

**AN ATLAS OF**  
***DROSOPHILA* GENES**  
*SEQUENCES AND*  
*MOLECULAR FEATURES*

**GUSTAVO MARONI**

**An Atlas of *Drosophila* Genes**

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# **An Atlas of *Drosophila* Genes**

Sequences and Molecular Features

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## Preface

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The time is long past when all workers in the field knew the main characteristics of all of the *Drosophila* genes that have been sequenced. My objective in preparing this book was to bring together the available molecular information concerning *Drosophila melanogaster* genes and thereby to make that information more readily accessible.

In Part I of this volume, I describe the main molecular features of genes for which the sequence of the entire transcription unit is available (with a special dispensation for *Ubx*). This sample includes 90 genes, approximately half of all the *Drosophila* genes that fulfill the condition mentioned above and that were listed in the GenBank and EMBO databases early in 1992. In organizing the voluminous data, I have tried to develop a form that would facilitate, and perhaps encourage, a comparative approach for future studies.

Part II includes four chapters that consider different aspects of gene organization as they occur in the *Drosophila* genome. These chapters cover: (1) size correlations among various genetic elements; (2) splicing signals (by S. M. Mount); (3) translation initiation signals (by D. R. Cavener and B. A. Cavener); and (4) codon bias (by P. M. Sharp and A. T. Lloyd). These last three chapters are not restricted to the genes covered in the first part of the book. On the contrary, the authors' analyses cover as much of the available data as possible.

Many people helped me by reviewing individual chapters, pointing out deficiencies and suggesting improvements. Some of these colleagues also made unpublished material available. For such help, I am very grateful to Paul D. Boyer, Carlos V. Cabrera, Sean B. Carroll, Robert S. Cohen, Allan Comer, Victor G. Corces, Winifred W. Doane, Wolfgang Driever, Marshal Edgell, James Fristrom, Eric Fyrberg, Donal A. Hickey, Jay Hirsh, Dan Hultmark, David Ish-Horowicz, Clyde Hutchison, Herbert Jäckle, Allen S. Laughon, Judith A. Lengyel, Michael Levine, John T. Lis, John C. Lucchesi, J. Lawrence March, Elliot M. Meyerowitz, Markus Noll, Christiane Nüsslein-Volhard, Mark Peifer, William H. Petri, Michael Rosbash, Georgette Sass, Lillie L. Searles, Stephen Small, Wayne Steinhauer, Alain Vincent, Gail L. Waring, Pieter Wensink, Theodore R. F. Wright and Ray Wu.

The internal consistency of the material in this book, as well as the clarity of its presentation benefited greatly from the editing of my wife,

Donna Maroni. I am grateful to her for her patience and generosity and for her support.

## Format and Conventions

I have tried to be consistent in presenting equivalent data for different genes using the same format. All chapters in Part I are arranged according to the following plan:

- Product
- Structure
- Function
- Tissue distribution
- Mutant phenotype
- Gene organization and expression
- Developmental pattern
- Promoter

The sections *Tissue distribution* and *Developmental pattern* contain comparable information, except that the former reflects results obtained from studies of the protein product and the latter from studies at the RNA level. In some cases, when a group of genes are considered as part of a cluster or a gene family, there may be other sections within the chapter.

The section *Promoter* includes information on all *cis*-acting regulatory regions.

Some of the conventions I used are the following: *Nomenclature, cytogenetic, and genetic map position* follow *The Genome of Drosophila melanogaster* by Lindsley and Zinn (New York: NY: Academic Press, 1992). The names of proteins are abbreviated by using the same letters of the corresponding gene, capitalized and non-underlined, i.e. ADH for *Adh* and ACT5C for *Act5C*.

## Sequences

All nucleotide sequences are numbered with A at the proposed site of translation initiation as position 1. The position immediately upstream of the initiation ATG is 0. Dots above the sequence mark the decades. Positions in the polypeptide chain obtained by virtual translation are indicated along the right-hand margin in parentheses.

The sequence figures were prepared using programs of the Genetics Computer Group of the University of Wisconsin (Madison, Wisconsin). Most of the sequence data were obtained from the GenBank and EMBL databases and the Accession numbers are given. In some cases, segments with no defined function at the 5' and 3' ends of a published sequence were omitted in the interest of space.

The site of transcription initiation is identified by the first dash of a three-character arrow ( $\dashrightarrow$ ); it should be remembered that the resolution in defining this site experimentally is usually no better than  $\pm 2$  nucleotides.

The Hogness–Goldberg box and the polyadenylation signal are marked with double underlining (-----). If a segment exists that matches the CAAT box sequence (or its reverse complement) 60–100 bp upstream of the transcription initiation site, it is also doubly underlined.

The polyadenylation site is marked by  $|(A)_n$  below the sequence, where  $|$  indicates the last transcribed position or the last nucleotide before a string of

A's? Introns in non-coding regions are delimited by brackets,  $\lrcorner$  and  $\llcorner$  marking

the end of one exon and the beginning of the next. Introns in coding regions can be identified by discontinuities in the amino acid sequence.

Short segments of interest such as promoter and enhancer elements are marked by dashes below the sequence (---). Arrowheads are often used to distinguish a certain sequence from its reverse complement ( $\dashrightarrow = 5'TAA3'$ ,  $\dashleftarrow = 5'TTA3'$ ).

Longer segments are delimited by  $|-|$  below the sequence line, usually with some designation or label between the delimiters or after the second vertical line.

Base substitutions are indicated above the line followed by = followed by the designation of the mutant allele (e.g., A = n11 marks the position where an A for G substitution is found in the *Adh*<sup>n11</sup> allele). Larger rearrangements are delimited by  $|-|$  above the sequence with a label describing the type of mutation (deletion, duplication, etc.).

Amino acid sequences are always the outcome of virtual translation. The initiation ATG is chosen according to the proposal of the original investigators. When confirmation of the amino acid sequence is available from direct protein sequencing, this fact is noted in the “product” section. In most cases, the positions of introns are derived exclusively from the comparison of cDNA and genomic sequences. TATA boxes and polyadenylation signals are indicated according to the proposals of the original investigators; these are usually based on sequence data alone. For other features, transcription initiation and termination sites, regulatory regions, etc., I indicate in the text the methods used to ascertain those features.

### Gene Diagrams

The transcription initiation sites are marked by  $\lrcorner$  for units in which transcription is from left to right, and by  $\llcorner$  for units in which transcription is from right to left. The boxes downstream of these symbols represent exons with the black boxes representing coding regions. The lines between exons represent introns.

### *Sequence Comparison Figures*

In the case of some gene families, a comparison of polypeptide sequences is included to highlight differences or similarities between different members of the family. When the sequence of putatively homologous proteins from distant groups, mammals in particular, were available, a sequence comparison figure is included. The sequence alignments were done with the program *Pileup* of the Genetics Computer Group.

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I

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# I

## The *achaete-scute* Complex: *ac*, *sc*, *lsc*, *ase*

### Chromosomal Location:

<i>ac</i>	X, 1B2-3
<i>sc</i> ; <i>lsc</i>	X, 1B3-4
<i>ase</i>	X, 1B3-4

### Map Position:

1-0.0
1-0.0
1-0.0

### Organization of the Complex

The *achaete-scute* complex is proximal to *yellow* (*y*) in a 90 kb segment that includes eight or nine transcription units; the units have been designated *T1* through *T9* (*T6* corresponds to *y*) (Fig. 1.1). Four of these are thought to be responsible for the *ac-sc* genetic function, the *scute* family. Within the *sc* family the following correspondence has been suggested: *T5* = *ac*; *T4* = *sc*; *T3* = *lsc* and *T8* = *ase*. Each of these four genes is transcribed toward the centromere. (Campuzano et al. 1985; Villares and Cabrera 1987; Alonso and Cabrera 1988; González et al. 1989, and references therein; see Ghysen and Dambly-Chaudière 1988 for a review).

### Products

DNA-binding regulatory proteins of the basic helix-loop-helix (bHLH) type that promote neuroblast differentiation.

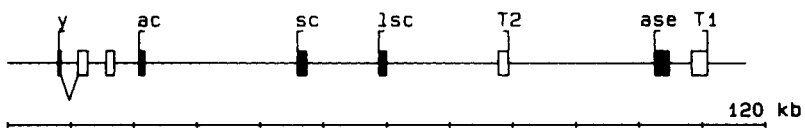


FIG. 1.1. The *ac-sc* complex and *y*. The open box to the left of *ac* corresponds to unidentified embryonic transcripts. *T7* (immediately to the right of *sc*) and *T9* (between *ase* and *T1*) have been omitted; there are conflicting reports on the existence of *T9* (Alonso and Cabrera 1988; González et al. 1989)

### Structure

Sequence comparisons show that, in the region of the HLH domain, the products of *ac*, *sc*, *lsc* and *ase* are similar to each other and to the products of the mammalian oncogene *myc*, the myogenic gene *MyoD*, and the *Drosophila* genes *daughterless* (*da*), *Enhancer of split*, *extramacrochaetae* (*emc*), *hairy* (*h*), and *twist* (Fig. 1.2). In these proteins, the hydrophobic surface of each helix is involved in dimer formation; the amino acids in these regions are particularly well conserved. The basic amino acids in the vicinity of the helices, which effect DNA binding, are also conserved (Villares and Cabrera 1987; Alonso and Cabrera 1988; Murre et al. 1989a, 1989b; Harrison 1991). PEST elements, regions rich in Pro, Glu, Ser and Thr and thought to be important in protein degradation, are common to the various proteins; however, these are not correlated with sequence similarities (González et al. 1989).

Three genes in the complex, *ac*, *sc* and *lsc* share certain sequence elements that distinguish them from *ase*. Particularly noteworthy is the occurrence of a Tyr at the end of a run of acidic amino acids (position 394 in Fig. 1.2; a similar arrangement is found at position 222 of *ase*). A Tyr so associated with acidic residues is reminiscent of a motif found in substrates for protein tyrosine kinases (Villares and Cabrera 1987; Alonso and Cabrera 1988; González et al. 1989).

### Function

Products of the *scute* family are transcriptional activators that promote transcription of genes involved in neuroblast differentiation. They act by binding to regulatory DNA sequences in association with ubiquitous helix-loop-helix proteins such as DA, the product of *da*. *In vitro*, AC, SC and LSC form heterodimers with DA. These complexes bind with high affinity to a DNA segment with the core sequence CANNTG, a sequence that is also found in the immunoglobulin kappa chain enhancer (Murre et al. 1989b), in the *hunchback* (*hb*) zygotic (proximal) promoter and at three positions in the *ac* promoter (Cabrera and Alonso 1991; Van Doren et al. 1991). In yeast cells, LSC/DA heterodimers induce transcription of a reporter gene bearing the *hb* target sequence in its promoter (Cabrera and Alonso 1991).

*ac-sc* function is counteracted by EMC, the product of *emc*. EMC, an HLH protein lacking the basic DNA-binding region, competes with the *ac-sc* products for DA binding. Thus, deficiency of EMC leads to excessive *ac-sc* function and the occurrence of ectopic sensory organs (see below; Ellis et al. 1990; Garrell and Modolell 1990; Van Doren et al. 1991).

All cells that express the LSC protein develop into neuroblasts, but this is not true of all cells in which *lsc* RNA is detected. There seems to be considerable degree of post-transcriptional regulation in that the LSC protein appears significantly later than the corresponding transcript and in a much more restricted subset of cells. Mutations in the neurogenic genes *Notch* and *Delta* (whose normal function is to limit neuronal differentiation to a single cell in a cluster of potential precursors) lead to the presence of LSC in all cells with *lsc*

```

1
1sc .....M TSICSSKF.Q QQHYQLT.NS NIFLLQHQH. ....HHQT QQHQLIAPKI PLGTSQL..Q NMQSQQ... ..SNVGM LSSQKKKfNT
sc MKNNNTTKS TTMSSSVLST NETFPTTINS ATKIFRYQHI MPAPSPILPG GNQNQPAGTM PIKTRKY.TP RGMALTR... ..CSESVSS LSPGSSPAPY
ac .....
ase IRK1RDFGML GAVQSAAST TNT..TPISS QRK....RP LGESQKQNRH NQQNQQLSKT SVPACKCKTN KKLAVERPCK AGTISHPKS QSDQSFQTP.
CON -----S-----S-----S-----A-----S-----S-----

101                                     150                                     200
1sc NNMPYGEQLP SVARRNARER NRVKQVNNGF VNLRQHLPQT VVNSLS.... NGGRGSSKKL SKVDTLRIAV EYIRGLQDML D.....DGT ASSTRHIYN.
sc N...VDQSQ SVQRRNARER NRVKQVNNSF ARLRQHIPS IITDLT...K GGGRGPHKKI SKVDTLRIAV EYIRSLQDLV D.....DLN GGSNIGANNA
ac N.....GP SVIRRNARER NRVKQVNNGF SQLRQHIPAA VIADLSNGRR GIGPGANKKL SKVSTLKMAY EYIRRLQKVL H.....E.....NDQ
ase .GRKGLPLPQ AVARRNARER NRVKQVNNGF ALLREKIPEE VSEAFE..AQ GAGRGASKKL SKVETLRMAV EYIRSLKLL GDFPPLNSQ GNSGSGGDS
CON N-----SV-RRNARER NRVKQVNNGF --LRQHIP-- V---L-----G-GRG--KKL SKV-TLR-AV EYIR-LQ--L -----S-----
--*---* ---*--- Helix I --*---* ---*--- Helix II

201                                     250                                     300
1sc .....SADE SSNDGSSYND YNDS..... LD SSQQ.....F..... LTGAT QSAQSRSYHS
sc VTQLQLCLDE SSSSHSSSST CSSSGHNTTY QNRIS...VS PVQQQQQLQR .....QQ FNHQPLTALS LNTNLVGTSV PGGDA.GCVS
ac QKQKQLHLQ. .... QQHLHFQQ .....QQ .QHQLYAWH QELQL.....
ase FFMKDFEFD C LDEHFDLSL NYEMDEQTV QQLSEMDLN PPOQADLLPS LTLNLGLQYI RIPGTNTYQL LTTDLLGDLS HEQKLEETA SAQLSRSPVP
CON -----D----- --QQ-----Q-----L-----L-----

301                                     350                                     400
1sc A.....S PTPSYSGSEI S.....G GG..... Y IKQELQEQ.. .D.LKFDSFD SFSDEQ... PDDEELL... ..DYISSWQE
sc TSKNQQTCHS PTSSFNSS.M SFDGTYEGV PQQ..... ISTHLDRLDH LDNELHTHSQ LQ.LKFEPYE HFQLDEEDCT PDDEEIL... ..DYISLWQE
ac ...QSPTGS TSSC..NSIS SYCKPATSTI PGA..... TPP..... .NNFHTKLE .....ASFE DYNNSCSCSG TEDEDIL... ..DYISLWQD
ase QKVVRSPCSS PVSPVASTEL LLQTQTCATP LQQQVIKQEY VSTNISSSSN AQTSPQQQQ VQNLGSSPIL PAFYDQEPVS FYDNVVLPGF KKEFSDILQQ
CON -----S-P-S---S---S-----L-----DE--L-----DYIS-WQ-

401                                     439
1sc Q*.....
sc Q*.....
ac DL*.....
ase DQPNNTTAGC LSDESMIDAI DWWEAHAPKS NGACTNLSV
CON -----

```

FIG. 1.2. *sc* family polypeptide sequences. The residues involved in the two helices are underlined, the conserved hydrophobic positions are marked with asterisks. The CON(sensus) sequence indicates positions where at least three of the sequences are identical. Alignment was done using the University of Wisconsin Genetics Computer Group *Gap* program. The first residue shown in this figure corresponds to amino acid 29 in the *ase* sequence.

mRNA and, thus, to the development of ectopic neural derivatives (Cabrera 1990).

SC distinguishes itself from the three other products in this family in that it plays a role in sexual development. This function was indicated by the ability of  $sc^+$  to complement *sisterless b* (*sisb*) mutations (*sisb* is one of the “numerators” used to measure the X:autosome ratio that controls sex determination and dosage compensation early in embryonic development). This prompted the realization that *sc* and *sisb* are one and the same gene. The role of SC in sex determination is likely to involve the formation of a heterodimer between SC and DA. In embryos with two copies of *sc*, enough product is generated to form heterodimers capable of inducing transcription of *sex lethal* (*sxl*) and thus leading to female development. In embryos with only one *sc* copy (males), not enough heterodimer exists to induce *sxl* expression (Parkhurst et al. 1990; Erickson and Cline 1991).

### *Mutant Phenotypes*

Mutations in the complex affect the development of sensory organs and central nervous system: *ac* and *ase* affect different subsets of larval and adult sensory organs while *sc* affects only a subset of adult sensory organs. Amorphic mutations that involve both *ac* and *sc* ( $sc^{10-1}$ ) lead to the absence of all macro- and microchaetae except for those of the wing margin and eye. *lsc* mutations are embryonic lethals that lead to degeneration of the larval peripheral and central nervous systems; chaetae are, however, present. Also, in *lsc* mutant embryos, there is reduced expression of *hb* in neuroblasts (see *Function*). Mutations that increase expression of *ac* and *sc*, such as the dominant gain-of-function allele *Hairy-wing* (*Hw*), are associated with supernumerary chaetae at ectopic sites (Campuzano et al. 1986, and references therein). Amorphic mutations of *ase* cause abnormalities in the development of the adult optic lobes as well as alterations in peripheral neurons and chaetae (González et al. 1989).

### *ac* (*achaete*)

Synonym: *T5*

### **Gene Organization and Expression**

Open reading frame, 201 aa; expected mRNA length, 912 bases. The 5' end was determined by primer extension, RNase protection and sequencing of a cDNA clone; the 3' end was determined from the sequence of the cDNA clone. There are no introns (*ac* Sequence) (Villares and Cabrera 1987).

ac

-939 GAATTCGAAATAATGGGACCTCCTAAATGCTTTCAAATGCTTTCGGCTGAGAGGAACAACTGATACGTTGGGCATAAAGGCCCCGGGG -850

-849 CATTAGAAGTGTTAATAGAAAAGCTCCGGCTGATCAGGTTTCGTTGCAGGACCGAATGGATCGCCGCTGAGGTGTTGATGAGCTGGC -760

-759 CTTGAAAATTCCTACGACTTTGGAGTCGAGCGACAATGGTCTAGTGTAAAGATAATGCCAATGATCCAGGGATCGGAAGGTCATCAG -670

-669 TACATAAAAATAAATAAATTAATGTATTAACATAAAAATTAAGATTTTTTAAAAGCTAAAATACCTAGCCTTGTTAATTAAGATTAT -580

-579 TTTTCGTAACACTTTTGGTAGTGATAAATGTAAATGTCCCATTTTTATAATGTAATGACAGCTATTCCACTAATTTGTTGTA -490

-489 TTTTGTAGTTATAAAAATGGATGGCCACTTTCAATAGGAGATACAGCTTTTACTTCGGAGGTGTTTTACTTGGCTCTGATGCTGG -400

-399 ACCTGTTCCTTTTTAAACCGGTTGGCAGCCGGCAGCGACAGGGCCAGGTTTCGTTTGGGGACGACAGGCAGCTGAAAATGAACAAA -310

----- e3

-309 AACACTCAGAAACTCTCCCACTCGACAACGGGAACACTCAGGTCACCAACAGCTGCGTTTTACAGAGAGAACGAGAGATAATTTACTA -220

----- e2

-219 CCTCTCTATTAATCAGAGAAAACACTCATCTCAAGAGACGATCCTTCAGTGATGATGCTGTTGCACCTTTCCAGGGCAGGTAGGTA -130

-->-62

-129 GTCACGCAGGTGGGATCCCTAGGCCCTGATACCTATAAATAGCCTGAACGGAAACGGGAAGGGCATCAGAACAGAGCCAGCGCTGAAGCA -40

----- e1

-39 AGGAGCATCGTCACACAATAACGTTATACTATCTCTTAAAATGGCTTTGGGCAGCGAAAATCACTCTGTTTTCAACGACGACGAGGAGTC 50  
MetAlaLeuGlySerGluAsnHisSerValPheAsnAspAspGluGluSe (17)

51 ATCTTCGGCCTTTAATGGACCTCTGTTATCCGGAGAAATGCCGGGAACGCACCCGCTAAAGCAGGTCAACAATGGCTTCAGCCAAC 140  
rSerSerAlaPheAsnGlyProSerValIleArgArgAsnAlaArgGluArgAsnArgValLysGlnValAsnAsnGlyPheSerGlnLe (47)

141 ACGACAACATATCCCTCGGGCGTAATAGCCGATTTAAGCAATGGTCGCCGGGAATGGTCCCGGGCCCAATAAAAACCTGAGCAAAGT 230  
uArgGlnHisIleProAlaAlaValIleAlaAspLeuSerAsnGlyArgArgGlyIleGlyProGlyAlaAsnLysLysLeuSerLysVa (77)

231 TAGCACACTGAAAATGGCAGTAGAGTACATACGGCGCTTGCAGAAAGTCTTCATGAAAACGACCAGCAGAAACAGAAAACAGTTGCATTT 320  
lSerThrLeuLysMetAlaValGluTyrIleArgArgLeuGlnLysValLeuHisGluAsnAspGlnGlnLysGlnLysGlnLeuHisLe (107)

||=Hw-1

321 GCAGCAGCAACATTTGCACCTTTCAGCAGCAGCAACAGCATCAACACTTATACGCTGGCACCAAGAGTTGCAGTTGCAATCTCCAACCTGG 410  
uGlnGlnGlnHisLeuHisPheGlnGlnGlnGlnHisGlnHisLeuTyrAlaTrpHisGlnGluLeuGlnLeuGlnSerProThrGl (137)

411 CAGCACAAGTTCCTGCAACAGCATTAGCTCTTATTGCAAGCCAGCAACATCGACGATTCGGGGAGCAACACCTCTAACAAATTTTCATAC 500  
ySerThrSerSerCysAsnSerIleSerSerTyrCysLysProAlaThrSerThrIleProGlyAlaThrProProAsnAsnPheHisTh (167)

501 CAAGTTGGAAGCCAGTTTTGAAGACTACCGTAACAATTCCTGCAGTTCTGGTACTGAAGATGAGGACATCCTCGACTATATCACTCTG 590  
rLysLeuGluAlaSerPheGluAspTyrArgAsnAsnSerCysSerSerGlyThrGluAspGluAspIleLeuAspTyrIleSerLeuTr (197)

591 GCAGGACGACCTGTAATAAACAGATCAAAATCTTCAGCTATTGCTAGTCGACCCCAACCATAAACACATCAAAACATTGATTGGCCAAC 680  
pGlnAspAspLeuEnd (201)

681 AAGTATTACCTCAGCCACAAGTATTTATATTCCTAGAACTACCTTTTTGCCTATAAATTAGTATTTAAGGTTTTATATAGTTTCTAA 770

771 GGATAGTTTCTAATGGAAGACAATTTATATTTAAGTTTTTTTTATAGCATACATTCAGGACATTAACCTGATATATATAAAAATTTTAAA 860

----- } (A)<sub>n</sub>  
(continued)



861	TGAATTTTATTGTAACAAAATTAACGGTAATTAAGTGAAACAATTTATGTACAAAAGGAGTAAAATTCAGAAAAGTTTTAATGAA	950
951	CAAATGCTTTATGAATATGGGCGTAGCAATGTTTTGATACAACTTGATCCTGTCTGTATACCACAGGACACGCTTCTTTTACCTGGT	1040
1041	ACATTCCTTTAAACGATCTAGTATACGCTTTATTCGGGGTAAGCCGAAAAAAGTATTCGAAACTGTAACCGTTAAGTATTACAGATC	1130
1131	ACTAGCCAATGAAGATAAATTACAATAACATTTTGTAAACACTTTTGATCGAAAAACGCCGATTTGCATAAATAAGTTGGATTGAGTAGG	1220
1221	GTGAAAAAGGAAAATATTTACCTGCTGCATTTTGCATATGAACCGGTCAAGGTAATAAGATCCTGAGAATTC	1293

*ac* SEQUENCE. Strain, *Canton S*. Accession M17120 (DROASC1). e1, e2 and e3, AC/DA binding sites (Van Doren et al. 1991). The dominant allele *Hw*<sup>1</sup> is caused by insertion of a *gypsy* element after nucleotide 368; termination occurs within the transposon's terminal repeat, one codon after the insertion (R. Villares and C. V. Cabrera, personal communication).

### *Developmental Pattern*

The expression patterns of *ac* and *sc* and *lsc* are very similar. Before blastoderm formation, expression is uniform throughout the embryo. Later, in early gastrula, transcripts begin to accumulate in stripes restricted to ectodermal cells. During the period of fast germ-band extension (stages 8 and 9), a pattern of two stripes per metamere develops; soon thereafter, when neuroblasts segregate from the ectoderm, transcription is restricted to the neurogenic cells and ceases in epidermal precursors. At the end of stage 9, when neuroblasts begin to divide, transcripts fade (Cabrera et al. 1987).

As development proceeds, expression appears restricted to small clusters of cells that are distributed in a more complex pattern. Even so, the general design outlined above persists: as waves of neuroblast differentiation occur throughout the embryo, transcripts appear immediately before and during the segregation of neuroblasts from the ectoderm; then, the transcripts disappear again, first from the epidermal precursor cells, and finally from the dividing neuroblasts. During germ-band shortening, as differentiation of the neural precursors is completed, expression ceases in the segmented portion of the embryo. After germ-band shortening, expression persists in the primordia for the optic lobes and stomatogastric nervous system (Cabrera et al. 1987). In third instar larvae and early pupae, these genes are expressed in imaginal discs in groups of cells from which the sensory organ mother cells will develop (Romaní et al. 1989). In wing imaginal discs, *ac* and *sc* are expressed with very similar distributions, although mutations affect different sensory organs. Experiments with a reporter gene in transgenic flies indicate that *ac* and *sc* are initially expressed in different clusters of cells; but their products stimulate transcription of each other, so that the ranges of expression soon overlap. As a consequence, in mutants for only one of the two genes, expression of both genes is affected, albeit in different subsets of clusters (Martínez and Modolell 1991).

Differences in expression among the genes are: (1) *ac* stripes are slightly offset from those of *lsc* and *sc*; and (2) during the later stages of expression (stages 10, 11 and 12), transcription of *ac* is more intense than that of *sc* and *lsc*, but *lsc* RNA occurs in more cells.

SC

-659 AAAAAATTTTGATCCTTTTGATAATTAATTGGAGAAATAAGTGAATTTGTTGAACACCTTTAGGGAGCGTACTCCGAATGTCTAATAA -570

-569 GGAGGATCCCAGGATCGGCTGTCGATCCCTTGGATCCGTCGGCGCTAATGAATAGAAGCGTGCGTGAGCTGCACATAAAAATTGGCGATC -480

-479 GCGACTTTTGCTAAGTTAATTAACACAGAAATCAAATTCCTGGCGTGCCGTAGCAAAAAGAGCCCTCACTCAGATACCTTGATCGTTTTT -390

-389 CGATATTTGCGAGTTGATATTTTGAGTTAAAATTTGAGTGTTCCTTTGGACTGTCGAGTGAGAACAGTTTCTGTGGGACTACGAGT -300

-299 ACCTGAGACAGAGAAAGAGAGAGACTACCTGTGGCTCACTCACTTCGAGTTCCTACCTGTGCAGGCAGCTTTGCCGTCCTCTCTC -210

-209 TCTCTCTTCTCTCCGATTCTCTCGCCCGTTTCTCTGCTGAGTGTGTGCAGAGAGTTGCATAAAGGGTACATAACGCGAGGGTTAGG -120

-----

--> --> -116/-111

-119 ACGAAGGGACTCATTCTGTGTAAGGTGCAAAACGATCAAGTCAAGTATTGTACTCTGTTTCATTTATTTTTTCTGTGATCGTTATCC -30

-29 GGAAAGTGAAGAAAGCTCCGAGTGTGTTAATGAAAAACAATAATAACAACGAAAAGCACTACCATGTCATCGAGTGTGCTGCCACC 60  
MetLysAsnAsnAsnAsnThrThrLysSerThrThrMetSerSerSerValLeuSerThr (20)

61 AACGAAACGTTTCCAACGACCATCAATTCGGCAACGAAGATCTTTCGTTATCAGCACATAATGCCAGCCCTAGTCCATTAATTCCCGGT 150  
AsnGluThrPheProThrThrIleAsnSerAlaThrLysIlePheArgTyrGlnHisIleMetProAlaProSerProLeuIleProGly (50)

151 GGCAATCAAATCAACCCGCTGGCAACAATTAAGACTCGCAAGTATACACCAAGGGGTATGGCACTGACCAGATGCTCTGAATCA 240  
GlyAsnGlnAsnGlnProAlaGlyThrMetProIleLysThrArgLysTyrThrProArgGlyMetAlaLeuThrArgCysSerGluSer (80)

241 GTATCATCTCTATCGCCTGGTTCCTCGCCGGCTCCATATAATGTAGACCAATCCCAGTCGGTCCAAGGCCAATGCTAGAGAACAATA 330  
ValSerSerLeuSerProGlySerSerProAlaProTyrAsnValAspGlnSerGlnSerValGlnArgArgAsnAlaArgGluArgAsn (110)

331 CGTGTAACAGCGTGAACAACAGCTTCGCCAGGTGCGGGCAACATATACCACAATCCATAATCACGGATTTGACAAAAGGGTGGTGGTCA 420  
ArgValLysGlnValAsnAsnSerPheAlaArgLeuArgGlnHisIleProGlnSerIleIleThrAspLeuThrLysGlyGlyGlyArg (140)

T=sc-10.1

421 GGACCTCACAAAAGATCTCCAAGTAGACACACTGCGCATTGCCGTCGAGTACATCCGAGCCTCAGGATCTGGTGGATGACCTAAAT 510  
GlyProHisLysLysIleSerLysValAspThrLeuArgIleAlaValGluTyrIleArgSerLeuGlnAspLeuValAspAspLeuAsn (170)

End

511 GGGGGCAGCAATATTGGTGCCAACAATGCAGTCACCCAGCTTCAACTTTGTTGGATGAGTCCAGCAGTACAGTTCGAGCAGCAGTACT 600  
GlyGlySerAsnIleGlyAlaAsnAsnAlaValThrGlnLeuGlnLeuCysLeuAspGluSerSerSerHisSerSerSerSerSerThr (200)

601 TGCAGTTCCTCAGGGCATAATACCTACTATCAAAACAGGATCTCTGTCAGTCTGTGCAACAACAGCAGCAGCTACAGAGGCAGCAGTTC 690  
CysSerSerSerGlyHisAsnThrTyrTyrGlnAsnArgIleSerValSerProValGlnGlnGlnGlnGlnLeuGlnArgGlnGlnPhe (230)

691 AATCACAACCGCTGACAGCGCTCTCATTAAATACCAACTTGGTGGGCACATCCGTACCAGGTGGAGATGCAGGATGCGTATCCACCAGC 780  
AsnHisGlnProLeuThrAlaLeuSerLeuAsnThrAsnLeuValGlyThrSerValProGlyGlyAspAlaGlyCysValSerThrSer (260)

781 AAAAACAGCAAACCTGCCACTCGCCAACATCATTCATCAACTCCAGCATGTCTTTGATTACGGCACCTACGAAGGAGTTCCCAAACA 870  
LysAsnGlnGlnThrCysHisSerProThrSerSerPheAsnSerSerMetSerPheAspSerGlyThrTyrGluGlyValProGlnGln (290)

}}=Hw-Ua

871 ATATCCACCCACCTGGATCGTCTGGATCATCTGGACAACGAATTACACAGCACTCCCAACTTCAGTAAAATTTGAACCGTACGAACAT 960  
IleSerThrHisLeuAspArgLeuAspHisLeuAspAsnGluLeuHisThrHisSerGlnLeuGlnLeuLysPheGluProTyrGluHis (320)

961 TTTCAATTAGACAGGAGGACTGCACCCCGACGACGAGGAGATTTGGACTACATCTCTCTATGGCAGGAGCAGTACTTAATCCCAA 1050  
PheGlnLeuAspGluGluAspCysThrProAspAspGluGluIleLeuAspTyrIleSerLeuTrpGlnGluGlnEnd (345)

(continued)

1051	AATTTACCACCACGCCCTATTTTCTTAGTCAATGTTGAGTTGAACCAAGTGCCTCAAATTGTAATAACACTAATACAAAAACAACAT	1140
1141	ACCCCAATTTTTTTTCTTACTTTAAGCTATTTTTTACATTGTTAAGAACCACGAGACCAGTTTCAAATTTATATATTTATGAAATAA	1230
1231	CTATAGCATGGAAACGAAAACATATTTTTTGGCTAATACAATTTTATGTTAATTAGTTTTGGTGGAAAAATAAAATGAAAAATTAAC	1320
	-----	
1321	GAAAAATAATATTTAAGTTTTTTGTACAAAGGGGATCCATCTATTGCATCAGGTTTGTA AACATTCGGGTACTACTTGCATTGCCTTG  (A) <sub>n</sub>	1410
1411	CAGTGCCGATGGGACCATGTGCAGCCGTTATGTACATTGGTTGCTTTGCATTGGTTTTCCA	1471

*sc* SEQUENCE. Strain, *Canton S*. Accession M17119 (DROASC2). The base substitution at 487 in the null allele *sc*<sup>10-1</sup> is indicated; this mutation also involves a breakpoint that inactivates *ac*. The dominant allele *Hw*<sup>Ua</sup> is caused by insertion of a *copia* element after nucleotide 899; termination occurs within the transposon's terminal repeat, 21 codons after the insertion (R. Villares and C. V. Cabrera, personal communication).

### Promoter

A segment of 0.9 kb upstream of the transcription initiation site is sufficient for nearly normal expression of *ac* (Ruiz-Gómez and Modollel 1987). Within that segment, there are three binding sites apparently responsible for autocatalysis: binding of heterodimers of *ac-sc* and *da* products has been detected at three copies of the element CANNTG (sites marked e in the *ac* sequence at -327, -259 and -123). This binding is blocked by the simultaneous presence of EMC (Van Doren et al. 1991).

### *sc* (*scute*)

Synonyms: *T4* and *sisb*

### Gene Organization and Expression

Open reading frame, 345 aa; expected mRNA length, 1,437 and 1,432 bases. The 5' ends were determined by primer extension; sequencing of a cDNA clone provided the 3' end. There are no introns. Translation might initiate at any of five in-frame AUGs in the mRNA. In the *sc* Sequence, translation is depicted as starting at the first of those ATGs, but the best fit to the initiation of translation consensus is next to the fifth ATG (Villares and Cabrera 1987).

### Developmental Pattern (see *ac*)

Product from the blastoderm period of *sc* expression is probably associated with the *sisb* function.

## lsc

-302	CTGAGTAGGAATAGAGGCCACCCACCACAGAAAAGAACCCTAGAAAAGAGAGAAAATGTACGATCACTTGTGCAAAGGACTTAGGTCC	-213
-212	CGGTTTTTCGAGGGCAGGTAGCCAGGATCCGACCCCGTACCAACCCCTGTAGCTCCTCTGCCGAAGTCGCTGCCTCTGTGCGCGCGCCTT	-123
-122	TCCCTCTGCCACTGGCCGGGTATTTAAAGCCCTAGATCAGAACAGCAATTATCATTGCGGAATCTGATTCCACACAGTCAACATCTGTAA	-33
	!-26	
-32	ACTAAATCTTAGAAAACCTCACAAGGATTACCATGACGAGCATTGTGCAGCAGCAATTCAGCAGCAGCATTACCAGCTGACCAACAGT	57
	MetThrSerIleCysSerSerLysPheGlnGlnGlnHisTyrGlnLeuThrAsnSer	(19)
58	AACATTTCTTGTGCAACATCAGCATCACCATCAAACGCAGCAGCACCAGTTGATTGTCTCCGAAAATACCTTTGGGTACCAGCCAACCTG	147
	AsnIlePheLeuLeuGlnHisGlnHisHisHisGlnThrGlnGlnHisGlnLeuIleAlaProLysIleProLeuGlyThrSerGlnLeu	(49)
148	CAGAATATGCAGCAGAGTCAACAGTCCAATGTTGGACCCATGTTGTCTCCAGAGAAGAAGTTCAACTACAATAACATGCCCTATGGC	237
	GlnAsnMetGlnGlnSerGlnGlnSerAsnValGlyProMetLeuSerSerGlnLysLysLysPheAsnTyrAsnAsnMetProTyrGly	(79)
238	GAGCAATTGCCATCGGTAGCCAGACGAAATGCCCGTGAACGCAATCGCGTGAAGCAGGTGAACAATGGATTTCGTCATCTCCGCCAGCAT	327
	GluGlnLeuProSerValAlaArgArgAsnAlaArgGluArgAsnArgValLysGlnValAsnAsnGlyPheValAsnLeuArgGlnHis	(109)
328	TTGCCTCAAACCTGTGGTAAACTCGCTGTCCAATGGAGGACGTGGTAGCAGCAAGAAGTTATCCAAGTGGACACACTCGGAATCGCCGTT	417
	LeuProGlnThrValValAsnSerLeuSerAsnGlyGlyArgGlySerSerLysLysLeuSerLysValAspThrLeuArgIleAlaVal	(139)
418	GAATATATTCGAGGACTACAGGACATGCTTGATGATGGCACTGCTTCATCAACTCGTCACATCTACAATTCGCCGATGAAAGTAGCAAC	507
	GluTyrIleArgGlyLeuGlnAspMetLeuAspAspGlyThrAlaSerSerThrArgHisIleTyrAsnSerAlaAspGluSerSerAsn	(169)
508	GATGGCAGCAGCTATAACGATTACAACGATAGTTGGACAGTTCGCAACAGTCTTTCGACGGGAGCCACCCAGTCTGCCAATCCCGCTCG	597
	AspGlySerSerTyrAsnAspTyrAsnAspSerLeuAspSerSerGlnGlnPheLeuThrGlyAlaThrGlnSerAlaGlnSerArgSer	(199)
598	TACCACTCCGCTCGCCACGCGCTGACTCCGGATCCGAGATTTCCGGAGGTGGCTATATCAAACAGGAACACAAGAGCAGGACCTC	687
	TyrHisSerAlaSerProThrProSerTyrSerGlySerGluIleSerGlyGlyGlyTyrIleLysGlnGluLeuGlnGluGlnAspLeu	(229)
688	AAATTCGACTCCTTTGATAGCTTCAGTGACGAGCAGCCAGATGACGAGGAGCTACTCGATTATTTTCATCTTGGCAAGAGCAGTGAAGG	777
	LysPheAspSerPheAspSerPheSerAspGluGlnProAspAspGluGluLeuLeuAspTyrIleSerSerTrpGlnGluGlnEnd	(257)
778	GGTCTTACTAAAAGTCCCAAAACAAACAAATATTGTACAACCTGTAATAACCTAAATGTTGCCCTAGTGAGTGTAAAACCAAGTCTCAA	867
868	ATTTACATTAGCCTCTAAGTTACCCCATATTTTTTTTTATTATATTTTAAACGCAATGGAAGACAATGATAGAAAACCACATATTTTTTT	957
958	CATAGTTATAAGTTTGTATAAGCATGGAAGACTAAACTAACTACTTTTAAAGCCAAAATAAAAACATATTGATAAAATTAATTCCAA	1047
1048	TGTTTTTACTGAAATCACTTACTCGTAAATATATTCAGATCGTCATGTAGGGTAATTACAACGAGTTCTCGTTCTCATACCAGCATCAG	1137
1138	AGCCAAAAAGTTTTTAAACAATCTGCATTTTGAAGCATTGCTTGTACTATATATATGTTATGATATTCGTTTTTAAATTTATGATTTTT	1227
	(A) <sub>n</sub>	
1228	ATATTATTATTATTTTTTGTAGCTTAGCTGTTTTGGCCCTCAGGCTTAATAATGGTACTAGCGATAGAAAATAAATTTCCAAAAAAGT	1317
1318	TACCAATTTATTATTTATATTCAATTAATTTTGGAGCGTGGACATGACTCACTCAAAATTCGTAACCAACATAGAGTTAAGCACCTGAC	1407
1408	AGGAAACAACAGCGAATATTTTCATGATTGGTTCCTAACGAGCTACAATTCGGCCGGGAATTGTTAATGGCGGTAAATAGCCCGGAAA	1497
1498	TAGGCAGTCACGCTGAGAGGATGAAATGTCTTAGTCCAAGG	1540

*lsc* SEQUENCE. Strain, *Canton S*. Accession, X12549, Y00846 (DROASCA).  
The exclamation mark at -26 indicates the 5' end of a cDNA.

*Promoter*

An *sc* construction with approximately 1 kb of DNA upstream of the transcribed region and 3 kb downstream is sufficient to provide *sisb* function but not *sc* function (Erickson and Cline 1991). The *cis*-acting regulatory region of *sc* is likely to extend for tens of kilobases.

*lsc*  
(*lethal of scute*)

Synonym: *T3*

**Gene Organization and Expression**

Open reading frame, 257 amino acids; expected mRNA length, ca. 1,184 bases. A cDNA sequence and low resolution S1 mapping were used to define the 5' and 3' ends. There are no introns (*lsc* sequence) (Alonso and Cabrera 1988).

*Developmental Pattern*

*lsc* expression follows the general pattern of *ac* and *sc* expression (see *ac*) except that the expression of *lsc* seems to be more extensive than that of the other two genes and persists longer in both epidermal precursors and neuroblasts (Cabrera et al. 1987).

*ase*  
(*asense*)

Synonym: *T8*

**Gene Organization and Expression**

Open reading frame, 486 amino acids; expected mRNA, 2,263 bases. The 5' end was defined by primer extension and the 3' end by a cDNA sequence. There are no introns (*ase* sequence) (Alonso and Cabrera 1988, partial sequence; González et al. 1989).

ase

-1942 GGATCCAGTATGTTCCACGCTAGCGTCAATTCGGTTTACTCATCTGTTTCATTACCAATTTGGCGTTTCTCTCGTCAAAGATATTTTCC -1853

-1852 CATTGAAATCAATGCGTTTTTAAAATGCAAAATAAACAGAAACCAGAAACCATTATAAATTTGATTTGCCTAGATTGGAACATTTTCGAT -1763

-1762 CCGCCAAAGATAACAGCCAAAAAATATATAAAAAAAGAGTGACAGGACTACAACGCAGGTTCTTAATTTTACGAACGTGGGAGTAAAT -1673

-1672 TCGAAAAGGTATGCCCGCTTTGGGGCAAACCTTTTTGAAACCGTTTACATGTAATATTTTTGGAATCGCTACTTTTATGTATGGTTAA -1583

-1582 TTTAATTATGAACATTTTTCTTGCAGTGCACGAAAGGCGTGGCTGGGGCAAGGAACAGTTCCTTGAGATGAGTGCTGCGATCGCTCGTCA -1493

-1492 AAGTGGGACGCAACCGAGTCAAATCCTCTAGGACAACAAAGGACGCCGAGCAGTACTTCCCAGTACTCAAATACTCCTCAGTACGCACAA -1403

-1402 GCGTTGACTCCTTTTTCTTGTAGAGCTCGTCTGCATAATGAGGAATGAGGACGTGGCATCTGGATCAAAAACCGGTAGTCGGTCTGCGC -1313

-1312 AAGTTTCTTCTCCCGCCGGTTATCCTGCGCTCAAGTCTTTTTCTTGAACCTTTTAAAGTGAACCAAGTTTATAAATGTGCAGCAAGT -1223

-1222 ACAACACACACACATATGTATACTCCTCTATTTACTCAGTGTGTTGGAAACTACCTGAGAGGAAGGATCAAAAAGTTTATTGTAGCAC -1133

-1132 TCGAAAACCTGTTTGCACAGATAGACCTAAGCTCCAAAAAAGAATAGATTATAGAAATAAAGCACTGGTAAAATTTTTCCACCAGG -1043

-1042 ACTAATGCCAGAAATTTTTGCCAGACGTAGAAAAACAAGAATGTAGAGAAGGATGGGTGATTTCTACCCCTTAAAGGATTTAAAT -953

-952 GGCTCTCTGGCATCTTGTCAATTTCCAACATAAATTTAGCCCTGTGAATACCTAAGACATTACTTTCGCAGTATATACTTGTCTGTTTA -863

-862 TTAGCTTAACAATAGGAAAATGTTTTGCCAATGCAAGTCTGGTAAATATATGTATATATTATCCAGTACGAGTTTTTGAAAAGTTAAC -773

-772 AATAGGTGATCCCGAGACATTTTCGAATGAAGTAGAAACCAGTCTTGGTTTTAGCTATAAGCTAAAAATAAGATTTCGATGCATTTCT -683

-682 GCGTTTTACATGACGAATATTGGAGGCTAAGGTGATCTATTAGGATATTTGCAAACTCCTAGGTGGTAGGGCATTCTTGAAAACCG -593

-592 GGCTGAAAAGCTCCCGGGAATAACTTTTTATTATATGCATACGTATATGTTTATTATAATGTCCATATATTAATAGGGCGGTATA -503

-->-455

-502 TAAGCATAATGTTGTGCTGCCGATAAATAATGAGAGAGCGTGGCGAAATCCACCCCTGAACCCAGGTGGACTTTTGGCTCGAGTTGA -413

-----

-412 TCAGATGTTAGTTTTCCCAAAAGCGTACTGTATATAATATATATATATGATAAAGGTGTATGTGTGAGCGATC6AGAAGGACACCCAG -323

-322 TCTAGGGGATGAAAAGTCAGGCCCTTACATAAGGGATACGCAGGACCTCAAATGCCTTCTGTTTTGTATGTGTGTCTGTGTGTGT -233

-232 ATGTCAGTCAACGAAGTCACTTCCGTTGGGTTTGCCTTTAGTTTGAGTTCGGAGTTTAGGGGCACCGACACAGAGCGCCAGCAGCTGT -143

-142 CCTGATGCAAGGACACGGAAACCATATTACATCAGTACCAGTTAACATTCCTCAAGAAGGACTAACTTGCTAAAAGTACACCCGCAAT -53

-52 CGCCACCAGTTTTCTCCCGCCCTCAAAAAGCCACGAATCAAAAACCTTAATTTATGCGCCCTTAAGCTTCAGCCCATCACCTCTCCAA 37  
MetAlaAlaLeuSerPheSerProSerProProProl (13)

38 AAGAAAACCCCAAGGAAAACCCCAATCCAGGAATAAAAACCAGTGAACCTTTTGGAAAGATTACCGTTCACAATGTTTAAAGTGAGA 127  
ysGluAsnProLysGluAsnProAsnProGlyIleLysThrThrLeuLysProPheGlyLysIleThrValHisAsnValLeuSerGluS (43)

128 GTGGCCCAACGCCTTGCAACAGCATATAGCCAATCAGAACCACCATTTCCGAAAGATCCGGAGCTTTGGCATGCTGGGCGCTGTCAA 217  
erGlyAlaAsnAlaLeuGlnGlnHisIleAlaAsnGlnAsnThrIleIleArgLysIleArgAspPheGlyMetLeuGlyAlaValGlnS (73)

(continued)

218	GTGCCGACGACCACTAACACCACCCATATCCAGTCAACGGAAGAGGCCCTGGGGAATCCCAAAGCAGAACCAGCACCAACC erAlaA1aA1aSerThrThrAsnThrThrProIleSerSerGlnArgLysArgProLeuGlyGluSerGlnLysGlnAsnArgHisAsnG	307 (103)
308	AGCAGAATCAACAGCTTAGTAAAACATCAGTGCCTGCTAAAAATGCAAGACCAACAAGAAGTTGGCGGTTGAAAGCCCCAAAAGCAG 1nGlnAsnGlnGlnLeuSerLysThrSerValProAlaLysLysCysLysThrAsnLysLysLeuAlaValGluArgProProLysAlaG	397 (133)
398	GAACTATAAGCCACCCTCATAAAAGCCAAAGCGATCAGAGTTTTGGGACTCCTGGAAGAAAGGGTTTGCCTTTGCCACAAGCCGTTGCC lyThrIleSerHisProHisLysSerGlnSerAspGlnSerPheGlyThrProGlyArgLysGlyLeuProLeuProGlnAlaValAlaA	487 (163)
488	GTAGAAACGCTAGGGAAAGAAATCGCGTGAAGCAGGTTAAACATGGATTGCTTTACTCCGGGAGAAGATCCAGAAGAAGTATCTGAGG rgArgAsnAlaArgGluArgAsnArgValLysGlnValAsnAsnGlyPheAlaLeuLeuArgGluLysIleProGluGluValSerGluA	577 (193)
578	CTTTTGAGGCCACGGGGCGGTAGAGGAGCAAGCAAGAGCTATCAAAGTGGAGACCCCTCCGATGGCCGTAGAGTACATAAGAAGTT 1aPheGluAlaGlnGlyAlaGlyArgGlyAlaSerLysLysLeuSerLysValGluThrLeuArgMetAlaValGluTyrIleArgSerL	667 (223)
668	TGAAAAACTGCTGGGATTTGATTTTCCACCTCTCAACAGTCAGGGGAATAGTTCTGGTTCGGCGATGATAGCTTTATGTTTATTAAAG euGluLysLeuLeuGlyPheAspPheProProLeuAsnSerGlnGlyAsnSerSerGlySerGlyAspAspSerPheMetPheIleLysA	757 (253)
758	ACGAATTCGATTGTCTGGATGAACATTTTCGACGACTCGCTGAGCAACTACGAAATGGATGAGCAACAGACTGTCCAACAACTTTATCCG spGluPheAspCysLeuAspGluHisPheAspAspSerLeuSerAsnTyrGluMetAspGluGlnGlnThrValGlnGlnThrLeuSerG	847 (283)
848	AGGATATGCTAAACCTCCGCAAGCCAGTGATCTCTGCCTAGTTTGACTACATTAATGGGTGCAATACATCAGAATACCAGGAACCA 1uAspMetLeuAsnProProGlnAlaSerAspLeuLeuProSerLeuThrThrLeuAsnGlyLeuGlnTyrIleArgIleProGlyThrA	937 (313)
938	ACACCTACCAACTGCTGACGACTGACTTATTGGGCGATTGAGTCACGAGCAAAAACCTGGAAGAAACAGCTGCTTCGGGCCAGTTATCGC snThrTyrGlnLeuLeuThrThrAspLeuLeuGlyAspLeuSerHisGluGlnLysLeuGluGluThrAlaAlaSerGlyGlnLeuSerA	1027 (343)
1028	GATCGCCGTGCCACAAAAGTGGTAAGAAGTCCCTGCTCTTCTCCAGTTTACCTGTGCCTCGACTGAATTGCTGTTACAGACACAGA rgSerProValProGlnLysValValArgSerProCysSerSerProValSerProValAlaSerThrGluLeuLeuLeuGlnThrGlnT	1117 (373)
1118	CGTGTGCCACACCGCTGCAACAGCAAGTAATCAAACAGGAATACGTCAGTACCAACATTAGCAGCAGCAGCAACGCACAGACTTCCCCGC hrCysAlaThrProLeuGlnGlnGlnValIleLysGlnGluTyrValSerThrAsnIleSerSerSerSerAsnAlaGlnThrSerProG	1207 (403)
1208	AGCAGCAGCAGCAAGTTCAGAACCTGGGATCGTCGCCTATTTTACC CGCTTCTACGACCAGGAGCCCGTGAGCTTCTACGACAACGTAG 1nGlnGlnGlnGlnValGlnAsnLeuGlySerSerProIleLeuProAlaPheTyrAspGlnGluProValSerPheTyrAspAsnValV	1297 (433)
1298	TCCTTCCGGATTCAAGAAGGAATTCAGCGATATTTTGCAGCAAGATCAGCCCAACAATACAACCGCTGGCTGCCCTTCGGACGAGAGCA alLeuProGlyPheLysLysGluPheSerAspIleLeuGlnGlnAspGlnProAsnAsnThrThrAlaGlyCysLeuSerAspGluSerM	1387 (463)
1388	TGATCGATGCCATTGACTGGTGGGAGGCACATGCACCTAAATCTAATGGTGCATGCACCAATCTGTCCGTTTAGCCGAATTTTTTCACAT etIleAspAlaIleAspTrpTrpGluAlaHisAlaProLysSerAsnGlyAlaCysThrAsnLeuSerValEnd	1477 (486)
1478	CACGCATCTCGGAAAAGCCGATTGCATTTTTTGGCATACTTTTTAAATGATTTTTAAATCCTCACAGCATAAGTCTGTGGCAGGCCATTCT	1567
1568	ATCTAAAGTTTTTTTTAATCAAGCCATGACTGAGTCATTGTGTAATATCAATTTAAGCCGAGAAAGGAGGATAAATCTCGCCAGCCGAA	1657
1658	GCTTATATACCTTTGCTGTTAAAACCATGTATTTAATATGAAAGTTCGCACAATTTTCGATGAAGTTTATCACAATTTACGATTTTCATCA	1747
1748	AGATTTGTATATTCTCCAATTTCTAAAAATATATGTACATTTTTGATCTTGTCTATGGTACTTGTACGTATGATATTGTTGATCGATCC	1837
	-----   (A) <sub>n</sub>	
1838	TGCCCGAGTCACCTTTTATATCACCAGACATGCCGATCATGAATTTTATTGATGTGGGGCTGGAA	1903

ase SEQUENCE. Strain, *Canton S*. Accession X51532 (DROASE).

## Developmental Pattern

The pattern of *ase* expression is very different from that of the other three genes in the *sc* family. Expression does not initiate until the extending germ-band stage (late stage 8 embryos); then *ase* transcripts occur in neuroblasts after they have segregated from the ectoderm. After germ-band retraction (stage 13), expression in the segmented region of the embryo ceases, but *ase* transcripts persist in the presumptive optic and procephalic lobes (Alonso and Cabrera 1988; González et al. 1989). Expression of *ase* is also evident in late third instar larvae, occurring throughout the central nervous system in many of its actively proliferating cells (González et al. 1989).

## References

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# 2

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## The Actin Genes: *Act5C*, *Act42A*, *Act57B*, *Act79B*, *Act87E*, *Act88F*

### Chromosomal Location:

<i>Act5C</i>	X, 5C3-4
<i>Act42A</i>	2R, 42A
<i>Act57B</i>	2R, 57B
<i>Act79B</i>	3L, 79B
<i>Act87E</i>	3R, 87E9-12
<i>Act88F</i>	3R, 88F

### Map Position:

1-[14]
2-[55.4]
2-[97]
3-[47.5]
3-[52.3]
3-57.1

### Products

Actins, cytoskeletal and contractile proteins.

### Structure

There is great similarity between *Drosophila* and mammalian actin amino acid sequences. Vertebrates have two distinct families of actins, one family as cytoplasmic filaments and the other occurring in muscle fibers. All *Drosophila* actins are more similar to vertebrate cytoskeletal actins than to muscle actins, but *Act5C* and *Act42A* are especially so (Fig. 2.1) (Fyrberg et al. 1981; Sanchez et al. 1983).

### Tissue Distribution and Function

*Act5C* and *Act42A* encode cytoplasmic actins present in all tissues; *Act57B* and *Act87E* encode larval and adult intersegmental muscle actins; *Act79B* encodes thoracic and leg muscle actins and *Act88F* flight muscle actin (Fyrberg et al. 1983; Sanchez et al. 1983; see also Fyrberg et al. 1991 and Sparrow et al. 1991).

Act88F	C	DD	AG	I	MC	S	T	I	I	
Act79B	C	EE	AS	V	MC	C	S	I	V	
Act57B	C	DE	VA	V	MC	S	T	I	I	
Act87E	C	DE	VA	V	MC	S	T	I	I	
Act5C	C	EE	VA	V	MC	S	T	V	I	
Act42A	C	EE	VA	V	MC	S	T	V	I	
Muscyt	D	XX	IA	V	MC	S	T	V	I	
Musmus	C	EDET	C	LV		S	T	I	I	
CON	M-D-----AL	V-DNGSG--K	AGFAGDDAPR	AVFPSIVGRP	RHQGVVMVGMG	QKD-YVGDEA	QSKRGIL-LK	YPIEHGI-TN	WDDMEK-WHH	TFYNELRVAP
	1					50				100

Act88F	V			S		L	S T	F	L	D
Act79B	V			S		L	S T	Y	L	H
Act57B	V			S		L	S T	Y	L	D
Act87E	V			A		L	S T	Y	L	D
Act5C	V			T		L	S T	Y	L	D
Act42A	V			T		L	S T	Y	L	D
Muscyt	V			T		M	T T	Y	L	D
Musmus	T			V		L	T N	Y	M	D
CON	EEHP-LLTEA	PLNPKANREK	MTQIMFETFN	-PAMYVAIQA	VLSLYASGRT	TGIV-DSGDG	V-H-VPIYEG	-ALPHAI-RL	DLAGR-LTDY	LMKILTERGY
	101					150				200

Act88F	T T	T	D	A T	C	A Q L	SC I	V YN	V	S
Act79B	S T	I	Q	A T	T	A Q L	SC I	V YQ	V	N
Act57B	S T	I	Q	A T	C	S Q L	SC I	V YN	V	I
Act87E	S T	I	Q	A T	C	S Q L	SC I	V YN	V	I
Act5C	S T	I	Q	S S	C	A H L	SC I	T YN	V	T
Act42A	S T	I	Q	S S	C	S Q L	AC L	T YN	V	T
Muscyt	S T	I	Q	S S	C	A Q L	SC I	T FN	V	T
Musmus	S V	I	N	S S	C	T Q I	SA I	T YN	I	N
CON	-F-TTAEREI VRD-KEKLCY VALDFE-EMA TAA-S-SLEK SYELPDGQVI TIGNERFR-P E-LF-PSF-G ME--G-HET- --SIMKCD-D IRKDLNAN-V									
	201				250					300
Act88F	L	T	I	I	L	L	IS Q	S S	*	
Act79B	L	A	M	I	S	L	IS Q	S G	*	
Act57B	M	S	I	I	S	L	IS E	S G	*	
Act87E	M	A	I	I	S	L	IS Q	S G	*	
Act5C	L	A	M	I	S	S	TS Q	S S	*	
Act42A	L	A	M	V	S	L	IS Q	S S	*	
Muscyt	L	A	M	I	S	L	IS Q	S S	*	
Musmus	M	A	M	I	S	L	IT Q	A S	*	
CON	-SGGTTMYPG IADRMQKEIT -LAPST-KIK I-APPERKYS VWIGG-ILAS -STFQQMW-- K-EYDE-GP- IVHRKCF-									
	301				350					378

FIG. 2.1. Comparison of the six *Drosophila* actins to the mouse striated muscle and cytoskeletal actins. The CON(sensus) line displays all positions for which there is total agreement among the sequences. Where there is no such agreement, the residues occupying that position in each sequence is indicated. The sequence of *Act57B* is known from a cDNA. There is 98% overall identity between the *Drosophila* and mouse cytoskeletal proteins.

### *Mutant Phenotype*

Mutations in *Act88F* affect only the development of indirect flight muscles, and mutants are viable (Karlik et al. 1984; Mahaffey et al. 1985; Okamoto et al. 1986). Some mutations, such as *Act88F*<sup>KM88</sup> and *Act88F*<sup>KM129</sup>, are recessive hypomorphs producing severely altered proteins that fail to accumulate. Other alleles, those with more subtle changes such as *Act88F*<sup>KM75</sup>, are antimorphs; they are dominant even in the presence of two normal alleles and often result in the expression of heat-shock genes, probably induced by the accumulation of denatured muscle proteins (Okamoto et al. 1986; Drummond et al. 1991).

### **Common Features of Gene Organization and Expression**

Open reading frame, 376 amino acids. Although coding sequences are 85–95% conserved among all *Drosophila* actins, the position of introns is not constant (Fyrberg et al. 1981; Fig. 2.2). Transcription from the six genes is differentially modulated during development, in accordance with the tissue distribution of their products (Bond-Matthews and Davidson 1988; Burn et al. 1989; Tobin et al. 1990).

### *Act5C*

#### **Gene Organization and Expression**

Determination of 5' and 3' ends was by S1 mapping and by RNase protection studies, primer extension and sequencing of several cDNAs. Transcription occurs from two main initiation sites. The upstream site is preceded by a putative TATA box, and the position of the 5' end seems to be quite invariant. The downstream initiation site lacks a canonical TATA box, and there is some microheterogeneity in the 5' end, although the main site seems to be –712. Both leaders have introns with donor sites at –1,675 and –602 and a common acceptor site at –7 (*Act5C* Sequence and Fig. 2.2). Three major and two minor alternative poly-A sites exist, and it is probable that all possible combinations of initiation and polyadenylation sites are used. The major classes of mRNAs would range from 1,524 to 1,919 bases. Three mRNA bands resolved by northern analysis are 1.8 kb, 2.0 kb and 2.3 kb long (Fyrberg et al. 1981; Bond and Davidson 1986; Vigoreaux and Tobin 1987; Chung and Keller 1990a).

#### *Developmental Pattern*

The gene for the cytoplasmic actin 5C is, as would be expected, transcribed in all tissues. Its maternal mRNA is uniformly distributed in preblastoderm embryos. During blastoderm formation this mRNA becomes localized in a peripheral layer; and, as tissue differentiation proceeds, it remains present in

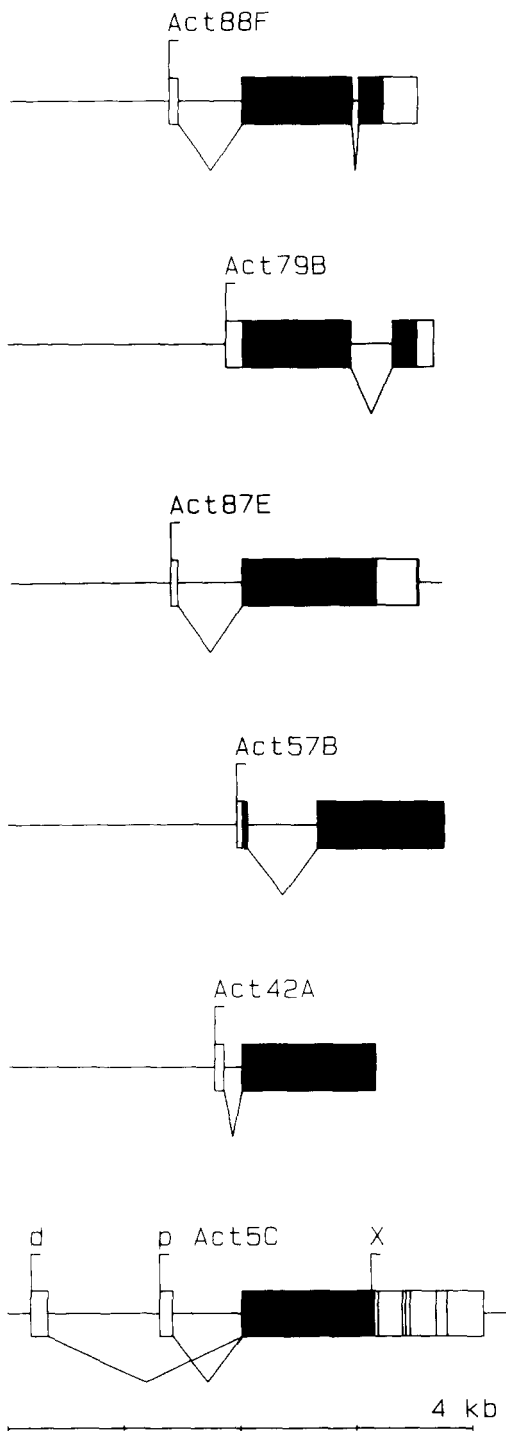


FIG. 2.2. Organization of the six actin genes.

*Act5C*

-3735 ATTTTCTACAAAACATGTTATCTATAGATAATTTTGTGCAAAATATGTTGACTATGACAAAGATTGTATGTATACCTTTAATGTAT -364  
 -3645 TCTCATTTTCTTATGTATTATAATGGCAATGATGATACTGATGATATTTAAGATGATGCCAGACCACAGGCTGATTTCGCGTCTTTT -355  
 -3555 GCCGAACGCAGTGCATGTGCGGTTGTTGTTTTTGGAAATAGTTTCAATTTTCGGACTGTCGCTTTGATTTTCAGTTTCTTGGCTTATTCA -346  
 -3465 AAAAGCAAAGTAAAGCCAAAAAAGCGAGATGGCAATACCAATGCGGCAAAACGGTAGTGAAGGAAAGGGGTGCGGGGCAGCGGAAGGA -337  
 -3375 AGGGTGGGGCGGGGCGTGGCGGGTCTGTGGCTGGGCGCGACGTACCAGCTGGAGCCACTCCTTTGACCATGTGTGCGTGTGTAT -328  
 -3285 TATTCGTGTCTCGCCACTCGCCGGTTGTTTTTCTTTTTATCTCGCTCTCTAGCGCCATCTCGTACGCATGCTCAACGCACCGCATG -319  
 -3195 TTGCCGTGTCTTTATGCGTCAATTTGGCTCGAAATAGGCAATTTTAAACAAGATTAGTCAACGAAAACGCTAAAAATAAATAAGTCT -310  
 -3105 ACAATATGTTACTTATTGCCATGTGTGCAGCCAACGATAGCAACAAAAGCAACAACACAGTGGCTTCCCTCTTCTACTTTTTGTTT -301  
 -3015 GCAAGCGCGTGCAGCAAGACGGCAGCACCAGGCAAAACGCAATTACGCTGACAAAGAGCAGACGAAGTTTTGGCCGAAAAACATCAAGCG -292  
 -2925 CCTGATACGAATGCAATTTGCAATAACAATTCGCATATTTAATATTGTTTATGAAGCTGTTTGACTTCAAAAACACAAAAAATAA -283  
 -2835 AACAAATTATTTGAAAGAGAATTAGGAATCGGACAGCTTATCGTTACGGGCTAACAGCACACCGAGACGAAATAGCTTACCTGACGTAC -274  
 -2745 AGCCTCGGAAGAACTGCCCCAAGCAGACGATGCAGAGGACGACACATAGAGTAGCGGAGTAGGCCAGCGTAGTACGCATGTCTTGTG -265  
 -2655 TGTGAGGCGTCTCTCTCTCGTCTCTGTTTGCGCAACGCATAGACTGCACGTGAGAAAATCGATTACCTATTTTTATGAATGAATATT -256  
 -2565 TGCATATTACTATTCAAACATTAAGATAGCAATCACATTCATAGCCAAATCTATACCACCTGAGCGATGCAACGAAATGATCAAT -247  
 -2475 TTGAGCAAAAATGCTGCATATTTAGGACGGCATCATTATAGAAATGCTTCTTGCTGTGACTTTTCTCTGCTGGCAGCTGTTTCGCCG -238  
 -2385 TTATTGTTAAAACCGGCTTAAGTTAGGTGTTTTCTACGACTAGTGATGCCCTACTAGAAGATGTGTGTTGCACAAATGTCCTGAAT -229  
 -2295 AACCAATTTGAAGTGCAGATAGCAGTAAACGTAAGCTAATATGAATATTTTAACTGTAATGTTTAAATATCGCTGGACATTACTAATA -220  
 -2205 AACCCACTATAAACACATGTACATATGTATTGTTTTGGCATACAATGAGTAGTTGGGAAAAAATGTGTAAAAGCACCGTGACCATCACA -211  
   -----A5Ce3      -----cA5Ce3  -----A5Ce2  
 -2115 GCATAAAGATAACCAGCTGAAGTATCGAATATGAGTAAACCCCAAATTGAATCACATGCCCAACTGATAGGACCCATGGAAGTACACTC -202  
 -2025 TCATGGCGATATACAAGACACACACAAGCAGCAACACCCAGTTGCGGAGGAAATCTCCGTAATGAAAACCCAATCGGCGAACAATTCA -193  
 -1935 TACCATATATGGTAAAGTTTTGAACGCGACTTGAGAGCGGAGACGATTGCGGCTGATAAGGTTTTAGCGCTAAGCGGGCTTTAATAAA -184  
   -->-1821  -----  
 -1845 ACGGGCTGCGGGACCAGTTTTCATATCACTACCGTTTGAGTTCTGTGCTGTGGATACTCCTCCGACACAAAAGCCGCTCCATCAGCC -175  
 -1755 AGCAGTCGTCTAATCCAGAGACACCAAACCGAAAGACTTAATTTATATTTAATTAATTTAATAAAACACACCAAATGTAAGTAGC -166  
 -1665 TTTCCCTTCCCAACAACAAAACACCATCGAACCCTCCCACCAAGAAAAAGCAATAATCGAGAAAAGCCGCGGAAAAATGTGTGATTTTT -157  
 -1575 TTTGTAACAAAATTTTTTATGTGCCAGTGTGAAAGTGATCAAAAATACTAGCCACGAGCTAAGAGTTATTGTATTGACCAAAACT -148

-1485 CCAAAAATACCCAAGTTTGGCCCTAAATGTCAATCAAATACCAATAGGTCGAAAGACATCAAAATTAACAAAACCAGGGTTTCAAATA -1396

-1395 CCATAACTCAAGAATCAGGATTACAACCTGCAGATTTTCAGGATATATACATACAAAATTATAGCAAATATAAAAACCAAAGCAATTCATAG -1306

-1305 CCCCAACTCAAATGTTAGGATCTAATATAGTGTAAAGCCAAGCTCGCTGATGTGGGCGTGCACGATTTACCACAAAGATATGCCAAA -1216

-1215 TTACGAATTGCAAAATCAATTC6CCAACACTTCTTTTTTCCACGCCTAAAAACAGATCATCAAAATGTACATACATACAGTATATGC -1126

-1125 ATATTATAACTGTAAACTAGATCAGGTTCTTAAAAATAGTGACGTAGGAGCCGTTTTGGCTGAAGCAGAAATTTTGGCCGTTTTTCAA -1036  
---

-1035 AGTTGTAGTTGCAAAAATGGAGAAAACCTTCGAGCATTCTGTTTCATATACACACACTCACGCACAAAATAACGAGAGAGAGTGTATGTGTG -946  
-----A5Ce1

-945 TGTGAGAGAGCGAAAGCCAGACGACGGTTTGCTTTTCGCCTCGAAACATGACCATATATGGTCACAAAACCTGGCCGCCCAATTCAACA -856

-855 CACCAGCGCTCTCCTTCGCACCCATAGCGACCATGGCGCGGAGCGAGAGTGGCGAGAGCGAGCGCTATGGCAGCTCGACGCAG -766  
<----> ----> ---->  
-->-712 -->-704 (minor)

-765 GCAGCGATTGAAAAACGAGTTAACTGGCATTCAACATTCACCAGCCACTTTCAGTCGGTTTATCCAGTCATTCTTTCAAACCGTGGC -676

-675 GTCGCTTAGCTCAGCCTCGCCACTTGCCTTTACAGTAGTTTTACGCCTTGAATTTGTTAAATCGAACAAAAAGGTTAAAGTTAACTAGC -586  
|

-585 TTTGAAAAGTTTCGTGGCTCTTAATTTGTTAAATTTCTAGAGTGCCTTAGTGTTTTTTTTTTTTTATTTGTAATGTTAATTTTCGGG -496  
|

-495 TTCCAATTCGAGTTTTAGGCAGCCGCCATTTTAAAGGGCGCATACACAGGCAACTGTGCTCTCTTTGCGGCTTCTTTTGCACCGGCAT -406

-405 TCGTTAAGCTGCTGCTAGAAAGCTTCTCCCTCCCTTTTCGCATATTCGTATTGTGGTTTTAATTTTTCGGGCGGGGCTTCTATTTTG -316

-315 TAACTGTCTTTTAAATTTCTTATTACAATTCGATCGCAAGTAAAATCAGTTTTCAATCGGAAAAGTATTTTTTATGAAATTTTTTTTT -226

-225 GTCCAAGATTAATAATTTTGTACTAAAAAACGTACATTGCATTGAGTGATTTTTAATTTGACACGAAAAACAAGTTAGTTGTTATGACA -136

-135 ATTGTACTTTGGTAGACCAGCGAGTCCAAGGAGACCACGCAAAATCTCAGTTTTTTTTTGGCATTCTACATTACCAATAAGGTAAC -46

-45 CAAAACTAATGGGAAATCCGCATCTTTCCATTGCAGCTTACAAAAATGTGTGACGAAGAAGTGTCTGCTCTGGTTGTCGACAACGGCTC 44  
| MetCysAspGluGluValAlaAlaLeuValValAspAsnGlySe (15)

45 TGGCATGTGCAAGGCCGGATTTGCCGGAGACGATGCTCCCGCGCCGTCTCCCATCGATTGTGGACGTCCTCCGTCACCAGGGTGTGAT 134  
rGlyMetCysLysAlaGlyPheAlaGlyAspAspAlaProArgAlaValPheProSerIleValGlyArgProArgHisGlnGlyValMe (45)

135 GGTGGCATGGGCCAGAAGGACTCGTACGTGGTGATGAGGCGCAGAGCAAGCGTGGTATCCTCACCCCTGAAGTACCCATTGAGCACGG 224  
tValGlyMetGlyGlnLysAspSerTyrValGlyAspGluAlaGlnSerLysArgGlyIleLeuThrLeuLysTyrProIleGluHisGln (75)

225 TATCGTGACCAACTGGGACGATATGGAGAAGATCTGGCACCACACCTTCTACAATGAGCTGCGTGTGGCACCCGAGGAGCACCCCGTGT 314  
yIleValThrAsnTrpAspAspMetGluLysIleTrpHisHisThrPheTyrAsnGluLeuArgValAlaProGluGluHisProValLe (105)

315 GCTGACCGAGGCCCGCTGAACCCCAAGGCCAACCGTGAGAAGATGACCCAGATCATGTTCCGAGACCTTCAACACACCCGCCATGTATGT 404  
uLeuThrGluAlaProLeuAsnProLysAlaAsnArgGluLysMetThrGlnIleMetPheGluThrPheAsnThrProAlaMetTyrVa (135)

(continued)



405	GGCCATCCAGGCTGTGCTCTCGCTGTACGCTTCGGGTCGTACCACCGGTATCGTCTGGACTCCGGCGATGGTGTCTCCACACCGTGCC 1A1a1IeGlnA1aValLeuSerLeuTyrA1aSerGlyArgThrThrGlyI1eValLeuAspSerGlyAspGlyValSerHisThrValPr	494 (165)
495	CATCTACGAGGGTTATGCCCTCCCCATGCCATCCTGCGTCTGGATCTGGCTGGTCGCGATTTGACCGACTACCTGATGAAGATCCTGAC oI1eTyrGluGlyTyrA1aLeuProHisA1aI1eLeuArgLeuAspLeuA1aGlyArgAspLeuThrAspTyrLeuMetLysI1eLeuTh	584 (195)
585	CGAGCGGGTACTCTTTCCACCACCCTGCTGAGCGTGAATCGTCCGTGACATCAAGGAGAAGCTGTGCTATGTTGCCCTCGACTTTGA rGluArgGlyTyrSerPheThrThrA1aGluArgGluI1eValArgAspI1eLysGluLysLeuCysTyrValA1aLeuAspPheG1	674 (225)
675	GCAGGAGATGGCCACCGCTGCCAGCAGCTCCTCGTTGGAGAAGTCTACAGCTGCCGACGCGAGGTGATCACCATCGGCAACGAGCG uGlnGluMetA1aThrA1aA1aSerSerSerSerLeuGluLysSerTyrGluLeuProAspGlyGlnValI1eThrI1eGlyAsnGluAr	764 (255)
765	TTTCCGCTGCCCGAGGCCCTGTCCATCCCTCGTTCCTTGGGATGGAGCTTGGCGCATCCACGAGACCACCTACAACCTCCATCATGAA gPheArgCysProGluA1aLeuPheHisProSerPheLeuGlyMetGluSerCysGlyI1eHisGluThrThrTyrAsnSerI1eMetLy	854 (285)
855	GTGTGATGTGGATATCCGTAAGGATCTGTATGCCAACCCGTGCTGTCGGTGGCACCACCATGTACCTGGCATCGCCGACCGTATGCA sCysAspValAspI1eArgLysAspLeuTyrA1aAsnThrValLeuSerGlyGlyThrThrMetTyrProGlyI1eA1aAspArgMetG1	944 (315)
945	GAAGGAGATCACCGCCTGGCCACCGTCGACCATGAAGATCAAGATCATTGCCCGCCAGAGCGCAAGTACTCTGTCTGGATCGGTGGCTC nLysGluI1eThrA1aLeuA1aProSerThrMetLysI1eLysI1eI1eA1aProProGluArgLysTyrSerValTrpI1eGlyGlySe	1034 (345)
		-->1108 (X)
1035	CATCTGCGTTCGCTGTCCACCTTCCAGCAGATGGATCTCCAAGCAGGAGTACGACGAGTCCGGCCCTCCATTGTGCACC6CAAGTG rI1eLeuA1aSerLeuSerThrPheGlnGlnMetTrpI1eSerLysGlnGluTyrAspGluSerGlyProSerI1eValHisArgLysCy	1124 (375)
1125	CTTCTAAGAAGGATCGCTTGCTGGGCAAGAGGATCAGGATCGGGATGGTCTTGATTCTGCTGGCAGGAGGAGGAGGAGAAGTCGAGGAA sPheEnd (376)	MetValLeuI1eLeuLeuA1aGlyGlyGlyGlyGluValGluGlu (15)
1215	GCAGCAGCGAAAGTCAAGTCCGAGTGGTGGAAAGTTTGGAGTGCAGCACAAACAAATCAACAACAACACCAACTACAAGATGAAAAGAGC A1aA1aA1aLysValGlnValArgValValGluValTrpSerA1aA1aGlnGlnAsnGlnGlnHisGlnLeuGlnAspGluLysSer	1304 (45)
1305	GGAACCACTGCCACACCATCATCACTATCATCGTTTTGGGCGCATGTTGTGTGGTTCACGCGTATTAATAATAATATTTATCCAA GlyThrThrCysHisThrI1eI1eThrI1eI1eI1eValLeuGlyA1aCysCysValValProA1aTyrEnd	1394 (68)
		-----
1395	TGAGATATGATATGATATACTATGTATTTTTTGTTTTTTTTTTATTGTAAACCTTTAATAAACAAGAAGTACAAAAAGTGAAAATGA  (A) <sub>n</sub>  (A) <sub>n</sub> ----- (X)  (A) <sub>n</sub> (X)	1484
1485	GCGAAAATGCATATTTGCCATTCCACACACACCAACAACACCCAAACACACGACACCCCAAGCTTACACACACACACATTCCGCGGC	1574
1575	ATGACAAGGACATCAAGATAAAGAAGAACTTAAGAAGATATTTCCCAAAGCGCAAAAAGAACACACACACATTGCAAAAACACAAACAAC	1664
1665	ACACTAGCGTTTTGTACAATTCGTCAGCAACCTTATGTATTATTTTTAATTATGATGAATTATAAACAAGTGAAAACAAAATATGAA  (A) <sub>n</sub> -----	1754
1755	AACAAAAAGGAAAATCAAATCTGCTTCTCTTTCTCCCGCTCTCCTCGCTCTCTGCTGCTAACCTCGCCCTCTCTCTCATCTTTTTG  (A) <sub>n</sub>	1844
1845	TCTGTCTCTTCCACATTTTTGGCGCCGGCAAAAATAAACCCACACACTCACACTTGGCTGCAGTTTCGCGTGCATATTCACACA	1934
1935	CATTCAAGCATACATATGTATTTTTTTTTTATTGTACACTTTTCTAATTGCATGCGTATCGATTGATAAGTTTACGCTGAAAAATG	2024
2025	TTAATTAATAATGTGAAAATGCAACTGAAAACTGATGAAATGAAACAACAACAAGCGAACAA	2086

all organs. There is a slightly greater accumulation of *Act5C* mRNA in the anterior and posterior segments of the prospective midgut, apparently due to increased transcription from the distal initiation site (Burn et al. 1989). Both cytoplasmic actin genes, *Act5C* and *Act42A*, are the only actin genes transcribed in Kc cells, with *Act5C* transcripts being 6–8 fold more abundant. The level of *Act5C* transcript increases 3–5 fold in response to 20-hydroxyecdysone treatment (Couderc et al. 1987). Most *Act5C* mRNA is associated with polysomes (Rao et al. 1988).

### Promoter

The two transcription initiation sites respond to independent regulatory regions, as shown by the expression of a reporter gene in cultured cells (Bond-Matthews and Davidson 1988). The distal promoter is the stronger and is developmentally regulated; the proximal promoter is uniformly expressed in all cell types (Vigoreaux and Tobin 1987; Burn et al. 1989).

*Distal Promoter* The controlling elements of the distal promoter include one that extends between 2,071 and 1,866 bp upstream of the transcription initiation site and several others that lie within 540 bp of the 5' end. These were identified by reporter gene expression essays performed in cultured cells (Bond-Matthews and Davidson 1988; Chung and Keller 1990b). A bipartite element between –2,343 and –2,182 strongly represses expression, and three elements with a positive effect on expression are found between –2,182 and –2,099, between –2,068 and –2,040 and between –1,911 and –1,864. The segment between –2,182 and –2,099 has the strongest effect, and footprinting and mutational analysis identified A5Ce2 (*Act5C* Sequence) as the main regulatory element in this region. *In vitro* mutagenesis identified two other elements, A5Ce3 and cA5Ce3 (Chung and Keller 1990b).

*Proximal Promoter* The proximal promoter contains three elements involved in the control of transcription, which were identified by band-shift assays, footprinting and expression of a reporter gene (Chung and Keller 1990a): (1) a 14-bp segment between –1,038 and –1,025 (A5Ce1 in the *Act5C* Sequence) that is necessary for full expression; (2) the 98 base pairs between –872 and –774 whose effect is probably due to the presence of three copies of the GAGA transcription factor binding sites (Biggin and Tjian 1988); and (3) the segment

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*Act5C* SEQUENCE (opposite). Mostly from Canton S. Accession, X15730 (DROACT5CB), X06382 (DRO5CACT1), X06383 (DRO5CACT2), X06384 (DRO5CACT3), M13586 (DROACT5C2). Two bases, –819 and –820, were corrected as suggested by Chung and Keller (1990a). Arrows between –855 and –766 underline potential binding sites for the GAGA factor. The initiation and termination of the 3' transcriptional unit are marked by X.

*Act42A*

-513	TCGAATTTTGAACACTGCATAATTTTTAAATGCATTTTCAAGGATCTTAGATCATTCTAATTTGTTGATAACACGTCAGTATACCA	-424
-423	ATGAATAAAAAATTTTAAAAAAGTCCGCTCTCCAGTCTCCACCGTTTCCAACCTTATCGCACATTTATTGTTGGTGGAGTCACCTTCGGAA	-334
	-----	
	-->-257	
-333	GTAAAAAGACCATAATTTTATGCGTATATGGTCACACTACTTTTCAACACTTAACTCGAAAAGTAGCGTCGTCATTAATCTTAAAG	-244
-243	CGTCTGCATTGTGCTAAGTGTGTGCAGCGGATAACTAGAACTACTCCTACATATTTCCATAAAAAGGTAAGACTCCTGCCAACACTTT	-154
-153	TTTTTGTCTGTGCGGTCAATTATTCTCTTCTGGAAGGGGTGGTCCCGTCTCGCTCTTTTTACGTAGCCGCTGCTGCTCTCTCT	-64
-63	TGTTTTTTAGTGACACATCCAGATTTCTTTCTCTTGCGATCCAAATAAAATTTCTACAAAATGTGTGACGAGAGGTTGCAGCTTT	26
	MetCysAspGluGluValAlaAlaLe	(9)
27	AGTGGTCGACAACGGATCCGGCATGTGCAAAGCCGGCTTTGCCGGTGTGACGACCCGCGTCAGTTTTCTTCTATTGTGCGCCGCTCC	116
	uValValAspAsnGlySerGlyMetCysLysAlaGlyPheAlaGlyAspAspAlaProArgAlaValPheProSerIleValGlyArgPr	(39)
117	ACGTCAACGAGGCGTAATGGTAGGAATGGGACAAAAGGACTCTTATGTGCGGCGATGAGGCACAGAGCAAACGTTGATCCTTACCCTGAA	206
	oArgHisGlnGlyValMetValGlyMetGlyGlnLysAspSerTyrValGlyAspGluAlaGlnSerLysArgGlyIleLeuThrLeuLy	(69)
207	GTACCCCATGAGCAGCGTATCGTGACTAACTGGGACGACATGGAGAAGATCTGGCATCACACTTCTACAACGAGCTTCGTGTGGCCCC	296
	sTyrProIleGluHisGlyIleValThrAsnTrpAspAspMetGluLysIleTrpHisHisThrPheTyrAsnGluLeuArgValAlaPr	(99)
297	GGAGGAGCACCCCGTCTTGCTTACTGAGGCTCCTTTGAACCCCAAGGCTAATCGCGAAAAGATGACTCAGATTATGTTGAAACCTTCAA	386
	oGluGluHisProValLeuLeuThrGluAlaProLeuAsnProLysAlaAsnArgGluLysMetThrGlnIleMetPheGluThrPheAs	(129)
387	CACTCCGCCATGTATGTTGCCATCCAAGCGTGTCTTCTCTACGCCCTCCGGCCGTACCACAGGTATCGTGTGGACTCCGGGGACGG	476
	nThrProAlaMetTyrValAlaIleGlnAlaValLeuSerLeuTyrAlaSerGlyArgThrThrGlyIleValLeuAspSerGlyAspGl	(159)
477	TGCTCCCATACCGTGCCATCTATGAGGGCTACGCTCTGCCGCACGCTATCTCCGCTGGATCTAGCCGGTCGCGATTTAACCGACTA	566
	yValSerHisThrValProIleTyrGluGlyTyrAlaLeuProHisAlaIleLeuArgLeuAspLeuAlaGlyArgAspLeuThrAspTy	(189)
567	CCTGATGAAGATTCTTACTGAGCGGGTTACAGCTTACCACCACCGCCGAGCGTAAATTTGCGCGACATCAAGGAGAAGCTGTGCTA	656
	rLeuMetLysIleLeuThrGluArgGlyTyrSerPheThrThrAlaGluArgGluIleValArgAspIleLysGluLysLeuCysTy	(219)
657	CGTGGCCTTGACTTCGAGCAGGAGATGGCCACGCGCCGCTTCAAGCTCGTCCCTGGAGAAGTCGTACGAGTTGCCGATGGACAGGTCAT	746
	rValAlaLeuAspPheGluGlnGluMetAlaThrAlaAlaSerSerSerSerLeuGluLysSerTyrGluLeuProAspGlyGlnValIl	(249)
747	CACCATCGAAATGAGCGATTCCGTTGCCCGAATCGCTGTTCCAGCCGTCGTTCTCCGGCATGGAGGCCCTGTGGACTTCACGAGACCAC	836
	eThrIleGlyAsnGluArgPheArgCysProGluSerLeuPheGlnProSerPheLeuGlyMetGluAlaCysGlyLeuHisGluThrTh	(279)
837	CTACAACCTCAATCATGAAGTGTGACGCTGCATCCGTAAGGATCTGTACGCCAACACTGTGCTGTCCGGCGGCACCACCATGTACCCGGG	926
	rTyrAsnSerIleMetLysCysAspValAspIleArgLysAspLeuTyrAlaAsnThrValLeuSerGlyGlyThrThrMetTyrProGl	(309)
927	AATCGCTGACCGCATGCAAAGGAAATCACGGCGTTGGCTCCGTCACCATGAAGATTAAGATTGTTGCCCGCCAGAACGCAAGTACTC	1016
	yIleAlaAspArgMetGlnLysGluIleThrAlaLeuAlaProSerThrMetLysIleLysIleValAlaProProGluArgLysTyrSe	(339)
1017	TGTTTGGATCGCGGCTCCATCCTAGTCTCGCTGTCTACTTTCCAGCAGATGTGGATCTCGAAGCAAGGATACGACGAGTCGGGCCCCCTC	1106
	rValTrpIleGlyGlySerIleLeuAlaSerLeuSerThrPheGlnGlnMetTrpIleSerLysGlnGluTyrAspGluSerGlyProSe	(369)
1107	CATTGTTCAACGCAAGTCTCTAA	1131
	rIleValHisArgLysCysPheEnd	(376)

between -770 and -744, the position that a TATA box would normally occupy.

### Transcription unit X

This transcription unit overlaps the last few codons and 3' untranslated region of *Act5C*.

### Gene Organization and Expression

Open reading frame, 68 amino acids; mRNA, 368 bases, in agreement with a 0.45 kb band detected by northern analysis. S1 mapping and primer extension were used to determine the 5' end. S1 mapping was used to determine the 3' end (see *Act5C* sequence). This mRNA is found in polysomes and has the same tissue and developmental distribution as *Act5C* mRNA. Its function is unknown (Rao et al. 1988).

### *Act42A*

### Gene Organization and Expression

The 5' end was determined by S1 mapping; there is no obvious TATA box in its neighborhood. The 3' end has not been determined. There is a leader intron with a donor site at -177 and an acceptor site at -21. Because most of the coding sequence was determined from a cDNA, the presence of other small introns cannot be ruled out (*Act42A* sequence) (Fyrberg et al. 1981; Couderc et al. 1987).

### *Developmental Pattern*

During embryonic development, *Act42A* transcription follows a pattern similar to that of *Act5C*. The accumulation of transcripts is greatest in the midgut, central nervous system and gonads (Tobin et al. 1990). *Act42A* is expressed in

Act79B

-517	AGCTTACAAGTGTGTGCGGACCAAATCTCAACATAAACAAGACTTACAACCTACAAAAACAATTATTTATATCGAAATCCAGTACC	-428
-427	AATTTAGTTGCTCTAAGTTGTGGCTTAACTAGGGTCTTTAATTCGTAATCCAACCTGTTGCCGTAGGCATACCCGAAATCGGAACAATT	-338
-337	TTTGTGAAATCGAAATGATGTCGATCCGACCACCTCCCCGGAACGCCGTATCCCAAGCCAGCTTACATATCGCGGAATTCATCAACAT	-248
-247	GTTACTAGATGAACAATTGTTCCGAGATGACAGGGACATGGGCGTGGGGCGGGGGGAGACAAGCTTATTTAAATGCAGCTGCCGGA	-158
	----- -->-146	
-157	GCGCATAACGAATCACTCTGATGCTGCTGCTGTTGGATTTACACGTCGTGAGTGTAGTCTTGTCCGCCATCCGAAATCCGTAACCCGC	-68
-67	ATAAGGGATAACCGATTCTGTTGTACCCCTGTACCCCTGTGTACCGCCCCGACCAAACTAACCAAACTGTGTGACGAAGAAGCATCAG	22
	MetCysAspGluGluAlaSerA	(8)
23	CCCTGGTCGTAGACAACGGCTCCGGCATGTGCAAGGCCGGATTGCGCGAGACGACGCGCCCGCGGGTATCCCTCGATCGTAGGCC	112
	1aLeuValValAspAsnGlySerGlyMetCysLysAlaGlyPheAlaGlyAspAspAlaProArgAlaValPheProSerIleValGlyA	(38)
113	GTCCCGTACCACGGCGTGATGGTGGGATGGGTGAGAAAGACTGCTACGTGGGCGACGAGCGCAAAGCAAGCGGGTATCCTGTCGC	202
	rgProArgHisGlnGlyValMetValGlyMetGlyGlnLysAspCysTyrValGlyAspGluAlaGlnSerLysArgGlyIleLeuSerL	(68)
203	TGAAGTACCCATCGAACACGGCATTATCACCAACTGGGATGACATGGAGAAGGTCTGGCCACACACCTTCTACAACGAGCTGCGTGTGG	292
	euLysTyrProIleGluHisGlyIleIleThrAsnTrpAspMetGluLysValTrpHisHisThrPheTyrAsnGluLeuArgValA	(98)
293	CCCCGAGGAGCACCCCGTTCGCTGACCGAGGCTCCCTTGAACCCCAAGGCCAACCGCGAGAAGATGACCCAGATCATGTTCCGAGACGT	382
	1aProGluGluHisProValLeuLeuThrGluAlaProLeuAsnProLysAlaAsnArgGluLysMetThrGlnIleMetPheGluThrP	(128)
383	TCAACTCCCGGCCATGTACGTGGCCATCCAGGCCGTGCTCTCCCTGTATGCTTCCGGCCGTACCACCCGGTATGCTCTGGACTCCGGTG	472
	heAsnSerProAlaMetTyrValAlaIleGlnAlaValLeuSerLeuTyrAlaSerGlyArgThrThrGlyIleValLeuAspSerGlyA	(158)
473	ACGGTGTCTCCACACCGTGCCCATCTATGAGGGCTATGCCCTGCCACGCCATCCTTCGTCTAGATCTGGCCGGTCGCCATTAACCG	562
	spGlyValSerHisThrValProIleTyrGluGlyTyrAlaLeuProHisAlaIleLeuArgLeuAspLeuAlaGlyArgHisLeuThrA	(188)
563	ACTACCTGATGAAGATCCTCACCGAGCGGGCTACAGCTTACCACCACCGCCGAGCGAGATTGTGCGCGACATCAAGGAGAAGCTGT	652
	spTyrLeuMetLysIleLeuThrGluArgGlyTyrSerPheThrThrThrAlaGluArgGluIleValArgAspIleLysGluLysLeuC	(218)
653	GCTACGTGCGCTGGACTTCGAGCAGGAGATGGCCACTGCCGCCCTCCACCTCCCTGGAGAAGTCTTACGAGCTGCCGATGGCCAGG	742
	ysTyrValAlaLeuAspPheGluGlnGluMetAlaThrAlaAlaAlaSerThrSerLeuGluLysSerTyrGluLeuProAspGlyGlnV	(248)
743	TAATCACCATCGGCAACGAGCGCTCCGCCACCCCGAGGCCCTTCCAGCCATCGTTCCTAGGCCATGGAGTCTGCGGCATCCACGAGA	832
	aIleIleThrIleGlyAsnGluArgPheArgThrProGluAlaLeuPheGlnProSerPheLeuGlyMetGluSerCysGlyIleHisGluT	(278)
833	CCGCTACCAGTCCATCATGAAGTGCAGCTGGACATCCGCAAGGATCTGTATGCCAAATGTGCTGTCTGGCGGCACTACCATGTATC	922
	hrValTyrGlnSerIleMetLysCysAspValAspIleArgLysAspLeuTyrAlaAsnAsnValLeuSerGlyGlyThrThrMetTyrP	(308)
923	CAGGTACGTAGTCTTAATTATTTAGGACCATAAAGTTCAGAGGAAATCTTCCGAGGGAATGGGATCAAACATATGCGGGATACTAAAA	1012
	roG	(309)
1013	AAAAAAACAAGTGTACTTTATACATTCATTTGGCAGAGAGCAAATCTTTAAATAATAAAGCCTAAATATTTAGCTGAGGTTTGTAAATA	1102
1103	CAGTTAAAAAAATCTTATGAAAGTAGTATTACAAAAAAGAAATTCACCTAACGGGTTATATCCTTTCCCTATATCTCATAT	1192
1193	TCATGCATGCTATTATTAATGTATGTAATGAGTACACCAAAGCTCCACGGTCCGTAGCACCAACCAATGGATTCTATTTCCGCCCTCT	1282

1283	CAGGTATCGCTGACCGTATGCAAAAGGAAATCACCGCACTTGCCCGTCCACCATGAAGATCAAGATCATGCCCCGCCAGAGCGCAAGT lyIleAlaAspArgMetGlnLysGluIleThrAlaLeuAlaProSerThrMetLysIleLysIleIleAlaProProGluArgLysT	1372 (338)
1373	ACTCCGCTCGGATCGGTGGCTCCATCCTGGCTTCGTTGTCCACCTTTCAGCAGATGTGGATCTCCAAGCAAGAGTATGACGAGTCCGGTC yrSerValTrpIleGlyGlySerIleLeuAlaSerLeuSerThrPheGlnGlnMetTrpIleSerLysGlnGluTyrAspGluSerGlyP	1462 (368)
1463	CGGGCATCGTCCACCGCAAGTGTCTTAAGCATCCAGGCCACCCAACCAAGGTC AACATCTCCTCGAGGCGCGGCCCTGGTGTTTGTCTC roGlyIleValHisArgLysCysPheEnd	1552 (376)
1553	CAGCGTAAGACATCCGACTAGGCGTCGGCGCACAGGGTCCGAGGACCGCAGTTCACTGAAAAGATCCTTAAATAACATTTAGTCGATGAA	1642
1643	GAAGTTTTAACA	1654

*Act79B* SEQUENCE. Strain, *Canton S*. Accession, M18829 (DROACT79B).

Kc cells and transcription is enhanced 6–8 fold in the presence of 20-hydroxyecdysone (see *Act5C*; Couderc et al. 1987).

### *Act57B*

#### Gene Organization and Expression

The 5' and 3' ends were not determined. There is an intron in the Gly-14 codon. Most of the coding sequence was determined from cDNA clones only; and the presence of other small introns cannot be ruled out [Fyrberg et al. 1981; Accession, K00672 (DROACT7A1) and K00673 (DROACT7A3)]. The amino acid sequence is shown in Fig. 1. In embryos, transcripts are detectable in the developing musculature of the future larval body wall (Tobin et al. 1990).

### *Act79B*

#### Gene Organization and Expression

The 5' end was determined by S1 mapping. The 3' end has not been determined. There is an intron within the Gly-309 codon (*Act79B* Sequence) (Fyrberg et al. 1981; Sanchez et al. 1983).

#### *Developmental Pattern*

Transcription is undetectable in embryos (Tobin et al. 1990), it increases during the first larval instar, peaks during the second instar and diminishes in the third instar and in prepupae. Another small burst of transcription occurs during pupation (Sanchez et al. 1983). Studies of transcript distribution and the pattern of expression of a reporter gene controlled by 4 kb of the *Act79B* promoter region showed that transcription starts in midpupae (at 168 h) and continues

*Act87E*

-981	TATTAGAAAACCATCACACAATAGAAAAAGGTACAAAAATAGATAATTTTCATCCATCATATGCGCTTTACAAAAATCTATATTTTCTC	-982
-891	ATAACATATTTTGGCCATCTTTCTGCAAGTGCACCATCTGGGAAATATGAACGAAGCGAGCAGAAGTCCAAAAGCAAAAAATCCTACGA	-802
-801	AAACAAATTATTTTAAAAGAACTCAGAATCTCCCCCGCCGGCGCAATGTGCATCCATGTGCACATGTGTGCCGAGAGGGCGATTGAGT	-712
		----- -->-637.
-711	GTGCGTGCGGAAAATATCTAAAACGACTGAGGGTCGCCAGAATGGTATAAAATATTAGCGCATCTCGTCCAGCGACCACTCGCAGTTCTA	-622
		----- 
-621	CAGCGAAAGTGTGATTGGATTCTAGTTTTCTTCGTCTAACGGTTAGTATACTCCACATCCACCAATCCGTCTGTCTGGTTGACTT	-532
-531	TTACCAATCCGATGCTGGATCCAGTGTACAGTGCCCAACTTTCTGAAAAGAAAGATACTTTGAAAAGATAGAGATCTCAACAAACAACA	-442
-441	TATTTGACAAGGAGCAGAAAAAGTTCAATCAACGATCCTTAAATGTTTGGTTTTTAAATAGTGACTAACTTTTGTTTAAAAAACATCTCT	-352
-351	TAAAATGTTAAAACAGAAAAATATTAGTTGTTGATCTTAAATCAAAAATTATAATTAATTAATAACTTTTATTAAGTATGTAATATCGT	-262
-261	CAAAAAGTTGAAGCAGCCCTTTGTTAATTATCCACGTTTCGATTAATTTTAAAGATTGCTCCTCTGCAAAAGATACTTTCTTTTAAAGT	-172
-171	CATACATGTTCTGAGGCAACACCTACACGTATTCATAATTTACACTTACACACAAGATTACAATTAATAATCCATACCAATCCGATTC	-82
-81	CCGAAAGCCCACTTCTCACTTCTCCTTCTAAAACCGCCTCCGTTCTCGTTGTTGTTGCAGTGAAAACAGCCAGTAGCCAAGATGTGTGA	8
		MetCysAs (3)
9	CGATGAGGTTGCCGCAATTGGTCGTGGACAATGGTCCGGAATGTGCAAAGCAGGATTCGCCGCGATGATGCGCCTCGCGCCGTCTTCCC	98
	pAspGluValAlaAlaLeuValValAspAsnGlySerGlyMetCysLysAlaGlyPheAlaGlyAspAspAlaProArgAlaValPhePr	(33)
99	CTCGATTGTGGGTCGTCCCGTCATCAGGCGTAATGGTGGGCATGGGACAGAAGGACTCCTATGTTGGTATGAGGCCAGAGCAAGCG	188
	oSerIleValGlyArgProArgHisGlnGlyValMetValGlyMetGlyGlnLysAspSerTyrValGlyAspGluAlaGlnSerLysAr	(63)
189	TGGTATCCTCACCTGAAATACCCATCGAGCAGCGCATCATCAACCACTGGGACGATATGGAGAAGATGCGCACCACACTTTCTATAA	278
	gGlyIleLeuThrLeuLysTyrProIleGluHisGlyIleIleThrAsnTrpAspAspMetGluLysIleTrpHisHisThrPheTyrAs	(93)
279	CGAGCTGCGCGTCGCCCGAGGAACACCCGTCCTGCTGACCGAGGCCCCCTGAACCCCAAGGCCAATCGCGAGAAGATGACCCAGAT	368
	nGluLeuArgValAlaProGluGluHisProValLeuLeuThrGluAlaProLeuAsnProLysAlaAsnArgGluLysMetThrGlnI	(123)
369	CATGTTGAGACCTTCAACGCACCCGCCATGTATGTGGCCATCCAGGCTGTGCTCTCGCTGTACGCCCTCCGGTGTACCACCGGATTGT	458
	eMetPheGluThrPheAsnAlaProAlaMetTyrValAlaIleGlnAlaValLeuSerLeuTyrAlaSerGlyArgThrThrGlyIleVa	(153)
459	CCTCGACTCCGGTGACGGTGTCTCCACACCGTGCCATCTACGAGGGTTACGCCCTGCCACGCCATCTCGCTGTGGATCTGGCTGG	548
	lLeuAspSerGlyAspGlyValSerHisThrValProIleTyrGluGlyTyrAlaLeuProHisAlaIleLeuArgLeuAspLeuAlaGl	(183)
549	TCGCGATTTGACCGACTACCTGATGAAGATCCTGACCGAGCGCGGTTACTCATTACCACCACCGCTGAGCGTAAATCGTTCGCGACAT	638
	yArgAspLeuThrAspTyrLeuMetLysIleLeuThrGluArgGlyTyrSerPheThrThrThrAlaGluArgGluIleValArgAspI	(213)
639	CAAGGAGAAGCTGTGCTATGTTGCCCTGGACTTTGAGCAGGAGATGGCCACCGCCGCCCTCCACATCCTGGAGAAGTCATACGAGCT	728
	eLysGluLysLeuCysTyrValAlaLeuAspPheGluGlnGluMetAlaThrAlaAlaAlaSerThrSerLeuGluLysSerTyrGluLe	(243)
729	TCCCGACGGACAGGTGATCACCATCGGCAACGAACGTTCCGCTGCCAGAGTCGCTGTCCAGCCCTCTTCTCGGGAATGGAATCGTG	818
	uProAspGlyGlnValIleThrIleGlyAsnGluArgPheArgCysProGluSerLeuPheGlnProSerPheLeuGlyMetGluSerCy	(273)

819	CGGCATCCACGAGACCGTGTACAACCTCGATCATGAAGTGCATGTGGACATCCGTAAGGATCTGTATGCTAACATCGTCATGTCGGGTGG sGlyIleHisGluThrValTyrAsnSerIleMetLysCysAspValAspIleArgLysAspLeuTyrAlaAsnIleValMetSerGlyG	908 (303)
909	TACCACCATGTACCCTGGTATTGCCGATCGTATGCAGAAGGAGATCACCGCCCTGGCCCCGTCACCATCAAGATCAAGATCATTGCCCC yThrThrMetTyrProGlyIleAlaAspArgMetGlnLysGluIleThrAlaLeuAlaProSerThrIleLysIleLysIleIleAlaPr	998 (333)
999	ACCGGAGCGCAAGTACTCCGCTGGATCGGTGGCTCCATCTCCGCTCCCTGTCACCTTCCAGCAGATGTGGATCTCCAAGCAGGAGTA oProGluArgLysTyrSerValTrpIleGlyGlySerIleLeuAlaSerLeuSerThrPheGlnGlnMetTrpIleSerLysGlnGluTy	1088 (363)
1089	CGACGAGTCCGGCCAGGAATCGTCCACCGAAGTCTTCTAAGCGATCTAAACACCACAGACTGCAAACCACACGGGCATTGAGACC rAspGluSerGlyProGlyIleValHisArgLysCysPheEnd	1178 (376)
1179	CAACCACACCACGCCACAGAACACCACACAACAACAAGAACAACATGAACAGCAACAACCAAATACCAAATCAAGATCTATAGCCTA	1268
1269	GTGCTATTGATGATTAATCTTAAGTTAAACCTCTTGCTGCCCTGCCATCCAAAAGAAAACCGAAGGAACCGGATTGTAACAGCATGTAT	1358
1359	TATACTTATATTAATATTTATTGGAGAGCCGCTTGATGGCGCTGAAGGAGGAGTTGAGGAGACACAAGAATGCAAAATTTTACAGTTTTA	1448
1449	AAAATAAATTATACTAGCATCTCTATAAAATTAATCTAAATTTTAAACGAAACGTATCTTTTATTCGCTGAAGCGGCATGCTATGCGA -----   (A) <sub>n</sub>   (A) <sub>n</sub>   (A) <sub>n</sub>	1538
1539	TTATTTTTAGCGACGCACAGGAAATTACGAAATTTGTCACGCCCACTGCAAAGAGCGAAATCTGGAGGTGGATCTCCTCGACTGGGGTGC	1628
1629	ACATACATATGTACATATGTGGCTGGGGATGAGCACGGTAATCCAGCATAGACGCCCTCCAAGACAGTCCATTTTGGCCATTGCCAGTC	1718
1719	GGTGCAGGAGCTGCCCCCTCGTGGATCTAAAAATACAGGCCAAAGGAAACAACAAGGCGGCAAATCAACATGCCGAAGTATTAAC	1808
1809	AAATGTCTTCTAAGACTACAGTCAACCCACAGTAGATTGAACAATATGTGACTTTGAATGTCAGAATGTCAACTTTAAAGGGATTGCAA	1898
1899	AATATATATTTTTAAACTAAACTAATTAGGAATACAAGAGCTC	1942

*Act87E* SEQUENCE. Strain, Oregon R. Accession, X12452 (DROACT87EA), K00674 (DROACT87E).

in young adults. *Act79B* RNA is present in the various tubular-type muscles of the thorax: direct flight muscles, leg muscles and muscles that support the head and abdomen. *Act79B* transcripts are also present in muscles surrounding the male genitalia, but not in indirect flight muscles (Courchesne-Smith and Tobin 1989).

### *Act87E*

#### Gene Organization and Expression

Expected mRNA sizes range between 1,568 and 1,580 bases. The 5' end was determined by S1 mapping, by primer extension and by sequencing of several cDNA clones. Three poly(A) sites have been identified in five cDNA sequences. There is a leader intron with a donor site at -577 and an acceptor site at -20.



*Act88F*

-2066 TCTAGAATGCACAATAGGCAAATTTAGTTAAGATATGAATTTTTAAATAAATGGTGAGCCCAATCAATTCAGTGGTTGAATGACTTTTCA -1977  
 -1976 TAAATTAATAAATAAAGATAAAGATGGTGAACAATTCCTGTTTCGACGCCAATAACCTCTTGCTCAATACACGTGCAATCAAGGCAATCCA -1887  
 -1886 AATAAAACGCTTTGGGAATGCCACCAATTCACCTCCGAGCATCAGTTCTATCTTTAGCCAACCGATTTCGATTATTTTCATGTGGGCAAGC -1797  
 -1796 AATAAAACGTAATAAGAGAAGTAAAAATAATTAATCTACATAAAGGAATAATACAGTTCGATTGAGAAAAACATTTTCGCTCGG -1707  
 -1706 TCTGGCTGGCAATGGTTGGTTAATTGCACTGATAAATGGTCGGCAGGTGATTTGCAACTTCGGGATTGCATCGGCGCCGCAATGCAAAA -1617  
 -1616 GTGCAGCAGCATTCTGTAGAATGCGATTGCAAAATGGATGCAGCTTCTCGAGCACCGC GCGGAGATCTGATCAACCTTGCCTGTTG -1527  
 -1526 ATTTATCGGTGCCGCTCTGCTTGGCGCGTCTATTTTAGATTCCGCTCGCTGCGTGCCCGTTGAAATGTCCCAATTCCTCCAGTCCCTGCCG -1437  
 -1436 CGGATGCCAATGTCTTGCCTGGTCTTCTAAGGTCGGTTCCTATTTCCGAAGCTCTCAGCACCGAATGAGTCGTCGCGCCGACGAGT -1347  
 -1346 CGCCCATTTGGCAGCAGGATTGGGACAGAGATGGGACGGAGATGGGGCTAATGGCCGCTCGAGAGTCTGATTGCCGTTTAGGTGGCCC -1257  
 -1256 ATACACCCTATCACGCACCTCTGCTAATCACTCGGCTATGGCGTTCTCTTATCTTTTCGAGAGCTTTCTCTCTCGGCACTCCCTACAA -1167  
 -1166 ATAATGAATAGGGTCTAAGATTGATAGCTTACTTCCATCATATATGTCAATTAATTAATATTTTCAGGATTAATAATGAAACGAATT -1077  
 -1076 GAACATAAAGTTTCTACTACATAGTTATTTAAGCTGTTATATGTTATGAGACCATTTTCTCAGGATTTGTACCTACTAACAATGTGAAAA -987  
 -986 AAATATAAATTTGCATATTTTCGCAGTTTGGAAATTCCTCGTTTATTGAATTTATTGGTAATCTTAATAAATGATTCTATGCTTTATT -897  
 -896 AAGTATTTAATGTGTGGCTTCCTTTTTTTTGTGAAAGCGCATTAAATGAGTCGCTTCGTCGAATGAGGCATCAAACCTCTGACATG -807  
 -806 CTCGGCCAGAAGTCTGAAAACGCTTATATGGATCGGTTTCGAGTTGATTGTTCCGCAGCACTTTCGCTCAATCTTTTTCTCAGTGCCGCA -717  
 -716 CTGGCATCCAATCAAAATCGCTTCGAGGGAGAGCCGAGATATAAAAGGCCAGGACAGACCATCGGCGTGCCATTTGTTGTTGAATCTAGTT -627  
 -626 GTCAACAGGAATCGAAGCTGCGACTCTATCCAATTTTTCTCCTTTCGTTGACCTAAAGGTTGTGTGAGTGCACCTCAATGTCGAAGGAT -537  
 -536 CCAAGGATTATTACAGAAAAAGCCAGAGGACTAAGGATATTAACACTTTTTTAATAAGTTCGGATTGTTGATGGATTTTTCTACAAG -447  
 -446 TCACTAATCGGTCTTCGAAAGTCAATATCTAAATATAAAGTGAAGAGTAATTGCAACGAAACGATTTTCAATTAATTTGATACGTTTA -357  
 -356 AATTAAGTTCTATGAACATTTCTTTCCGATATTTTAGAGCACTGATTTAGTTTCAAGTGAATAACCAATTAGCATGACTCAAAGGAA -267  
 -266 ATGGAATATACCAATTTTGGCAATTTTTCATGGTTTTATTACTGAAATGTGCTCAAATGGACAATAGAGTTTCACCTCACTTCTCAAT -177  
 -176 ATCTTAAAAAGTTAAATATTTCTTGAGACACAAATAGTTTTCTATGTTGCTAATAAGTAGTAGAATTTAAGAATTTGAGATGTAGGT -87  
 -86 GGGAGCTATAAACTTTACATATATAATCGACAGATCGAGCTAACCGAGTGCCTTCCATCTCCCTTCAGATAAACAACTGCCAAGATG 3  
 Met (1)  
 4 TGTGACGATGATCGGGTGCATTAGTTATCGACAACGGATCGGGCATGTGCAAAAGCCGGCTTCGCGGTGATGACGCTCCCGTGCCTGTC 93  
 CysAspAspAspAlaGlyAlaLeuValIleAspAsnGlySerGlyMetCysLysAlaGlyPheAlaGlyAspAspAlaProArgAlaVal (31)

94	T T C C C C T C A A T T G T G G G T C G T C C C C G A C A C C A G G G T G T G A T G G T G G G T A T G G G T C A G A A G G A C T C G T A C G T G G G C G A C A G G C G C A A A G C	183
	PheProSerIleValGlyArgProArgHisGlnGlyValMetValGlyMetGlyGlnLysAspSerTyrValGlyAspGluAlaGlnSer	(61)
	A=KM88	
184	A A G C G C G G T A T C C T G A C G C T G A A G T A C C C C A T C G A G C A C G G C A T C A T C A C G A A C T G G G A C G A C A T G G A G A A G A T C T G G C A T C A C A C C T T C	273
	LysArgGlyIleLeuThrLeuLysTyrProIleGluHisGlyIleIleThrAsnTrpAspAspMetGluLysIleTrpHisHisThrPhe	(91)
	End	
274	T A C A A C G A G C T G C G C G T G G C C C C C G A G G A G C A T C C A G T A T T A T T G A C C G A G G C T C C A C T G A A C C C C A A G G C C A A T C G C G A G A A G A T G A C C	363
	TyrAsnGluLeuArgValAlaProGluGluHisProValLeuLeuThrGluAlaProLeuAsnProLysAlaAsnArgGluLysMetThr	(121)
364	C A G A T C A T G T T C G A G A C C T T C A A C T C G C C G G C C A T G T A C G T G C C A T C C A G G C C G T C T C C C T G T A C G C C T C C G G T C G T A C C A C C G G T	453
	GlnIleMetPheGluThrPheAsnSerProAlaMetTyrValAlaIleGlnAlaValLeuSerLeuTyrAlaSerGlyArgThrThrGly	(151)
454	A T T G T G C T G G A C T C C G G C G A T G G T G T C T C C A C A C C G T G C C A T C T A T G A G G G C T T C G C C T G C C C C A C G C C A T T C T G C G T C T G G A T C T G	543
	IleValLeuAspSerGlyAspGlyValSerHisThrValProIleTyrGluGlyPheAlaLeuProHisAlaIleLeuArgLeuAspLeu	(181)
544	G C T G G T C G C G A T C T G A C C G A T T A C C T G A T G A A G A T C C T G A C G G A G C G C G G C T A C A G C T T C A C C A C C A C C G C G A G C G T G A G A T C G T G C G C	633
	AlaGlyArgAspLeuThrAspTyrLeuMetLysIleLeuThrGluArgGlyTyrSerPheThrThrThrAlaGluArgGluIleValArg	(211)
634	G A C A T C A A G G A A A G C T G T G C T A C G T G G C T C T G G A C T T C G A G C A G G A G A T G G C C A C C G C T G C C G C C C A C C T C G C T G G A G A A G T C G T A C	723
	AspIleLysGluLysLeuCysTyrValAlaLeuAspPheGluGlnGluMetAlaThrAlaAlaAlaSerThrSerLeuGluLysSerTyr	(241)
724	G A G T T G C C T G A C G C C A G G T G A T C A C C A T T G G C A A C A G A G C G C T T C C G C T G C C C G A G G C C C T G T T C A G C C C T C G T T C C T G G G C A T G G A G	813
	GluLeuProAspGlyGlnValIleThrIleGlyAsnGluArgPheArgCysProGluAlaLeuPheGlnProSerPheLeuGlyMetGlu	(271)
814	T C G T G C G G C A T C C A C G A G A C C G T C A C A A C T C G A T C A T G A A G T G C G A C G T G G A C A T C C G C A A G G A T C T G T A T G C C A A C T C C G T G C T G T C C	903
	SerCysGlyIleHisGluThrValTyrAsnSerIleMetLysCysAspValAspIleArgLysAspLeuTyrAlaAsnSerValLeuSer	(301)
	>=DefKM129	
904	G G C G G T A C C A C C A T G T A C C T G G T A C A C G G A T C G T T C G T T C A G C A G T T G C A C T T G T G C T T A A T C C T T T G G T G C A C T T T C A G G T A T T G C C	993
	GlyGlyThrThrMetTyrProGlyIleAla	(311)
994	G A T C G T A T G C A G A A G G A G A T C A C T G C C C T G G C C C A T C G A C C A T C A A G A T C A A G A T A T T G C G C C A C C G A G A G G A A G T A C T C C G T C T G G	1083
	AspArgMetGlnLysGluIleThrAlaLeuAlaProSerThrIleLysIleLysIleIleAlaProProGluArgLysTyrSerValTrp	(341)
	.A=KM75	A=HH5
1084	A T C G G T G G C T C C A T C T G G C C T C G T G T C C A C C T T C A G C A G A T G G G A T C T C G A A G C A G G A G T A C G A C A G T C C G G C C C C G G A A T C G T T	1173
	IleGlyGlySerIleLeuAlaSerLeuSerThrPheGlnGlnMetTrpIleSerLysGlnGluTyrAspGluSerGlyProGlyIleVal	(371)
	End	Ser
1174	C A C C G C A A A T G C T T T T A A G T C T T C G C C C C G C G A A A A G C T C T T C A A A G G C A G C A A C C A G C A G C G A C C A C A A G C A T C C A T C G A C C T T A	1263
	HisArgLysCysPheEnd	(376)
1264	C C C A A C A A C C T C G G C T C G G A C A G T G A T A G A C A A A A G C A G C G A A C C C A T C G C G A C A A C A A T T A T C A T C C A A C T C A G A T T C A T A G C A G A T A A	1353
1354	T C A G A G G C A A C C T C G G T T G T C G G T G G T A T C T T A T G C A T T T C A T C G G C A G C G G T A T A G C G G A T T T T A T T T T G A A G A A C T A A T C G T A A T	1443
1444	C G T A A G A G T C G T G G T C T G C T C A G G	1467

Act88F SEQUENCE. Strain, Canton S. Accession, M18830 (DROACT88F), and M13925 (DROACT88H). There are several discrepancies among published sequences, even within the coding regions; these could be due either to natural polymorphisms

(continued)

Transcription is directed toward the telomere (*Act87E* Sequence) (Fyrberg et al. 1981; Manseau et al. 1988).

### *Developmental Pattern*

In embryos, transcripts are detectable in the developing musculature of the future larval body wall; the level of *Act87E* transcript is 5–10 times lower than for *Act57B* (Tobin et al. 1990).

## *Act88F*

### **Gene Organization and Expression**

The 5' end was determined by primer extension and by cDNA sequencing (Geyer and Fyrberg 1986; Okamoto et al. 1986). The 3' end has not been mapped. There is a leader intron with a donor site at –568 and an acceptor site at –15; there is another intron in the Gly-309 codon (*Act88F* Sequence) (Fyrberg et al. 1981; Sanchez et al. 1983).

### *Developmental Pattern*

Transcription is undetectable in embryos (Tobin et al. 1990); it increases during the first larval instar, peaks during the second instar and diminishes during the third instar and in prepupae. There is another larger peak of expression during pupation (Sanchez et al. 1983); at this stage, transcription is most prominent in the indirect flight muscles (Geyer and Fyrberg 1986).

### *Promoter*

Approximately 1,000 bp of 5' flanking DNA are sufficient for normal levels of RNA production and for complementation of the *raised* mutation (*rsd*). A putative enhancer element was identified between –1,565 and –1,286 (Geyer and Fyrberg 1986).

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(*continued*) or to sequencing errors; I report the results of Geyer and Fyrberg (1986) with the modifications of Mahaffey et al. (1985) and Okamoto et al. (1986). These seem to correspond to the more common allele in *Canton S*. The nature of several mutations are shown (Karlik et al. 1984; Okamoto et al. 1986).

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# 3

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## *Alcohol dehydrogenase: Adh, Adh-dup*

**Chromosomal Location:**  
2L, 35B2-3

**Map Position:**  
2-50.1

### **Product**

Alcohol dehydrogenase (ADH; alcohol:NAD<sup>+</sup> oxidoreductase, EC 1.1.1.1) (Grell et al. 1965).

### *Structure*

ADH is a homodimer with subunits of 27.4 kD; the polypeptide is 255 amino acids long with Acetyl-Ser at the amino terminus. There are two common allozymes, Slow (S) and Fast (F), that differ in electrophoretic mobility due to a threonine/lysine substitution at position 192.

Unlike the ADH of other species, *Drosophila* ADH does not use Zn<sup>++</sup> as a cofactor. Amino acid sequence comparisons reveal significant differences between *Drosophila* ADH on one hand and ADH from yeast or horse liver on the other (the latter two being quite similar); these observations suggest that the *Drosophila* protein is not homologous to other ADHs (Thatcher 1980; Benyajati et al. 1981). Rather, sequence comparisons show similarities between *Drosophila* ADH and *Klebsiella* ribitol dehydrogenase (Jörnvall et al. 1981). The evolution of ADH in the genus *Drosophila* has been discussed by Sullivan et al. (1990).

### *Function*

ADH is more active on alcohols of 3–5 carbons than on ethanol and more active on secondary than on primary alcohols (Sofer and Ursprung 1968).

### *Tissue Distribution*

ADH activity increases very rapidly from the second larval instar to immediately before pupariation; it declines during the pupal stages and increases again

for the first 4–5 days after emergence of the adult. In larvae, the enzyme is distributed approximately equally between fat bodies and midgut (although it is absent from the middle midgut). In adults, most of the activity is in the fat tissues, with much lower levels in the Malpighian tubules and the male reproductive system (Ursprung et al. 1970; Maroni and Stamey 1983).

### *Mutant Phenotype*

Null mutants are quite sensitive to a 5% ethanol solution. Even without an ethanol supplement, such mutants sometimes die as first instar larvae in cultures with very active yeast. *Adh* mutants, however, are more tolerant than wild-type flies to unsaturated secondary alcohols (O'Donnell et al. 1975).

### **Gene Organization and Expression**

Open reading frame, 256 amino acids; expected mRNA length, 1,071 bases (distal promoter) and 1,010 bases (proximal promoter). The different-sized transcripts carry the same open reading frame but different 5' untranslated regions (Benyajati et al. 1983). S1 mapping and primer extension sequencing of mRNA were used to determine 5' ends while S1 mapping and cDNA sequences defined the 3' end. Much of the extra length of the distal promoter transcript is in an intron with donor site at –690 and acceptor site at –35 (*Adh* Sequence and Fig. 3.1). *Adh* also has two small introns in the coding region. The first is after the codon corresponding to Lys-33 in the middle of the presumptive NAD<sup>+</sup>-binding domain and the second after the codon corresponding to Ala-168 near the boundary between the presumptive NAD<sup>+</sup>-binding and catalytic domains (Benyajati et al. 1981).

### *Developmental Pattern and Promoter*

The upstream, distal promoter is expressed primarily in adults while the proximal promoter is used during larval stages (Savakis et al. 1986). Two

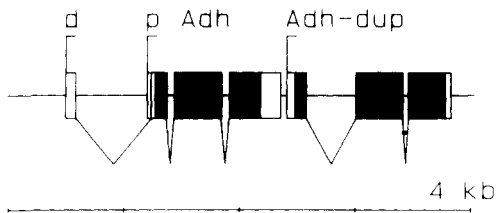


FIG. 3.1. Diagram of the organization and expression of *Adh* and *Adh-dup*

*Adh*

-1559 AGCTGCATTCGAAACCGCTACTCTGGCTCGGCCACAAGTGGGCTTGGTCGCTGTTGCGGACAAGTGAGATTGCTAATGAGCTGCTTTTA -1470

-1469 GGGGGCGTGTGTGCTTGCCTTCCAACCTTTCTAGATTGATTCTACGCTGCCTCCAGCAGCCACCCCTCCCATCCCCATCCCCATCACCA -1380

-1379 TCCAGTCCCCTGGCTCCCAGTCACAGTATTACACGTATGCAAATTAAGCCGAAGTCAATTGCGACCCGAGCAACAACACGATCTTTCT -1290  
 -----dep4=aef-1  
 -----

-1289 ACACTTCTCCTTGCTATGCTTGACATTCACAAGGCTCAAAGCTCTTAATATTCTGGCTTGTGCCCTACACTGTAAGAAATTACTATAGAA -1200  
 ---c/ebp -----dep1-2 -----dep3

-1199 ATAACGGTACACGGAATAAGATATTTTTTTAGTCCATATGCTTTTAACAAATGTGTTTTGAGTTTATGTTATATTGTTAGAAAACA -1110

-1109 GGTGTTTTTTTTAAATCGGTTAAAAAATACTACGAGAGAAAAATACAAATTTTGAAATAAGATTGACTCTTTTTCGATTTTGAATA -1020

-1019 TTTTCATTCATTTTATGTTTTACGTTTTCACTTATTTGTTTTCTCAGTGCACTTTCTGGTGTCCATTTTCTATTGGGCTTTTACCCCG -930

-929 CATTGTGTGCAGATCACTTGTTCGCGATTTTTATTGCATTTTACATATTACACATTATTGAACGCCGCTGCTGCTGCATCCGTCGAC -840  
 -----  
 --->-776

-839 GTCGACTGCACTCGCCCCACGAGAGAACAGTATTTAAGGAGCTGCGAAGGTCCAAGTACCAGATTATTGCTCAGTGCACTTGTGAGTT -750  
 -----d1 -----

-749 GCAGTTCAGCAGACGGGCTAACGAGTACTTGCATCTCTTCAAATTTACTTAATTGATCAAGTAGCAAAAAGGGCACCAATTAAGG -660  
 -----

-659 AAATCTTGTTAATTGAATTTATTATGCAAGTGCAGAAATAAAATGACAGTATTAATTAGTAAATATTTGTAAATCATATATAATCA -570

-569 AATTTATTCAATCAGAACTAATTCAAGCTGTACAAGTAGTGCAGACTCAATTAATTGGCATCGAATTAATAATTTGGAGGCTGTGCCGC -480

-479 ATATTCGCTTGGAAAATCACCTGTTAGTTAACTTCTAAAAATAGGAATTTTAAACATAACTCGTCCCTGTTAATCGGCGCCGTGCCCTCG -390

-389 TTAGCTATCTCAAAGCGAGCGCTGCAGACGAGCAGTAATTTCCAAGCATCAGGCATAGTTGGGCATAAATTATAAACATACAAACCG -300  
 -----p2

-299 AATACTAATATAGAAAAGCTTTGCCGGTACAAAATCCCAACAAAAACAACCGTGTGTGCCGAAAAATAAAAAATAACCATAAAGCTAG -210  
 -----

-209 GCAGCGCTGCCGTCCGGCTGAGCAGCTGCGTACATAGCCGAGATCGCGTAACGGTAGATAATGAAAAGCTCTACGTAACCGAAGCTT -120  
 -----p1 -----p0

-119 CTGCTGTACGGATCTTCTATAAAATACGGGGCCGACACGAACTGGAACCAACAACCTAACGGAGCCCTCTTCCAATTGAAACAGATCGAA -30  
 -----  
 --->-69

-29 AGAGCCTGCTAAAGCAAAAAGAAGTACCATGTCTGTTACTTTGACCAACAAGAACGTGATTTTCGTTGCCGGTCTGGGAGGCATTGGT 60  
 MetSerPheThrLeuThrAsnLysAsnValIlePheValAlaGlyLeuGlyGlyIleGly (20)  
 Asp

61 CTGGACACCAGCAAGGAGCTGCTCAAGCGGATCTGAAGGTAACATGCGATGCCACAGGCTCCATGCAGCGATGGAGGTTAATCTCGT 150  
 LeuAspThrSerLysGluLeuLeuLysArgAspLeuLys (33)  
 .CGATC|-def|=fn6 |-

(continued)



	def .- G=fn4 . . . . . A=UF . . . . . A=F'	
151	GTATTCAATCCTAGAACCCTGGTGATCCTCGACCATTGAGAACCCGGCTGCCATTGCCGAGCTGAAGCAATCAATCCAAGGTGACCG AsnLeuVal11leLeuAspArgIleGluAsnProAla1a1leAlaGluLeuLysAla1leAsnProLysVal1ThrV	240 (59)
	Asp . . . . . Glu	
	- def - =fn24 . . . . . T=n4	
241	TCACCTTCTACCCCTATGATGTGACCGTGCCATTGCCGAGACCACCAAGCTGCTGAAGACCATCTTCGCCAGCTGAAGACCGTGCATG a1ThrPheTyrProTyrAspVal1ThrVal1ProIleAlaGluThrThrLysLeuLeuLysThrIlePheAlaGluLeuLysThrVal1AspV	330 (89)
	Ter	
331	TCCTGATCAACGGAGCTGGTATCCTGGACGATCACCAGATCGAGCGCACCATTGCCGCAACTACACTGGCCTGGTCAACACCACGACGG a1Leu1leAsnGlyAlaGlyI1leLeuAspAspHisG1n1leGluArgThrI1leAlaVal1AsnTyrThrGlyLeuVal1AsnThrThrA	420 (119)
421	CCATTCTGGACTTCTGGACAAGCGCAAGGGCGGTCCCGTGGTATCATCTGCAACATTGGATCCGCTACTGGATTCAATGCCATCTACC 1a1leLeuAspPheTrpAspLysArgLysGlyGlyProGlyGlyI1le1leCysAsn1leGlySerVal1ThrGlyPheAsnAla1leTyrG	510 (149)
511	AGGTGCCCGTCTACTCCGGCACCAGGCCCGCTGGTCAACTTCACCAGCTCCCTGGCGGTAAGTTGATCAAAGGAAACGCAAGTTTTC 1nValProValTyrSerGlyThrLysAlaAlaVal1Val1AsnPheThrSerSerLeuAla	600 (168)
601	AAGAAAAACAACAACTAATTTGATTATAACACCTTTGAAAACCTGGCCCCATTACCGCGTGACCGCTTACACCGTGAACCCCGGCATC LysLeuAlaProIleThrGlyVal1ThrAlaTyrThrVal1AsnProGlyI1e	690 (185)
	C=F . . . . . T=F	
691	ACCCGCACCACCCTGGTGACAAAGTTCAACTCCTGGTGGATGTTGAGCCCCAGGTTGCTGAGAAGCTCCTGGCTATCCACCACGACCA ThrArgThrThrLeuVal1HisLysPheAsnSerTrpLeuAspVal1GluProGlnVal1AlaGluLysLeuLeuAlaHisProThrGlnPro	780 (215)
	Thr . . . . . Ser	
	. . . . . A=D . . . . . A=nB . . . . .   -	
781	TCGTTGGCCTGCGCCGAGAACTTCGTCAAGCTATCGAACTGAACAGAACGAGCCATCTGAAACTGGACTTGGGCACCTGGAGGCC SerLeuAlaCysAlaGluAsnPheVal1LysAla1leGluLeuAsnGlnAsnGlyAla1leTrpLysLeuAspLeuGlyThrLeuGluAla	870 (245)
	Glu Ter	
	def .- =fn23	
871	ATCCAGTGGACCAAGCACTGGGACTCCGGCATCTAAGAAGTGATAATCCCAAAAAAAAAACATAACATTAGTTCATAGGTTCTGCGAAC 1leGlnTrpThrLysHisTrpAspSerGlyI1eEnd	960 (256)
961	CACAAGATATTCACGCAAGGAATAAGGCTGATTCGATGCACACTCACATTCTCTCCTAATACGATAATAAAACTTTCATGAAAAATA	1050
	-----	
1051	TGGAAAAATATATGAAAATGAGAAATCCAAAAAAGCTGATAAACGCTCTACTTAATATAATAGATAAAATGGGAGCGGCAGGAATGGCGG  (A) <sub>n</sub> -----	1140 -->1132 ( <i>Adh-dup</i> )
1141	AGCATGGCCAAGTTCCTCCGCAATCAGTCGTA AACAGAAAGCTGGAAGCGGATAGAAAGAAATGTTGATTTGACGGGCAAGCATGT MetPheAspLeuThrGlyLysHisVa	1230
1231	CTGCTATGTGGCGGATGCGGAGGAATTGCACTGGAGACCAGCAAGGTTCTCATGACCAAGAATATAGCGGTGAGTGAGCGGGAAGCTCG 1CysTyrVal1AlaAspCysGlyGlyI1leAlaLeuGluThrSerLysVal1LeuMetThrLysAsn1leAla	1320
1321	GTTTCTGTCCAGATCGAACTCAAACCTAGTCCAGCCAGCTGCTGTGCAAACATAATTAAGTAAATGAGTTTTTCATGTTAGTTTCCGCTG	1410
1411	AGCAACAATTAAGTTTATGTTTCAGTTCGG	1440

*Adh* SEQUENCE. Slow allele from *Canton S*. Accession M14802 (DROADHA). Several other alleles have been sequenced and are listed under DROADH\* in GenBank. Several mutations are indicated (Benyajati et al. 1982; Martin et al. 1985; Place et al. 1987; Thatcher 1980). Indicated under the sequence in the promoter regions are binding sites for various regulatory proteins. For the *Adh-dup*, initiation of transcription and translation, at 1,132 and 1,205, respectively, are suggested by sequence comparison to *Adh* (Schaeffer and Aquadro 1987).

enhancers that control expression of the two promoters were identified (Posakony et al. 1985):

*Larval Enhancer and Proximal Promoter* The larval enhancer is located between 5,000 and 1,845 bp upstream of the distal transcription initiation site; it can stimulate transcription from the proximal (but not the distal) promoter at all developmental stages (Corbin and Maniatis 1989a).

In the proximal promoter, three protein-binding regions were identified ( $p_0$ ,  $p_1$  and  $p_2$  between  $-340$  and  $-140$  in the *Adh* Sequence) (Heberlein et al. 1985). Functional assays of promoter deletions demonstrated that those are the only regions in the neighborhood of the proximal promoter necessary for expression (Shen et al. 1989, 1991).

*Adult Enhancer and Distal Promoter* The adult enhancer is located between 600 and 450 bp upstream of the distal transcription initiation site (approximately  $-1,375$  and  $-1,225$  in the *Adh* Sequence); it stimulates transcription from both promoters but only during the late third larval instar and in adults (Corbin and Maniatis 1989a).

DNA-binding assays and *in vitro* transcription experiments defined a *cis*-acting region that extends from  $-860$  to  $-820$  as necessary for transcription from the distal promoter; a specific factor, ADF-1 (*Adh* distal factor 1), binds to this region ( $d_1$  in the *Adh* Sequence) (Heberlein et al. 1985; England et al. 1990). In addition, a general transcription factor similar to human transcription factor SP2 is required (Heberlein et al. 1985).

Four distal enhancer binding proteins were obtained from cultured-cell nuclear extracts (DEP1–4) (*Adh* Sequence). DEP1 and DEP2 have partly overlapping binding sites (*dep1* and *dep2*) in a segment that is required for full expression. DEP1 is FTZ-F1, a member of the steroid hormone receptor superfamily also involved in the control of the *fushi tarazu* (*ftz*) “zebra element” (Ayer and Benyajati 1992). The site *dep4*, also called *aef-1*, was identified as the binding site of a repressor (Falb and Maniatis 1992). Partly overlapping *aef-1* is a binding site for mammalian C/EBP, and the authors suggest that the *Drosophila* homolog of C/EBP acts to stimulate transcription in fat body; competition between C/EBP and AEF-1 (=DEP4?) would determine the level of transcriptional activity. Overlapping C/EBP and QEF-1 binding sites were found in the regulatory sequences of another gene expressed in fat body, *Yp1*, one of the yolk protein genes (Falb and Maniatis 1992).

Down-regulation of the proximal promoter in adults is dependent on expression of the distal promoter, an apparent instance of transcriptional interference (Corbin and Maniatis 1989b). Transcriptional interference and the stage and promoter specificity of the two enhancers could explain the major promoter switch that occurs between larval and adult stages (Corbin and Maniatis 1989b).

*Adh-dup*

The putative 5' end of this gene is positioned very near the 3' end of *Adh* and probably originated as a duplication (*Adh* Sequence; Fig. 3.1). It is present in other *Drosophila* species (including those of the *pseudoobscura* group). The amino acid sequence of the two genes is approximately 38% identical, and the coding region introns are similarly positioned. The nature or function of the product is not known (Schaeffer and Aquadro 1987; Kreitman and Hudson 1991).

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# 4

## The $\alpha$ -Amylase Genes: *AmyA*, *AmyB*

**Chromosomal Location:**  
2R, 54A

**Map Position:**  
2-77.7

**Product**  
 **$\alpha$ -Amylase (EC 3.2.1.1)**

### *Structure and Function*

$\alpha$ -Amylase is a monomeric enzyme of  $M_r$  54.5 kD, which acts in the hydrolysis of starch. The mature protein is thought to be 476 amino acids long, with its N terminus, a derivatized Gln, being the 19th amino acid of the translation product. The first 18 amino acids of the translation product are thought to constitute the transport signal peptide. There is 55% identity between *Drosophila*  $\alpha$ -amylase and  $\alpha$ -amylase of the mouse pancreas (Fig. 4.1) (Boer and Hickey 1986).

### *Tissue Distribution*

$\alpha$ -Amylase is most abundant in the midgut where it occurs in characteristic patterns under the genetic control of the *map* gene (Doane et al. 1975, 1983).

