages are dynamic, such a division would necessitate taxonomic changes on the basis of shifting temporal boundaries. In

light of the presumed (but not yet fully demonstrated) ancestor–descendant relationship of the two forms (McDonald 1995), it may be preferable to synonymize the taxa or delimit the lineage based on morphologic rather than chronologic criteria. As used by McDonald, *P. harlani* and *G. chapadmalense* differ greatly in size. We use the size criterion here in order to preserve the effective taxonomy of McDonald, but because the mammal age boundaries proposed here differ from those used by McDonald, *P. harlani* is known not only from the Irvingtonian and Rancholabrean but also from the latest Blancan.

### TEMPORAL DIVISIONS OF THE MAMMAL AGES

Temporal divisions of the Blancan and Irvingtonian mammal ages were first proposed in the 1970s and can be categorized generally based on two operational approaches. Figure 7.3 summarizes the different approaches and their current relationships to one another. The first approach used broad-scale changes in mammalian faunas as the basis for temporal division. This was the foundation for the recognition (Kurtén 1971; C. Schultz et al. 1977, 1978) of two major divisions of the Blancan (Rexroadian and Senecan) and two of the Irvingtonian (Sappan and Sheridanian).

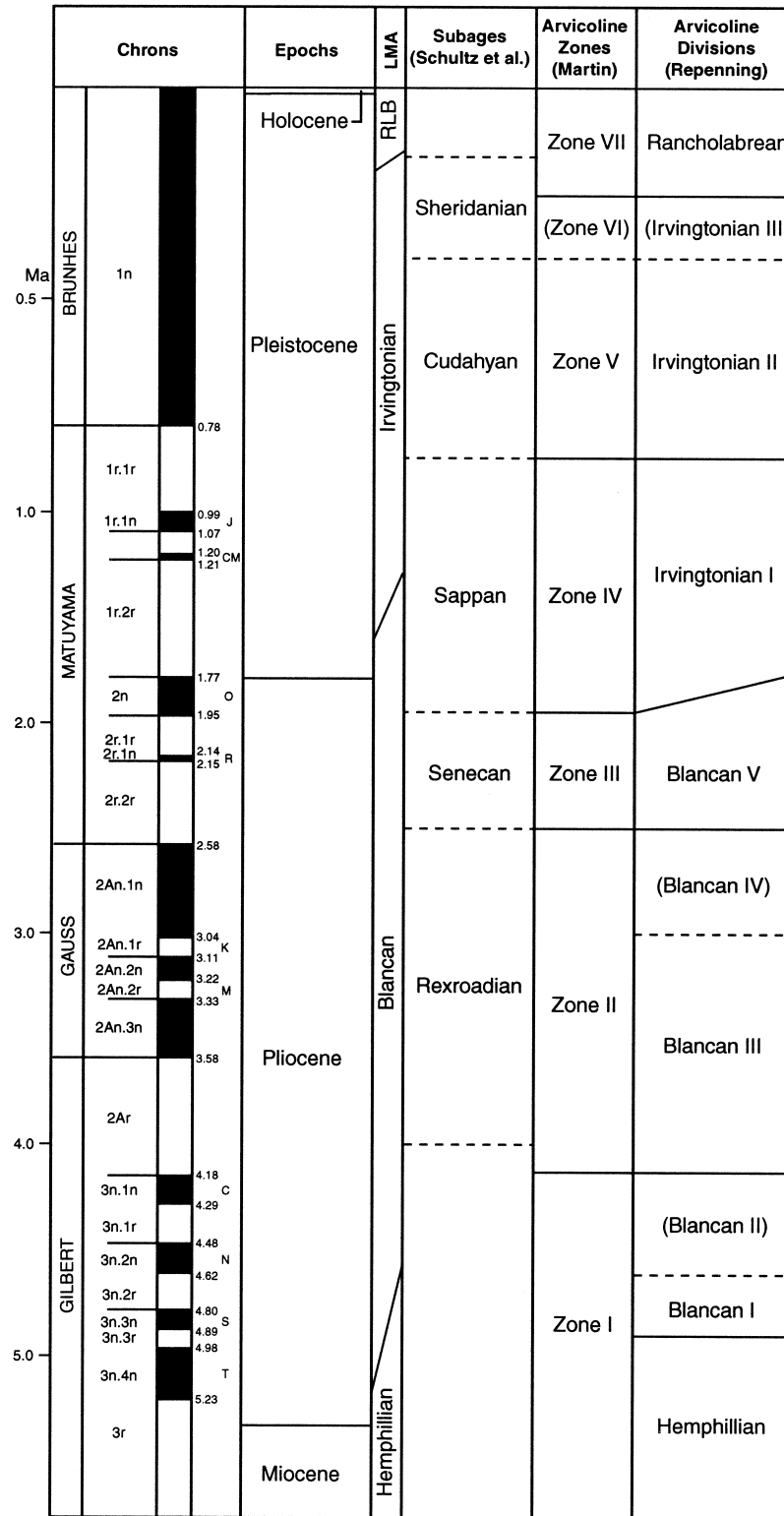
The second approach was based on evolutionary changes and dispersal history within a single lineage of mammals, the arvicoline rodents (voles, lemmings, muskrats). Following the initial explorations of the biostratigraphic utility of arvicoline rodents for faunal correlation (Hibbard 1944, 1949a, 1959; Hibbard and Zakrzewski 1967), they were soon recognized to be among the most useful biostratigraphic tools in many terrestrial settings. They are abundant, reproduce at a phenomenal rate, and appear to evolve rapidly. Several lineages demonstrate a number of significant evolutionary trends in the dentition, including the increase in hypsodonty and height of dentine tracts, the reduction, fusion, or loss of roots, the addition of cement in the reentrant angles, and an increase in crown complexity and length of the lower first molar (Hinton 1926; L. Martin 1979; Repenning 1987; Fejfar and Repenning 1992). These rodents are not well represented in deposits in the southern part of North America, thus limiting their applicability to biochronologies in lower latitudes. Longitudinal effects have been proposed also, but some may be a result of inadequate sampling and all need further critical evaluation.

Two alternative frameworks for dividing the Blancan and Irvingtonian based extensively on arvicoline rodents

were proposed almost simultaneously (Repenning 1978; L. Martin 1979). An outline of the structure for each is provided here, but the details are discussed under the relevant mammal ages.

L. Martin (1979) established a series of arvicoline rodent zones for the late Cenozoic faunas of the United States based primarily on faunas from the Great Plains. He proposed seven zones for the late Cenozoic based on first and last appearances of taxa and stages of progressive evolution within lineages. Zones were numbered consecutively from oldest to youngest. Zone I was considered to be Hemphillian, zones II and III were within the Blancan, zones IV through VI spanned the Irvingtonian, and zone VII represented an undifferentiated Rancholabrean and Holocene. Early attempts to include faunas from outside the Great Plains in these zones and preliminary attempts to correlate the proposed zones with the Eurasian sequence were explored by L. Martin (1979), but the zones were and are used almost exclusively in the Great Plains. The proposed zones and their boundaries have not been critically reevaluated in recent years, but the taxa used to define boundaries in Martin's proposal were nearly identical to those used in Repenning's model; therefore changes over the last two decades in our understanding of the evolutionary history and temporal range of these taxa apply equally to both models (figure 7.3).

A more elaborate arvicoline biochronology was outlined by Repenning and Fejfar (1977) and subsequently expanded in an extended abstract by Repenning (1978). Continued development and modification in subsequent years (Repenning 1980, 1984, 1987, 1992; Repenning et al. 1990, 1995; Fejfar and Repenning 1992; Bell and Repenning 1999; Bell 2000) resulted in a current biochronology that differs in many key aspects from that originally proposed. From its inception, this scheme relied extensively on external age control from radioisotopic dating, paleomagnetic correlations, and climatic correlations. Repenning proposed five divisions of the Blancan, Blancan I through Blancan V, numbered consecutively from oldest to youngest. The early papers (through 1987) outlining this scheme recognized two temporal divisions of the Irvingtonian and two of the Rancholabrean, again numbered I and II from oldest to youngest within each mammal age. Rancholabrean I included faunas that were acknowledged to be pre-*Bison* in age. Following the recommendation by Lundelius et al. (1987) that *Bison* be used as the defining taxon for the Rancholabrean, Rancholabrean I was abandoned by Repenning et al. (1990); faunas previously placed in Rancholabrean I were accommodated by a new division, Irvingtonian III, thus leaving an undifferentiated Rancholabrean.



**FIGURE 7.3** Summary chart showing temporal boundaries of the late Hemphillian, Blancan, Irvingtonian, and Rancholabrean, relative to defined epochal boundaries and the geomagnetic polarity time scale of Berggren et al. (1995). The subages proposed by C. Schultz et al. (1978), the arvicoline rodent zones of L. Martin (1979), and the arvicoline divisions proposed by Repenning (1987; Repenning et al. 1990) are shown. Dashed lines represent uncertain boundary placements or undefined temporal divisions. Temporal divisions in parentheses are not recognized in this chapter (see text).

## GEOGRAPHIC RESTRICTION

There are problems with defining mammal ages that are intended to be used over entire continents and to have isochronous boundaries. Dispersal is by nature a time-transgressive phenomenon, and when the temporal duration of dispersal of defining taxa can be resolved, mammal age boundaries may become time-transgressive (see also Walsh 1998). The diachronous boundaries that result may, in some cases, be a more accurate reflection of our understanding of faunal dynamics (see discussion in Fejfar and Repenning 1992; Repenning 1992; Repenning et al. 1995), but they also present additional complications that remain largely unexplored. The issues and questions surrounding mammalian faunal provinciality in the Pliocene and Pleistocene are discussed in greater length in this chapter, but one persistent problem we encountered in our efforts to prepare this chapter must be clarified at the outset.

Sufficient evidence is available to indicate that the higher latitudes in North America warrant an independent biochronology for the Pliocene and Pleistocene. In recognition of that fact, and in an effort to stimulate development of such a chronology, we formally recommend that use of the terms *Blancan*, *Irvingtonian*, and *Rancholabrean* be restricted geographically to the portions of the continent south of 55°N latitude. This boundary is arbitrary and must be evaluated critically, but it is drawn along a “data-free zone” where no relevant mammal faunas are known between those of southern Canada and the United States and those of Alaska and the Yukon Territory. This geographic restriction is consistent with our understanding of faunal change at higher latitudes and conforms with existing operational practice of members of this committee and many of our colleagues.

In the case of the Blancan, Irvingtonian, and Rancholabrean, we suggest that Alaska be regarded as part of the Beringian faunal region (*sensu* Fejfar and Repenning 1992). Faunas located north of 55°N latitude may be considered temporal equivalents of their counterparts to the south but clearly need independent biochronologies.

## BLANCAN

### HISTORICAL CONTEXT

The term *Blancan* was first proposed by Wood et al. (1941), based on the fauna at Mt. Blanco and the adjoining draws, near the “old rock house” north of Crawfish

Draw, Crosby County, Texas. The Blanco Fauna was at different times regarded as Pliocene (Osborn and Matthew 1909; Osborn 1910; Matthew 1924; Simpson 1933) and early Pleistocene (Meade 1945; Hibbard 1958). It is now considered to be largely Pliocene in age, but latest Blancan faunas (those dating to between 1.77 and 1.35 Ma) are Pleistocene.

### HISTORICAL CHARACTERIZATION

In the following paragraphs, we review the historical characterization of the Blancan. The definition and characterization as currently understood appear under the next heading.

In the original characterization of the Blancan, Wood et al. (1941) listed *Borophagus*, *Ceratomeryx*, *Ischyrosmilus*, and *Plesippus* (now sometimes identified as *Equus*) as “index taxa” for the Blancan (the taxa known only from the Blancan as of 1940). The following additional taxa were noted by Wood et al. (1941) to appear for the first time during the Blancan (but were not limited to it): glyptodonts, *Megalonyx*, *Paramylodon*, hystricomorphs, *Canis*, *Felis*, *Camelops*, *Tanupolama*, *Platygonus*, *Cervus*, and *Odocoileus*. In their article, the last appearance of *Anancus*, *Lutravus*, *Megatylopus*, *Nannippus*, and ?*Neohipparion* was during the Blancan.

Several of the taxa involved in the characterization provided by Wood et al. were reevaluated in subsequent years, and changes in their taxonomy and known stratigraphic range resulted in changes in the characterization of the Blancan. For example, the small Blancan mylodont sloth (“*Paramylodon*” of Wood et al. 1941) is now recognized as *Glossotherium* (McDonald 1995), *Tanupolama* is now recognized as *Hemiauchenia* (Webb 1974b), and *Megalonyx* (Hirschfeld and Webb 1968) and *Platygonus* (Voorhies 1990) are now known from the Hemphillian.

Further discussion of the historical treatment of the Blancan was provided by Lundelius et al. (1987). In that report, the Blancan was not formally defined (*sensu* Woodburne 1977) but was characterized by the presence of *Nannippus peninsulatus*, “*Equus (Dolichohippus) simplicidens*,” *Stegomastodon*, *Borophagus diversidens*, *Trigonic-tis*, *Nekrolagus*, *Procastoroides*, cotton rats in the *Sigmodon medius*–*Sigmodon minor* lineage (all here considered *S. minor*), and arvicolines with rooted teeth (e.g., *Ophiomys* and *Nebraskomys*). The Blancan was distinguished from the Hemphillian by the absence of *Machairodus*, *Agriotherium*, *Plesiogulo*, *Osteoborus*, *Osbornoceras*, *Prosthenmops*, *Pliohippus*, *Astrohippus*, *Dinohippus*, *Neohipparion*, and the rhinoceroses *Teleoceras* and *Aphelops*. After that publication *Dinohippus* sp. was reported from low in

the Blancan portion of the Palm Springs Formation of southern California (Downs and Miller 1994), a rhinoceros was reported by Madden and Dalquest (1990) from the Blancan Beck Ranch Fauna of Texas, and *Teleoceras* was reported from the Pipe Creek Sinkhole in Indiana (Farlow et al. 2001), a fauna that also includes *Ogmodontomys* (and is therefore Blancan by our definition). *Teleoceras* was also listed without comment from the Saw Rock Canyon Fauna in Kansas by Prothero and Manning (1987), Prothero et al. (1989), and Prothero (1998); we are not aware of any published description or other records of that occurrence, but the Saw Rock Canyon Fauna contains *Ogmodontomys* and is therefore Blancan.

### CURRENT DEFINITION AND CHARACTERIZATION

The Blancan is currently defined by the first appearance in North America south of 55°N latitude of arvicoline rodents in the genera *Mimomys*, *Ogmodontomys*, and *Ophiomys*. *Ogmodontomys sawrockensis* and *Ophiomys mcknighti* were used by Repenning (1987) and Repenning et al. (1990) to define the Blancan (reported under the names *Mimomys (Cosomys) sawrockensis* and *Mimomys (Ophiomys) mcknighti*). The earliest appearance of these two species was reported to be at approximately 4.8 Ma in the Upper Alturas Fauna in California (Repenning 1987). An alternative species, *Mimomys panacaensis*, was proposed by Lindsay et al. (2002). The enamel microstructure of this species is similar to that of Eurasian *Mimomys* and differs from the North American lineages *Cosomys*, *Ogmodontomys*, and *Ophiomys* (Mou 1998). This species was recovered from the Panaca Fauna in Nevada, where its earliest appearance is at approximately 4.98 Ma.

The characterization of the Blancan includes taxa that are limited to the Blancan, those that appear in earlier mammal ages but are common in Blancan faunas, and those that make their first appearance in the Blancan but persist into younger mammal ages. These are listed separately here, followed by comments justifying our treatment of some of the taxa and relevant faunas.

Taxa limited to the Blancan include *Megalonyx leptostomus*, *Glossotherium chapadmalense*, *Sylvilagus webbi*, *Aluralagus*, *Pewelagus dawsonae*, *Pratilepus kansasensis*, *Dipoides rexroadensis*, *Procastoroides*, *Guildayomys*, *Hibbardomys*, *Nebraskomys*, *Pliophenacomys*, *Sigmodon curtisi*, *Sigmodon minor*, *Mictomys vetus*, *Ogmodontomys*, *Ophiomys*, *Pliopotamys*, *Ondatra idahoensis*, *Pliolemmus*, *Borophagus diversidens*, *Canis lepophagus*, *Ursus abstrusus*, *Platygonus bicalcaratus*, *Platygonus pearcei*, and *Nannipus peninsulatus*.

Taxa that make their first appearance in earlier mammal ages but persist into the Blancan include *Alilepus*, *Hypolagus*, *Nekrolagus*, *Paenemarmota*, *Prodipodomys*, *Satherium*, *Borophagus hilli*, *Trigonictis*, *Megantereon*, and *Rhynchotherium*.

Taxa that first appear in the Blancan but persist into the Irvingtonian or younger include *Glyptotherium*, *Blarina*, *Sylvilagus*, *Miracinonyx inexpectatus*, *Smilodon gracilis*, and *Stegomastodon*. Taxa that first appear in latest Blancan faunas (those that are elsewhere often considered to be early Irvingtonian) include *Nothrotheriops*, *Paramylodon harlani*, *Holmesina*, *Lepus*, *Allophaiomys pliocaenicus*, *Microtus* with five closed triangles on the lower first molar, *Mictomys kansasensis/meltoni*, *Neofiber*, *Ondatra annectens*, *Phenacomys*, *Synaptomys*, *Sciurus*, *Canis edwardii*, *Chasmaporthetes*, *Lutra canadensis*, *Homotherium*, *Platygonus vetus*, and *Mammuth americanum*.

A Hemphillian record of *Nekrolagus* from Florida was mentioned by White (1991b), but the list of referred specimens in that article does not include any Hemphillian fossils or any specimen from Florida. The single specimen catalogued as *Nekrolagus* in the Florida Museum of Natural History is referable to *Alilepus* based on dental morphology (D. R. Ruez, pers. obs., 2001).

*Stegomastodon* survived into the Irvingtonian, and it is found with *Mammuthus* in several faunas, but the few localities with external age control suggest that the time of overlap was brief. Co-occurrences were reported from the Gilliland Fauna of Texas (Hibbard and Dalquest 1966); the Martin Ranch Fauna at the base of the Tule Formation in Briscoe County, Texas (G. Schultz 1990b; Dalquest and Schultz 1992), at the same stratigraphic level as a volcanic ash dated at 1.2–1.3 Ma (Izett 1977, 1981; Izett et al. 1981); possibly Holloman, Oklahoma (Hay and Cook 1930; Meade 1953; but see Madden 1983); Tortugas Mountain, New Mexico (Lucas et al. 1998); Fullerton, Nebraska (Madden 1986); near Taylor, Arizona (Madden 1986); and Faunule C in Mesilla Basin, New Mexico (Vanderhill 1986).

Our restriction of *Pliophenacomys* to the Blancan is based in part on the transfer of *Pliophenacomys parkeri* to *Propliophenacomys* by L. Martin (1994); *Pliophenacomys osborni* was reported from the Wellsch Valley faunas (Stalker and Churcher 1982; Repenning 1987) and was shown schematically to be associated with *Borophagus diversidens*, *Mammuthus*, and *Microtus paroperarius* on the Stalker and Churcher wall chart. Until the Wellsch Valley faunas are published in their entirety with adequate stratigraphic data, we discount these occurrences in our evaluation of temporal ranges of taxa.

We are aware of only one Irvingtonian occurrence of *Chasmaporthetes*. It is found with *Mammuthus* in the El Golfo Fauna in Mexico (Shaw 1981; Lindsay 1984).

### HEMPHILLIAN-BLANCAN BOUNDARY DETERMINATION

In recent years, additional work on latest Hemphillian and earliest Blancan faunas yielded new information on the age of the Hemphillian–Blancan boundary. The beginning of the Blancan now appears to fall between approximately 4.6 and 5.2 Ma. This age range is based on nine vertebrate localities with radiometric dates or paleomagnetic data, discussed individually later in this chapter. If these age estimates for the boundary are correct, then the transition from Hemphillian faunas to Blancan ones took place somewhat later than the 5.335 Ma Miocene–Pliocene boundary as defined in the Mediterranean marine section (Van Couvering et al. 2000).

The Pinole and Santee faunas are the youngest radiometrically dated Hemphillian faunas. The Pinole Fauna in California was recovered from an ash with a K–Ar date of  $5.2 \pm 0.1$  Ma (Evernden et al. 1964; Sarna-Wojcicki 1976). The Pinole Fauna contains *Machairodus*, *Plesiogulo*, *Osteoborus*, *Dinohippus*, *Sphenophalos*, and *Megalonyx* and therefore is considered to be late Hemphillian in age (Stirton 1939, 1951; Tedford et al. 1987). The Santee Fauna in Nebraska (Boellstorff 1978; Voorhies 1990) is overlain by an ash dated  $5.0 \pm 0.2$  Ma by fission-track methods on glass shards (Voorhies 1977). The Santee Fauna contains *Hesperoscalops mcgrewi*, *Dipoides* sp., mylagaulids, megalonychids, a rhinoceros, an early cervid, and *Protopliophenacomys parkeri* (=“*Propliophenacomys*” of L. Martin 1975 and “*Cseria parkeri*” of Voorhies 1990; see Korth 1994 and L. Martin 1994) and is considered to be late Hemphillian (Voorhies 1977, 1990; L. Martin 1975).

The Palmetto Fauna from the Upper Bone Valley Formation of central Florida includes *Felis rexroadensis*, *Megantereon hesperus*, *Mylohyus elmorei*, *Hexobelomeryx*, *Nannippus minor*, *Neohipparion phosphorum*, and *Calippus* sp. It is considered to be latest Hemphillian (Wright and Webb 1984). The earliest North American appearance of an antlered deer, *Eocoileus gentryorum*, was also reported from this fauna (Webb 2000). The Upper Bone Valley Formation intertongues with the marine Tamiami Formation. The phosphatic sands and gravels of the Upper Bone Valley Formation represent a high-energy backfilling episode, correlated on the basis of the marine fauna of the Tamiami Formation with an immediately post-Messinian (Zanclean) sea level rise in the early

Pliocene (Webb et al. 1978). This interpretation makes the Upper Bone Valley Formation correlative with chron C3n at approximately 5.0 Ma.

The oldest stratigraphic horizon containing the arvicoline rodent *Ogmodontomys sawrockensis* in the Upper Alturas Fauna of California is 21 m beneath a basalt flow dated  $4.7 \pm 0.5$  Ma (Repenning 1987). This occurrence led Repenning (1987) to place the Hemphillian–Blancan boundary at 4.8 Ma.

The San Timoteo Badlands in southern California contain a stratigraphic section that spans the Hemphillian–Blancan boundary (Albright 1999). The basal unit of the section contains *Repomys gustelyi* reported to be in chron C3An.2n, dating about 6.3 Ma. The Mt. Eden Fauna from near the top of the Mt. Eden Formation contains *Megacamelus merriami*, *Teleoceras*, *Agriotherium gregoryi*, *Dinohippus osborni*, and *Pediomeryx hemphillensis* (Webb 1983; Albright 1999), and was interpreted to be Hemphillian; it was correlated to chron C3r and assigned a tentative age of approximately 5.6 Ma (Albright 1999). The earliest taxon indicative of a Blancan age, *Thomomys*, occurs well up in the San Timoteo Formation in chron C2Ar at about 3.8 Ma (the only Hemphillian records of *Thomomys* of which we are aware are from the Santee and Devils Nest Airstrip faunas; Voorhies 1990). Other characteristic Blancan taxa occur at various positions above this. Thus, although superposed faunas are present, detailed resolution of the boundary is not possible. In this section the Hemphillian–Blancan boundary lies somewhere between 5.6 and 3.8 Ma.

Fission-track and  $^{40}\text{Ar}/^{39}\text{Ar}$  dates from sediments containing both Blancan and Hemphillian faunas in Guajuato, Jalisco, and Hidalgo, Mexico, were reported by Kowallis et al. (1998). The fission-track dates from Blancan sediments range from  $3.9 \pm 0.3$  to  $4.6 \pm 0.3$  Ma, whereas the fission-track dates associated with the Hemphillian faunas range from  $4.8 \pm 0.2$  to  $4.4 \pm 0.3$  Ma. The  $^{40}\text{Ar}/^{39}\text{Ar}$  dates for the Blancan range from  $3.36 \pm 0.04$  to  $4.74 \pm 0.14$  Ma. The one  $^{40}\text{Ar}/^{39}\text{Ar}$  date for the Hemphillian is  $4.89 \pm 0.16$  Ma. These dates place the Hemphillian–Blancan boundary in Mexico between  $4.74 \pm 0.14$  and  $4.89 \pm 0.16$  Ma.

In Chihuahua, Mexico, the late Hemphillian Yepómera Fauna is overlain by the early Blancan Concha Fauna (Lindsay et al. 1984; Lindsay and Jacobs 1985). These faunas are separated by approximately 25 m of section in the same reversed magnetozones, interpreted to be chron C3n.2r dated between 4.62 and 4.80 Ma (Berggren et al. 1995) between the Sidufjall (chron C3n.3n) and Nunivak (chron C3n.2n). No discernible breaks in the sequence were recorded (Lindsay and Jacobs 1985). The late

Hemphillian Yepómera Fauna contains *Machairodus*, *Megantereon*, *Pseudaelurus*, *Agriotherium*, *Taxidea*, canids, *Paenemarmota*, *Spermophilus*, *Notolagus*, *Prosthennops*, *Hexobelomeryx*, *Megatylopus*, *Astrohippus stocki*, *Dinohippus mexicanus*, *Nannippus*, *Neohipparion*, *Teleoceras*, and a proboscidean (Ferrusquia-Villafranca 1978; Lindsay et al. 1984; Lindsay and Jacobs 1985). The Concha Fauna is approximately 25 m higher in the section and contains the arvicoline rodent *Pliophenacomys wilsoni* and the gopher *Geomys minor*; Lindsay and Jacobs (1985) considered it to be early Blancan. The Concha Fauna also contains a horse that Lindsay and Jacobs (1985) referred to *?Equus* sp. The boundary in this section presumably falls somewhere between 4.62 and 4.80 Ma. This placement is younger than boundary determinations in other sections.

Recent work by Lindsay et al. (2002) demonstrated the presence of superposed Blancan and Hemphillian faunas in eastern Nevada. The Panaca Formation previously produced a Hemphillian fauna with *Pliohippus*, *?Teleoceras*, and *Hemiauchenia* (Stock 1921), but Lindsay et al. did not find Hemphillian fossils from the Panaca Formation. Blancan small mammals were recovered from a reversed magnetozone they assigned to chron C<sub>3n.3r</sub> (the reversed interval between the Sidufjall and Thvera). The fauna includes *Hypolagus edensis*, *Hypolagus tedfordi*, *Hypolagus* cf. *H. ringoldensis*, *Pewelagus dawsonae*, *Lepoides lepoides*, *Nekrolagus progressus*, *Pliogeomys parvus*, *Perognathus mclaughlini*, *Prodipodomys minor*, *Prodipodomys tiheni*, *Repomys panacaensis*, *Repomys* n. sp., and *Mimomys panacaensis* (Lindsay et al. 2002). The Hemphillian fauna from the Panaca beds (of Stock 1921) was attributed to a normal magnetozone exposed in the area where Stock made his collections. On this basis they place the Hemphillian–Blancan boundary at about the top of the Thvera (chron C<sub>3n.4n</sub>) dated to approximately 4.98 Ma (Berggren et al. 1995).

In western Nevada, a 280-m-thick section on the western slope of the Pine Nut Mountains, near Carson City, produced six magnetozones correlated by an included pumice zone dated isotopically at  $4.96 \pm 0.02$  Ma. *Ursus abstrusus* was recovered from a reversed magnetozone (chron C<sub>3n.3r</sub>) above the pumice zone (Kelly 1994; Lindsay et al. 2002; this is the earliest dated occurrence of *Ursus*). A rhinoceros was later recovered (Kelly 1997) 100 m below the pumice zone (and the *Ursus* specimen) in a reversed magnetozone assigned by Lindsay et al. (2002) to chron C<sub>3r</sub>. These occurrences support the placement of the Hemphillian–Blancan boundary in Nevada in chron C<sub>3n.3r</sub>, between 4.89 and 4.98 Ma (Berggren et al. 1995).

## TEMPORAL EXTENT

Current age estimates of early Blancan faunas indicate that the beginning of the Blancan dates to between 4.6 and 5.2 Ma. The upper limit of the Blancan is marked by the lower limit of the Irvingtonian, here defined by the earliest appearance of *Mammuthus* south of 55°N latitude. As currently understood, the earliest known specimens of *Mammuthus* appear at approximately 1.35 Ma. Most of the Blancan is therefore within the Pliocene Epoch (5.335–1.77 Ma), but the youngest Blancan faunas are earliest Pleistocene in age (according to the established epochal boundaries discussed earlier).

## TEMPORAL DIVISIONS OF THE BLANCAN

Several efforts have been made in the last 30 years to achieve finer temporal resolution in the Blancan. Most of them rely on first and last appearances of certain mammalian taxa in specified stratified sequences or isolated localities. These appearances reflect immigration and extinction events and progressive species evolution within particular lineages. Such efforts may be hampered by problems of provinciality; latitudinal, regional, elevational, and ecological factors lead to faunal provincialism that together with lack of radiometric and paleomagnetic control can make correlation of faunas difficult. Despite such difficulties, some progress has been made in dividing the Blancan, although to date no single method or proposal is universally adopted or considered universally applicable.

One of the earliest attempts to recognize temporal divisions in the Blancan was carried out by Claude Hibbard and his students in the Great Plains. Extensive use of screenwashing techniques (Hibbard 1949c) resulted in the recovery of tremendous numbers of fossils of small mammals, providing a new data set with which to analyze and compare fossil faunas. Through careful analysis of faunal composition, reference to stratigraphic marker beds (as then understood), and tentative correlation to the classic glacial–interglacial sequence, Hibbard and his colleagues proposed an ordered sequence of faunas (Hibbard et al. 1965; see also Zakrzewski 1975b). Although this ordered sequence implicitly recognized change in mammalian faunas through time, Hibbard never proposed formal, named temporal divisions. Nevertheless, his pioneering efforts in this regard laid a solid foundation for subsequent temporal correlations and divisions of Blancan and later times. Not surprisingly, the Great Plains sequence continues to play an important role in biochronologic studies in the late Cenozoic.



**Rexroadian and Senecan** Other early proposals for dividing the Blancan centered on the initial recognition of two major temporal divisions: an early Blancan and a late Blancan. One such proposal was based on the immigration of South American mammals after the development of the Panamanian isthmus and the great American faunal interchange. Faunas predating the interchange (at approximately 2.5 to 2.7 Ma; Marshall et al. 1979; Webb and Rancy 1996) were proposed to be early Blancan faunas; those after the interchange were late Blancan (Kurtén and Anderson 1980).

An alternative proposal was based on the diverse and well-known Rexroad faunas in Kansas. These were recognizably distinct from other faunas in the region, and Kurtén (1971) proposed recognition of a separate mammal age, the Rexroadian, that would be based on these faunas. This proposal was rejected by C. Schultz et al. (1977, 1978), but the Rexroadian was retained as a unit in their proposed twofold division of Blancan faunas in the Great Plains. The older unit, the Rexroadian of Kurtén (1971), was expanded from the original concept to include the Sand Draw, Broadwater, Lisco, Blanco, and Hagerman, as well as the Rexroad faunas. The Rexroadian was characterized by what was then thought to be the first appearance in North America of *Megantereon* (now known from the late Hemphillian of Florida; Berta and Galiano 1983), *Chasmaporthetes*, *Ursus*, *Trigonictis* (now known from the Hemphillian; Voorhies 1990), *Satherium*, *Stegomastodon*, *Mammot*, *Glyptotherium*, true cervids, and, in part, by certain arvicoline rodents lacking cement in their reentrant angles (e.g., *Nebraskomys*, *Pliopotamys*, *Ophiomys*, and *Pliolemmus*) and giant beavers that lack distinct ridges on their incisor enamel (*Procastoroides sweeti*).

The upper part of the Blancan, for which C. Schultz et al. (1977, 1978) proposed the name *Senecan*, includes the Grand View, White Rock, Dixon, and Seneca faunas and part of the Mullen assemblage described by L. Martin (1972). Senecan faunas are characterized by the first appearance of *Synaptomys* (*Pliotomys*), *Mictomys* (*Metaxymys*), *Ondatra*, *Loupomys monahani* (= *Mimomys monahani* of L. Martin 1972; see Koenigswald and Martin 1984), and *Procastoroides idahoensis*, which has ridged incisors. Based on faunas considered to be Senecan, the Rexroadian–Senecan boundary appears to fall near the Gauss–Matuyama (chron C2An–chron C2r) boundary. The Blanco Fauna, included by C. Schultz et al. (1978) in the Rexroadian, may be more properly considered Senecan. Although it lacks the taxa used to characterize the Senecan (Dalquest 1975), it lies in reversely magnetized sediments interpreted by Lindsay et al. (1975) as representing the early Matuyama chron (chron C2r.2r). The

Guaje ash bed, which overlies the Blanco Formation and fauna, is dated to between  $1.77 \pm 0.44$  Ma (Boellstorff 1976) and  $1.4 \pm 0.2$  Ma (Izett et al. 1972).

The terms *Rexroadian* and *Senecan* were based on faunas from the Great Plains sequence and the Snake River Plain. They are most commonly applied to faunas in the Great Plains, and their potential application in other regions remains largely unexplored but seems to be limited.

**Divisions Based on Arvicoline Rodents** The most successful attempts to temporally divide the Blancan were based on the taxonomic composition and stage of evolution of arvicoline rodents. Arvicoline zones II and III were the two temporal divisions of the Blancan proposed by L. Martin (1979). Zone II was marked by the first appearance of muskrats (*Pliopotamys*) and arvicoline rodents with rootless teeth (*Pliolemmus*). The following species were included in this zone: *Ogmodontomys poaphagus*, *Nebraskomys rexroadensis*, *Nebraskomys mcgrewi*, *Cosomys primus*, *Ophiomys taylori*, *Ophiomys meadensis*, *Pliophenacomys finneyi*, *Pliophenacomys primaevus*, *Pliopotamys minor*, *Pliopotamys meadensis*, and *Pliolemmus antiquus*. Martin included the Fox Canyon, Rexroad, Broadwater, and Sand Draw faunas in this zone. Zone III was marked by the first records of bog lemmings and the last records of *Mimomys*, *Ophiomys*, *Ogmodontomys*, and *Pliolemmus* in North America. Martin listed the Grand View, White Rock, Dixon, Seneca, and Mullen b faunas as falling in this zone.

An alternative arvicoline rodent biochronology was presented and refined by Charles Repenning in a series of articles between 1977 and 2001. In its most recent iterations, Repenning's scheme presents a fivefold temporal division of the Blancan. Divisions are numbered consecutively from oldest to youngest and are named Blancan I through Blancan V. Two of the divisions (Blancan II and Blancan IV) currently are not defined (*sensu* Woodburne 1977) but are characterized by perceived evolutionary changes within species groups. We provisionally accept three of Repenning's temporal divisions in this chapter (Blancan I, Blancan III, and Blancan V), but in an effort to avoid terminological confusion, we do not propose alternative names. Similarly, we do not adopt the numbered dispersal event terminology proposed by Repenning (1987), and used in his subsequent reports. The ungainly problems and nomenclatorial confusion that can result from additional (purported or real) dispersal events limit the utility of that convention (see, for example, the terminological adjustments proposed by Repenning et al. 1995:16 to accommodate a purported record of the Eurasian muskrat *Dolomys* in California, the identi-

fication of which was subsequently changed to *Pliopotamys* [Repenning 1998:49]).

The recognition of the defined temporal divisions in Repenning's chronologic framework is based on purported immigration events into North America across the Bering Land Bridge, but not all presumed immigrants have a fossil record in Beringia that predates their appearance in the United States. Multiple-taxon definitions were and are used for Repenning's arvicoline divisions. Potential intercontinental dispersal corridors and intra-continental dispersal pathways were discussed most recently by Repenning (1998). The reported temporal extent, definition, and characterization of the Blancan divisions are provided later in this chapter and are based on the most current published versions of Repenning's scheme (Repenning et al. 1990, 1995; Repenning 1998, 2001; Bell 2000). For the sake of completeness, we present Repenning's framework in its full form, but our consensus is that the Blancan II and IV should not be recognized at this time.

The temporal spans presented here for the divisions are somewhat different from those given by Repenning (1987), Repenning et al. (1990), and Bell (2000). These differences result from different conventions in the use of age calibrations of the Geomagnetic Polarity Time Scale; for the purposes of standardization in this volume we follow Berggren et al. (1995) for the dates of geomagnetic polarity boundaries.

#### BLANCAN I (APPROXIMATELY 4.9-4.62 MA)

The base of this division is defined by the immigration of *Ophiomys* and *Ogmodontomys* (both given as subgenera of *Mimomys* by Repenning 1987; Repenning et al. 1990). *Ophiomys mcknighti* is reported from several localities in the western United States, and it is found with *Ogmodontomys sawrockensis* [= *Mimomys (Cosomys) sawrockensis* of Repenning 1987 and Repenning et al. 1990] in the Upper Alturas Fauna of California, dated to approximately 4.8 Ma (21 m below a basalt flow dated at  $4.7 \pm 0.5$  Ma; Repenning 1987); *O. sawrockensis* is also found in the Maxum Fauna of California and the Saw Rock Canyon Fauna in Kansas (May 1981; Repenning 1987; Hibbard 1957), but external age control is unavailable at those localities. *Mimomys panacaensis* is another early Blancan arvicoline rodent that appears, on the basis of enamel microstructure, to be more closely related to Eurasian *Mimomys* species than to the North American lineages *Ophiomys*, *Ogmodontomys*, and *Cosomys* (Mou 1997, 1998). It is known from the Panaca Fauna in Nevada

and was assigned an age of 4.98 Ma (Lindsay et al. 2002). It provides an additional taxon by which the Blancan I can be defined.

In the earlier papers of Repenning, *Nebraskomys* also was used to define this interval. The only potential Blancan I specimen (from the Verde Fauna in Arizona) of this rare arvicoline is not referable to that taxon (it cannot be identified reliably to any genus; Czaplewski 1990; Winkler and Grady 1990; see also the discussion in Bell 2000).

*Pliophenacomys* appears in North America for the first time during the Blancan I; its earliest appearance is in the Concha Fauna in Mexico (Lindsay and Jacobs 1985). The identification of *Pliophenacomys* from the Verde Fauna in Arizona (Repenning 1987) is in error; the material in question was reidentified as *Ogmodontomys poaphagus* by Czaplewski (1990). The Verde Fauna is situated approximately 56 m above a basalt dated at 5.6 Ma and is placed at the base of the Nunivak subchron (chron C3n.2n; Bressler and Butler 1978; Czaplewski 1990), now recalibrated to about 4.62 Ma (Berggren et al. 1995). This fauna thus sits at the boundary of the Blancan I and II as recognized by Repenning et al. (1990).

#### BLANCAN II (APPROXIMATELY 4.62-4.1 MA)

The Blancan II is currently undefined. Its recognition was based on perceived evolutionary transitions in *Ophiomys* in the Pacific Northwest and in *Pliophenacomys* and *Ogmodontomys* in faunas east of the Rocky Mountains (Repenning 1987). An expanded justification for the Blancan II was provided by Repenning et al. (1990), whose support for this interval was based in part on a desire to "maintain equivalent biochronologic units" between Europe and North America (Repenning et al. 1990:386, 398). To this end, the age of the earliest record of *Germanomys* in Asia was used to draw an arbitrary boundary for the Blancan II in North America (Repenning et al. 1990:397-398), although no North American immigration event can be definitively identified. The *Germanomys* record used by Repenning et al. (1990) was from Yushe Basin in China, where its first stratigraphic appearance is just below the Nunivak normal magnetic polarity chron (chron C3n.2n). An age of 4.2 Ma was given by Repenning et al. (1990), but recalibration of the GPTS and the Yushe Basin faunas yields a new age of 4.62 Ma for the base of the Nunivak (Berggren et al. 1995; Flynn 1997). We preserve the intent of Repenning et al. (1990) by using the *Germanomys* record from Yushe Basin to denote the beginning of the Blancan II but note that the arbitrary temporal boundary would now be placed at 4.62 Ma.

The Blancan II is further characterized by a vole with a morphology on the lower first molar that is intermediate between *Ophiomys mcknighti* and *Ophiomys taylori* (known from the Blancan III) and by *Ogmodontomys poaphagus* and *Pliophenacomys finneyi*.

### BLANCAN III (APPROXIMATELY 4.1-3.0 MA)

The first appearance of the muskrat lineage defines the beginning of the Blancan III (Repenning 1987; Repenning et al. 1990). The earliest known North American muskrat is a single specimen from the Etchegoin Formation in the Kettleman Hills in California (Repenning et al. 1995). This specimen was originally identified as the Eurasian muskrat *Dolomys* (Repenning et al. 1995) but was subsequently reidentified as *Pliopotamys* (Repenning 1998:49). This specimen is stratigraphically below a volcanic tuff identified as the Lawlor Tuff, which is dated elsewhere at  $4.1 \pm 0.2$  Ma (Sarna-Wojcicki et al. 1991).

Other early records of *Pliopotamys* include *Pliopotamys minor* from near the base of the section in the Hagerman Fossil Beds National Monument in Idaho (Zakrzewski 1969), which may be as old as 4.0 Ma (G. McDonald et al. 1996). It is also known from the Taunton Fauna in Washington (Repenning 1987), which may be approximately correlative with Hagerman (Gustafson 1985) or slightly older (Repenning 1987). *Pliopotamys meadensis* is found in the Great Plains in the Deer Park (Kansas; Hibbard 1956) and Sand Draw (Nebraska; Hibbard 1972) faunas, but reliable external age control for these localities is lacking.

The Blancan III is characterized by *Ophiomys magilli*, *Ophiomys taylori*, *Ogmodontomys poaphagus*, *Cosomys primus*, *Nebraskomys*, and *Pliophenacomys primaevus*. The earliest occurrence of *Pliolemmus antiquus* probably is also from Blancan III, but external age control is lacking for most of the localities from which this species is known. Its earliest occurrence probably is in the Bender locality in Kansas, possibly as old as 3.2 Ma (Hibbard 1972:106; R. Martin et al. 2000). Its youngest occurrence is in either the Sanders or Cita Canyon fauna.

The identification of *Nebraskomys* from the Verde Fauna (Repenning 1987) is in error (Czaplewski 1990; Winkler and Grady 1990). Its earliest appearance seems to be in the Rexroad 3 locality in Kansas, where it occurs in normally magnetized sediments (Lindsay et al. 1975) that R. Martin et al. (2000) tentatively correlated with chron C2An.3n (early Gauss). That normal polarity interval is now dated to between 3.58 and 3.33 Ma (Berggren et al. 1995).

### BLANCAN IV (APPROXIMATELY 3.0-2.5 MA)

The Blancan IV is undefined. The lower temporal boundaries proposed by Repenning (1987; at  $3.2 \pm 0.2$  Ma) and Repenning et al. (1990; at 3.0 Ma) were drawn arbitrarily. The 1990 age of 3.0 Ma was chosen to achieve balance in correlation between Europe and North America and to accommodate the entire Hagerman fossil sequence within the Blancan III.

The Blancan IV is characterized by an *Ophiomys* with a lower first molar morphology intermediate between that of *O. taylori* and *O. parvus* (Repenning 1987; Repenning et al. 1990). *Ophiomys magilli* in the Great Plains apparently is replaced by *O. meadensis*. *Pliolemmus* is still found in faunas in the Great Plains, and *Ogmodontomys* and *Pliophenacomys* persist.

### BLANCAN V (APPROXIMATELY 2.5-1.9 OR 1.72 MA)

The beginning of the Blancan V is defined by the immigration from Eurasia of the bog lemmings. To the west of the Rocky Mountains the bog lemming of the Blancan V faunas is *Mictomys vetus*, which appears in several faunas in Idaho at approximately 2.5 Ma (Repenning et al. 1995). Additional and approximately contemporaneous records are from 111 Ranch in Arizona (Galusha et al. 1984; Tomida 1987) and from Cita Canyon in Texas, where a different lemming (*Pliotomys rinkerii*) is found. The age estimation for these localities is based on paleomagnetic data; all of these faunas are near the Gauss–Matuyama boundary (the boundary between chrons 2An.1n and 2r.2r), the age of which was given as 2.58 Ma by Berggren et al. (1995). An age of 2.56 Ma for the base of Blancan V was reported by Repenning et al. (1995), based in part on climatic correlations (see Repenning and Brouwers 1992). *Pliotomys* also is found at approximately 2.4 Ma in the Fish Creek Fauna in Alaska (Repenning et al. 1987).

In latest Blancan V, *Mictomys* appeared in the Great Plains (as *M. landesi*), and *Pliotomys* became extinct. Based on traditional taxonomic treatments of muskrat taxa, the extant genus *Ondatra* appeared during Blancan V. A recent proposal that all North American muskrats, including *Pliopotamys*, be synonymized with the extant *Ondatra zibethicus* and that traditional species be recognized as “chronomorphs” (R. Martin 1996) results in a somewhat cumbersome terminology. Martin’s proposal illustrates the problem of defining species boundaries in lineages with rich fossil records, but it has not been adopted elsewhere, and we retain traditional terminology here.

Blancan V is characterized by *Ondatra idahoensis*, *Ophiomys parvus*, *Pliophenacomys osborni*, *Mictomys vetus*, and *Mictomys landesi*. The end of Blancan V is marked by the beginning of Irvingtonian I. In the most recent versions of Repenning's chronology, Irvingtonian I is diachronous and is defined in different regions based on different taxa.

We accept Blancan I, III, and V, as just discussed, and consider Blancan II to be essentially a continuation (and further characterization) of Blancan I. Similarly, Blancan IV is a continuation and further characterization of Blancan III. For our purposes, the recognized divisions, as defined by Repenning, span the following intervals: Blancan I (approximately 4.9–4.1 Ma), Blancan III (approximately 4.1–2.5 Ma), and Blancan V (approximately 2.5–1.9 or 1.72 Ma).

## GEOGRAPHIC DISTRIBUTION

Blancan faunas occur primarily in the United States west of the Mississippi River, but important faunas are also found in Mexico and Florida, and temporal equivalents are known from Alaska (figure 7.4). Since the appearance of the earlier edition of this volume, we have learned a great deal about Blancan faunas in Mexico and the United States. The most complete sequences of Blancan faunas are in the Snake River Plain in Idaho, the Anza-Borrego Desert in southern California, the San Pedro Valley in southern Arizona, and in the Great Plains, especially Kansas. Where available, the dates of most of these sequences were established by a combination of radiometric and paleomagnetic methods (Johnson et al. 1975; Lindsay et al. 1975; Opdyke et al. 1977; Neville et al. 1979; Lindsay et al. 1984; Lundelius et al. 1987; Repenning 1992; Repenning et al. 1995; G. McDonald et al. 1996; Cassiliano 1999; R. Martin et al. 2000). First and last known stratigraphic appearances of many taxa were noted within the more complete sequences, and these sequences traditionally served as standards for the dating of other Blancan faunas. A summary of the key faunas from the major physiographic regions of North America and the recent literature pertaining to their correlation is presented in this section.

**Pacific Northwest** Sediments of the Ringold Formation in south-central Washington preserved a series of vertebrate faunas recovered from numerous localities that span the late Hemphillian (River Road Fauna with *Teleoceras*; Gustafson 1977, 1978) through early Blancan (White Bluffs and Blufftop faunas; Gustafson 1978, 1985; Repenning 1987; Repenning et al. 1995) and middle Blancan (Taunton Fauna; Tedford and Gustafson 1977; J. Morgan and Morgan 1995). These faunas, together with those from the

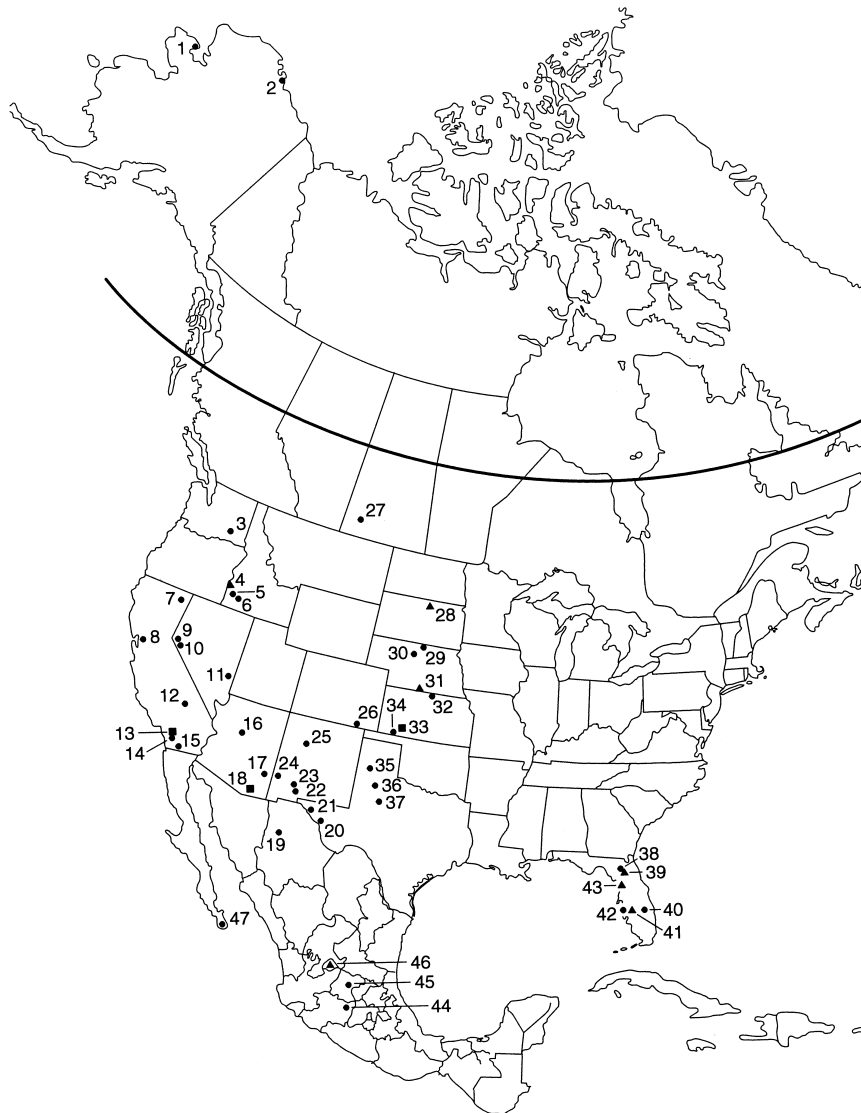
northern Snake River Plain, provide the best stratigraphic information available for the Pacific Northwest.

The White Bluffs Fauna includes *Hypolagus*, *Nekrolagus*, *Dipoides rexroadensis*, *Ophiomys mcknighti*, *Megalonyx rohrmanni*, *Borophagus hilli* (identification by Wang et al. 1999), *Trigonictis cookii*, *Platygonus pearcei*, *Megatylopus*, and *Hemiauchenia*. The age of this fauna is not well established. Paleomagnetic data for the Ringold Formation were presented by Packer (1979) and were used by Repenning to tentatively place the White Bluffs Fauna just above the Sidufjall subchron (chron C3n.3n). The fauna includes what are probably the earliest North American occurrences of *Ophiomys*, *Dipoides rexroadensis*, and *Platygonus pearcei*.

The Blufftop Fauna contains *Sorex powersi*, *Paracryptotis rex*, and *Ophiomys mcknighti-taylori* (Gustafson 1985; Repenning et al. 1995). This fauna was recovered from reversely magnetized sediments near a magnetic polarity transition that Gustafson (1985) interpreted to be the Gauss–Gilbert boundary (with an acknowledgment that it could be an older event). An older-age interpretation was favored by Repenning (1987), who placed the fauna just below the Cochiti subchron (chron C3n.1n; Repenning's Haymaker's Orchard locality is part of the Blufftop Fauna).

The Taunton Fauna includes *Megalonyx leptostomus*, *Hypolagus gidleyi*, *Hypolagus edensis*, *Hypolagus furlongi*, *Alilepus vagus*, *Procastoroides*, *Paraneotoma*, *Ophiomys taylori*, *Pliopotamys minor*, *Satherium piscinaria*, *Borophagus diversidens*, *Canis lepophagus*, *Trigonictis cookii*, *Parailurus*, *Capromeryx*, and “*Equus (Dolichohippus) simplicidens*” (Tedford and Gustafson 1977; Gustafson 1985; White 1987, 1991b; J. Morgan and Morgan 1995; Repenning et al. 1995). The Taunton Fauna was originally reported to be approximately correlative with Hagerman faunas in Idaho (Gustafson 1985). Subsequent placement in chron C2Ar (Repenning 1987) was considered too old by Repenning et al. (1995), who suggested a younger age, approximately correlative with Sand Point, which would place the Taunton Fauna in the Kaena subchron (chron C2An.1r) at approximately 3.05 Ma.

**Snake River Plain** A series of faunas along the Snake River range in age from early Blancan (e.g., Hagerman) to latest Blancan (e.g., Froman Ferry) and occur over a distance of more than 160 km (100 miles) within the Glenn's Ferry Formation. The Glenn's Ferry Formation exposed at the Hagerman Fossil Beds National Monument (Hibbard 1969; Zakrzewski 1969; Bjork 1970; G. McDonald et al. 1996) spans a stratigraphic interval of approximately 183 m. Lava flows and ash units in the section yielded K–Ar dates ranging from 3.2 to 3.57 Ma and



**FIGURE 7.4** Blancan localities and their temporal equivalents discussed in the text. The bold line represents 55°N latitude. Independent biochronologies should be established for the region north of this line, and the term *Blancan* should not be applied to faunas there. Localities with external chronologic control indicating an age between approximately 5.0 Ma and 1.35 Ma are considered to be temporal equivalents of Blancan faunas elsewhere in North America. Triangles = latest Blancan faunas (elsewhere often considered to be Irvingtonian; see text for discussion); squares = sequences or regions including Blancan and latest Blancan faunas. 1, Cape Deceit, AK; 2, Fish Creek, AK; 3, White Bluffs, Blufftop, and Taunton faunas, WA; 4, Froman Ferry Faunal sequence, ID; 5, Grand View faunas (*sensu* Repenning et al. 1995), ID; 6, Hagerman, ID; 7, Upper Alturas, CA; 8, Maxum, CA; 9, Buckeye Creek, NV; 10, Fish Springs Flat and Topaz Lake, NV; 11, Panaca and Muddy Valley, NV; 12, Coso Mountains, CA; 13, San Timoteo Badlands (in part), CA; 14, Elsinore Fault Zone (in part) and Temecula Arkose, CA; 15, Anza-Borrego Desert (in part), CA; 16, Verde, AZ; 17, 111 Ranch and Safford and Duncan basins, AZ; 18, San Pedro Valley sequence, AZ; 19, Yepómera (Hemphillian) and Concha, Chihuahua, Mexico; 20, Red Light and Hudspeth, TX; 21, Hueco Bolson, TX; 22, Mesilla Basin, NM; 23, Camp Rice Formation, NM; 24, Mangas Basin (Buckhorn fauna), NM; 25, Santa Domingo and Albuquerque Basins, NM; 26, Donnelly Ranch, CO; 27, Wellsch Valley (in part), Saskatchewan, Canada; 28, Java, SD; 29, Sand Draw, NB; 30, Seneca, NB; 31, Sappa, NB; 32, White Rock, KS; 33, Meade Basin faunas (in part), KS; 34, Saw Rock Canyon, KS; 35, Cita Canyon, TX; 36, Blanco, TX; 37, Beck Ranch, TX; 38, Santa Fe River 1, FL; 39, Haile 15A and Haile 7C, FL; 40, Kissimmee River, FL; 41, De Soto Shell Pit, FL; 42, Macaspah Shell Pit, FL; 43, Inglis 1A and Inglis 1C, FL; 44, La Goleta (in part), Michoacan, Mexico; 45, Rancho El Ocote and Rancho Viejo, Guanajuato, Mexico; 46, Cedazo (in part), Aguascalientes, Mexico; 47, Las Tunas, Baja California Sur, Mexico.

a zircon fission-track date of  $3.75 \pm 0.36$  Ma (Evernden et al. 1964; Izett 1981). The most current interpretation is that the entire sedimentary sequence at Hagerman dates to between 4.0 and 3.2 Ma (G. McDonald et al. 1996). Paleomagnetic studies by Neville et al. (1979) indicate that the Hagerman sequence corresponds to the upper part of the Gilbert reversed chron (chron C2Ar) and the lower part of the Gauss normal chron (chron C2An.3n). The Sand Point Fauna (Hibbard 1959), 48 km west of Hagerman, is preserved in reversely magnetized sediments that lie stratigraphically above normally magnetized sediments; Repenning (1987) tentatively placed it in chron C2An.2r (Mammoth subchron) but did not rule out the possibility that it could be in chron C2An.1r (the Kaena subchron). White and Morgan (1995:373) appear to have considered Sand Point to be in chron C2An.2r. The Grand View Fauna (Hibbard 1959; Shotwell 1970), from localities 120 to 160 km west of Hagerman, spans the upper part of the Gauss normal chron (chron C2An.1n) and the lower part of the Matuyama reversed chron (chron C2r.2r) (Conrad 1980). No fewer than 10 faunas were included in an expanded "Grand View Faunal Sequence" by Repenning et al. (1995:59; including Jackass Butte, Birch Creek [see also Hearst 1998, 1999], Chattin Hill, Black Butte, Castle Butte, Ninefoot Rapids, Oreana area, Poison Creek, Unnamed Butte, and Wild Horse Butte). Additional faunas from the region (e.g., Flatiron Butte, Tyson Ranch, Three Mile East) provide additional information on faunal succession and diversity through the Blancan (Conrad 1980; Repenning et al. 1995; Sankey 1996, 2002). The Jackass Butte Fauna includes the youngest known occurrence of *Procastoroides*.

The Hagerman faunal sequence includes the oldest published faunas in the Glenn's Ferry Formation. It includes *Paracryptotis gidleyi*, *Hypolagus limnetus*, *Hypolagus vetus*, *Pratilepus vagus*, *Trigonictis macrodon*, *Trigonictis cookii*, *Ursus abstrusus*, *Mustela rexroadensis*, *Satherium piscinaria*, *Pliopotamys minor*, *Ophiomys taylori*, *Cosomys primus*, *Castor californicus*, and "Equus simplicidens" ("Plesippus shoshonensis" of Repenning et al. 1995). The sequence also includes the youngest known occurrences of *Borophagus hilli* and *Platygonus pearcei* and the earliest known occurrences of *Megalonyx leptostomus*, *Procastoroides*, and *Canis lepophagus*. Chronologic control is provided by a series of radiometric dates on lava flows and ashes. An ash correlated to the Horse Quarry was dated at Ma  $3.2 \pm ?$  (Evernden et al. 1964). Lower in the sequence, the Deer Gulch lava flow yielded an age of  $3.48 \pm 0.27$  Ma (Evernden et al. 1964). The Peters Gulch Ash is the lowest dated ash in the sequence, and Izett (1981) reported a fission-track age of  $3.75 \pm 0.36$

Ma. The base of the sequence may be as old as 4.0 Ma (G. McDonald et al. 1996).

The Froman Ferry faunal sequence (Repenning et al. 1995) is the youngest faunal sequence in the Glenn's Ferry Formation. A small mammalian fauna was recovered from reversely magnetized sediments that are capped by the Pickles Butte basalt with an  $^{40}\text{Ar}/^{39}\text{Ar}$  date of  $1.58 \pm 0.085$  Ma (Repenning et al. 1995). Although an unconformity of unknown magnitude separates the basalt from the underlying fossiliferous marsh deposits, the fact that the entire marsh section shows reversed orientation indicates that the top of the Olduvai (chron C2n) is not present and that the section cannot be older than the end of the Olduvai (given as 1.65 Ma in age by Repenning et al. 1995 and as 1.77 Ma in Berggren et al. 1995). The temporal span of the section is between 1.77 and 1.58 Ma. *Phenacomys gryci* makes its first appearance in the section at approximately 744 m (2440 feet) above the base of the section and was used by Repenning et al. (1995) to define the base of the Irvingtonian I arvicoline division in Idaho. Use of the paleomagnetic calibration of Berggren et al. (1995) yields a date of 1.72 Ma for the first appearance of *Phenacomys*. *Mictomys vetus*, *Ophiomys parvus*, *Ondatra idahoensis*, *Hypolagus gidleyi*, *Gigantocamelus spatula*, "Plesippus," and *Stegomastodon mirificus*, traditionally considered to be Blancan taxa, are also in the fauna, and at least some of them, including *Stegomastodon*, are found in the upper part of the section overlapping the occurrence of *Phenacomys gryci*. The entire sequence is here considered to be Blancan. *Stegomastodon* also is found in stratigraphic association with *Mammothus* (the taxon used here to define the Irvingtonian) in several faunas in the Great Plains and the desert Southwest. The Froman Ferry faunal sequence contains the youngest dated occurrences of *Hypolagus*, *Ophiomys*, *Mictomys vetus*, and *Ondatra idahoensis*. It includes the earliest dated occurrence of *Phenacomys*.

**California** In the Imperial and Palm Springs formations at Anza-Borrego Desert State Park, California, the fossiliferous section spans 4300 m from below the base of the Cochiti subchron (chron C3n.1n) to the Jaramillo subchron (chron C1r.1n) (Cassiliano 1999). The traditional interpretation (e.g., Downs and White 1968) that the sedimentary sequence in the Vallecito–Fish Creek section contained a series of superposed faunas (the Layer Cake, Arroyo Seco, and Vallecito Creek) was recently challenged by Cassiliano (1999), who suggested that these faunal terms be abandoned. In place of the traditional terms, he proposed 10 local range zones. Approximate ages for these range zones are taken from Cassiliano, but

his use of *lowest stratigraphic datum* (LSD) and *highest stratigraphic datum* (HSD) terminology is equivalent to the more precise LSD<sub>k</sub> and HSD<sub>k</sub> of Walsh (1998; referring to the lowest and highest known stratigraphic occurrence in the specified section). The latter terminology is adopted here. The significant stratigraphic occurrences noted by Cassiliano include *Sigmodon* LSD<sub>k</sub>, from a reversed chron below the Cochiti (chron C3n.1n), at approximately 4.32 Ma; the *Equus* sp. LSD<sub>k</sub>, from a normal chron in the lower Gauss (chron C2An.3n), approximately 3.46 Ma (these specimens cannot be identified to subgenus or species); *Dinohippus* HSD<sub>k</sub>, from the top of the Kaena (chron C2An.1r), approximately 3.04 Ma (placing this taxon in the late early Blancan, the youngest occurrence in North America); *Erethizon stirtoni* LSD<sub>k</sub>, from the top of a normal chron above the Kaena, approximately 2.58 Ma (this is the earliest South American immigrant to reach the Anza-Borrego area); *Navahoceros* LSD<sub>k</sub>, early in the reversed chron (C2r.1r) below the Olduvai at approximately 2.11 Ma; *Equus* (*Equus*) LSD<sub>k</sub>, early in the reversed chron (chron C2r.1r) below the Olduvai, approximately 2.09 Ma (this may be the earliest appearance in North America); *Lepus* LSD<sub>k</sub>, base of the Olduvai (chron C2n) at approximately 1.95 Ma; *Pewelagus* HSD<sub>k</sub>, in the middle of chron C2n (Olduvai), at approximately 1.92 Ma (or slightly younger, using the calibration of Berggren et al. 1995); *Smilodon* LSD<sub>k</sub>, top of Olduvai, 1.77 Ma; and *Euceratherium* LSD<sub>k</sub>, late in the Matuyama, at approximately 1.13 Ma.

We attempt no final determination of the utility of the traditional faunal terms or of the newly proposed range zones, pending a thorough analysis of the stratigraphic distribution of taxa in the sequence. The faunal lists provided by Remeika et al. (1995) and Cassiliano (1999) and those in the database at the Anza-Borrego Desert State Park (G. Jefferson, pers. comm., May 2000) are not concordant, so the resolution of this question must wait.

The sequence contains the earliest temporally constrained North American occurrences of *Nothrotheriops*, *Sylvilagus*, *Microtus* with five closed triangles on the lower first molar, and *Euceratherium* (*Euceratherium* from the Topaz Lake Fauna in Nevada may be older). It also includes the youngest occurrence of *Borophagus diversidens*.

External (nonbiostratigraphic) age control in the Anza-Borrego sequence includes a fission-track date (on zircons from a volcanic tuff) of  $2.3 \pm 0.4$  Ma (Johnson et al. 1983). This date helped to anchor the magnetic polarity stratigraphy in the section and confirmed the original polarity determination at the 3.6-km level as representing the Gauss–Matuyama boundary (chron C2An–2r boundary; Opdyke et al. 1977; Johnson et al. 1983). A subsequent chal-

lenge to the original interpretation of the higher portion of the magnetic sequence at Anza-Borrego (Repenning 1992) was based on the presence, high in the section, of the vole *Microtus* (= *Terricola*) *meadensis*. Repenning (1992:72) argued that the youngest appearance of this species indicated that the normal polarity recorded in the Vallecito Creek–Fish Creek sequence represented the Brunhes and not the Jaramillo, as originally interpreted by Johnson et al. (1983). Resolution of this question will require additional field work and a careful magnetic sampling in the higher portion of the section, but the single specimen used by Repenning (1992) to argue for the alternative interpretation is associated with contradictory locality data and may not be from the Vallecito–Fish Creek sequence in the park (L. K. Murray and C. J. Bell, pers. obs., 1999).

A small fauna from the Coso Mountains of southern California includes *Hypolagus*, *Cosomys primus*, *Borophagus diversidens* (originally identified as *Borophagus solus*; see Wang et al. 1999), *Platygonus* sp., and “*Plesippus francescana*” (Wilson 1932; J. Schultz 1937). K–Ar dates on a series of basalt flows and rhyolites in the Coso Formation indicate that the fauna dates to at least 3 Ma (Bacon et al. 1979).

Two other sequences in southern California also yielded significant collections of Blancan mammals in the last decade. Hemphillian and Blancan faunas from the San Timoteo Badlands (discussed in part earlier) were reported by Albright (1999), who recognized three main stratigraphic sections (Eden Hot Springs–Jack Rabbit Trail, Riverside County Landfill–El Casco, and San Timoteo Canyon–Live Oak Canyon). The Hemphillian Mt. Eden Fauna is late Hemphillian and contains *Teleoceras*, *Dinohippus*, *Pediomyx hemphillensis*, and *Agriotherium*; Albright placed it in chron C3r. The section above this lies entirely in the Blancan with *Thomomys* appearing in chron C2Ar, *Sigmodon minor* in chron C2An.3n, and *Neotoma* in chron C2An.1r. At the top of the section *Ophiomys*, “*Plesippus idahoensis*,” and *Erethizon* are in chron 2r.2r. The Riverside County Landfill–El Casco section contains Blancan deposits extending from the upper part of the Gilbert (chron C2Ar) and includes *Neotoma* (*Paraneotoma*) *fossilis*, *Prodipodomys*, *Thomomys*, and *Sigmodon minor* from chron C2An.1n and possibly from C2An.2n. The El Casco Fauna in the San Timoteo Badlands of southern California was assigned to the Irvingtonian by Albright (1999). The fauna includes “*Plesippus francescana*” (the youngest known record of “*Plesippus*”) and lacks *Mammuthus* (which appears higher in the San Timoteo Badlands sequence; Albright 1999). *Microtus*, an advanced form of *Ondatra idahoensis*, *Mictomys kansansensis*, *Neotoma*, *Canis edwardii*, and *Erethizon ?cascoensis* are also present and provide faunal evidence for

assignment to the Irvingtonian as it is traditionally recognized in California. This part of the section is reversely magnetized and was assigned by Albright to chron C1r.2r. Albright suggested that the position of the fauna within this interval indicates an age of 1.3–1.4 Ma. The reported age of the El Casco Fauna is temporally consistent with an assignment to either the latest Blancan or the earliest Irvingtonian as they are recognized here. We treat the fauna as latest Blancan, primarily to exclude “*Plesippus*” from Irvingtonian faunas.

Vertebrate faunas recovered from the Temecula Arkose and an unnamed sandstone unit in the vicinity of Murrieta (Riverside County) include a diverse assemblage of small mammals that record faunal change in the Elsinore Fault Zone from early through late Blancan. These faunas and their correlation were discussed at length by Golz et al. (1977), Reynolds et al. (1991), and Pajak et al. (1996).

**Great Basin** Little is known of Blancan mammals from the Great Basin. In western Nevada two areas produced superposed Blancan faunas (Kelly 1994). Sediments in the Carson Valley–Pine Nut Mountains (recently given the name *Sunrise Pass Formation* by Trexler et al. 2000) area produced one Hemphillian and two Blancan faunas in superposition. The older Blancan Buckeye Creek Fauna contains *Platygonus* cf. *P. pearcei*, *Alilepus vagus*, *Hypolagus gidleyi*, *Megalonyx leptostomus*, *Ursus abstrusus*, “*Equus simplicidens*,” *Hemiauchenia* cf. *H. blancoensis*, and *Odocoileus*. *U. abstrusus* is limited to the early Blancan, and *H. gidleyi* and *A. vagus* are most common in the early Blancan (Kelly 1994).

The younger Fish Springs Flat Fauna (Kelly 1994) contains *Hypolagus furlongi*, *Satherium ingens*, “*Equus simplicidens*,” *Equus giganteus*, *Gigantocamelus spatula*, *Thomomys carsonensis*, *Mammot americanum*, and *Spermophilus howelli*. A late Blancan age assessment is based on the joint occurrence of *Hypolagus furlongi*, *Spermophilus howelli*, *Satherium ingens*, and “*Equus simplicidens*.”

Two superposed faunas from an unnamed formation in the Wellington Hills–Antelope Valley area of Douglas County, Nevada, were reported by Kelly (1997). The Blancan Wellington Hills Fauna is depauperate but includes “*Equus idahoensis*” and *Spermophilus wellingtonensis*. The Topaz Lake Fauna includes *Ondatra* cf. *O. idahoensis*, “*Equus idahoensis*,” *Equus* cf. *E. giganteus*, *Hemiauchenia*, and *Euceratherium collinum* (Kelly 1997). Kelly (1997) interpreted the Topaz Lake Fauna to be early Irvingtonian based on the presence of *Euceratherium*. This may be the oldest record of *Euceratherium*; the oldest record associated with external age control is in the Vallecito

Creek–Fish Creek sequence at approximately 1.1 Ma (Casiliano 1999). The association with a primitive muskrat is unique (the youngest record of *O. idahoensis* associated with external age control is from Froman Ferry, Idaho, at approximately 1.72 Ma [Repenning et al. 1995]).

Late Cenozoic vertebrates were reported from eastern Nevada by Stock (1921), who called the deposits in Meadow Valley the Panaca beds and those in Muddy Valley the Muddy Valley beds. He considered the Panaca beds to be “early Pliocene” (now considered to be Hemphillian and late Miocene) on the basis of the recovered horse, camel, and rhinoceros remains and the Muddy Valley beds to be slightly older. An age determination, ranging from Barstovian to Clarendonian (Muddy Valley Fauna) and Clarendonian to Hemphillian (Panaca), was suggested by MacDonald and Pelletier (1958). The Meadow Valley Fauna from the Panaca Formation (Reynolds and Lindsay 1999; Mou 1997; Lindsay et al. 2002) contains a number of Blancan taxa such as *Sorex meltoni*, *Hypolagus edensis*, *Hypolagus tedfordi*, *Hypolagus* cf. *H. ringoldensis*, *Pewelagus dawsoni*, *Nekrolagus progressus*, *Pliogeomys parvus*, *Repomys panacaensis*, *Mimomys panacaensis*, *Borophagus diversidens*, *Taxidea* sp., ?*Martinogale*, *Bassariscus casei*, *Felis* sp., ?*Cuvieronius*, *Dinohippus*, “*Equus* cf. *E. simplicidens*,” *Equus* (*Hemionus*) sp., *Platygonus* sp., *Megatylopus* sp., *Hemiauchenia*, and an unidentified Caprini species. The earlier reported presence of *Ophiomys mcknighti*, *Ophiomys magilli*, and *Ondatra* or *Pliopotamys* (Repenning 1987; Reynolds and Lindsay 1999) in this fauna was in error (see Lindsay et al. 2002). The *Dinohippus* specimen and a rhinoceros (?*Teleoceras*) are limited geographically and stratigraphically in Meadow Valley; it appears that they came from low in the section and indicate that the lowest exposures of the Panaca beds in Meadow Valley are of Hemphillian age, whereas most of the exposures yield only Blancan fossils. Paleomagnetic and radiometric data from the sequences in both eastern and western Nevada were obtained and used by Lindsay et al. (2002) to date the Hemphillian–Blancan boundary. This boundary was discussed more fully earlier in the section on the Hemphillian–Blancan transition.

The White Narrows Fauna south of Panaca may also represent the earliest Blancan (Reynolds and Lindsay 1999). Definitive species-level identifications for most taxa are not available, but the fauna includes *Sorex*, *Peromyscus valensis*, and *Dipodomys gidleyi*. *Sorex* is known only from Blancan and younger faunas (McKenna and Bell 1997), *D. gidleyi* only from the Blancan, and *P. valensis* only from the Hemphillian. The best available age assessment for the White Narrows Fauna is that it repre-



sents the early Blancan (Reynolds and Lindsay 1999; Mead and Bell 2001).

**Southwestern United States** In the San Pedro Valley in southern Arizona, the St. David Formation is nearly 120 m (400 feet) thick and extends from the upper part of the Gilbert reversed chron (chron C2Ar) to the lower part of the Brunhes normal chron (chron C1n) (Johnson et al. 1975; Lindsay et al. 1975). Within these sediments a series of faunas range in age from the early Blancan Benson Fauna within the Gauss normal chron to the latest Blancan Curtis Ranch Fauna at the Olduvai normal subchron (Gidley 1922, 1926; Gazin 1942; Lindsay et al. 1975). Radiometric control in the paleomagnetic sequence is provided by a zircon fission-track date of  $3.1 \pm 0.7$  Ma on an ash bed immediately below the Post Ranch (type Benson) faunal horizon and by identification of the Huckleberry Ridge (formerly Pearlette B) Ash in the California Wash section by G. Izett (1981); the age of this ash is somewhere between 2.0 and 2.1 Ma (Izett and Honey 1995; Gansecki et al. 1998). A proposed boundary between the Blancan and Irvingtonian was arbitrarily drawn between about 1.8 and 2.0 Ma at the lowest known local stratigraphic occurrence of *Lepus* in the Curtis Ranch Fauna (Lindsay et al. 1975). The Curtis Ranch Fauna occurs in sediments that straddle the base of the Olduvai subchron at about 1.95 Ma and includes *Ondatra idahoensis*, the earliest local record of *Dipodomys*, and a late record of *Stegomastodon*. Specimens from the fauna originally assigned to *Lepus* were subsequently reidentified as *Sylvilagus* (White 1991b:78). Extrapolating from paleomagnetic data presented by Johnson et al. (1975; with modified calibrations for polarity transitions from Berggren et al. 1995), *Ondatra idahoensis* and *Glyptotherium arizonae* appear in the local sequence (California Wash Fauna) between about 2.2 and 2.1 Ma, whereas *Nannippus* disappears at approximately 2.6 Ma (Johnson et al. 1975). The Curtis Ranch Fauna was traditionally considered to be early Irvingtonian but is here considered to be latest Blancan; neither *Lepus* nor *Mammuthus* is recorded from the Curtis Ranch Fauna.

Pliocene fossiliferous sediments on the flanks of Dry Mountain on the 111 Ranch were studied by Lance (1960), who recognized two faunas, the Flat Tire and Tusker claims, later modified by P. Wood (1960, 1962) to Flat Tire and Tusker faunas. The lower Flat Tire Fauna was considered to be Blancan on the basis of the presence of *Nannippus*, whereas the upper Tusker was assigned an Irvingtonian age because of the absence of *Nannippus*. Later studies in the San Pedro Valley to the southwest showed the presence of a fauna that lacked *Nannippus* su-

perposed over a fauna with *Nannippus*, but both faunas were best considered Blancan (Johnson et al. 1975). On the basis of these faunas and the resemblance of the Tusker rodents to the Benson and Curtis Ranch sites of the San Pedro Valley, the Tusker assemblage was assigned to the Blancan (Galusha et al. 1984). No justification for two distinct faunas was found, and the names *Flat Tire* and *Tusker* were abandoned. The entire sequence now should be called simply the 111 Ranch Fauna. One hundred meters of section span the later Gauss and the early (pre-Olduvai) part of the Matuyama (Galusha et al. 1984; see also Seff 1960; P. Wood 1960). An ash bed yielded pyrogenic zircons with fission-track dates of  $2.33 \pm 0.24$  Ma (Dickson and Izett 1981; date given as  $2.32 \pm 0.15$  Ma by Galusha et al. 1984). The fauna consists of at least 41 mammal taxa that are not evenly distributed through the section. The highest occurrence of *Nannippus* is just below the Gauss–Matuyama boundary. The first local occurrences of *Glossotherium*, *Glyptotherium*, *Neochocerus*, and the arvicoline rodents *Ondatra idahoensis*, *Pliophenacomys*, and *Mictomys vetus* are also located in the uppermost Gauss and identify a short time interval that is useful for correlation in the southwest. This record is among the earliest of *Mictomys*, used to define the base of the Blancan V of Repenning (1987; Repenning et al. 1990).

A new fauna, the Artesia Road Fauna, in the nearby Whitlock Mountains and San Simon Valley was included in an expanded “111 Ranch beds” by McCord et al. (2002). This fauna produced the first records of *Castor* and *Tapirus* from the 111 Ranch area. A new occurrence of cervid was also reported (as “*Bretzia* or *Odocoileus*”). The relationships of this fauna with the traditional 111 Ranch are not well established.

Four stratigraphic sequences with extensive faunas from the Safford and Duncan basins in Arizona were reported by Tomida (1987). The Duncan Fauna produced at least 25 taxa, including 14 small mammals. The rodents include *Geomys* (*Nerterogeomys*) cf. *G. persimilis*, *Perognathus gidleyi*, *Peromyscus hagermanensis*, *Reithrodontomys rexroadensis*, *Baiomys minimus*, *Calomys* (*Bensonomys*) sp., *Sigmodon minor* (originally identified as *Sigmodon medius*, but see Harrison 1978; R. Martin 1986), *Neotoma* (*Paraneotoma*) *fossilis*, *Pliopotamys*, and *Ophiomys* cf. *O. taylori*. Large mammals include *Nannippus peninsulatus*, *Equus* (*Dolichohippus*), *Equus* (*Asinus*), and camelids. The presence of *Nannippus* suggests an age no younger than the Gauss–Matuyama boundary. The lower part of the Duncan section has normal polarity, the upper has reversed polarity; the section was correlated with the base of the Mammoth subchron (chron C2An.2r)

by Tomida (1987). The Country Club Fauna contains only a few taxa of small mammals. The section shows a reversed–normal–reversed paleomagnetic pattern and was interpreted to extend from the Mammoth through the Kaena subchron (2An.1r) in the Gauss chron (Tomida 1987). The joint presence of *Sigmodon*, *Neotoma quadriplicata*, and *Equus* precludes an age older than the Gauss chron. The Bear Springs section produced too little material to determine its age other than Blancan. The presence of *Nannippus*, *Geomys*, and *Sigmodon* suggests placement in the Gauss or early Matuyama chrons (the youngest occurrence of *Nannippus* is in the Macasphalt Shell Pit Fauna in Florida at approximately 2.1 Ma). The thick sequence of normally magnetized sediments at Bear Springs could represent either the long normal chron (chron C2An.3n) below the Mammoth or the long normal chron (chron C2An.1n) above the Kaena.

A number of fossil vertebrate sites on the Pearson Mesa in the Duncan Basin were reported by Tedford (1981), Tomida (1987), and Morgan and Lucas (2000a). Forty-three different sites produced vertebrate fossils on Pearson Mesa (Morgan and Lucas 2000a). Fossils were collected from 60 m of section, but the lower 15 m contains *Nannippus peninsulatus*, "*Equus simplicidens*," *Equus* cf. *E. cummingsii* (see Cope 1893:67 for correct spelling of the species), *Platygonus bicalcaratus*, *Glossotherium* cf. *G. chapadmalense*, and *Hemiauchenia* cf. *H. blancoensis*, all suggesting a Blancan age. *Geomys* cf. *G. persimilis*, *Stegomastodon mirificus*, *Nannippus peninsulatus*, and *Equus* sp. from Pearson Mesa were reported by Tomida (1987). All of Tomida's paleomagnetic sites are of normal polarity, which he interpreted to fall in the long normal interval (C2An.1n; 2.581–3.040 Ma) in the Gauss chron between the Kaena subchron and the Matuyama chron. The faunal assemblage and paleomagnetic data indicate a middle Blancan age.

The Verde Fauna, from Yavapai County in central Arizona (Czaplewski 1987, 1990), contains *Ogmodontomys poaphagus*, *Geomys (Nerterogeomys) minor*, *Calomys (Bensonmys) arizonae*, and *Sigmodon minor*. *Copemys*, a taxon typically found in Hemphillian and earlier faunas, is present in low abundance in the Verde Fauna and was interpreted by Czaplewski (1990) to be a temporal extension of this taxon into the Blancan. The identification of the rare arvicoline *Nebraskomys* in the Verde Fauna (Repenning 1987) was subsequently shown to be erroneous (Winkler and Grady 1990; Czaplewski 1990); the specimen in question cannot be identified reliably to any arvicoline genus. The Verde Fauna is situated approximately 56 m above a basalt dated at 5.6 Ma and is placed at the base of the Nunivak subchron (chron 3n.2n) (Bressler and Butler

1978; Czaplewski 1990) at about 4.62 Ma (Berggren et al. 1995). The fauna includes the earliest occurrence of *Ogmodontomys poaphagus*.

Many of the Blancan faunas of New Mexico were reviewed by Tedford (1981). Subsequent work by Morgan and Lucas (1999, 2000a, 2000b) and Morgan et al. (1997, 1998) added new localities and new material. Blancan faunas are located in two areas of New Mexico: sedimentary basins along the Rio Grande Valley and structural basins in southwestern New Mexico. The Santo Domingo Basin north of Albuquerque, considered by Morgan and Lucas (2000b) to be a northward extension of the Albuquerque Basin, produced a fauna including *Equus scotti* and an equid similar to *Equus calobatus* (Tedford 1981); a late Blancan age was suggested based on the similarity of the horses to those of the Tule Formation of Texas (here considered to be Irvingtonian). The subsequent discovery of *Nannippus* by Morgan and Lucas (2000b) confirmed Tedford's placement of the fauna in the Blancan. Tedford noted that these deposits are interbedded with the Santa Ana Mesa Basalt dated ~2.4 Ma (Bachman and Mehnert 1978). The deposits are overlain by the lower bed of the Otowi Member of the Bandelier Tuff dated to approximately 1.61 Ma (Izett and Obradovich 1994; Spell et al. 1996; Morgan and Lucas 2000b). This assemblage apparently is late Blancan.

Ten Blancan localities were recently recognized in the Albuquerque Basin (Morgan and Lucas 1999, 2000b). None are clearly early Blancan, but the diversity is low. One fauna, the Belen, contains *Scalopus (Hesperoscalopus)*, *Geomys (Nerterogeomys)* cf. *G. paenebursarius*, *Equus* cf. *E. calobatus*, and *Stegomastodon mirificus*. An age range of 2.5–3.7 Ma was suggested by Morgan and Lucas (2000b). The Pajarito locality (= Laguna site of Tedford 1981) produced dentaries of *Geomys (Nerterogeomys)* that are similar in size to those of the Belen site. An  $^{40}\text{Ar}/^{39}\text{Ar}$  date of  $3.12 \pm 0.10$  Ma on a pumice clast (Maldonado et al. 1999) from the Pajarito locality led Morgan and Lucas (2000b) to suggest that *G. (N.) paenebursarius* also occurs in the middle Blancan, but the clast establishes only a maximum age for the unit. The Tijeras Arroyo area has Blancan and Irvingtonian in superpositional relationship (Logan et al. 1984; Lucas et al. 1993; Morgan and Lucas 1999, 2000b). The lower part of the section has *Hypolagus* cf. *H. gidleyi* and *Equus* cf. *E. cummingsii* and was assigned a middle to late Blancan age (2.2–3.5 Ma) by Morgan and Lucas (2000b).

Two areas in southwestern New Mexico recently produced Blancan faunas. In the Mangas Basin, the Buckhorn Fauna was recovered from 14 sites (Morgan et al. 1997). It contains *Nannippus*, *Ogmodontomys* cf. *O. poa-*

*phagus*, *Repomys* cf. *R. panacaensis*, and “*Equus* (*Dolichohippus*) *simplicidens*,” but Neotropical taxa are not present. The presence of “*E. simplicidens*” seems to preclude a very early Blancan age, and Neotropical taxa would be expected if the fauna were of late Blancan age. The rodents suggest a late–early to early–mid-Blancan age, but definitive species identifications are lacking.

The Camp Rice Formation in Doña Ana County, New Mexico, produced the Tonuci Mountain Fauna (Morgan et al. 1998). The fauna includes *Canis lepophagus*, *Borophagus*, *Nannippus* cf. *N. peninsulatus*, “*Equus simplicidens*,” *Equus scotti*, *Platygonus* cf. *P. bicalcaratus*, *Hemiauchenia blancoensis*, and *Cuvieronius*. Biostratigraphic correlation and paleomagnetic data constrain the age to between 3.6 and 3.0 Ma (Morgan et al. 1998).

The Mesilla Basin in south-central New Mexico contains a sedimentary sequence that spans the Blancan–Irvingtonian boundary. Vanderhill (1986) divided the sequence into three biostratigraphic zones. The lowest zone (“Faunule A” of Vanderhill) contains the Blancan taxa *Nannippus peninsulatus*, *Hemiauchenia blancoensis*, and *Glyptotherium*. The presence of *Glyptotherium* suggests at least a late Blancan age (the earliest appearance of *Glyptotherium* probably is in the 111 Ranch sequence; Galusha et al. 1984). Paleomagnetic data place this zone in the late Gauss, between the Kaena event (chron C2An.1r) and the Gauss–Matuyama (chron C2An–2r boundary). Faunule B in the Mesilla Basin is derived from sediments that span the base of the Matuyama to the Olduvai event. It lies above the highest occurrence of *Nannippus* and below the lowest occurrence of *Mammuthus* and contains a mixture of taxa traditionally used to characterize both the Blancan and Irvingtonian. Characteristic Blancan forms include *Gigantocamelus* and *Blancocamelus*. Characteristic Irvingtonian forms are *Paramylodon harlani* and *Smilodon*. The uncertain stratigraphic position of some specimens collected before Vanderhill’s field work make it impossible at this time to determine the position of the Blancan–Irvingtonian boundary in the Mesilla Basin, but *Mammuthus* was recovered from chron C1r.2r (between the Cobb Mountain and Olduvai events) within faunule C. Faunule C in Mesilla Basin was derived from sediments that span the time from just after the Olduvai event to the early part of the Brunhes, but most of the specimens were reported to come from sediments that predate the Jaramillo event. (Faunule C is discussed later in this chapter under “Irvingtonian.”)

The Hueco Bolson, a topographic basin southwest of the Hueco Mountains in the western part of Trans-Pecos Texas, contains sediments that produced a Blancan fauna that extends through the Fort Hancock and Camp Rice

formations (Strain 1966). Although the two formations are separated by an unconformity, the material from both units was combined as the Hudspeth Fauna. Small animals were recovered from the Fort Hancock Formation and large mammals from the Camp Rice Formation. The combined fauna contains *Nannippus peninsulatus*, “*Equus idahoensis*,” “*Equus simplicidens*,” *Gigantocamelus*, *Glyptotherium*, *Megalonyx*, and *Sigmodon hudspethensis*. Paleomagnetic data for this sequence were obtained by Vanderhill (1986). Most of the Fort Hancock and the lower part of the Camp Rice formations fall in a normal interval that Vanderhill identified as the late Gauss chron (presumably chron C2An.1n). The age is constrained by the presence of the Huckleberry Ridge Ash (between 2.0 and 2.1 Ma; see table 7.1) high in the section.

The sediments of the Red Light Bolson, which lies between the Eagle and Indio mountains on the northeast and the Quitman Mountains on the southwest, in the southernmost part of Hudspeth County, Texas, produced two faunas (Akersten 1972). The Aguila Fauna was recovered from the upper part of the Bramblett Formation and the Red Light Fauna from the overlying Love Formation. The former is represented by a single fish, unidentified birds, and an indeterminate rodent and is not biostratigraphically useful. The Red Light Fauna contains a diverse Blancan fauna including *Megalonyx*, “*Paramylodon*” sp., *Glyptotherium texanum*, *Borophagus*, *Nannippus peninsulatus*, *Hemiauchenia blancoensis* (originally listed as *Tanupolama*, but see Webb 1974b), and *Platygonus bicalcaratus*. The presence of *Glyptotherium* suggests that it is late Blancan. Elsewhere in western North America *Nannippus peninsulatus* disappears from faunas by approximately 2.5 Ma. In Florida, *N. peninsulatus* persists until approximately 2.1 Ma (Macasphalt Shell Pit). There are at present no radiometric or paleomagnetic data bearing on the age of this fauna.

**Mexico** Blancan faunas are known from a number of localities in Mexico, and a few contribute information on the Hemphillian–Blancan boundary. The boundary between Hemphillian and Blancan faunas in Chihuahua was documented by Lindsay et al. (1984) and Lindsay and Jacobs (1985; summarized earlier).

In central Mexico, a number of localities produced faunas of Blancan age. The La Goleta locality (Arellano and Azcón 1949; Repenning 1962; Miller and Carranza-Castañeda 1984) in Michoacan has a fauna that contains *Rhynchotherium falconeri*, *Paenemarmota barbouri*, *Nannippus* cf. *N. montezuma*, *Nannippus peninsulatus*, *Pliohippus*, *Equus* (*Dolichohippus*), a hyaenid, a tapir, a peccary, a camel, and an antilocaprid. A tentative referral to

the Hemphillian was proposed by Repenning (1962) on the basis of *Pliohippus*, but he acknowledged a written communication from R. Tedford that suggested the fauna was Blancan on the basis of the hyaenid and horses. Miller and Carranza-Castañeda (1984) suggested that both Hemphillian and Blancan components may be present. The fauna probably includes the earliest record of *N. peninsulatus*. The uppermost beds of the La Goleta locality include remains of *Mammuthus* and *Equus* (*Equus*), indicating that an Irvingtonian or Rancholabrean deposit caps the sequence.

The Rancho Viejo Fauna in Guanajuato contains *Glyptotherium*, *Glossotherium garbanii*, *Hypolagus mexicanus*, *Pratilepus ?kansasensis*, *Paranotolagus complicatus*, *Paenemarmota*, *Neochoerus cordobai*, *Neochoerus holmesi*, *Borophagus diversidens*, *Nannippus peninsulatus*, and *Equus* (*Dolichohippus*) sp. (Carranza-C. and Miller 1980; Miller and Carranza-Castañeda 1984; Carranza-Castañeda and Miller 1988). A Blancan age was proposed by Miller and Carranza-Castañeda (1984). A fission-track date of 3.6 Ma was derived from an ash stratigraphically 1 m below the *Borophagus* specimen (Kowallis et al. 1986; Miller and Carranza-Castañeda 1998b). This is the oldest well-dated record of *Glyptotherium* and *Borophagus diversidens* (Wang et al. 1999).

The Rancho El Ocote Fauna in Guanajuato is primarily Hemphillian (Carranza-Castañeda and Ferrusquía-Villafranca 1978; Dalquest and Mooser 1980; Miller and Carranza-Castañeda 1984; Montellano 1989; Carranza-Castañeda and Walton 1992; Carranza-Castañeda and Miller 2000), but there is a Blancan component to the fauna with *Rhynchotherium*, cf. *Glossotherium*, and *Nannippus peninsulatus* (Miller and Carranza-Castañeda 1984; Carranza-Castañeda and Miller 2000). *Equus* (*Dolichohippus*) was reported from the Rancho El Ocote by Miller and Carranza-Castañeda (1984) but was excluded from the faunal list provided by Carranza-Castañeda and Miller (2000:43). Early attempts to obtain zircon fission-track ages for the fauna were unsuccessful because of mixing or contamination of the dated materials (Kowallis et al. 1986), but more recent efforts using both fission-track and  $^{40}\text{Ar}/^{39}\text{Ar}$  successfully differentiated the components and yielded ages for the fauna of between  $4.6 \pm 0.3$  and  $4.70 \pm 0.07$  Ma (Kowallis et al. 1998).

One well-studied fauna from Baja California Sur, the Las Tunas (Miller 1980), contains *Hypolagus vetus* (see White 1987), *Borophagus hilli* (originally identified as "*Borophagus ?diversidens*;" identification changed by Wang et al. 1999), *Rhynchotherium* cf. *R. falconeri*, and "*Equus* cf. *E. simplicidens*." No external age control is

available, but a tentative assignment to the early Blancan was proposed.

In the state of Aguascalientes, a sequence of three stratigraphic units produced the Cedazo faunas, which probably represent a series of faunas ranging from Blancan through Rancholabrean (Mooser Barendun 1958; Mooser and Dalquest 1975a, 1975b; Montellano-Ballesteros 1992). The age of the faunas is difficult to assess; much of the collection was reported to be in private hands (see Mooser and Dalquest 1975b), and the stratigraphic relationships of the reported species were not published "for the sake of brevity" (Mooser and Dalquest 1975b:784). A more recent attempt to place the faunas in stratigraphic context resulted in the tentative recognition of three discrete units and faunas. The lower unit contains *Holmesina*, cf. *Aluralagus*, *Pappogeomys*, *Equus conversidens*, and *Platygonus* sp. and was assigned an Irvingtonian age by Montellano-Ballesteros (1992) on the basis of the tentative identification of cf. *Aluralagus* and on the morphologic similarity of the *Holmesina* material to that recovered from Inglis 1A, Florida. We consider the lower unit to be latest Blancan. Complete faunal lists for the other units can be found in Mooser and Dalquest (1975a, 1975b) and Montellano-Ballesteros (1992), but age relationships of the taxa are poorly understood. At least one part of the sequence contains *Bison* and is therefore Rancholabrean. It is not clear whether a discrete Irvingtonian fauna is present.

**Great Plains** In the Great Plains, especially in southwestern Kansas and the Texas panhandle, the faunal sequence must be compiled from shorter local sections using mainly paleomagnetic and biostratigraphic criteria because few radiometric dates are available. Careful biostratigraphic studies enabled C. W. Hibbard and his coworkers to compile an impressive succession of Blancan faunas in southwestern Kansas. A large number of Blancan faunas are known from this region, but few are discussed here; ongoing work in the region by R. Martin and colleagues (2000) is building on the foundation laid by Hibbard and expanding our understanding of the taxonomic composition and age relationships of the mammalian faunas. With the adoption of *Mammuthus* as the defining taxon for the Irvingtonian, many faunas previously considered to be Irvingtonian are here considered to be latest Blancan.

The Saw Rock Canyon Fauna in Seward County, Kansas (Hibbard 1949b, 1953, 1964, 1967), lacks paleomagnetic data but is considered one of the earliest faunas in the Rexroad Formation and is near the Hemphillian–Blancan boundary. The approximate age placement of

the Saw Rock Canyon Fauna (at 4.3 Ma) given by Lundelius et al. (1987, figure 3) may be too young according to R. Martin et al. (2000), who gave an age estimate of approximately 5.0 Ma. The mammalian fauna is dominated by small-bodied species and includes several rodent species that are more primitive than species recovered from the early Blancan Fox Canyon and Rexroad faunas (Hibbard 1949b, 1953, 1964, 1967; Zakrzewski 1967; R. Martin et al. 2000). The Saw Rock Canyon Fauna contains a beaver (*Dipoides wilsoni*), a gopher with rooted teeth (possibly *Pliogeomys*), a primitive arvicoline *Ogmodontomys sawrockensis*, and *Borophagus hilli* (originally identified as a species of *Osteoborus*; identification changed by Wang et al. 1999), but no horses. An evolutionary transition from *O. sawrockensis* to *Ogmodontomys poaphagus* was suggested by Zakrzewski (1967) based on material from Saw Rock Canyon.

Several other Great Plains faunas contain characteristic early Blancan taxa. The Fox Canyon Fauna (Meade County, Kansas; Hibbard 1950; Hibbard and Zakrzewski 1972) occurs in reversely magnetized sediments (Lindsay et al. 1975) and is one of the oldest Blancan faunas in the Great Plains area. A recent age range estimate between 4.8 and 4.3 Ma was given by R. Martin et al. (2000). Small mammals are abundant, but large mammals are rare. The fauna contains taxa typical of Blancan and younger faunas (e.g., *Pliophenacomys finneyi*, *Geomys*, and *Odocoileus*) as well as *Paenemarmota*, a sciurid also known from the Hemphillian. The earlier report of *Hypolagus vetus* in the Fox Canyon Fauna (Lundelius et al. 1987) was based on a personal communication from John White but cannot be confirmed at this time; it is not listed or discussed from Fox Canyon by White (1987, 1991b) or R. Martin et al. (2000). Recent collecting efforts in Kansas produced new faunas (e.g., Argonaut, Red Fox, Fallen Angel) that may be older than Fox Canyon but younger than Saw Rock Canyon (R. Martin et al. 2000).

The Rexroad 3 Fauna is another diverse early Blancan fauna but is considered to be younger than the Fox Canyon Fauna based on the presence of *Nannippus peninsulae*, *Stegomastodon*, *Canis lepophagus*, *Trigonictis*, and *Borophagus diversidens* in the Rexroad 3 (R. Martin et al. 2000). The fauna was recovered from normally magnetized sediments tentatively correlated by Martin et al. (2000) with chron C2An.3n (early Gauss), now dated to between 3.58 and 3.33 Ma (Berggren et al. 1995). It includes the youngest known occurrence of *Megantereon* and the earliest known occurrences of *Nebraskomys*, *Platygonus bicalcaratus*, and *Stegomastodon*.

The White Rock Fauna in Republic County, Kansas, was described by Eshelman (1975). External age control is

lacking, but the fauna includes *Megalonyx leptostomus*, *Hypolagus* cf. *H. furlongi*, *Procastoroides* sp., *Peromyscus cragini*, *Sigmodon minor*, *Nebraskomys mcgrewi*, *Ophiomys meadensis*, *Ogmodontomys* sp., *Pliophenacomys*, and one of the earliest records of *Ondatra idahoensis*. All of these taxa are limited to the Blancan.

The Borchers Fauna figures prominently in chronology of the Great Plains sequence because it lies immediately over the Huckleberry Ridge Ash, dated between  $2.003 \pm 0.014$  (Ganseccki et al. 1998) and  $2.10 \pm 0.02$  Ma (Honey et al. 1998). The Borchers Fauna is depauperate but contains *Lepus*, *Geomys*, *Neotoma taylori*, *Sigmodon minor*, *Ondatra idahoensis*, *Mictomys landesi*, *Prodipodomys*, *Dipodomys hibbaridi*, *Perognathus gidleyi*, *Canis lepophagus*, *Urocyon*, *Camelops*, and *Stegomastodon* (Hibbard 1942; R. Martin et al. 2000). It lacks *Allophaiomys*, which is sometimes used to define the beginning of the Irvingtonian in the Great Plains (Repenning 1992; Repenning et al. 1995; R. Martin et al. 2000). The Borchers Fauna was previously suggested to be indicative of a warm, mild climate (Zakrzewski 1988). The reported occurrence of *Lepus* in the fauna (Hibbard 1941; R. Martin et al. 2000) must be verified. If it is correct, it represents the earliest record of *Lepus* in North America; preliminary notice and an illustration of the occlusal surface of the teeth of the only reported specimen were provided by Hibbard (1941:216–217, plate 2, figure 9). No additional discussion of the material or its identification was provided by R. Martin et al. (2000). We tentatively accept the Borchers record as the oldest dated occurrence of *Lepus*. The fauna also includes the youngest dated occurrence of *Prodipodomys*. Additional potentially young records of *Prodipodomys* include the Java Fauna, South Dakota (which lacks external age control), and an occurrence stratigraphically above the Bishop Ash in California Oaks (Reynolds et al. 1991). Subsequent clarification of the California Oaks faunas, California (Pajak et al. 1996), revealed that three of the four putative records of *Prodipodomys* were questionably identified or came from mixed faunas; the fourth record is not in stratigraphic association with the Bishop Ash and lacks any age control.

There are four faunas stratigraphically above the Borchers; these were ordered from oldest to youngest by R. Martin et al. (2000) as the Nash Fauna, Aries A (= "Aries" of Izett and Honey 1995), Rick Forester, and Aries B. We consider all of these faunas to be latest Blancan. The Nash Fauna (Meade County, Kansas; Bayne 1976) was recovered from a channel fill deposit in the Crooked Creek Formation (Eshelman and Hibbard 1981). The Aries A locality is the original quarry mentioned by Izett and Honey (1995; see R. Martin et al. 2000). The Aries B locality is

stratigraphically just above Aries A and just below the Cerro Toledo B Ash (named, but not dated, by Izett 1981; thought to be associated with the eruption of the Cerro Toledo Rhyolite between 1.2 and 1.4 Ma [table 7.1]; Izett and Honey 1995; see R. Martin et al. 2000 for relative stratigraphic placement of the faunas). These localities are situated between the Huckleberry Ridge Ash ( $2.11 \pm 0.01$  Ma) and Cerro Toledo B Ash. Nash, Aries A, and Rick Forester all contain *Allophaiomys pliocaenicus* [= *Pitymys* (“*Allophaiomys*”) of Zakrzewski 1988 and *Microtus pliocaenicus* of R. Martin et al. 2000], which is missing from the Borchers Fauna. According to Zakrzewski (1988), the presence of *Mictomys kansasensis* and *Allophaiomys* suggests the beginning of climatic cooling in this area. If the relative stratigraphic (and temporal) placement of these faunas by R. Martin et al. (2000) is correct, the Nash Fauna would record the earliest known appearance of *Allophaiomys* and *Mictomys kansasensis*.

The Sand Draw mammalian fauna in Nebraska was described by Hibbard (1972). It includes *Prodipodomys centralis*, *Dipoides rexroadensis*, *Procastoroides*, *Sigmodon minor* (originally identified as *Sigmodon medius*, but see Harrison 1978; R. Martin 1986), *Nebraskomys mcgrewi*, *Ogmodontomys poaphagus*, *Ophiomys magilli*, *Ophiomys fricki*, *Pliopotamys meadensis*, *Pliophenacomys*, *Pliolemmus*, *Canis lepophagus*, *Borophagus diversidens*, *Trigonicitis cookii*, *Stegomastodon*, *Gigantocamelus spatula*, *Nannippus peninsulatus*, and “*Equus simplicidens*.” External age control is lacking, but the Sand Draw Fauna often is cited in studies attempting to correlate Great Plains faunas. It includes what may be the youngest records of *Dipoides rexroadensis*, *Ogmodontomys poaphagus*, and *Pliopotamys*.

The Seneca Fauna in southern Nebraska is not published in its entirety, but the small mammals include *Planisorex dixonensis*, *Procastoroides idahoensis*, *Zapus sandersi*, *Ondatra idahoensis*, *Ophiomys parvus*, *Pliophenacomys osborni*, and *Mictomys* cf. *M. vetus* (L. Martin and Schultz 1985). External age control is lacking, but faunal correlations suggest placement in the late Blancan, sometime after 2.56 Ma and before about 2.0 Ma. The fauna may record the earliest occurrence of *O. idahoensis* (another possible early occurrence is in the White Rock Fauna; Eshelman 1975).

The Sappa Fauna (Harlan County, Nebraska) was collected from sediments approximately 2 m below the Mesa Falls Ash, dated to between 1.2 and 1.3 Ma (Naeser et al. 1971, 1973; L. Martin and Schultz 1985; Gansecki et al. 1998). A preliminary report on the fauna was published by C. Schultz and Martin (1970), with a more expansive discussion, especially of the small mammals, provided by

L. Martin and Schultz (1985). Their faunal list includes *Stegomastodon*, *Geomys* cf. *G. tobinensis*, *Ondatra* cf. *O. annectens*, *Allophaiomys*, and *Mictomys kansasensis*. This fauna may be early Irvingtonian but is here considered latest Blancan.

The Blanco Fauna, which originally formed the basis of the Blancan, is derived from deposits in the vicinity of Mount Blanco near the eastern edge of the Llano Estacado in Crosby County, Texas. Comprehensive studies of this fauna began with Gidley (1903a, 1903b). This work was followed by studies by Matthew (1925), Meade (1945), and Dalquest (1975). The last study significantly increased our knowledge of the small-bodied species in the fauna. The fauna includes *Glyptotherium texanum*, *Megalonyx leptostomus*, *Hypolagus* sp., *Prodipodomys centralis*, *Paenemarmota* (the youngest known record in North America), *Sigmodon minor* (originally identified as *Sigmodon medius*, but see Harrison 1978; R. Martin 1986), *Canis lepophagus*, *Borophagus diversidens*, *Rhynchotherium praecursor*, *Nannippus peninsulatus*, and *Platygonus bicalcaratus*. Early estimates of the age of the Blanco Fauna were variously given as Pliocene (Gidley 1903a; Matthew 1925; Osborn 1936) and early Pleistocene (Meade 1945; Evans and Meade 1945; Hibbard 1958). The age was better constrained by dates of two ash beds in the Mount Blanco Section. A volcanic ash bed (informally named the “Guaje ash bed” by Izett et al. 1972) is located in the overlying Blackwater Draw Formation, 9 m above the fossiliferous level of the Blanco Formation (G. Schultz 1990a). This ash bed was correlated with the Guaje pumice bed in the Jemez Mountains, New Mexico, on the basis of petrographic comparisons (Izett et al. 1972). Two fission-track dates on glass shards were reported for this ash:  $1.4 \pm 0.2$  Ma (Izett et al. 1972; Izett 1981) and  $1.77 \pm 0.44$  Ma (Boellstorff 1976). A second ash, the Blanco Ash, located approximately 7.5 m below the Guaje ash bed and thus stratigraphically closer to the fossiliferous Blanco beds, yielded a fission-track age (based on glass shards) of  $2.8 \pm 0.3$  Ma (Boellstorff 1976). No normally magnetized sediments were found in the Mount Blanco Section by Lindsay et al. (1975), but if the date of the Blanco Ash is correct, it would fall within a normal polarity interval, chron C2An.1n. On our correlation chart (figure 7.1) we plot the ash within a reversed-polarity interval (chron C2r.2r) based on the paleomagnetic analysis of Lindsay et al. (1975), but we also provide its published fission-track age.

The Cita Canyon faunas were derived from exposures at the head of Cita Canyon in Randall County, Texas. The first report of the fauna was by Johnston (1938). A subsequent analysis revealed characteristic Blancan taxa such as *Megalonyx* cf. *M. leptostomus*, *Glyptotherium* cf. *G. tex-*

*anum*, *Hypolagus* cf. *H. regalis*, *Borophagus*, *Canis lepophagus*, *Chasmaporthetes johnstoni*, *Platygonus bicalcaratus*, “*Equus simplicidens*,” and *Nannippus peninsulatus* (Johnston and Savage 1955; more extensive faunal lists were provided by G. Schultz 1977 and Kurtén and Anderson 1980). Two mammal-bearing stratigraphic units are separated by a sand that produced no mammalian fossils. Although stegomastodonts and glyptodonts are limited to the upper unit (Savage 1955), the faunas from the two units were listed together because many taxa are common to both. The lower fossiliferous sequence is normally magnetized and was referred to chron C2An.1n (the upper part of the Gauss chron), but the upper unit is reversed and was placed in chron C2r.2r (the lower Matuyama; Lindsay et al. 1975). Cita Canyon includes the earliest occurrence of *Miracinonyx inexpectatus*.

The Beck Ranch Fauna in Scurry County, Texas, includes a diverse assemblage of mammals (58 species; Dalquest 1978). It shares *Felis lacustris*, *Ogmodontomys poaphagus*, *Hypolagus regalis*, and *Nannippus* with the Rexroad faunas in Kansas and *Sigmodon minor* (originally identified as *Sigmodon medius*, but see Harrison 1978; R. Martin 1986), *Borophagus diversidens*, and *Platygonus bicalcaratus* with the Blanco Fauna (Dalquest 1978). Dalquest (1978) considered the Beck Ranch Fauna to be older than the Rexroad and Blanco faunas and younger than the Fox Canyon Fauna. A report of a rhinoceros tooth fragment from Beck Ranch (Madden and Dalquest 1990) remains to be verified.

The Donnelly Ranch Fauna in southeastern Colorado was recovered from normally magnetized sediments and includes *Sigmodon minor*, *Sigmodon curtisi*, *Paramylodon*, “*Equus (Dolichohippus) simplicidens*,” *Tapirus haysii* (originally identified as *Tapirus copei*, but see Ray and Sanders 1984), *Hemiauchenia blancoensis* (originally listed as *Tanupolama*, but see Webb 1974b), *Gigantocamelus*, and an unidentified gomphothere (Hager 1975). The fauna was correlated with the Wolf Ranch Fauna in the St. David Formation of Arizona. The age assessment of 2.5 Ma given by Hager (1975) was derived from paleomagnetic correlations; Hager considered the fauna to be derived from normally magnetized sediments in the Gauss chron (chron C2An.1n). New paleomagnetic age calibrations yield an age of slightly older than 2.58 Ma (Berggren et al. 1995).

The Java Fauna of South Dakota generally is considered early Irvingtonian (R. Martin 1989a; Repenning 1992). No external age control is available. The Java Fauna includes *Allophaiomys*, *Guildayomys*, *Hibbardomys marthae*, *Ondatra annectens*, *Pliophenacomys*, *Mimomys dakotaensis* (closely related to and possibly synonymous with *Mi-*

*momys virginianus*), *Mictomys kansasensis*, and several zaptodid rodents (R. Martin 1973, 1975, 1989a, 1989b). *Mammuthus* is absent. The Java Fauna contains a unique association of taxa, and recent reanalysis suggests that the fauna may be mixed (R. Martin, pers. comm., 2002). For now we consider the fauna to be latest Blancan, but it may be early Irvingtonian. It includes the earliest known occurrence of *Ondatra annectens* (another early record is in the Kentuck Fauna).

