SECOND EDITION

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Acquisitions Editor: Lisa McAllister Managing Editor: Kerry Barrett Project Manager: Fran Gunning Senior Manufacturing Manager: Benjamin Rivera Marketing Manager: Angela Panetta Creative Director: Stephen Druding Production Services: Nesbitt Graphics, Inc. Printer: R.R. Donnelley, Shenzhen, China

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530 Walnut Street Philadelphia, PA 19106 USA LWW.com

First edition, © 1998 Lippincott-Raven Publishers

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Printed in China

Library of Congress Cataloging-in-Publication Data

Greenspan, Adam.

Differential diagnosis in orthopaedic oncology / Adam Greenspan, Gernot Jundt,

Wolfgang Remagen.—2nd ed.

p.; cm.

Rev. ed. of: Differential diagnosis of tumors and tumor-like lesions of bones and joints/

Adam Greenspan and Wolfgang Remagen. 1997.

Includes bibliographical references and index.

ISBN 978-0-7817-7930-2 (alk. paper)

1. Bones—Tumors—Diagnosis. 2. Joints—Tumors—Diagnosis. 3. Diagnosis, Differential. I. Jundt, Gernot. II. Remagen, Wolfgang. III. Greenspan, Adam. Differential diagnosis of tumors and tumor-like lesions of bones and joints. IV. Title.

[DNLM: 1. Bone Neoplasms—diagnosis. 2. Diagnosis, Differential. 3. Joint Diseases—diagnosis. 4. Soft Tissue Neoplasms—diagnosis. WE 258 G815d 2007]

RC280.B6G74 2007 616.99'471075-dc22

010.99 4/10/5-dc2

2006020800

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To my wife Barbara, my children Michael, Samantha, and Luddy, and my grandchildren Anna and Sydney, with love; and to the memory of my mother Eugenia, and my father Bernard, a brilliant physician, who taught me my ABC's of the medical profession.

-A.G.

To Inge for her constant support and encouragement while I took time off from her to work on this book.

-G.J.

To my wife Karin for her patience, understanding, and encouragement. -W.R.

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Preface

The first edition of this text, titled Differential Diagnosis of Tumors and Tumor-like Lesions of Bones and Joints, was published in 1998. The past eight years have brought significant changes to the imaging of musculoskeletal lesions, as well as changes in the pathologic classification of many of the tumors. In particular, further technological advances in the field of radiology, mainly technical improvements in orthopaedic imaging of bone and soft tissue tumors using positron emission tomography (PET), computed tomography (thin-section CT and 3-D CT studies), and magnetic resonance imaging (MRI), as well as wide use of enzyme histochemistry, immunohistochemistry, and genetics (flow cytometry, cytogenetics, and molecular cytogenetics) in pathology, prompted the need for a new edition of this book. The title has been changed to Differential Diagnosis in Orthopaedic Oncology, and although the main emphasis is on lesions of the bones and joints, some of the soft tissue tumors have also been included (the scope of this text precluded discussion of all the soft tissue tumors). A new co-author has been added; he is Gernot Jundt, M.D., Professor of Pathology at the Bone Tumor Reference Center of the Institute of Pathology, University Hospital in Basel, Switzerland. Dr. Jundt is a panelist of the Working Group of the World Health Organization (WHO), the body that, in 2002, revised the old classification and proposed a new classification of several tumors of soft tissue and bone. This addition guarantees inclusion of the most upto-date information on histopathologic classification and differential diagnosis of musculoskeletal lesions.

As in the first edition, the thrust of this text is facilitation of the complex differential diagnosis of bone and joint tumors and tumor-like lesions that confronts the radiologist, pathologist, and orthopaedic oncologist. Again, the emphasis is on radiologic and histopathologic approaches to the same lesion, because the differential diagnosis of a given tumor considered by the radiologist may vary from that considered by the pathologist. The text provides an overview of the diverse radiologic and histopathologic appearances of benign and malignant musculoskeletal lesions, including typical and atypical features. Each chapter presents information concerning clinical presentation of the lesion, imaging techniques and radiologic features, histopathology, and radiologic and pathologic differential diagnosis. Pathology sections stress the use of immunohistochemistry and genetics as they are applied to differential diagnosis. As in the

previous edition, the chapters are richly illustrated with radiographs, CT and MR images, photomicrographs in color, and schematic drawings. Important diagnostic features are provided in concise tables, and a summary of differential diagnoses (both radiologic and pathologic) is included at the end of each section in the form of a schematic drawing that also depicts the less likely possibilities not discussed in the text.

There are many changes, additions, and improvements in this new edition. The book has received a new design, and the Table of Contents is now more detailed. The captions for the illustrations have been improved, with the diagnosis placed at the beginning of the legend in boldface type. Technically suboptimal figures have been either deleted or substituted with better-quality images (particularly in the pathology sections). Outdated text and references have been deleted and replaced with current ones. The text has been revised to include additional MRIs, CTs, and 3-D CT images. Several new sections have been added to almost every chapter. For example, in Chapter 1 the latest information on PET has been included. Also revised are the pathology sections, with addition of basic techniques and methods of decalcification, special stains, enzyme histochemistry, immunohistochemistry, and genetics in bone tumors. Chapter 2 has been completely rewritten, with addition of new material, particularly on bone island, osteosarcoma associated with syndromes, and description of new developments in cytogenetics and molecular cytogenetics of bone-forming tumors, both of which are on the cutting edge of research into these lesions. Moreover, the new classification of osteosarcomas has been added in the form of a diagram. Chapter 3 includes new text on myxoid chondrosarcoma, and significant changes have been made to the section discussing extraskeletal chondrosarcomas. In Chapter 4 more information on Jaffe-Campanacci and Mazabraud syndromes has been provided, along with new material on focal fibrocartilaginous dysplasia of long bones, pachydysostosis of the fibula, and liposclerosing myxofibrous tumor of bone. The reader will also find the recently revised concepts on fibrosarcoma and malignant fibrous histiocytoma. Because of the latest viewpoints on the so-called small, blue round cell tumors of bone and their classification, this section in Chapter 5 had to be substantially rewritten to include differential diagnoses based on genetic alterations. Similarly, because of new approaches to benign

and malignant vascular lesions, particularly the recent redefinition of biological classification of vascular anomalies and a recent attempt by WHO to clarify the existing nomenclature for malignant vascular tumors, Chapter 6 required substantial revision. Minor changes have been also made in Chapters 7 to 9; however, all chapters have received up-to-date references.

The histopathologic material, with a few exceptions, was obtained from the Swiss Bone Tumor Reference

Center at the Institute of Pathology, University of Basel, Switzerland.

This text has been written primarily for the audience of radiologists, pathologists, and orthopaedic oncologists, although other physicians interested in orthopaedic oncology may also find it useful.

Adam Greenspan Gernot Jundt Wolfgang Remagen

Acknowledgments

We would like to acknowledge the guidance and help received in the preparation of this text from our friends at Lippincott Williams & Wilkins in Philadelphia, Pennsylvania. In particular, Lisa McAllister, Executive Editor, was instrumental in launching the second edition of this book, and Kerry Barrett, Senior Managing Editor, demonstrated effective coordination skills during all phases of preparation of this volume. Special thanks go to Stephen Druding, Design Coordinator, to Joseph DePinho, who designed the cover, and to Bill Donnelly at WT Design, who redesigned the interior of the book.

We also would like to thank several individuals from the Department of Radiology, UC Davis Medical Center in Sacramento, California. A special note of acknowledgment goes to Deborah Hoang for timely preparation of the manuscript, to Angela Michelier for her invaluable secretarial assistance, and to Julie Ostoich, senior photographer, and Aaron Peterson, for help in creating some digital illustrations.

Thomas Schürch and Jan Schwegler from University Hospital in Basel, Switzerland, are acknowledged for electronic reproduction of the radiographs and gross pathologic sections, and Petra Huber for constant help with finding cases, histopathologic slides and radiographs, and preparation of references. We are grateful to Sharon Rule for her meticulous editing of some of the sections of this book. Finally, we would like to express our gratitude to Joanne (Bonnie) Boehme, a Project Manager from Nesbitt Graphics, for supervision and help during the final composition of this text.

As with the previous edition, this project could not have been successfully completed without the prudent and dutiful efforts of the many individuals acknowledged here.

CHAPTER 1

Radiologic and Pathologic Approach to Bone Tumors

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Molecular Cytogenetics 34 The ideal therapeutic goals for managing patients with primary or metastatic lesions can be incorporated into a triad of important factors: (a) do not overtreat a benign bone tumor; (b) do not undertreat a malignant bone tumor; and (c) do not take an incorrect biopsy approach to the lesion, because this may suggest a need for radical rather than more conservative surgery (117,133,155). Achieving these goals depends on close cooperation among the radiologist, the pathologist, and the tumor surgeon (7,75,83).

Several factors make precise diagnosis of osseous tumors difficult. Although radiographic features have a high correlation with malignancy, benignity, and sometimes even an exact histologic diagnosis, radiographic determination of these characteristics is based on statistical probabilities. Moreover, errors in the radiologic interpretation may occur. These can usually be ascribed to failure to recognize a specific pathologic finding or to misinterpretation of normal structures as pathologic. Regardless of how persuasive a radiologist's clinical assessment of the biologic potential of a lesion may be to a pathologist colleague, any lesion, no matter how radiologically typical it appears, may represent an entirely different entity on histologic examination (68). A confident radiologic diagnosis may not outweigh the microscopic appearance of the lesion.

Both the radiologist and the pathologist play important roles by providing the diagnosis and/or differential diagnosis of a bone tumor, thus aiding the clinician in the complex process of patient management. Before a final diagnosis or differential diagnosis is made, clinical information, radiologic imaging, and pathologic material should be carefully studied and correlated. The radiologist has the significant advantage of being able to view the three-dimensional extent of a bone tumor, whereas the pathologist can view only a small biopsy specimen of the lesion selected by the surgeon (133), which may not reflect the histology of the entire lesion. Obviously, viewing the entire resected specimen enhances the final pathologic evaluation. Many years ago, Ewing commented that "the gross anatomy (as revealed in radiographs) is often a safer guide to a correct clinical conception of the disease than the variable and uncertain nature of a small piece of tissue" (48). It is important to emphasize that the differential diagnosis contemplated by the radiologist may be identical to or different from that contemplated by the pathologist. For example, a radiologist who is evaluating a lesion that looks like an aneurysmal bone cyst must include in the differential diagnosis the possibility of telangiectatic osteosarcoma. Likewise, the pathologist who is looking at the histologic sections of the same lesion must also consider the possibility of telangiectatic osteosarcoma because both lesions have several histopathologic similarities. On the other hand, in the radiologic evaluation of a purely lytic lesion at the articular end of a long bone that looks like a giant cell tumor, plasmacytoma might enter into the differential diagnosis, whereas for a pathologist examining the same lesion (which in fact is a giant cell tumor), the differential diagnosis will obviously include other lesions that may contain giant cells. However, the pathologist would not consider plasmacytoma as one of the differential possibilities because of the completely different histologic pattern of this tumor.

For teaching purposes, the diagnostic approach to bone tumors is discussed separately from the radiologic and the pathologic point of view.

Radiology

In general, the imaging of musculoskeletal neoplasms can be considered from three standpoints: detection, diagnosis and differential diagnosis, and evaluation (staging) (Fig. 1-1). Detection of a bone tumor does



Figure 1-1 Imaging of tumors. Imaging of musculoskeletal neoplasms can be considered from three aspects: detection, diagnosis (or differential diagnosis), and evaluation.

not always require the expertise of a radiologist. The clinical history and the physical examination are often sufficient to raise the suspicion of a tumor, although radiography is the most common means of revealing one. Skeletal scintigraphy can also pinpoint lesions, but its findings are nonspecific.

Despite the dramatic advances in imaging technology that have occurred in recent decades, especially the introduction of cross-sectional imaging modalities such as computed tomography (CT) and magnetic resonance imaging (MRI), radiography remains the single most important modality for establishing a diagnosis (69) and serves as the basis for differential diagnosis (37,115,136). It yields the most useful information about the location and morphology of a lesion, particularly concerning the type of bone destruction, calcifications, ossifications, and periosteal reaction. Conventional tomography, although presently rarely implemented, can be a useful diagnostic tool, particularly on those occasions when questions arise regarding cortical destruction, periosteal reaction, or mineralization of the tumor matrix. It can also detect occult pathologic fracture (1). Scintigraphy is only occasionally helpful in making a specific diagnosis but can be valuable in distinguishing, for example, multiple myeloma from similar-appearing metastases (121) or for distinguishing a benign or malignant sclerotic tumor from a bone island (70).

The imaging advances of the past decade have been most profoundly felt in the evaluation (staging) of bone tumors (46,102,144,145). The multiplanar capabilities and unsurpassed soft tissue contrast offered by CT (24,114) and MRI (10,11,103) have rendered these modalities indispensable in tumor staging (15,33,57,64, 148). They have enabled radiologists to determine tumor size, location, and configuration more accurately than ever before and have facilitated demonstration of intra- and extramedullary extension of tumors and the relationship of tumors to individual muscles, muscle compartments, fascial planes, and neurovascular bundles, as well as to neighboring joints and organs (12,13, 189). MRI, in fact, affords more accurate anatomic staging than any other imaging method (19,20,32). Nevertheless, it is rarely helpful in differentiating benign from malignant tumors, despite attempts to identify signal intensity patterns, the appearance of tumor margins, the presence of edema, and neurovascular bundle involvement as markers for this essential determination (76,93).

Other techniques also play roles in evaluation of bone tumors. These include scintigraphic techniques using technetium (Tc-99), gallium citrate (Ga-67), and indium (In-111)–labeled white blood cells, as well as single photon emission computed tomography (SPECT), among others (12,13,47,55,92,121,189). Because these imaging methods use radiopharmaceutical labeling, they can provide information about the pathophysiologic function of bone and surrounding soft tissues, as well as the size and location of a lesion (120).

Most recently the application of positron emission tomography (PET) for diagnosis of musculoskeletal neoplasms gained a worldwide acceptance (2,21,104,182). PET differs from other single-photon radionuclide scans in its ability to correct for tissue attenuation signal loss and its relatively uniform spatial resolution (52). This technique contributes unique information regarding metabolism of musculoskeletal lesions (51) based on observations of high glycolysis rates in malignant tissues (186). In particular, whole-body ¹⁸F-labeled 2-fluoro-2-deoxyglucose (¹⁸FDG)-PET scanning is a valuable adjunct in identifying primary, recurrent, and metastatic cartilage malignancies (52).

Arteriography is used mainly to map out bone lesion and to assess the extent of disease. It is also used to demonstrate the vascular supply of a tumor and to localize vessels suitable for preoperative intraarterial chemotherapy as well as to demonstrate the area suitable for open biopsy, because the most vascular parts of a tumor contain the most aggressive components. Occasionally, arteriography can be used to demonstrate abnormal tumor vessels, corroborating findings revealed by radiography. In selected cases, this technique may help to differentiate osteoid osteoma from bone abscess.

Ultrasonography only rarely contributes information useful in differential diagnosis of malignant and benign musculoskeletal tumors (18).

Radiography

Conventional radiographs continue to provide a wealth of information whose implications can be helpful in the precise diagnosis of a bone tumor (69,96,106,140, 192,210). However, radiography does not provide a diagnosis in every case (86). Some types of tumor can be diagnosed with certainty by their radiographic appearance alone, other types can be diagnosed only with various degrees of probability, and still others may have an appearance compatible with that of more than one type of tumor and therefore allow only a differential diagnosis to be made (134,163). The information yielded by radiography includes (Fig. 1-2):

- Topography of the lesion (location in the skeleton and in the individual bone)
- Borders of the lesion (the so-called zone of transition)
- Type of bone destruction
- Type of periosteal response to the lesion (periosteal reaction)
- Type of matrix of the lesion (composition of the tumor tissue)
- Nature and extent of soft tissue involvement

Patient age and determination of whether a lesion is solitary or multiple are the starting approaches in the diagnosis of bone tumors (Fig. 1-3). The age of the patient is the single most important item of clinical data that can be used in conjunction with the radiographic findings to establish a diagnosis (99). Certain tumors occur almost exclusively in a specific age group (Fig. 1-4). For example, aneurysmal bone cyst, chondromyxoid fibroma, and chondroblastoma rarely occur in individuals older than 20 years. Conversely, giant cell tumor of bone almost invariably arises after growth plate closure, and metastatic lesions, myeloma, and conventional chondrosarcoma are rarely encountered in patients younger than 40 years



Figure 1-2 Radiographic features of tumors and tumorlike lesions of bone.

(105,108). Some bone tumors associated with specific age groups may have different radiographic presentations and appear in atypical locations, when they arise outside of their usual age population. Simple bone cysts, for example, are found almost exclusively in the long bones (proximal humerus, proximal femur) before skeletal maturity. After skeletal maturity they may arise in the pelvis, scapula, or calcaneus, among other sites, and may exhibit unconventional radiographic features with progressing age (141).

Determining the number of lesions also has important implications. Benign lesions tend to involve multi-



Figure 1-3 Analytic approach to evaluation of the bone neoplasm. Pragmatic analysis must include patient age, multiplicity of a lesion, location in the skeleton and in the particular bone, and radiographic morphology.



Figure 1-4 Peak age incidence of some benign and malignant tumors and tumor-like lesions.

ple sites, as in polyostotic fibrous dysplasia, enchondromatosis, multiple osteocartilaginous exostoses, Langerhans cell histiocytosis (eosinophilic granuloma), hemangiomatosis, and fibromatosis. In contrast, primary malignancies, such as osteosarcoma, Ewing sarcoma, fibrosarcoma, and malignant fibrous histiocytoma, rarely present as multifocal disease. Multiple malignant lesions usually indicate metastatic disease, multiple myeloma, or lymphoma.

Site of the Lesion

Some tumors have a predilection for specific bones or specific sites in bone (Table 1-1 and Fig. 1-5). This loca-

tion is determined by the laws of field behavior and developmental anatomy of the affected bone, a concept first popularized by Johnson (84,133). Parosteal osteosarcoma, for example, arises with very rare exceptions in the posterior aspect of the distal femur, a feature so characteristic that it alone can suggest the diagnosis (Fig. 1-6). The same can be said of chondroblastoma, which has a strong preference for the epiphysis of long bones before skeletal maturity (Fig. 1-7). Adamantinoma and osteofibrous dysplasia have a specific predilection for the tibia (Fig. 1-8; see also Figs. 4-49, 4-50, and 7-64). A lesion's location can also exclude certain entities from the differential diagnosis.

	A. Skeletal Predilection of Benign Osseous Neoplasms and Tumor-like Lesions	B. Skeletal Predilection of Malignant Osseous Neoplasms	
Axial skeleton Appendicular skeleton	 Skull and facial bones: Osteoma, osteoblastoma, Langerhans cell histiocytosis, fibrous dysplasia, solitary hemangioma, osteoporosis circumscripta (lytic phase of Paget disease) Jaw: Giant cell reparative granuloma, myxoma, ossifying fibroma, desmoplastic fibroma Spine: Aneurysmal bone cyst, osteoblastoma, Langerhans cell histiocytosis, hemangioma Long tubular bones: Osteoid osteoma, simple bone cyst, aneurysmal bone cyst, osteochondroma, 	Skull and facial bones: Mesenchy- mal chondrosarcoma, chor- doma, multiple myeloma, metastatic neuroblastoma, metastatic carcinoma Mandible: Osteosarcoma, myeloma Spine: Chordoma, myeloma, metastases Long tubular bones: Osteosarcoma (all variants), adamantinoma,	
	enchondroma, periosteal chondroma, chondroblastoma, chondromyxoid fibroma, nonossifying fibroma, giant cell tumor, osteofibrous dysplasia, desmoplastic fibroma, intraosseous ganglion Hands and feet: Giant cell reparative granuloma, florid reactive periostitis, enchondroma, glomus tumor, epidermoid cyst, subungual exostosis, bizarre parosteal osteochondromatous lesion	malignant fibrous histiocytoma, fibrosarcoma, primary lym- phoma, chondrosarcoma, angiosarcoma Hands and feet: None	
Specific predilections	Simple bone cyst—proximal humerus, proximal femur Osteofibrous dysplasia—tibia, fibula (anterior cortex) Focal fibrocartilaginous dysplasia of long bones—proximal metaphysis of tibia Osteoid osteoma—femur, tibia Chondromyxoid fibroma—tibia, metaphyses Chondroblastoma—epiphyses Giant cell tumor—articular ends of femur, tibia, radius Liposclerosing myxofibrous tumor— intertrochanteric region of femur	Adamantinoma—tibia, fibula Parosteal osteosarcoma—distal fe- mur (posterior cortex) Periosteal osteosarcoma—tibia Chondrosarcoma (conventional)— femur, humerus, pelvic bones Clear cell chondrosarcoma— proximal femur and humerus Chordoma—sacrum, clivus, C2 Multiple myeloma—pelvis, spine, skull	

Table 1-1 Predilection of Tumors for Specific Sites in the Skeleton

Modified from Fechner RE, Mills SE. Tumors of the bones and joints, 3rd ed., Vol. 8. Washington, DC: Armed Forces Institute of Pathology, 1993:1–16.

Giant cell tumor, for example, should not be seriously considered if the lesion does not affect the articular end of a bone because very few of these tumors are found anywhere else.

Equally important in the evaluation of a lesion's site is its location in relation to the central axis of the bone (Fig. 1-9). This is particularly true when the lesion is located in a long tubular bone, such as humerus, radius, femur, or tibia (71). For example, simple bone cyst, enchondroma, or a focus of fibrous dysplasia always appears centrally located (see Figs. 3-6B, 3-8, 3-9, 4-25, 7-17, and 7-18). In contrast, eccentric location is characteristically observed in aneurysmal bone cyst, chondromyxoid fibroma, and nonossifying fibroma (see Figs. 3-74, 3-75, 4-2B, 4-3, 4-4, 7-31, and 7-33).

Borders of the Lesion

The borders or margins of a lesion are crucial factors in determining the growth rate of a lesion and hence whether it is benign or malignant (113). Three types of lesion margins are encountered: (a) a margin with sharp demarcation by sclerosis between the peripheral aspect of the tumor and the adjacent host bone (IA margin), (b) a margin with sharp demarcation without sclerosis around the periphery of the lesion (IB margin), and (c) a margin with an ill-defined region (either the entire circumference or only a portion of it) at the interface between lesion and host bone (IC margin) (Fig. 1-10). Slow-growing lesions are marked by sharply outlined, sclerotic borders, and this narrow zone of transition usually indicates that a tumor is benign (133). Benign lesions such as nonossifying fibroma, simple bone cyst, and chondromyxoid fibroma almost invariably exhibit sclerosis at their borders (Fig. 1-11A). Indistinct borders (a wide zone of transition), on the other hand, are typical of malignant or aggressive lesions (Fig. 1-11B). Primary malignancies such as fibrosarcoma, malignant fibrous histiocytoma, lymphoma, multiple myeloma (or solitary plasmacytoma), and metastases from primary tumors of the kidney, thyroid, and gastrointestinal tract usually lack a sclerotic border, as does the giant cell tumor of bone. Radio- or chemotherapy of malignant bone tumors can alter their appearance, causing them to exhibit sclerosis and a narrow zone of transition (54).



Figure 1-5 Site of the lesion. A: Distribution of various lesions in a long tubular bone in a growing skeleton (growth plate is open). **B:** Distribution of various lesions in a long tubular bone after skeletal maturity (growth plate is closed). **C:** Distribution of various lesions in a vertebra. Malignant lesions are seen predominantly in its anterior part (body), whereas benign lesions predominate in its posterior elements.

The more well defined the margin (e.g., IA, IB), the slower the biologic activity, and therefore the more likely that the lesion is benign. Conversely, the less well defined the margin (IC), the greater the biologic activity, and therefore the more likely that the lesion is malignant (79,107).

Type of Bone Destruction

Destruction of bone represents not only a direct effect of tumor cells but also reflects a complex mechanism in which normal osteoclasts of the host bone respond to pressure generated by the enlarging mass and by active hyperemia associated with the tumor (134). Cortical bone is destroyed less rapidly than trabecular bone. However, loss of cortical bone appears earlier on radiography because its density is highly homogeneous compared with that of trabecular bone. In the latter, greater amounts of bone must be destroyed (about 70% loss of mineral content) before the loss becomes radiographically evident (134). Like the borders of a lesion, the type of bone destruction caused by a tumor indicates its growth rate. Bone destruction can be described as geographic (type I), moth-eaten (type II), and permeative (type III) (105,107) (Fig. 1-12). Although none of these features are pathognomonic for any specific neoplasm, the type of destruction may suggest a benign or a malignant process. Geographic bone destruction is characterized by a uniformly destroyed area usually within sharply defined borders. It typifies slow-growing, benign lesions, such as simple bone cyst, enchondroma, chondromyxoid fibroma, or giant cell tumor. On the other hand, moth-eaten (i.e., characterized by multiple, small,



Figure 1-6 Site of the lesion. Parosteal osteosarcoma has a predilection for the posterior aspect of the distal femur.



Figure 1-8 Site of the lesion. The tibia is a common site for adamantinoma.



Figure 1-7 Site of the lesion. Chondroblastoma has a predilection for the epiphysis of a long bone.



Figure 1-9 Site of the lesion. A: Location of the epicenter of the lesion usually determines the site of its origin, whether medullary, cortical, periosteal, soft tissue, or in the joint. **B:** Eccentric versus central location of the similar-appearing lesions is helpful in differential diagnosis. **C:** Cortical and parosteal (periosteal) lesions.



Figure 1-10 Borders of the lesion. Borders of the lesion determine its growth rate. *1A,* sharp sclerotic; *1B,* sharp lytic; *1C,* ill-defined. (Modified from Madewell JE, Ragsdale BD, Sweet DE. Radiologic and pathologic analysis of solitary bone lesions. Part I. Internal margins. *Radiol Clin North Am* 1981;19:715–748.)



Figure 1-11 Borders of the lesion. A: Sclerotic border or narrow zone of transition from normal to abnormal bone typifies a benign lesion, as in this example of nonossifying fibroma in the distal femur. **B:** A wide zone of transition typifies an aggressive or malignant lesion, in this case a plasmacytoma involving the pubic bone and supraacetabular portion of the right ilium. (Reprinted with permission from Greenspan A. *Orthopedic imaging*, 4th ed. Philadelphia: Lippincott Williams & Wilkins, 2004.)











Figure 1-12 Patterns of bone destruction. A: Three types of bone destruction determine the lesion's growth rate. (Modified from Madewell JE, Ragsdale BD, Sweet DE. Radiologic and pathologic analysis of solitary bone lesions. Part I. Internal margins. *Radiol Clin North Am* 1981; 19:715–748.) **B:** The geographic type of bone destruction is characterized by a uniformly affected area within sharply defined borders, in this case a giant cell tumor. **C:** The moth-eaten bone destruction is characteristic of rapidly growing infiltrating lesions, in this case myeloma. **D:** The permeative type of bone destruction is characteristic of round cell tumors, in this case Ewing sarcoma. Note the almost imperceptible destruction of the metaphysis of the femur by a tumor that has infiltrated the medullary cavity and cortex and extended into the surrounding soft tissues, forming a large mass.

D



Figure 1-13 Types of periosteal reaction. An uninterrupted periosteal reaction usually indicates a benign process, whereas an interrupted reaction indicates a malignant or aggressive nonmalignant process.

often clustered lytic areas) and permeative (i.e., characterized by ill-defined, very small oval radiolucencies or lucent streaks) types of bone destruction mark rapidly growing, infiltrating tumors, such as myeloma, lymphoma, fibrosarcoma, or Ewing sarcoma. However, some nonneoplastic lesions may demonstrate this aggressive pattern. For example, osteomyelitis can exhibit both type II (moth-eaten) and type III (permeative) patterns of destruction (133). Similarly, hyperparathyroidism can cause a permeative pattern (113). The distinction between a moth-eaten and a permeative pattern of destruction may be subtle; often the two patterns coexist in the same lesion.

Periosteal Response

Like the pattern of bone destruction, the pattern of periosteal reaction is an indicator of the biologic activity of a lesion (209). Bone neoplasms elicit periosteal reactions that can be categorized as uninterrupted (continuous) or interrupted (discontinuous) (160) (Fig. 1-13 and Table 1-2). Any widening and irregularity of bone contour may represent periosteal activity. The solid periosteal reaction represents a single solid layer or multiple closely apposed and fused layers of new bone attached to the outer surface of the cortex (160) (Fig. 1-14). The resulting pattern is often referred to as cortical thickening (49). Although no single periosteal response is unique for a given lesion, an uninterrupted periosteal reaction indicates a long-standing (slowgrowing), usually indolent, benign process. There are several types of solid periosteal reaction: a solid buttress, such as is frequently seen accompanying aneurysmal bone cyst and chondromyxoid fibroma; a solid smooth or elliptical layer, such as is seen in osteoid osteoma and osteoblastoma; an undulating type, most frequently seen in long-standing varicosities, pulmonary osteoarthropathy, chronic lymphedema, periostitis, and, rarely, with neoplasms (160); and a single lamellar reaction, such as accompanies osteomyelitis,

 Table 1-2 Examples of Nonneoplastic and Neoplastic Processes Categorized by Type of Periosteal Reaction

Uninterrupted Periosteal Reaction					
Benign tumors and tumor-like lesions	Nonneoplastic conditions				
Osteoid osteoma	Osteomvelitis				
Osteoblastoma	Langerhans cell histiocytosis				
Anoury small hone syst	Healing fracture				
Alleurysmai bolle cyst					
Chondromyxoid fibroma	Juxtacortical myositis ossificans				
Periosteal chondroma	Hypertrophic pulmonary osteoarthropathy				
Chondroblastoma	Hemophilia (subperiosteal bleeding)				
	Varicose veins and peripheral vascular insufficiency				
Malianant tumors	Caffey disease				
Chandra constants (name)	Thumaid a service show				
Chondrosarcoma (rare)					
	Ireated scurvy				
	Pachydermoperiostosis				
	Gaucher disease				
Inte	errupted Periosteal Reaction				
Malianant tumors	Nonneoplastic conditions				
Osteosarcoma	Osteomyelitis (occasionally)				
Ewing sarcoma	Langerbans cell histiocytosis (occasionally)				
Chondrosarcoma	Subperiosteal hemorrhage (occasionally)				
Lymphoma (rare)					
Fibrosarcoma (rare)					
MFH (rare)					
Metastatic carcinoma					
Wetastatic Carcinoma					

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Figure 1-14 Solid type of periosteal reaction. A: An uninterrupted solid periosteal reaction is characteristic of benign lesions, in this case a cortical osteoid osteoma. B: An uninterrupted periosteal reaction typifies changes of hypertrophic pulmonary osteoarthropathy as seen here in the distal radius and ulna and bones of the hand in a patient with carcinoma of the lung.

Langerhans cell histiocytosis, and stress fracture. An interrupted periosteal response, on the other hand, is commonly seen in malignant primary tumors and less commonly in some metastatic lesions and highly aggressive nonmalignant processes. In these tumors, the periosteal reaction may appear in a sunburst ("hair-onend") (Fig. 1-15A) or onion-skin (lamellated) pattern (Fig. 1-15B,C). When the tumor breaks through the cortex and destroys the newly formed lamellated bone, the remnants of the latter on both ends of the breakthrough area may remain as a triangular structure known as a Codman triangle (Fig. 1-15D,E).

Type of Matrix

The matrix represents the intercellular material produced by mesenchymal cells and includes osteoid, bone, chondroid, myxoid, and collagen material (196). Assessment of the type of matrix allows differentiation of some similar-appearing lesions and, in particular, the tumor matrix provides a useful means of differentiating osteoblastic from chondroblastic processes. Although it should be kept in mind that tumor bone is often radiographically indistinguishable from reparative new bone deposited secondary to bone destruction by a reactive sclerosis or callus formation, the presence of irregular, not fully mineralized bone matrix within or adjacent to an area of bone destruction strongly suggests osteosarcoma (Fig. 1-16). Similarly, cottony or cloudlike densities within the medullary cavity and in the adjacent soft tissues are likely to represent tumor bone and hence osteosarcoma.

Calcifications in the tumor matrix, on the other hand, point to a chondroblastic process. These calcifications typically appear punctate (stippled), irregularly shaped (flocculent), or curvilinear (annular or comma-shaped, rings and arcs). In benign or welldifferentiated malignant tumors they may reflect the process of endochondral ossification (196) (Fig. 1-17A). Differential diagnosis of stippled, flocculent, or ring-and-arc calcifications includes enchondroma (Fig. 1-17B), chondroblastoma, and chondrosarcoma (Fig. 1-17C). Sometimes it is difficult to distinguish osseous matrix from cartilaginous matrix. The former usually appears radiographically more organized and structural (trabecular), whereas the latter is more amorphous, with frequently distinguished characteristic calcifications (described previously).

A completely radiolucent lesion may be either fibrous or cartilaginous in origin, although hollow structures produced by tumor-like lesions, such as simple bone cysts or intraosseous ganglion, can also present as radiolucent areas (Table 1-3).

Text continues on page 17



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Figure 1-15 Interrupted type of periosteal re-action. A: Highly aggressive and malignant le-sions may present radiographically with a sun-burst pattern of periosteal reaction, as seen in this case of osteosarcoma. **B:** Another pattern of interrupted periosteal reaction is the lamellated or oping skin type (grow) as seen here in Ewing or onion-skin type (*arrow*) as seen here in Ewing sarcoma involving the proximal femur. **C**: Radiographs of the slab sections (coronal at left and transverse at right) of the resected specimen from Ewing sarcoma demonstrate the lamellated type in more detail (continued).

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D

Figure 1-15 *Continued* **D**: Codman triangle (*arrow*) also reflects an aggressive, usually malignant type of periosteal response. **E**: Schematic representation of periosteal reactive bone formation. (a) Tumor (*red dashes*) growing against the endosteal border of the cortex blocks vascular supply and provokes reactive hyperemia of periosteum with secondary formation of a single layer of bone. (b) With progressive destruction of cortical bone, additional layers of periosteal bone form lamellated ("onion-peel") appearance. (c) After the rapidly growing tumor has destroyed cortex and newly formed periosteal bone, remnants of the latter at the tumor border form a Codman triangle. (d) Slowly growing tumor leaves time to form perpendicular reactive periosteal bone (spiculae, sunburst appearance).

Е

Table 1-3Tumors and Pseudotumors That May Present asRadiolucent Lesions

A. Solid	B. Cystic
Osteoblastic (osteoid osteoma, osteoblastoma, osteosarcoma, low- grade central osteosarcoma, telangiectatic osteosarcoma) Cartilaginous (enchondroma, chondroblastoma, chondromyxoid fi- broma, chondrosarcoma) Fibrous and histiocytic (fibrous cortical defect, nonossifying fibroma, fibrous dysplasia, osteofibrous dysplasia, desmoplastic fibroma, fibrosarcoma, malignant fibrous histiocytoma) Intraosseous lipoma Lymphoma Myeloma (plasmacytoma) Ewing sarcoma Metastatic (from lung, breast, gastrointestinal tract, kidney, thyroid) Giant cell tumor Giant cell reparative granuloma Langerhans cell histiocytosis Paget disease (osteolytic phase— osteoporosis circumscripta)	Simple bone cyst Aneurystmal bone cyst Various bone cysts (synovial, degenerative) Hydatid cyst Brown tumor of hyperparathyroidism Vascular lesions Hemophilic pseudotumor Intraosseous ganglion Bone abscess Cystic tuberculosis



Figure 1-16 Types of matrix: osteoblastic. The matrix of a typical osteoblastic lesion is characterized by the presence of either fluffy, cotton-like densities within the medullary cavity, such as in this case of osteosarcoma of the distal femur **(A)**, by the presence of the wisps of tumor-bone formation, like in this case of osteosarcoma of the sacrum **(B)**, or by the presence of a solid sclerotic mass, such as in parosteal osteosarcoma **(C)**.

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Figure 1-17 Types of matrix: chondroid. A: Schematic representation of various appearances of chondroid matrix calcifications. (Modified from Sweet DE, Madewell JE, Ragsdale BD. Radiologic and pathologic analysis of solitary bone lesions. Part III. Matrix patterns. *Radiol Clin North Am* 1981;19:785–814.) **B:** Enchondroma displays a typical chondroid matrix. **C:** Chondrosarcoma with characteristic chondroid matrix.



Figure 1-18 Soft tissue mass. Radiographic features of soft tissue extension characterizing malignant or aggressive bone lesions and benign processes.

Soft Tissue Mass

With few exceptions—giant cell tumor, aneurysmal bone cyst, and desmoplastic fibroma among the more common examples—benign bone tumors usually do not have an associated soft tissue mass, which is an invariable feature of many advanced malignant and aggressive lesions (94) (Fig. 1-18). Nevertheless, it is important to note that some nonneoplastic conditions also manifest with a soft tissue component (e.g., osteomyelitis). In such cases, however, the associated mass is usually poorly defined and the fatty tissue layers appear obliterated. This is in sharp contrast to the soft tissue extensions typical of malignant processes, in which a defined mass are usually intact.

A bone lesion associated with a soft tissue mass should prompt the question of which came first. Is the soft tissue lesion an extension of a primary bone tumor, or is it a primary soft tissue tumor invading bone? Some clues may help to answer this question, but these observations are by no means absolute (Fig. 1-19). A large soft tissue mass with a smaller bone lesion usually indicates secondary bone involvement. That said, it is worth noting that Ewing sarcoma and less commonly other malignant tumors may exhibit a large soft tissue mass accompanying a small primary bone malignancy. Another clue to help determine the primary malignancy may be found in the periosteal response. Primary soft tissue tumors adjacent to bone usually destroy the neighboring periosteum without eliciting a periosteal response. Primary bone malignancies, however, typically prompt a periosteal



Figure 1-19 Differential diagnosis of soft tissue mass. Certain radiographic features of bone and soft tissue lesions may help differentiate a primary soft tissue tumor invading the bone from a primary bone tumor invading soft tissues. (From Greenspan A. Orthopedic imaging, 4th ed. Philadelphia: Lippincott Williams & Wilkins, 2004.)



Figure 1-20 Radiographic features that may help differentiate benign from malignant lesions.

reaction when they grow into the cortex and extend into adjacent soft tissues (69,133,160).

Benign Versus Malignant Nature

Although it is sometimes very difficult to establish a lesion as benign or malignant by radiography alone, the clusters of features that can be gathered from radiographs can help in favoring one designation over the other (Fig. 1-20). Benign lesions usually have well-defined sclerotic borders and exhibit a geographic type of bone destruction; the periosteal reaction is solid and uninterrupted, and there is no soft tissue mass. In contrast, malignant tumors often exhibit poorly defined borders with a wide zone of transition; bone destruction appears in a moth-eaten or permeative pattern, and the periosteum shows an interrupted, sunburst, or onion-skin reaction with an adjacent soft tissue mass. It should be kept in mind, however, that some benign lesions may also exhibit aggressive features (Table 1-4).

Scintigraphy (Radionuclide Bone Scan)

The radionuclide bone scan is an indicator of mineral turnover. Because there is usually enhanced deposition of bone-seeking radiopharmaceutical agents in areas undergoing change and repair, a bone scan is useful in localizing tumors and tumor-like lesions in the skeleton (58). This is particularly true in such conditions as fibrous dysplasia, Langerhans cell histiocytosis, and metastatic cancer, in which more than one lesion is encountered. Technetium-99m methyl diphosphonate (MDP) scans are used primarily to determine whether a lesion is monostotic or polyostotic. Such a study is therefore essential in staging a bone neoplasm. It is important to remember that although the degree of abnormal uptake may be related to the aggressiveness of the lesion, this does not correlate well with the histologic grade (Fig. 1-21).

Radionuclide bone scan also plays an important role in localizing small lesions such as osteoid osteoma, which may not always be visible on radiography (Fig. 1-22). Although skeletal scintigraphy is a highly sensitive method for detection of bone neoplasms, its specificity is low. In most instances it cannot distinguish benign lesions from malignant tumors, because increased blood flow with increased isotope deposition and increased osteoblastic activity take place in both benign and malignant conditions. Nevertheless, it can sometimes achieve such differentiation in benign lesions that do not absorb the radioactive isotope. In addition, the radionu-

Lesion	Radiographic Presentation	Lesion	Radiographic Presentation
Osteoblastoma (aggressive)	Bone destruction and soft tissue extension similar to osteosarcoma	Osteomyelitis	Bone destruction, aggressive periosteal reaction. Occasionally, features resembling osteosarcoma, Ewing sarcoma, or lymphoma
Desmoplastic fibroma	Expansive, destructive lesion, frequently trabeculated, mimics fibrosarcoma and chondrosarcoma	Langerhans cell histiocytosis	Bone destruction, aggressive periosteal reaction. Occasionally, features resembling Ewing sarcoma
Periosteal desmoid	Irregular cortical outline, mimics osteosarcoma or Ewing sarcoma	Pseudotumor of hemophilia	Bone destruction, periosteal reaction, occasionally mimics malignant tumor
Giant cell tumor	Occasionally, aggressive features such as osteolytic bone destruction, cortical penetration, and soft tissue extension	Myositis ossificans	Features of parosteal or periosteal osteosarcoma, soft tissue osteosarcoma, or liposarcoma
Aneurysmal bone cyst	Soft tissue extension, occasionally mimics malignant tumor such as telangiectatic osteosarcoma	Brown tumor of hyperparathyroidism	Lytic bone lesion, resembling malignant tumor

Table 1-4 Benign Lesions with Aggressive Features



Figure 1-21 Scintigraphic activity. A: Radionuclide bone scan shows markedly increased uptake of radiotracer in the proximal humerus in a 26-year-old woman suspected of developing a malignant tumor. **B:** Anteroposterior radiograph shows, however, a benign-appearing lesion with sclerotic border (*arrows*), consistent on excision biopsy with healing benign fibrous histiocytoma. (Reprinted with permission from Greenspan A. *Orthopedic imaging*, 4th ed. Philadelphia: Lippincott Williams & Wilkins, 2004.)

clide bone scan is sometimes useful for differentiating multiple myeloma, which usually shows no significant uptake of the tracer, from metastatic bone cancer, which usually does. Scans using Ga-67 may show uptake in a soft tissue sarcoma and may help to differentiate a sarcoma from a benign soft tissue lesion.

Imaging with thallium-201 chloride has recently been tried in the diagnosis of bone and soft tissue lesions (26,44,112,171,190,206). Although the role of this technique for staging of bone tumors and for differentiation of benign from malignant lesions is limited, thallium-201 scintigraphy appeared useful in detection of primary and metastatic neoplasms, such as synovial sarcoma (112) and some of the cartilaginous tumors (29). Furthermore, the combined use of TI-201 and pentavalent dimercaptosuccinic acid (DMSAV) functional nuclear scanning proved to be promising in the differential diagnosis of benign and malignant cartilage lesions and their grading (29).

The essential factors in the evaluation of a bone lesion are intra- and extraosseous extension, and presence (or absence) of metastases. Although radiography can yield significant information about tumor extension, skeletal scintigraphy is an indispensable technique for evaluating whether a tumor has spread beyond its site of origin (12,15,28,62,152,194). Radionuclide bone imaging better demonstrates the extent of intramedullary involvement by tumor than radiography, but it tends to show a larger than actual area of extension because the radiopharmaceutical also localizes to areas of hyperemia and edema adjacent to the tumor (15). Therefore, bone scans are not adequate for evaluating the exact level of intramedullary invasion. However, scintigraphy has no competitor (except for MRI) for identifying additional remote skeletal lesions, so-called skip lesions, and intraosseous metastases (55).

Scintigraphy and MRI or CT are truly complementary examinations that provide quite different staging information for the patient who presents with a bone tumor that requires biopsy (167–169). The main role of scintigraphy is not so much for local staging as for evaluating the remainder of the skeleton. Although scintigraphy provides information on the intraosseous extent of disease, it cannot entirely be relied on for



Figure 1-22 Effectiveness of skeletal scintigraphy. A: Anteroposterior radiograph of the left hip of a 16-year-old boy with a typical history of osteoid osteoma is equivocal, although there is the suggestion of abnormal radiolucency in the supraacetabular portion of the ilium. **B:** Radionuclide bone scan shows an increased uptake of radiopharmaceutical agent (*arrow*) corresponding to questionable radiolucency seen on radiography. Osteoid osteoma was diagnosed on excision biopsy. (Reprinted with permission from Greenspan A. *Orthopedic imaging*, 4th ed. Philadelphia: Lippincott Williams & Wilkins, 2004.)

local extent of the lesion [because of the augmented uptake that has been described in osteosarcomas (28,79)], its inability to match MRI in differentiating normal from abnormal marrow, and its inability to adequately demonstrate extracompartmental disease. Increased uptake of radionuclides is highly nonspecific. Any pathologic process in bone that leads to new bone formation (reactive or tumor bone), increased blood flow, or bone turnover, will show increased radionuclide uptake (58). Therefore, a bone scan is usually not reliable for identifying the specific type of tumor or for differentiating malignant from benign processes. Despite these factors, scintigraphy is still an important technique for evaluation of solitary bone tumors. Because scintigraphy is the most sensitive examination for imaging the entire skeleton, it should always be performed to determine whether skeletal involvement is solitary or multiple. Metastases are the most common malignant tumors of bone and can frequently present as a solitary abnormality. In the pediatric age group, as in adults, metastases may mimic solitary tumors of bone. Metastatic neuroblastoma and leukemia may resemble true solitary lesions such as Ewing sarcoma, Langerhans cell histiocytosis, and acute osteomyelitis. Because synchronous and delayed skeletal metastases may occur, scintigraphy is recommended at initial presentation and in follow-up of patients after extirpation of primary Ewing tumor (62). Scintigraphy can suggest the presence or absence of disseminated skeletal disease. In the proper clinical setting its appearance can be quite specific for the diagnosis of osteoid osteoma (172) (see Fig. 2-35).

Positron Emission Tomography

PET is a diagnostic imaging technique that allows identification of biochemical and physiologic alterations in the body and assesses the level of metabolic activity and perfusion in various organ systems. The process produces biologic images based on detection of gamma rays that are emitted by a radioactive substance, such as ¹⁸FDG. One of the main applications of this technique is in oncology, including detection of primary and metastatic tumors and recurrences of the tumors after treatment (Fig. 1-23). In fact, PET has been found to be more sensitive than CT and MRI in this respect. Although some promising results have been reported using this technique (8), detection of bone marrow involvement is still controversial, because physiologic bone marrow uptake can be observed on ¹⁸FDG PET images. Moreover, although highly sensitive, PET scanning has relatively low specificity because FDG may also accumulate in benign aggressive and inflammatory lesions.

Recently, Zhang et al. conducted investigations of the utility of ¹¹C-methionine (MET) PET in the imaging of chordoma (215). Their study has demonstrated that MET PET is feasible for imaging of chordoma, showing a sensitivity of 80% in the visualization of all tumors, and 100% for the recurrent lesions.

Computed Tomography and Magnetic Resonance Imaging

Both CT and MRI are highly accurate for determining the presence of neoplasm within the cortex, trabecular bone, or marrow cavity, as well as soft tissue invasion by a tumor (98,150,188,193,199,212,217). Although CT by itself is rarely helpful in making a specific diagnosis, it can provide a precise evaluation of the extent of a bone lesion and may demonstrate breakthrough of the cortex and involvement of surrounding soft tissues (Fig. 1-24). Moreover, CT is very helpful in delineating a bone tumor in a complex anatomic structure such as scapula, pelvis, or sacrum (Fig. 1-25). At times, the three-dimensional reformation of CT images is used to better and more comprehensively demonstrate the tumors (25,53, 97). This technique can be useful in depicting the surface lesions of bone, such as osteochondroma (Fig. 1-26; see also Fig. 3-50), periosteal chondroma, parosteal osteosarcoma, or juxtacortical chondrosarcoma. The use of CT versus MRI is based on the radiographic findings: if there is no definite evidence of extension of osseous tumor into the soft tissues, then CT is preferred for detecting subtle cortical invasion and periosteal reaction, while providing an accurate means of determining the intraosseous extension of the neoplasm. If, however, the radiographs suggest cortical destruction and soft tissue mass, then MRI would be the more desirable modality because it provides an excellent soft tissue contrast and can determine the extraosseous extension of the tumor much better than CT. In addition, MRI provides valuable information regarding the in-



Figure 1-23 Positron emission tomography (PET) scan. ¹⁸FDG whole body PET scan of a 37-year-old woman with known fibrous dysplasia shows multiple skeletal deformities of the long bones. Red areas represent large hypermetabolic foci. (Courtesy of Drs. Frieda Feldman and Ronald van Heertum, New York, New York.)

traosseous extent of neoplasms based on its multiplanar imaging ability (sagittal, coronal, axial, and oblique) and lack of beam-hardening artifacts from cortical bone. Because of superior ability to characterize soft tissue masses, MRI has distinct advantages over CT (11,31,216) (Fig. 1-27).

A number of pulse sequences can be used to evaluate musculoskeletal tumors (66,154). These include spinecho (SE) sequences, inversion recovery (IR) sequences, short time inversion-recovery (STIR) sequences, gradient echo (GRE) sequences, and fast T2- or fat-suppressed T2-weighted sequences (11,43,65,158,183). SE sequences are the most effective for identification and staging of most skeletal tumors (10,216). The signal intensity for normal tissues is predictable using these sequences. On T1-weighted images, fat and bone marrow have a high signal intensity that changes to intermediate intensity on T2 weighting. Muscles have intermediate signal intensity on both T1 and T2 sequences. Cortical bone and fibrocartilage have low signal on both sequences. Fluid has intermediate signal on T1-weighted and high signal intensity on T2-weighted images (Table 1-5). Most bone tumors have increased T1 and T2 relaxation times compared with normal tissue and therefore are imaged as areas of low or intermediate signal intensity on T1-weighted SE sequences and high signal intensity on T2-weighted sequences (11,63,216). T1-weighted SE sequences enhance tumor contrast with bone, bone marrow, and fatty tissue, whereas T2-weighted SE or T2weighted gradient echo images enhance tumor contrast with muscle and accentuate peritumoral edema (73,132,151,191,195,207) (Fig. 1-28).

Several investigators have stressed the advantage of contrast enhancement of MR images using intravenous injection of gadopentate dimeglumine [gadolinium diethylenetriaminepentaacetic acid (Gd-DTPA)] (74,178). This paramagnetic contrast enhancement on conventional T1-weighted sequences decreases the T1 relaxation time in tumor tissue causing the lesion to be of higher signal intensity (hence making the demarcation of the tumor more obvious) (Fig. 1-29). In particular, enhancement was found to give better delineation of tumor in richly vascularized parts, in compressed tissue immediately surrounding the tumor, and in atrophic but richly vascularized muscle (153). Both static and dynamic Gd-DTPA studies show significant promise for evaluation of musculoskeletal tumors (47,98,208). Areas that demonstrate contrast enhancement on T1 weighting are typically more vascular, whereas those without enhancement usually represent necrotic tissue (47). In addition, utilization of a chemical shift or fat suppression technique is even more effective (128) because the fat signal becomes markedly suppressed, whereas the abnormal contrast-enhanced tumor displays a high signal intensity (127,197,198). This combination improves the interface between tumor and peritumoral edema, the interface between tumor and reactive zone, and the interface between tumor and adjacent muscle (179,197,198).



Figure 1-24 Effectiveness of computed tomography (CT) scanning. A: Conventional radiograph shows destructive changes in the proximal diaphysis of the left fibula (*arrows*) of a 12-year-old boy. B: CT scan demonstrates involvement of the bone marrow of the fibula (*arrow*) and extension of the tumor (which proved to be a Ewing sarcoma) into the soft tissues (*arrowheads*).

Coronal-plane MR images, in particular, are often superior to axial CT scans in providing details of extraand intramedullary tumor extent and its relationship to surrounding structures (61). Combining axial and coronal plane imaging has also been helpful in assessing important vascular structures adjacent to tumors. In addition, MRI offers better visualization of tissue planes surrounding a lesion and can evaluate neurovascular involvement without the use of intravenous contrast (45). Particularly in tumors of the extremities, the ability of MRI to demarcate normal from abnormal tissue more sharply than CT can reliably delineate the spatial boundaries of tumor masses, encasement and displacement of major neurovascular bundles, and the extent of joint involvement (1,100).

In several respects, however, CT unquestionably rivals MRI. MR images do not clearly depict calcifications in the tumor matrix or allow it to be characterized as readily as CT. In fact, large amounts of calcification and ossification have occasionally gone almost undetected by MRI (195). In addition, MRI demonstration of cortical destruction and periosteal reaction is less satisfactory than that of CT or even of radiography (67,191). Although neither technique is usually suitable for establishing the precise nature of a bone tumor (except perhaps the characteristic MRI features of intraosseous lipoma and hemangioma), much faith has been placed in MRI in particular as a method capable of distinguishing benign from malignant lesions (16,47,76,93,109). An overlap between the classic characteristics of benign

and malignant tumors is often observed (110). Moreover, some malignant bone tumors can appear misleadingly benign on MR images and, conversely, some benign lesions may exhibit a misleadingly malignant appearance (36,110). Attempts to formulate precise criteria for correlating MRI findings with histologic diagnosis have been largely unsuccessful (195). Tissue characterization on the basis of MRI signal intensities is still unreliable. Because of the wide spectrum of bone tumor composition and their differing histologic patterns, as well as in tumors of similar histologic diagnosis, signal intensities of histologically different tumors may overlap or there may be variability of signal intensity in histologically similar tumors. MRI is sensitive for delineating the extent of disease but, unfortunately, is relatively nonspecific (164,165,191).

Pathology

The task of categorizing an osseous lesion as benign or malignant is even more complex for the pathologist than for the radiologist. In bone tumor pathology, about 70 entities must be differentiated, most of them representing a very low incidence. In fact, primary malignant bone tumors comprise only 1% of all malignancies, and about half of these represent multiple myeloma. Accordingly, the diagnostic density, even in a large institution, may not be sufficient to ensure diagnostic security. Therefore, an opinion from a bone



Figure 1-25 Effectiveness of computed tomography (CT) scanning. Standard radiographs (not shown here) were ambiguous in this 70-year-old man with a palpable mass over the right scapula. However, two CT sections show a destructive lesion of the glenoid portion and body of the scapula (*arrows*) (**A**), with a large soft tissue mass containing chondroid calcifications and extending to the rib cage (*arrowheads*) (**B**). The lesion proved to be a chondrosarcoma.

tumor reference center should be obtained in all doubtful cases. For the pathologist, it is mandatory to view at least the radiographs (and the cross-sectional studies of tumors affecting the spine, pelvis, and shoulder girdle) and to consider all the radiologic criteria (see previous discussion), together with the data supplied by the clinician. Ideally, the pathologist should also consult with both the radiologist and the orthopedic surgeon. Although it may be obvious to a clinician that a child with a rock-hard, tender, 15-cm swelling of the femur almost undoubtedly has a tumor, a 4-mm sampling of tissue from the same lesion, unaccompanied by a history and radiologic studies, is fraught with diagnostic risk (68). Without consideration of all available correlative measurements before an attempt is made to interpret histologic sections,

there is no assurance that a biopsy specimen is representative of the lesion or, in fact, that the actual abnormality has been sampled. This is particularly true for chondroblastic tumors, in which benign tissue may be dispersed within the malignant portion of a tumor, and vice versa.

Basic Techniques and Decalcification

After results of the necessary clinical correlation studies are considered, the histologic diagnosis is rendered. For technical details of the different staining procedures mentioned later, the reader is referred to standard textbooks of histochemistry and histotechnology (6,17,78,157). The tumor cells and their arrangement, as well as the intercellular matrix *Text continues on page 28*



В

Figure 1-26 Effectiveness of three-dimensional computed tomography (CT). A: Conventional CT section through the chest shows an osteochondroma at the site of the anterolateral portion of the right fourth rib (*arrow*). It is difficult to determine if the lesion is sessile or pedunculated. **B:** Three-dimensional CT in maximum intensity projection (MIP) allows one to characterize the internal architecture of the lesion and delivers a much more informative image of osteochondroma. Note typical chondroid matrix of the tumor. **C:** Three-dimensional CT in shaded surface display (SSD) renders better conspicuity of the lesion; the pedicle of osteochondroma (*arrow*) is now clearly demonstrated.

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Figure 1-27 Effectiveness of magnetic resonance imaging (MRI) versus computed tomography (CT). A: Anteroposterior radiograph of the lower leg of a 14-year-old boy with Ewing sarcoma of the distal fibular diaphysis shows moth-eaten destruction of the bone and aggressive periosteal reaction. Soft tissue mass is not well demonstrated. B: CT scan effectively demonstrates cortical destruction of the fibula and periosteal reaction, but the soft tissue mass is not well characterized. C: Axial T2-weighted fat-suppressed MRI clearly shows a large soft tissue mass adjacent to the osseous tumor.



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Figure 1-28 Magnetic resonance imaging (MRI): T1 versus T2 weighting. Sagittal **(A)** and axial **(B)** spin-echo T1-weighted MRIs of a 29-year-old woman with malignant fibrous histiocytoma in the left tibia show excellent demarcation of low-signal-intensity tumor from high-signal-intensity bone marrow. **C:** Axial spin-echo T2-weighted MRI shows to better advantage the demarcation between the surrounding soft tissues and a heterogeneous-high-signal-intensity tumor that is breaking through the cortex.

Table 1-	5 N	Magnetic	Resonance	Imaging	Signal	Intensities	of	Various	Tissues
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	Image	
Tissue	T1-weighted	T2-weighted
Hematoma, hemorrhage (acute, subacute) Hematoma, hemorrhage (chronic) Fat, fatty marrow Muscle, nerves, hyaline cartilage Cortical bone, tendons, ligaments, fibrocartilage, scar tissue, air Hyaline cartilage Red (hematopoietic) marrow Fluid Proteinaceous fluid Tumors (generally) Lipoma Hemangioma	High/intermediate Low High Intermediate Low Intermediate Low Intermediate High Intermediate-to-low High Intermediate (slightly higher than muscle)	High Low Intermediate Intermediate Low Intermediate High High High Intermediate High



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Figure 1-29 Magnetic resonance imaging (MRI): contrast enhancement. A: Coronal T1-weighted MRI of a 14-year-old boy with osteosarcoma in the distal left femur shows a low-signal-intensity tumor in the bone marrow associated with a soft tissue mass. **B:** After intravenous administration of gadolinium (Gd-DTPA) there is a heterogeneous enhancement of both intramedullary tumor and soft tissue mass.

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produced, are compared with the appearance of radiographs and other imaging studies to verify adequate and representative sampling. The histologic patterns are then compared and matched with similar or identical histologic patterns of known examples of bone tumors (200). For daily diagnostic requirements, standard hematoxylin and eosin (H&E) staining after decalcification is usually sufficient (Fig. 1-30). Only for some histochemical and immunocytochemical methods does previous decalcification represent an obstacle. Acidic decalcifiers such as formic acid (5%-10% solutions) are ordinarily used. These agents provide good results within a short period of time. Because formic acid is not as rapid a decalcifier as nitric acid, the tissue structure is well preserved and good staining for H&E is retained (185). Most immunohistochemical protocols can also be applied. However, genetic analyses are often seriously hampered. Decalcification by EDTA, a chelating agent, does not pose these problems but is much more time-consuming (181). However, its use in combination with ultrasound leads to a reduction in time (126). Recently developed automated techniques of EDTA decalcification combined with ultrasound allow much more rapid decalcification, at least for small samples such as core biopsies (38). An earlier method, introduced about 20 years ago, of embedding the bone specimen in plastic and cutting it without previous decalcification, which is necessary for histomorphometric purposes, does not substantially contribute to bone tumor diagnostics.

Special Stains and Enzyme Histochemistry

Among the special stains available is van Gieson stain, which is more commonly used in Europe. This stain helps to identify the presence and amount of collagen in bone and other connective tissues by staining it intensely red (Fig. 1-31). This method is particularly effective to identify osteoid, whose major component is type I collagen. Giemsa stain should be used in the differentiation of small, blue, round cell tumors, particularly the lymphomas (Fig. 1-32). It also intensely stains the arrest lines in the bone matrix, thus making apposition and reconstruction processes clearer in an H&E preparation. We have found it useful in the differentiation of bone and cartilage when both tissues are intermingled because of the metachromatic properties. The Alcian blue stain can be useful in a similar manner (Fig. 1-33). Reticulin fibers are usually stained with Gomori stain, which is helpful in the differentiation against collagen fibers (Fig. 1-34). We prefer the variation of this stain described by Novotny, in which the fibers stand out more clearly (Fig. 1-35). Periodic acid-Schiff (PAS) staining is effective in demonstration of glycogen in the cytoplasm (Fig. 1-36). Another helpful tool is enzyme histochemistry, in particular the demonstration of alkaline phosphatase as marker for osteoblasts (Fig. 1-37A) and acid phosphatase as marker for osteoclasts (Fig. 1-37B).



Figure 1-30 Hematoxylin and eosin (H&E) stain. Chondrosarcoma. The cortex of the femur (*bottom*) shows large resorption lacunae (*center and right*) with cartilaginous tumor tissue invading the bone. Some osteoclasts that created the excavations are still visible (*center*). The cartilage stains pink; the bone stains red (H&E, original magnification \times 100).

Immunohistochemistry

A more recent method is immunohistochemistry (IHC), which demonstrates by immunologic methods specific antigens that serve as markers on the cell surface and its inner structures (60,130,174). By choosing the appropriate detection methods, including antigen retrieval techniques, almost any available antibody can be used to label a cell or a tissue (34,41). Studies using IHC are extremely helpful in differentiating among tissues that have similar morphology or whose histologic origins are uncertain. IHC can identify the antigenic factors in cells that would not other



Figure 1-31 Van Gieson stain. Fibrous dysplasia. Note irregular bony trabeculae typical for this disorder. Collagenous fibers are stained intensely red; particularly Sharpey fibers are highlighted in perpendicular arrangement to the trabecular surface (van Gieson, original magnification \times 200).



Figure 1-32 Giemsa stain. Diffuse large B-cell lymphoma. DNA, RNA, and basophilic cytoplasmic components are stained purple to grey-blue, helping to identify nuclear details of lymphoma cells (Giemsa, original magnification \times 630).



Figure 1-34 Gomori stain. Diffuse large B-cell lymphoma. Reticulin fibers appear as fine dark-brown to black network encircling lymphoma cells in a bone marrow biopsy specimen (Gomori, original magnification \times 630).

wise be evident in histologic sections stained by routine methods. Nevertheless, although highly specific antibodies have been developed, as well as sensitive systems for immune and enzymatic detection, there remain problems with false-positive and false-negative results (60). IHC relies on the use of enzyme-linked antibodies for detection of tissue antigens. The enzyme antibody converts a colorless substrate into a stained product that is precipitated on the slide at the site of the reaction. Continuing development of techniques for IHC has led to the use of enzyme labeling with alkaline phosphatase and peroxidase. Labeling with colloidal gold is also used in IHC reactions for both light and electron microscopy. However, the avidin-biotin-complex (ABC) method has now become the most sensitive and widely used technique for immunohistochemical staining (60).

For diagnostic purposes, the application of IHC in bone tumor pathology is necessary only in a limited number of cases, mainly Ewing tumor family [Ewing sarcoma and primitive neuroectodermal tumor (PNET)], hematopoietic tumors and histiocyte disorders, vascular tumors, chordoma, adamantinoma of long bones and, of course, metastatic disease. The IHC reaction pattern that characterizes a lesion is discussed in the appropriate chapters.

It cannot be overemphasized that a positive immunoreaction is not a substitute for a diagnosis of a



Figure 1-33 Alcian blue–periodic acid–Schiff (PAS) stain. Extraskeletal myxoid chondrosarcoma. Alcian-positive pale blue mucin pools contain acidic glycoproteins. Basement membrane material of capillaries (*upper left*) stains pink with PAS (original magnification \times 200).



Figure 1-35 Novotny stain. Ewing sarcoma. Reticulin fibers that stain black are absent within tumor cell areas, thus excluding diagnosis of malignant lymphoma and small cell osteosarcoma (Novotny, original magnification \times 12).



Figure 1-36 Periodic acid–Schiff (PAS) stain. Ewing sarcoma. Glycogen droplets (*center*) and glycoprotein-containing basement membrane material of capillaries (*bottom left*) stain pink to red. Because glycogen is water-soluble, it may partly be removed by formalin fixation (PAS, original magnification \times 630).

specific bone tumor, which should be established only by taking into account the additional clinical, radiologic, and histologic findings. The most commonly used antibodies are briefly described in the following sections.

Antibodies Against Intermediate Filaments

Intermediate filaments (IFs) are structural proteins with a diameter of 8 to 10 nm (nanometers) and a molecular weight of 40 to 110 KD (kilodalton). Together with microtubuli (24 nm) and microfilaments (6 nm), the IFs form the cytoskeleton (116). IFs can be separated into five distinct types on the basis of sequence identify and tissue distribution (27). For diagnostic purposes, the most important members of the IF family are the cytokeratins (CKs), desmin, neurofilaments, glial fibrillary acidic protein (GFAP), and vimentin.

With the use of two-dimensional gel electrophoresis, 20 different CKs have been identified and numbered. They constitute two groups of basic (CK 1–8) and acidic (CK 9–20) CKs that form heterodimers (27,56,129). CKs are found predominantly in epithelial cells but have also been detected in other tissues. Therefore, a positive immunoreaction for CKs does not unequivocally prove the epithelial nature of a tumor under investigation. In bone tumors, CKs are typically expressed in adamantinomas of long bones, in chordoma (Fig. 1-38), in osteofibrous dysplasia, in epithelioid hemangioendothelioma, and in metastases of extraosseous cancer. However, "aberrant" expression can be found in almost all types of nonepithelial tumors, including osteosarcoma (142).

Vimentin is present in all mesenchymal and some epithelial cells. Its IHC detection in mesenchymal tumors is rarely of diagnostic value (34).

Desmin is present in all muscle tissues, stabilizing the contractile apparatus. It is found in both smooth muscle and skeletal muscle tumors and also in lesser amounts in myofibroblasts (175). Antibodies to desmin are used mainly for the differential diagnosis of spindle cell tumors (161).

Neurofilaments (NFs) are present in neural cells in three different forms: high molecular weight NF-H (200 KD), medium weight NF-M (160 KD), and low weight NF-L (68 KD). They can be detected in small amounts in tumors of the Ewing family, such as PNETs or classical Ewing sarcomas, and especially in neuroblastomas (149,201).

GFAP is present in glial cells and is almost never used in studies of bone tumor pathology (162).

Antibodies Against Hematopoietic and Lymphoid Cells

Antibodies against hematopoietic and lymphoid cells are widely used in the diagnosis of lymphomas and



Figure 1-37 Enzyme histochemistry. A: Alkaline phosphatase. A small round bone trabecula (*center*) is surrounded by stroma typical for fibrous dysplasia. All cells, in the stroma as well as the osteoblasts at the bone border and osteocytes within the bone, are more or less positive, indicating their bone-forming capability (alkaline phosphatase, original magnification \times 50). **B:** Acid phosphatase. In fibrous dysplasia, with reaction for acid phosphatase an osteoclast with four nuclei (*left center*) shows intense activity. The surrounding mononuclear cells show a weak, finely granulated activity. The bone trabecula (*bottom*) shows negative reaction to acid phosphatase marker (acid phosphatase, original magnification \times 500).



Figure 1-38 Immunohistochemistry. Chordoma. Tumor cells react positive with an antibody-cocktail (CK22) directed against low molecular weight and high molecular weight cytokeratins. Lymphocytes (*center*) and cartilage cells (*upper left*) react negative. Methylene-blue counterstaining (original magnification \times 400).

hematologic malignancies (82). Most antibodies against these largely cell surface–based antigens have been categorized in workshops and statistically assigned to a socalled cluster of differentiation (CD) number that designates the respective antigen (9,218). These antigens are present not only on lymphatic or hematopoietic (as in Langerhans cell histiocytosis) cells (Fig. 1-39), but also in Ewing sarcoma and synovial sarcoma (CD99) (Fig. 1-40), and in endothelial and vascular tumors (CD31 and CD34). Applied as a panel that must be compiled according to the suspected morphologic dif-



Figure 1-40 Immunohistochemistry. Ewing sarcoma. Tumor cells show a positive membrane-bound reaction for CD99. Nuclei are counterstained with methylene-blue. Centrally located capillary with endothelial cells serve as negative control (original magnification \times 400).

ferential diagnoses, these antibodies are of great diagnostic help.

Antibodies Against Vascular Antigens

Endothelial cells and endothelial cell-derived tumors are characterized by the expression of Factor VIII related antigen or von Willebrand factor (Fig. 1-41). This factor is a glycoprotein that is also synthesized in megakaryocytes (170). Because background staining is often seen, especially in hemorrhagic tissue, its use in combination with other endothelial markers, such as



Figure 1-39 Immunohistochemistry. Langerhans cell histiocytosis. Langerhans cells show a strong cytoplasmic immunoreactivity for antibodies against CD1a. Accompanying lymphocytes, macrophages, and giant cells are negative. Methylene-blue counterstaining (original magnification ×400).



Figure 1-41 Immunohistochemistry. Epithelioid angiosarcoma. Note strong intracytoplasmic immunoreactivity for Factor VIII–related antigen. Methylene-blue counterstaining (original magnification \times 400).

CD31 (or platelet-endothelial adhesion molecule) and CD34 (or hematopoietic progenitor cell antigen), is advisable. Both are transmembrane glycoproteins that are expressed in almost all endothelial cells and related tumors (39,124). However, CD34 in particular is also expressed in spindle cell tumors such as hemangiopericytomas, gastrointestinal stromal tumors (GISTs), or solitary fibrous tumors (34).

Antibodies Against Muscle Antigens

For characterization of muscle tissue, antibodies to actins can be used in addition to desmin IHC. Actin is present in all types of cells. By the use of isoelectric focusing, three different groups of actin molecules can be separated-alpha, beta, and gamma actins-consisting of six different isoforms that have been identified by amino acid sequencing as four alpha actins (two sarcomeric-alpha-skeletal and alpha-cardiac muscle actins, and two smooth muscle actins-alpha and gamma, present in the contractile apparatus, and, ultrastructurally, in thin filaments of the respective cells), and two (beta and gamma) nonmuscle cytoskeletal actins (89,204,205). Antibodies to muscle-specific actins are widely used for characterization of muscle tumors (125,166,176,177,184). However, positive immunoreactions have also been observed in osteoblasts, bone marrow stromal cells, osteosarcomas, and malignant fibrous histiocytoma of bone (59,91,202).

Other Useful Antibodies in Bone Tumor Pathology

Enolases are enzymes of the glycolytic pathway. They consist of three different subunits: alpha, beta, and gamma. The functional enzyme is a dimer consisting of two identical subunits (118). The gamma subunit, or neuron-specific enolase (NSE), is found in neural and neuroendocrine cells. Aberrant expression, e.g., in smooth muscle cells, has also been observed (72). NSE positivity is found in PNETs belonging to the Ewing tumor family and in neuroblastomas and melanomas (5,40,147,149).

Synaptophysin is localized in the membrane of presynaptic vesicles. It is a glycosylated acidic protein of 38 KD present in neurons, at neuromuscular junctions, in neuroendocrine cells, and also in PNETs (149,213,214).

In 1965, Moore isolated a protein from bovine brain that was termed S-100 protein because of its solubility in 100% ammonium sulfate. Later it became clear that it comprised a mixture of two structurally related proteins, S-100A and S-100B1 (80,81,131). These belong to an expanding family of small calcium-binding proteins (S-100 protein family) with a molecular weight of 10 to 12 KD, forming homo- and heterodimers (42,119). S-100 proteins are present within cells and nuclei not only in neural tissues but also in Langerhans cells of the epidermis, melanocytes and melanomas, cartilagederived cells and their tumors, and chordomas (30,137–139,143).

Antibodies to cell cycle–related proteins have recently been applied in bone tumor IHC. Alterations of the cell cycle are considered fundamental to cancer development (88). Particularly in low-grade osteosarcomas and parosteal osteosarcomas, CDK4, a cell-cycle regulator, and MDM2, a regulator of TP53, which normally are not expressed in amounts detectable by IHC, are often overexpressed (159).

The proliferating activity of a tumor can be assessed by identifying all cells that are not in G0-phase of the cell cycle. These cells express a cell stage–related nuclear protein that can be visualized by the MIB1 antibody (90,173).

It should be remembered that IHC is a rapidly expanding field. New markers are described almost every month. They are first considered to be and are announced as "specific" for some tumors. With increasing experience, they become only "characteristic," and are eventually found in a number of different entities. Interpretation of staining results without considering the morphologic context will lead to erroneous assumptions.

Electron Microscopy

At present, the contribution of electron microscopy to the diagnosis of bone tumor is only minimal (Fig. 1-42) (85,203). Ultrastructural investigations can still be of use in the evaluation of small cell neoplasms, spindlecell tumors, and metastatic disease (111,156). However, these studies are useful only if precise questions, emerging from the evaluation of the routine H&E slides, are addressed, e.g., detection of myofilaments and basal lamina material for discriminating leiomyosarcomas from other spindle-cell sarcomas or Birbeck granules (Fig. 1-43) for identifying a lesion as Langerhans cell histiocytosis (146).

Genetics in Bone Tumors

Genetic investigations have contributed to a better understanding of cancer development. Moreover, some



Figure 1-42 Electron microscopy. Fibrous cortical defect. Cell extension (*middle*) containing numerous organelles and the somewhat distorted nucleus (*upper left corner*) is surrounded by stroma with densely arranged immature fibrils. Only a few mature-looking collagen fibrils are seen (*lower right corner*).



Figure 1-43 Electron microscopy. Langerhans cell histiocytosis. Rod-like and tennis racket–shaped cytoplasmic structures (Birbeck granules; *center*) that present a zipper-like internal core are characteristic for this disorder (original magnification \times 64,000).

genetic changes may represent specific cancer signatures such as the translocation of genetic material from chromosome 11 to chromosome 22, which is highly characteristic for Ewing sarcomas. Recent technical advances have dramatically reduced the time interval from the arrival of material at the pathology laboratory to the completion of a genetic analysis as an adjunct to morphologic diagnosis. Some of these methods are briefly discussed in the following sections.

Flow Cytometry

Flow cytometry (FCM) is a quantitative automated method used to analyze the DNA content and the proliferation rate of isolated cells (35,211). To determine the DNA content, the DNA is stained stoichiometrically with specific fluorescent dyes and the emitted fluorescent signal is measured as the nuclei pass one by one through a nozzle-equipped chamber (flow cell). The intensity of the signal is proportional to the amount of DNA in the isolated nuclei. DNA distribution and respective number of cells are calculated, compared with a standard (usually lymphocytes), and then assigned to each phase of the cell cycle (95).

In FCM the total amount of DNA is assessed without considering the distribution of single chromosomes. Therefore, a terminology different from that of cytogenetics has to be applied. The prefix DNA, along with the ploidy terms, is used for FCM investigations. Cells or tumors with the same DNA content as normal control cells, e.g., lymphocytes, are by convention called DNA diploid, whereas cells with less or more DNA are described as DNA aneuploid. Somatic cells ordinarily contain an amount of DNA that is "2C," or diploid, a term reserved for cytogenetic evaluation (77). In the G0 phase, cells are DNA diploid (2C). They then enter the pre-division period, G1 phase, which is followed by

the phase of DNA synthesis, the S phase. The number of cells in the S phase of the cell cycle represents the proliferating cell fraction (S-phase fraction, SPF). The amount of DNA gradually doubles until the cells are DNA tetraploid (4C). The cells have reached the premitotic G2 phase, after which mitotic division is performed during the M phase. DNA aneuploid cells contain neither a 2C nor a 4C DNA concentration.

From investigations of epithelial cells and tumors, it is known that DNA aneuploidy and often an increased SPF are found in most carcinomas (50). DNA aneuploidy and increased SPF are also linked to malignancy in cartilaginous tumors (Fig. 1-44). However, studies of endocrine and musculoskeletal tumors have shown that benign aneuploid lesions do exist (3,4). Therefore, the value of FCM as an additional tool for judging proliferating lesions and predicting their biological behavior must be interpreted differently for each tumor type.

Cytogenetics

Cytogenetics is the morphologic study of chromosomes with the use of karyotyping (22). Classical karyotyping still plays a role in the scientific study of many tumors. For identification of numerical and structural aberrations, tumor cells must be freshly obtained, immediately shipped, and cultured in appropriate media. Mitoses of tumor cells can be artificially interrupted during metaphase and the spreaded chromosomes can be stained with special dyes (e.g., Giemsa). This generates a specific transverse pattern ("banding") of each chromosome that can be analyzed. A metaphase chromosome is divided into two arms by a central region, the centromere (the constricted site on the chromosome at which the fibers of the mitotic spindle attach). The short arm is labeled with the letter p, the long arm with the letter q. The chromosomal ends (ter) are called telomeres (tel). The Giemsa-stained, microscopically visible cytogenetic bands or regions can be subdivided at higher magnifications into sub-bands. Both are numbered consecu-



Figure 1-44 Flow cytometry. Chondrosarcoma. Flow cytometry histogram of chondrocytes of grade 2 tumor presents multiploid DNA pattern.

tively beginning at the centromere. For example, the gene for type I collagen, CollA1, is located on 17q21.31–q22, i.e., on chromosome 17, long arm, band 21, sub-band 31 to band 22. For more detailed information the reader is referred to the International System for Human Cytogenetic Nomenclature (180).

Molecular Cytogenetics

With the development of fluorescent in situ hybridization (FISH), genetic analyses of interphase nuclei became possible, even in fixed and paraffin-embedded material, by application of differentially labeled centromere-specific and sequence-specific probes to tissues and cells (122). Multicolor labeling techniques made it possible to identify more than one DNA sequence simultaneously (187). Break-apart probes, differentially labeled DNA fragments flanking, e.g., the EWS breakpoint region on chromosome 22q12 in Ewing sarcomas, indicate translocations by the splitting of the normally paired green and red (or fused yellow) signal (Fig. 1-45) (23). Because results can be obtained within 48 hours, FISH methods are increasingly used as diagnostic tools.

Comparative genomic hybridization (CGH) also utilizes DNA hybridization procedures. Tumor DNA and differentially labeled reference DNA are hybridized to normal metaphase chromosomes. Changes in the ratio of the two fluorochromes (e.g., red for tumor DNA, green for normal DNA) reflect losses or gains of tumor DNA that may be caused by amplifications, deletions,



Figure 1-45 Fluorescence in situ hybridization (FISH). Ewing sarcoma. FISH using a commercially available breakapart probe involving the EWSR1 (Ewing sarcoma breakpoint region 1) gene on chromosome 22q12. Two probes, labeled in red and green, are applied flanking the breakpoint region from both sides. If no translocation is present, both colors should be in close contact. If, however, a translocation is present, the green and the red signal is separated. The cell in the center shows one fusion and one rearrangement (original magnification \times 1,000).

or duplications along the respective chromosomes (87). CGH is therefore a very useful tool to detect changes in DNA copy number. However, limitations exist because, for example, balanced aberrations cannot be detected.

The polymerase chain reaction (PCR) developed by K.B. Mullis is a powerful technique, for which he was honored in 1993 with the Nobel prize (135). This technique allows automatic generation of many copies of even extremely low amounts of target DNA by using a mixture of the four nucleotides that are present in normal DNA (adenine, guanine, cytosine, and thymidine), an oligonucleotide sequence as a starting molecule (primer), a DNA template and, as a synthesizing enzyme, thermostable DNA polymerases. This product can then be further analyzed by comparing the amplified target DNA with known sequences that code for a specific protein. In this way, point mutations (single nucleotide disparities) can be detected, e.g., point mutations in a signal-transducing membrane-bound protein, GS alpha, that is involved in the development of fibrous dysplasia. With reverse transcription PCR (RT-PCR), messenger RNA (mRNA) can be amplified. Initially, mRNA is converted by the enzyme reverse transcriptase into cDNA (complementary DNA: DNA without introns, i.e., non-protein-coding DNA sequences that must be removed-spliced out-before translation into mRNA), which then will be amplified. Using this method, fusion transcripts from chromosomal translocations, e.g., in Ewing sarcomas, can be detected (Fig. 1-46) (123). For details and other applications, such as quantitative (real-time) PCR, the reader is referred to special textbooks (101).

Despite the use of the previously mentioned studies, a small percentage of bone tumors are not easily categorized. The pathologist must then attempt to assess the biologic potential of the neoplasm, even if it cannot be given a specific name. In such cases, the classic signs of nuclear pleomorphism and hyperchromaticity, mitotic rate and the presence of atypical mitoses, and the identification of spontaneous necrosis are sought to help distinguish benign from malignant lesions. However, it must be kept in mind that in Ewing sarcoma, for example, the tumor with the highest grade of malignancy, mitoses are almost never found in the histologic preparation. This is probably because in a tumor with a high growth rate mitoses take place so rapidly as to be completed before the tumor tissue, after its excision, has been subjected to fixation. Observation of the interface area of the tumor with normal bone may reveal subtle infiltration of intertrabecular marrow spaces or Haversian canals by tumor cells and thus point to malignancy.

In the final analysis, only close cooperation among the radiologist, the pathologist, and the orthopedic surgeon in the review of history, imaging studies, and biopsy materials may lead to an accurate diagnosis of a bone lesion. In most cases, careful analysis of all of these variables makes it possible to identify the various benign and malignant tumors and tumor-like conditions.



Figure 1-46 Polymerase chain reaction (PCR). Because of variable chromosomal breakpoint locations, the EWS gene fusion with FLI-1 t(11;22)(q24;q12) and ERG t(21;22)(q21;q12) genes are highly heterogeneous, resulting in different sizes of the amplification products. This can be detected by PCR. A: RNA control: All samples (1-5: patients' samples; lane 6, HTB cells-osteosarcoma-derived cell line; lane 8, A673 cells-Ewing sarcoma-derived cell line) except lane 7 (water instead of RNA) contain the 738-bp fragment of the mRNA of the ubiguitously expressed transcription factor YY1 (M, molecular weight marker). B: Nested PCR (special PCR technique that allows detection of gene transcripts present in low copy numbers) with primers that specifically detect EWS-FLI-1 and EWS-ERG transcripts. Lanes 1-5, patients' samples as in A showing transcripts of different size; lane 6, the osteosarcoma-derived HTB cells used as control are negative as expected; lane 7, negative control (water); lane 8, the Ewing sarcoma-derived A673 cells also show fusion transcripts as expected (M, molecular weight marker). (For more details see ref. 123.) (Modified from Meier VS, Kuhne T, Jundt G, Gudat F. Molecular diagnosis of Ewing tumors: improved detection of EWS-FLI-1 and EWS-ERG chimeric transcripts and rapid determination of exon combinations. Diagn Mol Pathol 1998;7:29-35.)

REFERENCES

- Aisen AM, Martel W, Braunstein EM, McMillin KI. MRI and CT evaluation of primary bone and soft tissue tumors. Am J Roentgenol 1986;146:749–756.
- Alavi A, Lakhani P, Mavi A, et al. PET: a revolution in medical imaging. *Radiol Clin North Am* 2004;42:983–1001.

- Alho A, Connor JF, Mankin HJ, et al. Assessment of malignancy of cartilage tumors using flow cytometry. J Bone Joint Surg Am 1983;65A:779–785.
- Alho A, Skjeldal S, Pettersen EO, et al. Aneuploidy in benign tumors and nonneoplastic lesions of musculoskeletal tissues. *Cancer* 1994;73:1200–1205.
- Amann G, Zoubek A, Salzer-Kuntschik M, et al. Relation of neurological marker expression and EWS gene fusion types in MIC2/CD99-positive tumors of the Ewing family. *Hum Pathol* 1999;30:1058–1064.
- 6. An Y, Martin K, eds. *Handbook of histological methods for bone and cartilage*. Totowa, NJ: Humana Press, 2003.
- Anderson MW, Temple HT, Dussault RG, Kaplan PA. Compartmental anatomy: relevance to staging and biopsy of musculoskeletal tumors. *Am J Roentgenol* 1999;173:1663–1671.
- Aoki J, Watanabe H, Shinozaki T, et al. FDG PET of primary benign and malignant bone tumors: standardized uptake value in 52 lesions. *Radiology* 2001;219:774–777.
- 9. Bernard A, Boumsell L. The clusters of differentiation (CD) defined by the First International Workshop on Human Leucocyte Differentiation Antigens. *Hum Immunol* 1984;11:1–10.
- Berquist TH. Magnetic resonance imaging of musculoskeletal neoplasms. *Clin Orthop* 1989;244:101–118.
- 11. Berquist TH. Magnetic resonance imaging of primary skeletal neoplasms. *Radiol Clin North Am* 1993;31:411–424.
- Bloem JL. Radiological staging of primary malignant musculoskeletal tumors. A correlative study of CT, MRI, ^{99m}Tc scintigraphy and angiography. The Hague: A. Jongbloed, 1988.
- Bloem JL, Bluemm RG, Taminiau AHM, et al. Magnetic resonance imaging of primary malignant bone tumors. *RadioGraphics* 1987;7:425–445.
- Bloem JL, Reiser MF, Vanel D. Magnetic resonance contrast agents in evaluation of the musculoskeletal system. *Magn Res Q* 1990;6:136–163.
- Bloem JL, Taminiau AHM, Eulderink F, et al. Radiologic staging of primary bone sarcoma: MR imaging, scintigraphy, angiography, and CT correlated with pathologic examination. *Radiology* 1988;169:805–810.
- Bloem JL, Van der Woude HJ, Giernaerdt M, et al. Does magnetic resonance imaging make a difference for patients with musculoskeletal sarcoma? *Br J Radiol* 1997;70:327–337.
- Böck P, ed. Romeis Mikroskopische Technik, 17th ed. München: Urban & Schwarzenberg, 1989.
- Bodner G, Schocke MFH, Rachbauer F, et al. Differentiation of malignant and benign musculoskeletal tumors: combined color and power Doppler US and spectral wave analysis. *Radiology* 2002;223:410–416.
- Bohndorf K, Reiser M, Lochner B. Magnetic resonance imaging of primary tumors and tumor-like lesions of bone. *Skeletal Radiol* 1986;15:511–517.
- Boyko OB, Cory DA, Cohen MD, et al. MR imaging of osteogenic and Ewing's sarcoma. Am J Roentgenol 1987;148: 317–322.
- Bredella MA, Caputo GR, Steinbach LS. Value of FDG positron emission tomography in conjunction with MR imaging for evaluating therapy response in patients with musculoskeletal sarcomas. *Am J Roentgenol* 2002;179:1145–1150.
- 22. Bridge JA, Sandberg AA. Cytogenetic and molecular genetic techniques as adjunctive approaches in the diagnosis of bone and soft tissue tumors. *Skeletal Radiol* 2000;29:249–258.
- 23. Bridge RS, Rajaram V, Dehner LP, et al. Molecular diagnosis of Ewing sarcoma/primitive neuroectodermal tumor in routinely processed tissue: a comparison of two FISH strategies and RT-PCR in malignant round cell tumors. *Mod Pathol* 2005;Sep 2 [Epub ahead of print].
- 24. Brown KT, Kattapuram SV, Rosenthal DI. Computed tomography analysis of bone tumors: patterns of cortical destruction and soft tissue extension. *Skeletal Radiol* 1986;15:448–451.
- Calhoun PS, Kuszyk BS, Heath DG, et al. Three-dimensional volume rendering of spiral CT data: theory and method. *Radio-Graphics* 1999;19:745–764.
- Caluser CI, Abdel-Dayem HM, Macapinlac HA, et al. The value of thallium and three-phase bone scans in the evaluation of bone and soft tissue sarcomas. *Eur J Nucl Med* 1994;21:1198–1205.

- Chang L, Goldman RD. Intermediate filaments mediate cytoskeletal crosstalk. *Nat Rev Mol Cell Biol* 2004;5:601–613.
- Chew FS, Hudson TM. Radionuclide bone scanning of osteosarcoma: falsely extended uptake patterns. Am J Roentgenol 1982;139:49–54.
- Choong PFM, Kunisada T, Slavin J, et al. The role of thallium-201 and pentavalent dimercaptosuccinic acid for staging cartilaginous tumors. *Int Semin Surg Oncol* 2004;1:10.
- Cocchia D, Michetti F, Donato R. Immunochemical and immuno-cytochemical localization of S-100 antigen in normal human skin. *Nature* 1981;294:85–87.
- Coffre C, Vanel D, Contesso G, et al. Problems and pitfalls in the use of computed tomography for the local evaluation of long bone osteosarcoma. Report on 30 cases. *Skeletal Radiol* 1985;13:147–153.
- Cohen EK, Kressel HY, Frank TS, et al. Hyaline cartilage-origin bone and soft-tissue neoplasms: MR appearance and histologic correlation. *Radiology* 1988;167:477–481.
- Cohen MD, Weetman RM, Provisor AJ, et al. Efficacy of magnetic resonance imaging in 139 children with tumors. *Arch Surg* 1986;121:522–529.
- Coindre JM. Immunohistochemistry in the diagnosis of soft tissue tumours. *Histopathology* 2003;43:1–16.
- Couturier J. Cytogenetics. In: Forest M, Tomeno B, Vanel D, eds. Orthopedic surgical pathology: diagnosis of tumors and pseudotumoral lesions of bone and joints. Edinburgh: Churchill Livingstone, 1988:39–44.
- Crim JR, Seeger LL, Yao L, et al. Diagnosis of soft-tissue masses with MR imaging: can benign masses be differentiated from malignant ones? *Radiology* 1992;185:581–586.
- Davies MA, Wellings RM. Imaging of bone tumors. Curr Opin Radiol 1992;4:32–38.
- Delling G. Diagnostik von Knochentumoren. Verh Dtsch Ges Pathol 1998;82:121–132.
- Deshpande V, Rosenberg AE, O'Connell JX, Nielsen GP. Epithelioid angiosarcoma of the bone: a series of 10 cases. *Am J Surg Pathol* 2003;27:709–716.
- Dhillon AP, Rode J, Leathem A. Neurone specific enolase: an aid to the diagnosis of melanoma and neuroblastoma. *Histopathology* 1982;6:81–92.
- Domagala W, Osborn M. Immunocytochemistry. In: Koss LG, Woyke S, Olszewski W, eds. Aspiration biopsy. Cytologic interpretation and histologic bases, 2nd ed. New York: Igaku-Shoin, 1992:55–108.
- Donato R. S-100: a multigenic family of calcium-modulated proteins of the EF-hand type with intracellular and extracellular functional roles. *Int J Biochem Cell Biol* 2001;33:637–668.
- Dwyer AJ, Frank JA, Sank VJ, et al. Short-TI inversion-recovery pulse sequence: analysis and initial experience in cancer imaging. *Radiology* 1988;169:827–836.
- Elgazzar AH, Malki AA, Abdal-Dayem HM, et al. Role of thallium-201 in the diagnosis of solitary bone lesions. *Nucl Med Comm* 1989;10:477–485.
- Elias DA, White LM, Simpson DJ, et al. Osseous invasion by softtissue sarcoma: assessment with MR imaging. *Radiology* 2003;229:145–152.
- Enneking WF. Staging of musculoskeletal neoplasms. Skeletal Radiol 1985;13:183–194.
- Erlemann R, Reiser MF, Peters PE, et al. Musculoskeletal neoplasms: static and dynamic Gd-DPTA-enhanced MR imaging. *Radiology* 1989;171:767–773.
- Ewing J. A review and classification of bone sarcomas. Arch Surg 1922;4:485–533.
- 49. Fechner RE, Mills SE. *Tumors of the bones and joints.* Washington, DC: Armed Forces Institute of Pathology, 1993:1–16.
- Feichter GE, Maier H, Adler D, et al. S-phase fractions and DNA-ploidy of oropharyngeal squamous epithelium carcinomas compared with histologic grade, stage, response to chemotherapy and survival. *Acta Otolaryngol (Stockh)* 1987;104:377–384.
 Feldman F, van Heertum R, Manos C. ¹⁸FDG PET scanning of
- Feldman F, van Heertum R, Manos C. ¹⁸FDG PET scanning of benign and malignant musculoskeletal lesions. *Skeletal Radiol* 2003;32:201–208.
- Feldman F, van Heertum R, Saxena C, Parisien M. ¹⁸FDG-PET applications for cartilage neoplasms. *Skeletal Radiol* 2005;34: 367–374.

- Fishman EK, Magid D, Ney DR, et al. Three dimensional imaging. *Radiology* 1991;181:321–337.
- Fletcher BD. Response of osteosarcoma and Ewing sarcoma to chemotherapy: imaging evaluation. Am J Roentgenol 1991;157: 825–833.
- Frank JA, Ling A, Patronas NJ, et al. Detection of malignant bone tumors: MR imaging vs. scintigraphy. *Am J Roentgenol* 1990; 155:1043–1048.
- Franke WW, Schiller DL, Moll R, et al. Diversity of cytokeratins. Differentiation specific expression of cytokeratin polypeptides in epithelial cells and tissues. *J Mol Biol* 1981;153:933–959.
- Frouge C, Vanel D, Coffre C, et al. The role of magnetic resonance imaging in the evaluation of Ewing sarcoma. *Skeletal Radiol* 1988;17:387–392.
- Galasko CS. The pathological basis for skeletal scintigraphy. J Bone J Surg 1975;57B:353–359.
- 59. Galmiche MC, Koteliansky VE, Briere J, et al. Stromal cells from human long-term marrow cultures are mesenchymal cells that differentiate following a vascular smooth muscle differentiation pathway. *Blood* 1993;82:66–76.
- Gao Z, Kahn LB. The application of immunohistochemistry in the diagnosis of bone tumors and tumor-like lesions. *Skeletal Radiol* 2005;34:755–769.
- Gillespy T III, Manfrini M, Ruggieri P, et al. Staging of intraosseous extent of osteosarcoma: correlation of pre-operative CT and MR imaging with pathologic macroslides. *Radiology* 1988;167:765–767.
- 62. Gold RH, Bassett LW. Radionuclide evaluation of skeletal metastases: practical considerations. *Skeletal Radiol* 1986;15:1–9.
- Golfieri R, Baddeley H, Pringle JS, et al. Primary bone tumors. MR morphologic appearance correlated with pathologic examinations. *Acta Radiol* 1991;32:290–298.
- Golfieri R, Baddeley H, Pringle JS, et al. MR imaging in primary bone tumors: therapeutic implications. *Eur J Radiol* 1991;12: 201–207.
- Golfieri R, Baddeley H, Pringle JS, Souhami R. The role of the STIR sequence in magnetic resonance imaging examination of bone tumors. *Br J Radiol* 1990;63:251–256.
- Graif M, Pennock JM, Pringle J, et al. Magnetic resonance imaging: comparison of four pulse sequences in assessing primary bone tumors. *Skeletal Radiol* 1989;18:439–444.
- Greenfield GB, Warren DL, Clark RA. MR imaging of periosteal and cortical changes of bone. *RadioGraphics* 1991;11:611–623.
- Greenspan A, Klein MJ. Radiology and pathology of bone tumors. In: Lewis MM, ed. *Musculoskeletal oncology. A multidisciplinary approach.* Philadelphia: WB Saunders, 1992:13–72.
- Greenspan A. Pragmatic approach to bone tumors. Semin Orthop 1991;6:125–133.
- Greenspan A, Stadalnik RC. Bone island: scintigraphic findings and their clinical application. *Can Assoc Radiol J* 1995;46: 368–379.
- Greenspan A, Stadalnik RC. Central versus eccentric lesions of long tubular bone. *Semin Nucl Med* 1996;26:201–206.
- Haimoto H, Takahashi Y, Koshikawa T, et al. Immunohistochemical localization of gamma-enolase in normal human tissues other than nervous and neuroendocrine tissues. *Lab Invest* 1985;52:257–263.
- Hanna SL, Fletcher BD, Parham DM, Bugg MR. Muscle edema in musculoskeletal tumors: MR imaging characteristics and clinical significance. *J Magn Reson Imag* 1991;1:441–449.
- Hanna SL, Langston JW, Gronemeyer SA, Fletcher BD. Subtraction technique for contrast-enhanced MR images of musculoskeletal tumors. *Magn Reson Imag* 1990;8:213–215.
- Hau MA, Kim JI, Kattapuram S, et al. Accuracy of CT-guided biopsies in 359 patients with musculoskeletal lesions. *Skeletal Radiol* 2002;31:349–353.
- Hayes CW, Conway WF, Sundaram M. Misleading aggressive MR imaging appearance of some benign musculoskeletal lesions. *RadioGraphics* 1992;12:1119–1134.
- Hiddemann W, Schumann J, Andreef M, et al. Convention on nomenclature for DNA cytometry. Committee on Nomenclature, Society for Analytical Cytology. *Cancer Genet Cytogenet* 1984;13:181–183.
- Horobin R, Kierman J, eds. Conn's biological stains. A handbook of dyes and fluorochromes for use in biology and medicine, 10th ed. Oxford: BIOS Scientific Publishers Ltd., 2002.

- Hudson TM. Radiologic-pathologic correlation of musculoskeletal lesions. Baltimore: Williams & Wilkins, 1987.
- Isobe T, Okuyama T. The amino-acid sequence of S-100 protein (PAP I-b protein) and its relation to the calcium-binding proteins. *Eur J Biochem* 1978;89:379–388.
- Isobe T, Okuyama T. The amino-acid sequence of the alpha subunit in bovine brain S-100a protein. *Eur J Biochem* 1981;116:79–86.
- 82. Jaffe E, Harris N, Stein H, Vardiman J, eds. *Tumours of haematopoietic and lymphoid tissues*. Lyon: IARC Press, 2001.
- Jelinek JS, Murphey MD, Welker JA, et al. Diagnosis of primary bone tumors with image-guided percutaneous biopsy: experience with 110 tumors. *Radiology* 2002;223:731–737.
- Johnson LC. A general theory of bone tumors. Bull NY Acad Med 1953;29:164–171.
- Joy DC, Pawley JB. High-resolution scanning electron microscopy. Ultramicroscopy 1992;47:80–100.
- Jurik AG, Möller SH, Mosekilde L. Chronic sclerosing osteomyelitis of the iliac bone: etiological possibilities. *Skeletal Radiol* 1988;17:114–118.
- Kallioniemi A, Kallioniemi OP, Sudar D, et al. Comparative genomic hybridization for molecular cytogenetic analysis of solid tumors. *Science* 1992;258:818–821.
- Kastan MB, Bartek J. Cell-cycle checkpoints and cancer. *Nature* 2004;432:316–323.
- 89. Kedes L, Ng SY, Lin CS, et al. The human beta-actin multigene family. *Trans Assoc Am Physicians* 1985;98:42–46.
- 90. Key G, Becker MH, Baron B, et al. New Ki-67-equivalent murine monoclonal antibodies (MIB 1-3) generated against bacterially expressed parts of the Ki-67 cDNA containing three 62 base pair repetitive elements encoding for the Ki-67 epitope. *Lab Invest* 1993;68:629–636.
- Kinner B, Spector M. Expression of smooth muscle actin in osteoblasts in human bone. J Orthop Res 2002;20:622–632.
- Kloiber R. Scintigraphy of bone tumors. In: Current concepts of diagnosis and treatment of bone and soft tissue tumors. New York: Springer-Verlag, 1984:55–60.
- Kransdorf MJ, Jelinek J, Moser RP Jr, et al. Soft-tissue masses: diagnosis using MR imaging. Am J Roentgenol 1989;153:541–547.
- Kransdorf MJ, Murphey MD. Radiologic evaluation of softtissue masses: a current perspective. Am J Roentgenol 2000; 175:575-587.
- Kreicbergs A. DNA cytometry of musculoskeletal tumors. Acta Orthop Scand 1990;61:282–297.
- Kricun ME. Radiographic evaluation of solitary bone lesions. Orthop Clin North Am 1983;14:39–64.
- Kuszyk BS, Heath DG, Bliss DF, Fishman EK. Skeletal 3-D CT: advantages of volume rendering over surface rendering. *Skeletal Radiol* 1996:25:207–214.
- Lang P, Honda G. Roberts T, et al. Musculoskeletal neoplasm: perineoplastic edema versus tumor on dynamic postcontrast MR images with spatial mapping of instantaneous enhancement rates. *Radiology* 1995;197:831–839.
- 99. Larsson SE, Lorentzon R. The incidence of malignant primary bone tumors in relation to age, sex and site. A study of osteogenic sarcoma, chondrosarcoma, and Ewing's sarcoma diagnosed in Sweden from 1958–1968. J Bone Joint Surg Br 1974;56B:534–540.
- Lee JK, Yao L, Wirth CR. MR imaging of solitary osteochondromas: report of eight cases. Am J Roentgenol 1987;149:557–560.
- 101. Leonard D, ed. *Diagnostic molecular pathology*. Philadelphia: Saunders, 2003.
- 102. Lewis MM, Sissons HA, Norman A, Greenspan A. Benign and malignant cartilage tumors. In: Griffin PP, ed. Instructional course lectures. Chicago: American Academy of Orthopaedic Surgeons, 1987:87–114.
- Lindsay RL. The use of MRI in the evaluation of musculoskeletal neoplasms. *Applied Radiol* 1993;65 (suppl):67.
- Lodge MA, Lucas JD, Marsken PK, et al. A PET study: 18-FDG uptake in soft tissue masses. *Eur J Nucl Med* 1999;26:22–30.
- 105. Lodwick GS. A systematic approach to the roentgen diagnosis of bone tumors. In: M.D. Anderson Hospital and Tumor Institute— Clinical conference on cancer: tumors of bone and soft tissue. Chicago: Year Book, 1965:49–68.
- Lodwick GS. Solitary malignant tumors of bone: the application of predictor variables in diagnosis. *Semin Roentgenol* 1966;1:293–313.

- 107. Lodwick GS, Wilson AJ, Farrell C, et al. Determining growth rates of focal lesions of bone from radiographs. *Radiology* 1980;134:577–583.
- Lodwick GS, Wilson AJ, Farrell C, et al. Estimating rate of growth in bone lesions. Observer performance and error. *Radiology* 1980;134:585–590.
- 109. Ma LD, Frassica FJ, McCarthy EF, et al. Benign and malignant musculoskeletal masses: MR imaging differentiation with rim-tocenter differential enhancement ratios. *Radiology* 1997;202: 739–744.
- Ma LD, Frassica FJ, Scott WW Jr, et al. Differentiation of benign and malignant musculoskeletal tumors: potential pitfalls with MR imaging. *RadioGraphics* 1995;15:349–366.
- 111. Mackay B, Ordonez NG. Pathological evaluation of neoplasms with unknown primary tumor site. *Semin Onco.* 1993;20:206–228.
- 112. Mackie GC, Schlicht SM. Scintigraphic findings in synovial sarcoma with structural correlation. *Aust Radiol* 2004;48:466–472.
- 113. Madewell JE, Ragsdale BD, Sweet DE. Radiologic and pathologic analysis of solitary bone lesions. Part I: Internal margins. *Radiol Clin North Am* 1981;19:715–748.
- 114. Magid D. Two-dimensional and three-dimensional computed tomographic imaging in musculoskeletal tumors. *Radiol Clin North Am* 1993;31:425–447.
- 115. Manaster BJ, Ensign MF. The role of imaging in musculoskeletal tumors. *Semin US CT MR* 1989;10:498–517.
- 116. Manjo G, Joris I. Cells, tissues, and disease, 2nd ed. New York, Oxford: Oxford University Press, 2004:1005.
- 117. Mankin HJ, Lange TA, Spanier SS. The hazards of biopsy in patients with malignant primary bone and soft-tissue tumors. J Bone Joint Surg 1982;64:1121–1127.
- Marangos PJ, Schmechel DE. Neuron specific enolase, a clinically useful marker for neurons and neuroendocrine cells. *Annu Rev Neurosci* 1987;10:269–295.
- 119. Marenholz I, Heizmann CW, Fritz G. S-100 proteins in mouse and man: from evolution to function and pathology (including an update of the nomenclature). *Biochem Biophys Res Commun* 2004;322:1111–1122.
- McCook BM, Sandler MP, Powers TA, et al. Correlative bone imaging. In: *Nuclear medicine annual*. New York: Raven Press, 1989:143–177.
- McNeil BJ. Value of bone scanning in neoplastic disease. Semin Nucl Med 1984;14:277–286.
- 122. McNeil N, Ried T. Novel molecular cytogenetic techniques for identifying complex chromosomal rearrangements: technology and applications in molecular medicine. *Expert Rev Mol Med* 2000;2000:1–14.
- 123. Meier VS, Kuhne T, Jundt G, Gudat F. Molecular diagnosis of Ewing tumors: improved detection of EWS-FLI-1 and EWS-ERG chimeric transcripts and rapid determination of exon combinations. *Diagn Mol Pathol* 1998;7:29–35.
- 124. Meis-Kindblom JM, Kindblom LG. Angiosarcoma of soft tissue: a study of 80 cases. *Am J Surg Pathol* 1998;22:683–697.
- Miettinen M. Antibody specific to muscle actins in the diagnosis and classification of soft tissue tumors. Am J Pathol 1988; 130:205–215.
- Milan L, Trachtenberg MC. Ultrasonic decalcification of bone. *Am J Surg Pathol* 1981;5:573–579.
- 127. Mirowitz SA. Fast scanning and fat-suppression MR imaging of musculoskeletal disorders. Am J Roentgenol 1993;161: 1147–1157.
- 128. Mirowitz SA, Apicella P, Reinus WR, Hammerman AM. MR imaging of bone marrow lesions: relative conspicuousness on Tlweighted, fat-suppressed T2-weighted, and STIR images. *Am J Roentgenol* 1994;162:215–221.
- 129. Moll R, Franke WW, Schiller DL, et al. The catalog of human cytokeratins: patterns of expression in normal epithelia, tumors and cultured cells. *Cell* 1982;31:11–24.
- Montero C. The antigen-antibody reaction in immunohistochemistry. J Histochem Cytochem 2003;51:1–4.
- Moore BW. A soluble protein characteristic of the nervous system. Biochem Biophys Res Commun 1965;19:739–744.
- Moore SG, Bisset GS, Siegel MJ, Donaldson JS. Pediatric musculoskeletal MR imaging. *Radiology* 1991;179:345–360.
- Moser RP, Madewell JE. An approach to primary bone tumors. Radiol Clin North Am 1987;25:1049–1093.

- Mulder JD, Kroon HM, Schütte HE, Taconis WK. Radiologic atlas of bone tumors. Amsterdam: Elsevier, 1993:9–46.
- Mullis KB, Faloona FA. Specific synthesis of DNA in vitro via a polymerase-catalyzed chain reaction. *Methods Enzymol* 1987; 155:355–350.
- Murphy WA Jr. Imaging bone tumors in the 1990s. Cancer 1991;67:1169–1176.
- 137. Nakajima T, Watanabe S, Sato Y, et al. An immunoperoxidase study of S-100 protein distribution in normal neoplastic tissues. *Am J Surg Pathol* 1982;6:715–727.
- Nakamura Y, Becker LE, Marks A. S-100 protein in human chordoma and human and rabbit notochord. *Arch Pathol Lab Med* 1983;107:118–120.
- Nakamura Y, Becker LE, Marks A. S-100 protein in tumors of cartilage and bone. An immunohistochemical study. *Cancer* 1983;52:1820–1824.
- 140. Nelson SW. Some fundamentals in the radiologic differential diagnosis of solitary bone lesions. *Semin Roentgenol* 1966;1:244–267.
- Norman A, Schiffman M. Simple bone cyst: factors of age dependency. *Radiology* 1977;124:779–782.
- 142. Okada K, Hasegawa T, Yokoyama R, et al. Osteosarcoma with cytokeratin expression: a clinicopathological study of six cases with an emphasis on differential diagnosis from metastatic cancer. *J Clin Pathol* 2003;56:742–746.
- Okajima K, Honda I, Kitagawa T. Immunohistochemical distribution of S-100 protein in tumors and tumor-like lesions of bone and cartilage. *Cancer* 1988;61:792–799.
- 144. Oliveira AM, Nascimento AG. Grading in soft tissue tumors: principles and problems. *Skeletal Radiol* 2001;30:543–559.
- 145. Olson P, Everson LI, Griffith HJ. Staging of musculoskeletal tumors. *Radiol Clin North Am* 1994;32:151–162.
- 146. Ordonez NG, Mackay B. Electron microscopy in tumor diagnosis: indications for its use in the immunohistochemical era. *Hum Pathol* 1998;29:1403–1411.
- 147. Osborn M, Dirk T, Kaser H, et al. Immunohistochemical localization of neurofilaments and neuron-specific enolase in 29 cases of neuroblastoma. *Am J Pathol* 1986;122:433–442.
- 148. Panicek DM, Gatsonis C, Rosenthal DI, et al. CT and MR imaging in the local staging of primary malignant musculoskeletal neoplasms: report of the Radiology Diagnostic Oncology Group. *Radiology* 1997;202:237–246.
- 149. Parham DM, Hijazi Y, Steinberg SM, et al. Neuroectodermal differentiation in Ewing's sarcoma family of tumors does not predict tumor behavior. *Hum Pathol* 1999;30:911–918.
- Petasnick JP, Turner DA, Charters JR, et al. Soft-tissue masses of the locomotor system: comparison of MR imaging with CT. *Radiology* 1986;160:125–133.
- 151. Pettersson H, Eliasson J, Egund N, et al. Gadolinium-DTPA enhancement of soft tissue tumors in magnetic resonance imaging—preliminary clinical experience in five patients. *Skeletal Radiol* 1988;14:319–323.
- 152. Pettersson H, Gillespie T III, Hamlin DJ, et al. Primary musculoskeletal tumors: examination with MR imaging compared with conventional modalities. *Radiology* 1987;164:237–241.
- Pettersson H, Slone RM, Spanier S, et al. Musculoskeletal tumors: T1 and T2 relaxation times. *Radiology* 1988;167:783–785.
- Pettersson H, Spanier S, Fitzsimmons JR, et al. MR imaging relaxation measurements in musculoskeletal tumors and surrounding tissue. *Radiology* 1985;157(P):109.
- Pettersson H, Springfield DS, Enneking WF. Radiologic management of musculoskeletal tumors. New York: Springer-Verlag, 1987:9–13.
- 156. Peydro-Olaya A, Llombart-Bosch A, Carda-Batalla C, Lopez-Guerrero JA. Electron microscopy and other ancillary techniques in the diagnosis of small round cell tumors. *Semin Diagn Pathol* 2003;20:25–45.
- 157. Prophet E, Mills B, Arrington J, Sobin L, eds. *Laboratory methods in histotechnology*. Washington, DC: Armed Forces Institute of Pathology, 1992.
- 158. Pui MH, Chang SK. Comparison of inversion recovery fast spinecho (FSE) with T2-weighted fat-saturated FSE and T1-weighted MR imaging in bone marrow lesion detection. *Skeletal Radiol* 1996;25:149–152.
- 159. Ragazzini P, Gamberi G, Benassi MS, et al. Analysis of SAS gene and CDK 4 and MDM 2 proteins in low-grade osteosarcoma. *Cancer Detect Prev* 1999;23:129–136.

- 160. Ragsdale BD, Madewell JE, Sweet DE. Radiologic and pathologic analysis of solitary bone lesions. Part II: Periosteal reactions. *Radiol Clin North Am* 1981;19:749–783.
- Rangdaeng S, Truong LD. Comparative immunohistochemical staining for desmin and muscle-specific actin. A study of 576 cases. *Am J Clin Pathol* 1991;96:32–45.
- 162. Reifenberger G, Szymas J, Wechsler W. Differential expression of glial- and neuronal-associated antigens in human tumors of the central and peripheral nervous system. Acta Neuropathol (Berl) 1987;74:105–123.
- Reinus WR, Wilson AJ. Quantitative analysis of solitary lesions of bone. *Invest Radiol* 1995;30:427–432.
- Reiser M, Rupp N, Biehl T. MR in diagnosis of bone tumors. *Eur J Radiol* 1985;5:1–7.
- Richardson ML, Kilcoyne RF, Gillespie T III, Genant HK. Magnetic resonance imaging of musculoskeletal neoplasms. *Radiol Clin North Am* 1987;24:259–267.
- Roholl PJ, Elbers HR, Prinsen I, et al. Distribution of actin isoforms in sarcomas: an immunohistochemical study. *Hum Pathol* 1990;21:1269–1274.
- Rosenthal DI. Computed tomography of orthopedic neoplasms. Orthop Clin North Am 1985;16:461–470.
- Rossleigh MA, Smith J, Yeh SD. Scintigraphic features of primary sacral tumors. *J Nucl Med* 1986;27:627–630.
- Rotte KH, Schmidt-Peter P, Kriedemann E. CT-evaluation of osseous tumors. *Eur J Radiol* 1986;6:5–8.
- 170. Ruggeri ZM. Structure and function of von Willebrand factor. Thromb Haemost 1999;82:576–584.
- 171. Sato O, Kawai A, Ozaki T, et al. Value of thallium-201 scintigraphy in bone and soft tissue tumors. J Orthop Sci 1998;6: 297–303.
- 172. Schajowicz F. Tumors and tumorlike lesions of bone. Pathology, radiology, and treatment, 2nd ed. Berlin: Springer-Verlag, 1994:1–21.
- 173. Schluter C, Duchrow M, Wohlenberg C, et al. The cell proliferation-associated antigen of antibody Ki-67: a very large, ubiquitous nuclear protein with numerous repeated elements, representing a new kind of cell cycle-maintaining proteins. *J Cell Biol* 1993;123:513–522.
- 174. Schulz A, Jundt G, Berghauser KH, et al. Immunohistochemical study of osteonectin in various types of osteosarcoma. *Am J Pathol* 1988;132:233–238.
- 175. Schurch W, Seemayer TA, Gabbiani G. The myofibroblast: a quarter century after its discovery. *Am J Surg Pathol* 1998;22: 141–147.
- 176. Schurch W, Skalli O, Lagace R, et al. Intermediate filament proteins and actin isoforms as markers for soft-tissue tumor differentiation and origin. III. Hemangiopericytomas and glomus tumors. *Am J Pathol* 1990;136:771–786.
- 177. Schurch W, Skalli O, Seemayer TA, Gabbiani G. Intermediate filament proteins and actin isoforms as markers for soft tissue tumor differentiation and origin. I Smooth muscle tumors. *Am J Pathol* 1987;128:91–103.
- 178. Seeger LL, Widoff BE, Bassett LW, et al. Preoperative evaluation of osteosarcoma: value of gadopentetate dimeglumine-enhanced MR imaging. Am J Roentgenol 1991;157:347–351.
- 179. Sepponen RE, Sipponen JT, Tanttu JI. A method for chemical shift imaging: demonstration of bone marrow involvement with proton chemical shift imaging. J Comput Assist Tomogr 1984;8:585–587.
- 180. Shaffer L, Tommerup N, eds. An international system for human cytogenetic nomenclature (2005). Basel: Karger, 2005.
- 181. Shibata Y, Fujita S, Takahashi H, et al. Assessment of decalcifying protocols for detection of specific RNA by non-radioactive in situ hybridization in calcified tissues. *Histochem Cell Biol* 2000;113:153–159.
- Shreve PD, Anzai Y, Whal RL. Pitfalls in oncologic diagnosis with FDG PET imaging: physiologic and benign variants. *RadioGraphics* 1999;19:61–77.
- 183. Shuman WP, Patten RM, Baron RL, et al. Comparison of STIR and spin-echo MR imaging at 1.5T in 45 suspected extremity tumors: lesion conspicuity and extent. *Radiology* 1991;179:247–252.
- Skalli O, Gabbiani G, Babai F, et al. Intermediate filament proteins and actin isoforms as markers for soft tissue tumor differentiation and origin. II. Rhabdomyosarcomas. *Am J Pathol* 1988; 130:515–531.

- 185. Skinner R. Decalcification of bone tissue. In: An Y, Martin K, eds. *Handbook of histological methods for bone and cartilage*. Totowa, NJ: Humana Press, 2003:167–184.
- 186. Som P, Atkins HL, Bandaypadhyay D, et al. A fluorinated glucose analog 2-fluoro-2-glucose (F18): nontoxic tracer for rapid tumor detection. *J Nucl Med* 1980;21:670–675.
- 187. Speicher MR, Gwyn Ballard S, Ward DC. Karyotyping human chromosomes by combinatorial multi-fluor FISH. *Nat Genet* 1996;12:368–375.
- 188. Stacy GS, Heck RK, Peabody TD, Dixon LB. Neoplastic and tumorlike lesions detected on MR imaging of the knee in patients with suspected internal derangement: Part I, intraosseous entities. *Am J Roentgenol* 2002;178:589–594.
- Steinbach LS. MRI of musculoskeletal tumors. Contemp Diagn Radiol 1989;12:1–6.
- 190. Sumiya H, Taki J, Higuchi T, et al. Nuclear imaging of bone tumors: thallium-201 scintigraphy. *Semin Musculoskel Radiol* 2001;5:177–182.
- 191. Sundaram M, McDonald DJ. Magnetic resonance imaging in the evaluation of the solitary tumor of bone. *Radiology* 1990;2: 697–702.
- 192. Sundaram M, McDonald DJ. The solitary tumor or tumor-like lesion of bone. *Top Magn Reson Imag* 1989;1:17–29.
- 193. Sundaram M, McGuire MH. Computed tomography or magnetic resonance imaging for evaluating the solitary tumor or tumor-like lesion of bone. *Skeletal Radiol* 1988;17:393–401.
- 194. Sundaram M, McGuire MH, Herbold DR, et al. Magnetic resonance imaging in planning limb-salvage surgery for primary malignant tumors of bone. *J Bone Joint Surg* 1986;68A: 809–819.
- 195. Sundaram M, McLeod R. MR imaging of tumor and tumorlike lesions of bone and soft tissue. *Am J Roentgenol* 1990;155:817–824.
- 196. Sweet DE, Madewell JE, Ragsdale BD. Radiologic and pathologic analysis of solitary bone lesions. Part III: Matrix patterns. *Radiol Clin North Am* 1981;19:785–814.
- 197. Szumowski J, Eisen JK, Vinitski S, et al. Hybrid methods of chemical shift-imaging. *Magn Reson Med* 1989;9:379–388.
- Szumowski J, Plewes D. Fat suppression in the time domain in fast MR imaging. J Magn Reson Med 1988;8:345–354.
- 199. Tehranzadeh J, Mnaymneh W, Ghavam C, et al. Comparison of CT and MR imaging in musculoskeletal neoplasms. *J Comput As*sist Tomogr 1989;13:466–472.
- 200. Thomas C. Sandritter histopathology. Textbook and color atlas, 8th ed. Toronto: BC Decker, 1989:305–309.
- 201. Triche TJ. Neuroblastoma and other childhood neural tumors: a review. *Pediatr Pathol* 1990;10:175–193.
- 202. Ueda T, Araki N, Mano M, et al. Frequent expression of smooth muscle markers in malignant fibrous histiocytoma of bone. *J Clin Pathol* 2002;55:853–858.
- 203. Vaher-Lavenu MC. Electron microscopy. In: Forest M, Tomeno B, Vanel D, eds. Orthopedic surgical pathology: diagnosis of tumors

and pseudotumoral lesions of bone and joints. Edinburgh: Churchill Livingstone, 1988:33-37.

- 204. Vandekerckhove J, Weber K. At least six different actins are expressed in a higher mammal: an analysis based on the amino acid sequence of the aminoterminal tryptic peptide. *J Mol Biol* 1978;126:783–802.
- 205. Vandekerckhove J, Weber K. The complete amino acid sequence of actins from bovine aorta, bovine heart, bovine fast skeletal muscle, and rabbit slow skeletal muscle. A proteinchemical analysis of muscle actin differentiation. *Differentiation* 1979;14:123–133.
- 206. Van der Wall H, Murray IP, Huckstep RL, et al. The role of thallium scintigraphy in excluding malignancy in bone. *Clin Nucl Med* 1993;18:551–557.
- 207. Vanel D, Verstraele KL, Shagero LG. Primary tumors of the musculoskeletal system. *Radiol Clin North Am* 1997;35:213–237.
- 208. Verstraete KL, De Deene Y, Roels H, et al. Benign and malignant musculoskeletal lesions: dynamic contrast-enhanced MR imaging—parametric first-pass images depict tissue vascularization and perfusion. *Radiology* 1994;192:835–843.
- 209. Volberg FM Jr, Whalen JP, Krook L, Winchester P. Lamellated periosteal reactions: a radiologic and histologic investigation. *Am J Roentgenol* 1977;128:85–87.
- Watt I. Radiology in the diagnosis and management of bone tumours. Review article. J Bone Joint Surg Br 1985;67B:520–529.
- Weinberg DS, Carey JL. Flow and imaging cytometry. In: Damjanov I, Linder J, eds. Anderson's pathology, 10th ed. St. Louis: CV Mosby, 1996:258–280.
- Wetzel LH, Levine E, Murphey MD. A comparison of MR imaging and CT in the evaluation of musculoskeletal masses. *Radio-Graphics* 1987;7:851–874.
- 213. Wiedenmann B, Franke WW. Identification and localization of synaptophysin, an integral membrane glycoprotein of Mr 38,000 characteristic of presynaptic vesicles. *Cell* 1985;41: 1017–1028.
- 214. Wiedenmann B, Franke WW, Kuhn C, et al. Synaptophysin: a marker protein for neuroendocrine cells and neoplasms. *Proc Natl Acad Sci U S A* 1986;83:3500–3504.
- Zhang H, Yoshikawa K, Tamura K, et al. Carbon-11-methionine positron emission tomography imaging of chordoma. *Skeletal Radiol* 2004;33:524–530.
- Zimmer WD, Berquist TH, McLeod RA, et al. Bone tumors: magnetic resonance imaging versus computed tomography. *Radiology* 1985;155:709–718.
- Zimmer WD, Berquist TH, Sim FH, et al. Magnetic resonance imaging of aneurysmal bone cyst. *Mayo Clin Proc* 1984;59: 633–636.
- 218. Zola H, Swart B, Boumsell L, Mason DY. Human Leucocyte Differentiation Antigen nomenclature: update on CD nomenclature. Report of IUIS/WHO Subcommittee. *J Immunol Methods* 2003;275:1–8.

2

Bone-Forming (Osteogenic) Lesions

BENIGN LESIONS 40 Osteoma 40 Enostosis (Bone Island) 51 Osteoid Osteoma 59 Osteoblastoma 74

MALIGNANT TUMORS 84

Osteosarcomas 84 Primary Osteosarcomas 89 A. Intraosseous Osteosarcomas 89 Intramedullary (Conventional) Osteosarcomas 89 Malignant Fibrous Histiocytoma-like (Fibrohistiocytic) Osteosarcoma 91 Giant Cell-Rich Osteosarcoma 99 Small Cell Osteosarcoma 99 Telangiectatic Osteosarcoma 102 Low-Grade (Well-Differentiated) Central Osteosarcoma 104 Gnathic Osteosarcoma 105 Multicentric (Multifocal) Osteosarcomas 108 Osteosarcomas with Unusual Clinical Presentation 108 B. Intracortical Osteosarcomas 111 C. Surface Osteosarcomas 111 Parosteal Osteosarcoma 113 Dedifferentiated Parosteal Osteosarcoma 114 Periosteal Osteosarcoma 114 High-Grade Surface Osteosarcoma 118 D. Soft Tissue (Extraskeletal) Osteosarcomas 118 Secondary Osteosarcomas 127 Paget Sarcoma 128 Osteosarcoma Arising in Fibrous Dysplasia 128 Osteosarcoma Arising in Bone Infarct 128 Postirradiation Osteosarcoma 128

Whether benign or malignant, bone-forming neoplasms are characterized by the formation of osteoid or mature bone directly by the tumor cells (27,28). Only one malignant tumor, osteosarcoma, is capable of doing this. The other bone-forming lesions are benign: osteoma, enostosis, osteoid osteoma, and osteoblastoma.

Benign Lesions

Osteoma

Osteoma is a benign, slow-growing lesion consisting of well-differentiated mature bone tissue with a predominantly lamellar structure, commonly arising in the frontal and ethmoid sinuses (approximately 75% of cases) (41). It may also occur on the surface of the outer table of the calvaria ("ivory exostosis"), the bones of the jaw (46), and, rarely, on the surface of the flat bones and the long and short tubular bones (2,30,32,36,49,50) (Fig. 2-1). Extracranial parosteal osteomas compose only 0.03% of biopsied primary bone lesions (34). Exceptionally, osteoma can exhibit endosteal extension (24).

Although not regarded as a neoplastic lesion by the World Health Organization (15), osteoma is included in this chapter because traditionally it has been regarded as a bone-forming lesion (5,13,16,34,52), and because of its importance in differential diagnosis of these lesions.

Clinical Presentation

Osteomas have been reported in patients from 10 to 79 years of age, with most in the fourth and fifth decades (13,46). Males and females are equally affected. The signs and symptoms associated with osteoma vary



Figure 2-1 Osteoma: skeletal sites of predilection, peak age range, and male-to-female ratio.

depending on the size of the lesion and its location (37). Small lesions are usually asymptomatic and incidental findings. Large paranasal sinus tumors in particular may produce a variety of manifestations. Attached to the sinus wall and covered by the sinus mucoperiosteum (38), they often block the nasal ducts, causing mucoceles, sinusitis, nasal discharge, headache, and pain. At times a patient may lose the sense of smell (38). Occasionally, paranasal osteomas exhibit more dramatic features such as orbital or intracranial invasion (41). Tumors near the orbit can produce exophthalmos, double vision, and pressure on the optic nerve that may result in loss of vision. At other sites, they can erode the wall of the cranial fossa, perforate the dura, and compress the frontal lobe. Paranasal osteomas are usually discovered in the third and fourth decades, and the finding is most often incidental. One retrospective study involving 16,000 patients found a frequency of 0.4% for these tumors (11). They occur twice as frequently in males as in females and are believed to have a growth peak at the time of maximum skeletal development; they are only occasionally found before puberty (47).

Osteomas seldom develop in the long and short tubular bones (1,5,20,49) and are extremely rare in the flat bones (7,33,48). Tumors at these sites are known as parosteal osteomas. Like osteomas elsewhere, they are usually asymptomatic. Most parosteal lesions measure 1 to 4 cm in diameter, rarely reaching the dimensions reported by Baum et al. $(16.5 \times 4.2 \text{ cm})$ and Mirra et al. $(6 \times 5 \times 5 \text{ cm})$ (1,33). Many long bone lesions are multiple.

It is worth noting the association of osteomas with the familial adenomatous polyposis syndrome (Gardner syndrome), an autosomal-dominant disorder with 25% new mutations, frequently seen in Mormons in Utah (10,18). It is caused by mutations in the APC gene, which maps to chromosome 5q21-q22 (3). In addition to bone lesions, the syndrome is marked by multiple cutaneous and subcutaneous lesions (e.g., sebaceous cysts, skin fibromas, and desmoid tumors) and intestinal, particularly colonic, polyposis (17). The bone tumors may precede the appearance of the intestinal polyps, which have a marked propensity to carcinomatous change (8,17). Osteomas have also been reported in association with tuberous sclerosis, an autosomal-dominant disorder affecting TSC1 (9q34) or TSC2 (16p13) gene, characterized by mental retardation, epilepsy, cutaneous hamartomas, and benign juvenile or inflammatory polyps of the colorectum (3,33). Finally, osteomas of the soft tissues (so-called osseous choristomas) are rare occurrences that have been described in the thigh, in the intraoral soft parts, and in the bronchi (12,29,42,45).

Imaging

Conventional radiography is the basis for the diagnosis of osteoma. Radiographs typically show a dense, ivory-like sclerotic mass with sharply demarcated borders attached to the bone (Fig. 2-2). Conventional tomography (Fig. 2-3) and computed tomography (CT) (Fig. 2-4) are effective in demonstrating lack of



Figure 2-2 Osteoma. A: Anteroposterior radiograph of the humerus shows sclerotic, ivory-like mass attached to the cortex. (Reprinted with permission from Greenspan A. Benign bone-forming lesions: osteoma, osteoid osteoma, and osteoblastoma. *Skeletal Radiol* 1993; 22:485–500.) **B:** A small osteoma of the proximal phalanx of the middle finger.





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Figure 2-3 Osteoma: conventional tomography. Conventional tomogram shows a large osteoma arising from the coronoid process of the right mandible. The tumor abuts the floor of the orbit and maxillary sinus, but there is no evidence of invasion in the adjacent structures. (Reprinted with permission from Greenspan A. Benign bone-forming lesions: osteoma, osteoid osteoma, and osteoblastoma. *Skeletal Radiol* 1993;22:485–500.)



Figure 2-4 Osteoma: computed tomography (CT). CT of the skull shows a small osteoma attached to the outer table (*arrow*).

cortical invasion (50), which not infrequently is a feature of parosteal osteosarcoma, or lack of cortical and medullar continuity with a host bone, a feature of osteochondroma (see "Differential Diagnosis," later). Magnetic resonance imaging (MRI) shows low signal intensity on both T1 and T2 sequences, consistent with cortical bone (50).

Histopathology

Histologically, two types of osteoma may be encountered: cancellous and compact (51). Most investigators consider the former to be the forerunner of the latter. The cancellous (or trabecular, or spongy) osteoma reveals a cancellous, trabecular architecture. Trabeculae are thin with fatty marrow present in the intertrabecular spaces (25) (Fig. 2-5A). They usually contain more woven bone and show evidence of active bone formation with transformation to lamellar bone (39). Woven bone consists of a more or less mature matrix with a plait of collagen fibers; it contains roundish osteocyte lacunae in high numbers per space unit. Lamellar bone is composed of narrow parallel layers of mature matrix with densely packed collagen fibers; it contains spindleshaped osteocyte lacunae in smaller numbers than woven bone. Compact osteoma, on the other hand, consists of dense, compact, mature lamellar bone (9) (Figs. 2-5B, C). No Haversian systems are present and only occasionally marrow spaces are visible. A compact osteoma usually exhibits empty osteocyte lacunae in varying numbers, probably as a result of increased distance between the small marrow spaces and the cells that interferes with cell nourishment by diffusion.



Figure 2-5 Histopathology of osteoma. A: Cancellous osteoma. Irregularly formed bone trabeculae consist mostly of lamellar bone (hematoxylin and eosin, original magnification $\times 25$). **B:** Cancellous osteoma. Intertrabecular spaces enclose highly vascular fibrous marrow tissue (hematoxylin and eosin, original magnification $\times 50$). **C:** Compact osteoma. Dense, compact, mostly lamellar bone surrounds narrow marrow spaces. The surface is rather smooth (van Gieson, whole-mount section). **D:** Compact osteoma. Some osteocyte lacunae are empty as a result of osteocyte decline due to inefficient nourishment (hematoxylin and eosin, original magnification $\times 200$).

Differential Diagnosis

Radiology

The radiologic differential diagnosis of solitary osteoma should include *parosteal osteosarcoma*, *sessile osteochondroma, juxtacortical myositis ossificans, periosteal osteoblastoma, ossified parosteal lipoma*, and focus of *melorheostosis* (21) (Fig. 2-6 and Table 2-1). Among these, *parosteal osteosarcoma* is the most important entity that needs to be excluded, and that may be a difficult task radiographically because both lesions appear as ivory-like masses attached to the bone's surface (23) (Fig. 2-7). The keys to recognizing osteoma, however, are its usually exquisitely smooth borders and well-circumscribed, intensely homogeneous sclerotic appearance on radiographs. Parosteal osteosarcoma, in contrast, may show a zone of decreased density at the periphery and usually appears less dense and homogeneous than osteoma (53).



Figure 2-6 Schematic representation of differential possibilities of similarly appearing juxtacortical and cortical lesions.

Condition (Lesions)	Radiologic Features	Pathologic Features
Parosteal osteoma	Ivory-like, homogenously dense sclerotic mass, with sharply demarcated borders, in- timately attached to cortex. No cleft be- tween lesion and adjacent cortex.	Mature lamellar bone (either consisting of concentric rings of compact bone, or parallel plates of cancellous bone), lack of active fibrous stroma.
Parosteal osteosarcoma	Ivory-like, frequently lobulated sclerotic mass, homo- or heterogeneous in density with more radiolucent areas at periphery. Incom- plete cleft between lesion and adjacent cor- tex occasionally present.	Streamers of woven to woven-lamellar bone with heavily collagenized stroma. Moderately cellular foci with nuclei exhibiting slight pleomorphism.
Sessile osteochondroma	Cortex of host bone merges without interruption with cortex of lesion and respective cancellous portions of adjacent bone and osteochondroma communicate.	Cartilaginous cap composed of hyaline cartilage arranged similarly to growth plate. Beneath zone of endochondral ossification with vascular invasion and replacement of calcified cartilage by newly formed bone. Intertrabecular spaces may contain fatty or hematopoietic marrow.
Juxtacortical myositis ossificans	Zonal phenomenon: radiolucent area in center of lesion and dense zone of mature ossifica- tion at periphery. Frequently thin radiolucent cleft separates ossific mass from adjacent cortex.	Trabecular bone and fibrous marrow. Histologic zonal phenomenon: immature bone in the center with proliferating osteoblasts, fibroblasts, and areas of hemorrhage and necrosis; mature bone at the periphery.
Periosteal osteoblastoma	Round or ovoid heterogenous in density mass attached to cortex.	Trabeculae of women bone, numerous dilated capillaries, exuberant in number osteoblasts, osteoclasts, and fibroblasts.
Ossified parosteal (periosteal) lipoma	Lobulated mass containing irregular ossifica- tions and radiolucent area of fat. Hyper- ostosis of adjacent cortex occasionally present.	Formation of mature bone within adipose tissue. Occasionally foci of necrosis and calcifications.
Melorheostosis (monostotic)	Cortical thickening resembling wax dripping down one side of a candle. Commonly ex- tends to the joint.	Thickened cortical bone containing irregularly arranged Haversian canals surrounded by cellular fibrous tissue. Osteoblastic activity usually present.

Table 2-1 Differential Diagnosis of Parosteal Osteoma



Figure 2-7 Parosteal osteosarcoma. A: Lateral radiograph shows ossific mass is attached to the posterior cortex of the distal femur. B: Computed tomography shows lack of invasion of bone marrow.



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Figure 2-8 Sessile osteochondroma. Radiographic hallmarks include uninterrupted cortical and medullary continuity between the lesion and the host bone (humerus).

Sessile osteochondroma can usually be identified by its characteristic radiographic features: the cortex of the lesion merges without interruption with the cortex of the host bone, and the cancellous portion is continuous with the host medullary cavity of the adjacent metaphysis or diaphysis (Fig. 2-8).

A well-matured focus of *juxtacortical myositis ossificans* may occasionally mimic osteoma. The radiographic hallmark of myositis ossificans is the so-called zonal phenomenon, characterized by a radiolucent area in the center of the lesion that indicates immature bone formation and a dense zone of mature ossification at the periphery. Often a thin radiolucent cleft separates the ossific mass from the adjacent cortex (Fig. 2-9A). Occasionally, however, a lesion (frequently, but not always, a mature one) may adhere to and fuse with the cortex, thus mimicking parosteal osteoma (Fig. 2-9B). In these instances CT may demonstrate the classic zonal phenomenon of the lesion (Fig. 2-9C).

Periosteal osteoblastoma, an extremely rare boneproducing lesion (31,44), appears radiographically as a round-to-ovoid juxtacortical mass (19). Its less intense and less homogeneous density (Fig. 2-10) distinguishes it from osteoma.

Ossified parosteal (or periosteal) lipoma is a rare lesion, which in the majority of cases measures 5 to 7 cm at its greatest dimension (4,14,40,41,54). Pain is an unusual symptom; most patients give a history of a slow-growing, painless mass that has been present for many years (26). Consistent with fatty tumors generally, lobulation is the chief radiographic feature of the mass, which contains low-density fat in contrast to the surrounding muscle tissue that it displaces (35) (Fig. 2-11).

Melorheostosis is a rare form of mixed sclerosing dysplasia that on radiography appears as a long segment of cortical thickening (flowing hyperostosis), often resembling wax dripping down one side of a candle (6,22). A typical focus of monostotic melorheostosis usually exhibits both parosteal and endosteal involvement and

the lesion commonly extends into the articular end of the bone (Fig. 2-12), features rarely present in a parosteal osteoma.

Pathology

The difficulty in distinguishing between parosteal osteosarcoma and osteoma may extend to the histopathology. Conventional parosteal osteosarcoma is characterized by streamers of bone that appear woven to woven-lamellar and are produced by low-grade atypical, sometimes polar, cells of a fibroblastic stroma (33). Moderately cellular foci of stroma are noted, and nuclei may exhibit slight pleomorphism. Some lesions may exhibit a fairly cellular stroma containing neoplastic osteoid and bone, which clearly indicates malignancy. It may extend to and invade the underlying bone marrow. The heavily collagenous stroma of parosteal osteosarcoma is the key feature that distinguishes it from parosteal osteoma (51) (Fig. 2-13). In contrast, the presence of mature bone and the absence of an active fibrous stroma are the distinctive features of parosteal osteoma (1).

The microscopic features of *sessile osteochondroma* are similar to its radiographic appearance and are likewise diagnostic, with merging of the cortices and imperceptible blending of the spongiosa of the lesion and the host bone (Fig. 2-14). In addition, the spongiosa of sessile osteochondroma frequently contains remnants of calcified cartilage that was not completely replaced by bone during the growth phase of the lesion (33), a finding not present in osteoma. A cartilaginous cap of variable thickness on the surface of the osteochondroma is a common finding. Because a sessile-type lesion may lose this cap by ossification in late adulthood, it may be confused with osteoma; however, the latter is always attached to the intact cortex.

The remaining entities that may create difficulties in the differential diagnosis for the radiologist can easily be excluded by histopathologic characteristics. For example, microscopic examination of *myositis ossificans circumscripta* reveals a correlative histologic zonal phenomenon. In contrast to osteoma, the lesion consists of trabecular bone and marrow fat, not solid cortical bone (Fig. 2-15).

Small trabeculae of woven bone, numerous dilated capillaries, and exuberant hyperplasia of osteoblasts, osteoclasts, and, occasionally, fibroblasts are characteristic histopathologic findings of *periosteal osteoblastoma* (34) (Fig. 2-16). A thin shell of newly formed periosteal bone may cover the lesion (43).

Histologically, *ossified parosteal lipoma* can easily be distinguished from osteoma because it usually exhibits the typical features of lipoma, composed of adipose lobulated tumor tissue with mature fat cells and formation of woven bone trabeculae, and, at later stages, formation of lamellar bone with or without foci of necrosis and calcification (43).

Also, *melorheostosis* can easily be distinguished histologically from an osteoma. The former is characterized by thickened trabeculae containing irregularly arranged Haversian canals surrounded by cellular *Text continues on page 51*



Figure 2-9 Juxtacortical myositis ossificans. A, B: Radiographic hallmarks of the lesion include zonal phenomenon: a radiolucent area in the center of the lesion, which indicates immature bone formation, and a dense zone of mature ossification at the periphery. Note also a thin, radiolucent cleft separating the lesion from the adjacent cortex. **C:** Focus of mature post-traumatic myositis ossificans is tightly adherent to the cortex of the left femur (*arrow*). The radiolucent cleft that usually separates the lesion from the cortex is absent. **D:** In another patient with a post-traumatic focus of myositis ossificans that was firmly attached to the cortex, computed tomography (CT) section shows classic zonal phenomenon.



Figure 2-10 Periosteal osteoblastoma. Lateral radiograph of the femur shows a juxtacortical lesion with radiolucent center and more dense periosteal shell (*arrow*).



Figure 2-11 Ossified parosteal lipoma. Large ossific mass is attached to the medial cortex of the right femur. The radiolucent character of the fatty tissue (distal part of the tumor) is apparent. (Reprinted with permission from Greenfield GB, Arrington JA. *Imaging of bone tumors.* Philadelphia: JB Lippincott, 1995, Fig. 4-19.)



Figure 2-12 Forme fruste of melorheostosis. Lateral radiograph of the elbow shows a flowing hyperostosis of the anterior cortex of the distal humerus. Note the extension of the lesion into the joint.

CHAPTER 2 Bone-Forming (Osteogenic) Lesions - 49



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Figure 2-13 Histopathology of parosteal osteosarcoma. A: A low-power photomicrograph shows the typical appearance of a heavily collagenized, fibrous matrix with irregular trabeculae of bone (hematoxylin and eosin, original magnification \times 50). B: A higher power photomicrograph shows the cellular fibrous stroma with islands of bone tissue bordered by spindle cells with minimal atypia (hematoxylin and eosin, original magnification \times 400). C: The fibrous stroma contains atypical elongated cells, some with hyperchromatic nuclei and discrete mitotic activity (hematoxylin and eosin, original magnification \times 400).

Figure 2-14 Histopathology of sessile osteochondroma. Whole-mount section of a sessile lesion shows a cartilaginous cap covered by perichondrium. Broad-based stalk is completely filled with fatty marrow (hematoxylin and eosin).





Figure 2-15 Histopathology of myositis ossificans. A: Low-power photomicrograph shows the fibrous cellular center (*darkly stained*) (zone 1) surrounded by layers of more mature bone (zone 2). At the periphery of the lesion (zone 3), mature bone formation is present (hematoxylin and eosin, original magnification $\times 25$). **B:** At slightly higher magnification, mature lamellar bone borders the connective tissue and muscle (*right*). Toward the center of the lesion (*left*), less mature bone, consisting mainly of woven bone, borders almost boneless fibrous tissue (van Gieson, original magnification $\times 50$).





Figure 2-17 Histopathology of melorheostosis. A: A low-power photomicrograph reveals extensive asymmetric sclerosis of the cortex, with encroachment on the endosteal surface, representing periosteal and endosteal bone formation (hematoxylin and eosin, original magnification \times 9). **B:** Higher magnification of the abnormal cortex reveals that the new bone is largely lamellar in architecture, reflecting slow modeling (hematoxylin and eosin, polarized light, original magnification \times 50). (Courtesy of Michael J. Klein, M.D., Birmingham, Alabama.)

fibrous tissue (Fig. 2-17). Osteoblastic activity is usually present (6). This is in contrast to the thin trabeculae of the cancellous osteoma and the dense lamellar bone of the compact variant of this lesion.

The radiologic and pathologic differential diagnosis of parosteal osteoma is depicted in Figure 2-18 and Table 2-1.

Enostosis (Bone Island)

Enostosis, known also as a bone island, is the endosteal compatriot of osteoma: a focus of cortical bone within cancellous (trabecular) bone (68). Although not in the true sense a "bone-forming" lesion, it is included in this chapter because it exhibits some morphologic features closely related to those of osteoma and some radiologic features similar to the other "true" bone-forming neoplasms (13,60).

Since its earliest descriptions in the literature, first by Stieda in 1905 and 7 years later by Fischer (68), enostosis has been variously named and defined. Stieda referred to the small, dense circumscribed shadows he observed inside the cancellous portion of short tubular bones and in the articular ends of long bones as "compact bone nuclei" (*kompakte Knochenkerne*). It was Fischer, however, who described these lesions as compact "islands" and emphasized their importance in differential diagnosis. Others have offered further names, including "calcified island in medullary bone" by Steel (68), "sclerotic bone island" by Meschan (72b), "focal sclerosis" by Caffey (59), and "end-osteoma" by Schmorl and Junghanns (77) (describing vertebral lesions). Definitions of this entity, as the names imply, have been similarly varied. Although some continue to classify enostosis among tumor-like conditions (58,60,62), Mirra (73) regards it as "misplaced, hamartomatous cortical bone." In addition, recent investigations suggest that foci of mature compact bone within the spongiosa, which are characteristic of this condition, represent areas that failed to resorb during endochondral ossification (68,69). Probably of developmental or congenital origin, enostosis therefore represents an anomaly that appears closely related to osteopoikilosis (68). The importance of recognizing this benign lesion, which can be virtually diagnosed on the basis of its characteristic clinical and radiologic features, lies in the need to differentiate it from clinically more significant bone lesions, such as primary or metastatic tumors, when it manifests itself uncharacteristically by being very large (66,73) and showing activity on skeletal scintigraphy (69).

В

Clinical Presentation

Typically asymptomatic, a bone island is often an incidental finding on radiography performed for another purpose. It is discovered more commonly in adults than in children and shows no sexual predilection. The



Figure 2-18 Radiologic and pathologic differential diagnosis of parosteal osteoma.

pelvis, the femur, and other long bones are preferential sites of involvement, although the lesion may be found anywhere in the skeleton, including the carpal and tarsal bones as well as the ribs (74). The spine is a rare site of involvement (57,77), accounting for only three (1.4%) of 209 bone islands reviewed by Onitsuka (75). These vertebral bone islands involved the thoracic and lumbar segments.

Imaging

Regardless of its site or size, a bone island exhibits a consistent radiographic picture. The lesion appears as an ovoid, round, or oblong focus, homogeneously dense and sclerotic, in the cancellous bone (spongiosa). It is commonly oriented with the long axis of the bone parallel to the cortex. Highly distinctive for this lesion are radiating bony streaks, referred to as "thorny radiation" (70,71) or "pseudopodia" (67), aligned with the long axis of the host bone's trabeculae, that merge with the trabeculae in a feathered or brush-like fashion (Figs. 2-19 and 2-20). In the great majority of cases, bone islands range in size from 1 mm to 2 cm (79). "Giant" bone islands, defined as lesions larger than 2 cm (65), have also been reported (Fig. 2-21). The largest lesion on record, described by Brien et al. (56) measured $10.5 \times 4.8 \times 4.0$ cm and was located in the proximal femur. Another notable giant bone island reported by Park et al. (76) measured $10.0 \times 1.7 \times 1.0$ cm, located in the tibia. An island reported by Gold et al. (65) measured $5 \times 5 \times 4.5$ cm, also located in the tibia. Similarly, a bone island reported by Smith (79) measured 3.5×4 cm and was located in the ilium, and by Greenspan and Klein (66) measured $4 \times 3 \times 2.5$ cm and was located in the distal femur. In addition to showing the same radiographic features as smaller lesions, giant bone islands may also give the "cumulus cloud" impression (73) (Fig. 2-22).

Bone islands usually do not show radiologic evidence of change in size over time. Nevertheless, several investigators have reported size changes in bone islands, some of which exhibited metabolic activity (55,64,71,72a,74,75). In the large series of patients who underwent multiple examinations described by Onitsuka (75), 44 of 138 bone islands (31.9%) showed changes in radiographic size over periods ranging from 3 to 23 years. After exclusion of 19 of these lesions because they had enlarged proportionally during adolescent growth, 21 of the remainder had increased in size and four had become smaller. Other evidence of growing bone islands is offered by Blank and Lieber (55), who reported six cases, by Ngan (74), who described



Figure 2-19 Enostosis (bone island). A: Anteroposterior radiograph of the right hip of a 10-year-old boy shows a bone island (*arrow*) in the femoral neck. (Reprinted from Greenspan A. *Orthopedic imaging*, 4th ed. Philadelphia: Lippincott Williams & Wilkins, 2004:954, Fig. 33.40.) **B:** Anteroposterior radiograph of the left hip in a 32-year-old woman shows a bone island (*arrow*) in the ilium.



Figure 2-20 Enostosis (bone island). A: Sclerotic lesion in the distal tibia shows distinctive radiating streaks (*arrows*), aligned with the trabeculae of the host bone. **B:** Radiograph of the femoral head specimen shows "thorny radiation" of the bone island, blending with the normal trabeculae of the host bone.





Figure 2-21 Giant bone island. A large sclerotic lesion occupies more than 50% of the middle phalanx of the small finger.

three cases and emphasized the possible confusion of such lesions with sclerotic metastases, and by Hoffman and Campbell (71), who reported the unstable appearance of a bone island. In the latter work the instability of a bone island was demonstrated by its disappearance and reappearance over a period of 6 years in a patient with hyperparathyroidism.

On CT a bone island appears as a low-attenuation focus (63), exhibiting (as on radiography) its characteristic brush borders (Fig. 2-23A). Occasionally the lesion's "pseudopodia" may show more rounded contours (Fig. 2-23-B). On all MRI pulse sequences a bone island exhibits the low signal intensity characteristics of cortical bone (Figs. 2-24 and 2-25).

On skeletal scintigraphy a distinctive feature of bone islands is that they usually exhibit no activity. This is generally ascribed to the fact that their level of metabolic activity is about the same as that of the surrounding cancellous bone. Therefore, radionuclide imaging has been and continues to be the means of differentiating bone islands from more aggressive bone lesions. Nevertheless, reports in the literature of histologically confirmed bone islands that showed increased radiotracer uptake on bone scan (61,65,67,69,78) have raised a note of caution about the usefulness of scintigraphy in distinguishing a bone island from clinically more significant lesions (Fig. 2-26).

The phenomenon of scintigraphically active bone islands has prompted a number of explanations of the pathomechanism, which is still unclear. It has been linked to the large size of reported bone islands (78) and to their growth, based on the observation that uptake of the radiopharmaceutical agent is roughly proportional to the volume of a lesion. However, two large bone islands that were undetectable on bone scans, as reported by Hall et al. (70), indicate that a lesion's size alone is not always a factor in this phenomenon. Others have pointed to a giant bone island's histologic morphology and increased metabolic activity relative to the surrounding cancellous bone as the probable explanation for its striking radionuclide activity (65). Because tracer uptake implies greater metabolic activity and, usually, increased blood flow, it is logical to assume that a bone island will show increased activity if it becomes metabolically active. However, reports of radiographically documented growing bone islands (one with a positive and the other with a negative bone scan) that showed no histologic evidence of metabolic activity raise doubts about this pathomechanism (55,72).

A contribution by Greenspan et al. (67,69), correlating the radiologic and pathologic findings in six cases of bone island, suggested that the increased tracer uptake observed in two bone islands appeared to be directly related to the higher degree of bone remodeling and osteoblastic activity exhibited by these lesions in comparison with the scintigraphically "cold" lesions. On histologic examination, these "hot" bone islands were marked by a mixture of compact and trabecular bone,



Figure 2-22 Giant bone island. Radiograph of the left ilium shows a large sclerotic lesion with the brush borders, assuming configuration of the "cumulus cloud."



Figure 2-23 Bone island: computed tomography (CT). A: Axial CT section through the knee joint shows a large lowattenuation lesion in the medial femoral condyle that exhibits characteristic brush borders. **B:** Axial CT section through the proximal tibia in another patient shows a sclerotic focus in the medial aspect of the bone with rounded "pseudopodia."



Figure 2-24 Bone island: magnetic resonance imaging (MRI). A: Anteroposterior radiograph of the right hip joint shows a typical bone island in the intertrochanteric area (*arrow*). **B:** On coronal spin-echo T1-weighted MRI the lesion exhibits low signal intensity.







Figure 2-25 Bone island: magnetic resonance imaging (MRI). **A:** Frontal radiograph of the right knee shows a large sclerotic lesion with "thorny radiation" (*arrows*). **B:** Coronal spin-echo T1-weighted MR image shows the bone island to be of low signal intensity. **C:** On coronal spin-echo T2-weighted sequence, the lesion continues to exhibit low signal intensity.



Figure 2-26 Bone island: scintigraphy. A: Anteroposterior radiograph of the right knee in a patient with breast carcinoma shows a sclerotic lesion in the proximal tibia (*arrow*). B: Skeletal scintigraphy shows increased uptake of radiotracer by the lesion, suspected to be a metastatic focus. Excision biopsy revealed a bone island.

with substantial amounts of woven bone. Vasculature was abundant, and osteoblastic activity with remodeling was significant. In contrast, the scintigraphically isoactive bone islands were composed solely of compact bone or mixed compact and trabecular bone, and showed negligible cellular activity and remodeling.

Histopathology

Α

Microscopic examination of a bone island reveals a histologic picture that correlates with the radiographic findings. Enostosis is a focus of compact (cortical) bone in the spongiosa that shows thorn-like, thickened trabeculae radiating from the lesion and merging with the surrounding trabeculae of the host bone (58) (Fig. 2-27). Higher power magnification reveals mature lamellar configuration encircling Haversian systems of nutrient canals (Fig. 2-28). Bone islands occasionally contain foci of woven nonlamellar bone and, as reported in some scintigraphically active bone islands, they may consist of a mixture of lamellar and woven bone (65,67). Foci of osteoblastic and osteoclastic activity are rare (Fig. 2-29).

Differential Diagnosis

Radiology

In addition to its inclusion in the differential diagnosis of *calcifying enchondroma* and *medullary bone infarct*, as well as healing *nonossifying fibroma*, which may occasionally mimic it, an enostosis should be considered in the differential diagnosis of sclerotic medullary lesions that carry far more significant clinical implications than bone island, including osteoid osteoma, osteoblastoma, osteosarcoma, and sclerotic metastases. Scintigraphic activity in a lesion such as an enostosis that is generally believed to be "cold" on bone scan raises the possibility that a benign entity will be misdiagnosed as a more serious abnormality. The possibility also exists, regardless of activity on radionuclide imaging, that a tumor might be misdiagnosed as a bone island (73). Despite the continuing usefulness of bone scintigraphy in distinguishing bone islands from more aggressive lesions, finding activity in an otherwise suspected bone island cannot be a valid factor in the differential considerations (67).

R

In general, the presumptive diagnosis of a bone island can easily be made if the individual clinical and radiologic findings, together with follow-up examinations, are taken into consideration (56,80). The guides to the correct diagnosis are found in the lesion's morphologic features as demonstrated on radiography, CT, and MRI, without reliance on scintigraphic findings. Thus, if a patient is found to have an asymptomatic isolated sclerotic bone lesion that exhibits the typical features of a bone island, with brush or feathered borders, the most likely diagnosis is an enostosis, whatever its size or its activity on bone scanning. Even the discovery of a sclerotic lesion in a patient with a



Figure 2-27 Bone island: histopathology. Slab radiograph **(A)** and low-power photomicrograph **(B)** of the femoral head show typical features of enostosis. Note characteristic thorn-like trabeculae radiating from the lesion. (From Bullough PG. *Atlas of orthopedic pathology with clinical and radiologic correlations,* 2nd ed. New York: Gower Medical Publishing, 1992, with permission.)



В



Figure 2-29 Bone island: histopathology. A: High-power photomicrograph (hematoxylin and eosin, original magnification $\times 230$) of the bone island (same patient as in Fig. 2-26) shows large vascular spaces and osteoblastic cells along the bone surfaces. **B:** In another patient, a high-power photomicrograph (hematoxylin and eosin, original magnification $\times 250$) shows osteoblastic activity and bone remodeling. **C:** Under polarized light the osteoblasts and parallel lamellae of mature bone surrounding the Haversian canal are well demonstrated.



R



known neoplasm would strongly suggest a bone island if this lesion's characteristic features are observed and the bone scan is normal. However, when a patient is symptomatic or the lesion appears "hot" on scintigraphy, misdiagnosis may occur in difficult cases. Such a circumstance demands careful observation with followup imaging studies. An open biopsy is in order if, as Mirra (73) suggested, the lesion's growth exceeds 25% of its diameter within 6 months or 50% within 1 year.

Pathology

The histologic differential diagnosis of enostosis rarely creates a serious problem, particularly when a pathologist is aware that the specimen has been obtained from the cancellous bone. Conversely, if only a minute fragment of bone island is submitted for examination, the microscopic picture may be indistinguishable from a *normal cortical bone*. Small biopsy specimen from the focus of *melorheostosis* may exhibit some histologic similarities to enostosis; however, correlation with radiographs clarifies the correct diagnosis. Likewise, the histologic differentiation of enostosis from *osteopoikilosis* (which in fact consists of multiple bone islands) must be based on imaging features.

The radiologic and pathologic differential diagnosis of enostosis is shown in Figure 2-30.

Osteoid Osteoma

Osteoid osteoma is a relatively common lesion of bone (4% of all primary bone tumors excluding myeloma, and 10% of all benign bone lesions), characterized by a nidus of osteoid tissue, that is usually less than 1 cm in diameter (93). The nidus, consisting of cellular, highly vascularized tissue that contains osteoid, can be entirely radiolucent or it may have a sclerotic center. In many instances the nidus is surrounded by a zone of reactive bone formation (98,114). Very rarely, an osteoid osteoma may present with more than one nidus. When two or more nidi are present within a single bone, the name multifocal osteoid osteoma is given, whereas multicentric osteoid osteoma refers to multiple nidi in different bones (107,134,138,143). Another variant of this



Figure 2-30 Radiologic and pathologic differential diagnosis of enostosis.

lesion has recently been reported, a "beaded" osteoid osteoma, which most likely represents transition between solitary and multifocal tumor (90).

Clinical Presentation

Patients with osteoid osteoma almost invariably present with a complaint of nocturnal pain that may be severe enough to cause awakening (96,103). This is apparently due to the presence of prostaglandins that are demonstrated in the nidus but not in the surrounding bone (106,127,153). Recently, cyclooxygenase 1 and 2 (Cox-1 and Cox-2), apparently the source of these prostaglandins, have also been detected in osteoid osteoma by immunohistochemical methods (116,130). Salicylates, such as aspirin, are remarkably effective in relieving the pain, usually in less than half an hour. This classical presentation is typical in greater than 75% of patients and provides an important clue to the diagnosis (148). Local swelling and point tenderness are present in some patients (81,85,137). There may be also neurologic signs and symptoms, which can include muscle atrophy, a decrease in deep tendon reflex responses, and variable degrees of sensory loss (149). In most cases, osteoid osteoma arises in the age range of 10 to 35 years (89,121). There is a marked predominance in males, with a number of studies reporting a male-to-female ratio of 2:1 to 4:1 (34,93,100, 113-115, 119-121) (Fig. 2-31).

Osteoid osteoma can arise in virtually any bone. The long bones are preferentially affected (approximately 65% of the lesions occur in the long bones), particularly the femur and tibia [over 53% of all osteoid osteomas are located in these bones with the proximal femur being most frequently affected (91)], in which the lesions usually occur near the end of the shaft. Less commonly affected sites are the phalanges of hands and feet (21%), the vertebrae (9%), the humerus (122), and the scapula (88).

Depending on its location in the particular part of the bone, the lesion can be classified as cortical, medullary (cancellous), or subperiosteal. Osteoid osteomas can be further subclassified as extra- or intracapsular (intraarticular) (98,121). Recently, Kayser et al. (117) hypothesized that many osteoid osteomas arising in the tubular bones may in fact originate in a subperiosteal site and later appear as the intracortical lesions.

Intraarticular osteoid osteomas preferentially affect the hip (149). However, lesions in the elbow, foot, wrist, knee, and vertebral facet joints have also been reported (101,105). These tumors are usually characterized by nonspecific symptoms that are indicative of an inflammatory synovitis (82,144). Sclerosis may be lacking and the periosteal reaction is absent. On physical examination, a joint effusion and synovitis may be identified (82,92). Intracapsular lesions may also cause a precocious appearance of arthritis. This can provide an important diagnostic clue in cases when the history is typical of osteoid osteoma but the nidus has not been confirmed by radiography (132). If a lesion is located near the growth plate, particularly in a young child, it may cause accelerated bone growth.

Osteoid osteoma involves the axial skeleton in 10% of cases (118,119,123,131,135). Osteoid osteoma of the spine most commonly involves the lumbar segment (59%). Less frequently affected are the cervical (27%), thoracic (12%), and sacral (2%) vertebrae (34,120,121, 142). In patients with lesions of the neural arch painful


Figure 2-31 Osteoid osteoma: skeletal sites of predilection, peak age range, and male-to-female ratio.

scoliosis often occurs (frequently as a result of a spasm), the concavity of the curvature being toward the side of the lesion (118).

Cytogenetic analysis of osteoid osteoma showed clonal chromosomal changes in one out of six studied cases (94). Two other investigated tumors revealed structural alterations of 22q13, a region containing genes that are involved in cell cycle regulation (86).

Imaging

The probability of identifying osteoid osteoma by conventional radiography is dependent on the location of the lesion. Cortical lesions are usually characterized by a radiolucent nidus (representing the tumor itself) with a surrounding area of reactive sclerosis. A periosteal reaction may or may not be identified (Fig. 2-32A). An intramedullary nidus is less easily identified on radiography because very little or no reactive sclerosis is present (Fig. 2-32B). Occasionally, the nidus may exhibit a focus of central calcification (sclerotic center). The nidus of a subperiosteal lesion may be visualized either as a central radiolucent or sclerotic focus, with or without reactive sclerosis (Fig. 2-32C), or as a shaggy, crescent-like focus of periosteal reaction (120). The radiographic presentation of intracapsular lesions is frequently characterized by periarticular osteoporosis and in some instances by premature osteoarthritis (Fig. 2-32D) (132).

In some instances conventional tomography is used to evaluate osteoid osteoma, with better visualization of the nidus than can be obtained with conventional radiography (Fig. 2-33). It can also assist in confirming or excluding the presence of a sinus tract and is therefore useful when the differential diagnosis includes a possible bone abscess.

Skeletal scintigraphy is a highly sensitive method for detecting the nidus of osteoid osteoma (125,139,152). This modality can be particularly helpful in cases for which the symptoms are atypical and the initial radiographs appear normal (146). A three-phase radionuclide bone scan, using technetium-99 as a scanning agent, has been suggested (111), which can be especially valuable when intraarticular or intramedullary lesions are not clearly visualized by conventional radiography. Radionuclide tracer activity can be observed on both immediate and delayed images (Fig. 2-34). Osteoid osteoma is occasionally characterized by an interesting scintigraphic feature, the so-called double-density sign. This phenomenon is believed to be related to increased vascularity of the nidus (110,111). In agreement with the appearance of the lesion on radiography, the double-density sign is seen as a small focus of increased activity associated with the nidus; this focus is surrounded by a larger area of less intense activity, related to the reactive sclerosis that surrounds the nidus (Fig. 2-35). Identification of the double-density sign can help to differentiate an osteoid osteoma from a bone abscess.

Ultrasound can be helpful in diagnosis of intraarticular lesions (97), and Doppler duplex color technique can be used to localize the nidus (102).

As the definite study for diagnosis of osteoid osteoma, CT is recommended. Not only can CT identify the lesion (151) but it can also accurately determine its *Text continues on page 65*



Figure 2-32 Osteoid osteoma. A: Cortical lesion. Radiolucent nidus is surrounded laterally by reactive sclerosis and medially by a solid periosteal reaction (*arrow*). **B:** Intramedullary lesion. Intramedullary nidus (*arrows*) shows no evidence of reactive sclerosis. **C:** Subperiosteal lesion. Sclerotic, subperiosteal nidus on the surface of talar bone (*arrow*) shows minimal periosteal reaction without reactive sclerosis. **D:** Intracapsular lesion. Faintly discernible radiolucent nidus is present in the left femoral neck (*arrow*) without evident perilesional sclerosis. Periarticular osteoporosis and early osteoarthritic changes (osteophytes) (*arrowheads*) are the presumptive features of osteoid osteoma in this 14-year-old boy.



Figure 2-32 *Continued* **E:** Intracapsular lesion. Anteroposterior radiograph of the right hip in a 28-year-old man reveals osteoid osteoma in the medial femoral neck (*arrow*). **F:** Tomography of the same hip clearly shows osteoarthritic changes: collar osteophyte (*arrowheads*) and slight narrowing of the weight-bearing segment of the hip joint.



Figure 2-33 Osteoid osteoma: conventional tomography. A: Trispiral tomogram reveals a radiolucent nidus of osteoid osteoma in the proximal humerus (*arrow*), which had not been apparent on conventional radiography. **B:** Trispiral tomogram of the thoracolumbar junction demonstrates a nidus of osteoid osteoma with perilesional sclerosis in the left pedicle of L1.