### **Organization of the Cluster**

There are two divergently transcribed *Amy* genes separated by approximately 3.7 kb (Fig. 4.2). *AmyA* is the centromere proximal gene and *AmyB* the centromere distal one (Levy et al. 1985). The duplicated segments extend from approximately 130 bp upstream of the translation initiation site to the polyadenylation site. Within this region, divergence between the two genes is low in the coding region (the frequency of silent substitutions is ca. 1%) but it is considerable upstream and downstream of the coding region (frequency of substitutions, 30%). This observation led to the suggestion that gene conversions

```

1                               50                               100
Dm MFLAKSIVCL ALLAVANAQF DTNYASGRSG MVHLFEWKWD DIAAECENFL GPNYGAGVQV SPVNEHAV.. KDSRPWERY QPISYKLETR SGNEEQFASM
Mouse ...MKFWLLL SLIGFCWAQY DPHTSDGRTA IVHLFEWRWV DIAKECERYL APKGFGGVQV SPPNENVVVH NPSRPWERY QPISYKICTR SGNEDEFROM
CON ----K----L -L-----AQ- D-----GR-- -VHLFEW-W- DIA-ECE--L -P-G--GVQV SP-NEN-V-- --SRPWERY QPISYK--TR SGNE--F--M

101                               150                               200
Dm VKRCNAVGVVR TYVDVFNHM AADG..GTY GTGGSTASPS SKSYPGVPYS SLDFN...PT CAISNYNDAN EVRNCLEVLG RDLNQGNSYV QDKVVEFLDH
Mouse VTRCNVGVVR IYVDVINHM CGAGNPAGTS STCGSYLNPN NREFPAVPYS AWFNDNKCEN GEIDNYNDAY QVRNCRLTGL LDLALEKDYV RTKVADYMMNH
CON V-RCN-VGVVR -YVD-V-NHM ---G---GT- -T-GS---P- ----P-VPYS --DFN----- --I-NYNDAN- -VRNC-L-GL -DL-----YV --KV-----H

201                               250                               300
Dm LIDLGVAGFR VDAAKHMWPA DLAVIYGRLE NLNTDHGFAS GSKAYIVQEV IDMGGEAISK SEYTGGLGAIT EFRHSDSIGK VFR..GKDQL QYLTNWGTAW
Mouse LIDIGVAGFR LDAAKHMWPR DIKAVLDKLE NLNTKW.FSQ GSRPFIFQEV IDLGGEAIKG SEYFGNGRVT EFKYGAKLGT VIRKWNGEKM SYLKNWGEGW
CON LID-GVAGFR -DAAKHMWP- D-----L- NLNT---F-- GS---I-QEV ID-GGEAI-- SEY-G-G--T EF-----G- V-R----- -YL-NWG--W

301                               350                               400
Dm GFAASDRSLV FVDNHDNQRG HGAGGADVLT YKVPKQYKMA SAFMLAHPFG TPRVMSSFSF .....TDTQ ....GPPTTD GHNIASPIFN SDNSCSGGWV
Mouse GLVPSDRALV FVDNHDNQRG HGAGGSSILT FWDARMYKMA VGFMLAHPYG FTRVMSSYRW NRRNFQNGKQD NDWIGPPNNN GVTKEVTI.N ADITTCGNDWV
CON G---SDR-LV FVDNHDNQRG HGAGG---LT -----YKMA --FMLAHP-G --RVMSS--- -----DQ ----GPP--- G-----I-N -D--C---WV

401                               450                               500
Dm CEHRWRQIYN MVAFRNTVGS DEIQNWWDNG SNQISFSRGS RGFVAFNNDM YDLNSSLQTG LPAGTYCDVI SGSKSGSSCT GKTVTVGS DG RASINIGSSE
Mouse CEHRWRQIRN MVAFRNVVNG QPFSNWDNN SNQVAFSRGN RGFIVFNDD WALSATLQTG LPAGTYCDVI SGDKVDGNCT GLRVNVGSDG KAHFISISNSA
CON CEHRWRQI-N MVAFRN-V-- ---NWDN- SNQ--FSRG- RGF--FNDD- --L---LQTG LPAGTYCDVI SG-K-----CT G--V-VGSDG -A---I--S-

501                               514
Dm DDGVLAIHVN AKL*
Mouse EDPFIAIHAD SKL*
CON -D---AIH-- -KL-

```

FIG. 4.1. Comparison of the mouse (Accession, V00718) and *Drosophila* (Dm) *AmyA* sequences. There is 55% overall identity between the two proteins. Sequences aligned with the GCG *Pileup* program.

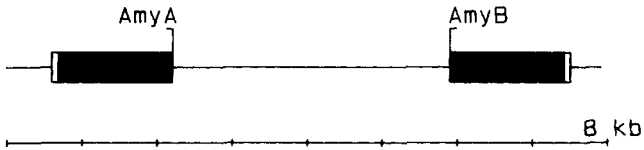


FIG. 4.2. The two *Amy* genes.

in the coding regions maintain a high degree of conservation (Hickey et al. 1991).

## *AmyA*

### Gene Organization and Expression

Open reading frame, 494 amino acids; predicted mRNA length, 1,601 bases. The 5' end was determined by primer extension and the 3' end from the sequence of one cDNA clone. There are no introns (Boer and Hickey 1986) (*AmyA* and *AmyB* Sequence).

### *Developmental Pattern*

The methods used do not distinguish between *AmyA* and *AmyB* RNA. *Amy* transcription is subject to glucose repression: larvae grown in 10% glucose accumulate only 1% as much *Amy* mRNA as larvae grown in the absence of glucose (Benkel and Hickey 1987).

### *Promoter*

An *AmyA* segment that extends from  $-142$  to  $-50$  in the *Amy* Sequence is sufficient to drive the glucose suppressible expression of *Adh* as a reporter gene. Deletion analysis showed that elements between  $-142$  and  $-125$  are required for full gene expression and that the sequences necessary for glucose repression are between  $-125$  and  $-50$  (Magoulas et al. 1992). Upstream *Amy* sequences have similarities with *cis*-acting elements that mediate glucose repression in yeast (Boer and Hickey 1986), and the *Drosophila AmyA* promoter is subject to glucose repression when introduced into yeast cells (D. A. Hickey, personal communication). Linker scanning mutations were used to identify functional CAAT and TATA boxes (Magoulas et al. 1992).

Amy

```

A T ----- AGCG GT A T AAAA TC TTGC TA A T GCAA TC AG GTG TACATG TTAC G TGGT T
-565 CACTTCAGAACCCAGAGATCAAGTGCCGCCAGTCAAGGCCAGAAGTACAGTATTCAGAGAAGCGCGCAGCCAAAGCTTCAACAAAAA -476

A ATA TGAA A TT AC TATAA C CCA T AC TGCA TA GTG A ATTAG C AT T TCCTTG
-475 TCGCTTGCTACCTTTATTTCAACATTTTTAGGCGATATTGCATGATTTCAATGCTTTCAAAATACGCTAAAAAATCCAATAAC----- -386

TAGGCCAA G G T G T A - TG C A --C G G C CT TATC A A TG
-385 -----AATTCACAGTAAACC CGCTCTAGGAGCGTGAACGTAAATAAATAGTCAATAAATCCCAACTGAAACC GATTTCAAAGGAAT -296

G TT -T C T GTCG AC TT T T AC AGT A TAGG T -----C G CT G T
-295 GCATTTTCCCGATGAGTTATTGATACAAATATAACGAAAAATAAGCCGACTCACTAATCATCAGCGAAAAATTGCGATCTCCAGTCAATAC -206

AA ACGC TT C C A TC ----- ATA AT CAA A CGT G GAC TA G -----A T
-205 GTCTGCTCGGAATTGTGATTTGACAAACTAATCGCCAGTCAGACCCCATGCGTGAAAAAACCCCTTAGGGAGCGATAAGATCCCATGCAG -116

. . . . . -->-32
C G G A -----GAATAGGT T TCATC C T A GACAC C TTA T
-115 TCACAAACTACTCCCCGGAAGCCCTCAGATAAAGTAGCAGTGGGGTCCACTATATAAGGAGCGGC-TCTGAGTAGTCCGACCAGAGTG -26

-----
T T G C AA ||G=null-d
-25 AAAGTGAATCCATCCTGGAATCATCATGTTCTTGCCAAAGACATAGTGTGCCTCGCCCTCTGGCGGTGGCCAACGCCAATTCGACA 64
MetPheLeuAlaLysSerIleValCysLeuAlaLeuLeuAlaValAlaAsnAlaGlnPheAspT (22)

. . . . . G G
65 CCAACTACGCATCCGGTCGTAGTGGAAATGGTCCACCTCTTCGAGTGGAAAGTGGGACGACATCGCTGCCGAGTGCGAAAACTTCTCTGGAC 154
hrAsnTyrAlaSerGlyArgSerGlyMetValHisLeuPheGluTrpLysTrpAspAspIleAlaAlaGluCysGluAsnPheLeuGlyP (52)

. . . . . G A
155 CCAATGGCTACGCGGTGTTCAAGTCTCCCTGTGAACGAGAACCGCGTCAAGGACAGCCGCCCTGGTGGGAACGTTACCAGCCATCT 244
roAsnGlyTyrAlaGlyValGlnValSerProValAsnGluAsnAlaValLysAspSerArgProTrpTrpGluArgTyrGlnProIleS (82)
Arg

. . . . . G
245 CCTACAAGCTGGAGACCCGCTCCGGAACGAAGAGCAGTTCCGCCAGCATGGTCAAGCGCTGCAACGCCGTGGAGTGGCACCTACGTGG 334
erTyrLysLeuGluThrArgSerGlyAsnGluGluGlnPheAlaSerMetValLysArgCysAsnAlaValGlyValArgThrTyrValA (112)

. . . . . G A=null-d
335 ACGTGGTCTTCAACCACATGGCCGCGCAGGAGGCACCTACGGCAGTGGCGGCAGCACC GCCAGCCAGCAGCAAGAGCTATCCGGAG 424
spValValPheAsnHisMetAlaAlaAspGlyGlyThrTyrGlyThrGlyGlySerThrAlaSerProSerSerLysSerTyrProGlyV (142)
Gly End

. . . . . C=Canton S G
425 TGCCCTACTCTCGTGGACTTCAACCCGACCTGCGCCATCAGCAACTACAACGACGCCAACGAGGTGCGCAACTGCGAGCTGGTCCGGTC 514
alProTyrSerSerLeuAspPheAsnProThrCysAlaIleSerAsnTyrAsnAspAlaAsnGluValArgAsnCysGluLeuValGlyL (172)
His Arg

. . . . . C A=Canton S C
515 TGCGCGACCTTAACCGAGGCAACTCTACGTGCAGGACAAGTGGTTCGAGTTCCTGGACCATCTGATTGATCTCGCGGTGGCCGGATTCC 604
euArgAspLeuAsnGlnGlyAsnSerTyrValGlnAspLysValValGluPheLeuAspHisLeuIleAspLeuGlyValAlaGlyPheA (202)
Asn

```

(continued)



	null-p=A      T      G	
605	GCGTGGACGCCGCCAAGCACATGTGGCCCGCCGACTGGCCGTCATCTATGGCCGCTCAAGAACCTAAACACCGACCACGGCTTCGCCT rgValAspAlaAlaLysHisMetTrpProAlaAspLeuAlaValIleTyrGlyArgLeuLysAsnLeuAsnThrAspHisGlyPheAlaS	694 (232)
	End	
	A	
695	CGGGATCCAAGGCGTACATCGTCCAGGAGGTCATCGACATGGCGGCGAGGCCATCAGCAAGTCCGAGTACACCGGACTGGCGCCATCA erGlySerLysAlaTyrIleValGlnGluValIleAspMetGlyGlyGluAlaIleSerLysSerGluTyrThrGlyLeuGlyAlaIleT	784 (262)
	A      T	
785	CCGAGTTCGCCACTCCGACTCCATCGGCAAGGCTTCCGCGGCAAGGACCAGTGCAGTACCTGACCAACTGGGACCCGCCCTGGGGCT hrGluPheArgHisSerAspSerIleGlyLysValPheArgGlyLysAspGlnLeuGlnTyrLeuThrAsnTrpGlyThrAlaTrpGlyP	874 (292)
	Asn	
	C      G	
875	TCGCTGCCTCCGACCGCTCCCTGGTATTCTGTGCACAACCACGACAATCAGCGGGACATGGAGCAGGAGGCGCCGACGTTCTGACCTACA heAlaAlaSerAspArgSerLeuValPheValAspAsnHisAspAsnGlnArgGlyHisGlyAlaGlyGlyAlaAspValLeuThrTyrL	964 (322)
	A      T      C      T	
965	AGGTGCCCAAGCAGTACAAGATGGCTCCGCCCTTACGTGGCGCACCCCTTCGGCACTCCCCGCGTGTGTCTCTCTCTTCCTTCACGG ysValProLysGlnTyrLysMetAlaSerAlaPheMetLeuAlaHisProPheGlyThrProArgValMetSerSerPheSerPheThrA	1054 (352)
1055	ACACCGATCAGGGCCCGCCACCACCGACGGCCACAACATCGCCTCGCCCATCTTCAATAGCGACAACCTCTGCAGCGGGCTGGGTGT spThrAspGlnGlyProProThrThrAspGlyHisAsnIleAlaSerProIlePheAsnSerAspAsnSerCysSerGlyGlyTrpValC	1144 (382)
	C      G      G      C	
1145	GTGAGCACCGCTGGCGCCAGATCTACAACATGGTGGCTTCCGAAACACCGTGGGCTCGGACGAGATCCAGAACTGGTGGGACAACGGCA ysGluHisArgTrpArgGlnIleTyrAsnMetValAlaPheArgAsnThrValGlySerAspGluIleGlnAsnTrpTrpAspAsnGlyS	1234 (412)
	Ala      Ala	
1235	GCAACCAGATCTCTTTCAGCCGAGGACGCCGGCTTCGTGGCCTTCAACAACGACAACCTACGACTGAACAGCTCCCTGCAGACGGGCC erAsnGlnIleSerPheSerArgGlySerArgGlyPheValAlaPheAsnAsnAspAsnTyrAspLeuAsnSerSerLeuGlnThrGlyL	1324 (442)
	C	
1325	TGCCCGCCGGCACCTACTGCGAGTCTATCCGGCTCCAAGAGCGGTTCCTCTGCAGGGCAAGACCGTACCCTCGGATCCGACGGAC euProAlaGlyThrTyrCysAspValIleSerGlySerLysSerGlySerSerCysThrGlyLysThrValThrValGlySerAspGlyA	1414 (472)
	A      CAAAGACCA	
1415	GGGCTTCCATCAACATTGGCAGCTCCGAGGACGACGGAGTGTGGCCATTACGTC AACGCCAAGTTGTAACAGCTGGGG.....AGC rgAlaSerIleAsnIleGlySerSerGluAspAspGlyValLeuAlaIleHisValAsnAlaLysLeuEnd	1504 (494)
	G C   GA   GA   T      C   - TTA      T C G      A A AGGAAGA G GC	
1505	ATGGCGAACAGCCAGGCAATTAATTGAGATTATAAATGTACGAAATATATATGATGAGATTATAAACACACAACTTTTATTCGCAAG -----   (A) <sub>n</sub>	1594
1595	TA C GT CA T ATGGA AATG AAAT TTAT TACTTAAAATTGACCACAATAACTGTTACGCATAATATGGCAAAAAAC GGATGATAAGACTAATATATATATTATCTGGGCTAAGCTGA-----	1684
1685	AACTTATGCGTGACCTTAAAAGCGTGCCTTTTCATCTCGGTATTCAGCGTGATT ----- 1739	

*AmyA* AND *B* SEQUENCE. The sequence on the numbered line corresponds to the proximal gene (*A*) of *Oregon R* (allele *Amy*<sup>1</sup>). This sequence combines the nonoverlapping regions of two GenBank entries: Accession X04569 (DROAMYAG1)

(continued)

*AmyB***Gene Organization and Expression**

Open reading frame, 494 amino acids; predicted mRNA length, 1,606 bases. The 5' and 3' ends of *AmyB* were deduced from sequence similarity to *AmyA* (Okuyama and Yamazaki 1988; D. A. Hickey, personal communication) (*AmyA* and *AmyB* Sequence).

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(continued) and Accession Y00438 (DROAMYAR). On the line immediately above is the sequence of the distal gene of strain *Makokou*; only in those positions where there is a difference from the proximal gene is the base indicated. There are differences in six amino acid residues between these two sequences. In four of those six positions (Gly-121, Arg-156, Asn-278 and Ala-398), the *Makokou* proximal gene (not shown) has the same residue as the *Makokou* distal gene, reinforcing the idea that there is intergenic correction between these genes (Hickey et al. 1991). The *Makokou* sequences were kindly provided by Donal A. Hickey. A *Canton S* allele with two amino acid substitutions (Tyr-144 and Tyr-181) has the same electrophoretic mobility as *AmyA*<sup>1</sup>. An *Amy*-null strain has two mutations in the distal gene, the addition of a G between positions 3 and 4, and a nonsense mutation at position 375 and one mutation in the proximal gene, with a nonsense mutation at position 654. This null strain apparently also has an inversion within the intergenic segment (Okuyama and Yamazaki 1988). The vertical bar marks the end of the signal peptide.

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# 5

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## The Andropin and Cecropins Gene Cluster: *Anp*, *CecA1*, *CecA2*, *CecB*, *CecC*

**Chromosomal Location:**  
3R, 99E

**Map Position:**  
3-[101]

### **Products**

Antibacterial peptides.

### *Structure*

Sequence analysis suggests that each polypeptide may fold into two amphipathic  $\alpha$ -helices separated by a four-amino-acid loop (Samakovlis et al. 1991).

By analogy to the better-characterized cecropins of the moth *Hyalophora cecropia*, processing is predicted to include the removal of the signal peptide and of an additional dipeptide at the N-terminus, and cleavage of the terminal Gly plus amidation at the C-terminus. These changes would give rise to mature cecropins 39 amino acids long (Kylsten et al. 1990, see Sequences).

Cecropins A1 and A2 are identical to each other and to the main cecropin from the flesh fly *Sarcophaga peregrina*. Cecropin B differs from A1 and A2 by four conservative substitutions in the mature protein (Arg-27, Ile-36, Ser-44 and Val-47) and four others in the signal peptide (Kylsten et al. 1990). Cecropin C is intermediate in sequence between A and B (Fig. 5.1) (Tryselius et al. 1992). The sequence similarities between andropin and the cecropins is restricted to the signal peptide (Samakovlis et al. 1991).

### *Tissue Distribution and Function*

For the most part, cecropins are synthesized in response to bacterial infection and released in the hemolymph. Cecropins disrupt the cell membrane of Gram-positive and Gram-negative bacteria (Dunn 1986; Boman and Hultmark 1987). The related andropin is synthesized constitutively in the ejaculatory duct of males (Samakovlis et al. 1991).

	1					50		63
Anp	MKYFVVLVVL	ALILAISVGP	SDAVFIDILD	KVENAIHNAA	QVGIGFAKPF	EKLINPK*..	....	
CecB	MNFNKIFVFV	ALILAISLGN	SEAGWLRKLG	KKIERIGQHT	RDASIQVLGI	AQQAANVAAT	ARG*	
CecC	MNFYKIFVFV	ALILAISIGQ	SEAGWLKCLG	KRIERIGQHT	RDATIQLGI	AQQAANVAAT	ARG*	
CecA	MNFYNIFVFV	ALILAITIGQ	SEAGWLKKG	KKIERVQHT	RDATIQLGI	AQQAANVAAT	ARG*	
CON	MNF--IFVFV	ALILAI--G-	SEAGWL-K-G	K-IER-GQHT	RDA-IQ-LGI	AQQAANVAAT	ARG-	
				^^				

FIG. 5.1. Aligned cecropin and andropin peptide sequences. The vertical line under Ser-21 marks the last amino acid of the signal peptide, and, under Ala-23, the dipeptidase cleavage site. A caret marks the intron positions. The CON(sensus) line indicates positions in which three of the four sequences agree.

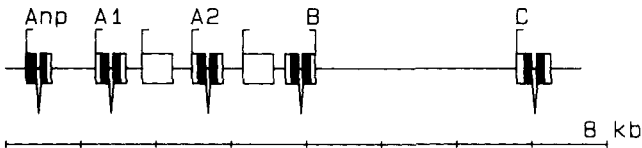


FIG. 5.2. The *Cecropin* cluster. Open boxes indicate the two pseudogenes.

## Organization and Expression of the Cluster

Five genes and two pseudogenes are clustered in approximately 8.0 kb of DNA (Fig. 5.2). The pseudogenes contain vestiges of exons, introns and TATA boxes; but they also include numerous nonsense mutations, and they have lost the splicing signals.

### Developmental Pattern

Transcription of *CecA*, *CecB*, and *CecC* is induced by injection or feeding of bacterial pathogens. Cecropin mRNAs, undetectable before infection, begin to accumulate 1 h after injection of bacteria, reach a maximum 2–6 h after injection, and soon thereafter they begin to decline. Twenty-four hours after injection, the RNAs return to their basal levels. *A1* and *A2* are expressed at high level in larval, pupal and adult stages. Transcription occurs primarily in the fat tissue, and the proteins accumulate in the hemolymph. *B* and *C*, by contrast, are inducible to a much lower extent than *A1* and *A2* in larvae and adults. They are active mainly during the early pupal stages in localized regions of tissues undergoing lysis, and this activation of *B* and *C* occurs in the absence of external agents (Kylsten et al. 1990; Tryselius et al. 1992).

AT-rich segments in the 3' untranslated region of the mRNAs may play a role in their selective degradation (Kylsten et al. 1990).

*Anp*

```

-287 TAACCTACAGAATTGTAGAACCTTAATTAATCTATAGAACACTATTGAATGAAAACCTTAGTAACCTTGTGAGGTTTTAGTAATTCCAAGAAA -198
-197 TATGCTCTTGAATAAAAAACCTTTTTAAGTCTCTTTCAATGCAAAAACACGAGTCTTTTTTTTTTACATATTGTAATTAATATGTTAAG -108
-107 GTCTAATTATTATTGTAACGTTTTTCGGTGGGTTGATTGCCTATAAAGCCACTTGTTTTTTCAGTCTAAATCATCAGTGTAATAATTCGGA -18
-17 AAACCCAGCGATCTAGTTATGAAATACCTTTGTGGTCCCTGTCGTCCTGGCCCTCATTTTGGCCATCAGCGTGGGTCCTTCGGATGCAGTA 72
MetLysTyrPheValValLeuValValLeuAlaLeuIleLeuAlaIleSerValGlyProSerAspAlaVal (24)
73 TTTATTGATATTCTTGACAAAGTGGTTTGTCTTCTTTAAACAATTGTAGTTTACAATGAAGCTTAAACATTTGTATTCTACAGGAAA 162
PheIleAspIleLeuAspLysVal GluA (34)
163 ACGCAATACACAATGCTGCTCAAGTGGGAATTGGCTTTGCTAAGCCCTTTGAAAAATTGATCAATCCGAAGTAATCTGCACTGCAATTT 252
snAlaIleHisAsnAlaAlaGlnValGlyIleGlyPheAlaLysProPheGluLysLeuIleAsnProLysEnd (57)
253 AATTAATGTATCGTTAACGAAAATAACACAAAATTTAAAATCTGAAAAACAATAAGTTACTAACGCAAGACTTTTAGTTAAGTTAGT 342
343 TAATATAGACCGAGATGTATGTACATACATACCCTTTCGCTTACAATAAAATGTTAAATAAGTTTTAGATTCAGATTCGTCAGTCTCAGTAAA 432
    
```

*Anp* SEQUENCE. Strain, *Canton S*. Accession, X16972 (DROCECPN).

*Anp*

**Gene Organization and Expression**

Open reading frame, 57 amino acids; expected mRNA length, 278 bases. Primer extension and sequence features were used to identify two 5' ends, the upstream site being the major one. The 3' end was obtained from a cDNA sequence. There is an intron after the Val-32 codon (*Anp* Sequence) (Samakovlis et al. 1991).

*Developmental Pattern*

Transcription is restricted to the ejaculatory ducts. mRNA level reaches a plateau 24 h after eclosion of the adult male and remains stable in virgin males; mating, however, causes a rise in the steady-state level of *Anp* mRNA (Samakovlis et al. 1991).

*CecA1*

**Gene Organization and Expression**

Open reading frame, 63 amino acids; expected mRNA length, 346 bases, in agreement with the 0.4 kb RNA detected in northern analysis of all cecropin

*CecA1*

433	CAATTATTTTTATTGTCATTTAATGCCTATTGAAATTTTCAAACCTAATTTAGTGCCCTTAGTAAAATATTGTAGTGATTCCTCCGCA	522
523	AAATACCACAAATGGATGCGTTTTATGTAATAAATGCCCCTTGAGTGATAGAGTAAATTTGAATTTGACTGTCTTAGAAAGATAGAAAG	612
613	AGATCAATTCAAAATGCCAAAAGGATAGAGTTATTAAGCTCTAATTCAAATTTGCCCCAGAACCCTTTAAAGGATATTACAATTTGTAAT	702
703	TTACATATTTGGATTATAGCATTGAAATCCCCGATTGTTCCCTAGATGTGCAGATGTGTGCTTGGAAATCAGATCGGTTACCTTCAGTGTA	792
793	CTTTTCTCTGCAAAAATCCCCGTGCATGCCTTATCTGTCAATTTGTTTTCAAGCTGTGTTGCGCTATAAAAAGCTCTCGCCTTTGTAT	882
	-----	
	A1 -->890	963
883	CGCAGTCATCAGTCGCTCAGACCTCACTGCAATATCAATATCTTTAGCTTCTCCTAAGAAAAAATCAAGAAAATATCACCATGAACCTCT	972
		MetAsnPheT (4)
973	ACAACATCTTCGTTTTCGTCGCTCTCATTCTGGCCATCACCATTGGACAATCGGAAGCTGGGTGGCTGAAGAAAATGGCAAGAAAATCG	1062
	yrAsnIlePheValPheValAlaLeuIleLeuAlaIleThrIleGlyGlnSerGluAlaGlyTrpLeuLysLysIleGlyLysLysIle	(33)
1063	TAAGTCTTCCATTTGAAATCTGTTAAGACGGAACTAACTGACTAACTCTTTTCGAAGGAACCGGTTGGTCAGCACACTCGGATGCC	1152
	GluArgValGlyGlnHisThrArgAspAla	(43)
1153	ACAATCCAGGGACTGGGAATCGCTCAACAAGCCGCAATGTGCGCCGCAACTGCCCGAGGTTGACCACGATGATTTATAATTATTTAT	1242
	ThrIleGlnGlyLeuGlyIleAlaGlnGlnAlaAlaAsnValAlaAlaThrAlaArgGlyEnd	(63)
1243	TTAAAGATCTATTTATTCTGTGCTCCCTGTAATAAAAACAATTTAAAAATTTAAAGAATTCTATTCAAACCTTGTTTTTAAAGAGTT	1332
	-----   (A) <sub>n</sub>	
1333	GGAGAAAAGCGAACTCTTGAATTTATACACACATTTAAATCACTTAAGAGGCATTATTATACAGGATATTACAAATCGCTCTTTTC	1422
1423	CGATTTGGAAAGGCCGAGATTATGTCTTATCTGTTGAAATATAATTCGTTTCACCTATAAAAGGACCAGTCTTTTAGITTTAAATTATCAG	1512
	-----Psi1	

*CecA1* SEQUENCE. Strain, *Canton S*. Accession, X16972 (DROCECPN). The numbering system continues from *Anp* Sequence. Psi1 downstream of *A1* marks the TATA box of a pseudogene.

genes. Primer extension and sequence features were used to define the 5' end. The 3' end was obtained from cDNA sequences. There is an intron after the Ile-33 codon (*CecA1* Sequence).

Sequence similarity between *A1* and *A2* occurs in an interval that extends between 40 bp upstream of the 5' end and 50 bp downstream of the 3' end (Kylsten et al. 1990).

*CecA2***Gene Organization and Expression**

Open reading frame, 63 amino acids; expected mRNA length, 354 bases. Primer extension and sequence features were used to define the 5' end. The 3' end was

*CecA2*

1513	TCGCTTGCAAATACTGAAACAATTAGATTAATTTGTGGATTTTATTTGTCTCATCTGACCACCTATTGGCCACAATTGGAAGCTGGC	1602
1603	TTCGACGGGACATTAGTAAGCTTAGTCATTTTAAAAGATTCTTTGCATCTAACTATGATTCTAAATCCTCAGAAGGACGTTGGTCTATA	1692
1693	CACCCATAAATGCTACCTGCAAGTTGCTGAAGTCGCTTCGAAAGCAGCCAATGTGGCAATCACTGCCAGGGGATAAACTTAAGTTAGGGT	1782
1783	ATTATTTATAAGAAATAAATTAATAGATTTTATTTTATATATTTTTTGTATATTGTTATTCAAACCTGATAATGTAATATACGCTTTTCA	1872
1873	AACGATCATTCCAATCAGTTGTGGGCTTATCGCAATGATTCGTAGTGTTTTTATTTTGATTGATTCAAAGAAGGGGTTTCTCTCTG	1962
1963	ATTCTTAGTCTCCCGCATTGACGAGGTAAAAATCCCTATGCATATGAAATATGCAAAATTTAAAAATCCCCAATCCGACAGGTTGGTTT	2052
2053	TGATCGGTTGGATTCTCTCGTACTTTTCAGCCATAAAAAATCCCTTTTCGAGCCTTATCAGGCGCTGAACCTAAGCTGATTCGCCTA	2142
		==
	-->2172 .	
2143	TAAAAGCTCTCGGGCTTCTGGTGAATCAACAGTCGATCACTTTCCATTGCAACAGCAACATCAGAGCTATAGCTACTCTTGCAAAATC	2232
	----	
	2253 .	
2233	TAAAGTCAAATAAAACCACCATGAACTTCTACAACATCTTCGTTTTCGTCGCTCTCATTCTGGCCATCACCATTGGACAATCGGAAGCTG	2322
	MetAsnPheTyrAsnIlePheValPheValAlaLeuIleLeuAlaIleThrIleGlyGlnSerGluAlaG	(24)
2323	GTTGGCTAAAGAAAATTGGCAAGAAAATCGTAAGTCCTATCTATTTGAAATTTGTTAAACCGGAAACTAACTAACTCCTTTTCATAGGAA	2412
	IyTrpLeuLysLysIleGlyLysLysIle	Glu (34)
2413	CGTGTGGTCAGCACACTCGCGACGCCACAATCCAGGGACTGGGAATCGCTCAACAGGCCCAATGTTGCAGCCACTGCTCGAGGTAA	2502
	ArgValGlyGlnHisThrArgAspAlaThrIleGlnGlyLeuGlyIleAlaGlnGlnAlaAlaAsnValAlaAlaThrAlaArgGlyEnd	(63)
2503	CCACGATGACTATCAATAAATATTTATACAAAATCTTATTTATTTTTTTGATCTAAGTAAATAAAACATTGGGAAAATCAATCTTTTG	2592
	----- (A) <sub>n</sub>	
2593	TCTTCTCTAAAGATCTATTACGCGAATAGTTGTGAATGAAAAGTGATTATAAATCCTATCTATAGTTTTAGGAGCGCACGTGCGAAA	2682
2683	AATATATATACAACATAAATCCCACTAATTAATTTTGTGTATTGTATAGATTGAAATCTAATGATAATATTTTCGACTGGGAAAATCC	2772
2773	ACAAAATATGCGTTATCTCCAAAAGTAGAAGATAGTTTCGCTATAAAAAGATCTAAGTCTAAGCTGTGAGCTTCAGTCCAAAAATAAC	2862
	-----Psi2	
2863	ATTAGCAAACAACATTTGCTGCTTTTTCCAGCTGTGAATTATATATTACTTAATATGAACCTTAGCCATATTTTTTTGTTGCTTTTCAT	2952
2953	CATCCTGACAATTAACCTTGCAACTCGCATGCCGGTGGCTGACGGATATAGTAATCTAAGACCGATCTAACTTAACCTCCCTTCACA	3042
3043	GAAGAAGAAATCTGAGGAGACTTTTAAATACTTAAAAACGCAGCATTGGAGGTCAATTGACGTCGGCCAAAAGCCGGGATTTTGTCTG	3132
3133	CATTGCCAGGGGACAGAAAAGTAGATCTCTACCAGATTTTTCTTGATGAGCTACAATTGCTGCAAAATTTTAAATAAAATCAAAAAGTAT	3222

*CecA2* SEQUENCE. Strain, *Canton S*. Accession, X16972 (DROCECPN). The numbering system continues from *CecA1* Sequence. Psi2 downstream of *A2* marks the TATA box of a pseudogene.



*CecB*

```

-809 GAATTCATTATGCTGGGAGTGGATAAATGGGATAAATGAGTGTACAATAAATGGATAATGCCATGTTGATTGAGGGGATTTCTTATGTCC -720
-719 AGGAAATATCATATTTCTACTGACTGTGTAAAGTGTGTACCTTTTATTCTGGGCTATAGAAAAATAATATATTAGTGTATAAAA -630
-629 TAACATTTTCTTGGAGTATTTATTGCAATTTGCTTCAATCTCCGACTTATTAACCTGCTGATAATTCAGTCCATTGCGAACTAAGTG -540
-539 ACTGATAGTCTTATAAATTTCTAAAAAAAAGAATACAGCATCTGTGACTGTAAAACGATGACAAATGGGATTTTGTCTGTAAAAAATAA -450
-449 TAAAAATTAAAAATAATAAAAAATTACGGGAGGCTTGTCTTACGGGAATACTATATAGGGAAAAACACTACACTTTAGTGTATGTTC -360
-359 CCCTAAAAGTTTAAAAAGTAATGTTTCATTATAATTACTTTGTTTTTAATTGTAGTTTTACGTTATTTTTAAGCTAGTTAAATCATCAT -270
-269 AATTCAATAGATTAATCAAATCATAGCTTGCAACCAACCAGTACTCTGAAATATCACTTGAGTAAGTCACTTTCATGGCGGTTCCGAAC -180
-179 TGAGTCCATCTGCTGGTGAACCTTTTGTCCCGCAGCAAAAAATCCCGCTGTGTCAGCCGTAGCATCTGTTGGTATCGCTATATAAGCTCA -90
      -----
-89 ATCTCTTCGATGTCCAATCATCAGTCGCACAGTTCCTACTGCAACAGCTTAAGCTTTCTTTCAATCCGATCGTAAGCCAACAATCTCGTC 0
  1 ATGAAGCTCAACAAGATCTTCGCTTTGTGGCACTCATCTCGCCATCAGCCTGGGAAACTCAGAGGCTGGTTGGCTTAGGAAGCTGGGA 90
  MetAsnPheAsnLysIlePheValPheValAlaLeuIleLeuAlaIleSerLeuGlyAsnSerGluAlaGlyTrpLeuArgLysLeuGly (30)
      |      |
  91 AAAAAATCGTATGGATTCCCTTCAAACCTAAACAAAATGAATTATTAATTTGATTTTCTTTTAGGAACGCATTGGTCAGCATACCAG 180
  LysLysIle                                     GluArgIleGlyGlnHisThrAr (41)
  181 GGATGCCTCAATCCAGGTCCTCGGAATCGCCACAGCCGCAATGTTGCAGCCACCCTCGAGGTTGAAATCAAGTCTCGAAGATCCT 270
  gAspAlaSerIleGlnValLeuGlyIleAlaGlnGlnAlaAlaAsnValAlaAlaThrAlaArgGlyEnd (63)
  271 CGACCCGCTCATTCTCTTATTATTATTAATGCATTAGGAAGATTAACATAATGAAAATAGATACTCAATGCCAATGTCAAATTATTAA 360
  361 AATATAAGCAAGCAGATATTAATAAAAACAAATTAAGACACTATATACAACAATAAGAAATGGTGAAAATATATTTCCCTGTAGGCTTAT 450
  451 CAAGATGTAATCGCACAAGCTGGTTACTGGTTAAATTTAAATAGAATTTGGAGGTTCTTATTATTTTATACTTTTTGATTTTATTAAT 540
  541 ATTTGCAGCAATTGTAGCTCATCAAGAAAAATCTGGTAGAGATCTACTTTTTCTGTCCCTGGCAATG 608

```

*CecB* SEQUENCE. Strain, *Canton S.* Accession, X16972 (DROCECPN).

obtain from cDNA sequences. There is an intron after the Ile-33 codon (*CecA2* Sequence; See *CecA1*) (Kylsten et al. 1990).

*CecB***Gene Organization and Expression**

Open reading frame, 63 amino acids; expected mRNA length, ca. 400 bases. Primer extension and sequence features were used to define the 5' end. The 3'

*CecC*

```

-324 GAAATATTGTTTAGAAGAAGTTAGCTATTGCTTTTGCACACATGAGAGCTAAGCGAAGAACCTCCATTTTACTAGCAGCTGCTCAA -235
-234 ACAGATTACCGAAGACAGCTCTTCGTCTAACAAAGAAGGGGATCCACTGCAGCTCTTCTCTCTCGCTGCCGAAAAGTTCCCCGTCGTCGCC -145
                                     I-91
-144 TTATCGGCATCGCATTCTTCGTATATAAAGCCGCTGTGCCAGAAGTCCAGTCATCAGTCGCTCAGTTTCCACAGCAGCTAAACAGCTAA -55
                                     -----
-54 ATCGCAATCTATATATATATATACTAAGGAATTAACCTAGAAAATTCACCATGAACTTCTACAAGATCTTCGTTTTCTGTCGCCCT 35
                                     MetAsnPheTyrLysIlePheValPheValAlaLe (12)
36 CATCCTGGCCATCAGCATTGGACAATCGGAAGCCGGTTGGCTGAAGAACTTGGCAAGAGAATCGTAAGTTCAGCAACAAAATATATTAA 125
uIleLeuAlaIleSerIleGlyGlnSerGluAlaGlyTrpLeuLysLysLeuGlyLysArgIle (33)
126 ATACTTGCAAATTTACTAATTTGTTTTATATTACTTGGCAAAGGAGCGCATTGGCCAGCACACCCGGGATGCAACCATTCAAGGACTGGG 215
GluArgIleGlyGlnHisThrArgAspAlaThrIleGlnGlyLeuGl (49)
216 AATTGCGCAACAGCCGCCAATGTGGCAGCCACCGCCAGAGGATGAGCCTTAATGTCCATCAAAGGACTCTACCAGGATAACGCGCGTT 305
yIleAlaGlnGlnAlaAlaAsnValAlaAlaThrAlaArgGlyEnd (63)
306 TAATTATACACACTTATTTATTTACCAGCCATAGAAAATAAAGTAGCTTACATCCCCGTAATTT 368
                                     -----

```

*CecC* SEQUENCE. Strain, *Canton S*. Accession, Z11167 (DROCECCG).

end was not determined. There is an intron after the Ile-33 codon (*CecB* Sequence) (Kylsten et al. 1990).

*CecC*

**Gene Organization and Expression**

Open reading frame, 63 amino acids; expected mRNA length, ca. 380 bases. Sequence features were used to define the 5' end, The 3' end was not determined. There is an intron after the Ile-33 codon (*CecC* Sequence) (Tryselius et al. 1992).

**References**

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# 6

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## *bicoid: bcd*

**Chromosomal Location:**  
3R, 84A

**Map Position:**  
3-[47.5]

### **Product**

The following discussion refers to the 489 amino acid product of the major transcript, BCD. It is a DNA-binding regulatory protein of the homeodomain type. BCD controls the expression of early developmental genes in the anterior half of the embryo (Gehring 1987; Driever and Nüsslein-Volhard 1989; Hayashi and Scott 1990; Harrison 1991). For a review see Driever (1992).

### *Structure*

The *bicoid* protein is a 55–58 kD protein, rich in Pro (10%) and probably phosphorylated. It has several sequence features of potential functional significance (Berleth et al. 1988):

1. The codons in the first exon include the PRD-repeat, alternating Pro and His, a pattern also found in the *paired* protein and other embryogenesis genes (Frigerio et al. 1986).

2. The amino-terminal region of the third exon (Pro-97 to Ser-156) encodes a homeodomain having weak (ca. 40%) similarity to other homeodomains.

3. There are several PEST sequences (rich in Pro, Ser and Thr), the most significant between amino acids 170 and 203. Such sequences are found in proteins of short half-life and are thought to be degradation signals (Rogers et al. 1986); although in this particular case, their deletion does not affect BCD stability (Driever 1992).

4. The carboxy half of the third exon is a Gln-rich region that results from the presence of repeated CAG (the M- or opa-repeat).

5. Further downstream, between positions 347 and 414 there is an acidic region.

Experiments with chimeric and mutant proteins in transgenic organisms established that the homeodomain is responsible for DNA binding and

sequence recognition and that the carboxy-terminal two thirds of the protein are necessary to effect transcriptional activation. However, no single localized region of BCD seems unequivocally responsible for the latter function (*bcd* Sequence) (Struhl et al. 1989; Driever 1992 and references therein).

The ten residues from 138 to 147 constitute the *recognition alpha helix* of the homeodomain (helix 3, which corresponds to the second helix of the prokaryotic helix-turn-helix repressor proteins). The Lys at position 9 of the recognition helix provides the specificity that distinguishes the *bcd* homeodomain from the *Antp* class homeodomain in which a Gln occurs in that position (Hanes and Brent 1989; Treisman et al. 1989).

### *Function*

The concentration of *bicoid* product determines "position" in the anterior embryo via regulatory action on other genes; that is, BCD is the "anterior morphogen" (Driever and Nüsslein-Volhard 1988b; Struhl et al. 1989).

BCD binds to the *hunchback* (*hb*) proximal promoter where it acts as a positive transcriptional regulator (Tautz 1988; Driever and Nüsslein-Volhard 1989). The BCD binding sites that occur in the *hb* promoter have the consensus TCTAATCCC; in this segment, the central TAAT is the core necessary for homeodomain protein binding, and the C in position 7 ensures that BCD, but not ANTP, binds (Driever and Nüsslein-Volhard 1989; Hanes and Brent 1991).

BCD is also involved in the regulation of *Krüppel* (Hoch et al. 1990, 1991, 1992), *even-skipped* (Small et al. 1991; Stanojevic et al. 1991) and probably other early genes. A less-well-understood function of *bcd* is its role in the formation of the *caudal* RNA and protein gradients, since this is a post-transcriptional process (Mlodzik and Gehring 1987; Driever 1992).

### *Tissue Distribution*

Production of BCD starts at the anterior tip of the egg shortly after oviposition (regardless of whether the egg is fertilized or not) and involves translation of a localized, pre-existing maternal message. By the syncytial blastoderm stage, the protein is localized in nuclei and distributed in a steep exponential gradient with the highest concentration at the anterior tip of the embryo and undetectable levels in the posterior 30% (Appendix, Fig. A.2). BCD reaches a maximum 2–4 h after oviposition; it begins to decline during blastoderm cellularization; and it is practically undetectable after gastrulation (Driever and Nüsslein-Volhard 1988a).

### *Mutant Phenotype*

This is a maternal-effect gene: offspring of homozygous *bcd*<sup>-</sup> females are inviable. In the absence of BCD, structures in the anterior half fail to differentiate; neither head nor thorax develops, and the terminal acron is transformed into a second telson (Frohnhöfer and Nüsslein-Volhard 1986).

*bcd*

-1414 GTCGACTGGAGTGTCTGTGAATTGACTTTTGTGCCAGTTGGCAGCGGCAGAAAGCCCGGCCAACAGCAACAAGCTCTTGCCA -1325

-1324 GATCCCAAAAGCAAAACACGACAATTATTTGGCAAATGTCATTAATAAATATTTTCACTTAAGGCCTTGCAGACTTGCCTAAAGTCAACT -1235

-1234 GGCTCGTTGGGTGTGTTTTAAATGTTAAAGCTTGGCCAAATGCACCTGAGCAACTTAATGCTTGTAGATATTTACACAATATTCTTCAAC -1145

-1144 GCTAAACATATCGAATTTTCCAATATGGAGCTGAAAATAAATATGCCAACTCTAGCTTAAATCAGAAATGAGTAGAACAACTTAAA -1055

-1054 AAAATTAACAAAAGAAATCGAACGCTACAGCTAATTAACTCGACAACTGGTTACCTTTTATCTTCTAATACATTTTATAATGCAGCTGCCT -965

-964 AACAGGTACAGATAGCAAGCACTATATGCTGTCTTACAAAACGATTATATGATATTTCTTTCTGTACGTAGCCGTTTGAGATCATTGGA -875

-874 AAAACAACTCGATCTCCACCATCTTATTCTTTGTCCAAAGTCTTATATATCTCGGATACTAAGATGAATAATGTAGTTATTAATA -785

-784 GCGGAAGTATGTAACAGAATAAACTACAAGTGACATTTTGTCAATTCAGGCTGGACTGGAGCATATTAATATTATAATATTA -695

-694 ACAAAAATTCAAATTAACATTCGACACTTGTCTAATTGATTCCATAAATTTGGGGTGCCTGTTTGTTAATTAATGTTAATATTATGAAG -605

-604 TTCCAAACAGAGCAAAGAGTTAAGTTAATTTGGTTCTACTTATTTGTTACAATATTCAGCTTTTTTTATTATTCTCAAAATGCAAA -515

-514 TCTCTACAAATAAATAAACCTCCGACGTTTTAGAACATTACCTTTTGTGAGTGCAGCACAACCTTCAATACAGCCCGCAGGGGGCTCT -425

-424 CTACTGCTGTCTTTCACGCCCTGGTGAACGCTGTGCACTCAATCGTTTGCAGCTTTGCCGTACTGTTTCGATTAATAAACTTTTAA -335

-334 ATTAGAGGCAAAACATTTAAAAATAAAATGTCCAAATATTTGTCTAAAATGTAATTTAGACGCTTATTGATTTTTAAATTACTCAAAGAA -245

!-168

-244 TGTTTCATCGAGGGAGGGCCGCAATTGTGCCATCTCTACATCTCTTCGCTCATCCCTAAATAACGGCACTCTGCAGATGCGAAGCAGTGG -155

-154 ATCGCAAAAACGCAAAATGTGGCGAAATAAGTTCGCGAGCGTCTGAAAGTAACGGTTACTGAAAATACAAGAAAGTTCCCACTCC -65

-64 TTTGCCATTTTTCCGCGCGGCGCTTGGAAATTCGTAAGATAACGCGCGGAGTGTGGGGAAAATGGCGCAACCGCCAGATCAAA 25  
MetAlaGlnProProProAspGlnA (9)

26 ACTTTTACCATCATCCGCTGCCCCACAGCACACATCCGCATCCGCACTCCCATCCGCATCCGCACTCGCATCCGCACCCACATCACC 115  
snPheTyrHisHisProLeuProHisThrHisThrHisProHisProHisSerHisProHisProHisSerHisProHisProHisHisG  
----- (39)

116 AACATCCGAGCTTCAGTTGCCGCCACAATCCGAAATCCCTTCGATTTGGTGAGTCCCATCGCAGCAGAGAAGGGCTCTGTGCCAGG 205  
lnHisProGlnLeuGlnLeuProProGlnPheArgAsnProPheAspLeu (55)  
----- PRD REPEAT

206 AAAGCTACAGTACAGATTCCTATGTTGAACAACAACAGTGCATCACTGATGACCATAAACATTTATTGAGCCGAGCAAATGTGTT 295

296 TCTAGAACATAGGGCGAAATCTTCTATTATCTTGTGTTGTGACTTTTAAAGTATCGTAGCAGAATCTAAATAACAATGATATTATTAATC 385

386 GTTACAGTTAGTATAGTATATAATTGTATATGAATTGTGGGGCAACATGTTATTAGTGATTTGCCGAAATGTTCTAAAAGATGTTTCATT 475

476 GAAATGGACGAATGTTAAACCTGTTGCACCTCACACCGAATATCAGTAATGTCTATTTTTCAAAGCCACATCTATGCCACTGGGTATAC 565

566 ATTATTGACTTAATACACTTTCATACAACATATTTCAAACAACAGCATGTTGTCTCGATGATGATTAGTGAAGTAATATTGCAAGAT 655

656	TCGGTCCCCGAAGCGAATCGTCTTTTACGTTTTATATAAAGACAGTGTACCCCTTGATTCTTTGAAAGCTTTTCGATGAGCGAACGGGA LeuPheAspGluArgThrGly	745 (62)
746	GCGATAAACTACAACTACATACGTCCGTATCTGCCCAACCAGATGCCCAAGCCAGGTGAGCTCAAAGCCAACAAAGTCAGCCATCGTCTT AlaIleAsnTyrAsnTyrIleArgProTyrLeuProAsnGlnMetProLysProA	835 (81)
836	ATCAGATGCTTTCCCTCAGAGGAGCTGCCGACTCTCTGGTGTGCGGCGACCACGTCGACCCCGCACCACCTTTTACCAGCTCTCAAAT spValPheProSerGluGluLeuProAspSerLeuValMetArgArgProArgArgThrArgThrThrPheThrSerSerGlnI1  alternate acceptor   DefE6= - T=E4 .T=E3   =DefE6	925 (109)
926	AGCAGAGCTGGAGCAGCACTTTCTGCAGGGAGCATACCTCACAGCCCCCGACTTGCGGATCTGTGAGCAAAGTACCCCTGGGCACAGC eAlaGluLeuGluGlnHisPheLeuGlnGlyArgTyrLeuThrAlaProArgLeuAlaAspLeuSerAlaLysLeuAlaLeuGlyThrAl Phe Leu -----*-----H1 * -----*-----H2 * -----	1015 (139)
1016	CCAGGTGAAGATATGGTTTAAAGAACCGTCGGCGTCGCACAAGATCCAATCGGATCAGCACAAGGACCAGTCTACGAGGGGATGCCTCT aGlnValLysIleTrpPheLysAsnArgArgArgArgHisLysIleGlnSerAspGlnHisLysAspGlnSerTyrGluGlyMetProLe End -----*-----*-----*H3* * *   HOMEODOMAIN DefE1= - .T=GB	1105 (169)
1106	CTCGCCGGGTATGAAACAGAGCGATGGCGATCCCCCAGCTTGCAGACTCTTAGCTGGGTGGAGGAGCCAGCCCAACGCTTTGACTCC uSerProGlyMetLysGlnSerAspGlyAspProProSerLeuGlnThrLeuSerLeuGlyGlyGlyAlaThrProAsnAlaLeuThrPr End AA- =DefE1	1195 (199)
1196	GTCACCCAGCCCTCAACGCCACTGCACACATGACGGAGCACTACAGCGAGTCATTCAACGCCCTACTACAACACTACAATGGAGGGCACAA oSerProThrProSerThrProThrAlaHisMetThrGluHisTyrSerGluSerPheAsnAlaTyrTyrAsnTyrAsnGlyGlyHisAs	1285 (229)
1286	TCACGCCAGGCCAATCGTCACATGCACATGCAGTATCTTCCGAGGGGGGCCAGGACCTGGGTGCAACCAATGTCAATGGCGGCCAGTT nHisAlaGlnAlaAsnArgHisMetHisMetGlnTyrProSerGlyGlyGlyProGlyProGlySerThrAsnValAsnGlyGlyGlnPh	1375 (259)
1376	CTTCCAGCAGCAGCAGGTCCATAATCACCAGCAGCAACTGCACCACCAGGGCAACCAGCTGCCCGCACCAGATGCAGCAGCAGCAACAGCA ePheGlnGlnGlnGlnValHisAsnHisGlnGlnGlnLeuHisHisGlnGlyAsnHisValProHisGlnMetGlnGlnGlnGlnGlnGln End End -----	1465 (289)
1466	GGCTCAGCAGCAGCAATACCATCACTTTGACTTCCAGCAAAAGCAAGCCAGCGCTGTGCGCTCTGGTCAAGGACGAACCGGAGGCCGA nAlaGlnGlnGlnGlnTyrHisHisPheAspPheGlnGlnLysGlnAlaSerAlaCysArgValLeuValLysAspGluProGluAlaAs ----- OPA REPEATS	1555 (319)
1556	CTACAACCTTCAACAGCTCTACTACATGCGATCGGGAATGTCTGGCCCACTGCATCGGCATCCGCTGTGGCCCGAGGGCTGCCCTGCC pTyrAsnPheAsnSerSerTyrTyrMetArgSerGlyMetSerGlyAlaThrAlaSerAlaSerAlaValAlaArgGlyAlaAlaSerPr	1645 (349)
1646	GGGCTCCGAGGTCTACGAGCCATTAACACCCAAGAAATGACGAAAGTCCGAGTCTGTGTGGCATCGGCATCGGGGACCTTGGCCATCGC oGlySerGluValTyrGluProLeuThrProLysAsnAspGluSerProSerLeuCysGlyIleGlyIleGlyGlyProCysAlaIleAl	1735 (379)
1736	CGTTGGCGAGACGGAGGCGGCCAGCAGACATGGACGACGGAACGAGCAAGAAGACGACGCTACAGGTGAGGCATGAGTCCACAACCTTTTT aValGlyGluThrGluAlaAlaAspAspMetAspAspGlyThrSerLysLysThrThrLeuGln	1825 (399)
1826	TGATCTCTTGATTCTGAGTGTGGCGTTTATAAATTGAAGCTTTAAGCTTTGTAACCTTCAAACCTGTCTGGTTTGAGATGTTATTCTGAAA	1915

1916	GTACTTCTATTTCCGATCGATGAGATTGGGAGTTCCTCAATATTTAACATTTAACTTATTAAGTTTTGTCTTAAATTAGCATGGC	2005
2006	ATTTCGAAAGGGAAGTACAAGTGTTAAAGATGATTTTAATATAGAATTTGTATCAAAGGTTAAGATTTCAACCGTTTGAAAGCCCTTA	2095
2096	GTTTTCAGGGTTTTTACTTTTTTATTCTGTAATCACTCTTAATACACTGCAAGTTAAAAATAGCATTCTTTGACCAGAAAAAATAGAA	2185
2186	TCTATGCATTTTAAAGTGAAAACAGACTCATATGCTGATGAACATTTTTAGCTATAAATTTGTAACAATAATTTAGCAATTTCAATTGAA	2275
2276	TTTATTTATGTTCTAAATGCGTTCGCTCTCTCCCTAGATCTTGGAGCCTTTGAAGGCTTGGACAAGAGCTGCGACGATGGCAGTAGCGA	2365
	IleLeuGluProLeuLysGlyLeuAspLysSerCysAspAspGlySerSerAs	(418)
2366	CGACATGAGCACC6GAATAAGAGCCTTAGCAGGAACCGGAAATCGTGGAGCGGCATTTGCCAAATTTGGCAAGCCTTCGCCCCCAAGG	2455
	pAspMetSerThrGlyIleArgAlaLeuAlaGlyThrGlyAsnArgGlyAlaAlaPheAlaLysPheGlyLysProSerProProGlnGln	(448)
	A=2-13	
2456	CCCTCAGCCGCCCTCGGGATGGGGGGCGTGGCCCTGGGCGAATCGAACCAATATCAATGCACGATGGATACGATAATGCAAGCGTATAA	2545
	yProGlnProProLeuGlyMetGlyGlyValAlaLeuGlyGluSerAsnGlnTyrGlnCysThrMetAspThrIleMetGlnAlaTyrAs	(478)
	His	
2546	TCCCCATCGGAACGCCGGGCAACTCGCAGTTTGCCCTACTGCTTCAATTAGCCTGGACGAGAGGCGTGTTAGAGAGTTTCATTAGCTTT	2635
	nProHisArgAsnAlaAlaGlyAsnSerGlnPheAlaTyrCysPheAsnEnd	(494)
2636	AGGTTAACCACTGTGTCTCGTATGTACAATAACCAAGTGATTGTAGATATCTACCGGTAGAAAGTTAGGCTAGTCTAAGATCCGGT	2725
	---	
2726	TAAATGGTTCACAGGGAAGTTTTATGTAAGCTAGCTAGTCAGCAGGCCGCACGGATTCCAGTGATATCTTAGTGATACTCCAGTTAACTC	2815
2816	TATACTTCCCTGCAATACGCTATTCGCCTTAGATGTATCTGGGTGGCTGCCACTAAAGCCGGGAATATGCAACCAAGTTACATTTGA	2905
2906	GGCCATTTGGGCTTAAGCGTATTCATGGAAAGTTATCGTCCCAATTTGGAAATATATATCCGAGCCAGCAAGAAAATCTTCTCTGTT	2995
2996	ACAAATTTGACATAGCTAAAACTGACTAATCAAAATGAAAATGTTTCTCTTGGGCGTAATCTCATAAATGATTACCCTTAAAGATCG	3085
3086	AACATTTAAACAATAATATTTGATATGATATTTCAATTTCTATGCTATGCCAAAGTGCTGACATAATCAACATTTGCGCATTCTTTG	3175
3176	ACCAAGAATAGTCAGCAAATTTGATTTTTCAATCAATGCAGACCATTTGTTTCAGATTTCTGAGATTTTTGCTGCCAAACGGAATAACTAT	3265
3266	CATAGCTCACATTCTATTTACATCACTAAGAAGAGCATTGCAATCTGTTAGGCCTCAAGTTTAAATTTAAATGCTGCACCTTTGATGTT	3355
	---  LOCALIZATION ELEMENT	
3356	GTCTCTTAAAGCTTGTATTTTTAATTACGAAAATATATAAGAACTACTCTACTCGGGTAAATTTGACTAACTACACATAACTACATAC	3445
	-----   (A) <sub>n</sub>	
3446	TTAGCCCATATTTCCGTCCTTTCTAGAATGAACGAAAACAGTATCTGGTTTTCCCGAAAATCTTATGAATTTAAAAATGCACCTTTATTG	3535
3536	CACATACTCACATGCTGCCATAAAAATATGATTCGCGATTTTTCCGCGAACACCCGCGGATCATAAAACATTTGCACCAGCTGCCTGT	3625
3626	GTTTATTCACCTACCTGAAACCATACTCTTATCGCCTGATCCTCGCGGGTGCACACTATTTAGGTAGACACTGTACAGGCAGCACTAGC	3715

*bcd* SEQUENCE. Strain, *Oregon R*. Accession, X07870 (DROBCDG). An exclamation mark at -168 indicates the 5' end of the longest cDNA. Dashes underline the region of PRD and OPA repeats. The boundaries of the RNA localization element and the homeodomain are indicated with vertical bars below the sequence. Within the homeodomain (Pro-97 to Ser-156), asterisks indicate conserved amino acids and dashes underline the presumptive helices. Mutations *bcd*<sup>E3</sup>, *bcd*<sup>E4</sup> and *bcd*<sup>E6</sup> (which

(continued)



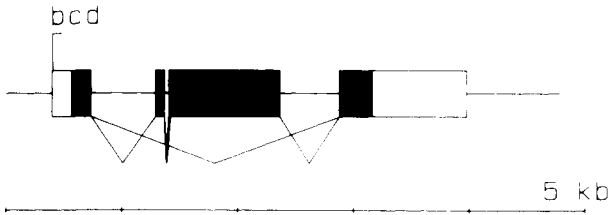


FIG. 6.1. Organization of *bcd*.

### Gene Organization and Expression, Major Transcript

Open reading frame, 489 (the most abundant) or 494 amino acids depending on the acceptor site of the second intron (*bcd* Sequence); expected mRNA length, ca. 2,453 bases, in agreement with the prevalent 2.6 kb RNA band (Berleth et al. 1988). Minor Transcript: Open reading frame, 149 amino acids; expected mRNA length, ca. 1,436 bases in agreement with a 1.6 kb RNA band (Berleth et al. 1988).

The 5' end of the longest cDNA is indicated in the Sequence at -168. No canonical TATA box is found in the appropriate position. The 3' end was determined from the sequence of two cDNAs. There are three introns: after the Leu-55 codon, within the codon for Asp-81 (or Glu-81), and after the Gln-399 codon. There are three alternative splicing forms. Two of them represent the major 2.6 kb transcript that carries four exons. They differ with respect to the acceptor site of the second intron; the two sites are in frame and the difference is a five amino acid segment (*bcd* Sequence). In the minor transcript, the second and third exons are spliced out (Fig. 6.1) (Berleth et al. 1988). The mRNA that codes for the 489-amino-acid protein is sufficient for all the *bcd* functions and is probably the functional form (Driever 1992).

This gene is 35–40 kb closer to the centromere than *Deformed* (*Dfd*) in the *Antennapedia* complex, and it is transcribed toward the centromere (Berleth et al. 1988).

### Developmental Pattern

Transcription of *bcd* begins early in oogenesis and seems to be restricted to the nurse cells. The RNA is transferred to the anterior region of the oocyte, together

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(continued) affect the homeodomain) encode proteins unable to bind to *hb* sequences in yeast cells. Mutation *bcd*<sup>GB</sup> (which truncates the polypeptide immediately downstream of the homeodomain) binds *hb* sequences, but it is unable to stimulate transcription in yeast cells (it is a strong allele *in vivo*). Mutations *bcd*<sup>085</sup> and *bcd*<sup>E5</sup> (which truncate further downstream) have some activating function left and are weaker alleles, specially *bcd*<sup>E5</sup> (Struhl et al. 1989).

with other maternal RNAs, by passage through the ring canals. A special feature of *bcd* RNA is its ability to remain strictly localized or “anchored” in the anterior 20% of the oocyte, in the cortical zone. A discrete *cis*-acting segment necessary for this localization is present in the 3' untranslated region of the *bcd* message. A 627-base segment (from 2,691 to 3,317) is sufficient to anchor mRNA to the anterior egg cap and includes sequences with the potential for extensive secondary structure (Macdonald and Struhl 1988). The *bcd* RNA remains highly localized until after the last embryonic cleavage division; then it is degraded, disappearing completely by blastoderm cellularization (Berleth et al. 1988). Microtubules and the products of maternal effect genes *swallow* (*swa*), *exuperantia* (*exu*) and *staufer* appear to be involved in the anchoring process (Schübpbach and Wieschaus 1986; Pokrywka and Stephenson 1991).

### Promoter

A 4.0 kb segment in front of the gene is sufficient for normal expression (Berleth et al. 1988).

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# 7

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## Blastoderm-specific gene at 25 D: *Bsg25D*

**Chromosomal Location:**  
2L, 25D3

**Map Position:**  
2-[16]

### **Product**

Unidentified. Codon translation yields a 741-amino-acid protein with two regions of similarity to known products. A 95-amino-acid stretch (positions 250–344) shows significant similarity (22%) to a portion of the product of the *fos* oncogene. The other segment is 21 amino acids long (509–529) and shows similarity to the repeating segments of rabbit tropomyosin that are thought to bind F actin molecules (Boyer et al. 1987).

### **Gene Organization and Expression**

Open reading frame, 741 amino acids; expected mRNA length, 2,645–2,774 bases when one of the proximal poly-A sites is used and approximately 4,749 bases when a distal site is used; this is in agreement with poly(A) + RNA bands of 2.7 and 4.5 kb. There are two introns in the coding region, one in the codon for Asp-78 and another after the codon for Gln-159. The 5' end was determined by S1 mapping and primer extension; 3' ends were determined by S1 mapping. Three of the proximal poly(A) sites scored have no corresponding poly(A) signals upstream; it is not clear whether these represent technical artifacts or whether they are true termini. A third RNA of 3.0 kb hybridizes to a *Bsg25D* cDNA, but it is very rare and could not be mapped (*Bsg25D* Sequence) (Boyer et al. 1987).

### *Developmental Pattern*

The 3.0 and 4.5 kb RNAs are expressed during the first 8 h of embryogenesis, and the 2.7 kb RNA is blastoderm-stage specific (Roark et al. 1985).

*Bsg25D*

-420	ATCAATCTAACGATAGTGATAAACGATAGGAACAATGGTCCACGATATGGCCACCTCCGTGCAAGTTTGCTTAATGCCCTCCAGAGCGCG	-331
	----- -->-295	
-330	CCACCGTGCTCGCTATACTGCATTAATGTGTTTTTATCAACTCGCTAGAAATACGCTATCCCAAAAAACCGCAAAACCCGCGATGTTTATG	-241
-240	TTGCGTCCGAAAGTCATATCATAGATTAGTAGTAGTAGTAACCCCTCAAACAGCCTGCTGCCAAAAACACGCGTGATTCGCCGCCA	-151
-150	CCCACGCACATAGACCCCGATATTTCACTTTTCTTGTGTTTCGACCCCTGACTGCGTTTGTGGATTTCCCCCAAGAAAAAAGCGAA	-61
-60	GTGAAAACGCAATTGAGCAGCCGATCGATTGGAACGGCAGGAATCCCCGGGTACGGATAATGGAGGTATCCGCCGATCCGTACGAGCA	29
	MetGluValSerAlaAspProTyrGluGlu	(10)
30	GAAGCTCTACCAAATGTTCCGACGTGCGAGACGCAGTGTGGACTTCTGGACGAGAAGTCCCTGCTGAAGCTCTGCTCAGTGTGGAGCT	119
	nLysLeuTyrGlnMetPheArgSerCysGluThrGlnCysGlyLeuLeuAspGluLysSerLeuLeuLysLeuCysSerLeuLeuGluLeu	(40)
120	CCGGGATCAGGGATCCGCACTGATCGCCAGCCTGGCGCGCAGCCATCAGCTGGGCGTGTCTTTGGCCAGTTCAAGGAGGCGCTACTCAA	209
	uArgAspGlnGlySerAlaLeuIleAlaSerLeuGlyGlySerHisGlnLeuGlyValSerPheGlyGlnPheLysGluAlaLeuLeuAs	(70)
210	CTTCCTGGGCTCCGAGTTCGATGGTAATACGTATCGGGTTTCATTGGTGAGATAGCACAAGAATCGATCACGCTATAGATTAACCTAT	299
	nPheLeuGlySerGluPheAspA	(78)
300	ATAGTATAAAGATAATATTGCTATAAGCTAACGCGCAGGTTCCGCATAAAACAACATACGTTTTATCTGTAATTGCGCTTTAATTACCC	389
390	ATCAAGCAACATCAGATAATTACGGAATGTTGCCAGCCACTTATTAGAGATAGTAATTCATTTTGACACGGATTGGAACCGTGTGGG	479
480	TTTCCCTATTAATAAAACACTGATCTAATGAACACATTCTAGCAGCTATAGATGAACAAGCCATTACTTAATACTCAAAGAAGTGCT	569
570	ACCATCTACGTGCTAATTTGCAAGGATTATGCACATTTACTTCAAACCTCCGCTTATCTGATTTGGAACTTCTGGGCAAATTTAGGACA	659
660	CCTTAGGGTACGAATATCATAATCAGCACGCGGATTAGCACGCGCAGCTGGCGATCAAAAATCATAGATGCAATTGACACTTTTTTAC	749
750	GACTCCCACTGTTCTCGACTACCTGATCCTGCATGATCCTTATCAGGTAGATGGTTACAATGTCCTGTATAAATACGCGACACATTCAC	839
840	CTGGGCAGTTTAGTCTAAATCAAATGGGAACACGATTGTATTACCGCCGATCCGGCGGTCAAGTAAACAGATCCGATAAATGAGAAGCTA	929
930	GCCGCTCGTTTTGGTAGCCACCTAAGATCCATACAACCTCTCCAGTCTCTGCTAACTTATATCTATTGAATCTTCCAGAGCGTTCACCTG	1019
	spArgSerLeu	(81)
1020	GTGATTACGGATGAGCCGCTAAACAACACATACATCGAGAGTCCGCCGGAGTCTCCGATCGCGAGGTTTCAACCAAACCTCGTCTGGGGC	1109
	ValIleThrAspGluProLeuAsnAsnThrTyrIleGluSerProProGluSerSerAspArgGluValSerProLysLeuValValGly	(111)
1110	ACCAAGAAATACGGTCGCGGTCTAGGCCACAGCAGGGAATCTACGAGTTATCCGTACGGACTCGGACAATACGGACGAGGACCAAGTTG	1199
	ThrLysLysTyrGlyArgArgSerArgProGlnGlnGlyIleTyrGluLeuSerValThrAspSerAspAsnThrAspGluAspGlnLeu	(141)
1200	CAGCAGCAGCAAAATCAGCGAAGCCTCAACGGATGCGATGAGCTGGGAGTTCCAGTGAGTGTGTTTTGCAAGTCACGTACGAAGTGCGG	1289
	GlnGlnGlnGlnAsnGlnArgSerLeuAsnGlyCysAspGluLeuGlyValGln	(151)
1290	ATACAACCTCTGGTATGTATGCAAAATTCATAGTAACAGATTTTGTTTAATCGTTATTATTGCTGATACAGTAGAGCATGCCTAAGTA	1379
1380	GCACTACCAAAGCAAACAATATCTTAATATACATCATGATCATCATAAGCATTTATTTTTCCAAACCACACAGGTGCAACGTTCCCT	1469
1470	CGTCCCAGAGCGATCTTCCTGGCAGCGGCGTCTCGGTCCTCCACACCAGCGGGAGCAAACCTGAAGCGTTGTGCTTCACTGCCAGCCC	1559

1560	GCCGGAAGATGAACAGCAACACCACGGAGCCACTACATCACCAGCGCAGCGG6CCAAGTTGAAACAGCTTCCATCCAGAGCCAGGCGCA	1649
1650	GCACAGCAGCAGCGTGGAACTCACTGGTAAGTTTCTCTGGCCAGACCAGCTTTGGCTAGCCGATCCCCCTGTCCCTGCCACCCCTCTGT	1739
1740	TGTTGTTAGCCAAAAATGCCAAAATTACGTTTGAAGCAATGTTAAAAGCAAACACTGTTTGTGCGGTACACACCAGTACGCCTCGCTGG	1829
1830	CCACCAATCCCAGCCGTCGTCGGAGCAGCTGGAGATGCTACCAGCGCGCCGTTGGTCACTGCTCAAAGGTTGTGCGCTCTGAAGCAAT	1919
1920	TGTCAACACCCCTCACACCCACCGAATCCCCAACCCAGTCATTCGGTATCTAATCGCACCCATGTAGCCGCACATTTGATTCGTTTTTTT	2009
2010	TACTCGTATAATAACATATCTACATTTTCAACCCCTTAGTAATGCTGTAATGCATTGACAATCAATTTAATTAAGGATTTTCATATAAATC	2099
2100	AATTTCAAGTTAGAAAGGATATTTACTTATAAATTTGTTCTATTTCTTGATTTATTAGTTTCTACCTCTTAAATAACACGGCAAAAATTT	2189
2190	CTCATTCTAAAAGCCATTTGATATAGAGAAATAACAAACTTTCGGCGCTTTTGGTTACACCATCGACACACACACACCCCTCCCCAC	2279
2280	TCCCAATCCCAATCCAATCCCACACCCACCTGGTATCTTGGGCTATATGTATAAAAATGTGTATATACAACAGCGAAGCCAATCTCATT	2369
2370	GTCCCACGCTAATGTTAATTGCCATGATTTACAGACACCGTGACGCCGAGCAATGGAGAGCATCTCAGTGATAGCATTATGGAAGC GlnLeuGluThrIleSerValHisSerIleMetGluAl	2459 (172)
2460	CTGGGAGCTGGCCAGCATTCCCAACTCGCAACCTACTTCACGTCCTGGGATTCGATGAGGAGGAGGAGGTAACCTGCAGCAGCTAAC aTrpGluLeuAlaSerIleProAsnThrArgAsnLeuLeuHisValLeuGlyPheAspGluGluGluValAsnLeuGlnGlnLeuTh	2549 (202)
2550	TAAGGCATTGGAGGAGGAGCTGCGGGGATCGATGGGGATCAGAGCAATCGAATATGTTGCGCGCTCTGGCTGCTCGACGGCCACCGA rLysAlaLeuGluGluGluLeuArgGlyIleAspGlyAspHisGluGlnSerAsnMetLeuArgAlaLeuAlaAlaLeuGlnAlaThrGln	2639 (232)
2640	GTTGGGCACTACAGACTTGCCATATAGCAGCAGCATGAGGAGAACCCTAAGCTGAGGGCCGATAATAAGGCGGCCAACCAAAGGGTGGC uLeuGlyAsnTyrArgLeuAlaTyrArgGlnGlnHisGluGluAsnLeuLysLeuArgAlaAspAsnLysAlaAlaAsnGlnArgValAla	2729 (262)
2730	TTTGCTGCGGTGGAAGTGGATGAGCGGCATGCGTCGCTGGAGGATAACTCCAAGAAGCAGGTGCAGCAGCTGGAGCAAAGACACGCCAG aLeuLeuAlaValGluValAspGluArgHisAlaSerLeuGluAspAsnSerLysLysGlnValGlnGlnLeuGluGlnArgHisAlaSe	2819 (292)
2820	CATGGTGCGTGAAATAACGCTGCGGATGACTAATGACCCGATCACTGGACCAGCATGACGGGAAAGCTGGAGGCACAGCTTAAATCGCT rMetValArgGluIleThrLeuArgMetThrAsnAspArgAspHisTrpThrSerMetThrGlyLysLeuGluAlaGlnLeuLysSerLe	2909 (322)
2910	TGAGCAGGAGGAGATCCGCTGAGAACGGAACCTGAACTGGTGCGCACTGAGAACACGGAGCTTGAGTCGGAGCAGCAAAAGGCTCACAT uGluGlnGlnGluIleArgLeuArgThrGluLeuGluLeuValArgThrGluAsnThrGluLeuGluSerGluGlnGlnLysAlaHisIl	2999 (352)
3000	CCAAATCACAGAGCTTCTCGAACAGAACATTAAGCTCAACCAGGAACCTGGCCCAAAGGTCGAGCAGCATTGGTGGCACCCCGGAGCACAG eGlnIleThrGluLeuLeuGluGlnAsnIleLysLeuAsnGlnGlnLeuAlaGlnArgSerSerSerIleGlyGlyThrProGluHisSse	3089 (382)
3090	TCCATTGCGACCGAGAAGGCATAGCGAGGACAAGGAGGAGAGATGCTCCAGCTAATGGAGAAGCTGGCTGCTCTTCAATGGAGAACGC rProLeuArgProArgArgHisSerGluAspLysGluGluGluMetLeuGlnLeuMetGluLysLeuAlaAlaLeuGlnMetGluAsnAl	3179 (412)
3180	CCAGCTGCGTGACAAGACTGACGAACGACCATCGAAATCGAGAGCTTAAATGTGGAACATAATTCGCTGCAAAAACAAAGGCTAAAAAGCA aGlnLeuArgAspLysThrAspGluLeuThrIleGluIleGluSerLeuAsnValGluLeuIleArgSerLysThrLysAlaLysLysGln	3269 (442)
3270	AGAAAACAGGAGAAACAAGAGGACCAGGAGTTCGGCGGCCACGGCTACCAAAGGCGTGGGATTCGCGGAGCAAAACACATCTAACAGA nGlnLysGlnGlnLysGlnGlnAspGlnGlnSerAlaAlaThrAlaThrLysArgArgGlyAspSerProSerLysThrHisLeuThrGln	3359 (472)
3360	GGAGAGCCCTCGCTTGGGGAACAGCGCAAGTGACCCGAAGGAGAGCAGAGCGATGCCAGCAACAGCGGAGATTGGTTGGCTCTAAACTC uGluSerProArgLeuGlyLysGlnArgLysCysThrGluGlyGluGlnSerAspAlaSerAsnSerGlyAspTrpLeuAlaLeuAsnSe	3449 (502)

(continued)

3450	CGAGCTGCAAAGAGTCAAAGCCAGGATGAGGAGCTAACAGCCCTTAGACAGCGGGTTGCTGAGCTAGAGGAGGAAGCTCAAGGCTGCAAA rGluLeuGlnArgSerGlnSerGlnAspGluGluLeuThrSerLeuArgGlnArgValAlaGluLeuGluGluLeuLysAlaAlaLy	3539 (532)
3540	GGAAGGCAGATCTCTACCCCGAAAGCCGTTTCAAGGAAGCTGGAGACCACTAGAGCAATGCAGCGTCCTATGAGATTGCGAGGA sGluGlyArgSerLeuThrProGluSerArgSerLysGluLeuGluThrSerLeuGluGlnMetGlnArgAlaTyrGluAspCysGluAs	3629 (562)
3630	CTACTGGCAAACGAACTTAGCGAGGAGCGCAGCTGTTTGAAGAGCGACAGATCTACAAGATGAGCAGCAGAGAGCGACAAGAA pTyrTrpGlnThrLysLeuSerGluGluArgGlnLeuPheGluLysGluArgGlnIleTyrGluAspGluGlnHisGluSerAspLysLy	3719 (592)
3720	GTTACCGAGCTGATGGAAAAGTGCAGGAGTACGAGGAGCAAGTTCAGCAAGGATGGCCGCTCTCGCCATTGATGAGCGCATATGCT sPheThrGluLeuMetGluLysValArgGluTyrGluGluGlnPheSerLysAspGlyArgLeuSerProIleAspGluArgAspMetLe	3809 (622)
3810	GGAACAGCAGTACTCGGAATTGGAGGCAGAGCCAGCCAGCTGCCTCGAGTTCCATTCAATGCTCGAGGAGAAGGCTCAGGAAATCAG uGluGlnGlnTyrSerGluLeuGluAlaGluAlaAlaGlnLeuArgSerSerSerIleGlnMetLeuGluGluLysAlaGlnGluIleSe	3899 (652)
3900	CTCACTGCAATCGGAGATCGAGATTGCGCAGAGATTGGGTGAGAGCGTTGAGATCCTTACAGGCGCTGTGAACTCACCTCGGAGTC rSerLeuGlnSerGluIleGluAspLeuArgGlnArgLeuGlyGluSerValGluIleLeuThrGlyAlaCysGluLeuThrSerGluSe	3989 (682)
3990	GGTAGCCCAACTGAGTCCGAGGCGGAAAGTCCAGCCAGCTACCCATCAGCTACCTTGCTGACAGCACCATCCAAGAGCCAGC rValAlaGlnLeuSerAlaGluAlaGlyLysSerProAlaSerSerProIleSerTyrLeuTrpLeuGlnSerThrIleGlnGluProAl	4079 (712)
4080	GAAATCGCTTGCCGATTCCAAGGATGAAGCCACCAGCCAGTCCATCGAATTGCTCGGAGGCTCACCATCGCACAGACAGCCAGCCGGTG aLysSerLeuAlaAspSerLysAspGluAlaThrAlaSerAlaIleGluLeuLeuGlyGlySerProSerHisLysThrAlaSerArgEn	4169 (741)
4170	AGTATGAGAAGCCCTCTCGGTGTGCTTGGTGTGAGCATCCCTGTGTCTTCTCATAAATTTGCACTGTATGCTGTATATATGTTTCAG d	4259
4260	TTTGTCCCTCACATCTAACCATGTCTAATATAAGCTAAATTAATCCTTTTAAATGTATGTTTGTGCTGTTTAAATAAATAAATTTATAT   (A) <sub>n</sub>   (A) <sub>n</sub>   (A) <sub>-----</sub>	4349
4350	TCATATAGAAATTCATCACATTATCGAAATTCATTGATTTATGATTTCAATAAATATACATTTAATATTTTACAAAAAATTACTCTTTT   (A) <sub>n</sub> -----   (A) <sub>n</sub>   (A) <sub>n</sub>	4439
4440	TCGGTTATGAAATGGCTGCTGGAAATGGTTTTGTTGCTTATTTTACATTTGTATCATTACACGTTTTGCATCTTTATGTTACATCT	4529
4530	TCAATCGTTTTTATTTTGTAAATCATGCCATTTAATGGTCCCTTAAACAGCAATAACCTCACCCTCGGAAACATCCATCTTTAGCACT	4619
4620	ACACCCTTCGAAAGCTCTCAGTCGGTCTTCGCCACGAACAGTGGCAACAGCAACGCTACGGCCAATCCCGGCCAGCTCCGATCAG	4709
4710	CAAGCCCAAGCGGTCCAGAGTCCCAACAGCGGGTGCATCGGAGGGAGAGATAGCCGATTGCGAGACGTCGTGACGGCGTCCGGCAA	4799
4800	AAGCTTCGAATCCAACAGTAAACGCTTGGCTTAGCCACGAGAAGTGCAGCAGTCCGTGGCCTGAAAGGAGGAAGTGAAGCGCCTTAA	4889
4890	GTTCTTCGAGCTCTCCCTCAAGGAGCAATCAAGGATCTGAGTCTGCAGCGGGACGGTCTGGTCATGGAACGTCAGCAGTTGCAAGGAGGC	4979
4980	GCGACCCGTGCTCGAGAAGCCATGCGGTGAGCCATAGACTTTGTTGATCAGGGAACATATTCTAATCGTATCTGTGGACTCTTCTTTA	5069
5070	GCGAACCAACGCATCCAACGCTTACGACGCACTGAACCAATTTGAGCTGCGAAATCGCCATCTGCAGAATGTCATCAAGCAGCAGCAGCA	5159
5160	TTACACGGAGTCCCTGATGCAGCGTAAGTTGAAAACTACCTACTGACCAAGAATGATAATGTAATATTTATTATTAGAACTCCTGGCGG	5249
5250	CAGCATCAAGTGGAGCTCAACGATTTGCATAGCCGAATCGAGACCAGGGTGTCTTACTGGCCGATCAGACACAGCGATTGCAAGAGTGCCG	5339

5340 ACATCCTGGTGAAGGATCTATATGTGGAGAACTCCCATCTGACGGCCACGGTGCAGCGTTGGAGCAGCAACGAGCTAGGGTGAACCTCA 5429

5430 TTCACCAGCAGCAGCAACAGCAGCGCCTTGTGGGCGGTGGACTGCCTGGCATGCCTTAGTTTGCACCCACGGCAAACGTATATAGTTTAT 5519

5520 AGATAATTATGAAAAAGACAAACCTGAGGAGGGAGTGGTGTCTCAGCATCGGCAGACATCGAACATGCACCTAACCATAGATCCTTATGAA 5609

5610 TGTTTAGACATATAACAATTCTCGGTAGATTAAGTTTGATACCCGTCGTATTTCGTATTCGTACGTTGCGTTTTTTTTGTGAATGAATGTG 5699

5700 AATCCATGTTGTTTCGACACGAGAGCACAGCAATAACTAAAGTGACTTTAAACTAAACTTAAACTCACCCACGCGCAAATGAGGAACA 5789

5790 ATCCCACTAGTGTACCAATTTGTAACACATCTAGTAATCGAATCGACTAAACTATTTACAGGAGTACAGGACATATACGATGAAGTAC 5879

5880 CCACGTAGTATATGTTTCGTCAATGTTGACCTTACTAATTGACTACTGAAACAGTTATCGTATATTAATTATATTAGAAGAAACAGTATT 5969

5970 TTAATTTGTTTATGCGTCTGAGTAGGCGAGCACGTTTATCAATGTTTATCACGTGCCAATCAAATGCATCGGAATTGTTGTTAATTTTA 6059

6060 TTGATAGAGAAAATGGAATGAGCGTAAAAAATGATCTATGATATTGATATTGATGTAATATTTAACGACAAAAGACCTGTAAGCTGTA 6149

6150 ACCATACACGAACTATGTATTTAAATTCGATCTAAGTTAGCCAACTCTTCAATATTGCTTTTTCGAAACGCGACTTTTTGTTATA 6239

6240 TCTTCATTCGTCCAATAACTCACTCGATTTATGTAAGAAAAAATACTCAACCTCAATCACAGATATCGTGAATCAGTGCTTAA 6329

6330 ATCAACTTTTCGATCAAAATAGAAGTTTACTTTTTAAAAGTATAAAAAATAATACAACAAAAAACCAATATACAATTTTATAAAA 6419

----- | (A)<sub>n</sub>

6420 CCAAATTGTGATAACTCGTCTTTATTCTAAATAGTTATTAATGTTGCGGGAATATAAACTATTGTTCAATAAACAACCTGCTTATC 6509

6510 AGTTTTTGGAAATGTTAAGATTTGTTCTTATTAATTAATGTTATTAATTAAGATTTATGAAGTTTAAATATATATTGATA 6599

6600 CGATAAACAATTTATTTTATTGCTTCAAAATATACATTACTTTTTTTAGGAATTTAAATATCCGTTTTAAGTCTTTAATTTTAATTA 6689

6690 GGTTTTAAATATAATATCGATTAATAGTTGACTCCATTGGAATATCGATACCGCGTCGATGTTTCTCCAGCTCTATCGGGCACGCGCT 6779

6780 GTTAAAGTTTATTGACTGTTAACGCGAATTGGATTAATGTTTGTGTTTGTGTTAGTTTTCGGTGTAATGTGGTTTTAGTGCACTT 6869

6870 AACACTGGTGAGTGTGCGTTATAGTAAATGAACCTAAATGCAATTACCGAATTC 6924

*Bsq25D* SEQUENCE. Accession, X04896 (DROBSG25D).

## References

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- Roark, M., Mahoney, P., Graham, M. and Lengyel, J. (1985). Blastoderm differential and blastoderm specific genes of *Drosophila melanogaster*. *Dev. Biol.* **109**:476-488.



# 8

## Chorion Protein Genes: *Cp36*, *Cp38*, *Cp15*, *Cp16*, *Cp18*, *Cp19*

### X-chromosome Cluster *Cp36* and *Cp38*

**Chromosomal Location:**  
X, 7F1-2  
Synonyms: *S36* and *S38*

**Map Position:**  
1-[23]

### Products

CP36 and CP38 (chorion proteins of 36 kD and 38 kD) are two of the six major protein components of the egg chorion; the other four are the product of genes on chromosome 3 (Petri et al. 1976). CP36 and CP38 are probably the main components of the innermost chorionic layer and the internal region of the thick endochorion (Parks and Spradling 1987; Orr-Weaver 1991). An N-terminal segment of approximately 20 amino acids is probably a signal peptide that is cleaved upon protein secretion (Waring and Mahowald 1979). As is true for all major chorion proteins, CP36 and CP38 are rich in Gly, Ala, Pro and Ser (in CP36, these amino acids constitute 40% of the residues and in CP38, 50%) and in Tyr, an amino acid that is extensively cross-linked in the mature chorion (Petri et al. 1976). Both proteins have runs of Ala and Gly-His, but overall sequence similarity is not striking.

### Organization and Expression of the Cluster

*Cp36* and *Cp38* lie within a 13 kb segment of DNA and are part of a cluster that includes six tandem transcription units. *Cp36* is positioned centromere distal, upstream of *Cp38*; both genes are transcribed toward the centromere. Downstream of *Cp38*, approximately 1.4 kb away and transcribed in the opposite orientation is *ovarian tumor* (see *otu* Fig. 23.1). The function of the

## Cp36

-802 TTTACATTGAGACGAAACAATCCACCGAAAAATCCATAAAATATAAGAATGTTGCATTTTATTTTTAAAAATAAAGATGCCTTTTAAGAG -713

-712 GAATAACTTAAATGTCTTTAATACCTTTGAATTTAATTATATGGCTAATAACACAACTTAAAGCTTAAACTGCATCGAATTGAATGC -623

-622 GGTATAAATGTACTTATATATCTAATAATAATCTGCTAATATGGTTTACATGGTATATCTTTCTCGGAAATTTTACAAAAATTATCTAT -533

-532 TCATATATCTCGAGCGTAAGATATTTATCAGTTTATAGATAACATCTTTAAATTTGGTGATTAATAAAAAACATTCGTGCAGGGCATGT -443

-442 TTATGTACACATTTTCAGTATAAGTCCCAAGTTAAAATGCAATGTAACAAACATATAAAGGATATTAACCTCAAACCCAAAGGATTGCAGAG -353

-352 AGATTGCAGCACAGCTGTAATCATCGCAACAAGGCAACCAAAACGAGACTCTCGTAGCGTTGGCATCATATTCGATCTTTGGAAGAGCTA -263

-262 TGATTCAAGCCAAGGGAACAACACTGCCAAAAAATAGAAGATTGCGACGAGCGAAAGCAGAGTGGTGCACCACGGTGCATAGGTGCATAG -173

-172 GAGGTTGGTTGTCTAGAGATCTGGGCACGATGGCGAGACAAGATGCGGCGCAAAATCGGAAATGGAGATGGATCACGTAGCCGCCCATG -83

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-->-30

-82 GCGGGCAGCGATTATAGCGCTATAAAGAGCCGGAGTCATCGCCAGACGCGAGCAGTAGACGATCCACACGTAACGGCAACATGCAAC 7  
MetGlnL (3)

-----

8 TCGGTCCTCGGTTTGGCATTTTGGCCATCGCCGCCGCGCGGTTAGTAGCTTTTATCCATGGCATTTCGATTGGGCATCCCCTAATCTC 97  
euGlyLeuTrpPheGlyIleLeuAlaIleAlaAlaAlaPro (16)

98 GCAAACCTCTCTCTCTCTCTCTTACCTCCTCCTATTAGCTGGTGAGCGCTAACTATGGTCCCGCTGGCGGACACGGACACGGGACAT 187  
LeuValSerAlaAsnTyrGlyProAlaGlyGlyHisGlyHisGlyHis (32)

188 GGACATGGACATGGACAGTACCTGTCCGGTCCAATGCCGACTCGAGGAGTACGTGAATGTGGCGTCTGGTGGCAACAGCAGGCTGCC 277  
GlyHisGlyHisGlyGlnTyrLeuSerGlyProAsnAlaGlyLeuGluGluTyrValAsnValAlaSerGlyGlyAsnGlnGlnAlaAla (62)

278 AATCAGATCGCCTCACAGGCCGAGATCCAGCCACGCGGAGGAGCCCGTCTTTGGGTGCGTCCAGGCCAACTTCAGGCCCTCAAC 367  
AsnGlnIleAlaSerGlnAlaGluIleGlnProThrProGluGluAlaArgArgLeuGlyArgValGlnAlaGlnLeuGlnAlaLeuAsn (92)

368 GCCGATCCCACTACCAGAAGTGAAGAATCCGAGGATATTGCCGAATCTCTGGCCGAGACCAATCTGGCCAGCAATATCCGTACGGGC 457  
AlaAspProAsnTyrGlnLysLeuLysAsnSerGluAspIleAlaGluSerLeuAlaGluThrAsnLeuAlaSerAsnIleArgGlnGly (122)

458 AAGATTAAGGTGGTGTCCGCACAGTTCGTTGACCAGCATCTGTCCGCTCCCTGTTGGTGCCATCGGGCCACAACAACCACAGGTGATC 547  
LysIleLysValValSerProGlnPheValAspGlnHisLeuPheArgSerLeuLeuValProSerGlyHisAsnAsnHisGlnValIle (152)

548 GCCACCAGCCCTGCCACCAATCTGTCACCAGCCTGGTGCACCACAGCCCATGTGAACAGCGGCCACCAGACTGTGGTGCAGCGGC 637  
AlaThrGlnProLeuProProIleIleValHisGlnProGlyAlaProProAlaHisValAsnSerGlyProProThrValValArgGly (182)

638 AATCCGGTGATCTACAAGTCAAGCCCTCGGTATCTACCAACAGGAGGTGATCAACAAGGTGCCACTCCGCTGAGCCTCAACCCCGTC 727  
AsnProValIleTyrLysIleLysProSerValIleTyrGlnGlnGluValIleAsnLysValProThrProLeuSerLeuAsnProVal (212)

728 TACGTGAAGGTCTACAAGCCGGCAAGAAGATCGAGGCTCCACTGGCCCGGTGGTTGCACCCGCTACAGCCAGCCAGGGAGTACAGC 817  
TyrValLysValTyrLysProGlyLysLysIleGluAlaProLeuAlaProValValAlaProValTyrSerGlnProArgGluTyrSer (242)

818 CAGCCCCAGGGTTATGGTAGTCCGCCAGCTGCTTCCCTCCGCCCGGTCGCCCTCTCTGCGGATGGCAATGCCTACGGCAACGAGGCT 907  
GlnProGlnGlyTyrGlySerAlaGlyAlaAlaSerSerAlaAlaGlyAlaAlaSerSerAlaAspGlyAsnAlaTyrGlyAsnGluAla (272)

908 CCACTGTACAACAGCCCGCCCTATGGCCAGCCCACTACTAAGGTGCTCATCTGGGCATGGGTTGTTCTCTCAGCTGCGACAGCTGG 997  
ProLeuTyrAsnSerProAlaProTyrGlyGlnProAsnTyrEnd (286)

```

998  TTTAATTTAAATTTTTGTTTTTTTTTCTTTGCCGAACACTGAGCGCAAATAAATGAAAAAAAAAACTTGAAACGTAACGTTTTGT 1087
      . . . . . ----- | (A)n
1088  CATCAATTATTTTCGACCGGAAGGGGCTACCTGAGTGGCAATGAAGCACACAGATTAAGCACATATTTATGAATATATAAATATATACGA 1177
1178  ATGCATGAGAGAACAAAAATTATATCTAGTTTTCTTCAAAAATAAATAATAAAGGGGAAAAATGGTATAAAGTATGAACATAAAATTATG 1267
1268  AACGAGTATAAATAAGTTTGAATTGAAATCTCTACGGTCATACAAGTATTTTAACTATTCTATAAATATGCATAAACATGGTACGCATT 1357
1358  TTATGAGATACAATTCGAAAGATATTGGATAGCATTATCATGCATGAAATTATTTACACTCTTGTGATGGTAGCTATCTTATTCCAGTT 1447
1448  ATCGTTTAATTGCAAAAGATGGGATCAAAGGTCATCTTTATCAACATATTTGTTGATTCCGGAATGAATAAATAAATAACATAACTAT 1537
1538  GAATTAATGGAGCAACTATAATTTTACGGCCTCTTTTCTTTTAAACAAGAATATAGCACTTTTAATGCATTAATACGTATTTAAACCT 1627
1628  TTTCTTTTGAAACGCCAAATTCATATTAGAGTTTCATAAGATTGTTTTAAACATAACAACATAATAATTGAAGAATTGGAATCTTTTT 1717
      . . . . .            EcoRI
1718  AGGTGTTTGTAAAGCCTTTGA 1737

```

*Cp36* SEQUENCE. Accession, X05245 (DROCHORS3). The *Cp36* sequence ends at the *EcoRI* site at which the *Cp38* sequence begins. Underlined are the regulatory chorion hexanucleotides, approximately 60 bp upstream of the transcription initiation site (see *Cp15 Promoter*).

three other transcription units in this cluster is unknown (Spradling et al. 1980).

The 13-kb segment is at the core of an 80–100-kb region that undergoes DNA amplification in the polyploid follicle cells prior to the time of programmed expression of *Cp36* and *Cp38*. This amplification results in a 15-fold increase in copy number (Spradling 1981). The amplification control element (ACE1), a *cis*-acting element, resides within a 3-kb segment that includes *Cp38*; a necessary portion of ACE1, at its upstream end, extends from –580 to –80 in the *Cp38* Sequence. In this region of *Cp38* are found the repeating pentanucleotide AATAC and related sequences (similar sequences are found in ACE3, the amplification control element of the third chromosome chorion–gene cluster). Whether other sequences within *Cp38* are also necessary for amplification is not known (Spradling et al. 1987). The mutation *ocelliless* (*In(1)oc*), is an inversion with one breakpoint 5 kb upstream of ACE1. Although homozygotes for this mutation amplify *Cp36* and *Cp38* in the new location, they do so to a reduced extent. The genes upstream of the breakpoint, which are left in place, fail to amplify but are correctly regulated (Spradling et al. 1979; Parks et al. 1986).

### *Developmental Pattern*

All of the genes in the X-chromosome cluster are expressed exclusively in ovarian egg chambers during the last 6 h of oogenesis, a time when these cells are actively involved in the synthesis and deposition of the egg shell. *Cp36* and

*Cp38* are transcribed during stages 10–13 of oogenesis (the chorion genes of the third chromosome cluster are expressed mainly during stages 13 and 14). Individual genes, however, have distinct temporal and spatial patterns of expression within the stages and cells mentioned, suggesting that each gene is independently regulated. With respect to *Cp36* and *Cp38*, in particular, *Cp38* transcripts accumulate in stages 11 and 12 while *Cp36* RNA is highest a little later, during stages 12 and 13 (Spradling and Mahowald 1979, Mahowald and Kambysellis 1980; Parks et al. 1986; Parks and Spradling 1987; Fenerjian et al. 1989).

A precise series of bursts of protein synthesis ensures that the different chorionic proteins are secreted in quick succession; this is accomplished by very fast mRNA turnover rates. Massive synthesis of each protein, on the other hand, depends on high levels of the corresponding mRNA. Because the mRNAs are short-lived, their accumulation depends on differential gene amplification in follicle cells, as described above (Mahowald and Kambysellis 1980; Parks et al. 1986; Parks and Spradling 1987).

### *Promoters*

Approximately 60 bp upstream of the start of transcription, both *Cp36* and *Cp38* carry the sequence TCACGT, the chorion hexanucleotide, which is thought to be involved in the regulation of all major chorion genes in *Drosophila* as well as other insects (Kalfayan et al. 1985; Kafatos et al. 1985).

## *Cp36*

### **Gene Organization and Expression**

Open reading frame, 286 amino acids; expected mRNA length, 1,004 bases. The approximate position of the 5' end was defined by primer extension; the exact position was suggested on the basis of sequence elements. The 3' end was determined from a cDNA sequence. There is one 91-base intron after the Pro-16 codon (Spradling et al. 1987). There is a well-defined region of transcription termination between 0 and 210 bp downstream of the poly-A addition site (*Cp36* Sequence) (Osheim et al. 1986).

### *Promoter*

An 84-bp segment (–162 to –79), sufficient for correct temporal expression, was defined by studies of germ line transformants carrying a reporter gene and fragments of the 5' regulatory region. The reporter gene consisted of *lacZ* associated with the *Hsp70* basal promoter. These studies also suggest that the 84-bp segment may contain two or more regulatory elements: while the upstream half of this segment controls expression at the posterior pole of the

Cp38

EcoRI  
 -822 ATTCCTAATTGGAATAGCTAAAGATCCATATTTTCATCTTCAAATCTCTTTGCAACTAGAGATTATTTTTATCTGGCAATTATTAAGTATA -733  
 -732 CATTTTTATATGTACTTTAAACTGATGGTTTAAATCAGTTACATGGATTTTCTAAATTA AAAATGGTCATGTGAAGATAGCCACTCTCT -643  
 -642 AACAACTAATCACATTTATAGTAAGAAATACAATACAATACAATACAATACAATACAATACAATACAATAGAAGACAATCGAATCTGC -553  
 -----  
 -552 GCAATCCGTGTGAAATCAAGGACTACAGCTGGGTGGCTAATCATTTCCCCCTATCCACTTACACCTCGGATTACCTCTTATTCGACTC -463  
 -462 CCGGAGTCTTGTGTCTGCCAATGCGGAECTATTTTCGCTATCTGAACAGACGTTCCGACCTCGATATGCGGCAAGATTACAGCCCCGGC -373  
 -372 TGTGATTCCGATTCCGGTGGCAATGTGTTCTGTTGTTATTGTAAAACGGGCAATGGCAACTGGGCAGTGGGCAGTGGGGTTTTCGGGTTGT -283  
 -282 GGCTCTACGTAAGTGAAGAGACGCCGTGATATGCGCTGGCAGCGATGCGTGCATCTATCAACATTTGGCCATGTTCTGTTGCGATCG -193  
 -192 CGTGGGCCCGGAGCGGAACAGCCGCGACCCGGAGTTGGCATCAATCCAATGTACAGTACCCGGAGCCGGAGACGCGTCCGGAGCATATTT -103  
 -----  
 -->-76.  
 -102 AAAGTAGTCGGCCACCAATGGAGGGCAGCAGAAGACAGCAGACAGTCCAAGCGGGAGCACACCAGAAGCCGAAGAGCAACTGGAAGTCA -13  
 ---  
 -12 ACTGGGAGACAAGATGACGAGATCGACCTACATTTGGGCGCTGGCCGCTGCCTGATCGTAAGTGTTCAGCTCGAAATCTTAGGATTAA 77  
 MetThrArgSerThrTyrIleTrpAlaLeuAlaAlaCysLeuIle (15)  
 78 TCCAAGAAAACCAAGTCTATCAATCTGACTGCTTTCGTTTGCCATGTAATCGTACATGAAAAGCAAATTGACTTTCCTTTAAATT 167  
 168 ACTTGA AACGGAATCAAGTATCTATCGATGCTAGACTTATTTAAGTATATGTATATGTCGATCCAATTCTAATCCACCCCCCCCCCT 257  
 258 CAATTTACTTTTAGGCCTGTGCAAGCGCAACTACGGCAGTCCCAGGGCTATGGACCCGAGTCCGGAAGCGGTGCCTCCGATGGCCGGTG 347  
 AlaCysAlaSerAlaAsnTyrGlySerSerGlnGlyTyrGlyProGluSerGlySerGlyAlaSerAspGlyGlyAla (41)  
 348 CTGATGCCGCTTCAGCGGCCGACGAGCTGCCGGCGGTGCCGGTGGAGCTGGTGGCAGTACGGTGGTCAACGCCGGTGCCTGGTGCCTC 437  
 AlaAspAlaAlaSerAlaAlaAlaAlaAlaAlaAlaGlyGlyAlaGlyGlyGlyAlaGlyGlyGlyTyrGlyGlyAlaAsnAlaGlyAlaGlyAlaL (71)  
 438 TCGAATCCGGAGCCGATGCCGCCGGTGTGGCACAGGCTGCCAGACGCTACGGATCCGACCGAAGCATTCCGTACAAGCCGGTGAACA 527  
 euGlySerGlyAlaAspAlaAlaGlyValAlaGlnAlaGlyGlnSerSerTyrGlySerAspGlnAsnIleProTyrLysProValAsnT (101)  
 528 CCAAGGGTAAACCCTGACCTCATCGATCACCTACCCGAGACAAGGGCGAGATCCTCATCCATCGTCCCGCTCCCATCATTGTCAAGC 617  
 hrLysGlyAsnThrLeuThrSerSerIleThrTyrProGlnAsnLysGlyGlyIleLeuIleHisArgProAlaProIleIleValLysA (131)  
 618 GTCCGCCACCAAGGTGCTGGTGAACCATCCACATTGGTGGTTAAGCCCGCTCCCGTGGTGCCTCCACAAGCCCCAGCAATCGTTCTCC 707  
 rgProProThrLysValLeuValAsnHisProProLeuValValLysProAlaProValValLeuHisLysProProAlaIleValLeuA (161)  
 708 GCAAGGTCTACGTCAAGCACCACCCAGCTCGCTCAAGGTTGAGCCCGTTCGTCATGTGGTCAAGCCCCAGCAGAGAAGTACTTTG 797  
 rgLysValTyrValLysHisHisProArgArgValLysValGluProValPheValAsnValValLysProProAlaGlyLysTyrPheV (191)  
 798 TCAACGAGAACAAGCAGGGCTACGGACAGGGCTCGCAGTCCCACGGACACGGCCATGGACACGGTGGCCATGGACACGGACACAGCGGAC 887  
 aAlAsnGluAsnLysGlnGlyTyrGlyGlnGlySerGlnSerHisGlyHisGlyHisGlyHisGlyHisGlyHisSerGlyHis (221)  
 888 ACGGACACGGTGGACACGGTGTGGACCCCATGGTCTGGACCCCATGACGGTGGCGGTGCTCTGCCCGCTTACGCTTCCGGGAGCTGATT 977  
 isGlyHisGlyGlyHisGlyAlaGlyProHisGlyProGlyProHisAspGlyGlyArgAlaLeuProAlaTyrAlaSerGlyAlaAspS (251)

978	CCGCTGCCGCCAGCGCTGGCTATCAGCTGCTCCAGAGCGGCAACCAGGGTCTGTCCGCTCTTGCCAACATCGCCGGCGAGCGTGAGGGTC	1067
	erAlaAlaAlaSerAlaGlyTyrGlnLeuLeuGlnSerGlyAsnGlnGlyLeuSerAlaLeuAlaAsnIleAlaGlyGluArgGluGlyP	(281)
1068	CCTATGGTCCCGCTCCAAGCCATCAGCACTATAGCGCCGGTCCAGCCGGACATGGCGGCTATGCTGCTCCCGCTATTAGGTAACAGATG	1157
	roTyrGlyProAlaProSerHisGlnHisTyrSerAlaGlyProAlaGlyHisGlyGlyTyrAlaAlaProAlaTyrEnd	(306)
1158	CGGAGGAGTTACGGATTGGATGACTGCTGCGGCTCCGGAATCAACTGAAGCGGCTGGTTTAGTCATTTCGCTTATCCGGCTGATTAGTTAC	1247
1248	TATGTTTTTTTTTACAAAAAACAACACAGCCTGATCGACCAACGCCCATGCCTACGCCACGCCCACTCATGCACACCCCAA	1337
.338	TACCACCCACTCACCCATTCAACGGCCAGGAGGGCGTGGCACTCAGGTTTCTTTGCAAAAACAAATAAAAAATTTGAACAAAAA	1427
428	AACAATTATACCCAAGCTGACTGTTGTTTTTCGATGAAGGGTAAATCTAGA	1478
	(A) <sub>n</sub>	

*Cp38* SEQUENCE. Accession, X05245 (DROCHORS3). The *Cp38* sequence begins at the *EcoRI* site at which the *Cp36* sequence ends. The bases underlined between -615 and -572 are part of ACE1. Also underlined are the regulatory chorion hexanucleotides, approximately 60 bp upstream of the transcription initiation site (see *Cp15 Promoter*).

egg chamber and the proximal half controls expression at the anterior pole, expression over the entire egg chamber requires the intact segment. A more distal element (-1,243 to -457), even though apparently not required, was found to allow weak expression (Tolias and Kafatos, 1990).

## *Cp38*

### Gene Organization and Expression

Open reading frame, 306 amino acids; expected mRNA length, 1,290 bases. The position of the 5' end was determined by primer extension and S1 nuclease mapping. The 3' end was obtained from a cDNA sequence. There is one 226-base intron after the Ile-15 codon (Spradling et al. 1987). There is a well-defined region of transcription termination between 220 and 585 bp downstream of the poly-A addition site (*Cp38* Sequence) (Osheim et al. 1986).

### Chromosome 3 Cluster *Cp15, Cp16, Cp18 and Cp19*

Synonyms: *S15, S16, S18 and S19*

```

          1                               50                               100
Cp15 .MKYLIVCVT LALFAYINAS PAYGNRGGYG .....GGYGGGYG. ....PVQR VVYEEVPAYG PSRG....Y NSYP...RSL RSEGNNG...
Cp18 MMKFM.CIC LCAISAVSAN SYGRPRGGYG .....GAPVGGYAY QVQPALTVKA IVPSYGGGYG GNHGGYGGAY ESVPVPVSSV YSGANVGSQY
Cp16 ....MSATLR LLCLMACCVA LAVANRPHYG .....G..... .....SGYG ASYGDVVKAA ETAEAQASAL TNA.....
Cp19 MNKFATLAVI FCACIVGSCY ANYGGQSYG QRSYGDSSA ASAASSAAA GAEGQRYER PVEIIAGGYR GSYAPEILRP IQVSGGYGGE RRGYNGGNHR
CON --K----- L----- ----R--YG ----- -G----- ----V---GYG -S-G----- ----- --N-G---

          101                               150                               193
Cp15 SAA..... AAAAASAAV NPGTYKQYAI PSYELDGARG YEIGHGYGQR AY*.....
Cp18 SGS..... GYGGAPPVDA QAIALAKLAL AAPSAGAPLV WKEAPRYAQP VYPPTSIVNQ EYGHSEKVKG GSAAAAASSV AAGKKGYKRP SY*
Cp16 .GA..... AASAAKLDGA DWYALNRYGW EQGRPLLAKP YGPLDPLYAA ALPPRSFVAE VDPVFKKSQY GGSYGENAYL KTDAKLGVVA I*.
Cp19 RAGYGPRWTV QPAGATLLYP GQNNYKAYVS PPEYSKVILP IRPAAPVAKL FVPENQYGNQ YVSQYSAPRS SGY*.....
CON ----- --A----- --Y----- --P-----

```

FIG. 8.1. Comparison of amino acid sequences for the chorion proteins in the chromosome 3 cluster. The sequences were aligned using the GCG *Pileup* program. The CON(sensus) line indicates positions at which three or more of the sequences agree.

**Chromosomal Location:**  
3L, 66D11-15

**Map Position:**  
3-[26.5]

## Products

CP15, CP16, CP18 and CP19 (chorion proteins of 15, 16, 18 and 19 kD) are four of the six major chorion proteins; the other two are products of *Cp36* and *Cp38*, which occur on the X-chromosome (Petri et al. 1976). These proteins are localized mainly in the exochorion and in the outer portion of the endochorion (Parks and Spradling 1987). The 20 or so N-terminal amino acids in each protein probably represent signal peptides (Waring and Mahowald 1979). These basic proteins are rich in Gly, Ala, Pro and Ser (residues that represent approximately 50% of the total) and Tyr (Petri et al. 1976). As in chorion proteins CP38 and CP36, there are Ala-rich stretches but no pattern of strong sequence similarity (Fig. 8.1).

## Organization and Expression of the Cluster

This cluster comprises four transcription units arranged in tandem (Fig. 8.2). In size, developmental expression and differential amplification, it is quite comparable to the X-chromosome chorion-gene cluster (Spradling et al. 1980).

The conserved position of introns (in all chorion genes but *Cp16*) and the presence of certain sequence elements in the 5' regions of the major chorion protein genes are suggestive of a common phylogenetic origin for all chorion genes in this cluster (Levine and Spradling 1985; Spradling et al. 1987; Wong et al. 1985). Although various *Drosophila* species show considerable divergence with respect to specific chorion-gene sequences, the disposition of the genes and general organization of the two clusters are remarkably conserved (Fenerjian et al. 1989).

Amplification (see Chorion-Gene Cluster on the X-chromosome) reaches 60-fold in the third chromosome cluster, and the amplification control element, ACE3, resides in a 3.8 kb fragment that includes the genes *Cp15* and *Cp18* (Levine and Spradling 1985). Within this segment, ACE3 sequences essential for amplification have been localized to the interval -673 to -163 of *Cp18* (*Cp18* Sequence). A 440 bp segment is capable of autonomous amplification,

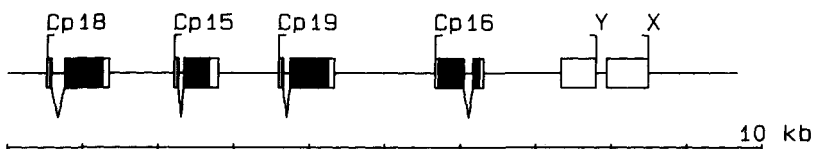


FIGURE 8.2. Chromosome-3 cluster organization. X and Y are two nonchorionic transcription units.



*Cp18*

-563	AAGCTTAGTGCGGCAGTTTGAAAGTGAACGGTTGTGTTATAATTTTATTGTAATTTTATCTCAATTTTTTTGCTTTTGTATATAAA	-474
	-----	
-473	TTCTACCAACGCAGCAGAATTTTCAGGCCACTGCCTTGACTTCACTGTGTCCTGAAAAATCGGTGTCAAGCTCTCGGCACCGTGGGGCA	-384
-383	AAGCAACTGCAATACTGATCGAAACTATGCGGATCCGGAGCACGAAGAGTCATGCGGTGCGAATCTTACGTAATGGGTCTCGTCTCTGGT	-294
-293	AGACGATGGCGTAAGCACAGACGCCCTGCTATCTGGACCGCCGAATTGAGAGCCAGCATTTTGGCCAGTGGGATTCGGCTGGCTGCA	-204
-203	CGTCTCCGGCGGCTCTCAAGATTGCTGGACAAAGAGGCGAGGCTGGAAC TCGCTCTCCGGGAACCGGAGACCCGAACTTGCATCAT	-114
	-->-43	
-113	ATTCGTCACGTAAGAGTTGGGCTCTGCCTGGATCTGGTATAAAAAACAAACATTGCGCCAGAATAAGACATTAGTTACCTTCGCATCGA	-24
	-----	
-23	TCAACTAACCAACTCAGCCTCAGAATGATGAAGTTCATGGTAAGCTTAAGTTCCAATATTGTTTCACCTCAACACCTCAACTGGCTCCAG	66
	MetMetLysPheMet	(5)
67	TATGATCCTTTTAAATAAAATATAACTACATATTATAATAATTGAAATAATATGATTGGATCTTCTTTTCTAATGCACCTTCAGGCATA	156
157	CCCAAATTAATGAAATTTTTCTTGAATCCCTTAGTGATCTGCCTCTGCGCCATCTCTGCCGTTTCGGCCAACCTCTACGGACGCTCCCC	246
	CysIleCysLeuCysAlaIleSerAlaValSerAlaAsnSerTyrGlyArgProA	(24)
247	GTGGTGGATACGGTGGTGGCCAGTCGGTGGCTATGCCTACCAGGTGCAGCCTGCCCTGACCCTTAAGGCGATCGTTCCTCATAAGGTG	336
	rgGlyTyrGlyGlyAlaProValGlyGlyTyrAlaTyrGlnValGlnProAlaLeuThrValLysAlaIleValProSerTyrGlyG	(54)
337	GTGGATACGGCGAAACCATGGAGGATATGGCGGTGCCACGAGTCGGTGCCTGTGCCGTGCTCTGTCTACAGCGGTGCCAATGTGG	426
	lyGlyTyrGlyGlyAsnHisGlyGlyTyrGlyGlyAlaTyrGluSerValProValProValSerSerValTyrSerGlyAlaAsnValG	(84)
427	GATCTCAGTACTCCGGTCCGGCTACGGCGTCCCCACCAGTCGATGCCAGGCCATTGCCCTCGCCAAGCTCGCCCTGGCCGCTCCCA	516
	lySerGlnTyrSerGlySerGlyTyrGlyGlyAlaProProValAspAlaGlnAlaIleAlaLeuAlaLysLeuAlaLeuAlaAlaProS	(114)
517	GCGCTGGAGCTCCTCTGGTCTGGAAGGAGGCTCCCGCTACGCCAGCCCGTCTATCCCCCACCAGCTACGTGAACCAGGAGTACGGAC	606
	erAlaGlyAlaProLeuValTrpLysGluAlaProArgTyrAlaGlnProValTyrProProThrSerTyrValAsnGlnGluTyrGlyH	(144)
607	ACAGCGAGAAGGTGAAGGGAGGCTCCGACGCCGCTGCTGCCAGCTCCGTGGCCGCCGAAAGAGGGCTACAAGAGGCCAGCTACTAAG	696
	isSerGluLysValLysGlyGlySerAlaAlaAlaAlaAlaSerSerValAlaAlaGlyLysLysGlyTyrLysArgProSerTyrEnd	(172)
697	TGGCAAAACGTTGAACAGTGAACCAAAAACCTTACCTGCCAATAAGGAACTAGGTCATAATAAAAAGCCAAAACATCAAGACTTAAAAT	786
	-----   (A) <sub>n</sub>	
787	TTTGAGTACTGTATTCTTGGTGGGTTTTAGTTTCGGGCCAAGAGTTGAG	836

*Cp18* SEQUENCE. Strain, *Oregon R*. Accession, X02497 (DROCHORSG). The underlined bases between -530 and -500 represent a segment that is a part of ACE3. Also underlined are the regulatory chorion hexanucleotides. The *Cp15*, *Cp18* and *Cp19* sequence segments occur contiguously in genomic DNA in the order shown in Fig. 8.2.

but sequences outside of it also seem to influence the process (Orr-Weaver and Spradling 1986; Carminati et al. 1992).

### *Developmental Pattern*

Transcription of these genes occurs during oogenesis, a little later than transcription of Cp36 and Cp38: Cp16, Cp18 and Cp19 are expressed mainly during stage 13 and to a lesser extent during stage 14; Cp15 is expressed almost exclusively during stage 14 (Mahowald and Kambysellis 1980; Parks and Spradling 1987; Fenerjian et al. 1989).

### *Promoters*

As in Cp36 and Cp38, the sequence TCACGT is found approximately 60 bp upstream of the transcription initiation site (except for Cp16, in which it is found 80 bp from the 5' end). Other sequence elements in the neighborhood of this hexanucleotide are also present in Cp18, Cp15 and Cp19 (Levine and Spradling 1985; Wong et al. 1985).

## *Cp15*

### **Gene Organization and Expression**

Open reading frame, 115 amino acids; expected mRNA length, 519 bases. One 71-base intron is present after the Leu-4 codon. The approximate position of the 5' end was determined by S1 mapping, and the first nucleotide transcribed was assigned on the basis of similarities to canonical *Drosophila* sequences. The 3' end was determined from the sequence of a cDNA clone (Cp15 Sequence) (Levine and Spradling 1985; Wong et al. 1985).

### *Promoter*

The 73-bp segment of DNA from -162 to -90 seems to be necessary and sufficient for correct tissue and temporal specificity in the expression of this gene; sequences between -858 and -162 may contribute to an elevation of the transcription rate. The TCACGT chorion hexanucleotide from -104 to -99 is indispensable for transcription as well as for follicular specificity. Another positive *cis*-acting element, between -116 and -107, activates expression late in oogenesis (stages 13 and 14). Element(s) between -162 and -124 act negatively to suppress early transcription (stages 11, 12 and early 13) (Mariani et al. 1988; Shea et al. 1990). By gel retardation assays, two protein-DNA complexes were detected that involve the -116/-107 and -104/-99 sites; there is partial overlap of the binding sites. Two cDNAs produce proteins that bind specifically to these sites: chorion factor I (CFI) binds to the chorion hexanucleotide while CFII binds to the late activator site (-116/-107). Both

*Cp15*

-857	ATCTGCATATCTTAGCTGAATTGGCAAAGACTTGCGGTTTCATTGCAATGCCAAGCGATACTTTGAGCCAGCAAAAATTTCTTGGTTTCGT	-768
-767	AGTTAAATGAAAATGCTGCTTAAAGTGCTAAAGAATAAATGTGCATGGCGAATGAAGCTGCAAAGCTAAAACATAAATTTGTGGGGCC	-678
-677	AATTTAAAACATATAGTTTGTCAAAGAGCCTTGACTTTTTTAAGTCACCATAAGTAAAGAATCTATTACATAAAAACGCGATTAGATAGAA	-588
-587	TATAGTTTTGCTTGAATTTATGTTTTTGTAAAATTTCAAATGATTGAAATACTTTAAAATGTTTTAGTTATAATTTAAGTTTTGTATG	-498
-497	TGACTAGTAATCACTTTAAAGGAATGACTCTATATAGGTTTTATCAGAAAAACCGCTGGAACCAAGTTCTAGAAGAATCCTCACTTAGAC	-408
-407	AAGCCAAGTCCGGACACAACCGATCTGGAAACCATTACCCCGAGAATGTGGATAATATAAAGTTCAATTCACAATAATTTGGAGTGTA	-318
-317	TTCGAAAAAACAACGCGTTCGTGGTTCACATTGGAAGAGTCGCGTTCGTAGTGTATCACCACCAACACCCGGTAGAATAGCACATC	-228
-227	GCGTAACCAAGCGATTTTATAATGGCTTGACAACAAGTACATAAATCAAATGTGAGTATATCCAGCCGGGCAATTAATAATGCCATTT	-138
-137	CTGGGCTGAAACAGAAACATTAGTGTATATAGGTCACGTAATGTCCAGGCTAAAATTTGCGTATAAAAAGCGAGCGTTTCTGGTCGGTAA	-48
	-----cf2-----cf1	
	-->-44	
-47	ATCATAGTTTGATTGATTACCCCAAACCAACAAAACTAAGCACTACCATGAAGTACCTGGTAAGTTGTGGTAGTCCCGTAGAAGGAGTG	42
	MetLysTyrLeu	(4)
43	GCAGCCAACGTATCCTCCGGATTTCCCTTTTACCTTCAGATTGCTGTGTTACCTGGCCCTTTTCGCCTACATCAACGCCAGCCCAAG	132
	IleValCysValThrLeuAlaLeuPheAlaTyrIleAsnAlaSerProA	(21)
133	CGTACGGCAACCGTGGAGGTTATGGTGGTGGCTACGGTGGTGGCTACGGTCTGTTACGCGCGTCTACGAGGAGGTGCCCGCCTACG	222
	1aTyrGlyAsnArgGlyGlyTyrGlyGlyGlyTyrGlyGlyGlyTyrGlyProValGlnArgValValTyrGluGluValProAlaTyrG	(51)
223	GACCATCCCGTGGCTACAACAGCTATCCCGCAGCCTGCGATCGGAGGGTAATGGAGGAAGTCCGCTGCCGCTGCCGCGCTCCGCGG	312
	1yProSerArgGlyTyrAsnSerTyrProArgSerLeuArgSerGluGlyAsnGlyGlySerAlaAlaAlaAlaAlaAlaAlaSerAlaA	(81)
313	CTGCCGTGAATCCCGAACCTACAAGCAGTACGCCATTCCTCCTACGAGTTGGATGCCGCTCGTGGCTACGAGATCGGACACGGCTACG	402
	1aAlaValAsnProGlyThrTyrLysGlnTyrAlaIleProSerTyrGluLeuAspGlyAlaArgGlyTyrGluIleGlyHisGlyTyrG	(111)
403	GCCAACGTGCTTACTAATTTCTCGCTTCATCGGCAGTGAATGAACTATCGACTCCTTGCTAAAATCCTCGAGTGGCTGTCATGGCGAAAC	492
	1yGlnArgAlaTyrEnd	(115)
493	TCTGAGAATCAGTGAATAAAAGCAGCTTGAACGCAATGGAAAATACCGAAAAGAAATACGTATTGTGTGTTTTGCATTGTGACATACTTT	582
	-----	{(A) <sub>n</sub>
583	TCAGCGCATT	592

*Cp15* SEQUENCE. Strain, *Oregon R*. Accession, X02497 (DROCHORSG). Underlined is the regulatory chorion hexanucleotide. cf1 (which overlaps the chorion hexanucleotide) and cf2 indicate the binding sites of chorion factors I and II respectively. The *Cp15*, *Cp18* and *Cp19* sequence segments occur continuously in genomic DNA in the order shown in Fig. 8.2.

CFI and CFII RNAs are more abundant in follicle extracts than in extracts from other tissues, and CFII protein is more abundant in nuclear extracts from late follicles than in extracts from early follicles. CFI corresponds to the product

*Cp16*

-922 TGGCATCGAGTGGCCACAATTCTTGGGAAAACCTGTGCGTTGAAATTAACCACATGTGTGATAGAGTTTGTCTGTTGAATAATTTTAA -833

-832 TTTTCGTAAAAGTGAATTTATGTTTTGTGTAAAGCCGAAATATAAATAAAGTTTAAATATTACTAACTAACCGTACGATCGTTTTTC -743

-742 ATAAAACAGCTTAAATTTGGTATTAGCAACATTGTAATATTACATTAATAAATAATAAGAATGAAATCTAATAAAAAGGCATAACT -653

-652 TAAATGCCAATGTATTGAAACATAACTTAGAATATTGACGTAATAATCCACTTTGTTGCTATGCACATTTTTGTCCATTTTTAAATAAA -563

-562 TTCATAGAAGTGTGTTACGATCCACAACTTTTCAAAAACATTGTCAGCTTTAAAACCTAGAGTTGCCCGACGTCCAAGACTTTCGT -473

-472 ACGTCGCTGGTCCGGTTAGTGTTCATTGATCGGTGGCTTAACCCAGTTGGCCCGATTTCCGATGCGTGCATGGGCCGGATCGCCGTGG -383

-382 AGTACGCCAAAGCCCGATACCGCACACACCAGAAGCGAACAGAGCGTGCCGAGCAGGGGGAAGTCGTATCAATGGAGCAGCTTCTCGG -293

-292 GGTCCCGGGTTGTGGCAGTCCGCAATTTGGCGCAATTTCAATGAGCATAGAAATGGAGACGATCCCGTGGCCTTATCGCCCTGG -203

-202 GCCGGAGGGGACGGAGGGGGCTGGAGATGCTGCCAGTGGCGGCCCCCGAAAGTGACTGGTCATCGAGGTGGTTGGTCACGTCGTGGTGA -113

-----

-112 GCTCACAAATCGCGGAGCAGCTCAATGGTGTGCTATAAAAAGCAATTTGGACACACGCTCTGGTTAATTAGTTTTCGAAACAGTCCGTT -23

-----

-22 CCTCGCACCACCACAAAAAAATGTCCGCCACCTACGCCCTCTCTGCCTGATGGCTGCTGCGTCGCCCTGGCTGTGGCCAAATCGCC 67  
MetSerAlaThrLeuArgLeuLeuCysLeuMetAlaCysCysValAlaLeuAlaValAlaAsnArgP (23)

68 CCCACTACGGCGGATCCGGATACGGAGCCAGCTACGGCGATGTGGTTAAGGCCGCTGAGACCGCCGAGGCTCAGGCTTCTGCCCTGACCA 157  
roHisTyrGlyGlySerGlyTyrGlyAlaSerTyrGlyAspValValLysAlaAlaGluThrAlaGluAlaGlnAlaSerAlaLeuThra (53)

158 ACGCCGCCGGAGCAGCTGCCTCCCGCCCAAGCTGGACGGTGTGACTGGTATGCCCTCAACCGTTACGGATGGGAGCAGGGTCGCCAC 247  
snAlaAlaGlyAlaAlaAlaSerAlaAlaLysLeuAspGlyAlaAspTrpTyrAlaLeuAsnArgTyrGlyTrpGluGlnGlyArgProL (83)

248 TTCTGCCAAGCCCTACGGTCTCTGGACCCGCTATACGCTGCTGCTGCCACCACGCTCCTTCGTGGCTGAGGTCGATCCAAGTGGGT 337  
euLeuAlaLysProTyrGlyProLeuAspProLeuTyrAlaAlaAlaLeuProProArgSerPheValAlaGluValAspProV (111)

338 TCCTAAGCTAAGCTACAACATGGATAATATTGTTTATCCTTATGATTTGGATTGACTTCATAGCACCCGCTTGGCACCATACTTACCTT 427

428 CTTTTGTATCGTCTCTACCTTTCAGTCTTCAAGAAGAGCCAATACGGCGGATCTTACGGCGAGAATGCGTACCTGAAGACCGACGCCAAA 517  
aPheLysLysSerGlnTyrGlyGlySerTyrGlyGluAsnAlaTyrLeuLysThrAspAlaLys (132)

518 CTGGGTGTTGTGGCCATCTAAGAGCTGGATTGTATAGCTCCAAAAGTGTTAATAAATAGTGATAGCTTAAAGCAATATAAATCAATGGAA 607  
LeuGlyValValAlaAlaIleEnd ----- (138)

608 AATTCATTTATGGGCTGGGAAACCAAATGAGCGAATCTTTATTTGCAAATGAGAATGTTGTTTACTCCGACAACTTCTGCCTATTT 697

698 TTGAATGCCATGAACTTTAGATGGTTAAAAAAAACCTCAAAAACCTTGGATTGGCTATGCCAAACTTCACTTCTTACTTGAAGTCCACTAAG 787

788 GTTCGCAGCTACACCATTTCTTGAATCTTGAAGACCCCCCAATTAGTAAAACCGAATTCACCTTACAATTTCTTATTGTTATTATTAT 877

878 GAATAGAATTTGTTTTATTGCAAGATACAAAGTAAAAATGTGAAAATGCTCAGTTTTGTTGATGCTGATTTAATGTAAAAATCAAA 967

968 TTCGTTACGAGCACAGAAATTTACCTACTAAACATAAAGTGAACAAAAACAAATAGTAGAAGCGGTTGAACTCGGTTAACTCGATG 1057

1058 CTGCGGTGGCGTCTTAGTGGGATATTCGGTGACGATTATCATTCCATTTCAAGTTATTAAAGTTTGTGCTTTTCGTTCAAAATGGGCT 1147

(continued)

*Cp19*

-790	CGGGTTAAGATTTAGCGGTGGGCTATTATTATTATCCACACACAAGATGGGTTTCAAAGTGGGGCAGCTAGAATATTCCTGCGGCAGA	-701
-700	TTGTACAATACTATATAGAAGTACTATTGCACTTTAAGCTACAAGTCGACAGGTTAAGCTTCAGTGACTCAAGAATTTAGTCACCTATG	-611
-610	AAACCCCTAGTTTCACTAATAGATTCTTAGACGAACATCTTAAATGTATAATCAACAACAAATGGCTATGTATATTTACAATAACATATT	-521
-520	TGCCAATGTGCAAAAAGGCATAGACTTTGAAGTTATGTTTTATCGTTAAAATTTGGTTTGTCTGTTTACTTGAAGGTATAGATAATATT	-431
-430	ATAGAATCCATATCCAATAACCATTTGGTCAGTTGTGGGCCGTTATCCATTAAACCCGCTGGCTTCCCACACGCCCAAATGCAACCA	-341
-340	TTGATTTTGGGCCTCAGTTGGGAGCATCTGCATCTGCCACCCCAACGAAAGTCAACCGCGAATGGAGGCGATACGATACGCTGCGGTG	-251
-250	AGCAACCTGCTCGAGCCGAAACGAGCTCAACGTGGAGCCCGATATCTGGCTAGGAAAAGCTAGAAATCCACAGAAAAGTCCCCCAACAAA	-161
-160	CTGGCCGAGAAGAGACGGCGAAGCCAGCTCTTGAGCCGTGATAAATTTCTGGGCGAGATCACGTTTCGAGTGCAACAATAAATTTGCTTA	-71
	-----	
-70	TATAAAGAAGTGTGCTTGCCATTTAATATGTTAATTCAGCCAACGTGCCAAAACCCATACATCATAGCCATGAACAAGTTCGCTGTAA	19
	-----	MetAsnLysPheAla (5)
20	GTGTCCCTGAGAACCCTCCGTATTCCTGCCGCTTTTTCATTTCCGGACTTATGCTAACTGAAAGTTTCTGTATTTCCAGACTCT	109
		ThrLe (7)
110	GGCAGTCATCTTCTGCGCTGCATCGTGGGCAGCTGCTACGCCAACTACGGTGGCCAGCAGAGCTACGGACAGCATCTTACGGTCAGGA	199
	uAlaValIlePheCysAlaCysIleValGlySerCysTyrAlaAsnTyrGlyGlyGlnGlnSerTyrGlyGlnArgSerTyrGlyGlnAs	(37)
200	TAGCTCCGCCCTCCGCCCCAGCTCAGCAGCTGCTGCTGGAGCCGAGGGTCAGCAGGTTATGAGCGCCCGTGGAGATCATCGCCGG	289
	pSerSerAlaAlaSerAlaAlaSerSerAlaAlaAlaAlaGlyAlaGlyGlyGlnGlnArgTyrGluArgProValGluIleIleAlaGly	(67)
290	CGTTACC CGCGCAGCTATGCCCCGAGATCCTGCTCCATCCAGGTCAGCGGTGGATATGGCGGTGAGCGACGTGGCTACAACGGTGG	379
	yGlyTyrArgGlySerTyrAlaProGluIleLeuArgProIleGlnValSerGlyGlyTyrGlyGlyGluArgArgGlyTyrAsnGlyGly	(97)
380	CAACTACCGCTGTCGGCTACGGACCCCGTTGGACTGTCCAGCCCGCGGTGCCACCCCTCTGTACCCCGCCAGAACAACTACAAGGC	469
	yAsnTyrArgArgAlaGlyTyrGlyProArgTrpThrValGlnProAlaGlyAlaThrLeuLeuTyrProGlyGlnAsnAsnTyrLysAl	(127)
470	TTACGTCTCGCCCCGGAGTACAGCAAGGTATCCTGCCATCCGCCCGCTGCTCCAGTGGCCAAGCTTTTCGTCCCAGAGAACCAGTA	559
	aTyrValSerProProGluTyrSerLysValIleLeuProIleArgProAlaAlaProValAlaLysLeuPheValProGluAsnGlnTy	(157)
560	TGGCAACCAGTACGTTAGCCAGTACTCTGCACCCCGCAGCAGCGGCTACTAAGCGCATACATGATTATCCCAGCCAACCTGGCGGATAC	649
	rGlyAsnGlnTyrValSerGlnTyrSerAlaProArgSerSerGlyTyrEnd	(173)
650	TTGATCTCAGCCTGATCGTGTACATAATAACAACAAGAAAAAATCATAATCATATTTGGAATATATATTTTTCGGGGCTTTTAGGTTAT	739
	-----	(A) <sub>n</sub>
740	TTTTTATATCTATGAGAAAAAAATTTTCGGGCTTTTCGAGCTCAAATGCAGCTGCAGCAGCTGTTTCAGAGTGGGTGGAGCATGTTTCAT	829
830	TTGATTGCACTGCCACCGGAATGTCTTTGAGTGGCTCGGCGGAAACGTGCTCCGGATTGCTTGCCTCCGGTTCGCTCGCAAATCACTC	919
920	AGCAAGCCATAAACATTCAATATTTATTGTGTGCTCAGTCAGTCAATAATCTTGGGGCCAGAAAGCGCCACAGTTCGCCGAGTTTCCCGATT	1009
1010	GCCGCGCTCATTTTCATATTTCTGTATTCTGGCTGGTAAGCAATCGCATCTGCTGACTTGTTTGGGGCCAAACTCTTGGCCAAGAGCTT	1099
1100	CAATGCTGCTGGCCATCGCTTGACATTCGAGTCGAGCGTGAATCACGGCAAGAATTC	1156

of the gene *ultraspiracle*, a steroid hormone receptor protein; and CFII contains C<sub>2</sub>H<sub>2</sub> zinc finger motifs (Shea et al. 1990).

### *Cp16*

#### Gene Organization and Expression

Open reading frame, 138 amino acids. One intron is present within the Val-111 codon. The position of the 5' end was assigned on the basis of similarities to canonical *Drosophila* sequences (*Cp16* Sequence) (Fenerjian et al. 1989).

### *Cp18*

#### Gene Organization and Expression

Open reading frame, 172 amino acids; expected mRNA length, 649 bases. One 176-base intron is present after the Met-5 codon. The approximate position of the 5' end was determined by S1 mapping, and the first nucleotide transcribed was assigned on the basis of similarities to canonical *Drosophila* sequences. The 3' end was determined from the sequence of a cDNA clone (*Cp18* Sequence) (Levine and Spradling 1985; Wong et al. 1985).

### *Cp19*

#### Gene Organization and Expression

Open reading frame, 173 amino acids; expected mRNA length, 653 bases. One 89-base intron is present after the Ala-5 codon. The approximate position of the 5' end was determined by S1 mapping, and the first nucleotide transcribed was assigned on the basis of similarities to canonical *Drosophila* sequences. The 3' end was determined from the sequence of a cDNA clone (*Cp19* Sequence) (Wong et al. 1985).

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*Cp16* SEQUENCE (page 83). Accession, X16715 (DROCHORS16). Underlined is the regulatory chorion hexanucleotide.

*Cp19* SEQUENCE (opposite). Strain, Oregon R. Accession, X02497 (DROCHORSG). Underlined is the regulatory chorion hexanucleotide. The *Cp15*, *Cp18* and *Cp19* sequence segments occur continuously in genomic DNA in the order shown in Fig. 8.2.

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# 9

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## Cuticle Protein Genes: *Lcp1*, *Lcp2*, *Lcp3*, *Lcp4*, *Pcp*, *Edg78E*, *Edg84A*, *Edg91A*

### Larval Cuticle Protein Gene Cluster on Chromosome 2: *Lcp1-Lcp2-Lcp3-Lcp4*

**Chromosomal Location:**  
2R, 44D

**Map Position:**  
2-[58]

#### Products

Members of the cutin family. These are four of the five major protein components of the third-instar larval procuticle (the main layer of the cuticle) (Fristrom et al. 1978; Silvert et al. 1984).

#### *Structure and Function*

These proteins bind chitin and can be solubilized from untanned cuticles with 7 M urea; upon tanning of the cuticle, they become cross-linked and insoluble. The solubilized (untanned) proteins have an apparent  $M_r$  of 8–20 kD. The only detectable modification of these proteins is the excision from each of them of the first 16 amino acids, the signal peptide; the resulting N-terminus is unmodified. Direct amino acid sequencing of 50–75% of the residues confirmed the sequence predicted from nucleic acids data (Fristrom et al. 1978; Snyder et al. 1982; Silvert et al. 1984).

#### *Tissue Distribution*

Like the other components of the cuticle, LCPs are secreted by epithelial cells, the epidermis, probably in response to the steroid 20-hydroxyecdysone (20-HE). During its life cycle, *Drosophila* produces five different cuticles, three

larval, one pupal and one adult. LCP1–4 contribute only to the third larval instar cuticle: LCP3 and LCP4 accumulate early in the third instar while LCP1 and LCP2 synthesis predominates late in the third instar (Chihara et al. 1982; Kimbrell et al. 1988).

### Organization and Expression of the Cluster

The four genes are clustered in less than 8 kb of DNA, and they are best regarded as two pairs: *Lcp1* and *Lcp2* versus *Lcp3* and *Lcp4*. The two pairs are transcribed divergently (Fig. 9.1). In the coding regions, the similarity within the *Lcp1*–2 gene pair is 91%, within the *Lcp3*–4 pair it is 85%; the similarity between pairs is approximately 60% (Fig. 9.2). Considerable similarities also occur in the 5' untranslated regions and in the 200 bp just upstream of the site of transcription initiation. The observed similarities suggest that the four-gene cluster evolved via an inverted duplication that gave rise to two ancestral genes followed by direct duplications of each of the two ancestral genes to give rise to the two pairs (Snyder et al. 1982).

A pseudogene carrying numerous disabling mutations lies between genes 1 and 2. It was probably generated by unequal crossing over between *Lcp1* and *Lcp2* (Snyder et al. 1982).

#### Developmental expression

*Lcp1* and *Lcp2* are transcribed primarily late in the third larval instar while *Lcp3* and *Lcp4* are transcribed primarily earlier, as might be expected from the pattern of protein synthesis (Snyder et al. 1982).

#### Gene Organization and Expression

Transcription initiation sites were defined by primer extension and sequence features. The 3' ends have not been determined (Snyder et al. 1982).