**Florida** A number of important Blancan faunas were traditionally recognized from localities in Florida (Santa Fe River 1, Haile 15A, Macasphalt Shell Pit, Kissimmee River, and Haile 7C; Morgan and Ridgway 1987; Morgan and Hulbert 1995). Many additional faunas that are elsewhere considered Irvingtonian (Morgan and Hulbert 1995; Ruez 2001) are here considered to be latest Blancan (based on our adoption of *Mammuthus* as the defining taxon for the Irvingtonian). These faunas include Inglis 1A, Inglis 1C, De Soto Shell Pit, and Haile 16A. Low topographic relief results in a lack of exposed extensive stratigraphic sequences, and suitable material for radiometric dating is lacking (a possible exception is helium–uranium dating applied to corals that was used to help bracket the age of the Leisey Shell Pit Fauna; Bender 1973; Morgan and Hulbert 1995). External age control is possible for some faunas by use of the relationship of fossiliferous deposits to eustatic sea level changes supplemented by paleomagnetic data and strontium isotopic data (Webb 1974a; Webb et al. 1978, 1989; Jones et al. 1991).

A diverse assemblage of species is known from these faunas (see summary in Morgan and Hulbert 1995). The characteristic taxa provided by Morgan and Hulbert for the faunas they considered to be Blancan includes *Dasybus bellus*, *Holmesina floridanus*, *Glyptotherium arizonae*, *Glossotherium chapadmalense*, *Megalonyx leptostomus*, *Ermootherium eomigrans*, *Trigonictis macrodon*, *Sigmodon minor* (originally reported as *Sigmodon medius*, but see Harrison 1978; R. Martin 1986), *Neochoerus dichroplax*, *Platygonus bicalcaratus*, *Hemiauchenia blancoensis*, and *Nannippus peninsulatus*. *Cormohipparion emsleyi* is an endemic Florida species known from the Macasphalt Shell Pit and Haile 15A faunas, the only known post-Hemphillian records of that hipparionine taxon (Hulbert 1987). The De Soto Shell Pit Fauna includes the youngest record of *Megalonyx leptostomus*, and *Glossotherium chapadmalense* makes its latest known appearance in the Macasphalt Shell Pit Fauna. Two Florida Blancan faunas (Haile 15A, Santa Fe River 1) contain associations of taxa such as *Smilodon gracilis* and *Nannippus peninsulatus* that are not known to overlap chronologically in the western

United States. The Santa Fe River 1 record of *S. gracilis* is the oldest in North America. It was identified as *Ischyromilus* by Churcher (1984b) and was included in *Megantereon* by L. Martin (1980, 1998); we follow Berta (1985, 1987) for its taxonomic placement.

*Nannippus* occurs in several Florida Blancan faunas (Haile 15A, Santa Fe River 1, Macasphalt Shell Pit, and Kissimmee River) along with South American immigrant taxa (*Holmesina floridanus*, *Dasyopus bellus*, and *Glossotherium chapadmalense*). The ages of these faunas are estimated to be between 2.5 and 2.0 Ma (Jones et al. 1991; Morgan and Hulbert 1995), and the youngest occurrence of *Nannippus* is in the Macasphalt Shell Pit at approximately 2.1 Ma. According to Tedford (1981), the last appearance of *Nannippus* and the first appearance of South American taxa in New Mexico were nearly synchronous at the Gauss–Matuyama (chrons 2An/2r) boundary (2.58 Ma; Berggren et al. 1995). *Nannippus* disappears from the 111 Ranch sequence just below that boundary, and South American species first appear 20 m lower in the section (Galusha et al. 1984). The concurrent range is entirely of normal geomagnetic polarity and probably corresponds to chron C2An.1n (Galusha et al. 1984).

“Earliest Irvingtonian” faunas of Morgan and Hulbert (1995) and Ruez (2001) were provisionally interpreted to range in age between approximately 2.0 and 1.6 Ma and are here considered to be latest Blancan. These faunas include the Inglis 1A, Inglis 1C, and De Soto Shell Pit localities; their temporal range is based on correlations with faunas outside Florida that have radiometric dates. They contain several characteristic Blancan taxa such as *Megalonix leptostomus*, *Trigonictis macrodon*, *Chasmaporthetes ossifragus*, *Capromeryx arizonensis*, and *Ondatra idahoensis*. Taxa such as *Smilodon gracilis*, *Platygonus vetus*, *Paramylodon harlani*, and *Canis edwardii* are found in latest Blancan faunas but are generally more characteristic of Irvingtonian faunas. The records of *Paramylodon harlani* and *Platygonus vetus* in Inglis 1A are the earliest known in North America. The Inglis 1C Fauna (Ruez 2001) appears to be intermediate in age between the older Inglis 1A and younger De Soto Shell Pit faunas. It contains the earliest Florida records of *Peromyscus polionotus*, *Reithrodontomys humulis*, and *Atopomys texensis*; late records of *Reithrodontomys wetmorei* and *Ondatra idahoensis* in Florida are also recorded in the fauna (Ruez 2001). The fauna also contains the youngest North American record of *Platygonus bicalcaratus*. *Dasyopus bellus*, *Holmesina floridanus*, *Paramylodon harlani*, *Sylvilagus webbi*, *Sigmodon curtisi*, *Orthogeomys propineta*, *Smilodon gracilis*, *Palaeolama mirifica*, *Hemiauchenia macrocephala*, *Platygonus bicalcaratus*, and *Tapirus haysii* are also pres-

ent. Based on biochronologic data, Ruez (2001) gave a likely chronologic span of 2.01–1.78 Ma, with an age of approximately 1.9 being most likely.

The Haile 16A Fauna is allied with the other latest Blancan faunas based on the shared presence of *Sylvilagus webbi*, *Orthogeomys propineta*, and *Atopomys texensis*, all of which are confined to the Blancan in Florida. Our removal of Haile 16A from the group of “late early Irvingtonian” faunas proposed by Morgan and Hulbert (1995) is consistent with the opinion of Morgan and White (1995), based on examination of the microfauna. The fauna records the earliest known occurrences of *Megalonix wheatleyi* and *Synaptomys*.

### CHRONOLOGIC ASPECTS OF THE GREAT AMERICAN BIOTIC INTERCHANGE

As noted earlier, a number of southern United States localities produce assemblages of several genera of late Blancan land mammal immigrants from source areas in South America. These southern species, their affiliations with sister groups in South America, and the chronologic evidence regarding the reciprocal cohort that moved southward across the Panamanian land bridge were treated by Marshall et al. (1979), Webb (1985, 1991), and Webb and Rancy (1996). The absence of radiometric dates from Florida sites leads to some loose correlations of key sites, based largely on biostratigraphy and aided, in some cases, by paleomagnetic evidence. For that reason it is fortunate that several of the late Blancan Neotropical taxa extend geographically beyond Florida, appearing concurrently in western sites, thus supporting and refining the biostratigraphic control on the Florida record. These immigrants and their earliest known North American records include *Glossotherium* in the Lower Cita Canyon (G. Schultz 1977) and Blanco (Dalquest 1975) faunas of Texas, the 111 Ranch Fauna in Arizona (Galusha et al. 1984), the Donnelly Ranch Fauna of Colorado (Hager 1975), and the Haile 15A and Santa Fe River 1 faunas of Florida (Webb 1974a; Robertson 1976); *Glyptotherium* in Upper Cita Canyon (Johnston and Savage 1955), Blanco (Meade 1945), Hudspeth (Strain 1966), and Red Light (Akersten 1972) faunas of Texas, the 111 Ranch Fauna in Arizona (Gillette and Ray 1981 as “Tusker Fauna,” but see Galusha et al. 1984), and Santa Fe River 1 in Florida (Gillette and Ray 1981); *Holmesina floridanus* (originally described as *Kraglievichia* but transferred to *Holmesina* by Edmund 1987) and *Dasyopus* in Haile 15A and Santa Fe River 1B faunas in Florida (Webb 1974a; Robertson 1976); *Nechoerus* at 111 Ranch, Arizona (Lindsay and Tessman 1974; Ahearn and Lance 1980; Galusha et al. 1984); and



*Erethizon* in the Grand View Fauna in Idaho (Wilson 1935), the Haile 7C and Inglis 1A faunas in Florida (Frazier 1982; Morgan and Hulbert 1995), the Wolf Ranch Fauna in Arizona (Harrison 1978; Frazier 1982), and the San Timoteo Badlands of California (Albright 1999). The records listed here suggest that these immigrants were in southwestern faunas by the end of chron C2An (Gauss) at approximately 2.5 Ma (Berggren et al. 1995).

It is noteworthy that late Blancan Neotropical land mammals of North America provide the best available datum establishing the existence of a functional Panamanian land bridge in the Pliocene. Studies of tectonism and volcanism in the isthmian region (Coates and Obando 1996) provide local observations and cannot demonstrate continuous terrestrial terrain on a convincing scale. On the other hand, although marine micropaleontology and molluscan studies indicate decreasing water depth in Pacific and Atlantic sediments during the Miocene and Pliocene (Cronin and Dowsett 1996), they approach but cannot record the final establishment of a land bridge that would permit the passage of terrestrial mammals. Subaerial exposure of the isthmus may have occurred sporadically between approximately 3.1 and 2.0 Ma (Cronin and Dowsett 1996; Webb and Rancy 1996). For that reason, the marine studies give older estimates, usually between 3 and 4 Ma, than do the mammal data. The main thrust of the Pliocene migration of North American taxa into South America occurred by the early Uquian South American mammal age. South American faunas containing the immigrants closely postdate the Gauss–Matuyama (chron C2An/2r) boundary (Webb and Rancy 1996) at 2.58 Ma (Berggren et al. 1995).

## IRVINGTONIAN

### HISTORICAL CONTEXT

The Irvingtonian mammal age was originally defined by Savage (1951) based on a fauna recovered from gravel pits southeast of Irvington, Alameda County, California (Stirton 1939; Savage 1951; Firby 1968). These gravel pits now are abandoned, and portions are overlain by Interstate Highway 680. Strata adjacent to the pits and at the level where the fossils were recovered are reversely magnetized, and the fauna was referred to the upper part of the Matuyama chron below the Jaramillo subchron by Lindsay et al. (1975; the placement would now presumably be in chron C1r.2r). The primary criteria proposed by Savage for recognition of the Irvingtonian were the

absence of *Bison* and the presence of mammalian species less advanced than related forms from the Rancholabrean and the Holocene. Savage used the appearance of *Bison*, among other criteria, to define the subsequent Rancholabrean. This usage effectively meant that two consecutive mammal ages were defined by the presence or absence of a single taxon, *Bison*. This situation is further complicated by the difficulty in establishing an adequate chronology for the arrival of *Bison* in North America (see “Rancholabrean” later in this chapter).

### HISTORICAL DEFINITION AND CHARACTERIZATION

The ambiguity of the original definition of the Irvingtonian (“the Irvingtonian is marked by the absence of *Bison*”; Savage 1951:289) led to unstable and inconsistent approaches to defining the Irvingtonian. Several authors proposed alternative definitions based on either the composite fauna from the type Irvington locality (Firby 1968) or on one or more members of the fauna (e.g., *Mammuthus*; Kurtén and Anderson 1980). No explicit definition was recommended by Lundelius et al. (1987), but they provided a discussion of advantages and disadvantages of using alternative taxa.

### CURRENT DEFINITION AND CHARACTERIZATION

**Definition** In our committee deliberations we failed to reach unanimous consensus on the definition of the Irvingtonian. The majority decision was that the Irvingtonian is best defined by the first appearance of *Mammuthus* in North America south of 55°N latitude. This definition previously was not proposed explicitly but is in keeping with a widespread use that emerged among Pleistocene mammalian paleontologists over the last 20 years. It is also in keeping with the historical context of the Irvingtonian; the presence of *Mammuthus* was considered by Savage (1951:236) to “offer some of the best evidence for assignment of a Pleistocene age to Irvington,” but he did not include *Mammuthus* in his discussion of the initial proposal for the Irvingtonian mammal age (Savage 1951:289).

We recognize the benefits of single-taxon boundary definitions (Woodburne 1977, 1987) but also acknowledge the complications that can arise from such definitions. The use of *Mammuthus* to define the Blancan–Irvingtonian boundary certainly suffers from deficiencies, but advantages of using *Mammuthus* to define the Irvingtonian include the facts that it was widespread in North America, is easy to identify, and is difficult to miss (because of its

large size) in faunas where it is present. However, although *Mammuthus* was widespread in North America in the Pleistocene it often is absent from faunas that are known or taken to be Irvingtonian based on radiometric dating or biochronologic correlation. Important examples include many cave deposits (where *Mammuthus* probably is absent for taphonomic reasons) and the San Pedro Valley (Arizona) and Meade County (Kansas) sequences, portions of which generally are interpreted to be Irvingtonian. In these cases assignment of faunas to the Irvingtonian must rely on characterizing taxa, especially those that are limited to the Irvingtonian.

With the adoption of *Mammuthus* to define the base of the Irvingtonian, many faunas elsewhere considered to be early Irvingtonian (based on alternative definitions) are here considered to be latest Blancan. These include the Inglis 1A, Inglis 1C, Haile 16A, and De Soto Shell Pit faunas in Florida; the Curtis Ranch Fauna in the San Pedro Valley sequence in Arizona; the Nash, Aries A, Rick Forester, and Aries B faunas in the Meade Basin of Kansas; the Sappa Fauna in Nebraska; the Java Fauna in South Dakota; the Froman Ferry faunal sequence in Idaho; the El Casco Fauna in California; and the lower portion of the Cedazo faunal sequence in Mexico. Not only do these faunas lack *Mammuthus*, but their known (or inferred) temporal ranges predate the established arrival of *Mammuthus* elsewhere (accepted herein as 1.35 Ma). The informal designation *latest Blancan* can be used to refer collectively to the faunas affected by the new definition. Because of uncertainties in their correlation, the Haile 16A, Rick Forester, Aries B, Sappa, and El Casco faunas may be either late Blancan or earliest Irvingtonian (they may or may not predate the earliest arrival of *Mammuthus*).

**Characterization** The characterization of the Irvingtonian includes a few taxa that are limited to the Irvingtonian, many that appear in earlier mammal ages but are common in Irvingtonian faunas, and many that make their first appearance in the Irvingtonian but persist into the younger Rancholabrean. These are listed separately later in this chapter, followed by comments justifying our treatment of certain taxa and relevant faunas.

Taxa limited to the Irvingtonian include *Microtus llanensis*, *Microtus meadensis*, *Microtus paroperarius*, *Canis armbrusteri*, and *Tetrameryx irvingtonensis*.

Taxa that make their first appearance in the Blancan (or earlier) but persist into the Irvingtonian include *Glyptotherium*, *Blarina*, *Sylvilagus*, *Miracinonyx inexpectatus*, *Smilodon gracilis*, *Arctodus pristinus*, *Ursus*, and *Mammut americanum*. Taxa that first appear in latest Blancan fau-

nas (those that are elsewhere often considered to be early Irvingtonian) include *Nothrotheriops*, *Paramylodon harlani*, *Holmesina*, *Lepus*, *Allophaiomys pliocaenicus*, *Microtus* with five closed triangles on the lower first molar, *Microtus kansasensis/meltoni*, *Neofiber*, *Ondatra annectens*, *Phenacomys*, *Synaptomys*, *Sciurus*, *Canis edwardii*, *Lutra canadensis*, *Homotherium*, and *Platygonus vetus*.

Taxa that first appear in the Irvingtonian and persist into Rancholabrean or younger faunas include *Didelphis*, *Brachylagus idahoensis*, *Sylvilagus palustris*, *Clethrionomys*, *Lemmings curtatus*, *Ondatra zibethicus*, *Marmota flaviventris*, *Marmota monax*, *Cynomys gunnisoni*, *Cynomys ludovicianus*, *Panthera onca*, *Smilodon populator*, *Mustela erminea*, *Brachyprotoma*, *Conepatus*, *Canis latrans*, *Canis lupus*, *Arctodus simus*, *Euceratherium*, *Oreamnos*, and *Mammuthus*.

The *Canis* in the Cudahy Fauna in Kansas was described by Getz (1960) as "*Canis* sp."; the identification was changed to *Canis edwardii* by Nowak (1979). *Rangifer tarandus* was reported from the Cape Deceit Fauna in Alaska, here considered an Irvingtonian temporal equivalent (originally identified as "*Rangifer* sp." by Guthrie and Matthews 1971, but see McDonald et al. 1996). The earliest known records from south of 55°N latitude are in the Rancholabrean Medicine Hat and Carlton Bar faunas, both in southern Canada (J. McDonald et al. 1996). The first Irvingtonian record of *Oreamnos* was recently discovered in Porcupine Cave, Colorado (Mead and Taylor 1998; originally identified as "cf. *Oreamnos harlingtoni*," but the "cf." was dropped by Jass et al. 2000).

## THE ARRIVAL OF MAMMUTHUS

The oldest reported reliable date for *Mammuthus* is from the Leisey Shell Pit Fauna from the Bermont Formation in Florida, which includes the largest sample of early Pleistocene mammoths in North America (Webb and Dudley 1995). An evaluation of strontium isotope data, paleomagnetic data, and biostratigraphy (Webb et al. 1989; Hulbert et al. 1995) yielded an age range for the Bermont Formation of between 1.55 and 1.1 Ma; the major vertebrate-bearing units are considered to be less than 1.3 Ma (Morgan and Hulbert 1995:30), the age we accept for the *Mammuthus* material.

A series of vertebrate fossil localities in the vicinity of Tijeras Arroyo in Bernalillo County, New Mexico, include both Blancan and Irvingtonian faunas (Logan et al. 1984; Lucas et al. 1993; Morgan and Lucas 2000b). The geologic formation from which the faunas were recovered is uncertain (but was tentatively referred to the Sierra Ladrones Formation by Lucas et al. 1993). Reworked

pumice fragments deposited in strata containing Irvingtonian faunas were interpreted to be derived from the Guaje Pumice, the lower bed of the Otowi Member of the Bandelier Tuff, dated elsewhere to approximately 1.61 Ma (Izett and Obradovich 1994; Spell et al. 1996). The Irvingtonian component has a fauna including *Mammuthus*, *Glyptotherium arizonae*, and *Equus scotti* and dates to between 1.0 and 1.6 Ma (Morgan and Lucas 2000b). Three localities in Tijeras Arroyo were reported to contain *Mammuthus*; the oldest recorded occurrence from this area was interpreted to be “no older than 1.61 Ma” (Lucas et al. 1993:8), but a more precise age is not available.

Another early record of *Mammuthus* was reported from the Rock Creek Fauna in the Tule Formation, Briscoe County, Texas. *Mammuthus* remains are in close stratigraphic association with a volcanic ash identified as the Cerro Toledo X Ash, with an age of 1.2 to 1.3 Ma (Izett 1977; G. Schultz 1986). The stratigraphic section illustrated by G. Schultz (1986:83) plots *Mammuthus* below the Cerro Toledo X ash bed (the section was republished with minor modification by G. Schultz 1990b).

Until recently, there were no specimens of *Mammuthus* from the Vallecito Creek–Fish Creek sequence in the Anza-Borrego Desert (Cassiliano 1999), but molar fragments referred to *Mammuthus* are now reported from three localities in that sequence, with a maximum age of 1.2 Ma (McDaniel and Jefferson 1999). Additional material is known from the Borrego Badlands in the northern part of the Anza-Borrego Desert State Park. The oldest possible age for this material is between approximately 1.0 Ma and 1.25 Ma (Remeika and Jefferson 1993; Jefferson and Remeika 1994).

Another often-cited early occurrence of *Mammuthus* is from the Bruneau Formation in Idaho, where Malde and Powers (1962) reported *Mammuthus* tooth fragments from a gravel bed 3.2 km (2 miles) southeast of Glenn’s Ferry. Based on their map (Malde and Powers 1962, plate 1), these tooth fragments were recovered from an area of “sediments and interbedded basalts.” The age of the “Bruneau Basalt *Mammuthus*-bearing fauna” is traditionally given as 1.36 Ma, an age derived from a K–Ar date reported by Evernden et al. (1964); magnetic polarity data are unavailable for the deposit that yielded the *Mammuthus* (Neville et al. 1979:519). Additional work by Armstrong et al. (1975) yielded a variety of ages for Bruneau Basalt flows, with an average of approximately 1.4 Ma. The complex stratigraphic relationships of former stream channels and the lava flows that dammed them were discussed by Malde (1991). The dates on the lava flow dams range in age from 2.06 Ma to 0.78 Ma, making the history of deposition and the age of associ-

ated discrete mammalian fossil deposits difficult to assess (Repenning et al. 1995).

Controversial reports of *Mammuthus* from the Merced Formation at Thornton Beach, California, are more problematic (Madden 1980, 1995; Lucas 1995). The age, stratigraphic position, and taxonomic affinities of the Thornton Beach mammoth are disputed. The mammoth material from Thornton Beach apparently was collected about 10 m below a horizon dated at  $1.5 \pm 0.8$  Ma (Hall 1965; Lucas 1995; Madden [1995] stated that the fossil came from above the dated horizon, but this appears to be an error), but much younger age estimates for the Merced Formation ( $0.45 \pm 0.08$  Ma) were reported by Meyer et al. (1980) from localities to the north and east of Thornton Beach itself. Other reported early occurrences include a single tooth from New Bern, North Carolina, with unknown stratigraphic and age determinations (Madden 1985) and a record from Santa Fe River 1B in Florida that was reported to be in association with Blancan taxa (Lambert et al. 1995; according to Cassiliano [1999] the association may be the result of mixing of fossils of different ages).

The appearance of *Mammuthus* in North America therefore appears to have been essentially contemporaneous over much of what is now the United States and can be bracketed at approximately 1.3 to 1.4 Ma. The differences in age between the first appearances in various regions are small enough to be attributable to discontinuous deposition and the vagaries of association between good fossil sites and good dates. Slightly older ages are possible for the *Mammuthus* material from Leisey Shell Pits in Florida and the Tijeras Arroyo in New Mexico.

An additional early report of *Mammuthus* in the Wellsch Valley faunal sequence from southern Saskatchewan (Stalker and Churcher 1972, 1982; Churcher 1984a; Barendregt et al. 1991, 1998) may represent the earliest North American appearance. Detailed stratigraphic data for the vertebrate species from Wellsch Valley are not published yet, and the stratigraphic position of *Mammuthus* must be confirmed. The fossils were reported from reversely magnetized sediments attributed to the upper part of the Matuyama (chron C1r.1r) by Barendregt et al. (1991).

**A Need for a Small Mammal Definition?** A small mammal definition for the Irvingtonian is also possible (and perhaps advisable) but is not endorsed here. If such a definition were adopted, the first appearances of *Allophaiomys* and *Microtus* (*sensu* Repenning 1992) seem to be the best choices for defining taxa; both are listed earlier in our characterization of the Irvingtonian. The ear-

liest purported *Allophaiomys* associated with external age control is from Hansen Bluff (Colorado) and was recovered from a core hole. The specimens in question are not definitively diagnostic; referral to *Allophaiomys* was based on the general morphology of nondiagnostic molars and stratigraphic position near chron C2n (the Olduvai), dated to approximately 1.9 Ma (Rogers et al. 1992). The Nash Fauna in Kansas may be slightly older (R. Martin et al. 2000), and the specimens permit a more reliable identification. We now consider both records to be latest Blancan. Other early records include Java, South Dakota (R. Martin 1975, 1989a), and Wellsch Valley, Saskatchewan (Churcher 1984a; Barendregt et al. 1991; Repenning 1992). *Allophaiomys* is fairly common in faunas from the Great Plains but is known from only three localities in or west of the Rocky Mountains: Porcupine Cave, Colorado (Bell and Barnosky 2000); Little Dell Dam, Utah (Gillette et al. 1999); and Cathedral Cave, Nevada (Bell 1995; Bell and Barnosky 2000). The earliest reported occurrence of *Microtus* (*sensu* Repenning 1992) is from the Anza-Borrego Desert (Zakrzewski 1972), where its earliest occurrence may be between approximately 1.4 and 1.6 Ma (Repenning 1992:59; Repenning et al. 1995). Additional very early *Microtus* material was reported from the El Casco Fauna in southern California (Albright 1999), with a reported age of between 1.3 and 1.4 Ma. *Microtus* persists into the modern fauna and includes a diverse assemblage of extant species (Hall 1981).

**Alternative Hypotheses** There are at least five operational (though not necessarily explicit) definitions of the Irvingtonian in the recent literature, many of which were specifically proposed to account for provinciality in restricted geographic areas. Because these various alternative methods for defining the Irvingtonian continue to be used, we provide a brief summary in the following sections.

Savage's initial reliance on the absence of *Bison* as a primary means of recognizing Irvingtonian faunas was itself provincial in nature and specifically addressed faunas in the San Francisco Bay area in California (to quote Savage, "*Bison* is not represented in the Irvington Fauna. In view of this fact it may seem rather strange that this genus should enter into a discussion of the age of the assemblage. I believe, however, that the dating of the Irvington and all other terrestrial Pleistocene vertebrate faunas in the Bay region must hinge on the presence or absence of this genus"; Savage 1951:277). This method of recognizing Irvingtonian faunas is still occasionally invoked in California (Dundas et al. 1996; Pajak et al. 1996:38), but it does not appear to be used extensively elsewhere.

The first appearances of one or more of the following taxa also are used sometimes to mark the beginning of the Irvingtonian: *Smilodon*, *Lepus*, *Soergelia*, and *Euceratherium* (Johnson et al. 1975; Lundelius et al. 1987; Cassiliano 1999). There are problems associated with each. Most of them (*Smilodon*, *Soergelia*, and *Euceratherium*) are of limited utility because they are not common in fossil faunas (Lundelius et al. 1987; Cassiliano 1999). *Lepus* recently was proposed as the best taxon for defining the Irvingtonian in the Vallecito Creek–Fish Creek sequence in the Anza-Borrego Desert (Cassiliano 1999), but it is difficult to distinguish from other leporid taxa (e.g., *Sylvilagus*; White 1984, 1991b). The earliest occurrence of *Smilodon* is in the Santa Fe River 1 Fauna in Florida, reported to be between 2.5 and 2.0 Ma by Morgan and Hulbert (1995). *Lepus* first appears in the Borchers Fauna at approximately 2.0 Ma. The oldest known *Soergelia* is from the Rock Creek Fauna, younger than 1.2 Ma. The earliest known occurrence of *Euceratherium* is in the Vallecito Creek–Fish Creek sequence in Anza-Borrego at approximately 1.1 Ma.

Three additional definitions were proposed by Repenning (1992; Repenning et al. 1995) and differ in philosophy from others in that they explicitly seek to recognize provincial changes in faunal compositions as the basis for definition. In Repenning's approach, these perceived provincial changes would result in diachronous boundaries for the Irvingtonian. The first appearance of *Allophaiomys* (*sensu* Repenning 1992) was proposed to define the Irvingtonian for faunas east of the Rocky Mountains (Repenning 1992; Fejfar and Repenning 1992; Repenning et al. 1995) at approximately 1.9 Ma (based on the core hole specimen from Hansen Bluff; Rogers et al. 1992; Repenning 1992); the Nash Fauna in Kansas may be somewhat older, and the presence of *Allophaiomys* there is based on definitively identifiable material (the Hansen Bluff core specimens are nondiagnostic teeth; see Rogers et al. 1992). The first appearance of *Phenacomys* was proposed to define the Irvingtonian for the portion of the northern United States west of the Rocky Mountains (Repenning 1992; Fejfar and Repenning 1992; Repenning et al. 1995). Its first appearance there is marked by the appearance of *P. gryci* in the Froman Ferry sequence in Idaho, at approximately 1.7 Ma (Repenning et al. [1995] gave the age as approximately 1.6 Ma, based on their use of 1.65 Ma as the end of the Olduvai event [chron C2n], now dated at 1.77; Berggren et al. 1995). That species is known from much earlier deposits in Fish Creek, Alaska (reported to be approximately 2.4 Ma by Repenning et al. 1987), and also from younger localities in Porcupine Cave, Colorado, and Cathedral Cave, Nevada (Bell and Barnosky

2000). The beginning of the Irvingtonian in the southern part of the United States west of the Rocky Mountains was recognized by Repenning (1992; Repenning et al. 1995) based on the first appearance of *Microtus* (*sensu* Repenning 1992) in the Anza-Borrego sequence at approximately 1.4 Ma to 1.6 Ma (Repenning 1992; Fejfar and Repenning 1992; Repenning et al. 1995; also see Pajak et al. 1996).

External age control for the three taxa in the various provinces indicates that there is as much as 0.5 Ma (1.9 Ma to 1.4 Ma) difference in their first appearance; therefore, based on these records, the Irvingtonian would not begin at the same time in every province, and provincial definition would be based on different taxa. Tentative faunal provinces were outlined by Fejfar and Repenning (1992; see map in Bell 2000) but must be critically reevaluated. Major additional complications with this framework include the determination of temporal and spatial boundaries for provinces and the establishment of rigorous criteria on which such determinations should be based. These proposals warrant additional study and evaluation and highlight an important arena for new research into the nature and scope of mammal ages in the context of faunal provinciality.

## TEMPORAL EXTENT

As defined here, the Irvingtonian begins with the earliest appearance of *Mammuthus* in North America south of 55°N latitude. The actual first appearance is almost undoubtedly earlier than our oldest reliably dated fossil, but as currently understood the early records of *Mammuthus* place the boundary at approximately 1.35 Ma. The end of the Irvingtonian is marked by the beginning of the subsequent Rancholabrean age (currently defined by the first appearance of *Bison*). Our understanding of the timing of arrival of *Bison* is inadequate; the end of the Irvingtonian may be as old as 210 ka or as young as 160 ka (discussed under "Rancholabrean" later in this chapter). The varied provincial definitions proposed by Repenning and his colleagues (based on *Microtus* at 1.4 Ma, *Phenacomys* at 1.7 Ma, and *Allophaiomys* at approximately 1.9 Ma; Repenning 1992:59; Repenning et al. 1995; *Phenacomys* age modified to conform to Berggren et al. 1995) provide slightly to considerably older ages for the beginning of the Irvingtonian.

## TEMPORAL DIVISIONS OF THE IRVINGTONIAN

**Sappan, Cudahyan, and Sheridanian** Various efforts to refine temporal resolution in the Irvingtonian have

been proposed (figure 7.3). Two divisions of the Irvingtonian, the Sappan (older) and the Sheridanian (younger), were proposed by C. Schultz et al. (1977, 1978) and were based primarily on faunas from the Great Plains. Faunas considered to be Sappan (C. Schultz et al. 1978; Lundelius et al. 1987) include the type Sappa Fauna in Harlan County, Nebraska, situated 2 m beneath the Mesa Falls Ash, dated to between 1.27 and 1.29 Ma (C. Schultz and Martin 1970; Naeser et al. 1971; Izett 1981; Izett and Wilcox 1982; L. Martin and Schultz 1985; Gansecki et al. 1998; this fauna lacks *Mammuthus* and may be more properly considered latest Blancan); the Nash and Aries faunas (the latter now Aries A, both here considered to be latest Blancan) in Meade County, Kansas; the Wathena Fauna in Doniphan County, Kansas (Van der Meulen 1978); the Kentucky Fauna in McPherson County, Kansas (Hibbard 1952; Semken 1966; R. Martin 1975), deposited in a channel fill cut into the Huckleberry Ridge Ash (Izett 1981); and the Java Fauna, Walworth County, South Dakota (R. Martin 1973, 1975, 1989a, 1989b). These faunas were grouped together originally because they included the earliest known records of the arvicolines *Mictomys* and *Allophaiomys* and because of a low diversity of arvicoline rodents (C. Schultz et al. 1978). That the faunas seem dominated by heteromyid and cricetine rodents was suggested by Lundelius et al. (1987), who noted that the subage could be characterized by *Ondatra annectens*, *Allophaiomys*, and *Mictomys kansasensis*. *Mammuthus* also was reported to appear during the Sappan (e.g., Gilliland Fauna in Texas), and *Stegomastodon* made its last appearance (C. Schultz et al. 1978). At least some of the faunas traditionally considered to be Irvingtonian (e.g., Sappa, Nash, Aries A) are here considered to be latest Blancan. The Sappan division therefore spans the latest Blancan and early Irvingtonian as they are used here.

The recognition that some Irvingtonian faunas were intermediate in age between the Sappan and the younger Sheridanian (C. Schultz et al. 1978) led Lundelius et al. (1987) to propose the term *Cudahyan* for these faunas. Cudahyan faunas in the Great Plains include Cudahy Ash Pit, Sunbright Ash Pit, Tobin, and Wilson Valley (all in Kansas) and the Vera, Gilliland, Woody Draw, Bull Draw, and Deadman's Creek faunas (all in Texas). These faunas are closely associated with the Lava Creek B Ash (formerly Pearlette Type O Ash), dated between 0.60 and 0.67 Ma (Izett and Honey 1995; Gansecki et al. 1998), and are characterized by the following arvicoline species: *Microtus paroperarius*, *Microtus meadensis* (= *Pitymys* or *Terricola* of some authors), *Microtus llanensis* (= *Pedomys*), *Mictomys meltoni*, *Ondatra annectens*, and *Phenacomys*. Cudahyan faunas also can be recognized outside the Great

Plains; examples include Conard Fissure, Arkansas (Brown 1908; Graham 1972); Cumberland Cave, Maryland (Gidley 1913, 1920a, 1920b; Gidley and Gazin 1933, 1938; Nicholas 1953; Van der Meulen 1978); Trout Cave No. 2, West Virginia (Pfaff 1990, 1991); and possibly Port Kennedy Cave, Pennsylvania (Wheatley 1871; Cope 1871; Daeschler et al. 1993; Hibbard 1955b). The Pit Fauna in Porcupine Cave, Colorado (Bell and Barnosky 2000); Cathedral Cave, Nevada (Bell 1995; Bell and Barnosky 2000); and some of the faunal components of Hansen Bluff, Colorado (Rogers et al. 1985, 1992), and SAM Cave, New Mexico (Rogers et al. 2000), may also be considered to be Cudahyan.

The Sheridanian subage (C. Schultz et al. 1977, 1978) primarily was based on the Hay Springs, Rushville, Gordon, and Angus faunas (Nebraska; C. Schultz et al. 1978). These faunas were characterized by *Ondatra nebrascensis* (often erroneously spelled “*nebrascensis*”; see type description in Hollister 1911; now considered a junior synonym of *Ondatra zibethicus*; see Stephens 1960; R. Martin 1996) rather than *Ondatra annectens*, *Microtus pennsylvanicus* rather than *Microtus paroperarius*, *Microtus ochrogaster* (= *Pedomys*) rather than *Microtus llanensis* (= *Pedomys*), and *Smilodon populator* (formerly *Smilodon fatalis*; see Berta 1985) rather than *Smilodon gracilis* (Lundelius et al. 1987). Although the Irvington Fauna was also included in the Sheridanian by C. Schultz et al. (1978), its preservation in reversely magnetized sediments of the upper Matuyama chron (Lindsay et al. 1975) argues for an older age than other Sheridanian faunas, all of which are found in the Brunhes chron (chron C1n). Other faunas considered by Lundelius et al. (1987) to be Sheridanian include Kanopolis (Hibbard et al. 1978), Rezabek (Hibbard 1943), Sandahl (Semken 1966), and Adams (G. Schultz 1969) from Kansas, Berends (Starrett 1956) from Oklahoma, and Slaton (Dalquest 1967) from Texas. The Kanopolis, Rezabek, and Slaton faunas contain *Neofiber leonardi* and were considered to be approximately the same age (Hibbard and Dalquest 1973; Hibbard et al. 1978; Lundelius et al. 1987). These faunas lack external age control, and their placement on figure 7.1 is tentative. The Kanopolis and Rezabek records of *Ondatra zibethicus* are among the earliest known occurrences.

**Divisions Based on Arvicoline Rodents** As discussed earlier in this chapter, Charles Repenning and Larry Martin gave alternative proposals for dividing the Irvingtonian on the basis of changes in taxonomic composition and stage of evolution in arvicoline rodents. However, unlike the Blancan, the two proposals for the Irvingtonian are more similar. Each recognized three temporal di-

visions based on the appearance of new immigrant taxa or changes in species of endemic North American lineages and used similar taxa to recognize temporal divisions, but they differed in their intended geographic scope. Martin’s proposal was centered on the biostratigraphy of Great Plains faunas and is not widely used outside that region. Repenning’s original proposal was intended to encompass most of North America and was centered on developing a North American system that corresponded to a similar European biochronology. Repenning’s initial proposal was subsequently modified extensively in light of new discoveries and interpretations (Repenning 1980, 1983, 1984, 1987, 1992, 1998, 2001; Repenning et al. 1990, 1995; Bell and Repenning 1999; Bell 2000). Because the taxa used in both schemes are so similar, many of the changes applied to the Repenning scheme have equal impact on the Martin proposal (although this has never been explicitly stated or explored in the literature). For that reason, we center our discussion on the Repenning chronology. The relationships of the two proposals are illustrated in figure 7.3.

Initially Repenning’s proposal (1984, 1987) recognized three divisions of the Irvingtonian, numbered consecutively from oldest to youngest: Irvingtonian I, and II, and “Rancholabrean I.” “Rancholabrean I” faunas were noted to predate the arrival of *Bison* in the United States (Repenning 1987). The formal adoption of *Bison* as the defining taxon for the Rancholabrean (established by Lundelius et al. 1987) led Repenning et al. (1990) to change “Rancholabrean I” to “Irvingtonian III.” Only the Irvingtonian I and Irvingtonian II are recognized here. To facilitate ready interpretation of the literature, we provide a complete review of the Repenning proposal in its full form, including brief discussion of the rejected Irvingtonian III. Temporal divisions of the Irvingtonian, like those of the Blancan, were based on purported immigration events, but not all taxa have a documented presence in Asia or North American Beringia.

#### IRVINGTONIAN I (APPROXIMATELY 1.9 MA OR 1.72 MA TO APPROXIMATELY 0.85 MA)

Three provincial definitions of the Irvingtonian I are currently in use, and the defining taxa and initiation time are different in all three provinces. In the southern United States west of the Rocky Mountains it is defined on the first appearance of a *Microtus* with five closed triangles on the lower first molar (*Microtus californicus* of Repenning 1987, but many North American species show an essentially identical morphology; see Bell and Barnosky 2000)

between 1.4 and 1.6 Ma (Repenning 1992; Repenning et al. 1995). In the northern United States, west of the Rocky Mountains, it is defined by the first appearance of *Phenacomys* at approximately 1.72 Ma (Repenning et al. 1995). The age given here is somewhat older than that of Repenning et al. (1995), whose age of 1.6 Ma was based on an age calibration of 1.65 Ma for the top of the Olduvai event (chron C2n), which we calibrate at 1.77 Ma (following Berggren et al. 1995). The earliest appearance of *Phenacomys* was said to be approximately 0.05 Ma younger than the top of the Olduvai, hence our age of 1.72. East of the Rocky Mountains it is defined by the first appearance of *Allophaiomys*, possibly as early as 1.9 Ma in Colorado (Rogers et al. 1992; Repenning et al. 1995), but the Nash Fauna in Kansas may be slightly older (R. Martin et al. 2000). The Irvingtonian I is characterized by *Mictomys kansasensis*, *Ondatra annectens*, and *Proneofiber guildayi*.

#### IRVINGTONIAN II (APPROXIMATELY 0.85 MA TO APPROXIMATELY 0.4 MA)

The Irvingtonian II was originally defined on the first appearance of *Microtus meadensis* (= *Pitymys* or *Terricola* of some authors) and the first *Clethrionomys*. *Lasiopodomys* (*sensu* Repenning 1992) was added as a defining taxon by Repenning et al. (1990). All of these taxa were considered by Repenning et al. (1990) to have entered North America as immigrants at approximately 850 ka. All of the purported early records of *Clethrionomys* that were originally cited to support its presence in the Irvingtonian II subsequently were shown to be misidentified specimens of *Ophiomys parvus* (southern California records; see Scott and Cox 1993) or *Phenacomys* (Hansen Bluff; see Bell and Barnosky 2000:124). The report of *Clethrionomys* from Cumberland Cave (Guilday 1971; reiterated by Repenning 1987) has not been verified since the recognition of *Mimomys virginianus* in Irvingtonian deposits in the eastern United States (Repenning and Grady 1988). The two taxa are superficially similar in that both have rooted molars with cementum in the reentrant angles, and the Cumberland Cave material may represent *Mimomys*. The recent report of *Clethrionomys* from reversely magnetized sediments in SAM Cave, New Mexico (Rogers et al. 2000), confirms its presence in the western United States before about 780 ka, but additional records are lacking.

*Lasiopodomys* was reported from several localities in the eastern and central United States including Hamilton Cave, West Virginia (Repenning and Grady 1988), Cumberland Cave, Maryland (Repenning 1992), and the

County Line Fauna in Illinois (B. Miller et al. 1994). Each of the three records from the western United States is based on a single isolated lower first molar; these are from the Anza-Borrego Desert (Repenning 1992), the Pit locality in Porcupine Cave, Colorado (Bell and Barnosky 2000), and Cathedral Cave, Nevada (Bell and Barnosky 2000). The paucity of material and the possibility that these specimens are actually population variants of *Microtus paroperarius* (see Repenning 1992; Bell and Barnosky 2000) reduces our confidence in the use of this taxon. *Lasiopodomys* was reported originally from Cape Deceit, Alaska (as *Microtus deceitensis*; Guthrie and Matthews 1971).

The complications with *Clethrionomys* and *Lasiopodomys* suggest that they are inadequate for defining the Irvingtonian II, and they are here considered as characterizing taxa of the Irvingtonian II. We therefore consider this interval to be defined by the first appearance of *Microtus meadensis* (= *Terricola* or *Pitymys* of others). The history of this species in North America was discussed at length by Repenning (1983, 1992). It is widespread throughout the central and western United States but appears never to have crossed the Mississippi River. The earliest records appear to be from the Hansen Bluff and Porcupine Cave faunas in Colorado (Rogers et al. 1985; Bell and Barnosky 2000), but an occurrence in the Anza-Borrego Desert in reversely magnetized sediments of unknown age (Repenning 1992) may predate the Colorado records. The Hansen Bluff material was assigned an age of approximately 820 ka (Repenning 1992:72); the older specimens from the Pit locality in Porcupine Cave are interpreted to be between 750 and 850 ka (Bell and Barnosky 2000).

Recognition of a *Mimomys*-like arvicoline in the North American Pleistocene came as early as 1972 (L. Martin 1972) with the description of *Loupomys monahani* (originally named *Mimomys monahani*; see Koenigswald and Martin 1984). *Loupomys* is known only from the type locality in the Mullen assemblage in Nebraska (L. Martin 1972). It has a unique enamel microstructure (Koenigswald and Martin 1984; R. Martin 1989a), and although its relationships are poorly understood, it seems best to consider it a late *Mimomys*-like immigrant into North America (Repenning and Grady 1988; R. Martin 1989a).

Another arvicoline lineage appears in North America during the Irvingtonian II and is presumed an immigrant from Asia. Members of this lineage are classified as *Mimomys* (Repenning and Grady 1988 placed them in the subgenus *Cromeromys*), and two species are currently named. *Mimomys virginianus* was described from Hamilton Cave in West Virginia (Repenning and Grady 1988)

and was tentatively identified in the Pit Fauna from Porcupine Cave and from Cathedral Cave in Nevada (Bell and Barnosky 2000). *Mimomys dakotaensis* was first described from the Java fauna in South Dakota (R. Martin 1989a) and was tentatively identified from Little Dell Dam in Utah (Gillette et al. 1999).

The Irvingtonian II is characterized by the presence of *Atopomys*, *Clethrionomys*, *Lasiopodomys*, *Lemmiscus curtatus*, *Mictomys kansasensis*, *Mictomys meltoni*, *Mimomys virginianus*, *Mimomys dakotaensis*, *Neofiber leonardi*, *Ondatra annectens*, *Pedomys llanensis*, *Phenacomys* sp., and *Synaptomys cooperi*.

### IRVINGTONIAN III (APPROXIMATELY 0.4-0.15 MA)

The Irvingtonian III (originally named Rancholabrean I but changed by Repenning et al. 1990) was originally defined by the first appearance of the sagebrush voles of the genus *Lemmiscus* and by three species of *Microtus*: *M. mexicanus*, *M. montanus*, and *M. pennsylvanicus* (Repenning 1987; Repenning et al. 1990). These taxa were thought to have entered North America between 400,000 and 450,000 years ago (Repenning 1987; Repenning et al. 1990). The recognition of *Lemmiscus* in faunas predating the Brunhes (chron C1n; 780 ka) (e.g., SAM Cave, New Mexico [Repenning 1992; Rogers et al. 2000], and the Pit Fauna in Porcupine Cave [Bell and Barnosky 2000]) indicates that this taxon first appears in the North American fossil record much earlier than previously thought (its immigrant status is unconfirmed). The purported Irvingtonian occurrence of *M. mexicanus* cannot be verified (Bell and Repenning 1999); the oldest material tentatively identified as such is from Papago Springs Cave, Arizona (Skinner 1942), with a maximum age of approximately 246,000 years (Czaplewski et al. 1999b), but these specimens were recently reevaluated, and species-level identification was not possible (Czaplewski et al. 1999a). The other two *Microtus* species in question share common dental morphologies with each other and with several other North American species and cannot be identified reliably from isolated dental elements (Bell and Repenning 1999; Bell and Barnosky 2000). The traditional Irvingtonian III interval is thus undefined and was rejected by Bell and Repenning (1999) as a meaningful and distinct temporal interval. That interpretation is followed here.

Arvicoline rodent biochronology deteriorates by the late Irvingtonian, and effective temporal resolution declines. The arvicoline fauna is taxonomically modern by the end of the Irvingtonian, and an undifferentiated Rancholabrean was included in the schemes of both L.

Martin (1979) and Repenning (1987; Repenning et al. 1990). The appearance of *Dicrostonyx* and *Lemmus* outside the Beringian region was used by Repenning (1987) as a means of recognizing Rancholabrean faunas on the basis of arvicolines, but these taxa never penetrate deeply into southern latitudes (Foley and Raue 1987; E. Mead and Mead 1989; Eger 1995; R. Slaughter and Jones 2000). Their utility therefore is quite limited.

We accept the Irvingtonian I as discussed earlier and accept an extended Irvingtonian II that extends temporally from approximately 0.85 Ma to what we consider the latest occurrence of the characteristic Irvingtonian arvicolines *Microtus paroperarius* and *Microtus meadensis*, both of which last appear in the Salamander Cave Fauna at approximately 252 ka (Mead et al. 1996; an additional reported Rancholabrean record of *M. meadensis* in the Kennewick sequence in Washington [Rensberger et al. 1984; Rensberger and Barnosky 1993] is here considered Irvingtonian).

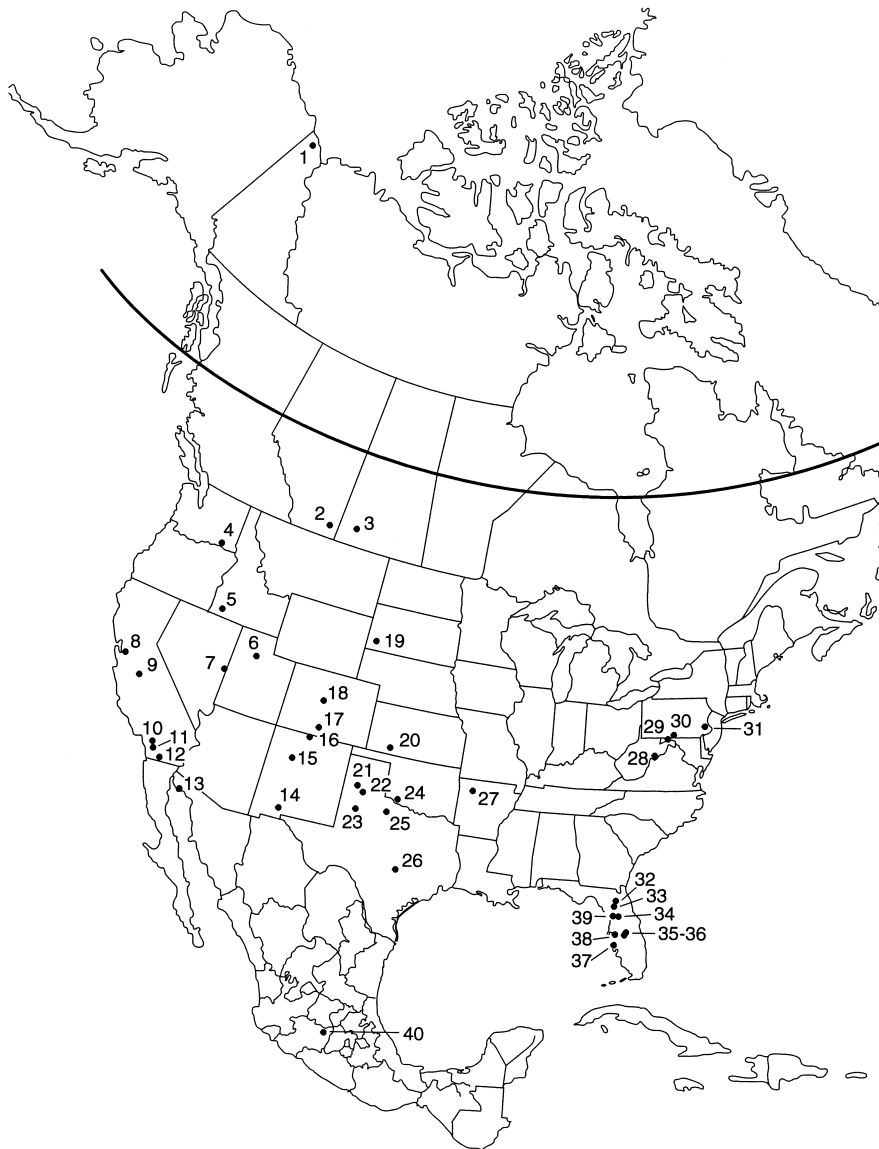
### GEOGRAPHIC DISTRIBUTION

Irvingtonian faunas are distributed widely across much of North America (figure 7.5). In contrast to Blancan faunas, many important Irvingtonian faunas were recovered from cave and fissure fill deposits. These deposits are in many respects mixed blessings: Although providing valuable information, they are also subject to complex stratigraphy, and it is often difficult to obtain reliable external age controls and paleomagnetic samples from friable cave sediments. Many Irvingtonian cave faunas lack reliable external age control, and their age estimates rely on varying interpretations of biochronology.

**Snake River Plain** Apart from the Bruneau Basalt *Mammuthus*-bearing locality discussed earlier in this chapter (see "Current Definition and Characterization"), there are no significant Irvingtonian faunal sequences in the Snake River Plain region. The sequence near Froman Ferry in Idaho was reported to be earliest Irvingtonian by Repenning et al. (1995). Their assessment was based on Repenning's arvicoline divisions of the mammal ages, in which the earliest appearance of *Phenacomys* defined the lower boundary of the Irvingtonian in the northwestern United States. The Froman Ferry sequence is here considered to be latest Blancan and is discussed earlier under "Blancan."

**California** The most extensive stratified sequence of Pliocene and Pleistocene sediments in North America is





**FIGURE 7.5** Irvingtonian localities and their temporal equivalents discussed in the text. The bold line represents 55°N latitude. Independent biochronologies should be established for the region north of this line, and the term *Irvingtonian* should not be applied to faunas there. Localities with external chronologic control indicating an age between approximately 1.35 and 0.21 Ma are considered to be temporal equivalents of Irvingtonian faunas elsewhere in North America. 1, Old Crow Basin, Yukon, Canada; 2, Medicine Hat sequence (in part), Alberta, Canada; 3, Wellsch Valley (in part), Saskatchewan, Canada; 4, Kennewick sequence (in part), WA; 5, Bruneau Formation, ID; 6, Little Dell Dam, UT; 7, Cathedral Cave, NV; 8, Irvington, CA; 9, Fairmead Landfill, CA; 10, San Timoteo Badlands (in part), CA; 11, Elsinore Fault Zone, CA; 12, Anza-Borrego Desert (in part), CA; 13, El Golfo, Sonora, Mexico; 14, Mesilla Basin, NM; 15, Tijeras Arroyo, NM; 16, SAM Cave, NM; 17, Hansen Bluff, CO; 18, Porcupine Cave, CO; 19, Salamander Cave, SD; 20, Meade County faunas (in part), KS; 21, Woody Draw and Bull Draw, TX; 22, Rock Creek, TX; 23, Slaton, TX; 24, Holloman, OK; 25, Vera and Gilliland, TX; 26, Fyllan Cave, TX; 27, Conard Fissure, AR; 28, Hamilton Cave and Trout Cave No. 2, WV; 29, Cumberland Cave, MD; 30, Hanover Quarry No. 1, PA; 31, Port Kennedy Cave, PA; 32, Haile 21A, FL; 33, McLeod, FL; 34, Coleman 2A; 35, Pool Branch, FL; 36, Payne Creek Mine, FL; 37, Rigby Shell Pit, FL; 38, Leisey Shell Pit, FL; 39, Crystal River, FL; 40, La Goleta (in part), Michoacan, Mexico.

beautifully preserved in the Anza-Borrego Desert State Park in southern California. Despite a long, apparently continuous and closely sampled sequence demonstrating that a characteristic Irvingtonian fauna overlies a Blancan fauna, the Blancan–Irvingtonian boundary in this

area is still uncertain. This problem was discussed in detail by Cassiliano (1999), who pointed out that this uncertainty is the result of the lack of a defining taxon for the boundary. Some of the taxa that are used to characterize the Irvingtonian, and some that are proposed as

taxa to define the boundary, have their lowest known stratigraphic appearance ( $LSD_k$  of Walsh 1998) in the Vallecito Creek–Fish Creek section at different places. The inferred times associated with these appearances are summarized here: *Navahoceros* early in the reversed chron (chron C2r.1r) below the Olduvai subchron at approximately 2.05 Ma, *Equus* (*Equus*) early in the reversed chron (chron C2r.1r) below the Olduvai event at approximately 2.09 Ma (this may be the earliest appearance in North America), *Lepus* at the base of the Olduvai (chron C2n) at approximately 1.95 Ma, *Smilodon* at the top of Olduvai at 1.77 Ma, and *Euceratherium* at approximately 1.13 Ma. At the time Cassiliano completed his report, mammoths were not present in the section, but molar fragments referred to *Mammuthus* are now reported from three localities in the sequence, with a maximum age of about 1.2 Ma (McDaniel and Jefferson 1999). Additional material from the Borrego badlands may be as old as 1.25 Ma (Remeika and Jefferson 1993; Jefferson and Remeika 1994).

Two faunas in the San Timoteo Formation were assigned to the Irvingtonian by Albright (1999). One of these, the El Casco Fauna, contains "*Plesippus francescana*" and lacks *Mammuthus*; it is here considered to be latest Blancan in age. The Shutt Ranch Fauna is stratigraphically much higher than the El Casco Fauna. It contains *Sorex*, *Microtus californicus*, and *Neotoma* close to *N. fuscipes* and was recovered from sediments that appear to be reversely magnetized. Albright interpreted this, along with the faunal data, to indicate placement in chron C1r.1r below the Brunhes. The only record of *Mammuthus* in this area is one tooth that is not well placed stratigraphically but is above the top of one transect that ends in chron C1r.1r (0.78–0.99 Ma) and is therefore below the Brunhes.

The Irvington Fauna is the type fauna for the Irvingtonian mammal age. A preliminary report of the fauna was given by Stirton (1939) and a more elaborate treatment by Savage (1951). Surprisingly, portions of this fauna have yet to be formally published (see Firby 1968). The original collection localities are now beneath Highway 680 and are inaccessible. Sediments adjacent to the original localities and at the stratigraphic level where fossils were originally recovered are reversely magnetized, and the Irvington Fauna was interpreted to be in the upper part of the Matuyama chron (chron C1r) by Lindsay et al. (1975). The fauna includes *Megalonyx*, *Mammuthus*, *Equus*, *Camelops*, *Hemiauchenia* (originally listed as *Tanupolama*), *Tetrameryx irvingtonensis*, *Euceratherium*, and *Microtus* with five closed triangles on the lower first molar (Savage 1951). *Mammut americanum*, *Arctodus*, *Capromeryx*, *Sorex*, *Scapanus*, *Paramylodon harlani*,

*Taxidea taxus*, *Panthera*, *Smilodon*, *Homotherium serum*, and an otariid seal were identified by Firby (1968) in an unpublished master's thesis.

An extensive and taxonomically diverse Irvingtonian fauna from near Fairmead in Madera County, California, provides an important complement to the type Irvington Fauna (Dundas et al. 1996). The Fairmead Landfill locality produced thousands of vertebrate fossils recovered from normally magnetized sediments tentatively correlated with the upper Turlock Lake Formation (Dundas et al. 1996). The fauna includes *Paramylodon harlani*, *Nothrotheriops*, *Megalonyx*, *Canis armbrusteri*, *Smilodon*, *Homotherium*, *Mammuthus columbi*, *Camelops*, *Hemiauchenia*, *Tetrameryx irvingtonensis*, and *Capromeryx*.

A sedimentary sequence from an unnamed sandstone in the Elsinore Fault Zone in Riverside County, California, produced many vertebrate fossil localities that included Blancan and Irvingtonian faunas (Reynolds et al. 1991; Scott and Cox 1993; Pajak et al. 1996). The Irvingtonian faunas were identified by the presence of *Mammuthus* and *Microtus* and included *Scapanus*, *Equus bautistensis*, *Arctodus simus*, *Camelops*, and *Hemiauchenia*. Several earlier reports (Reynolds et al. 1990, 1991; Repenning 1987) of *Clethrionomys* in the Elsinore Fault Zone were in error; the specimens are *Ophiomys parvus* (see discussion in Pajak et al. 1996). The Elsinore Fault Zone faunas were recovered during salvage operations to mitigate impact of construction activities on fossil resources; the deposits from which the fossils were collected are gone.

**Great Basin** The Topaz Lake Fauna from Douglas County, Nevada, contains few taxa but was considered an Irvingtonian fauna based on the presence of *Euceratherium* (Kelly 1997). The fauna also includes *Ondatra* cf. *O. idahoensis* and was discussed earlier with the Blancan Wellington Hills Fauna. The only known Great Basin Irvingtonian assemblage is from Cathedral Cave in White Pine County, Nevada (Bell 1995, 2000). A complete description of the fauna is not published, but the arvicoline rodent fauna is almost identical to that from the Pit Fauna in Porcupine Cave, Colorado, and includes *Mimomys* cf. *M. virginianus*, *Phenacomys gryci*, *Phenacomys* sp. (not *P. gryci*), *Mictomys meltoni* or *Mictomys kansanensis* (an earlier identification of *Mictomys borealis* was in error; Mead et al. 1992; Bell 1995), *Allophaiomys*, *Lemmiscus curtatus* (with both four and five closed triangles on the first lower molar), *Microtus paroperarius*, *Microtus meadensis*, and complex *Microtus* first lower molars with five closed triangles (Bell 1995; Bell and Barnosky 2000).

**Rocky Mountain Region** One of the most significant newly discovered Irvingtonian localities is Porcupine Cave in Park County, Colorado. The cave is situated in a large intermontane basin at 2900 m elevation. At least 26 discrete fossil localities are known in the cave (Barnosky and Bell in press), and a fantastically diverse vertebrate fauna is now recorded by tens of thousands of specimens recovered from the cave. A preliminary faunal list was provided by Barnosky and Rasmussen (1988), and reports of the arvicoline rodents and carnivorans from various localities are published in some detail (Barnosky and Rasmussen 1988; Anderson 1996; Barnosky et al. 1996; Bell and Barnosky 2000). At present, external age control is published only for the Pit locality, where paleomagnetic data indicate that the Pit sequence straddles the Brunhes–Matuyama boundary (Bell and Barnosky 2000). A unique assemblage of taxa was reported from the Pit locality (Barnosky and Rasmussen 1988; Anderson 1996; Bell and Barnosky 2000), and this is especially true for the arvicoline rodents. At least 10 arvicoline species were found in the Pit: *Phenacomys gryci*, *Phenacomys* sp. (not *Phenacomys gryci*) *Mimomys* cf. *M. virginianus*, *Ondatra*, *Mictomys* cf. *M. meltoni*, *Allophaiomys pliocaenicus*, *Microtus paroperarius*, *Microtus meadensis*, *Microtus* sp. (not *M. paroperarius* or *M. meadensis*), and *Lemmings curtatus* (with both four- and five-closed-triangle forms). Nine of these taxa were found in direct association in a single stratigraphic level of that deposit (the only one absent from level 4 was *Phenacomys* sp.). There is no evidence that the deposit is mixed, but the association of these taxa is unique. Attempts to reconcile paleomagnetic and biochronologic data resulted in an age estimate of 750–850 ka for the middle section of the Pit (levels 4–8; Bell and Barnosky 2000). The fauna includes the youngest known occurrence of *Allophaiomys pliocaenicus* and the earliest known records of *Microtus meadensis* (another early record is from the Hansen Bluff sequence) and *Lemmings curtatus* (an additional early record is from SAM Cave, New Mexico). The high elevation of the site and the species assemblage from the Pit suggest the possibility that high-elevation sites may warrant independent biochronologies. This hypothesis cannot be tested until additional high-elevation sites are discovered and studied.

The Hansen Bluff sequence includes a diverse faunal assemblage recovered from the Alamosa Formation in the San Luis Valley, south-central Colorado. The valley sits at 2300 m, and fossils were recovered primarily from five sections along a 4-km stretch of the bluff (Rogers et al. 1985). External chronologic control was established primarily by paleomagnetic data and the presence of the

Bishop Ash in two of the sections (the average age of the Bishop Ash is now calculated at  $758.9 \pm 1.8$  ka; Sarna-Wojcicki et al. 2000). Paleomagnetic samples were analyzed from Section A and from isolated portions of other sections. All sediments above unit A-14c have normal polarity and were interpreted to represent the Brunhes (chron C1n); those from A-14c and below have reversed polarity and represent the Matuyama (chron C1r) (Rogers et al. 1985). The Bishop Ash is present in sections C-9 and E-9 at Hansen Bluff. No mammalian fossils were found in association with the Bishop Ash. Mammals recovered from reversed sediments include *Spermophilus*, *Geomys*, *Reithrodontomys*, *Mictomys meltoni*, *Microtus meadensis*, *Microtus paroperarius*, *Ondatra annectens*, and *Equus* cf. *E. scotti*. The record of *M. meadensis* is among the earliest known occurrences of the species.

A 127-m core was drilled beneath section D at Hansen Bluff in 1986 (Rogers et al. 1992). The Huckleberry Ridge Ash (2.0–2.1 Ma) was intersected at 78.3 m depth in the core. Paleomagnetic data indicate that the sediments in the core preserve the Jaramillo (chron C1r.1n), Olduvai (chron C2n), and Reunion (chron C2r.1n) normal polarity events and the top of the Gauss chron (chron C2An.1n). Mammalian fossils were scarce in the core samples, but the recovery of *Spermophilus*, *Reithrodontomys*, and two arvicoline rodent teeth was reported (Repenning 1992; Rogers et al. 1992). The arvicoline teeth were assigned to *Allophaiomys* (Rogers et al. 1992; neither tooth is diagnostic of *Allophaiomys*, and the referral to that taxon was based on the ever-growing condition and the overall dental pattern; see Repenning 1992; Rogers et al. 1992). The oldest of the two arvicoline specimens from the core was recovered from a depth of 73.1 m, just above the base of the Olduvai event (chron C2n; Rogers et al. 1992).

SAM Cave is a lava tube cave located at 2737 m elevation in Rio Arriba County, New Mexico. Six discrete faunas were reported from the cave by Rogers et al. (2000). The four oldest of these localities have no external age control but contain *Allophaiomys*, *Lemmings curtatus*, *Microtus* (with five closed triangles on m1), *Mictomys kansasensis*, *Cynomys*, and *Spermophilus tridecemlineatus*. The “LB” locality, interpreted to be somewhat younger, was excavated following natural stratigraphy and preserves the Brunhes–Matuyama boundary. Biochronologic interpretation of the basal portion of the sequence yielded an age estimate of 820–840 ka. Based on biochronology and paleomagnetism, the middle section of the sequence (the lowest part with normal polarity) was interpreted as dating between 780 and 740 ka (Rogers et al. 2000). *Clethrionomys* was recovered from both above and below the Brunhes–Matuyama (chron C1n–1r) boundary; this

record represents the only confirmed Irvingtonian occurrence of *Clethrionomys*. *Mictomys kansasensis*, *Microtus*, and *Lemmiscus* are found only in the reversed sediments, but sample sizes appear to be small (the upper levels contained no mammalian fossils).

Two small Irvingtonian faunas were recently reported from the Little Dell Dam site in Salt Lake County, Utah (Gillette et al. 1999). Locality 1 contained *Peromyscus*, *Lemmiscus curtatus*, *Microtus paroperarius*, and an unidentified species of *Equus*. Locality 2 included *Mimomys* cf. *M. dakotaensis* (a *Mimomys* with roots and cementum in the reentrant angles, elsewhere known only from the Java Fauna in South Dakota; R. Martin 1989a), *Allophaiomys*, *Mictomys meltoni* or *Mictomys kansasensis*, and *Phenacomys* cf. *P. gryci*. The Little Dell faunas lack *Mammuthus* but are considered to be Irvingtonian based on the presence of *Microtus paroperarius*, a species limited to the Irvingtonian. The recovery of *Mimomys*, *Microtus paroperarius*, and *Allophaiomys* at Little Dell and Cathedral Cave in Nevada provides the first evidence of these taxa west of the Rocky Mountains in the contiguous United States. Similar *Mimomys* material is known from the Beringian region at the Old Crow Basin in the Yukon and Cape Deceit, Alaska, and at several localities east of the Rocky Mountains (Repenning, pers. comm., 1999).

**Southwestern United States** The Mesilla Basin in south-central New Mexico was divided into three biostratigraphic zones by Vanderhill (1986). "Faunules" A and B were discussed earlier in this chapter (under "Blancan"). Faunule C in Mesilla Basin was derived from sediments that span the time from just after the Olduvai event to the early part of the Brunhes (i.e., chron C1r), but most of the specimens were reported to come from sediments that predate the Jaramillo event (in chron C1r.2r; 1.77–1.21 Ma). It contains *Mammuthus* and the characteristic Irvingtonian taxa *Paramylodon harlani*, *Smilodon*, and *Canis armbrusteri*. It also contains *Glyptotherium*, *Equus scotti*, *Equus calobatus*, *Cuvieronius*, and *Stegomastodon*. The *Mammuthus* and *Stegomastodon* material was recovered from sediments in chron C1r.2r, *Smilodon* from reversed sediments below the Olduvai (chron C2n). The presence of *Stegomastodon* in association with *Mammuthus* is unusual but not unique.

The Tijeras Arroyo locality in New Mexico was discussed earlier and includes an early record of *Mammuthus*. The Curtis Ranch Fauna from the San Pedro Valley in Arizona is traditionally considered to be Irvingtonian. The fauna lacks *Mammuthus* and straddles

the base of chron C2n (the Olduvai). It is here considered to be latest Blancan.

**Mexico** The El Golfo Fauna in Sonora contains a diverse vertebrate assemblage and provides an important perspective on mammalian faunas from northern Mexico during the Irvingtonian. The fauna includes *Myrmecophaga tridactyla*, *Megalonyx wheatleyi*, *Nothrotheriops*, *Sigmodon curtisi*, *Neotoma (Hodomys) sp.*, *Chasmaportetes johnstoni* (the only Irvingtonian record of this genus), "*Cuvieronius*" sp., *Mammuthus imperator*, *Equus*, *Tapirus*, *Camelops*, *Hemiauchenia*, and *Odocoileus* (Shaw 1981; Shaw and McDonald 1987). Radioisotopic and paleomagnetic information are not available for the El Golfo Fauna.

**Great Plains** In our usage here, the Great Plains is an extensive physiographic province that covers much of the midcontinent region of North America and extends southward to include faunas on the Edwards Plateau region in Texas (Atwood 1940). Many Irvingtonian faunas are reported from the Great Plains region, but thick superposed stratigraphic sequences are few, and this lack hindered the establishment of a reliable faunal sequence. Although some superposed sequences of faunas can be demonstrated, much of the early work ordered the faunas through time on the basis of stage of evolution of mammals and the climatic interpretation of the faunas that was used to relate them to the classic glacial sequence. The use of the now-outdated "four glacial stage" scheme and the mistaken interpretation of multiple ashes as a single "Pearlette" ash led to many problems that only recently began to be sorted out. The understanding that the "Pearlette ash" is in fact three ash beds (Huckleberry Ridge, Mesa Falls, and Lava Creek B; see table 7.1 for dates) and the identification and dating of additional ash beds (e.g., those associated with the eruptions that produced the Cerro Toledo Rhyolites; table 7.1) led to major advances in our understanding of Irvingtonian faunas in the region. Ongoing paleomagnetic and faunal analyses by R. Martin and colleagues promise to clarify the faunal sequence in this region. Some of the most important Irvingtonian localities in the Great Plains are discussed in this section, but a more thorough discussion must await new data.

A sequence of six tills with interbedded sands, silts, and clays exposed along the South Saskatchewan River near Medicine Hat, Alberta, Canada, produced a superposed sequence of Pleistocene faunas that extend from the Irvingtonian into the Rancholabrean (Stalker and Churcher

1970, 1972, 1982; Stalker 1996). The base of the Pleistocene sequence rests unconformably on late Cretaceous rocks. The two units at the base of the section (originally referred to the "Kansan") contain few taxa but include *Mammuthus*, *Equus calobatus*, *Equus scotti*, *Camelops minidokae*, and a ground sloth questionably referred to *Nothrotheriops*. Although it is a small assemblage, the presence of mammoth and the absence of *Bison* suggest an Irvingtonian age.

Wellsch Valley is another fossiliferous sequence, approximately 220 km east of Medicine Hat. It contains a diverse assemblage of faunas and a complex stratigraphy that have hindered attempts to interpret the faunal sequence. The association of *Borophagus*, *Hypolagus*, and *Mammuthus* was reported in the first published discussion of the fauna (Stalker and Churcher 1972). *Allophaiomys*, *Microtus paroperarius*, *Mictomys kansasensis*, and *Pliophenacomys osborni* were added by Stalker and Churcher (1982). The Wellsch Valley Fauna appears to have been derived from several outcrops, and reworking of fossils is possible in some instances (Barendregt et al. 1991, 1998), but a detailed summary of vertebrate paleontological data is lacking. The Jaw Face section of the Wellsch Valley locality is the best studied (Barendregt et al. 1991). Vertebrate paleontological data are published in part, but summary charts lack sufficient detail to resolve important questions. The Jaw Face section as a whole spans the lower Brunhes (chron C1n) and upper Matuyama (chron C1r.1r; Barendregt et al. 1998). Rodent bones were reported to be concentrated in units II and III of the section, stratigraphically below the large mammal bones from unit IV (in the upper portion of the reversely magnetized part of the section; Barendregt et al. 1991). Without detailed data on the stratigraphic positions of the mammalian taxa, especially of *Mammuthus*, *Borophagus*, *Allophaiomys*, and *Microtus*, biochronologic significance of this section cannot be evaluated adequately. The faunal sequence appears to include both Blancan and Irvingtonian components, but faunal, stratigraphic, and paleomagnetic data are not consistent and are difficult to reconcile with radiometric dates (Westgate et al. 1978; Westgate and Gorton 1981) and electron spin resonance dates (Zymela et al. 1988).

The Horse Room Fauna from Salamander Cave in the Black Hills of South Dakota produced a small fauna including *Mictomys* cf. *M. meltoni*, *Microtus paroperarius*, *Microtus meadensis*, *Canis* cf. *C. dirus*, *Equus*, *Antilocapra americana*, and *Camelops* (Mead et al. 1996). The three arvicoline species suggest a possible Cudahy-equivalent age for the deposit (ca. 602–670 ka), but a uranium se-

ries date from the stratigraphic layer that produced the fauna yielded an age of  $252 \pm 30$  ka. In this chapter, we accept that the date is correct (see discussion in Mead et al. 1996:463–464), and consider the fauna latest Irvingtonian. Therefore the fauna includes the youngest radiometrically dated material of *M. paroperarius* and *M. meadensis*, and possibly the youngest record of *Mictomys meltoni*. The record of *Canis* cf. *C. dirus* is the only potential Irvingtonian record of that species (Dundas 1999).

The Hall Ash and Courtland Canal faunas (Jewell County, Kansas) were reported by Eshelman and Hager (1984). The Hall Ash Fauna was recovered from below an ash dated at  $0.706 \pm 0.017$  Ma and tentatively correlated with the Hartford Ash dated elsewhere at approximately 0.74 Ma (Boellstorff 1973, 1978). It contains *Ondatra annectens*, *Mictomys* cf. *M. meltoni*, *Microtus paroperarius*, *Phenacomys* cf. *P. intermedius*, *Geomys*, and *Zapus sandersi*. The Courtland Canal Fauna may be somewhat older and includes *Castoroides* cf. *C. ohioensis*, *Ondatra annectens*, *Mictomys meltoni*, a tentatively identified specimen of *Allophaiomys*, *Platygonus* cf. *P. vetus*, *Titanotylopus* sp., and *Soergelia mayfieldi*.

The Holloman gravel pit located near Frederick, Oklahoma, produced a fauna containing *Mammuthus*, *Gigantocamelus* (= *Titanotylopus*) *spatula*, *Platygonus vetus*, several horse species, and possibly *Stegomastodon* (Dalquest 1977). This locality is another in which both *Mammuthus* and *Stegomastodon* may co-occur, but the identification of *Stegomastodon* from Holloman probably is in error, and the material may actually represent *Teleobunomastodon* (= *Cuvieronius*; see Madden 1980, 1983). The only small mammal recovered was *Cynomys* sp. External age control is not available, but the fauna closely resembles the Gilliland Fauna in Texas (Dalquest 1977).

The Cudahy Fauna in Meade County, Kansas, is situated immediately beneath the Lava Creek B Ash (= "Cudahy Ash" of Hibbard 1944, "Pearlette Ash" of Paulson 1961, and "Pearlette-O" of Izett et al. 1981), the source of which is variably dated to between 602 and 670 ka (table 7.1). This close association of the fauna with a radiometrically dated ash bed made the Cudahy an important fauna for regional correlations for many years. Details of the excavation were provided by Hibbard (1976). The mammalian fauna was discussed by Hibbard (1944), Paulson (1961), and Bell and Repenning (1999) and includes *Geomys tobinensis*, *Ondatra annectens*, *Mictomys meltoni*, *Microtus paroperarius*, *Microtus meadensis*, *Microtus llanensis*, *Equus scotti*, *Megalonyx* sp., and *Mammuthus*. This fauna includes the youngest well-dated and definitive occurrences of *Megalonyx wheatleyi* (an addi-

tional young record is in the McLeod Fauna in Florida), *Ondatra annectens*, and *Mictomys meltoni*, but that species was tentatively identified in a significantly younger fauna in Salamander Cave, South Dakota (Mead et al. 1996).

Several localities with similar faunas in Kansas (Sunbright Ash Pit, Tobin, Wilson Valley) and Texas (the Vera Fauna) were united by Hibbard (1976) into an expanded "Cudahy Fauna"; that term was later restricted to the type Cudahy locality at the Cudahy Ash Pit (Dalquest and Carpenter 1988; Bell and Repenning 1999). The Vera Fauna is also directly beneath Lava Creek B Ash in Knox County, Texas. The depauperate Vera Fauna includes *Geomys tobinensis*, *Microtus paroperarius*, *Microtus llanensis*, and *Ondatra annectens* (Dalquest and Schultz 1992).

The Gilliland Fauna was recovered from the Seymour Formation in Knox and Wilbarger Counties, north-central Texas (Hibbard and Dalquest 1966). The fossils of the Gilliland Fauna were recovered from sands and gravels below the level of the Vera Fauna, but the age of the Gilliland deposits is difficult to assess (Hibbard and Dalquest 1973; Dalquest and Carpenter 1988; Dalquest and Schultz 1992). The most recent age estimate suggests that the Gilliland Fauna may predate the Vera Fauna (which is situated immediately beneath the Lava Creek B Ash) by as much as 400,000 years (Dalquest and Schultz 1992:18). The fauna includes *Nothrotheriops shastensis*, *Paramylodon harlani*, *Geomys tobinensis*, *Sigmodon curtisi*, *Microtus paroperarius*, *Microtus llanensis*, *Proneofiber guildayi*, *Canis armbrusteri*, *Miracinonyx*, *Homotherium*, *Cuvieronius*, *Stegomastodon*, *Mammuthus imperator*, *Tapirus haysii* (originally identified as *Tapirus copei*, but see Ray and Sanders 1984), *Equus scotti*, *Platygonus vetus*, *Camelops*, *Hemiauchenia*, *Capromeryx*, and *Tetrameryx* (Dalquest and Schultz 1992).

Fossils have been known for years along Rock Creek (Briscoe County, Texas), and were derived from several discrete localities during a complicated excavation history (see Dalquest and Schultz 1992); more recent work showed that there are several faunas and two volcanic ash beds in superpositional relationship in the Tule Formation (G. Schultz 1990b). At the base of the sequence is the Martin Ranch Fauna, with *Mammuthus*, *Stegomastodon*, *Paramylodon harlani*, "*Equus (Dolichohippus) simplicidens*," and *Camelops* (Dalquest and Schultz 1992). Elements of this fauna were collected from both below and above the Cerro Toledo X Ash Bed (1.2–1.3 Ma; Izett 1977). The locality is significant because *Mammuthus* is present as early as 1.2–1.3 Ma. About 27 m above Martin Ranch is Gidley's Horse Quarry (Gidley 1900), and 5 m above that level is the Sloth–Camel quarry that produced the other elements of the Rock Creek Quarry reported by Troxell

(1915a, 1915b). This material includes *Paramylodon harlani* (originally identified as *Myiodon*), *Canis armbrusteri*, *Arctodus simus*, *Platygonus vetus* (the original report of *Platygonus compressus* was in error; see Dalquest and Schultz 1992), *Mammuthus* (originally identified as *Elephas*), *Soergelia* (originally identified as *Preptoceras*), and several camelids and equids. At the top of the section, 5 m above the Horse Quarry, is a bed of the Lava Creek B Ash (602–670 ka) that directly overlies the Mayfield Ranch Fauna, a fauna reported by G. Schultz (1990b) and Dalquest and Schultz (1992) to contain *Geomys tobinensis*, *Peromyscus cragini*, *Ondatra annectens*, *Microtus paroperarius*, *Microtus meadensis*, *Microtus llanensis*, and *Mictomys meltoni*.

Several additional faunas are now known to occur directly beneath the Lava Creek B Ash in Texas. The Woody Draw Fauna (Randall County, Texas) includes *Geomys tobinensis*, *Microtus paroperarius*, *Microtus meadensis*, *Mammuthus*, *Equus*, and *Camelops* (Dalquest and Schultz 1992). The Bull Draw Fauna (also Randall County) includes *Geomys tobinensis*, *Ondatra annectens*, *Microtus paroperarius*, *Microtus llanensis*, *Microtus meadensis*, *Mictomys meltoni*, *Mammuthus*, *Platygonus vetus*, and *Camelops*; the Deadman's Creek Fauna is slightly older (4 m below the Lava Creek B Ash) and produced the same rodents (except *M. llanensis*), *Equus conversidens*, and *Camelops* (Dalquest and Schultz 1992).

The Fyllan Cave Fauna in Travis County, central Texas, is the only Irvingtonian fauna from that region (Patton 1965; Taylor 1982; Winkler and Tomida 1988; Winkler and Grady 1990). Fossils were recovered from a narrow fissure fill exposed in a quarry wall and include *Didelphis*, *Dasyus bellus*, *Sylvilagus* cf. *S. hibbardi* (see White 1991a), *Azlanolagus agilis* (Winkler and Tomida 1988), *Ondatra hiatidens* or *Ondatra annectens*, *Microtus guildayi* (= *Allophaiomys* or *Pedomys* or *Pitymys*), *Atopomys texensis*, and *Sigmodon* cf. *S. curtisi*. The sediments are reversely magnetized, which places a minimum age of 780 ka on the fauna (predating chron C1n). The co-occurrence of *Didelphis* with *Sylvilagus* cf. *S. hibbardi*, *Atopomys texensis*, and *Sigmodon* cf. *S. curtisi* is unique. Other records of *Didelphis* are confined to the latest Irvingtonian (e.g., Coleman 2A in Florida; R. Martin 1974) or Rancholabrean. The temporal ranges of the various taxa include late Blancan (if the *Sigmodon* and *Sylvilagus* identifications can be strengthened) through Rancholabrean. We consider the fauna Irvingtonian based primarily on the co-occurrence of *Didelphis*, *Ondatra*, *Microtus*, and *Atopomys*.

The Slaton locality (Lubbock County, Texas) produced a diverse mammalian fauna including *Holmesina septen-*

*trionalis*, *Lepus*, *Geomys bursarius*, *Canis latrans*, *Canis armbrusteri*, *Homotherium serum*, *Mammuthus*, *Platygonus vetus*, *Equus*, *Tetrameryx shuleri*, and *Neofiber leonardi* (Dalquest 1967; Dalquest and Schultz 1992). The absence of *Bison* and the presence of *Mammuthus* and *Platygonus vetus* suggests an Irvingtonian age. The most recent age estimate dated the fauna between 50 and 610 ka (Dalquest and Schultz 1992). The younger age estimate seems too young, based on the presence of *Platygonus vetus*, *Canis armbrusteri*, *Neofiber leonardi*, and *Tetrameryx shuleri*; we suspect a greater antiquity for the fauna. Previous correlations considered Slaton to be approximately equivalent with the Rezabek and Kanopolis faunas in Kansas (Hibbard et al. 1978; Lundelius et al. 1987).

**Florida** Several Irvingtonian faunas were recovered from fissure fill, shallow pond, marsh, and stream deposits in Florida. Useful paleomagnetic data are scarce, there are no igneous rocks on which to base radiometric ages, and superpositional sequences are rare. The ages of these faunas were determined based largely on biostratigraphy. Because Florida is a low-lying, tectonically stable region, relationships of fossil localities to sea level can be used to relate the bone-bearing deposits to former sea levels that are related to the glacial–interglacial cycles, with low sea levels being correlated to glacial stages and high sea levels being related to interglacial stages (Webb 1974a).

These faunas were grouped into four major temporal groups by Morgan and Hulbert (1995), but our designations are slightly different (because we use *Mammuthus* to define the Irvingtonian). “Earliest Irvingtonian” faunas of Morgan and Hulbert (1995) and Ruez (2001) were provisionally interpreted to range in age between approximately 2.0 and 1.6 Ma and are here considered to be latest Blancan. These faunas include the Inglis 1A, Inglis 1C, and De Soto Shell Pit localities and were discussed earlier in this chapter. The Haile 16A Fauna was included with other “late early Irvingtonian” faunas by Morgan and Hulbert (1995), but we discussed it earlier under “Blancan.”

“Late early Irvingtonian” faunas of Morgan and Hulbert (1995) were provisionally interpreted to range in age between approximately 1.6 and 1.0 Ma and include Leisey Shell Pit, Pool Branch, Crystal River, Payne Creek Mine, Rigby Shell Pit, and Haile 21A. Only the Leisey Shell Pit and Payne Creek Mine faunas include *Mammuthus*. Haile 21A includes *Geomys pinetis*, which is found only in Irvingtonian and Rancholabrean faunas in Florida. The Pool Branch, Crystal River, and Rigby Shell Pit faunas are de-

pauperate but were correlated by Morgan and Hulbert (1995) with the Leisey Shell Pit on the basis of overall faunal similarity. We follow that arrangement here. These faunas are characterized by *Nothrotheriops texanus*, *Ondatra annectens*, and *Mammuthus hayi*. These faunas record the first occurrences of *Sylvilagus palustris*, *Geomys pinetis*, *Erethizon dorsatum*, *Castoroides*, and *Canis armbrusteri*. Together with the Haile 16A Fauna, they include early records of *Megalonyx wheatleyi*, *Synaptomys*, *Tapirus haysii* (in all but Rigby; characteristic of Irvingtonian), and *Sigmodon libitinus*. We treated the Haile 16A Fauna earlier in our discussion of Florida Blancan localities, but its faunal similarities, especially with the Leisey Shell Pit, clearly indicate its proximity to the Blancan–Irvingtonian boundary.

The Leisey Shell Pit Fauna (Webb et al. 1989; Hulbert et al. 1995) was dated by a combination of strontium isotope data, paleomagnetic data, and biostratigraphy. When all relevant data were considered, the age range for the Bermont Formation (containing the Leisey Shell Pit Fauna) was interpreted to be between 1.55 and 1.1 Ma (Morgan and Hulbert 1995:30); the vertebrate sites are considered to be less than 1.3 Ma. *Mammuthus* is present in this fauna and thus is among the earliest occurrences of this taxon in North America.

The only “middle Irvingtonian” fauna (provisionally interpreted to range in age between approximately 1.0 and 0.6 Ma) currently recognized in Florida is the McLeod Fauna. This time interval was characterized by Morgan and Hulbert (1995) by the presence of *Neofiber leonardi* and an advanced *Smilodon gracilis* (probably the youngest North American occurrence) and by the first appearance of *Holmesina septentrionalis*, *Panthera onca*, and *Sigmodon bakeri*. This interval records the last occurrence of *Megalonyx wheatleyi*, *Smilodon gracilis*, and *Tapirus haysii* in Florida; the *M. wheatleyi* and *S. gracilis* records are among the youngest known in North America.

A single “late Irvingtonian” Florida fauna, the Coleman 2A (whose age lies between 0.6 and 0.3 Ma), is recognized. This interval was marked in Florida by the limited occurrence of *Microtus aratai* (= *Pitymys*) and by the first Florida occurrence of *Didelphis virginiana* (an earlier record exists in Fyllan Cave, Texas; Taylor 1982), *Neofiber alleni*, and *Tapirus veroensis* (also found in the Kanopolis Fauna in Kansas; Hibbard et al. 1978). The Coleman 2A Fauna was considered by Morgan and Hulbert (1995) to be Irvingtonian based on the presence of three taxa: *Arctodus pristinus*, *Canis armbrusteri* (originally identified as *Canis lupus* by R. Martin 1974, but see discussion in Nowak 1979), and *Platygonus cumberlandensis* (R. Martin 1974; the latter species is now referred to *Platygonus* cf. *P. vetus*; D. Wright

1995). This is one of the youngest occurrences of *C. armbrusteri* (Berta 1995; another late record is Slaton Quarry in Texas; Dalquest and Schultz 1992).

**Eastern Caves** A number of faunas from cave deposits in the eastern United States provide an important perspective on the taxonomic composition of Irvingtonian mammalian faunas in that region. External age control is lacking for these sites, and estimates of their age vary widely. Three such deposits were discovered in the late nineteenth and early twentieth centuries and provided early glimpses of middle Pleistocene mammals from the east: Port Kennedy Cave (perhaps more properly considered as a sinkhole) in Pennsylvania (Wheatley 1871; Cope 1871; Daeschler et al. 1993), Conard Fissure in Arkansas (Brown 1908; Graham 1972), and Cumberland Cave in Maryland (Gidley 1913, 1920a, 1920b; Gidley and Gazin 1933, 1938; Nicholas 1953; Van der Meulen 1978). The Port Kennedy Cave Fauna includes *Megalonyx wheatleyi*, *Arctodus pristinus*, *Panthera onca*, *Miracinonyx inexpectatus*, *Smilodon gracilis* (possibly the youngest occurrence in North America), *Tapirus haysii*, *Mylohyus nasutus*, and *Mammuthus americanum* (Berta 1987; Daeschler et al. 1993; Van Valkenburgh et al. 1990; Seymour 1993). The Conard Fissure Fauna contains *Microtus paroperarius*, *Microtus llanensis* (= *Pedomys*), *Ondatra annectens*, *Miracinonyx inexpectatus*, *Smilodon populator* (possibly the earliest known occurrence), and possibly *Panthera onca* (Graham 1972; Van Valkenburgh et al. 1990; Seymour 1993). The Cumberland Cave Fauna contains *Canis armbrusteri*, *Arctodus pristinus*, *Miracinonyx inexpectatus*, *Panthera onca*, *Smilodon*, *Platygonus vetus*, *Microtus paroperarius*, *Microtus guildayi* (= *Allophaiomys* of Repenning 1987, 1992), *Ondatra annectens*, *Atopomys*, *Mammuthus*, and *Tapirus* (Gidley and Gazin 1938; Van der Meulen 1978; Repenning 1987; Van Valkenburgh et al. 1990; Seymour 1993). These cave occurrences of *M. inexpectatus* are the youngest known records. The Cumberland Cave Fauna records the youngest known occurrence of *P. vetus*.

Three additional faunas are also noteworthy. The Hanover Quarry No. 1 Fauna (Pennsylvania) was reported to contain *Miracinonyx inexpectatus*, *Panthera onca*, *Smilodon gracilis*, *Microtus paroperarius*, and *Microtus guildayi* (Guilday et al. 1984; Seymour 1993). The Cheetah Room Fauna in Hamilton Cave in West Virginia includes *Miracinonyx inexpectatus*, *Panthera onca*, *Mimomys virginianus*, *Allophaiomys pliocaenicus*, *Lasiopodomys decessiensis* (= *Microtus* of Guthrie and Matthews 1971), *Ondatra annectens*, *Synaptomys*, *Atopomys*, and the earliest

known occurrence of *Microtus paroperarius* (Repenning and Grady 1988; Winkler and Grady 1990; Van Valkenburgh et al. 1990; Seymour 1993). Trout Cave Entrance (Guilday 1967b) and Trout Cave No. 2 (Pfaff 1990, 1991) in West Virginia are poorly known faunas. The Trout Cave Entrance (terminology following Pfaff 1990; this locality was previously called "Trout Cave" by Guilday 1967b; Zakrzewski 1975a; and Kurtén and Anderson 1980) Fauna includes *Ondatra annectens* and *Atopomys*. Trout Cave No. 2 includes *Ochotona*, *Microtus paroperarius*, *Microtus llanensis* (= *Pedomys*), *Atopomys*, and *Ondatra annectens* (Pfaff 1990, 1991).

## RANCHOLABREAN

### HISTORICAL CONTEXT, DEFINITION, AND CHARACTERIZATION

The Rancholabrean was established by Savage (1951) and is named for the Rancho La Brea Fauna of California. Rancholabrean faunas were defined and characterized by Savage (1951) by the presence of the Eurasian immigrant *Bison*, many extinct (late Pleistocene) large mammal taxa (e.g., *Equus*, *Camelops*, *Mammuthus*, and *Mammuthus*), and numerous extant species of smaller mammals, especially carnivorans and rodents.

### CURRENT DEFINITION AND CHARACTERIZATION

The Rancholabrean is defined by the first appearance of *Bison* in North America south of 55°N latitude and ends with the extinction of the megafaunal species in the same region. The characterization of the Rancholabrean includes a few taxa that are limited to the Rancholabrean, many that appear in earlier mammal ages but are common in Rancholabrean faunas, and a few that make their first appearance in the Rancholabrean but persist into the modern fauna. The lack of clear distinction between Irvingtonian and Rancholabrean faunas was, in at least one instance, used to justify treating the Rancholabrean as a temporal division within an expanded Irvingtonian (L. Martin 1985, 1989), but most authors continue to recognize it as a distinct mammal age.

Taxa that are limited to the Rancholabrean include *Megalonyx jeffersonii*, *Platygonus compressus*, *Canis dirus*, *Panthera atrox*, and *Miracinonyx trumani*. A questionable record of *M. jeffersonii* from Holloman (a broken humerus



fragment) was reported by Dalquest (1977); it is not clear whether this specimen is sufficient for reliable species allocation. The Papago Springs Cave specimens of *Platygonus* are the oldest dated records of the species (age taken here as 107 ka; see Czaplewski et al. 1999b); they were originally identified as *P. alemanii* (Skinner 1942), which Slaughter (1966) considered a junior synonym of *P. compressus*. *Canis* cf. *C. dirus* is recorded in the Salamander Cave Fauna in South Dakota (Mead et al. 1996) and is associated with a radiometric date of  $252 \pm 30$  ka; if the material actually represents *C. dirus*, it would constitute a latest Irvingtonian record for the species. There are two records of *Ovis* that predate the Rancholabrean, but neither can be identified reliably to species. *Ovis* cf. *O. canadensis* was reported from the Pit Fauna Porcupine Cave (Barnosky and Rasmussen 1988), and *Ovis* sp. was reported from the El Golfo Fauna (Lindsay 1984).

Taxa that first appear in earlier mammal ages but are common in Rancholabrean faunas include *Didelphis*, *Nothrotheriops*, *Paramylodon harlani*, *Glyptotherium*, *Brachylagus idahoensis*, *Sylvilagus*, *Clethrionomys*, *Lemmiscus curtatus*, *Ondatra zibethicus*, *Marmota flaviventris*, *Marmota monax*, *Cynomys gunnisoni*, *Cynomys ludovicianus*, *Panthera onca*, *Smilodon populator*, *Mustela erminea*, *Brachyprotoma*, *Conepatus*, *Canis latrans*, *Canis lupus*, *Arctodus simus*, *Ursus*, *Euceratherium*, *Oreamnos*, *Mammut americanum*, and *Mammuthus*.

Taxa that first appear in the Rancholabrean and persist into the modern fauna include *Aplodontia rufa*, *Vulpes velox*, *Felis concolor*, *Alces*, *Bison*, *Ovis canadensis*, *Rangifer tarandus* (known from an Irvingtonian equivalent fauna in Cape Deceit, Alaska), and *Homo sapiens*.

## ARRIVAL OF *BISON*

*Bison* was previously reported to appear in Alaska earlier than in the coterminous United States (Péwé 1975, 1989; Péwé and Hopkins 1967; Harington 1978, 1984). However, the time of first appearance of *Bison* in Alaska is now a confusing issue because of problematic stratigraphic attributions. *Bison* was reported from late Pliocene to early Pleistocene deposits in central Alaska (Péwé 1975, 1989; Péwé and Hopkins 1967). The fossils in question were reported to come from the "Fox Gravel" and the Gold Hill Loess. Recent data, including ages of volcanic ash beds, suggest that the "Fox Gravel" is time transgressive or represents multiple units (Hamilton et al. 1988; Westgate et al. 1990; Preece et al. 1999). Unfortunately, Péwé and Hopkins (1967) and Péwé (1989) did not identify specific localities from which their *Bison*

specimens were recovered. To our knowledge, none of these fossils were curated in museums, so it is impossible to identify specific localities or verify the reported identifications.

R. H. Tedford (pers. comm., 2001) indicated that the Frick collection of Alaskan Pleistocene mammals in the American Museum of Natural History contains specimens of *Bison* that were recovered with those of *Xenocyon lycaenoides*, *Alces* cf. *A. latifrons*, *Praeovibos recticornis*, *Praeovibos priscus*, *Saiga tatarica*, and *Bootherium bombifrons* when the lower sediments of the Cripple Creek Sump were being dredged. These taxa are characteristic of the Olyorian faunas of the Kolyma Lowland of Siberia, which are dated between the late Matuyama and early Brunhes (1.5 to 0.5 Ma; (Repenning 1992; Sher 1986). Although the *Bison* remains could have been mixed with the other taxa when dredged, the association suggests contemporaneity.

Previous attempts to establish a reliable date for the arrival of *Bison* in the coterminous United States depended to a large extent on the correlation of faunas containing *Bison* to the classic (four-fold) glacial sequence. Many seemingly early faunas that contain *Bison* were previously assigned to the late "Illinoian" glacial stage because many workers regarded *Bison* to have first appeared at that time, an unfortunately circular argument. No *Bison* are known from any fauna in close association with the 0.60–0.67 Ma Lava Creek B Ash (table 7.1). It probably is significant that *Bison* remains are not known from the rich "Sheridanian" faunas from either northern (e.g., Hay Springs, Rushville, Gordon) or southern (Angus) Nebraska. These faunas lie below the Loveland Loess (Repenning 1987), for which thermoluminescent (TL) dates indicate an age between 200 and 120 ka (Forman et al. 1992; Muhs et al. 1999). These data strongly suggest that *Bison* appeared on the central Great Plains after 200 ka.

Several authors recently reported putative *Bison* remains in association with radiometric dates or in sedimentary contexts that suggest that *Bison* entered the coterminous United States sometime before 120 ka. These purported records suffer from dubious identification, unclear stratigraphic position, or unsatisfactory chronologic control. The localities in question include the Macasphalt Shell Pit and Inglis 1A localities in Florida (see J. McDonald and Morgan 1999; the age of Macasphalt Shell Pit was estimated between 2.5 and 2.0 Ma by Jones et al. 1991 and Morgan and Hulbert 1995; the age of Inglis 1A was estimated between approximately 1.9 to 1.6 Ma by Webb et al. 1989, Jones et al. 1991, and Morgan and Hulbert 1995). Additional questionable reports include younger records

from Teichert Gravel Pit near Sacramento, California, and the Camp Cady faunas in southern California. The Teichert Quarry material consists of a single isolated second phalanx in the collection of the University of California Museum of Paleontology; the age of the locality was estimated at 130 ka by Repenning et al. (1990). The Camp Cady Fauna ranges in age between 350 and 19 ka (Jefferson 1987); the purported *Bison* material consists of fragmentary vertebral elements that were recently reidentified as belonging to *Camelops* (Scott and Cox 2002). The specimens are from questionable stratigraphic provenience (see discussions and data presented in Jefferson 1968, 1985, 1987, 1991).

A report of early *Bison* from unit c1 at Jones Spring, Missouri, has more reliable stratigraphic control and was in close stratigraphic association with a radiometric date. A uranium series date of 160 ka was obtained by C. R. McKinney from *Mammut* enamel from the contact of unit c1 with underlying gray clay (Haynes 1985:22; Saunders 1988:131).

Perhaps the most reliable early *Bison* date is from layers B and E of the American Falls Formation in Idaho. Twenty-one cranial elements referred to *B. latifrons* from layer E were recovered from a coarse alluvium conformably underlying fine-grained alluvium (Hopkins et al. 1969; Scott et al. 1982; Pinsof 1991). The fine-grained alluvium accumulated behind a lava dam of Cedar Butte basalt, dated at  $72,000 \pm 14,000$  years B.P. (Scott et al. 1982). This sedimentary package is superposed on Crystal Springs basalt, dated at  $210,000 \pm 60,000$  yr B.P., thus bracketing the age of these specimens.

## TEMPORAL EXTENT

The temporal extent of the Rancholabrean is poorly established. The first appearance of *Bison* in North America south of  $55^{\circ}\text{N}$  latitude defines the beginning of the Rancholabrean, but the exact time of its arrival in this region is not known. The American Falls *Bison* records bracket the time of arrival between  $210 \pm 60$  and  $72 \pm 14$  ka. The Jones Spring (Missouri) date provides a minimum arrival time of 160 ka (Saunders 1988).

Because no younger mammal age has been established, the end of the Rancholabrean is operationally defined by the extinction of large-bodied mammal species near the end of the Pleistocene (e.g., C. Schultz et al. 1978). The date of the end of the Rancholabrean therefore must be based on the youngest radiocarbon dates of now-extinct large mammals. A number of compilations and analyses of these terminal dates are available (Hester 1960, 1967;

Kurtén and Anderson 1980; Meltzer and Mead 1983, 1985; Lundelius et al. 1983; Mead and Meltzer 1984; Barnosky 1989; Grayson 1989, 1991), but recent developments in radiocarbon dating technology, primarily using accelerator mass spectrometry  $^{14}\text{C}$  (Stafford 1990), show that many of the previously reported post-Pleistocene dates (i.e., younger than 10 ka) may be erroneous, and all should be viewed with caution.

Using the criteria outlined by Meltzer and Mead (1985), 10 large mammal taxa with "good dates" were identified by Grayson (1991). Available data demonstrate that the extinction of large mammals was time-transgressive, ranging from approximately  $23.23 \pm 0.49$  ka (*Glyptotherium*; latest record from Laubach 3 in Texas; Lundelius 1985) and  $20.45 \pm 0.46$  ka (*Paramylodon*; latest reliable record from Rancho La Brea; Meltzer and Mead 1985) to approximately 10 ka for several taxa (Grayson 1991). The youngest good date for eight of these taxa (*Palaeolama*, *Tapirus*, *Smilodon*, *Mammuthus*, *Mammut*, *Equus*, *Camelops*, and *Nothrotheriops*) is younger than 12 ka (Grayson 1991). With adequate consideration of the error bar on the youngest of these dates, the latest survivor of an extinct large-bodied mammal could be approximately 9.5 ka. Based on these data, the end of the Rancholabrean can be considered to be as recent as 9500 years ago, a time slightly younger than the 10,000-year date recognized earlier as the lower boundary of the Holocene (see discussion of the Pleistocene–Holocene boundary earlier in this chapter).

Our discussion of the extinction event (and hence the end of the Rancholabrean) does not include the remarkable records of *Mammuthus* from Wrangel Island, where it survived until approximately 4000 years ago (Vartanyan et al. 1993, 1995; Lister 1993b; P. Martin and Stuart 1995; Arslanov et al. 1998). We justify this exclusion not by special pleading but as a direct result of our geographic restriction of use of the term *Rancholabrean* to faunas from latitudes south of  $55^{\circ}\text{N}$  latitude.

Many possible causes of the extinction have been proposed over the last three decades. These include overkill by early humans, seasonally out-of-step mating periods, epidemics, climatic stress, loss of habitat, vegetation changes, and food shortages (P. Martin 1967, 1973; Axelrod 1967; Guilday 1967a; B. Slaughter 1967; Dreimanis 1968; Van Valen 1970; Alford 1974; Mosimann and Martin 1975; L. Martin and Neuner 1978; Graham and Lundelius 1984; Delcourt 1985; Owen-Smith 1987; Barnosky 1989; Webb and Barnosky 1989; Stuart 1991; Flannery 1995; Beck 1996; Alroy 1999, 2001; Klein 2000). A detailed consideration of this subject is beyond the

scope of this chapter, but recent reviews of the subject can be found in MacPhee (1999).

### TEMPORAL DIVISIONS OF THE RANCHOLABREAN

There is no basis for a numeric chronology within the Rancholabrean except for its terminal ("Wisconsinan") phase, which falls within the range of radiocarbon dating techniques. The age lies completely within the Brunhes chron (chron C1n), during which polar wander is recorded (Opdyke and Channell 1996), but there are no known events of reversed magnetic polarity suitable for boundary markers. Recent efforts to document fluctuations in geomagnetic paleointensity (Stoner et al. 1998, 2002) show great promise, but these data are not yet developed sufficiently to permit temporal boundary recognition. Thus for most of the Rancholabrean, chronologic control necessarily relies on faunal relationships, radiometric dates derived from associated igneous deposits, and new dating techniques applied to paleosols and loess deposits. Attempts to fit faunas into the fourfold glacial–interglacial framework previously used were perhaps marginally successful, but climatologists now recognize as many as 20 significant cooling events in the last 2 million years (Shackleton and Opdyke 1976; Richmond and Fullerton 1986; Raymo 1992), and terrestrial mammal faunal correlation should not be made solely on the basis on climatic inferences (see Lindsay 1997 and the following discussion).

**Bison Chronology** The temporal duration of many mammalian species is greater than the short span of the Rancholabrean. Thus most schemes proposed to divide the Pliocene and Pleistocene mammal ages (C. Schultz et al. 1978; L. Martin 1979; Repenning 1987; Repenning et al. 1990) leave an undifferentiated Rancholabrean. Attempts to divide the Rancholabrean based on perceptions of *Bison* evolution produced varying results (Hibbard 1955a; C. Schultz and Martin 1970; J. McDonald 1981; C. Schultz and Hillerud 1977), and no one system is generally accepted. The disagreements result from uncertainty in the age relationships of relevant faunas and lack of consensus in the interpretation of *Bison* phylogeny and taxonomy (Wilson 1974a, 1974b; J. McDonald 1981). Significant sexual dimorphism, intraspecific and individual variation, and the purported presence of shifting geographic and temporal clines (C. Schultz and Hillerud 1977; J. McDonald 1981; Gingerich 1993) combine to form an intractable systematic and taxonomic problem.

Chronologic control for many purported early *Bison* faunas is derived from alternative techniques such as presumed superpositional relationships and temporal ordering of terrace sequences in the central part of the continent (C. Schultz et al. 1972). Analyses of sequential terrace deposits suggest that large-horned *Bison* (e.g., *B. latifrons*, *B. alaskensis*, and *B. alleni*) are, in general, older than the short-horned *B. bison antiquus* or *B. b. occidentalis* (Skinner and Kaisen 1947; Guthrie 1970; C. Schultz et al. 1972; C. Schultz and Hillerud 1977), and that these, in turn, are older than *B. b. bison*. However, *B. latifrons* survived into the latest Pleistocene, at least in California and Idaho, and was contemporaneous with *B. b. antiquus* (Wyman 1922; Miller 1968, 1971; Miller and Brotherson 1979; Pinsof 1991; Gust 1993). In Florida, *B. latifrons* was reported to predate the appearance of *B. b. antiquus*; temporal ordering was based on a eustatic–hydrologic approach (Webb 1974a).

**Loess Stratigraphy** Recent work on loess stratigraphy in the upper Midwest indicates that the widespread loess deposits across the Great Plains may be used as proxies for glacial advance. Numeric ages on loess deposits beyond the Laurentide ice sheet margin are becoming more reliable and may provide a detailed numeric chronology for the last two glacial–interglacial cycles. TL dates from the Loveland Loess type section in western Iowa suggest that the Loveland Loess accumulated between approximately 180 and 120 ka (Forman et al. 1992). TL ages from eastern Colorado and western Nebraska indicate a similar age for the Loveland Loess (between approximately 200 and 120 ka; Muhs et al. 1999), as do TL ages from the Mississippi Valley (Forman et al. 1992; Markewich et al. 1998; Rodbell et al. 1997).

Application of numeric dating techniques also recently increased the usefulness of paleosols as stratigraphic markers in the midcontinent Quaternary sequence. Several paleosols are widespread, and they developed during both stadial (glacial) and interglacial intervals. Using  $^{10}\text{Be}$  inventories on soils, the beginning of the last glacial in Illinois was placed after 59 ka by Curry and Pavich (1996), the last interglacial was placed between 59 and 130 ka, and the penultimate glacial was placed between 130 and 190 ka. Similar studies were applied to paleosols in the Middle Mississippi Valley loess sequence (Markewich et al. 1998). The potential for dating loess deposits and paleosols associated with vertebrate-bearing sediments offers hope of obtaining a better numeric chronology for Rancholabrean faunas. Relating Rancholabrean faunas to the location of the glacial front is essential for improving

chronology and interpreting glacial and interglacial paleoenvironments.

**Radiocarbon Interval** The last 40 to 55 ka (Trumbore 2000) of the Rancholabrean is known in greater detail than the earlier part because of the applicability of radiocarbon dating and the greater frequency of specimen recovery. As a result, changes in the fauna during that interval are better dated than for any other part of the Pliocene and Pleistocene, and the most commonly used temporal division of the Rancholabrean is “pre-” versus “post-radiocarbon.”

### GEOGRAPHIC DISTRIBUTION

Rancholabrean faunas are widespread across North America south of 55°N latitude (see FAUNMAP Working Group 1994; figure 7.6). We center our discussion here on the three available faunal sequences that reveal faunal change through that time in one place. Of the many Rancholabrean faunas known from North America, only a few show a superpositional relationship that spans a sufficient period of time to provide a faunal sequence that could help to subdivide the Rancholabrean.

**Medicine Hat Sequence** The Medicine Hat sequence (Stalker 1969, 1996; Stalker and Churcher 1970, 1972) is one of the few Rancholabrean sequences in North America. There are seven faunal levels above the Irvingtonian faunas. The lowest of these faunas, separated from the Irvingtonian component by a till, is diverse (33 taxa) and contains *Bison* (therefore, by definition, it is a Rancholabrean fauna). Other taxa include *Ondatra zibethicus*, *Erethizon dorsatum*, *Cynomys ludovicianus*, *Canis latrans*, *Canis lupus*, *Canis dirus*, *Panthera atrox*, *Equus conversidens*, *Equus niobrarenensis*, *Cervus canadensis*, *Ovis canadensis*, *Rangifer tarandus*, ?*Alces*, and *Mammuthus jeffersonii*. The fauna was assigned to an interglacial time.

This fauna is followed by a very small assemblage (three taxa) consisting of *Mammuthus primigenius* (the first appearance of *M. primigenius* in the sequence; it may indicate cold conditions), *Equus giganteus*, and *Antilocapra*. This fauna was assigned to the early “Wisconsinan.”

Separated from the first two faunas by two tills and an unconformity are two faunal horizons of 11 and 7 taxa, respectively; the lowest of these includes *Smilodon populator* (formerly *Smilodon californicus*; see Berta 1985), *Mammuthus*, *Equus* cf. *E. giganteus*, *Equus conversidens*, *Hemiauchenia macrocephala* (formerly *Tanupolama stevensi*, but see Webb 1974b), *Camelops hesternus*,

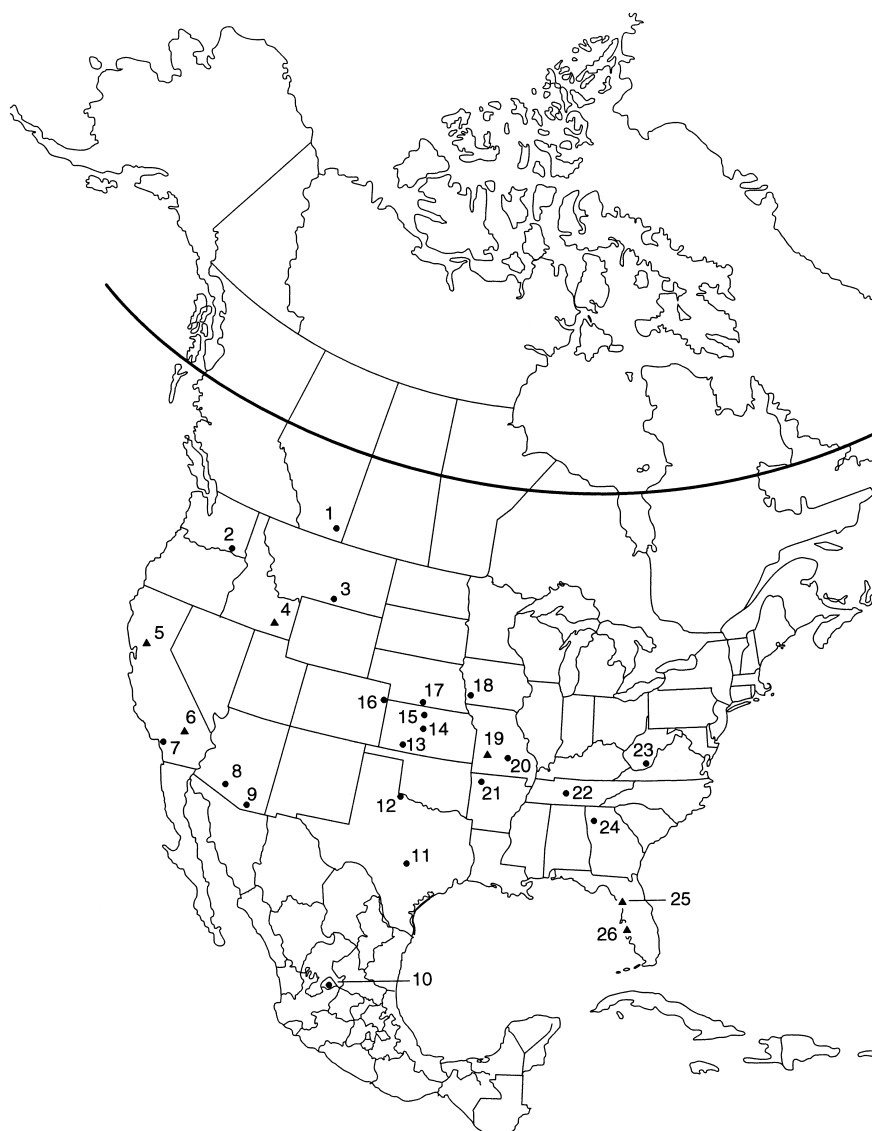
*Odocoileus* sp., and *Bison* sp. The upper fauna is separated from the lower by an unconformity and includes *C. hesternus*, *S. populator*, and *E. conversidens*.