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Figure 2-34 Osteoid osteoma: scintigraphy. A: In the first phase of a three-phase radionuclide bone scan, 1 min after intravenous injection of 15 mCi (555 MBq) technetium-99m-labeled MDP, there is increased activity in the right iliac and femoral vessels. Discrete activity in the area of the medial femoral neck (*arrow*) is related to the nidus. **B:** In the third phase, 2 hours after injection, there is accumulation of a bone-seeking tracer in the right femoral neck lesion (*arrow*). (Reprinted with permission from Greenspan A. Benign bone forming lesions: osteoma, osteoid osteoma, and osteoblastoma. *Skeletal Radiol* 1993;22:490.)



Figure 2-35 Osteoid osteoma: scintigraphy. Scintigraphy of tibial osteoid osteoma shows a characteristic "double-density" sign (*arrow*): increased uptake in the center (related to the nidus of osteoid osteoma) and a less intensely active area at the periphery (denoting reactive sclerosis).

extent, thus enabling exact measurement of the size and location of the nidus (103,105,112, 140,141) (Fig. 2-36). The nidus usually appears as a well-defined area of low attenuation, which is surrounded by a variably sized region of high attenuation reactive sclerosis (Fig. 2-37). To delineate the lesion, thin (preferably 2-mm) contiguous sections are most appropriate (Fig. 2-38). CT is particularly useful to define lesions that arise in the axial skeleton, whose complex anatomy is less clear on routine radiography and conventional tomography (145) (Fig. 2-39).

The suitability of MRI for detection of osteoid osteoma remains unclear and published reports have had mixed results (99,109,150,155,157). Goldman et al. (105) reported on four cases of intracapsular osteoid osteoma of the femoral neck, in which the lesions were evaluated with bone scintigraphy, CT, and MRI. Although in all cases abnormal findings were apparent in the MR images, the nidi could not be identified prospectively. On the basis of MRI findings of secondary bone marrow edema or synovitis, several incorrect diagnoses were made, which included Ewing sarcoma, osteonecrosis, stress fracture, and juvenile arthritis. In these cases, it is noteworthy that the correct diagnoses were made only after review of the radiographs and thin-section CT studies. Another report by Woods et al. (154) involved three patients with a highly unusual association of osteoid osteoma with a reactive soft tissue mass. In these cases, MRI studies might have led to confusion of osteoid osteoma with osteomyelitis or a malignant tumor. Moreover, in each case the nidus displayed different signal characteristics. In one case the intensity of signal was generally low on all pulse sequences but mild enhancement was seen after administration of gadolinium. In another case, the signal was of intermediate intensity and administration of gadolinium revealed inhomogeneous enhancement of the nidus. For the third case in which plain films showed the nidus to be intracortical, MRI could not identify the nidus distinctly. More recently, Davies et al. analyzing the MR imaging findings in 43 patients with osteoid osteoma concluded that reliance on MRI alone may lead to misdiagnosis (95). They suggested additional application of skeletal scintigraphy and CT in unclear cases, particularly when unexplained areas of bone marrow edema are encountered.

However, some reports do suggest the effectiveness of MRI for demonstrating the nidus of osteoid osteoma (84,104,156) (Figs. 2-40 and 2-41). Bell et al. (87) clearly demonstrated an intracortical nidus on MRI that had not been seen on scintigraphy, angiography, or CT scans. The nidus exhibits high signal intensity on T2-weighted sequences, most likely due to abnormal Cox-1 or Cox-2 levels (116). Recently, Liu et al. (126) advocated imaging of osteoid osteoma with dynamic gadolinium-enhanced MRI for greater conspicuity.

Histopathology

In lesions of osteoid osteoma, the nidus is small, well circumscribed, and self-limited. It is composed of osteoid tissue or mineralized immature woven bone (Figs. 2-42A–C). The osteoid matrix and bone form small and irregular trabeculae, with a thickness ranging from thin and delicate to broad and sclerotic (120). The trabeculae are surrounded by a highly vascular fibrous stroma that exhibits prominent osteoblastic and osteoclastic activity (Fig. 2-42D). The sclerosis surrounding the lesion is composed of dense bone that displays a variety of maturation patterns. From the ultrastructural viewpoint, the morphology of osteoblasts of osteoid osteoma is



Figure 2-36 Osteoid osteoma: computed tomography (CT). A: Anteroposterior radiograph of the left elbow of a 31-yearold man with the typical clinical symptoms of osteoid osteoma shows periarticular osteoporosis. There is the suggestion of a lesion in the capitellum. B: Sagittal CT section unequivocally demonstrates a subarticular nidus, measuring 6.5 mm.



Figure 2-37 Osteoid osteoma: computed tomography (CT). Axial (A) and reformatted sagittal (B) CT sections clearly demonstrate the nidus (*arrow*) in the anterior cortex of the tibia in a 16-year-old boy. Note the significant amount of perilesional reactive sclerosis.



Figure 2-38 Osteoid osteoma: computed tomography (CT). A: Osteoid osteoma in the medial femoral cortex in an 18-year-old woman (*arrow*). **B:** Two-millimeter-thin CT sections through the lesion were obtained. **C:** Low-attenuation nidus of osteoid osteoma is surrounded by a high-attenuation zone of reactive sclerosis.



Figure 2-39 Osteoid osteoma: computed tomography (CT). A: Radiograph of the cervical spine does not clearly demonstrate the suspected lesion of osteoid osteoma. B: CT section through C6 shows a low-attenuation nidus with a sclerotic center, located in the left pedicle and extending into the lamina. (Reprinted with permission from Greenspan A. Benign bone forming lesions: osteoma, osteoid osteoma, and osteoblastoma. *Skeletal Radiol* 1993;22:492.)



Figure 2-40 Osteoid osteoma: magnetic resonance imaging (MRI). A: Conventional radiograph shows a sclerotic area localized to the medial aspect of the proximal femoral shaft. The nidus is not apparent. **B:** Axial T1-weighted MRI clearly demonstrates the high-intensity nidus (*arrow*) within a low-intensity sclerotic cortex. (Courtesy of Lynne S. Steinbach, M.D., San Francisco, California. Reprinted with permission from Greenspan A. Benign bone forming lesions: osteoma, osteoid osteoma, and osteoblastoma. *Skeletal Radiol* 1993;22:492.)



Figure 2-41 Osteoid osteoma: magnetic resonance imaging (MRI). A: Coronal T1-weighted (SE; TR 600, TE 20) MRI shows an osteoid osteoma in the lateral aspect of the neck of the left femur. (Reprinted with permission from Stoller DW. *Magnetic resonance imaging in orthopedics and sports medicine,* 2nd ed. Philadelphia: JB Lippincott, 1993:1234.) **B:** Coronal T1-weighted (SE; TR 600, TE 20) MRI shows an osteoid osteoma in the medial cortex of the left tibia. (Reprinted with permission from Stoller SW. *Magnetic resonance imaging in orthopedics and sports medicine,* 2nd ed. Philadelphia: JB Lippincott, 1993:1234.) **B:** Coronal T1-weighted (SE; TR 600, TE 20) MRI shows an osteoid osteoma in the medial cortex of the left tibia. (Reprinted with permission from Stoller SW. *Magnetic resonance imaging in orthopedics and sports medicine,* 2nd ed. Philadelphia: JB Lippincott, 1993: 1236.)

generally similar to that of normal osteoblasts. However, these osteoblasts possess irregular, indented nuclei (indicative of high metabolic activity), glycogen particles, an abundance of fine intracytoplasmic fibers, and occasional iron-containing lysosomes (147). Moreover, some of these osteoblasts possess atypical mitochondria with a lobulated or "honeycomb" appearance. The areas of mineralized matrix exhibit the morphology of coarse woven bone. In two of the cases reported by Steiner (147), the osteoid contained, in addition to collagen, fine granular material, probably representing polysaccharides. Applying immunohistochemistry, nerve fibers positive for S-100, PGP 9.5 and/or neurofilament could be demonstrated in the fibrous zone and also in the nidus of osteoid osteoma, but not in other tumors including osteoblastoma (108,133). This finding in combination with prostaglandin and Cox-1/Cox-2 synthesis may explain the almost pathognomonic clinical presentation of this lesion.

Differential Diagnosis

Radiology

Even when a cortical lesion exhibits the classic radiographic appearance of osteoid osteoma, the differential diagnosis must consider an *osteoblastoma* (159,176), a *cortical stress fracture*, a focus of *infection* (124), and *intracortical osteosarcoma* (123). Only exceptionally osteoid osteoma may simulate an osteocartilaginous exostosis (128). The main differential factor to be considered in distinguishing osteoid osteoma from osteoblastoma is the size of the lesion. Generally, osteoblastomas are larger, exceeding a diameter of 2 cm (see discussion under heading "Osteoblastoma"). Periosteal reaction may be more prominent than encountered in osteoid osteomas. In stress fracture, the radiolucency is usually more linear than that of osteoid osteoma, running perpendicular to or at an angle to the cortex rather than parallel to it (Fig. 2-43). A cortical bone abscess may have a similar radiographic appearance to that of osteoid osteoma, but it can usually be differentiated by a linear, serpentine tract that extends away from the abscess cavity (Fig. 2-44). In some instances, intracortical osteoid osteoma must be differentiated from intracortical osteosarcoma, a rare boneforming malignancy that arises solely within the cortex of bone and grossly involves neither the medullary cavity nor the soft tissues (123,129,136). On radiography, the latter tumor appears as a radiolucent focus within the cortex (femur or tibia), surrounded by zone of sclerosis. Occasionally, fluffy densities can be demonstrated in the radiolucent zone (129). The cortex at the site of the lesion may bulge slightly or may be thickened. The reported size of the lesion varies from 1.0 to 4.2 cm (83).

In intramedullary lesions, the differential must again consider an *osteoblastoma*, a *bone abscess* (Brodie abscess) and, in a lesion with calcified nidus, a *bone island*



D

B

Figure 2-42 Histopathology of osteoid osteoma. A: Low-power magnification shows a well-demarcated nidus composed of osteoid and trabeculae of immature woven bone (*red, center*), surrounded by a narrow, cell-rich outer zone (*blue*). At the periphery of the nidus, bone shows partial sclerosis (*left half*) (hematoxylin and eosin, original magnification \times 6). **B:** Low-power magnification shows the nidus to be composed of irregular woven bone trabeculae with dense osteoblastic rimming. From the left and right the nidus is clearly delineated by the adjacent bone (hematoxylin and eosin, original magnification \times 50). **C:** At higher magnification, osteoblastic rimming of woven bone trabeculae is conspicuous. In addition, some osteoclasts are also present (hematoxylin and eosin, original magnification \times 100). **D:** High-power photomicrograph of the nidus shows interconnecting small trabeculae distributed within a fibrovascular stroma. Note the prominent osteoblastic activity (hematoxylin and eosin, original magnification \times 400). **E:** S-100 positive (*dark brown, top and center*) small peripheral nerves in the tan-stained stroma of the nidus. Trabeculae appear light blue because of immunohistochemistry methylene-blue counterstaining (immunohistochemistry, ABC method, original magnification \times 200).

С

Ε



Figure 2-43 Cortical stress fracture. Lateral radiograph of the tibia shows the perpendicular direction of radiolucency to the long axis of the tibial cortex (*arrow*).

(enostosis). Osteoblastoma, as mentioned previously, is a

larger lesion than osteoid osteoma, and, in addition,

exhibits considerably less peritumoral reactive sclerosis.

A *medullary bone abscess* exhibits a central radiolucency

very similar to that of osteoid osteoma, but, similarly to

the cortical focus of infection, it can usually be

Figure 2-44 Cortical bone abscess. Lateral tomogram of the tibia shows a radiolucent, serpentine tract (*arrow*) that was originally misdiagnosed as osteoid osteoma.

differentiated by the serpentine tract that extends from the abscess cavity toward the nearest growth plate (Figs. 2-45 and 2-46). A *bone island* is characterized on radiography by the lesion's brush-like borders, which blend with surrounding trabeculae in a pattern likened to "thorny radiation" or "pseudopodia" (67,68) (Fig. 2-47;



Figure 2-46 Epiphyseal bone abscess. A: A 7-year-old boy experienced intermittent pain in the left knee for 3 weeks; the pain was worse at night and was promptly relieved by salicylate. On the basis of clinical history and radiography showing a round radiolucency in the epiphysis with mild perilesional sclerosis, osteoid osteoma was considered in the differential diagnosis. **B:** Computed tomography (CT) section through the lesion reveals cortical disruption and the serpentine configuration of the radiolucent tract that extends into the articular cartilage. This is characteristic of bone abscess.



Figure 2-47 Enostosis (bone island). Note characteristic features of this lesion (*arrows*): a homogeneously dense, sclerotic focus in the cancellous bone with distinctive radiating bony streaks ("thorny radiation") that blend with the trabeculae of the host bone, creating a feathered, or brush-like, border.

see also Figs. 2-19 and 2-20). In addition, bone islands usually exhibit no increased activity on radionuclide bone scan (69) (Fig. 2-48).

A schematic representation of differential possibilities of cortical and medullary osteoid osteoma is shown in Figure 2-49.

Pathology

Histopathologic differential diagnosis of osteoid osteoma should include primarily *osteoblastoma* and *osteosarcoma* because the histologic appearances of *stress fractures, intracortical osteomyelitis, Brodie abscesses,* and *bone islands* are unmistakably characteristic and do not create identification problems (Table 2-2).

Probably the most difficult task is to differentiate osteoid osteoma from osteoblastoma. In fact, some authorities (44) consider these entities to represent the same lesion in different stages of development because they show such remarkable histologic similarities. On the other hand, some investigators contend that osteoid production is usually greater in osteoblastoma than in osteoid osteoma and that the former lesion shows greater vascularity. Whereas in osteoid osteoma there seems to be a more organized structure with maturation of the nidus toward its periphery, in osteoblastoma the distribution of osteoid and trabecular bone has a less organized pattern: the whole lesion may be in the same stage of development (43). The difference in the size of the nidus (1.0 cm or less for osteoid osteoma and greater than 2 cm for osteoblastoma) may be an artificial division because osteoblastoma in the early stage of development has to be smaller than 2 cm. In the spine that division may be even more artificial, because the lesions of osteoblastoma are often smaller than 2 cm. The presence of more or less marked perilesional sclerosis in osteoid osteoma may allow this rather arbitrary differentiation.



Figure 2-48 Enostosis (bone island). A 32-year-old woman presented with nocturnal pain localized in the wrist area. **A:** Dorsovolar radiograph of the wrist shows a dense, round lesion in the scaphoid (*arrow*), and a diagnosis of osteoid osteoma was considered. **B:** Bone scan reveals normal uptake of the radiopharmaceutical tracer, ruling out osteoid osteoma, which is invariably associated with increased isotope uptake. The lesion proved to be a bone island. The pain was related not to the bone lesion but to tenosynovitis.

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Figure 2-49 Schematic representation of differential possibilities of (A) cortical and (B) medullary osteoid osteoma.

Condition (Lesions)	Radiologic Features	Pathologic Features
Cortical osteoid osteoma	Radiolucent nidus, round or elliptical, surrounded by radiodense reactive sclerosis. Solid or laminated (but not interrupted) periosteal reaction. Scintigraphy invariably shows increased uptake of radiotracer. "Double-density" sign.	Nidus composed of osteoid tissue or mineralized immature bone. Osteoid matrix and bone form irregular islets and are surrounded by a richly vascular fibrous stroma. The islets have a trabecular structure, whose thickness ranges from thin and delicate to broad and sclerotic. The stroma shows prominent osteoblastic and osteoclastic activity. Perilesional sclerosis composed of dense bone exhibiting various maturation patterns.
Medullary osteoid osteoma	Radiolucent (or with central calcification) nidus, without or with only minimal perinidal sclerosis. Usually no or only minimal periosteal reaction. Scintigraphy—as above.	
Subperiosteal osteoid osteoma	Central radiolucent or sclerotic nidus with or without reactive sclerosis. Occasionally shaggy, crescent-like focus of periosteal reaction. Scintigraphy—increased uptake of radiotracer.	As above.
Intracapsular (periarticular) osteoid osteoma	Periarticular osteoporosis. Premature onset of osteoarthritis. Nidus may or may not be visualized. Scintigraphy— as above.	
Osteoblastoma Stress fracture (cortical)	See Table 2-3. Linear radiolucency runs perpendicular or at an angle to the cortex. Scintigraphy—increased uptake of radiotracer.	See Table 2-3. Features of bone repair: osteoid and cartilaginous callus, osteoblastic and osteoclastic activity.
Bone abscess (Brodie)	Irregular in outline radiolucency, usually with a sclerotic rim, associated with serpentine, linear tract. Predilection for metaphysis and the ends of tubular bones. Scintigraphy—increased uptake of radiotracer. MRI—on T1 WI a well- defined low-to-intermediate-signal lesion outlined by a low-intensity rim. On T2 WI a very bright homogenous signal, outlined by a low-signal rim.	Necrotic tissue, giant cells, granulocytes, lymphocytes, plasma cells, and histiocytes.
Bone island (enostosis)	Homogeneously dense, sclerotic focus in cancellous bone with distinctive radiating streaks (thorny radiation) that blend with the trabeculae of the host bone. Scintigraphy—usually no increased uptake. MRI—low-intensity signal on T1 and T2 WI.	Focus of mature, compact bone with thickened peripheral trabeculae that blend with trabeculae of the spongiosa. Wide bands of parallel or concentric lamellae; marrow spaces resembling Haversian canals.
Intracortical osteosarcoma	Intracortical radiolucent focus surrounded by zone of sclerosis. Occasionally central "fluffy" densities. Cortex thickened or bulged. Scintigraphy— increased uptake of radiotracer.	Consistent with an osteoblastic osteosarcoma with focal evidence of chondroid and fibroblastic differentiation. Permeation of Haversian systems. "Trapping" of lamellar bone within the tumor.

Table 2-2 Differential Diagnosis of Osteoid Osteoma



Figure 2-50 Histopathology of bone island. A: Low-power photomicrograph demonstrates "pseudopodia" of the lesion. Note the multiple marrow spaces in the center, very similar to the Haversian systems of nutrient canals (hematoxylin and eosin, original magnification $\times 16$). **B:** A high-power photomicrograph reveals compact bone with wide bands of parallel lamellae and concentric rings (hematoxylin and eosin, original magnification $\times 90$).

However, genetic analysis revealed chromosomal alterations involving chromosome 22 in osteoid osteoma that were different from alterations observed in osteoblastomas (86,193).

Histologic differentiation of osteoid osteoma from *osteosarcoma* (mainly from the intracortical variant) is less challenging. The latter tumor is rich in osteoid and woven bone, with minimal anaplasia due to so-called normalization of nuclei. Trapping of host cortical lamellar bone within the tumor, however, is an indicator of malignancy (129).

Differentiation of osteoid osteoma from a *bone island* is not difficult. The histopathologic features of bone island are characteristic: a dense, sclerotic focus of compact bone within the cancellous bone exhibits thickened peripheral trabeculae blending with the trabeculae of the spongiosa. In addition, the lesion shows parallel or concentric bands of mature compact lamellar bone, and marrow spaces having the appearance of Haversian canals (67,68) (Fig. 2-50; see also Fig. 2-28).

The radiologic and pathologic differential diagnosis of osteoid osteoma is depicted in Figure 2-51 and Table 2-2.

Osteoblastoma

Α

Osteoblastoma accounts for approximately 1% of all primary bone tumors and 3% of all benign bone lesions (176, 177). Jaffe and Mayer are credited with identifying it as an entity in 1932 (178). Later it was described as "giant osteoid osteoma," underscoring its close histologic resemblance to osteoid osteoma and its larger size (167). The lesion is often more than 2 cm in diameter (177). Lichtenstein termed it an "osteogenic fibroma of bone," subsequently changing the name to "benign osteoblastoma" (30,31,182). The histologic similarities between osteoid osteoma and osteoblastoma are striking, and differentiation is often

difficult if not impossible (163). Some authors subscribe to the concept that they represent different clinical expressions of the same pathologic process (44,170). In most of the world literature, however, osteoid osteoma and osteoblastoma are considered separate and distinct clinical entities, mainly because of their different clinical presentations and different radiologic characteristics. Their natural histories also differ: whereas osteoid osteoma tends toward regression, osteoblastoma tends toward progression and even malignant transformation (113) (although the possibility of the latter event remains controversial). Genetic findings are also in favor of separating these lesions (94,192).

Clinical Presentation

Like osteoid osteoma, osteoblastomas are most often found in patients in the first to third decades with the peak incidence in the second decade (184); "aggressive" tumors are usually seen in an older age group (average age 33 years) (180,187). The male-to-female ratio appears to be approximately 2:1 (122,175). The long tubular bones are frequently affected, but the lesion shows a distinct predilection for the axial skeleton in general and the vertebral column in particular (13,34,164,180,181) (Fig. 2-52). Approximately 30% to 40% are localized in the spine (131), more or less equally in the cervical, thoracic, and lumbar segments, mainly in the posterior elements including the arch and spinous processes (186). In one series of 123 tumors, 39 (32%) were located in the spine and 23 (19%)in the jaw; the remaining 61 tumors were distributed throughout the appendicular skeleton with a predilection for the femur and tibia (187). Periosteal osteoblastomas have occasionally been reported (190), but multifocal lesions are quite rare (158).

The clinical presentation of osteoblastoma is different from that of osteoid osteoma (159,176). Some pa-



Figure 2-51 A: Differential diagnosis of intracortical osteoid osteoma. B: Differential diagnosis of medullary osteoid osteoma.

tients are asymptomatic, and the lesion is found incidentally (183). Symptomatic patients in one study most often experienced a dull, localized pain, with a median duration of symptoms of 6 months before they presented for treatment. The pain very rarely interfered with sleep. Although the use of salicylates for pain relief is not mentioned in the study by Kroon and Schurmans (180), the pain of osteoblastoma does not appear to be as readily relieved by salicylates as the pain in osteoid osteoma. Tenderness at the tumor site is a consistent physical finding (186), and swelling may be present, particularly when the lesion is near the surface (180).



Figure 2-52 Osteoblastoma: skeletal sites of predilection, peak age range, and male-to-female ratio.

Patients with lumbar vertebral lesions will very commonly present with scoliosis. Scoliosis also commonly occurs in patients whose tumors arose in the ribs (169,172). Spinal lesions in particular may cause neurologic deficits ranging from muscle weakness to paraplegia.

Toxic osteoblastoma, a rare variant of this tumor, has recently been recognized (166). It is associated with systemic manifestations, including diffuse periostitis of multiple bones, fever, and weight loss (191).

Osteoblastomas rarely extend into the soft tissues and do not metastasize, although they do tend to recur locally.

Imaging

Conventional radiography is the basis for the diagnosis of osteoblastoma. In a review of the spectrum of osteoblastoma in 123 patients by McLeod et al. (187), most tumors (75%) were either spherical or slightly oval, with the remainder being longitudinal; the tumor margin was well defined (83%). In half of the appendicular tumors, there was considerable reactive sclerosis; 13% of these tumors, however, had only a sclerotic rim, and 37% showed no surrounding sclerosis. The interior of the lesions was most often radiolucent (64%), whereas various degrees of ossification were present in the remaining cases. The authors speculated that lesions tend to be radiolucent when they are younger, ossifying as they mature. In the great majority of tumors, the cortex was either perfectly normal or intact, although expanded and thinned (75%); the cortical expansion was usually eccentric. Destruction or penetration of the cortex suggesting malignancy was present in a minority (20%) of tumors. The great majority of osteoblastomas showed periosteal new bone formation, which in some cases was extremely prominent. The periosteal reaction was predominantly benign and of the solid type (86%), whereas the remainder showed spiculation or multilamination, suggesting a more aggressive lesion.

In a review of the range of manifestations of osteoblastoma by Marsh et al. (186), the majority of tumors showed a shell of reactive bone separating the tumor from the surrounding normal bone. Osteoblastomas of the spongy bone of the spine, ilium, or talus exhibited less prominent reactive bone formation, and those arising in the cortex more pronounced reactive bone, than is seen in osteoid osteoma.

Four distinctive types of osteoblastomas can be identified, based on the radiographic findings (23). (a) The lesion may be almost identical to osteoid osteoma, although much larger (usually >2 cm in diameter). This type occasionally exhibits less reactive sclerosis than osteoid osteoma, but the periosteal reaction is more prominent. (Fig. 2-53). (b) Osteoblastoma may also appear as a blow-out expansive lesion similar to an aneurysmal bone cyst, with small radiopacities in the center. This pattern is particularly common in tumors involving the spine (Fig. 2-54). (c) The tumor may occasionally appear as an aggressive lesion, simulating a malignant neoplasm (Fig. 2-55). Many of the so-called aggressive osteoblas-



Figure 2-53 Types of osteoblastoma: giant osteoid osteoma. The osteoblastoma of the proximal humerus in this 8-year-old boy appears similar to osteoid osteoma. However, the lesion is larger (2.5 cm in its largest dimension), and there is a more pronounced periosteal response.

tomas belong to this group. (d) Exceptionally rare, osteoblastoma is juxtacortical (periosteal) in location, as described in 6 of 42 tumors reported by Schajowicz and Lemos (44) and in 2 of 20 tumors reported by Lichtenstein and Sawyer (31). These lesions generally lacked perifocal bone sclerosis and in addition, a thin shell of newly formed periosteal bone was noted covering the lesion (19,34,44) (Fig. 2-56; see also Fig. 2-10).

Conventional tomography, although nowadays rarely used, may be valuable for determining the extent of the tumor and whether and to what degree intralesional bone formation has occurred. It is particularly useful for spinal lesions, which may be difficult to interpret on routine radiographs. Tomography may also help gauge the size of the nidus, thus facilitating the distinction from osteoid osteoma (Fig. 2-57).

Skeletal scintigraphy invariably demonstrates intense focal accumulation of bone-seeking radiopharmaceutical agents (Fig. 2-58). Angiography is infrequently performed as part of the evaluation of osteoblastoma





Figure 2-54 Types of osteoblastoma: blow-out expansion. A: Expansive blow-out osteoblastoma with several small central opacities in the lamina of C6 (*arrows*). **B:** Large osteoblastoma originating in the lamina and involving the pedicle, transverse process, and half of the vertebral body of L5 (*arrows*) shows aneurysmal-like expansion with central radiopacities. (**B** reprinted with permission from Greenspan A. Benign bone-forming lesions. *Skeletal Radiol* 1993;22:494.)

because of its questionable importance as a preoperative assessment tool (180).

CT, as in osteoid osteoma, is the most important imaging modality for facilitating the diagnosis of osteoblastoma and demonstrating its exact size and location (174) (Fig. 2-59). In addition, areas of calcifications and ossifications within the lesion (Fig. 2-60), as well as cortical destruction and soft tissue extension, are well delineated on CT sections (180). CT is particularly helpful in working up lesions of the spine (159) (Fig. 2-61).

Very few reports on the usefulness of MRI in the diagnosis and evaluation of osteoblastoma can be found in the literature (180). The strength of MRI would appear to be similar to that of CT, namely, defining the extent of bone and soft tissue involvement. Most osteoblastomas demonstrate signal intensity patterns on MR scans



Figure 2-55 Types of osteoblastoma: aggressive. Dorsovolar (A) and lateral (B) radiographs of the right hand show aggressive osteoblastoma. Note the destruction of the entire fourth metacarpal with massive bone formation. Although very similar in appearance to osteosarcoma, the lesion appears still to be contained by a shell of periosteal new bone.

similar to those of the majority of bone neoplasms: spinecho T1-weighted images show a low to intermediate intensity signal, whereas with T2-weighting signal intensity is intermediate to high. More sclerotic lesions are of low signal intensity on all pulse sequences (Fig. 2-62). MRI effectively reveals peritumoral edema in the adjacent



Figure 2-56 Types of osteoblastoma: periosteal. Anteroposterior radiograph shows a juxtacortical lesion of the mandible.

marrow and in soft tissues. A recent report by Crim et al. (165) of a 19-year-old man with osteoblastoma described an unusual "flare phenomenon" that caused a misleading appearance on MR scans, simulating a malignant process such as lymphoma or Ewing sarcoma. The phenomenon represented a widespread inflammatory response to the lesion that led to diffuse, reactive inflammatory infiltrates in two vertebrae, the adjacent ribs, and the paraspinal soft tissues. It is noteworthy that only CT-myelography was diagnostic in this case.

Histopathology

Osteoblastoma is marked by a very active new formation of osteoid and immature bone trabeculae produced by compact masses of large osteoblasts (44). Osteoid production is generally greater in osteoblastoma than in osteoid osteoma, and the lesion is more vascularized (185) (Fig. 2-63). Furthermore, osteoblastoma exhibits a less organized pattern of osteoid and trabecular bone distribution than the nidus of osteoid osteoma, which invariably appears as a circumscribed, organized structure maturing toward its periphery; in effect, the lesion of osteoblastoma is at the same stage of development throughout (44). Very rarely, cartilaginous matrix may

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Figure 2-57 Osteoblastoma: conventional tomography. A: Radiograph of the right scapula shows a faint radiolucent focus surrounded by a sclerotic area, accompanied by shaggy periosteal reaction at the axillary border. **B:** Conventional tomogram clearly demonstrates a radiolucent nidus with a sclerotic center resembling an osteoid osteoma. However, the size of this lesion (3 cm \times 3 cm) marks it as an osteoblastoma, the diagnosis proven by excisional biopsy.

Figure 2-58 Osteoblastoma: scintigraphy. Radionuclide bone scan, obtained after injection of 15 mCi (555 MBq) ^{99m}Tc-labeled methyl diphosphonate (MDP) in a 15-year-old girl with osteoblastoma in the proximal humerus, reveals an increased uptake of tracer localized to the site of the lesion.

Δ





Figure 2-59 Osteoblastoma: computed tomography (CT). A: Radiograph of the left shoulder shows a radiolucent lesion in the sternal segment of the left clavicle in this 4-year-old girl. The exact location and the nature of this lesion are not clear. **B:** CT section reveals the location and size of the lesion. Central ossifications are typical for osteoblastoma, the diagnosis confirmed on the histopathologic examination. (Courtesy of Dr. A. A. DeSmet, Madison, Wisconsin.)



Figure 2-60 Osteoblastoma: computed tomography (CT). A: Lateral radiograph of the skull shows a well-defined lytic lesion in the frontal bone (*arrows*). B: CT section shows, in addition, a focus of bone formation within the lesion, typical of osteoblastoma.

be present (161). Occasionally, spindle-shaped hyperchromatic cells with uniform nuclei and irregular eosinophilic cytoplasm may be interdispersed among bony trabeculae (173). In contrast to most forms of osteosarcoma, osteoblastomas characteristically show variable numbers of osteoclasts at the surfaces of the bone trabeculae and seams of osteoblasts rimming the periphery of the lesion (Figs. 2-64A–C). In addition, they do not infiltrate or permeate the normal trabeculae at the interface with the lesion, as occurs in osteosarcoma (34), but usually are separated from them by a narrow but distinct layer of bone-free fibrous tissue. Some atypical osteoblastomas may, however, closely resemble an osteosarcoma (Fig. 2-64D). Some osteoblastomas contain "epithelioid" osteoblasts, twice the size of ordinary osteoblasts (171). The cells are rounded and have large nuclei containing one or more prominent nucleoli; their cytoplasm is usually abundant. Bone trabeculae are wider and more irregular than in conventional osteoblastoma, and cement lines are usually absent (179) (Fig. 2-65). These variants have been denoted as "aggressive" osteoblastomas. Della Rocca and Huvos (168) stated recently that a clinically aggressive behavior of osteoblastomas is not correlated with histologic qualities but with a localization within the skeleton that is more difficult to approach surgically. Schajowicz and Lemos (195) described eight patients with "malignant" osteoblastoma whose tumors



Figure 2-61 Aggressive osteoblastoma. A: Anteroposterior radiograph of the lumbar spine shows a destructive lytic lesion affecting the right half of the vertebral body of L3 and extending to the pedicle (*arrows*) in a 65-year-old man who presented with insidious onset of pain in the lower back radiating to the right lower extremity. **B:** Computed tomography (CT) section demonstrates focal areas of bone formation within the lesion and invasion of the cortex. Subsequent biopsy revealed an aggressive osteoblastoma. (Courtesy of Dr. Ibrahim F. Abdelwahab, New York, New York. Reprinted with permission from Greenspan A. Benign bone-forming lesions. *Skeletal Radiol* 1993;22:494.)



Figure 2-62 Osteoblastoma: magnetic resonance imaging (MRI). A: Radiograph of the left proximal humerus (same patient as in Fig. 2-58) shows a large, sclerotic lesion (*arrow*), diagnosed on biopsy as osteoblastoma. B: Axial spinecho, T1-weighted MRI demonstrates that the low-intensity lesion is located posteriorly. The cortex is destroyed and the tumor extends into the soft tissues. C: Axial spin-echo, T2-weighted MRI shows that the lesion remains of low signal intensity, indicating osteoblastic matrix.

appear to be similar to "aggressive osteoblastoma." They noted on microscopic examination more frequent atypical mitoses and increased numbers of osteoclast-like giant cells. In addition, there were bone spicules of regular shape (dark blue staining with hematoxylin), closely resembling those observed in osteosarcoma. Another pseudomalignant or pseudoanaplastic variant of osteoblastoma contains bizarre multinucleated cells completely devoid of mitotic activity (160,187,189).

Differential Diagnosis

Radiology

To distinguish osteoblastoma from *osteoid osteoma* can be very difficult, if not impossible, on both radiologic and histologic grounds. Some authorities believe that because of its striking histologic similarity to osteoid osteoma, osteoblastoma represents a variant of clinical expression of the same pathologic process. Both lesions are considered separate entities, however, because their clinical manifestations and skeletal distribution are distinct: osteoid osteoma is invariably accompanied by nocturnal pain promptly relieved by salicylates, whereas osteoblastoma is larger than osteoid osteoma



and has a predilection for the axial skeleton. In addition, osteoid osteoma has a tendency to regress spontaneously, whereas osteoblastoma tends to progress and (although this is still a controversial speculation of some investigators) may undergo malignant transformation. The most useful differential factor is the size of a lesion: osteoblastoma usually measures 2 cm or more.

The differential diagnosis of osteoblastoma should also include *bone abscess* (Brodie abscess), *aneurysmal bone cyst*, and *osteosarcoma*. The same criteria distinguishing a bone abscess from osteoid osteoma apply in relation to osteoblastoma (see "Differential Diagnosis" under "Osteoid Osteoma"). The key to making the distinction is usually the serpentine track in bone abscess that extends toward the growth plate (see Fig. 2-45). At times the distinction between these two lesions may be difficult (Fig. 2-66A). Osteoblastoma located in a short tubular bone may be misdiagnosed as *enchondroma* (Fig. 2-66B, C). Expansive osteoblastoma, particularly in spinal location, on radiography may occasionally mimic an *aneurysmal bone cyst* (see Fig. 2-54) or a *giant cell tumor*.

Aggressive osteoblastoma should be differentiated from *osteosarcoma* (162) (see Fig. 2-55). Cross-sectional imaging modalities such as CT and MRI are helpful in

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Figure 2-64 Histopathology of osteoblastoma. A: Densely arranged small trabeculae made up of woven bone are surrounded by osteoblasts. In the narrow marrow spaces, some capillaries and giant cells of the osteoclast type are seen. These features, together with very regular appearing bone formation, militate against the diagnosis of osteosarcoma (hematoxylin and eosin, original magnification $\times 25$). **B:** Osteoblastoma shows woven bone formation in plates rimmed with densely arranged osteoblasts and giant cells of the osteoclast type (hematoxylin and eosin, original magnification $\times 400$). **C:** At high magnification, the uniform-appearing osteoblasts line the newly formed trabeculae (hematoxylin and eosin, original magnification $\times 400$). **D:** Atypical osteoblastoma with crowded, large epithelioid stromal cells and irregular foci of woven bone formation resembles an osteosarcoma (hematoxylin and eosin, original magnification $\times 400$).

making the distinction by demonstrating the soft tissue component of osteosarcoma.

Intracortical osteosarcoma, because it may have a benign radiographic appearance, can mimic periosteal and intracortical osteoblastoma (188). The principal means for distinguishing these entities are histologic.

Pathology

Osteoblastoma may be confused with *osteoid osteoma*, unless clinical and radiologic factors are considered. Although the nidus of osteoid osteoma is quite similar to that of osteoblastoma, some authorities postulate that there are minor microscopic differences (191,196): at the periphery of a nidus of osteoid osteoma a fibrovascular rim is often present, a feature only occasionally encountered in osteoblastoma. The latter tumor, on the other hand, often has a lobulated or multifocal outer margin (13). Moreover, the nidus of an osteoid osteoma often exhibits a distinct zonal pattern with central maturation to more mineralized woven bone.

Aneurysmal bone cyst can be easily distinguished from osteoblastoma. The former lesion consists of multiple blood-filled sinusoid spaces alternating with more solid areas. The solid tissue is composed of fibrous elements containing numerous giant cells and is richly vascular. The sinusoids have fibrous walls, often containing osteoid tissue or even lamellae of mature bone, though not in quantities seen in osteoblastoma (see Figs. 7-41 to 7-43). Furthermore, unlike in osteoblastoma, focal or diffuse collections of hemosiderin or reactive foam

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Figure 2-65 Histopathology of "aggressive" osteoblastoma. A: Aggregates of large osteoblasts are producing osteoid and irregular bone trabeculae accompanied by dilated capillaries (hematoxylin and eosin, original magnification ×200). **B:** Large epithelioid-like osteoblasts with brightened paranuclear Golgi fields surround an unorganized osteoid matrix (hematoxylin and eosin, original magnification ×400). **C:** In another area, foci of osteoid (*red*) are bordered by large epithelioid osteoblastic tumor cells (van Gieson, original magnification ×400).



cells may be seen in the fibrous septa of aneurysmal bone cyst (194,197).

Giant cell tumor characteristically demonstrates related dual populations of giant cells and mononuclear stromal cells. Giant cells are evenly distributed within the intervening spindle-celled stromal tissue. Osteoid and bone formation may occasionally occur but only in small amounts, never dominating the microscopic picture.

Diagnosis of *osteosarcoma* should always be the differential possibility. Histopathologic examination can make the distinction (see "Histopathology"). Although in osteoblastoma a certain number of mitotic figures may be found, the lack of areas of necrosis and lack of cellular pleomorphism helps to distinguish this lesion from osteosarcoma (43).

Intracortical osteosarcoma exhibits distinctive characteristics (83,123). According to Mirra et al. (188), the following histopathologic features are critical for diagnosis of intracortical osteosarcoma: (a) permeation of the Haversian systems with attenuation and trapping of cortical lamellar bone, (b) atypical mitoses or anaplasia, and (c) presence of tumor cartilage.

In the jaw, *cementoblastoma* can be distinguished from osteoblastoma by its intimate contact to the root of a

(molar) tooth that always is surrounded by lesional tissue (198).

The radiologic and pathologic differential diagnosis of osteoblastoma is depicted in Figure 2-67 and Table 2-3.

Malignant Tumors

Osteosarcomas

Osteosarcomas comprise a family of connective tissue tumors with various degrees of malignant potential. All these tumors share the characteristic ability to produce bone or osteoid directly from neoplastic cells (392). Osteosarcomas constitute about 20% of all primary bone malignancies (214,335). The collection of large series of osteosarcomas has led to the identification of an increasing number of subtypes (431).

Almost all osteosarcomas harbor complex cytogenetic and molecular genetic alterations (229,342, 374,375). However, no specific finding has emerged that might be used as a molecular or cytogenetic marker for the diagnosis of osteosarcoma (390). As extensively discussed by Sandberg and Bridge, conven-

B, C



Figure 2-66 A: Bone abscess. Focus of infection in the proximal shaft of the fourth metatarsal in a 16-year-old boy was originally diagnosed as osteoblastoma. **B, C: Enchondroma-like osteoblastoma.** In another patient, dorsovolar and lateral radiographs of the small finger reveal a radiolucent lesion with scalloped borders in the middle phalanx, resembling an enchondroma. Small radiopacities in the center represent bone formation rather than chondroid matrix. The lesion proved to be an osteoblastoma.



Condition (Lesions)	Radiologic Features	Pathologic Features
Cortical and medullary osteoid osteoma-like osteoblastoma (giant osteoid osteoma)	Radiolucent lesion, spherical or oval, with well-defined margins. Frequent perilesional sclerosis. Abundant periosteal reaction. Size of the nidus >2 cm.	Active formation of osteoid and immature bone trabeculae. Less organized pattern of osteoid and reticular bone distribution than seen in osteoid osteoma. Hypertrophic osteoblasts. Increased yascularity in
Aneurysmal bone cyst–like expansive osteoblastoma	Blow-out lesion, similar to aneurysmal bone cysts, but with central opacities.	the stroma. Occasionally spindle- shaped hyperchromatic cells with uniform nuclei and irregular eosinophilic cytoplasm interdispersed among bony trabeculae. Variable number of giant cells on the surface of bone trabeculae
Aggressive osteoblastoma (simulating malignant neoplasm)	Ill-defined borders, destruction of the cortex; aggressive-looking periosteal reaction; occasionally soft tissue extension.	Large, "epithelioid" osteoblasts. Rounded cells with large nuclei containing one or more prominent nucleoli; abundant cytoplasm. Bone trabeculae wider and more irregular than in other types of osteoblasma. Cement lines usually absent. Atypical mitoses. Bone spicules staining dark blue with bematoxylin-eosin
Periosteal osteoblastoma	Round or ovoid heterogenous in density mass attached to cortex.	Trabeculae of woven bone, numerous dilated capillaries, exuberant in number of osteoblasts, osteoclasts, and occasionally fibroblasts.
Osteoid osteoma	See Table 2-2.	See Table 2-2.
Aneurysmal bone cyst	Blow-out, expansive lesion. In long bone buttress of periosteal reaction. This shell of reactive bone frequently covers the lesion, but may be absent in rapidly growing lesions. Soft tissue extension may be present.	Multiple blood-filled sinusoid spaces separated by fibrous septae displaying lamellae of primitive woven bone; may contain hemosiderin and reactive foam cells; solid areas composed of fibrous elements containing irregular bone trabeculae and giant cells, sometimes in great numbers
Osteosarcoma	Permeative or moth-eaten bone destruction; wide zone of transition; tumor-bone in form of cloud-like opacities; aggressive periosteal reaction; soft tissue mass.	Permeation of cortical bone; attenuation and "trapping" of lamellar bone; atypical mitoses or anaplasia; hyperchromatism and pleomorphism of cells and nuclei; tumor bone and tumor cartilage formed by malignant cells.

Table 2-3 Differential Diagnosis of Osteoblastoma

tional osteosarcomas reveal complex, unbalanced cytogenetic alterations, with pronounced variations in chromosome number and/or form, very often within the same tumor. Structural abnormalities are most often found in genes 1p11–p13, 1q11–q12, 1q21–q22, 11p14–p15, 14p11–p13, 15p11–p13, 17p, and 19q13 (229). Loss of chromosomes 9, 10, 13, and 17 and gain of chromosome 1 are the most common abnormalities (390). Interphase fluorescent in situ hybridization (FISH) studies demonstrated chromosomal instability as a characteristic finding in osteosarcomas (obviously related to defects in chromosomal segregation mechanisms and *TP53* alterations) that may be responsible for this tumor's extreme aneuploidy (208).

Inactivation of tumor suppressor genes and overexpression of oncogenes are present in many osteosarcomas (e.g., *RB1*, *TP53*, *INK4A*, *CDK4*, *MDM2*, *CCND1/CyclinD1*) and lead to functional impairment of the cell cycle by affecting both the retinoblastoma (*RB1*) and/or the *TP53* pathway. Other activated or overexpressed oncogenes not related to *RB1* or *TP53* pathways are *FOS*, *MYC*, *MET*, *SAS*, and *ERBB2* [for details see reference (390)]. Genetic alterations may also contribute to drug resistance, especially to methotrexate, by influencing the drug's uptake or target (324).

Recent retrospective studies on mechanisms of telomere maintenance [i.e., lengthening of the repetitive end sequences of linear chromosomes—telomeres—by enzyme activity of telomerase (TA)] or by other mechanisms in the absence of TA, so-called alternative lengthening of telomeres (ALT) have provided evidence for a better prognosis of TA- and ALT-negative cases The etiology of the vast majority of osteosarcomas is unknown, and they are therefore considered to be idiopathic or primary. A smaller number of tumors can be related to known factors that predispose to malignancy, such as Paget disease of bone, bone infarct, fibrous dysplasia, external ionizing irradiation, and ingestion of radioactive substances. These lesions are classified as secondary osteosarcomas (244).

All types of osteosarcomas can also be subclassified, according to their anatomic site, as either axial or appendicular (202,316). They can be further classified, on the basis of their location in bone, as intraosseous, intracortical, or surface (juxtacortical). A separate and very small group consists of primary osteosarcomas that originate in the soft tissues (so-called extraskeletal or soft tissue osteosarcomas). In general, intraosseous osteosarcomas are much more common than juxtacortical tumors and tend to have a higher grade of malignancy (276,277).

On the basis of clinical, radiographic, and histopathologic data, the World Health Organization (WHO) categorizes osteosarcomas as follows: conventional (further subdivided according to the predominantly produced matrix as chondroblastic, fibroblastic, or osteoblastic) and unusual forms that do not exhibit a different biological behavior but histologically may imitate other bone tumors, e.g., malignant fibrous histiocytoma (MFH)–like or giant cell–rich osteosarcomas, telangiectatic, small cell, low-grade central, secondary, parosteal, periosteal, and high-grade surface osteosarcoma (15) (Table 2-4). For differential diagnosis, however, it appears appropriate to use a modified classification scheme that takes into account etiology (primary

Table 2-4 Classification of Osteosarcomas According to World Health Organization (WHO) (2002)

(Conventional osteosarcoma (OS)			
	Chondroblastic			
	Fibroblastic			
	Osteoblastic			
	Unusual histologic forms:			
	Sclerosing OS			
	OS resembling osteoblastoma			
	Chondromyxoid fibroma–like OS			
	Chondroblastoma-like OS			
	Clear-cell OS			
	MFH-like OS			
	Giant cell-rich OS			
	Epithelioid OS			
٦	Telangiectatic OS			
5	Small cell OS			
Low-grade central OS				
Secondary OS				
Parosteal OS				
F	Periosteal OS			
ł	High-grade surface OS			

and secondary), localization and bone-specific topography (intraosseous, intracortical, surface, and soft tissue), and histology according to WHO (Fig. 2-68).

Clinical Presentation

The clinical presentation varies from case to case and is largely dependent on the type of osteosarcoma, its anatomic site, and the age of the patient (316). Pain and development of a soft tissue swelling or mass are the most common presenting symptoms (265,338). Rarely, a pathologic fracture is the first sign of malignancy (345). At first the pain may be slight and intermittent, but gradually it becomes more severe and constant (233). In juxtacortical tumors, the presence of a hard, slowly enlarging, painless mass is the most common clinical finding. Although pulmonary metastases (frequently present as micrometastases at the time of diagnosis) are the most usual and most significant complication in the majority of high-grade osteosarcomas, they are rare in osteosarcomas of the jaw and are a late complication in multicentric osteosarcomas. Osteosarcoma metastasizing to skeletal muscle is exceedingly rare (369).

Imaging

For diagnostic purposes, conventional radiography usually provides adequate information. The findings of bone destruction associated with sclerotic foci of tumor bone formation, an aggressive (sunburst-type, lamellated, or Codman triangle) periosteal reaction, and a soft tissue mass are highly suggestive of osteosarcoma. Conventional tomography, although presently rarely performed, may aid in delineating the osseous matrix and cortical destruction and in detection of occult pathologic fracture. Arteriography is usually reserved for mapping of the tumor and its vascular supply, and for identification of the area most suitable for an open biopsy, although recently this technique has been replaced by MR angiography (MRA). Radionuclide bone scan can assist in diagnosis of intraosseous metastases and skip lesions. CT and MRI are crucial for determining the intraosseous and extraosseous extension of a tumor (227,400,416,464). PET imaging of osteosarcoma is not yet widely accepted, although it appears to be a promising technique (228).

Histopathology

Like the imaging findings, the histopathologic features of osteosarcoma vary according to the particular subtype. However, a common characteristic of all subtypes is the presence of tumor osteoid or tumor bone formed by the malignant cells.

Identification of osteoid relies on histomorphology. Although attempts have been made to define osteoid by biochemical or immunohistochemical means, accepted criteria are not yet established (398). On microscopy, osteoid is located intercellularly as a dense amorphous material, eosinophilic in hematoxylin and eosin (H&E) and fuchsinophilic in van Gieson stain. It must be differentiated from collagen, fibrin, and amyloid. Identification is sometimes extremely difficult or even arbitrary. The mode of deposition may be of help, **88** — Differential Diagnosis in Orthopaedic Oncology



Figure 2-68 Classification of osteosarcomas. MFH, malignant fibrous histiocytoma.

because osteoid is laid down afibrillarly and curvilinearly with ramifications and bud-like densities. It may also be deposited on normal bone trabeculae.

Osteosarcomas can be graded on the basis of their cellularity, cytologic atypia (nuclear pleomorphism), and mitotic activity. Grading has clinical, therapeutic, and prognostic importance (242,416). Three different grading systems comprising four grades, three grades, or two grades (low and high grade) are presently in use (Table 2-5). In Broders' four-grade system (230), the numerical grade (1 to 4) indicates the degree of malignancy (grade 1 indicating the least undifferentiated tumor with extensive formation of tumor bone and grade 4 the most undifferentiated tumor with only sparse amount of tumor osteoid) (Table 2-6). In the three-grade system, well differentiated (grade 1, correspon-

ding to Broders' grade 1 and 2), moderately differentiated (grade 2, corresponding to Broders' grade 3), and poorly differentiated tumors (grade 3, corresponding to Broders' grade 4) are separated, whereas the two-

Table 2-5 Grading of Osteosarcomas

Two-Grade System	Three-Grade System	Four-Grade System (Broders')
Low grade	Grade 1	Grade 1 Grade 2
High grade	Grade 2 Grade 3	Grade 3 Grade 4

Table 2-6 Histologic Grading of Osteosarcoma^a

Grade	Histologic Features
1	Cellularity: slightly increased Cytologic atypia: minimal to slight Mitotic activity: low Osteoid matrix: regular
2	Cellularity: moderate Cytologic atypia: mild to moderate Mitotic activity: low to moderate Osteoid matrix: regular
3	Cellularity: increased Cytologic atypia: moderate to marked Mitotic activity: moderate to high Osteoid matrix: irregular
4	Cellularity: markedly increased Cytologic atypia: markedly pleomorphic cells Mitotic activity: high Osteoid matrix: irregular, abundant

^a According to Unni KK, Dahlin DC. Grading of bone tumors. *Semin Diagn Pathol* 1984;1:165–172.

grade system merely discriminates between low- (grade 1 or Broders' grade 1 and 2) and high-grade lesions (grade 2 and 3 or Broders' grade 3 and 4) (see Table 2-5). For example, well-differentiated central osteosarcomas and parosteal osteosarcomas are regarded as Broders' grade 1, rarely grade 2 tumors; periosteal osteosarcomas and gnathic osteosarcomas as Broders' grade 2, rarely grade 3; and conventional osteosarcomas as grade 3 or 4. Telangiectatic osteosarcomas, osteosarcomas developing in pagetic bone, postirradiation osteosarcomas, and multifocal osteosarcomas are usually Broders' grade 4 tumors. Almost 90% of all osteosarcomas are high-grade lesions.

Primary Osteosarcomas

A. Intraosseous Osteosarcomas

Intramedullary (Conventional) Osteosarcoma

Intramedullary osteosarcoma is the most common type of osteosarcoma, constituting approximately 85% of all forms of osteosarcoma (349,379). Formation of osteoid or bone by tumor cells is the criterion for the histologic diagnosis (241). The incidence of medullary osteosarcoma is greatest during the second decade of life, and males are affected slightly more often than females. The knee (distal femur, proximal tibia) is the most commonly affected site, followed by the proximal humerus (243) (Fig. 2-69). The metaphysis is more frequently affected than the diaphysis, and the growth plate usually serves as a barrier to tumor spread into the epiphysis, although invasion of the physis and involvement of the epiphysis and the joint have been reported (359,407). Primary osteosarcoma of the spine is rare (299). The most common complications of conventional osteosar-



Figure 2-69 Conventional osteosarcoma: skeletal sites of predilection, peak age range, and male-to-female ratio.

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comas are pathologic fracture and development of pulmonary metastases.

Imaging

The distinctive radiologic features of conventional osteosarcoma, as demonstrated by radiography, are medullary and cortical bone destruction, an aggressive periosteal reaction, a soft tissue mass, and the presence of tumor bone either within the destructive lesion or on its periphery, as well as within the soft tissue mass itself, often assuming a spiculated or sunburst pattern (244,321) (Fig. 2-70). In some instances the type of bone destruction is not so obvious (249,385), but patchy densities, representing tumor bone, and an aggressive periosteal reaction provide clues to the diagnosis (Fig. 2-71). The degree of radiopacity in the tumor reflects a combination of the amount of tumor bone production, calcified matrix, and osteoid. Tumors may present as purely sclerotic lesions or purely osteolytic lesions (248), but mostly a combination of both (Fig. 2-72). The borders are usually indistinct, with a wide zone of transition. The type of bone destruction is either moth-eaten or permeative, only rarely geographic. The most common periosteal reactions observed with this tumor are the sunburst type and the Codman triangle; the lamellated (onion-skin) type of periosteal response is less common

(Fig. 2-73). The presence of a soft tissue mass, which may contain sclerotic foci (tumor bone formation), is a common finding (Fig. 2-74).

CT is invaluable for evaluation of these tumors (397), particularly for determining the degree of tumor extension in the bone marrow. CT using Hounsfield units can accurately determine the attenuation coefficient of the medullary cavity (Fig. 2-75). The normal bone marrow has a fat density of -20 to -80 Hounsfield units. With tumor infiltration, this density is significantly increased (275).

MRI is equally if not more effective for evaluating intraosseous tumor extension and soft tissue involvement (227,364,388,438,442). On T1-weighted spin-echo images, the mineralized components of osteosarcoma display a low signal intensity, whereas the solid, nonmineralized portions of the tumor usually appear as areas of low to intermediate signal intensity. On T2-weighted spin-echo images and T2* gradient sequences, the mineralized portions of tumor display a low signal intensity, and nonmineralized areas and soft tissue mass are of high signal intensity (Figs. 2-76 and 2-77). Osteosclerotic tumors demonstrate a low signal intensity on both T1- and T2-weighted sequences (225). MRI may also effectively demonstrate the peritumoral edema, which will image as bright areas adjacent to the tumor on T2-



Figure 2-70 Conventional osteosarcoma. Anteroposterior (A) and lateral (B) radiographs demonstrate the typical features of this tumor in the distal femur of a 19-year-old woman. Medullary and cortical bone destruction is present, in association with aggressive periosteal reaction of the velvet and sunburst types. A soft tissue mass contains tumor bone.



Figure 2-71 Conventional osteosarcoma. Although no gross bone destruction is evident in the distal femur of this 16-year-old girl, the patchy densities in the medullary portion of the femur and the sunburst appearance of the periosteal response are clues to the diagnosis of osteosarcoma. Note also the presence of a Codman triangle (*arrow*).

or T2*-weighted sequences. In addition to staging both the extraosseous and intraosseous extent of the tumor, MRI can aid in the identification of various components within the tumor, such as cellular areas, and regions of necrosis and liquefaction. Cartilaginous components are particularly easy to identify by virtue of the septumlike enhancement seen on gadolinium-enhanced MR images (223,268).

Arteriography is rarely used, as it is primarily reserved for mapping of the vascular supply of the tumor, and sometimes for the identification of the areas most desirable for biopsy (Fig. 2-78). Scintigraphy will invariably show an increased uptake of the radiopharmaceutical tracer (Fig. 2-79). In addition, as indicated previously, radionuclide bone scan can assist in the identification of intraosseous metastases (Fig. 2-80).

Histopathology

On the basis of the dominant histologic features, conventional osteosarcoma can be subdivided into three main histologic subtypes: osteoblastic, chondroblastic, and fibroblastic (Fig. 2-81). As stated previously, each of these is characterized by the formation of osteoid tissue or bone by the neoplastic cells. The specific appearance of osteosarcoma reflects its histologic grade. Microscopic

examination usually reveals highly pleomorphic spindleshaped and polyhedral tumor cells, producing different forms of osteoid (Fig. 2-82). Mitotic figures are commonly seen. Malignant giant cells may occasionally be found. Bone matrix formation may be subtle, displaying sporadic wisps in a lace-like pattern resembling a doily or a lotus root (277). The matrix sometimes takes the form of bulky deposits, which have been described as massive osteoid. Necrotic areas and hemorrhage may be present. The intercellular material may be fibrous, cartilaginous, osteoid, or osseous, with various amounts of calcification (349). If osteoid deposition is plentiful, the tumor cells and their nuclei may become inconspicuous, a process referred to by Jaffe as normalization (303). In some instances, the tumor cells are so undifferentiated that cytologic study fails to clearly determine if they are mesenchymal or epithelial in origin, a variant of conventional osteosarcoma sometimes referred to as epithelioid osteosarcoma (246,318,431) (Fig. 2-83). This appearance may lead to an incorrect diagnosis of metastatic carcinoma. The diagnosis in such cases can usually be deduced on the basis of the patient's age, the production of obvious osteoblastic tumor matrix, the immunohistochemical demonstration of the mesenchymal character of the tumor cells, and a radiographic appearance typical of osteosarcoma (277). Occasionally, conventional osteosarcoma may exhibit large numbers of cytologically bland, reactive stromal giant cells that may obscure the underlying sarcomatous stroma (13). This variant is known as giant cell-rich osteosarcoma (see later).

Malignant Fibrous Histiocytoma–like (Fibrohistiocytic) Osteosarcoma

This variant, which resembles MFH, has recently been described in the literature (216). Although only small series have been published thus far (216,284,310, 354,462), the MFH-like (or fibrohistiocytic) subtype comprises 3% to 23% of all osteosarcomas (296,345). This variant can sometimes be confused with true MFH of bone because both of these tumors tend to arise at an older age than conventional osteosarcoma, usually after the third decade. Both tend to involve the articular ends of long bones, and less periosteal reaction is typically present than in conventional osteosarcoma. Although on radiography both of these lesions tend to be radiolucent and therefore to resemble giant cell tumor and fibrosarcoma, the MFH-like osteosarcoma usually exhibits areas of bone formation resembling cotton balls or cumulus clouds, whereas MFH does not (245).

In a recent study of osteosarcomas in patients older than age 40, Kawaguchi et al. (310) demonstrated immunoreactivity for the p53 gene in one of eight MFHlike subtypes, which also showed a missense mutation of p53, whereas MDM2 was negative, in contrast to conventional osteoblastic osteosarcomas (5/15 and 3/15) and MFH of bone (8/18 each). In addition, the latter were associated with mutations of p53 in four cases each. When such areas are identified on radiologic studies, a diligent search should be made for tumor bone in the resected specimen (316). Histologically, MFH-like osteosarcoma is characterized by pleomor-*Text continues on page 99*



Figure 2-72 Various presentations of osteosarcoma. A: Purely sclerotic osteosarcoma. Anteroposterior (A1) and lateral (A2) radiographs show sclerotic variant of osteosarcoma in the proximal tibia. B: Purely lytic osteosarcoma. An anteroposterior radiograph shows destructive lesion in the distal humerus, which proved to be a fibroblastic osteosarcoma. C: Mixed osteosarcoma. Areas of bone formation are present within a destructive lesion in the distal femur.



Figure 2-73 Types of periosteal reaction in osteosarcoma. A1, A2: The sunburst type is the most common (continued).

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Figure 2-74 Conventional osteosarcoma. A soft tissue mass containing tumor bone is a common finding in osteosarcoma, as seen in this 20-year-old man with a lesion in the right proximal fibula.





Figure 2-75 Conventional osteosarcoma: computed tomography (CT). A: Anteroposterior radiograph of the left proximal femur of a 12-year-old boy demonstrates an osteolytic lesion in the intertrochanteric region, with a poorly defined margin and amorphous densities in the center, associated with a periosteal reaction medially. These features suggest osteosarcoma, which was confirmed on biopsy. Because a limb salvage procedure was contemplated, a CT scan was performed to determine the extent of marrow infiltration and the required level of bone resection. The most proximal section (**B**) shows obvious gross tumor involvement of the marrow cavity of the left femur (*continued*).



Figure 2-75 *Continued* A more distal section (C) shows no gross marrow abnormality, but a positive Hounsfield value of 52 units indicates tumor involvement of the marrow, which was not shown on the standard radiographs. By comparison, the section of the right femur shows a normal Hounsfield value of 26 for bone marrow.











Figure 2-76 Conventional osteosarcoma: magnetic resonance imaging (MRI). Coronal **(A1)** and sagittal **(A2)** spin-echo, T1-weighted MRI (TR 800, TE 20) in this 17-year-old boy with osteosarcoma of the left proximal tibia shows inhomogeneous appearance of a tumor. The mineralized parts of osteosarcoma display low signal intensity, whereas nonmineralized areas are of intermediate signal intensity. **B:** On spin-echo, T2-weighted MRI (TR 2000, TE 80), the sclerotic parts of the tumor remain of low signal intensity, whereas a nonmineralized part of the lesion (distally) shows high signal intensity. Likewise, the soft tissue extension of osteosarcoma displays high signal intensity.

A2


Figure 2-77 Conventional osteosarcoma: magnetic resonance imaging (MRI). Coronal (**A1**) and sagittal (**A2**) spin-echo, T1-weighted MRI (TR 800, TE 20) in a 29-year-old man with osteosarcoma of midshaft of the left femur shows a tumor displaying predominantly low signal intensity. Coronal (**B1**) and sagittal (**B2**) T1-weighted (TR 650, TE 19) MR images obtained after intravenous injection of gadolinium (gadopentetate dimeglumine), using the fat suppression technique, show tumor enhancement in the medullary portion of bone and in the soft tissues. Heavily mineralized portions of tumor remain of low signal intensity.



Figure 2-78 Conventional osteosarcoma: angiography. Arteriography reveals increased vascularity of a tumor involving the left tibial metaphysis. Also noted are abnormal tumor vessels in the soft tissue mass. Biopsy is usually performed from the most viable (vascular) areas.



Figure 2-79 Conventional osteosarcoma: scintigraphy. A: Anteroposterior radiograph shows a sclerotic variant of conventional osteosarcoma in the proximal tibia. B: After injection of 15 mCi (555 MBq) of ⁹⁹mTc-labeled methylene diphosphonate (MDP), there is significantly increased uptake of the tracer in the tumor.

В



Figure 2-80 Conventional osteosarcoma of the proximal humerus with intraosseous metastases ("skip lesions"). A: Scintigraphy shows markedly increased uptake of the radiopharmaceutical agent in the proximal humerus, corresponding to the primary tumor. In addition, two small foci of activity are seen distal to the main lesion (*arrows*). This finding, representing skip lesions (*arrows*), was confirmed by radiography (**B**) and magnetic resonance imaging (**C**).

phic spindle cells and giant cells, many with bizarre nuclei. This lesion therefore resembles giant cell–rich osteosarcoma. An inflammatory background is not unusual, and the storiform or spiral nebular arrangement characteristic of MFH, although sometimes a dominant feature, may be less prominent or may be replaced by areas of large pleomorphic cells arranged in diffuse sheets (216). As in all other subtypes of osteosarcoma, the distinction from other sarcomas depends on the demonstration of osteoid or bone formation by malignant cells in the very typical patterns seen in osteosarcomas (245,297,383,453).

Giant Cell–Rich Osteosarcoma

This is a rare variant of osteosarcoma, which appears as an undifferentiated sarcoma with an overabundance of osteoclasts (giant cells) and a paucity of tumor osteoid or tumor bone. It composes approximately 3% of all osteosarcomas and histologically is related to telangiectatic osteosarcoma and MFH-like osteosarcoma. Many of the classical radiologic features of osteosarcoma are absent, and differentiation from a benign lesion is sometimes difficult (219). In most cases the lytic lesion shows poorly defined borders. The typical location is the metaphysis or diaphysis of a long bone, usually the tibia or femur. A periosteal reaction is absent or scant, and a soft tissue mass is usually not present. The lesion has a striking histologic resemblance to giant cell tumor (Fig. 2-84). Tumor osteoid is usually scant and is therefore difficult to identify.

Small Cell Osteosarcoma

Originally described by Sim et al. (406), small cell osteosarcoma accounts for approximately 1.5% of all osteosarcomas. This subtype usually presents as a lucent lesion exhibiting permeative borders and a large soft tissue mass. Its appearance therefore mimics round cell tumors of bone, such as Ewing sarcoma. In some instances, small cell osteosarcoma presents as a sclerotic lesion that is indistinguishable from conventional osteosarcoma (256). The majority of cases display an aggressive periosteal reaction (221). The distal femur, proximal humerus, and proximal tibia are preferential sites (215). Almost half of the patients are in their second decade at the time of clinical presentation (223,355).

On histopathologic study, these lesions usually exhibit loose aggregations of small round cells separated by collagenous bands of a fine eosinophilic matrix that resembles Ewing sarcoma (254) (Fig. 2-85). To blur the distinction further, this tumor may also exhibit a positive staining for glycogen, as described for Ewing sarcoma. However, reticulin fibers that are almost always absent or extremely rare in Ewing sarcoma have been demonstrated in small cell osteosarcoma





Figure 2-81 Histologic subtypes of osteosarcoma. A: Osteoblastic. Irregular woven bone trabeculae are rimmed by pleomorphic tumor osteoblasts with hyperchromatic nuclei (hematoxylin and eosin, original magnification $\times 100$). **B:** Chondroblastic. The tumor consists mainly of cartilaginous tissue, with only scanty islands of tumor-osteoid (*lower right*) (hematoxylin and eosin, original magnification $\times 50$). **C:** Fibroblastic. **(C1)** Predominantly spindle fibroblast-like tumor cells with deeply stained nuclei are arranged in sheaths (hematoxylin and eosin, original magnification $\times 200$). **(C2)** In another field, malignant cells produce osteoid adjacent to remnants of bone trabeculum (hematoxylin and eosin, original magnification $\times 200$).

Figure 2-82 Histopathology of conventional osteosarcoma. Markedly pleomorphic tumor cells are separated by lace-like osteoid (hematoxylin and eosin, original magnification \times 400).



R



Figure 2-83 Histopathology of epithelioid osteosarcoma. A: Tumor cells are arranged in a nesting pattern, resembling epithelial tumor (hematoxylin and eosin, original magnification $\times 100$). **B:** At higher magnification there is evidence of osteoid formation by the tumor cells (hematoxylin and eosin, original magnification $\times 200$).



Figure 2-84 Giant cell–rich osteosarcoma. A: Anteroposterior radiograph of the left proximal humerus shows a destructive lesion affecting the medullary portion of the bone, breaking through the medial cortex into the soft tissues. Bone formation is seen in the soft tissue mass (*arrow*). **B:** Photomicrograph demonstrates polymorphous tumor cells accompanied by osteoclast-like giant cells similar to giant cell tumor (hematoxylin and eosin, original magnification ×100). **C:** At high-power magnification atypical cells with pleomorphic nuclei, irregular mitoses, and tumor-osteoid formation are visible (van Gieson, original magnification ×400).



Figure 2-85 Histopathology of small cell osteosarcoma. A: Tumor tissue consisting of medium-sized round and oval cells, with indistinct cytoplasm and slightly pleomorphic and hyperchromatic nuclei resembling those of Ewing sarcoma. There is no obvious osteoid matrix formation in this field (hematoxylin and eosin, original magnification ×400). **B:** Small cell osteosarcoma contains glycogen (*red, left*) similar to Ewing sarcoma, that is removed by enzyme (diastase) pretreatment (*right*) (Best reaction, original magnification ×400). **C:** In contrast to Ewing sarcoma, small cell osteosarcoma contains reticulin fibers staining dark (Novotny, original magnification ×400). **D:** Tumor-osteoid formation by small cell osteosarcoma (van Gieson, original magnification ×400).

(221,406), and chromatin, although occasionally fine, is usually coarse or granular. Furthermore, the presence of ovoid nuclei and definite spindling of the tumor cells in some areas, and the often focal production of osteoid wisps or bone formation, should direct pathologists away from Ewing sarcoma and toward a confident diagnosis of small cell osteosarcoma (277). The amount of cytoplasm varies considerably and is dependent on the degree of cellularity (215). In some instances, malignant cartilage can be detected.

Telangiectatic Osteosarcoma

A very aggressive and unusual type of osteosarcoma, the telangiectatic variant affects males twice as often as females and occurs predominantly during the second and third decades (298). Originally described by Paget in 1854, and subsequently redefined by Gaylord in 1903 as a "malignant bone aneurysm" and by Campanacci in 1971 as a "hemorrhagic osteosarcoma" (234), this variant accounts for less than 5% of osteosarcomas (261,337). Most of these tumors arise in the femur or tibia (223). The tumor grossly resembles a bag of blood, characterized by blood-filled spaces, necrosis, and hemorrhage. These features account for its radiographic presentation as a purely lytic and destructive radiolucent lesion, almost devoid of sclerotic changes (Fig. 2-86A, B). A soft tissue mass may be present (Fig. 2-86E, F). An aggressive periosteal reaction (lamellar, sunburst, Codman triangle) is present in most patients, reflecting the malignant nature of this



Figure 2-86 Telangiectatic osteosarcoma. (**A**) Purely destructive lesion is seen in the femoral diaphysis of this 17-year-old girl. Note the velvet type of periosteal reaction (*arrow*). The sclerotic changes usually seen in conventional osteosarcoma are absent, and there is no radiographic evidence of tumor bone. (**B**) Lateral radiograph of the proximal tibia in a 21-year-old man shows a lytic lesion with relatively narrow zone of transition and no visible periosteal reaction. Coronal (**C**) and sagittal (**D**) T1-weighted (SE, TR 400, TE 10) magnetic resonance (MR) images show the tumor to be predominantly of intermediate signal intensity with central areas of high signal. No definite soft tissue mass is demonstrated (*continued*).

С



Figure 2-86 Continued Coronal (E) and axial (F) inversion recovery (FMPIR/90, TR 4000, TE 54 EF, TI 140) images show the extension of the tumor into the soft tissues and presence of peritumoral edema.

tumor (223) (Fig. 2-87), and a pathologic fracture is not uncommon in the case of large lesions (Fig. 2-88). On radiography the lesion is osteolytic, with or without matrix mineralization, and occasionally exhibits narrow zone of transition. Recently, Murphey et al. (353) reviewed 40 cases of pathologically confirmed telangiectatic osteosarcomas. Geographic pattern of bone destruction was present in 94% of cases, whereas motheaten or permeative destruction was seen in only 6% of patients. Wide transition zone was present in 83% of patients, and narrow zone of transition without sclerosis—only in 17%. Expansive bone remodeling was common, being present in 75% of cases, and periosteal reaction with aggressive characteristics was frequent (72%). Sixty-one percent of patients exhibited a pathologic fracture. Cortical destruction and soft tissue mass were a common finding. It was of interest to note that matrix mineralization was identified on radiography only in 58% of patients. On MRI, telangiectatic osteosarcoma often exhibits areas of a high signal intensity on T1-weighted images, owing to the presence of methemoglobin (223) (see Fig. 2-86C, D). On T2weighted images signal intensity is usually heterogeneous, although with predominantly high-signal areas containing foci of low signal intensity representing islands of mineralized matrix. Fluid-fluid levels can occasionally be seen (Fig. 2-89). These fluid-fluid levels indicate the presence of hemorrhage and are sometimes observed in other lesions that contain blood-filled cavities, such as aneurysmal bone cysts.

Two histologic patterns may be observed. The tissue usually consists of blood-filled spaces divided by septa with few solid areas, a feature also shared by the major differential possibility, aneurysmal bone cyst (Fig. 2-90A, B, D). However, high-power microscopy of the cells lining the blood-filled spaces reveals significant pleomorphism, many mitotic figures, giant cells (benign and malignant), and focal osteoid production to a much greater extent than occurs in aneurysmal bone cysts (Fig. 2-90C). Some tumors exhibit only a minimal amount of osteoid production. However, according to Dahlin and Unni (244), if a tumor produces septa it should be classified as telangiectatic osteosarcoma, even in the absence of osteoid.

Low-Grade (Well-Differentiated) Central Osteosarcoma

This rare form of osteosarcoma, representing less than 2% of all osteosarcomas, usually affects patients older than those afflicted with conventional osteosarcoma (average age 30 years) (301). Its sites of occurrence are similar, with a predilection for the femur and tibia (436). Flat and short tubular bones are rarely affected (461). Unlike patients with conventional osteosarcoma, patients with this type of tumor often present with a long history of nonspecific aching, discomfort, or swelling over months or even years before the diagnosis is established. In many cases, the initial diagnosis on both a radiologic and a histologic basis is that of fibrous dysplasia (450). Although low-grade central osteosarcoma is occasionally indistinguishable on radiography from conventional osteosarcoma, more often the lesion appears benign, with a well-defined sclerotic rim (211,258). Expansion of the lesion is common, and the cortex is often thinned (Figs. 2-91 and 2-92). Periosteal reactions are rare, as is soft tissue extension of the tumor. This type of osteosarcoma is a slow-growing tumor and therefore has a better prognosis. Its radiographic presentation sometimes closely resembles that of fibrous dysplasia, giant cell tumor, desmoplastic fibroma, or even nonossifying fibroma (405).

The histologic diagnosis of low-grade osteosarcoma is extremely difficult. It may resemble fibrous dysplasia, desmoplastic fibroma, nonossifying fibroma, osteoblastoma, and chondromyxoid fibroma (220,301), or nonspecific reactive lesions. Even the usual histology of



Figure 2-87 Telangiectatic osteosarcoma. Anteroposterior **(A)** and lateral **(B)** radiographs show an ill-defined lesion that exhibits a permeative type of bone destruction in the middle portion of the right femur in a 41-year-old man. Note the velvet type of aggressive periosteal reaction.

low-grade parosteal osteosarcoma, with formation of mature-appearing bone trabeculae arranged in parallel arrays, has been reported (431). On microscopic examination, the tumor stroma consists of spindle cells (which usually do not appear obviously malignant) arranged in interlacing bundles (Fig. 2-93). The nuclei may exhibit some minimal atypia but the pleomorphism commonly observed in conventional osteosarcoma is absent. Mitotic figures are rare and inconspicuous. Variable amounts of collagen and bone are produced, and the stromal cells may invade and permeate the marrow cavity and trabeculae of cancellous and cortical bone, occasionally extending to and penetrating the periosteum (322). The lesion may resemble desmoplastic fibroma at one end of the spectrum and fibrous dysplasia at the other, except that the bone spicules formed in this lesion are usually bulkier and lack the delicate, curvilinear shapes seen in fibrous dysplasia (316). Foci of atypical cartilage are only rarely observed. Giant cells may be present but are usually rare. As a consequence, a diagnosis should be made on the basis of histologic and radiologic correlation.

Unlike in conventional osteosarcoma, low-grade central osteosarcoma shows minor genetic alterations, with gains at 12q13–14,12p, and 6p21 resulting in overexpression of *CDK4* and *MDM2* and amplification of *SAS* (sarcoma-amplified sequence) as identified by comparative genomic hybridization (CGH) studies (a method for the analysis of copy number changes in



Figure 2-88 Telangiectatic osteosarcoma. A: Predominantly lytic tumor associated with periosteal reaction is seen in the distal femoral diaphysis of this 6-year-old girl. B: On the lateral view, an oblique pathologic fracture is well demonstrated (*arrow*). (Courtesy of Dr. K. K. Unni, Rochester, Minnesota.)

the DNA of tumor cells) (420,451). In a study by Park et al. (365), a mutation of *TP53* present in about 20% of all conventional osteosarcomas was found in only one case, whereas amplification of the *MDM2* gene was present in four out of 21 investigated low-grade central osteosarcomas. The same group of investigators analyzing 20 cases reported a strong protein expression of MDM2 (35%) and CDK4 (65%) by immunohistochemistry methods, and *SAS* amplification by the quantitative PCR (**p**olymerase **c**hain **r**eaction) method (15%) (376). In addition, ring chromosomes with material derived from chromosome 1 have also been described (451).

Gnathic Osteosarcoma

Osteosarcoma arising in the maxilla or mandible is unlike osteosarcoma arising elsewhere in the skeleton because it occurs in older patients (fourth to sixth decades, with a mean age of 35 years) (328). It is usually a well-differentiated tumor with a low mitotic rate, possessing a predominantly cartilaginous component in a high percentage of cases (Fig. 2-94), and with less malignant potential and a better prognosis than for other forms of osteosarcoma (240,277,308,333,368).







Figure 2-89 Telangiectatic osteosarcoma. A: Frontal radiograph shows permeated focal bone destruction in the proximal fibula associated with bone formation in the soft tissue (*arrow*). **B:** Coronal fat-suppressed T1-weighted magnetic resonance imaging (MRI) obtained after intravenous administration of gadolinium shows slight enhancement of both the intramedullary tumor and periphery of a large soft tissue mass (*arrows*). **C:** Axial fat-suppressed T2-weighted image shows fluid-fluid level (*open arrow*).

CHAPTER 2 Bone-Forming (Osteogenic) Lesions - 107



Figure 2-90 Histopathology of telangiectatic osteosarcoma. A: Fibroblastic tumor stroma with scanty woven bone formation (red) and large, blood-filled capillary spaces (van Gieson, original magnification ×12). B: At higher magnification, a moderate amount of bone formation, characteristic of osteosarcoma, is clearly demonstrated. Large, blood-filled vascular spaces are typical of the telangiectatic variant (hematoxylin and eosin, original magnification ×25). C: Cellular areas of the tumor reveal bone formation by malignant cells (hematoxylin and eosin, original magnification $\times 25$). **D:** In another section, there is almost complete lack of tumor bone formation, except for the areas in the upper left corner (van Gieson, original magnification $\times 25$). **E:** A pseudocyst of telangiectatic osteosarcoma is surrounded by atypical malignant cells (hematoxylin and eosin, original magnification $\times 200$). F: At higher magnification observe scattered pleomorphic tumor cells (hematoxylin and eosin, original magnification $\times 400$).



A, B



Figure 2-91 Low-grade central osteosarcoma. Anteroposterior **(A)** and lateral **(B)** radiographs of the distal leg in an 18-year-old woman were originally interpreted as showing fibrous dysplasia of the distal tibia. Note a benign-appearing radiolucent lesion exhibiting a geographic type of bone destruction with a narrow zone of transition and no evidence of periosteal reaction. Sagittal **(C)** and axial **(D)** T1-weighted (SE, TR 600, TE 20) magnetic resonance imaging (MRI) demonstrates intermediate to low signal intensity of the lesion and lack of a soft tissue mass. (Courtesy of Dr. K. K. Unni, Rochester, Minnesota.)

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Multicentric (Multifocal) Osteosarcoma

This very rare type of osteosarcoma (1.5%) of all osteosarcomas), in which lesions develop simultaneously in multiple bones, is recognized at present as having two variants: synchronous and metachronous (205,232, 269,280,326,380). A slight male predominance exists among patients of 18 years and younger and a much greater (3:1) male predominance in patients of 19 years and older. The most commonly encountered sites in decreasing order of frequency are the distal end of the femur, proximal end of the tibia, proximal end of the femur, pelvis, and the proximal end of the humerus (243,294) (Figs. 2-95 and 2-96). Whether this type of osteosarcoma is truly a separate entity or whether it actually represents multiple bone metastases from a primary conventional osteosarcoma remains controversial (294). Because bone is a well-recognized site of metastasis for primary osteosarcoma, the possibility that a patient with multifocal involvement has a primary osteosarcoma with bone metastases should always be considered. When multiple bones are affected in the absence of pulmonary involvement, however, this situation probably indicates the independent development of multiple primary bone osteosarcomas. Amstutz (210) categorized multifocal osteosarcomas into three clinical patterns: type I are synchronous lesions occurring in younger patients, type II are low-grade synchronous lesions limited to the axial skeleton in adults, and type III are cases of metachronous lesions. Some of the patients in this group were believed to develop metastatic lesions from a solitary primary osteosarcoma.

Osteosarcomas with Unusual Clinical Presentation

There are numerous genetic disorders, marked by chromosome instability, associated with development of various tumors including osteosarcomas. Among these conditions are Rothmund-Thomson syndrome, Werner syndrome, Li-Fraumeni syndrome, retinoblastoma syndrome, and Bloom syndrome (264,273).



Figure 2-92 Low-grade central osteosarcoma. A: Lytic lesion with the geographic type of bone destruction and a narrow zone of sclerosis in the intertrochanteric region of the left femur has a benign appearance. Anteroposterior **(B)** and lateral **(C)** radiographs of another patient show a radiolucent lesion in the proximal tibial diaphysis, slightly expanding the lateral cortex.

The Rothmund-Thomson syndrome, also known as congenital poikiloderma, is a hereditary disease with a male predominance of 2:1, appearing in the first year of life, and characterized by erythematous and maculopapular skin lesions with areas of hyperpigmentation (218,332,404). These lesions are associated with a variety of other abnormalities such as sensitivity to light, juvenile cataracts, short stature, growth retardation, premature baldness, hypogonadism, and development of skin malignancies (particularly basal and squamous cell carcinoma). Conventional osteosarcoma develops in about 30% of cases (particularly at a younger age) (332), although the presence of multicentric osteosarcoma has also been reported (257,404,413). The syndrome, which is inherited in an autosomal recessive manner, has been attributed to mutations of RECQL4 located at chromosome band 8q24.3, and coding for a DNA-helicase that unfolds double-stranded into single stranded DNA (314,315,372). In a cohort study, Wang et al. (448) were able to demonstrate that truncating mutations leading to loss of function of the gene are associated with a high risk of osteosarcoma in patients with this syndrome.

The **Werner syndrome**, also known as adult progeria, is a rare autosomal recessive genetic disorder caused by mutations in the *WRN* gene that has been mapped at chromosome band 8p12–p11 (274,346,350,396). The WRN protein belongs to the ReQ family, being both a DNA helicase and exonuclease. Alterations of the helicase function may lead to chromosomal instability. This syndrome is characterized by the premature aging including graying of hair, alopecia, cataracts, sclerodermalike skin changes, osteoarthritis of peripheral joints, short stature, hypogonadism, osteoporosis, diabetes mellitus, and atherosclerotic cardiovascular disease. The patients with this syndrome are also at risk to develop epithelial neoplasms, melanoma, thyroid cancer, and osteosarcoma. Patients with osteosarcoma present at an older age (fifth decade of life) and at atypical sites (426).

The Li-Fraumeni syndrome is a rare, autosomal dominant inherited disorder associated with genetic heterozygous germline R156H, R267Q, and R290H mutation in the TP53 tumor suppressor gene (260,330, 373,424). Germline mutations of TP53 on 17p13.1 are found in about 70% of families with this syndrome (266). This disorder is characterized by multiple primary neoplasms in children and young adults, particularly soft tissue sarcomas, osteosarcomas, breast cancer, brain tumors, and leukemias (267,281,330,370). In a recent study of multiple primary tumors in patients with osteosarcoma, Hauben et al. (286) contended that these patients are at relative risk of 2.4 to develop another malignancy compared to the general population. Furthermore, these investigators postulated that diagnosis of unusual subtype of osteosarcoma should draw attention to a possible genetic predisposition (Li-Fraumeni syndrome or other inherited cancer syndromes) of the respective patients.

The **retinoblastoma syndrome** consists of the malignant tumor of the retina originating from the embryonic neural retina associated with the following dysmorphic abnormalities: microcephaly, broad and prominent nasal bridge, hypertelorism, ptosis, protrud-



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Figure 2-93 Histopathology of a low-grade central osteosarcoma. A: The cellular stroma contains irregularly formed woven bone trabeculae (*right*), resembling fibrous dysplasia. Observe subperiosteal extension of the tumor (*left*) (hematoxylin and eosin, original magnification $\times 25$). **B:** At higher magnification, benign-appearing spindle-cell tumor tissue is seen with formation of primitive woven bone pseudotrabeculae containing roundish osteocytes (*center*). Also present are large vascular spaces (hematoxylin and eosin, original magnification $\times 25$). **C:** At high magnification, the malignant nature of the tumor is better appreciated. Hyperchromatic tumor cells exhibit enlarged nuclei, a feature not present in fibrous dysplasia (hematoxylin and eosin, original magnification $\times 400$). **D:** Enlarged atypical spindle cell exhibiting hyperchromatism and mitosis (*upper right*) is present in the stroma of fibrous dysplasia-like low-grade central osteosarcoma (hematoxylin and eosin, original magnification $\times 400$).

ing upper incisors, micrognathia, short neck, low-set ears, facial asymmetry, genital malformation, and mental retardation (235,423). Retinoblastoma is in 60% of patients nonhereditary and unilateral. However, 40% of the cases are inherited in an autosomal dominant manner with almost complete penetrance, and 25% of these patients present with bilateral tumors. The syndrome is caused by a genetic mutation found in the tumor suppressor gene *RB1* located on the long arm of chromosome 13 (13q14.1) (235). Germline mutations of one allele followed by a somatic mutation in the hereditary form or somatic mutations of both alleles in the nonhereditary form lead to loss of function of this tumor suppressor gene, hence tumor formation (235). Osteosarcoma is the most common second malignancy in patients with hereditary retinoblastoma (348). Furthermore, these mutations also increase the risk for developing an irradiation-induced secondary osteosarcoma (428,444).

The **Bloom syndrome**, also known as Bloom-German syndrome, is an autosomal recessive disorder characterized by congenital telangiectatic erythema of the face resembling lupus erythematodes, dolichocephaly with malar hypoplasia, sensitivity to sunlight, low birth weight and well-proportional dwarfism, immunoglobulin deficiency, limb abnormalities (including syndactyly, polydactyly, and clinodactyly), and propensity for development of malignant tumors, particularly osteosarcoma (264). This syndrome has been attributed to the functional alteration of DNA-helicase gene of the



Figure 2-94 Gnathic osteosarcoma. A: Oblique radiograph of the mandible shows mixed sclerotic and lytic lesion destroying the cortex associated with a spiculated sunburst type of periosteal reaction. **B:** The tumor composed of chondroblastic tissue destroys adjacent bone. Osteoid-producing malignant cells (*bottom right*) are also visible (hematoxylin and eosin, original magnification $\times 100$). **C:** In other areas, purely cellular tissue with a moderate degree of pleomorphism of the cells and nuclei is present. Plate-like bone formation (*lower right*) confirms the diagnosis of osteosarcoma (hematoxylin and eosin, original magnification $\times 50$). (Reprinted with permission from Prein J, Remagen W, Spiessl B, Uehlinger E. *Atlas of tumors of the facial skeleton.* Berlin: Springer-Verlag, 1986.)

RecQ-family, *BLM*, located on chromosome band 15q26.1 (224,292).

B. Intracortical Osteosarcomas

These are the rarest forms of osteosarcomas. Only 20 cases have thus far been reported (83,123,136, 222,252,279,285,290,302,333,334,344,345,352,385,399, 441,446), with an age range of 9 to 43 years (average 21 years) and a male predominance. The presenting symptom is pain, often associated with activity, local tenderness, and occasionally a mass. In some patients, a history of previous trauma has been elicited. The tumor involves the cortex, especially of the mid-tibia or femur, without extension into the medullary portion of the bone or the soft tissues. The radiographic presentation is that of a radiolucent lesion with surrounding cortical sclerosis. The size of the lesion varies from 1.0 to 4.2 cm (83,323). In some instances the lesion

mimics osteoid osteoma or intracortical osteoblastoma. Histologic studies show the tumor cells to be round or spindle-shaped, pleomorphic, with vesicular nuclei containing prominent nucleoli. Mitotic figures may be present. The bony matrix produced by malignant cells assumes a lace-like pattern with tumor osteoid. Low-grade intracortical osteosarcoma and a case associated with oncogenic rickets have also been reported (222,285,290).

C. Surface Osteosarcomas

The term "surface" (formerly "juxtacortical") is a general designation for a group of osteosarcomas that arise on the surface of a bone (311,329) (Fig. 2-97). In general, these lesions are much rarer (4%–10% of all osteosarcomas) and present a decade later than their intraosseous counterparts (410). The great majority of surface osteosarcomas are low-grade tumors (378), al-

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Figure 2-96 Synchronous multicentric osteosarcoma: magnetic resonance imaging (MRI). In another patient, MRI shows involvement of the left femur and distal right femur.



Figure 2-97 Variants of surface osteosarcomas.

though there are moderately and highly malignant variants (291,304,437). Low-grade surface osteosarcoma is usually termed parosteal osteosarcoma. The lesion arises on the outside surface of the bone, virtually never provoking a periosteal reaction (361,418,431). Histologically, the proliferating portion of the neoplasm consists of a fibrous stroma, considered to be derived from the outer fibrous periosteal laver. Dedifferentiated parosteal osteosarcoma is a high-grade variant of the former tumor (393,455). Periosteal osteosarcoma belongs to an intermediate grade of surface lesions (434). This lesion may cause periosteal response and usually consists mostly of cartilage (381). The lesion is believed to be derived from the inner, or cambium, layer of periosteum. High-grade surface osteosarcoma is a lesion located on the periphery of a bone. It has a histology identical to that of conventional medullary osteosarcoma and a similar metastatic potential (363,454,460).

Parosteal Osteosarcoma

This type of surface osteosarcoma, which accounts for about 4% of all osteosarcomas, is seen in patients in their third and fourth decades, and characteristically affects the posterior aspect of the distal femur (414) (Fig. 2-98). Other locations include the tibia, the fibula, and the proximal portions of the femur and humerus (331). Rarely are other bones affected (306,312). Because the tumor is slow growing and usually involves only the surface of the bone, the prognosis is much better than for the other types of osteosarcoma.

On radiologic studies, the neoplasm presents as a dense oval or spherical mass attached to the surface of the cortical bone (Fig. 2-99). The surface is lobulated and may be less dense than the base of the tumor (Fig. 2-100). The mass appears sharply delineated from the peripheral soft tissues. The medullary cavity is usually spared unless the lesion is of long duration. A linear, translucent cleft usually forms an incomplete separation of the bulk of the tumor from the underlying cortex (Fig. 2-101). Larger lesions, however, tend to encircle the host bone (361). Although these features can be adequately demonstrated by conventional radiography, CT (Fig. 2-102) or MRI (Fig. 2-103) is often necessary to determine the extent of cortical penetration and intramedullary invasion by tumor (295).

The histology of this tumor is characterized by a high degree of structural differentiation and consists of immature (woven) bone in different stages of maturation to lamellar bone (395). The lamellar trabeculae, which predominate in most areas, are separated by a bland, well-differentiated fibrous stroma composed of spindle cells. The latter show minimal cytologic atypia, are minimally pleomorphic, and rarely exhibit mitotic activity (Fig. 2-104A–C). The surface may be covered by hypercellular and slightly atypical cartilage mimicking osteochondroma (Fig. 2-104D). Cartilaginous tumor nodules may also be present within the lesion. Determining malignancy may present some difficulty, as the indolent nature of this tumor is also evidenced by lack of significant tumor hemorrhage or necrosis. Multiple sections are usually required to reveal regions of prominent



Figure 2-98 Parosteal osteosarcoma: skeletal sites of predilection, peak age range, and male-to-female ratio.

osteoblastic activity around bony trabeculae (432). Applying the grading criteria to parosteal osteosarcoma elaborated by Ahuja et al., most tumors are low-grade lesions (grade 1 and 2) that have a good prognosis, and require simple surgery for treatment (204,401,435).

Genetically, parosteal osteosarcomas lack the complex aberrations found in conventional osteosarcomas. Supernumerary ring chromosomes containing chromosome 12 material have been found, as well as coamplification of *CDK4*, *SAS*, and *MDM2*, which are located at 12q13–15 (417,458).

Dedifferentiated Parosteal Osteosarcoma

Dedifferentiated parosteal osteosarcoma, the more common variant of surface-based high-grade osteosarcomas, usually arises as a conventional low-grade parosteal osteosarcoma (455). Then, either spontaneously (in which case both low- and high-grade components can be identified histologically) or secondarily (in recurrences of inadequately resected low-grade parosteal tumors), the lesion undergoes transformation to a high-grade malignancy (367,452). The latter situation is more common. Unni (431) cites a dedifferentiation rate of 20% in such recurrences. Only exceptionally rare, as reported by Shuhaibar and Friedman (402), the tumor at initial presentation shows coexistence of both the low-grade and high-grade sarcomas. In radiologic and histologic studies, dedifferentiated parosteal osteosarcoma mimics the features of conventional parosteal osteosarcoma. There are, however, some characteristics of a high-grade sarcoma, such as radiographically identifiable cortical destruction and histologically identifiable pleomorphic tumor cells with hyperchromatic nuclei and a high mitotic rate. Crosssectional imaging reveals medullary involvement in 22% of patients and adjacent soft tissue invasion in 46% (361) (Fig. 2-105). Hence, the prognosis is much worse than for parosteal osteosarcoma.

Periosteal Osteosarcoma

This very rare type of osteosarcoma (accounting for 1%-2% of all osteosarcomas and 25% of all surface osteosarcomas) grows on the surface of a bone and is usually located in the midshaft of a long bone; it is most common in adolescence (283). Although the age range varies, most lesions occur during the second decade of life. There is a slight female preponderance, and the majority of lesions affect the tibia and femur (85%-95%) followed by the ulna and humerus (5%-10%) (351).

The radiologic characteristics were defined by de-Santos et al. (250). These include heterogeneous tumor matrix with calcified spiculations interspersed with areas of radiolucency representing uncalcified matrix; occasional periosteal reaction in the form of a Codman triangle (Fig. 2-106); thickening of the periosteal surface of the cortex at the base of the lesion, with sparing of the endosteal surface; extension of the tumor into the soft tissues; and sparing of the medullary cavity (Figs. 2-107 and 2-108).

By microscopy, these tumors are of intermediate grade malignancy and are composed mainly of lobu-*Text continues on page 118*





Figure 2-100 Parosteal osteosarcoma. A sclerotic, lobulated mass is attached to the posterior cortex of distal femur. The periphery of the tumor is slightly less dense than the base.



Figure 2-101 Parosteal osteosarcoma. A large, sclerotic tumor partially attached to the medial cortex of left femur displays inferiorly a radiolucent cleft (*arrows*).

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Figure 2-102 Parosteal osteosarcoma: computed tomography (CT). A: Contrast-enhanced CT section demonstrates that the medullary portion of the bone has not been invaded (same patient as in Fig. 2-99A). **B:** CT section through the attached portion of the tumor (same patient as in Fig. 2-101) shows lack of invasion of the medullary cavity.



Figure 2-103 Parosteal osteosarcoma: magnetic resonance imaging (MRI). A: Lateral radiograph shows a tumor (*arrow*) involving the posterior aspect of the medial femoral condyle in a 22-year-old woman. It is not possible to determine whether the cortex has been violated. **B:** Sagittal MRI (SE;TR 517, TE 20) demonstrates invasion of the medullary cavity (*arrows*). (Reprinted with permission from Greenspan A. Osteosarcoma. *Contemp Diagn Radiol* 1993;16:1–6.)



Figure 2-104 Histopathology of parosteal osteosarcoma. A: The tumor originates at the surface of cortical bone (van Gieson, original magnification $\times 25$). **B:** At higher magnification, parallel arranged trabeculae of woven bone are separated by a fibrous stroma containing irregularly shaped spindle cells (hematoxylin and eosin, original magnification $\times 100$) (*continued*).



Figure 2-104 Continued C: Spindle cells with enlarged hyperchromatic nuclei form tumor-bone (hematoxylin and eosin, original magnification ×400). **D:** On its surface, parosteal osteosarcoma may be covered by a cartilaginous cap resembling osteochondroma; however, subchondral areas contain spindle cells, and hematopoietic or fatty marrow is absent (hematoxylin and eosin, original magnification ×25).

lated chondroid tissue with moderate cellularity (Fig. 2-109). They characteristically exhibit large amounts of cartilage matrix undergoing calcification or endochondral ossification, as well as small amounts of fine lace-like osteoid surrounding malignant cells (Fig. 2-110). Areas of moderately differentiated osteoblastic osteosarcoma are also present. The ultrastructural features consist of round or polyhedral chondroblast-like cells. The nuclei are rounded and have prominent nucleoli, and the nuclear envelope shows discrete indentations. The cytoplasm contains conspicuous rough endoplasmic reticulum, lakes of glycogen, and lipid vacuoles. The cytoplasmic membrane displays scanty, thin projections. The cells are surrounded by an abundant intercellular matrix of low electron density, with thin fibrils and granules (336). The prognosis for periosteal osteosarcoma is somewhat better than for conventional osteosarcoma but worse than for parosteal osteosarcoma (395).

Only few genetic studies have been reported with complex chromosomal aberrations in four cases and a numerical gain of chromosome 17 in one case (229).

High-Grade Surface Osteosarcoma

A small percentage of surface osteosarcomas are highgrade tumors and they carry the same prognosis as conventional osteosarcoma. The less common of these lesions, high-grade surface osteosarcoma, accounting for less than 1% of all osteosarcomas in a Mayo Clinic study (455), exhibits radiologic features similar to those of parosteal or periosteal osteosarcoma (289) (Fig. 2-111). Men are affected more frequently than women (M/F = 1.6:1). The most frequent site for this tumor is the femur (46%), followed by the humerus (16%) (363). It is histologically indistinct from conventional osteosarcoma (Fig. 2-112) and, like the latter, it has a high potential for distant metastases (455). Although intramedullary extension of the tumor is uncommon, the cortex may be involved and pathologic fractures may occur.

D. Soft Tissue (Extraskeletal) Osteosarcomas

Osteosarcoma of the soft tissues (extraskeletal, extraosseous) is an uncommon malignant tumor of mesenchymal origin, with the capacity to form neoplastic osteoid, bone, and cartilage (217,327,384). Middleaged and elderly individuals are most often affected, with a mean age at presentation of 54 years (238,412), although at times this tumor may present in a younger individual (408). Extraskeletal osteosarcoma is considerably less common than osteosarcoma of bone, accounting for only 4% of all osteosarcomas (206). The preferential sites for this neoplasm are in the lower extremities and buttocks (262,278,319,382,403,459). Although most of these tumors are of a high-grade malignancy, few cases of a low-grade lesion have been reported (362). The lesion may also develop in a variety of soft tissues such as breast, lung, thyroid, renal capsule, urinary bladder, and prostate, and even within the mesentery and pelvic retroperitoneum (237). Occasionally, a soft tissue osteosarcoma arises after radiation therapy. Exceptionally it may arise in myositis ossificans (317).

The most common clinical presentation is that of a slowly enlarging mass, with or without pain. The radiographic hallmarks include a soft tissue mass with scattered, amorphous calcifications and ossifications. The mass exhibits a disorganized arrangement of osteogenic elements in the center of the tumor (Figs. 2-113 and 2-114A). If the mass develops in close proximity to bone, it may invade the cortex.

CT usually reveals a heavily mineralized soft tissue mass, occasionally with areas of necrosis (255). This modality may demonstrate better than radiography the *Text continues on page 127*



Figure 2-105 Dedifferentiated parosteal osteosarcoma. A 24-year-old woman presented with pain and a palpable mass above the popliteal fossa of 2 months' duration. Three years prior to the current complaints a parosteal osteosarcoma had been resected from her distal femur. **A:** Anteroposterior radiograph of the distal femur shows an aggressive type of periosteal reaction and a large soft tissue mass with foci of bone formation. **B:** Lateral radiograph shows in addition the remnants of the previously resected parosteal osteosarcoma (*arrow*). **C:** The proximal computed tomography (CT) section shows a surface tumor exhibiting bone formation and a large soft tissue mass with foci of tumor bone. At this level the bone marrow is not invaded. **D:** The more distal section reveals in addition the invasion of the medullary cavity, a feature not consistent with a conventional parosteal osteosarcoma. **E:** Coronal T1-weighted (SE, TR 600, TE 25) magnetic resonance imaging (MRI) demonstrates the extent of both intramedullary invasion and a soft tissue mass. **F:** Axial T2-weighted (SE, TR 2000, TE 90) MRI shows heterogeneous signal of a large soft tissue mass. At the level of this section the bone marrow is not involved by a tumor (*continued*).



Figure 2-105 *Continued* **G:** Low-power photomicrograph of dedifferentiated parosteal osteosarcoma shows cellular island (*top*) at the periphery of the tumor (hematoxylin and eosin, original magnification $\times 25$). **H:** At higher magnification observe poorly differentiated malignant cells forming tumor-osteoid (hematoxylin and eosin, original magnification $\times 200$).



D



Figure 2-107 Periosteal osteosarcoma. A: Frontal radiograph of the right knee of a 12-year-old girl with discomfort in the upper leg for 2 months shows poorly defined calcifications and ossifications in a mass attached to the surface of the lateral tibial cortex. **B:** Anteroposterior tomography shows the ossified mass. Although attached to the tibia proximally (*open arrow*), it is partially separated from the lateral cortex by a narrow radiolucent cleft (*arrows*), very similar to that seen in the juxtacortical myositis ossificans. **C:** Computed tomography (CT) section obtained through the distal part of the lesion shows the extent of the soft tissue mass. Note low attenuation of the tumor with scattered high-attenuation ossifications. **D:** CT section obtained through the proximal part of the lesion clearly demonstrates its attachment to the tibial cortex. The medullary cavity is not affected.







Figure 2-109 Histopathology of periosteal osteosarcoma. The superficial part of the tumor consists of cellular cartilage with increasing cell density at the proliferation interface with covering fibrous tissue, indicating its malignant character. Endochondral ossification is also present (hematoxylin and eosin, original magnification $\times 25$).



Figure 2-110 Histopathology of periosteal osteosarcoma. A: Low-power photomicrograph shows predominantly cartilaginous matrix, which is a major component of the tu-mor. Areas of osteoid formation (*lower left corner*) confirm the diagnosis (hematoxylin and eosin, original magnification \times 40). **B:** At higher magnification observe metachromatically stained cartilaginous matrix of the tumor (Giemsa, original magnification $\times 100$). C: At high magnification note scanty osteoid formation in the peripheral cellular parts of the tumor (hematoxylin and eosin, original magnification \times 200).



Figure 2-111 High-grade surface osteosarcoma. A: Lateral radiograph of the distal leg shows a tumor arising from the surface of the posterior tibial cortex in a 24-year-old man. Poorly defined ossific foci are seen within a large soft tissue mass. Note the similarity of this tumor to periosteal osteosarcoma (compare with Fig. 2-107). B: Computed tomography (CT) section demonstrates the extent of the lesion. Characteristically, the marrow cavity is not affected.



Figure 2-112 Histopathology of high-grade surface osteosarcoma. A: High cellularity and marked pleomorphism of the stromal cells together with infiltration of the skeletal muscle and vascular invasion are characteristic of this tumor. The histologic appearance is indistinguishable from that of high-grade central osteosarcoma (hematoxylin and eosin, original magnification $\times 100$). **B:** High cellularity, numerous mitoses, and areas of tumor-osteoid formation indicate high-grade tumor (hematoxylin and eosin, original magnification $\times 200$).



Figure 2-113 Extraskeletal (soft tissue) osteosarcoma. A: Lateral radiograph of the right knee of a 51-year-old woman shows a poorly defined soft tissue mass situated above the patella merging with the quadriceps muscle. Approximately in the center of the lesion amorphous calcifications and ossifications are noted. The suprapatellar bursa appears unaffected. **B:** Radiograph of the specimen of the tumor depicted in **A** demonstrates foci of ossification in the center of the tumor, surrounded by the radiolucent zone at the periphery ("reverse zoning"). (Reprinted with permission from Greenspan A, Steiner G, Norman A, et al. Case report 436. Osteosarcoma of the soft tissues of the distal end of the thigh. *Skeletal Radiol* 1987;16: 489–492.)

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Figure 2-114 Extraskeletal (soft tissue) osteosarcoma. A 68-year-old woman presented with a progressively enlarging soft tissue mass in the popliteal region of the right knee. **A:** Lateral radiograph shows a large soft tissue mass, sharply outlined in its distal extent, but poorly delineated at the proximal end. Calcifications and ossifications are present throughout the tumor. **B:** One of the computed tomography (CT) sections through the distal portion of the tumor shows predominantly peripheral mineralization of the mass, very similar to that of myositis ossificans. **C:** The more proximal CT section clearly demonstrates reverse zoning, typical for soft tissue osteosarcoma. **D:** Axial T1-weighted (SE, TR 700, TE 19) magnetic resonance imaging (MRI) shows the mass exhibiting predominantly low-intensity signal. A smaller extension of the tumor is seen anterolaterally (*continued*).



Figure 2-114 *Continued* **E:** Axial fast spin-echo (FSE, TR 5200, TE 102 Ef) MRI reveals heterogeneity of the tumor, displaying variation of signals, from high to intermediate intensity. Coronal **(F)** and sagittal **(G)** inversion recovery images further reveal the internal septa and heterogeneous character of the tumor. **H:** Histologic (whole-mount) section of extraskeletal osteosarcoma in another patient shows reversed zonal phenomenon. Peripheral blue areas represent cellular component of the tumor, and the central area is occupied by plate-like tumor-osteoid (hematoxylin and eosin, whole-mount section). **I:** Centrally located tumor-matrix that contains collagen stains bright red (van Gieson, whole-mount section).



Figure 2-114 Continued J: Low-power magnification shows cellular tumor tissue containing tumor-bone beneath the skin (*right*) (Giemsa, original magnification \times 6). **K:** Plate-like tumor-osteoid is present adjacent to cellular areas of the tumor containing some osteoclast-like giant cells (hematoxylin and eosin, original magnification \times 200). **L:** Periphery of the tumor exhibits increased cellularity (*left*) and central bone formation (hematoxylin and eosin, original magnification \times 25). **M:** At higher magnification observe cell-rich peripheral part of the tumor with fibrosarcomatous areas and tumor-osteoid formation (hematoxylin and eosin, original magnification \times 100).

pattern of central ossification or the reverse zoning phenomenon (377) (Fig. 2-114B, C). It also demonstrates a lack of attachment of the mass onto the bone. MRI usually reveals a mass with heterogeneous low signal intensity on T1-weighted images (Fig. 2-114D) and heterogeneous but predominantly high signal intensity on T2weighted and inversion recovery sequences (443) (Fig. 2-114E–G). After injection of gadolinium (Gd-DTPA), noted is an intense enhancement of the tumor, and visualization of a pseudocapsule (255,308a). Scintigraphy using ^{99m}Tc-HMDP shows extraskeletal uptake of the radiopharmaceutical tracer within the tumor. Intense and heterogeneous uptake of ²⁰¹Tl in the extraskeletal osteosarcoma on both early and delayed images has recently been reported (308a).

The histopathology of soft tissue osteosarcoma is indistinguishable from that of a conventional osteosarcoma (449) (Fig. 2-114H–J). Only single cases have been investigated genetically, revealing moderate to complex aberrations as in conventional osteosarcoma (384).

Secondary Osteosarcomas

The term secondary applied to osteosarcoma refers to a tumor that arises in association with a preexisting lesion of the bone, which may or may not be neoplastic, such as pagetic bone, fibrous dysplasia, medullary bone infarct, chronic osteomyelitis, osteogenesis imperfecta, and metallic implants (200,282,352,425,433,439). Treatment-related postirradiation osteosarcoma also belongs to this group. In contrast to primary osteosarcomas, secondary lesions predominantly affect an older population. Whatever their etiology, the histopathology of secondary osteosarcomas is identical to that of pri-

mary osteosarcomas, and the prognosis for these tumors is equally ominous.

Paget Sarcoma

A large proportion of secondary sarcomas arise as a complication of Paget disease and characteristically develop in pagetic bone (339,347). Most cases of transformation to osteosarcoma occur in polyostotic Paget disease (394). Osteosarcoma can develop in any part of any bone affected by Paget disease (409), although the pelvic bones and femur are the preferential sites for this complication (339). The other sites commonly affected are the humerus and the tibia (371). Patients with advanced disease are at greater risk (347,394,447). The radiographic changes typically observed in malignant transformation of Paget disease include a destructive lesion in the affected bone, the presence of tumor bone in the lesion, pathologic fracture, and an associated soft tissue mass (411) (Figs. 2-115, 2-116, and 2-117). Periosteal reaction is only rarely encountered.

On microscopy, most Paget sarcomas are histologically conventional, high-grade osteoblastic intramedullary osteosarcomas (Fig. 2-118). Other types of osteosarcoma, such as fibrohistiocytic, chondroblastic, and fibroblastic, are also commonly observed. Osteosarcoma in these patients must be differentiated from metastases to pagetic bone from primary carcinoma elsewhere in the body, most commonly the prostate, breast, or kidney (see later). Nevertheless, metastases are a most uncommon event in Paget disease, despite the increased vascularity of pagetic bone (231).

Familial occurrence of Paget disease and secondary osteosarcoma have been reported by Wu et al. (457). Recently, susceptibility genes for Paget disease have been linked to chromosome 5q35 (293,325) and chromosome 18q (239). In a LOH-analysis of sporadic osteosarcomas and osteosarcomas in Paget disease, Nellissery et al. (356) identified a putative tumor-suppressor gene that closely maps to the same region on 18q. This region was more precisely defined by Johnson-Pais et al. to a 450kb sequence on 18q21.33 excluding *RANK* (coding for Receptor Activator of Nuclear Factor Kappa B, a cell membrane receptor involved in osteoclast formation) and *BCL2* (coding for an anti-apoptotic protein initially found in B-cell lymphoma) as candidate genes (307).

Osteosarcoma Arising in Fibrous Dysplasia

Osteosarcoma developing in fibrous dysplasia is a rare event occurring in about 0.5% to 1% of patients with this disorder, most commonly (in descending order) in craniofacial bones, proximal femur, humerus, pelvis, tibia, and scapula. Clinical symptoms such as swelling and increasing pain may point to the onset of sarcomatous change (386). Of 18 cases reported from the Mayo Clinic, 12 had previous irradiation before developing osteosarcomas. Six of these 18 patients had polyostotic fibrous dysplasia (430). Radiologically, a developing soft tissue mass or bone destruction, best visible on MRI, may aid in establishing the diagnosis. Most osteosarcomas are high-grade lesions exhibiting the same histologic findings as in primary osteosarcomas.

Only single genetic analysis has been reported so far, revealing a normal karyotype for uninvolved bone and fibrous dysplasia-tissue by comparative genomic hybridization (CGH); however, gains and losses were seen in the associated osteosarcoma comparable to changes seen in conventional osteosarcoma (305).

Osteosarcoma Arising in Bone Infarct

Secondary osteosarcomas associated with medullary bone infarcts seem to be even rarer than sarcomas developing in fibrous dysplasia. About 50 patients with sarcomas arising in bone infarcts have been reported, diagnosed in order of frequency as MFHs, osteosarcomas, fibrosarcomas, and angiosarcomas (199,247,425). Clinically, the patients present with pain. On radiologic imaging, residues of a bone infarct in combination with a soft tissue mass and bone destruction are found (263).

Postirradiation Osteosarcoma

Radiation therapy for an unrelated previous condition also may give rise to secondary osteosarcoma (207). These malignancies account for 3% to 5% of all osteosarcomas and 50% to 60% of radiation-induced sarcomas. The estimated risk of developing osteosarcoma in irradiated bone is 0.03% to 0.8%. The threshold is approximately 800 to 1,000 cGy, but usually a dose of at least 3,000 cGy administered within 3 weeks is required (270). The latent period is between 4 and 42 years, with an average of 11 years (343). The criteria for diagnosis of postirradiation sarcoma include the following (46): (a) the initial disease and the postirradiation sarcoma must not be of the same histologic type, (b) the site of the new tumor must be within the field of irradiation, and (c) at least 3 years must have elapsed since the previous radiation therapy.

Postirradiation osteosarcomas may develop either after radiation therapy for benign bone lesions, such as fibrous dysplasia (Fig. 2-119) and giant cell tumor, or after irradiation of malignant processes in the soft tissues, such as breast carcinoma and lymphoma. These tumors may also develop as a result of ingestion and intraosseous accumulation of radioisotopes, as has been described in painters of radium watch dials. Differentiation of radiation-induced sarcoma from osteonecrosis secondary to irradiation may be difficult. Progression and pain militate in favor of sarcoma. The double-line sign (parallel low- and high-signal-intensity lines) revealed with MRI indicates osteonecrosis. The presence of a soft tissue mass on MRI indicates the development of an osteosarcoma. The viability of the lesion can be confirmed with gadolinium-enhanced MRI (225).

In contrast to sporadic osteosarcomas, GCH studies revealed more losses than gains in radiation-induced osteosarcomas, especially at 1p21–p31 (57% vs. 3%), although the mean number of aberrations was similar. An additional finding was gains at 8qter, which in sporadic osteosarcomas are associated with a poor prognosis (421,422).



Figure 2-115 Osteosarcoma complicating Paget disease. Anteroposterior radiograph of the right hip in a 66-year-old man who had extensive skeletal involvement by Paget disease shows destruction of the cortex of the ischial bone, associated with a soft tissue mass containing tumor bone.

Differential Diagnosis

Radiology

On the basis of the subtype of osteosarcoma and the radiographic appearance of different variants, several lesions should be considered in the differential diagnosis. As mentioned previously, the radiologic differential possibilities do not always correspond to the histologic differential diagnosis, and they may vary according to the subtype of osteosarcoma. In radiographic studies, the **conventional medullary osteosarcoma** should be differentiated mainly from *Ewing sarcoma* (Figs. 2-120)



Figure 2-116 Osteosarcoma complicating Paget disease. A: Anteroposterior radiograph of the left hip in a 73-year-old man with polyostotic Paget disease shows destructive lesion of the ilium associated with a pathologic fracture. A soft tissue mass containing tumor bone is seen inferiorly to the sacroiliac joint. **B:** Computed tomography (CT) section demonstrates to better advantage the pathologic fracture and a large soft tissue mass with central necrosis, imaged as lower attenuation areas.

B

and 2-121). *Ewing sarcoma* usually affects the diaphysis of a long bone or the flat bones, such as the innominate bone and scapula. The lesion is poorly defined and is characterized by a permeative or moth-eaten type of bone destruction (see Figs. 5-15A and 5-16). The periosteal reaction is usually that of an aggressive, onion-skin (lamellated) type, although occasionally a sunburst type may be encountered. A soft tissue mass is almost invariably present, and at times there may be a disproportion between a small bone lesion and a large soft tissue mass (see Figs. 5-15A, B and 5-16). Occasionally, however, Ewing sarcoma may exhibit an unusual *Text continues on page 133*



Figure 2-117 Paget sarcoma. A: Anteroposterior radiograph of the pelvis of a 70-year-old woman shows extensive involvement of the left ilium, pubis, and ischium by Paget disease. There is also destruction of the cortex and a large soft tissue mass accompanied by bone formation, typical findings for osteosarcoma. **B:** Computed tomography (CT) scan demonstrates the bone formation in the soft tissue mass more clearly (*arrows*).



Figure 2-118 Histopathology of Paget sarcoma. A: There is clear separation between deformed bone trabeculae pathognomonic for Paget disease (*right*) and osteosarcoma (*left*) (hematoxylin and eosin, original magnification ×15). **B:** Pagetic bone with giant osteoclasts and osteoblastic rimming at the surface of deformed trabeculae shows infiltration of intertrabecular spaces by osteoblastic osteosarcoma (hematoxylin and eosin, original magnification ×100). **C:** Pagetic bone with mosaic pattern is invaded by pleomorphic osteosarcoma producing scant osteoid (*left half, red dots*) (van Gieson, original magnification ×100). **D:** At higher magnification observe irregular osteoid deposition against dilated small vessels (hematoxylin and eosin, original magnification ×200).

Α



Figure 2-119 Postirradiation osteosarcoma. A: Lateral radiograph of the skull of a 35-year-old woman with polyostotic fibrous dysplasia, who 11 years before this examination underwent radiation treatment of the mandible, shows predominant involvement of the frontal bones and the base of the skull. **B:** Oblique radiograph of the mandible shows an expansive lytic lesion in the body of the left mandible, with partial destruction of the cortex (*arrow*). Biopsy confirmed an osteosarcoma.



Figure 2-120 Osteosarcoma resembling Ewing sarcoma. Anteroposterior **(A)** and lateral **(B)** radiographs of the midshaft of the left humerus in this 17-year-old boy show a lesion that exhibits a moth-eaten pattern of bone destruction associated with Codman triangle of periosteal reaction and an adjacent soft tissue mass, similar to that of Ewing sarcoma. Histopathologic examination was consistent with grade 2 fibroblastic osteosarcoma.



Figure 2-121 Osteosarcoma resembling Ewing sarcoma. Anteroposterior radiograph of the distal femur of a 7-yearold girl shows a lesion in the metaphysis of the distal femur that exhibits a destructive, permeative, and moth-eaten pattern very similar to that seen in Ewing sarcoma. There is no evidence of significant bone formation. Aggressive periosteal reaction, in the form of Codman triangle, is present at the proximal extension of the tumor *(arrow)*. Biopsy was consistent with a fibroblastic osteosarcoma.
amount of ossification (see Fig. 5-19), thus closely resembling an osteosarcoma. It is noteworthy that Ewing sarcoma is extremely rare in black subjects, a helpful point to remember in making the differential diagnosis. Osteosarcoma developing after skeletal maturity should be differentiated from chondrosarcoma, fibrosarcoma, and MFH. Osteosarcoma affecting an epiphysis must be differentiated from *clear cell chondrosarcoma*, chondroblastoma, and epiphyseal enchondroma (427). Some osteosarcomas with atypical radiologic features should be differentiated from osteoblastoma (419). The central low-grade osteosarcoma should be differentiated from fibrous dysplasia. This may be difficult because both lesions may show the same radiographic characteristics (211,300). The former lesion is expansive; the cortex is thinned, frequently without signs of invasion; and the medullary component may have a narrow zone of transition with a ground-glass appearance (405) (see Figs. 2-91 and 2-92). CT and MRI are not helpful in this respect. However, neither periosteal reaction nor cortical destruction is present in fibrous dysplasia. For telangiectatic osteosarcoma the differential diagnosis includes fibrosarcoma, MFH, giant cell tumor, and aneuryssome mal bone cyst. Because telangiectatic osteosarcomas may closely resemble aneurysmal bone cyst on both imaging and histopathologic examination (389), it is of paramount importance to distinguish these two lesions. Although on radiography both lesions may show geographic pattern of bone destruction and narrow zone of transition, telangiectatic osteosarcoma more commonly will exhibit a wide zone of transition and aggressive type of periosteal reaction. In addition, most of the aneurysmal bone cysts will show a well-organized buttress of a new bone formation at the periphery of the lesion (see Figs. 7-31 and 7-32). MRI is usually not helpful, because heterogeneous signal intensity on T1 weighting with interspersed high-intensity areas (indicating presence of methemoglobin) and high signal intensity with several cystic foci and fluidfluid levels on T2-weighted images are present in both lesions. Some investigators, however, contended that an intact rim of low-intensity signal surrounding the lesion indicates a benign process (464). Multicentric osteosarcoma must be differentiated from metastatic osteosarcoma or other osteoblastic metastases. Finally, parosteal osteosarcoma should be differentiated among others from juxtacortical osteoma (see Fig. 2-2), soft tissue osteosarcoma (see Fig. 2-114A), sessile osteochondroma (see Fig. 3-39B), parosteal lipoma with ossifications (see Fig. 2-11), and well-matured juxtacortical myositis ossificans that became attached to the cortex (see Fig. 2-9C). The most common confusion in diagnosis involves the differentiation of this tumor from myositis ossificans (360) and sessile osteochondroma. Unlike parosteal osteosarcoma, myositis ossificans exhibits the distinctive radiologic features of a zonal phenomenon and a cleft that completely separates the ossific mass from the cortex (209,358) (see Fig. 2-9A, B). Osteochondroma, on the other hand, demonstrates uninterrupted cortical continuity and medullary communication between the lesion and the host bone (see Fig. 2-8), features that

are not present in parosteal osteosarcoma. In addition, osteochondromas usually contain fatty or even hematopoietic marrow in their intertrabecular spaces, features that can be demonstrated with MRI. Recent reports suggest that PET scanning may be useful in the ambiguous cases. ¹⁸F-labeled 2-fluoro-2-deoxyglucose (¹⁸FDG) PET can be used to determine cellular metabolic activity of the lesion. A high standardized uptake value (SUV) focus would suggest parosteal osteosarcoma rather than myositis ossificans (236).

Periosteal osteosarcoma may resemble *myositis ossificans* and *periosteal chondrosarcoma* (456). Based only on radiologic examination, differential diagnosis may be difficult. However, together with the young age of the patients, the radiologic presence of calcification and ossification in the base of the lesion (see Figs. 2-106, 2-107, and 2-108) and the histologic identification of at least some osteoid or bone formation by tumor cells are helpful in differentiating this tumor from periosteal chondrosarcoma (434).

Rarely, a **high-grade surface osteosarcoma** can be mimicked by so-called *gossypiboma*, a cotton-induced pseudotumor consisting of a foreign body granuloma. On both CT and MRI, the latter lesion may have a confusing appearance that suggests malignancy. Scintigraphy with thallium-201 is the most effective technique in such instances because, unlike in malignancy, the study will show no uptake of radiotracer in the mass. Exceptionally, as reported by Vanel et al. (440), high-grade surface osteosarcoma may mimic a *periosteal osteosarcoma*.

Paget osteosarcoma must be differentiated from *bone metastases* such as from primaries in the breast, prostate, or kidney, localized either in an area of pagetic bone or in unaffected bone (409). The metastatic foci may be either lytic (Fig. 2-122) or blastic (Fig. 2-123).

Postirradiation osteosarcoma should be differentiated from *postirradiation osteonecrosis*. MRI is useful in this respect showing so-called double-line sign characteristic for osteonecrosis, whereas the presence of soft tissue mass adjacent to the lesion indicates malignancy.

In the differential diagnosis of **extraskeletal osteosarcoma** the possibilities include *myositis ossificans, synovial sarcoma, extraskeletal chondrosarcoma, liposarcoma of soft tissues* with ossification, and *pseudomalignant osseous tumor* of soft tissue.

Myositis ossificans is a benign reactive lesion of the soft tissues, predominantly observed in adolescents and young adults (251). The zoning phenomenon represents the maturation pattern of the lesion, which is undifferentiated and cellular in the center, and exhibits an increasingly mature ossification toward the periphery; this is the histologic hallmark of this condition (203,358). On radiography the zoning phenomenon is characterized by a radiolucent center of the lesion and a more dense and sclerotic periphery (Fig. 2-124) (271). A radiolucent cleft often separates the mass from the adjacent cortex (Fig. 2-125). The evolution of myositis ossificans can be correlated well with the lapse of time since the trauma (320,358) (see also the



Figure 2-122 Paget disease complicated by lytic metastases. Anteroposterior radiograph of the pelvis of a 55-year-old woman shows extensive osteolytic destruction of the right ilium, ischium, and pubis secondary to metastatic renal cell carcinoma (hypernephroma). Note the characteristic features of Paget disease affecting the left hemipelvis. This metastatic lesion should not be mistaken for Paget sarcoma.



Figure 2-123 Paget disease complicated by blastic metastases. A 79-year-old man was diagnosed with prostate cancer. The anteroposterior radiograph of the lower pelvis and proximal femora shows classic features of Paget disease consisting of thickening of the cortex and coarse trabecular pattern of cancellous bone. Scattered foci of increased bone density represent osseous metastases.



Figure 2-124 Juxtacortical myositis ossificans. A coneddown radiograph of the right ribs shows a large ossific mass in the soft tissues. Note characteristic pattern of zoning phenomenon (i.e., increased density on the periphery of the lesion and more radiolucent center) and the narrow radiolucent cleft separating the mass from the ribs. (Courtesy of Dr. Alex Norman, New York, New York.)



Figure 2-125 Juxtacortical myositis ossificans. **A:** Oblique radiograph of the right shoulder shows an ossified lesion abutting lateral cortex of the proximal humeral shaft. **B:** Computed tomography (CT) section demonstrates the zoning phenomenon of myositis ossificans. The cleft that separates the lesion from the cortex is shown to the better advantage. **C:** Axial T1-weighted (SE, TR 600, TE 20) magnetic resonance imaging (MRI) shows the center of the lesion to be of high signal intensity, whereas the periphery exhibits low-to-intermediate signal.

previous discussion on differential diagnosis of juxtacortical periosteal osteosarcoma).

Synovial sarcoma is a lesion with a predilection for younger individuals (13 to 55 years) (259). It is usually located near a joint, especially in the lower extremities and particularly in the area around the knee. The radiologic appearance is that of a lobulated mass, with amorphous calcifications present in 25% of cases. Ossification is extremely rare in the mass of synovial sarcoma (445). In approximately 15% to 20% of patients, a periosteal reaction and/or erosion of adjacent bone structures is noted (see Fig. 9-34). Osteoporosis of the affected limb may be present secondary to disuse.

Chondrosarcoma of soft tissue, a very rare malignant tumor, is much less common than extraskeletal osteosarcoma. It is characterized by a soft tissue mass with ring-like, punctate calcifications. It can be radiographically distinguished from osteosarcoma of soft tissues by the lack of bone formation.

Liposarcoma of soft tissues tends to affect older adults, with a higher incidence in men. The lesion may closely mimic osteosarcoma of soft tissues, particularly when ossification is present. The type of ossification, however, is usually more organized, unlike that observed in osteosarcoma of soft tissues, and fatty tissue can usually be identified. The thigh, leg, and gluteal region are commonly affected. Growth of the liposarcoma may be exceedingly slow, over many years, and erosion of adjacent bone is a common feature (259).

Pseudomalignant osseous tumor of soft tissues was first described by Jaffe and later by Fine and Stout (262). These lesions are rare, are more common in females, and are located in the muscles and subcutaneous tissues. They are probably of infective origin, although

this has not been unequivocally confirmed. Some lesions may represent unrecognized foci of myositis ossificans (212,287).

С

Pathology

From the histopathologic standpoint, the differential diagnosis of osteosarcoma should be based on three aspects. First, the differences in histologic and cytologic appearances of malignancy and consecutive grading of the tumor have to be taken into account. Second, the location within a given bone (whether central, intracortical, or surface) is of importance. Finally, different forms of histologic differentiation of osteosarcoma and similarities and differences compared to the other tumors must be considered.

Histology and Cytology

Conventional medullary osteosarcoma (grade 3) represents the overwhelming majority of all cases, and it is this type that gives rise to the poor prognosis for osteosarcomas in general. Its different histologic types have been discussed previously. Because grade 2 tumors still display unequivocal signs of cellular and nuclear malignancy, in most cases the diagnosis is not difficult. Grade 1 tumors, however, exhibit only discrete signs of malignancy; therefore, in the differential diagnosis certain benign tumors and tumor-like lesions must be considered (Fig. 2-126). One of the tumors that must be distinguished from this malignancy is osteoblastoma. Although in most cases the distinction between osteosarcoma and osteoblastoma is straightforward, some osteosarcomas may resemble osteoblastomas (162,226), and conversely, some osteoblastomas (in particular, the epithelioid variant formerly called aggressive osteoblastoma) bear some resemblance



Figure 2-126 Histopathology of low-grade central osteosarcoma, resembling fibrous dysplasia. A: Irregular woven bone trabeculae in a moderately cellular stroma resemble those of fibrous dysplasia; however, observe infiltration of fatty marrow not present in the latter condition (hematoxylin and eosin, original magnification \times 200). B: In another field of view, the fibrous component shows pleomorphism, hyperchromatic nuclei, and mitoses (hematoxylin and eosin, original magnification \times 200).

to osteosarcoma (see Fig. 2-64D) (13,187). Probably the best distinction is the extremely rare formation of cartilage by osteoblastoma (13,161,171,345), and especially the lack of infiltration of the host bone by these tumors (13).

Δ

Another good example of this situation is a rare type of central osteosarcoma with a low grade of malignancy, which is well-differentiated, low-grade central osteosarcoma (436). The histologic picture of this tumor resembles fibrous dysplasia, osteofibrous dysplasia (Kempson-Campanacci lesion), adamantinoma, desmoplastic fibroma, and other lesions (see previous section). Very discrete focal signs of cellular atypia found in this variant may distinguish this lesion from the previously mentioned benign conditions (Fig. 2-127). In addition, invasion of bone marrow and permeation of trabeculae of cancellous and cortical bone by the tumor cells are constant and important features of low-grade osteosarcoma. Desmoplastic fibroma may infiltrate surrounding trabeculae of bone at their periphery, creating a similar picture to that of well-differentiated osteosarcoma (13). However, in the central portions of desmoplastic fibroma there will be no osteoid present and there will be no evidence of tumor bone production. Another low-grade tumor, intracortical osteosarcoma, should also be differentiated from *fibrous dys*plasia and osteofibrous dysplasia. In this case, the cytologic indicators of malignancy are usually more pronounced. However, pseudoanaplastic cytologic features do exist in benign lesions such as osteoblastoma, fibrous dysplasia, and nonossifying fibroma, probably related to degenerative nuclear changes. Careful search for mitoses, which are generally rare in the latter lesions in contrast to the true malignant tumors, is mandatory (160). In general, for therapeutic purposes it is essential to differentiate grade 1 tumors from those of higher grades because only the latter are likely to respond to chemotherapy.

Location

Within a given bone, osteosarcoma may be located centrally (i.e., in the medullary portion of bone), in the cortex, or beneath or outside the periosteum (juxtacortical). Therefore, the radiologic findings are of utmost importance and must always be compared with the histologic sections. An osteosarcoma on the bone surface must be differentiated from several other entities. Periosteal osteosarcoma is usually of a grade 2 malignancy. It differs from *periosteal chondrosarcoma* by the admixture of tumor bone within the mostly cartilaginous tumor matrix (Fig. 2-128). Fine lace-like osteoid can be demonstrated among the islands of chondroid tissue. In contrast, chondrosarcoma is purely chondroblastic and may show some reactive bone formation, but there is no tumor bone apposed to malignant cells (see Fig. 3-112). Fracture callus is occasionally difficult to distinguish from surface osteosarcoma because during the healing stage there is pronounced osteogenesis, and new cartilage and bone formation of callus may mimic tumor bone (Fig. 2-129). In addition, minimal histologic and cytologic signs suggesting malignancy may be present. Therefore, radiography is helpful and the radiographic appearance should take precedence over the histologic findings.

High-grade surface osteosarcoma is a very rare tumor. It is a grade 3 malignancy, corresponding to the high-grade osteosarcoma of the central type. Therefore, it is easier to differentiate this variant from other surface lesions, such as periosteal osteosarcoma and fracture callus. **Parosteal osteosarcoma**, on the other hand, is usually a low-grade tumor. The bone trabeculae usually display a regular alignment and a transformation from woven into lamellar bone. This tumor



Α

Figure 2-128 Histopathology of periosteal osteosarcoma and periosteal chondrosarcoma. A: In periosteal osteosarcoma, the tumor consists of a superficial layer of tumor cartilage and, unlike the case in periosteal chondrosarcoma, a base of partly cancellous tumor bone. The border resembles the zone of endochondral ossification in enchondroma (hematoxylin and eosin, original magnification $\times 12$). B: In periosteal chondrosarcoma lobulated chondroblastic tumor tissue partly cellular, partly with matrix resembling hyaline cartilage and with scattered calcifications (*upper center*) is seen. There is no evidence, however, of osteoid formation (hematoxylin and eosin, original magnification $\times 12$).



B

Figure 2-129 Histopathology of fracture callus. A: Callus with irregular arrangement of fibrous cartilage (clearer areas) and woven bone (darker areas) resembles chondrosteoid of osteosarcoma. Osteoblastic rimming of trabeculae (upper left and lower right) and the presence of loose fibrous tissue in the marrow spaces helps in differentiation from low-grade osteosarcoma (hematoxylin and eosin, original magnification $\times 25$). **B**: Marrow at some distance from the fracture line exhibits small foci of necrosis (right) and new irregular trabeculae of lamellar bone filling former vascular sinus. These findings constitute an expression of low-grade vascular disturbances after fracture and are not found in osteosarcoma (hematoxylin and eosin, original magnification $\times 25$). **C:** The fracture line is seen crossing the newly formed bone (upper right to lower middle). New cancellous bone is present in the marrow space. Here the reparative character of the new bone is obvious (hematoxylin and eosin, original magnification $\times 25$).

should be differentiated from *myositis ossificans* (358), *Nora lesion* (357), and *sessile osteochondroma* (13).

Myositis ossificans is fairly easy to distinguish when the lesion has been resected as a whole. The maturation pattern of the bone starting in the periphery (more mature), with fibroblastic tissue in the center producing immature bone by metaplasia, is in sharp contrast to surface osteosarcoma, in which the bone formation starts in the center. In fact, the most prominent feature in the diagnosis of myositis ossificans is the recognition of this zoning pattern, which was first described by Ackerman (203). It consists of a tumor-like structuring in three areas, reflecting different degrees of cellular maturation (see Fig. 2-15). The central zone is composed of an undifferentiated mesenchyme with a high-grade mitotic activity (Fig. 2-130A); the intermediate zone is composed of a variable amount of unmineralized woven bone intermingled with fibroblasts and osteoblasts (Fig. 2-130B); and in the peripheral zone the osteoid undergoes calcification and is remodeled into mature lamellar bone (Fig. 2-130C). The histologic distinction of parosteal osteosarcoma from myositis ossificans is analogous to the radiologic zonal phenomenon. The heterotopic ossification of myositis ossificans matures in a centripetal fashion, the most mature portion of the lesion being the outermost. In parosteal osteosarcoma and higher grade malignant tumors, the outer, advancing portion of the lesion usually is the least mature. As for fracture callus, the problem with the microscopic diagnosis of myositis ossificans involves the interpretation of early stage biopsies when complete maturation has not yet occurred and the tissue resembles that of sarcoma because of slight signs of pleomorphism in the cartilage and the large basophilic osteoblasts. However, in myositis ossificans the cells lack anaplasia, even in the zone where mitotic figures are numerous (203). The other important feature of myositis ossificans is a rather orderly production of bone, especially at the periphery of the lesion (203,358).

Bizarre parosteal osteochondromatous proliferation (BPOP, Nora lesion) (341,357) is a rare lesion, regarded as a form of heterotopic ossification [although cytogenetic changes that have recently been described put in question the lesion's nonneoplastic nature (463)], and should not be mistaken for parosteal osteosarcoma. It most commonly affects the bones of the hands and feet, but the long bones are affected in about 25% of reported cases. Radiographs typically show a heavily calcified or ossified mass with a broad base attached to the underlying cortex, which is normal in appearance, although occasionally may show signs of invasion (288). The contour of the mass is usually smooth, but it may be slightly lobulated (Fig. 2-131A,B). In contrast to osteosarcoma, the mass is smaller, shows bizarre cartilage forma-





Figure 2-130 Histopathology of myositis ossificans. A: Central zone of myositis ossificans shows predominantly spindle cell tissue with a disorderly arrangement, producing collagen matrix (hematoxylin and eosin, original magnification \times 50). B: The intermediate zone reveals immature bone matrix formation. The cellularity of this tissue might cause concern and lead to an erroneous diagnosis of osteosarcoma (hematoxylin and eosin, original magnification \times 50). **C**: At the periphery of myositis ossificans, there is mature lamellar bone formation (hematoxylin and eosin, original magnification \times 12).

С

tion and irregular short bone trabeculae, and does not invade the adjacent soft tissues. Histologic examination reveals a large amount of hypercellular cartilage showing transformation to trabecular bone, with frequent spindle cells in the intertrabecular spaces (Fig. 2-131C). Binucleated cells are common and there may be a considerable osteoblastic activity. This appearance can lead to an incorrect diagnosis of parosteal osteosarcoma.

Occasionally, parosteal osteosarcoma must be differentiated from *osteochondroma*, because this tumor may contain chondroid areas arranged like a cap of an osteochondroma (see Fig. 2-104D). According to Dahlin and Unni (9), the presence of an intertrabecular spindle cell component in the former tumor is an important differential point.

Histologic Differentiation

As mentioned previously, high-grade central osteosarcoma is by far the most common form of osteosarcoma. It is defined as a tumor with cytologic signs of highgrade malignancy, which forms a tumor bone matrix in very different quantities, from very little, as in telangiectatic osteosarcoma, to abundant, as in the sclerotic osteoblastic form of medullary osteosarcoma.

In osteoblastic osteosarcoma the formation of woven bone occurs in a mostly primitive trabecular or more plate-like manner (see Fig. 2-81A). If bone matrix formation is very conspicuous, resembling that of compact bone (Fig. 2-132), and with a corresponding radiographic image of very high and homogeneous density, a multicentric osteosarcoma must be suspected and sought by use of various radiologic modalities. It should be kept in mind, however, that in the monostotic osteoblastic form of osteosarcoma the bone matrix is not necessarily mineralized and that this tumor may radiographically present as a lytic lesion. Giant cell reparative granuloma, or so-called solid aneurysmal bone cyst, may cause considerable difficulty in differentiation from an osteosarcoma of low or intermediate grade malignancy with moderate bone formation, because the fibroblasts may show some polymorphism and mitoses and the bone formation may appear to be quite irregular. However, atypical mitoses are never seen. The radiographic features are usually helpful. Fibrous dysplasia can be difficult to differentiate from low-grade osteosarcoma with a high grade of differentiation (see previous discussion). The lack of osteoblasts rimming trabeculae in fibrous dysplasia is helpful in making this distinction.

In addition to formation of osteoblastic matrix by osteosarcomas, other types of matrix formation may be found. Chondroblastic osteosarcoma usually displays a fairly well-differentiated cartilaginous matrix, often with only moderate cytologic signs of malignancy but without the typical lobular pattern and the hyaline cartilage of well-differentiated chondrosarcoma (366). A characteristic feature is the indistinct transition from the

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Figure 2-131 Nora lesion. Anteroposterior **(A)** and lateral **(B)** radiographs of the small finger show an ossific mass adjacent to the posteromedial cortex of the proximal phalanx. **C:** Hyaline cartilage (*top*) borders somewhat irregular bone trabeculae formed by endochondral ossification and containing remnants of cartilage matrix, closely resembling osteochondroma. Loose fibrous tissue within the marrow spaces contains wide sinusoid capillaries (hematoxylin and eosin, original magnification ×12).

cartilaginous matrix into bony matrix (Fig. 2-133), the latter pointing to the osteoblastic nature of the tumor. This substance with intermediate characteristics between cartilage and osteoid is called chondrosteoid by most authors (345), and it may be also present in fracture callus. Another tumor that must be differentiated from chondroblastic osteosarcoma is dedifferentiated chondrosarcoma, in which features of a high-grade osteosarcoma are found side by side with a well-differentiated chondrosarcoma (Fig. 2-134). The demarcation is very clearly delineated, unlike chondroblastic osteosarcoma, in which the delineation is not sharp and chondroblastic and osteoblastic matrices may merge into one another (see Fig. 2-133B). In clear cell chondrosarcoma bone formation may be substantial. However, this bone does not represent tumor bone but rather is reactive (Fig. 2-135). Furthermore, the cartilage cells are larger than those in conventional chondrosarcoma and chondroblastic osteosarcoma, and their cytoplasm ranges from perfectly clear to eosinophilic.

Fibroblastic osteosarcoma consists of a three-dimensional interwoven pattern of collagen fibers formed by malignant spindle cells of variable pleomorphism and quantity. This tumor must be differentiated from *fibrosarcoma*. In particular, areas of spindle cells and homogeneous eosinophilic material, as encountered in fibroblastic osteosarcoma, may be indistinguishable from fibrosarcoma (223), as the latter, especially its sclerosing epithelioid variant, occasionally contains areas of homogeneous eosinophilic material that can resemble osteoid (201,213,340). In this situation special stains such as van Gieson or Goldner can be helpful. On the other hand, the formation of tumor osteoid in osteosarcoma may be scant and must be carefully searched for (Fig. 2-136). Usually, the typical herring-bone pattern of true fibrosarcoma is either absent or indistinct, which may be helpful in distinguishing osteosarcoma sparse in osteoid from fibrosarcoma. *Malignant fibrous histiocytoma (MFH)* rarely shows any bone formation and in most cases is



Figure 2-132 Histopathology of osteoblastic osteosarcoma. In the marrow spaces around a trabecula (*lower center*), there are newly formed, irregular, and smaller tumor bone trabeculae, which are fully mineralized and are surrounded by tumor cells (non-decalcified preparation, Goldner trichrome, original magnification \times 50).

В



Α

Figure 2-133 Histopathology of chondroblastic osteosarcoma. A: Osteoblastic tumor tissue with moderate osteoid formation (delicate red structures) extending into the periosteum (dense, reddish diagonal structure in upper right corner) (van Gieson, original magnification $\times 25$). B: Marrow space with two old trabeculae (upper right and lower middle) and tumor chondrosteoid (osteoid and cartilage matrix merging into one another without clear delineation) is typical of many osteosarcomas (hematoxylin and eosin, original magnification \times 50).





Figure 2-134 Histopathology of dedifferentiated chondrosarcoma with an osteosarcoma component. A: An area of well-differentiated chondrosarcoma (upper right) is sharply demarcated from dedifferentiated tumor showing osteoblastic differentiation within remnants of cancellous bone (lower left) (hematoxylin and eosin, original magnification $\times 25$). **B:** Almost entirely cellular, dedifferentiated tumor tissue with scanty tumor bone formation and remnants of cancellous bone (*lower right*) (hematoxylin and eosin, original magnification \times 25). C: The dedifferentiated portion of tumor exhibits a delicate network of tumor osteoid (red) (van Gieson, original magnification $\times 25$).



Α

Figure 2-135 Histopathology of clear cell chondrosarcoma resembling osteosarcoma. A: A considerable amount of new reactive bone (not produced by tumor cells) makes differentiation from osteosarcoma almost impossible. Meticulous search for areas typical of clear cell chondrosarcoma, such as depicted in Figures 3-96 and 3-97, is essential (hematoxylin and eosin, original magnification \times 50). B: In other areas there is evidence of small woven bone trabecula formation, mimicking that of osteosarcoma. Identification of cartilage tumor cells is essential for the diagnosis of chondrosarcoma (hematoxylin and eosin, original magnification \times 50).

characterized by the cartwheel or storiform pattern of fibers and cell arrangement (Fig. 2-137). In the giant cell–rich variant of MFH, giant cells may be numerous, unlike fibrosarcoma and fibroblastic osteosarcoma, in which they are sparse or absent. In this case, distinction from the giant cell–rich variant of osteosarcoma may create some problems (219). Both giant cell–rich osteosarcoma and MFH may contain giant cells in equal numbers and forms, so that some authors consider MFH to represent only a variant of osteosarcoma. *Desmoplastic fibroma* is easy to differentiate from osteosarcoma because in the former the fibroblasts are monomorphic and very evenly distributed within the tissue, and no tumor bone formation is present (see Fig. 4-58). Telangiectatic osteosarcoma and giant cell-rich osteosarcoma resemble each other in their moderate content of giant cells but differ in the number and size of blood-containing spaces resembling blood vessels, which are conspicuous in the former. Both tumors usually contain very little bone. In differentiating *giant cell tumor* from these two malignancies, it is important to note that the number and size of giant cells in giant cell tumor are much greater than in the two mentioned variants of osteosarcoma, whereas bone formation in a giant cell tumor is rarely present and is usually scant. In some giant cell tumors, however, the number and size of the giant cells may be reduced to the point that makes differentiation extremely difficult. In this situation the radiologic findings are essential. Telangiectatic R

R



Figure 2-136 Histopathology of fibroblastic osteosarcoma resembling fibrosarcoma. A: Areas without bone formation are very much like those of high-grade fibrosarcoma (hematoxylin and eosin, original magnification $\times 25$). **B:** In another section of this tumor, the tumor tissue is composed of spindle cells with distinct pleomorphism (*upper right*). Formation of tumor woven bone trabeculae (*left*) points to the diagnosis of fibroblastic osteosarcoma (hematoxylin and eosin, original magnification $\times 25$).



Figure 2-137 Histopathology of malignant fibrous histiocytoma. A: Densely arranged spindle cells, showing considerable pleomorphism and hyperchromatism of the nuclei resembling a cartwheel-like pattern (*upper left*), may help to distinguish this tumor from fibroblastic osteosarcoma (hematoxylin and eosin, original magnification \times 50). **B:** In another area of this tumor, loose myxoid arrangement of tumor tissue (*middle*) may be helpful in the differential diagnosis (hematoxylin and eosin, original magnification \times 50).

osteosarcoma contains enlarged capillary vessels surrounded by highly pleomorphic tumor tissue and may also contain elements of a true aneurysmal bone cyst surrounded by normal connective tissue. Differentiation from *aneurysmal bone* cyst may be difficult in such instances. In fact the confusion between telangiectatic osteosarcoma and aneurysmal bone cyst represents one of the most treacherous pitfalls in tumor pathology (272,309) (Fig. 2-138). The hallmark of osteosarcoma is the presence of anaplastic cells with atypical mitoses that are not present in the aneurysmal bone cyst (Fig. 2-139). Some authorities suggested that morphometric studies may be of help in differentiating these two entities (387).

Small cell osteosarcoma must be differentiated from Ewing sarcoma, primitive neuroectodermal tumor (PNET), lymphoma, and mesenchymal chondrosarcoma (253). It is essential to look for a sparse bone formation in osteosarcoma (see Figs. 2-85, 5-25). However, in some Ewing sarcomas, reactive woven bone formation may be found, and this may be misinterpreted as tumor bone formation, thus causing an erroneous diagnosis of small cell osteosarcoma (43). Immunocytochemical methods and molecular analyses are helpful in distinguishing these tumors from one another, because osteosarcomas in contrast to Ewing sarcomas do not show a t(11;22) translocation as can be revealed by FISH. CD99 (Mic 2), however, expressed in almost all cases of Ewing sarcoma family, and in the past thought to be specific for Ewing tumor may also be positive in small cell osteosarcoma (415).

Mesenchymal chondrosarcoma may contain a population of small cells indistinguishable from that of small cell osteosarcoma. However, according to Fechner and Mills (13), mesenchymal chondrosarcoma has sharply demarcated nests of low-grade cartilage and small cell osteosarcoma contains osteoid. *Lymphoma* can be eliminated by the absence of panleukocytic markers such as CD45 [formerly known as leukocyte common antigens (LCA)], and *PNET of bone* will demonstrate evidence of neuroendocrine differentiation in the form of positive staining for neuron specific enolase, Leu 7, synaptophysin, chromogranin, or other neural markers, and may demonstrate Homer Wright-type rosettes (13).



Figure 2-138 Histopathology of aneurysmal bone cyst resembling telangiectatic osteosarcoma. A cystic area (*up-per*) is bordered by a seam of flat undifferentiated cells intermingled with some small giant cells. Immediately below there is formation of considerable amounts of primitive woven bone rimmed with osteoblasts. Bone formation extends far down into the solid fibrous tissue (*lower left*), which appears myxoid. Here several larger giant cells of the osteoclast type lie near the bone. Bone formation may be erroneously interpreted as osteosarcoma (hematoxylin and eosin, original magnification ×25).



Figure 2-139 Histopathology of telangiectatic osteosarcoma resembling aneurysmal bone cyst. A: A low-power photomicrograph shows vascular areas very similar to those characteristic of aneurysmal bone cyst (hematoxylin and eosin, original magnification \times 15). B: At high magnification solid areas of the tumor show significant pleomorphism of malignant cells and hyperchromatism of their nuclei. Also seen is osteoid production by the tumor cells (hematoxylin and eosin, original magnification \times 400).

Low-grade intraosseous osteosarcoma should be differentiated from fibrous dysplasia and desmoplastic fibroma. Bertoni et al. (220) reported two such tumors that on histopathologic examination exhibited irregular bony seams in a spindle cell stroma simulating "Chinese characters" or "psammoma-like" appearance of bone formation typical for *fibrous dysplasia*. Although in both low-grade central osteosarcoma and fibrous dysplasia the fibroblastic stroma has a bland appearance and variable amounts of bone production is present, one can usually separate these lesions (300). The most helpful histologic features for distinguishing these entities are the presence of permeation, subtle cytologic atypia, and mitotic activity in osteosarcoma. Furthermore, the spindle-shaped nuclei in the latter tumor tend to be longer and more slender than the short plump nuclei of fibrous dysplasia (300). Occasionally, a prominent spindle cell proliferation may be present in low-grade central osteosarcoma accompanied by scattered thin-walled gaping vessels and heavy collagenization as seen is *desmoplastic fibroma* (220). However, the distinguishing feature is a small amount of osteoid production that is generally present in osteosarcoma, but is absent in fibrous dysplasia.

Osteosarcomas secondary to Paget disease and those evolving in bone infarcts usually do not cause problems in differential diagnosis because the primary lesions have characteristic appearances. In some cases, however, these characteristics may be obscured by the tumor tissue and must then be meticulously sought. As always, correlation with radiologic imaging features, including studies obtained prior to tumor development, are very important.

The radiologic and pathologic differential diagnosis of various subtypes of osteosarcoma is depicted in Figures 2-140 through 2-147.





Figure 2-140 Radiologic and pathologic differential diagnosis of conventional osteosarcoma.

osteosarcoma.



Figure 2-142 Radiologic and pathologic differential diagnosis of telangiectatic osteosarcoma.



Figure 2-143 Radiologic and pathologic differential diagnosis of lowgrade central osteosarcoma.











Figure 2-146 Radiologic and pathologic differential diagnosis of periosteal osteosarcoma.





CHAPTER 2 Bone-Forming (Osteogenic) Lesions - 149

REFERENCES

Osteoma

- Baum PA, Nelson MC, Lack EE, Bogumill GP. Case report 560. Parosteal osteoma of tibia. *Skeletal Radiol* 1989;18:406–409.
- Bertoni F, Unni KK, Beabout JW, Sim FH. Parosteal osteoma of bones other than of the skull and face. *Cancer* 1995;75: 2466–2473.
- 3. Bronner MP. Gastrointestinal polyposis syndromes. Am J Med Genet 2003;122:335–341.
- Bui-Mansfield LT, Myers CP, Chew FS. Parosteal lipoma of the fibula. Am J Roentgenol 2000;174:1698.
- Bullough P. Orthopaedic pathology, 4th ed. Edinburgh: Mosby, 2004:363–397.
- Campbell CJ, Papademetriou T, Bonfiglio M. Melorheostosis. A report of the clinical, roentgenographic, and pathological findings in fourteen cases. *J Bone Joint Surg* 1968;50A:1281–1304.
- Cervilla V, Haghighi P, Resnick D, Sartoris DJ. Case report 596. Parosteal osteoma of the acetabulum. *Skeletal Radiol* 1990;19: 135–137.
- Chang CH, Piatt ED, Thomas KE, Watne AL. Bone abnormalities in Gardner's syndrome. Am J Roentgenol 1968;103: 645–652.
- Dahlin DC, Unni KK. Osteoma. In: Bone tumors. General aspects on 8,542 cases, 4th ed. Springfield, IL: Charles C Thomas, 1986:84–87, 308–321.
- Dolan K, Seibert J, Seibert R. Gardner's syndrome. Am J Roentgenol 1973;119:359–364.
- Eckel W, Palm D. Statistische und röntgenologische Untersuchungen zu einigen Fragen des Nebenhöhlenosteoms. Arch Ohr Nas Kehlkopfheilk 1959;174:440–457.
- Enzinger FW, Weiss SW. Osseous tumors and tumor-like lesions of soft tissues. In: Soft tissue tumors, 2nd ed. St. Louis: Mosby, 1988:882–905.
- Fechner RE, Mills SE. Atlas of tumor pathology. Tumors of the bones and joints. Washington, DC: Armed Forces Institute of Pathology, 1993:25–77.
- Fleming RJ, Alpert M, Garcia A. Parosteal lipoma. Am J Roentgenol 1962;87:1075–1084.
- Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:225–226.
- Freyschmidt J, Ostertag H, Jundt G. Knochentumoren. Klinik, Radiologie, Pathologie, 2nd ed. Berlin: Springer-Verlag, 1998: 91–104.
- Gardner EJ, Richards RC. Multiple cutaneous and subcutaneous lesions occurring simultaneously with hereditary polyposis and osteomatosis. *Am J Hum Genet* 1953;5:139–147.
- Gardner EJ, Plenk HP. Hereditary pattern for multiple osteomas in a family group. *Am J Hum Genet* 1952;4:31–36.
- Gentry JF, Schechter JJ, Mirra JM. Case report 574. Periosteal osteoblastoma of rib. *Skeletal Radiol* 1989;18:551–555.
- Geschickter CF, Copeland MM. Parosteal osteoma of bone: a new entity. Ann Surg 1951;133:790–807.
- 21. Greenspan A. Benign bone-forming lesions: osteoma, osteoid osteoma, and osteoblastoma. *Skeletal Radiol* 1993;22:485–500.
- Greenspan A. Sclerosing bone dysplasias—a target-site approach. Skeletal Radiol 1991;20:561–583.
- Greenspan A. Orthopedic imaging: a practical approach, 4th ed. New York: Lippincott Williams & Wilkins, 2004:571–575, 584–594.
- Houghton MJ, Heiner JP, DeSmet AA. Osteoma of the innominate bone with intraosseous and parosteal involvement. *Skeletal Radiol* 1995:24:445–457.
- Huvos AG. Bone tumors: diagnosis, treatment, and prognosis, 2nd ed. Philadelphia: WB Saunders, 1991.
- Jacobs P. Parosteal lipoma with hyperostosis. *Clin Radiol* 1972;23:196–198.
- Jacobson HG. Dense bone—too much bone: radiological considerations and differential diagnosis. Part I. Skeletal Radiol 1985;13:1–20.
- Jacobson HG. Dense bone—too much bone: radiological considerations and differential diagnosis. Part II. Skeletal Radiol 1985;13:97–113.

- Krolls SO, Jacoway JR, Alexander WN. Osseous choristomas (osteomas) of intraoral soft tissues. *Oral Surg* 1971;32:588–595.
- 30. Lichtenstein L. Bone tumors, 5th ed. St Louis: CV Mosby, 1977:11.
- Lichtenstein L, Sawyer WR. Benign osteoblastoma: further observations and report of twenty additional cases. *J Bone Joint Surg Am* 1964;46A:755–765.
- Meltzer CC, Scott WW Jr, McCarthy EF. Case report 698. Osteoma of the clavicle. *Skeletal Radiol* 1991;20:555–557.
- Mirra JM, Gold RH, Pignatti G, Remotti F. Case report 497. Compact osteoma of iliac bone. *Skeletal Radiol* 1988;17:437–442.
- Mirra JM, Picci P, Gold RH. Bone tumors: clinical, pathologic, and radiologic correlations. Philadelphia: Lea and Febiger, 1989: 226-248.
- 35. Murphey MD, Johnson DL, Bhatia PS, et al. Parosteal lipoma: MR imaging characteristics. *Am J Roentgenol* 1994;162:105–110.
- O'Connell JX, Rosenthal DI, Mankin HJ, Rosenberg AE. Solitary osteoma of a long bone. J Bone Joint Surg 1993; 75A: 1830–1834.
- Peyser AB, Makley JT, Callewart CC, et al. Osteoma of the long bones and the spine: a study of eleven patients and a review of the literature. *J Bone Joint Surg Am* 1996;78A:1172–1180.
- Pool JL, Pontanos JN, Krueger EG. Osteomas and mucoceles of the frontal paranasal sinuses. J Neurosurg 1962;19:130–135.
- Prein J, Remagen W, Spiessl B, Uehlinger E. Atlas of tumors of the facial skeleton. Berlin: Springer-Verlag, 1986:99–101.
- Ramos A, Castello J, Sartoris DJ, et al. Osseous lipoma: CT appearance. *Radiology* 1985;157:615–619.
- Sadry F, Hessler C, Garcia J. The potential aggressiveness of sinus osteomas. A report of two cases. *Skeletal Radiol* 1988;17: 427–430.
- 42. Saglik Y, Kendi TK, Yildiz HY, et al. Clavicular osteoma associated with bronchial osteomas. *Skeletal Radiol* 2004;33:234–236.
- Schajowicz F. Tumors and tumorlike lesions of bone: pathology, radiology and treatment, 2nd ed. Berlin: Springer-Verlag, 1994:30–32, 52–56, 406–411.
- Schajowicz F, Lemos C. Osteoid osteoma and osteoblastoma closely related entities of osteoblastic derivation. *Acta Orthop Scand* 1970;41:272–291.
- Schweitzer ME, Greenway G, Resnick D, et al. Osteoma of soft parts. Skeletal Radiol 1992;21:177–180.
- 46. Spjut HJ, Dorfman HD, Fechner RE, Ackerman LV. Tumors of bone and cartilage. In: Firminger HI, ed. *Atlas of tumor pathology*, 2nd series, fascicle 5. Washington, DC: Armed Forces Institute of Pathology, 1971:117–119.
- 47. Spencer MG, Mitchell DB. Growth of a frontal sinus osteoma. *[Laryngol Otol* 1987;101:726–728.
- Steinberg I. Huge osteoma of the eleventh left rib. JAMA 1959;170:1921–1923.
- Stern PJ, Lim EVA, Krieg JK. Giant metacarpal osteoma. A case report. J Bone Joint Surg Am 1985;67A:487–489.
- Sundaram M, Falbo S, McDonald D, Janney C. Surface osteomas of the appendicular skeleton. Am J Roentgenol 1996;167: 1529–1533.
- Tamarito LV, Pardo J. Parosteal osteoma: a clinicopathological approach. *Pathol Annu* 1977;1:373–387.
- Unni KK. Dahlin's bone tumors. General aspects and data on 11,087 cases, 5th ed. Philadelphia: Lippincott-Raven, 1996:117–120.
- Unni KK, Dahlin DC, Beabout JW, Ivins JC. Parosteal osteogenic sarcoma. *Cancer* 1976;37:2644–2675.
- Wilner D. Radiology of bone tumors and allied disorders. Philadelphia: WB Saunders, 1982:629–638.
- Blank N, Lieber A. The significance of growing bone islands. Radiology 1985;85:508–511.

Enostosis (Bone Island)

- Brien EW, Mirra JM, Latanza L, et al. Giant bone island of femur. Case report, literature review, and its distinction from low grade osteosarcoma. *Skeletal Radiol* 1995;24:546–550.
- Broderick TW, Resnick D, Georgen TG, Alazraki N. Enostosis of the spine. *Spine* 1978;3:167–169.
- Bullough PG. Atlas of orthopedic pathology with clinical and radiologic correlations, 2nd ed. New York: Gower Medical Publishing, 1992:14.15–14.16.

- Caffey J. Focal sclerosis of spongiosa. Multiple generalized and scattered normal variants. In: *Pediatric X-ray diagnosis*, 6th ed., vol. 2. Chicago: Year Book Medical Publishers, 1972:960–963.
- Chew FS. Benign bone-forming tumors. Contemp Diagn Radiol 2001;24:1–6.
- 61. Davies JAK, Hall FM, Goldberg RP, et al. Positive bone scan in a bone island. *J Bone Joint Surg Am* 1979;61A:943–945.
- Dorfman HD, Czerniak B. Sclerosing bone lesions. In: Dorfman HD, Czerniak B, eds. *Bone tumors*. St. Louis: Mosby, 1997: 1087–1110.
- 63. Ehara S, Kattapuram SV, Rosenberg AE. Giant bone island. Computed tomography findings. *Clin Imaging* 1989;13:231–233.
- Go RT, El-Khoury GY, Wehbe MA. Radionuclide bone image in growing and stable bone island. *Skeletal Radiol* 1980;5:15–18.
- Gold RH, Mirra JM, Remotti F, Pignatti G. Case report 527. Skeletal Radiol 1989;18:129–132.
- Greenspan A, Klein MJ. Giant bone island. Skeletal Radiol 1995;24:59-60.
- 67. Greenspan A, Steiner G, Knutzon R. Bone island (enostosis): clinical significance and radiologic and pathologic correlations. *Skeletal Radiol* 1991;20:85–90.
- Greenspan A. Bone island (enostosis): current concept. Skeletal Radiol 1995;24:111–115.
- Greenspan A, Stadalnik RC. Bone island: scintigraphic findings and their clinical application. *Can Assoc Radiol J* 1995;46:368–379.
- Hall FM, Goldberg RP, Davies JA, Faisinger MH. Scintigraphic assessment of bone islands. *Radiology* 1980;135:737–742.
- Hoffman RR Jr, Campbell RE. Roentgenologic bone island instability in hyperparathyroidism. *Radiology* 1972;103:307–308.
- 72. Kim SK, Barry WF. Bone islands. Radiology 1968;99:77-78.
- 72a. Kim SK, Barry WF. Bone island Am J Roentgenol 1964;92: 1301–1306.
- 72b. Meschan I. Normal variant sclerotic bone island. In: Meschan I, ed. *Roentgen signs in clinical diagnosis*. Philadelphia: WB Saunders, 1957:256.
- 73. Mirra JM. Bone tumors. Philadelphia: Lea and Febiger, 1989:182–190, 303–309.
- 74. Ngan H. Growing bone islands. Clin Radiol 1972;23:199-201.
- Onitsuka H. Roentgenologic aspects of bone islands. Radiology 1977;123:607–612.
- 76. Park HS, Kim JR, Lee SY, Jang KY. Symptomatic giant (10-cm) bone island of the tibia. *Skeletal Radiol* 2005;34:347–350.
- 77. Schmorl G, Junghanns H. *The human spine in health and disease*, 2nd ed. New York: Grune and Stratton, 1971:327.
- Sickels EA, Genant HK, Hoffner PB. Increased localization of 99mTc-pyrophosphate in a bone island: case report. *J Nucl Med* 1976;17:113–115.
- 79. Smith J. Giant bone islands. Radiology 1973;107:35-36.
- Trombetti A, Noel E. Giant bone islands: a case with 31 years of follow-up. *Joint Bone Spine* 2002;69:81–84.

Osteoid Osteoma

- 81. Adil A, Hoeffel C, Fikry T. Osteoid osteoma after a fracture of the distal radius. *Am J Roentgenol* 1996;167:145–146.
- Alani WO, Bartal E. Osteoid osteoma of the femoral neck simulating an inflammatory synovitis. *Clin Orthop* 1987;223:308–312.
- Anderson RB, McAlister JA Jr, Wrenn RN. Case report 585. Intracortical osteosarcoma of tibia. *Skeletal Radiol* 1989;18: 627–630.
- Assoun J, Richardi G, Railhac J-J, et al. Osteoid osteoma: MR imaging versus CT. *Radiology* 1994;191:217–223.
- Baron D, Soulier C, Kermabon C, et al. Ostéomes ostéoides post-traumatique: à propos de deux cas et revue de la litérature. *Rev Rhum Mal Osteoartic* 1992;59:271–275.
- Baruffi MR, Volpon JB, Neto JB, Casartelli C. Osteoid osteomas with chromosome alterations involving 22q. *Cancer Genet Cyto*genet 2001;124:127–131.
- Bell RS, O'Connor GD, Waddell JP. Importance of magnetic resonance imaging in osteoid osteoma: a case report. *Can J Surg* 1989;32:276–278.
- Blacksin MF, Benevenia J. Neoplasms of the scapula. Am J Roentgenol 2000;174:1729–1735.
- Campanacci M. Bone and soft tissue tumors. New York: Springer-Verlag, 1990:355–373.