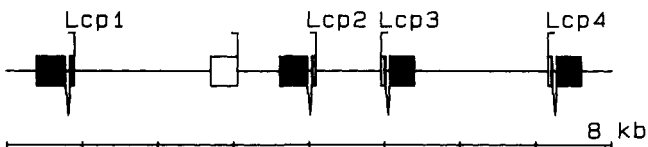


FIG. 9.1. *Lcp* cluster organization. Open box, pseudogene.

```

1                               50                               100
Lcp1 MFKFVMICAV LGLAVANPPV PHSLGRSEDV HADVLSRSDD VRADG...FD SSLHTSNGIE QAASGDAHGN IHGNFGWISP EGEHVEVKYV ANENGYQPSG
Lcp2 MFKFVMILAV VGVATALAPV ...SRSDDV HADVLSRSDD VRADG...FD SSLHTSNGIE QAASGDAHGN IHGNFGWISP EGEHVEVKYV ANENGYQPSG
Lcp3 MFKILLVCSL AALVAANA.. ..NVEVKELVND VQPDG...FV SKLVLDGSA SSATGDIHGN IDGVFEWISP EGVHVRVSYK ADENGYQPQS
Lcp4 MFKILLVCSL VALVAANE.. ..NPEVKELVND VQADG...FV SKLVLDNGSA ASATGDVHGN IDGVFEWISP EGEHVRVSYK ADENGYQPQS
CON1 MFK----CA- --L--AN--- -----V---D V-ADG---F S-L---NG-- --A-GD-HGN I-G-F-WISP EGEHV-V-Y- A-ENGYQP--

Edg78 MYKYLFCLAL IGCACADNI. ....NK DAQIRSFQND .ATDAEGNYQ YAYETSNGI. QIQEAGNANG ARGAVAYVSP EGEHISLTYT ADEEGYHPVG
Pcp MYLLVNFIVA LAVLQVQAGS SYIP...DS DRNRTLQND LQVERDGKYR YAYETSNGIS ASQEGLGGVA VQGGSSYTSP EGEVISVNYV ADEFGYHAHI
CON2 M-K-----AL --A-A-A-- -----D -A---S--ND -Q-D----- ---TSNGI- QS--G----- --G-----SP EGEH---YV ADE-GYQP-G
      ^           |

101                               150                               193
Lcp1 AWIPTPPPIP EAIGRAVAVL ESHPPAPEHP RHH*.....
Lcp2 AWIPTPPPIP EAIRAVAVL ESHPPAPEHP RHH*.....
Lcp3 DLLPTPPPIP AAILKAIAYI EANPSKN*..
Lcp4 DLLPTPPPIP EAILKAIAYI QAHPske*..
CON1 ---PTPPPIP EA1--A-A-- E-HP-----

Edg78 DHLPTPPVPP AVVLRALEYI RTHP..... PAPAQKEQ Q*.....
Pcp .....PQVP DYILRSLEYI RTHPYQIKDY YTGELKTVEH DAAAFNVYTR NIQDHTIPQS RPSTTPKTIY LTHPPTTSR PLRQRALPT H**
CON2 ---PTPPP-P --ILR---YI --HP-----

```

FIG. 9.2. Comparison of the four larval (LCP1–4) and two pupal EDG78E and PCP) cutins. The sequences were aligned with the GCG program *Pileup*. CON1 indicates positions where at least three LCP proteins have the same residue. CON2 indicates positions where the pupal proteins agree with the larval ones. Ala-16 is the last amino acid of the signal peptide. A caret under residue 4, indicates the presence of an intron in all the genes discussed in this chapter with the exception of *Edg91A*.

*Lcp1*

```

-707 AACATTAGGTTTTCTTAACAACTTTAATTGTCGCTAAAAAAGTATTTATTTCCGGTAGCCTCTATATTGAACAGGCTTTATTATCTTA -618
-617 TTTAATTTTACGAAATATAAAAAAATAATATAAGGCCTATGCATATGAAATATAATGTA AACACACACTTGAATTTGTTTAAAAGCAAAC -528
-527 TGCAACCTGTTGTCGAGAGGAATTGATAAAAAAAGAAGAAATGGTGCCAAGGTAGAGACACACGTTTATATATAACAAAACATCGAGT -438
-437 CTAAGAAGTCGGCGATGCTTTGTAGTCCATGGAGTCTTGATGGGACTACAAAAGTGGTTCACGGCCTGGCAATGCCAAGTCAAGCTCAA -348
-347 GGAGGGGATTTAATGAAGGGGCGGGTCAAACCTCGTTTCGATTTGGGATGCCACCCGACCCGTTTGCCCTTATTGATGCGATTGTTTCA -258
-257 TTTTAGCATCTATTAAGCGATTATATAGTACTTATCCCGTTGTTGGCATTGCTAAGCTGTCGCATGTGACGATGCTTTTTAATGGG -168
-167 TGTGGGCGCATCCGCGAAGTCAACCCATAACTCAGCGAACCAATGAAATGCAAGATGTAGAGTTTGTATATGGGTTCACTTTGGGTGGCA -78

          -----
          -->-41
-77 ATCATATAAAAAGGCTCTGCCGACCACAATCAGTTATCAGTCAACGTTTCGTTCTCGACAGACAGAAGTCAAGCAATATGTTCAAGTTT 12
          -----
          MetPheLysPhe (4)

13 GTAAGTGTCCGCAGGATACGAACCAACATACTCGATCCCTAACGAATGCCTATTTCTCCTTCAGGTCATGATCTGCGCAGTTTTGGGCCT 102
          ValMetIleCysAlaValLeuGlyLe (13)

103 GCGGTGGCCAAACCCCGGTGCCCATTCCTAGGCCGTTTCGGAGGATGTCACGCCGATGCTCTTCCCGATCCGATGATGTTCTGTC 192
    uAlaValAlaAsnProProValProHisSerLeuGlyArgSerGluAspValHisAlaAspValLeuSerArgSerAspAspValArgAl (43)

193 CGATGGATTGATTCCAGCCTGCACACCTCCAACGGAATCGAGCAGGCCGCCAGCGGTGATGCCATGGCAACATCCACGGCAACTTCGG 282
    aAspGlyPheAspSerSerLeuHisThrSerAsnGlyIleGluGlnAlaAlaSerGlyAspAlaHisGlyAsnIleHisGlyAsnPheGl (73)

283 CTGGATCTCACCCGAGGGCGAGCAGTCGAGGTTAAGTACGTCCCAATGAGAACGGATACCAGCCCTCGGGAGCCTGGATCCCCACTCC 372
    yTrpIleSerProGluGlyGluHisValGluValLysTyrValAlaAsnGluAsnGlyTyrGlnProSerGlyAlaTrpIleProThrPr (103)

373 TCCTCCAAATCCAGAGCCATCGGCCGCGCTCGCCTGGCTAGAGTCCCACCCACCAGCACCCGAGCACCCCGTCATCACTAGAACCT 462
    oProProIleProGluAlaIleGlyArgAlaValAlaTrpLeuGluSerHisProProAlaProGluHisProArgHisHisEnd (130)

463 CTATGAAAGCGGATCGCCTACGGACTGTTCCCGAAGACCTTTCGAACTATTAGCTTAAGTAATCGTACTGTTTGTAATAACACGCAA 552

553 TTGTTAACGGCAGAAACAGTTTGCAACCTTGACTTTGAATTTGGCAAACAACTGTAACGGTTTCGAACCCGCTACCCGTTTACCACC 642

          -----
          EcoRI
643 TTCGATTACTAGTTGTTTAGCAGTTCAGTACAATATGGTAATGTGGTCTCTACCTGGACCGTAAACCGAATTC 718
    
```

*Lcp1* SEQUENCE. Canton S strain. Accession, J01080 (DROCTCL1). The sequence of *Lcp1* extends to the first *EcoRI* site downstream of that gene.

*Lcp1*

Open reading frame, 130 amino acids; expected mRNA size, ca. 545 bases. There is one 64-base intron after the Phe-4 codon (*Lcp1* Sequence) (Snyder et al. 1982).

*Lcp2*

	_____HindIII	
-568	AACTCTGCCAAAAGCTTTGCGGGTTTTTTAAATTAACAGTGACATCCAAAATATTGAGATACAACAAAATGTCATAGGCAACTAGCA	-479
-478	CGTTAATATGCAGTATCACTTGCAGAAATCGTTTATTCGGGTATATTGTTTATTACCACCTTCGGAACCTTTTAAAATAGATGGGACTGCTA	-389
-388	TCAAGTGAAGTGTATTGGGTTTTTGATTTGTACAGGCATGATTGATAAAGACTTGGTCAACTCGAAACGTCATCGATGAGCACAGAAT	-299
-298	CCGAAAACCGTACTCCATCGCCCTACAAAATTTCTACCGAAGCATGTTTCATTTGGAACTGTTCAGCAGCGCAAGACTTGTTTTTTG	-209
-208	ACATTTGTATCGCAGAGTCAAGTGGAGAATTTATGGGCCCTGCCTTTGTTGGCATCATGGGCGTTTCGTGATAACTTAGATTTGGCCCA	-119
	-----	
-118	AAAAGTAATAAGCAATTGTTGGAAAAGCAACAAATGGGAATCATATAAAAAGACTCTGTGCACCAAAGTCAGTTATCAGTCAACGTT	-29
	-->-41	
	-----	
-28	CGTTCTCGACCAGACAGAAATCAGCCAACATGTTCAAGTTTGTGAGTGGCTCAGGACATTTATGAACTCGCCATCTAATTGGTATCAT	61
	MetPheLysPhe	(4)
	-----	
62	TTCTCTATCCAGGTGATGATTCGCGGTTGTGGGAGTGGCTACCGCCCTAGCCCCAGTTCCCGCTCCGATGATGTACACGCTGATGT	151
	ValMetIleLeuAlaValValGlyValAlaThrAlaLeuAlaProValSerArgSerAspAspValHisAlaAspVa	(30)
152	CCTTCCGATCGGACGACGTTTCGTGCCAGCGATTGCATCCAGCTGCACACCTCAAACGGAATCGAGCAGGCCGCCAGCGGTGATGC	241
	lLeuSerArgSerAspAspValArgAlaAspGlyPheAspSerSerLeuHisThrSerAsnGlyIleGluGlnAlaAlaSerGlyAspAl	(60)
242	CCATGGCAACATCCAGGCAACTTCGGCTGGATCTCACCAGGGGCGAGCACGTTGAGGTAAGTACGTCGCGAATGAAAACGGATACCA	331
	aHisGlyAsnIleHisGlyAsnPheGlyTrpIleSerProGluGlyGluHisValGluValLysTyrValAlaAsnGluAsnGlyTyrGl	(90)
332	GCCCTCGGGAGCCTGGATCCCCACTCTCCTCCAATCCCAGAGGCCATCGCCCGCCGTTGCTTGGCTGGAGTCTACCCCCAGCACC	421
	nProSerGlyAlaTrpIleProThrProProProIleProGluAlaIleAlaArgAlaValAlaTrpLeuGluSerHisProProAlaPr	(120)
422	CGAGCACCCCGTCACTACTAGGACTCGTACCAGGATCCCGACCCTACACGGACTGTTCTCCGAAAACAAATCGCCCAAGTTGTTTA	511
	oGluHisProArgHisHisEnd	(126)
512	GCTGTACTTCTTGACTTTCAAAAAAATACATGCACCTGCTTATAGCAGTAAAAATGTGTGTCTTCACTTGCACCTTTTAGGTAGTCC	601
602	TGTAATAATACGAGCTTTTATACCTCTACCTTCGCTGGGAATGCTTCTCTACCTTTTATATTCGATTCACTAAATCCATTTATCAAAA	691
692	ATGAGTATATGTGCCATAAAGAAAAGATGTGCTGAATTAA	732

*Lcp2* SEQUENCE. *Canton S* strain. Accession, J01081 (DROCTCL2). The sequence of *Lcp2* starts in the neighborhood of a *HindIII* site between *Lcp2* and *Lcp3*.

*Lcp2*

Open reading frame, 126 amino acids; expected mRNA size, ca. 533 bases. There is one 62-base intron after the Phe-4 codon (*Lcp2* Sequence) (Snyder et al. 1982).

### Promoter

Approximately 800 bp upstream of the *Lcp2* transcription initiation site is sufficient for correct developmental regulation, but other sequences still farther upstream may also be necessary for full expression. A 270-bp segment does not support any detectable transcription in transgenic animals (Kimbrell et al. 1989). It should be noted that the distance between the divergently transcribed genes *Lcp2* and *Lcp3* is approximately 870 bp.

### *Lcp3*

Open reading frame, 112 amino acids; expected mRNA size, ca. 494 bases. There is one 56-base intron after the Ile-4 codon (*Lcp4* Sequence) (Snyder et al. 1982).

### *Lcp4*

Open reading frame, 122 amino acids; expected mRNA size, ca. 494 bases. There is one 57-base intron after the Ile-4 codon (*Lcp4* Sequence) (Snyder et al. 1982).

## ***Pupal Cuticle Proteins: Pcp, Edg78E, Edg84A and Edg91A***

### *Pcp*

(*Pcp* in the *ade3* gene intron 1)

**Chromosomal Location:**

2L, 27D1-3

**Map Position:**

2-20

Synonym: *Pcpgart*

### Product

Probably a pupal cuticle protein. The amino acid sequence shows clear similarities to larval and pupal cuticular proteins (Fig. 9.2), including the presence of a putative signal peptide (Silvert et al. 1984; Henikoff et al. 1986).

### Gene Organization and Expression

Open reading frame, 184 amino acids; expected mRNA length, 718 bases, in agreement with an RNA band of 0.9 kb. Primer extension, mRNA sequencing and the sequence of two cDNAs were used to define the 5' end. The 3' end was obtained from a cDNA sequence. There is an intron after the Leu-4 codon (*Pcp*

*Lcp3* and *Lcp4*

-650	GTGCTCATGATGACGTTTCAGATTGACCAAGTCTTTATCAATCATGCCTGTACAAAAACAAAAACCAATACACTTCACCTTGATAGCA	-561
-560	GTCCCATCTATTTTAAAAGGTTCCGAAGTGGTAATAAACAAATATACCGGAATAAACGATTTTCGCAAGTGATACTGCATATTAACGTGCTA	-471
-470	GTTGCCATGACATTTTGTGTATCTCAATATTTTGGATGTCAGCTGTTTAATTTAAAAAACCCGCAAGCTTTTGGCCAGAGTTGTCAA	-381
	HindIII.	
-380	CGTGCCACACACCAAAATGAAACACCGAAAACCTATGCTATGCTTAAGTTAGTTTCATATTGAAGTTGAATTTTAGAAAAATTAATATTGTA	-291
-290	CTGCTTAATAAATATTCTGGTTTCTGGTCCGGTTTGCCTTTCGTTAGACTAGGGCGAATATTTTCAGTTGAATAAATAACTAAGA	-201
-200	ATGCTCATCTCCTAATGAAAGTGGTTAAGCCATCTCAAGTCGACTAATTTGCATCCAGAGCGGTTTTTATTATATGCATCACATTGACTT	-111
	HMS Beagle insertion	----
	=n1	-->-44 Lcp3
-110	AATTATAATACGCACATTGCATCAGCTTTTGGATGATATATAAACACCGATTGAGCATAGATTGCATCAGTCTTAGAAGATTCTAGTC	-21
	-----	
-20	CGACAATCCACCAAAATCAAATGTTCAAGATCGTAAGTATGCCTTGAGGAGCATAGTGACTTCGCAGTCTAATCCTGGATTATCCTAGC	69
	MetPheLysIle	L (5)
70	TGCTTGTCTGTTCTCTCGCCCGCTGGTGGCCGCCAACGCTAATGTGGAGGTCAAGGAGCTGGTCAACGATGTCAGCCGATGGCTTTG	159
	euLeuValCysSerLeuAlaAlaLeuValAlaAlaAsnAlaAsnValGluValLysGluLeuValAsnAspValGlnProAspGlyPheV	(35)
160	TCAGCAAGTTGGTCTCGACGACGGATCGCTCCTCCGCCACCGAGACATCCACGGCAACATCGACGGAGTCTTCGAGTGGATCCTCC	249
	aSerLysLeuValLeuAspAspGlySerAlaSerSerAlaThrGlyAspIleHisGlyAsnIleAspGlyValPheGluTrpIleSerP	(65)
250	CCGAGGGTGTCCATGTGCGAGTGAGCTACAAGGCTGACGAGAACGGATACCAGCCAGAGTGACTGCTGCCACTCCTCCTCCGATCC	339
	roGluGlyValHisValArgValSerTyrLysAlaAspGluAsnGlyTyrGlnProGlnSerAspLeuLeuProThrProProProIleP	(95)
340	CAGCTGCCATCCTGAAGGCTATCGCTACATCGAGGCTAACCCAGCAAGAACTAAGTGAACCCGCCGACTAGGAACATGAAAGATTGGA	429
	roAlaAlaIleLeuLysAlaIleAlaTyrIleGluAlaAsnProSerLysAsnEnd	(112)
430	GACAGCTAGGTTGAGTTTGATAAATTTCTTACCAGTTGTTTTAAATTTAAGGAAAATGTTATCGAATTCGAAAATAAATTAACCTTGCA	519
	-----	
520	ATATAAACCAAGTGCATGTTTTACAAATCTGACAGTTCGATTTAAGAGAAGGCTCCCGGATTATATGGTATAAGAAGGTACAATTAGAA	609
610	GATTAAGTAAATCAAAAGACACTTTGGCCTTCATTAATTAACAATTGTGTTGTTATAGTATAGTACGAAATTAATTTAAATACAAAAATC	699
700	TTTAAAGCATCTAAAAATAATGTAACATTACAAAAACCTTACCTGGACAAGCCGATATCTCCTGCATTAATTTTCATATTTCCGAAAAAC	789
790	TGGGTATAACTAGTTATTATTTAAGTTAAGTTATAGGCAGCCACAAGTAATTAATGTTGCCAACCTGATGCATCCAGATAAGATC	879
880	GCAATGATGATAAACGACGAGGAAGCTTTTTATATCTATTATTTGTAGAGGATAAGGGTACACTTGAATTTGTAACGCATGTCGGTA	969
970	TTATGGGTATTAAAGGTATTATGAAGCGTTTTTCGAACCTAAAAAGTATGTATATTATCATCATATTACACTAGCCGAGTCAAAATTTTGT	1059
1060	TTTTATATTGGCTTTTTTGAATATAACTGAAATGGCATTATAAGCCTAGATGTAATAAATAAATTTCTCATCTTTTTTTAACTTTT	1149
1150	TTTAAATAGTCATACAGTAACAAAAATAACACAGACTTCCCTGAGGTTACACGGTTATAAGATCTTGTAGTGATTTTTGGAGAAATAT	1239
1240	CAATCAAAATGCTGTGCTTTTCGGATTTTGTATTATATTGATATTGTAACCTAAGTGTTTTAATAGTGATTGTATAAGTAAGAAATCGTAT	1329

1330	ACATTGTATATGTCAAACCTCCCGGGGAATGTTTCATATTGACTTAACGGAACTAGAGATAAAATATACACAATGTTTTTTTTTATTAA	1419
1420	ACGAAATTATTTACAATAATTTAATTGACTAGCAATAGTACGCTCTTCTTAGGCAACCCAATCTTATCGGTATCAATTTAAACTATTCTT	1509
1510	AATATCTATGTTATTTACAAAGGTTATATGAGTAAAGAGTTTTTGAGGAATAGAATGTTTATGCAGATTTTAATTTAGTAGGATTTATGTC	1599
1600	AAGTCCCGGTCAGTCAAGCTTGTAGGGTGGTGAACACAGAATGTTAGATTCCATAAACCCGTTCCAGTCATTTCGCAGATAGAAACCAA	1689
1690	ATGATGCTCCGAAAGGTATGCTGGATCTACAAGCGGTTCCGAAAAAGTTTTGTTTTCTAGTTATTTTTACCTCCTAATAATTAACCTT	1779
1780	CTACTATCAGCAGCTTAGACATTATTCAATCAAGTTATTTTTATATGATTTGTCTGGAGTAATCAAAGTTATCTGACTAAATATTCCGG	1869
1870	AAGATGTTAAATTTTCAATGAGAAGGTGGACTTACCCTTTCCGAGTAACCCGATCTTTTTAGAAATAATTACGGTAGCGATTTGCAT	1959
1960	AGACAAATAGAAATCAAAAAGAGTGCAGCAGACGATTTTTATCGCCACCAAGCATGTCACCTGAACCAGTCCGTAACCAACGAGACCT	2049
2050	ATGCTGGCCGAAATGTTAATTA AAAACGGGTTGCATCAGCTTTTGATCAGCTTTAAGATTTCTGTTGGGAGTGCGTATAAGCTATAAAAAG	2139
	-----	
	-->2163 Lcp4	
2140	CCGACGAGTGATCCCGAATTGGCATCAGTCTCAGGAGTCTTTAGTCTGACAATCTAACCAAGTCAAAATGTTCAAGATCGTAAGTATCT	2229
	MetPheLysIle	(4)
2230	GAAGTTTAAAGCCGGACAGTTCATGAGTAAATCCCGGAATATCCTAGCTGCTGTCTGCGCCCTTGTCGCCCTGGTGGCCGCAACGAGA	2319
	LeuLeuValCysAlaLeuValAlaLeuValAlaAlaAsnGluA	(19)
2320	ATCCCGAGGTCAAGGAACTGGTCAACGATGTCCARGCCGATGGCTTCGTAAGCAAGTTAGTCTTGACAACGGTTCGCCGTCTTCTGCTA	2409
	snProGluValLysGluLeuValAsnAspValGlnAlaAspGlyPheValSerLysLeuValLeuAspAsnGlySerAlaAlaSerAlaT	(49)
2410	CCGGAGATGTCCACGGAACATCGACGGAGTTTTCGAGTGGGTCTCCCGAGGGCGAACACGTCGGTGTGAGCTACAAGGCCGACGAGA	2499
	hrGlyAspValHisGlyAsnIleAspGlyValPheGluTrpValSerProGluGlyGluHisValArgValSerTyrLysAlaAspGluA	(79)
2500	ACGGATACCAGCCCAGAGCGACCTCTGCCACTCTCTCCAATCCAGAGGCCATCCTGAAGGCCATCGCCTACATCCAGGCCCATC	2589
	snGlyTyrGlnProGlnSerAspLeuLeuProThrProProProIleProGluAlaIleLeuLysAlaIleAlaTyrIleGlnAlaHisP	(109)
2590	CCAGCAAGGAATAAGCAATCGACACGACGACCCACATTGCAATCGGAGGTGCAACTCCAAGACCTTGCCTCTAACCTTAGAATT	2679
	roSerLysGluEnd	(112)
2680	TAAACAGCATGCAGACATTATAAATGATTATCGAGTTAGGAAATAAAATTCGATACTCTTTGGCAACAAATCTATTTAGATATGGTCTTAA	2769
	-----	
2770	TTTCCCTGACGGCAGCAGGAGTTACCTTGTATTGGCTGATTTATTTGGCCGAGGGAACAAACGCTGCTTCAGATTATCACGAACTGTC	2859
2860	TGGATGTTACGTGATTGATCTTAGCCAATAGTAACCTGTTAATTAGCGATACATAAAGTGAAGACCATCAAACCAGATTTAGGTATAAA	2949
2950	TTCGGTCTGTTTATTACAGTTTTAAATGCAATAAAATTTTCATTAACAAAAGTCATGGCTGAGCAAAATATAACCGGATTGGAATTGC	3039
3040	TTGCGTTACTCTTCATCTTCATATTGTTAAAAGAACAGTAAAGAACGGTATAGTAAAATTTTGAATACTTATTATTATTACTCGGT	3129
3130	TAAATGTTGGTGGTACCCGATAGAAATTTGCAAGAAAAAGTTAAAATAACCATTTTTTTGAAAGAAATTCGGTGCCAAAATGAGACG	3219
3220	GTTTGAGAGCGTTACTGGAAAAAAAACCCGATGCAACATGGCTTTAACGATCGACTACCTGTTATAACAATCCCTTCACATTGTCAA	3309
3310	TCATCTAGTATAAACTTCAAATCTAGGAGTAGAGAGTTGGTAAAAACATCCTTGAAGATGTTAATGGACTAGCTGTTATCATGATTATAT	3399

*Lcp3-4* SEQUENCES. Canton S strain. Accession, J01081 (DROCTCL2). The sequence of *Lcp3-4* (the opposite strand of the two previous sequences) starts near the same *HindIII* site. Indicated is a mutation of *Lcp3* caused by an insertion in its TATA box.



*Pcp*

-244	AAAATCATTTTATTTATGACTGACTAAGGCGACCAGCAGCGATGAGATGTTTGTAGATGGAGACGATCATGACGATGACGAGCGGAGATG	-155
-154	GAGATGGAGACGGCAACGGCAACGGCAACGGCAACTCGGAACGGGTTTCCGAGGCGATGTATAGCCAAAAATCCGCTGGTGAGCGGATG	-65
	-->-32	
-64	GATATAAAAACGAAAGCGTCCGAGAAGCAGGCAAGCAGTTTAGAACCAAACCTCGAACGCGACACCATGTATTTGCTTGTAAAGCATCAGCT	25
	----- MetTyrLeuLeu	(4)
26	GGGAATTTCCCGAAAATGGATTATAATCGCCGACTCTCGTCTCGAATCCCGCCCACAGGTGAACCTTCATCGTTGCGCTGGCCGTGCTGCA	115
	ValAsnPheIleValAlaLeuAlaValLeuG	(15)
116	GGTGCAAGCCGGCTCATCTACATTCGGACTCGGATCGCAACACACGACCCTGCAGAACGATCTGCAGGTGGAGCGGGATGGCAAGTA	205
	nValGlnAlaGlySerSerTyrIleProAspSerAspArgAsnThrArgThrLeuGlnAsnAspLeuGlnValGluArgAspGlyLysTy	(45)
206	TCGGTATGCCTACGAGACCTCCAATGGCATTTCGCATCGCAGGAGGGATTGGGTGGCGTGGCCGTACAGGGCGGCAGTAGTTACACATC	295
	rArgTyrAlaTyrGluThrSerAsnGlyIleSerAlaSerGlnGluGlyLeuGlyGlyValAlaValGlnGlyGlySerSerTyrThrSe	(75)
296	ACCCGAGGGCGAAGTAATTAGTGTGAACTATGTGGCCGATGAGTTTGGCTATCATCCCGTGGCGCACATATACCCAGGTGCCGGACTA	385
	rProGluGlyGluValIleSerValAsnTyrValAlaAspGluPheGlyTyrHisIleProGlnValProAspTy	(105)
386	CATACTGCGCTCCCTGGAGTACATTAGGACGCATCCCTACCAGATCAAGGACTACTACACCGGGGAGCTGAAGACCGTGGAGCAGATGC	475
	rIleLeuArgSerLeuGluTyrIleArgThrHisProTyrGlnIleLysAspTyrTyrThrGlyGluLeuLysThrValGluHisAspAl	(135)
476	AGCCGCTTCAATGTGTACACACGCAACATTCAGGATCATACGATCCCCCAATCCCGACCGAGCACCACGCCAAGACCATATACCTCAC	565
	aAlaAlaPheAsnValTyrThrArgAsnIleGlnAspHisThrIleProGlnSerArgProSerThrThrProLysThrIleTyrLeuTh	(165)
566	CCATCCGCCACGACCACGTCGCGACTCTGCGCCAGAGACGAGCTCTCCGACGCACTGATGATCGATGGACGTGACTCTATGGCGGGG	655
	rHisProProThrThrThrSerArgProLeuArgGlnArgArgAlaLeuProThrHisEndEnd	(184)
656	CAAGGGGCTGGTCTCTTCGCGGCCAGCGGGCGAATCTGTGAATTTTGATCTAAACAATTAATTAAGCCACGAACAATAAATAGAAGTGC	745
	-----	
746	TAAGCAACATAAGCTAAAGTGAATCGATCTGTGAGTGTCTGCTGGGGATCATGGATCACATCATGGAGCGACATAAACAATTTTGG	835
	(A) <sub>n</sub>	
836	GTATTCGATTCTGTTTATGGC	856

*Pcp* SEQUENCE. Accession, J02527 (DROGART).

Sequence). The *Pcp* gene is completely within the long first intron of *ade3* (*Gart*), a gene that encodes two polypeptide chains involved in purine biosynthesis. *Pcp* and *ade3* are transcribed from opposite strands (Henikoff et al. 1986).

*Developmental Pattern*

*Pcp* RNA is present in prepupae and possibly in larvae and pupae as well. *In situ* hybridization in 11 h prepupae, shows *Pcp* RNA to be present in the larval

epidermal cells that secrete abdominal cuticle, and to a lesser extent in the imaginal cells that secrete cephalic and thoracic cuticle (see *Edg78E*) (Henikoff et al. 1986).

### *Edg* (*Ecdysone dependent genes*)

These genes were identified because their transcripts accumulate in imaginal discs in response to a pulse of the steroid 20-HE (Fetchel et al. 1988).

### *Edg78E*

**Chromosomal Location:**  
3L, 78E

**Map Position:**  
3-[47]

### **Product**

Pupal cuticle protein (Fetchel et al. 1988, 1989; Apple and Fristrom 1991).

### *Structure and Function*

Sequence features indicate a signal peptide at the N-terminus. Other sequence features characterize *Edg78E* as a member of the cutin family of cuticle proteins (Fig. 9.2) (Apple and Fristrom 1991). It is immunoprecipitated by antibodies against low molecular weight pupal cuticle proteins (L-PCP) (Fetchel et al. 1988).

### *Tissue Distribution*

The pupal procuticle is produced in the prepupal stage. It is subdivided into the exocuticle, secreted between 8 and 12 h after puparium formation, and the endocuticle, secreted between 12 and 20 h. The main protein components of the exocuticle are of low molecular weight (L-PCP;  $M_r$ , 8–25 kD). Six L-PCPs have been identified by gel electrophoresis, but it is not known which one of them corresponds to EDG78. Because the endocuticle is characterized by high molecular weight proteins (H-PCP;  $M_r$ , 40–82 kD), it is inferred that EDG78 is localized in the exocuticle (Fetchel et al. 1988, 1989 and references therein).

Edg78E

-1000 CTACCTGGGCTGGGAAAAATATACCATTTTATGTACGTTTATTTCTGGGTCGTTTGGCGATTCTTGAATCGAAGTCTACACATATGTA -911

-910 GAGAGATAAGTGTGAACACATTTAATTACTAGCTTACTTCGGATTTTGCACACTTCCTTGTACCAGAAACGATCTCAGCAATTAACAG -821

-820 CAGTTTCAATGGTTGATGGTGTGGCCAGCTAATTTCGAAACAAAAAATATCTTCGTTTCAACCTATCGCTGCGTCCATTGCAACCA -731

-730 ATCGATTGCCGAAGATCAAAGTGAACAATTAATTAAGTCATAAATGTAGGGTATCAGAAGATCACACGTAACATCGCACTGCATGGCT -641

-640 GGATCATCTTCGGCGCGCTCCGGGTGCATGCTGATGCTGCCCGATGACCTTGTCCATGTTTCAACAGCTTTCCAGGGGCACAAGGTAT -551

-550 ACTCGACCATACTAGACCATCGCACCTGCCACTCCATTGGGAAGCCTCGAGCCAGGGCGCAACTCCAATTGAAAGTTGTAACAAGAAA -461

-460 TCTTCAGCTCGTGTGGGAATTTCCAACGCTGTTTTGAATGGCCGAAAGCGTCACATTAACAGCAATATTTATTTCGATTGTAATTCAA -371

-370 CAGTTAATGGTATCTCGTGCAGAACCGAAACCGAAATCGAACTGAAACCGAATTAAGCATACAATATAAATGTTGGCAAAAT -281

-280 GACTCATGATTTTTAACTATAGGCCAGCCGAGCACTGTTGTTATTGTTGCCAATATTTGGTGGTATGATAAGAAGACATTTTGGCAATTA -191

-190 TACTCACCTCTCGGGTCTTGTGATTCCACGAGAAAAAAGCTGTTTAGCTGCTAAACTAAAGAGACTAAAGACAAGGGTCTCATCCATAT -101

---

-->>>-75/-72

-100 AAAAGACGCCTTGAGCTGATCAAATTAACAGTTGCACTGCAAGCACCATCATCACAGCATCACCGCTTTAAGAGAAGAAAAATCCCAA -11

---

-10 TTCCCATCATCATGTACAAATATGTAAGTTCGGTTGGACTTGGCACGCCATACCCAGAGTACCAATACTGATCATTGTACTTTGATC 79  
MetTyrLysTyr (4)

80 CCAAAAGCTGTTCTGCTTCTCATCGGCTGCGCCTGCGCCGACAACATCAACAAGGATGCCAGATCCGCAGCTTCCAGAACGACGC 169  
LeuPheCysLeuAlaLeuIleGlyCysAlaCysAlaAspAsnIleAsnLysAspAlaGlnIleArgSerPheGlnAsnAspAl (32)

170 TACCGATGCTGAGGGCAACTACCAGTACGCCTACGAGACCAGCAATGGCATCCAGATCCAAGAGGGGGCAACGCCAACGGAGCAGCTGG 259  
aThrAspAlaGluGlyAsnTyrGlnTyrAlaTyrGluThrSerAsnGlyIleGlnIleGlnGluAlaGlyAsnAlaAsnGlyAlaArgGly (62)

260 TGCCGTGGCTTACGTGTGCCCCGAGGGCGAGCACATCTCGTGACATACACC GCCAGGAGGGCTACCATCCAGTGGGTGACCACCT 349  
yAlaValAlaTyrValSerProGluGlyGluHisIleSerLeuThrTyrThrAlaAspGluGluGlyTyrHisProValGlyAspHisLe (92)

350 GCCCACC CGCCCCAGTTCCGGCTTACGTTCTCCGTCCCTGGAATATATCCGACCCATCCCCGGCGCCGCCAGAGGAGCAGCA 439  
uProThrProProProValProAlaTyrValLeuArgAlaLeuGluTyrIleArgThrHisProProAlaProAlaGlnLysGluGlnGln (122)

440 GTAATCTGGAGTAGCACCAGCACTCCAAAGCAGCAACCCACACTAAACTGCGGCCAGTCATTGTTATTTAGGTAGTTATCGTTAATAA 529  
nEnd

530 AGGATTTGCATACAGATCATTTTCGTTTTTAGTAATGTAGTAAAGATGGAAAATAAATGTTTCATGTATATGTATTTCATATGTAATGAA 619

620 CATATGTATAGTTCTTCGAAAAATATAGAAGCGTACACTATCTTCAATAGAAACAAATTTACAGGCGGATGGAGTTACATTTTGAACAT 709

710 TTCTTTATCTTAACATGCTCTTTTTCTTCAAATGAACAATTTGAAGAATGTATATGTTAGTTAATGATTTCCGCGACCCAGTAATTGT 799

800 ATAAAACCATTTATCTATGTAATAGATTTTGATTTATGTCATTTATTTTCCACTTTCATTTATACTCAACGCATTATGATTTCCGAACT 889

890 ACAATAGTTAAATTTTTGAAAACCAATCCAGCGGTGATGCACAGATGAGATAAAATAAAAGAAACAAAATCTCGTAGATGAGATAAATTA 979

----- (A)<sub>n</sub>

## Gene Organization and Expression

Open reading frame, 122 amino acids; predicted mRNA length, 962–966 bases, somewhat larger than the 0.6 kb band detected by northern analysis. Primer extension was used to define the 5' ends (there seem to be four clustered transcription initiation sites). The 3' end was obtained from a cDNA sequence that included a poly-A tail. There is an intron after the Tyr-4 codon (*Edg78E* Sequence) (Fetchel et al. 1988; Apple and Fristrom 1991).

### *Developmental Pattern*

As would be expected for a secreted protein, the *Edg78E* mRNA is preferentially associated with the membrane-bound polysome fraction. Low levels of this RNA are detected only in prepupal stages (Fetchel et al. 1988). By *in situ* hybridization, *Edg78E* RNA can be detected both in the larval epidermal cells that secrete abdominal cuticle and in the imaginal cells that secrete cephalic and thoracic cuticle. The peak of accumulation is in 10 h prepupae (Fetchel et al. 1989).

In imaginal discs in culture, *Edg78E* transcription is stimulated by a pulse of 20-HE, 6 h in 1 µg/ml hormone and 8.5 h without hormone. Transcription, however, is inhibited if the hormone treatment is continuous or if hormone is re-added to the medium after an original pulse that stimulates transcription. This hormonal regimen mimics the endocrine status during the larva-to-pupa molt. Thus, a 20-HE peak would stimulate *Edg78E* expression, and its product would presumably contribute to the exocuticle being produced at that time. A second rise in hormone titer, which signals the transition from exo- to endocuticle production, would repress *Edg78E* and induce expression of other genes whose products are characteristic of the endocuticle (Fetchel et al. 1988; Apple and Fristrom 1991).

## *Edg84A*

**Chromosomal Location:**  
3R, 84A

**Map Position:**  
3-[47]

### **Product**

Probably a cuticular protein.

### *Structure and Function*

It has sequence features that indicate a signal peptide and sequence similarities to cuticular proteins of *Hyalophora cecropia* and *Locusta migratoria* but not to cutins (Apple and Fristrom 1991).

### **Gene Organization and Expression**

Open reading frame, 188 amino acids; in northern analysis, a 0.9 kb band is detected. Primer extension was used to define the 5' ends (there seem to be three clustered transcription initiation sites). The 3' end was not determined. There is an intron after the Lys-4 codon (*Edg84A* Sequence) (Fetchel et al. 1988; Apple and Fristrom 1991). *Edg84A* is part of a cluster of small genes with related sequences located within the Antennapedia Complex, between *labial* and *proboscipedia* (Pultz et al. 1988; Fetchel et al. 1988).

### *Developmental Pattern*

As would be expected for a secreted protein, the *Edg84A* mRNA is preferentially associated with the membrane-bound polysome fraction. This RNA is detected only in prepupal stages (Fetchel et al. 1988). By *in situ* hybridization, *Edg84A* RNA can be detected only in the imaginal cells that secrete cephalic and thoracic cuticle but not in the larval epidermal cells that secrete abdominal cuticle. The peak of accumulation is in 10 h prepupae (Fetchel et al. 1989).

As for *Edg78E*, *Edg84A* transcription is stimulated by a pulse of 20-HE in imaginal discs in culture (Fetchel et al. 1988; Apple and Fristrom 1991).

## *Edg91A*

**Chromosomal Location:**  
3R, 91A

**Map Position:**  
3-[64]

### **Product**

Probably a cuticular protein.

### *Structure and Function*

It has sequence features that indicate a signal peptide and sequence similarities to insect egg-shell and egg-casing structural proteins. It also has some similarities to vertebrate cytokeratins. EDG91 is a hydrophobic protein with very high (32%) Gly content (Apple and Fristrom 1991).

*Edg84A*

```

-818 GAATTCCTTTTTATTAATTTTAAAGTTACATTTTTCTAAATAACACATATTTTACGATGGAAATATAAAACATTTTTGTAACCATT  -729
-728 TGTTACCTGTATATATGTATTGTTGATTTATTTATAAGGAAAGCGAAATCAGGAAATTTAGCACCCCTGTTGGTCAGCAAGAAAAA  -639
-638 TATTCTGCATACTTTTGGGCTGACTATGAATATTCAAAAAATGCTCCCAATGGTAATGGTTTTTTATTTCCGGCTAATACTACA  -549
-548 AATGAGCCATAGCAGTACATTATAAATTCGAAGTATGTCTTGGCATAGGGCTTATATTTGGGCGACATATTTGAGCAGTCTGCAACA  -459
-458 ATCGGCAAAATTTTATAAAAAATGTTCCCTGTCTTAGTTACAATATCATCAATTTGAAATGAGCAAGGCGATTATTATTATTTGCAAG  -369
-368 TTGTCCTTAAATAAGGAAGTAAATAAAAAACATACAAATATCAAAATTTGGTGAGGAATGACTCCGCGAAATATGGACGGAGCCCAT  -279
-278 ATCCCGGACAGCAAGTAAAAACGGTCTGAAAACCTGCCGATTGCCGATAAACTTGTGGGGCATCTCAACGCCAATTAAGCGGTCTAC  -189
-188 AAAGTGACTGGGCTGGAGTCCCGCGATGACCTGTTAAGATCCAGATGCAGAAACAGGCCACTGTGGCACTGGGTCGACGGCAAGGAA  -99

                                     ---> -->-60/-59, -55 .
-98 GCCGCCTATAAAAGCCGATGTGAGTACCGTAGTGAACCTGTGTAATACTCACTACCAGGAGCAAACTAATTCATCAACCTAAAAAT  -9
      -----
-8  TCGATCAGCATGTTGGTTAAGGTATATCATGTGTTATTTACAAGTTGGCTTGCCTTTATCCTAGTCCTTTAACCCAGTACAGACTGCGCT  81
      MetLeuValLys                                     ThrAlaLe  (7)

82  ATTTGTGACCCTCATCGGCTTGGCTCAAGCTGGTCCACTGCCCGCGAAATCATCTGGAAGTGAGGACACCTATGATTCTCATCCGAGTA  171
      uPheValThrLeuIleGlyLeuAlaGlnAlaGlyProLeuProAlaLysSerSerGlySerGluAspThrTyrAspSerHisProGlnTy  (37)

172 CTCATTTAACTATGATGTTCCAGGATCCAGAGACAGGAGATGTTAAGTCCAGTCCGAGTCTCGGGATGGCGATGTAGTCCACGGTCAGTA  261
      rSerPheAsnTyrAspValGlnAspProGluThrGlyAspValLysSerGlnSerGluSerArgAspGlyAspValValHisGlyGlnTy  (67)

262 CAGCGTGAATGATGCCGATGGTTACAGACGAACCGTGGACTACACGGCCGATGATGTCCGTGGATTCAACGCCGTGGTGCCTGTGAACC  351
      rSerValAsnAspAlaAspGlyTyrArgArgThrValAspTyrThrAlaAspAspValArgGlyPheAsnAlaValValArgArgGluPr  (97)

352 ACTTTCCAGTGCCGCGGTGGTTGTGAAGCCACAGGCTACAGCAGTCTGTTCCAAAAGTTCAGTTAAAGCCTCTGAAGAAGTTGCCAGCCCT  441
      oLeuSerSerAlaAlaValValValLysProGlnAlaThrAlaValValProLysValGlnLeuLysProLeuLysLysLeuProAlaLe  (127)

442 GAAGCCGCTTTCTCAGGCATCGGCTGTGGTGACCCGATCCTTTGCACCGGTGGTCCACCATGCCCCAGTGACCCATGTCGTGCACCACGC  531
      uLysProLeuSerGlnAlaSerAlaValValHisArgSerPheAlaProValValHisHisAlaProValThrHisValValHisHisAl  (157)

532 AGTCCCGCGCATCTTTCTGCTCTCACCAGTCCCGTGTGAAGACTACCGTGCACCACGCCCATCATCCCATGCCATTTTCATATGT  621
      aAlaProAlaHisSerPheValSerHisHisValProValLeuLysThrThrValHisHisAlaHisHisProHisAlaIleSerTyrVa  (187)

622 GTTCTAGA  629
      lPheEnd  (188)
    
```

*Edg84A* SEQUENCE. Strain, *Canton S*. Accession, M71249 (DROEDG84A).

**Gene Organization and Expression**

Open reading frame, 159 amino acids; mRNA length, 581–591 bases. Primer extension was used to define the 5' ends (there seem to be three clustered

*Edg91A*

-1116	CTGCAGGTCGATTAAGGCTCGATTGACCAAAATGAAAAATCCCAAATAAGAAAGACTTTACTCGTTGAGTTTTTGTAAAGAACTAATTTT	-1027
-1026	ATTTGGAATATCTTCGGTTTAAATAGGTGACATGAGAATCGCATCTTAAAGTAAATGGCCACGAGAGGCTAAGTAAATAGTCCCCGC	-937
-936	CTTATCGAGGTCACGCTCGGGCACATCTGCCTATCTTGAGCGGCAGGACCTTATCTGTGGTCTCCCACTAAGGGACTATTTTAGGAG	-847
-846	GCGGGGAACGATCTCAAGTGACTGACTCATGTAGTGTGCACTTAAATTACATTTTTGAGCAATGCACCCATGTCGCCTTGGATAACAAAA	-757
-756	TCCTAAATATAATTTATCGCTCTCGATTCAATTTACATAAGATGAACGGAGCCAAAATTGTAAGTCTTTAAATATATTCGTGTTTCATG	-667
-666	TGTGAACAACAAGCATTGGGTTTAAACCCTGCTATTGTAACCCATTAAAAGAAATATTTTATCAAATTAATATTTATAAAATATTTATA	-577
-576	TAGCCTTTAAATACTCCTTTTCATTCTGATTTGAAGTGGCTAAATTAATAGGTAATTTATTTTATCAACTCATACTTTTAAGAAATTAG	-487
-486	TTTCTTTACATTGAAATTTTTTAAAGATAGCTTAGTTTAAATTTTATATTTTTTAAATGCAGAGTCATCTATCGGTTACAGTGGAAATA	-397
-396	TTATATTCGTATTTCAACATTTTTCTGGTTGGTCTTGAATACCAGGATGTTGAGTAGTACGCATCGCTCAGTGATATTTTTATGGTTCA	-307
-306	CGATCTTGATGACCGCAACTAAGACAACCTCAAAAATGATAATTAGTTGGCCCTGTGACTTCAAGAAATTAACGCGTTCTGGGGCCAAG	-217
-216	TGAAGCACTGGTAGGCAAAGTGCTCTTGGGGGATCCAAAGTACGTCACAACTGGTTTCGCTTTCGCCGTGTTGTGTCTGCAATTTG	-127
-126	CGTAGAATCACTTGGCAATGCGTAGCGCTACTTGAGCTTCTTGGCCAGATTGAAGCGCGGTATAAAAAGCGGTGGGCACCTTCAAACTT	-37
	--- -->>-33/-34-->-23	
-36	GCAATTTAGTTTCATCCAAGAAGCGCTCGTTATCGCAATGGCTCTGGTTCGCGTGAGTGTGTAAAGTCCGGCTGCTATTTCCGCTCCGAT	53
	MetAlaLeuValArgValSerCys	(8)
54	TGGGATGCACTGAATCGATTTGGTTACCTTGCAAGTGGTGGCCCTTTTGTCTGATTCGCCGGTCAAGGTCAAGGTCAGGCGGCCTGGTGAAGACCGA	143
	MetLeuAlaLeuLeuLeuIleAlaGlyGlnGlyGlnAlaAlaProValLysThrG	(27)
144	AGGTCGCACCTTGGGCTTCTGGGCGGTGGATTTGGTGGCAGTGTAGGACTTAGTGCCGGCATCGAGTGGTGGTGGCCTGTATAGCGG	233
	uGlyArgThrLeuGlyLeuLeuGlyGlyGlyPheGlyGlySerValGlyLeuSerAlaGlyIleGlyValGlyGlyGlyLeuTyrSerG	(57)
234	TTTCGGAGGCGGTGGCTATCCTGGTGGCTATGCGAGTGGATACCCAGGTGGATATGGTGGTGGCTACTCAGGCTATAACGGCTACGGAGG	323
	yPheGlyGlyGlyGlyTyrProGlyGlyTyrAlaSerGlyTyrProGlyGlyTyrGlyGlyGlyTyrSerGlyTyrAsnGlyTyrGlyG	(87)
324	CAGTGGATTCGGAGGTGGCTACTATCCAGGAGGAGTTACTCCGGCTTTGGACACAGGCCGATTACCACGGAGGATACTATCCGGGCGG	413
	ySerGlyPheGlyGlyGlyTyrTyrProGlyGlyGlyTyrSerGlyPheGlyHisArgProHisTyrHisGlyGlyTyrTyrProGlyG	(117)
414	TGGATCGTACCACAATCAGGCGGATCTTATGGCGGCCACTATAGTCAGTCACAGTACTCGAATGGATATTACGGAGGTGGTGGCTATGG	503
	yGlySerTyrHisAsnGlnGlyGlySerTyrGlyGlyHisTyrSerGlnSerGlnTyrSerAsnGlyTyrTyrGlyGlyGlyGlyTyrG	(147)
504	AGGCGGTGGCTATGGAGGCAATGGCTTCTTTGGAAGTAAAGATGCCAAATCTGGCACCAGGATAGTTAAGTACTTGTGATTGACCCCTT	593
	yGlyGlyGlyTyrGlyGlyAsnGlyPhePheGlyLysEnd	(159)
594	TGTAGATTGTAATAAACAAGAAAAACATAAACCAGATTTAGTAAGCTCAATTCAAGGCCTTAAAAATCCGGTTTTCTGTGGAAATAT	683
	----- } (A) <sub>n</sub>	
684	TGTCCTTGGCGCTGCCTTTGTGGTTATTCTCTCACTGATTTTTATGAAGCAGACGCGACGTGCATAAATTTAATGCCAAAAGATCCAAGA	773
774	TTTATGCGCAAGTCTGACTAATCCATTGCCTCGAAATATCTGGGAATTC	823

*Edg91A* SEQUENCE. Strain, Canton S. Accession, M71250 (DROEDG91A).

transcription initiation sites). The 3' end was obtained from a cDNA sequence. There is an intron after Cys-8 (*Edg91A* Sequence) (Apple and Fristrom 1991).

### Developmental Pattern

As is true for *Edg78E*, *Edg91* is expressed during the time of pupal exocuticle synthesis (8–12 h after pupariation) in both larval and imaginal epidermal cells. Also as for *Edg78E*, a 20-HE pulse in imaginal discs *in vitro*, induces transcription of *Edg91A* (Apple and Fristrom 1991).

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# 10

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## The Cytochrome c Gene Cluster: *Cytc1, Cytc2*

**Chromosomal Location:**

2L, 36A10-11

**Map Position:**

2-[52]

Synonyms: DC4 and DC3

**Product**

Cytochromes c, small heme-binding proteins important in the mitochondrial electron-transport chain.

*Structure and Function*

Two Cys residues near the N-terminus bind the heme group. Another region near the N-terminus has a primary role in the import of cytochromes c into mitochondria *in vitro*; other portions of the molecule are also necessary for this transport (Sprinkle et al. 1990). These proteins are ubiquitous among eukaryotes and, judging from comparisons made among cytochromes c from 30 species, they are highly conserved. The CYTC1 sequence is very similar to the consensus sequence for other eukaryotic cytochromes c: at every position, the residue present in CYTC1 is found also in some other eukaryotic cytochrome c. CYTC2, on the other hand, is more divergent and has some unique characteristics: at 12 positions, the residues found in CYTC2 are not represented in any other eukaryotic cytochrome c (Fig. 10.1) (Limbach and Wu 1985). It is not known whether the two *Drosophila* proteins have specialized functions.

**Organization of the Cluster**

The two genes are arranged in tandem with approximately 2.5 kb between the 3' end of *Cytc2* and the 5' end of *Cytc1*. These are probably the only genes

```

          1                               50                               100 101       111
Dm c1  .MGVPAGDVE KGKCLFVQRC AQCHTVEAGG KHKVGNLHG LIGRKTGQAA GFAYTDANKA KGITWNEDTL FEYLENPKKY IPGTMIFAG LKKPNERGDL IAYLKSATK* .
Human  ....MGDVE KGKIFIMKC SQCHTVEKGG KHKTGPNLHG LFGRKTGQAP GYSYAANKN KGIWGEDTL MEYLENPKKY IPGTMIFVG IKKKEERADL IAYLKKATNE *
Dm c2  ...MGS6DAE NGKKIFVQKC AQCHTYEVGG KHKVGNLGG VVGRKCGTAA GYKYTDANIK KGVWTEGNL DEYLNPKKY IPGTMVFAG LKKAERADL IAFLLKSNK* .
Yeast  MTEFKAGSAK KGATLFKTRC LQCHTVEKGG PHKVGPNLHG IFGRHSGQAE GYSYTDANIK KNVLWDENNM SEYLTNPKKY IPGTMVAFGG LKKEKDRNDL ITYLKACE* .
CON    -----G--- -G---F---C -QCHT-E-GG -HK-GPNL-G --GR--G-A- G--YT-AN-- K---W-E--- -EYL--PKKY IPGTM-F-G -KK---R-DL I--LK----- -

```

FIG. 10.1. Comparison of the human (Accession, M22877), yeast (Accession, V01298) and *Drosophila* (Dm) sequences. The CON(sensus) line displays all positions for which there is agreement among the four sequences. There are 77% and 67% overall identities between the human protein and CYTC1 and CYTC2, respectively. Sequences aligned with the GCG *Pileup* program

*Cytcl*

-766	TCTGATGACGTTGCGACGCCCTCCACGCGCGTATTAGTGAGAGCAAAGTATGTGGGTTAAAAAGGGGGTGGCCGCAAAATGGAATGCAGA	-677
-676	CTACGTTAGATAATAATTTTCGGGCTTATCAGAAACAACAGCCGACTAATGCACCTAGCATGAGCAATTTAATAATTCGGTTCCGCGAG	-587
-586	GAGCTTATCAATTGTTTACATAACGGGGCAAGGGGACAAATATTAATTCACGGTCCATAACTACCTACATTAACCCATTATTTCCAAGC	-497
-496	ATGGAAATTTTGTAGATATAAAGACGTTATTATTTTAAATACCTTAAAAATATAATATTATATAAGTAACTGGGAAATCAACTGGT	-407
-406	TAATAAATTTTAAATTTTCGGGTTTATTTATTCAATAATCTTTTGATAATGTATGGCTGAAAGTGAAGCTTTTATCAGTATCTACACAATG	-317
-316	GTTCAATTGTGGCTAATAATAAATGGTATCAAATATCGTATAACTATTTTTGTCAGTGAACCAGAAATTCGGACTAAGTACATAAGCAA	-227
-226	TGATATAAAATATATATTGTAATCAATTTATCAGAATAGAACAATAATTGGTAGCAGTTATGAACCTTGAACCTTTAGTAGCCAGTT	-137
		-->-677.
-136	TTTTAAGTTTTTCAAACCTAAGATGTAAGATAACAGATATATTGGTTACCCCTGTTTTATGAACCCTCATTAATAACAACAACTTCTTT	-47
	-----	
-46	TTACAGTCGAGTCGGTAAACACATTAATTAACCACATAATCCATAATGGGCGTTCCTGCTGGTGATGTTGAGAAGGGAAAGAAGCTGT	43
	MetGlyValProAlaGlyAspValGluLysGlyLysLysLeuP	(15)
44	TCGTGCAGCGCTGCGCCAGTGCCACACCGTTGAGGCTGGTGCAAGCACAAGGTTGGACCAATCTGCATGGTCTGATCGGTCGCAAGA	133
	heValGlnArgCysAlaGlnCysHisThrValGluAlaGlyGlyLysHisLysValGlyProAsnLeuHisGlyLeuIleGlyArgLysT	(45)
	***            ***	
134	CCGGACAGCGCGCGGATTTCGCTACACGGACGCCAACAGGCCAAGGGCATCACCTGGAACGAGGACCCCTGTTGAGTACCTGGAGA	223
	hrGlyGlnAlaAlaGlyPheAlaTyrThrAspAlaAsnLysAlaLysGlyIleThrTrpAsnGluAspThrLeuPheGluTyrLeuGluA	(75)
224	ACCCCAAGAAGTACATCCCCGGCACCAAGATGATCTTCGCCGGTCTGAAGAAGCCCAACGAGCGCGGCGATCTGATCGCCTACCTGAAGT	313
	snProLysLysTyrIleProGlyThrLysMetIlePheAlaGlyLeuLysLysProAsnGluArgGlyAspLeuIleAlaTyrLeuLysS	(105)
314	CGGCGACCAAGTAAATGGTGTGCCATCAACTTACCCACAACAAGTGCAGGATGTCAAACCTGTATTATTGTGTTCAAGTCCAGTCCGGCA	403
	erAlaThrLysEnd	(108)
404	CGCAATGCAGCAGCAGCAACAACCTACAACCTACAATCAACATAGTACAGACCTAAAGAACTACAATTATGTTAATTATAAAGTTTAAAT	493
494	AGGACAATTTATTATTTAATTTAAATAAAAAGTGAATATTTAATCAAACCCGATGAGAATTGTGACATCCACAAAAAAGTTAATAAT	583
	-----	
584	AAAAAAGAAGTAAAAATGATATAAAATCTGTTTTATGCGAGGACCTGGTTTTTTGTAGCTCGCAGGTCAAAAAGAAATAAAAAAGCTTC	673
674	TTCAGATTTTGTACTCGGGCAACTCAAATAAAAATAAGAGATACCAATCATATTTATAAAAACAAATGTCTTGCAATTTCTATCAATAG	763
764	GTATCTGTTAGTCGTCAAACTCGACTGCG	792

*Cytcl* SEQUENCE. Strain, *Canton S.* Accession, X01760 (DROCYCDC4).

*Cytc2*

-916 GTAATATAAATATATAAATAATATCATCTCTGAAAAATATCAAATGCACTCTTGTAAATTTAAACAATTTAATTTTAAAGATAATTGG -827

-826 TTGAGATAAACATAGTTAATATTTTCAATTGATCCTTTAAATTTTAAATGTCAGGTGAATATCATCCCTGTGTGACCGTGTATGCGGCA -737

-736 TGGTTCCATGTCTCTTCCCGTTATTCATTTCCCTCTGCTTTGTTTTTTTTTTTTTTTTGGTTTTGTTATGCGGTGGCATCTGTTTTG -647

-646 CCACAGGAAAAATGTTAAGAGAGGGGAAGGCAGGGGGCGAAAAACGGAGAGTGCCTAAATGCGTTTTAATTGGAAGGAAATGTTCAAAGC -557

-556 ACATGTGCATCTGCTAGTCAACGAATGGTTGGAAAGGGGTGAAAAAGGGTTGCAAGCCGAATGTGTCTGCTAATTGAATTACTTTC -467

-466 GGTGCTTTTCCCATAGAAAGTCCGCCAAGTTCTCGAGCTGCTTGTGTCTTTTCATTTAATACCCATTTTGATTTAATTTTCGTTTTT -377

-376 CCTATTTTCTGACCAATTTGTTTTGCTTTCGTGCATTAGCAGCTGTCTGTCTATCGCTGTGACCCAAGAGAGTGACCAAGAGAA -287

-286 ACGCTCTCTCTCTCTCAGTTGTCCAGGACTGCACATTTTCAAACGGTTTTTTAGGACACTGAAACAATGAATCTGTTTTTCTTT -197

-196 TCTATCAAATTTTAGTCTACACTTTTCTTTTTCTTTTTTTTTTTTTTTCGGAATCAACCAATTTCTATTGATCCAATAAACA -107

-----  
 -->-43?

-106 AAAACAATAACAAAAATTAATAATATAGAAATAAAAGTCGATAAAAAGTTGAATCTAAATCATAAATATCATTTTCCCTATTTG -17

-----

-16 TCTTTCAGGCTTCCAAGATGGGTTCTGGTGATGCGAGAACGGCAAGAAGATATTTGTGCGAAGTGCGCCAGTCCACACCTACGAAG 73  
 MetGlySerGlyAspAlaGluAsnGlyLysLysIlePheValGlnLysCysAlaGlnCysHisThrTyrGluV (25)  
 \*\*\* \*\*

74 TGGGGGGCAACACAAGGTGGGCCAAATCTTGGCGGGTCTGGGTCGCAAGTGTGGCACAGCAGCGGATACAAGTATACCGATGCCA 163  
 a1GlyGlyLysHisLysValGlyProAsnLeuGlyGlyValValGlyArgLysCysGlyThrAlaAlaGlyTyrLysThrAspAlaA (55)

164 ATATAAAGAAGGGCGTTACCTGGACAGAGGGGAATTTGGACGAGTACCTCAAGGACCCGAAGAAATACATTTCCCGGAACAAAGATGGTGT 253  
 snIleLysLysGlyValThrTrpThrGluGlyAsnLeuAspGluTyrLeuLysAspProLysLysTyrIleProGlyThrLysMetValP (85)

254 TCGCAGGCTTAAAAAGGCTGAGGAGCGGGCCGATTGATTGCCTTCTCAAGTCAAACAAGTAGAATCGCTGCGAAACAACAAGATCG 343  
 heAlaGlyLeuLysLysAlaGluGluArgAlaAspLeuIleAlaPheLeuLysSerAsnLysEnd (105)

344 GCCACCATGCTATCCAGAAAACCTGCGCTTAAAGACTACAACATATTTCAAAGATGACGTATTTCACTTGGATTTCGAAACTTTGATTGG 433

434 GAATGGTCGAGCTCAAATACATTTCAAAAAGTTTACTTTTCACTTTAGCCAATTAAGTTGATAAACCACCAACCTCTTCTTAATCAA 523

524 GTTGTGTGCGACGCGGGTGGAGGAAAGTGTGTACCAATCAGCTTTGGTCACAGTTGGTTTTATGGTCTACTAGCAAAATGTAATAAAT 613  
 -----

614 TGGAGAAGCTTGTTAAATAATGCAAATTTCCAGAGGCTTTCCAATATAGTCCCTTAATAGGGGAAAAAATTACTTATACGCCGTGG 703

704 TGGATAAATACGGGTACAAAAGCTT 728

*Cytc2* SEQUENCE. Strain, Canton S. Accession, X01761 (DROCYCDC3).

responsible for cytochrome c production in *Drosophila* (based on Southern analysis) (Limbach and Wu 1985).

### *Cytc1*

#### **Gene Organization and Expression**

Open reading frame, 108 amino acids. The 5' and 3' ends have not been identified, a tentative site of transcription initiation was indicated based on sequence elements. A putative TATA box at -99 and a polyadenylation signal at 517 suggest a mRNA of approximately 600 bases, in reasonable agreement with an observed RNA of 0.9 kb bases. There are no introns in the coding region (*Cytc1* Sequence) (Limbach and Wu 1985).

#### *Developmental Pattern*

Expression is highest in first instar larvae and adults and lowest in third instar larvae. In adults, expression is higher in the muscle-rich thorax than in the head or abdomen. Expression of *Cytc1* is 25–150 times higher than that of *Cytc2* (Limbach and Wu 1985).

### *Cytc2*

#### **Gene Organization and Expression**

Open reading frame, 105 amino acids. The 5' and 3' ends have not been identified, a tentative site of transcription initiation was indicated based on sequence elements. A putative TATA box at -80 and a polyadenylation signal at 607 suggest that the mRNA is approximately 700 bases long, but the only transcript detected by northern analysis is 2.1 kb long; this indicates that the elements described here do not constitute the whole gene. There are no introns in the coding region (*Cytc2* Sequence) (Limbach and Wu 1985).

#### *Developmental Pattern*

*Cytc2* is present uniformly in all postembryonic stages and in adult head, thorax and abdomen. Expression is at very low levels relative to that of *Cytc1* (Limbach and Wu 1985).

#### **References**

- Limbach, K. J. and Wu, R. (1985). Characterization of two *Drosophila melanogaster* cytochrome c genes and their transcripts. *Nucl. Acids Res.* **13**:631–644.
- Sprinkle, J. R., Hakvoort, T. B. M., Koshy, T. I., Miller, D. D. and Margoliash, E. (1990). Amino acids sequence requirements for the association of apocytochrome c with mitochondria. *Proc. Natl. Acad. Sci. (USA)* **87**:5729–5733.

# 11

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## The *Dopa decarboxylase* Cluster: *Ddc*, *l(2)amd*, *Cs*, *DoxA2*

Chromosomal Location:		Map Position:
<i>Ddc</i>	2L, 37C1-2	2-54
<i>l(2)amd</i>	2L, 37B13-C2	2-54
<i>Cs</i>	2L, 37B13-C2	2-54
<i>DoxA2</i>	2L, 37B10-13	2-53.9

### Organization of the Cluster

The *Ddc* cluster is arbitrarily defined as those genes that fail to complement *Df(2L)TW130*, 37B9–C1 to 37D1–2, an 8–12-band deletion in the left arm of chromosome 2. The cluster contains 18 genetically identified genes plus three transcription units for which no mutations are known. Some of the genes in this cluster seem to be functionally related, most of them being involved in the formation, sclerotization and pigmentation of cuticle. Several genes in the cluster have mutant alleles that are female sterile. For three genes, *Ddc*, *l(2)amd* and *DoxA2*, some of the gene-product biochemistry is known; these genes are involved in catecholamine metabolism (Fig. 11.1) (Wright 1987).

Most of the genes are grouped in two very dense subclusters. The centromere-proximal sub-cluster contains nine elements in 25 kb of DNA, 70% of which is transcribed; the distal sub-cluster includes seven genes in 22 kb (Fig. 11.1) (Wright 1987).

The sequences of *Ddc* and *l(2)amd* are related, and it is probable that the genes originated by duplication. It appears unlikely, however, that all the genes in the cluster are members of a single family; the sequences of *l(2)37Cc* and *Cs*, for example, are not obviously related to *Ddc* or *l(2)amd*. Three genes in the proximal cluster and one in the distal cluster are presented here.

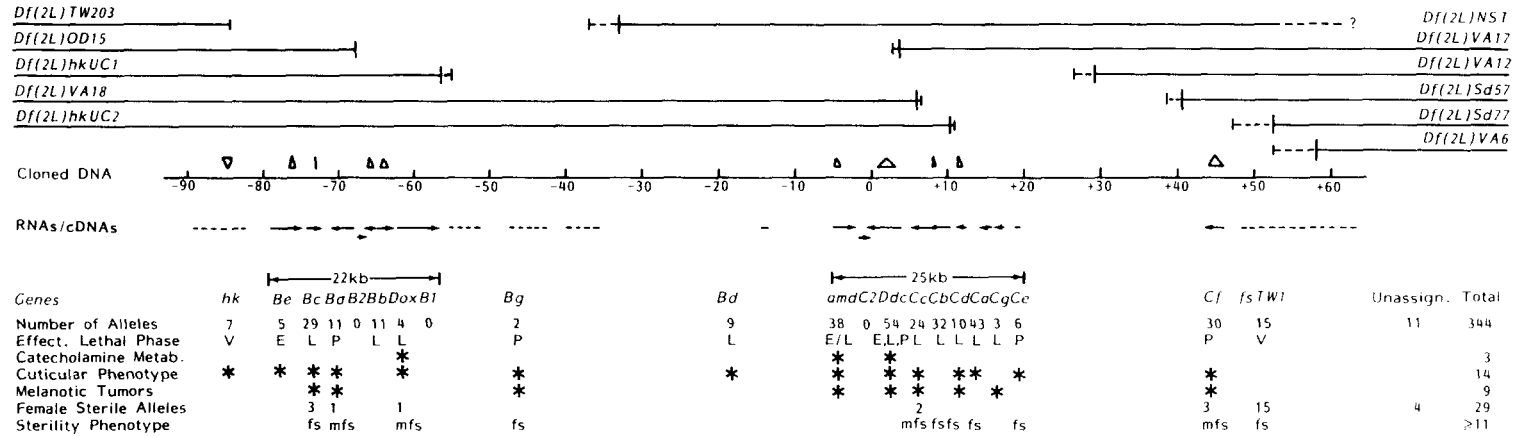


FIG. 11.1. *Ddc* cluster (centromere to the right), from Wright (1987), updated in 1992 by T. R. F. Wright: "The genetic and molecular organization of the *Ddc*-region. Deficiencies: *Solid lines* represent deleted DNA with *dashed lines* indicating uncertainty of the position of the breakpoint. Cloned DNA coordinates in kb from Gilbert et al. (1984). *Small triangles* above the cloned DNA line physically locate small deletion mutations and *short lines underneath* designate regions which hybridize to mRNAs or cDNAs with *arrowheads* representing direction of transcription. Transformed DNA lines indicate the segments of DNA that have been transformed by P elements. All the gene symbols except *hk*, *Dox = DoxA2*, *Bh*, *amd = 1(2)amd*, *Cs*, *Ddc*, and *fsTW1 = fs(2)TW1* should be preceded by '1(2)37', e.g., 1(2)37*Ba*. Effective lethal phase designations: *E* embryonic; *L* larval; *P* pupal; *V* viable. *Asterisks* underneath a gene symbol indicate the mutant alleles of that gene alter catecholamine metabolism, express a mutant cuticular phenotype, or produce melanotic tumors. Sterility phenotype: Individuals hemizygous for female sterile, *ts*, or hypomorphic alleles or heterozygous for complementing heteroalleles are female sterile = *fs* or both male and female sterile = *mfs*. See text for the sources of the information included in this figure." The transcription unit *Cs* is designated *C2* in this figure. The transcription unit *Cf* is actually transcribed toward the centromere.

***Ddc***  
**(*Dopa decarboxylase*)**

**Product**

Dopa decarboxylase (DDC, EC 4.1.1.26).

***Structure***

DDC is a homodimer of 54 kD subunits (Clark et al. 1978). Two forms of the enzyme, which are generated by alternative splicing, have been isolated; one is found in the central nervous system and the other in the epidermis (Morgan et al. 1986).

The amino acid sequence has considerable similarity with the DDC of mammals (Fig. 11.2) (Scherer et al. 1992), and prokaryotes (Jackson 1990). The heptapeptide consisting of residues 332 through 338 has similarities with the pyridoxal binding sites of porcine DDC and feline glutamate decarboxylase. Lys-337 is probably the pyridoxal-binding residue. See also *1(2)amd* below.

***Function and Tissue Distribution***

This enzyme catalyzes the decarboxylation of dopa (3,4-dihydroxy-L-phenylalanine) to dopamine, and of 5-hydroxytryptophan to serotonin. DDC is involved in tanning of the cuticle, and most of the enzyme is found in the epidermis where its activity peaks during the molting episodes. DDC is also involved in the synthesis of neurotransmitters and is present in a group of 150 serotonergic and dopaminergic neurons of the central and visceral nervous system (Wright et al. 1976a, 1976b; Konrad and Marsh 1987; Beall and Hirsh 1987; for reviews see Wright 1987 and Hirsh 1989).

***Mutant Phenotype***

Amorphic mutations are lethal; death occurs mostly in late-embryonic and larval stages. A few individuals survive to the pupal stage. Survivors have cuticular structures that are characteristically incompletely pigmented and sclerotized.

**Gene Organization and Expression**

Open reading frame, 475 or 510 amino acids; expected mRNA size, 2,067 or 1,923 bases, depending on splicing. The 5' end was defined by S1 mapping and primer extension. The 3' end was defined by cDNA sequencing. There are three introns, one in the leader, spanning -692 through -57, one after the Ser-33 codon and one in the Arg-62 codon. Two alternative splicing products have



```

      1                               50                               100
Dm  MSHIPISENTI PTKQTDGNGK ANISPOKLDP KVSIDMEAPE FKDFAKTMVD FIAEYLENIR ER.VLPEVKP GYLKPLIPDA APEKPEKWQD VMQDIERVIM
Rat  .....
CON  -----M---E F---K-MVD -IA-YL--I- -R-V-P-V-P GYL--LIP-- AP--PE---D ---DIE--IM

      101                               150                               200
Dm  PGVTHWHSPK FHAYFPTANS YPAIVADMLS GATACIGFTW IASPACTELE VVMMDWLGMK LELPAEFLAC SGGKGGGVIQ GTASESTLVA LLGAKAKKLK
Rat  PGVTHWHSPY FFAYFPTASS YPAMLADMLC GAIGCIGFSW AASPACTELE TVMMDWLGMK LELPEAFLAG RAGEGGGVIQ GSASEATLVA LLAARTKMIR
CON  PGVTHWHSP- F-AYFPTA-S YPA--ADML- GAI-CIGF-W -ASPACTELE -VMMDWLGMK LELP--FLA- --G-GGGVIQ G-ASE-TLVA LL-A--K---

      201                               250                               300
Dm  EVKELHPEWD EHTILGKLVG YCSDQAHSSV ERAGLLGGVK LRSVQSE.NH RMRGAALEKA IEQDVAEGLI PFYAVVTLGT TNSCAFDYLD ECGPVGKMKH
Rat  QLQAASPELT QAALMEKLVY YTSDAQHSSV ERAGLIGGVK IKAIPSDGHY SMRAALREA LERDKAAGLI PFFVVVTLGT TSCCSFDNLL EVGPICNQEG
CON  -----PE-- -----KLV- Y-SDQAHSSV ERAGL-GGVK -----S--N- -MR-AAL--A -E-D-A-GLI PF--VVTLGT T--C-FD-L- E-GP--N---

      301                               350                               400
Dm  LWIHVDAAYA GSAFICPEYR HLMKGIESAD SFNFNPHKWM LVNFDCSAMW LKDPSSVVVA FNVDPYLKX DMQGSA..PD YRHWQIPLGR RFRALKLWV
Rat  VWLHIDAAYA GSAFICPEFR YLLNGVEFAD SFNFNPHKWL LVNFDCSAMW VKKRTDLTEA FNMDPVYLRH SHQDSGLITD YRHWQIPLGR RFRSLKMWV
CON  -W-H-DAAYA GSAFICPE-R -L--G-E-AD SFNFNPHKW- LVNFDCSAMW -K-----A FN-DP-YL-H --Q-S----D YRHWQIPLGR RFR-LK-WFV

      401                               450                               500
Dm  LRLYGVENLQ AHIRRHCFNA KQFGDLCVAD SRFELAAEIN MGLVCFRLKG SNERNEALLK RINGRGIHL VPAKIKDVYF LAMAICSRFT QSEMEYSWK
Rat  FRMYGVKGLQ AYIRKHWKLS HEFESLVRQD PRFEICTEVI LGLVCFRLKG SNQLNETLLQ RINSAKIHL VPCRLRDKFV LRFVAVCSRTV ESAHVQLAWE
CON  -R-YGV--LQ A-IR-H---- --F--L---D -RFE---E-- -GLVCFRLKG SN--NE-LL- RIN---IHL VP---D--- L--A-CSR-- -S-----W-

      501                               516
Dm  EVSAAADEME QEQ*..
Rat  HIRDLASSVL RAEKE*
CON  -----A-----

```

FIG. 11.2. Comparison of the rat (Accession, M27716) and *Drosophila* (Dm) DDCs. There is 60% overall identity between the two proteins. Sequences aligned with the GCG *Pileup* program.



FIG. 11.3. Organization of the genes in the immediate vicinity of *Ddc*.

been detected. One is a 2.3 kb RNA in which all exons are present. The other, the most abundant, is a 2.1 kb RNA produced when the small second exon is spliced out together with the first two introns (*Ddc* Sequence). In the latter case the leader is spliced, in frame, onto the middle of the original open reading frame, and translation seems to start from an AUG six bases downstream of the splice site (Met-36) (Eveleth et al. 1986; Morgan et al. 1986). Transcription is toward the telomere (Fig. 11.3) (Spencer et al. 1986a).

Another gene in this cluster, *Cs*, is located immediately downstream of *Ddc*. The two genes are transcribed convergently and their untranslated 3' ends overlap by 76 bp (*Ddc* Sequence) (Spencer et al. 1986a; Eveleth and Marsh 1987).

### Developmental Pattern

The splicing reaction is tissue-specific with the 2.3 kb RNA occurring in embryos and in the nervous system and the 2.1 kb RNA involved in cuticular tanning. The 2.1 kb RNA is the predominant form during larval development; it is found in the integument fraction, and its level fluctuates according to the intensity of cuticle deposition (Eveleth et al. 1986; Morgan et al. 1986; Krieger et al. 1991).

### Promoter

**Proximal Elements** P-element-mediated transformation of genes carrying 5' deletions established that the 209 bp upstream of the transcription initiation site (up to position -1,093 in the *Ddc* Sequence) are sufficient for normally regulated full expression of *Ddc* in the epidermis. Deletions that leave only 25 bp of the 5' region (up to position -909 in the *Ddc* Sequence) result in much lower levels of mRNA production, but transcription is started correctly despite the absence of the TATA box (Hirsh et al. 1986). Progressively lower levels of DDC are produced when deletions are introduced in the segment between -1,093 and -922. In that segment, five putative regulatory elements have been identified on the basis of sequence similarities between the distant species *D. melanogaster* and *D. virilis*. Each of the putative regulatory elements includes the consensus sequence C(A/T)GCG(G/A) (Scholnick et al. 1986). In addition, a dimer of this consensus sequence, designated element I and lying between positions -970 and -957 is necessary for central nervous system expression in both glial cells and neurons. Element I is totally conserved in the two species, and this is the only segment of the proximal promoter region that is protected

*Ddc*

-2521	CCAATTAATTACAGATCGACTCTAAACGAATCTAATCACTTGCCCATATCATATAGATTTCAGACTAAATACGTGACCTATTGAAGCTCA	-2432
-2431	GCGATGTGATGTGTACACCAAAACACCCGCTCGTTTATCTCTGCCCTTGTTCACCCCATATGATGCCTGTTTATGCAATCCCCCTCTCAAA	-2342
	-----Df6	
-2341	GCGCCATTGACCCCTATAAGCGGAGAATACTTTCGCAATTCATTCGCAATCTAAGATGGTCATAAATCAAATTTAGTAAACTTCGCCT	-2252
	-----cf1	
-2251	CAATCGACCCGAACCTCCAGCCACCCGTAAGCAGCATAATGTGGTGGGTAGTTGGGCGACTGGTGGCTGGTGGCTGTTGGCTGGCGTGT	-2162
	-----bf2 -----uf3	
-2161	GGTGGAGCACCCAGCGCATTAAAAATCGAAAGCAGAGCCGTTGGCATGGCCGTATAAAATCTGTTGATTCAGCCAAGTGATTGCCAAAAT	-2072
-2071	GCTTCTGTTGAAATGTCAGGCACACGCACCTTGTCTGGCAGCTCAGCAACAGTTGGACCACCCGAGGATTCTTAGCAGCCCTACACTGAA	-1982
-1981	AGAAATTAATTTCTTTTGTGCTAGGCTAAAAATGTTTACTTGATCTTTTAAATAGTAATTAAGGAAGAGAATGATTTCTCTGCTGAT	-1892
	-----uf7 -----	
-1891	TCCAGGATCATTAGCCGAGCCGATATACCCATGTTGTCTGTCCGTATAAACTTCGAGATTTTGGGAACITTTAAAAAATAAAGGTC	-1802
	-----uf8	
-1801	CGAAACAGTTTTGAAAAATATTTGAATTTTTGTATTATATCTCTCGATATATTTGGCATAAACATTTAAGCCACATATTTATGTTTC	-1712
	-----uf9 -----uf10	
-1711	TTGCCAATTTCTATTGATATTTCAACTGAATTTTGAATTCGGCCAAAGTAACTGGCATCCAAAAGCTTTCTATAGTAATTTGAATTTT	-1622
-1621	TCTCAGTGATGCGGAACTGCCCGCTCAAAGGCTCAACCTAGCCCACTTCCCTCAGCAATGCGAAAGTGAGTGAGAGCATTGGATTA	-1532
-1531	TTTGACGTCACAATCCATGAGCGGTTCAAAAAGCAGCTCATATGTGGTCTCTAATAACCGGTTTCCAAGATGCGCGTAAAGCTGCCAT	-1442
-1441	TCCACGGCTTAATCAATTTCTGTCTTCTACGAATATAACTTTGTTTACATTTTTTGCCTGATTTTTTCTCGGGAGTCCAAGAAAA	-1352
-1351	ACCCTGTTTCGAGTGACTATAATTTGGGGATTCTCTGACGAGATCGCTCTTTCCACAATTCGAGTTGGGAACGACGTGAGCAGAATT	-1262
-1261	CAAAATGTTTTGCTTGTGTTTTAAATATCACTAGGTTCTCAAACATAATTTCAAAAATAATCAAATTAAGTTACAGAGCTGGCAAATAA	-1172
-1171	AATGTAATAGCTTGATGATGATATATATATATATTTTTTAAATCTAAATAAATCCATGAAAAATAATGCCTTTGATATCCAGTTACT	-1082
-1081	GATTCAGCGCCAATTAATGCATGTTCCAAAAAGTGTCAAAAAACGTGCACAAATCAAACGAGAGCAGAATTTGTTTTACGACAGCGG	-992
-991	CTGCGATTGGAAGTTCAGCGGCTGCGGACTGCGATTGAACCGTCTGCGGAATGGCAGCGCTGCTGGACGGGCTTTAAAGCCATGGC	-902
	I ----- -->-883 -----	
-901	CAAGAGCCGGGAGCGCTCAGTTAAGAGGAGAAGCCAAAGCGCACAGCAATCAGCACCAGAAATATCAGCATCGAAATATCAGCAAATAA	-812
-811	TATTAGCTGTTCTAAACAGGAGGGCAAACTGAACCTGGAGCAAAGATTTAGTTCGGAACGGAAGTAAAGCTCGGCAACAAGTGCAAACA	-722
	-----   ----- 	
-721	ATTAAGAGCAGGTTAACTAAAGTGAACCGTGAGAGACGAAAGTGGCTCCTCAACAGCCTCAGCTGCCTGAAGTCTGGCCAAACATA	-632
	----- 	
-631	ATGAGTGCATGTTGCATGCGAAAGATTCATTTGCGGGCTAACGCTGCGTATACGTAATGTGTATCTAAACTGGGCATATACTATAGCCT	-542



1529	CGCCAGTCCCAGTGCACGGAACTCGAGGTGGTTCATGATGGATTGGCTGGGCAAGATGCTGGAGCTGCCGGCAGAGTTCCTGGCCTGTTC eAlaSerProAlaCysThrGluLeuGluValValMetMetAspTrpLeuGlyLysMetLeuGluLeuProAlaGluPheLeuAlaCysSe	1618 (171)
1619	GGCGGCAAGGGTGGCGGTGCATCCAGGGCACGGCCAGTGAAGTCCACACTGGTGGCTCTGCTGGGAGCCAAGGCCAAGAAGTGAAGGA rGlyGlyLysGlyGlyValIleGlnGlyThrAlaSerGluSerThrLeuValAlaLeuLeuGlyAlaLysAlaLysLysLeuLysGly	1708 (201)
1709	GGTGAAGGAGCTCCATCCGGAGTGGGATGAGCACACCATCTGGGCAAGTGGTGGGCTACTGCTCGGACCAGGCTCACTCATCCGTGGA uValLysGluLeuHisProGluTrpAspGluHisThrIleLeuGlyLysLeuValGlyTyrCysSerAspGlnAlaHisSerSerValGly	1798 (231)
1799	GCGGGCTGGTCTTCTGGGCGGAGTAAAGCTCCGTTCCTGCAGTCCGAGAATCACAGAATGCGTGGTGGTGCCTGGAAAAGGCCATCGA uArgAlaGlyLeuLeuGlyGlyValLysLeuArgSerValGlnSerGluAsnHisArgMetArgGlyAlaAlaLeuGluLysAlaIleGly	1888 (261)
1889	ACAGGATGTGGCCGAGGGTTGATTCCTTCTACGCGGTGGTACCCTGGCCACCACCACTCTGCGCCTTCGACTACTTGGATGAGTG uGlnAspValAlaGluGlyLeuIleProPheTyrAlaValValThrLeuGlyThrThrAsnSerCysAlaPheAspTyrLeuAspGluCys	1978 (291)
1979	TGGACCGGTGGAAACAAGCACAATTTGGATCCATGTGGACGCTGCCTATGCCGGATCCGCTTTCATTGCCCGAGTATGCCACCT sGlyProValGlyAsnLysHisAsnLeuTrpIleHisValAspAlaAlaTyrAlaGlySerAlaPheIleCysProGluTyrArgHisLe	2068 (321)
2069	GATGAAGGGCATCGAATCAGCAGACTCTTCAATTTCAATCCACACAAATGGATGCTGGTGAACCTTGGACTGCTCGGCCATGTGGCTGAA uMetLysGlyIleGluSerAlaAspSerPheAsnPheAsnProHisLysTrpMetLeuValAsnPheAspCysSerAlaMetTrpLeuLys -----PYR	2158 (351)
2159	GGATCCCAAGTGGGTGGTCAACGCGTTCAATGTGGACCCTTTACCTGAAGCACGACATGCAGGGATCAGCTCCGGACTATCGTCACTG sAspProSerTrpValValAsnAlaPheAsnValAspProLeuTyrLeuLysHisAspMetGlnGlySerAlaProAspTyrArgHisTr	2248 (381)
2249	GCAATCCCACTTGGACGGCGATTGAGGGCATGAAGCTCTGGTTCGTCCTCCGGCTGTACGGTGTGAGAATCTCCAGGCCACATCCG pGlnIleProLeuGlyArgArgPheArgAlaLeuLysLeuTrpPheValLeuArgLeuTyrGlyValGluAsnLeuGlnAlaHisIleAr	2338 (411)
2339	CAGACACTGCAACTTTGCCAAGCAGTTCGGGGATCTCTGCGTGGCGGACTCCAGATTTGAACGTGGCCGCCAGATCAATATGGGATTGGT gArgHisCysAsnPheAlaLysGlnPheGlyAspLeuCysValAlaAspSerArgPheGluLeuAlaAlaGluIleAsnMetGlyLeuVa	2428 (441)
2429	CTGCTCCGGCTGAAGGGCAGCAACGAGCGGAAACGAAGCTCTTCTCAAGCGAATCAATGGACGCGGCCACATCCACTTGGTTCGCCCAA lCysPheArgLeuLysGlySerAsnGluArgAsnGluAlaLeuLeuLysArgIleAsnGlyArgGlyHisIleHisLeuValProAlaLys	2518 (471)
2519	GATCAAGGATGTCTACTTCTCGCGATGGCCATTTGCTGCGGATCACCCAGTCCGAGGACATGGAGTACTGTTGGAAGGAGGTGAGCGC sIleLysAspValTyrPheLeuAlaMetAlaIleCysSerArgPheThrGlnSerGluAspMetGluTyrSerTrpLysGluValSerAl	2608 (501)
2609	CGCTGCCGACGAGATGGAACAGGAGCAGTAAAGTGGTGTGCGAGTCTGTCCGTGTTAGTATATAAATTAATATAGTAACTTAAATT aAlaAlaAspGluMetGluGlnGluGlnEnd	2698 (510)
2699	GGACCAGTATGATATATAATGCATTGTGACTTGAACCCGGAACAGACCATACACTTCCACTTGCACATGTTTAGGGAATTTACATCG	2788
2789	CAACAAAAGATGGTTCGTCATCGCTACATTATATTTATAGTATCCTATCATTTGATCATTGATGTTTCATGATTTTTATTGTTAACG Cs <sub>n</sub> (A)	2878
2879	TTATGCGCCTAATTAACAATGTATTCTGCTTAAAAATACAACGAATTGTAACATATAAATTTTGACTAGTTTTTCGTGTTGATATACA ----- (A) <sub>n</sub> Ddc	2968
2969	CTGTACATTTAGCAGCCATTTCGGATTTCCATTTCACT 3006	

*Ddc* SEQUENCE. Accession, X04661 (DRODDC). The sequence was corrected by J. Hirsh by addition of a G at position -932. The acceptor site of the leader intron is 15 bases upstream from the position proposed by Morgan et al. (1986) (Shen and Hirsh, personal communication). Footprints in the promoter region are indicated by underlining; there are eight in the distal region and one in the proximal region. *B-ORF*,

(continued)

from nuclease digestion by an extract from embryonic nuclei (Bray et al. 1988, 1989). It has been reported that *Ddc* and *Ubx* may have a regulatory protein in common (Biggin and Tjian 1988).

**Distal Elements** In addition to the proximal elements, expression in the central nervous system also requires certain more distal *cis* sequences located in an 863-bp segment between  $-2,506$  and  $-1,643$  (Johnson et al. 1989). Eight protein-binding sites were detected within that segment (*Ddc* Sequence) by nuclease protection assays. Partial deletions of the distal promoter region, re-introduced into transgenic organisms, showed that *uf8*, *uf9* and *uf10* are not essential for neuronal expression. On the other hand, deletion of either *uf7* or *bf2* and *uf3* leads to complete loss of neuronal activity. The element *cf1* appears to be essential for expression in the medial dopaminergic neurons (Johnson et al. 1989). The gene for a POU/homeobox protein that binds to *cf1* has been cloned (Johnson and Hirsh 1990). A 40-bp segment between  $-2,519$  and  $-2,479$  is necessary for expression in serotonergic neurons (Johnson et al. 1989).

***l(2)amd***  
**( $\alpha$ -methyl dopa sensitive)**

**Product**

Unknown.

**Structure**

The sequence of the coding regions show 55% identity with the dopa decarboxylase sequence. The amino acid sequence similarity is particularly high near a putative pyridoxal-binding site (starting at position 298) (Eveleth and Marsh 1986; Marsh et al. 1986).

**Function**

AMD is thought to be involved in the metabolism of catecholamines judging by the  $\alpha$ -methyl dopa sensitivity of mutants (for a review, see Wright 1987).

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(*continued*) between positions 84 and 85, is a 4-bp insertional mutation that alters the reading frame of the second exon and leads to the absence of DDC in the central nervous system but not in the epidermis (Morgan et al. 1986). The putative pyridoxal-binding site starting at Asn-332 is underlined. The poly(A) site,  $_{n}(A)$  (near 2,850) of the partly overlapping gene *Cs*, is indicated.



1406	ACTTCGATTGCTCGCCATGTGGCTAAGGGATGCCAACAAAGGTGGTCGACAGCTTCAATGTGGATCGCATCTATCTGAAGCACAAAGCACG snPheAspCysSerAlaMetTrpLeuArgAspAlaAsnLysValValAspSerPheAsnValAspArgIleTyrLeuLysHisLysHisG	1495 (388)
1496	AGGGTCAGTCGCAAAATCTAGACTTCGCATTTGGCAAATCCCCTGGGTCGCGCTCCGAGCTCTAAAAGTCTGGATCACATTCCGCA luGlyGlnSerGlnIleProArgLeuProSerLeuAlaAsnProLeuGlyArgArgPheArgAlaLeuLysValTrpIleThrPheArgT	1585 (368)
1586	CTCTGGAAGCCGAGGGATTGCGAAACCATGTCGCGAAGCACATCGAGTTGGCCAAACAGTTTGAGCAGCTTGTGCTCAAGGATTCCGCGAT hrLeuGluAlaGluGlyLeuArgAsnHisValAlaLysHisIleGluLeuAlaLysGlnPheGluGlnLeuValLeuLysAspSerArgP	1675 (398)
1676	TCGAGCTGGTGGCTCCTCGTGCCTGGGACTGGTTTGTTCGGGCCAAAGGTGACAAATGAGATTACCACCCAGTTGCTGCAACGGCTTA heGluLeuValAlaProArgAlaLeuGlyLeuValCysPheArgProLysGlyAspAsnGluIleThrThrGlnLeuLeuGlnArgLeuM	1765 (428)
1766	TGGATCGAAAGAAGATCTACATGGTTAAGGCCGAGCATCGGGTCGTCAGTTTCTGCGATTGCTGCTGATGCGGCATGGACACCAAAGCCT etAspArgLysLysIleTyrMetValLysAlaGluHisAlaGlyArgGlnPheLeuArgPheValValCysGlyMetAspThrLysAlaS	1855 (458)
1856	CCGATATTGATTTGCCTGGCAGGAGATCGAGTCTCAACTGACGGACCTGCAGGCGGACGAATCCTTGGTGGCCCGCAAATCGGGAAACG erAspIleAspPheAlaTrpGlnGluIleGluSerGlnLeuThrAspLeuGlnAlaAspGluSerLeuValAlaArgLysSerGlyAsnV	1945 (488)
1946	TCGGCGATCTTGGCAGCAGACTTCAGATCCATCTGAGCACCGAAAATGCAACGACGAGAAAATCTCAGTGAGAAAAACGGATAAACTATT alGlyAspLeuAlaHisAspPheGlnIleHisLeuSerThrGluAsnAlaThrHisGluLysSerGlnEnd	2035 (510)
2036	TATGTTTAGGGACAACCTAGTTAGTTTGCATGTAGTTTTTAACCTTCCACATGTTTATAAATAAAGTGAAATTAATGTACGATCATT ----- (A) <sub>n</sub>	2125
2126	GGCAGCTTTTCTATAAAGGTAGAGTGGTTTTTCCCTGTCATTTTTTTTTGTGCAAAAAGTTCCCAACATCTCTGTAAACTTTCTGCCG	2215
2216	AGGCTTTAGTTTTTAAAGCATTACAATATCGTCGACTTTTTTAAAAATTTAAACCAAAATTTTCGCGGCTTAGTGTGACTGCATT	2305
2306	GGTTATGAATCGATACACTTCTTCATCGCCCTTCGATAAGTTCGCCAAGGTCTATCGTCATGTGCGGATCCGAGGGCAACAGCTGTTT	2395
2396	CTCCAATTTGGACCACCTGATATCGGTTAAATAACAAAGATAAAACAAAACAAAATATCTGTTTCCCTTAATTCATATTTTGATTAG EcoRI	2485
2486	CTTTGAATAGCGTTTAGTGCTATTTCTCATAAATATAGAATAGAAGCAGCCGCGGCTCGCCTTTGTACGAATTC	2559

*l(2)amd* SEQUENCE. Strain, *Canton S*. Accession, X04695 (DROL2AMD). The sequence ends at the *EcoRI* site at which the *Cs* Sequence, begins. The exclamation mark indicates the 5' end of the longest cDNA sequenced.

## Gene Organization and Expression

Open reading frame, 510 amino acids; expected mRNA size, 1,782 bases. The 5' end was tentatively identified on the basis of sequence features in the neighborhood of the 5' end of a cDNA clone. The 3' end was identified from the sequence of two cDNA clones. There is one intron after the Trp-105 codon. The distance from the polyadenylation site of *l(2)amd* to the transcription initiation site of *Cs* is 682 bp.

Although the length of the coding region is the same as that of *Ddc*, and the sequence is similar, the position of the introns in the two genes do not match. In the aligned sequences, the 5' end of *amd* coincides approximately with the second *Ddc* intron; and the *amd* intron is approximately 250 bp away from the position of the third *Ddc* intron (Fig. 11.3 and *l(2)amd* Sequence) (Eveleth and Marsh 1986).



## Cs

-658	_____EcoRI GAATTCTCAGATTTCTGTGAGTAAATAATCATATATGTAACATACAATAACATCCGAATTACTATAACCCCTTCAGCCAAAGTTTGAATTA	-569
-568	GACACCCAAACTGCCAATTGGATAACCGGCCACCATTGTGGTGGATACTATGATTCGCTTTTTAAAAACAACCTGGACGTGTGGCACTT	-479
-478	TAAAGCCTATACGCCTCTCAGCCTGTCAACAAATATTAATAAAATCTGGCAAAATCTAAAAATAACTTGATTACTTTCCGGAACCTCCAG	-389
-388	GAAACTAGGCCCGCTTGCCATGCAATGGTAAGTGAACAGCTCCAGCGGATTTGAATGTGCAAACTAAACCTTCTCTGTGATCCCCGCAGT	-299
-298	TTTAAACTGGCCAGCAGCGCGAGCTTATACAATGCACGGGTTCTACAGCGGATAACATCGCGACAAGCAACGCAGTCCAGATCTGGAG	-209
-208	CGCGCGCCAAAATACCCAGATAGTGGTCTGGGGCCAGGACTCGCCGGTCTCTCGCGGCCAGCACCTTGTGTCGACGGCTTCGCGC	-119
-118	GCACGTGTATCTGGAGGCCACAGATCGTTATGGCGGCAGGATTAACACCCAGCGCTTTGGTGACACCTACTGTGAAC TAGGCGCCAAGT	-29
-28	GGGTAAGATCGATGGATCGCAGGATTCGATGTATGAACTGCTACGCAACACGGAAGGCTTGGGAAGCAGATAAAGCAGGCCGGATCGG	61
	MetTyrGluLeuLeuArgAsnThrGluGlyLeuGlyLysGlnIleLysGlnAlaGlySerG	(21)
62	GCCACCTATCTTCAGGATGGAAGCCGCATCAATCCAGCCATGGTCGAGCTTATCGACACGCTATTTGGCAGCTTTGCCAGGCTTCAAGG	151
	lyHisLeuSerSerGlyTrpLysProHisGlnSerSerHisGlyArgAlaTyrArgHisAlaIleSerAlaAlaLeuProGlyPheLysV	(51)
152	TCTCCGAACGAGTTAAAACGGGTGGTGACCTGCACTCGCTGGACAATGTCATGAACTACTTTAGAACAGAAAGCGATCGCATCATTGGCG	241
	a1SerGluArgValLysThrGlyGlyAspLeuHisSerLeuAspAsnValMetAsnTyrPheArgThrGluSerAspArgIleIleGlyV	(81)
242	TCTCCTCCAGCATCTCAAGGATCAACTGGCGGCACGCGAGATCTCCAATCGCTGTCAAGGAGTTCGGCAGCATCTTGGGATGCTGCC	331
	a1SerPheGlnHisProLysAspGlnLeuAlaAlaArgGluIlePheGlnSerLeuPheLysGluPheGlySerIleLeuGlyCysCysL	(111)
332	TGGAGTACGTGAACATCGAACACATAACCAAGTGCCAGTGCAGCAGGAACAGCGCCCGCTTATGTGCCACTGGTCTAGATAATGTAG	421
	euGluTyrValAsnIleGluHisIleThrLysCysProValGlnGlnGluGlnArgProArgTyrValProThrGlyLeuAspAsnValV	(141)
422	TGGACGATCTCATTAGAACATGGACAAGCGCAGCTGCAGACCGGAAAGCCTGTGGCCAGATACAGTGGACACCAGCGCCGATGAAAA	511
	a1AspAspLeuIleGlnAsnMetAspLysAlaGlnLeuGlnThrGlyLysProValGlyGlnIleGlnTrpThrProAlaProMetLysS	(171)
512	GTGTGGTTGCCGTGGATGGCAGTCTTTACAACGCCGATCACATAATATGCACCCTCGCGCTCGGGGTGCTCAAAGCTTTGGCGGTTCT	601
	erValGlyCysLeuAspGlySerLeuTyrAsnAlaAspHisIleIleCysThrLeuProLeuGlyValLeuLysSerPheGlyAlaPheC	(201)
602	GTTTCGACCCACGCTGCCGCTGGACAAGATGCTGGCTATCACGCAACCTCGGCTTTGGCAATCCCCTCAAGATATATCTCTCTACAAG	691
	ysPheAspProArgCysArgTrpThrArgCysTrpLeuSerArgAsnLeuGlyLeuTrpGlnSerProGlnAspIleSerLeuLeuGlnG	(231)
692	AAGCCATCTGGTGGCTAAAGGGAAGCTGCCCATGGAACGTTCTGAATCTTCTGAGAGCAGCAACCGAACGCAACTGGACGCGAGGTT	781
	luAlaIleLeuValAlaLysGlyLysLeuArgHisGlyThrPheEnd	(245)
782	CGTGGAGATAGCCAGGTGCCACAGCTCAGCATGTGCTGGAGGTGCATGTGGTGGCGGATACTACGAGGAGATCGAGAAGCTGCCGATG	871
872	AGGAGCTGCTGGAGCAGATAAATGCTCTGCTAAGGCGCTGCGTGAGCAGTACCCTGGTGGCGTACCCACAGGAACCTGCTGCTTCCAAC	961
962	GGAGCACCTCGGCTGCTACCTCGGCCGTCGCTCTTACTTCTCCACCAACAGCAGTGCCCGGATGTCCAGCGACTGGCCGCTCCGGC	1051
1052	TGGCGGAGAAGTCCGGGCTGCTCTTGTCTGGGATGCAACCTCGCTGAAAAGGCTTTGGAACATTGATGCCGCCACGTCAGTGGCAT	1141
1142	CCGAGAAGCCCAATGTATCATTGACTACTATCTGAAAAGCGTGCACCTGCGGTTAAGTAAAATGGGAAATCCGAATGGGCTGCTAAATTGT	1231