Two stratigraphically higher faunas, both assigned to the late “Wisconsinan,” are separated from the previous two faunas by a till reported to date to 20 ka. The faunas have similar taxa except for the species of mammoth. The lower has *Mammuthus imperator*, and the upper has *Mammuthus primigenius*. A Holocene assemblage is found at the top of the sequence with *Bison bison* and *Antilocapra*.

Although some of the faunas are small, they clearly show shifts in environmental conditions from bottom to top. At present, reliable numeric age control on this sequence is not extensive (see Stalker 1996). A number of electron spin resonance dates from various units and localities in this area were reported by Zymela et al. (1988), but the results are too variable to be helpful.

**Meade Basin, Kansas** In the Meade Basin, a sequence of faunas is interpreted to span at least two glacial stages and include one interglacial stage (Hibbard et al. 1965; R. Martin et al. 2000). These are, from oldest to youngest, Adams, Butler Spring, Mt. Scott, Cragin Quarry, Jinglebob, Jones (dated to between 26.7 and 29.0 ka; Hibbard 1970), and Robert (approximately 11.1 ka; G. Schultz 1969). The Butler Spring and Adams microfaunas includes predominantly modern species (G. Schultz 1965, 1967; R. Martin et al. 2000), supporting their placement in the Rancholabrean. *Bison* first appears in this sequence in the Mt. Scott Fauna, is absent from Cragin Quarry, and reappears in the Jinglebob and Jones faunas. The Cragin Quarry Fauna was previously interpreted to represent an interglacial stage (Hibbard and Taylor 1960; Hibbard 1963), suggesting the possibility that the absence of *Bison* was somehow related to environmental change.

**Kennewick Sequence, Washington** A temporally long, stratigraphically superposed sequence of Rancholabrean localities near Kennewick was described by Rensberger et al. (1984; see also Rensberger and Barnosky 1993). The assemblages were assigned a Rancholabrean age based on the large percentage of Recent species. *Bison* is absent. The top of the section is clearly dated by Mazama Ash to around 7 ka. Dating the lowest part of the sequence is more problematic, with the best evidence coming from the estimated rate for formation of calcrete horizons. The calcrete chronology probably brackets the base of the section to between 40 and 328 ka, although using a very slow rate for calcrete formation would make the base of the section up to a few hundred thousand



**FIGURE 7.6** Putative early *Bison* localities and significant Rancholabrean localities discussed in the text. The bold line represents 55°N latitude. Independent biochronologies should be established for the region north of this line, and the term *Rancholabrean* should not be applied to faunas there. Localities with external chronologic control indicating an age between approximately 210 and 9.5 ka are considered to be temporal equivalents of Rancholabrean faunas elsewhere in North America. Circles = Rancholabrean faunas; triangles = putative early *Bison* localities; see text for discussion. 1, Medicine Hat sequence (in part), Alberta, Canada; 2, Kennewick sequence (in part), WA; 3, False Cougar Cave, MT; 4, American Falls, ID; 5, Teichert Gravel Pit, CA; 6, Camp Cady Fauna, CA; 7, Rancho La Brea, CA; 8, Ventana Cave, AZ; 9, Papago Springs Cave, AZ; 10, Cedazo, Aguascalientes, Mexico; 11, Cave Without a Name, TX; 12, Howard Ranch, TX; 13, Meade County faunas (in part), KS; 14, Kanopolis, KS; 15, Rezabek, KS; 16, Jones-Miller site, CO; 17, North Cove, NB; 18, Waubonsie, IA; 19, Jones Spring, MO; 20, Little Beaver Cave, MO; 21, Peccary Cave, AR; 22, Cheek Bend Cave, TN; 23, Patton Cave, WV; 24, Ladds Quarry, GA; 25, Inglis 1A (Blancan), FL; 26, Macasphalt Shell Pit (Blancan), FL.

years older. Rensberger and Barnosky (1993) argued against slower-rate calculations for the calcrete and proposed that the entire section be assigned to the Rancholabrean, but their placement of the Rancholabrean–Irvingtonian boundary was at 450 ka (Rensberger and Barnosky 1993:322). *Microtus meadensis* (= *Terricola* or *Pitymys*) is present in the lowest part of the Kennewick

sequence. Its youngest known occurrence elsewhere is in Salamander Cave in South Dakota, with an associated date of  $252 \pm 30$  ka (Mead et al. 1996). Using the calcrete calculations that yield ages up to 328 ka and the presence of *M. meadensis* in the basal part of the section, it seems likely that the lower part of the Kennewick is Irvingtonian.

## CHRONOCLINES AND EVOLUTION

Phyletic sequences or chronoclines in the Pliocene and Pleistocene have been proposed and evaluated for several mammalian lineages (e.g., *Megalonix*, *Ogmodontomys*, *Ondatra*, *Ophiomys*, *Mictomys*, *Microtus pennsylvanicus*, *Platygonus*, *Bison*, and *Mammuthus*). Studies on evolutionary changes in these lineages examine morphologic change at various chronologic scales and contribute to temporal divisions in the Pliocene and Pleistocene mammal ages. If they can be calibrated independently or demonstrated to be in superpositional context, evolutionary changes in species help provide regional faunal characterizations. In lineages for which an especially rich fossil record is available, it can be difficult to determine adequate species boundaries (see R. Martin 1996). Studies of such lineages, when considered with other works that establish morphologic stasis over long periods of time (e.g., Barnosky 1987; Lich 1990) contribute significantly to our understanding of the range of morphologic variation within and between closely related species and provide a rich data resource that is now being used to address questions of evolutionary tempo and mode (Barnosky 1987; Gingerich 1993; R. Martin and Barnosky 1993). The phylogenetic affinity of many of these fossils with the modern biota also provides a crucial bridge between paleontology and neontology because the consequences of the incomplete nature of the fossil record can be assessed at least tentatively in comparison with the more adequately understood extant biota. Detailed studies of morphologic variation within and between various lineages thus assume a broader importance in the contexts of determination of mosaic evolution in different morphologic systems (e.g., dentition versus hind limb morphology in muskrats; Viriot et al. 1993; R. Martin 1996; Flint 1998), reliable identification of isolated fossil remains (e.g., teeth), and the interplay of these and other factors in establishing reliable and stable taxon definitions for mammal ages and their temporal divisions (Woodburne 1996). By providing a means for definitive identification of fossils that is independent of modern geographic distribution, these studies also help to offset the use of modern geographic distribution to refine taxonomic identifications. That practice is still common in studies of late Pleistocene and Holocene mammals but should be avoided because it introduces circularity in discussions of faunal response to climatic changes.

Published explorations of morphologic variation in Pliocene and Quaternary mammals encompass a broad range of taxa and provide a crucial database on morphologic change and variation through time and space. The

voluminous literature on morphologic variation in Quaternary rodents was reviewed by R. Martin (1993). Studies on additional groups not covered in his review include *Holmesina* (Hulbert and Morgan 1993), *Blarina* (Guilday 1957; Graham and Semken 1976; Jones et al. 1984), *Sorex* (Jammot 1972; Junge and Hoffmann 1981; Carraway 1990, 1995), *Allophaiomys* (Repenning 1992), *Dicrostonyx* (Eger 1995), *Guildayomys* (Zakrzewski 1984), *Hibbardomys* (Zakrzewski 1984), *Microtus* (Bell and Repenning 1999), *Microtus pennsylvanicus* (Barnosky 1993; Gordon 1999), *Ogmodontomys* (Zakrzewski 1967), *Ondatra* (Viriot et al. 1993; R. Martin 1996), *Ophiomys* (Hibbard and Zakrzewski 1967), *Pliophenacomys* (Hibbard and Zakrzewski 1972; Zakrzewski 1984), *Neotoma* (Zakrzewski 1993), *Zapus* (Klingener 1963; R. Martin 1994), *Dipodomys* (Carrasco 2000), *Cynomys* (Goodwin 1993), *Canis* (Nowak 1979), *Panthera* (Seymour 1993), *Smilodon* (Berta 1985, 1987), *Odocoileus* (Purdue and Reitz 1993), and *Mammuthus* (C. Schultz et al. 1972; Lister 1993a; Webb and Dudley 1995).

## GLACIAL CLIMATES: HARMONIOUS AND DISHARMONIOUS FAUNAS

Perceived changes in mammalian biogeography in response to alternating glacial–interglacial environments provide a potential means to refine Pleistocene chronology. The recovery, in deposits south of their modern distribution, of mammal species that today are limited to boreal and tundra environments usually is interpreted as signifying glacial conditions at the time of deposition. Conversely, the presence of southern species in localities north of their present range is interpreted to indicate interglacial intervals (Blair 1958). These geographic shifts sometimes result in direct stratigraphic associations of species that are today entirely allopatric; the apparent sympatry in the fossil record results in species assemblages that have no modern analog. Such species assemblages have been recognized in the Great Plains for decades (Hibbard 1944, 1949a). These nonanalog assemblages, which vary in species richness (Graham and Semken 1976), are variously called intermingled (Graham 1985), disharmonious (Semken 1988; Lundelius 1989), mixed (Markova 1992), mosaic (Guthrie 1982), and extraprovincial (Roy et al. 1995). Conversely, Holocene interglacial faunas with a species richness similar to that in modern biomes are sometimes called depauperate (L. Martin and Hoffmann 1987) or impoverished (R. Martin and Webb 1974; Semken 1974, 1984).

Nonanalog faunas sometimes are used as biostratigraphic tools to separate late Pleistocene glacial faunas from Holocene faunas, the latter of which rarely have more than two allopatric species (Semken 1988). This practice has its origin in the mid-twentieth century when Claude Hibbard, Dwight Taylor, and their colleagues attempted to order a diverse suite of molluscan and mammalian faunas from the Great Plains into a meaningful temporal sequence. In conjunction with stratigraphic position and the perceived stage of evolution of certain species, Hibbard et al. (1965) used the relative abundance of "cool summer" (boreal) and "mild winter" (temperate) faunal elements to place a series of faunas in southwestern Kansas in biostratigraphic succession. The absence of any Meade County Fauna directly comparable to the modern led Hibbard and Taylor (1960) and Hibbard et al. (1965) to regard the Holocene as "geologically atypical" for the Great Plains Quaternary. Pre-Holocene faunas with wholly boreal exotic species (not found today in Meade County; e.g., Jones, Butler Spring, and Cudahy) were assigned to glacial intervals, and those with only temperate exotics (Cragin Quarry and Borchers) were assigned to interglacial times. These faunas generally are regarded as nonanalog faunas now (but see Alroy 1999). Other nonanalog faunas (e.g., Jinglebob and Mt. Scott) were difficult to place because they contained both boreal and temperate exotics. With misgivings (Hibbard, pers. comm., 1965), the Jinglebob Fauna was assigned to an interglacial and the Mt. Scott Fauna to a glacial age based on the relative abundance of boreal and temperate taxa.

An increased reliance on paleomagnetic and radioisotopic data for faunal correlation has reduced the prevalence of such paleoecologic correlations, but faunal paleoecology is still used in some cases to assign strata to or exclude them from certain intervals of time, especially those of the late Quaternary. This practice assumes that factors controlling modern geographic distributions of species are adequately understood and that ecological tolerances and preferences of extant species were static through time. These assumptions were explicitly stated by Hibbard (1949a) but have not received adequate critical evaluation since that time, especially in light of modern alternative philosophical and methodologic approaches to paleoecologic reconstructions (Owen et al. 2000; Bell and Gauthier 2002). Although climatic correlations may be applicable to faunas in the latest Pleistocene and Holocene, their application to older faunas is questionable and should be avoided (see Lindsay 1997).

Radiocarbon dating of late Pleistocene nonanalog faunas with boreal exotics in association with modern residents can be used to test Hibbard's model generally and

late Pleistocene faunas specifically. Late Pleistocene nonanalog faunas are defined by the presence of a few totally allopatric species (usually north boreal or tundra); widespread coexistence of boreal, prairie, and deciduous forest species now sympatric only along ecotones separating biotic provinces; and a species richness that may be twice that living around the site today (FAUNMAP Working Group 1996; Graham 1985; L. Martin and Hoffmann 1987; Semken 1988). A recent study of multiple accelerator mass spectrometry  $^{14}\text{C}$  dates conducted by Stafford et al. (1999) demonstrated contemporaneity of nonanalog species assemblages in discrete stratigraphic levels from Peccary Cave, Arkansas (Semken 1984) and Cheek Bend Cave, Tennessee (Klippel and Parmalee 1984).

Although the quantity and quality of radiocarbon dates from different sites vary widely, additional examples of radiocarbon-dated late Pleistocene glacial nonanalog faunas document the wide geographic distribution of such faunas. These include Patton Cave, West Virginia (13.3 ka; Grady 1988); Ladds Quarry, Georgia (10.9–10.3 ka; Lipps and Ray 1967; Holman 1985a, 1985b); Little Beaver Cave, Missouri (11.0 ka; Schubert 1997; Schubert and Graham 2000); the Waubonsie Fauna, Iowa (14.8 ka; Rhodes 1984); Cave Without a Name, Texas (10.9 ka; Lundelius 1967); Howard Ranch, Texas (19.1–16.8 ka; Dalquest 1965; Dalquest and Schultz 1992); North Cove, Nebraska (12.8–11.0 ka; Stewart 1987); False Cougar Cave, Montana (14.6–10.5 ka; M. Graham et al. 1987); Jones–Miller, Colorado (16.6 ka; Graham 1987); and Ventana Cave, Arizona (12.6 ka; Haury 1975; Mehringer 1967).

The significance and interpretation of nonanalog faunas was recently challenged on the basis of both empirical and methodologic concerns (Alroy 1999). These concerns centered primarily on the temporal association of (and possible causal relationships between) climate change, the breakup of nonanalog mammal communities, and the late Pleistocene megafaunal extinction in North America. The arguments raised by Alroy (1999) pose important challenges to traditional interpretations of Quaternary mammal faunal dynamics and provide incentive for reevaluating traditional hypotheses.

## NORTH AMERICAN PLIOCENE AND PLEISTOCENE FAUNAL PROVINCES

The mammal ages treated in this chapter, and their proposed temporal divisions, are defined and characterized by appearances and disappearances of certain species.

Earlier in this chapter we noted several instances in which perceived geographic differences in faunas play a role in the interpretation of temporal and spatial extent of mammal ages or their temporal divisions. The potential significance of these considerations affords us an opportunity to present some preliminary considerations regarding faunal provinciality and its potential impact on biochronologic interpretations and mammal age boundaries with respect to the Pliocene and Pleistocene.

That species are not distributed uniformly over the surface of the globe but rather can be seen as displaying varying degrees of provinciality is one of the most pervasive ideas in biogeography. Faunal provinciality has important implications for biochronology, some of which were noted by W. D. Matthew early in the twentieth century (Matthew 1915). The complex nature of faunal dynamics throughout the late Cenozoic, combined with the possibility that faunal provinces may persist through long periods of time (FAUNMAP Working Group 1996), forces the consideration that different faunal regions or provinces may need different biochronologies. For example, Repenning et al. (1995:12) recently stated, "The 'North American mammal ages' cannot be used in Alaska without complete redefinition." This difference in the faunal character of Alaska is tacitly acknowledged in much of the literature on North American mammalian biochronology and is one of the primary reasons for our geographic restriction of the Blancan, Irvingtonian, and Rancholabrean. However, few authors have addressed the immense complexities and potential ramifications of temporally variable faunal provinciality.

These issues are perhaps of greatest significance in the late Cenozoic, when temporal resolution may be sufficient to detect (and potentially resolve) the time-transgressive nature of species dispersal. The diverse range of geochronologic techniques available to scientists working in the Pliocene and, especially, the Pleistocene (Noller et al. 2000) affords a unique opportunity to resolve time intervals of short duration.

Climatic perturbations during the Pliocene, Pleistocene, and Holocene triggered varied, often individualistic responses among mammalian taxa. Mammalian faunal dynamics during these times certainly included significant (in some cases dramatic) geographic range adjustments of individual species. Some of the complications and challenges of recognizing faunal provinciality in the late Pleistocene were discussed by L. Martin and Neuner (1978), Graham (1979), and L. Martin and Hoffmann (1987). More recently, the FAUNMAP database documented that the rate, timing, breadth, and direction of geographic range adjustments varied signifi-

cantly among different mammal species in North America (FAUNMAP Working Group 1996). Despite the response of individual species to changing conditions and the lack of modern analogs for community compositions in the Pleistocene, cluster analyses of species revealed that mammalian faunal provinces of the late Pleistocene were related to moisture and temperature gradients, as were Holocene provinces (FAUNMAP Working Group 1996). The Pleistocene provinces differed because there were as many as twice the number of species in any given area during the Pleistocene as are found at present (Semken 1988).

The extent of North American mammalian faunal provinces during the Pliocene through middle Pleistocene has not been explored in detail (Fejfar and Repenning 1992; Repenning 1987, 1992; Repenning et al. 1995; Bell 2000). The Holarctic arvicoline faunal regions proposed by Fejfar and Repenning (1992) were based on overall faunal similarity across large geographic areas, but the possibility that the regions could be subdivided into smaller faunal provinces was discussed briefly. Five North American arvicoline faunal regions were proposed by Fejfar and Repenning (1992). The Beringian faunal region is in the extreme northwest portion of the continent. The Canadian and Mexican faunal regions essentially follow geopolitical boundaries but were postulated on the basis of latitudinal climatic differences. The eastern United States and western United States faunal regions are divided by the Rocky Mountains (the western United States region was renamed the Pacific faunal region by Repenning et al. 1995). The temporal duration and spatial extent of these faunal regions was not stated explicitly, but the authors implied that the regions were biogeographically pertinent throughout the Blancan and Irvingtonian mammal ages. A preliminary map illustrating the possible geographic limits of these arvicoline regions was provided by Bell (2000), but further critical evaluation of these limits (and their potential change through time) is needed for all relevant time intervals.

The problem is perhaps best illustrated by the complicated state of affairs surrounding current proposed definitions of the Irvingtonian mammal age. At least five operational definitions have been proposed for the base of the Irvingtonian. The differences between them stem from disagreement as to potential defining taxa and their apparently diachronous regional appearances. Three of these definitions were proposed with explicit recognition of diachrony for the beginning of the Irvingtonian on either side of the Rocky Mountains (Repenning 1992; Repenning et al. 1995). The potential complications of this situation are not adequately addressed in the litera-



ture (see Repenning et al. 1995; Bell 2000). The presence of faunal provinces with shifting boundaries throughout the Pliocene and Pleistocene raises some fundamental issues regarding the concept of mammal ages, usually considered to be continentwide in scale. In cases where provincial differences include taxa that are considered to define a mammal age, reevaluation of the temporal divisions of the age may be needed. If a mammal age is defined by the appearance of a single taxon that disperses slowly or unevenly, the base of that mammal age will be time-transgressive. A similar argument pertains to the termination of the Rancholabrean mammal age, the upper boundary of which is not defined by a subsequent mammal age but is based instead on the latest occurrence of large-bodied extinct mammals, a similarly time-transgressive phenomenon. The discussions in this chapter pertaining to the appearance of *Bison* in North America and the late occurrence of *Mammuthus* on Wrangel Island provide good examples. If provinciality in the past was as well marked as in the present, no single taxon may be suitable to define an isochronous boundary in all regions; in fact, a taxon well suited to mark a boundary in one region may not appear in another region. Perception of such provinciality led to the explicitly diachronous definitions proposed for the beginning of the Irvingtonian (Repenning 1992; Repenning et al. 1995).

These considerations take on special importance when we consider the proposed divisions of the Pliocene and Pleistocene mammal ages. Temporal divisions proposed by C. Schultz et al. (1978) and subsequently modified by Lundelius et al. (1987) were based on changes in the mammalian faunas of the Great Plains and may not have broad application outside that region. The temporal divisions based on taxonomic composition and stage of evolution of the arvicoline rodents proposed by Repenning (1978, 1980, 1984, 1987; Repenning et al. 1990) were established with an implicit or acknowledged recognition of provinciality.

The provincial nature of North American mammalian distribution is best exemplified by the extant biota and by fossils from the most recent geologic time intervals. The Rancholabrean is unique among the mammal ages because of its recency, the fact that many of the species that characterize the age are extant, and the widespread geographic distribution of Rancholabrean faunas. These factors combine to permit a greater understanding of the complexities of mammalian faunal dynamics, the interplay of biotic and abiotic factors in shaping mammalian community structure, the response of mammalian species to changes in local climate or vegetation, and the range of patterns thus potentially discernible in the fossil record. The opportunity to study these issues is especially great

for the Holocene and the part of the Rancholabrean that falls within the range of radiocarbon dating. Apart from merely providing a ready means of determining precisely the age, or age range, of a fossil deposit, radiocarbon dating may also permit detailed documentation of the rates of change in faunal communities. Well-dated sites also document the positive and negative aspects of time averaging in fossil deposits (Hadly 1999) and provide a temporal perspective on the persistence (or lack thereof) of faunal communities in time and space (e.g., Hadly 1999; Hadly et al. 1998; Hadly and Maurer 2001).

Late Pleistocene and Holocene biogeographic provinces based on differences in North American mammal faunas were proposed by L. Martin and Neuner (1978), Graham (1979), and L. Martin and Hoffmann (1987). Although different names were applied to the provinces, their geographic boundaries were remarkably similar. The most comprehensive study of Quaternary faunal provinces in North America is that of the FAUNMAP Working Group (1996), which was based on a larger number of taxa than was the case with previous studies. The FAUNMAP database recorded more than 2500 faunas from the late Pleistocene through the late Holocene in the contiguous United States. This database makes it possible to investigate changes in geographic distributions of species in greater detail than for any other period of time (FAUNMAP Working Group 1994, 1996). Shifts in the geographic distribution of species apparently took place individually, resulting in changes in overall community composition throughout the time interval covered by the FAUNMAP database. These data can be used for a preliminary evaluation of the effect of provinciality on biochronology. Examination of the FAUNMAP data set revealed that provincial boundaries at a given time were based on faunal aggregates of various species, but the aggregates on which the provinces were based were different at the two time intervals examined (late Pleistocene and late Holocene). A more robust (denser and richer) record of faunas from relevant time intervals across purported faunal regions must be obtained before reliable provincial definitions of mammal ages (e.g., Fejfar and Repenning 1992; Repenning 1992; Repenning et al. 1995) can be accepted for earlier time intervals (e.g., Irvingtonian and Blancan).

The faunal regions proposed by Fejfar and Repenning (1992) were based on preliminary examination of faunas from throughout the Holarctic and were erected based primarily on perceived provinciality in the arvicoline fauna. For the North American regions (see map in Bell 2000) we accept that the Beringian faunal region is largely valid (for the mammalian fauna generally, not just the

arvicolines), but its southern boundary is not defined in time or space. Our use of the 55°N latitude line is an attempt to provide a southern boundary (admittedly arbitrary). The Canadian region is essentially a data-pauperate zone for most of the Pliocene and Pleistocene and therefore should be abandoned. The western and eastern United States boundary must be carefully reevaluated. Comparisons of approximately contemporaneous faunas in Texas, the Great Plains, Arizona, New Mexico, and California will be needed to evaluate the temporal and spatial nature of that boundary. Many important Blancan faunas are situated in Arizona and New Mexico, where the distinction between the eastern and western United States regions is unclear (see map in Bell 2000). The validity of that boundary also must be tested for the Irvingtonian; preliminary analysis suggests that its recognition, based only on arvicolines during that time interval, may be premature. Two of the postulated differences between eastern and western faunas involve *Allophaiomys* and *Microtus paroperarius*, taxa that previously were not known west of the Rocky Mountains but are now both known from Porcupine Cave in Colorado (central Rockies), the Little Dell Dam faunas (near Salt Lake City), and Cathedral Cave (east-central Great Basin).

Limits of the Mexican faunal region are not well established or characterized. It was originally postulated on the basis of latitudinal climatic differences rather than faunal character (Fejfar and Repenning 1992). Florida faunas were included in the Mexican region by Bell (2000), but this must be reevaluated. Florida has long been known to contain a unique combination of northern, western, tropical, and endemic taxa (Webb and Wilkins 1984). The FAUNMAP data confirm that Floridian faunas were distinct in the late Pleistocene and Holocene. The similarity between Florida faunas and those in Texas during the late Pleistocene may reflect an effect of the Gulf of Mexico coastal plain, either climatic or simply due to greater subaerial exposure during periods of lowered sea level. Texas coastal plain faunas from earlier time intervals are unknown.

The FAUNMAP Working Group (1996) recognized eight fine-scale provinces for the late Holocene, grouped under two major geographic divisions (essentially breaking out along the hundredth meridian). Geographic limits of faunal provinces in the late Pleistocene and Holocene were similar, despite different species composition and community organization. The study of late Holocene fossil faunas provides the key data for evaluating the complexities of faunal dynamics as they appear in the fossil record because of their proximity in time to the modern biota and their preservation in depositional

contexts similar to those of older deposits. With extensive and detailed radiocarbon control and fine-scale stratigraphic excavation it is possible to tease apart details of morphologic and geographic response of mammals to climatic changes of varying intensities and durations. Studies of this nature, conducted by E. Hadly in Yellowstone National Park, provide a valuable insight into the dynamics of mammal communities and their response to short-term climatic changes (E. Barnosky 1994; Hadly 1996, 1997, 1999; Hadly et al. 1998; Hadly and Maurer 2001).

## SUMMARY

Approximately the last 5 million years of mammalian evolution are encompassed by three mammal ages, named, from oldest to youngest, the Blancan, Irvingtonian, and Rancholabrean.

Because of their young age and the application of a variety of dating methods independent of faunal composition, these mammal ages are among the best dated in North America. Close phylogenetic affinities with the extant mammalian fauna permit detailed evaluation of faunal dynamics over short time intervals. Mammalian response, especially geographic range adjustments, to climatic changes over the last 2 million years was used extensively in prior correlations of these North American faunas. The advent of independent dating and correlation techniques (such as radiometric dating of volcanic deposits and the development of the Geomagnetic Polarity Time Scale) provide a more reliable basis for establishing age relationships of these faunas. Climatic correlations of faunas from the late Pleistocene and Holocene are still commonly applied, but their application for earlier time intervals is discouraged.

Pronounced faunal provinciality in the extant, Holocene, and late Pleistocene mammalian fauna is well documented. There is now sufficient evidence to demonstrate that Pleistocene faunas from northern latitudes in North America are of a different overall character than their temporal equivalents at lower latitudes, and similar patterns now are emerging for Pliocene faunas. Mammal ages that are defined by faunas at lower latitudes are rendered effectively inoperable at higher latitudes, where independent biochronologies should be developed. For these reasons, we recommend that the terms *Blancan*, *Irvingtonian*, and *Rancholabrean* be restricted to faunas in North America south of 55°N latitude. Finer-scale faunal provincialism for the Blancan and Irvingtonian was pro-

posed nearly a decade ago but has yet to be demonstrated unequivocally in terms of both faunal character and the recognition of provincial boundaries through time and space. This is a fertile area for future research on Pliocene and Pleistocene biostratigraphy and biochronology.

The Blancan is defined by the first appearance in North America south of 55°N latitude of the arvicoline rodent *Mimomys* or the *Mimomys*-like arvicolines *Ogmodontomys* and *Ophiomys*. The beginning of the Blancan now lies between 5.2 and 4.6 Ma, depending on the arvicoline species used.

The Irvingtonian mammal age is defined by the first appearance in North America south of 55°N latitude of the elephant *Mammuthus*. The earliest records of *Mammuthus* with external chronologic control indicate that mammoths appear between 1.25 and 1.36 Ma across most of North America south of 55°N latitude, from California to Florida. An age of 1.35 is accepted here as the lower temporal limit of the Irvingtonian as it is currently understood. Following the adoption of *Mammuthus* as the defining taxon for the Irvingtonian, no fewer than 14 localities that were previously considered early Irvingtonian are here considered latest Blancan. These faunas include Inglis 1A, Inglis 1C, Haile 16A, and De Soto Shell Pit in Florida; Nash, Aries A, Rick Forester, and Aries B in Kansas; Sappa in Nebraska; Java in South Dakota; Curtis Ranch in Arizona; Froman Ferry in Idaho; El Casco in California; and the lower portion of the Cedazo Fauna in Mexico.

The Rancholabrean mammal age is defined by the first appearance in North America south of 55°N latitude of the bovid *Bison*. Well-dated early records of *Bison* are rare, and although several publications suggest the presence of *Bison* in the United States before 160 ka, we accept the Jones Spring, Missouri (160 ka), and American Falls, Idaho (between 210 and 72 ka), records as the earliest well-dated *Bison*. No mammal age younger than Rancholabrean is established, but the megafaunal extinction near the end of the Pleistocene represents a significant faunal change that we use to mark the termination of the Rancholabrean and the long sequence of North American mammal ages. The youngest reliable radiocarbon date from extinct megafauna in North America south of 55°N latitude could be as young as 9.5 ka, and that age is used here to mark the end of the Rancholabrean.

Divisions of the Blancan and Irvingtonian mammal ages permit finer-scale faunal correlations and temporal resolution for much of the Pliocene and Pleistocene. Proposals to divide the mammal ages in finer units include one based on overall faunal character and two

based on the evolutionary history of the arvicoline rodents. We recognize some, but not all, of the proposed divisions.

The Miocene–Pliocene and Pliocene–Pleistocene boundary stratotypes are now formally designated in Italy, permitting evaluation of the correspondence of North America mammal ages with the epochal boundaries. The base of the Pliocene is 5.335 Ma, and the base of the Pleistocene is slightly younger than 1.77 Ma. The Holocene has no formal boundary definition, but its beginning is here operationally recognized at 10 ka. Thus the Blancan mammal age as defined here is mostly Pliocene, but faunas from the latest Blancan are early Pleistocene in age. The Irvingtonian as defined here is entirely Pleistocene. The Rancholabrean is mostly Pleistocene in age, but its terminal phase extends approximately 500 years into the early Holocene.

#### ACKNOWLEDGMENTS

*A project of this scope is always more collaborative than authorship reflects. We are grateful to all of our colleagues who share our interest in the latest Cenozoic mammal ages and on whose work we drew so heavily in preparing this chapter. We are especially grateful to Charles Repenning for stimulating conversations, for helping us to clarify our thinking and our prose, and for his devotion to the science of stratigraphic paleontology. Our job was made easier by the assistance of Bill Akersten, Arthur Bettis, Nick Czaplewski, Walt Dalquest, Ismael Ferrusquia-Villafranca, Fred Grady, Elizabeth Hadly, Jason Head, Richard Hulbert, Robert Martin, Jim Mead, Gary Morgan, Charles Repenning, Amos Salvador, Jeff Saunders, Gerald Schultz, Eric Scott, Kevin Seymour, Richard Tedford, James Vanderhill, and Alisa Winkler.*

*The manuscript was directly improved by constructive criticism from Barry Albright, Judy Lundelius, Jim Mead, Charles Repenning, and Geraldine Swartz; they do not necessarily agree with our conclusions, but their input was valuable and is appreciated. Mike Woodburne responded graciously when we missed our first deadline. We are especially grateful to Dennis Trombatore, Jim McCulloch, and Melissa Van Ostran in the Geology Library at the University of Texas at Austin for their enduring enthusiasm, tolerance, and assistance during all phases of this project. Jeffrey Horowitz drafted the digital figures.*

*We were saddened by the recent death of Elaine Anderson, whose contributions to Pliocene and Pleistocene paleomammalogy helped to shape the science in the later part of the twentieth century. It is to her memory that we dedicate this work.*

#### REFERENCES

- Ahearn, M. E. and J. F. Lance. 1980. A new species of *Nechoeris* (Rodentia: Hydrochoeridae) from the Blancan (Late Pliocene) of North America. *Proceedings of the Biological Society of Washington* 93:435–442.

- Akersten, W. A. 1972. Red Light Local Fauna (Blancan) of the Love Formation, southeastern Hudspeth County, Texas. *Bulletin of the Texas Memorial Museum* 20:1–53.
- Albright, L. B. III. 1999. Biostratigraphy and vertebrate paleontology of the San Timoteo Badlands, southern California. *University of California Publications in Geological Sciences* 144:1–121, Plates 1–8.
- Alford, J. J. 1974. The geography of mastodon extinction. *The Professional Geographer* 26:425–429.
- Alroy, J. 1999. Putting North America's end-Pleistocene megafaunal extinction in context: Large-scale analyses of spatial patterns, extinction rates, and size distributions. In *Extinctions in near time: Causes, contexts, and consequences*, ed. R. D. E. MacPhee. New York: Kluwer Academic/Plenum Publishers, pp. 105–143.
- . 2001. A multispecies overkill simulation of the end-Pleistocene megafaunal mass extinction. *Science* 292:1893–1896.
- Anderson, E. 1996. A preliminary report on the Carnivora of Porcupine Cave, Park County, Colorado. In *Palaeoecology and palaeoenvironments of late Cenozoic mammals: Tributes to the career of C. S. (Rufus) Churcher*, ed. K. M. Stewart and K. L. Seymour. Toronto: University of Toronto Press, pp. 259–282.
- Arellano, A. R. V. and E. Azcón. 1949. Pre-*Equus* horses from Goleta (Morelia) Michoacan, Mexico. *Bulletin of the Geological Society of America* 60:1871.
- Armstrong, R. L., W. P. Leeman, and H. E. Malde. 1975. K–Ar dating, Quaternary and Neogene volcanic rocks of the Snake River Plain, Idaho. *American Journal of Science* 275:225–251.
- Arslanov, K. A., G. T. Cook, S. Gulliksen, D. D. Harkness, T. Kankainen, E. M. Scott, S. Vartanyan, and G. I. Zaitseva. 1998. Consensus dating of mammoth remains from Wrangel Island. *Radiocarbon* 40:289–294.
- Atwood, W. W. 1940. *The physiographic provinces of North America*. Boston: Ginn and Company.
- Axelrod, D. I. 1967. Quaternary extinctions of large mammals. *University of California Publications in Geological Sciences* 74:1–42.
- Azzaroli, A. 1995. A synopsis of the Quaternary species of *Equus* in North America. *Bollettino della Società Paleontologica Italiana* 34:205–221.
- Azzaroli, A. and P. Ambrosetti. 1970. Late Villafranchian and early mid-Pleistocene faunas in Italy. *Palaeogeography, Palaeoclimatology, Palaeoecology* 8:107–111.
- Azzaroli, A. and A. Berzi. 1970. On an upper Villafranchian fauna at Imola, northern Italy, and its correlation with the marine Pleistocene sequence of the Po plain. *Palaeontographia Italica* 66:1–12.
- Azzaroli, A., M. L. Colalongo, H. Nakagawa, G. Pasini, D. Rio, G. Ruggieri, S. Sartoni, and R. Sprovieri. 1997. The Pliocene–Pleistocene boundary in Italy. In *The Pleistocene boundary and the beginning of the Quaternary*, ed. J. A. Van Couvering. New York: Cambridge University Press, pp. 141–155.
- Azzaroli, A. and M. R. Voorhies. 1993. The genus *Equus* in North America. The Blancan species. *Palaeontographia Italica* 80:175–198.
- Bachman, G. O. and H. H. Mehnert. 1978. New K–Ar dates and the late Pliocene to Holocene geomorphic history of the central Rio Grande region, New Mexico. *Geological Society of America Bulletin* 89:283–292.
- Bacon, C. R., D. M. Giovannetti, W. A. Duffield, and G. B. Dalrymple. 1979. New constraints on the age of the Coso Formation, Inyo County, California. *Geological Society of America Abstracts with Programs* 11:67.
- Barendregt, R. W., E. Irving, E. A. Christiansen, E. K. Sauer, and B. T. Schreiner. 1998. Stratigraphy and paleomagnetism of late Pliocene and Pleistocene sediments in the Wellsch Valley and Swift Current Creek areas, southwestern Saskatchewan, Canada. *Canadian Journal of Earth Sciences* 35:1347–1361.
- Barendregt, R. W., F. F. Thomas, E. Irving, J. Baker, A. M. Stalker, and C. S. Churcher. 1991. Stratigraphy and paleomagnetism of the Jaw Face section, Wellsch Valley site, Saskatchewan. *Canadian Journal of Earth Sciences* 28:1353–1364.
- Barnosky, A. D. 1987. Punctuated equilibrium and phyletic gradualism: Some facts from the Quaternary mammalian record. In *Current mammalogy*, Vol. 1, ed. H. H. Genoways. New York: Plenum, pp. 109–147.
- . 1989. The late Pleistocene event as a paradigm for widespread mammal extinction. In *Mass extinctions: Processes and evidence*, ed. S. K. Donovan. New York: Columbia University Press, pp. 235–254.
- . 1993. Mosaic evolution at the population level in *Microtus pennsylvanicus*. In *Morphological change in Quaternary mammals of North America*, ed. R. A. Martin and A. D. Barnosky. New York: Cambridge University Press, pp. 24–59.
- Barnosky, A. D. and C. J. Bell. In press. Spatial distribution and taphonomic overview of vertebrate fossils in Porcupine Cave. In *Biodiversity response to climate change in the middle Pleistocene: The Porcupine Cave Fauna from Colorado*, ed. A. D. Barnosky. Berkeley: University of California Press.
- Barnosky, A. D. and D. L. Rasmussen. 1988. Middle Pleistocene arvicoline rodents and environmental change at 2900-meters elevation, Porcupine Cave, South Park, Colorado. *Annals of Carnegie Museum* 57:267–292.
- Barnosky, A. D., T. I. Rouse, E. A. Hadly, D. L. Wood, F. L. Keesing, and V. A. Schmidt. 1996. Comparison of mammalian response to glacial–interglacial transitions in the middle and late Pleistocene. In *Palaeoecology and palaeoenvironments of late Cenozoic mammals: Tributes to the career of C. S. (Rufus) Churcher*, ed. K. M. Stewart and K. L. Seymour. Toronto: University of Toronto Press, pp. 16–33.
- Barnosky, E. H. 1994. Ecosystem dynamics through the past 2000 years as revealed by fossil mammals from Lamar Cave in Yellowstone National Park, USA. *Historical Biology* 8:71–90.
- Bassett, M. G. 1985. Towards a “common language” in stratigraphy. *Episodes* 8:87–92.
- Bayne, C. K. 1976. Early and medial Pleistocene faunas of Meade County, Kansas. In Guidebook 24th annual meeting, Midwestern Friends of the Pleistocene: Stratigraphy and faunal sequence—Meade County, Kansas, May 22–23, 1976, by C. K. Bayne. The University of Kansas, Kansas Geological Survey, Guidebook Series 1:1–25.
- Beck, M. W. 1996. On discerning the cause of late Pleistocene megafaunal extinctions. *Paleobiology* 22:91–103.
- Bell, C. J. 1995. A middle Pleistocene (Irvingtonian) microtine rodent fauna from White Pine County, Nevada, and its implications for microtine rodent biochronology. *Journal of Vertebrate Paleontology* 15(suppl. 3):18A.
- . 2000. Biochronology of North American microtine rodents. In *Quaternary geochronology: Methods and applications*, ed. J. S. Noller, J. M. Sowers, and W. R. Lettis. AGU Reference Shelf 4. Washington, DC: American Geophysical Union, pp. 379–406.
- Bell, C. J. and A. D. Barnosky. 2000. The microtine rodents from the Pit locality in Porcupine Cave, Park County, Colorado. *Annals of Carnegie Museum* 69:93–134.

- Bell, C. J. and J. Gauthier. 2002. North America Quaternary Squamata: Re-evaluation of the stability hypothesis. *Journal of Vertebrate Paleontology* 22(suppl. 3):35A.
- Bell, C. J. and C. A. Repenning. 1999. Observations on dental variation in *Microtus* from the Cudahy Ash Pit Fauna, Meade County, Kansas and implications for Irvingtonian microtine rodent biochronology. *Journal of Vertebrate Paleontology* 19:757–766.
- Bender, M. L. 1973. Helium–uranium dating of corals. *Geochimica et Cosmochimica Acta* 37:1229–1247.
- Bennett, K. D., S. G. Haberle, and S. H. Lumley. 2000. The last glacial–Holocene transition in southern Chile. *Science* 290:325–328.
- Bennike, O. 1997. Quaternary vertebrates from Greenland: A review. *Quaternary Science Reviews* 16:899–909.
- Berggren, W. A. and J. A. Van Couvering. 1974. The late Neogene: Biostratigraphy, geochronology and paleoclimatology of the last 15 million years in marine and continental sequences. *Palaeogeography, Palaeoclimatology, Palaeoecology* 16:1–216.
- Berggren, W. A., D. V. Kent, C. C. Swisher III, and M.-P. Aubry. 1995. A revised Cenozoic geochronology and chronostratigraphy. In *Geochronology, time scales and global stratigraphic correlation*, ed. W. A. Berggren, D. V. Kent, and M.-P. Aubry. *SEPM (Society for Sedimentary Geology) Special Publication* 54:129–212.
- Berta, A. 1985. The status of *Smilodon* in North and South America. *Natural History Museum of Los Angeles County Contributions in Science* 370:1–15.
- . 1987. The sabercat *Smilodon gracilis* from Florida and a discussion of its relationships (Mammalia, Felidae, Smilodontini). *Bulletin of the Florida State Museum, Biological Sciences* 31:1–63.
- . 1995. Fossil carnivores from the Leisey Shell Pits, Hillsborough County, Florida. In *Paleontology and geology of the Leisey Shell Pits, early Pleistocene of Florida*, ed. R. C. Hulbert Jr., G. S. Morgan, and S. D. Webb. *Bulletin of the Florida Museum of Natural History* 37(Part II):463–499.
- Berta, A. and H. Galiano. 1983. *Megantereon hesperus* from the late Hemphillian of Florida with remarks on the phylogenetic relationships of machairodonts (Mammalia, Felidae, Machairodontinae). *Journal of Paleontology* 57:892–899.
- Bjork, P. R. 1970. The Carnivora of the Hagerman Local Fauna (late Pliocene) of southwestern Idaho. *Transactions of the American Philosophical Society* 60(7):1–54.
- Blair, W. F. 1958. Distributional patterns of vertebrates in the Southern United States in relation to past and present environments. In *Zoogeography*, ed. C. L. Hubbs. Washington, DC: American Association for the Advancement of Science Publication 51:433–468.
- Boellstorff, J. D. 1973. Fission-track ages of Pleistocene volcanic ash deposits in the central plains, U.S.A. *Isochron/West* 8:39–43.
- . 1976. The succession of late Cenozoic volcanic ashes in the Great Plains: A progress report. In *Guidebook: 24th annual meeting, Midwestern Friends of the Pleistocene: Stratigraphy and faunal sequence—Meade County, Kansas, May 22–23, 1976*, by C. K. Bayne. Kansas Geological Survey Guidebook Series 1:37–71.
- . 1978. North American Pleistocene stages reconsidered in light of probable Pliocene–Pleistocene continental glaciation. *Science* 202:305–307.
- Bressler, S. L. and R. F. Butler. 1978. Magnetostratigraphy of the late Tertiary Verde Formation, central Arizona. *Earth and Planetary Science Letters* 38:319–330.
- Brown, B. 1908. The Conard Fissure, a Pleistocene bone deposit in northern Arkansas: With descriptions of two new genera and twenty new species of mammals. *Memoirs of the American Museum of Natural History* 9:155–208, Plates 13–24.
- Bryson, R. A., D. A. Baerreis, and W. M. Wendland. 1970. The character of late-glacial and post-glacial climatic changes. In *Pleistocene and Recent environments of the central Great Plains*, ed. W. Dort Jr. and J. K. Jones Jr. Department of Geology, University of Kansas Special Publication 3. Lawrence: University of Kansas Press, pp. 53–74.
- Cande, S. C. and D. V. Kent. 1995. Revised calibration of the geomagnetic polarity timescale for the Late Cretaceous and Cenozoic. *Journal of Geophysical Research* 100(B4):6093–6095.
- Carleton, M. J. and G. G. Musser. 1984. Muroid rodents. In *Orders and families of Recent mammals of the world*, ed. S. Anderson and J. K. Jones Jr. New York: John Wiley & Sons, Pp. 289–379.
- Carranza-C., O. and W. E. Miller. 1980. The earliest capybara record in North America. *Geological Society of America Abstracts with Programs* 12:399.
- Carranza-Castañeda, O. and I. Ferrusquía-Villafranca. 1978. Nuevas investigaciones sobre la fauna Rancho El Ocote, Plioceno medio de Guanajuato, Mexico: Informe preliminar. *Universidad Nacional Autónoma de México, Instituto de Geología, Revista* 2:163–166.
- Carranza-Castañeda, O. and W. E. Miller. 1988. Roedores caviomorfos de la Mesa Central de México, Blancano temprano (Plioceno tardío) de la fauna local Rancho Viejo, Estado de Guanajuato. *Universidad Nacional Autónoma de México, Instituto de Geología, Revista* 7:182–199.
- . 1996. Hemphillian and Blancan felids from central Mexico. *Journal of Paleontology* 70:509–518.
- . 1998. Paleofaunas de vertebrados de las cuencas sedimentarias del Terciario tardío de la Faja Volcánica Transmexicana. In *Avances en investigación, paleontología de vertebrados*, ed. O. Carranza Castañeda and D. A. Córdoba Méndez. *Universidad Autónoma del Estado de Hidalgo, Instituto de Investigaciones en Ciencias de la Tierra, Publicación Especial* 1:85–95.
- . 2000. Selected late Cenozoic vertebrate localities in the State of Hidalgo, and Guanajuato, central Mexico. In *Guide book of the Field Trips, Society of Vertebrate Paleontology 60nd [sic] Annual Meeting, Universidad Nacional Autónoma de México, Mexico City, 2000. Avances en Investigación, Universidad Autónoma del Estado de Hidalgo, Ciudad Universitaria, Pachuca, Hgo. Special Publication* 3:3–47.
- Carranza-Castañeda, O. and A. H. Walton. 1992. Cricetid rodents from the Rancho El Ocote Fauna, late Hemphillian (Pliocene), state of Guanajuato. *Universidad Nacional Autónoma de México, Instituto de Geología, Revista* 10:71–93.
- Carrasco, M. A. 2000. Species discrimination and morphological relationships of kangaroo rats (*Dipodomys*) based on their dentition. *Journal of Mammalogy* 81:107–122.
- Carraway, L. N. 1990. A morphologic and morphometric analysis of the “*Sorex vagrans* species complex” in the Pacific Coast region. *Special Publications, The Museum, Texas Tech University* 32:1–76.
- . 1995. A key to recent Soricidae of the western United States and Canada based primarily on dentaries. *Occasional Papers of the Natural History Museum, The University of Kansas* 175:1–49.
- Cassiliano, M. L. 1999. Biostratigraphy of Blancan and Irvingtonian mammals in the Fish Creek–Vallecito Creek section, south-

- ern California, and a review of the Blancan–Irvingtonian boundary. *Journal of Vertebrate Paleontology* 19:169–186.
- Chaline, J. 1987. Arvicolid data (Arvicolidae, Rodentia) and evolutionary concepts. *Evolutionary Biology* 21:237–310.
- Churcher, C. S. 1984a. Faunal correlations of Pleistocene deposits in western Canada. In *Correlation of Quaternary chronologies*, ed. W. C. Mahaney. Norwich, England: Geo Books, pp. 145–158.
- . 1984b. The status of *Smilodontopsis* (Brown, 1908) and *Ischyrosmilus* (Merriam, 1918). A taxonomic review of two genera of sabretooth cats (Felidae, Machairodontinae). *Royal Ontario Museum Life Sciences Contributions* 140:1–59.
- Cita, M. B. 1975. The Miocene/Pliocene boundary: History and definition. In *Late Neogene epoch boundaries. Symposium on Late Neogene Epoch Boundaries, 24th International Geological Congress, Montreal, 21–30 August 1972*, ed. T. Saito and L. H. Burckle. Micropaleontology Special Publication Number 1. New York: Micropaleontology Press, American Museum of Natural History, pp. 1–30.
- Coates, A. G. and J. A. Obando. 1996. The geologic evolution of the Central American Isthmus. In *Evolution & environment in tropical America*, ed. J. B. C. Jackson, A. F. Budd, and A. G. Coates. Chicago: University of Chicago Press, pp. 21–56.
- Conrad, G. S. 1980. The biostratigraphy and mammalian paleontology of the Glenns Ferry Formation from Hammett to Oreana, Idaho. Unpublished Ph.D. dissertation, Idaho State University, Pocatello.
- Conroy, C. J. and J. A. Cook. 2000. Molecular systematics of a Holarctic rodent (*Microtus*: Muridae). *Journal of Mammalogy* 81:344–359.
- Cope, E. D. 1871. Preliminary report on the Vertebrata discovered in the Port Kennedy Bone Cave. *Proceedings of the American Philosophical Society Held at Philadelphia for Promoting Useful Knowledge* 12:73–102.
- . 1893. A preliminary report on the vertebrate paleontology of the Llano Estacado. In *Fourth annual report of the Geological Survey of Texas, 1892, Part II, Paleontology and natural history*, E. T. Dumble, state geologist. Austin: Ben C. Jones & Co., pp. 1–136, Plates 1–23.
- Corbet, G. B. 1978. *The mammals of the Palaearctic region: A taxonomic review*. London: British Museum (Natural History).
- Cox, A., R. R. Doell, and G. B. Dalrymple. 1963. Geomagnetic polarity epochs and Pleistocene geochronometry. *Nature* 198:1049–1051.
- Cronin, T. M. and H. J. Dowsett. 1996. Biotic and oceanographic response to the Pliocene closing of the Central American Isthmus. In *Evolution & environment in tropical America*, ed. J. B. C. Jackson, A. F. Budd, and A. G. Coates. Chicago: The University of Chicago Press, pp. 76–104.
- Curry, B. B. and M. J. Pavich. 1996. Absence of glaciation in Illinois during marine isotope stages 3 through 5. *Quaternary Research* 46:19–26.
- Czaplewski, N. J. 1987. Middle Blancan vertebrate assemblage from the Verde Formation, Arizona. *Contributions to Geology, The University of Wyoming* 25:133–155.
- . 1990. The Verde Local Fauna: Small vertebrate fossils from the Verde Formation, Arizona. *San Bernardino County Museum Association Quarterly* 37(3):1–39.
- Czaplewski, N. J., J. I. Mead, C. J. Bell, W. D. Peachey, and T.-L. Ku. 1999a. Papago Springs Cave revisited, Part II: Vertebrate paleofauna. *Occasional Papers of the Oklahoma Museum of Natural History* 5:1–41.
- Czaplewski, N. J., W. D. Peachey, J. I. Mead, T.-L. Ku, and C. J. Bell. 1999b. Papago Springs Cave revisited, Part I: Geologic setting, cave deposits, and radiometric dates. *Occasional Papers of the Oklahoma Museum of Natural History* 3:1–25.
- Daeschler, E., E. E. Spamer, and D. C. Parris. 1993. Review and new data on the Port Kennedy Local Fauna and Flora (late Irvingtonian), Valley Forge National Historical Park, Montgomery County, Pennsylvania. *The Mosasaur* 5:23–41.
- Dalquest, W. W. 1965. New Pleistocene formation and local fauna from Hardeman County, Texas. *Journal of Paleontology* 39:63–79.
- . 1967. Mammals of the Pleistocene Slaton Local Fauna of Texas. *The Southwestern Naturalist* 12:1–30.
- . 1975. Vertebrate fossils from the Blanco Local Fauna of Texas. *Occasional Papers, The Museum, Texas Tech University* 30:1–52.
- . 1977. Mammals of the Holloman Local Fauna, Pleistocene of Oklahoma. *The Southwestern Naturalist* 22:255–268.
- . 1978. Early Blancan mammals of the Beck Ranch Local Fauna of Texas. *Journal of Mammalogy* 59:269–298.
- Dalquest, W. W. and R. M. Carpenter. 1988. Early Pleistocene (Irvingtonian) mammals from the Seymour Formation, Knox and Baylor counties, Texas, exclusive of Camelidae. *Occasional Papers, The Museum, Texas Tech University* 124:1–28.
- Dalquest, W. W. and O. Mooser. 1980. Late Hemphillian mammals of the Ocote Local Fauna, Guanajuato, Mexico. *The Pearce-Sellards Series (Texas Memorial Museum)* 32:1–25.
- Dalquest, W. W. and G. E. Schultz. 1992. *Ice age mammals of northwestern Texas*. Wichita Falls, TX: Midwestern State University Press.
- de Jong, J. D. 1965. Opening notes and final resolutions of the Subcommittee for the Study of the Holocene. In *International Association on Quaternary Research, Report of the VIth International Congress on Quaternary, Warsaw 1961, Vol. I, Report of the secretary general, plenary sessions, commissions and subcommissions*, ed. J. Dylik, J. Kondracki, and S. Krajewski. Łódź, Poland, pp. 393–396.
- Delcourt, P. A. 1985. The influence of late-Quaternary climatic and vegetational change on paleohydrology in unglaciated eastern North America. *Ecologia Mediterranea* 11:17–26.
- de Queiroz, K. 1992. Phylogenetic definitions and taxonomic philosophy. *Biology and Philosophy* 7:295–313.
- Dickson, J. J. and G. A. Izett. 1981. Fission-track ages of air-fall tuffs in Pliocene basin-fill sediments near 111 Ranch, Graham County, Arizona. *Isochron/West* 32:13–15.
- Downs, T. and G. J. Miller. 1994. Late Cenozoic equids from the Anza-Borrego Desert of California. *Natural History Museum of Los Angeles County Contributions in Science* 440:1–90.
- Downs, T. and J. A. White. 1968. A vertebrate faunal succession in superposed sediments from late Pliocene to middle Pleistocene in California. In *International Geological Congress report of the twenty-third session, Czechoslovakia, 1968. Proceedings of Section 10: Tertiary/Quaternary boundary*, ed. J. Tejkal. Prague: Academia, pp. 41–47.
- Dreimanis, A. 1968. Extinction of mastodons in eastern North America: Testing a new climatic–environmental hypothesis. *The Ohio Journal of Science* 68(6):257–272.
- Dundas, R. G. 1999. Quaternary records of the dire wolf, *Canis dirus*, in North and South America. *Boreas* 28:375–385.
- Dundas, R. G., R. B. Smith, and K. L. Verosub. 1996. The Fairmead Landfill locality (Pleistocene, Irvingtonian), Madera County,

- California: Preliminary report and significance. In *The uses of vertebrate fossils in biostratigraphic correlation*, ed. C. J. Bell and S. S. Sumida. *PaleoBios* 17(2-4):50-58.
- Edmund, A. G. 1987. Evolution of the genus *Holmesina* (Pamphathiidae, Mammalia) in Florida, with remarks on taxonomy and distribution. *Pearce-Sellards Series* (Texas Memorial Museum) 45:1-20.
- Eger, J. L. 1995. Morphometric variation in the Nearctic collared lemming (*Dicrostonyx*). *Journal of Zoology* (London) 235:143-161.
- Eshelman, R. E. 1975. Geology and paleontology of the early Pleistocene (late Blancan) White Rock fauna from north-central Kansas. Claude W. Hibbard Memorial Volume 4. *University of Michigan, The Museum of Paleontology Papers on Paleontology* 13.
- Eshelman, R. and M. Hager. 1984. Two Irvingtonian (medial Pleistocene) vertebrate faunas from north-central Kansas. In *Contributions in Quaternary vertebrate paleontology: A volume in memorial to John E. Guilday*, ed. H. H. Genoways and M. R. Dawson. *Carnegie Museum of Natural History Special Publication* 8:384-404.
- Eshelman, R. E. and C. W. Hibbard. 1981. Nash Local Fauna (Pleistocene: Aftonian) of Meade County, Kansas. *Contributions from the Museum of Paleontology, The University of Michigan* 25:317-326.
- Evans, G. L. and G. E. Meade. 1945. Quaternary of the Texas High Plains. *Contributions to Geology, 1944: The University of Texas Publication* 4401:485-507, Plate 47.
- Evernden, J. F., D. E. Savage, G. H. Curtis, and G. T. James. 1964. Potassium-argon dates and the Cenozoic mammalian chronology of North America. *American Journal of Science* 262:145-198.
- Fairbridge, R. W. 1974. A regional lower boundary stratotype for the Holocene in Quebec. In *American Quaternary Association Abstracts of the Third Biennial Meeting, July 30-August 1, 1974*, University of Wisconsin-Madison, p. 108.
- Farlow, J. O., J. A. Sunderman, J. J. Havens, A. L. Swinehart, J. A. Holman, R. L. Richards, N. G. Miller, R. A. Martin, R. M. Hunt Jr., G. W. Storrs, B. B. Curry, R. H. Fluegeman, M. R. Dawson, and M. E. T. Flint. 2001. The Pipe Creek Sinkhole biota, a diverse late Tertiary continental fossil assemblage from Grant County, Indiana. *American Midland Naturalist* 145:367-378.
- FAUNMAP Working Group. 1994. FAUNMAP: A database documenting late Quaternary distributions of mammal species in the United States. *Illinois State Museum Scientific Papers* 25:1-690.
- . 1996. Spatial response of mammals to late Quaternary environmental fluctuations. *Science* 272:1601-1606.
- Fejfar, O. and W.-D. Heinrich (eds.). 1990. *International symposium: Evolution, phylogeny and biostratigraphy of arviculids (Rodentia, Mammalia)*. Prague: Geological Survey.
- Fejfar, O., W.-D. Heinrich, and E. H. Lindsay. 1998. Updating the Neogene rodent biochronology in Europe. In *The dawn of the Quaternary: Proceedings of the SEQS-EuroMam Symposium, Kerkrade 16-21 June 1996*, ed. T. van Kolfschoten and P. L. Gibbard. *Mededelingen Nederlands Instituut voor Toegepaste Geowetenschappen TNO* 60:533-553.
- Fejfar, O. and C. A. Repenning. 1992. Holarctic dispersal of the arviculids (Rodentia, Cricetidae). In *Mammalian migration and dispersal events in the European Quaternary*, ed. W. v. Koenigswald and L. Werdelin. *Courier Forschungsinstitut Senckenberg* 153:205-212.
- Ferrusquía-Villafranca, I. 1978. Distribution of Cenozoic vertebrate faunas in middle America and problems of migration between North and South America. In *Conexiones terrestres entre Norte y Sudamerica: Simposio interdisciplinario sobre paleogeografía Mesoamericana*, ed. I. Ferrusquía-Villafranca. *Universidad Nacional Autónoma de México, Instituto de Geología Boletín* 101:193-321.
- Firby, J. B. 1968. *Revision of the middle Pleistocene Irvington Fauna of California*. Unpublished M.A. thesis, University of California at Berkeley.
- Flannery, T. F. 1995. *The future eaters: An ecological history of the Australasian lands and people*. New York: George Braziller.
- Flint, M. E. T. 1998. Evolution of the hind limb and degree of aquatic adaptation in the *Pliopotamys-Ondatra* lineage. Unpublished M.S. thesis, Idaho State University, Pocatello.
- Flint, R. F. 1971. *Glacial and Quaternary geology*. New York: John Wiley & Sons.
- Flynn, L. J. 1997. Late Neogene mammalian events in north China. In *Actes du Congrès Biochron'97, Montpellier, 14-17 Avril: Biochronologie mammalienne du Cénozoïque en Europe et Domaines Reliés*, ed. J.-P. Aguilar, S. Legendre, and J. Michaux. *Mémoires et Travaux de l'Institut de Montpellier* 21:183-192.
- Foley, R. L. and L. E. Raue. 1987. *Lemmus sibiricus* from the late Quaternary of the midwestern United States. *Current Research in the Pleistocene* 4:105-107.
- Forbes, E. 1846. On the connexion between the distribution of the existing fauna and flora of the British Isles, and the geological changes which have affected their area, especially during the epoch of the Northern Drift. *Memoirs of the Geological Survey of Great Britain and of the Museum of Economic Geology in London* 1:336-432.
- Forman, S. L., E. A. Bettis III, T. J. Kemmis, and B. B. Miller. 1992. Chronologic evidence for multiple periods of loess deposition during the Late Pleistocene in the Missouri and Mississippi River Valley, United States: Implications for the activity of the Laurentide Ice Sheet. *Palaeogeography, Palaeoclimatology, Palaeoecology* 93:71-83.
- Frazier, M. K. 1982 ("1981"). A revision of the fossil Erethizontidae of North America. *Bulletin of the Florida State Museum, Biological Sciences* 27:1-76.
- Galusha, T., N. M. Johnson, E. H. Lindsay, N. D. Opdyke, and R. H. Tedford. 1984. Biostratigraphy and magnetostratigraphy, late Pliocene rocks, 111 Ranch, Arizona. *Geological Society of America Bulletin* 95:714-722.
- Ganseccki, C. A., G. A. Mahood, and M. McWilliams. 1998. New ages for the climactic eruptions at Yellowstone: Single-crystal <sup>40</sup>Ar/<sup>39</sup>Ar dating identifies contamination. *Geology* 26:343-346.
- Gazin, C. L. 1942. The late Cenozoic vertebrate faunas from the San Pedro Valley, Ariz. *Proceedings of the United States National Museum* 92:475-518.
- Gervais, P. 1869 ("1867-1869"). *Zoologie et paléontologie générales. Nouvelles recherches sur les animaux vertébrés vivants et fossiles. Première série*. Paris: Arthus Bertrand, Libraire-Éditeur, Libraire de la Société de Géographie, Paris, 263p, Plates 1-50.
- Getz, L. L. 1960. Middle Pleistocene carnivores from southwestern Kansas. *Journal of Mammalogy* 41:361-365.
- Gidley, J. W. 1900. A new species of Pleistocene horse from the Staked Plains of Texas. *Bulletin of the American Museum of Natural History* 13:111-116.
- . 1903a. The fresh-water Tertiary of northwestern Texas. American Museum Expeditions of 1899-1901. *Bulletin of the American Museum of Natural History* 19:617-635, Plates 52-58.

- . 1903b. On two species of *Platygonus* from the Pliocene of Texas. *Bulletin of the American Museum of Natural History* 19:477–481.
- . 1913. Preliminary report on a recently discovered Pleistocene cave deposit near Cumberland, Maryland. *Proceedings of the United States National Museum* 46:93–102.
- . 1920a. A Pleistocene cave deposit of western Maryland. *The Smithsonian Report for 1918*:281–287, Plates 1–6.
- . 1920b. Pleistocene peccaries from the Cumberland Cave deposit. *Proceedings of the United States National Museum* 57:651–678, Plates 54–55.
- . 1922. Preliminary report on fossil vertebrates of the San Pedro Valley, Arizona, with descriptions of new species of Rodentia and Lagomorpha. *U.S. Geological Survey Professional Paper* 131:119–131, Plates 34–35.
- . 1926. Fossil Proboscidea and Edentata of the San Pedro Valley, Arizona. *U.S. Geological Survey Professional Paper* 140:83–95, Plates 32–44.
- Gidley, J. W. and C. L. Gazin. 1933. New Mammalia in the Pleistocene fauna from Cumberland Cave. *Journal of Mammalogy* 14:343–357.
- . 1938. The Pleistocene vertebrate fauna from Cumberland Cave Maryland. *United States National Museum Bulletin* 171:1–99, Plates 1–10.
- Gignoux, M. 1910. Sur la classification du Pliocène et du Quaternaire dans l'Italie du Sud. *Comptes Rendus Hebdomadaires des Séances de l'Académie des Sciences* 150:841–844.
- . 1916 (“1914”). L'Étage Calabrien (Pliocène supérieur marin) sur le versant nord-est de l'Apennine, entre le Monte Gargano et Plaisance. *Compte Rendu Sommaire et Bulletin de la Société Géologique de France*, series 4, 14:324–348.
- Gillette, D. D., C. J. Bell, and M. C. Hayden. 1999. Preliminary report on the Little Dell Dam fauna, Salt Lake County, Utah (middle Pleistocene, Irvingtonian Land Mammal Age). In *Vertebrate paleontology in Utah*, ed. D. D. Gillette. *Utah Geological Survey Miscellaneous Publication* 99-1:495–500.
- Gillette, D. D. and C. E. Ray. 1981. Glyptodonts of North America. *Smithsonian Contributions to Paleobiology* 40:1–255.
- Gingerich, P. D. 1993. Rates of evolution in Plio–Pleistocene mammals: Six case studies. In *Morphological change in Quaternary mammals of North America*, ed. R. A. Martin and A. D. Barnosky. New York: Cambridge University Press, pp. 84–106.
- Golz, D. J., G. T. Jefferson, and M. P. Kennedy. 1977. Late Pliocene vertebrate fossils from the Elsinore Fault Zone, California. *Journal of Paleontology* 51:864–866.
- Goodwin, H. T. 1993. Patterns of dental variation and evolution in prairie dogs, genus *Cynomys*. In *Morphological change in Quaternary mammals of North America*, ed. R. A. Martin and A. D. Barnosky. New York: Cambridge University Press, pp. 107–133.
- Gordon, C. L. 1999. Morphological variation in the dentition of late Pleistocene meadow voles (*Microtus pennsylvanicus*) from Yarbrough Cave, Bartow County, Georgia. *Paludicola* 2:207–231.
- Grady, F. 1988. A preliminary account of the Pleistocene mammals from Patton Cave, Monroe County, West Virginia. *National Speleological Society Bulletin* 50:9–16.
- Graham, M. A., M. C. Wilson, and R. W. Graham. 1987. Paleoenvironments and mammalian faunas of Montana, southern Alberta, and southern Saskatchewan. In *Late Quaternary mammalian biogeography and environments of the Great Plains and Prairies*, ed. R. W. Graham, H. A. Semken Jr., and M. A. Graham. *Illinois State Museum Scientific Papers* 22:410–459.
- Graham, R. W. 1972. Biostratigraphy and paleoecological significance of the Conard Fissure Local Fauna with emphasis on the genus *Blarina*. Unpublished M.S. thesis, University of Iowa, Iowa City.
- . 1979. Paleoclimates and late Pleistocene faunal provinces in North America. In *Pre-Llano cultures of the Americas: Paradoxes and possibilities*, ed. R. L. Humphrey and D. Stanford. Washington, DC: The Anthropological Society of Washington, pp. 49–69.
- . 1985. Diversity and community structure of the late Pleistocene mammal fauna of North America. *Acta Zoologica Fennica* 170:181–192.
- . 1987. Late Quaternary mammalian faunas and paleoenvironments of the southwestern plains of the United States. In *Late Quaternary mammalian biogeography and environments of the Great Plains and prairies*, ed. R. W. Graham, H. A. Semken Jr., and M. A. Graham. *Illinois State Museum Scientific Papers* 22:24–86.
- Graham, R. W. and E. L. Lundelius Jr. 1984. Coevolutionary disequilibrium and Pleistocene extinctions. In *Quaternary extinctions: A prehistoric revolution*, ed. P. S. Martin and R. G. Klein. Tucson: The University of Arizona Press, pp. 223–249.
- Graham, R. W. and H. A. Semken. 1976. Paleoecological significance of the short-tailed shrew (*Blarina*), with a systematic discussion of *Blarina ozarkensis*. *Journal of Mammalogy* 57:433–449.
- Grayson, D. K. 1989. The chronology of North American late Pleistocene extinctions. *Journal of Archaeological Science* 16:153–165.
- . 1991. Late Pleistocene mammalian extinctions in North America: Taxonomy, chronology, and explanations. *Journal of World Prehistory* 5:193–230.
- Gromov, I. M. and I. Y. Polyakov. 1992. *Fauna of the USSR. Mammals. Vol. III, No. 8. Voles (Microtinae)*. Washington, DC: Smithsonian Institution Libraries and The National Science Foundation. [Translated from the original Russian; originally published 1977.]
- Guilday, J. E. 1957. Individual and geographic variation in *Blarina brevicauda* from Pennsylvania. *Annals of Carnegie Museum* 35:41–68.
- . 1967a. Differential extinction during late-Pleistocene and Recent times. In *Pleistocene extinctions: The search for a cause*, ed. P. S. Martin and H. E. Wright Jr. New Haven, CT: Yale University Press, pp. 121–140.
- . 1967b. Trout fishing. *The Netherworld News* (Pittsburgh Grotto of the National Speleological Society) 15:188–192.
- . 1971. The Pleistocene history of the Appalachian mammal fauna. In *The distributional history of the biota of the southern Appalachians, Part III: Vertebrates. A Symposium Sponsored by Virginia Polytechnic Institute and State University and the Association of Southeastern Biologists, Held at Blacksburg, Virginia June 25–27, 1970*, ed. P. C. Holt, R. A. Paterson, and J. P. Hubbard. *Virginia Polytechnic Institute and State University Research Division Monograph* 4:233–262.
- Guilday, J. E., J. F. P. Cotter, D. Cundall, E. B. Evenson, J. B. Gatewood, A. V. Morgan, A. Morgan, A. D. McCrady, D. M. Peete, R. Stuckenrath, and K. Vanderwal. 1984. Paleoecology of an early Pleistocene (Irvingtonian) cenote: Preliminary report on the Hanover Quarry No. 1 Fissure, Adams County, Pennsylvania. In *Correlation of Quaternary chronologies*, ed. W. C. Mahaney. Norwich, England: Geo Books, pp. 119–132.



- Gust, S. 1993. *Bison latifrons* and *Bison antiquus* at Rancho La Brea. *Journal of Vertebrate Paleontology* 13(suppl. 3):39A.
- Gustafson, E. P. 1977. First record of *Teleoceras* (Rhinocerotidae) from the Ringold Formation, Pliocene of Washington. *PaleoBios* 27:1-4.
- . 1978. The vertebrate faunas of the Pliocene Ringold Formation, south-central Washington. *Bulletin of the Museum of Natural History, University of Oregon* 23:1-62.
- . 1985. Soricids (Mammalia, Insectivora) from the Blufftop Local Fauna, Blancan Ringold Formation of central Washington, and the correlation of Ringold Formation faunas. *Journal of Vertebrate Paleontology* 5:88-92.
- Guthrie, R. D. 1970. Bison evolution and zoogeography in North America during the Pleistocene. *Quarterly Review of Biology* 45:1-15.
- . 1982. Mammals of the mammoth steppe as paleoenvironmental indicators. In *Paleoecology of Beringia*, ed. D. M. Hopkins, J. V. Matthews Jr., C. E. Schweger, and S. B. Young. New York: Academic Press, pp. 307-326.
- Guthrie, R. D. and J. V. Matthews Jr. 1971. The Cape Deceit Fauna—Early Pleistocene mammalian assemblage from the Alaskan Arctic. *Quaternary Research* 1:474-510.
- Hadly, E. A. 1996. Influence of late-Holocene climate on northern Rocky Mountain mammals. *Quaternary Research* 46:298-310.
- . 1997. Evolutionary and ecological response of pocket gophers (*Thomomys talpoides*) to late-Holocene climatic change. *Biological Journal of the Linnean Society* 60:277-296.
- . 1999. Fidelity of terrestrial vertebrate fossils to a modern ecosystem. *Palaeogeography, Palaeoclimatology, Palaeoecology* 149:389-409.
- Hadly, E. A. M. H. Kohn, J. A. Leonard, and R. K. Wayne. 1998. A genetic record of population isolation in pocket gophers during Holocene climatic change. *Proceedings of the National Academy of Sciences of the United States of America* 95:6893-6896.
- Hadly, E. A. and B. A. Maurer. 2001. Spatial and temporal patterns of species diversity in montane mammal communities of western North America. *Evolutionary Ecology Research* 3:477-486.
- Hager, M. W. 1975 ("1974"). Late Pliocene and Pleistocene history of the Donnelly Ranch vertebrate site, southwestern Colorado. *University of Wyoming Contributions to Geology Special Paper* 2:1-62.
- Hall, E. R. 1981. *The mammals of North America*. 2nd ed. New York: John Wiley & Sons.
- Hall, N. T. 1965. Late Cenozoic stratigraphy between Mussel Rock and Fleishhacker Zoo, San Francisco Peninsula. In *INQUA International Association for Quaternary Research VIIIth congress, guidebook for field conference I: Northern Great Basin and California*, ed. C. Wahrhaftig, R. B. Morrison, and P. W. Birkeland. Lincoln: The Nebraska Academy of Sciences, pp. 151-158.
- Hallberg, G. R. 1980. Status of Pre-Wisconsinan Pleistocene stratigraphy in Iowa. *Geological Society of America Abstracts with Programs* 12:228.
- Hallberg, G. R. and J. D. Boellstorff. 1978. Stratigraphic "confusion" in the region of the type areas of Kansan and Nebraskan deposits. *Geological Society of America Abstracts with Programs* 10:255-256.
- Hamilton, T. D., J. L. Craig, and P. V. Sellmann. 1988. The Fox permafrost tunnel: A late Quaternary geologic record in central Alaska. *Geological Society of America Bulletin* 100:948-969.
- Harrington, C. R. 1978. Quaternary vertebrate faunas of Canada and Alaska and their suggested chronological sequence. *Sylogues* 15:1-105.
- . 1984. Mammoths, bison and time in North America. In *Quaternary dating methods*, ed. W. C. Mahaney. Developments in Palaeontology and Stratigraphy 7. Amsterdam: Elsevier Science Publishers B.V., pp. 299-309.
- . 1989. Pleistocene vertebrate localities in the Yukon. In *Late Cenozoic history of the interior basins of Alaska and the Yukon*, ed. L. D. Carter, T. D. Hamilton, and J. P. Galloway. *U.S. Geological Survey Circular* 1026:93-98.
- . 1990a. Ice age vertebrates in the Canadian Arctic Islands. In *Canada's missing dimension: Science and history in the Canadian Arctic Islands*, Vol. I, ed. C. R. Harrington. Ottawa: Canadian Museum of Nature, pp. 140-160.
- . 1990b. Vertebrates of the last interglaciation in Canada: A review, with new data. *Géographie Physique et Quaternaire* 44:375-387.
- Harrington, C. R. and F. V. Clulow. 1973. Pleistocene mammals from Gold Run Creek, Yukon Territory. *Canadian Journal of Earth Sciences* 10:697-759.
- Harrison, J. A. 1978. Mammals of the Wolf Ranch Local Fauna, Pliocene of the San Pedro Valley, Arizona. *Occasional Papers of the Museum of Natural History, The University of Kansas* 73:1-18.
- Haury, E. W. 1975. *The stratigraphy and archaeology of Ventana Cave*. Tucson: The University of Arizona Press.
- Hay, O. P. and H. J. Cook. 1930. Fossil vertebrates collected near, or in association with, human artifacts at localities near Colorado, Texas; Frederick, Oklahoma; and Folsom, New Mexico. *Proceedings of the Colorado Museum of Natural History* 9:4-40, Plates 1-14.
- Haynes, C. V. Jr. 1985. Mastodon-bearing springs and late Quaternary geochronology of the Lower Pomme de Terre Valley, Missouri. *Geological Society of America Special Paper* 204:1-35.
- Hearst, J. M. 1998. Depositional environments of the Birch Creek Local Fauna (Pliocene: Blancan), Owyhee County, Idaho. In *And whereas . . . Papers on the vertebrate paleontology of Idaho honoring John A. White*, Vol. 1, ed. W. A. Akersten, H. G. McDonald, D. J. Meldrum, and M. E. T. Flint. *Idaho Museum of Natural History Occasional Paper* 36:56-93.
- . 1999. The mammalian paleontology and depositional environments of the Birch Creek Local Fauna (Pliocene: Blancan), Owyhee County, Idaho. Unpublished Ph.D. dissertation, University of Kansas.
- Hester, J. J. 1960. Late Pleistocene extinction and radiocarbon dating. *American Antiquity* 26:58-77.
- . 1967. The agency of man in animal extinctions. In *Pleistocene extinctions: The search for a cause*, ed. P. S. Martin and H. E. Wright Jr. New Haven, CT: Yale University Press, pp. 169-192.
- Hibbard, C. W. 1937. An upper Pliocene fauna from Meade County, Kansas. *Transactions of the Kansas Academy of Science* 40:239-265.
- . 1941. The Borchers Fauna, a new Pleistocene interglacial fauna from Meade County, Kansas. *University of Kansas Publications, State Geological Survey of Kansas Bulletin* 38:197-220.
- . 1942. Pleistocene mammals from Kansas. *State Geological Survey of Kansas Bulletin* 41:261-269.
- . 1943. The Rezabek fauna, a new Pleistocene fauna from Lincoln County, Kansas. *University of Kansas Science Bulletin* 29(II):235-247.
- . 1944. Stratigraphy and vertebrate paleontology of Pleistocene deposits of southwestern Kansas. *Bulletin of the Geological Society of America* 55:707-754, Plates 1-3.

- . 1949a. Pleistocene vertebrate paleontology in North America. *Bulletin of the Geological Society of America* 60:1417–1428.
- . 1949b. Pliocene Saw Rock Canyon fauna in Kansas. *Contributions from the Museum of Paleontology, University of Michigan* 7:91–105.
- . 1949c. Techniques of collecting microvertebrate fossils. *Contributions from the Museum of Paleontology, University of Michigan* 8:7–19, Plates 1–4.
- . 1950. Mammals of the Rexroad Formation from Fox Canyon, Meade County, Kansas. *Contributions from the Museum of Paleontology, University of Michigan* 8:113–192, Plates 1–5.
- . 1952. Vertebrate fossils from late Cenozoic deposits of central Kansas. *University of Kansas Paleontological Contributions, Vertebrata* 2:1–14.
- . 1953. The Saw Rock Canyon Fauna and its stratigraphic significance. *Papers of the Michigan Academy of Science, Arts, and Letters* 38:387–411.
- . 1955a. The Jinglebob interglacial (Sangamon?) fauna from Kansas and its climatic significance. *Contributions from the Museum of Paleontology, University of Michigan* 12:179–228, Plates 1–2, Chart 1.
- . 1955b. Notes on the microtine rodents from the Port Kennedy Cave deposit. *Proceedings of the Academy of Natural Sciences of Philadelphia* 107:87–97.
- . 1956. Vertebrate fossils from the Meade Formation of southwestern Kansas. *Papers of the Michigan Academy of Science, Arts, and Letters* 41:145–200, Plates 1–2.
- . 1957. Two new Cenozoic microtine rodents. *Journal of Mammalogy* 38:39–44.
- . 1958. Summary of North American Pleistocene mammalian local faunas. *Papers of the Michigan Academy of Science, Arts, and Letters* 43:3–32.
- . 1959. Late Cenozoic microtine rodents from Wyoming and Idaho. *Papers of the Michigan Academy of Science, Arts, and Letters* 44:3–40.
- . 1963. A late Illinoian fauna from Kansas and its climatic significance. *Papers of the Michigan Academy of Science, Arts, and Letters* 48:187–221.
- . 1964. A contribution to the Saw Rock Canyon Local Fauna of Kansas. *Papers of the Michigan Academy of Science, Arts, and Letters* 49:115–127.
- . 1967. New rodents from the late Cenozoic of Kansas. *Papers of the Michigan Academy of Science, Arts, and Letters* 52:115–131.
- . 1969. The rabbits (*Hypolagus* and *Pratilepus*) from the Upper Pliocene, Hagerman Local Fauna of Idaho. *Michigan Academician* 1:81–97.
- . 1970. Pleistocene mammalian local faunas from the Great Plains and central lowland provinces of the United States. In *Pleistocene and Recent environments of the central Great Plains*, ed. W. Dort Jr. and J. K. Jones Jr. Department of Geology, University of Kansas Special Publication 3. Lawrence: University of Kansas Press, pp. 395–433.
- . 1972. Class Mammalia. In *Early Pleistocene pre-glacial and glacial rocks and faunas of north-central Nebraska*, ed. M. F. Skinner and C. W. Hibbard. Bulletin of the American Museum of Natural History 148:77–130.
- . 1976. The localities of the Cudahy Fauna, with a new ground squirrel (Rodentia, Sciuridae) from the fauna of Kansas (late Kansan). In *Athlon: Essays on palaeontology in honour of Loris Shano Russell*, ed. C. S. Churcher. Royal Ontario Museum, Life Sciences, Miscellaneous Publication. Toronto: Royal Ontario Museum, pp. 278–286.
- Hibbard, C. W. and W. W. Dalquest. 1966. Fossils from the Seymour Formation of Knox and Baylor counties, Texas, and their bearing on the late Kansan climate of that region. *Contributions from the Museum of Paleontology, The University of Michigan* 21:1–66, Plates 1–5.
- . 1973. *Proneofiber*, a new genus of vole (Cricetidae: Rodentia) from the Pleistocene Seymour Formation of Texas, and its evolutionary and stratigraphic significance. *Quaternary Research* 3:269–274.
- Hibbard, C. W., D. E. Ray, D. E. Savage, D. W. Taylor, and J. E. Guilday. 1965. *Quaternary mammals of North America*. In *The Quaternary of the United States: A review volume for the VII congress of the International Association for Quaternary Research*, ed. H. E. Wright Jr. and D. G. Frey. Princeton, NJ: Princeton University Press, pp. 509–525.
- Hibbard, C. W. and D. W. Taylor. 1960. Two late Pleistocene faunas from southwestern Kansas. *Contributions from the Museum of Paleontology, The University of Michigan* 16:1–223, Plates 1–16.
- Hibbard, C. W. and R. J. Zakrzewski. 1967. Phyletic trends in the late Cenozoic microtine *Ophiomys* gen. nov., from Idaho. *Contributions from the Museum of Paleontology, The University of Michigan* 21:255–271.
- . 1972. A new species of microtine from the late Pliocene of Kansas. *Journal of Mammalogy* 53:834–839.
- Hibbard, C. W., R. J. Zakrzewski, R. E. Eshelman, G. Edmund, C. D. Griggs, and C. Griggs. 1978. Mammals from the Kanopolis Local Fauna, Pleistocene (Yarmouth) of Ellsworth County, Kansas. *Contributions from the Museum of Paleontology, The University of Michigan* 25:11–44.
- Hilgen, F. J. and C. G. Langereis. 1993. A critical re-evaluation of the Miocene/Pliocene boundary as defined in the Mediterranean. *Earth and Planetary Science Letters* 118:167–179.
- Hinton, M. A. C. 1926. *Monograph of the voles & lemmings (Microtinae) living and extinct*, Vol. I. London: The British Museum (Natural History).
- Hirschfeld, S. E. and S. D. Webb. 1968. Plio–Pleistocene megalonychid sloths of North America. *Bulletin of the Florida State Museum, Biological Sciences* 12:213–296.
- Hollister, N. 1911. A systematic synopsis of the muskrats. *North American Fauna* 32:1–47, Plates 1–6.
- Holman, J. A. 1985a. Herpetofauna of Ladds Quarry. *National Geographic Research* 1:423–436.
- . 1985b. New evidence on the status of Ladds Quarry. *National Geographic Research* 1:569–570.
- Honey, J. G., M. R. Hudson, and J. D. Obradovich. 1998. The occurrence of the Reunion II subchron in lacustrine beds in the Beaver Basin, Utah. In *Advances in vertebrate paleontology and geochronology*, ed. Y. Tomida, L. J. Flynn, and L. L. Jacobs. *National Science Museum Monographs, Tokyo, Japan* 14:17–38.
- Hopkins, D. M. 1975. Time-stratigraphic nomenclature for the Holocene epoch. *Geology* 3:10.
- Hopkins, M. L., R. Bonnicksen, and D. Fortsch. 1969. The stratigraphic position and faunal associates of *Bison* (*Gigantobison*) *latifrons* in southeastern Idaho, a progress report. *Tebiiwa* 12:1–8.
- Hulbert, R. C. Jr. 1987. A new *Cormohipparion* (Mammalia, Equidae) from the Pliocene (latest Hemphillian and Blancan) of Florida. *Journal of Vertebrate Paleontology* 7:451–468.

- Hulbert, R. C. Jr. and G. S. Morgan. 1993. Quantitative and qualitative evolution in the giant armadillo *Holmesina* (Edentata: Pampatheriidae) in Florida. In *Morphological change in Quaternary mammals of North America*, ed. R. A. Martin and A. D. Barnosky. New York: Cambridge University Press, pp. 134–177.
- Hulbert, R. C. Jr., G. S. Morgan, and S. D. Webb (eds.). 1995. Paleontology and geology of the Leisey Shell Pits, early Pleistocene of Florida, Parts I and II. *Bulletin of the Florida Museum of Natural History* 37.
- Hürzeler, J. 1967. Nouvelles découvertes de mammifères dans les sédiments fluvio-lacustres de Villafranca d'Asti. *Colloques Internationaux du Centre National de la Recherche Scientifique 163 Problèmes Actuels de Paléontologie (Évolution des Vertébrés):* 633–636.
- Izett, G. A. 1977. Volcanic ash beds in continental deposits of the southern High Plains: Their bearing on the time of the Blancan–Irvingtonian faunal transition. *Geological Society of America Abstracts with Programs* 9:1034.
- . 1981. Volcanic ash beds: Recorders of upper Cenozoic silicic pyroclastic volcanism in the western United States. *Journal of Geophysical Research* 86(B11):10200–10222, Plate 1.
- Izett, G. A. and J. G. Honey. 1995. *Geologic map of the Irish Flats NE Quadrangle, Meade County, Kansas*. U.S. Geological Survey Miscellaneous Investigations Series Map I-2498.
- Izett, G. A. and J. D. Obradovich. 1994.  $^{40}\text{Ar}/^{39}\text{Ar}$  age constraints for the Jaramillo normal subchron and the Matuyama–Brunhes geomagnetic boundary. *Journal of Geophysical Research* 99(B2):2925–2934.
- Izett, G. A. J. D. Obradovich, C. W. Naeser, and G. T. Cebula. 1981. Potassium–argon and fission-track zircon ages of Cerro Toledo Rhyolite tephra in the Jemez Mountains, New Mexico. *U.S. Geological Survey Professional Paper* 1199:37–43.
- Izett, G. A. K. L. Pierce, N. D. Naeser, and C. Jaworowski. 1992. Isotopic dating of Lava Creek B tephra in terrace deposits along the Wind River, Wyoming—Implications for post 0.6 Ma uplift of the Yellowstone Hotspot. *Geological Society of America Abstracts with Programs* 24(7):A102.
- Izett, G. A. and R. E. Wilcox. 1982. Map showing localities and inferred distributions of the Huckleberry Ridge, Mesa Falls, and Lava Creek ash beds (Pearlette family ash beds) of Pliocene and Pleistocene age in the western United States and southern Canada. U.S. Geological Survey Miscellaneous Investigations Series Map I-1325.
- Izett, G. A., R. E. Wilcox, and G. A. Borchardt. 1972. Correlation of a volcanic ash bed in Pleistocene deposits near Mount Blanco, Texas, with the Guaje Pumice Bed of the Jemez Mountains, New Mexico. *Quaternary Research* 2:554–578.
- Izett, G. A. R. E. Wilcox, H. A. Powers, and G. A. Desborough. 1970. The Bishop Ash bed, a Pleistocene marker bed in the western United States. *Quaternary Research* 1:121–132.
- Jammot, D. 1972. Principes et methodes d'une etude moderne des insectivores. Application aux Soricidae fossiles. *Mammalia* 36:435–448.
- Jass, C. N., J. I. Mead, and L. E. Logan. 2000. Harrington's extinct mountain goat (*Oreamnos harringtoni* Stock 1936) from Musko Cave, New Mexico. *Texas Journal of Science* 52:121–132.
- Jefferson, G. T. 1968. *The Camp Cady Local Fauna from Pleistocene Lake Manix, Mojave Desert, California*. Unpublished M.A. thesis, University of California, Riverside.
- . 1985. Stratigraphy and geologic history of the Pleistocene Manix Formation central Mojave Desert, California. In *Cajon Pass to Manix Lake: Geological investigations along Interstate 15 Cajon Pass to Manix Lake, California*, comp. R. E. Reynolds. Redlands, CA: San Bernardino County Museum, pp. 157–169.
- . 1987. The Camp Cady Local Fauna: Paleoenvironment of the Lake Manix Basin. *San Bernardino County Museum Association Quarterly* 34(3&4):3–35.
- . 1991. The Camp Cady Local Fauna: Stratigraphy and paleontology of the Lake Manix Basin. In *Inland southern California: The last 70 million years*, ed. M. O. Woodburne, R. E. Reynolds, and D. P. Whistler. *San Bernardino County Museum Association Quarterly* 38(3&4):93–99.
- Jefferson, G. T. and P. Remeika. 1994. The mid-Pleistocene stratigraphic co-occurrence of *Mammuthus columbi* and *M. imperator* in the Ocotillo Formation, Borrego Badlands, Anza-Borrego Desert State Park, California. *Current Research in the Pleistocene* 11:89–92.
- Johnson, N. M., C. B. Officer, N. D. Opdyke, G. D. Woodard, P. K. Zeitler, and E. H. Lindsay. 1983. Rates of late Cenozoic tectonism in the Vallecito–Fish Creek Basin, western Imperial Valley, California. *Geology* 11:664–667.
- Johnson, N. M., N. D. Opdyke, and E. H. Lindsay. 1975. Magnetic polarity stratigraphy of Pliocene–Pleistocene terrestrial deposits and vertebrate faunas, San Pedro Valley, Arizona. *Geological Society of America Bulletin* 86:5–12.
- Johnston, C. S. 1938. Preliminary report on the vertebrate type locality of Cita Canyon, and the description of an ancestral coyote. *American Journal of Science* 235:383–390.
- Johnston, C. S. and D. E. Savage. 1955. A survey of various late Cenozoic vertebrate faunas of the panhandle of Texas. Part I. Introduction, description of localities, preliminary faunal lists. *University of California Publications in Geological Sciences* 31:27–49.
- Jones, C. A., J. R. Choate, and H. H. Genoways. 1984. Phylogeny and paleobiogeography of short-tailed shrews (genus *Blarina*). In *Contributions in Quaternary vertebrate paleontology: A volume in memorial to John E. Guilday*, ed. H. H. Genoways and M. R. Dawson. *Carnegie Museum of Natural History Special Publication* 8:56–148.
- Jones, D. S., B. J. MacFadden, S. D. Webb, P. A. Mueller, D. A. Hodell, and T. M. Cronin. 1991. Integrated geochronology of a classic Pliocene fossil site in Florida: Linking marine and terrestrial biochronologies. *The Journal of Geology* 99:637–648.
- Junge, J. A. and R. S. Hoffmann. 1981. An annotated key to the long-tailed shrews (genus *Sorex*) of the United States and Canada, with notes on Middle American *Sorex*. *Occasional Papers of the Museum of Natural History, The University of Kansas* 94:1–48.
- Kelly, T. S. 1994. Two Pliocene (Blancan) vertebrate faunas from Douglas County, Nevada. *PaleoBios* 16(1):1–23.
- . 1997. Additional late Cenozoic (latest Hemphillian to earliest Irvingtonian) mammals from Douglas County, Nevada. *PaleoBios* 18(1):1–31.
- Klein, R. G. 2000. Human evolution and large mammal extinctions. In *Antelopes, deer, and relatives: Fossil record, behavioral ecology, systematics, and conservation*, ed. E. S. Vrba and G. B. Schaller. New Haven, CT: Yale University Press, pp. 128–139.
- Klingener, D. 1963. Dental evolution of *Zapus*. *Journal of Mammalogy* 44:248–260.
- Klippel, W. E. and P. W. Parmalee. 1984. Armadillos in North American late Pleistocene contexts. In *Contributions in Quaternary*

- vertebrate paleontology: A volume in memorial to John E. Guilday, ed. H. H. Genoways and M. R. Dawson. *Carnegie Museum of Natural History Special Publication* 8:149–160.
- Koenigswald, W. v. and L. D. Martin. 1984. The status of the genus *Mimomys* (Arvicolidae, Rodentia, Mamm.) in North America. *Neues Jahrbuch für Geologie und Paläontologie Abhandlungen* 168:108–124.
- Korth, W. W. 1994. *The Tertiary record of rodents in North America*. New York: Plenum Press.
- Kowallis, B. J., J. S. Heaton, and K. Bringhurst. 1986. Fission-track dating of volcanically derived sedimentary rocks. *Geology* 14:19–22.
- Kowallis, B. J., C. C. Swisher, O. Carranza-Castañeda, W. E. Miller, and D. G. Tingey. 1998. Preliminary radiometric dates in selected late Tertiary vertebrate faunas from Mexico. In *Avances en investigación, paleontología de vertebrados*, ed. O. Carranza Castañeda and D. A. Córdoba Méndez. *Universidad Autónoma del Estado de Hidalgo, Instituto de Investigaciones en Ciencias de la Tierra, Publicación Especial* 1:103–108.
- Kurtén, B. 1971. *The age of mammals*. New York: Columbia University Press.
- Kurtén, B. and E. Anderson. 1980. *Pleistocene mammals of North America*. New York: Columbia University Press.
- Lambert, W. D., D. Cordier, and R. Chandler. 1995. *Mammuthus* from the late Blancan Santa Fe River 1B locality: The oldest record of *Mammuthus* in North America. *Journal of Vertebrate Paleontology* 15(suppl. 3):40A.
- Lance, J. F. 1960. Stratigraphic and structural position of Cenozoic fossil localities in Arizona. *Arizona Geological Society Digest* 3:155–159.
- Lich, D. K. 1990. *Cosomys primus*: A case for stasis. *Paleobiology* 16:384–395.
- Lindsay, E. H. 1984. Late Cenozoic mammals from northwestern Mexico. *Journal of Vertebrate Paleontology* 4:208–215.
- . 1997. The Pliocene–Pleistocene boundary in continental sequences of North America. In *The Pleistocene boundary and the beginning of the Quaternary*, ed. J. A. Van Couvering. New York: Cambridge University Press, pp. 278–289.
- Lindsay, E. H. and L. L. Jacobs. 1985. Pliocene small mammal fossils from Chihuahua, Mexico. *Universidad Nacional Autónoma de México, Instituto de Geología, Paleontología Mexicana* 51:1–53.
- Lindsay, E. H., N. M. Johnson, and N. D. Opdyke. 1975. Preliminary correlation of North American land mammal ages and geomagnetic chronology. In *Studies on Cenozoic paleontology and stratigraphy in honor of Claude W. Hibbard*, Claude W. Hibbard Memorial Vol. 3, ed. G. R. Smith and N. E. Friedland. *University of Michigan, The Museum of Paleontology Papers on Paleontology* 12:111–119.
- Lindsay, E. H., Y. Mou, W. Downs, J. Pederson, T. S. Kelly, C. Henry, and J. Trexler. 2002. Recognition of the Hemphillian/Blancan boundary in Nevada. *Journal of Vertebrate Paleontology* 22:429–442.
- Lindsay, E. H., N. D. Opdyke, and N. M. Johnson. 1984. Blancan–Hemphillian land mammal ages and late Cenozoic mammal dispersal events. *Annual Review of Earth and Planetary Sciences* 12:445–488.
- Lindsay, E. H. and N. T. Tesson. 1974. Cenozoic vertebrate localities and faunas in Arizona. *Journal of the Arizona Academy of Science* 9:3–24.
- Lipps, L. and C. E. Ray. 1967. The Pleistocene fossiliferous deposit at Ladds, Bartow County, Georgia. *Bulletin of the Georgia Academy of Science* 25:113–119.
- Lister, A. M. 1993a. Evolution of mammoths and moose: The Holarctic perspective. In *Morphological change in Quaternary mammals of North America*, ed. R. A. Martin and A. D. Barnosky. New York: Cambridge University Press, pp. 178–204.
- . 1993b. Mammoths in miniature. *Nature* 362:288–289.
- Logan, T. R., S. G. Lucas, and J. C. Sobus. 1984. Blancan–Irvingtonian boundary in the Ceja Member of the Santa Fe Formation, Tijeras Arroyo, Albuquerque area, New Mexico. *New Mexico Geology* 6:14.
- Lucas, S. G. 1995. The Thornton Beach mammoth and the antiquity of *Mammuthus* in North America. *Quaternary Research* 43:263–264.
- Lucas, S. G., G. S. Morgan, and G. H. Mack. 1998. Early Pleistocene (early Irvingtonian) co-occurrence of the proboscideans *Cuvieronius*, *Stegomastodon*, and *Mammuthus* at Tortugas Mountain, Doña Ana County, New Mexico. In *Las Cruces country II*, ed. G. H. Mack, G. S. Austin, and J. M. Barker. *New Mexico Geological Society guidebook, 49th annual field conference*, p. 34.
- Lucas, S. G., T. E. Williamson, and J. Sobus. 1993. Plio–Pleistocene stratigraphy, paleoecology, and mammalian biochronology, Tijeras Arroyo, Albuquerque area, New Mexico. *New Mexico Geology* 15:1–8, 15.
- Lundelius, E. L. Jr. 1967. Late-Pleistocene and Holocene faunal history of central Texas. In *Pleistocene extinctions: The search for a cause*, ed. P. S. Martin and H. E. Wright Jr. New Haven, CT: Yale University Press, pp. 287–319.
- . 1985. Pleistocene vertebrates from Laubach Cave. In *Edwards Aquifer–Northern segment, Travis, Williamson, and Bell counties, Texas*, coord. C. M. Woodruff Jr., F. Snyder, L. de la Garza, and R. M. Slade Jr. *Austin Geological Society Guidebook* 8:41–45.
- . 1989. The implications of disharmonious assemblages for Pleistocene extinctions. *Journal of Archaeological Science* 16:407–417.
- Lundelius, E. L. Jr., C. S. Churcher, T. Downs, C. R. Harington, E. H. Lindsay, G. E. Schultz, H. A. Semken, S. D. Webb, and R. J. Zakrzewski. 1987. The North American Quaternary sequence. In *Cenozoic mammals of North America: Geochronology and biostratigraphy*, ed. M. O. Woodburne. Berkeley: University of California Press, pp. 211–235.
- Lundelius, E. L. Jr., R. W. Graham, E. Anderson, J. Guilday, J. A. Holman, D. W. Steadman, and S. D. Webb. 1983. Terrestrial vertebrate faunas. In *Late-Quaternary environments of the United States*, ed. H. E. Wright Jr. Vol. 1, *The late Pleistocene*, ed. S. C. Porter. Minneapolis: University of Minnesota Press, pp. 311–353.
- Lyell, C. 1833. *Principles of geology, being an attempt to explain the former changes of the earth's surface, by reference to causes now in operation*. Vol. III. London: John Murray.
- MacDonald, J. R. and W. J. Pelletier. 1958. The Pliocene mammalian faunas of Nevada, U.S.A. In *Paleontología, taxonomía y evolución*, ed. A. G. Rojas. Congreso Geológico Internacional, XXth Sesión, Ciudad de México, 1956. Mexico, D. F., pp. 365–388.
- MacFadden, B. J. 1984. Systematics and phylogeny of *Hipparion*, *Neohipparion*, *Nannippus*, and *Cormohipparion* (Mammalia, Equidae) from the Miocene and Pliocene of the New World. *Bulletin of the American Museum of Natural History* 179:1–195.

- MacPhee, R. D. E. ed. 1999. *Extinctions in near time: Causes, contexts, and consequences*. New York: Kluwer Academic/Plenum Publishers.
- Madden, C. T. 1980. The Proboscidea of South America. *Geological Society of America Abstracts with Programs* 12:474.
- . 1983. More earliest isotopically dated *Mammuthus* from North America. *Quaternary Research* 19:275–277.
- . 1985. On a single elephant tooth: Most primitive mammoth from coastal plain of southeastern United States. *Southeastern Geology* 25:207–211.
- . 1986. *Stegomastodon* associated with *Mammuthus* in Arizona during the Quaternary. *Quaternary Research* 26:266–271.
- . 1995. Even more earliest isotopically dated *Mammuthus* from North America. *Quaternary Research* 43:265–267.
- Madden, C. T. and W. W. Dalquest. 1990. The last rhinoceros in North America. *Journal of Vertebrate Paleontology* 10:266–267.
- Malde, H. E. 1991. Quaternary geology and structural history of the Snake River Plain, Idaho and Oregon. In *Quaternary nonglacial geology: Conterminous U.S.*, ed. R. B. Morrison. The Geology of North America Volume K-2. Boulder, CO: The Geological Society of America, pp. 251–281.
- Malde, H. E. and H. A. Powers. 1962. Upper Cenozoic stratigraphy of western Snake River Plain, Idaho. *Geological Society of America Bulletin* 73:1197–1219, Plate 1.
- Maldonado, F., S. D. Connell, D. W. Love, V. J. S. Grauch, J. L. Slate, W. C. McIntosh, P. B. Jackson, and F. M. Byers Jr. 1999. Neogene geology of the Isleta Reservation and vicinity, Albuquerque Basin, central New Mexico. In *Albuquerque geology*, ed. F. J. Pazzaglia and S. G. Lucas. *New Mexico Geological Society guidebook, 50th annual field conference*, pp. 175–188.
- Mangerud, J., S. T. Andersen, B. E. Berglund, and J. J. Donner. 1974. Quaternary stratigraphy of Norden, a proposal for terminology and classification. *Boreas* 3:109–127.
- Markewich, H. W., D. A. Wysocki, M. J. Pavich, E. M. Rutledge, H. T. Millard Jr., F. J. Rich, P. B. Maat, M. Rubin, and J. P. McGeehin. 1998. Paleopedology plus TL, <sup>10</sup>Be, and <sup>14</sup>C dating as tools in stratigraphic and paleoclimatic investigations, Mississippi River Valley, U.S.A. *Quaternary International* 51/52:143–167.
- Markova, A. K. 1992. Influence of paleoclimatic changes in the middle and late Pleistocene on the composition of small mammal faunas: Data from eastern Europe. In *Mammalian migration and dispersal events in the European Quaternary*, ed. W. v. Koenigswald and L. Werdelin. *Courier Forschungsinstitut Senckenberg* 153:93–100.
- Marshall, L. G., R. F. Butler, R. E. Drake, G. H. Curtis, and R. H. Tedford. 1979. Calibration of the great American interchange. *Science* 204:272–279.
- Martin, L. D. 1972. The microtine rodents of the Mullen assemblage from the Pleistocene of north central Nebraska. *Bulletin of the University of Nebraska State Museum* 9:173–182.
- . 1975. Microtine rodents from the Ogallala Pliocene of Nebraska and the early evolution of the Microtinae in North America. In *Studies on Cenozoic paleontology and stratigraphy in honor of Claude W. Hibbard*, Claude W. Hibbard Memorial Vol. 3, ed. G. R. Smith and N. E. Friedland. *University of Michigan, The Museum of Paleontology Papers on Paleontology* 12:101–110.
- . 1979. The biostratigraphy of arvicoline rodents in North America. *Transactions of the Nebraska Academy of Sciences* 7:91–100.
- . 1980. Functional morphology and the evolution of cats. *Transactions of the Nebraska Academy of Sciences* 8:141–153.
- . 1985. Tertiary extinction cycles and the Pliocene–Pleistocene boundary. In *Institute for Tertiary–Quaternary Studies, TER–QUA symposium series*, Vol. 1, ed. W. Dort Jr. Lincoln: Nebraska Academy of Sciences, pp. 33–40.
- . 1989. Plio–Pleistocene rodents in North America. In *Papers on fossil rodents in honor of Albert Elmer Wood*, ed. C. C. Black and M. R. Dawson. *Natural History Museum of Los Angeles County Science Series* 33:47–58.
- . 1994. A new genus of Miocene vole possibly related to *Phenacomys*. In *Institute for Tertiary–Quaternary Studies, TER–QUA symposium series*, Vol. 2, ed. W. Dort Jr. Lincoln: Nebraska Academy of Sciences, pp. 129–130.
- . 1998. Felidae. In *Evolution of Tertiary mammals of North America*, Vol. 1: *Terrestrial carnivores, ungulates, and ungulate-like mammals*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, pp. 236–242.
- Martin, L. D. and R. S. Hoffmann. 1987. Pleistocene faunal provinces and Holocene biomes of the central Great Plains. In *Quaternary environments of Kansas*, ed. W. C. Johnson. *University of Kansas, Kansas Geological Survey Guidebook Series* 5:159–165.
- Martin, L. D. and A. M. Neuner. 1978. The end of the Pleistocene in North America. *Transactions of the Nebraska Academy of Sciences and Affiliated Societies* 6:117–126.
- Martin, L. D. and C. B. Schultz. 1985. Small mammals of the Seneca and Sappa local faunas (post-Ogallala of Nebraska). In *Institute for Tertiary–Quaternary Studies, TER–QUA symposium series*, Vol. 1, ed. W. Dort Jr. Lincoln: Nebraska Academy of Sciences, pp. 163–179.
- Martin, P. S. 1967. Prehistoric overkill. In *Pleistocene extinctions: The search for a cause*, ed. P. S. Martin and H. E. Wright Jr. New Haven, CT: Yale University Press, pp. 75–120.
- . 1973. The discovery of America. *Science* 179:969–974.
- Martin, P. S. and A. J. Stuart. 1995. Mammoth extinction: Two continents and Wrangel Island. *Radiocarbon* 37:7–10.
- Martin, R. A. 1973. The Java Local Fauna, Pleistocene of South Dakota: A preliminary report. *Bulletin of the New Jersey Academy of Science* 18:48–56.
- . 1974. Fossil mammals from the Coleman IIA Fauna, Sumter County. In *Pleistocene mammals of Florida*, ed. S. D. Webb. Gainesville: The University Presses of Florida, pp. 35–99.
- . 1975. *Allophaiomys* Kormos from the Pleistocene of North America. In *Studies on Cenozoic paleontology and stratigraphy in honor of Claude W. Hibbard*, Claude W. Hibbard Memorial Vol. 3, ed. G. R. Smith and N. E. Friedland. *University of Michigan, The Museum of Paleontology Papers on Paleontology* 12:97–100.
- . 1986. Energy, ecology, and cotton rat evolution. *Paleobiology* 12:370–382.
- . 1989a. Arvicolid rodents of the early Pleistocene Java Local Fauna from north-central South Dakota. *Journal of Vertebrate Paleontology* 9:438–450.
- . 1989b. Early Pleistocene zapodid rodents from the Java Local Fauna of north-central South Dakota. *Journal of Vertebrate Paleontology* 9:101–109.
- . 1993. Patterns of variation and speciation in Quaternary rodents. In *Morphological change in Quaternary mammals of North America*, ed. R. A. Martin and A. D. Barnosky. New York: Cambridge University Press, pp. 226–280.
- . 1994. A preliminary review of dental evolution and paleogeography in the zapodid rodents, with emphasis on Pliocene

- and Pleistocene taxa. In *Rodent and lagomorph families of Asian origins and diversification*, ed. Y. Tomida, C. Li, and T. Setoguchi. *National Science Museum Monographs, Tokyo, Japan* 8:99–113.
- . 1996. Dental evolution and size change in the North American muskrat: Classification and tempo of a presumed phyletic sequence. In *Palaeoecology and palaeoenvironments of late Cenozoic mammals: Tributes to the career of C. S. (Rufus) Churcher*, ed. K. M. Stewart and K. L. Seymour. Toronto: University of Toronto Press, pp. 431–457.
- Martin, R. A. and A. D. Barnosky, eds. 1993. *Morphological change in Quaternary mammals of North America*. New York: Cambridge University Press.
- Martin, R. A., J. G. Honey, and P. Peláez-Campanones. 2000. The Meade Basin rodent project: A progress report. *Paludicola* 3:1–32.
- Martin, R. A. and S. D. Webb. 1974. Late Pleistocene mammals from the Devil's Den fauna, Levy County. In *Pleistocene mammals of Florida*, ed. S. D. Webb. Gainesville: The University Presses of Florida, pp. 114–145.
- Matthew, W. D. 1915. Climate and evolution. *Annals of the New York Academy of Sciences* 24:171–318.
- . 1924. Correlation of the Tertiary formations of the Great Plains. *Bulletin of the Geological Society of America* 35:743–754.
- . 1925. Blanco and associated formations of northern Texas. *Bulletin of the Geological Society of America* 36:221–222.
- May, S. R. 1981. *Repomys* (Mammalia: Rodentia gen. nov.) from the late Neogene of California and Nevada. *Journal of Vertebrate Paleontology* 1:219–230.
- Mayle, F. E. and L. C. Cwynar. 1995. A review of multi-proxy data for the Younger Dryas in Atlantic Canada. *Quaternary Science Reviews* 14:813–821.
- McCord, R., L. C. Thrasher, R. S. White, B. J. Tegowski, A. Skaff, G. McCullough, and E. Schirtzinger. 2002 (“2001”). New fossil records from the Blancan 111 Ranch beds, Graham County, Arizona: Results of cooperative efforts by a federal agency, public and private museums and the amateur community. *Mesa Southwest Museum Bulletin* 8:113–120.
- McDaniel, G. E. Jr. and G. T. Jefferson. 1999. Distribution of proboscideans in the Anza-Borrego Desert State Park region, California. In *Abstracts from proceedings, the 1999 Desert Research Symposium, San Bernardino County Museum, Redlands, California*, comp. J. Reynolds. *San Bernardino County Museum Association Quarterly* 46(2):61–62.
- McDonald, G. H., P. K. Link, and D. E. Lee. 1996. An overview of the geology and paleontology of the Pliocene Glenns Ferry Formation, Hagerman Fossil Beds National Monument. *Northwest Geology* 26:16–45.
- McDonald, H. G. 1995. Gravigrade xenarthrans from the early Pleistocene Leisey Shell Pit 1A, Hillsborough County, Florida. In *Paleontology and geology of the Leisey Shell Pits, early Pleistocene of Florida*, ed. R. C. Hulbert Jr., G. S. Morgan, and S. D. Webb. *Bulletin of the Florida Museum of Natural History* 37(Part II):345–373.
- McDonald, J. N. 1981. *North American bison: Their classification and evolution*. Berkeley: University of California Press.
- McDonald, J. N. and G. S. Morgan. 1999. The appearance of *Bison* in North America. *Current Research in the Pleistocene* 16:127–129.
- McDonald, J. N. C. E. Ray, and F. Grady. 1996. Pleistocene caribou (*Rangifer tarandus*) in the eastern United States: New records and range extensions. In *Palaeoecology and palaeoenvironments of late Cenozoic mammals: Tributes to the career of C. S. (Rufus) Churcher*, ed. K. M. Stewart and K. L. Seymour. Toronto: University of Toronto Press, pp. 406–430.
- McKenna, M. C. and S. K. Bell. 1997. *Classification of mammals above the species level*. New York: Columbia University Press.
- Mead, E. M. and J. I. Mead. 1989. Quaternary zoogeography of the Nearctic *Dicrostonyx* lemmings. *Boreas* 18:323–332.
- Mead, J. I. and C. J. Bell. 2001. Pliocene amphibians and reptiles from Clark County, Nevada. *Bulletin of the Southern California Academy of Sciences* 100:1–11.
- Mead, J. I., C. J. Bell, and L. K. Murray. 1992. *Mictomys borealis* (northern bog lemming) and the Wisconsin paleoecology of the east-central Great Basin. *Quaternary Research* 37:229–238.
- Mead, J. I., C. Manganaro, C. A. Repenning, and L. D. Agenbroad. 1996. Early Rancholabrean mammals from Salamander Cave, Black Hills, South Dakota. In *Palaeoecology and palaeoenvironments of late Cenozoic mammals: Tributes to the career of C. S. (Rufus) Churcher*, ed. K. M. Stewart and K. L. Seymour. Toronto: University of Toronto Press, pp. 458–482.
- Mead, J. I. and D. J. Meltzer. 1984. North American late Quaternary extinctions and the radiocarbon record. In *Quaternary extinctions: A prehistoric revolution*, ed. P. S. Martin and R. G. Klein. Tucson: University of Arizona Press, pp. 440–450.
- Mead, J. I. and L. H. Taylor. 1998. *Oreamnos* (Caprinae, Bovidae, Artiodactyla), from the Irvingtonian (Pleistocene) of Porcupine Cave, Colorado, North America. In *Advances in vertebrate paleontology and geochronology*, ed. Y. Tomida, L. J. Flynn, and L. L. Jacobs. *National Science Museum Monographs, Tokyo, Japan* 14:181–189.
- Meade, G. E. 1945. The Blanco Fauna. *Contributions to Geology, 1944: The University of Texas Publication* 4401:509–556, Plates 48–55.
- . 1953. An early Pleistocene vertebrate fauna from Frederick, Oklahoma. *The Journal of Geology* 61:452–460, Plate 1.
- Mehring, P. J. Jr. 1967. The environment of extinction of the late-Pleistocene megafauna in the arid southwestern United States. In *Pleistocene extinctions: The search for a cause*, ed. P. S. Martin and H. E. Wright Jr. New Haven, CT: Yale University Press, pp. 247–266.
- Meltzer, D. J. and J. I. Mead. 1983. The timing of late Pleistocene mammalian extinctions in North America. *Quaternary Research* 19:130–135.
- . 1985. Dating late Pleistocene extinctions: Theoretical issues, analytical bias, and substantive results. In *Environments and extinctions: Man in late glacial North America*, ed. J. I. Mead and D. J. Meltzer. Orono: Center for the Study of Early Man, University of Maine, pp. 145–173.
- Meyer, C. E., M. J. Woodward, A. M. Sarna-Wojcicki, and C. W. Naeser. 1980. Zircon fission-track age of 0.45 million years on ash in the type section of the Merced Formation, west-central California. *U.S. Geological Survey Open-File Report* 80-1071:1–9.
- Miller, B. B., R. W. Graham, A. V. Morgan, N. G. Miller, W. D. McCoy, D. F. Palmer, A. J. Smith, and J. J. Pilny. 1994. A biota associated with Matuyama-age sediments in west-central Illinois. *Quaternary Research* 41:350–365.
- Miller, W. E. 1968. Occurrence of a giant bison, *Bison latifrons*, and a slender-limbed camel, *Tanupolama*, at Rancho La Brea. *Los Angeles County Museum Contributions in Science* 147:1–9.
- . 1971. Pleistocene vertebrates of the Los Angeles Basin and vicinity (exclusive of Rancho La Brea). *Bulletin of the Los Angeles County Museum of Natural History, Science* 10:1–124.