- Chiou Y-Y, Rosenthal DI, Rosenberg AE. "Beaded" osteoid osteoma: a possible transition between solitary and multicentric tumor. *Skeletal Radiol* 2003;32:412–415.
- Cohen MD, Harrington TM, Ginsburg WW. Osteoid osteoma: 95 cases and a review of the literature. *Semin Arthritis Rheum* 1983;12:265–281.
- Corbett JM, Wilde AH, McCormack LJ, Evarts CM. Intra-articular osteoid osteoma. A diagnostic problem. *Clin Orthop* 1974; 98:225–230.
- Dahlin DC, Unni KK. Bone tumors: general aspects and data on 8542 cases, 4th ed. Springfield, IL: Charles C Thomas, 1987: 88–101.
- Dal Cin P, Sciot R, Samson I, et al. Osteoid osteoma and osteoblastoma with clonal chromosome changes. Br J Cancer 1998;73:344–348.
- Davies M, Cassar-Pullicino VN, Davies AM, et al. The diagnostic accuracy of MR imaging in osteoid osteoma. *Skeletal Radiol* 2002;31:559–569.
- Dockerty MB, Ghormley RK, Jackson AE. Osteoid osteoma: clinicopathologic study of 20 cases. Ann Surg 1951;133:77–89.
- Ebrahim FS, Jacobson JA, Lin J, et al. Intraarticular osteoid osteoma: Sonographic findings in three patients with radiographic, CT, and MR imaging correlation. *Am J Roentgenol* 2001;177:1391–1395.
- Edeiken J, DePalma AF, Hodes PJ. Osteoid osteoma. (Roentgenographic emphasis). *Clin Orthop* 1966;49:201–206.
- Ehara Š, Rosenthal DI, Aoki J, et al. Peritumoral edema in osteoid osteoma on magnetic resonance imaging. *Skeletal Radiol* 1999;28:265–270.
- 100. Freiberger RH, Loitman BS, Helpern M, Thompson TC. Osteoid osteoma: a report of 80 cases. Am J Roentgenol 1959;82: 194–205.
- Gamba JL, Martinez S, Apple J, et al. Computed tomography of axial skeletal osteoid osteomas. *Am J Roentgenol* 1984;142: 769–772.
- Gil S, Marco SF, Arenas J, et al. Doppler duplex color localization of osteoid osteoma. *Skeletal Radiol* 1999;28:107–110.
- Goldberg VM, Jacobs B. Osteoid osteoma of the hip in children. Clin Orthop 1975;106:41–47.
- 104. Glass RB, Poznanski AK, Fisher MR, et al. MR imaging of osteoid osteoma. J Comput Tomogr 1986;10:1065–1067.
- 105. Goldman AB, Schneider R, Pavlov H. Osteoid osteomas of the femoral neck: report of four cases evaluated with isotopic bone scanning, CT, and MR imaging. *Radiology* 1993;186: 227–232.
- Greco F, Tamburelli F, Ciabattoni G. Prostaglandins in osteoid osteoma. *Int Orthop* 1991;15:35–37.
- 107. Greenspan A, Elguezabel A, Bryk D. Multifocal osteoid osteoma. A case report and review of the literature. Am J Roentgenol 1974;121:103–106.
- Hasegawa T, Hirose T, Sakamoto R, et al. Mechanism of pain in osteoid osteomas: an immunohistochemical study. *Histopathol*ogy 1993;22:487–491.
- Hayes CW, Conway WF, Sundaram M. Misleading aggressive MR imaging appearance of some benign musculoskeletal lesions. *RadioGraphics* 1992;12:1119–1134.
- Helms CA. Osteoid osteoma: the double density sign. Clin Orthop 1987;222:167–173.
- Helms CA, Hattner RS, Vogler JB III. Osteoid osteoma: Radionuclide diagnosis. *Radiology* 1984;151:779–784.
- Herrlin K, Ekelund L, Lövdahl R, Persson B. Computed tomography in suspected osteoid osteomas of tubular bones. *Skeletal Radiol* 1982;9:92–97.
- Jackson RP, Reckling FW, Mants FA. Osteoid osteoma and osteoblastoma. Similar histologic lesions with different natural histories. *Clin Orthop* 1977;128:303–313.
- 114. Jaffe HL. Osteoid osteoma of bone. Radiology 1945;45:319-334.
- Kalil RK, Antunes JS. Familial occurrence of osteoid osteoma. Skeletal Radiol 2003;32:416–419.
- 116. Kawaguchi Y, Hasegawa T, Oka S, et al. Mechanism of intramedullary high intensity area on T2-weighted magnetic resonance imaging in osteoid osteoma: a possible role of Cox-2 expression. *Pathol Int* 2001;51:933–937.
- 117. Kayser F, Resnick D, Haghighi P, et al. Evidence of the subperiosteal origin of osteoid osteomas in tubular bones: analysis by CT and MR imaging. *Am J Roentgenol* 1998;170:609–614.

- 118. Keim HA, Reina EG. Osteoid osteoma as a cause of scoliosis. *J Bone Joint Surg Am* 1975;57A:159–163.
- Kendrick JL, Evarts CM. Osteoid osteoma: a critical analysis of 40 tumors. *Clin Orthop* 1967;54:51–59.
- Klein MH, Shankman S. Osteoid osteoma: radiologic and pathologic correlation. *Skeletal Radiol* 1992;21:23–31.
- Kransdorf MJ, Stull MA, Gilkey FW, Moser RP Jr. Osteoid osteoma. *RadioGraphics* 1991;11:671–696.
- Kricun ME. Imaging of bone tumors. Philadelphia: WB Saunders, 1993:121–125.
- 123. Kyriakos M. Intracortical osteosarcoma. *Cancer* 1980;46: 2525–2533.
- 124. Lee GK, Kang IW, Lee ES, et al. Osteoid osteoma of the tarsal cuboid mimicking osteomyelitis. Am J Roentgenol 2004;183: 341–342.
- Lisbona R, Rosenthall L. Role of radionuclide imaging in osteoid osteoma. Am J Roentgenol 1979;132:77–80.
- Liu PT, Chivers FS, Roberts CC, et al. Imaging of osteoid osteoma with dynamic gadolinium-enhanced MR imaging. *Radiol*ogy 2003;227:691–700.
- Makley JT, Dunn MJ. Prostaglandin synthesis by osteoid osteoma. *Lancet* 1982;2:42.
- Marinelli A, Giacomini S, Bianchi G, et al. Osteoid osteoma simulating an osteocartilaginous exostosis. *Skeletal Radiol* 2004;33: 181–185.
- 129. Mirra JM, Dodd L, Johnston W, Frost DB. Case report 700. Primary intracortical osteosarcoma of femur, sclerosing variant, grade 1 to 2 anaplasia. *Skeletal Radiol* 1991;20:613–616.
- Mungo DV, Zhang X, O'Keefe RJ, et al. Cox-1 and Cox-2 expression in osteoid osteoma. J Orthop Res 2002;20:159–162.
- Murphey MD, Andrews CL, Flemming DJ, et al. Primary tumors of the spine: radiologic-pathologic correlation. *RadioGraphics* 1996;16:1131–1158.
- Norman A, Abdelwahab IF, Buyon J, Matzkin E. Osteoid osteoma of the hip stimulating an early onset of osteoarthritis. *Radiology* 1986;158:417–420.
- O'Connell JX, Nanthakumar SS, Nielsen GP, Rosenberg AE. Osteoid osteoma: the uniquely innervated bone tumor. *Mod Pathol* 1998;11:175–180.
- 134. O'Dell CW Jr, Resnick D, Niwayama G, et al. Osteoid osteomas arising in adjacent bones: report of a case. J Can Assoc Radiol 1976;27:298–300.
- Pettine KA, Klassen RA. Osteoid osteoma and osteoblastoma of the spine. J Bone Joint Surg Am 1986;68A:354–361.
- 136. Picci P, Gherlinzoni F, Guerra A. Intracortical osteosarcoma: rare entity or early manifestation of classical osteosarcoma? *Skeletal Radiol* 1983;9:255–258.
- 137. Pinto CH, Taminiau AHM, Vanderschueren GM, et al. Technical considerations in CT-guided radiofrequency thermal ablation of osteoid osteoma: tricks of the trade. *Am J Roentgenol* 2002;179:1633–1642.
- 138. Rand JA, Sim FH, Unni KK. Two osteoid-osteomas in one patient. A case report. J Bone Joint Surg A 1982;64A:1243.
- Roger B, Bellin M-F, Wioland M, Grenier P. Osteoid osteoma: CTguided percutaneous excision confirmed with immediate followup scintigraphy in 16 outpatients. *Radiology* 1996;201:239–243.
- 140. Rosenthal DI, Hornicek FJ, Wolfe MW, et al. Percutaneous radiofrequency coagulation of osteoid osteoma compared with operative treatment. *J Bone Joint Surg Am* 1998;80A:815–821.
- Rosenthal DI, Springfield DS, Gebhardt MC, et al. Osteoid osteoma: percutaneous radio-frequency ablation. *Radiology* 1995;197:451–454.
- 142. Sabanas AO, Bickel WH, Moe JH. Natural history of osteoid osteoma of the spine: review of the literature and report of three cases. *AmJ Surg* 1956;91:880–889.
- 143. Schai P, Friederich NB, Krüger A, et al. Discrete synchronous multifocal osteoid osteoma of the humerus. *Skeletal Radiol* 1996;25:667–670.
- 144. Schlesinger AE, Hernandez RJ. Intracapsular osteoid osteoma of the proximal femur: findings on plain film and CT. Am J Roentgenol 1990;154:1241–1244.
- Sim FH, Dahlin DC, Beabout JW. Osteoid-osteoma: diagnostic problems. J Bone Joint Surg 1975;57A:154–159.
- Smith FW, Gilday DL. Scintigraphic appearances of osteoid osteoma. *Radiology* 1980;137:191–195.

- 147. Steiner GC. Ultrastructure of osteoid osteoma. Hum Pathol
- 1976;7:309–325.
- 148. Strach EH. Osteoid osteoma. Br Med J 1953;1:1031.
- Swee RG, McLeod RA, Beabout JW. Osteoid osteoma. Detection, diagnosis, and localization. *Radiology* 1979;130:117–123.
- 150. Thompson GH, Wong KM, Konsens RM, Vibhakars S. Magnetic resonance imaging of an osteoid osteoma of the proximal femur: a potentially confusing appearance. J Pediatr Orthop 1990;10:800–804.
- Vanderschueren GM, Taminiau AHM, Oberman WR, Bloem JL. Osteoid osteoma: clinical results with thermocoagulation. *Radiology* 2002;224:82–86.
- Winter PF, Johnson PM, Hilal SK, Feldman F. Scintigraphic detection of osteoid osteoma. *Radiology* 1977;122:177–178.
- Wold LE, Pritchard DJ, Bergert J, Wilson DM. Prostaglandin synthesis by osteoid osteoma and osteoblastoma. *Mod Pathol* 1988;1:129–131.
- 154. Woods ER, Martel W, Mandell SH, Crabbe JP. Reactive soft-tissue mass associated with osteoid osteoma: correlation of MR imaging features with pathologic findings. *Radiology* 1993;186:221–225.
- 155. Yamamura S, Sato K, Sugiura H, et al. Magnetic resonance imaging of inflammatory reaction in osteoid osteoma. Arch Orthop Trauma Surg 1994;114:8–13.
- 156. Yeager BA, Schiebler ML, Wertheim SB, et al. Case report: MR imaging of osteoid osteoma of the talus. J Comput Assist Tomogr 1987;11:916–917.
- 157. Youssef BA, Haddad MC, Zahrani A, et al. Osteoid osteoma and osteoblastoma: MRI appearances and the significance of ring enhancement. *Eur Radiol* 1996;6:291–296.

Osteoblastoma

- Adler C-P. Multifocal osteoblastoma of the hand. Skeletal Radiol 2000;29:601–604.
- 159. Azouz EM, Kozlowski K, Marton D, et al. Osteoid osteoma and osteoblastoma of the spine in children. Report of 22 cases with brief literature review. *Pediatr Radiol* 1986;16:25–31.
- Bahk WJ, Mirra JM. Pseudoanaplastic tumors of bone. Skeletal Radiol 2004;33:641–648.
- Bertoni F, Unni KK, Lucas DR, McLeod RA. Osteoblastoma with cartilaginous matrix. Am J Surg Pathol 1993;17:69–74.
- Bertoni F, Unni KK, McLeod RA, Dahlin DC. Osteosarcoma resembling osteoblastoma. *Cancer* 1985;55:416–426.
- Bettelli G, Tigani D, Picci P. Recurring osteoblastoma initially presenting as a typical osteoid osteoma. Report of two cases. *Skeletal Radiol* 1991;20:1–4.
- 164. Byers PD. Solitary benign osteoblastic lesions of bone. Osteoid osteoma and benign osteoblastoma. *Cancer* 1968;22:43–57.
- Crim JR, Mirra JM, Eckardt JJ, Seeger LL. Widespread inflammatory response to osteoblastoma: the flare phenomenon. *Radiology* 1990;177:835–836.
- 166. Dale S, Breidahl WH, Baker D, et al. Severe toxic osteoblastoma of the humerus associated with diffuse periostitis of multiple bones. *Skeletal Radiol* 2001;30:464–468.
- 167. Dahlin DC, Johnson EW Jr. Giant osteoid osteoma. J Bone Joint Surg Am 1954;36A:559–572.
- Della Rocca C, Huvos AG. Osteoblastoma: varied histological presentations with a benign clinical course. 55 cases. Am J Surg Pathol 1996;20:841–850.
- 169. Denis F, Armstrong GW. Scoliogenic osteoblastoma of the posterior end of the rib: a case report. *Spine* 1984;9:74–76.
- DeSouza, Diaz L, Frost HM. Osteoid osteoma—osteoblastoma. Cancer 1974;33:1075–1081.
- 171. Dorfman HD, Weiss SW. Borderline osteoblastic tumors: problems in the differential diagnosis of aggressive osteoblastoma and low-grade osteosarcoma. *Semin Diagn Pathol* 1984; 1:215–234.
- 172. Fabris D, Trainiti G, Di Comun M, Agostini S. Scoliosis due to rib osteoblastoma: report of two cases. J Pediatr Orthop 1983; 3:370–375.
- Fanning JW, Lucas GL. Osteoblastoma of the scaphoid: a case report. J Hand Surg 1993;18A:663–665.
- 174. Farmlett EJ, Magid D, Fishman EK. Osteoblastoma of the tibia: CT demonstration. J Comput Assist Tomogr 1986;10:1068–1070.
- Gitelis S, Schajowicz F. Osteoid osteoma and osteoblastoma. Orthop Clin North Am 1989;20:313–325.

- Healey HJ, Ghelman B. Osteoid osteoma and osteoblastoma. Clin Orthop 1986;204:76–85.
- 177. Jaffe HL. Benign osteoblastoma. Bull Hosp Joint Dis 1956;17: 141–151.
- Jaffe HL, Mayer L. An osteoblastic osteoid tissue-forming tumor of a metacarpal bone. *Arch Surg* 1932;24:550–564.
- Kenan S, Floman Y, Robin GC, Laufer A. Aggressive osteoblastoma. A case report and review of the literature. *Clin Orthop* 1985;195:294–298.
- Kroon HM, Schurmans J. Osteoblastoma: clinical and radiologic findings in 98 new cases. *Radiology* 1990;175:783–790.
- Kumar R, Guinto FC Jr, Madewell JE, et al. Expansile bone lesions of the vertebra. *RadioGraphics* 1988;8:749–769.
- 182. Lichtenstein L. Benign osteoblastoma. A category of osteoidand bone-forming tumors other than classical osteoid osteoma, which may be mistaken for giant-cell tumor or osteogenic sarcoma. *Cancer* 1956;9:1044–1052.
- Lucas DR, Unni KK, McLeod RA, et al. Osteoblastoma: clinicopathologic study of 306 cases. *Hum Pathol* 1994;25:117–134.
- 184. Malcolm AJ, Schiller AL, Schneider-Stock R. Osteoblastoma. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:259–285.
- Marcove RC, Alpert M. A pathologic study of benign osteoblastoma. *Clin Orthop* 1963;30:175–180.
- Marsh BW, Bonfiglio M, Brady LP, Enneking WF. Benign osteoblastoma: range of manifestations. J Bone Joint Surg Am 1975;57A:1–9.
- 187. McLeod RA, Dahlin DC, Beabout JW. The spectrum of osteoblastoma. Am J Roentgenol 1976;126:321–325.
- Mirra JM, Dodd L, Johnston W, Frost DB. Case report 700: Primary intracortical osteosarcoma of femur, sclerosing variant, grade 1 to 2 anaplasia. *Skeletal Radiol* 1991;20:613–616.
- Mitchell ML, Ackerman LV. Metastatic and pseudomalignant osteoblastoma: a report a two unusual cases. *Skeletal Radiol* 1986;15:213–218.
- Nakatani T, Yamamoto T, Akisue T, et al. Periosteal osteoblastoma of the distal femur. *Skeletal Radiol* 2004;33:107–111.
- 191. Picci P, Campanacci M, Mirra JM. Osteoid osteoma. Differential clinicopathologic diagnosis. In: Mirra JM, ed. Bone tumors: clinical, radiologic, and pathologic correlations. Philadelphia: Lea and Febiger, 1989:411–414.
- 192. Radig K, Schneider-Stock R, Mittler U, et al. Genetic instability in osteoblastic tumors of the skeletal system. *Pathol Res Pract* 1998;194:669–677.
- 193. Radig K, Schneider-Stock R, Oda Y, et al. Mutation spectrum of p53 gene in highly malignant human osteosarcomas. *Gen Diagn Pathol* 1996;142:25–32.
- 194. Rutter DJ, van Rijssel TG, van der Velde EA. Aneurysmal bone cysts. A clinicopathological study of 105 cases. *Cancer* 1977; 39:2231–2239.
- 195. Schajowicz F, Lemos C. Malignant osteoblastoma. J Bone Joint Surg 1976;58B:202–211.
- Steiner GC. Ultrastructure of osteoblastoma. Cancer 1977;39: 2127–2136.
- 197. Tillman BP, Dahlin DC, Lipscomb PR, Stewart JR. Aneurysmal bone cyst: an analysis of ninety-five cases. *Mayo Clin Proc* 1968;43:478–495.
- 198. van der Waal I. Cementoblastoma. In: Barnes L, Eveson JW, Reichart P, Sidransky D, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of head and neck. Lyon: IARC Press, 2005:318.

Osteosarcomas

- Abdelwahab IF, Klein MJ, Hermann G, Springfield D. Angiosarcomas associated with bone infarcts. *Skeletal Radiol* 1998; 27:546–551.
- 200. Abdelwahab IF, Klein MJ, Hermann G, et al. Osteosarcoma arising in a desmoplastic fibroma of the proximal tibia. Am J Roentgenol 2002;178:613–615.
- Abdulkader I, Cameselle-Teijeiro J, Fraga M, et al. Sclerosing epithelioid fibrosarcoma primary of the bone. *Int J Surg Pathol* 2002;10:227–230.

- 202. Abdulrahman RE, White CS, Templeton PA, et al. Primary osteosarcoma of the ribs: CT findings. *Skeletal Radiol* 1995;24: 127–129.
- 203. Ackerman LV. Extra-osseous localized non-neoplastic bone and cartilage formation (so-called myositis ossificans). J Bone Joint Surg Am 1958;40A:279–298.
- 204. Ahuja SC, Villacin AB, Smith J, et al. Juxtacortical (parosteal) osteogenic sarcoma: histological grading and prognosis. *J Bone Joint Surg Am* 1977;59A: 632–647.
- 205. Aizawa T, Okada K, Abe E, et al. Multicentric osteosarcoma with long-term survival. *Skeletal Radiol* 2004;33:41–45.
- Allan CJ, Soule EH. Osteogenic sarcoma of the somatic soft tissues. Clinicopathologic study of 26 cases and review of literature. *Cancer* 1971;27:1121–1133.
- 207. Alpert LI, Abaci IF, Werthamer S. Radiation-induced extraskeletal osteosarcoma. *Cancer* 1973;31:1359–1363.
- Al-Romaih K, Bayani J, Vorobyova J, et al. Chromosomal instability in osteosarcoma and its association with centrosome abnormalities. *Cancer Genet Cytogenet* 2003;144:91–99.
- Amendola MA, Glazer GM, Adha FP, et al. Myositis ossificans circumscripta: computed tomographic diagnosis. *Radiology* 1983; 149:775–779.
- Amstutz HC. Multiple osteogenic sarcomata—metastatic or multicentric? *Cancer* 1969;24:923–931.
- 211. Andersen KJ, Sundaram M, Unni KK, Sim FH. Imaging features of low-grade central osteosarcoma of the long bones and pelvis. *Skeletal Radiol* 2004;33:373–379.
- 212. Angervall L, Stener B, Stener I, Ahren C. Pseudomalignant osseous tumor of soft tissue. A clinical, radiological and pathological study of five cases. *J Bone Joint Surg Br* 1969;51B:654–663.
- 213. Antonescu CR, Rosenblum MK, Pereira P, et al. Sclerosing epithelioid fibrosarcoma: a study of 16 cases and confirmation of a clinicopathologically distinct tumor. *Am J Surg Pathol* 2001; 25:699–709.
- 214. Arndt CAS, Crist WM. Common musculoskeletal tumors of childhood and adolescence. *N Engl J Med* 1999;341:342–352.
- 215. Ayala AG, Ro JY, Raymond AK, et al. Small cell osteosarcoma. A clinicopathologic study of 27 cases. *Cancer* 1989;64:2162–2173.
- Ballance WA Jr, Mendelsohn G, Carter JR, et al. Osteogenic sarcoma. Malignant fibrous histiocytoma subtype. *Cancer* 1988; 62:763–771.
- 217. Bane BL, Evans HL, Ro JY, et al. Extra-skeletal osteosarcoma. A clinico-pathologic study of 26 cases. *Cancer* 1990;65:2762–2770.
- 218. Baró PR, Bastart FM, Bartrina JR, et al. Case report 529. Osteosarcoma of calcaneus with Rothmund-Thomson syndrome (RTS). Skeletal Radiol 1989;18:136–139.
- 219. Bathhurst N, Sanerkin N, Watt I. Osteoclast-rich osteosarcoma. Br J Radiol 1986;59:667–673.
- 220. Bertoni F, Bacchini P, Fabbri N, et al. Osteosarcoma: low-grade intraosseous-type osteosarcoma, histologically resembling parosteal osteosarcoma, fibrous dysplasia, and desmoplastic fibroma. *Cancer* 1993;71:338–345.
- 221. Bertoni F, Present D, Bacchini P, et al. The Instituto Rizzoli experience with small cell osteosarcoma. *Cancer* 1989;64: 2591–2599.
- 222. Blasius S, Link TM, Hillmann A, et al. Intracortical low grade osteosarcoma. A unique case and review of the literature on intracortical osteosarcoma. *Gen Diagn Pathol* 1996;141:273–278.
- 223. Bloem JL, Kroon HM. Osseous lesions. Radiol Clin North Am 1993;31:261–278.
- 224. Bloom D. Congenital telangiectatic erythema resembling lupus erythematosus in dwarfs; probably a syndrome entity. *Am J Dis Child* 1954;88:754–758.
- 225. Bohndorf K, Reiser M, Lochner B, et al. Magnetic resonance imaging of primary tumors and tumor-like lesions of bone. *Skeletal Radiol* 1986;15:511–517.
- 226. Bonar SF, McCarthy S, Stalley P, et al. Epiphyseal osteoblastomalike osteosarcoma. *Skeletal Radiol* 2004;33:46–50.
- 227. Boyko OB, Cory DA, Cohen MD, et al. MR imaging of osteogenic and Ewing's sarcoma. Am J Roentgenol 1987;148:317–322.
- Brenner W, Bohuslavizki KH, Eary JF. PET imaging of osteosarcoma. J Nucl Med 2003;44:930–942.
- 229. Bridge JA, Nelson M, McComb E, et al. Cytogenetic findings in 73 osteosarcoma specimens and a review of the literature. *Cancer Genet Cytogenet* 1997;95:74–87.

- 230. Broders AC. The microscopic grading of cancer. In: Pack CT, Ariel IM, eds. *Treatment of cancer and allied diseases*, 2nd ed., Vol 1. New York: Paul B. Hoeber, 1958:55.
- Burgener FA, Perry P. Solitary renal cell carcinoma metastasis in Paget's disease simulating sarcomatous degeneration. Am J Roentgenol 1977;128:835–855.
- 232. Buzzoni R, Della Torre S, Cortinovis D, Catena L. [Case report of synchronous multicentric osteosarcoma and review of the literature: the importance of autopsy for diagnosis.] *Tumori* 2005;91:90–92.
- Campanacci M, Cervellati G. Osteosarcoma: a review of 345 cases. Ital J Orthop Traumatol 1975;1:5–22.
- Campanacci M, Pizzoferrato A. Osteosarcoma emorragico. Chir Organi Mov 1971;60:409–421.
- 235. Cavanee W, Bögler O, Hadjistilianou T, Newsham I. Retinoblastoma syndrome. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002: 363–364.
- Chew FS, Richardson ML. Radiological reasoning: a benign-appearing bone mass. Am J Roentgenol 2005;184:S169–S174.
- Choudur HN, Munk PL, Nielson TO, Ryan AG-MJ. Primary mesenteric extraskeletal osteosarcoma in the pelvic cavity. *Skeletal Radiol* 2005;34:649–652.
- 238. Chung EB, Enzinger FM. Extraskeletal osteosarcoma. *Cancer* 1987;60:1132–1142.
- 239. Cody JD, Singer FR, Roodman GD, et al. Genetic linkage of Paget disease of the bone to chromosome 18q. *Am J Hum Genet* 1997;61:1117–1122.
- 240. Daffner RH, Fox KR, Galey RK. Fibroblastic osteosarcoma of the mandible. *Skeletal Radiol* 2002;31:107–111.
- Dahlin DC. Malignant osteoblastic tumors. In: Taveras JM, Ferucci JT, eds. *Radiology: diagnosis, imaging, intervention*, vol 5. Philadelphia: JB Lippincott, 1986:1–10.
- Dahlin DC. Grading of bone tumors. In: Unni KK, ed. Bone tumors. New York: Churchill-Livingstone, 1988:35–45.
- 243. Dahlin DC, Coventry MB. Osteogenic sarcoma: a study of six hundred cases. *J Bone Joint Surg* 1967;49A:101–110.
- 244. Dahlin DC, Unni KK. Osteosarcoma of bone and its important recognizable varieties. *Am J Surg Pathol* 1977;1:61–72.
- 245. Dahlin DC, Unni KK, Matsuno T. Malignant (fibrous) histiocytoma of bone—fact or fancy? *Cancer* 1977;39:1508–1516.
- Dardick I, Schatz JE, Colgan TJ. Osteogenic sarcoma with epithelial differentiation. Ultrastruct Pathol 1992;16:463–474.
- 247. Desai P, Perino G, Present D, Steiner GC. Sarcoma in association with bone infarcts. Report of five cases. Arch Pathol Lab Med 1996;120:482–489.
- deSantos LA, Edeiken B. Purely lytic osteosarcoma. Skeletal Radiol 1982;9:1–7.
- deSantos LA, Edeiken BS. Subtle early osteosarcoma. Skeletal Radiol 1985;13:44–48.
- deSantos LA, Murray JA, Finkelstein JB, et al. The radiographic spectrum of periosteal osteosarcoma. *Radiology* 1978;127: 123–129.
- DeSmet AA, Norris MA, Fisher DR. Magnetic resonance imaging of myositis ossificans: analysis of seven cases. *Skeletal Radiol* 1992;21:503–507.
- 252. Destouet JM, Gilula LA, Murphy WA. Computed tomography of long-bone osteosarcoma. *Radiology* 1979;131:439–435.
- 253. Devaney K, Vinh TN, Sweet DE. Small cell osteosarcoma of bone: an immunohistochemical study with differential diagnostic considerations. *Hum Pathol* 1993;24:1211–1225.
- 254. Dickerson GR, Rosenberg AE. The ultrastructure of small cell osteosarcoma with a review of light microscopy and differential diagnosis. *Hum Pathol* 1991;221:267–275.
- 255. Doud TM, Moser RP Jr, Giudici MAI, et al. Case report 704. Extraskeletal osteosarcoma of the thigh with several suspected skeletal metastases and extensive metastases to the chest. *Skeletal Radiol* 1991;20:628–632.
- Edeiken J, Raymond AK, Ayala AG, et al. Small-cell osteosarcoma. Skeletal Radiol 1987;16:621–628.
- 257. el-Khoury JM, Haddad SN, Atallah NG. Osteosarcomatosis with Rothmund-Thomson syndrome. *Br J Radiol* 1997;70:215–218.
- 258. Ellis JH, Siegel CL, Martel W, Weatherbee L, Dorfman H. Radiologic features of well-differentiated osteosarcoma. Am J Roentgenol 1988;151:739–742.

- CHAPTER 2 Bone-Forming (Osteogenic) Lesions 153
- 259. Enzinger F, Weiss S. Soft tissue tumors. St. Louis: CV Mosby, 1983.
- 260. Evans SC, Mims B, McMasters KM, et al. Exclusion of a p53 germline mutation in a classic Li-Fraumeni syndrome family. *Hum Genet* 1998;102:681–686.
- 261. Farr GH, Huvos AG, Marcove RC, et al. Telangiectatic osteogenic sarcoma: a review of twenty-eight cases. *Cancer* 1974; 34:1150–1158.
- Fine G, Stout AP. Osteogenic sarcoma of the extraskeletal soft tissues. *Cancer* 1956;9:1027–1043.
- 263. Forest M, Tomaneo B, Vanel D. Orthopedic surgical pathology, 1st ed. Edinburgh: Churchill Livingstone, 1998:795.
- Fuchs B, Pritchard DJ. Etiology of osteosarcoma. Clin Orthop Relat Res 2002;397:40–52.
- 265. Fukuda K, Ushigome S, Nikaidou T, et al. Osteosarcoma of the metatarsal bone. *Skeletal Radiol* 1999;28:294–297.
- Garber JE, Offit K. Hereditary cancer predisposition syndromes. *J Clin Oncol* 2005;23:276–292.
- 267. Garcia GA, Barros F, Bouzas ML, Penaranda JM. Li-Fraumeni syndrome and osteosarcoma of the maxilla. J Oral Maxillofac Surg 1998;56:1106–1109.
- 268. Geirnaerdt MJA, Bloem JL, van der Woude H-J, et al. Chondroblastic osteosarcoma: characterization by gadolinium-enhanced MR imaging correlated with histopathology. *Skeletal Radiol* 1998;27:145–153.
- Gherlinzoni F, Antoci B, Canale V. Multicentric osteosarcomata (osteosarcomatosis). *Skeletal Radiol* 1983;10:281–285.
- 270. Glicksman AS, Toker C. Osteogenic sarcoma following radiotherapy for bursitis. *Mt Sinai J Med* 1976;43:163–167.
- Goldman AB. Myositis ossificans circumscripta: a benign lesion with a malignant differential diagnosis. Am J Roentgenol 1976;126:32–40.
- 272. Gomes H, Menanteau B, Gaillard D, Behar C. Telangiectatic osteosarcoma. *Pediatr Radiol* 1986;16:140–143.
- Goto M, Miller RW, Ishikawa Y, Sugano H. Excess of rare cancers in Werner syndrome (adult progeria). *Cancer Epidemiol Biomarkers Prev* 1996;5:239–246.
- 274. Goto M, Rubenstein M, Weber J, et al. Genetic linkage of Werner's syndrome to five markers on chromosome 8. *Nature* 1992;355:735–738.
- Greenfield GB, Arrington JA. Imaging of bone tumors. A multimodality approach. Philadelphia: JB Lippincott, 1955:48–91.
- 276. Greenspan A. Osteosarcoma. Contemp Diagn Radiol 1993;16: 1-6.
- 277. Greenspan A, Klein MJ. Osteosarcoma: radiologic imaging, differential diagnosis, and pathological considerations. *Semin Orthop* 1991;6:156–166.
- 278. Greenspan A, Steiner G, Norman A, et al. Case report 436. Osteosarcoma of the soft tissues of the distal end of the thigh. *Skeletal Radiol* 1987;16:489–492.
- Griffith JF, Kumta SM, Chow LTC, et al. Intracortical osteosarcoma. Skeletal Radiol 1998;27:228–232.
- Gunawardena S, Chintagumpala M, Trautwein L, et al. Multifocal osteosarcoma: an unusual presentation. *J Pediatr Hematol On*col 1999;21:58–62.
- 281. Hainaut P, Eeles R, Ohgaki H, Oliver M. Li-Fraumeni syndrome. In: Tavassoli F, Devilee P, eds. *Tumours of the breast and female genital organs*. Lyon: IARC Press, 2003:351–354.
- 282. Hall MB, Sclar AG, Gardner DF. Albright's syndrome with reactivation of fibrous dysplasia secondary to pituitary adenoma and further complicated by osteogenic sarcoma. Report of a case. *Oral Surg Oral Med Oral Pathol* 1984;57:616–619.
- Hall RB, Robinson LH, Malawer MM, Dunham WK. Periosteal osteosarcoma. *Cancer* 1985;55:165–171.
- 284. Hasegawa T, Hirose T, Seki K, et al. Histological and immunohistochemical diversities, and proliferative activity and grading in osteosarcomas. *Cancer Detect Prev* 1997;21:280–287.
- 285. Hasegawa T, Shimoda T, Yokoyama R, et al. Intracortical osteoblastic osteosarcoma with oncogenic rickets. *Skeletal Radiol* 1999;28:41–45.
- 286. Hauben EI, Arends J, Vandenbroucke JP, et al. Multiple primary malignancies in osteosarcoma patients. Incidence and predictive value of osteosarcoma subtype for cancer syndromes related with osteosarcoma. *Eur J Hum Genet* 2003;11:611–618.
- 287. Heinrich SD, Zembo MM, MacEwen GD. Pseudomalignant myositis ossificans. *Orthopedics* 1989;12:599–602.

- Helliwell TR, O'Connor MA, Ritchie DA, et al. Bizarre parosteal osteochondromatous proliferation with cortical invasion. *Skeletal Radiol* 2001;30:282–285.
- Hermann G, Abdelwahab IF, Kenan S, et al. Case report 795. High-grade surface osteosarcoma of the radius. *Skeletal Radiol* 1993;22:383–385.
- 290. Hermann G, Klein MJ, Springfield D, et al. Intracortical osteosarcoma; two-year delay in diagnosis. *Skeletal Radiol* 2002;31:592–596.
- 291. Heul RO van der, Ronnen Jr von. Juxtacortical osteosarcoma. Diagnosis, differential diagnosis, treatment, and an analysis of eighty cases. *J Bone Joint Surg Am* 1967;49A:415–439.
- Hickson ID. RecQ helicases: caretakers of the genome. Nat Rev Cancer 2003;3:169–178.
- 293. Hocking LJ, Herbert CA, Nicholls RK, et al. Genomewide search in familial Paget disease of bone shows evidence of genetic heterogeneity with candidate loci on chromosomes 2q36, 10p13, and 5q35. *Am J Hum Genet* 2001;69:1055–1061.
- Hopper KD, Moser RP Jr, Haseman DB, et al. Osteosarcomatosis. *Radiology* 1990;175:233–239.
- 295. Hudson TM, Springfield DS, Benjamin M, et al. Computed tomography of parosteal osteosarcoma. *Am J Roentgenol* 1985; 144:961–965.
- 296. Huvos AG. Bone tumors. Diagnosis, treatment, and prognosis, 2nd ed. Philadelphia: WB Saunders, 1991:784.
- 297. Huvos AG. Osteogenic sarcoma of bones and soft tissues in older persons. *Cancer* 1986;57:1442–1449.
- 298. Huvos AG, Rosen G, Bretsky SS, Butler A. Telangiectatic osteosarcoma: a clinicopathologic study of 124 patients. *Cancer* 1982;49:1679–1689.
- 299. Ilaslan H, Sundaram M, Unni KK, Shives TC. Primary vertebral osteosarcoma: imaging findings. *Radiology* 2004;230:697–702.
- Inward CY. Low-grade central osteosarcoma versus fibrous dysplasia. Pathol Case Rev. 2001;6:22–27.
- 301. Înward CY, Knuutila S. Low grade central osteosarcoma. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:275–276.
- Jaffe HL. Intracortical osteogenic sarcoma. Bull Hosp Joint Dis 1960;21:180–197.
- 303. Jaffe HL. Tumors and tumorous conditions of the bones and joints. Philadelphia: Lea and Febiger, 1968.
- Jelinek JS, Murphey MD, Kransdorf MJ, et al. Parosteal osteosarcoma: value of MR imaging and CT in the prediction of histologic grade. *Radiology* 1996;201:837–842.
- 305. Jhala DN, Eltoum I, Carroll AJ, et al. Osteosarcoma in a patient with McCune-Albright syndrome and Mazabraud's syndrome: a case report emphasizing the cytological and cytogenetic findings. *Hum Pathol* 2003;34:1354–1357.
- 306. Johnson K, Davies AM, Mangham DC, Grimer RJ. Parosteal osteosarcoma of a metatarsal with intramedullary invasion. *Skeletal Radiol* 1999;28:111–115.
- 307. Johnson-Pais TL, Nellissery MJ, Ammerman DG, et al. Determination of a minimal region of loss of heterozygosity on chromosome 18q21.33 in osteosarcoma. *Int J Cancer* 2003;105: 285–288.
- 308. Jundt G, Prein J. Knochentumoren und tumorähnliche Läsionen im Kiefer. Erfahrungen aus dem Basler DÖSAK-Referenzregister. Mund Kiefer Gesichtschir 2000;4(suppl 1): S196–S207.
- 308a. Kajihara M, Sugawara Y, Hirata M, et al. Extraskeletal osteosarcoma in the thigh: a case report. *Radiat Med* 2005;23:142–146.
- 309. Kaufman RA, Towbin RB. Telangiectatic osteosarcoma simulating the appearance of an aneurysmal bone cyst. *Pediatr Radiol* 1981;11:102–104.
- 310. Kawaguchi K, Oda Y, Sakamoto A, et al. Molecular analysis of p53, MDM2, and H-ras genes in osteosarcoma and malignant fibrous histiocytoma of bone in patients older than 40 years. *Mod Pathol* 2002;15:878–888.
- Kenan S, Abdelwahab IF, Klein MJ, et al. Lesions of juxtacortical origin (surface lesions of bone). *Skeletal Radiol* 1993;22:337–357
- Kenan S, Abdelwahab IF, Klein MJ, et al. Case report 835. Parosteal osteosarcoma involving the left radius. *Skeletal Radiol* 1994;22:229–231.
- Kido A, Schneider-Stock R, Hauptmann K, Roessner A. Telomerase activity in benign bone tumors and tumor-like lesions. *Pathol Res Pract* 1999;195:753–757.

- Kitao S, Lindor NM, Shiratori M, et al. Rothmund-Thomson syndrome responsible gene, RECQL4: genomic structure and products. *Genomics* 1999;61:268–276.
- 315. Kitao S, Shimamoto A, Goto M, et al. Mutations in RECQL4 cause a subset of cases of Rothmund-Thomson syndrome. *Nat Genet* 1999;22:82–84.
- Klein MJ, Kenan S, Lewis MM. Osteosarcoma: clinical and pathological considerations. Orthop Clin North Am 1989;20: 327–345.
- 317. Konishi E, Kusuzaki K, Murata H, et al. Extraskeletal osteosarcoma arising in myositis ossificans. *Skeletal Radiol* 2001;30:39–43.
- 318. Kramer K, Hicks D, Palis J, et al. Epithelioid osteosarcoma of bone. Immunocytochemical evidence suggesting divergent epithelial and mesenchymal differentiation in a primary osseous neoplasm. *Cancer* 1993;71:2977–2982.
- 319. Kransdorf MJ, Meis JM. Extraskeletal osseous and cartilaginous tumors of the extremities. *RadioGraphics* 1993;13:853–884.
- Kransdorf MJ, Meis JM, Jelinek JS. Myositis ossificans: MR appearance with radiologic-pathologic correlation. *Am J Roentgenol* 1991;157:1243–1248.
- 321. Kumar N, David R, Madewell JE, Lindell MM Jr. Radiographic spectrum of osteogenic sarcoma. *Am J Roentgenol* 1987;148: 767–772.
- 322. Kurt AM, Unni KK, McLeod RA, Pritchard DJ. Low-grade intraosseous osteosarcoma. *Cancer* 1990;65:1418–1428.
- Kyriakos M, Gilula LA, Besich MJ, Schoeneker PL. Intracortical small cell osteosarcoma. *Clin Orthop Rel Res* 1992;279:269– 280.
- 324. Ladanyi M, Gorlick R. Molecular pathology and molecular pharmacology of osteosarcoma. *Pediatr Pathol Mol Med* 2000;19: 391–413.
- 325. Layfield R, Hocking LJ. SQSTM1 and Paget's disease of bone. Calcif Tissue Int 2004;75:347–357.
- 326. Lee HJ, Kim IO, Kim WS, et al. Metachronous multifocal osteosarcoma: a case report and literature review. *Clin Imaging* 2002;26:63–68.
- 327. Lee JS, Fetsch JF, Wasdhal DA, et al. A review of 40 patients with extraskeletal osteosarcoma. *Cancer* 1995;76:2253–2259.
- 328. Lee YY, Van Tassel P, Nauert C, et al. Craniofacial osteosarcomas: plain film, CT and MR findings in 46 cases. *Am J Roentgenol* 1988;150:1397–1402.
- Levine E, De Smet AA, Huntrakoon M. Juxtacortical osteosarcoma: a radiologic and histologic spectrum. *Skeletal Radiol* 1985;14:38–46.
- 330. Li FP, Fraumeni JF, Jr, Mulvihill JJ, et al. A cancer family syndrome in twenty-four kindreds. *Cancer Res* 1988;48:5358–5362.
- Lindell MM Jr, Shirkhoda A, Raymond AK, et al. Parosteal osteosarcoma: radiologic-pathologic correlation with emphasis on CT. Am J Roentgenol 1987;148:323–328.
- 332. Lindor NM. Rothmund-Thomson syndrome. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:365.
- 333. Logan PM, Mitchell MJ, Munk PL. Imaging of variant osteosarcomas with an emphasis on CT and MR imaging. Am J Roentgenol 1998;171:1531–1537.
- 334. Lopez BF, Rodriquez PJL, Gonzalez LJ, et al. Intracortical osteosarcoma. A case report. *Clin Orthop Rel Res* 1991;268: 218–222.
- 335. Malcolm AJ. Osteosarcoma: classification, pathology, and differential diagnosis. *Semin Orthop* 1988;3:1–12.
- Martinez-Tello FJ, Navas-Palacios JJ. The ultrastructure of conventional, parosteal, and periosteal osteosarcoma. *Cancer* 1982;50:949–961.
- Matsuno T, Unni KK, McLeod RA, Dahlin DC. Telangiectatic osteogenic sarcoma. *Cancer* 1976;38:2538–2547.
- McDonald DJ. Limb-salvage surgery for treatment of sarcomas of the extremities. *Am J Roentgenol* 1994;163:509–513.
- McKenna RJ, Schwinn CP, Soong KY, Higinbotham NL. Osteogenic sarcoma arising in Paget's disease. *Cancer* 1964;17:42–66.
- Meis-Kindblom JM, Kindblom LG, Enzinger FM. Sclerosing epithelioid fibrosarcoma. A variant of fibrosarcoma simulating carcinoma. *Am J Surg Pathol* 1995;19:979–993.
- 341. Meneses MF, Unni KK, Swee RG. Bizarre parosteal osteochondromatous proliferation of bone (Nora's lesion). Am J Surg Pathol 1993;17:691–697.

- Mertens F, Albert A, Heim S, et al. Clonal structural chromosome aberrations in fibrous dysplasia. *Genes Chromosomes Cancer* 1994;11:271–272.
- 343. Mindell ER, Shah NK, Webster JH. Postradiation sarcoma of bone and soft tissues. Orthop Clin North Am 1977;8:821–834.
- 344. Mirra JM, Dodd L, Johnston W, et al. Case report 700: Primary intracortical osteosarcoma of femur, sclerosing variant, grade 1 to 2 anaplasia. *Skeletal Radiol* 1991;20:613–616.
- 345. Mirra JM, Gold RH, Picci P. Osseous tumors of intramedullary origin. In: Mirra JM, ed. Bone tumors. Clinical, radiologic, and pathologic correlations. Philadelphia: Lea and Febiger, 1989: 143–438, 940.
- 346. Monnat RJ Jr. Werner syndrome. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:366–367.
- 347. Moore TE, King AR, Kathol MH, et al. Sarcoma in Paget disease of bone: clinical, radiologic, and pathologic features in 22 cases. *Am J Roentgenol* 1991;156:1199–1203.
- Moppett J, Oakhill A, Duncan AW. Second malignancies in children: the usual suspects? *Eur J Radiol* 2001;38:235–248.
- Mulder JD, Schütte HE, Kroon HM, Taconis WK. Radiologic atlas of bone tumors. Amsterdam: Elsevier, 1993:51–76.
- 350. Murata K, Hatamochi A, Shinkai H, et al. A case of Werner's syndrome associated with osteosarcoma. *J Dermatol* 1999;26: 682–686.
- Murphey MD, Jelinek JS, Temple T, et al. Imaging of periosteal osteosarcoma: radiologic-pathologic comparison. *Radiology* 2004;233:129–138.
- 352. Murphey MD, Robbin MR, McRae GA, et al. The many faces of osteosarcoma. *RadioGraphics* 1997;17:1205–1231.
- Murphey MD, wan Jaovisidha S, Temple HT, et al. Telangiectatic osteosarcoma: radiologic-pathologic comparison. *Radiology* 2003; 229:545–553.
- 354. Naka T, Fukuda T, Shinohara N, et al. Osteosarcoma versus malignant fibrous histiocytoma of bone in patients older than 40 years. *Cancer* 1995;76:972–984.
- Nakajima H, Sim FH, Bond JR, Unni KK. Small cell osteosarcoma of bone. Review of 72 cases. *Cancer* 1997;79:2095–2106.
- 356. Nellissery MJ, Padalecki SS, Brkanac Z, et al. Evidence for a novel osteosarcoma tumor-suppressor gene in the chromosome 18 region genetically linked with Paget disease of bone. Am J Hum Genet 1998;63:817–824.
- 357. Nora FE, Dahlin DC, Beabout JW. Bizarre parosteal osteochondromatous proliferations of the hands and feet. *AmJ Surg Pathol* 1983;7:245–250.
- Norman A, Dorfman H. Juxtacortical circumscribed myositis ossificans: evolution and radiographic features. *Radiology* 1970; 96:301–306.
- Norton KI, Hermann G, Abdelwahab IF, et al. Epiphyseal involvement in osteosarcoma. *Radiology* 1991;180:813–816.
- 360. Nuovo MA, Norman A, Chumas J, Ackerman LV. Myositis ossificans with atypical clinical, radiographic, or pathologic findings: a review of 23 cases. *Skeletal Radiol* 1992;21:87–101.
- Okada K, Frassica FJ, Sim FH, et al. Parosteal osteosarcoma. A clinicopathological study. J Bone Joint Surg Am 1994;76A: 366–378.
- Okada K, Ito H, Miyakoshi N, Sageshima M, et al. A low-grade extraskeletal osteosarcoma. *Skeletal Radiol* 2003;32:165–169.
- 363. Okada K, Kubota H, Ebina T, et al. High-grade surface osteosarcoma of the humerus. *Skeletal Radiol* 1995;24:531–534.
- 364. Onikul E, Fletcher BD, Parham DM, Chen G. Accuracy of MR imaging for estimating intraosseous extent of osteosarcoma. Am J Roentgenol 1996;167:1211–1215.
- 365. Park H-R, Jung WW, Bertoni F, et al. Molecular analysis of p53, MDM2 and H-ras genes in low-grade central osteosarcoma. *Pathol Res Pract* 2004;200:439–445.
- 366. Park H-R, Min SK, Cho HD, et al. Osteosarcoma of the ethmoid sinus. *Skeletal Radiol* 2004;33:291–294.
- 367. Partovi S, Logan PM, Janzen DL, et al. Low-grade parosteal osteosarcoma of the ulna with dedifferentiation into high-grade osteosarcoma. *Skeletal Radiol* 1996;25:497–500.
- 368. Patel SG, Meyers P, Huvos AG, et al. Improved outcomes in patients with osteogenic sarcoma of the head and neck. *Cancer* 2002;95:1495–1503.

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- Peh WCG, Shek TWH, Wang S-c, et al. Osteogenic sarcoma with skeletal muscle metastases. *Skeletal Radiol* 1999;28:298–304.
- 370. Porter DE, Holden ST, Steel CM, et al. A significant proportion of patients with osteosarcoma may belong to Li-Fraumeni cancer families. *J Bone Joint Surg Br* 1992;74B:883–886.
- 371. Price CHG, Goldie W. Paget's sarcoma of bone: a study of eighty cases from the Bristol and Leeds bone tumor registries. J Bone Joint Surg Br 1969;51B:205–224.
- 372. Pujol LÅ, Erickson RP, Heidenreich RA, Cunniff C. Variable presentation of Rothmund-Thomson syndrome. Am J Med Genet 2000;95:204–207.
- 373. Quesnel S, Verselis S, Portwine C, et al. *p53* compound heterozygosity in a severely affected child with Li-Fraumeni syndrome. Oncogene 1999;18:3970–3978.
- 374. Radig K, Schneider-Stock R, Oda Y, et al. Genetic instability in osteoblastic tumors of the skeletal system. *Pathol Res Pract* 1998;194:669–677.
- 375. Radig K, Schneider-Stock R, Oda Y, et al. Mutation spectrum of p53 gene in highly malignant human osteosarcomas. *Gen Diagn Pathol* 1996;142:25–32.
- 376. Ragazzini P, Gamberi G, Benassi MS, et al. Analysis of SAS gene and CDK4 and MDM2 proteins in low-grade osteosarcoma. *Cancer Detect Prev* 1999;23:129–136.
- 377. Rao U, Cheng A, Didolkar MS. Extraosseous osteogenic sarcoma: clinicopathological study of eight cases and review of literature. *Cancer* 1978;41:1488–1496.
- 378. Raymond AK. Surface osteosarcoma. *Clin Orthop Rel Res* 1991;270:140–148.
- 379. Raymond AK, Ayala AG, Knuutila S. Conventional osteosarcoma. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:264–270.
- 380. Resnick D, Kyriacos M, Greenway GD. Tumors and tumor-like lesions of bone: imaging and pathology of specific lesions. In: Resnick D, ed. *Diagnosis of bone and joint disorders*. Philadelphia: Saunders, 1995:3697–3699.
- Ritts GD, Pritchard DJ, Unni KK, et al. Periosteal osteosarcoma. *Clin Orthop* 1987;219:299–307.
- Robinson LH, Pitt MJ, Jaffe KA, Siegal GP. Small cell osteosarcoma of the soft tissue. *Skeletal Radiol* 1995;24:462–465.
- 383. Roessner A, Hobik HP, Grundmann E. Malignant fibrous histiocytoma of bone and osteosarcoma: a comparative light and electron microscopic study. *Pathol Res Pract* 1979; 164: 385–401.
- 384. Rosenberg A, Heim S. Extraskeletal osteosarcoma. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:182–183.
- 385. Rosenberg ZS, Lev S, Schmahmann S, et al. Osteosarcoma: subtle, rare, and misleading plain film features. *Am J Roentgenol* 1995;165:1209–1214.
- Ruggieri P, Sim FH, Bond JR, Unni KK. Malignancies in fibrous dysplasia. *Cancer* 1994;73:1411–1424.
- 387. Ruiter DJ, Cornelisse CJ, van Rijssel TG, van der Velde EA. Aneurysmal bone cyst and telangiectatic osteosarcoma. A histopathological and morphometric study. *Virchows Arch A* 1977;373:311–325.
- 388. Saifuddin A. The accuracy of imaging in the local staging of appendicular osteosarcoma. *Skeletal Radiol* 2002;31:191–201.
- 389. Saito T, Oda Y, Kawaguchi K-i, et al. Five-year evolution of a telangiectatic osteosarcoma initially managed as an aneurysmal bone cyst. *Skeletal Radiol* 2005;34:290–294.
- 390. Sandberg AA, Bridge JA. Updates on the cytogenetics and molecular genetics of bone and soft tissue tumors: osteosarcoma and related tumors. *Cancer Genet Cytogenet* 2003;145:1–30.
- 391. Sanders RP, Drissi R, Billups CA, et al. Telomerase expression predicts unfavorable outcome in osteosarcoma. J Clin Oncol 2004;22:3790–3797.
- 392. Sanerkin NG. Definitions of osteosarcoma, chondrosarcoma and fibrosarcoma of bone. *Cancer* 1980;46:178–185.
- 393. Sauer DD, Chase DR. Case report 461. Dedifferentiated parosteal osteosarcoma. *Skeletal Radiol* 1988;17:72–76.
- 394. Schajowicz F, Araujo ES, Berenstein M. Sarcoma complicating Paget's disease of bone: a clinicopathological study of 62 cases. J Bone Joint Surg Br 1983;65B:299–307.

- 395. Schajowicz F, McGuire MH, Araujo ES, et al. Osteosarcomas arising on the surfaces of long bones. J Bone Joint Surg Am 1988;70A:555–564.
- Schellenberg GD, Martin GM, Wijsman EM, et al. Homozygosity mapping and Werner's syndrome. *Lancet* 1992;339:1002.
- 397. Schreiman JS, Crass JR, Wick MR, et al. Osteosarcoma: role of CT in limb-sparing treatment. *Radiology* 1986;161:485–488.
- 398. Schulz A, Loreth B, Battmann A, et al. [Bone matrix production in osteosarcoma.] *Verh Dtsch Ges Pathol* 1998;82:144–153.
- 399. Scranton PE Jr, DeCicco FA, Totten RS, Yunis EJ. Prognostic factors in osteosarcoma. A review of 20 year's experience at the University of Pittsburgh Health Center Hospitals. *Cancer* 1975; 36:2179–2191.
- 400. Seeger LL, Eckardt JJ, Bassett LW. Cross-sectional imaging in the evaluation of osteogenic sarcoma: MRI and CT. *Semin Roentgenol* 1989;24:174–184.
- 401. Sheth DS, Yasko AW, Raymond AK, et al. Conventional and dedifferentiated parosteal osteosarcoma. Diagnosis, treatment, and outcome. *Cancer* 1996;78:2136–2145.
- 402. Shuhaibar H, Friedman L. Dedifferentiated parosteal osteosarcoma with high-grade osteoclast-rich sarcoma at presentation. *Skeletal Radiol* 1998;27:574–577.
- Sievert LJ, Elwing TJ, Evans ML. Primary pulmonary osteogenic sarcoma. Skeletal Radiol 2000;29:283–285.
- 404. Sim FH, DeVries EM, Miser JS, Unni KK. Case report 760. Osteoblastic osteosarcoma (grade 4) with Rothmund-Thomson syndrome. *Skeletal Radiol* 1992;21:543–545.
- 405. Sim FH, Kurt A-M, McLeod RA, Unni KK. Case report 628. Lowgrade central osteosarcoma. *Skeletal Radiol* 1990;19:457–460.
- 406. Šim FH, Unni KK, Beabout JW, Dahlin DC. Osteosarcoma with small cells simulating Ewing's tumor. J Bone Joint Surg Am 1979;61A:207–215.
- 407. Simon MA, Bos GD. Epiphyseal extension of metaphyseal osteosarcoma in skeletally immature individuals. J Bone Joint Surg Am 1980;62A:195–204.
- 408. Sirikulchayanonta V, Jaovisidha S. Soft tissue telangiectatic osteosarcoma in a young patient: imaging and immunostains. *Skeletal Radiol* 2005;34:295–298.
- 409. Sissons HA, Greenspan A. Paget's disease. In: Taveras JM, Ferrucci JT, eds. *Radiology: diagnosis, imaging, intervention*, vol 5. Philadelphia: JB Lippincott, 1986:1–14.
- 410. Smith J, Ahuja SC, Huvos AG, Bullough PG. Parosteal (juxtacortical) osteogenic sarcoma: a roentgenological study of 30 patients. *J Can Assoc Radiol* 1978; 29: 167–174.
- Smith J, Botet JF, Yeh SDJ. Bone sarcoma in Paget's disease: a study of 85 patients. *Radiology* 1984;152:583–590.
- Sordillo PP, Hajdu SI, Magill GB, Goldbey RB. Extraosseous osteogenic sarcoma. A review of 48 patients. *Cancer* 1983;51:727–734.
- 413. Spurney C, Gorlick R, Meyers PA, et al. Multicentric osteosarcoma, Rothmund-Thomson syndrome, and secondary nasopharyngeal non-Hodgkin's lymphoma: a case report and review of the literature. *J Pediatr Hematol Oncol* 1998;20:494–497.
- Stevens GM, Pugh DG, Dahlin DC. Roentgenographic recognition and differentiation of parosteal osteogenic sarcoma. Am J Roentgenol 1957;78:1–12.
- 415. Stevenson AJ, Chatten J, Bertoni F, Miettinen M. CD99 (p30/32MIC2) neuroectodermal/Ewing's sarcoma antigen as an immunohistochemical marker. *Appl Immunohistochem* 1994; 2:231–240.
- Sundaram M, McGuire MH, Herbold DR. Magnetic resonance imaging of osteosarcoma. *Skeletal Radiol* 1987;16:23–29.
- 417. Szymanska J, Mandahl N, Mertens F, et al. Ring chromosomes in parosteal osteosarcoma contain sequences from 12q13-15: a combined cytogenetic and comparative genomic hybridization study. *Genes Chromosomes Cancer* 1996;16:31–34.
- 418. Taconis WK. Osteosarcoma and its variants. J Med Imag 1988; 2:276–285.
- Tani T, Okada K, Shoji K, et al. Osteoblastoma-like osteosarcoma. Skeletal Radiol 2000;29:656–659.
- 420. Tarkkanen M, Böhling T, Gamberi G, et al. Comparative genomic hybridization of low-grade central osteosarcoma. *Mod Pathol* 1998;11:421–426.
- 421. Tarkkanen M, Elomaa I, Blomqvist C, et al. DNA sequence copy number increase at 8q: a potential new prognostic marker in high-grade osteosarcoma. *Int J Cancer* 1999;84:114–121.

- 422. Tarkkanen M, Wiklund TA, Virolainen MJ, et al. Comparative genomic hybridization of postirradiation sarcomas. *Cancer* 2001;92:1992–1998.
- 423. Tateishi U, Hasegawa T, Miyakawa K, et al. CT and MRI features of recurrent tumors and second primary neoplasms in pediatric patients with retinoblastoma. *Am J Roentgenol* 2003;181:879– 884.
- 424. Tokunaga A, Onda M, Matsukura N, Kato S. [Li-Fraumeni syndrome.] Nippon Rinsho 1995;53:2797–2802.
- 425. Torres FX, Kyriakos M. Bone infarct-associated osteosarcoma. *Cancer* 1992;70:2418–2430.
- 426. Tsuji Y, Kusuzaki K, Kanemitsu K, et al. Calcaneal osteosarcoma associated with Werner syndrome. A case report with mutation analysis. *J Bone Joint Surg Am* 2000;82A:1308–1313.
- 427. Tsuneyoshi M, Dorfman HD. Epiphyseal osteosarcoma: distinguishing features from clear cell chondrosarcoma, chondroblastoma and epiphyseal enchondroma. *Hum Pathol* 1987;18: 644–651.
- 428. Tucker MA, D'Angio GJ, Boice JD Jr, et al. Bone sarcomas linked to radiotherapy and chemotherapy in children. N Engl J Med 1987;317:588–593.
- 429. Ulaner GA, Huang HY, Otero J, et al. Absence of a telomere maintenance mechanism as a favorable prognostic factor in patients with osteosarcoma. *Cancer Res* 2003;63:1759–1763.
- 430. Unni KK. Dahlin's bone tumors. General aspects and data on 11,087 cases, 5th ed. Philadelphia: Lippincott-Raven; 1996:463.
- Unni KK. Osteosarcoma of bone. In: Unni KK, ed. Bone tumors. New York: Churchill Livingstone, 1988:107–133.
- 432. Unni KK, Dahlin DC. Grading of bone tumors. Semin Diagn Pathol 1984;1:165–172.
- Unni KK, Dahlin DC. Premalignant tumors and conditions of bone. Am J Surg Pathol 1979;3:47–60.
- Unni KK, Dahlin DC, Beabout JW. Periosteal osteogenic sarcoma. Cancer 1976;37:2476–2485.
- Unni KK, Dahlin DC, Beabout JW, Ivins JC. Parosteal osteogenic sarcoma. *Cancer* 1976;37:2466–2475.
- Unni KK, Dahlin DC, McLeod RA. Intraosseous well-differentiated osteosarcoma. *Cancer* 1977;40:1337–1347.
- 437. Van Der Heul RO, von Ronnen JR. Juxtacortical osteosarcoma. Diagnosis, differential diagnosis, treatment, and an analysis of eighty cases. *J Bone J Surg Am* 1967;49A:415–439.
- 438. van der Woude H-J, Bloem JL, Verstraete KL, et al. Osteosarcoma and Ewing's sarcoma after neoadjuvant chemotherapy: Value of dynamic MR imaging in detecting viable tumor before surgery. *Am J Roentgenol* 1995;165:593–598.
- Vanel D, Picci P, De Paolis M, Mercuri M. Osteosarcoma arising in an exostosis: CT and MR imaging. *Am J Roentgenol* 2001;176: 259–260.
- 440. Vanel D, Picci P, De Paolis M, Mercuri M. Radiological study of 12 high-grade surface osteosarcomas. *Skeletal Radiol* 2001;30: 667–671.
- 441. Vanhoenacker F, De Beuckeleer LH, De Schepper AM. Intracortical osteosarcoma: is MRI useful? *ROFO* 2001;173:959–960.
- 442. van Trommel MF, Kroon HM, Bloem JL, et al. MR imaging based strategies in limb salvage surgery for osteosarcoma of the distal femur. *Skeletal Radiol* 1997;26:636–641.
- 443. Varma DGK, Ayala AG, Guo S-Q, et al. MRI of extraskeletal osteosarcoma. J Comput Assist Tomogr 1993;17:414–417.
- 444. Vasquez E, Castellote J, Piqueras P, et al. Second malignancies in pediatric patients: imaging findings and differential diagnoses. *RadioGraphics* 2003;23:1155–1172.
- 445. Verela-Duran J, Enzinger FM. Calcifying synovial sarcoma. *Cancer* 1982;50:345–352.
- Vigorita VJ, Jones JK, Ghelman B, Marcove RC. Intracortical osteosarcoma. Am J Surg Pathol 1984;8:65–71.
- 447. Vuillemin-Bodaghi V, Parlierk-Cuau C, Cywiner-Golenzer C, et al. Multifocal osteogenic sarcoma in Paget's disease. *Skeletal Radiol* 2000;29:349–353.
- 448. Wang LL, Gannavarapu A, Kozinetz CA, et al. Association between osteosarcoma and deleterious mutations in the RECQL4 gene in Rothmund-Thomson syndrome. J Natl Cancer Inst 2003;95:669–674.
- 449. Waxman M, Vuletin JC, Saxe BI, Monteleone FA. Extraskeletal osteosarcoma. Light and electron microscopic study. *Mt Sinai J Med* 1981;48:322–329.

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- 450. Wenger DE, Sundaram M, Unni KK, et al. Microscopic correlation of radiographically disparate appearing well differentiated osteosarcoma. *Skeletal Radiol* 2002;31:488–492.
- 451. Werner M, Rieck J, Delling G. [Cytogenetic changes in low grade central osteosarcomas.] Verh Dtsch Ges Pathol 1998;82: 189–194.
- 452. Wines A, Bonar F, Lam P, et al. Telangiectatic dedifferentiation of a parosteal osteosarcoma. *Skeletal Radiol* 2000;29:597–600.
- Wold LE. Fibrohistiocytic tumors of bone. In: Unni KK, ed. Bone tumors. New York: Churchill Livingstone, 1988:183–197.
- 454. Wold LE, Unni KK, Beabout JW, Pritchard DJ. High-grade surface osteosarcomas. *Am J Surg Pathol* 1984;8:181–186.
- 455. Wold LE, Unni KK, Beabout JW, et al. Dedifferentiated parosteal osteosarcoma. *J Bone Joint Surg* 1984;66A:53–59.
- 456. Wong KT, Haygood T, Ďalinka MK, Kneeland B. Chondroblastic, grade 3 periosteal osteosarcoma. *Skeletal Radiol* 1995;24: 69-71.
- 457. Wu RK, Trumble TE, Ruwe PA. Familial incidence of Paget's disease and secondary osteogenic sarcoma. A report of three cases from a single family. *Clin Orthop Relat Res* 1991;265:306–309.

- 458. Wunder JS, Eppert K, Burrow SR, et al. Co-amplification and overexpression of CDK4, SAS and MDM2 occurs frequently in human parosteal osteosarcomas. *Oncogene* 1999;18:783–788.
- Wurlitzer F, Ayala A, Romsdahl M. Extraosseous osteogenic sarcoma. Arch Surg 1972;105:691–695.
- 460. Yamaguchi H, Nojima T, Yagi T, et al. High-grade surface osteosarcoma of the left ilium. A case report and review of the literature. *Acta Pathol Jpn* 1988;38:235–240.
- 461. Yamaguchi T, Shimizu K, Koguchi Y, et al. Low-grade central osteosarcoma of the rib. *Skeletal Radiol* 2005;34:490–493.
- 462. Yoshida H, Yumoto T, Minamizaki T. Osteosarcoma with features mimicking malignant fibrous histiocytoma. Virchows Arch A Pathol Anat Histopathol 1992;421:229–238.
- 463. Zambrano E, Nose V, Perez-Atayde AR, et al. Distinct chromosomal rearrangements in subungual (Dupuytren) exostosis and bizarre parosteal osteochondromatous proliferation (Nora lesion). Am J Surg Pathol 2004;28:1033–1039.
- 464. Zimmer WD, Berquist TH, McLeod RA, et al. Bone tumors: magnetic resonance imaging versus computed tomography. *Radiology* 1985;155:709–718.

CHAPTER 3

Cartilage (Chondrogenic) Lesions

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Diagnosis of a bone lesion as originating from cartilage is usually a simple task for the radiologist. The lesion's radiolucent matrix, scalloped margin, and annular, punctate, or comma-shaped calcifications usually suffice to establish its chondrogenic nature (Fig. 3-1). However, whether a cartilage tumor is benign or malignant is sometimes extremely difficult for the radiologist and the pathologist to determine (33,49,62,70).

All cartilage tumors, whether benign or malignant, exhibit a positive reaction for S-100 protein (54). This is sometimes diagnostically useful, as most other bone tumors do not display this reaction. Histologically, cartilage tumors are usually recognized by the features of their intercellular matrix, which has a uniformly translucent appearance and contains less collagen compared with the intercellular matrix of osteoblastic tumors, such as osteoblastoma and osteosarcoma. The tumor cells themselves are usually located in rounded spaces (lacunae), as in normal cartilage. In benign cartilage tumors, such as enchondroma, the tissue is sparsely cellular (Fig. 3-2). The cells, typically with small, dark-staining nuclei, fail to show the cytologic features that are characteristic of chondrosarcoma, such as large nuclei; pleomorphism; large, swollen chondroblastic and double nuclei; and mitoses. The tumor tissue is usually avascular, and areas of degenerate and calcified matrix are common. Calcification is sometimes followed by vascularization and replacement by bone, as in normal endochondral ossification. Because of their slow growth, enchondromas erode and expand the overlying cortical bone (Fig. 3-3) but do not invade it, unlike chondrosarcomas. Nevertheless, some slow-growing, low-grade chondrosar-



Figure 3-1 Cartilage-forming tumor. The radiologic characteristics of a radiolucent matrix, scalloped margin, and annular and comma-shaped calcifications establish the chondrogenic nature of this lesion (in this example—chondrosarcoma).



Figure 3-2 Histopathology of a benign cartilage lesion. Sparsely cellular tissue and uniform-sized chondrocytes located in rounded lacunae are characteristic features of enchondroma (hematoxylin and eosin, original magnification ×235).



Figure 3-3 Histopathology of enchondroma. A: Wholemount section (hematoxylin and eosin) shows the tumor eroding the cortex (endosteal scalloping). B: On higher magnification there is no evidence of invasion (hematoxylin and eosin, original magnification $\times 183$).



comas can masquerade as enchondromas, and some aggressive-looking benign cartilage lesions may be mistakenly diagnosed as chondrosarcomas. Moreover, it is important to be aware of the capricious biological behavior displayed by cartilage tumors. Some benign-appearing lesions of cartilage may behave in an aggressive or malignant manner, whereas tumors that present an ominous appearance on radiologic or histologic examination may show limited progression.

Clinically, malignancy is suggested by a variety of signs, including development of pain in a previously asymptomatic lesion (e.g., malignant transformation of enchondroma to chondrosarcoma), development of swelling or a mass at the site of a lesion, or rapid growth of a lesion (e.g., malignant transformation of osteochondroma to chondrosarcoma). Radiologically, the presence of a periosteal reaction, destruction of the cortex, and a soft tissue mass are almost pathognomonic features of malignancy (25). The histopathologic features of a malignant cartilage tumor include the presence of hypercellular and pleomorphic tumor tissue, appreciable numbers of plump cells with large or double nuclei, invasion (permeation) of bone trabeculae, and infiltration of Haversian systems and bone marrow (Fig. 3-4). However, these obvious features of malignancy may be not so prominent in biopsy specimen, or only a few areas in such a specimen may reveal the diagnostic features of malignancy. As Dahlin and Unni (13) pointed out, in the pathologic evaluation it is necessary to examine many fields in these tumors to be certain that there is sufficient evidence for a diagnosis of malignancy because evidence that a tumor is a chondrosarcoma rather than a benign enchondroma is frequently found in isolated areas within the mass. Therefore, it is of paramount importance to obtain biopsy specimens from several areas of a tumor. Close cooperation among the clinician, the radiologist, and the pathologist in reviewing the history, the radiologic examination, and the biopsy specimens is crucial to reaching a conclusion on the exact nature of a cartilage lesion.

Figure 3-4 Histopathology of chondrosarcoma. Note invasion of trabeculae by markedly cellular tissue of chondrosarcoma (hematoxylin and eosin, original magnification $\times 25$). (Courtesy Dr. M. J. Klein, Birmingham, Alabama.)

From the radiologic standpoint, the differential diagnosis of cartilage lesions should focus on the following three points:

- 1. Distinguishing a benign cartilage tumor from the noncartilaginous lesions (e.g., calcifying enchondroma from bone infarct; enchondroma located near the articular end of bone from giant cell tumor, osteoarthritic geode, or intraosseous ganglion; or chondromyxoid fibroma from aneurysmal bone cyst).
- 2. Distinguishing a benign cartilage tumor from a malignant one (e.g., enchondroma from low-grade chondrosarcoma; osteochondroma from exostotic chondrosarcoma; periosteal chondroma from periosteal chondrosarcoma; or synovial chondromatosis from synovial chondrosarcoma).
- 3. Distinguishing a malignant cartilage tumor from a noncartilaginous lesion [e.g., chondrosarcoma at the articular end of bone from giant cell tumor or from malignant fibrous histiocytoma (MFH); or periosteal chondrosarcoma from periosteal osteosarcoma].

From the histopathologic standpoint, because the microscopic features of cartilage lesions are obvious and characteristic, focus should be directed to the following:

- 1. Recognition of the varieties of benign cartilage tumors (e.g., enchondroma, periosteal chondroma, chondroblastoma, chondromyxoid fibroma) and various subtypes of chondrosarcoma (e.g., clear cell, mesenchymal, or periosteal).
- Recognition of benign versus malignant tumor (e.g., enchondroma vs. well-differentiated chondrosarcoma; chondromyxoid fibroma vs. myxoid chondrosarcoma; or chondroblastoma vs. mesenchymal chondrosarcoma).
- 3. Differentiation of chondrosarcoma from other malignant tumors that contain cartilage (e.g., from chondroblastic osteosarcoma or periosteal osteosarcoma).

Benign Lesions

Enchondroma

Enchondroma is the second most common benign tumor of bone and is characterized by the formation of mature hyaline cartilage. It constitutes about 10% of all benign bone tumors and represents the most common tumor of the phalanges of the hand. Several theories have been postulated regarding its etiology. Among these is that the enchondroma develops from abnormal zones of dysplastic chondrocytes in the growth plate. These abnormal foci fail to undergo normal endochondral ossification; instead, they are deposited within the metaphysis and, as the bone grows, are displaced into the diaphysis (49). Customarily, a lesion located centrally in the bone is called an enchondroma, whereas one located outside the cortex is called a chondroma (periosteal or juxtacortical). Separate groups consist of lesions (a) located in the soft tissues and (b) located in the joint, bursa, or in the tendon sheath. The former are called soft tissue chondromas and the latter synovial chondromatosis (69).

Clinical Presentation

Although the tumor may occur at any age, most cases arise between the second and fourth decades; there is no sex predilection. The short, tubular bones of the hand and foot (phalanges, metacarpals, and metatarsals) are preferred sites, followed in frequency by the femur, humerus, tibia, and ribs (Fig. 3-5). The lesion is often asymptomatic and usually is discovered incidentally due to a pathologic fracture through the tumor.

The single most important complication of the lesion is malignant transformation to chondrosarcoma. In a solitary enchondroma this occurs predominantly in a long or flat bone and almost never in a short tubular bone. An important clinical sign is development of pain at the site of the lesion in a previously asymptomatic patient and in the absence of a fracture.

Imaging

Radiography is usually sufficient to demonstrate the lesion and to establish its chondroid character. In short, tubular bones, the lesion is often entirely radiolucent (Fig. 3-6), but in long bones it may display visible calcifications (24). This flocculent mineralization usually assumes the form of dots, rings, and arcs, giving a "popcorn" appearance (Fig. 3-7). If the calcifications are extensive, the lesion is called a calcifying enchondroma (Fig. 3-8). The cortex is usually thinned and expanded in a symmetric, fusiform fashion. The often cloudy or hazy appearance of a lesion provides a clue to its chondroid composition (18). The lesion can also be recognized by scalloped inner cortical margins, reflecting the lobular growth pattern of cartilage (Fig. 3-9). Computed tomography (CT) and magnetic resonance imaging (MRI) may further delineate the tumor and more precisely localize it in the bone (Fig. 3-10). Because in long bones enchondroma may present as a radiolucent lesion with poorly defined margins, particularly in areas where trabeculae are sparse or absent (diaphysis), MRI may be extremely helpful in revealing the exact size and extent of the tumor (Fig. 3-11) (74). Like most cartilaginous and fibrous lesions, enchondroma exhibits low to intermediate signal intensity on T1-weighted images and high signal intensity on T2-weighted sequences. On gradient echo sequences the lesion may appear less bright than on conventional T2-weighted images (see Fig. 3-11E). The calcifications image either as signal void or as lowsignal-intensity foci. After administration of gadolinium, enchondroma demonstrates a typical pattern of signal enhancement, which is helpful in distinguishing this lesion from noncartilaginous tumors (52,218).

Histopathology

On histologic examination, enchondroma consists of lobules of hyaline cartilage of variable cellularity and can be recognized by the features of its intercellular matrix, which has a uniformly translucent appearance



Figure 3-5 Enchondroma: skeletal sites of predilection, peak age range, and male-to-female ratio.



Figure 3-6 Enchondroma in short tubular bones. A: Typical, purely radiolucent lesion at the base of the proximal phalanx of the ring finger in a 37-year-old woman. Note the marked attenuation of the ulnar side of the cortex. **B:** Radiolucent lesion at the base of the proximal phalanx of the thumb with a pathologic fracture.



Figure 3-7 Enchondroma in a long bone. Anteroposterior **(A)** and lateral **(B)** radiographs of the femur show a radiolucent lesion with "popcorn" calcifications.

В



Figure 3-8 "Calcifying enchondroma." A heavily calcified lesion in the proximal humerus of a 58-year-old woman.

and contains relatively little collagen (Fig. 3-12A). The lesion may slightly erode the endosteal surface of cortical bone; however, no invasion of Haversian systems is present. The tumor cells are located in lacunae. The tissue is sparsely cellular and the cells contain small, dark-staining nuclei (Fig. 3-12B). Binucleated cells and small nucleoli are sometimes present (51) (Fig. 3-12C). Calcifications are common and correspond to matrix calcifications or actual endochondral ossifications at the periphery of cartilage lobules (Fig. 3-13). Enchondromas located in the short tubular bones of hands and feet may exhibit increased cellularity, plump nuclei, and occasionally even binucleated cells.

Only a few cytogenetic studies with similar results have been reported on enchondromas, periosteal chondromas, and soft tissue chondromas, which are therefore discussed together. Flow cytometry studies revealed DNA-diploid peaks, although DNA-aneuploidy has rarely been observed (2,35). Most chondromas contain clonal chromosomal abnormalities involving chromosomes or chromosomal regions 4q, 5, 7, 11,14q, 16q22–q24, 20, and particularly rearrangements of chromosome 6 and 12q12–q15 (8). Tumor-specific anomalies, however, were not detected.

Periosteal (Juxtacortical) Chondroma

Periosteal or juxtacortical chondroma, a slow-growing lesion, affects the surface of a bone in or beneath the periosteum. Preferred sites include the proximal humerus, femur, tibia, and the phalanges of the hand



Figure 3-9 Enchondroma. A radiolucent lesion exhibits central calcifications and scalloped inner cortical margins, reflecting the lobular growth pattern of cartilage.

(52). This is an uncommon neoplasm, accounting for about only 0.66% of bone tumors in the Mayo Clinic series (56). Periosteal chondroma was first described by Lichtenstein and Hall in 1952 (39). It occurs in children and adults (most commonly in the second and third decades), who usually present with a history of pain and local tenderness, often accompanied by swelling at the involved site.

The radiographic appearance is rather characteristic (6,65). The lesion is small and is located on the surface of the bone (Fig. 3-14). As it enlarges, it erodes the underlying cortex in a saucer-like fashion, displaying a sclerotic rim and producing a buttress of periosteal new bone formation (15). The lesion may contain calcifications (Fig. 3-15). CT may show the scalloped cortex, matrix calcification (Fig. 3-16), and cortical shell (28,36). It also may demonstrate the separation of a lesion from the medullary cavity, an important feature in differentiation from osteochondroma (26). MRI findings correspond to radiographic findings, depicting the cartilaginous soft tissue component (76). If periosteal chondroma affects the medullary canal, MRI may be useful in depicting the extent of involvement (Fig. 3-17). Fat suppression







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Ε







Figure 3-12 Histopathology of enchondroma. A: At low magnification the lesion exhibits hyaline cartilage of low to moderate cellularity (hematoxylin and eosin, original magnification \times 50). **B:** At higher magnification the chondrocytes with darkly stained nuclei are seen to be located in the lacunae (hematoxylin and eosin, original magnification \times 100). **C:** Occasionally binucleated cells may be present (hematoxylin and eosin, original magnification \times 150) (**C**, courtesy of Dr. K. K. Unni, Rochester, Minnesota.)

В



Figure 3-13 Histopathology of enchondroma. Cartilage lobules are surrounded by a narrow rim of bone, representing endochondral ossification (hematoxylin and eosin, original magnification $\times 250$).

С



Figure 3-14 Periosteal chondroma. A: Radiolucent lesion erodes the external surface of the cortex of the proximal humerus (*arrow*) of a 24-year-old man. **B:** Well-defined, saucer-like erosion of the cortex of the proximal phalanx is characteristic of periosteal chondroma. (Reprinted from Bullough PG. *Atlas of orthopedic pathology,* 2nd ed. New York: Gower, 1992:16.22.) **C:** Metaphyseal periosteal chondroma affecting the fifth metacarpal bone erodes the cortex and evokes the buttress of periosteal reaction (*arrow*).



В



Figure 3-15 Periosteal chondroma. A lesion displaying calcifications erodes the medial cortex of the femoral neck (*arrow*). The buttress of a periosteal reaction seen at the inferior border of the lesion (*open arrow*) is characteristic of periosteal chondroma.





Figure 3-16 Periosteal chondroma: computed tomography (CT). A: Radiograph shows a lesion with calcifications eroding the medial cortex of the distal fibula (*arrow*). CT using a bone window (**B**) and a soft tissue window (**C**) better demonstrates the extent of the lesion and the distribution of the calcifications.

С

Α


or enhanced gradient echo sequences may improve tumor-marrow contrast (5). The potential pitfall of MRI is marrow edema mimicking tumor invasion or vice versa (76). Unlike enchondroma and osteochondroma, periosteal chondroma may continue to grow after skeletal maturation (20). Some lesions may attain a large size (up to 6 cm) and may resemble osteochondromas (26) (Fig. 3-18; also see Fig. 3-34). Some lesions may mimic an aneurysmal bone cyst. Very rarely the lesion may encase itself intracortically, thus mimicking other intracortical lesions such as intracortical angioma, intracortical fibrous dysplasia, intracortical bone abscess (1), or osteoid osteoma (63,67).

On histologic examination, the findings are identical to those of enchondroma, although the lesion sometimes exhibits a higher level of cellularity, occasionally with atypical and binucleated cells (Fig. 3-19). Well-formed lamellar bone or fibrous connective tissue separates the cartilaginous nodules, and in some instances deposition of calcium can be observed (16).

Enchondromatosis, Ollier Disease, and Maffucci Syndrome

Enchondromatosis is a condition marked by multiple enchondromas, usually involving the metaphyses and diaphyses (21). If skeletal involvement is extensive (42), and particularly if the involvement is unilateral [resembling the original case described by Ollier (50,58,59,72)], the term "Ollier disease" is applied



Figure 3-18 Periosteal chondroma. Large lesion eroding the cortex of the proximal humerus resembles a sessile osteochondroma. Note the periosteal reaction and separation of the tumor from the medullary cavity by a cortex, features that help in the differentiation from osteochondroma. (Courtesy Dr. K. K. Unni, Rochester, Minnesota.)

(68). There is no hereditary or familial tendency in this disorder, which some investigators consider to be developmental rather than neoplastic, classifying it as a form of bone dysplasia caused by failure of normal endochondral ossification (45). The femur, tibia, and ilium are the most commonly affected bones (72), followed by the phalanges, metacarpals, and metatarsals. Much less often the lesions involve the facial bones, skull, spine, carpal, and tarsal bones (17,48) (Fig. 3-20).

Maffucci syndrome is a congenital, nonhereditary disorder, characterized by enchondromatosis and soft tissue angiomatosis (hemangiomatosis) (19). The latter may occur anywhere in the skin and subcutaneous tissue. The hemangiomas are usually cavernous and may be unilateral or bilateral, localized or extensive. The skeletal lesions in Maffucci syndrome have predilection for tubular bones and have the same distribution as those in Ollier disease, with a strong predisposition for one side of the body (75). The most frequent sites of enchondromas are the metacarpals and the phalanges of the hands (37).

Recent investigations suggest that Ollier disease and Maffucci syndrome represent two entities within a continuum of the disease multiple enchondromatosis and that both conditions bear the risk of mesodermal as well as nonmesodermal malignancy (27).

Clinical Presentation

The condition has a strong preference for involvement of one side of the body. The clinical manifestations, such

as knobby swellings of the digits or gross disparities in limb length, are commonly recognized during childhood and adolescence. The affected child often begins to limp during the second year of life as a disparity in leg length becomes apparent (long bones that are affected are always shorter because the growth plate is involved). Pain is uncommon and when present is usually due to a pathologic fracture (52). Ollier disease is often arrested at puberty but may occasionally progress after that time. The most common and severe complication of Ollier disease is malignant transformation of one or more enchondromas to chondrosarcoma (34,41) (see Figs. 3-118 and 3-119). Schajowicz (68) believed that about 30% of the patients with Ollier disease will develop chondrosarcoma, but Jaffe (30) and Mirra et al. (49) have estimated that risk to be as high as 50%. However, because patients with enchondromatosis or Ollier disease may develop hundreds of enchondromas, the statistical risk of one given lesion to undergo malignant degeneration is probably not higher than in solitary enchondroma. Unlike solitary enchondromas, in patients with enchondromatosis even lesions in the short tubular bones may undergo sarcomatous changes. This is particularly true in patients with Maffucci syndrome (73).

Imaging

The radiographic appearance of enchondromatosis involving the hands and feet is quite characteristic. Radiolucent masses of cartilage with foci of calcifications closely resembling solitary enchondromas markedly deform the bones (Fig. 3-21). Enchondromas in this location may be intracortical and periosteal (50). They sometimes protrude from the shaft of the short or long bone, thus resembling osteochondroma (Fig. 3-22). However, unlike osteochondroma, these protruding enchondromas have neither a cartilaginous cap nor a bony stalk. Also unlike osteochondroma, the protrusions do not point away from the adjacent joint (50). In the long bones, columns of radiolucent streaks extend from the growth plate into the diaphysis. Coalescence of metaphyseal enchondromas often causes asymmetric enlargement of the long bones and flaring of the metaphyses. Interference with the growth plate causes foreshortening and deformity of the bones (Fig. 3-23). In the pelvis, and particularly in the iliac bones, a fan-like linear pattern is characteristic (Fig. 3-24). Maffucci syndrome is recognized radiographically by multiple calcified phleboliths in addition to the typical alterations of enchondromatosis (Fig. 3-25) (45).

Histopathology

Histologically, the lesions in enchondromatosis are essentially indistinguishable from those of solitary enchondromas, although they sometimes tend to be predominantly myxoid, more cellular, and may exhibit more cellular atypia (Fig. 3-26). Mitoses and prominent nucleoli are uncommon, and anaplasia is not observed (13). However, binucleated cells are often present.

Both Ollier disease and Maffucci syndrome occur sporadically although familial cases have been described (44). In a molecular genetic study Hopyan et al. (29) demonstrated a mutation in the gene of the parathyroid





Figure 3-19 Histopathology of periosteal chondroma. A: At the interface with adjacent cortex, the cartilaginous tumor (*right*) erodes the bone (hematoxylin and eosin, whole-mount section). B: The tumor tissue is more cellular than conventional enchondroma, and some cells appear atypical (hematoxylin and eosin, original magnification $\times 100$). C: Hyaline cartilage with a rather dense cell population, slight cell pleomorphism, and signs of cell proliferation. These histologic variations are compatible with a benign cartilage lesion in a periosteal location (hematoxylin and eosin, original magnification $\times 200$).



С



Figure 3-20 Enchondromatosis (Ollier disease): skeletal sites of predilection, peak age range, and male-to-female ratio.



Figure 3-21 Enchondromatosis (Ollier disease). Large, lobulated cartilaginous masses markedly deform the bones of the hand.

hormone receptor 1 (*PTHR1*), one germline and one somatic mutation, in two of six patients with Ollier disease leading to an abnormal signaling and, in transgenic mice, to enchondroma-like lesions. However, in an extensive study on 23 enchondromas and 18 chondrosarcomas from 31 patients with enchondromatosis, Rozeman et al. (66) could not confirm these findings or demonstrate any different mutation in the *PTHR1* gene.

Only two other genetic analyses in chondrosarcomas developing in enchondromatosis have been reported so far. Using cytogenetics, Ozisik et al. (60) showed an interstitial deletion [del(1)(p11p31.2)] at the short arm of chromosome 1, whereas Bovée et al. (7) observed loss of heterozygosity (LOH: loss of one allele in the tumor DNA compared to normal germline DNA of an individual) at 13q14, the locus of the tumor suppressor gene *RB1* (retinoblastoma gene), and at 9p21 among others where two cell cycle regulating genes (p15/p16) are located.

Differential Diagnosis

Radiology

The main differential diagnosis in **solitary enchondroma**, particularly when it is located in long bones **Figure 3-22 Enchondromatosis (Ollier disease).** Multiple lesions are observed in the phalanges and metacarpals. The intracortical lesion in the metaphysis of the fourth metacarpal protrudes from the bone (*arrow*), thus resembling an osteochondroma.



Figure 3-23 Enchondromatosis (Ollier disease). Anteroposterior radiograph of both legs of a 17year-old boy shows growth stunting and marked deformities of the tibiae and fibulae.





Figure 3-24 Enchondromatosis (Ollier disease). Anteroposterior radiograph of the pelvis demonstrates crescent-shaped and ring-like calcifications in tongues of cartilage extending from the iliac crests and the proximal femora, classic features of Ollier disease.



Figure 3-25 Maffucci syndrome. Radiograph of the hand reveals typical changes of enchondromatosis, accompanied by calcified phleboliths in soft tissue hemangiomas. (Reprinted from Bullough PG. *Atlas of orthopedic pathology,* 2nd ed. New York: Gower, 1992:14.9.)

and exhibits substantial calcifications ("calcifying enchondroma"), is *medullary bone infarct* (Fig. 3-27). At times the two lesions are difficult to distinguish from one another because of similar calcifications, particularly when the enchondroma is small. A number of radiographic features can be helpful in the differential diagnosis. These include lobulation of the margins of an enchondroma, punctate or annular calcifications within the matrix, and absence of the peripheral sclerotic fibro-osseous wall commonly observed in bone infarcts (18). Mineral deposits also tend to exhibit a more central distribution, and the calcifications often have a



Figure 3-26 Histopathology of enchondromatosis. Lobular and cellular appearance of cartilaginous nodules. These lesions usually exhibit more cellularity than is seen in solitary enchondroma (hematoxylin and eosin, original magnification \times 100).







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markedly stippled appearance (Fig. 3-28). Although the ring-like calcifications typical of enchondroma are occasionally present in medullary bone infarct, they are usually larger and have thicker walls. As a rule, bone infarct, unlike enchondroma, does not expand bone contours (51) except in rare cases of encystification (57) (see Fig. 7-54). The distribution of the medullary bone infarcts that commonly affect the proximal and distal femur and the proximal tibia may be helpful in the differential diagnosis.

The most difficult task for the radiologist is to distinguish a large solitary enchondroma from a slowly growing *low-grade chondrosarcoma* (53). One of the most significant findings pointing to a chondrosarcoma in the early stage of development is localized thickening of the cortex (Fig. 3-29). Deep endosteal scalloping (greater than two thirds of cortical thickness) is another sign of possible malignancy (53). The size of the lesion should also be taken into consideration: lesions longer than 4 cm are suggestive of malignancy (Fig. 3-30). In advanced lesions, destruction of the cortex, periosteal reaction of aggressive type, and the presence of a soft tissue mass are the hallmarks of malignancy.

It is equally important to distinguish sarcomatous transformation in patients with Ollier disease and Maffucci syndrome. Radiography will reveal intralesional radiolucencies and cortical destruction associated with a soft tissue mass (see Fig. 3-118).

If enchondroma, particularly in a short, tubular bone, extends to the articular end of the bone, *giant*



Figure 3-28 Enchondroma. Typical appearance of annular and stippled calcifications within a radiolucent lesion.



Figure 3-29 Low-grade chondrosarcoma. A 48-year-old woman presented with pain in the upper leg. Radiograph shows a radiolucent lesion in the proximal tibia with a wide zone of transition and central calcifications. Note the thick-ening of the cortex, a feature that distinguishes chondrosarcoma from similarly appearing enchondroma.

cell tumor should be included in the differential diagnosis. The latter contains no calcifications, an important differential clue. However, some enchondromas may not display visible calcifications. In addition, giant cell tumor usually lacks a sclerotic border. In the long, tubular bones, this distinction is much easier because enchondroma, unlike giant cell tumor, only rarely extends into the articular end of bone (Fig. 3-31). Enchondroma may mimic *solitary bone cyst*, although the latter is a rare occurrence in the short, tubular bones. If an enchondroma is found in the distal phalanx of the hand and does not contain calcifications, an *epidermoid cyst* is a differential possibility.

An **intracortical enchondroma**, or an enchondroma that causes nonsymmetric cortical expansion, may be mistaken for *osteochondroma* (see Fig. 3-22). However, the



Figure 3-30 Low-grade chondrosarcoma. A: Conventional radiograph in a 48-year-old man shows a cartilaginous lesion in the proximal right humerus. Although the cortex is not thickened, the length of the lesion (15 cm) suggests malignancy. Coronal T1-weighted **(B)** and T2-weighted **(C)** magnetic resonance imaging shows heterogeneous signal intensity, which may be seen in both enchondroma and chondrosarcoma. On biopsy, the tumor proved to be a low-grade chondrosarcoma.

distribution of the mineral matrix visualized within enchondroma usually helps to distinguish the two entities.

Enchondroma protuberans may create a problem in diagnosis (10). Enchondroma protuberans has been defined as an exophytic enchondroma of a long bone, although it may affect a rib as well (31). It arises in the medullary cavity but penetrates the cortex, forming a prominent exophytic mass on the surface of bone, and thus can mimic any surface lesion (e.g., *osteoma, osteo-chondroma, periosteal chondroma, myositis ossificans*). It also should be distinguished from *chondrosarcoma*, either primary or one developing in a preexisting enchondroma. Further distinction must be made from the simultaneous occurrence of an osteochondroma and an enchondroma in the same bone (64) (Fig. 3-32).

Ollier disease almost never creates a diagnostic problem. Only occasionally the Jansen type of *metaphyseal chondrodysplasia*, a skeletal disorder manifested by radiologically detectable, generalized, symmetric metaphyseal abnormalities, may mimic the former condition. Widening and flaring of the metaphyses and irregularity of the zones of provisional calcification may be mistaken for enchondromas. However, other features of this dysplasia, such as rhizomelic



Figure 3-31 Enchondroma resembling giant cell tumor. The lesion extends into the articular end of the distal radius and therefore may mimic a giant cell tumor. However, unlike the latter, there are a few faintly visualized calcifications and a sclerotic margin.

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Figure 3-32 Differential diagnosis of enchondroma protuberans.

short-limb dwarfism, enlarged joints, craniofacial changes including large skull with frontonasal hyperplasia, and hyperostosis of the cranial vault and base of the skull, should be helpful in arriving at the correct diagnosis.

Periosteal chondroma should be radiographically differentiated from a variety of other periosteal lesions (e.g., *periosteal ganglion, periosteal Ewing sarcoma*), as well as from *myositis ossificans* and *tumoral calcinosis*. Features of *myositis ossificans* have been described in the section on extraskeletal osteosarcoma. The pattern of zoning phenomenon is diagnostic of the former lesion. Features of *tumoral calcinosis* (Fig. 3-33) have been described in the section on synovial sarcoma (see Chapter 9).

A large periosteal chondroma may closely resemble *sessile osteochondroma* and vice versa. The key to distinguishing between these two lesions is the separation of periosteal chondroma from the medullary portion of the bone by intervening cortex (26) (Fig. 3-34). Conversely, in a typical osteochondroma the cortex of the host bone merges with the cortex of the lesion, and continuity exists between the medullary cavity of the host bone and the exostosis (see Fig. 3-38B). Another useful characteristic is the value of attenuation coefficient of the lesion, as determined from CT scans: the base of the osteochondroma, composed



Figure 3-33 Tumoral calcinosis. Although soft tissue calcification may look very similar to that of periosteal chondroma, there is no bone erosion and no periosteal buttress formation.

mainly of trabecular bone, shows higher numbers for this variable than does the cartilaginous matrix of periosteal chondroma.

Pathology

The main problem is to distinguish benign **enchondroma** from *low-grade chondrosarcoma*. In the majority of cases, particularly if the histologic picture is typical and the lesion is not cellular, this distinction does not create a problem (see Figs. 3-2 and 3-3). However, if the lesion is cellular, and particularly if it affects the long bones, (Fig. 3-35), the pathologist must take care not to confuse an enchondroma with a low-grade chondrosarcoma and vice versa.

Cell atypias and uneven distribution of cells may be found to a higher degree in enchondromas of the hands and feet without prognostic significance. However, when an enchondroma is located in long, tubular bones, particularly in the humerus and femur, or in bones of the trunk, slight histologic atypias have prognostic significance. From observations of large series over long periods it is known that tumors with these characteristics may recur once or several times and may progress to a chondrosarcoma of intermediate or high



Figure 3-34 Periosteal chondroma resembling osteochondroma. A: Lateral radiograph shows a bony excrescence arising from the posterior cortex of the distal femur that looks like a sessile osteochondroma (continued).



Figure 3-34 *Continued* **B:** Conventional tomography shows calcifications at the base of the lesion and lack of interruption of the posterior cortex of the femur. **C:** Computed tomography section demonstrates lack of communication between the medullary portion of the femur and the lesion, thus excluding the diagnosis of osteochondroma. (**A** and **C**, reprinted with permission from Greenspan et al. Periosteal chondroma masquerading as osteochondroma. *Can Assoc Radiol J* 1993;44:205–210.) **D:** In another patient the lateral radiograph of the proximal leg demonstrates a lesion in the fibula that may represent either a sessile osteochondroma or a periosteal chondroma.

В



Figure 3-34 *Continued* **E:** Sagittal T1-weighted (SE, TR 600, TE 20) magnetic resonance imaging (MRI) reveals that the lesion is brighter than the fibula's bone marrow and the anterior fibular cortex does not interrupt medullary continuity, suggesting a diagnosis of osteochondroma. **F:** This diagnosis is further confirmed with axial T2-weighted (SE, TR 2000, TE 80) MRI that shows unequivocally not only the merging of the medullary portions of the lesion and the host bone, but a cartilaginous cap (*bright signal*), characteristic of osteochondroma (*arrow*).



Α

Figure 3-35 Low-grade chondrosarcoma. A: Chondrocytes with a subtle degree of pleomorphism and abnormal nuclei with open chromatin and visible nucleoli are irregularly distributed within a chondroid matrix (hematoxylin and eosin, original magnification $\times 100$). **B:** Destruction of the bone trabeculae surrounded by chondroblastic tumor tissue that infiltrates the marrow spaces, and a dense arrangement of chondrocytes correspond to grade 2 tumor (hematoxylin and eosin, original magnification $\times 50$).

grade, or even to a dedifferentiated chondrosarcoma. Furthermore, enchondroma lobules are often encased by newly formed lamellar bone.

The microscopic distinction between benign cartilage of enchondromatosis and *low-grade chondrosarcoma* is difficult (16). Necrosis and myxoid stroma strongly favor the diagnosis of chondrosarcoma (23).

Histologically, **periosteal chondroma** must be differentiated from *periosteal chondrosarcoma* (see Figs. 3-112 and 3-113) and *periosteal osteosarcoma* (see Figs. 2-109 and 2-110). The latter lesion, however, is usually grade 2 or 3, exhibits a diaphyseal location, and contains osteoid or, less commonly, mature bone.

The radiologic and pathologic differential diagnosis of enchondroma and periosteal chondroma is depicted in Figure 3-36.

Soft Tissue Chondroma

These rare lesions occur preferentially in the soft tissues of hands and feet. A soft tissue chondroma located close to a joint is known as a paraarticular chondroma (22,47). Because these lesions exhibit a wide range of cytologic variations, as in the case of cartilage tumors of bone, they are commonly misdiagnosed as chondrosarcomas (18). They have no predilection for either gender and most patients are 20 years or older, with a mean occurrence during the fourth and fifth decades (9). These lesions are usually small, ranging from 2 to 4 cm. On radiographic studies they present as well-defined soft tissue masses, often (33% to 77%) with calcifications similar to those seen in enchondroma (78). They are not attached to the periosteum or cortex of a bone and are not found within the confines of a joint capsule or tendon sheath (3,12,77). On histopathologic examination soft tissue chondromas consist of masses of hyaline cartilage, which are commonly lobulated and sometimes partially myxoid (18). In addition, some lesions are hypercellular and exhibit hyperchromatic nuclei. These features, which are also seen in enchondromas of small tubular bones, are not indicative of malignancy. Soft tissue chondromas may contain focal areas of fibrosis, hemorrhage, necrosis, calcification, ossification, or granuloma formation (3,77). Electron microscopy shows typical features of cartilage cells, with abundant rough endoplasmic reticulum, free ribosomes (55), and short irregular microvillous processes surrounded by aggregates of calcium crystals (9).

Recently, nonrandom clonal alterations of chromosomes 6, 11, and 12 have been reported in soft tissue chondromas, including supernumerary ring chromosomes containing material from chromosome 12 (14,43,71). Molecular analysis has shown that the socalled *HMGA2* gene located at 12q15 (*high mobility* group gene, coding for a small non-histone-chromatinassociated protein implicated in the architectural organization of chromatin, thus influencing transcription and playing a role in growth and development) appears to be involved in soft tissue chondroma and other cartilaginous tumors, and also in lipomas (11,40).

Differential Diagnosis

Radiology

The main differential possibility is periosteal chondroma. This lesion is, however, attached to the periosteum or cortex and usually produces a cortical erosion and buttress of periosteal reaction. Myositis ossificans can result in a mineralized mass; however, it exhibits a classic zoning phenomenon, whereas calcifications in the soft tissue chondroma are distributed in a haphazard pattern. Synovial chondromatosis is a condition affecting joints and tendon sheaths, and CT or MRI is usually able to identify intra-articular or within the tendon sheath location. Benign mesenchymoma of soft tissue may contain calcifications. These lesions are composed primarily of mature hyaline cartilage. The difference between soft tissue chondroma and mesenchymoma is that the latter may also contain adipose and vascular elements (46). Lesions reported in the literature as soft tissue osteochondromas may appear similar to soft tissue chondromas, although in addition to calcifications they also may contain bony elements (38).

Tumoral calcinosis usually presents on radiography as a well-defined lobulated calcific mass, exhibiting a layering effect when imaged with a horizontal beam (4,61) (see Fig. 3-33).

Soft tissue chondrosarcomas are rare lesions (77). Unlike soft tissue chondromas they almost never occur in the hands and feet, their preferred sites being in the proximal parts of extremities and buttocks. Synovial sarcoma exhibits a soft tissue mass and, in approximately 25% to 30% of reported cases, calcifications. It is usually accompanied by destruction of adjacent bones.

Pathology

In general, soft tissue chondromas are so characteristic that problems in differential diagnosis rarely arise, although hypercellular lesions with hyperchromatic nuclei may be confused with *chondrosarcomas*. However, extraskeletal chondrosarcomas are almost always of myxoid or mesenchymal type, are rarely located in the hands and feet, and contain a prevailing small-cell component. Because no extraosseous low-grade chondrosarcoma composed of pure hyaline cartilage has yet been reported, this diagnosis can be excluded (32). Nevertheless, soft tissue extensions of intraosseous grade 1 chondrosarcoma must be ruled out.

Calcifying aponeurotic fibroma, mainly occurring in the hands and feet of children, contains foci of cartilage. However, in contrast to well-demarcated and lobulated soft tissue chondromas, these lesions are ill-defined and tend to infiltrate the surrounding soft tissues. *Synovial chondromatosis* is, by definition, located within synovial tissue and consists of multiple cartilaginous nodules. Myxoid changes in soft tissue chondroma may lead to the erroneous diagnosis of *extraskeletal myxoid chondrosarcoma*, which, however, preferentially occurs in the lower extremities and not in the hands and feet. In addition, this tumor is larger and more cellular, containing round cells with scant eosinophilic cyto-



Figure 3-36 Radiologic and pathologic differential diagnosis of enchondroma (A) and periosteal chondroma (B).



Figure 3-37 Radiologic and pathologic differential diagnosis of soft tissue chondroma.

plasm arranged in cords and clusters. These features are absent in soft tissue chondroma (77).

The radiologic and pathologic differential diagnosis of soft tissue chondroma is shown in Figure 3-37.

Osteochondroma (Osteocartilaginous Exostosis)

Clinical Presentation

Osteochondroma, the most common benign bone lesion (representing about 45% of all benign bone tumors and 12% of all bone tumors) (13), is a cartilage-capped bony projection on the external surface of a bone. Usually diagnosed before the third decade, it most commonly involves the metaphyses of long bones, particularly around the knee and the proximal humerus. In general, the lower extremities are more often affected than the upper extremities. The flat bones, including the scapula, ilium, and clavicle, are much less commonly affected (Fig. 3-38). The lesion, which possesses its own "growth plate," usually stops growing at skeletal maturity. Only sporadic cases of spontaneous resolution have been reported (113). Osteochondroma is asymptomatic unless it causes pressure on adjacent muscles, nerves, or blood vessels (97), as well as on adjacent bone, occasionally with consequent fracture (116). Other complications have been reported, such as fracture of the lesion itself (84) and inflammatory changes of the bursa exostotica covering the cartilaginous cap (79,86,92) (see Figs. 3-54 and 3-55). Malignant transformation to chondrosarcoma is very rare, occurring in less than 1% of solitary lesions (see Fig. 3-52). Pain (in the absence of a fracture, bursitis, or pressure on nerves) and a growth spurt or continued growth of the lesion beyond skeletal maturity are highly suspicious for this complication (110) (Table 3-1).

Variants of osteochondroma include subungual exostosis, turret exostosis, traction exostosis, bizarre parosteal osteochondromatous proliferation (BPOP), florid reactive periostitis, and dysplasia epiphysealis hemimelica (also known as intraarticular osteochondroma) (88,107,115).

Imaging

Radiographic presentation of osteochondroma depends on the type of lesion (107). The pedunculated osteochondroma manifests with a slender pedicle, which is usually directed away from the neighboring growth plate or joint (Fig. 3-39A). The sessile osteochondroma exhibits a broad base attached to the cortex (Fig. 3-39B). In either type, the most important identifying features are that the cortex of the host bone merges without interruption with the cortex of the osteochondroma and that the cancellous portion of the lesion is continuous with the medullary cavity of the adjacent diaphysis. Calcifications in the chondroosseous portion of the stalk of the lesion are also typical features (Fig. 3-40). CT



Figure 3-38 Osteochondroma: skeletal sites of predilection, peak age range, and male-to-female ratio.

scanning can establish unequivocally the continuity of cancellous portions of the lesion and the host bone (Fig. 3-41). These characteristics distinguish this lesion from the occasionally similar-appearing bone masses of osteoma, juxtacortical osteosarcoma, soft tissue osteosarcoma, and juxtacortical myositis ossificans (see Fig. 2-6). CT or MRI examination demonstrates the cartilaginous cap (51) (Fig. 3-42). On MRI the cartilaginous cap shows a high signal intensity on T2-weighted and gradient echo sequences. A narrow band of low signal intensity surrounding the cap represents the perichondrium (91,102) (Fig. 3-43). Small areas of various degrees of signal void and low signal intensity represent cartilage calcifications. Gadolinium-enhanced MRI may be helpful in demonstrating peripheral enhancement in osteochondroma, corresponding to fibrovascular tissue covering the nonenhancing cartilage cap (91) (Fig. 3-44).

Ultrasound (sonography) may also be a valuable modality for measuring the thickness of the cartilaginous cap of osteochondroma (107). In fact, the investigations conducted by Malghem et al. (105) revealed that this technique was more accurate than CT and quite similar to MRI in evaluation of cap thickness. Disadvantages of ultrasound included operator dependence, inability to evaluate deep lesions, and inability to evaluate the osseous component of osteochondroma.

Scintigraphy shows variably increased uptake by the lesion (87). The intensity of activity is directly correlated with the degree of chondral ossification (101). However, the main use of scintigraphy is to search for multiple lesions, when only a single osteochondroma has been discovered by radiography.

Histopathology

Histologically, the cap of the osteochondroma is composed of hyaline cartilage arranged similarly to a growth plate. A zone of provisional calcification is present, corresponding to the areas of calcifications in the chondroosseous portion of the stalk (Fig. 3-45). Deeper parts of the stalk show fatty marrow that merges with normal hematopoietic marrow of the host bone. The thickness of the cartilaginous cap ranges from 1 to 3 mm and rarely up to 1 cm. Greater thickness may imply the possibility of transformation to chondrosarcoma (18). A delicate, fibrous membrane known as the perichondrium, representing a continuation of the periosteum of the adjacent cortex, overlies the cartilaginous cap (see Fig. 3-45D). With age the cap atrophies and in some instances disappears completely (51).

Multiple Hereditary Osteochondromata (Hereditary Osteochondromatosis)

Classified by some authorities among the bone dysplasias, multiple osteochondromata, also known as multiple osteocartilaginous exostoses, familial or hereditary osteochondromatosis, or diaphyseal aclasis, represent an autosomal dominant, hereditary disorder (96), with incomplete penetrance in females (103). Approximately 66% of affected individuals have a positive

Clinical Features	Radiologic Findings	Imaging Modality
Clinical Features Pain (in the absence of fracture, bursitis, or pressure on nearby nerves) Growth spurt (after skeletal maturity)	Radiologic FindingsEnlargement of the lesion Indistinct or irregular lesion's surfaceFocal areas of radiolucency within the lesionDevelopment of a bulky cartilaginous cap, usually 2-3 cm thickDispersed calcifications in the cartilaginous cap Development of a soft tissue mass with or without	Imaging ModalityRadiography (comparison with earlier radiographs)RadiographyCT, MRIRadiography Conventional tomography Radiography, CT, MRI
	calcifications Increased uptake of isotope after closure of growth plate (not always reliable)	Scintigraphy (planar and SPECT)

	Table 3-1	Clinical ar	nd Radiologic	Features	Suggesting	Malignant	Transformation	of (Osteochond	roma
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CT, Computed tomography; MRI, magnetic resonance imaging; SPECT, single-photon emission computed tomography.



Figure 3-39 Osteochondroma. A: Pedunculated variant. The typical pedunculated type of osteochondroma is seen arising near the proximal growth plate of the right humerus in a 13-year-old boy. **B:** Sessile variant. Broad-based osteochondroma is seen here arising from the medial cortex of the proximal diaphysis of the right humerus in a 14-year-old boy. Note that in both lesions the cortex of the host bone merges, without interruption, with the cortex of the lesions and the medullary portion of both lesions and the medullary cavities of the adjacent bones communicate.





Figure 3-40 Osteochondroma. The lesion exhibits calcifications in the chondroosseous zone of the stalk.

family history (81). The specific genetic abnormalities have recently been detected, with three distinct loci on chromosomes 8, 11, and 19 (81,104,117). There is a decided 2:1 male predilection (Fig. 3-46). The lesions are usually discovered at about 2 years of age. The knees,

hips, ankles, and shoulders are the most commonly affected sites and growth disturbances are often present, primarily in the forearms and legs (111). There is evidence of defective metaphyseal remodeling, with deformation of affected bones and asymmetric retardation of longitudinal bone growth (16). The radiologic features are similar to those of a solitary osteochondroma; the sessile form of the lesion is more common (Figs. 3-47 to 3-51).

The Langer-Giedion syndrome, also known as trichorhinophalangeal syndrome type II, is characterized in addition to multiple osteochondromas by craniofacial dysmorphism and mental retardation. It is due to deletion of 8q24. Similar to the former disorder is Potocki-Shaffer syndrome, characterized by multiple osteochondromata, enlarged parietal foramina, and sometimes craniofacial dysostosis and mental retardation (80).

The pathologic features of multiple osteochondromata are the same as those of solitary lesions (99). Malignant transformation to chondrosarcoma [and in rare instances to other types of sarcoma (80)] in osteochondromatosis is more common than in solitary lesions, not only because of the greater number of lesions present but because of the higher risk for malignant transformation of each lesion. However, estimated figures from older literature (ranging from 5% to 25%) seem to be too high, mainly because they have been obtained from large tumor referral centers. Recent investigations point to a much lower percentage (1%-3%) of sarcomatous transformation of these lesions (111,114). Lesions at the shoulder girdle and around the pelvis are usually at greater risk for this complication. Malignant transformation before the age of 20 is uncommon.

As previously stated, hereditary osteochondromatosis is a heterogeneous autosomal dominant disorder in *Text continues on page 193*



Figure 3-41 Osteochondroma: computed tomography (CT). A: Lateral radiograph of the knee shows a calcified lesion at the posterior aspect of the proximal tibia (*arrows*). The exact nature of this lesion can not be ascertained. **B:** CT clearly establishes the continuity of the cortex, which extends without interruption from the osteochondroma into the tibia. Note also that the medullary portion of the lesion and the tibia communicate.





Figure 3-42 Osteochondroma: computed tomography (CT). A: Radiograph shows a large osteochondroma arising at the intertrochanteric area of the right femur. B: CT section shows the heavily calcified, cauliflower-like mass and a thin, cartilaginous cap.









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Figure 3-45 Histopathology of osteochondroma. A: Gross specimen of sessile osteochondroma exhibits characteristic features of the lesion: continuity of the cortex and medullary cavity and a thin, cartilaginous cap. (Reprinted with permission from Bullough PG. *Atlas of orthopedic pathology*, 2nd ed. New York: Gower, 1992:14.10.) **B:** The tumor consists of the hyaline cartilage cap with strong metachromasia of the matrix. A broad zone of endochondral ossification borders the cancellous bone containing the remnants of cartilaginous matrix (*center*) (Giemsa, original magnification ×6). **C:** At higher magnification areas of calcification are seen in the chondroosseous portion of the lesion. These areas correspond to a zone of provisional calcification in the growth plate (hematoxylin and eosin, original magnification ×30). **D:** Hyaline cartilage cap is seen at the periphery of the lesion, and beneath it transformation of cartilage into bone by endochondral ossification. A thin fibrous membrane (perichondrium) loosely overlies the cartilage (hematoxylin and eosin, original magnification ×60).

D







Figure 3-47 Multiple osteocartilaginous exostoses. A: Radiograph of the distal forearm of an 8-year-old boy with multiple osteochondromas shows growth disturbance in the distal radius and ulna. **B:** In another patient, a 21-yearold woman, observe growth disturbance of the proximal fibula.

Figure 3-46 Multiple osteocartilaginous exostoses: skeletal sites of predilection, peak age range, and male-to-female ratio.



Figure 3-48 Multiple osteocartilaginous exostoses. A: Anteroposterior radiograph of both knees of a 17-year-old boy shows numerous sessile and pedunculated lesions. B: Anteroposterior radiograph of the hips shows numerous sessile osteochondromas affecting proximal femora.

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Figure 3-49 Multiple osteocartilaginous exostoses. A: Anteroposterior radiograph of the shoulder of a 22-year-old man with familial multiple osteochondromas demonstrates multiple sessile lesions involving the proximal humerus and scapula. **B:** Involvement of the distal femur and proximal tibia in the same patient is characteristic of this disorder.

which three gene loci have been identified on chromosomes 8q24.1 (EXT1), 11p11-p12 (EXT2), and 19p (EXT3). So far, EXT1 and EXT2 have been cloned. These loci encode glycoproteins that are involved in biosynthesis of heparan sulfate proteoglycans. Several mutations have been detected. On the basis of analogue findings in the fruit fly Drosophila melanogaster, it appears that alteration of heparan sulfate biosynthesis may interfere with the normal maturation of chondrocytes, leading to premature chondrocyte differentiation and subsequent ossification (80,85). Because genotype-phenotype studies have shown that patients with an EXT1 mutation, as compared to patients with an EXT2 alteration, are at increased risk for development of severe disease and sarcomas, screening for EXT1 mutations in these patients appears to be advisable (89,112).

Differential Diagnosis

Radiology

The most important differential diagnosis is the distinction between benign osteochondroma and *exostotic chondrosarcoma* arising in previous exostosis. It is important to recognize early the features that suggest malignant transformation of osteochondroma to chondrosarcoma. Radiologic evaluation may reveal a constellation of features suggesting this complication: development of a thick, bulky cartilaginous cap (usually more than 2 to 3 cm in thickness) (95,105); development of a soft tissue mass with or without calcifications; and dispersed calcifications within the cartilaginous cap, separate from those contained in the stalk (Fig. 3-52), a sign described by Norman and Sissons (110). The most reliable imaging modalities for evaluating possible malignant transformation are radiography, CT, and MRI (83,150). Radiographs usually demonstrate containment of the calcifications of osteochondroma within the stalk of the lesion, but occasionally conventional tomography can also be helpful (Fig. 3-53). Dispersement of the calcifications in the cartilaginous cap and increased thickness of the cap, the cardinal signs of malignant degeneration, can be demonstrated by any of the previously mentioned modalities. Radionuclide bone scan can also be performed and may reveal increased uptake of radiopharmaceutical agent at the site of a lesion. Exostotic chondrosarcoma often exhibits greater intensity of uptake than a benign exostosis. However, a number of investigators believe that this is not always a reliable distinguishing feature of malignant transformation (94). The increased activity of a benign exostosis on bone scan is related to endochondral ossification within the cartilaginous cap, whereas that of exostotic chondrosarcoma represents active ossification, osteoblastic activity, and hyperemia within the cartilage and bony stalk of the

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Figure 3-50 Multiple osteocartilaginous exostoses: three-dimensional computed tomography (3-D CT). A: Conventional sagittal reformatted CT section shows osteochondromas arising from the posterior aspects of distal femur and proximal tibia. **B, C:** Three-dimensional CT images viewed from lateral and posterior aspects of the knee show spatial distribution of numerous osteochondromas. **D:** 3-D CT image of the distal femur in maximum intensity projection (MIP) shows interior architecture of one of the sessile osteochondromas.

R



Figure 3-51 Multiple osteocartilaginous exostoses: magnetic resonance imaging (MRI). Coronal **(A)** and axial **(B)** T1-weighted (SE, TR 600, TE 20) MR images show several sessile osteochondromas affecting the proximal femora. Observe abnormal tubulation of the bones. **C:** In another patient with multiple cartilaginous exostoses MRI was performed because one of the osteochondromas continued to enlarge. Sagittal T1-weighted (SE, TR 400, TE 12) image demonstrates intact cortex covering the lesion. **D:** Axial fast-spin echo (FSE, TR 4000, TE 102 Ef) MRI shows high-intensity thin cartilaginous cap of osteochondroma without malignant changes. **E:** Lack of malignant transformation was confirmed on axial inversion recovery (FMPIR/90; TR 4000, TE 51 Ef, TI 140) MRI, which shows no soft tissue extension of the lesion (*arrows*).

tumor (94). Occasionally the clinical and radiologic features of inflammatory changes occurring in bursa exostotica that covers the cartilaginous cap of osteochondroma can mimic malignant transformation to chondrosarcoma (Figs. 3-54 and 3-55).

On radiographic examination, the signs pointing to malignant transformation of multiple cartilaginous exostoses to chondrosarcoma are identical with those present in malignant transformation of solitary osteochondroma (90). After skeletal maturation has occurred, alteration in a lesion's size must be considered a potential indicator of malignant transformation. Particularly suggestive are alterations in the size and irregularity of contour of the cartilage cap, or the presence of mineral deposition beyond the previous contour as documented by radiography (18). When such signs are associated with signs of bone destruction at the base or neck, or when a soft tissue mass is present, the diagnosis of malignancy is certain (Fig. 3-56).

An interesting lesion that can be confused with osteochondroma is the so-called *epiphyseal* or *intraarticular osteochondroma*, better known as *Trevor-Fairbank disease* or *dysplasia epiphysealis hemimelica* (89,98). This is a developmental disorder characterized by asymmetric cartilaginous overgrowth of one or more epiphyses in the lower and occasionally the upper extremities. The talus, distal femur, and distal tibia are the most common sites of involvement (98). The lesion typically arises on one side of the affected limb and deforms the bone. Males are affected approximately three times as often as females. The basic pathologic process is abnormal cartilage proliferation in the epiphysis, and on histologic examination the lesion is almost identical with osteochondroma. Dysplasia epiphysealis hemimelica has been noted in association with conventional osteochondroma, chondroma (93), and Ollier disease (58). The clinical features are pain, deformity, and restricted motion in the affected joints. Radiography reveals an irregular, bulbous overgrowth of the epiphysis to one side of the ossification center (Fig. 3-57). Although CT may demonstrate the continuity of the mass with the underlying epiphysis, MRI is more effective in assessment of this abnormality. As the lesion enlarges the joint deformity increases (100).

Finally, sessile osteochondroma should be differentiated from *periosteal chondroma* (26). The latter is sepa-

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Figure 3-52 Exostotic chondrosarcoma. A 28-year-old man developed pain in the popliteal region and also noted an increase in a mass of which he had been aware for 15 years. **A:** Lateral radiograph of the knee shows a sessile-type osteochondroma arising from the posterior cortex of the distal femur. Calcifications are present not only in the stalk of the lesion but are also dispersed in the cartilaginous cap. **B:** Computed tomography section confirms the increased thickness of the cartilaginous cap (2.5 cm) and dispersed calcifications within the cap (*arrow*). These features are consistent with a diagnosis of malignant transformation to chondrosarcoma, which was confirmed by histopathologic examination.



Figure 3-53 Osteochondroma resembling exostotic chondrosarcoma. A: Lateral radiograph of the ankle of a 26-year-old woman with a painful osteochondroma shows a sessile lesion arising from the posterior aspect of the distal tibia. In the interpretation of this radiograph, uncertainties were raised that some of the calcifications might not be contained within the stalk, and tomographic examination was suggested. B: Tomographic section demonstrates lack of separation of calcifications from the main mass, suggesting a benign lesion. The osteochondroma was resected and histopathologic examination confirmed the lack of malignant transformation.

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Figure 3-54 Bursa exostotica. A 25-year-old man with a known solitary osteochondroma of the distal right femur presented with gradually increasing pain. Malignancy was suspected and arteriography was performed. The capillary phase of the arteriogram reveals a huge bursa exostotica. Inflammation of the bursa, with accumulation of a large amount of fluid (bursitis), was the cause of the patient's symptoms.





Figure 3-55 Bursa exostotica. A 12-year-old girl presented with pain in the popliteal fossa. **A:** Coronal T1-weighted (SE, TR 650, TE 25) magnetic resonance imaging (MRI) demonstrates a large osteochondroma arising from the posterolateral aspect of the distal femur (*arrow*). **B:** Axial T2-weighted (SE, TR 2200, TE 70) MRI shows a bursa exostotica distended with high-intensity fluid (*arrows*).



rated from the host bone by intervening cortex (see also section on differential diagnosis of periosteal chondroma). Differentiation of sessile osteochondroma from occasionally similarly appearing parosteal osteosarcoma, soft tissue osteosarcoma, and myositis ossificans should not create a significant problem (Fig. 3-58).

Pathology

Malignant transformation of osteochondroma to chondrosarcoma is the most important histopathologic differential diagnosis. On histologic examination, malignant transformation is recognized by greater cellularity of

the cartilage tissue, uneven distribution of cells without cord- or column-like arrangement, pleomorphism, and cellular and nuclear atypia (Fig. 3-59). Additional criteria include destruction of bone trabeculae of the stalk with invasion of marrow spaces, and invasion of the overlying soft tissues.

B

The other differential possibility of osteochondroma is so-called reactive cartilaginous exostosis, which differs from the former by its smaller size and the metaplastic formation of cartilage from the fibrous tissue covering a bone (108) (Fig. 3-60). The resulting fibrocartilage undergoes endochondral ossification, as



Figure 3-57 Dysplasia epiphysealis hemimelica. A 12-year-old girl presented with pain and limitation of motion in the ankle joint. Anteroposterior (A) and lateral (B) radiographs of the ankle demonstrate deformity and enlargement of the medial malleolus, talus, and navicular bone, features typical of Trevor-Fairbank disease.

in osteochondroma. Continuing formation is the result of ongoing metaplasia from the surrounding fibrous tissue on its surface, whereas in osteochondroma growth is generated by mitotic division of the chondrocytes.

Bizarre parosteal osteochondromatous proliferation (BPOB or *Nora lesion)* can be excluded by its feature of not being in continuity with the marrow cavity of underlying bone. Furthermore, Nora lesion is characterized by a spindle-cell stroma, irregular ossifications, and cap-like irregularly structured cartilage; however, cellular atypia is absent (82,106,109) (see Fig. 2-131C).

An osteochondroma in which intracartilaginous ossification has completely consumed the cartilage may be mistaken for an *osteoma* if the base of the stem is not available for histologic examination.

Chondroblastoma (Codman Tumor)

Clinical Presentation

Representing less than 1% of all primary bone tumors and 9% of benign bone tumors, this rare lesion typically presents in the epiphysis of long bones, such as the humerus, tibia, and femur (121,177,178). So far, only one case of diaphyseal chondroblastoma in a long bone has been reported (118a). The proximal humerus, dis-

tal femur, and proximal tibia are the preferred sites of involvement (153) (Fig. 3-61). Occasionally the patella, which is considered equivalent to an epiphysis, is involved (164). Ten percent of chondroblastomas involve the small bones of the hands and feet, the talus and calcaneus representing the most common sites (125,130, 154,158,166). Only 1% of all chondroblastomas have been reported to involve the skull (152). Involvement of the vertebrae is exceedingly rare (144, 157). The lesion is usually seen before skeletal maturity, but some cases have been reported after obliteration of the growth plate (129). Males are almost twice as frequently affected as females (52), although in the series from the Armed Forces Institute of Pathology (AFIP) the ratio was 3:1 (164). In general, the clinical symptoms are nonspecific: pain, often related to the nearest joint, and swelling, usually lasting for several months, are the most common complaints (123,132). Approximately one third of patients exhibit joint effusion (174). Pathologic fractures occur only rarely.

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Imaging

Radiographically, the lesion is usually located eccentrically (168). It is radiolucent and well defined, usually with a thin sclerotic border, and it exhibits a geographic pattern of bone destruction (141) (Fig. 3-62).



Figure 3-58 Differential diagnosis of osteochondroma. Radiographic features characterizing lesions similar in appearance to osteochondroma. (Reprinted from Greenspan A. *Orthopedic imaging – a practical approach,* 4th ed. Philadelphia: Lippincott Williams & Wilkins, 2004, Fig. 18.28).

R



Figure 3-59 Histopathology of exostotic chondrosarcoma. A: A photomicrograph (same patient as in Fig. 3-52) shows cellular chondroid tissue exhibiting slight increase in size and variation in shape of the nuclei, consistent with grade 1 chondrosarcoma (hematoxylin and eosin, original magnification $\times 100$). **B:** At higher magnification the malignant cells are clearly depicted (hematoxylin and eosin, original magnification $\times 300$).

The margins are smooth in 63% of cases, scalloped in 27%, and lobulated in 10% (161). Internal trabeculation can sometimes be observed (156). In 25% to 50% of cases, stippled calcifications are present (Fig. 3-63). If calcifications are not apparent on standard radi

Δ



Figure 3-60 Reactive cartilaginous exostosis. The lesion consists of a cap of fibrocartilage exhibiting weak metachromatism of the matrix. It is formed and grows by metaplasia from the overlying fibrous connective tissue, with a zone of endochondral ossification against the base of cancellous bone that contains some remnants of cartilage matrix (*center*) (compare this lesion with osteochondroma, Fig. 3-45B) (Giemsa, original magnification \times 6).

ographs, conventional tomography or CT can be helpful (138) (Fig. 3-64; see also Fig. 3-62C). The latter modality may also assess the extent of the lesion (170). The presence of a solid or layered periosteal reaction (123), usually located in some distance from the tumor, was reported on conventional radiographs in more than 50% of 214 cases in the series by Brower et al. (126) (Fig. 3-64A; see also Fig. 3-65A). These authors suggest an inflammatory response to the tumor as the cause of this phenomenon. MRI usually reveals a larger area of involvement than can be seen on radiography, including prominent soft tissue and regional bone marrow edema (128,139,181) (Fig. 3-65). This strikingly aggressive appearance can lead to an incorrect diagnosis of a malignant tumor (155,180). The combination of highly cellular chondroid matrix and calcification in chondroblastoma presumably accounts for the observed decrease in relaxation time on T2-weighted imaging (128). Low to intermediate heterogeneous signal intensity, lobular internal architecture, and fine lobular margins were detected with high-resolution T2-weighted MRI (180). Focal lobules of low, intermediate, and high signal intensity are superimposed and most likely correspond to calcification, chondroid matrix, and fluid in the lesion. In addition, MRI may demonstrate a rare extension of tumor to the soft tissues and may also reveal a joint effusion (Fig. 3-66).

Histopathology

Before the recognition of chondroblastoma by Jaffe and Lichtenstein in 1942, this tumor was usually referred to as epiphyseal chondromatous giant cell tumor (127). The tumor does contain giant cells and in fact is still occasionally misdiagnosed as giant cell tumor. However, Jaffe and Lichtenstein (146) felt that these giant cells were not part of the primary tumor pattern and instead pointed out its relationship to cartilage. However, its relationship to cartilage-forming tumors has recently been challenged. In an immunohistochemistry (IHC) analysis of the matrix composition in



Figure 3-61 Chondroblastoma: skeletal sites of predilection, peak age range, and male-to-female ratio.

56 chondroblastomas, Aigner et al. (118) were unable, in contrast to Edel et al. (131), to demonstrate type II collagen, the major matrix component of hyalin cartilage, whereas type I collagen, normally present in bone but not in cartilage, and type III collagen were found, the latter of which is also present in cartilage in small amounts. Furthermore, these authors detected aggrecan proteoglycan core protein, which is a major component of hyalin cartilage and is also present in other tissues. On the basis of these findings, the authors suggested that chondroblastoma should be considered a bone-forming tumor (118). However, in a study of 12 chondroblastomas, Romeo et al. (173) demonstrated that signaling molecules of chondrocyte differentiation [Indian Hedgehog-IHh/PTHrP/fibroblast growth factor (FGF), and fibroblast growth factor receptor (FGFR/bcl-2 and p21)] are expressed in chondroblastomas at the protein level. These authors concluded that chondroblastoma is a neoplasm committed toward chondrogenesis.

Most chondroblastomas are DNA diploid, as revealed by flow cytometry. However, DNA near-diploid and DNA aneuploid forms exist (151). Clonal abnormalities have been reported, including recurrent structural alterations in chromosomes 5 and 8 with rearrangements of band 8q21 in two "aggressive" chondroblastomas (124,179). Recently, Sjögren et al. (344) found recurrent breakpoints at 2q35, 3q21–q23, and 18q21 in an analysis of six otherwise diploid or near-diploid chondroblastomas.

Histologically, chondroblastoma is composed of nodules of fairly mature cartilage matrix surrounded by a highly cellular, relatively undifferentiated tissue comprising round and polygonal chondroblast-like cells, sometimes having indistinct outlines (176). At the borders of the differentiated areas, giant cells of the osteoclast or chondroclast type are usually found (174). The nuclei of the cells are of uniform shape and moderate size (125) (Fig. 3-67). They may be round but usually are indented. Mitotic figures are occasionally observed (162). Foci of intercellular calcifications are typically present. Particularly characteristic are fine, lattice-like matrix calcifications surrounding apposing chondroblasts and having a spatial arrangement that resembles the hexagonal configuration of chicken wire (125,138), a diagnostically useful, feature but not mandatory for diagnosis (Fig. 3-67C, E). Although chondroid matrix is relatively scant in chondroblastoma, the cells exhibit immunocytologic staining reaction for S-100 protein identical to that of other cartilage tumors, as well as for vimentin and neuron-specific enolase (NSE). Occasionally, chondroblastoma may exhibit focal expression of osteonectin and cytokeratins (131). Larger tumors may break into the metaphysis or through the cortex without signs of malignancy.

Pulmonary metastases have been reported in a minority of cases of chondroblastoma (49,129,137, 165a), but no histologic evidence of malignancy has been reported in either the primary bone tumor or the pulmonary metastases (142,143). However, single cases of spontaneous malignant transformation of chondroblas-



Figure 3-62 Chondroblastoma. Anteroposterior **(A)** and lateral **(B)** radiographs of the right knee of a 14-year-old boy show the typical appearance of this tumor in the proximal epiphysis of the tibia (*arrows*). Note the radiolucent, eccentrically located lesion with a thin, sclerotic margin. **C:** Frontal tomogram in another patient with chondroblastoma in the tibial epiphysis (*arrows*) reveals faint calcifications.



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Figure 3-63 Chondroblastoma. Anteroposterior radiograph of the left knee of a 17-year-old boy shows a large lesion crossing the scarred growth plate of the proximal tibia (*arrows*). The lesion is sharply demarcated and exhibits calcifications of the matrix.







Figure 3-64 Chondroblastoma. A: Anteroposterior radiograph of the right shoulder of a 16-year-old boy shows a lesion in the proximal humeral epiphysis (*open arrow*), but calcifications are not well demonstrated. Note the well-organized layer of periosteal reaction at the lateral cortex (*arrow*). **B:** Computed tomography section shows the calcifications clearly.







Figure 3-65 Chondroblastoma: magnetic resonance imaging (MRI). A: Conventional radiograph shows a lesion (*arrowheads*) with a narrow zone of transition in the left humeral head of an 18-year-old man. Note benign-appearing layer of periosteal reaction along the lateral cortex (*arrow*). B: Coronal T1-weighted (SE, TR 600, TE 20) MRI shows significant amount of perilesional edema. C: Axial T2-weighted (SE, TR 2000, TE 80) MRI shows the lesion exhibiting a heterogeneous signal.

Α


Figure 3-66 Chondroblastoma: magnetic resonance imaging (MRI). Sagittal proton-density (SE, TR 2000, TE 28) (A) and axial T2-weighted (SE, TR 2000, TE 80) (B) MRI of the lesion depicted in Figure 3-63 shows extension of the tumor posteriorly into the soft tissues (*arrow*).

toma have been reported (169,342) and, in one case, with DNA-aneuploidy and immunohistochemical proof of TP53 overexpression in the recurrent, and then becoming malignant tumor (165).

Differential Diagnosis

Radiology

If the lesion is in the classic epiphyseal location, only a few possibilities may be included in the radiologic differential diagnosis. The radiographic findings in chondroblastoma may be nonspecific when calcifications are not visible or when atypical features, such as extension into the metaphysis after closure of the growth plate, or extension into the soft tissues are encountered (135). *Clear cell chondrosarcoma* may be indistinguishable from chondroblastoma because it may exhibit the same benign pattern and, moreover, like chondroblastoma the former tumor may be seen in the immature skeleton (120,147).

Bone abscess is rare in an epiphyseal location (119,134) (see Fig. 2-46). It may demonstrate extension into the growth plate through a serpentine, radiolucent tract. The surrounding zone of sclerosis is usually thicker than that observed in chondroblastoma. In addition, no calcifications are present in this lesion and periosteal reaction is noted only in exceptional cases.

Osteonecrosis of either the humeral or the femoral head, although occasionally resembling chondroblastoma, usually exhibits classic radiographic findings, such as a crescent sign and a rather homogeneous density adjacent to it (136). MRI findings are also typical, although they vary depending on the stage of osteonecrosis.

Intraosseous ganglion is a rare lesion near the articular surface, invariably eccentric in location, with knee, ankle, and shoulder joints being preferred sites (Figs. 3-68 and 3-69). It is almost never seen before skeletal maturation. *Giant cell tumor*, with very few exceptions, is a lesion of the mature skeleton, and if seen before skeletal maturity it is metaphyseal in location (122,167,175). In addition, it shows no calcifications (148), and sclerotic margin is only rarely encountered.

Osteoblastoma of the epiphysis is extraordinarily rare (159,171), its usual location being the metaphysis or diaphysis. Enchondroma is rare in the epiphysis but at times may appear similar to chondroblastoma. Langerhans cell histiocytosis, like osteoblastoma and enchondroma, is extremely rare in the epiphysis and, unlike chondroblastoma, never presents with central calcifications.

Recently reported intracortical chondroblastoma of the femoral condyle radiologically mimicked *intraarticular osteoid osteoma* (145).

Occasionally, a *normal radiolucency in the proximal humerus* (representing an increased amount of cancellous bone in the region of greater tuberosity seen on end) may be mistakenly diagnosed as chondroblastoma (140,149,172) (Fig. 3-70). Knowledge of the existence of this normal variant may prevent the radiologist from making this erroneous diagnosis.

Pathology

Histologically, chondroblastoma must be differentiated from *chondrosarcoma* and *giant cell tumor*. Because large nuclei of variable size and shape are occasionally present in chondroblastoma, this and the presence of mitotic figures may suggest malignancy. Overall lowpower histopathologic appearance and correlation with radiography should help in arriving at the correct diagnosis (125).

Because osteoclast-like giant cells usually form a part of the histologic pattern of chondroblastoma, being present particularly at the border of the chondroid areas of this tumor, it may be mistaken for *giant cell tumor*. The giant cells of chondroblastoma, however, can be distinguished from the same cells in giant cell tumor by the

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Ε

Figure 3-67 Histopathology of chondroblastoma. A: Primitive chondroblastic tissue is seen in islands surrounded by densely arranged roundish cells. Note also many giant cells of osteo- and chondroclastic type (Giemsa, original magnification \times 25). **B:** At higher magnification, there is pleomorphism of chondrocytes within the cartilaginous matrix, with signs of resorption by giant cells (Giemsa, original magnification \times 50). **C:** The chondrocytes, some with ovoid and some with elongated nuclei, are embedded in a chondroid matrix. Surrounding the cells are fine, intercellular calcifications forming a chicken-wire image (hematoxylin and eosin, original magnification $\times 235$). **D**: At high magnification the chondroblasts, some well-demarcated, and some with ill-defined borders, display nuclei of various sizes, mostly roundish in shape (hematoxylin and eosin, original magnification ×400). E: Chicken-wire-like calcifications are present around the tumor cells (von Kossa, original magnification ×400). F: Tumor cells and adjacent hyaline cartilage are positive (nuclear and cytoplasmic) for S-100 protein (biotin-streptavidin peroxidase, anti S-100, original magnification ×400).



Figure 3-68 Intraosseous ganglion. Anteroposterior radiograph of the right knee shows a radiolucent lesion with a sclerotic border eccentrically located in the articular end of the proximal tibia, resembling a chondroblastoma.

rounded appearance of the intervening cells, in contrast to the spindle-shaped stromal cells of giant cell tumor. The presence of the chondroid areas indicates the true nature of the tumor (Fig. 3-71). When further doubt exists, the matrix stain aldehyde-fuchsin at pH 1.0 is useful, especially when combined with immunostaining for S-100 protein (163). The chondroid matrix of chondroblastoma is stained by aldehyde-fuchsin purple, as are the "chicken wire" calcifications often present around the tumor cells. Staining for S-100 protein is present in the cytoplasm of chondroblastoma but is absent in giant cell tumor mononuclear cells (125). Furthermore, immunohistochemical methods may be useful in the distinction of chondroblastoma from other noncartilaginous lesions that contain giant cells, such as giant cell reparative granuloma (133). Similarly to giant cell tumor, the latter lesion's mononuclear cells are usually strongly positive for histiocytic markers such as CD68 and alpha-1-chymotrypsin (133,160).

The radiologic and pathologic differential diagnosis of chondroblastoma is depicted in Figure 3-72.

Chondromyxoid Fibroma

Clinical Presentation

This rare cartilage tumor, consisting of a mixture of fibromyxoid tissue and cartilage tissue in variable proportions and accounting for 0.5% of all primary bone tumors and 2% of benign bone tumors, occurs predominantly in adolescents and young adults (most patients are younger than 30 years) and more commonly in males than in females (2:1) (13). This tumor has a predilection for the bones of the lower extremities, usually the proximal tibia (Fig. 3-73). Other locations, including vertebrae, are rare (184,189,193,195). The juxtacortical and intracortical sites for chondromyxoid





Figure 3-69 Intraosseous ganglion. A: Anteroposterior radiograph of the right knee of a 24-year-old man with an 8-week history of pain shows an oval, radiolucent eccentric lesion in the proximal tibia, resembling a chondroblastoma. B: Computed tomography section shows a low-attenuation area with ramifications, surrounded by a zone of reactive sclerosis.

fibroma are very unusual (188,194,201). The clinical symptoms include the presence of a peripherally located mass, together with local pain and swelling (182,186, 206).

Imaging

R

On radiographic imaging, the lesion may range from 1 to 10 cm in size, with an average of 3 to 4 cm (13,186). It is a round or ovoid, lucent, and eccentrically located lesion exhibiting a geographic pattern of bone destruction and a scalloped, sclerotic margin (196,207) (Fig. 3-74). Occasionally, ridges or septa are present within the lesion (52) (Fig. 3-75). It often erodes or balloons out from the cortex (186), and often a buttress of periosteal new bone can be ob-



Figure 3-70 Pseudotumor of the greater tuberosity of the humerus. A radiolucent area in the region of the greater tuberosity (*arrows*) is a normal variant that resembles a chondroblastoma.

served (138) (Fig. 3-76). Calcifications are not apparent on radiography but may be demonstrated on CT scans (206,208). MRI reveals characteristics of most cartilaginous tumors: intermediate to low signal intensity on T1-weighted and high signal intensity on T2weighted sequences (Fig. 3-77). This technique can also effectively demonstrate soft tissue extension of chondromyxoid fibroma (193,198).

Histopathology

Chondromyxoid fibroma is characterized by large lobulated areas of spindle-shaped or stellate cells distributed within abundant myxoid or chondroid intercellular material. They are separated by septum-like zones of more cellular tissue that may contain variable numbers of giant cells (209) (Fig. 3-78). A characteristic finding is the increased cellularity of the tissue near the septa (13) (Fig. 3-79). As the term "chondromyxoid fibroma" implies, the variety of tissues are characteristic of this lesion. Two types of matrix can be identified: dense chondroid matrix, which stains basophilic, and loose myxoid matrix, which stains eosinophilic, the latter be-



ing predominant in most chondromyxoid fibromas. Genuine hyaline cartilage is not seen in these tumors. Mild nuclear atypia may be observed. Not infrequently, large pleomorphic and hyperchromatic cells are observed and can lead to confusion in diagnosis with chondrosarcoma (203). Before the description of this entity by Jaffe and Lichtenstein in 1948 (190), many examples had probably been regarded as chondrosarcomas. Some cases reported as myxomas or myxofibromas of long bones are probably chondromyxoid fibromas (186). Histochemical studies demonstrate glycogen granules in many chondroblastic elements and in the stellate cells of the myxoid areas. The cytoplasm of giant cells of the osteoclast type exhibits a discrete amount of periodic acid-Schiff-positive material that is resistant to ptyalin digestion and is strongly positive for acid phosphatase, as has been observed in other giant cell lesions (202). The cells are reported to be positive for S-100 protein (183). In addition to the presence of type II collagen, type I, type III, and type VI were also found in the matrix of chondromyxoid fibroma on the protein level and in the matrix-associated cells on the mRNA level, whereas aggrecan was present throughout the tumor (204). As shown by Romeo et al. (200), signaling molecules regulating spatial expression of proteins involved in normal cartilage differentiation and proliferation are also expressed in chondromyxoid fibromas-however, in a different pattern. Recent reports indicate that chondromyxoid fibroma expresses smooth muscle antigen (CMA) and CD34 in regions peripheral to the lesion lobules but not elsewhere in this tumor (197). Ultrastructural studies of the stellate cells in this lesion reveal irregular cell processes, scalloping of the cell membrane, and frequently the presence of abundant cytoplasmic fibrils (205).

Differential Diagnosis

Radiology

The classic radiographic findings, including round or oval shape, eccentric metaphyseal location, lobulation, septation, and peripheral sclerosis, are usually diagnostic of chondromyxoid fibroma. Occasionally, however, the lesion may be radiographically indistinguishable from an

Figure 3-71 Histopathology of chondroblastoma. A highly cellular area of the tumor contains several giant cells (*upper right*). However, the juxtaposition of an area of chondroid matrix (*lower left*) indicates the true nature of this tumor (hematoxylin and eosin, original magnification $\times 100$). (Reprinted with permission from Bullough PG. Atlas of orthopedic pathology, 2nd ed. New York: Gower, 1992:16.24.)



Figure 3-72 Radiologic and pathologic differential diagnosis of chondroblastoma.

aneurysmal bone cyst (see Fig. 3-76). Other differential possibilities include nonossifying fibroma and fibrous dysplasia.

Nonossifying fibroma, unlike chondromyxoid fibroma, rarely displays cortical ballooning or cortical destruction, and a periosteal reaction is present only in lesions that have sustained a pathologic fracture. *Fibrous dysplasia*, in contrast to chondromyxoid fibroma, is centrally located rather than eccentric, rarely shows internal septations, and does not elicit a periosteal reaction.

In the tibia, *adamantinoma* and *osteofibrous dysplasia* (*Kempson-Campanacci lesion*) are the main differential possibilities, and when the lesion extends to the articular end of bone, *giant cell tumor* should be considered.

Adamantinoma is usually a much more destructive lesion, composed of multiple, sharply defined, radiolucent foci interspersed with sclerotic changes. It frequently involves the anterior cortex. Almost all cases of adamantinoma of long bones have been reported in the tibia and fibula. Periosteal reaction is usually much more aggressive than that observed in chondromyxoid fibroma. In addition, the peak age incidence of adamantinoma is between 30 and 50 years, in contrast to chondromyxoid fibroma, which occurs mostly in children and adolescents.

Osteofibrous dysplasia is usually limited in extent to the anterior cortex of the tibia (rarely in the fibula), is much

more sclerotic than chondromyxoid fibroma, and does not elicit a periosteal reaction. The tibia is usually anteriorly bowed. The peak incidence of osteofibrous dysplasia is in the first decade, whereas that of chondromyxoid fibroma is in the second decade.

In the mature skeleton, particularly if the lesion extends to the articular end of the bone, *giant cell tumor* should be a consideration in the differential diagnosis. The latter lesion, however, normally presents with no sclerotic margin (unlike chondromyxoid fibroma, which is demarcated from normal bone by a distinct sclerotic border).

Smaller lesions, particularly those located in the metaphysis, should be differentiated from *bone abscess* (*Brodie abscess*).

Pathology

On histologic examination, chondromyxoid fibroma may resemble *myxoid chondrosarcoma, chondroblastoma,* and *enchondroma* (185,190). However, in chondromyxoid fibroma the stellate cells on the myxoid background lack mitotic activity, and the cells at the periphery of the lobules are usually similar in shape to chondroblastoma cells. Conversely, in *myxoid chondrosarcoma* the mitotic activity is found at the peripheries of the lobules. In addition, myxoid chondrosar-



Figure 3-73 Chondromyxoid fibroma: skeletal sites of predilection, peak age range, and male-to-female ratio.





Figure 3-74 Chondromyxoid fibroma. Anteroposterior **(A)** and lateral **(B)** radiographs of the knee of a 12-year-old girl show a radiolucent, slightly lobulated lesion with a thin sclerotic margin in the proximal tibial diaphysis. Note the lack of visible calcifications.

В



Figure 3-75 Chondromyxoid fibroma. Anteroposterior **(A)** and lateral **(B)** radiographs of the proximal left leg of an 8-yearold girl demonstrate a radiolucent lesion extending from the metaphysis into the diaphysis of the tibia, exhibiting a geographic type of bone destruction, a sclerotic scalloped border, and internal septa.

Α



Figure 3-76 Chondromyxoid fibroma. A: Anteroposterior radiograph of the knee of an 18-year-old woman shows a lesion located in the lateral aspect of the proximal tibia (*arrows*). B: The lesion balloons out from the cortex and is supported by a solid periosteal buttress (*arrow*), better appreciated on a tomographic cut. This feature closely resembles aneurysmal bone cyst.







Figure 3-77 Chondromyxoid fibroma: magnetic resonance imaging (MRI). A: Sagittal T1-weighted (SE, TR 600, TE 19) MRI in a 10-year-old girl shows a well-demarcated lesion in the plantar aspect of the calcaneus, displaying low signal intensity. B: Two axial T1-weighted (SE, TR 600, TE 17) images show significant amount of peritumoral edema. C: Sagittal T2-weighted (SE, TR 2000, TE 80) MRI shows the lesion displaying high signal intensity. A sclerotic border is imaged as a rim of low signal intensity.

B

coma exhibits intranuclear details such as a fine chromatin pattern and nucleoli (199). When large areas of chondroid matrix are present, chondrosarcoma is suggested, but the absence of such matrix has no significance (16). Small areas that resemble chondroblastoma are sometimes observed in chondromyxoid fibroma. However, the diagnosis can be confirmed by the lack of calcifications and the presence of a myxoid component in the latter tumor.

In particular, anatomic sites, such as the skull base and the spine, chondromyxoid fibroma may be mistaken for chordoma. In the spine, however, chondromyxoid fibroma unlike chordoma is only rarely located within the vertebral body; therefore, correlation with imaging features will help to make a correct diagnosis (184,192). In the skull base, these lesions are not strictly oriented around the midline and they lack the positive IHC reaction for cytokeratins typical of chordomas (187,191).

The radiologic and pathologic differential diagnosis of chondromyxoid fibroma is depicted in Figure 3-80.

Synovial (Osteo)chondromatosis

Synovial chondromatosis (also known as synovial osteochondromatosis or synovial chondrometaplasia) is an uncommon benign disorder marked by metaplastic proliferation of multiple cartilaginous nodules in the synovial membrane of the joint, bursa, or tendon sheath. It is almost invariably monoarticular; rarely, multiple joints may be affected. This condition is discussed in Chapter 9.

Malignant Tumors

Chondrosarcomas

Chondrosarcoma, a malignant tumor of the connective tissue, is characterized by formation of cartilage matrix by the tumor cells (265). Chondrosarcomas comprise the third most common primary malignant tumors of bone, after multiple myeloma and osteosarcoma. Accounting for approximately 20% of all primary bone



С

Figure 3-78 Histopathology of chondromyxoid fibroma. A: Band-like peripheral cellular condensation surrounds and divides the pale-staining matrix-rich central parts of tumor (hematoxylin and eosin, whole-mount section). B: Cancellous bone with dilated marrow space (*center*) contains tumor tissue separated from bone by a layer of connective tissue (hematoxylin and eosin, original magnification $\times 6$). C: At higher magnification, the edge of the tumor (*right*) exhibits a denser arrangement of cells than the central portion. Several giant cells are scattered within a loose fibrous stroma. Resorption of the trabecula is seen at left (hematoxylin and eosin, original magnification $\times 50$). D: In another area, less cellular stroma (*left*) merges with more cellular tissue containing several giant cells and extravasated red blood cells (hematoxylin and eosin, original magnification $\times 50$).



Figure 3-79 Histopathology of chondromyxoid fibroma. A photomicrograph reveals the typical lobulated appearance of this tumor. Lobules of chondromyxoid tissue (*left and right*) are separated by a septum (*center*) of cellular fibrous tissue, with occasional giant cells (hematoxylin and eosin, original magnification \times 50).

sarcomas, chondrosarcoma is twice as common in men as in women (13). The peak age of incidence is late adulthood, and very few cases of chondrosarcoma have been reported in children and adolescents (219,264,370). These tumors, when they occur in young people, often appear in unusual sites and may have a more ominous prognosis (219,273,370).

D

Chondrosarcoma preferentially affects the flat bones, limb girdles, and proximal portions of long tubular bones. Anatomic location can be classified as either intramedullary (central) or surface (peripheral). It is rare for central chondrosarcomas to develop in preexisting benign lesions; they are therefore referred to as primary chondrosarcomas. Peripheral chondrosarcomas, conversely, often arise from underlying benign lesions, such as osteochondroma, and are termed secondary chondrosarcomas (285). The other type of peripheral lesions are those developing on the surface of bone, known as juxtacortical or periosteal chondrosarcomas (311). Classification of various types of chondrosarcomas is shown in Figure 3-81. **214** — Differential Diagnosis in Orthopaedic Oncology



Figure 3-80 Radiologic and pathologic differential diagnosis of chondromyxoid fibroma.

Reports on genetic studies of chondrosarcomas often present divergent results because data about the exact type that was analyzed tumor (e.g., central/ conventional vs. peripheral/periosteal, primary vs. sec-ondary) are not given (332). Reported karyotypes in chondrosarcoma encompass single aberrations and complex rearrangements. It is noteworthy that some alterations at certain chromosomal regions appear to be recurrent, e.g., losses of complete chromosomes or chromosomal regions 1p36, 1p13-p22, 4,5q13-q31, 6q22-qter, 9p22-pter, 10p, 10q24-qter, 11p13-pter, 11q25, 13q21-qter, 14q24-qter, 18p, 18q22-qter, and 22q13 or gains of 7p13-pter, 12q15qter, 19, 20pter-q11, and 21q. In contrast to peripheral chondrosarcomas, central conventional chondrosarcomas show LOH at 9p and an extra copy of chromosome 22. Losses of 13q sequences have been associated with development of metastases, independent of tumor grade or size (298,333). In a CGH study from Scandinavia on 50 samples from 45 chondrosarcoma patients, Larramendy et al. (291) demonstrated that gains of chromosomal material, usually entire chromosomes, were five times more common than losses, whereas 14 samples did not reveal any alteration, irrespective of histologic grade. The common gains in primary tumors involved chromosomal regions 20q12–qter (37%), 20q (32%), and 8q24–qter (27%). In recurrent and metastatic lesions, gains of chromosome 7, 5q14-q32, 6p, and 12q were observed. Gains of 8q24-qter or 14q24-qter appeared to be associated with reduced overall survival. However, in multivariate analyses only tumor grade emerged as significant.

Clinical Presentation

In primary chondrosarcomas, the symptoms are usually of long duration. Pain usually develops insidiously, but no mass is apparent. Conversely, secondary chondrosarcomas arising at peripheral sites may be asymptomatic despite the presence of a large mass. Peripheral chondrosarcomas that occur at sites with large potential spaces (e.g., the pelvis) may not be clinically detected until they reach an immense size.

Chondrosarcomas exhibit a wide range of behaviors (334). Some are slow growing and relatively benign, whereas others are highly malignant neoplasms with associated metastases. Most of these tumors, however, are indolent and of low grade. They tend to recur locally, and metastases are uncommon or occur late in the course of disease. Nevertheless, because they are often located close to the trunk, abdomen, or pelvis, a fatal outcome resulting from repeated local recurrence associated with compromise of vital functions is not unusual. In contrast to lowgrade tumors, metastasis is common with high-grade chondrosarcomas (250).

Imaging

Radiologic evaluation of a chondrosarcoma may require a range of imaging modalities (135), including conventional radiography, arteriography, radionuclide imaging (scintigraphy, bone scan), CT, and MRI (214,280,351).



Figure 3-81 Classification of the types of chondrosarcoma.

The single most important modality for establishing the diagnosis is radiography, which provides the most useful diagnostic information about radiologic morphology of the tumor, including the specific type of bone destruction, endosteal scalloping, calcifications (usually arcand-ring pattern), and periosteal reaction. Conventional tomography occasionally may help to better delineate the matrix calcifications, evaluate the cortex and periosteal reaction, and detect occult pathologic fracture. Arteriography is an essential technique for mapping the tumor and its vascular supply and for choosing the area most suitable for open biopsy.

In the evaluation of intra- and extraosseous extension of a tumor, CT and MRI are crucial (227,230,247, 304,327). Both modalities are highly accurate in determining the presence or absence of soft tissue invasion by tumor (83,321). MRI may be superior to CT, particularly in the coronal plane, for delineating the intramedullary extent of tumor and its relationship to surrounding structures (214). Because it reveals a sharper demarcation between normal and abnormal bone tissue than CT, MRI, particularly in evaluation of the extremities, can reliably identify the spatial boundaries of tumor masses, encasement and displacement of major neurovascular bundles, and the extent of joint involvement (270). Axial and coronal images in combination have been used to determine the extent of soft tissue invasion in relation to important vascular structures (214,351). Compared with CT, however, MR images do not clearly depict or allow characterization of calcification in the tumor matrix; in fact, large amounts of calcification or ossification may be almost undetectable (320,321). Moreover, MRI is less satisfactory than CT, or even radiography, for demonstration of cortical destruction (230,244). The use of the inversion recovery mode to obtain strongly T1-weighted images and the development of the spin-echo mode with variation of the pulse sequences have improved the evaluation of chondrosarcoma by MRI (91,247,277).

Histopathology

The histologic hallmark of chondrosarcoma is production of malignant cartilage by the tumor cells accompanied by infiltration of the marrow cavity and entrapment of preexisting bone trabeculae, destruction of normal bone, and infiltration of Haversian systems (221) (see Figs. 3-4 and 3-35). Assessment of the degree of malignancy of cartilage tumors is difficult (340). This reflects the lack of precisely defined criteria for grading the malignancy, as many tumors exhibit overlapping of the features used for gradation. Therefore, each histologic appearance of a cartilage lesion should be correlated with the radiographic appearance. The degree of malignancy of chondrosarcoma is determined by several histologic criteria. These include structural characteristics (numbers of cells and appearance of matrix), cytologic findings (size of cells, pleomorphism, nuclear detail, and presence or absence of bizarre cell forms), and replicate activity (bizarre forms, mitotic figures, and binucleated or multinucleated cells).

There is a decided correlation between the histologic structure and the clinical behavior of these tumors (335). It is therefore important to differentiate among low-grade, intermediate, and high-grade chondrosarcomas (grades 1, 2, and 3, respectively). Such differentiation is based on several histologic characteristics, including the cellularity of the tumor tissue (e.g., hypocellular, hypercellular), the degree of pleomorphism observed in cells and nuclei (e.g., the presence of double- and multinucleated cells, bizarre-shaped nuclei, large nuclei), and the degree of hyperchromasia of the nuclei (e.g., hyperchromatism) (Table 3-2 and Fig. 3-82). Certain other histologic signs are also indicative of malignancy, such as invasion of trabeculae by tumor tissue, infiltration of bone marrow and Haversian systems, and permeation through the cortex. However, these features, which are essential for the discrimination between enchondroma and low-grade chondrosarcoma, are not an integral part of an accepted grading system.

Some investigators distinguish only two grades of chondrosarcoma, low grade and high grade (300,330). Low-grade tumors have lower cell density, abundant matrix, limited pleomorphism, minimal mitotic activity, and few bizarre cells. High-grade tumors are characterized by hypercellularity (often in myxoid, fibrous, or mixed stroma), moderate to marked pleomorphism, double nuclei, bizarre forms, and high mitotic activity (Fig. 3-83). In addition, so-called pushing margins distinguish low-grade chondrosarcoma from highgrade chondrosarcoma that exhibits infiltrative margins (18,300).

The grading system published by the World Health Organization (WHO) is based on the criteria proposed by Evans et al. and considering nuclear size, nuclear staining (hyperchromasia), and cellularity:

Grade 1 tumors, in general very similar to enchondroma, are moderately cellular and contain uniformly sized plump hyperchromatic nuclei including some binucleated cells.

Grade 2 tumors are more cellular and present a greater degree of nuclear atypia, hyperchromasia, and nuclear size.

Grade 3 tumors are more cellular, pleomorphic, and atypical than grade 2 tumors and show easily detectable mitoses (221,250).

More recently, a simple grading system based on nuclear features (nuclear size, polymorphism, and chromatin structure including visibility of nucleoli) with good clinical correlation and an interobserver agreement of 80% has been proposed by Welkerling et al. (366).

Several types of chondrosarcoma are recognized, each having particular clinical, radiologic, and pathologic features (338).

Grade	Histologic Features
0.5 (borderline) 1 (low-grade)	Histologic features similar to enchondroma, but radiographic features more aggressive. Cellularity: slightly increased. Cytologic atypia: slight increase in size and variation in shape of the nuclei; slightly increased hyperchromasia of the nuclei. Binucleation: few binucleate cells are present. Stromal myxoid change: may or may not present.
2 (intermediate)	Cellularity: moderately increased. Cytologic atypia: moderate increase in size and variation in shape of the nuclei; moderately increased hyperchromasia of the nuclei. Binucleation: large number of double- and tri-nucleated cells. Stromal myxoid change: focally present.
3 (high-grade)	Cellularity: markedly increased. Cytologic atypia: marked enlargement and irregularity of the nuclei; markedly increased hyperchromasia of the nuclei. Binucleation: large number of double- and multinucleated cells. Stromal myxoid change: commonly present. Other: small foci of spindling at the periphery of the lobules of chondrocytes.

Table 3-2 Histologic Grading of Chondrosarcoma

Modified from Dahlin and Unni, 1988.



Figure 3-82 Histologic grades of chondrosarcoma. A1: Grade 1. Low-cellular tumor shows slightly abnormal nuclei (hematoxylin and eosin, original magnification ×156). A2: Grade 1. At high magnification the variation in shape of the nuclei and hyperchromasia are evident (hematoxylin and eosin, original magnification ×400). B1: Grade 2. The tumor exhibits moderately increased cellularity and several double-nucleated cells are present (hematoxylin and eosin, original magnification ×25).
B2: Grade 2. At high magnification focal myxoid changes of the stroma are evident (hematoxylin and eosin, original magnification ×25).
B2: Grade 2. At high magnification focal myxoid changes of the stroma are evident (hematoxylin and eosin, original magnification ×50).
C1: Grade 3. The tumor is markedly cellular, showing pleomorphism and hyperchromatism of the nuclei. In addition to crowding of the cells, they are lying in a myxoid matrix (hematoxylin and eosin, original magnification ×50).
C2: Grade 3. Irregularly distributed cells with marked pleomorphism of nuclei. Numerous bi-and trinucleated cells are present (hematoxylin and eosin, original magnification ×50).



Figure 3-83 High-grade chondrosarcoma. Note markedly pleomorphic cells, some with double nuclei, and some exhibiting mitosis (hematoxylin and eosin, original magnification ×235).

Primary Chondrosarcomas

Conventional Medullary Chondrosarcoma

This is the most common type of chondrosarcoma, accounting for about 80% of all cases (356). It usually occurs in men older than the age of 30 years (346), most commonly in the fourth and fifth decades of life (303). The sites most commonly affected are the pelvis and the long bones, particularly the femur and the humerus, but the tumor may develop in any part of the skeleton (268,271) (Fig. 3-84). The small bones of the hands and feet are rarely affected (237).

Imaging

Radiography is usually sufficient to establish the diagnosis. The typical chondrosarcoma exhibits a characteristic expansion of the medullary portion of the bone (368) (Fig. 3-85). Thickening of the cortex and endosteal scalloping are frequently observed, often associated with popcorn-like, comma-shaped, arc-like, or annular calcifications (302) (Fig. 3-86). The periosteal reaction may be absent, may have a noninterrupted benign appearance, or may display very aggressive features, including Codman triangle and sunburst characteristics (323). In some instances a soft tissue mass is present (Figs. 3-87 and 3-88). In exceptional cases, particularly in the early stage, the tumor may be indistinguishable from an enchondroma. Therefore, all central cartilage tumors that affect the long bones, particularly those in adults, should be considered malignant until they have been proven benign (323).

CT and MRI are invaluable techniques for evaluation and staging of primary chondrosarcomas, particularly in determining tumor extension in the bone marrow and the extent of soft tissue involvement (304) (Figs. 3-89 to 3-92). On CT examination, chondrosarcoma usually appears as a lobulated, intramedullary mass with scattered low-attenuation calcifications (Fig. 3-91). The Hounsfield values of the



Figure 3-84 Conventional chondrosarcoma: skeletal sites of predilection, peak age range, and male-to-female ratio.



Figure 3-85 Chondrosarcoma. Anteroposterior radiograph of the proximal left femur in a 52-year-old woman shows, typical for this tumor, expansion of the medullary portion of the bone due to destruction of the endocortex and periosteal new bone apposition.

bone marrow affected by the tumor are in the positive range (high attenuation), in contrast to the negative values of normal fatty bone marrow (low attenuation) (see Fig. 3-89B).

The MRI appearance of chondrosarcoma is not specific (270). In general, the tumor appears to be lobulated, with intermediate to low signal intensity on T1-weighted images and high signal intensity on T2-weighted images (268,361). Calcifications display low signal or signal void (Fig. 3-92). The hyperintense, usually homogeneous signal corresponds to areas of hyaline cartilage matrix with uniform composition and high water content (214,227,229). Visual analysis of signal characteristics on routine MR images usually does not enable the histologic type or grade of chondrosarcoma to be distinguished. MRI in the coronal plane provides a more graphic and precise definition of the intramedullary extent of the tumor, with a sharper demarcation between normal and abnormal bone tissue, than does CT (see Fig. 3-92C). For determination of the extent of tumor in the soft tissue and its relationship to important vascular structures, axial images are more effective (see Fig. 3-92D). Intravenous administration of gadolinium diethylenetriamine-penta-acetic acid (Gd-DTPA) before



Figure 3-86 Conventional chondrosarcoma. Oblique radiograph of the distal femur in a 46-year-old man shows the characteristic features of this tumor. Within the destructive lesion in the medullary portion of the bone noted are annular and comma-shaped calcifications. The thickened cortex, which is due to periosteal new bone formation in response to invasion of the cortex by the cartilaginous tumor, shows the typical endosteal scalloping.