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1232 ACAGTGATATATCAACACGAAAAGTGTCAAATTTATAGTTACATTCGTTTGTATTTTAAGCAGATACATTGTTTTAATTAGGCGCAT 1321
      Ddc  n(A)|
1322 AACGTTAACAAATAAAATCATGAACAACATCAATGATACAATGATAGGATACTATAAAATATAATGTAGCGATGGACGAACCATCTTTTGT 1411
      ----- |(A)n
1412 TGCATGTAAATCCCTAAACATGTCGCAAGTGGAAGTGTATGGTCTGTTCCGGGTTCCAAGTCACAATGCATTATATATCATACTGGT 1501
1502 CCAATTTAAGTTACTATATTAATTTATATACTAAACACGGAAACAGACCTGCACAACCACTTTACTGCTCTGTTCATCTCGTCGGCAG 1591
1592 CGGCGGTGACCTCCTCCACGAGTACTCCA 1621

```

*Cs* SEQUENCE. Strain, *Canton S*. Accession, X05991 (DROCSG). The sequence starts with the *EcoRI* site at the end of the *1(2)amd* Sequence. The exclamation mark indicates the 5' end of the longest cDNA sequenced. The poly(A) site,  $n(A)|$  (at 1,267) of the partly overlapping gene *Ddc*, is indicated.

### Developmental Pattern

A 2 kb RNA is detected in 8–16 h embryos and, at a much lower level, in adults.

## *Cs*

### Product

Unknown. It has been questioned whether this protein is ever synthesized, although the corresponding mRNA is found in association with polysomes. No mutations have been recovered in this transcription unit despite intensive screens involving the region (Eveleth and Marsh 1987 and references therein).

### Gene Organization and Expression

The longest open reading frame is 245 amino acids; but several smaller open reading frames exist, some with the starting codon upstream of the longest one. The presence of those upstream AUGs and the very poor codon bias displayed by this mRNA suggest that translation may be very inefficient. The expected mRNA length is 1,696 bases, in agreement with a 1.9 kb band detected in gels. A cDNA sequence was used to define the 5' end. The 3' end was obtained from the sequence of two cDNAs that included poly(A) tails. There is a leader intron at –361/–300 (*Cs* Sequence) (Eveleth and Marsh 1987).

### Developmental Pattern

Transcription of *Cs* occurs mainly in the first 8 h of embryonic development; the highest levels of transcript are detected in 3 h embryos (Spencer et al., 1986b).

*DoxA2*

1(2)37Bb. <--

-399 GTGCGATGTTATCGGAGTATCGATATCGAAAAGCCTTAACGGAATTGTGGTAATGTTTTATTGCAATTTAAATAAAAAACGCTTTTACT -310

-309 GAGTGCCTAACTTAAGAAATTCCTAACCCAAATTAAGCAATAGATACATTTACTGTAAAACATTAATAATAAATTCCTATAACATATGA -220

-219 GACTGAAAGTTCGCTCAGTGTACCGTAAACGTATCGATAAATGAAACGTAAACGGCTTAACAGCTCTGTTAACCAACTAAATTTACCAG -130

-----  
 --->-89

-129 CACTGCCTGTAGCCGAAAACGAATAAGAAGAAGAAGCAGACATTACTAGGCATTTTTGATTGGGATTGAGAAAAACAAAAGAAAAGTCGGC -40

-39 TATATTTGTGACCCAGTAAATTGAGAGTTCATTACAAAATGACCAACGCAACGGACATCGGTGCTAACGACGTGGAGATGGAGGTGGA 50  
 MetThrAsnAlaThrAspIleGlyAlaAsnAspValGluMetGluValAs (17)

51 TCCAACGGCGGAGACGCTGGCTGACGAGAAGAAGAACCAAGATGTGGCCGCCGTGCAGGAGATCCGCGAGCAGATTCTGCAGATTGAGAA 140  
 pProThrAlaGluThrLeuAlaAspGluLysLysAsnGlnAspValAlaAlaValGlnGluIleArgGluGlnIleArgGlnIleGluLy (47)

141 GGGGGTAGCCTCGAAAAGAGTCGCGGTGAGTAGTGCAAGAATTAATACTTGTCCTTCTTATTATGGCTCATTTCCGCCAACAGCTTCA 230  
 sGlyValAlaSerLysGluSerAr gPheI (57)

231 TCCTGCGCTCCTTCGCAATTTGCCAACACTCGTCGCAAGCTGAACGGCGTCGCTTCCGGAATCTTGACACAGAGATTTACCCCGCTG 320  
 leLeuArgValLeuArgAsnLeuProAsnThrArgArgLysLeuAsnGlyValValPheArgAsnLeuAlaGlnSerIleTyrProAlaG (87)

321 GTGCAGATCGTAGGGCGGCGTGGCTTTGATGCCCGCTGTGGAGAAAGACGCCACCGAGCTGCCGATGTTCCAAAAACAAGTTGCCA 410  
 lyAlaAspArgGluAlaAlaValAlaLeuMetProAlaValGluLysAspAlaThrGluLeuProAspValProLysLysGlnValAlaT (117)

411 CCAAGGCTCCAATCGCGAGGTCGATGCCTACTTCTACCTGCTCTGCTGGTCAAGCTCATCGACGCCAGTGATTTAAAGCGGGCCGGAA 500  
 hrLysAlaProIleAlaGluValAspAlaTyrPheTyrLeuLeuLeuValLysLeuIleAspAlaSerAspLeuLysArgAlaGlyI (147)

501 TTAGCGCCGACGCCCTAATGGCCAAAATCTCCATCCAAAACCGACGCCACCCCTTGATCTGATTGGTGCCAAAGTCTACTTCTATTTTCAA 590  
 leSerAlaAspAlaLeuMetAlaLysIleSerIleGlnAsnArgArgThrLeuAspLeuIleGlyAlaLysSerTyrPheTyrPheSerA (177)

591 GAGTGGCGGAGCTAAAACTCACTGGAAGGCATACGCTCGTTCCTGACGCTCGTCTGCGCACCGCTACGCTGCGTAATGATTTTGAAG 680  
 rgValAlaGluLeuLysAsnSerLeuGluGlyIleArgSerPheLeuHisAlaArgLeuArgThrAlaThrLeuArgAsnAspPheGluG (207)

681 GCCAGGCGGTGCTTATTAACGTGTTGCTCCGCAACTACTTGCACTATGCTTTGTACGACCAAGCCGACAAGCTGGTAAAGAAATCCGCTCT 770  
 lyGlnAlaValLeuIleAsnCysLeuLeuArgAsnTyrLeuHisTyrAlaLeuTyrAspGlnAlaAspLysLeuValLysLysSerValT (237)

771 ACCCGGAATCGGCCAGCAACAATGAATGGGCGCTTTCCCTGTACTATCTAGGTCGGATTAAAGCCGCTAAGCTGGAGTACAGCGATGCC 860  
 yrProGluSerAlaSerAsnAsnGluTrpAlaArgPheLeuTyrTyrLeuGlyArgIleLysAlaAlaLysLeuGluTyrSerAspAlaH (267)

861 ACAAGCATCTGGTCCAGGCCCTGCGTAAGTCGCCGACGCTGCCATCGGCTTTCGTCAGACGGTTCAAAGCTAATTATCGTTGTGG 950  
 isLysHisLeuValGlnAlaLeuArgLysSerProGlnHisAlaAlaIleGlyPheArgGlnThrValGlnLysLeuIleIleValValG (297)

951 AGCTGCTTTTGGGCAACATCCCGGAGCGTGTGGTTCGCGCAAGCGGCTCTCGCCAACTCTTTGGTGCTACTTCCAGCTCACGACGG 1040  
 luLeuLeuLeuGlyAsnIleProGluArgValValPheArgGlnAlaGlyLeuArgGlnSerLeuGlyAlaTyrPheGlnLeuThrGlnA (327)

1041 CCGTGCTGTGGGCAACTGAAGCGCTTCGCGGACGCTGGTATCCCAATACGGACCAAGTTCCAAGTGGACCACACATTCACCTGATTA 1130  
 laValArgLeuGlyAsnLeuLysArgPheGlyAspValValSerGlnTyrGlyProLysPheGlnLeuAspHisThrPheThrLeuIleI (357)

1131 TCCGGCTGCGCCACAATGTGATCAAGACGCGCAATCCGCTCCATCGGACTATCGTACTCACGCATCTCGCCGCAAGACATTGCCAAGCGGC 1220  
 leArgLeuArgHisAsnValIleLysThrAlaIleArgSerIleGlyLeuSerTyrSerArgIleSerProGlnAspIleAlaLysArgL (387)

1221	TAATGCTAGACTCCGCGGAGGATGCCGAGTTTATTGTATCGAAGGCTATACGGGACGGCGTGATTGAGGCTACGTTGGACCCAGCCCAGA euMetLeuAspSerAlaGluAspAlaGluPheIleValSerLysAlaIleArgAspGlyValIleGluAlaThrLeuAspProAlaGlnA	1310 (417)
1311	ATTTTCATGCGCAGCAAGGAAAGTACGGACATCTACAGCACCCGGGAACCGCAGCTGGCCTTTCACGAGCGCATCTCGTTCTGCCTGAACC snPheMetArgSerLysGluSerThrAspIleTyrSerThrArgGluProGlnLeuAlaPheHisGluArgIleSerPheCysLeuAsnL	1400 (447)
1401	TGCACAACCCAGAGCGTTAAGGCCATGCGCTATCCCCAAAGTCTACGGCAAGGATTTGGAGAGCGCCGAGGAGAGACGCGAGCGGGAGC euHisAsnGlnSerValLysAlaMetArgTyrProProLysSerTyrGlyLysAspLeuGluSerAlaGluGluArgArgGluArgGluG	1490 (477)
1491	AGCAGGACCTTGAGCTGGCCAAGGAGATGGCCGAGGATGATGAGGATGGTTTCTAAGCGGCTGATTCTGCAAATTAATTTGTGCTTGCAT lnGlnAspLeuGluLeuAlaLysGluMetAlaGluAspAspGluAspGlyPheEnd	1580 (494)
1581	TCATTTTTATAGAAATATAATCCGCAATTAATAAGTTACAATAATTTTCGGAACCTTTTAATTAGGTATTGGAATCAAATAGTTCAGAAC -----      -----       (A) <sub>n</sub>  (A) <sub>n</sub>	1670
1671	TGATCTTCTTTATTCAAGCAAAGTTGTATGTTGTTGTTGGTAGACATCAAATTCATCGTAGAATGAACATTAAGTTCATTCTG	1754

*DoxA2* SEQUENCE. Accession, M63010 (DRODOXA2). At -364 is indicated the 5' end of the neighboring gene *1(2)37Bb*, which is transcribed in the opposite direction.

### *DoxA2* (Diphenol oxidase component A2)

#### Product

Component A2 of phenol oxidase (PO) (EC 1.10.3.1).

#### Structure

Sequence comparisons involving entire amino acid sequences show 57% identity between DOXA2 and the mouse *tum*<sup>-</sup> transplantation antigen P91A; the similarity is even greater in the C-terminal two-thirds of the protein (Pentz and Wright 1991).

#### Function

PO has three components: A1 acts on monophenols, A2 and A3 on diphenols, including dopa and its derivatives; it is involved in the oxidation of catecholamines to quinones, compounds that are subsequently utilized to produce melanin or to cross-link cuticular proteins. Thus, PO plays a central role in eggshell and cuticular sclerotization, in melanization and in defense against pathogens. A2 (like A1 and A3) is synthesized as a proenzyme and activated, probably by proteolysis, via an activation cascade.

#### Mutant Phenotypes

Homozygous *DoxA2* mutants die primarily during the larval stages; however, rare pharate adults can be recovered, and these are totally unpigmented (Pentz et al. 1986 and references therein).

## Gene Organization and Expression

Open reading frame, 494 amino acids; expected mRNA length, 1,649 and 1,657 bases in agreement with a 1.7 kb band detected in gels. There are two alternate 3' ends; the positions of these were obtained from two cDNA sequences terminating in poly(A) tails. S1 mapping and a cDNA sequence were used to define the 5' end. There is no apparent TATA box. There is an intron in the Arg-55 codon (*DoxA2* Sequence) (Pentz and Wright 1991).

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# 12

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## Elongation Factor Genes: *Ef1 $\alpha$ 1*, *Ef1 $\alpha$ 2*

### Chromosomal Location:

*Ef1 $\alpha$ 1* 2R, 48D  
*Ef1 $\alpha$ 2* 3R, 100E

### Map Position:

2-[64]  
3-[102]

Synonyms: *Ef-1 $\alpha$ F1* and *Ef-1 $\alpha$ F2*

### Products

Translation elongation factor 1 alpha (EF1 $\alpha$ ), one of three components of elongation factor 1.

### Structure

There is remarkable conservation of the amino acid sequence in very distant species (Fig. 12.1). The similarities are particularly noteworthy in a region near the N-terminus that is thought to be the GTP binding site and in the neighborhoods of Ala-92, Lys-244 and Lys-273, residues that are considered important for tRNA binding (Walldorf et al. 1985; Hovemann et al. 1988).

### Function

EF-1 $\alpha$  is involved in the GTP-dependent binding of charged tRNAs to the acceptor site of the ribosome. A decrease in EF-1 $\alpha$  levels after emergence of adults seems to play a role in the aging process (Webster 1985). Conversely, increased expression of *Ef1 $\alpha$ 1* under the control of a heat-shock promoter leads to extended life spans (Shepherd et al. 1989).

### Comparison Between *Ef1 $\alpha$ 1* and *Ef1 $\alpha$ 2*

There is 90.5% identity and 93.3% similarity between the *Drosophila* sequences. Differences between the amino acid sequences of EF1 $\alpha$ 1 and EF1 $\alpha$ 2 are comparable to the interspecific differences found between the fly and rat





*Ef1 $\alpha$ 1*

-1881	CTCAAGCTTCCATTGTTATTTAAAGTTCTATTACGTTAGGGTTCACATACAAATTAAGTGGCAGGTTCTATCTCAAAACATTCGTTCAA	-1792
-1791	AATGCGGACTACTAATGCAATTGTTATTGTTTTACATATTAAGATATGTGTTCCAATATTACGTATAGAAATATAGACATCGTTTT	-1702
-1701	GTAGAAAATACTTTTGGAATCACTGATTATTTAGTTTTTCATATAAAAACAATGTCGAGCAAACAAGGTTTTTAAATTCCTCAATCTTT	-1612
-1611	AGGTTATTGTATTTGGCCACTTTCATCACTTAAATTTCAATAAAATGAAGTGCTTCATTGCGCGTAGTGGAACACCCGAGTGGGAAC	-1522
-1521	ACG6TTTCTGCTCTTTTGACAGTTGCGTAGCTTCGGTCACACCATGTGTCAAACGAGGCTTCCTGTGCTGAGCTCTGCCGAACGCTCGTT	-1432
	-->-1431	
-1431	CACTTTGTTCGAATCCGTCGCCGCTTAGACTTCGTGATTTCTCATTACGCTTATTAGAGAGTAAGTTTTACCTGCGAGGCTATAATTAAG	-1342
-1341	TGATTTCTGCAAAAAAAGTGCAGGGGGGAAACAATTTATAAACAATATGCAGCTGAGACGCCGAATTTGTGCATATTTCCAGTGTTTTT	-1252
-1251	CCTGTGTGTGTGTAATAAACCCGGAGATAACCTCTAACTCGGTTTTCCAAGTGAAAGTGGCCATAGAAGCAAACACGTGGCAAGTCT	-1162
-1161	GCAAAGGCAAAAATTTAACTGGCGTCCCAAGTAAAGTCCCAAGCATCTCAAATAATTTCCGGCTTTCCGGCCGCAATTTTCGCC	-1072
-1071	TGCAATATGGTGCACCTAGCGTGTAATTACTTTGCCACGCCACGCCGGACACAGAGGTCAATCCACAGATGTGCTCATTAAACCGAGAAA	-982
-981	AAAAACGTGCTTTCTCTTTGCCTTTGTCATGGCTATAGATATTCCTATTCTTTCTTTTTGCGGCATGGAATTTAAAATGGCGACC	-892
-891	CAGTGGCGTAGTCAAGTGGGCGAAAAAATTCGCCGGAACAAGCGAAAAATGTGCTTTTTGGGTTCCAGCCATTAGCATATCTG	-802
-801	GTGTAATGGCACTCGCATCAGCTATTTTCGCCATTTCCAACCGACTCAATAATTTGGTTTTGGTAAAATGGCTGCCGCTGCACTACGTTCTT	-712
-711	GATTAATTCGTTGTGCCCCCTCTTTTTTCATTTCTTTCCAATTACCAATTGTGCCACCGGGCGGAGACGCTTGCAATTTGTACAAGTC	-622
-621	ACACACGCACACTAATGCACATCCGCCATTTTGGTCTCTCTCTCTCTCTTACTTTTTCCGGCCGGCAACAGCGTCCACAAATACA	-532
-531	CAGGCATAGATATACACACGCATAGCCAGATAAGCACATGTGATTTTGCGAATTAATTTGCTGGAATTTTCCTTTGGACTCTTCGATTT	-442
-441	AACATGATGATGATTTTTAGTTCTGCTACTGAAGAGAGTTGACAGAAAGCAAAAATACCAAAATCACTGAAACAAAATCGAGTTCCAT	-352
-351	ATGGAATTTTATTGACAGCTCTTTCTGTAGTTGCGCCCCACTCGTTTTACCCACACCCCTACATGCGGGCACTGGTCTAACCTCAAA	-262
-261	AAACACGTTTTGTACGGCTGCAAGAGTTGAGGTTAGGTTGTGCTCGCGCATGCAAAACAAAAGTCGAACGTACGTAAGGAAATGAGAAA	-172
-171	GTGTTATACCCACTAATAATTGTAGTTGTAATCCACCGAATTGTTTTACCCTTTGTTTATCCAACCTCTTGTCTCGCAACCCGCCG	-82
-81	AACCCGCAACCTTCCAATGTTCCAAGTTCGGTTAATCCAACACTCGAATACACACAACAGCCATAGTGTAATCATCCAACATGGGCAA	8
		MetGlyLy (3)
9	GGAAAAGATTACATTAACTTGTGCGTATCGGACACGTCGATTCGGTAAGTCGACCACCACCGGACACTTGATCTACAAGTGGCGGTGG	98
	sGluLysIleHisIleAsnIleValIleGlyHisValAspSerGlyLysSerThrThrThrGlyHisLeuIleTyrLysCysGlyGly	(33)
99	TATCGACAAGCGTACCATCGAGAAGTTCGAGAAGGAGCCAGGAGATGGGAAAGGATCCTTCAAGTACGCTGGGTTTTGGATAAGTT	188
	yIleAspLysArgThrIleGluLysPheGluLysGluAlaGlnGluMetGlyLysGlySerPheLysTyrAlaTrpValLeuAspLysLe	(63)

189	GAAGGCTGAGCGCGAGCGTGGTATCACCATCGATATCGCCCTGTGGAAGTTCGAAACTGCCAAGTACTACGTGACCATCATTGATGCCCC uLysAlaGluArgGluArgGlyIleThrIleAspIleAlaLeuTrpLysPheGluThrAlaLysTyrTyrValThrIleIleAspAlaPr	278 (93)
279	CGGACACAGGGATTTCATCAAGAACATGATCACTGGTACCTCGCAGGCCGATTGCGCCGTGCAGATTGACGCCCGGAACCGGAGAATT oGlyHisArgAspPheIleLysAsnMetIleThrGlyThrSerGlnAlaAspCysAlaValGlnIleAspAlaAlaGlyThrGlyGluPh	368 (123)
369	CGAGGCCGGTATCTCGAAGAACGACCAGACCCGCGAGCAGCCCTGCCTGCCCTTACCCTGGGTGTGAAGCAGCTGATCGTTGGTGTGAA eGluAlaGlyIleSerLysAsnAspGlnThrArgGluHisAlaLeuLeuAlaPheThrLeuGlyValLysGlnLeuIleValGlyValAs	458 (153)
459	CAAGATGGACTCCTCCGAGCCACCATACAGCGAGGCCCGTTATGAGGAAATCAAGAAGGAGTGTCTCTTACATCAAGAAGGTCGGCTA nLysMetAspSerSerGluProProTyrSerGluAlaArgTyrGluGluIleLysLysGluValSerSerTyrIleLysLysValGlyTy	548 (183)
549	CAACCCAGCCGCGTTCCTTCGTGCCATTTCGGATGGCAGCCGACAAACATGTTGGAACCCCTACCAACATGCCCTGGTTCAGGG rAsnProAlaAlaValAlaPheValProIleSerGlyTrpHisGlyAspAsnMetLeuGluProSerThrAsnMetProTrpPheLysGl	638 (213)
639	ATGGGAAGTGGGACGCAAGGAGGGTAACGCTGACGGCAAGACCCCTGGTCGATGCCCTCGATGCCATCCTTCCCCAGCCCGTCCCACCGA yTrpGluValGlyArgLysGluGlyAsnAlaAspGlyLysThrLeuValAspAlaLeuAspAlaIleLeuProProAlaArgProThrAs	728 (243)
729	CAAGGCCCTGCGTCTGCCCTGACGGATGTGTACAAAATGGCGGATTTGGAACAGTACCCGTGGTGGTGTGGAGACTGGTGTCTGAA pLysAlaLeuArgLeuProLeuGlnAspValTyrLysIleGlyGlyIleGlyThrValProValGlyArgValGluThrGlyValLeuLy	818 (273)
819	GCCCGTACCGTGTGGTCTTCGCCCTGTAACATCACCCTGAGGTCAAGTCCGTGGAGATGCACCACGAGGCCCTGCAGGAGGCCGT sProGlyThrValValValPheAlaProAlaAsnIleThrThrGluValLysSerValGluMetHisHisGluAlaLeuGlnGluAlaVa	908 (303)
909	TCCCGAGACAACGTTGGCTCAACGTCAGAAGCGTCCGTGAAGGAGCTCGTGTGGCTACGTTGCCGGTACTCCAAGGCTAACCC lProGlyAspAsnValGlyPheAsnValLysAsnValSerValLysGluLeuArgArgGlyTyrValAlaGlyAspSerLysAlaAsnPr	998 (333)
999	CCCCAAGGAGCCGCGACTTACCGCCAGGTCATCGTGTGAACACCCCGGTGAGATTGCCAACGGCTACACCCAGTGTGGATTG oProLysGlyAlaAlaAspPheThrAlaGlnValIleValLeuAsnHisProGlyGlnIleAlaAsnGlyTyrThrProValLeuAspCy	1088 (363)
1089	CCACACCGCTCACATTGCTTGAAGTTCGCTGAGATCTTGGAGAAGGTCGACCGTCTCCGGCAAGACCACCGAGGAGAACCCCAAGTT sHisThrAlaHisIleAlaCysLysPheAlaGluIleLeuGluLysValAspArgArgSerGlyLysThrThrGluGluAsnProLysPh	1178 (393)
1179	CATCAAGTCTGGCGATGCTGCCATCGTCAACCTGGTGCCCTCAAGCCCTGTGCGTGGAGGCCCTCCAGGAGTTCACCCCTCTGGGTCG eIleLysSerGlyAspAlaAlaIleValAsnLeuValProSerLysProLeuCysValGluAlaPheGlnGluPheProProLeuGlyAr	1268 (423)
1269	CTTCGCTGTGCGTGACATGAGGCAGACCGTGGCTGTGCGTGCATTAAAGGCTGCACTTCAAGGATGCCTCCGGTGGCAAGGTCAACAA pPheAlaValArgAspMetArgGlnThrValAlaValGlyValIleLysAlaValAsnPheLysAspAlaSerGlyGlyLysValThrLy	1358 (453)
1359	GGCCCGCAGAGAAGCCACCAAGGGCAAGAAGTAGCTGGTTGCTTCCACTCAACAACAACAACAACACGAGTAGTAGCAGCAACAACAA sAlaAlaGluLysAlaThrLysGlyLysLysEnd	1448 (463)
1449	GCATATAACCAACATCATAATGCAGCCAACAACACCCTCAATAATACCAGCAACAGCAGCAGCGAACACAATAGTAGTATAACCAAC	1538
1539	ACCTGTCTCGCAGAGATGACCGATAAGATGATGTTTCAGCAGAAGCATAAAGTTAATTTCTTCCATCGAAAGGAGTTTCGACGGATACG	1628
1629	AATGCTAAATGCAGACGAGGCCCTTACTGGGAAATCGGTGGATCCCAAGGATAAGAGTGCACACTGGGAAAACACTTGCATTATGC	1718
1719	ATCCACTCCTCATCCACTTCCCGTCGATCTTTAGTTTACTAAATATGGTATGATGCACGCAGTGTGACTTCGTTTTATCATATCATATAT	1808
1809	AGGAATCCTCTGTAGCATTATGATATCGTTTAAATTAACCTTTATACTTTGATATGATCATTTATCTTACCCTACTTTTGCACACACT	1898
1899	ACTTTGTACACAAGAAAAGAACAGAATAGAAGCGATAAACTATATTTACAAAAAAAATAAAAAACCCCTATTTTTGTATTCTTTTGT	1988
1989	TACCACCCAGCCGTAAGAGGACTCTCTTTTTGGTTGTGCTCCCGATT	2041

Efl $\alpha$ 1 SEQUENCE. Strain, *Canton S*. Accession, X06869 (DROEF1AF1).

sequences (Fig. 12.1); this suggests that the two genes in *Drosophila* originated as an ancient duplication. The sequence similarity between the two genes outside of the coding regions is very limited; and there is great discrepancy in the number of introns (Walldorf et al. 1985; Hovemann et al. 1988).

### *Ef1 $\alpha$ 1*

#### **Gene Organization and Expression**

Open reading frame, 463 amino acids; expected mRNA size, 2,054 bases, in agreement with a single RNA band of 2 kb. The 5' end was defined by primer extension, cDNA sequencing and RNA sequencing. There is no apparent TATA box. The 3' end was obtained from S1 mapping and the sequence of a cDNA clone. There is one intron at -1,371/-20, in the leader (*Ef1 $\alpha$ 1* Sequence) (Walldorf et al. 1985; Hovemann et al. 1988).

#### *Developmental Pattern*

Expression is high throughout development, but it declines with age in adults (Webster 1985). It is also 5-10 times higher in adult females than in males (Walldorf et al. 1985; Hovemann et al. 1988).

#### *Promoter*

At -1,804 (373 bp upstream of the transcription initiation site) there is a sequence very similar to the HOMOL1 box of yeast. In yeast, this sequence occurs upstream of several genes for translation factors and ribosomal proteins (Walldorf et al. 1985; Hovemann et al. 1988).

### *Ef1 $\alpha$ 2*

#### **Gene Organization and Expression**

Open reading frame, 462 amino acids; expected mRNA size, 2,555/2,558 bases, in agreement with a single RNA band of 2.5 kb. The 5' end was defined by primer extension and by sequencing of a cDNA. There is no apparent TATA box. The 3' end was obtained from a cDNA sequence. There are four introns: two in the leader, at -1,811/-567, and -479/-30, -27 (this intron has two acceptor sites, and both are used), one in the Gly-275 codon and one after the Gln-343 codon (*Ef1 $\alpha$ 2* Sequence) (Walldorf et al. 1985; Hovemann et al. 1988).

*Efl $\alpha$ 2*

-2156 TAAGCGAATAGTGTGCACAATGTCTTTTGAATTAGTGGTGAATGTCATACTTTAGTGACAGTCCGTGAAAGTACTATATATTTTATC -2067

-2066 TGCAAAAGACTCAGTTTAAGAGAATATAAAATATCCATGAATGGTAGTAAAATTGTATTACTATTTTTATTTGGTACGTTTTATACTT -1977

-1976 AAGGGATGAAACTTTATTTAAGTCAAGAAATCCGCATAATGCAATAGGAAACCCAAGGCCCTGTGCATACATGGAATCCTGTGCCATCT -1887

----- -->-1833

-1886 CTAGGTCGGAATCAGTTCAGCTCCGTTACCTCAGCATCGTTGCTTTTTCCGGTCTTCCGTTTTGTGATTTCGAGGTAAGTGCACGCAGA -1797

-1796 GCTCCCGTTAAAATTGTGAAAATATTAATAGGCATTGATTAGTTGTGGAAATGTAAAAAGGGAAAAGTCCCAGAATCCCTACCCGTGCATT -1707

-1706 ATTAGGCGAATTTTCGGTTCGATTTCCAACCTAAAGAAAGTTCTAAAGTAAAGAAAGTTCCGAAAAGTGAGAGAGTGAAGTATTGCGC -1617

-1616 TGCCGGCCGGTCTTCTCATTCTTTTGCATAATAGCTGTGTAATCGATTGCAATTGGAAATGGTTTTCCAGCGACCTAAATGCAAG -1527

-1526 TAAATTAATAAAGTTGCATAGACTTTGCAATTTCAACATGGCGACCGGCTGCATGTGTGCGCGTTCGATTTGCCTGGATTGTACCCG -1437

-1436 TTTCTCCTTCCCGTTCTCAAGCGTTTTATCCCGAGTAGTTCTATTGGAATTCGAGGCAAAAAAAAAAATATCCGCGCATGATGGCAC -1347

-1346 ATGGTTAGCAGATTATTTCTTGCCTGCATCTCTGACGAAGTATTTGCATATCTTTCCCCCTTCATTCCCATGCTTCTTCCAATTT -1257

-1256 GCACTTCGATGCAAAACAAAGATTTAAAAATGGCATGCAGGAAAATCGGCAAGTAAACTGTCACTGGGGTAGAAAATAATCACAAACG -1167

-1166 CCCTGCAGTTCTCGCCGTCTCTTCCCTTCTTCTGTCATGACCAGCAAGTGCATGCGCCCGTTCGCCGTCCCTTTCTCTCCGCTCT -1077

-1076 CTCATCTCCCTCTACAGTTTTTACCCTTTGGAATCGCGGGATTTTCGCCGACGACCGCCACCGAATGCCGATGCTTTTGGCCATTTT -987

-986 CCTTTGGATTTTCTTCCACCGTCTGCGAAAGTTGCCAAATTTCCGCATTTGCACATTTGGCTTAATTGAAATCCGTTTGGGTGTGCGAT -897

-896 TTTTATTGGTTTTCCCACTAAAAACGCCGGCCGGCACATTTTCGCCATGCATGCGCACTTCCCGGCTTCCGACGAGGGTTTCTCTTC -807

-806 GGCTTAATCCTCTCCAGCCGAGGAGAGTGCATTTTCCAGTACGCACACTTCGGCTCCATTCTGTTCTGTCTGGGGCTCGTATTGATTT -717

-716 TTCGCCGGTGCATTCGGCAGAGGATATACACGGCAGTCTTAAACCAACAGACACTTGGCCCGTCTGGTCCGGCTGCAGAGTACGGA -627

-626 AGATCCGCATAGAGTTAAAAAAGTCCATTTTTATGACAACGATTTCTTCTAATTCTAGGATATAGCGTCCGCTGGGTTGTGATCAGT -537

-536 TTCTAAGTGCAGTTCGCGAGTAATAAGAACTCTAGAAAGTCTCGTAAAACAGGTGAGTTTTCTGCTGTAAATCTTGTGTCAT -447

-446 AGATTTGTGGGCAAAAATATTATGGGAATATGGGTGATTTTCTCAATCGTACACATTAGTGTCCATAAGAGTCCGTAAAAACATACATGT -357

-356 GTATTATATTTTTCTTATTATCAGTATAAGGCTTAATTTGAACTAATGGTAACTTTTCGCGTATTTCTGTTTACTCTTGAATT -267

-266 GTTTAAAATTCGATTTTTCGAAATATAAAAGTTCAACGGTTTTCCCTGTGTACGTTTGTCCGTCGGTATGAAGTGTGCTTTTGGTGTG -177

-176 CCACCACGATGACACGACCCACAGCATAACAGACGTCCTGTCGACCCACCATTAAGTTCAGACCCACATTGGCATGCTACCTCCCCG -87

(continued)

-86	AGTACGGAAACCACCCACTTTGCTCATCCGAATACCTGCATCCCTTCTGTCTCCCAGCAGCTCTAAAAATAGCTTAATCTGCAAGGATG	3
	 	Met (1)
4	GGCAAGGAGAAGATCCATATTAACATTGTGGTCATTGGCCATGTGGACTCCGGCAAGTCGACGACCACCGGCCACTTGATCTACAAATGC GlyLysGluLysIleHisIleAsnIleValValIleGlyHisValAspSerGlyLysSerThrThrThrGlyHisLeuIleTyrLysCys	93 (31)
94	GGCGGCATCGACAAGCGTACGATTGAGAAGTTCGAGAAGGAGGCCAGGAAATGGGAAAAGGCTCCTTTAAGTACGCTTGGGTACTGGAC GlyGlyIleAspLysArgThrIleGluLysPheGluLysGluAlaGlnGluMetGlyLysGlySerPheLysTyrAlaTrpValLeuAsp	183 (61)
184	AAGCTGAAGGCAGAGCGGGAGCGGGGCATCACCATCGACATTGCCCTATGGAAGTTCGAGACGTCCAAGTACTATGTGACCATCATCGAT LysLeuLysAlaGluArgGluArgGlyIleThrIleAspIleAlaLeuTrpLysPheGluThrSerLysTyrTyrValThrIleIleAsp	273 (91)
274	GCCCTGGTCACAGGGATTTTCATCAAGAACATGATTACCGGTACCTCTCAGGCCGATTTGTGCGGTGCTGATCGACGCCGCCGGAATGGA AlaProGlyHisArgAspPheIleLysAsnMetIleThrGlyThrSerGlnAlaAspCysAlaValLeuIleAspAlaAlaGlyThrGly	363 (121)
364	GAGTTCGAGGCCGGGATCTCGAAGAACGGCCAGACCCGCGAGCACGCCCTTCTGGCATTACGCTGGGCGTGAAGCAGCTTATTGTGGGC GluPheGluAlaGlyIleSerLysAsnGlyGlnThrArgGluHisAlaLeuLeuAlaPheThrLeuGlyValLysGlnLeuIleValGly	453 (151)
454	GTCACAAAGATGGACTCCACTGAGCCGCCGTACACGCGAGGCCCGCTACGAGGAGATCAAGAAGGAGGTGCTCTCGTACATCAAGAAGATC ValAsnLysMetAspSerThrGluProProTyrSerGluAlaArgTyrGluGluIleLysLysGluValSerSerTyrIleLysLysIle	543 (181)
544	GGCTACAATCCGGCTCGTGGCCCTTCGTGCCATCTCCGGATGGCACGGCGACAATATGCTGGAGCCGTCGAGAAAGATGCCCTGGTTC GlyTyrAsnProAlaSerValAlaPheValProIleSerGlyTrpHisGlyAspAsnMetLeuGluProSerGluLysMetProTrpPhe	633 (211)
634	AAGGGATGGTCCGTGGAGCGCAAGGAAGGCAAGGCGAGGGCAAGTGCTTGATCGACGCGCTGGACGCGATCTTCCACCCACGCGTCCC LysGlyTrpSerValGluArgLysGluGlyLysAlaGluGlyLysCysLeuIleAspAlaLeuAspAlaIleLeuProProGlnArgPro	723 (241)
724	ACCGACAAGCCGCTGCGCTGCCGCTCCAGGACGCTACAAGATCGGAGGCATCGGAACCGTACCAGTAGGTCGTGGAGACTGGTCTC ThrAspLysProLeuArgLeuProLeuGlnAspValTyrLysIleGlyGlyIleGlyThrValProValGlyArgValGluThrGlyLeu	813 (271)
814	CTCAAGCCAGGTAAGGCTCCGGGTGATGAGGTCCGGGTGGGCCCTCTTTCTCTTTGGGCACCTTCATACATGTATTCTGCAAAATTTG LeuLysProG	903 (275)
904	GGTCGACAGTGGGTGGCATCCAACAGCCACCGCTCCAAAGCGGAGCCGCAACGAAGTCTTGCAGTGTATGCATTATTGAGCGAACGT	993
994	CTTCGTCGAGAGCGAGACCCCTCACCTCATGCACTTGGTGAATTTCTCACTCCGAAGAGCTTCCATTTTCAACATGAAAGTGAAGGCCA	1083
1084	TAAAAATAAAATAACCTAGCTAACATATTAATATATGTAGAGCTATTGATTCAAATAAAAAATAAATGGAGTAGTTCGAATAATATCG	1173
1174	CTCCACGTTTCTCTCTGTATGCACCCACCCCATCAAATGTCTACACATAACGTCGGGATATGTAACCTCGTTTCGGTCGCTTCGTT	1263
1264	TCCGGTTTCGTTTCAGGCATGGTCGTCACATTTGCGCCGGTCAACCTGGTCACCGAAGTAAAGTCTGGAGATGCACCACGAGGCTCTC lYmetValValAsnPheAlaProValAsnLeuValThrGluValLysSerValGluMetHisHisGluAlaLeu	1353 (299)
1354	ACCGAAGCCATGCCGCGCAACAAGTTGGCTTCAACGTGAAGAACGTGTCCTGTAAGGAGCTCCGTCGTGGCTATGTGGCCGCGGATCC ThrGluAlaMetProGlyAspAsnValGlyPheAsnValLysAsnValSerValLysGluLeuArgArgGlyTyrValAlaGlyAspSer	1443 (329)
1444	AAGAACAATCCTCTAGGGGAGCAGCCGACTTACCCTCAGGTAGGTAACAAGATGAGAAATCTTTGATAGTTGAACACTCATCTTTGT LysAsnAsnProProArgGlyAlaAlaAspPheThrAlaGln	1533 (343)
1534	TTGGTTTTTTTTTTCTTTTTGCCACAGGTGATTGTGCTCAACCATCCGGCCAGATCGCCAATGGGTACACTCCCGCTTGGATTGC ValIleValLeuAsnHisProGlyGlnIleAlaAsnGlyTyrThrProValLeuAspCys	1623 (363)

1624	CACACGGCGCACATTGCCTGCAAGTTTTCCGAGATCAAGGAGAAGTACGACCGCGTACGGGGCGGAACCACCGAAGACGGGCGGAAGGCT HisThrAlaHisIleAlaCysLysPheSerGluIleLysGluLysTyrAspArgArgThrGlyGlyThrThrGluAspGlyProLysAla	1713 (393)
1714	ATCAAGTCCGGGATGCGGCCATCATTGTGCTGGTCCCAGCAAGCCGTTGTGCGTAGAGAGCTTCCAGGAGTCCACCAGCTGGGACGG IleLysSerGlyAspAlaAlaIleIleValLeuValProSerLysProLeuCysValGluSerPheGlnGluPheProProLeuGlyArg	1803 (423)
1804	TTCGCTGTGCGGCACATGAGGCAGACCGTGGCCGTGGGCGTCATCAAGTCGGTGAACCTTAAAGAGACGACCTCGGGCAAGGTGACAAAA PheAlaValArgAspMetArgGlnThrValAlaValGlyValIleLysSerValAsnPheLysGluThrThrSerGlyLysValThrLys	1893 (453)
1894	GCCGCTGAGAAGGCACAGAAGAAGAAATAACTAGGGTACCAGCAGAACAACGTCATCACTCGAACCCCAACAACAACAAAAACAGACGGCT AlaAlaGluLysAlaGlnLysLysLysEnd	1983 (462)
1984	AGAGCAACAGCAGCAACAACACACAACAATAACACATGTCAAATTATAATACCCACTCGACGATCAAATTCACACCTTGACTCCATG	2073
2074	GCAAGAGAGACCAATTACTACTATTACTAGCTGCTGGGAGAAGCGGCAGATATAACCGAAATCGAGCAGATTATACCCATATAATA	2163
2164	ACCACACGTACGATTAGCGAGGAGAGGAGCATCAGGTGCAGCGAGGATGCGAAGGAGGAGCCCTCCAGCCTCGCCGGGTCGGTTTTGGT	2253
2254	CGCCTTCGCGGTGGTCTACTGCAGCTATCTGAACATGTATCGTCACCGAAGTCCTTCGTAGGAAACCACCCGCTAGCCACTCCGC	2343
2344	AGAGTGGATAGGGCCCTCCGAGCACTGCTGTAGCCCGCCCTTCGATATATACTCATCTCTAAACTAACCTTACACTTGATTAGCAGC	2433
2434	CACACATCCGGTCGCATCCACCTGTTTCGAATGGATTTTAAACACTTTTTATACTTTTGATAAGTCAAGTCGGAGGCATTGATTTAAA	2523
2524	TCTATTGAAATATGTAATTTCCGAATTTAGTTTTAAACCAGCTCCGCGCTCCCAAAAATCCCCGAACCGAAAAGACTACATTGCGGATG	2613
2614	AATTCAAAATTTCTCTTGAACCAAAAAAACAATGCTTAAAGATATTACAAAAAGAAATCAACATTACACACATAATCATGCGGTT	2703
2704	TTTGAAAACATTATAAATGTTAATCGAGCCTCATTGTCATTTGCATATTACATAATATACGTTAGCCACATGTCATCTCATTGCCATA	2793
2794	ATAACCTGCATCTGCATATTATACAGTTAATCTCACACTCTGAATTTATACAAACCGAAGACAATTGTAACCGACACCAGAACAATTC	2883
2884	TTGGATACAGAACATGTTGGCTTGATAAAAAGATCTTTTAAATGATGAGAAAAATAAAGGAAGCTTAACCGTAAAAATACCACACACGAACG	2973
2974	CCTTTTAATTGAAAAATACTTGAATATCTATGAAGAAAATGAATTC 3019	

*Ef1 $\alpha$ 2* SEQUENCE. Strain, *Canton S*. Accession, X06870 (DROEF1AF2).

### Developmental Pattern

The level of expression is lower than that of *Ef1 $\alpha$ 1*, and it peaks during the pupal stages (Walldorf et al. 1985; Hovemann et al. 1988).

### Promoter

There are no obvious similarities with the promoter region of *Ef1 $\alpha$ 1* (Walldorf et al. 1985; Hovemann et al. 1988).

## References

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- Walldorf, U., Hovemann, B. and Bautz, E. K. F. (1985). F1 and F2: Two similar genes regulated differently during development of *Drosophila melanogaster*. *Proc. Natl Acad. Sci. (USA)* **82**:5795–5799.
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# 13

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## *even-skipped: eve*

**Chromosomal Location:**  
2R, 46C3-11

**Map Position:**  
2-58

### **Product**

A DNA-binding regulatory protein of the homeodomain type important in establishing the segmentation pattern of the embryo.

### *Structure*

The homeodomain occurs toward the N-terminus (Val-70 to Arg-129). The Gln residue in position 9 of the third homeodomain helix (*eve* Sequence, H3) makes EVE a homeoprotein of the *Antennapedia* (*Antp*) class (Hanes and Brent 1991). Another noteworthy sequence feature is the Ala-rich segment spanning Ala-146 to Ala-179. Similar Ala repeats have been found in the genes *caudal*, *engrailed* (*en*), *Ultrabithorax* (*Ubx*) and *Krüppel* (*Kr*); in the *Kr* product the Ala-rich region seems to be associated with the repressor function of that protein (Macdonald et al. 1986; Hoey et al. 1988; Biggin and Tjian 1989; Licht et al. 1990; Harrison 1991).

### *Function*

Binding sites for EVE have been found in the region of the *eve* promoter proximal to the site of transcription initiation and in the *en* promoter. The sequences of the binding sites are quite different in the two promoters. The consensus for the EVE binding site of the *en* promoter is TCAATTAAAT; this is similar to binding sites of other homeodomain proteins of the *Antp* class and was designated as class I (Levine and Hoey 1988; Hanes and Brent 1991). In contrast, the EVE binding sites near *eve* have in common the sequence TCAGCACCG and were designated as class II (Hoey and Levine 1988). EVE binding sites with segments combining features of both class I and class II sequences also exist in the *eve* autoregulatory region, 5.4–5.2 kb upstream of





(--> -->)-98/-93

-112 GGGTGGCTGAGAGCAGCACACTCGAGCTGTGACCGCCGCACAGTCAACAACTAAGCTTCGTTAATATCCTTTGAATAAGCAACTTT -23  
 ///>1

-22 GAATCACAAGACGCATACCAACATGCACGGATACCGAACCTACAACATGGAGAGCCACCATGCCATCACGACGCCAGTCCCGTGGACC 67  
 MetHisGlyTyrArgThrTyrAsnMetGlySerHisHisAlaHisHisAspAlaSerProValAspG (23)

68 AGAAGCCCTGGTGTGGACCTCTTGGCCACCCAGTACGGCAAGCCCGACACCCGCTCCCTCGCCAAATGGTAAGTTAAAGATAAAG 157  
 lNlysProLeuValValAspLeuLeuAlaThrGlnTyrGlyLysProGlnThrProProProSerProAsnG (47)

158 CCGAGCAAACGTGACGAGTTACTTACACCAATCTTCTCCTGTCCAAAACAGAATGCCTATCCAGTCCGGATAACTCCTTGAACGGCAG 247  
 lCysLeuSerSerProAspAsnSerLeuAsnGlySe (59)

248 CCGCGGCTCGGAGATTCCCGCCGACCCGTCGGTACGCCGCTATCGCACCGCTTACCCCGTGACCAGCTGGGTCGCTTGGAGAAGGAGTT 337  
 rArgGlySerGluIleProAlaAspProSerValArgArgTyrArgThrAlaPheThrArgAspGlnLeuGlyArgLeuGluLysGluPh (89)  
 | \* \* \* -----\*-----\*

338 CTACAAGGAGAACTACGTGTCCCGTCCCGTCCGCTGCGAACTGGCCGCCAGCTGAACCTCCCGGAGAGCAGCATCAAGGTGTGGTTCCA 427  
 eTyrLysGluAsnTyrValSerArgProArgArgCysGluLeuAlaAlaGlnLeuAsnLeuProGluSerThrIleLysValTrpPheG (119)  
 ---H1 \* -----\*-----H2 \* -----\*-----\*-----\*

428 GAACCGCCGCATGAAGGACAAGCGTCAGAGGATCGCCGTCGCCTGACCCTACGCAGCCGCTACTCCGATCCCGCCTTCGCCGCCCTCCAT 517  
 nAsnArgArgMetLysAspLysArgGlnArgIleAlaValAlaTrpProTyrAlaAlaValTyrSerAspProAlaPheAlaAlaSerI (149)  
 -\*--\*H3\* \* \* |HOMEDDOMAIN

518 CCTCCAGGCGCCGCAACAGCGTGGCGTACCCCTATCCGCCCTACGCCCCCGCTGCTGCCGCGCTGCTGCCGCGCCGCTGCGGTGGC 607  
 eLeuGlnAlaAlaAlaAsnSerValGlyMetProTyrProProTyrAlaProAlaAlaAlaAlaAlaAlaAlaAlaAlaAlaAlaAla (179)

608 CACCAATCCGATGATGCCACCGGAATGCCCCGATGGGATGCCCCAGATGCCACAATGCAGATGCCGGACACTCGGGACATGCCGG 697  
 aThrAsnProMetMetAlaThrGlyMetProProMetGlyMetProGlnMetProThrMetGlnMetProGlyHisSerGlyHisAlaG (209)

698 CCATCCCTGCCCCACGGACAGTACCGCTACACGCCCTACCACATCCCCGCCGCGCCGGCCGCCACATCCCGCTCGGTCTCATATGCA 787  
 yHisProSerProTyrGlyGlnTyrArgTyrThrProTyrHisIleProAlaArgProAlaProProHisProAlaGlyProHisMetHi (239)

788 TCATCCGCACATGATGGGATCCAGCGCCACGGGATGTCGTACTCCGCGGTGCCGCCGCTTTTGGGCGCTCTGCCCTCCGCCACCTG 877  
 sHisProHisMetMetGlySerSerAlaThrGlySerSerTyrSerAlaGlyAlaAlaGlyLeuLeuGlyAlaLeuProSerAlaThrCy (269)

878 CTATACCGGACTGGGTGTGGGTGTGCCAAGACCCAGACGCCCGCTGGATCTGCAGTCGTCGTCATCGCCGACTCCTCCACGCTGTC 967  
 sTyrThrGlyLeuGlyValGlyValProLysThrGlnThrProProLeuAspLeuGlnSerSerSerSerProHisSerSerThrLeuSe (299)

968 CGTCTCGCCAGTGGGATCCGATCACGCCAAGGTGTTCGACCGCAGTCCAGTGGCTCAATCCGCTCCATCAGTTCCTGCTCCCGCTCCACT 1057  
 rValSerProValGlySerAspHisAlaLysValPheAspArgSerProValAlaGlnSerAlaProSerValProAlaProAlaProLe (329)

1058 GACCACCACAGCCCGCTGCCCGCTCCCGGCTCCTGATGCCAGTGCCAAGCGGCTGCCTCCGACATGTCCGCCGCGCCGACGACAAC 1147  
 uThrThrThrSerProLeuProAlaProGlyLeuLeuMetProSerAlaLysArgProAlaSerAspMetSerProProProThrThrTh (359)

1148 TGTGATTGCGGAGCCCAAGCCGAAGCTCTTCAAGCCCTACAAGACTGAGGCGTAAGCCCGCATCCACACACTCTCGCCCCCCCCC 1237  
 rValIleAlaGluProLysProLysLeuPheLysProTyrLysThrGluAlaEnd (376)

1238 CTGCTCCCCAAAGATTGTACAAACTAGTCTTAGTCAGCCTCATCTATTTATTTCCGAAGATTGTACAGATTGTAGAGTAGCTAATTGTA 1327

1328 GTCATAATTAAGGCGCAAATCAAATTAAGAAATAAATGCGAAAATAACATTGAAAATATACGACACACTGTATTATTGCACTACCT 1417  
 ----- (A)<sub>n</sub>

1418 GGTACC 1423

the transcription initiation site (see *Promoter*). The binding of EVE to these sites is required for autoregulatory function as shown by germline transformation experiments (Jiang et al. 1991).

EVE is important in establishing segmentation in the early embryo. The anterior borders of EVE stripes define the anterior border of the corresponding *en* stripes at the anterior borders of odd-numbered parasegments. In the trailing edge of the stripes, EVE represses *fushi tarazu* (*ftz*) expression, thus defining the anterior border of FTZ stripes in even-numbered parasegments (Lawrence et al. 1987; Ish-Horowicz et al. 1989). EVE seems to act directly on *eve*, *ftz*, *en* and *wingless* (Macdonald et al. 1986; Harding et al. 1986; Frasch et al. 1988).

### *Tissue Distribution*

As detected by antibody staining, EVE protein is localized in nuclei; and it peaks briefly during the cellular blastoderm and gastrulation stages of embryonic development. EVE is first detectable in division-cycle-12 nuclei throughout the embryo; by cycle 13, staining disappears from the poles and becomes restricted to a band that extends from 70% to 20% egg length. Soon afterwards, the striped pattern along the antero-posterior axis of the embryo develops (Appendix, Fig. A.3). After germ band elongation, EVE protein persists only in neurogenic cells. The developmental pattern of EVE protein follows closely the distribution of *eve* transcript (Frasch et al. 1987; see below).

### *Mutant Phenotypes*

*eve* is one of the pair-rule genes, hypomorphic *eve* mutants are embryonic lethals having only half the correct number of segments. The missing elements correspond to the posterior region of T2 and the anterior of T3, the posterior of A1 and anterior of A2, etc.; i.e., every other segment boundary and neighboring areas (corresponding to odd-numbered parasegments) are missing. In amorphic mutants, the bands of ventral denticles are replaced with a uniform "lawn" of denticles, so that all obvious trace of segmentation is lost (Nüsslein-Volhard and Wieschaus 1980; Nüsslein-Volhard et al. 1985; Akam 1987).

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(*previous pages*) *eve* SEQUENCE. The segment from -202 to 1,423 has accession number M14767 (DROEVE). The segment -498 to -203 is from Read et al. (1990). The segment -1,601 to -931, the stripe 2 element, is from Stanojevic et al. (1991). The segment -5,400 to -5,200, the autoregulatory region, is from Jiang et al. (1991). GAGA (////>) and TCCT (====>), cores of the GAGA and TKK regulatory protein-binding sites, are underlined and numbered; dashes (---) underline EVE, BCD, HB, GT and KR binding sites. The limits of the homeodomain are marked by vertical lines under the sequence, asterisks indicate conserved amino acids, and dashes underline the presumptive helices.

## Gene Organization and Expression

Open reading frame 376 amino acids; expected mRNA length, 1,416/1,421 bases. There is a small uncertainty about the position of the 5' end of the transcript: RNase protection experiments localized the 5' end at position -93 (Macdonald et al. 1986) while S1 mapping and primer extension indicate it is at position -98 (Frasch et al. 1987). RNase protection was used to define the 3' end. There is an intron within the Glu-47 codon (*eve* Sequence) (Macdonald et al. 1986; Frasch et al. 1987).

### *Developmental Pattern*

This section was excerpted from Macdonald et al. (1986) and Frasch et al. (1988). The level of *eve* transcript is very low in 0–2 h embryos, it peaks in 2–4 h embryos and then persists in ever-decreasing amounts until the first larval instar. Early in nuclear cycle 13 (syncytial blastoderm), the transcript is localized in the peripheral cortical region of the embryo forming a broad band, as indicated by *in situ* hybridization. Over the next 30 min, this band intensifies and expands until it covers most of the future segmented portion of the embryo (20–70% egg length). As it expands, the band becomes subdivided into two, then four and then seven stripes to produce the “zebra” pattern of expression that is characteristic of pair-rule genes.

By the middle of nuclear cycle 14A (late syncytial blastoderm), expression is localized in seven stripes, six of them being five- or six-nuclei wide while the seventh posterior-most stripe is 6–8 nuclei wide; the stripes are separated by 2–3 nuclei wide spacers. Each *eve* stripe is asymmetric, with the anterior cells showing the highest level of expression; this is the first sign of segment polarity. Some transcript is also detectable in the yolk nuclei occupying the central region of the embryo. During blastoderm cellularization, the stripes narrow to a width of 2–3 nuclei.

At the beginning of gastrulation, the most anterior *eve* stripe is positioned immediately anterior to the cephalic fold. The *ftz* transcripts, which also display a seven-stripe pattern, are shifted posteriorly relative to *eve* such that the two genes are expressed in alternating parasegments. As gastrulation proceeds, seven minor *eve* stripes appear between the major ones. A similar pattern of alternating major and minor stripes is also exhibited by *en*; however *en* major stripes occur in even-numbered parasegments while *eve* major stripes are localized in odd-numbered parasegments. At this stage, *eve* expression seems to be localized to the anterior region of each of the 14 parasegments.

During germ band elongation, the segmented expression of *eve* disappears, and a new site of accumulation appears posterior to the last major stripe. This new site corresponds to cells of the proctodeal primordium, cells that also express *en*, *hairy* and *paired*.

After gastrulation, *eve* expression can be detected only in small clusters of neural ganglion mother cells in each parasegment; *eve* expression continues in the nerve cord until late in embryogenesis. The rapid disappearance of *eve*

transcript and protein suggest that these molecules have very short half-lives. In the grasshopper, expression of the *eve* cognate gene occurs in neuroblasts that occupy equivalent positions to those that express *eve* in *Drosophila*; the “zebra” pattern of expression of early embryos, however, is absent. This suggests that the role of *eve* in short germ band embryos is restricted to neurogenesis, and the pair rule function was acquired secondarily during the evolution of higher insects (Patel et al. 1992).

In null mutations, the sites of major and minor stripes, i.e., odd-numbered parasegments and the anterior regions of even-numbered parasegments, are missing. Thus, only the posterior regions of even parasegments are left; they correspond to the denticle belts of T2, A1, A3, etc., which, becoming fused without any naked cuticle to separate them, form the denticle “lawn” mentioned above. In weaker alleles, only the sites of major stripes, i.e., the odd-numbered parasegments, are missing (Nüsslein-Volhard et al. 1985; Macdonald et al. 1986).

### *Promoter*

Regulation of the seven major stripes was investigated in some detail. The production of the striped pattern seems to occur in two phases, an early phase when seven regions of expression are established and a late phase when these regions become narrower and expression intensifies such that stripes become more sharply defined. The early phase is regulated by the gap gene products and the maternal morphogen BCD (product of *bicoid*), all of which are expressed in broad, non-periodic and partly overlapping areas (Appendix, Fig. A.2). The late phase is controlled by the pair-rule gene products, EVE included, which are distributed periodically along the antero-posterior axis of the embryo (Goto et al. 1989; Harding et al. 1989 and references therein).

Early expression in stripes 1, 4, 5, and 6 seems to require unidentified *cis*-acting elements located more than 8.0 kb upstream of the transcription initiation site. An element located between 3.8 and 3.0 kb upstream of the transcription initiation site is required for early expression in stripe 3; and elements between 1.65 and 1.15 kb upstream of the transcription initiation site are required for expression in stripes 2 and 7 (Goto et al. 1989; Harding et al. 1989).

The late or autoregulatory function is controlled by a segment between 5.9 and 5.2 kb upstream of the transcription initiation site. A construction in which the 5.9–5.2 kb segment is linked to a reporter gene is expressed in all seven stripes only if the host organism is wild-type for all pair-rule genes. In the absence of the stripe-specific, early control elements, however, expression, is much weaker (Goto et al. 1989; Harding et al. 1989).

In the segment regulating transcription in stripe 2, there are the following protein binding sites: five for BCD, three for the *hunchback* (*hb*) product (HB), three for the *giant* (*gt*) product (GT) and six for the *Krüppel* (*Kr*) product (KR) (*eve* Sequence). In the stripe 3 promoter segment, there are 18 HB binding sites. The BCD binding sites have the consensus GGGATTAGA; KR binding sites

are derivatives of the decamer AACGGGTAA and the HB binding sites have the consensus G/CA/CATAAAAAA (Stanojevic et al. 1989; Small et al. 1991).

When KR binding sites are inserted into the promoter region of a reporter gene, the expression of the reporter is repressed by KR (Licht et al. 1990). Studies on cultured cells transfected with one or more of the putative regulatory genes (*bcd*, *hb*, *gt* or *Kr*) under the control of the *Actin 5C* promoter and co-transfected with a reporter gene under the control of the stripe 2 regulatory segment showed that BCD and HB are activators and that GT and KR are repressors of stripe 2 transcription. These results suggest that relatively high levels of BCD and HB in a region that includes the second stripe stimulate *eve* transcription; posterior to stripe 2, the band of KR accumulation represses transcription, thus defining the posterior boundary of stripe 2 (Appendix, Fig. A.2 and Fig. A.3); anteriorly, a region of GT accumulation defines the anterior border. The interactions between the regulatory factors seem to occur through direct competition for binding sites. Eight binding sites in the stripe 2 segment are sufficient for proper regulation, and these sites are arranged in two clusters: the proximal cluster includes a BCD and an HB site, overlapped respectively by KR and GT binding sites; the distal cluster includes two BCD sites also overlapped by a KR and a GT site (*eve* Sequence) (Small et al. 1991). This view of the regulation of stripe 2 expression is supported by studies on transgenic organisms carrying various binding-site mutations (Stanojevic et al. 1991).

The results described above are consistent with genetic studies indicating that KR is a repressor of *eve* and with the finding that establishment of the "zebra" pattern (early phase) requires the function of gap genes *hb*, *Kr*, *Knirps* and *tailless* (Frasch and Levine 1987).

Maintenance and refinement of the striped pattern (late phase) is dependent on the pair-rule genes *eve*, *hairy* and *runt* (Frasch and Levine 1987) and the autoregulatory region, 5.9–5.2 kb upstream of the transcription initiation site. EVE, in cooperation with the general transcription factor GAGA (and possibly with a zinc-finger protein coded by the gene *tramtrack* [*ttk*]), seems to interact directly with a 200-bp segment in the autoregulatory region (*eve* Sequence) (Jiang et al. 1991; Read and Manley 1992). The GAGA binding site has a sequence related to GAGAG (Biggin and Tjian 1988) while the putative binding site for the *ttk* product includes the octamer GGTCCTGC (see below) (Jiang et al. 1991).

Two other clusters of EVE binding sites in the *eve* promoter are necessary for transcription, one in a region 3.1–2.9 kb upstream of the transcription initiation site and the other in a proximal region, 295–44 bp upstream of the transcription initiation site (e4 and e5 in the *eve* Sequence). The proximal ones belong to the class II of EVE binding sites as already discussed (Hoey and Levine 1988). Sites e4 and e5 also bind the product of *prd*; e5 comprises two sections and can bind two PRD molecules, one through the homeodomain and the other through the paired domain (Treisman et al. 1991).

*In vitro* assays in the presence of proteins from embryonic nuclear extracts showed that sequences between 179 and 72 bp upstream of the transcription initiation site are required for transcription. Protein-binding assays identified

12 binding sites in the segment between 574 bp upstream of the transcription initiation site and 175 bp downstream of the transcription initiation site. Eleven of those sites are between 390 and 0 bp upstream of the transcription initiation site and one is 45 bp downstream. Eight of the eleven upstream sites probably bind the GAGA factor (1–6, 9, and 10 in the *eve* Sequence) while the remaining three (7, 8, and 11) seem to bind a different factor and share sequences related to GGTCCTGC. The GAGA-binding protein is relatively constant through development; but the TCCT-binding factor, the product of *ttk*, is apparently restricted to developmental stages when *eve* is active (Read et al. 1990; Read and Manley 1992).

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# 14

## *fushi-tarazu: ftz*

**Chromosomal Location:**  
3R, 84B1-2

**Map Position:**  
3-47.5

### **Product**

A DNA-binding regulatory protein of the homeodomain type important in establishing the segmentation pattern of the embryo.

### *Structure*

The homeodomain occurs between Ser-257 and Arg-316 (Laughon and Scott 1984). Like *Antennapedia's* (*Antp*) homeodomain (ANTP-HD), the *ftz* homeodomain (FTZ-HD) has a Gln in position 9 of helix 3 (*ftz* Sequence, H3); this gives FTZ a binding specificity that distinguishes it from the *bicoid* (*bcd*) and *paired* (*prd*) products, in which Lys and Ser respectively occupy position 9 (Treisman et al. 1989). FTZ occurs as a family of phosphorylated isoforms; 19 differently charged forms were detected. Given the numerous Ser and Thr residues available for modification, the total number of specific isoforms could be much larger. Some isoforms are specific to certain embryonic stages (Krause and Gehring 1989).

### *Function*

The binding of FTZ-HD to the *engrailed* (*en*) promoter binding site bs2 is similar to the binding of ANTP-HD to this site ( $K_D = 6-8 \times 10^{-10}$  M); in particular, the Gln in position 9 of H3 interacts with CC in the bs2 sequence GCCATTAGA (Percival-Smith et al. 1990). *In vitro*, FTZ binds as a monomer to 10-12 bp binding sites. Six of those base pairs are critical, the optimal sequence being C/TAATTA with an equilibrium dissociation constant of  $2.5 \times 10^{-11}$  (Florence et al. 1991).

FTZ is required for embryonic expression of the *Antp* proximal promoter

## ftz

-1020 AAGCTTTATATTCTCAACAATATTATGCTATTAATAATTGCTGGTTTTCTGCTGTTATAGAATCATTTTTAAAAGTATAACGTAACAAAAA -931

-930 TAAAAAAAAC TAGTATTCATTTGAAAAATCAGCGGCATATAATTTATATCATATTTTTAAAATTTTCGGCAAAGGATGTTGCATAAAG -841

-840 TTTTACTGTTTACTAGTCATTTTGGAAAGTCGTTTTGTTGGTTTTTAGGCAAATACCGGCACAGGAGTGAGTTTGGGAATCGGGAGTTG -751

-750 CGCACTTGCTTGCCACGAGGGCAAACAAAAGCGCAAACACGCGACCTCGGCCACGCGTATTCTGATCCAGGGATCGGACGTAATG -661

-660 TTATCCTTTGGCCGCCAGTGCCACGAAATAAATTCGGAGGGAAAGGGCATCGGGTTCGGAACTGCGACCGAGTCTTCGGTGT -571

-570 GCGCGCTGGCAAAATCCAGAGAAATTTTAGGGAACCATAAACGGCCGGGGAAAAAGCCTCTGCCCGAAGGAACGTTTTTCAGCAACA -481  
-----ae2a

-480 GTTTACAGTTTTTATGCTTTTATGATTATGCAATTAGAGGAGATCGGCTGAGAGTCGCGCCCTCTCGCTCTGCGCACCTCATAGGTAGG -391  
-----ae2b -----ae3

-390 CACCTCATGGCCGTAATTACTGCAGCACCGTCTCAAGGTCGCCGAGTAGGAGAAGCGCGGGCGGATAAATCGCGATGATAATGGGCGC -301  
-----re1 -----\*\*-----de1=ftz-f1 -----re2

-300 GATGGGTAGGTAATAAGCCGCGCAGCAGGTAGGCCACCGTACGGATAAAGTGCCAGGACCTCGGATAACTTCCCTCTCGTGCTGCAA -211  
-----\*\*-----re3=f2 I -----  
-----\*----- ftz-f3

-210 GGACATTCGCGGGAGGGGTGGCTGCGAACAGCAGCCGCAAAGTGTCATGCGCAGGGATATTTATGCGCTATAACGGCGAGCGTGTGCC -121  
\*\*-----de2=ftz-f2 II

-----69

-120 GAGGGCTCTCTGATTTTGCATATATGCAAGGATCGCCGAGGACCAGCTCATTGCAAACTCACCAGCGTTGCGTGACATCGCAGAGT -31  
-----

-30 TAGAGAAGAAATCTAGCAATACACATCCGATATGGCTACCACAAACAGCCAGGCCACTACAGCTACGCCACAACATGAACATGTACAA 59  
MetAlaThrThrAsnSerGlnSerHisTyrSerTyrAlaAspAsnMetAsnMetTyrAs (20)

60 CATGTATCACCCACAGCCTGCCGCCACCTACTACGATAATTCAGGCAGCAATGCCTACTATCAGAACACCTCCAATTACCACAGCTA 149  
nMetTyrHisProHisSerLeuProProThrTyrTyrAspAsnSerGlySerAsnAlaTyrTyrGlnAsnThrSerAsnTyrHisSerTy (50)

150 TCAGGGTACTATCCCAGGAGGTTACTCGGAGAGCTGCTACTACTACAACAATCAGGAGCAGGTGACCACCCAGAGCTGTACCGCCGT 239  
rGlnGlyTyrTyrProGlnGluSerTyrSerGluSerCysTyrTyrTyrAsnAsnGlnGluGlnValThrThrGlnThrValProProVa (80)

240 GCAACCCACACCCCGCCGCCAAGGCCACCAAGCGCAAGGCCGAAGATGATGCTGTCCATCATCGCCGCCGTGGAGGAGCGACCCAG 329  
lGlnProThrThrProProProLysAlaThrLysArgLysAlaGluAspAspAlaAlaSerIleIleAlaAlaValGluGluArgProSe (110)

330 CACACTGAGGGCTCTGCTCACAATCCCGTGAAGAAGCTGAAGTACACCCCGACTATTTCTACACAACCGTCGAGCAGGTGAAGAAGGC 419  
rThrLeuArgAlaLeuLeuThrAsnProValLysLysLeuLysTyrThrProAspTyrPheTyrThrThrValGluGlnValLysLysAl (140)

420 TCCGCGCTAACCAAGGTACCCGCGAGCCCGCTCCAGCTACGACCAAGAGTACGTGACTGTGCCACGCCAGCCAGCGCCTCCGAGGA 509  
aProAlaValThrThrLysValThrAlaSerProAlaProSerTyrAspGlnGluTyrValThrValProThrProSerAlaSerGluAs (170)

510 TGTCGACTACTTGAGCTCTACTCGCCAGTCGACAGCAGAAGCTGAAGAATGGCGACTTTGCCACCCCTCCGCCAACCCAGCCAC 599  
pValAspTyrLeuAspValTyrSerProGlnSerGlnThrGlnLysLeuLysAsnGlyAspPheAlaThrProProProThrThrProTh (200)

(continued)

. T=Ua12 . T=Ua11 .

600 CTCTCTGCCGCCCTCGAAGGCATCAGCACGCCACCCCAATCGCCGGGGAGAAATCGTGTGACGTGTCAGCCAGGAGATCAATCATCG 689  
 rSerLeuProProLeuGluGlyIleSerThrProProGlnSerProGlyGluLysSerSerSerAlaValSerGlnGluIleAsnHisAr (230  
 Leu Leu

690 AATTGTGACAGCCCCGAATGGAGCCGGCGATTTC AATTGGTCGCACATCGAGGAGACTTTGGCATCAGGTAGGCATCACACAGATTAAC 779  
 gIleValThrAlaProAsnGlyAlaGlyAspPheAsnTrpSerHisIleGluGluThrLeuAlaSerA (253

780 AACCCCTAAAAATACACTTTGAAAAATATGAAAAATATGTTTTGTATACATTTTTTGATATTTTCAAACAATACGCAGTTATAAAAGCTCA 869

870 TTGAGCTAACCCATTTTTCTTTTGTCTTACGTTACAGATTGCAAAGACTCGAAACGCACCCGTGACAGCTACACCCGCTACCAGACCCCT 959  
 spCysLysAspSerLysArgThrArgGlnThrTyrThrArgTyrGlnThrLe (270  
 | \* \* \* \* \*  
 T=47ts

960 GGAGCTCGAGAAGGAGTTCACCTTCAATAGATACATCACCCGGCGTGTGCGATCGATATCGCCAATGCCCTGAGCCTGAGCGAAAGGCA 1049  
 uGluLeuGluLysGluPheHisPheAsnArgTyrIleThrArgArgArgArgIleAspIleAlaAsnAlaLeuSerLeuSerGluArgG1 (300  
 -----\*-----H1 \* -----\*-----Val-----H2 \* -----

. ||=Rp1 .

1050 GATCAAGATCTGGTTCCAAACCAGCGCATGAAGTCAAGAAAGGATCGCACGCTGGACAGCTCCCCGGAGCAGCTGTGGTGCCGGCTACAC 1139  
 nIleLysIleTrpPheGlnAsnArgArgMetLysSerLysLysAspArgThrLeuAspSerSerProGluHisCysGlyAlaGlyTyrTh (330  
 -\*-----\*-----\*-----\*H3\* \* \* | HOMEODOMAIN

1140 CGCGATGCTGCCGCCACTGGAGGCCACAAGCACCACCACCACCGGGCCACCATCGTGCCAGTGCCCATGTACCACCACCACCAACCAC 1229  
 rAlaMetLeuProProLeuGluAlaThrSerThrAlaThrThrGlyAlaProSerValProValProMetTyrHisHisHisGlnThrTh (360

1230 CGCCGCTACCCGCTTACAGCCACAGTCACAGTCATGGTTATGGCCTGCTCAATGATTACCCTCAGCAGCAGACCCACCAGCAGTACGA 1319  
 rAlaAlaTyrProAlaTyrSerHisSerHisSerHisGlyTyrGlyLeuLeuAsnAspTyrProGlnGlnGlnThrHisGlnGlnTyrAs (390

1320 TGCCCTACCCGAGCAGTACCAACAGCAGTGCAGCTACCAGCAACATCCACAGGACCTTACCATCTGTCTTGAGGTCCGGCAGTGTCTCAG 1409  
 pAlaTyrProGlnGlnTyrGlnGlnCysSerTyrGlnGlnHisProGlnAspLeuTyrHisLeuSerEnd (413

1410 TTACTCTCTCCCCAGAGCGGAACCGAAAGCCGTACCGCCACGAAACCGAAGCGCACTTCTCTGACCATTGTAGGTGACACGCAAATG 1499

1500 ACACAGCCGAGAACGAAGCTGCGACGCGATGAGTTGCACAGTAGAGGGCGCACTCCCTACGGTGCCAGGACATTTTGGGCACAAGGACG 1589

1590 AGTGCBCAAGTGCAGAAGGCAGAGGCAAAAGAGGCAGCGCAACAGAAAAGGAGCCTTGTGTCGCGCGGAACCCAGTGGCTGGCCATGAT 1679

1680 GGGTTCTCAGCGATCGATTAGCTGCGGCCAAAACACAAGCCAAAACACTCAGCTGGGAGTGATAATGCCAAGAGACTTGGAGACTGACA 1769

1770 CACATGTTTTGTACATATAGTAGTTAAGATATTCTCATAGAAATCTATTTATTAATAATACAGAGTAAAGTAAATCGATCGAATTT 1859  
 -----

1860 AAAACAATCAAGTTGAACATTCATTTGGCAATTTGTGAAGAAGAGTCTTGGGCATGCTGCAATTTGACTGCTTTAAAAATTTAACTTA 1949

1950 TAGGCCGTGGCGCGTATGTTGAATACATTTTCATATGTATATGTGTTGAAATACAATTAATGCCTTTCAATGATAACTACTCAATAAACT 2039

2040 TCCGAACATTATACGAAACGCAACGATTTAATGTTGAGCAGCAATCGTACAAATTCGAGCAGCTGCATTTTGTGCTTCAGTCCCCCTCA 2129

2130 TCCCTGACCCATTGCTGTCTCCCGGATTTTCTATTAATGCACCTTTTTCGCCAGAGAAAATGTCACATTTTGGTCTGGCTTCGGGGCAT 2219

2220 ATCTACCACCGCATCCCTGCTCCCTTCTCCCTCCGACGCTGCACGTTCTCTATTGAAGTGAAGACTTGATTGGTAATTTTTCATTGCA 2309

2310 CATCCGTGACAGTTATGGGTAAACGCAACGCAAAAGGAAAAGCCGGTGCGGAATCGGATTCGGAATCAGAATCAATATCAAAGGCAAAAG 2399

(P2) and *Ultrabithorax* (*Ubx*) (Ingham and Martínez-Arias 1986). In experiments carried out using cultured cells, FTZ stimulates *Ubx* transcription if a segment of *Ubx* that extends from 225 to 292 bp downstream of the transcription initiation site (*Ubx* downstream element U-B) is present (Winslow et al. 1989). Two FTZ-HD binding sites were detected in *Antp* by DNase I protection assays; these are approximately 500 bp upstream of the P2 transcription initiation site. Other homeodomain binding sites were detected near the distal transcription initiation site, but it is less certain that they are functional. The consensus sequence of these binding sites is CAATTA (Nelson and Laughon 1990).

FTZ is also required for expression of the *ftz* gene itself (see *Promoter*) (Hiromi and Gehring 1987; Ish-Horowicz et al. 1989), and it is also involved in the regulation of the segment polarity genes *en* and *wingless* (*wg*) (Howard and Ingham 1986; Lawrence et al. 1987; Ingham et al. 1988).

*ftz* is one of the pair-rule segmentation genes. Its overall function is thought to be to define the anterior border of even-numbered parasegments (Lawrence et al. 1987).

### *Tissue Distribution*

FTZ is first detectable by antibody-staining after the 13th nuclear division; it is localized in nuclei in seven stripes each approximately four nuclei wide (the spacing between stripes is also four nuclei). During gastrulation, the stripes narrow to three nuclei wide; FTZ stripes disappear just before the germ band is fully extended (Carroll and Scott 1985). FTZ and EVE (product of *even-skipped*) accumulate during approximately the same time in development. At first, the areas of EVE and FTZ accumulation overlap somewhat; but, as the stripes become narrower and better defined, the two products end up in an alternating pattern, FTZ in even- and EVE in odd-numbered parasegments (Appendix, Fig. A.3) (Frasch and Levine 1987). In embryos with fully extended germ bands, antibody staining is visible in 15 metameric clusters of nuclei within the developing ventral nervous system; this staining disappears soon after germ band shortening is completed (10–12 h of development). In 12–15 h embryos, FTZ reappears in the developing hindgut (Carroll and Scott 1985; Krause et al. 1988).

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*ftz* SEQUENCE (*opposite*). Strain carrying marker  $p^P$ . Accession, X00854 (DROANTCF) (modified by adding a G at –259 as per Brown et al. 1991). The following mutations are indicated: *ftz*<sup>Ual3</sup>, Pro-215 to Ser-215; *ftz*<sup>f47ts</sup>, temperature-sensitive, Ala-291 to Val-291; and the chromosome 2 translocation *ftz*<sup>Rp1</sup>, with a breakpoint after position 1,091 (Laughon and Scott 1984). The “zebra” element regulatory sites *ftz*-f1, *ftz*-f2 and *ftz*-f3 are from Ueda et al. (1990) and Brown et al. (1991). Sites *ae*2–*ae*3 (to which activators bind), *re*1–*re*3 (to which repressors bind) and *de*1–*de*2 (to which both activators and repressors bind) are from Topol et al. (1991). *ae*2a and *ae*2b correspond to the CAD-binding sites, *cdre* of Dearolf et al. (1989b).

### *Mutant Phenotypes*

Homozygotes for null alleles of *ftz* show severe developmental abnormalities that become evident at about the time *ftz* should be expressed. These mutants have half the correct number of segments due to the absence of regions corresponding to even-numbered parasegments (Wakimoto et al. 1984).

The dominant gain-of-function mutations Ual1, Ual2 and Ual3 cause substitutions in Pro-211 and Pro-215 (*ftz* Sequence) which increase the half-life of FTZ from < 10 min to 40 min. The increase in level and persistence of the protein and the concomitant expansion of its domain result in the corresponding suppression of *eve* expression in odd-numbered parasegments; this leads to abnormalities in the parasegments where *ftz* is normally not expressed, the anti-*ftz* phenotype. The Ual mutations affect a segment of the polypeptide (Thr-210 to Ser-221) that seems to be conserved in other early development genes (*hb*, *eve* and *prd*) as well as in *myc*. It has been suggested that those 12 residues serve as a signal for protein degradation (Kellerman et al. 1990). PEST-like sequences, also thought to be involved in protein degradation, are present in that region (Rogers et al. 1986).

### **Gene Organization and Expression**

Open reading frame, 413 amino acids; expected mRNA length, approximately 1,770 bases (assuming it extends for 20–30 bases beyond the putative poly(A) signal highlighted in the *ftz* Sequence), in agreement with an observed RNA of 1.8 kb (Laughon and Scott 1984). Primer extension analysis was used to identify the 5' end (Dearolf et al. 1989a; Ueda et al. 1990). The 3' end was not determined. There is an intron in the Asp-253 codon (Laughon and Scott 1984).

*ftz* is centromere-proximal to *Antp*, separated from it by about 30 kb, and transcribed in the opposite orientation (Weiner et al. 1984; Wakimoto et al. 1984).

### *Developmental Pattern*

*ftz* transcripts appear in embryos after the 11th nuclear division; they accumulate along the periphery of the embryo between 65% and 15% egg length. Between this stage and the end of nuclear cycle 13, the signal intensifies and becomes less uniform along the antero-posterior axis. Eventually, in nuclear-elongation-stage embryos (cycle 14), *ftz* RNA becomes localized in seven stripes positioned between 65% and 15% egg length, and it remains so through the completion of blastoderm cellularization. The anterior-most stripe is positioned posterior to the cephalic furrow. Stripes are 3–5 cells wide, and they are separated from one another by 3–5 cells. This segmented pattern persists through the early stages of gastrulation, but by the time the germ band is fully extended, *ftz* transcripts are no longer detectable.

The strongest embryonic expression of *ftz* is restricted to the period between 2 h and 4 h of development approximately (Hafen et al. 1984). The turnover rate of *ftz* mRNA is extremely high (half-life, 7–14 min) (Edgar et al. 1986); and the phenotype of a gain-of-function mutation  $T(2;3)ftz^{Rp1}$  seems to be the result of increased mRNA stability, possibly because of the loss of degradation signals in the 3' untranslated region (Kellerman et al. 1990).

The developmental pattern of *ftz* expression was also studied using the promoter region of *ftz* and  $\beta$ -galactosidase as a reporter enzyme. This method demonstrates that the seven stripes are sharper and more intense at the anterior border and that they fade posteriorly. The sharp anterior edge of each stripe coincides with the anterior edge of *en* expression in even-numbered parasegments, and it thus defines the anterior edge of these parasegments. (The same kind of pattern is observed for *eve* expression, except that it is the odd-numbered parasegments that are involved.) The  $\beta$ -galactosidase method also demonstrates segmental staining of prospective ventral ganglia neuroblasts in fully extended germ-band embryos (Hiromi et al. 1985; Lawrence et al. 1987).

### Promoter

Approximately 6 kb of 5' sequences are required for normal *ftz* expression (as measured by the ability of fragments of various sizes to rescue *ftz* mutant embryos in transgenic experiments). However, fusions of the promoter to the reporter gene *lacZ* showed that the most proximal 0.62 kb of 5' sequences ("zebra" element) are sufficient to produce the "zebra" pattern of expression. A segment between 2.45 and 0.62 kb upstream of the transcription initiation site is required for expression in the ventral nervous system. A segment further upstream, between 6.1 and 2.45 kb of the transcription initiation site, functions as an enhancer of expression of the "zebra" pattern. In the absence of this distal enhancer element, the striped pattern of expression is weaker, mostly restricted to the mesoderm and extended anteriorly, so that one or two extra stripes appear anterior to the cephalic furrow (Hiromi et al. 1985).

*The "Zebra" Element* The striped pattern of *ftz* expression seems to be established through a combination of generalized activation of the gene throughout the embryo and a specific pattern of repression. Two systems of repression contribute to the *ftz* expression pattern: one system represses expression in the anterior and posterior poles of the embryo, and the other represses in the inter-stripe regions of the "zebra" pattern (Edgar et al. 1986). Several activator and repressor sub-regions were identified within the "zebra" element by promoter deletion analysis, and they were found to correspond to protected regions in footprinting analysis (*ftz* Sequence) (Dearolf et al. 1989a; Topol et al. 1991).

A search for *ftz*-promoter-binding proteins yielded three fractions: FTZ-F1, FTZ-F2 and FTZ-F3.

FTZ-F1 first appears in 1.5–4.0 h embryos (at the time the *ftz* stripes occur);

it then diminishes, to reappear after 13 h of development and in larval and adult stages. FTZ-F1 binds to four sites in the *ftz* gene: site I is a 21-bp segment from -362 to -343 (*ftz* Sequence), sites II and III are in the coding region, and site IV (to which binding is 10 times weaker) partly overlaps the binding site of FTZ-F2 (see below). Sites I, II and III have the consensus sequence YCAAGGYCRCCR. Close contact with FTZ-F1 seems to be made by the two consecutive Gs of the top strand (marked by an asterisk in the *ftz* Sequence) and the two Gs on the bottom strand that are opposite the Cs at positions 8 and 10. Expression of a construction containing the "zebra" element attached to *lacZ* in transgenic embryos showed that mutations of site I that abolish FTZ-F1 binding lead to overall reduced expression of *ftz*, in particular in stripes 1, 2, 3, and 6 (Ueda et al. 1990). The sequence of FTZ-F1 has similarities with proteins of the steroid receptor superfamily both in the putative DNA-binding region and in the putative ligand-binding domain (Lavorgna et al. 1991).

FTZ-F2 is present at low levels in 1.5–4.0 h embryos, and its concentration rises after 4.0 h as expression of *ftz* diminishes. FTZ-F2 affords protection against nuclease digestion to two sites within the "zebra" element that share the sequence TGCNAGGACNT (*ftz* Sequence): *ftz*-f2 I (abbreviated f2 I) and *ftz*-f2 II, located between -260 and -200. The two adjacent Gs marked with asterisks seem to interact directly with an FTZ-F2 residue as indicated by methylation interference. Mutant *ftz*-f2-binding sites are unable to bind FTZ-F2. When such mutations are part of a "zebra"-element-*lacZ* construction, there is continuous *lacZ* expression along the antero-posterior axis; i.e., the repression of the *ftz* promoter in the inter-stripe regions fails. These mutations also lead to precocious expression of *ftz*, as early as the third nuclear division (Brown et al. 1991). FTZ-F2 is probably the product of *tramtrack* (*ttk*), a Zn-finger protein (Harrison and Travers 1990; Brown et al. 1991; Read and Manley 1992).

FTZ-F3 also bind to the "zebra" element, partly overlapping the FTZ-F2 binding sites (Brown et al. 1991).

CAD, the product of the segmentation gene *caudal* (*cad*), a homeodomain protein that forms a gradient of increasing concentration from the anterior to the posterior pole, participates in the regulation of *ftz* expression. CAD activates expression of *ftz* in the posterior regions of the embryo through its binding to the hexanucleotide TTTATG that is present in the protein binding sites *ae2a* and *ae2b* of the "zebra" element (*ftz* Sequence) (Dearolf et al. 1989a, 1989b).

*Distal Upstream Enhancers* A DNA segment that extends from approximately 6.1 to 3.5 kb upstream of the transcription initiation site can direct transcription of the basal *Hsp70* promoter and an associated reporter gene in a seven-stripe pattern in both ectoderm and mesoderm. The 2,574-bp segment contains multiple regulatory regions; from distal to proximal they are: (1) the most upstream 330 bp portion of this segment, which seems to be an activator of parasegment 4 expression; (2) the Distal Enhancer, extending from 331 to 1,502,

which is capable of directing expression in seven mesodermal stripes; (3) the 583-bp Element A of the Proximal Enhancer, between 1,780 and 2,363, which can direct expression in seven stripes in the ectoderm and mesoderm; (4) the 211-bp Element B of the Proximal Enhancer, which is required, in conjunction with element A, for ectodermal expression (Pick et al. 1990). There is also a scaffolding attachment region that occurs in an AT-rich segment between positions 575 and 763 of this distal upstream regulatory region (Amati et al. 1990).

FTZ itself seems to interact with the Distal Enhancer region to activate transcription (Hiromi and Gehring 1987; Ish-Horowicz et al. 1989). Numerous FTZ-binding sites are found within the Distal and Proximal Enhancers, and two independent autoregulatory loops seem to control expression (Harrison and Travers 1988; Pick et al. 1990). The product of *ttk* binds to DNA in the distal upstream region (Harrison and Travers 1988, 1990).

The pattern of *ftz* expression also depends on the products of gap genes and other pair-rule genes, *eve* and *h* in particular (Carroll and Scott 1986; Howard and Ingham 1986; Harding et al. 1986; Frasch and Levine 1987; Ish-Horowicz and Pinchin 1987).

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# 15

## *hairy: h*

**Chromosomal Location:**  
3L, 66D8-15

**Map Position:**  
3-26.5

### **Product**

DNA-binding regulatory protein of the basic helix-loop-helix (bHLH) type involved in embryonic segmentation and neurogenesis.

### *Structure*

Sequence comparisons indicate that the HLH motif extends from Ala-45 to Arg-90 (*h* Sequence), with 26% identity to a region of the mammalian oncogene *myc*. The helices have the amphipathic nature characteristic of dimer-forming regulatory proteins such as the products of the genes *daughterless (da)*, *Enhancer of split [E(spl)]*, *extramacrochaetae (emc)*, *twist* and of the *achaete-scute* complex genes *ac*, *sc*, *lsc* and *ase*. In the *h* and *E(spl)* proteins, the sequences of the basic regions, adjacent to and upstream of the HLH domain are more closely related to each other than to those in the *da* and AS-C proteins (Rushlow et al. 1989; Harrison 1991; Van Doren et al. 1991).

Within the *h* sequence, the OPA (CAG) repeat occurs several times and results in stretches of Gln, Ala or Ser in the C-terminal half of the protein. Near the C-terminus there are also regions of similarity to PEST sequences (segments rich in Pro, Glu, Ser and Thr that may be degradation signals). There are three potential glycosylation sites, at Asn-9, Asn-209 and Asn-296 (Rushlow et al. 1989).

### *Function*

During embryonic development, the HAIRY product seems to act as a repressor of *fushi tarazu (ftz)* helping to define the posterior border of *ftz* stripes. In *h* mutants, *ftz* expression occurs, but the striped pattern fails to develop (Howard and Ingham 1986; Carroll and Scott 1986; Ish-Horowicz and Pinchin 1987).

*h*

-3210 CGGC GCGTGGGGTTCTGTCGCTGTTAACTCGCAACGTTGCTGTTAAAAGAGCCCTACGATCAACAGTATACATAGTATAGTATATAT -3121  
 -3120 AGTATAGTTTATGGATACTATATATAAAATAATATCAACATAGTTAGTATCTAAGATAAGATTTTCTGTATATTTTAAACTTAATAGTCAA -3031  
 -3030 AACTATATGGTATTTTGAGTCTAGTGAATAACCATTTTGAATGATAATGGCACACAAATGAATTCATTGATCTTATAAAATACAAGCAA -2941  
 -2940 ATAATAATACCTATAATATTATACATATGCTATAATGTTATTATCAACGCCTTTACGATTATTAATAGTTAACCAACATGGTCCAAAAT -2851  
 -2850 GATTGCAAAATACCTCAAGGGGTTCTTTATCGTACCACCAACGTGTTGTTTGTGTTTTCAGTGATCAGGGCTCCCGGGGCTTATGGGGAA -2761  
 -2760 TCTGGGGGATCTGGCGCTGCTAATTTTAGACGCAATTAGCAATGCGCACATTTTTGTTGTTGCTTGCCTTTTTCGACTATAAATTTTT -2671  
 -2670 GCCACAGTTTATTTAGAAAGTGCATGTGATCGGGTCCGCCAACCAACAATGGGGCGTCAAATGGGGCTTCAACGCACAAACAACGT -2581  
 -2580 CGAGTGTATCTGTATCTGTGACTGTATCTTTAGCGTTGTATCCGTGAGATACATCCACACCTTTGGCTGTTTTTTGGCCAGCTAGCATGA -2491  
 -2490 TGTAGCTAGCATGATGTAACGCGCCCAACGTTTTCCGACCTCTCGTTTTTTTTCTTTTTGTTTTATTCTTTTTGCTTTTCGTTG -2401  
 -2400 CAATTAATGGCATGCACAAGTCCGCTCTGCCGCCGACACCGCTCTGCCGACGTGACCGGGGGGGCCGCTTTTGATCGGCTGC -2311  
 -2310 CAAATTGAATTGGAACGCGAAGGTGTTGTCGACGTCGCCACTACCGTCTATATATATATGTATCCATATTGTTGGGGCATGTGTTCTC -2221  
 -2220 GGCATAACAACCTTCTCGCGCACCACAAAACGCAACACTTTAGACAACCTCAAAAATTCAGAAATTCCTTAACCTTTTTGAGTAT -2131  
 -2130 TTTACGAATCGATAGATATGCATATTTGTAAGACGTGATTGTTGATTAAGTTAATTTCAATTTAGTTATTAAGCGAAATTAAGTGTAG -2041  
 -2040 TAAATCAAATTAACCTTAACAGTTTTTTACTCATCTTCATTAGAGTCAACTTTATTAGTTTCTATAAAAACACTGCCAGTGTTTC -1951  
 -1950 GTTATAAAAAAATATTGTAACACCCGTTTTTAGCCAATTTAATGTTAAAGCCTGACTGACTCATTCCAATGTAACCTTTGTTGACGA -1861  
 -1860 TTCGTGTTTTGGTATAACTTCACTAATCAGTGGTCAGAGTCCAAGTCAAGGCTTTAAAAATTTTCCAAGAACAACGTCAAAGATAAC -1771  
 -1770 GTAATTTCTCTTTATAGATCGTGAACCTAAATATGTGCATCTACCTTTACTGAGCTCAGCCTGGTTAAACTAATTACATGGTTATTAC -1681  
 -1680 CATTCTTAGAACTTAACCCATATTTGTAGATAATAGAAGGCTTAAGCAGTTATTTAAAATATCACTTTGCGTTGTAACCAATGTGTG -1591  
 -1590 TGACGCACCTTTGGCTTTTTACTACCAATAAACAATATAATTTAAGCTTCATTTTACCGTAATATTTCCAGTTTTTACAGCAATGCCCC -1501  
 -1500 TCTTCTCATTCTGCTAATGAATGGTTAGTTTTCTGATGCCGACTATTCCGCGTGC GCGTAATTATAGTCAACCTTCGATTAATCATT -1411  
 -1410 CTCCAAAACAAAACAACAATAATATGAAAACGTAAGAAATCCAACGCTGCACGTAGAAGCCATCAAGCTGAATCTAAGCGTCCGGCG -1321  
 -1320 GAGCACGTGTATCCACGACGCTTGTCCACAGCGATTTCATTTTCAATTTAGCCCGTTGGCGGCTATCGATCAAAAGCCAAAAGGGCGAC -1231  
 -1230 CTTCACTTAATTGAGGCGTACGGCATGCTGAATGAGTCGGTTGTACAGACTGGTCTGGAAAAATGCTAGGGGGATAACTATAGCCACCACC -1141  
 -1140 CACTGCCGATCGCCCAACACCAACCACTTCCGCTAGCGTGCACACAACCTTGTGATCTGTTTACTGTTTAGCGACCCCGA -1051  
 -1050 GCCGCAGATACACAGTACACAGCACAAAAACCGAACCTGTGCGACTGGGGTGGCGTCATATAGCCAGTATTTTCACTTCTATGGGAC -961  
 -960 GTCGTCGCGTTGGCCGATGAATCAGCAAACCAAGCGGACGACCAGAAACCAGCAGAAACCAACCAACCAACCCG -871  
 -870 GACCATCACAACAGCACAGCCAGAAACACAGCCTCTTGTGAATCCCTCAGTTAGCAGAGCCAGCAGAGTCAAGCCAAACCGATCGCTG -781

*(continued)*





During adult development, *h* seems to counteract the function of *ac-sc* complex genes in the development of sensory organs (Botas et al. 1982; Ingham et al. 1985b). It has not been possible, however, to demonstrate direct interaction of HAIRY with any of the *ac-sc* complex products involved (Van Doren et al. 1991).

### *Tissue Distribution*

HAIRY is intranuclear, as revealed by antibody staining. In cellular-blastoderm-stage embryos HAIRY-containing nuclei are distributed in eight stripes. After the onset of germ band extension, HAIRY rapidly disappears from the seven posterior stripes. (This pattern of occurrence is quite similar to that observed at the RNA level, see below.) A little later, in embryos having fully extended germ bands, HAIRY is transiently detectable in cells associated with pairs of tracheal pits (parasegments 4–13); still later, during germ band retraction and the following stages, HAIRY appears in the mesoderm, proctodeum and anal plates (Carroll et al. 1988; Hooper et al. 1989; a detailed comparison of the metameric distribution of HAIRY and other pair-rule-gene products is presented in these references). HAIRY also occurs in the imaginal discs of third-instar larvae and early pupae. In the eye-antennal disc, HAIRY is transiently present in a band of cells just anterior to the morphogenetic furrow. In leg discs, HAIRY is localized in groups of cells that evolve into longitudinal rows during disc eversion. In wing discs, expression occurs along presumptive wing veins. In all these imaginal structures, HAIRY is excluded from peripheral neurons and sensory organs (Carroll and Whyte, 1989).

### *Mutant Phenotypes*

*h* belongs to the pair-rule class of segmentation genes. In amorphic *h* embryos, certain metameric elements fail to develop in alternating segments. The missing structures correspond to the regions where gene expression is detectable (see below). Hypomorphic mutations are viable; they result in extra microchaetae and other sensory organs in the adult epidermis. In these hypomorphic mutants, the adult phenotype can be rescued by expression of *h* coding sequences under the control of a heat-shock promoter 6–11 h after pupariation (Ingham et al. 1985b; Carroll et al. 1988; Hooper et al. 1989; Rushlow et al. 1989).

## **Gene Organization and Expression**

The open reading frame that is thought to produce active protein is 337 amino acids long. However, the Met at position 10 occurs within a very good

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*h* SEQUENCE (previous pages). Accession, X15904 (DROHAIRG). The amino acids underlined constitute the HLH domain. Asterisks mark the hydrophobic residues thought to participate in the formation of dimers.

translation initiation context and so may serve as an alternative initiation site. There are two mRNAs,  $\alpha 1$ , with an expected size of 2,335 bases and  $\alpha 2$ , with an expected size of 2,139 bases; this is in agreement with the results of northern analysis. The two different sites of transcription initiation involved in production of the two mRNAs are 196 bp apart, and neither one has a canonical TATA box. Primer extension and S1 mapping were used to define the 5' ends. The 3' end was obtained from a cDNA sequence that included a poly(A) tail. There are introns after the Arg-33 and Asp-65 codons (*h* Sequence) (Rushlow et al. 1989).

### *Developmental Pattern*

*h* transcripts are first detectable in 2–4 h embryos, and they remain present throughout larval development.  $\alpha 1$  mRNA is prevalent up to 4 h, then both mRNAs are equally represented until the end of larval development, except for late third-instar larvae when  $\alpha 2$  becomes more abundant. In pupae, *h* mRNAs nearly disappear, but they become abundant in adults, with the two RNAs occurring in nearly equal amounts (Rushlow et al. 1989).

*In situ* hybridization shows that *h* mRNA in cell-cycle 12 embryos (syncytial blastoderm) is nearly uniformly distributed around the periphery of the embryo. Labeling then differentiates an anterior dorsal region (region 0 or AD) that extends for 12–15 nuclei at 95–85% egg length (Appendix, Fig. A.3) and a region of continuous labeling from 75% to 20% egg length. In the latter region labeling becomes discontinuous; before the completion of cellularization (mid-cycle 14), the labeling is distributed in seven evenly spaced stripes, each approximately 3–4 nuclei wide. In the abdominal region, the stripes of expression correspond to the posterior portion of the odd-numbered segments and the anterior portion of the even-numbered ones. This pattern is carried forward into the thoracic and cephalic regions, with the AD patch corresponding to the labrum (Appendix, Fig. A.3). When gastrulation starts, the cephalic fold invaginates between *h* stripes 1 and 2. Soon thereafter, the striped pattern disappears; and, by the time of germ band elongation, *h* transcripts are most evident in the hindgut and the foregut (Ingham et al. 1985a).

### *Promoter*

As in the case of *even-skipped* (*eve*), the *cis*-acting regulatory region is very extensive, >14 kb; and the striped pattern is the result of independent regulation of the individual stripes by various segments of the regulatory region. Thus each section of the regulatory region responds to unique positional cues along the antero-posterior axis of the embryo to activate transcription and produce a particular stripe. The positional cues are given by maternal products, gap genes and other pair-rule genes such as *eve*.

A construction that carries 14 kb of upstream sequences and the coding region of *h* is sufficient, in germline transformants, to rescue the embryonic mutant phenotype, but the adults that result exhibit a severe hairy phenotype.



This suggests that the region controlling *h* expression in adults is located more than 14 kb upstream of the transcription initiation site (TIS) (Rushlow et al. 1989). The 14 kb of upstream sequence was further subdivided into stripe-specific segments using *lacZ* as a reporter gene in germline transformation experiments. The whole 14 kb segment resulted in expression in the seven posterior stripes but not in the AD zone. Expression in individual stripes requires the following segments: stripe 1, 4.9–4.0 kb upstream of the TIS; stripe 2, several elements dispersed between 9.4 and 4.0 kb upstream of the TIS; stripes 3 and 4, several elements dispersed between 14.0 and 6.4 kb upstream of the TIS and elements further upstream; stripe 5, a segment between 6.8 and 4.0 kb upstream of the TIS; stripe 6, a segment between 9.1 and 5.2 kb upstream of the TIS; stripe 7, a segment between 11.0 and 9.4 kb upstream of the TIS. The positions of stripes produced by many of these artificial promoters are shifted slightly relative to positions of normal *h* stripes. Thus, with the possible exception of stripe 1, sequences other than those listed here for each stripe are required for normal expression (Howard and Struhl 1990; Pankratz et al. 1990; Riddihough and Ish-Horowicz, 1991).

The products of the gap genes *knirps* (KNI) and *Krüppel* (KR) bind, with varying affinities, to several regions of the *h* promoter. For example, KR, which is thought to act as a repressor, binds with high affinity to the region responsible for stripe 6, and KNI, which is thought to act as an activator, binds with low affinity to the same region. The formation of stripe 6 then, probably results because there is only one zone along the axis of the embryo where KNI is in high enough concentration to stimulate *h* transcription while KR concentration is so low that it does not repress *h*; this zone is in the posterior region of the embryo that corresponds to stripe 6 (Appendix, Figs A.2 and A.3). By this argument, the anterior border of stripe 6 is defined by the posterior slope of KR's bell-shaped concentration distribution. More posteriorly, stripe 7 may arise by similar interactions involving KNI and the product of the gap gene *tailless* (TLL). In this case TLL would act as a positive regulator at high concentration; and KNI would act as a repressor, defining the anterior border of stripe 7 through its posterior concentration gradient (Pankratz et al. 1990).

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# 16

## *hunchback: hb*

**Chromosomal Location:**  
3R, 85A3-B1

**Map Position:**  
3-48

### **Product**

DNA-binding regulatory protein of the Zn-finger type involved in the earliest stages of embryonic pattern determination.

### *Structure*

The amino acid sequence of HB suggests that there are two Zn-finger domains, one with four fingers, the other with two. Two short segments near the N-terminus (boxes A and B, *hb* Sequence) have some similarity to the *Krüppel* (*Kr*) protein (KR, another finger protein) and to the retrovirus HIV-1 *pol* product (Tautz et al. 1987; Evans and Hollenberg 1988; Harrison 1991).

### *Functions*

HB participates in the transcriptional regulation of several developmentally important genes; it recognizes a sequence distinguished mainly by a run of 6 As:

1. HB binding sites have been found in the *hb* promoter itself where it is thought to stimulate transcription (Treisman and Desplan 1989).

2. Binding sites for HB have also been demonstrated in the *even-skipped* (*eve*) promoter elements responsible for two of the embryonic stripes (#2 and #3) in which *eve* is expressed; here again HB probably acts as a positive regulator of transcription. (Stanojevic et al. 1989, 1991; Small et al. 1991).

3. HB has been demonstrated to repress *Kr* expression (Hoch et al. 1991). *Kr* expression normally occurs in an embryonic band immediately posterior to the area of HB accumulation (Appendix, Fig. A.2). HB is thought to be a repressor of *Kr* at high concentration (thus defining the anterior edge of the *Kr* zone of expression), and an inducer at low concentrations (Hülskamp et al. 1990).

*hb*

-4682 CATACAAATAATAAGTTATCCTTTTGTATTGTATAGAGAAAAAAGGTTTTTACCAATGAACATATGAATAATGAATAATAATAAGTTT -4593  
 <-

-4592 TTTTTTTTAGTCCAAAATTTGCATTAAACCTAGTTAGAACAATCGCTCCTAATTTATCATTCTAAAAGCGAACATTCCGCTTGGGAAAA -4503  
 ----- hb8 -----

-4502 AAATTGGTCTAAACCGAATGATACTATTAATGATATGCATTATTGCTAACCATAATCCTTGTCAAGCTAACATGATACATTTTCCGAA -4413  
 -----> hb7

-4412 ATTAGCTTAAAAAGGTGGAATACACCCAATATGCACAACTACCTTAAGGAGATTGGAATTCGAATGCTAATTGTGGCAAAGCTTTGC -4323  
 -----> hb6

-4322 CCAAATTAAGTTAACACGCACAGCAACAGGAAAAATGTGTTAAAGCAACAAGGAATCCTCTGGCCCAAACCTCCATCGTCCCAATTGCAG -4233

-4232 TTGGCTAAGTTGTTAATGTGTCTGGGCTTAAAGTGTCCCAAAAAACAATTTGGCGAAGGCCCCCATCTTCTCCATTTCGGCTCTCTCAC -4143  
 -----> hb5

-4142 TTTTGGGCCAGAAATCAATAATCAATAGTGAAGCGGAGATGCCAAAAACGGCAAAGAGCCAAAAGGCAGCTGCATTGGCCAAAATGC -4053  
 -----> hb4

-4052 AGCGCCAGAAAAATGCAAAAAGGATAAAATGAGCGAGTCAGAGCGAGAGTGGGTGAGTGAGTGAAAGAGCGAACGCCACGAAAGGGATGC -3963

-3962 TGTTTAGTTATTGCTTTTGGGGATGGGGAAGTCACTCAGATTTACAGCTAGCATCCGATCCGTTTTGAGTGAGTTTCGTCGTACGTT -3873

-3872 GTTGATGCTCTCCGGCTGCTCTCTCATTTTCGATTTCTGCTTCCGTTAACGGCTCTCGTGCCTTTGTGTTGTTGCACTTCTGGCAT -3783

-3782 TTGTTTTGCTTTGCAATTGAGATTTTATGATTTGAGTTCTTTTTTGGCGAGTGCCTGTTGTGAGGAGCAGTGAGGAGATTTTCAGCTAT -3693  
 -----  
 --->3662 ---

-3692 TAGAAGAGCCCGTGAGCGTGAGTTTGGTCAGTTGTGCTCCGAGTCCGAAAACGAAAGTCGCCAGCATTGACAGGCAGCCACGGAATA -3603  
 ==

-3602 CAAAAATAACCAACATCCAAGGACGAAACGTAACCTGCTATCAAAAACAATATTGCCATTAATACAATTAACCTTCGTGCTTGTGCTA -3513

-3512 AAAGATAACCAATTGCAAAAAGACTTTTGTCCCGAAAACTTATTTTTTGCAAGACCACATCCCACATGCGCGAATTCGCGGAAAAA -3423

-3422 GAAAGCACAAAAGCAAGCTAAAAGCGAGGCCCAAAAAATAGACAAAAACGAAGAGCAAGGAGCCCCACATCGCCGCTCCCCCTCGCAC -3333

-3332 GCACTGTGCGTGTGGTCTAACGGTAACCGTGCCCGGTCGAAGCGAGAGAGTGGGAAAGAGAGAGTCCGCGAGCGAAAAGCGAAAACTTA -3243

-3242 CGTGCTCCTGCTCCTGCTCCCGGTTCCCGTTTCCAATTTCCCGCAGAATTGCCTTGGACTGTCCGAAGTAAGTAACTGGGCGTAC -3153  
 -|

-3152 CATCCAATATCCTAGTTATACACTGCATTTCGACCTCCAATAAATCGTAAAAACACATGGAGGTAGAAAATTCGCAAAAGCTTTCCGCGGA -3063  
 -|

-3062 TAAACAAATAACAAGAATTACAATCGCTTTTGC GGAGCAAAATGCCAAAATGTTGGGGTATCCAGAATATACACAGTTTTTGTGAGGA -2973

-2972 TTTACATACGCCCTGTAATTTTAAATTTAGTTCTCAATTGATACGAAATCTGTTTTTTTTTATAGAATATATTCGATATATAGTTGTTTA -2883  
 <-----

-2882 ATTATAATTCATCATGTGATATACTTTCAAAAAGAACAGATTTAAATAGTTCGTTTATATGCTATTATGCACTATGCTTAATGTATTTTA -2793  
 --- hb3

(continued)



632 ATTTTTGTTTCTGCTCTAATCCAGAATGGATCAAGAGCGCAATCCTCAATCCGCGATCCGTGATCCTCGATTCCCGACCGATCCGCGAC -543  
-----hb1 -----> bcd-A2

542 CTGTACCTGACTTCCCGTCACCTCTGCCATCTAATCCCTTGACGCGTGATCCTGCTACCTGAGCGATATATAAATTAATGCCTGTTGC -453  
-----> bcd-A3  
--> -->-444/-440

452 AATTGTTCACTGACGACGAGTTGTATTACACTGCGACAACACAAGAGCAGCACCAATAATATACTGCAAAATCCTTACGAAAATCC -363

362 CGACAAATTTGGAATATACTTCGATACAATCGCAATCATACGCACTGAGCGCCACGAAACGGTAGGATATTGTTAGCCATTACCAAGTG -273

272 TCTCCATTTTGAACACAAAATCACTCAAAATCGCCTTCAGGGGGTGGGTGCCGCCACGCCACCCCTGACGTATTTTTGTTAGGGGTGGTG -183

182 CCGCAAGCACACCAAAAAAGAGAAAAAAAATAAAGCGAGGAAAAATAAAATGAAAAACAAGCGGAAAAAAGAGGAAAAAATCGA -93

92 CGCAGGCGCAGTGCATGAATGAATAAATGAATATGCCACTAACCCACTCTCTCTGTTTTCTTATCCATTACAGCCGCTAGAGCCGCC -3  
|

2 AAGATGCAGAACTGGGAGACGACGCCACCAACTACGAGCAGCACAAACGCTGGTACAACAGCATGTTTCGGGCAAAATATCAAAACAG 87  
MetGlnAsnTrpGluThrThrAlaThrThrAsnTyrGluGlnHisAsnAlaTrpTyrAsnSerMetPheAlaAlaAsnIleLysGln (29)  
|- box A

88 GAGCCAGGTCATCATCTCGACGGGAATAGCGTGGCCAGCAGTCCGCGCAATCGCCATTCCTCGACCAATCACCTGGAACAGTTCCTC 177  
GluProGlyHisHisLeuAspGlyAsnSerValAlaSerSerProArgGlnSerProIleProSerThrAsnHisLeuGluGlnPheLeu (59)  
-|

178 AAGCAGCAGCAGCAGCAGCTTCAGCAGCAACCCTAGGATACCTGTGCGCATTGACCCATACCCAGCCAAACGATCAAAACAGCCTG 267  
LysGlnGlnGlnGlnGlnLeuGlnGlnGlnProMetAspThrLeuCysAlaMetThrProSerProSerGlnAsnAspGlnAsnSerLeu (89)  
|- box B -|

268 CAGCATTACGATGCTAACTTGACAGCAACAGTTGCTGACGCAACAGCAGTACCAGCAGCATTTCAGGCGCCAGCAGCAACATCATCAC 357  
GlnHisTyrAspAlaAsnLeuGlnGlnGlnLeuLeuGlnGlnGlnGlnTyrGlnGlnHisPheGlnAlaAlaGlnGlnGlnHisHisHis (119)

358 CATCACCATCTGATGGGTGGATTCAATCCGCTGACGCCACCTGGTCTGCCAAATCCCATGCAAGCCTTCTATGGCGGCAATCTGCGACCC 447  
HisHisHisLeuMetGlyGlyPheAsnProLeuThrProProGlyLeuProAsnProMetGlnHisPheTyrGlyGlyAsnLeuArgPro (149)

448 AGTCCGACGCCACGCCACATCTGCCTCCACAATTGCGCCCGTTGCAAGTGGCCACTGGCAGCAGCAGAGAAGTTGCAGGCCTAACACCA 537  
SerProGlnProThrProThrSerAlaSerThrIleAlaProValAlaValAlaThrGlySerSerGluLysLeuGlnAlaLeuThrPro (179)

538 CCCATGGATGTCACACCGCCTAAGTCGCCGCCAAGTCGAGTCAGTCAATATTGAGCCGGAGAAGGAGCAGCAGATGATGTCGAACTCC 627  
ProMetAspValThrProProLysSerProAlaLysSerSerGlnSerAsnIleGluProGluLysGluHisAspGlnMetSerAsnSer (209)

628 AGCGAGGACATGAAGTACATGGCCGAGTCCGAGGACGATGATACCAACATCCGGATGCCATCTACAATTCGCACGGCAAGATGAAGAAC 717  
SerGluAspMetLysTyrMetAlaGluSerGluAspAspAspThrAsnIleArgMetProIleTyrAsnSerHisGlyLysMetLysAsn (239)

718 TACAAGTGCAAGACTGCGGGCTGGTGGCCATCACCAAGTGGACTTCTGGGCGCACACCCGCCACATGAAACCAGACAAGATCCTG 807  
TyrLysCysLysThrCysGlyValValAlaIleThrLysValAspPheTrpAlaHisThrArgThrHisMetLysProAspLysIleLeu (269)  
--- --- ---

808	CAGTGCCCGAAGTGCCCGTTTCGTCACCGAGTTCAAGCACCACTTGGAGTACCATATCCGGAAGCACAAGAACAAAAGCCCTTCCAGTGC GlnCysProLysCysProPheValThrGluPheLysHisHisLeuGluTyrHisIleArgLysHisLysAsnGlnLysProPheGlnCys	89 (2)
898	GACAAATGCAGCTACAGCTGTGTCAACAAATCCATGCTAAACTCGCACCCGAAGTCGCACAGTTCTGTGTATCAGTACCGTTGTGCGGAT AspLysCysSerTyrThrCysValAsnLysSerMetLeuAsnSerHisArgLysSerHisSerSerValTyrGlnTyrArgCysAlaAsp	98 (3)
988	TGTGATTACGCCACCAAGTATTGCCACAGCTTCAAGCTGCATCTCGCAAGTATGGTCACAAGCCCGCATGGTTTTGGACGAGGATGGC CysAspTyrAlaThrLysTyrCysHisSerPheLysLeuHisLeuArgLysTyrGlyHisLysProGlyMetValLeuAspGluAspGly	10 (3)
1078	ACCCCGAATCCCTCGTTGGTCATCGATGTTTACGGCACGCGTCGTGGTCCGAAGAGCAAGAATGGTGGACCGATTGCCAGTGGAGGAAGT ThrProAsnProSerLeuValIleAspValTyrGlyThrArgArgGlyProLysSerLysAsnGlyGlyProIleAlaSerGlyGlySer	11 (3)
1168	GGCAGCGGCAGCCGGAAGTCAAATGTTGCAGCTGTGCTCCGCAGCAACAGCAATCTCAGCCAGCTCAGCCAGTCGCCACATCTCAGCTG GlySerGlySerArgLysSerAsnValAlaAlaValAlaProGlnGlnGlnGlnSerGlnProAlaGlnProValAlaThrSerGlnLeu	12 (4)
1258	AGTGCCGCCCTGCAAGGATTCCTCTGGTTCAAGGCAACTCCGCTCTCCGGCGGCATCTCCAGTGTCTCCGCTGCCGCCCTCTCTCTGCC SerAlaAlaLeuGlnGlyPheProLeuValGlnGlyAsnSerAlaProProAlaAlaSerProValLeuProLeuProAlaSerProAla	13 (4)
1348	AAGAGTGTGGCCAGTGTGGAACAGACGCCAGCTTGCCAGTCCAGCCAATCTTCTGCCCTCTGGCCAGCCTTCTGCAGCAGAACCGC LysSerValAlaSerValGluGlnThrProSerLeuProSerProAlaAsnLeuLeuProProLeuAlaSerLeuLeuGlnGlnAsnArg	14 (4)
1438	AACATGGCCTTCTCCCTACTGGAACCTCAATCTCCAGATGCTGGCCGCCAACACAGCCGCTGTCTTGGCCCAATGTGCCCAAGA AsnMetAlaPhePheProTyrTrpAsnLeuAsnLeuGlnMetLeuAlaAlaGlnGlnGlnAlaAlaValLeuAlaGlnLeuSerProArg	15 (5)
1528	ATGCGAGAGCAACTGCAGCAACAGAACCAGCAGCAGAGCGACAATCAGGAGGAGGAGCAGGACGATGAGTACGAGCGTAAGTCACTGGAC MetArgGluGlnLeuGlnGlnGlnAsnGlnGlnGlnSerAspAsnGlnGluGluGluGlnAspAspGluTyrGluArgLysSerValAsp	16 (5)
1618	TCTGCCATGGATCTGTCCCAAGGAACGCCAGTGAAGGAGGATGAGCAGCAGCAACAACCCGACGAGCCGCTGCCATGAATCTCAAGGTG SerAlaMetAspLeuSerGlnGlyThrProValLysGluAspGluGlnGlnGlnGlnProGlnGlnProLeuAlaMetAsnLeuLysVal	17 (5)
1708	GAGGAGGAGGCCACGCCTCTGATGAGCAGCTCGAATGCCTCGAGAGCGCAAGGGACGCGTCTCAAGCTGGACACCCCTGTTTCAACTGCGA GluGluGluAlaThrProLeuMetSerSerSerAsnAlaSerArgArgLysGlyArgValLeuLysLeuAspThrLeuLeuGlnLeuArg	17 (5)
1798	TCGGAGGCCATGACATCTCCGAGCAACTGAAAGTACCCAGCACACCCATGCCAATGCATCTCGCCATTGCGGACGCAAACCCATG SerGluAlaMetThrSerProGluGlnLeuLysValProSerThrProMetProThrAlaSerSerProIleAlaGlyArgLysProMet	18 (6)
1888	CCCGAGGAGCACTGCTCGGCCACCAAGTTCGGCAGATGAGTCGATGGAGACGGCCATGTCGCCAGGCCAATACCAGTGCCAGTTCGACG ProGluGluHisCysSerGlyThrSerSerAlaAspGluSerMetGluThrAlaHisValProGlnAlaAsnThrSerAlaSerSerThr	19 (6)
1978	GCGTCCAGCTCGGGGAACAGCTCCAATGCCAGCAGCAATAGCAACGGCAACAGCAGCAGCAATCCAGCAGCAATGGAACCACTCAGCG AlaSerSerSerGlyAsnSerSerAsnAlaSerSerAsnSerAsnGlyAsnSerSerSerAsnSerSerSerAsnGlyThrThrSerAla	20 (6)
2068	GTTGACGCTCCTCCATCCGGAACCTCCGGCGGCGGGTCCATCTACGAGTGAAGTACTGTGATATCTTCTCAAGGACGCCGTGCTC ValAlaAlaProProSerGlyThrProAlaAlaAlaGlyAlaIleTyrGluCysLysTyrCysAspIlePhePheLysAspAlaValLeu	21 (7)
2158	TACACCATTACATGGGCTACCACAGCTGCGACGATGTGTTCAAGTGAACATGTGCGGCGAGAAGTGCACGGACCCGTCGGCTCTTC TyrThrIleHisMetGlyTyrHisSerCysAspValPheLysCysAsnMetCysGlyGluLysCysAspGlyProValGlyLeuPhe	22 (7)

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2248 GTTCACATGGCCAGGAATGCTCACTCCTAAGTCCCCATCACCATCACCTTGTTATTATTATTTACTACTATTATCATATAATCGTTGTC 2337
ValHisMetAlaArgAsnAlaHisSerEnd (758)
    ---
2338 CAGAATTGTATATATTCGTAGCATAAGTTTTCCAAACATTATTTTGTGTCGAAAATTGTACATAAGCCAATTAAGCCGCTAATTCTAGA 2427
2428 CCTAAGTTTATCTAACTATCCTAACTGTATTGAACTGTAGCCACCTTTCAATCTGTCTCCTATACACTCTTGTTATTTTCGAAATCGACTA 2517
2518 AAAACCTGAAAACGGTTTAAAACTATCATAAATGCATGGAGAAACATAAGCCTAAGTTAAATCTAATTTGTAAGTTGAGTCAAGCGAA 2607
2608 ACAACCAACAATAACCAACAGTCCAAGTCAAATTAATAAAATATAGTTTATAACATATACATAATGAGTATGTTTTCTAAAATAATTA 2697
    -----
2698 TTAGTCTTATTTAACCTAACATATTCGTATATGCGCATAAACACTCAGTCTTTCTCTGATATTATTCTCTCAGTATTTTGTAGTTGAAA 2787
2788 GCGAATTCGAATCGAACGAAATCAAATCAAATAAATCCAATTATTCAATATAATTTCAAGTTTTTCGCTTTTTTTTTTTGTTGTTAACC 2877
    ----- | (A)n
2878 TTTTGCCAATAATGACAATATTTTCGATGCAACTGAAACTGACGAAAGAAGAAGTACAATTTAGAGATTTTTAAAGAGTAGCTAAGAT 2967
2968 GCGCGAAATCTGAGCAACGGATCAAATTAG 2997

```

*hb* SEQUENCE. Strain, *Canton S*. Accession, Y00274 (DROHBG). Binding sites for DA/LSC (*da*, *lsc* products heterodimer), BCD, HB and KR are indicated by underlining the short sequences that match the consensus (or its complement) in each binding site. The presumptive Zn-binding Cys and His residues are underlined.

4. HB-binding sites exist in the *knirps* (*kni*) regulatory region (Pankratz et al. 1992) where HB may act as a repressor at intermediate concentrations, thus positioning the anterior border of *kni* expression more posteriorly than the anterior border of *Kr* expression (Hülkamp et al. 1990).

5. HB binds to a *bithorax* region enhancer (BRE) and thereby represses the expression of *Ultrabithorax* in the anterior half of the embryo (Qian et al. 1991).

### *Tissue Distribution*

HB is a nuclear protein localized initially in the anterior half of the embryo. It does not appear until the *hb* RNA antero-posterior gradient is apparent (see below), and thereafter it follows the general distribution of this RNA (Tautz 1988). After gastrulation, HB is detectable in four longitudinal rows of cells (6–8 cells per row per segment) that correspond to the first wave of differentiating neuroblasts (Cabrera and Alonso 1991).

### *Mutant Phenotype*

This gene belongs to the gap class of segmentation genes. In amorphic *hb* embryos, gnathal and thoracic segments are absent, and there are abnormalities in abdominal segments 7 and 8; it is an embryonic lethal (Nüsslein-Volhard and Wieschaus 1980; Ingham 1988).



## Gene Organization and Expression

Open reading frame, 758 amino acids. There are three mRNAs: the two transcribed from a proximal promoter have an expected size of 2,996 and 3,000 bases, and the third, transcribed from a distal promoter, has an expected size of 3,348 bases. These expected sizes are consistent with the two RNAs of approximately 2.9 kb and 3.2 kb detectable by northern analysis. Of the three transcription initiation sites, the most upstream was deduced from Southern analysis and sequence features while the other two were defined by S1 mapping and sequence features (*hb* Sequence and Fig. 16.1) (Tautz et al. 1987).

The two proximal initiation sites are under the control of a single promoter included within the leader intron of the distal transcription unit that extends between  $-3,170$  and  $-18$ . The proximal transcripts have leader introns that extend between  $-300$  and  $-18$ . There are no introns in the coding region (Tautz et al. 1987).

The 3' end was deduced from Southern analysis and sequence features. All transcripts have the same protein-coding capacity (Tautz et al. 1987).

The proximal breakpoint of the deficiency *Df(3R)p-XT104* is within the transcribed region and transcription is toward the centromere (Tautz et al. 1987).

### Developmental Pattern

Overall, expression of *hb* is restricted to oogenesis and the first 8 h of embryonic development.

The distal promoter is first expressed during oogenesis, and the mRNA persists after fertilization. The 3.2 kb maternal RNA is uniformly distributed in newly laid eggs. Between the 8th and 11th rounds of embryonic nuclear divisions (Appendix, Fig. A.1), an anterior-posterior gradient develops, probably by differential degradation, and under the control of *oskar* (Tautz et al. 1987; Tautz 1988).

The first embryonic expression of *hb* is from the proximal promoter, and it starts at the 11th or 12th nuclear divisions under the control of the *bicoid* gene (*bcd*) product (BCD). A combination of threshold effect and BCD gradient leads to uniform transcription of the 2.9 kb RNA in the anterior 45% of the embryo, with a sharp posterior boundary. Initiation of transcription by the proximal *hb* promoter is one of the earliest transcriptional events in embryogenesis. After cycle 14, with the beginning of gastrulation, the 2.9 kb RNA

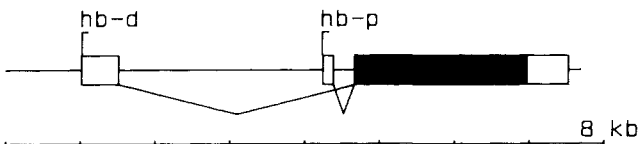


FIG. 16.1. Gene organization

disappears (Driever and Nüsslein-Volhard 1988, 1989; Schröder et al. 1988; Struhl et al. 1989).

Beginning at cycles 13–14 the 3.2 kb RNA is transcribed in a band at approximately 53% egg length (Appendix, Figs A.2 and A.3) and in a region of the embryo that corresponds to abdominal segments 7 and 8. During gastrulation, the spatial distribution of the 3.2 kb RNA increases in complexity; and after germ band extension, it becomes undetectable (Tautz et al. 1987; Schröder et al. 1988).

### Promoter

A 1.5-kb segment of DNA upstream of the distal transcription initiation site is insufficient for correct expression of the 3.2 kb transcript. On the other hand, a considerably smaller segment, one that extends between 50 and 300 bp upstream of the proximal site of transcription initiation is sufficient for correct developmental expression of the 2.9 kb transcript (Schröder et al. 1988; Driever and Nüsslein-Volhard 1989). The active core of the proximal promoter is a 100 bp segment that extends between –540 and –640 bp (Struhl et al. 1989), although some binding sites for the homeodomain protein BCD, as well as for the finger proteins HB and KR, are found further upstream in the proximal promoter region (*hb* Sequence).

The consensus sequence of the BCD-binding sites is TCTAATCCC. While the central TAAT seems to be the most conserved element (Driever and Nüsslein-Volhard 1989), the terminal CCC is important for discrimination between the BCD and Antennapedia homeodomains (Hanes and Brent 1991). Transcription from the proximal promoter in the posterior half of the embryo is repressed by KR, for which there are two binding sites in this promoter (Treisman and Desplan 1989; Licht et al. 1990). The existence of numerous binding sites for *hb* product (hb1–hb8) seems to indicate that this gene is also autoregulated (Stanojevic et al. 1989; Treisman and Desplan 1989).

At –847 there is a binding site for heterodimers of the helix-loop-helix proteins DA (*daughterless*) and products of the *achaete-scute* complex. This site may be responsible for activation of *hb* in neuroblasts, an activation that requires the *lethal of scute* product (Cabrera and Alonso 1991).

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# 17

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## The Heat-shock Gene Cluster at 67B: *Hsp22, Hsp23, Hsp26, Hsp27, HspG1,* *HspG2, HspG3*

**Chromosomal Location:**

2L, 67B

Synonyms for *HspG1, HspG2* and *HspG3*: *Gene1, Gene2* and *Gene3*

**Map Position:**

2-[28]

### Products

Small heat-shock proteins (HSPs): proteins of 22, 23, 26, and 27 kD, and three other small heat-inducible proteins.

### Structure

The small HSPs of *Drosophila* are thought to be homologous to those of many other species, from bacteria to mammals and higher plants. Although diverse in sequence, they all share the following features: (1) heat-inducibility; (2) some structural characteristics; and (3) the ability to form polymeric aggregates. In some species, *Drosophila* included, these proteins are phosphorylated and associated with RNA (see Lindquist and Craig (1988) for a review).

The polypeptides encoded by six of the genes in this cluster (*HspG2* is the exception) have two regions of similarities: (1) the 15 N-terminal amino acids, a hydrophobic segment with some resemblance to signal peptides; and (2) a segment of approximately 108 amino acids near the C-terminus with sequence similarities that range between 45% and 75%. In the latter segment, the first 83-amino-acid stretch matches approximately 50% of the mammalian  $\alpha$ -crystallin B2 chain; in *HspG3*, the crystallin-like region is only 50 amino acids long (Fig. 17.1) (Ayme and Tissières 1985; Ingolia and Craig 1982; Southgate et al. 1983; Pauli and Tonka 1987). The sequence similarities exhibited by these six genes, their uniform lack of introns, and their clustering, suggest that they are evolutionarily related to one another.

	1		50		100												
Hsp22	MRSLPMFWRM	AEEMARMPRL	SSPFHAFFHE	PPVWSVALPR	NWQHIAWQE	QELAPPATVN	.....K	DGY	KLTL	LDVKDY.							
Hsp23	MANIPLLSL	ADDLGRMSM	PFYEYYCQR	QRNPYLALVG	PMEQQLRLE	KQVGASSGSS	GAVSKIG...	.....K	DGF	QVCM	DVSHFK						
Hsp26	MSLSTLLSLV	DELQEPRSPI	YELGLGLPH	SRYVLP	PLGTQ	QRRSINGCPC	ASPICPSSPA	GQVLALRREM	ANRNDIHWPA	TAHVG	.K	DGF	QVCM	DVAQFK			
Hsp27	MSIIPLLHLA	RELDHDYRTO	WGHLLEDDFG	FGVHAHDLFH	PRRLLLPNTL	GLGRRRYSY	ERSHGHNQM	SRRASGGPNA	LLPAVGK	DGF	QVCM	DVSQFK					
HspG1	MSLIPFIDL	AEELHDFNRS	LAMIDDSAG	FGLYPLEATS	QLPQLSRGV	AWECNDVGAH	QGSVGGHRSI	AIIRTIVWPE	PRLAAISR	W	SKRNW	AIR					
HspG3	MPDIPVFLNL	DSPDSMYGH	DMFPNRM	YRRLHSRQHHLDL	LHTLGLIARM	GAHAHLVAN	KRNGELAALS	RGGASNKQGN	FEVHL	DVGLF	QP	GELT	VKLV				
CON	MS-IPLLL-L	AE-----	-----	-----L--	-----LR--	-----	-----	-----	-----K	DGF	QVCM	DV--FK					
	101		150		200												
Hsp22	.SELKVKVLD	ESVVLVEAKS	EQQEAEQGGY	SSRHFLGRYV	LPDGYEADKV	SSSLSDDGVL	TISVNP	PPGV	QET.....								
Hsp23	PSELVVKVD	NSV.LVEG.N	HEEREDDHGF	ITRHFVRRYA	LPPGYEADKV	ASTLSSDGVL	TIKVPP	PAI	EDK.....								
Hsp26	PSELNVKVD	DSI.LVEGK.	HEERQDDHGH	IMRHVRRYK	VPDGYKAEQV	VSQLSSDGVL	TVSIPK	QAV	EDK.....								
Hsp27	PNELTVKVD	NTV.VEGK.	HEEREDDHGH	IQRHFVRKYT	LPKGFDPNEV	VSTVSSDGVL	TLKAPPP	PSK	EQA.....								
HspG1	ARPGQARPV	ANGASKSAYS	VVNRRGFQVS	MNVKQFAANE	LTVKTDN	CI	VVEGQHDEKE	DGHG	VISRHF	IRKYI	LP	KGY	DPNEV	HSTLS	SDGIL	TVKAP	
HspG3	NECIVVEG..	.....K	HEEREDDHGH	VSRHFVPAVS	AAQGVRF	GCH	CFHFVGGWSS	QYHG	STISFQ	GGAQ	GAH	HTH *	.....				
CON	PSEL-VKV-D	-SV-LVEGK-	HEER-DDHG-	I-RHFVRRY-	LP-GY-A--V	VS-LSSDGVL	T---P-PP--	E-K-----	-----	-----	-----	-----	-----	-----	-----	-----	-----
	201		250														
Hsp22	.....	.LKERE	VTIE	QTGEP	AKKSA	EEP	KOKTASQ *	.....									
Hsp23	.....	.GNERI	VIIQ	QVGP	AHLNVK	ENPK	EAVEQD	NGNDK*	.....								
Hsp26	.....	.SKERI	I	IIQ	QVGP	AHLNVK	ANESEV	KGKE	NGAP	NGKDK*	.....						
Hsp27	.....	.KSERI	VIIQ	QTGP	AHLNVK	APAPE	AGDGK	AENG	S	G	E	K	M	E	TSK*		
HspG1	.....	QPLP	VVKG	SL	ERQ	ERIV	DIQ	QIS	QQ	KDKD	AHR	QSR	Q*	.....			
HspG3	.....	.....	.....	.....	.....	.....	.....	.....	.....	.....	.....	.....	.....	.....			
CON	-----	---ERIV	QIQ	Q-GPAHL	-VK	A---E-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----

FIG. 17.1. Comparison of six of the sequences in the 67B cluster. A residue is indicated in the CON(sensus) if three or more polypeptides agree in that position.

## Function

The specific function of small HSPs is unknown, but they seem to protect cells from heat damage. An extensive mutagenesis screen focused on the 67A-D region failed to uncover mutations in any of the small HSP genes. This failure and the sequence similarities among the genes in the cluster suggest functional equivalency and redundancy (Leicht and Bonner 1988).

HSP27 is localized in nuclei (Beaulieu et al. 1989). During development, the level of this protein parallels the transcription profile of *Hsp27* (Arrigo and Pauli 1988).

## Organization and Expression of the Cluster

The seven heat-inducible genes are clustered within 13–14 kb (Fig. 17.2).

In the absence of heat shock, all seven genes are expressed late in the third larval instar and during early pupation under the control of  $\beta$ -ecdysone (Thomas and Lengyel 1986). The level of expression is not uniform for the various genes: *Hsp23* is the most active gene; *Hsp26*, *Hsp27*, *HspG1* and *HspG3* are intermediate in activity and *Hsp22* and *HspG2* are the least active (Sirotkin and Davidson 1982; Mason et al. 1984; Ayme and Tissieres 1985). The seven genes are also expressed individually at other times in development.

All seven genes respond to heat shock (optimal temperature 35–36°C) at every stage of development except for early embryogenesis (Zimmerman et al. 1983), with *HspG2* response being lower than that of the others.

Transcriptional response to heat shock depends on the presence of at least two copies of a short, nearly palindromic sequence known as the heat-shock element, hse: CTNGAANNTTCNAG (Pelham 1985). In different genes, the position of the hse's varies considerably; their effect is independent of position so long as they lie within several hundred bp of the TATA box (see *Hsp70*).

## *Hsp22*

### Gene Organization and Expression

Open reading frame, 174 amino acids; expected mRNA length, 957 bases. Primer extension and S1 mapping were used to define the 5' end. S1 mapping was used to define the 3' end. There are no introns (*Hsp22* Sequence)



FIG. 17.2. Cluster organization

*Hsp22*

```

-764 GAATAAATGAAGATTTAATATTAATAGCTAAAAAAAACAGAAAACCTAAATATTTGTTAATATTAAGCTGATTTTTCATATATCTC -6:
-----| (A)n Gene2
-----HindIII
-674 AAGTTCTAGACTGCCATGCAAGCTTATCAATACACACAGTATACACTCGCACTCAGAAAGCTGTGCACTCCCACAAAACCTCTCTCTCC -5:
-584 CACTCTCTAATCGAGCTCTCTCAATGTGTCTCTCTGCGTATGGAACTGACCTTCCCAAGCGCAACAGCGAGAGAGAACTTCGCTAAA -4:
-494 TGCTAAAATAAAGGTAAATAAAGTAATATTTGGACACCCAGAGAGCCCCAGAAAACCTCCACGGAGTTCGCTAAAGAACAGTGAACAACC -4:
-----hse3
-404 CCTAACTAAATGCCATTGCCCGATTTCCAGGCAAAGCGGAAAATTCATCAGCAAAGGGCGAAGAAAATTCGAGAGAGTCCGGTATTTTC -3:
-----hse2
-->-250
-314 TAGATTATATGGATTTCTCTCTGTCAAGAGTATAATAGCCACCGGTTGGACACTACGCTCTCAGTTCAAAAAACCAACCAACTGCT -2:
---hse1-----
-224 AACAACCTCGAAGAAAGTCAACTAAATTAATAATTTGCCAGCTAAATAGAAATTCATACGATTGAAACCTCAGACAACAAGATTATCTTC -1:
-134 GAAACATAGAGGAAAAATTTAAAAAAAAGCCAGAAGTATTTCAAGATAACAATTTGGACGGAAATTCATCAAATTATTCGAATTTGCA -4:
-44 TAAGAAGCTTTATTGGAAAAACCAAGTTACCTTATCAACTACAATGCGTTCCTTACCGATGTTTTGGCGGATGGCCGAGGAGATGGCA 45
MetArgSerLeuProMetPheTrpArgMetAlaGluGluMetAla (15)
46 CGGATGCCACGCCTCCTCGCCCTTTCACGCCTTCTCCACGAGCCGCCGTTTGAGGTGTGGCGCTACCGAGGAACTGGCAGCATATT 13:
ArgMetProArgLeuSerSerProPheHisAlaPhePheHisGluProProValTrpSerValAlaLeuProArgAsnTrpGlnHisIle (45)
136 GCCCGCTGGCAGGAGCAGGAGTTGGCTCCGCCGCCACCGTCAACAAGGATGGCTACAAACTCACCTTGGACGTCAGGACTACAGCGAG 22:
AlaArgTrpGlnGluGlnGluLeuAlaProProAlaThrValAsnLysAspGlyTyrLysLeuThrLeuAspValLysAspTyrSerGlu (75)
226 CTGAAGGTCAAGGTGCTGGACGAGAGCGTGGTCTGTTGGTGGAGGCAAAATCGGAGCAGCAGGAGGCCGAACAAGGTGGCTATAGTTCCAGG 31:
LeuLysValLysValLeuAspGluSerValValLeuValGluAlaLysSerGluGlnGlnGluAlaGluGlnGlyGlyTyrSerSerArg (10)
316 CACTTCCTCGGCGATACGTTCTGCGGATGGATACGAGGCGGACAAGGTGTCCTCGTCGTCGAGCAGCAGCGGCTTCTGACCATCAGT 40:
HisPheLeuGlyArgTyrValLeuProAspGlyTyrGluAlaAspLysValSerSerSerLeuSerAspAspGlyValLeuThrIleSer (13)
406 GTGCCCAATCCTCCAGGCGTGCAGGAGACTCAAGGAGCGTGAGGTGACCATCGAGCAGACTGGCGAGCCG6CAAGAAAGTCCGCCGAG 49:
ValProAsnProProGlyValGlnGluThrLeuLysGluArgGluValThrIleGluGlnThrGlyGluProAlaLysLysSerAlaGlu (16)
496 GAGCCAAAAGACAAAACCGCCAGTCAGTAGAAAATAAGTTGAGATTACTAAAACCGATAAAAATGCTAGTGAACCTCTATGTTTAGATAT 58:
GluProLysAspLysThrAlaSerGlnEnd (17)
586 TCCAAAACCTATCAAATTTAAGTCTCTGTTAAATTAACAAGTTAATTTTAAAACAATTGTGATTCGGTAGCCGCAAGCCCAATAATTTT 67:
-----| (A)n
676 ATTTAGAAGAAAATAAATATTTGAAAAGACTATGATCAAATATTTACTTTNATTGGTTGGGTGGGAACACATTTGATATGGATAGTAT 76:
-----| (A)n
766 TATAGATTATATATATATCTGTCAAGTCT 795

```

*Hsp22* SEQUENCE. Strain, *Oregon R*. Accession, J01098 (DROHSP671). Dashes underline bases that match the consensus hse sequence. *HspG2* is immediately upstream of *Hsp22*: its poly(A) signal (-763) and last poly(A) site (-702) are indicated.

(Holmgren et al. 1981; Ingolia and Craig 1981, 1982; Southgate et al. 1983).

### *Developmental Pattern*

*Hsp22* is expressed in the third larval instar and, at barely detectable levels, in early pupae (Mason et al. 1984).

### *Promoter*

A 209-bp segment upstream of the transcription initiation site (to position -458) includes three hse's, and is necessary for full developmental and heat-inducible expression, as was demonstrated by study of 5' deletions (Klemenz and Gehring 1986). These studies also suggest that the segment between -443 and -383 is involved with hormonal induction. The first 26 bp of the leader seem to be important for transcription and for the preferential translation of *Hsp22* mRNA at high temperature (Hultmark et al. 1986).

## *Hsp23*

### **Gene Organization and Expression**

Open reading frame, 186 amino acids; expected mRNA length, 874 bases. Primer extension and S1 mapping were used to define the 5' end. S1 mapping was used to define the 3' end. There are no introns (*Hsp23* Sequence) (Holmgren et al. 1981; Ingolia and Craig 1981, 1982; Southgate et al. 1983).

### *Developmental Pattern*

*Hsp23* is expressed in late third instar larvae as well as in early pupae, when it is the *Hsp* gene that is most abundantly transcribed. *Hsp23* transcript reappears transiently in newly eclosed adults (Mason et al. 1984; Ayme and Tissières 1985).

### *Promoter*

Deletion analysis of the promoter region suggests that heat inducibility is controlled by a segment of the promoter region between -260 and -729. This segment includes five of the six hse's that occur within the promoter (Pauli et al. 1986). A segment between -250 and -490 is responsible for ecdysterone induction (Mestril et al. 1986).



*Hsp23*

-613	TTTCCCCTACAGAGCCCAATCTTGGATATTAATTAAGTTAATAGCTTAATGCCAGGCCATAAAAAGAAGAACTGTTCTGCTGTCT	-5:
-523	CGAAGTTTCGCGAATTTACTCCATCCTTCGTGGAATATACTCCAACCTTCCTATCTGCTATGATGTACATACATACGTGCTTACATACG --- -- - hse6 -- - --- - hse5 -- -- --- hse4	-4:
-433	TACATCTATACATACATAAATATTGCCGGTCTGATGCGACTTATCACTCCACCAGGCCTTTTTCATCCCACCTCCCTTAGGAGATTGC	-3:
-343	TCATTTTCCATAGCGATACTCTCACTTTCATGCGAGATAATGCGTAATTGCGGCAAATTCGAGAAGCTCTGCGATATTTTCAGCCCGAGA -- - --- - hse3 - - - --- - hse2 - - -	-2:
-253	AGTTTCGTGTCCCTTCTCGATGTCGATGTTTGTGCCCTTAGCACACAGACACGCGCACACACAGCGCCGACGGGCGCCGCACAC - --- - hse1	-1:
-163	TTCGACAGCAAGCGGTTGTATAAATATCCGGCACTTTCGTGCAACCGCGTCAGTTGAATCAAAAAGCCAAAGCGATAACAGCTAAAGC ----- -->-111	-7:
-73	GAAAGTAACCTATTAACAAAAGAAGTTTATTCTTTGAAGGAGGAGAATCATCTTGAAGCAATTAACAAAACAAAATGGCAAAATATCCAT MetAlaAsnIleProL (6)	16
17	TGTTGTTGAGCCTTGCCGACGATTTGGCCGAATGTCGATGGTGCCTTCTATGAGCCCTACTACTGCCAGCGCCAGAGGAATCCCTACT euLeuLeuSerLeuAlaAspAspLeuGlyArgMetSerMetValProPheTyrGluProTyrTyrCysGlnArgGlnArgAsnProTyrL (36)	106
107	TGGCCCTGGTTGGACCGATGGAGCAGCAGCTGCGCCAGCTGGAGAAACAGGTGGCGCCTCGTCGGGATCGTCGGGAGCCGTGTCGAAAA euAlaLeuValGlyProMetGluGlnGlnLeuArgGlnLeuGluLysGlnValGlyAlaSerSerGlySerSerGlyAlaValSerLysI (66)	196
197	TCGGAAAGGATGGCTCCAGGCTGCATGGATGTCGCACTTCAAGCCCAGCAGCAACTGGTGGTCAAAGTGCAGGACAACCTCCGCTCTGG IeGlyLysAspGlyPheGlnValCysMetAspValSerHisPheLysProSerGluLeuValValLysValGlnAspAsnSerValLeuV (96)	286
287	TGGAGGGCAACCACTAGGAGCGCGAAGATGACCATGGCTTTCATCACTCGTCACCTTTGTCCCGCCTATGCTCTGCCACCCGGTTATGAGG aIGluGlyAsnHisGluGluArgGluAspAspHisGlyPheIleThrArgHisPheValArgArgTyrAlaLeuProProGlyTyrGluA (126)	376
377	CTGATAAGGTGGCTCCACCTTGTCTCCGATGGTCTCCTGACCATCAAGGTGCCAAGCCACCGCAATCGAGGATAAGGGCAACGAGC IaAspLysValAlaSerThrLeuSerSerAspGlyValLeuThrIleLysValProLysProProAlaIleGluAspLysGlyAsnGluA (156)	466
467	GCATCGTTAGATCCAGCAGGTGGGACCCGCCATCTCAATGTGAAGGAGAATCCCAAGGAGCGGTGGAGCAGGACAATGGCAACGATA rgIleValGlnIleGlnGlnValGlyProAlaHisLeuAsnValLysGluAsnProLysGluAlaValGluGlnAspAsnGlyAsnAspL (186)	556
557	AGTAGAGGACTCGTTCCGGGAGATGCCCTGCATTATTAACCATTATCAAAGTCATACATCTGTTTTATAAGCTGTAGTTATCCAAGGAC ysEnd	646
647	ACTTCACTCATACACAATAGCCATTAAGGGTCTCTGCTTTAATCTTAGTTTGGAAATGTATTACTAAATTTGGCGAAATTAATATTACC	736
737	CATAAAAATAAATAACAAGTACACTTACTTATAATTGTGTTGGTCTGTTTTCTGGTGGTTATGGGTTACTATTACTATTACTATTAC -----  (A) <sub>n</sub>	826
827	TTCGGGAATTGTTTGGTAGCTCGGCCCTTTTTCTGTGATCCCGTTCTAGATTACTTTCTGCATTGTATATTGCATTGTTGTGTCAC	916
917	GTA AAAATGGCATT TTTATTTAATGTTGTTGTGTACATAACTGACTTTTACATTACTTCGGTAAAGAGTCTTGAAGCTATGAATGTAA	100
1007	GGA ACTCCAGTCAAGGTTAAATCCTTATGTAAGCATGCATAAGTACATCATGTACATACATACGTACATAAAAATATACATCCCTTTTC	109

## *Hsp26*

### Gene Organization and Expression

Open reading frame, 208 amino acids; expected mRNA length, 949 bases. Primer extension and S1 mapping were used to define the 5' end. S1 mapping was used to define the 3' end. There are no introns (*Hsp26* Sequence) (Holmgren et al. 1981; Ingolia and Craig 1981, 1982; Southgate et al. 1983).

### *Developmental Pattern*

In addition to being expressed in late third instar and early pupae, this gene is active in ovarian nurse cells in egg chambers at stages 7-10; the transcripts are transferred to the oocyte where they persist until the blastoderm stage of embryogenesis (Zimmerman et al. 1983; Mason et al. 1984). *Hsp26* promoter expression in several other tissues, including spermatocytes was detected using *lacZ* as a reporter gene (Glaser et al. 1986).

### *Promoter*

The effects of partial promoter deletions on *Hsp26* gene expression, as well as the localization of DNA-binding proteins suggest that *hse1-2* and *hse6* (*Hsp26* Sequence) are the *cis*-acting sequences responsible for heat-inducible expression of *Hsp26* (Cohen and Meselson 1985; Pauli et al. 1986; Simon and Lis 1987; Thomas and Elgin 1988). Nuclease protection studies identified (1) a constitutive footprint overlapping the TATA box and a fixed-position nucleosome between *hse1-2* and *hse6* (Thomas and Elgin 1988) and (2) a footprint produced by the GAGA-binding factor that extends from -312 to -264 (Gilmour et al. 1989).

Further upstream, from -704 to -534 there occurs a *cis*-acting region necessary for ovarian expression. All of the necessary information for ovarian, larval, pupal and heat shock expression is contained within the segment -910 to -169 (Cohen and Meselson 1985). Within that segment, two copies of the ovary-specific regulatory sequence (-704 to -534) are required to stimulate transcription of a basal-promoter/reporter-gene. Stimulation is very specific to the nurse cells and oocytes in egg chambers from stage 6 onwards. Footprinting experiments with ovarian nuclear proteins identified two binding sites in this 171-bp fragment (*onf1a* and *onf2a* in the *Hsp26* Sequence). Integrity of these sites is required for maintenance of the regulatory activity of the 171-bp

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*Hsp23* SEQUENCE (*opposite*). From -613 to 995: strain, *Oregon R*. Accession, J01100 (DROHSP673) with additions from Pauli et al. (1986) (see also V00210, DROHS09). From 996 to 1461, strain, *Canton S*, Hoffman and Corces (1986). Dashes underline bases that match the consensus *hse* sequence.

**Hsp26**

PstI

-942	TGCAGCAAACCAGGAACTGGCCAAGTGAAGTCGAACTAAAAGAAAAGAACATAAATAGTAATTAAGACAAAATAAATCTGCACGGGT	-85
	(A) <sub>n</sub> (minor) Gene1	
-852	AGGCGTGCGTTTTATCCATACGTTGTTCTGTGGTTTTCTTTCTTCATTTCACACAAAAAAGGAGGAGAAAGCTGACGGGAAA	-76:
	-----onf1b	
-762	AGCACTCAATTACTAATAGTGGAGATTGCGGGCGTTATATGTATGATTTCCATAAAACATATGTGACAACAACAAGTATTCC	-67:
	-----onf2a	
-672	CAAAGTAAACTTAAGACAGAAACACGAAATAATGTACTTAATAAAGAGAAAACCAGAATAAAAAAAGCTGACGTTTTGTTTGTTC	-58:
	-----onf1a	(A) <sub>n</sub> (maj) Gene
-582	CGTTAGCCGGCTGTTCTTTGCGCTCTTTCTAGAAAATTCACAACAACCTCTAGAAAACCTCGGCTCTCTCACTCATACAGGCGCACTAG	-49:
	-- -- -- hse7 -- -- -- hse6	
	- f6 -	
-492	CTCTGCTTTTGCAGTACGACAACAACACTTTAAAATTTCTCGAAAACCTATGGCATTATTGGGAAGGTTAGTTAGTTTTATTTTTG	-40:
	- - - - - hse4-5	
	-----onf2b	
-402	TTTTAGAGCAGCATTCAATTTAGACTTTTATAAAAAAATTTCTAATTTGATCCCTCGTTTATCAAACGATACAAAGCTATATTCATAA	-31:
	-- -- -- hse3	
-312	TTTTTCTCTCTGTGCAGTCT	-22:
	- GAGA protein -     - f1-2 -	hse1-2
	-- -- -- 183	
-222	CCAGCGGGTATAAAGCAGCGTCGTTGACGAACAGAGCAGATCGAATCAAAAAATCGAGCAGTGAACAACCTAAAGCAACTTTGCGC	-13:
	- FT -	
-132	AAAAGCAAACCTCAAACGAGAAAAAAGGATTA AAAACCTTGGCTTACAAGTCAAACAAGTTCAATCAACTTAACCAAGAAAAATA	-43
-42	TTTCAATCTCGAAAAGGAACATAACCTAAAGGAAACGAAAAATGTCGATCTACTCTGCTTTCGCTTGTGGATGAACCTCAGGAGCC	47
	MetSerLeuSerThrLeuLeuSerLeuValAspGluLeuGlnGluPr	(16)
48	CCGCAGCCCATCTACGAGCTTGGACTGGGATTGCATCCGCATCCCGCTACGTGTGCCCCCTGGCACTCAGCAGCGCCGTTCATCAA	137
	oArgSerProIleTyrGluLeuGlyLeuHisProHisSerArgTyrValLeuProLeuGlyThrGlnGlnArgArgSerIleAs	(46)
138	CGGATGCCCTTGCATCGCCGATATGCCATCGTCGCCGCGCCAGGTTTTGGCTTTGCGGCGGAGATGGCCAACCCGCAACGCAT	227
	nGlyCysProCysAlaSerProIleCysProSerSerProAlaGlyGlnValLeuAlaLeuArgArgGluMetAlaAsnArgAsnAspI	(76)
228	TCACTGGCCGCAACCCGCCATGTTGGCAAGGATGGATTCCAGGTGTGCATGGACGTCGCCAGTTCAAGCCAGTGAGCTCAACGTGAA	317
	eHisTrpProAlaThrAlaHisValGlyLysAspGlyPheGlnValCysMetAspValAlaGlnPheLysProSerGluLeuAsnValLy	(106)
318	GGTGGTGGACGACTCCATCTTGGTCGAGGGCAAGCATGAGAACGCCAGGACGACCATGGTCACATCATGCGCCACTTTGTGCGCCGCTA	407
	sValValAspAspSerIleLeuValGluGlyLysHisGluGluArgGlnAspAspHisGlyHisIleMetArgHisPheValArgArgTy	(136)
408	CAAGGTTCCCGATGGCTACAAGGCGGAGCAAGTGGTCTCGCAGCTGTCTGCGATGGCTGCTCACCGTGATTTCCCAAGCCGAGGC	497
	rLysValProAspGlyTyrLysAlaGluGlnValValSerGlnLeuSerSerAspGlyValLeuThrValSerIleProLysProGlnAl	(166)

498	C6TCGAGGACAAGTCCAAGGAGCGCATCATTCAAATTCAGCAAGTGGGACCCGCTCACCTCAACGTTAAGGCCAAATGAAAGCGAGGTGAA	587
	aValGluAspLysSerLysGluArgIleIleGlnIleGlnGlnValGlyProAlaHisLeuAsnValLysAlaAsnGluSerGluValLys	(196)
588	GGGCAAGGAGAACGGAGCACCCAACGGCAAGGACAAGTAAAGGAGCCATCATCATCCAACATCATCCATCATCATTCCCCTACTTAATTG	677
	sGlyLysGluAsnGlyAlaProAsnGlyLysAspLysEnd	(208)
678	TTCCTAATTTATTGCATTGTATTGTAAATGAGCTAAAGACTAGAATACTCATATTAATTTAATAAACTCTTTTGTTCACCTGGTGGAA	767
	-----   (A) <sub>n</sub>	
768	AATTAATAATGTTGCGACTTTTGATATGAAAGTTGGTTTTGAAAGAGGCCAAATATTTGGAATCGATCCGAAGATTGAATTGGGCGC	857
858	GACGAGGTGAAGACCCATTTCGTAACACCAAGTGTCTACCAAAATATTTATGCGCATTATTATATCAACTACGGGTACAATTTGTATT	947
948	TATTTATGTTTGAATCCAATTTAAATGTTCCGCTGCAATTGCTTGGTGTCCGAAAATAGTTACCTTGAGTTAGGCGCATTTCGATGGTT	1037
1038	GGGATTTGGGTTTGGTAAACACACATTCACTGCTTGCCCTTCTGATTCTGACACATGGTCCACTATTTCCAGGGCAGGGCCAGCTTTCC	1127
1128	GGTTTCATGAACCGGACCAATCTCTCCGGCGTGTAGTACTTGGCTGGCGGGGGTGGTGGAGCCTTGATGGTGAGGATGCCATCGCT	1217
1218	GGATATGTCCGAGATTACCTCATTGGCATTGTATCCGCGGGGCGAGAAGTACTTCTCACAAAGTGCCGCTCCACTAGGCCATTGGAACC	1307
1308	CTCGTCGCGACGATTGTGATTTCCCTGGAGCATGACATAGTCGTCATTGGTTTTGACCACAATGTCGTTGGGGATGAAATTGTCGTATCGA	1397
1398	T	1398

*Hsp26* SEQUENCE. Strain, *Oregon R*. The segment -672/1,398 is from GenBank: Accession, J01099 (DROHSP672), as modified by Thomas and Elgin (1988) (see also X03890, DROHSP26G). The segment -942/-839 is from *HspG1* Sequence. The segment -838/-673 was kindly supplied by R. S. Cohen. Dashes underline bases that match the consensus hse sequence. fT is the footprint associated with the TATA box; f1-2 and f6 are footprints associated with hse's. The onf's are ovarian-nuclear-factor binding sites. The polyadenylation sites of *HspG1* are indicated.

fragment. Second copies of these binding sites occur at -798 (onf1b) and -474 (onf2b). The nuclear factors that bind to onf1 and onf2 are ovary-specific (Frank et al. 1992).

## *Hsp27*

### Gene Organization and Expression

Open reading frame, 213 amino acids; expected mRNA length, approximately 1 kb. Primer extension and S1 mapping were used to define the 5' end. The 3' end has not been defined. There are no introns (*Hsp27* Sequence) (Holmgren et al. 1981; Ingolia and Craig 1981, 1982; Southgate et al. 1983).

*Hsp27*

-698	CGGCAAACATGAGGAGCAGCACGAAGCGAGACAAGGGTTCATGCACCTGTGCCAATGAAAAATACAAGCTCTGTTGCACTCTGAAAAGACT	-60'
-608	GCTTTTAAAGCGCGATAAGAGAAGAAAAATGTTTTAAATAATACATATATCTGCATATATACGTACATGTACATATGTATGTACTGCAT	-51'
-518	TTTAACTGTTGTTTTGCTTTTTATTTCGCAAAGAGAAACTCCCAGAAAAAGAAATGTCAAGAAGTTTCTGGTCTTTCTCCCTCTCTCTAT	-42'
	hse5 - - - - - - - - - - - hse3-4	
-428	GAAAAGCCGCTGTGCCAGAAAGAGCCAGAAGATGCAGAGAAAAACTGTTTGTGAATTACGGGGCGTATTCAAAGGGGCTTTTAAATGTC	-33'
	- - - - - hse1-2	
-338	GCTTAAATTTTAAAGTTTGACAGGCTAATAATTGCTTGCTATATCTAAATATTATTATATTGCATTAGGGGATCATAGGGAAAACTTC	-24'
-248	TCTGCAGGCCAAAATCTAACGAAGATGGCAACCCCCATCATTTTATTAAAGTTCGGTCCCTGGTGCCATGCACCTAGTGTGTGTGAGCCC	-15'
	-->-118	
-158	AGCGTCAGTATAAAAGCCGGCGTCAACGTCGCCCAGCACAGCTCAAACCTGAAAAATGGAAGGCAAACGTTGAAGCAAACCTCGCTAAAA	-69
	-----	
-68	AAATTCGAAAAGCAAAAAAATTCCTTTGTCTAGACAGGGTTGTGAATAAGAGAAAAAAAATCAAAAATGCAATTATACCCTGCTG	21
	MetSerIleIleProLeuLeu (7)	
22	CACTTGGCCCAGGTTGGATCATGACTACCGACCGACTGGGGCATTGCTGGAGGATGACTTCGGTTTTGGCGTCCATGCCACGAT	111
	HisLeuAlaArgGluLeuAspHisAspTyrArgThrAspTrpGlyHisLeuLeuGluAspAspPheGlyPheGlyValHisAlaHisAsp (37)	
112	CTGTCCATCCGCGTCGCTGCTACTGCCCAACACCCTGGGACTGGGTCGCTGCCTTATTCGCCGTACGAGAGGCCATGGCCACCAC	201
	LeuPheHisProArgArgLeuLeuLeuProAsnThrLeuGlyLeuGlyArgArgArgTyrSerProTyrGluArgSerHisGlyHisHis (67)	
202	AATCAAAATGTCACGTCGCGCGTGGGGGGTCCAAACGCTCTGCTGCCCGCCGTTGGGCAAGATGGCTCCAGGTGTGCATGGATGTGTG	291
	AsnGlnMetSerArgArgAlaSerGlyGlyProAsnAlaLeuLeuProAlaValGlyLysAspGlyPheGlnValCysMetAspValSer (97)	
292	CAGTTCAGCCCACGAGCTGACCGTCAAAGTGGTGGACAACACCGTGGTGGTAGAGGGGAAGCACAGGAGCGCGAGGACGGCCATGGA	381
	GlnPheLysProAsnGluLeuThrValLysValValAspAsnThrValValValGlyLysHisGluGluArgGluAspGlyHisGly (127)	
382	ATGATCCAGCGTCACTTTGTGCGCAAGTATACCTGCCCAAGGGCTTTGACCCCAACGAGGTAGTGTCCACTGTCTCATCCGACGGTGTG	471
	MetIleGlnArgHisPheValArgLysTyrThrLeuProLysGlyPheAspProAsnGluValValSerThrValSerSerAspGlyVal (157)	
472	CTGACCCTCAAGGCCCGCCCGCCAGCAAGGAACAGGCCAAGTCGGAGCGCATGTTCCAGATCCAGCAAAACGGGGCTGCCATTG	561
	LeuThrLeuLysAlaProProProSerLysGluGlnAlaLysSerGluArgIleValGlnIleGlnGlnThrGlyProAlaHisLeu (187)	
562	AGCGTCAAGGCACCGCACCCGAGGCTGGCGATGGAAGAACCCGAAAAATGGCAGCGGCGAGAAAATGGAGACTAGCAAGTAAAAGACGAAA	651
	SerValLysAlaProAlaProGluAlaGlyAspGlyLysAlaGluAsnGlySerGlyGluLysMetGluThrSerLysEnd (213)	
652	AGAGGAAGAAGACTAGGAGATGAAGAAGACGAGAAGAGGAAGAAGACTAGAAGAGGAAGAAGTCGTGAAGGAGGAAGAAGACGAGAT	741
742	TCGCTGGCGAAGCAGGAGAGAAAAGAAGAATTTAAAAAGAAACCGGGAGTGTGCCCGCTGCTCGGAGAGAGCAAGACTAAAAAGGACA	831
832	CACCACAACACCCAATGTATTACATTCACACACATCACATCATATACATCATAACATCCTAGACTAAGTGATTTTAACTCCA	921
922	TTTATCATAATGCATAAAAAAACAAAATTTT 953	

### *Developmental Pattern*

The pattern of expression during late third instar, early pupal stages and oogenesis is similar to that of *Hsp26* (Zimmerman et al. 1983; Mason et al. 1984).

### *Promoter*

Studies of 5' deletions established that the 579 bp upstream of the transcription initiation site (to position -696) are sufficient for full response to induction by ecdysterone (late third instar expression) or heat. The effect of the two treatments is mediated by two independent regions of the promoter: the hormonal-response segment extends from -696 to -572, and the heat-induction segment from positions -486 to -345. This latter segment includes five hse's in two clusters (Riddihough and Pelham 1986; slightly different results were reported by Hoffman and Corces 1986). The positions of the hormonal and heat-inducible regulatory regions are well correlated with DNase hypersensitive sites (Costlow and Lis 1984).

## *HspG1*

### **Gene Organization and Expression**

Open reading frame, 238 amino acids; from the major 5' end (at -92), the expected mRNA lengths are 1,423 and 1,904 bp, in agreement with bands of 1.6 and 1.9 kb seen in gels (the 1.9 kb band is the stronger). Primer extension and S1 mapping were used to define the 5' ends. S1 mapping and cDNA sequences were used to define the 3' ends. There are no introns (*HspG1* Sequence) (Ayme and Tissières 1985; Vázquez 1991).

### *Developmental Pattern*

*HspG1* is expressed in late third instar larvae, in white pupae and in freshly eclosed adults (Ayme and Tissières 1985). Heat shock causes a weak response in embryos and adults but a 10-100 times stronger response in pupae. This developmental response seems to be hormonally controlled, because cells in culture respond much more strongly to heat shock if ecdysterone is present (Vázquez 1991).

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*Hsp27* SEQUENCE (*opposite*). Strain, *Oregon R*. Accession, J01101 (DROHSP674) as modified by Riddihough and Pelham (1986). This sequence follows immediately after position 1,461 in the *Hsp23* Sequence (Hoffman and Corces 1986). Dashes underline bases that match the consensus hse sequence.

*HspG1*

-1196 TTTTATTACTATGTACAAGGGGCGATCTCGTACGCAGCATGCTCTGAAGTTTTGCTCTTCCGACTGCAGCTGGCATATACCATATCA -1

-1106 ATACAACATACTATATAATATAATATATGCCCACACAGAATTGTATCCCGCAGCTGAGTTCCGGGCCCCAGTAAATTTTAGCAAAGTC -1

-1016 TCCACTGTCTGGCCTCCGTCTGGATGTTGTTGGTGTGTTGTTGTTTTGCATTTTGGAGCTTTTCAACCGGTTGCCATCGCTTGCACCT -5

-926 TGGCTATGTAACCACATACGAATCCAGCAATATCATCATCATCTGTGTGGCAGGGTACATACATATGTATGTAGATACAAATGTATATGC -8

-836 CGACACCATATGTATGTTGCCCCAGACGCTGCTACTGCCATGTTTACGCGACGCCGGTCCCAATCTCCAGCTCTGACAACAGCGGA -7

-746 TTTGTAGCTTCCAGGCGCCCTGCCAGCCAGCCAGCCAGCTGTTGTTGTAGTTGTTTATCGCCGGCGGACTCGAATTCGCATCGGCA -6

-656 AGCCGGCACGAGACTCAGACCTCTCAGCTGTTTCGCTCAATGCCGGCAGTGGAATTCAGCTGCAACACGGACCCTTTACATATACCCCG -5

-566 TCTATATGGATATTGTATATATGAGTACATATATGTATATCGCCGGTACAAGGAAGATGGCATCTCTTGGGGGGGATATTCGTGCATAT -4

-476 ATGCTTCGATTTCAAGCCGGTTTGCCCTCTTTACTTATCTTTTTTATTGTTTTGCAACGTTGCAGTTTGGTTGTCTGGTTCTCGC -3  
 -->(minor)

-386 CGACTACGAGTACGAGTACTTTCTTTGTTCTCTGGCTATCTGCGGTAGAGGAAAAGTATCTCTTATTTCTGTATATAGCAGAAAATGGC -2

-296 ATAGTACATGGCTTGACTGACTGTTTTAATGGGTAGCCCTCCCTTGGCTGAGGCTTCTCTGGAGGATTCGATTAGTTTTTCGCCCTGG -2

-206 GAGCTGGCCTGGAAGCCGACTGGAAGTGACCAGGTTTTCCATTCAGCGCTGCACAGCCGCTTAAAAGCGTCGACATTACGCCATAAGGGC -1  
 -->(minor)

-116 TCAAACGCAGTCCAGTTGGAGGCCAGAACGGATCGCCGCCGGTCCAGACGACACCAATCCCCGCAAGACCTAAAAATAAAGATATA -2  
 --->92

-26 TCTTAGCCAGATAGGAAGAAAGTGAAAATGTCGCTGATACCGTTCATACTAGATTTGGCCGAGGAGCTGCACGATTTCAATCGCAGCCTG 63  
 MetSerLeuIleProPheIleLeuAspLeuAlaGluGluLeuHisAspPheAsnArgSerLeu (21)

64 GCAATGGATATAGATGATTCGCGGGATTTCGGGTTGTATCCACTGGAGGCCACCTCACAGTTGCCACAGCTGAGTCGTGGCCTGGGGCG 15  
 AlaMetAspIleAspAspSerAlaGlyPheGlyLeuTyrProLeuGluAlaThrSerGlnLeuProGlnLeuSerArgGlyValGlyAla (51)

154 TGGGAATGCAATGATGTGGGTCGCCATCAAGGGTCAGTCGGCGGCCATCGCAGCATCGCCATCATCCGTACAATCGTGTGGCCGGAGCCA 24  
 TrpGluCysAsnAspValGlyAlaHisGlnGlySerValGlyGlyHisArgSerIleAlaIleIleArgThrIleValTrpProGluPro (81)

244 AGACTGCTTGTGCAATAAGTCGCTGGTGGAGCTGGAAGGAAATGGGGCATAAGGGCAGCTCGGGGCAAGCGGCACGACCAGTGGCC 33  
 ArgLeuLeuAlaAlaIleSerArgTrpTrpSerTrpLysArgAsnTrpAlaIleArgAlaArgProGlyGlnAlaAlaArgProValAla (11)

334 AACGGGGCCAGCAAATCCGCCTACTCCGTGGTGAATAGGAACGGCTTCCAGGTGAGCATGAATGTGAAGCAGTTCCGCCCAACGAACTG 42  
 AsnGlyAlaSerLysSerAlaTyrSerValValAsnArgAsnGlyPheGlnValSerMetAsnValLysGlnPheAlaAlaAsnGluLeu (14)

424 ACCGTCAAGACCATCGATAACTGCATCGTGGTCGAGGGTCAGCACGACGAGAAGGAGGATGGCCACGGGGTATCTCGCCCACTTCATC 51  
 ThrValLysThrIleAspAsnCysIleValValGluGlyGlnHisAspGluLysGluAspGlyHisGlyValIleSerArgHisPheIle (17)

514 CGCAAGTACATCTGCCCAAGGGCTATGATCCCAACGAGGTGCACTCGACCCTCTCCTCGGACGGCATTCTGACGGTGAAGGCGCCGACG 60  
 ArgLysTyrIleLeuProLysGlyTyrAspProAsnGluValHisSerThrLeuSerSerAspGlyIleLeuThrValLysAlaProGln (20)

604 CCATTCAGTCTCAAGGCAGCCTGGAACGACAGGAGCGCATCGTAGACATCCAGCAGATATCGCAGCAGCAGAAGGATAAGGATGCG 69  
 ProLeuProValValLysGlySerLeuGluArgGlnGluArgIleValAspIleGlnGlnIleSerGlnGlnGlnLysAspLysAspAla (23)

394	CACCGCCAAAGCCGTCAGAGGTAGAGCAGCAGGCGCACGTCAGTGCACCACCTCCACTTTAAATCCGACTGCACCCACACCCTCCTTCG HisArgGlnSerArgGlnArgEnd	783 (238)
784	CTCTCGCTCACTCTCGCCGAGAGCAACGCAAGGTCAGGAAGAGACAGAGATGGAAATGCCGGCTGTTTCGCCATTTCATGAGGCTGC	873
374	TGTCGCCGTGCTGTTGCGATGGAAGCCCTCCACCGCAGGAACCCTCCAGTGCCAACAATGGCGTTCGAGAACCAGAATCAGAGTCC	963
364	ATGGAAGTGGCGTTGGCCAAAAACGAGAGACTGCCAATGTGGATGAACCCACACCCAATCCCCTTATAAGCTACGAAGAGGAGCAAAG	1053
354	GCAGAGGATGCAAATGCCAACGAAGTGCCCGTTGCCTCGAATAACGGCAATGGAGCAGTCGCAGCAGCCGAGGATGTAAATGCCGCTGG	1143
144	CCAAGAACCAGAAATCTCCACGGAAGACAGCAAAGAGGAGCAGGCGGAGAAGTTGATAAAGTAGAGAAATGGAGGAGAAGGGCGGCGAGG	1233
234	CAACTGGCAGCCGTAGAATGCGGCCATTCTACTGGCCAAAAACCAAGGCGAAATGGAGCCACTGCAGCAAAACCAGGAAGTGGCCAAG _____PstI	1323
324	TGAAGTCGAACTAAAAGAAAGAACATAAATAGTAATTAAGACAAAATAAATCTGCACGGGTAGGCGTGCCTTTATTCCATACGTGTT  (A) <sub>n</sub> (minor)	1413
414	TCTGTGGTTTTCTTTCTTTCATTTTCACACAAAAAAAAGAAGCAGAAAGCTGACGGGAAAAGCACTCAATTACTAATAGTGGGAGAT	1503
504	TGCGGGCGTTATATGTATGTATGATTTCCATAAAACATATGTGACAACAACACAAAGTATTCATACGTGTTCTTGTGGTTTTCTTTTC	1593
594	TTGCATTTACACAAAAAAAAGAAGCAGAAAGCTGACGGGAAAAGCACTCAATTACTAATAGTGGGAGATTGCGGGCGTTATATGTAT -----onf1b/Hsp26	1683
584	GTATGATTTCCATAAAACATATGTGACAACAACACTCAAGTATTCCAAAGTAAAACCTTAAAGACAGAAACACGAAATAATGTACTTAATA -----onf2a/Hsp26	1773
774	AAGAGGAAAACCAGAATAAAAAAACTGACGTTTTGTTTTTTGTC 1818 ----- (A) <sub>n</sub> (major) -----onf1a/Hsp26	

*HspG1* SEQUENCE. Strain, *Oregon R*. The segment -1,196/1,400 is from GenBank: Accession, M26267 (DROHSP1). Downstream of the *Pst*I site, the sequence continues in the *Hsp26* Sequence. Several changes were introduced between 1,311 and 1,400 following R. S. Cohen (personal communication). The binding sites of *Hsp26* ovarian nuclear factors are indicated. Dashes underline bases that match the consensus hse sequence.

## *HspG2*

### Gene Organization and Expression

Open reading frame, 111 amino acids; expected mRNA length, 465-622 bases depending on which of the multiple polyadenylation sites and promoters are used, or approximately 2 kb when a polycistronic mRNA is made in response to heat shock. The corresponding RNA bands are observed in northern blots. Two transcription initiation sites were defined by primer extension analysis and



*HspG2*

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-489 GTGGAGTTTAACGGTTGTCTGCGCCCTTTTATAGAGACGGAAGAGCTTTGCCCATGCCACAGAGCTTTCTGGAGCAGCAACTCGTTG -4
      << -----Gene3
-399 TTTCTGTTGATTCTAGGGAGACAACTGGGAACCTTCTGGGGCCAAAGCTTTCGTAGACCGTAAACTGTTATATGTGATCTGCTTTAAGGTA -3
      - - - - - hse4 - - - - - hse3
                                     -->-252
-309 TGTACATACATTGTATGTATAAAGTGGGTACAGATAGCAAGCTCTGTATTGGAGATCATACCATAAGATTTAATTTTAAATTTCAAAGTG -2
      -----
-219 AAATCGGATACGGATGAGAGACGCAATCTCCAGTGTAGCTGGCAAACAAGACGACACCAGATAATCAAATGCGATAAGCAAGTACG -1
      -----
-129 GACATACAAATGTACATACCCGAATCTTTAATCTTGAACCTCATAAATGGATCATCTGCGCCAGCTGGCAAGTCAGTTGTTATTCAGCT -4
      hse2 - - - - - x - - - - - hse1
-39  GGCGAACCGGTTGAAATTCGTGCTCCGCCCATTAACAATGCCACGTACGAACAGGTTAAGGATGTTCCCAACCATCCGGATGTGTA 50
      MetAlaThrTyrGluGlnValLysAspValProAsnHisProAspValTy (17)
51  TCTTATCGACGTTTCGACGGAAGGAGCTCCAGCAGACGGGCTTCATCCAGCCAGCATCAATATACCCTGTAATAAACTACTTTCCCT 14
      rLeuIleAspValArgArgLysGluGluLeuGlnGlnThrGlyPheIleProAlaSerIleAsnIleProL (41)
141 AGTATTTGCTTTATTACCAATTTGTTTTATTACTATTTTTTACTTAGTGGATGAACGTGGACAAGGCTCTAAATCTGGATGGATCTGCTT 23
      euAspGluLeuAspLysAlaLeuAsnLeuAspGlySerAlaP (55)
231 TTA AAAACAAGTACGGAAGATCGAAACCGGAGAAGCAGTCGCCAAATCATATTCACCTGCCGGTCGGGAAATCGAGTCTTGGGAAGCAGAGA 32
      heLysAsnLysTyrGlyArgSerLysProGluLysGlnSerProIleIlePheThrCysArgSerGlyAsnArgValLeuGluAlaGluL (85)
321 AAATTGCCAAAAGTCAGGGATACAGCAAGTGAGCTTTAAAAGTTTATTATAGTGC AACTTTTATATCGGATACATATACATATGTATG 41
      ysIleAlaLysSerGlnGlyTyrSerAs (94)
411 CTCATTTTGTGTGTGATCTACAAAGGCTCCTGGAATGAATGGGCTCAAAGGAGGGACTTTAACGATAAAACGTCGCTATATTTCTGAA 50
      nValValIleTyrLysGlySerTrpAsnGluTrpAlaGlnLysGluGlyLeuEnd == (11)
501 TAAATGAAGATTAAATTAATTAATTAATTAATTAATTAATAGCTAAAAAAAACAGAAAACCTAAATTTATTTGTTAATATTAAAGCTGTAT 59
      --- | (A)n | (A)n | (A)n
591 TTTTCATATATCTCAAGTCTTGACTACGCCCATGGCAAGCTT 633
      -----HindIII

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*HspG2* SEQUENCE. Strain, *Oregon R*. Accession, X07311 (DROHSG2). In the first line, double underlining marks the inverse complement of the TATA box of *HspG3*. The sequence ends in the *HindIII* site located at -650 in the *Hsp22* Sequence. Dashes underline bases that match the consensus hse sequence. hse3 and hse4 are the same segments labeled hse2 and hse1, respectively, in *HspG3*. The x-mark under the TATA box marks a nucleotide that also belongs to hse1-2 (two partly overlapping hse's).

*HspG3* SEQUENCE (*opposite*). Strain, Schneider cell line 3. Accession, X06542 (DROHSPG3). The inverse complement of the *HspG2* distal TATA box is at -370/-365. Dashes underline bases that match the consensus hse sequence; hse1 and hse2 correspond to hse4 and hse3, respectively, of *HspG2*.