- . 1980. The late Pliocene Las Tunas Local Fauna from southernmost Baja California, Mexico. *Journal of Paleontology* 54:762–805.
- Miller, W. E. and J. D. Brotherson. 1979. Size variation in foot elements of *Bison* from Rancho La Brea. *Natural History Museum of Los Angeles County Contributions in Science* 323:1–19.
- Miller, W. E. and O. Carranza-Castañeda. 1984. Late Cenozoic mammals from central Mexico. *Journal of Vertebrate Paleontology* 4:216–236.
- . 1998a. The importance of late Tertiary Carnivora from central Mexico. In *Avances en investigación, paleontología de vertebrados*, ed. O. Carranza Castañeda and D. A. Córdoba Méndez. *Universidad Autónoma del Estado de Hidalgo, Instituto de Investigaciones en Ciencias de la Tierra, Publicación Especial* 1:96–102.
- . 1998b. Late Tertiary canids from central Mexico. *Journal of Paleontology* 72:546–556.
- Montellano, M. 1989. Pliocene Camelidae of Rancho El Ocote, central Mexico. *Journal of Mammalogy* 70:359–369.
- Montellano-Ballesteros, M. 1992 (“1990”). Una edad del Irvingtoniano al Rancholabreano para la fauna Cedazo del Estado de Aguascalientes. *Universidad Nacional Autónoma de México, Instituto de Geología, Revista* 9:195–203.
- Moore, D. W. and L. L. Janecek. 1990. Genic relationships among North American *Microtus* (Mammalia: Rodentia). *Annals of Carnegie Museum* 59:249–259.
- Mooser, O. and W. W. Dalquest. 1975a. A new species of camel (genus *Camelops*) from the Pleistocene of Aguascalientes, Mexico. *The Southwestern Naturalist* 19:341–345.
- . 1975b. Pleistocene mammals from Aguascalientes, central Mexico. *Journal of Mammalogy* 56:781–820.
- Mooser Barendun, O. 1958. La fauna “Cedazo” del Pleistoceno en Aguascalientes. *Anales del Instituto de Biología, Universidad Nacional Autónoma de México* 29:409–452.
- Morgan, G. S. and R. C. Hulbert Jr. 1995. Overview of the geology and vertebrate biochronology of the Leisey Shell Pit Local Fauna, Hillsborough County, Florida. In *Paleontology and geology of the Leisey Shell Pits, early Pleistocene of Florida*, ed. R. C. Hulbert Jr., G. S. Morgan, and S. D. Webb. *Bulletin of the Florida Museum of Natural History* 37(Part I):1–92.
- Morgan, G. S. and S. G. Lucas. 1999. Pliocene (Blancan) vertebrates from the Albuquerque Basin, north-central New Mexico. In *Albuquerque geology*, ed. F. J. Pazzaglia and S. G. Lucas. *New Mexico Geological Society guidebook, 50th annual field conference*, pp. 363–370.
- . 2000a. Biostratigraphic significance of late Neogene vertebrate fossils from the Gila Group, Pearson Mesa, southwestern New Mexico and southeastern Arizona. In *Southwest passage: A trip through the Phanerozoic*, ed. T. F. Lawton, N. J. McMillan, and V. T. McLemore. *New Mexico Geological Society guidebook, 51st annual field conference*, pp. 211–220.
- . 2000b. Pliocene and Pleistocene vertebrate faunas from the Albuquerque Basin, New Mexico. In *New Mexico's fossil record 2*, ed. S. G. Lucas. *New Mexico Museum of Natural History and Science Bulletin* 16:217–240.
- Morgan, G. S., S. G. Lucas, and J. W. Estep. 1998. Pliocene (Blancan) vertebrate fossils from the Camp Rice Formation near Tonuco Mountain, Doña Ana County, southern New Mexico. In *Las Cruces country II*, ed. G. H. Mack, G. S. Austin, and J. M. Barker. *New Mexico Geological Society guidebook, 49th annual field conference*, pp. 237–249.
- Morgan, G. S. and R. B. Ridgway. 1987. Late Pliocene (late Blancan) vertebrates from the St. Petersburg Times Site, Pinellas County, Florida, with a brief review of Florida Blancan faunas. *Florida Museum of Natural History, Papers in Florida Paleontology* 1:1–22.
- Morgan, G. S., P. L. Sealey, S. G. Lucas, and A. B. Heckert. 1997. Pliocene (latest Hemphillian and Blancan) vertebrate fossils from the Mangas Basin, southwestern New Mexico. In *New Mexico's fossil record 1*, ed. S. G. Lucas, J. W. Estep, T. E. Williamson, and G. S. Morgan. *New Mexico Museum of Natural History and Science Bulletin* 11:97–128.
- Morgan, G. S. and J. A. White. 1995. Small mammals (Insectivora, Lagomorpha, and Rodentia), from the early Pleistocene (Irvingtonian) Leisey Shell Pit Local Fauna, Hillsborough County, Florida. In *Paleontology and geology of the Leisey Shell Pits, early Pleistocene of Florida*, ed. R. C. Hulbert Jr., G. S. Morgan, and S. D. Webb. *Bulletin of the Florida Museum of Natural History* 37(Part II):397–461.
- Morgan, J. K. and N. H. Morgan. 1995. A new species of *Capromeryx* (Mammalia: Artiodactyla) from the Taunton Local Fauna of Washington, and the correlation with other Blancan faunas of Washington and Idaho. *Journal of Vertebrate Paleontology* 15:160–170.
- Morlan, R. E. 1984. Biostratigraphy and biogeography of Quaternary microtine rodents from northern Yukon Territory, eastern Beringia. In *Contributions in Quaternary vertebrate paleontology: A volume in memorial to J. E. Guilday*, ed. H. H. Genoways and M. R. Dawson. *Carnegie Museum of Natural History Special Publication* 8:184–199.
- . 1996. Late and middle Pleistocene vertebrate fossils from Old Crow Basin, Locality CRH 15, northern Yukon Territory. In *Palaeoecology and palaeoenvironments of late Cenozoic mammals: Tributes to the career of C. S. (Rufus) Churcher*, ed. K. M. Stewart and K. L. Seymour. Toronto: University of Toronto Press, pp. 483–521.
- Morrison, R. B. 1991. Introduction. In *Quaternary nonglacial geology: Conterminous U.S.*, ed. R. B. Morrison. The Geology of North America Volume K-2. Boulder, CO: The Geological Society of America, pp. 1–12.
- Mosimann, J. E. and P. S. Martin. 1975. Simulating overkill by Paleoindians. *American Scientist* 63:304–313.
- Mou, Y. 1997. A new arvicoline species (Rodentia: Cricetidae) from the Pliocene Panaca Formation, southeast Nevada. *Journal of Vertebrate Paleontology* 17:376–383.
- . 1998. Schmelzmuster of *Mimomys panacaensis*. In *Advances in vertebrate paleontology and geochronology*, ed. Y. Tomida, L. J. Flynn, and L. L. Jacobs. *National Science Museum Monographs, Tokyo, Japan* 14:79–90.
- Muhs, D. R., J. B. Swinehart, D. B. Loope, J. N. Aleinikoff, and J. Been. 1999. 200,000 years of climate change recorded in eolian sediments of the High Plains of eastern Colorado and western Nebraska. In *Colorado and adjacent areas*, ed. D. R. Lageson, A. P. Lester, and B. D. Trudgill. *Geological Society of America Field Guide* 1:71–91.
- Musser, G. G. and M. D. Carleton. 1993. Family Muridae. In *Mammal species of the world: A taxonomic and geographic reference*, 2nd ed., ed. D. E. Wilson and D. M. Reeder. Washington, DC: Smithsonian Institution Press, pp. 501–755.
- Naeser, C. W., G. A. Izett, and R. E. Wilcox. 1971. Zircon fission-track ages of Pearlette-like volcanic ash beds in the Great Plains. *Geological Society of America Abstracts with Programs* 3:657.

- . 1973. Zircon fission-track ages of Pearlette family ash beds in Meade County, Kansas. *Geology* 1:187–189.
- Neustadt, M. I. 1967. The lower Holocene boundary. In *Quaternary paleoecology*, ed. E. J. Cushing and H. E. Wright Jr. New Haven, CT: Yale University Press, pp. 415–425.
- Neville, C., N. D. Opdyke, E. H. Lindsay, and N. M. Johnson. 1979. Magnetic stratigraphy of Pliocene deposits of the Glens Ferry Formation, Idaho, and its implications for North American mammalian biostratigraphy. *American Journal of Science* 279:503–526.
- Nicholas, G. 1953. Recent paleontological discoveries from Cumberland Bone Cave. *Scientific Monthly* 76:301–305.
- Noller, J. S., J. M. Sowers, and W. R. Lettis (eds.). 2000. *Quaternary geochronology: Methods and applications*. AGU Reference Shelf 4. Washington, DC: American Geophysical Union.
- Nowak, R. M. 1979. North American Quaternary *Canis*. *Monograph of the Museum of Natural History, University of Kansas* 6:1–154.
- Obradovich, J. D. and G. A. Izett. 1991.  $^{40}\text{Ar}/^{39}\text{Ar}$  ages of upper Cenozoic Yellowstone Group tuffs. *Geological Society of America Abstracts with Programs* 23(2):84.
- Opdyke, N. D. and J. E. T. Channell. 1996. *Magnetic stratigraphy*. International Geophysics Series, Vol. 64. San Diego: Academic Press.
- Opdyke, N. D., E. H. Lindsay, N. M. Johnson, and T. Downs. 1977. The paleomagnetism and magnetic polarity stratigraphy of the mammal-bearing section of Anza-Borrego State Park, California. *Quaternary Research* 7:316–329.
- Osborn, H. F. 1910. *The age of mammals in Europe, Asia and North America*. New York: The Macmillan Company.
- . 1936. *Proboscidea: A monograph of the discovery, evolution, migration and extinction of the mastodonts and elephants of the world, Vol. I, Mæratherioidea, Deinotherioidea, Mastodontoidea*. New York: The American Museum Press, The American Museum of Natural History.
- Osborn, H. F. and W. D. Matthew. 1909. Cenozoic mammal horizons of western North America with faunal lists of the Tertiary Mammalia of the West. *U.S. Geological Survey Bulletin* 361:1–138, Plates 1–3.
- Owen, P. R., C. J. Bell, and E. M. Mead. 2000. Fossils, diet, and conservation of black-footed ferrets (*Mustela nigripes*). *Journal of Mammalogy* 81:422–433.
- Owen-Smith, N. 1987. Pleistocene extinctions: The pivotal role of megaherbivores. *Paleobiology* 13:351–362.
- Packer, D. R. 1979. Paleomagnetism and age dating of the Ringold Formation and loess deposits in the State of Washington. *Oregon Geology* 41:119–132.
- Pajak, A. F. III, E. Scott, and C. J. Bell. 1996. A review of the biostratigraphy of Pliocene and Pleistocene sediments in the Elsinore Fault Zone, Riverside County, California. In *The uses of vertebrate fossils in biostratigraphic correlation*, ed. C. J. Bell and S. S. Sumida. *PaleoBios* 17(2–4):28–49.
- Pasini, G. and M. L. Colalongo. 1997. The Pliocene–Pleistocene boundary-stratotype at Vrica, Italy. In *The Pleistocene boundary and the beginning of the Quaternary*, ed. J. A. Van Couvering. New York: Cambridge University Press, pp. 15–45.
- Patton, T. H. 1965. A new genus of fossil microtine from Texas. *Journal of Mammalogy* 46:466–471.
- Paulson, G. R. 1961. The mammals of the Cudahy Fauna. *Papers of the Michigan Academy of Science, Arts, and Letters* 46:127–153.
- Péwé, T. L. 1975. Quaternary geology of Alaska. *United States Geological Survey Professional Paper* 835:1–145.
- . 1989. Quaternary stratigraphy of the Fairbanks area, Alaska. In *Late Cenozoic history of the interior basins of Alaska and the Yukon*, ed. L. D. Carter, T. D. Hamilton, and J. P. Galloway. *U.S. Geological Survey Circular* 1026:72–77.
- Péwé, T. L. and D. M. Hopkins. 1967. Mammal remains of pre-Wisconsin age in Alaska. In *The Bering Land Bridge*, ed. D. M. Hopkins. Stanford, CA: Stanford University Press, pp. 266–270.
- Pfaff, K. S. 1990. Irvingtonian *Microtus*, *Pedomys*, and *Pitymys* (Mammalia, Rodentia, Cricetidae) from Trout Cave No. 2, West Virginia. *Annals of Carnegie Museum* 59:105–134.
- . 1991. *An Irvingtonian mammalian fauna from Trout Cave No. 2, Pendleton County, West Virginia*. Unpublished M.S. thesis, University of Minnesota.
- Pinsof, J. D. 1991. A cranium of *Bison alaskensis* (Mammalia: Artiodactyla: Bovidae) and comments on fossil *Bison* diversity in the American Falls area, southeastern Idaho. *Journal of Vertebrate Paleontology* 11:509–514.
- Preece, S. J., J. A. Westgate, B. A. Stemper, and T. A. Péwé. 1999. Tephrochronology of late Cenozoic loess at Fairbanks, central Alaska. *Geological Society of America Bulletin* 111:71–90.
- Prothero, D. R. 1998. Rhinocerotidae. In *Evolution of Tertiary mammals of North America, Vol. 1, Terrestrial carnivores, ungulates, and ungulatelike mammals*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, pp. 595–605.
- Prothero, D. R. C. Guérin, and E. Manning. 1989. The history of the Rhinoceroidea. In *The evolution of perissodactyls*, ed. D. R. Prothero and R. M. Schoch. *Oxford Monographs on Geology and Geophysics* 15. New York: Oxford University Press, pp. 321–340.
- Prothero, D. R. and E. M. Manning. 1987. Miocene rhinoceroses from the Texas Gulf Coastal Plain. *Journal of Paleontology* 61:388–423.
- Purdue, J. R. and E. J. Reitz. 1993. Decrease in body size of white-tailed deer (*Odocoileus virginianus*) during the late Holocene in South Carolina and Georgia. In *Morphological change in Quaternary mammals of North America*, ed. R. A. Martin and A. D. Barnosky. New York: Cambridge University Press, pp. 281–298.
- Ray, C. E. and A. E. Sanders. 1984. Pleistocene tapirs in the eastern United States. In *Contributions in Quaternary vertebrate paleontology: A volume in memorial to John E. Guilday*, ed. H. H. Genoways and M. R. Dawson. *Carnegie Museum of Natural History Special Publication* 8:283–315.
- Raymo, M. E. 1992. Global climate change: A three million year perspective. In *Start of a glacial*, ed. G. J. Kukla and E. Went. NATO Advanced Science Institutes Series I, Global Environmental Change, Vol. 3. Berlin: Springer-Verlag, pp. 207–223.
- Remeika, P. and G. T. Jefferson. 1993. The Borrego Local Fauna: Revised basin–margin stratigraphy and paleontology of the western Borrego Badlands, Anza-Borrego Desert State Park, California. In *Ashes, faults and basins*, ed. R. E. Reynolds and J. Reynolds. *San Bernardino County Museum Association Special Publication* 93-1:90–93.
- Remeika, P., G. T. Jefferson, and L. K. Murray. 1995. Fossil vertebrate faunal list for the Vallecito–Fish Creek and Borrego–San Felipe Basins, Anza-Borrego Desert State Park and vicinity, California. In *Paleontology and geology of the western Salton Trough Detachment, Anza-Borrego Desert State Park, California: Field trip guidebook and volume for the 1995 San Diego Association of Geologist's field trip to Anza-Borrego Desert State Park*, Vol. 1, ed. P. Remeika and A. Sturz. San Diego: San Diego Association of Geologists, pp. 82–93.



- Rensberger, J. M. and A. D. Barnosky. 1993. Short-term fluctuations in small mammals of the late Pleistocene from eastern Washington. In *Morphological change in Quaternary mammals of North America*, ed. R. A. Martin and A. D. Barnosky. New York: Cambridge University Press, pp. 299–342.
- Rensberger, J. M., A. D. Barnosky, and P. Spencer. 1984. Geology and paleontology of a Pleistocene-to-Holocene loess succession, Benton County, Washington. *Eastern Washington University Reports in Archaeology and History* 100-39:1–105.
- Repenning, C. A. 1962. The giant ground squirrel, *Paenemarmota*. *Journal of Paleontology* 36:540–556.
- . 1978. Faunal exchanges between Siberia and North America. In *American Quaternary Association abstracts of the fifth biennial meeting, September 2–4, 1978, University of Alberta, Edmonton, Alberta*, pp. 40–55.
- . 1980. Faunal exchanges between Siberia and North America. *Canadian Journal of Anthropology* 1:37–44.
- . 1983. *Pitymys meadensis* Hibbard from the Valley of Mexico and the classification of North American species of *Pitymys* (Rodentia: Cricetidae). *Journal of Vertebrate Paleontology* 2:471–482.
- . 1984. Quaternary rodent biochronology and its correlation with climatic and magnetic stratigraphies. In *Correlation of Quaternary chronologies*, ed. W. C. Mahaney. Norwich, England: Geo Books, pp. 105–118.
- . 1987. Biochronology of the microtine rodents of the United States. In *Cenozoic mammals of North America: Geochronology and biostratigraphy*, ed. M. O. Woodburne. Berkeley: University of California Press, pp. 236–268.
- . 1992. *Allophaiomys* and the age of the Olyor Suite, Krestovka Sections, Yakutia. *U.S. Geological Survey Bulletin* 2037:1–98.
- . 1998. North American mammalian dispersal routes: Rapid evolution and dispersal constrain precise biochronology. In *Advances in vertebrate paleontology and geochronology*, ed. Y. Tomida, L. J. Flynn, and L. L. Jacobs. *National Science Museum Monographs, Tokyo, Japan* 14:39–78.
- . 2001. Beringian climate during intercontinental dispersal: A mouse eye view. *Quaternary Science Reviews* 20:25–40.
- Repenning, C. A. and E. M. Brouwers. 1992. Late Pliocene–early Pleistocene ecologic changes in the Arctic Ocean borderland. *U.S. Geological Survey Bulletin* 2036:1–37.
- Repenning, C. A., E. M. Brouwers, L. D. Carter, L. Marincovich Jr., and T. A. Ager. 1987. The Beringian ancestry of *Phenacomys* (Rodentia: Cricetidae) and the beginning of the modern Arctic Ocean borderland biota. *U.S. Geological Survey Bulletin* 1687:1–35.
- Repenning, C. A. and O. Fejfar. 1977. Holarctic correlations of microtine rodents. In *Project 73/1/24 Quaternary glaciations in the Northern Hemisphere, report no. 4 on the session in Stuttgart, Federal Republic of Germany, September 1976*, ed. V. Šibrava, ed. Prague: Geological Survey, pp. 234–250.
- Repenning, C. A., O. Fejfar, and W.-D. Heinrich. 1990. Arvicolid rodent biochronology of the northern hemisphere. In *International symposium: Evolution, phylogeny and biostratigraphy of arvicolids (Rodentia, Mammalia)*, ed. O. Fejfar and W.-D. Heinrich. Prague: Geological Survey, pp. 385–417.
- Repenning, C. A. and F. Grady. 1988. The microtine rodents of the Cheetah Room Fauna, Hamilton Cave, West Virginia, and the spontaneous origin of *Synaptomys*. *U.S. Geological Survey Bulletin* 1853:1–32.
- Repenning, C. A., T. R. Weasman, and G. R. Scott. 1995. The early Pleistocene (latest Blancan–earliest Irvingtonian) Froman Ferry Fauna and history of the Glenns Ferry Formation, southwestern Idaho. *U.S. Geological Survey Bulletin* 2105:1–86.
- Reynolds, R. E., L. P. Fay, and R. L. Reynolds. 1990. California Oaks Road: An early–late Irvingtonian land mammal age fauna from Murrieta, Riverside County, California. In *Abstracts of proceedings, 1990 Mojave Desert Quaternary research symposium*, comp. J. Reynolds. *San Bernardino County Museum Association Quarterly* 37(2):35–36.
- Reynolds, R. E. and E. H. Lindsay. 1999. Late Tertiary basins and vertebrate faunas along the Nevada–Utah border. In *Vertebrate paleontology in Utah*, ed. D. D. Gillette. *Utah Geological Survey Miscellaneous Publication* 99-1:469–478.
- Reynolds, R. E., R. L. Reynolds, and A. F. Pajak III. 1991. Blancan, Irvingtonian, and Rancholabrean(?) land mammal age faunas from western Riverside County, California. In *Inland southern California: The last 70 million years*, ed. M. O. Woodburne, R. E. Reynolds, and D. P. Whistler. *San Bernardino County Museum Association Quarterly* 38(3&4):37–40.
- Rhodes, R. S. II. 1984. Paleocology and regional paleoclimatic implications of the Farmdalian Craigmile and Woodfordian Waubonsie mammalian local faunas, southwestern Iowa. *Illinois State Museum Reports of Investigations* 40:1–51.
- Richmond, G. M. and D. S. Fullerton. 1986. Summation of Quaternary glaciations in the United States of America. In *Quaternary glaciations in the Northern Hemisphere*, ed. V. Šibrava, D. Q. Bowen, and G. M. Richmond. *Quaternary Science Reviews* 5:183–196.
- Rio, D., R. Sprovieri, and R. Thunell. 1991. Pliocene–lower Pleistocene chronostratigraphy: A re-evaluation of Mediterranean type sections. *Geological Society of America Bulletin* 103:1049–1058.
- Robertson, J. S. Jr. 1976. Latest Pliocene mammals from Haile XV A, Alachua County, Florida. *Bulletin of the Florida State Museum, Biological Sciences* 20:111–186.
- Rodbell, D. T. 2000. The Younger Dryas: Cold, cold everywhere? *Science* 290:285–286.
- Rodbell, D. T., S. L. Forman, J. Pierson, and W. C. Lynn. 1997. Stratigraphy and chronology of Mississippi Valley loess in western Tennessee. *Geological Society of America Bulletin* 109:1134–1148.
- Rogers, K. L., E. E. Larson, G. Smith, D. Katzman, G. R. Smith, T. Cerling, Y. Wang, R. G. Baker, K. C. Lohmann, C. A. Repenning, P. Patterson, and G. Mackie. 1992. Pliocene and Pleistocene geologic and climatic evolution in the San Luis Valley of south-central Colorado. *Palaeogeography, Palaeoclimatology, Palaeoecology* 94:55–86.
- Rogers, K. L., C. A. Repenning, R. M. Forester, E. E. Larson, S. A. Hall, G. R. Smith, E. Anderson, and T. J. Brown. 1985. Middle Pleistocene (late Irvingtonian: Nebraskan) climatic changes in south-central Colorado. *National Geographic Research* 1:535–563.
- Rogers, K. L., C. A. Repenning, F. G. Luiszer, and R. D. Benson. 2000. Geologic history, stratigraphy, and paleontology of SAM Cave, north-central New Mexico. *New Mexico Geology* 22:89–100, 113–117.
- Rowe, T. 1987. Definition and diagnosis in the phylogenetic system. *Systematic Zoology* 36:208–211.
- Rowe, T. and J. Gauthier. 1992. Ancestry, paleontology, and definition of the name *Mammalia*. *Systematic Biology* 41:372–378.

- Roy, K., D. Jablonski, and J. W. Valentine. 1995. Thermally anomalous assemblages revisited: Patterns in the extraprovincial latitudinal range shifts of Pleistocene marine mollusks. *Geology* 23:1071–1074.
- Ruez, D. R. Jr. 2001. Early Irvingtonian (latest Pliocene) rodents from Inglis 1C, Citrus County, Florida. *Journal of Vertebrate Paleontology* 21:153–171.
- Sankey, J. T. 1996. Vertebrate paleontology and magnetostratigraphy of the upper Glens Ferry (latest Pliocene) and lower Bruneau (Pliocene–Pleistocene) formations near Murphy, southwestern Idaho. *Journal of the Idaho Academy of Science* 32:71–88.
- . 2002. Vertebrate paleontology and magnetostratigraphy of the Glens Ferry and Bruneau formations (Plio–Pleistocene), near Murphy, southwestern Idaho. In *And whereas. . . Papers on the vertebrate paleontology of Idaho honoring John A. White*, Vol. 2, ed. W. A. Akersten, M. E. Thompson, D. J. Meldrum, R. A. Rapp, and H. G. McDonald. *Idaho Museum of Natural History Occasional Paper* 37:52–100.
- Sarna-Wojcicki, A. M. 1976. Correlation of late Cenozoic tuffs in the central coast ranges of California by means of trace- and minor-element chemistry. *U.S. Geological Survey Professional Paper* 972:1–30.
- Sarna-Wojcicki, A. M., K. R. Lajoie, C. E. Meyer, D. P. Adam, and H. J. Rieck. 1991. Tephrochronologic correlation of upper Neogene sediments along the Pacific margin, conterminous United States. In *Quaternary nonglacial geology: Conterminous U.S.*, ed. R. B. Morrison. The Geology of North America Volume K-2. Boulder, CO: The Geological Society of America, pp. 117–140.
- Sarna-Wojcicki, A. M., M. S. Pringle, and J. Wijbrans. 2000. New  $^{40}\text{Ar}/^{39}\text{Ar}$  age of the Bishop Tuff from multiple sites and sediment rate calibration for the Matuyama–Brunhes boundary. *Journal of Geophysical Research* 105(B9):21431–21443.
- Saunders, J. J. 1988. Fossiliferous spring sites in southwestern Missouri. In *Late Pleistocene and early Holocene paleoecology and archeology of the eastern Great Lakes region*, ed. R. S. Laub, N. G. Miller, and D. W. Steadman. *Bulletin of the Buffalo Society of Natural Sciences* 33:127–149.
- Savage, D. E. 1951. Late Cenozoic vertebrates of the San Francisco Bay region. *University of California Publications, Bulletin of the Department of Geological Sciences* 28:215–314.
- . 1955. A survey of various late Cenozoic vertebrate faunas of the panhandle of Texas. Part II. Proboscidea. *University of California Publications in Geological Sciences* 31:51–72.
- Savage, D. E. and G. H. Curtis. 1970. The Villafranchian Stage-Age and its radiometric dating. In *Radiometric dating and paleontologic zonation*, ed. O. L. Bandy. *Geological Society of America Special Paper* 124:207–231.
- Schubert, B. W. 1997. *Paleontology and paleoecology of a terminal Pleistocene mammalian fauna, Little Beaver Cave, central Ozarks, Missouri*. Unpublished M.S. thesis, Northern Arizona University, Flagstaff.
- Schubert, B. W. and R. W. Graham. 2000. Terminal Pleistocene armadillo (*Dasypus*) remains from the Ozark Plateau, Missouri, USA. *PaleoBios* 20(1):1–6.
- Schultz, C. B. and J. M. Hillerud. 1977. The antiquity of *Bison latifrons* (Harlan) in the Great Plains of North America. *Transactions of the Nebraska Academy of Sciences and Affiliated Societies* 4:103–116.
- Schultz, C. B. and L. D. Martin. 1970. Quaternary mammalian sequence in the central Great Plains. In *Pleistocene and Recent environments of the central Great Plains*, ed. W. Dort Jr. and J. K. Jones Jr. Department of Geology, University of Kansas Special Publication 3. Lawrence: University of Kansas Press, pp. 341–353.
- Schultz, C. B., L. D. Martin, L. G. Tanner, and R. G. Corner. 1977. Provincial land mammal ages for the North American Quaternary. In *Abstracts, X INQUA Congress, Birmingham, 1977*, p. 408.
- . 1978. Provincial land mammal ages for the North American Quaternary. *Transactions of the Nebraska Academy of Sciences and Affiliated Societies* 5:59–64.
- Schultz, C. B., L. G. Tanner, and L. D. Martin. 1972. Phyletic trends in certain lineages of Quaternary mammals. *Bulletin of the University of Nebraska State Museum* 9:183–195.
- Schultz, G. E. 1965. Pleistocene vertebrates from the Butler Spring Local Fauna, Meade County, Kansas. *Papers of the Michigan Academy of Science, Arts, and Letters* 50:235–265.
- . 1967. Four superimposed late-Pleistocene vertebrate faunas from southwest Kansas. In *Pleistocene extinctions: The search for a cause*, ed. P. S. Martin and H. E. Wright Jr. New Haven, CT: Yale University Press, pp. 321–336.
- . 1969. Geology and paleontology of a late Pleistocene basin in southwest Kansas. *Geological Society of America Special Paper* 105:1–85.
- . 1977. Blancan and post-Blancan faunas in the Texas Panhandle. In *Guidebook: Field conference on late Cenozoic biostratigraphy of the Texas panhandle and adjacent Oklahoma, August 4–6, 1977*, ed. G. E. Schultz. Killgore Research Center, West Texas State University Department of Geology and Anthropology Special Publication 1:105–145.
- . 1986. Stop 18: Biostratigraphy and volcanic ash deposits of the Tule Formation, Briscoe County, Texas. In *Geomorphology and Quaternary stratigraphy of the Rolling Plains, Texas panhandle*, ed. T. C. Gustavson. *The University of Texas at Austin, Bureau of Economic Geology Guidebook* 22:82–89.
- . 1990a. Stop 5: Blanco Local Fauna and the Blancan land mammal age. In *Tertiary and Quaternary stratigraphy and vertebrate paleontology of parts of northwestern Texas and eastern New Mexico*, ed. T. C. Gustavson. *The University of Texas at Austin, Bureau of Economic Geology Guidebook* 24 Pp. 44–51.
- . 1990b. Stop 8: Biostratigraphy and volcanic ash deposits of the Tule Formation, Briscoe County, Texas. In *Tertiary and Quaternary stratigraphy and vertebrate paleontology of parts of northwestern Texas and eastern New Mexico*, ed. T. C. Gustavson. *The University of Texas at Austin, Bureau of Economic Geology Guidebook* 24:60–64.
- Schultz, J. R. 1937. A late Cenozoic vertebrate fauna from the Coso Mountains, Inyo County, California. *Carnegie Institution of Washington Publication* 487:75–109, Plates 1–8.
- Scott, E. and S. M. Cox. 2002. Late Pleistocene distribution of *Bison* in the Mojave Desert, southern California and Nevada. *Journal of Vertebrate Paleontology* 22(suppl. 3):104A–105A.
- . 1993. *Arctodus simus* (Cope, 1879) from Riverside County, California. In *New additions to the Pleistocene vertebrate record of California*, ed. R. G. Dundas and D. J. Long. *PaleoBios* 15 Pp. 27–36.
- Scott, W. E., K. L. Pierce, J. P. Bradbury, and R. M. Forester. 1982. Revised Quaternary stratigraphy and chronology in the American Falls area, southeastern Idaho. In *Cenozoic geology of Idaho*, ed. B. Bonnicksen and R. M. Breckenridge. *Idaho Department of Lands, Bureau of Mines and Geology Bulletin* 26:581–595.

- Seff, P. 1960. Preliminary report of the stratigraphy of the 111 Ranch beds, Graham County, Arizona. *Arizona Geological Society Digest* 3:137–139, Plate 1.
- Semken, H. A. Jr. 1966. Stratigraphy and paleontology of the McPherson *Equus* beds (Sandahl Local Fauna), McPherson County, Kansas. *Contributions from the Museum of Paleontology, The University of Michigan* 20:121–178.
- . 1974. Micromammal distribution and migration during the Holocene. In *American Quaternary Association abstracts of the third biennial meeting, July 30–August 4, 1974, University of Wisconsin–Madison*, p. 25.
- . 1984. Paleoeology of a late Wisconsinan/Holocene micro-mammal sequence in Peccary Cave, northwestern Arkansas. In *Contributions in Quaternary vertebrate paleontology: A volume in memorial to John E. Guilday*, ed. H. H. Genoways and M. R. Dawson. *Carnegie Museum of Natural History Special Publication* 8:405–431.
- . 1988. Environmental Interpretations of the “Disharmomious” late Wisconsinan biome of southeastern North America. In *Late Pleistocene and early Holocene paleoecology and archeology of the eastern Great Lakes region*, ed. R. S. Laub, N. G. Miller, and D. W. Steadman. *Bulletin of the Buffalo Society of Natural Sciences* 33:185–194.
- Semken, H. A. Jr. and C. R. Falk. 1987. Late Pleistocene/Holocene mammalian faunas and environmental changes on the northern plains of the United States. In *Late Quaternary mammalian biogeography and environments of the Great Plains and Prairies*, ed. R. W. Graham, H. A. Semken Jr., and M. A. Graham. *Illinois State Museum Scientific Papers* 22:176–313.
- Seymour, K. 1993. Size change in North American Quaternary jaguars. In *Morphological change in Quaternary mammals of North America*, ed. R. A. Martin and A. D. Barnosky. New York: Cambridge University Press, pp. 343–372.
- Shackleton, N. J. and N. D. Opdyke. 1976. Oxygen-isotope and paleomagnetic stratigraphy of Pacific core V28-239 late Pliocene to latest Pleistocene. In *Investigation of late Quaternary paleoceanography and paleoclimatology*, ed. R. M. Cline and J. D. Hays. *Geological Society of America Memoir* 145:449–464.
- Shaw, C. A. 1981. *The middle Pleistocene El Golfo Local Fauna from northwestern Sonora, Mexico*. Unpublished M.S. thesis, California State University, Long Beach.
- Shaw, C. A. and H. G. McDonald. 1987. First record of giant anteater (*Xenarthra*, Mymecophagidae) in North America. *Science* 236:186–188.
- Sher, A. V. 1986. On the history of mammal faunas of Beringida. *Quartärpaläontologie* 6:185–193.
- Shotwell, J. A. 1970. Pliocene mammals of southeast Oregon and adjacent Idaho. *University of Oregon Museum of Natural History Bulletin* 17:1–103.
- Simpson, G. G. 1933. Glossary and correlation charts of North American Tertiary mammal-bearing formations. *Bulletin of the American Museum of Natural History* 67:79–121.
- . 1945. The principles of classification and a classification of mammals. *Bulletin of the American Museum of Natural History* 85:1–350.
- Skinner, M. F. 1942. The fauna of Papago Springs Cave, Arizona, and a study of *Stockoceros*, with three new antilocaprine from Nebraska and Arizona. *Bulletin of the American Museum of Natural History* 80:143–220.
- Skinner, M. F. and O. C. Kaisen. 1947. The fossil *Bison* of Alaska and preliminary revision of the genus. *Bulletin of the American Museum of Natural History* 89:123–256, Plates 8–26.
- Slaughter, B. H. 1966. *Platygonus compressus* and associated fauna from the Laubach Cave of Texas. *American Midland Naturalist* 75:475–494.
- . 1967. Animal ranges as a clue to late-Pleistocene extinction. In *Pleistocene extinctions: The search for a cause*, ed. P. S. Martin and H. E. Wright Jr. New Haven, CT: Yale University Press, pp. 155–167.
- Slaughter, R. W. and S. P. Jones. 2000. Late-glacial record of *Dicrostonyx* from Honey Dipper Den, Jones County, east-central Iowa. *Current Research in the Pleistocene* 17:133–135.
- Spell, T. L., I. McDougall, and A. P. Dougeris. 1996. Cerro Toledo Rhyolite, Jemez Volcanic Field, New Mexico: <sup>40</sup>Ar/<sup>39</sup>Ar geochronology of eruptions between two caldera-forming events. *Geological Society of America Bulletin* 108:1549–1566.
- Stafford, T. W. Jr. 1990. Late Pleistocene megafauna extinctions and the Clovis culture: Absolute ages based on accelerator <sup>14</sup>C dating of skeletal remains. In *Megafauna & man: Discovery of America's heartland*, ed. L. D. Agenbroad, J. I. Mead, and L. W. Nelson. *The Mammoth Site of Hot Springs, South Dakota, Inc. Scientific Papers* 1:118–122.
- Stafford, T. W. Jr., H. A. Semken Jr., R. W. Graham, W. F. Klippel, A. Markova, N. G. Smirnov, and J. Southon. 1999. First accelerator mass spectrometry <sup>14</sup>C dates documenting contemporaneity of nonanalogue species in late Pleistocene mammal communities. *Geology* 27:903–906.
- Stalker, A. M. 1969. Quaternary stratigraphy in southern Alberta report II: Sections near Medicine Hat. *Geological Survey of Canada Paper* 69-26:1–28.
- . 1996. Origin of the vertebrate fossil sites near Medicine Hat, Alberta. In *Palaeoecology and palaeoenvironments of late Cenozoic mammals: Tributes to the career of C. S. (Rufus) Churcher*, ed. K. M. Stewart and K. L. Seymour. Toronto: University of Toronto Press, pp. 247–258.
- Stalker, A. M. and C. S. Churcher. 1970. *Deposits near Medicine Hat, Alberta, Canada*. *Geological survey of Canada*. Wall chart, one sheet.
- . 1972. Glacial stratigraphy of the southwestern Canadian prairies: The Laurentide record. In *International Geological Congress, twenty-fourth session, section 12: Quaternary geology*, conv. J. G. Fyles, A. M. Stalker, and W. O. Kupsch. Montreal: International Geological Congress, pp. 110–119.
- . 1982. *Ice age deposits and animals from the southwestern part of the Great Plains of Canada*. Geological Survey of Canada, Miscellaneous Report 31. Wall chart, one sheet.
- Starrett, A. 1956. Pleistocene mammals of the Berends Fauna of Oklahoma. *Journal of Paleontology* 30:1187–1192.
- Stephens, J. J. 1960. Stratigraphy and paleontology of a late Pleistocene basin, Harper County, Oklahoma. *Bulletin of the Geological Society of America* 71:1675–1702, Plate 1.
- Stewart, J. D. 1987. Latitudinal effects in Wisconsinan mammalian faunas of the plains. In *Quaternary environments of Kansas*, ed. W. C. Johnson. *University of Kansas, Kansas Geological Survey Guidebook Series* 5:153–158.
- Stirton, R. A. 1939. Cenozoic mammal remains from the San Francisco Bay region. *University of California Publications, Bulletin of the Department of Geological Sciences* 24:339–409.

- . 1940. Phylogeny of North American Equidae. *University of California Publications, Bulletin of the Department of Geological Sciences* 25:165–197.
- . 1951. Prehistoric land animals of the San Francisco Bay region. In *Geologic guidebook of the San Francisco Bay counties: History, landscape, geology, fossils, minerals, industry, and routes to travel*, ed. O. P. Jenkins. *State of California Division of Mines Bulletin* 154:177–186.
- Stock, C. 1921. Later Cenozoic mammalian remains from the Meadow Valley region, southeastern Nevada. *The American Journal of Science* 202:250–264.
- Stoner, J. S., J. E. T. Channell, and C. Hillaire-Marcel. 1998. A 200 ka geomagnetic chronostratigraphy for the Labrador Sea: Indirect correlation of the sediment record to SPECMAP. *Earth and Planetary Science Letters* 159:165–181.
- Stoner, J. S., C. Laj, J. E. T. Channell, and C. Kissel. 2002. South Atlantic and North Atlantic geomagnetic paleointensity stacks (0–80 ka): implications for inter-hemispheric correlation. *Quaternary Science Reviews* 21:1141–1151.
- Strain, W. S. 1966. Blancan mammalian fauna and Pleistocene formations, Hudspeth County, Texas. *Bulletin of the Texas Memorial Museum* 10:1–55.
- Stuart, A. J. 1991. Mammalian extinctions in the late Pleistocene of northern Eurasia and North America. *Biological Reviews* 66:453–562.
- Taylor, A. J. 1982. *The mammalian fauna from the mid-Irvingtonian Fyllan Cave local fauna, Travis County, Texas*. Unpublished M.A. thesis, The University of Texas at Austin.
- Tedford, R. H. 1981. Mammalian biochronology of the late Cenozoic basins of New Mexico. *Geological Society of America Bulletin*, Part I, 92:1008–1022.
- Tedford, R. H., T. Galusha, M. F. Skinner, B. E. Taylor, R. W. Fields, J. R. Macdonald, J. M. Rensberger, S. D. Webb, and D. P. Whistler. 1987. Faunal succession and biochronology of the Arikareean through Hemphillian interval (late Oligocene through earliest Pliocene epochs) in North America. In *Cenozoic mammals of North America: Geochronology and biostratigraphy*, ed. M. O. Woodburne. Berkeley: University of California Press, pp. 153–210.
- Tedford, R. H. and E. P. Gustafson. 1977. First North American record of the extinct panda *Parailurus*. *Nature* 265:621–623.
- Tomida, Y. 1987. *Small mammal fossils and correlation of continental deposits, Safford and Duncan Basins, Arizona, USA*. Tokyo: National Science Museum.
- Trexler, J. H. Jr., P. H. Cashman, C. D. Henry, T. Muntean, K. Schwartz, A. TenBrink, J. E. Faulds, M. Perkins, and T. Kelly. 2000. Neogene basins in western Nevada document the tectonic history of the Sierra Nevada-Basin and range transition zone for the last 12 Ma. In *Great Basin and Sierra Nevada*, ed. D. R. Lageson, S. G. Peters, and M. M. Lahren. *Geological Society of America Field Guide* 2:97–116.
- Troxell, E. L. 1915a. A fossil ruminant from Rock Creek, Texas, *Preptoceras mayfieldi* sp. nov. *The American Journal of Science* 190:479–482.
- . 1915b. The vertebrate fossils of Rock Creek, Texas. *The American Journal of Science* 189:613–638, Plate 9.
- Trumbore, S. E. 2000. Radiocarbon geochronology. In *Quaternary geochronology: Methods and applications*, ed. J. S. Noller, J. M. Sowers, and W. R. Lettis. AGU Reference Shelf 4. Washington, DC: American Geophysical Union, pp. 41–60.
- Van Couvering, J. A. (ed.). 1997. *The Pleistocene boundary and the beginning of the Quaternary*. New York: Cambridge University Press.
- Van Couvering, J. A., D. Castradori, M. B. Cita, F. J. Hilgen, and D. Rio. 2000. The base of the Zanclean Stage and of the Pliocene Series. *Episodes* 23:179–187.
- Vanderhill, J. B. 1986. *Lithostratigraphy, vertebrate paleontology, and magnetostratigraphy of Plio–pleistocene sediments in the Mesilla Basin, New Mexico*. Unpublished Ph.D. dissertation, The University of Texas at Austin.
- Van der Meulen, A. J. 1978. *Microtus* and *Pitymys* (Arvicolidae) from Cumberland Cave, Maryland, with a comparison of some New and Old World species. *Annals of Carnegie Museum* 47:101–145.
- Van Valen, L. 1970. Late Pleistocene extinctions. In *Proceedings of the North American Paleontological Convention, Field Museum of Natural History, Chicago, September 5–7, 1969*, Vol. 1, Part E, *Evolution of communities*, ed. E. L. Yochelson. Lawrence, KS: Allen Press, pp. 469–485.
- Van Valkenburgh, B., F. Grady, and B. Kurtén. 1990. The Plio–Pleistocene cheetah-like cat *Miracinonyx inexpectatus* of North America. *Journal of Vertebrate Paleontology* 10:434–454.
- Vartanyan, S. L., K. A. Arslanov, T. V. Tertychnaya, and S. B. Chernov. 1995. Radiocarbon dating evidence for mammoths on Wrangel Island, Arctic Ocean, until 2000 BC. *Radiocarbon* 37:1–6.
- Vartanyan, S. L., V. E. Garutt, and A. V. Sher. 1993. Holocene dwarf mammoths from Wrangel Island in the Siberian Arctic. *Nature* 362:337–340.
- Viriot, L., J. Chaline, A. Schaaf, and E. Le Boulenge. 1993. Ontogenetic change of *Ondatra zibethicus* (Arvicolidae, Rodentia) cheek teeth analyzed by digital image processing. In *Morphological change in Quaternary mammals of North America*, ed. R. A. Martin and A. D. Barnosky. New York: Cambridge University Press, pp. 373–391.
- Voorhies, M. R. 1977. Fossil moles of late Hemphillian age from northeastern Nebraska. *Transactions of the Nebraska Academy of Sciences and Affiliated Societies* 4:129–138.
- . 1990. Vertebrate biostratigraphy of the Ogallala Group in Nebraska. In *Geologic framework and regional hydrology: Upper Cenozoic Blackwater Draw and Ogallala formations, Great Plains*, ed. T. C. Gustafson. Austin: The University of Texas at Austin, Bureau of Economic Geology, pp. 115–151.
- Walsh, S. L. 1998. Fossil datum and paleobiological event terms, paleontostratigraphy, chronostratigraphy, and the definition of land mammal “age” boundaries. *Journal of Vertebrate Paleontology* 18:150–179.
- Wang, X., R. H. Tedford, and B. E. Taylor. 1999. Phylogenetic systematics of the Borophaginae (Carnivora: Canidae). *Bulletin of the American Museum of Natural History* 243:1–391.
- Webb, S. D. 1974a. Chronology of Florida Pleistocene mammals. In *Pleistocene mammals of Florida*, ed. S. D. Webb. Gainesville: The University Presses of Florida, pp. 5–31.
- . 1974b. Pleistocene llamas of Florida, with a brief review of the Lamini. In *Pleistocene mammals of Florida*, ed. S. D. Webb. Gainesville: The University Presses of Florida, pp. 170–213.
- . 1983. A new species of *Pediomeryx* from the late Miocene of Florida, and its relationships within the subfamily Cranioceratinae (Ruminantia: Dromomerycidae). *Journal of Mammalogy* 64:261–276.

- . 1985. Late Cenozoic mammal dispersals between the Americas. In *The great American biotic interchange*, ed. F. G. Stehli and S. D. Webb. New York: Plenum Press, pp. 357–386.
- . 1991. Ecogeography and the great American interchange. *Paleobiology* 17:266–280.
- . 2000. Evolutionary history of New World Cervidae. In *Antelopes, deer, and relatives: Fossil record, behavioral ecology, systematics, and conservation*, ed. E. S. Vrba and G. B. Schaller. New Haven, CT: Yale University Press, pp. 38–64.
- Webb, S. D. and A. D. Barnosky. 1989. Faunal dynamics of Pleistocene mammals. *Annual Review of Earth and Planetary Sciences* 17:413–438.
- Webb, S. D. and J. P. Dudley. 1995. Proboscidea from the Leisey Shell Pits, Hillsborough County, Florida. In *Paleontology and geology of the Leisey Shell Pits, early Pleistocene of Florida*, ed. R. C. Hulbert Jr., G. S. Morgan, and S. D. Webb. *Bulletin of the Florida Museum of Natural History* 37(II):645–660.
- Webb, S. D., G. S. Morgan, R. C. Hulbert Jr., D. S. Jones, B. J. MacFadden, and P. A. Mueller. 1989. Geochronology of a rich early Pleistocene vertebrate fauna, Leisey Shell Pit, Tampa Bay, Florida. *Quaternary Research* 32:96–110.
- Webb, S. D. and A. Rancy. 1996. Late Cenozoic evolution of the Neotropical mammal fauna. In *Evolution & environment in tropical America*, ed. J. B. C. Jackson, A. F. Budd, and A. G. Coates. Chicago: University of Chicago Press, pp. 335–358.
- Webb, S. D. and K. T. Wilkins. 1984. Historical biogeography of Florida Pleistocene mammals. In *Contributions in Quaternary vertebrate paleontology: A volume in memorial to John E. Guilday*, ed. H. H. Genoways and M. R. Dawson. *Carnegie Museum of Natural History Special Publication* 8:370–383.
- Webb, S. D., S. W. Wise Jr., and R. Wright. 1978. Late Miocene glacio-eustatic cycles in Florida: Marine and fluvio-estuarine sequences. *Geological Society of America Abstracts with Programs* 10:513.
- Westgate, J. A., N. D. Briggs, A. M. Stalker, and C. S. Churcher. 1978. Fission-track age of glass from tephra beds associated with Quaternary vertebrate assemblages in the southern Canadian plains. *Geological Society of America Abstracts with Programs* 10:514–515.
- Westgate, J. A. and M. P. Gorton. 1981. Correlation techniques in tephra studies. In *Tephra studies*, ed. S. Self and R. S. J. Sparks. NATO Advanced Study Institutes Series. Series C: Mathematical and Physical Sciences, Vol. 75. Dordrecht, Holland: D. Reidel Publishing Company, pp. 73–94.
- Westgate, J. A., B. A. Stemper, and T. L. Péwé. 1990. A 3 m.y. record of Pliocene–Pleistocene loess in interior Alaska. *Geology* 18:858–861.
- Wheatley, C. M. 1871. Notice of the discovery of a cave in eastern Pennsylvania, containing remains of post-Pliocene fossils, including those of mastodon, tapir, *Megalonyx*, *Myiodon*, etc. *The American Journal of Science and Arts* 101:235–237.
- White, J. A. 1984. Late Cenozoic Leporidae (Mammalia, Lagomorpha) from the Anza-Borrego Desert, southern California. In *Papers in vertebrate paleontology honoring Robert Warren Wilson*, ed. R. M. Mengel. *Carnegie Museum of Natural History Special Publication* 9:41–57.
- . 1987. The Archaeolaginae (Mammalia, Lagomorpha) of North America, excluding *Archaeolagus* and *Panolax*. *Journal of Vertebrate Paleontology* 7:425–450.
- . 1991a. A new *Sylvilagus* (Mammalia: Lagomorpha) from the Blancan (Pliocene) and Irvingtonian (Pleistocene) of Florida. *Journal of Vertebrate Paleontology* 11:243–246.
- . 1991b. North American Leporidae (Mammalia: Lagomorpha) from late Miocene (Clarendonian) to latest Pliocene (Blancan). *Journal of Vertebrate Paleontology* 11:67–89.
- White, J. A. and N. H. Morgan. 1995. The Leporidae (Mammalia, Lagomorpha) from the Blancan (Pliocene) Taunton local fauna of Washington. *Journal of Vertebrate Paleontology* 15:366–374.
- Wilmarth, M. G. 1925. The geologic time classification of the United States Geological Survey compared with other classifications and accompanied by the original definitions of era, period and epoch terms. *U.S. Geological Survey Bulletin* 769:1–138, Plate 1.
- Wilson, M. 1974a. The Casper Local Fauna and its fossil bison. In *The Casper Site: A Hell Gap bison kill on the high plains*, ed. G. C. Frison. New York: Academic Press, pp. 125–171.
- . 1974b. History of the bison in Wyoming, with particular reference to early Holocene forms. In *Applied geology and archaeology: The Holocene history of Wyoming*, ed. M. Wilson. *The Geological Survey of Wyoming, Report of Investigations* 10:91–99.
- Wilson, R. W. 1932. *Cosomys*, a new genus of vole from the Pliocene of California. *Journal of Mammalogy* 13:150–154, Plate 12.
- . 1935. A new species of porcupine from the later Cenozoic of Idaho. *Journal of Mammalogy* 16:220–222.
- Winans, M. C. 1989. A quantitative study of North American fossil species of the genus *Equus*. In *The evolution of perissodactyls*, ed. D. R. Prothero and R. M. Schoch. Oxford Monographs on Geology and Geophysics 15. New York: Oxford University Press, pp. 262–297.
- Winkler, A. J. and F. Grady. 1990. The middle Pleistocene rodent *Atopomys* (Cricetidae: Arvicolinae) from the eastern and south-central United States. *Journal of Vertebrate Paleontology* 10:484–490.
- Winkler, A. J. and Y. Tomida. 1988. New records of the small leporid *Aztilanolagus agilis* Russell and Harris (Leporidae: Leporinae). *Southwestern Naturalist* 33:391–396.
- Wood, H. E. II, R. W. Chaney, J. Clark, E. H. Colbert, G. L. Jepsen, J. B. Reeside Jr., and C. Stock. 1941. Nomenclature and correlation of the North American continental Tertiary. *Bulletin of the Geological Society of America* 52:1–48, Plate 1.
- Wood, P. A. 1960. Paleontological investigations in the 111 Ranch area. *Arizona Geological Society Digest* 3:141–143.
- . 1962. *Pleistocene fauna from 111 Ranch area, Graham County, Arizona*. Unpublished Ph.D. dissertation, University of Arizona, Tucson.
- Woodburne, M. O. 1977. Definition and characterization in mammalian chronostratigraphy. *Journal of Paleontology* 51:220–234.
- . 1987. Principles, classification, and recommendations. In *Cenozoic mammals of North America: Geochronology and biostratigraphy*, ed. M. O. Woodburne. Berkeley: University of California Press, pp. 9–17.
- . 1996. Precision and resolution in mammalian chronostratigraphy: Principles, practices, examples. *Journal of Vertebrate Paleontology* 16:531–555.
- Wright, D. B. 1995. Tayassuidae of the Irvingtonian Leisey Shell Pit Local Fauna, Hillsborough County, Florida. In *Paleontology and geology of the Leisey Shell Pits, early Pleistocene of Florida*, ed. R. C. Hulbert Jr., G. S. Morgan, and S. D. Webb. *Bulletin of the Florida Museum of Natural History* 37(II):603–619.

- Wright, D. B. and S. D. Webb. 1984. Primitive *Mylohyus* (Artiodactyla: Tayassuidae) from the late Hemphillian Bone Valley of Florida. *Journal of Vertebrate Paleontology* 3:152–159.
- Wright, H. E. Jr. and D. G. Frey (eds.). 1965. *The Quaternary of the United States: A review volume for the VII congress of the International Association for Quaternary Research*. Princeton, NJ: Princeton University Press.
- Wyman, L. E. 1922. Notes on the Pleistocene fossils obtained from Rancho La Brea asphalt pits. *Los Angeles Museum of History, Science and Art, Department of Natural Sciences Miscellaneous Publications* 2 (revised):1–35.
- Zakrzewski, R. J. 1967. The primitive vole, *Ogmodontomys*, from the late Cenozoic of Kansas and Nebraska. *Papers of the Michigan Academy of Science, Arts, and Letters* 52:133–150.
- . 1969. The rodents from the Hagerman Local Fauna, upper Pliocene of Idaho. *Contributions from the Museum of Paleontology, The University of Michigan* 23:1–36.
- . 1972. Fossil microtines from late Cenozoic deposits in the Anza-Borrego Desert, California, with the description of a new subgenus of *Synaptomys*. *Natural History Museum of Los Angeles County Contributions in Science* 221:1–12.
- . 1975a. The late Pleistocene arvicoline rodent *Atopomys*. *Annals of Carnegie Museum* 45:255–261.
- . 1975b. Pleistocene stratigraphy and paleontology in western Kansas: The state of the art, 1974. In *Studies on Cenozoic paleontology and stratigraphy in honor of Claude W. Hibbard*, Claude W. Hibbard Memorial Vol. 3, ed. G. R. Smith and N. E. Friedland. University of Michigan, *The Museum of Paleontology Papers on Paleontology* 12:121–128.
- . 1984. New arvicolines (Mammalia: Rodentia) from the Blancan of Kansas and Nebraska. In *Contributions in Quaternary vertebrate paleontology: A volume in memorial to John E. Guilday*, ed. H. H. Genoways and M. R. Dawson. *Carnegie Museum of Natural History Special Publication* 8:200–217.
- . 1988. Plio–Pleistocene rocks, Borchers Badlands, Meade County, southwestern Kansas. In *Centennial field guide*, Vol. 4, *South-Central Section of the Geological Society of America*, ed. O. T. Hayward. Boulder, CO: Geological Society of America, pp. 69–74.
- . 1993. Morphological change in woodrat (Rodentia: Cricetidae) molars. In *Morphological change in Quaternary mammals of North America*, ed. R. A. Martin and A. D. Barnosky. New York: Cambridge University Press, pp. 392–409.
- Zymela, S., H. P. Schwarcz, R. Grün, A. M. Stalker, and C. S. Churcher. 1988. ESR dating of Pleistocene fossil teeth from Alberta and Saskatchewan. *Canadian Journal of Earth Sciences* 25:235–245.

## 8

## Global Events and the North American Mammalian Biochronology

Michael O. Woodburne

*The prime tasks of modern historical geology are to separate the local signals from the global ones, to plot the relationships of global patterns both to time and to each other, and to search for the forces that drive these varied processes.*

—A. G. Fischer 1984, p. 129

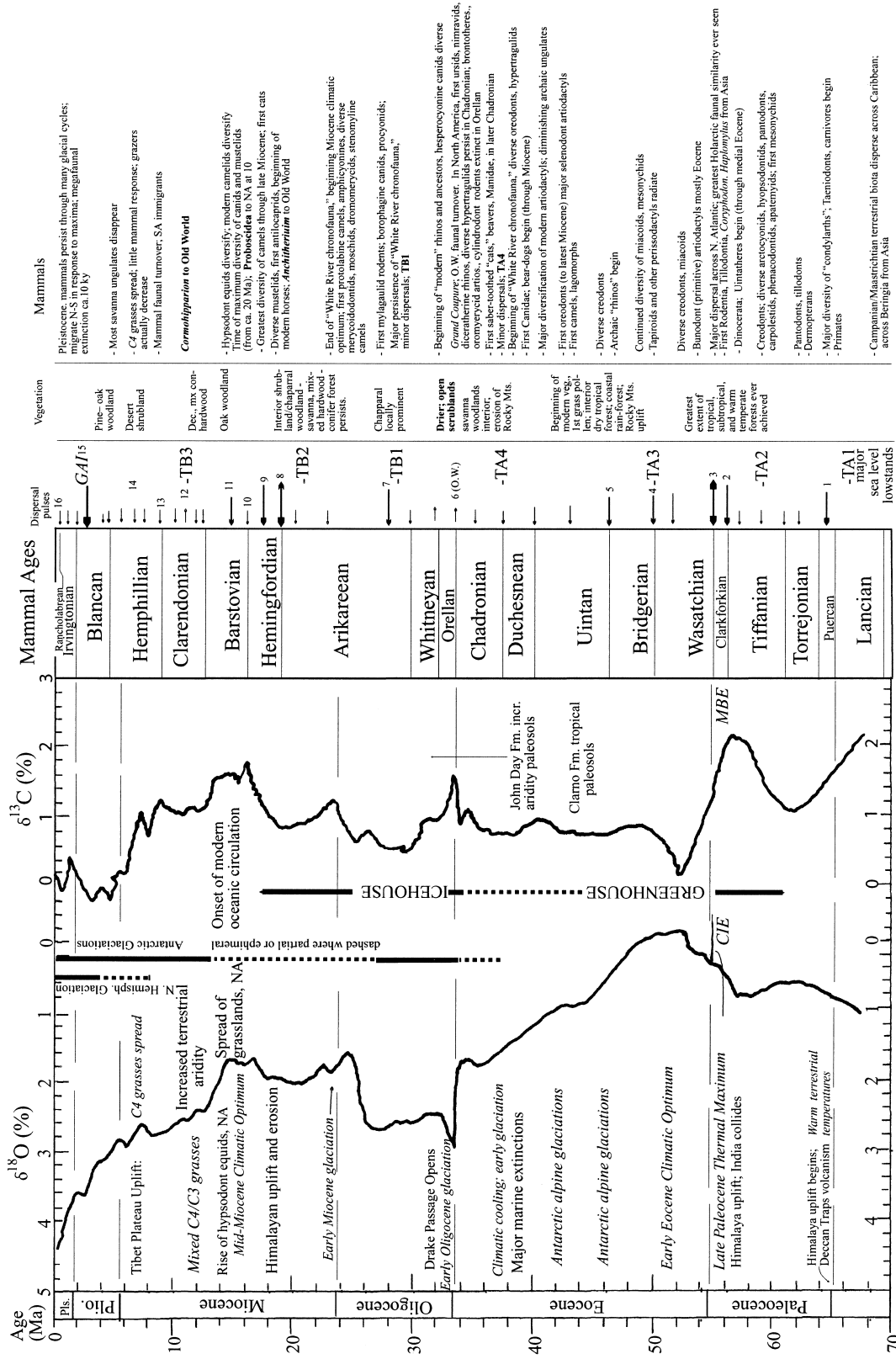
**FISCHER (1984) INTRODUCES** the concept of icehouse and greenhouse worlds to generalize global climatic settings in which glaciations were either preeminent or virtually absent. In the icehouse world, levels of atmospheric CO<sub>2</sub> would be those of preindustrial levels of this century; mean annual sea and land temperatures would be depressed; there would be greater poleward negative gradients in temperature and severity in climatic zonations; convective oceanic circulation would be active, with highly oxygenated oceans; and conditions would be conducive for the development of land and sea ice. In the greenhouse world, atmospheric CO<sub>2</sub> levels would be substantially greater than now; polar temperatures would be more equable and global temperatures elevated, with a decrease in the polar temperature–climatic gradient; oceanic circulation would be sluggish; and marine anoxia would be expected, as would be the absence of glacial ice. In general, sea level would be low during icehouse conditions, higher during greenhouse times.

Fischer (1984) correctly recognized that such cycles were of first-order magnitude (Abreu and Haddad 1998), or on the order of 50–300 m.y. in duration. The net ebb and flow of these global sea level fluctuations has been attributed to cycles in mantle convection (Fischer 1984), to assembly and dispersal of continents (Dewey and Pitman 1998), or to generally eustatic–tectonic origin (Jacquin and Graciansky 1998). Duval et al. (1998 and references cited therein) recognize two subsets of the 50- to 300-m.y. category and describe first-order (>50 m.y.) and second-order (3–50 m.y.) cycles as resulting from major continental encroachment associated with the breakup

of supercontinents and from transgression and regression reflecting changes in the rate of tectonic subsidence, respectively. Haq et al. (1988) and Graciansky et al. (1998, chart 1) suggest that the unconformities that bound these sequence packages can be correlated globally and therefore are potentially important for mammalian chronology. In a recent summary (Duval et al. 1998), third-order cycles (duration of 0.5–3 m.y.) are considered likely to be of glacio-eustatic origin, as are the fourth-order parasequence cycles (0.01–0.5 m.y. duration). The third-order sequence is the basic unit of sequence stratigraphy according to Jacquin and Graciansky (1998). Because of their global extent, such cycles are inferred to have a glacio-eustatic control, even when there is no independent evidence of the presence of major ice caps (Stoll and Schrag 1996; Gale et al. 2002). Jacobs and Sahagian (1993) proposed a mechanism whereby climatically induced fluctuations of major terrestrial lacustrine bodies could affect global sea level in nonglacial times.

Glacio-eustacy is a major factor affecting sea level changes during the Age of Mammals (here used as the time span embraced from the Late Cretaceous to the present), but aspects of tectonism and oceanic circulation also contribute importantly to climatic and other factors that affect the evolution and dispersal of terrestrial mammals. Some of these factors are summarized in figure 8.1.

Woodburne and Swisher (1995) examined the proposal that land mammal dispersals to between North America and other continents were linked to sea level lowstands and found that the tectonically active western margin of North America generally overrode the potential effect of



**FIGURE 8.1** Cenozoic global deep sea oxygen and carbon isotope records, mammal ages, climatic and tectonic events. After Zachos et al. (2001). Dispersal of mammalian taxa indicated by single- or double-headed arrows, with the length and thickness of the bar proportional to the magnitude of dispersal (see figures 8.4–8.7). Mean annual temperature and vegetation summary after Graham (1999). Mammal events from various sources, including chapters 2–7. CIE, carbon isotope excursion; GAI, Great American Interchange; MBE, Mammal Biotic Event; NA, North America; O.W., Old World; SA, South America.



the rise and fall of sea level in this regard. On the other hand, see Knox (1998) for a discussion of the interplay between tectonism and sea level change that probably resulted in the mammalian dispersal at the beginning of the Eocene across the northern Atlantic region. In addition to the tectonic influence of the Great American Interchange (GAI) at ca. 2.5 Ma (Iturralde-Vinent and MacPhee 1999), the Late Cretaceous to early Cenozoic exchange of land vertebrates between North and South America (Rage 1978; Gayet et al. 1992) also can be attributed to tectonic evolution of the Caribbean Plate (Pindell 1994; Villamil and Pindell 1998).

Abreu and Haddad (1998) provide a recent summary of the variations of isotopes of oxygen and carbon during glacial or nonglacial times. Fluctuations in isotopic ratios of both elements serve as proxies for glaciation and sea level independent of other methods or criteria.

Briefly, during glacial times the amount of  $^{18}\text{O}$  in sea water (and in foraminifera and other organisms that incorporate oxygen in their  $\text{CaCO}_3$  tests) increases as an increased amount of the lighter isotope  $^{16}\text{O}$  (which preferentially flows to the atmosphere) is removed from the oceans and trapped in glacial ice. Thus the  $\delta^{18}\text{O}$  ratio ( $^{18}\text{O}/^{16}\text{O}$ ) increases. In general, when sufficiently large  $\delta^{18}\text{O}$  ratios are measured in the  $\text{CaCO}_3$  of tests of fossil marine organisms, or even in bulk rock analyses of limestone, it is possible to infer that glaciations have taken place on land (see also Graham 1999:86–92).

Regarding carbon, the heavy isotope  $^{13}\text{C}$  is discriminated against during photosynthesis, so that all organic matter is enriched in the light isotope  $^{12}\text{C}$ . Any change in the organic carbon reservoir in sea water alters the  $^{13}\text{C}/^{12}\text{C}$  ratio recorded there. During interglacial times, when sea level is high, large amounts of  $^{12}\text{C}$  are stored in the terrestrial biomass and in sediments deposited in the flooded shelves; organisms living in the oceans record the concurrently higher (positive)  $^{13}\text{C}/^{12}\text{C}$  ratio. In glacial times, when shelves are exposed and eroded, formerly trapped  $^{12}\text{C}$  is released to the ocean and could show up in, say, bathyal organisms, with a lower (negative)  $\delta^{13}\text{C}$  ratio. Still, this relationship does not always hold true because the deep sea isotopic record commonly shows a positive shift in  $\delta^{13}\text{C}$  during periods of ice growth, coeval with the positive shift shown in the  $\delta^{18}\text{O}$  ratios, apparently caused by ventilation of the deep sea water masses during glacial periods. In the following discussion, ratios of these and other organic elements are considered as proxies for climatic change that could affect the faunal succession of Late Cretaceous and Cenozoic mammals. Figure 8.1 shows the oxygen and carbon isotopic curves for the past 70 m.y.

## “MEDIAL” THROUGH LATE CRETACEOUS

Before the Albian, a generally greenhouse world prevailed in the Cretaceous Period. As indicated by Hardenbol et al. (1998), the Late Albian through Late Campanian was overall a time of major sea level high and presumably a greenhouse world without significant polar ice, although sea surface temperatures apparently were lower than at present at 125 Ma (Frakes 1999). Huber (1998) suggests that sea surface temperatures were about  $12^\circ\text{C}$  at about  $60^\circ\text{S}$  in the Albian, rose to an average of about  $24^\circ\text{C}$  in the Turonian–Coniacian (ca. 93–86 Ma), and dropped to about  $8^\circ\text{C}$  in the Maastrichtian (ca. 67 Ma), with a rise to about  $12^\circ\text{C}$  toward the end of the Cretaceous. In this time interval Hardenbol et al. (1998) show distinctive drops in sea level at ca. 111 Ma and 102 Ma (Early and Late Albian), 95 Ma (Late Cenomanian), 91 Ma (Late Turonian), 79 and 72 Ma (Early and Late Campanian), and 67 Ma (Late Maastrichtian), for which a glacial-eustatic cause would be difficult to propose in a greenhouse world, and Frakes (1999:51) states that “there is no direct evidence for Cretaceous continent-wide glaciation anywhere on the planet.” Still, Abreu et al. (1998) suggest that oxygen isotope fluctuations in the Aptian to lower Albian and Campanian to Maastrichtian intervals (also Barrera and Savin 1999) resemble those of the medial Eocene. In that evidence has been marshaled to suggest the presence of Antarctic glaciation in the medial Eocene, the pattern similarity opens the question of a glacial cause for those oxygen isotope fluctuations in the Cretaceous. However, Immenhauser and Scott (1999) demonstrate that at least for the Albian, correlation of sea level fluctuations on a global basis is very difficult, so the pattern should be viewed with caution. Gale et al. (2002) continue to rely on glacio-eustasy as a mechanism to account for short-term fluctuations in the stratigraphic record of Cenomanian (ca. 98.9–93.5 Ma) marine strata in southeastern India and northwestern Europe, possibly driven by limited high-altitude Antarctic glaciation. And Miller et al. (1999) propose an Antarctic glacial origin for a drop in sea level (ca. 20–40 m; New Jersey) synchronous with a major global increase in  $\delta^{18}\text{O}$  at ca. 71 Ma (base of chron C31r) for the early Maastrichtian. These events apparently are coeval with a negative global  $\delta^{13}\text{C}$  excursion, which Miller et al. (1999) interpret as being caused by increased weathering of organic-rich sediments exposed by sea level lowering on continental shelves. Huber et al. (2002) evaluate deep and surface sea temperatures from the Albian through Maastrichtian from both mid- to high northern and southern paleolatitudes and conclude that both

realms show conditions that were mostly as warm as or warmer than present conditions and too warm to conform to scenarios that invoke Antarctic ice sheets, even for the cooler intervals discussed herein.

Whereas evidence of sea level fluctuation over the time span indicated earlier seems clear, as does the evidence of concurrent isotopic variations in oxygen and carbon, the effect, if any, of sea surface temperatures on continental atmosphere still is tenuous. Frakes (1999) reviews evidence in favor of episodes of cool sea surface temperatures at ca. 97, 89, and 68 Ma but cautions that the admittedly limited data on continental air temperatures do not track the marine record well. Continental air temperature data indicate a warming trend from the Early Cretaceous to a Turonian peak without the cooling in the Cenomanian (97 Ma) seen in the marine record and continued warming thereafter in higher latitudes. Dettman and Lohmann (2000) studied  $\delta^{18}\text{O}$  values of river water in Late Cretaceous and Paleogene basins of western North America and suggest that mountainous relief there was comparable to the 2.5–3 km seen today and that montane snow was present year-round at times, but whether this was controlled by or was only contemporaneous with oceanic cooling remains to be demonstrated. Upchurch et al. (1999) demonstrate the difficulties (and the power) in modeling continental climates in the Late Cretaceous, even when assumptions as to the impact of vegetation are supplied.

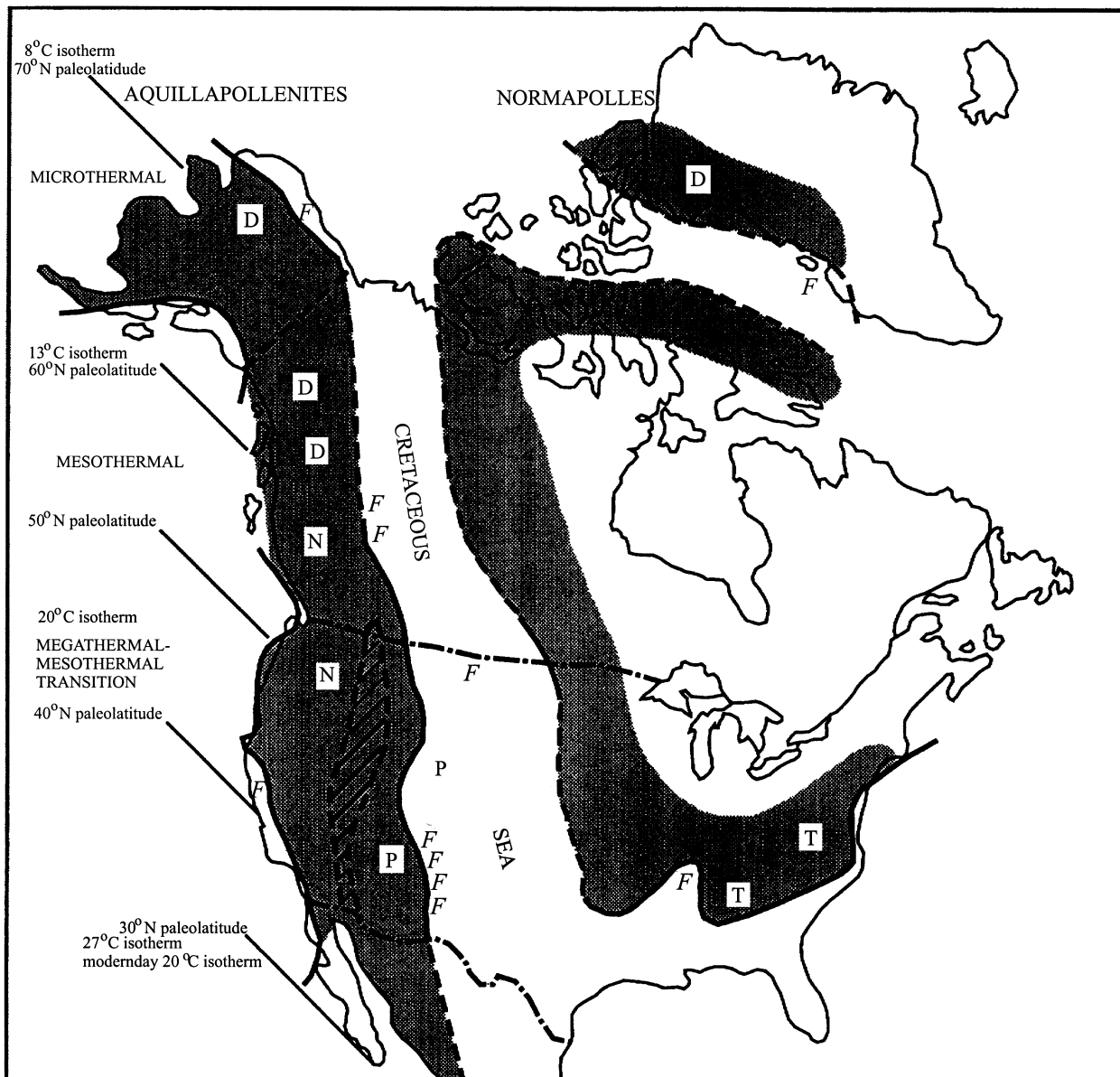
The problem remains that a glacio-eustatic solution to a drop in sea level is difficult to reconcile with general greenhouse conditions. This is exacerbated by the lack of independent evidence of glaciations proposed on a combination of eustatic and oxygen isotope data (Barrera and Savin 1999) and the still meager means of correlating atmospheric with sea surface temperatures. The evidence suggests that oceanic temperatures suffered periods of deterioration correlative with sea level drops in the mid-Campanian (ca. 79 Ma), at about the Campanian–Maastrichtian boundary (ca. 72 Ma), and in the late Maastrichtian (ca. 67 Ma) but also that climates warmed just before the beginning of the Cenozoic, possibly in response to Deccan Traps volcanism. Thus, although sea temperatures were cooler in the Maastrichtian than previously, the overall climate still was a cool greenhouse (Huber et al. 2002).

The terrestrial plant record from the Late Cretaceous through early Eocene is summarized from Graham (1999). In the Late Cretaceous, the prevailing epicontinental seaway, sited roughly athwart the present location of the Rocky Mountains, divided the terrestrial flora into an east-

ern *Normapolles* pollen group and a western *Aquilapollenites* pollen group, as summarized in figure 8.2. The *Normapolles* pollen group extended east of the Cretaceous Sea, the *Aquilapollenites* group to the west. Sparse megafossil plants were preserved mostly along the margins of the Late Cretaceous Seaway and in the Appalachian region. A polar microthermal broadleaved deciduous forest developed north of about the 60°N paleolatitude (northern Canada and Alaska), with a mean annual temperature (MAT) of 8–15°C; sea surface temperatures are estimated at 6–8°C. The poleward climatic gradient was much shallower than at present, and the deciduous nature of the vegetation suggests a seasonal regime, both as to temperature and as to light, in an overall cool-temperate climate, with north slope areas having a diverse understory and forest habitat. In southern Canada and the Pacific Northwest, this gave way to a mesothermal notophyllous broadleaved evergreen forest (MAT of about 13°C) between about 50 and 60° paleolatitude. South of about 45° paleolatitude, growth rings are absent to poorly developed, consistent with the southern (and southwestern) United States being dominated by a tropical or paratropical forest developed under a subhumid, largely aseasonal climate with an MAT of 20–25°C.

Lancian faunas best represent terrestrial mammal communities of the Late Cretaceous (chapter 2, table 2.3), with approximately 12 genera of multituberculates (cimolodontids, eucosmodontids, neoplagiulacids, and ptilodontids), a metatherian, 10 marsupials (alphadontids, “pediomyids,” and stagodontids), and placentals (cimolestids, gypsonictopids, soricomorphs, and unguatomorphs), mostly limited to sites along the western margin of the epicontinental sea (latitudinal range comparable to that of the fossil plant sites in figure 8.2). These sites fall within the areas dominated by notophyllous broadleaved evergreen forests on the north and by paratropical rainforest on the south. This appears to be reflected in northern and southern biofacies among the mammals (Weil and Williamson 2000; Clemens 2002), with the northern biofacies characterized by a greater diversity of eutherians. Most of the taxa apparently were derived endemically, with only two, *Batodon* and *Glasbius*, being allochthonous (Clemens 2002).

Case et al. (in press) include *Glasbius* as one of a small number of Lancian or Judithian marsupial lineages having a potential southern (equatorial) theater of origin or differentiation. Although the origin of *Batodon* is unclear, it appears unlikely that it was in equatorial America, based on the geographic distribution of potential sister taxa (McKenna and Bell 2000).



**FIGURE 8.2** General paleogeographic setting for North America in the Late Cretaceous, with pollen provinces (*Aquilapollenites*, *Normapolles*), distribution of floral types, Cretaceous Sea, after Graham (1999). Sites are better represented in the western than in the eastern facies but still sparse. Both show a polar broadleaved deciduous forest (D) and a southern tropical (T) or paratropical (P) forest in the south, with an intervening notophyllous broadleaved evergreen forest (N) documented in the west. This is summarized as ranging from a microthermal to mesothermal transition and megathermal climate. Isotherms and paleolatitudes are approximate. After Wolfe (1977), the polar broadleaved deciduous forest has a mean annual temperature (MAT) of ca. 8–15°C, with large, thin-textured leaves inhabiting a mesothermal to microthermal moist climate. The notophyllous broadleaved evergreen forest reflects an MAT of about 13°C and a mean coldest-month temperature of ca. 1°C, with sclerophyllous vegetation lacking dip tips and having 40–60% entire-margined leaves. The paratropical rainforest has an MAT of 20–25°C, lacks a dry season, and is composed mainly of broadleaved evergreen trees, with some deciduous and with 50–75% entire-margined leaves. The tropical forest has an MAT of about 25°C in a sub-humid, aseasonal climate, with broadleaved evergreen, single-tiered open-canopy vegetation, mostly entire-margined leaves. F, fossil plant site.

## PALEOGENE

As indicated in figure 8.1, oceanic temperatures generally rose across the Cretaceous–Tertiary boundary and progressed toward the early Eocene climatic optimum, by which time the sea temperature was about 12°C. Kennett and Stott (1995, figure 5.1) indicate that the temperature of Antarctic surface waters was ca. 8–10°C based on  $\delta^{18}\text{O}$  data in planktonic foraminiferans in the early Paleocene and increased to ca. 13–14°C just before the late Paleocene thermal maximum (LPTM; Zachos et al. 1993), associated with the global carbon isotope excursion (CIE), dated at 55.5 Ma (Bains et al. 1999). During the LPTM, deep ocean water temperatures were more than 15°C, surface waters in the high latitudes more than 20°C (Corfield and Norris 1998). The oceanic carbon isotope ( $\delta^{13}\text{C}$ ) values dropped about 2.5 per mil during the CIE, perhaps in three steps during an interval of about 10,000–20,000 years (Bains et al. 1999). The CIE implies that the global ocean experienced an abrupt input of a large amount of isotopically light carbon at this time, which Rea (1998) and Bains et al. (1999) suggest having resulted from the release of large amounts of biogenic methane into the ocean. The best candidate for this gas is proposed to be methane hydrates preserved in sediments beneath the sea floor in the subtropical western North Atlantic Ocean, with release promoted by erosion- or impact-triggered failure of the sea floor along continental slopes, perhaps aided by warming of deep ocean temperatures, which converted the once-solid hydrates to the gaseous state. Hudson and Magoon (2002) propose a second but not necessarily alternative source for the generation of atmospheric methane: heating of subducted hydrocarbon-producing sediments along coastal Gulf of Alaska from about 65 to 55 Ma. However, Clift and Bice (2002) elaborate that methane was not the sole trigger for global warming in the late Paleocene to early Eocene, with an increase of atmospheric  $\text{CO}_2$  being a likely candidate from volcanic or tectonic sources. Thomas et al. (2002) favor a model wherein a general late Paleocene trend in warming of oceanic surface and intermediate-depth waters led, by downwelling or subduction, to thermally induced dissociation of methane hydrates in an unspecified deeper source location. In any case, the onset of the CIE was associated with the Benthic Foraminiferal Extinction (BFE), the Kaolinitic Event (KE), and the Eolian Dust Event (EDE) and coincided with the Mammal Biotic Event (MBE) mammal dispersal event at Wao (Berggren et al. 1998; Zachos et al. 2001).

Thomas (1998) describes the BFE as a major, rapid global extinction wherein 30–50 percent of benthic

foraminifera at middle bathyal or greater depths became extinct in a few thousand years. Factors contributing to the extinction include changes in deep sea circulation, increased corrosion of  $\text{CaCO}_3$  related to the strong increase in isotopically light carbon, increased temperatures, low oxygenation, and changes in productivity patterns.

As discussed by Rea (1998), a dramatic reduction in size of eolian dust grains (the EDE) from North Atlantic and southern Indian oceanic deep sea cores at the time of the LPTM signifies a strong reduction in global wind intensity, in sharp contrast to the vigorous atmospheric circulation patterns interpreted for the Late Cretaceous and all but latest Paleocene. Based on these data, atmospheric circulation remained sluggish for most of the Eocene, apparently reflecting a significant reduction in the pole-to-equator temperature gradient, but began to reinvigorate in the icehouse world of the later Eocene and Oligocene. The KE is the sharp increase of kaolinitic clays that occurred globally in synchrony with the LPTM, apparently indicating an episode of even higher humidity and intensity of chemical weathering than otherwise characteristic of the early Eocene (Sloan and Thomas 1998:139).

Thus the LPTM saw a major reorganization of global patterns largely attributed to global warming but also to the CIE as isotopically light carbon was released into the oceans and then into atmosphere. Depending on the definition of the series–epoch boundary, these events transpired at the end of the Paleocene or beginning of the Eocene (see chapters 2 and 3).

The bolide impact at the K–T boundary (Alvarez et al. 1980) probably contributed to but arguably was not the major cause of the mammalian extinctions at that time, with impact dust–induced curtailment of photosynthesis viewed with some skepticism (Pope 2002; also see Clemens 2002 for an important review). Regardless of cause, Clemens (2001, 2002) records that the mammal fauna of the beginning Puercan (Pu1) was reduced in diversity by about 60 percent and that about 18 percent of the mammalian fauna survived from the Lancian. Early Paleocene floras in the vicinity of the southern Rocky Mountains (New Mexico) show the fern spike, followed by Paleocene recolonization by angiosperms (Wolfe 1987; Wolfe and Upchurch 1987), indicating a paratropical rainforest with an MAT of about 22°C, whereas farther north (Alberta) plant diversity decreases across the K–T boundary, but a fern spike is absent. The notophyllous broadleaved evergreen forest (figure 8.2) of the Late Cretaceous was replaced in those paleolatitudes by the broadleaved deciduous forest, although the climate still was mesothermal. Still, the overall climatic setting of the

Late Cretaceous was not markedly different from that of the early Paleocene, at least in the areas represented by fossil mammals, with rainfall on the order of 1000 mm annually. Graham (1999, table 5.5) indicates that the southeastern United States supported a tropical rainforest, with an MAT of 27°C, that gave way to the northeast to a notophyllous broadleaved evergreen to polar broadleaved deciduous forest, with an MAT of 15–20°C. Farther inland, early to late Paleocene floras of the eastern Rocky Mountains ranged from polar broadleaved deciduous forests in the north to paratropical rainforest and notophyllous broadleaved evergreen forests in the south (MAT 13–15°C to 18°C). In Alberta and the Yukon, polar broadleaved deciduous and notophyllous broadleaved evergreen forests lived under an MAT of 10–15°C. In general, a warming trend extends from the Tiffanian (MAT of 10°C) to the Clarkforkian (MAT of ca. 13.5°C) in the Rocky Mountains district (e.g., Bighorn Basin, Wyoming, figure 8.3), with mesothermal–megathermal broadleaved deciduous vegetation giving rise to megathermal evergreen tropical forests. The earlier Paleocene floras from the Ravenscrag Formation (figure 8.3) of Saskatchewan (i.e., north of about 50° paleolatitude) indicate a microthermal climate with near-freezing temperatures reached in January. Overall, floral indicators show a discernible north–south zonation in the Paleocene but also are consistent with the general warming trend toward the early Eocene climatic optimum (figure 8.1). From a tectonic standpoint, Jerzykiewicz and Sweet (1988) indicate pulses of uplift in at least the northern Rocky Mountains during the interval from 80 to 60 Ma, and by the early Eocene the range generally had been elevated to about half its present height (Graham 1999), so the previous lowlands and epicontinental sea were disrupted. A comparable scenario can be applied to the Front Range and southern Rocky Mountains as well (Pazzaglia and Kelley 1998; see also Dettman and Lohmann 2000).

As indicated in chapter 3 (table 3.2), early Puercan faunas continue to share with Lancian ones the ptilodontid, eucosmodontid, and taeniolabidid multituberculates, didelphid and glasbiid marsupials, and cimolestid didelphodontan, cimolestan, and lipotyphlan placentals (also Clemens 2002; classification follows McKenna and Bell 1997). The marsupials declined strongly from the Lancian in Pu1, with only *Peradectes* representing the group that formerly included alphasodontines (*Alphadon*, *Protalphadon*, and *Turgidodon*), pediomyids (“*Pediomyis*”), glasbiids (*Glasbius*), and stagodontids (*Didelphodon*). New groups for Pu1 include leptictids (?*Prodiacodon*) and a variety of ungulates (*Oxyprimus*, *Protungulatum*, *Baiococonodon*, and *Mimatuta*), although

their novelty may be more local than revolutionary. Clemens (2001) indicates that about 70 percent of the Pu1 mammals of northeastern Montana were aliens (lineages not known in the Lancian of the western northern interior), so their point of origin is not clear; they include *Acheronodon*, *Catopsalis*, *Stygimys*, ?*Prodiacodon*, *Protungulatum*, *Oxyprimus*, *Baiococonodon*, and *Mimatuta*. *Stygimys* may be represented in the Campanian El Gallo Fauna (Baja California del Norte), so the group to which it belongs can be considered to be known from pre-Pu1 faunas of North America generally, but not in the western northern interior. Similarly, *Protungulatum*, *Oxyprimus*, *Baiococonodon*, and *Mimatuta* also have Campanian representatives in southern districts (Baja California and Utah; Clemens 2002). The multituberculate *Catopsalis* and primitive ungulates of possible Lancian age occur in Saskatchewan, suggesting that western interior faunas were different from precursor or possibly contemporaneous suites in peripheral areas such as these. Whereas climatic indicators may indicate a biofacies contribution from that source, the factors that contributed to the Baja California element are less clear. Lying outside the San Andreas fault zone, Baja California was at least 300 km southeast of its present location in the Late Cretaceous. Thus, although immigration played a role in the development of early Puercan mammal faunas in the western interior, the home of the immigrants may not have been outside of North America in most (possibly all) cases.

About 400,000 years later, Pu2 saw the most dramatic increase in mammal diversification for the Paleocene, and, again, a major component of this increase resulted from immigration. Compared with Pu1, new taxa cited in chapter 3 for Pu2 are as follows (\*\*= alien, R = resident, U = unknown in Clemens 2002; \* = immigrant in 1987 volume). The multituberculates include *Neoplagiaulax* U, *Ectypodus*\*\*, *Parectypodus*\*\*, *Kimbetohia*, *Ptilodus*\*\*, *Xyronomys* U, and *Microcosmodon* R. Eutherians include the cimolestid *Alveugena*; the taeniodonts *Onychodectes*, *Schochia*, and *Wortmania*; the primates *Purgatorius*\* and *Pandemonium*\*; the oxyclaenids *Carcinodon*\*, *Chriacus*\*, and *Oxyclaenus*\*; the arctocyonids *Platymastus*, *Desmatoclaenus*, *Loxolophus*, and *Mimotricentes*; the hyposodontid *Litomylus*\*\*; the mioclaenids *Ellipsodon*, *Choeroclaenus*, *Bubogonia*\*\*, *Tiznatzinia*, and *Promioclaenus*; and the periptychids *Anisonchus*, *Haploconus*, *Mithrandir*, *Hemithlaeus*, *Ectoconus*, *Alticonus*, *Periptychus*, *Tinuviel*, *Oxyacodon*, and *Conacodon*. Based on Clemens (2002) it appears that cimolestids, periptychids, mioclaenids, arctocyonids, and taeniodonts were generally endemic to North America, but other taxa



**FIGURE 8.3** Map of western North America showing location of paleofloral sites discussed in the text. After Graham (1999). Also shows south coastal region of California that yields Eocene mammals.

(multituberculates, primates, oxyclaenids, hyopsodontids, and mioclaenids) were immigrants, probably from elsewhere in North America, if not Holarctica. The (neoplagiulacoid and eucosmodontid) multituberculates are otherwise endemic to North America in the Late Cretaceous and early Paleocene (including Torrejonian). Primates, oxyclaenids, and hyopsodontids are shared with other regions (McKenna and Bell 1997), but still may have their earliest Paleocene records in North America. Pu2 records a strong immigration pulse at a time (1, figures 8.1, 8.4) between major episodes of lowered sea level. If that pulse is relative only to interior North America, sea level may be irrelevant. For Pu3, the multituberculate *Taeniolabis*\*\* apparently is an immigrant, but the euthe-

rians (the leptictids *Prodiacodon* and *Palaeictops*, the cimolestan *Ravenictis*, the hyopsodontid *Haplaletes*, and the triisodontid *Goniacodon*) are not.

In the Torrejonian and Tiffanian, cimolestids, apatemyids, pantolestids, and arctocyonids are diverse; hyopsodontids undergo a radiation from the Puercan; pantodonts, erinaceomorphs, soricomorphs, plesiadapid primates, paromomyid dermopterans, carpolestid euprimates, and phenacodontids diversify in the Tiffanian (from Torrejonian first occurrences); and periptychids diminish relative to their Puercan abundance (e.g., chapter 3, table 3.2). Immigration was of minor importance in the Torrejonian and Tiffanian (figures 8.1 and 8.4), despite a major sea level drop (Ta2) at about Ti3. As for

TIME (Ma)	CHRONS	POLARITY	EPOCH	AGE	OTHER DATA	MAMMAL AGES	DISPERSAL/TAXA	
52	C23	Black	EOCENE EARLY	YPPESIAN	Early Eocene Climatic Optimum	Lostcabinian Wa7	<i>Lambdotherium, Eotitanops</i>	
		Lysitean Wa6				<i>Heptodon</i>		
		Wasatchian Wa5						
54	C24	Black			Carbon isotope excursion; Late Paleoc. Thermal Max.	Greybullian Wa4	<i>Homogalax</i>	
				Sandcoulean Wa2		3 <i>Apatemys*, Macrocranion, Cantius, Palaeosinopa*, Didelphodus, Arfia, Prototomus, Didymictis*, Palaeonictis*, Miacis, Pachyaena**, Hyopsodus*, Hyracotherium, Diacodexis</i>		
				Clarkforkian Wa0/Wa1		2 <i>Rodentia, Tillodontia, Coryphodon</i>		
56						Cf3		
						Cf1		
58	C25	White	PALEOCENE LATE	SELIANDIAN	Ta2 -	Ti6		
							Ti5	<i>Prodinoceras, Arctostylops (Asia)</i>
							Ti4	
60	C26	Black				Ti3	<i>Chiromyoides</i>	
						Ti2		
62	C27	White	PALEOCENE EARLY	DANIAN	Ta1 -	Ti1	<i>Plesiadapis</i>	
							To3	
							Torrejonian To2	<i>Dissacus, Ankalagon, Pantolambda, Deltatherium</i>
64	C28	Black				63.90 ± 0.04 To1		
						64.11 ± 0.02 Pu3	<i>Taeniolabis</i>	
						64.77 ± 0.06 Pu1	1 <i>?Purgatorius, Pandemonium, Carcinodon, Chriacus, Litomytus?</i>	
66	C29	White					- <i>?Oxyclaenus</i>	
68	C30	Black	CRETACEOUS	MAESTRICHIAN		Lancian	1, 2 = major dispersal event	
	C31n	Black					<i>Glasbius, Batodon</i>	

FIGURE 8.4 Paleocene time scale, mammal ages, and dispersal of mammalian taxa. Time, chrons, polarity, epochs, and ages after Berggren et al. (1995). Mammal ages and dispersal/taxa after chapters 2–4. Ta1 and Ta2 are times of major sea level fall, after Haq et al. (1988) for terminology and Hardenbol et al. (1998) for chronology. \* = taxa known from the Paleocene of North America, \*\* = from the Paleocene of Asia but involved in Holarctic dispersal at Wao.

North America, the Paleocene faunas of Asia are largely endemic (Ting 1998). Following chapter 3, members of the Mesonychia (*Ankalagon*, *Dissacus*), Pantodonta (*Pantolambda*), and Tillodontia (*Deltatherium*) are likely immigrants from Asia during the Torrejonian (To2), suggesting a correlation with the Shanghuanian mammal age (Ting 1998). The Nongshanian mammal age (Ting 1998) includes the first occurrence in Asia of Arctostylopida, Dinocerata, Phenacolphidae, and Ernandontidae (?Edentata), and the first two of these groups first appear in T15 of North America at ca. 57 Ma, about 2 m.y. later than the time of a major sea level lowstand (Ta2; figures 8.1 and 8.4). As indicated in chapter 3, the Asian first occurrence of these taxa may not be synchronous with the North American record, but their immigrant status seems clear in North America. The Clarkforkian heralds a major dispersal event from Asia, with Rodentia, Tillodontia, and Pantodonta (*Coryphodon*) making a dramatic appearance (2, figures 8.1 and 8.4). According to chapter 3 (table 3.2), Rodentia include *Alagomys*, *Acritoparamys*, *Paramys*, and *Apatosciuravus*; Tillodontia include *Esthonyx*. Other elements of Clarkforkian faunas record persistence of groups that occurred earlier, but oxyaenids, nyctitheriids, micro-momyids, and pauromomyids are more diverse than in Tiffanian faunas, and new hyopsodontid condylarths appear.

Eberle and Lillegraven (1998) point out that in the Puercan, mammal species of the Hanna Basin, Wyoming (figure 8.3), were more similar (54 percent) to those of southern districts than to those to the north, exemplified by both multituberculates and eutherians (“ungulatomorphs”). This is approximately coincident with the boundary between the paratropical dry forest and evergreen tropical rainforest in the southern part of the United States and the northern microthermal polar broadleaved deciduous forest (see *P* in the Cretaceous Sea on figure 8.2 for location of that boundary, if not marine conditions in the Paleocene). As indicated in chapter 3, the geographic distribution of North American Paleocene mammal faunas younger than Puercan is limited, so currently there is little documentation of paleozoogeographic facies for this time frame. In Torrejonian interval To3 plesiadapid and carpolestid primates may be dominant in northern faunas, whereas *Tetraclaenodon*, *Periptychus*, and *Mixodectes* may dominate in southern faunas.

Chapter 4 illustrates that there is some disagreement about the location of the Paleocene–Eocene boundary not only with respect to stratotypic relationships in the London/Paris Basin but also with respect to the continental mammal record in North America. Figures 8.4 and 8.5

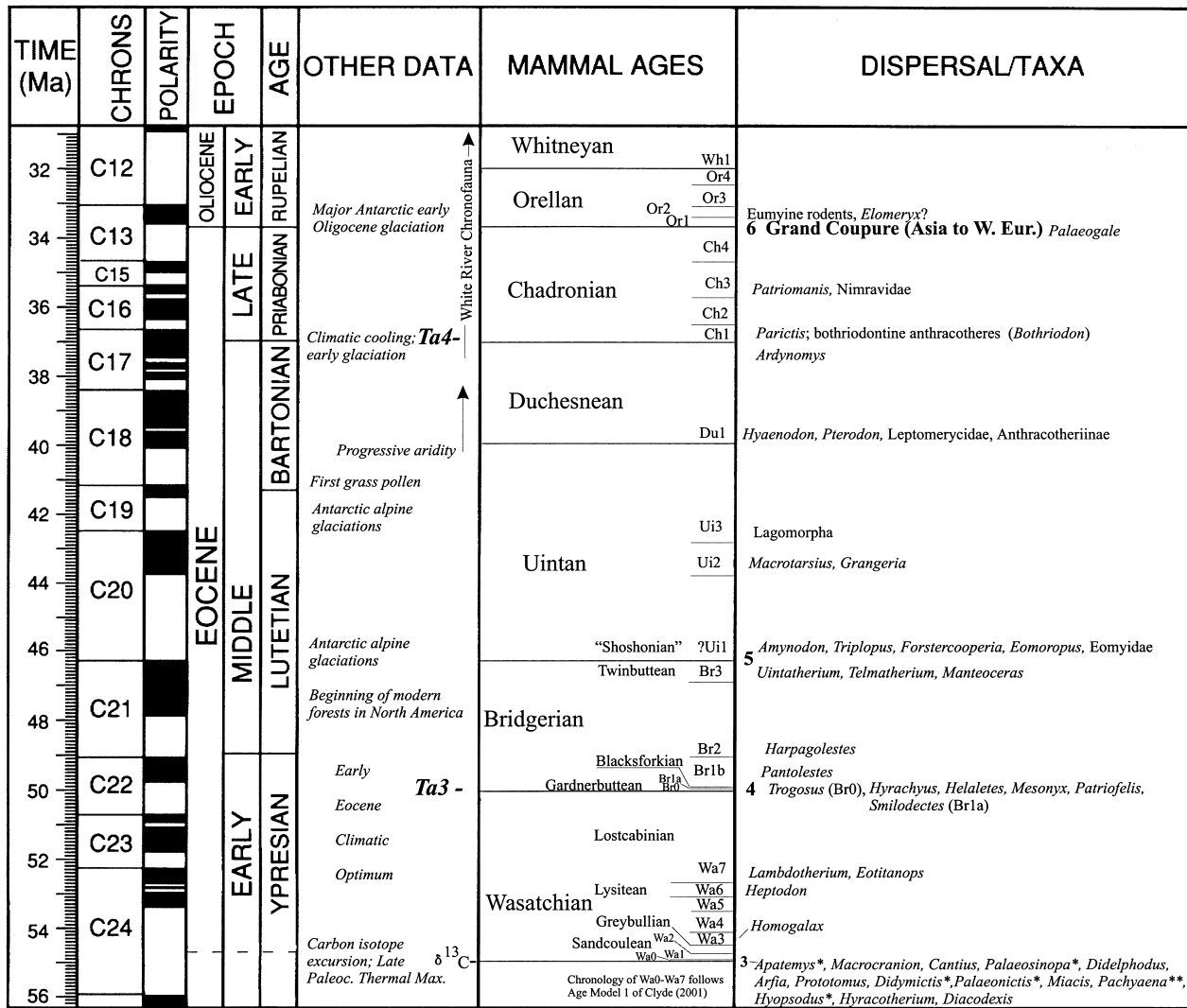
indicate that the base of the Eocene corresponds to the base of the Ypresian Stage/Age at ca. 54.7 Ma, whereas the Clarkforkian–Wasatchian boundary corresponds closely with the CIE, dated at ca. 55.5 Ma. Under that scenario, the earliest Wasatchian (Wao) is latest Paleocene in age. Alternatively, if the Paleocene–Eocene boundary is correlated to the CIE, then Wao would be earliest Eocene (see also Aubry et al. 1999; Aubry 2000). Regardless of a decision on this topic, Wao faunas witnessed a major intercontinental dispersal between North America and Europe and Asia, to which a combination of plate tectonic and local geologic events contributed (McKenna 1983a, 1983b; Knox 1998), but apparently was not coincident with a major sea level lowstand (figure 8.5).

The early Eocene is warmest period in the Paleogene, culminating the trend from the later Paleocene. Sea temperatures around Antarctica still are warm (Ehrmann and Mackensen 1992), and in Australia this is the time of maximum development of nonseasonal megathermal rainforests and year-round high humidity consistent with global warming (MacPhail et al. 1994:242–246). The interpretation for a generally broad interval of warmth during the early Eocene (ca. 55–49 Ma, early Eocene climatic optimum, figure 8.1) is based on low  $\delta^{18}\text{O}$  global sea water values (Browning et al. 1996:639, 640; Zachos et al. 2001).

As summarized in Graham (1999), tropical, subtropical, and warm temperate forests dominate the United States interior in the early Eocene. Gunnell (1997) indicates that the Wasatchian climate was characterized by warm, subtropical MATs (12–18°C) and forests and by closed conditions in the Bighorn Basin, Wyoming (figure 8.3). This climatic setting was at least conducive to the wave of dispersals that resulted in the greatest similarity of Holarctic mammal faunas ever achieved (Savage and Russell 1983:67), apparently along with a significant immigration of floral elements from Europe (Frederiksen 1988), with both possibly coeval with the LPTM.

Wasatchian faunas reflect an early phase in the progressive modernization of land mammals in North America and a reduction in some typical Paleocene groups. Thus these faunas show a loss of about 50 percent of multituberculate families, a diminution of archaic giants and primitive Paleocene elements (taeniodonts, pantodonts, and pantolestans), the last oxyclaenid and arctocyonine, and diminished hyopsodontid condylarths. These changes were countered by an increase in herpetotheriine marsupials; a development of more modern rodent groups (castorimorphans, sciruravids, and cylindrodontids); a radiation of new creodonts, miacid carnivores, phenacolemurine dermopterans, and notharctine and omomyoid primates; and the beginning of the artiodactyl and perissodactyl radia-





**FIGURE 8.5** Eocene time scale, mammal ages, and dispersal of mammalian taxa. Time, chrons, polarity, epochs, and ages after Berggren et al. (1995). Mammal ages and dispersal/taxa after chapters 4–5. Ta3 and Ta4 are times of major sea level fall. \* = taxa known from the Paleocene of North America, \*\* = from the Paleocene of Asia but involved in Holarctic dispersal at Wao). *Palaeogale* is first known from the Chadronian but apparently dispersed to Europe at about the Whitneyan (Baskin and Tedford 1996; also see this reference for *Parictis*). Nimravidae as immigrants suggested by Bryant (1996). *Patriomanis* after and Gaudin and Emry (2002).

tions, including the first equids, brontotheres, chalicotheres, hyracodontid rhinos, and helaletid and iscolophid tapiroids.

According to Gingerich (1989, 2001) and Strait (2001), Artiodactyla (*Diacodexis*), Perissodactyla (*Hyracotherium*), Primates (*Cantius*), and hyaenodontid Creodonta (*Prototomus*, *Arfia*) appear in Wao faunas (3, figures 8.1, 8.4, and 8.5), have no local precursors, and probably are of European origin. Bowen et al. (2002) suggest an Asian origin for those groups, however, so the North Atlantic avenue to Europe (McKenna 1983a, 1983b) may not have been the sole or most important dispersal pathway at this time. Other Wao novelties include *Macro-*

*cranion* (erinaceomorph lipotyphlan), *Miacis* (miacid carnivoran), and *Pachyaena*\*\* (mesonychid), all of which are shared with Europe. Additional Wao taxa (*Apatemys*\*, *Palaeonictis*\*, and *Didymictis*\*) are known from the Paleocene of North America but apparently dispersed to Europe at Wao time (\* = taxa known from the Paleocene of North America, \*\* = from the Paleocene of Asia). Wao taxa *Hyopsodus*\* and *Diacodexis* also occur in Asia (Ting 1998). In chapter 4, Wasatchian taxa, *Didelphodus*, and *Palaeosinopa*\* are indicated as shared with Europe. To summarize, the early Wasatchian witnessed a major overland dispersal across Holarctica, but the vectors are not always clear. Subsequently, *Homogalax* (Graybullian) is

shared with Asia at that time (Ting 1998); *Heptodon* is shared with Asia in the Lysitean (Ting 1998), so the very limited late Wasatchian Holarctic dispersals reflect European isolation relative to North America.

In the medial and later Eocene (49–36 Ma), a general cooling trend culminated in the early Oligocene with a reduction in surface Antarctic water temperatures by 10–12°C (figure 8.1) and deeper waters by 3–5°C (Ehrmann and Mackensen 1992). In Australia this is the time of maximum development of nonseasonal mesothermal rainforests, associated with global cooling (MacPhail et al. 1994), and at 46 Ma (early medial Eocene), ice-rafted debris in cores adjacent to East Antarctica indicate at least alpine glaciation with ice down to within 1000 m above sea level (Ehrmann and Mackensen 1992). In New Jersey (Browning et al. 1996), hiatuses at 48 and 46.5–44 Ma apparently are associated with increases in  $\delta^{18}\text{O}$  levels, indicating glacial action and cooler sea temperatures. Sea level falls at these times are considered to have had an Antarctic glacial origin, as are sea level drops at ca. 43–41 Ma (indicated as times of glaciation, but not major sea level drops on figure 8.5). All of these would significantly predate the generally accepted date of the opening of the Drake Passage at ca. 34–36 Ma but would be approximately consistent with the evidence of Ehrmann and Mackensen (1992).

Whereas the present Rocky Mountain region was a seaway or otherwise lowland from the Cretaceous (e.g., figure 8.2) into the Paleocene, the growth and development of a continental divide are manifested by floras at least by Bridgerian time. The Green River flora (figure 8.3; ca. 47 Ma) of northwestern Colorado and adjacent areas was a woodland–savanna with affinities to seasonally dry subtropical forests of the Gulf region, in contrast to the Germer flora of Idaho (figure 8.3), which is a mixed deciduous hardwood–conifer forest with affinities to more western associations. Altitudinally zoned elements of the Green River flora indicate the presence of lake margins, swamps, and floodplains at about 300 m elevation (according to MacGinitie 1953, but see Wolfe et al. 1998 for an estimate of 2.9 km), giving way to drier, better-drained savanna oak–pine woodland, followed by broadleaved deciduous forest, overlapped with a final mixed hardwood–coniferous forest zone from 1000 m to about 1700 m. Seasonal rainfall is thought to have reached ca. 700 m (Graham 1999). According to Graham (1999) the Germer flora (ca. 46 Ma) was deposited at about 600 m (but possibly three times as high) and shares elements (*Abies* [fir] and *Picea* [spruce]) of the present western montane coniferous forest with other sites to the west (Thunder Mountain, figure 8.3), where the MAT is as low as 8.5°C at ele-

vations of ca. 2200 m at treeline and precipitation of summer rain and light winter snow reaches 1100–1500 mm annually. Wolfe et al. (1998) indicate that the Salmon flora (figure 8.3), dated at ca. 41 Ma (and from the same Challis volcanics as the Germer flora), is estimated to have lived at elevations up to about 2 km with an MAT of 2–10°C. This elevation is compatible with the MAT for the Thunder Mountain flora, Idaho, and Wolfe et al. (1998) reconstruct paleoelevations of 2.0–2.9 km for sites ca. 48–50 m.y. old on both sides of the modern continental divide, with MATs ranging from 8.3°C on the northwest (One Mile Creek, southern British Columbia, figure 8.3) to 15–17°C on the southeast (Kisinger Lakes, western Wyoming, figure 8.3). With elements of the Green River flora (*Ephedra* [Mormon tea], *Celtis* [Hackberry], and *Ocotea* [Laurel]) having affinity with those from subhumid habitats more characteristic of subsequently more arid settings, both provinces record the continuing modernization of regional floras during the later Eocene.