T1-weighted spin-echo images are obtained can demonstrate enhancement of scalloped margins (reflecting hyaline cartilage lobules surrounded by fibrovascular bundles) and enhanced curvilinear septa with a distinct ring and arc pattern (reflecting the typical lobulated growth pattern of chondrosarcoma), yielding an appearance similar to that of enchondroma (218). The nonenhancing areas consist of paucicellular hyaline cartilage, cystic mucoid tissue, and necrosis (91). Some authorities point to the fact that septal enhancement of chondrosarcoma is more common in

Α



Figure 3-87 Conventional chondrosarcoma. Oblique radiograph of the knee in a 58-year-old woman shows a tumor in the proximal fibula and a soft tissue mass (*arrows*) containing chondroid calcifications.



Figure 3-88 Conventional chondrosarcoma. Anteroposterior (A) and lateral (B) radiographs of the right elbow in a 55year-old man show a tumor arising from the proximal ulna that is accompanied by a huge soft tissue mass containing chondroid calcifications.



Figure 3-89 Chondrosarcoma: computed tomography. CT evaluation of intraosseous extension of chondrosarcoma includes the following: **A:** Several contiguous axial sections, preferably 1 cm in thickness, of both affected and unaffected limbs are obtained. **B:** Hounsfield values of bone marrow are measured to determine the distal extent of tumor in the medullary cavity. A value of +85 in this example indicates the presence of tumor; a value of -48 is normal for fatty marrow. **C:** The linear measurement is obtained from proximal articular end of the bone **(A)** to the point located 5 cm distally to the tumor margin **(B).** Point C corresponds to the most distal axial section that still shows tumor in the marrow.



low-grade tumors (91); however, the validity of this has been challenged (241).

Scintigraphy of chondrosarcoma invariably shows increased uptake of radiotracer (269) (Fig. 3-93). This technique may demonstrate the extent of intramedullary involvement of the cartilage tumor better than radiography. However, it is not as accurate as MRI because the exact level of intramedullary spread of the tumor cannot be adequately evaluated. A radionuclide bone scan usually shows the extent of the bone lesion to be larger than it actually is because the areas of hyperemia and edema adjacent to the tumor also demonstrate increased activity.

Clear Cell Chondrosarcoma

This rare variant of chondrosarcoma was first described in 1976 by Unni et al. (358). It is considered a low-grade malignant tumor that exhibits clinical behavior less aggressive than that of conventional chondrosarcoma (220). Accounting for only 2% of all chondrosarcomas and 0.2% of all primary bone tumors (289), it is more common in males than in females (2:1) and usually affects patients in the third to fifth decades (120). Clear cell chondrosarcoma preferentially affects the articular ends (epiphysis) of long tubular bones (the femur and the humerus are most commonly involved), with resultant joint pain, effusions, and a limited range of motion. Flat bones are rarely affected (240). On radiography the lesion is predominantly lytic and sometimes expansive (275). There is often a sharp interface between tumor and surrounding normal bone, with a sclerotic border and occasionally with central calcifications and ossifications (Figs. 3-94 and 3-95). Endosteal scalloping is occasionally seen. Periosteal reaction is rare, as is soft tissue extension of the tumor (120,358). Because of its location and the narrow zone of transition with normal bone, the tumor resembles chondroblastoma. However, MRI appearance of clear cell chondrosarcoma differs from that of chondroblastoma, which usually shows low signal intensity on T1- and T2-weighted sequences.



Figure 3-90 Chondrosarcoma: computed tomography (CT). A: Radiograph of the right shoulder of a 62-year-old man is not adequate for demonstrating the soft tissue extension of the tumor in the proximal humerus. B: CT section through the lesion demonstrates cortical destruction and an extensive soft tissue mass (arrows).





Figure 3-91 Chondrosarcoma: computed tomography (CT). CT section through the lesion in the proximal femur of a 69-year-old man shows thickening of the cortex, endosteal scalloping, and a large soft tissue mass. The calcifications of the tumor image as low-attenuation foci.

Clear cell chondrosarcoma, on the other hand, exhibits homogeneous intermediate signal intensity on T1 weighting, which becomes heterogeneous but mostly bright on T2-weighted images (253).

To date only a few cases of clear cell chondrosarcoma have undergone genetic analysis. Most were karyotypically normal, being diploid or near-diploid, but some presented recurrent anomalies, e.g., loss or structural aberrations of chromosome 9 in four cases and gain of chromosome 20 in three cases (315). Recurrent alterations of chromosome 9, including LOH at 9p21, are also seen in conventional chondrosarcomas (333). The cell cycle-regulating gene *CDKN2A/p16*, located at 9p21, does not appear to be altered (359). However, in a case report on a laryngeal clear cell chondrosarcoma in a 57-year-old patient, Kleist et al. (286) were able to demonstrate methylation, an epigenetic mechanism (i.e., changes in gene function without alteration of the nuclear DNA sequence, leading to transcriptional inactivation of a gene) of the CDKN2A/p16 promoter region. However, because methylation may occur not only in cancer but also in aging, the importance of this finding, also observed in conventional chondrosarcomas, awaits further study (248,326,359).

Histologic studies of clear cell chondrosarcomas reveal regions of obvious cartilaginous matrix and clusters of large chondrocytes with distinct cell bound-



Figure 3-92 Chondrosarcoma: magnetic resonance imaging (MRI). Anteroposterior **(A)** and lateral **(B)** radiographs of the distal femur show typical appearance of central medullary chondrosarcoma. The cortex is destroyed, and there is a large soft tissue mass projecting posteriorly (*arrows*). **C:** Coronal T1-weighted (SE, TR 700, TE 20) MRI shows the tumor to be of low signal intensity. The calcifications display signal void. **D:** Axial T2-weighted (SE, TR 2000, TE 80) image shows intramedullary tumor displaying a high signal intensity, whereas calcifications are of low signal. The soft tissue mass shows heterogeneous signal.



Figure 3-93 Chondrosarcoma: scintigraphy. Radionuclide bone scan obtained after intravenous injection of 15 mCi (555 MBq) of technetium-labeled methylene diphosphonate (^{99m}Tc-MDP) shows increased uptake of tracer localized to the site of the tumor (same patient as in Fig. 3-92).

aries. These chondrocytes possess rather small nuclei and abundant clear or eosinophilic cytoplasm (238) (Fig. 3-96). Osteoclast-like giant cells and foci of reactive bone formation are also observed. Because osteoid trabeculae are occasionally present in clear cell chondrosarcoma (Fig. 3-97), this tumor should not be diagnosed as atypical osteoblastoma (13). In IHC studies, clear cell chondrosarcomas were positive for S-100 protein and type II collagen and, in addition, revealed immunoreactivity for type X collagen and osteonectin, both of which are found in hypertrophic chondrocytes, thus distinguishing this special type of tumor from conventional chondrosarcoma (211,231,286).

A review by Weiss and Dorfman (364) states that metastases develop in only 10% of patients with clear cell chondrosarcomas. However, in an earlier published series, late local recurrence and pulmonary and bone metastases were not uncommon. In fact, Bjornsson et al. (228) reported that 7 of 47 patients with clear cell chondrosarcoma (15%) died with metastatic disease. In a recent review of 16 cases treated at one institution, metastases developed in 3 of 16 patients in a span of 4 to 16 years (276).

It is worthwhile to note that recently three cases of dedifferentiated clear cell chondrosarcoma with an undifferentiated pleomorphic or spindle cell component have been reported (279).

Mesenchymal Chondrosarcoma

Mesenchymal chondrosarcoma, first described in 1959 by Lichtenstein and Bernstein (293), is a rare lesion, representing less than 3% of all primary chondrosarcomas, with a peak incidence in the second and third



Figure 3-94 Clear cell chondrosarcoma. A: Anteroposterior radiograph of the right hip shows a radiolucent lesion with calcifications in the femoral head. B: Computed tomography section shows a lytic character of the tumor and central chondroid calcifications.

Figure 3-95 Clear cell chondrosarcoma. Anteroposterior (A) and lateral (B) radiograph of the distal femur show unusual diaphyseal location of the tumor in a 32-year-old man. Note sharp demarcation of the lesion from the normal bone (narrow zone of transition) and its lytic character with central calcifications.





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Figure 3-96 Histopathology of clear cell chondrosarcoma. A: Coronal section of resected femoral head and neck (same patient as in Fig. 3-94) shows extension of the tumor from articular cartilage to the base of the neck. In the proximal part of the femoral head some uninvolved cancellous bone is seen (hematoxylin and eosin, original magnification \times 0.2). B: Large cells with clear or slightly eosinophilic cytoplasm and small nuclei are typical for this variant of chondrosarcoma. In addition, large eosinophilic deposits are present, resembling osteoid (hematoxylin and eosin, original magnification \times 200). C: Homogeneous area of large polygonal cells is in solid arrangement displaying a clear or slightly eosinophilic cytoplasm and small, dark, roundish nuclei. Some giant cells (*left and right middle*) are also present. Note small areas of necrosis (*center and bottom*) (hematoxylin and eosin, original magnification \times 50).





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Figure 3-97 Histopathology of clear cell chondrosarcoma. A: Large and vacuolated cells interspersed in the intercellular cartilaginous matrix. Note small osteoid trabeculae (hematoxylin and eosin, original magnification $\times 150$). **B:** In another area, vacuolated cells are seen in close proximity of reactive bone trabeculae (hematoxylin and eosin, original magnification $\times 75$).

decades (223). Males and females are equally affected. In about 30% of cases the tumor develops in an extraskeletal site (312). Long-standing pain, swelling, and a soft tissue mass are typical clinical findings. This tumor exhibits no characteristic radiologic pattern. In some instances it may resemble conventional chondrosarcoma or another type of more highly malignant tumor, characterized by aggressive, lytic bone destruction or a permeative pattern of bony infiltration associated with a soft tissue mass (Figs. 3-98 and 3-99). It appears to have a predilection for the axial skeleton (particularly mandible and maxilla), femur, fibula, and ribs (313). On histologic examination, mesenchymal chondrosarcoma consists of two components: (a) small, round, uniform-sized cells of mesenchymal tissue with round or ovoid nuclei that resemble those seen in Ewing sarcoma, occasionally intermingled with spindleshaped cells, alternating with (b) areas of welldifferentiated cartilage containing foci of calcification and occasionally metaplastic bone encasement (331), representing endochondral ossification. The microscopic appearances of these components are strikingly different (Fig. 3-100). An electron microscopic study by Steiner confirmed the bistructural characteristics of this tumor (348). In some portions the cells may be clustered, often arranged in an alveolar pattern resembling that observed in malignant lymphoma or in embryonal rhabdomyosarcoma and other small cell tumors (18) or may have the antler-like arrangement of capillary vessels seen in hemangiopericytoma (Fig. 3-100D). In addition, reticulin fibers surround individual or small groups of tumor cells (Fig. 3-100E). However, the chondroid component is the differentiating feature.

To date, six patients have been investigated cytogenetically, two of whom presented with a Robertsonian translocation [a translocation of two acrocentric chromosomes (chromosomes with a centromere at the terminal region as chr 13, 14, 15, 21, and 22)by joining of the long arms, while both extremely short arms, containing no relevant genes, are lost] involving chromosomes 13 and 21 [der(13;21)q10;q10)] (314). Further studies are needed to determine whether or not this represents a tumor-specific finding. According to Park et al. (318), the IHC overexpression of TP53 appears to be an epigenetic phenomenon because only a few cases have revealed low



Figure 3-98 Mesenchymal chondrosarcoma. Anteroposterior **(A)** and lateral **(B)** coned-down radiographs of the right leg of a 43-year-old woman with a 6-month history of intermittent pain in the right calf show a destructive lesion at the midportion of the fibula. The central portion of the lesion exhibits annular and comma-shaped calcifications typical of a cartilage tumor, but its periphery shows a permeative type of bone destruction characteristic of round cell tumors.



Figure 3-99 Mesenchymal chondrosarcoma: magnetic resonance imaging (MRI). A: Axial T1-weighted (SE, TR 600, TE 20) MRI shows a focus of intermediate signal within low signal of the lateral cortex of the fibula (*arrow*). A soft tissue mass displays a signal slightly higher than that of muscles but lower than that of subcutaneous fat (*open arrows*). Axial **(B)** and coronal **(C)** T2-weighted (SE, TR 2000, TE 80) images better demonstrate a destruction of the fibular cortex. On these sequences the soft tissue mass becomes bright. **D:** On T1-weighted image after intravenous administration of gadolinium there is enhancement of the tumor.

or reduced levels of the *TP53* gene, indicating genomic deletions.

The undifferentiated tumor cells of mesenchymal chondrosarcoma are positive by IHC for vimentin, type IIA collagen, and the transcription factor Sox9, a regulator of chondrocyte differentiation, and for CD99 (Fig. 3-100F), whereas the cartilaginous islands are positive for S-100, aggrecan, type IIB collagen, and type X collagen (212,260,309,362).

Biologically, the behavior of mesenchymal chondrosarcoma is unpredictable. Although it tends to be aggressive and to exhibit a high rate of metastasis, some patients may not develop metastatic spread for 15 to 20 years after removal of the tumor, whereas other patients develop distant metastases soon after diagnosis. At present, no clinicopathologic features have been described that can distinguish this different biological behavior for these histologically identical tumor types.

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Figure 3-100 Histopathology of mesenchymal chondrosarcoma. A: Islands of fairly well differentiated chondroblastic tissue are surrounded by densely arranged small spindle cells (hematoxylin and eosin, original magnification $\times 25$). B: At higher magnification the clear-cut distinction between two components, spindle cell (left) and chondroblastic (right), is obvious. A narrow band of necrotic tissue lies side-by-side with the chondroblastic area (middle top) (hematoxylin and eosin, original magnification ×50). C: A high magnification photomicrograph shows the interface between cartilaginous and mesenchymal components of the tumor (hematoxylin and eosin, original magnification $\times 400$). **D**: Hemangiopericytomalike pattern is occasionally seen due to numerous slit-like capillary vessels (hematoxylin and eosin, original magnification ×200). E: Cellular areas contain reticulin fibers encircling single and small nests of tumor cells (Novotny, original magnification ×200). F: Small and round tumor cells are positive for CD99, similar to Ewing sarcoma; however, the cartilaginous component (upper left) excludes the latter diagnosis (ABC-Elite, original magnification \times 400).

Myxoid Chondrosarcoma

This variant, occasionally referred to as chordoid sarcoma, represents approximately 12% of all chondrosarcomas of bone (255,272,283,347). Lichtenstein and Bernstein (293) were the first to describe this kind of lesion under the term solitary chondroblastic sarcoma. Enzinger and Shiraki first proposed the term myxoid chondrosarcoma for low-grade malignant neoplasms of soft tissue, similar to those arising primarily in bone, which were lobulated and composed of round stellate cells with abundant mucoid matrix and histochemical characteristics of cartilage (249,365). Myxoid chondrosarcoma of bone affects predominantly males, with a wide range of age, between 9 and 76 years. In about 50% of reported cases (283) the femur was the site of occurrence. There are no radiologic hallmarks of this neoplasm. Most commonly the tumor is radiolucent, sharply circumscribed, and lobulated and extends into the surrounding soft tissues (347) (Fig. 3-101A-C). Despite its locally aggressive features, it may be mistaken for a benign tumor, such as chondromyxoid fibroma or desmoplastic fibroma.

The histologic features of myxoid chondrosarcoma include round and stellate cells, some with acidophilic cytoplasm, separated by an abundant and pale basophilic matrix (Fig. 3-101D,E). A few mitotic figures may sometimes be seen (347). The presence of acid sulfated mucopolysaccharides consistent with cartilage may be proved by Alcian blue staining of the matrix, which remains unchanged after digestion with testicular hyaluronidase.

Dedifferentiated Chondrosarcoma

Originally described by Dahlin and Beabout in 1971 (243), this tumor represents approximately 10% of all

chondrosarcomas and overall has the worst prognosis. Histologically it is characterized by a relatively low-grade chondrosarcoma with juxtaposed dedifferentiated, i.e., less differentiated, noncartilaginous sarcomatous components including osteosarcoma, fibrosarcoma, MFH, rhabdomyosarcoma, angiosarcoma, giant cell tumor, and undifferentiated sarcoma (224,236,274,278,308, 354). Rare cases of dedifferentiated peripheral chondrosarcoma and dedifferentiated clear cell chondrosarcoma have also been reported (224,279). The age group affected and the anatomic predilection are the same as that of conventional chondrosarcoma (306), and men and women are equally affected. Patients with this tumor often present with rapid onset of swelling and local tenderness, superimposed on a long history of less severe pain (254,290). The chronic pain may be associated with a low-grade malignant lesion, and the sudden change in the pattern of symptoms probably reflects the emergence of a more malignant tumor (324). Pathologic fractures are not uncommon, and metastasis to distant organs is common. Radiography often reveals remodeling expansion of the involved segment with variable degrees of mineralization; evidence of a long-standing, low-grade cartilage tumor; and a superimposed, more destructive moth-eaten or permeative pattern (Fig. 3-102). Similarly, CT and MRI reveal two distinct areas with different intrinsic characteristics, reflecting the presence of lowgrade and high-grade chondrosarcomatous elements (296,311) (Fig. 3-103). Often, there is an accompanying soft tissue mass (319,343) (Fig. 3-104). Histologic studies demonstrate that the tumor usually arises from the site of a grade 1 chondrosarcoma, either spontaneously or after surgical treatment and recurrence of the latter. It is composed of low-grade cartilage tissue surrounded by a high-grade sarcomatous component, with a clear



Figure 3-101 Myxoid chondrosarcoma. A: Lateral radiograph of the hindfoot of a 65-year-old woman shows a large destructive lesion in the calcaneus. The tumor extends into the soft tissues (*arrow*). B: Lateral tomogram shows soft tissue mass more effectively (*arrows*) (*continues*).



Figure 3-101 Continued C: Lateral radiograph of the amputated specimen demonstrates in addition a pathologic fracture (*arrow*). **D:** Photomicrograph shows a myxoid pattern of cells uniform in size, separated by abundant and pale basophilic matrix. The tumor invades bone trabeculum (*left*) (hematoxylin and eosin, original magnification ×25). **E:** At higher magnification the tumor cells are rounded or elongated, fairly uniform in size with prominent nuclei. Observe cartilaginous and myxoid features of the tumor (hematoxylin and eosin, original magnification ×640).

delineation between the two components (Fig. 3-105). The high-grade tumor tissue is usually a fibrosarcoma, MFH, or osteosarcoma (236,343) (Figs. 3-106 and 3-107). The problem is further complicated by the fact that in a considerable number of cases (up to 33% in larger series) the chondrosarcoma arises on the site of a usually calcified enchondroma, so that the manifestations of all three tumors, one benign and two malignant (enchondroma, chondrosarcoma, and dedifferentiated chondrosarcoma) may be present side by side (Fig. 3-108). Although the term "dedifferentiation" implies that the high-grade tumor elements arise from genetic alterations in the chondrocytes of the low-grade cartilage component that cause them to grow more rapidly, these tumors may theoretically also derive from a neoplastic cell clone that by genomic instability develops into a differentiated and undifferentiated (dedifferentiated) tumor population (301). Recent cytogenetic and molecular genetic findings obtained by analysis of the welldifferentiated and the undifferentiated parts of such a sarcoma in comparison support the latter assumption (232,329,333). It was demonstrated that both components share an identical p53 mutation, not belonging to the frequently mutated hotspots and therefore indicating a monoclonal origin, as well as LOH at the same alleles of chromosomes 17p and 13 (232). Similar results have been obtained by Ropke et al. (329) in a study on methylation of cell cycle-related, apoptosis-related, and cell adhesion-related genes. However, no specific cytogenetic aberrations were demonstrated in dedifferentiated chondrosarcoma. Support for this hypothesis has also come from an immunohistochemical and biochemical study of cultured human chondrosarcoma, in which the existence of four different clonal cell lines within a parent tumor was demonstrated (257a). Patients with dedifferentiated chondrosarcoma usually do not survive for more than 2 years after diagnosis, even with multimodal treatment strategies (245).

Periosteal (Juxtacortical) Chondrosarcoma

Although most primary chondrosarcomas arise centrally in bone, in rare instances a primary tumor originates in a periosteal (juxtacortical) location (222). Initially described in 1955 by Lichtenstein (292) and later by Scha**Figure 3-102 Dedifferentiated chondrosarcoma.** Anteroposterior radiograph of the left shoulder of a 70-year-old woman shows a lytic lesion with chondroid calcifications in the humeral neck extending into the proximal shaft. The proximal part of the lesion is consistent with a slow-growing tumor (*arrow*), whereas the distal part exhibits a more aggressive appearance, including cortical destruction and a soft tissue mass (*open arrow*).





Figure 3-103 Dedifferentiated chondrosarcoma: magnetic resonance imaging (MRI). A: Anteroposterior radiograph of the left hip of a 70-year-old woman shows a mixed radiolucent and sclerotic lesion in the intertrochanteric region of the femur with central chondroid calcifications (*arrows*). B: Axial T1-weighted MRI shows tumor (*arrows*) to exhibit heterogeneous signal. C: Coronal T1-weighted MRI obtained after intravenous injection of gadolinium shows peripheral enhancement of the tumor. The mineralized areas representing low-grade sarcoma remain of low signal intensity.





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Figure 3-104 Dedifferentiated chondrosarcoma: magnetic resonance imaging (MRI). A: Anteroposterior radiograph of the proximal right femur of a 60-year-old man shows a predominantly osteolytic, destructive lesion in the subtrochanteric region (*arrows*). **B:** Coronal T2-weighted (STIR) MRI shows the high-signal-intensity tumor breaking through the medial cortex to form a large soft tissue mass.



Figure 3-105 Histopathology of dedifferentiated chondrosarcoma. A photomicrograph shows low-grade chondrosarcoma (*left*) that is clearly separated from the pleomorphic tumor tissue (*right*) (hematoxylin and eosin, original magnification \times 50).







Figure 3-107 Histopathology of dedifferentiated chondrosarcoma. A: In one part of the tumor there is a grade 2 chondrosarcoma with irregular distribution of chondrocytes (hematoxylin and eosin, original magnification \times 50). **B:** The other areas show dedifferentiated part of tumor with spindle-cell tissue and tumor-bone formation (*center*) typical for osteosarcoma. Some giant cells are also present (hematoxylin and eosin, original magnification \times 50) (*continued*).

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Figure 3-107 *Continued* **C:** Another field of view shows differentiation of giant cell–containing spindle cell tissue, consistent with malignant giant cell tumor (hematoxylin and eosin, original magnification 350).



Figure 3-108 Histopathology of dedifferentiated chondrosarcoma. A: Coronal section of the specimen of the distal femur shows calcified enchondroma (*round whitish area in center*), well-differentiated chondrosarcoma (*darker area medially*), and dedifferentiated tumor (*rose area distally*). Site of the biopsy is marked by hemorrhagic area proximally. **B:** Histologic overview of the medial half of the distal femur shows three different tumors: calcified enchondroma (*proximal middle*), well-differentiated chondrosarcoma (*section close to the medial cortex on the left*), and dedifferentiated tumor with osteoblastic differentiation (*distal middle*) (hematoxylin and eosin, original magnification \times 0.2). The details of osteoblastic differentiations are depicted in Figure 2-134.



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Figure 3-108 Continued **C:** Calcified enchondroma (*upper half*) is bordered by well-differentiated chondrosarcoma with myxoid component (*lower half*) (hematoxylin and eosin, original magnification $\times 25$). **D:** Two different tissues are clearly separated: on the left dedifferentiated part of tumor formed of spindle cells and some giant cells, abutting the well-differentiated chondrosarcoma on the right (Giemsa, original magnification $\times 50$).

jowicz (336), and accounting for only 4% of all malignant cartilage tumors, periosteal chondrosarcoma arises on the surface of a bone to which it is firmly attached but with no associated exostosis. Only one case, in a 9-yearold girl, has been reported of periosteal chondrosarcoma with associated multiple osteochondromata (363). On rare occasions this tumor may invade the medullary cavity (263,328). This malignancy, which most commonly affects adults in the third and fourth decades of life and has slight male predilection (311), presents clinically as an asymptomatic or a slightly painful, slowly growing mass. On radiologic and pathologic studies, it exhibits the same general features as a central chondrosarcoma (336,360) (Figs. 3-109 to 3-112) with nodular invasion of surrounding soft tissues (221). At times the underlying cortex may be thickened with associated saucerization or Codman triangles at the lesion margins (222,311). Most of these tumors represent well-differentiated grade 1 or 2 cartilage malignancy (Fig. 3-113), and the prognosis after surgical excision is therefore good. Metastases develop only rarely. On radiography they are occasionally mistaken for periosteal osteosarcomas, which also occur on the external surface of long bones and consist largely of cartilage.

Synovial Chondrosarcoma

This exceptionally rare tumor, which arises from the synovial membrane, may be primary or secondary. Primary lesions arise *de novo* in the joint synovium without evidence of previous synovial chondromatosis (226,367). In radiographic studies, synovial chondrosarcoma may mimic synovial chondromatosis or periosteal chondrosarcoma. Secondary tumors are believed to arise from malignant transformation of synovial chondromatosis (261) (see Chapter 9).

Because very few cases of synovial chondrosarcoma have been documented, it is difficult to compare the behavior of these tumors to that of chondrosarcoma of bone. However, the former appears to be characterized by a spectrum ranging from low-grade neoplasms to high-grade tumors (259,284,297,299).

Extraskeletal (Soft Tissue) Chondrosarcoma

These tumors develop in the soft tissues and are independent of bone, periosteum, or cartilage. It is usually not possible to determine the exact site at which they arise. However, it appears that these neoplasms originate in muscle, fascia, bursa, and other connective tissue structures. Patients typically present with a slowly enlarging soft tissue mass, and approximately one third of cases are characterized by pain or tenderness (215,249,316). On radiography and CT scans this type of



Figure 3-109 Periosteal chondrosarcoma. Anteroposterior radiograph of the lumbar spine shows a large calcified mass attached to the lateral aspect of the third lumbar vertebra. (Reprinted with permission from Bullough PG. *Atlas of orthopedic pathology*, 2nd ed. New York: Gower, 1992:16.31.)



Figure 3-110 Periosteal chondrosarcoma: magnetic resonance imaging (MRI). A: Sagittal T1-weighted MRI shows an intermediate signal intensity lesion arising from the cortex of the clavicle and extending into the soft tissues (*arrows*). **B:** Protondensity MRI demonstrates that the lesion now exhibits a bright signal (*open arrow*).



Figure 3-111 Periosteal chondrosarcoma. A: Anteroposterior radiograph of the right knee of a 30-year-old woman shows a parosteal calcified mass at the medial cortex of distal femur. B: Radionuclide bone scan obtained after intravenous administration of 15mCi (555MBq) of 99mTc-labeled methylene diphosphonate shows markedly increased uptake of radiotracer within the mass.

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Figure 3-111 *Continued* **C:** Coronal T1-weighted magnetic resonance imaging (MRI) shows the mass to be isointense with the surrounding muscles displaying intermediate signal intensity. **D:** On coronal T2-weighted MRI the mass becomes bright, but the central calcifications exhibit low signal intensity.



Figure 3-112 Histopathology of periosteal chondrosarcoma. A: Lobulated chondroblastic tumor is clearly separated from adjacent connective tissue (*top*) (hematoxylin and eosin, original magnification $\times 12$). **B:** At higher magnification there is obvious pleomorphism of the cells and hyperchromatism of the nuclei, proving malignancy (hematoxylin and eosin, original magnification $\times 50$).



Figure 3-113 Histopathology of periosteal chondrosarcoma. A: Hyaline cartilage with irregular cell arrangement and peripheral condensation of tumor cells exposes malignant character of the tumor (hematoxylin and eosin, original magnification \times 50). **B:** At higher magnification in another area, high cellularity and conspicuous pleomorphism of the cells and nuclei are consistent with grade 2 chondrosarcoma (hematoxylin and eosin, original magnification \times 100).

chondrosarcoma appears as a soft tissue mass with mineralization similar to that observed in conventional intramedullary chondrosarcomas (Fig. 3-114). However, calcifications occur only in 30% to 50% of cases (311,341) and ordinarily exhibit a fine granular pattern in focal or uniform distribution, particularly in the mesenchymal variant (262). Adjacent bone destruction may or may not be present (Fig. 3-115). On MRI examination, the mass is usually isointense to the surrounding tissues on T1-weighted sequences, and of a high signal intensity on T2 weighting (242,353). After administration of Gd-DTPA, heterogeneous enhancement of the tumor is observed on T1-weighted images in unmineralized and mineralized areas (262,287). Scintigraphy demonstrates increased uptake of technetium-labeled methylene diphosphonate (99mTc-MDP) that is related to hyperemia, calcifications, endochondral ossification, and reactive new bone formation in these tumors (210, 294).

On histologic examination, the lesion is characterized by undifferentiated mesenchymal cells with only rare islands of well-differentiated cartilage (18,352). The tumor may consist of well- or poorly differentiated hyaline cartilage (conventional soft tissue chondrosarcoma), may contain mesenchymal tissue (mesenchymal chondrosarcoma), or may be partially or entirely myxoid in appearance (myxoid chondrosarcoma) (77,341). The latter is the most common type of extraskeletal chondrosarcoma.

Extraskeletal myxoid chondrosarcoma, first described by Stout and Verner in 1953 (349), was originally believed to be derived from multipotential mesenchymal progenitor cells, because the cells are similar to chondroblasts histologically, histochemically, and ultrastructurally (249,252,305). In addition, S-100 protein and type II collagen have been detected by IHC in at least some portions of the tumors investigated (252,355). However, recent reports point to a neuroendocrine differentiation of the tumor cells (246,258). In addition, Aigner et al. (213) found an only partially expressed chondrocytic immunoprofile in 2 of 14 sarcomas investigated, leading to the assumption that myxoid chondrosarcoma is a tumor derived from primitive mesenchymal cells with multidirectional differentiation, including a chondrocytic phenotype.

Myxoid chondrosarcomas, as other types of extraskeletal chondrosarcomas, develop in the soft tissues and are independent of bone, periosteum, or cartilage. Although intraosseous cases have been described, molecular and ultrastructural differences clearly distinguish cartilaginous tumors, with their appearance similar to extraskeletal myxoid chondrosarcoma, from this entity (217). Extraskeletal myxoid chondrosarcoma represents



Figure 3-114 Intrathoracic mesenchymal chondrosarcoma. CT section of the chest of a 23-year-old man shows a large prevertebral calcified mass.



Figure 3-115 Extraskeletal chondrosarcoma. Oblique radiograph of the fifth finger of the left hand of a 47-year-old woman shows a soft tissue mass invading and destroying the proximal phalanx. There are no visible calcifications. The biopsy revealed myxoid chondrosarcoma.

about 2% of all soft tissue sarcomas (288). More than 80% of soft tissue chondrosarcomas affect the lower extremities, particularly in the regions of the thigh and knee (216,369). These are usually deep-seated tumors, which preferentially occur in the fifth or sixth decade (48%; range 6 to 89 years), and have a predilection for middle-aged men (267,281).

On histologic examination, the lesion is characterized by undifferentiated mesenchymal cells, with rare islands of well-differentiated cartilage (352). This tumor is typically surrounded by a pseudocapsule, formed by compression of normal connective tissue that presents a glistening, multinodular architecture on its cut surface. Histology reveals that the nodules are divided by poorly vascularized fibrous septa and show a pale chondroid to myxoid stroma rich in proteoglycans. The cells are well circumscribed, with eosinophilic cytoplasm and round to oval nuclei with fine chromatin and usually small nucleoli. Mitoses are not common. The cells are characteristically interconnected, forming cords, net-like structures, or aggregates. Spindle-shaped tumor cells may also be present, as well as cells with rhabdoid features (Fig. 3-116A-D). In some instances, anaplastic areas with pleomorphic cells and atypical mitoses are present. By IHC, only vimentin is constantly expressed. S-100 protein, epithelial membrane antigen (EMA), and cytokeratins may focally be found (Fig. 3-116E). Positivity for NSE and synaptophysin has also been reported (258,295).

CHAPTER 3 Cartilage (Chondrogenic) Lesions — 239

In 1985, Hinrichs et al. (266) described a translocation in extraskeletal myxoid chondrosarcoma that involved chromosomes 9 and 22. This nonrandom t(9;22)(q22;q12) translocation has been observed in about 50% of extraskeletal myxoid chondrosarcomas and, when present, is considered diagnostic for this tumor (332). At the molecular level, this translocation leads to the fusion of the NR4A3 gene (Nuclear Receptor subfamily 4, group A, member $\overline{3}$), which is involved in regeneration and proliferation, with the EWS gene, which was identified in Ewing sarcoma and encodes a protein (EWS protein) that participates in transcriptional regulation. Variant translocations have also been observed in single cases, always involving chromosomal region 9q22 (332). Gene expression profiling studies have provided evidence that targets of the fusion transcripts in extraskeletal myxoid chondrosarcoma are genes that are involved either in neural and neuroendocrine differentiation or in chondrogenesis (344,350). Recurrent abnormalities, especially trisomies 1q, 7, 8, 12, and 19, were also observed. 1q aberrations occurred only in tumors that also exhibit the t(9;22)(q22;q12) translocation (332,344).

Extraskeletal mesenchymal chondrosarcoma is histologically composed of two elements: primitive mesenchymal cells and well-differentiated cartilaginous tissue (262). The exact molecular mechanism is still uncertain, but recent cytogenetic studies have suggested that the chromosomal translocation der(13;21)(q10;q10) may be a causative factor (314). For additional details see previous discussion on mesenchymal (osseous) chondrosarcoma.

Secondary Chondrosarcomas

The term secondary chondrosarcoma is applied to any tumor that arises as a result of malignant transformation of a benign cartilaginous lesion, such as osteochondroma or enchondroma. It also applies to the rare cases of Paget disease (345) or irradiated bone that lead to development of a cartilaginous malignancy. These tumors, which constitute about 20% of all chondrosarcomas, may be either central or peripheral. The most common are peripheral lesions, which develop as malignant transformation of either hereditary multiple exostoses (about 3% of patients with this condition develop chondrosarcoma) or solitary osteochondroma (<1% of patients are at risk). Central secondary chondrosarcomas develop as malignant transformation of enchondroma (rarely in patients with a solitary lesion and much more commonly in patients with multiple enchondromatosis, e.g., Ollier disease and Maffucci syndrome). Exceptionally, secondary chondrosarcoma develops in the joint as a malignant transformation of synovial chondromatosis (310).

In malignant transformation of osteochondroma to chondrosarcoma (so-called exostotic chondrosarcoma), the most reliable clinical and radiologic features include pain (in the absence of fracture, bursitis, or pressure on nearby nerves), the growth spurt of osteochondroma (after skeletal maturity), development of a bulky cartilaginous cap (thicker than 2 cm), dispersed **240** — Differential Diagnosis in Orthopaedic Oncology


calcifications in the cartilaginous cap (which in benign exostosis are confined to the osteochondral junction), and development of a soft tissue mass (110) (Fig. 3-117; see also Figs. 3-52, 3-56, and Table 3-2). Scintigraphy is not a reliable modality for differentiation of benign exostoses from exostotic chondrosarcoma.

Some chondrosarcomas are believed to originate from benign enchondromas (18,324,325). However, it

is almost impossible to establish the evolution of a preexisting enchondroma to a chondrosarcoma by histologic means alone, particularly when the former is a solitary lesion. Some investigators believe that in these rare cases the preexisting enchondroma may in fact represent a low-grade chondrosarcoma that masquerades as a benign lesion. Nevertheless, malignant transformation of a calcified enchondroma has been well







B

С

documented in patients with dedifferentiated chondrosarcoma, with Ollier disease, and with Maffucci syndrome (234) (Figs. 3-118 and 3-119).

The radiologic signs of malignant transformation of enchondroma to chondrosarcoma include thickening or destruction of the adjacent cortex, deep endosteal scalloping, appearance of a periosteal reaction without evidence of a pathologic fracture, and development of a soft tissue mass. Pain at the site of the lesion in the absence of fracture is an important clinical sign of evolving malignancy.

Differential Diagnosis

Radiology

Radiologic differential diagnosis of **conventional chondrosarcoma** includes mainly an *enchondroma* (256). Among other clinical signs, the malignancy of a lesion is suggested by the development of pain in a previously asymptomatic lesion (e.g., malignant transformation of enchondroma to chondrosarcoma) or by development of swelling at the site of a lesion. The most obvious signs of malignancy are thickening of the cortex, deep endosteal scalloping, cortical destruction, periosteal reaction, and a soft tissue mass

(see Figs. 3-85 to 3-88). If any of these features are present, the diagnosis can be made unequivocally. A more difficult task is to differentiate low-grade chondrosarcoma from enchondroma. In some cases this is almost impossible, although the focally thickened cortex may point to the former tumor (Fig. 3-120; see also Fig. 3-29). It must be stressed that neither CT nor MRI is usually suitable for establishing the precise nature of a cartilage lesion. In particular, too much faith has been placed in MRI as a method helpful in distinguishing benign cartilage lesions from malignant ones. Despite the use of various criteria, the application of MRI to tissue diagnosis has rarely achieved satisfactory results (371) because benign and malignant chondrogenic lesions have the same MR characteristics. Both exhibit a low to intermediate signal intensity compared with normal bone marrow on T1-weighted images and a high signal intensity on T2-weighted images (128,229). Quantitative determination of relaxation times is of little clinical value in identifying cartilage tumor types (247), although it is an important technique in the staging of chondrosarcoma (225). Some investigators suggested that gadolinium-enhanced MR images may distinguish malignant cartilage tumors from benign lesions (257).



Α

B

Figure 3-118 Malignant transformation of Ollier disease. A: A 38-year-old woman with known long-standing Ollier disease since childhood presented with pain and a slowly enlarging mass at the dorsomedial aspect of the fifth finger of the right hand. The radiograph shows typical changes of enchondromatosis in the hand. Also noted is destruction of the cortex of the proximal phalanx of the small finger and a soft tissue mass containing chondroid calcifications, indicating malignant transformation to chondrosarcoma. **B:** A 30-year-old man with enchondromatosis presented with pain and an enlarging mass at the base of the ring finger of the right hand. Radiograph shows typical changes of enchondromatosis. Also noted is destruction of the cortex of the proximal and middle phalanges of the ring finger associated with adjacent soft tissue masses (*arrows*). The open biopsy revealed malignant transformation of enchondroma into chondrosarcoma.



Figure 3-119 Malignant transformation of Ollier disease. A: A 48-year-old watchmaker with a long history of Ollier disease developed pain in the ring finger. An oblique radiograph of the right hand demonstrates characteristic changes of this disorder consisting of multiple enchondromas and large lobulated cartilaginous masses in all fingers. The lesion of the middle phalanx of the ring finger shows destruction of the cortex and the extension of the tumor into the soft tissues. Excision biopsy revealed chondrosarcoma. **B:** Longitudinal section of the distal phalanx of the left thumb with nail (*clear blue lamella*) of another patient shows cancellous bone with large islands of chondrosarcoma (*purple*) proliferating into the surrounding soft tissue (*center*) (Giemsa, original magnification \times 5). **C:** At a higher magnification, the malignant cartilage exhibiting spindle cells in dense arrangement, with clearly polymorphous, hyperchromatic nuclei and a myxoid matrix, borders the fibrous connective tissue of the distal phalanx (*upper left*) (Giemsa, original magnification \times 20).



Figure 3-120 Low-grade chondrosarcoma. Focal thickening of the cortex at the greater trochanter and expanded character of the lesion point to malignancy.

They postulated that enchondromas show peripheral enhancement, whereas chondrosarcomas demonstrate more diffuse enhancement (218,244,257). However, Murphey et al. (311), in a retrospective study of 92 enchondromas and 95 chondrosarcomas of the appendicular skeleton, challenged this contention. Recently, investigations by Feldman et al. (251) suggested that application of positron emission tomography (PET) scanning with ¹⁸fluoro-2-deoxy-Dglucose (¹⁸FDG) may be a promising technique for distinguishing benign from malignant cartilage neoplasms.

Occasionally, **central chondrosarcoma** located in the proximal femur or humerus may mimic *osteonecrosis*. Careful examination of the radiograph for the presence of cartilage calcification and MRI findings typical of an osteonecrotic pattern (i.e., a clear demarcation at the reactive interface between live and dead bone), as well as MRI characteristics for each class of osteonecrosis (239), should help in making this distinction. Because **clear cell chondrosarcoma** can arise before skeletal maturity, it may be indistinguishable from chondroblastoma (235), although the former tumor is usually larger in size (average 5.2 cm, vs. 2.3 cm for chondroblastoma). MRI may be somewhat helpful in differential diagnosis, despite sharing similar features of heterogeneous signal by both lesions. After injection of gadolinium, contrast enhancement tends to be more intense with clear cell chondrosarcoma, whereas chondroblastoma is more frequently associated with peritumoral bone marrow edema and synovitis (147). Clear cell chondrosarcoma also can mimic a giant cell tumor, as both these tumors preferentially affect the articular end of a bone (Fig. 3-121). Giant cell tumor may be distinguished by the lack of matrix calcifications (although clear cell chondrosarcoma sometimes presents also as a purely lytic lesion). Mesenchymal chondrosarcoma, because of a permeative or moth-eaten type of bone destruction, may be mistaken for a round cell tumor. Myxoid chondrosarcoma, despite its locally aggressive features, could be mistaken for a benign tumor such as chondromyxoid fibroma or desmoplastic fibroma. Periosteal chondrosarcoma may be indistinguishable from periosteal osteosarcoma (see text on osteosarcoma). Occasionally, periosteal chondroma may be similar in appearance to periosteal chondrosarcoma but usually is much smaller (average 2 cm) in size (328). Soft tissue chondrosarcoma may mimic soft tissue osteosarcoma, myositis ossificans, and tumoral calcinosis, and vice versa (Fig. 3-122). Although synovial chondrosarcoma may be difficult to distinguish from synovial chondromatosis, MRI is occasionally helpful in this respect (317) (see Fig. 9-36).



Figure 3-121 Clear cell chondrosarcoma resembling giant cell tumor. A lytic lesion in the femoral head in a 39-year-old woman was thought to represent a giant cell tumor. Excision biopsy revealed clear cell chondrosarcoma.

Pathology

Histopathologic differential diagnosis of chondroblastic tumors is sometimes difficult because some tumors may exhibit indistinct histologic and cellular characteristics (307). The diagnosis is, however, dependent on the location of a given tumor within the skeleton.

The pathologic features of a malignant cartilage tumor, which include the presence of hypercellular areas and pleomorphic cells, are not unequivocal. The diagnosis is then dependent on the anatomic location of a given tumor. Appreciable numbers of plump cells with large vesicular ("chondroblastic") nuclei, or double nuclei, and invasion (permeation) of bone marrow and trabeculae, as well as infiltration of compact bone are clear signs of malignancy.

A cartilage tumor with histologic signs that clearly indicate malignancy, such as marked atypia of chondrocytes, cells arranged densely in clusters, and hyperchromasia of nuclei (grades 2 and 3), is easy to diagnose, as there are no reasonable differential problems. An important differentiation point is the relation between



Figure 3-122 Extraskeletal (soft tissue) mesenchymal chondrosarcoma. Conventional radiograph shows a large mineralized mass adjacent to the lower ribs. Although the mass resembles myositis ossificans, the calcifications are not at the periphery (as in the former lesion) but centrally located (compare with Fig. 2-124), whereas extraskeletal osteosarcoma can be excluded because calcified matrix exhibits typical chondroid features (dots, rings, and arcs in "O" and "C" configuration).

cartilage tissue and trabecular bone. In enchondroma the lobuli are usually surrounded by a narrow rim of lamellar bone (Fig. 3-123A) or dense fibrous tissue, whereas the cartilage of a chondrosarcoma "flows" into the marrow spaces of the trabecular bone, with secondary induction of bone resorption (Fig. 3-123B). Differentiation of conventional chondrosarcoma from chondroblastic osteosarcoma may be difficult if the latter shows only scant bone formation. In such instances the presence of hyaline matrix in chondrosarcoma (at least in the grade l and 2 tumors) may be helpful, in contrast to the fibrocartilage of osteosarcoma. Myxoid transformation of the matrix is in itself not a sign of malignancy. However, differentiation from chondromyxoid fibroma may cause a problem if the cell atypias of the malignant tumor are not clear-cut.

A special histopathologic problem in differential diagnosis is posed by dedifferentiated chondrosarcoma. This tumor may be confused with high-grade chondrosarcoma containing areas of spindle cells, mesenchymal chondrosarcoma, chondroblastic osteosarcoma, MFH, and fibrosarcoma (16). The two latter diagnoses can be eliminated by the presence of cartilage in dedifferentiated chondrosarcoma. The areas of spindle cells seen in high-grade chondrosarcomas exhibit multiple zones of transition and intermingling of the spindle cell foci with the hyaline cartilage. Conversely, in dedifferentiated chondrosarcoma the foci of spindle cells and the hyaline cartilage are juxtaposed at the interface between the two tumor components, with no intermingling of components throughout the lesion. In a similar manner, the cartilaginous component of mesenchymal chondrosarcoma appears as specks throughout the tumor and is mingled with the major component of spindle cells or small round cells (16). In *chondroblastic osteosarcoma*, tumor bone or tumor osteoid is usually irregularly mixed with cartilage and the two malignant elements are not as distinctly separated as in dedifferentiated chondrosarcoma, but may merge onto one another in the form of chondrosteoid. Differentiation of these tumors is further complicated by the fact that in a number of cases dedifferentiated chondrosarcoma arises at the site of a usually calcified enchondroma or a low-grade chondrosarcoma, so that all three tumor manifestations, one benign and two malignant, may be found side by side (see Fig. 3-108). It therefore follows that the term "dedifferentiated chondrosarcoma" is not accurate, although well established.

Clear cell chondrosarcoma consists of rather large cells with a water-clear or more eosinophilic cytoplasm and fairly small nuclei. They are arranged in clusters, with intermingled small trabeculae or newly built reactive bone (Fig. 3-124A). Bone formation may be so extensive as to mimic an osteosarcoma (233) (Fig. 3-124B). In such instances, the location in the epiphysis and the size and characteristics of the cartilage cells are helpful. However, sporadic tumors located in the diaphysis have been observed. Histologic differential diagnosis should also include chondroblastoma. This tumor may have cells with clear cytoplasm, although they are not a predominant feature. In addition, in the latter tumor the cell nuclei are indented or cleaved, whereas in clear cell chondrosarcoma they are round (322). Furthermore, in most cases of clear cell chondrosarcomas areas typical of conventional chondrosarcoma are present. Metastatic clear cell carcinoma (e.g., from the



Figure 3-123 Histopathologic distinction of enchondroma from chondrosarcoma. A: In enchondroma, the cartilage lobules are commonly surrounded by a narrow rim of bone (hematoxylin and eosin, original magnification $\times 250$). (Reprinted with permission from Bullough PG. *Atlas of orthopedic pathology,* 2nd ed. New York: Gower, 1992:16.21.) **B:** In chondrosarcoma, the tumor is more cellular and invades the bone trabeculae (hematoxylin and eosin, original magnification $\times 100$). (Reprinted with permission from Bullough PG. *Atlas of orthopedic pathology,* 2nd ed. New York: Gower, 1992:16.32.)



В



Α

Figure 3-124 Histopathology of clear cell chondrosarcoma resembling osteosarcoma. A: Clear cell chondrosarcoma exhibiting bundles and cords of fairly large, slightly polymorphic cells with water-clear cytoplasm and small hardly polymorphic nuclei. Minimal reactive bone is present (*right upper corner*) (hematoxylin and eosin, original magnification ×50). **B:** Tumor with extremely marked bone formation mimics osteosarcoma. Clear cells in small clusters within the marrow spaces identify this lesion as clear cell chondrosarcoma (hematoxylin and eosin, original magnification ×25).

kidneys) may be difficult to distinguish from clear cell chondrosarcoma solely on the basis of its histologic appearance. The glandular architecture and lumen formation are characteristic features of the former and are not seen in clear cell chondrosarcoma. The clinical setting and the presence of chondroid differentiation are essential in confirming the latter diagnosis (322). In addition, on immunohistochemical studies renal cell carcinomas show a coexpression of vimentin and cytokeratin.

Mesenchymal chondrosarcoma may be difficult to distinguish from malignant mesenchymoma of bone because both tumors demonstrate the same biphasic pattern (337,339). In malignant mesenchymoma, however, a third component (osteosarcoma) is always present, whereas in mesenchymal chondrosarcoma the matrixproducing component is always chondrosarcoma. The nonmatrix cellular component of malignant mesenchymoma consists of liposarcoma, rhabdomyosarcoma, or both (282). The nonmatrix component of mesenchymal chondrosarcoma consists of undifferentiated round or spindle cells, frequently arranged in a hemangiopericytomatous pattern and accompanied by reticulin fibers (see Fig. 3-100D,E). Areas containing broad sheaths of small cells may mimic *Ewing sarcoma* (16). Moreover, both tumors contain cytoplasmic glycogen. In Ewing sarcoma, however, spindle cells and cartilage formation are absent. Positivity for CD99, also seen in Ewing sarcoma, in combination with Sox9 immunoreactivity is further helpful in diagnosis, as is the Ewing sarcoma-specific translocation that can also be visualized on paraffin sections with fluorescence in situ hybridization (FISH) techniques. Areas containing primarily spindle-cell tissue may resemble low-grade fibrosarcoma (Fig. 3-125). Small cell osteosarcoma may occasionally mimic mesenchymal chondrosarcoma because the background population of small cells in both tumors may be indistinguishable. However, the presence of osteoid in osteosarcoma is diagnostic.

Myxoid chondrosarcoma requires differentiation from other myxoid tumors of bone and soft tissue, such as *myxoma* and *myxoid liposarcoma*. Myxoid chondrosarcoma is a more cellular lesion than *myxoma* and contains acid sulfated mucopolysaccharides in the matrix. *Myxoid liposarcoma* can be excluded because it lacks the well-differentiated features of cartilage, such as lacunar formation around the chondroblasts.

Periosteal chondrosarcoma is occasionally difficult to distinguish from the *exostotic chondrosarcoma*. In such cases the continuity of the cancellous bone of osteochondroma with that of the adjacent host bone may



Figure 3-125 Histopathology of mesenchymal chondrosarcoma resembling fibrosarcoma. In areas without chondroblastic differentiation and capillaries, distinction from low-grade fibrosarcoma may be difficult (hematoxylin and eosin, original magnification $\times 25$).

indicate malignant transformation rather than a periosteal tumor. To make a distinction between periosteal chondrosarcoma and periosteal osteosarcoma is extremely difficult. In fact, some investigators (68) have previously considered these lesions to be identical. Although periosteal osteosarcoma is predominantly a chondroblastic tumor, unlike the large lowgrade cartilage present in periosteal chondrosarcoma, the chondroid element in periosteal osteosarcoma is usually high grade (16). Moreover, tumor osteoid is formed by the malignant neoplastic cells of periosteal osteosarcoma, a feature that is absent in periosteal chondrosarcoma (357) (Fig. 3-126). Periosteal chondroma may mimic periosteal chondrosarcoma and vice versa. There is overlap in the histologic appearance of both lesions. Mirra et al. (49) contended that periosteal chondrosarcomas are usually wider than they are long, in contrast to periosteal chondromas.

The radiologic and pathologic differential diagnosis of conventional chondrosarcoma, clear cell chondrosarcoma, mesenchymal chondrosarcoma, dedifferentiated chondrosarcoma, and periosteal chondrosarcoma is depicted in Figure 3-127.



Figure 3-126 Histopathology of periosteal chondrosarcoma resembling periosteal osteosarcoma. Fairly regularly appearing chondroblastic tissue borders loose connective tissue (*top*) and is supported by trabecular bone (*below*) with features of endochondral ossification, which is not part of the tumor. That reactive bone formation mimics tumor bone of osteosarcoma (hematoxylin and eosin, original magnification ×6).



Figure 3-127. A: The radiologic and pathologic differential diagnosis of conventional medullary chondrosarcoma (continued).



Figure 3-127 *Continued* **B:** The radiologic and pathologic differential diagnosis of clear cell chondrosarcoma. **C:** The radiologic and pathologic differential diagnosis of mesenchymal chondrosarcoma.





Figure 3-127 *Continued* **D:** The radiologic and pathologic differential diagnosis of dedifferentiated chondrosarcoma. **E:** The radiologic and pathologic differential diagnosis of periosteal chondrosarcoma.

REFERENCES

Enchondroma, Periosteal Chondroma, Enchondromatosis, Soft Tissue Chondroma

- Abdelwahab IF, Hermann G, Lewis MM, Klein MJ. Case report 588. Intracortical chondroma of the left femur. *Skeletal Radiol* 1990;19:59–61.
- Alho A, Skjeldal S, Pettersen EO, et al. Aneuploidy in benign tumors and nonneoplastic lesions of musculoskeletal tissues. *Cancer* 1994;73:1200–1205.
- Bansal M, Goldman AB, DiCarlo EF, McCormack R. Soft tissue chondromas: diagnosis and differential diagnosis. *Skeletal Radiol* 1993;22:309–315.
- Barton DL, Reeves RJ. Tumoral calcinosis. Report of three cases and review of the literature. Am J Roentgenol 1961;86:351–358.
- Beltran J, Chandnani V, McGhee RA Jr, Kursunoglu-Brahme S. Gadopentetate dimeglumine-enhanced MR imaging of the musculoskeletal system. *Am J Roentgenol* 1991;156:457–466.
- Boriani S, Bacchini P, Bertoni F, Campanacci M. Periosteal chondroma. A review of twenty cases. J Bone Joint Surg Am 1983; 65A:205–212.
- Bovée JV, van Roggen JF, Cleton-Jansen AM, et al. Malignant progression in multiple enchondromatosis (Ollier's disease): an autopsy-based molecular genetic study. *Hum Pathol* 2000;31: 1299–1303.
- Buddingh EP, Naumann S, Nelson M, et al. Cytogenetic findings in benign cartilaginous neoplasms. *Cancer Genet Cytogenet* 2003; 141:164–168.
- 9. Chung EB, Enzinger FM. Chondromas of soft parts. *Cancer* 1978;41:1414–1424.
- Crim JR, Mirra JM. Enchondroma protuberans. Report of a case and its distinction from chondrosarcoma and osteochondroma adjacent to an enchondroma. *Skeletal Radiol* 1990;19:431–434.
- 11. Dahlen A, Mertens F, Rydholm A, et al. Fusion, disruption, and expression of HMGA2 in bone and soft tissue chondromas. *Mod Pathol* 2003;16:1132–1140.
- Dahlin DC, Salvador AH. Cartilaginous tumors of the soft tissues of the hands and feet. *Mayo Clin Proc* 1974;49:721–726.
- Dahlin DC, Unni KK. Bone tumors. General aspects and data on 8542 cases, 4th ed. Springfield, IL: Charles C Thomas, 1986:18, 33–51, 227–259.
- Dal Cin P, Qi H, Sciot R, van den Berghe H. Involvement of chromosomes 6 and 11 in a soft tissue chondroma. *Cancer Genet Cyto*genet 1997;93:177–178.
- deSantos LA, Spjut HJ. Periosteal chondroma: a radiographic spectrum. Skeletal Radiol 1981;6:15–20.
- Fechner RE, Mills SE. *Tumors of the bones and joints*. Washington, DC: Armed Forces Institute of Pathology, 1993.
- Feldman F. Cartilaginous lesions of bones and soft tissues. CRC Crit Rev Clin Radiol Nucl Med 1974;4:477–554.
- Feldman F. Cartilaginous tumors and cartilage-forming tumorlike conditions of the bones and soft tissues. In: Ranniger K, ed. *Bone tumors.* Berlin: Springer-Verlag, 1977:83–242.
- Flach HZ, Ginai AZ, Oosterhuis JW. Maffucci syndrome: radiologic and pathologic findings. *RadioGraphics* 2001;21:1311– 1316.
- Freiberg TA, Hembree JL, Laine W. Periosteal chondroma: a review of the literature and case report. *J Foot Surg* 1986;25:54–57.
- 21. Gabos PG, Bowen JR. Epiphyseal-metaphyseal enchondromatosis. J Bone Joint Surg Am 1998;80A:782–792.
- González-Lois C, García-de-la-Torre JP, SantosBriz-Terrón, et al. Intracapsular and para-articular chondroma adjacent to large joints: report of three cases and review of the literature. *Skeletal Radiol* 2001;30:672–676.
- Goodman SB, Bell RS, Fornasier VS, et al. Ollier's disease with multiple sarcomatous transformation. *Hum Pathol* 1984;15: 91–93.
- 24. Greenfield GB, Arrington JA. Imaging of bone tumors. A multimodality approach. Philadelphia: JB Lippincott, 1995.
- Greenspan A. Tumors of cartilage origin. Orthop Clin North Am 1989;20:347–366.
- Greenspan A, Unni KK, Matthews J II. Periosteal chondroma masquerading as osteochondroma. *Can Assoc Radiol J* 1993; 44: 205–210.
- 27. Hofman S, Heeg M, Klein J-P, Krikke AP. Simultaneous occurrence of a supra- and an infratentorial glioma in a patient with

Ollier's disease: more evidence for non-mesodermal tumor predisposition in multiple enchondromatosis. *Skeletal Radiol* 1998; 27:688–691.

- Holder SF, Grana WA. Periosteal chondroma. Orthopaedics 1987; 10:1997–1998.
- 29. Hopyan S, Gokgoz N, Poon R, et al. A mutant PTH/PTHrP type I receptor in enchondromatosis. *Nat Genet* 2002;30:306–310.
- Jaffe HL. Tumors and tumorous conditions of the bones and joints. Philadelphia: Lea & Febiger, 1968.
- Keating RB, Wright PW, Staple TW. Enchondroma protuberans of the rib. *Skeletal Radiol* 1985;13:55–58.
- Kempson RL, Fletcher CD, Evans HL, et al. Cartilaginous and osseous tumors. In: Atlas of tumor pathology. Tumors of soft tissues. Washington, DC: Armed Forces Institute of Pathology, 2001.
- Kendell SD, Collins MS, Adkins MC, et al. Radiographic differentiation of enchondroma from low-grade chondrosarcoma in the fibula. *Skeletal Radiol* 2004;33:458–466.
- 34. Kosaki N, Yabe H, Anazawa U, et al. Bilateral multiple transformation of Ollier's disease. *Skeletal Radiol* 2005;34:477–484.
- Kusuzaki K, Murata H, Takashita H, et al. Usefulness of cytofluorometric DNA ploidy analysis in distinguishing benign cartilaginous tumors from chondrosarcomas. *Mod Pathol* 1999;12:863– 872.
- Lewis MM, Kenan S, Yabut SM, et al. Periosteal chondroma. A report of ten cases and review of the literature. *Clin Orthop* 1990; 256:185–192.
- Lewis RJ, Ketcham AS. Maffucci's syndrome: functional and neoplastic significance—case report and review of the literature. J Bone Joint Surg Am 1973;55A:1465–1479.
- Li C, Arger PH, Dalinka MK. Soft tissue osteochondroma. A report of three cases. *Skeletal Radiol* 1989;18:435–437.
- Lichtenstein L, Hall JE. Periosteal chondroma: a distinctive benign cartilage tumor. J Bone Joint Surg Am 1952;34:691–697.
- 40. Ligon AH, Moore SD, Parisi MA, et al. Constitutional rearrangement of the architectural factor HMGA2: a novel human phenotype including overgrowth and lipomas. *Am J Hum Genet* 2005; 76:340–348.
- Liu J, Hudkins PG, Swee RG, Unni KK. Bone sarcomas associated with Ollier's disease. *Cancer*1987;59:1376–1385.
- Ly JQ, Beall DP. A rare case of infantile Ollier's disease demonstrating bilaterally symmetric extremity involvement. *Skeletal Radiol* 2003;32:227–230.
- Mandahl N, Heim S, Arheden K, et al. Chromosomal rearrangements in chondromatous tumors. *Cancer* 1990;65:242–248.
- McKusick V. Online Mendelian inheritance in man, OMIM (TM). Johns Hopkins University, Baltimore, MD. MIM Number: #166000, updated: 1/10/2005. Available at http://www.ncbi. nlm.nih.gov/omim/ (accessed October 2005).
- 45. Mertens F, Unni KK. Enchondromatosis: Ollier disease and Maffucci syndrome. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002: 356–357.
- Milchgrub S, McMurry NK, Vuitch F, Dorfman HD. Chondrolipoangioma. A cartilage-containing benign mesenchymoma of soft tissue. *Cancer* 1990;66:2636–2641.
- Milgram JW, Dunn EJ. Para-articular chondromas and osteochondromas. A report of three cases. *Clin Orthop* 1980;148: 147– 151.
- Mills TD, Hiorns MP, Hall CM. Symmetrical enchondromatosis without vertebral involvement and with cone-shaped phalangeal epiphyses. *Skeletal Radiol* 2001;30:346–349.
- 49. Mirra JM, Picci P, Gold RH. Bone tumors: clinical, radiologic and pathologic correlations. Philadelphia: Lea & Febiger, 1989.
- Mitchell ML, Ackerman LV. Case report 405. Ollier disease (enchondromatosis). *Skeletal Radiol* 1987;16:61–66.
- Moser RP, Gilkey FW, Madewell JE. Enchondroma. In: Moser RP, ed. Cartilaginous tumors of the skeleton. AFIP atlas of radiologic-pathologic correlation, vol 2. Philadelphia: Hanley and Belfus, 1990: 8–34.
- 52. Mulder JD, Schütte HE, Kroon HM, Taconis WK. Radiologic atlas of bone tumors. Amsterdam: Elsevier, 1993.
- Murphey MD, Flemming DJ, Boyea SR, et al. Enchondroma versus chondrosarcoma in the appendicular skeleton: differentiating features. *RadioGraphics* 1998;18:1213–1237.

- Nayler S, Heim S. Soft tissue chondroma. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:180–181.
- Nojima T, Unni KK, McLeod RA, Pritchard DJ. Periosteal chondroma and periosteal chondrosarcoma. *Am J Surg Pathol* 1985;9: 666–677.
- Norman A, Steiner GC. Radiographic and morphological features of cyst formation in idiopathic bone infarction. *Radiology* 1983;146:335–338.
- Oestreich AE, Mitchell CS, Akeson JW. Both Trevor and Ollier disease limited to one upper extremity. *Skeletal Radiol* 2002;31: 230–234.
- 59. Ollier L. De la dyschondroplasie. Bull Soc Chir Lyon 1900;3:2-27.
- Ozisik YY, Meloni AM, Spanier SS, et al. Deletion 1p in a lowgrade chondrosarcoma in a patient with Ollier disease. *Cancer Genet Cytogenet* 1998;105:128–133.
- 61. Palmer PES. Tumoral calcinosis. Br J Radiol 1966;39:518-525.
- Ragsdale BD, Sweet DE, Vinh TN. Radiology as gross pathology in evaluating chondroid tumors. *Hum Pathol* 1989;20: 930–951.
- Ramnath RR, Rosenthal DI, Cates J, et al. Intracortical chondroma simulating osteoid osteoma treated by radiofrequency. *Skeletal Radiol* 2002;31:597–602.
- 64. Resnick CS, Levine AM, Aisner SC, et al. Case report 522. Concurrent adjacent osteochondroma and enchondroma. *Skeletal Radiol* 1989;18:66–69.
- Robinson P, White LM, Sundaram M, et al. Periosteal chondroid tumors: radiologic evaluation with pathologic correlation. *Am J Roentgenol* 2001;177:1183–1188.
- 66. Rozeman LB, Sangiorgi L, Briaire-de Bruijn IH, et al. Enchondromatosis (Ollier disease, Maffucci syndrome) is not caused by the PTHR1 mutation p.R150C. *Hum Mutat* 2004;24:466–473.
- Rudman DP, Damron TA, Vermont A, Mathur S. Intracortical chondroma. *Skeletal Radiol* 1998;27:581–583.
- Schajowicz F. Cartilage-forming tumors. In: Schajowicz F, ed. Tumors and tumorlike conditions of bone. New York: Springer-Verlag, 1994:141–256.
- Schajowicz F, Ackerman LV, Sissons HA. *Histological typing of bone tumours*. International Histological Classification of Tumors, No. 6. Geneva: World Health Organization, 1972.
- Schajowicz F, McGuire M. Diagnostic difficulties in skeletal pathology. *Clin Orthop* 1989;240:281–308.
- 71. Shadan FF, Mascarello JT, Newbury RO, et al. Supernumerary ring chromosomes derived from the long arm of chromosome 12 as the primary cytogenetic anomaly in a rare soft tissue chondroma. *Cancer Genet Cytogenet* 2000;118:144–147.
- Shapiro F. Ollier's disease. An assessment of angular deformity, shortening, and pathological fracture in twenty-one patients. J Bone Joint Surg Am 1982;64:95–103.
- Sun TC, Swee RG, Shives TC, Unni KK. Chondrosarcoma in Maffucci's syndrome. J Bone J Surg Am 1985;67:1214–1219.
- Sundaram M, McLeod RA. MR imaging of tumor and tumor-like lesions of bone and soft tissue. *Am J Roentgenol* 1990; 155:817–824.
- Unger EC, Kessler HB, Kowalyshyn MJ, et al. MR imaging of Maffucci syndrome. Am J Roentgenol 1988;150:351–353.
- Varma DGK, Kumar R, Carrasco CH, et al. MR imaging of periosteal chondroma. *J Comput Assist Tomogr* 1991;15:1008–1010.
- Weiss SW, Goldblum JR. Cartilaginous soft tissue tumors. In: Enzinger and Weiss's soft tissue tumors, 4th ed. Philadelphia: Mosby-Harcourt, 2001:1361–1388.
- Zlatkin MB, Lander PH, Begin LR, Hadjipavlou A. Soft-tissue chondromas. Am J Roentgenol 1985;144:1263–1267.

Osteochondroma, Multiple Hereditary Osteochondromatosis

- Borges AM, Huvos AG, Smith J. Bursa formation and synovial chondrometaplasia associated with osteochondromas. *Am J Clin Pathol* 1981;75:648–653.
- Bovée JVMG, Hogendorn PCW. Multiple osteochondromas. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology G genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:360–362.

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- Carroll KL, Yandow SM, Ward K, Carey JC. Clinical correlation to genetic variations of hereditary multiple exostoses. *J Pediatr Orthop* 1999;19:785–791.
- Choi JH, Gu MJ, Kim MJ, et al. Fibrosarcoma in bizarre parosteal osteochondromatous proliferation. *Skeletal Radiol* 2001;30:44–47.
- Cohen EK, Kressel HY, Frank TS, et al. Hyaline cartilage-origin bone and soft-tissue neoplasms: MR appearance and histologic correlation. *Radiology* 1988;167:477–481.
- Davids JR, Glancy GL, Eilert RE. Fracture through the stalk of pedunculated osteochondromas. A report of three cases. *Clin Orthop Rel Res* 1991;271:258–264.
- 85. Duncan G, McCormick C, Tufaro F. The link between heparan sulfate and hereditary bone disease: finding a function for the EXT family of putative tumor suppressor proteins. *J Clin Invest* 2001;108:511–516.
- El-Khoury GY, Bassett GS. Symptomatic bursa formation with osteochondromas. Am J Roentgenol 1979;133:895–898.
- Epstein DA, Levin EJ. Bone scintigraphy in hereditary multiple exostoses. *Am J Roentgenol* 1978;130:331–333.
- Fairbank TJ. Dysplasia epiphysealis hemimelica (tarso-epiphyseal aclasis). J Bone Joint Surg Br 1956;38:237–257.
- Francannet C, Cohen-Tanugi A, Le Merrer M, et al. Genotypephenotype correlation in hereditary multiple exostoses. J Med Genet 2001;38:430–434.
- Garrison RC, Unni KK, McLeod RA, et al. Chondrosarcoma arising in osteochondroma. *Cancer* 1982;49:1890–1897.
- Geirnaerdt MJA, Bloem JL, Eulderink F, et al. Cartilaginous tumors: correlation of gadolinium-enhanced MR imaging and histopathologic findings. *Radiology* 1993;186:813–817.
- Griffiths HJ, Thompson RC Jr, Galloway HR, et al. Bursitis in association with solitary osteochondromas presenting as mass lesions. *Skeletal Radiol* 1991;20:513–516.
- 93. Hensinger RN, Cowell HR, Ramsey PL, Leopold RG. Familial dysplasia epiphysealis hemimelica associated with chondromas and osteochondromas. Report of a kindred with variable presentations. J Bone Joint Surg Am 1974;56:1513–1516.
- Hudson TM, Chew FS, Manaster BJ. Scintigraphy of benign exostoses and exostotic chondrosarcomas. *Am J Roentgenol* 1983; 140:581–586.
- Hudson TM, Spriengfield DS, Spanier SS, et al. Benign exostoses and exostotic chondrosarcomas: evaluation of cartilage thickness by CT. *Radiology* 1984;152:595–599.
- 96. Jaffe HL. Tumors and tumorous conditions of the bones and joints. Philadelphia: Lea & Febiger, 1968.
- Karasick D, Schweitzer ME, Eschelman DJ. Symptomatic osteochondromas: imaging features. Am J Roentgenol 1997;68:1507– 1512.
- Kettelkamp DB, Campbell CJ, Bonfiglio M. Dysplasia epiphysealis hemimelica. A report of fifteen cases and a review of the literature. *J Bone Joint Surg Am* 1966;48:746–766.
- Khurana J, Abdul-Karim F, Bovée J. Osteochondroma. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:234–236.
- Lang IM, Azuz EM. MRI appearance of dysplasia epiphysealis hemimelica of the knee. *Skeletal Radiol* 1997;26:226–229.
- Lange RH, Lange TA, Rao BK. Correlative radiographic, scintigraphic, and histological evaluation of exostoses. J Bone Joint Surg Am 1984;66:1454–1459.
- Lee JK, Yao L, Wirth CR. MR imaging of solitary osteochondromas: report of eight cases. *Am J Roentgenol* 1987;149:557–560.
- Legeai-Mallet L, Munnich A, Maroteaux, P, Le Merrer M. Incomplete penetrance and expressivity skewing in hereditary multiple exostoses. *Clin Genet* 1997;52:12–16.
- 104. Le Merrer M, Legeai-Mallet L, Jeannin PM, et al. A gene for hereditary multiple exostoses maps to chromosome 19 p. *Hum Mol Genet* 1994;3:717–722.
- 105. Malghem J, Vande Berg B, Noël H, Maldague B. Benign osteochondromas and exostotic chondrosarcomas: evaluation of cartilage cap thickness by ultrasound. *Skeletal Radiol* 1992;21:33–37.
- 106. Meneses MF, Unni KK, Swee RG. Bizarre parosteal osteochondromatous proliferation of bone (Nora's lesion). Am J Surg Pathol 1993;17:691–697.
- Murphey MD, Choi JJ, Kransdorf MJ, et al. Imaging of osteochondroma: variants and complications with radiologic-pathologic correlation. *RadioGraphics* 2000;20:1407–1434.

- 108. Nidecker A, Remagen W, Elke M. Korrelation radiologischer und pathologischer Befunde bei Tumoren und tumorähnlichen Läsionen der Hand. *Radiologe* 1982;22:222–229.
- Nora FE, Dahlin DC, Beabout JW. Bizarre parosteal osteochondromatous proliferation of the hands and feet. *Am J Surg Pathol* 1983;7:245–250.
- Norman A, Sissons HA. Radiographic hallmarks of peripheral chondrosarcoma. *Radiology* 1984;151:589–596.
- Peterson HA. Multiple hereditary osteochondromata. Clin Orthop 1989;239:222–230.
- 112. Porter DE, Lonie L, Fraser M, Dobson-Stone C, et al. Severity of disease and risk of malignant change in hereditary multiple exostoses. A genotype-phenotype study. *J Bone Joint Surg Br* 2004; 86:1041–1046.
- Reston SC, Savva N, Richards RH. Spontaneous resolution of solitary osteochondroma in the young adult. *Skeletal Radiol* 2004;33:303–305.
- Schmale GA, Conrad EU 3rd, Raskind WH. The natural history of hereditary multiple exostoses. J Bone Joint Surg Am 1994;76: 986–992.
- 115. Sundaram M, Wang L, Rotman M, et al. Florid reactive periostitis and bizarre parosteal osteochondromatous proliferation: pre-biopsy imaging evolution, treatment and outcome. *Skeletal Radiol* 2001;30:192–198.
- Uri DS, Dalinka MK, Kneeland JB. Muscle impingement: MR imaging of a painful complication of osteochondromas. *Skeletal Radiol* 1996;25:689–692.
- 117. Wu YQ, Heutink P, de Vries BB, et al. Assignment of a second locus for multiple exostoses to the pericentromeric region of chromosome 11. *Hum Mol Genet* 1994;3:167–171.

Chondroblastoma

- Aigner T, Loos S, Inwards C, et al. Chondroblastoma is an osteoid-forming, but not cartilage-forming neoplasm. J Pathol 1999;189:463–469.
- 118a. Azorín D, González-Mediero I, Colmenero I, et al. Diaphyseal chondroblastoma in a long bone: first report. *Skeletal Radiol* 2006;35:49–52.
- Azouz EM, Greenspan A, Marton D. CT evaluation of primary epiphyseal bone abscesses. *Skeletal Radiol* 1993;22:17–23.
- Björnsson J, Unni KK, Dahlin DC, et al. Clear-cell chondrosarcoma of bone: observation in 47 cases. *Am J Surg Pathol* 1984;8: 223–230.
- Bloem JL, Mulder JD. Chondroblastoma: a clinical and radiological study of 104 cases. *Skeletal Radiol* 1985;14:1–9.
- Bogumill GP, Schultz MA, Johnson LC. Giant-cell tumor—a metaphyseal lesion. (Scientific exhibit). J Bone Joint Surg 1972; 54A:1558.
- 123. Braunstein E, Martel W, Weatherbee L. Periosteal bone apposition in chondroblastoma. *Skeletal Radiol* 1979;4:34–36.
- 124. Bridge JA, Bhatia PS, Anderson JR, Neff JR. Biologic and clinical significance of cytogenetic and molecular cytogenetic abnormalities in benign and malignant cartilaginous lesions. *Cancer Genet Cytogenet* 1993;69:79–90.
- 125. Brower AC, Moser RP, Gilkey FW, Kransdorf MJ. Chondroblastoma. In: Moser RP, ed. Cartilaginous tumors of the skeleton. AFIP atlas of radiologic-pathologic correlation, vol 2. Philadelphia: Hanley and Belfus, 1990:74–113.
- 126. Brower AC, Moser RP, Kransdorf MJ. The frequency and diagnostic significance of periostitis in chondroblastoma. *Am J Roentgenol* 1990;154:309–314.
- 127. Codman EA. Epiphyseal chondromatous giant cell tumors of the upper end of the humerus. Surg Gynecol Obstet 1931;52: 543–548.
- Cohen EK, Kressel HY, Frank TS, et al. Hyaline cartilage-origin bone and soft-tissue neoplasms: MR appearance and histologic correlation. *Radiology* 1988;167:477–481.
- Dahlin DC, Ivins JC. Benign chondroblastoma: a study of 125 cases. *Cancer* 1972;30:401–413.
- Davila JA, Amrami KK, Sundaram M, et al. Chondroblastoma of the hands and feet. *Skeletal Radiol* 2004;33:582–587.
- Edel G, Ueda Y, Nakanishi J, et al. Chondroblastoma of bone. A clinical, radiological, light and immunohistochemical study. Virchows Arch A Pathol Anat Histopathol 1992;421:355– 366.

- 132. Erickson JK, Rosenthal DI, Zaleske DJ, et al. Primary treatment of chondroblastoma with percutaneous radiofrequency heat ablation: report of three cases. *Radiology* 2001;221: 463–468.
- Gao Z, Kahn LB. The application of immunohistochemistry in the diagnosis of bone tumors and tumor-like lesions. *Skeletal Radiol* 2005;34:755–770.
- 134. Gardner DJ, Azouz EM. Solitary lucent epiphyseal lesions in children. *Skeletal Radiol* 1988;17:497–504.
- Giudici MA, Moser RP Jr, Kransdorf MJ. Cartilaginous bone tumors. *Radiol Clin North Am* 1993;31:237–259.
- Gohel VK, Dalinka MK, Edeiken J. Ischemic necrosis of the femoral head simulating chondroblastoma. *Radiology* 1973;107: 545–546.
- 137. Green P, Wittaker RP. Benign chondroblastoma. Case report with pulmonary metastasis. *J Bone Joint Surg Am* 1975;57:418–420.
- 138. Greenspan A, Klein MJ. Radiology and pathology of bone tumors. In: Lewis MM, ed. *Musculoskeletal oncology. A multidisciplinary approach.* Philadelphia: WB Saunders, 1992:13–72.
- Hayes CW, Conway WF, Sundaram M. Misleading aggressive MR imaging: appearance of some benign musculoskeletal lesions. *RadioGraphics* 1992;12:1119–1134.
- 140. Helms C. Pseudocyst of the humerus. Am J Roentgenol 1979;131: 287–292.
- Hudson TM, Hawkins IF Jr. Radiological evaluation of chondroblastoma. *Radiology* 1981;139:1–10.
- 142. Huvos AG. Chondroblastoma and clear cell chondrosarcoma. In: Huvos AG, ed. Bone tumors. Diagnosis, treatment and prognosis, 2nd ed. Philadelphia, WB Saunders, 1991:295–318.
- 143. Huvos AG, Higinbotham NL, Marcove RC, O Leary P. Aggressive chondroblastoma: Review of the literature on aggressive behavior and metastases with a report of one new case. *Clin Orthop* 1977;126:266–272.
- Ilaslan H, Sundaram M, Unni KK. Vertebral chondroblastoma. Skeletal Radiol 2003;32:66–71.
- Ishida T, Goto T, Motoi N, Mukai K. Intracortical chondroblastoma mimicking intra-articular osteoid osteoma. *Skeletal Radiol* 2002;31:603–607.
- 146. Jaffe HL, Lichtenstein L. Benign chondroblastoma of bone: Reinterpretation of so-called calcifying or chondromatous giant cell tumor. *Am J Pathol* 1942;18:969–991.
- 147. Kaim AH, Hügli R, Bonél HM, Jundt G. Chondroblastoma and clear cell chondrosarcoma: radiological and MRI characteristics with histopathological correlation. *Skeletal Radiol* 2002;31: 88–95.
- 148. Kaufman RA, Wakely PE, Greenfield DJ. Case report 224. Giant cell tumor of ossification center of the distal end of the fibula, growing into the metaphysis. *Skeletal Radiol* 1983;9:218–222.
- 149. Keats TE. Normal Roentgen variants that may simulate disease. Chicago: Year Book Medical, 1980.
- Kenney PJ, Gilula LA, Murphy WA. The use of computed tomography to distinguish osteochondroma and chondrosarcoma. *Radiology* 1981;139:129–137.
- 151. Kilpatrick S, Parisien M, Bridge J. Chondroblastoma. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:241–242.
- 152. Kobayashi Y, Murakami R, Toba M, et al. Chondroblastoma of the temporal bone. *Skeletal Radiol* 2001;30:714–718.
- 153. Kricun ME. Imaging of bone tumors. Philadelphia: WB Saunders, 1993.
- 154. Kricun ME, Kricun R, Haskin ME. Chondroblastoma of the calcaneus: radiographic features with emphasis on location. Am J Roentgenol 1977;128:613–616.
- 155. Kroon HM, Bloem JL, Holscher HC, et al. MR imaging of edema accompanying benign and malignant bone tumors. *Skeletal Radiol* 1994;23:261–269.
- Kurt AM, Unni KK, Sim FH, McLeod RA. Chondroblastoma of bone. *Hum Pathol* 1989;20:965–976.
- 157. Leung LYJ, Shu SJ, Chan MK, Chan CHS. Chondroblastoma of the lumbar vertebra. *Skeletal Radiol* 2001;30:710–713.
- Ly JQ, LaGatta LM, Beall DP. Calcaneal chondroblastoma with secondary aneurysmal bone cyst. *Am J Roentgenol* 2004;182: 130.
- Marsh BW, Bonfiglio M, Brady LP, Enneking WF. Benign osteoblastoma: range of manifestations. *J Bone Joint Surg Am* 1975; 57:1–9.

- Masui F, Ushigome S, Fujii K. Giant cell tumor of bone: an immunohistochemical comparative study. *Pathol Int* 1998;48: 355–361.
- McLeod RA, Beabout JW. The roentgenographic features of chondroblastoma. Am J Roentgenol 1973;118:464–471.
- Mirra JM, Ulich TR, Eckardt JJ, Bhuta S. "Aggressive" chondroblastoma. Light and ultramicroscopic findings after en bloc resection. *Clin Orthop* 1983;178:276–284.
- Monda L, Wick MR. S-100 protein immunostaining in the differential diagnosis of chondroblastoma. *Hum Pathol* 1985;16: 287–293.
- 164. Moser RP, Brockmole DM, Vinh TN, et al. Chondroblastoma of the patella. *Skeletal Radiol* 1988;17:413–419.
- 165. Ostrowski ML, Johnson ME, Truong LD, et al. Malignant chondroblastoma presenting as a recurrent pelvic tumor with DNA aneuploidy and p53 mutation as supportive evidence of malignancy. *Skeletal Radiol* 1999;28:644–650.
- 165a.Ozkoc G, Gonlusen G, Ozalay M, et al. Giant chondroblastoma of the scapula with pulmonary metastases. *Skeletal Radiol* 2006; 35:42–48.
- 166. Peh WCG, Shek TWH, Ip WY. Metadiaphyseal chondroblastoma of the thumb. *Skeletal Radiol* 2000;29:176–180.
- Picci P, Manfrini M, Zucchi V, et al. Giant-cell tumor of bone in skeletally immature patients. J Bone Joint Surg Am 1983;65:486– 490.
- Plum GE, Pugh DG. Roentgenologic aspects of benign chondroblastoma of bone. *Am J Roentgenol* 1958;79:584–591.
- Pösl M, Werner M, Amling M, et al. Malignant transformation of chondroblastoma. *Histopathology* 1996;29:477–480.
- Quint LE, Gross BH, Glazer GM, et al. CT evaluation of chondroblastoma. J Comput Assist Tomogr 1984;8:907–910.
- Raymond AK, Raymond PG, Edeiken J. Case report 531. Epiphyseal osteoblastoma distal end of femur. *Skeletal Radiol* 1989; 18:143–146.
- 172. Resnick D, Cone RO III. The nature of humeral pseudocyst. *Radiology* 1984;150:27–28.
- 173. Romeo S, Bovée JV, Jadnanansing NA, et al. Expression of cartilage growth plate signaling molecules in chondroblastoma. *J Pathol* 2004;202:113–120.
- 174. Schajowicz F, Gallardo H. Epiphyseal chondroblastoma of bone: a clinicopathological study of sixty-nine cases. *J Bone Joint Surg Br* 1970;52:205–226.
- Schütte HE, Taconis WK. Giant cell tumor in children and adolescents. Skeletal Radiol 1993;22:173–176.
- Sissons HA, Murray RO, Kemp HBS. Orthopaedic diagnosis. Berlin: Springer-Verlag; 1984.
- 177. Springfield DS, Capanna R, Gherlinzoni F, et al. Chondroblastoma: a review of seventy cases. J Bone Joint Surg Am 1985;67: 748–755.
- Steiner GC. Benign cartilage tumors. In: Taveras JM, Ferrucci JT, eds. *Radiology: diagnosis, imaging, intervention*, vol 5. Philadelphia: JB Lippincott, 1986:chap 78, 1–6.
- 179. Swarts SJ, Neff JR, Johansson SL, et al. Significance of abnormalities of chromosomes 5 and 8 in chondroblastoma. *Clin Orthop Relat Res* 1998;349:189–193.
- Weatherall PT, Maale GE, Mendelsohn DB, et al. Chondroblastoma: classic and confusing appearance at MR imaging. *Radiol*ogy 1994;190:467–474.
- Yamamura S, Sato K, Sugiura H, Iwata H. Inflammatory reaction in chondroblastoma. *Skeletal Radiol* 1996;25:371–376.

Chondromyxoid Fibroma

- Beggs IG, Stoker DJ. Chondromyxoid fibroma of bone. Clin Radiol 1982;33:671–679.
- Bleiweiss IJ, Klein MJ. Chondromyxoid fibroma: report of six cases with immunohistochemical studies. *Mod Pathol* 1990;3: 664–666.
- 184. Bruder E, Zanetti M, Boos N, von Hochstetter AR. Chondromyxoid fibroma of two thoracic vertebrae. *Skeletal Radiol* 1999;28:286–289.
- Dahlin DC. Chondromyxoid fibroma of bone, with emphasis on its morphological relationship to benign chondroblastoma. *Cancer* 1956;9:195–203.
- Feldman F, Hecht HL, Johnston AD. Chondromyxoid fibroma of bone. *Radiology* 1970;94:249–260.

- 187. Feuvret L, Noel G, Calugaru V, et al. Chondromyxoid fibroma of the skull base: differential diagnosis and radiotherapy: two case reports and a review of the literature. *Acta Oncol* 2005;44: 545–553.
- Fujiwara S, Nakamura I, Goto T, et al. Intracortical chondromyxoid fibroma of humerus. *Skeletal Radiol* 2003;32:156–160.
- Hau MA, Fox EJ, Rosenberg AE, Mankin HJ. Chondromyxoid fibroma of the metacarpal. *Skeletal Radiol* 2001;30:719–721.
- 190. Jaffe HL, Lichtenstein L. Chondromyxoid fibroma of bone: a distinctive benign tumor likely to be mistaken especially for chondrosarcoma. *Arch Pathol* 1948;45:541–551.
- 191. Keel SB, Bhan AK, Liebsch NJ, Rosenberg AE. Chondromyxoid fibroma of the skull base: a tumor which may be confused with chordoma and chondrosarcoma. A report of three cases and review of the literature. *Am J Surg Pathol* 1997;21:577–582.
- 192. Kikuchi F, Dorfman HD, Kane PB. Recurrent chondromyxoid fibroma of the thoracic spine 30 years after primary excision: case report and review of the literature. *Int J Surg Pathol* 2001; 9:323–329.
- 193. Macdonald D, Fornasier V, Holtby R. Chondromyxoid fibroma of the acromium with soft tissue extension. *Skeletal Radiol* 2000; 29:168–170.
- 194. Marin C, Gallego C, Manjón P, Martinez-Tello FJ. Juxtacortical chondromyxoid fibroma: imaging findings in three cases and a review of the literature. *Skeletal Radiol* 1997;26:642–649.
- Mitchell ML, Sartoris DJ, Resnick D. Case report 713. Chondromyxoid fibroma of the third metatarsal. *Skeletal Radiol* 1992; 21:252–255.
- 196. Murphy NB, Price CHG. The radiological aspects of chondromyxoid fibroma of bone. *Clin Radiol* 1971;22:261–269.
- 197. Nielsen GP, Keel SB, Dickersin GR, et al. Chondromyxoid fibroma: a tumor showing myofibroblastic, myochondroblastic, and chondrocytic differentiation. *Mod Pathol* 1999;12:514– 517.
- 198. O'Connor PJ, Gibbon WW, Hardy G, Butt WP. Chondromyxoid fibroma of the foot. *Skeletal Radiol* 1996;25:143–148.
- 199. Ribalta T, Ro JY, Carrasco CH, et al. Case report 638. Chondromyxoid fibroma of a sesamoid bone. *Skeletal Radiol* 1990;19: 549–551.
- 200. Romeo S, Bovée JV, Grogan SP, et al. Chondromyxoid fibroma resembles in vitro chondrogenesis, but differs in expression of signalling molecules. *J Pathol* 2005;206:135–142.
- Schajowicz F. Chondromyxoid fibroma: report of three cases with predominant cortical involvement. *Radiology* 1987;164:783–786.
- 202. Schajowicz F, Cabrini RL. Histochemical studies on glycogen in normal ossification and calcification. J Bone Joint Surg Am 1958; 40:1081–1092.
- 203. Schajowicz F, Gallardo H. Chondromyxoid fibroma (fibromyxoid chondroma) of bone. J Bone Joint Surg Br 1971;53:198–216.
- 204. Soder S, Inwards C, Muller S, et al. Cell biology and matrix biochemistry of chondromyxoid fibroma. Am J Clin Pathol 2001; 116:271–277.
- 205. Ushigome S, Takakuwa T, Shinagawa T, et al. Chondromyxoid fibroma of bone. An electron microscopic observation. *Acta Pathol Jpn* 1982;32:113–122.
- 206. White PG, Saunders L, Orr W, Friedman L. Chondromyxoid fibroma. Skeletal Radiol 1996;25:79–81.
- 207. Wilson AJ, Kyriakos M, Ackerman LV. Chondromyxoid fibroma: Radiographic appearance in 38 cases and in a review of the literature. *Radiology* 1991;179:513–518. Also Erratum. *Radiology* 1991;180:586.
- Yamaguchi T, Dorfman HD. Radiographic and histologic patterns of calcification in chondromyxoid fibroma. *Skeletal Radiol* 1998;27:559–564.
- Zillmer DA, Dorfman HD. Chondromyxoid fibroma of bone: thirty-six cases with clinicopathologic correlation. *Hum Pathol* 1989;20:952–964.

Chondrosarcomas

- Abello R, Lomena F, Garcia A, et al. Unusual metastatic chondrosarcoma detected with bone scintigraphy. *Eur J Nucl Med* 1986;12:306–308.
- 211. Aigner T. Towards a new understanding and classification of chondrogenic neoplasias of the skeleton—biochemistry and cell biology of chondrosarcoma and its variants. *Virchows Arch* 2002;441:219–230.

- Aigner T, Loos S, Muller S, et al. Cell differentiation and matrix gene expression in mesenchymal chondrosarcomas. *Am J Pathol* 2000;156:1327–1335.
- Aigner T, Oliveira AM, Nascimento AG. Extraskeletal myxoid chondrosarcomas do not show a chondrocytic phenotype. *Mod Pathol* 2004;17:214–221.
- 214. Aisen AM, Martel W, Braunstein EM, et al. MRI and CT evaluation of primary bone and soft-tissue tumors. *Am J Roentgenol* 1986;146:749–756.
- 215. Amir D, Amir G, Mogle P, Pogrund H. Extraskeletal soft tissue chondrosarcoma. Case report and review of the literature. *Clin Orthop* 1985;198:219–223.
- Angervall L, Enerback L, Knutson H. Chondrosarcoma of soft tissue origin. *Cancer* 1973;32:507–513.
- 217. Antonescu CR, Argani P, Erlandson RA, et al. Skeletal and extraskeletal myxoid chondrosarcoma: a comparative clinicopathologic, ultrastructural, and molecular study. *Cancer* 1998; 83:1504–1521.
- 218. Aoki JA, Sone S, Fujioka F, et al. MR of enchondroma and chondrosarcoma: rings and arcs of Gd-DTPA enhancement. *J Comput Assist Tomogr* 1991;15:1011–1016.
- 219. Aprin H, Riseborough EJ, Hall JE. Chondrosarcoma in children and adolescents. *Clin Orthop Rel Res* 1982;166:226–232.
- 220. Bagley L, Kneeland JB, Dalinka MK, et al. Unusual behavior of clear cell chondrosarcoma. *Skeletal Radiol* 1993;22:279–282.
- 221. Bertoni F, Bacchini P, Hogendoorn PC. Chondrosarcoma. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:247–251.
- 222. Bertoni F, Boriani S, Laus M, Campanacci M. Periosteal chondrosarcoma and periosteal osteosarcoma. Two distinct entities. *J Bone Joint Surg Br* 1982;64:370–376.
- Bertoni F, Picci P, Bacchini P, et al. Mesenchymal chondrosarcoma of bone and soft tissues. *Cancer* 1983;52:533–541.
- 224. Bertoni F, Present D, Bacchini P, et al. Dedifferentiated peripheral chondrosarcomas. A report of seven cases. *Cancer* 1989; 63:2054–2059.
- 225. Bertoni F, Present DA, Enneking WF. Staging of bone tumors. In: Unni KK, ed. *Bone tumors*. New York: Churchill Livingstone, 1988:47–83.
- Bertoni F, Unni KK, Beabout JW, Sim FH. Chondrosarcomas of the synovium. *Cancer* 1991;67:155–162.
- 227. Berquist TH. Magnetic resonance imaging of primary skeletal neoplasms. *Radiol Clin North Am* 1993;31:411–424.
- Bjornsson J, Unni KK, Dahlin DC, et al. Clear cell chondrosarcoma of bone. Observations in 47 cases. *Am J Surg Pathol* 1984;8: 223–230.
- Bloem JL, Bluemm RG, Taminiau AH, et al. Magnetic resonance imaging of primary malignant bone tumors. *RadioGraphics* 1987;7:425–445.
- Bohndorf K, Reiser M, Lochner B, et al. Magnetic resonance imaging of primary tumors and tumor-like lesions of bone. *Skeletal Radiol* 1986;15:511–517.
- 231. Bosse A, Ueda Y, Wuisman P, et al. Histogenesis of clear cell chondrosarcoma. An immunohistochemical study with osteonectin, a non-collagenous structure protein. J Cancer Res Clin Oncol 1991;117:43–49.
- 232. Bovée JVMG, Cleton-Jansen AM, Rosenberg C, et al. Molecular genetic characterization of both components of a dedifferentiated chondrosarcoma, with implications for its histogenesis. *J Pathol* 1999;189:454–462.
- 233. Brien EW, Mirra JM, Ippolito V, Vaughn L. Clear-cell chondrosarcoma with elevated alkaline phosphatase, mistaken for osteosarcoma on biopsy. *Skeletal Radiol* 1996;25:770–774.
- 234. Brien EW, Mirra JM, Herr R. Benign and malignant cartilage tumors of bone and joints: Their anatomic and theoretical basis with an emphasis on radiology, pathology, and clinical biology. *Skeletal Radiol* 1997;26:325–353.
- Cannon CP, Nelson SD, Seeger LL, Eckardt JJ. Clear cell chondrosarcoma mimicking chondroblastoma in a skeletally immature patient. *Skeletal Radiol* 2002;31:369–372.
- Capanna R, Bertoni F, Bettelli G, et al. Dedifferentiated chondrosarcoma. J Bone Joint Surg Am 1988;70:60–69.
- 237. Cawte TG, Steiner GC, Beltran J, Dorfman HD. Chondrosarcoma of the short tubular bones of the hands and feet. *Skeletal Radiol* 1998;27:625–632.

- Chan YF, Yeung SH, Chow TC, Ma L. Clear cell chondrosarcoma: case report and ultrastructural study. *Pathology* 1989;21:134–137.
- Chang CC, Greenspan A, Gershwin ME. Osteonecrosis: current perspectives on pathogenesis and treatment. *Semin Arthritis Rheum* 1993;23:47–69.
- 240. Collins MS, Koyama T, Swee RG, Inwards CY. Clear cell chondrosarcoma: radiographic, computed tomographic, and magnetic resonance findings in 34 patients with pathologic correlation. *Skeletal Radiol* 2003;32:687–694.
- 241. Crim JR, Seeger LL. Diagnosis of low-grade chondrosarcoma. *Radiology* 1993;189:503–504.
- 242. Crim JR, Seeger LL, Yao L, et al. Diagnosis of soft-tissue masses with MR imaging: can benign masses be differentiated from malignant ones? *Radiology* 1992;185:581–586.
- 243. Dahlin DC, Beabout JW. Dedifferentiation of low-grade chondrosarcomas. *Cancer* 1971;28:461–466.
- 244. De Beuckeleer LHL, De Schepper AMA, Ramon F. Magnetic resonance imaging of cartilaginous tumors: is it useful or necessary? *Skeletal Radiol* 1996;25:137–141.
- 245. Dickey ID, Rose PS, Fuchs B, et al. Dedifferentiated chondrosarcoma: the role of chemotherapy with updated outcomes. *J Bone Joint Surg Am* 2004;86:2412–2418.
- 246. Domanski HA, Carlen B, Mertens F, Akerman M. Extraskeletal myxoid chondrosarcoma with neuroendocrine differentiation: a case report with fine-needle aspiration biopsy, histopathology, electron microscopy, and cytogenetics. *Ultrastruct Pathol* 2003; 27:363–368.
- 247. Ehman RL, Berquist TH, McLeod RA. MR imaging of the musculoskeletal system: a 5-year appraisal. *Radiology* 1988;166: 313–320.
- 248. Ehrlich M. DNA methylation in cancer: too much, but also too little. *Oncogene* 2002;21:5400–5413.
- Enzinger FM, Shiraki M. Extraskeletal myxoid chondrosarcoma: an analysis of 34 cases. *Hum Pathol* 1972;3:421–435.
- Evans HL, Ayala AG, Romsdahl MM. Prognostic factors in chondrosarcoma of bone: a clinicopathologic analysis with emphasis on histologic grading. *Cancer* 1977;40:818–831.
- 251. Feldman F, van Heertum R, Saxena C, Parisien M. 18FDG-PET applications for cartilage neoplasms. *Skeletal Radiol* 2005;34: 367–374.
- Fletcher CD, Powell G, McKee PH. Extraskeletal myxoid chondrosarcoma: a histochemical and immunohistochemical study. *Histopathology* 1986;10:489–499.
- 253. Fobben ES, Dalinka MK, Schiebler ML, et al. The MRI appearance at 1.5 tesla of cartilaginous tumors involving the epiphysis. *Skeletal Radiol* 1987;16:647–651.
- 254. Frassica FJ, Unni KK, Beabout JW, Sim FH. Dedifferentiated chondrosarcoma. A report of the clinicopathological features and treatment of seventy-eight cases. *J Bone Joint Surg Am* 1986;68:1197–1205.
- 255. Fu YS, Kay S. A comparative ultrastructural study of mesenchymal chondrosarcoma and myxoid chondrosarcoma. *Cancer* 1974;33:1531–1542.
- 256. Geirnaerdt MJA, Hermans J, Bloem JL, et al. Usefulness of radiology in differentiating enchondroma from central grade I chondrosarcoma. *Am J Roentgenol* 1997;169:1097–1104.
- Geirnaerdt MJA, Hogendoorn PCW, Bloem JL, et al. Cartilaginous tumors: fast contrast-enhanced MR imaging. *Radiology* 2000;214:539–546.
- 257a. Gitelis S, Block JA, Inerot SE. Clonal analysis of human chondrosarcoma. The 35th Annual Meeting, Orthopedic Research Society. *Orthop Trans* 1989;13:443.
- 258. Goh YW, Spagnolo DV, Platten M, et al. Extraskeletal myxoid chondrosarcoma: a light microscopic, immunohistochemical, ultrastructural and immuno-ultrastructural study indicating neuroendocrine differentiation. *Histopathology* 2001;39:514–524.
- Goldman RL, Lichtenstein L. Synovial chondrosarcoma. Cancer 1964;17:1233–1240.
- Granter SR, Renshaw AA, Fletcher CD, et al. CD99 reactivity in mesenchymal chondrosarcoma. *Human Pathol* 1996;27:1 273–1276.
- Hamilton A, Davis RI, Hayes D, Mollan RA. Chondrosarcoma developing in synovial chondromatosis. J Bone Joint Surg Br 1987;69:137–140.
- 262. Hashimoto N, Ueda T, Joyama S, et al. Extraskeletal mesenchymal chondrosarcoma: an imaging review of ten new patients. *Skeletal Radiol* 2005;34:785–792.

- 263. Hatano H, Ogose A, Hotta T, et al. Periosteal chondrosarcoma invading the medullary cavity. *Skeletal Radiol* 1997:26:375–378.
- 264. Haygood TM, Teot L, Ward WG, et al. Low-grade chondrosarcoma in a 12-year-old boy. *Skeletal Radiol* 1995;24:466–468.
- 265. Henderson ED, Dahlin DC. Chondrosarcoma of bone: a study of 280 cases. J Bone Joint Surg Am 1963;45:1450–1458.
- 266. Hinrichs SH, Jaramillo MA, Gumerlock PH, et al. Myxoid chondrosarcoma with a translocation involving chromosomes 9 and 22. *Cancer Genet Cytogenet* 1985;14:219–226.
- 267. Hisaoka M, Hashimoto H. Extraskeletal myxoid chondrosarcoma: updated clinicopathological and molecular genetic characteristics. *Pathol Int* 2005;55:453–463.
- 268. Hudson TM. Medullary (central) chondrosarcoma. In: Hudson TM, ed. Radiologic pathologic correlations of musculoskeletal lesions. Baltimore: Williams & Wilkins, 1987:153–175.
- Hudson TM, Chew FS, Manaster BJ. Radionuclide scanning of medullary chondrosarcoma. Am J Roentgenol 1982;139:1071–1076.
- 270. Hudson TM, Hamlin DJ, Enneking WF, Petterson H. Magnetic resonance imaging of bone and soft-tissue tumors: early experience in 31 patients compared with computed tomography. *Skeletal Radiol* 1985;13:134–146.
- 271. Hudson TM, Moser RP, Gilkey FW, Aoki J. Chondrosarcoma. In: Moser RP, ed. *Cartilaginous tumors of the skeleton. AFIP atlas of radiologic-pathologic correlation*, vol 2. Philadelphia: Hanley and Belfus, 1990:155–204.
- 272. Huvos AG. Chondrosarcoma including spindle-cell (dedifferentiated) and myxoid chondrosarcoma: mesenchymal chondrosarcoma. In: Mitchell J, ed. *Bone tumors: diagnosis, treatment, and prognosis,* 2nd ed. Philadelphia: Saunders, 1991:343–393.
- 273. Huvos AG, Marcove RC. Chondrosarcoma in the young: a clinicopathologic analysis of 79 patients younger than 21 years of age. Am J Surg Pathol 1987;11:930–942.
- 274. Ishida T, Dorfman HD, Habermann ET. Dedifferentiated chondrosarcoma of humerus with giant cell tumor-like features. *Skeletal Radiol* 1995;24:76–80.
- 275. Ishida T, Yamamoto M, Goto T, et al. Clear cell chondrosarcoma of the pelvis in a skeletally immature patient. *Skeletal Radiol* 1999;28:290–293.
- 276. Itälä A, Leerapun T, Inwards C, et al. An institutional review of clear cell chondrosarcoma.. *Clin Orthop Relat Res* 2005;440: 209–212.
- 277. Janzen L, Logan PM, O'Connel JX, et al. Intramedullary chondroid tumors of loose: correlation of abnormal peritoneal narrow and soft tissue MRI signal with tumor type. *Skeletal Radiol* 1997;26:100–106.
- Johnson S, Tetu B, Ayala AG, Chawla SP. Chondrosarcoma with additional mesenchymal component (dedifferentiated chondrosarcoma). A clinical study of 26 cases. *Cancer* 1986;58:278–286.
- Kalil RK, Inwards CY, Unni KK, et al. Dedifferentiated clear cell chondrosarcoma. Am J Surg Pathol 2002;24:1079–1086.
- Kaufman JH, Cedermark BJ, Parthasarathy KL, et al. The value of 67Ga scintigraphy in soft-tissue sarcoma and chondrosarcoma. *Radiology* 1977;123:131–134.
- 281. Kawaguchi S, Wada T, Nagoya S, et al. Extraskeletal myxoid chondrosarcoma. A multi-institutional study of 42 cases in Japan. *Cancer* 2003;97:1285–1292.
- 282. Kessler S, Mirra JM, Ishii T, et al. Primary malignant mesenchymoma of bone: case report, literature review, and distinction of this entity from mesenchymal and dedifferentiated chondrosarcoma. *Skeletal Radiol* 1995;24:291–295.
- 283. Kilpatrick SE, Inwards CY, Fletcher CD, et al. Myxoid chondrosarcoma (chordoid sarcoma) of bone: a report of two cases and review of the literature. *Cancer* 1997;79:1903–1910.
- King JW, Spjut HJ, Fechner RE, Vanderpool DW. Synovial chondrosarcoma of the knee joint. J Bone Joint Surg Am 1967;49: 1389–1396.
- 285. Klein MJ. Chondrosarcoma. Semin Orthop 1991;6:167-176.
- Kleist B, Poetsch M, Lang C, et al. Clear cell chondrosarcoma of the larynx: a case report of a rare histologic variant in an uncommon localization. *Am J Surg Pathol* 2002;26:386–392.
- 287. Kransdorf MJ, Jelinek JS, Moser RP Jr, et al. Soft-tissue masses: diagnosis using MR imaging. Am J Roentgenol 1989;153:541–547.
- 288. Kransdorf MJ, Meis JM. Extraskeletal osseous and cartilaginous tumors of the extremities. *RadioGraphics* 1993;13:853–884.
- Kumar R, David R, Cierney G III. Clear cell chondrosarcoma. *Radiology* 1985;154:45–48.

- 290. Kumta SM, Griffith JF, Chow LTC, Leung PC. Primary juxtacortical chondrosarcoma dedifferentiating after 20 years. *Skeletal Radiol* 1998;27:569–573.
- Larramendy ML, Mandahl N, Mertens F, et al. Clinical significance of genetic imbalances revealed by comparative genomic hybridization in chondrosarcomas. *Hum Pathol* 1999;30:1247–1253.
- 292. Lichtenstein L. Tumors of periosteal origin. *Cancer* 1955;8: 1060–1069.
- 293. Lichtenstein L, Bernstein D. Unusual benign and malignant chondroid tumors of bone. *Cancer* 1959;12:1142–1157.
- Lin W-Y, Wang S-J, Yeh S-H. Radionuclide imaging in extraskeletal myxoid chondrosarcoma. *Clin Nucl Med* 1995;20:524–527.
- 295. Lucas D, Heim S. Extraskeletal myxoid chondrosarcoma. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:213–215.
- 296. MacSweeney F, Darby A, Saifuddin A. Dedifferentiated chondrosarcoma of the appendicular skeleton: MRI-pathological correlation. *Skeletal Radiol* 2003;32:671–678.
- 297. Mahajam H, Lorigan JG, Shirkhoda A. Synovial sarcoma: MR imaging. *Magn Reson Imag* 1989;7:211–216.
- 298. Mandahl N, Gustafson P, Mertens F, et al. Cytogenetic aberrations and their prognostic impact in chondrosarcoma. *Genes Chromosomes Cancer* 2002;33:188–200.
- 299. Manivel JC, Dehner LP, Thompson R. Case report 460. Synovial chondrosarcoma of left knee. *Skeletal Radiol* 1988;17:66–71.
- Mankin JH, Cantley KP, Lippiello L, et al. The biology of human chondrosarcoma. I. Description of the cases, grading, and biochemical analyses. *J Bone Joint Surg Am* 1980;62:160–176.
- McCarthy EF, Dorfman HD. Chondrosarcoma of bone with dedifferentiation: a study of eighteen cases. *Hum Pathol* 1982;13: 36–40.
- McFarland GB, McKinley LM, Reed RJ. Dedifferentiation of low-grade chondrosarcomas. *Clin Orthop* 1977;122:157–164.
- McLeod RA. Chondrosarcoma. In: Taveras JM, Ferrucci JT, eds. Radiology: diagnosis, imaging, intervention, vol 5. Philadelphia: JB Lippincott, 1993:1–12.
- 304. McLeod RA, Berquist TH. Bone tumor imaging: contribution of CT and MRI. In: Unni KK, ed. *Bone tumors*. New York: Churchill Livingstone, 1988:1–34.
- 305. Meis-Kindblom JM, Bergh P, Gunterberg B, Kindblom LG. Extraskeletal myxoid chondrosarcoma. A reappraisal of its morphologic spectrum and prognostic factors based on 117 cases. *Am J Surg Pathol* 1999;23:636–650.
- Mercuri M, Picci P, Campanacci M, Rulli E. Dedifferentiated chondrosarcoma. *Skeletal Radiol* 1995;24:409–416.
- 307. Mirra JM, Gold R, Downs J, Eckardt JJ. A new histologic approach to the differentiation of enchondroma and chondrosarcoma of the bones. *Clin Orthop Rel Res* 1985;1:214–237.
- Mirra JM, Marcove RC. Fibrosarcomatous dedifferentiation of primary and secondary chondrosarcomas. J Bone Joint Surg Am 1974;56:285–296.
- 309. Muller S, Soder S, Oliveira AM, et al. Type II collagen as specific marker for mesenchymal chondrosarcomas compared to other small cell sarcomas of the skeleton. *Mod Pathol* 2005;18: 1088–1094.
- Mullins, F, Berard CW, Eisenberg SH. Chondrosarcoma following synovial chondromatosis. A case study. *Cancer* 1965;18: 1180–1188.
- Murphey MD, Walker EA, Wilson AJ, et al. Imaging of primary chondrosarcoma: radiologic-pathologic correlation. *RadioGraphics* 2003;23:1245–1278.
- 312. Nakashima Y, Park Y, Sugano O. Mesenchymal chondrosarcoma. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:255–256.
- 313. Nakashima Y, Unni KK, Shives TC, et al. Mesenchymal chondrosarcoma of bone and soft tissue. A review of 111 cases. *Cancer* 1986;57:2444–2453.
- 314. Naumann S, Krallman PA, Unni KK, et al. Translocation der (13;21)(q10;q10) in skeletal and extraskeletal mesenchymal chondrosarcoma. *Mod Pathol* 2002;15:572–576.
- Nishio J, Reith JD, Ogose A, et al. Cytogenetic findings in clear cell chondrosarcoma. *Cancer Genet Cytogenet* 2005;162:74–77.
- Okamoto S, Hara K, Sumita S, et al. Extraskeletal myxoid chondrosarcoma arising in the finger. *Skeletal Radiol* 2002;31:296–300.

- Ontell F, Greenspan A. Chondrosarcoma complicating synovial chondromatosis: findings with magnetic resonance imaging. *Can Assoc Radiol J* 1994;45:318–323.
- Park YK, Park HR, Chi SG, et al. Overexpression of p53 and rare genetic mutation in mesenchymal chondrosarcoma. *Oncol Rep* 2000;7:1041–1047.
- Park Y-K, Yang MH, Ryu KN, Chung DW. Dedifferentiated chondrosarcoma arising in an osteochondroma. *Skeletal Radiol* 1995;24:617–619.
- 320. Pettersson H, Gillespy T III, Hamlin DJ, et al. Primary musculoskeletal tumors: examination with MR imaging compared with conventional modalities. *Radiology* 1987;164:237–241.
- Pettersson H, Slone RM, Spanier S, et al. Musculoskeletal tumors: T1 and T2 relaxation times. *Radiology* 1988;167:783–785.
- 322. Present DA, Bacchini P, Pignatti G, et al. Clear cell chondrosarcoma of bone. A report of eight cases. *Skeletal Radiol* 1991;20: 187–191.
- Reiter FB, Ackerman LV, Staple TW. Central chondrosarcoma of the appendicular skeleton. *Radiology* 1972;105:525–530.
- 324. Remagen W, Nidecker A, Dolanc B. Case report 368. Enchondroma of the tibia with extensive myxoid degeneration occurrence with secondary highly differentiated chondrosarcoma. *Skeletal Radiol* 1986;15:330–333.
- 325. Resnick D, Kyriakos M, Greenway GD. Tumors and tumor-like lesions of bone: imaging and pathology of specific lesions. In: Resnick D, Niwayama G, eds. *Diagnosis of bone and joint disorders*, 3rd ed. Philadelphia: WB Saunders, 1988.
- Richardson B. Impact of aging on DNA methylation. Ageing Res Rev 2003;2:245–261.
- 327. Richardson ML, Kilcoyne RF, Gillespy T III, et al. Magnetic resonance imaging of musculoskeletal neoplasms. *Radiol Clin North Am* 1986;24:259–267.
- Robinson P, White LM, Sundaram M, et al. Periosteal chondroid tumors. Am J Roentgenol 2001;177:1183–1188.
- 329. Ropke M, Boltze C, Neumann HW, et al. Genetic and epigenetic alterations in tumor progression in a dedifferentiated chondrosarcoma. *Pathol Res Pract* 2003;199:437–444.
- Rosenthal DJ, Schiller AL, Mankin HJ. Chondrosarcoma: correlation of radiological and histological grade. *Radiology* 1984;150: 21–26.
- Salvador AH, Beabout JW, Dahlin DC. Mesenchymal chondrosarcoma—observations on 30 new cases. *Cancer* 1971;28: 605–615.
- Sanberg AA. Genetics of chondrosarcoma and related tumors. Curr Opin Oncol 2004;16:342–354.
- 333. Sandberg AA, Bridge JA. Updates on the cytogenetics and molecular genetics of bone and soft tissue tumors: chondrosarcoma and other cartilaginous neoplasms. *Cancer Genet Cytogenet* 2003;143:1–31.
- Sanerkin NG, Gallagher P. A review of the behaviour of chondrosarcoma of bone. *J Bone Joint Surg Br* 1979;61:395–400.
- Sanerkin NG. The diagnosis and grading of chondrosarcoma of bone. *Cancer* 1980;45:582–594.
- Schajowicz F. Juxtacortical chondrosarcoma. J Bone Joint Surg Br 1977;59:473–480.
- 337. Schajowicz F, Cuevillos AR, Silberman FS. Primary malignant mesenchymoma of bone. *Cancer* 1966;19:1423–1428.
- 338. Schajowicz F, Sissons HA, Sobin LH. The World Health Organization's histologic classification of bone tumors. A commentary on the second edition. *Cancer* 1995;75:1208–1214.
- Scheele PM, von Kuster LC, Krivchenia G. Primary malignant mesenchymoma of bone. Arch Pathol Lab Med 1990;114:614–617.
- Schiller AL. Diagnosis of borderline cartilage lesions of bone. Semin Diagn Pathol 1985;1:42–62.
- 341. Shapeero LG, Vanel D, Couanet D, et al. Extraskeletal mesenchymal chondrosarcoma. *Radiology* 1993;186:819–826.
- 342. Sirsat MV, Doctor VM. Benign chondroblastoma of bone. Report of a case of malignant transformation. J Bone Joint Surg Br 1970;52:741–745.
- Sissons HA, Matlen JA, Lewis MM. Dedifferentiated chondrosarcoma. Report of an unusual case. J Bone Joint Surg Am 1991;73: 294–300.
- 344. Sjögren H, Orndal C, Tingby O, et al. Cytogenetic and spectral karyotype analyses of benign and malignant cartilage tumours. *Int J Oncol* 2004;24:1385–1391.
- Smith J, Botet JR, Yeh SDJ. Bone sarcomas in Paget's disease. A study of 85 patients. *Radiology* 1984;152:583–590.

- 346. Spjut HJ, Dorfman HD, Fechner RE, Ackerman LV. Tumors of bone and cartilage. In: *Atlas of tumor pathology*, 2nd series, fascicle 5. Washington, DC: Armed Forces Institute of Pathology, 1971.
- Steiner G, Greenspan A, Jahss M, Norman A. Myxoid chondrosarcoma of the os calcis: a case report. *Foot Ankle* 1984;5: 84–91.
- Steiner GC, Mirra JM, Bullough PG. Mesenchymal chondrosarcoma. A study of the ultrastructure. *Cancer* 1973;32:926–939.
- Stout AP, Verner EW. Chondrosarcoma of the extraskeletal soft tissues. *Cancer* 1953;6:581–590.
- 350. Subramanian S, West RB, Marinelli RJ, et al. The gene expression profile of extraskeletal myxoid chondrosarcoma. *J Pathol* 2005;206:433–444.
- 351. Sundaram M, McGuire MH, Herbold DR, et al. Magnetic resonance imaging in planning limb-salvage surgery for primary malignant tumors of bone. *J Bone Joint Surg Am* 1986;68:809–819.
- Sundaram M, Percelay S, McDonald DJ, Janney C. Case report 799. Extraskeletal chondrosarcoma. *Skeletal Radiol* 1993;22:449– 451.
- 353. Tateishi U, Hasegawa T, Nojima T, et al. MRI features of extraskeletal myxoid chondrosarcoma. *Skeletal Radiol* 2006;35: 27–33.
- 354. Tetu B, Ordonez NG, Ayala AG, Mackay B. Chondrosarcoma with additional mesenchymal component (dedifferentiated chondrosarcoma). *Cancer* 1986;58:287–298.
- 355. Ueda Y, Okada Y, Nakanishi I. Cellular variant of extraskeletal myxoid chondrosarcoma of abdominal wall—a case report with comparative immunohistochemical study on cartilaginous collagenous proteins in various myxoid mesenchymal tumors. *J Cancer Res Clin Oncol* 1992;118:147–151.
- 356. Unni KK. Dahlin's bone tumors. General aspects and data on 11,087 cases, 5th ed. Philadelphia, Lippincott-Raven,1996.
- 357. Unni KK, Dahlin DC, Beabout JW. Periosteal osteosarcoma. *Cancer* 1976;37:2476–2485.
- 358. Unni KK, Dahlin DC, Beabout JW, Sim FH. Chondrosarcoma: Clear-cell variant: a report of 16 cases. J Bone Joint Surg Am 1976;58:676–683.
- 359. van Beerendonk HM, Rozeman LB, Taminiau AH, et al. Molecular analysis of the INK4A/INK4A-ARF gene locus in conventional (central) chondrosarcomas and enchondromas: indication of an important gene for tumour progression. *J Pathol* 2004;202:359–366.
- Vanel D, De Paolis M, Monti C, et al. Radiological features of 24 periosteal chondrosarcomas. *Skeletal Radiol* 2001;30:208–212.
- Varma DGK, Ayala AG, Carrasco CH, et al. Chondrosarcoma: MR imaging with pathologic correlation. *RadioGraphics* 1992;12: 687–704.
- 362. Wehrli BM, Huang W, De Crombrugghe B, et al. Sox9, a master regulator of chondrogenesis, distinguishes mesenchymal chondrosarcoma from other small blue round cell tumors. *Hum Pathol* 2003;34:263–269.
- Weinberg J, Miller TT, Handelsman JE, et al. Periosteal chondrosarcoma in a 9-year-old girl with osteochondromatosis. *Skeletal Radiol* 2005;34:539–542.
- 364. Weiss AC, Dorfman HD. Clear cell chondrosarcoma: a report of 10 cases and review of the literature. Surg Pathol 1988;1:123–129.
- Weiss SW. Ultrastructure of the so-called "chordoid sarcoma" evidence supporting cartilaginous differentiation. *Cancer* 1976;37: 300–306.
- 366. Welkerling H, Kratz S, Ewerbeck V, Delling G. A reproducible and simple grading system for classical chondrosarcomas. Analysis of 35 chondrosarcomas and 16 enchondromas with emphasis on recurrence rate and radiological and clinical data. *Virchows Arch* 2003;443:725–733.
- 367. Wenger DE, Sundaram M, Unni KK, et al. Acral synovial chondrosarcoma. *Skeletal Radiol* 2002;31:125–129.
- West OC, Reinus WR, Wilson AJ. Quantitative analysis of the plain radiographic appearance of central chondrosarcoma of bone. *Invest Radiol* 1995;30:440–447.
- Wu KK, Collon DJ, Guise ER. Extra-osseous chondrosarcoma. Report of five cases and review of the literature. *J Bone Joint Surg Am* 1980;62:189–194.
- Young CL, Sim FH, Unni KK, McLeod RA. Chondrosarcoma of bone in children. *Cancer* 1990;66:1641–1648.
- 371. Zimmer WD, Berquist TH, McLeod RA, et al. Bone tumors: magnetic resonance imaging versus computed tomography. *Radiology* 1985;155:709–718.

CHAPTER 4

Fibrogenic, Fibroosseous, and Fibrohistiocytic Lesions

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Fibrogenic, fibroosseous, and fibrohistiocytic lesions of bone represent a clinical spectrum ranging from very innocent lesions requiring no treatment at all to highly malignant neoplasms (36) (Table 4-1). All of these lesions have a common dominant constituent cell, the fibroblast (8,21). In general, fibrous lesions are composed of spindle cells (i.e., fibroclasts and myofibroblasts with characteristic ultrastructural features), which produce a collagenous matrix and so-called ground substance consisting of glycosaminoglycans, whereas fibrohistiocytic lesions may or may not produce a collagenous matrix (35). The term "fibrohistiocytic" is no longer clearly defined. It was introduced for lesions believed to be derived from histiocytes that could act as facultative fibroblasts. Although this concept has been seriously questioned, the term is still in use for descriptive purposes (176).

Although some fibrous lesions, such as nonossifying fibroma (NOF) or periosteal desmoid, are not regarded as tumors or are categorized as miscellaneous lesions by the World Health Organization (WHO), they are included in this text for the purpose of differential diagnosis (14).

Benign Lesions

Fibrous Cortical Defect and Nonossifying Fibroma

Fibrous cortical defect (also called metaphyseal defect) and NOF (formerly called nonosteogenic fibroma) are the most common fibrous lesions of bone and have histologically identical characteristics. Some authors prefer the term fibroxanthoma for both lesions (2,21), whereas Schajowicz (29) preferred the term histiocytic xanthogranuloma. These lesions are not true neoplasms but are considered developmental defects (17), arising in the region of the primary trabeculae of a tubular bone and wandering toward the diaphysis as the bone grows in length (26).

Only two cases of NOF have been so far investigated cytogenetically, revealing a deletion of the short arm of chromosome 4 [edl(4)(p14)] and a clonal chromosomal

Table 4-1	Spectrum of Fibrous,	Fibroosseous,	and		
Fibrohistiocytic Lesions of Bone					

Condition			Histopathology	/
Fibrous corti	cal defect		Benign	
Nonossifying	fibroma		Benjan	
Benign fibrohistiocytoma			Benign	
(fibroxant	homa)		g.	
Congenital f	ibromatosis		Benign	
Periosteal de	smoid		Benign	
Fibrous dysp	lasia		Benign	
Monostoti	C			
Osteofibrous	dysplasia (Ken	nnson-	Benjan	
Campanao	ci lesion)	1103011-	Denigh	
Desmoplasti	c fibroma		Intermediate	
Fibrosarcom	a		Malignant	
Malignant fil	<mark>orous hist</mark> iocyto	oma	Malignant	
Nonossifying Benign fibro (fibroxantl Congenital fi Periosteal de Fibrous dysp Monostoti Polyostotic Osteofibrous Campanac Desmoplasti Fibrosarcom Malignant fil	j fibroma histiocytoma homa) ibromatosis smoid lasia c dysplasia (Ken cci lesion) c fibroma a brous histiocyto	npson-	Benign Benign Benign Benign Benign Intermediate Malignant Malignant	

translocation t(1;4)(p31:q34) involving the short arm of chromosome 1 and the long arm of chromosome 4 (25,34).

Clinical Presentation

Seen predominantly in childhood and adolescence and more commonly (2:1) in boys (5), both lesions have a predilection for the long bones, particularly the femur and the tibia (Fig. 4-1) (8). Fibrous cortical defect is a small, asymptomatic lesion (31) that occurs in 30% of the normal population during the first and second decades of life (7,11,22). The term NOF is applied to this lesion if it enlarges and encroaches on the medullary portion of the bone (28). Depending on age, the distance of these lesions from the growth plate varies (26). When the longitudinal axis of a fibrous metaphyseal defect projects in the direction of the epiphysis it encounters a tendon or ligamentous structure that inserts at the epiphyseal line (27). On the basis of this finding, some investigators believe that such an insertion, along with additional pathogenetic factors, may be related in some degree to the development of this lesion (27). Recent investigations have demonstrated characteristic radiomorphologic stages of progression and regression of fibrous cortical defects (26). At stage A they are located near the growth plate (31), have a round to oval shape, and exhibit a fine sclerotic margin. In stage B the lesion has wandered into the metaphysis and represents a polycyclic form, also delineated by a distinct sclerotic margin, giving the lesion a grape-like appearance. Stage C represents the beginning of the healing process and shows increased mineralization, which characteristically begins at the diaphyseal end of the lesion and progresses toward the growth plate (19,26). Stage D represents complete healing, and the lesion becomes sclerotic.

Most NOFs are asymptomatic, but larger ones may undergo a pathologic fracture (1). Although multifocal presentation is rare (3,12), Moser et al. (24) found that about 8% of patients exhibit a multicentric localiza-



Figure 4-1 Fibrous cortical defect and nonossifying fibroma: skeletal sites of predilection, peak age range, and male-to-female ratio.



Figure 4-2 Fibrous cortical defect and nonossifying fibroma. A: Fibrous cortical defect, seen here in the lateral cortex of the distal tibia in a 13-year-old boy, typically presents as a radiolucent lesion demarcated by a thin zone of sclerosis (*arrow*). **B:** When a fibrous cortical defect encroaches on the medullary cavity, it is called a nonossifying fibroma. Note the similarity of the lesion to that in **A.** The only difference is the size.

tion. The multiple lesions occur in four patterns: (a) clustered together, usually around the knee; (b) nonclustered, at opposite ends of long bones; (c) several small lesions gradually coalescing into one large lesion; and (d) a metachronous appearance, with new lesions eventually appearing near the original lesion (24). Occasionally such multiple NOFs are associated with neurofibromatosis and café-au-lait spots, known as **Jaffe-Campanacci syndrome** (23,30,32).

Imaging

Radiography is usually diagnostic (15). NOFs present as elliptic, radiolucent defects confined to the cortex of a long bone near the growth plate, frequently resembling a bunch of grapes and demarcated by a thin sclerotic margin (Fig. 4-2). They range in size from 0.5 to 7 cm, with their long axes aligned with the long axis of the affected bone (17). The larger defects are usually scalloped and have a wider zone of sclerosis. Most foci undergo spontaneous involution (healing) by the remodeling process of the tubular bone with diaphyseal diminution of the diameter finally resulting in osteosclerosis. A few lesions may continue to grow and as they encroach on the medullary cavity they characteristically display a scalloped, sclerotic border and are located eccentrically within the bone (Figs. 4-3 and 4-4). Some of the larger lesions may be complicated by a fracture (Fig. 4-5).

Skeletal scintigraphy shows a minimal to mild increase in activity (4). During the healing phase, mild hyperemia may be seen on the blood pool image, and the positive delayed scan reflects the osteoblastic activity (16). Computed tomography (CT) may demonstrate to better advantage the cortical thinning and medullary involvement (Fig. 4-6) and may more precisely delineate early pathologic fracture. Hounsfield attenuation values for NOF are higher than for normal bone marrow. Magnetic resonance imaging (MRI), usually performed for another reason, shows a low signal intensity on



Figure 4-3 Nonossifying fibroma. The lesion, seen here in the distal tibia in an asymptomatic 15-year-old boy, appears eccentrically located in the bone and has a scalloped sclerotic border.



Figure 4-4 Nonossifying fibroma. A, B: Anteroposterior and lateral radiographs show a large elliptical, lobulated lesion in the proximal tibial diaphysis exhibiting well-defined sclerotic border. Note the eccentric location of the lesion, which only slightly scallops the endosteal aspect of the anteromedial cortex.



Figure 4-5 Nonossifying fibroma: a pathologic fracture. A large radiolucent lesion in the distal tibia of a 10-year-old boy is complicated by a pathologic fracture.



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Figure 4-6 Nonossifying fibroma (NOF): computed tomography (CT). **A:** Oblique radiograph of the right tibia of a 14-year-old girl shows an elliptical radiolucent lesion with sclerotic border. **B:** Axial CT section shows the intramedullary lesion with well-defined sclerotic border extending into the anterior cortex of the tibia. **C:** Coronal reformatted CT shows the full extent of NOF. Observe thinning of the anterior cortex.







T1-weighted sequences (17) (Figs. 4-7 and 4-8B). On T2 weighting the lesion exhibits either low signal (more common) or high signal intensity (less common) (20) (Fig. 4-8C). After gadolinium diethylenetriamine–penta-acetic (DTPA) injection, NOFs invariably exhibit a hyperintense border and signal enhancement (27) (Fig. 4-8D). Mineralization of the lesion during healing (stages C and D) appears predominantly as low signal intensity on MR images.

Histopathology

Histologically identical regardless of size, fibrous cortical defect and NOF are composed of rather bland monomorphous spindle cells admixed with histiocytic cells that often possess a clear or foamy cytoplasm (33). The cells are frequently arranged in a swirling storiform pattern (derived from Greek and implying resemblance to a woven mat), typical of fibrohistiocytic lesions, but a fascicular arrangement may also be seen (Fig. 4-9). A variable number of lipid-bearing xanthomatous cells and of hemosiderin pigment-laden histiocytes may be present (29) (see Fig. 4-9C). In some cases, unusually heavily lipidized fibroxanthomatous areas are predominant. Such lesions have been referred to as xanthoma or fibroxanthoma by some investigators (2). Xanthomas in bone may also rarely be associated with juvenile xanthogranuloma or xanthoma disseminatum, a cutaneous

form of non-Langerhans cell histiocytosis, and with hyperlipidemia, usually found at older ages and in atypical skeletal sites, such as flat bones, or as diaphyseal lesions (9,10,13). In hyperlipidemia, cutaneous xanthelasmas and detectable disturbances in lipid values may be absent at the time of bone manifestations (9). Osteoclast-like giant cells are usually present in NOF (Fig. 4-10) and, when numerous, may lead to confusion with giant cell tumor of bone (18). Typically, variable numbers of inflammatory cells, especially lymphocytes, and a few plasma cells are scattered in the background. The amount of collagen production may vary, but no bone or cartilage formation should be present unless a fracture or previous surgical disruption of the lesion has occurred. Exceptionally, bizarre pseudosarcomatous nuclear features may be regarded as degenerative or ischemic changes that may raise the suspicion of malignancy when radiologic and clinical details are not taken into consideration (6).

Differential Diagnosis

Radiology

Both fibrous cortical defect and NOF have rather characteristic imaging features (see previous discussion) that make radiography diagnostic. Rarely, a fibrous cortical defect affecting the posterior aspect of the distal



Figure 4-8 Nonossifying fibroma (NOF): magnetic resonance imaging (MRI). A: Anteroposterior radiograph shows a radiolucent lesion with sclerotic border abutting the posteromedial cortex of the right femur. **B**: Sagittal T1-weighted MRI shows predominantly low signal intensity of the lesion. **C**: Sagittal T2-weighted image shows heterogeneous but mostly high signal intensity of NOF. D: Sagittal T1-weighted fat-suppressed MRI after intravenous injection of gadolinium shows slight heterogeneous enhancement of the lesion.

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Figure 4-9 Histopathology of nonossifying fibroma. A: Fibrous stroma displays a storiform arrangement. Several giant cells are present (hematoxylin and eosin, original magnification $\times 100$). **B:** The fibrous stroma exhibits a storiform arrangement of slender cells (hematoxylin and eosin, original magnification $\times 200$). **C:** At high magnification several foam cells are conspicuous (hematoxylin and eosin, original magnification $\times 400$).

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Figure 4-10 Histopathology of nonossifying fibroma. A: Siderin-containing macrophages are irregularly distributed within a fairly dense spindle cell tissue (Giemsa, original magnification $\times 200$). **B:** In another area the number and the size of the giant cells containing up to 50 nuclei resemble that of a true giant cell tumor of bone (hematoxylin and eosin, original magnification $\times 50$).



Figure 4-10 Continued C: At high magnification the resemblance of giant cells to those of a giant cell tumor is striking (hematoxylin and eosin, original magnification \times 400).

femur may be mistaken for *periosteal desmoid*, and a NOF in long or flat bones may be mistaken for a focus of *fibrous dysplasia* or *desmoplastic fibroma*. An atypical sclerotic presentation of NOF or healing stage of this lesion may occasionally be confused with a *medullary bone infarct, sclerotic metastasis*, or a *giant bone island* (Fig. 4-11). If a lesion is located in thin bones (e.g., fibula, ulna) and occupies the entire diameter of the bone, its characteristic eccentric location is lost and it may therefore mimic a *simple bone cyst* (Fig. 4-12), an *aneurysmal bone cyst* (Fig. 4-13), or a *Langerhans cell histiocytosis*. Whenever NOF affects the anterior aspect of the tibia,



Figure 4-11 Healed nonossifying fibroma. A lesion that healed spontaneously may persist as a sclerotic patch. In this sclerosing phase, nonossifying fibroma should not be mistaken for osteoblastic tumor or bone island.



Figure 4-12 Nonossifying fibroma resembling a simple bone cyst. Multiloculated lesion in the distal fibula mimics a simple bone cyst. Periosteal reaction at the lateral cortex is secondary to a healed pathologic fracture.

osteofibrous dysplasia should be considered in differential diagnosis (Fig. 4-14).

Pathology

NOF can be confused with benign fibrous histiocytoma because both lesions have almost identical histopathologic features, differing mainly in their clinical and radiologic presentation. A storiform architecture in a hypercellular NOF or pseudosarcomatous nuclear changes can lead to confusion with malignant fibrous histiocytoma (MFH) or unclassified sarcoma if the pathologist is unfamiliar with the clinicopathologic spectrum of fibrous cortical defect and NOF. Less likely differential possibilities are giant cell tumor, fibrous dysplasia, osteofibrous dysplasia, or *low-grade intraosseous osteosarcoma*. Although giant cells are almost always present in NOF, they are not uniformly distributed throughout the lesion as in giant cell tumor (see Figs. 4-10 and 7-10). Because bone formation is not a feature of NOF, all three lesions-fibrous dysplasia, osteofibrous dysplasia, and low-grade osteosarcoma-can easily be distinguished. If xanthoma cells are numerous and a storiform pattern of fibrohistiocytic cells is not obvious, disturbances of lipid metabolism (in elderly patients) or non-Langerhans cell histiocytic disorders (in children or adolescents, ordinarily involving also the dermis) with osseous manifestations must be considered.

The radiologic and pathologic differential diagnosis of NOF is depicted in Figure 4-15.



Figure 4-13 Nonossifying fibroma resembling an aneurysmal bone cyst. A large lesion in the fibula exhibits an expansive character and thus resembles an aneurysmal bone cyst. Note, however, the absence of a periosteal reaction that is invariably present in the latter lesion.

Benign Fibrous Histiocytoma

The term benign fibrous histiocytoma (sometimes called xanthofibroma or fibrous xanthoma) may be controversial (36,42,47), but it is useful in an organizational sense for subclassification of lesions with histologic features similar to those of NOF but having an atypical location and an atypical clinical or radiographic pattern (45,46,48). This is a rare lesion characterized by a spindle cell fibrous tissue arranged in a storiform pattern. The tissue contains variable numbers of giant cells, hemosiderin, and lipid-laden histiocytes (45).

Clinical Presentation

The lesion, which is often symptomatic (painful), is usually diagnosed in older patients who are in the age range from 15 to 60 years. The usual location is the shaft or (in contrast to NOF) the articular end of a long bone, the pelvis, and the ribs. Rarely, the clavicle, spine, or skull is affected (29) (Fig. 4-16). Clinically, the lesion can exhibit a locally aggressive behavior, with recurrences after curettage or excision.



Figure 4-14 Nonossifying fibroma resembling osteofibrous dysplasia. Lateral radiograph of the leg shows a long lesion with sclerotic border affecting the anterior cortex of the tibia and extending into the medullary portion of bone. Because this is a preferential location for osteofibrous dysplasia, the latter diagnosis must be considered.

Imaging

Radiographically, benign fibrous histiocytoma may also appear to be more aggressive than NOF, although some features are similar. Radiographs show a welldefined, radiolucent lesion with sclerotic borders, occasionally exhibiting some degree of expansion (41) (Fig. 4-17). Internal trabeculations may also be present (38). The lesion may be situated either centrally in the bone or eccentrically, the latter presentation being more common at the articular end of bone (43). On skeletal scintigraphy, benign fibrous histiocytoma exhibits a moderately increased uptake (41). MRI shows the lesion to be of intermediate signal intensity (isointense with the muscles) on spin-echo (SE) T1-weighted sequences and of a high signal intensity on T2- and inversion time-inversion recovery (TI-IR) images (37,41).

Histopathology

Benign fibrous histiocytoma usually has histologic features quite similar to those of NOF, although the storiform spindle cell stroma is more distinctive (29,44)



Figure 4-15 Radiologic and pathologic differential diagnosis of nonossifying fibroma.

(Fig. 4-18). The background may contain larger numbers of histiocytic cells and Touton-type giant cells seen in xanthomatous tissue reactions (39) or in MFH. Although frankly malignant histologic criteria are absent and no cytologic atypia is seen, mitotic activity may be present (40). Reactive bone formation may be seen at the periphery of the lesion or in association with a pathologic fracture through the lesion (36).

Differential Diagnosis

Radiology

From the radiologic standpoint, the main differential possibilities include NOF and giant cell tumor, particularly when benign fibrous histiocytoma is situated at the articular end of bone. The radiographic features of NOF may be indistinguishable from those of benign fibrous histiocytoma, although the more aggressive appearance of the latter (such as an expanding border) and the location, which is atypical for NOF, are the clues to the diagnosis. Giant cell tumor only rarely exhibits a rim of reactive sclerosis, whereas this is a common feature of benign fibrous histiocytoma. Although intraosseous ganglion is usually much smaller than the average size of benign fibrous histiocytoma, the former should be considered in differential diagnosis if the lesion occurs at the articular end of a bone. Osteoblastoma, unlike benign fibrous histiocytoma, usually evokes a periosteal reaction and exhibits central opacities of bone formation.

Pathology

The differentiation of NOF and fibrous cortical defect from benign fibrous histiocytoma must be made on a clinical and radiographic basis rather than on histologic grounds (36) because the microscopic appearance of all three lesions is almost identical (Fig. 4-19).

Giant cell tumor always must be considered in the differential diagnosis. It is particularly difficult to distinguish a giant cell tumor that undergoes regressive changes and, conversely, some giant cell tumors with regressive features may erroneously be diagnosed as benign fibrous histiocytoma (38). Furthermore, the healing reaction associated with a pathologic fracture in a giant cell tumor can lead to proliferation of spindle cells with associated lipid- and hemosiderin-laden macrophages, giving the appearance of benign fibrous histiocytoma (36). Finally, some investigators consider benign fibrous histiocytoma a regressive stage of a burned-out giant cell tumor (43).

The radiologic and pathologic differential diagnosis of benign fibrous histiocytoma is depicted in Figure 4-20.

Periosteal Desmoid

Periosteal desmoid (also known as avulsive cortical irregularity, distal metaphyseal femoral defect, cortical desmoid, or medial supracondylar defect of the femur) (52) is a tumor-like fibrous proliferation of



Figure 4-16 Benign fibrous histiocytoma: skeletal sites of predilection, peak age range, and male-to-female ratio.



Figure 4-17 Benign fibrous histiocytoma. A: A 37-year-old man presented with occasional pain in the right knee. An oblique radiograph of the knee shows a lobulated radiolucent lesion with a well-defined sclerotic border, located eccentrically in the proximal tibia. **B:** A 16-year-old boy presented with a painful tibial lesion that on radiography looked like a nonossifying fibroma. Because of persistent symptoms the lesion was biopsied and proved to be consistent with a benign fibrous histiocytoma.



Figure 4-18 Histopathology of benign fibrous histiocytoma. A: Benign-appearing fibrous tissue displays a storiform arrangement (hematoxylin and eosin, original magnification $\times 100$). **B:** Giant cells and some inflammatory cells are present. The spindle cells do not exhibit atypical features (hematoxylin and eosin, original magnification $\times 200$).

the periosteum, which has a striking predilection for the posteromedial cortex of the medial femoral condyle (50). It was first described in 1951 by Kimmelstiel and Rapp (54) as a benign fibrous lesion originating beneath the periosteum and associated with erosion of the underlying bone. Although the term periosteal desmoid is widely accepted, it has an unfortunate similarity to the term desmoid tumor of soft tissue and therefore is suggestive of an aggressive lesion (35). Periosteal desmoid usually occurs between the ages of 12 and 20 years, mainly in boys, and most lesions disappear spontaneously by the end of the second decade. It is considered to represent an asymptomatic normal variant that does not require biopsy or treatment. Although many patients have a history of injury, trauma does not necessarily predispose to the development of this lesion (56). The radiologic features of periosteal desmoid are similar to those of fibrous cortical defect, except for the specificity of its location. Its radiographic hallmarks are a saucer-like radiolucent defect with sclerosis at its base. The lesion may erode the cortex (55) and appear as a cortical irregularity (Fig. 4-21). Occasionally, because of its poorly defined and irregular bony margin and the presence of small bony spiculae at the cortical surface, it may simulate an aggressive or even a malignant tumor, such as osteosarcoma or Ewing sarcoma (49,50). The bone scan usually is normal but sometimes may show a focal increase in activity (17,53,57). On MRI the lesion appears hypointense on T1- and hyperintense on T2weighted images, with a dark rim on both sequences at or near the sites of the bone attachment of the medial head of the gastrocnemius muscle (58). Histopathologic examination of the lesion demonstrates fibroblastic spindle cells that produce large amounts of collagen (51)



Figure 4-19 Histopathology of benign fibrous histiocytoma. A: Densely arranged spindle cells exhibit a cartwheel-like arrangement (*center*). In contrast to nonossifying fibroma, giant cells are rarely encountered (hematoxylin and eosin, original magnification \times 50). **B:** Spotty increase of collagen fiber formation (*red*) resembles primitive osteoid (van Gieson, original magnification \times 20).





(Fig. 4-22). Large areas of hyalinization and fibrocartilage and small fragments of bone may be scattered within the fibrous tissue. Newly formed capillaries may also be present. Unlike NOF or fibrous cortical defect, giant cells or xanthomatous cells are not present. The most important point in the differential diagnosis is not to mistake this lesion for a malignancy. Its characteristic radiographic appearance and location should serve as clues to the correct diagnosis.

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The radiologic and pathologic differential diagnosis of periosteal desmoid is shown in Figure 4-23.

Fibrous Dysplasia

Fibrous dysplasia, occasionally termed fibrous osteodystrophy, osteodystrophia fibrosa, and osteitis fibrosa disseminata (70), is a fibroosseous lesion that may affect one bone (monostotic form, 70% to 80% of patients) or

B



Figure 4-21 Periosteal desmoid. A: Oblique radiograph of the left knee of a 12-year-old boy shows the classic appearance of periosteal desmoid. Note the elliptical radiolucency eroding the medial border of the distal femoral metaphysis at the linea aspera and producing cortical irregularity (*arrow*). **B:** In another patient, an 11-year-old boy, a saucer-like defect is present in the medial cortex of the distal femoral metaphysis (*arrow*).



Figure 4-22 Histopathology of periosteal desmoid. The cortical bone (lower left) is made up of coarse plates of cellrich woven bone merging with benign-appearing fibrous tissue (hematoxylin and eosin, original magnification $\times 12$).

several bones (polyostotic form, 20% to 30%) (98). Classified as a developmental abnormality by some authorities, it is now considered as a genetically based sporadic disorder and is characterized by the replacement of normal lamellar cancellous bone by an abnormal fibrous tissue that contains small, abnormally arranged trabeculae of immature woven bone (17,91,118), formed by metaplasia of the fibrous stroma. Fibrous dysplasia presents with an equal gender distribution and occurs in all ethnic groups. Although monostotic fibrous dysplasia is usually diagnosed in young adults and polyostotic forms in children and adolescents, syndrome-associated lesions may be detected as early as in infancy (118).

A mutation in the GNAS1 gene that codes for the alpha-subunit of a signal transducing membrane protein (Guanine Nucleotide-finding Protein, Alpha-Stimulating Activity Polypeptide 1) is involved in fibrous dysplasia. This protein belongs to the group of guanine nucleotide binding proteins, or G-proteins, involved in second messenger cascades. Because mutations in this autosomal gene located on 20q13 are lethal, only postzygotic mutations leading to mosaicism, i.e., to the existence of two cell lines with a different genetic makeup within one individual, are observed (81,82). Depending on whether it occurs during embryonic development or postnatal life, it is believed that the mutation will lead to monostotic or polyostotic fibrous dysplasia or to Mc-Cune-Albright syndrome. The GNASI mutation is of the activating type, causing increased activity of the enzyme adenylcyclase, which leads to overproduction of cyclic adenosine monophosphate (cAMP) (117). This affects proliferation and differentiation of preosteoblasts by increasing the expression of genes that contain cAMPresponsive elements, such as c-fos, c-jun, IL-6, and Il-11 (65, 67, 101, 102, 113).

Clonal chromosomal alterations have been reported in 8 of 11 investigated cases of monostotic fibrous dysplasia, with structural recurrent aberrations of chro-



Figure 4-23 Radiologic and pathologic differential diagnosis of periosteal desmoid.

mosome 12 (12p13) and trisomy 2 in three cases each, suggesting that fibrous dysplasia may be a neoplastic process (73).

Monostotic Fibrous Dysplasia

Clinical Presentation

Most commonly affecting the femur (with a predilection for the femoral neck), the tibia, the ribs (representing the most common benign lesion of the rib), and the base of the skull, the lesion of monostotic fibrous dysplasia arises centrally in the bone; it usually spares the epiphysis in children and the articular end of the bone in adults (83). The incidence in men and women is the same (Fig. 4-24). Because these lesions are usually asymptomatic, most of them are discovered incidentally on radiographs obtained for other reasons. Rarely, pain accompanied by local swelling and deformity is present. The most common complication is a pathologic fracture through the structurally weakened bone. This may be the initial presentation, particularly for lesions located in the lower extremity (29).

Imaging

The radiographic appearance of the lesion depends on the proportion of osseous to fibrous tissue. Lesions with higher degrees of ossification appear more dense and sclerotic. More fibrous lesions exhibit a greater radiolucency, with a characteristic "ground-glass" or even cystic appearance (Fig. 4-25). Some lesions may contain scattered calcifications similar to those of chondroid lesions. A solitary focus of fibrous dysplasia may be surrounded by a characteristic thick band of reactive bone, creating a "rind" sign (Fig. 4-26). This appearance of fibrous dysplasia must be differentiated from the very similar presentation of a bone infarct (see Fig. 4-43B). When the lesion enlarges, it expands the medullary cavity. Although the cortex may be attenuated and even remodeled around the lesion, it is rarely directly involved by the lesion. In extremely rare instances, lesions of fibrous dysplasia may protrude from the bone (the so-called exophytic variant of fibrous dysplasia) (75).

Scintigraphy is helpful in determining the activity (Fig. 4-27) and the potential multicentricity of the lesion (86). Machida et al. (100) reported that although a high incidence of increased uptake of radiopharmaceutical was seen in 59 patients with fibrous dysplasia, 10% of the lesions with a ground-glass appearance failed to show similarly increased uptake.

The lesion of fibrous dysplasia shows a variety of appearances on MRI (94). Some lesions show a decreased signal on both T1 and T2 sequences and some low signal on T1 but either mixed or high signal on T2 images (90). The sclerotic rim (rind sign) is invariably imaged as a band of low signal intensity on both T1 and T2 sequences (122).

Histopathology

Histologically, fibrous dysplasia consists of an aggregate of moderately dense fibrous connective tissue composed



Figure 4-24 Monostotic fibrous dysplasia: skeletal sites of predilection, peak age range, and male-to-female ratio.



Figure 4-25 Monostotic fibrous dysplasia. A: Anteroposterior radiograph of the distal leg of a 17-year-old girl shows a monostotic focus of fibrous dysplasia in the diaphysis of the tibia. Observe the slight expansion and thinning of the cortex and the partial loss of trabecular pattern in the cancellous bone, which gives the lesion a "ground-glass" or "smoky" appearance. **B:** Anteroposterior radiograph of the right hip in a 25-year-old man shows the focus of fibrous dysplasia in the femoral neck that exhibits a more sclerotic appearance than that seen in **A**.

of spindle cells arrayed in intersecting, whorled bundles containing haphazardly distributed bone trabeculae. Rather than being stress oriented, as in normal cancellous bone, these trabeculae tend to be irregularly curved and branching with sparse interconnections (62,107). The collagen fibers of the stroma are characteristically continuous with those of the bone trabeculae, reminiscent of Sharpey fibers. The low-power photomicrographic picture has been likened to Chinese characters or even alphabet soup (Fig. 4-28). Although bone trabeculae contain osteocytes, they exhibit no evidence of osteoblastic activity. In some areas, cementicles-like rounded trabeculae may be present (Fig. 4-29) and scattered osteoclasts may be found. Areas of hyaline cartilage may rarely be present. Under polarized light, the bone trabeculae are seen to be composed of woven immature bone (Fig. 4-30). The cells of the stroma and of

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the trabeculae show a strong alkaline phosphatase activity, indicating that they are all potential osteoblasts. It appears that an organizational defect, now identified as mutation of the GNAS1 gene, prevents them from differentiating into fully mature, polarized osteoblasts capable of forming normal lamellar bone. The lack of seams of polarized osteoblasts at any stage in their maturation ("naked trabeculae") and the direct passage of the collagen fibers from the stroma into the bone trabeculae are the most important characteristics in differential diagnosis (see Fig. 4-29). Therefore, the bone formed is considered immature (see Fig. 4-30). Furthermore, typical findings on electron microscopy consist of fairly extended areas with collagen fibrils in the precollagen stage and in continuation with normal-appearing mature collagen fibrils and calcification (Remagen, unpublished observations, 1996) (Fig. 4-31).



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Figure 4-26 Monostotic fibrous dysplasia. A: In this presentation of fibrous dysplasia the thick peripheral rim and a radiolucent center create the image of a "rind" sign. B: In another patient a similar rind sign marks a focus of fibrous dysplasia. C: Computed tomography section through the lesion shows a thick rim of peripheral sclerosis (arrows).



Figure 4-27 Monostotic fibrous dysplasia: scintigraphy. A: A 24-year-old woman presented with mild discomfort in the right leg. Anteroposterior radiograph shows a radiolucent lesion with a "smoky" appearance in the midshaft of the tibia, associated with thinning of the cortex and slight expansion. **B:** Radionuclide bone scan shows markedly increased uptake of the radiopharmaceutical agent, indicating metabolic activity of fibrous dysplasia.

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Figure 4-28 Histopathology of fibrous dysplasia. A: Metaplastic trabeculae are haphazardly arranged in loose, fibrous stroma. Note lack of osteoblastic activity (hematoxylin and eosin, original magnification \times 200). **B:** Collagen, reminiscent of Sharpey fibers, extends into the stroma surrounding the trabeculae. Note the absence of osteoblastic rimming (van Gieson, original magnification \times 200)

Figure 4-29 Histopathology of fibrous dysplasia. Variant of classic histologic appearance with small, discrete, almost osteocyte-free foci of calcified matrix that resemble the cementicles usually seen in cementifying fibromas of the jaw. The scanty spindle cell stroma shows the typical passing of the collagen fibers into the bone (hematoxylin and eosin, original magnification \times 25).





Figure 4-30 Histopathology of fibrous dysplasia. A: In a background of collagenized fibrous tissue, irregularly shaped spicules of immature woven bone have been formed. No polarized differentiated osteoblasts are seen (hematoxylin and eosin, original magnification \times 50). **B:** Under polarized light the woven structure of the collagen fibers within the bone matrix is conspicuous (hematoxylin and eosin, polarized light, original magnification \times 25).



Figure 4-31 Electron microscopy of fibrous dysplasia. Mature cross-striated collagen fibrils are seen (*upper right corner*) as well as an extended area with immature collagen fibrils. Mineralization of the matrix progresses (*lower left*) regardless of the structure and maturation of the collagen.

Polyostotic Fibrous Dysplasia

Clinical Presentation

Although radiographically similar to the monostotic form, polyostotic fibrous dysplasia often exhibits a somewhat more aggressive appearance. Moreover, the lesions are distributed differently in the skeleton, showing a striking predilection (more than 90% of cases) for one side of the body. They often involve the pelvis, followed in frequency by the long bones, skull, and ribs; the proximal end of the femur is a common site of occurrence (Fig. 4-32). In general, the lesions of polyostotic fibrous dysplasia progress in number and size until skeletal maturity is reached, after which they become quiescent. Only 5% of lesions continue to enlarge. Unlike the monostotic form, this variant of fibrous dysplasia is usually symptomatic. In most patients the presenting symptoms, such as a limp, leg pain, or a pathologic fracture, result from skeletal involvement. However, at a very early age, associated endocrine abnormalities (such as vaginal bleeding) usually appear first (119).

Polyostotic fibrous dysplasia that is associated with endocrine disturbances (i.e., premature sexual development, gigantism or acromegaly, hyperthyroidism, hyperparathyroidism, and Cushing syndrome) and skin pigmentation (café-au-lait spots) constitutes the disorder called **McCune-Albright syndrome** (59,106). This condition, which is caused by somatic mutations in the



Figure 4-32 Polyostotic fibrous dysplasia: skeletal sites of predilection, peak age range, and male-to-female ratio.
GNAS1 gene located in chromosome 20q13 (68), appears to predominantly affect girls, who exhibit true sexual precocity resulting from accelerated gonadotropin release by the anterior lobe of the pituitary. However, a recent study indicates that, in a substantial number of cases, children treated for isolated fibrous dysplasia actually have unrecognized McCune-Albright syndrome, which becomes apparent when appropriate tests are performed (80). Typically, the café-au-lait spots of McCune-Albright syndrome show irregular, ragged ("coast of Maine") borders, in contrast to the smoothly marginated ("coast of California") café-au-lait markings seen in neurofibromatosis. Mazabraud syndrome is a condition in which multiple fibrous and fibromyxomatous soft tissue tumors occur in association with polyostotic fibrous dysplasia (79,89,120). In this syndrome, which was first described by German pathologist F. Henschen in 1926 (84) and later reemphasized by French physician A. Mazabraud in 1967 (105), it is important to recognize the soft tissue masses as benign myxomas (60,63,95,109) and not to confuse them with malignant soft tissue tumors that may develop de novo (e.g., MFH, malignant mesenchymoma, or liposarcoma), or those that may be present in cases of malignant transformation of fibrous dysplasia.

The most common complication of polyostotic fibrous dysplasia is a pathologic fracture that, when it occurs in the femoral neck, frequently leads to a "shepherd's crook" deformity (Fig. 4-33). Occasionally, accelerated growth of a bone or hypertrophy of a digit may be observed. The development of a sarcoma in fibrous dysplasia is extremely rare, but it may occur either spontaneously (93,114,116,123) or, more commonly, after radiation therapy (115,123).

Imaging

The characteristic radiographic changes of polyostotic fibrous dysplasia may occur in any portion of the long bones, and involvement of the bone by the lesion may be limited or extensive. The lower extremities are most commonly affected, with a preferentially unilateral distribution. As in the monostotic form, the articular ends of bones are usually unaffected. This latter phenomenon has given rise to speculations that a defect of the primary ossification center may lead to impairment of the production and maintenance of mature cancellous bone. The cortex, as in the solitary form, is usually intact but is often thinned because of the expansive nature of the lesion. The borders of the lesion are well defined, and the inner cortical margins may exhibit scalloping. The variable amount of bone in the fibrous tissue that replaces the cancellous bone imparts a radiographic picture that ranges from lucency to increased density (Fig. 4-34A). More fibrous lesions exhibit greater lucency, occasionally with a loss of the trabecular pattern, and a groundglass, milky, or smoky appearance (Fig. 4-34B). Massive formation of cartilage may be observed in the lesion, accompanied by secondary calcification and ossification patterns, known as fibrocartilaginous dysplasia or fibrochondrodysplasia (76,85,88,97), that can easily be confused radiographically with a cartilaginous neoplasm



Figure 4-33 Polyostotic fibrous dysplasia. A "shepherd'scrook" deformity, seen here in the proximal femur in a 12-year-old boy, is often the result of multiple pathologic fractures.

(Fig. 4-35) (72,74). This condition should not be confused with so-called *focal fibrocartilaginous dysplasia of long bones* (66), which occurs mainly in children and young adults. Characteristically, it affects the proximal tibia, although other long bones, such as the ulna and femur, may sometimes be involved. Radiographic examination ordinarily shows a radiolucent lesion in the medial metadiaphyseal segment of the bone and perilesional cortical thickening (Fig. 4-36).

Skull lesions in fibrous dysplasia affect primarily the base, causing thickening and sclerosis of the sphenoid wings, sella, and the vertical portion of the frontal



Figure 4-34 Polyostotic fibrous dysplasia. A: Anteroposterior radiograph of the right hip of an 18-year-old woman shows unilateral involvement of the ilium and femur. Some lesions are more radiolucent, some more sclerotic. **B:** In this anteroposterior radiograph of the left leg of a 5-year-old girl with precocious puberty whose left upper and lower extremities were affected (McCune-Albright syndrome), note slight expansion of the tibia and fibula associated with thinning of the cortex and "ground-glass" appearance of the medullary portion of these bones.



Figure 4-35 Fibrocartilaginous dysplasia. A: Anteroposterior radiograph of the right femur of a 10-year-old boy with polyostotic fibrous dysplasia exhibits typical appearance of a massive formation of cartilage. **B:** Coned-down view of the proximal femur displays annular calcifications typical for a cartilaginous matrix.

Figure 4-36 Focal fibrocartilaginous dysplasia of long bones. Anteroposterior radiograph of the left knee of a 2-year-old boy shows a well-defined radiolucent lesion in the medial diaphysis of the tibia associated with focal thickening of the cortex. (Courtesy of Dr. Peter Salamon, Sacramento, California.)



bone (17,64). Less commonly, the vault of the skull is affected (112). The severe form of craniofacial fibrous dysplasia has been termed "leontiasis ossea," because marked deformities combined with bone enlargement resemble a lion's face (78).

Because of increased uptake on skeletal scintigraphy, radionuclide bone scanning is the most rapid way to determine the distribution of skeletal lesions (69,100) (Fig. 4-37). This examination may also disclose unsuspected sites of involvement (Fig. 4-38). CT can accurately delineate the extent of bone involvement (71) (Fig. 4-39). Tissue attenuation values, as measured by Hounsfield units (HU), are usually within the 70- to 400-HU range (17), apparently reflecting the presence of calcium and microscopic ossification throughout the abnormal tissue. On MRI, fibrous dysplasia exhibits homogeneous, moderately low signal intensity on T1-weighted images, whereas on T2 weighting the signal is bright (116,122) or mixed (77,87,90). After gadolinium infusion, the majority of lesions show central contrast enhancement and some peripheral rim enhancement (90) (Figs. 4-40 and 4-41). In general, signal intensity on T1- and T2weighted images and the degree of contrast enhancement on T1 sequences depend on the amount and degree of bone trabeculae, collagen, and cystic and hemorrhagic changes (90).

Histopathology

The histologic appearance of polyostotic fibrous dysplasia is identical to that of the monostotic form. The presence of small trabeculae of woven bone of various sizes and shapes, scattered within a fibrous tissue without evidence of osteoblastic activity, is diagnostic for this disorder (17,42,47).

Differential Diagnosis

Radiology

If the lesion of fibrous dysplasia contains cartilage (fibrocartilaginous dysplasia) and displays visible calcifications, the differential diagnosis includes *enchondroma* (if the lesion is solitary) or *enchondromatosis* (if the lesion is polyostotic). In exceptional circumstances, a monostotic presentation of fibrous dysplasia may mimic *desmoplastic fibroma* (Fig. 4-42). If a solitary focus is present in the tibia, *osteofibrous dysplasia* and *adamantinoma* are the diagnostic possibilities. In the early phase of development, particularly if the lesion is entirely radiolucent and is situated in the proximal humerus or

Text continues on page 286



Figure 4-37 Polyostotic fibrous dysplasia: scintigraphy. A: Anteroposterior radiograph of the right proximal femur shows several radiolucent lesions particularly in the intertochanteric and subtrochanteric areas. B: Total body radionuclide bone scan demonstrates additional sites of involvement.



Figure 4-38 Polyostotic fibrous dysplasia: scintigraphy. A: Radiograph of the right hip, obtained to exclude a fracture, demonstrates an asymptomatic focus of fibrous dysplasia in the femoral neck. **B**, **C:** To determine other sites of involvement, a radionuclide bone scan was obtained. In addition to the focus in the femoral neck, increased uptake of the tracer was demonstrated at various other sites, but predominantly in the right leg. **D:** Subsequently obtained radiograph of the right lower leg confirmed the presence of multiple foci of polyostotic fibrous dysplasia.

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Figure 4-39 Polyostotic fibrous dysplasia: computed tomography (CT). A: Anteroposterior radiograph of the pelvis shows multiple lesions in the left ilium and proximal left femur. The involvement of the sacrum is not well demonstrated. **B:** CT of the pelvis precisely shows the extent of involvement of the left ilium and the sacrum. **C:** CT of the chest shows foci of fibrous dysplasia in the posterior ribs. **D:** Magnified coned-down CT image of one of the affected ribs shows details of this condition: observe the multiloculated appearance, expansion of the bone, pseudosepta, thinning of the cortex, and a pathologic fracture.



Figure 4-40 Polyostotic fibrous dysplasia: magnetic resonance imaging (MRI). A: Anteroposterior radiograph of the proximal left femur of a 23-year-old woman shows a geographic radiolucent lesion in the subtrochanteric region of the bone. **B:** Coronal T1-weighted MRI shows the full extent of the lesion, which is of low signal intensity. Note that the lesion is much larger than shown by radiography (*continued*).

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Α









Figure 4-41 Polyostotic fibrous dysplasia: magnetic resonance imaging (MRI). A: Anteroposterior radiograph of the proximal right leg of a 23-year-old woman shows a long lesion in the proximal tibia exhibiting a "ground-glass" appearance. The bone is mildly expanded and the cortex is thin. **B:** Coronal T1-weighted MRI shows the lesion to be multifocal with isointense signal similar to that of the skeletal muscles. **C:** Coronal T2-weighted MRI shows heterogeneous signal of the lesion ranging from intermediate to high intensity. **D:** Coronal T1-weighted fat-suppressed MRI after intravenous injection of gadolinium demonstrates slight enhancement of the lesion.



Figure 4-42 Fibrous dysplasia resembling desmoplastic fibroma. Oblique (A) and lateral (B) radiographs of the left lower leg of a 32-year-old woman demonstrate a large, trabeculated radiolucent lesion in the distal tibia. Because of its aggressive features, it was thought to be a desmoplastic fibroma. However, biopsy proved it to be a fibrous dysplasia, a rare lesion at this location in adults.

proximal femur, *a simple bone cyst* should be considered. When the lesion markedly expands the cortex, an *aneurysmal bone cyst* is a possibility. On those rare occasions when the lesion exhibits an eccentric location, *NOF* must be excluded. Monostotic presentation with a thick sclerotic rim at the periphery (rind sign), particularly in the proximal femur, must not be mistaken for

Α

medullary bone infarct (Fig. 4-43) (111). Another lesion that may mimic fibrous dysplasia in this location, particularly in the intertrochanteric region, is so-called *liposclerosing myxofibrous tumor* (96). It is a benign fibroosseous lesion characterized by a complex mixture of histologic elements that include lipoma, fibroxanthoma, myxoma, myxofibroma, fat necrosis, bone, and

B



Figure 4-43 Fibrous dysplasia versus bone infarct. A: Two separate foci of fibrous dysplasia affecting the right femur display a rind sign (proximal lesion) and central calcifications (distal lesion). This presentation resembles a bone infarct. **B:** Bone infarct located in the neck of the right femur exhibits a rind sign. Observe also osteonecrosis of the femoral head (*arrow*). cartilage (111). Radiography usually reveals a welldefined, geographic lytic lesion, with a sclerotic margin and, commonly, with a mineralized matrix (Fig. 4-44). Scintigraphy shows increased uptake of the radiopharmaceutical agent by the lesion (Fig. 4-45B). CT and MRI are useful for further characterization of this tumor and demonstration of its complex morphology (Figs 4-45 and 4-46). On T1-weighted images the lesion is relatively homogeneous, with signal intensity similar to that of skeletal muscle (Fig. 4-45). T2 weighting shows heterogeneous signal with intensity equal to or greater than that of fat (96) (Figs. 4-45D and 4-46B). An interesting lesion that can mimic fibrous dysplasia in the fibula is *pachydysostosis* ("pachy" from the Greek,



Figure 4-44 Liposclerosing myxofibrous tumor of bone. Anteroposterior radiograph of the left hip of an asymptomatic 30-year-old man shows a mostly sclerotic lesion with a narrow zone of transition (*arrows*) in the intertrochanteric region of the femur. The lesion exhibits central amorphous mineralization. (Reprinted with permission from Kransdorf MJ, Murphey MD, Sweet DE. Liposclerosing myxofibrous tumor: a radiologic-pathologic-distinct fibro-osseous lesion of bone with a marked predilection for the intertrochanteric region of the femur. *Radiology* 1999;212:693–698.)

meaning broad). This entity, only recently described (103) in four children of ages 4 weeks to 4 years and affecting exclusively the fibula, consists of widening and elongation of this bone associated with lateral or posterior bowing (Fig. 4-47). Predominantly lytic presentation of fibrous dysplasia affecting the articular end of bone may be mistaken for a *giant cell tumor* (110).