## HspG3

374 CCACTTTATACATACAATGTATGTACATACCTTAAAGCAGATCACATATAACAGTTTACGGTCTACGAAAGCTTGGCCCCAGAAAGTTTC -285  
 << -----Gene2 - - - - - hse2

284 CCAGTTGTCTCCCTAGAAATCAACGAAACACGAGTTGCTGCTCCAGAAAAGCTCTGTGGCAATGGGCAAAGCTCTCCGTCTCTATAAAA -195  
 - - - - - hse1 -----

194 GGGCGCAGACAAACCGTTAAACTCCACATTCGAGTCGGAAAAGTCAAGGTGAATTGTGCGCCAAACTGCAGCTGAGATTGTGGATTACACA -105  
 --->-167.

104 CCGCTGGCAAATCAACCCCTGGATACTTTTAAAGGAAAACAGGTCGTCGGTTCAACGAACTCTTCCGCCAGATACACAAGAAAGTTAA -15

-14 GCAAAGAAAAGTAAATGCCAGATATCCCTTTGTCTTGAATTTGGACTCCCGGACTCCATGTACTACGGCCACGATATGTTCCCGAAT :75  
 MetProAspIleProPheValLeuAsnLeuAspSerProAspSerMetTyrTyrGlyHisAspMetPheProAsn (25)

76 CGCATGTACAGCGGATTCATTGCGCGCAGCATCATGATCTTGATTTGCACACCTGGGCTGATTGCCCGGATGGGTGCACATGCCCAT 165  
 ArgMetTyrArgArgLeuHisSerArgGlnHisHisAspLeuAspLeuHisThrLeuGlyLeuIleAlaArgMetGlyAlaHisAlaHis (55)

166 CACCTGGTGGCCAAATAAAGGAACGGAGAGCTGGCTGCATTGAGCGCGGTGGAGCCTCAAATAAGCAGGGCAATTCGAGGTCCACTCG 255  
 HisLeuValAlaAsnLysArgAsnGlyGluLeuAlaAlaLeuSerArgGlyGlyAlaSerAsnLysGlnGlyAsnPheGluValHisLeu (85)

256 GATGTGGGACTTTTCAGCCAGGTGAACTGACCGTCAAAGTGGTCAACGAGTGCAATTTGGTGCAGGAAAAACACGAGGAGCCGAGGAC 345  
 AspValGlyLeuPheGlnProGlyGluLeuThrValLysLeuValAsnGluCysIleValValGluGlyLysHisGluGluArgGluAsp (115)

346 GATCATGGACATGTATCCCGCATTTTGTTCGGCCGATCCCGTGCCCAAGGAGTTCGATTCGGATGCCATTGTTCCACTTTGTCCGGA 435  
 AspHisGlyHisValSerArgHisPheValProAlaValSerAlaAlaGlnGlyValArgPheGlyCysHisCysPheHisPheValGly (145)

436 GGTGGAGTTCTCAATATCACGTTCCACCATTAGTTTCCAAGGAGGAGCTCAAGGAGCGCATATACCCATTAAGCATGTGGGTCCATC 525  
 GlyTrpSerSerGlnTyrHisGlySerThrIleSerPheGlnGlyGlyAlaGlnGlyAlaHisHisThrHisEnd (169)

526 GGTCTCTTCCAGGAATGGAAACGGTCATAAGGAGGCCGGTCCCGCAGCTTCTGCTTCAGAGCCAGAAGCCAAGTGAAGAGCCCCCTCT 615

616 AAAGATTGCAGCCTAAGCAGCCAAGTGATTTCCCAAGACTCTCGTTTATCGTTGCACCAAAAAAAAAAGTCCAAGAAAGTATCGCACAAAT 705

706 CGTATTATATTATTAATTTATTATTAGCTACATTTTAAACAGTCCAATCAAATTTTTAAGACTAATCGAAATCCAGTATTAATAAGGA 795  
 -----

796 ATATGAATGTCTCAGTAATCAAAGACTTTTACTAATATTTAAGAGCTTAATTCATATCAAAAAGCAGCAATCCAATTTTGGGTACAAT 885  
 |(A)<sub>n</sub>

886 ATTAACCTTTCCTTTGTTTCGATTAGACAGGTATTAAGCTGTGCATATTAATAATAGTCCCGGATGTCAATCTACTTAAAAAGCTT 975

976 TGGTTAGCCTTTCCAGGTGCGATTGAGTGAACCTTTGAACTTTGAAATGAAAGCCGCCATAAGTGAATATTCGATAGCTTTTAGTC 1065

1066 ATCTTTCCAAAACATCTATCGAAGTAAACAGTTTTTAAACAGTGGTAAAGTCAACGATAAATTTAATAAAAAGAACTAACATTTAATTACA 1155

1156 CAAAGTATATATATTTTTTAAAGTTATTTAGCAGATGGAGTACATTATAAACAATTTATTTGTTGGGATTATAATCTGAATAATA 1245

1246 AAAACCCGTGACATATTGCATGTTGCCATCTCCAGCTGGCAGTGAACCTCAAAAAAATGTTTGTGTTACTTTTGCGCCGCTCTGCAGT 1335

1336 TCATAATTCCTGCAAAATTAATCAGTAAACAGATTGCCAAGCCCGCTTCTAACACACCCCAACAATGCTCTGCACAACCACAATACGT 1425

1426 AAGTGGGAGCCTTTAAACCTACAGAATCATCACTATATTATGCCGAAAACCCCACTGATTTATGAAATTCGGTTGATTTTACAGCGCGG 1515

1516 CGGCATGGCGAGTTCGAATGGCAGGATCCAAGTCCACGGATGAAATGTAAGTCCCTAGAGAGAAGCTAATGTACACAATATAACCAAG 1605

cDNA sequences. Sequencing of multiple cDNA clones suggested three different 3' termini; at least one of which may be an artifact of cDNA cloning resulting from the presence of a stretch of As. Transcripts from the distal promoter have an intron between -135 and -64, and transcripts from both promoters have introns within the Leu-41 and Asn-94 codons (*HspG2* Sequence) (Pauli et al. 1988).

### *Developmental Pattern*

The distal transcript is testes specific; it appears first in early pupae and persists in adult males. The proximal transcript appears first in 7-h embryos, reaches a maximum at 10–12 h and persists through the second larval stage. It drops to very low levels in third instar larvae and adults, but rises to a second pronounced peak in early pupae.

Heat shock induces transcription from the proximal promoter but normal termination fails so that the *HspG2* heat-shock transcripts extend to the next polyadenylation site, that of *Hsp22* (*Hsp22* Sequence and Fig. 17.2). However, the amount of *Hsp22* transcript in the 2 kb RNA is a small fraction of that derived from the *Hsp22* promoter. Whether or not the polycistronic nature of this mRNA is functionally significant is not known. The two introns are properly excised. Of the genes in the cluster, *HspG2* is the least responsive to heat shock (Pauli et al. 1988).

### *Promoter*

There are 430 bp between the divergent, heat-inducible, transcription initiation sites of *HspG2* and *HspG3*, and there are four putative hse's in that region. Whether some hse's are allocated to one gene and some to the other or whether they are shared is not known.

## *HspG3*

### **Gene Organization and Expression**

Open reading frame, 169 amino acids; expected mRNA length, 979 bases, in agreement with the observed 1.0 kb major RNA. Upon induction, two minor RNA bands (1–2% of the major band) are also detectable; they are 1.6 kb and 2.3 kb long and appear to result from downstream extension of the major RNA. The site of transcription initiation was determined by primer extension, S1 mapping and the sequence of a cDNA clone. The polyadenylation site was obtained from the sequence of a cDNA clone. There are no introns (*HspG3* Sequence) (Pauli and Tonka 1987).

### Developmental Pattern

During embryogenesis, the expression of *HspG3* is first detectable at 7–8 h and reaches a peak at 10–12 h. No message is detectable through most of the larval period, but it reappears in the late third instar and peaks in early pupae. *HspG3* responds strongly to heat shock (Pauli and Tonka 1987).

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# 18

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## The *Hsp70* Gene Family: *Hsp70A7d*, *Hsp70A7p*, *Hsp70C1d1*, *Hsp70C1d2*, *Hsp70C1p*

### Chromosomal Location:

*Hsp70A7d*, *Hsp70A7p* 3R, 87A7  
*Hsp70C1d1/2*, *Hsp70C1p* 3R, 87C1

### Map Position:

3-[51]  
3-[51]

### Products

Heat-shock proteins of 70 kD, HSP70s, the most abundant type of heat-shock proteins.

### Structure

The sequence of *Drosophila* HSP70s is 70–80% identical to heat-shock proteins of groups as distant as vertebrates and vascular plants (Fig. 18.1), and some of the properties discussed below are from studies in other organisms. The different members of the *Drosophila* HSP70 family are no less than 97% identical. Two distinct regions have been identified in these proteins. The more highly conserved region is near the N-terminus; it contains an ATP-binding site and has weak ATPase activity. The other region, closer to the C-terminus, is more variable, and it has sites important for nucleolar localization. It has been suggested that a hydrophobic pocket on the protein surface is the site of HSP70 binding to hydrophobic residues of partly denatured proteins (Lindquist and Craig 1988; Schlesinger 1990 and references therein).

### Function

Organisms subjected to mildly elevated temperatures become more tolerant of subsequent high-temperature exposure. It has been suggested that HSP70 may be capable of preventing the denaturation of cellular proteins; and, with the

```

Hsp70-C1 ...M          Y      N Y          S  N EP          R          YD KIAE          VS . G
Hsp70-A7 ...M          Y          S  EP          R          YD KIAE          VS . G
Pig  MAKSV          F          S  T  DA          L Q          FG VVQG          R IN . D
Petunia ..EG          W DR          G  T  DA          I          RFS SVQS I L          IPGP D
CON  ---PAIGID LGTTYSCVGV -QH GKVEIIA NDQGNRTTPS YVAFTD-ERL IG--AKNQVA MNP-NTVFDA KRLIGRK--D P----DMKHW PFKV--D-G-
1                                     50                                     100

Hsp70-C1      G E   S R          T XX  ES TD          H          L   N L ..
Hsp70-A7      G E   S R          T A   ES TD          H          L   N L ..
Pig   VQ S   T G Y          I . G  HP VSN          V          I   RT G ..
Petunia  M V T   E Q A          I .   TT KN V          V   M          I   K ASSA K
CON  KPKI-V-YKG E-K-FAPEEI SSMVLTKMKE -A-EAYLG-- I--AVITVPA YFNDSQRQAT KDAG-IAGLN VLRIINEPTA AA-AYGLDK- -K--GERNVL
101                                     150                                     200

Hsp70-C1          S L RS          T LAE  Y LRS          A          E   A   Q
Hsp70-A7          S L RS          T LAD  Y LRS          A          E   A   Q
Pig           D . I KA          N FVE  H YSQ K V          C          Q SL  S   I
Petunia          L E . I KA          M N FVQ  N ISG          C          TAQT  S Y I S
CON  IFDLGGGTFD VSILTIDEG- -FEV--TAGD THLGGEDFDN RLV-H---EF KRK-KKD--- NPRALRRLRT A-ERAKRTLS SST-ATIEID -LFEG-DFYT
201                                     250                                     300

Hsp70-C1. KVS          AN  N Q          N   G  I          S  E H  N  L          Q  G I  V  V
Hsp70-A7  KVS          A  N  Q          N   G  I          S  H  N  L          Q  G I  V  V
Pig  SIT          S  S  E          R  L  A  L          K  N  RD  K          M  K  ENV  L  L
Petunia TIT          NM  KCME          C R          SSV  V          Q  N  E  CK          EGN  E  V  L  L
CON  ---RARFEEL C-DLFR-TL- PVEKAL-DAK MDK-QIHD-V LVGGSTRIPK VQ-LLQDFF- GK-LN-SINP DEAVAYGAHV QAAILSGD-S -K-QD-LL-D
301                                     350                                     400

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Hsp70-C1      I      K E CR C  KT      S      A T D      L      KEM
Hsp70-A7      I      K E CR C  KT  A    S      A T D      L      KEM
Pig           L      A K ST T  QI T    L      R L R E      I      T TDK
Petunia      T  L      G V P  TT T KE QV    L      R L K E      T C I      EDKT
CON  VAPLSLG-ET AGGVMT-LI- RN--IP-KQT --FSTYSDNQ PGV-IQVYEG ERAMTKDNN- LG-F-LSGIP PAPRGVPQIE VTFD-DANGI LNVSA---ST
401                                     450                                     500

Hsp70-C1      KN K      QA D N  AD KH Q  ITR      VF V QS Q AP.A D  NSV  N T R  S T E D
Hsp70-A7      KN K      QA D N  AD KR Q  TSR      H VL V QA Q AP.A D  NSD  N DT R  S T E D
Pig           NK T      KE E Q  KA IQ E  GAK      AF M SV D EGLK IS  KKV  Q V S  A L D E
Petunia      QKNK T      KE E Q  KS ELKKK EAK      N AY MRNTIKD DKINSQ SA  KRIE AID A K  N Q L AD ED
CON  GKA--ITI-N DKGRLS--EI -RMV-EAEKY --EDE--R-R V---NALESY --N-K--VE- ----GKL-EA DK---LDKC- E-I-WLD-NT -AEK-EF-HK
501                                     550                                     600

Hsp70-C1      ME TRH S  MT H Q  AAG .P N CGQQAG FGG YS  V  *
Hsp70-A7      LE TRH S  MT H Q  AGA GP N CGQQAG FGG YS R V  *
Pig           RK EQV N  ISGLY GAG PGP GF P DLKGG S.. .. I  .
Petunia      MK ESI N  IA Y G  GATMDEDGP SVGGSA SQT GA  KI  *
CON  --EL---C-P I--KM-Q-GA G---G--GA- -----G---- --GPT-EEVD -
601                                     650

```

FIG. 18.1. Comparison of HSP70s from *Drosophila*, the pig (Accession, M69100) and *Petunia* (Accession, X13301). The CON(sensus) line indicates positions at which all four sequences agree. Where there is no such agreement, the residue occupying that position in each sequence is indicated. There is 89% overall identity between Hsp70A7 and the porcine sequence. Sequences aligned with the GCG *Pileup* program.



expenditure of ATP, it may be involved in the renaturation of denatured proteins and the dissociation of abnormal protein complexes. HSP70 is related to other non-heat-shock proteins known as “molecular chaperones”. Molecular chaperones are involved in the translocation of proteins across membranes and also seem to have a role in controlling denaturation and renaturation of proteins (Schlesinger 1990; Gething and Sambrook 1992).

### *Tissue Distribution*

HSP70 is present at low levels in untreated flies. During heat stress, *Hsp70* transcription increases, and HSP70 becomes prominent in the nucleus and the nucleolus where it forms insoluble complexes. After return to normal temperatures, HSP70 levels remain high for some time, but the protein returns to the cytoplasm (Velazquez and Lindquist 1984; Schlesinger 1990).

### **Organization and Expression of the Clusters**

The two *Hsp70* genes at 87A7 are separated by a 1.6 kb spacer, and they are divergently transcribed. At 87C1, a centromere proximal gene, *Hsp70C1p*, is separated from two centromere-distal genes, *Hsp70C1d1* and *Hsp70C1d2*, by 40 kb of DNA. The distal genes are tandemly transcribed toward the telomere while the proximal gene is transcribed in the opposite direction. None of the *Hsp70* genes have introns (Fig. 18.2).

A large portion of the spacer between the proximal and distal copies at 87C1 is made up of simple sequences designated alpha, beta and gamma; these are arranged in various repeat patterns. The gamma element includes a copy of the *Hsp70* regulatory region and, in response to heat shock, it promotes transcription of the spacer sequences. As far as is known, the spacer transcripts have no coding capacity and are non-functional (Ish-Horowitz and Pinchin 1980; Hackett and Lis 1981).

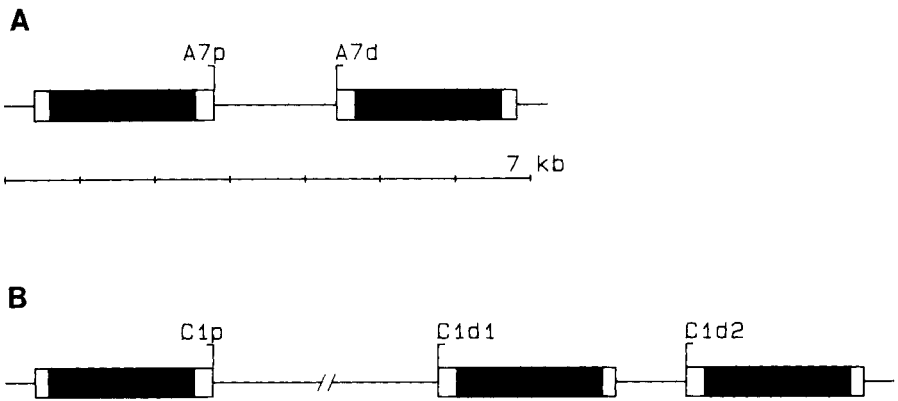


FIG. 18.2. Organization of the *Hsp70* clusters. (A) Cluster at 87A7. (B) Cluster at 87C1.

In *D. simulans* and *D. mauritiana*, there are only four *Hsp70* genes; that is, two divergently transcribed genes occur at each of two loci corresponding to 87C1 and 87A7. Thus, it appears that duplication of the distal gene at 87C1 and multiplication of the simple spacer sequences are recent events unique to *D. melanogaster* (Leigh-Brown and Ish-Horowitz 1981; see reviews in Schlessinger et al. 1982).

All copies of *Hsp70* are very similar, especially in the coding and 5' regions. In a segment that extends from -610 to 1 (the first codon), the various genes present the following frequencies of base substitution, addition, or deletion relative to *Hsp70C1d1*: *Hsp70C1d2*, 0.5%; *Hsp70C1p*, 1.4%; *Hsp70A7d*, 6.5%. In contrast, at the 3' end of the genes, *Hsp70C1d1* is much more similar to *Hsp70C1p* than to *Hsp70C1d2*. Within the segment -610/1 *Hsp70A7d* and *Hsp70A7p* differ by 3%, but further upstream the two sequences appear unrelated. Because sequence similarities and repeats occur in blocks having no apparent functional significance, it has been suggested that much of the sequence conservation of the *Hsp70* genes may be due to intergenic corrections rather than negative selection against deleterious mutations (Török et al. 1982).

### *Developmental Pattern and Promoter*

Transcription of the *Hsp70* genes in *Drosophila* occurs only in response to heat and other stressful conditions usually associated with protein denaturation; i.e., no developmentally related expression occurs (Mason et al. 1984).

Most studies of transcription regulation were carried out with *Hsp70A7* promoter sequences. However, given the great deal of sequence conservation in the promoter regions, the available information about transcriptional regulation probably applies equally to all *Hsp70* genes. As is true for other *Hsp* genes, the heat-shock response seems controlled by the heat-shock element (hse) consensus sequence CTNGAANN TTCNAG that must be present in at least two adjacent copies (Pelham 1985; Bienz and Pelham 1987).

Germline transformations involving 5' deletions demonstrated that the 97 bp upstream of the transcription initiation site (to coordinate -348), a segment that includes hse1 and hse2 (*Hsp70* Sequences), are sufficient for normal levels of heat-induced transcription (an approximately 100-fold increase as compared to the uninduced state) (Dudler and Travers 1984). Thus, these two hse's seem to be the main functional regulatory elements. Repositioning a 51-bp segment that includes the two hse's at various distances from the TATA box does not affect expression very much (Simon and Lis 1987). This flexibility contrasts with the sequence conservation noted earlier.

More detailed studies involving *in vitro* mutagenesis, germline transformation, and *in vitro* binding assays led to a reassessment of the sequence elements responsible for heat induction. The conclusion from those studies is that hse's possess alternating repeats of the 5-bp unit NGAAN and its reverse complement, NTTCN. There are three or four such units in the hse's of the *Hsp70* genes (Xiao and Lis 1988; Perisic et al. 1989).

Transcription is activated by a heat-shock transcription factor (HSF). HSF

*Hsp70-C1d1* and *Hsp70-A7d*

A7d TGTCA AG TCCATAGGCC----- C G GACAAC C - - C C T A C  
 C1d1 TCAGACATTATTGGTTTAGAAGCGCAGTATTTTTTGGCA----ATACGCATAACAAGCGCTTCGATTATCTTTAACATAAGTTAT -568

TATATATATAAATAAA C G

-567 TTAAGCAGCCGTATTTATAAAGAAATTTCCAAAATAAAGC-----GAATATTCTAGAATCCCAAACAAA-CTGGTTATT -478  
 \*\*\* \*\* hse4  
 |- f4 -|

C CG - C TC -----G T A  
 -477 GTGGTAGGTCATTGTTGGCAGAAAGAAAACCGAGAAATTTCTCTGGCCGTTATTCGTTATTTCTCTCTTTCTTTTGGGCTCTCCC -388  
 \*\* \*\* hse3  
 |- f3 -| -| GAGA -| -|

T T G CT AT C  
 -387 TCTCTGCACTAATGCTCTCTCACTCTGTCCACAGTAAACGGCATACTGCTCTCGTTGGTTTCGAGAGAGCGCCCTCGAATGTTCCGGAA -298  
 \* \* \*\* \*\* hse2 \*\*\* \*\* hse1  
 GAGA -| -| f2 -| -| f1  
 -| GAGA -|

G T -->-251 A G --  
 -297 AAGAGCGCCGGAGTATAAATAGAGGCGCTTCGTCTACGGAGCGACAATTCAAATCAACAAGCAAAAGTGAACACGCTCGCTAAGCGAAAGC -208  
 -| -----

G C - A A  
 -207 TAAGCAATAAACAAGCGCAGCTGAACAAGCTAAACAATCTGCAGTAAAGTCAAGTAAAGTGAATCAATTAAGTAAACGACGACCA -118

TTAAACT AA AA C G GTC  
 -117 AGTAA-----ATCAACTGCACTACTGAAATCTGCCAAGAAGTAAATATTGAATACAAGAAGAGAAGTCTGAATACTTTCAACAA--- -28

G --- A  
 -27 GTTACCAGAAAGAAGAACTCACACACAATGCCTGCTATTGGAATCGATCTGGGCACCACCTACTCTCGTGGGTGCTACCAGCATGG 62  
 MetProAlaIleGlyIleAspLeuGlyThrThrTyrSerCysValGlyValTyrGlnHisG1 (21)

G T A C T C C  
 63 CAAGGTTGAGATTAACGCCTATGACCAGGGCAACCGCACCACGCCCTCTACGTGGCTTTCACAGACTCGGAACGCCTCAATGGTGAACC 152  
 yLysValGlnIleAsnAlaTyrAspGlnGlyAsnArgThrThrProSerTyrValAlaPheThrAspSerGluArgLeuAsnGlyGluPr (51)  
 Ile Asn Ile

C G  
 153 GGCCAAGAACCAGGTGGCCATGAACCCAGAAACACAGTGTGTTGACGCCAAGCGACTCATCGGCCGAAAATACGACGATCCCAAAATCGC 242  
 oAlaLysAsnGlnValAlaMetAsnProArgAsnThrValPheAspAlaLysArgLeuIleGlyArgLysTyrAspAspProLysIleAl (81)

G G C  
 243 AGAGGACATGAAGCACTGGCCTTTCAAAGTTGTAAGCGATGGCGGAAAGCCCAAGATCGGGGTGGAGTATAAGGGTGAGTCCAAGAGATT 332  
 aGluAspMetLysHisTrpProPheLysValValSerAspGlyGlyLysProLysIleGlyValGluTyrLysGlyGluSerLysArgPh (111)

C C CCG A A C  
 333 TGCTCCCAGGAGATCAGTTCGATGGTCTGACCAAGATGAAGGAGACGGCGG---AGGCGTATCTGGCGAGAGCATCACGGATGCAGT 422  
 eAlaProGluGluIleSerSerMetValLeuThrLysMetLysGluThrAlaG---luAlaTyrLeuGlyGluSerIleThrAspAlaVa (141)  
 AlaGlu

423	CATCACAGTTCACGCTTACTTCAACGACTCTCAGCGCCAGGCTACCAAAGACCCGGTCACATCGCCGGCTGAATGTGCTCCGCATCAT 11leThrValProAlaTyrPheAsnAspSerGlnArgGlnAlaThrLysAspAlaGlyHisileAlaGlyLeuAsnValLeuArgle11	512 (171)
513	CAATGAGCCACGGCGGACAGCTTGGCTACGGACTGGACAAGAATCTCAAGGGTGAGCGCAATGTGCTTATCTTCGACTTGGCGGCGG eAsnGluProThrAlaAlaLeuAlaTyrGlyLeuAspLysAsnLeuLysGlyGluArgAsnValLeuIlePheAspLeuGlyGlyG1	602 (201)
603	CACCTTCGATGTCTCCATCCTGACCATCGACGAGGGATCTCTGTTCGAGGTGCGCTCCACAGCCGGAGACACACTTGGCGGCGAGGA yThrPheAspValSerIleLeuThrIleAspGluGlySerLeuPheGluValArgSerThrAlaGlyAspThrHisLeuGlyGlyGluAs	692 (231)
693	CTTTGACAACCGCTAGTCACCCACCTGGCGGAGGAGTTCAAGCGCAAGTACAAGAAGGATCTGCGCTCCAACCTCGCGCCCTACGCG pPheAspAsnArgLeuValThrHisLeuAlaGluGluPheLysArgLysTyrLysLysAspLeuArgSerAsnProArgAlaLeuArgAr Asp	782 (261)
783	CCTCAGAACAGCAGCTGAACGGGCAAGCGCACACTCTCCTTAGCAGCGAGGCCACCATCGAGATCGACGCATTGTTGAGGGCCAAGA gLeuArgThrAlaAlaGluArgAlaLysArgThrLeuSerSerSerThrGluAlaThrIleGluIleAspAlaLeuPheGluGlyGlnAs	872 (291)
873	CTTCTACACAAAGTAAGCCGTGCCAGGTTTGAGGAGCTGTGCGCAACCTCTCCGCAACACCCGTCAGCCTGTGGAGAAGGCCCTCAA pPheTyrThrLysValSerArgAlaArgPheGluGluLeuCysAlaAsnLeuPheArgAsnThrLeuGlnProValGluLysAlaLeuAs Asp	962 (321)
963	CGATGCCAAGATGGACAAGGGTCAGATCCACGACATCGTGCTGTCGCGGATCCACTCGCATTCCAAGGTGCAAAGTCTGCTGCAGGA nAspAlaLysMetAspLysGlyGlnIleHisAspIleValLeuValGlyGlySerThrArgIleProLysValGlnSerLeuLeuGlnG1 As	1052 (351)
1053	GTTCTTCCACGGCAAGAACCTCAACCTATCCATCAACCCAGACGAGGCGAGTGGCATACGGAGCTGCTGTGCAAGGCCGCTATCCTCAGCGG uPhePheHisGlyLysAsnLeuAsnLeuSerIleAsnProAspGluAlaValAlaTyrGlyAlaAlaValGlnAlaAlaIleLeuSerG1 P	1142 (381)
1143	AGACCAGAGCGCAAGATCCAGGACGTGCTGCTGGTGGACGTGGCCCCACTTTCATTGGGAATTGAGACCCTGGAGGTGAATGACCAA yAspGlnSerGlyLysIleGlnAspValLeuLeuValAspValAlaProLeuSerLeuGlyIleGluThrAlaGlyGlyValMetThrLy	1232 (411)
1233	GCTGATCGAGCGCAACTGTGCAATCCGTCGAAGCAGACTAAGACGTTCTCCACGTACTCGGACAACCAGCCGGAGTCTCCATCCAGGT sLeuIleGluArgAsnCysArgIleProCysLysGlnThrLysThrPheSerThrTyrSerAspAsnGlnProGlyValSerIleGlnVa Ala	1322 (441)
1323	GTATGAGGGCAACGTGCGATGACGAAGGACAACAATGCATTGGGCACCTTCGATCTGTCGGCATCCACCTGCACCAAGGGGTGTC 1TyrGluGlyGluArgAlaMetThrLysAspAsnAsnAlaLeuGlyThrPheAspLeuSerGlyIleProProAlaProArgGlyValPr	1412 (471)
1413	CCAGATAGAAGTAACCTTCGACTTGGACGCCAATGGAATCCTGAACGTCAGCGCCAAGGAGATGAGTACGGGCAAGGCCAAGAATCAC oGlnIleGluValThrPheAspLeuAspAlaAsnGlyIleLeuAsnValSerAlaLysGluMetSerThrGlyLysAlaLysAsnIleTh	1502 (501)

(continued)

1503 GATCAAGAACGACAAGGGACGCCTCTCGCAGGCCGAGATTGATCGCATGGTGAACGAGGCTGAGAAGTACGCCGACGAGGACGAAAAGCA 159;  
 rIleLysAsnAspLysGlyArgLeuSerGlnAlaGluIleAspArgMetValAsnGluAlaGluLysTyrAlaAspGluAspGluLysHi (531)  
 Ar  
 AG C C CC T G G A A T  
 1593 TCGCCAGCGCATAACCTCTAGAAATGCTCTGGAGAGCTACGTATTCAACGTAAGCAGTCCGTGGAGCAGGCCCGCTGGCAAACCTGGA 168;  
 sArgGlnArgIleThrSerArgAsnAlaLeuGluSerTyrValPheAsnValLysGlnSerValGluGlnAlaProAlaGlyLysLeuAs (561)  
 g Val His Leu Ala  
 T A C C G T  
 1683 CGAGGCCGACAAGAACTCCGCTTGGACAAGTGAACGAAACTATTCGATGGCTGGACAGCAACACCACCGCCGAGAAGGAGGAGTTCTGA 177;  
 pGluAlaAspLysAsnSerValLeuAspLysCysAsnGluThrIleArgTrpLeuAspSerAsnThrThrAlaGluLysGluGluPheAs (591)  
 Asp Asp  
 C C C T T GA CT GGT  
 1773 CCACAAGATGGGAGAGCTCACTCGCCACTGCTCCCCTATCATGACCAAGATGCATCAGCAGGGAGCGGGAGCAGCTGGGGGT---CCGGG 186;  
 pHisLysMetGluGluLeuThrArgHisCysSerProIleMetThrLysMetHisGlnGlnGlyAlaGlyAlaGlyGly---ProGl (621)  
 Leu GlyAla Gly  
 A C G G A G G G G TCTAAT TT  
 1863 AGCCAAGTGTGGCCAACAGGCCGAGGATTTGGCGGCTACTCTGGACCCACAGTCGAGGAGGTCGACTAAGCCAAATAGAAATTATTTCA 195;  
 yAlaAsnCysGlyGlnGlnAlaGlyGlyPheGlyGlyTyrSerGlyProThrValGluGluValAspEnd (643)  
 Arg  
 ATCAA GG A A C TA GGT ATA AA T TTTA GTTTTGAG CTG T AG A GT T GATCGA A CCA  
 1953 GTTCTGGCTTAAGTTTTAAAAGTGATATTATTTATTTGGTGTGAACCAACCAAAAGAATGTAATAACTAATACATAATTATGTAGTT 2042  
 AG CAACAAT GT T ACC AA TA C AG CTTAATT A CAA ATGT TTGCT AG AAA TA ATTA TTA G AAT T  
 2043 TTAAGTTAGCAACAAATGATTTTAGCTATATTAGCTACTTGGTTAATAAATAGAATATATTTATTTAAAGATAATTCGTTTTTATTGTC 2132  
 -----(87C1) -----(87A7)  
 AA TCAACT  
 2133 AGGGAGTGAGTTTGCTTAAAACCTGTTTAGATCTGTCTCGAGAAATTATTTATTTAAATGCGATGGAGAGCCGGCCGCAATCGAAAA 2222  
 |(A)<sub>n</sub> (87A7)  
 2223 CTTTACGCGCTTAAAAGCACGAGTTGGCATCCCTAGTAACAGCTGTTTCGTGAAGATATGCAGTCAAACGAAAAACCCGCTACAAATA 2312  
 2313 TTGTTATTTTGATTAGATTACGGATTACAGAATGGAACCGCCGTTCCGCCCGCTAAGTGAGTCTGCACCAAGGCGTGGCGACAGGTGT 2402  
 2403 ACGAGAAATGTAAGCTGGCCTCGCAGGAGATCCGTCATCCCAATTGGGAAATGTAATCTTTGCCAGAAATGGTTACGGAGTTCAACAACA 2492  
 2493 AAACAGTCTATAGAAATAATAGCCTTTCCTTCTCATATGTATGTAATATGTAATAAAGTCACAACAAATTTCTAATACACTTCTCA 2582  
 2583 GTCTAAATTAATTTTATCGTATATTAACACAGAAGAAAGTCCGTTAATCGTTGATTCGTTAACTAAAAGTACAAAATAATCTTTAATC 2672  
 | Ca. coordinate -640 of Hsp70-C1d2  
 2673 TTTAGAAGCGCAGCAATGTT 2692

*Hsp70* SEQUENCES. Accession, J01104, J01105 (*DROHSP7D1*) and J01103 (*DROHSP7A2*). The numbered line shows the sequence of *Hsp70C1d1*; where the sequence of *Hsp70A7d* differs, the changed bases are indicated above and the amino acid substitutions below the *Hsp70C1d1* sequence. Dashes represent gaps in one sequence relative to the other. Asterisks below the sequence mark positions that match the hse consensus.

has an apparent  $M_r$  of 110 kD and binds with high affinity (dissociation constant,  $4 \times 10^{-12}$ ) to two contiguous segments designated f1 and f2 in the *Hsp70* Sequences; these binding sites extend from  $-315$  to  $-290$  and from  $-340$  to  $-315$ , respectively (between 40 and 90 bp upstream of the transcription initiation site). Two additional binding sites occur farther upstream, at  $-440$  to  $-415$  (f3) and at  $-510$  to  $-485$  (f4). The binding of HSF to these secondary sites has a minor effect on *in vitro* transcription; it is not clear what their *in vivo* role might be. All the binding sites overlap hse's (Wu et al. 1987; Topol et al. 1985).

HSF seems to preexist as an unbound monomer in all cells. In response to heat, it is reversibly changed to the active form capable of specific DNA binding; that change includes the formation of oligomers (Westwood et al. 1991). Heat treatments as short as 30 s are sufficient to induce detectable binding of HSF to *Hsp70* promoter fragments (Zimarino and Wu 1987). Binding of HSF to hse is highly cooperative, and the cooperativity is itself temperature-dependent (Xiao et al. 1991).

At normal temperatures, RNA polymerase II binds to the region around the transcription initiation site of *Hsp70* (coordinates  $-186$  to  $-263$ ), and transcription is initiated but blocked. It is only after heat shock, and presumably after binding of HSF, that the transcription block is released and RNA polymerase II becomes detectable along the whole length of the gene (Gilmour and Lis 1985, 1986; Rougvié and Lis 1988).

Another factor, a 66 kD protein, seems to associate with the segments of alternating CT (or GA) sequence found between positions  $-415$  and  $-360$  and between  $-325$  and  $-319$ . This same protein, the GAGA, factor binds sequences upstream of the histone genes *His3* and *His4*, the heat shock gene *Hsp26* and *Ultrabithorax* (Gilmour et al. 1989).

*Hsp70* mRNA is very stable and efficiently translated at  $36^\circ$ , but has a half-life of only minutes at  $25^\circ$ . This insures that when *Drosophila* flies or cells are returned to  $25^\circ$  after a heat shock, HSP70s cease to be synthesized. AU-rich sequences in the 3' untranslated region of the mRNA are responsible for the specificity of temperature-dependent degradation of *Hsp70* mRNA (Petersen and Lindquist 1989).

### *Hsp70A7d* (Distal gene at 87A7)

#### Gene Organization and Expression

Open reading frame, 643 amino acids, expected mRNA length, 2,389 bases. Primer extension and S1 mapping were used to define the 5' end. The 3' end was obtained by S1 mapping (*Hsp70* Sequences) (Karch et al. 1981; Török and Karch 1980; Török et al. 1982).

The transcription initiation site of *Hsp70A7p* (the proximal gene at 87A7)

is approximately 1,630 bp upstream of the *Hsp70A7d* transcription initiation site. Only a few hundred bp at the 5' and 3' ends of *Hsp70A7p* have been sequenced. Assuming conservation of the intervening coding region, the segment of sequence similarity between *Hsp70A7d* and *Hsp70A7p* extends approximately from coordinates -600 to 2,130 (*Hsp70* Sequences and Fig. 18.1). This would leave, between the inverted repeats, a spacer of approximately 940 bp made up largely of blocks of simple sequence DNA (Mason et al. 1982). The two genes do not seem to share regulatory elements, i.e., each has its own *cis*-acting hse's.

### *Hsp70C1d1* (first distal gene at 87C1)

#### Gene Organization and Expression

Open reading frame, 641 amino acids; expected mRNA length, ca. 2,360 bases. Primer extension and S1 mapping were used to define the 5' end. The 3' end was not defined (*Hsp70* Sequences) (Ingolia et al. 1980; Karch et al. 1981).

The repeat containing *Hsp70C1d2* (the second distal gene at 87C1) begins 576 bp downstream of the *Hsp70C1d1* termination codon (ca. coordinate 2510). The two genes are part of a tandem duplication of approximately 2,900 bp from coordinates -820 to 2,080 of *Hsp70C1d1*. Such alignment leaves a spacer of approximately 430 bp between the repeats (from 2,080 to 2,510) (*Hsp70* Sequences and Fig. 18.2); it is not clear whether this spacer originated at the time of the duplication or whether it arose by sequence divergence at one or both ends of the repeat. *Hsp70C1d2* has been only partially sequenced but it appears very similar to *Hsp70C1d1*. The most extensive sequence divergence occurs within the last 100 bp at the 3' end, especially around the polyadenylation signals (Török et al. 1982).

The region of sequence similarity between *Hsp70C1d1* and *Hsp70C1p* (for which only the 5' and 3' end-sequences are available) starts near coordinate -600 and extends to coordinate 2,540. The overlap of *Hsp70A7* with *Hsp70C1p* sequences extends from -600 to 1,940.

#### Related Genes

*Hsp68* at chromosomal location 95 D is a heat-shock gene related to the *Hsp70* family. Hybridization data indicate that *Hsp68* and *Hsp70* are 75–85% identical.

Seven other genes identified by cross-hybridization are the *heat shock cognate genes*, *Hsc1–Hsc7*. They are expressed very strongly during development but are not heat-inducible or clustered (Lindquist and Craig 1988).

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## *janus: janA, janB*

**Chromosomal Location:**  
3R, 99D4-8

**Map Position:**  
3-[101]

### **Products**

The properties and functions of *janus* products are unknown. Allowing for nine gaps, there is 37% sequence identity between JANA and JANB.

### **Organization of the *janus* Cluster**

*janus* is a small but complex locus that includes two partly overlapping transcription units: *janA* is upstream and its 3' untranslated region contains the transcription initiation site of *janB* (*jan* Sequences and The *Serendipity* Gene Cluster Fig. 28.1) genes probably originated by duplication, judging from their sequence similarities and the comparable positions of two of the three introns in each gene (Yanicostas et al. 1989).

### *janA*

#### **Gene Organization and Expression**

The structure of all the transcription products is not yet clear; a final description will be possible only after more cDNA sequences become available. There are two initiation sites, 18 bp apart, two polyadenylation sites, 5 bp apart, and a facultatively spliced intron that spans part of the leader and part of the coding region. The open reading frames are 119 and 135 amino acids long, and the expected mRNA length is between 638 and 731 bases depending on which 5' and 3' ends occur and whether or not the leader intron is spliced. These sizes are in agreement with a 0.8 kb band observed in RNA gels (but see below). Primer extension and a cDNA sequence were used to define the upstream 5'

*janA* and *janB*

-351	ATTCGGCTTAAACAATTTAATTTGTGTATATTTTGTGTGAACGCCAGAGCTGTGCCGATAGTGCCGATAGTATCGACTGCGTGTCTCG	-262
	<-- Sry-beta	
-261	GCGTAATCGATAAATTTGCTGTCACGTATAACAACGTTTTCTTTTTGAGTTTATTAATTTACTAAAAATACTGAGTAGTACAAAAA	-172
	-----	
-171	CGTTTTCCCAATGTACTAAAGAAATACTGAATATATAATTTTAAATAGTATCGATACATAAGGTGAACGAGAATAAAAGTATCTGGTC	-82
	-----	
	-->-81 (janA)      -->-63	
-81	ACATTGCTGGACTAAAGCAGCGTTTTTGGAAAAATTTGCCGGTGGTAAGACATTAATTTCTGTTTTCAAACACTTTTCCACAATGAATCG	8
	MetAsnAr	(3)
9	CCTCCAACGTCTTCCAAAGGACTACGACTGATTCACAAAAATGTCGGAGGAAGCACTTGCCGGCTGCCACTGGTGACATCAGTCCAGA	98
	gLeuGlnLeuLeuSerLysGlyLeuArgLeuIleHisLysMetSerGluGluAlaLeuAlaGlyValProLeuValHisIleSerProG	(33/17)
99	GGGCATCTTCAAGTATGTCATGATCAATGCTCTCGATGGAGGAGATGCTTCAAAGGCGGTGATCCGCGGATTGCGGACTGCACATGGCA	188
	uGlyIlePheLysTyrValMetIleAsnValPheAspGlyGlyAspAlaSerLysAlaValIleArgGlyPheAlaAspCysThrTrpHi	(63/47)
189	TGGTAAGTCGGATCCTCATCACCATCAAGTCCCACTTAGCTTGGTTACTGTCCACAGCCGACATCTCGAGCGCAGGAGGAGGCT	278
	sA	IaAspIlePheGluArgGluGluGluValP
		(74/58)
279	TTAAAAAAGTGGGCTGCGGGCCGAGTGTCTGGCGGGGTGCGATTGAACACAATCCCAGAGAAGAAGTACTTGAAGGTCTACGGATACT	368
	heLysLysLeuGlyLeuArgAlaGluCysProGlyGlyGlyArgIleGluHisAsnProGluLysLysTyrLeuLysValTyrGlyTyrS	(104/8)
369	CGCAGGTGGGTCTATTCTTGGATAAAGGGTGCCTGGGCAGTGGATGGACTGATGTATCTAACTACTTGAAAAATTTCTGGAGTCTA	458
	erGln	(105/8)
459	ATCAAGTCTTCTATTTAAGGGCTTTGGAAAAGCTGATCAGCGCAGACCAAACGCATCTTGGCCACCAAAATACCCGGACTACACGATCG	548
	GlyPheGlyLysAlaAspHisAlaGlnThrLysArgIleLeuAlaThrLysTyrProAspTyrThrIleG	(129/1)
549	AAATCTCCGATGAGGGATATTAGTGCATCAACGAGAGAAGACTCCACATAAGCACACTGACATGAATTTATACCATTGGCTTCGATCC	638
	IuIleSerAspGluGlyTyrEnd	(135/1)
	-----	
	-->696 (janB)	
639	TGTGTGCCATGATTTTATTGGAAATGGCATTAAAATGGAGAAATACTCTGAAAGGCAGTTAGTCTGTAGCTTTGCAACTGCTCGCACTA	728
	-----	
729	AACCTTTTCGGATCTAAATTAATCAGTTGTACACAAATTTCTGTTCTTTCTTTTGGTTAAATAAAATGAAAATGTTCAAGTCATTGCG	818
	MetLysMetPheLysSerLeuAr	(8)
	(A) <sub>n</sub>  (A) <sub>n</sub> (janA)	
819	TCTGTTCTCATATGTTTCTCGTTTCGTAAGGCTTAGGAATATCAATATTAAGATTACAAGCCCTAATATACTTGGTTTTAGAAA	908
	gLeuLeuProHisIleValSerProPheG	lnL
		(19)
909	AATGTTACTCAACGATTTGATAAGTTTGGTAGGCGTCCCCGGGTCAAGATAACCAAGGGTCAGAATCGTATTGTTGGTGAAATTC	998
	ysCysTyrSerThrAspLeuIleSerLeuValGlyValProArgValLysIleThrLysGlyGlnAsnArgTyrLeuLeuValAsnIleH	(49)
999	ATACGCATGGCTTACGAAATGGAAGATTATTGTCGGTGGCCGATGTTGACAATCACTGTGAGTTCCACTGCTGGACGCTTAAC	1088
	isThrHisGlyPheThrLysTyrGlyArgValIleValArgGlyAlaAspValAspAsnHisL	(70)
1089	CTTGAGCAGTCTTACAAATCCTTCTTTCAGTGGCGGTCTTCGACTCGATTTTGGAGGAGCTGGAACCCGAGGGCATATGTCGCAAAATCC	1178
	euAlaValPheAspSerIleLeuGluGluLeuGluProGluGlyIleCysAlaLysIleL	(90)

1179	TCGGTGGTGAAGGATTCTCAACGAGGCAGAAAATAAAAAAATTAAGATCTATGGCACCTCCAGGGTAAGTAGAGGATCCTTGGTCCTTG euGlyGlyGlyArgIleLeuAsnGluAlaGluAsnLysLysIleLysIleTyrGlyThrSerArg	1268 (111)
1269	AAGCACC GGCTAATGGTCTTGTATGGGCTCCCTAGACTTTCGGCGGTGCTGATCACACAAGGACAAGGAATATACTTCAAGCGTGGACC ThrPheGlyGlyAlaAspHisThrArgThrArgAsnIleLeuGlnAlaTrpThr	1358 (129)
1359	ACTTATAAGGACTTTAAGATAACCGTTAAACAATAAAGTTGCATAAATTCGAAAATGGAAATTCAGTACTAATAAAAAGAAAATAGAAT ThrTyrLysAspPheLysIleThrValLysGlnEnd	1448 (140)
1449	ATAAACTAGCGCTCTTTCATATTATTAAGGGGTAATCGACAGCGGATTGTAATTTGGCTTCGATCCTGTGTGCCATCATTTTATTGGA  (A) <sub>n</sub> (janB)	1538

*jan* SEQUENCES. Strain, *Canton S*. Accession, M27033. The TATA box and transcription initiation site of *Sryβ* are indicated (near -200).

end; and primer extension was used to define the downstream 5' end. The 3' ends were obtained from two cDNA sequences. There is a leader intron starting at -41, with an acceptor site at +29. One cDNA in which this intron was spliced out, and one in which it was not, were sequenced. If this intron is spliced out, translation might start with Met-17 at +49. There are also introns in the Ala-64 and after the Gln-105 codons (*jan* Sequences) (Yanicostas et al. 1989).

### *Developmental Pattern*

The 0.8 kb *janA* transcript is present at all developmental stages in both sexes; it is particularly high in 0-12 h embryos and in the ovaries of adult females. In addition, there is a 0.95 kb transcript that differs from the 0.8 kb transcript only in the length of its poly(A) tail. The 0.95 kb transcript is sex-specific, occurring only in males from the third larval instar onward; the highest levels are in the adult male, where it is found in the gonads (Yanicostas et al. 1989).

### *Promoter*

The gene *Sryβ* is upstream of *janA* and transcribed in the opposite direction (see *Sry*, Fig. 28.1). Less than 100 bp separate the putative TATA boxes of *Sryβ* and *janA*; and since they are both expressed at high level in ovaries, it is likely that the two genes share regulatory sequences (Yanicostas et al. 1989).

## *janB*

### Gene Organization and Expression

Open reading frame, 140 amino acids; expected mRNA length, 579 bases. Primer extension, S1 mapping and a cDNA sequence defined the 5' end. The 3' end was obtained from S1 mapping and a cDNA sequence. There are introns

in the Gln-18 and Leu-70 codons and after the Arg-111 codon (*jan* Sequences) (Yanicostas et al. 1989).

### *Developmental Pattern*

*janB* transcripts are present only in males from the third larval through the adult stages; the highest levels occur in adults. Expression appears to be restricted to the gonads. The leader region of *janB* has striking sequence similarity with the leader element of *mst(3)g1-9*, a gene that is thought to mediate spermatid-specific translation (Yanicostas et al. 1989).

### *Promoter*

Accurate and tissue-specific transcription requires no more than 175 bp upstream of the transcription initiation site of *janB* (Yanicostas et al. 1989; Yanicostas and Lepesant 1990). When there is active transcription of *janA*, there is a reduced accumulation of RNA from the *janB* transcription initiation site; this is probably a case of transcription interference similar to that observed in *Adh* (Yanicostas and Lepesant 1990).

## References

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## *knirps* and Related Genes: *kni*, *knrl*, *egon*

### Chromosomal Location:

*kni* 3L, 77E1-2  
*knrl* 3L, 77E1-2  
*egon* 3L, 79B

### Map Position:

3-[46]  
3-[46]  
3-[47]

### Products

DNA-binding regulatory proteins of the steroid/thyroid hormone receptor superfamily that includes receptors for vitamin D and retinoic acid in vertebrates.

### Structure

Throughout this superfamily of proteins, the region extending from Cys-5 to Arg-81 is conserved, with 20 amino acids being identical in all of the related proteins and 40 others common to several of them (*kni* Sequence). In the *kni*-group proteins, as in other proteins in the superfamily, the conserved region is divided into two putative finger domains: one with four Cys ( $C_4$ ) and one with five ( $C_5$ ) (Evans 1988; Evans and Hollenberg 1988; Nauber et al. 1988; Harrison 1991). The three proteins encoded by the *kni*-group genes are more than 80% identical in the finger regions and identical for a group of 19 amino acids adjacent to the fingers (KNI box), but they are completely divergent in other regions. *kni* and *knrl* have several segments of short repeats (Fig. 20.1) (Rothe et al. 1989).

It should be noted that the classification of KNI-group proteins with the hormone receptor superfamily is based on similarities in the DNA-binding

*kni*

-2577	GAATTCCTCTGCCTGATGCAACAAATGAAAGTCAAATGGAAAACTTCTGGGAAGTCAGCTAACGAGTTTTTTGTTAAGAGTATACCTTAG	-248
-2487	ACATGGTTTAGTACATCGGTTGAAGTTTTATATTTATAATACTAGCCACACTTCGGAGTGAAAAAGTCAAGGTTCCGTCTTTGGGTCT	-239
-2397	GAACAACCCCTTTGGTACAATGCGCGCCCAATAAAGGGTTAAGCAGTCCGTTAGCGGCATAAAAGGGTTAAACAGGTAGCTCCTCTTT	-230
	----->kr2	----->kr1
-2307	CTTTTGGCTTTGAGCAAAACAATAAATATTCATAAAAAGAGCTTAAGTGCCGCCATAAGGCTCCTTGTTTACACAAAAGGAGAAATTA	-221
-2217	TGTTGGAAGTTGACTTTTAAAGGGTTACAATTAATTCGATTGATATTTGATTTTTATTGAGTATAATGATGGTGAAGGTGGGATAAG	-212
-2127	AAAGTTTATAATATTTAAGAATAATATAATTTTATGATTATTTGCGAAATATTACTTACTAAAAGTGTAATATAATAAAATTATTA	-203
-2037	AAATAATATAATATATTTCTATTCATATTGAACCTGTATGGTTAAACCTATTTTTGTATGCTATTTTGAACCGACTTGCAAAATCAAC	-194
-1947	TACTTTAATATGAATCAATCTGAATCCGGGTAATAGCCCGTCTAAATAGTATTTTTTATAACTTTTCGGACGCAATTACATACTCAATAA	-185
-1857	TACTCAACTATCGTTTTTGGCTATGAATCAATGCAGATCTCTTATTGATTAAGTTCAATTAAGCGTTTCAATTTATTGCCAAGTCGC	-176
-1767	GGTTATGCAAAATTTAACACATTTTATGAAATCTTGAGAATCAGTTTGTGAATCACACAGAAAGTGGGAATATTTCCCGCGGAAAAAGGT	-167
-1677	TTTGAAATCAAACCTAGGTGTTAGGCATACAGGCAACTCTAAATGTACCCTAAAACCGCGGACTTTGAAAAGAAAACCCCAAGCGAATT	-158
-1587	GGCCTCAAACCATTTTCGATTTTCGAGCAGCAAAAACCGTCCGCCATGCCAAAAAATGAGCAGCTGTTAAAAATGAAGTCAATAGCTTAG	-149
-1497	TCAATGTGGTGTGTGTGTGTGTGTGTGTGTGTCGAGTGTGAGAAATCCAGCCGCCCTTAGCACGCGAGTATCTTAAATAAATAAACGA	-140
-1407	ATAACGAATAATATCAGGGCCATGCAAATAGCCGTGATTACAGGAACTCAAATCGAGAGAGAGAGAGACTGAGCGTGTGATCTGAAT	-131
-1317	GAGTGTGTGTGTTTTCTATTCATTAACAACAGAGCGTTAAACATTCGCTAACATTTTCGCTCGAGTGGGGTTCGAACTCAATGCGCAT	-122
-1227	GTGTGCGTGCTCGATCGCTCTCCTCACGATCCGAGCTTAAAGGTGGTGGTTTTAGCCGCGTATTATCAGAGAGCTGGGGCTGAAAACT	-113
-1137	GGTAAGTTTGCTTTTGTGTGTGTGTGTGTGTGTGTGTGTGAGTGTGAGTGTGAGTGTGAGTGTGAGTGTGAGTGTGAGTGTGAGTGTGAGT	-104
-1047	TCTCAGCCAGAAATCAGTGTGTGAGTGTAGCCGAAGTGTGAGTTACACACATCGAGGATCCCAACCGCGTTGTTTGGGGATCAAAA	-958
-957	ACCATAATCACAATAAATCGATTCCAAATTTATAATCAAAGTGTGAAAACCTGAATCACTCAGACTGATCGAAAAGTCTTGCAAACC	-868
-867	GAAAACAAAACAAAACAAAAAAATAAACAGACACAACCTGCCAAAGTGTGAAAACGTGCAGCAACGATCGCGAATCGCTGCAATTAATA	-778
-777	AGTCGTATAAACCTAGTGCCATCCAACAAAAGCAAAAAGGTAAGTTCTGAGCAGGGGAGACAACATGAGATCAAAGTCAATTTCA	-688
-687	CGGTTCTTGTTTTGGGTGCGGGCCGTGGGCAAAATTTACGATCCCTTGTGCACTACTTTTATTTTTTACTGTTTACGCGAAAAACTAA	-598
-597	CGGCCACATTTCCATCTTTTCTTATTTTTTGCCTCCGAGGCGGCAAAAATCACAATTTTATAAGCGAAATTTCTATTTTT	-508
-507	TGTTTGACTCGGAAAAATCGCGGTGAGTTTTATGGCCGAACGTCAAGGCTGGACTGCTGCTGCAATTTTTTAGGGAACACTATTTTC	-418
-417	GCAGCACTCGTAATGACTTCAATAAAAAAGAAAATCTACCTGAGTTTTATGACTGGACAGCGCAAAAATGAAAATGAACGCATAGG	-328

-327	GTTGCATAATCCGGCAGCTTAAGTTTTTTGGCCTGTTGTGATCATAAAAAACGCCATCCTGTCTAGTTTTTCCCAGTTTCCTATATATA	-238
-237	TCCTGGCCCGCTAGAGCTCAGCATCAGTTGCTCAGCAGCATTCCAAGCGAACAGATCATAACAGCAGATCCTCACAGCGATCGTGAGAAA	-148
-147	AGCATTCAAATTCACAAATATCATTCCAAAATGGTGTCAACTGGTGTAGTGCCAGAGCCGTTCCGCTACTTGTGGATTACACATA	-58
-57	TACCTCTCGACCAGTGGATTAACCCCTTACTAACCGCGATTTCTTTACAATCTCCAGATGAACCAGACATGCAAAGTGTCCGGTGAGCC	32
	MetAsnGlnThrCysLysValCysGlyGluPr	(11)
	*** ----**	
33	GGCGGGGGCTTCCATTTTGGCGCCTTACCTGCGAGGGCTGCAAGGTAAGTTGTGTCTCAAGAAATCCATTGACAATAAATTAGCAGA	122
	oAlaAlaGlyPheHisPheGlyAlaPheThrCysGluGlyCysLys	(26)
	-----***-----**	
123	ACGTAACCCCAAGGGGTAGTTTTAGAAATGTTTCGAGGAACAGGCCATCGGCGATTCAAATCGTTGTACCATTGGCCTTAAGTCTTGA	212
213	ATGAATTTCTGCTCTCTTTTGTCAATCAGTGTGCATACCACATACTAAGCCACATTCGTCCTTCCCTTCGCCATTGCAGTCTCTCTTTG	302
	SerPhePheG	(30)
	-----	
303	GCCGCTTTACAACAACATCAGCACCATCAGCGAGTGAAGAACGAGGGCAAGTGCATCATCGACAAGAAGAACCGCACCCTGCAAGG	392
	lyArgSerTyrAsnAsnIleSerThrIleSerGluCysLysAsnGluGlyLysCysIleIleAspLysLysAsnArgThrThrCysLysA	(60)
	-----***-----***-----***-	
393	CGTGCCGCTTGAGGAAGTGTACAACGTGGGCATGTGGAAGGGGGATCCCGCTACGGACGTCGCTCCAAGTGGTTCAAGATCCATTGTC	482
	laCysArgLeuArgLysCysTyrAsnValGlyMetSerLysGlyGlySerArgTyrGlyArgArgSerAsnTrpPheLysIleHisCysL	(90)
	*****-----**	
483	TGTCGAGGAGCACGAACAGGCCGCGCAGCGCGGGCAAGCGCCCTCATTAGCGGGTGGCGTATCGGGTGGGTGGTCCCGTCGGCCT	572
	euLeuGlnGluHisGluGlnAlaAlaAlaAlaAlaGlyLysAlaProProLeuAlaGlyGlyValSerValGlyGlyAlaProSerAlaS	(120)
573	CTTCCCGGTGGGCTCGCCACACTCCCGGATTTGGGGACATGGCCGCCATTTGCACCACCATCATCAGCAGCAGCAGCAGCAGG	662
	erSerProValGlySerProHisThrProGlyPheGlyAspMetAlaAlaHisLeuHisHisHisHisGlnGlnGlnGlnGlnGlnGln	(150)
663	TGCCGCTCATCCACATATGCCTCTGCTGGGCTATCCAGCTATCTGTCCGACCCATCCGCCGCCCTGCCCTTCTTCAGCATGATGGCG	752
	a1ProArgHisProHisMetProLeuLeuGlyTyrProSerTyrLeuSerAspProSerAlaAlaLeuProPhePheSerMetMetGlyG	(180)
753	GTGTACCGCACCAGTCGCCCTTCCAGCTGCCCCACACCTCCTCTCCAGGCTACCATGCAAGTGTGCCGCTGCAGCGGCTTCTGCTG	842
	lyValProHisGlnSerProPheGlnLeuProProHisLeuLeuPheProGlyTyrHisAlaSerAlaAlaAlaAlaAlaAlaSerAlaA	(210)
843	CCGATCCGCTTACCGCAGGAGATGTACAAGCACCCAGAGCGTGGATTCCGTTGAGTCGCAAGAACCGCTTAGTCCCGCCAGCCAGC	932
	laAspAlaAlaTyrArgGlnGluMetTyrLysHisArgGlnSerValAspSerValGluSerGlnAsnArgPheSerProAlaSerGlnP	(240)
933	CACCAAGTGGTGCAGCCCACTCCTCGGCCCGCCAGTCGCCCATCGATGTCTGCCTGGAGGAGGATGTTCACTCCGTGCACAGCCATCAGT	1022
	roProValValGlnProThrSerSerAlaArgGlnSerProIleAspValCysLeuGluGluAspValHisSerValHisSerHisGlnS	(270)
1023	CGTCCGCAAGCCTCTGCATCCCATTTGCCATCCGAGCCACGCCAACCACTCCGACTAGCAGCAGCCCGCTGAGTTTTGCGCCAAGATGC	1112
	erSerAlaSerLeuLeuHisProIleAlaIleArgAlaThrProThrProThrSerSerSerProLeuSerPheAlaAlaLysMetG	(300)
1113	AGAGCTTGTGCCCGTTTCGGTTTGCTCCATTGGCGCGAAACCACCAGCGTTGTACCAGTGCATCTCCACCCTTCCGCTCAAGAAG	1202
	lnSerLeuSerProValSerValCysSerIleGlyGlyGluThrThrSerValValProValHisProProThrValSerAlaGlnGluG	(330)

(continued)



1203	GACCCATGGATCTGAGCATGAAGACCTCGCGGAGCTCCGTGCACAGCTTCAACGACAGCGGCTCCGAGGATCAAGAAGTGGAGGTGGCTC lyProMetAspLeuSerMetLysThrSerArgSerSerValHisSerPheAsnAspSerGlySerGluAspGlnGluValGluValAlaP	1292 (360)
1293	CGCGCCGGAAGTTCTACCAACTGGAGGCCGAGTGCCTGACCACCAGCAGCAGCTTCTCCCACTCCGCGGCCCACTACCGAACACCA roArgArgIysPheTyrGlnLeuGluAlaGluCystLeuThrThrSerSerSerSerSerHisSerAlaAlaHisSerProAsnThrT	1382 (390)
1383	CCACCGCCCATGCGGAAGTCAAGCGGCAGAAGCTAGGTGGTGAGAGGCCACCCACTTCGGTGGCTTCGCGGTGGCCACAATGCGGCTA hrThrAlaHisAlaGluValLysArgGlnLysLeuGlyGlyAlaGluAlaThrHisPheGlyGlyPheAlaValAlaHisAsnAlaAlaS	1472 (420)
1473	GTGCCATGAGGGGAATATTCGTGTGTCTAAGTACACGGCGAAAAACCAAGTGGGAGGAGTCGCCCCAAAAACCTCGTTGTTATTT erAlaMetArgGlyIlePheValCysValEnd	1562 (429)
1563	TTTGTTACTTAAAGAAAATGTAATTTATTCGTGTGCTCGCTCACACTTAGGGAAGTGAAAAGAGATAGGGACAGACAGGTTTTGCTGGA	1652
1653	AAGAGACAGCCTGACCAGTTAGTTGCATTGCACTCGCACACATACACCTATATACCACCACACACACTCACACTCACCTATTGAGCTC	1742
1743	GGATCCAAAAATATTTTTTATGAAAACGTTAAAATTGTAATATATCTTTGAGCTTGTTTGCAATTGTATTTTAAAGTTAGCCGGCGGA	1832
1833	AGAGCCGTAGAAGTAGTAATCATTCACCCTCAAATGCTATTGTACATACAATTTGTTAAGTCTAAGAATGATCTTTATTGTCTCTAAG	1922
1923	TATTTATTCTATTATAGCTCTAGTTATGGTATGCTAAAGATTGGCATTTAGGTTTTATACAAGAAAAATAAAACTATTAAAAATTA ----- (A) <sub>n</sub>	2012
2013	AACTTTGTGTTTCCAATGCTTTTCGTGTATTTCAAGAATACACAAATTCATATTTGAAGTTTTGCTTATGGATAATGAACTAECTT	2102
2103	ATTTACAATGTGCTCTGAAGCTCATAATTCACGGCAGCTTACTTCGGTGTGCTTTTATTGATTTAATTTTAGTTGTGACATCATA	2192
2193	GAAAAGTGATTTAATTACAAAACAACACTTTAAGAAAATATTTAAAAATACTCATACAACGTATTTCGTTGTACCTTAAAGTTAACGA	2282
2283	ACTCTTCTGATTTGTTTAAAGCACATTATTATGGACTATATGCTGGTGCAAATCTCTTCGGATGTATCTGCTGGATGTATCAACGAAG	2372
2373	TTTGTCGGCTACCGCATTCTATAGGATCAAAGCCAATAATTTTACGTATTCAGCTACCGCTCTGTTTCAAATCAGTTCTGTT	2462
2463	CAAACATGCGAGCATATATCCATATACATATATCTGATCGGGGTGCTTTGGCTCGATGTTGTTAACCACGGGCCAAATGGCGTGGCC	2552
2553	TGCAATGGCAACACAAAAAGAGACGGAAAAACAATGCTTTGGCATAAATCAATCAACATTCGGTTGCAAGTCGATCGGCGATGGCCG	2642
2643	ATAATAAACCGATATAGCAGCCGTTAAGTGCTTTGCTGTGCTGCCATTGCACTCGCGATTGCTGTAAGGCAGTTGTGTAGTAAATTA	2732
2733	AAAATGCCACAAATGTTACGCACAGAAATTCGATGCAACCCCC	2776

*kni* SEQUENCE. Accession, X14153 (DROKNR1). *kr1* and *kr2* are two KR binding sites. An exclamation sign at -1,003 marks the 5' end of a cDNA. Dashes under the amino-acid sequence mark conserved positions in the C<sub>4</sub>/C<sub>5</sub> finger regions, and asterisks, the relevant Cys residues.

regions only; the C-terminal regions of KNI-type proteins bear no resemblance to the C-terminal regions of the mammalian receptors to which hormones bind. Further, there is no evidence that function of the KNI-type proteins in *Drosophila* requires the presence of a ligand, as is the case for the steroid/thyroid hormone receptors.

## *kni*

### Product

#### *Functions*

KNI plays an important role in the early stages of embryonic pattern determination in the posterior region of the embryo. The consensus binding site of KNI is AA/TCTAA/GATC (Hoch et al. 1992).

1. KNI is one of the regulators of the embryonic "zebra" pattern of expression of the pair-rule gene *hairy* (*h*): two of the functions of KNI appear to be the activation of stripe 6 of *h* and the repression of anterior expansion of stripe 7. Strong binding of KNI to the *h* promoter in the stripe-7 regulatory element and weak binding to that of stripe 6, has been observed (Pankratz et al. 1990).

2. KNI has a binding site in *cd1*, a *cis*-acting regulatory region of *Kr*. This binding site partly overlaps a *bicoid* protein (BCD) binding site and the two regulatory proteins compete for binding: excess KNI prevents BCD from activating *Kr* (Hoch et al. 1992).

#### *Tissue Distribution*

At blastoderm stage, the KNI protein is localized in a band that extends approximately between 43% and 27% egg length (Appendix, Fig. A.2).

#### *Mutant Phenotypes*

This is one of the gap genes. In embryos homozygous for a null allele, abdominal segments A1–A7 are fused and replaced by a single segment with a broad band of ventral denticles (embryonic lethal) (Nüsslein-Volhard and Wieschaus 1980; Ingham 1988).

### Gene Organization and Expression

Open reading frame, 429 amino acids; expected mRNA length, ca. 2,068 bases. A cDNA sequence provides the only information on the 5' and 3' ends. There are two introns at –732/0 and after the Lys-26 codon. These parameters agree with an RNA of 2.2 kb detected in northern blots. A second RNA of 2.5 kb has been reported; it is not clear if this is generated by alternative splicing or by alternative initiation or termination. *kni* is transcribed toward the telomere (*kni* Sequence) (Nauber et al. 1988).

#### *Developmental Pattern*

Accumulation of *kni* RNA is first evident in 2–4 h embryos and reaches a maximum by 4–6 h. After 8 h the RNA level is very low and it becomes

```

1                               50                               100
EGON .....M NQLCKVCGEP AAGFHFGAFT CEGCKSFFGR TYNNIAAIG CKHNGDCVIN KKNRTACKAC RLRKCLLVGM SKSGSRYGRR SNWFKIHCLL
KNRL MNQDNPYAM NQTCKVCGEP AAGFHFGAFT CEGCKSFFGR SYNLLSSISD CKNNGECIIN KKNRTACKAC RLLKCLMVGM SKSGSRYGRR SNWFKIHCLL
KNI .....M NQTCKVCGEP AAGFHFGAFT CEGCKSFFGR SYNNIISTISE CKNEGKCIID KKNRTTCKAC RLRKCYNVGM SKGGSRYGRR SNWFKIHCLL
CON -----M NQ-CKVCGEP AAGFHFGAFT CEGCKSFFGR -YNN---I-- CK--G-C-I- KKNRT-CKAC RL-KC--VGM SK-GSRYGRR SNWFKIHCLL
                * *                * *                * *                * *                *

```

```

101                               150                               200
EGON QEQQ.....TTSGL GGGSSVGS GS GGGVSSASLE QLARLQQASN QARQTYQDKT NPC...IKSA TATTSPRIEG AAVGTGIGGG .....
KNRL QEQQQAVAA MAAHNSQQA GGGSSGSGG GQGMPNGVKG MSGVPPAAA AAALGMLGHP GYPGLYAVA NAGGSSRSKE ELMMLGLDGS VEYGSKHHPV
KNI QEHEQAAAAA ...GKAPPL AGGVSVGGAP SASSPVGSPH TPGFGDMAAH LHHHQQQQ QVPRHPHMP LLGYPSYLSD .....PS
CON QE----- -GG-S-G-- -----A-- -----
| KNI BOX

```

```

201                               250                               300
EGON .ASPSFLQAA KLHHQRQLKL DSRLSN....TPSDSGAS SAGD..... PNEGVTSVL GGQIATPSST NATSLPKLDL RHPNFPATSE PDA.DMQQR
KNRL VASPSVSSPD SHNSDSSVEV SSVRGNPLLH LGGKSNSSGGS SGA..... DGSHSGGGG GGGGVTGPRP PQM...RKDL S.PFLPLFP GLA.SMPVMP
KNI AALPFFSMMG GVPHQSPFQL PPHLLFPGYH ASAAAAAASA ADAAYRQEMY KHRQSVDSVE SQNRFSPASQ PPVVQPTSSA RQSPIDVCLE EDVHSHVSHQ
CON -A-P----- -P-----

```

```

301                               350                               400
EGON HQELLE... IFRSHSEPLY SSFAPFSLP PVLLAAGVPQ LPI...FKDQ FKAELLFPTT SSPELEEPID LSFRRADHA SPMAHNSNSP SLSEPAASH
KNRL PPAFLPPSHL LFPGYHPALY SHHQGLLKPT PEQQQAQVAA AAVQHLFNSS GAGQRFAPGT SPFANHQQHH KEEDQPAPAR SPSTHANNH LLTNGGADE
KNI SSASLLHPA IA IRATPTTPTS SSPLSFAAKM QLSLSPVSCS IG..... .GET TSVVPHPTT VSAQEGPMDL SMKTSRSSVH SFNDSGSEDQ
CON ---L----- S-----V-- ---T----- S-----

```

```

                                401                                450                                500
EGON  ....CLGES TNFVRKSTPL DLTLV.....SQTLTG *.....
KNRL  LTKRFYLDAV LKSQQSQSPPP TTKLPPHSKQ DYSISALVTP NSESGRERVK SRQNEEDDEA RADGIIDGAE HDDEEEDLVV SMTPPHSPAQ QEERTPAGED
KNI   .....EVE VAPRRKFYQL EAECLTTSSS SSSHSAAHSP NTTTAHAEVK RQKLGGAEAT HFGGFAVAHN AASAMRGIFV CV*.....
CON   -----S-----

                                501                                550                                600
EGON  .....
KNRL  PRPSPGQDMP IDLSMKTTGS SLSSKSSSPE IEPETEISSD VEKNDTDDDD EDLKVTPEEE ISVRETADPE IEEDHSSTTE TAKTSIENTH NNNNSISNNN
KNI   .....
CON   -----

                                601                                650
EGON  .....
KNRL  NNNNNNNNSI LSDSEASETI KRKLEDELIEA SSENGKRLRL EAPVKVATSN ALDLTTKV*
KNI   .....
CON   -----

```

FIG. 20.1. Alignment of the KNI-related polypeptides by the GCG program *Pileup*. Asterisks mark Cys in the C<sub>4</sub>/C<sub>5</sub> finger domains. The KNI box is underlined. The CON(sensus) sequence identifies residues identical in the three sequences.

undetectable in larval stages. The 2.2 kb transcript is present transiently during the blastoderm stage while the 2.5 kb transcript predominates in the later stages (Nauber et al. 1988). RNA is first detectable, by *in situ* hybridization, after the 11th round of embryonic nuclear division when it appears in a broad band centered at 40–35% egg length (Appendix, Figs A.1–A.3). Soon thereafter, RNA appears at the anterior tip; and, still later, during blastoderm cellularization, a third zone of expression becomes evident as a narrow stripe at 75–70% egg length. Expression in the posterior domain diminishes during gastrulation and eventually ceases altogether. In the anterior tip, on the other hand, expression persists through gastrulation when it exhibits a complex pattern. In yet older embryos, *kni* transcription is limited to distinct areas of the epidermis and gut (Rothe et al. 1989).

### Promoter

The expression of *kni* is stimulated by the *Krüppel* protein (KR) either directly or indirectly, and there are two KR binding sites between –2,300 and –2,400 (Pankratz et al. 1989; Capovilla et al. 1992). Anteriorly, transcription of *kni* seems mainly regulated by the product of *hunchback* (HB) (and perhaps the product of *bicoid*), being repressed at intermediate and high concentrations, and stimulated at low concentrations. Similarly, *kni* is repressed by the *tailless* product (TLL) which is present at the posterior end of the embryo. It is proposed that these interactions explain the expression of *kni* in a broad band immediately posterior to the band of *Kr* expression in the mid-section of the embryo (Appendix, Figs A.2 and A.4) (Hülskamp et al. 1990).

A 4.4 kb fragment upstream of the transcription initiation site is sufficient for normal *kni* expression. Deletion mapping of this DNA segment indicates the presence of several sub-regions in whose absence *kni* expression in embryos expands either anteriorly or posteriorly. The presence of HB and TLL binding sites in those sub-regions led to the suggestion that *kni* expression is activated throughout the embryo and that the broad band of *kni* transcription in the posterior half of the embryo is achieved through repression by HB (anteriorly) and TLL (posteriorly) (Pankratz et al. 1992).

### *knrl* (*knirps-related*)

#### Product

Unknown. No mutations are known in this gene (Fig. 20.1).

#### Gene Organization and Expression

Open reading frame, 647 amino acids. One cDNA of 3,505 bases was sequenced; a single band of 3.8 kb is detected by northern analysis. That cDNA

sequence provides the only information on the 5' and 3' ends (Oro et al. 1988).

### *Developmental Pattern*

A low level of maternal *knrl* RNA is uniformly distributed throughout pre-blastoderm embryos. After the 12th nuclear division a posterior band forms as is also the case for *kni* RNA; and afterwards expression of the two genes is almost the same. However, *knrl* transcription never ceases altogether; a low level of expression is maintained in all stages (Oro et al. 1988; Rothe et al. 1989).

### *egon* (*embryonic gonad*)

### **Product**

Unknown. No mutations are known in this gene (Fig. 20.1).

### **Gene Organization and Expression**

Open reading frame, 373 amino acids. There is one intron after Lys-26.

### *Developmental Pattern*

Transcripts are restricted to late embryogenesis and they are 10-fold less abundant than for *kni* or *knrl*. After germ band shortening, transcripts appear only in the gonadal primordia that form in abdominal segment 5, as demonstrated by *in situ* hybridizations (Rothe et al. 1989).

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# 21

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## *Krüppel: Kr*

**Chromosomal Location:**  
2R, 60F3

**Map Position:**  
2-107.6

### **Product**

DNA-binding regulatory protein of the Zn-finger type that plays a central role in the early stages of embryonic pattern determination in the mid-section of the embryo.

### *Structure*

The protein sequence can be divided into three regions. Two of the regions, the amino-terminal and carboxy-terminal segments (221 and 108 amino acids, respectively) are >30% Ala, Ser, and Pro. The third region, at the middle of the protein, is made up of four and a half repeats of a 28-amino-acid segment; these segments have the characteristics of C<sub>2</sub>H<sub>2</sub> Zn-fingers. A potential glycosylation site is found near the C-terminus. Short segments near the N-terminus show similarities with the *hunchback* (*hb*) protein (Rosenberg et al. 1986; Evans and Hollenberg 1988; Harrison 1991).

### *Function*

Distinct DNA-binding and repressor domains have been identified in KR. KR finger domains bind to AANGGGTTAA decamers, sequences that are known to occur in the promoters of several genes controlled by KR (Pankratz et al. 1989; Stanojevic et al. 1989; Treisman and Desplan 1989). Transcriptional repression is effected through an Ala-rich region of the protein included in the segment from amino acids 26–110 (Licht et al. 1990).

KR acts as a repressor of the anterior gap gene *hb* (*hb*), the pair-rule gene *eve-skipped* (*eve*) (Licht et al. 1990) and probably *giant* (*gt*) (Kraut and Levine 1991). In the posterior regions of the embryo, it interacts with the pair-rule gene *hairy* (*h*) and with the gap gene *knirps* (*kni*), which it activates, perhaps



indirectly, by repressing the expression of *gt*, a repressor of *kni* (Capovilla et al. 1992). KR-binding sites exist in several of the stripe-specific regulatory elements in the promoters of *eve* and *h*, as well as in the promoters of *hb* and *kni*. These interactions play important roles in the periodic expression of the primary pair-rule genes *eve* and *h* and consequently, in the future segmentation pattern along the antero-posterior axis of the embryo (Pankratz et al. 1989, 1990; Small et al. 1991; Stanjojevic et al. 1989, 1991; Treisman and Desplan 1989).

KR may be involved in developmental processes other than segmentation. After gastrulation, the protein can be seen in the nuclei of some neuroblasts, Malpighian tubule anlagen, amnion serosa and other cells; in some sites, KR persists until the end of embryogenesis (Gaul et al. 1987).

### *Tissue Distribution*

The *Kr* protein is localized in nuclei with a pattern of accumulation that agrees roughly with what would be expected from the regions and stages of embryonic *Kr* transcription (see below). The correspondence is not exact, however; the protein begins to appear 30 min later than mRNA, during the 13th nuclear division, suggesting that there exists a mechanism of post-transcriptional control. During the blastoderm stage, when metamer determination is taking place, KR accumulates in a bell-shaped concentration profile. That is, the protein is detectable between approximately 60 and 33% egg length (Appendix, Figs A.2 and A.3) with the concentration being maximum in the middle of the embryo and declining in steep exponential fashion toward each pole (Knipple et al. 1985; Gaul et al. 1987).

### *Mutant Phenotypes*

This is one of the gap genes. Embryos amorphic for *Kr* lack the three thoracic and first five abdominal segments; *Kr* is an embryonic lethal without maternal effect (Nüsslein-Volhard and Wieschaus 1980; Ingham 1988).

## **Gene Organization and Expression**

Open reading frame, 466 amino acids. Primer extension and cDNA sequencing were used to define the 5' end. Two cDNAs having different 3' ends (368 bp apart) were sequenced. Spliced and unspliced RNAs are abundant. Thus it might be expected that RNAs of several sizes ranging between 1,851 and 2,591 bases would be observed. However, only two bands of approximately 2.5 kb are detectable; one has an intron, the other does not. The intron is in the Thr-13 codon, and there are several short open reading frames within the intron that might serve for translational control (*Kr* Sequence) (Gaul et al. 1987).

When a genomic library was searched extensively with a *Kr* probe consisting of only the finger domains, eight cross-hybridizing clones were identified. One

Kr

Kr730 element  
BamHI

-3267 GGATCCTAAGTTAACTATAATCCAGGCTTAATCACTGGATCAATAACTAAGTAGCATTTCCTGGGATGGAATATGAAGTTACCTGCATA -3178

-3177 TGACCTACCGATCCTGAAAACGCTTTAACTTAATCGACATGCATGATCATAAAAAGCAATTTGCTACAATTTATATTTTTTGCCTTTTC -3088  
 -----t111 //

-3087 CTTCTTTTAAGCATCTGGGATCTGGATCAGAAAAGAAAAAGTGAACGCCTACCTTCAGAAACGGATTAATATTTTTTCAGACAAAATAATC -2998  
 //hb3 -----t112 -----  
 //hb4

-2997 CAGCCTTAAGCATGGTGATTAAGCTTGATCCCCACCAAGGGGCGTAATATTGACGGATTTTCCTTAAATCCGCTCGTTAATCTCCGGCT -2908  
 -t113 -----t114 -----t115  
 //bcd2-3 //

-2907 TAGAGCGCGACGCGTTTTTCGCGACTCCGCCTGCATTGTTTTTTTTTCAGTTTCTCAATTGCAAGAAGGCAGGCCTATGGACCGAAT -2818  
 //hb5 //hb6

-2817 GAGGATCATAATTATGAATTCTAAATAAACTAAGAAGGGCAGTCGGCATAGTATTGATCTACCTGTAAGCGTGGGTTCTATCTTTGCC -2728

-2727 CCTCGCATTGAGACTCTCTAGTCACAGGTAGACTGTATACCAGCCTTGAGTTCGTCGGCAATTAAGAAGTCAAATTTCTCTTAAAAACA -2638  
 -----t116  
 //

-2637 ACAAAAAATGTCAAAGTAAAAACAATGCAAAAAAGATGTGTAAGTGAAGTAAATCCGGCTTAGGATCTTGCGTCATAAACGTGACTAGG -2548  
 //hb7-10 -----kni1 -----t117  
 //bcd6  
 //\gt

-2547 TAGCC -2543

-----184

-267 AATATAATCGAATGAAATTTCAACTACCTCATTTTGTAGTCNGTAGACTTTTATAAAAGACAATTTTGTGAAATCTCTCTACCTCAA -178  
 -----

-177 AGTACAAAAGTGTGTACAAAAATTATTCATATCCCTGAAAGTGCACAAAATCTCAAATGAAATTTTGTGTCTAAAAACTAAGCTCCA -88

-87 AAATCACTAAGCGAATATTATAGGTGTTTTCTGTGTGCGGGAAAACATTGCGCGACACAAAATAGGAGCACAAGAAGAAATTTGTTGAT 2  
 Me (1)

3 GTCCATATCAATGCTTCAAGACGCACAAACGCGAAGTAAAGTATAGACCAAATAAAAATATCCCAAGAAAGTAAACTATCTAGAAGTTC 92  
 tSerIleSerMetLeuGlnAspAlaGlnThrArgT (13)

93 TAGTGTCCCGATCACTTTCTCATTATTAACAGTCCGATGCTTTAGGATAGAAAAACAAAATGTAATGTAATTGCAGCACAATACCGAT 182

183 TAGTTGAATTTGTTTACATGTTTGACAGGAACCGGCACTTAACCTGTTATCGACAAAACAAAACAGTATTAGACGAAAAATAGAGAGCT 272

273 GCGAAAACACTAAGAGTTCGCTCCGACGAAACTTCTCTCACACATGAATCATATGTAAAATTTTTTCTCTTTAAGCGGTGCTCTT 362

363 AAGACATTTCCAAATGAAAACATACTAECTTATGATTTTTTTTTAGCCTTAGCTGCTGCATTAGCTGGCATAAAAACAGAGGACGTTCA 452  
 hrLeuAlaAlaAlaLeuAlaGlyIleLysGlnGluAspValHi (27)

453 TCTAGACCGTTCCATGTCGCTATCGCCCCCATGTGCGCCAACACATCAGCTACAAGCGCCGCTGCGATTATCCAGCTATGGGTCTCCA 542  
 sLeuAspArgSerMetSerLeuSerProProMetSerAlaAsnThrSerAlaThrSerAlaAlaAlaIleTyrProAlaMetGlyLeuG1 (57)

(continued)

543	ACAGGCGCCGCTGCCTCAGCTTTTGGAAATGCTATCACCTACCCAACCTCTGCGTGCAAACCGTCAAGCTGCCGATTCATGGCCCAACT nGlnAlaAlaAlaAlaSerAlaPheGlyMetLeuSerProThrGlnLeuLeuAlaAlaAsnArgGlnAlaAlaAlaPheMetAlaGlnLe	632 (87)
633	GCCCATGAGCACATTGGCCAACACTCTCTTTCCACACAATCCGGCGGCTTTGTTTGGGGCTTGGGTGCCCAACAGTCGCTCCCGCCCA uProMetSerThrLeuAlaAsnThrLeuPheProHisAsnProAlaAlaLeuPheGlyAlaTrpAlaAlaGlnGlnSerLeuProProG	722 (117)
723	GGGTACGCATTTACATTGCGCCGACGACCCCGCACTGCGCGCTGCCACTCTTTAGGTAGTGGAAGCACCATTAAATCCCCCAA nGlyThrHisLeuHisSerProProAlaSerProHisSerProLeuSerThrProLeuGlySerGlyHisProLeuAsnSerProAs	812 (147)
813	CAGCACTCCCAGCACCATGAGCCAGCGAAGAAGGCTCGAAAGTTATCGGTTAAGAAGGAGTTTCAGACCGAGATCAGCATGAGTGATAA nSerThrProGlnHisHisGluProAlaLysLysAlaArgLysLeuSerValLysLysGluPheGlnThrGluIleSerMetSerValAs	902 (177)
903	CGATATGTACCTATCATCGGAGGCCAATATCTCCGCCITCCAGTGGCAGCTCTCCTAATTCACGCACGACGGAGCGGGTGGAAATGC nAspMetTyrLeuSerSerGlyGlyProIleSerProProSerSerGlySerSerProAsnSerThrHisAspGlyAlaGlyGlyAsnAl	992 (207)
993	TGGATGTGTCGGTCTCCAAGGATCCATCTCGCGACAAAAGCTTACCTGTAATACTGCTCAGCAGCTTTGGCTATAAGCACGTGCT aGlyCysValGlyValSerLysAspProSerArgAspLysSerPheThrCysLysIleCysSerArgSerPheGlyTyrLysHisValLe	1082 (237)
	-----	
1083	TCAGAACCACGAACGCCACCCACACCGGTGAGAAGCCTTTCGAATGTCGGAGTGCACAACCGGTTTACTCGGGACCATCACTTAAAAAC uGlnAsnHisGluArgThrHisThrGlyGluLysProPheGluCysProGluCysAspLysArgPheThrArgAspHisHisLeuLysTh	1172 (267)
	-----	
1173	CCACATGCGTTTGCATACTGGAGAAAAACCATATCATTGCTCGCACTGCGATCGTCAATTCGTTAGGTGGCCAATCTTAGACGACATTT rHisMetArgLeuHisThrGlyGluLysProTyrHisCysSerHisCysAspArgGlnPheValGlnValAlaAsnLeuArgArgHisLe	1262 (297)
	-----	
1263	GCGAGTCCACACTGGAGAGCGTCCCTATACTTGTGAAATCTGCGATGGCAAATTCAGTGACTCCAATCAGCTTAAGTCCCACATGCTGGT uArgValHisThrGlyGluArgProTyrThrCysGluIleCysAspGlyLysPheSerAspSerAsnGlnLeuLysSerHisMetLeuVa	1352 (327)
	-----	
1353	ACACACCGGTGAAAAGCCGTTTCGAGTGCGAACGGTGCACATGAAGTCCGACGGCGGCACCATCTGATGAATCACAAGTGTGGCATCCA lHisThrGlyGluLysProPheGluCysGluArgCysHisMetLysPheArgArgArgHisHisLeuMetAsnHisLysCysGlyIleG	1442 (357)
	-----	
1443	GTCGCGCCTACTCCCGCCTTTACCAGGCGATGAGTGGAGATTACCCCGTGGCAATCTCCGCAATGCTATCGAGGCATCCACGAATAG nSerProProThrProAlaLeuSerProAlaMetSerGlyAspTyrProValAlaIleSerAlaIleAlaIleGluAlaSerThrAsnAr	1532 (387)
	-----	
1533	ATTTGCGGCAATGTGTGCCACCTACGGAAGTTCGAATGAGTCGGTCGACATGGAAAAAGCGACACCGGAGACGATGGTCCATTGGATTG gPheAlaAlaMetCysAlaThrTyrGlySerSerAsnGluSerValAspMetGluLysAlaThrProGluThrMetValHisTrpIleCy	1622 (417)
	-----	
1623	TCTGAAGATGGAGCCAGCTCTGTGGATGGCCATTACAGCAACATCGCACGGCGCAAGGCACAGGACATTCGTCGGGTTTTCCGGCTGCCT sLeuLysMetGluProAlaLeuTrpMetAlaIleThrAlaThrSerHisGlyAlaArgHisArgThrPheValGlyPheSerGlyCysLe	1712 (447)
	-----	
1713	CCACCGCAAATCCCTCAGTACCCAGTGATATGCTGAGCAAACCGACGAGGATTTGAGCATGCATTCCTCGTTCTATCGGATCT uHisArgLysSerLeuThrTyrProValIleCysLeuSerLysProSerGlnArgIleEnd	1802 (466)
	-----	
1803	CACGAGCAAACCGATGATATTGACTTGTATGATTTAGATGATGCCCGGCTTCTTATATGGGCCATCAACAACATTAGGCCCAACCGAT	1892
	-----	
1893	CCGAATTGTACATAGCCCTAATCAGTTTTCAATTTGATGAAATGACTGGCATTATTAACACAAAATGAAAATTTGCTATTTCAAAGT	1982

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1983  GGAAAGTAAAAATTGTTGCAACAGGAATATAATGATAAGTACAAGTTTAAAAAATAACATACAAAAAGTCGAAATTGTACAAAGTAAGC  2072
      -----                                     |(A)n
2073  CATACGTATGCTTGTGTACGCCAAACCCACCAAATCAAATCGAAAAATGTCGTGCCATTCCTTTACCTTAAATTTAAGTTATATTCTTAGGTT  2162
2163  CGGAATCTTAAATGTACATATTAGCACTTACACAGCTGCCAATTGTAAGAATATCGGCCTCTAAACATGCTTGTTCGAGAAAAATAAAA  2252
2253  GACACAAAGGTTAATTAGGAAATCTATAACTAATTTTATTTAATTTATTACGCTTAATTTTTTTATAATTTAATCAAATCTTTAAGAA  2342
2343  AACAAATCGCAATAATCTCAAAACAAACTAAGTTCAAGTTAAATAATAAAAAACATTTGTTTGATAAATGTTCTGTTCGATTCTCTATTT  2432
      -----                                     |(A)n
2433  AAAACTATTATTAATAATAAAAAATTTAGTTAATCCTGTTTTTTTAAAGATC  2483

```

*Kr* SEQUENCE. The segment from  $-267$  to  $2,483$  is from GenBank, Accession, X03414 (DROKR). His and Cys of the Zn-finger repeats are underlined, as is a potential glycosylation site (Asn-399). The segment from  $-3,267$  to  $-2,543$  (Kr730) is from Hoch et al. (1992) and is numbered arbitrarily. This regulatory region starts at the *Bam*H1 site approximately 3 kb upstream of *Kr*. Symbols under the sequence indicate various footprints: ---tll, for TLL; |||bcd for BCD, ///hb for HB (Hoch et al. 1991, 1992) and \\ \\gt for GT (Capovilla et al. 1992).

of these was characterized in some detail. It was localized to the left arm of chromosome two in region 26A–B. Sequence analysis identified three finger domains of the *Kr* type; greatest similarity was found in the seven amino acids that separate adjacent fingers (the “H/C-link”; Schuh et al. 1986).

### Developmental Pattern

Both *Kr* transcripts are present primarily in 2–5 h embryos, blastoderm to gastrula stages (Rosenberg et al. 1986).

*Kr* transcripts are first detected in syncytial blastoderm embryos, after the 11th nuclear division (Appendix, Fig. A.1). RNA occurs in the peripheral cytoplasm confined to a band 8–10 nuclei wide in the mid-embryo (55–45% egg length; Appendix, Figs A.2 and A.4). By the cellular blastoderm stage (3.5 h of development), the level of transcript has greatly increased; the RNA appears in a band about 12–14 cells wide as well as in the cytoplasm of yolk cells. During this stage, *Kr* RNA also accumulates in a posterior cap; the cap is 10 cells wide and does not include the pole cells. Early in gastrulation, a third zone of gene expression develops in the anterior portion of the embryo; and, as gastrulation progresses, expression becomes yet more widespread. By the end of germ-band extension (6 h), *Kr* RNA occurs throughout the embryo, from the posterior edge of the cephalic furrow and through the thoracic and abdominal anlagen. The transcripts then begin to diminish; and, by the beginning of germ-band shortening (8 h), they reach near background level (Knipple et al. 1985).

### Promoter

An upstream segment of DNA 18-kb long is necessary for normal *Kr* expression. Within this region, there are at least seven independent *cis*-acting elements that, alone or in various combinations control *Kr* expression at each of the ten identified embryonic sites where *Kr* product is found.

Two of the *cis*-acting elements (cd1 and cd2), located from 1 to 3 kb upstream of the transcription initiation site, are primarily responsible for expression in the central domain of the embryo (Hoch et al. 1990). During the blastoderm stage, the central region of expression is, at least in part, defined by the gradients of *bicoid* (*bcd*) and *hb* gene products; *Kr* transcription appears to be stimulated by low concentrations and repressed by high concentrations of those proteins (Hülskamp et al. 1990). A 400-bp segment in cd1 is essential for expression of a reporter gene in the central region of the embryo. The *cis*-acting function of cd1 depends on the presence of wild-type alleles of *hb* (repressing *Kr* transcription) and *bcd* (activating transcription). Clustered in 730 bp of cd1 (the *Kr*730 element) are 10 HB and 6 BCD binding sites (Hoch et al. 1991). Seven binding sites for the product of *tll* (TLL) are also found in the *Kr*730 element. The TLL sites partly overlap BCD binding sites, and there is competition for occupancy such that the activating function of BCD can only occur if TLL concentration is low enough. Similar competition occurs between BCD and the *kni* product (KNI); but there is only one KNI binding site, so its effect does not appear to be so significant as TLL's (*Kr* Sequence) (Hoch et al. 1992).

The repressive action of BCD may be effected directly or through its activation of *gt*, which in turn would interact with HB to repress *Kr* (Kraut and Levine 1991). The repressive action of *gt* on *Kr*, if it occurs, would be mediated by *gt* protein binding sites in the regulatory regions cd1 (*Kr* Sequence) and cd2 (Capovilla et al. 1992).

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## The Metallothionein Genes: *Mtn*, *Mto*

### Chromosomal Location:

*Mtn* 3R, 85E10-15  
*Mto* 3R, 92

### Map Position:

3-48.8  
 3-[68]

### Products

Small, Cys-rich cadmium- and copper-binding proteins.

### Structure

MTN and MTO share properties with the metallothioneins (MT) of other invertebrate and vertebrate species: they are small, they lack aromatic amino acids and Cys residues constitute 25% or more of the protein (Lastowski-Perry et al. 1985; Mokdad et al. 1987). One striking feature of MTN is the arrangement of its 10 Cys residues in Cys-X-Cys groups that are distributed almost identically to the Cys-X-Cys groups in the N-terminal half of mammalian MT (Lastowski-Perry et al. 1985; Maroni 1990). Otherwise, sequence identity between MTN and MTO, or between either one of the *Drosophila* MTs and a mammalian MT is not extensive, being only 20–25% in all pairwise combinations.

Cu-MTs may be precursors of the copper- and sulfur-rich concretions that are detectable in the middle mid-gut of larvae fed on  $\text{Cu}^{++}$ -containing food (Tapp and Hockaday 1977; Maroni et al. 1986b; Lauverjat et al. 1989).

MTO has been purified and partially sequenced (Silar et al. 1990); but MTN has proven surprisingly intractable in this respect, and purification of the protein has not been achieved (Silar et al. 1990; G. Maroni, unpublished observations).

### Function

MTs are involved in metal tolerance as evidenced by the fact that flies with duplications for *Mtn* have increased tolerance to  $\text{Cu}^{++}$  and  $\text{Cd}^{++}$  in the medium. Such duplication-carrying flies have been obtained from many natural

populations where it is thought that elevated  $\text{Cu}^{++}$  level has acted as a selective agent (Otto et al. 1986; Maroni et al. 1987; Theodore et al. 1991). Also, cells in culture that had been selected for increased tolerance to  $\text{Cd}^{++}$  showed higher levels of MT (probably MTO) accumulation (Debec et al. 1985; Mokdad et al. 1987). Whether these proteins also serve a role in metal homeostasis is not known; null mutations are not available.

### *Tissue Distribution*

Synthesis of MT is stimulated by the presence of  $\text{Cd}^{++}$  or  $\text{Cu}^{++}$  in the food and the proteins accumulate primarily in the midgut of individuals so treated (Maroni and Watson 1985).

## ***Mtn***

### **Gene Organization and Expression**

Open reading frame, 40 amino acids. There are two common alleles: *Mtn*<sup>-3</sup>, thought to be closer to the ancestral allele, is expected to make an mRNA 387 bases long; *Mtn*<sup>1</sup> has lost 49 bases of the 3' untranslated region (*Mtn* Sequence) and is expected to make an mRNA 338 bases long. These estimates are in agreement with RNA bands of 0.4 and 0.5 kb detected by northern analysis. Primer extension and cDNA sequencing were used to define the 5' end. The 3' end was obtained from a cDNA sequence that included a poly(A) tail. There is an intron in the Gly-8 codon (*Mtn* Sequence) (Lastowski-Perry et al. 1985; Maroni et al. 1986a; Theodore et al. 1991).

Duplications occur in natural populations and in laboratory strains; they always involve the *Mtn*<sup>1</sup> allele. The two copies are in direct tandem repeats at a distance of 1–5 kb of each other (Otto et al. 1986; Maroni et al. 1987; Lange et al. 1990).

Flies carrying the allele *Mtn*<sup>-3</sup>, an allele that is present primarily in African populations, accumulate approximately 30% as much mRNA as those carrying *Mtn*<sup>1</sup>; the extra 49 bases in the 3' untranslated region of *Mtn*<sup>-3</sup> may increase its mRNA turnover rate (Theodore et al. 1991).

### *Developmental Pattern*

Cadmium, copper, mercury, silver and zinc induce transcription of *Mtn* in larval and adult mid-guts, zinc being the least effective of these metals. Treatment with high metal concentrations leads to expression in the fat bodies and other tissues as well. *Mtn* RNA is not detectable early in embryogenesis, but it is clearly present, even in the absence of a metal supplement, in 18–24 h embryos, larvae, and adults (Lastowski-Perry et al. 1985; Silar et al. 1990).



*Mtn*

```

      _____ EcoRI                      Begin Mtn-.3]
-496 GAATTCGTTGCAGGACAGGATGGTGCCCGATGTGACTAGCTCTTTGCTGCAGGCCCTCTATCCTCTGGTTCCGATAAGAGACCCAGA  -40:
-406 ACTCCGGCCCCCACCGCCACCGCCACCCCATACATATGTGGTACGCAAGTAAGAGTGCCTGCGCATGCCCATGTGCCCCACCAAGA  -31:
      C
-316 GTTTTGCATCCCATACAAGTCCCCAAAGTGGAGAACCGAACCAATCTTCGCGGGCAGAACAAAAGCTTCTGCACACGCTCTCCACTCGAA  -22:
      ----->
      G
-226 TTTGGAGCCGGCCGGCGTGTGCAAAGAGGTTGAATCGAACGAAAGACCCGTTGTAAAGCCGCTTCCAAAATGTATAAACCGAGAGC  -13:
      ---- <----- <----- -----
      -->-123
-136 ATCTGGCCAATGTGCATCAGTTGTGGTCAGCAGCAAATCAAGTGAATCATCTCAGTGCAACTAAAGGCCATAATAGCCCATACCTACCT  -47
-46 TTTTGTAAACAAGTGAACAAGTTCGAGGAAATACAACCTCAATCAAGATGCCTTGCCCATCGGAAAGCGGTAAAGTTCGCAGTCTGGTGTG  43
      MetProCysProCysGlySerG (8)
      44 ATCCTTTAGGATATCACAGATCTTTCAGAGAAATGGTATTATACTAGTATAAAAATCAATGGTGATTCAATAGTATAAAAATCAAGGC  133
      134 TGAACATATCTGCAAAGTGAATCTCTGAGTTCGTCTCTAAGAAAAGAAGTCTTCAACTGCGTTTTATAAAAATGGAACACTAATGTT  223
      224 ATATGGCTTATGGATTACAGGATGTACCAGCATGTACTAATTTTTAAATCTACTTCTTCCAGGATGCAAATGCGCCAGCCAGGCCACC  313
      l yCysLysCysAlaSerGlnAlaThr (16)
      A
      314 AAGGGATCCTGCAACTGCGGATCTGACTGCAAGTGCGGCGGCACAAGAAATCGGCTGCGGCTGCTCCGAGTGAGCTTCCCCCAAAA  403
      LysGlySerCysAsnCysGlySerAspCysLysCysGlyGlyAspLysLysSerAlaCysGlyCysSerGluEnd (40)
      Lys
      CGAACTGATTTCTGTATAACTCCCAATACTAAAACGACATGTTTTCTCA T
      404 AGATCTGGAGTAGAGGCGCTGCATCTGTCTC.....TCTACACAC  493
      494 CCTGCAATAAATGTCCAATAAAGTAATTGATGCCTAACTGCGCTTTTTCGGGTGCATAATCAATTGGTCTGCGGCATCTTAGGTTAGA  583
      ----- | (A)n
      | End Mtn-.3
      584 TTCGCTTTTATTGGAGGTAGCTTCTAGCTACGTGGTCGGCAATATGCGTCGTGGAAATGGGATGGTCAAGTGTTCACAAATGTGCATA  673
      674 TACATATGTACATAACACTAAAGTCAGTTGAGCAATATGGTAATCTGAGATGACTACTTCTGAAGCGACTGAGGGATGAGTTCAAACACA  763
      764 CGGCTGACCATGACTGTAGATAAAAAACAGTTCGGCGTTAGAATATAGCCGCTATCGAATGGATAATATTAAAGAATACTAGCTTTAGA  853
      854 AATAATAAAAAATATATTACCCATCAAATTTAAAACGATTTTAGGCATAACAACGAAATGGGTAATGAAAGTTCATATTTAAATCGGCTT  943
      944 CCATTATTTTATAGGTGATTCATAGAAATATATGATGTAGACTTATTATTGCTCAGTCTGTTTTGTGAAATGCCTCGTTTATAGCGCAA  1033
1034 AAGTGCCATATAGTTTTAGATGTAATATGATCGCGCAATTAACATGAAAGTGAAGAACC GG 1095

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*Mtn* SEQUENCE. Accession, M12964 (DROMETG). The numbered lines represent the sequence of the *Mtn*<sup>1</sup> allele, above it are the four base substitutions and the extra 49 bases present in the allele *Mtn*<sup>3</sup>. Between positions -250 and -170, the 8-bp cores of putative metal regulatory elements are underlined.

Mto

\_\_\_\_\_ SspI . . . . .

-1072 AATATTGAGTCTACAGGAATGTTCCAGGACTACACGGAGAAAAATCGAAGGACACTTTGGGGATGAGAGGATATTCATGCAATTTGTG -983

-982 GTAAGGAACGAAGTCATACTCTAACTGAACGGTCTGGCTGGTCAAGTTATATATGTTTATGTGATGTTAAATATATACCTTGTGGTCA -893

-892 ACACAAGCACAGAATCAATTATATATTCATTATACCCGTTTAAAGATATAGTAAGGTAAATAAAATGTAGAAGGATCAGGATAACTAGAC -803

-802 GACTTAAAGACTGCAGACAACCTTATCTAAGTCATCTTTTCGTTGCAGGTACACCTACCAAAAAACTATTCTATATTTGTTTTCGAAAAAC -713

-712 TTTTTTTTTACTAAAAGTCATAAATATATATAAAGTTGTTCCGGGTGTTGGTTTTCCGTGCAACGAACCTGTTTTCGTAGCTCCCAG -623

-622 AGCTTATAGTTTTGCCTAATTTGCAGCGCTTTTTCTCTATTAATTTTAGTTAGCTTCCACATGTGATTTTTATGGCATTTC -533

-532 GCTGGGTTTTTTTTGAAAAGAGTTTAGTCGTAAGCGTTTTTGACGCCAATATGAGCATTTAAATTTGTTTTACTACAGAAAGTCTTTT -443

-442 ATTTATTGTGAAAAACCCGCTGGGTAGCTGCCTGCGCTTTTCATGCTTTTTATTGTGTGCTTCTGGGCTGGGGCTGAGTCACGATACGC -353

-352 GCGTATACGCAACGTATACGCAACGTGGGCAGCTGATAAGCTGATGAGGAGTTCGTGTGCCAGGAGTTGGCGAGCAATCGCGTGCACAA -263

<----- <-----

-262 AAAGAATTGCCTGGCTATCGTCTGATAAATGCGAACCCTCGCCCAGGCTGCACACGACGTGATAAGTTGGGTCAACAAACAAAT -173

----->

--->-143

-172 TTGTTTTGGATTTGTGCAATTTGCACTCGTTCGAGTTCGAGGCAATCGAAGTGGGTATAAAAGTGGGGAGTTGCCGGACTGGGTATC -83

----->

-82 AGTTGAATAGCCAAGCAACAAGCAACAAGTGAATATCAGTTCGCCTCAGCCAAGTGAAAGTCGAGAAATAGATACATACAAGATGGTTT 7  
MetValC (3)

8 GCAAGGGTTGTGGAACAAGTAAGTGGTACAACGACGAGCAAGCTGTATAATTGACAATCGTTCTCGATTCCCTCGACAGACTGCCAGTGC 97  
ysLysGlyCysGlyThrA snCysGlnCys (12)

98 TCGGCCAAAAGTGCGGGGACAACCTGCGCTGCAACAAGGATTGCCAGTGCCTTTGCAAGAATGGGCCAAGGACCAGTGTGCAGCAAC 187  
SerAlaGlnLysCysGlyAspAsnCysAlaCysAsnLysAspCysGlnCysValCysLysAsnGlyProLysAspGlnCysCysSerAsn (42)

188 AAATAAGCGGGCCAACCTATATAACTAACTGTTTAACTTCTAAACTGGAGCTTAACTCCCAACGAGTTGGCCGCAATAAATAAAGTTTATA 277  
LysEnd -----/----- (43)

278 AAGATTTGAGCATTTAAAGTTTCTGCCGTTAACTTTTTGTTACTGGGCGGTGGTCACTTACCAAGCGATAATTATATTTTCGGCTT 367  
(A)<sub>n</sub>

368 TTGGCAGCTAAACCAATTATGGTAAAATAATAAACGTCAGCTGGCATTGAGTTAAGCAAACCGCAAAATAGAATTACATGAAAAATAAG 457

458 CAAACGCAATGCGACAATTTGGGCGGGATTGCAAAATTTGTATGTTTCGGGACAGCTGCACCGGAATTAATAATCCAATCCATCAGCCG 547

548 TGATTTGAGTAAAACTCACCAGAAAGTCCATTGAATTGTGCGCAAAACGGAACATAAATCGA \_\_\_\_\_ TaqI 610

Mto SEQUENCE. Strain, Oregon R. Accession, X52098 (DROMTOG). Between positions -300 and -140, the 8-bp cores of putative metal regulatory elements are underlined.

### *Promoter*

A fragment that extends from 373 bp upstream to 54 bp downstream of the transcription initiation site is sufficient for apparently full metal response and for control of the expression of reporter genes. The addition of 3,500 bp farther upstream does not seem to increase the metal-induced response. Within the 373-bp segment that precedes the transcription initiation site, there are several copies of a 12-bp sequence that is related to the mammalian metal regulatory element (*Mtn* Sequence). The *Drosophila Mtn* promoter is capable of supporting metal-regulated expression of a reporter gene transfected into baby hamster kidney cells (Maroni et al. 1986a; Otto et al. 1987).

## *Mto*

### **Gene Organization and Expression**

Open reading frame, 43 amino acids; expected mRNA length, 376 bases, in agreement with RNA detected in northern blots. Primer extension was used to define the 5' end. The 3' end was obtained from a cDNA sequence that included a poly(A) tail. There is an intron in the Asn-9 codon (*Mto* Sequence) (Mokdad et al. 1987; Silar et al. 1990).

### *Developmental Pattern*

Cadmium, copper, zinc, mercury and silver induce transcription of *Mto* in larvae and adults, zinc being the least effective inducer (Silar et al. 1990). RNA accumulations reach levels that are only 30–50% of the levels reached by *Mtn* when the same metals are used (G. Maroni and J. E. Young, unpublished observations). During embryonic and larval development, in the absence of a metal supplement, *Mto* RNA is present at approximately constant levels; in adult females it is barely detectable; and it is absent from males (Silar et al. 1990).

### *Promoter*

There is no canonical TATA box upstream of the transcription initiation site. As in the *Mtn* promoter, there are several short sequences related to the metal regulatory elements found in mammalian metallothionein promoters (Silar et al. 1990).

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# 23

## *ovarian tumor: otu*

Synonym: Transcription unit K of the chorion gene cluster on the X

**Chromosomal Location:**  
X, 7F1

**Map Position:**  
X-23.2

### **Products**

Proteins of 98 and 104 kD of uncertain function.

### *Structure*

In each case, the apparent  $M_r$  is slightly larger than predicted from the sequence, probably due to the skewed amino-acid composition. OTU proteins are largely hydrophilic and rich in Pro (approximately 10%) (Steinhauer et al. 1989; Steinhauer and Kalfayan 1992).

### *Tissue Distribution*

OTU proteins are localized in ovaries. The 104 kD form predominates in pupal stages when advanced stages of oocyte maturation are absent. The 98 kD form is the more abundant one in adult females, when most of the ovarian mass comprises egg chambers at more advanced stages (G. L. Sass and L. L. Searles, personal communication).

### *Mutant Phenotypes*

Mutations in *otu* lead to female-sterility; they have no effect on viability in either sex or male fertility. Null alleles of *otu* (the QUI alleles) result in the total absence of germ cell proliferation. Severely deficient alleles (the ONC alleles), seem to result in germ cell proliferation with little or no differentiation while more subtle mutations (DIF alleles) produce ovarioles with mixtures of egg chambers that have reached various degrees of differentiation (King et al. 1986; Steinhauer and Kalfayan 1992; Sass et al. 1993).

## otu

-1331 GAATTCATAGTCGTTGCGTTTTGCACACTCGCAAGATAACCAACTAACGACATTTACTAACAATAAACAAAAACATAACTTTACACGAGA -1242

-1241 ACACAAAAAACACAAAAAAAACAGGAAAAACAAAGGCACACACAGTCACACACTCACATCTCTCCAGACAACATTTGTGCGGGTAAC -1152

-1151 AGCGCGAACTGAAAGTTTGCTCCTGGCTTCATTGACTCGCAATTTGAACTGAGTCTGATGAACAAGAACAACAGTGCGCCGTGTGGAAA -1062

-1061 GCGGCATTTTCCACCCCTAAAAAGCGGCCAGCAACAACAGCAACGACAGTAAACAAGAACAATTTGAAGGTAACAGAAAATTTGGGGAT -972

-971 GACACGGAACAGATGATGCCGCTATCGGTGTCATCGATAGACGGCGATAACAGGAGTTTTTAAACCGCTCAGCAATATATTCAAGTATA -882

-881 TCATACACTTGTGATTTTCATTTAGAAAATTCAACAAGATCAGATATATTTATTTGTTGATAAAAATCACGAACCAACTCCATTGATT -792

-791 CATTTCGCACATCACTATTGCCCAATTTGTTGTCGGCATCCTCCAGGCACTGGAAGTTCGTTCTTATACTTTTCGTTGCATTCTA -702

-----  
 --> -----  
 -----  
 ||=P3  
 ! ||=P1,P2,P4 . -->-668 --->-659/658

-701 GTTCGCGGGTCTCTGAAAGGCTAGATCGCGCCATTCGCTCAATTTCTCGTGAACGGTGTAGGTGCGGATGCCAGTGTTATTTTTAA -612

-611 TTGTTAATTTAATTGTTAACTATTATAAAAATAGAAATTTGTACAACAGAAGACGACAGACAACCCGGTAATATCTCGATTTCGATT -522

-521 TAACTGTATTAGTTGAAACATTTATAGTAACGGTAATTTGTCAAGTGACGAAATTAECTAATTAAGCGCAGCATGAGAGGCTTTTAAATC -432

-431 ATTAATTTTAAACAAATATTTAATTTTCATCAGCTTCATCACATTTAATTTTGCTCTTTTGCTTCATTTGCCTTTCTACTGCGCCATCT -342

-341 TGAATTCGACAGTGATATTGTCATCTCGCTCGAAGCCCGCTTGATGGAGTCGGTTAATAATGGAATATATTTGTTATGCAGCAAA -252

-251 TTTGCTTTAAAACATTAAGTTAAAAAACTATAACAATAGTAAACATAAAAATAAGTAATAAAGCTTAGTATGCGCACTTCTTAGTGAAA -162

-161 CGACAATAGATAGCAGTTGAAAAGTGATTGTGAAGGTCAAATAGATCGAGGTCAGGGCCCTCTCTAACTGTTAATTGTGCAATACTTGT -72

-71 ATTTCAAAGGGAAAACATGACAAAAAAAATGAAATGAATAAAATTTAAGTTTCTCGATTCAGAGTCGCCATGGACATGCAAGTGACG -18  
 MetAspMetGlnValGln (6)

19 CGCCCCATTACGTACGGCAGCCGCGCAGGCCCGGATCCGTATGATCAGTATCTGGAGAGCCGTGGACTTACCCTAAGCACACGGCCCGG 108  
 ArgProIleThrSerGlySerArgGlnAlaProAspProTyrAspGlnTyrLeuGluSerArgGlyLeuTyrArgLysHisThrAlaArg (36)

109 GACGCCCTCAGTTTGTCCGTGTGATCGCCGAGCAGATGTACGACACCCAGATGCTGCCTACGAGATTCGGCTAGAGTGGCTCCGCTTC 198  
 AspAlaSerSerLeuPheArgValIleAlaGluGlnMetTyrAspThrGlnMetLeuHisTyrGluIleArgLeuGluCysValArgPhe (66)

199 ATGACCCTAAAACGACGCATCTTTGAGAAGGTAGGCCCTCTAACAATCACACATTTTGTAAAAAAAAGAAATAATTTTATATATCCC 288  
 MetThrLeuLysArgArgIlePheGluLys (76)

289 AGGAAATTCCTGCGCATTTTCGATAGTACATCGAGACATGTCCAAGCCCAAGACATATGGAACCATGACAGAACTACGCGCTATGCTCT 378  
 GluIleProGlyAspPheAspSerTyrMetGlnAspMetSerLysProLysThrTyrGlyThrMetThrGluLeuArgAlaMetSerC (106)

379 GCCTATATCGGTAATTAATCCTTAGTACTATTTTCTATTAACACTACAATATATATGATTTCTGTACGACTCCAGCCGCAATGTTATC 468  
 vsLeuTyrAr gArgAsnValIle (113)

(continued)

469	CTGTATGAGCCCTACAACATGGGACCAGCGTCGTTTTTAATCGTCGCTATGCGGAAAACCTCCGTGCTCTTCCAACAATGAGAATCAC LeuTyrGlu <u>Pro</u> TyrAsnMetGlyThrSerValValPheAsnArgArgTyrAlaGluAsnPheArgValPhePheAsnAsnGluAsnHis	558 (143)
559	TTTGATTCGGTTTATGACGTTGAATATATAGAAAGACGCCCATTTGTCAATGTACGTAGCCTATTAATATATCCAATTTGCTTTTTGT PheAspSerValTyrAspValGluTyrIleGluArgAlaAlaIleCysGlnS	648 (161)
649	ATATGTACGTTGCTTTCAGCAATGCCTTTAAGTTGCTGTACCAGAAGCTTTTCAAATGCTGACGTATCCTTTGCTGGGAGATTATG erIleAlaPheLysLeuLeuTyrGlnLysLeuPheLysLeu <u>Pro</u> AspValSerPheAlaValGluIleMet	738 (184)
739	TTGCATCCACACACCTTCAATTGGGATCGCTTCAATGTGGAGTTCGATGACAAGGGCTATATGGTTTCGATTTCATTGACCCGATGGACGA LeuHis <u>Pro</u> HisThrPheAsnTrpAspArgPheAsnValGluPheAspAspLysGlyTyrMetValArgIleHisCysThrAspGlyArg	828 (214)
829	GTTTTTAAGCTTGATCTGCCAGGGGACAAAACGCATACATGGAAAACATAAGCTGTGCAATTTCCATAGCACCAAATGGAAATCAGAGC ValPheLysLeuAspLeu <u>Pro</u> GlyAspThrAsnCysIleLeuGluAsnTyrLysLeuCysAsnPheHisSerThrAsnGlyAsnGlnSer	918 (244)
919	ATTAATGCTCGAAAGGGAGGCCGGCTGGAGATTA AAAACAGGAGGAGCGAAAGGCATCCGGCAGCAGTGGCCACGAACCAAACGATCTG IleAsnAlaArgLysGlyGlyArgLeuGluIleLysAsnGlnGluGluArgLysAlaSerGlySerSerGlyHisGlu <u>Pro</u> AsnAspLeu	1008 (274)
1009	TTGCCCATGTGCCAAACCGATTGGAGTCTGTGCCCGCAGCTGTAGATGATGGTCAGTAGAGGTGGTTTCAAACATCAAATGCTTAC Leu <u>Pro</u> MetCys <u>Pro</u> AsnArgLeuGluSerCysValArgGlnLeuLeuAspAspG	1098 (293)
1099	ATAATACTCTCTTTTAGTATCTCTCCGTTTCCCTACAAAGTGCCCAAGTCCATGGACCCCTATATGTATCGTAATATAGAATTTGATT IlyIleSer <u>Pro</u> Phe <u>Pro</u> TyrLysValAlaLysSerMetAsp <u>Pro</u> TyrMetTyrArgAsnIleGluPheAspC	1188 (317)
1189	GCTGGAACGATATGCGCAAGGAGGCCAAGCTTTATAATGTCTACATAAATGACTATAACTTTAAGGTAACCTGTGCAGAACATTTGGATTA ysTrpAsnAspMetArgLysGluAlaLysLeuTyrAsnValTyrIleAsnAspTyrAsnPheLys	1278 (338)
1279	TCGTTAGCACACATACACACGCACCAACACACGTTTCATGTCAACCACCCATCCAAATTAACACCCCTTTCATTTTGATCTATACACTG A=13	1368
1369	GATACACCTTATACTTTACTATACATGTATGTCTTCCCTATCCTTCCTCGTCTCGTCGCGGTGTATTGTTTCCAGGTGGCGCCAA ValGlyAlaLys A=11	1458 (342)
1459	GTGCAAGGTGGAATTGCCGAACGAAACGGAGATGTACACGTGCCACGTTCAAATATCTCCAAAGATAAGAATTA CTGCCACGCTTTGT sCysLysValGluLeu <u>Pro</u> AsnGluThrGluMetTyrThrCysHisValGlnAsnIleSerLysAspLysAsnTyrCysHisValPheVal Tyr	1548 (372)
1549	TGAGAGGATTGGCAAAGAGATAGTGGTACCTCTCTTTTATCTGATTTTCTAGACCCCTGCAGAGAAATGCAAAAATTTTCGATTAGAAA lGluArgIleGlyLysGluIleVal	1638 (380)
1639	CGATTATCATATTAACAATTAGTTAAATTTGTTAAAGTTAGTTAAAAGTATATTAATTTGTGGCCCAATGAACGGTATATAAGTCTAT	1728
1729	AAAATAATTGATCTGCAAGGGCTAAAAATGTCGGTATCCGAAGCTAATTGTAACATTTTCGCTTAAATAGAGACTTACTAATATACAA	1818
1819	ACATATCTGTTGGCTTAGTCCCGTATGAATCGCTCCATCCCTGCCGCCAGATGAGTACCGCCCATGGTGGTTCGCATTTCCGCTATCAT Val <u>Pro</u> TyrGluSerLeuHis <u>Pro</u> Leu <u>Pro</u> AspGluTyrArg <u>Pro</u> TrpSerLeu <u>Pro</u> PheArgTyrHis	1908 (404)
1909	CGCCAGATGCTCGCTTGGCTTGGCCAGTATGCCGGTAAGGCCAACCAAGTCTTCCAAATGGAAGAAGAACAAGCTTTCGAAATGGAC ArgGlnMet <u>Pro</u> ArgLeu <u>Pro</u> Leu <u>Pro</u> LysTyrAlaGlyLysAlaAsnLysSerSerLysTrpLysLysAsnLysLeuPheGluMetAsp	1998 (434)
1999	CAGTATTTTGAGCACAGCAAGTGTGATTTGATGCCCTACATGCCCGTGGACAAATGCTATCAGGGTGTGCACATTCAGGACGATGAGCAG GlnTyrPheGluHisSerLysCysAspLeuMet <u>Pro</u> TyrMet <u>Pro</u> ValAspAsnCysTyrGlnGlyValHisIleGlnAspAspGluGln	2088 (464)
2089	CGGGATCATAATGATCCTGAACAAAATGACCAGAACCAGACTACGGAGCAGCGGGATCGTGAAGAACCAGGACAGAGCAGAACACCCAG ArgAspHisAsnAsp <u>Pro</u> GluGlnAsnAspGlnAsn <u>Pro</u> ThrThrGluGlnArgAspArgGluGlu <u>Pro</u> GlnAlaGlnLysGlnHisGln	2178 (494)

2179	CGCACGAAGGCATCAAGGGTTCAGCCGAGAACTCGAGTTCCAGCCAAAACCAGGAGGTTTCGGGTTCGGCTGCCCGCCACCCACTCAG ArgThrLysAlaSerArgValGlnProGlnAsnSerSerSerSerGlnAsnGlnGluValSerGlySerAlaAlaProProProThrGln	2268 (524)
2269	TATATGAATTACGTGCCAATGATACCCAGTCTCTCGGTCATTTACCGCCACCTTGGCCTGCATCTCCGATGGCTATTGCCGAGGAGTTT TyrMetAsnTyrValProMetIleProSerArgProGlyHisLeuProProProTrpProAlaSerProMetAlaIleAlaGluGluPhe	2358 (554)
2359	CCGTTCCCATTTACGGAACCCCGCATCCACCGCCAACCGAAGGTTGTGTATACATGCCATTTCGGTGGTTATGGTCCACCACCCCGGGA ProPheProIleSerGlyThrProHisProProProThrGluGlyCysValTyrMetProPheGlyGlyTyrGlyProProProProGly	2448 (584)
2449	GCTGTTGCTTTATCGGGACCGCATCCATTTATGCCGCTTCCTTCTCCACCCTAAATGTTACCGAATTTGGCGAGCCACGCTGTTCTCTA AlaValAlaLeuSerGlyProHisProPheMetProLeuProSerProProLeuAsnValThrGlyIleGlyGluProArgArgSerLeu	2538 (614)
2539	CACCCAAACGGTGAAGATTTGCCCGTGGATATGGTGACTTTGAGATACTTCTACAACATGGGCGTGGATTTGCATTGGCGCATGTCGCAC HisProAsnGlyGluAspLeuProValAspMetValThrLeuArgTyrPheTyrAsnMetGlyValAspLeuHisTrpArgMetSerHis	2628 (644)
T=5		
2629	CACACGCCGCTGATGAACAGGAAATGTTTGATACCATCAGCAGAACAACACTGATCAACAGGCAGGACGGACTGTAGTCATTGGCGCC HisThrProProAspGluLeuGlyMetPheGlyTyrHisGlnGlnAsnAsnThrAspGlnGlnAlaGlyArgThrValValIleGlyAla	2718 (674)
End		
T=14		
2719	ACAGAGACAATTTGACTGCCGTGGAGTCAACACCACCACCTTCGCCAGAGGTGGCAAATGCCACAGAGCAGTCACCGCTTGAGAAAAGT ThrGluAspAsnLeuThrAlaValGluSerThrProProProSerProGluValAlaAsnAlaThrGluGlnSerProLeuGluLysSer	2808 (704)
End		
2809	GCCTACGCCAAGCGCAATTTGAATTCGGTTAAGGTGCGCGGCAACGTCGGAGCAGCTGCAAGATATTAAGGATTCGCTGGGGCCACCG AlaTyrAlaLysArgAsnLeuAsnSerValLysValArgGlyLysArgProGluGlnLeuGlnAspIleLysAspSerLeuGlyProAla	2898 (734)
2899	GCATTTTGGCCACTCCAACGCCATCGCCAAGCTCGAATGGCAGTCAGTTAGTTTCTATACTACTCCATCGCCGCATCATCACCTGATA AlaPheLeuProThrProThrProSerProSerSerAsnGlySerGlnPheSerPheTyrThrThrProSerProHisHisHisLeuIle	2988 (764)
2989	ACACCCGCGAGGTTGCTCCAACCGCCGCCACCCGATATTCTACCACAAGCGGGACCACCACAGCTAGGGGGAGCAGCTCAAGGA ThrProProArgLeuLeuGlnProProProProProIlePheTyrHisLysAlaGlyProProGlnLeuGlyGlyAlaAlaGlnGly	3078 (794)
3079	CAGGTAGGAGTGATACATGCACAAACAATTCAAAATATCTATAGGCAATCGACTCGACCATTTTTAGACTCCCTACGCCTGGGGCA Gln	3168 (802)
ThrProTyrAlaTrpGlyM		
3169	TGCCAGCTCCGGTGGTGTCCCCCTATGAGGTGATCAACACTATAACATGGACCCGTCGGCTCAGCCACAACAAGCAGCCAGCCCCCT etProAlaProValValSerProTyrGluValIleAsnAsnTyrAsnMetAspProSerAlaGlnProGlnGlnGlnProAlaProL	3258 (832)
3259	TGCAACCAGCTCCCTTATCTGTCCAATCTCAGCCGGCAGCTGTCTATGCTGCAACGCGTCATCACTAAACAAAGAAAGAAAAAAGG euGlnProAlaProLeuSerValGlnSerGlnProAlaAlaValTyrAlaAlaThrArgHisHisEnd	3348 (853)
3349	GAGCGGGGCAAAAACAGATCACTTGAAGAGAGAGGCCATACAGATCGAAGGCACTACATCCATTGCAATTAACGGCTTTTAAATTT 3438	
3439	TAATCTCACTTTTAAATTTGTAGTTAACTTTTATAGGCCATAAGCGTTGGCGCTCTATCATAAACCATTGAGCTTCTGTACAACAATCG 3528	
3529	ATTGCATAACCTAACGCAAATGTCAACCCAACCTTCAATTTAAAAATGTAATTTAACGTAATTTTATGCGAATTTTTTAAAGTTAGCCGT 3618	
3619	CACGAAATCAAAGAACCACCTATTTATATGATTTTAAAAACCTTCTAACAAAAATATCTACATACTACTACTATATATATACATA 3708	
3709	TATATATATATATATTTATGTGCTCGCTGTTCCGGCTAGAGACTCACCTATGTAAGGTGACCATCAAAAATTAACCATAAATAAAAA 3798	
3799	AGATTCACCTGCAG 3812	-----

|(A)<sub>n</sub>



By *in vitro* mutagenesis of *otu*, two constructions were prepared, one that could produce only the 104 kD protein and another that could produce only the 98 kD protein. When introduced into QUI mutants, the 104 kD protein restores fertility. The 98 kD protein is unable to rescue the QUI mutant phenotype but does restore some fertility to ONC or DIF type alleles. Thus, it would appear that the 104 kD protein is capable of carrying out all *otu* functions while the 98 kD protein can perform some of the late oocyte maturation functions but is unable to carry out early oocyte maturation functions or those required for controlled cell proliferation (A. R. Comer and L. L. Searles, personal communication).

### Gene Organization and Expression

Open reading frame, 811 (98 kD protein) or 853 (104 kD protein) amino acids depending on splicing; mRNA, 3,045–3,230 bases, depending on the start site and splicing. The most common RNA is approximately 3.2 kb, but other cross-hybridizing RNAs occur. The 5' end was defined by S1 mapping, primer extension, and the sequencing of two cDNA clones. Several sites are used for transcription initiation, the main ones being those at positions –668, –659 and –658 (*otu* Sequence). There is no TATA box associated with any of the 5' ends. The 3' end was defined from a cDNA sequence that contained a poly-A tail. There are eight introns: one is in the leader between positions –541 and –7, the others are after the Lys-76 codon, in the Arg-109, Ser-161 and Gly-293 codons, and after the Lys-338, Val-380 and Gln-795 codons. The 126-base exon starting with the Val-339 codon is often spliced out to produce mRNA that codes for the 98 kD protein (*otu* Sequence, Fig. 23.1) (Champe and Laird 1989; Steinhauer et al. 1989; Comer et al. 1992; Steinhauer and Kalfayan 1992).

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*otu* SEQUENCE (previous pages). Strain, Canton S. Accession, M30825 (DROOTUA) and X13693 (DROOTU). Arrows above the sequence, between –720 and –658, indicate possible sites of transcription initiation; the exclamation mark at –688 marks the 5' end of two independently obtained cDNAs. Several mutations are indicated in the sequence: *otu*<sup>5</sup> and *otu*<sup>14</sup> cause premature termination, and homozygotes accumulate smaller proteins (both alleles belong to the DIF class); *otu*<sup>13</sup> is unable to produce the 104 kD protein because it has a disabled acceptor site in exon 7; and *otu*<sup>11</sup> has an amino-acid substitution in exon 7 (both *otu*<sup>11</sup> and *otu*<sup>13</sup> affect the 104 kD protein but not the 98 kD protein and both are ONC alleles) (Steinhauer and Kalfayan 1992). The four *P* element insertions near the 5' end seem to affect transcription, and the severity of their phenotypes is generally proportional to the size of the insertion: *otu*<sup>P1</sup> (2.9 kb) is a QUI allele, *otu*<sup>P2</sup> (2.0 kb) is an ONC allele, and *otu*<sup>P3</sup> (0.6 kb) and *otu*<sup>P2</sup> (0.5 kb) are DIF alleles (Sass et al. 1993).

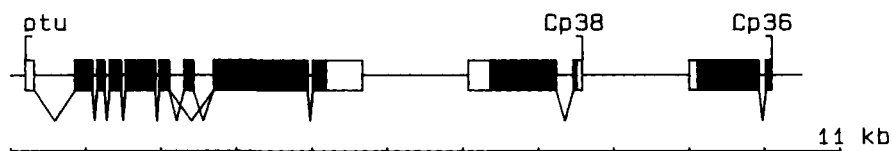


FIG. 23.1. *otu* and neighboring genes *Cp36* and *Cp38*

The *otu* gene is 0.06 map units away from the chorion protein gene *Cp38*, closer to the centromere, and transcribed convergently with *Cp38*, toward the telomere; the two 3' ends are approximately 1.4 kb apart (Fig. 23.1). *otu* is amplified, together with the chorion genes, in follicular cells, but it is not expressed in those cells (Parks and Spradling 1987; see also Chorion Protein Genes).

### *Developmental Pattern*

The predominant 3.2 kb transcript is present mainly in female pupae and adults. It occurs in nurse cells and oocytes, and the peak of expression is egg chambers between stages 8 and 10. This transcript is found at much lower levels in female heads and thoraxes and in male testes along with other cross-hybridizing transcripts. Given that null mutations have no effect other than on female sterility, it is likely that the non-ovarian transcripts lack any function (Mulligan et al. 1988).

### *Promoter*

Studies of a reporter gene under the control of an *otu* fragment that extends from 452 bp upstream of the transcription initiation site to the end of the first exon, showed expression, in ovaries, in nurse cells and oocytes as well as in the germarium. In males, expression was detected in the anterior tip of the testes, in the region of stem cells and primary spermatocytes (Comer et al. 1992).

Constructions with 310 bp of upstream sequence and the complete transcribed region produced apparently normal levels of 3.2 kb RNA and rescued *otu* mutations. Similar constructions with only 190 bp of the promoter region, however, were unable to support gene expression (Comer et al. 1992).

### **References**

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# 24

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## 6-Phosphogluconate Dehydrogenase Gene: *Pgd*

**Chromosomal Location:**

X, 2D4-6

**Map Position:**

1-0.6

**Product**

6-Phosphogluconate dehydrogenase (6-PGD) (E.C. 1.1.1.44), a member of the pentose shunt.

*Structure*

The sequence of *Drosophila* 6-PGD is 50% identical to prokaryotic 6-PGD and 60–70% identical to the porcine and ovine enzymes (Fig. 24.1) (Scott and Lucchesi 1991). 6-PGD is a homodimer; the monomer has a  $M_r$  of approximately 53 kD (Williamson et al. 1980).

*Function*

6-PGD is responsible for the oxidative decarboxylation of 6-phosphogluconate (6-PG) to yield ribulose-5-phosphate and reduced nicotinamide adenine dinucleotide phosphate (NADPH); these two products are important for the biosynthesis of ribose and lipids, respectively (Wood 1985).

*Tissue Distribution*

The specific activity of the enzyme increases during the larval stages to reach a maximum early in the third instar. Activity diminishes late in the third instar and early pupal stages, then climbs again in late pupae and adults (Williamson et al. 1980). In larvae, highest activity is observed in fat bodies and actively dividing imaginal cells (Gutierrez et al. 1989; Scott and Lucchesi 1991).

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1                               50                               100
Dm pgd MSGQADIALI GLAVMGQQLI LNMDEKGFVV CAYNRTVAKV KEFLANEAKD TKVIGADSLE DMVSKLKSPPR KVMLLVKAGS AVDDFIQQLV PLLSAGDVII
Ovine .MAQADIALI GLAVMGQQLI LNMNDHGFVV CAFNRTVSKV DDFLANEAKG TKVLGAHSLE EMVSKLKKPR RIILLVKAGQ AVDNFIEKLV PLLDIGDIII
CON ---QADIALI GLAVMGQQLI LNM---GFVV CA-NRTV-KV --FLANEAK- TKV-GA-SLE -MVSCLK-PR ---LLVKAG- AVD-FI--LV PLL--GD-II

101                             150                             200
Dm pgd DGGNSEYQDT SRRCDELAKL GLLFVGSVGS GGEGARHGP SLMPGGHEAA WPLIQIPFQA ICAK.ADGEP CCEWVGDDGA GHFVKMVHNG IEYGDMLIC
Ovine DGGNSEYRDT MRRCRDLKDK GILFVGSVGS GGEDGARYGP SLMPGGNKEA WPHIKAIFFQ IAAKVGTEGP CCDWVGDDGA GHFVKMVHNG IEYGDMLIC
CON DGGNSEY-DT -RRC--L--- G-LFVGSVGS GGE-GAR-GP SLMPGG---A WP-I--IFQ- I-AK---GEP CC-WVGD-GA GHFVKMVHNG IEYGDMLIC

201                             250                             300
Dm pgd EAYHIMKS.L GLSADQMADE FGKWSAELD SFLIEITRDI LKYKDGKG.Y LLERIRDTAG QKGTGKWTAI AALQYGVVPT LIGEAVFSRC LSALKDERVQ
Ovine EAYHLMKDV LGLGHKEMAKA FEEWNKTELD SFLIEITASI LKFQDADGKH LLPKIRDSAG QKGTGKWTAI SALEYGVVPT LIGEAVFARC LSSLKDERIQ
CON EAYH-MK--L GL----MA-- F--WN--ELD SFLIEIT--I LK--D--G-- LL--IRD-AG QKGTGKWTAI --AL-YGVVPT LIGEAVF-RC LS-LKDER-Q

301                             350                             400
Dm pgd ASSVLKGPST KAQVANLTKF LDDIKHALYC AKIVSYAQGF MLMREAAREN KWRLNYGGIA LMWRGGCIIR SVFLGNKDA YTSQPELSNL LLDFFFKKAI
Ovine ASKKLKGPN IPFEGDKSF LEDIRKALYA SKIISYAQGF MLLRQAATEF GWTLYNYGGIA LMWRGGCIIR SVFLGKIDA FDRNPGLQNL LLDFFFKSAV
CON AS--LKGP-- -----F L-DI--ALY- -KI-SYAQGF ML-R-AA-E- -W-LNYGGIA LMWRGGCIIR SVFLG-IKDA ----P-L-NL LLDFFFK-A-

401                             450                             485
Dm pgd ERGQDSWREV VANAFRWGIP VPALSTLSF YDGYRTAKLP ANLLQAQRDY FGAHTYELLG QEQGFHHTNW TGTGGNVSA SYQA*
Ovine ENCQDSWRR ISTGVQAGIP MPCFTTALS F YDGYRHAML ANLIQAQRDY FGAHTYELLA KPGQFIHTNW TGHGGSVSS SYNA*
CON E--QDSWR-- -----GIP -P---TALS F YDGYR-A-LP ANL-QAQRDY FGAHTYELL- --GQF-HTNW TG-GG-VS-S -Y-A-

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FIG. 24.1. Comparison of the sheep (Accession, 60195) and *Drosophila* (Dm) sequences. There is 72% overall identity between the proteins. Sequences aligned with the GCG *Pileup* program.