In addition to the inauguration of a continental divide, the middle Eocene also witnessed a diversification of floral associations as compared with the earlier Eocene. Graham (1999) indicates that an important modernization of North American land plant communities begins in the medial Eocene during the initial phases of transition from greenhouse to icehouse climates (figure 8.1). In the later part of the Bridgerian (Br<sub>3</sub>), MATs decreased somewhat (still in the range of 11–19°C; Gunnell 1997), possibly coincident with the Antarctic alpine glaciations recorded about then (figures 8.1 and 8.5), and more open wooded conditions may have prevailed. Graham (1999) describes the pollen record as indicating episodes of change about synchronous with the Antarctic alpine glaciations at about 46 and 42 Ma (figure 8.5), with the second interval recording not only the first grasses (pollen) but also the elimination of many of the previously common arborescent taxa (see also Frederiksen 1988 but use time scale calibrations of Berggren et al. 1995).

In general a semideciduous tropical dry forest occupied much of the continental interior as a reflection of reduced and more seasonally distributed rainfall than previously. The eastern coastal region (southeastern United States) experienced a middle Eocene MAT of 24°C, which decreased to about 13°C at Ellesmere Island, a gradient of 0.28°C/1° latitude (comparable to that of the Late Cretaceous but somewhat lower than for the early Eocene). This is generally comparable to the western coastal gradient, where an MAT of 27°C in northern Pacific California (Susanville, figure 8.3) decreased to 15–16°C in northern lowland Alaska (or even cooler: 12–13°C in montane Washington State; Graham 1999). In

Vancouver, British Columbia, mixed conifer, paratropical rainforest, and notophyllous broadleaved forests prevailed, with more northern regions (Alaska) clothed in a polar, broadleaved deciduous forest.

Tropical rainforest occupied west coastal California and Washington in the medial Eocene. The Susanville flora reflects a coastal rainforest environment, with such conditions persisting to the end of the Eocene. In central Oregon, the Clarno flora (ca. 44 Ma) is a paratropical association (e.g., above) with only a few temperate forms, living under humid conditions that lack a coastal influence (Coast Ranges and Cascade Mountains not yet present). The upper part of the Clarno Formation (ca. 44 Ma; figure 8.3) begins the base of a succession interpreted (Bestland et al. 1997) as recording a transition in soil-forming regimes that track the onset and development of more arid conditions from originally nearly tropical climates reflective of the early Eocene climatic optimum. The sequence is developed mostly in the lower part of the John Day Formation (figure 8.3), which unconformably overlies the Clarno, but the upper Clarno Formation records deeply weathered paleosols developed under a tropical to paratropical climate (Bestland et al. 1997:165) at ca. 44 Ma. Subsequently, subtropical conditions prevailed at ca. 43–42 Ma, followed by paleosols developed in the John Day Formation from ca. 40 Ma that show that subtropical to humid temperate climates obtained by 34 Ma and humid temperate to subhumid temperate conditions by 30 Ma (Bestland 2002). Bestland et al. (1997) interpret the paleosol evidence as reflecting a climatic change to drier and cooler conditions coincident with glacially controlled global cooling during the 44–30 Ma interval, comparable to the summary of Wolfe (1992, 1994b).

Middle Eocene floras of coastal southern California reflect the drying trend seen in the continental interior, with rainfall in the range of 500–1000 mm annually, supporting a savannalike vegetation with an understory of shrubs and herbs rather than grasses (the first megafossil evidence of which is of Miocene age, late middle Eocene grass pollen notwithstanding; figure 8.5), and a diversity of mesic trees in gallery forests. In contrast to southeastern U.S. floras, grass pollen was not represented in the southern California associations in the late Eocene, and at the very end of the Eocene these floras show the beginning of the Madrean scrubland or chaparral, woodland, and savanna vegetation characteristic of the present arid southwestern United States and adjacent Mexico.

Although Bridgerian mammal faunas still are almost nonexistent in southern California, the climatic settings discussed here apparently set the stage for provincial dif-

ferentiations found in the Uintan (below). The few taxa of late Bridgerian Clarno Nut Bed Fauna, Oregon (figures 4.1 and 8.3), show correspondence with those of the continental interior (Hanson 1996) and therefore do not illuminate mammalian response to the regional floral differentiation under way at that time, as summarized earlier in this chapter. The geographic clustering of existing Bridgerian faunas apparently leads to general discussion of faunal dynamics in a temporal rather than geographic context (Prothero and Heaton 1996; Janis et al. 1998a–1998c; Prothero 1999).

Bridgerian faunas can be contrasted with those of the Wasatchian by having the last pantodonts and the first leptocherid artiodactyls, along with a radiation of dichobunids, a diversity of sciuravid rodents, omomyid and notharctine primates, hyaenodontid creodonts, and brontotheriids, along with oxyaenid procreodi, tillodonts, helaletid tapiroids, and hyracodontid rhinos.

Middle Eocene mammalian dispersals were generally sporadic, except for pulses in the early Bridgerian (ca. 50 Ma; Smith et al. 2003) and latest Bridgerian–early Uintan (ca. 46 Ma). Correspondence between early Bridgerian dispersals (4, figures 8.1 and 8.5) and a major sea level lowstand (Ta<sub>3</sub>) is suggested. Most Bridgerian immigrants show affinities with Asian faunas.

By the late Eocene, vegetation records an overall shift to drier forests, at least in the United States, with the climatic setting in areas east of the Rocky Mountains reflecting seasonally dry, if tropical, conditions. The generally more arid conditions in the Clarno–John Day region (but still humid-temperate) have been reviewed earlier in this chapter. Graham (1999) and Wolfe et al. (1998) suggest that segments of the Rocky Mountains and Basin and Range provinces had achieved modern elevations at that time, if not in the medial Eocene. In that context, the more northerly situated Copper Basin flora of northeast Nevada (ca. 40 Ma; figure 8.3) contrasts with the approximately coeval Florissant flora of Colorado (ca. 35 Ma; figure 8.3) to the southeast. The Copper Basin flora is a microthermal mixed deciduous–conifer association that apparently occupied a montane, lakeside setting, with an interpreted MAT of 11°C at an elevation of ca. 1200 m (MAT 10.5°C, elevation ca. 2.0 km; Wolfe et al. 1998), and rainfall estimated between ca. 1300–1500 mm annually. The Florissant flora is summarized (Graham 1999) as a stream and lake margin mesic forest with a drier evergreen oak–pine woodland on the higher ground, comparable to the dry savanna associations of western Texas and adjacent Mexico. Annual rainfall (mostly in summer, with a dry winter) is estimated at 500–635 mm, with an MAT of 18°C. Even though the Florissant has been

subsequently interpreted as having an MAT 11.8°C and an elevation ca. 3.1 km (Wolfe et al. 1998, with an inferred high elevation for the southern Rocky Mountains at that time), the more southern and arid-adapted affinity of its taxa is compatible with floral and topographic diversity having been achieved by the Uintan and Duchesnean.

In addition to containing the first sespedectine erinaceomorphs; soricids; lagomorphs; eomyine rodents; muroid rodents (in North America); leptocherine, agriochoerid, hypertragulid, and camelid artiodactyls; and hyracodontine, amynodontine, and metamynodontine rhinos, Uintan faunas are characterized by a diversity of miacoid carnivores, bunomerycine dichobunid, oromerycid tylopod, and protoceratid artiodactyls. Collectively these more selenodont forms (especially artiodactyls) apparently evolved in reflection of the transition from the previous megathermal tropical rainforests to an overall semideciduous tropical dry forest, including a mosaic of paratropical and warm temperate taxa and upland temperate forms (Graham 1999:195). As indicated at 5 in figures 8.1 and 8.5, an immigration pulse at about the Bridgerian–Uintan boundary included uintatheres, brontotheres, amynodontine and hyracodontid rhinos, and eomoropine chalicotherioids. Limited later dispersals include omomyine primates and eomoropines shared with Asia at Ui2, lagomorphs at Ui3.

Walsh (1996) summarizes the mammal faunas of the Uintan and Duchesnean of California, the Rocky Mountains, and Texas. He suggests that the two distal regions are about equally distinct relative to the Rocky Mountains region (ca. 60 percent Simpson coefficient) but even more distinct relative to each other (ca. 49 percent Simpson coefficient).<sup>1</sup> California faunas contain a number of holdover taxa from the Bridgerian (e.g., *Scenopagus*, *Pantolestes*, *Palaeictops*, *Pauromys*, and *Lophiohyus*), but they are not unique to California. *Antiacodon* (dichobunid artiodactyl), *Aethomylos* (“proteutherian”), and *Crypholestes* (erinaceimorph) are unique to California, which, with *Stockia* and *Washakius* (and with the possible absence of otherwise common early Uintan groups such as peraceratheriine, triplopodine, and hyracodontine rhinos, isectolophid tapiroids, and bunomerycine homacodontines), appears to reflect a western facies distinct from more interior regions, but the faunas west of the Rocky Mountains are poorly known. The synopsis of floral distributions in this chapter is consistent with Walsh’s (1996) finding that the later Uintan faunas of California differed (mainly at the species level; Lillegraven 1979) from those of the interior largely because of vegetational and climatic patterns. For the later Uintan, washakiine

omomyid primates apparently persisted (*Dyseolemur*), which, with the presence of ailuravine (*Eohaplomys*) and simimiyid (*Simimys*) rodents and the absence of otherwise key groups such as rabbits and hypsodontid condylarths (*Hyopsodus*), continues to suggest a western facies during that time.

Duchesnean faunas continue the Uintan renovations in that the sciuravid rodents, taeniodonts, oxyaenid and limnocyonine creodonts, viverravid carnivores, microsypid and notharctine primates, Dinocerata, helohyid and homacodontine artiodactyls, and hyrachine rhinos are no longer present. *Heliscomys* is the first geomyoid rodent, *Hyaenodon*, *Ischyrognathus*, and *Hemipsalodon* the first hyaenodontine creodonts, *Hesperocyon* the first canid, and *Heptacodon* the first anthracothere. Selenodont artiodactyls (e.g., agriochoerid and merycoidodontine oreodonts and hypertragulids) and other advanced groups persist, and later in the Duchesnean, taxa having affinities with Chadronian faunas occur as early elements of the White River Chronofauna. In chapter 4, faunal provincialism is highlighted for the West Coast, West Texas, Gulf Coastal Plain, Great Basin, and Rocky Mountains (based mostly on rodents; Storer 1989), and this seems to be a part of the regional endemism also recognized for the Uintan as based on other mammalian groups also (Walsh 1996). This provincialism apparently does not persist in Chadronian and younger elements of the chronofauna (Storer 1989).

As far as Duchesnean dispersals are concerned (figure 8.5), *Hyaenodon* shows affinity with Europe and *Pterodon* with Europe and Asia (McKenna and Bell 2000). Leptomerycidae refers to *Hendryomeryx* (chapter 4), with potential Asian affinities (*Archaeomeryx*; Webb 1998; Webb and Taylor 1980); Anthracotheriinae is a likely Asian immigrant (Kron and Manning 1998), of which *Heptacodon* is the oldest North American representative (also chapter 4).

In the late Eocene (Chadronian), the White River Chronofauna persisted. As indicated in chapter 4, Chadronian innovations include Sciuridae, Nimravidae, Tayassuidae, and Ursidae (Baskin and Tedford 1996). Limited immigration from Asia produced the bothriodontine anthracothere *Bothriodon* (Emry et al. 1987; Kron and Manning 1999) and the amphicyodontine ursid *Parictis* (Emry et al. 1987; Hunt 1996, 1998b). The major sea level lowstand at ca. 37 Ma (Ta4; figures 8.1, 8.6) was largely unremarked by the North American indigenous fauna (*Ardynomys* being another immigrant about this time), but a coeval early phase of climatic cooling may correspond with the faunal turnover that marked the beginning of the White River Chronofauna, in the

context of general increase in climatic aridity and concomitant more modern, open floral associations. The chrono-faunal turnover begins in the late Duchesnean and is manifested by archaic groups such as microsypid, adapid, and omomyine primates, ailuravine rodents, oxyaenid and mesonychid creodonts, hyopsodontid condylarths, uintatheres, and viverravid carnivores giving way to more modern elements: *Leptictis* (leptictid epithere), *Eutypomys*\* (eutypomyid castorimorph; \* = but also in the Lac Pelletier Lower Fauna of the Cypress Hills, Saskatchewan [figure 8.3], which is arguably older than late Duchesnean; Storer 1996, 1998), *Ischyromys* (ischyromyine rodent), *Pseudocylindrodon*\* (cylindrodontine rodent), *Adjidaumo*\*, *Yoderimys* (eomyid rodents), *Hyaenodon*\*\* (creodont, \*\* = also from early Duchesnean; chapter 4), *Daphoenus*\*\* (amphicyonid carnivore), *Toxotherium*\*\* (amynodontine rhino), *Hyracodon*\*\* (hyracodontine rhino), *Mesohippus* (miohippine equid), *Agriochcerus*\*\* (agriochcerid oreodont), *Poabromylus*,

*Heteromeryx* (protoceratid artiodactyls), *Eotylopus*\*\* (oromerycid artiodactyl), *Aclistomycter*\*\* (oreodontine oreodont), *Hendryomeryx*\* (leptomerycid artiodactyl), *Sinclairiella*\* (apatemyid rodent), *Hesperocyon*\* (canid), *Heptacodon*\* (anthracotheriine artiodactyl), and hypertragulids\*\* (Prothero and Emry 1996).

This long interval of faunal integrity began in the late Duchesnean, as seen in the Porvenir (figure 8.3) and related units (Emry et al. 1987; Prothero and Emry 1996), coincident with the dispersal of *Ardynomys* (figure 8.5) between North America and Asia (Emry and Korth 1996). A second limited Asian dispersal in the Chadronian (figure 8.6) is represented by *Patriomanis* (Gaudin and Emry 2002) and Nimravidae (Bryant 1996).

The present icehouse world is heralded by the opening of the Drake Passage and onset of the modern regime of Antarctic continental glaciation at the beginning of the Oligocene (figure 8.1). The circum-Antarctic current was established at about 34–36 Ma. In Australia, this is coeval

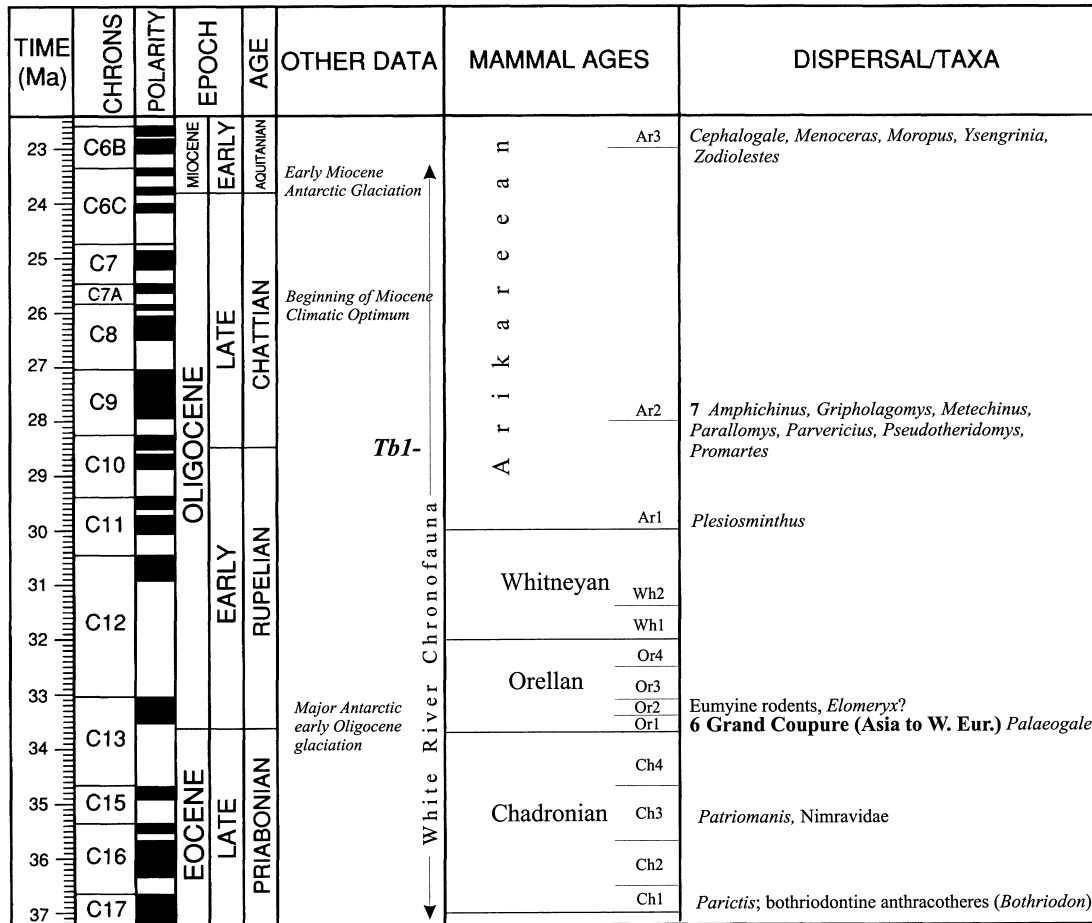


FIGURE 8.6 Oligocene time scale, mammal ages, and dispersal of mammalian taxa. Time, chrons, polarity, epochs, and ages after Berggren et al. (1995). Mammal ages and dispersal/taxa after chapter 5. Tb1 is time of major sea level fall. Other notes as for figure 8.4.

with the last occurrence of mesothermal–megathermal rainforests and the beginning of the dominance of *Nothofagus* as cooler climates prevail (MacPhail et al. 1994) and open forests are developed (Woodburne and Case 1996). In North America, the early Oligocene is a period of cooling and increased seasonal aridity (Graham 1999) with stable conditions reflected in the presence and persistence of the White River Chronofauna for contemporary mammals.

The Oligocene begins with the Priabonian–Rupelian boundary, at about 33.7 Ma, slightly below chron C13n and correlative with sequence boundary TA4.4 of Haq et al. (1988; Woodburne and Swisher 1995) and with oxygen isotope event Oi1 of Miller et al. (1991; Abreu et al. 1998:251), a major cooling event attributed to Antarctic glaciation. Global cooling continues during chron C13n (Brinkhuis 1994; sea surface data based on dinoflagellates, ca. 33.7 Ma; Hardenbol et al. 1998), with oxygen isotope event Oi2 (C11.1n, ca. 29.4 Ma; Abreu et al. 1998:252). In this context, sea level fluctuations from the late Eocene into the Oligocene appear to correspond to the Haq et al. (1988) intervals TA4.1–4.5, or from about 37.0 to 31.8 Ma (Hardenbol et al. 1998) as being glacially controlled. Abreu et al. (1998) continue this analysis further into the Miocene and Pliocene, but the main episode of sea level lowering concomitant with glaciation focused mainly on the Oligocene. Although oxygen isotope values fluctuate in the Miocene, consistent with glacially controlled climate and sea level, they regain the high levels of the late Oligocene only in the medial Miocene.

The modernization of paleofloras begun in the late Eocene proceeds in the Oligocene as far as ascertained from limited evidence. The late Oligocene Creede flora of southwestern Colorado (ca. 27 Ma; figure 8.3) is estimated to have lived under an MAT of ca. 4.5°C (compared with about 2°C at present) and consisted of a fir–spruce forest, a fir–pine forest, a pine–juniper forest or woodland, and a mountain mahogany chaparral community, with affinities to species in eastern and western North America and eastern Asia. Wolfe et al. (1998) indicate a paleoelevation of ca. 2.6 km for the Creede flora. Although the Creede flora is not considered to have a direct modern analog (Graham 1999), it shows that the modern montane deciduous forest continued to develop from middle Eocene forerunners in the Pacific Northwest (Washington, British Columbia) and Rocky Mountains (Green River). Prior tropical elements were absent by the Oligocene in interior North America.

The earlier Oligocene (ca. 32 Ma) Bridge Creek flora (and adjacent sites in central Oregon; same general locality as the middle Eocene Clarno flora, figure 8.3) is a tem-

perate broadleaved deciduous hardwood forest, indicating an MAT of 9–11°C, and reflects a cooler, drier, and more seasonal climate than inferred from the Clarno, possibly in part because of the early phase of Cascade Mountain elevation. Contemporaneous and later Oligocene floras of more western Oregon preserve the more humid warm temperate to temperate maritime conditions, with MATs ranging from 12–13°C (upland) to ca. 16°C (coastal). Farther northwest, cooler climates support temperate deciduous and coniferous forests (Graham 1999). The Oligocene mammalian record is biased toward that from the eastern flank of the Rocky Mountains (Montana, Wyoming) and adjacent plains (Dakotas, Nebraska), with an outlier in the Texas panhandle and another in Death Valley, California. The mammalian record thus does not yet reflect the geographic diversity seen in paleofloras of the time, so faunistics are couched mostly in temporal terms (Prothero and Heaton 1996; Janis et al. 1998a–1998c; Prothero 1999).

The mammalian record for the Orellan and Whitneyan is characterized by persistence of the White River Chronofauna, with assemblages dominated by hyaenodontids (*Hyaenodon*), rabbits (*Archaeolagus*, *Gripholagomys*, and *Megalagus*), rhinos (hyracodontine, *Hyracodon*; diceratheriine, *Amphicaenopus*, *Subhyracodon*, and *Diceratherium*; amynodontine, *Metamynodon*), and miohippine horses (*Miohippus* and *Mesohippus*). Oreodonts are represented by leptaucheniines (*Pseudocyclopius*, *Hadroleptauchenia*, *Leptauchenia*, and *Pithecestes*), oreodontines (*Aclistomycter*, *Merycoidodon*, *Otionohyus*, and *Paramerycoidodon*), miniochoerines (*Stenopschoerus* and *Miniochoerus*), desmatochoerines (*Prodesmatochoerus* and *Subdesmatochoerus*), and promerycochoerines (*Promesoreodon*). Camelids include the poebrodon-tines (*Poebrotherium*, *Paralabis*, and *Paratylopus*) and pseudolabines (*Pseudolabis*), with oromerycids represented by *Eotylopus*, *Malaquiferus*, and *Montanatylopus*. Ruminants are represented by leptotraguline (*Poabromylus* and *Heteromeryx*) and protoceratine (*Protoceras*) protoceratids, the hypertragulids (*Hypertragulus*, *Nanotragulus*, and *Hypisodus*), and leptomerycids (*Leptomeryx*; Lucas 1992). Canids include the hesperocyonines (*Prohesperocyon*, *Hesperocyon*, *Parenthydrocyon*, *Mesocyon*, and *Cynodesmus*), the borophagine *Otarocyon*, the daphoenine amphicyonids (*Daphoenus*), and nimravids (*Dinictis* and *Nimravus*). A diverse group of rodents includes the ischyromyids (*Ischyromys* and *Titanotheriomys*), the allomyines (*Prosciurus* and *Pelycomys*), the sciurine *Prosciurus*, the eutypomyoid *Eutypomys*, an eumyid (*Eumys*), the eomyids *Paradjidaumo* and *Eomys*, the eomyoids *Heliscomys*, *Tenudomys*, and *Proheteromys*, and

the sciuravid *Pipestoneomys*. Limited mammal dispersal during the Oligocene is suggested by eumyine rodents in Or2 (*Eumys elegans*) and *Elomeryx* (Fossil Bush Local Fauna, possibly about this age; Storer 1996; Kron and Manning 1999). The carnivore *Palaeogale* (Baskin and Tedford 1996) is known from the Chadronian of North America but apparently dispersed to Eurasia in the Orellan (6, figure 8.5, which otherwise signals the time of the Eurasian Grand Coupure).

In chapter 6, a general similarity is noted for the late Oligocene (early Arikareean) mammal faunas from the Great Plains (and Montana) to the southwest (Great Basin), in contrast to those from the John Day region of Oregon. Among these differences is a lack of leptauchenine oreodonts and protoceratids and abundance of *Entoptychus* species in Oregon, along with a diversity of aplodontid rodents, a characteristic group of small hypercarnivores, the presence of gentilicamelinae and paratylopine camels, and persistence of White River Chronofaunal relicts. The causes of this regional endemism are unclear. The John Day region was part of a major volcanic province that, along with the rising continental divide to the east, may have provided background for these faunal differences, as could the rising Cascade Mountains evidenced by the Rujada flora (below) and relatively arid conditions indicated by the Bridge Creek flora and John Day paleosols (above). The early Arikareean faunas of the Albuquerque area, New Mexico, and the Big Bend area, Texas, also illustrate endemism relative to those of the Great Plains, as do those of the Texas Gulf Coastal Plain, which flourished under subtropical to tropical conditions that contrast with the drier upland environments envisioned for the Great Plains (chapter 6).

Faunas from the early part of the Arikareean (figure 8.6) generally include the last elements of the White River Chronofauna. Members of persisting but dwindling to outgoing lineages include cylindrodontids, heterosoricine soricids (e.g., *Domnina*), hesperocyonine canids (Munthe 1998), daphoenine amphicyonids (Hunt 1998a), entelodonts (Effinger 1998), anthracotheres (Kron and Manning 1998), miniochoerine oreodonts (Lander 1998), hypertragulids and leptomerycids (Webb 1998), brachydont horses, early tapirids (Colbert and Schoch 1998), hyracodontid rhinos (Prothero 1998a), and diceratheriine rhinos (Prothero 1998c). Chapter 6 identifies *Geolabis*, *Centetodon*, *Domnina*, *Proscalops*, *Protosciurus*, *Eutypomys*, *Agnotocastor*, *Palaeocastor*, *Scottimus*, *Arikareomys*, *Heliscomys*, *Palaeolagus*, *Megalagus Leptomeryx*, *Elomeryx*, and *Diceratherium annectens* as among the White River relicts.

Early Arikareean faunas also record the first allomyid, mylagaulid, marmotine, and entoptychine rodents, as

well as canine (*Leptocyon*; Munthe 1998) and procyonid carnivores, characteristic of the Runningwater Chronofauna that begins in the late Arikareean (Webb and Opdyke 1995). Also during this interval an expansion of groups important to the Runningwater Chronofauna include beavers (Xu 1996), borophagine canids (Wang et al. 1999), mustelids (Baskin 1998), synthetocerine protoceratids (Prothero 1998b), and stenomyline camels (Honey 1998). Ar2 witnessed the major immigration of the Arikareean (7, figure 8.6), represented by erinaceids (*Amphechinus* and *Parvericius*), a brachyericine (*Metechinus*), an ochotonid (*Gripholagomys*), an aplodontid (*Parallomys*), an eomyid (*Pseudotheridomys*), and the mustelid *Promartes*. As indicated in chapter 6, faunas of the earlier part of the Arikareean were undergoing an incompletely documented revolution that probably was under way by about 24 Ma. By the time of the Runningwater Chronofauna of Janis et al. (1998a–1998c), in the later Arikareean, almost all remnants of the White River Chronofauna had been replaced.

## NEOGENE

Shackleton et al. (2000) suggest that the age of the Oligocene–Miocene boundary is revised to  $22.1 \pm 0.1$  Ma, 0.9 Ma younger than the usually accepted date of  $23.8 \pm 1$  Ma. For present purposes, the Miocene (23.8–5.3 Ma) includes the early occurrence of major phosphogenic episodes (21–20, 19–18, and 17–16 Ma) on the southeastern coast of the United States, apparently in response to changes in sea level, upwelling intensity, northward-aging Atlantic deep water, and high phosphorus burial rates possibly reflecting rapid uplift and erosion of the Himalayan–Tibetan plateau between 21 and 15 Ma (Raymo 1994; Compton and Mallinson 1996; figure 8.1 herein) and delivery of phosphorus to the North Atlantic ocean via deepwater formation in the northern Indian Ocean (Woodruff and Savin 1989), according to Mallinson and Compton (1997).

As indicated in figures 8.1 and 8.6, a period of climatic warming began in the late Oligocene at about 25 Ma and culminated in the medial Miocene climatic optimum at about 15 Ma. This interval generally witnessed a continuation of the warm temperate to subtropical conditions of the Oligocene. Erosion of the Rocky Mountains that began in the Oligocene continued to shed debris to the east in support of a diversity of stream border communities, open interfluves, and savanna–parkland environments (Hunt 1990).

Sparse early Miocene floras record a northern and Arctic element of mixed hardwood and coniferous forests with an MAT of 11–14°, giving way in the northeastern United States (Vermont; Brandon flora, ca. 24 Ma) to a warm temperate to subtropical climate with an MAT of about 17°C (frost free). Farther south, in the Great Plains, limited evidence suggests the presence and persistence of a shrubland–savanna community including the first megafossil evidence of grasses. For the Rocky Mountains, higher elevations resulted in the persistence of a western montane forest, with streamside components of broadleaved deciduous angiosperms living under conditions that fluctuated from dry to wet in the late early Miocene, from wet to dry in the early middle Miocene, and dry to wet in the late Miocene–Pliocene (Graham 1999), apparently synchronous with the oceanic oxygen isotope record for that interval (figure 8.1). The warm temperate conditions interpreted for the west coastal Rujada flora (Oregon; figure 8.3) reflect its maritime location, but the presence of fir trees in the assemblage suggests derivation from the possibly rising Cascade Ranges to the east, contributing a rain shadow effect (Graham 1999) represented by the more arid conditions inferred from the paleosols of the Clarno–John Day region (figure 8.3), conditions that plausibly persisted into the time of the Arikareean John Day faunas as well (Bestland et al. 1997; Bestland 2002).

The initiation of extensional faulting in the Basin and Range province about 17 Ma resulted in its elevation via isostatic processes (Wolfe et al. 1997). Floras from this region (Buffalo Canyon, ca. 15.6 Ma [figure 8.3]; Eastgate, ca. 15.5 Ma; Middlegate, ca. 15.5 Ma) are considered to have lived at elevations ranging from 2.8 to 3.2 km, under an MAT of ca. 10°C, made up of a mixed conifer–deciduous hardwood forest, with drier, south-facing slopes supporting a chaparral vegetation. The more arid, subtropical influence also is recorded by the Tehachapi flora (figure 8.3; ca. 17 Ma) to the southwest, in California, even at lower elevations. The Sierra Nevada still was low at this time as well, as reflected by the ca. 21 Ma Sutro flora (mixed mesophytic broadleaved evergreen and conifer) in westernmost Nevada (figure 8.3; Graham 1999). By the end of the early Miocene, the previous tropical dry forest and notophyllous broadleaved evergreen vegetation of the middle and late Eocene had been replaced by a southern coastal tropical community, a southern deciduous forest having a pine woods and southern mixed hardwood floodplain component; an Appalachian montane coniferous forest; a shrubland–chaparral woodland, savanna, and mixed hardwood–conifer forest in the interior; and a montane coniferous forest in the northwest (Graham 1999).

As indicated in chapter 6, faunas of Hemingfordian age generally show reasonable similarity across North America, even though those of the southern Great Basin and adjacent coastal California tend to be composed of fewer taxa than Great Plains contemporaries. This includes the sharp reorganization of the Miocene Chronofauna in the second part of the Hemingfordian. The new chronofauna persists to about the Hemphillian, but its Barstovian segment shows a sharp separation of Great Basin faunas relative to those of the Great Plains (and even in Montana on the west and the Florida Gulf and East coasts). The great diversification of midcontinent lineages such as mylagauline rodents, dromomerycids, cervids, and antilocaprids, the radiation of equine horses, and other features documented in chapter 6 only marginally affect Great Basin faunas of the late Hemingfordian through Barstovian, in contrast to the Great Plains affinities seen the faunal elements of the early Clarendonian Dove Spring Formation of California. These observations may reflect the elevation of the Great Basin at about 17 Ma and its collapse by about 13 Ma as discussed by Wolfe et al. (1997).

The Runningwater Chronofauna (from late Arikareean [Ar3] through early the Hemingfordian [He1]) reflects the opening floral landscape characterized as sustaining riparian forests, woodland, park savanna by Webb et al. (1995) and Webb and Opdyke (1995). This chronofauna is characterized by derived anchitherines (*Archaeohippus*, *Desmatippus*, and *Parahippus*) and diceratheriine, aceratheriine, and teleoceratine rhinos (immigrants are *Menoceras* [Ar3], *Floridaceras* [He1], and *Brachypotherium* [He1]). Paleomustelids immigrate (*Zodiolestes*; Ar3) and diversify, specialized hesperocyonine canids persist, and borophagines diversify. Amphicyonine amphicyonids undergo a minor radiation, aided by immigrants *Ysengrinia* (Ar3) and *Cynelos* (Ar4). The expanding ungulate component is represented by new and diverse camelids (protolabines, miolabines, and floridatragulines), which show their greatest breadth in the Barstovian and diminish subsequently (Webb et al. 1995). Leptauchenine oreodonts persist, and ticholeptines are new. Synthetocerine protoceratids first appear. Blastomerycines (*Problastomeryx*, Ar4) and aletomerycine (*Aletomeryx*, He1) and cranioceratine dromomerycids (*Barbouromeryx*, Ar3) immigrate and diversify; merycodontine antilocaprids appear in the late Arikareean (*Paracosoryx*, Ar4) and early Hemingfordian, with the dromomerycids achieving greatest diversity in the early Barstovian, the merycodontines in the late Barstovian. Dwindling groups include entelodonts, anthracotheres, and hypertragulids, and leptauchenine, merycochoerine, and phenacocoeline oreodonts do not



survive, or barely survive, the Arikareean (Janis et al. 1998a–1998c).

Minor immigrant pulses at Ar3 and Ar4 in the late Arikareean (figure 8.7) bracket a strong (and apparently ineffectual) sea level lowstand (Tb2) and predate a major pulse at the beginning of the Hemingfordian (8, figure 8.7). As indicated in chapter 6, this contingent of immigrants is dominated by mustelids (*Potamotherium*, *Craterogale*, *Leptarctus*, and *Miomustela*) but includes a soricid (*Antesorex*), plesiosoricid (*Plesiosorex*), rabbits (*Desmatolagus* and *Oreolagus*), a castorine beaver (*Euroxenomys*), procyonids (*Amphictis* and *Edaphocyon*), an amphicyonid (*Amphicyon*), a hemicyonine (*Phoberocyon*), an ursid (*Ursavus*), aceratherine (*Floridaceras*) and teleoceratine (*Brachypotherium*) rhinos, and aletomerycine artiodactyl (*Aletomeryx*) and merycodont antilocaprids (Janis and Manning 1998), suggesting a variety of habitats ranging from streamside or pond through more open country settings consistent with the vegetational changes summarized earlier.

The second Hemingfordian immigration episode is contemporaneous with the beginning of the mid-Miocene climatic optimum (Flower and Kennett 1995),

recorded between 17 and 15 Ma (figure 8.1); it includes the time of major radiation of North American mesodont to hypsodont equids (Hulbert and MacFadden 1991) and presumed spread of grasslands (Wang et al. 1994), and it heralds the beginning of the Clarendonian Chronofauna as used by Janis et al. (1998a–1998c), which persisted until the Hemphillian at ca. 9 Ma. After about 15 Ma, climate began to deteriorate, as reflected in floral and faunal change.

The 17–15-Ma warm interval is reflected in the eastern coastal area having a rich, warm temperate mixed mesophytic forest, with numerous conifers. The woodland–grassland vegetation of the Great Plains graded westward into a western montane coniferous forest and mixed mesophytic deciduous forest. The Pyramid flora of Nevada (15.6 Ma; figure 8.3) estimated to have lived at 2.8 km elevation and composed of a deciduous hardwood forest and floodplain association, with an MAT of 13°C (or less) and annual precipitation about 35–40 inches. The approximately coeval Mascall flora of Oregon (same site as John Day, figure 8.3) reflects a swamp cypress–deciduous forest in the lowlands and a hardwood–conifer forest in the upper slopes and probably was sited at lower eleva-

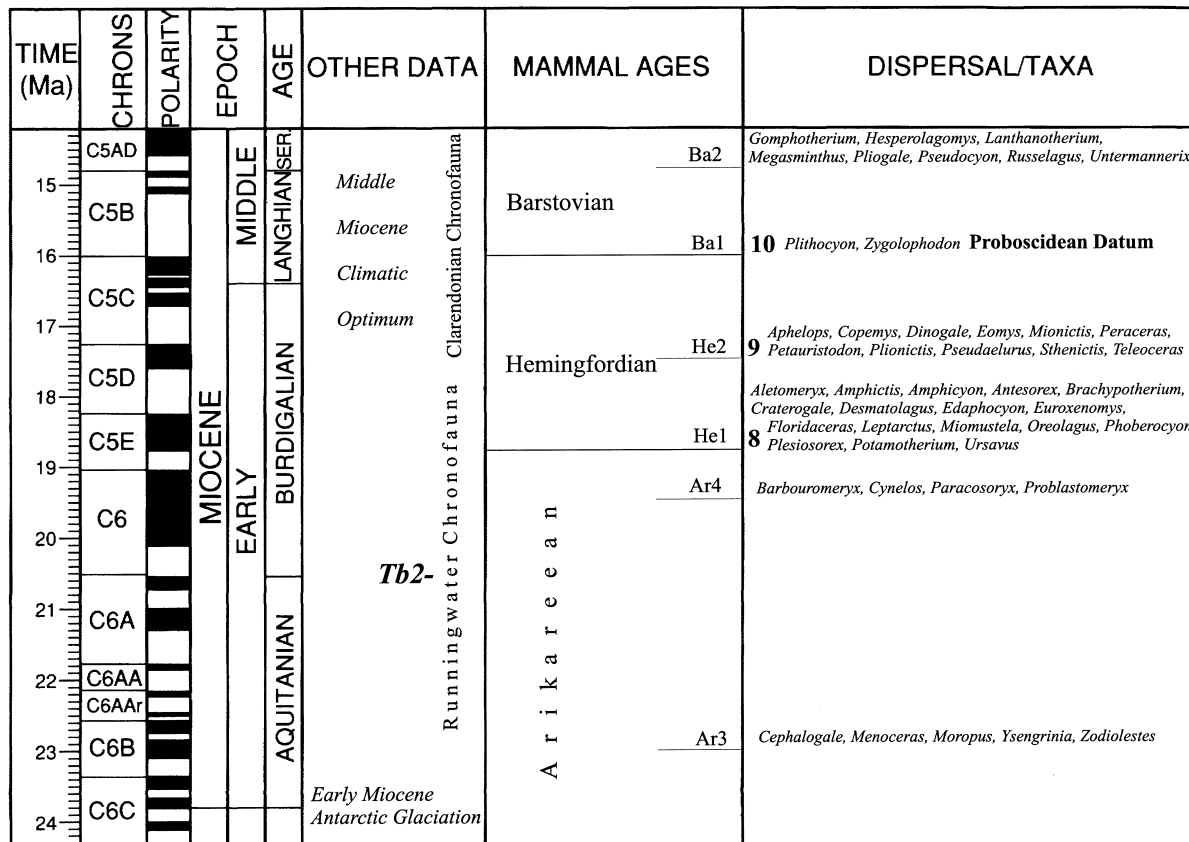


FIGURE 8.7 Early Miocene time scale, mammal ages, and dispersal of mammalian taxa. Time, chrons, polarity, epochs, and ages after Berggren et al. (1995). Mammal ages and dispersal/taxa after chapter 6. Tb2 is time of major sea level fall.

tion than the Pyramid. The *Clarkia* flora of Idaho (ca. 16 Ma; figure 8.3) shows a bottomland, swamp, and riparian forest community and a drier slope forest. These and other floras of comparable age in Nevada, Oregon, and Washington typically show abundant proportions of exotic deciduous hardwoods<sup>2</sup> that most resemble elements from modern eastern North America and Asia, whereas floras only about 1 m.y. younger either lack these exotic elements (e.g., Stewart Valley, Nevada, ca. 14.5 Ma; figure 8.3) or possess them in reduced numbers and apparently reflect the effects of decline in global temperatures and at least local rainfall that began at about 15 Ma (Graham 1999). Especially in the southern part of western North America, floras younger than 15 Ma apparently lived under regimes with less than 3.5 inches of rainfall in the summer.

The Clarendonian Chronofauna (Janis et al. 1998a–1998c) saw a diversification of the borophagines (Wang et al. 1999), neomustelids, and procyonines. Amphicyonids and hemicyonine ursids declined. Among the ungulates, many lineages were extinct, including anthracotheriids, entelodontids, most merycoidodontid oreodonts (except ticholeptines, which declined in the interval), hypertragulids, and most leptomerycids. Alomyine and promylagauline rodents, stenomyline camels, and all dromomerycini became extinct during the interval. First appearances include scalopin moles, dipodomysine rodents, castorini beavers (*Hystricops*), *Aepycamelus* (Lamini), *Procamelus* (Camelini), *Ramoceros*, *Plioceros* (Antilocapridae), tayassuines, and proboscideans. Equines horses begin a major radiation, composed of merychippines (*Merychippus*), hipparionines (*Cormohipparion*, *Hipparion*, *Neohipparion*, *Pseudhipparion*, and *Nannippus*), protohippines (*Protohippus* and *Calippus*), and pliohippines (*Parapliohippus*, *Acritohippus*, *Heteropliohippus*, and *Pliohippus* s.s.; Kelly 1995, 1998; Hulbert and MacFadden 1991; MacFadden 1998). Aceratherine and teloceratine rhinos persist.

The later Hemingfordian immigrant episode (9, figure 8.7) is dominated by mustelids (*Dinogale*, *Plionictis*, *Sthenictis*, and *Mionictis*) but also includes cricetid (*Copemys*), eomyid (*Eomys*), and sciurid (*Petauristodon*) rodents, a true felid (*Pseudaelurus*), and aceratherine (*Aphelops* and *Peraceras*) and teloceratine (*Teleoceras*) rhinos.

Climatic cooling at about 15 Ma may be linked to the modernization of global oceanic circulation. An evolutionary turnover of Pacific benthic foraminifera from about 17 to 14 Ma (Flower and Kennett 1995) is considered to reflect circulation changes of the deep oceans caused by a major expansion of the East Antarctic Ice Sheet and the spread of cooler deep ocean waters, and

Flower and Kennett (1993) identify 15 Ma as the approximate date of the beginning of modern oceanic circulation, temperature conditions, and influence of the cryosphere.

15 Ma also is the time when the vegetation and climate record of northwestern Canada and Alaska suggest a decline in temperature from a continental global warm peak (White et al. 1997). The early and middle Miocene Seldovian floral stage and correlative intervals in Japan and Siberia (Volkova et al. 1986; Itoigawa and Yamanoi 1990) showed the warmest climate of the last 24 m.y. This flora was rich in tree genera now living in temperate climates of Asia and North America (Wolfe 1966; Wolfe and Tanai 1980; Leopold and Liu 1994; White and Ager 1994). The temperate flora extended northward at 15.2 Ma (White and Ager 1994), but subsequent cooling reduced high-latitude floral diversity (Wolfe et al. 1966), positively correlated with a decrease in available energy (Currie and Paquin 1987).

European evidence of climatic cooling over the continent at about 14 Ma is based on terrestrial mammal records in Spain, correlated to the global increase in  $\delta^{18}\text{O}$  (Krijgsman et al. 1996). This is based on a change in rodent faunas, which show an increase in diversity of immigrant, cool-adapted northeastern taxa; this also results in a peak of new occurrences (Daams et al. 1999).

Hemicyonine bears (*Plithocyon*) and proboscideans are important immigrants at the beginning of the Barstovian (10, figure 8.7), with a former Barstovian indicator (*Copemys*) being recognized in the second Hemingfordian pulse (chapter 6). The Proboscidean datum of the early Barstovian apparently represents mammutids, followed by the later appearance of gomphotheriids in the later Barstovian, especially in the Great Plains and adjacent areas. In the later Barstovian (figure 8.7), other immigrants include the ochotonid rabbits *Hesperolagomys* and *Russellagus*, the erinaceids *Lanthanotherium* and *Untermannerix*, the zapodid rodent *Megasminthus*, the mustelid *Pliogale*, and the amphicyonine *Pseudocyon*.

By about 13 Ma, at about the midpoint of the duration of the Clarendonian Chronofauna, the trend toward colder and winter-dry climates resulted in a mosaic of open deciduous forest–woodland and grassland patches in the Great Plains region (Kilgore, Nebraska; figure 8.3), and intervening highlands tended to separate Great Plains floras (with affinities mainly to the east and south) from those of the Columbia Plateau. The latter tended to support a rich broadleaved deciduous forest at low to mid-elevations to montane woodlands, with summer-wet conditions in contrast to their southern (summer-dry) contemporaries. To the southwest of the Rocky Moun-

tains vegetation was largely piñon pine–juniper woodland and shrubland with evergreen, oak, a depauperate western montane coniferous forest in the higher elevations, and woodland–chaparral associations in southern exposures and drier situations.

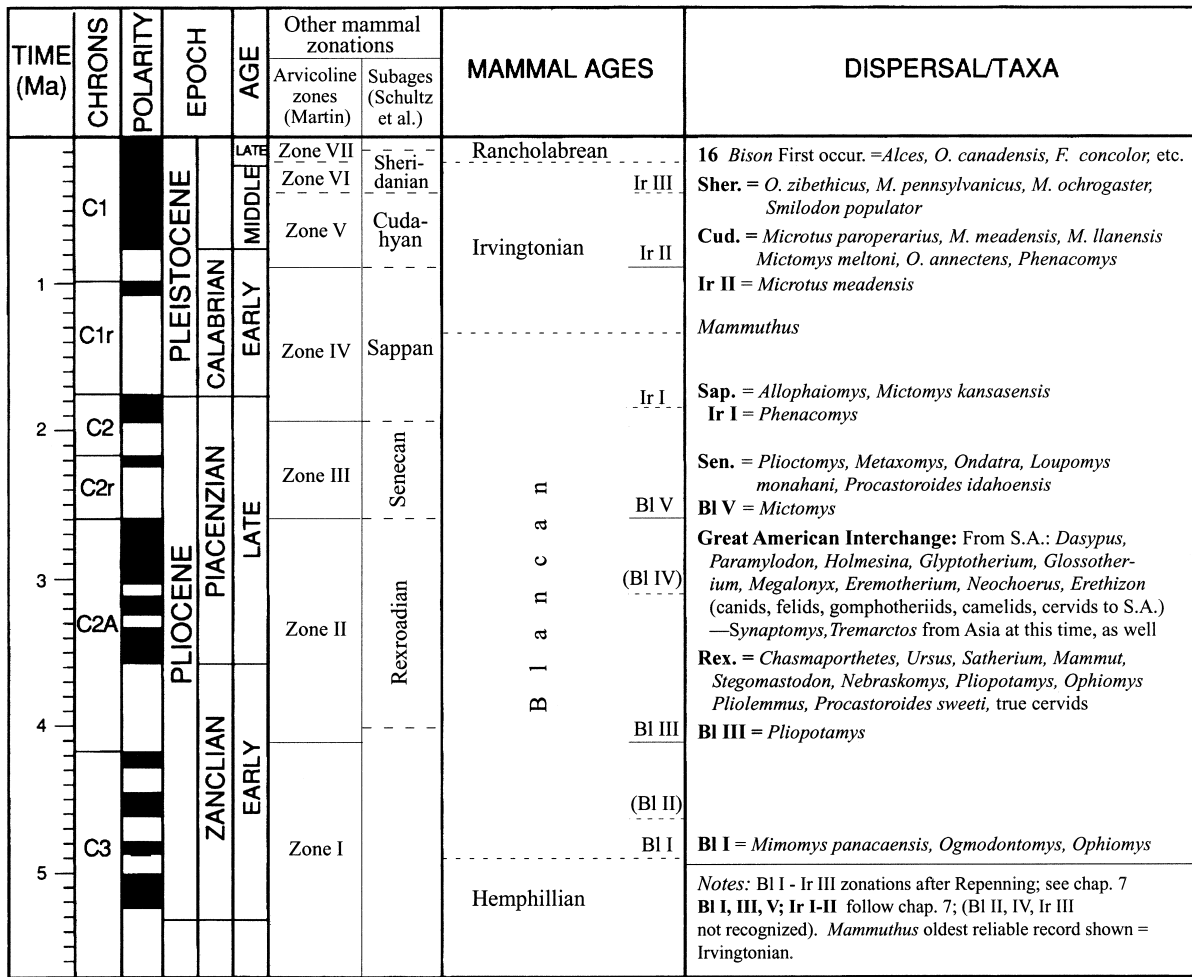
Limited immigrations define the subdivisions of the Clarendonian (figure 8.8), with the exit of *Cormohipparion* to the Old World (12, figure 8.8) forming the basis of the *Hippotherium* datum at the base of the Vallesian mammal age in that region (Woodburne 1996). Discussions in chapter 6 illustrate the faunal provincialism that besets the correlation of Arikareean through Barstovian mammal faunas from the Great Plains region to the Great Basin and Mojave Desert and the extension of key elements of the (mostly equid) fauna to the western province at the beginning of the Clarendonian. As summarized in Janis et al. (1998a–1998c), the Gulf Coastal region commonly held a distinct fauna (*Merychyus* the sole oreodont, *Prosynthetoceras* and *Floridatragulus* distinctive, and amphicyonids, canids, and antilocaprids largely absent).

The Miocene–Pliocene Chronofauna (Janis et al. 1998a–1998c) begins with the Hemphillian at ca. 9 Ma and continues through the Blancan (ca. 1.3 Ma; figure 8.9). The interval is distinctive in recording (13–14, figure 8.8) an increased communication with South America before and after the establishment of the Panamanian land bridge at ca. 3.5 Ma (Keigwin 1982). Haug et al. (2001) evaluate restricted surface water exchange between the tropical Atlantic and Pacific oceans to indicate that shoaling occurred in the Central American seaway as a result of Panamanian isthmus evolution by about 4.7 Ma. This is compatible with the increased influx of Neotropical taxa to North America during the Blancan and later, but the mammal record also shows periodic immigration of edentates in the Hemphillian (e.g., figure 8.8, with 13 and 14 generally embracing the edentate and other [boreal] dispersals from 9 to ca. 7 Ma).

The interval also witnesses at about 8–6 Ma a global increase in biomass of plants using C<sub>4</sub> photosynthesis as indicated by changes in carbon isotope ratios of fossil her-

TIME (Ma)	CHRONS	POLARITY	EPOCH	AGE		MAMMAL AGES	DISPERSAL/TAXA		
5	C3	Normal	PLIOCENE EARLY	ZANCLIAN	C <sub>4</sub> grasses spread	Blancan	<i>Eocoileus, Lutra, Megantereon, Miopetaurista, Mustela, Ogmodontomys, Protopliophenacomys, Sminthosinus, Trigonictis</i>		
6	C3A	Normal	MIOCENE LATE	MESSINIAN		Hemphillian	Hh4	14 <i>Agriotherium, Castor, Felis, Megalonyx, Ochotona, Plesiogulo, Pliozapus, Prosomys</i>	
7	C3B	Normal		TORTONIAN			Hh3		<i>Enhydritherium, Eomellivora, Indarctos, Lutravus, Machairodus, Neotragoceras, Simocyon, Thinobadistes</i>
8	C3Br	Normal					Hh2		
9	C4	Normal	MIOCENE LATE	TORTONIAN	Hemphillian	Hh1	<i>Crusafontina, Kansasimys, Lemoynea, Pliometanastes, 13 Paramicroscoptes, Pliotomodon</i>		
10	C4r	Normal				C13	<i>Actiocyon, Hoplictis, Platybelodon</i>		
11	C4A	Normal	MIOCENE MIDDLE	SERRAVALLIAN	Clarendonian	C12	12 <i>Cormohipparion Datum to Eurasia</i>		
12	C4Ar	Normal				C11	<i>Barbourofelis</i>		
13	C5	Normal	MIOCENE MIDDLE	SERRAVALLIAN	Clarendonian	C11	<i>Pseudoceras</i> ; Plains taxa invade Great Basin		
14	C5r	Normal				Barstovian	Ba2	<i>Gomphotherium, Hesperolagomys, Lanthanotherium, Megasminthus, Pliogale, Pseudocyon, Russellagus, Untermannerix</i>	
15	C5A	Normal							
	C5AA	Normal							
	C5AB	Normal							
	C5AC	Normal							
	C5AD	Normal							
	C5B	Normal							

FIGURE 8.8 Middle–Late Miocene time scale, mammal ages, and dispersal of mammalian taxa. Time, chrons, polarity, epochs, and ages after Berggren et al. (1995). Mammal ages and dispersal/taxa after chapter 6. Tb3 is time of major sea level fall.



**FIGURE 8.9** Pliocene–Pleistocene time scale, mammal ages, and dispersal of mammalian taxa. Time, chrons, polarity, epochs, and ages after Berggren et al. (1995). Mammal ages and dispersal/taxa after chapter 7. Cud., Cudahyan; GAI, Great American Interchange; Rex., Rexroadian; S.A., South America; Sap., Sappan; Sen., Senecan; Sher., Sheridanian.

bivore tooth enamel in Asia, Africa, and North and South America (MacFadden et al. 1994, 1996; Quade et al. 1989; Wang et al. 1994; Ojha et al. 2000). This apparently reflects a change in herbivore diet to C<sub>4</sub> grasses, rather than shrubs and leaves, concurrent with a mid-Hemphillian mass extinction of grazing equids and antilocaprids, browsing camelids, and gomphotheres (Webb et al. 1995). The vegetation change may be related to a decrease in atmospheric CO<sub>2</sub> below the threshold favored by plants using the C<sub>3</sub> photosynthetic pathway and concurrent increased seasonality in rainfall and lower temperatures. Raymo (1994) and Raymo and Ruddiman (1992) link atmospheric lowering of atmospheric CO<sub>2</sub> to increased rates of erosion since about 40 Ma, focused on the active Himalayan–Tibetan region. Garzzone et al. (2000) suggest that the Tibetan Plateau achieved its current elevation as early as ca. 7 Ma and the beginning of the change

to steppe conditions in Northern Hemisphere (Webb et al 1995; Webb and Opdyke 1995). The photosynthetic change is about coeval with a remarkable warming of Pacific Ocean marine climate at about 7.5–6.0 Ma (Climatic Optimum 3; Barron and Baldauf 1990; Ozawa and Tomida 1996) and was followed by the terminal Miocene bioevent in Pacific Ocean (Beu 1973), where 23 molluscan genera in New Zealand become extinct at about 5.5 Ma, probably caused by intensification of oceanic circulation around Antarctica.

As summarized by Graham (1999), overall climatic conditions for North America show increased aridity and a decrease in temperature during the final 10 m.y. of the Cenozoic Period, with a warm pulse at about 3 Ma. Elevation changes leading to the modern topography for the Coast Ranges, Sierra Nevada, Great Basin, Rocky Mountains, and Great Plains transpired during

this interval. Modern deserts appear at about 5 Ma, northern boreal forests at about 4 Ma, tundra and permafrost at about 2 Ma.

The Great Plains floras show increasing influence of grasses and prairie herbs in the interval beginning about 10 Ma, with a sparsely wooded grassland reconstructed for the Oklahoma Panhandle area at ca. 6 Ma, with about coeval sites in Nebraska (Ash Hollow) preserving remains of sedges, grasses, and borages (forget-me-not), including hackberry. Farther north (Jackson Hole, Wyoming) the Teewinot Lake flora (ca. 8 Ma; figure 8.3) is a semi-arid shrubland, with riparian elements along streams, giving way to a western montane coniferous forest in higher elevations. Floras in the Great Basin and Idaho–Washington also record the late Miocene through Pliocene change from humid and summer rain conditions to those of a more semiarid dry summer regime, as does the Mt. Eden flora (ca. 5 Ma; figure 8.3) in nearly coastal California, where semiarid conditions prevailed, with rainfall at about 12–18 inches annually (Graham 1999).

In the Columbia Plateau, the late Miocene floras flourished under warm temperate (summer rain) conditions. After the Hemphillian, progressively arid (dry summer) conditions developed on the leeward side of the rising (Reiners et al. 2002) Cascade Range and Sierra Nevada, with sparse pine and conifer forests having only rare hardwoods growing in the Snake River Plain at ca. 3 Ma. Vegetation about this old in northwestern Oregon (Oak Grove; figure 8.3), in northern California (Tulelake; figure 8.3), and in northern Utah suggest temperatures 2–4°C warmer than at present, but more arid conditions, with the Utah sites suggesting a more continental climate. The Palousie Prairie (Idaho, eastern Washington, eastern Oregon) records the occurrence of grasslands here about 10 m.y. later than in the Great Plains. In coastal locations of northern California, redwood forests were maintained, with rainfall estimated at 40 inches annually, with a greater summer component and an MAT ca. 3°C warmer than now at 3–4 Ma, whereas somewhat more interior sites record a pine–Douglas fir forest near sea level with an oak–woodland on the slopes.

Janis et al. (1998a–1998c) indicate that the Miocene–Pliocene Chronofauna began when climatic cooling relative to the medial Miocene thermal maximum was well advanced. The opening of the landscape apparently was amenable to the introduction and success of a variety of bone-cracking carnivores, an increase in canid and feloid diversity, and an acme of mustelid carnivores. A dramatic decrease in the ungulate component occurred in this interval, apparently coincident with the regional effects of increased aridity as summarized earlier in this chapter.

The interval saw the demise of protoceratids and oreodonts and a reduction of camelids (to camelines), with specialized diversity in a number of lineages (*Alforjas* and *Megacamelus* in the Hemphillian, *Titanotylopus* and *Gigantocamelus* in the Blancan) and the persistence of *Camelops*. Previously diverse antilocaprids dwindled to the extent that merycodontines were extinct and dromomeryids almost extinct by the Pliocene. However, cervids appeared in the early Blancan and persisted. Whereas browsing equids had been extirpated by the mid-Hemphillian mass extinction, browsing rhinos persisted (*Aphelops* and *Teleoceras*) at least through the age. In contrast to the survival and persistence (*Astrohippus*) of the ultimate monodactyl lineage leading to *Equus*, formerly typical grazing horses became extinct (*Protohippus* and *Hipparion*) or limited to a Gulf Coast refugium (*Cormohipparion*, *Neohipparion*, *Calippus*, and *Pseudhipparion*; Webb et al. 1995; MacFadden et al. 1999). Taxa in which the development of the preorbital fossa (presumptively housing a diverticulum of social significance) had been an important attribute (e.g., *Cormohipparion*) lost this feature, as also occurred independently at about the same time by various hipparionine groups in the Old World (Woodburne 1989), apparently as a result of increased hypsodonty (Webb et al. 1995). Continuing aridity apparently contributed to the demise of the shovel-tusked proboscideans by at least the Blancan, which witnessed a minor radiation of beak-jawed taxa (*Rhynchotherium* and derivatives, *Stegomastodon*, *Cuvieronius*), which, along with their mammutid cousin (*Mammut*), survived into the Pleistocene.

Edentates from South America form a major new aspect to North American faunas of Hemphillian and later times, with *Pliometanastes*, *Thinobadistes*, and *Megalonyx* known from Florida in the Hemphillian (chapter 7; figure 8.8). Each of these participates in the definition of subdivision of the age: *Pliometanastes* appears in the North American record at ca. 9 Ma, at the beginning of the Hemphillian, *Thinobadistes* at the medial Hemphillian (Hh2, ca. 7.3 Ma), and *Megalonyx* in the later Hemphillian (Hh3, ca. 6.8 Ma; figure 8.8). For Hh1, *Pliometanastes* is joined by *Crusafontina* (soricid), *Lemoynia* (desmanine mole), *Kansasimys* (eomyid), and the murids *Paramicrotoscopes* and *Pliotomodon*. In Hh2, immigrants coeval with *Thinobadistes* include a variety of carnivores (*Indarctos*, *Simocyon*, *Lutravus*, *Enhydritherium*, *Eomellivora*, and *Machairodus*) and the bovid ungulate *Neotragocerus*. For Hh3, immigrants associated with *Megalonyx* are a pika (*Ochotona*), a murid (*Prosomys*), a zapodid (*Pliozapus*), a beaver (*Castor*), and a variety of carnivores (*Agriotherium*, *Plesiogulo*, and *Felis*).

Hh4 immigrants are wholly Holarctic and, except for the flying squirrel *Miopetaurista*, the arvicolines *Protopliophenacomys* and *Ogmodontomys*, and the ungulate *Eocoileus*, continue to favor carnivores (*Mustela*, *Trigonic-tis*, *Smithosinis*, *Lutra*, and *Megantereon*; chapter 7).

Except for the latter part of Hh4, the Blancan is almost the sole representative of the Pliocene in North America and, as indicated in chapter 7, the interval that continues into the Pleistocene and modern times witnessed episodic and progressive climatic deterioration. These changes were largely of global scale but also were locally driven by tectonism that eventually resulted in the present diversity of landscape and climate setting. The interval was one of frequent immigration of new taxa, apparently mostly from the American polar regions with contribution from and also influence on Holarctic Asia. In fact, it is clear that a coherent discussion of mammal age relationships is enhanced by setting aside, for now, a treatment that includes the Boreal region. Discussion in chapter 7 illuminates the presence of zoogeographic provinces, with focus on those on either side of the Rocky Mountains and those of the Gulf Coastal region. The use of mammal evolution and dispersal in correlation may be more problematic regionally across North America in the Pliocene and Pleistocene than for any other part of the mammalian record, and chapter 7 demonstrates the positive role played by applying independent age information to the mammalian stratigraphic and biochronologic framework. The importance of assessing the relationship of faunal data to independent chronologic systems is a theme on which much of this book is based.

Chapter 7 demonstrates the presence of at least six distinct episodes of immigration from the Blancan through Rancholabrean, most of which are documented by the record of arvicoline rodents. At the same time, the Neotropical influence that began in the Hemphillian culminates in the Great American Interchange (GAI, figure 8.9), with a major overland dispersal attesting to the development of the Panamanian isthmus by about 2.5 Ma. The North American fauna received mostly edentates and rodents from the Neotropics but gave in return a diversity of carnivores and ungulates.

If identifying the source of the immigrants on both sides of the Panamanian isthmus can be said to be straightforward, in that the host taxa are well known or can be readily sought, the opposite is the case for the dispersals recorded from the Late Cretaceous onward that require input from areas north of the main fossil-producing districts of the coterminous United States. This has been alluded to or mentioned directly in many chapters of this book, as has been the generally disappointing

record found in the Yukon and other northern territories. Currently limited data from Ellesmere Island in the Wasatchian, from the "Arikarean" Houghton Astrobleme of the Canadian Arctic, and from the arvicoline rodent record of the Pliocene and Pleistocene (and the *Bison* problem) illustrate the tantalizing uniqueness and importance of the Boreal province to the faunal record for more equator-ward districts in North America. Perhaps a future volume will provide further enlightenment.

Another goal is to diversify the geographic coverage of North American fossil mammal sites in parity with that of the paleobotanical record, especially for the Paleogene. This, plus the improved correlation network now in place, will permit a more comprehensive appraisal than reviewed here of the mammalian response to our increasingly sophisticated understanding of the climatic and topographic evolution of the continent during the Age of Mammals. Only then will mammalian paleontologists be in a position to evaluate more precisely the role global (Prothero, 1999), as well as local, change plays in molding mammalian evolution.

## ENDNOTES

1. Simpson coefficient is calculated as  $C/N1 \times 100$ , where  $C$  = the number of taxa being compared between two faunas and  $N1$  = number of taxa in the smaller of the two being compared. See Simpson (1960).
2. Exotic elements include *Carya* (hickory), *Diospyros* (persimmon), *Eugenia* (myrtle), *Gymnocladus* (Kentucky coffeetree), *Quercus* (oak), *Robinia* (locust), *Sophora* (mescal bean), *Ulmus* (elm), and *Zelkova* (Japanese keaki tree) (after Graham 1999:262).

## REFERENCES

- Abreu, V. S. and G. Haddad. 1998. Glacioeustatic fluctuations: The mechanism linking stable isotope events and sequence stratigraphy from the early Oligocene to middle Miocene. In *Mesozoic and Cenozoic sequence stratigraphy of European basins*, ed. P.-C. de Graciansky, J. Hardenbol, T. Jacquin, and P. R. Vail. Tulsa, OK: SEPM Special Publication 60:246–259.
- Abreu, V. S., J. Hardenbol, G. Haddad, G. R. Baum, A. W. Droxler, and P. R. Vail. 1998. Oxygen isotope synthesis: A Cretaceous ice house? In *Mesozoic and Cenozoic sequence stratigraphy of European basins*, ed. P.-C. de Graciansky, J. Hardenbol, T. Jacquin, and P. R. Vail. Tulsa, OK: SEPM Special Publication 60:75–80.
- Alvarez, L. W., W. Alvarez, F. Asaro, and H. V. Michel. 1980. Extraterrestrial cause for the Cretaceous–Tertiary extinction. *Science* 208:1095–1108.
- Aubry, M.-P. 2000. Where should the Global Stratotype Section and Point (GSSP) for the Paleocene/Eocene boundary be located? *Bulletin de la Société Géologique de France* 17:461–476.

- Aubry, M.-P., W. A. Berggren, J. A. Van Couvering, and F. Steininger. 1999. Problems in chronostratigraphy: Stages, series, unit and boundary stratotypes, Global Stratotype Section and Point and tarnished golden spikes. *Earth-Science Reviews* 46:99–148.
- Bains, S., R. M. Corfield, and R. D. Norris. 1999. Mechanisms of climate warming at the end of the Paleocene. *Science* 285:724–727.
- Barrera, E. and S. M. Savin. 1999. Evolution of late Campanian–Maastrichtian marine climates and oceans. In *Evolution of the Cretaceous ocean–climate system*, ed. E. Barrera and C. C. Johnson. Boulder, CO: *Geological Society of America Special Paper* 332:245–282.
- Barron, J. A. and J. G. Baldauf. 1990. Development of biosiliceous sedimentation in the North Pacific during the Miocene and Early Pliocene. In *Pacific Neogene events*, ed. R. Tsuchi. Tokyo: University of Tokyo Press, pp. 43–63.
- Baskin, J. A. 1998. Mustelidae. In *Evolution of Tertiary mammals of North America*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, pp. 152–173.
- Baskin, J. A. and R. H. Tedford. 1996. Small arctoid and feliform carnivorans. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 486–497.
- Berggren, W. A., D. V. Kent, C. C. Swisher III, and M.-P. Aubry. 1995. A revised Cenozoic geochronology and chronostratigraphy. In *Geochronology, time scales, and global stratigraphic correlation: Unified temporal framework for an historical geology*, ed. W. A. Berggren, D. V. Kent, M.-P. Aubry, and J. Hardenbol. Tulsa, OK: SEPM Special Publication 54:129–212.
- Berggren, W. A., S. Lucas, and M.-P. Aubry. 1998. Late Paleocene–early Eocene climatic and biotic evolution: An overview. In *Late Paleocene–early Eocene climatic and biotic events in the marine and terrestrial records*, ed. M.-P. Aubry, S. G. Lucas, and W. A. Berggren. New York: Columbia University Press, pp. 1–17.
- Bestland, E. A. 2002. Fossil andisols identified with mass-balance geochemistry (Oligocene John Day Formation, Oregon, USA). *Journal of Sedimentary Research* 72(5):673–686.
- Bestland, E. A., G. J. Retallack, and C. C. Swisher. 1997. Stepwise climate change recorded in Eocene–Oligocene paleosol sequences from central Oregon. *Journal of Geology* 105:153–172.
- Beu, A. G. 1973. Nautiloids of the genus *Aturia* from the Uppermost Miocene of Australia and New Zealand. *Science Report, Tohoku University, Second Series (Geology)*, special volume 6 (Prof. Hatai Memorial Volume):297–308.
- Bowen, G. J., W. C. Clyde, P. L. Koch, S. Ting, J. Alroy, T. Tsubamoto, Y. Wang, and Y. Wang. 2002. Mammalian dispersal at the Paleocene/Eocene boundary. *Science* 295:2062–2065.
- Brinkhuis, H. 1994. Late Eocene to early Oligocene dinoflagellate cysts from the Priabonian type-area (northeast Italy): Biostratigraphy and paleoenvironmental interpretation. *Palaeogeography, Palaeoclimatology, Palaeoecology* 107:121–163.
- Browning, J. V., K. G. Miller, and D. K. Pak. 1996. Global implications of lower to middle Eocene sequence boundaries on the New Jersey coastal plain: The icehouse cometh. *Geology* 24(7):639–642.
- Bryant, H. 1996. Nimravidae. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 453–475.
- Case, J. A., F. J. Goin, and M. O. Woodburne. In press. South American marsupials from the Late Cretaceous of North America and the origin of marsupial cohorts. *Journal of Mammalian Evolution*.
- Clemens, W. A. 2001. Patterns of mammalian evolution across the Cretaceous–Tertiary boundary. *Mitteilungen aus dem Museum für Naturkunde in Berlin; Zoologische Reihe* 77(2):175–191.
- . 2002. Evolution of the mammalian fauna across the Cretaceous–Tertiary boundary in northeastern Montana and other areas of the western interior. In *The Hell Creek Formation and the Cretaceous–Tertiary boundary in the northern Great Plains: An integrated continental record of the end of the Cretaceous*, ed. J. H. Hartman, K. R. Johnson, and D. J. Nichols. Boulder, CO: *Geological Society of America Special Paper* 361:217–245.
- Clift, P. and K. Bice. 2002. Baked Alaska. *Nature* 419:129–130.
- Clyde, W. C. 2001. Mammalian biostratigraphy of the McCullough Peaks area in the northern Bighorn Basin. In *Paleocene–Eocene stratigraphy and biotic change in the Bighorn and Clark's Fork basins, Wyoming*, ed. P. D. Gingerich. *University of Michigan Papers on Paleontology* 33:109–126.
- Colbert, M. W. and R. M. Schoch. 1998. Tapiroidea and other morphomorphs. In *Evolution of Tertiary mammals of North America*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, pp. 569–582.
- Compton, J. S. and D. J. Mallinson. 1996. Geochemical consequences of increased late Cenozoic weathering rates and the global CO<sub>2</sub> balance since 100 Ma. *Paleoceanography* 11:431–446.
- Corfield, R. M. and R. D. Norris. 1998. The oxygen and carbon isotopic context of the Paleocene/Eocene epoch boundary. In *Late Paleocene–early Eocene climatic and biotic events in the marine and terrestrial records*, ed. M.-P. Aubry, S. G. Lucas, and W. A. Berggren. New York: Columbia University Press, pp. 124–137.
- Currie, D. J. and V. Paquin. 1987. Large-scale biogeographical patterns of species richness of trees. *Nature* 239:326–327.
- Daams, R., A. J. van der Meulen, P. Pealez-Campomanes, and M. A. Alvarez-Sierra. 1999. Trends in rodent assemblages from the Aragonian (early–middle Miocene) of the Calatayud–Daroca Basin, Aragon, Spain. In *Hominoid evolution and climate change in Europe. Vol. 1: The evolution of Neogene terrestrial ecosystems in Europe*, ed. J. Agusti, L. Rook, and P. Andrews. Cambridge: Cambridge University Press, pp. 127–139.
- Dettman, D. L. and K. C. Lohmann. 2000. Oxygen isotope evidence for high-altitude snow in the Laramide Rocky Mountains of North America during the Late Cretaceous and Paleogene. *Geology* 28(3):243–246.
- Dewey, J. F. and W. C. Pitman. 1998. Sea-level changes: Mechanisms, magnitudes and rates. In *Paleogeographic evolution and non-glacial eustasy, northern South America*, ed. J. L. Pindell and C. Drake. Tulsa, OK: SEPM Special Publication 58:1–16.
- Duval, B. C., C. Cramez, and P. R. Vail. 1998. Stratigraphic cycles and major marine source rocks. In *Mesozoic and Cenozoic sequence stratigraphy of European basins*, ed. P.-C. de Graciansky, J. Hardenbol, T. Jacquin, and P. R. Vail. Tulsa, OK: SEPM Special Publication 60:43–51.
- Eberle, J. J. and J. A. Lillegraven. 1998. A new important record of earliest Cenozoic mammalian history: Eutheria and paleogeographic/biostratigraphic summaries. *Rocky Mountain Geology* 33(1):47–117.
- Effinger, J. A. 1998. Entelodontidae. In *Evolution of Tertiary mammals of North America*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, pp. 375–380.
- Ehrmann, W. U. and A. Mackensen. 1992. Sedimentological evidence for the formation of an East Antarctic ice sheet in

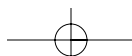
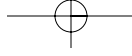
- Eocene/Oligocene time. *Palaeogeography, Palaeoclimatology, Palaeoecology* 9:85–112.
- Emry, R. J. and W. W. Korth. 1996. Cylindrodontidae. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 399–415.
- Emry, R. J., L. S. Russell, and P. Bjork. 1987. The Chadronian, Orellan, and Whitneyan North American land mammal ages. In *Cenozoic mammals of North America: Geochronology and biostratigraphy*, ed. M. O. Woodburne. Berkeley: University of California Press, pp. 118–152.
- Fischer, A. G. 1984. Two Phanerozoic supercycles. In *Catastrophes in Earth history*, ed. W. A. Berggren and J. A. Van Couvering. Princeton, NJ: Princeton University Press, pp. 129–150.
- Flower, B. J. and J. P. Kennett. 1993. Middle Miocene ocean–climate transition: High resolution oxygen and carbon isotopic records from Deep Sea Drilling Project site 588A, southwest Pacific. *Paleoceanography* 8:811–843.
- . 1995. Middle Miocene deepwater paleoceanography in the southwest Pacific: Relations with East Antarctic Ice Sheet development. *Paleoceanography* 10(6):1095–1112.
- Frakes, L. A. 1999. Estimating the global thermal state from Cretaceous sea surface and continental temperature data. In *Evolution of the Cretaceous ocean–climate system*, ed. E. Barrera and C. C. Johnson. Boulder, CO: Geological Society of America Special Paper 332:49–57.
- Frederiksen, N. O. 1988. Sporomorph biostratigraphy, floral changes, and paleoclimatology, Eocene and earliest Oligocene of the eastern Gulf Coast. *U.S. Geological Survey Professional Paper* 1448.
- Gale, A. S., J. Hardenbol, B. Hathway, W. J. Kennedy, J. R. Young, and V. Phansalkar. 2002. Global correlation of Cenomanian (Upper Cretaceous) sequences: Evidence for Milankovitch control on sea level. *Geology* 30(4):291–294.
- Garzzone, C. N., D. L. Dettman, J. Quade, P. G. DeCelles, and R. F. Butler. 2000. High times on the Tibetan Plateau: Paleoelevation of the Thakkhola graben, Nepal. *Geology* 28(4):339–342.
- Gaudin, T. J. and R. J. Emry. 2002. The late Eocene pangolin *Patriomanis* from North America, and a new genus of pangolin from the late Eocene of Nei Mongol, China (Mammalia, Pholidota). *Journal of Vertebrate Paleontology* (suppl. 3):57A.
- Gayet, M., J. C. Rage, T. Sempere, and P. Y. Gagnier. 1992. Modalités des échanges de vertèbres continentaux entre l’Amérique du Nord et l’Amérique du Sud au Crétacé Supérieur et au Paléocène. *Bulletin de la Société Géologique de France* 163:781–791.
- Gingerich, P. D. 1989. New earliest Wasatchian mammalian fauna from the Eocene of northwestern Wyoming: Composition and diversity in a rarely sampled high-floodplain assemblage. *University of Michigan Papers on Paleontology* 28:1–97.
- . 2001. Biostratigraphy of the continental Paleocene–Eocene boundary interval on Polecat Bench in the northern Bighorn Basin. In *Paleocene–Eocene stratigraphy and biotic change in the Bighorn and Clark’s Fork basins, Wyoming*, ed. P. D. Gingerich. *University of Michigan Papers on Paleontology* 33:37–72.
- Graciansky, P.-C. de, J. Hardenbol, T. Jacquin, and P. R. Vail. 1998. *Mesozoic and Cenozoic sequence stratigraphy of European basins*. Tulsa, OK: SEPM Special Publication 60.
- Graham, A. 1999. *Late Cretaceous and Cenozoic history of North American vegetation*. Oxford, UK: Oxford University Press.
- Gunnell, G. F. 1997. Wasatchian–Bridgerian (Eocene) paleoecology of the western interior of North America: Changing paleoenvironments and taxonomic composition of omomyid (Tarsiiformes) primates. *Journal of Human Evolution* 32:105–132.
- Hanson, C. B. 1996. Stratigraphy and vertebrate faunas of the Bridgerian–Duchesnean Clarno Formation, north-central Oregon. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 209–239.
- Haq, B. U., J. Hardenbol, and P. R. Vail. 1988. Mesozoic and Cenozoic chronostratigraphy and cycles of sea-level change. In *Sea-level changes: An integrated approach*, ed. C. K. Wilgus, B. S. Hastings, C. A. Ross, H. Posamentier, J. L. Van Wagoner, and C. G. St. C. Kendall. Tulsa, OK: SEPM Special Publication 42:71–108.
- Hardenbol, J., J. Thierry, M. B. Farley, T. Jacquin, P.-C. de Graciansky, and P. R. Vail. 1998. Mesozoic and Cenozoic sequence chronostratigraphic framework of European basins. In *Mesozoic and Cenozoic sequence stratigraphy of European basins*, ed. P.-C. de Graciansky, J. Hardenbol, T. Jacquin, and P. R. Vail. Tulsa, OK: SEPM Special Publication 60:Chart 1.
- Haug, G. H., R. Tieddemann, R. Zahn, and A. C. Ravelo. 2001. Role of Panama uplift on oceanic freshwater balance. *Geology* 29(3):207–210.
- Honey, J. G. 1998. Camelidae. In *Evolution of Tertiary mammals of North America*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, pp. 439–462.
- Huber, B. T. 1998. Tropical paradise at the Cretaceous poles? *Science* 282:2199–2200.
- Huber, B. T., R. D. Norris, and K. G. MacLeod. 2002. Deep-sea paleotemperature record of extreme warmth during the Cretaceous. *Geology* 30(2):123–126.
- Hudson, T. L. and L. B. Magoon. 2002. Tectonic controls on greenhouse gas flux to the Paleogene atmosphere from the Gulf of Alaska accretionary prism. *Geology* 30:547–550.
- Hulbert, R. C. Jr. and B. J. MacFadden. 1991. Morphological transformation and cladogenesis at the base of the adaptive radiation of Miocene hypsodont horses. *American Museum of Natural History Novitates* 3000.
- Hunt, R. M. Jr. 1990. Taphonomy and sedimentology of Arikaree (lower Miocene) fluvial, eolian, and lacustrine paleoenvironments, Nebraska and Wyoming: A paleobiota entombed in fine-grained volcanoclastic rocks. In *Volcanism and fossil biotas*, ed. M. G. Lockley and A. Rice. *Geological Society of America Special Paper* 244:69–111.
- . 1996. Biogeography of the order Carnivora. In *Carnivore behavior, ecology, and evolution*, ed. J. L. Gittleman. Ithaca, NY: Cornell University Press, pp. 485–541.
- . 1998a. Amphicyonidae. In *Evolution of Tertiary mammals of North America*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, pp. 196–227.
- . 1998b. Ursidae. In *Evolution of Tertiary mammals of North America*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, pp. 174–195.
- Immenhauser, A. and R. W. Scott. 1999. Global correlation of middle Cretaceous sea-level events. *Geology* 27(6):551–554.
- Itoigawa, J. and T. Yamanoi. 1990. Climatic optimum in the mid-Neogene of the Japanese Islands. In *Pacific Neogene events*, ed. R. Tsuchi. Tokyo: University of Tokyo Press, pp. 3–14.



- Iturralde-Vinent, M. A. and R. D. E. MacPhee. 1999. Paleogeography of the Caribbean region: Implications for Cenozoic biogeography. *American Museum of Natural History Bulletin* 238:1–95.
- Jacobs, D. A. and D. L. Sahagian. 1993. Climate-induced fluctuation in sea-level during non-glacial time. *Nature* 361:710–712.
- Jacquin, T. and P.-C. de Graciansky. 1998. Transgressive/regressive (second order) facies cycles: The effects of tectono-eustasy. In *Mesozoic and Cenozoic sequence stratigraphy of European basins*, ed. P.-C. de Graciansky, J. Hardenbol, T. Jacquin, and P. R. Vail. Tulsa, OK: SEPM Special Publication 60:31–42.
- Janis, C. M., J. A. Baskin, A. Berta, J. J. Flynn, G. F. Gunnell, R. M. Hunt Jr., L. D. Martin, and K. Munthe. 1998a. Carnivorous mammals. In *Evolution of Tertiary mammals of North America*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, pp. 73–90.
- Janis, C. M., M. W. Colbert, M. C. Coombs, W. D. Lambert, B. J. MacFadden, B. J. Mader, D. M. Prothero, R. M. Schoch, J. Shoshani, and W. Wall. 1998b. Perissodactyla and Proboscidea. In *Evolution of Tertiary mammals of North America*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, pp. 196–227.
- Janis, C. M., J. A. Effinger, J. A. Harrison, J. G. Honey, D. G. Kron, B. Lander, E. Manning, D. R. Prothero, M. S. Stevens, R. K. Stucky, S. D. Webb, and D. B. Wright. 1998c. Artiodactyla. In *Evolution of Tertiary mammals of North America*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, pp. 337–357.
- Janis, C. M. and E. Manning. 1998. Antilocapridae. In *Evolution of Tertiary mammals of North America*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, pp. 491–507.
- Jerzykiewicz, J. and A. R. Sweet. 1988. Sedimentological and palynological evidence of regional climatic changes in the Campanian to Paleocene sediments of the Rocky Mountain foothills, Canada. *Sedimentary Geology* 59:29–76.
- Keigwin, L. D. Jr. 1982. Isotopic paleoceanography of the Caribbean and East Pacific: Role of Panama uplift in late Neogene time. *Science* 217:350–353.
- Kelly, T. S. 1995. New Miocene horses from the Caliente Formation, Cuyama Valley Badlands, California. *Natural History Museum of Los Angeles County, Contributions in Science* 455:1–33.
- . 1998. New middle Miocene equid crania from California and their implications for the phylogeny of the Equini. *Natural History Museum of Los Angeles County, Contributions in Science* 473:1–43.
- Kennett, J. P. and L. D. Stott. 1995. Terminal Paleocene mass extinction in the deep sea: Association with global warming. In *Effects of past global change on life*, ed. S. M. Stanley, A. H. Knoll, and J. P. Kennett. National Research Council Studies in Geophysics. Washington, DC: National Academy Press, pp. 94–107.
- Knox, R. W. O. 1998. The tectonic and volcanic history of the North Atlantic region during the Paleocene–Eocene transition: Implications for NW European and global biotic events. In *Late Paleocene–early Eocene climatic and biotic events in the marine and terrestrial records*, ed. M.-P. Aubry, S. G. Lucas, and W. A. Berggren. New York: Columbia University Press, pp. 91–102.
- Krijgsman, W., M. Garcés, C. G. Langereis, R. Daams, J. van Dam, A. J. van der Meulen, J. Agustí, and L. Cabrera. 1996. A new chronology for the middle to late Miocene continental record in Spain. *Earth and Planetary Science Letters* 142:367–380.
- Kron, D. G. and E. Manning. 1998. Anthracotheriidae. In *Evolution of Tertiary mammals of North America*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, pp. 381–388.
- Lander, B. 1998. Oreodontoidea. In *Evolution of Tertiary mammals of North America*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, pp. 402–425.
- Leopold, E. and G. Liu. 1994. A long pollen sequence of Neogene age, Alaska Range. *Quaternary International* 22/23:103–140.
- Lillegraven, J. A. 1979. A biogeographical problem involving comparisons of late Eocene terrestrial vertebrate faunas of western North America. In *Historical biogeography, plate tectonics, and the changing environment*, ed. J. Gray and A. J. Boucot. Corvallis: Oregon State University Press, pp. 333–347.
- Lucas, S. G. 1992. Redefinition of the Duchesnean land mammal “age,” late Eocene of western North America. In *Eocene–Oligocene climatic and biotic evolution*, ed. D. R. Prothero and W. A. Berggren. Princeton, NJ: Princeton University Press, pp. 88–105.
- MacFadden, B. J. 1998. Equidae. In *Evolution of Tertiary mammals of North America*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, pp. 537–559.
- MacFadden, B. J., T. E. Cerling, and J. Prado. 1996. Cenozoic terrestrial ecosystem evolution in Argentina: Evidence from carbon isotopes of fossil mammal teeth. *Palaaios* 11:319–327.
- MacFadden, B. J., N. Solounias, and T. E. Cerling. 1999. Ancient diets, ecology, and extinction of 5-million-year-old horses from Florida. *Science* 283:824–827.
- MacFadden, B. J., Y. Wang, T. E. Cerling, and F. Anaya. 1994. South American fossil mammals and carbon isotopes: A 25 million year sequence from the Bolivian Andes. *Palaeoecology, Palaeoecology* 107:257–268.
- MacGinitie, H. D. 1953. Fossil plants from the Florissant beds, Colorado. *Carnegie Institute of Washington* 599:1–198.
- MacPhail, M. K., N. F. Alley, E. M. Truswell, and I. R. K. Sluiter. 1994. Early Tertiary vegetation: Evidence from spores and pollen. In *History of the Australian vegetation: Cretaceous to Recent*, ed. R. S. Hill. Cambridge, UK: Cambridge University Press, pp. 189–261.
- Mallinson, D. J. and J. S. Compton. 1997. Linking phosphogenic episodes on the southeast U.S. margin to marine  $^{13}\text{C}$  and  $^{18}\text{O}$  records. *Geology* 25:103–106.
- McKenna, M. C. 1983a. Cenozoic palaeogeography of North Atlantic Land bridges. In *Structure and development of the Greenland–Scotland Ridge*, ed. M. H. P. Bott, A. Saxov, M. Talwani, and J. Thiede. New York: Plenum, pp. 351–399.
- . 1983b. Holarctic landmass rearrangement, cosmic events, and Cenozoic terrestrial organisms. *Annals of the Missouri Botanical Garden* 70:459–489.
- McKenna, M. C. and S. K. Bell. 1997. *Classification of mammals above the species level*. New York: Columbia University Press.
- . 23 October 2000. *Mammal classification*. [ftp://ftp.amnh.org/people/mckenna/Mammalia.txn.sit](http://ftp.amnh.org/people/mckenna/Mammalia.txn.sit). Read, displayed, and searchable with Unitaxon browser. CD-ROM. 2001. Boulder, CO: Mathemaethetics, Inc.
- Miller, K. G., E. Barrera, R. K. Olsson, P. J. Sugarman, and S. M. Savin. 1999. Does ice drive early Maestrichtian eustasy? *Geology* 27(9):763–786.

- Miller, K. G., J. D. Wright, and R. G. Fairbanks. 1991. Unlocking the ice house: Oligocene–Miocene oxygen isotopes, eustasy, and margin erosion. *Journal of Geophysical Research* 96(B4): 6829–6848.
- Munthe, K. 1998. Canidae. In *Evolution of Tertiary mammals of North America*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, pp. 124–143.
- Ojha, T. P., R. F. Butler, J. Quade, P. G. DeCelles, D. Richards, and B. N. Upreti. 2000. Magnetic polarity stratigraphy of the Neogene Siwakik Group at Khutia Khola, far western Nepal. *Geological Society of America Bulletin* 112(3):424–434.
- Ozawa, T. and S. Tomida. 1996. Occurrence of *Aturia coxi* (Cephalopoda: Nautilida) from the uppermost Miocene of Japan and its implications for late Miocene climate in the northwestern Pacific. *Journal of Paleontology* 70(5):795–798.
- Pazzaglia, F. J. and S. A. Kelley. 1998. Large-scale geomorphology and fission-track thermochronology in topographic exhumation reconstructions of the southern Rocky Mountains. *Rocky Mountain Geology* 33(2):229–257.
- Pindell, J. L. 1994. Evolution of the Gulf of Mexico and the Caribbean. In *Caribbean geology: An introduction*, ed. S. K. Donovan and T. A. Jackson. Jamaica: The University of West Indies Publishers' Association, Mona, pp. 13–39.
- Pope, K. O. 2002. Impact dust not the cause of the Cretaceous–Tertiary mass extinction. *Geology* 30(2):99–102.
- Prothero, D. R. 1998a. Hyracodontidae. In *Evolution of Tertiary mammals of North America*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, pp. 375–380.
- . 1998b. Protoceratidae. In *Evolution of Tertiary mammals of North America*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, pp. 431–438.
- . 1998c. Rhinocerotidae. In *Evolution of Tertiary mammals of North America*, ed. C. M. Janis, K. M. Scott, and L. L. Jacobs. New York: Cambridge University Press, pp. 595–605.
- . 1999. Does climate change drive mammalian evolution? *GSA Today* 9(9):1–7.
- Prothero, D. R. and R. J. Emry. 1996. Summary. In *The terrestrial Eocene–Oligocene transition in North America* ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 664–683.
- Prothero, D. R. and R. H. Heaton. 1996. Faunal stability during the early Oligocene climatic crash. *Palaeogeography, Palaeoclimatology, Palaeoecology* 127:257–283.
- Quade, J., T. E. Cerling, and J. R. Bowman. 1989. Development of Asian monsoon revealed by marked ecological shift during the latest Miocene in northern Pakistan. *Nature* 342:163–166.
- Rage, J.-C. 1978. Une connexion continentale entre Amérique du Nord at Amérique du Sud au Crétacé supérieur? L'exemple des vertébrés continentaux. *Comptes Rendus Sommaire Séances Société Géologique de France* 6:281–285.
- Raymo, M. E. 1994. The Himalayas, organic carbon burial, and climate in the Miocene. *Paleogeography* 9:352–404.
- Raymo, M. E. and W. F. Ruddiman. 1992. Tectonic forcing of late Cenozoic climate. *Nature* 359:117–122.
- Rea, D. K. 1998. Changes in atmospheric circulation during the latest Paleocene and earliest Eocene epochs and some implications for the global climate regime. In *Late Paleocene–early Eocene climatic and biotic events in the marine and terrestrial records*, ed. M.-P. Aubry, S. G. Lucas, and W. A. Berggren. New York: Columbia University Press, pp. 118–123.
- Reiners, P. W., T. A. Ehlers, J. I. Garver, S. G. Mitchell, D. R. Montgomery, J. A. Vance, and S. Nicolescu. 2002. Late Miocene exhumation and uplift of the Washington Cascade Range. *Geology* 30(9):767–770.
- Savage, D. E. and D. E. Russell. 1983. *Mammalian paleofaunas of the world*. New York: Addison-Wesley.
- Shackleton, N. J., M. A. Hall, I. Raffi, L. Tauxe, and J. Zachos. 2000. Astronomical calibration age for the Oligocene–Miocene boundary. *Geology* 28(5):447–450.
- Simpson, G. G. 1960. Notes on the measurement of faunal resemblance. *American Journal of Science* 258-A:300–311.
- Sloan, L. C. and E. Thomas. 1998. Global climate of the late Paleocene Epoch: Modeling the circumstances associated with a climatic “event.” In *Late Paleocene–early Eocene climatic and biotic events in the marine and terrestrial records*, ed. M.-P. Aubry, S. G. Lucas, and W. A. Berggren. New York: Columbia University Press, pp. 138–157.
- Smith, M. E., B. Singer, and A. R. Carroll. 2003. <sup>40</sup>Ar/<sup>39</sup>Ar geochronology of the Eocene Green River Formation, Wyoming. *Geological Society of America Bulletin* 115:549–565.
- Stoll, H. M. and D. P. Schrag. 1996. Evidence for glacial control of rapid sea level changes in the early Cretaceous. *Science* 272:1771–1774.
- Storer, J. E. 1989. Rodent faunal provinces, Paleocene–Miocene of North America. In *Papers on fossil rodents in honor of Albert Elmer Wood*, ed. C. C. Black and M. R. Dawson. *Natural History Museum of Los Angeles County, Science Series* 33:1729.
- . 1996. Eocene–Oligocene faunas of the Cypress Hills Formation, Saskatchewan. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, p. 240–261.
- . 1998. The rodents of the Lac Pelletier Lower Fauna, late Eocene (Duchesnean) of Saskatchewan. *Journal of Vertebrate Paleontology* 8(1):84–101.
- Straight, S. G. 2001. New Wa-o mammalian fauna from Castle Gardens in the southeastern Bighorn Basin. In *Paleocene–Eocene stratigraphy and biotic change in the Bighorn and Clark's Fork basins, Wyoming*, ed. P. D. Gingerich. *University of Michigan Papers on Paleontology* 33:127–143.
- Thomas, D. J., J. C. Zachos, T. J. Bralower, E. Thomas, and S. Bohaty. 2002. Warming the fuel for the fire: Evidence for thermal dissociation of methane hydrate during the Paleocene–Eocene thermal maximum. *Geology* 30(12):1067–1070.
- Thomas, E. 1998. Biogeography of the late Paleocene benthic foraminiferal extinction. In *Late Paleocene–early Eocene climatic and biotic events in the marine and terrestrial records*, ed. M.-P. Aubry, S. G. Lucas, and W. A. Berggren. New York: Columbia University Press, pp. 214–243.
- Ting, S. 1998. Paleocene and early Eocene land mammal ages of Asia. In *Dawn of the age of mammals in Asia*, ed. K. C. Beard and M. R. Dawson. *Bulletin of the Carnegie Museum of Natural History* 34:124–147.
- Upchurch, G. R., B. L. Otto-Bliesner, and C. R. Scotese. 1999. Terrestrial vegetation and its effects on climate during the latest Cretaceous. In *Evolution of the Cretaceous ocean–climate system*, ed. E. Barrera and C. C. Johnson. *Geological Society of America Special Paper* 332:407–426.
- Villamil, T. and J. E. Pindell. 1998. Mesozoic paleogeographic evolution of northern South America: Foundations for sequence stratigraphic studies in passive margin strata deposited during non-glacial times. In *Paleogeographic evolution and non-glacial*

- eustacy, northern South America*, ed. J. L. Pindell and C. Drake. Tulsa, OK: SEPM Special Publication 58:283–318.
- Volkova, S., I. A. Kul'kova, and A. F. Fradkina. 1986. Palynostratigraphy of the non-marine Neogene in north Asia. *Review of Palaeobotany and Palynology* 48:415–424.
- Walsh, S. L. 1996. Middle Eocene mammal faunas of San Diego County, California. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 74–117.
- Wang, X., R. H. Tedford, and B. E. Taylor. 1999. Phylogenetic systematics of the Borophaginae (Carnivora: Canidae). *American Museum of Natural History Bulletin* 243:1–391.
- Wang, Y., T. E. Cerling, and B. J. MacFadden. 1994. Fossil horses and carbon isotopes: New evidence for Cenozoic dietary, habitat, and ecosystem changes in North America. In *Stable isotope and trace-element geochemistry of vertebrate fossils: Interpreting ancient diets and climates*, ed. B. J. MacFadden and J. D. Bryant. *Paleogeography, Paleoclimatology, Paleoeecology* Special Issue 107:269–280.
- Webb, S. D. 1998. Hornless ruminants. In *Evolution of Tertiary mammals of North America*, ed. C. M. Janis, K. M. Scott, and L. S. Jacobs. New York: Cambridge University Press, pp. 463–476.
- Webb, S. D., R. C. Hulbert, and W. D. Lambert. 1995. Climatic implications of large-herbivore distributions in the Miocene of North America. In *Paleoclimate and evolution: With emphasis on human origins*, ed. E. S. Vrba, G. H. Denton, T. C. Partridge, and L. H. Burckle. New Haven, CT: Yale University Press, pp. 91–108.
- Webb, S. D. and N. D. Opdyke. 1995. Global climatic influence on Cenozoic land mammal faunas. In *Studies in geophysics: Effects of past global change on life*. Board on Earth Sciences and Resources Commission on Geosciences, Environment and Resources National Research Council. Washington, DC: National Academy Press, pp. 184–208.
- Webb, S. D. and B. E. Taylor. 1980. The phylogeny of hornless ruminants and a description of the cranium of *Archaeomeryx*. *Bulletin of the American Museum of Natural History* 167:121–157.
- Weil, A. and T. Williamson. 2000. Diverse Maastrichtian terrestrial vertebrate fauna of the Naashoibito Member, Kirtland Formation (San Juan Basin, New Mexico) confirms “Lancian” faunal heterogeneity in western North America. *Geological Society of America Abstracts with Programs* 32(7):498.
- White, J. M. and T. A. Ager. 1994. Palynology, palaeoclimatology and correlation of middle Miocene beds from Porcupine River (Locality 90-1), Alaska. *Quaternary International* 22/23:43–77.
- White, J. M., T. A. Ager, D. P. Adam, E. B. Leopold, G. Liu, G. Jetté, and C. F. Schweger. 1997. An 18 million year record of vegetation and climate change in northwestern Canada and Alaska: Tectonic and global climatic correlates. *Palaeogeography, Palaeoclimatology, Palaeoecology* 130:293–306.
- Wolfe, J. A. 1966. Tertiary plants from the Cook Inlet region, Alaska. *U.S. Geological Survey Professional Paper* 398-B4–B32.
- . 1977. Paleogene floras from the Gulf of Alaska region. *U.S. Geological Survey Professional Paper* 997:1–108.
- . 1987. Leaf assemblages across the Cretaceous–Tertiary boundary in the Raton Basin, New Mexico and Colorado. *Proceedings of the National Academy of Science* 84:5096–5100.
- . 1992. Climatic, floristic, and vegetational changes near the Eocene/Oligocene boundary in North America. In *Eocene–Oligocene climatic and biotic evolution*, ed. D. R. Prothero and W. A. Berggren. Princeton, NJ: Princeton University Press, pp. 421–436.
- . 1994a. An analysis of Neogene climates in Beringia. *Palaeogeography, Palaeoclimatology, Palaeoecology* 108:207–216.
- . 1994b. Tertiary climatic changes at middle latitudes of western North America. *Palaeogeography, Palaeoclimatology, Palaeoecology* 108:195–205.
- Wolfe, J. A., C. E. Forest, and P. Molnar. 1998. Paleobotanical evidence of Eocene and Oligocene paleoaltitudes in midlatitude western North America. *Geological Society of America Bulletin* 110:664–678.
- Wolfe, J. A., D. M. Hopkins, and E. B. Leopold. 1966. Tertiary stratigraphy and paleobotany of the Cook Inlet Region, Alaska. *U.S. Geological Survey Professional Paper* 398-A.
- Wolfe, J. A., H. E. Schorn, C. E. Forest, and P. Molnar. 1997. Paleobotanical evidence for high altitudes in Nevada during the Miocene. *Science* 276:1672–1675.
- Wolfe, J. A. and T. Tanai. 1980. The Miocene Seldovia Point flora from the Kenai Group, Alaska. *U.S. Geological Survey Professional Paper* 1105:1–52.
- Wolfe, J. A. and G. R. Upchurch. 1987. North American nonmarine climates during the Late Cretaceous. *Palaeogeography, Palaeoclimatology, Palaeoecology* 61:33–77.
- Woodburne, M. O. 1989. Hipparion horses: A pattern of endemic evolution and intercontinental dispersal. In *The evolution of perissodactyls*, ed. D. R. Prothero and R. M. Schoch. *Oxford Monographs on Geology and Geophysics* 15:197–233. New York: Oxford University Press.
- . 1996. Precision and resolution in mammalian chronostratigraphy: Principles, practices, examples. *Journal of Vertebrate Paleontology* 16(3):531–555.
- Woodburne, M. O. and J. A. Case. 1996. Dispersal, vicariance, and the Late Cretaceous to early Tertiary land mammal biogeography from South America to Australia. *Journal of Mammalian Evolution* 3(2):121–161.
- Woodburne, M. O. and C. C. Swisher III. 1995. Land mammal high-resolution geochronology, intercontinental overland dispersals, sea level, climate, and vicariance. In *Geochronology, time scales, and global stratigraphic correlation: Unified temporal framework for an historical geology*, ed. W. A. Berggren, D. V. Kent, M.-P. Aubry, and J. Hardenbol. Tulsa, OK: SEPM Special Publication 54:335–364.
- Woodruff, F. and S. M. Savin. 1989. Miocene deepwater oceanography. *Palaeoceanography* 6:755–806.
- Xu, X. 1996. Castoridae. In *The terrestrial Eocene–Oligocene transition in North America*, ed. D. R. Prothero and R. J. Emry. New York: Cambridge University Press, pp. 417–432.
- Zachos, J. C., K. C. Lohmann, J. C. G. Walker, and S. W. Wise. 1993. Abrupt climate change and transient climates during the Paleogene: A marine perspective. *Journal of Geology* 101:191–213.
- Zachos, J. C., M. Pagani, L. Sloan, E. Thomas, and K. Billups. 2001. Trends, rhythms, and aberrations in global climate 65 Ma to present. *Science* 292:686–693.