Polyostotic fibrous dysplasia should be mainly differentiated from enchondromatosis and neurofibromatosis. In enchondromatosis, unlike in polyostotic fibrous dysplasia, the lesions may extend into the articular ends of the bone, and columns of radiolucent streaks extending from the growth plate into the metaphyses are characteristic of the former disorder. Neurofibromatosis affecting the skeleton usually exhibits long bone deformities without intramedullary changes typical of fibrous dysplasia. A problem in differential diagnosis may arise if neurofibromatosis is associated with multiple NOFs, so-called Jaffe-Campanacci syndrome, which can mimic the lesion of fibrous dysplasia (30). Faciocranial involvement by fibrous dysplasia can mimic Paget disease; however, the latter entity primarily affects the outer table and spares the facial bones. Another craniofacial disorder, known as cherubism, is frequently mistaken for fibrous dysplasia (61). It is characterized by multiple cystic giant cell lesions symmetrically affecting the mandible and maxilla, appearing in early childhood, with stabilization and remission after puberty. This rare hereditary autosomal dominant disorder is caused by mutations in the c-Abl-binding protein SH3BP2 (99), and represents in fact a form of giant cell reparative granuloma (124). The inherited trait has 100% penetrance in males and 50% to 70% penetrance in females (104). The genetic alterations in this condition have been detected and mapped to chromosome 4p16.3 (121), whereas mutations resulting in this disorder were identified in the gene SH3BP2 (99). Histologic examination reveals a nonneoplastic fibrous tissue with multiple multinucleated giant cells of osteoclastic type, and deposition of collagen (92,104).

Occasionally, osseous changes that are present in *Weismann-Netter-Stuhl syndrome*, a rare form of diaphyseal dysplasia, may mimic those seen in polyostotic fibrous dysplasia.

Pathology

Although in most cases fibrous dysplasia exhibits a very characteristic histologic picture, occasionally it can mimic *fibroblastic osteosarcoma* and *fibrosarcoma*. In *osteosarcoma* the cells and nuclei are usually distinctly pleomorphic and mitoses are frequent. In addition, tumor bone formation is more amorphous, without a distinctive trabecular structure. However, low-grade central osteosarcoma may present a real challenge to the pathologist (see the section "Osteosarcoma"). *Adamantinoma* may resemble fibrous dysplasia when it is accompanied by abundant fibroblastic tissue. However, the distinct epithelial cells of the former [which in more obscure cases can be identified using immunohistochemistry (IHC) with epithelial markers] are characteristic and diagnostic.

When the formation of bone is limited, as in some cases of fibrous dysplasia, there may be a problem with



Figure 4-45 Liposclerosing myxofibrous tumor of bone: scintigraphy and magnetic resonance imaging (MRI). A: Anteroposterior radiograph of the left hip of a 39-year-old man shows a round lesion in the intertrochanteric region of the femur with a thin, well-defined sclerotic border (*arrows*). There are foci of amorphous mineralized matrix within the lesion. B: Radionuclide bone scan obtained after intravenous injection of 99mTc methylene diphosphonate (MDP) shows a mildly increased uptake of radiopharmaceutical agent (*arrowheads*). C: Coronal T1-weighted MRI shows the lesion (*arrows*) to exhibit intermediate signal intensity similar to that of skeletal muscles. D: Coronal T2-weighted MRI shows the lesion (*arrows*) displaying predominantly high signal intensity with foci of intermediate signal medially. (Reprinted with permission from Kransdorf MJ, Murphey MD, Sweet DE. Liposclerosing myxofibrous tumor: a radiologic-pathologic-distinct fibro-osseous lesion of bone with a marked predilection for the intertrochanteric region of the femur. *Radiology* 1999;212:693–698.)



Figure 4-46 Liposclerosing myxofibrous tumor of bone: magnetic resonance imaging (MRI). A: Anteroposterior radiograph of the left hip of a 38-year-old woman presenting with vague hip pain shows a radiolucent lesion with a well-defined thick sclerotic border (*arrows*) in the intertrochanteric region of the femur. **B:** Coronal T2-weighted MRI shows the lesion (*arrows*) to exhibit heterogeneous signal intensity; however, sclerotic rind displays signal void. (Reprinted with permission from Kransdorf MJ, Murphey MD, Sweet DE. Liposclerosing myxofibrous tumor: a radiologic-pathologic-distinct fibro-osseous lesion of bone with a marked predilection for the intertrochanteric region of the femur. *Radiology* 1999;212:693–698.)

A

Figure 4-47 Pachydysostosis of the fibula. Anteroposterior **(A)** and lateral **(B)** radiographs of the left leg of a 1-year-old boy show medullary radiolucency of the distal fibula, thinning of the cortex, as well as widening and bowing deformity of the bone, resembling abnormalities seen in fibrous dysplasia. (Courtesy of Dr. Peter Salamon, Sacramento, California.) the differential diagnosis of *NOF* (108). In such instances, the absence of groups of osteoclast-like giant cells and of hemosiderin may be helpful. If in a biopsy specimen only the fibrous component is present, differentiation of fibrous dysplasia from *desmoplastic fibroma* may be impossible on histologic grounds only. However, when undecalcified or EDTA decalcified tissue is available, molecular analysis will reveal the mutation of the *GNAS1* gene.

The radiologic and pathologic differential diagnosis of monostotic and polyostotic fibrous dysplasia is depicted in Figure 4-48A and B, respectively.



Figure 4-48 Radiologic and pathologic differential diagnosis of monostotic (A) and polyostotic (B) fibrous dysplasia.

Osteofibrous Dysplasia (Kempson-Campanacci Lesion)

Osteofibrous dysplasia or Kempson-Campanacci lesion (formerly called ossifying fibroma) is a rare disorder with a decided preference for the tibia (129,142,143,153). It is characterized by the presence of fibrous connective tissue and trabeculae of immature nonlamellar bone rimmed by osteoblasts.

Clinical Presentation

With rare exceptions, the lesion is situated in the proximal or middle third of the tibia and is often localized to the anterior cortex (128,129). In more than 80% of patients, some degree of anterior bowing is observed. When the fibula is affected, the distal third of the diaphysis is usually involved (131,137). This lesion occurs predominantly in childhood (>60% of the patients affected are younger than 5 years), but occasionally it is first discovered in adolescence. Boys are slightly more commonly affected (129).

Osteofibrous dysplasia is usually asymptomatic but occasionally may be revealed clinically by the enlargement and bowing of the tibia (154). The disorder may sometimes be locally aggressive (150). It frequently recurs after local excision and, according to some authors, may coexist with the malignant lesion adamantinoma (125,147,149). Other investigators have suggested that osteofibrous dysplasia may be a precursor to adamantinoma (132,151,152). Weiss and Dorfman (155), and later Czerniak et al. (133), suggested that adamantinoma and osteofibrous dysplasia are on a continuum, with osteofibrous dysplasia on one end and adamantinoma on the other.

Recently familial occurrence has been described (138,140). Cytogenetic studies in osteofibrous dysplasia revealed trisomies for chromosomes 7, 8, 12, and 22 (126,127). Similar findings in adamantinomas of long bones point to a relationship of both conditions (136). Mutations of the *GNAS1* gene in fibrous dysplasia, however, have not been detected, thus genetically separating osteofibrous dysplasia from fibrous dysplasia (146).

Imaging

Radiographically, the Kempson-Campanacci lesion bears a striking similarity to NOF and fibrous dysplasia. It is radiolucent and possesses lobulated sclerotic margins, usually confined to the cortex (Fig. 4-49). Larger lesions may destroy the cortical bone and invade the medullary cavity (156) (Fig. 4-50). In the tibia the lesion rarely involves the entire circumference of the shaft, but in the fibula this may occur (125). Rarely, there are multiple osteolytic lesions over a large part of the diaphysis, simulating adamantinoma.

MRI findings in osteofibrous dysplasia differ in the reported cases (134,135). However, T1-weighted images generally show heterogeneous but predominantly low signal intensity of the lesion, whereas T2-weighted sequences show intermediate-to-high signal. After administration of gadolinium, significant enhancement of the lesion is noted (Fig. 4-51).



Figure 4-49 Osteofibrous dysplasia. The lesion in the anterior aspect of the right tibia of a 14-year-old girl was originally diagnosed as nonossifying fibroma. Although it is similar to a nonossifying fibroma and fibrous dysplasia, its site is typical of osteofibrous dysplasia, which was confirmed by biopsy.



Figure 4-50 Osteofibrous dysplasia. A, B: A cortical lesion in the distal tibia invades the medullary portion of the bone in a 2-year-old boy.

Histopathology

Osteofibrous dysplasia and fibrous dysplasia, as the similarity in their names suggests, have a remarkable histopathologic resemblance. Like fibrous dysplasia, osteofibrous dysplasia is characterized by a fibrous background containing a rather haphazard arrangement of incomplete trabeculae (145). In osteofibrous dysplasia, however, these bony trabeculae generally show appositional activity by polarized osteoblasts ("dressed trabeculae") (Fig. 4-52A,C). Such activity, which can sometimes be seen at the earliest stages of bone development, more often occurs surrounding larger trabeculae (144). Viewed under polarized light, these trabeculae consist of an inner portion of woven bone surrounded by an outer zone of lamellar transformation (Fig. 4-52B), a pattern sometimes referred to as zonal architecture (128,129). Because osteofibrous dysplasia and fibrous dysplasia exhibit almost identical radiographic and very similar histologic features, with the exception of the former lesion's predilection for the cortex of the tibia, the differences in the light and electron microscopy findings must be meticulously sought. Nevertheless, some investigators have speculated that osteofibrous dysplasia represents a regional or localized presentation of fibrous dysplasia (130). This, however, is excluded in view of the histomorphologic and molecular genetic findings (146).

Sissons et al. (148) reported two cases of fibroosseous lesions containing smaller aggregates of osseous material resembling cementum. They proposed the term "ossifying fibroma" for this lesion. The term "osteofibrous dysplasia," they suggested, should be reserved for fibrous cortical lesions of the tibia and fibula. To avoid confusion in terminology, the differential features of the various lesions are summarized in Table 4-2.

Differential Diagnosis

Radiology

The main differential possibilities include fibrous dysplasia and NOF. If the lesion is confined to the cortex and there is associated anterior bowing of the tibia, osteofibrous dysplasia is more likely. Conversely, particularly in older children and adolescents, if the lesion extends into the medullary portion of the bone and is bordered with a sclerotic rim, NOF is a more probable diagnosis. Centrally located lesions that exhibit thinning of the cortex rather than an intracortical multiloculated bubbly appearance more likely represent fibrous dysplasia. Finally, osteofibrous dysplasia should be differentiated from adamantinoma (139). The latter lesion is usually more extensive, with a more aggressive appearance. A helpful hint is that patients with adamantinoma are much older than those with osteofibrous dysplasia.

Pathology

The main differential diagnoses include fibrous dysplasia and adamantinoma. Fibrous dysplasia shows the characteristic absence of osteoblasts at the surface of trabeculae, whereas osteofibrous dysplasia exhibits dressed trabeculae. Adamantinoma is distinguished from osteofibrous dysplasia by the presence of single or small nests of epithelial cells within the fibroosseous stroma (141). More problems are encountered in distinguishing osteofibrous dysplasia from so-called osteofibrous dysplasia-like adamantinoma (149). From a practical standpoint, a diagnosis of osteofibrous dysplasia made on tissue removed by a small incisional biopsy cannot be certain because the biopsy material may not be representative of the entire lesion. One cannot rule out an osteofibrous dysplasia-like adamantinoma, which, according to the WHO, is characterized by isolated or small groups of epithelial cells intermingled with stromal cells (149,153). In such cases, immunohistochemical staining for markers of epithelial cells, especially cytokeratins, can be helpful.

Radiologic and pathologic differential diagnosis of osteofibrous dysplasia is depicted in Figure 4-53.

Desmoplastic Fibroma

Originally described by Jaffe in 1958, desmoplastic fibroma, also known as desmoid tumor of bone, is a rare, locally aggressive lesion, closely related to the



Figure 4-51 Osteofibrous dysplasia: magnetic resonance imaging (MRI). A: Sagittal T1-weighted MRI shows an oblong lesion involving the anterior cortex of the tibia, which exhibits heterogeneous signal intensity (*arrows*). **B, C:** Sagittal and axial T1weighted fat-suppressed sequences obtained after intravenous injection of gadolinium (Gd-DTPA) show significant enhancement of the lesion. (**A** and **B** reprinted from Greenspan A. *Orthopedic imaging – a practical approach,* 4th ed. Philadelphia: Lippincott Williams & Wilkins, 2004:642, Fig. 19.30.)



В







Figure 4-52 Histopathology of osteofibrous dysplasia. A: Plump, irregular bone trabeculae scattered in a moderate-density spindle cell stroma are made up in part of a cell-rich woven bone, but at the periphery display lamellar bone rimmed by osteoblasts (hematoxylin and eosin, original magnification $\times 25$). **B:** Viewed under polarized light, lamellar (*light green*) and woven (*darker areas*) bone is more clearly discernible (polarized light, original magnification $\times 25$). **C:** Under higher magnification osteoblastic rimming of the bone trabeculae is clearly visible (hematoxylin and eosin, original magnification $\times 200$).

B

desmoid tumor of abdominal wall, characterized by the formation of abundant collagen fibers by the tumor cells (19,45,160,167). Although generally classified as a benign bone tumor, some investigators classify this lesion in a so-called intermediate category between benign and malignant lesions (29). As in extraosseous desmoid-type fibromatosis and in extraabdominal (aggressive) fibromatosis, trisomies 8 and 20 have been observed in desmoplastic fibroma of bone; however, thus far their significance is not clear (160,163).

Clinical Presentation

Desmoplastic fibroma accounted for only about 0.06% of all bone tumors and 0.3% of benign bone tumors in the Mayo Clinic series (8). It usually arises in individuals younger than 40 years; half of the cases occur during the second decade (8). There is no sex predominance (Fig. 4-54). The clinical signs and symptoms are nonspecific (157). Because the onset is often insidious, the lesion tends to be relatively large before the patient begins to experience symptoms (108). In symptomatic patients, pain or swelling, sometimes of long duration, is the most common complaint (162). Rarely (in about 10% of cases), a pathologic fracture through the lesion

is the first presenting symptom (172). The long bones (humerus, femur, tibia), mandible, and pelvis are the most common sites of involvement (171). In the long bones the lesion is diaphyseal, sparing the epiphysis, but it often extends into the metaphysis and may even extend into the articular end of the bone after closure of the growth plate (174).

Imaging

There are no characteristic radiographic features of desmoplastic fibroma. The spectrum of radiologic features of this tumor overlap the radiographic patterns described for low-grade central osteosarcoma and, in some cases, fibrous dysplasia (161). In general, it is an expansive, radiolucent lesion, usually with nonsclerotic but sharply defined borders (Fig. 4-55A). Occasionally a wide zone of transition is present (169). The cortex is either thinned or thickened, but there is no significant periosteal response (159) (Fig. 4-55B). Internal septation or trabeculation is observed in approximately 75% of cases, yielding a honeycomb pattern of bone destruction (164) (Fig. 4-55C). Locally aggressive variants, which may simulate malignant bone tumors, are marked by bone destruction and invasion of the soft tissues.

Sex	Age	Location	Radiographic Appearance	Histopathology
M/F	Any age	Fibr	ous Dysplasia Radiolucent, ground glass, or	Woven (nonlamellar) type
,.	(monostotic) First to third decades (polyostotic)	Long bones Pelvis Ends of bones usually spared Polyostotic: unilateral in skeleton (frequent)	smoky lesion Thinning of cortex with endosteal scalloping "Shepherd's crook" deformity Accelerated growth	of bone in loose to dense fibrous stroma; bone trabeculae lacking osteoblastic activity ("naked trabeculae")
		Nono.	ssifying Fibroma	
M/F	First to third decades	Long bones (frequently posterior femur)	Radiolucent, eccentric lesion Scalloped, sclerotic border	Whorled pattern of fibrous tissue containing giant cells, hemosiderin, and lipid-filled histiocytes
		Osteofibrous Dysplasic	a (Kempson-Campanacci Lesion)	
M/F	First to second decades	Tibia (frequently anterior aspect) Fibula Intracortical (frequent)	Osteolytic, eccentric lesion Scalloped, sclerotic border Anterior bowing of long bone	Woven and mature (lamellar) type of bone surrounded by cellular fibrous spindle cell growth in whorled or matted pattern; bone trabeculae rimmed by differentiated osteoblasts ("dressed trabeculae")
		Ossifyir	ng Fibroma of Jaw	
F	Third to fourth decades	Mandible (90%) Maxilla	Expansive radiolucent lesion Sclerotic, well-defined borders	Uniformly cellular fibrous spindle cell growth with varying amounts of woven bone formation and small, round cementum-like bodies
		Ossifying Fil	broma (Sissons Lesion)	
M/F	Second decade	Tibia Humerus	Radiolucent lesion Sclerotic border Similar to osteofibrous dysplasia	Fibrous tissue containing rounded and spindle- shaped cells with scant intercellular collagen and small, partially calcified spherules resembling cementum-like bodies of ossifying fibroma of jaw
		Liposclerosir	ng Myxofibrous Tumor	
M/F	Second to seventh decades	Intertrochanteric region of femur	Radiolucent or partially sclerotic lesion with well- defined sclerotic border, occasional central matrix mineralization	Fibrous or myxofibrous areas with metaplastic curvilinear or circular woven bone ossicles and/or dystrophic mineralization in necrotic fat

Table 4-2 Differential realures of various ribiousseous Lesions with similar natiourabilit Abbearain	Table 4-2	Differential Features of	Various Fibroosseous	Lesions with Simila	r Radiographic Appearang
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In addition to radiography, radiologic evaluation of desmoplastic fibroma should include bone scintigraphy, CT, and MRI. Radionuclide bone scan shows an increase in the uptake of the radiopharmaceutical agent at the site of the lesion (178). CT is useful for evaluating cortical breakthrough and tumor extension into the soft tissues (161,177,179). MRI, also helpful in assessing intra- and extraosseous extension, can further characterize

the tumor (Figs. 4-56 and 4-57). The lesion appears well defined on MR images, exhibiting an intermediate signal intensity on T1 weighting and a heterogeneous pattern on T2 weighting, marked by an area of increased signal intensity mixed with foci of intermediate and low signal intensity (159,164,175). The hypointensity of the signal reflects the dense connective tissue matrix and relative acellularity of the tumor (178).

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Figure 4-53 Radiologic and pathologic differential diagnosis of osteofibrous dysplasia.

Histopathology

Histologically, the lesion is composed of very regular bundles of normal-looking, spindle-shaped, and occasionally stellate fibroblasts with elongated or ovoid nuclei interspersed within a densely collagenized matrix (Fig. 4-58). There is almost no evidence of mitotic activity (166). Although cellularity is variable, the collagenous matrix almost always constitutes the greater portion of the tumor. For this reason, the biopsy specimen, resembling scar tissue, is identical in appearance to a desmoid tumor of soft tissue. Large, thin-walled vessels similar to those observed in soft tissue desmoid tumors are usually apparent (35). On purely histologic grounds, the lesion most closely resembles periosteal desmoid, but correlation with the radiographs clarifies its true nature. As with desmoids of soft tissue, in desmoplastic fibroma of bone it is difficult to histologically differentiate the tumor tissue from the surrounding normal connective tissue. Recently, it has been shown that beta-catenin, a protein that may act as a modulator of transcription in the nucleus, is overexpressed in desmoid-type fibromatosis owing to a mutation in the *beta-catenin* gene or its regulating gene, the APC or adenoma polyposis coli gene. This gene is mutated in familial adenomatous polyposis syndrome and Gardner syndrome, both of which are associated with aggressive fibromatosis (168,170,173). Hauben et al. (165) were able to demonstrate that beta-catenin immunoreactivity was also present in 6 of 13 investigated cases of desmoplastic fibroma. However, mutations of the beta-catenin gene were absent.

Differential Diagnosis

Radiology

Desmoplastic fibroma is frequently difficult to distinguish radiographically from similar-appearing lesions. These include *simple bone cyst, aneurysmal bone cyst, fibrous dysplasia, chondromyxoid fibroma, giant cell tumor, low-grade central osteosarcoma, fibrosarcoma, MFH,* and *chondrosarcoma* (169,171,172,175,178). Giant cell tumor, fibrosarcoma, and MFH can be excluded if the lesion possesses sclerotic margins. A purely lytic appearance of desmoplastic fibroma excludes simple and aneurysmal bone cyst and chondromyxoid fibroma, as these entities possess sclerotic borders.

Pathology

Histopathologic differential diagnosis must include *NOF* and *fibrous dysplasia* (158). However, unlike the former, abundant intercellular substance is observed, and giant cells and lipid-bearing histiocytes are absent (29); lack of metaplastic bone formation distinguishes desmoplastic fibroma from the latter. In addition, the structure of the fibrous tissue in NOF, with its typical cartwheel or storiform pattern, as well as its content of giant cells and foam cells, makes the distinction easy. In equivocal cases, absence of *GNAS1* mutations and/or presence of beta catenin immunoreactivity rules out fibrous dysplasia. A very cellular lesion may be confused with *low-grade fibrosarcoma* or *MFH*, although the absence of mitotic activity and the lack of nuclear hyperchromatism and atypia in desmoplastic fibroma may help in this differ-



Figure 4-54 Desmoplastic fibroma: skeletal sites of predilection, peak age range, and male-to-female ratio.

entiation. In fibrosarcoma, the cells are usually more densely arranged, and slight pleomorphism and hyperchromatism of the nuclei should point to the correct diagnosis, although in a low-grade tumor the distinction may be very difficult. If the radiographs are confusing, the distinction usually can be made histologically on the basis of the quantity of collagen, the lack of nuclear atypia and hyperchromatism, and the paucity of mitotic activity in desmoplastic fibroma.

Fibroblastic osteosarcoma can be distinguished from desmoplastic fibroma by the tumor bone production in the former tumor.

The distinction between *desmoid of soft tissues* and desmoplastic fibroma of bone must be made on the basis of the clinical and radiologic features (169). In addition, mutational analysis of the *beta-catenin* or *APC* gene may be helpful because these are altered in desmoid-type fibromatosis.

The radiologic and pathologic differential diagnosis of desmoplastic fibroma is depicted in Figure 4-59.

Malignant Tumors

Fibrosarcoma and Malignant Fibrous Histiocytoma

Fibrosarcoma and malignant fibrous histiocytoma (MFH) are malignant tumors with similar radiographic presentations and similar histologic features. Fibrosarcoma is a malignant tumor characterized by the formation of interlacing bundles of collagen fibers by spindle-shaped tumor cells, and by lack of bone or cartilage formation. MFH is a malignant tumor with a pleomorphic spindle-cell structure that produces various amounts of collagen fibers but lacks any specific pattern of histologic differentiation (28,29,46,182,233). According to previous studies, there are no essential differences in the radiologic features, clinical behavior, and survival data for these tumors (185,189,192,193,210). Some authors therefore consider them to represent a single group (227,240). Nevertheless, the existence of MFH as a separate tumor entity has been questioned. With electron microscopy and increasing use of immunohistochemical and genetic investigations, it has become obvious that, at least in the soft tissues, tumors initially classified as MFH can be recognized as pleomorphic variants of other sarcomas, e.g., leiomyosarcomas, liposarcomas, fibrosarcomas, or extraskeletal osteosarcomas (196,200,201,223). However, because some pleomorphic sarcomas remain unclassified, even with the application of all available diagnostic techniques, the WHO retained the term MFH (synonymously used with undifferentiated high-grade pleomorphic sarcoma) for tumors that do not exhibit any line of differentiation (202).

Antonescu et al. (182) conducted a comparative ultrastructural study of fibrosarcomas and MFH of bone. These authors proposed reclassification of MFH as pleomorphic fibrosarcoma. Recent studies, however, have provided evidence for genetically based differ-







Figure 4-55 Desmoplastic fibroma. A: Radiolucent lesion with nonsclerotic, sharply defined borders is seen in a supraacetabular region of the right ilium *(arrows).* (Reprinted from Bullough PG, *Atlas of orthopedic pathology,* 2nd ed. New York: Gower, 1992:17.2.) **B:** Radiolucent, trabeculated lesion occupies the proximal end of the right fibula in a 17-year-old girl. Note lack of periosteal reaction. **C:** Radiolucent, trabeculated lesion is present in the metadiaphyseal region of the distal radius in a 15-year-old boy.



Figure 4-56 Desmoplastic fibroma. A: Anteroposterior radiograph of the pelvis in a 67-year-old man demonstrates an expansive, lytic lesion that involves the left ischium and pubis and extends into the supraacetabular portion of the ilium. **B:** Conventional tomography shows the lytic nature of the tumor and its expansive character. The involvement of the ilium is better demonstrated. **C:** Computed tomography section through the hip joint shows a lobulated appearance of the tumor and a thick, sclerotic margin. **D:** Axial T2-weighted (SE, TR 2000, TE 80) magnetic resonance imaging demonstrates nonhomogeneity of the signal from tumor. The bulk of the lesion displays low to intermediate signal intensity with central areas of high signal.



Figure 4-57 Desmoplastic fibroma: magnetic resonance imaging (MRI). A: Coronal T1-weighted MRI shows the lesion in the left femoral shaft breaking through the cortex and extending into the soft tissues (*arrows*). B: Axial proton density image demonstrates replacement of the bone marrow by the tumor, extension into the soft tissues, and peritumoral edema.



ences between fibrosarcoma of bone and MFH of bone. Hattinger and her group (206) analyzed seven fibrosarcomas of bone by comparative genomic hybridization and DNA microarrays. Gain of chromosome 22q was the most common detectable aberration. With DNA microarray technique and also by IHC, gains of the *platelet-derived growth factor beta (PDGF-B)* gene, localized at 22q12.3–q13.1, and expression of the PDGF-B protein could be demonstrated.

With respect to MFH of bone, in 1973 Arnold (183) described a family presenting diaphyseal necroses in the shaft of long tubular bones with subsequent development of sarcomas. These hereditary tumors, initially di-

agnosed as fibrosarcomas and later reclassified as MFH, are characterized by loss of heterozygosity at chromosome 9p21-22 (205,219). In sporadic MFH of bone, the same alteration was observed in five of seven analyzed cases, leading to the hypothesis that, in the pathogenesis of MFH, bone alterations of a putative tumor suppressor gene located at 9p21-22 may be involved (220).

В

Both fibrosarcoma and MFH can be either primary tumors or secondary to a preexisting benign condition, such as Paget disease, fibrous dysplasia, bone infarct, or chronic draining sinuses of osteomyelitis (183,194,203,204,211,212,224,225). These lesions may also arise in bones that have been previously irradi-



Figure 4-58 Histopathology of desmoplastic fibroma. A: Densely collagenized hypocellular stroma contains spindleshaped fibroblasts with uniform-appearing nuclei (hematoxylin and eosin, original magnification \times 40). **B:** At higher magnification the cells producing the abundant collagen show no evidence of atypia (hematoxylin and eosin, original magnification \times 150).



Figure 4-59 Radiologic and pathologic differential diagnosis of desmoplastic fibroma.

ated (226,234). Such lesions are termed secondary fibrosarcomas (or secondary MFH).

Rarely, fibrosarcoma can arise in a periosteal location (periosteal fibrosarcoma). However, some investigators postulate that these lesions represent primary soft tissue tumors abutting the bone and invading the underlying periosteum (185).

Clinical Presentation

Fibrosarcoma and MFH usually occur in the third to sixth decades of life and represent about 5% and 2%, respectively, of all malignant bone tumors (213,236). Their sites of predilection are the femur, humerus, tibia, and pelvic bones (197,210,246) (Fig. 4-60). The most common clinical symptoms are pain and localized swelling, ranging in duration from a few weeks to several months (199). In about 23% of reported cases, a pathologic fracture leads to incidental detection of the tumor (213,235).

Imaging

Radiographically, both lesions are marked either by an osteolytic area of destruction surrounded by a wide zone of transition or by a moth-eaten or permeative type of bone destruction (204). There is little or no reactive sclerosis, and usually no periosteal reaction (189) (Figs. 4-61 and 4-62). The lesions are often eccentrically located, close to or within the articular end of the bone. Pathologic fractures are not uncommon. A soft tissue mass is frequently present (198). These lesions may resemble giant cell tumor of bone or telangiectatic os-

teosarcoma, and they may be mistaken for metastatic carcinoma because of the age group in which they usually occur. Some authorities believe that an almost pathognomonic sign of fibrosarcoma is its tendency to allow the preservation of small sequestrum-like fragments of cortical bone and spongy trabeculae, which can be demonstrated on conventional radiographs or CT examination. On scintigraphy, both lesions typically exhibit an area of increased uptake (186,217), often around the periphery of the tumor (17).

On CT examination, both fibrosarcoma and MFH show a predominant density similar to that of normal muscle (207,232) and exhibit the nonspecific tissue attenuation values of Hounsfield units encountered in most nonmineralized tissues. Hypodense areas reflect areas of necrosis within tumor. MRI is useful to outline the intraosseous and extraosseous extension of these tumors, but there are no characteristic MRI findings for either one (181,186,207,229) (Fig. 4-63). Some investigators found the signal characteristics comparable to those of other lytic bone tumors (230,239). Signal intensity is intermediate to low on T1-weighted images and high on T2 weighting, frequently heterogeneous and varying with the degree of necrosis and hemorrhage within the tumor (17). Peripheral enhancement was noted after intravenous injection of Gd-DTPA (218).

Histopathology

The histologic criteria for the diagnosis of MFH for soft tissue lesions were outlined by Stout and Lattes in 1967 and by Kempson and Kyriakos in 1972, and



Figure 4-60 Fibrosarcoma and malignant fibrous histiocytoma: skeletal sites of predilection, peak age range, and male-to-female ratio.

have been applied to tumors arising in bone (184,193, 195,209,215,221,228,237). These include (a) bundles of fibers and of spindle-shaped, fibroblast-like cells arranged in a cartwheel or storiform pattern, exhibiting mitotic activity and features of atypia; (b) rounded and foam cells with histiocytoid features, exhibiting ovoid, and often grooved or indented nuclei and well-defined cytoplasmic borders; cells may contain bizarre and frequently pleomorphic multiple nuclei; some phagocytic activity with cytoplasmic inclusions of red blood cells, hemosiderin, and/or lipids is present; (c) typical and atypical giant cells of the osteoclastic type; and (d) a relatively conspicuous infiltration of inflammatory cells, predominantly lymphocytes (29,236,243).

Fibrosarcomas are graded from 1 to 4, depending on their cellularity and their nuclear atypia. The higher grade lesions are very cellular and show a greater mitotic activity, nuclear hyperchromatism, and variation in nuclear size and shape (188,191). The low-grade lesions are less cellular, show only slight anaplasia of the collagenproducing spindle cells, and demonstrate considerable fibrogenesis (231). Other investigators divide these tumors into three grades: well differentiated, moderately differentiated, and poorly differentiated (29,241). In well-differentiated tumors, the cells are elongated and spindle-shaped, with ovoid or elongated nuclei (187). Sometimes nuclei are hyperchromatic and plump, but no mitotic activity is present. The tumor cells are sparse compared with the relatively abundant intercellular collagen fibers, which are sometimes dense and hyalinized. The poorly differentiated type of tumor is highly cellular, with a marked cellular atypia and a high mitotic activity. The cells possess one or more bizarre, hyperchromatic nuclei. The intercellular matrix is also sparse and occasionally consists only of reticulin fibers (29). Areas similar to MFH are also present.

A rare variant, sclerosing epithelioid fibrosarcoma of bone, consisting of small epithelioid cells arranged in strands or nests and containing deeply eosinophilic thick collagenous bands, has recently been described (180,190).

Although both fibrosarcoma and MFH are composed of sarcomatous spindle cells that form variable amounts of collagen, each tumor has certain distinguishing features (196). The spindle-shaped cells of fibrosarcoma are largely arranged in long fascicles. These fascicles usually run at right angles to one another, so that any plane of section shows some bundles of tumor cells running transversely and others running longitudinally (Fig. 4-64A, B). In some areas of sectioning, all bundles may lie longitudinally, forming a "herringbone tweed" pattern in their spatial relationship (192) (Fig. 4-64C, D). The spindle cells have hyperchromatic nuclei with an increased ratio of nucleus to cytoplasm. Mitotic activity, including bizarre mitoses, may be present. Collagen production is usually evident in parts of the neoplasm, but it tends to be sparse compared with that of the benign fibroblastic tumors. It is important that there is no evidence of osteoid or bone formation, a feature that distinguishes fibrosarcoma from fibroblastic variants of osteosarcoma (242).



Figure 4-61 Fibrosarcoma. A: Oblique radiograph of the right knee of a 28-year-old woman shows a purely destructive osteolytic lesion in the intercondylar fossa of the distal femur (*arrows*). Note the absence of reactive sclerosis and lack of periosteal reaction. **B:** Anteroposterior radiograph of the left proximal humerus in a 62-year-old man shows an osteolytic lesion in the shaft of the bone with a pathologic fracture. A metastatic lesion was suspected but biopsy revealed a fibrosarcoma.



Figure 4-62 Malignant fibrous histiocytoma. A destructive osteolytic lesion in the distal femoral shaft shows poorly defined margins. (Reprinted from Greenfield GB, *Imaging of bone tumors.* Philadelphia: JB Lippincott Company, 1995: 113.)

In contrast to the long fascicles of tumor cells in fibrosarcoma, the spindle cells of MFH tend to be arranged in sweeping, curved fascicles and expanding nodules (221). Their histologic appearance has been compared to pinwheels, spiral nebulae, or cut, matted straw (storiform pattern) (193) (Fig. 4-65). The cells of this tumor are spindle-shaped, sometimes bearing a histiocytoid appearance, and are not so monotonous in shape as pleomorphic fibroblasts of fibrosarcoma (Fig. 4-66). The tumor cells may exhibit phagocytic activity. Although predominantly spindle-shaped, the tumor cells may also be round or polyhedral. In addition to the usual hyperchromatic nuclei, vesicular nuclei (similar to those in histiocytes) may be identified. Giant cells, sometimes in large numbers, may also be present. Areas of necrosis are a common finding, and it is not unusual to observe a background with scattered chronic inflammatory cells. Occasionally, collagen production by the fibroblastic type of tumor cells of MFH may be seen in broader hyaline-appearing bands resembling osteoid, which may lead to diagnostic confusion with osteosarcoma. In such cases, experience is necessary to make the differentiation from true osteosarcoma (36,242). Immunohistochemical studies are not helpful in the diagnosis of MFH (238,244,245); however, they may help to rule out other sarcomas, sarcomatoid carcinomas, and melanomas.

Huvos et al. (208,209) subdivide the histologic pattern seen in MFH into several variants, according to the

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predominant pathologic appearance, although in clinical practice these patterns frequently overlap: (a) a predominantly fibroblastic pattern (Fig. 4-67A), in which the fibroblast-like spindle cells are arranged in whorls of cartwheel or storiform pattern with only occasional giant cells of the osteoclastic type; (b) a predominantly histiocytic or xanthomatous morphologic variant, in which giant cells of the osteoclastic type, frequently with foamy cytoplasm, predominate (Fig. 4-67B); and (c) the malignant giant cell tumor type, in which there is an intimate admixture of fibroblasts, mononuclear histiocytes, and giant cells of the osteoclastic type (Fig. 4-67C).



Figure 4-63 Malignant fibrous histiocytoma: magnetic resonance imaging (MRI). A: Oblique radiograph of the right femur of a 16-year-old girl shows fusiform thickening of the cortex and permeative type of medullary bone destruction. **B:** Radionuclide bone scan (⁹⁹Tc-MDP) shows increased uptake of the tracer in the right femur. **C:** Coronal T1-weighted (SE, TR 500, TE 20) MRI demonstrates the extent of the tumor, which involves about 75% of the length of the femur. **D:** Coronal T2-weighted (SE, TR 2000, TE 80) MRI shows that the tumor exhibits high signal intensity. The soft tissue extension medially is also accurately depicted.



Figure 4-64 Histopathology of fibrosarcoma. A: Interwoven bundles of spindle cells with a moderate degree of pleomorphism characterize this grade 2 fibrosarcoma (hematoxylin and eosin, original magnification ×25). **B:** With a reticulin fiber stain the uneven distribution of the fibers is obvious (Novotny reticulin stain, original magnification ×50). **C:** The cells and fiber bundles are cut longitudinally (*center*) and across the axis (*left and right*), producing a herringbone pattern (Giemsa, original magnification ×50). **D:** At high magnification the herringbone pattern is conspicuous (hematoxylin and eosin, original magnification ×400).



Figure 4-65 Histopathology of malignant fibrous histiocytoma. Marked nuclear pleomorphism of malignant fibroblasts and scattered giant cells form the typical storiform or "starry-night" pattern (hematoxylin and eosin, original magnification \times 50).



Α

Figure 4-66 Histopathology of malignant fibrous histiocytoma. A: Moderately cellular tumor, predominantly composed of spindle cells, exhibits an allusively storiform pattern. Some preserved fat cells are seen within the tumor tissue (*upper right*) (Giemsa, original magnification \times 50). **B:** At higher magnification the pleomorphism of the spindle cells is obvious. In the lower center is seen one preserved fat cell (Giemsa, original magnification \times 100).

Differential Diagnosis

Radiology

Because pleomorphic fibrosarcoma and MFH have an almost identical radiologic appearance, both lesions are considered here as one in the differential diagnosis. All purely lytic lesions without marginal sclerosis should be considered in the differential diagnosis, including *giant cell tumor, lymphoma, solitary myeloma, metastasis,* and *brown tumor of hyperparathyroidism. Giant cell tumor* must be particularly considered in lesions affecting the articular end of a bone (Fig. 4-68). Occasionally, *chondrosarcoma* with





Figure 4-67 Histopathology of malignant fibrous histiocytoma. A: Fibroblastic pattern with typical storiform arrangement (van Gieson, original magnification ×25). **B:** Histiocytic pattern with giant cells (hematoxylin and eosin, original magnification ×25). **C:** Giant cell–rich pattern (hematoxylin and eosin, original magnification ×250).



Figure 4-68 Malignant fibrous histiocytoma. Anteroposterior radiograph of the left knee (A) and magnification study in the oblique projection (B) show an expanding, lytic lesion in the proximal end of the fibula in a 13-year-old girl. There is a buttress of periosteal new bone formation at the distal extent of the tumor. A giant cell tumor or an aneurysmal bone cyst had been suspected, but a biopsy revealed a malignant fibrous histiocytoma.



Figure 4-69 Secondary malignant fibrous histiocytoma. A: Anteroposterior radiograph of both knees in a 39-year-old woman with known multiple idiopathic bone infarcts shows the typical appearance of this condition in the distal femora. In the left femur there is, however, evidence of a lamellated periosteal reaction along the lateral cortex (*arrowheads*). **B:** Magnification study shows cortical destruction (*arrows*). On biopsy the suspicious lesion proved to be a malignant fibrous histiocytoma complicating a bone infarct (*continued*).

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Figure 4-69 *Continued* **C:** Radiograph of the resected specimen shows the aggressive periosteal reaction and a soft tissue mass more clearly. Note typical serpentine coarse calcifications of the medullary bone infarct.

С

minimal or no central calcification and lack of peripheral sclerosis may look like fibrosarcoma or MFH. In lesions located in the tibia, *adamantinoma* should be excluded. In the younger age group, osteolytic (either fibroblastic or telangiectatic) *osteosarcoma* may be a differential consideration and for patients in the first two decades, *Ewing sarcoma*.

The presence of calcifications at the periphery or within the tumor is strongly suggestive of secondary fibrosarcoma or secondary MFH resulting from malignant complications of a bone infarct (Fig. 4-69).

Pathology

Because pleomorphic fibrosarcoma and MFH appears very similar, the differential diagnosis may be extremely difficult (196,202,216). Their histopathologic differences, particularly the characteristic herringbone tweed pattern of a well-differentiated fibrosarcoma and the storiform pattern of MFH (214), have been outlined previously.

The histologic appearance of fibrosarcoma may resemble *fibroblastic osteosarcoma* (242). The distinctive difference is the absence of bone or osteoid formation in the former. *Metastases* or some *soft tissue sarcomas* may simulate sclerosing epithelioid fibrosarcoma, which is constantly positive only for vimentin (222). All such lesions can be ruled out by IHC, e.g., *synovial sarcoma* (CD99+, BCL-2+, EMA/CK+), *clear cell sarcoma* (S-100/HMB-45+), *metastatic carcinoma* (CK+), or *sclerosing lymphoma* (CD45+).

Primary leiomyosarcomas of bone, although very rare, can be differentiated from MFH by IHC, applying muscle-specific antibodies such as anti-desmin and anti-al-pha-smooth muscle actin. *Melanomas* are S-100-positive, and metastatic *sarcomatoid carcinoma* exhibits cytokeratin immunoreactivity. A careful search for osteoid production is mandatory to exclude *osteosarcoma* before the diagnosis of MFH can be established.

Desmoplastic fibroma may be a differential possibility when a well-differentiated type of fibrosarcoma is examined, although this lesion is less cellular than fibrosarcoma and its mitotic activity is very low.

The radiologic and pathologic differential diagnosis of fibrosarcoma and malignant fibrous histiocytoma is shown in Figure 4-70.





REFERENCES

Fibrous Cortical Defect and Nonossifying Fibroma

- Arata MA, Peterson HA, Dahlin DC. Pathological fractures through non-ossifying fibromas. J Bone Joint Surg Am 1981;63: 980–988.
- Bertoni F, Unni KK, McLeod RA, Sim FH. Xanthoma of bone. Am J Pathol 1988;90:377–384.
- Blau RA, Zwick DL, Westphal RA. Multiple nonossifying fibromas. J Bone Joint Surg Am 1988;70:299–304.
- Brenner RJ, Hattner RS, Lilien DL. Scintigraphic features of nonosteogenic fibroma. *Radiology* 1979;131:727–730.
- Caffey J.On fibrous defects in cortical walls of growing tubular bone: their radiologic appearance, structure prevalence, natural course and diagnostic significance. *Adv Pediatr* 1955;7: 13–51.
- Craver RD, Heinrich S, Mirra J. Fibrous cortical defect with bizarre nuclear features. Ann Diagn Pathol 1997;1:26–30.
- Cunningham BJ, Ackerman LV. Metaphyseal fibrous defects. J Bone Joint Surg 1956;38:797–808.
- Dahlin DC, Unni KK. Bone tumors. General aspects and data on 8,542 cases, 4th ed. Springfield, IL: Charles C Thomas, 1986.
- Dallari D, Marinelli A, Pellacani A, et al. Xanthoma of bone: first sign of hyperlipidemia type IIB: a case report. *Clin Orthop Relat Res* 2003;410:274–277.
- Dehner LP. Juvenile xanthogranulomas in the first two decades of life: a clinicopathologic study of 174 cases with cutaneous and extracutaneous manifestations. *Am J Surg Pathol* 2003;27: 579–593.
- Dunham WK, Marcus NW, Enneking WF, Haun C. Developmental defects of the distal femoral metaphysis. J Bone Joint Surg Am 1980;62:801–806.
- Evans GA, Park WM. Familial multiple non-osteogenic fibromata. J Bone Joint Surg Br 1978;60:416–419.
- Favara BE, Feller AC, Pauli M, et al. Contemporary classification of histiocytic disorders. The WHO Committee On Histiocytic/ Reticulum Cell Proliferations. Reclassification Working Group of the Histiocyte Society. *Med Pediatr Oncol* 1997;29:157–166.
- Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:225–226.
- Friedland JA, Reinus WR, Fisher AJ, Wilson AJ. Quantitative analysis of the plain radiographic appearance of nonossifying fibroma. *Invest Radiol* 1995;30:474–479.
- Greyson ND, Pang S. The variable bone scan appearances of nonosteogenic fibroma of bone. *Clin Nucl Med* 1981;6:242–245.
- Hudson TM, Stiles RG, Monson DK. Fibrous lesions of bone. *Radiol Clin North Am* 1993;31:279–297.
- Jaffe HL, Lichtenstein L. Non-osteogenic fibroma of bone. Am J Pathol 1942;18:205–221.
- Jaffe HL. Tumors and tumorous conditions of the bones and joints. Philadelphia: Lea & Febiger, 1968.
- Jee W-H, Choe B-Y, Kang H-S, et al. Nonossifying fibroma: characteristics at MR imaging with pathologic correlation. *Radiology* 1998;209:197–202.
- Kumar R, Madewell JE, Lindell MM, Swischuk LE. Fibrous lesions of bones. *RadioGraphics* 1990;10:237–256.
- 22. Kumar R, Swischuk LE, Madewell JE. Benign cortical defect: site for an avulsion fracture. *Skeletal Radiol* 1986;15:553–555.
- Mirra JM, Gold RH, Rand F. Disseminated nonossifying fibromas in association with café-au-lait spots (Jaffe-Campanacci syndrome). *Clin Orthop* 1982;168:192–205.
- Moser RP Jr, Sweet DE, Haseman DB, Madewell JE. Multiple skeletal fibroxanthomas: radiologic-pathologic correlation of 72 cases. *Skeletal Radiol* 1987;16:353–359.
- Nelson M, Perry D, Ginsburg G, et al. Translocation (1;4)(p31;q34) in nonossifying fibroma. *Cancer Genet Cytogenet* 2003;142:142–144.
- Ritschl P, Karnel F, Hajek PC. Fibrous metaphyseal defects determination of their origin and natural history using a radiomorphological study. *Skeletal Radiol* 1988;17:8–15.
- Ritschl P, Hajek PC, Pechmann U. Fibrous metaphyseal defects. Magnetic resonance imaging appearances. *Skeletal Radiol* 1989; 18:253–259.

- Schajowicz F. Histological typing of bone tumors. In: World Health Organization international histological classification of tumors. Berlin: Springer-Verlag, 1993.
- Schajowicz F. Tumors and tumorlike lesions of bone: pathology, radiology, and treatment, 2nd ed. Berlin: Springer-Verlag, 1994.
- Schwartz AM, Ramos RM. Neurofibromatosis and multiple nonossifying fibroma. Am J Roentgenol 1980;135:617–619.
- Selby S. Metaphyseal cortical defects in the tubular bones of growing children. J Bone Joint Surg Am 1961;43:395–400.
- Shelton CH, Nimityongskul P, Richardson PH, Brogdon BG. Progressive painful bowing of the right leg. *Acad Radiol* 1995;2: 351–353.
- Steiner GC. Fibrous cortical defect and non-ossifying fibroma of bone: a study of the ultrastructure. Arch Pathol 1974;97:205–210.
- Tarkkanen M, Kaipainen A, Karaharju E, et al. Cytogenetic study of 249 consecutive patients examined for a bone tumor. *Cancer Genet Cytogenet* 1993;68:1–21.
- Unni KK. Fibrous and fibrohistiocytic lesions of bone. Semin Orthop 1991;6:177–186.
- Wold LE. Fibrohistiocytic tumors of bone. In: Unni KK, ed. Bone tumors. New York: Churchill Livingstone, 1988:183–197.

Benign Fibrous Histiocytoma

- Arii Y, Moritani M, Hirakoh H. A case of benign fibrous histiocytoma of the femur. Orthop Surg (Seikeigeka) 1991;42:1248–1250.
- Bertoni F, Calderoni P, Bacchini P, Sudanese A. Benign fibrous histiocytoma of bone. J Bone Joint Surg Am 1986;68: 1225-1230.
- Clarke BE, Xipell JM, Thomas DP. Benign fibrous histiocytoma of bone. Am J Surg Pathol 1985;9:806–815.
- Dominok GW, Eisengarten W. Benignes fibroeses Histiozytom des Knochens. Zentralbl Pathol 1980;124:77–83.
- 41. Hamada T, Ito H, Araki Y, et al. Benign fibrous histiocytoma of the femur: review of three cases. *Skeletal Radiol* 1996;25: 25–29.
- 42. Huvos A. Bone tumors: diagnosis, treatment and prognosis, 2nd ed. Philadelphia: WB Saunders, 1991.
- Matsuno T. Benign fibrous histiocytoma involving the ends of long bone. *Skeletal Radiol* 1990;19:561–566.
- 44. Mesiter P, Konrad E, Hohne N. Incidence and histological structure of the storiform pattern in benign and malignant fibrous histiocytomas. *Virchows Arch A* 1981;393:93–101.
- 45. Schajowicz F, Ackerman LV, Sissons HA. *Histological typing of bone tumors*. Geneva: World Health Organization, 1972.
- Schajowicz F, Sissons HA, Sobin LH. The World Health Organization's histologic classification of bone tumors. A commentary on the second edition. *Cancer* 1995;75:1208–1214.
- 47. Spjut HJ, Dorfman HD, Fechner RE, Ackerman LV. Tumors of bone pathology. In: Firminger HI, ed. *Atlas of tumor pathology*, 2nd series, fascicle 5. Washington DC: Armed Forces Institute of Pathology, 1971.
- Statz EM, Pochebit SM, Cooper A, et al. Case report 525. Benign fibrous histiocytoma (BFH) of thumb. *Skeletal Radiol* 1989;18: 299–302.

Periosteal Desmoid

- 49. Bahk W-J, Kang Y-K, Lee A-H, Mirra JM. Desmoid tumor of bone with enchondromatous nodules, mistaken for chondrosarcoma. *Skeletal Radiol* 2003;32:223–226.
- Barnes GR Jr, Gwinn JL. Distal irregularities of the femur simulating malignancy. Am J Roentgenol 1974;122:180–185.
- Brower AC, Culver JE Jr, Keats TE. Histological nature of the cortical irregularity of the medial posterior distal femoral metaphysis in children. *Radiology* 1971;99:389–392.
- 52. Bufkin WJ. The avulsive cortical irregularity. Am J Roentgenol 1971;112:487-492.
- Burrows PE, Greenberg ID, Reed MH. The distal femoral defect: Technetium-99m pyrophosphate bone scan results. J Can Assoc Radiol 1982;33:91–93.
- 54. Kimmelstiel P, Rapp I. Cortical defect due to periosteal desmoids. Bull Hosp Joint Dis 1951;12:286–297.
- Pennes DR, Braunstein EM, Glazer GM. Computed tomography of cortical desmoid. *Skeletal Radiol* 1984;12:40–42.

- **310** Differential Diagnosis in Orthopaedic Oncology
- Resnick D, Greenway G. Distal femoral cortical defects, irregularities, and excavations: a critical review of the literature with the addition of histologic and paleopathologic data. *Radiology* 1982;143:345–354.
- 57. Velchik MG, Heyman S, Makler PT Jr, et al. Bone scintigraphy: differentiating benign cortical irregularity of the distal femur from malignancy. *J Nucl Med* 1984;25:72–74.
- Yamazaki T, Maruoka S, Takahashi S, et al. MR findings of avulsive cortical irregularity of the distal femur. *Skeletal Radiol* 1995;24:43–46.

Fibrous Dysplasia

- 59. Albright F, Butler AM, Hampton AO, Smith P. Syndrome characterized by osteitis fibrosa disseminata, areas of pigmentation and endocrine dysfunction, with precocious puberty in females: report of five cases. *N Engl J Med* 1937;216:727–746.
- Bancroft LW, Kransdorf MJ, Menke DM, et al. Intramuscular myxoma: Characteristic MR imaging features. Am J Roentgenol 2002;178:1255–1259.
- 61. Beaman FD, Bancroft LW, Peterson JJ, et al. Imaging characteristics of cherubism. *Am J Roentgenol* 2004;182:1051–1054.
- 62. Bullough PG. Atlas of orthopaedic pathology with clinical and radiologic correlations, 2nd ed. New York: Gower Medical Publishing, 1992.
- 63. Cabral CEL, Guedes P, Fonseca T, et al. Polyostotic fibrous dysplasia associated with intramuscular myxomas: Mazabraud's syndrome. *Skeletal Radiol* 1998;27:278–282.
- 64. Camilleri AE. Craniofacial fibrous dysplasia. J Laryngol Otol 1991;105:662–666.
- 65. Candeliere GA, Glorieux FH, Prud'Homme J, St.-Arnaud R. Increased expression of the c-fos proto-oncogene in bone from patients with fibrous dysplasia. *N Engl J Med* 1995;332:1546–1551.
- Choi IH, Kim CJ, Cho T-J, et al. Focal fibrocartilaginous dysplasia of long bones: report of eight additional cases and literature review. *J Pediatr Orthop* 2000;20:421–427.
- Cohen MM Jr, Howell RE. Etiology of fibrous dysplasia and Mc-Cune-Albright syndrome. Int J Oral Maxillofac Surg 1999;28: 366–371.
- Cohen MM Jr, Siegal GP. McCune-Albright syndrome. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:357–359.
- 69. Creagh MF, Nunan TO. Positive gallium-67 citrate uptake in a patient with polyostotic fibrous dysplasia. *Clin Nucl Med* 1988;13: 241–242.
- 70. Daffner RH. Fibrous dysplasia. Contemp Diagn Radiol 2002;25:1-5.
- Daffner RH, Kirks DR, Gehweiler JA Jr, Heaston DK. Computed tomography of fibrous dysplasia. Am J Roentgenol 1982;139: 943–948.
- Dahlin DC, Bertoni F, Beabout JW, Campanacci M. Fibrocartilaginous mesenchymoma with low grade malignancy. *Skeletal Radiol* 1984;12:263–269.
- Dal Cin P, Sciot R, Brys P, et al. Recurrent chromosome aberrations in fibrous dysplasia of the bone: a report of the CHAMP study group. Chromosomes and morphology. *Cancer Genet Cytogenet* 2000;122:30–32.
- DeSmet AA, Travers H, Neff JR. Chondrosarcoma occurring in a patient with polyostotic fibrous dysplasia. *Skeletal Radiol* 1981;7: 197–201.
- Dorfman HD, Ishida T, Tsuneyoshi M. Exophytic variant of fibrous dysplasia (fibrous dysplasia protuberans). *Hum Pathol* 1994;25:1234–1237.
- Drolshagen LF, Reynolds WA, Marcus NW. Fibrocartilaginous dysplasia of bone. *Radiology* 1985;156:32.
- Fisher AJ, Totty WG, Kyriakos M. MR appearance of cystic fibrous dysplasia. Case report. J Comput Assist Tomogr 1994;18:315–318.
- Fitzpatrick KA, Taljanovic MS, Speer DP, et al. Imaging findings of fibrous dysplasia with histopathologic and intraoperative correlation. *Am J Roentgenol* 2004;182:1389–1398.
- Gober GA, Nicholas RW. Case report 800. Skeletal fibrous dysplasia associated with intramuscular myxoma (Mazabraud's syndrome). Skeletal Radiol 1993;22:452–455.
- Hannon TS, Noonan K, Steinmetz R, et al. Is McCune-Albright syndrome overlooked in subjects with fibrous dysplasia of bone? *J Pediatr* 2003;142:532–538.

- Happle R. The McCune-Albright syndrome: a lethal gene surviving by mosaicism. *Clin Genet* 1986;29:321–324.
- Happle R. Lethal genes surviving by mosaicism: a possible explanation for sporadic birth defects involving the skin. J Am Acad Dermatol 1987;16:899–906.
- Henry A. Monostotic fibrous dysplasia. J Bone Joint Surg Br 1969; 51:300–306.
- Henschen F. Fall von Ostitis Fibrosa mit multiplen Tumoren in der umgebenden Musculatur. Verh Dtsch Ges Pathol 1926;21: 93–97.
- Hermann G, Klein M, Abdelwahab IF, Kenan S. Fibrocartilaginous dysplasia. *Skeletal Radiol* 1996;25:509–511.
- Hoshi H, Futami S, Ohnishi T, et al. Gallium-67 uptake in fibrous dysplasia of the bone. Ann Nucl Med 1990;4:35–38.
- Inamo Y, Hanawa Y, Kin H, Okuni M. Findings on magnetic resonance imaging of the spine and femur in a case of McCune-Albright syndrome. *Pediatr Radiol* 1993;23:15–18.
- Ishida T, Dorfman HD. Massive chondroid differentiation in fibrous dysplasia of bone (fibrocartilaginous dysplasia). *Am J Surg Pathol* 1993;17:924–930.
- Iwasko N, Steinbach LS, Disler D, et al. Imaging findings in Mazabraud's syndrome: seven new cases. *Skeletal Radiol* 2002;31: 81–87.
- Jee W-H, Choi K-H, Choe B-Y, et al. Fibrous dysplasia: MR imaging characteristics with radiopathologic correlation. Am J Roentgenol 1996;167:1523–1527.
- Jundt G. Fibrous dysplasia. In: Barnes L, Eveson J, Reichart P, Sidransky D, eds. *Pathology and genetics of head and neck tumours*. Lyon: IARC Press, 2005:321–322.
- Kaugars GE, Niamtu J III, Svirsky JA. Cherubism: diagnosis, treatment, and comparison with central giant cell granulomas, and giant cell tumors. *Oral Surg Oral Med Oral Pathol* 1992; 73:369–374.
- Kaushik S, Smoker WRK, Frable WJ. Malignant transformation of fibrous dysplasia into chondroblastic osteosarcoma. *Skeletal Radiol* 2002;31:103–106.
- Kransdorf MJ, Moser RP, Gilkey FW. Fibrous dysplasia. RadioGraphics 1990;10:519–537.
- Kransdorf MJ, Murphey MD. Case 12: Mazabraud syndrome. Radiology 1999;212:129–132.
- 96. Kransdorf MJ, Murphey MD, Sweet DE. Liposclerosing myxofibrous tumor: a radiologic-pathologic-distinct fibro-osseous lesion of bone with a marked predilection for the intertrochanteric region of the femur. *Radiology* 1999;212:693–698.
- 97. Kyriakos MK, McDonald DJ, Sundaram M. Fibrous dysplasia with cartilaginous differentiation ("fibrocartilaginous dysplasia"): a review, with an illustrative case followed for 18 years. *Skeletal Radiol* 2004;33:51–62.
- Lichtenstein L, Jaffe HL. Fibrous dysplasia of bone. Arch Pathol 1942;33:777–816.
- Lo B, Faiyaz-Ul-Haque M, Kennedy S, et al. Novel mutation in the gene encoding c-Abl-binding protein SH3BP2 causes cherubism. *Am J Med Genet* 2003;121A:37–40.
- Machida K, Makita K, Nishikawa J, et al. Scintigraphic manifestation of fibrous dysplasia. *Clin Nucl Med* 1986;11:426–429.
- Marie PJ. Cellular and molecular basis of fibrous dysplasia. *Histol Histopathol* 2001;16:981–988.
- 102. Marie PJ, de Pollak C, Chanson P, Lomri A. Increased proliferation of osteoblastic cells expressing the activating Gs alpha mutation in monostotic and polyostotic fibrous dysplasia. Am J Pathol 1997;150:1059–1069.
- Maroteaux P, Freisinger P, Merrer M. Pachydysostosis of the fibula. J Bone Joint Surg Br 1991;73:842–845.
- 104. Martinez-Tello FJ, Manjón-Luengo P, Martin-Pérez M, Montes-Moreno S. Cherubism associated with neurofibromatosis type 1, and multiple osteolytic lesions of both femurs: a previously undescribed association of findings. *Skeletal Radiol* 2005;34: 793–798.
- Mazabraud A, Semat P, Roze R. A propos de l'association de fibromyxomes des tissus mous à la dysplasie fibreuse des os. *Presse Med* 1967;75:2223–2228.
- McCune DJ, Bruch H. Progress in pediatrics: osteodystrophia fibrosa. Am J Dis Child 1937;54:806–848.
- 107. Mirra JM, Gold RH. Fibrous dysplasia. In: Mirra HM, Gold RH, Picci P, eds. *Bone tumors*. Philadelphia: Lea & Febiger, 1989.

- Mulder JD, Schütte HE, Kroon HM, Taconis WK. Radiologic atlas of bone tumors. Amsterdam: Elsevier, 1993:607–625.
- Murphey MD, McRae GA, Fanburg-Smith JC, et al. Imaging of soft-tissue myxoma with emphasis on CT and MR and comparison of radiologic and pathologic findings. *Radiology* 2002;225: 215–224.
- Okada K, Yoshida S, Okane K, Sageshima M. Cystic fibrous dysplasia mimicking giant cell tumor: MRI appearance. *Skeletal Radiol* 2000;29:45–48.
- Ragsdale BD. Polymorphic fibro-osseous lesions of bone: an almost site-specific diagnostic problem of the proximal femur. *Hum Pathol* 1993;24:505–512.
- Riley GM, Greenspan A, Poirier VC. Fibrous dysplasia of a parietal bone. J Comput Assist Tomogr 1997;21:41–43.
- Riminucci M, Fisher LW, Shenker A, et al. Fibrous dysplasia of bone in the McCune-Albright syndrome: abnormalities in bone formation. *Am J Pathol* 1997;151:1587–1600.
- 114. Rodenberg J, Jensen OM, Keller J, et al. Fibrous dysplasia of the spine, costae and hemipelvis with sarcomatous transformation. *Skeletal Radiol* 1996;25:682–684.
- 115. Ruggieri P, Sim FH, Bond JA, Unni KK. Malignancies in fibrous dysplasia. *Cancer* 1994;73:1411–1424.
- Schwartz DT, Alpert M. The malignant transformation of fibrous dysplasia. Am J Med Sci 1964;247:1–20.
- 117. Schwindinger WF, Francomano CA, Levine MA. Identification of a mutation in the gene encoding the alpha subunit of the stimulatory G-protein of adenylyl cyclase in McCune-Albright syndrome. *Proc Natl Acad Sci* 1992;89:5152–5156.
- 118. Siegal GP, Dal Cin P, Araujo ES. Fibrous dysplasia. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:341–342.
- 119. Sissons HA, Malcolm AJ. Fibrous dysplasia of bone: case report with autopsy study 80 years after the original clinical recognition of the bone lesions. *Skeletal Radiol* 1997;26:177–183.
- Sundaram M, McDonald DJ, Merenda G. Intramuscular myxoma: a rare but important association with fibrous dysplasia of bone. *Am J Roentgenol* 1989;153:107–108.
- 121. Tiziani V, Reichenberger E, Buzzo CL, et al. The gene for cherubism maps to chromosome 4p16. *Am J Hum Genet* 1999;65: 158–166.
- Utz JA, Kransdorf MJ, Jelinek JS, et al. MR appearance of fibrous dysplasia. J Comput Assist Tomogr 1989;13:845–851.
- 123. Yabut SM, Kenan S, Sissons HA, Lewis MM. Malignant transformation of fibrous dysplasia. *Clin Orthop* 1988;228:281–289.
- Yamaguchi T, Dorfman HD, Eisig S. Cherubism: clinicopathologic features. *Skeletal Radiol* 1999;28:350–353.

Osteofibrous Dysplasia

- 125. Alguacil-Garcia A, Alonso A, Pettigrew NM. Osteofibrous dysplasia (ossifying fibroma) of the tibia and fibula and adamantinoma. Am J Clin Pathol 1984;82:470–474.
- 126. Bridge JA, Dembinski A, DeBoer J, et al. Clonal chromosomal abnormalities in osteofibrous dysplasia. Implications for histopathogenesis and its relationship with adamantinoma. *Cancer* 1994;73:1746–1752.
- 127. Bridge JA, Swarts SJ, Buresh C, et al. Trisomies 8 and 20 characterize a subgroup of benign fibrous lesions arising in both soft tissue and bone. *Am J Pathol* 1999;154:729–733.
- Campanacci M, Laus M. Osteofibrous dysplasia of the tibia and fibula. J Bone Joint Surg Am 1981;63:367–375.
- Campanacci M. Osteofibrous dysplasia of the long bones. A new clinical entity. *Ital J Orthop Traumatol* 1976;2:221–237.
- Campbell CJ, Hawk T. A variant of fibrous dysplasia (osteofibrous dysplasia). J Bone Joint Surg Am 1982;64:231–236.
- Castelotte A, Garcia-Peña P, Lucaya J, Lorenzo J. Osteofibrous dysplasia. A report of two cases. *Skeletal Radiol* 1988;17: 483–486.
- Cohen DM, Dahlin DC, Pugh DG. Fibrous dysplasia associated with adamantinoma of the long bones. *Cancer* 1962;15:515–521.
- 133. Czerniak B, Rojas-Corona RR, Dorfman HD. Morphologic diversity of long bone adamantinoma. The concept of differentiated (regressing) adamantinoma and its relationship to osteofibrous dysplasia. *Cancer* 1989;64:2319–2334.

- Dominguez R, Saucedo J, Fenstermacher M. MRI findings in osteofibrous dysplasia. *Magn Reson Imag* 1989;7:567–570.
- Greenfield GB, Arrington JA. Imaging of bone tumors. A multimodality approach. Philadelphia: Lippincott, 1995:158–159.
- Hazelbag HM, Wessels JW, Mollevangers P, et al. Cytogenetic analysis of adamantinoma of long bones: further indications for a common histogenesis with osteofibrous dysplasia. *Cancer Genet Cytogenet* 1997;97:5–11.
- 137. Hisaoka M, Hashimoto H, Ohguri T, et al. Congenital (infantile) pseudoarthrosis of the fibula associated with osteofibrous dysplasia. *Skeletal Radiol* 2004;33:545–549.
- Hunter AG, Jarvis J. Osteofibrous dysplasia: two affected male sibs and an unrelated girl with bilateral involvement. *Am J Med Genet* 2002;112:79–85.
- 139. Kahn LB. Adamantinoma, osteofibrous dysplasia and differentiated adamantinoma. *Skeletal Radiol* 2003;32:245–258.
- 140. Karol LA, Brown DS, Wise CA, Waldron M. Familial osteofibrous dysplasia. A case series. J Bone Joint Surg Am 2005; 87:2297–2307.
- 141. Keeney GL, Unni K, Beabout JW, Pritchard DJ. Adamantinoma of long bones. *Cancer* 1989;64:730–737.
- 142. Kempson RL. Ossifying fibroma of the long bones. A light and electron microscopic study. *Arch Pathol* 1966;82:218–233.
- 143. Levine SM, Lambiase RE, Petchprapa CN. Cortical lesions of the tibia: characteristic appearances at conventional radiography. *RadioGraphics* 2003;23:157–177.
- 144. Markel SF. Ossifying fibroma of long bone. Am J Clin Pathol 1978;69:91–97.
- 145. Park Y, Unni KK, McLeod RA, Pritchard DJ. Osteofibrous dysplasia: clinicopathologic study of 80 cases. *Hum Pathol* 1993;24: 1339–1347.
- 146. Sakamoto A, Oda Y, Iwamoto Y, Tsuneyoshi M. A comparative study of fibrous dysplasia and osteofibrous dysplasia with regard to Gs alpha mutation at the Arg201 codon: polymerase chain reaction-restriction fragment length polymorphism analysis of paraffin-embedded tissues. *J Mol Diagn* 2000;2:67–72.
- 147. Sherman GM, Damron TA, Yang Y. CD99 positive adamantinoma of the ulna with ipsilateral discrete osteofibrous dysplasia. *Clin Orthop Relat Res* 2003;408:256–261.
- Sissons HA, Kancherla PL, Lehman WB. Ossifying fibroma of bone. Report of two cases. *Bull Hosp Joint Dis Orthop Inst* 1983; 43:1–14.
- 149. Springfield DS, Rosenberg AE, Mankin HJ, Mindell ER. Relationship between osteofibrous dysplasia and adamantinoma. *Clin Orthop* 1994;309:234–244.
- Sweet DE, Vinh TN, Devaney K. Cortical osteofibrous dysplasia of long bone and its relationship to adamantinoma. *Am J Surg Pathol* 1992;16:282–290.
- Ueda Y, Blasius S, Edel G, et al. Osteofibrous dysplasia of long bones—a reactive process to adamantinomatous tissue. J Cancer Clin Oncol 1992;118:152–156.
- Unni KK, Dahlin DC, Beaubout JW, Ivins JC. Adamantinoma of long bones. *Cancer* 1974;34:1796–1805.
- 153. Vigorita VJ, Ghelman B, Hogendoorn PCW. Osteofibrous dysplasia. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:343–344.
- Wang J, Shih C, Chen W. Osteofibrous dysplasia (ossifying fibromas of long bones). *Clin Orthop* 1992;278:235–243.
- 155. Weiss SW, Dorfman HD. Adamantinoma of long bones. *Hum Pathol* 1977;8:141–153.
- Zeanah WR, Hudson TM, Springfield DS. Computed tomography of ossifying fibroma of the tibia. J Comput Assist Tomogr 1983;7:688–691.

Desmoplastic Fibroma

- 157. Bertoni F, Calderoni P, Bacchini P, Campanacci M. Desmoplastic fibroma of bone: a report of six cases. J Bone Joint Surg Br 1984;66:265–268.
- 158. Bridge JA, Rosenthal H, Sanger WG, Neff JR. Desmoplastic fibroma arising in fibrous dysplasia. Chromosomal analysis and review of the literature. *Clin Orthop Rel Res* 1989;247: 272–278.
- 159. Crim JR, Gold RH, Mirra JM, et al. Desmoplastic fibroma of bone: radiographic analysis. *Radiology* 1989;172:827–832.

- 160. Fornasier V, Pritzker KPH, Bridge JA. Desmoplastic fibroma. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:288.
- 161. Frick MA, Sundaram M, Unni KK, et al. Imaging findings in desmoplastic fibroma of bone: distinctive T2 characteristics. Am J Roentgenol 2005;184:1762–1767.
- 162. Gebhardt MC, Campbell CJ, Schiller AL, Mankin HJ. Desmoplastic fibroma of bone. A report of eight cases and review of the literature. J Bone Joint Surg Am 1985;67: 732–747.
- 163. Goldblum JR, Fletcher JA. Desmoid-type fibromatosis. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:83–84.
- Greenspan A, Unni KK. Case report 787. Desmoplastic fibroma. Skeletal Radiol 1993;22:296–299.
- 165. Hauben EI, Jundt G, Cleton-Jansen AM, Yavas A, et al. Desmoplastic fibroma of bone: an immunohistochemical study including beta-catenin expression and mutational analysis for betacatenin. *Hum Pathol* 2005;36:1025–1030.
- Inwards CY, Unni KK, Beabout JW, Sim FH. Desmoplastic fibroma of bone. *Cancer* 1991;68:1978–1983.
- 167. Jaffe HL. Tumors and tumorous conditions of the bones and joints. Philadelphia: Lea & Febiger, 1958:298–303.
- 168. Li C, Bapat B, Alman BA. Adenomatous polyposis coli gene mutation alters proliferation through its beta-catenin-regulatory function in aggressive fibromatosis (desmoid tumor). Am J Pathol 1998;153:709–714.
- Lichtman EA, Klein MJ. Case report 302. Desmoplastic fibroma of the proximal end of the left femur. *Skeletal Radiol* 1985;13:160–163.
- 170. Montgomery E, Lee JH, Abraham SC, Wu TT. Superficial fibromatoses are genetically distinct from deep fibromatoses. *Mod Pathol* 2001;14:695–701.
- 171. Rabhan WN, Rosai J. Desmoplastic fibroma. Report of ten cases and review of the literature. J Bone Joint Surg Am 1968;50: 487–502.
- 172. Sugiura I. Desmoplastic fibroma. Case report and review of the literature. *J Bone Joint Surg Am* 1976;58:126–130.
- 173. Tejpar S, Nollet F, Li C, et al. Predominance of beta-catenin mutations and beta-catenin dysregulation in sporadic aggressive fibromatosis (desmoid tumor). *Oncogene* 1999;18:6615–6620.
- 174. Taconis WK, Schütte HE, van der Heul RO. Desmoplastic fibroma of bone: a report of 18 cases. *Skeletal Radiol* 1994;23: 283–288.
- 175. Vanhoenacker FM, Hauben E, De Beuckeleer LH, et al. Desmoplastic fibroma of bone: MRI features. *Skeletal Radiol* 2000; 29:171–175.
- 176. Weiss SW, Goldblum JR. Enzinger and Weiss's soft tissue tumors, 4th ed. St. Louis: Mosby, 2001.
- 177. West R, Huvos AG, Lane JM. Desmoplastic fibroma of bone arising in fibrous dysplasia. *Am J Clin Pathol* 1983;79:630–633.
- 178. You JS, Lawrence S, Pathria M, et al. Desmoplastic fibroma of the calcaneus. *Skeletal Radiol* 1995;24:451–454.
- 179. Young JWR, Aisner SC, Levine AM, et al. Computed tomography of desmoid tumors of bone: Desmoplastic fibroma. *Skeletal Radiol* 1988;17:333–337.

Fibrosarcoma and Malignant Fibrous Histiocytoma

- Abdulkader I, Cameselle-Teijeiro J, Fraga M, et al. Sclerosing epithelioid fibrosarcoma primary of the bone. *Int J Surg Pathol* 2002;10:227–230.
- Aisen AM, Martel W, Braunstein EM, et al. MRI and CT evaluation of primary bone and soft-tissue tumors. *Am J Roentgenol* 1986;146:749–756.
- 182. Antonescu CR, Erlandson RA, Huvos AG. Primary fibrosarcoma and malignant fibrous histiocytoma of bone. A comparative ultrastructural study: evidence of a spectrum of fibroblastic differentiation. *Ultrastruct Pathol* 2000;24:83–91.
- Arnold WH. Hereditary bone dysplasia with sarcomatous degeneration. Study of a family. Ann Intern Med 1973;78:902–906.
- 184. Bacci G, Picci P, Mercuri M, et al. Neoadjuvant chemotherapy for high grade malignant fibrous histiocytoma of bone. *Clin Orthop Relat Res* 1998;346:178–189.

- 185. Bertoni F, Capanna R, Calderoni P, et al. Primary central (medullary) fibrosarcoma of bone. *Semin Diagn Pathol* 1984;1: 185–198.
- 186. Bloem JL, Taminiau AH, Eulderink F, et al. Radiologic staging of primary bone sarcoma: MR imaging, scintigraphy, angiography, and CT correlated with pathologic examination. *Radiology* 1988;169:805–810.
- Boland PJ, Huvos AG. Malignant fibrous histiocytoma of bone. *Clin Orthop* 1986;204:130–134.
- Broders AC, Hargrave R, Meyerding HW. Pathological features of soft tissue fibrosarcoma. Surg Gynecol Obstet 1939;69:267–280.
- Capanna R, Bertoni F, Bacchini P, et al. Malignant fibrous histiocytoma of bone: the experience at the Rizzoli Institute. Report of 90 cases. *Cancer* 1984;54:177–187.
- 190. Chow LT, Lui YH, Kumta SM, Allen PW. Primary sclerosing epithelioid fibrosarcoma of the sacrum: a case report and review of the literature. *J Clin Pathol* 2004;57:90–94.
- Dahlin DC. Grading of bone tumors. In: Unni KK, ed. Bone tumors. New York: Churchill Livingstone, 1988:35–45.
- Dahlin DC, Ivins JC. Fibrosarcoma of bone: a study of 114 cases. Cancer 1969:23:35–41.
- Dahlin DC, Unni KK, Matsuno T. Malignant (fibrous) histiocytoma of bone—fact or fancy? *Cancer* 1977;39:1508–1516.
- 194. Dorfman HD, Norman A, Wolff H. Fibrosarcoma complicating bone infarction in a caisson worker: case report. *J Bone Joint Surg* Am 1966;48:528–532.
- 195. Enzinger FM, Weiss SW. Soft tissue tumors. St. Louis: CV Mosby, 1983.
- 196. Erlandson RA, Woodruff JM. Role of electron microscopy in the evaluation of soft tissue neoplasms, with emphasis on spindle cell and pleomorphic tumors. *Hum Pathol* 1998;29:1372–1381.
- 197. Eyre-Brook AL, Price CHG. Fibrosarcoma of bone: review of 50 consecutive cases from the Bristol Bone Tumor Registry. J Bone Joint Surg Br 1969;51:20–37.
- Feldman F, Lattes R. Primary malignant fibrous histiocytoma (fibrous xanthoma) of bone. *Skeletal Radiol* 1977;1:145–160.
- Feldman F, Norman D. Intra- and extraosseous malignant histiocytoma (malignant fibrous xanthoma). *Radiology* 1972;104: 497–508.
- 200. Fletcher CDM. Pleomorphic malignant fibrous histiocytoma: Fact or fiction? *Am J Surg Pathol* 1992;16:213–228.
- Fletcher CDM, Gustafson P, Rydholm A, et al. Clinicopathologic re-evaluation of 100 malignant fibrous histiocytomas: prognostic relevance of subclassification. *J Clin Oncol* 2001;19:3045–3050.
- 202. Fletcher CDM, van den Berg E, Molenaar WM. Pleomorphic malignant fibrous histiocytoma/undifferentiated high grade pleomorphic sarcoma. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:120–122.
- Galli SJ, Weintraub HP, Proppe KH. Malignant fibrous histiocytoma and pleomorphic sarcoma in association with medullary bone infarcts. *Cancer* 1978;41:607–619.
- 204. Greenfield GB, Arrington JA. Imaging of bone tumors. A multimodality approach. Philadelphia: JB Lippincott, 1995.
- 205. Hardcastle P, Nade S, Arnold W. Hereditary bone dysplasia with malignant change. Report of three families. *J Bone Joint Surg Am* 1986;68:1079–1089.
- 206. Hattinger CM, Tarkkanen M, Benini S, et al. Genetic analysis of fibrosarcoma of bone, a rare tumour entity closely related to osteosarcoma and malignant fibrous histiocytoma of bone. *Eur J Cell Biol* 2004;83:483–491.
- 207. Hudson TM, Hamlin DJ, Enneking WF, Pettersson H. Magnetic resonance imaging of bone and soft tissue tumors: early experience in 31 patients compared with computed tomography. *Skeletal Radiol* 1985;13:134–146.
- Huvos AG. Primary malignant fibrous histiocytoma of bone: clinicopathologic study of 18 patients. NY State J Med 1976; 552–559.
- 209. Huvos AG, Heilweil M, Bretsky SS. The pathology of malignant fibrous histiocytoma of bone. A study of 130 patients. *AmJ Surg Pathol* 1985;9:853–871.
- Huvos AG, Higinbotham NL. Primary fibrosarcoma of bone: a clinico-pathologic study of 130 patients. *Cancer* 1975;35:837–847.
- 211. Huvos AG, Higinbotham NL, Miller TR. Bone sarcomas arising in fibrous dysplasia. J Bone Joint Surg Am 1972;64:1047–1056.
- Huvos AG, Woodard HQ, Heilweil M. Postradiation malignant fibrous histiocytoma of bone: a clinicopathologic study of 20 patients. *Am J Surg Pathol* 1986;10:9–18.
- 213. Kahn LB, Vigorita VJ. Fibrosarcoma of bone. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:289–290.
- 214. Kahn LB, Webber B, Mills E, et al. Malignant fibrous histiocytoma (malignant fibrous xanthoma: xanthosarcoma) of bone. *Cancer* 1978;42:640–651.
- Kempson RL, Kyriakos M. Fibroxanthosarcoma of the soft tissues. A type of malignant fibrous histiocytoma. *Cancer* 1972;29: 961–976.
- Lawson CW, Fisher C, Gatter KC. An immunohistochemical study of differentiation in malignant fibrous histiocytoma. *Histopathology* 1987;11:375–383.
- 217. Lin WY, Kao CH, Hsu CY, et al. The role of Tc-99m MDP and Ga-67 imaging in the clinical evaluation of malignant fibrous histiocytoma. *Clin Nucl Med* 1994;19:996–1000.
- Link TM, Haeussler MD, Poppek S, et al. Malignant fibrous histiocytoma of bone: conventional X-ray and MR imaging features. *Skeletal Radiol* 1998;27:552–558.
- 219. Martignetti JA, Desnick RJ, Aliprandis E, et al. Diaphyseal medullary stenosis with malignant fibrous histiocytoma: a hereditary bone dysplasia/cancer syndrome maps to 9p21–22. *Am J Hum Genet* 1999;64:801–807.
- 220. Martignetti JA, Gelb BD, Pierce H, et al. Malignant fibrous histiocytoma: inherited and sporadic forms have loss of heterozygosity at chromosome bands 9p21–22—evidence for a common genetic defect. *Genes Chromosomes Cancer* 2000;27:191–195.
- McCarthy EF, Matsuno T, Dorfman HD. Malignant fibrous histiocytoma of bone: a study of 35 cases. *Hum Pathol* 1979;10:57–70.
- 222. Meis-Kindblom JM, Kindblom LG, van den Berg E, Molenaar WM. Sclerosing epithelioid fibrosarcoma. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:106–107.
- 223. Mertens F, Fletcher CD, Dal Cin P, et al. Cytogenetic analysis of 46 pleomorphic soft tissue sarcomas and correlation with morphologic and clinical features: a report of the CHAMP Study Group. Chromosomes and Morphology. *Genes Chromosomes Cancer* 1998;22:16–25.
- 224. Mirra JM, Bullough PG, Marcove RC, et al. Malignant fibrous histiocytoma and osteosarcoma in association with bone infarcts. J Bone Joint Surg Am 1974;56:932–940.
- 225. Mirra JM, Gold RH, Marafiote R. Malignant (fibrous) histiocytoma arising in association with a bone infarct in sickle-cell disease: coincidence or cause-and-effect? *Cancer* 1977;39:186–194.
- Murphey MD, Gross TM, Rosenthal HG. Musculoskeletal malignant fibrous histiocytoma: radiologic-pathologic correlation. *RadioGraphics* 1994;14:807–826.
- 227. Nakashima Y, Morishita S, Kotoura Y, et al. Malignant fibrous histiocytoma of bone. *Cancer* 1985;55:2804–2811.

- Nishida J, Sim FH, Wenger DE, Unni KK. Malignant fibrous histiocytoma of bone. A clinicopathologic study of 81 patients. *Cancer* 1997;79:482–493.
- 229. Pettersson H, Gillespy T, Hamlin DJ, et al. Primary musculoskeletal tumors: examination with MR imaging compared with conventional modalities. *Radiology* 1987;164:237–241.
- Pettersson H, Slone RM, Spanier S, et al. Musculoskeletal tumors: T1 and T2 relaxation times. *Radiology* 1988;167: 783–785.
- 231. Pritchard DJ, Sim FH, Ivins JC, et al. Fibrosarcoma of bone and soft tissues of the trunk and extremities. *Orthop Clin North Am* 1977;8:869–881.
- Ros PR, Viamonte M Jr, Rywlin AM. Malignant fibrous histiocytoma: mesenchymal tumor of ubiquitous origin. *Am J Roentgenol* 1984;142:753–759.
- 233. Seiss SW, Enzinger FM. Malignant fibrous histiocytoma: an analysis of 200 cases. *Cancer* 1978;41:2250–2260.
- 234. Smith J. Radiation-induced sarcoma of bone: clinical and radiographic findings in 43 patients irradiated for soft tissue neoplasms. *Clin Radiol* 1982;33:205–221.
- 235. Spanier SS, Enneking WF, Enriquez P. Primary malignant fibrous histiocytoma of bone. *Cancer* 1975;36:2084–2098.
- 236. Steiner GC, Jundt G, Martignetti JA. Malignant fibrous histiocytoma of bone. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:294–296.
- 237. Stout AP, Lattes R. Tumors of the soft tissues. In: Atlas of tumor pathology, 2nd fascicle series 1. Washington, DC: Armed Forces Institute of Pathology, 1967.
- 238. Strauchen JA, Dimitriu-Bona A. Malignant fibrous histiocytoma: expression of monocyte-macrophage differentiation antigens detected with monoclonal antibodies. *Am J Pathol* 1986;124: 303–309.
- 239. Sundaram M, McLeod RA. MR imaging of tumor and tumorlike lesions of bone and soft tissue. *Am J Roentgenol* 1990;155: 817–824.
- 240. Taconis WK, Mulder JD. Fibrosarcoma and malignant fibrous histiocytoma of long bones: radiographic features and grading. *Skeletal Radiol* 1984;11:237–245.
- 241. Taconis WK, Van Rijssel TG. Fibrosarcoma of long bones: a study of the significance of areas of malignant fibrous histiocy-toma. *J Bone Joint Surg Br* 1985;67:111–116.
- Unni KK. Osteosarcoma of bone. In: Unni KK, ed. Bone tumors. New York: Churchill Livingstone, 1988:107–133.
- Weiss SW, Goldblum JR. Malignant fibrous histiocytoma. In: Enzinger and Weiss's soft tissue tumors, 4th ed. Philadelphia: Mosby-Harcourt, 2001:917–954.
- Wood GS, Beckstead JH, Turner RR, et al. Malignant fibrous histiocytoma tumor cells resemble fibroblasts. *Am J Surg Pathol* 1986;10:323–335.
- 245. Wood GS, Beckstead JH, Turner RR, et al. Malignant fibrous histiocytoma cells do not express the antigenic or enzyme histochemical features of cells of monocyte/macrophage lineage. *Lab Invest* 1985;52:78 (abstract).
- 246. Yuen WWH, Saw D. Malignant fibrous histiocytoma of bone. *J Bone Joint Surg Am* 1985;67:482–486.

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Benign Lesions

Langerhans Cell Histiocytosis (Eosinophilic Granuloma)

Langerhans cell histiocytosis (LCH) is a lesion belonging to a group of disorders now classified by the World Health Organization (WHO) as histiocytic and dendritic cell disorders (39). The incidence in the United States is estimated at 0.05 to 0.5 per 100,000 children per year, with a 2:1 male predominance (20,45,49). This disorder represents less than 1% of all biopsyproven primary bone lesions (16,20,52). Lichtenstein (46,47) proposed the name histiocytosis X for the three conditions: eosinophilic granuloma (condition with bone lesions only); Hand-Schüller-Christian disease or xanthomatosis (characterized by the triad of bone lesions, diabetes insipidus, and exophthalmos), now considered as multifocal unisystem disease; and

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Letterer-Siwe disease or nonlipid reticulosis (in which disseminated disease is marked by wasting, lymphadenopathy, hepatosplenomegaly, and anemia), now considered by the WHO as multifocal multisystem disease (75). His rationale was the postulation that these lesions, being of unknown cause, appeared to represent an increasing spectrum of severity in which the common denominator was the presence of the histiocyte (9,76). Schajowicz and Polak proposed the general term histiocytic granuloma for this group of disorders, which may appear without or with only isolated eosinophils or, more commonly, with many eosinophilic leukocytes, and may involve a single or several bones (68). Recently, histiocytosis X has been given the name of Langerhans cell histiocytosis (23,48,50) because it has been verified that the primary proliferative element in this disease is the Langerhans cell, a mononuclear cell of the dendritic type that is found in the epidermis but is derived from precursors in the bone marrow (60,61,67,70). LCH is characterized by its ultrastructural (Birbeck granules) and immunohistochemical (positivity for CD1a, S-100, CD68, and Langerin/CD207, the latter of which is involved in Birbeck granule formation) properties (6,28,62,71,72). Although its causes and pathogenesis remain unsettled, LCH is now considered a disorder of immune regulation rather than a neoplastic process (59,80). However, some laboratory studies have shown that certain lesions exhibit a clonal proliferation of cells, suggesting a neoplastic origin (77,79). In addition, differences between normal epidermal Langerhans cells

and lesional Langerhans cells have been described (44). Familial clustering and studies in twins with 80%and 33% concordance of the disease in monozygotic and dizygotic twins, respectively, point to a genetic component in LCH (4). Molecular genetic studies using comparative genomic hybridization (CGH) and loss of heterozygosity (LOH) experiments have revealed chromosomal alterations, with predominant losses affecting chromosomes 1p, 5p, 6q, 9, 16, 17, and 22q in CGH and highest LOH frequencies on 1p and 17, leading to the hypothesis that loss of tumor suppressor genes located on chromosome 1p may be involved in development and progression of the disease (57). Other studies have demonstrated alterations of the cell cycle in lesional Langerhans cells (22). Because lesions in LCH represent the cellular composition of an innate immune response, Nezelof and Basset (59) hypothesize that a block in the cross-talk between innate and adaptive immune responses may give rise to large numbers of inflammatory molecules and to the proliferation of Langerhans cells and macrophages. On the basis of these assumptions, Egeler et al. (22) proposed a unifying concept comprising a combination of oncogenesis (underlying genetic defect in replication pathways or DNA repair of Langerhans cells) and immune dysregulation (blockage of maturation of Langerhans cells in lesions).

LCH exhibits a broad spectrum of clinical and radiologic abnormalities. It is characterized by an abnormal proliferation of Langerhans cells in various parts of the mononuclear system such as bone, lungs, central nervous system, skin, and lymph nodes.

Clinical Presentation

LCH may manifest as a solitary lesion or as multiple lesions (30,51). It occurs most often during childhood, with a peak incidence between the ages of 5 and 10 years (78). It is slightly more common in males (24). The most commonly affected sites are the skull (calvaria, skull base, and facial bones), the ribs, the pelvis, the spine, and the long bones (Fig. 5-1). Epiphyseal lesions are relatively rare (35,70). Clinical manifestations include local pain, tenderness, and swelling or a soft tissue mass adjacent to the site of the skeletal lesion (1). Fever, an elevated sedimentation rate, and leukocytosis may also be present (65). If a vertebra is affected, the patient may present with neurologic symptoms (16) resulting from a collapse of the vertebral body (31,41).

Imaging

Radiography remains the most effective modality for the diagnosis of LCH. In the mandible, radiolucent lesions and "floating teeth" secondary to the destruction of the alveolar bone are characteristic (Fig. 5-2). In the skull, a beveled lytic lesion is typical (Fig. 5-3) (25). In the center of the lytic lesion there is sometimes a small sclerotic focus referred to as a button sequestrum (11), "cockade" image (in European literature), or "bull's eye" simulating an infectious process (16,46,65). In the spine, so-called vertebra plana (or coin-on-edge appearance) is a usual manifestation and results from a collapsed verte-



Figure 5-1 Langerhans cell histiocytosis: skeletal sites of predilection, peak age range, and male-to-female ratio.



Figure 5-2 Langerhans cell histiocytosis. A 3-year-old girl with extensive skeletal involvement had in addition a large destructive lesion in the mandible. Note the characteristic appearance of a floating tooth, which results from destruction of supportive alveolar bone.

Α



Figure 5-3 Langerhans cell histiocytosis. Lateral radiograph of the skull of a $2\frac{1}{2}$ -year-old boy with disseminated process shows an osteolytic lesion in the frontal bone with a sharply outlined margin, giving it a punched-out appearance. Uneven involvement of the inner and outer tables results in its beveled presentation.

В



Figure 5-4 Langerhans cell histiocytosis. A: Vertebra plana represents collapse of vertebral body secondary to destruction of bone by granulomatous lesion. Note the preservation of the intervertebral disk space. **B:** In another patient, a sagittal T2-weighted magnetic resonance imaging (MRI) scan shows compression of the ventral aspect of the thecal sac by a fractured vertebral body (*arrow*).



Figure 5-5 Langerhans cell histiocytosis. Anteroposterior radiograph of the lower leg of a 4-year-old boy demonstrates a lesion in the diaphysis of the left tibia (*arrows*) exhibiting a permeative type of bone destruction and a lamellated (onion-skin) type of periosteal reaction similar to that seen in osteomyelitis and Ewing sarcoma.

bral body (10) (Fig. 5-4). This finding was for a long time mistakenly considered an osteochondrosis of the vertebra and was (and in the European literature still is) named Calvé disease after its proposer (10).

In the long bones, LCH presents as a radiolucent destructive lesion, commonly associated with a lamellated periosteal reaction (27,29,67) (Fig. 5-5). This appearance may mimic that of a round cell malignant tumor, such as lymphoma or Ewing sarcoma. The lesion may have well-defined or poorly defined margins, with or without sclerotic borders (11). Occasionally, a cluster of partially overlapping lesions may be observed. In such instances, the superimposed areas of destruction have the appearance of a hole within a hole (55,56). A distinct and characteristic feature of many lesions is slanting or beveling of the edges (38). Sometimes the lesion is poorly demarcated and its indistinct borders gradually merge with the adjacent normal bone. The lesion may affect only the cortical surface of bone (21), although more commonly it is intramedullary, encroaching on the endocortex and causing scalloping of the endosteal surface (34) (Fig. 5-6). In later stages the lesion becomes more sclerotic, with dispersed radiolucencies (Fig. 5-7). The periosteal reaction is either absent or more defined and appears nonaggressive. Uhlinger (73) and later Mirra and Gold (52) have distinguished three phases in the evolution of LCH: incipient, midphase, and late phase. During the early phases the lesions tend to have aggressive patterns, with a lamellated periosteal reaction and poorly marginated or permeative lytic lesions. During the late phase the lesions tend to become more circumscribed (52).

The distribution of the lesion may be determined by radionuclide bone scan, which can aid in the detection of silent lesions and in differentiating LCH from Ewing



Figure 5-6 Langerhans cell histiocytosis. Anteroposterior radiograph of the proximal femur of a 3-year-old boy with a limp and tenderness localized to the upper thigh shows an osteolytic lesion without sclerotic changes, causing scalloping of the lateral endocortex. There is fusiform thickening of the cortex and a solid uninterrupted type of periosteal reaction.



Figure 5-7 Langerhans cell histiocytosis. The healing stage of the lesion, as seen here in the distal humerus of a 16-year-old girl, exhibits predominantly sclerotic changes with interspersed radiolucent foci, thickening of the cortex, and a well-organized periosteal reaction. In this stage, the lesion mimics chronic osteomyelitis.

sarcoma, which rarely has multiple foci (43,69). The study has its limitations, however, because approximately 35% of lesions show a normal uptake of the radiopharmaceutical tracer (36). Bone destruction may sometimes result in inadequate residual bone to produce uptake, and areas of photon deficiency are exhibited (2). Recent reports indicate the usefulness of thallium-201 scintigraphy, which proved to be positive in a lesion that exhibited photon deficiency on technetium-99m methylene diphosphonate (^{99m}Tc-MDP) scintigraphy (26).

Computed tomography (CT) may be useful if radiography inadequately defines the extent of the process (53), particularly in cases of spine and pelvic involvement (33). This modality effectively demonstrates periosteal reaction, beveled edges, and reactive sclerosis.

There have been isolated reports of the usefulness of magnetic resonance imaging (MRI) in evaluating this condition (5,40). The MRI appearance varies and appears to correlate with the radiographic appearance (11). The MRI manifestations of LCH during the earlier stages are nonspecific and may simulate an aggressive lesion, such as osteomyelitis or Ewing sarcoma, and occasionally benign tumors, such as osteoid osteoma or chondroblastoma (5). After gadoliniumdiethylenetriamine-penta-acetic (DTPA) injection, the lesions show marked enhancement on T1-weighted images (19) (Fig. 5-8). Occasionally, MRI can demonstrate early bone marrow involvement in the absence of radiographic or scintigraphic abnormalities (45). In some studies, on T1-weighted sequences the lesions were isointense with adjacent structures (54,58). In the skull, lesions have been reported to show well-defined high-signal-intensity areas of marrow replacement on T2-weighted sequences. The most recent investigations have shown that the most common MRI appearance of LCH is that of a focal lesion, surrounded by an extensive, ill-defined signal from bone marrow and by soft tissue reaction with low signal intensity on T1-weighted images and high signal intensity on T2-weighted images, considered to represent bone marrow and soft tissue edema or the flare phenomenon (5,29) (Fig. 5-9).

Histopathology

Histologically, LCH consists of agglomerates of more or less densely arranged large rounded cells without cytoplasmic extensions, thus distinguishing them from epidermal Langerhans cells (Fig. 5-10). Electron microscopic observations proved that this pathognomonic cell was identical to the Langerhans cells seen in the skin (70,78). These cells are plump and pink, with a lightly granular cytoplasm and dendritic extensions. Their nuclei are translucent, ovoid, or kidney bean-shaped, have distinct nuclear membranes, and may exhibit longitudinal grooves. Langerhans cells can be specifically identified in electron microscopy by the presence of racquet-shaped cytoplasmic organelles known as Birbeck granules (6,70), also referred to as Langerhans cell granules or X granules (45) (see Fig. 1-43). In LCH, osteoclast-like giant cells are present, usually with five to seven nuclei (13). They may either represent osteoclasts, as indicated by their immunoprofile (positive for CD68, tartrate-resistant acid phosphatase, vitronectin receptor, cathepsin K, and metalloproteinase 9) or, because some giant cells also express CD1a, especially in nonbony lesions such as skin and lymph nodes, may be formed at a later stage when mononuclear bone marrow-based progenitor cells, capable of differentiating to osteoclasts or dendritic cells, have already switched to the dendritic pathway (3,66).

Langerhans cells express a variety of surface antigens. The IHC demonstration of CD1a, S-100 protein, and Langerin/CD207 and the electron microscopic (EM) identification of Birbeck granules are the most useful markers for Langerhans cells (8,20,32,62,74).

Zones of necrosis are common. In older or multiple lesions, lipid-bearing foam cells can also be observed (68) (Fig. 5-11). Special stains can reveal abundant droplets of sudanophilic fat peripherally or in the middle of the giant cell cytoplasm (so-called Touton cells) (68). The granulomatous lesions develop during the first (incipient) phase (52,73). During the second phase (midphase), more and more eosinophilic granulocytes



A,C

Figure 5-8 Langerhans cell histiocytosis: magnetic resonance imaging (MRI). A: Anteroposterior radiograph of the right femur of a 13-year-old boy shows a radiolucent lesion in the proximal femoral diaphysis exhibiting a lamellated periosteal reaction. **B:** Axial T1-weighted (SE, TR 600, TE 14) MRI demonstrates the lesion to be of low signal intensity. The cortex is markedly thickened. **C:** Coronal T1-weighted (SE, TR 500, TE 15) MRI obtained after intravenous injection of gadolinium shows marked enhancement of the lesion and the soft tissues adjacent to the thickened femoral cortex. **D:** Axial T1-weighted (SE, TR 700, TE 18) MRI obtained after intravenous injection of gadolinium shows enhancement of both granuloma and perilesional edema.

containing lobulated nuclei and coarse eosinophilic granules infiltrate the granuloma, finally forming dense agglomerates, the so-called eosinophilic pseudoabscesses (Fig. 5-12). Sparse infiltrates of lymphocytes and plasma cells may be found, as well as giant cells and foci of hemorrhage. The phagocytic capacity of mononuclear and multinucleated histiocytic cells can be demonstrated by their ingestion of erythrocytes, eosinophils, and hemosiderin granules (38). The cytoplasm of the phagocytic cells often exhibits double-refractile neutral fat deposition (38). The eosinophilic granulocytes then progressively disappear. Some of these features are displayed to better advantage with methylene blue and Giemsa preparations. In areas where the eosinophilic leukocytes are undergoing fragmentation, proteinaceous crystalline structures, the so-called Charcot-Leyden crystals, are demonstrable (68). In the third phase (late phase), fibroblasts, probably under the influence of transforming growth factor beta, a major

mediator of fibrosis (12,17), produce collagen with concomitant reduction and final loss of Langerhans cells. The lesion then becomes indistinguishable from a focus of unspecific chronic fibroblastic osteomyelitis.

п

The so-called Langerhans cell sarcoma represents an exceedingly rare but very aggressive form of LCH with multiorgan involvement (75). It can arise de novo or may progress from typical LCH. The cells present with obvious malignant features such as enlarged nucleoli, unevenly distributed nuclear chromatin, and a high mitotic rate. By immunohistochemistry (IHC) they express CD1a and S-100 but sometimes only focally (74).

Differential Diagnosis

Radiology

The differential diagnosis depends on the site and extent of bone involvement, the phase of the disease, and



Figure 5-9 Langerhans cell histiocytosis: magnetic resonance imaging (MRI). A: Anteroposterior radiograph of the left tibia of a 20-year-old woman shows an ill-defined osteolytic lesion in the proximal part of the bone (*arrow*). **B:** T1-weighted sagittal MRI shows a lobulated lesion exhibiting a low signal intensity. **C:** T1-weighted axial MRI shows that the lesion pene-trated the posterior cortex and extended into the soft tissues. **D:** T2-weighted axial MRI shows high signal intensity of the lesion in and outside the tibia. In addition, a high signal intensity soft tissue reaction surrounds the bone. (Reprinted with permission from Greenfield GB, Arrington JA. *Imaging of bone tumors: a multimodality approach.* Philadelphia: JB Lippincott, 1995:417.)

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Figure 5-10 Histopathology of Langerhans cell histiocytosis. A: Densely arranged large Langerhans cells and rounded giant cells are infiltrated by numerous eosinophilic granulocytes (hematoxylin and eosin, original magnification ×100). **B:** At higher magnification large pale histiocytes (Langerhans cells) and numerous roundish giant cells containing up to ten nuclei (*left*) are clearly discernible. In addition, sparse lymphocytes and eosinophils are present (hematoxylin and eosin, original magnification ×400). **C:** At high power, the indentations of oval nuclei are clearly visible (hematoxylin and eosin, original magnification ×630).







Figure 5-11 Histopathology of Langerhans cell histiocytosis. Mulberry-like foam cells (*left and upper right*), eosinophilic granulocytes, and hyaline bodies prevail in this field of view (hematoxylin and eosin, original magnification \times 50).

the age of the patient (63,70). In a solitary lesion in long or flat bones, the main diagnostic considerations are Ewing sarcoma and osteomyelitis (see Fig. 5-5). Both lesions may exhibit moth-eaten or permeative types of bone destruction and lamellated, onion-skin periosteal reaction. If a solitary lesion of LCH in a long tubular bone does not develop a periosteal reaction, a differential diagnosis of a *simple bone cyst* must be considered. However, the characteristic clinical features of the former lesion should always be taken into consideration: the so-called tempo phenomenon (38) is a useful sign, i.e., progression and disappearance of the bony lesion is very rapid. The differential diagnosis of a solitary skull lesion in a child or young adult should include osteomyelitis, hemangioma, and fibrous dysplasia. In an elderly patient, the considerations are the lytic phase of Paget disease (osteoporosis circumscripta), myeloma, and a metastasis. The beveled edge or double-contoured appearance of the lesion is always highly suggestive of LCH.

A lytic expansive lesion in the pelvis may mimic *fibrous dysplasia, aneurysmal bone cyst, brown tumor of hyperparathyroidism,* or *hemophilic pseudotumor.* Disseminated

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Figure 5-12 Histopathology of Langerhans cell histiocytosis. A: In a later stage of development, nonspecific inflammatory cells, particularly lymphocytes, predominate. Some giant cells are also present (hematoxylin and eosin, original magnification imes100). **B:** In another field, Langerhans cells exhibit a more spindle-shaped appearance. Scattered eosinophils and some giant cells are also present (hematoxylin and eosin, original magnification ×200). C: Tumor cells are positive for S-100 protein, both cytoplasmic and nuclear (biotin-streptavidin peroxidase, original magnification $\times 200$). **D**: On higher magnification observe cytoplasmic and membranous positivity for CD1a in Langerhans cells (ABC peroxidase, original magnification \times 400).

osseous lesions in infants should be distinguished from infantile myofibromatosis (64), in adolescents and young adults must be distinguished from multifocal osteomyelitis, leukemia, lymphoma, cystic angiomatosis, fibrous dysplasia, brown tumors of hyperparathyroidism, and in older patients from metastatic disease and multiple myeloma.

Pathology

The major histologic differential diagnostic condition is infection because osteomyelitis is among the diseases capable of producing a polymorphous infiltrate of several inflammatory cell types. Moreover, there is an inflammatory reaction that simulates osteomyelitis, particularly during the midphase of LCH, and it is therefore necessary to correlate histologic findings with the radiographic presentation before rendering a diagnosis. Furthermore, when Langerhans cells and eosinophils have disappeared in the late phase, chronic productive osteomyelitis cannot be differentiated by histologic means.

Another diagnostic consideration should be Hodgkin lymphoma, which can present as a solitary osseous defect with a mixed inflammatory infiltrate. The identification of Reed-Sternberg cells and the absence of Langerhans cells in the latter serve as positive diagnostic clues. Furthermore, because the presentation of Hodgkin lymphoma as a primary disease in bone is exceedingly rare, clinical history is helpful (37,42).

During the involutional stage of LCH, the eosinophils markedly diminish in number and disappear altogether as the lesion progresses (matures). The large mononucleated histiocytes may become lipid-laden in this stage. The presence of those foci of foam cells may lead to the mistaken diagnosis of xanthoma of bone, juve-



nile xanthogranuloma of bone, or Chester-Erdheim disease (7,14,15,18).

The radiologic and pathologic differential diagnosis of LCH is depicted in Figure 5-13.

Malignant Tumors

Round small cell malignancies of bone form a heterogeneous group of neoplasms, distinguished from most other malignancies of bone by the fact that the tumors form a pure cellular growth without production of a tumor matrix (90). The common denominator is an undifferentiated, round cell, basophilic, cytoplasmpoor, stroma-poor, highly cellular tumor (144). These malignancies include primary bone tumors such as Ewing sarcoma and primitive neuroectodermal tumor (PNET), lymphoma, mesenchymal chondrosarcoma, myeloma, metastatic neuroblastoma, and some other metastatic lesions (Table 5-1).

Ewing Sarcoma

Ewing sarcoma, along with Askin tumor and PNET, is now regarded as a single entity (Ewing sarcoma family of tumors) (113). These lesions are characterized by various degrees of neuroectodermal differentiation and by common histologic, immunohistochemical, and molecular properties (87,110,113,121,122,148).

Ewing sarcoma is the prototype of round, small cell malignancies of bone. It is the sixth most common malignant tumor, comprising approximately 11% (138)

to 12% (127) of all malignant tumors of bone, with a rather uniform histologic appearance. Conventional Ewing sarcoma is composed of densely packed small cells with round nuclei but without distinct cytoplasmic outlines (138). It exhibits no microscopic evidence of matrix production (97). Occasionally, similar tumors can arise as primary neoplasms of soft tissue. These are referred to as extraskeletal or soft tissue Ewing sarcomas (95). Even less commonly, Ewing sarcoma may exhibit periosteal location (82). There is also a rare form of Ewing sarcoma in which the cells are larger and more pleomorphic than those of conventional Ewing tumor. This form has been referred to as atypical Ewing sarcoma or large cell Ewing sarcoma (129). The clinical behavior and prognosis of large cell Ewing sarcoma appear similar to those of conventional tumor. Furthermore, single cases of so-called adamantinoma-like Ewing sarcomas have been described. These sarcomas present with tumor cell nests or anastomosing cords of tumor cells and strong cytokeratin positivity but reveal the translocation typical of Ewing sarcomas (see later), which is not present in adamantinoma of long bones (85,106). Very rarely, Ewing sarcoma may be multifocal (88).

All tumors of the Ewing family are characterized by recurrent chromosomal translocations involving chromosomes 11 and 22 [t(11;22)(q24;q12)] or chromosomes 21 and 22 [t(21;22)(q22;q12)] in about 85% or 15% of cases, respectively. In less than 1% of cases, translocations involving chromosomes 22 and chromosomes 2, 7, or 17 and inversions of chromosome 22 (i.e., breakage of a single chromosome with consecutive reversed insertion) have been described. These

Tumor	Histopathologic Features								
Ewing sarcoma	Small, uniform-sized cells with almost clear cytoplasm, round, slightly hyperchromatic nuclei. Cell borders indistinct.								
Large cell (atypical) Ewing sarcoma	Larger cells, with better delimited contours, more cytoplasm with eosinophilic "histiocytoid" appearance, nuclei may contain very prominent nucleoli; atypical variant merges into small cell Ewing sarcoma.								
Primitive (peripheral) neuroectodermal tumor (PNET) of bone	Similar to Ewing sarcoma. Focal Homer-Wright rosettes, often in tumor association with a fibrillary intercellular background. Reticulin fibers surrounding large groups of cells in a basket-like distribution.								
Askin tumor of bone (thoracopulmonary or thoracospinal variant of PNET)	Similar to Ewing sarcoma. Focal Homer-Wright rosettes. Compact sheets of cells with dark nuclei; nesting arrangement of cells with intervening fibrovascular stroma; serpiginous bands of cells with necrosis. May or may not display neural differentiation.								
Malignant lymphoma	Aggregates of malignant lymphoid cells, typically mixture of small lymphocytic cells and large histiocytic cells. Cells are rounded, ovoid, and pleomorphic, displaying basophilic cytoplasm. Nuclei contain scanty chromatin, sometimes cleaved or horseshoe-shaped with prominent nucleoli.								
Myeloma	Sheets of atypical plasmacytoid cells frequently bi- and trinucleated. Round nuclei possess dense, coarse chromatin with cartwheel-like distribution. Prominent nuclei eccentrically located in a strongly basophilic cytoplasm.								
Small cell osteosarcoma	Loose aggregations of small round cells with ovoid nuclei separated by collagenous bands of fine eosinophilic matrices. Occasional spindling of tumor cells. Focal production of osteoid or bone.								
Mesenchymal chondrosarcoma	Small, round, uniform-sized cells with round or ovoid nuclei and scant cytoplasm, occasionally interspersed with spindle-shaped cells. Areas of well-differentiated cartilage containing foci of calcification.								
Metastatic neuroblastoma	Very similar to PNET, including Homer-Wright rosettes; a fibrillary intercellular background; long, thin tapering cytoplasmatic extensions creating appearance of a "pear" or "carrot" cells; and higher degree of neural differentiation.								
Metastatic primitive alveolar rhabdomyosarcoma	Similar to Ewing sarcoma. Alternating areas of cellularity and myxoid changes. Round or oval, occasionally spindled or tapered cells with scant eosinophilic cytoplasm.								

Table 5-1 Small Round Cell Tumors of Bone

Modified from Triche T, Cavazzana A. Round cell tumors of bone. In: Unni KK, ed. *Bone tumors,* New York: Churchill Livingstone, 1988;199–223; from Meis-Kindblom JM, Stenman G, Kindblom L-G. Differential diagnosis of small round cell tumors. *Semin Diagn Pathol* 1996;13:213–214; and from Llombart-Bosch A, Contesso G, Peydro-Olaya A. Histology, immunohistochemistry, and electron microscopy of small round cell tumors of bone. *Semin Diagn Pathol* 1996;13:153–170.

transcripts give rise to fusions between EWS (for Ewing sarcoma) gene on chromosome 22q12 and members of the ETS family of transcription factors (ETS is derived from a sequence that was detected in an avian erythroblastosis virus, E26, where it formed a transforming gene; it was therefore called E26 transformation specific sequence or ETS) like *Fli1* (for *F*riend *l*eukemia virus integration 1; present in about 85%), ERG (early response gene; present in about 15%), ETV1 (or ETS translocation variant-1), E1A-F (for E1A factor, a binding protein of the adenovirus E1A gene), and FEV (for fifth Ewing variant), or ZSG (for zinc finger sarcoma gene), a transcription activator (136,148). As a result, chimeric proteins are expressed, which are believed to act as aberrant transcription factors that may upregulate or downregulate Ewing sarcoma target genes. The gene for the transforming growth factor (TGF) beta II receptor, a putative tumor suppressor gene, has been identified as such a target that is downregulated by the EWS/Fli1 transcript (105). Recently, additional putative target genes of EWS/Fli1 have been recovered from DNA of Ewing sarcomas (140).

In about 20% of cases of Ewing sarcoma, the second most common genetic alteration is the inactivation of the gene p16 or *INK4A*, which is involved in cell-cycle control and is responsible for preventing cells from entering the S-phase by suppressing the activity of the retinoblastoma protein (116). Wei et al. (152) suggested that p16 deletions represent a significant negative predictive factor in Ewing sarcoma. This has recently been confirmed by the same group and has been shown for mutations of *TP53*, which are present in about 10% of cases (108). In contrast, the most common type of *EWS/Fli1* fusion (type 1 fusion between *EWS* exon 7 and *Fli1* exon 6) is associated with a better prognosis (148).

Clinical Presentation

Ewing sarcoma occurs primarily in young people, most commonly in the second decade. Approximately 90% of cases present before the age of 20 years, with a median age of about 13 years (130). The tumor has a decisively male predominance (3:2), and only exceptionally occurs in black individuals (138). Clinically, Ewing sarcoma may present as a localized, painful mass or with systemic



Figure 5-14 Ewing sarcoma: skeletal sites of predilection, peak age range, and male-to-female ratio.

symptoms such as fever, malaise, weight loss, leukocytosis, and increased erythrocyte sedimentation rate (115), a constellation of findings that may lead to an erroneous diagnosis of osteomyelitis (99). It has been speculated that the presence of systemic symptoms indicates dissemination of the tumor, with a consequently worsened prognosis (83). Diaphyses of the long bones (predominantly femur, tibia, and humerus), the ribs, and the flat bones, such as the scapula and the pelvis, are preferred sites (90,118) (Fig. 5-14). Although metaphyses of the long bones occasionally may be affected (135), an epiphyseal location is rare (2%) (125,138). The small bones of the hands and feet can also be affected (81). According to Ilaslan et al. (109), 9.8% of all cases of Ewing sarcoma has a primary vertebral origin. Pathologic fractures are uncommon (97). Occasionally, Ewing sarcoma may metastasize to the other bones.

Imaging

In most cases, the radiographic presentation of Ewing sarcoma is quite characteristic. It consists of an illdefined lesion with a permeative or moth-eaten pattern of bone destruction associated with a lamellated periosteal new bone formation that has an onion skin (or "onion peel") appearance, or, less commonly, a "sunburst" ("trimmed whiskers" effect) configuration (101), and a large soft tissue mass (Fig. 5-15). Ewing sarcoma occasionally presents as a large area of geographic bone destruction, with or without matrix mineralization, simulating any other type of bone sarcoma. At times the lesion in the bone is almost imperceptible, and the only prominent finding is a soft tissue mass (Fig. 5-16). The lesion sometimes creates a typical "saucerization" of the cortex, a feature once believed to be virtually pathognomonic for this tumor (Fig. 5-17). The saucerization may be related to the combination of destruction of the periosteal surface of bone by the tumor and an associated extrinsic pressure effect of the large soft tissue mass (132). This sign has recently been reported in other tumors and even in osteomyelitis. However, the presence of saucerization associated with a permeative lesion and a soft tissue mass favors the diagnosis of Ewing sarcoma. When the lesion exhibits a permeative appearance, it may be indistinguishable from osteomyelitis (Fig. 5-18). Because Ewing sarcoma does not produce a matrix material, radiography reveals a lack of mineralization in the substance of the tumor. However, because there is often an abundant periosteal new bone formation, particularly in the flat bones, this appearance may mimic osteosarcoma, and differentiation between the two tumor types may be difficult (Fig. 5-19). Approximately one third of the cases affecting flat bones demonstrate a diffuse sclerosis. Unlike osteosarcoma, this sclerosis represents not a tumor bone but rather reactive bone to the tumor cell infiltration (139).

On radionuclide bone scan, Ewing sarcoma shows a very intense increase of technetium-99m methylene diphosphonate (^{99m}Tc-MDP) accumulation (128). Gallium-67-citrate (⁶⁷Ga-citrate) more readily identifies soft tissue tumor extension (94,96). Although scintigraphic findings are nonspecific, this technique



Figure 5-15 Ewing sarcoma. A: Lateral radiograph of a 12-year-old boy shows the typical appearance of this malignancy in the diaphysis of the fibula (arrows). The poorly defined lesion exhibits a permeative bone destruction associated with an aggressive periosteal reaction. **B:** CT section through the tumor demonstrates a large soft tissue mass (arrowheads), not well depicted on the routine study. Note the complete obliteration of the marrow cavity by tumor.



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Figure 5-16 Ewing sarcoma. A: Bone destruction (arrows) is almost imperceptible on this magnification study of the distal femoral diaphysis of a 10-year-old girl. **B:** Lateral radiograph of the distal femur, however, shows a large soft tissue mass adjacent to the posterior cortex.





Figure 5-16 *Continued* **C:** Computed tomography using a bone window demonstrates destruction of the medullary portion of the bone and invasion of the cortex.





Figure 5-17 Ewing sarcoma. Anteroposterior radiograph of the right femur of a 12-year-old girl shows saucerization of the medial cortex of the diaphysis, often seen in Ewing sarcoma. Note a large soft tissue mass (*arrowheads*).

Figure 5-18 Ewing sarcoma resembling osteomyelitis. A 24-year-old man presented with pain and swelling of the left ankle for 8 weeks; he also had a fever. Anteroposterior radiograph of the ankle demonstrates a lesion of the distal fibula exhibiting a permeative type of bone destruction and a lamellated periosteal reaction. The appearance is that of infection (osteomyelitis), but biopsy revealed Ewing sarcoma.

provides reliable information concerning the presence of skeletal metastases (112,127).

CT reveals the pattern of bone destruction, and attenuation values (Hounsfield units) provide information about the medullary extension (94). In addition, CT may help to delineate extraosseous involvement (149) (see Fig. 5-15B).

MRI is essential for definite demonstration of the extent of intraosseous and extraosseous involvement by this tumor (84,150) (Fig. 5-20). In particular, MRI may effectively reveal extension through the epiphyseal plate (100). T1-weighted images show intermediate to low signal intensity, which becomes bright on T2 weighting. Hypocellular regions and areas of necrosis are of lesser intensity (94). Imaging after injection of gadolinium (Gd-DTPA) reveals signal enhancement of the tumor on T1-weighted sequences (100). Enhancement occurs only in the cellular areas, allowing differentiation of the tumor from the peritumoral edema.

Histopathology

The gross appearance of Ewing sarcoma may be misleading (155). Because the lesion does not produce any matrix, its appearance can range from a soft and fleshy but solid mass to an almost liquid consistency. When a tumor of the latter consistency is cut into at the time of surgery, it may actually run like pus. This **328** — Differential Diagnosis in Orthopaedic Oncology



Figure 5-19 Ewing sarcoma resembling osteosarcoma. Anteroposterior (A) and lateral (B) radiographs of the left femur of a 17-year-old boy, displaying a significant sclerosis of the tumor that was originally interpreted as osteosarcoma.

can lead to a mistaken diagnosis of osteomyelitis, and the material may be sent for microbiologic culture rather than histopathologic examination, thus causing a delay in diagnosis (83).

The microscopic appearance of Ewing sarcoma is typical. At lower magnification, the tumor consists of small, uniform-sized cells characterized by an almost clear to light eosinophilic cytoplasm and nuclei that are round and slightly hyperchromatic. Prominent septa composed of fibrous connective tissue divide the tumor tissue into strands or lobules (138) (Fig. 5-21A). Because the cell borders are indistinct, it sometimes appears as if the tumor is composed of many nuclei floating in a sea of cytoplasm rather than being separated into discrete cells. The nucleoli are usually inconspicuous. Areas of necrosis and hemorrhage are often observed (120). At higher magnification, the nuclei appear round and uniform (Fig. 5-21B-D). Sometimes the cytoplasm contains a moderate amount of glycogen that can be demonstrated by periodic acid–Schiff (PAS) staining (137) (Fig. 5-22) or, more specifically, with Best carmine stain with and without diastase digestion prior to staining, which will remove glycogen. Mitotic figures are uncommon in classical Ewing sarcoma (138). Although no extracellular matrix is found between the tumor cells, and reticulin fibers are almost always absent (Fig. 5-22C), reactive new bone formation may be observed, especially at the periphery. This reactive bone is sometimes difficult to distinguish from tumor-bone formation, and small cell osteosarcoma may therefore be erroneously diagnosed.

In extremely rare cases, Ewing sarcoma exhibits an epithelial differentiation similar to adamantinoma of the long bones (93,103,124,126,141,151). Other primary tumors of bone known to have epithelial features are adamantinoma of long bones (89), malignant fibrous histiocytoma (153), and osteosarcoma (91,117).

Electron microscopy of this lesion reveals highly characteristic ultrastructural features (133,142). The predominant or principal cell type is polygonal. The cytoplasm exhibits few organelles, with sparse rough endoplasm reticulum, and few mitochondria. However, a conspicuous Golgi complex is often present. The nuclei of these cells are round or oval. Typically, the chromatin is fine and evenly distributed, and there are single or multiple small nucleoli. The cytoplasm may contain many glycogen granules. In addition to



Figure 5-20 Ewing sarcoma: magnetic resonance imaging (MRI). Anteroposterior (A) and lateral (B) radiographs of the right distal femur of a 7-year-old girl show permeative type of bone destruction in the metaphysis and diaphysis associated with a large soft tissue mass. Coronal (C) and sagittal (D) T1-weighted (SE, TR 750, TE 20) MR images demonstrate the intraosseous and extraosseous extent of the tumor. E: Axial T2-weighted (SE, TR 2000, TE 80) MRI shows heterogeneous but mostly high signal intensity of the soft tissue mass.

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Figure 5-21 Histopathology of Ewing sarcoma. A: Densely packed small cells with round nuclei and indistinct cell borders are characteristic for this tumor. To the right a prominent septum composed of fibrous connective tissue is visible (hematoxylin and eosin, original magnification \times 50). **B:** At high magnification the uniform aspect of the round nuclei without clear cytoplasmic outlines is striking (hematoxylin and eosin, original magnification \times 100). **C:** With Giemsa staining the cytoplasm is more distinct. The nuclei are darkly and homogeneously stained (Giemsa, original magnification \times 100). **D:** With alcohol fixation, in this case there was better preservation of cell details (hematoxylin and eosin, original magnification \times 100).

the principal cell, there may be a small number of socalled dark cells (reflecting a condensed chromatin) (119, 138). It is still undetermined whether this cell is a precursor or, more likely, a terminal form of the principal cell.

IHC reveals that almost all Ewing family tumors exhibit a positive membranous or cytoplasmic reaction for CD99 and vimentin, respectively (102) (Fig. 5-23). Because CD99 positivity [a transmembrane glycoprotein involved in cell adhesion, apoptosis and differentiation, encoded by the Mic2 gene which is located on both x and y chromosomes as a pseudoautosomal gene at the end of their short arms (114)] is not specific for Ewing tumors, additional markers covering the spectrum of "small round blue-cell" tumors must be applied (see later). To diagnose a given tumor as PNET, more than one finding of neural differentiation must be present, i.e., Homer-Wright rosettes by light microscopy or positive immunoreactions for two neural markers [neuron-specific enolase (NSE), synaptophysin, neurofilament, S-100, CD57, protein gene product 9.5 (PGP9.5)], at least in some tumor cells (Fig. 5-24), or neurosecretory granules within cytoplasmic processes must be demonstrated by ultrastructural analysis (131).

Differential Diagnosis

Radiology

On radiography, the most important differential consideration is to distinguish Ewing sarcoma from *osteomyelitis* and *LCH* because both of these entities may present with a similar aggressive moth-eaten or permeative pattern of bone destruction (see Fig. 5-18). It is not uncommon for all three conditions to appear identical. The clinical information, e.g., duration of the patient's symptoms, aids in radiographic differential diagnosis. In LCH, for example, the amount of bone destruction seen on radiography after 1 week of symptoms is usually the same as that seen after 4 to 6 weeks of symptoms in osteomyelitis and after 3 to 4 months in Ewing sarcoma (104). The other look-alike



Figure 5-22 Histopathology of Ewing sarcoma: periodic acid–Schiff (PAS) staining. A: Densely packed small cells with dark round nuclei infiltrate the muscle (*bottom*). The cells are somewhat artificially deformed. No glycogen has been preserved in the cells because of formaldehyde fixation (PAS, original magnification ×100). B: Fixed in ethanol, the glycogen granules in the cytoplasm of the tumor cells were stained red with PAS method (PAS, original magnification ×100). C: Tumor cells do not form reticulin fibers; only the preexisting fibers of the infiltrated tissue are stained dark brown to black (Novotny, original magnification ×200).





Figure 5-23 Histopathology of Ewing sarcoma: immunohistochemistry (IHC). IHC reaction with CD99 is positive, but there is no reaction in endothelial cells (*center*) (biotinstreptavidin peroxidase, original magnification ×400).



reaction, and displays (except for purely lytic lesions, such as telangiectatic or fibroblastic variants) bone formation within destructive lesions in the bone and within a soft tissue mass. It should be stressed, however, that Ewing sarcoma may occasionally exhibit reactive sclerotic changes, thus mimicking bone-forming tumors (see Fig. 5-19).

Lymphoma must also be included in the differential diagnosis, although this lesion usually occurs in older age groups. The important radiologic difference is usually the absence of a soft tissue mass in lymphoma (see Fig. 5-28), whereas in Ewing sarcoma a soft tissue mass is almost invariably present, often being disproportionally large compared with the amount of bone destruction (see Fig. 5-16). The distinction between Ewing sarcoma and *PNET* cannot be made on the basis of radiography (107,134), and differentiation between these two lesions must rely entirely on histologic examination (127). *Metastatic neuroblastoma*, which has a similar appearance on radiography and simulates Ewing sarcoma, invariably affects patients younger than 5 years.

Although recently Sundaram et al. (143) reported a case of Ewing sarcoma of the humerus mimicking *fibrous dysplasia* on imaging and biological behavior, from a practical point of view the latter lesion should not be seriously considered in the differential diagnosis.

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Figure 5-24 Histopathology of primitive neuroectodermal tumor (PNET). A: Observe newly formed periosteal bone with extensive infiltration (hematoxylin and eosin, original magnification ×50). B: Nuclear and cellular details are similar to Ewing sarcoma (hematoxylin and eosin, original magnification ×400). C: Almost all tumor cells are positive for neuron-specific enolase (NSE) (biotin-streptavidin peroxidase, original magnification ×200). D: Same tumor as in C; some cells also exhibit a positive reaction for synaptophysin (biotin-streptavidin peroxidase, original magnification \times 400).

Pathology

On histologic examination, Ewing sarcoma should be differentiated from malignant lymphoma. In the latter, the nuclei are larger and less uniform than those in Ewing sarcoma, with distinct indentations and lobulations and often with enlarged nucleoli. A more or less dense net of reticulin fibers usually surrounds the cells. This is in contrast to Ewing sarcoma, in which the fibers are usually limited to the septa of fibrous tissue (123). It is entirely possible for the large cell variant of Ewing sarcoma to be mistaken for large cell lymphoma, which is by far the most common subtype of primary lymphoma of bone. The presence of intracellular glycogen and the lack of reticulin fibers, except for those in the septa in Ewing sarcoma, should help in the diagnosis of Ewing sarcoma (97).

In addition to malignant lymphoma, the differential diagnosis of Ewing sarcoma includes other small round

cell malignancies, such as *metastatic neuroblastoma* (144, 145). However, neuroblastomas usually occur in the first 3 years of life, whereas Ewing sarcoma is uncommon in patients younger than 5 years. Furthermore, most neuroblastomas are characterized by a neural filamentous material that is absent in Ewing sarcoma, and the former malignancy is accompanied by elevated levels of urinary catecholamines. Among the light microscopic clues are the presence of Homer-Wright rosettes (which may also be present in PNET), a fibrillary intercellular background, and the tapering cytoplasmic processes that are present in metastatic neuroblastoma (97). Ewing sarcoma must also be differentiated from osteomyelitis. Chronic osteomyelitis with a preponderance of plasma cells or lymphocytes might occasionally be mistaken for Ewing sarcoma. Again, the variegated histologic picture with granulocytes, should allow distinction.

Other entities to be differentiated from Ewing sarcoma are small cell osteosarcoma and mesenchymal

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Figure 5-25 Histopathology of small cell osteosarcoma. A: Loosely arranged tumor cells with hyperchromatic nuclei and some inconspicuous needle-like osteoid particles (*upper left and right*) make recognition of bone-forming tumor difficult (hematoxylin and eosin, original magnification \times 50). B: On the left at higher magnification, the pleomorphism of the dark nuclei and the narrow cytoplasm seams betray the malignancy of the cells (hematoxylin and eosin, original magnification \times 100). On the right, with reaction for osteonectin, most cells show a more or less distinct positivity (ochreous yellow) pointing to their malignant osteoblastic character (osteonectin, original magnification \times 100). (Courtesy of Professor Klaus Remberger, University of Saarland, Germany.)

chondrosarcoma. The distinguishing feature of small cell osteosarcoma is the presence of foci of osteoid (Fig. 5-25; see also Fig. 2-85). Mesenchymal chondrosarcoma is characterized by foci of cartilaginous matrix, although they may be sparse. In such instances, the demonstration of the t(11;22) translocation by fluorescence in situ hybridization (FISH) or reverse transcriptase polymerase chain reaction (RT-PCR) is crucial.

Undifferentiated round cell areas of *synovial sarcoma* might also be mistaken for Ewing sarcoma. They usually contain a delicate network of reticulin fibers, which are not present in Ewing tumors. The same is true for

metastatic seminoma, which, on the basis only of histologic staining with hematoxylin and eosin (H&E), may be difficult to differentiate. The uniform appearance of the cells and nuclei, clear cytoplasm, conspicuous nucleoli, and interspersed lymphocytes may help in the diagnosis.

Differential diagnosis of so-called round, blue, small cell tumors by histopathologic examination is usually not possible without the use of additional methods, particularly IHC, electron microscopy, and, recently, molecular biology (98,127,132). For routine histologic diagnosis of these tumors, IHC has almost replaced electron microscopy (Table 5-2). Distinction between

Tumor	bcl-2	ск	Vim	Des	NF	CD99	CD57	NSE	CD45	Light Chains κ or λ	Syn	CD138
Ewing sarcoma		+/a	+	_	—/a	+	(+)	_		_		
PNET/Askin tumor			+	—	(+)	+	(+)	+		—	+	
Malignant												
lymphoma	+	—	+	—	—	(+)	(+)	—	+	+/-	—	(+)
Myeloma	+	—	-/+	—	—	(+)	—	—	-/+	+	—	+
Small cell												
osteosarcoma	—	—	+	—	—	_	—	_	—	_	—	—
Mesenchymai												
Motastatic			+	_		+	_	_		_		
neuroblastoma					+		(+)	+			+	
Rhabdomyosarcoma			(+)	+			(-)	(+)			<u> </u>	
Synovial sarcoma	+	+	+			+	_					(+)
Desmoplastic small						·						
round cell tumor		+	+	+		(+)	+	+	_	_	+	_
						. ,						

Table 5-2 Small Round Cell Tumors of Bone and Typical Immunohistochemistry Results

CK, cytokeratin; Vim, vimentin; Des, desmin; NF, neurofilament; NSE, neuron-specific enclose; Syn, synaptophysin; (+), weak reaction; PNET, primitive neuroectodermal tumor of bone.

^aPositive in single cases in a few cells.

Modified from Triche T, Cavazzana A. Round cell tumors of bone. In: Unni KK, ed. Bone tumors. New York: Churchill Livingstone, 1988:199-223.





Ewing sarcoma, large cell Ewing sarcoma, PNET, and other round small cell tumors is now possible with a fair degree of accuracy (92,111). In IHC, CD99 positivity appears to be highly characteristic for the Ewing tumor group (147,199) (see Fig. 5-23). However, because other round, blue, small cell tumors may also be CD99-positive, a panel of additional markers must be applied to rule out other primary or metastatic tumors, e.g., lymphoma (positive for CD45), rhabdomyosarcoma (positive for myogenin, a transcription factor for skeletal muscle-specific proteins, and desmin), neuroblastoma (strong positive reactions for neurofilament, NSE, synaptophysin), desmoplastic small round cell tumor (positive for coexpressed cytokeratins, vimentin and desmin; NSE), synovial sarcoma [positive for BCL2, epithelial membrane antigen (EMA), cytokeratins], and metastatic seminoma (positive for placenta-specific alkaline phosphatase, CD117, and OCT3/4, a transcription factor that is expressed in pluripotent embryonic germ cells) (98,146). Molecular probes are also available for most of these tumors (154). Other tumors, such as small cell osteosarcoma and mesenchymal chondrosarcoma which, to a lesser or greater extent, are also CD99 positive, need to be excluded by the demonstration of osteoid or cartilage, respectively, within the tumor.

Whenever possible, molecular genetic studies should be conducted to identify the diagnostic fusion transcripts of Ewing tumors. For this purpose, a small aliquot of biopsy material should be stored snap-frozen as recommended, for example, by the College of American Pathologists (86). Histochemical techniques and special stains are rarely employed for differential diagnosis. However, if no frozen material for molecular biology is available and IHC is equivocal, proof of intracytoplasmic glycogen by the PAS or Best reaction combined with diastase pretreatment, as well as absence of reticulin fibers from tumor tissue, may be of diagnostic value.

The radiologic and pathologic differential diagnosis of Ewing sarcoma is depicted in Figure 5-26.

Malignant Lymphoma

The term malignant lymphoma refers to a group of neoplasms that are composed of lymphoid cells in various stages of maturation (177). According to the WHO, malignant lymphomas of bone are subdivided into (a) those that affect one skeletal site, with or without involvement of regional lymph nodes, (b) those that affect multiple bones without lymph node or visceral involvement, (c) those that present as a primary bone tumor but reveal nodal or visceral lesions at staging examinations, and (d) those with a known lymphoma and consecutive positive bone biopsy. Groups (a) and (b) are considered primary lymphomas of bone (209). Primary lymphoma of bone is very uncommon (180). Unni (208) has reported a prevalence of 0.07% for malignant lymphoma of bone (primary and secondary) in the Mayo Clinic series of 8,591 primary malignant bone tumors, with only 3.1% of cases representing primary osseous involvement.

Lymphomas are subdivided into non-Hodgkin lymphoma and Hodgkin lymphoma. Although secondary

involvement of bones is relatively common in Hodgkin lymphoma and especially in non-Hodgkin lymphoma [skeletal involvement is present in approximately 13% of patients who have Hodgkin lymphoma and about 33% who are affected by the latter (186)], primary Hodgkin lymphoma of bone is extremely rare (184). A rare form of lymphoma, seen predominantly in children in tropical Africa, is known as Burkitt lymphoma (see Fig. 5-37). In this type of lymphoma, involvement of the facial bones is particularly characteristic (201). Lesions affecting long tubular bones and flat bones are less common.

Recently, WHO adopted the Revised European-American Classification of Lymphoid Neoplasms (REAL) that originally was proposed by the International Lymphoma Study Group (Table 5-3). This classification defines distinct disease entities based on a combination of morphologic, immunophenotypic, genetic, and clinical features (185a).

Non-Hodgkin Lymphoma

Primary lymphoma of bone is a rare tumor (comprising 3% to 4% of all malignant bone tumors). This lesion was formerly termed reticulum cell sarcoma or lymphosarcoma (158). Lymphomas are considered primary in bone only when a complete systemic workup reveals no evidence of extraosseous involvement other than a regional lymph node. Moreover, some authorities consider that no evidence of extraosseous or nodal disease must be present for at least 6 months after the discovery of the osseous focus for lymphoma to be considered primary in the bone. It should be pointed out, however, that the skeletal system may be secondarily involved in about 30% to 33% of malignant lymphomas (168,186). Non-Hodgkin lymphomas encompass a heterogeneous

spectrum of neoplasms that vary in their clinical and radiologic manifestations. The most common lymphoma subtype with bone marrow involvement, primary and secondary, is follicular lymphoma (about 39%) followed by diffuse large cell lymphoma (about 16%), mantlecell lymphoma (about 9%), low-grade B-cell lymphomas (NOS), and lymphoplasmacytic lymphomas (about 8% each) and T-cell lymphomas (about 6%) (156). The vast majority of primary lymphomas of bone are diffuse large B-cell lymphomas (92%), followed by diffuse follicle center-cell lymphomas, anaplastic large cell lymphomas (3% each), and lymphoplasmacytic lymphomas (2%) (200,206,209). In Southeast Asia, primary T-cell lymphomas in bone appear to be the second most common type following diffuse large B-cell lymphoma (181).

Clinical Presentation

This malignancy occurs in the second to seventh decades, usually involving young and older adults, with a peak incidence between the ages of 35 and 45 years. As a whole, the patient population is distinctly older than the population with Ewing sarcoma, and there is a decisively male predominance, with a reported ratio ranging from 1.5:1, to 2:1, to 3:2 (38,208). Patients may present with local symptoms, such as pain and swelling, or with systemic symptoms, such as fever and weight loss, although some patients may be asymptomatic. Some investigators pointed out that a disproportion between an extensive local disease and overall good clinical condition of the patient with malignant bone lymphoma can be a helpful feature in differential diagnosis (185a). The sites of predilection are the long bones (femur, tibia), the pelvis, the ribs, and the vertebrae (Fig. 5-27), although the small bones of the hands and feet can occasionally be

B-Cell Lymphomas	T-Cell and Natural Killer Cell Neoplasms	Hodgkin Disease
 Precusor B-cell neoplasm Precursor B-lymphoblastic leukemia or lymphoma Mature B-cell neoplasm B-cell chronic lymphocytic leukemia, prolymphocytic leukemia, small lymphocytic leukemia Lymphoplasmacytoid lymphoma Mantle cell lymphoma Follicle center lymphoma Marginal zone B-cell lymphoma Hairy cell lymphoma Diffuse large cell B-cell lymphoma Burkitt lymphoma High-grade B-cell lymphoma 	 Precursor T-cell neoplasm Precursor T-lymphoblastic lymphoma or leukemia Peripheral T-cell and natural killer cell neoplasm T-cell chronic lymphocytic leukemia Large granular lymphocyte leukemia Mycosis fungoides, Sezary syndrome Peripheral T-cell lymphoma Angioimmunoblastic T-cell lymphoma Angiocentric lymphoma Adult T-cell lymphoma Anaplastic large cell lymphoma 	Nodular lymphocyte predominance (paragranuloma) Nodular sclerosis Mixed cellularity Lymphocyte depletion Lymphocyte-rich classic Hodgkin disease

Table 5-3 Revised European-American Lymphoma Classification

Modified from Krishnan A, Shirkhoda A, Tehranzadeh J, et al. Primary bone lymphoma: radiographic–MR imaging correlation. *Radiographics* 2003;23:1371–1387, with permission.



Figure 5-27 Malignant lymphoma: skeletal sites of predilection, peak age range, and male-to-female ratio.

affected (180). Only rarely is the tumor present in a periosteal location (162). Pathologic fractures are observed in approximately 25% of cases (208). Neurologic symptoms may be present in lesions localized in the vertebrae.

Imaging

The roentgenographic appearance of malignant lymphoma can vary widely (159,185a,190,202). Lymphoma tends to involve a large portion of the affected bone and is usually localized in the shaft. The lesion may be marked by a large area of geographic, lytic bone destruction (Fig. 5-28A), may exhibit a moth-eaten pattern (Fig. 5-28B), or may have a very subtle permeative pattern involving the medullary canal (Fig. 5-28C) (165). In the latter case, radiographs may appear almost normal (Fig. 5-29). This lesion may also have an ivory bone appearance, a picture commonly seen in vertebrae (Fig. 5-30) or flat bones, although this presentation is not as common as in Hodgkin lymphoma. Although in the past it was thought that lymphoma rarely evokes any periosteal new bone formation and soft tissue mass, in distinct contrast to the presentation of Ewing sarcoma (174), the recent investigations suggest something different. Mulligan et al. (193), in reviewing 237 pathologically proven cases of primary lymphoma of bone, noted periosteal reaction in 58% and soft tissue masses in 48% of cases. Moreover, Manaster (189) observed a prominent endosteal reaction that is rather characteristic for lymphoma. This reaction, together with periosteal response, caused substantial thickening of the cortex. Furthermore, this author suggested that the character of a soft tissue mass may be helpful in the diagnosis of lymphoma. Whereas the soft tissue mass in most bone sarcomas is usually round or oval, displacing adjacent muscles and neurovascular bundles, the soft tissue mass in primary lymphoma of bone is often disproportionately large, exhibits an infiltrative appearance involving more than one compartment, extends over long distances, and does not appear to be pseudoencapsulated (189).

The radionuclide bone scan is usually positive (187,197) (see Fig. 5-32B), and the combination of a normal radiograph and positive scintigraphy should suggest the possibility of malignant lymphoma (83).

CT can reveal bone destruction and soft tissue involvement, often associated with osseous changes (171,188) (Figs. 5-31 and 5-32). It also can detect bony sequestra, which are estimated to occur in 11% of primary lymphomas of bone (192,193) (Fig. 5-33).

MRI may also be helpful in demonstrating the involvement of the bone marrow (167,169,203,207). On T1-weighted images, diffuse infiltration presents as an area of low signal intensity (211). On T2-weighted images, the affected areas are hyperintense compared with muscle but isointense compared with fat (178, 179,192,194,196). On short time inversion recovery (STIR) images, with suppression of the normally bright marrow fat, the foci of lymphomatous infiltration exhibit a high signal intensity (175, 210). After administration of gadolinium, T1-weighted images show areas of enhancement within the lesion (185a).



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Figure 5-28 Malignant lymphoma. A: Anteroposterior radiograph of the left elbow of a 42-year-old man shows a large osteolytic lesion in the distal humerus (*arrows*). **B:** Anteroposterior radiograph of the right femur of a 7-year-old girl who presented with pain in the upper thigh and a fever reveals a destructive lesion of the diaphysis extending to the growth plate. There is also a subtle lamellated periosteal reaction. **C:** Anteroposterior radiograph of the left shoulder in a 30-year-old man shows a permeative pattern of bone destruction in the proximal humerus accompanied by a lamellated periosteal reaction.

Figure 5-29 Malignant lymphoma. Oblique radiograph of the left distal forearm and wrist in a 63-year-old woman shows almost imperceptible lesion in the distal radius (*arrows*). Note that there is no periosteal reaction present. (Reprinted with permission from Greenfield GB, Arrington JA. *Imaging of bone tumors: a multimodality approach.* Philadel-phia: JB Lippincott, 1995:404.)



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Figure 5-30 Malignant lymphoma. Anteroposterior radiograph of the thoracic spine of a 32-year-old man shows sclerosis of T7 vertebra ("ivory vertebra"). Note bulging of the paraspinal line (*arrows*).



Figure 5-31 Malignant lymphoma: computed tomography (CT). A: Anteroposterior radiograph of the lumbar spine of a 45-year-old man shows a destructive lesion of the L3 vertebra (*arrows*). The extent of the tumor cannot be evaluated. **B:** CT section shows the full extent of the lesion in the bone and a large soft tissue mass.

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Figure 5-32 Malignant lymphoma: scintigraphy and computed tomography (CT). A: Anteroposterior radiograph of the right hip of a 36-year-old man shows a very subtle osteolytic lesion of the acetabulum (*arrow*). **B:** Radionuclide bone scan shows increased uptake of the tracer in the region of the right hip (*arrow*). **C, D:** Two CT sections demonstrate the involvement of the anterior column and roof of the acetabulum.



Figure 5-33 Malignant lymphoma: computed tomography (CT). A: A lateral myelogram in an 18-year-old woman, who presented with low back pain for several months with the clinical impression of herniated intervertebral disk, shows that the disk is normal, but the body of L5 vertebra exhibits a mottled appearance (*arrow*) and its posterior border is indistinct. **B:** CT section demonstrates a large osteolytic lesion extending from the anterior to the posterior margins of the vertebral body. Note small sequestra within the lesion frequently seen in lymphomas.

Histopathology

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Malignant lymphoma of bone appears grossly similar to lymphomas elsewhere in the body (e.g., in lymph nodes). Both lesions are soft and fleshy. Areas of necrosis are occasionally observed (198).

From the histologic standpoint, the appearance of lymphoma is also diverse (191). The lesion consists of aggregates of malignant lymphoid cells that fill the marrow spaces and induce resorption of cancellous bone trabeculae or reactive sclerosis. In primary non-Hodgkin lymphoma, diffuse growth pattern is invariably observed, typically exhibiting a mixture of small lymphocytic cells and a larger histiocytic cell component, whereas secondary lymphomas present with a mixture of paratrabecular, nodular, interstitial, and diffuse infiltration patterns (156). The cells in primary non-Hodgkin lymphoma are usually rounded, ovoid, and pleomorphic (Fig. 5-34A). The cytoplasm is basophilic and exhibits well-defined outlines (166) (Fig. 5-34B). Unlike the case in Ewing sarcoma, the nuclei are rather large, contain relatively scanty chromatin, and are sometimes indented (cleaved) or horseshoe-shaped, with prominent nucleoli (Fig. 5-35). Cytoplasmic glycogen is usually lacking and reticulin fibers are usually present, forming a more or less uniformly distributed net around the tumor cells (138) (Fig. 5-36). A fibroblastic component is often prominent in intraosseous lymphomas and is sometimes

associated with spindling of the neoplastic lymphoid cells (97). Depending on the type of lymphoma, the infiltrate may be monomorphic or polymorphic (Fig. 5-37). However, even in the polymorphic variant, the cells are characterized by malignant rather than inflammatory properties. T-cell lymphomas, in particular, tend to exhibit the widest admixture of cell types, and also tend to be infiltrated by eosinophils (156). The histologic differential diagnosis includes Ewing sarcoma/ PNET, metastatic neuroblastoma, and other small round cell tumors involving bone. Although the patient's age and the overall morphologic features are usually sufficient to categorize the tumor, the most important single procedure to distinguish lymphoma from all other major round cell tumors, as already mentioned, is the immunoreaction for CD45 or CD20 (see Fig. 5-34C) and CD3 (B-cell and T-cell markers) because lymphoid cells are the only cells that show positivity (for additional information see section on differential diagnosis).

When a lymphoma of bone is suspected, all special studies usually done in lymphoma affecting lymph nodes should be performed.

Hodgkin Lymphoma

Hodgkin lymphoma, or Hodgkin disease, is a malignant tumor of the lymph nodes, constituting approximately 30% of all malignant lymphomas. Males are



Figure 5-34 Histopathology of malignant lymphoma. A: In diffuse large B-cell lymphoma observe pleomorphic round cells with well-defined cytoplasmic outline and enlarged nuclei with one to three visible nucleoli (hematoxylin and eosin, original magnification ×400). **B:** With Giemsa staining, the basophilic darkly staining cytoplasm of the tumor cells and nuclear details are better delineated. Cleaved nuclei, nuclear polymorphism, and prominent nucleoli distinguish this tumor from Ewing sarcoma (Giemsa, original magnification ×630). **C:** The tumor cells react with the B-cell marker CD20 (biotin-streptavidin peroxidase, original magnification ×400).







Figure 5-35 Histopathology of malignant lymphoma. A: Densely arranged pleomorphic cells of medium size with roundish nuclei are present, some with a clear cytoplasm. The cells are larger and more pleomorphic than those in Ewing sarcoma (Giemsa, original magnification ×50). **B:** At high magnification the mixed cell population and the cleaved nuclei with prominent nucleoli mark the difference from Ewing sarcoma (hematoxylin and eosin, original magnification ×500). (From Bullough PG. *Atlas of orthopedic pathology*, 2nd ed. New York: Gower, 1992:17.15.)

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A



Figure 5-36 Histopathology of malignant lymphoma. A: With reticulin fiber staining, a fine black-appearing net surrounding individual cells is visible (Novotny, original magnification \times 50). **B:** At high magnification, abundant reticulin fibers form a dense network surrounding individual cells, or small groups of cells (Gomori silver stain, original magnification \times 100).

more commonly affected than females and whites more often than blacks (170). Most cases of Hodgkin lymphoma occur in patients aged 20 to 60 years, without an outstanding peak age (127). Secondary bone involvement is detected radiologically in 5% to 21% of patients (157,160,176). Primary Hodgkin lymphoma of bone is extremely rare (164,173) and usually involves the spine, humerus, and femur (97,195). Thus far, seven cases of multifocal primary osseous Hodgkin lymphoma have been described (172).

Symptoms include pain, often with an onset before the lesion becomes visible by radiography. Occasionally a swelling over the affected bone can be noted (163). In the majority of cases with secondary osseous involvement, the spine and other bones with abundant red bone marrow, such as ribs, pelvic bones, and femur, are affected. The lesion is frequently sclerotic (e.g., ivory vertebra) (Fig. 5-38) or mixed (lytic with sclerotic patches). A purely lytic geographic pattern is uncommon.

It is difficult to make a specific diagnosis of Hodgkin lymphoma of bone, largely because the histologic appearance may not be as classic as in cases involving the lymph nodes (183). In the latter, according to the current classification based on biological and clinical data and proposed by the WHO, two entities can be separated: nodular lymphocyte-predominant Hodgkin lymphoma (comprising 5%) and classical Hodgkin lymphoma (comprising about 95%) with its four subtypesnodular sclerosis, mixed cellularity, lymphocyte-rich, and lymphocyte-depleted (204). These subtypes differ in their sites, clinical features, and morphologic appearance but not in their immunophenotype. The prognostic significance of this histologic classification has lost its value with the advent of modern multimodal treatment modalities. Today, the stage and the presence of systemic symptoms are much more important (205). The tumor cell (Hodgkin cell) is derived from a large mature B cell with a large nucleus that possesses a conspicuous



Figure 5-37 Histopathology of Burkitt lymphoma. A: Small round cells are arranged in clusters separated by delicate reticulin fibers (PAS, original magnification \times 50). **B:** Large macrophages with clear cytoplasm containing cellular debris are intermingled with lymphoma cells, creating a "starry sky" appearance characteristic of this tumor (Giemsa, original magnification \times 400).



Figure 5-38 Hodgkin lymphoma. Anteroposterior radiograph of the lower thoracic spine of a 35-year-old man shows sclerotic T10 vertebra ("ivory vertebra"). (From Bullough PG. *Atlas of orthopedic pathology,* 2nd ed. New York: Gower, 1992:17.16.)

nucleolus. The histologic picture of Hodgkin lymphoma in the bone usually corresponds to the mixed-cell pattern, including lymphocytes, neutrophilic granulocytes, histiocytes, and plasma cells, in addition to the tumor cells (Fig. 5-39). Therefore, the differential diagnosis CHAPTER 5 Round Cell Lesions — 343

usually includes chronic osteomyelitis. It is noteworthy that patients who present with Hodgkin lymphoma in bone almost always have involvement of the lymph nodes, especially the retroperitoneal nodes.

Histologically, Hodgkin lymphoma is distinguished from other lymphomas by the presence of mononuclear Hodgkin cells and multinuclear Reed-Sternberg cells. The latter are distinctive large cells with sharply delineated abundant cytoplasm and a mirror-image double nucleus rich in chromatin, with a large, prominent central eosinophilic nucleolus (161) (Fig. 5-40A, B). These large nucleoli, almost the size of a small lymphocyte, give the cell a conspicuous and almost specific "owl's eyes" appearance. Although pathognomonic Reed-Sternberg cells may be difficult to find, the immunochemical marker CD30 is useful for distinguishing them (205) (Fig. 5-40C).

Differential Diagnosis

Radiology

The radiologic differential diagnosis must include other primary bone tumors. If the patient is an infant or a child (up to 10 years of age), metastatic neuroblastoma should be considered. Between the ages of 5 and 25 years, particularly if the lesion is osteolytic or has a motheaten or permeative pattern of bone destruction with lamellated or onion-skin periosteal reaction, Ewing sar*coma* may be difficult to exclude (127). In more sclerotic lesions, particularly if the "sunburst" pattern of periosteal reaction is present, osteosarcoma is a serious consideration. If a sclerotic lesion extends into the articular end of bone, Paget disease should be considered (Fig. 5-41). In patients older than the age of 40 or 50 years, metastatic carcinoma and plasmacytoma are differential possibilities. At any age, *osteomyelitis* should be a diagnostic consideration. In all of these differential possibilities, because the radiographic appearance of the lesion may be

Figure 5-39 Histopathology of Hodgkin lymphoma. Fibrous stroma with mixed cellular infiltrate of small round cells and large histiocytes is characteristic for this tumor (hematoxylin and eosin, original magnification \times 400). (From Bullough PG. *Atlas of orthopedic pathology*, 2nd ed. New York: Gower, 1992:17.16.)









Figure 5-40 Histopathology of Hodgkin lymphoma. A: Pleomorphic round cells are seen together with scattered Reed-Sternberg cells (hematoxylin and eosin, original magnification ×400). **B:** High-power photomicrograph demonstrates a binucleate Reed-Sternberg cell with prominent eosinophilic nucleoli (hematoxylin and eosin, original magnification ×1,000). **C:** Immunocytochemically, both Hodgkin and Reed-Sternberg cells are positive for CD30 (biotin-streptavidin peroxidase, original magnification ×400). **(A** and **B** from Bullough PG. *Atlas of orthopedic pathology,* 2nd ed. New York: Gower; 1992:17.16.)

В



Figure 5-41 Malignant lymphoma resembling Paget disease. Anteroposterior **(A)** and lateral **(B)** radiographs of the right knee of a 47-year-old woman show a mixed sclerotic and osteolytic lesion extending into the articular end of the tibia, associated with a pathologic fracture, originally diagnosed as Paget disease. Biopsy revealed a histiocytic lymphoma.

very similar, clinical and laboratory information must be taken into consideration.

Pathology

The main considerations are similarly appearing *Ewing* sarcoma, metastatic neuroblastoma, and metastatic carcinoma. Rarely, osteomyelitis with abundant lymphocytic cells is a differential possibility. The use of antibodies against CD45 and other lymphocytic antigens will almost invariably reveal the lymphoid character of lymphoma tumor cells (127,185).

Most lymphomas exhibit a variety of cytologic features within the lesion, making the histologic appearance that of a polymorphous growth. This is helpful in differentiating malignant lymphoma from Ewing sarcoma, which exhibits a growth of isomorphic small cells. The cells in lymphomas usually are less round than those in Ewing sarcoma, and they tend to have larger regular or even cleaved nuclei (see Fig. 5-35).

Another differential diagnostic consideration in a lesion suspected to be lymphoma of bone is a leukemic *infiltrate* of bone. However, all patients with chronic lymphocytic leukemia have peripheral blood abnormalities, and if the bones are affected the diagnosis should not be difficult. The same is true of patients with chronic myeloid (granulocytic) leukemia. Chronic and acute myeloic leukemia may present as a primary bone neoplasm, termed myeloic (granulocytic) sarcoma or chloroma. The clinical and radiologic features of this condition may be similar to those of lymphoma. Biopsy specimens will exhibit sheets of large blast-like cells, although it is very difficult to make a specific diagnosis of granulocytic sarcoma from routine sections of bone. When this diagnosis is suspected, it is necessary to do immunohistochemical studies such as the reaction for lysozyme and myeloperoxidase, which are considered the most helpful study because the blast cells will be positive. The acid chloracetate-esterase reaction is also helpful. Granulocytic sarcoma, however, may be positive for CD99 similar to Ewing sarcoma and some non-Hodgkin lymphomas.

The histologic differential diagnosis of malignant lymphoma also includes benign conditions such as chronic osteomyelitis and other types of small cell malignancies such as metastatic carcinoma and Ewing sarcoma. Chronic osteomyelitis is usually characterized by a mixture of inflammatory cells and no evidence of cytologic atypia. There is also a proliferation of capillaries in the background, as well as fibrosis. It may be difficult to distinguish a large cell lymphoma from metastatic carcinoma or amelanotic melanoma. In these instances, special reactions using CD45, S-100, and cytokeratin antibodies may be helpful in making the distinction. Positivity of the undifferentiated cells for CD45 is strong evidence for a malignant lymphoma, whereas positivity for anti-keratin and S-100 protein suggests respectively an epithelial origin of the tumor or melanoma.

Sometimes it is difficult to distinguish a lymphoma from a true sarcoma of bone. Lymphomas of bone may possess spindle cells that occasionally have a storiform pattern, suggesting the diagnosis of malignant fibrous histiocytoma. *Malignant fibrous histiocytomas* usually create large areas of bone destruction. If the lesion appears to be only permeative, i.e., it fills the marrow spaces and leaves behind normal bony trabeculae, the diagnosis of malignant lymphoma with spindling can be suspected. The immunoperoxidase reaction may be helpful in this instance because the spindle cells in malignant lymphoma react positively with CD45, whereas the spindle cells of malignant fibrous histiocytoma are negative. However, it should be noted that acidic decalcification of bone biopsy specimens may interfere with IHC, because certain antigenic determinants are denatured by brisk acid decalcification.

Occasionally, the eosinophilic component and the histiocytes may be such prominent features of Hodgkin lymphoma as to suggest the diagnosis of *LCH* (38). In this situation, the age of the patient is crucial: in a middle-aged or elderly patient, the diagnosis of Hodgkin lymphoma is much more likely and should be strongly considered (182). Additionally, cells in LHC are positive for CD1a and S-100 protein, and negative for CD30, whereas Hodgkin lymphoma exhibits the reverse pattern.

The radiologic and pathologic differential diagnosis of malignant lymphoma is depicted in Figure 5-42.

Multiple Myeloma (Plasmacytoma)

Myeloma, a tumor originating in the bone marrow (also known as multiple myeloma, plasma cell myeloma, or plasmacytoma), is the most common primary malignant tumor of bone and accounts for 27% of biopsied bone tumors (208). Most of these tumors are diagnosed by bone marrow biopsy. Myeloma is often associated with the presence of abnormal proteins in the blood and urine, and occasionally with the presence of amyloid in the tumor tissue or other organs (138,249).

Myeloma is frequently preceded by a precursor lesion, so-called monoclonal gammopathy of unknown significance (MGUS), that can be detected in about 1% of all adults older than 50 years. Each year, about 1% of MGUS cases will progress to myeloma. Myeloma cells do not initially reveal a high proliferation rate. DNA synthesis is observed in less than 1% of tumor cells (148). Solitary myeloma (or plasmacytoma) is a clinical variant (227) considered to represent an early stage of multiple myeloma (256), with stages of transition existing between the localized and the disseminated types (218,235,257).

During the course of the disease, myeloma cells acquire an increasing number of structural and numerical chromosomal aberrations that are believed to be connected to pathogenesis (238). CGH studies revealed gains of 1q, 3q, 9q, 11q, and 15q. Losses of chromosome 13 or at 13q14 are observed in about 60% of cases, leading to the hypothesis that a special tumor suppressor gene, which appears to be different from the *RB* (retinoblastoma) gene (248), might be involved in myeloma pathogenesis. Deletions of *TP53* at 17p13 are reported in 25% of cases and are believed to be related to poor outcomes (238).





In MGUS and early-stage multiple myeloma, chromosomal translocations occur involving the *VDJ* genes (*V*ariable gene segment, *D*iversity gene segment, *J*oining gene segment; these genes encode the variable region of the immunoglobulin heavy chain, which is responsible for immunoglobulin diversity) and oncogenes, putting the latter in close proximity to an immunoglobulin transcriptional activator. This may lead to dysregulation or overexpression of oncogenes. Additional translocations involving *MYC* and *RAS* have also been observed (221).

Myeloma in bone is characterized by the presence of local alterations in the bone marrow microenvironment. Myeloma cells directly and indirectly increase osteoclast formation and activation (via stimulation of bone marrow stromal cells and bone marrow endothelial cells) by producing or mediating the release of potent osteoclast activators such as RANKL (receptor activator of nuclear factor kappa B, which is also involved in osteoclastogenesis). Furthermore, myeloma cells decrease osteoblastic production of osteoprotegerin (OPG), which acts as a neutralizing soluble receptor for RANKL, thereby inhibiting osteoclast activity and consequent bone loss. In addition, myeloma cells bind OPG via syndecan (CD 138, a transmembrane glycoprotein that acts as a receptor for type I collagen, which is used as a marker for myeloma cells in IHC), and internalize and degrade it. They also interfere with the function and differentiation

of osteoblasts and induce apoptosis of osteoblasts (229,241).

Clinical Presentation

Myeloma usually occurs in patients older than 40 years and is most commonly seen between the fifth and seventh decades, with a median patient age at diagnosis of 64 years. Occurrence in the younger age group is exceptional (222,224,242,243,252) and is usually represented by cases of solitary myeloma (or plasmacytoma) (246). The incidence is 6.6 per 100,000 for males and 5.9 per 100,000 for females in western Europe (234). According to the Surveillance, Epidemiology and End Results (SEER) program, the incidence rates in the United States are 6.9 per 100,000 and 4.5 per 100,000, respectively, for males and females. In African Americans, the prevalence and incidence of MGUS and myeloma are about twofold higher than in white subjects (228,268). A clinical staging system based on the serum concentrations of immunoglobulin, calcium, and paraprotein; excretion of urinary Bence-Jones protein; and the number of skeletal lesions seen on radiography has been proposed by Durie and Salmon (231). However, this system has recently been re-evaluated, leading to a prognostic index that is based on the serum levels of beta-2microglobulin and albumin (237,240). Myeloma arises primarily in bones that contain red marrow and therefore the axial skeleton is most commonly affected. However, no bone is exempt from involvement (Fig. 5-43).



Figure 5-43 Myeloma: skeletal sites of predilection, peak age range, and male-to-female ratio.

Skeletal involvement distal to the elbow and knee occurs in only 10% of cases (276). Pain, mild and transient, worse during the day, and enhanced by weight bearing and activity, may be the initial complaint and is present in about 75% of patients (230). Therefore, multiple myeloma may masquerade as sciatica or intercostal neuralgia in the stages before the diagnosis is established. Malaise, fatigue, weight loss, fever, and bone pain are other commonly encountered symptoms. Low-back pain or sciatic pain may sometimes be accompanied by neurologic symptoms (213). Rarely, a pathologic fracture is the first sign of disease. Other common clinical features of multiple myeloma are related to a refractory, normocytic, normochromic anemia, resulting from bone marrow replacement by myeloma cells, to hypercalcemia resulting from widespread bone destruction, to renal dysfunction caused by renal tubule obstruction and damage by abnormal circulating proteins, and to decreased resistance to infection resulting from a deficiency of normal antibody production. Because the malignant cells represent a monoclonal proliferation of plasma cells, the tumor produces the specific immunoglobulin type associated with that particular neoplastic plasma cell. Characteristically, the ratio of serum albumin to globulins is reversed because of the high concentration of monoclonal immunoglobulin produced by the myeloma cells [most often IgG (65%), or IgA (20%), less frequently IgD (2%), and rarely IgM or IgE]. Because the monoclonal immunoglobulin is of the same molecular weight and charge, during serum electrophoresis it moves to a single spot, and a concentrated narrow band, the so-called M component, appears as a peak when densitometry is performed on the electrophoretic strip. Immunoelectrophoresis of the serum predominantly shows a monoclonal heavy chain (83%)and a monoclonal light chain (8%) (250). Although the total immunoglobulin level is elevated because of the monoclonal protein, the normal serum immunoglobulin concentrations are markedly depressed, resulting in impaired humoral immunity. If light chains are produced in excess, these pass into the renal tubules and are dimerized to form Bence-Jones protein in the urine (220). MGUS is characterized by an M-protein level in serum below 30 g/L, less than 10% clonal plasma cells in the bone marrow, no evidence of other B-cell disorders, and no organ impairment, especially no detectable bone lesions (245).

Although myeloma, being a neoplasm of mature B cells, is primarily originating in bone, many other organs (including lymph nodes) may be affected (244).