## Pgd

XhoI  
 -1206 CTCGAGCAGTTCAGTTCCTGAAGTGAGTTGCGCCACCTTTGTCTTCTGAGCGTTACCAATCTGTTCAACAACTTATTTCCCATAGC -1117  
 -1116 TCCCCCATTTCCGGATTTCCTTCTACATGCTCATCGAGACCTCGGGCAGCAACGGTGACCACGACGAGGAGAAGATCAACCAGTTCATT -1027  
 -1026 GGGGACGGTATGGAGCGTGGCGAGATCCAGGATGGCACCGTAACCGGTGATCCCGGCAAGGTGCAGGAGATCTGGAAGATCGCGAAATGG -937  
 -936 TGCCGCTGGGTCTGATCGAGAAGAGCTTCTGCTTCAAGTACGACATCTCGCTGCCTCGCGGACTTCTACAACATTGTGGACGTGATGC -847  
 -846 GAGAGAGGTGCGGTCCCTTGGCCACAGTTGTCTGCGGATACGGCCATCTGGGGGACTCTAATCTGCACCTGAACGCTCTCTGCGAGGAGT -757  
 -756 TTAACGACGAGATCTACAAGCGGGTGAACCCCTTCTGTCTACGATACACCTCCAAGCTGAAGGGCAGCATTATGGCGGAGCAGGCATTGG -667  
 -666 CTTCTGAAGAAGGACTACCTGCACTACTCCAAGGACCCGGTGGCCATTGGCTACATGCGCGAGATGAAGAAGCTGCTGGACCCCAACAG -577  
 -576 CATCTCAATCCCTACAAGGTGCTTAAGTGAAGGCTTCTACCTAATAGATTCTATTTTTTTTGTGTGTGAATTTTCATAACCTTATA -487  
 -486 ATACAGAAATGGCATTAGAAGTGAATTTTGTAACTTGTGAAGTTAAAAGGACCATCATATTTGGCAGCAAACCAATGGGCAAACTTA -397  
 -396 CTTATAAAATAGTCCGAAAAATAGTATATACCAGTTTTTACAGTACCACATTATAGTACTCGGAGGTAATAATAGAAAAACACTATC -307  
 -306 TTTGCATTTACTGTTACTACTACGAAGCACTATATTTAGTAGCAGTACTCATTAGAGTCCACTCACAAAATTAGCACCAACCGGAGTAAT -217  
 -216 TGGTCAAGGATCGGCATAGCTTCAAACCCGAAGTCAAAGTCAAACCTGCCGCCCTGCGAAAAGCTTCGCGAGTGGAGCTTTTCTGCACT -127  
 -126 TATCGATAGCTAACATTGTGGCGGACTATCGATCGACGAGCTGCCGCTTAACAGTGCCATATATAGATTGAACATTAGGAGCTCAAAT -37  
 ---  
 -->-34  
 -36 CATTGTTGGAACACAACCACAAGAACACAGAAACATGAGCGGGTGAAGTAGAGGGAAATTCCTTTTCCCGGAGTTTTCCGCGATCC 53  
 MetSerG1 (3)  
 54 TAACGTCGCCCATTTCCGGATTTCTTCCAGACAAGCGGATATTGCCCTCATCGGCCTGGCCGTCATGGGCCAAAACCTGATACTCAACAT 143  
 yGlnAlaAspIleAlaLeuIleGlyLeuAlaValMetGlyGlnAsnLeuIleLeuAsnMe (23)  
 144 GGACGAGAAGGGATTCTGGTGTGCGCTACAACCCACGGTGGCCAAAGGTCAAAGAGTTCTCGCCAATGAGGCTAAGGACACCAAAGT 233  
 tAspGluLysGlyPheValValCysAlaTyrAsnArgThrValAlaLysValLysGluPheLeuAlaAsnGluAlaLysAspThrLysVa (53)  
 234 GATTGGAGCCGACTCGCTCAGGACATGGTCTCCAAGCTGAAGAGCCCCGGAAAGTCAATGTGCTGGTCAAGGGTGAGTTGCATATCCA 323  
 IileGlyAlaAspSerLeuGluAspMetValSerLysLeuLysSerProArgLysValMetLeuLeuValLysA (78)  
 324 AATTCAGCGGCTGGGTAGCGCAGAGCATGAAAACCCATTGAAACCTGCTGCAAGCGATCGCTGTGTTGGTGACTCAACTTACATGTGTG 413  
 414 CGCGGTGCTGTGGAATTTGGTAAAAAGTGAAGCAAGTCAATGATGACGATTTTTGCGGCTCATATCCAATGTGCAAAGGGGAAC 503  
 504 GATAGGATAAGCAGGTGAGCTCAATGCTTAAAGTTTGAAGTCTATAAAGAGCTTGAATTTCTGTCTAGTTTTCAAGTCAAACCTATCGCA 593  
 594 TACAAAACCTACGAAATGCCATCCCTATCATTGTACAAAAAGAACTCCTAACCCAGACTTAGTGGTTAAGCCGAGCTCAATGATCTC 683  
 684 TAAACAGTTGTTTTTGTGTTTACTCCACCCCTCACCGTTTTCTGCGCTCCCTCTCTCTACTTCTCTTTAAAACCCGACTTCTGA 773  
 774 TAAAAGTTTATAAATGGATCAGTCCATTTTCGAAAACCGTAACCAAGTGTGGCGTGAGTTTTGTCTAATCACATAGTTGTGGTAAGC 863  
 864 TGCTCCACTTACCTAAACCATCGAGCGAACCCATCAGGTGATTTCCAGGTCACCTCACGCTTCGTCTACCACTCTCGCGTGTCCGAAAC 953

(continued)

954	TCTGCTCACCTCTAGATCGGCGTGCCCGGCTTATCTGTTCTGTCGAAAGCAACAACACGCGGCGCAGAGAGAAATCTTTGACATTCATA	1043
1044	ATAGGTCACACAAAATGGGCGATTTTCAGGTGGATTTACTCGGATTTGACCAGCCGAAAAACCTACATATTCCTCTTCTGCGAGTTGCCA	1133
1134	GGCCAGTGAGTCATTTCTGCTGGAGACTGCTCCTTAGAAGAATACAGTGCGGGTCAATAACATATGTACATAGCTCTGGAGGTTTTGTG	1223
1224	CTGAACATATGTAGATTTGAAAGTTGCGTGACAGGTTGTGCGAATCCACATTCACAGGGTGGGTGGGAGTAAGGATGACGACACAAAA	1313
1314	AGCTAGTTGGTCATTGAACAGAGCGAGTCCAACAATCTTGACCCTAGTGTGCCCCACAAACCACCACCAACGACCCTAGATAGATAGA	1403
1404	TCAATGGTAGTATCGCCACGACTCGTTGGCCTTATCTGGGTCCACTGCGCTGGAGAAGTGTCTACCCGGCGCTAGGGGAATTCCTCATCG	1493
1494	GGGTTCTCAAAGCTCAACTATCGTAGACTCATTTTCAAAGCGTCTTTAGCGAGCGCCAGTCTTTTAAAGTAAAGAAATCTTCGATTT	1583
1584	AGCCAGAAAAGTAGAGCGTGCATTGGACAAGGTCGGTTGGTTGCTTTTGGAAAAGTCACTGTTTTGGAGGTACCCCTGGTGGCGAGGCGTG	1673
1674	ATCTGCTTTAATCGACTTACGCTAATCAGATGTAACCTCGATACAATTTTCAGCTGGAAAGTGCAGTGCAGACTTCATCCAGCAGCTGGT	1763
	1aGlySerAlaValAspAspPheIleGlnGlnLeuVa	(90)
1764	GCCGCTGCTTCCGCGGCGATGTGATCATCGATGGTGGCAACTCGGAGTATCAGGACACATCTCGCCGCTGCAGCAG6TTAGCCAAACT	1853
	1ProLeuLeuSerAlaGlyAspValIleIleAspGlyGlyAsnSerGluTyrGlnAspThrSerArgArgCysAspGluLeuAlaLysLe	(120)
1854	TGGCTGCTTTCGTCGGATCCGGCTGAGCGGTGGCGAG6AGGGCGCCGCCACGGACCTCGCTGATGCCGGCG6ACACGAGGCCGC	1943
	uGlyLeuLeuPheValGlySerGlyValSerGlyGlyGluGluGlyAlaArgHisGlyProSerLeuMetProGlyGlyHisGluAlaAl	(150)
1944	GTGGCCCTTATCCAACCCATCTTCCAGCGGATCTCGCCCAAGGCCGAG6TGAACCTGCTGCGAGTGGTGGGCGATGGAGGCCCGGG	2033
	aTrpProLeuIleGlnProIlePheGlnAlaIleCysAlaLysAlaAspGlyGluProCysCysGluTrpValGlyAspGlyGlyAlaGly	(180)
2034	TCACTTCGTCAGATGGTGCACAACGGCATCGAATACGGTGACATGCAGCTGATCTGCGAGGCGTACCACATCATGAAGAGCCTGGGACT	2123
	yHisPheValLysMetValHisAsnGlyIleGluTyrGlyAspMetGlnLeuIleCysGluAlaTyrHisIleMetLysSerLeuGlyLe	(210)
2124	GTCGGCTGACCAGATGGCAGACGAGTTCCGCAAGTGGAACTCGGCCAAGTGGACTTCCTTCTCATTGAAATCACGCGTGATATTCTTAA	2213
	uSerAlaAspGlnMetAlaAspGluPheGlyLysTrpAsnSerAlaGluLeuAspSerPheLeuIleGluIleThrArgAspIleLeuLy	(240)
2214	GTACAAGGACGGCAAAGTTATCTGCTGGAGCGGATTCGCGATACCGCCGGCCAGAAGGGCAGGG6CAA8TGGACGGCAATCGCTGCTCT	2303
	sTyrLysAspGlyLysGlyTyrLeuLeuGluArgIleArgAspThrAlaGlyGlnLysGlyThrGlyLysTrpThrAlaIleAlaAlaLe	(270)
2304	GCAGTATGGAGTGCTGTGACGCTAATGGCGAGGCGGTCTTCTCGCATGCCGTCTGCCCTGAAGGACGAGCGCTCCAGGCCAGCAG	2393
	uGlnTyrGlyValProValThrLeuIleGlyGluAlaValPheSerArgCysLeuSerAlaLeuLysAspGluArgValGlnAlaSerSe	(300)
2394	CGTGCTGAAGGGACCTCGACCAAGGCGCAAGTGGCCAACCTCACCAAGTTCTCGACGACATCAAGCAGCTCTCTACTGCGCCAAGAT	2483
	rValLeuLysGlyProSerThrLysAlaGlnValAlaAsnLeuThrLysPheLeuAspAspIleLysHisAlaLeuTyrCysAlaLysIl	(330)
2484	CGTGCTCTACGCCAGGGATTATGCTCATGCGAGAGGCGCCAGGGAGAACAAGTGGAGACTTAATACGGCGGCATTGCGCTGATGTG	2573
	eValSerTyrAlaGlnGlyPheMetLeuMetArgGluAlaAlaArgGluAsnLysTrpArgLeuAsnTyrGlyGlyIleAlaLeuMetTr	(360)
2574	GCGTGGCGGCTGCATATCCGACGCTCTTCTGGGCAACATTAAGGACGCGTATACGTCGACGCGGAGCTGTCTAATCTGCTGCTGGA	2663
	pArgGlyGlyCysIleIleArgSerValPheLeuGlyAsnIleLysAspAlaTyrThrSerGlnProGluLeuSerAsnLeuLeuLeuAs	(390)
2664	TGACTTCTCAAGAAGGCCATCGAGCGCGCCAGGACTGCTGGCGGAGGTGGTGGCCAATGCCITCCGCTGGGGCATTCCCGTGCCGGC	2753
	pAspPhePheLysLysAlaIleGluArgGlyGlnAspSerTrpArgGluValValAlaAsnAlaPheArgTrpGlyIleProValProAl	(420)
2754	CCTGTCTACCGCCTAAGCTTCTACGACGCTACCGCACGGCCAAAGCTGCCAGCCAACCTTGCTGACGCGCCAGGGATTACTTCGGCGC	2843
	aLeuSerThrAlaLeuSerPheTyrAspGlyTyrArgThrAlaLysLeuProAlaAsnLeuLeuGlnAlaGlnArgAspTyrPheGlyAl	(450)

2844	CCACACCTATGAGCTGCTGGGCCAGGAGGGTCAGTTCACCACACGAACTGGACAGGCACCGCGGCAATGTGTCCCGCAGCACTTACCA aHisThrTyrGluLeuLeuGlyGlnGluGlyGlnPheHisHisThrAsnTrpThrGlyThrGlyGlyAsnValSerAlaSerThrTyrGlu	2933 (480)
2934	GGCGTAGGTTCCACCTGCTCCACTTCCCGTTACACATTCCATGTATTGGCGCCGGTGTCTTAGATGTTCTTTTTTTCTGGAGTAC nAlaEnd	3023 (481)
3024	TTTAGTACTTATTTATACCATTAATATATATGTATGTATATAGAAATTCATAAATGTTGTTAAACATAACATTAAATTTGGTGTTTTTTG -----	3113
3114	CTAGCAAATGATTTTGATTCCCTTAGTTTCATGAATGCAAGTGCCATTTAAAAACAACATGCGTGTGGTGGTGTGTGTGTGTGTGT  (A) <sub>n</sub>	3203
3204	GGGTCGAGTCTTTTCGAGTTGTGTCTTCATCTGGAGACGCCTCCTGCTCCTTCTACCCTCCTCCCTGCTATTGTACTCTCTTCAGCTAG	3293
3294	CGCGCTTTTTTCGCTCCGATTTCCCTTAGTCGTCGAGGGCTCAGGGTCTCTTGTCTCTATAACCAAGTTGTGAGCGGAATACAGG	3383
3384	TGGCCGATGATTACCTGTGGACATTCAAAGGTTAATAAACTCAACCGGCTGATAAGCGAAAAAGGGGCAAAATGGTTACTTTTCGATTTCT	3473
3474	AATAGGATGGTAATTGAGTTTTCCATCCCATATTTGCAAAATCAGATATATATGATAAAATCTACTTTAAATATACATTAATATT	3560 SspI

*Pgd* SEQUENCE. Strain, *Canton S*. Accession, M80598 (DROPGD).

### Phenotype of Mutations

Two electrophoretic variants (A and B) have been described (Kazazian et al. 1965). *Pgd* null mutations are lethal due to the accumulation of 6-PG; viability can be improved by dietary manipulations that reduce 6-PG synthesis or by the introduction of a null mutation on a second gene in the pentose shunt, *Zwischenferment* (*Zw*). *Zw* is the structural gene for glucose-6-phosphate dehydrogenase, G-6-PD, the enzyme that precedes 6-PGD in the pentose biosynthetic pathway, and it is required for the synthesis of 6-PG (Hughes and Lucchesi 1977, 1978).

### Gene Organization and Expression

Open reading frame, 481 amino acids; mRNA length, 1,659 bases, in agreement with an RNA of 1.7 kb observed in gels. Primer extension and S1 mapping were used to define the major 5' end (there seem to be several minor transcription initiation sites as well). The 3' end was obtained from a cDNA sequence. There is a short intron in the Gly-3 codon and a long one in the Ala-78 codon (*Pgd* Sequence) (Scott and Lucchesi 1991).

### Promoter

In transgenic animals, a 4.7 kb fragment that extends 1,172 bp upstream of the transcription initiation site and 442 bp downstream of the poly(A) site is sufficient for apparently normal expression of *Pgd* in larvae. Removal of the small first intron does not significantly affect expression, but removal of the



larger second intron leads to a 10-fold reduction in enzyme levels. The second intron is specifically required for expression in the fat body, but apparently not necessary for expression in actively dividing imaginal cells. Expression in imaginal cells requires only a 421-bp segment immediately upstream of the transcription initiation site (Scott and Lucchesi 1991).

## References

- Gutierrez, A. G., Christensen, A. C., Manning, J. E. and Lucchesi, J. C. (1989). Cloning and dosage compensation of the *6-phosphogluconate dehydrogenase* gene (*Pgd*<sup>+</sup>) of *Drosophila melanogaster*. *Dev. Genet.* **10**:155–161.
- Hughes, M. B. and Lucchesi, J. C. (1977). Genetic rescue of a lethal “null” activity allele of *6-phosphogluconate dehydrogenase* in *Drosophila melanogaster*. *Science* **196**:1114–1115.
- Hughes, M. B. and Lucchesi, J. C. (1978). Dietary rescue of a lethal “null” activity allele of *6-phosphogluconate dehydrogenase* in *Drosophila melanogaster*. *Biochem. Genet.* **16**:469–475.
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- Scott, M. J. and Lucchesi, J. C. (1991). Structure and expression of the *Drosophila melanogaster* *6-phosphogluconate dehydrogenase* gene. *Gene* **109**:177–183.
- Williamson, J. H., Krochko, D. and Geer, B. W. (1980) *6-Phosphogluconate dehydrogenase* from *Drosophila melanogaster*, I. Purification and properties of the A isozyme. *Biochem. Genet.* **18**:87–101.
- Wood, T. (1985). *The Pentose Phosphate Pathway* (New York: Academic Press).

# 25

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## *paired: prd*

**Chromosomal Location:**  
2L, 33C1-2

**Map Position:**  
2-45

### **Product**

A DNA-binding regulatory protein of the homeodomain type important in establishing the segmentation pattern in early embryos.

### *Structure*

The following potentially important sequence features occur.

1. The segment between residues 27 and 154 has great similarity to regions in both *gooseberry* genes and has been designated the “paired domain” (Bopp et al. 1986). This is a DNA-binding region (Treisman et al. 1991).

2. A homeodomain occurs between Gln-213 and His-272 (Frigerio et al. 1986; Harrison 1991). The sequence similarities between *prd* and *gooseberry* extends 18 amino acids upstream of the homeodomain (Bopp et al. 1986). A Ser in position 9 of the recognition helix (H3 in *prd* Sequence) differentiates the binding specificity of PRD from that of the products of *bicoid* (*bcd*), and *fushi tarazu* (*ftz*), which have Lys and Gln, respectively, in that position. *In vitro*, the PRD H3 does not bind sequences derived from the “standard” homeodomain binding site (TAAT). It is able to bind to the sequence TTTGACGT but only if the C-terminal region of the protein is removed. *In vivo*, the latter may be a regulatory region that is moved out of the way by interactions with other molecules (Treisman et al. 1989).

3. The C-terminus of PRD is characterized by a high proportion of His and Pro residues called the “PRD repeat”. Using a DNA fragment from the PRD repeat, 11 other cross-hybridizing sequences were identified, one of which was *bcd* (Frigerio et al. 1986).

*prd*

-495 AGCTGAGACGCCCTGGGCGCAGCGAGACGGTTGCTAAATGGTTCGAGTCGAGCCAGAGCGAGATGCCGTTGTGGAGAGCGCTGCGA -406

-405 TTGGTCCGCGTAGTGGTTACCTGCCAAGTGACTGTGGGATATGGCCGACGCTGGGCCGTGGCTTCACAGAAAGCGAACGATCTTGGCCG -316

-315 ACGTTCGGATGGTGAAGTCAGTCAGGCACAGACTGCGCAGCGAGCCACCCGCATCTCGTCTCGTTCCTCGCTTCGCCTTCGCCTCCGT !-244 -226

-225 TTCATCTTTCCCATCGAGATTGCGAACTCACAGATACTAGATATTCGAAGTGAACCTAATCGGTTAATCAATACCTCGCAACGCTTACT -136

-135 TATGACTTTGACAAAGTGTCCAGACATTGTCCAAAATAAAGTGATATAATCAAGTGATACACGAACTTCGAGACTGAGTTAACACCGGT -46

-45 TTTGTGCCGGGACAAGCTTACGCATCTTGGAGCTCTCCAGAAACTATGACCGTAACCGCCTTTGCTGCCGAATGCACAGACCTTCTT 44  
MetThrValThrAlaPheAlaAlaAlaMetHisArgProPhePh (15)

45 CAATGGATTTCTACGATGCAAGGTGAGTGTCTATCGATCTTATAGAATCCAGCAAAGTCACCTTTCACAATTTACTTACTAAATATC 134  
eAsnGlyTyrSerThrMetGlnA (23)

135 AAAGCCTAGTTGATCATTTCATATATCTCCATTTCTAAACCTACTACCCAAGATCCCCTAAAGATCTCAGTTTGGGCCAAGGCGTCGG 224

225 CTACTCTCTAATGGCCATTAGTGTGCCCGGCGGGAGAGTCGCGCGCCTCTGACCTTCGACCTTAGCTCCGAGTTTCCCGTCTCCCGGGAA 314

315 GTCACCTCCGGTCGAAGGTGTCGTAATCAAGTGACACGCGCTCCGCTCTACCTAGCTAGTATTGGAAAAGCCTCTAAAATTTCCATTTT 404

405 CTCATCTTCTCATTCCAGACATGAACAGCGGCCAGGGGCGCTCAATCAACTAGGTGGAGTTTTCATCAACGGTCGCTCTTTGCCAAC 494  
spMetAsnSerGlyGlnGlyArgValAsnGlnLeuGlyGlyValPheIleAsnGlyArgProLeuProAsn (46)

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|---|2.45.17

495 AATATTCGCTTAAAATCGTCGAGATGGCCGGCAGTGGCATTGCGCCCTGTGTGATCTCCAGACAGCTACGTGTATCCCATGGCTGCGTA 584  
AsnIleArgLeuLysIleValGluMetAlaAlaAspGlyIleArgProCysValIleSerArgGlnLeuArgValSerHisGlyCysVal (76)

585 TCGAAGATCCTGAATCGTACCAGGAGACTGGCTCCATTAGACCAGGTGTGATCGGTGGCTCCAAGCCGAGGATAGCCACGCCGAAATC 674  
SerLysIleLeuAsnArgTyrGlnGluThrGlySerIleArgProGlyValIleGlyGlySerLysProArgIleAlaThrProGluIle (106)

675 GAAAACCGAATTGAGGAGTACAAGCGCAGTAGCCCGGGCATGTTCTCGTGGGAGATCAGGAGAAGCTGATCCGCGAGGGGTGCTGCGAC 764  
GluAsnArgIleGluGluTyrLysArgSerSerProGlyMetPheSerTrpGluIleArgGluLysLeuIleArgGluGlyValCysAsp (136)

A

765 AGGAGCACAGCACCATCTGTGTCGCGCATATCGCGCTGGTGCAGCGCCGAGATGCTCCATTGGACAATGATATGTCTTCTGCTCTGGA 854  
ArgSerThrAlaProSerValSerAlaIleSerArgLeuValArgGlyArgAspAlaProLeuAspAsnAspMetSerSerAlaSerGly (166)

-|PRD DOMAIN Thr

855 TCTCCGCGGGTGATGGCACCAAAGCATCGAGTTCTGTGGCTCCGATGTCTCCGCGGCCATCACAACAACGGCAAGCCCTCCGATGAG 944  
SerProAlaGlyAspGlyThrLysAlaSerSerSerCysGlySerAspValSerGlyGlyHisHisAsnAsnGlyLysProSerAspGlu (196)

A

945 GACATCTCAGACTGCGAAAGTGAGCCGGGAATCGCCTTGAAGCGCAAACAGCGCCGCTGCAGGACCACCTTTTCCGCTTCCAGTTGGAC 1034  
AspIleSerAspCysGluSerGluProGlyIleAlaLeuLysArgLysGlnArgArgCysArgThrThrPheSerAlaSerGlnLeuAsp (226)

|- \* \* Ile\* -----\*

1035 GAACTGGAACGCGCCTTCGAGCGACCCCAATACCTGATATCTACACCGTGAGGAGCTGGCCAGCGCACCACCTCAGGAGGCGACGC 1124  
GluLeuGluArgAlaPheGluArgThrGlnTyrProAspIleTyrThrArgGluGluLeuAlaGlnArgThrAsnLeuThrGluAlaArg (256)

-----H1 \* -----H2 \*

1125	ATCCAGGTGTGGTTTCAGCAACCGCGTGCCTCGTCTCCGCAAGCAGCACACCTCGGTCTCAGGCGGAGCACCTGGCGGAGCAGCTGCCTCA IleGlnValTrpPheSerAsnArgArgAlaArgLeuArgLysGlnHisThrSerValSerGlyGlyAlaProGlyGlyAlaAlaAlaSer *-----*-*-*-----*-*-*H3 * * *  HOMEODOMAIN	1214 (286)
1215	GTAAGCCATGTGCGCGCTCCAGCTCTCTCCAGTGTGGTATCAAGTGTGCCAGCATGGCTCCGCTGGCCATGATGCCGGATCCCTG ValSerHisValAlaAlaSerSerSerLeuProSerValValSerSerValProSerMetAlaProLeuAlaMetMetProGlySerLeu	1304 (316)
1305	GATCCAGCCACTGTGTACAGCAGCAATACGATTCTACGCGAGTCACGCCAACATTTCCGTATCCGCCGAGCTCCAATGGCCAGTAGT AspProAlaThrValTyrGlnGlnGlnTyrAspPheTyrGlySerHisAlaAsnIleSerValSerAlaAlaAlaProMetAlaSerSer	1394 (346)
1395	AATCTATCGCCCGGAATTACAACCACGCCACCAGCCACCACCTCAGTTCTACAATCCAGCGCTAACACAGCCAGCTACATAATGCCGGGT AsnLeuSerProGlyIleThrThrThrProProHisHisHisGlnPheTyrAsnProSerAlaAsnThrAlaSerTyrIleMetProGly	1484 (376)
1485	GAGAAATGGCAACACCACACCACCGGGGAACATCATCGTCTCCAGCTATGAGACTCAGTTGGGTTCAAGTTTACGGCACCGAAACGGAAACC GluAsnGlyAsnThrThrProThrGlyAsnIleIleValSerSerTyrGluThrGlnLeuGlySerValTyrGlyThrGluThrGluThr	1574 (406)
1575	CACCAGACTATGCCACGCAACGAGAGCCCCAACGAGTCCGTCTCCGCCCTCGGGCAACTGCCACCCACACCCAACAGCCTTCCGCG HisGlnThrMetProArgAsnGluSerProAsnGluSerValSerSerAlaPheGlyGlnLeuProProThrProAsnSerLeuSerAla	1664 (436)
1665	GTGGTGTAGTGGAGCTGGTGTGACCTCCTCCAGTGGGGCCAACCTCGGGAGCCGATCCCTCGCAGTCGCTGGCCAAATGCCAGTGTGGAAGT ValValSerGlyAlaGlyValThrSerSerSerGlyAlaAsnSerGlyAlaAspProSerGlnSerLeuAlaAsnAlaSerAlaGlySer	1754 (466)
1755	GAGGAGCTATCGGCTGCCCTGAAAGTGAATCGGTGGACCTGATCGCGCCAGTCAGTCGAGTTGTACGGCGGATGGAGCTCCATGCAG GluGlnLeuSerAlaAlaLeuLysValGluSerValAspLeuIleAlaAlaSerGlnSerGlnLeuTyrGlyGlyTrpSerSerMetGln	1844 (496)
1845	GCACTGCGCCCAATGCGCCACTTTCGCGGAGGACTCGTGAACCTCCACGAGCTCGACCAGCCAGGCTCTGGATGTCACCGCCACCAG AlaLeuArgProAsnAlaProLeuSerProGluAspSerLeuAsnSerThrSerSerThrSerGlnAlaLeuAspValThrAlaHisGln	1934 (526)
1935	ATGTTCCATCCGTATCAGCATACGCCGAGTATGCATCCTATCCGGCACCAGGCCACGCCATTTCGCATCACGGACATCCCATGCGCGG MetPheHisProTyrGlnHisThrProGlnTyrAlaSerTyrProAlaProGlyHisAlaHisSerHisHisGlyHisProHisAlaPro  -	2024 (556)
2025	CATCCGCACGCACATCCGCATCCGAGTACGCAGGCGCACATCCGCACTATCCGCCGCCAGTTCTGTCGGCGCACCTTCATGCCCGAGAAC HisProHisAlaHisProHisProGlnTyrAlaGlyAlaHisProHisTyrProProProSerSerSerAlaHisPheMetProGlnAsn - PRD REPEAT	2114 (586)
2115	TTCAATGCCGCCCTTTCTTCGCCCTCGAAGGTCAACTACACAACGATGCCGCCACAGCCGTTCTATCCCTCTGGTACTAGAATCAA PheAsnAlaAlaAlaPheProSerProSerLysValAsnTyrThrThrMetProProGlnProPheTyrProSerTrpTyrEnd	2204 (613)
2205	AGAGACACGGATCCACCACCTACTCCTCCAGGAGCAGGAGCAGTGTACCAGATCCATGGTACAAGTCGCCAAAGATGTACATACCATA	2294
2295	GAGCAGGGGACGAAAATATAAATAACATTTTATTGTGGTGGAGCAGTACAGACATTTCCGTTTGAGAAAACCGCTGACAGACTCGCTC	2384
2385	CCAAACAATAACATATGTATTAGTTCCAATTCGTAGATGTAAGCTAGAAAATAGTACCAGCTTAGGATTAGATTTAAGATGATTAGC	2474
2475	CTAAGTAGCAAGTGCTCTTAAATAAAAAATATATCTATGCTAATTTACAACGTACTCCAATGATCTTTCAC 2546 -----   (A) <sub>n</sub>	

*prd* SEQUENCE. Accession No. M14548 (DROPRD). An exclamation mark at -244 marks the 5' end of the longest cDNA. Allele *prd*<sup>2,4,5,17</sup> is an insertion of 1.1 kb following position 569, with a concomitant 5-bp deletion of positions 569-573. In cDNA sequences, two natural variants were detected; these involve changes in the amino-acid sequence at codons 164 and 220. A homeodomain spanning Gln-213 to His-272 is delimited by vertical bars and conserved residues are marked with asterisks; within the domain, the three putative helices, H1, H2 and H3, are

(continued)

### Function

Treisman et al. (1989, 1991) have demonstrated direct binding of PRD to element e5 of the *even-skipped* (*eve*) promoter. The homeodomain and the paired domain bind to different sub-regions of e5.

### Mutant Phenotype

*prd* is one of the pair-rule genes. Null mutants are embryonic lethals with only half the correct number of segments. The missing elements correspond mainly to odd-numbered parasegments (Appendix, Fig. A.3); i.e., posterior region of T2 and the adjacent boundary to T3, the posterior of A1 and the adjacent boundary to A2, etc. (every other segment boundary and neighboring areas are missing). The pattern is similar to that affected by *eve*, but the position of the missing elements is shifted anteriorly by a fraction of a parasegment in *prd* as compared to *eve* (Nüsslein-Volhard and Wieschaus 1980; Nüsslein-Volhard et al. 1985). It would appear that, in mutants, the regions of the segmented embryo that are lacking are those in which *prd* is maximally expressed in normal embryos (see below).

### Gene Organization and Expression

Open reading frame, 613 amino acids; expected mRNA length, 2,417+ bases; in agreement with a 2.5 kb band detected by northern analysis); information on the 5' and 3' ends is from a cDNA sequence. There is an intron in the Asp-23 codon (*prd* Sequence) (Frigerio et al. 1986).

### Developmental Pattern

The *prd* transcript is absent from oocytes and barely detectable in 0–2 h embryos; it peaks in 2–4 h embryos and disappears soon afterward. The transcript is first detectable by *in situ* hybridization during nuclear cycle 12 (syncytial blastoderm) in the primordial cephalic region (77–63% egg length; Appendix, Figs A.1–A.3). By nuclear cycle 14 (late syncytial blastoderm), expression is localized in seven bands covering the area from the cephalic region to the eighth abdominal segment (75–20% egg length). These bands are more intense on the dorsal than on the ventral side of the embryo. In general terms, the seven bands of *prd* expression have a two-segment periodicity similar to that of other pair-rule genes such as *eve*, *ftz* and *hairy* (*h*). The *prd* bands,

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(continued) underlined; these were identified based upon their similarity to *Antennapedia* helical regions. The PRD repeat, spanning His-552 to His-572, and PRD domain, spanning Gly-27 to Asp-154, are also delimited by vertical bars.

however, are broader with the area covered by each band corresponding to more than one segment, i.e., they extend posteriorly from the middle of one segment to the posterior boundary of the next segment. The intensity of expression increases posteriorly within each band so that the regions of highest *prd* expression correspond to the posterior compartments of the mandibular, labial, T2, A1, A3, A5 and A7 segments. At this time expression starts in a new domain in the anterior pole of the embryo, at 93–87% egg length, but in the dorsal region only (Kilchherr et al. 1986; Akam 1987; Baumgartner et al. 1987).

Around the time of blastoderm cellularization, an eighth band appears posteriorly (at 13% egg length), and bands 2–7 of the original seven become double because transcripts disappear from the central portion of each band. Thus, in the segmented germ band region, there are 14 stripes; 13 of them are two-cell-wide bands that appear to correspond to the two most posterior cells of each segment in the region from the mandibular segment to the A7 segment. The 14th band is wider and includes A8 and A9. This banded pattern persists until the beginning of gastrulation but disappears soon thereafter (Kilchherr et al. 1986; Baumgartner et al. 1987). In later stages, expression is restricted to the head region and central nervous system (Gutjahr et al. 1993).

## References

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## *Ribosomal protein 49: Rp49*

**Chromosomal Location:**

3R, 99D4-8  
Synonym: *M(3)99D*

**Map Position:**

3-[101]

**Product**

Protein 49 of the large ribosomal subunit (Vaslet et al. 1980; O'Connell and Rosbash 1984). The syntheses of ribosomal proteins are coordinately regulated and at least part of that regulation occurs at the level of translation. For instance, while almost all *Rp49* mRNA is translated during oogenesis, only a small fraction is associated with polysomes early in embryogenesis (Al-Atia et al. 1985).

*Mutant Phenotype*

Heterozygotes for a deletion show a strong *Minute* phenotype (Kongsuwan et al. 1986).

**Gene Organization and Expression**

Open reading frame, 133 amino acids; mRNA length, ca. 520 bases in agreement with a 0.6 kb band from RNA blots. S1 mapping was used to define the 5' and 3' ends; the 3' end is near position 570. There is no apparent TATA box. There is an intron after Ser-31 (*Rp49* Sequence) (O'Connell and Rosbash 1984). *Rp49* is in the *Serendipity* cluster (Chapter 28, Fig. 28.1); it is transcribed convergently with *Sryδ*, the 3' ends of these genes being approximately 300 bp apart.



*Rp49*

-418	ACGACGTCGATGTTAAACCACAGCTTCTCTTCGCTCTCGTITCCGGCAAGGATGTGCCGTGATTTGGGCCACGTTGATGTCATT	-329
-328	ATTTTAAGCCGTAATGTCGTTTTGCGTTTCGAGTTGAAGCTGCGTTAGTCCTCGGGCTAGTGAAGTAGTTAGCAAGTAGTTCGGGCTAGT	-239
-238	ATTTACAGACCATTCTTGATTCTGTGAGCAGTTACTGCCGAATGGCTTCTGTGTTGCTGAATTCGGTATTCGATGTTTCGACATCACGGT	-149
-148	ACTGTCAATGGATACTGCCCAAGCAGCTAGCCCAACCTGGTTGAATTATGCATTAGTGGACACCTGTGTGTTATTAGCTTGATAAGTG	-59
	-->-8	
-58	ATATTTCCAGTGGGTCAGTGCACACTAATGGCTACACTTGTGTGTCCTACCAGCTTCAAGATGACCATCCGCCAGCATACAGGCCAAGA	31
	MetThrIleArgProAlaTyrArgProLysI	(11)
32	TCGTGAAGAAGCGCACCAGGACTTCATCCGCCACCAAGTCGGATCGATATGCTAAGCTGTCGGTGAAGTCCACGGATTGTGCCAAATGT	121
	IeValLysLysArgThrLysAspPheIleArgHisGlnSerAspArgTyrAlaLysLeuSer	(31)
122	ACCCGTGTTTTAATCAACATGTCTCCTTGACAGCACAATGGCCGAAGCCCAAGGGTATCGACAACAGAGTCGGTCGCCGCTTCAAGGGACA	211
	HisLysTrpArgLysProLysGlyIleAspAsnArgValGlyArgArgPheLysGlyGI	(51)
212	GTATCTGATGCCAACATCGGTTACGGATCGAACAGCGCACCCGCCACATGCTGCCACCAGGATTCAGAAGTTCCTGGTGCACAACGT	301
	nTyrLeuMetProAsnIleGlyTyrGlySerAsnLysArgThrArgHisMetLeuProThrGlyPheLysLysPheLeuValHisAsnVa	(81)
302	GCGCGAGCTGGAGGTCCTGCTCATGCAGAACCCGCGTTTACTGCGGAGATGCCACGGCGTCTCCTCCAAGAAGCAAGGAGATTATCGA	391
	IArgGluLeuGluValLeuLeuMetGlnAsnProArgLeuLeuArgGluMetProThrAlaSerProProArgSerLysGluIleIleGI	(111)
392	GCGGCCAAGCAGCTGTGCTCCGCTCACCAACCCCAACGGTCGCCCTGCGTCTCAAGAAGAACGAGTAAGCTTAAGATTCTTGAGAGTT	481
	uArgAlaLysGlnLeuSerLeuArgSerProThrProThrValAlaCysValSerArgArgThrArgEnd	(133)
482	CTGTAACTGGTCGGAATACACATTTGTAACGTTAATATACCGGACTTTTAGTTAAAAATGATGTCCAGTGCCAGTGCCAGTTCATGTC	571
	-----	
572	ATTTCTGAGATCGGGATAGCAGCACCATCGATAACATGTGCATTATCTGGATGGATATCAGTTAATCCAGACCATTGCGGCTTTCTTTC	661
662	TGATAGCAACTGCCTCGAGATATTAGACCAATATAAATCTTGACGTGCCAAAAGTAGACAGCATCAATCCTTATCAGGGAATTTGTGA	751
752	TATATTTACATTTTCCCCCTTAGTATTCAAAGAGGTTGTTTATATGAAATCATATATATTCGCAATTATTTTACAGAACAGTGTA	841

*Rp49* SEQUENCE. Accession X00848 (DRORP49).

## References

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# 27

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## *Salivary Gland Secretion Protein Genes: Sgs3, Sgs5, Sgs7, Sgs8*

### **Chromosomal Location:**

*Sgs3, Sgs7, Sgs8* 3L, 68C3-5  
*Sgs5* 3R, 90B3-8

### **Map Position:**

3-35.0  
3-[60]

### **Products**

Glue proteins in the salivary gland secretion of third-instar larvae.

### *Structure and Function*

Genes for seven proteins have been identified, SGS1 and SGS3-8. Proteins are numbered in order of increasing electrophoretic mobility except for SGS6 which is slightly slower than SGS3 (Velissariou and Ashburner, 1981). Partial sequences for SGS3, SGS7 and SGS8 confirmed the primary structure derived from nucleotide sequences and the existence of a 23-amino-acid signal peptide (Fig. 27.1) (Crowley et al. 1983). These proteins attach larvae to a solid substratum prior to pupariation. Cys residues and glycosylation appear to play a role in the function of SGS.

### *Tissue Distribution*

The glue proteins are synthesized in the larval salivary glands between 106 h and 120 h after fertilization, during the second half of the third instar (Beckendorf and Kafatos 1976; for reviews, see Berendes and Ashburner 1978; Ashburner and Berendes 1978).

### **Evolutionary Relationships**

The evolutionary relationships among the *Sgs* genes are not entirely obvious. It is clear from amino-acid sequence similarities, intron position, sequence of

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1                               50                               100
Sgs5 MFNIKLLLLL LAVSWFHHGQ AVQET.....
Sgs3 MKLIATIALA SILLIGSANV ANCCDCGCPT TTTTCAPRTT QPPCTTTTTT TTTTCAPPTQ QSTTQPPCTT SKPTTPKQTT TQLPCTTPTT TKATTTKPTT
Sgs7 MKLIAVTIIA CILLIGFSDL ALGGA.....
Sgs8 MKLLVAVIA CIMLIGFADP ASGCK.....
CON MKL-----A -I-LIG--- A-----
      ^                |
101                               150                               200
Sgs5 .....K IEEKVSEPE IESEIKNSTS VPSKNCIYYR
Sgs3 TKATTTKATT TKPTTTKQTT TQLPCTTPTT TKQTTTQLPC TTPTTTKPTT TKPTTTKPTT TKPTTTKPTT TKPTTTKPTT TKPTTTKPTT TKPTTTKPTT
Sgs7 .....
Sgs8 .....
CON -----
201                               250                               300
Sgs5 NYQWALQDCV CRCFQNECLM QIESDQRKKE GRSPFVPVTE ELCRSFICK CSVGFVVVAE FPIPAPCGCN RKPFSIATER FVSLCHLLKF SAENSKPFLT YSYCWPWF*
Sgs3 TKPTTTKPTT TKPTTTKPTT TKPTTTKPTT TKPTTTKPTT TKPTTTKPTT TKPTTTKPCG CKSCGPGGEP CNGCAKRDAL CQDLNGVLRN LERKIRQCVC GEPQWLL*
Sgs7 .....CE CQPCGPGGKA CTGCPEKPQL CQQLISDIRN LQQKIRKVCV GEQWMI*
Sgs8 .....DCS CVICGPGGEP CPGCSARVPV CKDLINIMEG LERQVRQCAC GEQWVLF*
CON -----C- C--CGPGG-- C-GC----- C--L----- L---R-C-C GE--W---
      ^                                     ^

```

Fig. 27.1. Comparison of SGS3, SGS7, SGS8 and SGS5 amino-acid sequences. Only positions in which three of the four sequences agree are represented in the CON(sensus). The vertical line at position 23 marks the last residue in the signal peptides of SGS3, SGS7 and SGS8 (Crowley et al. 1983). The caret at position 10 marks the intron in *Sgs3*, *Sgs7* and *Sgs8*, and at positions 234 and 297, the introns in *Sgs5*.

regulatory elements and clustering of the genes, that the three genes at 68C have a common ancestor (Martin and Meyerowitz 1988). On the other hand, *Sgs5* is similar to the other genes in the group only with respect to protein function and possibly some regulatory sequences (Fig. 27.1) (Shore and Guild 1986; Todo et al. 1990). It is likely that at least some SGS proteins are functionally equivalent since natural variants causing a deficiency in *SGS5* (Shore and Guild 1987), *SGS4* or *SGS6* (Velissariou and Ashburner 1981) have no obviously deleterious effect.

## Gene Expression and Developmental Pattern

The *Sgs* genes are expressed in salivary glands during the third larval instar (Meyerowitz and Hogness 1982). Transcription starts approximately 96–98 h after fertilization, reaches a plateau by approximately 112 h, and becomes undetectable by 120 h, the time of pupariation (Hansson and Lambertsson 1983; Georgel et al. 1991). An increase in ecdysterone level is necessary for the start of transcription in the middle third instar (Hansson and Lambertsson 1983). Subsequently, however, in late third instar larvae, high levels of this hormone repress transcription (Crowley and Meyerowitz 1984). Chromosomal puffing accompanies transcriptional activity, but the two processes seem to be somewhat independent of each other (Crowley et al. 1984; Hansson and Lambertsson 1983). There is considerable information on the expression of *Sgs4*; the complete sequence, however, is not available.

### Promoters

The consensus sequence TNTTTGN<sub>x</sub>TCCAT(T/A), in which N<sub>x</sub> represents a variable number of nucleotides (values between 18 and 39 have been observed), was identified as a tissue-specific, *cis*-acting regulatory element of *Sgs3*; such sequences were also found upstream of *Sgs5*, *Sgs7* and *Sgs8* (Todo et al. 1990; Hofmann et al. 1991).

## *Sgs* Gene Cluster at 68C: *Sgs3*, *Sgs7* and *Sgs8*

### Organization and Expression of the Cluster

The three genes are contained in less than 5 kb of DNA. The arrangement of the genes is shown in Fig. 27.2; *Sgs8* is centromere distal (Garfinkel et al. 1983). *Sgs7* and *Sgs8* are separated by 475 bp and they are transcribed divergently. The developmental expression of *Sgs7* and *Sgs8* seems to be controlled by common enhancer elements (Todo et al. 1990; Hofmann et al. 1991). The levels of RNA accumulation are comparable for *Sgs3* and *Sgs7* and an order of magnitude lower for *Sgs8* (Crowley and Meyerowitz 1984).

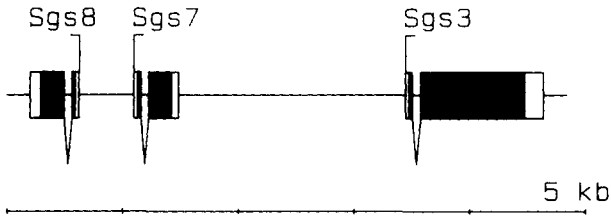


FIG. 27.2. Organization of the 68C cluster.

### *Sgs3*

#### Product

SGS3 is heavily glycosylated (Beckendorf and Kafatos 1976) and very rich in the likely target of glycosylation, Thr residues (45% in the mature peptide). SGS3 is most similar to SGS7 and SGS8 in the amino-terminal 20–25 residues and the carboxy-terminal 50 residues. In particular, the position of eight Cys is conserved among the three sequences. The middle segment of SGS3 is not represented in SGS7 or SGS8; this segment is 235 amino acids long and contains most of the Thr residues: the first 50 amino acids constitute a Thr- and Cys-rich region (residues 23 to 72), and the last 185 amino-acid segment (from 73 to 257) is composed of 37 repeats of the peptide Pro Thr Thr Thr Lys, and variants thereof (Garfinkel et al. 1983). Twenty of the repeats are lacking from a natural variant found in the strain *Formosa* (*Sgs3* Sequence) (Mettling et al. 1985).

#### Gene Organization and Expression

Open reading frame, 307 amino acids; expected mRNA length, 1,117 bases. The strain *Formosa* makes an mRNA that is 300 bases shorter due to an internal deletion (Mettling et al. 1985). Primer extension was used to define the 5' end. The 3' end was obtained from a cDNA sequence. There is an intron in the Ala-10 codon (Garfinkel et al. 1983).

#### Promoter

Two *cis*-acting regulatory elements were identified by *in vitro* mutagenesis and analysis of DNase-hypersensitive sites. Either element is sufficient for correct developmental regulation of transcription, albeit at reduced level; when both elements are present, the transcription level increases 20-fold (Martin et al. 1989a, 1989b; Meyerowitz et al. 1987; Roark et al. 1990). Mutational analysis of the proximal element established that it is bipartite: the critical sequences being TGTTTG (pa, at –120, in *Sgs3* Sequence) and TCCATT (pb at –96). Sequences related to these two hexanucleotides are also found in the promoter

*Sgs3*

-782	TCGTTGAATCAATGTCAAATTCGCTGTCAAAGTGCAAACGAAGCCCAAAATGTCTATCTCAATTCGAACCTAAAAATATATATTTTTTGA	-693	
-692	ATATGCAATACTATAAGATAAATGAATAGTTTTATGGGGCTTATTTGTAAAGCTAAATTAAGCTAAATTTAACTGTCCCTATTATATATTA	-603	
	----->da	----->db	
-602	TTATATTTACTCAGCCTATATTAAGACCTATTATTTATAGAATTTAACGCAGTTTGTCTGCAAAAACATCTCTACACCTTTTTCTACCCG	-513	
-512	TTACTCGTAGAGTAAAGGGGTACTCGTTCGCTGAGAAGTAAACAGGCAGAATATAAAGCATATATATTCTTGATTAGGGTCAATAGCC	-423	
-422	GAGTCGATCTGGCCATGTCCGCTGATTCTGTGTTGCCACTCCACATTTTTGAAAAATGTTTTATAATTTTTTCATATTTTTATTATCTA	-333	
-332	AATCTATCCCTCCACACCTTAGAGCATTAAATTTAATTTCTTTCCCCAATTTTACCAGATATTCGTGAAAAATGTTATACATTTTCCA	-243	
-242	TTTCACTGAACTAGTAAAGTAAACGGGTACTGTGTAGTCTGTTAGCGTTCTCTCTGTGTTTTAAAATAAAGCTAGGCGATCGAGTCGAC	-153	
-152	CCAAAAGTATCAAAACAAAGGGGAGAAGGCTTGTGTTTGCATAATCGAAACTGACTCCATTTTTAGAATTGCAGTTTCAGTGAAAGCGT	-63	
	----->pa	----->pb	----
	--> -28		
-62	ACCTATAAAAAGGTGAGGTATCCGCAAGAAAAGTATCAGTTTGTGGAGAATTAAGTAAAAACATGAAGCTGACCATTGCTACCCGCTA	27	
	-----	MetLeuLeuThrIleAlaThrAlaLeu	(9)
28	GGTAGGTTTCACCGAATGCTCTTGTTTCGGTATTTGAGCCACTGATATATTCATCCGTTTGCCITTCACACGCGAGCATCCTGCTTAT	117	
	A	IaSerIleLeuLeuI	(15)
118	TGGCTCCGCTAATGTTGCCAACTGTTGCGATTGTGGATGCCCCACAACACTACAACACTACTGTGCGCCACGTACCACGCAACCTCCGTGCAC	207	
	eGlySerAlaAsnValAlaAsnCysCysAspCysGlyCysProThrThrThrThrThrCysAlaProArgThrThrGlnProProCysTh	(45)	
208	AACTACGACAACAACAACCAACTACTTGTGCGCCACCCACACAACATCTACCACGCAACCTCCATGCAGCATCTAAGCCACCAC	297	
	rThrThrThrThrThrThrThrThrThrCysAlaProProThrGlnGlnSerThrThrGlnProProCysThrThrSerLysProThrTh	(75)	
	-		
298	ACCTAAGCAAACCTACCACGCAACTCCGTCACAACACCCACCACCCTAAGGCCACCACCACGAAGCCACCACCCTAAGCCACCAC	387	
	rProLysGlnThrThrThrGlnLeuProCysThrThrProThrThrThrLysAlaThrThrThrLysProThrThrThrLysAlaThrTh	(105)	
388	CACTAAGGCCACCACCCTAAGCCACCACCCTAAGCAAACCTACCACGCAACTCCGTCACAACACCCACCACCCTAAGCAAACCTAC	477	
	rThrLysAlaThrThrThrLysProThrThrThrLysGlnThrThrThrGlnLeuProCysThrThrProThrThrThrLysGlnThrTh	(135)	
	-  Deleted in <u>Formosa</u>		
478	CACGCAACTCCGTCACAACACCCACCACCCTAAGCCACCACCACGAAGCCACCACCACGAAGCCACCACCCTAAGCCACCAC	567	
	rThrGlnLeuProCysThrThrProThrThrThrLysProThrThrThrLysProThrThrThrLysProThrThrThrLysProThrTh	(165)	
	-		
568	CACGAAGCCACCACCACCAAGCCACCACCACGAAGCCACCACCCTAAGCCACCACCACGAAGCCACCACCCTAAGCCACCAC	657	
	rThrLysProThrThrThrLysProThrThrThrLysProThrThrThrLysProThrThrThrLysProThrThrThrLysProThrTh	(195)	
658	CACGAAGCCACCACCACGAAGCCACCACCCTAAGCCACCACCACGAAGCCACCACCCTAAGCCACCACCACGAAGCCACCAC	747	
	rThrLysProThrThrThrLysProThrThrThrLysProThrThrThrLysProThrThrThrLysProThrThrThrLysProThrTh	(225)	
	-  Deleted in <u>Formosa</u>		
748	CACTAAGCCACCACCACGAAGCCACCACCCTAAGCCACCACCACGAAGCCACCACCACGAAGCCACCACCCTAAGCCACCAC	837	
	rThrLysProThrThrThrLysProThrThrThrLysProThrThrThrLysProThrThrThrLysProThrThrThrLysProThrTh	(255)	

(continued)

838	ACCTAAGCCGTGCGGTTGCAAGAGCTGCGGTCTGGAGGAGAGCCATGCAATGGATGTGCTAAGAGGGATGCACGTGCCAGGATCTTAA rProLysProCysGlyCysLysSerCysGlyProGlyGlyGluProCysAsnGlyCysAlaLysArgAspAlaLeuCysGlnAspLeuAs	927 (285)
928	CGGCGTACTCCGCAATCTGGAGCGCAAGATCCGTCAATCGCTCTGCGGTGAACCGCAATGGTTGCTGTGAAGCGTCAAGGAGCGTCTAA nGlyValLeuArgAsnLeuGluArgLysIleArgGlnCysValCysGlyGluProGlnTrpLeuLeuEnd	1017 (307)
1018	TCCACTCCCGTACTGATCGATGTGACTGCACCCCTGCGAAATATATTCTGTGGGGGAGCTCGGCCAGGACTTTGACTACGCTTTGTTTTT	1107
1108	GTTATCATCAATTGATTTTACGTGTAAGAATAATAAAAATTAGTTAGACTGCATAAAATTTTAAAGCATTATTATTATTTTACTTGTAT -----   (A) <sub>n</sub>	1197
1198	TATTTATGACAAATTATTATTTATCTGTTGGGTTTTCGAAATGTTGGTCTAAATTAAGTTTGGCCATCTTTGATCGACTTTTTTCGAA	1287
1288	TGTATCTGTTACTTTTACCAATGCGTTGGCTTTGGCTCCTAGTTCTATGCGAAGTCTTAACTATCCGAGCTCTTATGACTTGGTCAACT	1377
1378	GTCTCAGCTAACTACTGTTGG	1398

*Sgs3* SEQUENCE. Strain, *Oregon R*. Accession, X01918 (DROSGS378). Arrows labeled da, db, pa and pb underline the a and b parts of the distal and proximal promoter elements. The *Formosa* strain deletions that occur in the repetitive middle portion of the coding region are delimited by vertical bars (Mettling et al. 1985).

region of *Sgs7* and *Sgs8* and within the distal element of *Sgs3*, at -651 (da) and -617 (db) (Todo et al. 1990). A DNase-hypersensitive site near -630 occurs only in the chromatin of salivary glands of third instar larvae, and DNase protection experiments identified two footprints overlapping da and db. There are three other hypersensitive sites near the 5' end of *Sgs3*, including one around -100; but these are not restricted to the tissue in which *Sgs3* is expressed (Georgel et al. 1991).

A 115-kD protein that binds specifically to the distal promoter element, the Glue Enhancer-Binding Factor, GEBF1, was isolated from nuclear extracts. It appears that GEBF1 binds to both parts of the distal promoter element (da and db). The amount of GEBF1 found in extracts rises in parallel with the transcriptional activity of the salivary gland secretion genes during the third instar. GEBF1 is absent from extracts of the *Broad Complex* mutant *l(1)t<sup>435</sup>* (located in region 2B5, the site of an early, ecdysone-irreducible, puff) (Georgel et al. 1991). This allele also reduces or eliminates expression of the glue genes (Crowley et al. 1984). These observations suggest that (a) a gene in 2B5 is, directly or indirectly, responsible for the synthesis of GEBF1; and that (b) ecdysone induces the glue genes indirectly, by inducing the appearance of a regulatory factor (Georgel et al. 1991).

## *Sgs7*

### Product

SGS7 is not glycosylated (Beckendorf and Kafatos 1976); it contains only 4% Ser/Thr (Shore and Guild 1986).





## Gene Organization and Expression

Open reading frame, 74 amino acids; expected mRNA length, 319 bases. S1 mapping was used to define the 5' end. The 3' end was obtained from a cDNA sequence. There is an intron in the Ala-10 codon (*Sgs7* Sequence) (Garfinkel et al. 1983).

### Promoter

A region between -243 and -75 is necessary for transcription of *Sgs7* and *Sgs8*. Within this region, the segment between -165 and -80 enhances transcription from the *Sgs3* promoter in promoter fusion experiments. This 85-bp segment contains two copies of the bipartite regulatory element defined experimentally for *Sgs3*: 1a, 1b, 2a and 2b (*Sgs7* Sequence) (Todo et al. 1990; Hofmann et al. 1991).

## *Sgs8*

### Product

SGS8 is not glycosylated (Beckendorf and Kafatos 1976); it contains only 4% Ser/Thr (Shore and Guild 1986).

## Gene Organization and Expression

Open reading frame, 75 amino acids; expected mRNA length, 353 bases. S1 mapping was used to define the 5' end. The 3' end was obtained from a cDNA sequence. There is an intron at Ala-10 (*Sgs8* Sequence) (Garfinkel et al. 1983).

### Promoter

The putative regulatory elements, between positions -452 and -370, are the same as those described above for *Sgs7* (Todo et al. 1990).

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(continued) -507 and the TATA box at -478 is double underlined. A base substitution (at 245) found in the strain *Formosa* is indicated (Mettling et al. 1985).

*Sgs8*

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-270 ATTTTATGAGTAATACTTCTTTCTTAGTAAAAAGGGCATGTAATAATGTATATTATATTAGTATGAACGATGACACAGAAAATGGTGAGAAC -181
-180 AAAGGGCGAAGTTTAATATCGTGTTCTATTGTGCTTAAGCACAAATGGTTATATATTTTATTTTTGGATGAATTCATGAAATTTCCATT -91
                                     ---
-90 GATCCTATAACACATTTGTTTTGCTATAAAAGGGGTAGGGTGTTCATAACGGTTACCATCTGGTAAAGTTAATTTGTTAAAGCAACAAC -1
  =                                     -----
  0 CATGAAGCTGCTCGTTGTGCGCGTCATTGGTAAGTGCCAAAAAGTACTATTTTTTATGTACCCAAATCCACTTAGCCATCCGTTTCATT 89
    MetLysLeuLeuValValAlaValIleA (10)
  90 TGACCCAGCGTGCATCATGCTCATCGGATTCGCGCATCCTGCCTCGGGCTGCAAGGATTGTTTCATGCGTGATTTGTGGACCTGGTGCCGA 179
    l aCysIleMetLeuIleGlyPheAlaAspProAlaSerGlyCysLysAspCysSerCysValIleCysGlyProGlyGlyGly (37)
  180 GCCGTGCTCGGGTGTCCGACAGGGTTCCTGCTGCAAAAGATCGATCAACATTATGGAGGGTCTTGAGCGGCAAGGTGCGTCAGTGCCG 269
    uProCysProGlyCysSerAlaArgValProValCysLysAspLeuIleAsnIleMetGluGlyLeuGluArgGlnValArgGlnCysAl (67)
  270 CTGCGGAGAGCAGGTTTGGCTGTCTAGAGATGTGCCCTCAACCTAATCGGCACTGACCTTTTATCTGCTGGCATTAAAACTGCTGTCT 359
    aCysGlyGluGlnValTrpLeuPheEnd (75)
  360 AATAAACTATTATCATTCTCGACGACCCAAACTCCTTTTCTTTGTTTTTAATTATTTATTTTCAGATGTATTGCTTAAAAAGTGTCA 449
                                     |(A)n
  450 GAACTAGTCTTTTCTGTGTGCACAATCACGATGGTCTTGTTGGCACCTCTTTGGATTCTTGCACTTCCGCTTGGGAATGCGGGTGTGGCA 539
  540 CCAATTGTTGCCCTCCAACAAAGCTTTTTGTGAGTGGAAAGCGGCGTGTGTTGTTGTTGTTGATGCTGCAGTAGTGGTTGTTGTTGT 629
  630 GGTGGTGGCATCCGTGGTTGTTGTTGTTACTAGCAGATGACGCAACCTGAATGGCCAGGGCAATAAGGGCAACCACGAAAAGATACTTCAT 719
  720 TTTGAAATAAGATTAGATTTTTTCGATACGACTGGAATTGAACGATCAGGTGTTGTGATTAATTTAAATCATACCCACTGCTTTTATAGCA 809
  810 AAACAAGCAGATTTCCGCACTCGCTTACTATGTTTTGCTTCCATAACGCATAAGCACATAAAAAGCGAGTACAATAGCAAAAAGCATT 899
  900 TAATAATCAAATGTTTGAACAGTAAGCAAAGACGGTTTTGTTGACATATTTGTAATATCAACAATTAATGGGTACTATTCTAAAAAA 989
  990 ATTCCCTAAAAAGTATGCAATAATGTTTACCCAGCAGCATTTGATTTCAATGTCAAACACTGCAACAGAAATAAAAAATATTTCAAAT 1079
1080 ATTCTAGAAGCTTTTGAAGAATATTACCCAGAAGAAAAAACACATTAAATTTGTTACATTT 1143

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*Sgs8* SEQUENCE. Strain, *Oregon R*. Accession, X01918 (DROSGS378).

*Sgs5***Product**

SGS5 is lightly glycosylated (Beckendorf and Kafatos 1976; it contains 12% Ser/Thr distributed throughout the sequence; Cys (6.75%) is also distributed without any apparent pattern (Shore and Guild 1986).

*Sgs5*

-205	AAGCTTTTTTTTGGAGTGGAAATTTATGGCTGTGTTTTTTTGCCAGTCAAGGTTGTTTGCCTGTCTGCAAACTTTTACTTTTCTAG	-116	
	<span style="margin-left: 100px;">A</span> <span style="margin-left: 200px;">A</span> <span style="margin-left: 300px;">G</span> <span style="margin-left: 400px;">A</span>		
-115	ATGCACTAAGTCAATAAAGCGCTTGGCCAACTGCTAAAACAGTGGAGGTATTCAATATAAATAGCCAAATGAGATATTATGGGGACA	-26	
	<span style="margin-left: 50px;">&gt;</span> <span style="margin-left: 100px;">A</span> <span style="margin-left: 150px;">G</span> <span style="margin-left: 200px;">A</span>		
-25	GTTATATTCTTAGCCACTTTTACGACATGTTCAATATTAATTTGCTGCTTTTGTATTGGCCGTTTCGTGGTTCACCATGGACAAGCCG	64	
	MetPheAsnIleLysLeuLeuLeuLeuLeuLeuLeuAlaValSerTrpPheHishHisGlyGlnAlaV	(22)	
	Gln		
65	TCCAGGAGACGAAATCGAAGAAAACCAGTATCAGAGCCTGAAATGAATCCGAAATAAAGAACTCTACGAGCGTCCCAAGTAAATGCA	154	
	alGlnGluThrLysIleGluGluLysProValSerGluProGluIleGluSerGluIleLysAsnSerThrSerValProSerLysCysA	(52)	
	Val	Ser	
155	ATATTACTATAGAACTACCAATGGGCTCTTCAGGATTGTGTCTGCCGTTGTTTCCAAAACGAATGCCTTATGCAAAATCGAGAGCGACC	244	
	snIleTyrTyrArgAsnTyrGlnTrpAlaLeuGlnAspCysValCysArgCysPheGlnAsnGluCysLeuMetGlnIleGluSerAspG	(82)	
245	AGCGCAAAAAGGAGGTAGATCCCGTAAAGTAAATTAACCAGTTAAGCAAAATGATTTTATTATAACTTGTAATACAGCATTGTGCC	334	
	lnArgLysLysGluGlyArgSerP	roPheValPr	(93)
335	CGTTACGGAGAACTCTGCCGTTCCCTTCATCTGCAAAAAGTGCAGCGTGGGTTTCCCGTGGTTGCTGAATTCGCCATTCGGGCTCCCTG	424	
	oValThrGluGluLeuCysArgSerPheIleCysLysLysCysSerValGlyPheProValAlaIleGluPheProIleProAlaProCy	(123)	
425	TGGATGCAATCGAAAGCCAGGATCAATGGCCACAGAGATTCTACAGTTTGTGCCACCTGCTGAAATCTCAGCGGAGAACGACGAAGCG	514	
	sGlyCysAsnArgLysProGlySerIleAlaThrGluArgPheTyrSerLeuCysHisLeuLeuLysPheSerAlaGluAsnSerLysP	(153)	
515	TAAGTCCAAGAATTGGTTCCAAATATATCGGTAATATACATTTTGTATCTTACAGCATTCCTGACTATTCTATTGTTGCCCTTC	604	
	roPheLeuThrTyrSerTyrCysTrpProPhe	(163)	
605	TAAGTGGAGTGGATTTCAGTTGGATCACGTTACTAATATCTTTGTTGTTTATTATTTTGGTATTGTTTCATTTAAAGGGAGATG	694	
	End		
695	GATTACAAATAAAGAAATATATTCAATGACGAGTGCATAAAATTTTTTGAATATGAAAATCTTTTATAGACTAAACAGCTATGCAT	784	
	<span style="margin-left: 100px;">&gt;</span> <span style="margin-left: 250px;">(A)<sub>n</sub></span>		
785	ATGTTTAAACATTGAAAAGCTT	806	

*Sgs5* SEQUENCE. Strain, *Oregon R*. Accession, X04269 (*DROSGS5*). The sequences with similarity to the *Sgs3* regulatory element are underlined by the arrows at -150 and -120. The natural variant *Sgs5*<sup>n1</sup>, found in strain *CA-2* fails to express this gene. The base substitutions that distinguish *CA-2* from *Oregon R* are shown above the *Oregon R* sequence (Shore and Guild 1987).

### Gene Organization and Expression

Open reading frame, 163 amino acids; expected mRNA length, 646–653 bases. The average polyadenylation tail is 100–150 bases long. Transcription appears to initiate with equal frequency at the first A or at any of the five Gs between -33 and -25. Nuclease protection was used to define the 5' and 3' ends. There are introns in the Pro-90 and Pro-153 codons (*Sgs5* Sequence) (Shore and Guild 1986).

## Promoter

A DNA fragment that extends from -205 to 806 is capable of autonomous expression in a somatic transformation assay. A segment that extends from -151 to -93 contains *cis*-acting sequences necessary for expression (Shore and Guild 1987). The shorter interval includes sequences that resemble the bipartite regulatory elements of *Sgs3* (Todo et al. 1990).

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# 28

## The *Serendipity* Gene Cluster: *Sry* $\alpha$ , *Sry* $\beta$ , *Sry* $\delta$

**Chromosomal Location:**  
3R, 99D4-8

**Map Position:**  
3-[101]

### Organization of the Cluster

*Sry* $\alpha$ , *Sry* $\beta$  and *Sry* $\delta$  are grouped in a dense cluster within an 8 kb segment that also includes *janA*, *janB* and the ribosomal protein gene *rp49* (Fig. 28.1). The three *Sry* genes are transcribed in the same direction; the distance between the poly-A signal in one gene and the TATA box of the next is a few hundred bp. In addition to the gene-specific transcripts, two other longer poly-A RNAs are detectable. These include sequences from neighboring genes: either  $\beta$  plus  $\alpha$  or  $\alpha$  plus  $\delta$  are combined. These longer RNAs are thought to be the consequences of transcription starting normally in one gene but then proceeding to “read-through” to the end of the next gene downstream (Vincent et al. 1984, 1985).

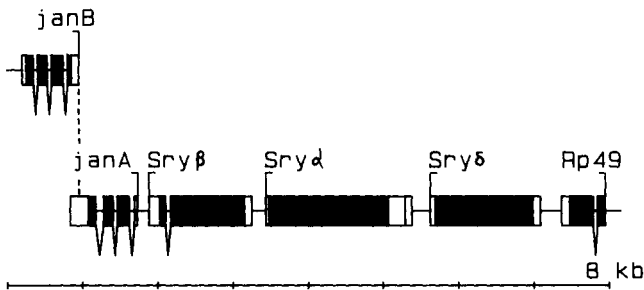


FIGURE 28.1. Organization of the *Sry* cluster. For the sake of clarity, *janB* was drawn on a separate line; it actually overlaps *janA*

## *Sryα*

### **Product**

A 58 kD protein without resemblance to other known proteins (Vincent et al. 1985).

### *Tissue Distribution*

It is present very briefly in embryos undergoing blastoderm cellularization. The *Sryα* protein (SR $Y\alpha$ ) accumulates sharply during nuclear cycle 14 and disappears as gastrulation proceeds. As the embryonic syncytium becomes partitioned into individual cells, SR $Y\alpha$  is concentrated at the leading edges of the invaginations of the plasma membrane; this intracellular distribution is very similar to that of actin filaments (Schweisguth et al. 1990).

### *Mutant Phenotypes*

In embryos homozygous for a deletion of *Sryα*, the process of cellularization is severely disrupted, surface invaginations of the plasma membrane are very irregularly distributed, and they often encompass multiple nuclei. Such *Sryα* mutation is an embryonic lethal (Schweisguth et al. 1990).

### **Gene Organization and Expression**

Open reading frame, 530 amino acids; expected mRNA length, 1,862 or 1,952 bases depending on whether the major or minor polyadenylation site is used. The 5' end was determined by S1 mapping and primer extension; the 3' end was defined by S1 mapping. There are no introns (*Sry* Sequences) (Vincent et al. 1985).

### *Developmental Pattern*

Expression of *Sryα* is restricted to 2–4 h embryos (Vincent et al. 1985). In accordance with the pattern of protein synthesis, *Sryα* mRNA is first detectable in syncytial blastoderm embryos, during nuclear division cycle 11; it peaks in cycle 14 and disappears soon afterwards (Schweisguth et al. 1989).

### *Promoter*

P elements that included 5' sequences from *Sryα* and  $\beta$ -galactosidase as a reporter gene, were used to define regions important for transcription. A 248-bp segment that extends from 118 bp upstream of the transcription initiation site to 130 bp downstream of the transcription initiation site is sufficient for specific

Sryβ

-580 GGATCCGACTTACCATGCCATGTGCAGTCCGCAAAATCCGCGGATCACCGCCCTTTGAAGCATCTCTCCATCGAAGACATTGATCATGACA -491

-490 TACTTGAAGATGCCCTCTGGACTGATGTGCACCAGTGGCACGCCGCAAGTGTCTCTCGGACATTTTGTGAATCAGTCGTAGTCTCTTG -401

-400 GAAAGCAGTTGGAGGCGATTTCATTGTGAAAAAGTGTGAAAAACAGAATTTAATGTCTTACCAACCGCAAAATTTCCAAAAACGCTGCT -311

. <-- janA. . . . .

-310 TTAGTCCAGCAATGTGACCAGATACTTTTATTCTCGTTCACCTTATGTATCGATACTATTAAAAATATAATATTCAGTTATTTCTTTAGT -221

. . . . . ----- . . . . . -->-144

-220 ACATTTGGGAAAACGTTTTTTGTACTACTCAGTATATTTTAGTAATAATTAATAAACTCAAAAAGAAAACGTTGTATCAGTGACAGCAA -131

-----

-130 ATTTATCGATTACGCCGACAGCACGAGTCGATACTATCGGCACATATCGGCACAGCTCTGGCGTTCACAACAAAATACACAAAATAAA -41

-40 TTGTTAAGCCGAATTTTCGATTGGATTCCACGGCAGCTAGATGAGCTCCACGCGTCCGTTTTGCTTCGTTTGC6GCAAGGAGAAGTCCG 49  
MetSerSerThrArgProPheCysPheValCysGlyLysGluLysSerV (17)

50 TGGGGGTGTCCAGCTGATAGAAGTAACTGCTTACGCCGACTCGAAAGTCTGATAGCCGACTTTTTACAGGCTGCATTGTGCC 139  
aIGlyValPheGlnLeuIleGluG lyCysIleValPr (29)

140 AGGAACCTTTAAGCCCATCAAGGATATACTGAAATCTCGAGAAGATCATAAAACAGCGGCTGGAGCTCCTGCCAACCTCGCCGCGCTG 229  
oGlyThrPheLysProIleLysAspIleLeuLysTyrPheGluLysIleIleAsnGlnArgLeuGluLeuLeuProAsnSerAlaAlaCy (59)

230 CCGGGACTGCCGAGTACCTCTTCAACTACGACAGGCTGGTGGAGAAATCTCAGCCAAGTGCAGCGCCAGATTGCGGACGACTGCTCGG 319  
sArgAspCysLeuGluTyrLeuPheAsnTyrAspArgLeuValArgAsnLeuSerGlnValGlnArgGlnIleAlaAspAlaLeuLeuGln (89)

320 CTGCAAGCAGGTGGAGGGCAAGGCGGAGACCAAGCAACAGGCGGCAAGAGGGCCCGCTCCAGGTGCCGGCCTTCAAGATCGTCCAGGC 409  
yCysArgGlnValGluGlyLysAlaGluThrLysGlnGlnAlaAlaLysArgAlaArgValGlnValProAlaPheLysIleValGlnAl (119)

410 CACCGCCCTCAAGGAGCCGAAAGGCAGCCGGGCGAGGAGGATGAGTGCAGGAATTCATGAAGGAGGAGATGCTGGACGAGGAGTTCCA 499  
aThrAlaLeuLysGluProGluArgGlnProGlyGluGluAspGluCysGluGluPheMetLysGluGluMetLeuAspGluGluPheGln (149)

500 GTTCAGCGAGCCGGACGACAGCATGCCCTGTCGGAGGAGGATTTCTACCAGAGACCACCGAGATACCTGCCATATCTGCGGCGAGAT 589  
nPheSerGluProAspAspSerMetProSerSerGluGluGluPhePheThrGluThrThrGluIleProCysHisIleCysGlyGluMe (179)

. . . . . --- ---

590 GTTTTCCAGCCAGGAGGTGCTCGAGCGGCACATCAAGGCGGACACCTGCCAGAAGAGCGAGCAGGCCACCTGCAACGTGTGTGGCTTGAA 679  
tPheSerSerGlnGluValLeuGluArgHisIleLysAlaAspThrCysGlnLysSerGluGlnAlaThrCysAsnValCysGlyLeuLy (209)

. . . . . --- ---

680 AGTGAAGGACGACGAGGTACTCGATCTGCATATGAACCTTGACAGGAGGCAAAACAGAACTTGAATGCCGCTACTGCGACAAAAGTTCTC 769  
sValLysAspAspGluValLeuAspLeuHisMetAsnLeuHisGluGlyLysThrGluLeuGluCysArgTyrCysAspLysLysPheSe (239)

. . . . . --- ---

770 GCACAAGCGGAACGTCCTGCGCCACATGGAGGTGCACTGGGACAAGAAGAAGTACCAGTGCAGCAAGTGC6GCGAACGCTTCTCGCTCTC 859  
rHisLysArgAsnValLeuArgHisMetGluValHisTrpAspLysLysLysTyrGlnCysAspLysCysGlyGluArgPheSerLeuSe (269)

. . . . . --- ---

860 CTGGCTGATGTACAACCTCTGATGCACCAGCACGCCAGGAGAACGCCCTGATCTGCGAGGTGTGCCACCAGCAGTTCAAGACCAAGCG 949  
rTrpLeuMetTyrAsnHisLeuMetArgHisAspAlaGluGluAsnAlaLeuIleCysGluValCysHisGlnGlnPheLysThrLysAr (299)

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(continued)



950	CACCTACAAGCACCACCTTGGCGACCCACCAGACGGACCGCGCTACCCTGCCCCGACTGCGAGAAATCGTTCGTGGACAAGTACAC gThrTyrLysHisHisLeuArgThrHisGlnThrAspArgProArgTyrProCysProAspCysGluLysSerPheValAspLysTyrTh	1039 (329)
	--- --	
1040	CCTGAAGGTGCACAAGCGGGTCCACCAGCCGGTCGAGAAGCCAGAGTCGCGGAGGCCAAGGAAGCCACCGTTCACGTTCTTTAGGGTAG rLeuLysValHisLysArgValHisGlnProValGluLysProGluSerAlaGluAlaLysGluAlaThrValThrPhePheEnd	1129 (356)
	---	
1130	TCCTTTGCTAGATTAATCTAAGAAGCCAGCTCATGGTGCCATTAGCGCGGTATGTATCATAAAATTAATCTAAACAATTATTGACGG	1219
	-----	
1220	AAAGTACCAGTTCCTTGGCGTCTTCGCCCAATTTCCGAAACCCAGTAGGTAAGTAGCGGATTTCCGCAATTTTCGCGGGTATGGCA  (A) <sub>n</sub>	1309
	<i>Srya</i>	
1310	ATAAAACAGGCGAGATGTTTTTAATCCCCAAAATAGGTCCTTCTACCTGTGCGCTTGGCAAAGTATATAAAGGTGTTGCGTCGTCGGCC	1399
	--->1407 ----- 1450 -----	
1400	AGAAGTACCTAGTTGAAACATTTCTGTTTCCCGAGCACATCTGATAGAACAGCATGGAACAGCTATTGGCCCAATTACACACTTGCAGTGAGC MetGluGlnLeuLeuAlaGlnLeuHisThrCysSerGluL	1489 (14)
1490	TGATTGCGAGGGCTACAGCAGCACCGCCAACATTGGCTGGCTGAACGAGTTCGCGCCACTTTCCTGGACTTCGCGAGCATCTGAAGG euIleAlaGluGlyTyrSerSerThrGlyAsnIleGlyTrpLeuAsnGluPheCysAlaThrPheLeuAspPheAlaSerAspLeuLysA	1579 (44)
1580	CTAGGCTGCCGGAGGTGGCGCCAGTGGCGCAAACCTTGATGTGGAGACCATCTCCTGTCCTCACCCAGGTGGTAACTGCATCACCC laArgLeuProGluValAlaProSerGlyAlaAsnLeuAspValGluThrIlePheLeuCysLeuThrGlnValIleThrCysIleThrH	1669 (74)
1670	ACCTAGAGCGGACCATCAGCATGGAGGCACCGCATATGACCAGGCAGCAGTTCCTCGACCCTGGACTGGTGTCTTCGCGGACTGCTCG isLeuGluArgThrIleSerMetGluAlaProHisMetThrArgGlnHisPheLeuAspArgLeuAspTrpCysLeuArgArgLeuLeuV	1759 (104)
1760	TCTCCTTGACGCAACTGGAAGGCAACGTGACCCAGTCAAGAACCAGGAGTACCTCCTCGTTGAGCTCATGGACCTGGCTCTGGACC alSerLeuThrGlnLeuGluGlyAsnValThrProValLysAsnLeuGluAspHisSerPheValGluLeuMetAspLeuAlaLeuAspH	1849 (134)
1850	ACTTGGATGACTACATGGAGAAGCTGGCCAGCAGAGAACAACCTCCTGCACATCTTAGAAGAGAGCTTCACGGAAGACACCTACCAGC isLeuAspAspTyrMetGluLysLeuAlaGlnGlnArgAsnAsnSerLeuHisIleLeuGluGluSerPheThrGluAspThrTyrGlnL	1939 (164)
1940	TGGCCAGCATAGTTAATCACATCGTTTCGCCAGCCCTGGCCCTTTCCTCAATGTGCCATTTCATTCGACAAGAAGGCTTTGACGGCTTTGT euAlaSerIleValAsnHisIleValArgHisAlaLeuAlaPheAlaAsnValAlaIleHisSerAspLysLysAlaLeuThrAlaLeuC	2029 (194)
2030	GCGAGACCTTGTCTGCCAAGTGTGCCACTTCCACGAGGAGGCGGGCGAGCCCAACAGTGGTTCATCGAAAGCTGGAGGCCCTCTCCCTGG ysGluThrLeuLeuAlaGluCysAlaThrPheHisGluGluAlaGlyGluProAsnSerGlyHisArgLysLeuGluAlaLeuSerLeuG	2119 (224)
2120	AACGTGCCCTCTATGCCGTGAATCCCTTCTCAATGAGCGCTGCTGCACCTTGTCTGTCAGTCTGATAGATCTGGAAAACGCTTCGG luArgAlaLeuTyrAlaLeuGluSerPheLeuAsnGluAlaLeuLeuHisLeuLeuPheValSerLeuIleAspLeuGluAsnAlaSerV	2209 (254)
2210	TGGAGAAGCTAAAGGATGCACTGCAAAAGGATCCTGCGGGAGCTCAGGAGCTAATCTCCGCAATTCGACACGAAACATGGATCGCATTACGC alGluLysLeuLysAspAlaLeuGlnArgAspProAlaGlyAlaGlnGluLeuIleSerAlaPheAspThrAsnMetAspArgIleGlnG	2299 (284)
2300	AGATTGGGTTCTGGCCATAGCCTTCTCGCAGGACATCAAACGAAGAGCATGTCAGGAGCTGCCTGGCCTCCTGGAATCCCTGGATG lnIleGlyValLeuAlaIleAlaPheSerGlnAspIleLysThrLysThrIleValArgSerCysLeuAlaSerLeuGluSerLeuAspA	2389 (314)
2390	CGTGCATTGTGCCGCTCTCCAGCTGCCAGAGTCCACTTCATCCGACACCACCGGGAGGTCTTCAGGAGCATTTAACCAGGAGCTGC laCysIleValProAlaLeuGlnLeuProGluSerThrSerSerAlaHisHisAlaGluValLeuGlnGluHisPheAsnGlnGluLeuL	2479 (344)

2480	TGATCTTTAGGAACGTCATCCACGAAATCATCGATAGCTGCTCCCTGATCAACAAC TACCTGGACATGCTGGGCGAGAGGATCCACGTAC euIlePheArgAsnValIleHisGluIleIleAspSerCysSerLeuIleAsnAsnTyrLeuAspMetLeuGlyGluArgIleHisValG	2569 (374)
2570	AGGACAAAAGCCATCTGAAGCTGATGTCAGAGGGGGCGAGTGGTGGTGGATCATCTTCGGCTGCCCGTCAATTACTCGGGACTCAGTG lnAspLysSerHisLeuLysLeuIleValGlnArgGlyGlyValValValAspHisPheArgLeuProValAsnTyrSerGlyLeuSerG	2659 (404)
2660	AAGATGGCAAGCGGGTGCACAAGGACCTCATTCTGATCTGCGCGAGTGCCAGGCCGCTGGTCAACCTGGACGTCCAGTGGATCCCAAGC luAspGlyLysArgValHisLysAspLeuIleLeuIleLeuArgGluCysGlnAlaValValAsnLeuAspValProValAspProLysA	2749 (434)
2750	GCATCGTGAAGCGCCTTAAGATACTGTACTCCGTGCTGGCCAAGCTGAGGGACTTGATATGCAGGGATAATCTGGAGCCCGATTCTCTCAG rgIleValLysArgLeuLysIleLeuTyrSerValLeuAlaLysLeuArgAspLeuIleCysArgAspAsnLeuGluProAspSerSerV	2839 (464)
2840	TTGCTTCGGAAGCTCAAGTGCCCTCAAGTGCAACCCGAACCTTTGTGCGGAGCAGTCGATCCTTTGGCAAACGGCATCGATCCTTTGTAA alAlaSerGluAlaGlnValProSerSerAlaThrArgThrPheValArgSerSerArgSerPheGlyLysArgHisArgSerPheValL	2929 (494)
2930	AACAAACCAGAAATTGCTCAGTTTTGCGGCCACAGGACTCACTTGCTGAATCCGGACACAGCGAAAGCGATCTTATTAGTTTCCAAATCA ysGlnThrGlyAsnCysSerValPheGlyProGlnAspSerLeuAlaGluSerGlyHisSerGluSerAspLeuIleSerPheGlnIleT	3019 (524)
3020	CTGAGATTCTTAGGTTAGATTGAGTGGGCGAGCCATATCTAAATACGCGGGCTTATCTGTATGAGATTTTTTTAATACTTCATTGGC hrGluIleLeuArgLeuAspEnd	3109 (530)
3110	TTGAAGTGCTTAATTAAGATTAACATATCCTTACAAGATTCTAATTAGCTACCTAAGTCAATTGTGTTCTTTACACTTATGTAATTAC	3199
3200	TTCATTAAGTTGAAGCCATTGCATAATTTATATAAAATACAATTAACATACCATTATAAAAAAATATATAATCAATTCAAATTTTT	3289
	-----   (A) <sub>n</sub> (major)	
3290	TTCAGGGAATCAATAAATTAATGCTACTCGTTTTTCATAACTAAAGAAACCAACACCACCATAATAATCAACAACAAATATGATTTAT	3379
	(A) <sub>n</sub> (minor)	

Sry $\delta$

3380	GCAAAATGAATATCCGTTTGCAAATATTGAGCAAAACACATATTTTTATTATTCAACTCATATATTCAGTTTTTCACACCCGTGCCATTCC	3469
3470	CACCGTCCCGTTCCTGGCATCAATCGGCATCGTTCGCCACGCGTGGTGGGCATTATGCCATGGTGCATTTTCACGCATTTTAGTATAGCT	3559
	--->3601	
3560	TCCGATATTCATCATTTTTGCCAACTCTATAAAATTCATACACAATTTAAAAGATTGTAACAACACGCCACGAAACGGAAATCGCTAAA	3649
	----- 3668 A=12	
3650	AGGACCATCGTCGGCGCAATGGATACTTGCTTCTTCTGCGGCGCCGTCGATCTGAGCGACACGGGCTCTCCAGTCCATGCCTACGAG MetAspThrCysPhePheCysGlyAlaValAspLeuSerAspThrGlySerSerSerSerMetArgTyrGlu Tyr	3739 (24)
3740	ACGCTGTCGGCCAAGGTGCCGTCGTCAGAAAACAGTGTCCCTGGTCTACCCACCTGGCCAAC TGCATCCAGACGCAGCTGGACCTG ThrLeuSerAlaLysValProSerSerGlnLysThrValSerLeuValLeuThrHisLeuAlaAsnCysIleGlnThrGlnLeuAspLeu	3829 (54)
3830	AAGCCCGGCCCGCCGGCTGTGTCCGCGCTGCTTCAGGAGCTCTCCGACTACGACACGATCATGGTGAACCTGATGACCAACCCAGAAGAGG LysProGlyAlaArgLeuCysProArgCysPheGlnGluLeuSerAspTyrAspThrIleMetValAsnLeuMetThrThrGlnLysArg	3919 (84)
3920	CTGACGACCAGCTAAAGGGCGCTCAAAGTCCGAGTTCGAGGTGCCGGAGTCCGGCGAGGACATACTCGTGAGGAGGTTGGAGATACC LeuThrThrGlnLeuLysGlyAlaLeuLysSerGluPheGluValProGluSerGlyGluAspIleLeuValGluGluValGluIlePro	4009 (114)

	G=SF1	
4010	CAAAGCGATGTCGAGACAGACGCCGATGCCGAGGCGGACGCCCTGTTCTGTTGGAGCTGGTCAAGGATCAGGAGGATCCGACACGGAGATA GlnSerAspValGluThrAspAlaAspAlaGluAlaAspAlaLeuPheValGluLeuValLysAspGlnGluGluSerAspThrGluIle Val	4099 (144)
4100	AAGAGAGAGTTCTGTGGACGAGGAGGAGGAGGAGGACGACGACGACGACGACGAGTTTCATCTGCGAGGACGTGGATGTGGGCGACTCC LysArgGluPheValAspGluGluGluGluGluAspAspAspAspAspGluPheIleCysGluAspValAspValGlyAspSer	4189 (174)
4190	GAGGCCCTGTATGGCAAGTCTCCGATGGCGAGGACAGGCCGACGAAGAAGCGCTCAAGCAGGAGTGCCTACCTGCGGCAAGGTGTAC GluAlaLeuTyrGlyLysSerSerAspGlyGluAspArgProThrLysLysArgValLysGlnGluCysThrThrCysGlyLysValTyr	4279 (204)
4280	AACTCCTGGTATCAACTGCGAAGCACATCAGCGAGGAGCAC TCCAAGCAGCCCAACCACATCTGCCCATCTGCGGGGTGATCCGGCGC AsnSerTrpTyrGlnLeuGlnLysHisIleSerGluGluHisSerLysGlnProAsnHisIleCysProIleCysGlyValIleArgArg	4369 (234)
4370	GACGAGGAGTACTTGGAGCTGCACATGAATCTGCACGAGGGCAAGACGGAAAAGCAATGCCGCTACTGCCCAAGAGCTTCTCGCGCCCG AspGluGluTyrLeuGluLeuHisMetAsnLeuHisGluGlyLysThrGluLysGlnCysArgTyrCysProLysSerPheSerArgPro Cys	4459 (264)
4460	GTGAACACCCCTGCGCCACATGCGCATGCCTGGGACAAGAAGTACCAGTGCAGAGAAGTGCGGCTGAGGTTCTCCGAGGACAACCTA ValAsnThrLeuArgHisMetArgMetHisTrpAspLysLysLysTyrGlnCysGluLysCysGlyLeuArgPheSerGlnAspAsnLeu Ile	4549 (294)
4550	CTCTACAACCACCGGCTGCGCCACGAGGCTGAGGAGAACCCATCATATGCAGCATCTGCAATGTGTCGTTCAAGTCGCGCAAGACCTTC LeuTyrAsnHisArgLeuArgHisGluAlaGluGluAsnProIleIleCysSerIleCysAsnValSerPheLysSerArgLysThrPhe	4639 (324)
4640	AACCATCACACGCTCATTCACAAGGAGAACCGCCCAAGACACTACTGCTCCGCTGCCCCAAGTCCTTACCAGAGCGCTACACCCTCAAG AsnHisHisThrLeuIleHisLysGluAsnArgProArgHisTyrCysSerValCysProLysSerPheThrGluArgTyrThrLeuLys	4729 (354)
4730	ATGCACATGAAGACCACGAGGGCGACGTCGTTTACGGGGTTCGCGAGGAGGCGCCCGGACGAGCAGCAGAGGTGGTGGAGGAGCTGCAT MetHisMetLysThrHisGluGlyAspValValTyrGlyValArgGluGluAlaProAlaAspGluGlnGlnValValGluGluLeuHis	4819 (384)
4820	GTGGACGTCGACGAATCGGAGGCGGCCCTACCCTCATCATGTCCGACAACGATGAGAACAGCGGCTTCTGTCTCATTGCAATACCACC ValAspValAspGluSerGluAlaAlaValThrValIleMetSerAspAsnAspGluAsnSerGlyPheCysLeuIleCysAsnThrThr	4909 (414)
4910	TTCGAGAACAAGAAGGAGCTCGAACACCCTTGC AATTTGATCAGCAGGTGGTCTTGAATAAGCTACATTGCCTACAATAAGTAATTGT PheGluAsnLysLysGluLeuGluHisHisLeuGlnPheAspHisAspValValLeuLysEnd	4999 (434)
5000	TTATCTTCCCTAGTGTATTTCTCCTCTTTGTACTTGATTATTGTAGATTCTACAAAATAATAATTTACTGGTATTTCAATTACTGCGT ----- (A) <sub>n</sub>	5089
5090	TTCATTTAGACAGAAGCATTTCCGATAATAATTGTAC	5126

*Sry* SEQUENCES. Accession X03121 (DRYOSRYG1). The Cys and His residues of the Zn-fingers are underlined. Four mutations in *Sryδ* are also indicated.

activation in blastoderm stage embryos, although at much reduced level. A segment extending between 311 and 118 bp upstream of the transcription initiation site is necessary to increase the level of transcript. The latter segment also seems to be responsible for repression of *Sry* $\alpha$  activity in the peripheral nervous system (Schweisguth et al. 1989).

## *Sry* $\beta$ and *Sry* $\delta$

### Products

DNA-binding proteins of the Zn-finger type.

### Structure

The amino-acid sequences of *Sry* $\beta$  and *Sry* $\delta$  proteins have some similarities to the *Xenopus* transcription factor TFIID and other Zn-finger proteins. There is a repeating unit of 28 or 29 amino acids characterized by Cys at positions 1 and 4 and His at positions 17 and 21/22 of the repeat (a C<sub>2</sub>H<sub>2</sub> finger). A Phe at position 8 is also frequent. *Sry* $\beta$  has six such repeats and *Sry* $\delta$  seven, both are in the C-terminal half of the molecule (*Sry* Sequences). Although residues in other positions are not conserved from one repeat to the next, the C-terminal regions of *SRY* $\beta$  and *SRY* $\delta$  are 50% identical; this suggests that the two genes were generated by a duplication. No sequence similarities are evident outside the coding regions (Vincent et al. 1985; Vincent 1986; Evans and Hollenberg 1988; Payre et al. 1990; Harrison 1991). An 18-amino-acid segment (residues 180–197) was identified as the nuclear localization signal of *SRY* $\delta$ ; within that segment, the heptapeptide Pro-188/Lys-194 has strong similarity to the nuclear localization signals of SV40 large T antigen and *c-myc* (Noselli and Vincent 1991).

### Function

The two proteins bind DNA, both in solution and in polytene chromosomes. *SRY* $\beta$  binding sites include the consensus sequence YCAGAGATGCGCA and *SRY* $\delta$  binding sites the sequence YTAGAGATGGRAA (Payre et al. 1990; Payre and Vincent 1991).

### Tissue Distribution

*SRY* $\alpha$  and *SRY* $\delta$  are maternally inherited and present in embryonic nuclei at the onset of zygotic transcription as well as in numerous cell types throughout development. Zygotic synthesis starts during the syncytial blastoderm stage (nuclear division cycles 12–13) for *SRY* $\beta$  and during germ band extension (stage 10 embryos) for *SRY* $\delta$  (Payre et al. 1989, 1990).

### *Mutant Phenotypes*

Four amino acid substitutions in *Sryδ* are lethal (*Sry* Sequences). These mutants can be rescued by germ line transformation with *Sryδ* but not by an extra copy of *Sryβ* sequences, an indication that the two genes have different functions (Crozatier et al. 1992).

## *Sryβ*

### **Gene Organization and Expression**

Open reading frame, 356 amino acids, expected mRNA length, 1,314 bases. The 5' end was determined by S1 mapping and primer extension; the 3' end was defined by S1 mapping. There is an intron within the Gly-25 codon (*Sry* Sequences) (Vincent et al. 1985; Payre et al. 1990).

### *Developmental Pattern*

See *Sryδ*.

## *Sryδ*

### **Gene Organization and Expression**

Open reading frame, 434 amino acids; expected mRNA length, 1,476 bases. The 5' end was determined by S1 mapping and primer extension; the 3' end was defined by S1 mapping. There are no introns (*Sry* Sequences) (Vincent et al. 1985).

### *Developmental Pattern*

Expression of *Sryδ* (and of *Sryβ*) is very high during oogenesis and early embryonic development; it remains significant, but lower, throughout the life cycle (Vincent et al. 1985; Payre et al. 1990).

*Sryδ* transcripts are abundant in nurse cells up to stage 10, at which time they begin to be transferred to the oocyte. Approximately 4 h after oviposition, transcripts from embryonic nuclei are added to the maternal complement. The total level of transcripts gradually decreases after germ band extension (Payre et al. 1989).

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# 29

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## *Ultrabithorax: Ubx*

**Chromosomal Location:**  
3R, 89E1-2

**Map Position:**  
3-58.8

### **Products**

DNA-binding regulatory proteins of the homeodomain type involved in the determination of segmental identity in the mid-section of the embryo.

### *Structure*

A family of at least five related polypeptides of approximately 40 kD, translated from alternatively spliced mRNAs (see Fig. 29.1B and discussion under Gene Organization and Expression). They all share the sequences encoded in exons at the 5' and 3' ends of the transcription unit; but they differ from each other with respect to whether they include one or more of three short internal segments, one nine amino acids long and the other two 27 amino acids each.

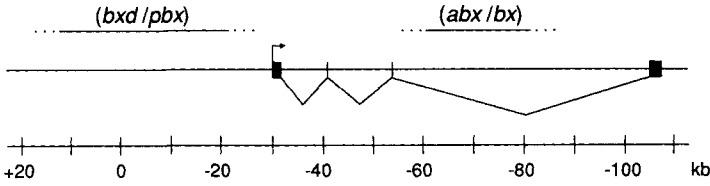
The homeodomain is near the C-terminus. Other sequence features include an alanine-rich segment near the homeodomain (see *eve*) and a glycine-rich segment between residues 111 and 129 (*Ubx* Sequence) (Weinzierl et al. 1987; O'Connor et al. 1988; Kornfeld et al. 1989). For a comparison of *Ubx* protein and DNA sequences in *D. melanogaster* and other species, see Wilde and Akam (1987).

All isoforms of UBX are multiply phosphorylated at Ser and Thr residues that occur between amino acids 39 and 183. Most of the phosphorylation is between residues 130 and 183 (Gavis and Hogness 1991).

### *Function*

UBX helps to define segment identity (Lewis 1978) by acting as a transcriptional regulator. There is evidence that UBX acts on homeotic genes. In particular, it stimulates its own transcription while repressing transcription of *Antennapedia*

A. *Ubx* Domain of the *BX-C*



B. *Ubx* mRNAs

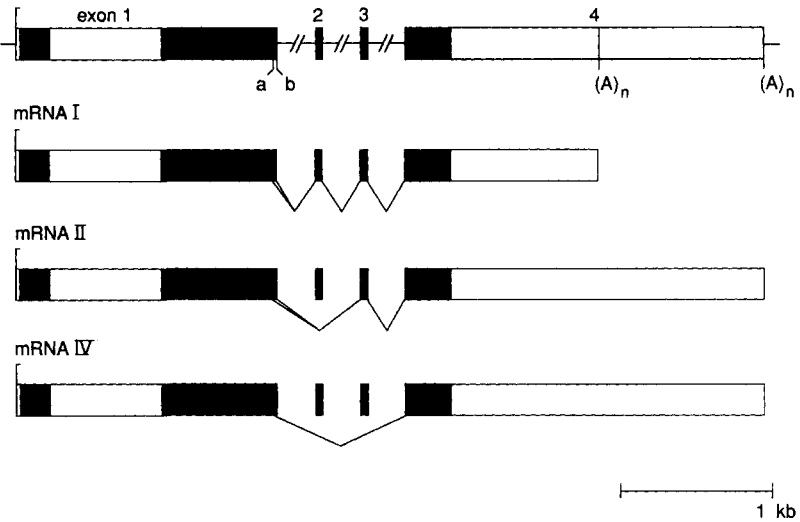


FIG. 29.1. Organization of the *Ubx* transcriptional unit: (A) based on Simon et al. (1990); and (B) based on O'Connor et al. (1988) and Kornfeld et al. (1989).

(*Antp*); this may serve to modulate the segmental distribution of the two products (Beachy et al. 1988; Biggin and Tjian 1989; Samson et al. 1989 and references therein). The identity of the UBX-controlled effector genes that are directly responsible for carrying out segmental differentiation is a subject of active current research.

UBX Ib, a member of the family that includes all three internal polypeptide segments, was produced in *E. coli* and cultured insect cells and tested for DNA-binding activity using DNA fragments from the neighborhoods of the *Antp* and *Ubx* genes. Five binding sites were found near *Antp*: A-1, A-2 and A-3 (approximately 6 kb upstream of the transcription initiation site P1), A-A and A-B (300–400 bp downstream of P1). Two binding sites, u-A and u-B, were detected near *Ubx*. These are 60 bp and 250 bp downstream of the transcription initiation site (*Ubx* Sequence). Multiple repeats of the trinucleotide TAA is a characteristic of all these binding sites (Beachy et al. 1988).



*Ubx*

	Hm	
-4111	ATGACAGAAAAAGTAAGAACAGTTAAGTTATTCAATTAATAAATGGATTATTAGTTTTAGGAAACTCCAAGCACTTGTAAAAATCGAATT	-402;
-4021	TGTTCAATAACTGCATGATGTAGCAAGAAGTAATGTATTTTTAAATATTATTGCCTTATAGCTATGGCCATTTTTAAGTATTTTTCCCA	-393;
-3931	GTGCACCATCTAACAGGTGCCGAGCCGCATCGAACAGAGAAGTCCGAAAGACACAGCCGAAATCCTTATAGACAATACGTAACAAGTC	-384;
-3841	GGAGAGTTCAGGCAGTATTTTGTGAACATTTCTGTGTAATAAACCTCGGGGCCACGAAAACCTTGTATCTCGAATGGAGGAGGGCAACT	-375;
-3751	AGTATACTCTGGTACTGGCCGTTTGATGTTCTGGACTGGCGTCAGCCGGCCGCTCCAGCTGCCAAATGCTGCTTTATTAGTCGCGTA	-366;
-3661	AGTGGCTCCCCCTGATTTTCTGCTTCCACCTGGAGCAAATGTATCTGTTTTGGACTATGATTAGATTGGGTGCACCATCGCACGCA	-357;
-3571	TACGGATGGCATCGCTCGATTTGAGCGATTGTGGCCAATAAACAGCGGGTGAGAAGGCAACAGCTGCCAAGTGGCAATTAACGGC	-348;
-3481	TTGTCTAATTGCCCTGCACAGTTCTCAACAGCGAATGGTGAACGGAGATGGAGGCCATCAATCAAGGCGTGGTGTGAAGGAGGAGTCA	-339;
-3391	GGTTTTGGCCACGGATTCGGCCGCTCGGGCTAATTGGCCACATTTAGCATTGTCCATATCCACTGGGCAACTGGTCAACCTCAGGCTA	-330;
-3301	CTTGGACAGGTGTGAGCTTTCATTTAATCCCCCTTTTCGCGAAACGGAAGCTCTCGTAAATGCTGCAACAAGCTACCGATGACAGTGA	-321;
-3211	AGCGGGGGCGCTGGTGGTGGCCATATGAAAAATGAGATCGCTTTGTATGCAAATGCCTGGGAATCGAATGCGAATCGGGAATCGTACAAT	-312;
-3121	CTCATTGCGACTTTATGCCAAGACAATCGATGCCTCCCTTTTCGGGTCGCTGGGCGTGGTGGTGGGGCTTCCATTGTTAAAACGTGTT	-303;
-3031	TACACATCCAGAAGAAAAGAAATAAATAAATGCTGCTGCTATTGAGAGATGTTACTAGTTTCTAAGTAAAAAGCTCTTTTCATCTTAATCG	-294;
-2941	TAATTTTCAAATTAATAGGATTGGTGAAGAACTCAAAAACGTTTCCACTTTCTGAAAGAATAGATTCTCAGAACTAAAATACATCAACT	-285;
-2851	CATAATCGAGCAGTAACACAAAACCTCCTCTTATTTACGCCAAGCTCGGGAATGCAGAACGGAGGAAAAACAATCATCGATGTCGAACA	-276;
-2761	AAAACAAAACCTTTCTGCAGGAGGAGGCTCTGCAGGAGTAACGAGAGCAACGAAAGGAGCAGAAGCCACAAGGGGAAGCTGGCGCTG	-267;
-2671	CCCCATTGCATACCCACCATAACGTAGCAAGTTTGAATATACTCGCACCCGTAAGATTTCCCGAGTATATTAGTAGCAAAAATTTTACG	-258;
-2581	AGCTCATTATGGCTCATTTCGGCGATTGTTGTGATCCTTTTATGCGCCTTCGAATGGCTTCAAATGGTTATGAGGCTATTTCTCTGTCAT	-249;
-2491	CCCCGCGAGTCTTCGCACTCGTGCTTCCCCGGGTCCAAATGCGGGATAGCGAGTTTCTGGGCTCGGATTTCCCGACTCAGGATAT	-240;
-2401	TTAGTTTGCAAGTTCGCACTGCGATTTTTACTTTTACTTTTGTCTTGGCTCTGGGCTTCTCGCCTTTTGGCTTCGCTTTTCTTGGGCTTTT	-231;
-2311	GGTCGCATATTTAGAAAAATGTCGCCGTGCTCCGAATGTGATCAAGTGTGTCAGTGTGTGTATGTGGTGGTGCATCTGTGTTT	-222;
-2221	TGTGCAAGTTTTGTATCTTTCGCTTGTGTGATTTTTAAACTTGGCACCGAAAATTTGGTCGCGGAAAAATGGCGGAGCAGGGTT	-213;
-2131	GGAAAAAGTAGAAAAAGTAAATTTCTTATAATAAGTTTATTTTGCAACACTTGTGGCAAGCAACTTGTGTTGTTGCCGTCGGTTTCT	-204;
-2041	GGCTTAATCTACGAGCCGTTTGTACGAATGTGGAAACTTGCAGAAATATTGCGAAACTTTCGCAATGGCGGGCGTTCTCGGTCTGCA	-195;
-1951	AGACCCGATCTATTTGCATATGCATATGAATTTATTAGCATCAATCCCACCATATTTACATTGACATTAATATCCCATTGAAAGTATT	-186;
-1861	GCCGAACGGATTGAACATTTTCGATTCGAATGCGAATTTGAATTTGAATAGGAAAAAGGAAAAACCGCAGCACACAAATTTTCCGGTCCCTG	-177;
-1771	GCTTCTATTACCGTACAATGCCGAGTTGGGGTTGACTTATTATCAATAAACTAATGCTAAACACGAGACTTTTAATAATAAAAAATAAT	-168;

-1681 GTCCTCATTAAATATATTATAATTCGTTTTTAAAGGCCAATAAAGCCTTAAAGTGATAAGTATCTTTTTATGCCATACCAAAATTGAATTA -1592

-1591 AGCTTATAGTTTTGAGGTAAGGTA AAAACGATTAATTTACTAATATTTTTCTAAATTGTA AAAACA AATTATACGATATTTACTTCCAT -1502

-1501 AGAAATTAATAATAATTACAATATTCGACTATTCTTTAAAAATTTGTTGTTGCAAATATTTTCCCAAAATATTAACAATTAATA -1412

-1411 ATGACCAGCTTAAACTGAAAAGAACAGACAAAGAATTTGCGTAAGCCGATTCTAGCACAAAGATTGGGAACCGAAACTGTAGTCATGA -1322

-1321 GTGCGCTAAAACCGAGCAAATACGGGATGCGCATCGTTTGGCGTGCAACTGGCAACTGGCGGGCAGCCCGTCGAGCGTTTTTCGCCACTC -1232

-1231 AGTTGAAGGAAAATCAGCCCTCCTCCATGATGAATTTCCGCGGCGAGCGCATTTTCTTCTCGCGAATGAATGAACGAGAGGCCCA -1142

/////////////////////////////////>g4

-1141 CCCCATAAACTTAAACTGAACGAACACTCAAGAGAGAGCGCAAGAGCGCTCAAAAACAATCTGGTTTTGAGCGTTTCGCTGGCTCTCTG -1052

/////////////////////////////////>g3 -----fp4 </////////////////////////////////g2

////////////////////////////////|z5 //////////////////////////////////|z4 //////////////////////////////////|z3

-->-->-966/-964

-1051 TTTCTGTTTTCCACTCGTTTTTAGGCCGAGTCGAGTGAGTTGAGTCGGCAGAGCAAAGTCAAAACACTGGCAACTGCGATTTGGTGCCAC -962

g2//////

/////////////////////////////////>g1

////////////////////////////////|z2 //////////////////////////////////|z1

-961 ATTCGTTTCGATGGCAACGATTGGATAACAGGCGCGCTTGTGTTTTATTATCCACATTATCAGCGCATTATGTTATTATTGGCCCTC -872

MetAlaThrAspTrpIleThrGlyAlaArgPheValLeuLeuSerThrLeuSerAlaAlaLeuLeuLeuLeuAlaLeu (27)

-----A ----- u-A

-871 AGCGCTTACCAGCTCGCCACGCGTCCGCCGTGAATGCCGCGGAAAAGTCGCTTCCACTAGATTGGCGTCCAGATTCGAGGAAATC -782

SerAlaLeuProLeuAlaHisAlaSerAlaArgGluCysArgAlaGluLysSerLeuSerThrArgLeuAlaSerArgPheGluGluIle (57)

-781 CGTCAGCAGACTCATTGCGCCCGTTTCGGTCAGCACTAAGGCTAATAATCGTTCAAATCGTTAAAACATAAAAAATAATAATTGCAA -692

ArgGlnGlnThrHisSerArgProPheGlyGlnHisEnd (69)

-----

-691 TAACAATAAACATAGTAATAATCGTAACGCTTACGAGCCTTGATAGTGCCAAGGCAAGCGCAATCCAAGTATTCAAATTCGAATTC AAT -602

----- u-B

-601 TAACAGCAAAGTGCAATTGGCTAAAAACCGAAACCAAAACGCAACAAGATACGAAACACTTGTGAAACCGTACAACAATTGTGGAAA -512

-511 AAAAAATAAAAGATTATTAAGATTGAAGTCTCAATAAACATTAGTGCTTAAATAAATTTAAAACGACCCGCGTGGAGAGTGCAATAAAAA -422

-421 GAATAACTTTTGAATAAATATTTACCAACAGAAAAATTTTTATAAATTTAAATAAGTGAAAAACA AATTGGTTACTCTGAAACAA -332

-331 AGAATATTCAAATTTGGTGCTAAAACAAGGAGAAAAAATTTCAAGAATTATTATACAATAATAAGACATATTTAACTATATAAAACCAA -242

-241 ACTTAATCAACAAGACAAAGGAGTGA AAAAAATAAAAAAATTTAAAAGAGTTAAAAA AATTTGTTTATATCCAAGGAGGCAAAG -152

-151 GAACAGCACAGAAAGCGAGGAAACACTCAAATAAAATCCGCCAAAAATCGCAGATCCCTGGAAACCAATTCGTGTGAAATCGGTCAAGCC -62

-61 CCCAACGACTTTTAGCCGCTCTCAGACGGAGCACCGCCAAGATTCTTACCGCCAGCAGCGCAATGA ACTCTACTTTGAACAGGCCCTCGG 28

MetAsnSerTyrPheGluGlnAlaSerG (10)

.-| - .-| Def 6.28

29 GCTTTTATGGCCATCCGACCAGGCCACC6GAATGGCAATGGGCGCGGTGGCCACCAGCAGACGCCAGTGCAGCGCGGCCGCGT 118

lyPheTyrGlyHisProHisGlnAlaThrGlyMetAlaMetGlySerGlyGlyHisHisAspGlnThrAlaSerAlaAlaAlaAlaT (40)

119	ACAGAGGATTCCTCTCTCGCTGGGCATGAGTCCCTATGCCAACCCACCATCTGCAGCGCACCACCAGGACTCGCCCTACGATGCCAGCA yrArgGlyPheProLeuSerLeuGlyMetSerProTyrAlaAsnHisHisLeuGlnArgThrThrGlnAspSerProTyrAspAlaSerI	208 (70)
209	TCACGGCCGCCTGCAACAAGATATACGGCGATGGAGCCGGAGCCTACAACAGGACTGCCTGAACATCAAGCGGATGCGGTGAATGGCT leThrAlaAlaCysAsnLysIleTyrGlyAspGlyAlaGlyAlaTyrLysGlnAspCysLeuAsnIleLysAlaAspAlaValAsnGlyT	298 (100)
299	ACAAAGACATTTGGAACACGGGCGGCTCGAATGGCGGGGGGGTGGCGGGAGGCGGTGGTGGCGGGAGCAGCGGGGAAACAGGTGGAG yrLysAspIleTrpAsnThrGlyGlySerAsnGlyGlyGlyGlyGlyGlyGlyGlyGlyGlyAlaGlyGlyThrGlyGlyA	388 (130)
389	CCGGCAATGCCAATGGCGGTAATGCGGCCAATGCAACCGGACAGAACATCCGGCGGGCGGTATGCCCGTTAGACCCTCCGCCTCGACCC laGlyAsnAlaAsnGlyGlyAsnAlaAlaAsnAlaAsnGlyGlnAsnAsnProAlaGlyGlyMetProValArgProSerAlaCysThrP	478 (160)
479	CAGATTCGGAGTGGGCGGCTACTTGGACCGTGGGCGGCGAGTCCCGTTAGCCATCGCGGCGGAGTCCGGCGGTAATGTGAGTGTCA roAspSerArgValGlyGlyTyrLeuAspThrSerGlyGlySerProValSerHisArgGlyGlySerAlaGlyGlyAsnValSerValS	568 (190)
569	GCGGCGGCAACGGCAACGCCGGAGGCGTACAGAGCGGCGTGGGCGTGGCCGGAGCGGGCACTGCCGGAATGCCAATTGCACCATCTCGG erGlyGlyAsnGlyAsnAlaGlyGlyValGlnSerGlyValGlyValAlaGlyAlaGlyThrAlaTrpAsnAlaAsnCysThrIleSerG	658 (220)
659	GCGCCGCTGCCAAAACGGCGGCCGCCAGAGTTTACACCAGGCCAGCAATCACACATTTACCCTGGATGGCTATCGCAGGTGAGTGC lyAlaAlaAlaGlnThrAlaAlaAlaSerSerLeuHisGlnAlaSerAsnHisThrPheTyrProTrpMetAlaIleAlaGlyGluCysP	748 (250)
749	CTGAAGATCCGACCAAAAGTGAGTGTCCTACTGCAGCA* INTRON 1 (10 KB) roGluAspProThrLysS	(251)
	*TTTCAGGTAAGATAAGATCTGATTTAACACAATACGGCGGCATATCAACAGA erLysIleArgSerAspLeuThrGlnTyrGlyGlyIleSerThrAs	838 (271)
839	CATGGGTAAGAAAATTTCCACTTTTATTTTCGTTACATTATTCGCTCTTAAGTTTTCCGAAAAATAGAGTATAAAGGTAGAGCAGGTCCA pMetG	928 (273)
929	CTAACAAACCGTAGAGAACTAATCCCATATGGTGTGGTGGCTAAAATATTGTAGTATTCGCTTTAAGGTGTGCAAAATTCATGAATC	1018
1019	AATGGGCGGGTCTGTGGTGGACCGGAAAACCTGGGGCCGCGTGTGAAATGATTGATTCACTGTCCGCA* INTRON 2 (13 KB)	
	*GACCTTTATAAACGTT	1108
1109	TTCTCTATTTTTTCCAGGTAAGAGATACTAGAATCTCTTGCGGGCTCACTTCTACCAGACTGGCTAGGTAAGTGAAGTTTTGTTAT lyLysArgTyrSerGluSerLeuAlaGlySerLeuLeuProAspTrpLeuG	1198 (290)
	A=195 End	
1199	ATTTTTTGAACCCC* INTRON 3 (50 KB)	
	*GGATCCTGTATTTTGTACCATTTCGTTAAGACTTTCTGAGAGATATGGCCGACAAATGCCATAAACTGAC	1288
1289	GCATCGCAATCTGTGACCTGTCACTGGCCAATTTCTGGCACCATAAATGGGCTTTTATAATTCGCAAGGCAGTTAAAAATAAAG	1378
1379	CCACATTAAGGAAATTTATCAGCATGCATGAGCGAGTCCATATAGATGATAATTTCTTGTGTCATCTGGCCATTTTATTTCCATATCA	1468 Def 9
1469	TTTTTATTGCTATAAAAATTTTTTCGCCATTTATTTCCACCACCCACACAATGCAATCGCTACGTACGCTGCACATGTGTGCTTTGTG	1558

1559	TGAGTGAAGATAAAATGCGTTTTACACTTTATGACTTTCGTGTCGGCATAAAATTTGTTAATACCTTTAGGCCAAATTTATAACATAATAAA	1648
1649	TGCTCATAATATTTAACTTAACATTGTGCTCGGGCCAGGAGAAAGACCTGGTCTCCAAAATGCCAAGTTAACATGGTCGAATGGGTGGG	1738
1739	TTGGTTGGTTGATATGGTGTGGTATGATTGGTATGGGTGATTTCGATAATATCAGACATTGTCTGGGCCCTCTTCTCGATGGGAGATG	1828
1829	GGCCAGAGACAGCTGCAGTGCATTTGCACACACACGAAATGAGTATTGCACTTGAAGGCCAAATTAACCTCATAAATATTTAAATCA	1918
1919	GAGATTAACACGGCATTGTTGCAACATGTTGATGCGACTTCTGGCTGCCCCGGCTCCCCTGGCTCCCCCGGATTCCCCTGGATTCCCC	2008
2009	TGCTCCTCTGCCCCATCTCGTCTCTCAGGTTGCCAATTAACGGGCATTATCTGGCATAACTGCAATTTAAGTAGCCACATTGCCCCATA	2098
2099	TCCCCAGTCCAATGCCACAACCGAGTGTCTGACAGTTCCTCTTTTCATTTTAATGTGGCTGCATCTGGGATCTGTGTATCTTTGTATC	2188
2189	TGAGGAACGTGGAACTGCGAATCTGGATGCAATGCAGCAGCGCAGCAACATTTGGCGGTGCAGCGGCAACCATCAATTTAAAGTAACG	2278
2279	ATCGCGCCGAGAAACAAAACCGCAACTGCAAACCTGGCAAACCTGGCAAATCTCGCGGATCTCGTAAGAGTAAATGATTTTTTTGCG	2368
2369	CTGAGATCCCCTTTCCTTTGCGATTGGGCCCTTTGCGAGCAATTGCGGCCACGTTCCGAGCTGCTGTATGATCGCTGGCAAAAAGGAGA	2458
2459	ATTTATATTTACGACTTGGCCAAATAACAACGGCGAACAGCAAACAATGTTGAGTTGCTCGTTGCAATTTTCAATTTAATTTGCCCGA	2548
2549	TGAAAGGCCAAAATATAAATACCCGAAAACACTCTGTCACTGCTGCTCAATATGACTCAAATTTGATGCTCATGTTCTCCTAAACGT	2638
2639	TAATATAACCAATTAATCACTTTTGTGGCGATTTATATAAATAATAGGCCAATAAATCGATAAAGATATATATCTACTTAGTCACTTT	2728
2729	GTCAATGTTTTCTAACACATATCTGCATTTTGTAGTGTGTTATGAGACACATATTTTTGATTGCAAAATGAAATGTATGTATTTCTG	2818
2819	CGATGCAGGTCCAAAATGAATAATATAAAGTTTAATAACTCTGGTTACTTACAAATGCTTTCCATTCCGATCTACAGGTACAATGGT	2908
		lyThrAsnGly (293)
2909	CTGCGAAGACGCGGCGACAGACATACACCCGCTACCAGAGCTCGAGCTGGAGAAGGAGTCCACACGAATCATTATCTGACCCGCGAGA	2998
	LeuArgArgArgGlyArgGlnThrTyrThrArgTyrGlnThrLeuGluLeuGluLysGluPheHisThrAsnHisTyrLeuThrArgArg	(323)
	- * * * * * - - - - - * - - - - - * - - - - - H1 * - - - - -	
		- AAATTT=Def 9.22
2999	CGGAGAATCGAGATGGCGCACGCGTATGCTGACGGAGCGGCAGATCAAGATCTGGTTCCAGAACC GGCGAATGAAGCTGAAGAAGGAG	3088
	ArgArgIleGluMetAlaHisAlaLeuCysLeuThrGluArgGlnIleLysIleTrpPheGlnAsnArgArgMetLysLeuLysLysGlu	(353)
	- - - - - * - - - - - H2 * - - - - - * - - - - - * - - - - - * - - - - - H3 * * *	
3089	ATCCAGGCGATCAAGGAGCTGAACGAACAGGAGAAGCAGGCGCAGGCCAGAAGGCGGCGGCGCAGCGGCTGCGGCGGCGGGTCCAA	3178
	IleGlnAlaIleLysGluLeuAsnGluGlnGluLysGlnAlaGlnAlaGlnLysAlaAlaAlaAlaAlaAlaAlaAlaAlaValGln	(383)
	HOMEODOMAIN	
3179	GGTGGCACTTAGATCAGTAGATCCTTAGATCCTTAGATCCTTAGATCCGTAGGTTGTATGTTGGATTGGCGAAATGACCGGAGACAG	3268
	GlyGlyHisLeuAspGlnEnd	(389)
3269	ATACAAAGCACTATATTGTAACAAATGAACTATTTACTTAAATGAATAATATTTAAATATTTTGGTACTTGTGCGAATACGAACT	3358
3359	TAACTAAATCGAACCTAATGGAATTTATTCAGCGTTTGAGCAGCAACCGAAAATACGTAATGAAACAAAACCTACAACTAATTAAC	3448
3449	AGGCTAAGTAAATAAAGTAGTGGAAAGGAGCGCAGATTATAAACCTACTTAGAATTAATGAGCAAACAAAACATTTAATTTAGTTCC	3538
3539	AAACGAAAAAAAATCAAGAGGATTCGCTCGAAATGAAAACCTCTGTCTGCCCTTTGTTGCTTACTGCTATGTTAAATTAATTTTCG	3628

(continued)

3629	CGAAAAATACTCAAAAATTGAAACACAAGAAACAACAAAAATGAAAGTATACCATTATAATGTTGAATGCGAGCAAAATCTGTTGATAT	3718
3719	GAATTTTTGGTAAAAACATGTTCTAAACCAATTTAAGATACGTAACGAAGGATGCAAAAACAAAATGAAAACATTAAACTTTAACTTAA	3808
3809	ATATAAATAGAATTTGTTAGCCAGTAACATATTACGACACGAAGAACAACGTTTTCCGGGAGTATCGAATATTTGAATGTGTATAGTT	3898
3899	TGTGCTTATTAATAAAAATAATGCAATTTTAGTTAACTCTGTTTATTGTAACGAATTTGTTTAGTCTCGCCCAACAGCTAGAGTGA	3988
3989	AGCTGTTCTTTAAGTAATGTGTAGTGTGTTACTTTTTAAATAAATTAATGCCTAATTTATTATTATTATGTTAGTTAATGACAA	4078
4079	GCGTTTATGAGATTATCCGACAGAAGCGGCGAGAAGAGGAGTGCACAAACCGTTTGCCCGCAACGCAATAAATTTATTGGTTTTGA	4168
	-----	
4169	AAAAATCTAAGAAAAACAAAAAACAATGAGAAATCGAATCCGATTGTTGTGTTATTATTTTAGTCTGCCATTGCGATTTCCG	4258
	(A) <sub>n</sub> proximal	
4259	TTCTCCAGTGTAATTAGACCTGAGTTGTTGTGAGAGAGTCTCGGGCTACCCGCTGCATGCGAAATGCTTTTGATCTCGTTTGGAC	4348
4349	CGTTAATTGATCGTGAGTTGTACGCTCTATAGAGATACCCATACCGATTAGCTATAACGATACCATACCGATACCAATACCATATATATA	4438
4439	GTTTAGTGGATCC 4451	

*Ubx* SEQUENCE. Accession, Y00206, X05723 (DROUBX1), X05724 (DROUBX2), X05725 (DROUBX3), X05727 (DROUBX5), X05427 (DROUBXG5). Discontinuities in the sequence at 785, 1,091 and 1,213 correspond to introns that have not been completely sequenced; those gaps are not reflected in the numbering system. Position 739 is the alternative donor site of the first exon. Underlining in the interval between -1,160 and -600 marks protein-binding sites. The various sites are associated with the following proteins: g1-g4, GAGA protein; z1-z5, ZESTE; u-A and u-B, UBX; fp4, a protein that also binds to the *Ddc* promoter; and A, an unidentified protein (Biggin and Tjian 1988). Marks above the sequence indicate the following mutations: a vertical bar (at -4,036), the breakpoint of translocation Hm, a regulatory mutation of the *bx*d/*pbx* type (Bienz et al. 1988); |- -|Def. 6.28 (between position 81 and 112) and |- -|Def 9.22 (between positions 1,468 and 3,046), two *Ubx* deletions; and an A-for-G base substitution (at position 1,173), a nonsense mutation; all but Hm are null mutations (Weinzierl et al. 1987). The limits of the homeodomain are indicated by vertical lines below Arg-295 and Ala-356; helices 1, 2 and 3 (H1, H2, H3) are underlined and asterisks mark conserved positions. Helix 4, is seven amino acids long and follows immediately after H3 by analogy to the ANTP homeodomain (Qian et al. 1989).

The homeodomain controls specificity of DNA binding (Gehring 1987; Hayashi and Scott 1990; Harrison 1991), while other region(s) of the protein act as effectors, either stimulating or repressing transcription (Kuziora and McGinnis 1989). The Gln at position 9 of helix 3 (H3), characterizes UBX as an *Antp* class homeodomain (*bicoid* class homeoproteins have a Lys in that position) (Hanes and Brent 1991). Amino acids within the homeodomain but outside of H3 must distinguish the DNA-binding specificities of UBX and the product of *Deformed*, another homeotic of the *Antp* class, since both proteins are identical in H3 but interact with different genes (Kuziora and McGinnis 1989).

The optimal *in vitro* binding sequence was identified by the following procedure: an affinity matrix containing the UBX homeodomain was used to select random-sequence oligonucleotides capable of binding. The bound oligonucleotides were eluted and amplified by the polymerase chain reaction. The process was repeated several times. The sequence of the selected oligonucleotide, TTAATGG is found near the *decapentaplegic* gene in seven near-perfect copies of that consensus; and these sequences are afforded protection from DNase I digestion by a 70-amino-acid polypeptide that includes the UBX homeodomain (residues 295–365) (Ekker et al. 1991).

### *Tissue Distribution*

Antibodies against an epitope common to all of the *Ubx* products were used to detect gene expression. UBX is first detectable in early stage 9 embryos (approximately 3 h 45 m of development) as a single band that occupies the posterior portion of parasegment 6 (anterior compartment of the first abdominal segment, A1a) (Appendix, Fig. A.3). Next UBX appears in parasegments 8, 10 and 12 and soon afterward in all parasegments between 5 and 13. In parasegment 7–12 UBX forms a repeating pattern wherein, in each parasegment, expression is weaker in the anterior portion and stronger in the posterior portion (Irvine et al. 1991). During the rest of embryogenesis UBX appears in a complex pattern that includes the nervous system; in larvae, UBX is found in imaginal discs. Highest antigen levels are observed in T3p and A1a structures (parasegment 6), in T2p and in the anterior compartment of A2–A7. UBX is localized in nuclei (Beachy et al. 1985).

The tissue distribution of UBX is in general agreement with the sites of gene transcription (see below) and with the sites of gene activity deduced from the effects of *Ubx* mutations.

### *Mutant Phenotypes*

*Ubx* is a homeotic gene. Null mutations transform structures of parasegment 5 and parasegment 6 origin to parasegment 4 type differentiation, and they also cause minor abnormalities of the abdominal segments. (Lewis 1978; Sánchez-Herrero et al. 1985; Duncan 1987; Akam 1987).

### **Organization of the Complex**

*Ubx* is part of the bithorax complex (BXC), a three-gene, 300-kb cluster. Approximately 60 kb upstream of *Ubx* is the 3' end of *abdominal A*, which extends for 25 kb, and 90 kb further upstream is *Abdominal B*. All three genes are transcribed toward the centromere (reviewed by Duncan 1987; Peifer et al. 1987). *Ubx* itself is spread over 77 kb of DNA, and not all of it has been sequenced.

## Gene Organization and Expression

The published *Ubx* sequence includes four exons and small sections of the neighboring introns. The open reading frames of several alternative splicing products vary between 346 and 389 amino acids. The expected size of mRNAs are 3,096 and 3,123 bases (Fig. 29.1B), forms Ia and Ib with polyadenylation at the proximal site) and approximately 4,100–4,200 (forms II and IV with polyadenylation at the distal site, see below). These sizes are in agreement with the occurrence of two main poly(A)<sup>+</sup> RNA bands of 3.2 and 4.3 kb detected by northern analysis. There are introns within the Ser-256, Gly-273 and Gly-290 codons (Saari and Bienz 1987; Weinzierl et al. 1987; O'Connor et al. 1988; Kornfeld et al. 1989). Primer extension, S1 mapping and a cDNA sequence were used to define the 5' end at -966/-964. There is no discernible TATA box appropriately positioned upstream of the *Ubx* transcription initiation site (Saari and Bienz 1987; O'Connor et al. 1988; Kornfeld et al. 1989). S1 protection was used to localize the two 3' ends 1.1 kb apart; the proximal 3' end was also identified by a cDNA sequence (Kornfeld et al. 1989).

Data from two studies (O'Connor et al. 1988; Kornfeld et al. 1989) on a total of 78 embryonic cDNAs indicates the following forms of splicing (see *Ubx* Sequence and Fig. 29.1B): Exon 1 has two donor sites, a and b; site a is used 80% of the time. Splicing can occur so that all four exons are included (forms Ia and Ib, 75%), or so that exon 2 is spliced out (forms IIa and IIb, 21%), or so that both exons 2 and 3 are spliced out (form IVa, 3%); IVb has not been observed. These alternative splicings introduce only small differences in the size of the mRNA: the two donor sites in exon 1 are only 27 bp apart while exons 2 and 3 (the two "micro" exons) are only 51 bp long. Thus, the main differences in the expected sizes of mRNAs depends on which polyadenylation site is used. As already mentioned, the proximal poly(A) site is used predominantly in form I RNA (the form that carries two micro exons) and the distal one in forms II and IV (one micro exon and no exon, respectively).

The unusually long leader region of this gene (1,066 bp) includes a potentially functional second open reading frame of 69 codons. The first 23 residues of this putative protein resemble a signal peptide; it has been suggested that translation of the leader peptide may be involved in regulating translation of the UBX protein (*Ubx* Sequence) (Saari and Bienz 1987).

In addition to the RNAs described above, there are other minor transcripts of uncertain function (O'Connor et al. 1988; Kornfeld et al. 1989).

### *Developmental Pattern*

*Ubx* expression is undetectable before fertilization; transcripts are first detected at the end of the syncytial blastoderm stage, immediately after the 13th nuclear division at approximately 2 h 30 m (Akam and Martínez-Arias 1985). There is a 60–75 min lag between the time of appearance of *Ubx* RNA and the time when protein is first detected (Irvine et al. 1991). This delay has been ascribed to the enormous size of the 77 kb transcript, and Kornfeld et al. (1989) proposed that

size may serve a regulatory function to insure the correct timing of *UBX* protein accumulation. *Ubx* expression increases dramatically between 3 h and 6 h of embryonic development, reaches a plateau by 9 h and remains at a high level until 15 h. The level of transcripts then decreases and remains relatively constant and low through to the adult stage (O'Connor et al. 1988).

The choice of splicing and polyadenylation sites are also developmentally regulated. Form I transcripts predominate (70–80% of *Ubx* transcripts) early in embryogenesis (3–8 h of development); they decrease during middle and late embryogenesis to approximately 30% and then rise once again to 50–60% during larval and adult stages. Form II rises from very low levels early in embryogenesis to 30–40% after 10 h of development and stays in that range. Form IV peaks late in embryogenesis and disappears after the second instar (O'Connor et al. 1988; Kornfeld et al. 1989).

Late in the cellular blastoderm stage (4 h), transcripts are detectable extending from 50% to 20% egg length (Appendix, Fig. A.3). The concentration of transcript is significantly higher in a zone that probably corresponds to parasegment 6 (between 50% and 45% egg length). With the onset of gastrulation, the distribution of transcripts becomes more complex. During the extended germ band stage (6–8 h) transcripts seem to accumulate in ectodermal and mesodermal derivatives of regions that correspond to parasegments 6–12. In parasegments 5 and 13, transcripts are more localized to ectodermal derivatives (Akam and Martínez-Arias 1985). In older embryos and larvae, *Ubx* expression is evident in ectodermal and many mesodermal (but not endodermal) derivatives. In 12–20 h embryos, strongest expression is in the nervous system. Expression is not uniform in all segments: in third instar larvae, expression in muscle extends primarily between A1 and A6; in the nervous system, highest RNA levels are detected in T3 and A1 (Akam 1983).

The mRNAs also display tissue specificity: form I predominates in embryonic myoblasts, while forms II and IV predominate in neuroblasts (O'Connor et al. 1988).

The pattern of *Ubx* expression early in development is determined by the action of maternal and segmentation genes. After the end of the germ band extension period, that pattern seems to be maintained through the rest of development by the products of genes of the *Polycomb* (*Pc*) group. It has been proposed that the *Pc* protein acts by modification of chromatin organization to prevent ectopic activation of *Ubx* (Paro and Hogness 1991 and references therein).

### *Promoter*

P-Element-mediated transformation experiments showed that a segment extending from 1.7 kb upstream of the transcription initiation site to the first codon, when attached to a reporter gene, supports transcription in embryonic ectoderm. The expression is evident along the entire length of the embryo in a segmented pattern and is called the "basal pattern" of expression. The intensity of the "basal pattern" depends on sequences within 626 bp of the transcription initiation site while the segmented nature of the expression is dependent on



regions of the *Ubx* leader that seem to coincide with homeoproteins binding sites (Bienz et al. 1988; *Ubx* Sequence).

*In vitro* transcription experiments defined a minimal promoter region that responds to nuclear extracts of staged embryos: a segment starting 154 bp upstream of the transcription initiation site and extending 41 bp into the leader is capable of supporting transcription in the presence of extracts from 8–12 h embryos (but not with extracts from 0–4 h embryos, where *Ubx* is not normally expressed). Proteins that bind 5' upstream sequences include the GAGA protein, the *zeste* product, and a factor that also binds to a promoter element of *Dopa decarboxylase* (Biggin and Tjian 1988; Biggin et al. 1988; *Ubx* Sequence). At least one element downstream of the transcription initiation site is also required for *in vitro* transcription (designated A, in the *Ubx* Sequence). Just beyond this element are segments u-A and u-B to which UBX binds specifically and which are thought to be important in transcriptional regulation (Beachy et al. 1988; Kuziora and McGinnis 1989). Experiments using cultured cells demonstrated that ANTP and FTZ (the product of *fushi tarazu*) require element u-B to stimulate *Ubx* transcription (Winslow et al. 1989).

In addition to the proximal DNA elements responsible for the “basal pattern”, there are at least two more distal regions that play a role (Fig. 29.1A). The *bx*d/*pbx* region, extending from 3 kb to >30 kb upstream of the transcription initiation site is thought to be involved in the regulation of *Ubx* expression in parasegments 5, 6, and perhaps also in the abdominal segments. A segment of DNA that extends from 35.4 kb upstream of the transcription initiation site to the eighth codon of *Ubx* can drive the expression of *lacZ* in an embryonic pattern identical to that of *Ubx*. A reporter gene construction in which the regulatory region extends 22.2 kb upstream of the transcription initiation site shows some deviations from normal *Ubx* expression; and, when only 5 kb of upstream DNA are included, *lacZ* is expressed in the “basal pattern” described above (Irvine et al. 1991).

The *abx/bx* regulatory region, found within the last intron of *Ubx* (Cabrera et al. 1985; White and Wilcox 1985; Peifer and Bender 1986) contains a 2–3-kb segment (approximately between –77 and –80 in Fig. 29.1A) that behaves as an enhancer and appears to be responsible for defining parasegment 5 as the anterior boundary of *Ubx* expression (Simon et al. 1990).

In a separate set of experiments, Qian et al. (1991) identified a 500-bp segment of the *bx* region (near coordinate –63 kb in Fig. 29.1A) containing an enhancer (called *bre*) that activates the minimal promoter to strong expression in parasegments 6, 8, 10 and 12 and represses its expression in the anterior half of the embryo. The *hunchback* product binds to three sites in the *bre* and this binding is necessary for repression of *Ubx* transcription in the anterior half of the embryo.

### Other Transcripts

The *bx*d region produces a 27-kb transcript early in embryogenesis, between 3 h and 6 h of development. This transcript includes at least 11 exons that are

spliced in different combinations to give rise to numerous distinct polyadenylated RNAs. It is doubtful, however, that these are functional mRNAs because their coding capacity is very poor as judged from the length of open reading frames and codon usage. Another *bxd* transcript is synthesized later, from the third larval instar onward. In contrast to the early transcripts, this is a simple, unspliced poly(A) + RNA with a 110-amino-acid open reading frame and good codon usage. It is not clear what role these upstream transcription units might play in the control of *Ubx* expression. It has been suggested that *bxd* transcripts are completely incidental, resulting because the strong *Ubx* enhancers can activate cryptic promoters (Lipschitz et al. 1987; Saari and Bienz 1987).

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# 30

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## *vermilion: v*

**Chromosomal Location:**  
X, 10A1-2

**Map Position:**  
1-33.0

### **Product**

Tryptophan oxygenase, TO (EC 1.13.1.12), an enzyme involved in the biosynthesis of brown eye pigment.

### *Structure*

A 150 kD protein, it requires a heme cofactor for activity (see review by Phillips and Forrest 1980).

### *Function*

TO converts tryptophan to N-formylkynurenine, the first step in the synthesis of xanthomatin from tryptophan. This is the major pathway for utilization of non-protein tryptophan in higher insects; and xanthomatin is the only brown eye pigment in *Drosophila* (Phillips and Forrest 1980 and references therein). There is considerable similarity between *Drosophila* and mammalian TO (Fig. 30.1).

### *Mutant Phenotypes*

Null alleles such as  $v^{36f}$  and  $v^{48a}$  have no enzymatic activity, do not accumulate xanthomatin and display bright red eyes when present alone, or pure white when combined with *brown* (*bw*), a mutation that blocks synthesis of the red pigment. Severe hypomorphic alleles such as  $v^1$  have a few percent of normal enzyme activity, accumulate a small amount of xanthomatin and develop a slightly off-white eye color when in combination with *bw*. Mutations in another gene, *suppressor of sable* (*su(s)*), cause  $v^1$  homozygotes to accumulate 20% of normal TO level and to develop normal eye pigmentation. *v* mutations are not cell-autonomous (Phillips and Forrest 1980 and references therein) (see below).