## Systematic Index

### Plants

*Aquilapollenites* 24R, 318R  
 calcareous nannofossils 89R  
*Celtis* 326R  
 coccoliths 124R  
 Coyote Creek flora 120L  
*Ephedra* 326R  
 nannoplankton 133R, 134L  
*Normapolles* 24R, 318R  
*Nothofagus* 330L  
*Ocotea* 326R  
*Wodehousia spinata* 32L

### Invertebrates

ammonite(s) 34L, R  
*Arctica islandica* 237L  
 cephalopods 25L  
 foraminifera(ns) 24L, 89R, 90L, 92L,  
 94R, 124R, 171R, 206R, 317L, 320L, R,  
 334L  
*Hoploscaphites birkelundi* 34R  
*H. aff. nicoletti* 34R  
*Jeletzkytes nebrascensis* 34L, R  
 mollusca(n) 124R, 269L, 291L, 336R

### Non-mammalian vertebrates

chondrichthyan(s) 92R  
*Glyptosaurus* 131L  
 ray 205L  
 shark 205L

### Mammals

*Aaptoryctes* 55, 79R, 84L  
*Absarokius* 112R, 113L, 114L, R, 120R  
 Aceratherinae 231L  
 aceratheriine rhino(s) 332L, 333L, 334L  
*Achaenodon* 117L, 120R, 121L, 127R  
*Acheronodon* 54, 63R, 64L, 65R, 80L

*Acidomomys* 86L, 87R  
*Aclistomycter* 119R, 129R, 329R, 330R  
*Acmeodon* 55, 71R, 73R, 75L, 76L, 79R,  
 81L, R, 82R  
*Acritohippus* 174L, 219R  
*A. isonesus* 178R, 188L, 189R, 190L,  
 192R, 199L, 206R, 207L  
*A. quinni* 174L  
*A. styloidontus* 176L, 190R  
*A. tertius* 173L, 175R  
*Acritoparamys* 86L, 87L, 88L, R, 324L  
*A. atavus* 86L  
*Actiocyon* 217R  
 adapid primate 329L  
 adapiform(s) (euprimates) 111R, 112R  
 adapisoricid(s) 82L, 126L  
*Adjidaumo* 119R, 164R, 165L, 329L  
*A. craigi* 119R  
*A. minutus* 164R  
*Adunator* 58, 71R, 75L, 76L, 79R, 81L, R,  
 82R, 83R, 84L, R, 86L, 87L, 88L  
*Aelurodon* 172L, 195L, 197R, 215L, 217L  
*A. aphobus* 171L  
*A. asthenostylus* 171R, 176L, 177R,  
 190L, R  
*A. ferox* 192R, 199R  
*A. saevus* 192R  
*A. taxoides* 171L, 198L, 200L, 217L  
*A. wheelerianus* 199R  
*A. (Strobodon)* 199R  
*A. (Strobodon) stirtoni* 199R, 215R,  
 217L  
 Aelurodontina 213R  
 aelurodontine borophagine canid(s)  
 195L  
*Aepyamelus* 172L, 173R, 174L, 175R,  
 188L, 189R, 195R, 199L, 200L, 208L,  
 213R, 218R

*A. elrodi* 190R  
*A. madisonensis* 190R  
*Aethomylos* 117R, 118L  
*Agnotocastor* 188L, 189R, 205L, 211L  
*A. praetereadens* 166L  
*A. readingi* 164L  
 agriochoerid(s) 117R, 130R, 133R, 328L,  
 R, 329L  
*Agriochoerus* 118R, 119R, 129R, 179R,  
 211L  
*Agriotherium* 200R, 218R, 249R, 252L,  
 259R  
*A. gregoryi* 175L, 251R  
*Aguascalientia* 201L, R  
*A. wilsoni* 201R  
 ailuravine rodent(s) 328R, 329L  
*Alagomys* 55, 86L, 87L, 324L  
*Alcidedorbignya* 93R  
*Alces* 285L, 288L  
*A. latifrons* 285R  
*Aletodon* 60, 79L, 82R, 83R, 84L, R,  
 86L, 87L, 88L  
*A. gunnelli* 86L, 87R  
 aletomerycine dromomerycid(s) 332R  
*Aletomeryx* 213L  
*A. occidentalis* 176R  
*Alforjas* 218L, R, 337R  
*Alilepus* 215R, 250R  
*A. vagus* 256R, 260L  
 allomyid, ine rodent(s) 330R, 331L, 334R  
*Allomys* 179R, 187L, R, 193L, 195R, 211L,  
 216L  
*Allophaiomys* 246L, 265R, 266L, R, 267L,  
 R, 273L, R, 275L, 278R, 279R, 280L,  
 281L, R, 282R, 284L, 290R, 294L  
*A. pliocoenicus* 250R, 270R, 279L  
*Alostera* 28R, 32, 33, 55, 63L, R, 64L, 65R  
*A. saskatchewanensis* 28R, 33

- Alphadon* 30, 54, 64L, R, 64L, R, 65R, 321L  
*A. eatoni* 28R  
*A. jasoni* 28R  
*A. wilsoni* 28R  
 alphadontid(s), alphadontine(s) (marsupial) 321L  
*Alphagaulus* 178R, 190L, R, 191L, 196L, 199L  
*A. douglassi* 191L  
*A. pristinus* 190L  
 Alticonodontinae 23L  
*Alticonus* 61, 63R, 67R, 321R  
*Aluralagus* 235, 250L, 264R  
*Alveugena* 56, 63R, 67L, 321R  
*Alwoodia* 179R, 188R, 189L, 193L, R, 196R, 211L  
*A. cristabrevis* 191L  
*A. harkseni* 211L  
*A. magna* 211L  
*Ambloctonus* 112R  
 amblypod(s) 132L  
*Amebelodon* 218R  
*Amelotabes* 56, 79R, 83R  
*Amphechinus* 188R, 196R, 205L, 211L  
*Amphicaenopus* 330R  
*Amphictis* 213L, 333L  
 amphicyodontine ursid 328R  
*Amphicyon* 171R, 192R, 199L, 208L, 213L, 215L, 333L  
*A. ingens* 176L, 199R  
*A. sinapius* 171R, 175R  
 amphicyonid(s), ine(s) 171R, 179R, 195L, 199R, 211R, 212L, 213L, R, 215L, R, 329L, 330R, 331L, 332R, 333L, 334L, R, 335L  
 Amphicyonidae 217R  
*Ampliconus* 61, 63R, 64R, 65R, 67R  
*Amynodon* 111R, 116R, 117L, 119R, 120R, 121L, 127R, 129R, 130R  
 amyodontine rhino(s) 328L, 329L, 330R  
*Amynodontopsis* 118R, 119L, 124R, 129R, 133R  
*Anacodon* 60, 79L, 83L, 84L, R, 86L, 87R, 112R  
 anagalid(s) 90R  
*Anancus* 249R  
*Anaptomorphus* 113L, 114R, 120R  
*A. westi* 115L  
*Anasazia* 59, 72L, 76L  
*Anchippodus* 132R  
*Anchippus* 204L, R, 205L  
*A. texanus* 205R  
 Anchitheriinae 217R  
 anchitherine(s) 176L, 200L, 332R  
*Anchitheriomys* 192R, 195R, 208L, 213R, 216L  
*Anchitherium* 178R, 195R, 199L, 203R, 208L, R, 216L  
*A. clarencei* 175L, 206R  
*Anconodon* 54, 71R, 75L, R, 76L, 79R, 80R, 81L, R  
*Angustidens* 190R  
*Anemorhysis* 112R  
*Anisonchus* 61, 62R, 63L, 67R, 70L, 72L, 73R, 75L, 76L, 78R, 79R, 80R  
*A. sectorius* 78L  
*Ankalagon* 61, 72L, 75L, 76L, 324L  
*Ankylodon* 117R  
*Anonymus* 119R  
*A. baroni* 119R  
*Anouroneomys* 218L  
*Antecalomys* 217R  
*Antesorex* 213L, 333L  
 anthracothere 189L, 211R, 328R, 331L, 332R  
 anthracotheriid, iine 328R, 332R  
 Anthracotheriidae 213L  
*Antiacodon* 113L, R, 114R, 115L, 117L, R, 328L  
*Antilocapra* 288L, R  
*A. americana* 281L  
 antilocaprid(s), artiodactyl 190R, 195L, R, 196L, 199R, 213R, 215L, 263R, 333L, 335L, 336L, 337R  
 Antilocapridae 212L, R, 213R, 334L  
 antilocaprine 200R, 216L, 216R, 218L, R, 219L  
 apatemyid(s) 82L, 323R, 329R  
*Apatemys* 56, 78R, 79L, 82R, 83R, 84L, R, 86R, 87L, 88L, 114R, 116R, 119R, 133L, 325R  
*Apatosciuravus* 55, 86L, 87L, 88L, 324L  
*Aphelops* 172L, 175R, 195L, 199L, 200L, 206R, 207R, 213R, 334L, 337R  
*A. jemezianus* 200L  
*A. mutilus* 218R  
*Apheliscus* 60, 86L, 87L, 88L, 113L  
*A. nitidus* 86L, 88L  
*Aphronorus* 56, 71R, 73R, 75L, R, 76L, 79R, 81L, R, 82R  
*A. ratatoski* 78L  
*Aplodontia* 285L  
*A. rufa* 285L  
 aplodontid(s) rodents 179L, 187L, R, 188R, 191L, 195R, 211L, 216L  
 Aplodontidae 211L  
*Apriculus* 117R  
*Apternodus* 119R  
*Archaeocyon* 173L  
*A. pavidus* 173L  
*Archaeohippus* 176L, 192R, 195R, 199L, 203R, 204L, 208L, 212L, 216L, 332L  
*A. blackbergi* 208L  
*A. minimus* 190L  
*A. mourningi* 172L, 174L, 176L  
*A. penultimus* 173R  
*A. stenolophus* 192L  
*A. ultimus* 188L  
 archaeolagine(s), lagomorph 187R, 198L, 218L  
*Archaeolagus* 172R, 179R, 188R, 189L, 192L, 193R, 196R, 198R, 199L, 201L, 330R  
*A. ennisianus* 211L  
*A. esmeraldensis* 191L  
*Archaeolambda* 91R  
*Archaeomeryx* 328R  
*Archaeotherium* 161R  
*A. coarctatum* 163R  
*A. mortoni* 165L  
*A. trippensis* 197L  
 archaic ungulates 36L, 67L  
 Arctocyoniidae 60, 62R  
 arctocyonid(s), arctocyonine(s) 62L, 82L, 321R, 323R, 324R  
*Arctodontomys* 58, 86L, 87L, 88L, 113L  
*Arctodus* 278L  
*A. pristinus* 270L, 283R, 284L, 285L  
*A. simus* 270L, 278R, 282R  
 arctoid(s), carnivore 199R  
*Arctomyoides* 190L  
*Arctonasua* 215L  
*Arctostylops* 60, 79L, 84L, R, 86L, 87R, 88L, 91R  
*A. steini* 120L  
 Arctostylovida, arctostylopid(s) 91L, R, 92L, 324L  
 Arctostylopidae 94R  
*Ardynomys* 119R, 328R, 329R  
*Arfia* 112R, 325L  
*Arikareomys* 193R, 205L, R, 211R  
*Armintodelphys* 113L  
*Arretotherium* 189L, 197L, 204R, 211R  
*A. acridens* 189L, 204R  
*A. fricki* 189L, 192L  
*A. leptodus* 189L, 204R  
 Artiodactyla 85R, 88L, 89R, 106L, 112R, 325L  
 artiodactyl(s) 90L, 120R 122L, 126L, 130R, 134R, 324R, 327R, 328L, R, 329R, 333L  
*Arvicola* 246L  
 Arvicolidae 239R  
 Arvicolinae 239R, 245L  
 arvicoline(s), rodent 198R, 218R, 219L, 232L, 239R, 245L, 247L, R, 249R, 250L, 252R, 253L, R, 254L, R, 258R, 261R, 262L, 265R, 276L, R, 278R, 279L, R, 281L, 292R, 293L, R, 294L, 295L, R, 338L, R  
*Asiostylops* 91L, R  
 astrapothere(s) 92R  
*Astrohippus* 219L, 249R, 337R  
*A. ansae* 200R, 218R  
*A. stocki* 200R, 252L  
*Atopomys* 276L, 282R, 284L  
*A. texensis* 268R, 282R  
*Aulolithomys* 119R

- Auraria* 61, 63R, 64R, 65R  
*Australocamelus* 199L, 200L, 213L  
*Auxontodon* 118L, 133L  
*Avunculus* 56, 72L, 75L, 76L  
*Aztlanolagus* 282R  
   *A. agilis* 282R  
*Azygonyx* 120L  
   *A. ancylion* 120L
- Baioconodon* 60, 62R, 63L, R, 64L, R, 65R, 67R, 69R  
*Baiomys* 200R, 219L, 261R  
   *B. minimus* 261R  
*Baiotomeus* 54, 64L, 71R, 75R, 76L, 79R, 81L, R, 82R  
*Barbourofelis* 171L, 172R, 216R, 217L, 218R  
   *B. lovei* 171L, 172R, 217R  
   *B. morrisoni* 217R  
   *B. osborni* 217L  
   *B. whitfordi* 217L  
*Barbouromeryx* 187L, 196L, 212L, 213R  
*Barylambda* 57, 78R, 82R, 83R, 84L, R, 86R, 87L, 88L  
*Bassariscus* 260R  
   *B. casei* 260R  
*Bathogenys* 120L, 161R, 162R  
*Bathyopsis* 114L, 115L  
   *B. fissidens* 114R  
   *B. middlewsi* 115L  
*Bathyopsoides* 78R  
*Batodon* 28L, 58, 63L, R, 64L, 65R, 318R  
   *B. tenuis* 28R  
*Batodontoides* 118L  
beaver 187R, 189L, 193L, 195L, R, 198L, 211L, 212L, 213L, R, 215L, R, 216L, 217L, R, 218L, R, 331R, 333L, 334L, 337R  
*Beckia* 217R  
*Bemalambda* 91L  
*Bison* 236, 245, 247R, 264R, 269R, 272L, 273L, 274R, 281L, 283L, 284R, 285–289, 290L, 293L, 295L  
   *B. alaskensis* 287R  
   *B. alleni* 287R  
   *B. antiquus* 287R  
   *B. bison* 287R, 288R  
   *B. b. antiquus* 287R  
   *B. b. bison* 287R  
   *B. b. occidentalis* 287R  
   *B. latifrons* 286L, 287R  
*Bisonalveus* 56, 79R, 80R, 82R  
*Blackia* 173R  
*Blancocamelus* 263L  
*Blarina* 250R, 270L, 290R  
blastomerycine artiodactyl(s) 332R  
*Blastomeryx* 188L, 191L, 192L, 200L, 206R, 208L, 212L, 217R  
   *B. elegans* 190R  
   *B. galushai* 208L  
   *B. gemmifer* 190R  
*Blickomylus* 196L, 199L, 213L  
*Bomburia* 62R  
*Bootherium* 285R  
   *B. bombifrons* 285R  
Borophagina 213R  
borophagine canid(s) 171R, 176L, 177R, 187L, 190R, 195L, 196L, 198L, 199L, R, 207L, 212L, 213L, R, 215L, R, 217L, R, 218L, R, 330R, 331R, 332R, 334L  
Borophagina 213R  
Borophagini 213R  
*Borophagus* 217L, 249R, 263R, 264L, 267L, 281L  
   *B. diversidens* 244, 249R, 250L, R, 259L, 260R, 264L, 265L, 266L, R, 267L  
   *B. dudleyi* 219L  
   *B. hilli* 219L, 244, 250R, 256R, 258L, 264L  
   *B. littoralis* 171L, 172L, R, 177L, 217L  
   *B. parvus* 218R  
   *B. pugnator* 218L  
   *B. secundus* 203R, 218L, 219L  
   *B. solus* 259R  
*Bothriodon* 328R  
bothriodontine anthracothere 328R  
*Bothriostylops* 91R  
*Bouromeryx* 172L, 178R, 187L, 196L, 206R, 207L, 213R, 216L  
   *B. americanus* 190R  
   *B. milleri* 174L  
*Brachycrus* 174L, 188L, 196L, 199R, 203R  
   *B. buwaldi* 176L  
   *B. rusticus* 190L  
brachyericine 211L  
*Brachyerix* 190R, 192R, 196L  
*Brachyhyops* 118R, 119R  
*Brachylagus* 236, 270R, 285L  
   *B. idahoensis* 236, 270R, 285L  
*Brachyops* 119R, 130R,  
*Brachypotherium* 213L, 332R, 333L  
*Brachyprotoma* 236, 270R, 285L  
*Brachypsalis* 217R  
*Brachyrhynchocyon* 161R  
   *B. dodgei* 161R  
*Bretzia* 261R  
brontothere(s), iid(s) 115L, 124R, 126L, 129R, 130R, 131R, 133L, 133R, 134L, 160L, R, 161L, R, 163R, 164L, 325L, 328L  
*Bubodens* 29, 33  
   *B. magnus* 28R, 33, 35L  
*Bubogonia* 60, 63R, 67R, 69R, 321R  
*Buisnictis* 198L, 219L  
bunodont artiodactyl(s) 120R  
bunomerycine(s) 328L  
*Bunomeryx* 117L, R, 118L  
*Bunophorus* 114L, R, 122L  
*Caenolambda* 57, 79R, 81R, 82R  
*Caenopus* 133R, 189L  
*Calippus* 192R, 195L, 197R, 202R, 203R, 215L, 218R, 251L, 334L, 337R  
   *C. cerasinus* 217R  
   *C. hondurensis* 203R  
   *C. proplacidus*, 207L  
   *C. (Grammohippus) martini*, 198L  
*Calomys* 198R, 200R, 261R, 262L  
   *C. (Bensonomys)* 198R, 200R, 218R, 261R  
   *C. (Bensonomys) arizonae* 262L  
camel(s), artiodactyl, 166L, 172L, 173R, 175L, 187L, 188L, 189R, 190R, 193L, 195L, R, 196L, 197L, 198R, 199L, R, 200L, R, 201L, R, 202L, R, 211L, R, 212L, 213L, 217R, 219L, 260R, 263L, 282L, 331L, R, 334L  
Camelidae 117R, 213R  
camelid(s) 117R, 118L, 130R, 173L, 195L, 199L, 201L, 202R, 206L, 207L, 211R, 213R, 215L, 261R, 282R, 328L, 330R, 332R, 336L, 337R  
*Camelops* 249R, 265R, 278L, R, 280R, 281L, 282L, R, 284R, 286L, R, 288L, 337R  
   *C. hesternus* 288L, R  
   *C. minidokae* 281L  
*Campestrallomys* 165L  
   *C. annectens* 165L  
canine(s), carnivore 195L, 196R, 199R, 331R  
*Canis* 235, 249R, 270R, 290R  
   *C. armbrusteri* 236, 270L, 278R, 280L, 282L, R, 283L, R, 284L  
   *C. dirus* 236, 281L, R, 284R, 285L, 288L  
   *C. edwardii* 250R, 259R, 268L, 270R,  
   *C. latrans* 236, 270R, 283L, 285L, 288L  
   *C. lepophagus* 250L, 256R, 258L, 263L, 265L, R, 266L, R, 267L,  
   *C. lupus* 236, 270R, 283R, 285L, 288L  
*Cantius* 90L, 112R, 113L, 114R, 115L, 120L, 121L, 132R, 325L  
*Capacikala* 211L  
   *C. gradatus* 211L  
*Capatanka* 211L  
   *C. cankpeopi* 211L  
Caprini 260R  
*Capromeryx* 256R, 278L, R, 282L  
   *C. arizonensis* 268L  
*Carcinodon* 60, 62R, 63R, 67R, 69R, 321R  
Carnivora 57, 91L, 171L  
carnivoran 91L, 279L, 284R, 325R  
carnivore(s) 82L, 92L, 134L, 190L, 196R, 198R, 291R, 202R, 205R, 216R, 218L, 324R, 328L, R, 329L, 331L, R, 337L, R, 338L

- canid(s), 118L, 171L, 173L, 176L, 179L, 190R, 195L, 196L, 198L, 199R, 200R, 205L, 211L, 212L, 213L, R, 215L, R, 217L, R, 218L, R, 219L, 252L, 328R, 329R, 330R, 331L, R, 332R, 335L, 337L
- Canidae 171L, 202R
- Canini 217R, 218L
- Carpocristes 59, 79R, 83R, 84L
- Carpocyon 195L, 199R, 215L, 219L  
*C. compressus* 178L, 197R  
*C. robustus* 217L
- Carpodaptes 59, 73L, 79R, 80R, 81R, 82R, 83R, 84L
- Carpolestes 59, 79L, 84L, R, 85R, 86L, 87L, 88L  
*C. nigridentis*, 86L
- carpolestid(s) 59, 75L, 82L, 323R, 324L
- Carpomegodon 59, 83R
- Carsiptychus 62R
- Castor 218R, 261R, 337R  
*C. californicus* 258L
- castorid (ine) beaver(s) 190R, 333L, 334L
- castorimorph(s) 329L
- Castoroides 244, 283R  
*C. ohioensis* 281R
- Catopsalis 54, 63L, 64L, 65R, 67R, 70L, 71R, 72L, 73R, 75L, R, 76L, 79R, 81L, R, 82R, 83L, 321R,
- Cedrocherus 58, 79R, 82R
- Cedromus 164R, 166L  
*C. wardi* 164R  
*C. wilsoni* 166L
- Centetodon 116L, 119R, 129R, 192L, 193R, 205L, R, 211R  
*C. marginalis* 165L  
*C. chadronensis* 162R
- Cephalogale 198R
- Ceratogaulus 215L  
*C. hatcheri* 218R
- Ceratomeryx 249R
- cervid(s) 175L, 178L, 207R, 251L, 253L, 261R, 332R, 337R
- Cervus 249R  
*C. canadensis* 288L
- Cete 61, 62R
- Ceutholestes 58, 86L, 87L, 88L
- Chaenohyus 211L
- Chadrolagus 163L  
*C. emryi* 163L
- Chalicomomys 58, 86L, 87L, 88L, R
- chalicotherid(s), oid(s) 195R, 196L, 211R, 328L
- Chalicotheriidae 216L
- Chasmaporthetes 235, 251L, 253L  
*C. johnstoni* 267L  
*C. ossifragus* 267L
- Chipetaia 117R
- Chiromyoides 58, 79L, 82R, 83R, 84L, 86L, 87L, R
- C. caesor* 83R  
*C. major* 87R  
*C. minor* 82R  
*C. potior* 84L
- Choeroclaenus 60, 63R, 67R, 69R, 321L
- Chriacus 60, 62R, 63L, 67R, 70L, 71R, 72L, 73R, 75L, 76L, 79R, 81L, R, 82R, 83R, 84L, 86R, 87L, 88L, 321R
- Chumashius 119R  
*C. balchi* 119R
- Cimexomys 23L, 29, 33, 54, 63R, 64L, 65R, 69R  
*C. gregoryi* 29  
*C. judithae* 29  
*C. minor* 28R, 29, 33
- Cimolestes 31, 33, 56, 63R, 65L, R, 67R, 69R, 92R  
*C. cerberoides* 28R, 31, 33  
*C. incisus* 28R, 31, 33  
*C. magnus* 28R, 31, 33  
*C. propalaeoryctoides* 28R, 31, 33  
*C. stirtoni* 28R, 31, 33
- cimolestid(s), cimolestan 64L, 92R, 318R, 321L, R, 323R
- Cimolodon 54, 63R, 64L, R, 65R, 67R, 69R
- cimolodontid (multituberculate) 318R
- Cimolomyidae, cimolomyid(s) 23R, 28R, 29, 33
- Cimolomys 28R, 29, 54, 63L, R, 64L, R, 65R  
*C. clarki* 24R, 29  
*C. gracilis* 28R, 29, 33  
*C. millensis* 29  
*C. trochuus* 28R, 29, 33
- Cimolodontidae, cimolodontid(s) 23L, R, 29, 33
- Claenodon 60, 71R, 75L, 78R, 79R, 81L, R, 82R, 83R, 84L
- Clemensodon 28R  
*C. megaloba* 28R
- Clethrionomys 236, 245, 275L, R, 276L, 278R, 279R, 280L, 285L
- Colodon 117R, 118L, 121L, 133L, R
- Colpoclaenus 60, 71R, 75L, R, 76L, 79R, 81L, R, 82R
- Comancheomys 198L
- Conacodon 61, 62R, 65R, 66R, 67R, 70L, 72L, 74L, 75L, 76L, 79R, 81L, R, 82R, 321R
- Conepatus 236, 270R, 285L
- condylarth(s, -ran) 62L, R, 71L, 86L, R, 92L, R, 132L, 133R, 324L, R, 329L
- Condylarthra 60, 62L, 85L, 93R
- Conoryctella 56, 73R, 74R, 75L
- Conoryctes 56, 72L, 75L, 76L
- Copecion 61, 79R, 82R
- Copedelphys 165L  
*C. stevensoni* 165L
- Copelemur 112R
- Copemys* 175R, 192R, 200R, 206R, 207L, 213R, 214R, 215L, 262L, 334L, R  
*C. dentalis* 174L  
*C. kelloggae* 191R  
*C. lindsayi* 198L  
*C. loxodon* 199R  
*C. nebrascensis* 190R  
*C. russelli* 177L
- Coriphagus 56, 76L, 79R, 80R
- Cormocyon 212L  
*C. copei* 205R
- Cormohipparion 195L, 203L, 207L, 208L, 334L, 337R  
*C. occidentale* 172L, 177R, 203R, 217L  
*C. emsliei*, 207R, 219L, 267R  
*C. paniense* 197R  
*C. quinmi* 191R, 203L
- Coryphodon 57, 85L, R, 86L, 87L, 88L, 90L, R, 91R, 107R, 112R, 113L, R, 114L, R, 120L, 121L, 129L, 130R, 132R, 134L, 324L
- Coryphodontidae 92L
- Cosomys 246L, 250L  
*C. primus* 246, 255L, 258L, 259R  
*C. sawrockensis* 246, 250L, 254L
- Cosoryx 177R, 197R, 200L, 215L, 217R
- Cranioceras 172L, R, 197R, 217L, R  
*C. teres* 200L
- cranioceratine dromomerycid, 172L, 174L, 332R
- Craseops 117R, 118L
- Craterogale 213L, 333L
- creodont(s) 82L, 92L, 122L, 324R, 327R, 328R, 329L
- Creodonta 57, 62L, R, 106L
- cricetid (s), rodent 187R, 190R, 192L, 198L, R, 214R, 215R, 217R, 218R, 334L
- Cridetidae 239R
- cricetine(s), rodent 175L, 211R, 213R, 217R, 218R, 219L
- cricetodontine murid(s), rodent 218L
- Cromeromys 246, 275R
- Crucimys 192L, 211R
- Crusafontina 218L
- Crypholestes 117R, 123R, 328L
- Cseria 246, 250L  
*C. parkeri* 246, 250L
- Cuyamalagus 173R
- Cupidinimus 175L, 188L, 190R, 191R, 198R, 213L  
*C. avawatzensis* 177L  
*C. nebrascensis* 191R  
*C. tertius* 177L  
*C. whitlocki* 191R
- Cuvieronius 260R, 263L, 280L, R, 281R, 282L, 337R
- cylindrodontid(s), cylindrodontine(s) 324R, 329L, 331L
- Cynarctina 213R
- cynarctine borophagine canid(s) 195L



- Cynarctoides* 176L, 196L, 198R, 199L, R  
*C. lemur* 205R  
*Cynarctus* 176L, 195L, 215L  
*C. crucidens* 217L  
*C. marylandica* 207R  
*C. saxatilis* 215R  
*C. voorhiesi* 217L  
*Cynelos* 208L, 212L  
*Cynomyoides* 190R  
*Cynorca* 204R, 205L, R, 206R, 211R, 215L  
*C. occidentale* 174L  
*C. proterva* 207R  
*C. sociale* 208L  
*Cynodesmus* 188R, 189R, 211L, 330L  
*C. martini* 211L  
*C. thoooides* 166L  
*Cynomys* 279R, 281R, 290R  
*C. gunnisoni* 236, 270R, 285L  
*C. ludovicianus* 236, 270R, 285L, 288L  
*Cyriacotherium* 57, 82R, 83R, 84L, R, 86L, 88L  
  
*Daeodon* 208R  
*D. leidyianum* 208R  
*Daemonelix* 212L  
*Daphoenictis* 120L  
*D. tedfordi* 162R  
*Daphoenocyon* 120L, 161R  
*Daphoenodon* 198R, 204R, 205R, 206L, 211R, 212L  
*D. notionastes* 204L, R, 205R, 206L  
*D. superbis* 204R  
daphoenine amphicyonid(s) 331L  
*Daphoenus* 119R, 329L, 330R  
*Dasyypus* 268R  
*D. bellus* 268L, 282R  
*Delahomeryx* 201L  
*Deltatherium* 57, 70L, 72L, R, 75L, 91R, 324L  
*Deltatheridium* 28R, 30, 33  
dermopteran 120L, 323R  
desmanine mole(s), insectivore 218L  
*Desmatippus* 178R, 191L, 212L, 215L, 332R  
*D. avus* 172L  
*D. crenidens* 189R  
desmatochoerine oreodont(s) 330R  
*Desmatochoerus* 197L, 212L  
*D. leidyi* 191L  
*D. megalodon* 171R, 173L, 179R, 189L, 193R, 196R, 197L  
*Desmatoclaenus* 60, 63L, 67R, 70L, 72L, 73R, 75L, 76L, 79R, 81L, R, 321R  
*Desmatolagus* 132R, 192L, 196R, 213L, 333L  
*D. schizopetrus* 196L  
*Desmocyon* 212L, 213L  
*D. thomsoni* 199L  
*Deuterogonodon* 60, 72L, 75L, 76L  
  
*Diacodexis* 112R, 113L, 114L, R, 120L, 131R, 325L  
*Diacodon* 55, 58, 63L, 69R, 71R, 72L, 74L, 75L, 76L, 79R, 81L, R, 82R, 83R, 84L, R, 86R, 87R, 88L, R, 112R  
diceratheriine rhino(s) 330R, 331L, 332R  
*Diceratherium* 166L, 173L, 179R, 188R, 192L, 193R, 197L, 198R, 203R, 204L, R, 208R, 212L, 330R  
*D. annectens* 204L, R  
*D. armatum* 204R  
*D. matutinum* 208R  
*D. tridactylum* 166L  
dichobunid(s) (artiodactyl) 327R, 328L,  
*Dicrostonyx* 276R, 290R  
*Didelphis* 236, 270R, 282R, 285L  
*D. virginiana* 283R  
*Didelphodon* 25L, 28R, 31, 33, 55, 63L, R, 64L, 65R, 321L  
*D. coyi* 31  
*D. padanicus* 31, 33  
*D. vorax* 31, 33  
*Didelphodus* 112R, 119R, 133L, 325R  
*Didymictis* 57, 71R, 79L, 84L, R, 86L, R, 87L, 88L, 112R, 114L, R, 325R  
*D. vanceleaveae* 114R  
*Dillerlemur* 59, 79L, 84L, R, 86R, 87L, 88L  
*Dilophodon* 114R, 117R, 120R  
*Dinictis* 166L, 179R, 330R  
*D. felina* 163L  
*Dinogale* 213R, 334L  
*Dinohippus* 171L, 178L, 200L, 217L, 219L, 249R, 251L, 259L, R, 260R  
*D. interpolatus* 200R, 218R  
*D. leidyianus* 218R  
*D. mexicanus* 200R, 219L, 252L  
*D. osborni* 175L, 251R  
*Dinohyus* 204R, 211L  
*D. hollandi* 175L  
*D. minimus* 197L  
*Diplacodon* 117L, 118L, 130R  
*Diplobunops* 117R, 118L, 119R, 133R, 134L  
dipodomysine rodent(s) 334L  
*Dipodomys* 235, 244, 250L, 261L, 290R  
*D. gidleyi* 260R  
*D. hibbardi* 265R  
*Dipoides* 218L, 251L  
*D. rexroadensis* 235, 244, 250L, 256R, 266L  
*D. stirtoni* 218L  
*D. williamsi* 200R  
*D. wilsoni* 265L  
*Diprionomys* 191L  
*Dipsalodon* 57, 79L, 84L, 86L, 87R, 88L  
*Dipsaladictides* 57, 86L, 87L, 88L  
*Dissacus* 61, 71R, 74R, 75L, 76L, 81L, R, 82R, 83R, 84L, R, 86L, 87L, 88L, 131R, 324L  
*D. praenuntius* 86L, 88L  
  
Dinocerata 91R, 94R, 117R, 324L, 328R  
dinoceratan(s) 92L  
dinosaur (s) 21R, 24R, 26R, 28L, 35R, 36R, 37L, R, 64R, 93R  
dog 195L  
*Dolichorhinus* 116L, R, 117L, R, 127R  
*Dolomys* 253R, 255L  
*Domnina* 118L, 192R, 193R  
*Domninooides* 190R, 198L  
*Dorraletes* 60, 79R, 82R, 83R, 84L  
*Downsimus* 188R, 192L  
*Draconodus* 59, 72L, 73R  
*Dracontolestes* 59, 72L, 73R  
*Drepanomeryx* 215L  
dromomerycid(s), artiodactyl 172L, 173R, 174L, 175L, 189R, 190R, 192L, 195L, R, 196L, 199R, 207L, 208L, 212L, 213L, R, 215L, 216L, 218R, 332R  
Dromomerycidae 212L, 218R  
*Dromomeryx* 178R, 191L, 195R, 213R, 216L  
*D. borealis* 189R, 190R  
*D. pawniensis* 197R  
*Duchesneodus* 118R, 119L, 125L, 126L, 129R, 133L, R  
*D. uintensis* 124L, 126L  
*D. primitivus* 133L  
*Duchesnehippus* 118R  
*D. intermedius* 118R, 133R  
dugongid(s) 205L  
*Dyseohyus* 201R, 215L  
*Dyseolemur* 117L, 118L, 124L, 328R  
*Dyseotylopus* 174R  
  
*Earendil* 62R  
*Ecclesimus* 165L  
*E. tenuiceps* 165L  
*Ectocion* 44L, 61, 73L, 77R, 80R, 81R, 82R, 84L, R, 85R, 86L, R, 88L, 112R, 113L, 114R, 122L, 131R  
*E. osbornianus* 86L  
*E. parvus* 133R  
*Ectoconus* 61, 62R, 63R, 65L, R, 66L, R, 67R, 321R  
*Ectoganus* 56, 79R, 83R, 84L, R, 86R, 87L, 88L, 112R  
*Ectopocynus* 166L  
*E. antiquus* 166L  
*Ectypodus* 54, 63L, 66R, 67R, 70L, 72L, 74L, 75L, 76L, 78R, 79R, 81R, 82R, 83R, 84L, R, 86L, R, 87L, 88L, 321R  
*E. szalayii* 78L  
*Edaphocyon* 213L, 215L, 333L  
Edentata 91R, 218L, 324L  
*Ekgmowechasala* 211L  
*Elephas* 282R  
*Ellipsodon* 60, 62R, 63L, 67R, 70L, 72L, 74L, 75L, 321R  
*Elomeryx* 192L, 331L  
*E. armatus* 173L

- Elpidophorus* 59, 76L, 79R, 81L, R, 82R, 83L  
*Elphidotarsius* 59, 71R, 76L, 79R, 81L, R, 82R  
*Enhydritherium* 218L  
*Enhydrocyon* 197L, 211L, 291L  
*E. pahinsintewakpa* 196R, 205L, 211L  
entelodont(s), artiodactyl 192L, 195L, 197L, 211L, R, 332R, 334L  
Entelodontidae 213L  
*Entomolestes* 58, 79R, 83L, 84L, R, 86R, 87R, 114R  
entoptychine 205L, 211L, R, 212L, 331L  
*Entoptychus* 179R, 187L, R, 211R  
*E. basilaris* 187R  
*E. fieldsi* 187R  
*E. grandiplanus* 193R  
*E. sheppardi* 187R  
*Eobasileus* 116R, 117L, R, 127R  
*Eocoileus* 219L  
*E. gentryorum*, 207R, 251L  
*Eoconodon* 61, 62R, 63R, 65L, R, 67R, 70L  
*Eodelphis* 23R, 31  
*E. browni* 31  
*E. cutleri* 31  
*Eoemys* 165L  
*Eohaplomys* 117L, 118L, 123R, 328R  
*Eomellivora* 218L  
eomoropine chalicothere(s) 328L  
*Eomoropus* 117R, 127R  
*Eomys* 173R, 213R, 330R, 334L  
eomyid(s), eomyine(s) rodent 118L, 120R, 198L, 205L, 211L, 213R, 215R, 217R, 218L, R, 328L, 329L, 330R, 331R, 334L, 337R  
Eomyidae 116L  
*Eotitanops* 112R, 131R, 134R  
*E. borealis* 114R  
*E. minimus* 114R  
*Eotitanotherium* 118L  
*Eotylopus* 124R, 163R, 329R, 330R  
*E. reedi* 163R  
*Epicyon* 172R, 207L, 217L  
*E. aphobus* 177L  
*E. haydeni* 177L, 200L, R, 218L  
*E. saevus* 217L  
*Epihippus* 116R, 117R, 118L, 120R, 121L, 130L, 133L, R  
*E. gracilis* 129R  
*Epitriplopus* 117R, 118L  
*Eporeodon* 179R, 212L  
*E. thurstoni* 173L  
equid(s), perissodactyl 191R, 196L, 202R, 203L, 208L, 211R, 212L, 213L, 215L, 216L, 217L, R, 218L, R, 219L, 325L, 329L, 333R, 335L, 336L, 337R  
Equidae 192R  
Equinae 213R  
equine(s) 178R, 194R, 202L, 332R, 334L  
Equini 213R  
*Equus*, 178L, 219L, 246R, 249R, 252L, 256R, 259L, 262L, 278L, 280L, R, 281L, 282R, 283L, 284R, 286L, 337R  
*E. bautistensis* 278R  
*E. calobatus* 262R, 280L, 281L  
*E. conversidens* 264R, 282R, 288L  
*E. cumminsii* 262L, R  
*E. francescana* 246L  
*E. giganteus* 260L, 288L  
*E. idahoensis* 246R, 260L, 263R  
*E. niobrarenensis* 288L  
*E. phlegon* 246R  
*E. scotti* 262R, 263L, 271L, 279R, 280L, 281L, R, 282L  
*E. simplicidens* 246R, 249R, 258L, 260L, R, 262L, 263L, R, 264L, 266L, 267L  
*E. (Asinus)* 261R  
*E. (Dolichohippus)* 261R, 263R, 264L  
*E. (Dolichohippus) simplicidens* 249R, 256R, 263L, 267L, 282L  
*E. (Equus)* 259L, 264L, 278L  
*E. (Hemionus)* 260R  
*Eremotherium* 267R  
*E. eomigrans* 267R  
*Erethizon* 259R, 269L  
*E. cascoensis* 259R  
*E. dorsatum* 283R, 288L  
*E. stirtoni* 259L  
erinaceid(s), insectivore 120R, 189R, 196L, 211L, 216L, 217R, 331R, 334R  
erinaceomorph 323R, 325R  
Erinaceidae 217R  
*Ernanodon* 91R  
Ernanodontidae (?Edentata) 91R, 324L  
*Escatepos* 62R  
*Escavadodon* 56, 72L, 76L  
*Essonodon* 28L, R, 29, 33, 36R, 54, 63L, R, 64L, 65R  
*E. browni* 29, 33  
*Esteslestes* 131R  
*E. ensis* 131R  
esthonychid 133R  
*Esthonyx* 57, 79R, 84R, 85R, 86R, 87L, 88L, 112R, 114L, R, 131R, 324L  
*E. ancylion* 86L, R, 87R, 88L  
*E. grangeri* 86R, 88L  
*E. xenicus* 86L, R, 87R  
*Eubelodon* 217L  
*Eucastor* 195L, 215R, 217R  
*E. dividerus* 217L  
*E. planus* 198L, 217L  
*E. tortus* 199R, 215R  
*Euceratherium* 236, 245, 259L, 260L, 270R, 278L, 285L  
*E. collinum* 260L  
*Eucosmodon* 54, 62L, 67R, 70L, 71R, 72L, 73R, 75L, 76L, 79R, 80R  
eucosmodontid(s) (multituberculate) 318R, 321L, 323L  
*Eucyon* 217R  
*E. davisii* 200R, 218L  
*Eudaemonema* 59, 71R, 75L, 76L, 81L, R, 82R  
*Euhapsis* 189L, 212L  
*E. platyceps* 194L  
eumyid rodent(s) 330R  
*Eumys* 166R, 189R, 211L, 330R  
*E. brachyodus* 166L  
*E. elegans* 164R, 331L  
*E. parvidens* 164R  
*Euoplocyon* 176L, 213L, 215L  
euprimate(s) 111R, 112R, 122L  
*Euroxenomys* 190R, 333L  
*Eusmilus* 211L  
*E. cerebralis* 166L, 173L  
Eutheria, eutherian(s) 23L, 24R, 28L, 31, 33, 37R  
eutypomyid rodent(s) 329L  
Eutypomyidae 211R  
*Eutypomys* 119R, 165L, 188R, 192L, 196R, 329L, 330R, 331L  
*E. acares* 119R  
*E. obliquidens* 119R  
*E. magnus* 163L  
*E. thomsoni* 164R  
*Fanimus* 211R  
felid(s), feloid (s), carnivore 195L, 199L, 202L, 217R, 218L, R, 334L, 337L  
Felidae 202R, 213R  
*Felis* 218R, 249R, 260R, 338L  
*F. concolor* 236, 285L  
*F. lacustris* 267L  
*F. rexroadensis* 251L  
florentiamyid 211L, R, 212L  
florentiamyine 213L  
*Florentiamys* 211R  
*Floresomys* 131R  
*F. guanajuatoensis* 131R  
*Floridaceras* 213L, 332R, 333L  
*Floridachoerus* 213L  
floridatragulid(s), artiodactyl 201R  
*Floridatragulus* 213L, 335L  
*Fossodontia* 188R  
*Fossorcastor* 196R, 197L, 212L  
*F. brachyceps* 211L  
*F. fossor* 211L, 212L  
fox 198L  
*Fractinus* 54, 80R  
*Franimys* 55, 86L, 87R, 88R  
*Frictops* 163L  
*F. emryi* 163L  
*Galbreathia* 190R, 196L, 215L

- Gallolestes* 32, 37R  
*G. agujaensis* 32  
*G. pachymandibularis* 32  
*Galushamys* 218R  
*Gelastops* 56, 71L, 75L, 76L, 79R, 81L, R, 82R  
*G. joni* 78L  
 gelocid(s), artiodactyl 200L, 207R, 216L, R  
 Gelocidae 219L  
*Gentilicamelus* 187L, 205L, 211R  
*G. sternbergi* 211R  
 geolabidid(s) 126L  
 Geolabididae 211R  
*Geolabis* 196R  
 geomyid rodent 190R, 191L, 217R, 218L, R  
 geomyoid rodent (s) 179L, 196L, 198L, 203R, 215R, 328R, 330R  
*Geomys* 262L, 262R, 265L, R, 279R, 281R, *G. bursarius* 283L  
*G. minor* 252L  
*G. pinetis* 283L, R  
*G. tobinensis* 266R, 281R, 282L, R  
*G. (Nerterogeomys) minor* 262L  
*G. (Nerterogeomys) persimilis* 261R, 262L  
*G. (Nerterogeomys) paenebursarius* 262R  
*Geringia* 192L, 211R  
*Germanomys* 254R  
*Gigantocamelus* 263L, R, 267L, 337R  
*G. spatula* 260L, 266L  
*Gillisonchus* 62R  
 glasbiid(s) (marsupial) 321L  
*Glasbius* 28L, R, 30, 33, 37L, 55, 63L, R, 64L, 65R, 318R, 321L  
*G. intricatus* 28R, 30, 33  
*G. twitchelli* 28R, 30, 33  
*Glossotherium* 249R, 261R, 264L, 268R  
*G. chapadmalense* 235, 244, 246R, 247L, 250L, 262L, 267R  
*G. garbanii* 264L  
 glyptodont(s) (edentate) 249R, 267L  
*Glyptotherium* 235, 244, 250R, 253L, 261R, 263L, R, 264L, 268R, 285L  
*G. arizonae* 261L, 267R, 271L  
*G. texanum* 263R, 266R  
 gomphothere 202R, 203L, 207L, 218R  
 gomphotheriid(s) 176L, 195L, 197R, 200L, 202R, 203L, 214L, R, 215L, R, 217L, R  
 Gomphotheriidae 215R  
*Gomphotherium* 174L, 197R, 202R, 203L, 214R, 219L  
*G. calvertensis* 207L, R, 208L  
*G. obscurum* 200L  
*G. productum* 200L  
 gondwanathere(s) 92R  
*Goniacodon* 61, 62R, 63L, 69R, 72L, 73R, 76L, 323R  
*Goniodontomys* 218L  
*Grangeria* 117R  
*Grangerimus* 173L  
*Gregorymys* 173L, 189L, 193R, 196R, 197L, 211R  
*G. montanensis* 189L  
*Griphologomys* 189L, 196R, 211L, 212L, 331R  
*Griphomys* 117R, 118L, 119R  
*Guanajuatomys* 132L  
*G. hibbardi* 132L  
*Guildayomys* 250L, 267L, 290R  
 gypsonictopid(s) 318R  
*Gypsonictops* 31, 33  
*G. clemensi* 31  
*G. hypoconus* 28R, 33  
*G. illuminatus* 28R, 33  
*G. lewisi* 31  
*Hadroleptauchenia* 330R  
*Hapalodectes* 114R  
*Haplaletes* 60, 63L, 69R, 72L, 74L, 75L, 76L, 79R, 81L, R, 82R, 83R, 84L, 323R  
*Haploconus*, 61, 62R, 63L, 67R, 70L, 71R, 72L, 75L, 76L, 321R  
*Haplohippus* 119L, 133R  
*Haplolambda* 57, 79R, 83L, 84L, R, 86L, 87L  
*Haplomyilus* 60, 85L, 86L, 87L, 88L, 90L, 113L, 121L, 122L, 132R  
*H. simpsoni* 86R, 88L  
*H. speirianus* 120R, 133R  
*Harpagolestes* 114R, 119R, 129R  
*Harrymys* 189R, 196L  
 hedgehog 190R, 198L, 215R  
*Helaletes* 113R, 114R, 120R, 134R  
 helaletid(s) 325L, 327R  
*Heliscomys* 119R, 165L, 192L, 196R, 205L, 328R, 330R  
*H. mcgrewi* 165L  
*H. vetus* 165L  
 helohyid artiodactyl(s) 328R  
*Helohyus* 114L, 116L, 117R, 122R, 129R  
*Hemiacodon* 114R, 116L, R, 117L, R, 127R, 128R  
*Hemiauchenia* 178L, 198L, 200L, R, 249R, 252L, 256R, 260L, R, 278L, R, 280R  
*H. blancoensis* 260L, 262L, 263L, R, 267L, R  
*H. macrocephala* 268L, 288L  
*Hemicyon* 192R, 199R, 208L, 217R  
*H. ursinus* 199R  
*Hemicyon (Plithocyon)* 176L  
*H. (P.) barstowensis* 176L  
*Hemicyon (Phoberocyon)*, 208L, 213L, 333L  
 Hemicyonidae 213L  
 hemicyonine 214R, 215L, 333L, 334L, R  
*Hemipsalodon* 119R, 133R, 328R  
*H. grandis* 130R  
*Hemithlaeus* 61, 62L, 63R, 65R, 66R, 67R, 69L, 321R  
*Hendryomeryx* 119R, 120R, 129R, 328R, 329R  
*Heptacodon* 120L, 133L, 328R, 329R  
*H. pellionis* 119R  
*Heptodon* 113L, 114R, 122L, 128L, 326L  
 herpetheriine(s) 324R  
*Herpetherium* 116L, R, 165L  
*H. fugax* 165L  
*Hesperhys*, 187L, 208L, 212L  
*H. antiquus* 208R  
*Hesperocamelus* 174L, 176R  
*H. alexandrae* 174L  
*Hesperocyon* 119R, 133L, 166R, 211L, 328R, 329R, 330R  
*H. coloradensis* 165L  
 Hesperocyoninae 215L  
 hesperocyonine canid(s) 195L, 199R, 211L, 330R, 331L  
*Hesperogaulis* 215L  
*Hesperologomys* 192R, 199R, 215R, 217R, 334R  
*Hesperolemur* 115L, 117L  
*Hesperoscalops* 198L, 218R  
*H. mcgrewi* 251L  
*Hessolestes* 119R  
*Heteromeryx* 119R, 329R, 330R  
 heteromyid(s) 190R, 198R, 201R, 215L, 218R  
 heteromyine(s), rodent 189R  
*Heteropliohippus* 334L  
 heterosoricine(s), shrew 188R, 192R, 331L  
*Hexameryx* 219L  
*Hexobelomeryx* 200R, 252L  
*Hexacodus* 113L  
*Hibbardomys* 250L, 290R  
*H. marthae* 267L  
*Hidrotherium* 119L, R  
*Hitonkala* 211R  
*Hipparion* 192R, 215L, 218R, 334L, 337R  
*H. forcei* 172R, 177L, 217R  
*H. plicatile* 203R  
*H. sanfondensis* 199R  
*H. shirleyi* 202R  
*H. tehonense* 172L, 177L, 198L, 217L  
 hipparionine(s), perissodactyl 191R, 202L, R, 219L, 334R, 337R  
 Hipparionini 213R  
*Hippidion* 218L  
*Hippotherium* 335L  
*H. peninsulatum* 246R  
*H. speciosum* 199R  
*Holmesina* 235, 250R, 264R, 268R, 270R, 290R

- Holmesina* (continued)  
*H. floridanus* 267R, 268L, R  
*H. septentrionalis* 282R, 283R  
*Homacodon* 114L  
*H. vagans* 115L  
homacodontine dichobunid(s) 328L, R  
*Homo* 236, 285L  
*H. sapiens* 236, 285L  
*Homogalax* 112R, 122L, 130L, 132R, 325R  
*H. protapirinus* 113L, 132L  
*Homotherium* 235, 250R, 270R, 278R, 282L  
*H. serum* 278R, 283L  
*Hoplictis* 217R  
*Hoplophoneus* 163L, R, 164R, 166L  
*H. dakotensis* 166L  
*H. mentalis* 163L, R  
*H. occidentalis* 164R  
horse(s) 166L, 171L, R, 173R, 174L, R, 175L, 176L, 177L, 178L, R, 187R, 188L, 189R, 190L, R, 193L, 194R, 195L, R, 197L, R, 198L, 199L, R, 200L, 201R, 202L, R, 205L, 207L, 208L, 212L, 213R, 215R, 218L, R, 219L, 246L, 252L, 258L, 260R, 262R, 264L, 265L, 281L, R  
*Huerfanodon* 56, 72L, 75L  
*Hyaenodon* 118R, 129R, 130R, 162R, 163L, 165L, 166L, 174R, 211L, 328R, 329L, 330R  
*H. brevirostris* 166L  
*H. crucians* 165L  
*H. horridus* 166L  
*H. megalodon* 163L  
*H. microdon* 163L  
*H. montanus* 162R  
hyaenid 263R  
hyaenodontid(s) ine(s) 74R, 90L, 106L, 122L, 325L, 327R, 328R  
Hyaenodontidae 89R, 112R  
*Hylomeryx* 117, 118L  
hyopsodontid(s) 132L, 133R, 321R, 323L, R, 324L, R, 329L  
*Hyopsodus* 60, 86L, 88L, 112R, 113L, 114R, 120L, 121L, 122R, 129R, 131R, 325R, 328R  
*H. lepidus* 115L, 128R  
*H. lovei* 120R  
*H. paulus* 120R  
*H. powellianus* 130R  
hypertragulid(s) 211L, 212L, 328L, R, 330R, 331L, 332R, 334L  
*Hypertragulus* 129R, 162L, 164L, R, 179R, 211L, 212L, 330R  
*H. calcaratus* 162L, 164L, R, 173L  
*H. hesperus* 172L  
*H. (Allomeryx) planiceps* 211L  
*Hypisodus* 120L, 211L, 330R  
hypocarnivore (ous), 187L  
*Hypohippus* 178R, 192L, 196L, 199L, 200L, 213L, 217R  
*H. affinus* 207L  
*H. equinus* 189R  
*H. osborni* 172L, 188L, 190R, 192R, 197R  
*H. pertinax* 197R  
*Hypolagus* 173R, 190R, 191L, 199R, 213R, 235, 244, 250R, 256R, 258R, 259R, 266R, 281L  
*H. edensis* 252L, 256R, 260R  
*H. furlongi* 256R, 260L, 265R  
*H. gidleyi* 178L, 256R, 258R, 262R  
*H. limnetus* 258L  
*H. mexicanus* 264L  
*H. regalis* 267L  
*H. ringoldensis* 252L, 260R  
*H. tedfordi* 252L, 260R  
*H. vetus* 198L, 218L, 258L, 264L, 256L  
*Hypsiops* 179R, 211R, 212L  
*H. brachymelis* 211R  
*H. erythroceps* 197L  
*H. latidens* 197L  
hyrachine rhino(s) 328R  
*Hyrachyus* 112L, 113R, 114L, R, 116L, 117L, R, 120R, 122R, 129R, 133R, 134R  
*H. eximius* 127R  
*Hyracodon* 118R, 119R, 211L, 329L  
*H. leidyianus* 166L  
*H. primus* 119R  
hyracodontid, ine, rhino(s) 130R, 325L, 327R, 328L, 329L, 330R  
*Hyracotherium* 112R, 113L, R, 114R, 121L, 129L, 130R, 131R, 132R, 325L  
*Hystriacops* 217R, 218R, 334L  
hystriacomorpha(s), rodent 249R  
*Ibarus* 117R, 118L, 120L  
*Icaronycteris* 58, 86L, 88L  
*Ictidopappus* 57, 70L, 74L, 75L  
*Ignacius* 59, 71R, 75L, 76L, 79R, 81L, R, 82R, 83R, 84L, R, 86R, 87L, 88L, R  
*Ignatiolambda* 57, 79R, 83L, 84L  
*Ilingoceros* 218R  
*Indarctos* 218L  
insectivoran(s) 133R  
insectivore(s) 23R, 126L, 189R, 196L, 218L  
*Ischnognathus* 119L, R  
ischyromyid(s) 128R  
*Ischyrocyon* 176L, 177R, 217R  
*Ischyrognathus* 328R  
ischyromyid, ine rodent(s) 329L, 330R  
*Ischyromys* 119R, 163L, 329L, 330R  
*I. parvidens* 164R, 165L  
*I. typus* 165L  
*I. veterior* 162R  
*Ischyrosmilus* 249R, 268L  
*Ischyrotomus* 117R  
isctolophid(s) 133R, 325L, 328L  
*Janimus* 117R, 118L, 119R, 133L  
*Jaywilsonomys* 119R  
*Jepsenella* 56, 71R, 75L, 76L, 79R, 80R  
*Kalobatippus* 212L, 213L  
*Kansasimys* 218L  
*K. dubius* 198L  
*Kimbetohia* 28R, 30, 33, 54, 63R, 65R, 67R, 70L, 321R  
*K. campi* 28R, 30, 33  
kyptocerine protoceratid(s), artiodactyl 212L  
*Kirkkomys* 211L  
*K. schlaikjeri* 211L  
*Knightomys* 114R  
Kollpaniinae 93R  
*Kraglievichia* 268R  
*Krauseia* 54, 71R, 76L, 79R, 80R  
*K. clemensi* 78L  
*Kyptoceras* 219L  
*K. amatorum* 207R  
kyptoceratine protoceratid(s), artiodactyl 202L, 212L  
*Labidolemur* 78R  
lagomorph(s) 117R, 187R, 190R, 328L  
*Lambdoceras* 231L  
*Lambdaotherium* 107R, 112R, 113L, R, 121R, 122L, 127L, 129L, 131L, 132L  
*Lambertocyon* 60, 79R, 82R, 83R, 84L, R, 86L, 87L  
*Lanthanotherium* 174L, 206R, 215R, 334R  
*Laredochoerus* 118L  
*Laredomys* 118L  
*Lasiopodomys* 246L, 275L, R, 276R  
*L. deceitensis* 246, 284L  
*Leidymys* 166R, 174R, 175L, 188R, 205L  
*L. blacki* 166R  
*L. nematodon* 172R, 179R  
*Leipsanolestes* 58, 86L, 88L, R  
*Lemmiscus* 276L, 280L  
*L. curtatus* 236, 245, 246, 270R, 276L, 279L, R, 280L  
*Lemmus* 235, 244, 276R  
*Lemoynia* 218L  
*Lepoides* 219L, 252L  
*L. lepoides* 178L, 252L  
leporid(s), lagomorph 178L, 191L, 198R, 211L  
leporine 198L  
*Leptacodon* 58, 63L, 69R, 72L, 75L, 76L, 78R, 79R, 81L, R, 82R, 83R, 84L, R, 86R, 88L, R  
leptictid(s) (placental) 92R, 321L, 323R, 329L  
*Leptictis* 83L, 165L, 329L  
*L. haydeni* 165L  
Leptictida 55, 93R  
leptictoid(s) 24R  
Leptarctinae 213L

- Leptarctus* 195L, 202L, 213L, 218R, 333L  
*L. primus* 190R, 197R  
*Leptauchenia* 165R, 166L, 189R, 193R, 196R, 330R  
*L. decora* 165R  
*L. major* 165R  
leptauchenine(s) 173L, 187L, 189R, 197L, 211R, 330R, 332R  
leptochoerid(s), ine(s) 114R, 327R  
*Leptochoerus* 166R, 211L  
*L. emilyae* 165L  
*Leptocyon* 195L, 196R, 199R  
*Leptodontomys* 173R, 192L, R, 218L, 215R, 218R  
leptomerycid(s), artiodactyl 188R, 195R, 200L, 212L, 216L, 329R, 330R, 331L, 334L  
Leptomerycidae 216L  
*Leptomeryx* 119R, 162R, 163L, 164L, R, 192L, 211L, 330R  
“L.” (?*Hendryomeryx*) *defordi* 119R  
*L. evansi* 164L, R  
*L. mammifer* 163L  
*L. speciosus* 163L  
*L. yoderi* 162R  
*Leptonysson* 55, 72L, 75L  
*Leptoreodon* 116L, R, 119R, 124L, 129R, 130L, 133L  
*L. edwardsi* 129R  
*L. major* 129R  
*L. pusillus* 129R  
*Leptotomus* 129R  
leptotraguline(s) 133R, 330R  
*Leptotragulus* 117L, 119R, 132R  
*Lepus* 235, 245, 250R, 259L, 261L, 265R, 270R, 272R, 278L, 283L  
*Lignimus* 191L, R, 192R, 215R  
*L. montis* 191R  
*L. transversus* 191R  
*Limaconyssus* 58, 79R, 81R, 82R, 83R, 84L, R, 87L, 88L  
*Limnocyon* 114R, 117R, 118L, 131R  
limnocyonid(s), limnocyonine(s) 118L, 328R  
*Limnoecus* 190R  
*Liodontia* 178R  
*L. alexandrae* 178R  
*Liotomus* 54, 79R, 80R  
Lipotyphla, lipotyphlan 23R, 37R, 321L, 325R  
*Litaletes* 60, 63L, 69R, 72L, 73R, 74L, 75L, 76L, 79R, 80R  
*L. disjunctus* 78L  
*Litocherus* 58, 71R, 76L, 79R, 81L, R, 82R, 83R, 84L  
*Litolestes* 58, 79R, 82R, 83R, 84L  
*Litomylus* 60, 63L, 67R, 70L, 72L, 74L, 75L, 76L, 79R, 81R, 82R, 83L, 321R  
litoptern(s) 92R  
*Litoyoderimys* 162R  
*L. lustrorum* 162R  
llama, lamini 178L, 198L, 334L  
*Longirostromeryx* 200L, 216L, 217R  
*Lophiohyus* 328L  
*Loupomys* 275R  
*L. monahani* 246, 253L, 275R  
*Loveina* 112R, 113L  
*Loxolophus* 60, 62R, 63L, 66R, 67R, 70L, 71R, 72L, 74L, 75L, 76L, 321R  
*L. pentacus* 67L, 68L  
*L. priscus* 68L  
*Lutra* 219L, 235, 250R, 270R  
*L. canadensis* 235, 250R, 270R  
*Lutravus* 218L, R, 249R  
*Machaeromeryx* 175L, 212L  
machairodont(s) 171L  
*Machairodus* 200R, 218L, R, 219L, 249R, 251R  
*Macrocranion* 112R, 117R, 118L, 325L  
*Macrognomomys* 198L  
*Macrotarsius* 116L, R, 117R, 121L  
*M. siegerti* 121L  
*Mahgarita* 119L, 129R  
*Maiorana* 61, 63R, 65L, R  
*Malaquiferus* 117R, 118L, 129R, 330R  
*Mammacyon* 196R, 211R, 212L  
*M. obtusidens* 205R  
*M. (Temnocyon)* 206L  
*Mammut* 253L, 284L, R, 286L, R, 337R  
*M. americanum* 236, 260L, 270L, 278L, 284L, 285L  
mammutid(s) 172L, 176L, 177R, 178L, R, 189R, 197R, 214L, R  
Mammutidae 189R  
*Mammuthus* 236, 245, 250R, 252R, 259R, 261L, 263L, 264L, R, 267R, 269R, 270L, R, 271L, R, 273L, R, 276R, 278L, 280L, 281L, R, 282L, R, 283L, R, 284L, R, 286R, 290L, R, 293L, 295L  
*M. columbi* 278R  
*M. hayi* 283R  
*M. imperator* 280R, 282L, 288R  
*M. jeffersonii* 288L  
*M. primigenius* 288R  
*Manitsha* 130L  
*Manteoceras* 114R, 115L  
*Marfilomys* 132L  
*M. aewoodi* 132L  
*Marmota* 217R, 236, 270R, 285L  
*M. flaviventris* 236, 270R, 285L  
*M. monax* 236, 270R, 285L  
*M. vetus* 219L  
marmotine(s), rodent 189R, 198L, 331L  
marsupial(s) 23L, R, 24R, 28L, 35L, 45R, 64L, 92R, 93R, 126L, 131R, 318R, 321L, 324R  
Marsupialia 30, 33, 54  
*Martes* 190R, 215L  
*Martinogale* 260R  
mastodon(t)(s), proboscideans 190L, 206R  
*Matthomeryx* 215L  
*Mediochoerus* 213L  
*M. mohavensis* 176L  
*Megabelodon* 217L  
*Megacamelus* 175L, 219L, 337R  
*M. merriami* 251R  
*Megadelphus* 113L, 114L, R  
*M. lundeliusi* 114R  
*Megahippus* 172L, 176L, 197R, 199L, 207L, 215L, 217R  
*M. matthewi* 177R, 200L, 217L  
*M. mckennai* 176L, 200L  
*Megalagus* 165L, 188R, 189R, 192L, 205L, 330R  
*M. brachyodon* 163L  
*Megalictis* 205R, 212L  
*Megalestonyx* 112R, 113L  
megalonychid sloths 175L, 251L  
Megalonychidae 218L  
*Megalonyx* 218R, 249R, 250L, 251L, 263R, 278L, R, 281R, 290L, 337R  
*M. jeffersonii* 236, 245, 284R  
*M. leptostomus* 235, 244, 256R, 258L, 260L, 265R, 266R, 267R  
*M. rohrmanni* 256L  
*M. wheatleyi* 245, 280R, 281R, 283R, 284L  
*Megantereon* 219L, 235, 244, 250R, 252L, 253L, 265L, 338L  
*M. hesperus* 251L  
*Megasmithus* 192R, 215R, 216L, 334R  
*Megatylopus* 200L, R, 249R, 252L, 256R, 260R  
*M. gigas* 218L  
*M. matthewi* 200R  
*Megoreodon* 187R, 188R, 211L  
*M. grandis* 197L, 211L  
*Melaniella* 56, 79R, 82R  
*Meliakrounomys* 163L  
*Meniscoessus* 25L, 29, 33, 54, 63L, R, 64L, R, 65R  
*M. collomensis* 29, 33  
*M. conquistus* 29, 33  
*M. intermedius* 24R, 29  
*M. major* 24R, 29  
*M. robustus* 28R, 29, 33, 35L  
*M. seminoensis* 28R, 29, 33  
*Meniscomys*, 174R, 179R, 187R, 211L  
*Meniscotherium* 61, 86L, 88L, 109R, 112R, 113L, R, 131R  
*Menoceras* 176R, 195L, 199L, 201R, 206L, 208R, 211R, 213L  
*M. arikareense* 206L, 208R  
*M. barbouri* 175L, 176R  
*Menops* 119R

- Mentoclaenodon* 60, 79R, 81R  
 mephitine(s), carnivore 215R  
*Merriamoceros* 176L, 213R, 215L  
*Meryceros* 175R, 200L, 213R  
   *M. joraki* 176L  
 merychippine equid(s) 334L  
*Merychippus* 171R, 202L, 213R, 217R, 334L  
   *M. brevidontus* 172L, 174L, 178R, 207L  
   *M. calamarius* 178L, 200L  
   *M. insignis* 188L, 197R  
 "Merychippus" 191L, 199L, 202R, 202L, 208L, 213L  
   "M." *californicus* 172L, 178L  
   "M." *goorisi* 207L  
   "M." *gunteri* 206R, 213L  
   "M." *intermontanus* 176L, 178L, 190R, 199L  
   "M." *primus* 196L, 206R  
   "M." *sumani* 176L  
*Merychius* 192L, 195R, 197L, 199R, 211R, 212L, 215L, 335L  
   *M. arenarum* 175L, 188L, 191R, 198R, 212L  
   *M. calaminthus* 201L  
   *M. elegans* 175R, 201L, R, 213L  
   *M. minimus* 202L  
   *M. relictus fletcheri* 175R  
   *M. (Metoreodon)* 188L  
*Merycobunodon* 117L, R  
 merycochoerine oreodont(s) 331R  
*Merycochoerus* 187L, 212L, 213L  
   *M. magnus* 191R, 194R  
   *M. matthewi* 194R  
   *M. proprius* 192L, 194R  
 merycodont(s), antilocaprid 177R, 195R, 200L, 202R, 333L  
 Merycodontinae 217R  
 merycodontine(s) 195R, 213R, 215L, 216L, 333R, 337L  
*Merycodus* 173R, 188L, 191L, 196L, 199L, R, 213R  
   *M. agilis* 190R  
   *M. cerroensis* 174L  
   *M. necatus* 190R, 192R  
   *M. nevadensis* 178R  
   *M. sabulonis* 192R  
*Merycoides* 179R, 189R, 211R  
*Merycoidodon* 119R, 120L, 166R, 330R  
   *M. bullatus* 165L  
   *M. culbertsoni* 163R  
   *M. dunagani* 161R  
   *M. major* 166L  
   *M. presidioensis* 162R  
 Merycoidodontidae 217R  
 merycoidontontid, ine 328L, 334L  
*Mesatirhinus* 114R, 115L, 127R  
*Mesocyon* 173L, 174R, 330R  
   *M. brachyops* 173L  
   *M. coryphaeus* 174R  
   *Mesodma* 23R, 29, 33, 54, 63R, 64L, 65L, R, 67R, 70L, 72L, 74L, 75L, 76L, 79R, 81L, R, 82R, 83L  
   *M. formosa* 28R, 29, 33  
   *M. hensleighi* 28R, 29, 33  
   *M. primaeva* 24R, 29  
   *M. senecta* 29  
   *M. thompsoni* 29, 33  
*Mesogaule* 189R, 213L  
*Mesohippus* 119R, 160R, 211R, 329L, 330R  
   *M. bairdi* 163R, 165L  
   *M. barbouri* 164R, 165L  
   *M. exoletus* 163R, 165L  
   *M. planidens* 192L  
   *M. westoni* 163L, R, 165L  
*Mesomeryx* 117L, R  
*Mesonychia* 91L, 324L  
 mesonychid(s) 61, 74R, 90R, 325R, 329L  
*Mesonyx* 114R, 117R, 118L, 120R  
*Mesoreodon* 166L, 174R, 187R, 189R, 205L, 211L  
   *M. minor* 166L, 196R, 211L  
*Mesoscalops* 189R, 191L, 196L  
*Metadjidaumo* 166L  
   *M. hendryi* 166L  
*Metamynodon* 129R, 330R  
*Metanoiamys* 117R  
   *M. korthi* 119R  
   *M. lacus* 119R  
*Metarhinus* 116R, 117L, R  
 metatherian 318R  
*Metatomarctus* 175L, 196L, 199L, 208L, 213L, R  
   *M. canavus* 176R, 199L, 208L  
*Metechinus* 211L, 217R  
 miacid, miacoid 133R, 134L, 324R, 325R, 328L  
*Miacis* 112R, 114R, 325R  
*Michenia* 175R, 192L, 199L, 200L, 211R, 212L, 218R  
   *M. agatense* 175L, 188L, 198R  
*Microclaenodon* 61, 72L, 75L, 76L  
*Microcosmodon* 54, 63L, 67R, 79L, 72L, 73L, 74L, 75L, 76L, 79R, 81L, R, 82R, 83R, 84L, R, 86L, 87L, R, 321R  
*Microeutypomys* 118L, 119R, 133L  
   *M. tilliei* 119R  
*Micromomys* 58, 79R, 82R, 84L, R, 86R, 88L, R  
*Microparamys* 55, 86L, 87R, 88R, 116L, 119R, 124L, 129R  
   *M. nimius* 1194  
*Microsus* 113L, 114L  
 microsypid primate(s) 328R, 329L  
*Microsypops* 112R, 113L, 114R, 117R, 118L, 120R, 129R, 130R  
   *M. annectens* 115L, 116L, R  
   *M. elegans* 115L  
 Microsypoidea 82L  
 Microtinae 239R  
 microtine(s), rodent 239R  
*Microtus* 235, 245, 246, 250R, 259L, R, 271R, 272L, 273L, 274R, 278L, R, 279L, R, 280L, 281L, 282L, R, 290R  
   *M. aratai* 246, 283R  
   *M. californicus* 274R, 278L  
   *M. deceitensis* 246, 275R  
   *M. guildayi* 246, 282L, R  
   *M. llanensis* 236, 246, 270L, 273R, 274L, 281R, 282L, R, 284L, R  
   *M. meadensis* 236, 245, 246, 259R, 270L, 273R, 275L, R, 276R, 278R, 279L, R, 281L, R, 282R, 289L  
   *M. mexicanus* 276L  
   *M. montanus* 276L  
   *M. ochrogaster* 274L  
   *M. paroperarius* 236, 245, 250R, 270L, 273R, 274L, 275R, 276R, 278R, 279L, R, 280L, 281L, R, 282L, R, 284L, R, 294L  
   *M. pennsylvanicus* 274L, 276L, 290L, 290R  
   *M. pliocoenicus* 246, 266L  
*Microtomarctus* 197R, 199R, 213L  
   *M. conferta* 176L, 178L, 199L  
*Mictomys* 253L, 255R, 261R, 273R, 290L  
   *M. borealis* 278R  
   *M. kansasensis* 235, 244, 246, 250R, 259R, 266L, R, 267R, 273R, 275L, 276L, 278R, 279R, 280L, 281L  
   *M. landesi* 256R, 265R  
   *M. meltoni* 235, 244, 246, 250R, 273R, 276L, 278R, 279L, R, 280L, 281L, R, 282L, R  
   *M. vetus* 235, 244, 250L, 256R, 258R, 261R, 266L  
*Mimatuta* 61, 62R, 63L, R, 64L, R, 65L, R, 67R, 321L, R  
*Mimetodon* 54, 71R, 75R, 76L, 79R, 81L, R, 82R, 83R, 84L  
*Mimomys* 246L, 250L, 253R, 254L, 275L, R, 280L, 295L  
   *M. dakotaensis* 246, 267L, 276L, 280L  
   *M. monahani* 246, 253L, 275R  
   *M. panacaensis* 250L, 252L, 254L, 260R  
   *M. virginianus* 246, 267R, 275L, R, 276L, 279L, 284L  
   *M. (Cosomys) primus* 246  
   *M. (Cosomys) sawrockensis* 246, 250L, 254L  
   *M. (Cromeromys) dakotensis* 246  
   *M. (Cromeromys) virginianus* 246  
   *M. (Ogmodontomys) poaphagus* 246  
   *M. (Ophiomys)* 246  
   *M. (Ophiomys) mcknighti* 250L  
*Mimoperadectes* 54, 86L, 88L

- Mimotricentes* 60, 63L, 67R, 70L, 74L, 75L, R, 76L, 78L, 79R, 81R, 82R, 83R, 84L, R, 321R  
*M. subtrigonus* 78L  
 miniochoerine oreodont(s) 330R, 331L  
*Miniochoerus* 163L, R, 164L, R, 165L, 166L, 330R  
*M. affinis* 164R  
*M. chadronensis* 163R, 164L  
*M. douglasensis* 164L  
*M. forsythae* 163L  
*M. gracilis* 164R  
*M. starkensis* 165L, 166L  
*Mingotherium* 55, 79R, 83R  
 mioclaenid(s) 62R, 93R, 321R, 323L  
 Mioclaenidae 60, 62R  
*Mioclaenus* 60, 72L, 73R, 76L  
*Miocyon* 119R, 133L  
*Mioheteromys* 217R  
 miohippine equid(s) 329L, 330R  
*Miohippus* 163L, 164L, 166L, 173L, 179R, 187L, R, 197L, 204L, 205L, 212L, 330R  
*M. annectens* 166L  
*M. equinanus* 166L, 197L  
*M. gidleyi* 166L  
*M. grandis* 163L, R, 164L  
*M. intermedius* 166L  
 miolabine camel(s), artiodactyl 212L, 217R, 332R  
*Miolabis* 172L, 200L, 202R, 212L  
*M. fricki* 174L  
*M. montanus* 190R  
*Miomastodon* 172L  
*Miomustela* 213L, 333L  
*M. madisonae* 190L  
*Mionictis* 213R, 217R, 334L  
*Miopetaurista* 207R, 219L  
*Miospermophilus* 173R, 192L, 196L, 19L  
*Miotapirus* 204R, 212L  
*Miotylopus* 173L, 174R, 197L, 211L, 212L  
*M. gibbi* 173L  
*Miracinonyx*, 282L  
*M. inexpectatus* 235, 244, 250R, 267L, 270L, 284L  
*M. trumani* 236, 284R  
*Mithrandir* 61, 62R, 63R, 66R, 67R, 70L, 321R  
*Mixodectes* 59, 72L, 75L, R, 76L, 324L  
*M. pungens* 72R, 74L, 75L  
 mixodectid(s) 59, 75R  
*Mojavemys* 191L, 215L  
 mole(s), insectivore 196L, 198L, 218R  
*Molinodus* 93R  
*Monosaulax* 171R, 193R, 195R, 215L, 216L  
*M. pansus* 171R, 199R  
*M. skinneri* 171R  
 monotreme(s) 92R  
*Montanatylopus* 120L, 330R  
*Mookomys* 173R, 189L, 190R  
*Moropus* 179R, 187L, 204R, 205R, 206L, 211R, 216L  
*M. elatus* 204R  
*M. merriami* 178R  
*M. oregonensis* 204R, 205R  
 moschid(s), artiodactyl 188L, 190R, 199R, 200L, 212L, 215L, 216L  
 Moschidae 212R, 217R  
 Multituberculata, multituberculates 23L, R, 24R, 25L, 29, 33, 35L, 36R, 37R, 54, 64L, 66L, 75L, 92R, 318R, 321L, R, 323L, 324L  
 murid(s), muroid(s) rodent 211L, 218R, 218L, 337R  
 Muridae 239R  
*Mustela* 219L  
*M. erminea* 236, 270R, 285L  
*M. rexroadensis* 258L  
 mustelid(s), carnivore 190R, 195L, 207L, 211L, 212L, 213L, 215L, 217R, 218L, R, 219L, 331R, 332R, 333L, 334L, R, 337L  
 Mustelinae 213L  
 musteline(s) 213R  
 musteloid(s) 205L  
 mylagaulid(s), rodent 189R, 190R, 196L, 212L, 213L, 251L, 331L  
 Mylagaulidae 211L, 218R  
 mylagauline(s), rodent 190R, 194R, 211R, 215L, 216L  
*Mylagaulodon* 187L, 192L, 212L  
*Mylagaulus* 199R, 206R, 215R  
*Myiodon* 282R  
 mylodont sloth 246R, 249R  
 Mylodontidae 218K  
*Mylohyus* 219L, 251L, 284L  
*M. elmorei* 251L  
*M. nasutus* 284L  
*Myrmecoboides* 55, 71R, 75L, 79R, 81L, R, 82R, 83R, 84L,  
*Myrmecophaga* 280R  
*tridactyla* 280R  
*Mysops* 113R, 114R, 116L, 117R, 129R  
*Mytonomeryx* 117L, R, 118L  
*Mytonomys* 119R, 129R, 134R  
*Mytonolagus* 117R, 118L  
*Nannasfiber* 212L  
*Nannippus* 218L, 249R, 252L, 261L, R, 262L, 263L, 267L, 268L, 334L  
*N. aztecus* 198L, 218R, 219L  
*N. lenticularis* 218R  
*N. minor* 251L  
*N. montezuma* 263R  
*N. peninsulatus* 235, 244, 246R, 249R, 261R, 262L, 263L, R, 264L, 265L, 266L, 267L, R  
*N. phlegon* 246R  
*Nannodectes* 59, 79R, 80R, 81R, 82R, 83R, 84L  
*N. gazini* 81R  
*N. gidleyi* 83R  
*N. intermedius* 77R, 78L, 80R  
*N. simpsoni* 82R  
*Nanodelphys* 165L  
*N. hunti* 165L  
*Nanotragulus* 166R, 171R, 174R, 197L, 204R, 211R, 212L, 330R  
*N. loomisi* 166R, 205L, R, 211R  
*N. ordinatus* 205L, R, 211R  
*Navahoceros* 259L, 278L  
*Navajovius* 58, 80R, 81R, 82R, 83R, 84L  
*Neotacastor* 204R, 205L, R, 208L  
*N. hesperus* 189L  
*Nebraskomys* 250L, 253L, 254R, 255L, 262L, 265L  
*N. mcgrewi* 253R, 265R, 266L  
*N. rexroadensis* 253R  
 nectogaline shrew(s), insectivore 218L  
*Nekrolagus* 235, 249R, 250R, 256R, 260R  
*N. progressus* 252L  
*Neochoerus* 261R, 268R  
*N. cordobai* 264L  
*N. dichroplax* 267R  
*N. holmesi* 264L  
*Neoclaenodon* 60, 72R, 76L, 79R, 81L  
*Neofiber* 235, 250R, 270R  
*N. allenii* 283R  
*N. leonardi* 274L, 276L, 283L, R  
*Neohipparion* 195L, 215R, 217R, 219L, 249R, 252L, 334L, 337R  
*N. affine* 217L  
*N. coloradense* 199R, 215R  
*N. eurystyle* 200R, 218R, 219L  
*N. leptode* 218L  
*N. phosphorum* 251L  
*N. repulicanus* 191R  
*N. trampasense* 217R  
*Neoliotomus* 54, 81R, 84L, R, 86R, 87L, 88L, R, 113L  
 neomustelid carnivore 334L  
 Neoplagiulacidae, neoplagialacid(s) 23L, R, 29, 33, 318R  
*Neoplagiaulax* 30, 33, 54, 63R, 65R, 67R, 70L, 72L, 74L, 75L, R, 76L, 81L, R, 82R, 83R, 84L, 321R  
*N. burgessi* 28R, 33  
*Neotoma* 198R, 259R, 278L, 290R  
*N. fuscipes* 278L  
*N. quadriplicata* 262L  
*N. taylori* 265R  
*N. (Hodomys)* 289R  
*N. (Paraneotoma)* 198R, 259R, 261R  
*N. (Paraneotoma) fossilis* 259R, 261R  
*Neotragocerus* 218L  
*Nexuotapirus* 196R, 204L, 211R, 213L  
*N. marslandensis* 204R  
*N. robustus* 179R

- Niglarodon* 187R, 188R, 211L  
*N. blacki* 191L  
 nimravid(s) 173L, 196R, 205R, 216R,  
 217R, 330R  
 Nimravidae 120L, 211R, 217L, 218R, 325,  
 328R, 329R  
*Nimravides* 218R  
*N. catocopis* 218L  
*N. galiani* 171L, 217R  
*N. thinobates* 171L, 172R, 217R  
*Nimravus* 166L, 179R, 196R, 211R, 330R  
*N. brachyops* 166L, 173L  
*Niptomomys* 58, 86L, 87R, 88R, 112R  
 nontherian (mammal) 92R  
 notharctine(s) 324R, 327R, 328R  
*Notharctus* 113L, 114R, 115L, R, 116L, R,  
 117L, R, 122R, 127R, 128R, 129R, 131L  
*N. pugnax* 115L, R  
*N. robinsioni* 114R  
*N. robustior* 127R  
*N. tenebrosus* 115L, 132R  
*Nothokemas* 204R, 205L  
*N. waldropi* 205L, R  
*Nothotylopus* 199L, 213R, 217R  
*Nothrotheriops*, 235, 244, 250R, 259L,  
 270R, 278R, 280R, 281L, 285L, 286R  
*N. shastensis* 282L  
*N. texanus* 283R  
*Notiotitanops* 134L  
*N. mississippiensis* 134L  
*Notolagus* 200R, 252L  
*Notoparamys* 112R  
 Notoungulata 93R, 94R  
 notoungulate(s) 92R  
 nyctitheriid(s) 58, 82L, 126L, 133R, 324L  
*Nyctitherium* 83L, 114R, 116R, 119R,  
 129R, 133L  
  
*Ocajila* 118R, 193L, 211L  
*Ochotona* 218R, 284R  
 ochotonid(s) 173R, 176R, 190R, 192R,  
 196L, 198L, 211L, 213L, 215R, 216L,  
 217R, 331R, 334R  
 Ochotonidae 211L  
*Odocoileus* 249R, 260L, 261R, 265L,  
 280R, 288L, 290R  
*Ogmodontomys* 235, 244, 246L, 250L,  
 253R, 254L, R, 255R, 262R, 265R,  
 290L, R  
*O. poaphagus* 244, 246, 253R, 255L,  
 262L, R, 265L, 266L, 267L  
*O. sawrockensis* 244, 246, 250L, 251R,  
 254L, 265L  
*Oklahomalagus* 198L  
*Oligoryctes* 116R, 117R  
*Oligospermophilus* 165L  
 omomyid(s), omomyine(s), omomy-  
 oid(s) 92L, 324R, 327R, 328L, 328R,  
 329L  
  
*Omomys* 112R, 114L, 116L, R, 119R, 120R,  
 128R, 129R  
*O. carteri* 114R  
*Ondatra* 253L, 255R, 260R, 279L, 282R,  
 290L, R  
*O. annectens* 235, 245, 246, 250R,  
 266R, 267R, 270R, 273R, 274L,  
 275L, 279R, 281R, 282L, R, 284L, R  
*O. hiatidens* 282R  
*O. idahoensis* 235, 244, 246, 250L,  
 256L, 258R, 259R, 260L, 261L, R,  
 265R, 266L, 268L  
*O. nebracensis* 246, 274L  
*O. zibethicus* 236, 245, 246, 255R,  
 270R, 274L, 285L, 288L  
*Onhippidion* 218R  
*O. galushai* 218R  
*Onychodectes* 56, 63R, 67R, 70L,  
 321R  
*Onychomys* 218L  
*Ophiomys* 235, 244, 246L, 249R, 250L,  
 253L, R, 254L, R, 255R, 256R, 258R,  
 259R, 290L, R, 295L  
*O. fricki* 266L  
*O. magilli* 255L, R, 260R, 266L  
*O. mcknighti* 250L, 254L, 255L, 256R,  
 260R  
*O. meadensis* 253R, 255R, 265R  
*O. parvus* 256L, 258R, 266L, 275L,  
 278R  
*O. taylori* 253R, 255L, R, 256R, 258L,  
 261R  
*Oreamnos* 236, 270R, 285L  
*O. harringtoni* 270R  
 oreodont(s), artiodactyl 162L, 164L, R,  
 165L, 166L, 171L, 176L, 178R, 187L, R,  
 188L, 189L, R, 190L, 191R, 193L, 195L,  
 R, 196L, R, 197L, 199L, R, 200L, 201R,  
 203R, 205L, 206L, 211L, R, 212L, 213R,  
 215L, 216L, 217L, 328R, 329L, R, 330R,  
 331L, 332R, 334L, 335L  
 oreodontine oreodont 329R  
*Oreodontoides* 196R, 212L  
*O. oregonensis* 179R, 197L, 211L, R  
*Oregonomys* 191R  
*O. agrarius* 191R  
*Oreolagus* 190R, 191L, 192L, 199L, 213L,  
 333L  
*O. nebrascensis* 196L  
*O. nevadensis* 178R, 190R  
*Orientalophus* 91R  
*Orohippus* 113L, R, 114R, 115L, 120R,  
 122R, 128R, 133R  
*O. major* 115L  
*O. sylvaticus* 115L  
 oromerycid(s) 328L, 329R, 330R  
*Oromeryx* 116R, 117L, R, 118L, 130L  
*Oropycitis* 166L  
*O. pediasius* 166L  
  
*Orthogeomys* 268L  
*O. propineta* 268L  
*Osbornoceras* 218L, R, 249R  
*Osbornodon* 199R, 205L, 215L  
*Osteoborus* 249R, 251L, 265L  
*O. diabloensis* 171L, 177L  
*Otarocyon* 330R  
*Otionohyus* 330R  
*Ourayia* 116L, R, 117L, R, 118L, 127R, 129R  
*Ovis* 285L  
*O. canadensis* 236, 285L, 288L  
*Oxetocyon* 166L  
*O. cuspidatus* 166L  
*Oxyacodon* 61, 62R, 63R, 66R, 67R, 70L,  
 321R  
*O. priscilla* 67L  
*Oxyaena* 57, 79R, 84L, R, 85R, R, 86R,  
 87L, 88L, R, 90R, 112R, 113R, 131R  
 oxyaenid(s) 117R, 324L, 327R, 328R,  
 329L  
*Oxyaenodon* 117L, 118L  
 oxyclaenid 321R, 323L, 324R  
 Oxyclaenidae 60, 62R  
*Oxyclaenus* 60, 62L, R, 63L, 65R, 67L,  
 70L, 72L, 73R, 321R  
*O. antiquus* 68L  
 oxydactyline camel(s), artiodactyl 212L  
*Oxydactylus* 198R, 202R, 205L, 212L,  
 213R  
*O. longipes* 188L, 195R  
*Oxyprimus* 60, 63L, R, 64L, R, 65L,  
 80L, R, 321L, R  
*Oxytomodon* 60, 72L, 73R  
  
*Pachyaena* 112R, 325R  
*Paciculus* 173L, 187R, 188R, 211R  
*P. woodi* 211L  
*Palaeonodon* 56, 86L, 87L, 88L, R  
 palaeonodont(s) 132L  
*Palaeochthon* 59, 72L, 73R, 75L, R, 76L,  
 78L, 79R, 81L, R, 82R  
*P. alticuspis* 78L  
*P. nacimienti* 78L  
*Palaeictops* 55, 63L, 69R, 72L, 74L, 75L,  
 76, 79R, 81L, R, 82R, 83R, 84L, R,  
 86R, 87R, 88L, R, 112R, 122R, 133L,  
 323R, 328L  
*Palaeolama* 286R  
*P. mirifica* 268L  
*Paenemarmota* 200R, 218R, 235, 244,  
 250R, 252L, 264L, 265L  
*P. barboursi* 263R  
*P. sawrockensis* 219L  
*Palaeoarctomys* 190L  
*Palaeocaster* 187R, 196R, 204L, 211L, R,  
 212R  
*P. nebrascensis* 166R, 211L  
*Palaeogale* 205L, R  
 Palaeolaginae 211R



- Palaeolagus* 120L, 173L, 179R, 187R, 188R, 193R, 196R, 204L, 211R  
*P. burkei* 165L, 166L  
*P. hypsodus* 166R, 189R, 211L  
*P. intermedius* 164L, R, 165L  
*P. primus* 162R  
*P. temnodon* 163L, R  
*Palaeolama* 286R  
*P. mirificus* 268L  
*Palaeonictis* 57, 86R, 88L, 325R  
*Palaeoryctes* 55, 72L, 76L, 79R, 81L, R, 82R, 83R, 84L, R, 86L, 87L, 88L, 92R  
palaoryctid(s) 92L, R  
*Palaeosinopa* 56, 78L, R, 79R, 81R, 82R, 83R, 84L, 86R, 87L, 88L, R, 114L, 325R  
*Palaeosyops* 112R, 113L, R, 114L, R, 120R, 121R, 122R, 134R  
*P. fontinalis* 114L  
*Palenochtha* 59, 72L, 75L, 76L, 79R, 80R  
*P. minor* 78L  
paleomustelid(s) 332R  
*Paleotomus* 56, 72L, 75L, 76L, 78L, 79R, 81L, R, 82R, 84L  
*P. milleri* 78L  
*Pandemonium* 58, 63R, 67R, 70L, 321R  
*Panolax* 199R  
*P. santaefidei* 199R  
*Panthera* 278R, 290R  
*P. atrox* 236, 284R, 288L  
*P. onca* 236, 270R, 283R, 284L, 285L  
pantodont(s) 82L, 83L, 89L, 90R, 92L, R, 93R, 131R, 323R, 324R, 327R  
Pantodonta 57, 85L, 91L, 93R, 94L, 324L  
*Pantolambda* 57, 70L, 72L, R, 75L, 76L, 324L  
*P. intermedium* 93R  
pantolambdid(s) 131R  
*Pantolestes* 113L, 114L, R, 117R, 134L, 328L  
pantolestid, pantolestan 133R, 318R, 323R, 324R  
*Pappogeomys* 264R  
*Parablastomeryx* 192L, 212L, 217R  
*P. mollis* 178R  
*Paracimexomys* 23L, 29, 33  
*P. priscus* 28R, 29, 33  
*P. magnus*, 29  
*Paracosoryx* 177L, R, 196L, 212L  
*P. alticornis* 192R  
*P. furcatus* 190R  
*P. furlongi* 172L, 174L, 177L  
*P. loxoceras* 178L  
*Paractyptotis* 256R, 258L  
*P. gidlevi* 258L  
*P. rex* 256R  
*Paracynarctus* 196L, 199L, R  
*P. kelloggi* 171R, 178L, R, 208L  
*Paradjidaumo* 132R, 165L, 166R, 330R  
*P. validus* 164R  
parahippine 207L  
*Parahippus* 173R, 178R, 188L, 191L, 192L, 197L, 212L, 215L, 332R  
*P. leonensis* 208L  
*P. pawniensis* 175L, 187L  
*P. tyleri* 191R, 199L  
*Parahyus* 129R  
*Parailurus* 256R  
*Paralabis* 166R, 330R  
*P. cedrensis* 166L  
*Parallomys* 192L, 193R, 211L  
*Paramerycoidodon* 330R  
*Paramerychys* 197L, 212L  
*P. harrisonensis* 197L  
*Paramicroscoptes* 218L  
*Paramiolabis* 174L, 199L, 200L, 213R, 215L  
*P. singularis* 175R, 188L  
*P. taylori* 164L  
*P. tenuis* 175R  
*Paramoceros* 176L, 200L  
paramyid(s) 130R  
*Paramylodon* 249R, 263L, R, 250R, 267L, 286R, 278R  
*P. harlani* 235, 245, 246R, 247L, 250R, 268L, 270R, 278L, R, 280L, 282L, R, 285L  
*Paramys* 55, 86L, 87L, 88L, R, 112R, 114R, 117R, 324L  
*P. delicatior* 115L  
*P. delicatus* 115L  
*Paranasua* 217R  
*Paraneotoma* 256R, 259R, 261R  
*Paranotolagus* 264L  
*P. complicatus* 264L  
*Paranycetoides* 23R, 28R, 32, 33, 37R  
*P. maleficus* 32  
*P. megakeros* 32  
*P. sternbergi* 32  
*Parapliohippus* 173R, 213R, 334L  
*P. carrizoensis* 173R, 175R  
*Parapliosacomys* 176L  
*Paratomarctus* 176L, 192R, 195L, 199R, 215L, 217R  
*P. euthos* 217L  
*P. temerarius* 177R, 190R  
*Paratylopus* 166R, 330R  
*P. labiatus* 165L  
*P. primaevus* 166L  
*Pararyctes* 55, 72L, 75R, 76L, 79R, 81L, R, 82R, 83L  
*Paratoceras* 203R, 217R  
*Paractypodus* 28R, 30, 33, 54, 65R, 67R, 72L, 74L, 75L, R, 76L, 79R, 81R, 82R, 83R, 84L, R, 86R, 87L, 88L, R, 321R  
*P. foxi* 28R, 30, 33  
*Parenhydrocyon* 197L, 330R  
*P. wallovianus* 201L  
*Pareumys* 117R, 119R  
*Parictis* 325, 328R  
*Parisectolophus* 114R  
*Paroligobunis* 205R  
*P. frazieri* 205R  
paromomyid 111L, 323R  
*Paromomys* 59, 72L, 73L, R, 75L, R, 76L, 79R, 81L, R, 82R  
*P. depressidens* 78L  
*Paronychomys* 177L, 218L  
*Paroreodon* 179R, 211R, 212L  
*Parvericius* 188R, 192L, 205L, R, 211L, 216L  
*Parvitragulus* 162R  
*P. priscus* 162R  
*Patriofelis* 114L, R, 133R  
*Patriolestes* 117L  
*Patriomanis* 162R, 325, 329R  
*P. americanus* 325, 329R  
pauromomyid(s) 324L  
*Pauromys* 113L, R, 114R, 117R, 129R, 328L  
*P. lillegraveni* 123R  
peccary, 187L, 201R, 208L, 211R, 213L, 215L, 291R  
*Pediomeryx* 175L, 218R  
*P. hemphillensis* 251R, 259R  
Pediomyidae, pediomyid(s) 23L, 24R, 31, 33, 318R, 321L  
“*Pediomys*” 31, 33, 321L  
“*P.*” *clemensi* 31  
“*P.*” *cooki* 28R, 31, 33  
“*P.*” *elegans* 28R, 31, 33, 35L  
“*P.*” *fasseti* 31  
“*P.*” *florencae* 28R, 31, 33  
“*P.*” *hatcheri* 28R, 31, 33  
“*P.*” *krejicii* 28R, 31, 33  
“*P.*” *prokrejicii* 31  
*Pediomys* 55, 63L, R, 64R, 65R  
*Pedomys* 246, 273R, 274L, 282R, 284R  
*P. llanensis* 276L, 284R  
*Pelycodus* 112R  
*Pelycomys* 165L, 330R  
*P. brulanus* 164R  
*P. placidus* 165L  
*Penetrigonias* 120L  
*Pentacemylus* 117R, 118L  
*Pentacodon* 56, 72L, 75L, 76R, 79R, 80R  
*Pentacosmodon* 54, 79R, 84L  
*Peraceras* 195R, 199R, 200L, 207L, 213R, 217R, 334L  
*P. hessei* 207L  
*P. superciliosum* 178L, 190L, R  
peraceratheriine rhino(s) 328L  
*Peradectes* 45R, 53L, R, 54, 63L, 64L, 65L, R, 67R, 70L, 74L, 75L, 76L, 79R, 81L, R, 82R, 83R, 84L, R, 86R, 88R, 321L

- Peratherium* 114R, 129R, 132R  
*Perchoerus* 179R, 188R, 211L  
*Peridiomys* 188L, 190R, R, 191L, 196L, 215L  
 periptychid 62L, 71L, R, 321R, 323R  
 Periptychidae 61, 62R  
*Periptychus* 61, 62R, 63L, 65R, 66R, 67R, 70L, R, 71R, 72L, 73R, 74L, 75L, 76L, 79R, 81L, R, 83L, 321R, 324L  
*P. carinidens* 62L, 63R, 67R, 71L, R, 72L, R, 73L  
*P. gilmorei* 71L  
*Perissodactyla* 89R, 91R, 112R, 325L  
 perissodactyl(s) 90L, 122L, 127R, 132L, 133R, 324R  
*Perognathus* 190R, 198L, R, 215L  
*P. furlongi* 174L  
*P. gidleyi* 261R, 265R  
*P. mclaughlini* 252L  
*Peromyscus* 198R, 218R, 260R, 280L  
*P. cragini* 265R, 282R  
*P. hagermanensis* 261R  
*P. polionotus* 268L  
*P. valensis* 269R  
*Petauristodon* 213R, 217R, 334L  
*P. uphami* 174L  
*Pewelagus* 259L  
*P. dawsonae* 235, 250L, 252L, 260R  
*Phaiomys* 246L  
*Phelosaccomys* 190R, 217R  
 phenacocoeline(s) 332R  
*Phenacocoelus* 179R, 205L, R, 212L  
*P. typus* 194L, 206L  
*P. luskensis* 206L  
 phenacodontid(s) 60, 82L, 86L, R, 323R  
*Phenacodus* 44L, 60, 72L, 73L, 76L, 78R, 79R, 81L, R, 82R, 83R, 84L, R, 86L, R, 87R, 88L, R, 112L, 113L, R, 114R, 121R, 122R, 128R, 129L, 131R, 132R  
*P. grangeri* 78, 83R  
*P. primaevus* 86L  
*P. vortmani* 86L  
*Phenacodaptes* 60, 79R, 83L, R, 84L, R, 86L  
*Phenacolemur* 59, 83L, 84L, R, 86R, 87L, R, 88L, R, 113L, 129L  
*P. pagei* 86R, 87L  
*P. praecox* 86R, 88L  
 phenacolemurine(s) 324R  
 Phenacolphidae 91R, 324L  
*Phenacomys* 235, 245, 250R, 258R, 270R, 272R, 273L, R, 275L, 276L, R, 278R, 279L  
*P. gryci* 258R, 278R, 279L, 280L  
*P. intermedius* 281R  
*Pitheciastes* 330R  
*Phlaocyon* 205L, 213R  
*P. achoros* 205R  
*P. annectens* 201L  
*P. leucosteus* 205R  
*P. minor* 204L  
 Phlaocyonini 213R  
*Phoberocyon* 208L, 213L, 333L  
*Phosphatherium* 92R  
*Picrodus* 59, 72L, 75L, 76L, 79R, 81L, R, 82R, 83R, 84L  
*Pipestoneomys* 156L, 331L  
 pitymyine species 246L  
*Pitymys* 246, 266L, 273R, 275L, R, 282R, 283R, 289L  
 placental(s) 94L  
 plagiaulacidan(s) 23L, R  
*Plagioctenodon* 58, 86L, 87L, 88R  
*Plagiomene* 59, 86L, 87R, 88R  
*Planetetherium* 59, 86R, 87L, R  
*Planisorex* 266L  
*P. dixonensis* 266L  
*Platybelodon* 217R, 218R  
*Platygonus* 218R, 249R, 259R, 260R, 264R, 285L, 290L  
*P. alemanii*, 285L  
*P. bicalcaratus* 235, 250L, 262L, 263L, R, 265L, 266R, 267L, R, 268L, R  
*P. compressus* 236, 245, 284R  
*P. cumberlandensis* 283R  
*P. pearcei* 235, 244, 250L, 256R, 258L, 260L  
*P. vetus* 235, 245, 250R, 268L, 270L, 281R, 282L, R, 283L, R, 284L  
*Platymastus* 60, 63R, 66R, 67R, 321R  
 plesiadapid(s) 58, 71L, 75R, 77R, 78L, 82L, 89L, 323R, 324L  
*Plesiadapis* 59, 71L, 73L, 76L, 78L, R, 79R, 80R, 81R, 82R, 83R, 84L, R, 85R, 86R, 87R, 88L, 90L, 113L  
*P. anceps* 44L, 80L, 81L, R  
*P. churchilli* 80L, 82L, 82R, 83R  
*P. cookei* 80L, 84L, 85R, 86L, R, 87L, R, 88L, 120L  
*P. dubius* 84R, 85R, 120L  
*P. fodinatus* 84L  
*P. gingerichi* 80L, 83R, 84L, R, 86R, 87L  
*P. praecursor* 44L, 72L, 73L, 75L, 77R, 78L, 80L, R  
*P. rex* 80L, 81L, 82L, R  
*P. simonsi* 80L, 82R, 83R, 84L  
*P. tricuspiciens* 89R  
*Plesictis* 196R  
*Plesippus* 246R, 249R, 258L, R, 260L  
*P. francescana* 246R, 259R, 278L  
*P. idahoensis* 246R, 259R  
*P. shoshoniensis* 258L  
*P. simplicidens* 246R  
*Plesiogulo* 200R, 218R, 249R, 251L  
*Plesiosorex* 213L, 333L  
 Plesiosoricidae 213L  
*Plesiolestes* 59, 72L, 73R, 75L, 76L, 79R, 80R  
*Plesiosminthus* 193L, 196L, 209R  
 pleurolicine 187L  
*Pleurolicus* 179R, 188R, 189L, 196R, 199L, 211R  
*P. sulcifrons* 211L  
*Plioceros* 200L, R, 127R, 218R, 334L  
*P. floblairi* 216L  
*Pliotomys* 253L, 255R  
*P. rinkerii* 255R  
*Pliocyon* 213R, 215L  
*P. ossifragus* 190L  
*Pliogale* 215R, 218R, 334R  
*Pliogeomys* 198L, 200R, 218R, 265L  
*P. parvus* 252L, 260R  
 pliohippine equid(s) 334L  
*Pliohippus* 200L, 215R, 218R, 249R, 252L, 264L, 334L  
*P. leardi* 171L, R, 172R, 177L  
*P. mirabilis* 215R  
*P. nobilis* 218L  
*P. pernix* 200L, 217L  
*P. spectans* 218R  
*P. tantalus* 177L  
*P. tehonensis* 172L, 177L  
*Pliolemmus* 235, 244, 250L, 253L, R, 255R, 266L  
*P. antiquus* 253R, 255L  
*Pliometanastes* 200R, 218L, 337R  
*Plionarctos* 218R  
*P. edensis* 218R  
*Plionictis* 190R, 213R, 334L  
*P. oaxacaensis* 202R  
*Pliophenacomys* 250L, R, 254R, 265R, 266L, 267L, 290R  
*P. finneyi* 253R, 255L, 265L  
*P. osborni* 250R, 256L, 266L, 281L  
*P. parkeri* 246, 251L  
*P. primaevus* 253R, 255L  
*P. wilsoni* 252L  
*Pliopotamys* 235, 244, 250L, 253L, R, 255L, R, 260R, 261R, 266L  
*P. meadensis* 246, 253R, 255L, 266L  
*P. minor* 253R, 255L, 256R, 258L  
*Pliosaccomys* 198L, 218L  
*Pliotaxidea* 218L  
*Pliotomodon* 218L  
*Pliozapus* 218R  
*Plithocyon* 175R, 176L, 214R, 215L, 334R  
*Poabromylus* 118R, 130R, 329L, 330R  
*Poebrodon* 117R, 118L  
 poebrodonline camelid(s) 330R  
*Poebrotherium* 120L, 330R  
*P. eximium* 163R, 164L  
*P. franki* 163R  
*Pontifactor* 58, 86L, 87R, 88R, 114R  
*Potamotherium* 213L, 333L  
*Praeovibos* 285R  
*P. priscus* 285R  
*P. recticornis* 285R  
*Pratilepus* 235, 250L, 258L, 264L

- P. kansansensis* 235, 250L, 264L  
*P. vagus* 258L  
*Premnoides* 59, 72L, 76L  
*Preptoceras* 282R  
*Presbymys* 119L  
*Princetonia* 60, 79R, 84L, R, 86R, 87R, 88L, R  
 primate(s) 58, 64L, 71L, 75R, 78R, 79R, 82R, 90L, 106L, 111R, 112R, 116R, 126L, 133L, 321R, 323L, R, 324L, R, 325L, 237R, 328L, R, 329L  
*Priscocamelus* 201L, 212L  
*Pristinictis* 57, 79R, 80R  
*Proantilocapra* 217R  
*Probarbouromeryx* 213L  
*Probassariscus* 213L  
*Probathyopsis* 78R  
*Problastomeryx* 212L, 215L  
*Proboscidea* 171R, 172L, 175L, 176L, 177R, 190L, 195L, 197R, 214R, 215L  
*proboscidean(s)* 92L, 189R, 195L, 200L, 203L, 206R, 214L, R, 215L, R, 219L, 252L, 334R, 337R  
*Procadurcodon* 133R  
*Procamelus* 176L, 195R, 200L, 215L, 218R, 334L  
*P. grandis* 203R  
*P. lacustris* 190R  
*Procaprolagus* 117R, 118L, 133L  
*Procastoroides* 244, 249R, 250L, 253L, 256R, 258L, 265R, 266L  
*P. idahoensis* 253L, 266L  
*P. sweeti* 253L  
*Procerberus* 56, 63L, R, 64L, 65R, 67R, 70L  
*Prochetodon* 54, 79R, 82R, 83R, 84L, R, 86L, 87R, 88L  
*Procranioceras* 192R, 216L  
*P. pawniensis* 197R  
*P. skinneri* 192R, 207L  
*Procreodi* 327R  
*Procoelius* 175L  
*Procynodontis* 117L, 118L, 121L  
*Procyon* 218R  
*procyonid(s), ine(s), carnivora* 215L, 331R, 333L, 334L  
*Procyonidae* 213L  
*Prodaphoenus* 117L, 118L  
*Prodesmatochoerus* 330R  
*Prodiacodon* 55, 63L, 69R, 72L, 74L, 75L, 76L, 79R, 81L, R, 82R, 83R, 84L, R, 86R, 87R, 88L, R, 321L, R, 323R  
*P. puercensis* 78L  
*Prodinoceras* 59, 78R, 84L, R, 86L, R, 87R, 88L, R, 90R  
*Prodipodomys* 198L R, 200R, 218R, 235, 244, 250R, 252L, 259R, 265R  
*P. centralis* 266L, R  
*P. minor* 252L  
*P. tiheni* 252L  
*Progeomys* 198R
- Prohesperocyon* 330R  
*Proheteromys* 174L 192L, 196L, 198R, 204R, 205L, R, 206R, 330R  
*P. sulcatus* 173R  
*P. magnus* 173R, 205R, 208L  
*P. maximus* 174L  
*Prolapsus* 117R, 129R  
*Prolimnocyon* 74R, 112R, 114R, 131R  
*Promartes* 196R, 198R, 199L, 211L  
*Promerycochoerus* 171R, 179R, 187L, 189R, 192L, 211R, 212L  
*P. carrikeri* 194L, 197L  
*P. hesperus* 173L  
*P. superbus* 191L, 197L  
*P. (Parapromerycochoerus) barbouri* 189L  
*Promesoreodon* 330R  
*prometheomyine murid(s), rodent* 218L  
*Promioclænus* 60, 63R, 66R, 67R, 70L, 72L, 74L, 75L, R, 79R, 81L, R, 82R, 321R  
*Promylagaulinae* 211L, 215L  
*promylaguline(s), rodent* 187R, 190R, 211R, 334L  
*Promylagaulus* 211L  
*Proneofiber* 275L, 282L  
*P. guildayi* 275L, 282L  
*Pronodens* 188R, 212L  
*Pronothodectes* 58, 75L, 76L, 79R, 80R  
*Pronotolagus* 198L, 215R  
*Propalaeonodon* 56, 79R, 84L  
*Propalaeosinopa* 72L, 75L, R, 76L, 79R, 81R, 82R, 83R, 84L  
*Proscalopidae* 189R, 211R  
*Proscalops* 196R, 211R  
*Prosciurus* 165L, 330R  
*P. magnus* 165L  
*P. vestustus* 163L  
*Prosigmodon* 198R, 200R, 218R  
*Prosomys* 218R  
*Prosthennops* 200R, 215L, 249R, 252L  
*P. niobrarenensis* 208L  
*P. serus* 203R, 218R  
*P. xiphodonticus* 207R  
*Prosynthetoceras* 195R, 204L, 206L, 208R, 211R, 216L, 335L  
*P. orthrionanus* 204R  
*P. texanus* 204R  
*Protadjidaumo* 117R, 118L, 132R  
*P. pauli* 119R  
*Protalphadon* 28R, 321L  
*P. foxi* 28R  
*P. lulli* 28R  
*Protapirus* 133R, 166R, 211L  
*P. obliquidens* 166L  
*Protentomodon* 56, 83L, 86R, 87L  
*Protepcyon* 215L  
*P. raki* 176L  
*Proterixoides* 117R, 118L, 119R, 123R, 124L
- proteutherian* 328L  
*Prothryptacodon* 60, 72L, 75L, 79R, 80R  
*Protictis* 57, 72L, 75L, 76L, 79R, 81L, 82R, 83R, 84L  
*Protictops* 119L  
*Protitanotherium* 117L, 118L  
*Protoceras* 165R, 166R, 330R  
*P. celer* 166L  
*protoceratid(s) ine(s)* 130R, 187L, 195R, 202L, R, 207L, R, 212L, 213L, 217L, R, 328L, 329R, 330R, 331L, 332R, 337R  
*Protoceratidae* 218R  
*protohippine(s), perissodactyl* 202R, 334L  
*Protohippus* 190L, 195L, 199L, 213R, 218R, 334L, 337R  
*P. gidleyi* 217R  
*P. perditus* 207L  
*P. supremus* 207L, 217L  
*Protolabinae* 218R  
*protolabine camel(s), artiodactyl* 212L, 332R  
*Protolabis* 173R, 175R, 176R, 188L, 195R, 196L, 199L, 200L, 202L, R, 212L, 218R  
*P. barstowensis* 199L  
*P. heterodontus* 203R  
*Protomarctus* 178R, 196L  
*P. optatus* 171R, 173R, 175R, 199L  
*Propliophenacomys* 219L, 250R  
*P. parkeri* 246, 251L  
*Protoprocyon* 217R  
*Protoptychus* 117L, R  
*Protoreodon* 116R, 117R, 119R, 127R, 129R, 130R, 133R  
*P. pacificus* 119R  
*P. petersoni* 129R  
*P. pumilus* 121L, 129R  
*Protosciurus* 165L, 171R, 205, 231L, 331L  
*Protoselene* 60, 62R, 63L, 68L, 69R, 71R, 72L, 74L, 75L, 76L, 79R, 81L, R, 82R  
*P. opisthacus* 71R, 72R, 73L, 74L  
*Protospermophilus* 189R, 192L, 196L, 204R, 206R, 217R  
*Prototomarctus* 172L, 196L, 213L, R  
*Prototomus* 112R, 325  
*Protungulatum* 31, 35R, 36L, 44L, 53L, R, 59, 61R, 62L, 63L, R, 64L, R, 65L, 67R, 321L, R  
*P. donnae* 21R, 28L, 36L  
*Protylopus* 116R, 117R, 119R, 124L, 127R  
*P. pearsonensis* 119R  
*Proviverra* 118L  
*Psalidocyon* 199R, 215L  
*Pseudadjidaumo* 191L  
*Pseudaelurus* 199L, 202L, 213R, 252L, 334L

- Pseudhipparion* 215R, 334L, 337R  
*P. gratum* 198L, 200L, 217L  
*P. retrusum* 217L  
*P. simpsoni* 207R, 219L  
*P. skinneri* 217R  
*Pseudoblastomeryx* 192L, 202L  
*P. advena* 175L  
*Pseudoceras* 200L, 203R, 207R, 216R, 217L, 219L  
*Pseudocyclopius* 330R  
*Pseudocylindrodon* 117R, 132R, 163R, 329L  
*Pseudocyon* 215R, 217R, 334R  
*Pseudodesmatochoerus* 212L  
pseudolabine camelid(s) 330R  
*Pseudolabis* 212L, 330R  
*P. dakotensis* 166L  
*Pseudopalaeocastor* 197L, 204L, 212L  
*P. barbouri* 204L, 212L  
*Pseudoparablastomeryx* 192L, 202L, 216L  
*Pseudoprotoceras* 119R, 120L, 133L  
*P. longinarius* 163L  
*P. semicinctus* 162R  
*P. taylori* 163R  
*Pseudotomus* 130L  
*P. timmys* 119R  
*Pseudotheridomys* 173R, 175L, 192L, 193R, 196R, 211L, 217R  
*Pseudotrimylus* 188R  
*Psittacotherium* 56, 72L, 75L, 76L, 79R, 81L, R, 82R, 83R, 84L  
*Pterodon* 328R  
*Pterogaulus* 215L  
ptilodontid(s) (multituberculate), 318R, 321L  
*Ptilodus* 54, 63L, 66R, 67R, 70L, 71R, 72L, 73R, 74L, 75R, 76L, 79R, 81L, R, 82L, R, 83R, 84L, 321R  
*Purgatorius* 58, 63L, R, 64L, 65R, 67R, 70L, 83L, 321R  
  
rabbit(s), lagomorph 198L, 211L, 213R, 328R, 330R, 333L, 334R  
*Ragnarok* 62R  
*Rakomeryx* 174L, 176L, 199R, 206R, 213R, 215L  
*R. kinseyi* 188L, 189R, 190R  
*R. raki* 174L  
*Rakomylus* 199L, R, 215L  
*Ramoceros* 174L, 176R, 195R, 200L, 215L, 216L, 334L  
*Rangifer* 270R  
*R. tarrandus* 236, 270R, 285L, 299L  
*Rapamys* 118L, 119R  
*Raphictis* 57, 79R, 82R, 83L  
*Ravenictis* 55, 63R, 70L, 323R  
*R. krausei* 91L  
*Reithrodontomys* 279R  
*R. humulis* 268L  
  
*R. rexroadensis* 261R  
*R. wetmorei* 268L  
*Reithroparamys* 55, 86L, 87R, 88R, 117R  
*Repomys* 175L, 219L, 252L  
*R. gustelyi* 219L, 251R  
*R. maxumi* 175L  
*R. panacaensis* 252L, 260R, 263L  
rhino, rhinoceros 172L, 175L, 176L, 178L, 189L, 190L, R, 192R, 193L, 195L, 198L, 199L, R, 200L, 201R, 202L, 203L, 207L, 212L, 213R, 217R, 218L, R, 250L, 251L, 252L, 260R, 267L, 325L, 327R, 328L, R, 329L, 330R, 331L, 332R, 333L, 334L, 337R  
rhinocerotid 211R  
*Rhynchotherium* 203R, 218L, 235, 250R, 263R, 264L, 337R  
*R. blicki* 203R  
*R. falconeri* 263R, 264L  
*R. praecursor* 266R  
rodent 44L, 85R, 87L, 113L, 120R, 126L, 128R, 130R, 131R, 133R, 134R, 171L, 174L, 179L, 187L, R, 190L, R, 191L, R, 192L, 193L, 194R, 198L, 199L, 201R, 203R, 205R, 209L, R, 211L, R, 212L, 213L, 215L, 218L, 232L, 239R, 246, 247L, 248, 251R, 252L, 253L, R, 254L, 261R, 263L, R, 265L, 267R, 273R, 274L, 276L, 278R, 279L, R, 281L, 282R, 284R, 290R, 293L, 295R, 324R, 327R, 328L, R, 329L, R, 330R, 331L, 332R, 334L, R, 338L  
Rodentia 44L, 55, 78R, 80L, 84L, R, 85L, R, 87L, 89L, 90L, 91R, 92L, 324L  
*Rooneyia* 119L  
*Russellagus* 192R, 199R, 215R, 217R, 334R  
  
*Saiga* 285R  
*S. tatarica* 285R  
*Sanctimus* 211R  
*Satherium* 235, 250R, 253L  
*S. ingens* 260L  
*S. piscinaria* 256R, 258L  
*Saxonella* 58, 79R  
*Scalopoides* 211L  
*Scalopus* 218R, 262R  
*S. (Hesperoscalopus)* 218R, 262R  
*Scapanus* 278R  
*Scenopagus* 114R, 116L, 117R, 129R, 129R, 328L  
*Schaubeumys* 191L, 192L  
*Schizodontomys* 175L, 179R, 187L, 192L, 196L  
*Schochia* 56, 63R, 66R, 67R, 321R  
Sciuravidae 112R  
sciuravid(s) 131R, 327R, 328R, 331L  
*Sciuravus* 113L, 114R, 116R, 117R, 118L, 120R  
  
sciurid(s), ine(s) 177L, 190L, R, 191L, 217R, 330R, 334L  
Sciuridae 120L, 328R  
*Sciurus* 325, 250R, 270R  
*Scottimus* 166R, 192L  
*S. lophatus* 166L  
*S. viduus* 163R  
selenodont artiodactyls 120R, 328L, R  
Semantorinae 213L  
*Sespedectes* 117R, 118L, 119R, 123R, 124L, 133L  
*S. singularis* 124R  
sespedectine(s) 328L  
*Sespemys* 173L  
*S. thurstoni* 173L  
*Sespia* 173L, 174R, 189R, 192L, 196R, 211L  
*S. nitida* 211L  
*Shoshonius* 113L, 114L  
*Shunkahetanka* 211L  
*S. geringensis* 211L  
*Sigmodon* 259L, 262L, 282R  
*S. bakeri* 283R  
*S. curtisi* 235, 250L, 267L, 268L, 280R, 282L, R  
*S. hudsouthensis* 263R  
*S. libitinus* 283R  
*S. medius* 249R, 261R, 266L, R, 267L, R  
*S. minor* 235, 249R, 250L, 259R, 261R, 262L, 265R, 266L, R, 267L, R  
sigmodontine(s), rodent 201R, 217R, 218R  
*Simiacritomys* 119R  
*S. whistleri* 119R  
*Simidectes* 118L, 119R, 129R  
*S. merriami* 119R  
*Similosciurus* 201L  
*Simimeryx* 117R, 118R, 119R  
*S. hudsoni* 119R  
simimyyid rodent 328R  
*Simimys* 117R, 118L, 119R, 123R, 124L, R, 328R  
*S. landeri* 119R  
*Simocyon* 218L  
*Simpsonictis* 57, 72L, 75L, R, 76L, 79R, 81R  
*Sinclairiella* 119R, 133L  
*S. dakotensis* 162R  
*Sinclairiomeryx* 213R  
*Sinopa* 114L  
*S. rapax* 115L  
*Sinostylops* 91R  
skunk(s), carnivore 198L, 219L  
sloth(s), edentate 175L, 200R, 201R, 218L, R  
*Smilodectes* 113L, 114L, R, 115R  
*S. gracilis* 115L  
*S. mcgrewi* 115L  
*Smilodon* 259L, 263L, 272R, 278L, R, 280L, 284L, 286R, 290R