Complications encountered with myeloma include pathologic fractures (of long bones, ribs, sternum, and vertebrae), amyloidosis (in about 15% of patients), increased incidence of infection, anemia, and a hyperviscosity syndrome. Most patients with solitary myeloma develop the systemic form of myeloma within 5 to 20 years (90).

Imaging

On radiography, multiple myeloma may present in a variety of patterns (Fig. 5-44). Most commonly it pre-



Figure 5-44 Myeloma: variants in the radiographic presentation.

sents either as a lesion with a geographic type of bone destruction or as multiple small foci of bone destruction (Fig. 5-45). In the skull, characteristic roundish, punched-out areas of bone destruction, usually uniform in size, are observed (Fig. 5-46).

In the spine, multiple myeloma may present simply as osteoporosis, with no definitely identifiable lesions. A more common presentation, however, is the presence of multiple lytic lesions scattered throughout the vertebral column. Multiple compression fractures of the vertebral bodies may be seen in both forms.

In the ribs, lace-like areas of bone destruction and small osteolytic (punched-out) lesions accompanied by soft tissue masses are occasionally observed (260). In the flat and long bones, areas of medullary bone destruction are noted and, if these abut the cortex, endosteal scalloping of the inner cortical margin (socalled rat bites) is seen (see Fig. 5-45B). Characteristically, there is no evidence of sclerosis and no periosteal reaction. Osteosclerotic lesions, even when mixed with lytic lesions, are rare in myeloma, and less than 1% of myelomas exhibit pure sclerotic lesions (sclerosing myelomatosis) (226,275).

Although scintigraphy in multiple myeloma is usually normal (217), increased uptake is occasionally demonstrated, presumably secondary to hyperemia, to increased turnover in adjacent bone, and perhaps even to some osteoblastic activity (261). On some occasions, areas of decreased uptake (cold foci) are detected, presumably caused by a replacement of normal bone by myeloma tissue, which does not take up the tracer (277).

CT is better than radiography for revealing the intraosseous extent of the tumor, cortical destruction, and extraosseous involvement (Fig. 5-47). Some vertebral


Figure 5-45 Multiple myeloma. A: Lateral radiograph of the distal femur of a 65-year-old woman shows multiple osteolytic lesions. **B:** Anteroposterior radiograph of the left elbow in the same patient shows multiple osteolytic lesions and endosteal scalloping of the cortex typical of diffuse myelomatosis.



Figure 5-46 Multiple myeloma. Involvement of the skull is a prominent feature of multiple myeloma of this 60-year-old woman. Note characteristic "punched-out," lytic lesions, most of which are uniform in size and lack sclerotic borders. Occasionally, this pattern may be seen in metastatic disease.

lesions that are not visible on radiographs can also be demonstrated (254,271).

MRI is more sensitive than radiography and CT for detection of skeletal abnormalities in multiple myeloma (219,236,259,273) (Fig. 5-48). In particular, MRI is effective in demonstrating spinal lesions (253,272,274). The MRI changes may be either diffuse or focal. On T1weighted images the lesions are visible as hypointense areas (usually of intermediate signal intensity), as contrasted with surrounding bone (Fig. 5-48B). On T2weighted and STIR images, the lesions display homogeneously high signal intensity (214) (Fig. 5-48C). The lesions show enhancement on turbo-fast low-angle shot (FLASH) images (263). On contrast-enhanced T1weighted SE images, persistence of paramagnetic contrast agent is seen (263) (Fig. 5-48D). Because the fatty tissue replacement that occurs within the vertebrae in older patients can mask the lesions, fat-suppressed MR images are of value for detecting myeloma (233,267).

Recently, Major et al. (255) reported an interesting MRI appearance of plasmacytoma affecting the vertebral body. In all 10 patients they examined, an expansive lesion exhibited low signal intensity on T1-weighted images, whereas on axial T2-weighted sections high signal was present involving the entire vertebral body. Within these bright areas were curvilinear structures with low signal intensity, resembling sulci of the brain.



Figure 5-48 Multiple myeloma: magnetic resonance imaging (MRI). A: Anteroposterior radiograph of the right proximal femur of a 53-year-old man shows osteolytic lesion in the subtrochanteric region. B: Coronal T1-weighted MRI shows the tumor to be of intermediate signal, isointense with the skeletal muscles.



Figure 5-48 *Continued* **C:** Coronal T2-weighted MRI shows the tumor to be of homogeneous high signal intensity. **D:** After intravenous administration of gadolinium there is a slight enhancement of myeloma.

The authors termed this phenomenon the "mini brain" appearance, suggesting that it may be a pathognomonic feature of plasmacytoma in this location.

It should be emphasized that the variable appearance of multiple myeloma on MRI is caused by a combination of factors. Although myeloma may involve the marrow diffusely, the involvement is not necessarily homogeneous. The appearance of the bone marrow depends both on the extent of the disease and on the degree of fatty replacement, which varies considerably, and is greater in the older age group (253).

At present, whole body ¹⁸F-labeled 2-fluoro-2-deoxyglucose (¹⁸F-FDG) positron emission tomography (PET) is included in the workup of myeloma and MGUS because bone marrow alterations can be detected by this method before they are visible on MRI (232).

An interesting variant of sclerosing myeloma is the so-called POEMS syndrome, first described in 1968 (270) but gaining wide acceptance only more recently (225,239,247,265,269). It consists of polyneuropathy (P); organomegaly (O), particularly of the liver and the spleen; endocrine disturbances (E), including amenorrhea, gynecomastia, impotence, diabetes mellitus, and hypothyroidism; monoclonal gammopathy (M), usually due to lambda light chains; and skin changes (S), such as hyperpigmentation and hirsutism. Additional associated abnormalities include papilledema, peripheral edema, ascites, and clubbing of the fingers (212). The exact pathogenesis of POEMS syndrome remains unsettled. It is still debatable whether the sclerotic changes are related to plasma cells that fail to secrete osteoclastactivating factor or perhaps are related to the secretion of an osteoblast-stimulating factor (216). Relatively few plasma cells are present in the marrow of patients with POEMS syndrome, and some investigators consider this entity a plasma cell dyscrasia rather than a true neoplasm (265). These patients do not have an increased incidence of amyloidosis and rarely have Bence-Jones proteinuria (225).

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Certain radiologic findings concerning the lesions in POEMS syndrome have been emphasized by Resnick et al. (265,266). These include irregular, spiculated, osseous proliferation involving multiple tendinous and ligamentous attachments (enthesopathy), especially of the posterior elements of the thoracic and lumbar spine, and lucent areas of bone destruction with a surrounding

rim of sclerosis. Some authors describe sclerotic foci within the vertebral body ranging from very dense bone, resembling ivory vertebrae, to fine sclerotic lesions with irregular peripheral spiculations. Some of these sclerotic foci may even resemble bone islands. Positive bone scans in patients with POEMS syndrome appear to be exceptional. When positive, the accumulation of the radionuclide in the osseous foci is probably due to a sclerotic bone reaction and to the cortical expansion combined with a periosteal reaction that occurs in some cases (223,239).

Probably the rarest presentation of multiple myeloma is that of an extraskeletal lesion in form of a soft tissue mass (258).

Histopathology

The gross appearance of myeloma is that of either a fleshy white lesion, very similar to the appearance of lymphoma, or a grayish-pink fleshy mass. However, the gross appearance alone is not diagnostic of myeloma.

The histologic appearance of myeloma is characteristic. The diagnosis is based on the presence of extended sheets of a population of atypical plasmacytoid cells that fill the trabecular bone marrow spaces and replace the normal fatty and hematopoietic marrow. The cytologic findings may vary. A high degree of differentiation is characterized by almost normalappearing plasma cells (Fig. 5-49A). Their round nuclei





Figure 5-49 Histopathology of multiple myeloma. A: In a higher degree of differentiation (grade 2), tumor cells exhibit plasma cell-like features with eccentric nuclei and enlarged eosinophilic cytoplasm with juxtanuclear halo. Few cells exhibit a "spoke-wheel" chromatin pattern; most tumor cells contain clearer and larger, slightly pleomorphic nuclei with eosinophilic nucleoli (hematoxylin and eosin, original magnification ×400). **B:** At high power basophilic cytoplasm and perinuclear halos are best demonstrated in Giemsa-stained preparations. Observe nuclear pleomorphism, peripherally condensed chromatin, and prominent nucleoli (Giemsa, original magnification ×630). **C:** By immunohistochemistry (IHC), tumor cells show a monoclonal expression of lambda light chains (biotin-streptavidin peroxidase, original magnification ×400). **D:** The collagen type I binding protein syndecan-1 (CD138) is typically present on myeloma cells (biotin-streptavidin peroxidase, original magnification ×400).



Figure 5-50 Histopathology of multiple myeloma. In grade 3 tumor, between the normal-appearing plasma cells (fewer than the number illustrated in Fig. 5-49A) are pleomorphic cells with clearer nuclei. Numerous giant cells with two and more nuclei are also present (Giemsa, original magnification \times 100).

possess a dense, coarse chromatin that has a typical cartwheel-like distribution. The prominent nucleolus is excentrically located in a strongly basophilic cytoplasm (Fig. 5-49B) that corresponds to the densely arranged rough endoplasmic reticulum as revealed by electron microscopy and displays a perinuclear lighter halo that represents the well-developed Golgi apparatus. In addition, a wide variety of cytoplasmic inclusion bodies of crystallite form may be found in the cells. Increasing dedifferentiation is accompanied by an increasing pleomorphism of the cells and the nuclei, with loss of the regimented arrangement of the cell organelles. Furthermore, there are increasing numbers of bi- and trinucleated cells (Fig. 5-50). Dedifferentiation is usually classified into three grades that are correlated with decreasing survival time for the patient (138,244). It may be difficult to differentiate a grade 3 myeloma from a lymphoma or from an undifferentiated carcinoma. Myeloma cells may occasionally be arranged in groups or nests surrounded by thin fibrovascular septa, imitating a neuroendocrine tumor. Moreover, pseudovascular spaces and stromal proliferations of blood vessels may be present, also resembling a vascular tumor. Myelomas consisting of signet-ring cells, clear cells, oncocytic cells, or spindle cells have been described but are exceedingly rare (215).

Although conventional myeloma is not associated with osteosclerosis, rare cases of myeloma are associated with this feature, known as sclerosing myeloma. These patients classically present with peripheral neuropathy. They also may have organomegaly, endocrine disturbances, monoclonal gammopathy, and skin lesions (POEMS syndrome; see previous discussion). They tend to be younger than patients with classical myeloma, and the disease progression is more indolent. The myeloma associated with peripheral neuropathy may be either localized or a part of multiple myeloma. The peripheral neuropathy may be arrested and may even improve with treatment of the underlying myeloma. Of all myelomas, 10% to 15% will exhibit the presence of amyloid within the tumor, in the form of eosinophilic hyaline masses or amyloid deposition within vascular walls. The amyloid may invoke a foreign body type of giant cell reaction. The presence of abundant amyloid may also obscure the underlying neoplasm (262). When amyloid is found in a bone biopsy specimen the pathologist should suspect a diagnosis of myeloma unless the patient has been receiving chronic hemodialysis because such patients may also deposit so-called amyloid pseudotumors in the skeleton.

IHC almost always reveals a monotypic cytoplasmic expression of kappa or lambda light chains. CD138 and/or syndecan 1 is also positive (see Fig. 5-49C,D). CD45 and CD20 are usually negative, discriminating plasmacytoma from plasmacytic lymphoma (238).

Differential Diagnosis

Radiology

When many osteolytic lesions in several bones are observed, the only alternative diagnosis is *metastatic* disease. This is particularly true if the patient is older than 40 or 50 years. In younger patients, brown tumors of hyperparathyroidism (Fig. 5-51) and Gaucher disease (Fig. 5-52) should be considered. Severe osteoporosis may sometimes mimic the diffuse type of multiple myeloma. A valuable differential clue is the lack of endosteal scalloping in this condition, unlike multiple myeloma, in which scalloping is invariably present (see Fig. 5-45B). When the spine is involved, which occurs rather commonly, multiple myeloma must be differentiated from metastatic carcinoma. In this respect, the vertebral-pedicle sign identified by Murray and Jacobson (260) may be helpful. These investigators contended that in the early stages of myeloma the pedicle, which does not contain as much red marrow as the vertebral body, is not initially involved, whereas in metastatic carcinoma the pedicle is affected to the same extent as the vertebral body, even in the early stages (Fig 5-53). In the late stages of multiple myeloma, however, the pedicle and the vertebral body are both affected. A more reliable distinction between these two processes is achieved with radionuclide bone scanning. Although the bone scan is invariably abnormal in cases of metastatic carcinoma, myeloma seldom produces an increased uptake of the radiopharmaceutical agent.

A solitary myeloma may create even more diagnostic problems. Because it is a purely radiolucent lesion, it may mimic other purely destructive processes such as *giant cell tumor, brown tumor of hyperparathyroidism, fibrosarcoma, malignant fibrous histiocytoma,* and a *solitary metastatic focus* from a carcinoma of the kidney, lung, thyroid, or the gastrointestinal tract. Moreover, only a small percentage of solitary myelomas are associated with production of detectable monoclonal immunoglobulins.

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Figure 5-51 Hyperparathyroidism. Anteroposterior radiograph of the lower legs of a 36-year-old woman with primary hyperparathyroidism shows multiple lytic lesions ("brown tumors") in both tibiae, resembling multiple myeloma.



Figure 5-52 Gaucher disease. Destructive changes seen here in the right humerus of a 52-year-old woman with the adult form of the disease may assume a honeycomb appearance simulating multiple myeloma.

Sclerosing myeloma with diffuse bone abnormalities should be differentiated from *sclerotic metastasis, mastocytosis, myelofibrosis,* and *renal osteodystrophy.* Focal sclerotic changes observed in myeloma should be differentiated from *tuberous sclerosis* and some *sclerosing dysplasias* (e.g., van Buchem disease; osteopoikilosis). Some myelomas with calcified amyloid depositions may be mistaken for *chondrosarcoma* (264). It must be pointed out that amyloidosis occurs in approximately 10% to 15% of cases of myeloma (251) and that calcifications are associated with both primary and secondary amyloidosis (262).

Pathology

The important differential diagnosis of myeloma includes a tumor and reactive forms of plasma cell proliferation, such as *plasmacytoid lymphoma*, *monoclonal gammopathy* of unknown significance (but only up to 10% monoclonal plasmocytes in the bone marrow), *plasma cell-rich osteomyelitis*, and *rheumatoid arthritis* (up to 10% polyclonal plasmocytes in bone marrow). The immunohistochemical demonstration of the monoclonal immunoglobulin produced by the myeloma cells allows a clear-cut distinction between the tumor and the reactive form. The same is true for the differentiation of chronic plasma cell–rich osteomyelitis. In addition, the presence of other types of inflammatory cells and the formation of granulation tissue with proliferating capillaries in the latter condition provide aids to diagnosis. As mentioned previously, distinguishing cases of undifferentiated myeloma or undifferentiated metastatic carcinoma from a lymphoma may be difficult, but the demonstration of the specific immunoglobulin in the former and of keratin markers in the latter will aid in making the diagnosis.

The radiologic and pathologic differential diagnosis of solitary plasmacytoma is depicted in Figure 5-54, and the differential diagnosis of multiple myeloma is shown in Figure 5-55.





Α

Figure 5-53 Multiple myeloma versus metastases. Anteroposterior **(A)** and lateral **(B)** radiographs of the spine of a 70-year-old man with multiple myeloma involving both the spine and appendicular skeleton show a compression fracture of the body of T8 vertebra (*arrows*); several other vertebrae show only osteoporosis. The pedicles are preserved in contrast to metastatic disease of the spine (which usually also affects the pedicles), as seen on this anteroposterior radiograph of the cervical spine **(C)** in a 65year-old man with colon carcinoma and multiple lytic metastases. Note the involvement of the right pedicle of C7 vertebra (*arrows*).



С





Figure 5-54 Radiologic and pathologic differential diagnosis of solitary plasmacytoma.



Figure 5-55 Radiologic and pathologic differential diagnosis of multiple myeloma.

REFERENCES

Langerhans Cell Histiocytosis (Eosinophilic Granuloma)

- Alexander JE, Seibert JJ, Berry DH, et al. Prognostic factors for healing of bone lesions in histiocytosis X. *Pediatr Radiol* 1988;18: 326–332.
- Antonmattei S, Tetalman MR, Lloyd TV. The multiscan appearance of eosinophilic granuloma. *Clin Nucl Med* 1979;4:53–55.
- Arai F, Miyamoto T, Ohneda O, et al. Commitment and differentiation of osteoclast precursor cells by the sequential expression of c-Fms and receptor activator of nuclear factor kappaB (RANK) receptors. *J Exp Med* 1999;190:1741–1754.
- Aricò M, Danesino C. Langerhans' cell histiocytosis: is there a role for genetics? *Haematologica* 2001;86:1009–1014.
- Beltran J, Aparisi F, Marti Bonmati L, et al. Eosinophilic granuloma: MRI manifestations. *Skeletal Radiol* 1993;22:157–161.
- Birbeck MS, Breathnach AS, Everall JD. An electron microscopic study of basal melanocytes and high-level clear cell (Langerhans cell) in vitiligo. *J Invest Dermatol* 1961;37:51–64.
- Bohne WHO, Goldman AB, Bullough P. Case report 96. Chester-Erdheim disease (lipogranulomatosis). *Skeletal Radiol* 1979;4:164–167.
- Chikwava K, Jaffe R. Langerin (CD207) staining in normal pediatric tissues, reactive lymph nodes, and childhood histiocytic disorders. *Pediatr Dev Pathol* 2004;7:607–614.
- 9. Cline MJ. Histiocytes and histiocytosis. Blood 1994;84:2840-2853.
- Compere EL, Johnson WE, Coventry MB. Vertebra plana (Calvé's disease) due to eosinophilic granuloma. J Bone Joint Surg 1954;36:969–980.
- Conway WF, Hayes CW. Miscellaneous lesions of bone. *Radiol Clin North Am* 1993;31:339–358.
- Corrin B, Butcher D, McAnulty BJ, et al. Immunohistochemical localization of transforming growth factor-beta 1 in the lungs of patients with systemic sclerosis, cryptogenic fibrosing alveolitis and other lung disorders. *Histopathology* 1994;24:145–150.
- da Costa CE, Annels NE, Faaij CM, et al. Presence of osteoclastlike multinucleated giant cells in the bone and nonostotic lesions of Langerhans cell histiocytosis. *J Exp Med* 2005;201: 687–693.
- Dalinka MK, Turner ML, Thompson JJ, Lee RE. Lipid granulomatosis of the ribs: focal Erdheim-Chester disease. *Radiology* 1982;142:297–299.
- Dallari D, Marinelli A, Pellacani A, et al. Xanthoma of bone: first sign of hyperlipidemia type IIB: a case report. *Clin Orthop Relat Res* 2003;410:274–277.
- David R, Oria RA, Kumar R, et al. Radiologic features of eosinophilic granuloma of bone. Am J Roentgenol 1989;153: 1021–1026.
- de Graaf JH, Tamminga RY, Dam-Meiring A, et al. The presence of cytokines in Langerhans' cell histiocytosis. *J Pathol* 1996;180: 400–406.
- Dehner LP. Juvenile xanthogranulomas in the first two decades of life: a clinicopathologic study of 174 cases with cutaneous and extracutaneous manifestations. *Am J Surg Pathol* 2003;27:579–593.
- De Schepper AM, Ramon F, Van Marck E. MR imaging of eosinophilic granuloma: report of 11 cases. *Skeletal Radiol* 1993; 22:163–166.
- De Young B, Unni K. Langerhans cell histiocytosis. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:345–346.
- Doyle AJ, Christie M, French G. Bone surface lesions [letter]. Radiology 1998;209:282–283.
- Egeler RM, Annels NE, Hogendoorn PC. Langerhans cell histiocytosis: a pathologic combination of oncogenesis and immune dysregulation. *Pediatr Blood Cancer* 2004;42:401–403.
- Favara BE. Langerhans cell histiocytosis—pathobiology and pathogenesis. Semin Oncol 1991;18:3–7.
- Favara BE, McCarthy RC, Mierau GW. Histiocytosis X. Hum Pathol 1983;14:663–676.
- Fisher AJ, Reinus WR, Friedland JA, Wilson AJ. Quantitative analysis of the plain radiographic appearance of eosinophilic granuloma. *Invest Radiol* 1995;30:466–473.

- Flores LG II, Hiroaki H, Nagamachi S, et al. Thallium-201 uptake in eosinophilic granuloma of the frontal bone: comparison with technetium-99m-MDP imaging. J Nucl Med 1995;36: 107–110.
- 27. Fowles JV, Bobechko WP. Solitary eosinophilic granuloma in bone. J Bone Joint Surg 1970;52B:238–243.
- Geissmann F, Lepelletier Y, Fraitag S, et al. Differentiation of Langerhans cells in Langerhans cell histiocytosis. *Blood* 2001;97: 1241–1248.
- Greenfield GB, Arrington JA. Imaging of bone tumors. A multimodality approach. Philadelphia: JB Lippincott, 1995.
- Grundy P, Ellis R. Histiocytosis X: a review of the etiology, pathology, staging, and therapy. *Med Pediatr Oncol* 1986;14: 45–50.
- Haggstrom JA, Brown JC, Marsh PW. Eosinophilic granuloma of the spine: MR demonstration. J Comput Assist Tomogr 1988;12: 344–345.
- Hajdu I, Zhang W, Gordon GB. Peanut agglutinin binding as a histochemical tool for diagnosis of eosinophilic granuloma. *Arch Pathol Lab Med* 1986;110:719–721.
- 33. Helms CA, Jeffrey RB, Wing VW. Computed tomography and plain film appearance of a bony sequestration: significance and differential diagnosis. *Skeletal Radiol* 1987;16:117–120.
- Hermann G, Abdelwahab IF, Sacher M, Klein MJ. The many faces of Langerhans cell histiocytosis. Osteologiai Közlemények 1998;6:202–205.
- Hindman BW, Thomas RD, Young LW, Yu L. Langerhans cell histiocytosis: unusual skeletal manifestations observed in thirtyfour cases. *Skeletal Radiol* 1998;27:177–181.
- Hudson TM. Radiologic-pathologic correlation of musculoskeletal lesions. Baltimore: Williams & Wilkins, 1987.
- 37. Hurwitz N, Birnbaum Y, Pellet D, et al. Hodgkin's disease with primary presentation in the bone marrow. *Leuk Lymphoma* 1992; 6:267–269.
- Huvos AG. Bone tumors: diagnosis, treatment, and prognosis, 2nd ed. Philadelphia: WB Saunders, 1991.
- Jaffe ES. Histiocytic and dendritic cell neoplasms: Introduction. In: Jaffe ES, Harris NL, Stein H, Vardiman JW, eds. World Health Organization classification of tumours. Pathology and genetics of tumours of haematopoietic and lymphoid tissues. Lyon: IARC Press, 2001:275–277.
- Kaplan GP, GR, Saifuddin A, Pringle JAS, et al. Langerhans' cell histiocytosis of the spine: use of MRI in guiding biopsy. *Skeletal Radiol* 1998;27:673–676.
- Kilpatrick SE, Wenger DE, Gilchrist GS, et al. Langerhans cell histiocytosis (histiocytosis X) of bone: a clinicopathologic analysis of 263 pediatric and adult cases. *Cancer* 1995;76:2471–2484.
- Krishnan A, Shirkhoda A, Tehranzadeh J, et al. Primary bone lymphoma: radiographic-MR imaging correlation. *RadioGraphics* 2003;23:1371–1387.
- Kumar R, Balachandran S. Relative roles of radionuclide scanning and radiographic imaging in eosinophilic granuloma. *Clin Nucl Med* 1980;5:538–542.
- Laman JD, Leenen PJ, Annels NE, et al. Langerhans-cell histiocytosis 'insight into DC biology'. *Trends Immunol* 2003;24:190–196.
- Leonidas JC. Langerhans cell histiocytosis. In: Taveras JM, Ferrucci JM, eds. *Radiology: diagnosis, imaging, intervention*, vol 5. Philadelphia: JB Lippincott, 1990:1–9.
- Lichtenstein L. Histiocytosis X. I. Integration of eosinophilic granuloma of bone, Letterer-Siwe disease, and Schüller-Christian disease as related manifestations of single nosologic entity. *Arch Pathol* 1953;56:84–102.
- Lichtenstein L. Histiocytosis X (eosinophilic granuloma of bone, Letterer-Siwe disease, and Schüller-Christian disease). *J Bone Joint Surg Am* 1964;46:76–90.
- Lieberman PH, Jones CR, Steinman RM, et al. Langerhans cell (eosinophilic) granulomatosis. A clinicopathologic study encompassing 50 years. *Am J Surg Pathol* 1996;20:519–552.
- Makley JT, Carter JR. Eosinophilic granuloma of bone. Clin Orthop 1986;204:37–44.
- Meyer JS, Harty MP, Mahboubi S, et al. Langerhans cells histiocytosis: presentation and evolution of radiologic findings with clinical correlation. *RadioGraphics* 1995;15:1135–1146.
- 51. Mickelson MR, Bonfiglio M. Eosinophilic granuloma and its variations. *Orthop Clin North Am* 1977;8:933–945.

- Mirra JM, Gold RH. Eosinophilic granuloma. In: Mirra JM, Gold RH, Picci P, eds. Bone tumors: clinical, radiologic, and pathologic correlations. Philadelphia: Lea & Febiger, 1989:1021–1039.
- Mitnick JS, Pinto RS. Computed tomography in the diagnosis of eosinophilic granuloma. J Comput Tomogr 1980;4:791–793.
- Moore JB, Kulkarni R, Crutcher DC, Bhimani S. MRI in multifocal eosinophilic granuloma: staging disease and monitoring response to therapy. *Am J Pediatr Hematol Oncol* 1989;11:174–177.
- Moseley JE. Bone changes in hematologic disorders (Roentgen aspects). New York: Grune and Stratton, 1963:161–179.
- Moseley JE. Patterns of bone change in the reticuloendothelioses. J Mt Sinai Hosp 1962;29:282–321.
- Murakami I, Gogusev J, Fournet JC, et al. Detection of molecular cytogenetic aberrations in Langerhans cell histiocytosis of bone. *Hum Pathol* 2002;33:555–560.
- Murayama S, Numaguchi Y, Robinson AE, Richardson DE. Magnetic resonance imaging of calvarial eosinophilic granuloma. *J Comput Tomogr* 1988;12:251–252.
- 59. Nezelof C, Basset F. An hypothesis Langerhans cell histiocytosis: the failure of the immune system to switch from an innate to an adaptive mode. *Pediatr Blood Cancer* 2004;42:398–400.
- Nezelof C, Basset F, Rousseau MF. Histiocytosis X: histogenetic arguments for a Langerhans cell origin. *Biomedicine* 1973;18: 365–371.
- Ostand ME. Histiocytosis X: Langerhans cell histiocytosis. Hematol Oncol Clin North Am 1987;1:737–751.
- 62. Pileri SA, Grogan TM, Harris NL, et al. Tumours of histiocytes and accessory dendritic cells: an immunohistochemical approach to classification from the International Lymphoma Study Group based on 61 cases. *Histopathology* 2002;41:1–29.
- Resnick D. Lipidoses, histiocytoses, and hyperlipoproteinemias. In: Resnick D, ed. *Bone and joint imaging*. Philadelphia: WB Saunders, 1989:687–702.
- Robbin MR, Murphey MD, Temple T, et al. Imaging of musculoskeletal fibromatosis. *RadioGraphics* 2001;21:585–600.
- Sartoris DJ, Parker BR. Histiocytosis X: rate and pattern of resolution of osseous lesions. *Radiology* 1984;152:679–684.
- 66. Servet-Delprat C, Arnaud S, Jurdic P, et al. Flt3+ macrophage precursors commit sequentially to osteoclasts, dendritic cells and microglia. *BMC Immunol* 2002;3:15.
- Siegelman SS. Taking the X out of histiocytosis X. Radiology 1997;204:322–324.
- Schajowicz F. Tumors and tumorlike lesions of bone: pathology, radiology, and treatment, 2nd ed. Berlin: Springer-Verlag, 1994: 552–566.
- Schaub T, Ash JM, Gilday DL. Radionuclide imaging in histiocytosis X. *Pediatr Radiol* 1987;17:397–404.
- Stull MA, Kransdorf MJ, Devaney KO. Langerhans cell histiocytosis of bone. *RadioGraphics* 1992;12:801–823.
- Valladeau J, Dezutter-Dambuyant C, Saeland S. Langerin/ CD207 sheds light on formation of Birbeck granules and their possible function in Langerhans cells. *Immunol Res* 2003;28: 93–107.
- Valadeau J, Ravel O, Dezutter-Dambuyant C, et al. Langerin, a novel C-type lectin specific to Langerhans cells, is an endocytic receptor that induces the formation of Birbeck granules. *Immunity* 2000;12:71–81.
- Uhlinger E. Das eosinophile Knochengranulom. In: Heilmeyer L, Hittmair A, eds. *Handbuch der Gesamten Haematologie*. Munich: Urban and Schwartzenberg Verlag, 1963:Band IV/2; 56.
- 74. Weiss L, Grogan T, Müller-Hermelink H-K, et al. Langerhans cell histiocytosis. In: Jaffe ES, Harris NL, Stein H, Vardiman JW, eds. World Health Organization classification of tumours. Pathology and genetics of tumours of haematopoietic and lymphoid tissues. Lyon: IARC Press, 2001:280–282.
- 75. Weiss L, Grogan T, Pileri SA, et al. Langerhans cell sarcoma. In: Jaffe ES, Harris NL, Stein H, Vardiman JW, eds. World Health Organization classification of tumours. Pathology and genetics of tumours of haematopoietic and lymphoid tissues. Lyon: IARC Press, 2001:283.
- Wester SM, Beabout JW, Unni KK, Dahlin DC. Langerhans cell granulomatosis (histiocytosis X) of bone in adults. *Am J Surg Pathol* 1982;6:413–426.
- Willman CL. Detection of clonal histiocytes in Langerhans cell histiocytosis: biology and clinical significance. *Br J Cancer* 1994; 70(suppl 23):S29–S33.

- Wilner D. Radiology of bone tumors and allied disorders. Philadelphia: Lea & Febiger, 1982.
- Yu RC, Chu C, Buluwela L, Chu AC. Clonal proliferation of Langerhans cells in Langerhans cell histiocytosis. *Lancet* 1994; 343:767–768.
- 80. Zelger B. Position paper. Langerhans cell histiocytosis: A reactive or neoplastic disorder? *Med Pediatr Oncol* 2001;37:543–544.

Ewing Sarcoma

- 81. Baraga JJ, Amrami KK, Swee RG, et al. Radiographic features of Ewing's sarcoma of the bones of the hands and feet. *Skeletal Radiol* 2001;30:121–126.
- Bator SM, Bauer TW, Marks KE, Norris DG. Periosteal Ewing's sarcoma. *Cancer* 1986;58:1781–1784.
- Bertoni F, Bacchini P, Ferruzzi A. Small round-cell malignancies of bone. Ewing's sarcoma, malignant lymphoma, and myeloma. *Semin Orthop* 1991;6:186–195.
- Boyko OB, Cory DA, Cohen MD, et al. MR imaging of osteogenic and Ewing's sarcoma. Am J Roentgenol 1987;148: 317–322.
- Bridge JA, Fidler ME, Neff JR, et al. Adamantinoma-like Ewing's sarcoma: genomic confirmation, phenotypic drift. *Am J Surg Pathol* 1999;23:159–165.
- 86. Carpentieri DF, Qualman SJ, Bowen J, et al. Protocol for the examination of specimens from pediatric and adult patients with osseous and extraosseous Ewing sarcoma family of tumors, including peripheral primitive neuroectodermal tumor and Ewing sarcoma. Arch Pathol Lab Med 2005;129:866–873.
- Cavazzana AO, Miser JS, Jefferson J, Triche TJ. Experimental evidence for a neural origin of Ewing's sarcoma of bone. *Am J Pathol* 1987;127:507–518.
- Coombs RJ, Zeiss J, McKann K, Phillips E. Multifocal Ewing's tumor of the skeletal system. *Skeletal Radiol* 1986;15:254–257.
- Czerniak B, Rojas-Corona RR, Dorfman HD. Morphologic diversity of long bone adamantinoma. The concept of differentiated (regressing) adamantinoma and its relationship to osteofibrous dysplasia. *Cancer* 1989;64:2319–2334.
- Dahlin DC, Unni KK. Bone tumors. General aspects and data on 8,542 cases, 4th ed. Springfield, IL: Charles C Thomas, 1986.
- Dardick I, Schatz JE, Colgan TJ. Osteogenic sarcoma with epithelial differentiation. Ultrastruct Pathol 1992;16:463–474.
- Dehner LP. Primitive neuroectodermal tumor and Ewing's sarcoma. Am J Surg Pathol 1993;17:1–13.
- Dellagi K, Lipinski M, Paulin D, et al. Characterization of intermediate filaments expressed by Ewing tumor cell lines. *Cancer* 1987;47:1170–1173.
- Eggli KD, Quiogue T, Moser RP. Ewing's sarcoma. Radiol Clin North Am 1993;31:325–337.
- Enzinger FM, Weiss SW. Soft tissue tumors. St. Louis: CV Mosby, 1988:951–958.
- Estes DN, Magill HL, Thompson EL, Hayes FA. Primary Ewing sarcoma: follow-up with Ga-67 scintigraphy. *Radiology* 1990;177: 449–453.
- 97. Fechner RE, Mills SE. *Tumors of the bones and joints.* Washington, DC: Armed Forces Institute of Pathology, 1993.
- Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002.
- Friedman B, Hanoaka H. Round cell sarcoma of bone. J Bone Joint Surg Am 1971;53:1118–1136.
- 100. Frouge C, Vanel D, Coffre C, et al. The role of magnetic resonance imaging in the evaluation of Ewing's sarcoma. A report of 27 cases. *Skeletal Radiol* 1988;17:387–392.
- Garber CZ. Reactive bone formation in Ewing's sarcoma. Cancer 1951;4:839–845.
- 102. Gould VE, Moll, R, Berndt R, et al. Immunohistochemical analysis of Ewing's tumors. *Lab Invest* 1987;56:28 (abst).
- 103. Greco A, Steiner GC, Fazzini E. Ewing's sarcoma with epithelial differentiation. Fine structural and immunocytochemical study. *Ultrastruct Pathol* 1988;12:317–325.
- 104. Greenspan A. Orthopedic imaging: a practical approach, 4th ed. Philadelphia: Lippincott Williams & Wilkins, 2004:728–731.
- 105. Hahm KB, Cho K, Lee C, et al. Repression of the gene encoding the TGF-beta type II receptor is a major target of the EWS-FLI1 oncoprotein. *Nat Genet* 1999;23:222–227.

- 106. Hauben E, van den Broek LC, Van Marck E, Hogendoorn PC. Adamantinoma-like Ewing's sarcoma and Ewing's-like adamantinoma. The t(11;22), t(21;22) status. *J Pathol* 2001;195:218–221.
- 107. Hindman BW, Gill HK, Zuppan CW. Primitive neuroectodermal tumor in a child with tuberous sclerosis. *Skeletal Radiol* 1997;26:184–187.
- Huang HY Illei PB, Zhao Z, et al. Ewing sarcomas with p53 mutation or p16/p14ARF homozygous deletion: a highly lethal subset associated with poor chemoresponse. *J Clin Oncol* 2005; 23:548–558.
- Ilaslan H, Sundaram M, Unni KK, Dekutoski MB. Primary Ewing's sarcoma of the vertebral column. *Skeletal Radiol* 2004; 3:506–513.
- Isayama T, Iwasaki H, Kikuchi M, et al. Neuroectodermal tumor of bone. Evidence for neural differentiation in a cultured cell line. *Cancer* 1990;65:1771–1781.
- 111. Jaffe R, Santamaria M, Yunis EJ, et al. The neuro-ectodermal tumor of bone. *Am J Surg* 1984;8:885–898.
- 112. Jones GR, Miller JH, White L, et al. Improved detection of metastatic Ewing's sarcoma with the use of bone marrow scintigraphy. *Med Pediatr Oncol* 1987;15:78–81.
- 113. Khoury JD. Ewing sarcoma family of tumors. *Adv Anat Pathol* 2005;12:212–220.
- Kim HY, Kim YM, Shin YK, et al. Solution structure of the cytoplasmic domain of human CD99 type I. *Mol Cells* 2004;18:24–29.
- 115. Kissane JM, Askin PB, Foulkes M, et al. Ewing's sarcoma of bone. Clinicopathological aspects of 303 cases from the intergroup Ewing's sarcoma study. *Hum Pathol* 1983;14:773–779.
- 116. Kovar H, Jug G, Aryee DN, et al. Among genes involved in the RB dependent cell cycle regulatory cascade, the p16 tumor suppressor gene is frequently lost in the Ewing family of tumors. *Oncogene* 1997;15:2225–2232.
- 117. Kramer K, Hicks D, Palis J, et al. Epithelioid osteosarcoma of bone. Immunocytochemical evidence suggesting divergent epithelial and mesenchymal differentiation in a primary osseous neoplasm. *Cancer* 1993;71:2977–2982.
- Levine E, Levine C. Ewing tumor of rib: Radiologic findings and computed tomography contribution. *Skeletal Radiol* 1983;9: 227–233.
- Llombart-Bosch A, Blache R, Peydro-Olaya A. Ultrastructural study of 28 cases of Ewing's sarcoma: Typical and atypical forms. *Cancer* 1978;41:1362–1373.
- 120. Llombart-Bosch A, Contesso G, Henry-Amar M, et al. Histopathological predictive factors in Ewing's sarcoma of bone and clinicopathologic correlations. A retrospective study of 261 cases. Virchows Arch A 1986;409:627–640 (erratum published in Virchows Arch A 1986;410:263).
- 121. Llombart-Bosch A, Lacombe MJ, Peydro-Olaya A, et al. Malignant peripheral neuroectodermal tumors of bone other than Askin's neoplasm: characterization of 14 new cases with immunohistochemistry and electron microscopy. *Virchows Arch A* 1988;412:421–430.
- 122. Llombart-Bosch A, Terrier LMJ, Peydro-Olaya A, Contesso G. Peripheral neuroectodermal sarcoma of soft tissue (peripheral neuroepithelioma): a pathologic study of ten cases with differential diagnosis regarding other small, round cell sarcomas. *Hum Pathol* 1989;20:273–280.
- Meis-Kindblom JM, Stenman G, Kindblom L-G. Differential diagnosis of small round cell tumors. *Semin Diagn Pathol* 1996;13: 213–241.
- 124. Meister P, Konrad E, Huebner G. Malignant tumor of humerus with features of adamantinoma and Ewing's sarcoma. *Pathol Res Pract* 1979;166:112–122.
- Mirra JM, Picci P, Gold RH, eds. Bone tumors: clinical, radiologic, and pathologic correlations. Philadelphia: Lea & Febiger, 1989.
- 126. Moll R, Lee I, Gould V, et al. Immunocytochemical analysis of Ewing's tumors. Patterns of expression of intermediate filaments and desmosomal proteins indicate cell type heterogeneity and pluripotential differentiation. *Am J Pathol* 1987;127:288–303.
- 127. Mulder JD, Schütte HE, Kroon HM, Taconis WK. Radiologic atlas of bone tumors. Amsterdam: Elsevier, 1993.
- 128. Nair M. Bone scanning in Ewing's sarcoma. J Nucl Med 1985;26:349–352.
- Nascimento AG, Unni KK, Cooper KL, Dahlin DC. A clinicopathologic study of 20 cases of large cell (atypical) Ewing's sarcoma of bone. *Am J Surg Pathol* 1980;4:29–36.

- 130. Nesbit ME Jr, Gehan EA, Burgert EO Jr, et al. Multimodal therapy for the management of primary, nonmetastatic Ewing's sarcoma of bone: a long-term follow-up of the First Intergroup study. J Clin Oncol 1990;8:1664–1674.
- 131. Parham DM, Hijazi Y, Steinberg SM, et al. Neuroectodermal differentiation in Ewing's sarcoma family of tumors does not predict tumor behavior. *Hum Pathol* 1999;30:911–918.
- 132. Reinus WR, Gilula LA. Ewing's sarcoma. In: Taveras JM, Ferruci JT, eds. *Radiology: diagnosis, imaging, intervention*, vol. 5. Philadelphia: JB Lippincott, 1987:Chap. 95.
- 133. Rice RW, Cabot A, Johnston A. The application of electron microscopy to the diagnostic differentiation of Ewing's sarcoma and reticulum cell sarcoma of bone. *Clin Orthop* 1973;97: 174–184.
- 134. Rousselin B, Vanel D, Terrier-Lacombe MJ, et al. Clinical and radiologic analysis of 13 cases of primary neuro-ectodermal tumors of the bone. *Skeletal Radiol* 1989;18:115–120.
- 135. Saifuddin A, Whelan J, Pringle JAS, Cannon SR. Malignant round cell tumours of bone: atypical clinical and imaging features. *Skeletal Radiol* 2000;29:646–651.
- 136. Sandberg AA, Bridge JA. Updates on cytogenetics and molecular genetics of bone and soft tissue tumors: Ewing sarcoma and peripheral primitive neuroectodermal tumors. *Cancer Genet Cytogenet* 2000;123:1–26.
- 137. Schajowicz F. Ewing's sarcoma and reticulum cell sarcoma of bone: with special reference to the histochemical demonstration of glycogen as an aid to differential diagnosis. J Bone Joint Surg Am 1959;41:349–356.
- 138. Schajowicz F. *Tumors and tumorlike lesions of bone*, 2nd ed. Berlin: Springer-Verlag, 1994.
- Shirley SK, Gilula LA, Segal GP, et al. Roentgenographicpathologic correlation of diffuse sclerosis in Ewing's sarcoma of bone. *Skeletal Radiol* 1984;12:69–78.
- 140. Siligan C, Ban J, Bachmaier R, et al. EWS-FLI1 target genes recovered from Ewing's sarcoma chromatin. *Oncogene* 2005;24: 2512–2524.
- 141. Steiner GC, Matano S, Present D. Ewing's sarcoma of humerus with epithelial differentiation. *Skeletal Radiol* 1995;24:379–382.
- Steiner GC. Neuroectodermal tumor versus Ewing's sarcoma. Immunohistochemical and electron microscopic observations. *Curr Top Pathol* 1989;80:1–29.
- 143. Sundaram M, Inwards CY, Shives TE, Anderson PM. Ewing's sarcoma of the humerus mimicking fibrous dysplasia on imaging and biological behavior. *Skeletal Radiol* 2005;34:285–289.
- Triche T, Cavazzana A. Round cell tumors of bone. In: Unni KK, ed. *Bone tumors*. New York: Churchill Livingstone, 1988:199–223.
- 145. Triche TJ, Askin FB, Kissane JM. Neuroblastoma, Ewing's sarcoma and the differential diagnosis of small, round blue cell tumors. In: Finegold M, ed. *Pathology of neoplasia in children and adolescents*. Philadelphia: WB Saunders, 1986:145–156.
- 146. Ulbright TM. Germ cell tumors of the gonads: a selective review emphasizing problems in differential diagnosis, newly appreciated, and controversial issues. *Mod Pathol* 2005;18(suppl 2): S61–S79.
- 147. Unni KK. Dahlin's bone tumors: general aspects and data on 11,087 cases, 5th ed. New York: Lippincott-Raven, 1996.
- 148. Ushigome S, Machinami R. Sorensen P. Ewing sarcoma/primitive neuroectodermal tumor (PNET). In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:298–300.
- Vanel D, Contesso G, Couanet D, et al. Computed tomography in the evaluation of 41 cases of Ewing's sarcoma. *Skeletal Radiol* 1982;9:8–13.
- 150. Vanel D, Couanet D, Leclerc J, Patte C. Early detection of bone metastases of Ewing's sarcoma by magnetic resonance imaging. *Diagn Imag Clin Med* 1986;55:381–383.
- 151. van Haelst UJ, de Haas van Dorsser AH. A perplexing malignant bone tumor. Highly malignant so-called adamantinoma or nontypical Ewing's sarcoma. *Virchows Arch A Pathol Anat Histol* 1975; 365:63–74.
- 152. Wei G, Antonescu CR, de Alava E, et al. Prognostic impact of INK4A deletion in Ewing sarcoma. *Cancer* 2000;89:793–799.
- Weiss SW, Bratthauer GL, Morris PA. Post-radiation malignant fibrous histiocytoma expressing cytokeratin: implications for the immunodiagnosis of sarcomas. *Am J Surg Pathol* 1988;12:554–558.

- Weiss SW, Goldblum JR. Enzinger and Weiss's soft tissue tumors, 4th ed. St. Louis: Mosby, 2001:1632.
- 155. Wilkins RM, Pritchard DJ, Burgert EO, Unni KK. Ewing's sarcoma of bone. Experience with 140 patients. *Cancer* 1986;58: 2551–2555.

Malignant Lymphoma

- 156. Arber DA, George TI. Bone marrow biopsy involvement by non-Hodgkin's lymphoma: frequency of lymphoma types, patterns, blood involvement, and discordance with other sites in 450 specimens. *Am J Surg Pathol* 2005;29:1549–1557.
- Beackley MC, Lau BP, King ER. Bone involvement in Hodgkin's disease. Am J Roentgenol 1972;114:559–563.
- Boston HC Jr, Dahlin DC, Ivins JC, Cupps RE. Malignant lymphoma (so-called reticulum cell sarcoma) of bone. *Cancer* 1974; 34:1131–1137.
- Bragg DG, Colby TV, Ward JH. New concepts in the non-Hodgkin lymphomas: radiologic implications. *Radiology* 1986; 159:289–304.
- Braunstein EM. Hodgkin's disease of bone: radiographic correlation with the histologic classification. *Radiology* 1980;137: 643–646.
- Bullough PG. Atlas of orthopedic pathology with clinical and radiologic correlations, 2nd ed. New York: Gower, 1992:17.1–17.29.
- 162. Campbell SE, Filzen TW, Bezzant SM, et al. Primary periosteal lymphoma: an unusual presentation of non-Hodgkin's lymphoma with radiographic, MR imaging, and pathologic correlation. *Skeletal Radiol* 2003;32:231–235.
- Castellino RA. Hodgkin disease: practical concepts for the diagnostic radiologist. *Radiology* 1986;159:305–310.
- 164. Chan K-W, Rosen G, Miller DR, Tan CTC. Hodgkin's disease in adolescents presenting as a primary bone lesion. A report of four cases and review of the literature. *Am J Pediatr Hematol Oncol* 1982;4:11–17.
- 165. Chao S, Mullins ME, Gallagher L, Slanetz PJ. Primary non-Hodgkin's lymphoma of the femur. Am J Roentgenol 2001;176: 1160.
- Clayton F, Butler JJ, Ayala AG, et al. Non-Hodgkin's lymphoma in bone: pathologic and radiologic features with clinical correlates. *Cancer* 1987;60:2494–2500.
- Cohen MD, Klatte EC, Smith JA, et al. Magnetic resonance imaging of lymphomas in children. *Pediatr Radiol* 1985;15:179–183.
- Coles WC, Schultz MD. Bone involvement in malignant lymphoma. *Radiology* 1948;50:458–462.
- Daffner RN, Lupetin AR, Dash N, et al. MRI in the detection of malignant infiltration of bone marrow. Am J Roentgenol 1986; 146:353–358.
- Fisher AMH, Kendall B, Van Leuven BD. Hodgkin's disease: a radiological survey. *Clin Radiol* 1962;13:115–127.
- 171. Fishman EK, Kuhlman JE, Jones RJ. CT of lymphoma: spectrum of disease. *RadioGraphics* 1991;11:647–669.
- 172. Gebert C, Hardes J, Ahrens H, et al. Primary multifocal osseous Hodgkin disease: a case report and review of the literature. J Cancer Res Clin Oncol 2005;131:163–168.
- 173. Gold RH, Mirra JM. Case report 101. Primary Hodgkin disease of humerus. *Skeletal Radiol* 1979;4:233–235.
- 174. Granger W, Whitaker R. Hodgkin's disease in bone, with special reference to periosteal reaction. *Br J Radiol* 1967;40:939–948.
- 175. Greco A, Jelliffe AM, Maher EJ, Leung AWL. MR imaging of lymphomas: impact on therapy. J Comput Assist Tomogr 1988;12: 785–791.
- 176. Guermazi A, Brice P, de Kerviler E, et al. Extranodal Hodgkin disease: spectrum of disease. *RadioGraphics* 2001;21:161–179.
- 177. Harris NL, Jaffe ES, Vardiman JW, et al. WHO classification of tumours of haematopoietic and lymphoid tissues: Introduction. In: Jaffe ES, Harris NL, Stein H, Vardiman JW, eds. World Health Organization classification of tumours. Pathology and genetics of tumours of haematopoietic and lymphoid tissues. Lyon: IARC Press, 2001:12–13.
- Hermann G, Klein MJ, Abdelwahab IF, Kenan S. MRI appearance of primary non-Hodgkin's lymphoma of bone. *Skeletal Radiol* 1997;26:629–632.
- 179. Hicks DG, Gokan T, O Keefe RJ, et al. Primary lymphoma of bone. Correlation of MRI features with cytokine production by tumor cells. *Cancer* 1995;75:973–980.

- Hillemanns M, McLeod RA, Unni KK. Malignant lymphoma. Skeletal Radiol 1996;25:73–75.
- 181. Hsieh PP, Tseng HH, Chang ST, et al. Primary non-Hodgkin's lymphoma of bone: a rare disorder with high frequency of T-cell phenotype in southern Taiwan. *Leuk Lymphoma* 2006;47:65–70.
- 182. Jaffe HL. Metabolic, degenerative, and inflammatory diseases of bones and joints. Philadelphia: Lea & Febiger, 1972.
- 183. Kaplan H. Hodgkin disease, 2nd ed. Cambridge, MA: Harvard University Press, 1980:85–92.
- Klein MJ, Rudin BJ, Greenspan A, et al. Hodgkin disease presenting as a lesion in the wrist. J Bone Joint Surg Am 1987;69: 1246–1249.
- Kremer M, Quintanilla-Martinez L, Nahrig J, et al. Immunohistochemistry in bone marrow pathology: a useful adjunct for morphologic diagnosis. *Virchows Arch* 2005;447:920–937.
- 185a. Krishnan A, Shirkhoda A, Tehranzadeh J, et al. Primary bone lymphoma: radiographic-MR imaging correlation. *RadioGraphics* 2003;23:1371–1387.
- 186. Lambertenghi-Deliliers G, Annaloro C, Soligo D, et al. Incidence and histological features of bone marrow involvement in malignant lymphomas. *Ann Hematol* 1992;65:61–65.
- 187. Leeson MC, Makely JT, Carter JR, Krupco T. The use of radioisotope scans in the evaluation of primary lymphoma of bone. Orthop Rev 1989;18:410–416.
- Malloy PC, Fishman EK, Magid D. Lymphoma of bone, muscle, and skin: CT findings. Am J Roentgenol 1992;159:805–809.
- Manaster BJ. Primary bone lymphoma: radiographic-MR imaging correlation. Invited commentary. *RadioGraphics* 2003;23: 1384–1386.
- 190. Melamed JW, Martinez S, Hoffman CJ. Imaging of primary multifocal osseous lymphoma. *Skeletal Radiol* 1997;26:35–41.
- 191. Mirra JM. Lymphoma and lymphoma-like disorders. In: Mirra JM, Picci P, Gold RH, eds. Bone tumors: clinical, radiologic, and pathologic correlations. Philadelphia: Lea & Febiger, 1989.
- 192. Mulligan ME, Kransdorf MJ. Sequestra in primary lymphoma of bone: prevalence and radiologic features. *Am J Roentgenol* 1993; 160:1245–1248.
- Mulligan ME, McRae GA, Murphey MD. Imaging features of primary lymphoma of bone. *Am J Roentgenol* 1999;173:1 691–1697.
- 194. Negendank W, Weissman D, Bey TM, et al. Evidence for clonal disease by magnetic resonance imaging in patients with hypoplastic marrow disorders. *Blood* 1991;78:2872–2879.
- Newcomer LN, Silverstein MB, Cadman EC, et al. Bone involvement in Hodgkin's disease. *Cancer* 1982;49:338–342.
- 196. Olson DO, Shields AF, Scheurich JC, et al. Magnetic resonance imaging of the bone marrow in patients with leukemia, aplastic anaemia and lymphoma. *Invest Radiol* 1986;21:540–546.
- 197. Orzel JA, Sawaf NW, Richardson ML. Lymphoma of the skeleton: scintigraphic evaluation. *Am J Roentgenol* 1988;150:1095–1099.
- Ostrowski ML, Unni KK, Banks PM, et al. Malignant lymphoma of bone. *Cancer* 1986;58:2646–2655.
- 199. Perlman EJ, Dickman PS, Askin FB, et al. Ewing's sarcoma routine diagnostic utilization of MIC2 analysis: a Pediatric Oncology Group/Children's Cancer Group Intergroup Study. *Hum Pathol* 1994;25:304–307.
- 200. Pettit CK, Zukerberg LR, Gray MH, et al. Primary lymphoma of bone. A B-cell neoplasm with a high frequency of multilobated cells. *Am J Surg Pathol* 1990;14:329–334.
- Resnick D, Haghighi P. Myeloproliferative disorders. In: Resnick D, ed. Bone and joint imaging. Philadelphia: WB Saunders, 1989: 703–714.
- 202. Ruzek KA, Wenger DE. The multiple faces of lymphoma of the musculoskeletal system. *Skeletal Radiol* 2004;33:1–8.
- 203. Salter M, Sollaccio RJ, Bernreuter WK, Weppelman B. Primary lymphoma of bone: the use of MRI in pretreatment evaluation. *Am J Clin Oncol* 1989;12:101–105.
- 204. Stein H. Hodgkin lymphomas: Introduction. In: Jaffe ES, Harris NL, Stein H, Vardiman JW, eds. World Health Organization classification of tumours. Pathology and genetics of tumours of haematopoietic and lymphoid tissues. Lyon: IARC Press, 2001:239.
- 205. Stein H, Delsol G, Pileri SA, et al. Classical Hodgkin lymphoma. In: Jaffe ES, Harris NL, Stein H, Vardiman JW, eds. World Health Organization classification of tumours. Pathology and genetics of tumours of haematopoietic and lymphoid tissues. Lyon: IARC Press, 2001:244–253.

- Stein H, Kaiserling E, Lennert K. Evidence for B-cell origin of reticulum cell sarcoma. Virchow Arch A 1974;A364:51–67.
- 207. Stiglbauer R, Augustin I, Kramer J, et al. MRI in the diagnosis of primary lymphoma of bone: correlation with histopathology. *J Comput Assist Tomogr* 1992;16:248–253.
- 208. Unni K. Dahlin's bone tumors: general aspects and data on 11,087 cases, 5th ed. Philadelphia: Lippincott-Raven, 1996:463.
- 209. Unni K, Hogendoorn PCW. Malignant lymphoma. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:306–308.
- Vincent JM, NG YY, Norton AJ, Armstrong PA. Case report. Primary lymphoma of bone: MRI appearances with pathological correlation. *Clin Radiol* 1992;45:407–409.
- White LM, Schweitzer ME, Khalili K, et al. MR imaging of primary lymphoma of bone: variability of T2-weighted signal intensity. *Am J Roentgenol* 1998;170:1243–1247.

Multiple Myeloma (Plasmacytoma)

- 212. Aggarwal S, Goulatia RK, Sood A, et al. POEMS syndrome: a rare variety of plasma cel dyscrasia. *Am J Roentgenol* 1990;155: 339–341.
- 213. Anguaco EJC, Fassas ABT, Walker R, et al. Multiple myeloma: clinical review and diagnostic imaging. *Radiology* 2004;231:11–23.
- 214. Avrahami E, Tadmor R, Kaplinsky N. The role of T2-weighted gradient echo in MRI demonstration of spinal multiple myeloma. *Spine* 1993;18:1812–1815.
- Banerjee SS, Verma S, Shanks JH. Morphological variants of plasma cell tumours. *Histopathology* 2004;44:2–8.
- 216. Bardwick PA, Zvaifler NJ, Gill GN, et al. Plasma-cell dyscrasia with polyneuropathy, organomegaly, endocrinopathy, M-protein and skin changes: the POEMS syndrome. Report on two cases and review of the literature. *Medicine* 1980;59:311–322.
- 217. Bataille R, Chevalier J, Ross M, Sany J. Bone scintigraphy in plasma-cell myeloma. *Radiology* 1982;145:801–804.
- 218. Bataille R, Sany J. Solitary myeloma: clinical and prognostic features of a review of 114 cases. *Cancer* 1981;48:845–851.
- 219. Baur A, Stäbler A, Lamerz R, et al. Light chain deposition disease in multiple myeloma: MR imaging features correlated with histopathological findings. *Skeletal Radiol* 1998;27:173–176.
- Bergsagel DE. Plasma cell myeloma. An interpretive review. Cancer 1972;30:1588–1594.
- Bergsagel PL, Kuehl WM. Molecular pathogenesis and a consequent classification of multiple myeloma. *J Clin Oncol* 2005;23: 6333–6338.
- Bernstein SC, Perez-Atayde AR, Weinstein HJ. Multiple myeloma in a child. *Cancer* 1985;56:2143–2147.
- 223. Bessler W, Antonucci F, Stamm B, et al. Case report 646. POEMS syndrome. *Skeletal Radiol* 1991;20:212–215.
- Bradwey TM, Murphy DA, Eyster RL, Cannon MW. Multiple myeloma in a 25-year-old woman. *Clin Orthop* 1993;294:290–293.
- 225. Brandon C, Martel W, Weatherbee L, Capek P. Case report 572. Osteosclerotic myeloma (POEMS syndrome). *Skeletal Radiol* 1989;18:542–546.
- 226. Brown TS, Paterson CR. Osteosclerosis in myeloma. J Bone Joint Surg Br 1973;55:621–623.
- Campanacci M. Plasmacytoma. In: Bone and soft tissue tumors. New York: Springer-Verlag, 1990:560.
- Cohen HJ, Crawford J, Rao MK, et al. Racial differences in the prevalence of monoclonal gammopathy in a community-based sample of the elderly. *Am J Med* 1998;104:439–444.
- De Raeve HR, Vanderkerken K. The role of the bone marrow microenvironment in multiple myeloma. *Histol Histopathol* 2005; 20:1227–1250.
- Dimopoulos MA, Moulopoulos A, Delasalle K, Alexanian R. Solitary plasmacytoma of bone and asymptomatic multiple myeloma. *Hematol Oncol Clin North Am* 1992;6:359–369.
- 231. Durie BGM, Salmon SE. A clinical staging system for multiple myeloma. *Cancer* 1975;36:842–854.
- Durie BG, Waxman AD, D'Agnolo A. Williams CM. Whole-body (18)F-FDG PET identifies high-risk myeloma. J Nucl Med 2002;43:1457–1463.
- Dwyer AJ, Frank SL, Sank VJ, et al. Short TI inversion recovery pulse sequence: analysis and initial experience in cancer imaging. *Radiology* 1988;168:827–836.

- 234. Ferlay J, Bray F, Pisani P, Parkin DM. Globocan 2000. Cancer incidence, mortality and prevalence worldwide. Lyon: IARC Press, 2001. [CD-ROM, last update: 2002; http://www.iarc.fr].
- 235. Frassica DA, Frassica FJ, Schray MF, et al. Solitary plasmacytoma of bone: Mayo Clinic experience. *Int J Radiat Oncol Biol Phys* 1989;16:43–48.
- 236. Fruehwald FX, Tschalakoff D, Schwaighofer B, et al. Magnetic resonance imaging of the lower vertebral column in patients with multiple myeloma. *Invest Radiol* 1988;23:193–199.
- 237. Greipp PR, San Miguel J, Durie BG, et al. International staging system for multiple myeloma. *J Clin Oncol* 2005;23: 3412–3420.
- 238. Grogan TM, van Camp B, Kyle RA, et al. Plasma cell neoplasms. In: Jaffe ES, Harris NL, Stein H, Vardiman JW, eds. World Health Organization classification of tumours. Pathology and genetics of tumours of haematopoietic and lymphoid tissues. Lyon: IARC Press, 2001:142–156.
- Hall FM, Gore SM. Osteosclerotic myeloma variant. Skeletal Radiol 1988;17:101–105.
- Harrouseau JL, Greil R, Kloke O. ESMO Minimum clinical recommendations for diagnosis, treatment and follow-up of multiple myeloma. *Ann Oncol* 2005;16(suppl 1):i45–i47.
- 241. Heider U, Hofbauer LC, Zavrski I, et al. Novel aspects of osteoclast activation and osteoblast inhibition in myeloma bone disease. *Biochem Biophys Res Commun* 2005;338:687–693.
- 242. Hermann H, Abdelwahab IF, Berson BD, et al. Case report 621. Multiple myeloma (IgD) in a 28 year old woman. *Skeletal Radiol* 1990;19:379–381.
- 243. Hewell GM, Alexanian R. Multiple myeloma in young persons. Ann Intern Med 1976;84:441–443.
- 244. Hübner K. The pathology of the multiple myeloma (plasmacytoma). Verh Dtsch Ges Pathol 1983;67:423–439.
- 245. International Myeloma Working Group. Criteria for the classification of monoclonal gammopathies, multiple myeloma and related disorders: a report of the International Myeloma Working Group. *Br J Haematol* 2003;121:749–757.
- Ishida T, Dorfman HD. Plasma cell myeloma in unusually young patients: a report of two cases and review of the literature. *Skeletal Radiol* 1995;24:47–51.
- 247. Kelly JJ Jr, Kyle RA, Miles JM, Dyck PJ. Osteosclerotic myeloma and peripheral neuropathy. *Neurology* 1983;33:202–210.
- Kuehl WM, Bergsagel PL. Multiple myeloma: evolving genetic events and host interactions. *Nat Rev Cancer* 2002;2:175–187.
- 249. Kyle RA. Diagnostic criteria of multiple myeloma. *Hematol Oncol Clin North Am* 1992;6:347–358.
- Kyle RA. Multiple myeloma: review of 869 cases. Mayo Clin Proc 1975;50:29–40.
- 251. Kyle RA, Bayrd ED. Amyloidosis: review of 236 cases. *Medicine* 1975;54:271–299.
- 252. Lazarus HM, Kellermeyer RW, Aikaisa M, Herrig RH. Multiple myeloma in young men: clinical course and electron microscopic studies of bone marrow plasma cells. *Cancer* 1980;46: 1397–1400.
- 253. Lipshitz HI, Malthouse SR, Cunningham D, et al. Multiple myeloma: appearance at MR imaging. *Radiology* 1992;182: 833–837.
- 254. Mahnken AH, Wildberger JE, Gehbauer G, et al. Multidetector CT of the spine in multiple myeloma: Comparison with MR imaging and radiography. *Am J Roentgenol* 2002;178:1429–1436.
- 255. Major NM, Helms CA, Richardson WJ. The "mini brain": Plasmacytoma in a vertebral body on MR imaging. Am J Roentgenol 2000;175:261–263.
- 256. Meis JM, Butler JJ, Osborne BM, Ordonez NG. Solitary plasmacytomas of bone and extramedullary plasmacytomas. A clinicopathologic and immunohistochemical study. *Cancer* 1987;59: 1475–1485.
- 257. Meyer JE, Schulz MD. Solitary myeloma of bone: a review of 12 cases. *Cancer* 1974;34:438–440.
- Mochizuki K, Katayama K, Kikuchi F, et al. Immunoglobulin D myeloma presenting as an extraosseous soft tissue tumor. *Skeletal Radiol* 2001;30:166–169.
- 259. Moulopoulos LA, Varma DGK, Dimopoulos MA, et al. Multiple myeloma: spinal MR imaging in patients with untreated newly diagnosed disease. *Radiology* 1992;185:833–840.
- Murray RO, Jacobson HG. The radiology of bone diseases, 2nd ed. New York: Churchill Livingstone, 1977.

- Nilsson-Ehle H, Holmdahl C, Suurkula M, Westin J. Bone scintigraphy in the diagnosis of skeletal involvement and metastatic calcification in plasma cell-related dyscrasia. *Acta Med Scand* 1982; 211:427–432.
- 262. Pambuccian SE, Horyd ID, Cawte T, Huvos AG. Amyloidoma of bone, a plasma cell/plasmacytoid neoplasm. Report of three cases and review of the literature. *Am J Surg Pathol* 1997;21: 179–186.
- 263. Rahmouni A, Divine M, Mathieu D, et al. Detection of multiple myeloma involving the spine: efficacy of fat-suppression and contrast-enhanced MR imaging. Am J Roentgenol 1993;160: 1049–1052.
- 264. Reinus WR, Kyriakos M, Gilula LA, et al. Plasma cell tumors with calcified amyloid deposition mistaken for chondrosarcoma. *Radiology* 1993;189:505–509.
- 265. Resnick D, Greenway GD, Bardwick PA, et al. Plasma cell dyscrasia with polyneuropathy, organomegaly, endocrinopathy, M-protein, and skin changes: the POEMS syndrome. *Radiology* 1981;140:17–22.
- 266. Resnick D, Haghighi P, Guerra J Jr. Bone sclerosis and proliferation in a man with multisystem disease. *Invest Radiol* 1984; 19:1–6.
- 267. Ricci C, Cova M, Kang YS, et al. Normal age-related patterns of cellular and fatty bone marrow distribution in the axial skeleton: MR imaging study. *Radiology* 1990;177:83–88.
- 268. Ries LAG, Eisner MP, Kosary CL, et al., eds. SEER Cancer Statistics Review, 1975-2002. Bethesda, MD: National Cancer Institute. Available at http://seer.cancer.gov/csr/1975_2002/, based on

November 2004 SEER data submission, posted to the SEER web site 2005.

- 269. Sartoris DJ, Pate D, Haghighi P, et al. Plasma cell sclerosis of bone: a spectrum of disease. *Can Assoc Radiol* J 1986;37:25–34.
- 270. Shimpo S. Solitary myeloma causing polyneuritis and endocrine disorders. *Jpn J Clin Med* 1968;26:2444–2456.
- 271. Solomon Ä, Řahamani R, Seligsohn U, Ben-Artzi F. Multiple myeloma: early vertebral involvement assessed by computerized tomography. *Skeletal Radiol* 1984;11:258–261.
- 272. Stäbler Å, Baur A, Bartl R, et al. Contrast enhancement and quantitative signal analysis in MR imaging of multiple myeloma: assessment of focal and diffuse growth patterns in marrow correlated with biopsies and survival rates. *Am J Roentgenol* 1996; 167:1029–1036.
- 273. Steiner RM, Mitchell DG, Rao VM, Schweitzer ME. Magnetic resonance imaging of diffuse bone marrow disease. *Radiol Clin North Am* 1993;31:383–409.
- 274. Tertti R, Alanen A, Remes K. The value of magnetic resonance imaging in screening myeloma lesions of the lumbar spine. Br J Haematol 1995;91:658–660.
- 275. Voss SD, Murphey MD, Hall FM. Solitary osteosclerotic plasmacytoma: association with demyelinating polyneuropathy and amyloid deposition. *Skeletal Radiol* 2001;30:527–529.
- 276. Wilner D. Radiology of bone tumors and allied disorders. Philadelphia: Lea & Febiger, 1982.
- 277. Woolfenden JM, Pitt MJ, Durie BGM, Moon TE. Comparison of bone scintigraphy and radiography in multiple myeloma. *Radiology* 1980;134:723–728.

CHAPTER 6

Vascular Lesions

BENIGN LESIONS 364 Intraosseous Hemangioma 364 Synovial Hemangioma 368 Cystic Angiomatosis 368 Lymphangioma and Lymphangiomatosis 371 Glomus Tumor, Glomangioma, and Glomangiomyoma 374

MALIGNANT TUMORS 374 Epithelioid Hemangioendothelioma 375 Angiosarcoma 378 Hemangiopericytoma 380

Vascular lesions of bone constitute a spectrum of pathologic entities. These range from congenital vascular malformations, i.e., errors of morphogenesis with nonproliferative capacity, which grow in parallel with the patient, such as angiomatosis, to benign neoplasms that may be present at birth, and highly malignant tumors that metastasize widely and have low survival rates (e.g., angiosarcoma) (28,101).

Recently, immunohistochemistry (IHC) studies have been conducted in attempts to discriminate between malformations and true tumors, the former being negative for WT1 [a transcription factor initially isolated from a hereditary Wilms tumor encoded by the socalled Wilms tumor 1 (*WT1*) gene (62)] and GLUT1 [an erythrocyte-type glucose transporter protein not present in normal vasculature and vascular malformations but found at sites of blood-tissue barriers and in vascular tumors (63,80)]. In addition, lymphatic lesions have been successfully identified by another monoclonal antibody, D2-40, that reacts with a formalinresistant epitope on lymphatic endothelium (36,56). Boye et al. (12) were able to demonstrate clonality of cells isolated from proliferating (cellular) hemangiomas as well as alterations of their in vitro behavior, leading to the hypothesis that somatic mutations may be involved in their pathogenesis. Clonality of juvenile hemangiomas was also proved by Walter et al. (105) in 12 of 14 investigated cases. These authors also found somatic mutations in genes coding for the receptors (VEGFR2 and VEGFR3) of the vascular endothelial growth factors (VEGFs), which play an important role in angiogenesis and vascular formation (19).

Benign vascular tumors are of either endothelial or pericytic origin. The first group includes hemangiomas (2). Cystic angiomatosis, lymphangiomas, and lymphangiomatosis, although malformations, are also included here for reasons of differential diagnosis. The second group includes glomus tumor and its variants, glomangioma and glomangiomyoma. The malignant and locally aggressive varieties are quite uncommon and are mostly of endothelial (hemangioendothelioma and angiosarcoma) origin. The existence of hemangiopericytoma as a distinct tumor entity is currently a matter of debate (137).

Benign Lesions

Intraosseous Hemangioma

Hemangioma is a benign lesion composed of newly formed blood vessels, often discovered incidentally on radiographs performed for other reasons. It composes approximately 2% of all benign and 0.8% of benign and malignant lesions of the skeletal system (77). On the basis of their site of origin, hemangiomas in general are classified as intraosseous, intracortical, periosteal, intraarticular (synovial), intramuscular, subcutaneous, or cutaneous. Depending on size of the vessels predominating within the lesion, hemangioma is subclassified into capillary or cavernous types (25). Capillary hemangiomas are composed of small vessels that consist merely of a flat endothelium, surrounded only by a basal membrane. In bone they most commonly occur in the vertebral body (86). Cavernous hemangiomas are composed of dilated, blood-filled spaces lined by the same flat endothelium with a basal membrane. Osseous cavernous hemangiomas most commonly involve the calvaria (29).

The biological classification of vascular anomalies has recently gained renewed attention (72). Based on a system introduced by Mulliken and Glowacki (78), and redefined by Enjorlas and Mulliken in 1997 (28), this classification takes into consideration cellular turnover and histology, as well as natural history and physical findings. It clearly separates hemangiomas of infancy, with their early proliferative and later involutional stages, from vascular malformations, which are characterized as arterial, venous, capillary, lymphatic, or combined (24,72,78). The terms "venous hemangioma" (a lesion composed of thick-walled vessels that possess a muscle layer and frequently contains phleboliths) and "arteriovenous hemangioma" (characterized by abnormal communications between arteries and veins) are no longer in use. These lesions occur exceedingly rarely in bone, involve almost exclusively the soft tissues and at present time are considered as vascular malformations (28).

Clinical Presentation

The incidence of intraosseous hemangiomas increases with age, and most patients present after their fourth decade. The majority of patients are asymptomatic and women are affected twice as often as men. The most common sites are vertebral bodies (particularly in the thoracic segment) and the skull. Hemangiomas of the long bones (e.g., femur, humerus) and short tubular bones are less often encountered (26,57,71) (Fig. 6-1). Calvarial hemangiomas most commonly affect the frontal and parietal regions, and account for 20% of all intraosseous hemangiomas (74). These lesions arise in the diploic space and cause expansion that often involves the outer table to a great extent (79). Vertebral hemangiomas are very common and are observed in 11% of autopsied cases (50). They account for 28% of all skeletal hemangiomas (74). In this location the lesion typically involves the vertebral body, although it may extend into the lamina and, rarely, into the spinous process. Multiple vertebrae are sometimes affected.



Figure 6-1 Hemangioma: skeletal sites of predilection, peak age range, and male-to-female ratio.



Figure 6-2 Intraosseous hemangioma. Dorsovolar radiograph of the right hand of an 11-year-old girl with hemangioma involving the middle finger shows the lace-like pattern and honeycombing characteristic of this lesion.

The majority of vertebral hemangiomas are asymptomatic and are discovered incidentally. The pathologic fractures are rare (35). Symptoms occur when the affected vertebra expands in the direction of the spinal cord and causes compression (34,70,86). This neurologic complication is most commonly associated with lesions in the midthoracic spine (43,59,75). Another mechanism considered responsible for compression of the cord, although less frequently, is a fracture of the involved vertebral body and the formation of a soft tissue mass or hematoma (5,9).

Imaging

On radiography, an osseous hemangioma in a long or short tubular bone is characterized by coarse striations or by multifocal lytic areas (60,93) (Fig. 6-2). Periosteal and cortical hemangiomas occur most commonly in the anterior tibial diaphysis. These lesions present as lytic, cortical erosions, occasionally accompanied by a periosteal reaction. In the vertebral body, hemangioma is characterized either by vertical striations or by multiloculated lytic foci (108). These appearances are referred to as a corduroy cloth pattern (Fig. 6-3A) or honeycomb pattern (Fig. 6-3B), respectively, and are considered virtually pathognomonic for this lesion (60). A calvarial hemangioma commonly appears as a lytic lesion that exhibits a radiating configuration of



Figure 6-3 Vertebral hemangioma. A: Anteroposterior tomography demonstrates vertical striations of hemangioma of L1 vertebra (*arrows*), referred to as a "corduroy cloth" pattern. **B:** Lateral radiograph of the lumbar spine demonstrates a "honeycomb" pattern of hemangioma of L2 vertebra (*arrowhead*).



Figure 6-4 Vertebral hemangioma: computed tomogra-phy (CT). CT section of a T10 vertebra demonstrates coarse dots that indicate reinforced vertical trabeculae of the cancellous bone, characteristic of hemangioma.

thickened trabeculae, frequently arranged in a spokewheel or web-like pattern (79).

On computed tomography (CT) scan, vertebral hemangioma characteristically exhibits a pattern of multiple dots (often referred to as the "polka-dot" appearance), representing a cross-section of reinforced trabeculae (Fig. 6-4). On magnetic resonance imaging (MRI), T1- and T2weighted images usually reveal areas of a high-intensity signal that correspond to the vascular components, adipocytes, and interstitial edema (7,48,89,102) (Fig. 6-5). Areas of trabecular thickening exhibit a low signal intensity regardless of the pulse sequence used (79). Both CT and MRI obtained after intravenous administration of contrast material demonstrate a lesion enhancement (16,79,97,110).

On scintigraphy, the appearance of osseous hemangiomas ranges from photopenia (37,68) to a moderate increase in the uptake of radiopharmaceutical tracer (54,76,83). A recent study of single photon emission computed tomography (SPECT) imaging of vertebral hemangiomas and their correlation with MRI showed that in most cases hemangiomas exhibited normal uptake on planar images. SPECT images were also normal, particularly if the lesions were less than 3 cm in diameter. This study also showed a disparity between SPECT images and MRI: there was no correlation between MRI signal intensity changes and patterns of uptake on bone imaging (47). Arteriography of the hemangioma is rarely indicated (64).

Histopathology

On histologic examination, most hemangiomas consist of simple channels formed by an endothelium-lined basal membrane, morphologically identical with normal capillaries (Fig. 6-6). The lining endothelial cells are small, flat, uniform, and inconspicuous (50). Mitotic figures are rarely seen (21). Some or all of the vascular channels may be enlarged and may have a sinusoidal appearance, in which case the lesion is



Figure 6-5 Vertebral hemangioma: magnetic resonance imaging (MRI). Sagittal T1-weighted (SE, TR 517, TE 12) (A) and T2-weighted (SE, TR 2000, TE 80) (B) MR images show high signal intensity of hemangioma of L4 vertebra.



Figure 6-6 Histopathology of capillary intraosseous hemangioma. Blood vessels of capillary character and partly of cavernous type lie between small bone trabeculae and border normal cancellous bone of the vertebrae containing hematopoietic marrow (*below*) (hematoxylin and eosin, original magnification $\times 25$).

referred to as cavernous type (Fig. 6-7), which incidentally is the most common, particularly in the calvaria (31). These lesions are composed of multiple large, thin-walled, dilated, blood-filled vascular spaces lined by endothelial cells (50). Endothelial cells of hemangiomas are consistently positive for CD31, CD34, and factor VIII-related antigen. It has recently been shown that the Fli-1 protein, a transcription factor involved not only in cell proliferation and tumorigenesis, e.g., in Ewing sarcoma/primitive neuroectodermal tumor (PNET) and lymphoma but also in formation of endothelial cells, including lymphatic endothelium, is expressed in the nucleus of normal endothelia and in endothelium-derived benign and malignant tumors (32,33). The hemangiomatous tissue often spreads between the preexisting bone trabeculae without distorting the original bone pattern (77).

The designation epithelioid hemangioma is used for vascular lesions that, unlike epithelioid hemangioendothelioma, are often arranged in a lobular fashion (66,98,99,107). According to O'Connell et al. (81,162), more than 50% of neoplastic vessels should be lined by epithelioid endothelial cells to establish the diagnosis. Well-formed vessels with open lumina contain enlarged eosinophilic endothelial cells, sometimes protruding into the lumen, with vesicular nuclei and occasionally with prominent nucleoli. However, overt nuclear atypia is absent. Typical mitoses can be detected, and cytoplasmic vacuoles may be seen. The stroma often contains an inflammatory infiltrate consisting of lymphocytes, eosinophils, and extravasated erythrocytes, but myxoid change or hyalinization is not present. However, the existence of an epithelioid hemangioma in bone similar to its soft tissue counterpart [which is also designated as angiolymphoid hyperplasia with eosinophilia or histiocytic hemangioma and which is regarded by some authors as being reactive (106)] has been questioned by others (25,30).

Differential Diagnosis

Radiology

Solitary hemangioma in a small tubular bone, such as a metacarpal or metatarsal, may exhibit an expansive appearance, thus mimicking an *aneurysmal bone cyst* (20). In a long tubular bone, particularly when the lesion is predominantly radiolucent, a focus of *fibrous dysplasia* may be the consideration.

In a vertebra, the differential possibilities include *Paget disease, myeloma (plasmacytoma)*, and *metastasis*, and in a younger patient hemangioma may mimic a *Langerhans cell histiocytosis*. In patients with a significant degree of *osteopenia*, the vertebrae sometimes exhibit vertical linear streaks, similar to the appearance of vertebral hemangioma (Fig. 6-8). In *Paget disease*, the vertebral body, unlike in hemangioma, is frequently enlarged,



Figure 6-7 Histopathology of cavernous intraosseous hemangioma. A: Cortical bone (*top*) and cancellous bone (*bottom*) stained red enclosing large, mostly empty vascular spaces (van Gieson, original magnification \times 6). B: At higher magnification large vascular spaces filled with blood cells lie in the marrow spaces of cancellous bone. Surfaces of the trabeculae are lined with osteoblasts (hematoxylin and eosin, original magnification \times 25).



Figure 6-8 Osteopenia. Fine vertical striations in the vertebral bodies secondary to osteopenia mimic hemangioma.

particularly dorsally, and the vertebral endplates are either indistinct or markedly sclerotic ("picture frame" appearance) (Fig. 6-9). If vertical striations are present, they are not so neatly arranged as in hemangioma (77). *Myeloma* and *metastasis* may exhibit lytic changes of the vertebral body, but there is no evidence of vertical striation. Conversely, hemangioma may present as a purely lytic lesion (Fig. 6-10). *Langerhans cell histiocytosis* usually presents as a collapsed vertebra (vertebra plana) (see Fig. 5-4), whereas hemangiomas of the vertebra exhibit pathologic fractures only in exceptional cases.

Pathology

On microscopic examination, **intraosseous hemangioma** cannot be distinguished from *skeletal angiomatosis* (58). Differentiation is therefore based on the extensive nature of the latter. Moreover, the presence of GLUT1 and/or WT1 immunoreactivity militates against a vascular malformation. The differential diagnosis from *epithelioid hemangioma, epithelioid hemangioendothelioma,* and *angiosarcoma* should not be too difficult because flat, normal-appearing endothelial cells usually line the vascular spaces of hemangioma (162).

Epithelioid hemangioma must be differentiated from epithelioid hemangioendothelioma because of the



Figure 6-9 Paget disease. Vertebral body of L2 is enlarged, and the vertebral endplates are thickened, creating a "picture frame" appearance.

different treatment modalities required (curettage or local excision vs. wide resection, eventually followed by chemotherapy). Epithelioid hemangioma consists of well-formed vessels, grows in a lobulated fashion, and contains inflammatory cells, including eosinophilic granulocytes and a nonhyalinized stroma. Epithelioid hemangioendothelioma presents with a peculiar myxoid- to chondroid-appearing stroma and an infiltrative growth pattern of moderate atypical cells growing in strands or nests rather than forming vascular channels.

The radiologic and pathologic differential diagnosis of intraosseous hemangioma is depicted in Figure 6-11.

Synovial Hemangioma

Synovial hemangioma is a rare benign lesion of the synovium that most commonly affects the knee articulation. This entity is discussed in Chapter 9.

Cystic Angiomatosis

Angiomatosis is defined as a diffuse involvement of bones by hemangiomatous lesions (14,29,40,41,46, 61,73,82), although some investigators also include cases of lymphangiomatosis (29,31,86). According to Enjolras and Mulliken (28), this lesion is a vascular malformation that, owing to its basic structural components, can be subclassified as either capillary, lymphatic, venous, arterial (rarely), or combined. In addition, further separation into slow-flow or fast-flow types appears to be clinically useful (44). The radiographic presentation of angiomatosis is that of lytic lesions, often with a honeycomb or lattice work ("hole-within-hole") appearance



Figure 6-10 Vertebral hemangioma. Lateral tomography shows a purely lytic hemangioma of the posterior part of vertebral body T6 (*arrows*). This presentation may be mistaken for metastasis or myeloma.



(13). When bone is extensively involved, the term cystic angiomatosis is applied (65,85). Some other terms used for this condition include diffuse skeletal hemangiomatosis (75,104), cystic lymphangiectasia (49), and hamartous hemolymphangiomatosis (91). This is a rare bone disorder characterized by diffuse cystic lesions of bone, frequently (60% to 70% of cases) associated with visceral involvement (52). Schajowicz (90) postulated that diffuse hemangiomatosis should be distinguished from cystic angiomatosis because of their different radio-logic and macroscopic aspects. On histologic examination, cystic angiomatosis is characterized by cavernous angiomatous spaces, indistinguishable from benign hemangioma of bone.

Another condition that must be distinguished is socalled Gorham disease of bone (18,39), also known by the names massive osteolysis (100), disappearing bone disease, and phantom bone disease (18,38). This condition is characterized by progressive, localized bone resorption (53), probably caused by multiple or diffuse cavernous hemangiomas or lymphangiomas of bone or by a combination of both (90). The radiographic presentation of Gorham disease consists of radiolucent areas in the cancellous bone or concentric destruction of the cortex, giving rise to a sucked-candy appearance (31). Eventually, the entire medullary cavity and the cortex are destroyed (4,103,111). On histologic examination, a marked increase is observed in intraosseous capillaries, which form an anastomosing network of endothelium-lined channels that are usually filled with erythrocytes or serum (90). Although some investigators claim that there is no evidence of osteoclasts in areas of bone resorption (38), recent studies suggest that osteoclastic activity plays a role in pathogenesis of Gorham disease (94).

Clinical Presentation

Patients with cystic angiomatosis usually present in the first three decades (79). There is 2:1 male-to-female predominance. The bones affected are most often those of the axial skeleton, as well as the femur, humerus, tibia, radius, and fibula (20,29,82) (Fig. 6-12). The bonerelated symptoms are usually secondary to pathologic fractures through the cystic lesions. Most of the symptoms, however, are related to visceral involvement.

Imaging

On radiography, the osseous lesions are usually osteolytic (Fig. 6-13), occasionally with a honeycomb appearance (Fig. 6-14). They are well defined and surrounded by a rim of sclerosis, and they vary in size (15,67) (Fig. 6-15). Although medullary involvement predominates, cortical invasion, osseous expansion, and periosteal reaction can occur (20). Rarely, sclerotic lesions may be present, and in these instances the condition may mimic osteoblastic metastases (51). On MRI the lesions usually show an intermediate signal intensity on T1-weighted images, and T2-weighted images with fat saturation show a mixture of high, intermediate, and low signal intensities (8).



Figure 6-12 Cystic angiomatosis: skeletal sites of predilection, peak age range, and male-to-female ratio.



Figure 6-13 Cystic angiomatosis. Several osteolytic lesions affect the shafts of the radius and ulna in this 25-year-old man.

Histopathology

Whether cystic angiomatosis is truly of blood vessel origin or of lymphatic derivation, or even represents a mixture of the two, is difficult to decide in a given case without IHC (50). On gross examination, the lesions of cystic angiomatosis resemble simple bone cysts (20). They consist of large cavities lined by a dull yellowishgray membrane. Multiple communicating cysts, separated by thickened trabeculae of mature lamellar bone but without osteoblastic rimming (107), may also be present, producing a honeycomb appearance (87). In some cases osteoid and immature woven bone formation with rimming of active osteoblasts was reported (51). Histologically, the lesions are indistinguishable from those observed in capillary or cavernous hemangiomas (20). They contain multiple dilated, thin-walled vascular channels lined by flat endothelial cells (107).

Differential Diagnosis

Radiology

The predominant differential diagnosis must include *polyostotic fibrous dysplasia, enchondromatosis, Langerhans cell histiocytosis, brown tumors of hyperparathyroidism,* and *pseudotumors of hemophilia* (95). In older patients, although the diagnosis of *metastatic disease* (42) and *multiple myeloma* must be considered, these conditions usually can be easily distinguished on the basis of both radiologic and clinical examinations. The osteosclerotic variant of cystic angiomatosis must be differentiated from *osteoblastic metastases* (1,17,51), *sclerosing variant of myeloma* (104), *lymphoma,* and *mastocytosis* (6). Careful interpretation of the radiographs may reveal the presence of mixed sclerotic and lytic lesions in cystic angiomatosis, a feature that is helpful in excluding some of the mentioned entities.

Pathology

Microscopically, the differential diagnosis of cystic angiomatosis should include *massive osteolysis (Gorham disease)*. Because both lesions are pathologically almost identical, the diagnosis is based on clinical findings (31).

The radiologic and pathologic differential diagnosis of cystic angiomatosis is depicted in Figure 6-16.

Lymphangioma and Lymphangiomatosis

Lymphangioma is a rare benign bone disorder, first described in 1947 by Bickel and Broders (10). The lesion is composed of sequestered, noncommunicating lymphoid tissue lined by lymphatic endothelium (29). The cause of this lesion is believed to be a congenital obstruction of lymphatic drainage. Lymphangiomas, similar to hemangiomas, are classified and subclassified, according to the size of the vessels, as capillary, cavernous, cystic, or mixed (22). The cystic variety is most common.

Clinical Presentation

There is no sex predilection and no hereditary tendency (69). The vast majority of these disorders are soft tissue lesions, with osseous involvement being rare (27,55). The most common sites of involvement are the diaphyses or metaphyses of the tibia and the humerus, as well as ilium, skull, mandible, and vertebrae (45,96). These lesions are usually discovered at birth (50% to 65% of cases) or at least within the first 2 years of life (90%) (11,79). The lesion may be solitary or multifocal (88). When the lesions are multiple, the term lymphangiomatosis is applied (96,109).

Imaging

On radiography, these lesions are indistinguishable from hemangiomas and hemangiomatosis (see previous



Figure 6-14 Cystic angiomatosis. A 28-yearold man with honeycomb pattern seen in the right ilium and both pubic bones.



Figure 6-15 Cystic angiomatosis. Several confluent lesions with peripheral sclerosis and cortical thickening are present in the right femur of a 20-year-old man.

discussion), exhibiting a multiple or multilocular expansive osteolytic appearance (3,23). In some cases the margin may have a "soap-bubble" appearance, whereas in others it may be quite poorly defined, with a moth-eaten pattern of bone destruction (92) (Fig. 6-17).

A CT scan can be useful in demonstrating the relative sparing of the medullary cavity in preference to the cortex (45,69). MRI demonstrates bone lacunae displaying a hypointense signal on T1-weighted and a hyperintense signal on T2-weighted images, which indicates the presence of a liquid-filled lesion. There is no enhancement after gadolinium injection (69). Lymphography is very effective for demonstration of abnormal and dilated lymphatic channels filling the cystic bone lesions (84).

Histopathology

The histopathology of lymphangioma is essentially the same as that of hemangioma. The most common is the cystic capillary form. Endothelia in lymphangioma are positive for D2-40 by IHC.

Differential Diagnosis

Radiology

Based on the radiographic appearance, the differential diagnosis must include *lymphoma*, *plasmacytoma*, and *fibrosarcoma* when the lesion is solitary, and *metastatic neuroblastoma*, *hemangiomatosis*, *Langerhans cell histiocytosis*, *polyostotic fibrous dysplasia*, *congenital fibromatosis*, and *Gaucher disease* when there is polyostotic involvement.

Pathology

Because the wall of a lymphatic vessel is structurally identical in histology to that of a blood capillary (see previous discussion), most investigators base their differential diagnosis on the presence or absence of erythrocytes in the lumen. Nevertheless, it is possible that an erythrocyte may be found in a lumen of a lymphatic



Figure 6-17 Skeletal lymphangiomatosis. A 2-year-old boy presented with multiple bone lesions, particularly affecting the pelvis and proximal femora. Note honeycomb and moth-eaten pattern of bone destruction. In addition to skeletal involvement, lymphangiomas were also present in the soft tissues.



vessel because of artificial translocation during the histologic preparation of the tissue. Therefore, this finding does not constitute an absolute proof of whether the vessel is lymphatic or not. By using IHC it is usually possible to discriminate between hemangiomas (positive for CD34, CD31, and FVIII, negative for D2-40) and lymphangiomas (positive only for D2-40). Hence, the differential diagnosis is the same as for hemangioma (see previous discussion).

Glomus Tumor, Glomangioma, and Glomangiomyoma

Glomus tumor and its variants, glomangioma and glomangiomyoma, are extremely rare benign lesions of bone composed of rounded uniform cells often arranged in a brick-work-like manner. They are intimately associated with vascular structures (29). Of the three, the glomus tumor is most common, whereas the other two are very rare. The occurrence of a glomus tumor primarily arising in the bone (only about 100 published cases), rather than originating in the adjacent soft tissues and eroding bone secondarily, is extremely rare (50). At one time this tumor was considered to derive from the previously mentioned neuromyoarterial glomus, a neurovascular structure. At present, however, it is believed that the tumor arises from modified smooth muscle cells. Outside of bone, glomus tumors with cellular atypia (so-called symplastic glomus tumors) and malignant glomus tumors have been described, both of which are exceedingly rare (116).

Recently, *glomulin*, a gene located on chromosome 1p21-22 and possibly involved in differentiation of vascular smooth muscle cells, has been shown to be mutated in familial glomuvenous malformations (glomangiomas) (113,114). Genetic alterations in sporadic glomus tumors have not been published to date (115).

Clinical Presentation

The vast majority of intraosseous glomus tumors occur at the fingertips (117), arising in the distal phalanges (112). Very rarely they affect the coccyx. Patients are adults in the third to fifth decade, with a female predilection. The main clinical symptoms include localized pain and exquisite point tenderness (120). Exposure to cold or minimal trauma may induce severe paroxysmal attacks of pain (86).

Imaging

On radiography, glomus tumor presents as a small, lytic bony defect with sharply marginated edges, usually located at the dorsal aspect of the distal phalanx. If the lesion arises primarily in the soft tissue and erodes bone secondarily, an adjacent soft tissue mass is present (79). On CT examination, a nonspecific subungual mass is demonstrated, with an attenuation value similar to that of other soft tissue lesions. On MR T1-weighted images the lesion exhibits either isointense or hyperintense signal, and strong enhancement after injection of gadolinium (121). On T2-weighted images, glomus tumor exhibits a homogeneous high signal intensity (79,119).

Histopathology

Histologically, the lesion is composed of lobules, strands, and broad sheets of rounded, uniform glomus cells, often arranged in the above-mentioned brick-work–like manner, with indistinct capillaries in the walls of the surrounding large blood vessels (26,29). Because each cell is encircled by a basal lamina, cell margins are clearly delineated by reticulin stains, toluidine blue, or periodic aid–Schiff (PAS) preparations. In glomangioma, the larger blood vessels are more conspicuous, with the tumor cells arranged in smaller areas within their walls. In glomangiomyoma, spindle cells resembling smooth muscle cells are found in variable numbers (31). By ICH, glomus tumor and its variants express smooth muscle actin and collagen type IV but usually no desmin or CD34 (106).

Differential Diagnosis

Radiology

On radiography the lesion closely resembles an *enchondroma* (118). Other lesions that should be included in the differential diagnosis are *epidermal inclusion cyst, metastasis* (particularly from a bronchogenic carcinoma), *aneurysmal bone cyst, sarcoidosis*, and *tuberous sclerosis* (112).

Pathology

The differentiation of this tumor from other entities is usually easy because of its conspicuous clinical behavior. Histologically, a *metastasis* from a small cell epithelial or mesenchymal tumor must be taken into account, but can be ruled out by appropriate IHC investigations (negativity for cytokeratins, desmin and myogenin, CD45 and CD99). The very regular arrangement of the glomus tumor cells and their connection with the walls of larger vessels are helpful in making the distinction.

The radiologic and pathologic differential diagnosis of glomus tumor is shown in Figure 6-18.

Malignant Tumors

The existing nomenclature for malignant vascular tumors is not uniform and therefore can be confusing (184). Different terms, including hemangiosarcoma (angiosarcoma), hemangioendothelioma, and hemangioendothelial sarcoma, have been used as synonyms. Moreover, the tumors have been classified into different grades, from grade I hemangioendothelioma (well-differentiated) to grade III hemangiosarcoma (poorly differentiated) (90). In contrast to the classification of their soft tissue counterparts (180,183), all aggressive and malignant vascular lesions in bone have recently been grouped by the World Health Organization (WHO) under the heading of angiosarcoma (164). However, because histologic and clinical differences exist, hemangioendothelioma and angiosarcoma of bone are discussed separately (30,162).





Epithelioid Hemangioendothelioma

Epithelioid hemangioendothelioma is regarded as a neoplasm with an unpredictable clinical course intermediate between that of hemangioma and conventional angiosarcoma (90,141,142,145,155,166,170,177,178). Tumor involving the soft tissues, bones, or other sites (liver, lung) share an identical histomorphology. However, angiocentricity or intravascular spread, as seen in soft tissues or in liver and lung, is not described in bone (106,162). Cytogenetic analysis of two tumors occurring in the soft tissues revealed a nonrandom translocation t(1;3) (p36;q25) involving chromosomes 1 and 3 (157). In bone, more than 50% of patients develop multiple lesions, either synchronously or metachronously, making the discrimination from metastatic lesions difficult if not impossible (148). Characteristic of this tumor is the formation of irregular nests or anastomosing cords of enlarged and vacuolated endothelial cells that are embedded in a myxoid substrate (25). In bone, other subtypes, such as kaposiform or retiform hemangioendotheliomas, are described only as single cases (123,153). Hemangioendotheliomas constitute less than 1% of all the primary malignant tumors of bone (179).

Clinical Presentation

Epithelioid hemangioendothelioma can occur at any age within a range of 10 to 75 years (122). The lesion may be solitary or multicentric (127,129). There is a slightly more common predilection for males in the second and third decades (79). Patients affected by multifocal disease are, on average, 10 years younger than those with a solitary lesion. Osseous epithelioid hemangioendotheliomas most commonly affect the calvaria, spine, and bones of the lower extremities. They may occur in any part of a bone, and it is not unusual to see multiple lesions in contiguous anatomic sites (e.g., in several carpal or tarsal bones).

Epithelioid hemangioendothelioma usually causes dull local pain and tenderness (169); some swelling may also be observed, and hemorrhagic joint effusion sometimes occurs (90). A vertebral location for this tumor is characterized by radiculopathy. Symptoms are quite variable in their duration, ranging from a few weeks to many years (average 5 months) (144). A soft tissue mass or a pathologic fracture is only rarely the presenting symptom (143).

Imaging

On radiography, epithelioid hemangioendothelioma exhibits an osteolytic appearance; it may be well circumscribed or may have a wide zone of transition (Fig. 6-19A). There may be variable degrees of peripheral sclerosis, sharply demarcating the lesion. Frequently a soap-bubble appearance with expansion of bone is observed, with occasional extension into the soft tissues (31). However, unless there is an associated pathologic fracture, a soft tissue mass is usually not present. Scintigraphy shows an increased uptake of the radiopharmaceutical tracer (125,147,149). On MRI the tumor shows a mixed signal on T1-weighted images with moderate increase in the signal intensity on T2 weighting (165,172) (Fig. 6-19B, C).

Histopathology

On histologic examination, epithelioid hemangioendothelioma reveals endothelial cells with abundant faintly eosinophilic or amphophilic cytoplasm (136, 174) (Fig. 6-20A). Individual cells may exhibit intracytoplasmic lumina of variable size, whose diameters often approximate that of an entrapped erythrocyte (Fig. 6-20B). Larger lumina appear to be formed as the result of fusion of their smaller intracytoplasmic counterparts. A residual strand of attenuated cytoplasm frequently bridges the opening. Although mitotic figures may be absent, one or two figures per 10 high-power fields are





Figure 6-19 Epithelioid hemangioendothelioma: magnetic resonance imaging (MRI). A: Anteroposterior radiograph of the proximal tibia shows a radiolucent destructive lesion affecting mainly the medial aspect of the bone. The cortex is destroyed. **B:** Coronal T1-weighted MRI shows a low-signalintensity tumor replacing bone marrow. Small foci of high signal represent hemorrhagic areas. **C:** Coronal T2-weighted MRI demonstrates an increase in the signal intensity of the tumor, which exhibits heterogeneous appearance. (Reprinted with permission from Greenfield GB, Arrington JA. *Imaging of bone tumors*. Philadelphia: JB Lippincott, 1995:161.)



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Figure 6-20 Histopathology of epithelioid hemangioendothelioma. A: A pale-staining myxoid stroma is filled with single or cord-like proliferating plump endothelial cells that exhibit lack of obvious pleomorphism or significant mitotic activity (hematoxylin and eosin, original magnification $\times 200$). **B:** Eosinophilic tumor cells contain intracytoplasmic vacuoles (hematoxylin and eosin, original magnification $\times 400$). **C:** Observe intracytoplasmic vacuoles and membranous CD34 immunoreaction on cell surface (biotin-streptavidin peroxidase, original magnification $\times 400$). **D:** Also positive to a varying degree are other endothelial markers such as FVIII-related antigen (biotin-streptavidin peroxidase, original magnification $\times 200$).

usually seen. The tumor cells are arranged in nests and cords. Primitive vascular channels are occasionally formed but are not a prominent feature. Small foci of hemorrhage or necrosis may be observed (182). The stroma typically varies from fibrous to myxoid, and may appear more hyalinized, thus resembling a hyaline cartilage–like matrix (31). That the cells composing these tumors are differentiated is evident from their frequent positive immunohistochemical staining for Factor VIII and for Fli-1, CD34, and CD31 (Fig. 6-20C, D).

Weiss et al. emphasized four major patterns, often characterized by transitional zones. These include (a) plump, polygonal, or elongated cells possessing large vacuoles that resemble signet ring cells; (b) aggregates of the above cells arranged into elongated cords or clusters; (c) formation of vascular lumina, which may be interconnected and may also contain erythrocytes; and (d) solid or spindle cell areas that sometimes contain nests of cells (134,182).

In bone, atypical histologic features such as mitotic rate, degree of atypia, presence of necrosis, and spindling of tumor cells, considered as prognostic indicators in soft tissue lesions (180), are not of value in predicting the clinical course (148).

Differential Diagnosis

Radiology

On radiologic studies it is very difficult to differentiate epithelioid hemangioendothelioma from other vascular lesions, both benign and malignant. A solitary osteolytic lesion may mimic a *metastasis*, *fibrosarcoma*,

malignant fibrous histiocytoma (MFH), plasmacytoma, or lymphoma. Lesions with a sclerotic border may resemble a Brodie abscess, intraosseous ganglion, enchondroma, or chondroblastoma, and those extending to the articular end of bone can be mistaken for a giant cell tumor. A multicentric presentation should be differentiated from other polyostotic conditions, such as brown tumors of hyperparathyroidism, pseudotumors of hemophilia, metastatic disease, and multiple myeloma. When lytic lesions are seen predominantly in the cortex, metastatic disease is a strong consideration (42,140). Because the radiologic presentation of hemangioendothelioma is usually nonspecific, clinical information may be helpful in narrowing the differential diagnosis.

Pathology

The differential diagnosis of epithelioid hemangioendothelioma includes metastatic adenocarcinoma, adamantinoma of the long bones, angiosarcoma ("malignant hemangioendothelioma"), myxoid chondrosarcoma, telangiectatic osteosarcoma, and melanoma. The differentiation between hemangioendothelioma and angiosarcoma is in many cases arbitrary. A clear distinction is not possible because areas with the characteristics of the first tumor may merge into areas with the characteristics of the latter (see later). However, tumor cells are more uniform and less atypical in epithelioid hemangioendothelioma than in angiosarcoma, and vascular channels tapered by overt atypical cells are much more prominent in the latter. Intraluminal budding and increased mitotic rates are also in favor of angiosarcoma. To exclude metastatic carcinoma or melanoma, IHC studies are helpful, revealing positivity for endothelial markers such as CD31, CD34, Fli-1, or factor VIII. However, cytokeratins may also be positive in epithelioid hemangioendotheliomas and epithelioid angiosarcomas, and CD31 has been reported positive in some carcinomas; therefore, a panel of antibodies must be applied (106,132,168). Unlike vascular tumors, melanomas are positive for melan A and S-100 [however, weak staining in some tumor cells in single cases of angiosarcoma has been reported (156)], but are negative for Fli-1 and CD31, whereas CD34 may also be positive in single melanomas (32,128,185).

Telangiectatic osteosarcoma may have areas that strongly resemble hemangioendothelioma. However, the presence of osteoid formation in the former is diagnostic.

Myxoid chondrosarcoma may exhibit the nodular arrangement and epithelioid appearance of the tumor cells (126,138). However, the myxoid background and the positive staining for S-100 protein are not features of hemangioendothelioma.

Adamantinoma exhibits a characteristic biphasic pattern consisting of epithelial strands intimately admixed with a fibrous stroma. In this tumor, however, the epithelial component is usually scant and is often difficult to discern, in contrast to hemangioendothelioma, in which the rather large epithelioid cells are conspicuous in size and number. Moreover, epithelial cells in adamantinoma do not react with CD34 or CD31.

Angiosarcoma

Angiosarcoma of bone represents the most malignant end of the spectrum of vascular tumors. This is an aggressive malignancy, characterized by frequent local recurrence and distant metastases (79,130,133).

Clinical Presentation

Angiosarcomas most commonly involve the skin (33%) and soft tissues (24%), with bone being involved in only 6% of cases (135). These lesions occur during the second to the seventh decade, although the peak is during the third to fifth decade (20). Males are affected twice as frequently as females. The most common osseous locations are the long bones (approximately 60% of cases), particularly the tibia (23%), femur (18%), and humerus (13%), as well as pelvis (7%) (79,86). The two most common symptoms are local pain and swelling. Metastases to the lungs and other parenchymal organs are found in about 66% of cases (130).

Angiosarcomas associated with chronic lymphedema particularly in the extremities are known as Stewart-Treves syndrome (160,173).

Imaging

On radiologic studies it is impossible to distinguish among hemangioendothelioma, hemangiopericytoma, and angiosarcoma. Angiosarcomas have radiographic features similar to those of hemangioendotheliomas. However, they are usually solitary lesions and almost invariably there is a wide transitional zone between tumor and uninvolved bone. Soft tissue masses are common. Angiosarcomas are predominantly lytic, and sometimes they exhibit a honeycomb or hole-withinhole pattern, similar to the appearance of hemangioma. More aggressive features include osseous expansion, cortical permeation, and an associated soft tissue mass (79) (Fig. 6-21). Multifocal involvement is common. Skeletal scintigraphy is invariably positive, revealing increased tracer uptake (125). The CT and MRI appearances of these lesions are decidedly nonspecific (20).

Histopathology

Histologically, angiosarcomas correspond to their soft tissue counterparts. They are composed of poorly formed blood vessels that exhibit complicated infolding and irregular anastomoses. The endothelial cells lining these blood vessels display features of frank malignancy, with the presence of often plump intraluminal cells resembling hobnails, frequent and atypical mitoses, and nuclear hyperchromatism (Fig. 6-22). Solid areas of the tumor may contain spindle cells and epithelioid cells. Spontaneous necrosis is often present, as well as sometimes extensive hemorrhage (164,183). According to Stout (174), two fundamental histologic criteria are necessary for diagnosing an angiosarcoma: (a) formation of greater number of atypical endothelial cells than would be necessary to line vessels with a simple endothelial membrane and (b) formation of vascular tubes and channels that exhibit a framework of reticulin fibers and that usually anastomose.

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Figure 6-22 Histopathology of angiosarcoma. A: Clusters of very large polygonal cells with large pleomorphic and hyperchromatic nuclei border blood-filled vascular spaces. The solid areas of the tumor resemble large cell metastatic cancer (hematoxylin and eosin, original magnification \times 50). **B:** In another field of view the vascular spaces are larger and more visible. The vascular character of the tumor is obvious (hematoxylin and eosin, original magnification \times 50). **C:** Reticulin fiber stain shows that the tumor cells lie corona-like within the rims of reticulin fibers (*black*). Note newly formed cortex made up of trabecular woven bone (Novotny, original magnification \times 25). **D:** At higher magnification the interconnected network of vascular spaces each bound by a rim of reticulin fibers (*black*) is seen, surrounding a seam of large tumor cells (Novotny, original magnification \times 50) (*continued*).



Figure 6.22 Continued **E:** Large epithelioid-appearing tumor cells line vascular spaces and protrude into the lumen (hematoxylin and eosin, original magnification \times 630). **F:** Tumor cells lining vascular spaces and infiltrating the surrounding stroma are positive for CD31. Note luminar shedding of tumor cells (biotin-streptavidin peroxidase, original magnification \times 200).

Some angiosarcomas present as epithelioid tumors, consisting predominantly or almost exclusively of enlarged eosinophilic epithelioid cells with prominent nucleoli. These clearly atypical cells line irregularly formed vascular channels and exhibit increased mitotic activity. Intraluminal papillary projections and diffuse infiltrating sheets of tumor cells may also be present (133) (Fig. 6-22E). Because antibodies to factor VIII yield inconsistent and sometimes negative results, reflecting poor tumor differentiation, additional endothelial markers (CD34 and CD31) must therefore be applied (Fig. 6-22F). Furthermore, angiosarcomas react with antibodies against the transcription factors WT1 and Fli-1 and against D2-40, directed toward the lymphatic endothelium (32,36,56,176a).

Differential Diagnosis

Radiology

Because the radiologic characteristics of angiosarcoma are not specific, several possibilities should be considered in the differential diagnosis, including *metastasis*, *plasmacytoma*, *lymphoma*, *fibrosarcoma*, and *MFH*. As always in such situations, the radiologist should rely on clinical information.

Pathology

As in other vascular tumors, differential diagnosis must include *hemangioendothelioma* and *hemangiopericytoma*. The myxoid- to cartilage-like matrix of epithelioid hemangioendothelioma is rarely found in angiosarcoma, which almost always presents a higher grade of nuclear atypia and pleomorphism. Because it is usually difficult to determine the tumor's vascular pattern, staining for reticulin fibers should always be utilized to identify the basal membrane in the vessel wall (77). In angiosarcoma, the tumor cells lie within the basal membrane whereas in hemangiopericytoma they lie outside. Some poorly differentiated but extremely vascular sarcomas may strongly resemble angiosarcoma. *Telangiectatic osteosarcoma* and *metastases from renal cell carcinoma* may contain areas that strongly resemble a vascular tumor. In such cases, studies with a panel of vascular markers (CD34, CD31, Fli-1) may be helpful for demonstration of the endothelial nature of tumor cells in the solid regions of angiosarcoma (32,77). Especially in epithelioid angiosarcomas, immunoreactivity for cytokeratins may be present, giving the erroneous impression of a metastatic carcinoma if only antibodies to cytokeratins are applied (133).

Hemangiopericytoma

With the emerging concept of solitary fibrous tumor, the existence of hemangiopericytoma as a single tumor entity has been seriously questioned, at least in the soft tissues (137). Hemangiopericytoma is regarded as a histologic pattern that is also present in a variety of other well-defined sarcomas and even in carcinomas (161). However, even with this approach some tumors are still classified as hemangiopericytomas because they cannot be identified as solitary fibrous tumor or other entities even with application of ancillary techniques (139). Furthermore, according to Miettinen et al. (159), genetic differences between solitary fibrous tumor and hemangiopericytoma appear to be revealed by comparative genomic hybridization (CGH) studies, which showed chromosomal gains and losses in solitary fibrous tumors but not in hemangiopericytomas (159). In addition, balanced translocations have been observed in hemangiopericytomas, and a trisomy 21 has been noted in an extrapleural solitary fibrous tumor (131,154).

Initially, hemangiopericytoma was believed to arise from the Zimmerman cells that are located around vessels (171,175). However, electron microscopic and IHC studies have demonstrated no convincing evidence for this assumption (106). This is a tumor of intermediate aggressiveness that has both benign and malignant potential (79,163).

Clinical Presentation

Hemangiopericytoma has been reported in patients ranging from 15 to 48 years of age (135,181). The tumor commonly affects the lower extremities (35%), particularly the soft tissues of the thigh and pelvis. Primary osseous lesions are rare, accounting for less than 1% of vascular tumors (150). Symptoms consist primarily of pain or swelling, or a combination of both, with a duration of 1 month to 1 year (31,152). Occasionally there is associated hypophosphatemic osteomalacia (151).

Imaging

The radiographic features of hemangiopericytoma are nonspecific. Approximately 70% of these tumors are purely lytic (Fig. 6-23), whereas various degrees of sclerosis are present in about 30% of cases (186). When sclerosis predominates, bone trabeculae become prominent, giving the lesion a honeycomb appearance similar to that of a hemangioma (20). Although the cortex may be expanded, a periosteal reaction is rare. Wold et al. (186) reported a correlation between the histologic malignancy and the radiographic aggressiveness of this tumor, although they found it very difficult, if not impossible, to distinguish benign from malignant lesions on the basis of radiographic appearance alone. Thus far, CT and MRI have also failed to facilitate this distinction (20,124), although Juan et al. (146) reported recently a "spokewheel" appearance of hemangiopericytoma on MR images that may suggest this diagnosis. Angiography does not provide additional useful information (187,188).

Histopathology

Histologically, hemangiopericytomas are composed of vascular channels lined at their inner side by normal endothelial cells not belonging to the tumor and surrounded by round to oval cells individually enclosed by a basket of reticulum fibrils (Fig. 6-24A). The perivascular pattern of these tumors can be best recognized in reticulin preparations (167). Silver reticulin staining of the basal membrane of the vessel reveals that the tumor cells lie outside this membrane (101) (Fig. 6-24B). The proliferation of tumor cells outside the vascular spaces distinguishes hemangiopericytoma from endothelial lesions in which the cells line the inner surface of the basal membrane of the vascular spaces (176). The tumor may be of low-grade malignancy or of a high grade, with anaplasia and multiple mitoses (144). Tumor cells are positive for CD34 and vimentin, and negative for CD31 and (with rare exceptions) for actins and desmin (158).

Differential Diagnosis

Radiology

Like hemangioendothelioma, hemangiopericytoma is also difficult to distinguish from other vascular lesions. If the lesions exhibit osteolytic features, the same differential diagnoses apply as for hemangioendothelioma. An expansive solitary lesion may mimic *aneurysmal bone cysts*, and lesions that extend to the articular end of bone should be differentiated from *giant cell tumor*. Sclerotic lesions may mimic *sclerosing hemangiomas* and even *osteoblastic metastases*.

Pathology

Hemangiopericytoma can be microscopically confused with a monophasic type of *synovial sarcoma*, when the latter exhibits a prominent hemangiopericytomatous



Figure 6-23 Hemangiopericytoma. Anteroposterior radiograph of the left humerus in a 30-year-old man shows an osteolytic lesion with wide zone of transition. (From Bullough PG. *Atlas of orthopedic pathology,* 2nd ed. New York: Gower, 1992:17.10.)



Α

Figure 6-24 Histopathology of hemangiopericytoma. A: Solid tumor tissue containing interconnected small blood vessel lumina are surrounded by polygonal cells of medium size. The blood vessels show a seam of flat endothelium different from the other cells (hematoxylin and eosin, original magnification \times 50). **B:** With reticulin fiber staining, the numerous capillary lumina containing a seam of normal endothelia and being surrounded by a sharp narrow band corresponding to the basal membrane become distinct. The tumor cells lie outside the basal membrane (Novotny, original magnification \times 25).

histologic growth pattern (31). However, the typical features of biphasic synovial sarcoma include glandular and pseudoglandular patterns, and the lesion actually arises in soft tissues with only secondary involvement of bone, both of which features are helpful in the differentiation (144). Moreover, synovial sarcomas (biphasic and monophasic) are immunohistochemically positive for epithelial membrane antigen (EMA), CD99, and BCL2.

Hemangiopericytoma may also mimic *glomus tumor*, but its cells are less differentiated and exhibit scant

cytoplasm and a poorly delineated border. In contrast to hemangiopericytoma, glomus tumors are positive for muscle markers (actins, desmin).

Intraosseous myofibroma and myofibromatosis must also be ruled out. These lesions occur particularly in children and have a nodular appearance. Histologically, a central highly vascularized component with plump spindle cells can be observed, which is accompanied by peripheral bundles of spindle-shaped myofibroblasts. Myofibromas are positive for actin and negative for desmin (106).





The radiologic and pathologic differential diagnoses of epithelioid hemangioendothelioma, hemangiopericytoma, and angiosarcoma are depicted in Figure 6-25.

REFERENCES

Intraosseous Hemangioma, Cystic Angiomatosis, Lymphangioma, and Lymphangiomatosis

- 1. Abdelwahab IF. Sclerosing hemangiomatosis: a case report and review of the literature. *Br J Radiol* 1991;64:894–897.
- Adler CP, Wold L. Haemangioma and related lesions. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:159–160.
- Asch MJ, Cohen AH, Moore TC. Hepatic and splenic lymphangiomatosis with skeletal involvement. Report of a case and review of the literature. *Surgery* 1974;76:334–339.
- Assoun J, Richardi G, Railhac JJ, et al. CT and MRI of massive osteolysis of Gorham. J Comput Assist Tomogr 1994;18:981–984.
- Baker ND, Greenspan A, Neuwirth M. Symptomatic vertebral hemangiomas: a report of four cases. *Skeletal Radiol* 1986;15: 458–463.
- Barer M, Peterson LFA, Dahlin DC, et al. Mastocytosis with osseous lesions resembling metastatic malignant lesions in bone. *J Bone Joint Surg Am* 1968;50:142–152.
- Baudrez V, Galant C, Vande Berg BC. Benign vertebral hemangioma: MR-histological correlation. *Skeletal Radiol* 2001;30: 442–446.
- Bergman AG, Rogero GW, Hellman B, Lones MA. Case report 841. Skeletal cystic angiomatosis. *Skeletal Radiol* 1994;23:303– 305.
- Bergstrand A, Hook O, Lidvall H. Vertebral haemangiomas compressing the spinal cord. Acta Neurol Scand 1963;39:59–66.
- Bickel WH, Broders AC. Primary lymphangioma of the ilium: report of a case. J Bone Joint Surg 1947;29:517–522.
- 11. Bill AH, Sumner DS. A unified concept of lymphangioma and cystic hygroma. Surg Gynecol Obstet 1965;120:79-86.
- Boye E, Yu Y, Paranya G, et al. Clonality and altered behavior of endothelial cells from hemangiomas. J Clin Invest 2001;107: 745–752.
- Boyle WJ. Cystic angiomatosis of bone. J Bone Joint Surg Br 1972; 54:626–636.
- Brower AC, Culver JE Jr, Keats TE. Diffuse cystic angiomatosis of bone. Report of 2 cases. Am J Roentgenol Radium Ther Nucl Med 1973;118:456–463.
- Brower AC, Culver JE, Keats TE. Diffuse cystic angiomatosis of bone: report of two cases. Am J Roentgenol 1973;118:456–463.
- Buetow PC, Kransdorf MJ, Moser RP Jr, et al. Radiologic appearance of intramuscular hemangioma with emphasis on MR imaging. *Am J Roentgenol* 1990;154:563–567.
- Bullough PG. Atlas of orthopedic pathology with clinical and radiologic correlation, 2nd ed. New York: Gower, 1992:15.12–15.14.
- Choma ND, Biscotti CV, Bauer TW, et al. Gorham's syndrome: a case report and review of the literature. Am J Med 1987;83: 1151–1156.
- 19. Cohen MM Jr. Vasculogenesis, angiogenesis, hemangiomas, and vascular malformations. *Am J Med Genet* 2002;108:265–274.
- Conway WF, Hayes CW. Miscellaneous lesions of bone. Radiol Clin North Am 1993;31:339–358.
- Cooper PH. Is histiocytoid hemangioma a specific pathologic entity? *Am J Surg Pathol* 1988;12:815–817.
- Cutillo DP, Swayne LC, Cucco J, Dougan H. CT and MRI imaging in cystic abdominal lymphangiomatosis. J Comput Assist Tomogr 1989;13:534–536.

- Daoud S, Lefort G, Munzer M, et al. Lymphangiomatose osseuse disséminée, à propos de six observations. *Chir Pédiatr* 1988;29:342–348.
- Donnelly LF, Adams DM, Bisset GS III. Vascular malformations and hemangiomas: A practical approach in a multidisciplinary clinic. *Am J Roentgenol* 2000;174:597–608.
- Dorfman H, Czerniak B. *Bone tumors*. St. Louis: Mosby, 1998: 1261.
 Dorfman HD, Steiner GC, Jaffe HL. Vascular tumors of bone.
- Hum Pathol 1971;2:349–376. 27. Ellis GL, Brannon RB. Intraosseous lymphangiomas of the
- mandible. *Skeletal Radiol* 1980;5:253–256.
- Enjolras O, Mulliken JB. Vascular tumors and vascular malformations (new issues). Adv Dermatol 1997;13:375–423.
- Enzinger FM, Weiss SW. Benign tumors and tumorlike lesions of blood vessels. In: Enzinger FM, Weiss SW, eds. Soft tissue tumors, 3rd ed. St. Louis: CV Mosby, 1995.
- 30. Evans HL, Raymond AK, Áyala AG. Vascular tumors of bone: a study of 17 cases other than ordinary hemangioma, with an evaluation of the relationship of hemangioendothelioma of bone to epithelioid hemangioma, epithelioid hemangioendothelioma, and high-grade angiosarcoma. *Hum Pathol* 2003;34:680–689.
- 31. Fechner RE, Mills SE. *Tumors of the bones and joints.* Washington, DC: Armed Forces Institute of Pathology, 1993.
- Folpe AL, Chand EM, Goldblum JR, Weiss SW. Expression of Fli-1, a nuclear transcription factor, distinguishes vascular neoplasms from potential mimics. *Am J Surg Pathol* 2001;25:1061–1066.
- Folpe AL, Hill CE, Parham DM, et al. Immunohistochemical detection of FLI-1 protein expression: a study of 132 round cell tumors with emphasis on CD99-positive mimics of Ewing's sarcoma/primitive neuroectodermal tumor. *Am J Surg Pathol* 2000; 24:1657–1662.
- Friedman DP. Symptomatic vertebral hemangiomas: MR findings. Am J Roentgenol 1996;167:359–364.
- Fujimoto H, Ueda T, Masuda S, Nosaka K. Blood-pool scintigraphic diagnosis of fractured lumbar vertebral hemangioma. *Skeletal Radiol* 2001;30:223–225.
- Fukunaga M. Expression of D2-40 in lymphatic endothelium of normal tissues and in vascular tumours. *Histopathology* 2005;46: 396–402.
- 37. Gerard PS, Wilck E. Spinal hemangioma. An unusual photopenic presentation on bone scan. *Spine* 1992;17:607–610.
- Gorham LW, Stout AP. Massive osteolysis (acute spontaneous absorption of bone, phantom bone, disappearing bone): its relation to haemangiomatosis. *J Bone Joint Surg Am* 1955;37:985–1004.
- Gorham LW, Wright AW, Shultz HH, Mexon FC Jr. Disappearing bones: a rare form of massive osteolysis. *Am J Med* 1954; 17: 674–682.
- Graham DY, Gonzales J, Kothari SM. Diffuse skeletal angiomatosis. Skeletal Radiol 1978;3:131–135.
- Gramiak R, Ruiz G, Campeti FL. Cystic angiomatosis of bone. Radiology 1957;69:347–353.
- Greenspan A, Norman A. Osteolytic cortical destruction: an unusual pattern of skeletal metastases. *Skeletal Radiol* 1988; 17: 402–406.
- 43. Greenspan A, Klein MJ, Bennett AJ, et al. Case report 242. Hemangioma of the T6 vertebra with a compression fracture, extradural block and spinal cord compression. *Skeletal Radiol* 1978;10:183–188.
- 44. Grevelink SV, Mulliken JB. Vascular anomalies. In: Freedberg IM, Eisen AZ, Wolff K, Austen KF, Goldsmith LA, Katz SI, Fitzpatrick TB, eds. *Fitzpatrick's dermatology in general medicine*, 5th ed. New York: McGraw-Hill, 1999:1175–1194.
- Griffin GK, Tatu WF, Fischer LM, et al. Systemic lymphangiomatosis: a combined diagnostic approach of lymphangiography and computed tomography. *J Comput Assist Tomogr* 1986; 10:335–339.
- Gutierrez RM, Spjut HJ. Skeletal angiomatosis: report of three cases and review of the literature. *Clin Orthop* 1972;85:82–97.
- Han BK, Ryu J-S, Moon DH, et al. Bone SPECT imaging of vertebral hemangioma. Correlations with MR imaging and symptoms. *Clin Nucl Med* 1995;20:916–921.
- Hawnaur JM, Whitehouse RW, Jenkins JPR, Isherwood I. Musculoskeletal haemangiomas: comparison of MRI with CT. Skeletal Radiol 1990;19:251–258.
- 49. Hayes JT, Brody GL. Cystic lymphangiectasis of bone. A case

report. J Bone Joint Surg Am 1961;43:107-117.

- Huvos AG. Hemangioma, lymphangioma, angiomatosis/ lymphangiomatosis, glomus tumor. In: Huvos AG, ed. Bone tumors: diagnosis, treatment and prognosis, 2nd ed. Philadelphia: WB Saunders, 1991.
- Ishida T, Dorfman HD, Steiner GC, Norman A. Cystic angiomatosis of bone with sclerotic changes mimicking osteoblastic metastases. *Skeletal Radiol* 1994;23:247–252.
- Jacobs JE, Kimmelstiel P. Cystic angiomatosis of the skeletal system. J Bone Joint Surg Am 1953;35:409–420.
- Johnson PM, McClure JG. Observations on massive osteolysis: a review of the literature and report of a case. *Radiology* 1958; 71: 28–42.
- Joseph UA, Jhingran SG. Technetium-99m labeled red blood cells in the evaluation of hemangioma. *Clin Nucl Med* 1987; 12: 845–847.
- Jumbelic M, Feuerstein IM, Dorfman HD. Solitary intraosseous lymphangioma: a case report. J Bone Joint Surg Am 1984;66: 1479–1480.
- 56. Kahn HJ, Bailey D, Marks A. Monoclonal antibody D2-40, a new marker of lymphatic endothelium, reacts with Kaposi's sarcoma and a subset of angiosarcomas. *Mod Pathol* 2002;15: 434–440.
- 57. Kaleem Z, Kyriakos M, Totty WG. Solitary skeletal hemangioma of the extremities. *Skeletal Radiol* 2000;29:502–513.
- Karlin CA, Brower AC. Multiple primary hemangiomas of bone. Am J Roentgenol 1977;129:162–164.
- Laredo JD, Assouline E, Gelbert F, et al. Vertebral hemangiomas: fat content as a sign of aggressiveness. *Radiology* 1990; 177:467–472.
- Laredo JD, Reizine D, Bard M, Merland JJ. Vertebral hemangiomas: radiologic evaluation. *Radiology* 1986;161:183–189.
- Lateur L, Simoens CJ, Gryspeerdt S, et al. Skeletal cystic angiomatosis. Skeletal Radiol 1996;25:92–95.
- Lawley LP, Cerimele F, Weiss SW, et al. Expression of Wilms tumor 1 gene distinguishes vascular malformations from proliferative endothelial lesions. *Arch Dermatol* 2005;141:1297–1300.
- Leon-Villapalos J, Wolfe K, Kangesu L. GLUT-1: an extra diagnostic tool to differentiate between haemangiomas and vascular malformations. *Br J Plast Surg* 2005;58:348–352.
- Levin DC, Gordon DH, McSweeney J. Arteriography of peripheral hemangiomas. *Radiology* 1976;121:625–630.
- Levey DS, MacCormack LM, Sartoris DJ, et al. Cystic angiomatosis: Case report and review of the literature. *Skeletal Radiol* 1996;25:287–293.
- Ling S, Rafii M, Klein M. Epithelioid hemangioma of bone. Skeletal Radiol 2001;30:226–229.
- Lomasney LM, Martinez S, Demos TC, Harrelson JM. Multifocal vascular lesions of bone: imaging characteristics. *Skeletal Radiol* 1996;25:255–261.
- Makhija M, Bofill ER. Hemangioma: a rare cause of photopenic lesion on skeletal imaging. *Clin Nucl Med* 1988;13:661–662.
- Martinat P, Cotten A, Singer B, et al. Solitary cystic lymphangioma. *Skeletal Radiol* 1995;24:556–558.
- Marymont JV, Shapiro WM. Vertebral hemangioma associated with spinal cord compression. *South Med* J1988;81:1586–1587.
- Matsumoto K, Ishizawa M, Okabe H, Taniguchi I. Hemangioma of bone arising in the ulna: imaging findings with emphasis on MR. *Skeletal Radiol* 2000;29:231–234.
- Meyer JS, Hoffer FA, Barnes PD, Mulliken JB. Biological classification of soft-tissue vascular anomalies: MR correlation. *Am J Roentgenol* 1991;157:559–564.
- Mirra JM, Arnold WD. Skeletal hemangiomatosis in association with hereditary hemorrhagic telangiectasia. J Bone Joint Surg Am 1973;55:850–854.
- Mirra JM. Vascular tumors. In: Mirra JM, ed. Bone tumors: clinical, radiologic, and pathologic considerations. Philadelphia: Lea & Febiger, 1989.
- Mohan V, Gupta SK, Tuli SM, Sanyal B. Symptomatic vertebral hemangiomas. *Clin Radiol* 1980;31:575–579.
- Moreno AJ, Reeves TA, Turnbull GL. Hemangioma of bone. Clin Nucl Med 1988;13:668–669.
- Mulder JD, Kroon HM, Schütte HE, Taconis WK. Radiologic atlas of bone tumors. Elsevier: Amsterdam, 1993:241–254, 507–516.
- 78. Mulliken JB, Glowacki J. Hemangiomas and vascular malforma-

tions in infants and children: a classification based on endothelial characteristics. *Plast Reconstr Surg* 1982;69:412–420.

- Murphey MD, Fairbairn KJ, Parman LM, et al. Musculoskeletal angiomatous lesions: radiologic-pathologic correlation. *Radio-Graphics* 1995;15:893–917.
- North PE, Waner M, Mizeracki A, Mihm MC Jr. GLUT1: a newly discovered immunohistochemical marker for juvenile hemangiomas. *Hum Pathol* 2000;31:11–22.
- O'Connell JX, Nielsen GP, Rosenberg AE. Epithelioid vascular tumors of bone: a review and proposal of a classification scheme. *Adv Anat Pathol* 2001;8:74–82.
- Paley D, Evans DC. Angiomatous involvement of an extremity: a spectrum of syndromes. *Clin Orthop Rel Res* 1986;206:215– 218.
- Pearce WH, Rutherford RB, Whitehill TA, Davis K. Nuclear magnetic resonance imaging: its diagnostic value in patients with congenital vascular malformations of the limbs. *J Vasc Surg* 1988;8:64–70.
- Peh WCG, Ngan H. Lymphography still useful in the diagnosis of lymphangiomatosis. Br J Radiol 1993;66:28–31.
- Reid AB, Reid IL, Johnson G, et al. Familial diffuse cystic angiomatosis of bone. *Clin Orthop Rel Res* 1989;238:211–218.
- 86. Resnick D, Kyriakos M, Greenway GD. Tumors and tumor-like lesions of bone: imaging and pathology of specific lesions. In: Resnick D, ed. *Diagnosis of bone and joints disorders*, 3rd ed. Philadelphia: WB Saunders, 1995:3628–3938.
- Ritchie G, Zeier FG. Hemangiomatosis of the skeleton and the spleen. J Bone Joint Surg Am 1956;38:115–122.
- Rosenquist CJ, Wolfe DC. Lymphangioma of bone. J Bone Joint Surg 1978;50:158–162.
- Ross JS, Masaryk TJ, Modic MT, et al. Vertebral hemangioma: MR imaging. *Radiology* 1987;165:165–169.
- Schajowicz F. Tumors and tumorlike lesions of bone. Pathology, radiology, and treatment, 2nd ed. Berlin: Springer-Verlag, 1994.
- Schajowicz F, Aiello CL, Francone MV, Giannini RE. Cystic angiomatosis (hamartous haemolymphangiomatosis) of bone. *J Bone Joint Surg Br* 1978;60:100–106.
- Schopfner CE, Allen RP. Lymphangioma of bone. *Radiology* 1961;76:449–453.
- 93. Sherman RS, Wilner D. The roentgen diagnosis of hemangioma of bone. *Am J Roentgenol* 1961;86:1146–1159.
- 94. Spieth ME, Greenspan A, Forrester DM, et al. Gorham's disease of the radius: radiographic, scintigraphic, and MRI findings with pathologic correlation. *Skeletal Radiol* 1997;26: 659–663.
- Stafford JM, James TT, Allen AM, Dixon LR. Hemophilic pseudotumor: radiologic-pathologic correlation. *RadioGraphics* 2003;23:852–856.
- Steiner GM, Farman J, Lawson JP. Lymphangiomatosis of bone. Radiology 1969;93:1093–1098.
- Suh JS, Hwang G, Hahn SB. Soft tissue hemangiomas: MR manifestations in 23 patients. *Skeletal Radiol* 1994;23:621–625.
- Suneja R, Davies AM, Deshmukh NS, et al. Metachronous haemorrhagic epithelioid and spindle cell haemangioma of bone. *Skeletal Radiol* 2002;31:475–478.
- Sung MS, Kim YS, Resnick D. Epithelioid hemangioma of bone. Skeletal Radiol 2000;29:530–534.
- Torg JS, Steel HH. Sequential roentgenographic changes occurring in massive osteolysis. *J Bone Joint Surg Am* 1969;51:1649–1655.
- 101. Unni KK. Dahlin's bone tumors. General aspects and data on 11,087 cases, 5th ed. New York: Lippincott-Raven, 1996.
- Vilanova JC, Barceló J, Smirniotopoulos JG, et al. Hemangioma from head to toe: MR imaging with pathologic correlation. *RadioGraphics* 2004;24:367–385.
- 103. Vinee P, Tanyu O, Havenstein KH, et al. CT and MRI of Gorham syndrome. *J Comput Assist Tomogr* 1994;18:985–989.
- 104. Waldron RT, Zeller JA. Diffuse skeletal hemangiomatosis with visceral involvement. J Can Assoc Radiol 1969;20:119–123.
- Walter JW, North PE, Waner M, et al. Somatic mutation of vascular endothelial growth factor receptors in juvenile hemangioma. *Genes Chromosomes Cancer* 2002;33:295–303.
- Weiss SW, Goldblum JR. Enzinger and Weiss's soft tissue tumors, 4th ed. Philadelphia: Mosby-Harcourt, 2001.
- 107. Wenger DE, Wold LE. Benign vascular lesions of bone: radiologic and pathologic features. *Skeletal Radiol* 2000;29:63–74.
- 108. Williams AG Jr, Mettler FA Jr. Vertebral hemangioma. Radionu-
clide, radiographic, and CT correlation. *Clin Nucl Med* 1985; 10:598.

- 109. Winterberger AR. Radiographic diagnosis of lymphangiomatosis of bone. *Radiology* 1972;102:321–324.
- Yamamoto T, Kurosaka M, Mizuno K. Juxta-articular hemangioma of long bone. *Skeletal Radiol* 2000;29:535–537.
- 111. Yoo SY, Hong SH, Chung HW, et al. MRI of Gorham's disease: findings in two cases. *Skeletal Radiol* 2002;31:301–306.

Glomus Tumor, Gliomangioma, and Glomangiomyoma

- 112. Bjorkengren AG, Resnick D, Haghighi P, Sartoris DJ. Intraosseous glomus tumor: report of a case and review of the literature. *Am J Roentgenol* 1986;147:739–741.
- 113. Brouillard P, Boon LM, Mulliken JB, et al. Mutations in a novel factor, glomulin, are responsible for glomuvenous malformations ("glomangiomas"). Am J Hum Genet 2002;70:866–874.
- 114. Brouillard P, Ghassibe M, Penington A, et al. Four common glomulin mutations cause two thirds of glomuvenous malformations ("familial glomangiomas"): evidence for a founder effect. *J Med Genet* 2005;42:313.
- 115. Folpe AL. Glomus tumours. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:136–137.
- 116. Folpe AL, Fanburg-Smith JC, Miettinen M, Weiss SW. Atypical and malignant glomus tumors: analysis of 52 cases, with a proposal for the reclassification of glomus tumors. *Am J Surg Pathol* 2001;25:1–12.
- Fornage BD. Glomus tumor in the fingers: diagnosis with US. Radiology 1988;167:183–185.
- Greenfield GB, Arrington JA. Imaging of bone tumors. A multimodality approach. Philadelphia: JB Lippincott, 1995.
- Kneeland JB, Middleton WD, Matloub HS, et al. High resolution MR imaging of glomus tumor. J Comput Assist Tomogr 1987; 11:351–352.
- Sugiura I. Intra-osseous glomus tumor: a case report. J Bone Joint Surg Br 1976;58:245–247.
- 121. Theumann NH, Goettmann S, Le Viet D, et al. Recurrent glomus tumors of fingertips: MR imaging evaluation. *Radiology* 2002:223:143–151.

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- Abrahams TG, Bula W, Jones M. Epithelioid hemangioendothelioma of bone. *Skeletal Radiol* 1992;21:509–513.
- 123. Aditya GS, Santosh V, Yasha TC, Shankar SK. Epithelioid and retiform hemangioendothelioma of the skull bone—report of four cases. *Indian J Pathol Microbiol* 2003;46:645–649.
- 124. Alpern MB, Thorsen MK, Kellman GM, et al. CT appearance of hemangiopericytoma. J Comput Assist Tomogr 1986;10: 264–267.
- 125. Anez LF, Gupta SM, Berger D, et al. Scintigraphic evaluation of multifocal hemangioendothelioma of bone. *Clin Nucl Med* 1993; 18:840–843.
- 126. Antonescu CR, Argani P, Erlandson RA, et al. Skeletal and extraskeletal myxoid chondrosarcoma: a comparative clinicopathologic, ultrastructural, and molecular study. *Cancer* 1998;83: 1504–1521.
- Boutin RD, Spaeth HJ, Mangalik A, Sell JJ. Epithelioid hemangioendothelioma of bone. *Skeletal Radiol* 1996;25:391–395.
- 128. Breza TS, Magro CM. CD34 expression in primary cutaneous malignant melanoma: apropos of a case and review of the aberrant melanoma phenotype. *J Cutan Pathol* 2005;32: 685–689.
- Campanacci M, Boriani S, Giunti A. Hemangioendothelioma of bone: a study of 29 cases. *Cancer* 1980;46:804–814.
- Coldwell DM, Baron RL, Charnsangavej C. Angiosarcoma: diagnosis and clinical course. *Acta Radiol* 1989;30:627–631.
- Dal Cin P, Sciot R, Fletcher CD, et al. Trisomy 21 in solitary fibrous tumor. *Cancer Genet Cytogenet* 1996;86:58–60.
- De Young BR, Frierson HF Jr, Ly MN, et al. CD31 immunoreactivity in carcinomas and mesotheliomas. *Am J Clin Pathol* 1998; 110:374–377.
- 133. Deshpande V, Rosenberg AE, O'Connell JX, Nielsen GP. Epithe-

lioid angiosarcoma of the bone: a series of 10 cases. Am J Surg Pathol 2003;27:709–716.

- 134. Enzinger FM, Weiss SW. Hemangioendothelioma: vascular tumors of intermediate malignancy. In: Enzinger FM, Weiss SW, eds. *Soft tissue tumors*, 3rd ed. St. Louis: CV Mosby, 1995.
- 135. Enzinger FM, Weiss SW. Malignant vascular tumors. In: Enzinger FM, Weiss SW, eds. Soft tissue tumors, 3rd ed. St. Louis: CV Mosby, 1995.
- Fulling KH, Gersell DJ. Neoplastic angioendotheliomatosis. Histologic, immunohistochemical, and ultrastructural findings in two cases. *Cancer* 1983;51:1107–1118.
- Gengler C, Guillou L. Solitary fibrous tumour and haemangiopericytoma: evolution of a concept. *Histopathology* 2006;48: 63–74.
- Greenspan A, Unni KK, Blake L, Rab G. Extraskeletal myxoid chondrosarcoma: an unusual tumor in a 6-year-old boy. *Can Assoc Radiol J* 1994;45:62–65.
- 139. Guillou L, Fletcher JA, Fletcher CDM, Mandahl N. Extrapleural solitary fibrous tumor and hemangiopericytoma. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:86–90.
- Hendrix RW, Rogers LF, Davis TM. Cortical bone metastases. *Radiology* 1991;181:409–413.
- 141. Hisaoka M, Aoki T, Kouho H, et al. Maffucci's syndrome associated with spindle cell hemangioendothelioma. *Skeletal Radiol* 1997;126:191–194.
- 142. Hisaoka M, Okamoto S, Aoki T, et al. Spinal epithelioid hemangioendothelioma with epithelioid angiosarcomatous areas. *Skeletal Radiol* 2005;34:745–749.
- Hudson TM. Radiologic-pathologic correlation of musculoskeletal lesions. Baltimore: Williams & Wilkins, 1987.
- 144. Huvos AG. Angiosarcoma of bone (epithelioid hemangioendothelioma, malignant hemangioendothelioma). In: Huvos AG, ed. *Bone tumors: diagnosis, treatment and prognosis,* 2nd ed. Philadelphia: WB Saunders, 1991.
- Ignacio EA, Palmer KM, Mathur SC, et al. Epithelioid hemangioendothelioma of the lower extremity. *RadioGraphics* 1999;19: 531–537.
- 146. Juan C-J, Huang G-S, Hsueh C-J, et al. Primary hemangiopericytoma of the tibia: MR and angiographic correlation. *Skeletal Radiol* 2000;29:49–53.
- 147. Kida T, Hujita Y, Munaka S, et al. Malignant hemangioendothelioma demonstrated by thallium imaging. *Clin Nucl Med* 1987; 11:886–887.
- 148. Kleer CG, Unni KK, McLeod RA. Epithelioid hemangioendothelioma of bone. *Am J Surg Pathol* 1996;20:1301–1311.
- Lee KS, Rossleigh MA, Fernandes VB, Morris JG. Scintigraphic features of malignant epithelioid hemangioendothelioma. *Clin Nucl Med* 1989;14:501–503.
- Lian Y-W, Yao M-S, Hsieh S-C, et al. MRI of hemangiopericytoma of the sacrum. *Skeletal Radiol* 2004;33:485–487.
- Linovitz RJ, Resnick D, Keissling P, et al. Tumor-induced osteomalacia and rickets: a surgically curable syndrome. Report of two cases. *J Bone Joint Surg Am* 1976;58:419–423.
- Lorigan JG, David CL, Evans HL, Wallace S. The clinical and radiologic manifestations of hemangiopericytoma. *Am J Roentgenol* 1989;153:345–349.
- 153. Mac-Moune Lai F, To KF, Choi PC, et al. Kaposiform hemangioendothelioma: five patients with cutaneous lesion and long follow-up. *Mod Pathol* 2001;14:1087–1092.
- 154. Mandahl N, Orndal C, Heim S, et al. Aberrations of chromosome segment 12q13-15 characterize a subgroup of hemangiopericytomas. *Cancer* 1993;71:3009–3013.
- 155. Maruyama N, Kumagai Y, Ishida Y, et al. Epithelioid hemangioendothelioma of the bone tissue. *Virchows Arch A* 1985;407: 159–165.
- 156. Meis-Kindblom JM, Kindblom LG. Angiosarcoma of soft tissue: a study of 80 cases. *Am J Surg Pathol* 1998;22:683–697.
- 157. Mendlick MR, Nelson M, Pickering D, et al. Translocation t(1;3) (p36.3;q25) is a nonrandom aberration in epithelioid hemangioendothelioma. *Am J Surg Pathol* 2001;25:684–687.
- 158. Middleton LP, Duray PH, Merino MJ. The histological spectrum of hemangiopericytoma: application of immunohistochemical analysis including proliferative markers to facilitate diagnosis

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and predict prognosis. Hum Pathol 1998;29: 636-640.

- 159. Miettinen MM, el-Rifai W, Sarlomo-Rikala M, et al. Tumor sizerelated DNA copy number changes occur in solitary fibrous tumors but not in hemangiopericytomas. *Mod Pathol* 1997;10: 1194–1200.
- 160. Nakazono T, Kudo S, Matsuo Y, et al. Angiosarcoma associated with chronic lymphedema (Stewart-Treves syndrome) of the leg: MR imaging. *Skeletal Radiol* 2000;29:413–416.
- 161. Nappi O, Ritter JH, Pettinato G, Wick MR. Hemangiopericytoma: histopathological pattern or clinicopathologic entity? *Semin Diagn Pathol* 1995;12:221–232.
- 162. O'Connell JX, Kattapuram SV, Mankin HJ, et al. Epithelioid hemangioma of bone. A tumor often mistaken for low-grade angiosarcoma or malignant hemangioendothelioma. *Am J Surg Pathol* 1993;17:610–617.
- 163. Ravenel JG, Goodman PC. Late pulmonary metastases from hemangiopericytoma of the mandible: unusual findings on CT and MR imaging. *Am J Roentgenol* 2001;177:244–246.
- 164. Roessner A, Boehling T. Angiosarcoma. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:322–323.
- 165. Rong Y, Sato K, Yamamura S, et al. Malignant hemangioendothelioma of the left calcaneus associated with fever and hematological abnormalities. *Skeletal Radiol* 1997;26:64–66.
- 166. Rosenthal DI, Treat ME, Mankin HJ, et al. Treatment of epithelioid hemangioendothelioma of bone using a novel combined approach. *Skeletal Radiol* 2001;30:219–222.
- 167. Sahin Akyar G, Fitöz S, Akpolat I, et al. Primary hemangiopericytoma of bone located in the tibia. *Skeletal Radiol* 1997;26: 47–50.
- 168. Sapino A, Bongiovanni M, Cassoni P, et al. Expression of CD31 by cells of extensive ductal in situ and invasive carcinomas of the breast. *J Pathol* 2001;194:254–261.
- 169. Shah ZK, Peh WCG, Shek TWH, et al. Hemangioendothelioma with an epithelioid phenotype arising in hemangioma of the fibula. *Skeletal Radiol* 2005;34:750–754.
- 170. Shin MS, Carpenter JT, Ho KJ. Epithelioid hemangioendothelioma: CT manifestations and possible linkage to vinyl chloride exposure. *J Comput Assist Tomogr* 1991;15:505–507.
- Staut AP, Murray MR. Hemangiopericytoma: a vascular tumor featuring Zimmermann's pericytes. *Ann Surg* 1942;116:26–33.
- Steinbach LS, Ominsky SH, Shpall S, Perkocha LA. MR imaging of spindle cell hemangioendothelioma. J Comput Assist Tomogr 1991;15:155–157.
- 173. Stewart FW, Treves N. Lymphangiosarcoma in postmastectomy

lymphedema: a report of six cases on elephantiasis chirurgica. *Cancer* 1948;1:64–81.

- 174. Stout AP. Hemangioendothelioma: a tumor of blood vessels featuring vascular endothelial cells. *Ann Surg* 1943;118:445–464.
- 175. Stout AP, Murray MR. Hemangiopericytoma: a vascular tumor featuring Zimmermann's pericytes. *Ann Surg* 1942;116:26–33.
- 176. Tang JS, Gold RH, Mirra JM, Eckardt J. Hemangiopericytoma of bone. *Cancer* 1988;62:848–859.
- 176a. Timar J, Meszaros L, Orosz Z, et al. TW1 expression in angiogenic tumours of the skin. *Histopathology* 2005;47:67–73.
- 177. Tsang WY, Chan JK. The family of epithelioid vascular tumors. *Histol Histopathol* 1993;8:187–212.
- 178. Tsuneyoshi M, Dorfman HD, Bauer TW. Epithelioid hemangioendothelioma of bone. A clinicopathologic, ultrastructural, and immunohistochemical study. *Am J Surg Pathol* 1986;10: 754–764.
- 179. Unni KK, Ivins JC, Beabout JW, Dahlin DC. Hemangioma, hemangiopericytoma, and hemangioendothelioma (angiosarcoma) of bone. *Cancer* 1971;27:1403–1414.
- 180. Weiss SW, Bridge JA. Epithelioid hemangioendothelioma. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:173–174.
- Weiss SW, Enzinger FM. Epithelioid hemangioendothelioma: a vascular tumor often mistaken for a carcinoma. *Cancer* 1982;50: 970–981.
- 182. Weiss SW, Ishak KG, Dail DH, et al. Epithelioid hemangioendothelioma and related lesions. *Semin Diagn Pathol* 1986;3: 259–287.
- 183. Weiss SW, Lasota J, Miettinen MM. Angiosarcoma of soft tissue. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:175–177.
- Wenger DE, Wold LE. Malignant vascular lesions of bone: radiologic and pathologic features. *Skeletal Radiol* 2000;29:619–631.
- 185. Wobser M, Siedel C, Schrama D, et al. Expression pattern of the lymphatic and vascular markers VEGFR-3 and CD31 does not predict regional lymph node metastasis in cutaneous melanoma. *Arch Dermatol Res* 2006;297:352–357.
- Wold LE, Unni KK, Beabout JW, et al. Hemangioendothelial sarcoma of bone. Am J Surg Pathol 1982;6:59–70.
- Wold LE, Unni KK, Cooper KL, et al. Hemangiopericytoma of bone. Am J Surg Pathol 1982;6:53–58.
- Yagmai I. Angiographic features of soft-tissue and osseous hemangiopericytomas. *Radiology* 1978;126:653–659.

CHAPTER 7

Miscellaneous Tumors And Tumor-like Lesions

BENIGN LESIONS 387 Giant Cell Tumor 387 Malignant Giant Cell Tumor 396 Simple Bone Cyst 399 Aneurysmal Bone Cyst 408 Solid Variant of Aneurysmal Bone Cyst 420 Intraosseous Lipoma 424 Fibrocartilaginous Mesenchymoma 425

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Benign Lesions

Giant Cell Tumor

Giant cell tumor (GCT) is a locally aggressive neoplasm characterized by richly vascularized tissue containing proliferating mononuclear stromal cells and many osteoclast-like, multinucleated giant cells randomly but evenly distributed throughout (23,58,74,82). GCT represents approximately 5% (94) to 8.6% (82) of all primary bone tumors and about 22.7% of benign bone tumors (94) and is the sixth most common primary osseous neoplasm (30).

The mononuclear component of GCT consists of two types of cells, fibroblast-like and histiocyte/monocytic-

like stromal cells (4,78,139). Roessner et al. (78) also demonstrated that only fibroblast-like stromal cells were able to incorporate [³H]-thymidine, thus representing the proliferating neoplastic cell pool. Recently, Wulling et al. (100,101) provided evidence from immunohistochemistry (IHC), tissue culture experiments, and molecular biology studies that the proliferating neoplastic stromal cells in GCT are derived from mesenchymal stem cells. In vitro, these stromal cells of GCT can still differentiate into osteoblasts, chondroblasts, and adipocytes. In addition, Atkins et al. demonstrated that these stromal cells, similar to osteoblasts, express factors necessary for osteoclast formation and differentiation (osteoclast differentiation factor/ODF or receptor activator of nuclear factor kappa B ligand/RANKL), leading to the hypothesis that GCT is a mesenchymal stem cell-derived tumor with the ability to attract monocytic precursors (the second mononuclear component in GCT) and to induce their transformation to osteoclastic giant cells (6,100). In cytogenetic studies, telomeric associations (end-to-end fusions of apparently intact chromosomes) involving chromosomes 11p, 13p, 14p, 15p, 19q, 20q, and 21p have been identified as the most commonly occurring chromosomal aberration (16,74).

Other structural chromosomal abnormalities have also been detected in single cases (57). In an analysis of 12 cases including primary, recurrent, metastatic, and malignant GCTs, Rao et al. (73) observed *loss of het*erozygosticy (LOH) of 1p, 3p, 5q, 9q, 10q, and 19q, whereas LOH of 17p and 9p occurred only in pulmonary metastases of GCTs (73). However, the possible roles of these findings in the evolution of GCT are not clearly understood.

In a study of c-myc expression (a transcription factor involved in cell proliferation and in cell growth and differentiation) in GCTs, Gamberi et al. (32) demonstrated a strong correlation between c-myc overexpression at the mRNA and protein levels and the occurrence of lung metastases.

Clinical Presentation

GCT occurs almost exclusively after skeletal maturity, when the growth plates are obliterated, and patients are usually between the ages of 20 and 40 years, with a female preponderance (2:1). At least 60% of cases arise in long bones and almost all extend to the articular end (29). Those unusual cases with presentation in the metaphysis are seen in skeletally immature patients (46,71). The most common skeletal sites are the distal femur, the proximal tibia, the distal radius, the proximal humerus, and the sacrum (23). Rarely, the tumor arises in other sites, such as the bones of hands and feet, the vertebral bodies, the ribs (14,18,42,89,94), the scapula (3), the ischium (83), and the skull (48) (Fig. 7-1). Very uncommonly, GCT may be multifocal (69,85,96), with a reported frequency of 0.04% to 1%(22,58,91,94). The majority of multifocal GCTs are associated with Paget disease of bone (31,41,72), and most have a predilection for the skull, face (43), and ilium (82). Multifocal GCTs in patients who do not exhibit Paget disease usually occur in the bones of the hand (7,8,21) and characteristically affect a slightly younger age group (27,98).

The symptoms most commonly reported include pain of increasing intensity, with local swelling and tenderness of the affected area. Limitation of motion of the adjacent joint is also present in many patients (82). Only in exceptional cases is a pathologic fracture the first sign of a GCT. Although a histologic grading system exists, the clinical behavior of GCT cannot be accurately predicted on the basis of histologic, clinical, or radiologic features (11,12,15,28). Recurrences after surgical curettage and bone grafting are common [reported recurrence rates range from 12% to 50% (40)].

Imaging

The characteristic radiographic features of GCT are those of a purely osteolytic lesion with a geographic type of bone destruction (50). The lesion is radiolucent, frequently expansive, and eccentrically located, without marginal sclerosis but usually with well-defined borders (Figs. 7-2 and 7-3). Internal trabeculations or pseudotrabeculations are occasionally present (82) (Fig. 7-4), most likely representing a nonuniform growth of the tumor (62). There is usually no periosteal reaction. Attempts have been made to assess the aggressiveness of GCT on the basis of radiologic criteria. According to some investigators (60), nonaggressive tumors exhibit a prominent trabeculation and no cortical expansion or soft tissue mass. Conversely, aggressive tumors exhibit a lack of trabeculation, and expansion or destruction of the cortex and soft tissue mass are present (Fig. 7-5).



Figure 7-1 Giant cell tumor: skeletal sites of predilection, peak age range, and male-to-female ratio.



Figure 7-2 Giant cell tumor. A: Anteroposterior radiograph of the right knee of a 30-year-old woman shows an osteolytic lesion eccentrically located in the proximal tibia and extending into the articular end of the bone. **B:** Radiolucent lesion in a 27-year-old woman affects almost the entire proximal end of the humerus. Note a pathologic fracture at the distal extent of the tumor.

Α



Figure 7-3 Giant cell tumor. Anteroposterior (A) and lateral (B) radiographs of the right knee of a 32-year-old man show a purely osteolytic lesion in the distal end of the femur. Note its eccentric location, the absence of reactive sclerosis, and the extension of the tumor into the articular end of the bone (*continued*).

В



Figure 7-3 *Continued* **C:** In another patient, an eccentric osteolytic lesion extends into the articular end of the distal radius.



Figure 7-4 Giant cell tumor. A: Anteroposterior radiograph of the right hip of a 27-year-old woman shows a radiolucent lesion with internal trabeculations in the femoral head. **B:** Anteroposterior radiograph of the left wrist of a 36-year-old woman shows a trabeculated lesion in the distal radius.



Figure 7-5 Giant cell tumor. Dorsovolar radiograph of the left wrist of a 56-year-old woman shows an osteolytic lesion in the distal radius that has destroyed the cortex and that extends into the soft tissues.

Conventional tomography, particularly trispiral or hypocycloidal, reveals the osseous margins of the lesion and the status of the cortex more clearly than radiographs. Arthrotomography is effective for the evaluation of tumor invasion through the subchondral plate and articular cartilage (39). When a soft tissue mass is present, computed tomography (CT) or magnetic resonance imaging (MRI) examination may be particularly helpful (24,56,75,90). CT may outline the tumor extent and better delineate areas of cortical destruction (Fig. 7-6). The attenuation coefficient of unenhanced GCT averaged 44 Hounsfield units, similar to measurements of other noncalcified bone lesions (39). However, enhancement after infusion of a contrast agent is striking according to Hudson et al. (39). Conversely, deSantos and Murray (25) did not observe a significant contrast enhancement.

On MRI, like most bone tumors, GCT exhibits a low to intermediate signal intensity on T1-weighted images and a high signal on T2-weighted sequences (Fig. 7-7). The intramedullary portion of tumor is best seen on T1 weighting, whereas its extraosseous component is more clearly observed on T2-weighted images (13,38) (Fig. 7-8). After intravenous injection of gadolinium, heterogeneous enhancement of the tumor is observed (Fig. 7-9). Some reports in the literature indicate that certain instances of GCT containing large amounts of hemosiderin may show different MRI characteristics (2). Aoki et al. (1) found in these instances a markedly decreased signal on both T1- and T2-weighted sequences and, in one case, the extraosseous tumor extension appeared as a signal void area on MR imaging. MRI is also effective in demonstrating subchondral breakthrough and extension of tumor into the adjacent joint (38). Fluid levels in the tumor have been reported (45,75) and are considered by some investigators to be secondary to an aneurysmal bone cyst (ABC) component (39,45).

Radionuclide bone scan rarely provides additional information because the degree of tracer uptake does not correlate with the histologic grade of the tumor (39,53,68,95). However, scintigraphy may help in detection of multiple foci if a multicentric GCT is clinically



Figure 7-6 Giant cell tumor: computed tomography (CT). A: Anteroposterior radiograph of the left knee of a 33-year-old woman shows an osteolytic lesion in the medial femoral condyle (*arrows*). There is no definite evidence of cortical break-through. B: Axial CT section through the tumor demonstrates destruction of the cortex and the presence of a soft tissue mass.



С



Α

С

extends laterally into the soft tissues (arrowhead).

В

Е





Figure 7-9 Giant cell tumor: magnetic resonance imaging (MRI). Coronal **(A)** and sagittal **(B)** T1-weighted MR images show a low-signal-intensity lesion in the medial femoral condyle of a 37-year-old woman. The tumor extends to the articular end of bone. On coronal **(C)** and axial **(D)** T2-weighted MR images the tumor exhibits heterogeneous signal.



Ε

Figure 7-9 *Continued* **E:** Axial T1-weighted fat-suppressed MRI obtained after intravenous administration of gadolinium shows significant enhancement of the lesion.

suspected. In a study by Hudson et al. (39), these investigators found in 49% of cases an abnormal uptake pattern resembling a doughnut: intense uptake around the periphery with relatively little activity in the central portion of the tumor. They contended that this appearance was due to uptake of the bone-seeking radiopharmaceutical agents predominantly by reactive new bone or by hyperemic bone around the tumor, with the tumor tissue retaining little tracer. Occasionally, an increased tracer activity can be detected across the adjacent joint (49). This phenomenon may be due to increased blood flow and to increased bone turnover secondary to disuse osteoporosis (86).

Although 5% to 10% of GCTs undergo malignant degeneration (or are malignant from their inception) (10,36,54), malignant GCT is unaccompanied by additional or specific radiologic characteristics and therefore cannot be diagnosed radiographically.

The radiographic presentation of GCT complicating Paget disease is usually that of an expansive lytic lesion, frequently accompanied by a soft tissue mass (53,65,81).

Histopathology

The histologic features of GCT include a dual population of fibrocytic or monocytic mononuclear stromal cells and of giant cells that are usually uniformly distributed throughout the tumor (Fig. 7-10). Morphologically, the giant cells somewhat resemble osteoclasts and, like all resorptive giant cells, exhibit marked acid phosphatase activity (5). However, they are much larger than normal osteoclasts and are not apposed to bone surfaces and therefore do not possess a ruffled border (88). The cells are round, oval, or fusiform and vary considerably in shape owing to their capacity for ameboid motility. In general, they exhibit large nuclei with little chromatin and very few inconspicuous nucleoli (74,82). The number of nuclei may vary widely but is usually 20 or more. Giant cells never exhibit mitotic activity. Areas of hemorrhage, and occasional groups of foam cells and of hemosiderin-laden macrophages, are frequently present (30).

In typical areas, the tumor matrix contains only sparse amounts of collagen. On histologic study, silver staining reveals a network of reticulin fibers that surrounds the individual cells but does not penetrate the



Figure 7-10 Histopathology of a giant cell tumor. A: Densely arranged tumor tissue consists of mononuclear cells intermixed with numerous uniformly distributed large giant cells containing up to 50 or more nuclei, much larger than those observed in other tumors (hematoxylin and eosin, original magnification ×100). **B:** At higher magnification the characteristic appearance of giant cells is better appreciated (hematoxylin and eosin, original magnification ×200).

giant cells (82). The tumor is rich in newly formed capillaries, with walls composed of only a single endothelial layer. Occasionally the vascular channels appear to be directly lined with tumor cells (82). This may be related to the production of proteases [urokinase-type plasminogen activator (u-PA) and matrix metalloproteinases (MMPs), which are capable of degrading proteins of the extracellular matrix] by the cells in GCT that are involved in vascular invasion (33,84,93,102). Elevated u-PA expression appears to be associated with increased tumor aggressiveness (33). Bone or osteoid formation can be seen especially at sites of previous pathologic fracture or curettage and, occasionally, surrounding a focus of recurrent tumor in the soft tissue. Vascular invasion has been observed in the tumor periphery and even in the surrounding soft tissues. However, no studies have demonstrated a correlation with clinical behavior or prognosis. Mitoses are always present and do not indicate malignancy in GCT. However, atypical mitoses are not usually found and, if present, should raise the suspicion of malignancy.

A fibroxanthomatous pattern is sometimes observed in GCT with only a few scattered giant cells. The presence of these lipoid-bearing foam cells, particularly in the vicinity of necrotic foci and with association of fibrous tissue, may indicate spontaneous regression and healing of the tumor (43) or, as others have suggested, a less aggressive type of tumor (82). When this pattern predominates, the GCT may be misdiagnosed as benign fibrous histiocytoma. Some authorities, however, consider the latter lesion to represent merely the end stage of a burned-out GCT (55).

Attempts to correlate the histologic appearance of the tumor with its biological behavior have proven unreliable and have now been abandoned (35). Histologic classification of GCT into three grades, corresponding to the number and size of giant cells and the degree of pleomorphism of the mononuclear cells (grade III being frankly malignant histologically), has been attempted in the past (44). In this historical classification, grade I tumor is characterized by a dense layer of huge giant cells with up to 100 or more nuclei, almost covering the tumor tissue proper of mononuclear cells. Grade II tumor exhibits a considerable reduction in the size and number of giant cells, and the mononuclear cells may be pleomorphic to some degree. In grade III tumors, the giant cells are further reduced in number and there is an increase in pleomorphic mononuclear cells. All three grades may be characterized by extended fields with spindle cells and practically no giant cells. At present, most investigators agree that grading of GCT is without practical value because prediction of the evolution of a GCT based on its histologic features is impossible (26,82,94), and histologic grading has not been shown to correlate with local recurrence rate or the incidence of pulmonary metastases (19,35,52,74).

Malignant Giant Cell Tumor

Although GCT is usually benign, malignant tumors have been reported (17,64,77). However, one must be

very cautious in making the diagnosis of malignant GCT. Several reported cases of "primary" malignant GCT later proved to be sarcomas rich in multinucleated giant cells of the osteoclast type, such as telangiectatic osteosarcomas, giant cell-rich osteosarcomas, fibrosarcomas, or malignant fibrohistiocytomas (82). Dahlin and Unni believe that primary malignant GCT is extremely unusual and that most of these cases actually represent another type of malignancy arising within GCT (94). Nevertheless, malignant transformation may occur spontaneously, after surgery (103), or after radiation therapy. McGrath (59) divided malignant GCT into three types: (a) primary, in which the tumor was malignant from the onset (de novo); (b) evolutionary, in which there was malignant transformation of originally benign GCT spontaneously or after multiple surgical resections for recurrent lesions, or after a long latency period; and (c) secondary, developing as a result of malignant transformation of an initially benign tumor after radiation therapy (59). In the latter group, the majority of sarcomas that occur are fibrosarcomas, malignant fibrous histiocytomas, or osteosarcomas (82). Exceedingly rare cases represent the so-called dedifferentiated GCT, in which there is the concomitant presence of a typical GCT of bone juxtaposed to a sarcoma [such as a malignant fibrous histiocytoma (61)]. Under the heading "Malignancy in giant cell tumor" the World Health Organization (WHO) only discriminates between primary malignancy, i.e., sarcomas arising in a GCT, or secondary malignancy, i.e., sarcomas arising at the site of a previously documented GCT (17). The diagnosis of malignant GCT requires either a frankly sarcomatous change in the lesion or a sarcoma arising in the previous site of a treated GCT (94). The histologic diagnosis of malignant GCT is established only when the stromal cells are frankly sarcomatous, as indicated by pleomorphism, atypia, and high mitotic activity (80,103). In addition, areas of typical benign GCT should be present in the tumor at the time of diagnosis (primary malignant GCT), or in tissues taken previously from the same lesion (secondary malignant GCT) (41,94). Histologic grading of GCT in an attempt to predict local recurrence and clinical prognosis has generally met with little success. Moreover, there are reports of completely benign-appearing GCT that were followed by distant metastases to the lung, having the histologic characteristics of classic GCT (47,52,66,76,92,94,99).