```

          1                               50                               100
Rat  MSGCPFSGNS VGYTLKNLSM EDNEEDGAQT GVNRASKGGL IYGDYLQLEK ILNAQELQSE IKGNKIHDEH LFIITHQAYE LWFKQILWEL DSVREIFQNG
Dm   .MSCPYAGNG ..... NDHDDS AVPLTTEVGK IYGEYLMLDK LLDAQMLSE EDKRPVHDEH LFIITHQAYE LWFKQIIFEF DSIRDML.DA
CON  ---CP--GN- -----D---- -V-----G- IYG-YL-L-K -L-AQ---SE -----HDEH LFIITHQAYE LWFKQI--E- DS-R-----

          101                               150                               200
Rat  HVRDERNMLK VMTRMHRVVV IFKLLVQQFS VLETMTALDF NDFREYLSPA SGFQSLQFRL LENKIGVLQS LRVYPYNRKHY RDNFEGDYNE LLLKSEQEQT
Dm   EVIDETKTLK IVKRLNRVVL ILKLLVDQVP ILETMTPLDF MDFRKYLAPE SGFQSLQFRL IENKLGVLTE QRVRYNQKYS DVFSDEEARN SIRNSEKPS
CON  -V-DE---L- ---R--RVV- I-KLLV-Q-- -LETMT-LDF -DFR-YL-PA SGFQSLQFRL -ENK-GVL-- -RV-YN-K-- -----SE----

          201                               250                               300
Rat  LLQLVEAWLE RTPGLEPHGF NFWGKFEKNI LKGLEEEFLK IQAKKDSEEK EEQMAEFRKQ KEVLLCLFDE KRHDYLLSKG ERRLSYRALQ GALMIYFYRE
Dm   LLELVQRWLE RTPGLEESGF NFWAKFQESV DRFLEAQVQS AMEEPVEKAK NYRLMDIEKR REVYRSIFDP AVHDALVRRG DRRFSHRALQ GAIMITFYRD
CON  LL-LV--WLE RTPGLE--GF NFW-KF---- ---LE----- -----K- -----K- -EV----FD- --HD-L---G -RR-S-RALQ GA-MI-FYR-

          301                               350                               400
Rat  EPRFQVPFQL LTSLMDIDL MTKWRYNHVC MVHRMLGSKA .GTGGSSGYQ YLRSTVSDRY KVFVDLFNLS SYLVPRHWIP KMPPIHKFL YTAEYSDSSY
Dm   EPRFSQPHQL LTLMDIDSL ITKWRYNHVI MVQRMIGSQQ LGTGGSSGYQ YLRSTLSDRY KVFLDLFNLS TFLIPREAIP PLDETIRKKL INKSV*...
CON  EPRF--P-QL LT-LMDID-L -TKWRYNHV- MV-RM-GS-- -GTGGSSGY- YLRST-SDRY KVF-DLFNLS --L-PR--IP -----I-K-L -----

          401
Rat  FSSDES*
Dm   .....
CON  -----

```

FIG. 30.1. Comparison of the rat (M55167) and *Drosophila* (Dm) sequences. There is 50% overall identity between the proteins. Sequences were aligned with the GCG *Pileup* program.

v

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-----EcoRI
1170 GAATTCCAAGCACATTGCAAGAATCCCAAATCAAAAAATCGCATGAAATTGCCCCCGTACCTTTTGCCTTTACTCCAGATGTAAGTCA  -1081
.
.
1080 ATTTTTTCTATGCAAAAGTAGTTGAAATATATAATAAAAAACGGATTAGAAAAACAAAACATACATATATATATAATAATATATATA  -991
.
-990 TATATTTAGACACACATCGACAGTATCCTATTCAATTGATTTCTTTGAGAAGTTTGATTTTGCAGTTTGGATATGCAGCAAGAAAAGTA  -901
-900 AAACCAACAACAGAAAATGTGTAAAGAATAGTATAAATAAGTTCGGATTATAATGCCCCGACATTACGCTATATGTATGTCTGGGCCAA  -811
.
-810 ACAGCAACAATCCAATAAAACAAGTAATTAACAACAACAACAACGGATGAAAACAACCAAGATTATGTAAGCGATGGATCGAAGTAAGA  -721
-720 ATTGATGCGAACGGCACAAGTATATAACAAATTTCAACAAGTATATGACTGAGCCAATGACTCAAAAATACATTTTAAATAAAAAGGGAAA  -631
-630 ACCGAGAAATATATGAAAAATATAAAAAACGATAAGCAAGTGAATGAAGCTCTTTTTCTTTTTGGGTTTTGGGAAGTCAAAATTATTGAT  -541
.
-540 TGTATGGAATGTTTTGTTTACCTATTTTGCATATGGTGGCATTGTATCAAAAACAGTTTGAATTATCAAAATGGTTCATTTATTTT  -451
-450 TATACAACCTTGACCTATTTTCAAGGACCAATAAGATTGGACCCACATTAECTTAGAAAACAATACTTGCCATGTTCAATTTTATTCCT  -361
-360 ACGCAGGGTTTATTTATTATACTATGTTAATCAAAAATAAAATGTTAATTTCTCAGTTATTTAACTACACCTTAGTAACTCTGA  -271
-270 TTTGGCATTCTCACTGAAGTACTACTGTAGACTACCTCCATTGAGGAAAATATTGTGTGCGCCGCACCTTCCACTCAAGTGATTG  -181
-180 ATAATCCCAGCCTATCTGGCAGTCCCATCGCCAGATCACCAGCTGTGCAATCAGTCGGAAGTGGAGCTCTCTCGCTCTGTTATCGGT  -91
.
.
.
-----
-->-56.
.
.
-90 TCGCTGGGGTCTCATCTCCGGTCCGTGGCGGAGATCAGTTCGCCAGCATCCGCCCTCGAGGAGTCACGATCTGATCTGAGCTGTGCAC  -1
.
.
.
0 CATGAGCTGTCCCTATGCAGGAAACGGGTGAGCACCAGCAGCTGTCTGTCAGGAATGCCAATCGATCTTCAGTTCTGCGATTCAATTCAA  89
MetSerCysProTyrAlaGlyAsnG1 (9)
.
.
.
90 ACCCATACAGAAACGATCAGCATGATTCGGCGGTGCCATTAACCACGGAAGTGGGCAAAATCTATGGAGAGTATCTGATGCTGGACAAAC  179
yAsnAspHisAspAspSerAlaValProLeuThrThrGluValGlyLysIleTyrGlyGluTyrLeuMetLeuAspLysL (36)
Phe Val
. A |- Def217. -| . G=kLTR |-|=Def208 .
180 TGCTGGATGCCAGTGTATGCTGTCAGGAGGACAAGCAGCCGTGCACGATGAGCATCTGTTTATCATCACGACACGGGTGAGTAGG  269
euLeuAspAlaGlnCysMetLeuSerGluGluAspLysArgProValHisAspGluHisLeuPheIleIleThrHisGlnA (63)
Gln Arg
.
.
.
270 TTTACAACCTTTGATGACAACACTCAATGGCATTAAAGTACCTTCGCCACAGCCTACGAGCTTTGGTTCAAGCAGATCATCTTTGAGTTG  359
laTyrGluLeuTrpPheLysGlnIleIlePheGluPheA (76)
End Arg Phe V
T . Def48a|- . C=257. T=225
360 ACTCCATACGAGACATGTTGGATGCGAGGTCATCGATGAAACCAAGACGCTGGAGATTGTCAAGCGACTGAACCGAGTGGTTCTGATTC  449
spSerIleArgAspMetLeuAspAlaGluValIleAspGluThrLysThrLeuGluIleValLysArgLeuAsnArgValValLeuIleL (106)
al Pro Phe
.
.
.
450 TAAAAGTGAGTGTCTTCTGAATCTCTTACCAAAATCCGTTTATAACTTCCTTTGTACAGCTCCTGGTGGACCAAGTGCCCATTTCTGGAGA  539
euLys LeuLeuValAspGlnValProIleLeuGluT (118)
Glu
.
.
.
Def226=|- -| C=270 234=A 252=|-.-| 253=G .A=245
540 CCATGACCCCGTAGACTTCATGGACTTCCGCAAGTACCTGGCACCCTGCTGGTTTTTTCAGTCGCTGAGTTCCGTTTGTGCGAACA  629
hrMetThrProLeuAspPheMetAspPheArgLysTyrLeuAlaProAlaSerGlyPheGlnSerLeuGlnPheArgLeuIleGluAsnL (148)
A-----laPro Thr TrpAsn

```

	- =Def48a . . . . . T=218 . . . . .	
630	AGCTGGGAGTTCTGACAGAGCAGCGGGTGAGATACAACCAAGTACTCGGATGCTTTAGCGACGAGGAGCGCGGAATTCGATTCGCA ysLeuGlyValLeuThrGluGlnArgValArgTyrAsnGlnLysTyrSerAspValPheSerAspGluGluA1aArgAsnSerI1eArgA	719 (17E)
	End	
		T=223
720	ACTCGGAGAAAGATCCCTCGCTACTGGAGCTAGTGCAGCGATGGCTGGAGAGGACGCCCGGACTGGAGGAGAGTGGCTCAACTTCTGGG snSerGluLysAspProSerLeuLeuGluLeuValGlnArgTrpLeuGluArgThrProGlyLeuGluGluSerGlyPheAsnPheTrpA	809 (20E)
	End	
810	CCAAGTTTCAGGAGAGCGTCGATCGATTCTGGAGCGCAGGTACAGAGCGCCATGGAGGAGCCGTGGAAAGGCGAAAAACACC GCC 1aLysPheGlnGluSerValAspArgPheLeuGluA1aGlnValGlnSerA1aMetGluGluProValGluLysA1aLysAsnTyrArgL	899 (23E)
		201=A=214 A=219. . . . .  - .Def210 - .
900	TCATGGACATTGAGAAGCGACGCGAGGTGTATCGCTCCATCTTTGATCCGGCAGTGCACGATGCATGGTGGTGGGGATCGCCGGT euMetAspI1eGluLysArgArgGluValTyrArgSerI1ePheAspProA1aValHisAspA1aLeuValArgArgGlyAspArgArgP	989 (26E)
	Asn Gly	
		A=244
990	TTAGCCATCGTCCCTTCAGGGAGCCATCATGATCACCTCTATAGGGATGAACCCAGGTTACGCCAACCCACCCAGTTGCCTCACCTGC heSerHisArgA1aLeuGlnGlyA1alleMetI1eThrPheTyrArgAspGluProArgPheSerGlnProHisGlnLeuLeuThrLeuL	1079 (29E)
	Lys	
		T=227 . . . . . T=266 . . . . .    =36f
1080	TCATGGACATCGACTCGTTAATAACCAAGTGGAGATGTAAGTATTGCATCTTTGATACTCTTTATAAATATATCTTATGTTAAAGACT euMetAspI1eAspSerLeuI1eThrLysTrpArgT	1169 (310)
	Val . . . . . SerE	
		250=A . . . . . T=246 . . . . . T
1170	GGTTTTCTAACCAAATACTTTCTATTCGCCGCGCAGACAATCACGTGATCATGGTGAACGCATGATTGGATCCCAACAGTTGGGCACT yrAsnHisValI1eMetValGlnArgMetI1eGlySerGlnGlnLeuGlyThr	1259 (327)
	nd Leu . . . . . Phe	
		-Def237 . . . . . -
1260	GGTGGCTCGTCTGGATATCAATATCTCGCTCCACTCTCAGGTGATCATCGCAGATGTGATTATATCGGGGATCAATGAACTCAAAGT GlyGlySerSerGlyTyrGlnTyrLeuArgSerThrLeuSe	1349 (341)
		242=AACAN A=267 . . . . .    =Def281
1350	TCTCCCTTGTTTTTTTTTGGITTCAGTGATGGTACAAAGGTGTTCTGGATCTGTTCAATCTGTCCACTTTTCTGATTCCCGCGAGGC rAspArgTyrLysValPheLeuAspLeuPheAsnLeuSerThrPheLeuI1eProArgGluA1	1439 (362)
		=H2a
1440	GATTCACCCTGGACGAGACCATTCGCAAGAAACTGATCAACAAAAGTGTCTGACAATCGGCAGGGTATCCAATGGTCAATGTTGGC alleProProLeuAspGluThrI1eArgLysLysLeuI1eAsnLysSerValEnd	1529 (379)
1530	TATGCGTGTGTTGTTCTGCCTACTGTTTTGTCGTTTTGGTGAATAAAATACTGTTTAGTCTTTGTTATCACATTTGATGTGTTCTT -----  (A) <sub>n</sub>	1619
1620	TTCTTTATGTCTGACATATAACATATAACATAACAAAATAAATATTCATATTTAGACATAAACAATTCATGGGAATGTGTGAGTC	1709
1710	AGCAGCCTGAAAGTAGACCATATATTTCTGGTTGCTTTCTCGCTCGTTTCTATTAGTTCGTTAGCAAATTAATCCATAATATTGG -----HindIII	1799
1800	TGGCAATACTGTCAAATAATAATGGTATAAGTGAATTTTAATTACAAAATACCGATTTAACAAAAGGCTTG 1873	

*v* SEQUENCE. Accession, M34147 (DROVERM). Mutations  $v^1$ ,  $v^2$ ,  $v^k$ ,  $v^{H2a}$  and  $v^{48a}$  are indicated as well as a mutation produced *in vitro*,  $v^{KLTR}$ ,  $v^{KLTR}$ , in which Met-32 and His-55 are replaced, codes for an inactive enzyme (*v* Sequence) (R. A. Fridell and L. L. Searles, personal communication). Allele  $v^{48a}$ , in which 50 amino acids are missing, accumulates normal levels of RNA, but produces no detectable enzymatic activity. The mutation  $v^{36f}$  is caused by insertion of a the transposable element *B104* and leads to a null phenotype. The mutation  $v^{H2a}$  is caused by insertion of a *P* element (Searles et al. 1990). Numerous mutations of *v* sequenced by Nivard et al. (1992), and designated by numbers between 200 and 299, are also shown.

## Gene Organization and Expression

Open reading frame, 379 amino acids; expected mRNA length, 1,306 bases, in agreement with an RNA of 1.4 kb detected by northern analysis. Primer extension and S1 mapping were used to define the major 5' end. The two longest cDNA clones identified extend 60–80 bp upstream of the major 5' end; these may represent minor transcription initiation sites. There are no correctly positioned TATA boxes. The 3' end was obtained from a cDNA sequence that included a poly(A) tail. There are 5 introns: in the Gly-9 and Ala-63 codons, after the Lys-107 codon and in the Tyr-310 and Ser-341 codons (*v* Sequence) (Searles et al. 1990).

Mutations  $v^1$ ,  $v^2$  and  $v^k$  are all the result of insertion of the transposable element 412 in the leader region. Homozygotes for these mutations accumulate trace amounts of a *v* RNA of almost normal size. This apparently functional RNA (its coding region is unaltered) is produced because transcription from the *v* promoter is normal, and because rare splicing events using cryptic splice sites near the ends of 412 remove most of the 412 sequences from the *v* transcript. Mutations in *suppressor of sable* (*su(s)*) lead to increased accumulation of these spliced RNAs and thus to suppression of the mutant phenotype (Fridell et al. 1990; Pret and Searles 1991). A similar mechanism of suppression is found in some *y* mutations.

### *Developmental Pattern*

*v* RNA begins to accumulate in 12–24 h embryos, it remains at a constant level between the first larval instar and the beginning of the third larval instar, becomes very low during the pupal stages and rises again in adults. Using a chimeric *v-lacZ* construction that included 1.1 kb upstream of the transcription initiation site and the *v* leader, it was determined that larval expression is restricted to the fat body (Fridell and Searles 1992).

### *Promoter*

Analysis of deletions of upstream and leader segments showed that sequences upstream of the 5' end plus a segment of the leader are necessary and sufficient for normal expression in transgenic animals. The upstream elements are located in the intervals –550 to –350 and –210 to –110 and the leader element is between positions –38 and –12 (Fridell and Searles 1992).

## References

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# 31

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## Vitelline Membrane Protein Genes: *Vm26Aa*, *Vm26Ab*, *Vm32E*, *Vm34C*, *Fcp3C*

### Chromosomal Location:

<i>Vm26Aa</i> , <i>Vm26Ab</i>	2L, 26A
<i>Vm32E</i>	2L, 32E
<i>Vm34C</i>	2L, 34C
<i>Fcp3C</i>	X, 3C

### Map Position:

2-[20]
2-[44]
2-[47]
1-[3]

### Products

The vitelline membrane is made up of 6–10 proteins that range in size from 10 to 100 kD; these proteins are secreted by the follicle cells that surround the developing oocyte.

### Structure

The complete sequences of four genes for vitelline membrane proteins (*Vm26Aa*, *Vm26Ab*, *Vm32E* and *Vm34C*) are available: all of these genes are in the left arm of the second chromosome. The predicted amino-acid sequences for the four proteins include a common 38-amino-acid segment: within this segment, the sequences of *Vm26Aa* and *Vm34C* are identical to each other and to the consensus sequence; the *Vm26Ab* sequence differs from the consensus in 10% of the positions and the *Dm32E* sequence differs by 24%. Outside of this region, the protein sequences are quite different, but putative signal peptides have been identified. *Vm26Ab* has 6–7 repeats of the octapeptide Tyr-Ser-Ala-Pro-Ala-Ala-Pro-Ala, a sequence that occurs only once in *Vm32E* and *Vm34C* (Fig. 31.1). These predicted sequences indicate the proteins are rich in Ala (10–27%) and Pro (9–16%) (Popodi et al. 1988; Scherer et al. 1988).

```

1                               50                               100
Vm34C MKCIAIVSTI CLLAAFVAAD KEDKMLGSSY G.....G GYGK.PAAA.....PAP SYSAPAAASP GLRAPAAPSY AAAPV.....
Vm26A1 MKSFVCIALV AFAAALASP TNVASATGST GSSVTTQDGE LEGVTGQGFQ DLRLRKSAY GGSSGGYGGS .....
Vm26A2 MAFNFGHLLI AGLVALSAYS SETIQLQPTQ GILIPAPLAE NIRVSRAAYG GYGAAPAAPS YSAPAAPAAQ AYSAPAAPAY SAPAAPAYSA PAAPAYSAPA
Vm32E MQI.VALTLV AFVAIA.....
CON M-----L- A--AA--A-- -----G-----G----- ----- --A-----

101                               150                               197
Vm34C .....SIPA PPCPKNYLFS CQPNLAPVPC SAPAPSYGSA GAYSQYAPVY APQIQW*.....
Vm26A1 .....SIPA PPCPKNYLFS CQPNLAPVPC SAPAPSYGSA GAYSSPVATY VAPNYGVPQH QQQLYSAYVP QTYGYQY*.....
Vm26A2 APAYSAPAAP AYSAPASIPS PPCPKNYLFS CQPSLQPVPL SAPAQSYGSA GAYSQYVPQY AVPFVREL*.....
Vm32E APAYSAPAA.....SSGYPA PPCPTNYLFS CQPNLAPAPC AQEAPAYGSA GAYTEQVPTT WTSNREQLQ QFHQRIGMAA LMEELRGLGQ GIQQQY*
CON -----SIPA PPCPKNYLFS CQPNLAPVPC SAPAPSYGSA GAYS--VP-Y-----

```

FIG. 31.1. Amino-acid sequence comparison of four vitelline membrane proteins. Gaps were introduced to highlight sequence features present in more than one protein. The CON(sensus) line indicates positions at which three of the four sequences agree.

### *Tissue Distribution*

Synthesis takes place during egg-chamber stages 8–11, i.e., immediately before the synthesis of the chorion proteins that will form the outer eggshell (Petri et al. 1976; Fargnoli and Waring 1982; Mindrinos et al. 1985).

This chapter describes genes that are expressed exclusively in follicle cells at the time of vitelline membrane synthesis and, in addition to vitelline membrane proteins, includes the gene *Follicle cell protein at 3C*.

## **Follicle Cell Gene Cluster at 26A**

### **Organization and Expression of the Cluster**

The cluster consists of four transcriptional units (TU) contained in a little over 7 kb of DNA (Fig. 31.2). TU2 and TU4 (*Vm26Aa* and *Vm26Ab*, respectively) have been sequenced. Their *in vitro* translation products comigrate with identified vitelline membrane proteins. The other two transcription units are expressed at much lower levels: TU1 produces a 1.3-kb transcript; TU3 produces a 0.7-kb transcript which may be translated *in vitro* into a 20-kD protein. All four genes in the cluster are expressed exclusively in the follicle cells of egg chambers during the period of vitelline membrane deposition (Popodi et al. 1988).

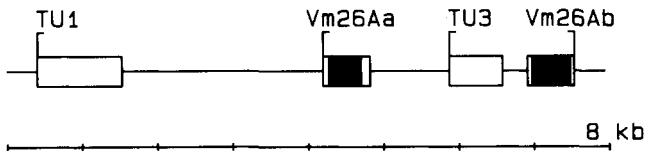


FIG. 31.2. Follicle cell gene cluster at 26A.

### *Vm26Aa*

#### **Product**

Vitelline membrane protein Sv17.5.

#### **Gene Organization and Expression**

Open reading frame, 141 amino acids; expected mRNA length, 629 bases, in agreement with the results of northern analysis. S1 mapping and sequence features were used to define the 5' end. The 3' end was obtained from a cDNA sequence. There are no introns (*Vm26Aa* Sequence) (Burke et al. 1987).

*Vm26Aa*

--&gt;-80

-122	GGAGAGCTATAAAAGATGGGAGGCCAATTGAATGGTATTGGCATCAGTCACCTTTGGTAACTACCAGCAGCCCAACCAGCTCCCATCCGC	-33
	-----	
-32	CTCCAGCTCAATCTTCAACCACCAACAACCAAGATGAAATCCTTCGTGTGCATCGCTCTGGTCGCCTTCGCCGCCCGCTCTGGCTTCG	57
	MetLysSerPheValCysIleAlaLeuValAlaPheAlaAlaAlaAlaLeuAlaSer	(19)
58	CCCACCAACGTGGCTTCGGCCACCGGCTCCACTGGCTCCTCGGTGACCACCCAGGACGGAGAGCTGGAGGGAGTGACCGGACAGGGATTCC	147
	ProThrAsnValAlaSerAlaThrGlySerThrGlySerSerValThrThrGlnAspGlyGluLeuGluGlyValThrGlyGlnGlyPhe	(49)
148	GGTGACCTGACCCGCTCCGTAAGTCTGCCTACGGCGGCAGCTCCGGCGGCTATGGCGGCTCCAGCATCCCAGCTCCTCCCTGCCCAAG	237
	GlyAspLeuThrArgLeuArgLysSerAlaTyrGlyGlySerSerGlyGlyTyrGlyGlySerSerIleProAlaProProCysProLys	(79)
238	AACTACCTGTTTCAGCTGCCAGCCCAACCTTGCCCCGGTCCATGCAGCGCTCCAGCTCCCAGCTACGGATCCGCCGGCGCTACTCCTCC	327
	AsnTyrLeuPheSerCysGlnProAsnLeuAlaProValProCysSerAlaProAlaProSerTyrGlySerAlaGlyAlaTyrSerSer	(109)
328	CCGGTGGCCACCTACGTCGCCCAACTACGGCGTGCCCCAGCACCAGCAGCAGCTGTACAGCGCCTACGTGCCCAAGACCTATGGCTAC	417
	ProValAlaThrTyrValAlaProAsnTyrGlyValProGlnHisGlnGlnGlnLeuTyrSerAlaTyrValProGlnThrTyrGlyTyr	(139)
418	CAGTACTAAGCACCTGCTCCGACTGCGACTCGATCATCGCCCAAGACCACGAACCGACTGCCGAGAAACATAAGCTTTGATGGATTTGA	507
	GlnTyrEnd	(141)
508	CAAAAAATATACCAAAAAATATGTACTGCAATTAATCACT	548
	(A) <sub>n</sub>	

*Vm26Aa* SEQUENCE. Accession, M18280 (DROVITA). The vertical bars at Val-23 and Ser-25 mark potential signal peptide cleavage sites.

*Developmental Pattern and Promoter*

High levels of RNA are evident in follicle cells between stages 8 and 11 (Burke et al. 1987). A 170-bp segment upstream of the site of transcription initiation controls developmental specificity (Jin and Petri, personal communication).

*Vm26Ab***Product**

Vitelline membrane protein Sv23 (Popodi et al. 1988). The female sterile mutation *fs(2)QJ42* is rescued by transformation with *Vm26Ab* DNA (Savant and Waring 1989).

**Gene Organization and Expression**

Open reading frame, 168 amino acids; expected mRNA length, ca. 625 bases, in agreement with the results of northern analysis. Primer extension was used

*Vm26Ab*

		-207	GTCGACTGGCGGTTGCAGGTG	-187
-186	GTCAGCAGATTTCGAGCCGGGGTCTCCATTTCATTTTTTCGGAACGCTGTCGTTCTACTCCGTCAGTGCATCAGCGTTTTCCGAG			-97
	-----			
		-->-61		
-96	TGGGCTATAAAGTGGATTGGCTGGGAGGCTACAATCAACAGTCAGCCTTCGTTTCGTCACCTTCAGCAGCAAGTAGAGACAGCTCAAGAACC			-7
	-----			
-6	ATCCGCAATGGCATTCAACTTTGGTCACCTCCTCATCGCCGGCCTCGTGCCCTGTCCGCGTGCTTCGGAGACCATCCAGTGCAGCC			83
	MetAlaPheAsnPheGlyHisLeuLeuIleAlaGlyLeuValAlaLeuSerAlaValSerSerGluThrIleGlnLeuGlnPr			(28)
84	CACTCAGGGCATCCTCATCCCCGCCCGCTGGCCGAGAACATCCGTGTGTCGCGTCCGCCCTACGGAGGATACGGCGCTGCCCCAGCCGC			173
	oThrGlnGlyIleLeuIleProAlaProLeuAlaGluAsnIleArgValSerArgAlaAlaTyrGlyGlyTyrGlyAlaAlaProAlaAl			(58)
174	CCCATCGTACTCCGCCAGCCGCTCCCGCTGCCAGGCTACTCTGCTCCCGCTGCCCCAGCCTACTCCGACCCGCTGCTCCCGCCTA			263
	aProSerTyrSerAlaProAlaAlaProAlaAlaGlnAlaTyrSerAlaProAlaAlaProAlaTyrSerAlaProAlaAlaProAlaTy			(88)
264	CTCCGACCCGCTGCTCCTGCTACTCTGCTCCCGCTGCCAGCCTACTCTGCCAGCCGACCCAGCTTACTCCGACCCGCTCCAT			353
	rSerAlaProAlaAlaProAlaTyrSerAlaProAlaAlaProAlaTyrSerAlaProAlaAlaProAlaTyrSerAlaProAlaSerIl			(118)
354	TCCGTCGCGCCGTGCCCAAGAACTACCTGTTACAGTGCCAGCCCTCCCTGCAGCCCGTGCCCTGTCCGCCAGCTCAGTCTACGG			443
	eProSerProProCysProLysAsnTyrLeuPheSerCysGlnProSerLeuGlnProValProLeuSerAlaProAlaGlnSerTyrGl			(148)
BamHI	-----			
444	ATCCGCCGGTGCCTACTCCCAGTACGTGCCCCAGTACGCCGTGCCCTTCGTCGCCGAACCTTAAGGATCGAACCGAATCTGACTTGACAT			533
	ySerAlaGlyAlaTyrSerGlnTyrValProGlnTyrAlaValProPheValArgGluLeuEnd			(168)
534	CTGAACCTAAGAATAAAGTAATGCTTTCATAAAA	567		
	-----	(A) <sub>n</sub>		

*Vm26Ab* SEQUENCE. -96 to 567, from Popodi et al. (1988); the segment from -207 to -97 was kindly supplied by Gail L. Waring. The vertical bar at Thr-23 marks a potential signal peptide cleavage site.

to define the 5' end, and S1 mapping gave the approximate position of the 3' end. There are no introns (*Vm26Ab* Sequence) (Popodi et al. 1988).

*Developmental Pattern and Promoter*

High levels of RNA are present in follicle cells of stage 8-10 egg chambers. *Vm26Ab* RNA is approximately half as abundant as *Vm26Aa* RNA, but it is 20-40 times more abundant than TU1 or TU3 transcripts (Popodi et al. 1988). One hundred and forty-seven bp upstream of the transcription initiation site seem sufficient for correct gene expression (Savant and Waring 1989).

*Vm32E*

**Product**

Vitelline membrane protein of approximately 12 kD (Gigliotti et al. 1989).

*Vm32E*

-->-28

```

-94 AAAAGTGCCGAGTTTTGTTATTAAAGTCAACGCATGAATGCTATAAAGATGCCACCATTGGTCACTAAATCGACAGTGTAATCATTAGT -5
      -----
-4  TCATCATGCAGATCGTTGCTCTCACCCCTCGTTGCGTTTGTGGCCATTGCCGGTGCCCTCTGCCCGTATGCAGCTCCAGCTCCAGCTTATT 85
      MetGlnIleValAlaLeuThrLeuValAlaPheValAlaIleAlaGlyAlaSerCysProTyrAlaAlaProAlaProAlaTyrS (29)
86  CAGCGCCCGCTGCTTCTTCTGGCTACCCGGCTCCACCATGCCCCACCACTACCTGTTCACTGCCAGCCCAATTTGGCCCCAGCTCCTT 175
      erAlaProAlaAlaSerSerGlyTyrProAlaProProCysProThrAsnTyrLeuPheSerCysGlnProAsnLeuAlaProAlaProC (59)
176 GTGCCCAGGAGGCCCCAGCCTATGGATCCGCCGGCGCTACACAGAACAGGTGCCCACTACGTGGACAAGTCCCAACCGAGAGCAGTTGC 265
      ysAlaGlnGluAlaProAlaTyrGlySerAlaGlyAlaTyrThrGluGlnValProThrThrTrpThrSerProAsnArgGluGlnLeuG (89)
266 AGCAATTTACCAGCGCATTGGAATGGCGGCTTTGATGGAGGAACTGCCGGCTTGGGCCAAGGAATCCAGGGTCAACAGTACTAGTGGC 355
      TnGlnPheHisGlnArgIleGlyMetAlaAlaLeuMetGluGluLeuArgGlyLeuGlyGlnGlyIleGlnGlyGlnGlnTyrEnd (116)
356 AAAAAAAATTCATGTGAAGAATGTTTTCGAATTAATCCGCTATGCTTTAATGGACTTTATACTATGGAACAAAAAAATTTGGGAT 445
      ----- | (A)n
446 TGGAGATAAGGAAACTGGTAAAAAAATAGGAGTTAAACTTATTTGTTGTTTTGTGCCTCTGGCTCCGATTCCTTTTCAAAGCCATA 535
536 AAGAACATTGCTCGTCTGTATTATATATTCTAAC 570

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*Vm32E* SEQUENCE. Strain, *Oregon R*. Accession, M27647 (DROVMP).

### Gene Organization and Expression

Open reading frame, 116 amino acids; expected mRNA length, 434 bases, in agreement with a 0.46-kb RNA detected by northern analysis. Primer extension and S1 mapping were used to define the 5' end. The 3' end was obtained from the sequence of several cDNA clones. There are no introns (*Vm32E* Sequence) (Gigliotti et al. 1989).

### Developmental Pattern

Transcription seems to be restricted to follicle cells in stage 10 egg chambers (Gigliotti et al. 1989).

### *Vm34C*

#### Product

Vitelline membrane protein of approximately 10–11 kD (Mindrinis et al. 1985).

### Gene Organization and Expression

Open reading frame, 119 amino acids. Northern analysis revealed an RNA of approximately 0.6 kb. Primer extension was used to define the 5' end.

*Vm34C*

```

-523   GTTGCTAGGCAAACATAAACGAATATTTTTTCCAATGACCCGCATATTCGGCACGCGATTACAAATCTTGTGGAAAATTAAG -440
-439   CTCATTGAACATAAATAATTTTTAGATATAAATAATTTATACACATATAATATTTATTTAATACATTTATTCGAATGTTTCAGTAAAA -350
-349   TAATGTAGCTCAATGCAAAGCTAAGTACATCAATCTTGGTGCTTCAACAATTTTTAGTCCGTTACTTCATTAATTTACATTTTTGGC -260
-259   ATGCGACAATTGTTTACTCAACAAGTTCAGTGGCCAAAAAAGTAGAGGAAATGTTTGTCTTTTCACTTTCTGTTGGCCGTGCAAAA -170
-169   AAAGCGCCACTCACGTCGACTTCGAGGGGTCGTTGGGTAAGTAAAACTGGTGCAGTCTGCATCTGCACCTTTTATGGCATTGCATC -80
-79   GGGTATATAAACCTCAAGTGTGGAAGCCAGAAGCATCGAGTCTGCTACCAACAGTCTAAGAAATCATCAACCAATCAACATGAAGTGCA 10
      -----
      MetLysCysI (4)
11   TCGCCATCGTCTCCACCATCTGCCTGCTGGCCGCTTTCGTTGCCCGCATAAAGAGGATAAGATGCTCGGCTCCCTCACGGTGGTGGCT 100
      leAlaIleValSerThrIleCysLeuLeuAlaAlaPheValAlaAlaAspLysGluAspLysMetLeuGlySerSerTyrGlyGlyGlyT (34)
      |
101  ACGGCAAGCCCGCCGCTGCTCCGGCTCCATCCTACTCCGCTCCGGCTGCCGCTTCCCCAGGCTACGCGCCACAGCTGCTCCATCCTACG 190
      yrGlyLysProAlaAlaAlaProAlaProSerTyrSerAlaProAlaAlaAlaSerProGlyLeuArgAlaProAlaAlaProSerTyrA (64)
191  CCGCGCTCCGGTCTCGATCCGGCTCCTCCTTGCCCCAAGAAGTACCTGTTAGCTGCCAGCCCAACCTGGCCCCAGTGCCATGCAGCG 280
      laAlaAlaProValSerIleProAlaProProCysProLysAsnTyrLeuPheSerCysGlnProAsnLeuAlaProValProCysSerA (94)
      |
      BamHI
281  CCCAGCTCCAGCTATGGATCCGCGGTGCTACTCGCAGTACGCCCGTCTACGCTCCTCAGCCCATCCAGTGGTAGGATGATCCAC 370
      laProAlaProSerTyrGlySerAlaGlyAlaTyrSerGlnTyrAlaProValTyrAlaProGlnProIleGlnTrpEnd (119)
371  AGACTTCACTAACCCCTGATCAACGACAAAAGCAATGCAATAAAAAAATAAAAGAAAAATATTTATGTTTAATCATAAAAAATTCATATGT 460
      -----
461  TTCAATTTGGGGATAATAGCGTGCCTAATAGCTGAACTAAAAACATTAATAATTAATGATCGAAGCTCGTCGTTATTCAAAGATTTTG 550
551  AAAAAAATATTGTTTTATTGATTCATACTTAATTCATAATTTTTAGAAATTAACAACCTTTTATAGATAATCTGGTAAGTTCCTCTTT 640
641  AATTGTCGAC 650
    
```

*Vm34C* SEQUENCE. Kindly supplied by W. H. Petri and L. J. Scherer and from Mindrinos et al. (1985). The vertical bar at Ala-19 marks a potential signal peptide cleavage site. Also indicated are a *Bam*HI site present near the 3' end of all *Vm* genes and a potential poly(A) signal.

The 3' end was not determined. There are no introns (*Vm34C* Sequence) (W. H. Petri and L. J. Scherer, personal communication; Mindrinos et al. 1985).

*Developmental Pattern*

High levels of RNA are present in follicle cells of stage 8–10 egg chambers (Mindrinos et al. 1985).



***Fcp3C***  
(*Follicle cell protein at 3C*)

**Product**

Unknown. The predicted amino-acid composition is relatively rich in Ser and Thr (11% each). The sequence shows no obvious similarity to other proteins (Burke et al. 1987).

**Gene Organization and Expression**

Open reading frame, 217 amino acids; expected mRNA length, 770/786 bases: two sites, 16 bp apart, were indicated to be the likely position of the 5' end by S1

*Fcp3C*

-211	AAAAGTAATATTAGCTAAAGAACACATTTTCATATCGTATATATTTTCATATATCAGGCGCCTTTAAAAATCCCTGCTGCTGCTGACACTC	-122
	<div style="display: flex; justify-content: space-around; align-items: center;"> <span>!-111</span> <span>!-95</span> </div> <div style="text-align: center; margin-top: 5px;"> <span style="border-bottom: 1px dashed black; width: 100px; display: inline-block;"></span> </div>	
-121	TCTGCTAGCCATCCATTGGAGAGCCATCCAGATAGTCTACAAGAAGCCGCTCTATGGCAATAGCAACATCATCAAGGACAAGCGTATAA	-32
-31	AGACGAAGCCCGTCAAACCTGAAACCAGCACCATGAGCAGCCTGGTGTGCAAGTAGCAGCACACAGCCGAAGAGGATTGGCCACGG	58
	MetSerSerThrGlyValAlaSerSerSerThrThrAlaGluGluAspTrpProThrA	(20)
59	CCGTTGAGTTTGTGATTATGACAACGCCCGAAGCGAATTGGAAGCCAGCACGGAAACCATTGGTAACAATGGCACCACCGAAACGACCG	148
	laValGluPheValIleMetThrThrProAlaSerGluLeuGluAlaSerThrGluThrIleGlyAsnAsnGlyThrThrGluThrThrV	(50)
149	TTGGCAGGACCCATCATCGGATCGTCGGAAGGATCCACACGATCGATGGAGCCAACCACCGGAGTCCGCTGATGAGCACAAACCCAT	238
	aGlyGluAlaProIleIleGlySerSerGluGlySerThrArgSerMetGluProThrThrAlaSerProLeuMetSerThrAsnProS	(80)
239	CGAGCAGCAGCAGTCTGGTTAGCACCATTCCCTTGCCACCAGCAGCGGACTACATGCGCAGGATAATCAGCCAGTCCGCTGCACATGCG	328
	erSerSerSerSerLeuValSerThrIleProLeuProProThrAlaGlyLeuHisAlaGlnAspAsnGlnProValProCysThrCysG	(110)
329	GCGTCTCTCTCTCGCAAAATCCCAAATGGCTTGCCGACAAAGCCACTTATCCACCAGGAATTGGATCATATGTTCCCTGCAATGCCA	418
	lyValPheLeuSerSerGlnIleProAsnGlyLeuProThrLysProLeuIleHisGlnGluLeuAspHisMetPheProCysAsnAlaI	(140)
419	TCGGTCGCAAGCAGTGTCAAACCAATGCCTAGAGACGGTGAGTACTGGGAAACGAGGAGGAAAACATCAGGAGAAGCGCTCTATAACT	508
	leGlyArgLysGlnCysGlnThrLysCysLeuGluThr	(152)
509	CACCAATTCGTCCTTTTAGATCGTACAACATCTGCCGAATCCGCAATATAGTATGCTCCGCACTGGGTACGATTGTCACAAGGAA	598
	IleValGlnHisLeuProAsnSerAlaAsnIleValCysSerAlaLeuGlyHisAspCysHisLysGlu	(182)
599	CGGGCCTATTTGTCATCAAGAACTGTCACAATCAATGGGTTAATACCAATCTGCAGGCGGGCAGGGAGTACTGTTGTCCTCGGCTTC	688
	ArgAlaTyrLeuPheIleLysAsnCysHisAsnGlnTrpValAsnThrAsnLeuGlnAlaGlyArgGluTyrCysCysArgLeuGlyPhe	(212)
689	CCTACCCTGGCCATTGATGGTTAAGCACTGTGCAAATGAAATAAATATATGATATGAC	747
	ProThrValAlaHisEnd <span style="margin-left: 100px;">-----</span> [(A) <sub>n</sub>	(217)

*Fcp3C* SEQUENCE. Accession, M18281 (DROVITB).

mapping and cDNA sequencing. The 3' end was obtained from a cDNA sequence. There is one intron after the Thr-152 codon (*Fcp3C* Sequence) (Burke et al. 1987).

### *Developmental Pattern*

Transcription occurs during vitellogenesis and is restricted to the follicle cells. RNA is first detectable in stage 9 egg chambers, it reaches a maximum during stages 10 and 11, and it is absent from stage 12 chambers (Burke et al. 1987).

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# 32

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## *yellow: y*

**Chromosomal Location:**

X, 1B1

**Map Position:**

1-0.0

**Product**

Unknown. It plays a role in the accumulation or deposition of melanins in larval and adult cuticles.

***Structure***

Several features of the *y* product are suggested by the predicted amino-acid sequence (*y* Sequence). A signal peptide-like segment is an indication that the protein is either secreted or incorporated into membrane. Two potential N-glycosylation sites (Asn-X-Thr/Ser) are present, occurring at Asn-144 and Asn-215. The widespread occurrence of Pro and Gly residues suggests that extensive regions of  $\alpha$ -helix or  $\beta$ -pleated sheet do not occur (Geyer et al. 1986).

***Mutant Phenotypes***

Mutations are classified into two groups. Type 1 alleles are probably amorphs; they show a uniform absence of melanin (yellow color) in all structures. Type 2 alleles show the mutant phenotype in some structures (body cuticles, wing blades) and the wild-type appearance in others (denticles, bristles, sex combs).

**Gene Organization and Expression**

Open reading frame, 541 amino acids; expected mRNA length, 1,985 bases. Primer extension and the sequences of two cDNA clones were used to define the 5' end. The 3' end was obtained from a cDNA sequence. There is an intron in the Gly-80 codon (*y* Sequence) (Geyer et al. 1986).

y

-3042 GTCGACTATTAATGATTATCGCCCGATTACCACATTGAGTGGTTAAAATAGCCATAAAAATGCAACTGACGATGGCTTAAGATAAAT -2953  
 -2952 ACGTCGCAGAGTCACTCATAAATTCGAACGCAGCCCGCTGATTACCTACCCCTCTAAACGATTCATAGTATATGTACGAGTATATCCA -2863  
 -2862 CTAAGCTTTTTCGAGCACTGATTTTTTCGCTTGCACGAGACAAGTGCACCACCGCAATTGCAGGCAAATATGTCTGAGGTAATGATTCC -2773  
 -2772 GTTTCGTGAAGATTACACAGAAATCAAATACGACAACCTTTATTTCAGTAAGCAAACAAAGCCCTTTGTTGGCATCTAATATCCACTT -2683  
 -2682 ATGGTTGCGATTCGGGAGCTACAATCGGTTTTGGTTTAGTATATCTAGCGAGTTCCTTGGCGACATTTAAAATTTACAAAATAAGGTTTC -2593  
 -2592 TCTATTCAATCGGAACAGTGGAAATGACTATTTTTATTTATATTAATGAACCTATTTTTAATTTGGCTTAAGTTACTAAGGGGTACTAAT -2503  
 -2502 AGTTTGAGCGCAGTGCATGTGCATGGGGACATGTGAATGTGTGTAACGGGAAGTGATCGCGCCTCCGGAATTTGGCCATGCCAAATA -2413  
 -2412 ATCCGAGCTCGAAAGGAGGGGACCCGGCGTCAAGGCCATGGACATTGAACTTGAAAAAACAACACAAAATATATAACACAAAACG -2323  
 -2322 GAAAATGCTGTGTACCGCTTATGTTAGAGAAGTTGAGCAACGGGTTTTTCGTTTTGCAGTCACGATGGATTTCCAAATTAGTGTAGGAGG -2233  
 -2232 GGGGAGGGGAGGGAGAGATAATGTCCAGGCTGCCATAAGTGGGAATAAGGAAAAAATAACATGAACACGGGTGGGCAATGTGATG -2143  
 -2142 CGGTATTCGGCTTTGCTTTCCGCCCAAGTTGAAGTGATCCTGTGTGTAATAATGTGCAATGTTGCGGGTCGGTTGCATAAAGCCTGGTC -2053  
 -2052 AATTATGGCCAAAGAGATCTGATTTGTGGAAGCTTTTTTGACCACTTAGCGCGCTCCGCTGATGTTGTTTTGTTTGTGCTGGGGCAGA -1963  
 -1962 AAACCTGTTTCAATTATTGGGAAAAGTGCGTATAAATCATTGCCGCAAGCTCTGAAAAGCGAAAAAGAAAACAGTAACCAACACAGACAA -1873  
 -1872 ACGCAGCATTCGCCACACAATTAAGCAAAAACCTGAAAACAAGTCAATTCGAAAAAATATAGGTTCAACGGCTGCAGCGATCGCATCA -1783  
 -1782 TTAGTTGCGTTTTTAGTAAATACACCATTTTCATTACACAACACACAATTAATTAATAAACTGTACTGTTATTCAAGTGTGCTTTT -1693  
 -1692 TAATAAGCCTGCCGATCGCAATAAATTCGAGCAGCATTGCCGGTAATTTGTGCAACATATTTTTTCGATTGCCACACCGGTTTGTTTAT -1603  
 -1602 TTTTTCTGTTGGTGAATGATTTAGAATGCGGGCAAGGGATCAAGTTGAACCCTTCTAAGAAAAAATAGACATTGCATAAATGATATAG -1513  
 -1512 AGTCCAAAACTACACCAATTCAAATAGCAGTAATGGTTACATTAGCTTTGAAATGTTTTTAGACATCCGAAGAAATAAGATTAAATTTA -1423  
 -1422 AACGGCATTCTTAATTTGATTTTTAATATTTTGAGAGGTTTTCCATTATTAAGTGTAGATTATTGAGGATTAATGCAATACCACCTTTA -1333  
 -1332 CCTGCGGAGGTCGTAACGATTTTTTACCATTGTCATGTTTATTATGCGTGTGCTGGTTGTTACTTTACTTTAAGTTTTGCAATTTT -1243  
 -1242 TTCTTTAGCAAGCAGGTGCATTTGGGCCAAGAGATATATGCGATCGCTTTCGGTTCGAATTTTAAACATTTACTTGGCGGATGGTCATT -1153  
 -1152 AGAGCATTACCCACTTAGGGCACCCCAACATCCAGTTGATTTTCAGGGACCACAATTTTTAAATAACAGCTAGTGAATTACCTAAAA -1063  
 -1062 GCGCTTTCGTTCTTTTTGAAATTTTATGTAACACTCAATATATTTATGTATATGTATGCTCAAATCACCTGCCAATAACTAGCGGAA -973  
 -972 ACCAAATATTTGACCCTCAGTGAATTTGGAATCATCGGTGACGCCAATCGAAATCCAATCCTAAGCAATTGAAACGAGCAGAGTTCCA -883  
 -882 ATTTAATAGTATACAAGGAAACACCTGCTTTAAATACTCTACATAGTACACGTTATAATAACGATTTATTGATATTTCTGGATTTTTGT  
 -792 CTGCATGATTTTCATATAATTTGATTTGATTTTTTAAATGAATTGAACTAAAAAATCATATTAGAACATTTTTGCGAGTCGCCGATAAA -703

||gypsy=2 .

(continued)

-702	GATGAACACAGTTCTCAGAACACAACACTGTCATGTATTAAGCTTTCAGATTTTCAGAAATTTGGAGAGCAATGCATTCTATGCACGAGCCT	-613
-612	CCTGGCCTTACAATTTACTTGTTTGAAATTAGATCGTCAAAATAAAGTCCCTAAAATAAATAAATAGTAGTCACAACCTTTAAAATAGGTC	-523
-522	TTAATCTTTTAGGGTACCGAAAAGTATTTTCGGCACAAATCAGCGCAGTTTTAAATGTCGATGAAGGCCAAAAATCATACCAAAACCCAGCG	-433
-432	AAAGGTGATGTCTGACTCATTAAATTTGGGGATTTCGAGTGTATTTATTAACATGCGTGAAAATCAATCATGGAAGACAAAACGCAAAGT	-343
-342	TGGCCGATCTATGGGAACAGCATAAGCCACCTGATTACCCGAACACTGAACCACCCGAATCACTAAAACCACCGAAGTTGGCGCGCGCCT	-253
		-->-170
-252	TCGTTTTCAATTTTCATTGGCCTGTCTTCGTCTTCGGAGAAAAAACCTTCATATAAAACGCGGCCGACATATTATGGCCACCGAGTCGTTA	-163
	-----	-----
-162	CCGCGCCACGGTCCACAGAAGAGGATTAATAAATATCACACAGCCGAAGGCTAGAGAAGAACCCCTATAGCTGAACATATATAAACAA	-73
		P=76d28
-72	ATATATTTTTTTTTATTGCCAACACACTTTGGCTTAAGTGTTAAGAGTGATTGTCAGCTTAGAGCTAAGTCAATGTTCCAGGACAAAAGG	17
		MetPheGlnAspLysGI (6)
18	GTGGATCCTTGACCCCTGATCACCTTGGTGACGCCGTCTGGGGTCTTACAACCTCAGGAGCGATATAGTTGGAGCCAGCTGGACTT	107
	yTrpIleLeuValThrLeuIleThrLeuValThrProSerTrpAlaAlaTyrLysLeuGlnGluArgTyrSerTrpSerGlnLeuAspPh	(36)
108	TGCTTTCCGAAATACCCGACTAAAGGACCAAGCTCTGGCTAGTGGAGATTATATCCGCAAAATGCTCTACCTGTTGGAGTCGAACACTT	197
	eAlaPheProAsnThrArgLeuLysAspGlnAlaLeuAlaSerGlyAspTyrIleProGlnAsnAlaLeuProValGlyValGluHisPh	(66)
198	TGGCAATCGTTATTTCGCTACTGTTCCCGCTGGCGTGATGGTAAGTGAAGTTAAATATGAAGCCCTTGGGGAGATCGTAAATGGGACA	287
	eGlyAsnArgLeuPheValThrValProArgTrpArgAspG	(80)
288	TTCTTACTTAGGCATCAGAGATATCTGATTGAGTGGTTGACAGTTTTATATGGCTTGTTTGACATGATGAAAAACACAAAATTCATTT	377
378	AGTTTAGGTATTCGAAATAAGAGCTTGTATTTATTTAGAATTTGGAGAACATTTTTTGTCTTCTACCCCTCTTAGAAAATAATATT	467
468	GTTTTGTACAATTTAATTTAACTAGTACAGACGAAAAATGTATTTTTTATTTGTATGCCTTTTACCATTTTTGGCAGAGGAATAAAT	557
558	ATGACAATATATTTGAGAGCACCCCTCATGTAAAGTTTTAGCGTGGCGACCTCTCATAAATCCGGTTGGTACCTGCGCGTTATTTTAAAC	647
648	ATTTTAAACAATTAACCGTTGTAATAACGAAGCCAATAGCATGGCATTGGCTTTTACTGTATTAATTTGTATTATATTACCATCCGAAT	737
738	TGTAAGACTTCTTCAGGGCCGCCACATAGAAATGGAATCCAATCACAAACAATAACTTATGGCATTAGCTATTAACCGACGATTAGCT	827
828	GTCAGTTCAACAAATGTAAGAGTGGCGAAATGTTTAAATGCGAAGGCATTGTTCTGTGACTCACGTTTTATTATTAATCACACAAATGAT	917
918	TTTGCTTCAAAATATTTGGCTTACACAATAACAAAATTTTTATGAAATAGTTGAAACACAAAACCTAGGAAATTTAAAAAGCAATGAA	1007
1008	ACTAAAAACCAATTTGTAAGATTATATGATGCGCATACAAATACTTCAGTACGCTAGGAATGCTTTTCGATGATTGATTAGTTTTTAT	1097
1098	GCATGGCTTACAATTTGGTATTTACACAGAAAAACACGGCTGTATCGATTCAAAATGCGATGTTAATAAATTTGTACATATGTTCTTAAG	1187
1188	CAGTCCGAAACCCCAAACTTCTGACTAAACTTAAAAAACACGCTCTTCAAGGATATCTTAATGTCACTTATAATGTAATGGTATTGTA	1277
1278	TTATTAAGTATATCAAAATATTTGGCCAGCCTTGGAGTCTTTTTAAAAAGATATCGACTGACTACCTCCAGTCAATGAAATAATAG	1367
1368	CCCAGAAGGCCAATCGGCAAAAAATAACCCCAAGTTACGGCAACAAAACATAGTGAAGTTGTGGCAAAGTGAACATTTAAAGGCA	1457

1458 TGCTTCAATGGCCATCGAAGCAAATCAATTAGTCAAAGCAAATCGGTAGTGGCAACAACAGGCTACAGAATACCTATAAGTGACAGTTAT 1547  
 1548 GGGGTATGATTAATTATAAATATTATCATTGACCACCAATGCTGGGCTCAATTGAAAAAATACTTCTATGAAGATTGAGTAAATAAATT 1637  
 1638 TTGATTTAAAAAGCCCATGGTTATCGCGACAACACTAGCTACGGGACAAGATTACTGTTTAAAAATCAAGTGTGAAATATCAAAATCAAAAT 1727  
 1728 CGGATTCCGATCGGGAAGTTGTATCCGATTCTGAAACTAAAACACAGAATGGCAACATTTCCGATATCGACTCAGCTCACGTATTTC 1817  
 1818 TACAGATTCATTAGGCCACCAGCCATTGAATAATATACCCAGTCAATTGAGCTACTCGATAGTTGATCAACTTAGCTTTGTCAACGAG 1907  
 1908 TGAACGCATAAACTACTACATCAACGATATTGCGGCCCATTTCCAAGCTAAAAGTTCATCTTAATTACAAAATAGATTAGAAAAAATATC 1997  
 1998 TGAATGAAAAAATGTTGAGACATATTTCTTTGGAAAAGGAGAACCCTCAAGACAGTCGAAAAAATGTTTACAATGAAAATGTTGAAAAT 2087  
 2088 CATGAAGCAGATAAATCTGTCAAGTTCGAGGTTTTAGGACTGAAAAGAGCACATGTCAAAATATAAATTTGTTCAAATACTTTATATTTGA 2177  
 2178 CTGAATTAGATTGTTATTTTAAAAGTTATGAATTAATAAAGATTGAAAGGTGCATTATGCTCAAATGTATATTTATCGCAACCCCGGT 2267  
 2268 TACTTTGTAAGCAAAAACGCTGGTTTGATTTTTAAGAAGTGGGTCGTAATCGATAAAGCTATATTTCTGGTCGTGCGAGCTC 2357  
 2358 ACTCGCCTGCTATAAAACATTAAAAGTCCCAGAAACAATAAATGTCTTTAAATCAATTAACGAAGAAATAAGAAAGGAAAAGAACTG 2447  
 2448 GAGCGGAAATCGGTCGAACTACTGCAATGGCCACATATACATTTAACAGCGATATATGGTATACATATTGATAATGATGTCAGACGCAA 2537  
 2538 TTGCTTCAGACGGCTAATGACATCGCAAAATGCACGCAACTGCAATAGTGC AATATGACTGAAGTACATATAGCCGGGGATCTTTTA 2627  
 2628 ACAATAAACTCCAGTAGATGTACAAGCAGAAAAAGAGCCATTAGCACGGCAGTTACCATTGCTTATGATTCTTGTGTCCAAAATAAT 2717  
 2718 GACAAATAGGTATATAAATAATTAATGCCAACATAAGCGATTCTAATTTACCTTTACATCTGTATGCATTACATATTATCCAGAAAA 2807  
 2808 CAGACAGCGATAACTTGCAACATTGCTTAGTATAATAATCAAAGAAGGAATTTAGCGCAAAATCCAGTTAATTAATATTCAAAACAA 2897  
 2898 ACTTTATTTAGTGCCTCAATAATAGTTTGGCCCTGCTAATCTCCTATTTATTTTTAGGGATTCCGGCCACTCTGACCTATATAACA 2987  
 yIleProAlaThrLeuThrTyrIleAsnM (90)  
 2988 TGGACCCGAGTTTGACGGGTTACCCGGAGCTAATCCGATTCAGATTGGCGCTCAAATACAGCTGGAGATTGCGCCAACAGTATTACCA 3077  
 etAspArgSerLeuThrGlySerProGluLeuIleProTyrProAspTrpArgSerAsnThrAlaGlyAspCysAlaAsnSerIleThrT (120)  
 3078 CTGCCTACCGCATTAAAGTGGATGAGTGTGGTCGGCTGTGGGTTTTGGACACTGGAACCGTGGGCATCGGCAATACCACCCTAATCCGT 3167  
 hrAlaTyrArgIleLysValAspGluCysGlyArgLeuTrpValLeuAspThrGlyThrValGlyIleGlyAsnThrThrThrAsnProC (150)  
 3168 GCCCCTATGCGGTAATGTCTTTGACTTGACCACGGATACGCGAATTCGGAGATACGAGTACCTGGCGTGGACACAAATCCAAATACTT 3257  
 ysProTyrAlaValAsnValPheAspLeuThrThrAspThrArgIleArgArgTyrGluLeuProGlyValAspThrAsnProAsnThrP (180)  
 3258 TCATAGCTAACATTGCCGTGGATATAGGCAAAAATTCGCATGATGCATATGCCTATTTTCCGATGAATGGGATACGGCTTGATTGCTT 3347  
 heIleAlaAsnIleAlaValAspIleGlyLysAsnCysAspAspAlaTyrAlaTyrPheAlaAspGluLeuGlyTyrGlyLeuIleAlaT (210)  
 3348 ACTCCTGGGAACGAAACAGTCTGGAGATTCTCGGCACATTCGATTTTTTCCCGATCCATTGAGGGGCGATTTC AATGTCGCTGGTA 3437  
 yrSerTrpGluLeuAsnLysSerTrpArgPheSerAlaHisSerTyrPhePheProAspProLeuArgGlyAspPheAsnValAlaGlyI (240)  
 3438 TTAACCTCCAATGGGGCAGGAGGGTATATTTGGTATGTCCTTTCCGCCATTCGATCGGATGGTTATCGTACCCTGTACTTTAGTCCGT 3527  
 leAsnPheGlnTrpGlyGluGlyIlePheGlyMetSerLeuSerProIleArgSerAspGlyTyrArgThrLeuTyrPheSerProL (270)

(continued)

3528	TAGCAAGTCATCGACAATTTGCCGTATCCACGAGGATTTTGGGGATGAAACCAGGACGGAAGATAGCTATCATGACTTTGTTGCCCTTAG euAlaSerHisArgGlnPheAlaValSerThrArgIleLeuArgAspGluThrArgThrGluAspSerTyrHisAspPheValAlaLeuA	361 (30)
3618	ATGAACGGGGTCCAAACTCCCATACCACTTCCAGTGTGATGAGCGATGATGGAATTGAGCTGTTCAATTTAATAGATCAAAATGCAGTGG spGluArgGlyProAsnSerHisThrThrSerArgValMetSerAspAspGlyIleGluLeuPheAsnLeuIleAspGlnAsnAlaValG	370 (33)
3708	GTTGCTGGCACTCATCAATGCCGTACTCACCGCAATTCATGGCATTGTGGATCGCGATGACGTTGGCTTAGTTTTCCGGCCGATGTGA lyCysTrpHisSerSerMetProTyrSerProGlnPheHisGlyIleValAspArgAspAspValGlyLeuValPheProAlaAspValL	379 (36)
3798	AAATTGATGAGAACAAAAACGTTTGGGTTCTATCCGATAGGATGCCGTTTTCTTGTGTCTGACTTGGATTATTCAGATACTAATTTCC ysIleAspGluAsnLysAsnValTrpValLeuSerAspArgMetProValPheLeuLeuSerAspLeuAspTyrSerAspThrAsnPheA	388 (39)
3888	GAATTTACACGGCTCCCTTGGCCACTTTAATTTGAGAATACTGTGTGTGATTTGAGGAATAACGCCTATGGCCGCAAAATACCGTTTCAA rgIleTyrThrAlaProLeuAlaThrLeuIleGluAsnThrValCysAspLeuArgAsnAsnAlaTyrGlyProProAsnThrValSerI	397 (42)
3978	TACCAAAAACAGCCGTTTTGCCAATGGGTCCACCGTTATATACGAAACAATATCGTCTGTCTGGCCACAGAAACCTCAGACCAGCTGGG leProLysGlnAlaValLeuProMetGlyProProLeuTyrThrLysGlnTyrArgProValLeuProGlnLysProGlnThrSerTrpA	406 (45)
4068	CTTCTCGCCGCTCCCTCCAAGTCGCACTTATTTGCCGCCAATTCAGGCAATGTAGTCTCCAGTATTAGTGCTCTACAATTTCTGTGG laSerSerProProProSerArgThrTyrLeuProAlaAsnSerGlyAsnValValSerSerIleSerValSerThrAsnSerValG	415 (48)
4158	GTCTCGCAGGAGTGGAGGTGCCAAAGCCCTATATTTCAACCAGCACACGGCATAAATTACGAGACAAGTGGTCCCATCTATTTCCCA lyProAlaGlyValGluValProLysAlaTyrIlePheAsnGlnHisAsnGlyIleAsnTyrGluThrSerGlyProHisLeuPheProT	424 (51)
4248	CCCATCAACCCGCCAACCGGGTGGCCAGGATGGTGGGTTAAAACTTATGTGAATGCCCGCAATCTGGGTGGTGGCATCATCAGCATC hrHisGlnProAlaGlnProGlyGlyGlnAspGlyGlyLeuLysThrTyrValAsnAlaArgGlnSerGlyTrpTrpHisHisGlnHisG	433 (54)
4338	AAGGTTAACATAATCCTACACACGGTACTTGGGTATATTCTCACACTCGATTGATGTAAGAATATTTAAAGACAACACATAGGGCA lnGlyEnd	442 (54)
4428	ACAGCGGTTAAAAAACACATGACGTATGAGCAAGTGGCAAATCAATCTTTATCTAGTTATGTTAAGCAAAAAATAACAATAAATCAA	451
4518	CTTTTTTTGAAGGTTAAGAGTTTACGCAATTTTCTTGAGCGGAAAAAGCGGAAAAAATGTAAGTATGC 4586	

|(A)<sub>n</sub>

*y* SEQUENCE. Strain, Canton S. Accession, X04427 (DROYELLOW) and X06481 (DROYELL5). An insertion of the transposable element *gypsy* following the A at -870 causes the mutation *y*<sup>2</sup>. Mutations *y*<sup>76d28</sup> and *y*<sup>1#7</sup> are both caused by insertion of a *P* element at the same site in the leader, but the insertions are in opposite orientations.

Most type 1 mutations occur in the transcribed region of the gene and likely result in non-functional *y* product (Chia et al. 1986; Geyer et al. 1986). Mutation *y*<sup>76d28</sup> is the result of a *P* element insertion in the leader (*y* Sequence). In this insertion, *P* is transcribed in the opposite orientation from *y* and the RNA produced is derived from the *y* promoter. Some of that RNA includes both *y* and *P* sequences and is not functional. In a small fraction of the RNA, however, splicing of most of the *P* sequences takes place through the use of cryptic splice signals in the *y* leader and the *P* element. This processed RNA codes for a small amount of normal *y* product that is responsible for a hypomorphic phenotype.

Mutations in *suppressor of sable* (*su(s)*) leads to increased accumulation of processed RNA and a more complete restoration of the normal phenotype (Geyer et al. 1991). This mechanism of suppression is similar to that observed in some *v* mutations.

*y* is less than 1 kb from *achaete*, centromere distal and transcribed toward the centromere (Fig. 1.1).

### *Developmental Pattern*

There are two broad peaks of expression, one beginning late in embryonic development (16–20 h) and lasting until the second larval instar, the other during the middle pupal stages, about 48 h after pupariation. Gene expression is detectable in epidermal structures in which pigmentation will develop (Parkhurst and Corces 1986; Martin et al. 1989).

### *Promoter*

Analysis of 5' deletions by germ line transformation identified 2,873 bp upstream of the transcription initiation site (up to -3,042) that are sufficient for full expression of *y*. The region between -3,042 and -2,038 controls expression in the wing blade and the adult abdominal cuticle. The region between -2,038 and -665 contains a *cis*-acting regulatory signal that also contributes to expression in the adult abdominal cuticle. Deletions that leave only 495 bp of the promoter region cause yellow body and wing blades but pigmented larval mouth parts and denticle belts and adult bristles and sex combs. The segment between -665 and 166 upstream of the transcription initiation site seems to control expression in larval mouth parts and denticle belts, and the segment between 166 and 95 appears to include elements that contribute to *y* expression in larval structures as well as elements that determine expression in the adult tarsal claws and sex combs. With 95 bp of the 5' region left, only bristles are pigmented normally (Geyer and Corces 1987; Martin et al. 1989).

The long intron contains enhancer-like sequences that seem to be responsible for increased transcript levels; they act in a position-independent manner (Geyer and Corces 1987; Martin et al. 1989).

Most type 2 mutations, including  $y^2$ , are associated with rearrangements in the 5' region of the gene; these seem likely to affect the regulation of *y* transcription (Chia et al. 1986; Geyer et al. 1986).

### **References**

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# 33

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## The Yolk Protein Gene Family: *Yp1, Yp2, Yp3*

### Chromosomal Location:

*Yp1* X, 8F-9B  
*Yp2* X, 8F-9B  
*Yp3* X, 12B-C

### Map Position:

1-30  
1-29.5  
1-44

### Products

Yolk proteins 1, 2 and 3 (YP1, YP2, YP3) of 46, 45 and 44 kD, respectively; also known as vitellogenins, when circulating in the hemolymph, and vitellins, when deposited in the oocyte.

### Structure

YP precursors contain signal peptides that are cleaved before secretion (Warren et al. 1979). Other post-translational modifications include the sulfation of Tyr residues (Tyr-172 in YP2) (Baeuerle and Huttner 1985; Baeuerle et al. 1988), glycosylation and phosphorylation (Minoo and Postlethwait 1985; Brennan and Mahowald 1982).

Judging from the predicted amino-acid sequence, the three yolk proteins have only moderate similarity (Fig. 33.1); sequence identities are 48–53% in pairwise comparisons over the whole lengths of the proteins and 73% if the comparisons are restricted to the C-terminal one-third (Hung and Wensink 1983; Garabedian et al. 1987; Yan et al. 1987).

The yolk proteins of higher dipterans seem to be related to the triacylglycerol lipase family of proteins rather than to the vitellogenins of vertebrates, nematodes and lower insects, which have a different common evolutionary origin (Terpstra and Geert 1988). Comparison to the yolk proteins of the Mediterranean fruit fly, *Ceratitis capitata*, shows that the most conserved region extends from residue 202 to 427 of YP1; in this segment there is 40% identity between the two species and 40% of the substitutions are conservative. In terms

```

1                               50                               100
Yp1 MNPMPRVLSLL ACLA.VAALA KP...NGRM DNSVQALKP SQWLSGSQLE AIPALDDFTI ERLENMNLER GAELLQQVYH LSQIHNVPEP NY..VPSGIQ
Yp2 MNPLRTLCLVM ACLLAVAMGN PQSGNRSRR SLSLDNVEQP SNWVNPREVE ELPNLKEVTL KKLQEMSMEE GATLLDKLYH LSQFNHVFKP DYTPEPSQIR
Yp3 MMSLRICLLA TCLL.VAAHA SK..... .DASNDR LKP TKWL TATELE WPSLNDITW ERLENQPLEQ GAKVIEKIYH VGQIKHDLTP SFVPSPSNVP
CON M---R-----CL--VA--- -----P --W-----E --P-L---T- --L----E- GA-----YH --Q--H---P ----PS---

101                             150                             200
Yp1 VYVYPGNGDK TVAPLNEMIQ RLKQKQNFGE DEVTIIVTGL PQTSETVKKA TRKLVQAYMQ RYNLQQQRQH GKNGNQDYQD QSNEQRKNQR TSSEEDY...
Yp2 GYIVGERGQK IEFNLNLVE KVKRQKQKFGD DEVTIFIQGL PETNTQVQKA TRKLVQAYQQ RYNLQP.... .YETT DYSNEEQSQR SSSEEQQTQR
Yp3 VWIISKNGQK VECKLNYYVE TAKAQPGFGE DEVTIVLTGL PKTSPAQKKA MRRLIQAYVQ KYNLQQLQ.. .KNAQEQQ QLKSSDYDY TSSEEAADQ.
CON -----G-K ---LN---- --K---FG- DEVTI---GL P-T---KA -R-L-QAY-Q -YNLQ----- -----SSEE-----

201                             250                             300
Yp1 .SEEVKNAKT QSGDIIVIDL GSKLNTYERY AMLDIEKTA KIGKWIVQMV NELDMPFDTI HLIQNVGAH VAGAAAQEFRT RLTGHKLRRV TGLDPSKIVA
Yp2 RKQNGEQDDT KTGDLIVIQI GNAIEDFEQY ATLNIERLGE IIGNRLVELT NTVNVPQEI HLIQSGPAAH VAGVAGRQFT RQTGHKLRLI TALDPTKIYG
Yp3 ...WKSAKA ASGDLIIIDL GSTLTNFKRY AMLDVLNTGA MIGQTLIDLT N.KGVPPQEI HLIQGISAH VAGAAGNKYT AQTGHKLRLI TGLDPAKVLK
CON ----- --GD-I-I-L G-----Y A-L-----G- -IG----- N---P---I HLIQ---AH VAG-A---T --TGHKLRR- T-LDP-K---

301                             350                             400
Yp1 KSKNTLTGLA RGDAEFVDAI HTSVYMGTP IRSGDVFDFP NGPAAGVPGA SNVVEAAMRA TRYFAESVRP GNERSFPAPV ANSLQQQYKQN DGFGRKRAYMG
Yp2 KPEERLTGLA RGDAFVDAI HTSAYMGTS QRLANVDFFP NGPSTGVPGA DNVEATMRA TRYFAESVRP GNERNFPSVA ASSYQEYKQN KGYGKRYMG
Yp3 KRPQILGGLS RGDAFVDAI HTSTFAMGTP IRCGDVDFFP NGPSTGVPGS ENVIEAARA TRYFAESVRP GSERNFPAPV ANSLKQYKEQ DGFGRKRAYMG
CON K----L-GL- RGDA-FVDAI HTS---MGT- -R---VDF-P NGP--GVPG- -NV-EA--RA TRYFAESVRP G-ER-FP-V- A-S---YK-- -G-GKR-YMG