- S. californicus* 288L  
*S. fatalis* 274L  
*S. gracilis* 235, 244, 267R, 268L, 270L, 274L, 283R, 284L  
*S. populator* 236, 245, 270R, 274L, 284L, 285L, 288L  
*Sminthosinus* 219L  
*Soergelia* 272R, 282R  
*S. mayfieldi* 272R  
*Sorex* 260R, 278L, 290R  
*S. meltoni* 260R  
*S. powersi* 256R  
soricid(s) 117R, 190R, 328L, 331L, 333L, 337R  
Soricinae 213L  
soricomorph, 32, 33, 37R, 318R, 323R  
spalacotheriid(s) 23L, R  
*Sparactolambda* 78R  
*Spermophilus* 190R, 191L, 198L, 252L, 279R  
*howelli* 260L  
*tridecemlineatus* 279R  
*wellingtonensis* 260L  
*S. (Buiscitellus) dotti* 198L  
*S. (Otospermophilus) primitivus* 190L  
*Sphenocoelus* 117L  
*Sphenophalos* 218R, 251L  
squirrel(s), rodent 173R, 196L, 198L, 207R, 213R, 219L  
Stagodontidae, stagodontid(s) 23L, R, 24R, 25L, 28R, 31, 33, 318R, 321L  
*Stegomastodon* 236, 244, 249R, 250R, 253L, 258R, 262L, 265L, R, 266L, R, 267L, 273R, 280L, 281R, 282L, 337R  
*S. mirificus* 258R, 262L, R  
stegomastodont 267L  
*Stelocyon* 61, 72L, 76L  
*Stenoechinus* 189L  
Stenomylinae 215L  
stenomyline(s), artiodactyl 199L, 211L, 331R, 334L  
*Stenomylus* 198R, 199L, 201L, 211R, 213L  
*Stenopsoschoerus* 330R  
*Sthenodectes* 129R  
*Sthenictis* 213R, 218R, 334L  
*Stibarus* 120L  
*S. obtusilobus* 166L  
*S. quadricuspis* 165L  
*Stilpnodon* 55, 72L, 75L, 76L, 79R, 81L, R, 82R  
*Stockia* 117L, R, 328L  
*Stygimys* 29, 54, 63R, 64L, 65R, 67R, 70L, 72L, 74L, 75L, 321R  
*S. cupressus* 29  
*S. kuszmauli* 37R  
*Stylinodon* 116L, 122R, 129R  
*Subantilocapra* 219L  
*Subdesmatochoerus* 330R  
*Subdromomeryx* 213R, 215L  
*S. antelopinus* 189R  
*Subhyracodon* 119R, 174R, 330R  
*S. occidentalis* 165L, 173L  
*Submeryceros* 195R, 213R  
*Submerycochoerus*, 212L  
*Swaindelphys* 54, 72L, 76L  
*Sylvilagus* 235, 244, 250R, 259L, 261L, 270L, 272R, 282R, 285L  
*S. hibbardi* 282R  
*S. palustris* 236, 270R, 283R  
*S. webbi* 250L, 268L  
symmetrodonts 23R  
*Synaptomys* 235, 245, 250R, 268R, 270R, 283R, 284L  
*S. cooperi* 276L  
*S. kansasensis* 246  
*S. meltoni* 246  
*S. (Pliotomys)* 253L  
*Syndyoceras* 194R, 212L  
*S. cooki* 202L  
*Synthetoceras* 207L, 217L  
synthetocerine protoceratid(s) 331R, 332R  
*Tachylagus* 119R  
*T. gawneae* 119R  
taeniodont 74R, 117R, 321R, 324R, 328R  
Taeniodonta, 56, 62R  
taeniolabidid 68R  
*Taeniolabis* 54, 62R, 63R, 65R, 66L, 67L, 68L, R, 69L, R, 70L, 323L  
*T. lamberti* 68R, 69L  
*T. taoensis* 65L, R, 67L, 68L, 69L  
Taligrada 62L, R  
*Talpavus* 119R, 133L  
talpid(s), (insectivore) 190R, 198L, 211L  
Talpinae 193L  
*Tamias* 190R  
*Tanupolama* 249R, 263R, 267L, 278L  
*T. stevensi* 288L  
*Tanymykter* 212L, 213L  
*T. brevidontus* 175L  
tapir 211R, 212L, 263R  
tapirid 331L  
tapiroid(s) 117R, 132L, 325L, 327R, 328L  
*Tapiravus* 208L, R  
*T. validus* 208L, R  
*Tapirus* 261R, 280R, 284L, 286R  
*T. copei* 267L, 282L  
*T. haysii* 267R, 268L, 282L, 283R, 284L  
*T. veroensis* 283R  
*Tapochoerus* 117R, 118L  
*Tapocyon* 117R, 118L, 129R  
*Tapomys* 117R, 118L  
*Tardontia* 191L, R  
*Tarka* 120R  
*Taxidea* 200R, 218R, 252L, 260R  
*T. taxus* 278R  
*Taxymys* 1145  
tayassuid, ine 207R, 212L, 219L, 328R  
Tayassuidae 120L, 328R  
*Teilhardina* 112R, 113L  
*Telacodon* 28R, 31, 33  
*T. laevis* 28R, 31, 33  
*Teleobunomastodon* 281R  
*Teleoceras* 175L, 178L, 195L, R, 198R, 199R, 200L, 200R, 203L, 207L, 213R, 250L, 252L, 334L, 337R  
*T. fossiger* 218L  
Teleoceratinae 213L, 332L, 333L  
*Telmatherium* 115L, 131L, 133R  
*Temnocyon* 173L, 206L, 212L  
*T. altigenis* 173L  
*Tenudomys* 165L, 330R  
*T. basilaris* 165L  
*Tephrocyon* 215L  
*Terricola* 246L, 259R, 273R, 275R, 289L  
*T. meadensis* 246, 259R  
*Tethyopsis* 115L, 125L, 127R  
*Tetoniuss* 112R, 132R  
*Tetraclaenodon* 60, 72L, R, 73R, 75L, R, 76L, 78L, R, 80R, 324L  
*T. puericensis* 78L  
*Tetrameryx* 282L  
*T. irvingtonensis* 236, 270L, 278L, R  
*T. schuleri* 283L  
*Texoceros* 218R  
*Texodon* 117R  
*Texomys* 204R, 205R, 206L  
*T. stewarti* 203R  
*Thangorodrim* 62R  
*Thelysia* 56, 86R, 87R  
theria (ian) 36R, 66R  
*Thinobadistes* 218L, 337R  
*Thinocyon* 114L  
*Thinohyus* 165L  
*T. lentus* 165L  
*Thisbemys* 116L, 117R, 129R  
*Thomomys* 219L, 251R, 259R  
*T. carsonensis* 260L  
*T. (Plesiothomomys)* 219L  
*Thryptacodon* 60, 72L, 75L, 77R, 78R, 79R, 79R, 80R, 81R, 82R, 83R, 84L, R, 86L, R, 87R, 88L, R, 114R  
*Thylacaelurus* 59L, 79R, 80R, 81R, 82R, 83R, 84R, 86R, 87R, 88L, R, 117R, 118L, 132R  
ticholeptine oreodont(s) 332R, 334L  
*Ticholeptus* 178R, 192R, 195R, 197R, 206R, 213R, 216L  
*T. zygomatikus* 173R, 188L, 189R, 190R  
tillodont(s) 66R, 67R, 85L, R, 90L, 132R, 133R, 327R  
Tillodontia 57, 91L, R, 324L, 327R  
*Tillodon* 115L  
*Tillomys* 113R, 114R, 120R  
*Tinimomys* 58, 86L, 87L, 88L, R  
*Tinuviel* 61, 63R, 65R, 67R, 70L, 321R  
*Titanoides* 57, 78R, 79R, 80R, 81R, 82R, 83R, 84L, R, 86L, 87L  
titanothera 160R

- Titanotheriomys* 330R  
*Titanotherium* 160R  
*Titanotylopus* 281R, 337R  
*Tiuclaenus* 93R  
*Tiznatzinia* 60, 63R, 67R, 70R, 321R  
 todralestid(s) 92L  
*Tomarctus* 192R, 199R, 215L  
   *T. brevirostris* 199R  
   *T. hippophaga* 174L, 176L, 199R  
*Toromeryx* 117R, 118L, 130L  
*Toxotherium* 119R, 129R, 163R, 329L  
 tragulid(s), artiodactyl 200L  
*Tregomys* 198L, 215R, 217R  
*Tribosphenomys* 91R  
*Tricentes* 60, 62R  
*Trigenicus* 119R, 133L  
   *T. profectus* 162R  
*Trigonias* 119R  
*Trigoniectis* 219L, 235, 249R, 250R, 253L, 265L  
   *T. cookii* 256R, 258L  
   *T. macrodon* 258L, 267R, 268L  
 trigonostylopoids 92R  
*Triisodon* 61, 73R, 75L  
 triisodontid 323R  
 Triisodontidae 61, 62R  
 Triconodontidae 23L, R  
*Trilaccogaulus* 187R, 193R, 211L  
 triplopodine rhinos 328L  
*Triplopus* 116L, R, 119R, 127R, 129R  
 trogonotheriine beaver(s) 213L  
*Trogolemur* 119R  
   *T. leonardi* 119R  
*Trogomys* 175L, 176R  
*Trogosus* 112R, 114L, R, 115L, 120R, 122R, 132R  
*Turgidodon* 24R, 30, 33, 54, 63L, R, 64L, 65R, 321L  
   *T. lillegraveni* 30  
   *T. madseni* 30  
   *T. parapraesagus* 30  
   *T. petaminis* 28R, 30, 33  
   *T. praesagus* 30  
   *T. rhiaster* 28R, 30  
   *T. russelli* 30, 33  
*Tylocephalonyx* 196L, 208L  
 tylopod 328L  
*Tytthaena* 57, 79R, 82R  
  
*Uintaceras* 116R, 117R, 121L  
*Uintacyon* 57, 86L, 87R, 88R, 129R  
   *U. scotti* 129R  
*Uintanius* 114L, R  
 uintathere(s) 89L, 328L, 329L  
*Uintatherium* 114R, 115L, 116L, 123L, 127R  
*Uintasorex* 114R, 116L, 119R, 128R, 129R, 133L  
*Umbogaulus* 215L, 216L  
 ungulate(s) 36L, 53L, R, 62L, 63R, 64L, R, 66R, 67L, R, 68L, 69L, 321L, R, 332L, 337L, R, 338L  
 ungulatomorph(s) 28L, 318R, 324L  
*Untermannerix* 198L, 215L, 217R, 334L  
*Unuchinia* 56, 72L, 75L, 76L, 79R, 81L, R, 82R, 83R, 84L  
*Urocyon* 265R  
*Ursavus* 192R, 213L, 215R, 333L  
   *U. pawniensis* 215R  
 ursid(s) 175L, 212L, 213L, 215R, 217R, 218R, 328R, 333L, 334L  
 Ursidae 213L, 328R  
*Ursus* 244, 252L, 253L, 270L, 285L  
   *U. abstrusus* 178L, 235, 250L, 252L, 258L, 260L  
*Ustatochoerus* 172L, 195R, 197R, 211R, 212L, 217R  
   *U. californicus* 172R, 217L  
   *U. leptoscelos* 201L  
   *U. major* 200L, 217L  
   *U. medius* 177R, 197R, 200L, 216L  
   *U. profectus* 177L  
*Utahia* 114L, R  
*Utemylus* 60, 79R, 83R  
  
*Viejadjidaumo* 119L  
*Viridomys* 54, 63R, 65R, 67R, 70L, 72L, 73R  
 viverravid carnivore(s) 328R, 329L  
*Viverravus* 57, 70R, 84L, R, 86R, 87R, 88L, R, 112R, 117R, 120R, 132L  
*Vulpavus* 112R, 114R, 134L  
   *V. australis* 125R  
*Vulpes* 198L  
   *V. stenognathus* 200R, 218L  
   *V. velox* 236, 285L  
 Vulpini 218L  
  
*Wallia* 117R  
*Wasatchia* 112R  
 washakiine omomyid primate(s) 328L  
*Washakius* 113, 114L, 116R, 117L, R, 120R, 122R, 328L  
   *W. insignis* 115L  
*Wilsonium* 165L  
   *W. planidens* 165L  
*Worlandia* 59, 86L, 87R, 88R  
*Wortmania* 56, 62R, 63R, 67R, 70L, 321R  
*Wyolestes* 131R  
*Wyonycteris* 58, 86R, 87L, 88L  
  
*Xanocromys* 54, 72L, 75L  
*Xenacodon* 55, 79R, 83  
 xenarthrans 92R  
*Xenicohippus* 111L, 112R  
*Xenocranium* 163R  
*Xenocyon* 285R  
   *X. lycanoides* 285R  
 xenungulates 92R  
*Xyromomys* 54, 63L, 67R, 72L, 75L, 321R  
  
*Yatkolamys* 175L  
*Yoderimys* 119R, 162R, 329L  
   *Y. stewarti* 162R  
 Yoderomyinae 211R  
 yoderomyine(s), rodent 193R  
*Yumaceras* 218R  
  
*Zanycteris* 59, 79R, 82R, 83L  
 zapodid(s), rodent 191L, 193L, 198L, 209R, 215R, 216L, 218R, 337R  
*Zapus* 290R  
   *Z. sandersi* 266L, 281R  
*Zemiodontomys* 163L  
   *Z. burkei* 163L  
*Ziamys* 198R  
*Zodiolestes* 212L  
*Zygodolophodon* 172L, 178R, 190L, 197R, 207L, 214R, R, 215L  
   *Z. brevidens* 189R, 190L  
   *Z. merriami* 178R, 214R  
   *Z. proavus* 192R  
 zygodolophodont 178L

## Subject Index

- A. ferox-Pantolambda cavirictum* Interval Zone 17L
- AB Tuff, John Day Formation, Oregon 211L, 220L
- Abundance (Acme) Zone xiR, 2, 5, 6L, R, 17L, 44L
- accumulation (rate) xviiiL, 8R, 10L, R, 11L, 12L
- accurate, accuracy 8L, 9L, 14L, 18L, 45L, 78L, 93L, 94L, 114L, 171R, 176R, 238L, 247R
- Acme Zones xiR, 17L, 44L, 86R, 87R, 88L
- Cf<sub>3</sub> acme zone 78R, 86R, 87L, 88L
- Across the River Tuff, Oregon 220R
- Adobe Town Member, Washakie Formation, Wyoming 108R, 109L, 115L, 116R, 117L, R, 127R
- lowest part equivalent to Washakie A of Granger 127R
- Africa 92L, R, 336L
- Aftonian interglacial stage (Pleistocene) 237R
- Agate Ash, Nebraska 194L, 220R
- Agate Bone Bed, Nebraska 194L
- Agate National Monument, Nebraska 194L
- Age or age viiR, xiiiL, xiiiL, R, xivL, R, xviiiL, R, xviiiL, 1L, R, 2L, 3L, 4, 7, 8L, R, 9L, R, 10L, R, 11L, R, 12R, 13R, 14L, R, 15L, 16L, R, 17L, R, 18L, 21–28L, R, 32L, R, 34L, R, 35L, R, 36L, R, 37R, 43–45L, R, 47, 53L, R, 61R, 62–66L, R, 67L, 68L, R, 69R, 70–73L, R, 74R, 75R, 76L, R, 77R, 78–79L, R, 80L, 81L, R, 83R, 84R, 85–87L, R, 88R, 89–94L, R, 95L, 96L, 106–107L, R, 111L, R, 113L, 115R, 116L, 118R, 119L, 120–121L, R, 122R, 123L, 123L, 124–138L, R, 158L, 159, 161L, R, 162R, 164R, 167L, 171R, 172R, 173L, R, 174R, 175L, R, 176R, 177–178L, R, 187–189L, R, 190R, 191L, R, 192R, 194L, 195R, 197R, 198L, R, 200–203L, R, 204R, 205–206L, R, 207L, 210L, R, 211L, 216L, R, 233L, R, 236–238L, R, 240, 241, 244, 246R, 247R, 249L, 250R, 251–252L, R, 255L, R, 256R, 257, 258L, R, 259L, 260–269L, R, 271–273L, R, 274L, 275–277L, R, 278L, 279L, R, 285L, R, 286L, 287L, R, 288R, 289, 291L, 292–293L, R, 294R, 295L, R, 315R, 321R, 324L, R, 327L, 331L, R, 332R, 334R, 335L, 337R, 338L
- age boundary not synchronous with lithologic boundary 107L, 115R, 116L, 159L–162L, 163R–164L, 165R, 213L
- Agua Fria area, Trans-Pecos, Brewster County, Texas 108, 116L, R, 129L, R
- Aguja Formation, Texas 24L
- Ahearn Member, Chadron Formation, South Dakota 158L, 162L, R
- Alamo Wash, San Juan Basin, New Mexico 45L, 65R, 66L, R, 67R
- Alamosa Formation, Colorado 279L
- Alaska 26R, 27, 95R, 111L, 132L, 239L, 249L, 255R, 256L, 270R, 272R, 275R, 280L, 285L, R, 292L, 318R, 230L, 326R, 327L, 334R
- Albian Stage/Age, faunas, rocks 22R, 23L, 317R
- Albuquerque Basin, New Mexico 196L, 198R, 199L, R, 257, 262R
- alien faunal elements 28L, 321R
- Allerød pollen zone 238R
- allochthonous 13R, 209L, 210, 211L, 212R, 214L, 318R
- Almagre beds, New Mexico 109R, 130L, R
- Almy Formation, Wyoming 107R
- American Falls Formation, Idaho (early *Bison*) 245, 286L, 289, 295
- Amynodon* sandstone, Utah 111R
- analog vs nonanalog faunas 290R, 291L, R
- Andean Basin, northwest Argentina, southern Bolivia 92R, 93L, R, 94R
- Animas Formation, Colorado 76L
- Antarctic(a) 317R, 318L, 320L, 324R, 326L, R, 329R, 330L, 334L, 3336R
- Antelope Valley, Nevada 260L
- Anxiety Butte, Saskatchewan, Canada 192L
- Anza-Borrego Desert, California 232R, 256L, 257, 259L, 271L, R, 272L, R, 371L, 275R, 277
- Anza-Borrego Desert State Park, California 258R, 259R, 271L, 277L
- Appalachian (region) 318R, 332L
- A-P Zone (= part of *Deltatherium* Zone, To2) 72R, 74L
- APTS (Astronomical Polarity Time Scale) xiL
- Aptian Stage/Age, faunas, rocks 23L
- Aquilan mammal age**, faunas, rocks 21L, R
- definition and characterization 23L, R
- Arcadia Formation, Florida 204R, 207L
- Archaeolambda* Interval Zone 91L
- Archusa Marl Member, Cook Mountain Formation, Mississippi 134L

- Ardath Shale, California 123L, 124R  
 Arenazzolo Formation, Messinian Stage, Italy 233R  
 Argile Plastique, Sparnacian, France 89L  
**Arikareean mammal age**, faunas, rocks 156R, 166L, R, 171R, 173L, 174R, 176R, 179L, 187L, R, 188–189L, R, 191–192L, R, 193L, 196L, 201L, 204L, R, 205L, R, 206L, 208R, 209R, 211–212L, R, 331–332L, R, 333L, 338R  
 definition and characterization 209R–211L  
 zonation 209R–212R  
 Arikaree Group, Great Plains, North America 173L, 179R, 188R, 189L, R, 192L, 193–194L, R, 196L, R, 197L, 209R, 211L, 212R, 213L  
 Arroyo Chijuillita Member, Nacimiento Formation, New Mexico 45R, 66R, 67R, 72R, 75L  
 arvicoline rodent biochronology, faunal zones 232L, 247L, R, 248, 250L, 253R, 254L, R, 258R, 274–276L, R, 278R, 279L, R, 281L, 292R, 293L, R, 294L  
 Ash Hollow Formation, Nebraska 177R, 195R, 216L, R, 220R, 337L  
 Asia 23R, 37R, 78R, 85R, 89R, 90–91L, R, 92L, 93R, 94R, 173R, 207R, 209L, 218L, 238L, 250L, 254–255L, R, 274R, 275R, 284R, 323, 324–326L, R, 328L, R, 329R, 330L, 334L, R, 336L, 338L  
 Asian origins for:  
 pantodonts 91L, 93R, 94R  
 rodents 85R  
 tillodonts 85R  
*Asiostylops* Interval Zone 91R  
 Assemblage Chron xiL, R, 5, 15L  
 Assemblage Fossizone or Fossilzone xiR, xiiR, 4, 5, 6L, R, 15L  
 Assemblage Zone xiL, R, xivR, 5, 6L, R, 15L, 32L, 113L, 121R, 170L, 177L, 181, 191L  
 Australian Creek beds, British Columbia, Canada 133L  
 astronomical precession cycles 233R  
 Australia 324R, 326L, 329R  
 autochthone, autochthonous 209L, 211L, 212L, 215L, 217R, 218R  
 Axel Heiberg Island, Nunavut Territory, Canada 108R, 109R, 111L, R, 133L, R  
 Aycross Formation, Bighorn Basin, Wyoming 108L, R, 114L, R, 120L, 122L  
 Baca Formation, New Mexico 109R, 115R, 130R, 131L  
 Badwater Creek, Wind River Basin, Wyoming 115L, R, 118R, 120L, R, 121R, 160L  
 Baja California del Norte, Mexico 25L, 37R, 108L, 111L, 131R, 138L, 321R  
 Baja California Sur 257, 264L  
 Bajo de la Palangana, Bolivia 94R  
 Bald Peak Volcanics, California 219R  
 Balm Creek, John Day region, Oregon 187L  
 Banco Negro Inferior, Argentina 94R  
 Bandelier Tuff, Jemez Mountains, New Mexico 234, 262R, 271L  
 Barstow Formation, California 9L, 171R, 173R, 174L, 175R, 177R, 178L, 191L, 213R, 214L, 215R  
**Barstovian mammal age**, faunas, rocks 171R, 172L, 173R, 174L, R, 175R, 176L, 177R, 178L, R, 188L, R, 189R, 190R, 191R, 192L, R, 195L, 196L, 197R, 199L, 200L, 202L, R, 203L, R, 204R, 206R, 207L, 208L, 209L, 213R, 214–215L, R, 216L, 217L, 260R, 332R, 334R, 335L  
 definition and characterization 215–216L  
 zonation 215L–216L  
*Peridiomys-Parahippus* Zone (Ba1) 191L  
*Lignimus-Tardontia* Zone (Ba2) 191L  
 Barstovian-Clarendonian boundary 217L  
 Bashi Formation, Mississippi 108L, 111L, 134L  
*Bathygenys* Interval Zone (earliest Chadronian) 162R  
 Bayfield, San Juan Basin, Colorado 83R  
 Bear Creek, Montana 50, 76R, 85R, 87L  
 Bear Formation, Montana 62L, 69R  
 Bearpaw Shale, Montana 24R  
 Beaver Divide, Wind River Basin, Wyoming 108R, 115L, R, 121R, 158R  
 Beaverhead Mountains, Idaho 187R  
*Bemalambda*, Interval Zone 91L  
 Bena Formation, California 172L  
 bentonite (itic) 63L, 123R, 134L  
 Beringia(n) (dispersals) 91R, 254L, 274L  
 faunal region 249L, 274R, 276R, 280L, 292R, 293R  
 Bermont Formation, Florida 270R, 283R  
 Betonnie-Tsosie wash, San Juan Basin, New Mexico 45R, 48, 66L, R, 71L, 72R, 73R  
 BFE (benthic foraminiferal extinction) 320L  
 Big Badlands, South Dakota 160L, 162L, 164R, 165L, R, 166L  
 Big Basin Member, John Day Formation, Oregon 179L  
 Big Bend area (National Park), Texas 69R, 77L, 80L, 82L, 83R, 85L, 108L, R, 129L, R, 134R, 201L, R, 211R  
 Big Cottonwood Creek Member, Chadron Formation, Nebraska 158L, 161L  
 Bighorn Basin, Wyoming 11R, 12L, 16L, 17L, R, 24L, 25R, 47, 50, 63R, 64L, 70R, 74R, 75R, 76R, 77L, R, 78R, 79L, 80L, R, 81L, 82–83L, R, 85L, 86L, 87L, R, 88L, 90L, 107R, 108L, R, 111R, 114L, 121, 122L, R, 122L, 125L, R, 132R, 135, 138L, 139L, 231L, 324R  
 “Big Horn Wasatch,” (= Willwood Formation) 107R  
 biocorrelation 14R  
 biochron xiL, R, xiiL, 4, 5, 7, 16R, 17L, 43R, 113R, 114R, 115L, 116L, R, 117L, R, 125R, 131L, 135, 136, 137, 176R, 217L  
 biochronologic (unit) xiL, xiiR, xiiiR, xivL, R, xviiiL, 1L, 2, 4, 5, 7, 9R, 10L, 13R, 14L, R, 15L, R, 16L, 17L, 18L, 43R, 45L, 46, 47, 53R, 63L, 64R, 66L, R, 68L, 69L, 70L, 75R, 76L, 79R, 90R, 95L, 106R, 109L, 111R, 112R, 131R, 138R, 139L, 171L, 175L, 179R, 193R, 194L, 195L, 197L, 198L, 204R, 205L, 209R, 239R, 252R, 254R, 268R, 270L, 279L, R, 281L, 292L  
 biochronology xiR, xviiR, 1L, R, 3R, 6L, 12R, 14L, R, 15L, 16L, 21L, 23L, 24R, 35R, 44R, 46, 68L, 106L, R, 112L, 160R, 169L, R, 173R, 192R, 198R, 208R, 213L, 232L, R, 239R, 247R, 249L, 253R, 274R, 276L, R, 279R, 292L, 293R, 295L  
 biogeographic (-graphy) 25L, 28L, 36R, 37L, 38L, 69R, 95L, 290R, 292L, R, 293R  
*Biorbia* seed assemblage, Ogallala Formation, Oklahoma 198L  
 biostratigraphic (unit) xiL, R, xiiiL, R, xivL, R, xviiL, xviiiL, 2, 3L, R, 4, 5, 6L, 9R, 13R, 14L, R, 15L, R, 16L, 17L, 18L, 25L, 36R, 53R, 65L, 66L, 74L, 75R, 79R, 89L, 95L, 112L, R, 116R, 122R, 127L, 156L, 161R, 162L, R, 163R, 164L, R, 165L, R, 166R, 169R, 170L, 177L, 179L, 188L, 192R, 193L, 197R, 207R, 208R, 214L, 217R, 233R, 239R, 247L, 259L, 263L, 264R, 268R, 291L  
 biostratigraphy viiL, xiR, 1L, 3L, R, 6L, 10L, 14R, 17R, 46, 53R, 90L, 158L, 162L, 164L, 173R, 176R, 177L, 179L, 187R, 191L, 193L, 196R, 197L, 200L, 207R, 232L, R, 239R, 268R, 270R, 283R, 295L  
 biotic/abiotic 232R, 293L  
 Biotite Tuff, Oregon 220R  
 biozone xiR, xiii, xiiiR, 2, 3R, 4, 7, 10R, 16R, 17R



- Bishop Ash, Long Valley Caldera, California 234, 265R, 279R
- Bison Basin, Wyoming 77L, 81L, 82L, 83R
- Bitter Creek, Wyoming 126R, 127L
- Black's Fork Member, Bridger Formation 108R, 111L, 115R
- Blacksforkian subage (Br 1b, Br-2), Bridgerian mammal age 115L, R
- Black Peaks Member, Tornillo Formation, Texas 74R, 77L, 79L
- Blackwater Draw Formation, Texas 266R
- Blaine County, Montana 23R
- Blancan mammal age**, faunas, rocks 175L, 178L, 200R, 201R, 207R, 218L, 232L, 233L, 235, 236, 239L, 246L, R, 247R, 248, 249–270, 271R, 272L, 273R, 274L, R, 276R, 277L, 278R, 280R, 281L, 282R, 283L, R, 289, 292L, R, 293R, 294–295L, R, 335R, 337R, 338L  
 definition and characterization 250L, R  
 geographic distribution 256L, R  
 Pacific Northwest 256L, R  
 Snake River Plain 256R–258R  
 California 258R–260L  
 Great Basin 260L–261L  
 Southwestern U.S. 261L–263R  
 Mexico 264R–264R  
 Great Plains 264R–276R  
 Florida 267R–268R  
 historical treatment 249L–250L  
 zonation 252L, 252R–256L  
 Blancan I subage 253R, 254L, R, 256L  
 Blancan II subage 253R, 254R–255L, 256L  
 Blancan III subage 253R, 255L, R, 256L  
 Blancan IV subage 253R, 254R–255R, 256L  
 Blancan V subage 253R, 255R–256L  
 Arvicoline Zones II and III 253R–254L  
 Rexroadian subage 253L, R  
 Senecan subage 253L, R  
 Blancan-Irvingtonian boundary 263L, 269R, 277L, 283L
- Blanco Ash, Blanco Formation, Texas 266R
- Blanco Formation, Texas 253R, 266R
- Bonanza, Utah 126L
- boundary stratotype 233L, R, 237L, 238L, R, 239L, 295R
- bolide impact 320R
- Bølling pollen zone 238R
- Box Butte Formation, Nebraska 194R
- Bracks Rhyolite, Texas 120L, 137, 138R
- Bramblett Formation, Texas 263R
- Branch Canyon Formation, Cuyama Valley, California 173R
- Brandon flora, Maine 332L
- Brennan Basin Member, Duchesne River Formation, Utah 109L, 112L, 117R, 118L, R, 126R, 136
- Bridge Creek flora, Oregon 330L
- Bridger A-D (Bridgerian) 111L, R, 114R–115R, 128L, R
- Bridger E (early Uintan) 108R, 111L, 116L, R, 117L, 128L, 129R, 136
- Bridger Formation 107L, 108R, 111L, R, 128L, R, 135, 136
- “Bridger Group” 111L
- Bridgerian mammal age**, faunas, rocks 106R, 107L–109R, 111L, 112R–117L, 120R, 121R, 122L, R, 123L, 124R, 126L, R, 127L, R, 128L, R, 129R, 130R, 131L, R, 132L, R, 133R, 134L, R, 135, 138L, R, 139L, 326L, R, 327L, R, 328L  
 definition and characterization 113L–114R  
 zonation 114R–115R
- Blacksforkian subage, (Br1b, Br2) 114R–115R
- Gardnerbuttean subage 107R, 113L, 114R, 115L, 121R, 126L, 135
- Twinbuttean subage 115L, R
- Bridger A 108R, 111L, R, 115L, R, 128L, R, 135
- Bridger B 108R, 111R, 115L, R, 128L, 135
- Bridger C 111L, 115L, R, 128L, 135
- Bridger D 115R, 116L, 128R
- Bridger E (early Uintan) 108R, 111L, 116L, R, 117L, 128L, 129R, 136
- Bro, Gardnerbuttean 107L, 108L, R, 113L, R, 114L, R, 115L, 121L, R, 126L, 127L, 131L, 134R
- Br1a, Gardnerbuttean 108L, R, 113R, 114L, R, 115L, 121L, 127L, 128R, 131L, R, 134L, 135R
- Br1b, Blacksforkian 108R, 114R, 115L, R, 128R, 135
- Br2, Blacksforkian 108L, R, 114R, 115L, R, 120R, 122R, 123L, 126L, 127R, 135
- Br3, Twinbuttean 107L, 108R, 114R, 115L, R, 116L, 117L, 120R, 125L, 127R, 131L, 135
- Bridgerian-Uintan boundary 115R, 116L, R, 117L, 129L, 131R, 138L
- British Columbia 108R, 109R, 132R, 326R, 327L, 330L
- Brougher Dacite, Nevada 220L
- Brown Siltstone Member (Bed), White River Group, Nebraska 158L, 166R, 193L, 196L, 209R
- Brule Formation, Nebraska, South Dakota 158L, 160L, R, 161L, 163R, 164L, 165R, 173L, 193L, 196L, 209L
- Bruneau Formation, Idaho 271L, 276R, 277
- Brunhes magnetic polarity chron 233L, 238L, 259R, 261L, 263L, 274L, 278L, 279L, R, 280L, 281L, 285R, 287L
- Buckshot Ignimbrite, Texas 120L, 130L, 137, 138R
- Buffalo Canyon flora, Nevada 332L
- Bug Creek, Montana 35L, 36L, 51, 53L, 63R, 64L, R
- “Bugcreekian” (mammal age) 35L, 43L, 53L
- Bull Draw fauna (Irvingtonian, Cudahyan), Texas 273R, 277, 282R
- Bullhead Member, Fox Hills Formation, South Dakota 34L
- Bumbanian mammal age, Asia (approx. corresponds to Wasatchian) 89R, 92L
- Bunophorus* Interval Zone 17R
- Burge Member, Valentine Formation, Nebraska 177R, 195R, 216L, R, 217L
- Burnt Fork Limestone, Bridger Formation, Wyoming 115R
- C<sub>3</sub>/C<sub>4</sub> photosynthesis 335R
- Cabbage Patch beds, Montana 187R, 188R, 189L
- Cabrillo Formation, California 122R
- Cady Mountains, California 176R
- Calabrian Stage, Italy 236R
- Calcaires de Rona, Transylvania, Romania, ?Clarkforkian equivalent 89L
- calibration viiR, xviiiL, 1R, 8R, 10L, 11L, 14L, 93L, 95R, 134R, 135, 136, 137, 138L, 156L, 162L, 166L, R, 167L, 169L, 171R, 172L, R, 175L, R, 176L, 177L, 178L, 187L, R, 191L, 195R, 196L, 200R, 206L, 207L, 210, 211R, 212L, 233L, 238L, 254L, 258R, 259L, 261L, 267L, 275L, 290L
- Calico Mountains, California 214R
- Caliente Formation, California 173L, R, 174L, R, 214L, 219R
- Caliente Range, California 173L, R
- California Coast Ranges 171L, 203L, 214L, R
- Calvert Formation, Maryland 207R, 208L
- Campanian Stage/Age, faunas, rocks 22R, 23L, R, 24L, R, 25L, 37R, 92R, 317R, 318L, 321R
- Camp Rice Formation, New Mexico, Texas 257, 263L, R

- Canada 23R, 26R, 37L, 45R, 46, 52, 62L, 63R, 68R, 69R, 71L, 81L, 83L, 95R, 106L, 108R, 109R, 111L, R, 132R, 191R, 192L, 239L, 249L, 257, 270R, 277, 280R, 289, 318R, 334R
- Cañon Rhyolite, Nevada 178R, 220L
- Canyada Pilares Member, Zia Formation, New Mexico 196L, 199L
- Cape Deceit, Alaska 257, 270R, 275R, 280L, 285L
- Capo Rossello, Sicily 233R
- Capote Mountain Tuff, Texas 160L
- Carbon Basin, Wyoming 77L, R, 80R, 82L
- carbon isotope (excursion) 32L, 36R, 43R, 89R, 90L, R, 92L, 106L, 138L, 316, 317L, 318L, 320L, R, 335R
- Caribbean (area, plate) 317L
- Carson Valley, Nevada 260L
- Carter, Wyoming 107R
- Carter County, Montana 26L
- Carter Mountain, Wyoming 122R
- Carthage-La-Joya Basin, New Mexico 131L
- Cascade Range, Oregon 187L, 327L, 330R, 331L, 332L, 337L
- Cathedral Bluffs Tongue of Wasatch Formation 107L, 108R, 112R, 113R, 114L, R, 115L, 126R, 127L, 128L, R
- Cedar Butte Basalt, Idaho 286L
- Cedar Creek Member, Orella Formation, Colorado 160L, 165L
- Cedar Mountain Formation, Utah 22R, 111L
- Cedar Mountain Member, Bridger Formation, Wyoming 111L
- Cedar Mountain, Wyoming 50, 51, 62L, 63R, 64L, 77R
- cement-bearing, hypsodont (equid) cheek teeth 199L
- Cenomanian Stage/Age, faunas, rocks 22L, R, 23L, 317R, 318L
- Cenozoic System/Period, faunas, rocks 43L, 90R, 93L, 122R, 194R, 202L, 209L, 232L, R, 233R, 234, 247R, 252R, 260R, 292L, 316, 317L, 318L, 336R
- Cernasian mammal age, France 89L, R, 90L
- Cernay Conglomerate, France 89L, R
- Cerro Conejo Fm., New Mexico 221L
- Cerro Redondo, Argentina 94R
- Cerro Toledo B Ash, Kansas 234, 240, 266L
- Cerro Toledo X Ash, Texas 234, 240, 271L
- Cerro Toledo Rhyolite, New Mexico 234, 266L, 280R
- Cerrotejonian Stage 171L, R, 172L, R, 174R, 177L, R
- Cfi-Cf<sub>3</sub> *see* Clarkforkian
- Chappo Member, Wasatch Formation, Wyoming 49, 77L, 82L, 85L
- Chadron A, Nebraska (informal subdivision of formation) 158L
- Chadron B, Nebraska (informal subdivision of formation) 158L
- Chadron C, Nebraska (informal subdivision of formation) 158L
- Chadron Formation, Nebraska, South Dakota 156R, 158L, 160R, 161L, R, 162L, 163R, 164L  
purplish-white layers 159, 160R, 161L  
“Chadronian-aspect” taxa 119L, R
- Chadronian mammal age**, faunas, rocks 106L, 109, 119L, R, 120L, 123R, 129R, 130L, R, 131L, 133L, R, 134L, R, 137, 138R, 156L, R, 158L, 159, 160–163L, R, 164L, 166R, 167L, 192L, 325, 328R, 329R, 331L  
definition and characterization 120L, 160L–162L  
zonation 162L–163R
- Earliest Chadronian 162R
- Late early Chadronian 162R
- Bathogenys* Interval Zone (earliest Chadronian) 162R
- Leptomeryx yoderi* Interval Zone (late early Chadronian) 162R
- Leptomeryx mammifer* Interval Zone (middle Chadronian) 163L
- Miniochoerus chadronensis* Interval Zone (late Chadronian) 163R
- Chadronian-Orellan boundary 160L, R, 161L, 162L, 163R, 164L
- Chalk Cliffs (Hepburn’s Mesa), Montana 190R, 191L
- Challis volcanics, Idaho 326R
- Chamberlain Pass Formation, South Dakota 158L
- Chambers Tuff, Texas 160L
- Chamisa Mesa Member, Zia Formation, New Mexico 198R
- Chamita Formation, New Mexico 200L, R
- Chanac Formation, California 172R  
characterize, characterization xil, R, xiiL, 1L, 3L, R, 5, 6L, 11, 12, 13L, R, 16L, R, 17R, 21R, 28L, 36R, 43R, 44L, 62L, 65R, 66R, 71L, R, 78R, 79L, 83L, 85R, 86L, 93L, 95L, 111R, 113L, 115L, 116R, 117L, 118L, 119L, 120L, 121R, 134R, 138L, R, 161R, 162R, 164R, 165L, 166L, 170L, 171L, 176L, 177L, 191L, 192R, 193L, R, 194L, 206L, 209L, R, 211L, 212L, R, 213L, R, 214L, R, 215L, R, 217L, R, 218R, 219L, 249R, 250L, 253L, R, 254L, R, 255L, R, 256L, 263L, 269R, 270L, 271R, 273R, 274L, 275L, 276L, R, 283R, 284R, 290L, 291R, 293L, 294L, 318R, 324R, 328L, 330R, 332R
- chemostratigraphy (ic) 10L, 32L, 36R
- Cheswold Sand, Calvert Formation, Maryland 208L
- Chiapas, Mexico 202R, 203L, 214R
- Chickaloon Formation, Alaska 132L
- Chimney Rock Ash, Nebraska 220R
- China 85R, 90R, 91R
- Chisos Formation, Texas 109L, 129R, 130L
- Choptank Formation, Maryland 207R, 208L
- Choteau County, Montana 23R
- Cripple Creek Sump, Alaska 285R
- chron xiL, R, xiiiL, xiiiL, R, xivL, R, 2L, 4, 5, 7, 12R, 15L
- chronocline(s) 290L
- chronocorrelation 14R
- chronofauna xiiL, 166R, 173L, 193L, 194L, R, 195L, 197R, 205L, 206L, 209L, R, 214L, 215R, 216R, 217L, 218L, 328R, 329L, 330L, R, 331L, R, 332R, 333R, 334L, R, 335R, 337L
- chronology viiR, xiiiL, xviiR, 1L, R, 2R, 3L, R, 6L, 7, 8L, R, 9R, 10L, 12R, 14L, R, 15L, 16L, 24L, 171L, 175L, 179L, 209L, 210, 213L, 249L, 255R, 265R, 269R, 274R, 287L, R, 288L, R
- chronologic(al) viiL, R, xiiiL, xviiL, xviiiL, 1L, R, 5, 7, 8R, 9R, 13R, 14L, R, 16L, 17L, 169L, 171L, 172R, 175L, R, 176R, 180, 187L, 194L, 197L, 198L, 204L, 210, 213R, 215L, 218R, 239L, R, 247L, 253R, 257, 258L, 267R, 268R, 277, 279L, 285R, 287L, R, 289, 290L, 295L, 338L
- chronometric 239L
- chronostratigraphic (unit) xiR, xiiL, R, xivL, xviiL, xviiiL, 1L, R, 2L, R, 3L, R, 5, 6L, R, 7, 8L, R, 13L, R, 14L, R, 15L, R, 18L, 21L, 43R, 47, 95L, R, 161R, 238L, 239L
- chronostratigraphy viiL, xiiL, 1L, 2R, 3L, 5, 6L, R, 8L, 10L, 14R, 138R, 161R, 167L, 177L
- chronozone xiR, xiiL, xivL, 2L, 5, 6L, 7, 8L, R, 14R, 71R, 72L
- CIE (carbon isotope excursion, at LPTM) 106L, 316, 320L, R, 324R
- Cita Canyon, Texas 244, 255L, R, 257, 266R, 267L, 268R
- cladogenesis 213R, 232L
- Clarendonian Chronofauna 333R, 334L, R
- Clarendonian mammal age**, faunas, rocks 174L, 176L, 177R, 178L, 195R, 197R, 198L, 200L, 207L, R, 216L, R, 217L, R, 218L, R, 260R, 332R, 333R, 334L, R, 335L  
definition and characterization 216L, R, 217L, R

- zonation 217L, R  
*Ustatochoerus profectus-Copemys russelli* Assemblage Zone (Cl1) 177L  
*Cupidiniimus avawatzensis-Paracosoryx furlongi* Assemblage Zone (Cl2) 177L  
*Epicyon haydeni-Hipparion forcei* Assemblage Zone (Cl2/3) 177L  
 Clarendonian-Hemphillian boundary 195R, 217R  
**Clarkforkian mammal age**, faunas, rocks 43R, R, 44L, R, 45L, 46, 49, 61, 76R, 78R, 83R, 84R, 84R-88L, 89L, R, 90L, 91R, 92L, R, 95R, 112R, 120L, 122L, 125L, R, 126R, 127L, 128L, 129L, 131R, 134L, R, 138L, 321L, 324L, R  
 definition and characterization 84R-86L  
 zonation 86R-88R  
 Rodentia/*Plesiadapis cookei* Interval Subzone (Cf1) 87L, R  
*Plesiadapis cookei* Lineage Zone (Cf2) 87R, 88L  
*Phenacodus-Ectocion* Acme Zone (Cf3) 88L, R  
 Clarkforkian Stage/Age 16L  
 Clarkforkian-Wasatchian boundary 134R, 138L, 324R  
 Clark Fork faunal zone, Wyoming 84R  
 Clark Fork Member, Polecat Bench Formation, Wyoming 84R  
 Clarkia flora, Idaho 334L  
 Clarks Fork Basin, Wyoming 16L, 17R, 46, 71R, 75R, 76R, 77L, 78R, 79L, 80L, R, 81R, 82R, 83L, R, 84L, R, 85L, R, 86L, R, 87L, R, 88L, 90L  
 Clarkston Basin, Montana 160L  
 Clarno flora, Oregon 327L, R, 330L  
 Clarno Formation, Oregon 108R, 109R, 133R, 327L, R, 332L  
 climate (atic) 232R, 237L, 238L, R, 266L, 280R, 286R, 287L, 290L, 291R, 292R, 293L, 294R, 315L, R, 316, 318L, R, 319, 320L, R, 321L, R, 324R, 326R, 327L, 328L, R, 329L, 330L, R, 331R, 332L, 333L, R, 334L, R, 336R, 337L, 338L  
 in correlation 232R, 237L, 238L, R, 247R, 266L, 280R, 287L, 290L, 291L, 292L, 320L, 324R, 333R  
 Climbing Arrow Formation, Montana 160L  
 Coalspur Formation, Alberta, Canada 71L, 75L  
 Coal Valley Formation, Nevada 220L  
 Cobb Mountain magnetic polarity subchron of Matuyama Chron 240, 263L  
 Cochiti magnetic polarity subchron of Gilbert Chron 240, 256R, 258R, 259L  
 Coldwater Formation/Sandstone, California 124R, 138R  
 Coffee Ranch Ash, Texas 221L  
 Colgate Member, Fox Hills Formation, South Dakota 34L  
 Colter Formation, Montana 191L, R  
 Colton Formation, Uinta Basin, Utah 108L, 114L, R, 125R  
 Colmena Formation, Texas 109L, 188R, 130L  
 Columbia Plateau, Nevada, Oregon 171R, 178L, R, 212R, 214R, 215L, 334R, 337L  
 concurrent-range chron xiiiL, 4, 15L  
 concurrent-range zone xiR, xiiiL, 4, 6L, 15L  
 Coniacian Stage/Age, faunas, rocks 23L, R, 317  
 Continental Divide, North America 187R, 188L, R, 189L, R, 326L, R  
 Copper Basin flora, Nevada 327R  
*Cupidiniimus avawatzensis-Paracosoryx furlongi* Assemblage Zone, Clarendonian (Cl2), California 177L  
 Cook Mountain Formation, Texas 134L  
 Cornell Dam Member, Valentine Formation, Nebraska 195L  
 correlation viiL, R, xiL, xiiL, R, xiiiR, xviiiL, xviiiL, 1L, R, 2R, 3R, 5, 7, 8L, R, 9R, 12R, 13L, R, 14L, R, 15L, R, 16L, R, 17L, 18L, 21R, 23R, 24L, R, 25L, R, 26R, 32L, R, 34L, R, 36L, R, 43L, R, 44R, 45L, R, 46, 47, 63L, 65L, R, 66R, 67L, R, 68L, 69L, R, 71L, R, 72L, R, 73L, R, 74L, R, 75R, 76L, 77L, 78L, 79L, 80R, 81R, 82L, R, 84L, 87L, R, 88L, R, 89L, R, 90R, 91L, R, 92L, 93L, R, 94L, R, 95L, R, 106L, 109L, 113R, 119L, 120R, 121, 122L, R, 123L, R, 124L, 125L, R, 127L, R, 129L, 130L, 131R, 133L, R, 134L, R, 136, 138L, R, 156L, R, 158L, R, 159, 160L, 162L, R, 164R, 165L, 166L, R, 169L, 170, 171R, 172R, 173L, R, 174L, R, 175L, 176R, 177L, R, 178L, 179L, R, 180R, 188L, R, 189L, 191R, 192L, R, 193R, 194L, R, 197R, 198R, 199R, 201L, 204L, 205L, 206R, 207L, R, 209L, 214R, 216L, R, 219L, 233L, R, 234, 237L, 238L, 239L, R, 240, 247R, 251R, 252R, 255L, R, 256L, R, 258L, 260L, 261R, 263L, 265L, 266L, R, 267L, 268L, R, 270L, 278R, 281R, 283L, 285R, 287L, 291L, 294R, 295L, 315R, 317R, 318L, 320R, 324L, 330L, 334R, 335L, 338L  
*Coryphodon* zone 107R  
 Coso Formation, California 257, 259R  
 Cougar Point Tuff V, California 177L, 220L  
 Cougar Point Tuff XIII, California 220L  
 Coyote Creek flora, Aycross Formation, Wyoming 120L  
 Cozy Dell Shale, California 124R  
 Crater Tuff-Breccia Member, Colter Formation, Wyoming 191R  
 Crazy Johnson Member, Chadron Formation, South Dakota 158L, 162L, 163L  
 Crazy Mountains (area, basin, field) 51, 69R, 70R, 71R, 73R, 74L, 76R, 77L, 80L, R, 81L, R, 82L  
 Creede flora, Colorado 330L  
 Cretaceous System/Period, rocks, faunas 21L, R, 22, 23L, R, 24L, 25R, 26R, 32L, R, 34, 35L, R, 36L, R, 37L, R, 43R, 53L, 62L, 64R, 89L, R, 92R, 93R, 95R, 111L, 116L, 122R, 130L, 315R, 317L, R, 318L, R, 319, 320L, R, 321L, R, 324L, 326L, R  
 Cretaceous-Paleocene boundary 43R  
 Cretaceous-Tertiary boundary 21R, 32L, R, 35R, 36L, R, 37L, 43R, 62L, 64R, 320L  
 Cronese Tuff, California 220L  
 Crooked Creek Formation, Kansas 265R  
 Crookston Bridge Member, Valentine Formation, Nebraska 191R, 195R  
 CTS (Classical Time Scale) xiiiL, 9L, R  
 Cuba Mesa Sandstone Member, San Jose Formation, New Mexico 45L, 48, 71R, 72R, 73R, 76L 130R  
 Cub Creek, Montana 50, 75R, 77L, 80R  
 Cub Mountain Formation, New Mexico 109R, 131L  
 Cucaracha Formation, Panama 203R  
 Cuchara Formation, Colorado 108L, 131L  
 Cudahyan subage, Irvingtonian mammal age 273L, R, 274L  
 Cudahy Ash 281R  
 Cuisian Stage/Age, Europe 89R  
 Cuyama Badlands, Cuyama Valley, California 173R, 174L, R  
 Cuyama Valley, California 172R  
 cyclostratigraphy viiR, xiiR, 10L  
 Cypress Hills Formation, Saskatchewan, Canada 109L, 118R, 119R, 160L, 192L  
 Cypress Hills Plateau, Saskatchewan, Canada 133L, 191R, 192L, R  
 Cypress Hills, Saskatchewan, Canada 26R, 329L  
 Dababiya section (Paleocene-Eocene boundary), Egypt 106R  
 Dakota Formation, Utah 22R  
 Dano-Montian Stage/Age 94R

- Dated Tuff, California 219R  
 datum xiiR, xiiiL, R, xivL, xviiiR, 5,  
 13R, 14L, R, 15L, R, 17L, 23R, 44L,  
 203L, 215L, 258R, 269L, 334R, 335L  
 Davis Ash, Ash Hollow Formation,  
 Nebraska 195R, 220R  
 $\delta^{13}\text{C}$  excursion 11R, 18L, 32L, 43R, 89R,  
 90L, R, 92L, 316, 317R, 320L  
 Debeque Formation, Colorado 85L,  
 108L, 125L, R  
 deciduous (forest) 318R, 319, 320R,  
 321L, 324L, 326R, 327L, R, 328L, 330L,  
 R, 332L, 333R, 334L, R  
 Deep Creek Tuff, John Day Formation,  
 Oregon 172R, 179R, 187R, 189R, 211L,  
 R, 220R  
 Deep River Formation, Montana 188L,  
 189R, 190R, 191L, 213R, 214R  
 Deer Gulch lava, Idaho 240, 258L  
 define (ing), definition viiL, xiL, R,  
 xiiiL, R, xiiiL, R, xivL, R, xviiR,  
 xv–iiR, 2, 3L, R, 4, 5, 6L, R, 7, 8L, R,  
 13L, R, 14L, R, 15L, R, 16R, 17L, R,  
 18L, 21R, 22, 24R, 25L, 26R, 28L, R,  
 36L, R, 43R, 44L, 45R, 53L, R, 62L,  
 63L, R, 64R, 65L, R, 66L, R, 67R, 70L,  
 R, 71L, 72L, R, 73L, 74L, R, 75L, 76L,  
 78L, R, 80L, 81L, 82L, R, 83R, 84L,  
 85L, R, 86R, 87R, 88, 89L, 90R, 91L,  
 R, 93L, 95L, 106L, 111R, 112L, R, 114L,  
 115R, 117L, R, 118R, 120R, 121R, 122L,  
 130R, 131L, 132R, 158L, 160L, R, 161L,  
 R, 162L, 164L, R, 165R, 166L, 175R,  
 177L, 191L, 192L, 193L, 194R, 195L,  
 203L, 205L, 209L, R, 211L, R, 212L, R,  
 312L, R, 214L, R, 215L, R, 216R, 217L,  
 R, 218L, R, 219L, 233L, R, 237L, R,  
 238L, 239L, 247R, 248, 249L, R, 250L,  
 251L, 252R, 253R, 254L, R, 255L, R,  
 256L, 258R, 260R, 261R, 263L, 264R,  
 265R, 267R, 269L, R, 270L, 271R,  
 272L, R, 273L, 274R, 275L, R, 276L, R,  
 277R, 278L, 281R, 283L, 284R, 286L,  
 288L, 290L, 291R, 292L, 293L, R,  
 294L, R, 29RL, R, 335L, 337R  
 base defines boundary 3R, 6L, 8R,  
 13L, 17R, 177L  
 Delaho Formation, Big Bend area,  
 Texas 201L  
 Delmar Formation, California 108R,  
 122R, 123L, 124R  
*Deltatherium* Chronozone 72L  
*Deltatherium* zone 70L, 72L, R  
 De-na-zin wash (Barrel Springs), San  
 Juan Basin, New Mexico 45R, 48,  
 65R, 66L, R, 67R, 71L, 72R, 73R  
 dentine tracts 247L  
 Denver Basin, Colorado 49, 63R, 66R  
 Denver Formation, Colorado 64R, 66R  
 Devil's Graveyard Formation, Trans-  
 Pecos, Texas 198R, 109L, R, 116L,  
 117L, R, 118R, 129L, R  
 Devil's Gulch Member, Valentine For-  
 mation, Nebraska 195R  
 diachroneity, diachronous, diachrony  
 xiiR, xiiiL, xivR, xviiiR, 10R, 36L,  
 138L, 239L, 249L, 256L, 272R, 292R,  
 293L  
*Dinohippus* HSD<sub>k</sub> 259L  
 dinosaur (s) 21R, 24L, R, 26R, 28L, 35R,  
 36R, 37L, R, 64R, 93R  
 Dinosaur Park, Alberta, Canada 24L  
 disjunctive overlap fossilzone 5  
 disperse, dispersal(s) 13R, 14L, 23R,  
 78R, 88R, 89L, 90R, 91L, R, 92R, 93R,  
 94L, R, 95L, 106L, 209L, 239L, 247L,  
 249L, 254L, 292L, 293L, 315L, R, 316,  
 317L, 320L, 323, 324L, R, 325, 326L,  
 327R, 328L, R, 329R, 331L, 333, 335R,  
 336, 338L  
 Dogtown Member, Torreya Forma-  
 tion, Florida 206R  
 Domingo Basin, New Mexico 257,  
 262R  
 Douglas, Wyoming 158R, 161L, R, 163L,  
 164L, R  
 Dove Spring Fauna, California 177L, R  
 Dove Spring Formation, California  
 175R, 176R, 177L, R  
 Dragonian Mammal Age (= To1) 43L,  
 49, 62R, 70L, R, 71L, R, 72L, R,  
 73L, R  
 Drake Passage 326L, 329R  
 Drewsey Fm., Oregon 220R  
 Drinkwater Basalt, Oregon 220R  
 Dry Canyon Tuff, California 219R  
 Dry Gulch Creek Member, Duchesne  
 River Formation, Utah 109R, 112L,  
 118R, 126R  
 Duchesnean-Chadronian boundary  
 120L, 161R  
**Duchesnean mammal age**, faunas,  
 rocks 106R, 107L, 112L, 118L, R, 119L,  
 R, 120L, 121, 122R, 123, 124L, R, 125L,  
 R, 126L, R, 127, 129L, R, 130L, R,  
 133L, R, 134R, 136, 137, 138R, 139L,  
 161R, 328L, R, 329L, R  
 definition and characterization  
 119L, R  
 faunas restricted to those from Dry  
 Gulch Creek and lower LaPoint  
 members 112L  
 time of faunal revolution and re-  
 placement 119L  
 time of provinciality 119L  
 Duchesne River Formation, Utah 107L,  
 109R, 111R, 112L, 116L, 117R, 118L, R,  
 126L, 136R  
 Dunbar Creek Formation, Montana  
 160L  
 Dunbar Creek Member, Renova For-  
 mation, Montana 160L  
 Duncan Basin, Arizona 257, 261R, 262L  
 Eagle Crag Ash, Nebraska 194L, 220R  
 Early Eocene Climatic Optimum 320L  
 early Wasatchian; see Sandcouleean  
 Eastern Idaho, North America 187R  
 Eastern Oregon, North America 178R  
 East Fork Basin, Wyoming 115R, 117L,  
 120R, 135  
 Eastgate flora, Nevada 332L  
 E-A Zone (= part of *Deltatherium*  
 Zone, To2) 72R, 74L  
*E. grangeri-Arctocyon ferox* Interval  
 Zone, 17L  
*Ectoconus-Taeniolabis taoensis* (Pu2)  
 biochron 16R  
*Ectoconus-Taeniolabis taoensis* Interval  
 Zone (Pu2) 63R, L, 65R, 66R  
 EDE (Eolian Dust Event, at LPTM)  
 320L, R  
 Eden Hot Springs, California 259R  
 “**Edmontonian**” mammal age, faunas,  
 rocks 21L, 24R, 25L, 26R, 28L, 32  
 definition and characterization 24R,  
 25L  
 Edwards Plateau, Texas 280R  
 egg shells (dinosaurs, birds) 93R  
 Egypt 106R  
 El Cameron Formation, Oaxaca, Mex-  
 ico 202R  
 El Casco, California 259R  
 “El Gallo” Formation, Baja California  
 del Norte 25L  
 Elk Creek facies, Willwood Formation  
 107R  
 El Kef, Tunisia 21R  
 Ellesmere Island, Canada 108R, 111L, R,  
 133L, 138L  
 El Molino Formation, Bolivia 94L  
 El Salvador 203L  
 Elsinore Fault zone, California 260L  
 endemic (endemism) 13R, 18L, 23L,  
 35L, 37L, 88R, 90L, R, 91L, R, 92R,  
 93R, 94R, 117L, R, 120R, 131R, 178R,  
 187L, 188R, 198L, 199L, 201L, 203R,  
 204L, R, 205L, R, 206L, 214L, 215R,  
 233R, 246L, 274R, 294L, 318R, 321R,  
 323L, 324L, 328R, 331L  
 environmental zone (zonation) 232R  
 Eocene-Oligocene boundary 133L,  
 160R, 164L  
 Eocene Series/Epoch, faunas, rocks 11L,  
 R, 16L, 17L, R, 43L, R, 76R, 79L, 89L,  
 R, 90L, R, 92L, R, 93L, 95R, 106L, R,  
 107R, 112L, 119L, 122L, R, 124L, R,

- 125R, 126L, 129L, 130R, 131L, R, 132L, 133L, 134L, R, 135, 156R, 160L, R, 161R, 164L, 172R, 192L, 317L, R, 318L, 320L, R, 321L, 322, 324L, R, 325, 326L, R, 327L, R, 328R, 330L, 332L
- Eotitanops borealis* Range Zone (Bro; lowest Gardnerbuttean) 113L, 114R
- Epicyon haydeni*-*Hipparion forcei* Assemblage Zone (Cl2/3) 177L
- Equus* (*Equus*) LSD<sub>k</sub> 259L
- Equus* sp. LSD<sub>k</sub> 259L
- Erethizon stirtoni* LSD<sub>k</sub> 259L
- Escavada Member, Nacimiento Formation, New Mexico 45R
- Escavada Wash, New Mexico 48, 74L, 75L
- Esmeralda Formation, Nevada 220L
- Española Basin, New Mexico 198R, 199L, R
- Etchegoin Formation, Kettleman Hills, California 255L
- Etla Member, Suchilquitongo Formation, Oaxaca, Mexico 202L
- Etla Tuff, Suchilquitongo Formation, Oaxaca, Mexico 202L, R, 221L
- Europe (an) 16L, 21L, 24L, 88R, 89L, R, 90L, R, 92R, 94R, 233L, 238L, R, 254R, 255R, 274R, 317R, 324R, 325L, R, 328R, 334R
- Evanston Formation, Wyoming 77L
- Evanston, Wyoming 127R, 128R
- evolutionary first occurrence xiiiR, 88L
- Euceratherium* LSD<sub>k</sub> 259L
- Eurasia(n) 173R, 207R, 209L, 247R, 250L, 254L, 255L, R, 284R, 331L
- Eureka Sound Group, Canada 108R, 109R, 133R, 138L
- extant 232L, 246L, 255R, 272L, 284R, 290L, 291L, 293L, 294R
- extinction(s) 37L, 38L, 89R, 90L, 92R, 160L, 179R, 193L, 194R, 195R, 209L, 212R, 213R, 218L, R, 238R, 244, 252R, 255R, 284R, 286L, R, 291R, 293L, 295L, 320L, R, 334L, 336L, R, 337R
- FAD (First Appearance Datum) xiiR, xiiiL, R, xivL, 13R, 14L, R, 15L, R, 23R, 28L, 35L, 36R, 37R, 44L, 65L, 77R
- Fairpoint Member, Fox Hills Formation, South Dakota 34L
- Fallon County, Montana 26L
- Farisita Formation, Colorado 108L, 131L
- Farrand Channel, Tullock Formation, Montana 51, 70R, 73L
- Fauna, faunal viiL, R, xiiR, xiiiR, xivR, xviiiL, R, 15L, 16L, R, 17R, 21L, R, 22L, R, 23L, R, 24L, R, 25L, R, 26L, R, 27, 28L, R, 32L, R, 34L, R, 35L, R, 36L, R, 37L, R, 38L, 43R, 44L, 45L, R, 46, 49, 50, 51, 53L, R, 61, 62–95L, 106–107L, R, 109L, R, 111L, R, 112L, 113L, R, 115–134L, R, 138L, 157, 158L, R, 160L, R, 161R, 162L, R, 163L, 164L, 166L, R, 167L, 169–219L, R, 232–234L, R, 236, 237R, 238–242L, R, 244, 246–294L, R, 295L, 317L, 318R, 230R, 321R, 324L, R, 325L, 327L, R, 328–329L, R, 330R, 332L, R, 333R, 334–335L, R, 337L, 338R
- faunal facies xiL, 35L, 66L, 173L, 194L, 197L, 318R, 321R, 324L, 328L, R
- faunal turnover 44R, 89R, 90L, 114L, 174R, 176R, 177R, 178L, 193L, 194L, R, 195L, 200R, 209L, R, 211R, 212R, 213L, R, 217L, 218L, 328R, 329L, 334L
- FAUNMAP 288L, 291R, 292L, R, 293R, 294L
- faunule xiiR, 22L, 23L, R, 24L, 25R, 26L, 27R, 36R, 250R, 263L, 280L
- fern spike 320R
- Ferris (=No. 1) Coal Seam, Saskatchewan, Canada 35R
- Ferris Formation, Hanna Basin, Wyoming 16R, 27L, 36L, 50, 53R, 61R, 62L, 65L, 66L, R, 68L
- first appearance(s) 21R, 23L, R, 24R, 28L, 34L, 36R, 44L, 45R, 53L, R, 62–63L, R, 64R, 65R, 67R, 71L, R, 73L, 74L, 75L, 78L, R, 80L, 81L, 82L, R, 83R, 84L, R, 85L, R, 86L, 86R, 87L, R, 88L, 91L, R, 91R, 112R, 113L, 114L, R, 116R, 117L, R, 118L, R, 119L, R, 120L, 122L, 133L, 161R, 162L, R, 163L, R, 164L, R, 165L, 166L, R, 174L, 176L, 179R, 187L, R, 188R, 189L, 193L, 194R, 195L, 196L, R, 200L, R, 204L, 209R, 211L, R, 212L, 214R, 215L, R, 216R, 217L, R, 218L, R, 247L, 250L, 252R, 253L, R, 254R, 255L, 258R, 261R, 268L, 269R, 270R, 271R, 272R, 273L, 274R, 275L, R, 276L, 280L, 283R, 284R, 285L, R, 286L, 288L, R, 295L, 323R, 324L, 325L, 327R, 328L, R, 331L, 332L, R, 324L
- First Division, Barstow succession 176L
- Flagstaff Limestone, Utah 125R
- Flagstaff Peak Formation, Utah 67L
- Flagstaff Rim, Bates Hole, Wyoming 158R, 161R, 162L, R, 163L, R
- Florissant flora, Colorado 327R
- Fly Tuff, Nevada 220L
- FOD (First Occurrence Datum) xiiR, xiiiL, R, xivL, 15R
- Foremost Formation, Alberta, Canada 24L
- Fort Hancock Formation, Texas 263R
- Fort Logan Formation 187R, 188R, 189R
- Fort Peck Reservoir, Montana 25R, 27, 28R, 32L, 34
- Fort Union Formation, Montana, Wyoming 11R, 17L, 45R, 50, 51, 62L, 63L, R, 64L, 67L, 70R, 71R, 73R, 76R, 77L, R, 80R, 85L, R, 107R, 122L, 126R, 132L
- Fort Union Group, Montana, Wyoming 70R, 71R, 77L
- Fossil Basin, Wyoming 77L, 107L, R, 108L, R, 127R, 128R
- Fossil Butte National Monument, Wyoming 128R
- Fowkes Formation, Wyoming 107R, 108R, 128R
- Fox Hills Formation, South Dakota 26R, 27, 32R, 34L, R, 35R
- Frick Laboratory (collections, American Museum of Natural History) New York 164R, 165R, 166R, 188L, 198R, 199L, 285R
- Fraction Tuff, Nevada 220L
- Fraser River, British Columbia, Canada 133L
- Frenchman Formation, Saskatchewan, Canada 26R, 27, 35R, 52, 53L, R, 64R
- Frenchman River, Saskatchewan, Canada 26L
- Friars Formation, San Diego area, California 10L, 9L, 115R, 117L, 118L, 123R, 124L
- Fruitland Formation, New Mexico 24L, 25L
- Galisteo Formation, New Mexico 108L, 109R, 115R, 130R
- Garbani Channel, Tullock Formation, Montana 51, 68R, 73L
- Gardnerbuttean subage (Bro, Bria), Bridgerian mammal age 107R, 113L, R, 114L, R, 115L, 121R, 126L, 135
- Garfield County, Montana 64L
- Gashatan, Asian mammal age (approx. correlative with Clarkforkian) 90R, 91R, 92L
- Gauss magnetic polarity chron 233L, 253L, 255L, R, 256R, 258L, 259L, 261L, R, 262L, 283L, R, 265L, 267L, 268L, 269L, 279L
- geochron 7R, 160R
- geochronologic (unit) viiL, xiR, xiiL, xivL, 1–2L, R, 5, 7, 8L, R, 14L, 15L, 24L, 25R, 43R, 46, 47, 93L, 94R, 95L, 134R, 161R, 169L, 189R, 192R, 208R, 209R, 215R, 217L, 232R, 238L, 292L
- geochronology xiR, xiiL, xiiiL, 1L, 2R, 3L, 7, 8L, 43L, 106L, 134R, 135, 138L, 216R, 232R

- geologic time 1R, 2L, R, 3L, R, 6R, 7L, 8R, 9R, 127L, 209L, 233L, 293L
- Gering Formation, Nebraska 158L, 179R, 187R, 188R, 189L, R, 193L, 209R, 211L
- Germer flora, Idaho 326L, R
- Gila River Valley, Arizona, New Mexico 232R
- Gilbert magnetic polarity chron 233L, 256R, 258L, 259R, 261L
- glacial, glacial stages, glaciation 191R, 232R, 236L, 237R, 238L, 252R, 280R, 283L, 285R, 287L, R, 288L, R, 290R, 291L, R, 315L, R, 317L, R, 318L, 326L, R, 327L, 329R, 330L
- glacial till 237R, 288L, R
- glacio-eustatic (sea level) changes 315R, 317R, 318L
- Glendive, Montana 67L
- Glenn's Ferry Formation, Idaho 256R, 258L, R, 271L
- global xiiiL, R, xiiiL, R, xviiR, 5, 7, 8L, R, 9R, 13R, 18L, 21R, 35R, 36R, 37L, 88R, 89R, 90L, 106R, 134R, 138L, 156L, R, 160L, 166R, 209L, 233L, R, 237L, 315L, R, 316, 317R, 320L, R, 324R, 326L, 327L, 330L, 334L, R, 335R, 338L
- Golden County, Montana 24L
- Golden Valley Formation, North Dakota 108L, 111L, 132L
- golden spike 7, 8L
- Goler Formation, California 53, 62R, 70R, 77L, 81L
- Gondwanan "Stage," Argentina 92R
- Gosport Sand, Claiborne Group, Alabama 108R, 134L
- GPTS (Geomagnetic Polarity Time Scale) xiiiL, 9L, R, 10L, 11L, R, 12R, 22L, 32R, 34L, 65L, 93R, 94L, 138L, 169L, 173L, 209L, 233L, R, 237L, 239L, 254R
- grass, grassland 326R, 327L, 332L, 333R, 334R, 336L, 337L
- Gray Bull beds 107R, 108L
- Graybullian subage (Wa3–Wa5) of Wasatchian mammal age 107L, 111L, 113L, 122L, 130R, 132L, 135, characterized 113L
- Great American Interchange (GAI), (biotic, faunal) 253L, 268R, 269L, 316, 317L, 336, 338L
- Great Basin 176L, 178L, 199R, 212R, 214L, 215R, 218L, 328R, 331L, 332R, 335R, 336R, 337L  
northern 175L–178L, 214L  
southern 169L, 198R–202L, 214L
- Great Divide Basin, Wyoming 107R, 126R, 127L, R, 158R
- Great Plains, North America 169L, 172L, 173L, R, 174R, 176R, 177R, 178L, R, 179R, 187L, R, 188L, R, 189L, R, 190R, 191L, R, 192L, R, 193L, R, 194L, 195L, R, 196R, 197R, 199R, 200L, R, 201L, 202R, 203R, 204L, R, 205L, R, 206L, R, 207L, R, 209L, 211L, R, 212L, R, 214L, 215L, R, 217L, R, 218L, 219L, 237R, 247R, 252R, 253L, 255L, R, 256L, 258R, 264R, 265L, R, 266L, 272L, 273R, 274L, R, 280R, 285R, 287R, 290R, 291L, 331L, 332L, R, 333R, 334R, 335L, 336R, 337L
- greenhouse, icehouse conditions 156R, 315L, 317R, 318L, 320R, 326R, 329R
- Greenland (dispersal) 90R, 239L
- Green River Basin, Wyoming 82L, 85L, 87R, 106R, 107L, 108L, R, 109L, 111R, 113L, R, 114L, R, 115R, 116L, 125R, 126L, 127L, R, 128L, R, 131L, 132L, 135, 136, 138L, 139L
- Green River flora 326L, R
- Green River, Formation, Wyoming 108L, R, 109R, 113R, 114L, R, 115L, 125L, R, 126L, R, 127L, 128L, R
- Green Valley Formation, California 171R
- Grey Tuff, Wilkins Peak Member, Green River Formation 113R, 114L, 128L
- GSSP (Global Boundary Stratotype Section and Point) 7, 8L, 90L, 106R, 134R, 237L
- Guaje ash, Mt. Blanco, Texas 234, 240, 253L, 266R
- Guaje Pumice, Otowi Member, Bandelier Tuff, Jemez Mountains, New Mexico 234, 266R, 271L
- Guanajuato, Mexico 108R, 131R, 201R, 218R, 257
- Hackberry Wash, California 176R
- Hagerman Fossil Beds National Monument 255L, 256R
- Halfway horizon, Duchesne River Formation 112L
- Hand Hills, Alberta, Canada 191R
- Hanna Basin, Wyoming 1L, 16R, 17R, 27L, 50, 61L, 63R, 65L, 66R, 70R, 73L, 75R, 77L, R, 78L, R, 80R, 81R, 82L, 107R
- Hanna Formation, Wyoming 70R, 77L, R
- Hannold Hill Formation, Texas 108L, 111L, 129L
- Haplomylus-Ectocion* Range Zone 17R
- harmonious (vs disharmonious) faunas 290R
- Harrison Formation, Nebraska 191L, R, 193R, 194L, R, 196R, 197L, 198R, 201L, 204L, 205R, 208R, 209R, 211R, 212L, R
- Hartford Ash 234, 237R, 240, 281R
- Hart Mine Formation, New Mexico 108R, 130R, 131L
- Hatchetigbee Formation, Alabama 108L, 134L
- Houghton Astrobleme, Devon Island, Canadian Arctic Islands 192R, 338R
- Hawthorn Formation, Florida 206L
- Hawthorn Group, Florida 206L, 207L
- Haystack Mountain, Wyoming 127R
- Haystack Valley, John Day region, Oregon 187L
- Haystack Valley Member, John Day Formation, Oregon 179L, R, 187L
- Hector Formation, California 176R, 219R
- Hell Creek Formation, Montana, North Dakota, South Dakota 24R, 25R, 26L, R, 27, 28R, 32L, R, 34L, R, 36L, R, 53L, R, 62L, 63L, R, 64L, 95R
- Hell Creek, Montana 32L
- Hemicyon* Tuff, Barstow Formation, California 215R, 219R
- Hemingfordian mammal age**, faunas, rocks 172L, 173R, 174R, 175L, R, 176R, 178L, R, 187L, R, 188R, 189L, 191R, 192L, R, 195L, R, 196L, 198R, 199L, 201L, R, 202L, 204R, 206R, 208L, 209L, 211R, 212R, 213L, R, 214L, R, 332R, 333L, 334L, R  
definition and characterization 212R, 213L, R  
zonation 213L, R
- Hemingford Group, Nebraska 212R
- Hemithlaeus kowalevskianus*–*Taeniolabis taoensis* Zone 16R
- Hemphill beds, Texas 200R
- Hemphillian mammal age**, faunas, rocks 175L, 177R, 178L, R, 195R, 198L, 200R, 201L, R, 203R, 207R, 216L, 217L, R, 218L, R, 219L, 232L, 244, 247R, 248, 249R, 250R, 251L, R, 252L, 253L, 256L, 257, 259R, 260L, R, 262L, 263L, 264L, R, 265L, 267R, 332R, 333R, 335R, 336L, 337L, R, 338L  
definition and characterization 217R–219L  
zonation 217L–219L  
*Paronychomys-Borophagus littoralis* Assemblage Zone (Hh1) 177L
- Hemphillian-Blancan boundary 178L, 251L–252L, 260R, 263R, 264R
- Hemphill Member, Ogallala Formation 217R
- Hendry Ranch Member, Wagon Bed Formation, Wyoming 109L, R, 121R
- Henry Mountains, Utah 23R
- Hepburn's Mesa, Montana 188L
- Hepburn's Mesa Formation, Montana 190R, 191L, R
- Heptodon* Range Zone 17R

- hiatus 2, 10R, 23R, 89R, 126R, 172R, 174L, 177L, 194L, 197L, R, 199R, 211R, 215R, 216R, 326L
- Hidalgo, Mexico 251R
- Hickey Mountain Limestone, Bridger Formation, Wyoming 115R
- Hickey Mountains, Wyoming 128L
- high latitudes; independent chronology needed 249R
- High resolution chronology, chronostratigraphy viiL, 1R, 8L, 156L, 165R, 167L
- Himalayan-Tibetan plateau 331R, 336L
- Hippotherium* Datum 13R, 335L
- HO (Highest Stratigraphic Occurrence) xiiiL, R, xivR, 71L, 74R, 174L
- Hoback Basin, Wyoming 49, 77L, 82L, 83R, 85L, R, 87R, 108L, 111L, 127R, 128R
- Hoback Formation, Wyoming 47, 77L
- Holarctic 90L, 292R, 293R, 323L, 324R, 325R, 326L, 338L
- Holocene 233L, 238L, R, 247R, 269R, 286R, 288R, 290L, R, 291L, 292R, 293R, 294L, R, 295R
- Honduras 203L, 217L
- Hoploscaphites birkelundi* Range Zone 34R
- Horseshoe Canyon Formation, Alberta, Canada 259L
- HSD (Highest Stratigraphic Datum) xiiiL, 259L
- HSD<sub>k</sub>, highest known stratigraphic occurrence 259L
- Huckleberry Ridge Ash, Yellowstone, Wyoming, western U.S. 234, 240, 261L, 263R, 265R, 266L, 273R, 279R, 280R, 44
- Heuco Bolson, Texas 257, 263L
- Huerfano A 131L
- Huerfano B 121R, 131L
- Huerfano Formation, Colorado 108L, R, 112R, 114R, 131L, R
- Huerfano Park, Colorado 113L, 114L
- Hurlbut Ash, Valentine Formation Nebraska 195L, 200L, 215R, 220R
- Hypertragulus calcaratus* Interval Zone (earliest Orellan) 164R
- hypodont(y) 247L
- Icehouse World *see* greenhouse
- ICS (International Commission on Stratigraphy) xiiiL, 233R
- IGCP 90R
- Illinoian glacial stage (Pleistocene) 237R, 238L, 285R
- IMBS (Integrated magnetobiostratigraphic scale) xiiiL, 9L, R, 10R, 11R
- immigrant, immigrate, immigration xivL, xviiR, 7, 13R, 17R, 18L, 37R, 78R, 85R, 91L, 92R, 114R, 174L, 193L, 194R, 195L, 196R, 199L, 204R, 206L, 207R, 209L, R, 210, 211R, 212L, 213L, R, 214L, R, 216R, 218L, R, 253L, 254L, R, 255R, 268L, R, 269L, 274R, 275L, R, 276L, 284R, 321R, 323L, R, 324L, R, 325, 327R, 328R, 331R, 332R, 333L, 334L, R, 335L, R, 337R, 338L
- Imperial Formation, California 258R
- index fossils (taxa) 16R, 44L, 62R, 63L, R, 64L, 65R, 67R, 68L, 70L, 71R, 72L, 73R, 75L, 76L, 78L, R, 79R, 80R, 81R, 82R, 83R, 84L, R, 85R, 86L, 87L, R, 88L, 112R, 117L, 118R, 119L, 133L, 162L, R, 163L, R, 165R, 167L, 209R, 249R
- intercontinental (correlation, dispersal) 44R, 88R, 138L, 139L, 209L, 254L, 324R
- interglacial(s), times 317L
- Interior Paleosol, South Dakota 158L
- International Geological Congress 236R, 237L
- International Geological Correlation Project 237L
- International Stratigraphic Guide (ISG) 43R, 44L
- International Union for Quaternary Research (INQUA) 237L, 238R
- Interval Chron xiiiL, xivL, 4, 15L
- Interval Subzone 79L, R, 80L, 84L, R, 85R, 87L
- Ti6 interval subzone 79L, R, 80L, 84L, R, 85R, 86R
- Cfi interval subzone 84L, R, 86R, 87L
- Interval Zone xiL, R, xiiiL, R, 4, 5, 6L, R, 15L, 16L, R, 17L, R, 35L, 44L, 45R, 53R, 61L, 63L, R, 64L, R, 65L, R, 66L, R, 67L, R, 68L, R, 69L, R, 70R, 71L, R, 72L, R, 73L, R, 74L, R, 75L, R, 76L, 79L, 80R, 87R, 91L, 93R, 95R, 162R, 163L, R, 164R, 165L, R, 166L
- Archaeolambda* Interval Zone 91L
- Asiostylops* Interval Zone 91R
- Bemalambda* Interval Zone 91L
- Puo interval zone 53R, 61L
- Pu1 interval zone 44L, 45R, 49–52, 53R, 54–60, 61L, 63–65L, R, 66R, 68L, 80L
- Pu2 interval zone 45R, 48–52, 54–60, 61L, 63–69L, R, 71L, 93R, 95R
- Pu3 interval zone 45R, 48–52, 54–60, 61L, 63L, R, 64R, 65L–69L, R, 70L, 71L, 73L, 95R
- Sinostylops* Interval Zone 91R
- T-P interval zone 67R
- To1 interval zone 71L, R, 72L, R, 73L, R
- To2 interval zone 72L, R, 73R, 74L, R, 91L
- To3 interval zone 72L, R, 73L, 74L, 75L, R, 76L, 79L, 80R
- intracontinental (correlation, dispersal) 138L, 254L
- iridium (anomaly) 9L, 21R, 28R, 32L, 35R, 43R
- Iron Lightning Member, Fox Hills Formation, South Dakota 27, 32R, 34L, R
- Iron Springs Formation, Utah 23L
- Irvington, Alameda County, California 269L
- Irvingtonian mammal age**, faunas, rocks 233L, 235, 236, 239L, 246L, R, 247R, 248, 249L, 250L, R, 252R, 256L, 257, 258R, 259R, 260L, 261L, 262R, 263L, 264L, R, 265R, 266R, 267L, R, 268L, 269L, R, 270L, R, 271R, 272L, R, 273L, R, 274L, R, 275L, R, 276L, R, 285L, 288L, 289L, R, 292L, R, 293L, R, 294L, R, 295L, R
- arrival of *Mammuthus* 270R–271R
- definition and characterization 269R–270R
- geographic distribution 276R–284R
- California 276R–278R
- Eastern Caves 284L, R
- Florida 283L, R
- Great Basin 278R
- Great Plains 280R–283L
- Rocky Mountain region 279L–280L
- Snake River Plain 276R
- Southwestern U.S. 280L, R
- Mexico 280R
- historical treatment 269L
- small mammal definition 271R, 272L
- zonation 273L–276L
- arvicoline rodents 274L, R
- Irvingtonian I subage 274R, 275L, R
- Irvingtonian II subage 275L, R, 276L
- Irvingtonian III subage 276L, R
- Sappan subage 273L, R
- Cudahyan subage 273L, R, 274L
- Sheridanian subage 273L, R, 274L
- isochron, isochroneity, isochronous xiiR, 3R, 6R, 7R, 8L, 14L, R, 15R, 215L, 249L, 293L
- Itaboraian mammal age 94L
- Itaboraian mammal subage 95L
- Itaboraí district, southeastern Brazil 92R, 93L, 94L
- Italy, Italian 237L, R, 238L, 295R
- IUGS (International Union of Geological Sciences) xiiiL, 237L
- Ixtapa Formation, Chiapas, Mexico 203L, 214R
- Jackrabbit Trail, California 259R
- Jackson Group, Arkansas 108R, 134L

- Jalisco, Mexico 251R  
 Jaramillo magnetic polarity subchron of Matuyama Chron 233L, 240, 258R, 259R, 263L, 269L, 279R, 280L  
 Javelina Member, Tornillo Formation, Texas 62L, 69R  
 Jaw Face section, Wellsch Valley, Saskatchewan, Canada 281L  
 Jbel Guersif Formation (Thanetian), Morocco 92L  
 Jefferson Basin, Montana 160L  
*Jeletzkytes nebrascensis* Range Zone 34L  
 John Day 'fauna,' Oregon 179R, 187L, 188R, 212R, 332L  
 John Day Formation, Oregon 172R, 173L, 174R, 179L, R, 187L, R, 189L, R, 193L, 204L, R, 206L, 209R, 211L, R, 327L  
 John Day region, Oregon 178R, 179L, 188R, 327R, 331L, 332L, 333R  
 John Henry Member, Straight Cliffs Formation, Utah 23L  
 Jones Spring, Missouri (early *Bison*) 286L, 289, 295L  
**Judithian mammal age**, faunas, rocks 21L, R, 23R, 24L, R, 25L, 28L, 37R, 318R  
 definition and characterization 23R–24R  
 Judith River Formation, Montana 23R, 24L, R  
 Judith River Group, Alberta and Saskatchewan, Canada 24L  
 Juntura Formation, Oregon 220R
- Kaena magnetic polarity subchron of Gauss Chron 240, 256R, 258L, 259L, 262L, 263L  
 Kaiparowits Formation, Utah 24L  
 Kaiparowits Plateau, Utah 23R  
 Kansan glacial stage (Pleistocene) 237R, 238L, 281L  
 Kansan till 237R  
 KE (kaolinitic event, at LPTM) 320L, R  
 Kemp Clay Formation, Texas 28L  
 Kilgore flora, Nebraska 334R  
 Kimbeto wash, San Juan Basin, New Mexico 45L, R, 48, 66R, 71L, 72R, 73R, 74L  
 Kinney Rim Member, Washakie Formation, Wyoming 108R, 115L, 127R  
 Kirkwood Formation, New Jersey 207R, 208L, R  
 Kirtland Formation, New Mexico 24L, 25L, 27, 28L, 34, 36R  
 Kishenehn Formation, British Columbia, Canada 109R, 132R  
 Kisinger Lakes flora, Wyoming 326R  
 Kneehills Tuff, Scollard Formation, Alberta, Canada 32R
- Knight Formation, Wyoming 107R, K-T boundary 9L, 21R, 32L, 35R, 36L, R, 37L, 38L, 320R  
 Kutz Canyon, New Mexico 48, 71R, 72L, R, 73R, 74
- LaBarge, Wyoming 108L, 128L  
 LAD (Last Appearance Datum) xiiiL, R, 14L  
 Laguna Umayo (area), southeastern Peru 92R, 93L, R  
 Lake Titicaca, Peru 93L  
*Lambdotherium* Range-Zone (Lost-cabinian, Wa-7) 113R, 121R  
 Lance Creek, Wyoming 25R, 34L, R  
 Lance Formation, Wyoming 25R, 26R, 27L, 34L, 36L  
**Lancian mammal age**, faunas, rocks 16R, 21L, R, 22L, 24R, 25–28L, R, 32L, R, 34–37L, R, 38L, 45R, 46, 53L, R, 61L, 62L, 63L, 64L, R, 92R, 318R, 320R, 321L, R  
 definition and characterization 28L, R  
 Lancian-Puercan boundary 21R, 35R, 36L, R, 62L, 64L  
 land mammal age(s) viiL, viiiL, xivL, R, 8R, 15R, 21L, R, 22L, 23–25L, R, 26R, 27, 28R, 35L, 36L, R, 43L, R, 76R, 90R, 91L, R, 93L, 95L, 106L, 139L, 156L, R, 158L, 166R, 169R, 175R, 232L, 240, 244, 246R  
 Laney Shale Member, Green River Formation, Wyoming 113R, 114R, 115L  
 La Palca, Bolivia 94L  
 Lapilli Tuff, California 220L  
 LaPoint Ash, LaPoint Member, Duchesne River Formation, Utah 118R, 119L, 136  
 LaPoint horizon, Duchesne River Formation 112L  
 LaPoint Member, Duchesne River Formation, Utah 109R, 112L, 118R, 119L, 126R  
 Laramie Basin, Wyoming 108L, 111L, 132R  
 Laramie Formation, Colorado 27R, 34  
 Laramie Range, Wyoming 158R  
 Laredo Formation, Texas 109L, 118R, 134L  
 Largo beds, New Mexico 109R, 130L, R  
 last appearance(s) xiiiR, 5, 16R, 23R, 28R, 36R, 44L, 62R, 63L, R, 65R, 67R, 69R, 71R, 72L, 73R, 75L, 76L, 78R, 79R, 60R, 81R, 82R, 83L, 84L, R, 86L, 87L, R, 88L, 112R, 113L, 114R, 115L, 117R, 118L, 119L, R, 163R, 164L, R, 165L, 166L, R, 176L, 187L, 189R, 190R, 196R, 199R, 200R, 204L, 207R, 209R, 211L, R, 212L, 213R, 215L, 216L, 217R, 218R, 219L, 247R, 249R, 252R, 253R, 256L, 268L, 273R, 276R, 283R, 324R, 327R, 331L  
 Late Cretaceous, faunas, rocks viiL, 16R, 21–22L, R, 23L, 26R, 32L, 36L, R, 37L, R, 38L, 62L, 64R, 92R, 93R, 315R, 317–318L, R, 319, 320–321L, R, 323L, 326R  
 Lava Creek B Ash (Pearlette-O) 234, 237R, 240, 273R, 280R, 281R, 282L, R, 285R  
 Laverne Formation, Oklahoma 197R, 198L  
 Lawlor Tuff, Etchegoin Formation, California 219R, 234, 255L  
 Lebo Formation, Montana 70R, 73L, R, 80R  
*Leptauchenia* beds, Brule Formation, Big Badlands, South Dakota 159, 165R, 166L  
*Leptauchenia major* Interval Zone (early Whitneyan) 165R  
*Leptomeryx mammifer* Interval Zone (middle Chadronian) 163L  
*Leptomeryx yoderi* Interval Zone (late early Chadronian) 168R  
*Lepus* LSD<sub>k</sub> 259L  
 Leslie Gulch Tuff, Sucker Creek Formation, Oregon 187R  
*Lignimus-Tardontia* Zone (Ba2), Hepburn's Mesa Formation, Montana 191L, R  
 Lineage Chron xiiiR, xivL, 4, 15L  
 Lineage Zone xiR, xiiiR, 2, 4, 5, 6L, 15L, 16L, 17L, 44L, 78R, 79–84L, R, 86R, 87L, R, 89L, 91R  
 T<sub>i1</sub> lineage zone 79L, 80L, R, 81L, 82L, 83R  
 T<sub>i2</sub> lineage zone 78R, 79L, 80L, R, 81L, R, 82L  
 T<sub>i3</sub> lineage zone 79L, 80L, 82L, R, 83L  
 T<sub>i4</sub> lineage zone 79L, 80L, 82R, 83L  
 T<sub>i5</sub> lineage zone 79L, 80L, 83L, R, 84L, 91R  
 T<sub>i6</sub>-C<sub>f1</sub> lineage zone 79L, R, 80L, 84L, R, 86R, 87L, 89L  
 C<sub>f2</sub> lineage zone, 86R, 87L, R  
 lithologic 2, 3L, 10R, 13L, 15R, 18L, 34, 45L, R, 70L, 94R, 107L, R, 111R, 112L, 113R, 115R, 116L, 124R, 128L, 131L, R, 158R, 160R, 187L, 196L, R, 200L  
 lithostratigraphic (unit) viiL, xviiiL, 2L, R, 3L, R, 7, 10L, 13L, 106R, 107R, 111R, 112L, 123R, 158L, 159, 161R, 162L, 163R, 164L, 165R, 170L, 179L, 187L, 193L  
 lithostratigraphy xivL, 3L, 156R, 160R, 164L, 166L, 172L, R, 176R, 179L, 193L, 197L, 198R, 209R



- Little Cove Point Unit, basal St. Mary's Formation, Maryland 208L
- Llajas Formation, California 124L, R
- Llano Estacado, Texas 266R
- Llaves Member, San Jose Formation, New Mexico 130R
- LMA (land mammal age) 106L, R, 107L, 111L, R, 112L, 113L, 114R, 115R, 119L, 128L, 138L
- LO (Lowest Stratigraphic Occurrence) xiiR, xiiiR, xivL, R, 6L, 14L, 15R
- Lobato Basalt, New Mexico 221L
- Local Fauna(s)** xiiR, xiiiR, 169R, 170L
- 111 Ranch fauna (Blancan, Blancan V), Arizona 244, 255R, 257, 261L, R, 263L, 268L, R
- Adams Fauna (Irvingtonian, Sheridanian; Rancholabrean), Kansas 274L, 288R
- Agate Springs Local Fauna (Arikareean, Ar4), Nebraska 209R, 212R
- Aguila Fauna (Blancan), Texas 263R
- Ahearn Member faunas (late early Chadronian), South Dakota 162R
- Airstrip Local Fauna (Chadronian), Texas 130L, 160L
- Alamos Canyon Local Fauna (Arikareean, Ar2), California 172R
- Anceney Local Fauna (Barstovian, Ba1), Montana 190R, 191L
- Angus Fauna (Irvingtonian, Sheridanian), Nebraska 274L, 285R
- Archusa Marl Fauna (Uintan), Mississippi 134L
- Argonaut Fauna (early Blancan), Kansas 265L
- Aries A Fauna (latest Blancan; Sappan), Kansas 265R, 266L, 270L, 273R, 295L
- Aries B Fauna (latest Blancan/Irvingtonian), Kansas 265R, 270L, 295L
- Arroyo Seco Fauna (middle Blancan), California 258R
- Artesia Road Fauna (Blancan), Arizona 261R
- Ash Hollow Formation faunas (Clarendonian-Hemphillian), Nebraska 177R, 195R, 216R
- Axtel Local Fauna (Hemphillian, Hh4), Texas 200R
- Ash Springs Local Fauna (late Chadronian), Texas 129R, 130L, 163L
- Balm Creek Member Fauna (Arikareean, Ar3), Oregon 187L
- Barker's Ranch Local Fauna (late Hemingfordian, He2), California 172L
- Barstow Fauna (Barstovian, Ba2), California, 171R, 174L, 176L, R, 177R, 215R
- Bear Creek Local Fauna (Clarkforkian, Cf1) 50, 76R, 85R, 87L
- Beaver Fauna (Clarendonian, Cl2), Oklahoma 197R, 198L
- Beaver Quarry Local Fauna (Clarendonian, Cl2), Oklahoma 198L
- Beck Ranch Fauna (Blancan), Texas 250L, 257, 267L
- Bee Canyon Landfill Fauna (Hemingfordian, He1), California 174R
- Belen Fauna (Blancan), Albuquerque Basin, New Mexico 262R
- Benson Fauna (early Blancan), Arizona 261L, R
- Berends Fauna (Irvingtonian, Sheridanian), Oklahoma 274L
- Big Spring Canyon Local Fauna (Clarendonian, Ca1), South Dakota 216L
- Birch Creek Fauna (late Blancan), Idaho 258L
- Black's Beach Local Fauna (late Bridgerian/early Uintan), California 108R, 123L
- Black Butte Fauna (late Blancan), Idaho 258L
- Black Hawk Ranch Local Fauna (Clarendonian, Montediablan, Cl3), California 171L, R, 172R
- Blacktail Local Fauna (Lancian), Montana 26L, 27
- Blacktail Deer Creek Fauna (Arikareean, Ar1/2), Montana 189L
- Blanco Fauna (early Blancan, Rexroadian), Texas 249L, R, 253L, R, 257, 266R, 267L, 268R
- Blufftop Fauna (early Blancan), Washington 256R
- Bolero Lookout Local Fauna (Hemingfordian, He1), California 174R
- Bone Valley Fauna (Hemphillian, Hh4) Florida 204R
- Borchers Fauna (late Blancan), Kansas 234, 245, 265R, 266L, 272R, 291L
- Boron Local Fauna (Hemingfordian, He1), California 176R
- Brea Canyon Local Fauna (late Uintan), Ventura County, California 109L, 124L, R, 125L
- Broadwater Fauna (early Blancan, Rexroadian; Arvicoline Zone II), Nebraska 253R
- Brooksville 2 Local Fauna (Arikareean, Ar2), Florida 205R
- Buckeye Creek Local Fauna (Blancan, Bl1), Nevada 178L, 257, 260L
- Buckhorn Fauna (Blancan), New Mexico 257, 262R
- Buckman Hollow Local Fauna (Clarkforkian, Cf2), Wyoming 49, 87R
- Buda Local Fauna (Arikareean, Ar2), Florida 205L, R, 206L
- Buis Ranch Local Fauna (Hemphillian, He3/4), Oklahoma 198L, R, 200R
- Burkeville Fauna (Barstovian, Ba2), Texas 214R
- Burge Fauna (Clarendonian, Cl1), Nebraska 177R, 195R, 216L, R, 217L
- Butler Spring Fauna (RanchoLabrean) 288R, 291L
- Cabbage Patch Fauna (Arikareean, Ar1/2), Montana 187R, 188R, 189L, R
- Calgary 2E Local Fauna (Torrejonian, To3?), Alberta, Canada 52, 75R
- California Wash Fauna (late Blancan), Arizona 261L
- Candelaria Local Fauna (Uintan, Ui3), Texas 109L, 118R, 130L
- California Oaks Fauna (Blancan), California 265R
- Camp Cady Fauna (RanchoLabrean), California 286L, 289
- Cape Deceit Fauna (Irvingtonian), Alaska 257, 270R, 275R, 280L, 285L
- Carlton Bar Fauna (Irvingtonian), Alberta, Canada 270R
- Casa Blanca Local Fauna (late Uintan), Texas 109L, 118R, 134L, R
- Castle Butte Fauna (late Blancan), Idaho 258L
- Castolon Fauna (Arikareean, Ar3), Texas 201L, R, 211R
- Cathedral Bluffs Fauna, (Wasatchian, Wa-7), Wyoming 128L
- Cathedral Cave Fauna (Irvingtonian, ?Cudahyan), Nevada 272L, R, 274L, 275R, 276L, 277, 278R, 280L, 294L
- Cedar Ridge Local Fauna (Whitneyan), Badwater Creek area, Wyoming 160L
- Cedar Run Local Fauna (Arikareean, Ar3), Texas 204L, R, 205R
- Cedazo Fauna (late Blancan/Irvingtonian), Aguascalientes, Mexico 257, 264R, 270L, 289, 295L
- Cerrillos Local Fauna (Wasatchian, Wa-5/Wa-6?), New Mexico 108L, 130R

**Local Fauna(s)** (*continued*)

- Cernay faunas (Cernasian, Thanetian; ?Ti6-Cfi equivalent, in part), France 89L, R
- Chattin Hill Fauna (late Blancan), Idaho 258L
- Cheetah Room Fauna (Irvingtonian), Pennsylvania 284L
- Chesapeake Bay Fauna (Barstovian), Maryland 207R, 208L, R
- Circle Local Fauna (Tiffanian, Ti4), Montana 51, 83L
- Cita Canyon Fauna (Blancan, Blancan III), Texas 244, 255R, 257, 266R, 267L, 268R
- Claw Butte Local Fauna (Lancian), Montana 26L
- Clarendon Local Fauna (Clarendonian), Texas 216L
- Clarks Fork Fauna (Clarkforkian, Cf2), Wyoming 76R, 84R
- Cochrane II Local Fauna (Tiffanian, Tii), Alberta, Canada 52, 76R, 80L
- Coffee Cup Local Fauna (Chadronian), Texas 129R
- Coffee Ranch Local Fauna (Hemphillian, Hh2), Texas 200R, 217R
- Coleman 2A Fauna (Irvingtonian), Florida 277, 282R, 283R
- Comanche Point Local Fauna (Clarendonian, Cl3, Montediblan) 172L, R
- Conard Fissure Fauna (Irvingtonian, Cudahyan), Arkansas 245, 274L, 277, 284L
- Concha Fauna (Blancan, Blancan I), Chihuahua, Mexico 252L
- Cook Ranch Local Fauna (Orellan), Montana 160L
- Corinto Fauna (Hemphillian, Hh1), El Salvador 201L, 203L, R
- Coso Mountains Fauna (middle Blancan), California 257, 259R
- Cotter Channel Fauna (early Duchesnean), Texas 109R, 129R
- Country Club Fauna (Blancan), Arizona 262L
- County Line Fauna (Irvingtonian), Illinois 275R
- Courtland Canal Fauna (Irvingtonian), Kansas 281R
- Cow House Slough Local Fauna (Arikareean, Ar2), Florida 204R, 205L, R, 206L
- Cragin Quarry Fauna (Rancholabrean), Kansas 288R, 291L
- Cragin Quarry Local Fauna (Clarendonian, Cl), Oklahoma 198L
- Crazy Johnson Member fauna (middle Chadronian), South Dakota 163L
- Crystal River Fauna (Irvingtonian), Florida 277, 283L
- Cudahy Fauna (Irvingtonian), Kansas 237R, 245, 270R, 281L, R, 282L, 291L
- Cumberland Cave Fauna (Irvingtonian, Cudahyan), Maryland 245, 274L, 275L, 277, 284L
- Cunningham Hill Fauna (Barstovian, Ba2), Wyoming 191R
- Curtis Ranch Fauna (latest Blancan), Arizona 261L, R, 270L, 280L
- Dad Local Fauna (Wasatchian, Wa7), Wyoming 108L, 127L
- Deadman's Creek Fauna (Irvingtonian, Cudahyan), Texas 273R, 282R
- Deep River Fauna (Barstovian, Ba1), Montana 188L, 189R, 190L, R, 191L, 213R, 214R
- Deer Park Fauna (Blancan, Blancan III), Kansas 255L
- De Soto Shell Pit Fauna (latest Blancan), Florida 244, 267R, 268L, 270L, 283L, 295L
- Devils Nest Airstrip Fauna (Hemphillian, Hh4), Nebraska 251R
- Diamond O Local Fauna (Duchesnean/Chadronian), Montana 109R, 134R
- Diss Local Fauna (Torrejonian, To3?), Alberta, Canada 52, 75L, R
- Dixon Fauna (late Blancan, Senecan; Arvicoline Zone III), Kansas 253L, R
- Doe Spring Canyon Fauna (Barstovian, Ba2), California 174L
- Donnelly Ranch Fauna (Blancan), Colorado 257, 267L, 268R
- Dragon Local Fauna (To1), Utah 49, 62R, 70L, R, 71L, R, 72L, 73L, R
- Douglass Draw Local Fauna (late Uintan), Montana 109L, 134R
- Duncan Fauna (Blancan), Arizona 257, 261R
- East Lake Local Fauna (early Arikareean), California 174R
- East Pilgrim 11 Local Fauna (Arikareean, Ar4), Wyoming 191R
- East Pilgrim 5 Local Fauna (Hemingfordian, He1), Wyoming 191R
- El Casco Fauna (latest Blancan/Irvingtonian), California 259R, 260L, 270L, 272L, 278L, 295L
- El Golfo Fauna (Irvingtonian), Sonora, Mexico 251L, 277, 280R, 285L
- El Gramal Fauna (Barstovian?, Ba2?), Oaxaca, Mexico 202R
- Elsinore Fault Zone Fauna (Irvingtonian), California 260L, 277, 278R
- Emerald Lake Fauna (Arikareean, Ar2), Wyoming 191L
- Eubanks Fauna (Barstovian, Ba1), Colorado 197R
- Fallen Angel Fauna (early Blancan), Kansas 265L
- Farmingdale Fauna (Arikareean, Ar3), New Jersey 208R
- Farrand Channel Local Fauna (To1), Montana 51, 70R, 73L
- Faunule A (Blancan), Mesilla Basin, New Mexico 263L
- Faunule B (Blancan), Mesilla Basin, New Mexico 263L
- Faunule C (Irvingtonian), Mesilla Basin, New Mexico 250R, 263L, 280L
- Feltz Ranch Fauna (Hemphillian, Hh1), Nebraska 195R
- Fish Creek Fauna (Blancan, Blancan V), Alaska 255R, 257, 272R
- Fish Lake Valley Fauna (Clarendonian, Cl2), Nevada 178L, 216L
- Fish Springs Flat Fauna (late Blancan), Nevada 257, 260L
- Flat Creek Local Fauna (Lancian), Montana 25R, 28L
- Flatiron Butte Fauna (late Blancan), Idaho 258L
- Flat Tire Fauna *see* 111 Ranch Fauna
- Fort Logan Fauna (Arikareean, Ar1/2), Montana 191L
- Fossil Bush Local Fauna (Orellan, Or2?), Saskatchewan, Canada 331L
- Four Mile Fauna (Wasatchian), Wyoming 108L, 126R
- Fox Canyon Fauna (early Blancan, Arvicoline Zone II), Kansas 253R, 265L, 267L
- Franklin Phosphate Pit 2 Local Fauna (Arikareean, Ar2), Florida 205R
- Friars Formation fauna (Uintan), California 109L, 115R, 117L, 118L, 123R, 124L
- Froman Ferry Fauna (latest Blancan), Idaho 245, 256R, 257, 258R, 260R, 272R, 276R, 295L
- Fullerton Fauna (Irvingtonian), Nebraska 250R
- Fyllan Cave Fauna (Irvingtonian), Texas 277, 282R, 283R
- Garbani Local Fauna (Puercan, Pu3), Montana 51, 68R, 69L, R, 73L

- Gas Tank Local Fauna (Puercan, Pu2?), Utah 49, 67L
- Gilliland Fauna (Irvingtonian, Cudahyan), Texas 250R, 273R, 277, 281R, 282L
- Gordon Fauna (Irvingtonian, Sheridanian), Nebraska 274R, 285R
- Gracias Fauna (Hemphillian, H1), Honduras 201L, 203L, R
- Grand View Fauna (late Blancan, Senecan; Arvicoline Zone III), Idaho 244, 253L, R, 257, 258L, 269L
- Grayson Ridge Fauna (Tiffanian, T11?), Wyoming 50, 77R
- Green Hills Fauna (Barstovian, Ba1), California 174L, 175R, 176L, 214R, 215L
- Gryde Local Fauna (Lancian), Saskatchewan, Canada 26R, 27
- Hackberry Fauna (Hemingfordian, He1), California 176R
- Hagerman Fauna (early Blancan, Rexroadian), Idaho 244, 253L, 255L, R, 256R, 257, 258L
- Haile 7C Fauna (Blancan), Florida 257, 267R, 269L
- Haile 15A Fauna (Blancan), Florida 257, 267R, 268L, R
- Haile 16A Fauna (latest Blancan/Irvingtonian), Florida 245, 267R, 268R, 270L, 283L, R, 295L
- Haile 21A Fauna (Blancan), Florida 277, 283L
- Hainin Local Fauna (Danian), Belgium 88R
- “Halfway Fauna” (Duchesnean), Utah 118R
- Halfway Fauna (Uintan, Ui-3), Utah 112L, 126R
- Halfway Hill Fauna (Tiffanian, T11?), Wyoming 50, 77R
- Hall Ash Fauna (Irvingtonian), Kansas 240, 281R
- Hamilton Cave Fauna (Irvingtonian), West Virginia 244, 277, 284L, 285L
- Harrison Formation fauna (Arikareean, Ar3), Nebraska 191L, R, 194L, R, 197L, 198R, 204L, 205R, 209R, 211R, 212L
- Hartman Ranch Local Fauna (Uintan/Duchesnean), California 109R, 124R, 125L
- Hatchetigbee Bluff Local Fauna (Wasatchian), Alabama 108L, 134L
- Hay Springs Fauna (Irvingtonian, Sheridanian), Nebraska 274L, 285R
- Hemphill Local Fauna (Hemphillian, He2/3), Texas 200R, 217R
- Hell’s Hollow Local Fauna (Puercan, Pu1), Montana 51, 63R, 64L
- Hepburn’s Mesa Fauna (Barstovian, Ba1/2), Montana 188L, 190R, 191L, R
- Hiatt Local Fauna (Puercan, Pu2), Montana 51, 65R, 67L, R
- Hidden Treasure Springs Fauna (Hemingfordian, He2), California 173R
- Higgins Local Fauna (Hemphillian, Hh1), Texas 198L, 217R
- Hill County Local Fauna (Judithian), Montana 24L
- Holloman Gravel Pit Fauna (Irvingtonian), Oklahoma 250R, 277, 281R, 284R
- Horse Room Fauna, Salamander Cave (Irvingtonian), South Dakota 281L
- Hough Draw Local Fauna (late Uintan), Montana 109L, 134R
- Hudspeth Fauna (Blancan), Texas 257, 263R, 268R
- Huerfano B fauna (Gardnerbuttean, Br1a), Wyoming 121R, 131L
- I-95 Local Fauna (Whitneyan), Florida 160L
- Inglis Fauna (Blancan), Florida 264R
- Inglis 1A Fauna (latest Blancan), Florida 245, 257, 267R, 268L, 269L, 270L, 283L, 285R, 289, 295L
- Inglis 1C Fauna (latest Blancan), Florida 245, 257, 267R, 268L, 270L, 283L, 295L
- Indian Meadows Fauna (Wa-2/Wa-3) Wyoming 108L, 120R
- Iron Canyon Fauna (Clarendonian, Cl1/2), California 177L, R, 217L
- Irvington Fauna (Irvingtonian), California 269L, R, 272L, 274L, 277, 278L
- Itaboraí Fauna (“middle” Paleocene), Brazil 93L, 94L, R
- Ixtapa Local Fauna (Barstovian, Ba1), Chiapas, Mexico 203L, 214R
- Jackass Butte Fauna (late Blancan), Idaho 258L
- Jacks Channel Local Fauna (Puercan, Pu1), Montana 51, 63R, 64L
- Java Fauna (latest Blancan; Sappan), South Dakota 245, 257, 265R, 267L, R, 270L, 272L, 273R, 276L, 280L, 295L
- Jeff’s Discovery Local Fauna (late Uintan), San Diego area, California 124L
- Jinglebob Fauna (Rancholabrean), Kansas 288R, 291L
- Jones Fauna (Rancholabrean), Kansas 288R, 291L
- Kanopolis Fauna (Irvingtonian, Sheridanian), Kansas 245, 274L, 283L, 283R, 289
- Kealey Springs Local Fauna (Arikareean, Ar2), Saskatchewan, Canada 192L
- Keefer Hill Local Fauna (Tiffanian, T11), Wyoming 50, 80L
- Kentuck Fauna (Blancan; Sappan), Kansas 245, 267R, 273R
- Keota Fauna (Barstovian, Ba2), Colorado 192R, 197R
- Kew Quarry Local Fauna (Arikareean, Ar1), California 173L
- Kishenehn Fauna (Duchesnean/Chadronian), British Columbia, Canada 109R, 132R
- LaBarge Fauna (Wasatchian, Wa-7), Wyoming 108L, 128L
- La Colonia Local Fauna (Campanian-Maastrichtian), Argentina 92R
- Lac Pelletier Lower Fauna (Duchesnean), Saskatchewan, Canada 109, 133L, 329L
- Lac Pelletier Upper Fauna (Duchesnean), Saskatchewan, Canada 109, 133L
- Lake Casa Blanca Local Fauna (Uintan, Ui-3), Texas 109L, 118R, 134L, R
- Laguna Umayo Local Fauna (Paleocene), Peru 92R, 93L, R
- Laguna Riviera Local Fauna (Uintan/Duchesnean), San Diego area, California 109R, 124L
- LaPoint Fauna (middle Duchesnean), Utah 118R, 120L, 133R
- Las Tunas Fauna (early Blancan?), Baja California del Sur, Mexico 257, 264L
- Laudate Local Fauna (Tiffanian, T11), California 53, 62R, 81L, R
- Layer Cake Fauna (early Blancan), California 258R
- Lebo Fauna (Torrejonian-Tiffanian), Montana 70R
- Leisey Shell Pit Fauna (Blancan/Irvingtonian), Florida 267R, 270R, 271R, 277, 283L, R
- Lisco Fauna (early Blancan, Rexroadian), Nebraska 235L
- Little Egypt Local Fauna (earliest Chadronian), Texas 120L, 130L, 138R, 160L, 161R, 162R
- Little Pipestone Creek Local Fauna (middle Chadronian), Montana 163L