Differential Diagnosis

Radiology

Various lesions may be mistaken for GCT and, conversely, GCT can mimic other lesions that affect the articular end of a bone (87). Primary *ABC* rarely affects the articular end of a bone and occurs in a younger age group. However, after obliteration of the growth plate at skeletal maturity, this lesion may extend into the subarticular region of a long bone, becoming indistinguishable from a GCT. Occasionally, if

a fluid-fluid level is demonstrated either on CT or MRI examination, this feature is more consistent with ABC (75). However, it should be noted that ABC may sometimes coexist with other lesions, among them the GCT. The so-called solid ABC, also referred to as a giant cell reparative granuloma (GCRG) when affecting the articular end of bone, may have the same radiologic characteristics as a conventional GCT (9,67,70). Benign fibrous histiocytoma, because of its frequent location at the end of a long bone, may appear identical to a GCT. However, as mentioned earlier, some investigators consider benign fibrous histiocytoma to be an end stage of a "burned-out" GCT (55). Brown tumor of hyperparathyroidism is yet another lesion that radiologically can mimic GCT. However, the former lesion is usually accompanied by other skeletal manifestations of hyperparathyroidism, such as osteopenia, cortical or subperiosteal resorption, resorptive changes at the distal phalangeal tufts, or loss of the lamina dura of the teeth (53). Occasionally, an unusually large intraosseous ganglion may be mistaken for GCT, although the former lesion invariably exhibits a sclerotic border (Fig. 7-11). Some malignant lesions, such as chondrosarcoma, may extend into the articular end of the



Figure 7-11 Intraosseous ganglion. Large lytic eccentric lesion extending into the articular end of the distal tibia of a 40-year-old man was originally thought to represent a giant cell tumor. The biopsy revealed an intraosseous ganglion. Note a distinct sclerotic border, rarely present in giant cell tumors.

bone and, particularly without radiographically identified calcifications, may closely mimic GCT. *Myeloma* and a *lytic metastasis* occupying subchondral segments of bone can usually be distinguished from GCT without much difficulty (the older age group in which the latter malignancies usually occur is a helpful hint), although at times the radiographic differences between the lesions may not be so obvious (Fig. 7-12A,B). Finally, on rare occasions, *fibrosarcoma, malignant fibrous histiocytoma*, and *fibroblastic osteosarcoma*, because of their purely lytic radiographic presentation, may exhibit some similarities to GCT.





Figure 7-12 Metastasis. A, B: Osteolytic metastasis from a renal cell carcinoma into the articular end of the tibia in this 42-year-old woman has an appearance of a giant cell tumor.

Multicentric GCT should be differentiated from *oste*olytic metastases, myeloma, brown tumor of hyperparathyroidism, and multicentric GCRG (20,34,37,51,70).

Pathology

Diagnosis of GCT, with its typical histology and cytology of giant cells, usually does not create a problem. The characteristic radiologic picture of an expansile, eccentric lytic lesion in the articular end of a long tubular bone adds to the security of the histologic diagnosis. For a tumor formerly considered as grade II or III, diagnosis has been and is still difficult. In such cases several lesions must be considered in the differential diagnosis. In the epiphysis, chondroblastoma has in the past been considered as a cartilage-containing GCT. However, it can now be easily differentiated from GCT by the presence of cleaved or indented nuclei of the mononuclear cells, "chicken-wire"-like calcifications, and cartilage islands that are only exceedingly rarely observed in GCT. Another giant cell-containing epiphyseal lesion, benign fibrous histiocytoma, which some authors consider to be a burned-out GCT (55), may create diagnostic difficulties because GCT may contain areas of storiform and cellular fibrous tissue intermingled with a few giant cells in a pattern exactly like that of benign fibrous histiocytoma. However, the latter tumor does not exhibit broad sheets of polygonal mononuclear cells with heavy interspersal of giant cells, which is a pattern diagnostic of GCT. It is possible for certain epiphyseal lesions to be arbitrarily diagnosed as GCT or as benign fibrous histiocytoma (30). Matsuno (55) reclassified three of five epiphyseal lesions with the latter diagnosis as being GCT. Clear cell chondrosarcoma is fairly easy to distinguish from GCT because it contains clusters of large clear cells (see Chapter 3).

If the giant cell-containing lesion is located in the metaphysis and the growth plate is still open, there are several possibilities in the differential diagnosis.

Nonossifying fibroma and, with a lower probability, fibrous cortical defect can be considered, but because of their giant cells arranged in small groups and exhibiting a resorption activity, and the more or less marked cartwheel-like pattern of the spindle-cell stroma, the histiocytic character of these lesions becomes obvious. Both lesions may be characterized by groups of foam cells. Microscopic fields within GCT may contain spindle cells, fibrosis, and clumps of foamy macrophages that cannot be histologically distinguished from nonossifying fibroma (30). In most cases these changes are only focal and the typical GCT pattern predominates. ABC can be differentiated because of its more or less large spaces bordered by flat undifferentiated cells and containing blood, as well as the presence of smaller and not evenly distributed giant cells, whereas in a true GCT the giant cells are larger and contain more nuclei (Fig. 7-13). However, it should be kept in mind that GCT, like many other tumors, may be associated with secondary ABC formation, so that constituents of both lesions may be found. Differentiation is particularly difficult in so-called solid ABC, also known as GCRG (9,63,97). This tumor contains giant cells, often in large numbers, and has a solid component that resembles the spindle-cell variant of GCT. Its smaller size and the clustered arrangement of the giant cells may assist in the differential diagnosis. The tissue of GCT usually contains less fibrosis and is largely composed of sheets of characteristic mononuclear cells (30). GCRG, on the other hand, is more fibrotic, and hemorrhage and hemosiderin deposition are more prominent. Both lesions may exhibit foci of reactive bone formation, but this is more commonly seen in GCRG (30). Chondromyxoid fibroma is usually not a diagnostic problem because large areas of the tumor possess typical myxoid characteristics. Brown tumor of hyperparathyroidism contains numerous giant cells, but these are smaller than in GCT and, instead of being uniformly distributed, are usually arranged in groups (Fig. 7-14).



Α

Figure 7-13 Giant cell tumor vs. aneurysmal bone cyst. A: True giant cell tumor is characterized by spindle-cell tissue containing numerous giant cells which are usually larger and contain more nuclei than in aneurysmal bone cyst (hematoxylin and eosin, original magnification \times 50). **B:** In a solid area of an aneurysmal bone cyst the giant cells are smaller and are not evenly distributed, but instead arranged in groups. In addition, the presence of osteoid (*bottom right*) and vascular spaces (*top*) makes the distinction obvious (hematoxylin and eosin, original magnification \times 100).



Figure 7-14 Brown tumor of hyperparathyroidism. A: The loose connective tissue with a group of osteoclasts (*upper right corner*) is bordered by immature bone trabeculae made up of woven bone and lined by dense seams of osteoblasts (*bottom*) (hematoxylin and eosin, original magnification ×100). **B:** At high magnification there is an obvious difference between these giant osteoclasts seen in hyperparathyroidism and giant cells of a giant cell tumor (compare with Fig. 7-10) (hematoxylin and eosin, original magnification ×200).

The differential diagnosis must also consider giant cell-containing malignant tumors. A particular problem is that of osteosarcoma with prominent giant cells (socalled *giant cell-rich osteosarcoma*), which emphasizes the fact that GCT is frequently a diagnosis of exclusion. The histologic examination of some osteosarcomas reveals that they consist predominantly of large sheets of giant cells, many or most of which are histologically bland. Production of osteoid may be minimal, detectable only by high-power microscopy as fine strands that surround a population of pleomorphic, mononuclear stromal cells. These cells may be undetectable at low power. However, on higher power examination the tissue consists of scattered hyperchromatic cells exhibiting many atypical mitotic figures. When radiographic studies suggest the presence of matrix production, a permeative growth pattern, a lesion not extending to the articular cartilage, or a lesion in a bone with an active growthplate, giant cell-rich osteosarcoma should be suspected and a very careful histologic examination is warranted (30). By the same token, the giant cell-rich variant of malignant fibrous histiocytoma should be included in the differential diagnosis, particularly when irregular arrangement of the usually smaller giant cells and pleomorphism and hyperchromasia of their nuclei are present. Telangiectatic osteosarcoma may be difficult to exclude unless special consideration is given to the conspicuous pleomorphism of the tumor tissue and the high number of blood-filled spaces. In all three of these malignant tumors, the usual location in the metaphysis may be of help in differentiation from GCT, which is usually located in the articular end of bone.

Δ

GCT may occasionally be confused with *metastatic carcinoma* containing giant cells. These lesions may contain large numbers of giant cells, indistinguishable from those of GCT. The thyroid, breast, and pancreas

(79) are the most common primary sites for these giant cell–rich metastatic lesions. IHC should demonstrate the epithelial quality of the mononuclear carcinoma cells and, in the case of thyroid carcinoma, may show evidence of thyroglobulin production.

R

As can be seen from this discussion, tumors and tumor-like lesions containing considerable numbers of giant cells are quite common (Table 7-1). Each of these must be taken into account and must be differentiated from true GCT when a giant cell–containing lesion is under investigation. As shown in Table 7-1, the giant cells are considered to have different functions. However, with the exception of tumor giant cells, which are formed by and belong to the tumor tissue proper, all are derived from the monohistiocytic branch of hematogenic stem cells and are very close relatives, with corresponding enzymatic pathways involved in their resorptive activity.

The radiologic and pathologic differential diagnosis of GCT is depicted in Figure 7-15.

Simple Bone Cyst

The simple bone cyst (SBC), also called unicameral bone cyst, is a tumor-like lesion of unknown cause, attributed to a local disturbance of bone growth (110,114,122,145). Although the pathogenesis is still unknown (130,131), the lesion appears to be reactive or developmental rather than to represent a true neoplasm (30). SBC consists of a solitary cavity lined by a membrane of variable thickness and filled with a clear yellow fluid (105). It represents approximately 3% of all primary bone lesions (111).

Until now, only two SBCs have been examined cytogenetically, revealing complex clonal rearrangements involving chromosomes 4, 6, 8, 16, 21, and 12 in one

Tumor or Tumor-like Lesion	Type of Giant Cells
Giant cell tumor of bone	Osteoclast-like gigantic GC
Osteoid osteoma	Osteoclast GC
Osteoblastoma	Osteoclast GC
Aneurysmal bone cyst	Osteoclast and resorptive GC
Giant cell reparative granuloma	Osteoclast and resorptive GC
Simple bone cyst	Osteoclast and resorptive GC
Chondrolastoma	"Chondroclast" GC
Chondromyxoid fibroma	"Chondroclast" GC
Clear cell chondrosarcoma	Osteoclast GC
Giant cell-rich osteosarcoma	Tumor and osteoclast GC
Telangiectatic osteosarcoma	Tumor GC
Nonossifying fibroma	Resorptive GC
(fibrous cortical defect)	
Malignant fibrous histiocytoma	Tumor GC
(giant cell_rich variant)	
Brown tumor of hyperparathyroidism	Osteoclast GC
Langerhans cell histiocytosis	Histiocytic GC
Paget disease	Osteoclast GC

 Table 7-1 Giant Cell Tumor and Lesions Containing Different

 Forms of Giant Cells (GC)



Figure 7-15 Radiologic and pathologic differential diagnosis of giant cell tumor.

case, and a translocation, (16;20) (p11.2;q13), in the second case (138,143). In case one, examination of the fourth recurrence by polymerase chain reaction (PCR) revealed a *TP53* mutation (144).

Clinical Presentation

The SBC is more common in males (3:1) and is usually detected during the first 2 decades of life (116). About 65% of these cysts occur in teenagers and an additional 20% in the first decade of life (30,135). The vast majority of SBCs are located in the proximal diaphysis of the humerus and the femur, especially when they occur in patients younger than 17 years (122,134). The symptoms include pain, swelling, or stiffness at the nearest joint (109). A pathologic fracture is often the first sign of the lesion (115,120). In fact, this is the most common complication of SBC and occurs in about 66% of cases. In older patients, the incidence of involvement of atypical sites, such as the calcaneus, talus, and ilium, rises significantly (117,135) (Fig. 7-16). In these sites the lesion is usually asymptomatic and is discovered by accident.

Imaging

Radiographically, the appearance of an SBC is that of a centrally located, well-circumscribed, radiolucent lesion with sclerotic margins. The lesion is located within the metaphysis or diaphysis of a long bone, abutting or being remote from the cartilaginous growth plate (Fig. 7-17). Epiphyseal extension is unusual (112,118).

The cortex is frequently thinned and cortical expansion may be present, but, unlike the ABC, the width of the SBC does not exceed the width of the neighboring growth plate (116). A periosteal reaction is absent unless there has been a pathologic fracture, and this feature distinguishes it from ABC, in which there is almost invariably some degree of periosteal response. Diagnosis is best based on conventional radiographs; conventional tomography and CT are used only exceptionally in equivocal cases.

Occasionally (in 20% of cases in the experience of Struhl et al.), one can identify a characteristic "fallen fragment" sign (128,141). This represents a piece of fractured cortex that is displaced into the interior of the lesion, indicating that the lesion is either hollow or is filled with fluid, as most SBCs are (Fig. 7-18). A variant of this phenomenon is the "trap door" sign (116) [also called a "forme fruste" of the fallen fragment by Reynolds (137)], in which the fragment remains attached to the periosteum but folds inward at the fracture site and floats in the fluid. It is important to obtain multiple views of the cyst to accurately identify this sign because a pathologic fracture through a solid lesion can also produce fragments that appear intramedullary on one view but are in fact adjacent to the outer cortex or are located in the soft tissue. Radiographs taken in both the erect and the recumbent position (133), or the observation of a movement of the fallen fragment under fluoroscopy, may demonstrate the free movement of the fragments within the cyst. These signs permit differentiation of SBC from radiographically similar radiolucent lesions containing solid fibrous or



Figure 7-16 Simple bone cyst: skeletal sites of predilection, peak age range, and male-to-female ratio. The left half of the skeleton shows sites of occurrence seen in an older patient population.



Figure 7-17 Simple bone cyst. A: Anteroposterior radiograph of the right shoulder in a 6-year-old boy shows the typical appearance of this lesion. Its location in the metaphysis and the proximal diaphysis of the humerus is also characteristic. The radiolucent lesion is centrally located and shows pseudosepta. Note the slight thinning of the cortex and lack of periosteal reaction. **B:** Anteroposterior radiograph of the left hip of an 11-year-old girl shows characteristic features of a simple bone cyst. Note the central location, narrow zone of transition, geographic type of bone destruction, pseudotrabeculation, and lack of periosteal reaction.

cartilaginous tissue, such as ABC, fibrous dysplasia, nonossifying fibroma, and enchondroma.

CT may be helpful when the appearance of an SBC is atypical or when the lesion is located in the pelvis. This modality may also assist in the identification of a fallen fragment sign (141). Attenuation values of 15 to 20 Hounsfield units have been reported in the center of fluid-filled cysts (108,117). Skeletal scintigraphy may demonstrate an increased peripheral uptake around the cyst and a decreased central activity, although this appearance is not specific for the SBC (120). Some cysts may be scintigraphically normal. MRI of a cyst shows the signal characteristics of fluid: a low to intermediate signal on T1-weighted images and a bright, homogeneous signal on T2 weighting (116) (Figs. 7-19 and 7-20).

Histopathology

Histologically, SBC is a diagnosis of exclusion. A vigorous surgical curettage, if the lesion is not simply injected, yields almost no solid tissue, but the walls of the cavity may show remnants of fibrous tissue or, occasionally, a flattened single-cell lining (Fig. 7-21). Granulation tissue, containing hemosiderin deposits and small lymphocyte infiltrates, is often present, along with scattered osteoclasts (30). Chemical examination of the fluid usually identifies an elevated alkaline phosphatase level (139). Furthermore, a cloudy-appearing, cell-free material is often found in the loose connective tissue surrounding the cysts (121). With stains for collagen, such as van Gieson stain, the material stains strongly and reveals loose and irregularly arranged fibers. It therefore cannot be fibrin, which it is considered by some authors to represent on the basis of hematoxylin and eosin (H&E) staining (Fig. 7-22).

B

Differential Diagnosis

Radiology

In a long bone, the main differential diagnosis is an *ABC*. The primary differences are that SBC is a centric solitary lesion, with minimal expansion, invariably lacks periosteal reaction, and never extends into the soft tissues. In contrast, an ABC is almost invariably an eccentric lesion, with a significant blown-up appearance, and is always accompanied by a solid periosteal reaction (usually in the form of a buttress) (see Figs. 7-31 to 7-34).



Figure 7-18 Simple bone cyst: "fallen fragment" sign. A: One of the most common complications of simple bone cyst is pathologic fracture, as seen here in the proximal humeral diaphysis of a 6-year-old boy. The presence of the "fallen fragment" sign (*arrow*) is characteristic for this lesion. **B:** Anteroposterior radiograph of the right ankle of a 5-year-old boy shows a radio-lucent lesion in the distal diaphysis of the fibula. There is a pathologic fracture through the lesion and the associated periosteal reaction. A radiodense cortical fragment in the center of the lesion (*arrow*) represents the "fallen fragment" sign, identifying this lesion as a simple bone cyst.

Α



Figure 7-19 Simple bone cyst: magnetic resonance imaging (MRI). A: Anteroposterior radiograph of the right shoulder of a 22-year-old man shows a radiolucent lesion with a narrow zone of transition in the proximal humeral shaft abutting the anatomic neck. **B:** On coronal T1-weighted MRI the lesion exhibits homogeneous intermediate signal intensity (*continued*).

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Figure 7-19 *Continued* **C:** Sagittal T2-weighted MRI demonstrates homogeneous high signal of the cyst.



Figure 7-20 Simple bone cyst: magnetic resonance imaging (MRI). A: Lateral radiograph of the foot of an 18-year-old man shows a radiolucent lesion with slightly sclerotic border in the calcaneus.



Figure 7-20 *Continued* **B:** Sagittal T1-weighted (SE, TR 850, TE 15) MRI demonstrates homogeneous intermediate signal intensity within the lesion, rimmed by low-signal-intensity sclerotic margin. **C:** Sagittal STIR MR image shows that the lesion now is of homogeneous high signal intensity. (From Greenfield GB, Arrington JA. *Imaging of bone tumors.* Philadelphia: JB Lippincott, 1995:217, 218.)



Figure 7-21 Histopathology of simple bone cyst. A: Flattened fibrous lining contains a few bone trabeculae (hematoxylin and eosin, original magnification $\times 25$). B: At higher magnification within a fibrous lining several giant cells are present (hematoxylin and eosin, original magnification $\times 100$). C: High-magnification photomicrograph shows more clearly the flat fibrous lining with scattered giant cells (hematoxylin and eosin, original magnification $\times 250$).



Fibrous dysplasia may mimic SBC, but usually it lacks trabeculation and exhibits a ground-glass, smoky appearance (see Fig. 4-35A). If a pathologic fracture occurs and the fallen fragment sign can be demonstrated, a diagnosis of a SBC is usually confirmed.



Figure 7-22 Histopathology of simple bone cyst. The cyst cavity (*left*) is bordered by an undifferentiated flat cell layer. In the loose connective tissue of the cyst wall there are extended areas of a "cloudy," loose, collagen-positive material bordered by some giant cells. Because of its collagenous characteristics the material cannot be fibrin (van Gieson, original magnification $\times 25$).

Nonossifying fibroma can usually be distinguished by its eccentric location and its well-defined, usually thick, sclerotic margin (see Fig. 7-44).

Brown tumor of hyperparathyroidism may at times appear similar to SBC, particularly if it is located in the proximal humerus or proximal femur. In this situation, one should look for other features of hyperparathyroidism, such as osteopenia and subcortical resorption. Bone abscess likewise may occasionally mimic SBC, particularly when it is located in sites of SBC predilection such as the proximal humerus or the proximal femur. However, the presence of a periosteal reaction and the frequent extension of the former lesion beyond the boundaries of a growth plate are important features indicating a bone abscess (Fig. 7-23).

Exceptionally rare, an *intraosseous ganglion* may resemble SBC (Fig. 7-24).

In the calcaneus, SBC is characteristically located at the base of the calcaneal neck on the lateral aspect of the bone, within a particular anatomic structure poor in trabeculae called Ward triangle. Most of the cysts in this location have a characteristic shape: the anterior margin is usually straight and vertically oriented, and the posterior border is typically curvilinear, paralleling the trabeculae in the posterior portion of the bone (104) (Fig. 7-25A, B). In this location, SBC must be differentiated from a *bone infarct*, an *ABC*, and several other benign bone tumors (106,136). Here are some



Figure 7-23 Brodie abscess. A bone abscess may mimic a simple bone cyst, as seen here in the proximal humerus of a 12-year-old boy. The periosteal reaction in the absence of pathologic fracture and the extension of the lesion into the epiphysis favor the former diagnosis.

helpful hints. A *bone infarct* often exhibits central calcifications, which are never present in an SBC (Fig. 7-26). *ABC* of the calcaneus is more expansive than SBC. The margin may be either poorly defined or sharp and thinly sclerotic (125) and is usually located toward the plantar and posterior aspect of the bone (107,119), although occasionally it may be found within Ward



Figure 7-24 Intraosseous ganglion resembling a simple bone cyst. An 18-year-old woman presented with left shoulder pain. Anteroposterior radiograph shows a radiolucent, trabeculated lesion in the glenoid, resembling a simple bone cyst. Excision biopsy revealed an intraosseous ganglion.

triangle. In the latter location it probably develops secondary to other lesions, such as bone infarct or an intraosseous hemorrhage. *Chondroblastoma* of the calcaneus is usually a subarticular lesion and is located in the subtalar region. It may exhibit a sclerotic, lobulated margin. Seven percent of chondroblastomas located in the calcaneus present with punctate calcifications (126). It is important to remember that Ward triangle normally contains fatty marrow, and this feature has been interpreted as an intraosseous lipoma by several authors (113,124,129). These *pseudotumors* are usually



Figure 7-25 Simple bone cyst. Lateral (A) and axial (B) views of the ankle in a 32-year-old man show a radiolucent lesion in the calcaneus. Typically, bone cysts occurring at this site are located in the anterolateral aspect of the bone.



Figure 7-26 Bone infarct. Radiolucent lesion in the region of Ward triangle exhibits central calcification resembling simple bone cyst and intraosseous lipoma.

found by serendipity, as they are not symptomatic (123,140,142) (Fig. 7-27).

Pathology

SBC is a diagnosis of exclusion, and pathologists must rely on the radiologic examination. In a long tubular bone the diagnosis is usually readily made. Most problems occur when the lesion is located in the calcaneus. In this instance, a *bone infarct* must be included in the differential diagnosis (127). The presence of characteristic microscopic features of dead bone marrow is diagnostic for the latter (Fig. 7-28).

If the membrane of the SBC contains giant cells of the osteoclastic type, *ABC* may be a consideration. However, fairly characteristic of SBC is the finding of focal deposits of an amorphous, fibrillar, cloudy eosinophilic material that stains strongly for collagen with special stains. This material resembles osteoid and may undergo calcification or even ossification (132).

The radiologic and pathologic differential diagnosis of SBC is depicted in Figure 7-29.

Aneurysmal Bone Cyst

The term aneurysmal bone cyst was first used by Jaffe and Lichtenstein (122) to describe two examples of blood-filled cysts in which tissue from the cyst wall contained conspicuous spaces, areas of hemosiderin deposition, giant cells, and occasional bone trabeculae. In a subsequent publication, Jaffe (176) chose the designation aneurysmal bone cyst as a descriptive term for this lesion to emphasize the blown-out appearance.



Figure 7-27 Pseudotumor. Radiolucent "lesion" in the region of Ward triangle exhibits no sharp margination, which is unlike the simple bone cyst, and in addition shows some bone trabeculae traversing the radiolucent area.



Figure 7-28 Histopathology of bone infarct. Necrotic remnants of fatty marrow with "oil lakes" (*center and left*) are seen in addition to bone trabeculae with empty osteocyte lacunae. A cell-rich and fiber-rich reparative connective tissue is growing in (*right*) (van Gieson, original magnification $\times 12$).

Although the cause of this lesion is unknown (159), alterations in local hemodynamics related to venous obstruction or arteriovenous fistula are believed to play an important role (155,191). Some investigators believe that the lesion is caused by a trauma (130,160,189,198). Dahlin and McLeod (162) believe that it may be similar to and related to reactive nonneoplastic processes, such as GCRG or traumatic reactions observed in periosteum and bone. ABC constitutes between 2.5% and 6% of the

primary lesions of bone with an incidence of 1.4 per million individuals (26,175,180). The lesion may arise de novo in bone, in which case no recognizable preexisting lesion can be demonstrated in the tissue, or it may be associated with various benign (e.g., GCT, osteoblastoma, chondroblastoma, chondromyxoid fibroma, fibrous dysplasia) and malignant (e.g., osteosarcoma, fibrosarcoma, or chondrosarcoma) lesions (156,177,185,192). The concept of ABC as a secondary phenomenon occurring in a preexisting lesion has been proposed by several investigators (178,181,186,187). Some investigators, however, regard ABC as a reparative process, probably the result of trauma or a tumor-induced anomalous vascular process (168,188). The typical ABC affects the medullary portion of a long bone (183). ABCs localized to the cortex (151) (see Fig. 7-36A) or in a subperiosteal location are unusual (158,202). Malignant transformation of ABC is extremely rare (179), and it is still a controversial issue. However, recent studies and cytogenetic findings permit the assumption that at least some ABCs are neoplastic in nature. A reciprocal translocation t(16;17)(q22;p13) has been detected in classic ABCs, as well as in solid extraosseous variants (163,169,197). Other chromosomal alterations have also been described, often involving chromosome 16 or 17 (149,152). Oliveira et al. (194,195) demonstrated that the translocation t(16;17)(q22;p13) initiates a fusion transcript in which the promoter region of the osteoblast cadherin 11 gene CDH11 (cloned from human osteosarcoma cell lines and involved in neoplastic processes) fuses with the entire sequence of the ubiquitin protease USP6 (which may



Figure 7-29 Radiologic and pathologic differential diagnosis of simple bone cyst.

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be involved in invasion), leading to upregulation of *USP6* by the highly active *CDH11* promoter. In a molecular cytogenetic study, Althof et al. (149) detected abnormalities of the 17p13.2 break-point region by fluorescent in situ hybridization (FISH) techniques, even in karyotypically normal ABCs, indicating that the t(16;17) (q22;p13) translocation may be useful for differential diagnostic exclusion of other tumors, e.g., telangiectatic osteosarcoma. In a comparative analysis of 52 primary and 17 secondary ABCs (associated with GCT, chondroblastoma, osteoblastoma, or fibrous dysplasia), *CDH11* and *USP6* rearrangements have not been detected in secondary ABCs, which may indicate that secondary ABCs are biologically different (reactive) lesions (196).

Clinical Presentation

ABC is seen predominantly in childhood, and 76% of cases occur in patients younger than 20 years (211). There is a slight female preponderance (200,209). The metaphysis of the long bones is the site of predilection, although the diaphysis, the flat bones, the short tubular bones (150), and even the spine may be involved (176) (Fig. 7-30). The pelvis accounts for about 50% of all flat bone lesions (212). When the spine is affected, the lesion is usually situated in the posterior elements (neural arch) (162,176,182), but the vertebral body may also be affected (209). In this location, ABC may occasionally cross the intervertebral disk to involve more than one vertebra (175). Lesions in a spinal location often produce clinical symptoms by compression of adjacent structures (the spinal cord or nerve roots) or as a result of a pathologic fracture (190,191). The most common symptoms in lesions located in the long tubular bones include pain and local swelling (161,167,190).

Imaging

The radiographic hallmark of ABC is a multicystic, eccentric expansion (blow-out) of the bone, with a buttress or thin shell of periosteal response (Fig. 7-31; see also Figs. 7-32 to 7-34). In the short tubular bones the lesion may appear more central, destroying the entire shaft of the bone (166,172). The lesion exhibits a geographic type of bone destruction with a narrow zone of transition and often a sclerotic margin (165,205) (Fig. 7-32). Sometimes ridges or trabeculations are observed within the lesion (Fig. 7-35). Rarely, internal calcifications may be present (204). A soft tissue extension is produced by bulging of the periosteum. This periosteal envelope may or may not be visible on radiography. The outer shell of bone may be partially absent (172) but, if seen, the diagnosis can frequently be ascertained. In the vertebral column the posterior elements are usually affected. Although radiographs are usually sufficient to evaluate the lesion, other modalities, such as radionuclide bone scan, CT, and MRI, can be of further assistance (Fig. 7-36).

CT is superior to radiography in defining these lesions, particularly in areas in which bone anatomy is complex (e.g., the pelvis). The cortical or periosteal shell surrounding ABC and its soft tissue extension, as



Figure 7-30 Aneurysmal bone cyst: skeletal sites of predilection, peak age range, and male-to-female ratio.



Figure 7-31 Aneurysmal bone cyst. Anteroposterior (A) and lateral (B) radiographs of the right lower leg in an 8-year-old girl show an expansive radiolucent lesion in the metaphysis of the distal tibia, extending into the diaphysis. Note its eccentric location in the bone and the buttress of periosteal reaction at the proximal aspect of the lesion.



Figure 7-32 Aneurysmal bone cyst. Anteroposterior (A) and lateral (B) radiographs of the left upper leg of a 17-year-old girl show a radiolucent lesion in the proximal fibula exhibiting a narrow zone of transition, a sclerotic margin, and a well-organized periosteal reaction.

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Figure 7-33 Aneurysmal bone cyst (ABC). Anteroposterior **(A)** and lateral **(B)** radiographs of the left proximal tibia of a 10-year-old girl show characteristic appearance of ABC, including eccentric location, expansive character, and buttress of solid periosteal reaction proximally and distally (*arrows*).



Figure 7-34 Aneurysmal bone cyst. A large, radiolucent expansive lesion in the proximal fibula of an 11-year-old girl reveals a buttress of periosteal reaction (*arrows*).



Figure 7-35 Aneurysmal bone cyst. Anteroposterior radiograph of the right shoulder of a 19-year-old woman shows an expansive, trabeculated lesion in the right clavicle. Note also a soft tissue extension.



Δ

Figure 7-36 Aneurysmal bone cyst: scintigraphy and computed tomography (CT). A: Anteroposterior radiograph of the distal femur of an 8-year-old boy with a 6-month history of pain in the lower right thigh shows a radiolucent expansive lesion located eccentrically in the femur and buttressed proximally and distally by a solid periosteal reaction. B: Radionuclide bone scan obtained after injection of 10 mCi (375 MBq) of technetium-99m-labeled methylene diphosphonate (MDP) demonstrates increased uptake of radiopharmaceutical tracer by the lesion. C: CT section shows intracortical location of the lesion, which balloons out from the lateral aspect of the femur but is contained within an uninterrupted shell of periosteal new bone.



В

well as the sharp interface between the extraosseous mass and the adjacent tissue planes, are well demonstrated with this technique (172,203). CT may also show ridges in the bony walls, described on radiographs as trabeculation or septation (172), and corresponding to the ridges found pathologically (Fig. 7-37). Attenuation coefficient values (Hounsfield units) range from 20 to 78. Fluid-fluid levels can also be demonstrated (119,171,173,174). These fluid levels are believed to represent the sedimentation of red blood cells and serum within the cystic cavities (153,210). To demonstrate this phenomenon, the patient must remain motionless for at least 10 minutes before scanning, and imaging must be performed in a plane perpendicular to the fluid levels.

Skeletal scintigraphy may occasionally be helpful because it reflects the vascular nature of the lesion (173,178). Some investigators have described an in-

creased uptake in a ring-like pattern around the periphery of an ABC (170,173,184,206). Although this phenomenon is not specific for the lesion, because it can also be observed in SBC, bone infarct, and other lesions (208), the scintigraphic findings corroborate the radiographic presentation. Hudson (173) found no correlation between the histologic features of the lesion, the amount and type of fluid contained within the cyst, and the scintigraphic pattern or intensity of uptake in his experience with 25 patients with ABC who underwent skeletal scintigraphy using technetium-99m methylene diphosphonate (^{99m}Tc-MDP) and ^{99m}Tc-pyrophosphate.

MRI findings are rather characteristic and usually allow a specific diagnosis of ABC (214). These include a well-defined lesion (Fig. 7-38), often with lobulated contours (191) (Fig. 7-39), cystic cavities with fluid-fluid levels of varying intensity (see Figs. 7-40E and 7-45B), multiple internal septations, (see Fig. 7-40C, D), and an 414 — Differential Diagnosis in Orthopaedic Oncology



Figure 7-37 Aneurysmal bone cyst: computed tomography (CT). Lateral **(A)** and oblique **(B)** radiographs of the right ankle of a 24-year-old woman show a radiolucent, trabeculated lesion in the talus. A coronal anterior CT section **(C)** and a coronal CT section **(D)** at the level of the middle facet demonstrate the internal ridges to better advantage.





Figure 7-38 Aneurysmal bone cyst (ABC): computed to-mography (CT) and magnetic resonance imaging (MRI). A: Anteroposterior radiograph of the right knee of a 9-year-old girl shows an eccentric radiolucent lesion in the medial metaphysis of the tibia abutting the growth plate. **B:** Axial CT section shows the pathologic fracture and extension of the lesion into the soft tissues (arrows), not well demonstrated on conventional radiography. **C**: Coronal T2-weighted MRI shows heterogeneous but predominantly high signal of the ABC.

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Figure 7-39 Aneurysmal bone cyst: magnetic resonance imaging (MRI). Sagittal T1-weighted (SE, TR 600, TE 16) MRI (same patient as in Fig. 7-37) demonstrates a lobulated low-signal-intensity lesion in the talus. Unlike on computed tomography examination, the septa are not well delineated.



intact rim of low-intensity signal surrounding the lesion (153,193). This rim has been described as an indicator of a benign process (213). Occasionally, small diverticulum-like projections arising in the walls of the cyst have been observed. The wide range of signal intensities within the cyst on both T1- and T2-weighted sequences is probably due to settling of degraded blood products and reflects intracystic hemorrhages of different ages (153) (Fig. 7-40).

Histopathology

Histologically, the lesion consists of multiple bloodfilled spaces alternating with more solid areas (162,207) (Fig. 7-41). These spaces are lined by a single layer of flat undifferentiated cells that react negatively with all endothelial markers (147,148). The solid tissue surrounding these spaces is composed of fibrous, richly vascular connective tissue, the blood vessels exhibiting a progression from small to large caliber (94). Mitotic figures may be observed in the fibroblasts (146,190). The fibrous lining contains many giant cells, usually in

Figure 7-40 Aneurysmal bone cyst: magnetic resonance imaging (MRI). Anteroposterior (A) and lateral (B) radiographs of the midshaft of the right femur of a 15-year-old girl show an expansive lesion arising eccentrically from the medial aspect of the bone. Note a thin shell of periosteal bone covering the lesion and a buttress of periosteal reaction at its proximal and distal extent.



Α

D



С

Figure 7-40 Continued C, **D**: Coronal T1-weighted (SE, TR 600, TE 20) MR images demonstrate heterogeneity of the lesion and internal septations. **E**: Axial T2-weighted (SE, TR 2000, TE 90) MRI shows the lesion to exhibit a high signal intensity. A fluid-fluid level, frequently seen in aneurysmal bone cyst, is well demonstrated (*arrows*).



Α

Figure 7-41 Histopathology of aneurysmal bone cyst. A: Vascular spaces are bordered with solid tissue containing spindle cells and giant cells (hematoxylin and eosin, original magnification $\times 100$). B: In another area blood-filled spaces are separated by narrow septa containing spindle fibroblasts bordering the vascular spaces with flat undifferentiated cells but without any blood vessel structures. Newly formed osteoid is also present (*lower left*) (hematoxylin and eosin, original magnification $\times 50$).

clusters (Fig. 7-42). Furthermore, lamellae of primitive woven bone may be present in the lining of the cyst as well as deeper in the connective tissue (Fig. 7-43). Deposits of iron pigment are a common finding (190).

Differential Diagnosis

Radiology

The conditions that should always be included in the differential diagnosis at any age are *SBC* and *chondromyxoid fibroma* and, after skeletal maturity, when the lesion extends into the articular end of bone, *GCT* (182). The most critical points in differentiation of ABC from *SBC* are that the former is an eccentric, expansive lesion, invariably associated with some degree of periosteal reaction (usually a solid layer or solid buttress), whereas the latter is a centrally located lesion,



Figure 7-42 Histopathology of aneurysmal bone cyst. Solid areas with densely arranged capillaries contain fairly large numbers of the giant cells osteoclast type, similar to those of a giant cell tumor. Here, however, they are grouped in clusters, are smaller, and are less numerous than in the latter lesion (hematoxylin and eosin, original magnification \times 50).

showing little if any expansion, and exhibiting periosteal reaction only when a pathologic fracture has occurred. In thin bones, such as the ulna, fibula, metacarpals, or metatarsals, the characteristic eccentricity of ABC may be lost, and, conversely, SBC may demonstrate expansive features. Because the former contains solid tissue, whereas SBC is a hollow structure filled with fluid, a fallen fragment sign (if present) is a good differential feature, pointing to the latter diagnosis. Chondromyxoid fibroma may be indistinguishable from ABC because both lesions are eccentric, are expansive, and usually affect the metaphysis, exhibiting a reactive sclerotic rim and the previously mentioned solid periosteal reaction (usually in the form of a buttress). CT and MRI are sometimes effective in making this distinction if they identify fluid-fluid levels, a phenomenon that points to the diagnosis of ABC because chondromyxoid fibroma is a solid lesion. In the mature skeleton, GCT may closely mimic ABC, although it usually is not associated with a periosteal reaction and rarely exhibits a zone of reactive sclerosis.

GCRG (so-called *solid ABC*) may be indistinguishable from the solid parts of the conventional ABC (37,157). This lesion, however, unlike true ABC, usually involves the short tubular bones of the hands and feet (34,70,97,199). The cortex is thin but is characteristically intact (164). Extension into the surrounding soft tissues is distinctly uncommon, and the periosteal reaction is usually absent (34).

Nonossifying fibroma, if it is predominantly radiolucent and is associated with a cortical expansion (a rare occurrence) (Fig. 7-44), may resemble ABC (see also Fig. 4-13). However, the former lesion never exhibits a periosteal reaction (unless a pathologic fracture has already occurred).

In smaller bones, such as the fibula, metacarpals, or metatarsals, ABC due to expansive growth may destroy the cortex, mimicking an aggressive tumor such as *osteosarcoma*. The less aggressive appearance of ABC in the small tubular bone of the hand can be mistaken for an *enchondroma* (Fig. 7-45).



Figure 7-43 Histopathology of aneurysmal bone cyst. A: Blood cyst (*bottom*) lined by a seam of flat undifferentiated cells and randomly scattered small giant cells. Noted above is formation of considerable amounts of primitive woven bone lined by osteoblasts (hematoxylin and eosin, original magnification ×40). B: Typical lamellar structure (*lower right*) is surrounded by fresh blood. Fairly extended trabeculae of primitive woven bone (*red*) are seen close to the surface of the cyst (*upper right*) (van Gieson, original magnification ×12). C: Solid areas are made up of fibrous tissue with large amounts of bone formation (*red*) and some small cysts (*upper right corner*). Significant fresh bleeding is present (van Gieson, original magnification ×12).





Figure 7-44 Nonossifying fibroma. In this 12-year-old boy a radiolucent trabeculated lesion in the proximal diaphysis of the right humerus abutting the growth plate was thought to represent either a simple bone cyst (SBC) or an aneurysmal bone cyst (ABC). Note, however, the lesion's eccentric location (bulging of the lateral cortex) that speaks against SBC, and lack of periosteal reaction that militates against ABC. On biopsy the lesion proved to be a nonossifying fibroma.

Pathology

In the pathologic differential diagnosis of ABC, an essential point to be kept in mind is the fact that, with the exception of SBC, all other entities to be differentiated may in fact contain some elements of ABC (secondary ABC engrafted on the other lesions) (Fig. 7-46). The most critical issue, however, is to distinguish ABC from telangiectatic osteosarcoma and from osteosarcoma with engrafted secondary ABC. In both of these lesions the presence of pleomorphism and of anaplastic cells is diagnostic. In telangiectatic osteosarcoma it is crucial to recognize pleomorphic tumor tissue immediately bordering the blood-filled cavity. In osteosarcoma with coexisting ABC, the benign connective tissue bordering the cavity of ABC must be distinguished from the pleomorphic tumor tissue of osteosarcoma. Atypical mitotic figures are often present in both types of osteosarcoma, and the osteoid, unlike that in ABC, has a more irregular, closely packed pattern (164).

So-called *solid ABC* should also be included in the differential diagnosis. The term, coined by Sanerkin et al. in 1983, is considered to represent a *GCRG* (67,162,201). Histologically, this is a predominantly spindle-cell lesion with a plump fibroblastic proliferation and occasional mitotic figures (175). The lesion contains a characteristic reticulated, lacy, chondroid-like material, similar to that seen in conventional ABC but without the typical



Figure 7-45 Aneurysmal bone cyst resembling enchondroma. A: Dorsovolar radiograph of the ring finger of a 6-year-old boy shows a radiolucent lesion with scalloped borders in the middle phalanx, suggesting a diagnosis of enchondroma. **B:** Ax-ial T2-weighted fat-suppressed MRI shows a fluid-fluid level (*arrow*), inconsistent with a diagnosis of a solid tumor. Excision biopsy revealed the aneurysmal bone cyst.

cyst-like cavities (178). Giant cells are numerous but are smaller and are clustered rather than being in a randomly even distribution, a characteristic feature of GCT (37). Fibrous cell proliferation is often associated with strands of irregular islands of osteoid or bone (154). Although the exact nature of solid ABC and its relation-



Figure 7-46 Osteoblastoma with secondary aneurysmal bone cyst. In the center of the lesion of osteoblastoma large blood-filled spaces are bordered by narrow septa of fibrous tissue with some bone formation (hematoxylin and eosin, original magnification $\times 25$).

ship to the solid areas of a true ABC are still not well understood (164), recent cytogenetic studies, revealing an identical t(16;17)(q22,p13) translocation in the classical and the solid variant of ABC, suggest a common origin of both lesions (163).

R

Differentiation from *SBC* may be a major problem if only small fragments of the wall are available for histopathologic diagnosis. The presence of cloudy collagenous material suggests SBC, whereas the presence of multiple blood-filled spaces suggests ABC. A *GCT* can usually be excluded by the lack of the characteristic findings for this neoplasm, i.e., the dual population of cells: mononuclear stromal cells and evenly distributed giant cells with a much greater number of nuclei (see previous discussion). One must remember, however, that there are cases of ABC containing unusually large numbers of giant cells with a greater number of nuclei (Fig. 7-47), and in such a situation there may be a problem in the histologic differentiation of these two entities.

The radiologic and pathologic differential diagnosis of ABC is depicted in Figure 7-48.

Solid Variant of Aneurysmal Bone Cyst

In 1953, Jaffe (219) reported on a nonneoplastic hemorrhagic process in bones, which he termed giant cell reparative granuloma. Subsequently, Lorenzo and Dorfman (221) reviewed the published literature up to


Figure 7-47 Histopathology of aneurysmal bone cyst resembling a giant cell tumor. Solid part of the aneurysmal bone cyst contains numerous giant cells of osteoclast type similar to the presentation of a grade 2 giant cell tumor. Distinction is based on group arrangement of the giant cells and their smaller size, as well as on the presence of blood-filled spaces (hematoxylin and eosin, original magnification \times 50).

1980 and added eight new cases, making reference to the lesion's similarity to ABC on the basis of morphology and biological behavior. In 1983, Sanerkin et al. (201) described a lesion they regarded as a "solid" variant of ABC in which the predominant histology was that of the solid components of a conventional ABC. The histopathologic appearance of this lesion was very similar to that of the lesions described by Jaffe, and by Lorenzo and Dorfman. At present, the terms "solid aneurysmal bone cyst" and "giant cell reparative granuloma" are used interchangeably. However, although only one case of GCRG has thus far been analyzed, cytogenetic findings [a t(x;4) (q22;q31.3) translocation in GCRG and a t(16;17)(q22;p13) translocation in solid ABC] do not support this view (163,216). Both lesions are considered reactive and nonneoplastic, although they can lead to a mistaken diagnosis of malignancy.

Clinical Presentation

GCRG occurs in children and adult patients, most commonly in the second and third decades, with a mean age of 18 years (218). In the reported cases,



Figure 7-48 Radiologic and pathologic differential diagnosis of aneurysmal bone cyst.

about 74% of patients were younger than 30 years at presentation. There is a slight female preponderance (1.4:1). Although these lesions are seen primarily in craniofacial and short tubular bones of the hands and feet (222), they may also occur in the long bones, such as the femur, tibia, and ulna (215,218,223). Multicentric presentations have been reported (217). Although the lesions are usually intramedullary, subperiosteal locations have been recorded (220). The patients are usually symptomatic. Some reported history of previous trauma and painful swelling at the site of the lesion comprise the most common complaints. Some lesions can be asymptomatic. Recurrence after curettage was reported as 23.5% in the Rizzoli Institute series (215) and 38.8% in the Mayo Clinic series (222).

Imaging

The radiography of GCRG shows a variably sized radiolucent lesion, predominantly expansive and frequently multilocular with internal trabeculation. The lesion's location is usually eccentric, and it may or may not extend into the articular end of bone (Fig. 7-49A). The cortex is thin and is usually intact, although an aggressive appearance with cortical destruction and soft tissue extension has been described (215). Limited matrix mineralization may be present. Pathologic

fractures are rare, and periosteal reaction is unusual. The appearance on CT is nonspecific, with attenuation values similar to that of muscle (63). On MRI the lesion appears to be solid and exhibits a signal intensity slightly higher than that of muscle on T1-weighted images and a heterogeneous, predominantly high signal on T2 weighting (218) (Fig. 7-49 B, C). Cystic areas within the lesion and fluid-fluid levels are sometimes present.

Histopathology

The histologic features of GCRG include a mixture of collagen, plump bland fibroblasts, and multinucleated giant cells surrounding foci of hemorrhage. Commonly observed are foci of osteoid production along hemorrhagic foci (Fig. 7-50).

Differential Diagnosis

Radiology

The main differential diagnosis should include conventional *ABC*, *GCT*, and *enchondroma*. *ABC* should be considered, particularly in children, when the lesion of GCRG is eccentric and expansive. Lack of periosteal reaction favors the latter diagnosis. In addition, MRI demonstrates a solid lesion, unlike the conventional ABC, with occasional foci of mineralization. *GCT* should



Figure 7-49 Solid variant of aneurysmal bone cyst: magnetic resonance imaging (MRI). A: Oblique radiograph of the left ankle of an 11-year-old girl shows a sharply marginated radiolucent lesion in the metadiaphysis of the tibia. **B:** On coronal T1-weighted MRI the lesion exhibits intermediate heterogeneous signal.



Figure 7-49 *Continued* **C:** On axial T2-weighted MRI the lesion exhibits heterogeneous but predominantly high signal intensity.

be considered, particularly in skeletally mature patients, when the lesion extends into the articular end of the bone. The presence of a sclerotic margin will favor GCRG. Conversely, the absence of mineralization of the matrix will suggest a GCT. Aggressive features of the lesion will be more compatible with GCT because aggressive radiologic features are rarely encountered in a GCRG (215). In the small bones of the hand and feet, particularly when the lesion is central in location, *enchondroma* should be strongly considered. The presence of a periosteal reaction without pathologic fracture and pseudotrabeculation would exclude this diagnosis. In a subperiosteal location, GCRG should be differentiated from *periosteal chondroma* (218).

Pathology

As with the radiologic differential diagnosis, the histologic appearance of GCRG is similar to that of GCT and ABC. GCT can easily be excluded if a dual population of mononuclear stromal cells and of uniformly distributed giant cells, characteristic for this tumor, are not present. In addition, the tissue of GCT usually exhibits less fibrosis, and lacks osteoid formation and mineralization of the matrix. Moreover, the giant cells of GCRG are smaller and are not evenly distributed throughout the lesion as in GCT but instead are arranged in groups (see Fig. 7-50). Hemosiderin deposition is more prominent in the former lesion. ABC shares many similar histologic features with GCRG, particularly its solid part. However, it contains a significant number of bloodfilled spaces, a finding that is not present in GCRG. The other lesions to be considered in differential diagnosis are brown tumor of hyperparathyroidism and low-grade central osteosarcoma. The histologic appearance of the former condition is similar to that of GCRG. However, laboratory data (serum calcium and phosphorous levels) will invariably point to the correct diagnosis. Lowgrade central osteosarcoma will exhibit a greater amount of collagen and bone formation than is seen in GCRG. If nuclear atypia and atypical mitoses are present, and the stromal cells invade the marrow spaces and trabeculae of cancellous and cortical bone, the diagnosis of osteosarcoma is certain.



Figure 7-50 Histopathology of solid variant of aneurysmal bone cyst (giant cell reparative granuloma). A: Multinucleated giant cells are accompanied by inflammatory cells and hemosiderin-loaded macrophages in a spindle-cell stroma. Note irregular bluish chondroosteoid (*right, center*) (hematoxylin and eosin, original magnification $\times 100$). B: At higher magnification, observe irregularly distributed giant cells in close proximity to bluish chondroosteoid. Inflammatory cells and spindleshaped stromal cells are also present (hematoxylin and eosin, original magnification $\times 200$).





The radiologic and pathologic differential diagnosis of GCRG is shown in Figure 7-51.

Intraosseous Lipoma

Lipomas can be categorized according to their location in the bone as intraosseous, cortical, or parosteal lesions (224,225,261). In the older (232,259,260,262) as well as in more recent (82) publications, intraosseous lipoma is considered to be an extremely rare tumor, although normal fatty tissue is so abundant in bone. Most cases of so-called intraosseous lipomas were considered to represent a simple hyperplasia of fat tissue rather than a true tumor (262). Particularly, socalled vertebral lipomas, which have been found in 0.6% of a large series of autopsy material, are considered to represent merely foci of fatty marrow (239,262). Also in the current literature, intraosseous lipoma is considered to be extremely rare, with an incidence of less than 1 in 1,000 primary bone tumors (82,230). Fewer than 200 cases have been reported (229, 248, 249, 256, 265).

To date, only one parosteal lipoma has been analyzed cytogenetically, revealing a t(3;12)(q28;q14)translocation similar to those observed in subcutaneous lipomas, which often show rearrangements of 12q13–15 and chromosome 3 (227,251). These rearrangements involve two genes, the *HMGIC* gene at 12q15 (*h*igh-*m*obility group *i*soform *c* gene, a member of the high-mobility group genes, representing an architectural transcription factor), and the LPP gene at 3q27–28 (*l*ipoma *p*referred *p*artner gene, believed to be involved in cell adhesion), leading to a fusion transcript, HMGIC/LPP, that can be detected by reverse transcriptase polymerase chain reaction (RT-PCR) specifically in lipomas (253).

In recent years, there have been an increasing number of reports of intraosseous lipoma, particularly that located in the intertrochanteric and subtrochanteric regions of the femur and in the calcaneus (113,124, 127,228,249,255). The authors of these publications apparently do not take into account the fact that in describing lesions in areas scanty of bone trabeculae but rich in fatty tissue, most of the time they are dealing with normal bone structures of the particular regions, known as Ward triangle in the anatomical (235) and in the radiologic literature (123,226,237,240,243,263). The calcified lipomas and bone cysts that have been described, particularly those located in the calcaneus, are lesions secondary to an infarct (82) and have probably developed because of the special anatomic arrangement of the vascular supply in and around the calcaneus (136,241); they do not represent true tumors.

Clinical Presentation

The tumor has no sex predilection and occurs in patients from 5 to 75 years old (106,249). It is usually an asymptomatic lesion, found on radiographic examinations performed for other reasons (250). Some investigators report a higher incidence of symptomatic patients (257); however, even when a patient is symptomatic, the symptoms are not necessarily related to the lesion. In the largest series to date, of 61 intraosseous lipomas (249), the most common site was the inter- and subtrochanteric region of the femur, followed by the calcaneus, ilium, proximal tibia, and sacrum (266). As is clear from the previous discussion, because the intertrochanteric area and Ward triangle in the calcaneus are inherent sites of a prominent amount of fatty tissue, the presence of intraosseous lipomas at these sites must be questioned.

Imaging

Intraosseous lipoma has a rather characteristic radiographic appearance. It is invariably a nonaggressive radiolucent lesion with sharply defined borders, associated with thinning and bulging of the cortex particularly in thin bones such as the fibula or rib (231,232, 234,238) (Fig. 7-52). The central calcifications and ossifications described in a very large number of cases (236,244) invariably represent a sign of fat necrosis that is seen in an infarct of the fatty marrow. CT may be helpful in the diagnosis of these lesions because the Hounsfield units are consistent with fat (254,257). MRI shows on T1- and T2-weighted images that the lesion has a signal similar to that of subcutaneous fat (225,247) (Fig. 7-53). After administration of intravenous gadolinium there is no enhancement of the lesion (248). MRI is very effective in demonstrating the exact intraosseous extension of the lesion.

Histopathology

Histologically, intraosseous lipomas are composed of lobules of mature adipose tissue and are characterized by the presence of mature lipocytes, which are slightly larger than nonneoplastic fat cells, in a background of fibroblasts with occasional foci of fat necrosis (245,257). A capsule may occasionally encompass all or part of the tumor mass (246) (see Fig. 7-52C), and in most cases reported, atrophic bone trabeculae are found throughout the lesion. In the nontumorous fatty marrow contained in the previously mentioned segments in the femur and the calcaneus, infarcts may arise for yet unknown reasons, although some investigators believe that circulatory abnormalities are the last link in the pathogenetic chain (258). The necrotic fat cells may calcify, and then ossify, and secondary bone cysts may form in the necrotic region (252). Usually, the calcifications exhibit a birch-broom-like picture, but dense aggregates of calcification can exhibit a Liesegang ring-like configuration (234).

Differential Diagnosis

Radiology

Because the MRI appearance of intraosseous lipoma is so characteristic, with this technique there is essentially no possibility of making a mistake in rendering this diagnosis (233). The same is true if the fatty marrow becomes necrotic such as in *medullary bone infarct*. In the latter lesion, however, the necrotic tissue exhibits a characteristic peripheral rim of fibrosis, separating the lesion from the viable bone. Occasionally, however, a *secondary cyst* may develop in the medullary bone infarct (252) that may mimic an intraosseous lipoma(Fig. 7-54). A case of an *SBC* has recently been described in which an intraosseous cavity partly filled with fat after involution of the cyst (264). The imaging appearance of this fatty replacement within the intraosseous defect resembled that of an intraosseous lipoma. So-called intraosseous lipomas of the calcaneus usually represent bone infarction, SBC, ABC, or pseudocyst in the fatty marrow of the normally radiolucent Ward triangle (136).

Pathology

Like the radiologic appearance, the histopathology of intraosseous lipoma is almost specific. The differential diagnosis becomes complex if there is a necrosis of the marrow presenting as a *bone infarct*, in which case the lesions characteristically exhibit necrotic marrow and empty osteocyte lacunae.

Another differential possibility is *intraosseous liposarcoma*. This extremely rare lesion is characterized by the pleomorphic appearance of the neoplastic cells with varying degrees of lipogenic differentiation. Signet ring cells are usually present (224,242), with a large, single globule of lipid filling the cytoplasm (234). Evidence of malignancy is the finding of lipoblasts which show a rather broad seam of cytoplasm with numerous fat vacuoles and an indented, somewhat hyperchromatic, nucleus.

The other entities that may be considered in the differential diagnosis are either primary or secondary *ABC* and *SBC*. If an ABC or an SBC develops in the area of bone infarction (252), the primary necrosis of the bone marrow may be covered by the secondary lesion.

The radiologic and pathologic differential diagnosis of intraosseous lipoma is depicted in Figure 7-55.

Fibrocartilaginous Mesenchymoma

Fibrocartilaginous mesenchymoma is an extremely rare tumor composed of two distinct tissues, one benign and cartilaginous, resembling an active growth plate, and the other resembling a low-grade fibrosarcoma (269). Mirra et al. (272) classify this lesion as desmoid tumor with enchondroma-like nodules. The number of reported cases is probably less than 20, although there may exist several unpublished cases (Remagen, personal communication).

Clinical Presentation

Fibrocartilaginous mesenchymoma has been reported in patients ranging from 9 to 23 years of age (mean age 13 years). Males are more frequently affected. The lesion is located usually in the epiphysis of a long bone, such as the fibula or humerus. The symptoms usually indicate a slow-growing tumor. They consist of slight discomfort and tenderness at the site of the lesion, and occasionally a palpable mass.

Imaging

On radiography, the lesion is radiolucent with scalloped borders, extending to or abutting the growth plate. After skeletal maturity the lesion may extend into the articular end of bone (Fig. 7-56). Occasionally the cortex is expanded and thinned. The cortex may be invaded, and in these cases the lesion extends into the soft tissues



Figure 7-52 Intraosseous lipoma. A: Anteroposterior radiograph of the left ankle shows a radiolucent lesion in the distal fibula exhibiting expansive character, thinning of the cortex, and central calcifications. (Reprinted from Bullough PG. *Atlas of orthopedic pathology,* 2nd ed. New York: Gower, 1992:16.20.) **B:** Lateral radiograph of the knee of another patient with intraosseous lipoma shows a radiolucent sharply defined lesion in the proximal tibia. **C:** Gross anatomic specimen of the tibia shows the lesion surrounded by a thin fibrous capsule. **D:** Whole-mount section shows replacement of trabecular bone by lipoma exhibiting central flocculent calcifications (*upper right*) (toluidine blue).



Figure 7-53 Intraosseous lipoma: magnetic resonance imaging (MRI). A: Anteroposterior radiograph of the right ankle in a 42-year-old man shows a radiolucent lesion in the distal tibia sharply delineated by a thin sclerotic margin (*arrows*). **B:** On the lateral radiograph there is a suggestion of a faint calcific body in the center of a radiolucent lesion (*arrowhead*). **C:** Coronal T1-weighted (SE, TR 685, TE 20) MRI demonstrates the lesion to be of a high signal intensity paralleling that of subcutaneous fat. A small focus of low signal is present within the lesion (*arrowhead*). **D:** Axial T2-weighted (SE, TR 2000, TE 70) MRI shows that the lesion becomes of lower intensity, again paralleling the signal of subcutaneous fat.



Figure 7-54 Cyst formation in bone infarction. Anteroposterior radiograph of the left humerus in a 31-year-old woman shows an expansive radiolucent lesion in the proximal shaft with central calcifications typical for bone infarction and a rim of reactive sclerosis. (Courtesy of Dr. Alex Norman, New York, New York)



Figure 7-55 Radiologic and pathologic differential diagnosis of intraosseous lipoma. Figure 7-56 Fibrocartilaginous mesenchymoma. Anteroposterior (A) and lateral (B) radiographs of the right knee of a 23-year-old man show a radiolucent trabeculated lesion in the proximal tibia, bulging the anterolateral cortex and extending into the articular end of the bone.



(Fig. 7-57). This can be effectively demonstrated with CT and MRI. Although a periosteal reaction is usually absent, when present it is sparse and of benign appearance. The tumor may contain visible calcifications typical of cartilaginous matrix (see Fig. 7-57B).

Histopathology

By microscopy, the lesion is composed of a tissue made up of intersecting bundles of spindle cells and collagen fibers. The tissue is fairly cellular, the nuclei are plump, and there is evidence of pleomorphism and hyperchromatism, with occasional mitotic figures (269). Superimposed on this background are well-defined islands of obviously benign cartilage, varying in density from case to case, from a cartilage island here and there to densely packed, bowed bands of cartilage that resemble the epiphyseal growth plate, with characteristic gradual columnation, cellular hypertrophy, and endochondral ossification (269) (Fig. 7-58). On H&E staining, this gives the tumor a very typical "shrimp cocktail" appearance, with the white cartilage ("shrimps") lying in the red spindle-cell tumor ("sauce") (Fig. 7-59). In other cases, the cartilage more closely resembles the islands of an enchondroma. Furthermore, variable amounts of detached trabeculae of woven bone similar to fibrous dysplasia may be found (Fig. 7-60). Therefore, these cases are considered to be an entity of their own, related to fibrous dysplasia and called "fibrocartilaginous dysplasia" (267,271). However, we have observed at least one case where fibrous dysplasia-like structures were combined with the presence of epiphyseal plate-like cartilage, not allowing that distinction (Remagen, unpublished observations, 1997). In its first description, the tumor was named fibrocartilaginous mesenchymoma with lowgrade malignancy (270). However, because metastases have never been observed thus far, the group at the Mayo Clinic later deleted that addition (268), calling it, simply, fibrocartilaginous mesenchymoma.

Differential Diagnosis

Radiology

Because the lesion is metaphyseal and often arises in a growing skeleton, benign lesions such as *ABC* and *chondromyxoid fibroma* should be included in the differential diagnosis. After obliteration of the growth plate, *GCT* and *chondrosarcoma* must be considered. Other tumors that may have a similar appearance include *desmoplastic fibroma*, *fibrosarcoma*, *malignant fibrous histiocytoma*, and *osteosarcoma*.

Pathology

The histologic differential diagnosis must include *fibrosarcoma, chondrosarcoma,* particularly mesenchymal and dedifferentiated variants, *fibrocartilaginous dysplasia* (fibrous dysplasia with cartilaginous metaplasia), and *desmoplastic fibroma* (Fig. 7-61).

The presence of the benign cartilaginous component allows differentiation from fibrosarcoma. This is also true for chondrosarcoma and the listed variants. In fibrous dysplasia, the spindle-cell stroma has a benign aspect and the cartilaginous component resembles an enchondroma, in contrast to fibrocartilaginous mesenchymoma, in which the cartilaginous component resembles the growth plate.

The radiologic and pathologic differential diagnosis of fibrocartilaginous mesenchymoma is depicted in Figure 7-62.



Figure 7-57 Fibrocartilaginous mesenchymoma: computed tomography (CT) and magnetic resonance imaging (MRI). A: Oblique radiograph of the left knee of a 14-year-old boy shows an osteolytic trabeculated lesion in the distal femur abutting the growth plate. The lateral cortex is destroyed. B: CT section through the tumor shows destruction of the posterolateral cortex and a large soft tissue mass containing calcifications. C: Coronal T1-weighted MRI shows heterogeneous signal of the tumor that violated the growth plate and extended into the distal femoral epiphysis. D: Axial T1-weighted MRI shows destruction of the cortex and a large soft tissue mass of intermediate signal. Calcifications within the mass display a low signal intensity. E: On axial T2-weighted MRI image the tumor becomes of high signal intensity. Pseudoseptation of the mass and its heterogeneous character are well demonstrated.



CONSISTENCE OF CONSIS

Figure 7-58 Histopathology of fibrocartilaginous mesenchymoma. Typical aspect of the tumor consists of two components: cartilage structured similar to the growth plate, and surrounding spindle-cell tissue exhibiting subtle to moderate pleomorphism and some collagen formation (van Gieson, original magnification $\times 12$).

Figure 7-59 Histopathology of fibrocartilaginous mesenchymoma. Numerous curved cartilage particles lie in scanty spindle-cell tissue, giving the tumor a "shrimp cocktail" appearance (hematoxylin and eosin, original magnification $\times 0.2$).



Figure 7-60 Histopathology of fibrocartilaginous mesenchymoma. In the areas free of cartilage the tumor resembles fibrous dysplasia because of formation of deformed Chinese character–like immature bone trabeculae (van Gieson, original magnification $\times 12$).



Figure 7-61 Histopathology of fibrocartilaginous mesenchymoma. In the areas free of cartilage and bone the tumor clearly resembles a desmoplastic fibroma; however, pleomorphism of the cells and nuclei is more pronounced (hematoxylin and eosin, original magnification $\times 10$).



Figure 7-62 Radiologic and pathologic differential diagnosis of fibrocartilaginous mesenchymoma.

Malignant Tumors

Adamantinoma of Long Bones

Adamantinoma is a rare malignant tumor, comprising about 0.4% of all bone tumors. It is characterized by the formation of apparently epithelial cells surrounded by a spindle-cell fibrous tissue (82,290). The tumor owes its name (given by Fischer in 1913) to its close morphologic resemblance to ameloblastoma of the jaw, which formerly was called adamantinoma (the name was later changed because no adamantine enamel is produced by the tumor cells) (283). The histogenesis of adamantinoma has long been debated. A number of authors have advocated an endothelial (291,299) or epithelial (306,314) derivation. However, recent advances in electron microscopy and IHC have provided strong evidence in support of an epithelial origin (290,293,312).

Clinical Presentation

Adamantinoma occurs with equal frequency in males and females between the second and fifth decades and has a predilection for the tibia in 90% of cases (275). Rarely, it can involve the fibula (304), the humerus, or the ulna (Fig. 7-63). The lesion is usually confined to the middle or distal third of the tibia, but a simultaneous occurrence in tibia and fibula has occasionally been reported (280,314). A history of trauma is obtained in approximately two thirds of cases (312,314). The most common presenting complaint is localized swelling or enlargement of the affected bone (302,303). Pain is mild at onset and becomes more severe as the size of the lesion increases. Physical examination notes a firm, tender mass or swelling, usually affixed to the underlying bone (305).

Imaging

The radiographic appearance of adamantinoma is that of well-delineated, elongated osteolytic defects of various sizes, separated by areas of sclerotic bone, occasionally having a soap-bubble appearance (Fig. 7-64A). A saw-toothed area of cortical destruction, when present, is quite distinctive for this tumor (Fig. 7-64B). One study noted that 80% of all lesions were greater than 5 cm in length (296). At times, the entire bone is involved by multiple satellite lesions. Some long-standing lesions may exhibit a more sclerotic, ground-glass appearance and therefore can be confused with fibrous dysplasia (Fig. 7-65). Occasionally, adamantinoma manifests as a single, large lytic lesion in the cortex, which expands and destroys the cortex and produces a soft tissue mass (116).

On MRI examination, a hypointense (compared to the normal bone marrow) signal on T1-weighted images and a hyperintense signal on T2-weighted images are noted. After intravenous administration of gadoliniumdiethylenetriamine-penta-acetic (Gd-DTPA), some investigators reported no significant enhancement of the lesion (319), whereas others (315) noted intense enhancement. The main purpose of MRI is not to diagnose the tumor but to determine the extent of intraosseous and extraosseous involvement (116).



Figure 7-63 Adamantinoma: skeletal sites of predilections, peak age range, and male-to-female ratio.



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Figure 7-64 Adamantinoma. A: Lateral radiograph of the left leg in a 64-year-old woman shows a destructive lesion in the midshaft of the tibia. The lesion is multiloculated and slightly expansive, with mixed osteolytic and sclerotic areas creating a soap-bubble appearance. **B:** Lateral radiograph of the right leg of a 28-year-old woman shows multiple, confluent lytic lesions involving almost the entire tibia. The articular ends are spared. The anterior cortex exhibits a predominantly saw-tooth type of destruction.



Figure 7-65 Adamantinoma. A single, predominantly sclerotic lesion in the midshaft of the tibia of this 20-year-old man was thought to represent fibrous dysplasia. A biopsy revealed adamantinoma.

Scintigraphy invariably reveals an increased uptake of the radiopharmaceutical tracer (298,312) (Fig. 7-66).

Histopathology

Histologically, the tumor is biphasic and consists of an epithelial component intimately admixed in various proportions with a fibrous component (Fig. 7-67A) (309,317). The epithelial component, usually consisting of islands of polyhedral cells in pavemented nests or in gland-like spaces, exhibits peripheral nuclear palisading and a looser, myxoid inner zone that resembles the stellate reticulum of odontogenic tissue. The palisading and columnar arrangement of the peripheral cells, accompanied by stellate central cells, somewhat resembles an ameloblastoma of the jaw (273) (Fig. 7-67B). Epithelium-like cells may have a variety of patterns (Fig. 7-67C,D). There are four classic forms, found in a variety of combinations: tubular, spindle, squamous, and basaloid, as well as a fifth form that is termed an osteofibrous dysplasia (OFD)-like pattern because of its similarity to OFD (281,286,290,310). The latter may contain only small clusters or even single dispersed epithelial cells (316). The cells of adamantinoma are strongly marked by antibodies to keratin (Fig. 7-67E,F), an IHC property that helps to distinguish this tumor from other similar lesions, such as OFD, fibrous dysplasia, or nonossifying fibroma (293,316). In particular, the differentiation between OFD-like adamantinoma and OFD is often impossible without cytokeratin IHC. Squamous metaplasia is sometimes present and, if extensive, the lesion may be erroneously interpreted as a metastasis of a squamous cell carcinoma. The fibrous component consists of spindle cells, which usually exhibit only slight atypia. The epithelial spaces and stroma sometimes become highly vascularized, and the lesion may resemble a vascular neoplasm. However, ultrastructural and IHC evidence points toward an epithelial nature of adamantinoma (282,297,306,307, 318). A coexpression of α -smooth muscle actin and cytokeratins in epithelial tumor cells has been described in some cases (293). The lesion may contain spicules of bone and may exhibit cystic cavities that contain blood or a straw-colored fluid (116).

A relationship of adamantinoma to OFD (Kempson-Campanacci lesion) and fibrous dysplasia has been suggested by some investigators because adamantinoma may contain a fibroosseous component that, on pathologic examination, resembles both fibrous dysplasia and OFD (281,286,292,294,310,311,313,317). Cytogenetic studies have provided evidence that similar clonal aberrations (trisomies 7, 8, and 12) exist in OFD and adamantinoma and are distinct from those described in one case of fibrous dysplasia (277,289). Czerniak et al. (281) suggested that classical adamantinoma may regress by a reparative process with OFD-like features leading to OFD-like or, as they termed it, differentiated adamantinoma. However, other authors have regarded OFD as a precursor lesion of adamantinoma on the basis of clinical course and of molecular and IHC analyses (276,285,286,300,310). Kanamori et al. (295) demonstrated that genetic aberrations in adamantinomas are more complex than those in OFD, possibly indicating a progression from OFD to adamantinoma. In addition, Hazelbag et al. (288) suggested that epithelial cells in adamantinoma may transform from osteofibrous tissue, based on an analysis of extracellular matrix (ECM) components. This view was supported by Maki and Athanasou (300), who analyzed factors involved in the mesenchymal-epithelial transition. Bovée et al. (276) demonstrated proliferative capacity only in the epithelial parts of classic and OFD-like adamantinomas but not in the stromal component. Furthermore, TP53 aberrations and DNA aneuploidy were restricted to the epithelial component of all investigated adamantinomas. These features are obviously responsible for the malignant behavior of the tumor. Despite these findings, it is still difficult to imagine how an epithelial neoplasm might arise as a primary in bone.

Differential Diagnosis

Radiology

The main differential diagnosis includes *fibrous dysplasia*, *OFD*, and *osteomyelitis*. *Fibrous dysplasia* is less aggressive looking and, unlike adamantinoma that predominantly involves the anterior cortex, the former is more centric in location. Moreover, osteolytic



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Figure 7-66 Adamantinoma: scintigraphy. A: Anteroposterior radiograph of the left leg in a 46-year-old woman shows multiple radiolucent lesions in the midshaft of the tibia. The lateral cortex is slightly thickened. **B:** Lateral radiograph shows a mixed sclerotic and osteolytic lesion predominantly affecting the anterior tibial cortex. Frontal **(C)** and lateral **(D)** radionuclide bone scans obtained after injection of 20 mCi (740 MBq) of ^{99m}Tc-labeled methylene diphosphonate (MDP) show markedly increased uptake of radiopharmaceutical tracer in the lesion.



Figure 7-67 Histopathology of adamantinoma. A: Admixed within a spindle-cell fibrous stroma is an island of small epithe-

Figure 7-67 Fistopathology of adamantinoma. A: Admixed within a spinole-cell hibrors stroma is an island of small epithe-lial cells containing an extended area of squamous cell metaplasia (hematoxylin and eosin, original magnification ×50). **B:** In another field of view, observe ameloblastoma-like areas with peripheral palisading of epithelial tumor cells (hematoxylin and eosin, original magnification ×200). **C:** In another area observe the spindle-cell pattern (hematoxylin and eosin, original mag-nification ×200). **D:** At high power, tumor cells are densely packed and exhibit oval to spindle-shaped nuclei (hematoxylin and eosin, original magnification ×400). **E:** Epithelial tumor cells are positive for cytokeratins [peroxidase-antiperoxidase (PAP) method, original magnification ×200]. **F:** Densely packed spindle-shaped tumor cells are positive for cytokeratin 5 (PAP method, original magnification ×200).

cortical defects are almost never encountered in fibrous dysplasia (301).

OFD is a lesion of infancy and childhood, whereas adamantinoma [with a few exceptions (319)] is never seen before skeletal maturity. The lesion of OFD is usually well defined, without the destructive features of adamantinoma, although, albeit rarely, larger lesions may destroy the cortical bone and invade the medullary cavity (see Fig. 4-50).

Osteomyelitis of the tibia may at times be difficult to distinguish from adamantinoma. The presence of a periosteal reaction and sequestra in the former may point to the correct diagnosis (Fig. 7-68).

Nonossifying fibroma is a lobulated, well-demarcated lesion with a sclerotic border, showing no evidence of cortical invasion, whereas a *fibrous cortical defect*, when it reaches the zone of metaphyseal diminution, may mimic a small focus of adamantinoma.

Desmoplastic fibroma may present as an aggressive lesion. It frequently exhibits internal septation or trabeculation, features not typical of adamantinoma.

Very small intracortical lesions may occasionally mimic a *Brodie abscess, osteoid osteoma*, or even a *stress fracture* (319).

Pathology

The most important point is to distinguish this lesion from OFD (279). The latter lesion does not possess the epithelial component within the fibro-osseous stroma that is characteristic of adamantinoma. Immunohistochemically, OFD, in contrast to adamantinoma, does not show any keratin-positive cells. However, more problems are encountered in distinguishing so-called OFD-like adamantinoma from conventional OFD (310). Moreover, in recent years, patients have presented with lesions that contained foci of epithelial tissue corresponding to adamantinoma within areas of OFD (274,281) (Fig. 7-69). Czerniak et al. (281) have termed such lesions "differentiated (regressing) adamantinomas." According to these authors, features characteristic of differentiated adamantinomas include onset during the first 2 decades of life, an exclusively intracortical location, uniform predominance of OFD, and scattered foci of cytokeratin-positive epithelial elements that are identical to those observed in classic adamantinoma. Other authors, however, question this concept because recurrences of OFD-like adamantinoma may progress to classic adamantinoma and may even metastasize (287,290). According to the WHO, cytokeratin-positive epithelial cells in a lesion with OFDlike features should lead to the diagnosis of OFD-like adamantinoma (316).

Fibrous dysplasia and *nonossifying fibroma* can usually be excluded by the immunohistochemical demonstration of the epithelial component of adamantinoma, which may consist of a very few epithelial cells or very small islands looking like blood vessels but which are positive for keratin markers (308).

If squamous metaplasia is present, and particularly if it is extensive, adamantinoma may be mistaken for *squamous cell carcinoma*. This is the only differential in which the epithelial and keratin markers are not helpful. In such cases, the absence of the fibrous stroma, the age of the patient, and radiologic differences should be considered.

Rarely, a *Ewing sarcoma* may be cytokeratin-positive ("adamantinoma-like Ewing sarcoma"), a point that raises differential diagnostic problems. However, Ewing sarcomas are positive for cytokeratins 8 and 18, which are consistently lacking in adamantinomas. In addition, the typical Ewing fusion transcripts *EWS/FL11* and *EWS/ERG* are absent from adamantinoma (278,284).

The radiologic and pathologic differential diagnosis of adamantinoma is depicted in Figure 7-70.

Chordoma

Chordoma is a rare malignant bone tumor that arises from developmental embryonic remnants of the notochord (which disappears during the second month of embryonal life) (373). This tumor is characterized by a lobular arrangement of the tissue with vacuolated socalled physaliphorous cells and mucoid intercellular material (346,380). Chordoma accounts for about 1% of all malignant bone tumors. The tumor occurs only at sites in which notochordal remnants are not normally present. Therefore, although it is a midline lesion of the axial skeleton, it never occurs in the nuclei pulposi.

Genetically, most of the investigated chordomas are hypodiploid or diploid with chromosome segments 1p31-pter, 3p21pter, 3q21-qter, 9p24-pter, and 17q11-qter most often affected. Gains of chromosomes 5q, 7q, and 20 have been reported by comparative genomic hybridization (CGH) investigation. A putative tumor suppressor gene was suspected at 1p36 in familial and sporadic chordomas by LOH analysis (359,361). In clivus chordomas, expression of the mRNA of human telomerase reverse transcriptase (an enzyme involved in telomere lengthening and thus in "immortalization" of cells) is correlated with fast tumor growth rates (363).

Clinical Presentation

The lesion is most frequently discovered between the fifth and seventh decades (mean age 56 years) and males are more commonly affected (M/F ratio 2:1) (343). The predilection of chordoma for the spinal axis is well documented (331–333). The three most common sites for this tumor are the sacrococcygeal area (55%), the sphenooccipital area, mainly the clivus (35%) (347), and the second cervical vertebra (8%)(Fig. 7-71). Chordoma is a locally aggressive lesion (321,342,343). In most series reported, metastases occurred usually late in the course of the disease (322). The incidence of reported distant metastases ranges from 10% to 43% (345). Because chordoma usually grows slowly, often the symptoms have been present for a year or more (344). The symptoms are closely related to the location of the lesion. The usual presenting complaint is sharp or dull pain affecting the lower back or the cervical spine, sacrum, or coccyx (323). Sphenooccipital lesions give rise to headache, cranial nerve palsies, and signs of endocrine dysfunction caused by



Figure 7-68 Osteomyelitis resembling adamantinoma. Anteroposterior **(A)** and lateral **(B)** radiographs of the right leg of a 55-year-old man with chronic bone infection show sclerotic changes of the proximal tibia and thickening of the anterior cortex that mimics adamantinoma. Sagittal T1- **(C)** and T2-weighted **(D)** magnetic resonance images (MRI) demonstrate small sequestra and intraosseous sinuses characteristic of osteomyelitis.

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Figure 7-69 Histopathology of adamantinoma with osteofibrous dysplasia-like component. A: Resected fragment of the anterior tibia shows irregular thickening and thinning of the cortex associated with metaplastic new bone formation (hematoxylin and eosin, reduced scale). B: Fibrous tissue of various density contains an island of adamantinoma (middle third). Metaplastic new bone formation (lower right) is seen; note also the inner limit of cortical bone with signs of remodeling (right) (hematoxylin and eosin, original magnification ×16). C: At higher magnification the tubular and solid structures of epithelial cells are conspicuous. Slight hyperchromatism and pleomorphism of the nuclei are present. Some capillaries (right middle and top) have to be differentiated from the epithelial structures (hematoxylin and eosin, original magnification ×200). **D:** Immunocytochemical marking of epithelial cells (brown) with a panepithelial marker displays groups and single positive cells (lower right) lying in the fibrous tissue [pan-cytokeratin antibody Lu5 peroxidase-antiperoxidase (PAP) method, original magnification ×100].

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Figure 7-71 Chordoma: skeletal sites of predilection, peak age range, and male-to-female ratio.

compression of the pituitary gland (365). Lesions of the cervical vertebrae usually cause cord compression. Lesions of the sacrococcyx produce increasing lower back pain, anorectal or bladder dysfunction, and paresthesias (336,366).

Imaging

The radiographic appearance of chordoma is that of a highly destructive, expansive lytic lesion, with irregular scalloped borders and occasionally with matrix calcifications, which are probably caused by extensive tumor necrosis (370) (Figs. 7-72 and 7-73). Sclerosis of the tumor edges may be observed, and a pathologic fracture is often present (362). Soft tissue masses are commonly associated with the lesion (337).

Although conventional radiography, including tomography, delineates the tumor well, CT and MRI are important to demonstrate the soft tissue extension, and myelography often demonstrates an invasion of the spinal canal (381). CT can reveal the extent of bone destruction, the presence of a soft tissue mass, growth within the spinal canal, and invasion of adjacent structures (348,357). The MRI characteristics (372), which include a low or intermediate signal intensity on T1-weighted images and a high signal on T2-weighted images (328), are not specific for chordoma (362,367). However, a very high signal intensity on T2 weighting, combined with a lobulated appearance of the tumor, suggests the diagnosis of chordoma (Fig. 7-74).

Scintigraphy reveals an increased uptake of radiopharmaceutical tracer around the periphery of the tumor. Areas of abnormally decreased activity due to complete replacement of bone by the tumor may also be observed (345). Lack of uptake of the tracer within the tumor itself is probably secondary to the absence of vascularity and the lack of ossification (345).

Histopathology

Chordoma may demonstrate considerable variations in its histologic appearance (327,329,364). Apart from the degree of differentiation, the histology is also determined by the extent of anaplasia. At present, on the basis of microscopic appearance, the tumor is divided into three subtypes: conventional, chondroid, and dedifferentiated (336).

Conventional chordoma microscopically resembles the fetal notochord. The tumor consists of cord-like arrays and lobules of large polyhedral cells with a vacuolated cytoplasm and vesicular nuclei (Fig. 7-75A–C), referred to as physaliphorous (from Greek for "bubble-bearing") (341). The vacuoles contain a mucinous substance with neutral mucopolysaccharides (320,362) and a mixture of weakly sulfated and carboxylated glycoproteins (376). The mucinous material is either secreted into the intercellular space or freed on the death of cells. In addition, cells of the signet ring type, in which the nucleus is displaced to one side by one or two vacuoles, are also observed (346). Some cells have an eosinophilic, nonvacuolated cytoplasm (336). Nuclear pleomorphism is usually mild, and mitotic fig-



Figure 7-72 Chordoma. A: Destructive osteolytic lesion in the lower sacrum, with scalloped borders and amorphous calcifications in the tumor matrix, is seen in this 60-year-old woman. B: CT shows extensive bone destruction and a large soft tissue mass.



Figure 7-73 Chordoma. Open-mouth anteroposterior conventional tomography of the cervical spine of a 52-year-old man demonstrates an osteolytic lesion in the body of C2 vertebra (*arrows*), destroying the left pedicle (*open arrow*).



Figure 7-74 Chordoma: computed tomography (CT) and magnetic resonance imaging (MRI). A: Anteroposterior radiograph of the pelvis of a 68-year-old woman shows a destructive lesion in the middle and lower part of the sacrum associated with a soft tissue mass.



Figure 7-74 *Continued* **B:** CT section demonstrates the low-attenuation tumor destroying the sacral bone (*arrows*). **C:** Axial T1-weighted (SE, TR 600, TE 20) MRI shows a large, heterogeneous tumor mass displaying predominantly intermediate signal intensity with areas of higher signal probably representing hemorrhage (*arrows*). **D:** Sagittal T1-weighted (SE, TR 650, TE 20) MRI shows the extraosseous extension of the tumor mass. **E:** On the sagittal T2-weighted (SE, TR 2000, TE 90) image the lobulated tumor displays a high signal and shows a great deal of heterogeneity. (Reprinted from Greenfield GB, Arrington JA. *Imaging of bone tumors*. Philadelphia: JB Lippincott, 1995:149–150.)



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ures are rarely discerned, predominantly seen in the most cellular areas. The fibrous septa contain the vascular channels and occasionally are infiltrated by lymphocytes (374).

Immunohistochemically, chordomas are positive for cytokeratins, S-100 protein (Fig. 7-75D,E), and epithelial membrane antigen (EMA) (361).

Chondroid chordoma, originally described by Falconer et al. (335), is a slowly growing variant of conventional chordoma. It is characterized by a substantial chondroid component, which resembles either chondroma or chondrosarcoma, intermixed with foci more characteristic of conventional chordoma (254,326). The cartilaginous tissue possesses cells within lacunae, separated by a matrix of hyaline cartilage (336).

Dedifferentiated chordoma, in addition to displaying features of conventional chordoma, exhibits features that resemble malignant fibrous histiocytoma, fibrosarcoma, osteosarcoma, or high-grade chondrosarcoma (323,353,356,358,371).

Differential Diagnosis

Radiology

The main radiologic considerations in differential diagnosis, when the tumor involves the sacral bone, are *chondrosarcoma* and *metastasis* (324,334,351). Both lesions may very closely mimic chordoma, and vice versa, although metastasis rarely is accompanied by a soft tissue mass (375). The purely lytic lesion may closely resemble *plasmacytoma*. In doubtful instances, positive scintigraphy is highly suggestive of chordoma (339). Osteomyelitis and *lymphoma* are two additional considerations. Because the radiologic appearance of these entities is very similar to that of chordoma, detailed clinical information may be a helpful aid in differential diagnosis. Finally, chordoma should be differentiated from recently reported giant notochordal intraosseous hamartoma (360).

Pathology

Benign notochordal cell tumors and giant notochordal rests may constitute a challenging differential diagnosis for the pathologist when no attention is paid to radiologic findings (325,378,379). Their distribution is similar to that of chordoma, but they are strictly confined to the vertebral body. Lobulation is absent, and the cells are arranged in sheets that fill the intertrabecular spaces. An intercellular matrix is lacking (349,378). These lesions are believed to be precursors of classical chordoma (378,379,380).

Metastatic adenocarcinoma of the rectosigmoid colon, especially of the mucinous type, is an occasional consideration in differential diagnosis. This is particularly true when the chordoma possesses many cells with single cytoplasmic vacuoles (336). Furthermore, both of these lesions may contain glycogen, mucin, cytokeratins, and EMA. In addition, both tumors may be positive for S-100 protein (362). However, colon carcinomas are positive for the nuclear transcription factor CDX2, which is absent in chordomas (350). Additional keys to the correct diagnosis are examination of several histologic sections and careful evaluation of the clinical details (346). The points to be stressed are that gland formation does not occur in chordomas and that physaliphorous cells are absent in adenocarcinomas.

Myxoid chondrosarcoma is another diagnostic possibility because of its lobular architecture, cord-like arrangement of cells, and myxoid matrix. This lesion is occasionally termed chordoid sarcoma because of its histologic similarity to chordoma (354,355,377). Helpful in the differential diagnosis is the fact that myxoid chondrosarcomas are extremely rare in the spine. They do not contain physaliphorous cells, and positivity for cytokeratin and EMA is only found focally (352). In addition, myxoid chondrosarcomas in about 50% of cases possess a characteristic nonrandom t(9;22) (q22;q12) translation that is diagnostic (368).

An interesting tumor to distinguish from chordoma is so-called *parachordoma*, first described by Dabska (330). This tumor exhibits a cytoplasmic vacuolization of the cells, which thus resemble the physaliphorous cells of chordoma. Parachordoma also contains fibrous septa similar to those of the latter tumor. This rare peripheral tumor is lobular and pseudoencapsulated, and it frequently occurs adjacent to tendons, synovium, and osseous structures. Occasionally it may arise in bone. According to Folpe et al. (340), the immunohistochemical profile of parachordomas is different from that of chordomas and extraskeletal myxoid chondrosarcomas; however, this view has been challenged by others (369). Although parachordoma is presently accepted as a distinct entity based on cytogenetic and immunohistochemical findings, its origin is still under investigation (338).

The radiologic and pathologic differential diagnosis of chordoma is depicted in Figure 7-76.

Leiomyosarcoma of Bone

Primary leiomyosarcomas of bone are very rare malignancies, with about 120 cases thus far reported (386,387,389,391–396,399,401–403,405). Berlin et al. (385) reported 16 cases and suggested that this tumor may be more common than previously recognized. Recently, a single case report described a radiation-induced leiomyosarcoma (383). It is a malignant, predominantly spindle-cell neoplasm exhibiting smooth muscle differentiation (336).

Clinical Presentation

The reported patients ranged from 9 to 80 years of age (mean 44 years). However, occurrence before the age of 20 is uncommon. Males are affected about twice as often as females. The usual presentation is pain of variable intensity and duration (382). A soft tissue mass is occasionally encountered (384). The most common sites are the distal femur, proximal tibia (336), and proximal humerus (395). Other bones that may be affected include the pelvis, clavicle, ribs, and mandible (392) (Fig. 7-77).

Imaging

Although there are no characteristic radiographic features of leiomyosarcoma, in the reported cases the tumor most often presented either as a lytic area of geographic bone destruction or with aggressive-looking,



Figure 7-76 Radiologic and pathologic differential diagnosis of chordoma.

ill-defined borders (Fig. 7-78). Some lesions exhibited an aggressive permeative and moth-eaten pattern. A periosteal reaction was present in about 50% of cases (385). On MRI the tumor is isointense relative to muscle on T1-weighted images, iso-hypointense relative to fat on fast spin-echo T2-weighted sequences, and hyperintense relative to muscle on fast spin-echo T2 weighting with fat saturation (400) (Fig. 7-79).

Histopathology

Microscopy reveals interlacing fascicles of spindleshaped cells that resemble leiomyosarcoma of the soft tissues (388,403). Cellularity varies from case to case. Most cellular areas exhibit little intervening stroma (395). Less cellular foci contain variably abundant intercellular collagen (Fig. 7-80). Rarely, a storiform-like pattern, reminiscent of malignant fibrous histiocytoma, is seen (395). The spindle cells have oval or elongated cigar-shaped nuclei and an eosinophilic (picrinophilic) cytoplasm, occasionally having a fibrillary pattern. The nuclei quite often exhibit a depression on both ends, caused by a clear vacuole. Giant cells are rarely encountered. When present, they contain multiple hyperchromatic nuclei, prominent nucleoli, and indistinct cytoplasmic borders. Mitotic figures are frequently observed. Immunoreaction for muscle-specific actin is invariably positive, and desmin marking also may be positive. Some cases exhibit positive keratin marking.

Ultrastructurally, most leiomyosarcoma cells are elongated; however, a few cells are round to polygonal (395). Nuclei often exhibit corrugated profiles. Most cells contain many intermediate smooth muscle filaments in their cytoplasm, with dense bodies along their course. They also display subplasmalemmal densities, and pinocytotic vesicles are present (395).

Differential Diagnosis Radiology

Leiomyosarcoma of bone does not have a characteristic radiologic presentation. Therefore, several possibilities should be considered in the differential diagnosis. Because of the frequently aggressive type of bone destruction exhibited by this tumor, *fibrosarcoma, malignant fibrous histiocytoma*, and *lymphoma* are primary considerations. *Solitary metastasis* in older patients is always a possibility. In younger patients, *Ewing sarcoma* and *Langerhans cell histiocytosis* must be considered.

Pathology

The most important differentiation is from a *metastasis* of an extraskeletal leiomyosarcoma, particularly from the uterus. Therefore, the diagnosis of a primary skeletal leiomyosarcoma is a diagnosis of exclusion, and clinical information about the case is crucial. Fibrosarcoma and malignant fibrous histiocytoma should be considered, particularly if areas of a herring-bone arrangement or a storiform pattern are present (398). Osteosarcoma must be a consideration because occasionally broad bands of hyalinized collagen may mimic osteoid. Because the histologic differentiation of leiomyosarcoma from these tumors with a reasonable degree of probability is almost impossible, immunocytochemical demonstration of a characterizing pattern of several smooth muscle cell markers in tumor cells is always mandatory (392), remembering, however, that some osteosarcomas may also exhibit actin positivity (390,397,404).

The radiologic and pathologic differential diagnosis of leiomyosarcoma of bone is depicted in Figure 7-81.



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Figure 7-78 Leiomyosarcoma of bone. A: Anteroposterior radiograph of the right knee of a 12-year-old boy shows an osteolytic lesion in the proximal tibial metaphysis destroying the medial cortex and extending into the soft tissues. **B:** CT section shows destruction of the medial aspect of the tibia and an associated soft tissue mass.

Figure 7-77 Leiomyosarcoma of bone: skeletal sites of predilection, peak age range, and male-to-female ratio.

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Figure 7-79 Leiomyosarcoma of bone: computed tomography (CT) and magnetic resonance imaging (MRI). A: Antero-posterior radiograph of the right knee of a 66-year-old woman shows a lytic eccentric lesion in the lateral aspect of the distal femur. B: Axial CT section shows destruction of the lateral cortex and soft tissue extension of the tumor. C: Coronal T1weighted MRI shows the lesion to be isointense with the skeletal muscles. D: On coronal T2-weighted MRI the tumor exhibits heterogeneous but predominantly high signal.

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Figure 7-80 Histopathology of leiomyosarcoma of bone (same patient as shown in Fig. 7-78). A: The tumor located in the metaphysis of the tibia destroyed the medial part of the growth plate and extended into the epiphysis, infiltrating the cancellous bone (hematoxylin and eosin, original magnification $\times 1.5$). **B:** Irregular bundles of elongated spindle cells producing intracellular collagen (red) show marked pleomorphism of the nuclei. Some tumor giant cells are present (center). Differentiation from high grade fibrosarcoma is almost impossible and should be done with immunohistochemical markers (van Gieson, original magnification \times 50). **C:** In another area the pseudoepithelial clear cell aspect of the tumor-a typical variant-helps in differentiation from fibrosarcoma (hematoxylin and eosin, original magnification \times 25).





Figure 7-81 Radiologic and pathologic differential diagnosis of leiomyosarcoma of bone.

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REFERENCES

Giant Cell Tumor

- Aoki J, Moriya K, Yamashita K, et al. Giant cell tumors of bone containing large amounts of hemosiderin: MR-pathologic correlation. J Comput Assist Tomogr 1991;15:1024–1027.
- 2. Aoki J, Tanikawa H, Ishii K, et al. MR findings indicative of hemosiderin in giant cell tumor of bone: frequency, cause, and diagnostic significance. *Am J Roentgenol* 1996;166:145–148.
- Aoki J, Moser RP Jr, Vinh TN. Giant cell tumor of the scapula. A review of 13 cases. *Skeletal Radiol* 1989;18:427–434.
- 4. Aparisi T, Arborgh B, Ericsson JL. Giant cell tumor of bone: detailed fine structural analysis of different cell components. *Virchows Arch A Pathol Anat Histol* 1977;376:273–298.
- Athanasou NA, Bliss E, Gatter KC, et al. An immunohistological study of giant-cell tumor of bone: evidence for an osteoclast origin of the giant cells. *J Pathol* 1985;147:153–158.
- Atkins GJ, Haynes DR, Graves SE, et al. Expression of osteoclast differentiation signals by stromal elements of giant cell tumors. *J Bone Miner Res* 2000;15:640–649.
- 7. Averill R, Smith R, Campbell CJ. Giant cell tumors of the bones of the hand. *J Hand Surg* 1980;5:39–50.
- Bacchini P, Bertoni F, Ruggieri P, Campanacci M. Multicentric giant cell tumor of skeleton. *Skeletal Radiol* 1995;24:371–374.
- Bertheussen KJ, Holck S, Schiodt T. Giant cell lesion of bone of the hand with particular emphasis on giant cell reparative granuloma. *J Hand Surg Am* 1983;8:46–49.
- Bertoni F, Bacchini P, Staals EL. Malignancy in giant cell tumor. Skeletal Radiol 2003;32:143–146.
- Bertoni F, Present D, Enneking WF. Giant cell tumor of bone with pulmonary metastases. J Bone Joint Surg Am 1985;67: 890–900.
- Bertoni F, Present D, Sudanese A, et al. Giant cell tumor of bone with pulmonary metastases: Six case reports and a review of the literature. *Clin Orthop* 1988;237:275–285.
- Brady TJ, Gebhardt MC, Pykett IL, et al. NMR imaging of forearms in healthy volunteers and patients with giant-cell tumor of bone. *Radiology* 1982;144:549–552.
- 14. Briccoli A, Malaguti C, Ianetti C, et al. Giant cell tumor of the rib. *Skeletal Radiol* 2003;32:107–110.
- Bridge JA, Neff JR, Bhatia PS, et al. Cytogenetic findings and biologic behavior of giant cell tumors of bone. *Cancer* 1990; 65:2697–2703.
- Bridge JA, Neff JR, Mouron BJ. Giant cell tumor of bone. Chromosomal analysis of 48 specimens and review of the literature. *Cancer Genet Cytogenet* 1992;58:2–13.
- Bullough PG, Bansai M. Malignancy in giant cell tumor. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone Lyon: IARC Press, 2002:313.
- Campanacci M, Baldini N, Boriani S, Sudanese A. Giant cell tumor of bone. J Bone Joint Surg Am 1987;69:106–114.
- Campanacci M, Giunti A, Olmi R. Giant-cell tumors of bone: a study of 209 cases with long-term follow-up in 130. *Ital J Orthop Traumatol* 1975;1:249–277.
- 20. Caskey PM, Wolf MD, Fechner RE. Multicentric giant cell reparative granuloma of the small bones of the hand. A case report and review of the literature. *Clin Orthop* 1985;193: 199–205.
- Dahlin DC. Giant cell bearing lesions of bone of the hands. Hand Clin 1987;3:291–297.
- Dahlin DC. Giant cell tumor of bone: Highlights of 407 cases. AmJ Roentgenol 1985;144:955–960.
- Dahlin DC, Cupps RE, Johnson EW Jr. Giant-cell tumor: a study of 195 cases. *Cancer* 1970;25:1061–1070.
- Denison GL, Workman TL. General case of the day. *RadioGraphics* 1997;17:545–547.
- deSantos LA, Murray JA. Evaluation of giant cell tumor by computerized tomography. *Skeletal Radiol* 1978;2:205–212.
- 26. Dorfman H, Czerniak B. Bone tumors St. Louis: Mosby, 1998.
- Dumford K, Moore TE, Walker CW, Jaksha J. Multifocal, metachronous, giant cell tumor of the lower limb. *Skeletal Radiol* 2003;32:147–150.

- Duncan CP, Morton KS, Arthur JS. Giant cell tumor of bone: Its aggressiveness and potential for malignant change. *Can J Surg* 1983;26:475–476.
- Eckhardt JJ, Grogan TJ. Giant cell tumor of bone. *Clin Orthop* 1986;204:45–58.
- Fechner RE, Mills SE. *Tumors of the bones and joints* Washington, DC: Armed Forces Institute of Pathology, 1993:173–186.
- Francis R, Lewis E. CT demonstration of giant cell tumor complicating Paget disease. J Comput Assist Tomogr 1983;7:917–918.
- Gamberi G, Benassi MS, Bohling T, et al. Prognostic relevance of C-myc gene expression in giant-cell tumor of bone. J Orthop Res 1998;16:1–7.
- Gamberi G, Benassi MS, Ragazzini P, et al. Proteases and interleukin-6 gene analysis in 92 giant cell tumors of bone. *Ann Oncol* 2004;15:498–503.
- 34. Glass TA, Mills SE, Fechner RE, et al. Giant-cell reparative granuloma of the hands and feet. *Radiology* 1983;149:65–68.
- Goldenberg RR, Campbell CJ, Bonfiglio M. Giant-cell tumor of bone. An analysis of two hundred and eighteen cases. J Bone Joint Surg Am 1970;52:619–664.
- Grote HJ, Braun M, Kalinski T, et al. Spontaneous malignant transformation of conventional giant cell tumor. *Skeletal Radiol* 2004;33:169–175.
- Hermann G, Abdelwahab IF, Klein MJ, et al. Case report 603. Giant cell reparative granuloma of the distal end of right femur. *Skeletal Radiol* 1990;19:367–369.
- Herman SD, Mesgarzadeh M, Bonakdarpour A, Dalinka MK. The role of magnetic resonance imaging in giant cell tumor of bone. *Skeletal Radiol* 1987;16:635–643.
- Hudson TM, Schiebler M, Springfield DS, et al. Radiology of giant cell tumors of bone: computed tomography, arthrotomography, and scintigraphy. *Skeletal Radiol* 1984;11:85–95.
- Hudson TM. Radiologic-pathologic correlation of musculoskeletal lesions Baltimore: Williams & Wilkins, 1987:209–237.
- Hutter RVP, Worcester JN Jr, Francis KC, et al. Benign and malignant giant cell tumors of bone. A clinicopathological analysis of the natural history of the disease. *Cancer* 1962;15:653–690.
- 42. Huvos AG. Bone tumors: diagnosis, treatment, and prognosis, 2nd ed. Philadelphia: WB Saunders, 1991.
- 43. Jaffe HL. Tumors and tumorous conditions of the bones and joints Philadelphia: Lea & Febiger, 1958.
- Jaffe HL, Lichtenstein L, Portis RB. Giant cell tumor of bone. Its pathologic appearance, grading, supposed variants and treatment. Arch Pathol 1940;30:993–1031.
- Kaplan PA, Murphy M, Greenway G, et al. Fluid-fluid levels in giant cell tumors of bone: report of two cases. *Computed Tomogr* 1987;11:151–155.
- Kransdorf MJ, Sweet DE, Buetow PC, et al. Giant cell tumor in skeletally immature patients. *Radiology* 1992;184:233–237.
- Ladanyi M, Traganos F, Huvos AG. Benign metastasizing giant cell tumors of bone: a DNA flow cytometric study. *Cancer* 1989;64:1521–1526.
- Lee JA, Bank WO, Gonzalez-Melendez M, Olan WJ, Tabbara SO. Giant cell tumor of the skull. *RadioGraphics* 1998;18: 1295–1302.
- Levine E, DeSmet AA, Neff JR, Martin NL. Scintigraphic evaluation of giant cell tumor of bone. *Am J Roentgenol* 1984;143: 343–348.
- Levine E, DeSmet AA, Neff JR. Role of radiologic imaging in management planning of giant cell tumor of bone. *Skeletal Radiol* 1984;12:79–89.
- Lorenzo JC, Dorfman HD. Giant-cell reparative granuloma of short tubular bones of the hands and feet. *Am J Surg Pathol* 1980;4:551–563.
- Maloney WJ, Vaughan LM, Jones HH, et al. Benign metastasizing giant-cell tumor of bone. Report of three cases and review of the literature. *Clin Orthop* 1989;243:208–215.
- Manaster BJ, Doyle AJ. Giant cell tumor of bone. Radiol Clin North Am 1993;31:299–323.
- Marai T, Yamamoto T, Yoshihara H, et al. De novo malignant transformation of giant cell tumor of bone. *Skeletal Radiol* 2001;30:104–108.
- Matsuno T. Benign fibrous histiocytoma involving the ends of long bone. *Skeletal Radiol* 1990;19:561–566.

- May DA, Good RB, Smith DK, Parsons TW. MR imaging of musculoskeletal tumors and tumor mimickers with intravenous gadolinium: experience with 242 patients. *Skeletal Radiol* 1997; 26:2–15.
- McComb EN, Johansson SL, Neff JR, et al. Chromosomal anomalies exclusive of telomeric associations in giant cell tumor of bone. *Cancer Genet Cytogenet* 1996;88:163–166.
- McDonald DJ, Sim FH, McLeod RA, Dahlin DC. Giant cell tumor of bone. *J Bone Joint Surg Am* 1986;68:235–242.
- 59. McGrath J. Giant-cell tumour of bone: an analysis of fifty-two cases. J Bone Joint Surg Br 1972;54:216–229.
- McInerney DP, Middlemis JH. Giant-cell tumors of bone. Skeletal Radiol 1978;2:195–204.
- Meis JM, Dorfman HD, Nathanson SD, et al. Primary malignant giant cell tumor of bone: dedifferentiated giant cell tumor. *Mod Pathol* 1989;2:541–546.
- Moser RP Jr, Kransdorf MJ, Gilkey FW, Manaster BJ. Giant cell tumor of the upper extremity. *RadioGraphics* 1990;10:83–102.
- Murphey MD, Nomikos GC, Flemming DJ, et al. Imaging of giant cell tumor and giant cell reparative granuloma of bone: radiologic-pathologic correlation. *RadioGraphics* 2001;21: 1283–1309.
- Nascimento AG, Huvos AG, Marcove RC. Primary malignant giant cell tumor of bone: a study of eight cases and review of the literature. *Cancer* 1979;44:1393–1402.
- Nusbacher N, Sclafani SJ, Birla SR. Case report 155. Polyostotic Paget disease complicated by benign giant cell tumor of left clavicle. *Skeletal Radiol* 1981;6:233–235.
- Nojima T, Takeda N, Matsuno T, et al. Case report 869. Benign metastasizing giant cell tumor of bone. *Skeletal Radiol* 1994; 23:583–585.
- 67. Oda Y, Tsuneyoshi M, Shinohara N. Solid variant of aneurysmal bone cyst (extragnatic giant cell reparative granuloma) in the axial skeleton and long bones: a study of its morphologic spectrum and distinction from allied giant cell lesions. *Cancer* 1992;70:2642–2649.
- O'Reilly M, Chew FS. The scintigraphic features of giant-cell tumors in relation to other imaging modalities. *Clin Nucl Med* 1996;21:43–48.
- Peimer CA, Schiller AL, Mankin HJ, Smith RJ. Multicentric giant cell tumor of bone. J Bone Joint Surg Am 1980;62:652–656.
- Picci P, Baldini N, Sudanese A, et al. Giant cell reparative granuloma and other giant cell lesions of the bones of the hand and feet. *Skeletal Radiol* 1986;15:415–421.
- Picci P, Manfrini M, Zucchi V, et al. Giant cell tumor of bone in skeletally immature patients. J Bone Joint Surg Am 1983;65: 486–490.
- Potter HG, Schneider R, Ghelman B, et al. Multiple giant cell tumors and Paget disease of bone: Radiographic and clinical correlations. *Radiology* 1991;180:261–264.
- Rao UN, Goodman M, Chung WW, et al. Molecular analysis of primary and recurrent giant cell tumors of bone. *Cancer Genet Cytogenet* 2005;158:126–136.
- 74. Reid R, Banerjee SS, Sciot R. Giant cell tumor. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone Lyon: IARC Press, 2002:310–312.
- Resnik CS, Steffe JW, Wang SE. Case report 353. Giant cell tumor of distal end of the femur, containing a fluid level as demonstrated by computed tomography. *Skeletal Radiol* 1986;15: 175–177.
- Rock MG, Pritchard DJ, Unni KK. Metastases from histologically benign giant-cell tumor of bone. J Bone Joint Surg Am 1984;66: 269–274.
- Rock MG, Sim FH, Unni KK, et al. Secondary malignant giantcell tumor of bone. Clinicopathological assessment of nineteen patients. *J Bone Joint Surg Am* 1986;68:1073–1079.
- Roessner A, Bassewitz BDV, Schlake W, et al. Biologic characterization of human bone tumors. III. Giant cell tumor of bone. *Path Res Pract* 1984;178:431–440.
- Rosai J. Carcinoma of pancreas simulating giant cell tumor of bone. Electron-microscopic evidence of its acinar cell origin. *Cancer* 1968;22:333–344.
- Sanerkin NG. Malignancy, aggressiveness and recurrence in giant cell tumor of bone. *Cancer* 1980;46:1641–1649.

- Schajowicz F, Slullitel J. Giant cell tumor associated with Paget's disease of bone. J Bone Joint Surg Am 1966;48:1340–1349.
- Schajowicz F. Tumors and tumorlike lesions of bone: pathology, radiology, and treatment, 2nd ed. Berlin: Springer-Verlag, 1994.
- Shankman S, Greenspan A, Klein MJ, Lewis MM. Giant cell tumor of the ischium. A report of two cases and review of the literature. *Skeletal Radiol* 1988;17:46–51.
- Si AI, Huang L, Xu J, et al. Expression and localization of extracellular matrix metalloproteinase inducer in giant cell tumor of bone. *J Cell Biochem* 2003;89:1154–1163.
- Sim FH, Dahlin DC, Beabout JW. Multicentric giant cell tumors of bone. J Bone Joint Surg Am 1977;59:1052–1060.
- Simon MA, Kirchner PT. Scintigraphic evaluation of primary bone tumors. J Bone Joint Surg Am 1980;62:758–764.
- Stacy GS, Peabody TD, Dixon LB. Mimics on radiography of giant cell tumor of bone. Am J Roentgenol 2003;181:1583–1589.
- Steiner GC, Ghosh L, Dorfman HD. Ultrastructure of giant cell tumor of bone. *Hum Pathol* 1972;3:569–586.
- Szendröi M, Antal I, Perlaky G. Mid-foot reconstruction following involvement of five bones by giant cell tumor. *Skeletal Radiol* 2000;29:664–667.
- Tehranzadeh J, Murphy BJ, Mnaymneh W. Giant cell tumor of the proximal tibia: MR and CT appearance. J Comput Assist Tomogr 1989;13:282–286.
- Tornberg DN, Dick HM, Johnston AD. Multicentric giant-cell tumors in the long bones. A case report. J Bone Joint Surg Am 1975;57:420–422.
- 92. Tubbs WS, Brown LR, Beabout JW, et al. Benign giant-cell tumor of bone with pulmonary metastases: clinical findings and radiologic appearance of metastases in 13 cases. *Am J Roentgenol* 1992;158:331–334.
- 93. Ueda Y, Imai K, Tsuchiya H, et al. Matrix metalloproteinase 9 (gelatinase B) is expressed in multinucleated giant cells of human giant cell tumor of bone and is associated with vascular invasion. *Am J Pathol* 1996;148:611–622.
- Unni KK. Dahlin's bone tumors: general aspects and data on 11,087 cases, 5th ed. New York: Lippincott-Raven, 1996.
- Van Nostrand D, Madewell JE, McNiesh LM, et al. Radionuclide bone scanning in giant cell tumor. J Nucl Med 1986;27: 329–338.
- Williams HT. Multicentric giant cell tumor of bone. *Clin Nucl Med* 1989;14:631–633.
- Wold LE, Dobyns JH, Swee RG, Dahlin DC. Giant cell reaction (giant cell reparative granuloma) of the small bones of the hands and feet. *Am J Surg Pathol* 1986;10:491–496.
- Wold LE, Swee RG. Giant cell tumor of the small bones of the hand and feet. *Semin Diagn Pathol* 1984;1:173–184.
- Wray CC, MacDonald AW, Richardson RA. Benign giant-cell tumor with metastases to bone and lung. J Bone Joint Surg Br 1990;72:486–489.
- Wulling M, Delling G, Kaiser E. The origin of the neoplastic stromal cell in giant cell tumor of bone. *Hum Pathol* 2003; 34:983–993.
- 101. Wulling M, Engels C, Jesse N, et al. The nature of giant cell tumor of bone. J Cancer Res Clin Oncol 2001;127:467–474.
- 102. Zheng MH, Fan Y, Panicker A, et al. Detection of mRNAs for urokinase-type plasminogen activator, its receptor, and type 1 inhibitor in giant cell tumors of bone with in situ hybridization. *Am J Pathol* 1995;147:1559–1566.
- 103. Zhu X, Steiner GC. Malignant giant cell tumor of bone: malignant transformation of a benign giant cell tumor treated by surgery. *Bull Hosp Joint Dis Orthop Inst* 1990;50:169–176.

Simple Bone Cyst

- Abdelwahab IF, Lewis MM, Klein MJ, Barbera C. Case report 515. Simple (solitary) bone cysts of the calcaneus. *Skeletal Radiol* 1989;17:607–610.
- 105. Baker DM. Benign unicameral bone cyst. *Clin Orthop* 1970;71: 140–151.
- Barcelo M, Pathria MN, Abdul-Karim FW. Intraosseous lipoma. A clinicopathologic study of four cases. Arch Pathol Lab Med 1992;116:947–950.
- Berlin SJ. A review of 2,720 lesions of the foot. JAm Podiatr Assoc 1980;70:318–324.

- 108. Blumberg ML. CT of iliac unicameral bone cysts. Am J Roentgenol 1981;136:1231–1232.
- Boseker EH, Bickel WH, Dahlin DC. A clinicopathologic study of simple unicameral bone cysts. *Surg Gynecol Obstet* 1968;127: 550–560.
- 110. Broder HM. Possible precursor of unicameral bone cysts. J Bone Joint Surg Am 1968;50:503–507.
- Campanacci M, Capanna R, Picci P. Unicameral and aneurysmal bone cysts. *Clin Orthop* 1986;204:25–36.
- 112. Capanna R, Van Horn J, Ruggieri P, Biagini R. Epiphyseal involvement in unicameral bone cysts. *Skeletal Radiol* 1986;15: 428–432.
- 113. Chow LT, Lee K. Intraosseous lipoma. A clinicopathological study of nine cases. *Am J Surg Pathol* 1992;16:401–410.
- 114. Cohen J. Etiology of simple bone cyst. J Bone Joint Surg Am 1970;52:1493–1497.
- Cohen J. Unicameral bone cysts: a current synthesis of reported cases. Orthop Clin North Am 1977;8:715–726.
- Conway WF, Hayes CW. Miscellaneous lesions of bone. Radiol Clin North Am 1993;31:339–358.
- 117. Goldberg RP, Genant HK. Case report 67. Solitary bone cyst of the right ilium. *Skeletal Radiol* 1978;3:118–121.
- 118. Haims AH, Desai P, Present D, Beltran J. Epiphyseal extension of a unicameral bone cyst. *Skeletal Radiol* 1997;26:51–54.
- 119. Hertzanu Y, Mendelsohn DB, Gottschalk F. Aneurysmal bone cyst of the calcaneus. *Radiology* 1984;151:51–52.
- 120. Hudson TM. Radiologic-Pathologic correlation of musculoskeletal lesions Baltimore: Williams & Wilkins, 1987:249–252.
- Huvos AG. Bone tumors: diagnosis, treatment, and prognosis, 2nd ed. Philadelphia: WB Saunders, 1991:713–726.
- 122. Jaffe HL, Lichtenstein L. Solitary unicameral bone cyst with emphasis on the roentgen picture, the pathologic appearance, and the pathogenesis. *Arch Surg* 1942;44:1004–1025.
- Keats TE. Atlas of normal roentgen variants that may simulate disease, 4th ed. Chicago: Year Book, 1988:699–700.
- Ketyer S, Braunstein S, Cholankeril J. CT diagnosis of intraosseous lipoma of the calcaneus. *J Comput Assist Tomogr* 1983; 7:546–547.
- Kricun ME. Tumors of the foot. In: Kricun ME, ed. Imaging of bone tumors Philadelphia: WB Saunders, 1993:221–225.
- 126. Kricun ME, Kricun R, Haskin ME. Chondroblastoma of the calcaneus: radiographic features with emphasis on location. *Am J Roentgenol* 1977;128:613–616.
- Lagier R. Calcaneus lipoma with bone infarct. Fortschr Röentgenstr 1985;142:472–474.
- McGlynn FJ, Mickelson MR, El-Khoury GY. The fallen fragment sign in unicameral bone cyst. *Clin Orthop* 1981;156:157–159.
- Milgram JW. Intraosseous lipomas. A clinicopathological study of 66 cases. *Clin Orthop* 1988;231:277–302.
- Moore TE, King AR, Travis RC, Allen BC. Post-traumatic cysts and cyst-like lesions of bone. *Skeletal Radiol* 1989;18:93–97.
- Morton KS. The pathogenesis of unicameral bone cyst. Can J Surg 1964;7:140–150.
- Mulder JD, Kroon HM, Schütte HE, Taconis WK. Radiologic atlas of bone tumors Amsterdam: Elsevier, 1993:579–590.
- Murray RO, Jacobson HG. *The radiology of skeletal disorders*, 2nd ed. New York: Churchill Livingstone, 1977.
- 134. Neer CS II, Francis KC, Marcove RC, et al. Treatment of unicameral bone cyst. A follow-up study of one hundred seventyfive cases. J Bone Joint Surg Am 1966;48:731–745.
- Norman A, Schiffman M. Simple bone cyst: factors of age dependency. *Radiology* 1977;124:779–782.
- 136. Remagen W, Lampérth BE, Jundt G, Schildt R. Das sogenannte osteolytische Dreieck des Calcaneus. Radiologische und pathoanatomische Befunde. Osteologie 1994;3:275–283.
- Reynolds J. The fallen fragment sign in the diagnosis of unicameral bone cysts. *Radiology* 1969;92:949–953.
- Richkind KE, Mortimer E, Mowery-Rushton P, Fraire A. Translocation (16;20) (p11.2;q13). Sole cytogenetic abnormality in a unicameral bone cyst. *Cancer Genet Cytogenet* 2002;137: 153–155.
- Schajowicz F. Tumors and tumorlike lesions of bone and joints, 2nd ed. New York: Springer-Verlag, 1994:505–514.
- 140. Smith RW, Smith CF. Solitary unicameral bone cyst of the calcaneus. A review of 20 cases. J Bone Joint Surg Am 1974;56: 49–56.

- 141. Struhl S, Edelson C, Pritzker H, et al. Solitary (unicameral) bone cyst. The fallen fragment sign revisited. *Skeletal Radiol* 1989;18:261–265.
- 142. Van Linthoudt D, Lagier R. Calcaneal cysts: a radiological and anatomical-pathological study. Acta Orthop Scand 1978;49: 310–316.
- 143. Vayego SA, De Conti OJ, Varella-Garcia M. Complex cytogenetic rearrangement in a case of unicameral bone cyst. *Cancer Genet Cytogenet* 1996;86:46–49.
- 144. Vayego-Lourenco SA. TP53 mutations in a recurrent unicameral bone cyst. *Cancer Genet Cytogenet* 2001;124:175–176.
- Weisel A, Hecht HL. Development of a unicameral bone cyst. J Bone Joint Surg Am 1980;62:664–666.

Aneurysmal Bone Cyst

- 146. Aho HJ, Aho AJ, Einola S. Aneurysmal bone cyst, a study of ultrastructure and malignant transformation. *Virchows Arch A* 1982;395:169–179.
- 147. Aho HJ, Aho AJ, Pellineimi LJ, et al. Endothelium in aneurysmal bone cyst. *Histopathology* 1985;9:381–387.
- Alles JU, Schulz A. İmmunohistochemical markers (endothelial and histiocytic) and ultrastructure of primary aneurysmal bone cysts. *Hum Pathol* 1986;17:39–45.
- 149. Althof PA, Ohmori K, Zhou M, et al. Cytogenetic and molecular cytogenetic findings in 43 aneurysmal bone cysts: aberrations of 17p mapped to 17p13.2 by fluorescence in situ hybridization. *Mod Pathol* 2004;17:518–525.
- Apaydin A, Özkaynak C, Yilmaz S, et al. Aneurysmal bone cyst of metacarpal. *Skeletal Radiol* 1996;25:76–78.
- 151. Barrett TJ, Beall DP, Ly JQ, Davis SW. Cortical aneurysmal bone cyst of the tibia. *Am J Roentgenol* 2004;182:740.
- 152. Baruffi MR, Neto JB, Barbieri CH, Casartelli C. Aneurysmal bone cyst with chromosomal changes involving 7q and 16p. *Cancer Genet Cytogenet* 2001;129:177–180.
- 153. Beltran J, Simon DC, Levy M, et al. Aneurysmal bone cysts: MR imaging at 1.5 T. *Radiology* 1986;158:689–690.
- 154. Bertoni F, Bacchini P, Capanna R, et al. Solid variant of aneurysmal bone cyst. *Cancer* 1993;71:729–734.
- Biesecker JL, Marcove RC, Huvos AG, Miké V. Aneurysmal bone cysts. A clinicopathologic study of 66 cases. *Cancer* 1970;26: 615–625.
- Bonakdarpour A, Levy WM, Aegerter E. Primary and secondary aneurysmal bone cysts: a radiological study of 75 cases. *Radiology* 1978;126:75–83.
- 157. Buirski G, Watt I. The radiological features of solid aneurysmal bone cysts. *Br J Radiol* 1984;57:1057–1065.
- Burnstein MI, De Smet AA, Hafez GR, Heiner JP. Case report 611. Subperiosteal aneurysmal bone cyst of tibia. *Skeletal Radiol* 1990;4:294–297.
- 159. Clough JR, Price CH. Aneurysmal bone cyst: pathogenesis and long term results of treatment. *Clin Orthop* 1973;97:52–63.
- 160. Dabezies EJ, D Ambrosia RD, Chuinard RG, Ferguson AB Jr. Aneurysmal bone cyst after fracture. A report of three cases. J Bone Joint Surg Am 1982;64:617–621.
- Dabska M, Buraczewski J. Aneurysmal bone cyst. Pathology, clinical course and radiologic appearances. *Cancer* 1969;23: 371–389.
- Dahlin DC, McLeod RA. Aneurysmal bone cyst and other nonneoplastic conditions. *Skeletal Radiol* 1982;8:243–250.
- 163. Dal Cin P, Kozakewich HP, Goumnerova L, et al. Variant translocations involving 16q22 and 17p13 in solid variant and extraosseous forms of aneurysmal bone cyst. *Genes Chromosomes Cancer* 2000;28:233–234.
- 164. Fechner RE, Mills SE. Tumors of the bones and joints Washington, DC: Armed Forces Institute of Pathology, 1993:181–184, 253–258.
- 165. Freeby JA, Reinus WR, Wilson AJ. Quantitative analysis of the plain radiographic appearance of aneurysmal bone cyst. *Invest Radiol* 1995;30:433–439.
- 166. Fush SE, Herndon JH. Aneurysmal bone cyst involving the hand: a review and report of two cases. J Hand Surg 1979;4A: 152–159.
- Garg NK, Carty H, Walsh HPJ, et al. Percutaneous Ethibloc injection in aneurysmal bone cysts. *Skeletal Radiol* 2000;29: 211–216.
- Godfry LW, Gresham GA. The natural history of aneurysmal bone cyst. Proc R Soc Med 1959;52:900–905.

- 169. Herens C, Thiry A, Dresse MF, et al. Translocation (16;17) (q22;p13) is a recurrent anomaly of aneurysmal bone cysts. *Cancer Genet Cytogenet* 2001;127:83–84.
- 170. Heyman S, Treves S. Scintigraphy in pediatric bone tumors. In: Jaffe N, ed. Bone tumors in children Littleton, MA: Wright, 1979:79–96.
- 171. Hudson TM. Fluid levels in aneurysmal bone cysts: a CT feature. *Am J Roentgenol* 1984;141:1001–1004.
- Hudson TM. Radiologic-pathologic correlation of musculoskeletal lesions Baltimore: Williams & Wilkins, 1987:261–265.
- 173. Hudson TM. Scintigraphy of aneurysmal bone cysts. Am J Roentgenol 1984;142:761–765.
- 174. Hudson TM, Hamlin DJ, Fitzimmons JR. Magnetic resonance imaging of fluid levels in an aneurysmal bone cyst and in anticoagulated human blood. *Skeletal Radiol* 1985;13:267–270.
- 175. Huvos AG. *Bone tumors: diagnosis, treatment, and prognosis,* 2nd ed. Philadelphia: WB Saunders, 1991:727–743.
- 176. Jaffe HL. Aneurysmal bone cyst. Bull Hosp Joint Dis 1950; 11:3-13.
- 177. Kinley S, Wiseman F, Wertheimer SJ. Giant cell tumor of the talus with secondary aneurysmal bone cyst. *J Foot Ankle Surg* 1993;32:38–46.
- Kransdorf MJ, Sweet DE. Aneurysmal bone cyst: concept, controversy, clinical presentation, and imaging. *Am J Roentgenol* 1995;164:573–580.
- 179. Kyriakos M, Hardy D. Malignant transformation of aneurysmal bone cyst, with an analysis of the literature. *Cancer* 1991;68: 1770–1780.
- Leithner A, Windhager R, Lang S, et al. Aneurysmal bone cyst. *Clin Orthop Relat Res* 1999;363:176–179.
- 181. Levy WM, Miller AS, Bonakdarpour A, Aegerter E. Aneurysmal bone cyst secondary to other osseous lesions. Report of 57 cases. *Am J Clin Pathol* 1975;63:1–8.
- Lichtenstein L. Aneurysmal bone cyst. A pathological entity commonly mistaken for giant cell tumor and occasionally for hemangioma and osteogenic sarcoma. *Cancer* 1950;3:279–289.
- Lichtenstein L. Aneurysmal bone cyst. Observations on fifty cases. J Bone Joint Surg Am 1957;39:873–882.
- Makhija MC. Bone scanning in aneurysmal bone cyst. Clin Nucl Med 1981;6:500–501.
- 185. Martinez V, Sissons HA. Aneurysmal bone cyst. A review of 123 cases including primary lesions and those secondary to other bone pathology. *Cancer* 1988;61:2291–2304.
- McCarthy EF, Dorfman HD. Vascular and cartilaginous hamartoma of the ribs in infancy with secondary aneurysmal bone cyst formation. *Am J Surg Pathol* 1980;4:247–253.
- 187. Mintz MC, Dalinka MK, Schmidt R. Aneurysmal bone cyst arising in fibrous dysplasia during pregnancy. *Radiology* 1987;165: 549–550.
- Mirra JM. Bone tumors: clinical, radiologic, and pathologic correlations Philadelphia: Lea & Febiger, 1989:1267–1311.
- Moore TE, King AR, Travis RC, Allen BC. Post-traumatic cysts and cyst-like lesions of bone. *Skeletal Radiol* 1989;18:93–97.
- Mulder JD, Kroon HM, Schütte HE, Taconis WK. Radiologic atlas of bone tumors Amsterdam: Elsevier, 1993:557–578.
- Munk PL, Helms CA, Holt RG, et al. MR imaging of aneurysmal bone cysts. Am J Roentgenol 1989:153:99–101.
- 192. Nguyen BD, Lugo-Olivieri CH, McCarthy EF, et al. Fibrous dysplasia with secondary aneurysmal bone cyst. *Skeletal Radiol* 1996;25:88–91.
- O'Donnell P, Saifuddin A. The prevalence and diagnostic significance of fluid-fluid levels in focal lesions of bone. *Skeletal Radiol* 2004;33:330–336.
- 194. Oliveira AM, Chou MM, Perez-Atayde AR, Rosenberg AE. Aneurysmal bone cyst: a neoplasm driven by upregulation of the USP6 oncogene. *J Clin Oncol* 2006;24:e1; author reply e2.
- 195. Oliveira AM, His BL, Weremowicz S, et al. USP6 (Tre2) fusion oncogenes in aneurysmal bone cyst. *Cancer Res* 2004;64: 1920–1923.
- 196. Oliveira AM, Perez-Atayde AR, Inwards CY, et al. USP6 and CDH11 oncogenes identify the neoplastic cell in primary aneurysmal bone cysts and are absent in so-called secondary aneurysmal bone cysts. *Am J Pathol* 2004;165:1773–1780.
- 197. Panoutsakopoulos G, Pandis N, Kyriazoglou I, et al. Recurrent t(16;17)(q22;p13) in aneurysmal bone cysts. *Genes Chromosomes Cancer* 1999;26:265–266.

- 198. Ratcliffe PJ, Grimmer RJ. Aneurysmal bone cyst arising after tibial fracture. A case report. J Bone Joint Surg Am 1993;75: 1225–1227.
- 199. Ratner V, Dorfman HD. Giant-cell reparative granuloma of the hand and foot bones. *Clin Orthop* 1990;260:251–258.
- 200. Ruiter DJ, van Rijssel TG, van der Velde EA. Aneurysmal bone cysts. A clinicopathological study of 105 cases. *Cancer* 1977;39: 2231–2239.
- 201. Sanerkin NG, Mott MG, Roylance J. An unusual intraosseous lesion with fibromyxoid elements: solid variant of aneurysmal bone cyst. *Cancer* 1983;51:2278–2286.
- 202. Schoedel K, Shankman S, Desai P. Intracortical and subperiosteal aneurysmal bone cysts: a report of three cases. *Skeletal Radiol* 1996;25:455–459.
- 203. Scully SP, Temple HT, O Keefe RJ, Gebhardt MC. Case report 830. Aneurysmal bone cyst. *Skeletal Radiol* 1994;23:157–160.
- 204. Sharma P, Elangovan S, Ratnakar C. Case report: calcification within aneurysmal bone cyst. *Br J Radiol* 1994;67:306–308.
- 205. Sherman RS, Soong KY. Aneurysmal bone cyst: its roentgen diagnosis. *Radiology* 1957;68:54–64.
- Simon MA, Kirchner PT. Scintigraphic evaluation of primary bone tumors. *J Bone Joint Surg Am* 1980;62:758–764.
- 207. Spjut HJ, Dorfman HD, Fechner RE, Ackerman LV. Tumors of bone and cartilage Washington, DC: Armed Forces Institute of Pathology, 1971.
- Thrall JH, Geslien GE, Corcoron RJ, Johnson MC. Abnormal radionuclide deposition patterns adjacent to focal skeletal lesions. *Radiology* 1975;115:659–663.
- Tillman BP, Dahlin DC, Lipscomb PR, Stewart JR. Aneurysmal bone cyst: an analysis of ninety-five cases. *Mayo Clin Proc* 1968; 43:478–495.
- Tsai JC, Dalinka MK, Fallon MD, et al. Fluid-fluid level: a nonspecific finding in tumors of bone and soft tissue. *Radiology* 1990;175:779–782.
- 211. Unni KK. Dahlin's bone tumors: general aspects and data on 11,087 cases, 5th ed. New York: Lippincott-Raven, 1996.
- Vergel De Dios AM, Bond JR, et al. Aneurysmal bone cyst. A clinicopathologic study of 238 cases. *Cancer* 1992;69:2921–2931.
- Zimmer WD, Berquist TH, McLeod RA, et al. Bone tumors: magnetic resonance imaging versus computed tomography. *Radiology* 1985;155:709–718.
- Zimmer WD, Berquist TH, Sim FH, et al. Magnetic resonance imaging of aneurysmal bone cyst. *Mayo Clin Proc* 1984;59: 633–636.

Solid Variant of Aneurysmal Bone Cyst

- Bertoni F, Biscaglia R, Bacchini P. Giant cell reparative granuloma of the phalanx of the hand with aggressive radiographic features. *Skeletal Radiol* 1998;27:584–587.
- Buresh CJ, Seemayer TA, Nelson M, et al. t(X;4) (q22;q31.3) in giant cell reparative granuloma. *Cancer Genet Cytogenet* 1999; 115:80–81.
- 217. Caskey PM, Wolf MD, Fechner RE. Multicentric giant cell reparative granuloma of the small bones of the hand. *Clin Orthop* 1985;193:199–205.
- Ilaslan H, Sundaram M, Unni KK. Solid variant of aneurysmal bone cysts in long tubular bones: Giant cell reparative granuloma. *Am J Roentgenol* 2003;180:1681–1687.
- 219. Jaffe HL. Giant-cell reparative granuloma, traumatic bone cysts, and fibrous (fibro-osseous) dysplasia of the jawbones. *Oral Surg Oral Med Oral Pathol* 1953;6:159–175.
- Kenan S, Lewis MM, Abdelwahab IF, Klein M. Subperiosteal giantcell reparative granuloma. *J Bone Joint Surg Br* 1994;76: 810–813.
- 221. Lorenzo JC, Dorfman HD. Giant-cell reparative granuloma of short tubular bones of the hands and feet. *Am J Surg Pathol* 1980;4:551–568.
- 222. Wold LE, Dobyns JH, Swee RG, Dahlin DC. Giant cell reaction (giant cell reparative granuloma) of the small bones of the hands and feet. *Am J Surg Pathol* 1986;10:491–496.
- 223. Yamamoto T, Marui T, Akisue T, Mizuno K. Solid aneurysmal bone cyst in the humerus. *Skeletal Radiol* 2000;29:470–473.

Intraosseous Lipoma

224. Addison AK, Payne SR. Primary liposarcoma of bone. Case report. J Bone Joint Surg Am 1982;64:301–304.

- 225. Blacksin MF, Ende N, Benevenia J. Magnetic resonance imaging of intraosseous lipomas: a radiologic-pathologic correlation. *Sheletal Radiol* 1995;24:37–41.
- 226. Bonnet J, Garreta L, Ducloux J-M. Les lacunes calcanéennes. (Calcaneal lacunae.) Ann Radiol 1968;11:171–184.
- 227. Bridge JA, DeBoer J, Walker CW, Neff JR. Translocation t(3;12)(q28;q14) in parosteal lipoma. *Genes Chromosomes Cancer* 1995;12:70–72.
- Bruni L. The cockade image: a diagnostic sign of calcaneum intraosseous lipoma. *Rays* 1986;11:51–54.
- 229. Campbell RSD, Grainger AJ, Mangham DC, et al. Intraosseous lipoma: report of 35 new cases and a review of the literature. *Skeletal Radiol* 2003;32:209–222.
- Dahlin DC, Unni KK. Bone tumors: general aspects and data on 8,542 cases, 4th ed. Springfield IL: Charles C Thomas, 1986: 181–185.
- DeLee JC. Intraosseous lipoma of the proximal part of the femur: case report. *J Bone Joint Surg Am* 1979;61:601–603.
- Dominok GW, Knoch HG. Knochengeschwülste und Geschwulstähnliche Knochenerkrankungen, 3rd ed. Jena: VEB Gustav Fischer Verl, 1971:267–275.
- Dooms GC, Hricak H, Sollitto RA, Higgins CB. Lipomatous tumors and tumors with fatty component: MR imaging potential and comparison of MR and CT results. *Radiology* 1985;157: 479–483.
- 234. Fechner RE, Mills SE. *Tumors of the bones and joints* Washington, DC: Armed Forces Institute of Pathology, 1993:203–209.
- Gray's anatomy. The anatomical basis of medicine and surgery, 38th ed. New York. Churchill Livingstone, 1995:452–468.
- Greenspan A, Raiszadeh K, Riley GM, Matthews D. Intraosseous lipoma of the calcaneus. *Foot Ankle Int* 1997;18:53–56.
- 237. Hall RL, Shereff MJ. Anatomy of the calcaneus. *Clin Orthop Relat Res* 1993;290:27–35.
- 238. Hart JAL. Intraosseous lipoma. J Bone Joint Surg Br 1973;55: 624–632.
- Junghanns H. Lipomas (fatty marrow areas) in the vertebral column. In: Handbuch der Speziellen Pathologischen Anatomie und Histologie, Tome IX/4. Berlin: Springer-Verlag, 1939:333–334.
- 240. Keats TE. Atlas of normal Roentgen variants that may simulate disease, 5th ed. St Louis: Mosby Year Book, 1992:637-648.
- Keats TE, Harrison RB. The calcaneal nutrient foramen: a useful sign in the differentiation of true from simulated cysts. *Skeletal Radiol* 1979;3:239–240.
- 242. Kenan S, Lewis MM, Abdelwahab IF, et al. Case report 652. Primary intraosseous low grade myxoid sarcoma of the scapula (myxoid liposarcoma). *Skeletal Radiol* 1991;20:73–75.
- 243. Köhler A, Zimmer EA. Borderlands of normal and early pathologic findings in skeletal radiography, 13th ed. Revised by Schmidt H, Freyschmidt J. Stuttgart: Thieme Verlag, 1993:797–814.
- 244. Kricun ME. Imaging of bone tumors Philadelphia: WB Saunders, 1993:236-237.
- Lagier R. Case report 128. Intraosseous lipoma. Skeletal Radiol 1980;5:267–269.
- Leeson MC, Kay D, Smith BS. Intraosseous lipoma. Clin Orthop 1983;181:186–190.
- Levey DS, Sartoris DJ, Resnick D. Advanced diagnostic imaging techniques for pedal osseous neoplasms. *Clin Pod Med Surg* 1993;10:655–682.
- 248. Levin MF, Vellet AD, Munk PL, McLean CA. Intraosseous lipoma of the distal femur: MRI appearance. *Skeletal Radiol* 1996;25:82–84.
- Milgram JW. Intraosseous lipoma: Radiologic and pathologic manifestations. *Radiology* 1988;167:155–160.
- Mueller MC, Robbins JL. Intramedullary lipoma of bone. Report of a case. J Bone Joint Surg Am 1960;42:517–520.
- 251. Nielsen GP, Mandahl N. Lipoma. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone Lyon: IARC Press, 2002:20–22.
- Norman A, Steiner GC. Radiographic and morphological features of cyst formation in idiopathic bone infarction. *Radiology* 1983;146:335–338.
- 253. Petit MM, Swarts S, Bridge JA, Van de Ven WJ. Expression of reciprocal fusion transcripts of the HMGIC and LPP genes in parosteal lipoma. *Cancer Genet Cytogenet* 1998;106:18–23.

- 254. Posteraro RH. Radiographic evaluation of pedal osseous tumors. *Clin Pod Med Surg* 1993;10:633–653.
- 255. Poussa M, Holmstrom T. Intraosseous lipoma of the calcaneus: report of a case and a short review of the literature. *Acta Orthop Scand* 1976;47:570–574.
- Propeck T, Bullard MA, Lin J, et al. Radiologic-pathologic correlation of intraosseous lipomas. *Am J Roentgenol* 2000;175: 673–678.
- 257. Ramos A, Castello J, Sartoris DJ, et al. Osseous lipoma: CT appearance. *Radiology* 1985;157:615–619.
- Remagen W. Pathologische Anatomie der Femurkopfnekrose. Orthopäde 1990;19:174–181.
- 259. Resnick D, Kyriakos M, Greenway GD. Tumors and tumor-like lesions of bone: imaging and pathology of specific lesions. In: Resnick D. ed. *Bone and joint imaging* Philadelphia: WB Saunders, 1989:1154–1156.
- 260. Resnick D, Niwayama J. *Diagnosis of bone and joint disorders* Philadelphia: WB Saunders, 1988:3782–3786.
- Salzer M, Gotzmann H. Parostale Lipome. (Parosteal lipomas.) Bruns Beitr Klin Chir 1963;206:501–505.
- Salzer M, Salzer-Kuntschik M. Zur Frage der sogenannten zentralen Knochenlipome. *Beitr Pathol Anat* 1965;132:365–375.
- Sirry A. The pseudo-cystic triangle in the normal os calcis. Acta Radiol 1951;36:516–520.
- 264. Wada R, Lambert RGW. Deposition of intraosseous fat in a degenerating simple bone cyst. *Skeletal Radiol* 2005;34:415–418.
- 265. Wilner D. Radiology of bone tumors and allied disorders Philadelphia: WB Saunders, 1982:387.
- 266. Zorn DT, Cordray DR, Randels PH. Intraosseous lipoma of bone involving the sacrum. J Bone Joint Surg Am 1971;53: 1201–1204.

Fibrocartilaginous Mesenchymoma

- 267. Bhaduri A, Deshpande RB. Fibrocartilaginous mesenchymoma versus fibrocartilaginous dysplasia: are these a single entity? *Am J Surg Pathol* 1995;19:1447–1448.
- Bulichova LV, Unni KK, Bertoni F, Beabout JW. Fibrocartilaginous mesenchymoma of bone. Am J Surg Pathol 1993;17: 830–836.
- 269. Campanacci M. Bone and soft tissue tumors New York: Springer-Verlag, 1986:345-348.
- 270. Dahlin DC, Bertoni F, Beabout JW, Campanacci M: Fibrocartilaginous mesenchymoma with low grade malignancy. *Skeletal Radiol* 1984;12:263–269.
- 271. Ishida T, Dorfman HD. Massive chondroid differentiation in fibrous dysplasia of bone (fibrocartilaginous dysplasia). *AmJ Surg Pathol* 1993;17:924–930.
- 272. Mirra JM. Intramedullary cartilage- and chondroid-producing tumors. In: Mirra JM, Picci P, Gold RH, eds. *Bone tumors: clinical, radiologic, and pathologic correlations,* vol 1. Philadelphia: Lea & Febiger, 1989:439–690.

Adamantinoma of Long Bones

- Adler C-P. Case report 587. Adamantinoma of the tibia mimicking osteofibrous dysplasia. *Skeletal Radiol* 1990;19:55–58.
- 274. Alquacil-Garcia A, Alonso A, Pettigrew NM. Osteofibrous dysplasia (ossifying fibroma) of the tibia and fibula and adamantinoma. *Am J Clin Pathol* 1984;82:470–474.
- 275. Baker PL, Dockerty MD, Coventry MB. Adamantinoma (socalled) of the long bones. J Bone Joint Surg Am 1954;36: 704–720.
- 276. Bovée JVMG, van den Broek LJCM, De Boer WJ, Hogendoorn PCW. Expression of growth factors and their receptors in adamantinoma of long bones and the implication for its histogenesis. *J Pathol* 1998;184:24–30.
- 277. Bridge JA, Dembinski A, DeBoer J, et al. Clonal chromosomal abnormalities in osteofibrous dysplasia. Implications for histopathogenesis and its relationship with adamantinoma. *Cancer* 1994;73:1746–1752.
- Bridge JA, Fidler ME, Neff JR, et al. Adamantinoma-like Ewing's sarcoma: genomic confirmation, phenotypic drift. *Am J Surg Pathol* 1999;23:159–165.
- Campanacci M. Osteofibrous dysplasia of long bones. A new clinical entity. *Ital J Orthop Traumatol* 1976;2:221–237.

- Campanacci M, Laus M, Giunti A, et al. Adamantinoma of the long bones. The experience at the Istituto Ortopedico Rizzoli. *Am J Surg Pathol* 1981;5:533–542.
- 281. Czerniak B, Rojas-Corona RR, Dorfman HD. Morphologic diversity of long bone adamantinoma. The concept of differentiated (regressing) adamantinoma and its relationship to osteofibrous dysplasia. *Cancer* 1989;64:2319–2334.
- Eisenstein W, Pitcock JA. Adamantinoma of the tibia: an eccrine carcinoma. Arch Pathol Lab Med 1984;108:246–250.
- 283. Fischer B. Über ein primäres Adamantinom der Tibia. Frankfurter Z Pathol 1913;12:422-441.
- 284. Hauben E, van den Broek LC, Van Marck E, Hogendoorn PC. Adamantinoma-like Ewing's sarcoma and Ewing's-like adamantinoma. The t(11;22), t(21;22) status. *J Pathol* 2001;195: 218–221.
- Hazelbag HM, Fleuren GJ, Cornelisse CJ, et al. DNA aberrations in the epithelial cell component of adamantinoma of long bones. *Am J Pathol* 1995;147:1770–1779.
- 286. Hazelbag HM, Fleuren GJ, van den Broek LJ, et al. Adamantinoma of the long bones: keratin subclass immunoreactivity pattern with reference to its histogenesis. *Am J Surg Pathol* 1993; 17:1225–1233.
- 287. Hazelbag HM, Taminiau AH, Fleuren GJ, Hogendoorn PC. Adamantinoma of the long bones. A clinicopathological study of thirty-two patients with emphasis on histological subtype, precursor lesion, and biological behavior. *J Bone Joint Surg Am* 1994;76:1482–1499.
- 288. Hazelbag HM, van den Broek LJ, Fleuren GJ, et al. Distribution of extracellular matrix components in adamantinoma of long bones suggests fibrous-to-epithelial transformation. *Hum Pathol* 1997;28:183–188.
- 289. Hazelbag HM, Wessels JW, Mollevangers P, et al. Cytogenetic analysis of adamantinoma of long bones: further indications for a common histogenesis with osteofibrous dysplasia. *Cancer Genet Cytogenet* 1997;97:5–11.
- 290. Hogendoorn PWC, Hashimoto H. Adamantinoma. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone Lyon: IARC Press, 2002:332–334.
- 291. Huvos AG, Marcove RC. Adamantinoma of long bones: a clinicopathological study of fourteen cases with vascular origin suggested. J Bone Joint Surg Am 1975;57:148–154.
- 292. Ishida T, Iijima T, Kikuchi F, et al. A clinicopathological and immunohistochemical study of osteofibrous dysplasia, differentiated adamantinoma, and adamantinoma of long bones. *Skeletal Radiol* 1992;21:493–502.
- 293. Jundt G, Remberger K, Roessner A, et al. Adamantinoma of long bones. A histopathological and immunohistochemical study of 23 cases. *Pathol Res Pract* 1995;191:112–120.
- 294. Kahn LB. Review article. Adamantinoma, osteofibrous dysplasia and differentiated adamantinoma. *Skeletal Radiol* 2003;32: 245–258.
- 295. Kanamori M, Antonescu CR, Scott M, et al. Extra copies of chromosomes 7, 8, 12, 19, and 21 are recurrent in adamantinoma. J Mol Diagn 2001;3:16–21.
- Keeney GL, Unni KK, Beabout JW, Pritchard DJ. Adamantinoma of long bones. A clinicopathologic study of 85 cases. *Cancer* 1989;64:730–737.
- 297. Knapp RH, Wick MR, Scheithauer BW, Unni KK. Adamantinoma of bone. An electron microscopic and immunohistochemical study. *Virchows Arch A* 1982;398:75–86.
- 298. Kumar D, Mulligan ME, Levine AM, Dorfman HD. Classic adamantinoma in a 3-year-old. *Skeletal Radiol* 1998;27: 406–409.
- Llombart-Bosch A, Ortuno-Pacheco G. Ultrastructural findings supporting the angioblastic nature of the so-called adamantinoma of the tibia. *Histopathology* 1978;2:189–200.
- 300. Maki M, Athanasou N. Osteofibrous dysplasia and adamantinoma: correlation of proto-oncogene product and matrix protein expression. *Hum Pathol* 2004;35:69–74.
- Markel SF. Ossifying fibroma of long bone. Its distinction from fibrous dysplasia and its association with adamantinoma of long bone. *Am J Clin Pathol* 1978;69:91–97.
- 302. Moon NF. Adamantinoma of the appendicular skeleton. A statistical review of reported cases and inclusion of 10 new cases. *Clin Orthop* 1965;43:189–213.

- Moon NF, Mori H. Adamantinoma of the appendicular skeleton —updated. *Clin Orthop* 1986;204:215–237.
- 304. Povýšil C, Kohout A, Urban K, Horák M. Differentiated adamantinoma of the fibula: a rhabdoid variant. *Skeletal Radiol* 2004;33:488–492.
- 305. Rock MG, Beabout JW, Unni KK, Sim FH. Adamantinoma. Orthopedics 1983;6:472–477.
- 306. Rosai J. Adamantinoma of the tibia: Electron microscopic evidence of its epithelial origin. *Am J Clin Pathol* 1969;51: 786–792.
- 307. Rosai J, Pinkus GS. Immunohistochemical demonstration of epithelial differentiation in adamantinoma of the tibia. *Am J Surg Pathol* 1982;6:427–434.
- Schajowicz F, Santini-Araujo E. Adamantinoma of the tibia masked by fibrous dysplasia. Report of three cases. *Clin Orthop* 1989;238:294–301.
- Schajowicz F. Tumors and tumorlike lesions of bone: pathology, radiology, and treatment, 2nd ed. Berlin: Springer-Verlag, 1994:468–481.
- Springfield DS, Rosenberg AE, Mankin HJ, Mindell ER. Relationship between osteofibrous dysplasia and adamantinoma. *Clin Orthop* 1994;309:234–244.
- 311. Sweet DE, Vinh TN, Devaney K. Cortical osteofibrous dysplasia of long bone and its relationship to adamantinoma. A clinicopathologic study of 30 cases. *Am J Surg Pathol* 1992;16:282–290.
- Tehranzadeh J, Fanney D, Ghandur-Mnaymneh L, et al. Case report 517. Ulcerating adamantinoma of the tibia. *Skeletal Radiol* 1989;17:614–619.
- 313. Ueda Y, Roessner A, Bosse A, et al. Juvenile intracortical adamantinoma of the tibia with predominant osteofibrous dysplasia-like features. *Pathol Res Pract* 1991;187:1039–1043.
- 314. Unni KK, Dahlin DC, Beabout JW, Ivins JC. Adamantinoma of long bones. *Cancer* 1974;34:1796–1805.
- 315. Van der Woude H-J, Hazelbag H-M, Bloem JL, et al. MRI of adamantinoma of long bones in correlation with histopathology. Am J Roentgenol 2004;183:1737–1744.
- 316. Vigorita VJ, Ghelman B, Hogendoorn PCW. Osteofibrous dysplasia. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone Lyon: IARC Press, 2002:343–344.
- 317. Weiss SW, Dorfman HD. Adamantinoma of long bone. An analysis of nine new cases with emphasis on metastasizing lesions and fibrous dysplasia-like changes. *Hum Pathol* 1977;8: 141–153.
- 318. Yoneyama T, Winter WG, Milsow L. Tibial adamantinoma: its histogenesis from ultrastructural studies. *Cancer* 1977;40: 1138–1142.
- Zehr RJ, Recht MP, Bauer TW. Adamantinoma. Skeletal Radiol 1995;24:553–555.

Chordoma

- Abenoza P, Sibley RK. Chordoma: an immunohistochemical study. *Hum Pathol* 1987;17:744–747.
- 321. Azzarelli A, Quagliuolo V, Cerasoli S, et al. Chordoma: natural history and treatment results in 33 cases. J Surg Oncol 1988;37: 185–191.
- Beaugié JM, Mann CV, Butler ECB. Sacrococcygeal chordoma. Br J Surg 1969;56:586–588.
- Belza MG, Urich H. Chordoma and malignant fibrous histiocytoma. Evidence of transformation. *Cancer* 1986;589:1082–1087.
- Byers PD. A study of histological features distinguishing chordoma from chondrosarcoma. *Br J Cancer* 1981;43:229–232.
- 325. Chauvel A, Taillat F, Gille O, et al. Giant vertebral notochordal rest: a new entity distinct from chordoma. *Histopathology* 2005;47:646–649.
- 326. Chu TA. Chondroid chordoma of the sacrococcygeal region. Arch Pathol Lab Med 1987;111:861–864.
- Coindre JM, Rivel J, Trojan M, et al. Immunohistological study in chordomas. J Pathol 1986;150:61–63.
- 328. Coombs RJ, Coiner L. Sacral chordoma with unusual posterior radiographic presentation. *Skeletal Radiol* 1996;25:679–681.
- Crawford T. The staining reactions of chordoma. J Clin Pathol 1958;11:110–113.
- Dabska M. Parachordoma. A new clinicopathologic entity. Cancer 1977;40:1586–1592.

- Dahlin DC, MacCarty CS. Chordoma. A study of fifty-nine cases. Cancer 1952;5:1170–1178.
- Dahlin DC, Unni KK. Bone tumors: general aspects and data on 8,542 cases Springfield, IL: Charles C Thomas, 1986:379–393.
- deBruine FT, Kroon HM. Spinal chordoma: radiologic features of 14 cases. Am J Roentgenol 1988;150:861–863.
- Disler DG, Miklic D. Imaging findings in tumors of the sacrum. Am J Roentgenol 1999;173:1699–1706.
- 335. Falconer MA, Bailey IC, Duchen LW. Surgical treatment of chordoma and chondroma of the skull base. J Neurosurg 1968;29:261–275.
- Fechner RE, Mills SE. *Tumors of the bones and joints* Washington, DC: Armed Forces Institute of Pathology, 1993.
- 337. Firooznia H, Printo RS, Lin JP, et al. Chordoma: radiologic evaluation of 20 cases. Am J Roentgenol 1976;127:797–805.
- 338. Fisher C. Parachordoma exists—but what is it? *Adv Anat Pathol* 2000;7:141–148.
- Fisher H-P, Alles JU, Stambolis C. Skeletal chordoma. Clinicopathological and differential diagnostic aspects. *Pathologe* 1983; 4:307–312.
- 340. Folpe AL, Agoff SN, Willis J, Weiss SW. Parachordoma is immunohistochemically and cytogenetically distinct from axial chordoma and extraskeletal myxoid chondrosarcoma. *Am J Surg Pathol* 1999;23:1059–1067.
- Friedman I, Harrison DFN, Bird ES. The fine structure of chordoma with particular reference to the physaliphorous cell. *J Clin Pathol* 1962;15:116–125.
- 342. Healey JH, Lane JM. Chordoma: a critical review of diagnosis and treatment. *Orthop Clin North Am* 1989;20:417–426.
- Higginbotham NL, Phillips RF, Farr HW, Hustu HO. Chordoma. Thirty-five-year study at Memorial Hospital. *Cancer* 1967; 20:1841–1850.
- 344. Hruban RH, Traganos F, Reuter VE, Huvos AG. Chordomas with malignant spindle cell components. Am J Pathol 1990; 137:435–447.
- Hudson TM. Radiologic-pathologic correlation of musculoskeletal lesions Baltimore: Williams & Wilkins, 1987:287–303.
- Huvos AG. Bone tumors: diagnosis, treatment, and prognosis, 2nd ed. Philadelphia. WB Saunders, 1991:599–624.
- 347. Kay S, Schatzki PF. Ultrastructural observations of a chordoma arising in the clivus. *Hum Pathol* 1972;3:403–413.
- Krol G, Sundaresan N, Deck M. Computed tomography of axial chordomas. J Computed Assist Tomogr 1983;7:286–289.
- Kyriakos M, Totty WG, Lenke LG. Giant vertebral notochordal rest: a lesion distinct from chordoma: discussion of an evolving concept. *Am J Surg Pathol* 2003;27:396–406.
- Li MK, Folpe AL. CDX-2, a new marker for adenocarcinoma of gastrointestinal origin. *Adv Anat Pathol* 2004;11:101–105.
- Llauger J, Palmer J, Amores S, et al. Primary tumors of the sacrum: diagnostic imaging. Am J Roentgenol 2000;174:417–424.
- 352. Lucas D, Heim S. Extraskeletal myxoid chondrosarcoma. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone Lyon: IARC Press, 2002:213–215.
- 353. Makek K, Leu HJ. Malignant fibrous histiocytoma arising in a recurrent chordoma. Case report and electron microscopic findings. *Virchows Arch A* 1982;397:241–250.
- 354. Martin RF, Melnick PJ, Warner NE, et al. Chordoid sarcoma. Am J Clin Pathol 1973;59:623–635.
- 355. Meis JM, Giraldo AA. Chordoma. An immuno-histochemical study of 20 cases. Arch Pathol Lab Med 1988;112:553–556.
- 356. Meis JM, Raymond AK, Evans HL, et al. Dedifferentiated chordoma. A clinico-pathologic and immunohistochemical study of three cases. *Am J Surg Pathol* 1987;11:516–525.
- 357. Meyer JE, Lepke RA, Lindfors KK, et al. Chordomas: their CT appearance in the cervical, thoracic and lumbar spine. *Radiol*ogy 1984;153:693–696.
- 358. Miettinen M, Karaharju E, Järvinen H. Chordoma with a massive spindle-cell sarcomatous transformation. A light- and electron-microscopic and immunohistological study. *Am J Surg Pathol* 1987;11:563–570.
- 359. Miozzo M, Dalpra L, Riva P, et al. A tumor suppressor locus in familial and sporadic chordoma maps to 1p36. *Int J Cancer* 2000;87:68–72.

- 360. Mirra JM, Brien EW. Giant notochordal hamartoma of intraosseous origin: a newly reported benign entity to be distinguished from chordoma. Report of two cases. *Skeletal Radiol* 2001;30:698–709.
- Mirra JM, Nelson SM, Della Rocca C, et al., eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone Lyon: IARC Press, 2002:316–317.
- Mulder JD, Kroon HM, Schütte HE, Taconis WK. Radiologic atlas of bone tumors Amsterdam: Elsevier, 1993:267–274.
- 363. Pallini R, Maira G, Pierconti F, et al. Chordoma of the skull base: predictors of tumor recurrence. *J Neurosurg* 2003;98: 812–822.
- 364. Pardo-Mind FJ, Guillen FJ, Villas C, Vasquez JJ. A comparative ultrastructural study of chondrosarcoma, chordoid sarcoma, and chordoma. *Cancer* 1981;47:2611–2619.
- 365. Rich TA, Schiller A, Suit HD, Mankin HJ. Clinical and pathologic review of 48 cases of chordoma. *Cancer* 1985;56:182–187.
- 366. Risio M, Bagliani C, Leli R, et al. Sacrococcygeal and vertebral chordomas. Report of three cases and review of the literature. J *Neurosurg Sci* 1985;29:211–227.
- 367. Rosenthal DI, Scott JA, Mankin HJ, et al. Sacrococcygeal chordoma: magnetic resonance imaging and computed tomography. Am J Roentgenol 1985;145:143–147.
- Sandberg AA. Genetics of chondrosarcoma and related tumors. Curr Opin Oncol 2004;16:342–354.
- Scolyer RA, Bonar SF, Palmer AA, et al. Parachordoma is not distinguishable from axial chordoma using immunohistochemistry. *Pathol Int* 2004;54:364–370.
- Smith J, Ludwig RL, Marcove RC. Sacrococcygeal chordoma. A clinicoradiological study of 60 patients. *Skeletal Radiol* 1987; 16:37–44.
- 371. Smith J, Reuter V, Demas B. Case report 576. Anaplastic sacrococcygeal chordoma (dedifferentiated chordoma). *Skeletal Radiol* 1989;18:561–564.
- 372. Sze G, Uichanco LS III, Brant-Zawadzki MN, et al. Chordomas: MR imaging. *Radiology* 1988;166:187–191.
- 373. Taylor JR. Persistence of the notochordal canal in vertebrae. J Anat 1982;11:211–217.
- 374. Thiery JP, Mazabraud A, Mignot J, Durigon M. Electron microscopic study of a sacral chordoma. Characterization of various evolutive stages of the tumoral cells. *Ann Anat Pathol* 1977; 22:193–204.
- 375. Turner ML, Mulhern CB, Dalinka MK. Lesions of the sacrum. Differential diagnosis and radiological evaluation. *JAMA* 1981; 245:275–277.
- Uhrenholt L, Stimpel H. Histochemistry of sacrococcygeal chordoma. Acta Pathol Microbiol Scand A 1985;93:203–204.
- Weiss SW. Ultrastructure of the so-called chordoid sarcoma. Evidence supporting cartilaginous differentiation. *Cancer* 1976;37: 300–306.
- 378. Yamaguchi T, Suzuki S, Ishiiwa H, et al. Benign notochordal cell tumors: A comparative histological study of benign notochordal cell tumors, classic chordomas, and notochordal vestiges of fetal intervertebral discs. *Am J Surg Pathol* 2004;28:756–761.
- 379. Yamaguchi T, Suzuki S, Ishiiwa H, Ueda Y. Intraosseous benign notochordal cell tumours: overlooked precursors of classic chordomas? *Histopathology* 2004;44:597–602.
- 380. Yamaguchi T, Yamato M, Saotome K. First histologically confirmed case of a classic chordoma arising in a precursor benign notochordal lesion: differential diagnosis of benign and malignant notochordal lesions. *Skeletal Radiol* 2002;31:413–418.
- Yuh WT, Flickinger FW, Barloon TJ, Montgomery WJ. MR imaging of unusual chordomas. J Comput Assist Tomogr 1988;12: 30–35.

Leiomyosarcoma of Bone

- Abdelwahab IF, Hermann G, Kenan S, et al. Case report 794. Primary leiomyosarcoma of the right femur. *Skeletal Radiol* 1993;22:379–381.
- Abdelwahab IF, Kenan S, Hermann G, et al. Radiation-induced leiomyosarcoma. *Skeletal Radiol* 1995;24:81–83.
- 384. Angervall L, Berlin O, Kindblom LG, Stener B. Primary leiomyosarcoma of bone. A study of five cases. *Cancer* 1980;46: 1270–1279.
- 385. Berlin O, Angervall L, Kindblom LG, et al. Primary leiomyosarcoma of bone. A clinical, radiographic, pathologic-anatomic, and prognostic study of 16 cases. *Skeletal Radiol* 1987;16: 364–376.
- Bush CH, Reith JD, Spanier SS. Mineralization in musculoskeletal leiomyosarcoma: Radiologic-pathologic correlation. *Am J Roentgenol* 2003;180:109–113.
- 387. Eady JL, McKinney JD, McDonald EC. Primary leiomyosarcoma of bone. *J Bone Joint Surg Am* 1987;69:287–289.
- 388. Fornasier VL, Paley D. Leiomyosarcoma in bone: primary or secondary? A case report and review of the literature. *Skeletal Radiol* 1983;10:147–153.
- Goto T, Ishida T, Motoi N, et al. Primary leiomyosarcoma of the femur. J Orthop Sci 2002;7:267–273.
- 390. Hasegawa T, Hirose T, Seki K, et al. Histological and immunohistochemical diversities, and proliferative activity and grading in osteosarcomas. *Cancer Detect Prev* 1997;21:280–287.
- Inoue S, Tanaka K, Sakamot A, et al. Primary leiomyosarcoma of the patella. *Skeletal Radiol* 2001;30;530–533.
- Jundt G, Moll C, Nidecker A, et al. Primary leiomyosarcoma of bone: report of eight cases. *Hum Pathol* 1994;25:1205–1212.
- 393. Kawai T, Suzuki M, Mukai M, et al. Primary leiomyosarcoma of bone. An immunohistochemical and ultrastructural study. Arch Pathol Lab Med 1983;107:433–437.
- 394. Meister P, Konrad E, Gokel JM, Remberger K. Case report 59. Leiomyosarcoma of the humerus. *Skeletal Radiol* 1978;2: 265–267.

- 395. Myers JL, Arocho J, Bernreuter W, et al. Leiomyosarcoma of bone. A clinicopathologic, immunohistochemical, and ultrastructural study of five cases. *Cancer* 1991;67:1051–1056.
- 396. Overgaard J, Frederiksen P, Helmig O, Jensen OM. Primary leiomyosarcoma of bone. *Cancer* 1977;39:1664–1671.
- 397. Remadi S, Samson J, Sando Z, Kuffer R. Mandibular osteosarcoma with unusual expression of alpha-actin smooth muscle antibody. *Pathol Res Pract* 1996;192:148–153.
- 398. Sanerkin NG. Primary leiomyosarcoma of the bone and its comparison with fibrosarcoma. A cytological, histological, and ultrastructural study. *Cancer* 1979;44:1375–1387.
- Shamsuddin AK, Reyes F, Harvey JW, Toker C. Primary leiomyosarcoma of bone. *Hum Pathol* 1980;11:581–583.
- 400. Shen S-H, Steinbach LS, Wang S-F, et al. Primary leiomyosarcoma of bone. *Skeletal Radiol* 2001;30:600–603.
- Sundaram M, Akduman I, White LM, et al. Primary leiomyosarcoma of bone. Am J Roentgenol 1999;172:771–776.
- 402. Von Hochstetter AR, Eberle H, Ruettner JR. Primary leiomyosarcoma of extragnathic bones. *Cancer* 1984;53:2194–2200.
- 403. Wang TY, Erlandson RA, Marcove RC, Huvos AG. Primary leiomyosarcoma of bone. Arch Pathol Lab Med 1980;104:100–104.
- 404. Yamamura H, Yoshikawa H, Tatsuta M, et al. Expression of the smooth muscle calponin gene in human osteosarcoma and its possible association with prognosis. *Int J Cancer* 1998;79: 245–250.
- 405. Young MPA, Freemont AJ. Primary leiomyosarcoma of bone. Histopathology 1991;19:257–262.

8

Metastases

OSSEOUS METASTASES 458

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Osseous Metastases

Skeletal metastases are the most common variety of bone tumors and should always be considered in the differential diagnosis, particularly in older patients (47,65). In a series of 1,000 consecutive autopsies of patients who died of carcinoma, metastases to bone were found in 27% (2). Approximately 70% of all malignant bone tumors are metastatic in origin (128). The incidence varies according to the type of primary neoplasm and the duration of disease. Some malignant tumors demonstrate a far greater predilection for osseous involvement than others. Cancers of the breast, prostate, lung, and kidney account for 80% of all metastatic cancers to bone (67). In men, carcinoma of the prostate is reported to account for almost 60% and carcinoma of the lung for 25% of all bone metastases. In women, carcinoma of the breast is responsible for almost 70% of all metastatic lesions, the remaining 30% being mainly due to carcinomas of the thyroid, uterus, and kidney (1,87). Other primary tumors responsible for bone metastases include carcinomas of the stomach, colon, urinary bladder, melanoma, and some neurogenic tumors (121). Some sarcomas, such as osteosarcoma and Ewing sarcoma, may occasionally metastasize to bones. In children aged 5 years and younger, neuroblastoma is usually the primary tumor responsible for metastatic disease. Because of their frequency, cancers of the breast, lung, and prostate are responsible for the majority of bone metastases.