**Local Fauna(s)** (*continued*)

- Little Stave Creek Local Fauna (Wasatchian/Bridgerian), Alabama 108R, 134L
- Littleton Local Fauna (Puercan, Pu1), Colorado 49, 63R, 64R
- Logan Mine Local Fauna (Hemingfordian, He1), California 176R
- Los Alamos Local Fauna (Campanian-Maastrichtian), Argentina 92R
- Love Bone Bed Local Fauna (Clarendonian, Cl3), Florida 171L
- Lower Cady Mountains Fauna (Hemingfordian, He1) 176R
- Lower Dome Spring Fauna (Barstovian, Ba1), California 174L
- Lower Hunter Wash Fauna (Judithian?), New Mexico 25L
- lower Monroe Creek Fauna (Arikareean, Ar2), Nebraska 193R
- Lower Snake Creek Fauna (Barstovian, Ba1), Nebraska 175R, 199R
- Macasphalt Shell Pit Fauna (Blancan), Florida 244, 257, 262L, 263R, 267R, 268L, 285R, 289
- MacPherson Local Fauna (Hemphillian, He4), Kansas 219L
- Madison Valley Fauna (Barstovian, Ba1), Montana 188L, 190L, R, 191L, 214R
- Mantua Lentil Local Fauna (Pu1), Wyoming 45R, 50, 63R, 64R, 65L
- Marsland fauna (Arikareean, Ar3/4-Hemingfordian, He1), Nebraska 212R
- Martin-Anthony Local Fauna (Arikareean, Ar2), Florida 206L
- Martin Ranch Fauna (Irvingtonian), Texas 244, 250R, 282L
- Mascall Fauna (Barstovian, Ba1), Oregon 171R, 187R, 213R
- Massacre Lake Local Fauna (Hemingfordian, He2), Nevada 178R, 214R
- Matatlán Local Fauna (Barstovian, Ba2), Oaxaca, Mexico 202L
- Mathews Ranch Fauna (Clarendonian, Cerrotejonian, Ch1/2), California 174L, R
- Maxum Fauna (Blancan, Blancan I), California 244, 254L, 257
- Mayfield Ranch Fauna (Irvingtonian), Texas 282R
- McCann Canyon Local Fauna (Arikareean, Ar2), Nebraska 193R, 205L
- McCarty's Mountain Local Fauna (late early Chadronian), Montana 162R
- McGuire Creek Local Faunas (Puercan, Pu1), Montana 51, 53L, R, 63R
- McLeod Fauna (Irvingtonian), Florida 244, 277, 282L, 283R
- Medicine Hat Fauna (Irvingtonian), Alberta, Canada 232R, 270R, 277, 280R, 281L, 288L, 289
- Merychippus* Zone Fauna (Barstovian, Ba1), California 171R
- Midway Fauna (Hemingfordian, He2), Florida 206R
- Minnechaduzza Fauna (Clarendonian, Cl2), Nebraska 216L, R
- Mollie Gulch Local Fauna (Arikareean, Ar3/4), Montana 188L
- Monroe Creek Fauna (Arikareean, Ar2), Nebraska, South Dakota 187R, 189L, 191L, 192L, 193R, 194L, 196R, 197L, 204L, 205L, R, 206L, 209R
- Morena Boulevard Local Fauna (Wasatchian, Wa-6/Wa-7), California 108L, 122R
- Mosquito Gulch Local Fauna (Torrejonian, To1), Montana 51, 73L
- Mount Eden Local Fauna (Hemphillian, Hh4), California 175L, 251R, 259R
- Mt. Eden *see* Mount Eden
- Mt. Blanco Fauna (Blancan), Texas 249L, 253L, R, 257, 266R, 267L, 268R
- Mt. Scott Fauna (Rancholabrean), Kansas 288R, 291L
- Muddy Creek Fauna (Arikareean, Ar2?), Wyoming 204R
- Muddy Tork Local Fauna (Lancian), Montana 26L
- Muddy Valley Fauna (Blancan), Nevada 257, 260R
- Mussentuchit Local Fauna (Albian-Cenomanian), Utah 22R
- Nash Fauna (latest Blancan; Sappan), Kansas 244, 265R, 266L, 270L, 272L, R, 273R, 275L, 295L
- Nettle Springs Fauna (Clarendonian, Cl2), California 174L, R
- Ninefoot Rapids Fauna (late Blancan), Idaho 258L
- Norden Bridge Local Fauna (Barstovian, Ba2), Nebraska 191R, 197R, 200L
- North Coalinga Local Fauna (Barstovian, Ba1), California 171R, 172L, 203L, 214R
- Northeast of Agate Local Fauna (Hemingfordian, He1), Nebraska 187L
- North Tejon Hills Fauna (Clarendonian, Montediablan, Cl3) 172L, R
- Oldman assemblage (Judithian), Alberta, Canada 24L
- Olive Local Fauna (Tiffanian, Ti4), Montana 51, 83L
- Oreana Fauna (early Blancan), Idaho 258L
- Oshkosh Fauna (Hemphillian, Hh2), Nebraska 195R
- Otay Fauna (early Arikareean, Ar1), California 174R
- Palmetto Fauna (Hemphillian, Hh4), Florida 207L, R, 251L
- Panaca Fauna (Blancan, Blancan I), Nevada 250L, 252L, 254L, 257, 260R
- Papago Springs Cave Fauna (Irvingtonian), Arizona 245, 276L, 285L, 289
- Pawnee Creek Fauna (Barstovian, Ba2), Colorado 197R, 213R
- Payne Creek Mine Fauna (Irvingtonian), Florida 277, 283L
- Peanut Peak Member fauna (late Chadronian), South Dakota 168L
- Pearson Mesa Fauna (middle Blancan), Arizona 262L
- Pearson Ranch Local Fauna (early Duchesnean), California 109R, 119R, 124R, 125L, 138R
- Peterson Creek Local Fauna (Arikareean, Ar1/2), Montana 187R, 188L
- Phillips Ranch Local Fauna (Hemingfordian, He2), California 175R
- Pinole Local Fauna (Hemphillian, Hh4), California 251L
- Pipestone Springs Local Fauna (middle Chadronian), Montana 160L, 163L
- Pit Fauna, Porcupine Cave (Irvingtonian, ?Cudahyan), Colorado 245, 270R, 272L, R, 274L, 275R, 276L, 278R, 279L, 294L
- Plateau Valley Local Fauna (Clarkforkian), Colorado 125L
- Poison Creek Fauna (late Blancan), Idaho 258L
- Police Point Local Fauna (Tiffanian, Ti3), Alberta, Canada 52, 82L
- Pollack Farm Local Fauna (Hemingfordian, He1), Delaware 206R, 208L, R
- Pool Branch Fauna (Irvingtonian), Florida 277, 283L
- Port Kennedy Cave Fauna (Irvingtonian, ?Cudahyan), Pennsylvania 244, 274L, 277, 284L
- Porvenir Local Fauna (late Duchesnean), Texas 109R, 120L, 130L, 133L, 138R, 160L, 161R, 329R

- Punta Peligro Fauna (late Paleocene), Argentina 92R, 93L, 94R
- Purgatory Hill Local Fauna (Puercan, Pu3), Montana 51, 68R, 69L, R
- Quarry A Local Fauna (Hemingfordian, He1), Colorado 200R
- Quincy Local Fauna (Hemingfordian, He2), Florida 206R
- Raben Ranch Local Fauna (middle Chadronian), Nebraska 163L
- Railway Quarry Local Fauna (Barstovian, Ba2), Nebraska 191R, 200L
- Rak Division Local Fauna (Hemingfordian, He2), California 175R, 176L, 200R, 215L
- Rancho del Oro Local Fauna (late Uintan), San Diego area, California 124L
- Rancho el Ocote Local Fauna (Hemphillian, He4), Guanajato, Mexico 178L, 201L, R, 218R (also Blancan) 257
- Rancho La Brea Fauna (Rancholabrean), California 284R, 286R, 289
- Rancho Gaitan Local Fauna (earliest Chadronian), Mexico 130L, 162R
- Rancho Viejo (Blancan), Guanajuato, Mexico 244, 257, 264L
- Randlett Fauna (Uintan, Ui-3), Utah 109L, 112L, 118R, 126R, 130L
- Red Division Quarry Local Fauna (Hemingfordian, He2), California 173R, 175R
- Red Fox Fauna (early Blancan), Kansas 265L
- Red Owl Local Fauna (Lancian), South Dakota 26R, 27, 32R, 34L, 35L
- Red Light Fauna (Blancan), Texas 257, 263R, 268R
- Rexroad Fauna (early Blancan, Rexroadian; Arvicoline Zone II), Kansas 253L, R, 267L
- Rexroad 3 Fauna (early Blancan), Kansas 244, 255L, 265L
- Rezabek Fauna (Irvingtonian, Sheridanian), Kansas 245, 274L, 283L, 289
- Ricardo Fauna (Clarendonian, Cl2/3), California 175R, 176R, 177L, 216L, 217L
- Rick Forester Fauna (latest Blancan/Irvingtonian), Kansas 265R, 266L, 270L, 295L
- Rigby Shell Pit Fauna (Irvingtonian), Florida 277, 283L, R
- Rio Chico Fauna (late Paleocene), Argentina 92R, 93L, 94R
- River Road Fauna (Hemphillian, Hh4), Washington 256L
- Robert Fauna (Rancholabrean), Kansas 288R
- Rock Bench Fauna (Torrejonian), Montana 50, 70R, 74L, R
- Rock Creek Fauna (Irvingtonian), Texas 271L, 272R, 277, 282L
- Roche Percée Local Fauna (Tiffanian, Ti4), Saskatchewan, Canada 52, 82R, 83L
- Runningwater Local Fauna (Hemingfordian, He1), Nebraska 175L, 187L, 191R, 192L, 194R, 199L, 206R, 208L, R, 212R
- Rushville Fauna (Irvingtonian, Sheridanian), Nebraska 274L, 285R
- Sand Canyon Fauna (Barstovian, Ba2), Colorado 197R
- Sand Draw Fauna (early Blancan, Rexroadian; Arvicoline Zone II; Blancan III), Nebraska 244, 253L, R, 255L, 257, 266L
- Sandahl Fauna (Irvingtonian, Sheridanian), Kansas 274L
- Sanders Fauna (Blancan, Blancan III), Kansas 244, 255L, 267L
- Sand Point Fauna (early Blancan), Washington 256R, 258L
- Sand Wash Fauna (Uintan, Ui-1), Wyoming 117L
- Santa Fe River 1 Fauna (Blancan), Florida 244, 257, 267R, 268L, R, 271R, 272R
- Santana Mesa Fauna (Arikareean, Ar3), Texas 201L, R, 211R
- Santee Local Fauna (Hemphillian, Hh4), Nebraska 195R, 251L, R
- Santiago Canyon Landfill Fauna (Hemingfordian, He1), California 175L
- Santo Domingo Basin Fauna (late Blancan), New Mexico 262R
- Sappa Fauna (latest Blancan/Irvingtonian; Sappan), Nebraska 257, 266L, 270L, 273R, 295L
- Saw Rock Canyon Local Fauna (Hemphillian, Hh3/4), Kansas 198R, (Blancan) 244, 250L, 254L, 257, 264R, 265L
- SB-1A Local Fauna (Arikareean, Ar2), Florida 205R, 206L
- Scabby Butte Local Fauna ("Edmontonian"), Alberta, Canada 25L
- School Well Local Fauna (Torrejonian) 51, 73R
- Seaboard Local Fauna (Hemingfordian, He1/2), Florida 206R, 207R
- Second Division Fauna (Barstovian, Ba1), California 174L, 176L
- Seneca Fauna (late Blancan, Senecan; Arvicoline Zone III), Nebraska 244, 253L, R, 257, 266L
- Sespe Creek Local Fauna (Duchesnean), California 124R, 125L
- Serendipity Local Fauna (Uintan, Ui-3), Texas 109L, 118R, 129R, 130L, 134R
- Sharktooth Hill Local Fauna (Barstovian, Ba1), California 172L, 214R
- Shutt Ranch Fauna (Irvingtonian), California 278L
- Sheep Creek Fauna (Hemingfordian, He2), Nebraska 175R, 194R, 199L, R, 212R
- Shoddy Springs Fauna (Duchesnean), Montana 109R, 134R
- Shiloh Local Fauna (Arikareean, Ar3), New Jersey 208L, R
- Shotgun Butte Fauna (early Wasatchian), Wind River Basin, Wyoming 121L
- Silver Coulee Local Fauna (Tiffanian, Ti2), Montana 76R
- Simi Valley Landfill Local Fauna (middle Duchesnean), California 109R, 119R, 120L, 124R
- Slaton Fauna (Irvingtonian, Sheridanian), Texas 274L, 277, 282R, 283L, 284L
- Slim Buttes Fauna (Duchesnean), South Dakota 109R, 133R
- Skull Springs Fauna (Barstovian, Ba1), Oregon 187L, R, 214R
- South Mountain Fauna (Arikareean, Ar1), California 173L
- South Tejon Hills Fauna (Clarendonian, Cerrotejonian, Cl1), California 172L, R, 174R
- Split Rock Fauna (Hemingfordian, He2), Wyoming 195R, 196L
- Standing Rock Quarry Local Fauna (Arikareean, Ar4) 198R
- Stewart Springs Fauna (Barstovian, Ba1), Nevada 171R, 177R, 178L
- Stonecrest Local Fauna (Uintan, Ui-3), California 109L, 123R
- Strathern Local Fauna, (Uintan/Duchesnean), Ventura County, California, 109R, 124L, R
- Suchilquitongo Local Fauna (Hemingfordian, He2), Oaxaca, Mexico 202L
- Sucker Creek Fauna (Barstovian, Ba1), Oregon 187R, 214R

**Local Fauna(s)** (*continued*)

- Swami's Point Local Fauna (Bridgerian, Br-2), California 108R, 122R
- Swift Current Creek Local Fauna (Uintan, Ui-3), Saskatchewan, Canada 109L, 118R, 133L
- Sycamore Creek Local Fauna (Clarendonian, Montediablan, Cl3), California 171R
- Tapo Canyon Local Fauna (Uintan, Ui-2), California 109L, 118R, 124R, 125L
- Taunton Fauna (middle Blancan, Blancan III), Washington 255L, 256R, 257
- The Breaks Local Fauna (Torrejonian-Tiffanian, To3-Ti3), Wyoming 50, 77R, 78L, 81R, 82L
- Thomas Farm Local Fauna (Hemingfordian, He1/2), Florida 206L
- Three Mile East Fauna (late Blancan), Idaho 258L
- Tiupampa Fauna (early Paleocene), Bolivia 92R, 93L, R, 94L, R, 95L
- Tobin Fauna (Irvingtonian, Cudahyan), Kansas 273R, 282L
- Toledo Bend Local Fauna (Arikareean, Ar3), Texas 204L, R, 205R
- Tonopah Local Fauna (Barstovian, Ba1), Nevada 177R
- Tonque Local Fauna (Duchesnean), New Mexico 109R, 130R
- Tonuci Mountain Fauna (Blancan), New Mexico 263L
- Topaz Lake Fauna (Blancan/Irvingtonian), Nevada 257, 259L, 260L, 278R
- Topham Local Fauna (Hemingfordian, He1), Saskatchewan, Canada 192L
- Tortugas Mountain Fauna (Irvingtonian), New Mexico 250R
- Tres Cruces Fauna (early Paleocene), Bolivia 92R, 93L, 94R
- Trinity River Local Fauna (Barstovian, Ba1), Texas 203R, 207L
- Trochu Local Fauna (Lancian), Alberta, Canada 26R, 27, 28L
- Trout Cave Entrance Fauna (Irvingtonian), West Virginia 284R
- Trout Cave No. 2 Fauna (Irvingtonian, Cudahyan), West Virginia 274L, 277, 284R
- Tusker Claims, Tusker Fauna (see 111 Ranch Fauna)
- Tyson Ranch Fauna (late Blancan), Idaho 258L
- Unnamed Butte Fauna (late Blancan), Idaho 258L
- Upper Alturas Fauna (Hemphillian/Blancan), California 244, 250L, 251L, 254L, 257
- Upper Cady Mountains Local Fauna (Hemingfordian, He2), California 175R
- Upper Bone Valley Fauna (Hemphillian, Hh4), Florida 207L
- Upper Dome Spring Fauna (Barstovian, Ba1), California 174L
- Upper Harrison Formation faunas (Arikareean, Ar4), Nebraska 191R, 194L, R, 196R, 198R, 212L, R
- Upper Oso Dam Fauna (Hemingfordian, He1), California 174R
- Uptegrove Fauna (Hemphillian, Hh3), Nebraska 195R
- Vallecito Creek Fauna (late Blancan/Irvingtonian), California 244, 258R, 259R, 260L, 271L, 272R
- Vedder Local Fauna (Hemingfordian, He2), California 173R
- Vera Fauna (Irvingtonian, Cudahyan), Texas 273R, 277, 282L
- Verde Fauna (Blancan, Blancan I), Arizona 244, 254R, 255L, 257, 262L
- Vim-Peetz Fauna (Barstovian, Ba2), Colorado 197R
- Virgin Valley Fauna (Barstovian, Ba1), Nevada 171R, 178L, R, 213R, 214R
- Waubonsie Fauna (Rancholabrean), Iowa 289, 291R
- Wagonroad Local Fauna (Puercan, Pu3), Utah 49, 67L, 68L
- Wannagan Creek Local Fauna (Tiffanian, Ti4), North Dakota 52, 83L
- Warm Springs Local Fauna (Hemingfordian, He1), Oregon 187L
- Washoe Local Fauna (Hemphillian, He4), Nevada 178L
- Wathena Fauna (Irvingtonian; Sappan), Kansas 273R
- Wellington Hills Fauna (Blancan), Nevada 260L, 278R
- Wellsch Valley Local Fauna (Irvingtonian), Saskatchewan, Canada 191R, 250R, 257, 271R, 272L, 277, 281L
- West Dry Canyon Fauna (Barstovian, Ba1), California 174L
- White Bluffs Fauna (early Blancan), Washington 244, 256L, R, 257
- White Narrows Fauna (early Blancan), Nevada 260R
- White Rock Fauna (late Blancan, Senecan; Arvicoline Zone III), Kansas 244, 253L, R, 257, 265L, 266L
- White Springs Local Fauna (Arikareean, Ar2), Florida 205L, R, 206L
- Wild Horse Butte Fauna (late Blancan), Idaho 258L
- Willacoochee Creek Local Fauna (Barstovian, Ba1), Florida 206R, 207L, 213R
- Wilson Valley Fauna (Irvingtonian, Cudahyan), Kansas 273R, 282L
- Whisenhunt Quarry Local Fauna (Clarendonian, Cl2), Oklahoma 198L
- Wolf Ranch Fauna (Blancan), Arizona 267L, 269L
- Wood Mountain Fauna (Barstovian, Ba2), Saskatchewan, Canada 192L, R
- Woody Draw Fauna (Irvingtonian, Cudahyan), Texas 273R, 277, 282R
- Wounded Knee Local Fauna (Lancian), Saskatchewan, Canada 26R
- Wounded Knee-Sharps Fauna (Arikareean, Ar1), South Dakota 196L, R
- Yepómera Local Fauna (Hemphillian, He4), Mexico 178L, 200R, 201L, R, 202L, 218R, 251R, 257
- Yoder Local Fauna (late early Chadronian), Wyoming 162R
- Z-line Channel Local Fauna (Puercan, Pu1), Montana 51, 63R, 64L
- Zoyotal Fauna (Arikareean, Ar3), Mexico 201L, R

**Localities, quarries, sites**

- .6 miles east of Junction, Trans-Pecos, Texas (Uintan, Ui-1) 116L, 129R
- 7-Up Butte, Montana (Tiffanian, Ti2) 51, 81L
- "A" sites, Transverse Ranges, California (Hemingfordian, He1) 174R
- Aaron's locality, Alberta, Canada (Tiffanian, Ti2?) 52, 81L
- Adrar Mgorn 1 locality, Morocco (Thanetian) 92L
- Agate Springs quarries, Nebraska (Arikareean, Ar4) 212R
- Agua Fria sites, Trans-Pecos, Texas (Uintan, Ui1) 116L
- Airport site, Wyoming (Tiffanian, Ti4) 50, 83L
- Alexander Locality, Colorado (Puercan, Pu1) 49, 64R
- AMNH locality I, Colorado (Bridgerian, Br1) 114R

- AMNH locality II, Colorado (Bridgerian, Br1) 114R
- AMNH locality III, Colorado (Bridgerian, Br1) 114R
- AMNH locality V, Colorado (Bridgerian, Br1) 114R
- AMNH locality VII, Colorado (Bridgerian, Bro) 114R
- AMNH locality 2, New Mexico (Puercan, Pu2, 3), 48
- AMNH locality 4, New Mexico (Puercan, Pu3), 48
- AMNH locality 5, New Mexico (Puercan, Pu2), 48
- AMNH locality 6, New Mexico (Puercan, Pu2), 48
- AMNH locality 7, New Mexico (Puercan, Pu2), 48
- AMNH locality 8, New Mexico (Torrejonian, To2) 74L
- AMNH locality 10, New Mexico (Torrejonian, To3) 48, 75L
- AMNH locality 230, New Mexico, (Torrejonian, To1) 71R
- AMNH locality 1482, New Mexico (Torrejonian, To2) 74L
- AMNH locality 2658, New Mexico (Torrejonian, To2) 74L
- Baca Formation sites, New Mexico (Uintan/Duchesnean) 109R, 115R, 130R, 131L
- Badwater locality 5, Wind River Basin, Wyoming (Uintan, Ui3) 121R
- Badwater locality 5A, Wind River Basin, Wyoming (Uintan, Ui3) 121R
- Badwater locality 6, Wind River Basin, Wyoming (Uintan, Ui3) 121R
- Badwater locality 7, Wind River Basin, Wyoming (Uintan, Ui3) 118R
- Badwater locality 17, Wind River Basin, Wyoming (early Bridgerian) 121R
- Badwater locality 18, Wind River Basin, Wyoming (early Bridgerian) 121R
- Badwater locality 20, Wind River Basin, Wyoming (early Duchesnean) 120L, 121R
- Bangtail locality, Montana (Tiffanian, Ti1) 51, 80L, R
- Battle Mountain locality, Wyoming (Tiffanian, Ti3) 49, 82L
- Bear Springs section, Arizona (Blancan) 262L
- Bechtold Site, Montana (Puercan, Pu3) 51, 68R, 69L
- Bender locality, Kansas (Blancan, Blancan III) 244, 255L, 267R
- Big Multi Quarry, Wyoming (Clarkforkian, Cf1?) 49, 87L
- Bingo locality, Montana (Tiffanian, Ti1) 51, 80L, R
- Bitter Creek sites, Wyoming (Clarkforkian/Wasatchian) 126R, 127L
- Black Hawk Ranch Quarry, California (Clarendonian, Montediablan, Cl3) 171L, R, 172R
- Blick Quarry, New Mexico (Hemingfordian, He1) 199L
- Blindman River localities DW-1, DW-2, DW-3, Alberta, Canada (Tiffanian, Ti3) 52, 82L
- Bone Bed A, Tepee Trail Formation, Wyoming (Uintan, Ui-1, Shoshonian) 108R, 116L, R, 117L, 120R
- Brown Ranch localities, North Dakota (Torrejonian, To2) 52, 74R
- Bug Creek, Montana (?Lancian) 35L, R, 36L, 53L, R, 64L
- Bug Creek Anthills, Montana (?Lancian; Puercan, Pu1) 35L, 51, 53L, R, 63R, 64L, R
- Bug Creek West, Montana (?Lancian; Puercan, Pu1) 35L, 51, 53L, 63R, 64L
- Campo site, Spain (Paleocene, Thanetian; ?Tiffanian Ti4 equivalent) 88R
- Camp San Onofre Locality, San Diego area, California (Uintan/Duchesnean) 109R, 124L
- Canoe assemblage A, Big Bend area, Texas (Uintan, Ui-1) 108R, 129L
- Canoe assemblage B, Big Bend area, Texas (Uintan, Ui-3) 109L, 129R, 130L
- Canyon Ski Quarry, Alberta, Canada (Tiffanian, Ti4) 52, 83L
- Carrot Top Quarry, Nebraska (Barstovian, Ba2) 195L
- Cathedral Cave, Nevada (Irvingtonian) 272L, R, 274L, 275R, 276L, 277, 278R, 280L, 294L
- Cave Without a Name, Texas (Rancholabrean) 289, 291R
- Cedar Point Quarry, Wyoming (Tiffanian, Ti3) 82L
- Chappo Type Locality, Wyoming (Tiffanian, Ti3) 49, 82L
- Cheek Bend Cave, Tennessee (Rancholabrean) 289, 291R
- Chris's Bonebed locality, Montana (Puercan, Pu1) 51, 63R, 64L
- Cochrane I locality, Alberta, Canada (Tiffanian, Ti1) 52, 80L
- Cochrane II locality, Alberta, Canada (Tiffanian, Ti1) 52, 76R, 88L
- Coffee Ranch Quarry, Texas, (Hemphillian, Hh2) 217R
- Corral Bluffs locality, Colorado (Puercan, Pu2-3?) 67L
- Crestomere School site, Alberta, Canada (Tiffanian, Ti4) 52, 83L
- Croc Pot site, Saskatchewan, Canada (Puercan, Pu3) 52, 69L, R
- Cub Creek 1 site, Montana (Tiffanian, Ti1) 75R, 77L, 80R
- Cub Creek 2 site, Montana (Torrejonian, To3) 50, 75R, 77L
- Cub Creek 3 site, Montana (Tiffanian, Ti1) 75R, 77L, 80R
- Cudahy Ash Pit, Kansas (Irvingtonian, Cudahyan) 273R, 282L
- Dell Creek Quarry, Wyoming (Tiffanian, Ti5) 49, 83R
- Divide Quarry, Wyoming (Tiffanian, Ti4) 50, 83L
- Dogie site, Texas (Puercan, Pu3), 48, 69R
- Dolichorhinus* Quarry, Utah (Uintan, Uinta B) 116L
- Donnybrook site, North Dakota (Torrejonian, To3?) 52, 75R
- Douglass Quarry, Montana (Tiffanian, Ti1) 51, 80L, R
- Dove Spring Formation basal faunal site, California (Clarendonian, Cl1) 177L
- Duchesneodus* Quarry, Utah, (Duchesnean) 118R
- Eagle Quarry, Montana (Tiffanian, Ti1) 75R, 77L, 80R
- Edentulous Jaw Site, California (Tiffanian, Ti2) 53, 81L, R
- Egelhoff Quarry, Nebraska (Barstovian, Ba2) 195L
- Egg Mountain, Montana (Judithian) 24L
- El Gallo, Baja California del Norte, Mexico 25L, 321R
- Erickson's Landing locality, Alberta, Canada (Tiffanian, Ti2) 52, 76R
- Eureka Sound Group, Canada (Wasatchian/Bridgerian) 108R, 109R, 133R, 138L
- Fairmead Landfill locality, California (Irvingtonian) 277, 278R
- False Cougar Cave, Montana (Rancholabrean) 289, 291R
- Ferron Mountain locality, Utah (Puercan, Pu3?) 67L
- Foggy Day beds, N. Fork Owl Creek, Wyoming (Uintan, Ui-1) 121L
- Fort Union Formation sites, Wyoming (Puercan, Pu1) 63R

**Localities, quarries, sites** (*continued*)

- Fr1 (Frenchman 1), Saskatchewan, Canada (Puercan) 35R, 52, 53L, R, 62L, 63R, 64R
- Ferguson Ranch locality, Montana (Puercan, Pu1) 51, 63R, 64L
- Fisher/Sullivan site, Virginia (Wasatchian, Wa-3–Wa-5) 108L, 111R
- Flagstaff Peak locality, Colorado (Puercan, Pu2?) 67L
- Foster Gulch locality, Wyoming (Clarkforkian, Cf2) 50, 87R
- Fowkes Formation sites, Wyoming (Bridgerian) 108R, 128R
- Garbani Quarry, Channel, site, Montana (Puercan, Pu3) 69L, R, 73L
- Gas Tank Hill locality, Utah (Puercan, Pu2?) 49, 67L
- Gidley Gidley Quarry, Montana (Torrejonian, To2) 51, 74L, R, 80R
- Gidley's Horse Quarry, Texas (Irvingtonian) 282L
- Glenn Eleven site, Texas (Puercan, Pu3) 48, 69R
- Glennie locality, Montana (Tiffanian, Ti1) 51, 80L, R
- Goler Formation sites, California (Tiffanian, Ti2) 53, 62R, 70R, 77L, 81L
- Hagerman Horse Quarry, Idaho (early Blancan) 258L, 282L, R
- Hancock Quarry, Oregon (Uintan/Duchesnean) 109R, 133R
- Hand Hills West (lower level), Alberta, Canada (Tiffanian, Ti2?) 52, 81L
- Hand Hills West (upper level), Alberta, Canada (Tiffanian, Ti3) 52, 82L
- Hanna Basin sites, Wyoming (Puercan, Pu1) 50, 61L, 63R, 65L, 66R, 73L, 75R, 77L, R, 78L, R, 80R, 81R, 82L
- Hannold Hill sites, Texas (Wasatchian, Wa-7) 108L, 111L, 129L
- Hanover Quarry No 1, Pennsylvania (Irvingtonian) 277, 284L
- Hansen Bluff, Colorado (Irvingtonian, Cudahyan) 245, 272L, R, 274L, 275L, R, 277, 279L, R
- Harbicht Hill, Montana (?Lancian; Puercan, Pu1) 35L, 51, 53L, 63R
- Hardscrabble Creek, Wyoming (Wasatchian) 120L
- Hares site, North Dakota (Torrejonian, To3?) 52, 75R
- Hart Mine Formation sites, New Mexico (Bridgerian, Br-3) 108R, 130R, 131L
- Haymaker's Orchard locality, Washington (early Blancan) 256R
- Herpajunk Promontory locality, Montana (Puercan, Pu1) 51, 63R
- Hidden Treasure Springs site, California (Hemingfordian, He2) 173R
- High Rock Lake sites, Nevada (Barstovian, Ba2) 178L, R, 214R
- Highway Blowout, Montana (Tiffanian, Ti2) 51, 81L
- Holy City beds, N. Fork Owl Creek, Wyoming (Uintan, Ui-1?) 121L
- Horse and Mastodon Quarry, Colorado (Barstovian, Ba2) 197R
- Horseshoe Canyon, Alberta, Canada ("Edmontonian") 25L
- Howard Ranch, Texas (Rancholabrean) 289, 291R
- Huerfano II, Colorado (Bridgerian) 114L
- Huerfano III, Colorado (Bridgerian) 114L
- Huerfano IV, Colorado (Wasatchian, Wa-7) 131L
- Huerfano V, Colorado (Bridgerian) 114L
- Huerfano VI, Colorado (Wasatchian, Wa-7) 131L
- Huerfano (AMNH) VII, Colorado (Bridgerian, Bro-Bria) 114L, R, 51
- Huerfano VIII, Colorado (Wasatchian, Wa-6) 131L
- Huerfano IX, Colorado (Wasatchian, Wa-6) 131L
- Huerfano XI, Colorado (Wasatchian, Wa-7) 131L
- Huerfano XII, Colorado (Wasatchian, Wa-6) 131L
- Huerfano XIII, Colorado (Wasatchian, Wa-5) 131L
- Ihadjamene locality, Morocco (Thanetian) 92L
- Iron Lightning locality, South Dakota (Lancian) 27, 32R, 34L, R
- Jeep Quarry, New Mexico (Hemingfordian, He1) 199L
- Joe's Bone Bed (TMM 41365–66), Texas (Tiffanian, Ti5) 48, 79L, 83R, 84L
- Joffre Bridge locality, Alberta, Canada (Tiffanian, Ti3) 52, 82L
- Jones-Miller site, Colorado (Rancholabrean) 289, 291R
- Junction locality, Trans-Pecos, Texas (Uintan, Ui-1) 108R, 116L, 118L, 129R
- Keefer Hill locality, Wyoming (Tiffanian, Ti1) 50, 80L
- Kemp Clay Formation, Texas (Lancian) 28L
- Kennewick sequence, Washington (Irvingtonian) 276R, 277, 288R, 289L, R
- Kissimmee River site, Florida (Blancan) 257, 267R, 268R
- KU locality 9, "Little Pocket" (Torrejonian, To2) 48, 74R
- L-41, Aycross Formation, Wyoming (Wasatchian/Bridgerian) 108L, 120R
- LACM [CIT] locality 104, California (Clarendonian, Montediablan, Cl3) 172R
- LACM[CIT] locality 292, California (Duchesnean) 124L
- LACM [CIT] locality 302, California (Clarendonian, Montediablan, Cl3) 172R
- LACM [CIT] locality 303, California (Clarendonian, Cerrotejonian, Cl1) 172L
- LACM [CIT] locality 304, California (Clarendonian, Cerrotejonian, Cl1) 172L
- LACM [CIT] locality 305, California (Clarendonian, Montediablan, Cl3) 172R
- LACM [CIT] locality 307, California (Clarendonian, Cerrotejonian, Cl1) 172L
- LACM [CIT] locality 315, California (Hemingfordian, He2) 173R
- Ladd's Quarry, Georgia (Rancholabrean) 289, 291R
- La Goleta, Michoacan, Mexico (Hemphillian-Irvingtonian) 244, 257, 263R, 264L, 277
- Laramie Formation, Colorado (Lancian) 27R
- Laubach 3, Texas (terminal Rancholabrean) 286R
- Ledge site, Wyoming (Tiffanian, Ti3) 49, 82L
- Leidy Quarry, Wyoming (Puercan, Pu1) 50, 63R
- Leptarctus B site, *Leptarctus* Quarry, Nebraska (Clarendonian, Cl2) 216L
- Little Beaver Cave, Missouri (Rancholabrean) 289, 291R
- Little Dell Dam site, Utah (Irvingtonian) 272L, 276L, 277, 280L, 294
- Little Muddy Creek locality, Wyoming (Tiffanian, Ti1) 49, 80L
- Live Oak site (SB-1A Local Fauna), Florida (Arikareean, Ar2) 205R
- Lloyd site, North Dakota (Torrejonian, To3?) 52, 75L



- Long Fall, Saskatchewan, Canada (Puercan) 26R, 35R, 36L, 52, 53L, 62L, 63R, 64R
- Love Bone Bed, Florida (Clarendonian, Cl3) 171L
- Lundbreck, Alberta, Canada ("Edmontonian") 25L
- Marfil, Mexico (Bridgerian/Uintan) 108R, 131R, 132L
- Martin-Anthony oreodont site, Florida (Arikareean, Ar2) 206L
- Mason Pocket (Quarry), Colorado (Tiffanian, Ti4) 48, 76R, 79L, 82R
- McKeever Ranch localities, Montana (Puercan, Pu1) 51, 63R, 64L
- Medicine Rocks 1-Mehling Site, Montana (Torrejonian, To3?) 51, 75R, 81L
- Mel's Place locality, Alberta, Canada (Tiffanian, Ti3) 52, 82L
- Mesa Prospect, New Mexico (Hemingfordian, He2) 199L
- Monument Blowout, Wyoming (Wasatchian, Wa-3) 132L
- Mount Laurel Formation, New Jersey (Lancian) 28L
- Morales 1 locality, Montana (Puercan, Pu1) 51, 63R, 64L
- Mullen assemblage, Kansas (late Blancan, Senecan; Arvicoline Zone III) 253L, R, 275R
- Naashoibito Member, Kirtland Formation, New Mexico (Lancian) 28L
- Nejapa Valley sites, El Cameron Formation, Oaxaca, Mexico (Barstovian, Ba3) 202L, R
- Newell's Nook Quarry (USGS D-2003), Montana (Tiffanian, Ti1) 51, 80R
- New Year Quarry, California (Barstovian, Ba1) 176L
- Norden Bridge Quarry, Nebraska (Barstovian, Ba2) 191R, 197R, 200L
- North Cove, Nebraska (Rancholabrean) 289, 291R
- North Horn Formation, Utah (Lancian) 27R
- Nut Bed, Clarno, Oregon (Bridgerian) 108R, 133R
- Owl Creek, Wyoming (Bridgerian) 109L, 120R
- Pajarito locality (Laguna site), New Mexico (Blancan) 262R
- Patton Cave, West Virginia (Rancholabrean) 289, 291R
- Peccary Cave, Arkansas (Rancholabrean) 289, 291R
- Pipe Creek Sinkhole, Indiana (Blancan) 250L
- Pita Flats locality, North Dakota (Puercan, Pu2-3) 52, 67L
- Pollack Farm site, Delaware (Hemingfordian, He1) 206R, 208L, R
- Pomerado Conglomerate sites, San Diego area, California (Uintan/Chadronian) 109L, 118R
- Porcupine Cave, Colorado (Irvingtonian) 244, 270R, 272L, R, 274L, 275R, 276L, 277, 278R, 279L, 285L, 294L
- Porcupine Hills Formation sites, Alberta, Canada (Torrejonian) 70R, 75L
- Post Ranch faunal horizon, Arizona (early Blancan) 261L
- Powder Wash sites, Utah (Wasatchian/Bridgerian) 108R, 126L
- Prince Creek, Alaska (Lancian) 26R
- Princeton Quarry, Wyoming (Tiffanian, Ti5) 50, 83R, 85R
- Railway Quarry, Nebraska (Barstovian, Ba2) 191R
- Rav-1, Saskatchewan, Canada (Puercan, Pu3?) 35R, 52, 69L, R (Rav W-1)
- Ray's Bonebed (TMM 40536-37), Texas (Tiffanian, Ti3) 48, 79L, 82L, R
- Red Creek, Wyoming (Clarkforkian) 120L
- Red Deer locality, Alberta, Canada, (Tiffanian, Ti2) 76R
- Red Division Quarry California (Hemingfordian, He2) 173R, 175R
- Red Hot Local Fauna, Mississippi (Wasatchian, Wa-1?) 108L, 133R
- Red Owl Quarry, South Dakota (Lancian) 26R, 27, 32R, 34L, 35L
- Rexroad 3 Locality, Kansas (Blancan, Blancan III) 244, 255L, 265L
- Rick Forester locality (late Blancan), Kansas 266L
- Ries locality, Wyoming (Clarkforkian, Cf2) 50, 87R
- Roche Percée localities, Saskatchewan, Canada (Tiffanian, Ti4) 52, 82R
- Rock Bench Quarry, Montana (Torrejonian, To2) 50, 70R, 74L, R
- Rock Creek Quarry, Texas (Irvingtonian) 277, 282L
- Rough Gulch locality, Wyoming (Clarkforkian, Cf2) 50, 87R
- Round Mountain Quarry, New Mexico (Clarendonian, Cl1) 200L, R
- Saddle Annex locality, Wyoming (Tiffanian, Ti3) 49, 82L
- Saddle locality, Wyoming (Tiffanian, Ti2) 49, 81L
- Salamander Cave, South Dakota (Irvingtonian) 245, 276R, 277, 281L, 282L, 285L, 289R
- SAM Cave, New Mexico (Irvingtonian, ?Cudahyan) 274L, 275L, 276L, 277, 279L, R
- Sand Creek fauna, Wyoming (Tiffanian, Ti3?) 50, 82L
- San Jose Formation Locality 18, New Mexico (Wasatchian, Wa-6?) 108L, 130L
- San Jose Formation sites, New Mexico (Wasatchian, Wa-6?) 111L
- Santiago Formation sites, San Diego area, California (Uintan) 109L, R, 118R, 124L
- Savage Canyon site (Stewart Springs Fauna), Nevada (Barstovian, Ba1) 177R
- Scarritt Quarry, Montana (Tiffanian, Ti2) 51, 80R, 81L, R
- Schiebout-Reeves Quarry, Texas (Tiffanian, Ti1) 48, 80L, R
- SDSNH Locality 3495, San Diego area, California (Uintan/Duchesnean) 124L
- Sebits Ranch Site 24B, Texas (Hemphillian, Hh1) 198L
- Sespe Formation sites, California (Uintan/Arikareean) 109L, R, 119R, 124R, 125L, 172R, 174R
- Silberling Quarry, Montana (Torrejonian, To2) 51, 74L, R, 80R
- Simpson localities 9, 65, 78, Montana (Torrejonian, To1?) 51, 73R
- Simpson locality 13, Montana (Tiffanian, Ti3) 51, 82R
- Simpson Quarry, Montana (Puercan, Pu2-3) 51, 69L, R
- Skyline Channels, Texas (early Duchesnean) 109R, 120L, 129R
- Sloth-Camel Quarry, Texas (Irvingtonian) 282L
- South Table Mountain sites, Colorado (Puercan, Pu1) 49, 64R
- Stadium Conglomerate sites, California (Uintan, Ui-2?) 109L, 118L, R, 123L, R
- Steepside Quarry, California (Barstovian, Ba1) 176L, 215L
- Stewart Valley sites, Nevada (Clarendonian, Cl2) 178L
- Sunbright Ash Pit, Kansas (Irvingtonian, Cudahyan) 273R, 282L
- "Sweetwater" Formation sites, San Diego area, California (Uintan/Duchesnean) 109R, 124L

**Localities, quarries, sites** (*continued*)

- Swain Quarry, Wyoming (Torrejonian, To2) 49, 62R, 74R
- Swan Hills site, Alberta, Canada (Tiffanian, Ti4) 52, 83L
- Swift Current Plateau, Saskatchewan, Canada (Uintan, Ui-3) 109L, 133L
- Thornton Beach, California (Irvingtonian) 271R
- Tijeras Arroyo, New Mexico (Blancan/Irvingtonian) 262R, 270R
- Titanoides* locality, Wyoming (Tiffanian, Ti5) 49, 83R
- TMM locality 40147, Texas (Torrejonian, To2?) 74R
- Togwotee Summit, Wyoming (Bridgerian, Br-2) 108R, 120R
- Tom's Top site, Texas (Puercan, Pu3) 48, 69R
- Tsentas Microsite (NMMNH locality L-312), New Mexico (Torrejonian, To2) 48, 74L
- UALP locality 7650, New Mexico (Torrejonian, To2) 74L
- UCM loc. 77039, Colorado (Wasatchian) 112R
- UCM loc. 77041, Colorado (Wasatchian, Wa-5) 131L
- UCM loc. 83120, Wyoming (Wasatchian/Bridgerian) 127L
- UCM loc. 84115, Wyoming (Bridgerian) 115L
- UCM loc. 84213, Wyoming (Bridgerian) 113R
- UCM loc. 84219, Wyoming (Bridgerian) 113R
- UCM loc. 84220, Wyoming (Wasatchian, Wa-7) 113R
- UCM loc. 84211, Wyoming (Wasatchian, Wa-7) 113R
- UCM loc. 84228, Wyoming (Wasatchian, Wa-7) 113R
- UCM loc. 88052, Wyoming (Wasatchian, Wa-3) 132L
- UCM loc. 92189, Wyoming (Uintan, Ui-1) 116L, 117L
- UCM loc. 96197, Colorado (Bridgerian) 112R
- UCMP locality V-5814, California (Uintan/Duchesnean) 124R
- UCMP locality V-72088, San Diego area, California (Uintan/Duchesnean) 124L
- University of Michigan locality I, II, III, V, New Mexico (Bridgerian, Gardnerbuttean) 131L
- UM locality 263, Wyoming (Tiffanian, Ti2) 50, 81L
- Valley View Quarry, California (Barstovian, Ba1) 176L
- Ventana Cave, Arizona (Rancholabrean) 289, 291R
- Verdigris Coulee (early Campanian), Alberta 23L, R
- Walbeck site, Germany (Paleocene, ?Tiffanian equivalent) 88R
- Wapiti Valley fauna (Lysitean-Bridgerian), Wyoming 122R
- West Bijou Creek-1 locality, Colorado (Puercan, Pu2-3?) 49, 67L
- West End site, Wyoming (Tiffanian, Ti3) 49, 82L
- Whistler Squat Quarry, Trans-Pecos, Texas (Uintan, Ui-1/Ui-2) 109L, 116L, 118L, 129R
- White Site, Montana (Tiffanian, Ti2) 51, 81L
- White's River Basin Survey Site, North Dakota (Tiffanian, Ti3) 52, 82L
- Wiggins Formation sites, Wyoming (Uintan) 109L, 117R, 118L, 121L, R, 122R
- Wind River Formation sites, Wind River Basin, Wyoming, (Uintan, Bridgerian) 107L, R, 108L, 113L, 114R, 121L, 122L, 132L, R
- Witter Quarry, Wyoming (Tiffanian, Ti4) 50, 52, 83L
- XMas and Kat quarries, Nebraska (Clarendonian, Cl2) 216L, R
- LOD, Last Occurrence Datum xiiiR
- Lomas de Tetas de Cabra Formation, Baja California 108L, 111L, 131R
- London Basin 324L
- Lost Cabin beds (*Lambdaotherium* level) 107R
- Lostcabinian subage (Wa-7) of Wasatchian mammal age 107L, R, 113L, R, 121L, 122L, R, 135 characterized 113L, R, 122L
- Lost Cabin Member, Wind River Formation 107R, 108L, 113R, 114R, 121L, 122L
- Loup Fork beds 190L
- Love Formation, Texas 263R
- Loveland Loess, Colorado, Iowa, Nebraska 285R, 287R
- Love Ranch Formation, New Mexico 131L
- Lower *Oreodon* Beds, Big Badlands, South Dakota 165L
- Lower Nodular Zone, Big Badlands, South Dakota 164R
- Lower Pleistocene Series 236R
- Lower Rosebud Formation, beds, Nebraska, South Dakota 196R, 209R
- Lower Tuffaceous Zone, New Mexico 221L
- Lower variegated sequence, Togwotee Pass, Wyoming 85L, 108L, 120L
- Lower Whitney Ash, Nebraska 159, 166L
- LPTM (late Paleocene Thermal Maximum) 320L, R, 324R
- LRD (lowest range datum) 17L
- LSD, Lowest Stratigraphic Datum xiiiL, R, xivL, 4, 5, 14L, R, 15R, 259R
- LSD<sub>k</sub>, lowest known stratigraphic occurrence 259L, 278L
- Ludlow Formation, Montana, North Dakota 62L, 63L, 65R, 67L, 68R, 71L, 74R
- Ludlow Member, Fort Union Formation, North Dakota 62L
- Luisian benthonic foraminiferal stage, California 171R, 172L, 214R
- Lusk, Wyoming 163L, 164L, R, 165L
- Lysite beds 107R, 108L
- Lysitean subage (Wa-6) of Wasatchian mammal age 107L, R, 111L, 113L, 122L, R, 130R, 135 characterized 113L
- Lysite Member, Wind River Formation, Wyoming 107R, 108L, 121L, 122L
- Ma (Megannum in the radioisotopic time scale) xivR
- Maastrichtian Stage/Age, rocks, faunas 21L, 24R, 35R, 37L, 92R, 93R, 317R, 318L
- macroevolutionary 37L
- Madison Valley Formation, Montana 188L, 190L, R
- Madrean shrubland, chaparral 327L
- magnetostratigraphic viiL, xivL, xviiiR, 10L, 16R, 17R, 18L, 35R, 44R, 46, 47, 63L, 69L, R, 71L, 74R, 75R, 76L, 79L, 86L, 88R, 93L, R, 94L, 95R, 123R, 134R, 138L, R, 159, 161R, 171L, 172R, 175L, 179L, 194L, 199R, 206R
- magnetostratigraphic polarity chrons, units, zones xivL, 9L, R, 10L
- Brunhes Chron 233L, 238L, 259R, 261L, 263L, 274L, 276L, 278L, 279L, R, 280L, 281L, 285R, 287L
- Gauss Chron 233L, 253L, 255L, R, 256R, 258L, 259L, 261L, R, 262L, 263L, R, 265R, 267L, 268L, 269L, 279R
- Kaena subchron of Gauss Chron 240, 256R, 258L, 259L, 262L, 263L
- Mammoth subchron of Gauss Chron 240, 258L, 261R, 262L
- Gilbert Chron 200R, 233L, 256R, 258L, 259R, 261L

- Cochiti subchron, Gilbert Chron  
240, 256R, 258R, 259L
- Nunivak subchron, Gilbert Chron  
240, 251R, 154R, 262L
- Sidufjall subchron, Gilbert Chron  
200R, 240, 251R, 252L, 256R
- Thvera subchron, Gilbert Chron  
233R, 240, 252L
- Matuyama Chron 233L, 253L, 255R,  
258L, 259L, 261R, 262L, 263L, 267L,  
268L, 269L, 271R, 274L, 278L,  
279L, R, 281L, 285R, 294L
- Cobb Mountain subchron of  
Matuyama Chron 240, 263L
- Jaramillo subchron of Matuyama  
Chron 233L, 240, 258R, 259R,  
263L, 269L, 279R, 280L
- Olduvai subchron of Matuyama  
Chron 233L, 237L, 240, 258R,  
259L, 261L, R, 263L, 272L, R,  
275L, 278L, 279R, 280L, R
- Reunion subchron, Matuyama  
Chron 240, 279R
- Chron C<sub>1n</sub>, Brunhes Chron 233L,  
238L, 261L, 274L, 276L, 279R, 281L,  
282R, 287L
- Chron C<sub>1r</sub> 258R, 278L, 279R, 280L
- C<sub>1r.1r</sub> 271R, 278L, 281L
- C<sub>1r.1n</sub>, Jaramillo subchron of  
Matuyama Chron 258R, 279R
- C<sub>1r.2r</sub> 260L, 263L, 269L, 289L
- Chron C<sub>2n</sub>, Olduvai subchron of  
Matuyama Chron 233L, 237L, 240,  
258R, 259L, 261L, R, 263L, 272L, R,  
275L, 278L, 279R, 280L, R
- Chron C<sub>2r</sub> 253L, 258L
- C<sub>2r.1r</sub> 259L, 278L
- C<sub>2r.1n</sub>, Reunion subchron of  
Matuyama Chron 279R
- C<sub>2r.2r</sub> 253L, 258L, 266R
- Chron C<sub>2An</sub> 253L, 259L, 263L, 269L  
C<sub>2An.1n</sub> 258L, 259R, 262L, 263R,  
266R, 267L, 268L, 279R
- C<sub>2An.1r</sub>, Kaena subchron of  
Gauss Chron 256R, 258L, 259L,  
R, 263L
- C<sub>2An.2r</sub>, Mammoth subchron of  
Gauss Chron 258L, 261R
- C<sub>2An.3n</sub> 255L, 258L, 259L, R,  
262L, 265L
- Chron C<sub>2Ar</sub> 251R, 256R, 258L, 259R,  
261L
- Chron C<sub>3n</sub> 251R
- C<sub>3n.1n</sub>, Cochiti subchron of  
Gilbert Chron 256R, 258R, 259L
- C<sub>3n.2n</sub>, Nunivak subchron of  
Gilbert Chron 251R, 254R
- C<sub>3n.2r</sub> 251R
- C<sub>3n.3n</sub>, Sidufjall subchron of  
Gilbert Chron 200R, 251R, 256R
- C<sub>3n.3r</sub> 252L
- C<sub>3n.4n</sub>, Thvera subchron of  
Gilbert Chron 233R, 252L
- Chron C<sub>3r</sub> 175R, 200R, 218R, 251R,  
252L, 259R
- Chron C<sub>3An</sub> 218R, 251R (3An.2n)
- Chron C<sub>3Bn</sub> 200R
- Chron C<sub>4r</sub> 171R (C<sub>4A</sub>)
- Chron C<sub>4Ar</sub> 172R, 177L
- Chron C<sub>5n</sub> 177L, R, 200R
- Chron C<sub>5r</sub> 172R, 174L, R, 177L
- Chron C<sub>5An</sub> 172R, 174L, R, 215R  
(C<sub>5An2</sub>) 199R
- Chron C<sub>5ABn</sub> 217L
- Chron C<sub>5ABr</sub> (C<sub>5AB</sub>) 200L
- Chron C<sub>5ACn</sub> 176L
- Chron C<sub>5ACr</sub> (C<sub>5AC</sub>) 200L
- Chron C<sub>5ADn</sub> 191L, 199R
- Chron C<sub>5ADr</sub> (C<sub>5AD</sub>) 176L
- Chron C<sub>5Bn</sub> 176L
- Chron C<sub>5Br</sub> 176L, 191L, 214R (C<sub>5B</sub>)  
199R, 207L
- Chron C<sub>5Cn</sub> 173R, 196L, 199L (C<sub>5C</sub>)  
199L (C<sub>5Cn2</sub>) 215L
- Chron C<sub>5Cr</sub> 173R, 175R
- Chron C<sub>5Dr</sub> 175L, 194R
- Chron C<sub>5Er</sub> 175L, 194L (C<sub>5E</sub>)  
194R
- Chron C<sub>6n</sub> 194L
- Chron C<sub>7r</sub> 193R
- Chron C<sub>7Ar</sub> 193R
- Chron C<sub>8n</sub> 193R
- Chron C<sub>8r</sub> 193R
- Chron C<sub>9n</sub> 193R
- Chron C<sub>9r</sub> 172L, R, 193R
- Chron C<sub>10n</sub> 173L, 179R
- Chron C<sub>10r</sub> 173L, 174R
- Chron C<sub>11n</sub> 166R
- Chron C<sub>11r</sub> 166L
- Chron C<sub>12r</sub> 165L, 166L
- Chron C<sub>13n</sub> 164R, 165L, 330L
- Chron C<sub>13r</sub> 163L, 164L, R
- Chron C<sub>15n</sub> 162R, 163L
- Chron C<sub>15r</sub> 163L
- Chron C<sub>16r</sub> 162R
- Chron C<sub>17n</sub> 138R
- Chron C<sub>17r</sub> 136,
- Chron C<sub>18n</sub> 125L, 138R
- Chron C<sub>18r</sub> 125L, 136, 138R
- Chron C<sub>19n</sub> 125L
- Chron C<sub>19r</sub> 123R, 125L, 136
- Chron C<sub>20n</sub> 123R, 125L, 136, 138L
- Chron C<sub>20r</sub> 116L, 117L, 120R, 123R,  
136, 138L
- Chron C<sub>21n</sub> 123L, R, 135, 138L
- Chron C<sub>21r</sub> 123L, 135
- Chron C<sub>22n</sub> 135
- Chron C<sub>22r</sub> 135, 138L
- Chron C<sub>23n.1n</sub> 135
- Chron C<sub>23n.1r</sub> 135
- Chron C<sub>23n.2n</sub> 135
- Chron C<sub>23r</sub> 131R, 135, 138L
- Chron C<sub>24n</sub> 131R
- Chron C<sub>24n.1n</sub> 135
- Chron C<sub>24n.1r</sub> 135
- Chron C<sub>24n.2n</sub> 135
- Chron C<sub>24n.2r</sub> 135
- Chron C<sub>24n.3n</sub> 135, 138L
- Chron 24r 12L, 79L, 86L, 87L, R, 88L,  
89R, 90L, 92L, 106L, 135, 138L
- Chron C<sub>25n</sub> 79L, 84L, R, 86L, 87L
- Chron C<sub>25r</sub> 79L, 82R, 83L, 84L, 89L,  
92L
- Chron C<sub>26n</sub> 79L, 82L, 83L, 88R
- Chron C<sub>26r</sub> 75R, 77L, 79L, 80R, 81R,  
82L, R, 93R, 94L
- Chron C<sub>27n</sub> 71R, 72R, 74L, 75R, 76L,  
77L, 78L, 79L
- Chron C<sub>27r</sub> 71L, 72R, 73R, 74L, R,  
79L, 91L, 93R, 94L
- Chron C<sub>28n</sub> 71L, R, 73R
- Chron C<sub>28r</sub> 63L, 68L, 69R, 71L, 73L,  
R, 93R, 94L
- Chron C<sub>29n</sub> 65R, 66L, 67R, 68L,  
69L, R, 71L
- Chron C<sub>29r</sub> 12R, 34R, 63L, 65L, R,  
67R, 91L, 93R
- Chron C<sub>30n</sub> 34R
- Chron C<sub>31n</sub> 34R
- Chron C<sub>31r</sub> 34R, 317R
- Chron C<sub>33n</sub> 25L
- magnetic polarity stratigraphy, magne-  
tostratigraphy viiR, xivL, 9L, R, 10L,  
32L, R, 34, 63L, 65L, R, 67R, 68L,  
69L, R, 71L, R, 72R, 73L, R, 74L, R,  
75R, 76L, 77L, 78L, 79L, 80R, 81R,  
82L, R, 83L, 84L, R, 86L, 87L, R, 88L,  
R, 89R, 91L, 92L, 93R, 94L, 117R,  
157R, 158L, 160L, 162L, R, 169L, 172R,  
173R, 175L, 177L, 179R, 191L, 193L, R,  
198R, 200R, 202L, 206R, 217R, 218R,  
233L, R, 271L
- magnetic polarity time scale viiR, 9R,  
10L, 117R
- Mammal age viiR, xiL, xivL, R
- Mammoth magnetic polarity subchron  
of Gauss Chron 258L, 261R
- Mantuan Mammal Age 43L
- Marshalltown Formation, New Jersey  
25R
- Marsland Formation, Nebraska  
212R
- Mascall flora, Oregon 333R
- Mascall Formation, Oregon 187L,  
220R
- Masuk Formation, Utah 23R
- MAT (mean annual temperature) 318L,  
319, 320R, 321L, 326L, R, 327R, 328L,  
330L, R, 332L, 333R, 337L
- Matilija Sandstone, California 124R

- Matuyama magnetic polarity chron  
233L, 253L, 255R, 258L, 259L, 261R,  
262L, 263L, 267L, 268L, 269L, 271R,  
274L, 278L, 279L, R, 281L, 285R, 294L
- Mazama Ash, Washington, Oregon  
288R
- MBE (Mammal dispersal event, at  
LPTM) 316, 320L
- McCone County, Montana 64L
- McCullough Peaks Formation,  
Wyoming 17R
- McGuire Creek, Montana 32L, 53L, R,  
64L
- Meade Basin, Kansas 257, 270L, 288R
- Meade County, Kansas 232R, 265L, R,  
270L, 273R, 277, 281R, 289, 291L
- Meade County, South Dakota 26R
- Medicine Hat sequence, Alberta,  
Canada 232R, 270R, 277, 280R, 281L,  
288L, 289
- Medicine Rocks, Montana 51, 81L
- Mediterranean Sea (area) 233R, 237L,  
251L
- megathermal climate, floras 319, 321L,  
324R, 327L, 328L, 330L, 332L
- Melville Formation, Montana 80R
- Merced Formation, California 271R
- Merycoiodon major* Interval Zone  
(late Whitneyan) 166L
- Mesa Chijuilla, New Mexico 72R, 75L
- Mesa Falls Ash, Nebraska 234, 266L,  
273R, 280R
- Mesa de Cuba, New Mexico 48, 71R,  
72R, 73R
- Mesa de Cuba-Mesa Portales 48
- “Mesaverde” Formation, Wyoming  
24L
- Mesilla Basin, New Mexico 250R, 257,  
263L, 277, 280L
- mesothermal climate, floras 218R, 319,  
320R, 321L, 326L, 330L
- Mesozoic Erathem 25R
- Messinian salinity crisis 233R, 251L
- Messinian Stage, Italy 233R
- methane hydrates 320L
- Mexico 37L, 106L, 130L, 131R, 162R,  
169L, 200R, 201L, R, 202L, R, 217L,  
218L, R, 219L, 239L, 251L, R, 254R,  
256L, 257, 263R, 270L, 277, 280R, 289,  
294L, 285L, 327L, R
- microthermal climate, floras 318R, 319,  
324L, 327R
- Middlegate flora, Nevada 332L
- middle Wasatchian *see* Graybullian
- Milk River Formation, Alberta,  
Canada 23R
- Minimal Overlap Assemblage Fossil-  
zone 5, 6R
- Miniochoerus affinis* Interval Zone (late  
early Orellan) 164R
- Miniochoerus chadronensis* Interval  
Zone (late Chadronian) 163R
- Miniochoerus gracilis* Interval Zone  
(early late Orellan) 164L
- Miocene Chronofauna 195L, 197R,  
209L, 218L
- Miocene (medial) climatic optimum  
331R, 333L
- Miocene-Pliocene boundary 218R,  
233R, 251L, 295R
- Miocene-Pliocene Chronofauna 335R,  
337L
- Miocene Series/Epoch, faunas, rocks  
176R, 188L, 191R, 192R, 195L, 197R,  
198R, 201L, R, 202L, 203L, R, 207L,  
R, 208L, 209L, 211R, 214L, 215R, 217R,  
218L, R, 233L, R, 251L, 260R, 269L,  
295R, 327L, 330L, 331R, 332L, R, 333L,  
334R, 335R, 336R, 337L,
- Miramar Sandstone Member, Pomer-  
ado Conglomerate, San Diego area,  
California 109L, 124L
- Mission Valley Formation, San Diego  
area, California 109L, 118R, 123R,  
124L, 136
- Mixodectes pungens/Plesiadapis prae-  
cursor* Interval Zone (To3) 72R, 75L
- Mixodectes pungens* Taxon-range Zone  
(To2) 16R, 17L
- Mojave Desert, California 172L, 174R,  
175L, R, 176R, 214R, 217L, 335L
- molecular 37L, 246L
- Molina Member, Debeque Formation,  
Colorado 125R
- Mongolia 90R
- Monroe Creek Canyon, Nebraska  
193R, 194L
- Monroe Creek Formation, Nebraska,  
South Dakota 189L, 191L, 192L, 193R,  
194L, 196R, 197L, 204L, 205R, 206L,  
209R
- Montediablan Stage, Clarendonian,  
Cl3, California 171L, 172R, 174R, 177L
- Moraga Formation, California 219R
- Morris Skinner, American Museum of  
Natural History, New York 160R,  
166R
- Mount Eden Formation, California  
175R, 251R
- Mount Laurel Formation, New Jersey  
28L
- Mount Soledad Formation, California  
122R
- Mt. Eden flora, California 337L
- Muddy Valley beds, Nevada 257, 260R
- Mud Hills, California 175L, 214R
- M.Y. (or m.y.) xivR
- Myton Member, Uinta Formation (= *U*  
Uinta C), Utah 109L, 111R, 117R,  
118L, 126R
- M Zone (= *Pantolambda* Zone, To3)  
72R
- Naashoibito Member, Kirtland Forma-  
tion, New Mexico 28L, 34, 36R
- Nacimiento Formation 45R, 62L, 63L,  
66R, 67R, 70L, R, 72R, 73L, R, 74R,  
75L, 76L, 78R
- NALMA xivL, R, 21L, 22, 34L, 35L, R,  
43L, R, 45L, R, 53R, 70L, 90R, 93L,  
106R, 107L, 112L, 134R, 135, 136, 137,  
138L, R, 158L, 169R, 209L, R, 210,  
212R, 214L, 218L
- Nambé Member, Tesuque Formation,  
New Mexico 199L, R
- Nanjemoy Formation, Virginia 108L,  
133R
- Nanxiong Basin, China 91L
- Navahoceros* LSD<sub>k</sub> 259L
- Nebraskan glacial stage (Pleistocene)  
237R
- Nebraskan till 237R
- Neogene xivR 237L, 331R
- Neotropical (taxa) 263L, 268R, 269L
- New Fork Tongue of Wasatch Forma-  
tion 113R, 128L
- Niland Tongue, Green River Forma-  
tion, Wyoming 180L, 127L
- Niobrara County, Wyoming 25R
- No. 4 White Ash, New Mexico 221L
- non-analog faunas (see analog)
- Nongshanian, Asian land mammal age  
(approx. correlative with Tiffanian/  
Torrejonian) 90R, 91R, 32RL
- Nonpareil Ash, South Dakota 166R,  
196L
- North American Land Mammal Age  
viiiL, 8R, 21L, 43L, R, 106L, 135, 158L,  
169R, 208R, 210, 232L
- North American Stratigraphic Code  
(NACSN) 43R, 44L, 164L
- Northern Great Basin, North America  
175L
- Northern Hemisphere 106L, 336R
- Northern Rocky Mountains, North  
America 179L, 187L, 188L, 191L, 214R
- North Great Plains, North America  
191R
- North Horn Formation, Utah 27R,  
20L, R, 21L, 67L, 68L, 70L, R, 71L, R,  
73R
- Northwestern Nevada, North America  
178L
- notophyllous broadleaved evergreen  
forest 318R, 319, 320, 321L, 327L, 332L
- NP (nannoplankton) zones 11L, R, 12L,  
89R, 133R, 134L
- Nunivak magnetic polarity subchron  
of Gilbert Chron 240, 251R, 254R,  
262L

- Oak Grove flora, Oregon 337L  
 Oaxaca, Mexico 202L  
 Ocean Point, Alaska 26R  
 Ogallala Formation, Great Plains 192L, 197L, R, 198L  
 Ogallala Group, Great Plains 188R, 194R, 213L  
 Ojo Caliente Member, Tesuque Formation, New Mexico 200L  
 Ojo Encino Member, Nacimiento Formation 45R, 72R, 75L  
 Olcott Formation, Nebraska 197L, 214L  
 Oldman Formation, Alberta, Canada 24L  
 Olduvai magnetic polarity subchron of Matuyama Chron 233L, 237L, 240, 258R, 259L, 261L, R, 263L, 272L, R, 275L, 278L, 279R, 280L, R  
 Oligocene Series/Epoch, faunas, rocks 133L, 156L, R, 160R, 164L, 172R, 174R, 176R, 198R, 205R, 209L, 320R, 326L, 329R, 330L, R, 331L, R  
 Oligocene-Miocene boundary 331R  
 Olyorian faunas, Siberia 285R  
 One Mile Creek flora, British Columbia, Canada 326R  
 Opal, Wyoming 111R  
 Orella A, (Big Cottonwood Creek Member, Chadron Formation), Nebraska 158L, 161L, 162L, 163R  
 Orella B, Nebraska 164R  
 Orella C, Nebraska 165L  
 Orella D, Nebraska 165L  
 Orella Member, Chadron Formation, Nebraska, Wyoming 158L, 160L, R, 163R, 164L  
**Orellan mammal age**, faunas, rocks 156L, R, 158L, 159, 160L, R, 161L, R, 162L, 163R, 164–165L, R, 330R  
 definition and characterization 164L  
 zonation 164L, 164R  
*Hypertragulus calcaratus* Interval Zone (earliest Orellan) 164R  
*Miniochoerus affinis* Interval Zone (late early Orellan) 164R  
*Miniochoerus gracilis* Interval Zone (early late Orellan) 164R, 165L  
*Merycoidodon bullatus* Interval Zone (latest Orellan) 165L  
 Orellan-Whitneyan boundary 166L  
 Oreodon beds, Brule Formation, Big Badlands, South Dakota 159, 163R  
 Oreodont Tuff, Barstow Formation, California 9L, 219R  
 Otay Formation, San Diego area, California 174R  
 Ouarzazate Basin, Morocco 92L  
 Ouled Abdoun Basin, Morocco 92L  
 Owyhee Basalt, Columbia Plateau, Oregon 187R, 214R  
 oxygen isotope sea water values 316, 317L, R, 318L, 320L, 330L, 332L  
 oxygen isotope stages 238R  
 Pacific Northwest 254R, 256L  
*Palaeosyops borealis* Assemblage-Zone (Br-o) 113L, 114R  
 Paleocene Series/Epoch, faunas, rocks xivR, xviiR, 1L, 11L, R, 16L, R, 17L, 26R, 35L, R, 36L, 37L, R, 43L, R, 44R, 45L, 64R, 76R, 77R, 78–79L, R, 80L, 83L, R, 85L, 88R, 89–95L, R, 96L, 320–321L, R, 323L, 324L, 325R  
 Paleocene-Eocene boundary 11L, R, 43L, R, 79L, 89L, R, 90L, R, 92L, 93L, 106L, R, 107R, 128R, 132L, 134L, R, 139L, 320L, R, 324L, R  
 Paleogene xivR, 202L, 318L, 324R  
 paleomagnetic viiiR, xiiiL, 1R, 8L, 14L, 15R, 17R, 26L, 34, 46, 63L, 66L, 68L, 71R, 73R, 74R, 78R, 79L, 81R, 82R, 83L, 87L, R, 88L, 89L, 90L, 91L, R, 92L, 94L, R, 95R, 106L, 120L, R, 123L, R, 125L, 131R, 208L, 214L, 237L, 238L, 239L, 240, 247R, 251L, 252R, 255R, 256L, R, 258L, R, 260R, 261L, 262L, 263L, R, 264R, 266R, 267L, R, 268R, 270R, 276R, 279L, R, 280R, 281L, 283L, R, 291L,  
 Paleontologically Distinct Lithozone 16R  
 Palm Park Formation, New Mexico 131L  
 Palm Springs Formation, California 250L, 258R  
 Palousie Prairie, northwest US 337L  
 Panaca Formation, Nevada 252L, 260R  
 Panama 203R, 212L  
 Isthmus of (Panamanian isthmus; land bridge) 253L, 268R, 269L, 335R, 338L  
 Panhandle, Texas, Oklahoma 264R  
*Pantolambda* Chronozone 72L  
*Pantolambda-Plesiadapis praecursor* (To3) biochron 17L  
*Pantolambda-Plesiadapis praecursor* Interval Zone (To3) 72L, R  
*Pantolambda* zone 70L, 71R, 72L  
 Pants Butte, Nebraska 193R, 194L  
 paratropical forest 319, 320R, 321L, 324L, 327L, 328L  
 Paris Basin 324L  
*Paronychomys-Borophagus littoralis* Assemblage Zone (Hh1) 177L  
 Paskapoo Formation, Alberta, Canada 76R, 77L, 81L, 82L, 83L  
 Pass Peak Formation, Wyoming 108L, 128R  
 Paunsaugunt Plateau, Utah 23R  
 Pawnee Buttes, Colorado 197L  
 Pawnee Creek Formation, Colorado 192R, 195L, 197L, R, 213R, 214L, 215R, 221L  
 P-E Zone (= part of *Deltatherium* Zone, To2) 7RR, 74L  
 Peace River Formation, Florida 207L  
 Peach Springs Tuff, Arizona, California 176R, 219R  
 Peanut Peak Member, Chadron Formation, South Dakota, Nebraska 158L, 162L, 163L  
 Pearlette Ash 237R, 280R, 281R  
 Pearlette B Ash 261L  
 Pearlette O Ash 273R, 281R  
 Peligrian mammal subage, South America 93L, 94R  
 Peninsular Ranges, California 174R, 175L, 176R  
 Penny Farms Formation, Florida 206L  
*Peridiomys-Parahippus* Zone (Ba1), Hepburn's Mesa Formation, Montana 191L  
*Periptychus-Loxolophus* Chronozone 71R, 72L  
*Periptychus-Tetraclaenodon* (To1) biochron 17L  
*Periptychus carinidens/Protoselene opisthacus* Interval Zone (To) 17L, 71R, 72R, 73L  
*Periptychus carinidens/Tetraclaenodon* Interval Zone (To1) 72L, R  
*Periptychus cavirictum-Mixodectes pungens* Interval Zone 17L  
*Periptychus opisthacus-Ellipsodon grangeri* Interval Zone 17L  
 Persistent White Layer (= purplish white layer, PWL) 159, 161L  
 Peters Gulch ash, Idaho 234, 240, 258L  
*Pewelagus* HSD<sub>k</sub> 259L  
*Phenacodus-Ectocion* Acme Zone (Cf3) 17L, 44L, 86R, 88L  
 Piceance Creek Basin, Colorado 85L, 87R, 88L, 108L, 109L, 111L, R, 112R, 114L, R, 115L, 125L, R  
 Pickles Butte basalt, Idaho 258R  
 Picture Gorge Ignimbrite, John Day Formation, Oregon 179R, 189L, 220R  
 Piedra Parada Member, Zia Formation 198L  
 Pine Nut Mountains, Nevada 178L  
 Pine Ridge Escarpment, Nebraska, Wyoming 158R, 194L, 196R  
 Pine Ridge Reservation, South Dakota 193R, 196R  
 Pinole Tuff, California 219R  
 planktonic foraminiferal zonations 89R, 124R, 133L, 134L, 206R, 237L  
 Pleistocene-Holocene boundary 238L, R, 247R

- Pleistocene Series/Epoch, faunas, rocks 232–239L, R, 240, 244, 246L, 249R, 252R, 266R, 269R, 270L, R, 272L, 275R, 276R, 280R, 281L, 284–287L, R, 288L, 290L, 291–295L, R, 336, 337R, 338R
- Plesiadapis anceps/Plesiadapis rex* Lineage Zone (Ti2) 80L, 81L, R
- Plesiadapis cookei* Lineage Zone (Cf2) 86R, 87R
- Plesiadapis churchilli/Plesiadapis simonsi* Lineage Zone (Ti4) 80L, 82R, 83L
- Plesiadapis gingerichi/Plesiadapis cookei* Lineage Zone (Ti6–Cfi) 80L, 84L, 86R, 87L
- Plesiadapis gingerichi*/Rodentia Interval Subzone (Cfi) 80L, 84L, R
- Plesiadapis praecursor/Plesiadapis anceps* Lineage Zone (Ti1) 44L, 80L–81L
- Plesiadapis rex/Plesiadapis churchilli* Lineage Zone (Ti3) 80L, 82L, R
- Plesiadapis simonsi/Plesiadapis gingerichi* Lineage Zone (Ti5) 80L, 83R, 84L
- Pliocene–Pleistocene boundary 233R, 237L, R  
in North America 237L, R
- Pliocene Series/Epoch, faunas, rocks 191R, 201L, 232–233L, R, 237L, R, 239L, R, 240, 244, 246L, 249R, 251L, 252R, 260R, 262L, 266R, 269L, 276R, 285L, 287L, 288L, 290L, 292L, R, 293L, 294L, R, 295R, 330L, 336, 337L, R, 338R
- Plum Point Member, Calvert Formation, Maryland 207R
- Pojoaque Member, Tesuque Formation, New Mexico 199R, 200L
- polar broad-leaved deciduous forest 318R, 319, 321L, 324L, 327L
- Polecat Bench, Wyoming 17R, 77L, 81L, 84R
- Poleslide Member, Brule Formation, South Dakota 160L, 165R, 166L
- Pomerado Conglomerate, San Diego area, California 109L, 118R, 123R, 124L
- Porcupine Hills Formation, Alberta, Canada 70R, 75L
- Powder River Basin, Montana, Wyoming 77L, 80R, 107R, 108L, 111L, 121, 132L, R, 158R
- Powder Wash, Ravens Ridge, Green River Basin, Utah 108R, 115L, 126L
- P–P zone (Dragonian, To1) 72L
- Preboreal pollen zone 238R
- precision (in correlation) xiL, xviiR, xviiiR, 6R, 8R, 9L, 10R, 13L, 16R, 26R, 36R, 85R, 92L, 106R, 121L, 132L, R, 160L, 167R, 194R, 206L, 209L, 214L, 233R, 236R, 259L, 271L, 293R
- Priabonian Stage 330L
- Principle of Paleontological Correlation xviiiL
- Principle of Superposition xviiL
- Principle of Original Horizontality xviiiL
- Principle of Original Continuity xviiiL
- proboscicean datum 171R, 172L, 176L, 177R, 189R, 190L, 195L, 197R, 200L, 203L, 206R, 214L, R, 215L, R
- Protoceras* beds, Brule Formation, Big Badlands, South Dakota 165R, 166L
- Protoceras-Leptauchenia* beds 165R
- Protoselene opisthacus/Mixodectes pungens* Interval Zone (To2) 72R, 74L–75L
- Protungulatum/Ectoconus* Interval Zone (Pui) 44L, 63R–65R
- provincial (ism, ity) 24R, 28L, R, 36L, R, 68R, 69L, R, 95L, R, 119L, 178L, 203R, 209L, 214L, 215R, 232R, 249L, 252R, 272L, R, 273L, 274R, 280R, 290R, 291R, 292–294L, R, 295L, 319, 326R, 327L, R, 328R, 331L, 332L, 335L, 338R
- Pui–Pu3 *see* Puercan
- Puercan mammal age**, faunas, rocks 16R, 17L, 21L, R, 26R, 27L, 28L, R, 32L, 35L, R, 36–37L, R, 43–45L, R, 46, 53L, R, 54, 61R, 62–70L, R, 71L, 73L, R, 78L, 91L, 92R, 94L, 95L, R, 320, 321L, R, 323R, 324L
- definition and characterization 62L–63R
- zonation 63R–70L
- Hanna Basin sole succession for superposition of all zones 66R
- Protungulatum/Ectoconus* Interval Zone (Pui) 63R–65R
- Ectoconus/Taeniolabis taoensis* Interval Zone (Pu2) 65R–67R
- Taeniolabis taoensis/Periptychus carinidens* Interval Zone (Pu3) 16R, 67R–70L
- Puercan–Torrejonian boundary 71L
- Puerco Formation 45L, R, 68L, 70L
- “Puerco Marls” 45L
- Pumpkin Buttes, Powder River Basin, Wyoming 132R, 158R
- Punta Peligro, Argentina 92R, 93L, 94R
- Punta Prieta, Baja California del Norte 131R
- PWL (Purplish White Layer; *see* Persistent White Layer; Ash J, Flagstaff Rim) 159, 161L, 163L, R, 164R, 165L
- Pyramid flora, Nevada 333R
- Quaternary System/Period, faunas, rocks 232R, 237L, 287R, 290L, 291L, R, 293R
- radiocarbon dates, dating 238L, R, 286L, R, 287L, 288L, 291L, R, 293R, 294R, 295L
- radioisotopic dates, dating viiL, R, xiiL, xiiiL, xivR, xviiiL, iR, 8R, 9R, 10L, 14R, 15R, 22L, 24L, 26R, 32R, 44R, 47, 91L, R, 94L, 95R, 106L, 120L, 134R, 138R, 169L, 172R, 176L, 177L, 178L, 179R, 193L, 206L, 208R, 214L, 233L, 247R, 280R
- fission-track method 192R, 194L, 195L, R, 201R, 219L, 237R, 251L, R, 258L, 259L, 261L, R, 264L, 266R
- <sup>40</sup>Ar/<sup>39</sup>Ar method viiR, xviiiL, 8R, 9L, 11L, 22R, 25L, 26R, 63L, 65R, 123R, 135–137, 156L, 158L, 161R, 162L, R, 163L, 164L, 166L, R, 173R, 177R, 178L, R, 179L, 187L, R, 195R, 197R, 199R, 200L, R, 251R, 258R, 264L, R
- K–Ar method viiiR, xviiiL, 8R, 9L, R, 26R, 132L, 173L, R, 187R, 194L, 195L, 201L, 202L, R, 203L, 208L, 251L, 256R, 259R, 271L
- radiometric 14L, 26R, 36R, 91L, R, 160L, 171R, 178L, 196L, 219L, 233L, 237R, 238L, 239L, 240, 251L, 252R, 256L, 258L, 260R, 263R, 264R, 267R, 268L, R, 270L, 281L, R, 283L, 285L, R, 286L, 287L, 294R
- Rak Tuff, California 219R
- Rancholabrean mammal age**, faunas, rocks 232L, 233L, 239L, 240, 244, 246L, R, 247R, 248, 249L, 264L, R, 269R, 270L, R, 273L, 274R, 276L, R, 280R, 282R, 283–288L, R, 289, 292L, 293L, R, 294R
- arrival of *Bison* 285L, R
- definition and characterization 284R–285L
- geographic distribution 288L–290L
- Kennewick sequence 288R, 289R
- Meade Basin sequence 288R
- Medicine Hat sequence 288L, R
- historical treatment 284R
- zonation 287L, R
- Bison* chronology 287L, R
- Loess stratigraphy 287R
- Randlett horizon, Duchesne River Formation 112L
- Randlett Point, Uinta Basin, Utah 118R
- Range Chron xiiL, xivR, 4, 5, 15L
- Range Zone xiL, R, 3R, 4, 5, 6L, R, 15L, 17R, 34L, R, 44L, 113R, 114R, 121R,

- 156L, 162L, 166R, 173L, 177L, 179R, 197R, 198L, 258R, 259L
- Raton Basin, Colorado 108L, R, 111L, R, 125R, 131L
- Rattlesnake Fm., Oregon 220R
- Raven Ridge, Uinta Basin, Utah 108L, R, 114R, 115L, 125–126L, R
- Ravenscrag Formation, Saskatchewan, Canada 26R, 35R, 53L, 62L, 64R, 69L, 76R, 321L
- Rawls Formation, Big Bend area, Texas 201L
- Recent (temporal interval) 238L, 288R
- Red Bird, Wyoming 34R
- Red Light Bolson, Texas 257, 263R
- Regina Member, San Jose Formation, New Mexico 130L
- Relizian benthonic foraminiferal stage, California 171R, 214R
- Relizian-Luisian boundary 171L, R, 214R
- Renova Formation, Montana 160L, 188R
- Reunion magnetic polarity subchron of Matuyama Chron 240, 279R
- resident faunal elements 321R
- Rexroad Formation, Kansas 264R
- Rexroadian subage, Blancan age 253L, R
- Ricardo Faunas, faunal sequence, California 175R, 176R, 177L, 216L, 217L
- Ricardo Group, California 175R, 176R
- Ringold Formation, Washington 232R, 256L, R
- Riochican mammal age, South America 93L, 94L
- Riochican mammal subage, South America 95L
- Rio Chico Formation, Argentina 93L, 94R
- Rio Grande rift, New Mexico 130L
- Riverside County Landfill, California 259R
- Robin's Egg Blue Tuff, Washakie Formation, Sand Wash Basin, Wyoming 116R, 127R
- Rock Springs Uplift, Wyoming 83L, R, 109L
- rock unit vs temporal unit 106R, 160R, 161R, 162L, 163R, 164L, 165R
- Rockyford Ash, South Dakota 166R, 196L
- Rocky Mountains 107L, 117L, R, 179L, 187L, 188L, 191L, 214R, 217L, 254R, 255R, 272L, R, 273L, 274R, 275L, 279L, 280L, 292R, 294L, 318L, 320R, 321L, 326L, 327R, 328L, R, 330L, R, 331R, 332L, 334R, 336R, 338L
- Rodentia/*Plesiadapis cookei* Interval Subzone (Cfi) 86L, 87L, R
- Rosebud Formation, beds, South Dakota 196R, 197L, 209R, 212L
- Rose Creek Member, John Day Formation, Oregon 187L
- Roundhouse Rock Ash, Nebraska 220R
- Rubio Peak Formation, New Mexico 131L
- Runningwater Chronofauna 331R, 332R
- Runningwater faunas, Nebraska 194R, 199L, 206R
- Runningwater Formation, Nebraska 175L, 187L, 191R, 192L, 194R, 208L, R, 212R
- Rupelian Stage 330L
- Saddleback Basalt, California 219R
- Safford Basin, Arizona 257, 261R
- Sage Creek Formation, Beaverhead County, Montana 108R, 134R
- Sage Creek Limestone, Bridger Formation 115R
- Sage Creek Mountain, Wyoming 115R, 128L
- Sage Creek White Layer (SCWL), Bridger Basin, Wyoming 115L, R
- Salamanca Formation, Argentina 94R
- Salmon flora, Idaho 326R
- San Andreas fault, California 175L
- Sand Coulee beds 107R, 108L
- Sandcouleean subage (Wa-0–Wa-2) of Wasatchian mammal age (early Wasatchian) 107L, 113L, 122L, 132L, 135
- characterized 113L
- Sand Creek facies, Willwood Formation 107R
- San Diego area, California 116L, R, 118R, 122R, 123R, 124L, R, 125L
- Sand Wash Basin, Colorado 107R, 108L, R, 115L, R, 116R, 117L, 126R, 127R
- San Francisco Bay Area 171L
- Sangamonian interglacial stage (Pleistocene) 237R, 238L
- San Jacinto fault, California 175L
- San Joaquin Valley, California 171R, 172L, R
- San Jorge Basin, southern Argentina 92R, 93L, 94R
- San Jose Formation, New Mexico 76L, R, 108L, 109R, 111L, 130L
- San Juan Basin, Colorado, New Mexico 16L, R, 24R, 27, 34, 36R, 45L, R, 63L, 64R, 65L, 66–69L, R, 70L, 71–74L, R, 75L, 76L, 78L, 83R, 93R, 94L, 95R, 107R 108L, 109R, 125R, 130L
- San Pablo Group, California 171L
- San Pedro Valley, Arizona 232R, 256L, 257, 261L, R, 270L, 280L
- Santa Ana Mountains, California 174L
- Santa Fe Marls, New Mexico 198R
- Santa Margarita Formation, California 172L
- Santee Ash, Ash Hollow Formation, Nebraska 195R, 220R
- Santiago Formation, San Diego area, California 109L, R, 118R, 124L
- San Timoteo Badlands, California 232R, 251L, 259R, 269L, 277,
- San Timoteo Formation, California 175L, 251R, 278L
- Santa Ana Mesa basalt, New Mexico 262R
- Santa Lucia Formation, Bolivia 93R
- Santonian Stage/Age, faunas, rocks 23L, R
- Sao Jose de Itaborai, Brazil 94L
- Sao Jose de Itaborai Formation, Brazil 94L
- Sappan subage, Irvingtonian age 247L, 273L, R
- Scollard Formation, Alberta, Canada 26R, 27, 32R, 34
- Scottsbluff National Monument, Nebraska 161L
- Scenic Member, Brule Formation, South Dakota 160L, 166L
- Scripps Formation, San Diego area, California 108R, 115R, 123L
- sea level 315L, R, 317L, R, 318L, 323–324L, R, 325, 326L, 327R, 328L, R, 329, 330L, 331R, 333L, 335, 337L
- Seaman Hills, Wyoming 160R, 161L, 163–164L, R
- sea surface temperatures 317R, 318L, R, 320L, 324R, 326L, 330L
- second Nonpareil Ash Zone of the “brown siltstone,” Nebraska 166L
- Seldovian (floral) stage, Alaska 334R
- Senecan subage, Blancan age 247L, 253L, R
- Sentinel Butte Formation, North Dakota 76R, 77L
- Sentinel Butte Shale Member of Fort Union Formation, North Dakota 132L
- sequence stratigraphy 315R
- Sespe Formation, California 109L, R, 118R, 119R, 123R, 124L, R, 125L, 138R, 172R, 173L, 174R, 175L
- Seymour Formation, Texas 282L
- Shanghuan(ian), Asian land mammal age 90R, 91L, R, 324L
- Shanyang Basin, China 91L
- Shark River Marls, New Jersey 132L
- Sharps Formation, South Dakota 166R, 179R, 187R, 188R, 189L, R, 193R, 196L, R, 197L, 209R
- Sheep Creek Formation, Nebraska 175R, 194R, 220R

- Sheep Mountain Table, Big Badlands, South Dakota 165L, 166L
- Sheridanian subage, Irvingtonian age 247L, 273L, R, 274L, 285R
- Shirley Basin, Wyoming 132R
- Shoshonian subage, Uintan mammal age 116L, R, 117L, 136, 138R
- Shotgun Butte, Wyoming 121L
- Shotgun Member, Fort Union Formation, Wyoming 77L
- Sidufjall magnetic polarity subchron of Gilbert Chron 200R, 240, 251R, 252L, 256R
- Siebert Tuff, Nevada 177R
- Sierra Ladrones Formation, New Mexico 270R
- Sierra Nevada 332L, 336R, 337L
- Siesta Formation, California 219R
- Sigmodon* LSD<sub>k</sub> 259L
- Simi Valley area, California 109R 119R, 120L, 124L, R, 125L, 172R, 173L
- Sinostylops* Interval Zone 91R
- Six Mile Creek Formation, Montana 188R
- Skull Ridge Member, Tesuque Formation, New Mexico 199R, 199L, 200L
- Slim Buttes, South Dakota 196R, 197L
- Slim Buttes Formation, South Dakota 109R, 133R
- Smilodon* LSD<sub>k</sub> 259L
- Smoky Hollow Member, Straight Cliffs Formation, Utah 23L
- Snake River Plain, Idaho 232R, 253R, 256L, 256R, 276R
- Soldier Meadow Tuff, Nevada 178L
- South America 37L, 91R, 92R, 93–94L, R, 95L, 218L, 253L, 259L, 268L, R, 269L, 316, 317L, 335R, 336L, 337R
- Southern Great Basin, North America 169L, 214L
- Southern San Joaquin Valley, California 171R, 172L
- South Mountain, California 173L
- South Tejon Hills, California 172R
- South Townsend Basin, Montana 160L
- South Pass, Green River Basin, Colorado 113L, 128R
- Spanish Peaks, Colorado 131L
- Sparnacian Stage/Age, faunas, rocks 16L, 89–90L, R
- Spain 88R, 89L, 334R
- Spitzbergen (dispersal) 90R
- Split Rock Formation, Wyoming 195R, 220R
- <sup>87</sup>Sr/<sup>86</sup>Sr isotope ratios 205–206L, R, 267R, 270R, 283R
- Stadium Conglomerate, California 109L, 118L, R, 123L, R
- Stage 1L, 2, 7, 8L, R, 16L, 171–172L, R, 172R, 174R, 233R, 236R, 234R, 334R
- Starr Flat Member, Duchesne River Formation, Utah (unfossiliferous) 112L
- stasis 290L
- St. David Formation, San Pedro Valley, Arizona 261L, 267L
- St. Mary River Formation, Alberta, Canada 25L
- St. Mary's Formation, Maryland 208L
- Steens Basalt, Oregon 220L
- Stewart Valley flora, Nevada 334L
- Straight Cliffs Formation, Utah 23L
- stratigraphic classification 1R, 2L
- stratigraphic (position) 1L–8R, 45L, 46, 47, 53L, R, 62L, 63L, 65L, 67L, 68L, 71R, 72L, R, 73R, 74L, R, 77L, R, 78L, 80R, 83L, 85L, 89L, R, 90L, 93R, 115R, 116L, R, 120L, 124R, 126L, 128L, 133R, 134R, 156R, 162L, 163R, 164L, 166R, 169L, R, 170L, 174L, 175R, 179L, 187R, 190L, R, 193L, R, 195L, 198–199L, R, 200L, 203L, R, 206R, 207L, 208R, 215L, 216L, R, 237R, 239L, 247L, 250R, 251L, 252R, 254R, 256L, R, 258–259L, R, 261L, R, 263L, 264L, R, 265R, 266L, R, 271L, R, 272L, 278L, 279L, 280R, 281L, R, 286L, 287R, 288R, 290R, 291L, R, 294R, 317R, 338R
- stratotype (type section, area) 1R, 2, 3R, 5, 7, 8L, R, 13L, 15R, 16L, R, 17R, 21R, 23L, R, 25R, 26R, 27L, 28L, 34L, 36R, 45L, R, 49, 53R, 64R, 65L, 66L, R, 67R, 69L, 70L, 71R, 72L, 75L, 79L, 82L, R, 84R, 85L, R, 86L, 87L, 89L, R, 90L, 106R, 107L, R, 111L, 115L, 118R, 120R, 123R, 126L, 127R, 134R, 139L, 158L, 160R, 161R, 162R, 163L, R, 164–165L, R, 166L, 167L, 175R, 178R, 187L, 189L, R, 190L, 193R, 194L, R, 195R, 197L, R, 198R, 203L, R, 206L, 208R, 213R, 233L, R, 237–238L, R, 239L, 261L, 269R, 273R, 275R, 278L, R, 282L, 287R, 295R
- Strict Overlap Assemblage Fossilzone 4, 5, 6L, R, 15L
- subtropical forests 324 R, 326L, 327L, 331L, R, 332L, 332L
- Summit Lake Tuff (Tuff of Big Basin), Nevada 178R
- superposition(al) xviiiL, 5, 16R, 18L, 27L, 45R, 65–66L, R, 68L, 71L, 73R, 74L, 75R, 77R, 94R, 95L, 118L, 121L, 129R, 131L, 133L, 169L, 171R, 172R, 174L, R, 178L, 188L, 198R, 200R, 212R, 215R, 216L, 217R, 251R, 252L, 258R, 260L, 262R, 280R, 282L, 283L, 286L, 287R, 288L, R, 290L
- Sunrise Pass Formation, Nevada 260L
- Susanville flora, California 326R, 327L
- Sutro flora, Nevada 332L
- Swallow Ash, Ash Hollow Formation, Nebraska 195R, 220R
- “Sweetwater” Formation, San Diego area, California 109R, 124L
- Swift Current Plateau, Saskatchewan, Canada 133L
- Suchilquitongo Formation, Mexico 202L
- Sucker Creek Formation, Oregon 187R
- Suwanee Limestone, Florida 205R
- Sycamore Canyon Formation, California 171L, 219R
- synchronous 21R, 32L, 37L, 91R, 106R, 176L, 268L, 317R, 320R, 324L, 236R, 332L
- Tabernacle Butte, Wyoming 108R, 113R, 128L, R
- Taeniolabis-Periptychus* (Pu3) biochron 16R
- Taeniolabis taoensis/Periptychus carinidens* Interval Zone (Pu3) 16R, 63R, 67R
- Talkeetna Mountains, Alaska 111L
- Tamiami Formation 251L
- Tampa Limestone Member, Arcadia Formation, Florida 204R
- Tapicitos Member, San Jose Formation, New Mexico 130R
- Tatman Formation, Wyoming 108R, 122L
- Taxon-range Chron xivR, 4, 5, 15L
- Taxon-range Zone xiR, 2, 4, 5, 6L, 15L, 16R, 17L
- Tecuya Formation, California 171L, 219R
- Teewinot Lake flora, Wyoming 337L
- Tehachapi flora, California 332L
- Tehachapi Mountains, California 175R
- Teichert Gravel Pit, California 286L
- Tejon Hills, California 172L, R
- Temblor Formation, California 171R
- Temecula Arkose, California 257, 260L
- temperate climate, flora 318R, 324R, 327L, R, 328L, 330R, 334R, 327L
- temperature gradient (poleward) 315L, 318R, 230R, 326R
- Tepee Trail Formation, East Fork Basin, Wyoming 108R, 116L, 117L, R, 118L, 120R, 121L, 122L
- Tertiary System/Period, faunas, rocks 9L, 21L, R, 32L, R, 35L, 35R, 36L, R, 37L, 43R, 62L, 64R, 115R, 116L, R, 127L, 129R, 171L, 188L, R, 191R, 195R, 202L, R, 232R, 230L
- Tesuque Formation, New Mexico 199L, R, 200L, 214L
- Tetraclaenodon/Pantolambda* Interval Zone (To2) 72L, R
- Texas Coastal Plain 204L



- Thanetian Stage/Age, Europe, Africa  
88R, 89L, R, 92L, R
- thermoluminescent (TL) dates 285R,  
287R
- Thousand Creek beds, Nevada 178R
- Three Forks Basin, Montana 160L
- Thunder Mtn. flora, Idaho 326R
- Thvera magnetic polarity subchron of  
Gilbert Chron 233R, 240, 252L
- Ti1–Ti6 *see* Tiffanian
- Tiffanian mammal age**, faunas, rocks  
17L, R, 43L, R, 44L, 48, 62R, 64L,  
70R, 71R, 73L, 75R, 76–80L, R, 321L,  
323R, 324L  
definition and characterization 78R,  
79L, R  
zonation 79R  
*Plesiadapis praecursor*/*P. anceps* Lin-  
eage Zone (Ti1) 79R, 80L–81L,  
82L, 83R  
*Plesiadapis anceps*/*P. rex* Lineage  
Zone (Ti2) 78L, 79L, 80L, R,  
81L–R, 82L  
*Plesiadapis rex*/*P. churchilli* Lineage  
Zone (Ti3) 77L, 79L, 81R, 82L–R,  
83L  
*Plesiadapis churchilli*/*P. simonsi* Lin-  
eage Zone (Ti4) 79L, 80L,  
82R–83R, 89L, 91R  
*Plesiadapis simonsi*/*P. gingerichi* Lin-  
eage Zone (Ti5) 79L, R, 80L, 83L,  
R–84L, 91R  
*Plesiadapis gingerichi*/Rodentia In-  
terval Subzone (Ti6) 79R, 80L,  
84L–R  
*Plesiadapis gingerichi*/*P. cookei* Lin-  
eage Zone (Ti6-Cfi) 79L, R, 80L,  
84L, R, 85R, 86R, 87L, 89L
- Tiffanian-Clarkforkian boundary 84L,  
85L, 85R, 86L, R
- Tiffany beds, Colorado 76L, R
- Tiffany, Colorado 76L
- Tijeras Arroyo, Bernalillo County,  
New Mexico 262R, 270R, 271L, 271R,  
277, 280L
- time-average(d) 35L, 293R
- Tin Roof Tuff, John Day Formation,  
Oregon 173L, 179R, 211R, 220R
- Tipton Tongue, Green River Forma-  
tion, Wyoming 127L
- Titanotherium* beds 160R
- Tiupampian mammal subage, South  
America 93L
- Tetraclaenodon-Pantolambda* (To2)  
biochron 17L
- To1–To3 *see* Torrejonian
- Toadstool Park, Nebraska 160R, 161L,  
162L, 164R, 165L, 166L
- Togwotee Pass, Wyoming 77L, 85L, R,  
87L, 87R, 108L, R, 111L, 120L, R
- Tongue River Formation, Montana,  
North Dakota; member of Fort  
Union Fm. 76R, 77L, 80R, 81L
- Tornillo Formation, Texas 62L, 69R,  
70R, 71R, 74R, 79L, 85L
- Tonopah Tuff, Nevada 220L
- Torrejon Formation 45L, R, 70L
- Torrejon Wash, New Mexico 45L, 74L
- Torrejonian mammal age**, faunas,  
rocks 16R, 17L, 43L, R, 44L, 45L, R,  
47, 48, 62R, 69R, 70–73L, R, 74R,  
75L, R, 76R, 77–79L, R, 80R, 81R,  
88R, 91L, R, 92R, 94L, 95L, 323L, R,  
342L  
definition and characterization  
71L–72L  
zonation 72L–76L  
*Periptychus carinidens*/*Protoselene*  
*opisthacus* Interval Zone (To1)  
17L, 63L, 71–73L, R, 74L  
*Protoselene opisthacus*/  
*Mixodectes pungens* Interval  
Zone (To2) 72L, R, 73R,  
74L–75L, 91L, 93R, 94L  
*Mixodectes pungens*/*Plesiadapis*  
*praecursor* Interval Zone (To3)  
72L, R, 73L, 74L, 75L–76L, 77L,  
R, 79L, 80R, 81R, 91R
- Torrejonian-Tiffanian boundary 75R,  
77L–79L, 80R
- Torrey Sandstone, California 123L
- Torreya Formation, Florida 206R
- Trail City Member, Fox Hills Forma-  
tion, South Dakota 34L
- Trans-Mexican Volcanic Belt, Mexico  
201L, R
- Trans-Pecos (area), Texas 112L, 115R,  
116L, 136, 138R, 160L, 161R, 162R
- Transverse Ranges, California 172L, R,  
174R
- Tres Cruces, Bolivia 92R
- Triceratops* Zone, Scollard Formation,  
Alberta, Canada 32R
- tropical forest 318R, 319, 321L, 324L, R,  
326–327L, R, 328L, 330L, 331L, 332L,  
335R
- Trout Peak Trachyandesite, Wyoming  
122R
- Trubi Formation (Trubi Marls), Italy  
233R
- Trunk Butte (= Big Cottonwood  
Creek) Member, Chadron Forma-  
tion, Nebraska, Wyoming 161L
- Tuff of Big Basin, Nevada 178R, 220L
- Tule Formation, Texas 250R, 262R,  
271L, 282L
- Tulelake flora, California 337L
- Tulloch Formation, Montana 28R, 36L,  
R, 53L, 62L, 63L, 64L, 68R, 70R, 71R,  
73L, 95R
- Turlock Lake Formation, California  
278R
- Turonian Stage/Age, faunas, rocks 23L,  
317R, 318L
- Turtle Butte Formation, South Dakota  
196R
- Turtle Cove Member, John Day For-  
mation, Oregon 179L
- Tusahoma Formation, Mississippi  
108L, 111L, 133R
- Turtle Bluffs Member, Bridger Forma-  
tion, Wyoming 108R, 111R, 117L
- Twinbuttean subage (Br-3), Bridgerian  
mammal age 115R
- Twin Buttes Member, Bridger Forma-  
tion 108R, 111L, 115L, R
- Two Medicine Formation, Montana 24L  
type locality 107R
- Uinta A-C 111R, 115R, 116L, 117L, 126L,  
R, 136
- Uinta B1 111R, 116L, 117L, R, 118L, 120R
- Uinta B2 111R, 118L
- Uinta Basin, Colorado, Utah 107L,  
108–109L, R, 111R, 112L, 114L, R, 115L,  
R, 118L, R, 125L, R, 126L, 131L, 136,  
138L
- Uinta Formation, Colorado, Utah 111R,  
112, 117R, 118L, 125L, R, 126L, R, 138R
- Uintan-Duchesnean boundary 118R,  
119L, 138R
- Uintan mammal age**, faunas, rocks  
107L, 109, 111R, 112L, 115–118L, R,  
119L, 120R, 121–124L, R, 125L, 126L,  
127R, 128L, 129L, R, 130R, 132L, 133L,  
R, 134L, R, 136, 138L, R, 327R, 328L, R  
definition and characterization  
117L, R  
zonation 117R–118R
- Shoshonian subage (Ui-1?) 116L, R,  
117L, 136, 138R
- Bridger E (early Uintan) 108L, 111L,  
116L, R, 117L, 128L, 129R, 136
- Uinta A 111R, 115R, 116L
- Uinta B 111R, 116L, 117R, 126L  
Uinta B1 109L, 111R, 116L, 117L, R,  
118L, 120R  
Uinta B2 109L, 111R, 117R, 118L
- Uinta C 111R, 116L, 117R, 126R, 136
- Uii, Shoshonian 116L, R, 117L, 136,  
138R
- Ui2 108, 109, 116L, 117L, R, 118L, 123R,  
126R, 127R, 129R, 136
- Ui3 108, 109, 116L, 117R, 118L, 121R,  
123R, 124L, 126R, 129R, 130L, 131R,  
133L, 134L, R, 136,
- Umayo Formation, Peru 92R
- unconformity (ies), unconformably  
xiiiL, xviiiR, 3L, 6R, 10L, R, 11L, 13L,  
18L, 113R, 116L, 122L, 129L, 130L, 131L,

- unconformity (*continued*)  
 177L, 187L, 188L, R, 193R, 198L, 202L,  
 233R, 258R, 263R, 276L, 281L, 288L, R  
 Upper Bone Valley Formation, Florida  
 251L  
 Upper John Day Formation, Oregon  
 179L  
 Upper Harrison beds, Nebraska 191R,  
 194L, R, 196R, 198R, 212L, R  
 Upper Harrison Formation, Nebraska  
 194L  
 Upper *Oreodon* Beds, Brule Forma-  
 tion, Big Badlands, South Dakota  
 159, 165L, R, 166L  
 Upper Nodular Zone, Big Badlands,  
 South Dakota 165L  
 Upper Tuffaceous Zone, New Mexico  
 221L  
 Upper Whitney Ash, South Dakota 166L  
 Uquian mammal age (South America)  
 269L  
*Ustatochoerus profectus/Copemys rus-*  
*selli* Assemblage Zone (Cl1) 177L  
 Uzbekistan 23R
- Valentine Formation, Nebraska 177R,  
 191R, 192R, 193R, 195L, R, 197L, R,  
 200L, 215R, 216L, R  
 Vallecito-Fish Creek section, Anza-  
 Borrego Desert State Park, Califor-  
 nia 244, 245, 258R, 259R, 260L, 271L,  
 272R  
 Valle de Oaxaca Gaben, Mexico 202L  
 Vaqueros Formation, California 174R  
 Ventura County area, California 118R,  
 122R, 124L, 125L, 172R  
 Vieja Group, Texas 130L, 160L  
 Vieja-Ojinaga area, Presidio County,  
 Texas 129L, R, 138R  
 Villafranchian Stage, Italy 236R,  
 Virgin Valley beds, Nevada 178R,  
 Virgin Valley Formation, Nevada 178R,  
 220L  
 Vista Member, Brule Formation, Col-  
 orado 160L, 166L  
 volcanoclastic deposit(s) 173L, 178L, R,  
 188R, 194L, R, 198R, 202L, 203R, 213L  
 Vrica, Italy 237L
- Wa-o–Wa7 *see* Wasatchian  
 Wagon Bed Formation, Wind River  
 Basin, Wyoming 108L, R, 109L, 115L,  
 118L, 121R, 132R, 134R  
 Wagonhound Member, Uinta Forma-  
 tion (= Uinta A and B), Utah 109L,  
 111R, 117R  
 Wahweap Formation, Utah 23R  
 Wapiti Formation, Wyoming 122L  
 Wapiti Valley, Wyoming 122R
- warm-temperate forests 324R, 328L,  
 330R, 331R, 332L, 333R, 337L  
 Wasatch beds 107R  
 Wasatch Formation, Wyoming, Utah  
 77L, 85L, 107–108L, R, 112R, 113L,  
 114–125L, R, 126R, 127L, R, 128R,  
 132L, R  
 type locality 107L, R  
 Wasatch Group 107L, 128R  
**Wasatchian mammal age**, faunas,  
 rocks 43R, 46, 74R, 77R, 85R, 86L, R,  
 87L, 89L, R, 90L, R, 92L, 106R, 107L,  
 R, 109, 111L, 112–114L, R, 115R, 120L,  
 R, 122L, R, 123L, 125–128L, R, 129L,  
 130–134L, R, 135, 138L, 139L, 324R,  
 235R, 326L, 327R, 338R  
 definition and characterization  
 112R–113L  
 zonation 113L  
 Sandcouleean subage 107L, 113L,  
 122L, 132L, 135  
 Graybullian subage 107L, 111L, 113L,  
 122L, 130R, 132L, 135, 325R  
 Lysitean subage 107L, R, 111L, 113L,  
 122L, R, 130R, 135  
 Lostcabinian subage 107L, R, 113L, R,  
 121L, 122R, 135  
 Wa0, Sandcouleean 107L, 113L, 122L,  
 125R, 131R, 132L, 135, 320L, 323,  
 324R, 325L, R  
 Wa1, Sandcouleean 125R, 131R, 133R  
 Wa2, Sandcouleean 108, 111L, 113L,  
 120R, 126L, 132L  
 Wa3, Graybullian 108, 113L, 120L,  
 122L, 127L, 131L, 132L, R, 133R  
 Wa4, Graybullian 113L, R, 135, 138L  
 Wa5, Graybullian, 108, 111L, 113L,  
 125R, 126L, 130L, 131L, 132R, 133R,  
 135, 138L  
 Wa6, Lysitean 111L, 113L, 122L, R,  
 130R, 135  
 Wa7, Lostcabinian 107L, R, 113L, R,  
 121L, 122R, 135  
 Wasatchian-Bridgerian boundary 113L,  
 R, 114L, 138L, 139L  
 Wasatchian Stage/Age 16L  
 Wasatch Station, Weber Canyon,  
 Summit County, Utah 107L, R  
 Washakie A (faunal level) 127R  
 Washakie B (faunal level) 127R  
 Washakie Basin, Wyoming 70R, 74R,  
 75R, 85L, 87L, 107R, 108L, 115L, R,  
 116R, 126R, 127L, R, 135, 136, 138L  
 Washakie Formation, Wyoming, Col-  
 orado 108R, 109L, 115L, 116R, 117L, R,  
 127L, R, 136  
 Western Interior 24R, 26R, 28L, 32L,  
 34L, 35R, 36L, R, 37L, R, 39L, 93R,  
 122R, 321R
- western montane coniferous forest  
 326L, 332L, 333R, 335L, 337L  
 Western Nevada, North America  
 177R  
 West Fork, Gallegos Canyon, San Juan  
 Basin, New Mexico 45R, 48, 66L,  
 67R, 72R  
 Weta Paleosol, South Dakota 158L  
 Wheatland County, Montana 24L  
 White Ash No. 4, Skull Ridge Member,  
 Tesuque Formation, New Mexico  
 199R  
 White River Chronofauna 166R, 173L,  
 179R, 193L, 205L, 206L, 209R, 328R,  
 330L, R, 331L, R  
 White River Group, Colorado, Ne-  
 braska, North Dakota, Montana,  
 South Dakota 156R, 158L, R, 160L,  
 166R, 167L, 179R, 188R, 193L, 194R,  
 196L, 209R  
 deposition 158R  
 White River relicts (taxa from White  
 River faunas) 171R, 196L, 204R,  
 205L, R  
**Whitneyan mammal age**, faunas,  
 rocks 156L, R, 159, 160L, 165–166L, R,  
 167L, 325, 330R  
 definition and characterization 165R  
 zonation 165R  
*Leptauchenia major* Interval Zone  
 (early Whitneyan) 165R, 166L  
*Merycoiodon major* Interval Zone  
 (late Whitneyan) 166L, R  
 Whitneyan-Arikarean boundary 166R,  
 209R  
 Whitney A, Brule Formation, Big Bad-  
 lands, South Dakota 166L  
 Whitney B, Brule Formation, Big Bad-  
 lands, South Dakota 166L  
 Whitney C, Brule Formation, Big Bad-  
 lands, South Dakota 166L  
 Whitney Member, Brule Formation,  
 Nebraska, Wyoming 158L, 160L, R,  
 165R, 166R  
 Wiggins Formation, Wyoming 109L,  
 117R, 118L, 121L, R, 122R  
 Wild Horse Mesa Tuff, California  
 176R, 219R  
 Willard Canyon Tuff, Sespe Forma-  
 tion, California 173L, 219R  
 Wilkins Peak Member, Green River  
 Formation, Wyoming 113R, 114L,  
 127L, 128L  
 Williamsburg Formation, South Car-  
 olina 77L  
 Williams Fork Formation, Colorado  
 25L  
 Williston Basin, Montana, North  
 Dakota 26L, 51, 52, 77L, 82L, 83L, 111L

- Willwood Formation, Wyoming 11R, 17L, R, 85L, R, 107L, R, 108L, 114L, 122L, R, 126R, 132L, R, 135
- Wind River Basin, Wyoming 24L, 77L, 80L, 83L, 84R, 87L, R, 107R, 108–109L, R, 111R, 113L, R, 114L, 118R, 120–121L, R, 122L, 125R, 131L, 132L, R, 134R, 158R
- Wind River Formation, Wyoming 107L, 108L, 113L, 114R, 121L, 122L, 132L, R
- Wisconsinan glacial stage (Pleistocene) 237R, 238L, 287L, 288L, R
- Wodehousia spinata* Assemblage Zone 32L
- woodland-savanna forest 326L, 327L, 332L
- Wood Mountain Plateau, Saskatchewan, Canada 192L
- Woods Mountain volcanic center, California 176R
- Yarmouthian interglacial stage (Pleistocene) 237R, 238L
- Yautepec Tuff, Mexico 221L
- Yellowstone National Park 111L, 191L
- Yepómera, Chihuahua, Mexico 178L, 200R, 201L, R, 202L, 218R
- Younger Dryas pollen zone 238R
- Ypresian Stage/Age, Europe 89R, 92R
- Yukon Territory 239L, 249L, 277, 280L, 321L
- Yushe Basin, China 254R
- Zanclean Stage, Capo Rossello, Sicily 233R, 251L
- Z coal complex 28R
- Zia Formation, New Mexico 196L, 198R, 199L
- zoogeography (ic); provinces 169L, 170L, 174R, 177R, 178L, 179L, 187R, 191L, 192R, 199L, R, 204R, 206L, 209L, 213L, 214L, 215R, 324L, 338R
- zone, zonation xi–xivL, R, xviiR, xv–iiL, 2–3L, R, 4, 5, 6–7L, R, 8L, 10L, 11L, 14R, 15–17L, R, 32L, R, 34L, R, 35L, 43R, 44L, R, 53R, 61L, R, 63–88L, R, 107L, 111L, R, 113L, R, 114R, 121R, 127R, 133R, 134L, 138L, 156L, 162R–163R, 164L–166L, 170L, 171R, 173L, 177L, R, 179R, 180, 188L, 191L, R, 198L, 206L, R, 247R, 248, 253R, 258R, 259L, 263L, 280L
- Zoyatal Tuff, Mexico 201R