In the past it was thought that the usual route of metastasis is via tumor emboli, mostly carried through the lymphatic system and the ductus thoracicus to the central blood circulation, and then by the arterial system into the periphery (11). This belief was in contrast to the ancient hypothesis of the British surgeon Stephan Paget (96), who, based on a study of more than 900 autopsy records, proposed that metastasis is not a random process, but the tumor cells (which he regarded as "seeds") need a special environment (the "soil") to grow and form metastatic lesions. In recent years this theory has gained wide acceptance and was substantiated by experimental data (for additional information see references 33, 83, 84, 106, and 115). Tumor cells seem to acquire a special "genetic signature" that enables them to metastasize (104). In addition, the microenvironment in bone, especially marrow stem cells, supports cancer cells in homing, differentiation, and survival. Conversely, cancer cells influence osteoblasts and osteoclasts by secreted factors such as parathyroid hormone-related peptide

(PTHrP) or endothelin I (17,84,129). This leads to osteolytic or osteoblastic metastases in bone; however, even osteoblastic metastases are accompanied by increased bone resorption, as is clinically evident by the treatment response to bisphosphonates in osteoblastic metastases of prostate cancer. Furthermore, bone contains an abundance of growth factors (e.g., transforming growth factor β , insulin-like growth factors I and II, fibroblast growth factors, and bone morphogenetic proteins) that are released into the microenvironment during the resorption process (55). Stimulated tumor cells release factors that induce osteoblasts to secrete RANK (receptor activator of nuclear factor kappa b)-ligand or RANKL, which is a potent factor for osteoclast formation and activity. Osteoclasts, in turn, resorb bone and thus release additional growth factors that enhance the accumulation of cancer cells (17).

Clinical Presentation

Metastases are relatively rare in patients younger than the age of 40. Therefore, patient age is an important discriminating factor in the diagnosis (81). Unlike primary bone tumors, metastases usually involve the axial skeleton (skull, spine, and pelvis) and the most proximal segments of limb bones. Only in extremely rare cases do metastatic tumors occur distal to the elbows and the knees (82,97). Most metastatic lesions are found in the vertebrae, ribs, pelvis, skull, femur, and humerus (81) (Fig. 8-1).

The majority of skeletal metastases are silent. When metastases are symptomatic, pain is the main clinical feature, although rarely a pathologic fracture through the lesion may focus attention on the disease (98). Clinical and laboratory examinations frequently disclose other signs of malignancy, such as weight loss, anemia, fever, and an elevated erythrocyte sedimentation rate. In addition, serum calcium and alkaline phosphatase levels are usually elevated (125). In the spine, metastases are frequently associated with neurologic symptoms caused by compression of the cord or nerve roots or even by complete obstruction of the thecal sac.

Imaging

Detection of skeletal metastases is not always possible on routine radiography, because destruction of the bone may not be visible (6,113,127). It has been estimated that 30% to 50% of normal bone mineral must be lost before a bone metastasis becomes visible on a radiograph (9). Radionuclide bone scan, including both planar and single proton emission computed tomography (SPECT) imaging (15), is the best screening method for early detection of metastatic tumors and can detect lytic and sclerotic (blastic) lesions (Fig. 8-2). Computed tomography (CT), magnetic resonance imaging (MRI), and fluorodeoxyglucose (FDG) positron emission tomography (PET) scanning may also be helpful, the latter modality being the most sensitive (23,29,113). On radiography, a metastatic lesion may resemble any of the benign or malignant lesions. There



Figure 8-1 Skeletal metastases: sites of predilection and peak age range.





Figure 8-2 Skeletal metastases: scintigraphy. A radionuclide bone scan was performed on a 68-year-old woman with breast carcinoma to determine the presence of metastases. After an intravenous injection of 15 mCi (555 MBq) of technetium-99m diphosphonate (MDP), an increased uptake of the radiopharmaceutical tracer is seen in the skull and cervical spine **(A)**, and lumbar spine and pelvis **(B)**, localizing the multiple metastases. **C:** In another patient, a 55-year-old man with bronchogenic carcinoma, a radionuclide bone scan disclosed metastases to the cervical and thoracic spine, sternum, and right femur.



Figure 8-3 Origin of osteolytic metastases.

are no radiographic characteristics of metastasis. The type of bone destruction may be geographic, motheaten, or permeative, and the margins may be well or poorly defined. A periosteal reaction and a soft tissue mass may or may not be present, although the latter situation is more common. Metastases can be solitary or multiple, and they can be further subdivided into purely lytic, purely sclerotic, and mixed lesions. This distinction can be of considerable aid in analysis of the site of origin of the primary tumor (27). The primary sources of these lesions are as follows:

- 1. *Lytic lesions*. Osteolytic metastases are the most common, representing about 75% of all metastatic lesions. The primary source is usually a carcinoma of the kidney, lung, breast, gastrointestinal (GI) tract, or thyroid (35) (Figs. 8-3 and 8-4).
- Sclerotic lesions. Osteoblastic metastases represent approximately 15% of all metastatic lesions. In men they are caused mainly by a prostatic gland cancer or a seminoma (21) (Fig. 8-5A). In women the primary source is usually carcinoma of the breast, uterus (particularly cervix), or ovary (93) (Fig. 8-5B). In both genders, metastases may originate from carcinoid tumors (88,100,101), bladder tumors (32), certain neurogenic tumors, including medulloblastoma (121), and osteosarcoma (47,73) (Fig. 8-6).



Figure 8-4 Osteolytic metastases. A: Osteolytic metastasis to the proximal femur from carcinoma of the colon in a 52-yearold woman. B: Osteolytic metastasis to the left ilium from carcinoma of the thyroid in an 83-year-old man.



Figure 8-5 Sclerotic metastases. A: Multiple sclerotic foci are scattered through the ilium, pubis, ischium, and proximal femur of a 55-year-old man with carcinoma of the prostate. B: Sclerotic metastases to the pubis, ischium, and proximal femur in a 57-year-old woman with breast carcinoma.

3. *Mixed lesions*. Mixed osteoblastic and osteolytic lesions represent approximately 10% of all metastatic lesions. Any primary tumor can give rise to mixed metastases, the most common primaries being breast and lung tumors. Occasionally, some primary neoplasms may give rise to both lytic and sclerotic metastases (Fig. 8-7).

Although metastases in the skeleton may appear similar, regardless of their primary source, in some instances their distribution, location, or morphologic appearance may suggest a site of origin (7,43,56,76,90, 112,116). For example, 50% of skeletal metastases distal to the elbows and the knees, although unusual, originate from either bronchogenic or breast carcinoma (85) (Fig. 8-8). Although extremely rare, metastases to the hands and feet, particularly those affecting the distal phalanges (acrometastases), again indicate a primary tumor in the lung, less commonly in the breast or kidney (97) (Fig. 8-9). An extensive osteolysis of the distal phalanx, often accompanied by a soft tissue mass, is the usual presentation of this type of metastasis from bronchogenic carcinoma. Some time ago, characteristic cortical metastases were reported from bronchogenic carcinoma (27,28,48-50). These lesions produce a type of

cortical destruction termed "cookie-bite" or "cookiecutter" lesions (Fig. 8-10). The spread of malignant cells to involve the skeleton usually takes place via the hematogenous route. In such instances, the bulk of the tumor lodges in the marrow and spongy bone. Therefore, the initial radiography of metastatic lesions in the skeleton reveals the destruction of cancellous bone; only after further tumor growth is the cortex affected. An anastomosing vascular system in the cortex, originating in the overlying periosteum (118), probably serves as the pathway by which malignant cells from bronchogenic carcinoma reach the compact bone and initiate cortical destruction (39). Primary carcinomas of the kidney and bladder and melanoma may also give rise to cortical metastases (20,36,59). It is of interest that the majority of cortical metastases affect the femur (20,50).

In certain instances, the morphologic appearance of a metastasis may suggest a specific site of origin or may at least narrow the differential diagnostic possibilities (27). For example, bubbly, highly expansive, so-called blow-out metastatic lesions originate from a primary carcinoma of the kidney or thyroid (Fig. 8-11). Multiple round, dense foci or diffuse increases in bone density are often seen in metastatic carcinoma of the prostate (see Fig. 8-5A).





Figure 8-6 Origin of osteoblastic metastases.

MRI is occasionally useful in determining the origin of the metastatic lesion. Recently, Choi et al. (18) reported a "flow-void" sign apparently characteristic for metastatic renal carcinoma. Reviewing 20 osseous metastatic lesions from renal cell carcinoma, the investigators noted within a mass of intermediate signal intensity the frequent appearance of multiple dot-like or tubular structures exhibiting low signal intensity on T1-weighted sequences. These structures showing the flow-void signal corresponded to dilated blood vessels (arteries supplying the hypervascular tumor and veins draining the lesion). Although the specificity of the flow-void sign for diagnosis of metastatic renal carcinoma is as yet unknown, its presence on MR images might suggest this diagnosis.

In general, metastatic bone disease is characterized by a combination of bone resorption and bone formation. Radiographic imaging of the lesions will reveal the predominant process (12,95). When osteolysis predominates, the lesions appear lytic, and when bone formation is dominant, they appear sclerotic (38,105). Multiple sclerotic metastases may present either in a focal pattern (multiple snowball appearance) or may have a diffuse pattern (generalized radiopacity of bones) (31,60). Bone formation in metastatic disease involves either a stromal or a reactive mechanism. Stromal new bone occurs only in lesions associated with the development of a surrounding fibrous stroma, as in carcinoma of the prostate, and reflects intramembranous ossification of areas within the stroma. In contrast, reactive bone represents an attempt to repair the injury caused by tumor cells. This type of formation occurs to some extent in all metastatic lesions, although it tends to be negligible in highly anaplastic or rapidly growing tumors (105). Bone destruction is always mediated by tumorinduced osteoclastic resorption. Primary tumors responsible for purely osteolytic metastases are usually located in the kidneys, lung, breast, thyroid, or GI tract, whereas primaries responsible for osteoblastic metastases are usually located in the prostate gland. It must be pointed out, however, that after treatment (radiation therapy, chemotherapy, or hormonal therapy), purely lytic lesions may become sclerotic.

Scintigraphy is almost invariably positive in bone metastases (91), and increased uptake is observed in both sclerotic and lytic lesions (16,19,30,44,78,79,94,99) (see Fig. 8-2). This phenomenon is secondary to the

Figure 8-7 Osteolytic and sclerotic metastases. Osteolytic metastasis in the medial end of the clavicle (*arrows*) and sclerotic metastasis in the humeral head (*open arrow*) in a 27-year-old woman with a bronchial carcinoid tumor.





Figure 8-8 Metastases distal to the elbows and the knees. A: Diffuse osteolytic metastases to the ulna in a 66-year-old woman with breast carcinoma. **B:** Osteolytic metastasis to the midshaft of the right fibula of a 41-year-old woman with hypernephroma.



Figure 8-9 Acrometastases. A: Osteolytic metastasis to the proximal phalanx of the left thumb in a 63-year-old man with bron-chogenic carcinoma. **B:** Osteolytic metastasis to the distal phalanx of the right thumb (*arrow*) in a 50-year-old woman with breast carcinoma.



465

F





Figure 8-11 Skeletal metastases. A 52-year-old man with renal cell carcinoma (hypernephroma) developed a solitary metastatic lesion in the acromial end of the left clavicle. **A:** Radiograph of the left shoulder shows an expansive blown-out lesion associated with a soft tissue mass destroying the distal end of the clavicle. **B:** Subtraction study of a selective left subclavian arteriogram demonstrates hypervascularity of the tumor, a characteristic feature of metastatic hypernephroma. **C:** In another patient, a 59-year-old woman with hypernephroma, a blown-out lesion is associated with a soft tissue mass destroying the acromial end of the right clavicle, acromion, and glenoid.

increased bone turnover and reactive repair at the periphery of the lesion (3). Radionuclide bone scan is helpful for distinguishing metastatic disease from multiple myeloma because the latter usually presents with a normal uptake of a tracer (77). Only a few cases have been reported that exhibited negative bone scintigraphy in metastatic disease of the skeleton despite positive radiographic findings (14,68). This can be explained by the fact that bone density, as imaged by conventional radiography, reflects net metabolic activity over an extended period of time and does not necessarily reflect current metabolic activity. Therefore, a radiographically sclerotic lesion with a low metabolic rate can produce a negative bone scan (25). Occasionally, widespread metastatic disease produces a diffusely increased uptake throughout the skeleton rather than discrete hot spots. This socalled superscan appearance is identified by the abnormally intense bone uptake or by the absence of a normally visible activity in the kidneys, bladder, and soft tissues (10). Sometimes metastases cause cold spots (photopenic defects) when there is bone destruction but insignificant reactive bone formation; this may be observed in metastases from lung and breast carcinoma (10,57,61,70).

С

CT can contribute to the diagnosis of metastatic disease by frequently revealing a definite destructive lesion in bone in patients who have an abnormal radionuclide bone scan, a normal or inconclusive radiograph, or both (13,54,58,69,72,80,102,103). This technique can also detect an associated soft tissue mass (Figs. 8-12 and 8-13).

MRI is more sensitive than technetium scans for detection of metastatic disease (8,26,29,42,117). Focal lytic metastases are characterized by a low signal on T1weighted sequences, becoming bright on T2-weighting (4,24,119) (Fig. 8-14). Focal sclerotic metastases, such as those from a primary carcinoma in breast or prostate, induce an osteoblastic reaction with new bone formation. Therefore, both T1- and T2-weighted images reveal a low signal intensity.

Histopathology

On microscopic examination, two aspects must be considered. The first is that of the tumor tissue itself, which depends on the origin of the primary tumor. Metastatic tumor is often histologically identical or very similar to the primary, thus enabling accurate identification. In some cases, however, the tissue may be highly undifferen-



Figure 8-12 Skeletal metastasis: computed tomography (CT). A: An anteroposterior radiograph of the left hip of a 50-year-old man with hypernephroma shows an osteolytic lesion almost completely destroying the ischium (*arrows*). **B:** CT section demonstrates the extent of bone destruction and a soft tissue extension of metastasis.



A,B

Α

Figure 8-13 Skeletal metastasis: computed tomography (CT). Anteroposterior **(A)** and lateral **(B)** radiographs of the distal arm of a 78-year-old man with bronchogenic carcinoma show an osteolytic lesion in the posterolateral cortex of the distal humerus associated with periosteal reaction. **C:** CT section shows cortical destruction, periosteal reaction, and a soft tissue mass.

В

С



Figure 8-14 Skeletal metastasis: magnetic resonance imaging (MRI). A: Anteroposterior radiograph of the left hip shows a diffuse osteolytic metastatic lesion in the proximal femur of a 60-year-old woman with breast carcinoma. B: Coronal T2*-weighted (MPGR, TR 550, TE 15, flip angle 15 degrees) MRI demonstrates increased signal of the lesion. The uninvolved bone marrow remains of low signal intensity.

tiated, and its primary source therefore cannot be determined (67,114) (Fig. 8-15). The second histologic aspect is the effect of the metastasis on the bone, which constitutes a combination of reactive bone destruction and reactive proliferation (111). Some metastatic tumors are characterized primarily by bone destruction, others by bone proliferation, and still others exhibit a mixed reaction (Fig. 8-16). These effects appear on radiography as lytic, blastic, or mixed lesions, respectively (61).

A variety of osteoclast-stimulating factors are secreted by malignant cells (32). Secretion of these substances by metastatic cells probably increases the number and activity of osteoclasts, and thus induces bone resorption (17). Although malignant cells can occasionally be observed in resorption bays on bone surfaces, no resorption lacunae corresponding to the size of tumor cells could be demonstrated by electron microscopy.

The increased bone apposition in osteoblastic metastases exhibits two patterns, which are usually observed in association with each other. New osteoid is produced on the surface of existing cancellous bone, resulting in trabeculae that are thicker than normal. Osteoblasts are also stimulated to form new trabeculae in marrow spaces that lie between preexisting medullary bone. Cultured prostate cancer cells have been demonstrated to secrete osteoblast-stimulating factor (32).



Figure 8-15 Histopathology of metastasis. A photomicrograph of a needle biopsy specimen obtained from a vertebral body with a sclerotic lesion suspected to be metastatic. There is evidence of reactive bone formation as well as fibrous scarring, and a conglomerate of atypical cells, strongly suggestive of a tumor; however, the definite diagnosis as to the origin of this lesion is difficult (hematoxylin and eosin, original magnification \times 400). (Reprinted from Bullough PG. *Atlas of orthopedic pathology*, 2nd ed. New York: Gower, 1992:17.28.)

B



Figure 8-16 Histopathology of metastasis. A: Osteoblastic metastasis from a moderately differentiated adenocarcinoma of the prostate shows tumor-filled marrow spaces and thick-ened trabeculae (hematoxylin and eosin, original magnification $\times 100$). **B:** Tumor cells react with an antibody against prostate-specific antigen (PSA) (biotin-streptavidin peroxidase, original magnification $\times 200$). **C:** Trabecula of lamellar bone (*center left to right*) with attached cell-rich layer of new woven bone on upper surface and rim of osteoblasts on lower surface. Metastasis from moderately differentiated adenocarcinoma of prostate is present in the marrow space (*top*) (van Gieson, original magnification $\times 200$).





Differential Diagnosis

Radiology

There are no characteristic radiologic features of metastasis. A metastatic lesion may look like a primary benign or malignant tumor, like a focus of infection, like a metabolic disease, or even like a post-traumatic abnormality. The length of the lesion is often a helpful clue because long lesions (10 cm or greater) frequently represent a primary malignant tumor, whereas most metastatic lesions are smaller, between 2 and 4 cm in length. A recent study evaluated the usefulness of MRI in discriminating osseous metastases from benign lesions. The so-called bull's eye sign (a focus of high signal intensity in the center of an osseous lesion) proved to be a sign of a benign disease, whereas a halo sign (a rim of high signal intensity around an osseous lesion) proved to be a sign of metastasis (109).

From the practical point of view and to facilitate differential diagnosis, a division of metastatic lesions into those with a single focus or multiple foci, and further subdivision into lytic, sclerotic, or mixed, with further classification according to the particular site in the bone, should be attempted.

SOLITARY METASTASIS

A single metastatic bone lesion must be distinguished from primary benign (and tumor-like) lesions and malignant tumors (74). In addition to the length of the lesion (see previous), which can help in distinguishing a primary from a metastatic tumor, other helpful features are the presence (or absence) of a periosteal reaction and a soft tissue mass. A metastatic lesion usually presents without or with only a small soft tissue mass, although exceptions to this rule have been reported (Fig. 8-17, see also Figs. 8-11 and 8-12). A periosteal reaction is usually absent unless the mass breaks through the cortex. In some reported series, however, more than 30% of metastatic lesions demonstrated a periosteal response (122), particularly metastases from carcinoma of the prostate (5,75,89). In the spine, metastatic lesions usually destroy the pedicle (Fig. 8-18), a useful feature for distinguishing this lesion from myeloma or neurofibroma eroding the vertebral body (5,64).

Lytic Metastasis

Osteolytic metastasis, particularly at the articular end of a long bone or affecting apophyseal parts (such as the femoral trochanter or humeral tuberosity), must be



Figure 8-17 Skeletal metastasis with soft tissue mass. A 70-year-old woman with breast carcinoma developed a skeletal metastasis to the thoracic vertebra. Note a large associated soft tissue mass.



Figure 8-18 Skeletal metastases. Metastases from bronchogenic carcinoma in a 45-year-old woman destroyed the right pedicles of vertebrae T8 and T10 (*arrows*).

differentiated from a *giant cell tumor* and an *intraosseous* ganglion. The latter invariably exhibits a sclerotic border, a rare feature of metastatic lesions. At other sites, the differential diagnosis must include primary malignant bone tumors such as *fibrosarcoma*, *malignant fibrous* histiocytoma (MFH), lymphoma, and plasmacytoma and some benign conditions such as hemangioma, brown tumor of hyperparathyroidism, and pseudotumor of hemophilia. As mentioned previously, differentiation solely on the basis of radiologic appearance is difficult, and clinical information is usually helpful.

Sclerotic Metastasis

A solitary osteoblastic metastasis should be differentiated from the *bone island* (*enostosis*). On radiography, bone islands exhibit characteristic thorny radiations that blend imperceptibly with the normal trabeculae of the host bone, a feature not present in metastasis (92). Further confirmation is provided by the radionuclide bone scan, which is invariably positive in metastatic disease but is usually normal in bone islands (51). Occasionally, however, errors in diagnosis can be made, particularly if a bone island is very large (giant bone island) (53) or if it exhibits an increased uptake of the radiopharmaceutical tracer (52) (see Chapter 2).

At times, a sclerotic metastasis with exuberant sunburst periosteal reaction (75,89), such as metastasis from a prostatic carcinoma, can simulate the appearance of *osteosarcoma* (62).

A single sclerotic lesion at the medial (sternal) end of the clavicle, which often is mistaken for a metastasis, may in fact represent *condensing osteitis* of the clavicle. Clinically, this condition manifests with pain, as well as local swelling and tenderness. Radiography reveals a homogeneously dense sclerotic patch, usually limited to the inferior margin of the medial end of the clavicle (Fig. 8-19A, B). Although this part of the clavicle may be slightly expanded, periosteal reaction and bone destruction are absent (46). CT reveals an obliteration of the marrow cavity, and radionuclide bone scan shows a significant focal increase in tracer uptake (Fig. 8-19C, D).

On rare occasions, a sclerotic metastatic lesion within the medial femoral condyle may mimic spontaneous osteonecrosis (37). A sclerotic vertebra ("ivory vertebra") resulting from metastasis should be differentiated from lymphoma, sclerosing hemangioma, and Paget disease (34,108,110). Involvement by a lymphoma is usually indistinguishable from metastatic disease, although the clinical and laboratory data may be helpful. In Hodgkin lymphoma there is an occasional anterior scalloping of the vertebral body, which accentuates the anterior vertebral concavity and thus provides a useful differentiating feature. Hemangioma often presents with typical vertical striations or a honeycomb pattern. Paget disease characteristically enlarges affected vertebrae and causes disappearance or coarsening of the vertebral endplates. If a picture frame appearance typical for Paget disease is present, metastasis can be safely ruled out (see Fig. 6.29). Conversely, in metastatic lesions to the vertebrae the endplates remain preserved (Fig. 8-20).



Figure 8-19 Condensing osteitis of the clavicle. A: Radiograph shows a sclerotic lesion in the inferior aspect of the right clavicle (*arrow*), originally thought to represent sclerotic metastasis. **B:** Trispiral tomography shows that the superior aspect of the clavicle is not affected. There is no evidence of periosteal reaction. **C:** Computed tomography section through the sternal ends of the clavicles shows homogeneous sclerosis of the right clavicular head and soft tissue swelling adjacent to it anteriorly. **D:** Radionuclide bone scan obtained after injection of 15 mCi (555 MBq) of 99m-methylene diphosphonate (MDP) shows increased uptake of the radiopharmaceutical tracer localized to the sternal end of the right clavicle. (Reprinted with permission from Greenspan A, Gerscovich EO, Szabo RM, et al. Condensing osteitis of the clavicle: a rare but frequently misdiagnosed condition. *Am J Roentgenol* 1991;156:1011–1013.)



Figure 8-20 Skeletal metastasis. Sclerotic metastasis to the lumbar vertebra of a 72-year-old man with prostatic carcinoma mimics Paget disease. Note, however, that the vertebral endplates are preserved and vertebral body is not enlarged.

Mixed Metastasis

A mixed (osteolytic and osteoblastic) solitary metastasis must be distinguished from a focus of *osteomyelitis*, *osteosarcoma*, and some primary round cell tumors, particularly *lymphoma*. Again, without additional clinical information it may be difficult to distinguish these lesions only on the basis of radiologic imaging.

MULTIPLE METASTASES

Lytic Metastases

Osteolytic metastases must be differentiated from multiple myeloma and brown tumors of hyperparathyroidism. In younger patients, Langerhans cell histiocytosis must be considered. Probably the best modality for distinguishing metastases from *multiple myeloma* is the radionuclide bone scan because, with only a few exceptions, metastatic lesions will exhibit an increased uptake of radiopharmaceutical tracer, whereas bone scan in multiple myeloma (except for the sclerotic variant) is usually normal. Helpful in distinguishing brown tumors of hyperparathyroidism are other hallmarks of this condition, such as diffuse osteopenia, loss of the lamina dura of the tooth sockets, subperiosteal bone resorption, and soft tissue calcifications. Laboratory data typical of hyperparathyroidism (particularly serum levels of calcium and phosphorus) will further enhance the difference.

Because of their expansive nature, multiple metastases from kidney and thyroid should be differentiated from *pseudotumors of hemophilia*. In such cases the clinical information and laboratory data characteristic of the latter condition are extremely helpful.

Sclerotic Metastases

Sclerotic metastases should be differentiated from osteopoikilosis (40). Osteopoikilosis is classified among the sclerosing dysplasias of endochondral failure of bone formation and remodeling (45). Sclerotic foci in osteopoikilosis are typically distributed near the large joints, such as hips, knees, and shoulders (Fig. 8-21). An additional helpful radiologic feature is the scintigraphic appearance of the lesion: osteopoikilosis, unlike sclerotic metastases, exhibits a normal radionuclide bone scan (45). Other entities that may be confused with sclerotic metastatic disease include polyostotic Paget disease (Fig. 8-22), mastocytosis, and secondary hyperparathyroidism, in which sclerotic bone changes may predominate. A rare form of sclerosing myeloma (representing <1% of cases of multiple myeloma) (107) and sclerosing hemangiomatosis (63) may have a radiographic appearance similar to that of sclerotic metastases. In this situation, the clinical and laboratory data typical for myeloma, along with a good general physical condition and absence of abnormal laboratory data in hemangiomatosis, are helpful. Erdheim-Chester disease, a rare form of histiocytosis, can radiographically mimic sclerotic metastases (Fig. 8-23). This condition usually exhibits bilateral, symmetric, patchy, or diffuse sclerosis of the medullary cavity of the long bones, sparing the epiphyses (120,123). The sclerotic lesions, similarly to metastases, show increased uptake of radiopharmaceutical agents on skeletal scintigraphy (71). On MRI the lesions exhibit low signal intensity on T1-weighted sequences, with heterogeneous signal on T2 weighting. After injection of gadolinium intense enhancement can be observed (126). Finally, osteosarcomatosis (synchronous or metachronous presentation of multifocal osteosarcoma) may also mimic metastatic disease (41). In such cases, a localization of the lesions in sites rarely affected by metastases, such as carpal and tarsal bones, metacarpals, epiphyses, and apophyses, as well as the radiographic morphology of the lesion consistent with osteosarcoma, leads to the correct diagnosis (see Fig. 2-95).



Figure 8-21 Osteopoikilosis. Anteroposterior radiograph of the right shoulder of a 34-year-old man shows typical periarticular distribution of sclerotic foci of osteopoikilosis.



Figure 8-22 Skeletal metastases. Diffuse sclerotic metastases to the pelvis and left femur causing a pathologic fracture in a 68-year-old man with prostate carcinoma mimic sclerotic changes of Paget disease.

CORTICAL METASTASES

A solitary cortical metastasis should be differentiated, among other possibilities, from *osteoid osteoma, cortical bone abscess, plasmacytoma, hemangioma,* and *cortical osteosarcoma.* Cortical involvement associated with a soft tissue mass must be differentiated from an *aneurysmal bone cyst* and a primary soft tissue tumor invading the bone, including *synovial sarcoma.* Multiple cortical metastases should be differentiated from *hemangiomatosis* and any vascular lesion involving the cortex. The patient's age and clinical history, as well as the radiologic morphology of the lesion, are useful in differ-



Figure 8-23 Erdheim-Chester disease resembling sclerotic metastases. Diffuse sclerosis of the radius may be mistaken for blastic metastases.

ential diagnosis. A metastatic origin should always be considered in differential diagnosis of an osteolytic cortical lesion of a long bone in older patients and in patients with a known primary malignant condition (20).

Pathology

Histologically, metastatic tumors are easier to diagnose than many primary tumors because of their essentially epithelial pattern. Biopsies of suspected metastases are useful for the diagnosis in patients with known primary tumors. With increasing use of ancillary techniques it is possible to identify the site of an unknown primary tumor (66). Occasionally, the only identifying feature in the marrow spaces is a desmoplastic fibrotic reaction to tumor. If a bone biopsy reveals predominantly mature bone trabeculae with intertrabecular fibrosis, the differential diagnosis of metastatic carcinoma should be considered. A metastatic lesion may exhibit a characteristic morphologic pattern that strongly suggests a primary tumor, such as the clear cells of renal carcinoma, follicular or giant cell carcinoma of the thyroid, or the pigment production of melanoma. Some neoplasms are associated with the production of enzymes or antigens that can be detected after the decalcification procedure. It is possible, for example, to perform a prostate-specific antigen study or a prostate-specific acid phosphatase by immunohistochemistry to identify a primary site in the prostate. Demonstration of nuclear transcription factors such as CDX2 (occurring in GI carcinoma) or TTF1 (occurring in lung and thyroid cancers); glycoproteins such as GCDFP15 (gross cystic disease fluid protein) present in breast or apocrine cells; pattern analysis of cytokeratin filaments (e.g., presence of CK 20 and absence of CK7 in intestinal but not in lung cancer) supplemented by additional immunoreactions, cytokeratins, CD20, CD99, and neuron-specific enolase (NSE) for the differentiation of small, round blue-cell tumors; and melanoma markers such as S-100, melan-A, and HMB-45 may also contribute to identification of an unknown primary tumor (22,66) (See Fig. 8-24C,D).

Osteoblastic metastases occasionally exhibit metastatic cells surrounded by rapidly proliferating osteoid, in a pattern that mimics *osteosarcoma* (Fig. 8-24A,B). However, careful study of the reactive bone will reveal the regular distribution of the osteocytes, even if the bone has been formed by metaplasia. The immunohistochemical marker for cytokeratin or for other epithelial markers can distinguish neoplastic epithelial cells from the reactive osteoblasts and from the tumor cells of epithelioid osteosarcoma (32).

When osseous metastases are composed of spindle cells, with or without associated epithelioid-like neoplastic cells, the diagnostic difficulties may be enormous. Differential diagnosis includes a number of primary osseous sarcomas, such as *fibrosarcoma*, *MFH*, and *leiomyosarcoma*. As a particular example, osseous metastases from spindle cell (sarcomatoid) renal cell carcinomas usually involve bone and may closely resemble MFH. Immuno-histochemistry is extremely useful for distinguishing

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Figure 8-24 Histopathology of skeletal metastases. A: Metastatic lesion from a well-differentiated papillary carcinoma of the thyroid exhibits formation of reactive bone that should not be mistaken for tumor bone of osteosarcoma (Giemsa, original magnification \times 50). **B:** Metaplastic formation of narrow woven bone trabeculae is seen in the stroma of a well-differentiated papillary thyroid carcinoma. The bone is reactive and not produced by malignant cells (van Gieson, original magnification \times 50). **C:** Metastasis from a melanoma resembles trabecular carcinoma; however, some tumor cells contain melanin pigment (hematoxylin and eosin, original magnification \times 200). **D:** Metastasis from melanoma resembles epithelioid tumor; however, tumor cells react strongly with anti-melanin A (biotin-avidin peroxidase, original magnification \times 200).

between these groups of lesions (32). However, some antigens are shared by both carcinomas and sarcomas. Vimentin, almost invariably present in sarcomas, may be synthesized by carcinoma cells and by those of malignant melanoma. Cytokeratin, present in most carcinomas, has been reported to occur in skeletal leiomyosarcoma (86) and other mesenchymal neoplasms (32,124). A useful immunohistochemical screening panel for the diagnosis of spindle cell lesions includes cytokeratin, epithelial membrane antigen, S-100, vimentin, desmin, and muscle-specific actin. Positive findings for epithelial membrane antigen or keratin are strongly suggestive of metastatic carcinoma. Diffuse positivity for S-100 protein suggests a diagnosis of malignant melanoma, when the light microscopic appearance is appropriate. This can be confirmed by staining with the melanocyte marker HMB-45 or melan-A (32).

If cytokeratin, epithelial membrane antigen, and S-100 markers are negative, the differential diagnosis must focus on primary sarcomas. In MFH, fibrosarcoma, and leiomyosarcoma, some cells are vimentin-positive. Furthermore, MFH and leiomyosarcoma may occasionally be positive for cytokeratin. However, the latter is strongly positive for muscle-specific actin, desmin, and smooth muscle actin (Table 8-1) (22,32,124).

The radiologic and pathologic differential diagnosis of solitary lytic metastasis is depicted in Figure 8-25 and of solitary blastic (sclerotic) metastasis in Figure 8-26, whereas the differential diagnoses of multiple osteolytic and multiple osteoblastic metastases are depicted in Figures 8-27 and 8-28, respectively. The radiologic and pathologic differential diagnosis of cortical metastases is shown in Figure 8-29.

Table 8-1 Panel of Immunohistochemical	Markers for Spindle Cell Neoplasms
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		Antigens						
Tumor	Desmin	Vim <mark>entin</mark>	СК	EMA	S-100	MSA	HMB-45	CD99
Spindle cell carcinoma Malignant melanoma Malignant fibrous histiocytoma Leiomyosarcoma Fibrosarcoma Synovial sarcoma	- -/+ + -	+/- + + +/- +	+ _/+ +/- _ +	+ - - - +	+/- + -/+ + -/+	_ _/+ + _	- + - -	- + - +

CK, cytokeratin; EMA, epithelial membrane antigen; MSA, muscle-specific actin; HMB-45, melanocytic marker; +/-, considerable number of positives; -/+, very few positives. Modified from Fechner RE, Mills SE. *Tumors of the bones and joints*, 3rd ed., Vol 8. Washington, DC: Armed Forces Institute of Pathology, 1993; and from Brooks JSJ. Immunohistochemistry in the differential diagnosis of soft-tissue tumors. In: Weiss SH, Brooks JSJ, eds. *Soft* tissue tumors. Baltimore: Williams and Wilkins, 1996:65.



Figure 8-25 Radiologic and pathologic differential diagnosis of solitary osteolytic metastasis.



Figure 8-26 Radiologic and pathologic differential diagnosis of solitary osteoblastic metastasis.



Figure 8-27 Radiologic and pathologic differential diagnosis of multiple osteolytic metastases.



Figure 8-28 Radiologic and pathologic differential diagnosis of multiple osteoblastic metastases.



Figure 8-29 Radiologic and pathologic differential diagnosis of cortical metastases.

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REFERENCES

- Abrams HL. Skeletal metastases in carcinoma. *Radiology* 1950;55:534–538.
- Abrams HL, Spiro R, Goldstein N. Metastases in carcinoma. Analysis of 1000 autopsied cases. *Cancer* 1950;3:74–85.
- Alazraki N. Radionuclide techniques. In: Resnick D, ed. Bone and joint imaging Philadelphia: WB Saunders, 1989:185–198.
- Algra PR, Bloem JL. Magnetic resonance imaging of metastatic disease and multiple myeloma. In: Bloem JL, Sartoris DJ, eds. *MRI and CT of the musculoskeletal system* Baltimore: Williams & Wilkins, 1992:218.
- Algra PR, Heimans JJ, Valk J, et al. Do metastases in vertebrae begin in the body or the pedicles? Imaging study in 45 patients. *Am J Roentgenol* 1992;158:1275–1279.
- Ardran GM. Bone destruction not demonstrable by radiography. Br J Radiol 1951;24:107–109.
- Asdourian PL, Weidenbaum M, Dewald RL, et al. The pattern of vertebral involvement in metastatic vertebral breast cancer. *Clin Orthop* 1991;250:164–170.
- Avrahami E, Tadmor R, Dally O, Hadar H. Early MR demonstration of spinal metastases in patients with normal radiographs and CT and radionuclide bone scans. *J Comput Assist Tomogr* 1989;13:598–602.
- Bachman AS, Sproul EE. Correlation of radiographic and autopsy findings in suspected metastases in the spine. *Bull NY Acad Med* 1940;44:169–175.
- Bassett LW, Steckel RJ. Imaging techniques in the detection of metastatic disease. *Semin Oncol* 1977;4:39–52.
- Berrettoni B, Carter JR. Mechanisms of cancer metastasis to bone. J Bone Joint Surg Am 1986;68:308–312.
- Bloom RA, Libson E, Husband JE, Stoker DJ. The periosteal sunburst reaction to bone metastases. A literature review and report of 20 additional cases. *Skeletal Radiol* 1987;16:629–634.
- Braunstein EM, Kuhns LR. Computed tomographic demonstration of spinal metastases. *Spine* 1983;8:912–915.
- Brown B, Laorr A, Greenspan A, Stadalnik R. Negative bone scintigraphy with diffuse osteoblastic breast carcinoma metastases. *Clin Nucl Med* 1994;19:194–196.
- Bushnell DL, Kahn D, Huston B, Bevering CG. Utility of SPECT imaging for determination of vertebral metastases in patients with known primary tumors. *Skeletal Radiol* 1995;24:13–16.
- Caluser CI, Scott AM, Schnieder J, et al. Value of lesion location and intensity of uptake in SPECT bone scintigraphy of the spine in patients with malignant tumors. *Radiology* 1992;185(S):315.
- Chirgwin JJM, Mohammad KKS, Guise TTA. Tumor-bone cellular interactions in skeletal metastases. J Musculoskel Neuronal Interact 2004;4:308–318.
- Choi J-A, Lee KH, Jun WS, et al. Osseous metastasis from renal cell carcinoma: "flow-void" sign at MR imaging. *Radiology* 2003; 228:629–634.
- Citrin DL, Bessent RG, Greig WR. A comparison of the sensitivity and accuracy of the 99m Tc-phosphate bone scan and skeletal radiograph in the diagnosis of bone metastases. *Clin Radiol* 1977;28:107–117.
- Coerkamp EG, Kroon HM. Cortical bone metastases. *Radiology* 1988;169:525–528.
- Cumming J, Hacking N, Fairhurst J, et al. Distribution of bony metastases in prostatic carcinoma. *Br J Urol* 1990;66:411–414.
- 22. Dabbs D. *Diagnostic immunohistochemistry*, 2nd ed. New York: Churchill-Livingstone, 2006:848.
- 23. Daldrup-Link HE, Franzius C, Link TM, et al. Whole-body MR imaging for detection of bone metastases in children and young adults: comparison with skeletal scintigraphy and FDG PET. *Am J Roentgenol* 2001;177:229–236.
- Daffner RA, Lupetin AR, Dash N, et al. MRI in the detection of malignancy infiltration of bone marrow. *Am J Roentgenol* 1986; 1146:353–358.
- Datz FL, Patch GG, Arias JM, Morton KA. Nuclear medicine. A teaching file St. Louis: Mosby Year Book, 1992:28–29.
- Delbeke D, Powers TA, Sandler MP. Negative scintigraphy with positive magnetic resonance imaging in bone metastases. *Skeletal Radiol* 1990;19:113–116.
- Deutsch A, Resnick D. Eccentric cortical metastases to the skeleton from bronchogenic carcinoma. *Radiology* 1980;137:49–52.

- Deutsch A, Resnick D, Niwayama G. Case report 145. Bilateral, almost symmetrical skeletal metastases (both femora) from bronchogenic carcinoma. *Skeletal Radiol* 1981;6:144–148.
- Eustace S, Tello R, DeCarvalho V, et al. A comparison of wholebody turboSTIR MR imaging and planar ^{99m}Tc-methylene diphosphonate scintigraphy in the examination of patients with suspected skeletal metastases. *Am J Roentgenol* 1997;169: 1655–1661.
- Even-Sapir E, Martin RH, Barnes DC, et al. Role of SPECT in differentiating malignant form benign lesions in the lower thoracic and lumbar vertebrae. *Radiology* 1993;187:193–198.
- Evison G, Pizey N, Roylance J. Bone formation associated with osseous metastases from bladder carcinoma. *Clin Radiol* 1981; 32:303–309.
- Fechner RE, Mills SE. *Tumors of the bones and joints*, 3rd series, fascicle 8. Washington, DC: Armed Forces Institute of Pathology, 1993.
- Fidler IJ. The pathogenesis of cancer metastasis: the 'seed and soil' hypothesis revisited. *Nat Rev Cancer* 2003;3:453–458.
- Fig LM, Gross MD. Metastatic prostate carcinoma mimicking Paget's disease on bone imaging. *Clin Nucl Med* 1989;14:777– 778.
- 35. Forbes G. Radiographic manifestations of bone metastases from renal carcinoma. *Am J Roentgenol* 1977;129:61–66.
- Foster D. Cortical skeletal metastasis in malignant melanoma. Australas Radiol 1982;20:545–560.
- 37. Fujimoto H, Nishimura G, Motoori K, et al. Metastatic bone tumor mimicking spontaneous osteonecrosis of the medial condyle of the femur: misleading appearance on MR imaging. *Skeletal Radiol* 2000;29:286–288.
- Galasko CSB. Mechanisms of lytic and blastic metastatic disease of bone. *Clin Orthop* 1982;69:20–27.
- Galasko CSB. The anatomy and pathways of skeletal metastases. In: Weiss L, Gilbert H, eds. *Bone metastasis* Boston: GK Hall, 1981:49–63.
- Ghandur-Mnaymneh L, Broder LE, Mnaymneh WA. Lobular carcinoma of the breast metastatic to bone with unusual clinical, radiologic, and pathologic features mimicking osteopoikilosis. *Cancer* 1984;53:1801–1803.
- 41. Gherlinzoni F, Antoci B, Canale V. Multicentric osteosarcomata (osteosarcomatosis). *Skeletal Radiol* 1983;10:281–285.
- Godersky JC, Smoker WR, Knutzon R. Use of magnetic resonance imaging in the evaluation of metastatic spinal disease. *Neurosurgery* 1987;21:676–680.
- Gold RI, Seeger LL, Bassett LW, Steckel RJ. An integrated approach to the evaluation of metastatic bone disease. *Radiol Clin North Am* 1990;28:471–483.
- Gosfield E, Alavi A, Kneeland B. Comparison of radionuclide bone scan and magnetic resonance imaging in detecting spinal metastases. *J Nucl Med.* 1994;34:2191–2198.
- Greenspan A. Sclerosing bone dysplasias—a target-site approach. Skeletal Radiol 1991;20:561–584.
- Greenspan A, Gerscovich EO, Szabo RM, Matthews II JG. Condensing osteitis of the clavicle: a rare but frequently misdiagnosed condition. *Am J Roentgenol* 1991;156:1011–1015.
- Greenspan A, Klein MJ. Radiology and pathology of bone tumors. In: Lewis MM, ed. *Musculoskeletal oncology: a multidisciplinary approach* Philadelphia: WB Saunders, 1992:13–72.
- Greenspan A, Klein MJ, Lewis MM. Case report 272. Skeletal cortical metastases in the left femur arising form bronchogenic carcinoma. *Skeletal Radiol* 1984;11:297–301.
- Greenspan A, Klein MJ, Lewis MM. Case report 284. Osteolytic cortical metastasis in the femur from bronchogenic carcinoma. *Skeletal Radiol* 1984;12:146–150.
- Greenspan A, Norman A. Osteolytic cortical destruction: an unusual pattern of skeletal metastases. *Skeletal Radiol* 1988;17: 402–406.
- Greenspan A, Stadalnik RC. Bone island: scintigraphic findings and their clinical application. *Can Assoc Radiol J* 1995;46: 368–379.
- 52. Greenspan A, Steiner G, Knutzon R. Bone island (enostosis): clinical significance and radiologic and histologic correlations. *Skeletal Radiol* 1991;20:85–90.
- Greenspan A, Klein MJ. Giant bone island. Skeletal Radiol 1996; 25:67–69.

- Hauschka PV, Mavrakos AE, Iafrati MD, et al. Growth factors in bone matrix. Isolation of multiple types by affinity chromatography on heparin-Sepharose. *J Biol Chem* 1986;261:12665– 12674.
- Healey JH, Turnbull ADM, Miedema B, Lane JM. Acrometastases. A study of twenty-nine patients with osseous involvement of hand and feet. *J Bone Joint Surg* 1986;68:743–746.
- Hellman RS, Wilson MA. Discordance of sclerosing skeletal secondaries between sequential scintigraphy and radiographs. *Clin Nucl Med* 1982;7:97–99.
- Helms CA, Cann CE, Brunelle FO, et al. Detection of bone-marrow metastases using quantitative computed tomography. *Radiology* 1981;140:745–750.
- Hendrix RW, Rogers LF, Davis TM Jr. Cortical bone metastases. Radiology 1991;181:409–413.
- Hove B, Gyldensted C. Spiculated vertebral metastases from prostatic carcinoma. *Neuroradiology* 1990;32:337–339.
- Hudson TM. Radiologic-pathologic correlation of musculoskeletal lesions Baltimore: Williams & Wilkins, 1987:421–440.
- 62. Igou D, Sundaram M, McDonald DJ, et al. Appendicular metastatic prostate cancer simulating osteosarcoma, Paget's disease, and Paget's sarcoma. *Skeletal Radiol* 1995;24:447–449.
- Ishida T, Dorfman HD, Steiner GC, Normatz A. Cystic angiomatosis of bone with sclerotic changes mimicking osteoblastic metastases. *Skeletal Radiol* 1994;23:247–252.
- Jacobson HG, Poppel MH, Shapiro JH, Grossberger S. The vertebral pedicle sign: a Roentgen finding to differentiate metastatic carcinoma from multiple myeloma. *Am J Roentgenol* 1958;80:817–821.
- Jaffe H. Tumors metastatic to the skeleton. In: *Tumors and tumorous conditions of the bones and joints* Philadelphia: Lea & Febiger, 1958:594–595.
- 66. Jambhekar NA, Borges A. Metastases involving bone. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone Lyon: IARC Press, 2002:334–335.
- Johnston AD. Pathology of metastatic tumors in bone. *Clin Orthop* 1970;73:8–32.
- Kattapuram SV, Khurana JS, Scott JA, el-Khoury GY. Negative scintigraphy with positive magnetic resonance imaging in bone metastases. *Skeletal Radiol* 1990;19:113–116.
- Kido DK, Gould R, Taati F, et al. Comparative sensitivity of CT scans, radiographs, and radionuclide bone scans in detecting metastatic calvarial lesions. *Radiology* 1978;128:371–375.
- Kim EE, Deland FH, Maruyama Y. Decreased uptake in bone scans ("cold lesions") in metastatic carcinoma. J Bone Joint Surg Am 1978;60:844–846.
- Klieger MR, Schultz E, Elkowitz DA, et al. Erdheim-Chester disease: a unique presentation with multiple osteolytic lesions of the spine and pelvis that spared the appendicular skeleton. *Am J Roentgenol* 2002;178:429–432.
- Kori SH. Computed tomographic evaluation of bone and soft tissue metastases. In: Weiss L, Gilbert H, eds. *Bone metastasis* Boston: GK Hall, 1981:245–257.
- Kumar N, David R, Madewell JE, Lindell MM Jr. Radiographic spectrum of osteogenic sarcoma. Am J Roentgenol 1987;148: 767–772.
- Legier JF, Tauber LN. Solitary metastases of occult prostatic carcinoma simulating osteogenic sarcoma. *Cancer* 1968;22:168– 172.
- Lehrer HZ, Maxfield WS, Nice CM. The periosteal sunburst pattern in metastatic bone tumors. Am J Roentgenol 1970;108: 154–161.
- Libson E, Bloom RA, Husband JE, Stocker DJ. Metastatic tumors of the bones of the hand and foot. A comparative review and report of 43 additional cases. *Skeletal Radiol* 1987;16:387–392.
- Ludwig H, Kumpan W, Sinzinger H. Radiography and bone scintigraphy in multiple myeloma: a comparative analysis. Br J Radiol 1982;55:173–181.
- Mall JC, Beckerman C, Hoffer PB, Gottschalk H. A unified radiological approach to the detection of skeletal metastases. *Radiology* 1976;118:323–328.

- McDougall IR, Kriss JP. Screening for bone metastases. Are only scans necessary? *JAMA* 1975;231:46–50.
- 80. Muindi J, Coombes RC, Golding S, et al. The role of computed tomography in the detection of bone metastases in breast cancer patients. *Br J Radiol* 1983;56:233–236.
- Mulder JD, Schütte HE, Kroon HM, Taconis WK. Radiologic atlas of bone tumors Amsterdam: Elsevier, 1993.
- 82. Mulvey RB. Peripheral bone metastases. Am J Roentgenol 1964;91:155–160.
- Mundy GR. Metastasis to bone: causes, consequences and therapeutic opportunities. *Nat Rev Cancer* 2002;2:584–593.
- 84. Mundy GR. Endothelin-1 and osteoblastic metastasis. *Proc Natl Acad Sci U S A* 2003;100:10588–105989.
- 85. Murray RO, Jacobson HG. *The radiology of skeletal disorders*, 2nd ed. New York: Churchill-Livingstone, 1977:585.
- Myers JL, Arocho J, Bernreuter W, et al. Leiomyosarcoma of bone. A clinicopathologic, immunohistochemical, and ultrastructural study of five cases. *Cancer* 1991;67:1051–1056.
- Napoli LD, Hansen HH, Muggia FM, et al. The incidence of osseous involvement in lung cancer, with special reference to the development of osteoblastic changes. *Radiology* 1973;108:17–21.
- Norman A, Greenspan A, Steiner G. Case report 173. Metastases from a bronchial carcinoid tumor. *Skeletal Radiol* 1981;7: 155–157.
- 89. Norman A, Ulin R. A comparative study of periosteal new-bone response in metastatic bone tumors (solitary) and primary bone sarcomas. *Radiology* 1969;92:705–708.
- Nystrom JS, Weiner JM, Wolf RM, et al. Identifying the primary site in metastatic cancer of unknown origin. *JAMA* 1979;241: 381–383.
- O'Mara RE. Bone scanning in osseous metastatic disease. JAMA 1974;229:1915–1917.
- Onitsuka H. Roentgenologic aspects of bone islands. *Radiology* 1977;123:607–612.
- Ontell FK, Greenspan A. Blastic osseous metastases in ovarian carcinoma. Can Assoc Radiol J 1995;46:231–234.
- Osmond TD III, Pendergrass HP, Potsaid MS. Accuracy of 99M Tc-diphosphonate bone scans and roentgenograms in the detection of prostate, breast, and lung carcinoma metastases. *Am J Roentgenol* 1975;124:972–975.
- Pagani JJ, Libshitz HI. Imaging bone metastases. Radiol Clin North Am 1982;20:545–560.
- Paget S. The distribution of secondary growths in cancer of the breast. *Lancet* 1889;133:571–573.
- Panebianco AC, Kaupp HA. Bilateral thumb metastasis from breast carcinoma. Arch Surg 1968;96:216–218.
- Papac RJ. Bone marrow metastases: a review. Cancer 1994;74: 2403–2413.
- Parthasarathy KL, Landsberg R, Bakshi SP, et al. Detection of bone metastases in urogenital malignancies utilizing 99m Tc-labeled phosphate compounds. *Urology* 1978;11:99–102.
- Peavy PW, Rogers JV Jr, Clements JL Jr, Burns JB. Unusual osteoblastic metastases from carcinoid tumors. *Radiology* 1973;107: 327–330.
- 101. Powell JM. Metastatic carcinoid of bone. Report of two cases and review of the literature. *Clin Orthop* 1988;230:266–272.
- Rafii M, Firooznia H, Golimbu C, Beranbaum E. CT of skeletal metastasis. Semin Ultrasound Comput Tomogr Magn Reson Imag 1986;7:371–379.
- Rafii M, Firooznia H, Kramer E, et al. The role of computed tomography in evaluation of skeletal metastases. *J Comput Tomogr* 1988;12:19–24.
- Ramaswamy S, Ross KN, Lander ES, Golub TR. A molecular signature of metastasis in primary solid tumors. *Nat Genet* 2003;33: 49–54.
- 105. Resnick D, Niwayama G. Skeletal metastases. In: Resnick D, ed. Diagnosis of bone and joint disorders, 3rd ed. Philadelphia: WB Saunders, 1995:3991–4064.
- Roodman GD. Mechanisms of bone metastasis. N Engl J Med 2004;350:1655–1664.
- 107. Sartoris DJ, Pate D, Haghighi P, et al. Plasma cell sclerosis of bone: a spectrum of disease. *Can Assoc Radiol J* 1986;37:25–34.
- Schajowicz F, Velan O, Santini Araujo E, Plantalech L, et al. Metastases of carcinoma in pagetic bone. *Clin Orthop* 1988;228: 290–296.

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- Schweitzer ME, Levine C, Mitchell DG, et al. Bull's-eyes and halos: useful MR discriminators of osseous metastases. *Radiology* 1993;188:249–252.
- Shih WJ, Riley C, Magoun S, Ryo UY. Paget's disease mimicking skeletal metastases in a patient with coexisting prostatic carcinoma. *Eur J Nucl Med* 1988;15:422–423.
- 111. Sim FH, Frassica FJ. Metastatic bone disease. In: Unni KK, ed. *Bone tumors* New York: Churchill-Livingstone, 1988:226.
- 112. Simon MA, Bartucci EJ. The search for the primary tumor in patients with skeletal metastases of unknown origin. *Cancer* 1986;58:1088–1095.
- 113. Söderlund V. Radiological diagnosis of skeletal metastases. *Eur Radiol* 1996;6:587–595.
- Spjut HJ, Dorfman HD, Fechner RE, Ackerman LV. Tumors of bone and cartilage, 2nd series, fascicle 5. Washington, DC: Armed Forces Institute of Pathology, 1971:347–390.
- 115. Steeg PS. Cancer biology: Emissaries set up new sites. *Nature* 2005;438:750-751.
- Thrall JH, Ellis BI. Skeletal metastases. Radiol Clin North Am 1987;25:1155–1170.
- 117. Traill Z, Richards MA, Moore NR. Magnetic resonance imaging of metastatic bone disease. *Clin Orthop Relat Res* 1995;312:76–88.
- 118. Trias A, Fery A. Cortical circulation of long bones. J Bone Joint Surg Am 1979;61:1052–1059.
- 119. Trillet V, Revel D, Combaret V. Bone marrow metastases in small cell lung cancer: detection with magnetic resonance imaging and monoclonal antibodies. *Br J Cancer* 1989;60:83–88.

- Veyssier-Belot C, Cacoub P, Caparros-Lefebvre D, et al. Erdheim-Chester disease: clinical and radiologic characteristics of 59 cases. *Medicine* 1996;75:157–169.
- Vieco PT, Azouz EM, Hoeffel J-C. Metastases to bone in medulloblastoma. A report of five cases. *Skeletal Radiol* 1989;18: 445–449.
- Vilar JL, Lezena AH, Pedrosa CS. Spiculated periosteal reaction in metastatic lesions in bone. *Skeletal Radiol* 1979;3:230–233.
- 123. Vinh TN, Sweet DE. Erdheim-Chester disease. In: Fletcher CDM, Unni KK, Mertens F, eds. World health organization classification of tumours. Pathology & genetics. Tumours of soft tissues and bone Lyon: IARC Press, 2002:347.
- 124. Weiss SW, Goldblum JR. Enzinger and Weiss's soft tissue tumors, 4th ed. Philadelphia: CV Mosby, 2001:1632.
- 125. Wilner D. Cancer metastasis to bone. In: Radiology of bone tumors and allied disorders Philadelphia: WB Saunders, 1982:3641–3908.
- Wright RA, Hermann RC, Parisi JE. Neurological manifestations of Erdheim-Chester disease. J Neurol Neurosurg Psychiatry 1999;66:72–75.
- 127. Yamaguchi T, Tamai K, Yamato M, et al. Intertrabecular pattern of tumors metastatic to bone. *Cancer* 1996;78:1388–1394.
- 128. Yochum TR, Rowe LJ. Tumor and tumor-like processes. In: Yochum TR, Rowe LJ, eds. *Essentials of skeletal radiology*, vol 2. Baltimore: Williams & Wilkins, 1987:699–919.
- 129. Yoneda T, Hiraga T. Crosstalk between cells and bone microenvironment in bone metastasis. *Biochem Biophys Res Commun* 2005;328:679–687.

Tumors and Tumor-like Lesions of the Joints

BENIGN LESIONS 481

Synovial (Osteo) chondromatosis 481 Pigmented Villonodular Synovitis 487 Localized Pigmented Nodular Tenosynovitis [Pigmented Giant Cell Tumor of the Synovium or the (Localized) Giant Cell Tumor of the Tendon Sheath] 494 Synovial Hemangioma 496 Lipoma Arborescens 500

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Benign Lesions

Synovial (Osteo)chondromatosis

Synovial osteochondromatosis (also known as synovial chondromatosis or synovial chondrometaplasia) is an uncommon benign disorder marked by the metaplastic proliferation of multiple cartilaginous nodules in the synovial membrane of the joints, bursae, or tendon sheaths (26,27,30,42). It is almost invariably monoarticular; rarely, multiple joints may be affected (40,43,44).

Clinical Presentation

The disorder is twice as common in men as in women and is usually discovered in the third to the fifth decade (27). The knee is a preferential site of involvement, with the hip, shoulder, elbow, and ankle accounting for most of the remaining cases (6,7,12,23,39) (Fig. 9-1). Patients usually report pain and swelling. Joint effusion, tenderness, limited motion in the joint, and a soft tissue mass are common clinical findings (11).

Imaging

The radiographic findings depend on the degree of calcification within the cartilaginous bodies, ranging from mere joint effusion to visualization of many radiopaque joint bodies, usually small and uniform in size (31,33) (Fig. 9-2). The best proof that the bodies are indeed intraarticular is achieved by arthrography or computed tomography (CT) (3) (Figs. 9-3 and 9-4). These modalities can visualize even noncalcified bodies (13). MRI may also be helpful: calcifications can be seen as signal void on T2-weighted images against the high-signal-intensity fluid and an inflamed hyperplastic synovium (22,24,25,44,45) (Figs. 9-5 and 9-6). In addition to revealing loose bodies in the joint, these modalities may demonstrate bone erosion (34) and extracapsular extension of the lesion (38).

Histopathology

By microscopy, many cartilaginous nodules are observed as they form beneath the thin layer of cells that line the surface of the synovial membrane (12,36) (Fig. 9-7). These nodules are highly cellular, and the cells *Text continues on page 486*



Figure 9-1 Synovial (osteo)chondromatosis: skeletal sites of predilection, peak age range, and male-to-female ratio.





Figure 9-2 Synovial (osteo)chondromatosis. Anteroposterior **(A)** and lateral **(B)** radiographs of the right elbow of a 23-year-old man who presented with pain and occasional locking in the elbow joint show numerous osteochondral bodies in the joint (*arrows*), which are regularly shaped and mostly uniform in size.



Figure 9-3 Synovial (osteo)chondromatosis: computed tomography (CT). A: Anteroposterior radiograph of the right hip of a 27-year-old woman shows multiple osteochondral bodies around the femoral head and neck. Note preservation of the joint space. B and C: Two CT sections one through the femoral head, another through the femoral neck—demonstrate unquestionably the intraarticular location of multiple osteochondral bodies.





С



Figure 9-4 Synovial (osteo)chondromatosis: computed tomography (CT). A: Anteroposterior radiograph of the left shoulder of a 36-year-old man shows multiple osteochondral bodies around the glenohumeral joint. B: Coronal CT section effectively shows location of the uniform in size calcified bodies in the glenohumeral joint and subacromial bursa (*continued*).



Figure 9-4 *Continued* **C:** Axial CT section also clearly shows location of the calcified bodies in the glenohumeral joint and subacromial bursa.





Figure 9-6 Synovial (osteo)chondromatosis: magnetic resonance imaging (MRI). A: Lateral radiograph of the left knee of a 50-year-old man shows multiple osteochondral bodies in and around the joint. **B:** Axial T2*-weighted (MPGR, TR 500, TE 20, flip angle 30 degrees) MRI demonstrates high signal joint effusion and multiple bodies of intermediate signal intensity, primarily located in a large popliteal cyst. Coronal fast-spin echo (FSE, TR 2400, TE 85 Ef) **(C)** and sagittal fast-spin echo (FSE, TR 3400, TE 85 Ef) **(D)** MR images show to better advantage the distribution of numerous osteochondral bodies.



Figure 9-7 Histopathology of synovial (osteo)chondromatosis. Photomicrograph of the synovium removed from the knee of a patient with primary synovial chondromatosis shows nodules of irregular cellular cartilage covered by a thin layer of synovium (hematoxylin and eosin, original magnification \times 6).

themselves may exhibit a moderate pleomorphism, with occasional plump and double nuclei (11). Although such cytologic atypia merely indicates an actively growing cartilaginous focus, an erroneous diagnosis of malignancy may sometimes be considered (Fig. 9-8). The cartilaginous nodules, which often are undergoing calcification and endochondral ossification, may detach and become loose bodies. The loose bodies continue to be viable and may increase in size as they receive nourishment from the synovial fluid (18), but they ossify no more.

Mitoses in synovial chondromatosis are rare. However, subpopulations of cells appear to proliferate, according to immunohistochemistry (IHC) results and those of studies using image cytometry (8,32). Robinson et al. (37) also demonstrated that proliferating cells in cartilaginous nodules of synovial chondromatosis simultaneously express *fibroblast growth factor receptor*



Figure 9-8 Histopathology of synovial (osteo)chondromatosis. High-power photomicrograph shows pleomorphic cells with dark pleomorphic nuclei arranged in clusters (hematoxylin and eosin, original magnification $\times 100$).

3 (FGFR3) and its ligand, fibroblast growth factor 9 (FGF9), found in the synovial fluid, obviously produced by synoviocytes. This would constitute an autocrine loop that might be expected in tumor cells but not in a benign lesion. In addition, cytogenetic studies revealed clonal chromosomal aberrations with involvement of chromosome 6 (losses or rearrangements) in five of eight thus far reported cases, indicating that synovial chondromatosis may be a neoplastic rather than a metaplastic lesion (4,41). The expression of FGFR3 is regulated by sonic hedgehog (SHH), an important gene involved in embryonic development and in tumorigenesis (1,29). A negative regulator of the SHH gene, Rab23, is localized on chromosome 6p11 (10). Buddingh et al. (4) hypothesized that rearrangements of chromosome 6 may inactivate Rab23, leading to activation of SHH that, in turn, acts on FGFR3 expression. The involvement of SHH signaling in the development of synovial chondromatosis was recently demonstrated by Hopyan et al. (19) using transgenic mice defective in a transcriptional repressor for SHH.

Differential Diagnosis

Radiology

This condition should be differentiated from the *sec-ondary osteochondromatosis* caused by osteoarthritis, particularly in the knee and hip joints, and from *synovial chondrosarcoma*, either primary (arising *de novo* from the synovial membrane) or secondary (due to malignant transformation).

Distinguishing primary synovial osteochondromatosis from *secondary osteochondromatosis* usually presents no problems. In the latter condition, there is invariably radiographic evidence of osteoarthritis with all of its typical features, such as narrowing of the radiographic joint space, subchondral sclerosis, and, occasionally, periarticular cysts or cyst-like lesions (Fig. 9-9). The loose bodies are fewer, larger, and invariably of different sizes. Conversely, in primary synovial chondromatosis the joint is not affected by any degenerative changes. In some cases, however, the bone may show erosions secondary to pressure of the calcified bodies on the outer aspects of the cortex. The intraarticular bodies are numerous, small, and usually of uniform size (see Fig. 9-2).

More difficult is to distinguish synovial chondromatosis from *synovial chondrosarcoma*. The clinical and radiographic features have not been useful in this differentiation and are equally ineffective in distinguishing a secondary malignant lesion arising in synovial chondromatosis (2). In addition, both entities tend to have a protracted clinical course, and local recurrence is common after synovectomy for synovial chondromatosis or local resection of synovial chondrosarcoma (17). The presence of frank bone destruction rather than merely erosions, and the association of a soft tissue mass, should always raise the concern about malignancy (14) (see Fig. 9-41). Although extension beyond the joint capsule should heighten the suspicion of malignancy, some cases of synovial chondromatosis have



Figure 9-9 Secondary (osteo)chondromatosis due to osteoarthritis. A: Lateral radiograph of the knee in a 58-year-old man with advanced osteoarthritis of the femoropatellar joint compartment shows multiple osteochondral bodies in the suprapatellar bursa and within the popliteal cyst. **B:** Radiograph of the left shoulder in a 68-year-old woman with osteoarthritis of the glenohumeral joint shows multiple intraarticular osteochondral bodies.

been reported to have an extraarticular extension (5,31).

The other conditions that can radiologically mimic synovial chondromatosis include *pigmented villonodular* synovitis (PVNS), synovial hemangioma, and lipoma arborescens (15,16).

In *PVNS* the filling defects in the joint are more confluent and less distinct. Magnetic resonance imaging (MRI) may show foci of decreased intensity of the synovium in all sequences because of the paramagnetic effects of deposition of hemosiderin (18) (see Fig. 9-14). *Synovial hemangioma* usually presents as a single soft tissue mass. On MRI, T1-weighted images show that the lesion is either isointense or slightly higher (brighter) in signal intensity than surrounding muscles but much lower in intensity than subcutaneous fat. On T2weighted images the mass is invariably much brighter than fat (15) (see Figs. 9-26 and 9-27). Phleboliths and fibrofatty septa in the mass are common findings that show low signal characteristics.

Lipoma arborescens is a villous lipomatous proliferation of the synovial membrane (35). This rare condition usually affects the knee joint but has occasionally been reported in other joints, including the wrist and ankle (16). The disease has been variously reported to have a developmental, traumatic, inflammatory, or neoplastic origin, but its true etiology is still unknown. The clinical findings include slowly increasing but painless synovial thickening as well as joint effusion with sporadic exacerbation. Radiographic studies reveal a joint effusion accompanied by various degrees of osteoarthritis. MRI is diagnostic, showing frond-like masses arising from the synovium that have the signal intensity of fat on all imaging sequences (see Fig. 9-30). Histologic examination demonstrates complete replacement of the subsynovial tissue by mature fat cells and the formation of proliferative villous projections.

Pathology

From the histopathologic standpoint it may be very difficult to distinguish *synovial chondrosarcoma* from synovial chondromatosis (20,21,28). The histopathologic distinction between primary and secondary malignant synovial lesions and synovial chondromatosis has been a matter of dispute. Occasional foci of increased cellularity showing hyperchromatic atypical cells, consistent with grade 1 chondrosarcoma, should not be considered sufficient evidence for a malignant change in synovial chondromatosis. However, evidence of aggressive growth (invasion), arrangement of atypical chondrocytes in sheets, spindling of cells in the periphery, presence of mitoses, myxoid change of the matrix, and lack of attachment of a lesion to the synovial lining support the diagnosis of malignancy (2,9) (see Fig. 9-43).

The radiologic and pathologic differential diagnosis of synovial chondromatosis is depicted in Figure 9-10.

Pigmented Villonodular Synovitis

PVNS is a locally destructive fibrohistiocytic proliferation, characterized by many villous and nodular synovial protrusions (11), which affects joints, bursae, and tendon sheaths (46,77,109). PVNS was first described by Jaffe, Lichtenstein, and Sutro (94) in 1941, who used



Figure 9-10 Radiologic and pathologic differential diagnosis of synovial (osteo)chondromatosis.

this name to identify the lesion because of its vellowbrown, villous, and nodular appearance. The yellowbrown pigmentation is due to excessive deposits of lipid and hemosiderin. This condition can be diffuse or localized. When the entire synovium of the joint is affected and there is a major villous component, the condition is referred to as diffuse PVNS. When a discrete intraarticular mass is present, the condition is called localized PVNS (84,88,110) or (in the European literature) pigmented giant cell tumor of articulations. When the process affects the tendon sheaths, it is called localized giant cell tumor of the tendon sheaths or nodular tenosynovitis (69,98,110,111,127,143). The diffuse form usually occurs in the knee, hip, elbow, or wrist and accounts for 23% of cases (103). The localized nodular form is often regarded as a separate entity. It consists of a single polypoid mass attached to the synovium. Nodular tenosynovitis is most often seen in the fingers and is the second most common soft tissue tumor of the hand, exceeded only by the ganglion (133). In the new revised classification of soft tissue tumors (2002), the World Health Organization (WHO) classifies localized intra- and extraarticular lesion as giant cell tumor of sheath (72), whereas diffuse intra- and extraarticular forms are categorized as diffuse-type giant cell tumor (keeping PVNS as a synonym) (71).

Both the diffuse and the localized forms of villonodular synovitis usually occur as a single lesion, mainly in young and middle-aged individuals of either sex (67,119). One of the most characteristic findings in PVNS is the ability of the hyperplastic synovium to invade the subchondral bone, producing cysts and erosions. Although the etiology is unknown (78) and is often controversial (86,87), some investigators have suggested an autoimmune pathogenesis (57). Trauma also is a suspected cause, because similar effects have been produced experimentally in animals by repeated injections of blood into the knee joint (133,148). Some investigators have suggested a disturbance in lipid metabolism as an etiologic factor (82,91). It has also been postulated by Jaffe et al. (94) that these lesions may represent an inflammatory response to an unknown agent, and by Stout and Lattes (138), and later by Ray et al. (128) that they are true benign neoplasms. Although the latter theory was supported by pathologic studies indicating that the histiocytes present in PVNS may function as facultative fibroblasts (116) and that foam cells may derive from histiocytes (73), relating PVNS to a benign neoplasm of fibrohistiocytic origin (86), it was presumed that these findings did not constitute definite proof that PVNS is a true neoplasm but rather a special form of a chronic proliferative inflammation process, as has already been postulated by Jaffe et al. (94). However, the most recent cytogenetic investigations have shed a new light on this subject. Clonal aberrations have been described in both localized and diffuse forms of PVNS, presenting a near- or pseudodiploid karyotype and rearrangements of chromosome 1p, preferentially at 1p11-p13 (122,132). Aneuploid forms (in 3 of 7 diffuse-type tenosynovial giant cell tumors of tendon sheaths and 1 of 15 in diffuse type tenosynovial giant cell tumor of the knee joint) have

been described in DNA flow cytometric studies (47,54). In addition, Berger et al. (54) demonstrated chromosomal gains in 5 of 15 cases by CGH, preferentially at 22q, 16p, and 16q. These data are in favor of a neoplastic origin of PVNS. Gains of chromosomes 5 and 7 have been reported only in diffuse forms of tendon sheath giant cell tumors, indicating possible differences between localized and diffuse types (71,117). However, because at least gains of chromosomes 5 and 7 have been reported in osteoarthritis and rheumatoid arthritis, the significance of these findings remains to be determined (68,114).

Clinical Presentation

Clinically, PVNS is a slowly progressive process that manifests as mild pain and joint swelling with limitation of motion (48,78,137,144,149). Occasionally, increased skin temperature is noted over the affected joint (73). The knee joint is most commonly affected, and 66% of patients present with a bloody joint effusion (97). In fact, the presence of a serosanguinous synovial fluid in the absence of a history of recent trauma should strongly suggest the diagnosis of PVNS (86). The synovial fluid contains elevated levels of cholesterol (59,121), and fluid reaccumulates rapidly after aspiration (89). Other joints may be affected, including hip, ankle, wrist, elbow, and shoulder the (49,50,58,66,124,131). There is a 2:1 predilection for females. Patients range from 4 to 60 years of age, with a peak incidence in the third and fourth decades (11,113) (Fig. 9-11). The duration of symptoms can range from 6 months to as long as 25 years (51, 70, 105, 106).

Although a few "malignant" PVNS have been reported in the literature (55,99), this diagnosis is still debatable. Recently, attention has been drawn to the extraarticular form of diffuse PVNS, also referred to as diffuse-type giant cell tumor. This condition is characterized by the presence of an infiltrative, extraarticular mass with or without involvement of the adjacent joint (100). This presentation of PVNS creates a real diagnostic challenge for both radiologist and pathologist because its extraarticular location, invasion of the bones, and more varied histologic infiltrative pattern may suggest malignancy (100,134).

Imaging

Radiography reveals a soft tissue density in the affected joint, frequently interpreted as joint effusion. However, the density is greater than that of a simple effusion, and it reflects not only a hemorrhagic fluid but also lobulated synovial masses (Fig. 9-12). A marginal, welldefined erosion of subchondral bone with a sclerotic margin may be present [incidence reported from 15% (61) to 50% (74)], usually on both sides of the affected articulation (75,150). Narrowing of the joint space has also been reported (86). In the hip, multiple cyst-like or erosive areas involving non-weight-bearing regions of the acetabulum, as well as the femoral head and neck, are characteristic (65). Calcifications are encountered only in exceptional cases (52).



Figure 9-11 Pigmented villonodular synovitis: skeletal sites of predilection, peak age range, and male-to-female ratio.

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Figure 9-12 Pigmented villonodular synovitis. Lateral radiograph of the knee of a 58-year-old man shows a large suprapatellar joint effusion and a dense, lumpy soft tissue mass eroding the posterior aspect of the lateral femoral condyle (*arrow*). Note that posteriorly the density is greater than that of a suprapatellar fluid.

Arthrography reveals multiple lobulated masses with villous projections, which appear as filling defects in the contrast-filled suprapatellar bursa (Fig. 9-13). CT effectively demonstrates the extent of the disease (60,102). The increase in iron content of the synovial fluid results in high Hounsfield values, a feature that can help in the differential diagnosis (129). MRI is extremely useful in making a diagnosis (53,56,107,126, 135,136,139,141,146) because on T2-weighted images the intraarticular masses demonstrate a combination of high-signal-intensity areas, representing fluid and congested synovium, interspersed with areas of intermediate to low signal intensity, secondary to random distribution of hemosiderin in the synovium (Fig. 9-14) (76,96). In general, MRI shows a low signal on both T1and T2-weighted images because of hemosiderin deposition and thick fibrous tissue (24,120) (Fig. 9-15). In addition, within the mass, signals consistent with fat can be noted, which are due to clumps of lipid-laden macrophages (93). Other MRI findings include hyperplastic synovium and occasionally bone erosions. Administration of gadolinium in the form of Gd-DTPA





Figure 9-13 Pigmented villonodular synovitis: arthrography. A: Lateral radiograph of the knee of a 25-year-old woman shows what appears to be a suprapatellar effusion (*arrows*). The density of the fluid, however, is increased, and there is some lobulation evident. B: Contrast arthrogram of the knee shows lobulated filling defects in the suprapatellar pouch, representing lumpy synovial masses. Joint aspiration yielded thick bloody fluid, which explains the increased density of the soft tissue mass seen on radiography.



leads to a notable increase in overall heterogeneity, which tends toward an overall increase in signal intensity of the capsule and septae (93). This enhancement of the synovium allows it to be differentiated from the fluid invariably present, which does not enhance. Apart from its diagnostic effectiveness, MRI also is useful in defining the extent of the disease (92,112).

Histopathology

On histologic examination, PVNS reveals a tumor-like proliferation of the synovial tissue. A dense infiltration of mononuclear histiocytes is observed, accompanied by plasma cells, lymphocytes, and variable numbers of resorptive giant cells (Fig. 9-16A, B). The villi are made up of a matrix that contains collagen and reticulin fibers, with a varied cell population. The villi are also covered by hyperplastic and reactive-appearing synovial cells that may contain a large amount of hemosiderin (11). The synovial layer merges gradually with the underlying cell infiltrate that occupies the central core of the nodules (11). The stroma of the nodules contains variable amounts of collagen fibers and foci of large, epithelioid histiocytic cells that exhibit a macrophagic activity for hemosiderin and lipids (mainly cholesterol) (61,118) (Fig. 9-16C). Between these areas giant cells are present, some of which contain up to 50 nuclei (61,74). Dilatation of the blood vessels is often observed, and the histiocytic elements may contain variable amounts of hemosiderin or lipids (150). Mitotic figures are occasionally seen in the stromal and synovial cells (11). Electron microscopic studies have demonstrated that the histiocytic cell population of





Figure 9-15 Pigmented villonodular synovitis: magnetic resonance imaging (MRI). Coronal (A) and sagittal (B) T1-weighted (SE, TR 600, TE 12) MR images of the knee of a 40year-old man show lobulated low-signal-intensity masses mainly localized to the popliteal fossa. C: Sagittal T2-weighted (SE, TR 2300, TE 80) MRI shows high-intensity fluid in the suprapatellar bursa. The lobulated masses of pigmented villonodular synovitis remain of low signal intensity.

R

PVNS is not uniform but contains two basic cell types: (a) fibroblasts that produce collagen and proteoglycan and (b) macrophages that contain hemosiderin and lipids (85,86,130).

The pathologic spectrum of PVNS parallels the clinical features. Over time, the cellularity of the synovium diminishes and the degree of fibrosis increases (74). As the stroma becomes more fibrous, the numbers of foamy macrophages decrease and they may eventually disappear. Lesser numbers of lymphocytes and plasma cells remain, often in a perivascular distribution. Hemosiderin also persists, although much of it usually disappears (74).

Differential Diagnosis

Radiology

The most common diagnostic possibilities include *hemophilia*, synovial chondromatosis, synovial hemangioma, osteoarthritis, rheumatoid arthritis, and tuberculous arthritis

(7,63,83,108,123). MRI is very effective in the differential diagnosis because it can reveal the presence of hemosiderin deposition in PVNS (76) (see Fig. 9-14C), whereas neither monoarticular *rheumatoid arthritis* nor *synovial hemangioma* clearly exhibits this feature. Some investigators, however, postulate that both hemophilia and synovial hemangioma may be associated with an intraarticular bleeding, and hence the hemosiderin deposition (119). Detection of diffuse hemosiderin clumps, synovial irregularity and thickening, and distention of the synovial sac usually suggest the diagnosis of PVNS (140).

Synovial chondromatosis may manifest with pressure erosions of the bone similar to those of PVNS, but it can be distinguished by the presence of multiple joint bodies, calcified or uncalcified (18) (see Fig. 9-6). Synovial hemangioma is usually associated with the formation of phleboliths (15). Synovial sarcoma tends to have a shorter T1 and longer T2 on MRI compared with PVNS, and when calcifications are present the latter di-






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Figure 9-16 Histopathology of pigmented villonodular synovitis. A: Extended strands of roundish histiocytic cells and scattered giant cells having formed bundles of condensed collagen (*center and upper right*). In other areas (*lower right*) the cells are more spindled (hematoxylin and eosin, original magnification $\times 25$). B: At higher magnification there is accumulation of mononuclear cells with interspersed giant cells, which have frequently peripherally arranged nuclei (*center*). Condensed collagen fibers sometimes resemble bone matrix (*bottom*) (hematoxylin and eosin, original magnification $\times 50$). C: In another field of view there are extended areas of lipid-laden macrophages and scattered hemosiderin-containing macrophages (hematoxylin and eosin, original magnification $\times 25$).

agnosis can be excluded. *Lipoma arborescens* has a very characteristic appearance on MRI, with its signal compatible with fatty tissue and its typical frondlike synovial projections (see Fig. 9-30). In addition, there is very little or no enhancement of the mass after administration of gadolinium (16,90).

Hemophilia, unlike PVNS, usually affects multiple joints. The growth disturbances typical of this disorder are not a feature of PVNS. *Tuberculous arthritis*, like PVNS, favors the knee joint and the hip and tends to be monoarticular. On the radiographs, characteristic subchondral cysts and articular erosions are present, with relative preservation (at least in the early stages) of the joint space. However, the most consistent radiographic finding of tuberculosis arthritis is severe juxtaarticular osteoporosis, which is not seen in PVNS.

Monoarticular osteoarthritis (degenerative joint disease) commonly occurs in an older population. Osteoarthritis affecting large joints, such as the knee and hip, may be characterized by cyst-like bony lesions that resemble those of PVNS. The presence of juxtaarticular soft tissue masses is helpful in distinguishing PVNS from osteoarthritis and the former usually exhibits a lack of joint space narrowing. The formation of hypertrophic spurs that often occurs in degenerative joint disease is lacking in PVNS (74). Differentiation between *rheumatoid arthritis* and PVNS is sometimes more difficult because this arthropathy causes bony erosions secondary to the formation of a synovial pannus and to a chronic elevation of intraarticular pressure, which is a similar mechanism in both conditions (74). The lack of hypertrophic bone changes in rheumatoid arthritis constitutes another similarity to PVNS. Again, periarticular osteoporosis and the occasional formation of so-called rice bodies in rheumatoid arthritis serve as important differentiating points. Clinical information is also crucial in this situation.

Pathology

Rheumatoid arthritis may be characterized by synovial hypertrophy and proliferation of villi, which, however, are usually less prominent than in PVNS and during the acute phase contain an inflammatory infiltrate with many plasma cells. These villi also lack conspicuous hemosiderin deposits. *Hemophilia* is sometimes characterized by large hemosiderin deposits, but these are generally confined to the cells that line the synovium. The subsynovial tissue, also unlike the case with PVNS, almost completely lacks hemosiderin (11).

When PVNS invades the neighboring bone and radiologically appears like a primary epiphyseal bone tumor, the histologic presentation may mimic *chondroblastoma*, mainly because of the roundish appearance of histocytic elements (Remagen, unpublished observations, 1998) (Fig. 9-17).

Malignant forms of diffuse giant cell tumor are exceptions and therefore rarely pose a problem in differential diagnosis. However, nodular invasion of soft tissues, sheath-like arrangement of uniform tumor cells, more extensive necrosis, and atypical mitoses are features indicating malignancy (55,145,147). **494** — Differential Diagnosis in Orthopaedic Oncology





Figure 9-17 Histopathology of chondroblastoma-like pigmented villonodular synovitis. A: Cancellous bone from articular end of tibia shows infiltration by large roundish cells exhibiting some pleomorphism and nuclear hyperchromatism, originally mistaken for chondroblastoma (hematoxylin and eosin, original magnification ×100). B: With Giemsa stain the matrix even more resembles a chondroblastic tumor (Giemsa, original magnification ×100). C: The tissue obtained from the interior of the knee joint shows typical appearance of PVNS (hematoxylin and eosin, original magnification ×100).

B

Localized Pigmented Nodular Tenosynovitis [Pigmented Giant Cell Tumor of the Synovium or the (Localized) Giant Cell Tumor of the Tendon Sheath]

Clinical Presentation

The most common presentation of the localized nodular form of villonodular synovitis occurs in the tendon sheaths, mainly on the palmar aspect of the phalanges, and occasionally in the articular synovium (142). Unlike PVNS, there is a 2:1 male predominance (11), although some investigators report a slight female predominance (104,142). It is a well-circumscribed lesion that affects a small area of the synovium (95) or the tendon sheath (104). The main clinical symptom is pain, although the lesion is frequently asymptomatic (81,86).

Imaging

On radiography, localized nodular tenosynovitis is seen as a well-circumscribed, dense soft tissue mass, often in close approximation to a bony erosion (Fig. 9-18) (79). Pathologic fractures are exceedingly rare (151). Occasionally, the iron stored in histocytes may appear as fine dispersed densities within the soft tissue mass (Remagen, unpublished observations, 1998). MRI is rather characteristic. On T1- and T2-weighted images, lesions show predominantly low signal intensity equal to or slightly darker than skeletal muscle. On gadolinium-enhanced

T1-weighted images, strong homogenous enhancement is seen (69,95). These findings reflect the underlying histologic composition of the lesion: Hemosiderin deposition in xanthoma cells and abundant collagenous proliferation are responsible for low signal intensity on T1- and T2-weighted images. Strong homogenous enhancement reflects the presence of numerous proliferative capillaries in the collagenous stroma (67,95,101,104). Scattered reports have recently suggested the usefulness of sonography for evaluation of giant cell tumors of the tendon sheath. In general, this technique can distinguish between solid and cystic masses. In the reported cases the lesion under discussion appeared as a solid homogeneous (most common) or heterogeneous (less common) mass, exhibiting hypoechoic or hyperechoic characteristics (115), and occasionally displaying posterior acoustic enhancement (125). Sonography was effective in excluding the diagnosis of ganglion cyst, which may mimic the giant cell tumor of the tendon sheath, based on the former lesion's characteristic sonographic appearance as a cystic mass (62). In addition to characterizing giant cell tumor as a solid-appearing mass, ultrasound provides information about the extent of contact and the circumferential involvement of the tendon (115).

Histopathology

Pathologically, the tumor is firm, round, oval, or lobulated and is usually contained within a dense fibrotic



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Figure 9-18 Giant cell tumor of the tendon sheath. A: Lateral radiograph of the index finger of a 58-year-old man shows a soft tissue mass at the site of the proximal interphalangeal joint. A small erosion is present at the base of the middle phalanx (*arrow*). **B:** Anteroposterior radiograph of the toes in a middle-aged man shows a soft tissue swelling of the second toe associated with several osteolytic defects in the middle phalanx (*arrows*). (Reprinted from Bullough PG. *Atlas of orthopedic pathology*, 2nd ed. New York: Gower, 1992:17.25.)



Figure 9-19 Histopathology of giant cell tumor of the tendon sheath. The histiocytic character of the tumor tissue prevails. The collagen septa are less conspicuous (van Gieson, original magnification \times 50).

capsule. The size of the tumor varies from a few millimeters to several centimeters in diameter. Microscopic examination reveals collagenous connective tissue with a wide variation in the proportion of matrix to tumor cells (Fig. 9-19). The round to polygonal synovial-like cells are the basic component of this tumor. Mitoses may be present, but rarely are numerous. Although giant cells are a constant feature, they vary in number. Foam cells and hemosiderin-laden macrophages, although often identified, are not a constant feature. Xanthomatous cells often contain hemosiderin granules (67,142). The presence of hemosiderin results from the breakdown of erythrocytes that have extravasated into the tissue.

Differential Diagnosis

Radiology

The differential diagnosis includes *synovial sarcoma*, gout, amyloidosis, dermoid (dermal) inclusion cyst, glomus tumor, enchondroma, periosteal chondroma, and fibroma of the tendon sheath.

Synovial sarcoma is more destructive, the soft tissue mass is usually larger than in giant cell tumor of the tendon sheath, and, when true calcifications are present (imaging much coarser than the fine densities caused by hemosiderin deposits), the latter condition can safely be excluded. *A gouty tophus* may mimic nodular tenosynovitis, but it usually contains radiographically visible calcifications. The bone erosion has a characteristic overhanging edge, and the affected joint is partially preserved.

Enchondroma exhibits a centric location and possesses lobulated borders, with frequent matrix calcifications. *Periosteal chondroma* may closely mimic giant cell tumor of the tendon sheath but, again, almost invariably it displays visible soft tissue calcifications.

Fibroma of the tendon sheath is a rare benign lesion that most commonly affects the extremities (98%), particularly the hands and wrists (82%) (80). The lesion is

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Figure 9-20 Histopathology of giant cell tumor of the tendon sheath. Sclerotic bundles of collagen fibers (*red structures*) resembling osteoid such as produced by malignant cells of osteosarcoma surround clusters of fairly large histiocytes with several giant cells (van Gieson, original magnification \times 50).

composed of tightly packed spindle cells surrounded by collagen fibers. It closely mimics the giant cell tumor of the tendon sheath because of its similar clinical presentation, size, location, and gross appearance at pathology (64). However, the lesion can be distinguished from the latter on MRI, which shows a focal nodular mass adjacent to a tendon sheath with decreased signal intensity on all pulse sequences, but exhibiting little or no enhancement after application of gadolinium, unlike giant cell tumor of the tendon sheath.

Pathology

The cleft-like structures in localized nodular tenosynovitis may suggest the glandular or biphasic form of *synovial sarcoma*. The large numbers of giant cells and the admixture of histiocytes help in the differential diagnosis to exclude the latter malignancy (11). In addition, the well-defined epithelioid cells characteristic of synovial sarcoma are not present. IHC is also helpful showing positivity for cytokeratins, epithelial membrane antigen (EMA), CD99, and BCL2 in synovial sarcoma (201). If the collagen produced appears very hyalinized and therefore resembles osteoid, the condition may at first glance be confused with *soft tissue osteosarcoma* poor in matrix (Fig. 9-20). The monomorphism of the cells and the lack of other malignant features in the nodular form of PVNS are then helpful in differentiation.

The radiologic and pathologic differential diagnosis of PVNS and a giant cell tumor of the tendon sheath is depicted in Figures. 9-21 and 9-22, respectively.

Synovial Hemangioma

Clinical Presentation

Synovial hemangioma is a rare benign lesion that most commonly affects the knee joint, usually involving the anterior compartment (163,165). This lesion has also been found in the elbow (162), wrist, and ankle (157),

Figure 9-21 Radiologic and pathologic differential diagnosis of pigmented villonodular synovitis.

Figure 9-22 Radiologic and pathologic differential diagnosis of giant cell tumor of the tendon sheath.

as well as in tendon sheaths (173). Most cases affect children and adolescents. In one study, 75% of patients with synovial hemangioma of the knee joint presented before the age of 16 years, and 35% had a history of trauma at the site (167). A more recent study involving 20 patients places the average age at the time of presentation at 25 years (range 9 to 49 years) (157). Almost all patients with synovial hemangioma are symptomatic, frequently presenting with a swollen knee or with mild pain or limitation of movement in the joint (170). Sometimes patients report a history of recurrent episodes of joint swelling and various degrees of pain of several years' duration. Synovial hemangioma is often associated with an adjacent cutaneous or deep soft tissue hemangioma. For this reason, some investigators classify knee joint lesions as intraarticular, juxtaarticular, or intermediate, depending on the extent of involvement (161). Synovial hemangioma is frequently misdiagnosed, and according to one estimate a correct preoperative diagnosis is made in only 22% of cases (157). Pathogenesis of this lesion is yet unclear, and the possibilities of it being either reactive, malformation, or tumor are still under consideration (159).

Imaging

Until recently, synovial hemangiomas were evaluated by a combination of radiography, arthrography, angiography, and contrast-enhanced CT. Although radiographs appear normal in at least half of the patients, they may reveal soft tissue swelling, a mass around the joint, joint effusion, or bone erosions (169,171) (Fig. 9-23). Phleboliths, periosteal thickening, advanced maturation of the epiphysis, and arthritic changes are also occasionally noted on the radiographs (167) (Fig. 9-24). Arthrography usually shows nonspecific filling defects with a villous configuration. Angiograms yield much more specific information than radiography (166). They can often reveal a vascular lesion (164) and can demonstrate pathognomonic features of hemangioma, leading to a presumptive diagnosis. Contrast-enhanced CT of the joint typically reveals a heterogeneous-appearing soft tissue mass that displays tissue attenuation approximating that of skeletal muscle and contains areas of decreased attenuation, some approaching that of fat (155) (Fig. 9-25). CT is effective for demonstrating phleboliths and revealing patchy enhancement around them, as well as enhancement of tubular areas and contrast pooling within the lesion. In some cases, CT reveals enlarged vessels feeding and draining the mass, as well as enlarged adjacent subcutaneous veins (160).

At present, MRI is the modality of choice for evaluation of hemangiomas because with this technique a presumptive diagnosis can be made. The soft tissue mass typically exhibits an intermediate signal intensity on T1-weighted sequences, appearing isointense with or slightly brighter than muscle but much less bright than fat. The mass is usually much brighter than subcutaneous fat on T2-weighted images and on fat suppression sequences (Fig. 9-26), and shows thin, often serpentine low-intensity septa within it (15,156,172) (Fig. 9-27). In general, the signal intensity characteristics of hemangiomas appear to be related to a number of factors, including slow flow, thrombosis, vessel occlusion, and increased free water in stagnant blood that

Figure 9-23 Synovial hemangioma. Anteroposterior **(A)** and lateral **(B)** radiographs of the right knee of a 7-year-old boy show articular erosions at femoropatellar and femorotibial joint compartments. Soft tissue masses are seen anteriorly and posteriorly. An incidental finding is a nonossifying fibroma in the posterior tibia (*arrows*). (Reprinted with permission from Greenspan A, Azouz EM, Matthews J II, Décarie J-C. Synovial hemangioma: imaging features in eight histologically proved cases, review of the literature, and differential diagnosis. *Skeletal Radiol* 1995;24:583–590.)

Figure 9-24 Synovial hemangioma. Dorsovolar radiograph of the left wrist of a 15-year-old boy shows narrowing of the radiocarpal joint, and erosions of the scaphoid and lunate bones (*arrows*). Also noted are irregular articular surfaces of the trapezium-scaphoid and fifth carpometacarpal joints. (Reprinted with permission from Greenspan A, Azouz EM, Matthews J II, Décarie J-C. Synovial hemangioma: imaging features in eight histologically proved cases, review of the literature, and differential diagnosis. *Skeletal Radiol* 1995;24: 583–590.)

Figure 9-25 Synovial hemangioma: computed tomography. Axial CT section through the knee (same patient as illustrated in Fig. 9.23) shows marginal erosions of the femoral condyles. Heterogeneous masses are seen anteriorly and posteriorly with hypodense areas representing fat. (Reprinted with permission from Greenspan A, Azouz EM, Matthews J II, Décarie J-C. Synovial hemangioma: imaging features in eight histologically proved cases, review of the literature, and differential diagnosis. *Skeletal Radiol* 1995;24: 583–590.)

Figure 9-26 Synovial hemangioma: magnetic resonance imaging (MRI). A: Sagittal T1-weighted (SE, TR 400, TE 11) MRI in a 9-year-old boy shows masses isointense with muscle involving the suprapatellar bursa and infrapatellar Hoffa fat. **B:** With fat-suppression technique the mass becomes very bright. The fluid-fluid level seen in the popliteal region (*arrowheads*) is typical for the cavernous type of synovial hemangioma. In another patient, a 16-year-old girl, axial T2*-weighted (MPGR, TR 500, TE 15, flip angle 30 degrees) MRI **(C)** and axial fast spin-echo (FSE, TR 5000, TE 85 Ef) image with fat-suppression technique **(D)** show the size and extent of the synovial hemangioma. (Reprinted with permission from Greenspan A, Azouz EM, Matthews J II, Décarie J-C. Synovial hemangioma: imaging features in eight histologically proved cases, review of the literature, and differential diagnosis. *Skeletal Radiol* 1995;24:583–590.)

pools in larger vessels and dilated sinuses, as well as to the variable amounts of adipose tissue in the lesion (155). After intravenous injection of gadolinium, there is evidence of enhancement of the hemangioma (156). In patients with a cavernous hemangioma of the knee, fluid-fluid levels are also observed (see Fig. 9-26B), a finding recently reported also in soft tissue hemangiomas of this type (158).

Histopathology

С

Originating in the subsynovial layer mesenchyme of the synovial membrane, synovial hemangioma is a vascular lesion that contains variable amounts of adipose, fibrous, and muscle tissue, as well as thrombi in the vessels. When the lesion is completely intraarticular, it is usually well-circumscribed and apparently encapsulated, attached to the synovial membrane by a pedicle of variable size and adherent to the synovium on one or more surfaces by separable adhesions (154). Grossly, the tumor is a lobulated soft, brown, doughy mass with overlying villous synovium that is often stained mahogany-brown by hemosiderin. On microscopic examination, the lesion exhibits arborizing vascular channels of different sizes and a hyperplastic overlying synovium, which may show abundant iron deposition in chronic cases with repeated hemarthrosis (153) (Fig. 9-28).

Figure 9-27 Synovial hemangioma: magnetic resonance imaging (MRI). Sagittal T2-weighted (SE, TR 2000, TE 80) MRI of a 47-year-old woman demonstrates thin, low-intensity fibrofatty septa within the lesion (*arrows*). (Reprinted with permission from Greenspan A, Azouz EM, Matthews J II, Décarie J-C. Synovial hemangioma: imaging features in eight histologically proved cases, review of the literature, and differential diagnosis. *Skeletal Radiol* 1995;24:583–590.)

Differential Diagnosis

Radiology

The differential diagnosis of synovial hemangioma includes *PVNS* and *synovial chondromatosis*. All proliferative chronic inflammatory processes, such as *rheumatoid arthritis, tuberculous arthritis*, and *hemophilic arthropathy*, should also be considered in the differential, but these

Figure 9-28 Histopathology of synovial hemangioma. The lesion consists of a network of connected blood-filled spaces within the loose connective tissue of the synovium (hematoxylin and eosin, original magnification \times 12.5).

conditions when involving the knee can usually be distinguished clinically. Because it is extremely uncommon, lipoma arborescens is rarely included in the differential diagnosis (152,174). MRI is diagnostic for this condition, showing typical frond-like projections of the lesion and fat characteristics (bright on T1- and intermediate on T2-weighted images) (183,186) (see Fig. 9-30). In PVNS, radiographs commonly reveal findings similar to those of synovial hemangioma, such as joint effusion and a mass in the suprapatellar bursa or popliteal fossa region (see Fig. 9-12). Radiographs may also demonstrate bone erosions on both sides of the joint, which in one study were reported in 26% of 81 patients (74). As in synovial hemangioma, arthrography in PVNS may demonstrate the synovial mass as a nonspecific filling defect (175) (see Fig. 9-13B). Without contrast enhancement, CT cannot identify decreased attenuation associated with fat within the tumor; contrast-enhanced CT, however, is diagnostic (60). The synovium in PVNS exhibits nodular thickening and masses of heterogeneous signal intensity. The majority of the lesion will display a higher signal intensity than muscle on both T1- and T2weighted sequences, with other portions exhibiting a low signal intensity on all sequences, reflecting the hemosiderin content of the tumor (135). Synovial chondromatosis can often be diagnosed by radiography when nodules are calcified. Intraarticular osteochondral fragments of uniform size are almost pathognomonic for this condition (see Figs. 9-3 and 9-4). CT may be helpful in demonstrating faint calcifications not otherwise seen, and MRI may be useful in delineating the extent of the disease. However, neither modality is of particular value in establishing the diagnosis of synovial chondromatosis (168).

Pathology

Synovial hemangioma in the great majority of cases can be diagnosed on a histopathologic basis with a high degree of confidence. Rarely, cases with atypical presentation may resemble *hemangioendothelioma*.

The radiologic and pathologic differential diagnosis of synovial hemangioma is depicted in Figure 9-29.

Lipoma Arborescens

Clinical Presentation

Lipoma arborescens, also known as villous lipomatous proliferation of the synovial membranes, is a rare intraarticular disorder characterized by a nonneoplastic lipomatous proliferation of the synovium (35,181). The term "arborescens" describes the characteristic treelike morphology of this lesion, which resembles a frond-like mass (177). Its cause remains unclear, although an association with osteoarthritis, rheumatoid arthritis, psoriasis, diabetes mellitus, and trauma has been postulated (184,188), suggesting a reactive rather than a neoplastic process (182). Lipoma arborescens is more prevalent in males, usually in the fourth and fifth decades. It is usually a monoarticular disease, most commonly affecting the knee joint, although involvement of other joints, such as shoulder,

Figure 9-29 Radiologic and pathologic differential diagnosis of synovial hemangioma.

hip, wrist, elbow, and ankle, has been sporadically reported by various authors (16,90,152,174,176,178, 179). Occasionally this condition may affect multiple joints (177,184). There have been sporadic reports of bursae involvement (180,185). Most patients present with a long-standing, slowly progressing swelling of the joint, initially painless but later associated with pain and limited motion.

Imaging

Radiography shows only a soft tissue density within the joint, commonly interpreted as "joint effusion." On CT, however, the synovial mass is obvious, and attenuation coefficient measurements are consistent with fat. There is very little or no enhancement of the mass after administration of intravenous contrast (90). The appearance on MRI is very characteristic and allows definite diagnosis of this condition (183,186,187,188). Joint effusion is invariably present, associated with frond-like masses arising from the synovium that have the signal intensity of subcutaneous fat (bright on T1 and intermediate on T2 weighting) on all imaging sequences (Fig. 9-30). Occasionally, a chemical shift artifact is present at the fat-fluid interface.

Pathology

Histopathologically, lipoma arborescens is characterized by hyperplasia of subsynovial fat, formation of mature fat cells, and the presence of proliferative villous projections. Osseous and chondroid metaplasia can occur (145).

Differential Diagnosis

Radiology

The differential diagnosis of lipoma arborescens includes mainly PVNS. synovial chondromatosis. and synovial hemangioma. In PVNS, radiography reveals findings similar to those of lipoma arborescens, i.e., joint effusion and an intraarticular mass. MRI, however, is diagnostic because of characteristic imaging of fatty areas in the latter condition, whereas PVNS exhibits on T2-weighted images the paramagnetic effects of hemosiderin deposition. Synovial chondromatosis is most commonly associated with a number of radiopaque or radiolucent intraarticular bodies and commonly with bone erosions. In contrast to lipoma arborescens, the joint space is preserved. Exclusion of synovial hemangioma can be based on CT and MRI. In lipoma arborescens the former modality will demonstrate obvious synovial masses and Hounsfield attenuation coefficient measurements consistent with fat. In addition, minimal or no enhancement of the mass after administration of intravenous contrast will be observed. MRI will show the previously discussed characteristic features of fatty proliferation in this condition.

Pathology

Lipoma arborescens poses no problems in differentiating this lesion from other conditions associated with synovial proliferation, such as *PVNS* or *hypertrophic synovitis*.

Figure 9-50 Lipoma arborescens: magnetic resonance imaging (MRI). A 54-year-old woman presented with history of fullness in the left knee for the past 5 months. Conventional radiography (not shown here) revealed knee joint effusion. **A:** Sagittal proton-density MRI shows numerous structures within suprapatellar bursa exhibiting signal intensity consistent with fat (*arrows*). Coronal (**B**) and sagittal (**C**) T2weighted fat-suppressed images demonstrate high signal intensity joint effusion. Hypertrophic synovial villa (*arrows*) show signal consistent with fat. (Reprinted from Greenspan A. *Orthopedic imaging – a practical approach*, 4th ed. Philadelphia: Lippincott Williams & Wilkins, 2004.)

The radiologic and pathologic differential diagnosis of lipoma arborescens in shown in Figure 9-31.

Malignant Tumors

Synovial Sarcoma

Synovial sarcoma (previously called "synovioma") is an uncommon malignant mesenchymal neoplasm, comprising about 8% to 10% of soft tissue sarcomas (201). Despite its name, it does not arise from synovium (217). It occurs mainly in paraarticular regions close to joint capsules, bursae, and tendon sheaths and also in regions without any relation to synovial structures, e.g., parapharyngeal or abdominal (197).

The etiology of synovial sarcoma is undetermined (201). A consistent finding, present in about 90% of cases, is a translocation involving chromosomes X and 18 [t(X;18) (p11;q11)], giving rise to a fusion transcript of the *SYT* (for synovial sarcoma translocation) gene on chromosome 18 with the *SSX* (for synovial sarcoma X chromosome breakpoint) gene on the X chromosome. Involved are three different breakpoint regions designated *SSX1*, *SSX2*, and *SSX4* (201). The fusion protein appears to be involved in gene transcription (231). For diagnosis, break-apart fluorescent in situ hybridization (FISH) probes for the *SYT* gene on 18q11.2 (see Fig. 9-37E) or molecular analysis by reverse transcriptase polymerase chain reaction (RT-PCR) is successfully employed in addition to IHC (201,205).

Clinical Presentation

The tumor usually occurs before the age of 50 (197), most commonly between the ages of 16 and 36 years (196), although cases in much younger pediatric patients have been reported (214). There is no sex predilection, although recently Fisher et al. (201) suggested slight male preponderance. The extremities account for 83% of synovial sarcomas, but the most common sites are around the knee and foot (197) (Fig. 9-32). In exceptional instances the tumor may be intraarticular (11,194,215). Synovial sarcoma is usually slow growing with an indolent course, although in late stages it may demonstrate aggressiveness. Metastases to the lung by the hematogenous route and to the soft tissue have been reported (216). Schajowicz (225) cited a local recurrence rate of greater than 50%. The clinical symptoms usually include soft tissue swelling or a mass and progressive pain. On physical examination, a diffuse or a discrete soft tissue mass is present, usually tender on palpation.

Imaging

The radiographic features of synovial sarcoma include a soft tissue mass, usually in close proximity to a joint (Fig. 9-33) and occasionally associated with bone invasion (193,206). A periosteal reaction may also be observed (229). The soft tissue calcifications, usually amorphous in type, are present in about 25% to 30% of cases (197,224) (Fig. 9-34).

CT effectively demonstrates the extent of the soft tissue mass, calcifications, and bone invasion (191). MRI shows the tumor to be a heterogeneous, septated mass of low to intermediate signal intensity with infiltrative margins on T1-weighted sequences, displaying a high signal on T2 weighting (204,211,212,220,230) (Figs. 9-35 and 9-36), and diffuse enhancement after administration of gadolinium (232,237). This modality can also demonstrate cyst formation within the tumor (222). The most extensive MRI study of synovial sarcoma in 34 patients showed that it tends to be deep, large (85% were >5 cm in diameter), and located in the extremities, with the epicenter close to a joint. The

Figure 9-32 Synovial sarcoma: skeletal sites of predilection, peak age range, and male-to-female ratio.

Figure 9-33 Synovial sarcoma. A lateral radiograph of the left ankle of a 71-year-old woman shows a large calcified mass located in the soft tissues anteriorly to the Achilles tendon not affecting the adjacent bones.

Figure 9-34 Synovial sarcoma. A dorsoplantar radiograph of the right forefoot of a 55-year-old woman shows a large mass exhibiting coarse calcifications, eroding the proximal phalanx of the second toe. (Courtesy of Dr. Harold Jacobson, The Bronx, New York.)

Figure 9-35 Intraarticular synovial sarcoma. External oblique radiograph of the left elbow **(A)** and axial T1weighted (SE, TR 600, TE 20) magnetic resonance imaging (MRI) **(B)** in an 8-year-old girl show a soft tissue mass arising from the elbow joint, displaying faint calcifications.

lesion was usually heterogeneous on T2-weighted images and was clearly delineated from surrounding tissues. Forty-four percent of the cases had a high signal on both T1- and T2-weighted sequences, consistent with a hemorrhage within the tumor. Fluid-fluid levels, best visualized on T2-weighted images, were present in 18% of patients. The tumor commonly involved the adjacent bone, invading, eroding, or touching it in 71% of patients (208).

Histopathology

Synovial sarcoma, unlike the term implies, does not arise from synovial tissue (217,227). Most of the tumors occur in extraarticular locations and bear no resemblance to synovium, neither ultrastructurally nor immunohistochemically (218). It has been suggested that synovial sarcoma originates from the pluripotential mesenchyme of the limb bud (234). Several studies

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Figure 9-36 Intraarticular synovial sarcoma. A: Anteroposterior radiograph of the left of a 37-year-old man shows an osteolytic lesion in the femoral neck bordered laterally by sclerotic margin (*arrows*). **B:** Scintigraphic (blood pool) examination demonstrates increased vascularity to the left hip joint (*arrows*) (*continued*).

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Figure 9-36 *Continued* **C:** Delayed radionuclide bone scan with 99m-technetium methylene diphosphonate (MDP) shows increased uptake of the radiopharmaceutical tracer in the femoral head and neck and around the hip joint. **D:** Coronal T1-weighted (SE, TR 850, TE 20) MRI shows a low-signal-intensity lesion affecting the medial aspect of the left femoral neck (*arrow*). **E:** Coronal T2-weighted (SE, TR 2000, TE 80) MRI demonstrates increased signal at the junction of femoral head and neck and in the medial and lateral aspects of the hip joint (*arrows*).

have demonstrated a close relationship between cells that differentiate into osteoblasts and those that differentiate into synovioblasts (190,198,209). This factor may explain the formation of calcifications and ossifications in some synovial sarcomas.

Several subtypes of synovial sarcoma are recognized (235). Among them are biphasic (fibrous and epithelial), monophasic, poorly differentiated (45,192), purely glandular, and calcifying types (145,201). The classical biphasic type exhibits distinct spindle-cell and epithelial components arranged in glandular or nestlike patterns (Fig. 9-37A). The monophasic synovial sarcoma is composed of interdigitating fascicles and "ball-like" structures formed by the spindle cells (Fig. 9-37B). Anastomosing vascular channels similar to those seen in hemangiopericytoma are occasionally present. Foci of calcification may also be observed, usually localized in areas of hyalinization within the spindle-cell elements of the tumor in the calcifying type, which may also present with ossification and is associated with a better prognosis (197,218) (Fig. 9-38A, B). At the ultrastructural level, abundant microvilli and many desmosomal attachments can be seen (192). The spindle cells are closely apposed to each other and appear polygonal or oval when viewed by electron microscopy. The cytoplasm demonstrates well-developed Golgi complexes, rough endoplasmic reticulum, polyribosomes, and glycogen particles (210). In the monophasic fibrous type, a spindle cell pattern predominates, with only a small number of epithelial cells. The monophasic epithelial type is rare and closely resembles metastatic carcinoma (192,200). This type exhibits glandular structures lined with epithelial cells. If pure glandular forms really exist, they could be diagnosed with certainty only by molecular methods. Practically, minute foci of spindle cells have

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Figure 9-37 Histopathology of synovial sarcoma. A: Typical biphasic appearance of synovial sarcoma with gland-like spaces (*left and below*) side-by-side with spindle-cell sarcomatous areas (hematoxylin and eosin, original magnification \times 80). **B:** In rare cases, the tumor has a monophasic appearance without any gland-like patterns, resembling fibrosarcoma (hematoxylin and eosin, original magnification \times 50). **C:** Monophasic fibrous synovial sarcoma shows focal positivity for epithelial membrane antigen (EMA) (biotin-avidin peroxidase, original magnification \times 400). **D:** In addition to positivity for CD99 and cytokeratins (not shown here), synovial sarcoma is also positive for BCL2 (biotin-avidin peroxidase, original magnification \times 200). **E:** Using a double-labeled fluorescent break-apart probe for the SYT-gene, red and green signal splitting indicates gene rearrangement (FISH, original magnification \times 1,000).

С

Figure 9-38 Histopathology of synovial sarcoma. A: Calcifying type of synovial sarcoma exhibits extensive ossification and mineralization (hematoxylin and eosin, original magnification $\times 100$). **B:** Calcifying type of synovial sarcoma mimics an osteosarcoma; however, observe uniformity of spindle cells and absence of overt pleomorphism (hematoxylin and eosin, original magnification $\times 400$).

been present in most of these cases, identifying them as biphasic synovial sarcomas, by definition (145,201). The poorly differentiated type contains cells resembling an intermediate form with characteristics of both epithelial and spindle cells (145). Immunohistochemical studies have shown a strong positivity for cytokeratin and EMA in the epithelial areas (see Fig. 9-37C) and a clear-cut positivity for the same markers, in addition to vimentin, in the spindle cell elements (11). Furthermore, synovial sarcomas express CD99 and BCL2 (see Fig. 9-37D), whereas CD34 is almost always absent (201).

Differential Diagnosis

Radiology

Α

The radiologic differential diagnosis should include *soft tissue chondroma, soft tissue chondrosarcoma, soft tissue osteosarcoma, soft tissue fibrosarcoma* or *malignant fibrous histiocytoma (MFH), myositis ossificans, gout, and tumoral calcinosis.* Except for fibrosarcoma and MFH, all of these entities exhibit calcifications. Therefore, if synovial sarcoma contains areas of mineralization, it may mimic the previously mentioned lesions (195,207,224,233).

Soft tissue chondroma usually presents as a small and well-defined mass. Unlike synovial sarcoma, the lesion never invades bone. Soft tissue chondrosarcoma is a rare tumor that usually arises in the proximal parts of the extremities and the buttocks (194). This neoplasm may display an aggressive behavior and hence may invade adjacent skeletal structures (226,238). Soft tissue osteosarcoma is also much less common than synovial sarcoma. The preferred locations are the upper thighs and the buttocks. Like chondrosarcoma, this lesion may invade the osseous structures. Myositis ossificans usually exhibits a distinctive zoning phenomenon, and juxtacortical lesions may evoke a well-organized pe-

riosteal reaction, although the cortex is not destroyed (189,203,223). A gouty tophus usually occurs adjacent to the joint capsule, and calcifications are less sharply defined. The articular erosion usually exhibits a characteristic overhanging edge. Tumoral calcinosis is a rare metabolic disease characterized by the presence of single or multiple lobulated cystic masses containing chalky material (202). The condition can be idiopathic or associated with an abnormal phosphate metabolism with a high calcium-phosphate product, often due to secondary hyperparathyroidism (221). Radiographs reveal well-demarcated, well-lobulated heterogeneous calcific masses located around the joints (236). A consistent radiologic finding on conventional radiographs and CT is that of a dense calcified mass that is homogeneous except for a "chicken wire" pattern of lucencies within the mass, giving the calcifications a "cobblestone" appearance (228) that is not seen in synovial sarcoma. The adjacent bones are not invaded, although a pressure erosion of the cortex adjacent to the calcified mass may occasionally occur (228). The diagnosis of idiopathic tumoral calcinosis is one of exclusion. All other causes of soft tissue calcifications must be considered and excluded before this diagnosis is made.

Pathology

The histologic differential diagnosis of synovial sarcoma includes *soft tissue osteosarcoma, parosteal osteosarcoma, periosteal fibrosarcoma, Ewing sarcoma,* and *metastatic carcinoma*. Some synovial sarcomas are characterized by amorphous calcifications accompanied by extensive ossification, sometimes having a ribbon-like pattern, that simulates osteosarcoma (218). It is important to recognize this variant of synovial sarcoma with bone formation (see Fig. 9-38) and to distinguish it from *extraskeletal* or *parosteal osteosarcoma*. The most important point in the recognition of synovial sarcoma is its biphasic pattern and the uniform appearance of the nuclei of the epithelial and spindle cells. Another factor that assists in the diagnosis of synovial sarcoma is the finding of amorphous concretions arranged in sheets and of small calcospherites located within spaces that are surrounded by flat or conspicuous epithelial cells (218). The cells are immunoreactive for cytokeratin, EMA, CD99, and BCL2. A synovial sarcoma with an extensive spindle cell component (see Fig. 9-37B) may be confused with *fibrosarcoma* (199). Poorly differentiated round cell areas may resemble Ewing sarcoma (145). Moreover, tumors with predominant epithelial glandular (213) or squamous (219) components are difficult to differentiate from metastatic carcinoma (218). In this situation, as mentioned previously, molecular genetic studies revealing the t(X:18) (p11;q11) translocation or the SYT-SSX fusion transcript will lead to a correct diagnosis.

The radiologic and pathologic differential diagnosis of synovial sarcoma is depicted in Figure 9-39.

Synovial Chondrosarcoma

Synovial chondrosarcoma is a rare tumor that originates from the synovial membrane. It may arise as a primary synovial tumor or it may develop as a malignant transformation of synovial chondromatosis (2,239,240). The concept of malignant degeneration of synovial chondromatosis is still controversial and the entity is rare: fewer than 40 well-documented cases are on record (17,21,45,241–248).

Clinical Presentation

Most synovial chondrosarcomas are located in the knee joint. Rarely other joints such as the hip, elbow, or ankle are affected. These malignancies show a slight predominance in men and the patients range in age from 25 to 70 years (Fig. 9-40). The symptoms include pain and swelling, with duration in most patients exceeding 12 months (11). In patients with primary synovial chondromatosis, malignant transformation to synovial chondrosarcoma should be clinically suspected if there is development of a soft tissue mass at the site of the affected joint.

Imaging

Radiologically the presence of chondroid calcifications within the joint, destruction of the adjacent bones, and a soft tissue mass are highly suggestive of a synovial chondrosarcoma. In patients with documented primary synovial (osteo) chondromatosis, a soft tissue mass and destructive changes in the joint should alert one to the development of a secondary synovial chondrosarcoma (Fig. 9-41). It has to be pointed out, however, that

Figure 9-39 Radiologic and pathologic differential diagnosis of synovial sarcoma.

Figure 9-40 Synovial chondrosarcoma: skeletal sites of predilection, peak age range, and male-to-female ratio.

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Figure 9-41 Secondary synovial chondrosarcoma. Anteroposterior **(A)** and lateral **(B)** radiographs of the right ankle of a 64-year-old man with a long history of synovial chondromatosis show a large soft tissue mass on the dorsal aspect of the ankle joint, eroding the talus. Multiple calcifications, uniform in size and shape, are noted laterally (*arrow*). **C:** After injection of 15 mCi (555 MBq) of 99m-technetium-labeled methylene diphosphonate (MDP) there is increased uptake of radiopharmaceutical tracer in the right ankle (*continued*).

Е

Figure 9-41 *Continued* **D**: Sagittal T1-weighted (SE, TR 400, TE 20) magnetic resonance imaging (MRI) shows the mass displaying intermediate signal intensity, isointense with the muscles. **E**: Parasagittal T1-weighted (SE, TR 400, TE 20) image demonstrates the mass to be well encapsulated. **F**: Coronal proton density (SE, TR 1800, TE 29) MRI shows that the mass is continuous with the ankle joint. **G**: Coronal T2-weighted (SE, TR 2000, TE 80) image demonstrates the mass to be of high signal intensity. Punctate areas of low signal intensity within the mass represent calcifications. (Reprinted with permission from Ontell F, Greenspan A. Chondrosarcoma complicating synovial chondromatosis: findings with magnetic resonance imaging. *Can Assoc Radiol J* 1994;45:319–320.)

frequently both uncomplicated synovial chondromatosis and synovial chondrosarcoma may exhibit similar features on radiography and MRI (246).

Histopathology

The histopathologic distinction between primary synovial chondromatosis and secondary malignancy in synovial chondromatosis has been a matter of dispute. Manivel et al. (28) suggested that histologic features equivalent to those of grade 2 or 3 central chondrosarcoma must be present before chondrosarcoma arising in synovial chondromatosis can be diagnosed. Occasional foci of increased cellularity showing hyperchromatic atypical cells, consistent with grade 1 chondrosarcoma, should not be sufficient evidence for a malignant change in synovial chondromatosis (40). However, evidence of aggressive growth (invasion) and a lesion's lack of attachment to the synovial lining, combined with hypercellularity and pleomorphisms of the cells, should support the diagnosis of malignancy (Figs. 9-42 and 9-43).

Bertoni et al. (2) have attempted to develop criteria for making this crucial distinction. They identified several microscopic features indicative of malignancy. The distinguishing features of synovial chondrosarcoma include the following: tumor cells arranged in sheets, myxoid changes in the matrix, hypercellularity with crowding and spindling of nuclei at the periphery, necrosis, and permeation of bone trabeculae. Remarking on the danger of misinterpreting synovial chondromatosis as chondrosarcoma on both radiographic and histopathologic examination, these investigators singled out pulmonary metastases as the only distinguishing feature.

Differential Diagnosis

Radiology

The main differential diagnosis is between synovial chondrosarcoma and *synovial (osteo)chondromatosis*. Frequently, the radiologic findings in both conditions are similar, although the development of destructive changes around the affected joint favors synovial chondrosarcoma. However, these destructive changes should be differentiated from periarticular erosions occasionally present in synovial chondromatosis (34). *PVNS* can usually be excluded without much difficulty because it does not exhibit calcifications and, in addition, shows

somewhat characteristic MRI features (see previous discussion). Exceedingly rare location of synovial chondrosarcoma in small joints of the hand can mimic *gout* (249). It should be emphasized, however, that even severe cellular atypia per se is not sufficient to render the diagnosis of synovial chondrosarcoma.

R

Pathology

The difficulties in distinguishing secondary synovial chondrosarcoma from *synovial chondromatosis* have been discussed previously.

The radiologic and pathologic differential diagnosis of synovial chondrosarcoma is depicted in Figure 9-44.

Figure 9-43 Histopathology of secondary synovial chondrosarcoma. A: The chondrocytes with clearly pleomorphic, sometimes large blastic nuclei (lower right corner) are unevenly distributed. These differences in the nuclear size and structure, however, do not indicate malignancy, and may be features of uncomplicated synovial chondromatosis (hematoxylin and eosin, original magnification \times 50). B: Smaller parts of the benign synovial lesion (lower right) border the cell-rich tissue of secondary chondrosarcoma of grade 2-3 (hematoxylin and eosin, original magnification \times 50). **C:** At the proliferation line of the tumor against the fibrous connective tissue there is more matrix formation of the tumor, which here is a grade 2 sarcoma (hematoxylin and eosin, original magnification \times 25).

Figure 9-44 Radiologic and pathologic differential diagnosis of synovial chondrosarcoma.

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REFERENCES

Synovial Chondromatosis

- Berman DM, Karhadkar SS, Maitra A, et al. Widespread requirement for hedgehog ligand stimulation in growth of digestive tract tumours. *Nature* 2003;425:846–851.
- Bertoni F, Unni KK, Beabout JW, Sim FH. Chondrosarcomas of the synovium. *Cancer* 1991;67:155–162.
- 3. Blacksin MF, Ghelman B, Freiberger RH, Salvata E. Synovial chondromatosis of the hip: evaluation with air computed arthrotomography. *Clin Imag* 1990;14:315–318.
- Buddingh EP, Krallman P, Neff JR, et al. Chromosome 6 abnormalities are recurrent in synovial chondromatosis. *Cancer Genet Cytogenet* 2003;140:18–22.
- Burnstein MI, Fisher DR, Yandow DR, et al. Case report 502. Intra-articular synovial chondromatosis of shoulder with extra-articular extension. *Skeletal Radiol* 1988;17:458–461.
- Chen A, Shih S-L, Chen B-F, Sheu C-Y. Primary synovial osteochondromatosis of the first metatarsophalangeal joint. *Skeletal Radiol* 2002;31:122–124.
- Conway WF, Hayes CW. Miscellaneous lesions of bone. Radiol Clin North Am 1993;31:339–358.
- 8. Davis RI, Foster H, Arthur K, et al. Cell proliferation studies in primary synovial chondromatosis. *J Pathol* 1998;184:18–23.
- Davis ŘI, Hamilton A, Biggart JD. Primary synovial chondromatosis: a clinicopathologic review and assessment of malignant potential. *Hum Pathol* 1998;29:683–688.
- Eggenschwiler JT, Espinoza E, Anderson KV. Rab23 is an essential negative regulator of the mouse Sonic hedgehog signalling pathway. *Nature* 2001;412:194–198.
- 11. Fechner RE, Mills SE. *Tumors of the bones and joints.* Washington, DC: Armed Forces Institute of Pathology, 1993.
- Feldman F. Cartilaginous tumors and cartilage-forming tumorlike conditions of the bones and soft tissues. In: Ranniger K, ed. *Bone tumors*. Berlin: Springer-Verlag, 1977:83–242.
- Ginaldi S. Computed tomography feature of synovial osteochondromatosis. *Skeletal Radiol* 1980;5:219–222.
- Goldman RL, Lichtenstein L. Synovial chondrosarcoma. Cancer 1964;17:1233–1240.
- Greenspan A, Azouz EM, Matthews J II, Décarie J-C. Synovial hemangioma: imaging features in eight histologically proved cases, review of the literature, and differential diagnosis. *Skeletal Radiol* 1995;24:583–590.
- Hallel T, Lew S, Bansal M. Villous lipomatous proliferation of the synovial membrane (lipoma arborescens). *J Bone Joint Surg Am* 1988;70:264–270.
- Hamilton A, Davis RI, Hayes D, Mollan RA. Chondrosarcoma developing in synovial chondromatosis. A case report. J Bone Joint Surg Br 1987;69:137–140.
- Hermann G, Abdelwahab IF, Klein MJ, et al. Synovial chondromatosis. Skeletal Radiol 1995;24:298–300.
- Hopyan S, Nadesan P, Yu C, et al. Dysregulation of hedgehog signalling predisposes to synovial chondromatosis. J Pathol 2005;206:143–150.
- Kindblom L-G, Angervall L. Myxoid chondrosarcoma of the synovial tissue: a clinicopathologic, histochemical, and ultrastructural analysis. *Cancer* 1983;52:1886–1895.
- King JW, Spjut HJ, Fechner RE, Vanderpool DW. Synovial chondrosarcoma of the knee joint. J Bone Joint Surg Am 1967;49: 1389–1396.
- Kramer J, Recht M, Dely DM, et al. MR appearance of idiopathic synovial chondromatosis. J Comput Assist Tomogr 1993;17: 772–776.
- Kudawara I, Aono M, Ohzono K, Mano M. Synovial chondromatosis of the acromioclavicular joint. *Skeletal Radiol* 2004;33:600–603.
- 24. Laorr A, Helms CA. MRI of musculoskeletal masses: a practical text and atlas. New York: Igaku-Shoin, 1997.
- Llauger J, Palmer J, Monill JP, et al. MR imaging of benign softtissue masses of the foot and ankle. *RadioGraphics* 1998;18: 1481–1498.
- Llauger J, Palmer J, Rosón N, et al. Nonseptic monoarthritis: Imaging features with clinical and histopathologic correlation. *RadioGraphics* 2000;20:S263–S278.

- Madewell JE, Sweet DE. Tumors and tumor-like lesions in or about joints. In: Resnick D, Niwayama M, eds. *Diagnosis of bone* and joint disorders Philadelphia: WB Saunders, 1988.
- Manivel JC, Dehner LP, Thompson R. Case report 460. Synovial chondrosarcoma of left knee. *Skeletal Radiol* 1988;17:66–71.
- Marigo V, Roberts DJ, Lee SMK, et al. Cloning, expression, and chromosomal location of SHH and IHH: two human homologues of the Drosophila segment polarity gene hedgehog. *Genomics* 1995;28:44–51.
- Maurice H, Crone M, Watt I. Synovial chondromatosis. J Bone Joint Surg Br 1988;70:807–811.
- Milgram JW. Synovial osteochondromatosis. A histopathological study of thirty cases. J Bone Joint Surg Am 1977;59:792–801.
- Mohr W. Is synovial osteo-chondromatosis a proliferative disease? Pathol Res Pract 2002;198:585–588.
- Murphy FP, Dahlin DC, Sullivan CR. Articular synovial chondromatosis. J Bone Joint Surg Am 1962;44:77–86.
- Norman A, Steiner GC. Bone erosion in synovial chondromatosis. *Radiology* 1986;161:749–752.
- Ryu KN, Jaovisidha S, Schweitzer M, et al. MR imaging of lipoma arborescens of the knee joint. *Am J Roentgenol* 1996;167: 1229–1232.
- Osburn AW, Bassett LW, Seeger LL, et al. Case report 609. Synovial (osteo)chondromatosis. *Skeletal Radiol* 1990;19:237–241.
- Robinson D, Hasharoni A, Evron Z, et al. Synovial chondromatosis: the possible role of FGF 9 and FGF receptor 3 in its pathology. *Int J Exp Pathol* 2000;81:183–189.
- Robinson P, White LM, Kandel R, et al. Primary synovial osteochondromatosis of the hip: extracapsular patterns of spread. *Skeletal Radiol* 2004;33:210–215.
- Robinson P, Whitehouse RW, Freemont AJ, Ellis LD. Synovial osteochondromatosis complicating pilon fracture of the tibia. *Skeletal Radiol* 2001;30:475–477.
- Schajowicz F. Synovial chondromatosis. In: *Tumors and tumorlike* lesions of bones and joints New York: Springer-Verlag, 1981:541– 545.
- Sciot R, Dal Cin P, Bellemans J, et al. Synovial chondromatosis: clonal chromosome changes provide further evidence for a neoplastic disorder. *Virchows Arch* 1998;433:189–191.
- Sugimoto K, Iwai M, Kawate K, et al. Tenosynovial osteochondromatosis of the tarsal tunnel. *Skeletal Radiol* 2003;32:99–102.
- Trias A, Quintana O. Synovial chondrometaplasia: review of world literature and study of 18 Canadian cases. *Can J Surg* 1976;19:151–158.
- Tuckman G, Wirth CZ. Synovial osteochondromatosis of the shoulder: MR findings. J Comput Assist Tomogr 1989;13:360–361.
- Wittkop B, Davies AM, Mangham DC. Primary synovial chondromatosis and synovial chondrosarcoma. a pictorial review. *Eur Radiol* 2002;12:2112–2119.

Pigmented Villonodular Synovitis

- Abdelwahab IF, Kenan S, Steiner GC, Abdul-Quader M. True bursal pigmented villonodular synovitis. *Skeletal Radiol* 2002;31: 354–358.
- Abdul-Karim FW, El-Naggar AK, Joyce MJ, et al. Diffuse and localized tenosynovial giant cell tumor and pigmented villonodular synovitis. A clinicopathologic and flow cytometric DNA analysis. *Hum Pathol* 1992;23:729–735.
- Abrahams TG, Pavlov H, Bansal M, Bullough P. Concentric joint space narrowing of the hip associated with hemosiderotic synovitis (HS) including pigmented villonodular synovitis (PVNS). *Skeletal Radiol* 1988;17:37–45.
- Aghasi MK, Robinson D, Reif RM, Halperin N. Pigmented villonodular synovitis of the talus in a child. *Foot Ankle* 1988;9: 139–142.
- Aglietti P, Di Muria GV, Salvati EA, Stringa G. Pigmented villonodular synovitis of the hip joint (review of the literature and report of personal case material). *Ital J Orthop Traumatol* 1983;9: 487–496.
- Atmore WG, Dahlin DC, Ghormley RK. Pigmented villonodular synovitis: a clinical and pathologic study. *Minn Med* 1956;39: 196–202.
- Baker ND, Klein JD, Weidner N, et al. Pigmented villonodular synovitis containing coarse calcifications. *Am J Roentgenol* 1989; 153:1228–1230.

- Balsara ZN, Staiken BF, Martinez AJ. MR image of localized giant cell tumor of the tendon sheath involving the knee. J Comput Assist Tomogr 1989;13:159–162.
- Berger I, Ehemann V, Helmchen B, et al. Comparative analysis of cell populations involved in the proliferative and inflammatory processes in diffuse and localised pigmented villonodular synovitis. *Histol Histopathol* 2004;19:687–692.
- 55. Bertoni F, Unni KK, Beabout JW, Sim FH. Malignant giant cell tumor of the tendon sheaths and joints (malignant pigmented villonodular synovitis). *Am J Surg Pathol* 1997;21:153–163.
- Besette PR, Cooley PA, Johnson RP, Czarnecki DJ. Gadoliniumenhanced MRI of pigmented villonodular synovitis of the knee. *J Comput Assist Tomogr* 1992;16:992–994.
- Bhawan J, Joris I, Cohen N, Majno G. Microcirculatory change in posttraumatic pigmented villonodular synovitis. *Arch Pathol Lab Med* 1980;104:328–332.
- Boyd AD Jr, Sledge CB. Evaluation of the hip with pigmented villonodular synovitis. A case report. *Clin Orthop* 1992;275: 180–186.
- Breimer CW, Freiberger RH. Bone lesions associated with villonodular synovitis. Am J Roentgenol 1958;80:618–629.
- Butt WP, Hardy G, Östlere SJ. Pigmented villonodular synovitis of the knee: computed tomographic appearances. *Skeletal Radiol* 1990;19:191–196.
- Byers PD, Cotton RE, Deacon OW, et al. The diagnosis and treatment of pigmented villonodular synovitis. J Bone Joint Surg Br 1968;50:290–305.
- Cardinal E, Buckwalter K, Braunstein EM, et al. Occult dorsal carpal ganglion: comparison of US and MR imaging. *Radiology* 1994;193:259–262.
- Cavanagh R, Schwamm HA. RPC of the month from AFIP. Radiology 1971;100:409–414.
- Chung EB, Enzinger FM. Fibroma of tendon sheath. Cancer 1979;44:1945–1954.
- Cotten A, Flipo R-M, Chastanet P, et al. Pigmented villonodular synovitis of the hip: review of radiographic features in 58 patients. *Skeletal Radiol* 1995;24:1–6.
- 66. Crawford GP, Offerman RJ. Pigmented villonodular synovitis in the hand. *Hand* 1980;12:282–287.
- Crosby EB, Inglis A, Bullough PG. Multiple joint involvement with pigmented villonodular synovitis. *Radiology* 1977;122: 671–672.
- 68. Dahlen A, Broberg K, Domanski HA, et al. Analysis of the distribution and frequency of trisomy 7 in vivo in synovia from patients with osteoarthritis and pigmented villonodular synovitis. *Cancer Genet Cytogenet* 2001;131:19–24.
- De Beuckeleer L, De Schepper A, De Belder F, et al. Magnetic resonance imaging of localized giant cell tumour of the tendon sheath (MRI of localized GCTTS). *Eur Radiol* 1997;7:198–201.
- Descamps F, Yasik E, Hardy D, et al. Pigmented villonodular synovitis of the hip. A case report and review of the literature. *Clin Rheumatol* 1991;10:184–190.
- 71. de St. Aubain Sommerhausen N, Dal Cin P. Diffuse-type giant cell tumour. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002:112–114.
- 72. de St. Aubain Sommerhausen N, Dal Cin P. Giant cell tumour of tendon sheath. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours. Pathology & genetics. Tumours of soft tissue and bone. Lyon: IARC Press, 2002: 110–111.
- Docken WP. Pigmented villonodular synovitis: a review with illustrative case reports. *Semin Arthritis Rheum* 1979;9:1–22.
- Dorwart RH, Genant HK, Johnston WH, Morris JM. Pigmented villonodular synovitis of synovial joints: clinical, pathologic, and radiologic features. *Am J Roentgenol* 1984;143:877–885.
- Erler K, Demiralp B, Ozdemir MT, et al. Giant cell tumor of tendon sheath simulating giant cell tumor of bone: report of a case. J Surg Orthop Adv 2004;13:124–127.
- Eustace SE, Harrison M, Srinivasen U, Stack J. Magnetic resonance imaging in pigmented villonodular synovitis. *Can Assoc Radiol J* 1994;45:283–286.
- 77. Enzinger FM, Weiss SW. Benign tumors and tumor-like lesions of synovial tissue. In: *Soft tissue tumors*. St. Louis, CV Mosby, 1988:638–658.

- Flandry F, Hughston JC. Pigmented villonodular synovitis. J Bone Joint Surg Am 1987;69:942–949.
- Fletcher AG Jr, Horn RC Jr. Giant-cell tumor of tendon sheath origin: a consideration of bone involvement and report of 2 cases with extensive bone destruction. *Ann Surg* 1951;133: 374–385.
- Fox MG, Kransdorf MJ, Bancroft LW, et al. MR imaging of fibroma of the tendon sheath. *Am J Roentgenol* 2003;180:1449–1453.
- Fraire AE, Fechner RE. Intra-articular localized nodular synovitis of the knee. Arch Pathol 1972;93:473–476.
- Galloway JDB, Broders AC, Ghormley RK. Xanthoma of tendon sheaths and synovial membranes. *Arch Surg* 1940;40: 485–538.
- 83. Gaary E, Gorlin JB, Jaramillo D. Pseudotumor and arthropathy in the knees of a hemophiliac. *Skeletal Radiol* 1996;25:85–87.
- Georgen TG, Resnick D, Niwayama G. Localized nodular synovitis of the knee: a report of two cases with abnormal arthrograms. *Am J Roentgenol* 1976;126:647–650.
- Ghadially FN, Labonde JMA, Dick CE. Ultrastructure of pigmented villonodular synovitis. J Pathol 1979;127:19–26.
- Goldman AB, DiCarlo EF. Pigmented villonodular synovitis. Diagnosis and differential diagnosis. *Radiol Clin North Am* 1988;26: 1327–1347.
- Granowitz SP, D Antonio J, Mankin HL. The pathogenesis and long-term end results of pigmented villonodular synovitis. *Clin Orthop* 1976;114:335–351.
- Granowitz SP, Mankin HJ. Localized pigmented villonodular synovitis of the knee. Report of five cases. J Bone Joint Surg Am 1967;49:122–128.
- Greenfield MM, Wallace KM. Pigmented villonodular synovitis. *Radiology* 1950;54:350–356.
- Grieten M, Buckwalter KA, Cardinal E, Rougraff B. Case report 873. Lipoma arborescens (villous lipomatous proliferation of the synovial membrane). *Skeletal Radiol* 1994;23:652–655.
- Hirohata K. Light microscopic and electron microscopic studies of individual cells in pigmented villonodular synovitis and bursitis. *Kobe J Med Sci* 1968;14:251–279.
- Huang G-S, Lee C-H, Chan WP, et al. Localized nodular synovitis of the knee: MR imaging appearance and clinical correlates in 21 patients. *Am J Roentgenol* 2003;181:539–543.
- Hughes TH, Sartoris DJ, Schweitzer ME, Resnick DL. Pigmented villonodular synovitis: MRI characteristics. *Skeletal Radiol* 1995;24:7–12.
- Jaffe HL, Lichtenstein L, Sutro CJ. Pigmented villonodular synovitis, bursitis and tenosynovitis. Arch Pathol Lab Med 1941;31: 731–765.
- Jelinek JS, Kransdorf MJ, Shmookler BM, et al. Giant cell tumor of the tendon sheath: MR findings in nine cases. *Am J Roentgenol* 1994;162:919–922.
- Jelinek JS, Kransdorf MJ, Utz JA, Hudson-Berry B Jr, et al. Imaging of pigmented villonodular synovitis with emphasis on MR imaging. *Am J Roentgenol* 1989;152:337–342.
- Jergesen HE, Mankin HJ, Schiller AL. Diffuse pigmented villonodular synovitis of the knee mimicking primary bone neoplasms. A report of two cases. J Bone Joint Surg Am 1978;60: 825–829.
- 98. Jones FE, Soule EM, Coventry MB. Fibrous xanthoma of synovium (giant-cell tumor of tendon sheath, pigmented nodular synovitis). A study of 118 cases. *J Bone Joint Surg Am* 1969;51:76–86.
- Kalil ŘK, Unni KK. Malignancy in pigmented villonodular synovitis. Skeletal Radiol 1998;27:392–395.
- Kallas KM, Vaughan L, Haghighi P, Resnick D. Pigmented villonodular synovitis of the hip presenting as a retroperitoneal mass. *Skeletal Radiol* 2001;30:469–474.
- Karasick D, Karasick S. Giant cell tumor of tendon sheath: spectrum of radiologic findings. *Skeletal Radiol* 1992;21:219–224.
- Keenan MG. Computed tomography in pigmented villonodular synovitis of the hip. *J Rheumatol* 1987;14:1181–1183.
- Kerr R. Diffuse pigmented villonodular synovitis. Orthopedics 1989;12:1008–1012.
- 104. Khan S, Neumann CH, Steinbach LS, Harrington KD. MRI of giant cell tumor of tendon sheath of the hand: a report of three cases. *Eur Radiol* 1995;5:467–470.
- Kindblom LG, Gunterberg B. Pigmented villonodular synovitis involving bone. Case report. J Bone Joint Surg Am 1978;60:830–832.

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- Klompmaker J, Veth RPH, Robinson PH, et al. Pigmented villonodular synovitis. Arch Orthop Trauma Surg 1990;109:205–210.
- 107. Kottal AR, Vogler JB, Matamoros A, et al. Pigmented villonodular synovitis: a report of MR imaging in two cases. *Radiology* 1987;163:551–553.
- Kursunoglu-Brahme S, Riccio T, Weisman MH, et al. Rheumatoid knee: role of gadopentetate-enhanced MR imaging. *Radiol*ogy 1990;176:831–835.
- 109. Lin J, Jacobson JA, Jamadar DA, Ellis JH. Pigmented villonodular synovitis and related lesions: The spectrum of imaging findings. *Am J Roentgenol* 1999;172:191–197.
- Llauger J, Palmer J, Rosón N, et al. Pigmented villonodular synovitis and giant cell tumors of the tendon sheath: Radiologic and pathologic features. *Am J Roentgenol* 1999;172:1087–1091.
- 111. Ly JQ, Carlson CL, LaGatta LM, Beall DP. Giant cell tumor of the peroneus tendon sheath. *Am J Roentgenol* 2003;180:1442.
- 112. Mandelbaum BR, Grant TT, Hartzman S, et al. The use of MRI to assist int he diagnosis of pigmented villonodular synovitis of the knee. *Clin Orthop* 1988;231:135–139.
- McMaster PE. Pigmented villonodular synovitis with invasion of bone. Report of six cases. J Bone Joint Surg Am 1960;42:1170– 1183.
- 114. Mertens F, Palsson E, Lindstrand A, et al. Evidence of somatic mutations in osteoarthritis. *Hum Genet* 1996;98:651–656.
- Middleton WD, Patel V, Teefey SA, Boyer MI. Giant cell tumors of the tendon sheath: analysis of sonographic findings. *Am J Roentgenol* 2004;183:337–339.
- 116. Miller WE. Villonodular synovitis: pigmented and nonpigmented variations. *South Med* J 1982;75:1084–1086.
- 117. Mitelman F, Johansson B, Mertens F. Mitelman database of chromosome aberrations in cancer. (2005) Available at: http://cgap.nci.nih.gov/Chromosomes/Mitelman.
- Mulder JD, Kroon HM, Schütte HE, Taconis WK. Radiologic atlas of bone tumors. Amsterdam: Elsevier, 1993.
- 119. Myers BW, Masi AT. Pigmented villonodular synovitis and tenosynovitis, a clinical epidemiologic study of 166 cases and literature review. *Medicine* 1980;59:224–238.
- Narváez JA, Narváez J, Ortega R, et al. Hypointense synovial lesions on T2-weighted images: Differential diagnosis with pathologic correlation. *Am J Roentgenol* 2003;181:761–769.
- Nilsonne U, Moberger G. Pigmented villonodular synovitis of joints: histological and clinical problems in diagnosis. *Acta Orthop Scand* 1983;24:67–70.
- 122. Ohjimi Y, Iwasaki H, Ishiguro M, et al. Short arm of chromosome 1 aberration recurrently found in pigmented villonodular synovitis. *Cancer Genet Cytogenet* 1996;90:80–85.
- Park JS, Ryu KN. Hemophilic pseudotumor involving the musculoskeletal system: Spectrum of radiographic findings. Am J Roentgenol 2004;183:55–61.
- Patel MR, Zinberg EM. Pigmented villonodular synovitis of the wrist invading bone; report of a case. J Hand Surg 1984;9:854– 858.
- 125. Peh WC, Wong Y, Shek TW, Ip WY. Giant cell tumor of the tendon sheath of the hand: a pictorial essay. *Australas Radiol* 2001;45:274–280.
- 126. Poletti SC, Gates HS III, Martinez SM, Richardson WJ. The use of magnetic resonance imaging in the diagnosis of pigmented villonodular synovitis. *Orthopedics* 1990;13:185–190.
- 127. Rao AS, Vigorita VJ. Pigmented villonodular synovitis (giant-cell tumor of the tendon sheath and synovial membrane). A review of eighty-one cases. *J Bone Joint Surg Am* 1984;66:76–94.
- Ray RA, Morton CC, Lipinski KK, et al. Cytogenetic evidence of clonality in a case of pigmented villonodular synovitis. *Cancer* 1991;67:121–125.
- Rosenthal DI, Aronow S, Murray WT. Iron content of pigmented villonodular synovitis detected by computed tomography. *Radiology* 1979;133:409–411.
- 130. Schumacher HR, Lotke P, Athreya B, Rothfuss S. Pigmented villonodular synovitis: light and electron microscopic studies. *Semin Arthritis Rheum* 1982;12:32–43.
- Schwartz GB, Coleman DA. Pigmented villonodular synovitis of the wrist and adjacent bone. Orthop Rev 1986;15:526–530.
- 132. Sciot R, Rosai J, Dal Cin P, et al. Analysis of 35 cases of localized and diffuse tenosynovial giant cell tumor: a report from the Chromosomes and Morphology (CHAMP) study group. *Mod Pathol* 1999;12:576–579.

- Sherry JB, Anderson W. The natural history of pigmented villonodular synovitis of tendon sheath. J Bone Joint Surg Am 1956;37:1005–1011.
- 134. Somerhausen NSA, Fletcher CDM. Diffuse-type giant cell tumor. Clinicopathologic and immunohistochemical analysis of 50 cases with extraarticular disease. *Am J Surg Pathol* 2000;24: 479–492.
- Spritzer CE, Dalinka MK, Kressel HY. Magnetic resonance imaging of pigmented villonodular synovitis: a report of two cases. *Skeletal Radiol* 1987;16:316–319.
- Steinbach LS, Neumann CH, Stoller DW, et al. MRI of the knee in diffuse pigmented villonodular synovitis. *Clin Imag* 1989;13: 305–316.
- 137. Stiehl JB, Hackbarth DA. Recurrent pigmented villonodular synovitis of the hip joint, case report and review of the literature. J Arthroplasty 1991;6:S85–S90.
- 138. Stout AP, Lattes R. Tumors of the soft tissue. In: Atlas of tumor pathology, 2nd series, fascicle 1. Washington DC: Armed Forces Institute of Pathology; 1967.
- Suh J, Griffith HJ, Galloway HR, Everson LI. MRI in the diagnosis of synovial disease. *Orthopedics* 1992;15:778–781.
- 140. Sundaram M, Chalk D, Merenda J, et al. Case report 563. Pigmented villonodular synovitis (PVNS) of knee. *Skeletal Radiol* 1989;18:463–465.
- Sundaram M, McGuire MH, Fletcher J, et al. Magnetic resonance imaging of lesions of synovial origin. *Skeletal Radiol* 1986;15:110–116.
- 142. Ushijima M, Hashimoto H, Tsuneyoshi M, Enjoji M. Giant cell tumor of the tendon sheath (nodular tenosynovitis). A study of 207 cases to compare the large joint group with the common digit group. *Cancer* 1986;57:875–884.
- 143. Villani C, Tucci G, DiMille M, et al. Extra-articular localized nodular synovitis (giant cell tumor of tendon sheath origin) attached to the subtalar joint. *Foot Ankle Int* 1996;17:413–416.
- Wagner ML, Spjut HJ, Dutton RV, et al. Polyarticular pigmented villonodular synovitis. *Am J Roentgenol* 1981;136:821–823.
- 145. Weiss SW, Goldblum JR. *Enzinger and Weiss's soft tissue tumors*, 4th ed. Philadelphia: CV Mosby, 2001.
- 146. Weisz GM, Gal A, Kitchener PN. Magnetic resonance imaging in the diagnosis of aggressive villonodular synovitis. *Clin Orthop* 1988;236:303–306.
- 147. Wu NL, Hsiao PF, Chen BF, Chen HC, Su HY. Malignant giant cell tumor of the tendon sheath. *Int J Dermatol* 2004;43:54–57.
- 148. Young JM, Hudacek AG. Experimental production of pigmented villonodular synovitis in dogs. Am J Pathol 1981;19: 379–392.
- 149. Yudd AP, Velchik MG. Pigmented villonodular synovitis of the hip. *Clin Nucl Med* 1985;10:441–442.
- Zwass A, Abdelwahab IF, Klein MJ. Case report 463. Pigmented villonodular synovitis (PVNS) of the knee. *Skeletal Radiol* 1988; 17:81–84.
- 151. Zwass A, Greenspan A, Green SM. Giant cell tumor of the tendon sheath with a pathologic phalangeal fracture: a rare association. *Bull Hosp Joint Dis Orthop Inst* 1985;45:87–93.

Synovial Hemangioma

- Armstrong SJ, Watt I. Lipoma arborescens of the knee. Br J Radiol 1989;62:178–180.
- Bullough PG. Atlas of orthopaedic pathology with clinical and radiologic correlations, 2nd ed. New York: Gower, 1992.
- 154. Brodsky AE. Synovial hemangioma of the knee joint. Bull Hosp Joint Dis 1956;17:58–69.
- 155. Buetow PC, Kransdorf MJ, Moser RP Jr, et al. Radiologic appearance of intramuscular hemangioma with emphasis on MR imaging. *Am J Roentgenol* 1990;154:563–567.
- 156. Cotten A, Flipo RM, Herbaux B, et al. Synovial haemangioma of the knee: a frequently misdiagnosed lesion. *Skeletal Radiol* 1995;24:257-261.
- 157. Devaney K, Vinh TN, Sweet DE. Synovial hemangioma: report of 20 cases with differential diagnostic considerations. *Hum Pathol* 1993;24:737–745.
- 158. Ehara S, Son M, Tamakawa Y, et al. Fluid-fluid levels in cavernous hemangioma of soft tissue. *Skeletal Radiol* 1994;23: 107–109.
- Enjolras O, Mulliken JB. Vascular tumors and vascular malformations (new issues). Adv Dermatol 1997;13:375–423.

- Hawnaur JM, Whitehouse RW, Jenkins JP, Isherwood I. Musculoskeletal haemangiomas: comparison of MRI with CT. Skeletal Radiol 1990;19:251–258.
- Jacobs JE, Lee FW. Hemangioma of the knee joint. J Bone Joint Surg Am 1949;31:831–836.
- 162. Larson IJ, Landry RN. Hemangioma of the synovial membrane. *J Bone Joint Surg Am* 1969;51:1210–1215.
- Lenchik L, Poznanski AK, Donaldson JS, Sarwark JF. Case report 681. Synovial hemangioma of the knee. *Skeletal Radiol* 1991;20:387–389.
- Levin DC, Gordon DH, McSweeney J. Arteriography of peripheral hemangiomas. *Radiology* 1976;121:625–630.
- Llauger J, Monill JM, Palmer J, Clotet M. Synovial hemangioma of the knee: MRI findings in two cases. *Skeletal Radiol* 1995;24:579–581.
- 166. Madewell JE, Sweet DE. Tumors and tumor-like lesions in or about joints. In: Resnick D, ed. *Bone and joint imaging*. Philadelphia: WB Saunders, 1989:1184.
- 167. Moon NF. Synovial hemangioma of the knee joint. A review of previously reported cases and inclusion of two new cases. *Clin Orthop* 1973;90:183–190.
- Osburn AW, Bassett LW, Seeger LL, et al. Case report 609. Synovial (osteo)chondromatosis. *Skeletal Radiol* 1990;19:237–241.
- 169. Ravin CE, Bergin D, Bisset GS III, et al. The Radiological Society of North American 86th Scientific Assembly and Annual Meeting. Image Interpretation, Session: 2000. *RadioGraphics* 2001;21: 267–287.
- Resnick D, Oliphant M. Hemophilia-like arthropathy of the knee associated with cutaneous and synovial hemangiomas. *Radiology* 1975;114:323–326.
- Silit E, Mutlu H, Pekkafali Z, et al. Synovial hemangioma of the knee invading the femur. *Skeletal Radiol* 2002;31:612–614.
- 172. Suh J-S, Hwang G, Hahn S-B. Soft tissue hemangiomas: MR manifestations in 23 patients. *Skeletal Radiol* 1994;23:621–625.
- 173. Waddell GF. A haemangioma involving tendons. J Bone Joint Surg Br 1967;49:138–141.
- 174. Weitzman G. Lipoma arborescens of the knee. J Bone Joint Surg Am 1965;47:1030–1033.
- 175. Wolfe RD, Giuliano VJ. Double-contrast arthrography in the diagnosis of pigmented villonodular synovitis of the knee. Am J Roentgenol 1970;110:793–799.

Lipoma Arborescens

- 176. Al-Ismail K, Torreggiani WC, Al-Sheikh F, et al. Bilateral lipoma arborescens associated with early osteoarthritis. *Eur Radiol* 2002;12:2799–2802.
- 177. Bejia I, Younes M, Moussa A, et al. Lipoma arborescens affecting multiple joints. *Skeletal Radiol* 2005;34:536–538.
- Cil A, Atay OA, Aydingoz U, et al. Bilateral lipoma arborescens of the knee in a child: a case report. *Knee Surg Sports Traumatol Arthrosc* 2005;13:463–467.
- Dinauer P, Bojescul JA, Kaplan KJ, Litts C. Bilateral lipoma arborescens of the bicipitoradial bursa. *Skeletal Radiol* 2002;31: 661–665.
- Doyle AJ, Miller MV, French JG. Lipoma arborescens in the bicipital bursa of the elbow: MRI findings in two cases. *Skeletal Radiol* 2002;31:656–660.
- Hallel T, Lew S, Bansal M. Villous lipomatous proliferation of synovial membrane (lipoma arborescens). J Bone Joint Surg Am 1988;70:264–270.
- 182. Kloen P, Keel SB, Chandler HP, et al. Lipoma arborescens of the knee. *J Bone Joint Surg Br* 1998;80:298–301.
- Laorr A, Peterfy CG, Tirman PF, Rabassa AE. Lipoma arborescens of the shoulder: magnetic resonance imaging findings. *Can Assoc Radiol J* 1995;46:311–313.
- Martin S, Hernandez L, Romero J, et al. Diagnostic imaging of lipoma arborescens. *Skeletal Radiol* 1988:27:325–329.
- Nisolle J-F, Blouard E, Baudrez V, et al. Subacromial-subdeltoid lipoma arborescens associated with a rotator cuff tear. *Skeletal Radiol* 1999;28:283–285.
- Parsonage S, Mehr A, Davies AM. Lipoma arborescens: a definitive MR imaging diagnosis. Osteologiai Közlemények 2001;9:80–82.
- 187. Soler T, Rodriguez E, Bargiela A, Da Riba M. Lipoma arborescens of the knee: MR characteristics in 13 joints. J Comput Assist Tomogr 1998;22:605–609.

 Vilanova JC, Barcelo J, Villalon M, et al. MR imaging of lipoma arborescens and the associated lesions. *Skeletal Radiol* 2003;32: 504–550.

Synovial Sarcoma

- 189. Ackerman LV. Extra-osseous localized non-neoplastic bone and cartilage formation (so-called myositis ossificans). Clinical and pathological confusion with malignant neoplasms. J Bone Joint Surg Am 1958;40:279–298.
- 190. Ashton BA, Eagleson CC, Bab I, Owen ME. Distribution of fibroblastic colony-forming cells in rabbit bone marrow and assay of their osteogenic potential by an in vitro diffusion chamber method calcification. *Tissue Int* 1984;36:83–86.
- 191. Azouz EM, Vicker DB, Brown KLB. Computed tomography of synovial sarcoma of the foot. J Can Assoc Radiol 1984;35:85–87.
- Blacksin M, Adesokan A, Benevenia J. Case report 871. Synovial sarcoma, monophasic type. *Skeletal Radiol* 1994;23:589–591.
- 193. Cadman NL, Soule EH, Kelly PJ. Synovial sarcoma: an analysis of 134 tumors. *Cancer* 1965;18:613–627.
- 194. Campanacci M. Bone and soft-tissue tumors. New York: Springer-Verlag, 1990:998–1012.
- 195. Chung EB, Enzinger FM. Extraskeletal osteosarcoma. Cancer 1987;60:1132–1142.
- Enterline HT. Histopathology of sarcomas. Semin Oncol 1981;8: 133–155.
- 197. Enzinger FM, Weiss SW. Soft tissue tumors, 3rd ed. St. Louis: CV Mosby, 1995:757–786.
- 198. Ernst M, Froesch ER. Growth hormone dependent stimulation of osteoblast-like cells in serum-free cultures via local synthesis of insulin-like growth factor 1. *Biochem Biophys Res Commun* 1988; 151:142–147.
- 199. Evans HL. Synovial sarcoma: a study of 23 biphasic and 17 probably monophasic examples. *Pathol Annu* 1980;15:309–313.
- 200. Farris KB, Reed RJ. Monophasic, glandular, synovial sarcomas and carcinomas of the soft tissues. *Arch Pathol Lab Med* 1982;106: 129–132.
- 201. Fisher C, de Bruijn DRH, Guerts van Kessel A. Synovial sarcoma. In: Fletcher CDM, Unni KK, Mertens F, eds. World Health Organization classification of tumours Pathology & genetics Tumours of soft tissue and bone. Lyon: IARC Press, 2002:200–204.
- 202. Geirnaerdt MJA, Kroon HM, van der Heul RO, Herfkens HF. Tumoral calcinosis. *Skeletal Radiol* 1995;24:148–151.
- 203. Goldman AB: Myositis ossificans circumscripta: a benign lesion with a malignant differential diagnosis. Am J Roentgenol 1976;126:32–40.
- Greenfield GB, Arrington JA, Kudryk BT. MRI of soft tissue tumors. *Skeletal Radiol* 1993;22:77–84.
- 205. Guillou L, Coindre J, Gallagher G, et al. Detection of the synovial sarcoma translocation t(X;18) (SYT;SSX) in paraffin-embedded tissues using reverse transcriptase-polymerase chain reaction: a reliable and powerful diagnostic tool for pathologists. A molecular analysis of 221 mesenchymal tumors fixed in different fixatives. *Hum Pathol* 2001;32:105–112.
- Horowitz AL, Resnick D, Watson RC. The roentgen features of synovial sarcomas. *Clin Radiol* 1973;24:481–484.
- 207. Ishida T, Iijima T, Moriyama S, et al. Intra-articular calcifying synovial sarcoma mimicking synovial chondromatosis. *Skeletal Radiol* 1996;25:766–769.
- Jones BC, Sundaram M, Kransdorf MJ. Synovial sarcoma: MR imaging findings in 34 patients. Am J Roentgenol 1993;161:827– 830.
- 209. Jotereau FW, LeDouarin NM. The developmental relationship between osteocytes and osteoclasts: a study using the quail-chick nucleus marker in endochondral ossification. *Dev Biol* 1978;63: 253–265.
- 210. Krall RA, Kostinovsky M, Patchefsky AS. Synovial sarcoma: a clinical, pathological, and ultrastructural study of 26 cases supporting the recognition of monophasic variant. *Am J Surg Pathol* 1981;5:137–151.
- Kransdorf MJ, Jelinek JS, Moser RP, et al. Soft-tissue masses: diagnosis using MR imaging. Am J Roentgenol 1989;153:541–547.
- Mahajan H, Lorigan JG, Shirkhoda A. Synovial sarcoma: MR imaging. Magn Reson Imag 1989;7:211–216.
- Majeste RM, Beckman EN. Synovial sarcoma with an overwhelming epithelial component. *Cancer* 1988;61:2527–2531.

518 — Differential Diagnosis in Orthopaedic Oncology

- McCarville MB, Spunt SL, Skapek SX, Pappo AS. Synovial sarcoma in pediatric patients. *Am J Roentgenol* 2002;179:797–801.
- McKinney CD, Mills SE, Fechner RE. Intraarticular synovial sarcoma. Am J Surg Pathol 1992;16:1017–1020.
- Meyer CA, Kransdorf MJ, Moser PP Jr, Jelinek JS. Case report 716. Soft-tissue metastasis in synovial sarcoma. *Skeletal Radiol* 1992;21:128–131.
- 217. Miettinen M, Virtanen I. Synovial sarcoma—a misnomer. Am J Pathol 1984;117:18–25.
- 218. Milchgrub S, Ghandur-Mnaymneh L, Dorfman HD, Albores-Saavedra J. Synovial sarcoma with extensive osteoid and bone formation. *Am J Surg Pathol* 1993;17:357–363.
- Mirra JM, Wang S, Bhuta S. Synovial sarcoma with squamous differentiation of its mesenchymal glandular elements. *Am J Surg Pathol* 1984;8:791–796.
- 220. Morton MJ, Berquist TH, McLeod RA, et al. MR imaging of synovial sarcoma. *Am J Roentgenol* 1991;156:337–340.
- Murphey MD, Sartoris DJ, Quale JL, et al. Musculoskeletal manifestations of chronic renal insufficiency. *RadioGraphics* 1993;13:357–379.
- 222. Nakanishi H, Araki N, Sawai Y, et al. Cystic synovial sarcomas: imaging features with clinical and histopathologic correlation. *Skeletal Radiol* 2003;32:701–707.
- 223. Nuovo MA, Norman A, Chumas J, Ackerman LV. Myositis ossificans with atypical clinical, radiographic, or pathologic findings: a review of 23 cases. *Skeletal Radiol* 1992;21:87–101.
- 224. Sánchez Reyes JM, Alcaraz Mexia M, Quiñonex Tapia D, Aramburu JA. Extensively calcified synovial sarcoma. *Skeletal Radiol* 1997;26:671–673.
- Schajowicz F. Tumors and tumorlike lesions of bone: pathology, radiology, and treatment, 2nd ed. New York: Springer-Verlag, 1994.
- Shapeero LG, Vanel D, Couanet D, et al. Extraskeletal mesenchymal chondrosarcoma. *Radiology* 1993;186:819–826.
- 227. Soule EH. Synovial sarcoma. Am J Surg Pathol 1986;10:78-82.
- Steinbach LS, Johnston JO, Tepper EF, et al. Tumoral calcinosis: radiologic-pathologic correlation. *Skeletal Radiol* 1995;24:573–578.
- Strickland B, Mackenzie DH. Bone involvement in synovial sarcoma. J Faculty Radiol 1959;10:64–72.
- Sundaram M, McLeod RA. MR imaging of tumor and tumorlike lesions of bone and soft tissue. Am J Roentgenol 1990;155:817–824.
- 231. Thaete C, Brett D, Monaghan P, et al. Functional domains of the SYT and SYT-SSX synovial sarcoma translocation proteins and co-localization with the SNF protein BRM in the nucleus. *Hum Bol Genet* 1999;8:585–591.
- van Rijswijk CSP, Hogendoorn PCW, Taminiau AHM, Bloem JL. Synovial sarcoma: dynamic contrast-enhanced MR imaging features. *Skeletal Radiol* 2001;30:25–30.

- Varela-Duran J, Enzinger FM. Calcifying synovial sarcoma. Cancer 1982;50:345–352.
- 234. Witkin GB, Miettinen M, Rosai J. A biphasic tumor of the mediastinum with features of synovial sarcoma. Am J Surg Pathol 1989;13:490–499.
- Wright PH, Sim FH, Soule EH, Taylor WF. Synovial sarcoma. J Bone Joint Surg Am 1982;64:112–122.
- Wilbur JF, Slatopolsky E. Hyperphosphatemia and tumoral calcinosis. Ann Intern Med 1968;68:1044–1049.
- 237. Wu JW, Kahn SJ, Chew FS. Paraspinal synovial sarcoma. Am J Roentgenol 2000;174:410.
- Wu KK, Collon DJ, Guise ER. Extraosseous chondrosarcoma. J Bone Joint Surg Am 1980;62:189–194.

Synovial Chondrosarcoma

- Dahlin DC, Unni KK. Chondrosarcoma. In: Bone tumors General aspects and data on 8542 cases, 4th ed. Springfield, IL: Charles C Thomas, 1986:227–259.
- Dunn EJ, McGavran MH, Nelson P, Greer RB III. Synovial chondrosarcoma. Report of a case. J Bone Joint Surg Am 1974;56: 811–813.
- 241. Hermann G, Klein MJ, Abdelwahab IF, Kenan S. Synovial chondrosarcoma arising in synovial chondromatosis of the right hip. *Skeletal Radiol* 1997;26:366–369.
- 242. Kaiser TE, Ivins JC, Unni KK. Malignant transformation of extra-articular synovial chondromatosis: report of a case. *Skeletal Radiol* 1980;5:223–226.
- Kenan S, Abdelwahab IF, Klein MJ, Lewis MM. Synovial chondrosarcoma secondary to synovial chondromatosis. *Skeletal Radiol* 1993;22:623–626.
- Milgram JW, Addison RG. Synovial osteochondromatosis of the knee. Chondromatous recurrence with possible chondrosarcomatous degeneration. *J Bone Joint Surg Am* 1976;58:264–266.
- 245. Mullins F, Berard CW, Eisenberg SH. Chondrosarcoma following synovial chondromatosis. A case study. *Cancer* 1965;18: 1180–1188.
- 246. Ontell F, Greenspan A. Chondrosarcoma complicating synovial chondromatosis: findings with magnetic resonance imaging. *Can Assoc Radiol J* 1994;45:318–323.
- 247. Perry BE, McQueen DA, Lin JJ. Synovial chondromatosis with malignant degeneration to chondrosarcoma. Report of a case. J Bone Joint Surg Am 1988;70:1259–1261.
- 248. Taconis WK, van der Heul RO, Taminiau AMM. Synovial chondrosarcoma: report of a case and review of the literature. *Skeletal Radiol* 1997;26:682–685.
- Wenger DE, Sundaram M, Unni KK, et al. Acral synovial chondrosarcoma. *Skeletal Radiol* 2002;31:125–129.

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