401                             450
Yp1 IDTAHDELDG YILQVNPKSP FGRNAPAQKQ SSYHGVMHQAW NTNQDSKDYQ *..
Yp2 IATDFDLQGD YILQVNSKSP FGRSTPAQKQ TGYHQVHQPW RQSSSNQGSR RQ*
Yp3 LQIDYDLRGD YILEVNAKSP FGQRSPAHKQ AAYHGMHHAQ N*.....
CON ----DL-GD YIL-VN-KSP FG---PA-KQ --YH--H--- -----

```

FIG. 33.1. Amino-acid sequence comparison of the three yolk proteins. The CON(sensus) sequence indicates positions in which there is identity in all three sequences.

of gene organization, the Mediterranean fruit fly has two types of yolk protein genes, one type with only one intron, as in *Drosophila Yp1* and *Yp2*, and the other with two introns at the same positions as in *Drosophila Yp3* (Rina and Savakis 1991).

### *Function*

Yolk proteins are the main protein component of the yolk platelets stored in mature oocytes.

### *Mutant Phenotype*

Mutation *Yp3*<sup>S1</sup> occurs in the signal peptide (*Yp3* Sequence) and blocks normal processing and secretion; as a consequence, YP3 fails to accumulate in oocytes (see below). Viability and fertility are normal, suggesting that *Yp3* has a redundant function (Liddell and Bownes 1991).

### *Tissue Distribution*

YP synthesis occurs only in adult females. The proteins are barely detectable in newly eclosed females, but the rate of synthesis increases steadily during the first 24 h after eclosion. The main sites of synthesis are the fat body and the follicle cells. In female fat bodies, YP can reach 20–30% of newly made proteins, and all three YPs are produced in comparable amounts. YPs are secreted into the hemolymph and then pinocytosed by the maturing oocytes. Follicle cells of stages 9 and 10 egg chambers also actively synthesize YPs; these are transferred to the oocyte through the intercellular matrix, without entering the hemolymph. Follicle cells contribute a significant proportion of YP1 and YP2, but YP3 synthesis is under-represented by four-fold in these cells (Brennan et al. 1982; Bownes 1986 and references therein). Synthesis of YPs is under hormonal control: 20-hydroxyecdysone stimulates fat bodies to synthesize all three YPs; juvenile hormone stimulates synthesis in fat bodies and ovaries, but the effect is more pronounced on YP1 and YP2 than on YP3 (Jowett and Postlethwait 1980).

## **Organization and Expression of the Cluster**

*Yp1* and *Yp2* are separated by 1,228 bp and transcribed divergently; *Yp3* is several hundred kb closer to the centromere (Fig. 33.2).

### *Developmental Pattern*

Transcription is limited to ovaries and fat bodies of adult females (Garabedian et al. 1985). Expression of *Yp1* and *Yp2* occurs, in general, in follicle cells lining

Yp3

-800 TTAATCTTTTGGTGATGTTGCCTATGTTTTGATTGAGCTCATCATTTTAGCAGTTGCTATGCTTTTGATATATAAAATATAATGCATTC -71

-710 ACCTGGCGGCTGGTCATTGATTCCAATTTGGCCGGCTTCCAATCGCTGGAGGTCAATGCCGGGTACACCCAGTTTCTCACTTGACGCGAGG -62

-620 TGTGCAAGTTTGTGCCAGTTCAATCTAATCAAGGGATCTGCACAAGTTGTTTCAATCAATCCGTACTAGAATACATTTTAAAGTGCAAG -53

-530 AGAACAAAAATTTGCATTACTTTGGGAATTATATGCATAAATCTGTAAGTGTGCTTTAAACCAAATGATAGTGATGATACAAAATATATCA -44

-440 CGATGCAATACTACTAGTGGTCAACGATTTTCCAATAATCTAAATCTTAACATTTTATGAATGGATTTTTTTTGCACATTTTTTGCCAA -35

-350 GTGTGAAGAGGTTCAAAAACCTTAGTGCATAAGAGAATAAATGGTTGGCAAACACACACACATGTGAAATAAATCCGGCTATTTGCAA -26

-260 TCAATTTTCCCTTGACTTGCACTTTATACACCGGCGACAGATCAGCAGAACGAAAGGGTGGGGAAAAAAGCTGGAAGCCTAGACAGCCGA -17

-170 CAACGACGACAACGACGACGACGACGACTTCTGTGGTCAGCAGAAAAATCGCTGGCAGTGCCTATCGGGAATCGGAGCTATATAAG -81

-----

-80 CCAGAGATGGGGCTGAAGGAAGCCATCAACAGTCGTTAGCGTTTGCCCTGATCTGATTCAATCCGGATTTGCACCAAAATGATGAGT 9  
MetMetSer (3)

A=S1

10 CTAAGGATTTGCCTGCTGGCCACCTGCCTCCTGGTGGCGCCCATGCCTCCAAGGATGCCTCCAATGACCGACTGAAGCCGACCAAGTGG 99  
LeuArgIleCysLeuLeuAlaThrCysLeuLeuValAlaAlaHisAlaSerLysAspAlaSerAsnAspArgLeuLysProThrLysTrp (33)  
Asp |

100 CTGACCGCCACCGAGCTGGAGAACGTCCTCCCTCAACGACATCACCTGGGAGCGTTTGGAGAATCAGCCGCTGGAGCAGGGCGCCAAG 189  
LeuThrAlaThrGluLeuGluAsnValProSerLeuAsnAspIleThrTrpGluArgLeuGluAsnGlnProLeuGluGlnGlyAlaLys (63)

190 GTGATCGAGAAGATCTGTGAGTAGAAACCGATGTTGCTGGAAATCTCCAGAGATAACCTCCTGTGAAATCACACCTAGACCACGTTGGCC 279  
ValIleGluLysIleT yrHisValGlyG (73)

280 AAATCAAGCAGCATCTGACCCCGAGCTTTGTGCCAGCCGAGCAATGTGCCGCTCGATTATCAAGTCCAATGGACAGAAGGTTGAGT 369  
IleIleLysHisAspLeuThrProSerPheValProSerProSerAsnValProValTrpIleIleLysSerAsnGlyGlnLysValGluC (10)

370 GCAAGTTGAACAACACTATGTGGAGACGGCCAAAGGCACAGCCGGATTGGCGAGGATGAGGTACCATTGTCTGACTGGTCTGCCAAGA 459  
ysLysLeuAsnAsnTyrValGluThrAlaLysAlaGlnProGlyPheGlyGluAspGluValThrIleValLeuThrGlyLeuProLysT (13)

460 CCAGCCCGCTCAGCAGAAGGCCATGCGCAGGTTGATCCAGGCCTACGTCCAGAAGTACAACCTCCAGCAGCTGCAGAAGAACGCCAGG 549  
hrSerProAlaGlnGlnLysAlaMetArgArgLeuIleGlnAlaTyrValGlnLysTyrAsnLeuGlnGlnLeuGlnLysAsnAlaGlnG (16)

550 AGCAGCAGCAGCAGCTCAAGAGCAGCGACTACGACTACACAGCAGCGAGGAGGCCGCTGACCAATGGAATCCGCCAAGGCTGCCAGCG 639  
luGlnGlnGlnGlnLysSerSerAspTyrAspTyrThrSerSerGluGluAlaAlaAspGlnTrpLysSerAlaLysAlaAlaSerG (19)

640 GCGATTTGATCGTAAGTTGGTCGCATTCTATATTTTCATAATTAACGTTACATATGGATATTTATGAAATTCAAATTCAGATCATTG 729  
lyAspLeuIle IleIleA (19)

730 ACCTCGGCTCCACCTGACCAACTTCAAACGCTACGCGATGCTGGATGTTCTGAACACCGCGCCATGATCGGCCAGACCTGATCGATC 819  
spLeuGlySerThrLeuThrAsnPheLysArgTyrAlaMetLeuAspValLeuAsnThrGlyAlaMetIleGlyGlnThrLeuIleAspL (22)

820 TGACCAACAAGGGTGTGCCCAGGAGATCATCCATCTGATCGCCAGGGAATCAGCGCCATGTGGCCGGAGCTGTGGCAACAAGTACA 909  
euThrAsnLysGlyValProGlnGluIleIleHisLeuIleGlyGlnGlyIleSerAlaHisValAlaGlyAlaAlaGlyAsnLysTyrT (25)

910 CCGCCCAACCGGACACAAGCTGCGCCGATCACCGGCTGGATCCCGCAAGGTGCTGCCAAGCGTCCCAAGCTCCCAAGCTCCCGGTTGGTCTGT 999  
hrAlaGlnThrGlyHisLysLeuArgArgIleThrGlyLeuAspProAlaLysValLeuSerLysArgProGlnIleLeuGlyGlyLeuS (28)

1000	CCC GCGCGCATGCTGACTTCGTTGATGCCATTACACATCGACCTTCGCCATGGGCACGCCCATCCGTTGCGGCGATGTTGACTTCTACC	1089
	erArgGlyAspAlaAspPheValAspAlaIleHisThrSerThrPheAlaMetGlyThrProIleArgCysGlyAspValAspPheTyrP	(319)
1090	CCAACGGACCGTCCACCGGTGCCGGCTCCGAGAATGTGATCGAGGCTGTGGCCCGTCCACCCGTTACTTTGCCGAGTCTGTGCGTC	1179
	roAsnGlyProSerThrGlyValProGlySerGluAsnValIleGluAlaValAlaArgAlaThrArgTyrPheAlaGluSerValArgP	(349)
1180	CCGGTAGCGAGCGCAATTTCCCGCGTTCGGCCAACTCGCTGAAGCAGTACAAGGAGCAGGATGGCTTTGGCAAGCGCGCTACATGG	1269
	roGlySerGluArgAsnPheProAlaValProAlaAsnSerLeuLysGlnTyrLysGluGlnAspGlyPheGlyLysArgAlaTyrMetG	(379)
1270	GTCTCCAGATCGACTACGATCTGCGCGGTGACTACATCTGGAGGTCAACGCCAAGAGCCCTTCGGTCAGCGCAGCCCTGCCACAAGC	1359
	lyLeuGlnIleAspTyrAspLeuArgGlyAspTyrIleLeuGluValAsnAlaLysSerProPheGlyGlnArgSerProAlaHisLysG	(409)
1360	AGGCCGCTACCATGGCATGCCACCAGCCAGAAGTACAGCGCCCATGGCCACGCCCTGGTTACCAGGGACGTTTCGATCGTCACGCAC	1449
	lnAlaAlaTyrHisGlyMetHisHisAlaGlnAsnEnd	(420)
1450	TTTCTGATAATCAGAAAAATAAAACCCGGAATGCGTAGTGTAGCTTAGAAGTTTCATCAAAACATCAAAAAAGAAAAATCTATAAAATCC	1539
1540	CATAAAAAATAAAGCTGCAAATTTTCGAAAAGTCAAGTTTTTAAATAGCAATAGCAATGGTTATTCTGGATTGGATTCTAACTTTTATGG	1629
	-----   (A) <sub>n</sub>	
1630	TATTA AAAAACACACACAAGAATTGCTGGGCACATTTTAGGCCACCCCTTCTGAAGTAAATAGAAAAATTTCCGAAAATATACATATTT	1719
1720	AACATAGTAAATCGGCCAAACAACTTAAATGAGCTAATAATAAAAAGATAAATGCATATATCACAGGTGATCTTAAGCAGATGCTTAACC	1809
1810	AAAAACAACACGATAAATAAAGCAACAAAAAGTCCATAAAATACAATTATGACACCTAATGAAAGGTACACGAAAGAAAAATGTAGATA	1899
1900	ATAAATAAACTGAAAAAGAAATTAGGAATAACTCATAAAAATCAAATTTAGAAAACTGTGCAGCTTGGTATTACTAGCACCCCTAGATGC	1989
1990	TTAACAGGATTGCGAAAGTGGGATGGAAATACGCACAACGAGATGGATGCATTGAGTGGCGGGAAGTGAGAGTGAGGCAACTAGTGCCG	2079
2080	TTGCCACTTGATGTGCACTCAATTA AAACCTGCATTGCGTTTATCGTTAGTGACTACTCGTTCAAAAATCACTGGGCAACCTGTGTAAC	2169
2170	TCAATTGTTCTTACAGTTTTGGGCATCGCGGTTAAATGTCAAAGTTGAACTTTATCAAATGCAATAGACAAACTAGAAAAGGCGAGC	2259
2260	GAAAACAGCAGAGTCGAAAATAGAGCGAGATAGGGAGCTGGAGTGACAGGAGCGGAATGACAACAGTTGGCGTCTTTTGTGTGCATGT	2349
2350	CGTGACATGTTTGCTTTGACTCTGACCGAACGGAAATGCGCCGTTAAGCTT	2399

*Yp3* SEQUENCE. Strain, Canton S. Accession, M15898 (DROY P3) and X04754 (DROY P3G) as corrected near the transcription initiation site by Liddell and Bownes (1991). The vertical line at Ala-19 marks the putative cleavage site of the signal peptide.

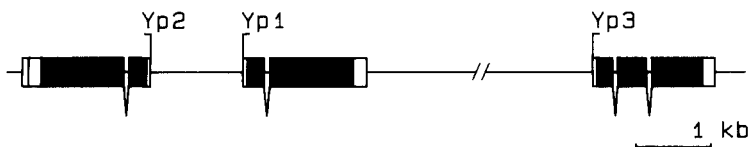


FIG. 33.2. *Yp* cluster, centromere to the right. Note that *Yp3* is many kbs from *Yp1/Yp2* and that the direction of transcription of the three genes relative to the centromere is not known

the maturing oocyte (stages 8–10) but not in the nurse cells (Logan and Wensink 1990) (see *Yp1 Promoter*).

## *Yp1*

### Gene Organization and Expression

Open reading frame, 439 amino acids; expected mRNA length, 1,559 bases, in agreement with northern analysis. S1 mapping, primer extension and sequence features were used to define the 5' end. The 3' end was obtained from S1 mapping. There is one intron in the Tyr-74 codon (*Yp1 Sequence*) (Hung and Wensink 1981; Hovemann and Galler 1982).

#### *Promoter*

There is evidence that the 1,228-bp segment separating *Yp1* and *Yp2* includes two *cis*-acting regulatory elements, one for ovarian and the other for fat body expression; these two elements control both *Yp1* and *Yp2*. The two genes were cloned separately into *P* elements; this split the 1,228-bp segment leaving 886 associated with *Yp1* and the remaining 342 with *Yp2*. In germline transformants, the fragment with *Yp1* was expressed only in fat bodies and the one with *Yp2* only in ovaries (Garabedian et al. 1985).

*Fat Body Enhancers* Deletion mapping and ligation of fragments to a heterologous promoter (*Hsp70*) and a reporter gene (*lacZ*) showed further that 125 bp of the 886-bp segment (from –378 to –253 in the *Yp1 Sequence*) was sufficient for stage-, sex- and tissue-specific expression in adult female fat bodies. This regulatory segment of DNA acts relatively independently of its orientation and distance from the genes, and it acts on both *Yp1* and *Yp2* (Garabedian et al. 1986; K. Coschigano and P. Wensink, personal communication). The rest of that segment, from –942 to –378 contains a weaker fat body enhancer (P. Wensink, personal communication).

Sex-specificity of expression seems to be controlled by the *doublesex* (*dsx*) gene products; these bind to three sites in the fat body enhancer, and all three binding sites contain sequences related to CTACAAAGT (Burtis et al. 1991). Binding sites A and B (between –378 and –253) direct male-specific repression (mediated by binding of DSX<sup>M</sup>, the product of *dsx* in males) and female-specific stimulation (mediated by binding of DSX<sup>F</sup>, the product of *dsx* in females) (K. Coschigano and P. Wensink, personal communication). Partly overlapping binding site A are binding sites for two regulatory proteins (AEF-1 and C/EBP) that are also involved in regulating *Adh* expression in fat body (*Yp1 Sequence*) (Falb and Maniatis 1992).

*Ovarian Enhancers* Expression of *Yp1* and *Yp2* in ovarian follicle cells is controlled by an enhancer, *oe1*, located in the interval between –1,242 and

Yp1

-1453 GCTGCTCCACATTGTCCAGGGAGTTGGATCGGCGACCGGAACGGTTACCAGACTGGGGATTACCCATGGCGACCCGAGAAGGCAGGCCA -1364  
 | -  
 <--Yp2  
 -1363 TAACGCAAAGGGTGCAGAGGATTCATTGTGGCTTCCAAGTTCGACTTTTTTCAGACACCGTACCAAATTTGACTGCATGCCACTGCTG -1274  
 Met(Yp2) -|oe2  
 -1273 CGACTCAATGCATTTTATACCCCTTGAATCGGTAGTCTATACACACTATAATGCACGCGCCGGAAGCAATTGATTTTCAGCAACCGATT -1184  
 | -  
 -1183 CTGGATCAGCACAAATGCATTGATTCGCAGCGTCAGTGATTTTGCAACACTTCTGATGAGCTCTAAAATTCGTTCCCTTTTTTTTTT -1094  
 -1093 TTTTTTTGGTTATTAAGTATCCATCGGGTAACAGGTAATGGGAAACTCTTTAAACCAGCACTTCATAACATAACAAAAGGTGGTCTG -1004  
 -1003 GCCATTAAGGGCTTGACAGTGGGGCACGACTTGAACATGCACAGGTCAAGATAAAGCTTTTGTGGAAAAAATATTTGGCAATTT -914  
 -|oe1  
 -913 TGTGAAATTTATGCAACTATTTAAGTGTGGCCAAAAGAATTGTCTAAATGTTCTATAAGCAGATAACACTTTCAGGGAAATGCAAAAT -824  
 -823 AAATATATTATAAATTATAATATTATAAATATAAATATTTACATCTATCGAAATATACATATATTTTAATAAGTAGAATGAGTTACATG -734  
 -733 AAATAGCATCGATAAGATCATATATTATAAAACGAATCCCGGATATTAATAAGATACTCCTTGAAAAACGTTTCCCTGAAATCAATTCA -644  
 -643 TTTCTAAAGTCCAAAAACAAATATAATCTTACTATCTTGCCTTGGAAACTACAACATTCCATCTTTTCGTATCAATGGCAAACTCTA -554  
 -553 GGAATCAATGAAGTGTATCGGCTTGAATTGAAAATGCAAAATATGACTTTTAATTAAGCAGAAGAAAAGTCCCAAATATAAATCTAC -464  
 -463 TTATAAACAAAAAAATCAATAAAATGTTGTATATAATAACCAACTAATGCCATGTAGATCTATATTTTATGCATTTATTTGATCAAA -374  
 -373 TCCGGTGCACAACATAATGTTGCAATCAGCGGAGCCTACAAAGTGATTACAATTAATAAATATCAGGCGGCAGCAGGTGCTGCTAAGTC -284  
 -----A -----B -----  
 -----aef-1  
 -----c/ebp  
 -283 ATCAGTGGGGTCAGCTATAGGTAGGCCCGGTGTCTATTTTGTATGTATAACAATTTATCCGCTATCGATAGCATATACACTCGATCCGAT -194  
 ----C  
 -193 TCCCAGGCACCCGAAAACCTTACTCAGCACAAAGTGACCGATTAAGGCCTGAGCCAGCGAAAAGCAAGTCGGAATGGGAAATCGCTCA -104  
 -->-57  
 -103 GCGTAAATTGTGGTATATAAACCACTCGTTGGATTGGAAGGCCAGTTCACACTCACTCAGTGTGAAGTCGCATCCGAGGACCAAAAT -14  
 -----  
 -13 CCCAAATCCGAACCATGAACCCATGAGAGTGCAGGCTTCTGGCTTGGCTTGGCGGTGCGCCGCTTGGCCAAGCCCAATGGCCGTATGG 76  
 MetAsnProMetArgValLeuSerLeuLeuAlaValAlaAlaLeuAlaLysProAsnGlyArgMetA (26)  
 |  
 77 ACAACTCCGTCAACCAAGCATTGAAGCCGTCGAGTGGCTCTCCGGATCCAGCTGGAGGCCATTCGCCCTCGACGATTTACCATTG 166  
 spAsnSerValAsnGlnAlaLeuLysProSerGlnTrpLeuSerGlySerGlnLeuGluAlaIleProAlaLeuAspAspPheThrIleG (56)  
 167 AGCCTCTGGAGAACATGAACCTGGAGCGTGGCGCCGAGCTGCTGCAGCAAGTCTGTGAGTAATCCTAGATGCAGATAAAAAAAAAAAAA 256  
 luArgLeuGluAsnMetAsnLeuGluArgGlyAlaGluLeuLeuGlnGlnValT (74)

(continued)



257	AAACATCGAATATTCCTATGGAATATATATATATCCCTTTGTAGACCACCTGTCGCAGATCCACCACAACGTTGAGCCCACTATGTGCCACG yrHisLeuSerGlnIleHisHisAsnValGluProAsnTyrValProSer	346 (90)
347	GGCATCCAGGTCATGTGCCAAGCCCAATGGTGACAAGACCGTTGCTCCCTGCAACGAGATGATCCAGCGCTGAAGCAGAAGCAGAAC GlyIleGlnValTyrValProLysProAsnGlyAspLysThrValAlaProLeuAsnGluMetIleGlnArgLeuLysGlnLysGlnAsn	436 (120)
437	TTTGGTGAGGATGAGGTGACCATCATTGTGACCGGACTGCCAGACCAGCGAGACCGTGAAGAAGCGACCAGGAAGCTGGTTCAGGCT PheGlyGluAspGluValThrIleIleValThrGlyLeuProGlnThrSerGluThrValLysLysAlaThrArgLysLeuValGlnAla	526 (150)
527	TACATGCAGCGCTACAATCTGCAGCAGCAGCGCCAGCAGCGCAAGAACGGCAACCCAGGACTACCAGGATCAGAGCAACGAACAGAGGAAG TyrMetGlnArgTyrAsnLeuGlnGlnGlnArgGlnHisGlyLysAsnGlyAsnGlnAspTyrGlnAspGlnSerAsnGluGlnArgLys	616 (180)
617	AACCAGAGGACCAGCAGCGAGGAGGACTACAGCGAGGAGGTTAAGAACGCCAAGACCCAAGCGGCGACATCATTGTGATCGATTGGGC AsnGlnArgThrSerSerGluGluAspTyrSerGluGluValLysAsnAlaLysThrGlnSerGlyAspIleIleValIleAspLeuGly	706 (210)
707	TCCAAGCTGAACACCTATGAGCGTTATGCCATGCTCGACATTGAGAAGACCGCGCCAAGATCGGCAAGTGGATCGTCCAGATGGTCAAC SerLysLeuAsnThrTyrGluArgTyrAlaMetLeuAspIleGluLysThrGlyAlaLysIleGlyLysTrpIleValGlnMetValAsn	796 (240)
797	GAGTTGGACATGCCCTTCGATACCATTACCTGATTGGCCAGAATGTGGGTGCCATGTTGCCGGTGCCGCTGCCAGGAATCACCCCGT GluLeuAspMetProPheAspThrIleHisLeuIleGlyGlnAsnValGlyAlaHisValAlaGlyAlaAlaAlaGlnGluPheThrArg	886 (270)
887	CTCACCAGCACAAAGCTGCGCCGTGTACCAGGCTGGATCCCTCCAAGATCGTGGCCAAGAGCAAGAACACCCCTGACCCGCTGGCTCGC LeuThrGlyHisLysLeuArgArgValThrGlyLeuAspProSerLysIleValAlaLysSerLysAsnThrLeuThrGlyLeuAlaArg	976 (300)
977	GGTGATGCTGAATTCGTTGACGCCATCCACACCTCGGTCTACGGCATGGGCCACCCCATCCGCTCCGGTGATGTTGACTTCTATCCCAAT GlyAspAlaGluPheValAspAlaIleHisThrSerValTyrGlyMetGlyThrProIleArgSerGlyAspValAspPheTyrProAsn	1066 (330)
1067	GGACCTGCCCGCGGTGTTCCCGAGCCAGCAACGTGGTGGAGGCCCATGCGTGCCACCCGCTACTTCGCCGAGTCCGTGCGTCCCGGA GlyProAlaAlaGlyValProGlyAlaSerAsnValValGluAlaAlaMetArgAlaThrArgTyrPheAlaGluSerValArgProGly	1156 (360)
1157	AACGAGAGGAGCTCCCGCCGTGCCAGCCAATCCCTCGACAGTACAAGCAGAACGATGGATTCCGCAAGCGTGCCATACATGGGCATC AsnGluArgSerPheProAlaValProAlaAsnSerLeuGlnGlnTyrLysGlnAsnAspGlyPheGlyLysArgAlaTyrMetGlyIle	1246 (390)
1247	GATACCGCTCACGATCTCGAGGGTGACTACTTCTGCAAGTGAACCCCAAGTCTCCTTTCCGGCCCAACGCACCCGCCAGAACGAGCAGC AspThrAlaHisAspLeuGluGlyAspTyrIleLeuGlnValAsnProLysSerProPheGlyArgAsnAlaProAlaGlnLysGlnSer	1336 (420)
1337	AGCTACCAGGTTGCCACCAGGCGTGAACACCAACCCAGGACAGCAAGGACTACCAGTAAGGATGAGTCTGCTTACTCTGGACACCTGGA SerTyrHisGlyValHisGlnAlaTrpAsnThrAsnGlnAspSerLysAspTyrGlnEnd	1426 (439)
1427	ATGGCAACTACCAAAACACCCACCAACACACAAACACTGTAGTCCCTAAGTGAACCCATATTGGCCCTTTCTTGAGATTACCTAAAC	1516
1517	ATTTAACGAGCACATCGCGAAATTCAGCAAATAAACGCTCGATAAAGAGCTTAAAAATATCTATTTTGTATCTTAAATCATTTAGGAA -----   (A) <sub>n</sub>	1606
1607	CTATAATAGTCTAATAGATCATCCAAAAAAGGGGAACAAAATCAAAAGTAAATATCGTAGTTGGTTTTGTAAACTTAGATTTATTTT	1696
1697	ATTGTTGTCGGTGTTTTGTGG	1718

*Yp1* SEQUENCE. Strain, Canton S. Accession, V00248 (DMYOLK), X01524, J01157 and M11170 (DROYPI2). The segment between -1,453 and -1,282 corresponds to the reverse complement of *Yp2*: sites of transcription and translation initiation are indicated. The vertical line at Ala-19 marks the putative cleavage site of the signal peptide. A, B and C indicate the footprints produced by the *dsx* products in the main fat body enhancer; and *af-1* and *c/ebp* are footprints of fat-body specific proteins.

Yp2

-761 GAAAAGTATGGAATGTTTGTAGTTTCCAAGGCAAGATAGTAAGATTATATTTGTTTTGGACTTTAGAAGTGAATTGATTCAGGGGAAAC -672  
 -671 GTTTTTCAAGGAGATTCATTTAATATCCGGGATTCGTTTTATAATATATGATCTTATCGATGCTATTTCATGTAACCTACTTACTTA -582  
 -581 TTAATAATATATGTATATTTTCGATAGATGTAATATTTTATAATTTATAATTTATAATATATTTATTTGCATTTCCCTGAAAG -492  
 -491 TGTTATCTGCTTATAGAACAATTTAGACAATCTTTTGGCAAACACTTAAATAGTGCATAAAATTCACAAAATTGCCAAATATTTTTTT -402  
 -401 CAAACAAAAGCTTTATCTTGACCTGTGCATGAGTCAAGTCGTGCCCCACTGTCAAGCCCCTTAATGCCAGACCACCTTTTGTTTATG -312  
 -311 TTATGAAAGTCTGGTTAAAGAAGTTTCCCATCTACCTGTTACCCGATGGATACTTAATAACCAAAAAAAAAAAAAAAAAAGGGGAACGA -222  
 -221 AATTTTAGAGCTCATCAGAAGTGTGCAAAATCACTGACGCTGCGAATCAATGCATTTGTGCTGATCCAGAAATCGGTTGCTGAAATCAA -132  
 --- -->-53  
 -131 TTGCTTCCGGCGCTGCATTATAGTGTGTATAGACTACCGATTCCAAGGGTATAAAATGCATTGAGTCGCAGCAGTGGGATGCAGTAC -42  
 ---  
 -41 AATTTGTACGGTGTCTGAAAAAGTCGAACCTGGGAAGCCACAATGAATCCCTCTGCGCACCCCTTTCGTTATGGCCTGCTTCTGGCGGTC 48  
 MetAsnProLeuArgThrLeuCysValMetAlaCysLeuLeuAlaVal (16)  
 49 GCCATGGGTAATCCCAGTCTGGTAACCGTCCGGTCGCGATCCAACCTCCCTGGACAATGTGGAGCAGCCAGCAACTGGGTCAACCCA 138  
 AlaMetGlyAsnProGlnSerGlyAsnArgSerGlyArgArgSerAsnSerLeuAspAsnValGluGlnProSerAsnTrpValAsnPro (46)  
 139 CGTGAAGTCGAGGAGCTGCCAACCTGAAGGAGTTACCCCTAAGAAGTCGAGGAGATGAGCATGGAGGAGGGCGCTACGCTGTGGAC 228  
 ArgGluValGluGluLeuProAsnLeuLysGluValThrLeuLysLysLeuGlnGluMetSerMetGluGluGlyAlaThrLeuLeuAsp (76)  
 229 AAGCTCTGTAAGTTCAAGGATCTCTAAAAGTCTACCAATCATGTTATATTTACACGCACATCTCTATCCCGCAGACCATCTGTCCCAGT 318  
 LysLeuT yrHisLeuSerGlnP (84)  
 319 TCAACCATGTCTTCAAGCCCATTACACCCCGGAACCCAGCCAGATCAGGGGTACATTGTGCGGCAGCGCGCCAGAAGATCGAGTTCA 408  
 heAsnHisValPheLysProAspTyrThrProGluProSerGlnIleArgGlyTyrIleValGlyGluArgGlyGlnLysIleGluPheA (114)  
 409 ACCTGAACACTTTGGTGGAGAAGGTTAAGCGCCAGCAGAAGTTCGGCGACGATGAGGTCACCATCTTATCCAGGGCTGCCCGAGACCA 498  
 snLeuAsnThrLeuValGluLysValLysArgGlnGlnLysPheGlyAspAspGluValThrIlePheIleGlnGluLeuProGluThrA (144)  
 499 ACACCCAAGTGCAGAAGGCTACCAGGAAGCTGGTGCAGGCTACCAGCAGCGTTACAACCTCCAGCCCTATGAGACCACCGACTACTCCA 588  
 snThrGlnValGlnLysAlaThrArgLysLeuValGlnAlaTyrGlnGlnArgTyrAsnLeuGlnProTyrGluThrThrAspTyrSerA (174)  
 589 ACGAGGAGCAGAGCCAGAGGAGTCCAGCGAGGAGCAGAAACGCAGCGCAGGAAGCAGAACGGTGAACAGGATGATACCAAGACCGGAG 678  
 snGluGluGlnSerGlnArgSerSerSerGluGluGlnGlnThrGlnArgArgLysGlnAsnGlyGluGlnAspThrLysThrGlyA (204)  
 679 ACCTGATTGTGATCCAGCTGGGCAATGCCATCGAGGACTTTGAGCAGTACGCCACCTGAACATTGAGCGTCTGGGCGAGATCATTGGCA 768  
 spLeuIleValIleGlnLeuGlyAsnAlaIleGluAspPheGluGlnTyrAlaThrLeuAsnIleGluArgLeuGlyGluIleIleGlyA (234)  
 769 ACCGTCGTGGTTGAGCTGACCAACCCGTGAACGTGCCCCAGGAGATCATCCATCTGATTGGCTCTGGACCCGCTGCCACGTTGCCGGAG 858  
 snArgLeuValGluLeuThrAsnThrValAsnValProGlnGluIleIleHisLeuIleGlySerGlyProAlaAlaHisValAlaGlyV (264)  
 859 TGGCTGGACGCCAGTTCACCCGTCAGACC GGACACAAGTTGCGCCGCATCACC GCCCTGGACCCCACTAAGATCTACGGCAAGCCCGAGG 948  
 aAlaGlyArgGlnPheThrArgGlnThrGlyHisLysLeuArgArgIleThrAlaLeuAspProThrLysIleTyrGlyLysProGluG (294)

(continued)

949	AGAGGCTGACCGGCTGGCCCGTGGTGTGCTGACTTCGTTGATGCCATCCACACCTCCGCCTACGGCATGGGTACCAGCCAGCGATTGG 1uArgLeuThrGlyLeuAlaArgGlyAspAlaAspPheValAspAla11eHisThrSerAlaTyrGlyMetGlyThrSerGlnArgLeuA	1038 (324)
1039	CCAACGTGGACTTCTTCCCAACGGACCTCGACCGGAGTGCCCGGAGCCGATAATGTCGTTGAGGCCACCATGCGTGCACCCCGCTACT 1aAsnValAspPhePheProAsnGlyProSerThrGlyValProGlyAlaAspAsnValValGluAlaThrMetArgAlaThrArgTyrP	1128 (354)
1129	TCGCCGAGTCTGTGCGTCTCTGGAAACGAGAGGAACTTCCCTCCGTGGCCGCCAGCTCGTACCAGGAGTACAAGCAGAACAAGGGCTATG heAlaGluSerValArgProGlyAsnGluArgAsnPheProSerValAlaAlaSerSerTyrGlnGluTyrLysGlnAsnLysGlyTyrG	1218 (384)
1219	GCAAGCGCGGATACATGGGCATCGCCACCGATTTTCGATCTGCAGGGCGATTACATTCTGCAGGTGAACTCCAAGAGCCCTTCGGCAGGA lyLysArgGlyTyrMetGlyIleAlaThrAspPheAspLeuGlnGlyAspTyrIleLeuGlnValAsnSerLysSerProPheGlyArgS	1308 (414)
1309	GCACTCCCGCCAGAAACAGACGGCTACCACCAAGTCCACCAGCCCTGGCCGACCTCTCTCCAACAGGGTTCCCGCGTCAGTAGA erThrProAlaGlnLysGlnThrGlyTyrHisGlnValHisGlnProTrpArgGlnSerSerSerAsnGlnGlySerArgArgGlnEnd	1398 (442)
1399	TCATCGCACAGTGATCCATCGATGACAACCAGATCGCACACCCCTCATGCGAGCGAACCCTCCAGCCCATCTCATCCAGCAGAACCCT	1488
1489	CTGCCAGTTGCATCCACTACGATTAGTTAGCTTTGTTTTTAACTCACAATAAAAAACGTTTGCATTTTTAAACATTCTAAAGAGTTCA -----   (A) <sub>n</sub>	1578
1579	GTTCAATATCGAAAAAAACCCAGTTC AATTTACAATAAAAAAATTGCTTATGTCGAAATATTTGAGAGTTCCAATGTCTCCTTATAT -----   (A) <sub>n</sub>	1668
1669	AAAAATATCCAAAACCAAAATTATGCAATGCCACTGAGGCCATAAAGAAGCACACAACAACATTTGGGT	1738

*Yp2* SEQUENCE. Strain, Canton S. Accession, X01524, J01157 and M11170 (DROY12). The vertical line at Gly-19 marks the putative cleavage site of the signal peptide.

– 942 in the *Yp1* Sequence, between 43 and 343 bp upstream of the transcription initiation site of *Yp2* (Logan et al. 1989). *oe1* is composed of multiple parts, each controlling the expression in various subsets of follicle cells (Logan and Wensink 1990).

*oe2*, located between – 1,389 and – 1,284 (the first 105 bp of the first exon of *Yp2*), is also necessary for expression of *Yp1* in ovaries (Logan et al. 1989).

*Other Regulatory Elements* Another *cis*-acting regulatory region was identified by its ability to bind YPF1, a heterodimer with subunits of 85 and 69 kD, very specifically and very tightly ( $K_D < 5 \times 10^{-16}$ ). This element occurs in the translated region of *Yp1* (between positions 82 and 126 in the *Yp1* Sequence) and is necessary for *Yp1* transcription (Mitsis and Wensink 1989a, 1989b).

## *Yp2*

### Gene Organization and Expression

Open reading frame, 442 amino acids; expected mRNA lengths, 1,546 or 1,630 bases depending on which of two polyadenylation sites is used. S1 mapping

and primer extension were used to define the 5' ends. The 3' ends were obtained by S1 mapping. There is an intron in the Tyr-79 codon (*Yp2* Sequence) (Hovemann and Galler 1982; Hung and Wensink 1983).

### Promoter

See discussion of the *Yp1* promoter, above.

## Yp3

### Gene Organization and Expression

Open reading frame, 420 amino acids; expected mRNA length, 1,488 bases. S1 mapping and cDNA sequencing were used to define the 5' and 3' ends. There are two introns, at Tyr-69 and after Ile-196 (*Yp3* Sequence) (Garabedian et al. 1987; Yan et al. 1987; Liddell and Bownes 1991).

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# II

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# 34

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## Size Variations Among the Elements that Constitute the Genes of *Drosophila* (Leader, coding region 3' untranslated region, exons, introns)

The discussion in this chapter centers around two questions: (1) what are the size ranges of the various elements that constitute a functional gene? and (2) is there a correlation between the size of one element and the size of another.

The data analyzed in this chapter are derived from 73 of the genes presented in Part I. Because 12 of those 73 genes have multiple transcripts, they encompass a total of 87 transcripts. Two partly overlapping datasets can be examined: Dataset A includes all 87 transcripts, but elements shared by different transcripts of the same gene are considered only once. For example, if two transcripts of a gene differ only with respect to the poly(A) site, both 3' untranslated regions (3' UTR) are included in the analysis, but the leader is counted only once, since it is the same for both transcripts. Dataset B includes only one representative from each family of related genes or from the group of multiple transcripts of a given gene. In this case the sample is reduced to 40 "unrelated" transcripts. The size of a few elements were found to be outside the expected size range suggested by statistical analyses and so were excluded from the analysis. These elements are the 3' UTR of *bsg25D* II and the leader, 3' UTR and introns of *Ubx*.

### **Coding Regions and Untranslated Regions**

The questions posed in the first paragraph are discussed as they apply to those parts of the gene that give rise to the mature mRNA: the leader, the coding region and the 3' untranslated regions (the size of these regions in bp will be represented by the symbols *Leader*, *CR* and *3'UTR*, respectively, and *mRNA* will be used for *mRNA* size. These elements are often encoded by segments in more than one exon; however, because they are the constitutive parts of the



mature message, they will be considered here as units. The size and position of exons and introns will be discussed in the next section.

## Size Distribution

Table 34.1 covers Dataset A and lists 87 transcripts arranged in order of increasing CR, values are given for *Leader*, *CR*, *3'UTR*, *mRNA*, and the fraction

TABLE 34.1. Dataset A

<i>Gene</i>	Leader	CR	3'UTR	mRNA	CR/mRNA
* Mtn	124	123	140	387	0.32
Mto	144	132	100	376	0.35
CecA1	73	192	81	346	0.55
* CecA2	81	192	81	354	0.54
CecB	71	192			
Sgs7	32	225	61	319	0.71
Sgs8	32	228	92	353	0.65
CytC2	44	318	311	673	0.47
* CytC1	68	327	212	607	0.54
* Hspg2	60	336	69	465	0.72
Hspg2 d	182	336	103	622	0.54
Lcp3	45	339			
S15	45	348	126	519	0.67
Vm32E	29	351	54	434	0.81
Lcp2	42	381			
* Lcp1	42	393			
* Rp49	9	402			
JanA	60	408	241	661	0.62
Cp16	46	417	52	515	0.81
* JanB	100	423	56	579	0.73
* Vm26A1	81	426	122	629	0.68
* Sgs5	33	492	129	653	0.75
Vm26A2	62	507	56	625	0.81
* Hspg3	168	510	301	979	0.52
Cp18	44	519	86	649	0.80
* Cp19	45	522	86	653	0.80
* Hsp22	251	525	181	957	0.55
Hsp23	112	561	201	874	0.64
ASC-ac	63	606	243	912	0.66
* Ddc-Cc	200	612	376	1,188	0.52
Hsp26	184	627	138	949	0.66
Hsp27	119	642			
* Fcs3C	111	654	41	786	0.83
Hspg1	93	717			
* Ddc-Cs	353	738	605	1,696	0.44
Adh d	123	771	173	1,067	0.72
* Adh p	70	771	173	1,014	0.76
* ASC-lsc	27	774	383	1,184	0.65
* Cp36	31	861	112	1,004	0.86

TABLE 34.1. *Continued*

<i>Gene</i>	Leader	CR	3'UTR	mRNA	CR/mRNA
Cp38	77	921	293	1,290	0.71
Sgs3	29	924	164	1,117	0.83
* h alpha1	491	1,014	830	2,335	0.43
h alpha2	295	1,014	830	2,139	0.47
* ASC-sc	117	1,038	283	1,438	0.72
* Ubx IVa	966	1,041	2,100	4,106	0.25
* Sryb	144	1,056	99	1,299	0.81
* Act5C I	155	1,131	184	1,560	0.73
Act5C II	155	1,131	543	1,919	0.59
Act5C III	119	1,131	184	1,524	0.74
Act5C IV	119	1,131	543	1,883	0.60
Act42A	102	1,131			
Act79B	147	1,131			
Act87E I	82	1,131	355	1,568	0.72
Act87E II	82	1,131	367	1,580	0.72
Act88F	95	1,131			
* eve	94	1,131	191	1,416	0.80
Ubx Ia	966	1,143	986	3,096	0.37
* ftz	70	1,242			
* Yp3	59	1,260	168	1,490	0.85
* kni	271	1,290	507	2,068	0.62
Sry d	67	1,293	104	1,464	0.88
Yp1	61	1,320	181	1,562	0.85
Yp2(I)	51	1,329	166	1,546	0.86
Yp2(II)	51	1,329	250	1,630	0.82
* EF-1AF2	138	1,389	1,030	2,558	0.54
EF-1AF1	80	1,392	582	2,054	0.68
* Kr(I)	185	1,401	265	1,851	0.76
Kr(II)	185	1,401	633	2,219	0.63
* Ddc I	197	1,428	298	1,923	0.74
* Pgd	35	1,446	178	1,659	0.87
ASC-ase	456	1,461	346	2,263	0.65
* Amy	33	1,485	83	1,601	0.93
* Ddc-DoxA	90	1,485	82	1,657	0.90
* bcd	169	1,485	817	2,471	0.60
Ddc II	232	1,533	298	2,064	0.74
Ddc-amd	150	1,533	99	1,782	0.86
* Sry a	43	1,593	226	1,862	0.86
* y	171	1,626	188	1,985	0.82
* prd	245	1,842	330	2,417	0.76
Hsp70C1d	242	1,926			
* Hsp70A7d	246	1,932	210	2,388	0.81
* bsg25D I	296	2,226	198	2,720	0.82
bsg25D II	296	2,226	2,227	4,749	0.47
hb d	510	2,277	561	3,348	0.68
* hb p	161	2,277	561	3,000	0.76
* otu2	122	2,436	486	3,045	0.80
otu1	171	2,562	486	3,220	0.80

Asterisks mark transcripts included in dataset B.

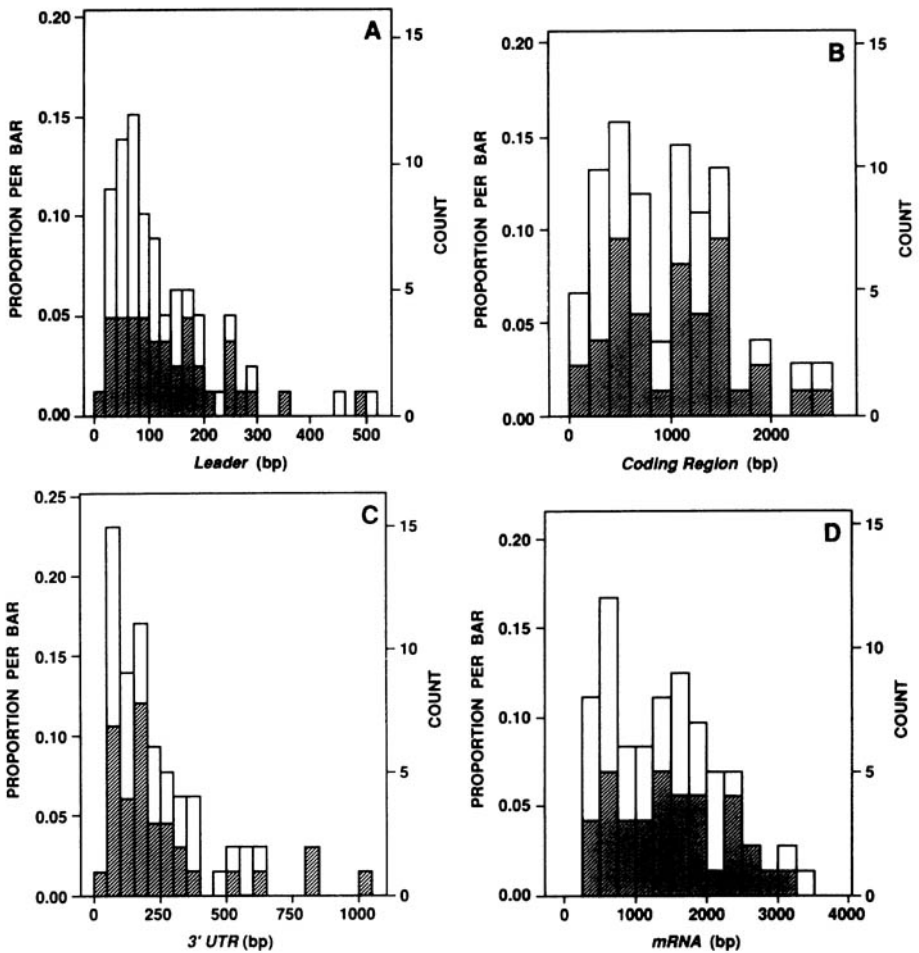


FIG. 34.1. Frequency distributions of size classes of: mRNA, coding regions, leaders and 3' UTRs. Open bars represent Dataset A, shaded bars represent Dataset B. The "Count" scale measures the absolute number of cases in each class and it applies to both datasets. The "proportion per bar" scale measures the fraction of the total in each class and it applies only to dataset A. The transcripts for *Ubx* and *bsg25D* II were excluded.

	N	Min.	Max.	Mean	St. Dev.
(A) Leader:					
Dataset A	79	9	510	127	103
Dataset B	39	9	491	137	101
(B) Coding Region:					
Dataset A	76	123	2,562	959	589
Dataset B	39	123	2,436	1,036	575
(C) 3' UTR:					
Dataset A	66	41	1,030	250	206
Dataset B	34	41	1,030	260	236
(D) mRNA:					
Dataset A	72	318	3,348	1,412	773
Dataset B	39	354	3,044	1,517	770

of the mature mRNA represented by *CR*. Fig. 34.1 shows frequency distributions for the size of these elements. For both datasets, *mRNA* and *CR* are broadly distributed; 90% of all *mRNA* values lie between 350 bp and 2,500 bp and 90% of all *CR* values are between 120 bp and 1,600 bp. For Dataset B, the *Leader* profile also forms a broad shoulder, but the *3'UTR* distribution is more skewed toward the smaller sizes. Both variables seem to have a threshold at the smaller end of the distribution, *Leader* at about 30 bp (with only 9 bp, *RP49* has the smallest leader) and *3'UTR* at about 50 bp (no *3'UTR* is smaller than 40 bp). Among the longer elements is found the leader of *Ubx* (966 bp) (excluded from the data in Fig. 34.1), which may contain a functional open reading frame (the leader associated with this secondary open reading frame is only 12 bp). The *3' UTR* of some *Ubx* and *bsg* transcripts are also outside the size normal range at approximately 2,100 and 2,200 bp, respectively.

## Size Correlations

When regression analyses were applied to Dataset A, significant correlations were observed for several pairs of variables (*3'UTR* vs *CR*, *Leader* vs *3'UTR*, etc.). However, many of these correlations were probably due to the inclusion of multiple members of the same family of transcripts. When the analysis was carried out using Dataset B, most of the correlations disappeared; the exceptions are as follows (Table 34.2):

1. There was a highly significant correlation ( $p < 0.001$ ) between *Leader* and *3'UTR*. Even when a single representative from each family of transcripts was considered, 31% of the variability in *Leader* was associated with changes in *3'UTR* ( $r^2 = 0.31$ ) (Fig. 34.2).

2. *Leader* ( $r^2 = 0.23$ ), *CR* ( $r^2 = 0.86$ ) and *3'UTR* ( $r^2 = 0.42$ ) were correlated to *mRNA*. This is as would be expected since the last variable is the sum of the first three.

TABLE 34.2. Size correlations for various pairs of genetic elements from Dataset B

	<i>Leader</i>	<i>CR</i>	<i>3'UTR</i>	<i>Exon1</i>	<i>Exon2</i>	<i>LastExon</i>	<i>mRNA</i>	<i>Intron1</i>	<i>Intron2</i>
<i>Leader</i>		NS	***	NS	NS	**	***	*	NS
<i>CR</i>			NS	NS	NS	***	***	*	NS
<i>3'UTR</i>				NS	NS	***	***	NS	NS
<i>Exon1</i>					NS	NS	NS	NS	NS
<i>Exon2</i>						NS	NS	NS	NS
<i>LastExon</i>							***	*	NS
<i>mRNA</i>								*	NS
<i>Intron1</i>									NS
<i>Intron2</i>									

The significance of each correlation is indicated by asterisks: \*,  $p < 0.05$ ; \*\*,  $p < 0.01$ ; \*\*\*,  $p < 0.001$ ; NS, indicates that the correlation is not significant. *Exon1*, *Exon2* and *Intron2* are not correlated with any of the variables. *Exon1* is not correlated with the number of exons either.

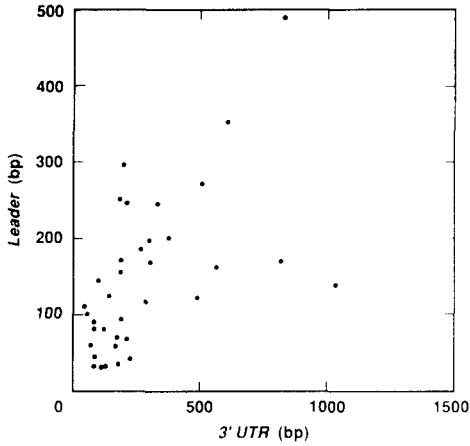


FIG. 34.2. Plot of leader size as a function of 3' UTR size for Dataset B (*Ubx* was excluded). Regression analysis is actually not permissible on the raw data because there is lack of variance homogeneity. To obviate this problem a logarithmic transformation was applied and significant correlation was observed between the transformed variables.

**Introns and Exons**

*The Number of Introns*

Fig. 34.3 shows the frequency distribution of transcripts according to the number of introns. For genes with 0 to 3 introns, there was no statistically significant correlation between number of introns and *Leader*, *CR*, *3'UTR* or

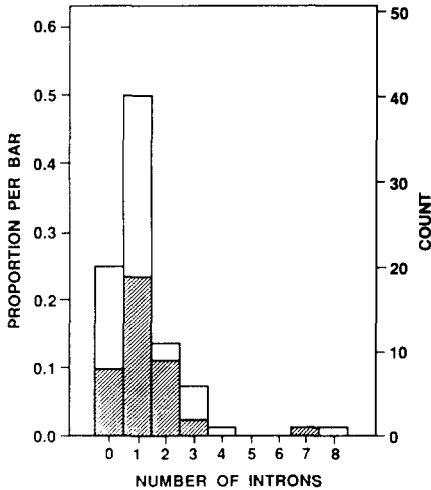


FIG. 34.3. Frequency distribution of transcripts classified according to the number of introns. See Fig. 34.1 legend.

TABLE 34.3. The size of the leader, the coding region, the 3' UTR and the mRNA in genes with various number of introns

	Number of introns			
	0	1	2	3
<i>Leader</i>	125 (20, 24)	108 (40, 15)	173 (11, 45)	144 (6, 25)
<i>3'UTR</i>	203 (17, 24)	222 (31, 32)	338 (10, 90)	281 (6, 113)
<i>CR</i>	881 (20, 129)	880 (40, 92)	1,164 (10, 166)	837 (6, 275)
<i>mRNA</i>	1,167 (17, 142)	1,306 (31, 142)	1,647 (10, 234)	1,244 (6, 336)

Mean size in bp. Numbers in parentheses indicate the number of observations and the standard error of each mean.

*mRNA* (Table 34.3). There was no correlation either between number of introns and exon sizes.

### The Size of Exons

In order to study the size distribution of exons, the last exons of all the genes were classified in a single category. The remaining exons were classified as exon 1, exon 2, exon 3, etc., starting at the 5' end; the size of the corresponding exons are designated *LastExon*, *Exon1*, *Exon2*, and *Exon3*. No significant differences were found among *Exon1*, *Exon2* and *Exon3*; meaningful comparisons among higher numbered exons were not possible because they are so few in numbers. *Exon1*, *Exon2* and *Exon3* (*UpstreamExons*), however, are significantly smaller than *LastExon*. The frequency distribution of *UpstreamExons* is shown in Fig. 34.4; the most frequent size class, is between 50 and 150 bp. *LastExon* shows a much broader distribution (Fig. 34.5).

In order to evaluate the frequency with which leader introns occur, a plot was prepared of the frequency distribution of genes according to the position of the first intron. As Fig. 34.6 shows, the distribution is fairly uniform around the AUG codon; i.e., there is no obvious cluster of genes possessing a leader intron. It would appear that there is a preferred location for the first intron in the neighborhood of the AUG codon, and whether it occurs to its right or to its left is a question of chance. For Dataset A, the position of the first intron is centered around the origin of translation, with more than 50% of transcripts having the first intron within 50 bp on either side of the AUG. For dataset B, however, the peak is not quite so sharp, and it is centered 50 bp downstream of the AUG codon. This preference for the first intron to be near the translation initiation site may be a simple coincidence of the average sizes of leaders and first exons, or it may be determined by certain sequence characteristics of that region. That the first explanation is most likely the correct one is suggested by

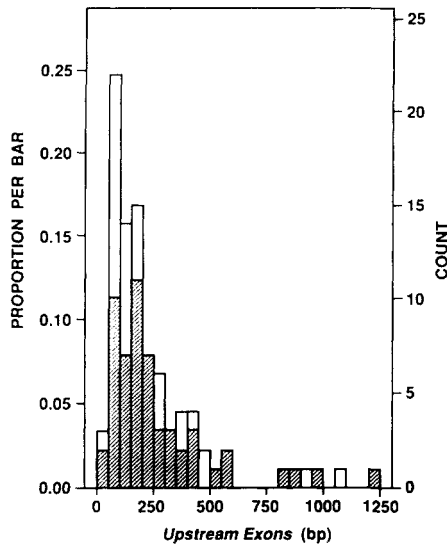


FIG. 34.4. Frequency distribution of upstream exons classified according to size. See Fig. 34.1 legend. *Ubx* was excluded.

	N	Min.	Max.	Mean	St. Dev.
Dataset A	89	22	1,245	245	240
Dataset B	55	22	1,245	264	242

the fact that leader introns seem to be more common among genes with longer leaders (Table 34.4).

### *The Size of Introns*

The size distribution of introns appears to be uniform across the various classes of introns (intron 1, intron 2, etc.), and values were pooled for Fig. 34.7A and B: 47% of all introns fall in the size class 50–75 bp; and 24% are between 60 and 70 bp. However, introns that are many thousands of bp long also occur, as in the case of *Ubx*.

*Size Correlations* Dataset B was used to estimate correlations between various pairs of variables with the following results (Table 34.2):

1. *Exon1* and *Exon2* were independent of the size of the mature mRNA, but *LastExon* was highly correlated to *mRNA* ( $r^2 = 0.64$ ).

2. As might have been expected from the *LastExon/mRNA* correlation, *CR* ( $r^2 = 0.55$ ) and *3'UTR* ( $r^2 = 0.22$ ) were correlated with *LastExon*. *Leader* was also correlated with *LastExon* ( $r^2 = 0.32$ ). This might not have been expected except for the observation that *Leader* was correlated with *3'UTR*, as was mentioned in the previous section.

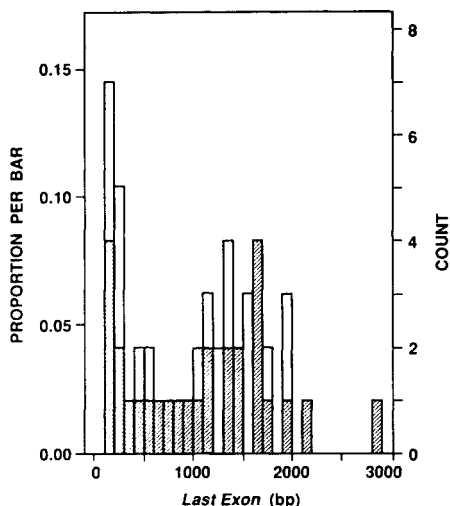


FIG. 34.5. Frequency distribution of the last exons classified according to size. See Fig. 34.1 legend. *Ubx* and *bsg* II were excluded.

	N	Min.	Max.	Mean	St. Dev.
Dataset A	50	124	2,856	1,046	713
Dataset B	39	124	2,856	1,109	698

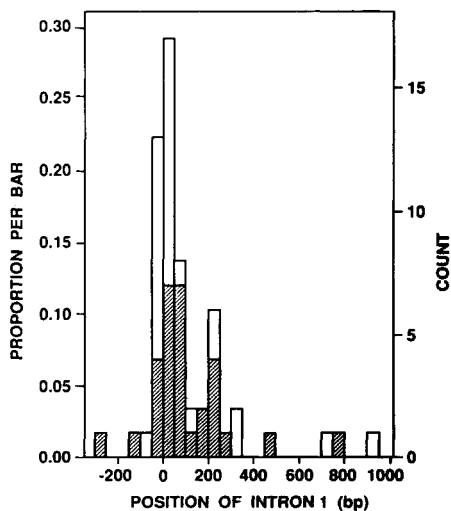


FIG. 34.6. Frequency distribution of transcripts classified according to the position of the first intron. Position 0 marks the AUG codon. See Fig. 34.1 legend. *Ubx* was excluded.



TABLE 34.4. Genes in dataset B classified by the size of intron 1

<i>Gene</i>	<i>Leader</i>	3'UTR	EXON1	INTRON1	LASTEX	mRNA	CR	<i>X(INT1)</i>	<i>EXON &gt; 1</i>	<i>INTRON &gt; 1</i>
Adh p	70	173	169	65	440	1,014	771	99	405	70
CecA2	81	81	180	58	174	354	192	99		
Ddc-Cs	353	605	53	62	1,643	1,696	738	- 300		
Ddc-DoxA	90	82	254	61	1,403	1,657	1,485	164		
eve	94	191	233	71	1,183	1,416	1,131	139		
Fcs3C	111	41	568	73	218	786	654	457		
Hspg2	60	69	181	67	124	465	336	121	160	72
JanB	100	56	152	58	146	579	423	52	156, 125	57, 61
Lcp1	42		54	64			393	12		
Pgd	35	178	43	75	1,392	1,659	1,446	8	224	1,419
Rp49	9		102	59			402	93		
S19	45	86	60	89	593	653	522	15		
S36	31	112	79	91	925	1,004	861	48		
Sgs5	33	129	301	56	164	653	492	268	188	60
Sryb	144	99	217	68	1,082	1,299	1,056	73		
Yp3	59	168	264	62	843	1,490	1,260	205	383	72

Amy	33	83				1,601	1,485			
ASC-sc	117	283				1,438	1,038			
CytC1	68	212				607	327			
Hsp22	251	181				957	525			
Hsp70A7d	246	210				2,388	1,932			
Hspg3	168	301				979	510			
Sry a	43	226				1,862	1,593			
Vm26A1	81	122				629	426			
Act5C I	155	184	147	1,667	1,413	1,560	1,131	-8		
bcd	169	817	334	559	1,102	2,471	1,485	165	76, 959	40, 513
bsg25D I	296	198	528	776	1,947	2,720	2,226	232	245	1,168
Ddc I	197	298	191	869	1,646	1,923	1,428	-6	86	1,029
Ddc-Cc	200	376	401	360	787	1,188	612	201		
EF-1AF2	138	1,030	22	1,245	1,390	2,558	1,389	-116	87, 853, 206	450, 456, 78
ftz	70		827	150			1,242	757		
h alpha1	491	830	590	1,021	1,649	2,335	1,014	99	96	136
hb p	161	561	144	283	2,856	3,000	2,277	-17		
Kr(I)	185	265	222	372	1,629	1,851	1,401	37		
kni	271	507	271	733	1,719	2,068	1,290	0	78	214
Mtn	124	140	146	265	241	387	123	22		
otu2	122	486	118	537	663	3,045	2,436	-4	233, 98, 155, 396, 137, 1,245	62, 67, 57, 53, 583, 68
prd	245	330	312	356	2,105	2,417	1,842	67		
y	171	188	409	2,719	1,576	1,985	1,626	238		

X(INT1) indicates the position of the first intron relative to the translation initiation site. EXON > 1 and INTRON > 1 contain the values of all exons and introns between the first and last ones. The top panel includes Class I genes, the bottom panel, Class II and the middle panel, Class III.

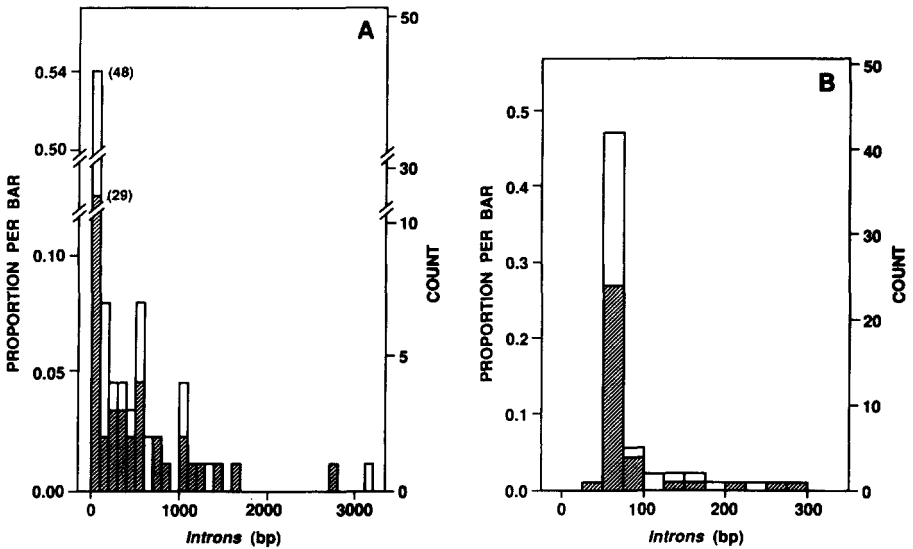


FIG. 34.7. Frequency distribution of introns classified according to size. See Fig. 34.1 legend. Panel B includes introns between 0 and 300 bp only. *Ubx* was not included.

3. Surprisingly, *Intron1* (the size of intron 1) had a significant, if not very strong, correlation with the size of several other elements. Naturally, some of these multiple associations may not have been independent of each other; i.e., *Intron1* might be correlated to *mRNA* because it was correlated to *LastExon*, which is in turn a determinant of *mRNA*.

### Classes of Genes

The correlation between *Intron1* and *mRNA* ( $r^2 = 0.19$ ) was not due to a smooth relationship between the two variables but rather to the fact that all introns 1 of small size were associated with mRNAs smaller than 1.7 kb while most larger introns were associated with mRNAs of more than 1.7 kb (Fig. 34.8). The same phenomenon explains the correlations between *Intron1* and the other variables and the correlation between *Leader* and *3'UTR*. In other words, all of the unexpected size correlations that were found are ascribable to the fact that genes with introns can be classified into two groups: class I (those having a first intron of less than 100 bp) and, class II (those having a first intron of more than 100 bp). As a group, class I genes have significantly smaller *Leader*, *3'UTR* and *CR* than class II genes. Within each class, none of the size correlations exist (Fig. 34.9, Table 34.4 and Table 34.5).

What is the biological or molecular significance of two such distinct classes of genes? One possibility is that some of the larger introns may contain segments important for the control of gene expression. Several instances of regulatory

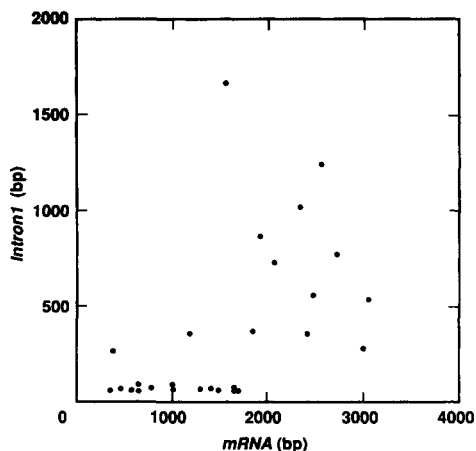


FIG. 34.8. Plot of intron 1 size as a function of mRNA size for Dataset B. Regression analysis is actually not permissible on the raw data because there is lack of variance homogeneity. To obviate this problem a logarithmic transformation was applied, and a significant correlation was observed between the transformed variables.

TABLE 34.5. The size of the leader, the coding region and the 3' untranslated region in three classes of genes

	Class I	Class II	Class III
Leader	85 (16, 80)	200 (15, 100)	126 (8, 87)
CR	760 (16, 408)	1,435 (15, 614)	980 (8, 621)
3'UTR	148 (14, 140)	444 (14, 278)	202 (8, 74)

Mean size in bp. Numbers in parentheses indicate the number of observations and the standard error of each mean. Analysis of variance indicates that in each case, Class II means are significantly different from Class I and Class III means ( $p = 0.05$ ).

sequences in transcribed but non-coding regions of genes have been documented (see, for example, *bcd*, *ftz*, *Hsp70*, *Pgd*, *Ubx*). But why should the presence of such regulatory elements be associated exclusively with larger coding regions? Alternatively, the explanation may rest entirely with the mechanics of mRNA transcription and processing (see Chapter 35). Another possible explanation, albeit one that does not seem to be borne out by the data, is that genes within each class are more closely related to one another than to genes of the other class and that the correlations presented here are just a consequence of "family resemblance".

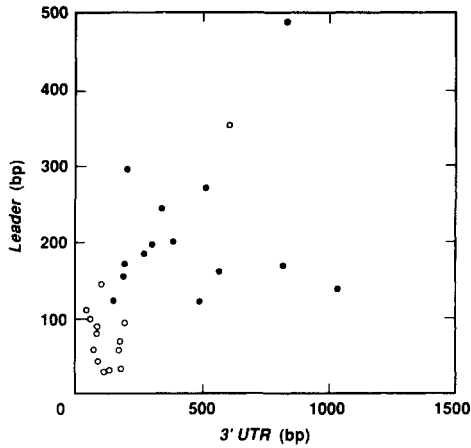


FIG. 34.9. Plot of leader size as a function of 3' UTR size for Class I (○) and Class II (●) genes in Dataset B. Regression analysis within each class showed no significant correlation between the variables.

In addition to the two classes of genes treated heretofore, there is a third class, those without introns. The mean values for *Leader*, *CR* and *3'UTR* in intronless genes fall in between the values for class I and class II genes. Statistically, however, those values are significantly smaller than the values for class II genes, and not significantly different from the values for class I (Tables 34.4 and 34.5).

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## Messenger RNA splicing signals in *Drosophila* genes

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This chapter provides a general description of introns in *Drosophila* genes, with emphasis on the genetic information responsible for the correct specification of boundaries between introns and exons. The problem of locating introns within unannotated DNA sequences is posed by any large genomic sequencing project, and provides a perspective for discussing the information that specifies their removal. I want to stress, however, that there may not be a single set of rules that can identify all introns in all tissues. Certainly, it has become clear that the rules for locating introns will differ between species, such as flies and humans, in different taxonomic classes. Here, I will attempt to describe in general terms both what is known about how introns are recognized by the splicing machinery, and how an investigator might go about identifying introns within the sequence of his favorite *Drosophila* gene. Ultimately, such searches will be carried out by computer. Most current software, however, is designed specifically or primarily for species other than *Drosophila* (one exception is the program GM (Fields and Soderlund 1990), which accepts organism-specific consensus matrices and codon asymmetry tables). I am currently developing computational applications of the ideas described here, and interested readers are encouraged to consult current releases of the electronic *Drosophila Information Newsletter*.

### **The Mechanism of Splicing**

To understand how genetic information specifies the removal of introns, one must understand splicing at the level of biochemical mechanism. To date, the biochemistry of splicing has been studied in extracts from HeLa cells or yeast

(reviewed by Smith et al. 1989; Green 1991; Guthrie 1991). However, *Drosophila* is becoming increasingly important to the study of messenger RNA splicing, primarily because of extremely promising genetic systems bearing on the regulation of alternative splicing (Laski et al. 1986; Boggs et al. 1987; Chou et al. 1987; Zachar et al. 1987; Bell et al. 1988; Nagoshi et al. 1988; Pongs et al. 1988; Schwartz et al. 1988; Siebel and Rio 1989; Collier et al. 1990; Geyer et al. 1991; Pret and Searles 1991; McAllister 1992; Steinhäuser and Kalfayan 1992; Hazelrigg, unpublished results). Extracts from *Drosophila* cells or embryos that are capable of accurate and efficient removal of introns from RNA substrates have been described (Rio 1988; Hodges and Bernstein 1992; Guo et al. 1992), and are certain to be used increasingly. However, the HeLa *in vitro* system will just as certainly continue to provide the biochemical paradigm, and most of the information in this section pertains to results derived using extracts from HeLa cells.

### The Chemistry of Splicing

The removal of introns from messenger RNA precursors occurs in a series of two cleavage–ligation reactions, each involving transesterification at a splice site phosphate (Fig. 35.1A). Thus, messenger RNA splicing resembles the

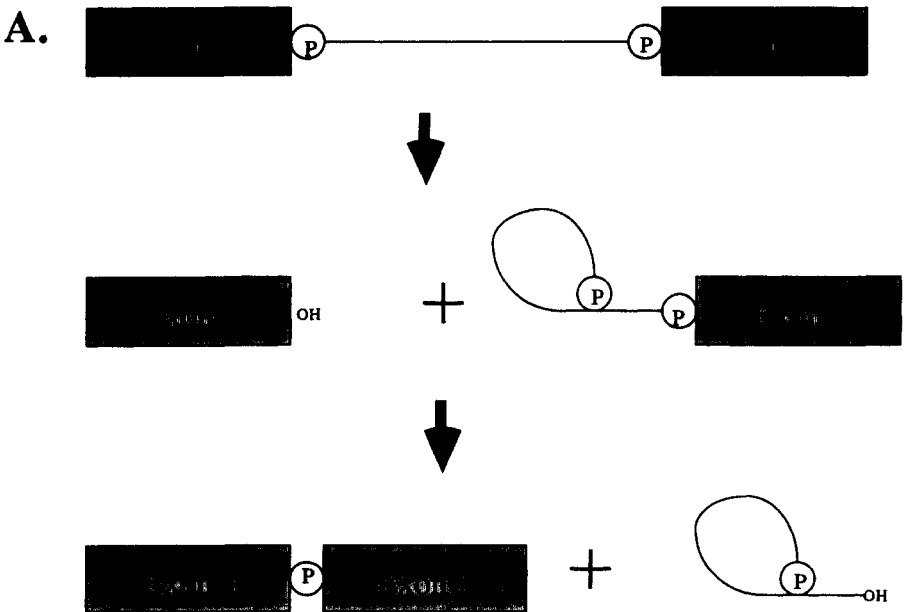


FIG. 35.1. Overview of the splicing mechanism. (a) Each of the chemically distinct steps in the splicing process is indicated. The first phosphotransfer reaction joins the 5' phosphate of the intron to a 2' hydroxyl group within the intron, resulting in a free upstream exon and a lariat intermediate. The second step of the splicing reaction joins the now free 3' hydroxyl of the upstream exon to the phosphate at the 3' splice site. (continued)

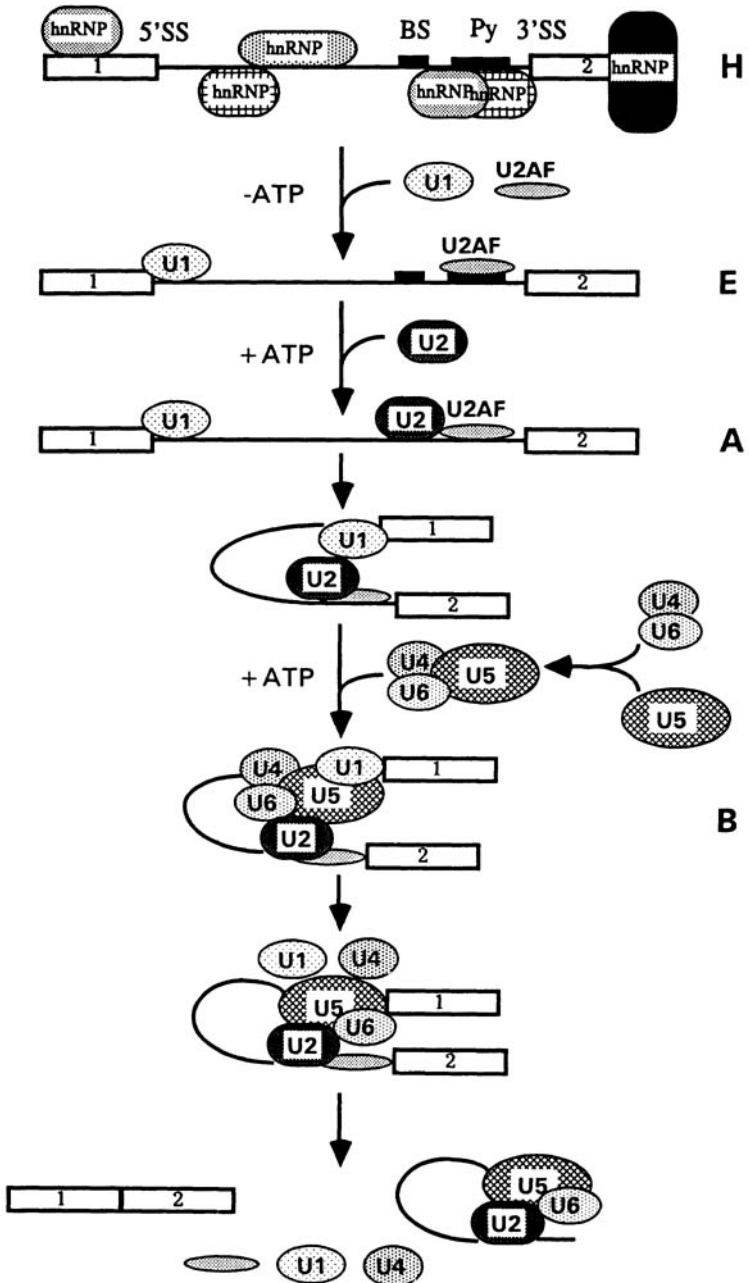
**B.**

FIG. 35.1 (continued). Overview of the splicing mechanism. (B) Spliceosome assembly involves the ordered addition of snRNPs and protein factors. The generally recognized series of steps in HeLa nuclear extract spliceosome assembly are shown and the complexes named (see text).



splicing of both Group I and Group II introns of the self-splicing type. In mRNA splicing and Group II splicing (but not Group I splicing), the phosphate at the 5' splice site reacts with a 2' hydroxyl group within the intron, resulting in a free upstream exon and a lariat that consists of nucleotides from the intron and the downstream exon. In the splicing of Group I introns, exemplified by the *Tetrahymena thermophila* ribosomal RNA intron, the 5' splice site phosphate reacts with a 3' hydroxyl group on a guanosine nucleotide, and no lariat is formed. These three classes of intron are similar in that the second step is carried out by attack of the now free 3' hydroxyl group of the upstream exon with the phosphate at the 3' splice site. Both steps of pre-mRNA splicing proceed with inversion of configuration at phosphorus (K. L. Maschoff and R. A. Padgett, M. J. Moore and P. A. Sharp, personal communication), which constitutes evidence for a concerted transesterification reaction, as had been previously described for Group I self-splicing introns (McSwiggen and Cech 1989; Rajagopal et al. 1989). The basic similarity between pre-mRNA splicing and splicing in which the intron participates in the catalysis of the splicing reaction has led to the speculation that pre-mRNA splicing is essentially RNA-catalyzed (Cech 1986; Guthrie 1991; Sharp 1991). It is supposed that in the case of pre-mRNA splicing the catalytic RNA is one or more of several small nuclear RNAs (snRNAs) that assemble onto nascent intron-containing transcripts as part of a large (40S–60S) complex of RNAs with at least 30 proteins known as the spliceosome.

### *The Spliceosome*

The spliceosome contains the pre-mRNA and a number of associated factors. The best understood of these factors are snRNPs (small ribonucleoproteins), complexes of one or more snRNAs and associated proteins. The most abundant spliceosomal snRNAs (U1, U2, U4, U5 and U6) are present in RNPs containing a number of common proteins recognized by antibodies from patients with a number of autoimmune diseases (for reviews of snRNPs and snRNP proteins, see Paterson et al. 1991; Birnstiel 1988). All of these RNAs carry a trimethyl guanosine cap at their 5' ends, with the exception of U6, which has a monomethyl cap. U1 and U2 snRNPs, each with a single U snRNA, are most abundant, and have well-defined roles in the splicing process (see Fig. 35.1 and the discussion below). U4 and U6 are normally found associated in a single snRNP, loosely associated with the U5 snRNP to form a tri-snRNP (Behrens and Lührmann 1991). Both the protein and RNA components of these U snRNPs are highly conserved. In particular, *Drosophila* U RNAs are highly conserved in sequence (Mount and Steitz 1981; Saba et al. 1986; Das et al. 1987; Lo and Mount 1991; see Mylinski et al. 1984; Guthrie and Patterson 1988; and Reddy and Busch 1988; for overviews of snRNA conservation). Furthermore, it is generally possible to make a one-to-one correspondence between HeLa cell and *Drosophila* snRNP proteins on the basis of mobility and antigenicity (Paterson et al. 1991), and those proteins involved in splicing whose sequences have been determined in *Drosophila* as well as in vertebrates are also highly conserved (Mancebo et al. 1990; Harper et al. 1992; Zahler et al. 1992).

A considerable number of specific interactions among various components of the spliceosome and the splicing substrate occur prior to the first step of splicing. Green (1991) divides spliceosome assembly into four steps: the U1 snRNP-binding reaction, the U2 snRNP binding reaction, the entry of the U4/U5/U6 tri-snRNP and the loss of U4 snRNP from the spliceosome (Fig. 35.1). A number of intermediates in this process can be separated on non-denaturing gels (Konarska and Sharp 1987) or on sizing columns (Michaud and Reed 1991), and some of the intermediate complexes have been named (Fig. 35.1). Prior to its assembly with spliceosomal components, the pre-mRNA can be found associated with heterogeneous nuclear ribonucleoprotein (hnRNP) proteins both *in vivo* (Dreyfuss 1986) and *in vitro* (Bennett et al. 1992). This early complex, known as the H complex, contains different hnRNP proteins on different substrates. A second complex, known as the E complex, consists of stably bound U1 snRNP, and can assemble in the absence of ATP (Michaud and Reed 1991). Subsequent addition of the U2 snRNP (which associates with the branchpoint) requires ATP and results in the formation of the A complex. A pre-existing complex of U4, U5 and U6 is added to the A complex to form the B complex. Then, the U4 snRNP (without U6) is either lost from the spliceosome (Lamond et al. 1988; Yean and Lin 1991) or destabilized (Blencowe et al. 1989), and splicing follows. Splice site recognition by snRNPs has recently been reviewed by Steitz (1992).

### *Recognition of 5' Splice Sites*

A 5' splice site that conforms to the consensus sequence  $\text{MAG}\underline{\text{GU}}\text{RAGU}$  (M = A or C; R = A or G), within which the underlined GU dinucleotide is invariant, is generally required for splicing (Aebi et al. 1986; Green 1986; Smith et al. 1989). The 5' splice site is recognized by the U1 snRNP (Mount et al. 1983; Black et al. 1985) via base-pairing with the 5' end of U1 RNA (Zhuang and Weiner 1986; Séraphin et al. 1988; Siliciano and Guthrie 1988), as originally proposed by Lerner et al. (1980) and by Rogers and Wall (1980). The 5' splice site is probably also recognized by additional factors (Siliciano and Guthrie 1988; Bruzik and Steitz 1990; Seraphin and Rosbash 1990; Stolow and Berget 1991), including the U5 snRNP (Newman and Norman 1991), which appears to recognize the exonic portions of both the 5' and the 3' splice sites (Newman and Norman 1992). The G at intron position 1 is required for the second step of splicing as well as for the first; mutations in this position can result in accumulation of lariat intermediates in both yeast (Newman et al. 1985; Vijayraghavan et al. 1986) and mammalian (Aebi et al. 1986) systems. Thus, it appears that nucleotides at the 5' splice site are recognized multiple times in the course of a single splicing event, and this may help to explain the observation that consensus sequences for the 5' splice site are highly conserved between species (Mount 1982; Shapiro and Senapathy 1987; Jacob and Gallinaro 1989; Fields 1990; Mount et al. 1992). It is of particular interest to this discussion that the *Drosophila* matrix is remarkably similar to those obtained from mammalian introns (Table 35.1).

TABLE 35.1. 5' splice site sequences

*Drosophila* (frequencies, as percentages).

	-5	-4	-3	-2	-1	1	2	3	4	5	6	7	8
A	33	34	37	52	9	0	0	60	71	9	11	39	27
C	24	21	29	15	8	0	0	1	9	2	14	13	21
G	14	23	15	11	71	100	0	35	9	82	6	19	20
T	29	22	19	21	12	0	100	4	11	6	68	29	32
consensus:			M	A	G	<u>G</u>	<u>T</u>	R	A	G	T	W	

Total (all species, dominated by mammals).

	-3	-2	-1	1	2	3	4	5	6
A	32	60	9	0	0	59	71	7	16
C	37	13	5	0	0	3	9	6	16
G	18	12	79	100	0	35	11	82	18
T	13	15	7	0	100	3	9	6	50
consensus:	M	A	G	<u>G</u>	<u>T</u>	R	A	G	T

*Drosophila* 5' splice site scoring table. Scores were calculated according to Hertz et al. (1990).

	-3	-2	-1	1	2	3	4	5	6	7
A	0.6	1.1	-1.4	-5.7	-5.7	1.3	1.5	-1.4	-1.1	0.6
C	0.2	-0.7	-1.6	-4.7	-4.7	-4.1	-1.4	-3.1	-0.8	-0.9
G	-0.7	-1.1	1.5	2.0	-5.7	0.5	-1.4	1.7	-1.9	-0.4
T	-0.4	-0.2	-1.0	-5.7	2.0	-2.5	-1.1	-2.0	1.5	0.2

*Recognition of Branchpoints, Pyrimidine Tracts, and 3' Splice Sites*

3' Splice sites conform to the consensus sequence YAG|G and are typically found at the site of the first AG dinucleotide downstream of the branchpoint. Mammalian branchpoints fit the consensus sequence UNCRAC (in which branch formation occurs at the underlined A) and usually reside between 18 and 38 nucleotides upstream of the 3' splice site (Noble et al. 1988; Reed and Maniatis 1988; Nelson and Green 1989). Between the branchpoint and the 3' splice site is a pyrimidine-rich region. The way in which sequences at the 5' splice site, the branchpoint, the pyrimidine-rich stretch, and the 3' splice site act together in mammalian splicing to specify intron boundaries has been investigated in detail and much is known of the factors that recognize these sites (Reed and Maniatis 1988; Smith et al. 1989; reviewed in Smith et al. 1989; Green 1991). The branchpoint is recognized by the U2 snRNP via base pairing (Parker and Patterson 1987; Nelson and Green 1989; Wu and Manley 1989; Zhuang et al. 1989; Zhuang and Weiner 1989). However, binding of the

U2 snRNP to the branchpoint requires a number of factors, including the U1 snRNP (Zillman et al. 1987; Ruby and Abelson 1988; Séraphin et al. 1988; Barabino et al. 1990) and U2AF, a factor that binds to the pyrimidine-rich stretch (Ruskin et al. 1988; Zamore and Green 1991).

There exists considerable evidence supporting the proposal that after a branchpoint has been selected (and possibly, but not necessarily, after the first step of splicing) a 3' splice site is selected at the first AG dinucleotide downstream of the branch. This model is supported by the result, observed in both yeast (Rymond and Rosbash 1985) and HeLa cell extracts (Smith et al. 1989), that the first step of splicing can proceed without an AG dinucleotide if certain conditions are met (see below). In particular, Reed (1989) has divided introns into two categories based on the relative importance of the branchpoint and the pyrimidine tract, and finds that a tract of 14 pyrimidines is sufficient to confer AG-independent splicing. In any event, the lack of AG dinucleotides in the region between the branchpoint and the 3' splice site (Mount 1982; Shapiro and Senapathy 1987; Gelfand 1989) is suggestive of some sort of microscanning model, as was noted very early (Mount 1982). Consistent with this, mutational analysis indeed indicates that the first AG downstream of such a branchpoint is used as the 3' splice site (Langford and Gallwitz 1983; Smith et al. 1989). In the mammalian case (Smith et al. 1989), CAG, UAG or AAG, introduced between the branchpoint and the genuine 3' splice site, were found to "capture" splicing, but GAG in the same position prevented splicing altogether, a result that is consistent with the lack of any recorded 3' splice sites with the sequence GAG.

Recently, Reich et al. (1992) observed that compensatory changes in U1 RNA can suppress mutations in the AG at the 3' splice site in *Schizosaccharomyces pombe*, indicating base pairing between U1 and the 3' splice site prior to the first step of splicing. Thus, U1 RNA interacts with both splice sites prior to the first step of splicing, at least for some introns (possibly all those introns that require the 3' splice site AG to complete the first step of splicing). This division of introns into categories based on AG-dependence can be extended to include a third category: those introns that do not require U1 at all (Bruzik and Steitz 1990). Thus, it is becoming apparent that the relative contributions of particular factors to intron recognition may vary among introns.

### Species-specificity of Splicing Signals

Although it is now clear that mRNA splicing is carried out by a universally conserved fundamental mechanism, it does not follow that there is conservation of splicing signals. In fact, both *in vivo* and *in vitro* systems splice introns derived from other phyla either inaccurately or not at all, and those interested in the expression of genes in *Drosophila* must keep in mind that there is no counterpart in *Drosophila* to the wealth of information available about splicing signals in yeast and mammalian cells. However, judicious consideration of *Drosophila*

intron sequences, the small but growing database of experimental results obtained in *Drosophila*, and selected results from other species, allows a good understanding of *Drosophila* splicing signals. In this section, I will review what is known about variation between species with respect to the nature and relative contribution of various splicing signals.

### *Exon Definition and Intron Retention*

What happens when a splice site is defective? Naively, one would think that the splice site would be ignored, resulting in retention of the intron whose excision is dependent upon that splice site (intron inclusion—Fig. 35.2). Alternatively, if there is information elsewhere that indicates that a splice should take place within any given region, then another site may be used for the splice (cryptic sites, Fig. 35.2). This result can also be explained by competition between the two sites—either alone would be sufficient to compel a splice, but the stronger site is better at recruiting factors that result in a commitment to splicing. In fact, the result of many mutations in mammalian splice sites is skipping of an entire exon that includes the affected splice site (Mitchell et al. 1986; reviewed in Robberson et al. 1990; see exon skipping, Fig. 35.2). This implies that exons, rather than introns, are recognized as a unit. Such results from mutational analyses have been used, in combination with results from the study of complex assembly *in vitro* on model substrates, including an association of U1 snRNP with the 3' half of introns (Zillman et al. 1987), to advance a theory of exon definition (Robberson et al. 1990). Exon definition implies that the productive assembly of spliceosomal components at splice sites is dependent upon the presence of functional sequences at both ends of each exon. This phenomenon has now been well documented experimentally (Talerico and Berget 1990; Grabowski et al. 1991). The strength of the 5' splice site at the 3' end of the internal exon has been shown to be critical for the efficiency of splicing in a manner that is independent of the strength of the upstream 5' splice site (Grabowski et al. 1991), implying that two U1 snRNPs, interacting with two distinct binding sites (5' splice site sequences) are critical for splicing.

*Drosophila* has relatively shorter introns (Hawkins 1988; Bingham et al. 1988; Mount et al. 1992), and relatively longer exons (Hawkins 1988; Maroni, this volume, Chapter 34), than do mammalian species, and exon definition may play a correspondingly smaller role in the determination of splicing patterns. Consistent with this, a two-intron adenovirus test substrate (an exon of 94 nucleotides flanked by an upstream intron of 120 nucleotides and a downstream intron of 89 nucleotides) reveals species-specific behavior when tested in splicing extracts from *Drosophila* and human cells. Mutation of the 5' splice site results in exon skipping in splicing extracts from HeLa cells (Talerico and Berget 1990), but intron inclusion in extracts from *Drosophila* cells (M. Talerico and S. Berget, personal communication). Intron inclusion has also been observed in response to similar mutations in a two-intron *Drosophila* substrate from the *zeste* gene assayed in *Drosophila* extracts (M. Talerico and S. Berget, personal communication). Finally, there are hints that intron inclusion may be

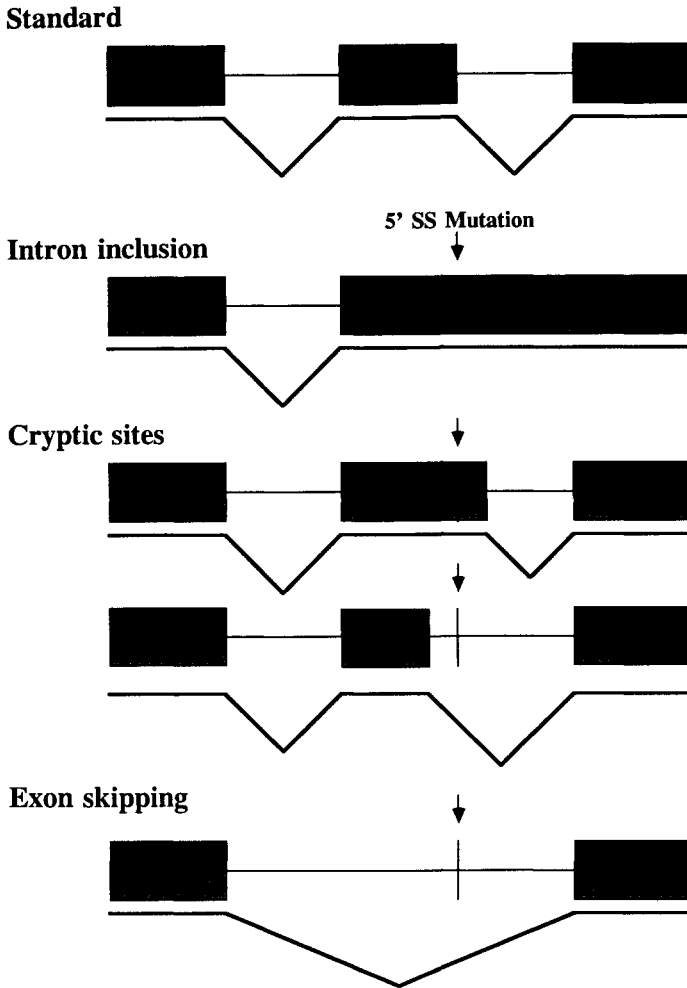


FIG. 35.2. Exon-skipping, intron inclusion, and the use of cryptic splice sites as responses to inactivation of a splice site. The splicing pattern of a typical gene segment including three exons and two introns is depicted in the top cartoon ("standard"), and altered patterns of splicing that can result from a mutation at one splice site (here the a mutation at the 5' splice site) are shown below: activation of cryptic splice sites, intron inclusion, exon skipping. Most alternative splicing can be explained in terms of one of these three responses to an inactivated (or activated) site.

accompanied by a greater stability of intron-containing RNA *in vivo*. The classical *in vivo* result of 5' splice site mutations in vertebrate systems is no RNA. In contrast, flies carrying 5' splice site mutations have been observed to accumulate intron-containing RNA (S. Wasserman, personal communication; unpublished data from the author's laboratory). These results are all consistent

with the suggestion that in *Drosophila* the intron, rather than the exon, is the unit of recognition during spliceosome assembly.

However, recognition of exons probably occurs as well. The term "micro-exon" was first applied by Beachy et al. (1985) to two, 51 nucleotide, alternatively spliced (O'Conner et al. 1988; Kornfeld et al. 1989), exons in *Ubx*. In fact, these exons are within the normal range of exon sizes (see Maroni, this volume, Chapter 34); what led to their being called microexons was their small size relative to the size of the introns flanking them. This makes them candidates for regulation of alternative splicing by regulation of exon definition/recognition, as is the case in the sex-specific autoregulation of *Sexlethal* (Bell et al. 1988), and alternative splicing of the *Drosophila* myosin heavy chain gene (Hodges and Bernstein 1992). True microexons have also been observed in *Drosophila* genes. One rather striking case is an exon of only six nucleotides that lies somewhere within 26 kb separating the first and third exons of the *invected* gene (Coleman et al. 1987). To my knowledge, the location of this microexon has never been ascertained. McAllister et al. (1992) describe two nine-nucleotide microexons whose inclusion in the *Drosophila fasciclin 1* gene is variable. In this case, the positions of the microexons have been determined (they reside within a stretch of only 2.7 kb), and the sequence flanking them provides a clue as to how exons of such a small size might be recognized. Each of the microexons is preceded by a long stretch (160 and 120 nucleotides) of sequence with reduced G content (less than 10%) and no AG dinucleotides. Thus, it is possible to propose that these microexons are recognized by formation of a complex at the microexon 5' splice site and a site greater than 100 nucleotides upstream. Once commitment to splicing (and possibly removal of the downstream exon) had occurred, microscanning (see above) could locate the appropriate 3' splice site. This model makes the experimentally testable prediction that microexons will generally use remote branchpoints.

### *Introns with High A + T Content*

Animal introns are not properly recognized in transfected plant cells (Weibauer et al. 1988). This is despite the observation that splice site consensus sequences are fairly similar between plants and animals (Brown 1986; Goodall and Filipowicz 1991; White et al. 1992). It appears that the relative A + T-richness of plant introns is critical to their proper recognition (Goodall and Filipowicz 1989), an effect that is more pronounced in introns from dicots than in introns from monocots. The upshot of considerable mutational analyses (assayed by transfection into tobacco (dicot) protoplasts) is that these cells will recognize as an intron almost any sequence that is extremely A + T-rich and is flanked by appropriate, short, consensus sequences (Goodall and Filipowicz 1991); branchpoint and pyrimidine tract sequences are not important to splicing. As would be predicted from the foregoing, deletion of intron sequences so as to move the boundary between A + T-rich and flanking sequences is sufficient to activate cryptic 3' splice sites that lie in the vicinity of the new boundary (Lou et al. 1992). *Drosophila* also has introns that are significantly richer in A + T

than are flanking exons (65% versus 48%; Mount et al. 1992), but the possible contribution of A + T content (or of critical subsequences composed of A and T) to intron recognition has not been demonstrated experimentally. In fact, a survey of base composition in introns versus exons (Csank et al. 1990) reveals that mammals are unique among species surveyed in their lack of a significant difference between introns and exons with respect to A + T content, and the yeast *Saccharomyces cerevisiae* is among species with the smallest difference in A + T content between introns and exons. Thus, a contribution of A + T content to the recognition of introns may be general, but poorly described because of the choice of experimental organisms by the pre-mRNA splicing community.

### *Variation in Intron Size*

Hawkins (1988) and Bingham et al. (1988) were the first to note that there are considerable differences between *Drosophila* and other species (notably mammals) with respect to the size of introns. Specifically, approximately half of all sequenced *Drosophila* introns are less than 80 nucleotides, with a modal length between 60 and 65 nucleotides (Mount et al. 1992). Thus, the typical *Drosophila* intron is smaller than all but a few mammalian introns (Hawkins 1988; Ge et al. 1990), and shorter than the length of approximately 80 nucleotides generally required for efficient splicing in mammalian cells (Wieringa et al. 1984; Ruskin et al. 1985). This strongly suggests species specificity in the recognition of introns (as opposed to the idea that, although smaller than most mammalian introns, the many short *Drosophila* introns would nevertheless be recognized by a mammalian splicing system). An experimental demonstration of species specificity with respect to size requirements has been obtained recently by Guo et al. (1992), working with a short (74 nucleotide) *Drosophila* intron that was properly recognized in homologous (*Drosophila* Kc cell), but not heterologous (HeLa cell) nuclear extracts. An even more extreme situation exists in *C. elegans*, where intron lengths of less than 50 nucleotides are common (Blumenthal and Thomas 1988). Consistent with these observations, a *C. elegans* intron of 53 nucleotides was efficiently spliced in HeLa cell nuclear extracts only when expanded to 84 nucleotides (Ogg et al. 1990).

The distance between the 5' splice site and the branchpoint and the distance between the branchpoint and the 3' splice site are presumably subject to different constraints, so it is of interest to know in which portion of the intron species-specific length preferences reside. The distribution of sequences that resemble the branchpoint, for example CTAA, within small introns indicates that distances between the branchpoint and the 3' splice site in *Drosophila* are very similar to those found in mammals, but 5' splice site to branchpoint distances are often shorter (typically 38–43 nucleotides; Mount et al. 1992). In the case of the *white* second intron, the experimentally observed branchpoint used by Kc cell extracts is 42 nucleotides from the 5' splice site (Guo et al. 1992). This 5' splice site to branchpoint distance is considerably less than that in mammalian introns. For example, manipulation of this distance in the small-t



intron (Fu et al. 1988) indicated that the wild-type distance in that case (48 nucleotides) is minimal; an intron with a distance of 46 nucleotides showed no splicing, while a distance of 53 nucleotides showed significantly increased splicing. In another study, Smith and Nadal-Ginard (1989) found 51 nucleotides too short, but 59 sufficient. In addition, a distance of 49 nucleotides between the 5' splice site and branchpoint was found too short to allow U4,U5,U6 tri-snRNP binding to an adenovirus E1A pre-mRNA *in vitro* (Himmelspach et al. 1991).

Intriguingly, although the distance between branchpoint and 3' splice site is conserved between mammals and fruit flies, it is not conserved in all species with small introns. For example, Prabhala et al. (1992) examined sequence data for introns from *Schizosaccharomyces pombe*, and found that over half of *S. pombe* introns appear to have less than 10 nucleotides between the branchpoint and the 3' splice site. In the nematode *C. elegans*, there is a modal intron size of roughly 45 nucleotides (Blumenthal and Thomas 1988), smaller than all but the very smallest *Drosophila* introns. Because no *C. elegans* branchpoint consensus can be discerned, it is unclear which half of the intron has altered size constraints.

The smallest intron in the *Drosophila* data set examined by Mount et al. (1992) is 51 nucleotides. However, a 36 nucleotide intron is described in the *vasa* gene by Lasko and Ashburner (1988), and the set of genes in this atlas contains a 40 nucleotide intron in the *bicoid* gene. In the latter case, the unusually small size may be due to overlap between the branchpoint and 3' splice site (CTTATCAG|A incorporates both the CTAAT branchpoint consensus and the YAG|G 3' splice site consensus). The distance between the putative branchpoint and the 3' splice site in this case would be only four nucleotides, which is an extremely short distance. However, there are a number of *Drosophila* introns with alternative 3' splice sites bearing a similar relationship. One site is very close to a putative branchpoint and the other is about 15 nucleotides further on. In the *bcd* example, the downstream 3' splice site corresponds to a short intron that is normal in every respect save one—the occurrence of an AG dinucleotide relatively close to the 3' splice site, at position -15. Other examples of this arrangement are the two 3' splice sites in the *Sxl* male-specific exon (Bell et al. 1988), where a branchpoint consensus is 10 nucleotides upstream of one 3' splice site and 28 nucleotides upstream of another; and Hrb98DE, where the corresponding distances are 5 and 17 nucleotides (Haynes et al. 1990).

The short but relatively constant distance between the 5' splice site and branchpoint of small *Drosophila* introns raises the possibility of direct contact between complexes at the 5' splice site and at the branchpoint. A model that incorporates the information summarized here is described in Fig. 35.3. Fig. 35.3A depicts the mechanism of splicing of large introns in *Drosophila* and follows information from splicing in HeLa cells presented in greater detail in Fig. 35.1. Fig. 35.3B presents a model involving direct interaction between a complex at the 5' splice site and a complex at the 3' splice site, indicating how accommodation to a shorter distance between the 5' splice site and branchpoint

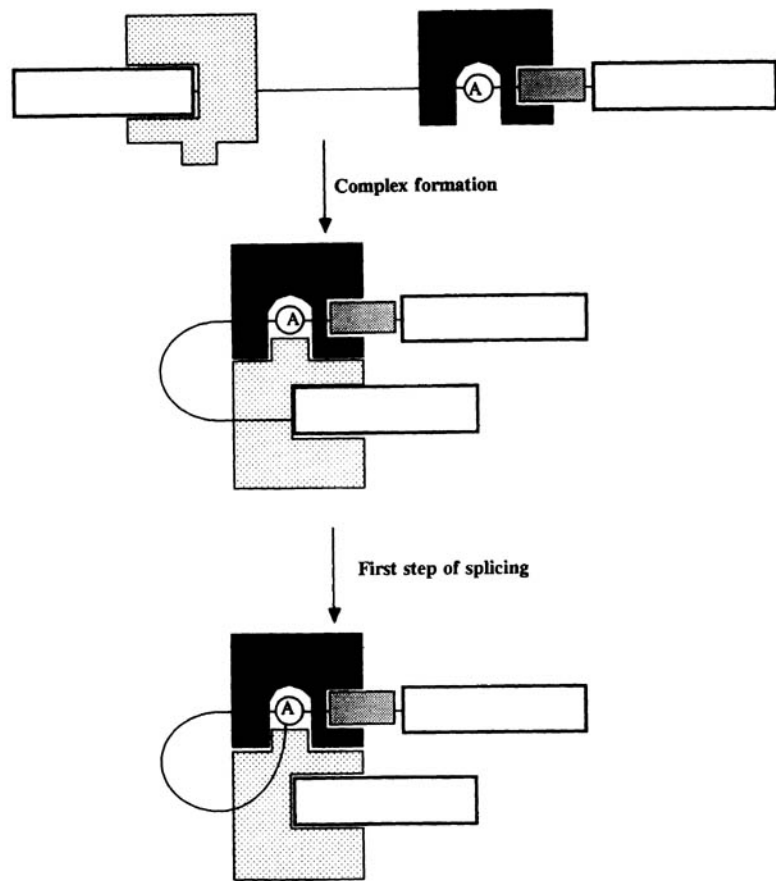
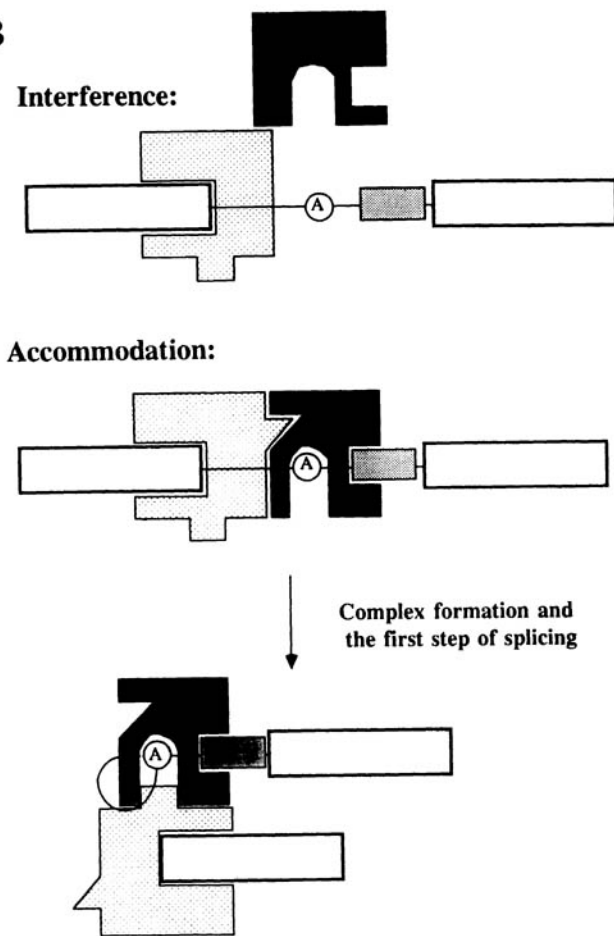
might lead to a novel, and possibly species-specific, interaction between complexes at the two sites.

### *Variation in Branchpoint Recognition*

Most mammalian introns are not spliced in yeast cells (Beggs et al. 1980; Langford and Gallwitz 1983). This is due, at least in part, to the fact that yeast introns almost always use the precise sequence UACUAAC as a branchpoint, and this sequence is the primary determinant of yeast 3' splice site selection (Jacquier et al. 1985; Newman et al. 1985; Parker and Guthrie 1985). In contrast, the branchpoint sequence of mammalian introns has greater flexibility (Keller and Noon 1984; Ruskin et al. 1984; Zeitlin and Efstratiatis 1984; Konarska et al. 1985; Reed and Maniatis 1988; Nelson and Green 1989) and the pyrimidine-rich stretch is relatively more important (Frendeway and Keller 1985; Reed 1989). This dichotomy is nicely illustrated by differences in the sequences required for the first step of the splicing reaction to take place in the absence of the second. In yeast, UACUAAC is sufficient (Rymond and Rosbash 1985), while the mammalian splicing machinery demands a significant stretch of pyrimidines for AG-independent splicing (Reed 1989; Smith et al. 1989).

Branchpoint recognition has not been carefully examined in *Drosophila*. In one case, *in vitro* splicing was observed to proceed, using a non-consensus branchpoint, when the wild-type site was mutated (Guo et al. 1992). In addition, the pyrimidine-rich stretch that is so prominent in the literature on mammalian intron splicing (cited above) is absent in a large fraction of *Drosophila* introns, implying that branchpoint recognition must occur in the absence of a significant pyrimidine tract. For example, 49% of short *Drosophila* introns lack even a single stretch of 12 nucleotides including 10 pyrimidines in the region between -50 and -3 relative to the 3' splice site. In the -26 to -5 region, the average content of pyrimidines in a mammalian intron is 72%. In *Drosophila*, this number is 66%. Perhaps more striking is the observation that A residues are actually more common than C residues (25 versus 22%). This region is high in T (44%) and low in G (9%). In fact, 78% of all *Drosophila* introns in that data set have TTT somewhere in the region between -35 and -3. However, there are introns with few Ts in the region, but no introns that are G-rich in this region. Counting the number of G residues in a 25 nucleotide window adjacent to 3' splice sites leads to rather striking results; only three out of 205 3' splice sites have more than five Gs in this region, and the average intron has less than three (Fig. 35.4A). Of 205 3' splice sites in the data set, only seven have more Gs in an adjacent exonic 25 nucleotide window than in this window. Thus, this region carries a lot of information that can be used to predict the location of a 3' splice site.

How is this region recognized by the splicing machinery in *Drosophila*? U2AF recognizes the pyrimidine tract and promotes recognition of the branchpoint by U2 in mammalian splicing, and U2AF activity has been found in *Drosophila* extracts (Zamore and Green 1991). It is possible that pyrimidine-poor *Drosophila* introns are indeed recognized by U2AF; the *Drosophila*

**A****B**

homologue of U2AF could simply have an altered sequence specificity, and bind to G-poor rather than pyrimidine-rich regions. Another possibility is that other factors are involved.

### A Strategy for the Identification of *Drosophila* Introns

This section is written for the *Drosophila* geneticist or developmental biologist who has just sequenced his favorite gene, but has not yet isolated cDNAs, or would like to assess the likelihood that additional spliced RNAs exist, and if so, which splice sites are likely to be used. The ideas presented here are being developed into computer programs that can be used by those involved in any large scale *Drosophila* genomic sequencing project.

Splice sites conform well to a consensus, and the identification of potential splice sites on the basis of conformity to frequency matrices, such as those in Tables 35.1–35.3, generated by tabulating known splice sites, would appear to be a straightforward matter. However, there is no universally accepted method for weighting specific nucleotides within such matrices. Lear et al. (1990) experimentally determined the strength, relative to a reference site, of 37 actual 5' splice sites within a common defined sequence context by HeLa cell transfection assays, and then compared those results to a compilation of primate data (Shapiro and Senapathy 1987) using a number of distinct scoring schemes. These techniques, including the increasingly applied "Senapathy score," calculated according to Shapiro and Senapathy (Shapiro and Senapathy 1987), performed comparably (giving coefficients of correlation between measured strength and score of between 0.68 and 0.76. However, it is entirely possible that scoring schemes not examined in that paper would yield better results. In particular, the log-likelihood scoring technique derived from information theory and embodied in the programs CONSENSUS and PATSER

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FIG. 35.3. A model for the splicing of small introns in *Drosophila*. Many *Drosophila* introns are smaller than the minimum size recognized by the mammalian splicing apparatus (Bingham et al. 1988; Hawkins 1988; Mount et al. 1992), and there are indications of a distinct mechanism for the splicing of small introns in *Drosophila* (Guo et al. 1992). (A) depicts the mechanism of splicing of large introns in *Drosophila*, following Fig. 35.1. Boxes around the splice sites and branchpoint represent complexes formed from splicing factors. (B) presents a model of direct contact between complexes at the 5' splice site and at the branchpoint that may explain those observations. "Interference" refers to the effect of too short a 5' splice site to branchpoint distance on mammalian splicing, as described by Himmelspach et al. (1991), who observed a lack of complexes involving U4/U5/U6 or U2. "Accommodation" depicts the idea that *Drosophila* spliceosomal components may have evolved to be compatible with assembly on introns with shorter 5' splice site to branchpoint distances, and indicates how that might have led to the observation of a preferred 5' splice site to branchpoint distance of approximately 40 nucleotides in small *Drosophila* introns (Mount et al. 1992).

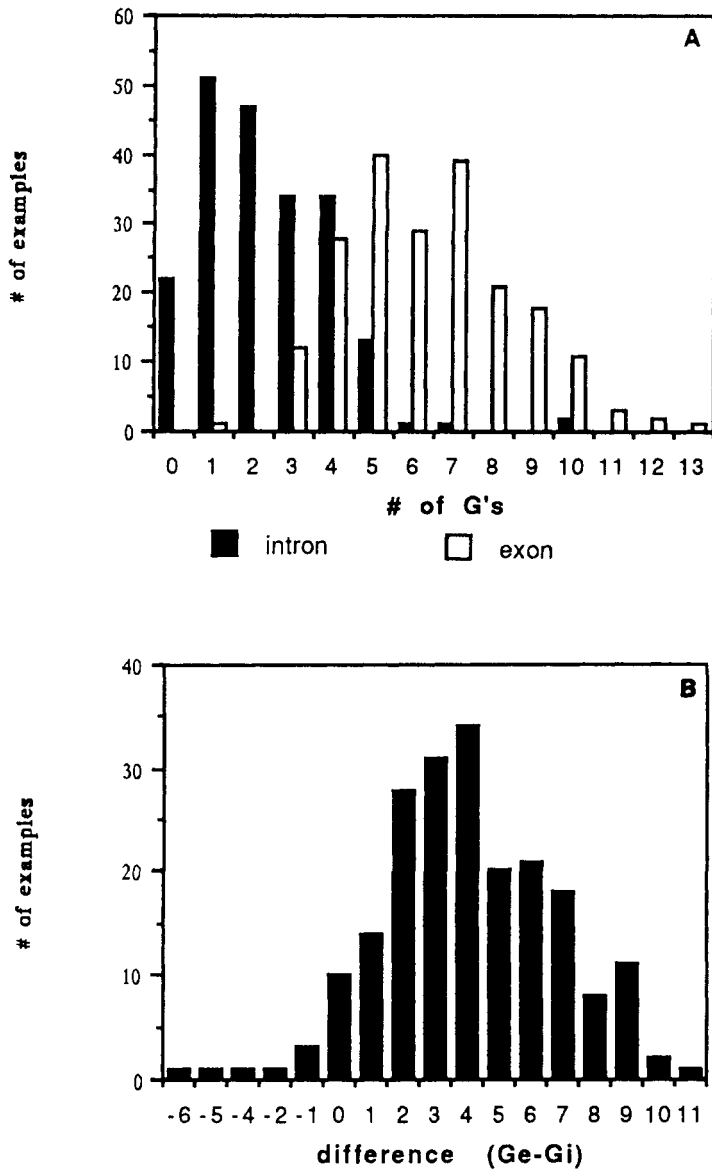


FIG. 35.4. Distribution of G content in 25 nucleotide windows flanking 3' splice sites. (A) The number of Gs in a 25 nucleotide window within the intron and adjacent to the 3' splice site (positions -29 to -5) was determined, and the number of examples in a dataset of 205 3' splice sites with a given number of Gs is indicated by black bars. Note that there are relatively few cases of introns with more than 4 Gs in this window, and only three cases of introns with more than 5 Gs in this window. The number of examples with a given number of Gs in a 25 nucleotide window in the adjacent exon is indicated by white bars. (B) The difference between the number of Gs in a 25 nucleotide window within the intron is subtracted from the number of Gs in a 25 nucleotide window within the flanking exon for each of 205 *Drosophila* 3' splice sites, and the distribution of results is plotted. Note that very few (seven out of 205) cases of 3' splice sites have more Gs in the flanking exon than in the intron.

TABLE 35.2. 3' splice site sequences

*Drosophila.*

	-14	-13	-12	-11	-10	-9	-8	-7	-6	-5	-4	-3	-2	-1	1	2	3
A	21	21	22	20	19	19	24	19	10	11	28	5	99	0	33	17	18
C	21	23	16	24	24	37	28	36	28	20	23	68	0	0	15	21	32
G	8	10	9	9	10	6	11	6	5	4	23	0	0	100	34	19	25
T	49	45	53	47	47	39	37	40	57	64	25	27	0	0	18	43	25
	T	T	T	T	T	Y	Y	Y	T	T		C	A	G	R	T	

Total.

	-14	-13	-12	-11	-10	-9	-8	-7	-6	-5	-4	-3	-2	-1	1
A	11	11	10	8	11	10	11	11	7	8	25	3	100	0	27
C	29	33	30	30	32	34	37	38	39	36	26	75	0	0	14
G	14	12	10	10	9	11	10	9	7	6	26	1	0	100	49
T	46	44	50	52	48	45	42	43	47	51	23	21	0	0	10
	T	Y	Y	Y	Y	Y	Y	Y	Y	Y		C	A	G	G

*Drosophila* 3' splice site scoring table. Scores were calculated according to Hertz et al. (1990).

	-22	-21	-20	-19	-18	-17	-16	-15	-14	-13	-12	-11	-10
A	0.5	0.5	0.1	0.3	0.0	0.2	-0.2	0.1	-0.2	-0.2	-0.2	-0.3	-0.4
C	-0.6	-0.6	-0.3	-0.3	-0.2	-0.3	-0.3	-0.4	-0.2	-0.1	-0.6	0.0	0.0
G	-1.1	-1.3	-1.1	-1.2	-0.9	-1.5	-1.8	-0.8	-1.5	-1.3	-1.4	-1.4	-1.3
T	0.6	0.6	0.8	0.7	0.7	0.8	1.1	0.7	1.0	0.9	1.1	0.9	0.9

	-9	-8	-7	-6	-5	-4	-3	-2	-1	1	2	3
A	-0.4	0.0	-0.4	-1.3	-1.2	0.2	-2.2	2.0	-5.7	0.4	-0.5	-0.5
C	0.6	0.2	0.5	0.2	-0.3	-0.1	1.5	-4.7	-5.7	-0.7	-0.2	0.4
G	-2.0	-1.2	-2.0	-2.1	-2.4	-0.1	-5.7	-5.7	2.0	0.5	-0.4	0.0
T	0.6	0.6	0.7	1.2	1.4	0.1	0.1	-5.7	-5.7	-0.4	0.8	0.0

(Hertz et al. 1990) was not tested. Tables 35.1–35.3 include scoring matrices in addition to frequency matrices for splice sites and branchpoints. Scores were calculated from the data set used by Mount et al. (1992) using the formula  $\log_2(4[N_b + 1]/N + 1)$ , where  $N_b$  is the frequency of a particular base and  $N$  is the number of examples that contribute to the matrix (Hertz et al. 1990).

It should be kept in mind that one cannot know the splicing pattern of a gene without looking at the mRNA from that gene. This is primarily because there are exceptions to each of the various features common to most *Drosophila* introns or splice sites. However, there are multiple signals involved in the specification of most introns, and these signals are usually in agreement. Furthermore, biochemical information summarized in the preceding section may serve as guidelines.

TABLE 35.3. Branchpoint consensus

Mammalian examples. Actual numbers, from Nelson and Green (1989).

	BP							
A	3	10	0	8	10	29	1	2
C	8	9	20	6	4	1	15	11
G	5	7	3	0	13	0	7	2
T	15	5	8	17	4	1	8	16
Consensus:	T	N	C	T	R	<u>A</u>	C	Y
Yeast sequence:	T	A	C	T	A	<u>A</u>	C	T

Branchpoints determined for *Drosophila* introns in homologous extracts.

ftz:	A	G	C	T	A	<u>A</u>	C	C	Rio (1988)
white:	T	C	T	T	A	<u>A</u>	T	A	Guo et al. (1992)
Myosin HC exon 19:	T	T	T	T	A	A	T	C	Hodges and Bernstein (1992)
Myosin HC exon 19:	A	A	C	T	A	A	T	T	Hodges and Bernstein (1992)
Myosin HC exon 6:	T	C	C	T	A	A	T	G	Hodges and Bernstein (1992)

*Drosophila* branchpoint matrix\* as determined by CONSENSUS (percentages).

A	36	41	7	20	92	86	1
C	1	10	42	10	2	4	10
G	8	3	2	9	5	1	1
T	55	45	48	60	0	9	88
Consensus:	W	W	Y	T	A	A	T

*Drosophila* branchpoint scoring table. Scores were calculated according to Hertz et al. (1990) using a weighted average of the matrix above and that in Mount et al. (1992).

A	0.6	0.7	-2.3	-0.2	1.7	1.8	-2.0
C	-1.8	-1.8	1.1	-0.7	-1.7	-2.2	-1.2
G	-1.6	-1.4	-1.2	-1.9	-1.9	-4.3	-4.7
T	0.9	0.8	0.4	1.2	-4.7	-1.7	1.7

Alternative *Drosophila* branchpoint scoring table. Scores were calculated according to Hertz et al. (1990) using the five experimentally determined branchpoints listed above.

A	1.0	0.4	-0.6	-0.6	2.0	2.0	-0.6
C	-0.6	1.0	1.4	-0.6	-0.6	-0.6	0.4
G	-0.6	0.4	-0.6	-0.6	-0.6	-0.6	-0.6
T	1.4	0.4	1.0	2.0	-0.6	-0.6	1.7

\* Different in detail from that reported in Mount et al. because a different (uniform) *a priori* base composition was assumed.

Splice sites will also generally occur at boundaries between DNA with roughly 50% A + T content (exons) and DNA with higher A + T content (introns). Introns in non-coding regions may be an exception to this rule. However, G content in the vicinity of 3' splice sites is an extremely reliable

predictor (Fig. 35.4; see below). Splice sites will generally conform to the matrices given in Tables 35.1 and 35.2. If the exons are coding, an open reading frame will generally be continued across the splice sites. Size is also an important clue—over half of the *Drosophila* introns that occur in GenBank are between 50 and 80 nucleotides in length, with the majority of those being between 60 and 66.

### *Identification of 5' Splice Sites*

5' splice sites are best identified by the invariant GT and a consensus matrix. The *Drosophila* 5' splice site matrix determined by Mount et al. (1992) is presented in Table 35.1. This matrix or one like it can be considered universal, reports of minor differences between species (Jacob and Gallinaro 1989) or between introns of different sizes (Fields 1990) notwithstanding. One example of such a difference is the greater frequency of T at position 6 in *Drosophila* as opposed to mammalian introns (68% versus roughly 50%). When the matrix shown in Table 35.1 is used to calculate scores for each of the 205 5' splice sites in the dataset used, the average score obtained is 6.8. Eighty per cent of actual sites score over 5.0, 95% score over 3.0 and only three sites have negative scores.

### *The Branchpoint, G-poor Region, and 3' Splice Site*

Two scoring matrices for the *Drosophila* branchpoint are given in Table 35.3. One matrix is based on the five branchpoints that have been experimentally determined in a homologous extract, and one is based on a weighted average of matrices derived using the program CONSENSUS (Hertz et al. 1990; Mount et al. 1992). Given the low number of experimentally determined branchpoints, it is unclear which scoring matrix is preferable, or, indeed, how much importance should be attached to finding a match to the branchpoint matrix.

A G-poor region should be found between the branchpoint and the 3' splice site, and is an extremely useful tool for locating 3' splice sites within unannotated sequence. Note that a large portion of this G-poor region is incorporated in the 3' splice site scoring matrix given in Table 35.2.

In summary, the strategy I propose is a simple one. First, determine open reading frames and plot A + T content and G content across the gene. Then look for splice sites and branchpoints using that information and the scoring matrices given in Tables 35.1–35.3, bearing in mind the size constraints and preferences described above. I am currently developing computational applications of the ideas described here, and interested readers are encouraged to consult current releases of the electronic *Drosophila Information Newsletter* for information about the availability of software. To add your name to the Newsletter distribution list, send e-mail to [LISTSERV@IUBVM.UCS.INDIANA.EDU](mailto:LISTSERV@IUBVM.UCS.INDIANA.EDU) with the message "SUB DIS-L *Your-real-name*." Statistics cited in this chapter were derived by the author and Lonny Sorkin using the data set described in Mount et al. (1992).



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## Translation Start Sites and mRNA Leaders

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### Introduction

A prototypical eukaryotic mRNA is often described as having a short (less than 100 nt) 5' untranslated leader sequence upstream of start codon containing a good consensus sequence (Lewin 1990). Translation initiation from such a mRNA follows the scanning model whereby: (1) a complex of proteins including the cap-binding protein (eIF-4E) associates with the 5' cap of the mRNA; (2) this complex in turn facilitates binding of the preinitiation complex (40S ribosomal subunits + eIF-2-GTP-tRNA<sup>met</sup>); (3) the preinitiation complex scans the mRNA searching for the start codon (the first AUG encountered in the prototypical mRNA); (4) the large ribosomal subunit (60S) joins the 40S subunit beginning translation (Kozak 1989). The first *Drosophila* mRNAs characterized (e.g., *Adh*, larval cuticle proteins, and the glue proteins) fit the eukaryotic prototype. However, in more recent years an increasing number of eukaryotic mRNAs have been discovered that contain unusual features. First, approximately 9% of the characterized vertebrate mRNAs contain long leader sequences with upstream open reading frames (Kozak 1987). The presence of upstream open reading frames present a dilemma for the scanning model. If the ribosome engages translation of an upstream open reading frame, terminates, and then dissociates from the mRNA, how is translation of the major coding region achieved? Kozak demonstrated two possible solutions. The scanning preinitiation complex can ignore an AUG codon in the leader if it is in a poor context for initiation or the ribosomes can engage translation of the URF, terminate, resume scanning (presumably in the form of the small ribosomal subunit), reload initiation factors, and reinitiate translation downstream at the start codon for the major coding region (Kozak 1989). Recently, Macejak and Sarnow (1991) have demonstrated a more radical solution: cap-independent, internal binding of the ribosome. Under the "internal



initiation" model, the ribosome can bind downstream of any offending URFs and then traverse the remaining leader sequence to the major start codon.

A reanalysis of the translation start site consensus sequence has also altered our view of the prototypical mRNA. Kozak (1984) initially argued that the sequence CCACCAUGG was the eukaryotic consensus sequence for translation initiation and showed the sequences that departed markedly from this consensus reduced translation initiation of the rat preproinsulin mRNA (Kozak 1986). However, similar experiments in yeast failed to show significant reduction in translation initiation from start codons with a "poor context" (Baim and Sherman 1988). Studies on the start codon context of the *Drosophila Adh* gene showed a significant effect of context intermediate to that observed in the rat and yeast studies (Feng et al. 1991). A further complication of the start codon context came from the finding that Kozak's consensus sequence was not based upon explicit quantitative criteria and did not represent a true consensus sequence for any major eukaryotic group (Cavener 1987; Cavener and Ray 1991). Moreover, various eukaryotic groups exhibit somewhat different consensus sequence for the translation initiation site. For example, yeast mRNAs exhibit relatively high frequencies of U at  $-2$  and  $-1$ ; Kozak had shown that Us at these positions were rare in vertebrates mRNA and detrimental to translation of the rat preproinsulin mRNA. Only the presence of A or G at the  $-3$  position is a consensus throughout eukaryotes (Cavener and Ray 1991).

## Data Acquisition and Analysis

We compiled the following data for *Drosophila* mRNAs: (1) length of the leader sequence; (2) method of determining the extent of the leader sequence; (3) the number of upstream start codons (uAUG); and (4) the start codon context from positions  $-6$  to  $+4$  for the major translation start sites and for a random sample of the uAUGs. Initially, most of the mRNA sequences were identified in GenBank Release 69 using the INTERBAS computer program (Cavener and Ray 1991). Sequences reported recently in several journals were added to this list. In the vast majority of cases the start codons are readily discernible from the GenBank records. However, information regarding the leader sequence is almost always inaccurate and/or incomplete in GenBank. In many cases the extent of the leader sequence has not been determined empirically. Consequently we examined the primary literature reporting each of the 403 mRNAs listed in Table 36.1 in order to ascertain the method for mapping the leader sequence and to verify the map features of the GenBank records. Since the 5' end of the leader sequence is defined by the presumptive start site of transcription, the precise limits of the leader sequence are only known in cases where extensive transcript mapping experiments have been conducted. Ideally, this involves a combination of comparing cDNA sequences with genomic DNA sequences, primer extension and nuclease protection experiments. For the majority of *Drosophila* mRNAs these data are incomplete. Typically, the extents of mRNA sequences are inferred only from the analysis of the longest cDNA

TABLE 36.1. Leader lengths (nt), number of upstream AUGs, and translation start site sequences from -6 to +4

<i>File</i>	<i>Encoded protein</i>	<i>Method</i>	<i>Leader lengths</i>	<i>Number of uAUGs</i>	<i>Start site</i>
5HTR	5HT serotonin receptor	b	894	15	CUGCUGAUGG
6DHR	RAD6 homolog	c	88	0	UGAAAAAUGU
ABDA	abd-A, abdominal-A, homeotic	b	668	0	AGCAAGAUGU
ABDBP3	abd-b, abdominal-B, P3	a	?	?	CCCGUCAUGC
ACHE	ace, acetylcholinesterase	b	993	6	UAUUCAAUGC
ACHRR	acetylcholinesterase receptor	b	254	6	AAAAUCAUGG
ACHRX	muscarinic acetylcholine receptor	b	32	0	CCGGCGAUGA
ASC1	ac, achaete	e	63	0	CUUAAAAUGG
ACS2	sc, scute	e	117	0	GUGUAAAUGA
ACT42A	actin 42A	e	102	0	UACAAAAUGU
ACT5CX	actin 5C	e	156	0	UACAAAAUGU
ACT79B	actin 79B	c	149	0	CCAAACAUGU
ACT87EA	actin 87E	e	82	0	GCCAAGAUGU
ACT88F	actin 88F	c	187	4	GCCAAGAUGU
ADF1A	adf-1 transcription factor	b	312	0	AUUGAGAUGG
ADHa	Adh, alcohol dehydrogenase distal protein	e	123	0	GUCACCAUGU
ADHb	Adh, alcohol dehydrogenase proximal protein	e	70	0	GUCACCAUGU
AFLI	arf-like, GTP binding protein	b	118	0	GUCAUCAUGG
ALSR	acetylcholine receptor alpha	b	1,282	7	CCUAAGAUGG
AMA	ama, amalgam	e	235	0	CCAGACAUGG
AMYAG1	amy, amylase	c	35	0	AUCAUCAUGU
ANNX	annexin	b	90	0	UGCAUAAUGG
ANP*	andropin	c	37	0	CUAGUUAUGA
ANTCA	Dfd, deformed homeotic	b	490	4	UCCGCAUGA
ANTCF	ftz, fushi tarazu	c	120	1	UCCGAUAUGG
ANTPa	Antp, antennapedia P1 mRNA	e	1,527	8	GCCACGAUGA
AnTPb	Antp, antennapedia P2 mRNA	e	1,729	15	GCCACGAUGA
ANTPS2	position-specific antigen 2	b	258	5	GACAAAAUGA
APRT	adenine phosphoribosyltransferase	b	89	0	AGAAAAUGA
ARMa	armadillo E16	e	135	1	ACCAAGAUGA
ARMb	armadillo E9	e	170	0	ACCAAGAUGA
ARR	arrestin-1	c	120	0	UCCAAAAUGG
ARRA	arrestin-2	c	116	0	UCCAAAAUGG
ASCA	T3 of achaete-scute	c	27	0	AUUACCAUGA
ASCB	T8 of achaete-scute	a	?	?	UUUGGCAUGC
ASE	ase, asense	c	456	8	UUAAUUAUGG
ATPA	Da-47, Na <sup>+</sup> /K <sup>+</sup> ATPase alpha subunit	b	12	0	AAUACAUGG
AWDR	awd, abnormal wing disc	e	25	0	GCGACAAUGG
B52*	B52 protein, NHCP	b	55	0	GUUAUCAUGG
BAM	bam, bag-of-marbles	c	186	1	AGAAUAAUGC
BCD16	bic, bicoid	b	169	2	GGGAAAUGG
BICD	bic <sup>D</sup> bicaudal-D	b	131	0	AUCAUCAUGU

(continued)

TABLE 36.1. *Continued*

<i>File</i>	<i>Encoded protein</i>	<i>Method</i>	<i>Leader lengths</i>	<i>Number of uAUGs</i>	<i>Start site</i>
BJ1G	BJ1, chromatin-binding protein	b	210	0	GCUAAAAUGC
BJ6	no-on transient A, Bj6	b	76	0	UAAAAAAUGG
BR*	br, broad	b	386	0	AUCGAGAUGG
BROWN	bw, brown	b	268	1	CUCGAAAUGC
BSG25D	bsg25D, blastoderm	e	296	1	CGGAUAAUGG
BX189A	pH189A ORF, BX-C	a	?	?	UCCUAAAUGU
BX189B	ph189B ORF, BX-C	c	1,019	5	UACCCGAUGG
BX200	pH200 gene, BX-C	c	494	1	UACAGAAUGG
C1A9	NHC, non-histone chromosomal protein	b	349	4	AACAAAAUGG
CACTTR	choline actyltransferase	c	406	0	GCGAACGUGG
CADA1a	cad, caudal zygotic	e	460	4	CCAGCCAUGG
CADA1b	cad, caudal maternal	e	301	3	CCAGCCAUGG
CAIM1	calmodulin	b	85	0	ACAAAAAUGG
CAPKCA	cAMP-dep protein, kinase catalytic	a	?	?	UCCAAGAUGG
CATHPO	catalase	b	87	1	AGCAAAAUGG
CCG	Cc gene, Ddc region	a	?	?	AGGAUAAUGG
CDC2P24	cdc2 homolog	b	55	0	UAAAUAUUGG
CHAB	potassium channel protein	b	406	5	GGUUGCAUGG
CHORS16	chorion, s16	?	46	0	AAAAAAAUGU
CHORS3	chorion, s36	c	31	0	GGCAACAUGC
CHORS3	chorion, s38	e	77	0	GACAAGAUGA
CHORSGa	chorion, S18-1	c	44	0	CUCAGAAUGA
CHORSGb	chorion, S15-1	c	45	0	CUCACCAUGA
CHORSGc	chorion, S19-1	c	45	0	AUAGCCAUGA
CID	ciD, cubitus interruptus dominant	b	415	6	AAUGAAAUGG
CLARET	claret non-disjunctional <sup>+</sup>	a	?	?	UUGGCGAUGG
CNC	cnc, segmentation protein	b	94	0	UGUCGCAUGG
COPO1	chaoptin	e	255	0	AGCAAAAUGG
CRN*	crn, crooked neck, cell cycle	b	80	0	CACAGCAUGG
CRPA	crumbs protein	b	213	4	GCGAUCAUGG
CSG	Cs, Ddc region	a	?	?	GAUUCGAUGU
CSKA	casein kinase II alpha	b	258	0	AGAAAAAUGA
CSKB	casein kinase II beta	b	22	0	AUCAAAAUGA
CSPAA	cysteine-string protein 29	b	150	0	AUCAGGAUGA
CSTAA	ctr, concertina	b	133	1	CCAGCGAUGU
CTCL1	cuticle protein I	c	42	0	GCGAAUAUGU
CTCL2a	cuticle protein II	f	42	0	GCCAACAUGU
CTCL2b	cuticle protein III	c	45	0	AUCAAAAUGU
CTCL2c	cutical protein IV	f	45	0	GUCAAAAUGU
CUT	cut	b	268	4	CCACGAAUGC
CYCA	cyclin A	b	296	5	CGCACAUGG
CYCC*	cyclin	b	93	0	UACGAAAUGG
CYCDC3	cytochrome c, DC3	a	?	?	UCCAAGAUGG
CYCDC4	cytochrome c, DC4	a	?	?	UCCAUAUUGG
CYCLB	cyclin B	b	123	0	AUCAAAAUGG
CYP1	cyp-1 protein, cyclophilin	a	?	?	UCAAGAUGA

TABLE 36.1. *Continued*

<i>File</i>	<i>Encoded protein</i>	<i>Method</i>	<i>Leader lengths</i>	<i>Number of uAUGs</i>	<i>Start site</i>
D1DE	insulin-degrading enzyme	b	297	1	CCCAAGAUGA
D1P	chromosomal protein D1	b	227	0	AGAGAAAUGG
DA2	D alpha-2 protein, D'2	b	492	1	GUCACCAUGG
DC1AB	DC1, putative protein kinase	b	92	3	GCUGUUAUGA
DC2	DC2, putative protein kinase	b	894	1	ACAGCGAUGU
DCKA	calmodulin-dependent protein kinase	b	250	0	AUCGCGAUGG
DCO	cAMP-protein kinase catalytic subunit	c	828	2	UCCAAGAUGG
DDC a	Ddc, dopa decarboxylase CNS	c	233	0	UCUGAAAUGA
DDC b	Ddc, hypoderm form	c	197	0	AUCGACAUGG
DDY3	Ddyn3, dynamin shibire locus	b	394	2	GCCGCAAUGG
DDYN4	Ddyn4, dynamin shibire locus	b	51	0	GCCGCAAUGG
DEC1A	dec-1 chorion-1 fc125	e	75	0	UACAGGAUGA
DELTA	D1, delta, neurogenic (DLG)	b	141	0	AUAAACAUGC
DFUR1	dfur1, furin-type protein	b	104	0	CCCACAAUGA
DG1A1	cGMP-dependent protein kinase	c	108	1	GGCAGAAUGG
DG2T1A3	cGMP-dependent protein kinase	e	97	0	GCCUGGAUGC
DG2T2A	cGMP-dependent protein kinase	e	776	9	UUCGUAAUGA
DG2T2B	cGMP-dependent protein kinase	e	338	1	UUCGUAAUGA
DGHTRL	da, daughterless	b	212	2	GCUGAAAUGG
DIPT	dipteracin	b	24	0	ACUGAGAUGC
DLGA	discs-large tumor suppressor	b	380	3	UGCGAUAUGA
DMYD	Dmyd, myogenic	b	262	2	UGAAAAAUGA
DNC	dnc, dunce	b	363	4	AGUCUUAUGA
DORSAL	dl, dorsal	b	274	2	CACAUAAUGU
DOXA2	A2 comp. of diphenol oxidase	c	90	0	UACAAAAUGA
DPPC	dpp, decapentaplegic	b	1,187	6	GCGACCAUGC
DRCII1	II-cAMP-dependent protein kinase regulatory subunit	e	402	1	AGCGAAAUGG
DRCIV1	IV-cAMP-dependent protein kinase regulatory subunit	c	182	1	AGCCCGAUGC
DRICI1	I-cAMP-dependent protein kinase regulatory subunit	e	565	3	UACCACAUGU
DSK	sulfated tyrosine kinin	a	?	?	CUGUUUAUGC
DSX*	doublesex, male and female	e	1,020	9	GGAAUCAUGG
E74A	E74A, ecdysone inducible	e	1,891	17	UCAGCGAUGC
E74B	E74B, ecdysone inducible	e	793	6	UGCAAAAUGA
E75A	E75A, ecdysone inducible	e	380	3	AGCAAAAUGU
E75B	E75B, ecdysone inducible	e	284	3	UCAAAUAUGG
EAG	putative potassium channel protein	b	463	2	GGCAAAAUGC

(continued)

TABLE 36.1. *Continued*

<i>File</i>	<i>Encoded protein</i>	<i>Method</i>	<i>Leader lengths</i>	<i>Number of uAUGs</i>	<i>Start site</i>
EAST	easter, putative serine protease	b	203	0	ACGAAAAUGC
ECR*	EcR, ecdysone receptor	b	1,068	11	CAGAGGAUGA
EDG78A	EDG-78 cuticle protein	c	76	0	AUCAUCAUGU
EDG84A	EDG-84 cuticle protein	c	61	0	AUCAGCAUGU
EDG91B	EDG-1, cuticle protein	c	34	0	AUCGCAAUGG
EF1AF1	elongation factor, F1	c	80	0	UCCAACAUGG
EF1AF2	elongation factor, F2	c	139	0	GCAAGGAUGG
EF2A	translation elongation factor 2	b	72	0	UCCAAAAUGG
EFSII	RNA pol II elongation factor	b	236	0	GCCAAAAUGA
EGFRA	epidermal growth factor receptor homolog	b	84	0	GAUAUCAUGA
EGFRB	epidermal growth factor receptor homolog	b	22	0	GCAACAAUGC
EIF2AL*	eIF-2 alpha subunit	a	?	?	UUUAACAUGG
EIF2BE*	eIF-2 beta subunit	b	> 99	1	GACACAAUGG
EIP28G	ecdysone inducible protein	e	65	1	GAAAUCAUGU
ELAVK	elav protein	b	491	1	AAAACAAUGG
ELF1	Elf1, DNA binding protein	b	920	7	CGUAUAAUGU
EMC	emc, extramacrochaetae	c	258	0	UCCAGAAUGA
ENGM	en, engrailed	b	168	0	AAACCAAUGG
ENHSPA	E(spl), enhancer of split	b	222	0	AACAACAUGU
ESPLM4	E(spl), m4 transcription unit	f	79	0	AUCAUCAUGU
ESPLM5	E(spl), m5 transcription unit	c	84	0	UACAAAAUGG
ESPLM7a	E(spl), m7 transcription unit	f	128	0	CACACAAUGG
ESPLM7b	E(spl), m8 transcription unit	f	96	0	ACAAAAAUGG
EST6	Est-6, esterase-6	b	24	0	AGCAACAUGA
EVE	eve, even skipped	c	94	0	CCAAACAUGC
F1GA	F1 50kd protein	b	200	2	UCCAACAUGG
FCN	fasciclin I	b	174	0	GCUAAAAUGC
FCNIII	fasciclin III	b	582	2	AAAAUCAUGU
FKH	fork head	b	707	3	GACAUCAUGC
FMRF	FMRFamide	b	18	1	GCCUUGAUGU
FOS*	fos homolog	b	772	5	GCAACAAUGA
FPS85D	dfps 85D	b	243	2	AGCAUCAUGG
FRZAC2	frizzled, AC2	b	709	8	UCCAAAAUGU
FS1YA	fs(1)Ya, nuclear env.	b	23	0	AGGUGUAUGU
FSHA	fsh membrane protein A	c	662	3	ACCACCAUGU
GADPH1	GAPDH-1	e	62	0	UCAGCCAUGU
GADPH2	GAPDH, glyceraldehyde-3-phosphate dehydrogenase	c	49	0	UUAACCAUGU
GART	Gart	c	160	0	GGAAUUAUGU
GART p	pcp, pupal cuticle gene Gart	b	33	0	GACACCAUGU
GIAA	guanine nucleotide binding, regulatory subunit	b	441	3	CACAAGAUGA
GLDGMC	Gld, glucose dehydrogenase	e	344	0	AUCAACAUGU
GLUEDA	Glued	b	360	6	UCCUCCAUGA
GNBPSA1	guanine nucleotide binding protein alpha	b	486	3	GCUGCGAUGG
GOALB*	G-o-alpha-like protein	b	519	2	CGCACCAUGG

TABLE 36.1. *Continued*

<i>File</i>	<i>Encoded protein</i>	<i>Method</i>	<i>Leader lengths</i>	<i>Number of uAUGs</i>	<i>Start site</i>
GPAMA*	G protein alpha mRNA type a	b	189	0	ACCACAAUGG
GPDHA	Gpdh, glycerol-3-phosphate dehydrogenase	e	136	0	CAAAAUAUGG
GTUB	gamma-tubulin	b	196	1	ACCACAAUGC
HAIRR	h, hairy	c	492	0	ACCGAAAUGG
HBGa	hunchback, maternal mRNA	c	511	1	GCCAAGAUGC
HBGb	hunchback, zygotic mRNA	c	165	0	GCCAAGAUGC
HELI	RNA helicase	b	33	0	UGAAUAAUGA
HGSG2	heat-shock 2, male specific	e	60	0	ACUACAAUGG
HISH1	histone, H1	c	36	0	AAAAAGAUGU
HLI*	HL, putative troponin I	b	134	0	CUCAAAAUGG
HMGCO	HMG CoA reductase	b	572	2	GCAGCCAUGA
HOXH20	H2.0 homeobox	b	205	0	CGGACAAUGU
HP1	Hp-1	c	169	0	ACAAAAAUGG
HRB87F*	Hrb87F, A/B hnRNP protein	c	132	0	GAGAGAAUGG
HREC2C	putative steroid hormone receptor	b	198	1	CCCAGGAUGG
HSC7A1	cognate of hsp70	a	?	?	GCCGACAUGC
HSP1	heat-shock protein 1	c	94	0	GUGAAAAUGU
HSP22G	hsp22, heat-shock protein	e	253	0	ACUACAAUGC
HSP27G	hsp27, heat-shock protein	e	121	0	UCAAAAAUGU
HSP4	hsp23	e	111	0	ACAAAAAUG
HSP7A2	hsp70	e	244	0	CACACAAUGC
HSP83A	hsp83	c	148	0	UUGCAGAUGC
HSPG3	heat-shock gene 3 from 67B	e	168	0	AGUAAAAUGC
HSPHEX	heat-shock transcription factor	b	228	0	CACUUUAUGU
IMP	IMP-E2, ecdysone inducible	b	75	0	GCGAUAAUGA
INT1HO	Dint-1	b	417	7	GCAAUAAUGG
INVR	invected	e	294	3	AAACUGAUGU
JUN	dJRA/Djun, jun homolog	b	207	0	GCAAACAUGA
K10G	K10 putative DNA-binding protein	e	191	0	CCUGCAAUGG
KINHCA	kinesin heavy chain	b	320	1	UAAGCAAUGU
KINLA	nod, kinesin-like protein	b	71	1	AUCUGCAUGG
KNIRPS	knirps	b	270	0	UUCGAGAUGA
KNR1	knirps-related protein	b	516	4	ACCAUAAUGA
KR	krueppel	d	185	1	UUGUGGAUGU
L2AMD	alpha-methyl dopa hypersen.	b	150	0	AGCGGUAUGG
LA9	LAP, DNA-binding protein	b	435	8	GUCAAAAUGG
LABG1	labial F24	d	239	0	GACAAUUGA
LAMB1	laminin B1	b	423	5	AUCGAGAUGU
LAMB2	laminin B2	b	227	2	CCCACCAUGA
LAMDMO	laminin, nuclear	b	130	0	GUGAACAUUGU
LAMIN	lamin	c	148	1	GUGAACAUUGU
LARM	DLAR, protein tyrosine phosphatase	b	117	0	GAAAUAUGG
LETHAL	lethal(1)2cb sarcoplasmic actinin	b	66	0	CACAAGAUGA

(continued)

TABLE 36.1. *Continued*

<i>File</i>	<i>Encoded protein</i>	<i>Method</i>	<i>Leader lengths</i>	<i>Number of uAUGs</i>	<i>Start site</i>
LGL2	lethal(2) giant (L2GLR)	b	474	6	CCAAUUAUGU
LOD*	lodestar, nucleotide triphosphate binding	b	84	1	CUAAAAAUGU
LSP1A5	Lsp larval serum protein, alpha	c	88	0	UCCAGGAUGA
LSP1B	Lsp larval serum protein, beta	c	85	0	GUCAACAUGA
LSP1C	Lsp larval serum protein, gamma	c	82	0	CCAAGGAUGA
MACE	muscarinic acetylcholine receptor	b	293	3	UCCGUCAUGG
MAP205	205kd microtubule-associated protein	e	420	0	UAAAGGAUGG
MASTER	mastermind	c	753	8	GCAUUUAUGG
MET	Met, metallothionein	b	123	0	AUCAAGAUGC
METO	Met, metallothionein	b	69	0	UACAAGAUGG
MEX1A	mex1	c	76	0	AUCACCAUGU
MLE*	mle, maleless	b	79	0	CUAAGAAUGG
MOV34	Mov34 protein	b	111	0	ACAAACAUGC
MP20	mp20, muscle-specific protein	c	70	0	UCAAACAUGU
MPP1	patched (PTCR)	b	772	7	ACCAUAAUGG
MSP316	msP316 male-specific protein	c	34	0	AUCAAAUGG
MST355a	msP355a male-specific protein	c	22	0	CUCGAAAUGA
MST355b	msP355b male-specific protein	c	25	0	UCCACAAUGA
MYBDR	D-myb oncogene homolog	b	605	7	CUUAAGAUGG
MYHB	myosin heavy chain	e	113	0	AGCAAGAUGC
MYL	myosin light chain	b	43	0	GACAAAAUGG
MYLA	myosin light chain 2	c	66	0	AGCACAAUGG
MYONMAa	non-muscle myosin heavy chain	c	93	0	AAACAAAUGA
MYONMAb	non-muscle 2nd start codon	c	228	1	GCCAAAAUGU
MYSP	myspheroid	b	93	0	AAAGCCAUGA
NCDA	ncd, non-claret disjunctional	b	65	0	UUGGCGAUGG
NEU*	neu, neuralized	b	273	2	ACUACCAUGG
NEUROT	neurotactin	b	508	1	GACAAUAUGG
NINAA	ninaA	a	?	?	AAAAUCAUGA
NINAC	ninaC	c	146	1	UAAGUCAUGA
NORPA	norpa, phospholipase C	b	652	5	GCAAUAAUGA
NOS*	nanos	c	261	1	UUCGCCAUGU
NOTCH1	Notch, ectodermal determinant	c	865	8	AACAAAAUGC
NRGAA	neuroglian	b	27	0	ACCAAAAUGU
NUMB	numb	b	791	5	ACAGGCAUGG
OPSA	ninaE, opsin	c	170	2	AACACAAUGG
OPSAA	Rh2, opsin	e	37	0	CUGAGCAUGG
OSKAR	oskar	c	15	0	CAAGCGAUGG
OTEDA	otefin	b	75	1	GCCAAAAUGC
OTUA	ovarian tumor (OTU)	c	154	1	GUCGCCAUGG
PABP	poly(A)-binding protein PABP	b	132	0	CCAAAUAUGG
PAH	pah, phenylalanine hydroxylase	b	84	0	GUGAAAAUGU
PCGENE	Pc, polycomb	b	109	0	UUAAAAAUGA
PCNA	proliferation cell nuclear antigen	d	89	0	UUCAACAUGU

TABLE 36.1. *Continued*

<i>File</i>	<i>Encoded protein</i>	<i>Method</i>	<i>Leader lengths</i>	<i>Number of uAUGs</i>	<i>Start site</i>
PEP*	pep, protein on ecdysone puffs	b	217	0	AAAAUUAUGG
PEPCK	PEPCK, phosphoenolpyruvate carboxylase	v	29	0	AACAAAAUGC
PERA	per, period	c	368	1	AGCACCAUGG
PKC53E	protein kinase C 53E	b	62	0	CUUUUAAUGG
PKC98F	protein kinase C 98F	b	398	7	GACCUCAUGC
PKCR	protein kinase C	b	886	17	GCAACAAUGU
PLC21A	plc-21, phospholipase c	b	824	11	GUGAGGAUGA
PMSH2	msh-2	b	289	0	GCGAGGAUGU
PN*	pn, prune	b	70	0	CUGGUAUUGG
POLO*	polo, putative protein kinase	b	219	1	AGCAAGAUGG
PP1A	phosphatase 1 alpha	b	129	0	GCAAAUUGG
PRD	paired	b	245	1	GAACUAUGA
PROS*	prospero, axonal growth regulation	b	301	0	GGCUUCAUGA
PROS281	proteasome subunit	b	60	0	AACAAGAUGU
PROS29	proteasome subunit	b	77	1	UUAGCAAUGG
PROS35	proteasome 35kd	b	70	0	AAAGUCAUGU
PTPM	tyrosine phosphatase DPTP	b	54	1	CAAGCAUGG
R118C	intronic R1 gene 18C	b	117	0	UGCAAAAUGA
RAB3	rab3, neuronal GTP-binding protein	b	586	3	GAUAAAAUGG
RAFPO	raf, proto-oncogene	b	84	0	GAACUAAUGG
RAS1	Dras1, proto-oncogene	b	167	0	AGCCAAAUGA
RAS21	Dras2, proto-oncogene	b	184	3	CUUAUAAUGU
RAS3	Dras3, proto-oncogene	b	57	0	GCCAGCAUGC
RCC1*	RRC1, regulator chromatin condensation	b	211	0	GCUAAAAUGC
RDG*	rdgB, retinal degeneration	b	180	2	GUCAACAUGC
REF2P	ref(2)p, sigma rhabdovirus multiplication	e	371	0	GCGAAAAUGC
RGPS14a	rp14, ribosomal protein S14 A	c	29	0	CCCAGAAUGG
RGPS14b	rp14, ribosomal protein S14 B	c	34	0	UGCAGAAUGG
RH3A	Rh3, opsin	e	22	0	CGGAGCAUGG
RH4A1	Rh4, opsin	c	87	0	ACCGAUUUGG
RM62RH	rm62, RNA helicase	b	482	2	GGAGUAAUGG
RNP70K	U1 70K snRNP	c	208	1	CACAAAAUGA
RNPOL2	RP140, RNA polymerase II 140 kilodalton subunit	c	168	4	AUUCAGAUGU
RP128	RNA polymerase III 128 kilodalton subunit	a	?	?	AACGAAAUGG
RP135	RNA polymerase III 135 kilodalton subunit	c	98	0	UACAACAUGC
RP21C	rp21C, A-type ribosomal protein	b	48	0	UUCGACAUGU
RP49	rp49, ribosomal protein 49	d	9	0	UUCAAGAUGA
RPA1R	rpA1, ribosomal protein	e	89	0	UUAAACAUGC

*(continued)*



TABLE 36.1. *Continued*

<i>File</i>	<i>Encoded protein</i>	<i>Method</i>	<i>Leader lengths</i>	<i>Number of uAUGs</i>	<i>Start site</i>
RPII	RNA polymerase II, 215 kilodalton subunit	d	435	3	ACCAGGAUGA
RPL1R	ribosomal protein L1	b	69	0	ACGAAAAUGA
RPS17	rp17, ribosomal protein S17	c	56	0	AACAUAUUGG
RRP1	Rrp1, strand transferase	b	132	0	UCCAUAUUGC
RUD1	rudimentary	e	11	0	UCCAAUAUGG
RUNTR	runt, segmentation gene	e	252	0	UACGAGAUGC
S12*	1(3)S12	a	?	?	UGCAGCAUGG
S1C4	beta-amyloid-like	b	152	0	CGAACAAUGU
S2ZSTM	suppressor-2 of zeste	b	149	1	AGAAAGAUGC
S59	S59 homeo box	b	67	0	CCAAAAUUGG
SAD	sad, nicotinic acetylcholine receptor	b	343	1	GUCACCAUGG
SAL	spalt	b	50	0	GCCACGAUGA
SAS	sas, putative cell adhesion receptor	b	44	0	ACCAAAUUGC
SCAa	sca, scabrous, 1st putative start	c	321	1	GUGUGAAUGA
SCAb	sca, scabrous, 2nd, in-frame start	c	396	2	GCAACAAUGG
SD*	Sd, segregation distortion	b	121	2	CGAGGCAUGU
SER2a	serine protease SER1	c	24	0	AACAAGAUGA
SER2b	serine protease SER2	c	24	0	ACCAAGAUGA
SERCA	sarcoplasmic/endoplasmic reticulum Ca <sup>2+</sup> -ATPase	b	32	0	AUCAAGAUGG
SEV	sevenless	c	229	2	GCCUCGAUGA
SGG	shaggy	b	280	0	GUUACGAUGA
SGS378a	Sgs-3, salivary gland protein	c	29	0	AAAAACAUGA
SGS378b	Sgs-7, salivary gland protein	c	33	0	AGAACCAUGA
SGS378c	Sgs-8, salivary gland protein	c	33	0	ACAACCAUGA
SGS4C1	Sgs-4, salivary gland protein	e	13	0	GUCAAGAUGC
SGS5	Sgs-5, salivary gland protein	d	33	1	UACGACAUGU
SHAKE2	shaker	b	269	1	GCCAAGAUGA
SHAKE3	shaker, larval	b	72	2	GCCUGUAUGG
SINA	seven in absentia	e	903	11	CUUCCAAUGU
SING2	sn, singed	b	739	1	AGCACCAUGA
SLIT	slit	f	314	1	GCCACAAUGG
SNAIL	snail	b	163	0	UCAAAAAUGG
SNAKE	snake	b	78	2	AAUAGAAUGA
SOD	Sod, superoxide dismutase	b	68	0	UUCGAAAUGG
SODCHA	para locus, sodium channel alpha	b	> 271	4	UAGACAAUGA
SOL	sol, small optic lobes gene	b	263	0	CGCGCAAUGG
SPCA	alpha-spectrin	b	270	1	AGCGAAAUGG
SPERM	mst(3)gl-9, spermatogenesis	b	97	0	UUAUAUUGU
SQH*	sqh, regulatory non-muscle myosin	c	221	0	GCAACCAUGU
SRC28C	Dsrc proto-oncogene	b	133	1	GGCAACAUGA
SRCC	Dsrc proto-oncogene	a	?	?	UAAGCCAUGG
SRYG1a	serendipity, beta	e	145	0	GACUAGAUGA

TABLE 36.1. *Continued*

<i>File</i>	<i>Encoded protein</i>	<i>Method</i>	<i>Leader lengths</i>	<i>Number of uAUGs</i>	<i>Start site</i>
SRYG1b	serendipity, alpha	e	43	0	AACAGCAUGG
SRYG1c	serendipity, gamma	e	67	0	GGCGCAAUGG
STAUFEN	staufen	b	274	3	AAGAAAAUGC
STELL	stellate	e	30	0	GGCAACAUGU
STGA	string, cdc25	b	391	3	AACAAAAUGC
STIMG	stimulatory G protein	b	299	2	GCUGCGAUGG
SUHW	suppressor of hairy wing	b	59	0	ACCAACAUGA
SUSG	suppressor of sable	e	507	4	UCGAUAAUGU
SVP1	seven-up protein, svp type 1	b	450	3	GGCGUCAUGU
SWA*	swallow	c	39	0	AAAGCGAUGA
SX1PS11	sex-lethal	e	425	1	CAGGAUAUGU
SYT	synaptotagmin	b	359	0	AACAAAAUGC
TAC*	tachykinin-like receptor	b	258	1	GCAGCGAUGG
TCP1	T complex protein Tcp-1	b	42	0	AGGAAAAUGU
TER	terminus protein	c	154	0	UCAAUCAUGU
TFIID	TATA-box binding protein TFIID	c	173	1	UGUAAGAUGG
TGA	transformer, sex determination	c	70	0	UUUCCGAUGA
TKABL1	abl, tyrosine kinase abelson homolog	c	96	0	UGGCAAAUGG
TKO	tko, technical knock-out	b	171	0	GAGAGCAUGA
TLD*	tolloid, dorsal/ventral pattern	b	72	0	CACGCAAUGA
TLL	tailless	b	177	0	AUCGGUAUGC
TMLPA	serrate (SER)	b	433	3	CCCAGAAUGU
TOLL	toll	b	574	4	GACAACAUGA
TORSO	torso, tyrosine kinase	f	195	0	AGGAAAAUGC
TRA2Aa	tra-2, transformer "A" non-sex determination	e	186	1	AGCCAGAUGG
TRA2Ab	tra-2, transformer "B" non-sex determination	e	488	2	AUCACUAUGU
TRA2Ac	tra-2, transformer "C" male germline	e	503	1	GAACGAAUGC
TROIIN	tropomyosin II, non-muscle	b	435	0	ACAAAAAUGA
TROPI2	tropomyosin I	c	103	0	AACACCAUGG
TROPT	wupA, troponin-T	a	?	?	GUAGCCAUGU
TRP	trp protein	c	191	3	GCAGAUUAUGG
TRPB	transient receptor pot	b	484	2	CGGAAGAUGG
TRYA	trypsin like, alpha	a	?	?	CCCAUCAUGU
TSH*	teashirt, ventral trunk development	b	1,008	9	UUAAAAAUGU
TTKFTZ	tramtrack (FTZF2)	b	251	3	CUCCCAAUGA
TU4A	TU-4 vitelline membrane	c	62	0	UCCGCAAUGG
TUBA1	alpha-tubulin-1	e	141	0	CUCAUAUGG
TUBA2	alpha-tubulin-1	e	96	0	AUCAUCAUGG
TUBA3	alpha-tubulin-3	e	504	0	AUCAUAUGC
TUBA4	alpha-tubulin-4	e	149	0	AAUAAAAUGG
TUBB2A	beta-tubulin-2	c	175	0	AUCAAAAAUGC
TUBE	tube	b	193	2	AACACCAUGG

*(continued)*

TABLE 36.1. *Continued*

File	Encoded protein	Method	Leader lengths	Number of uAUGs	Start site
TWISTG	twist	e	159	0	CACCAAAUGA
TYRDROG	tyramine receptor (OCR)	b	312	4	GGAAAGAUGC
UBS2AA	ubiquitin 52-AA extension protein	b	34	0	CGCAUUAUGC
UBIA	ubiquitin	e	139	6	UCCAAAAUGC
UBXG5	Ubx, ultrabithorax	e	697	2	CGUUCGAUGG
UROX	urate oxidase	e	33	0	GUCACAAUGU
VASA	vasa	b	131	1	AUCAAAUUGU
VERM	vermilion	e	57	0	UGCACCAUGA
VHATP	vacuolar H <sup>+</sup> ATPase	b	116	0	AGCAAAAUGU
VITA	vitelline membrane protein, 26A-1	c	81	0	ACCAAGAUGA
VITB	vitelline membrane protein, 3C-1	c	96	1	AGCACCAUGA
VMP	vitelline membrane protein	b	29	0	UUCAUCAUGC
WL	w, white	a	?	?	CCGGCAAUGG
XDH	ry, xanthine dehydrogenase	b	180	1	UUCACGAUGU
XR2C	xr2c, ultraspiracle	b	162	0	CCCAGGAUGG
YELLOW	y, yellow	c	171	0	AGTGCAAUGU
YOLK	yolk protein I	c	61	0	CGAACCAUGA
YP3	Yp3, yolk protein-3	c	59	0	ACCAAAAUGA
Z60MEX1a	z600	e	63	0	GUUAUUAUGU
Z60MEX1b	gld-F female specific	e	142	0	GUUAAGAUGG
Z60MEX1c	gld-M male specific	e	307	1	GUUAAGAUGG
ZPBA	trithorax	b	841	11	ACUAUUAUGG
ZESTE	zeste	d	964	4	ACUCAAAUGU
ZFH1	zinc finger homeobox protein 1	b	358	2	UUCCAAAUGU
ZFH2	zinc finger homeobox protein 2	b	369	4	UCUCCAAAUGU
ZIPR	zipper	b	261	3	AGCACGAUGA

Methods used to map the leader sequence: a = none or ambiguous data; b = 5' UTR of the longest cDNA; c = 5' UTR of longest cDNA and primer extension data or nuclease protection; d = primer extension or nuclease protection (genomic sequence only); e = primer extension and nuclease protection along with cDNA and/or genomic sequence data; f = presence of consensus TATA sequence and *Drosophila* consensus transcription start site plus partial cDNA sequence of leader.

\* An asterisk at the end of the file name indicates that this sequence was not included in GenBank Release 69. A temporary file name was assigned to such sequences by us.

Lower case letters in file names were used to uniquely identify multiple mRNAs present in a single GenBank file.

clone obtained from an exhaustive screening of a cDNA library. In many, if not most of these cases, the 5' end of the longest cDNA is likely to correspond to the transcription start site. A terminal G residue is often found at the 5' end of the cDNA that is not found at the corresponding position in the genomic sequence; it is thought that this G is copied from the 5' methyl G cap that is added post-transcriptional to the 5' end of eukaryotic mRNAs. Sequences from *Drosophila* species other than *D. melanogaster* were not included because their orthologous counterparts in *D. melanogaster* are almost always represented in

the database. Many *Drosophila* genes contain multiple mRNAs arising from alternative promoters and/or RNA processing. In cases where alternative leader sequences have been clearly documented, more than one leader sequence is listed for a particular gene. If such alternative leaders share the same translation start site, the start context (-6 to +4) is listed for just one of the leader sequences. Several genes have been characterized by more than one research group and reported to GenBank. We have arbitrarily used the information and GenBank file name given for one of the duplicate entries if the data are equivalent. Where data are not equivalent for the same gene we have chosen the data which are most strongly supported by experimental evidence.

### Leader Length and Upstream AUGs

Inspection of Table 36.1 reveals numerous mRNAs with leader sequences exceeding 100 nt and containing multiple upstream start codons (uAUGs). Indeed the average *Drosophila* leader sequence is 248 nt and the median is 156. The distribution of leader lengths is shown in Fig. 36.1. Forty-six of the leader sequences exceed 500 nt. The smallest size class (0-100 nt) is the largest containing 140 mRNA sequences. Many of the reported leader sequences are based upon the analysis of the longest cDNA obtained but may nonetheless

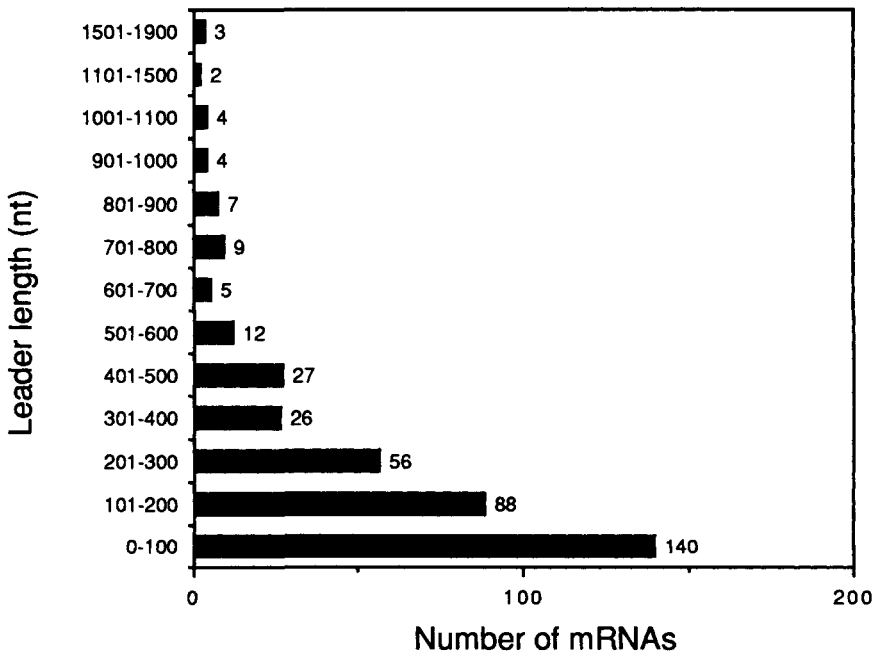


FIG. 36.1. Distribution of the number of nucleotides (nt) in the 5' untranslated leader sequences of *Drosophila* mRNAs.

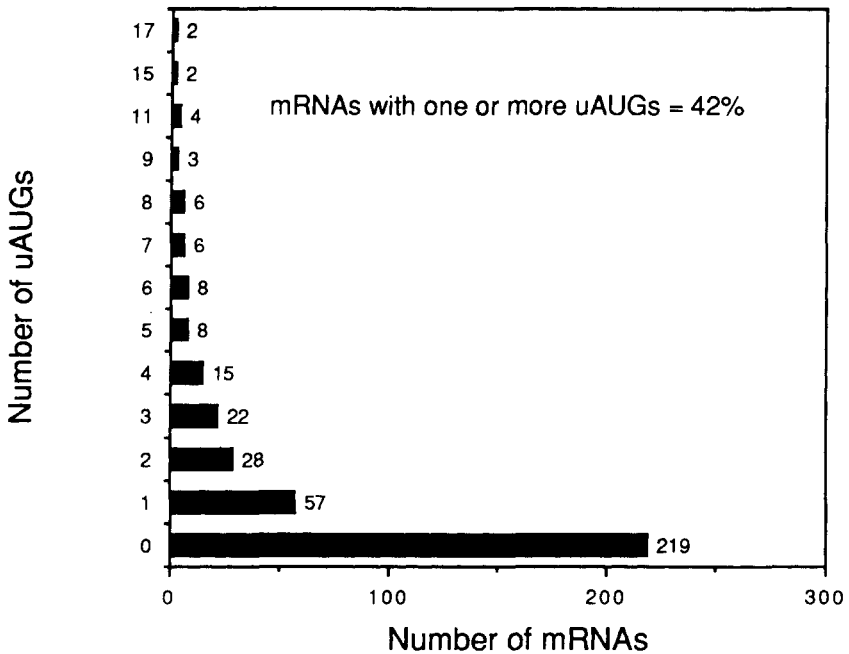


FIG. 36.2. Distribution of upstream AUGs in the 5' untranslated leader sequences of *Drosophila* mRNAs.

lack the complete 5' end. Therefore, these global leader sequence statistics most likely underestimate the true values.

Unquestionably the most surprising result of our analysis is that 42% of all *Drosophila* mRNAs contain one or more uAUGs in their leader sequence (Fig. 36.2). The majority of mRNAs containing uAUGs contain more than one. Indeed 10% of all *Drosophila* mRNAs surveyed contain five or more uAUGs. The vast majority of *Drosophila* uAUGs are followed by a short (ca. 1–100) open reading frame which terminates before reaching the major translation start site (data not shown). *Drosophila* uAUGs do not exhibit a similar preference for specific flanking nucleotides as exhibited by major start codons (see below). Nonetheless, many of the uAUGs (if not the majority) contain flanking sequences that are compatible with a good translation initiation site.

Previously, Kozak (1991) reported that approximately 9% of vertebrate mRNAs contain uAUGs and further noted that the majority of these unusual mRNAs encoded regulatory proteins (e.g., transcription factors and proto-oncogenes, receptors, and components of signal transduction). *Drosophila* appears similar to vertebrates in this respect as typified by the long leader-uAUG laden mRNAs encoding *Antennapedia*, ecdysone receptor, acetylcholine receptor, decapentaplegic, seven in absentia, and protein kinase C (Table 36.1). In general long leader-uAUG laden mRNAs encode low abundant proteins, particularly as compared to very short-leader mRNAs encoding such

proteins as the yolk, cuticle, and larval serum proteins. This dichotomy is consistent with the general finding that removal of long leader sequences typically increases translation initiation rates (e.g. Chinkers et al. 1989; Muller and Witte 1989). Long leader mRNAs may be tolerated as a consequence of the absence of natural selection to increase translation rates of proteins that are not needed in abundance. Alternatively, the presence of a long leader with multiple uAUGs may afford devices to regulate translation initiation. The paradigm par excellence in this regard is the yeast GCN4 gene. GCN4 mRNA is constitutively produced but the translation of GCN4 protein is highly regulated through the interaction of four upstream open reading frames, the scanning preinitiation complex, and some of the translation initiation factors that undergo changes in activity as a consequence of amino deprivation (Miller and Hinnebusch 1989; Ramirez et al. 1991; Dever et al. 1992). Whether other eukaryotic mRNAs that contain long leader and uAUGs are under similar control is unknown. For many of the long-leader *Drosophila* genes, mRNA and protein expression are temporally and spatially correlated suggesting the lack of translational regulation. However, it should be noted that translation initiation rates are almost never determined empirically. Consequently, the relative translation rates among different mRNAs cannot be compared at this time.

OH and coworkers have recently reported (OH et al. 1992; and personal communication) that the long-leader sequences of the *Antennapedia* and *Ultrabithorax* can promote internal ribosome binding in *Drosophila* cell culture. Since internal binding circumvents the requirement for the cap-binding protein (eIF-4E), it is likely that internal initiation also circumvents global translation control as mediated by altering the level and activity of eIF-4E. It will be interesting to see if *Antp* and *Ubx* use an internal mode of initiation in flies and whether internal initiation is used by other mRNAs with exceptionally long leader sequences.

## Translation Start Sites

Table 36.2 presents an update of the translation start sites from positions  $-6$  to  $+4$  relative to the start codon. The 50/75 consensus rule (Cavener and Ray 1991) was used to assign consensus nucleotides. The derived consensus sequence, C/A A N N AUG has not appreciably changed with the doubling of the database from that reported by Cavener and Ray (1991). Since long leader mRNAs are thought to be poorer substrates for translation initiation, such mRNAs may on average contain a poorer fit to the consensus sequence. To examine this question the mRNA sequences listed in Table 36.1 were divided into two groups: long leaders, exceeding the median leader length and short leaders, less than the median leader length (Table 36.2). The short leader mRNAs do exhibit a significantly stronger preference for A or G at the critical  $-3$  position as compared to the long leader mRNAs as might be expected. In addition, differences are observed between the short and long leader classes at

TABLE 36.2. Nucleotide frequencies flanking the start codons for the major protein coding regions and the start site consensus sequences

Total mRNA dataset.

	-6	-5	-4	-3	-2	-1	+1	+2	+3	+4
A	33	23	26	70	47	41	100	0	0	23
G	28	18	11	20	11	20	0	0	100	39
C	17	34	51	6	24	30	0	0	0	15
U	22	25	12	5	18	9	0	100	0	23
	a	c	C/A	A	a	a	A	U	G	g

Short leader mRNA dataset.

	-6	-5	-4	-3	-2	-1	+1	+2	+3	+4
A	34	22	29	77	51	39	100	0	0	26
G	24	16	9	15	14	17	0	0	100	35
C	18	30	54	3	18	35	0	0	0	14
U	23	32	8	4	16	9	0	100	0	25
	a	u	C/A	A	A	A/C	A	U	G	g

Long leader mRNA dataset.

	-6	-5	-4	-3	-2	-1	+1	+2	+3	+4
A	34	24	23	66	43	45	99	0	0	22
G	32	22	13	20	10	23	1	0	100	42
C	16	38	48	10	29	23	0	0	0	16
U	19	16	16	4	19	9	0	100	0	20
	a	c	c	A	a	a	A	U	G	g

Upstream AUGs (uAUGs).

	-6	-5	-4	-3	-2	-1	+1	+2	+3	+4
A	32	28	27	29	45	40	100	0	0	30
G	23	18	35	15	17	16	0	0	100	9
C	20	21	16	21	25	14	0	0	0	26
U	25	34	22	35	13	30	0	100	0	35
	a	u	g	u	a	a	A	U	G	u

The 50/75 Consensus Rule was applied: if the frequency of one nucleotide is greater than 50% and is greater than twice the frequency of the next highest nucleotide, it is assigned as the consensus and denoted as such with a capital letter (e.g., A). If the sum of the frequency of the two most frequent nucleotides is greater than 75% but neither meet the requirement for singular consensus, the two nucleotides are assigned as co-consensus nucleotides and denoted with capital letters (e.g., C/A). Lower case letters indicate the most frequent nucleotide at a particular position when no nucleotides meet the consensus criteria.

positions  $-4$ ,  $-2$  and  $-1$ . However, these are largely differences in the relative distribution of A and C, both favored at these positions in most eukaryotic groups (Cavener and Ray 1991). Overall, the differences between long and short leader mRNAs translation start sites are significant but minor. Among the long leader mRNAs occurs an exceptional GUG start codon for choline acetyltransferase. The data supporting the use of GUG as a start codon in this case are strong (Sugihara et al. 1990). Preliminary evidence indicates that the E74A gene uses CUG as a major alternative start codon (L. Boyd and C. Thummel, personal communication). Non-AUG start codons are likely to be more prevalent than currently recognized because most AUG translation start sites have not been empirically confirmed.

The sequence context flanking 151 upstream AUGs was examined in order to see if uAUGs lie in a poor context. It might be expected that uAUGs would exhibit a strong anti-consensus sequence to discourage translation initiation at these sites. However, the summary of these data in Table 36.1 indicates that uAUGs collectively neither show a good or poor fit to consensus relative to major translation start sites. At the critical  $-3$  position A or G is found in 44% of the cases. The frequency of A at  $-2$  and  $-1$  is relatively high similar to major translation initiation sites. Some unique biases are observed including a relatively high frequency of G at  $-4$  and a relatively low frequency of G at  $+4$ ; just the opposite biases are seen for major translation initiation sites. One possible explanation for the lack of consensus either opposite or similar to major translation initiation sites is that uAUG context data may contain a mixture of uAUG which are either selected for or against as initiation sites depending upon the mRNA. This assumes that some uAUGs may be involved in translational regulation but that others are not. Overall, a large fraction of uAUGs would appear to be in a reasonably good context. How the scanning preinitiation complex traverses a leader sequence burdened with uAUGs is an interesting mechanistic and regulatory question.

### A Caveat to Using the Translation Start Site Consensus

Comparing putative translation start sites of newly sequenced genes with the start site consensus sequence is a common practice. In some cases investigators have favored a downstream start codon over an in-frame upstream start codon based upon a better fit of the former to the consensus sequence. However, mutational analysis of the translation start site of *Adh* (Feng et al. 1991) and inspection of the diversity of start contexts in Table 36.1 demonstrates that start codons that exhibit a poor fit to the consensus can nonetheless serve as the major site of translation initiation. A good example of this is provided by the translation start site for *hsf* encoding the *Drosophila* heat-shock transcription factor (Table 36.1). The start codon context for *hsf* is UUUAUGU (Clos et al. 1990). Based upon mutational analysis and the consensus sequence, a UUUAUGU context is exceptionally poor. Changing the start codon context for *Adh* to this same sequence resulted in a 6–12-fold reduction in translation



depending upon the developmental stage. However, an appreciable level of ADH protein was still observed in this mutant. Thus a "poor" context may reduce but not necessarily eliminate translation initiation. Kozak's studies on the rat preproinsulin mRNA have clearly indicated that a poor context reduces the probability that the ribosome will initiate at that particular site. If initiation does not occur at a particular start codon, the preinitiation complex resumes scanning, a process called leaky scanning. The overall effect of leaky scanning may be the use of multiple start codons, particularly when two start codons are in-frame and within close proximity. An important perspective to bear in mind when analyzing translation start sites is that the sequence context may be adapted to down-regulate the rate of translation initiation. Moreover, sequence context effects are likely to be developmentally dependent (Feng et al. 1991) as a function of changes in the concentration and activities of the translation initiation factors (particularly eIF-2). These considerations are also relevant to the presence of upstream start codons in the leader sequence which are either out of frame with the major coding region or followed by an in-frame termination codon.

## Summary

*Drosophila* genes exhibit a diverse array of untranslated leader sequences and translation start sites. The presence of a long leader or multiple uAUGS or a poor sequence context surrounding the start codon should no longer be perceived as abnormal or unusual given the large fraction of *Drosophila* mRNAs which contain such features. In many cases these features will affect translation initiation rates. How they affect translation and what the physiological rationale is for these effects remain to be elucidated. Although it would appear that *Drosophila* mRNAs are more prone to long leaders and uAUGs than vertebrate genes, only a small fraction of all mRNAs have been characterized for either group. The current *Drosophila* and vertebrate databases are biased somewhat differently as a consequence of the types of genes and questions being analyzed using different systems. In particular the *Drosophila* database contains a larger fraction of genes encoding proteins that regulate development. Whether the current *Drosophila* database is a more representative sample than the vertebrate database is unknown. Fortunately our obsession for cloning and sequencing will eventually answer this question.

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## Codon Usage

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### **Introduction**

In most genes in most species, alternative synonymous codons are not used in equal frequencies (Aota et al. 1988)—*Drosophila* is no exception (Ashburner et al. 1984; O'Connell and Rosbash 1984; Shields et al. 1988). In Table 37.1 we present the total codon usage for 438 *D. melanogaster* genes. This dataset was extracted from the GenBank/EMBL/DDBJ DNA sequence data library (GenBank release 71) using the ACNUC sequence retrieval software (Gouy et al. 1985), and screened to remove duplicates and/or multiple alleles (making particular use of FlyBase; Ashburner 1992); the genes are listed in Appendix A. As we discuss below, there is considerable heterogeneity of codon usage patterns among genes, and so the values in Table 37.1 must be taken only as an overall, or average, guide to *D. melanogaster* codon usage. This may be useful in the design of oligonucleotide probes, or in the assessment of whether a novel open reading frame is actually a coding sequence: many genes approximate to this pattern, but particular genes may differ quite markedly. Note also that genes from transposable elements have rather different patterns of codon usage from “chromosomal” genes (Shields and Sharp 1989), and so they are presented separately in Table 37.1 (to be discussed in more detail below). (Codon usage in the *Drosophila* mitochondrial genome is completely different from nuclear genes (Clary and Wolstenholme 1985), and will not be discussed further.)

Why is codon usage in *Drosophila* biased, and why does it vary among genes? Clearly, the pattern of synonymous codon usage in a gene must reflect the net result of past evolutionary pressures: the two main influences are natural selection (some codons may be translated more accurately and/or efficiently than synonyms encoding the same amino acid), and mutational biases (which may give rise to strongly biased codon usage even in the absence of any selection). Thus, in general terms, codon usage can be considered as the result

TABLE 37.1. Codon usage in *Drosophila melanogaster*

	Total		T.E.			Total		T.E.			Total		T.E.			Total		T.E.					
	N	RSCU	N	RSCU		N	RSCU	N	RSCU		N	RSCU	N	RSCU		N	RSCU	N	RSCU	N	RSCU		
Phe	UUU	2,896	0.66	478	1.17	Ser	UCU	1,745	0.49	226	0.95	Tyr	UAU	2,610	0.69	341	0.96	Cys	UGU	1,396	0.55	167	0.82
	UUC	5,822	1.34	336	0.83		UCC	5,377	1.50	216	0.91		UAC	4,957	1.31	368	1.04		UGC	3,712	1.45	238	1.18
Leu	UUA	922	0.26	430	1.30		UCA	1,857	0.52	297	1.25	ter	UAA	220	1.60	2	0.60	ter	UGA	75	0.55	3	0.90
	UUG	3,787	1.05	319	0.96		UCG	4,705	1.31	153	0.64	ter	UAG	117	0.85	5	1.50	Trp	UGG	2,380	1.00	223	1.00
Leu	CUU	1,959	0.54	364	1.10	Pro	CCU	1,683	0.46	195	0.78	His	CAU	2,814	0.78	295	1.01	Arg	CGU	2,480	1.08	128	0.66
	CUC	3,410	0.94	256	0.77		CCC	5,087	1.39	204	0.82		CAC	4,363	1.22	292	0.99		CGC	4,866	2.13	134	0.69
	CUA	1,758	0.49	337	1.02		CCA	3,484	0.95	437	1.76	Gln	CAA	3,984	0.55	580	1.28		CGA	1,964	0.86	197	1.02
	CUG	9,835	2.72	283	0.85		CCG	4,436	1.21	160	0.64		CAG	10,537	1.45	328	0.72		CGG	1,941	0.85	103	0.53
Ile	AUU	4,065	0.97	583	1.16	Thr	ACU	2,232	0.60	344	1.04	Asn	AAU	5,565	0.84	747	1.10	Ser	AGU	2,704	0.75	255	1.07
	AUC	6,523	1.56	349	0.69		ACC	6,152	1.67	307	0.93		AAC	7,692	1.16	614	0.90		AGC	5,174	1.44	283	1.19
	AUA	1,947	0.47	581	1.15		ACA	2,773	0.75	513	1.55	Lys	AAA	3,759	0.51	1,115	1.36	Arg	AGA	1,041	0.45	405	2.09
Met	AUG	6,366	1.00	335	1.00		ACG	3,611	0.98	163	0.49		AAG	11,038	1.49	519	0.64		AGG	1,440	0.63	195	1.01
Val	GUU	2,757	0.72	290	1.07	Ala	GCU	4,022	0.78	343	1.11	Asp	GAU	7,274	1.05	497	0.97	Gly	GGU	4,344	0.92	222	1.05
	GUC	3,839	1.00	223	0.82		GCC	9,783	1.89	319	1.03		GAC	6,631	0.95	528	1.03		GGC	8,179	1.73	218	1.03
	GUA	1,450	0.38	300	1.11		GCA	3,265	0.63	409	1.33	Glu	GAA	4,778	0.58	808	1.30		GGA	5,208	1.10	294	1.39
	GUG	7,234	1.89	271	1.00		GCG	3,601	0.70	163	0.53		GAG	11,655	1.42	435	0.70		GGG	1,140	0.24	113	0.53

“Total” indicates summed codon usage for 438 nuclear chromosomal genes (i.e., excluding transposable elements), a total of 264,421 codons. “T.E.” indicates summed codon usage for 30 genes from 16 transposable elements (listed in Appendix 37.B), a total of 20,836 codons. Codon usage is presented as *N* (the observed number of occurrences) and RSCU (the relative synonymous codon usage, obtained by dividing *N* by the average value for the amino acid); the RSCU value is useful for comparing the level of bias among different amino acids, or among data sets of different sizes.

of a selection-mutation balance (Sharp and Li 1986; Bulmer 1991). However, while it is clear that selection among synonyms shapes codon usage in certain prokaryotes and unicellular eukaryotes (reviewed in Ikemura 1985; Andersson and Kurland 1990), it is not obvious how widespread selective codon usage may be in the genomes of multicellular organisms. In particular, it is not clear whether the long-term evolutionary effective population sizes of most multicellular species are large enough for the selective differences between alternative synonymous codons (which are expected to be very small) to overcome random genetic drift (Sharp 1989). In an earlier study (Shields et al. 1988), we concluded (somewhat to our surprise!) that the evidence suggests that codon usage in many *D. melanogaster* genes is influenced by natural selection. Here we briefly review that evidence, utilizing the much larger *D. melanogaster* gene sequence data set now available.

### Codon usage variation among genes

Codon usage patterns vary considerably among *D. melanogaster* genes (Shields et al. 1988). To take an extreme example, 92% (33/36) of the Leu residues in the enolase phosphoglycerate hydrolase gene (*Eno*) are encoded by CUG; in contrast, the cubitus interruptus Zn finger gene (*ci*) uses this codon in only seven of 91 cases (8%); differences are also seen for all other 17 amino acids where there is a choice of codons. Under the selection-mutation balance model, two possible reasons for this variation stand out. If selection among synonymous codons for translational efficiency occurs in *D. melanogaster*, then the strength of selection is likely to vary among genes, depending on their level (and perhaps also tissue and developmental stage) of expression. For example, in *Escherichia coli* (Gouy and Gautier 1982) and *Saccharomyces cerevisiae* (Sharp et al. 1986) the strength of codon usage bias in a gene is very highly correlated with the level of gene expression. Alternatively, or perhaps additionally, genes may be affected by different mutational biases. For example, mammalian genes vary greatly in base composition (G + C content) at silent sites (and thus in codon usage) depending on the local base composition of the chromosome (Bernardi et al. 1985; Ikemura 1985); this variation can be most simply explained as variation in the mutation pattern around the genome (Filipski 1988; Sueoka 1988; Wolfe et al. 1989).

To elucidate the situation in *Drosophila*, the first step is to characterize the nature of the codon usage variation among genes. Since the codon usage pattern of each gene is a composite of 59 values (one for each codon, less Met, Trp and stop codons), it is necessary to use multivariate statistical analyses. In codon usage studies the most commonly used method is correspondence analysis (pioneered by Grantham et al. 1981). It is not appropriate to go into any details of the method here, except to say that it allows definition of the major trends among genes—see Grantham et al. (1981) or Shields et al. (1988) for more discussion of this method. We applied this approach to 84 genes (Shields et al. 1988) and have also used it on a data set of 438 genes here. In each case, the

major variation in synonymous codon usage among genes is found to be strongly associated with G + C content at silent sites ( $GC_S$ ): genes at one end of the trend have relatively unbiased codon usage, while genes at the other end of the trend have very highly biased codon usage, and high values of  $GC_S$ .

This seems very like the situation found with mammalian (e.g., human) genes, but in fact there are several important differences (Shields et al. 1988). Some of these become apparent from a comparison of  $GC_S$ , the G + C content at silent third positions of codons (i.e., excluding Met and Trp) in a gene, and  $GC_1$ , the G + C content in the introns of a gene. First, in *D. melanogaster* (unlike humans)  $GC_S$  is not strongly correlated with  $GC_1$ . Second,  $GC_S$  values are generally much higher than  $GC_1$  values, particularly in genes with very biased codon usage. It is also noticeable that  $GC_S$  becomes reduced in pseudogenes (Shields et al. 1988; Moriyama and Gojobori 1992). Most of the *D. melanogaster* genes studied have been mapped, and there is no obvious relationship between  $GC_S$  and map position, although local variations in base composition on the scale that they are thought to occur in the human genome would be difficult to detect at this level. However, it is clear that neighboring genes can have quite different  $GC_S$  values. For example, in the highly biased alcohol dehydrogenase gene  $GC_S$  is 0.77, but in the relatively unbiased *Adh*-related gene (less than 300 bp away; Kreitman and Hudson 1991)  $GC_S$  is only 0.53. It is also noticeable that in human genes the trend in G + C content is due to similar changes in the frequency of both C and G, but in *D. melanogaster* the major trend is more specifically (though not exclusively) due to a change in the frequency of C. Thus, the major variation in  $GC_S$  in *Drosophila* does not appear to be due to regional chromosomal base composition differences.

On the other hand, the main trend in codon usage differences among genes may be correlated with expression level. We might expect that the highly biased genes at the G + C-rich extreme would be those under the most selection pressure, particularly as their  $GC_S$  values are the most different from noncoding DNA (i.e., introns). Of course, in a multicellular organism with a complex series of developmental stages, it is rather more difficult to quantify "expression level" than it would be in *E. coli* or yeast. Nevertheless, the G + C-rich genes do seem to include many genes that can be identified as highly expressed. For example, one is alcohol dehydrogenase: *Adh* mRNA "accounts for about 1–2% of the translational activity of mRNA from adult flies" (Benyajati et al. 1980), and must be considered a highly expressed gene. Others at this extreme include *Yp1* and *Yp2* encoding yolk proteins, the nine ribosomal protein genes in the data set, and genes encoding actins and cuticle proteins; all such genes were considered by O'Connell and Rosbash (1984) to be "abundantly expressed".

### **"Optimal" Codons in *Drosophila melanogaster***

If it is true that the major trend among genes in codon usage is associated with expression-level-mediated selection on codon usage, then contrasting the codon usage patterns for the genes at either end of this trend should reveal which

particular codons for each amino acid are favoured. Codon usage in 10% of genes at each extreme of this trend (as identified by multivariate statistical analysis) is presented in Table 37.2. There are 23 codons used with (significantly) higher frequency in the highly biased group of genes: 22 of these are here defined as “optimal” codons, the exception being GGU (for Gly), where the difference in RSCU values is small (though significant at the 5% level). These optimal codons are G + C-rich: of the 22, 15 end in C and six end in G—only one ends in U (CGU) and none end in A. Interestingly, CGU appears to be an optimal Arg codon in many other species (Sharp et al. 1992).

A simple measure of the strength of species-specific codon usage bias is given by the frequency of optimal codons ( $F_{op}$ ) in a gene (Ikemura 1985). We define a  $F_{op}$  for *D. melanogaster* as the number of occurrences of these 22 optimal codons (Table 37.2), divided by the total number of occurrences of codons for these 18 amino acids (i.e., excluding Met and Trp codons). (Calculation of  $F_{op}$  values is an option in the FORTRAN program CODONS (Lloyd and Sharp 1992), which is available from the authors on request.)  $F_{op}$  values for these 438 genes are given in Appendix A, and they range from 0.22 (*Scr* encoding the sex combs reduced homeobox protein) to 0.88 (*Lsp1-b* encoding  $\beta$ -larval serum protein).

While we have already alluded to the difficulties in discussing absolute expression levels, it is nevertheless possible to compare  $F_{op}$  values among genes whose relative expression levels have been described. There are two cytochrome c genes, and it is known that *Cyt-c2* ( $F_{op} = 0.77$ ) “is expressed at much higher levels than” *Cyt-c1* ( $F_{op} = 0.57$ ) (see Limbach and Wu 1985); among four  $\alpha$ -tubulin genes, the transcript of *Tuba84D* ( $\alpha$ -1) ( $F_{op} = 0.79$ ) is “much more abundant” than that of *Tuba84E* ( $\alpha$ -2) ( $F_{op} = 0.69$ ) (see Kalfayan and Wensink 1982); of two elongation factor 1 $\alpha$  genes, expression of *Efla100E* ( $F_{op} = 0.76$ ) is “generally markedly stronger” than that of *Efla48D* ( $F_{op} = 0.71$ ) (see Hovemann et al. 1988); and there are two lysozyme genes, *LysP* ( $F_{op} = 0.63$ ) whose expression was only detected in adults, and whose expression in the adult “was low compared to that of *LysD*” ( $F_{op} = 0.70$ ) (see Kylsten et al. 1992). In some cases these differences in  $F_{op}$  values are quite small—the genes’ similarity in sequence may reflect quite recent gene duplication events; nevertheless, the differences are all in the direction predicted.

In Table 37.2, it is interesting to note that the highly biased (and highly expressed?) genes favour the most A + T-rich stop codon (UAA), even though the rest of their codons are generally G + C-rich. The highly biased genes also appear to avoid UGA, which is more common in the low bias genes. This is reminiscent of the pattern of stop codon usage in genes of high and low expression in *E. coli*, *Bacillus subtilis*, and yeast (Sharp et al. 1992), and lends further credence to the idea that codon usage in *D. melanogaster* is influenced by natural selection.

## Transposable Element Genes

Codon usage in the open reading frames (ORFs) of the various transposable elements (TEs) found in the *D. melanogaster* genome (see Appendix B) is

TABLE 37.2. Codon usage in high and low bias genes in *D. melanogaster*

	<i>High</i>		<i>Low</i>			<i>High</i>		<i>Low</i>			<i>High</i>		<i>Low</i>			<i>High</i>		<i>Low</i>	
	N	RSCU	N	RSCU		N	RSCU	N	RSCU		N	RSCU	N	RSCU		N	RSCU	N	RSCU
Phe UUU	37	0.16	541	1.14	Ser UCU	73	0.51	407	0.99	Tyr UAU	59	0.25	365	1.03	Cys UGU	18	0.19	206	0.81
UUC*	421	1.84	406	0.86	UCC*	412	2.86	476	1.16	UAC*	420	1.75	345	0.97	UGC*	169	1.81	305	1.19
Leu UUA	6	0.04	334	0.93	UCA	13	0.09	423	1.03	ter UAA	32	2.29	24	1.76	ter UGA	0	0.00	10	0.73
UUG	101	0.60	503	1.40	UCG*	176	1.22	323	0.79	ter UAG	10	0.71	7	0.51	Trp UGG	147	1.00	196	1.00
Leu CUU	39	0.23	370	1.03	Pro CCU	44	0.30	298	0.87	His CAU	46	0.35	368	1.06	Arg CGU*	227	2.20	204	0.99
CUC*	149	0.88	190	0.53	CCC*	385	2.61	320	0.94	CAC*	215	1.65	324	0.94	CGC*	327	3.17	225	1.10
CUA	17	0.10	282	0.79	CCA	85	0.58	474	1.39	Gln CAA	44	0.16	699	0.97	CGA	16	0.16	264	1.29
CUG*	703	4.16	475	1.32	CCG	75	0.51	276	0.81	CAG*	501	1.84	743	1.03	CGG	13	0.13	146	0.71
Ile AUU	147	0.56	576	1.31	Thr ACU	76	0.38	390	0.96	Asn AAU	73	0.25	793	1.10	Ser AGU	17	0.12	426	1.04
AUC*	633	2.43	340	0.77	ACC*	637	3.21	425	1.05	AAC*	513	1.75	645	0.90	AGC	174	1.21	404	0.99
AUA	2	0.01	401	0.91	ACA	25	0.13	507	1.25	Lys AAA	38	0.09	775	0.99	Arg AGA	4	0.04	220	1.07
Met AUG	301	1.00	549	1.00	ACG	55	0.28	301	0.74	AAG*	813	1.91	798	1.01	AGG	32	0.31	172	0.84
Val GUU	129	0.54	446	1.18	Ala GCU	267	0.87	558	1.20	Asp GAU	284	0.77	903	1.29	Gly GGU	316	1.19	426	1.02
GUC*	346	1.46	300	0.79	GCC*	854	2.79	539	1.15	GAC*	457	1.23	493	0.71	GGC*	505	1.91	450	1.08
GUA	19	0.08	278	0.73	GCA	41	0.13	491	1.05	Glu GAA	75	0.17	885	1.10	GGA	230	0.87	619	1.49
GUG*	453	1.91	492	1.30	GCG	62	0.20	279	0.60	GAG*	809	1.83	728	0.90	GGG	7	0.03	169	0.41

"High" and "Low" denote groups of genes with high and low codon usage bias; they are the 10% of genes at each extreme of the major codon usage trend among genes (identified by multivariate statistical analysis). Twenty-two codons defined as "optimal" (see text) are indicated by \*. The High and Low groups each comprise 44 genes, and total 13,374 and 26,307 codons, respectively. *N* and RSCU are explained in Table 37.1.



different, overall, from that of "chromosomal" genes (Table 37.1). The TE ORFs are more similar to the low bias genes than the high bias genes (Table 37.2), and exhibit very little evidence of selected codon usage.

However, as with chromosomal genes, codon usage varies greatly among TE ORFs: in general, ORFs from the same TE have rather similar codon usage patterns, but ORFs from different TEs have different codon usage patterns (this is apparent, to some extent, in the GC<sub>s</sub> values in Appendix B, but see Shields and Sharp (1989) for more details). This observation is most simply explained if the TEs have been subject to different mutational biases, and we consider two possible scenarios. Since TEs appear to have been subject to occasional horizontal transfer among species, their base composition could reflect different mutation biases in different previous host genomes. However, it seems rather more likely that the differences reflect current/ongoing differences in mutation pattern. For many TEs, movement around the genome involves an RNA intermediate which is then subject to a (quite highly error prone) reverse transcription process. The different TEs have reverse transcriptases which differ considerably in their primary amino-acid sequences (Xiong and Eickbush 1990), and it is quite likely that each reverse transcriptase has a slightly different error propensity which leads to different mutational spectra, and ultimately to different base composition and codon usage (Shields and Sharp 1989).

## Conclusions

We have concluded above that *Drosophila melanogaster* genes are subject to different levels of codon selection. This seems to be corroborated by the observation that silent sites in genes with high codon usage bias have diverged to a lesser extent between *D. melanogaster* and other related species (e.g., *D. simulans* and *D. pseudoobscura*), suggesting that there is more constraint on codon usage in the highly biased genes (Sharp and Li 1989). In a recent examination of silent site base composition and substitution rates, Moriyama and Gojobori (1992) suggested that the variation in each can be explained by mutational biases, in a manner consistent with the situation in mammalian genes (Wolfe et al. 1989). However, we have outlined many discrepancies between the observations relating to *Drosophila* and mammals which make a similar explanation unlikely. We have detailed some cases where it seems that the strength of codon usage bias can be correlated with the level of gene expression—it will be of particular interest to investigate whether any of the heterogeneity in codon usage among genes can be related to the genes' tissue or time of expression. Certainly, while we have discussed the major pattern of codon usage variation among genes, we do not exclude the possibility that there are other (as yet undefined) trends which explain some further part of the heterogeneity in these data.

Another question of interest concerns the extent to which a similar pattern is found in other species of *Drosophila*. Codon usage differs among *Adh* genes derived from various *Drosophila* species (Starmer and Sullivan 1989).

Interestingly, Moriyama and Gojobori (1992) reported that in the *Adh* gene of Hawaiian *Drosophila*, GC<sub>s</sub> is low and the silent substitution rate is high (see also Thomas and Hunt 1991); these two observations can be consistently explained if codon selection has been relaxed in that lineage, due possibly to a small effective population size caused by several bottleneck events. It will be interesting to examine to what extent (and to ask why) codon usage patterns generally vary among *Drosophila* species.

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### Appendix 37.A: Codon Usage Bias in *D. melanogaster* Genes

Gene	Function/Product	Map	AA	GC <sub>S</sub>	GC <sub>I</sub>	F <sub>op</sub>	Acc.#
<i>Abd-A</i>	abdominal-A: homeodomain TF	3-58.8	330	0.72		0.62	X54453
<i>Abd-B</i>	abdominal-B: homeodomain TF	3-58.8	491	0.73		0.62	X16134
<i>Abl</i>	abl-oncogene analog: Tyr kinase	3-[44]	1,520	0.69	0.40	0.57	M19692
<i>ac</i>	achaete: T5 AHLH protein	1-0.0	201	0.52		0.41	M17120
<i>Ace</i>	acetylcholinesterase	3-52.5	649	0.74		0.64	X05893
<i>Acp70A</i>	male accessory gland protein	3-[40]	55	0.47		0.37	M21201
<i>Acr60C</i>	muscarinic acetylcholine receptor C	2-[107]	788	0.79		0.62	M23412
<i>Acr64B</i>	nicotinic acetylcholine receptor D	3-[8]	521	0.72		0.62	X04016
<i>Acr96Aa</i>	nicotinic acetylcholine receptor B	3-[83]	567	0.75		0.66	X07194
<i>Acr96Ab</i>	nicotinic acetylcholine receptor E	3-[83]	535*	0.70		0.61	X52274
<i>Act5C</i>	actin	1-[14]	376	0.78		0.78	K00667
<i>Act42A</i>	actin	2-[55.2]	376	0.68		0.64	K00670
<i>Act57A</i>	actin	2-[92]	376	0.76		0.79	K00673
<i>Act79B</i>	actin	3-[47]	376	0.82	0.33	0.80	M18829
<i>Act87E</i>	actin	3-[53]	376	0.76		0.77	K00674
<i>Act88F</i>	actin	3-57.1	376	0.80	0.48	0.79	M18830
<i>Actn</i>	sarcomeric $\alpha$ actinin	1-[0.5]	895	0.81		0.76	X51753
<i>ade3</i>	glycinamide ligase	2-[22]	434	0.65	0.42	0.55	J02527
<i>Adfl</i>	Adh distal factor 1: AHLH protein	2-[56]	253	0.74		0.67	M37787
<i>Adh</i>	alcohol dehydrogenase	2-50.1	256	0.81	0.39	0.77	J01066
<i>Adhr</i>	alcohol dehydrogenase related	2-50.1	272	0.53		0.43	
<i>Ald</i>	fructose-1,6-biphosphate aldolase	3-91.5	363	0.82	0.40	0.82	M76409
<i>ama</i>	amalgam protein	3-[47.5]	333	0.77	0.28	0.66	M23561
<i>amd</i>	$\alpha$ -methyl-dopa hypersensitivity	2-53.9	510	0.67	0.38	0.58	X04695
<i>Amy-d</i>	$\alpha$ -amylase 1	2-77.7	494	0.88		0.82	X04569
<i>AnnIX</i>	annexin IX	3-[70]	296*	0.87		0.81	M34068
<i>AnnX</i>	annexin X	1-[64]	321	0.87		0.78	M34069
<i>annon-77F</i>	histone-like protein	3-[46]	215	0.56	0.42	0.48	X16962
<i>Anr</i>	andropin: male-specific protein	3-[101]	57	0.46	0.26	0.39	X16972
<i>Antp</i>	antennapedia: homeodomain TF	3-47.5	378	0.75		0.63	X03791
<i>App1</i>	$\beta$ -amyloid-like gene	1-0.0	886	0.71		0.63	J04516
<i>Aprt</i>	adenine phosphoribosyltransferase	3-1.5	183	0.64		0.57	M18432
<i>arl</i>	arf-like: GTP-binding protein	3-[43]	180	0.81	0.40	0.74	M61127
<i>arm</i>	armadillo	1-[0.4]	843	0.64	0.47	0.57	X54468
<i>Arr1</i>	arrestin A/phosphorestin II	2-[53]	364	0.76	0.32	0.71	M30177
<i>Arr2</i>	arrestin B/phosphorestin I	3-[26]	401	0.76	0.34	0.70	M32141
<i>ase</i>	asense: T8 AHLH protein	1-0.0	396	0.52		0.41	X12550
<i>Atpa</i>	Na/K-ATPase $\alpha$ subunit	3-[70]	1,038	0.70		0.66	X14476

(continued)

APPENDIX 37.A. *Continued.*

<i>Gene</i>	<i>Function/Product</i>	<i>Map</i>	<i>AA</i>	<i>GC<sub>S</sub></i>	<i>GC<sub>I</sub></i>	<i>F<sub>op</sub></i>	<i>Acc.#</i>
<i>awd</i>	abnormal wing discs	3-[105]	153	0.89		0.86	X13107
<i>B</i>	Bar: homeodomain protein	1-57.0	543	0.70		0.56	M73079
<i>bam</i>	bag-of-marbles	3-[85]	442	0.64	0.33	0.54	X56202
<i>bcd</i>	bicoid: homeodomain TF	3-[47.5]	489	0.66		0.54	X14458
<i>Bd</i>	Beaded: EGF-like transmembrane P	3-92.5	1,408	0.66		0.55	X56811
<i>BicD</i>	bicaudal D $\alpha$ -helical coiled coil	2-52.9	782	0.67		0.59	M31684
<i>Bjl</i>	chromatin-binding protein	3-[20]	547	0.68	0.40	0.58	X58530
<i>boss</i>	bride of sevenless: transmembrane P	3-[89]	896*	0.61	0.35	0.53	X55887
<i>br</i>	broad: Zn finger protein	1-[0.4]	704	0.80		0.67	X54664
<i>brm</i>	brahma: homeotic regulator	3-43.0	1,638	0.62		0.52	M85049
<i>Bsg25D</i>	blastoderm-specific transcript	2-[16]	741	0.68	0.43	0.57	X04896
<i>bw</i>	brown	2-104.5	675	0.81		0.68	M20630
<i>cad</i>	caudal: homeodomain TF	2-[54]	472	0.74	0.35	0.58	M21070
<i>Cal</i>	calmodulin	2-[64]	152	0.67	0.38	0.63	X05951
<i>Cam</i>	CAM-kinase type II $\alpha$	4-[3]	490	0.32		0.28	M74583
<i>Cat</i>	catalase	3-[45]	506	0.76		0.68	X52286
<i>cdc2</i>	protein kinase	2-[40]	297	0.54		0.45	X57485
<i>cdc2c</i>	cdc2c protein kinase	3-[68]	314	0.60		0.53	X57486
<i>CecA1</i>	cecropin A1	3-[101]	63	0.56	0.34	0.52	X16972
<i>CecA2</i>	cecropin A2	3-[101]	63	0.51	0.31	0.52	X16972
<i>CecB</i>	cecropin B	3-[101]	63	0.61	0.26	0.56	X16972
<i>CecC</i>	cecropin C	3-[101]	63	0.64	0.26	0.59	Z11167
<i>Cfla</i>	chorion transcription factor 1 $\alpha$	3-[22]	549	0.65		0.54	X58435
<i>Cf2</i>	chorion transcription factor 2	2-[15]	235*	0.72		0.62	X53380
<i>Cg25C</i>	collagen $\alpha$ -1 type IV	2-[15]	1,775	0.52		0.47	M23704
<i>Cha</i>	choline acetyltransferase	3-64.6	728*	0.66		0.55	M13219
<i>chi</i>	chickadee: profilin	2-[18]	126	0.74		0.66	M84528
<i>chp</i>	chaoptin: cell surface glycoprotein	3-[102]	1,134	0.72		0.64	M19017
<i>ci</i>	cubitus interruptus: Zn finger P	4-0.0	1,377	0.30		0.23	X54360
<i>CkIIa</i>	casein kinase II $\alpha$ subunit	3-[47]	336	0.37		0.34	M16534
<i>CkIIb</i>	casein kinase II $\beta$ subunit	1-[36]	215	0.55		0.51	M16535
<i>cnc</i>	cap-n-collar: AHLH protein	3-81.2	533	0.64		0.52	M37495
<i>Cp15</i>	chorion protein S15	3-[26]	115	0.65	0.54	0.64	X02497
<i>Cp16</i>	chorion protein S16	3-[26]	138	0.69	0.40	0.66	X16715
<i>Cp18</i>	chorion protein S18	3-[26]	172	0.70	0.29	0.67	X02497
<i>Cp19</i>	chorion protein S19	3-[26]	173	0.74	0.45	0.71	X02497
<i>Cp36</i>	chorion protein S36	1-[23]	286	0.73	0.47	0.67	X05245
<i>Cp38</i>	chorion protein S38	1-[23]	306	0.64	0.36	0.60	X05245
<i>crb</i>	crumbs: transmembrane protein	3-82	2,139	0.63		0.55	M33753
<i>Csp</i>	cysteine-string protein 29	3-[47]	223	0.65		0.54	M63008
<i>ct</i>	cut: homeodomain protein TF	1-20.0	2,175	0.61		0.49	X07985
<i>cta</i>	concertina: G-protein- $\alpha$ 1-like	2-[54.8]	457	0.36		0.31	M63651
<i>CycA</i>	cyclin A	3-[36]	491	0.66		0.56	M24841
<i>CycB</i>	cyclin B	2-[101]	530	0.71		0.61	M33192
<i>CycC</i>	cyclin C	3-[55]	267	0.73		0.64	X62948
<i>Cyp1</i>	cyclophilin-1	---	165	0.82		0.77	M62398
<i>Cyt-c1</i>	cytochrome c DC3	2-[52]	105	0.65		0.57	X01761
<i>Cyt-c2</i>	cytochrome c DC4	2-[52]	108	0.81		0.77	X01760

APPENDIX 37.A. *Continued.*

<i>Gene</i>	<i>Function/Product</i>	<i>Map</i>	<i>AA</i>	<i>GC<sub>S</sub></i>	<i>GC<sub>I</sub></i>	<i>F<sub>op</sub></i>	<i>Acc.#</i>
<i>D1</i>	chromosomal protein D1	3-[49]	355	0.61		0.54	J04725
<i>da</i>	daughterless: AHLH protein	2-41.3	710	0.70		0.57	J03148
<i>Dbp73D</i>	D-E-A-D box protein 73D	3-[44]	572	0.51	0.31	0.44	M74824
<i>Ddc</i>	dopa decarboxylase	2-53.9	508	0.71	0.37	0.62	X04661
<i>dec1</i>	defective chorion-1	1-20.8	1,123	0.54		0.45	M35887
<i>Dfd</i>	deformed: homeodomain TF	3-47.5	590	0.64		0.50	X05136
<i>Dhod</i>	dihydroorotate dehydrogenase	3-48.0	51	0.50		0.46	X17297
<i>dim</i>	didymous: homeodomain protein	2-[46]	475	0.70		0.56	M65016
<i>disco</i>	disconnected: Zn finger protein	1-53.1	568	0.68		0.56	X56232
<i>Dl</i>	delta: EGF-transmembrane	3-66.2	833	0.67		0.57	Y00222
<i>dl</i>	dorsal: embryonic polarity	2-52.9	678	0.66		0.56	M23702
<i>Dlar</i>	protein Tyr phosphatase	2-[52]	2,029	0.65		0.56	M27700
<i>dlg1</i>	discs-large: guanate-cyclase-like	1-34.8	960	0.64		0.53	M73529
<i>Dms11</i>	RNA pol II elongation factor	—	313	0.77		0.70	X53670
<i>dnc</i>	dunce: cAMP phosphodiesterase	1-3.9	584	0.58		0.48	X55167
<i>Dox-A2</i>	diphenol oxidase A2	2-53.9	494	0.70	0.41	0.62	M63010
<i>dpp</i>	decapentaplegic	2-4.0	588	0.74		0.60	M30116
<i>Dpt</i>	dipteracin	2-[87]	106	0.57		0.47	M55432
<i>Dromsopa</i>	CAX (opa) repeat	3-[47]	69	0.78		0.75	X56491
<i>Dsk</i>	sulfated tyrosine-kinin	3-[47.1]	128	0.46		0.36	J03957
<i>ea</i>	easter: serine protease	3-57	392	0.76		0.64	J03154
<i>eag</i>	ether-a-gogo: K <sup>+</sup> channel protein	1-50.0	1,174	0.65		0.53	M61157
<i>EcR</i>	ecdysone receptor	2-[55.2]	878	0.68		0.56	M74078
<i>Edg78E</i>	pupal cuticle protein	3-[47]	122	0.77	0.46	0.73	M71247
<i>Edg84A</i>	pupal cuticle protein	3-[47.5]	188	0.55	0.38	0.49	M71249
<i>Edg91</i>	pupal cuticle protein	3-[62]	159	0.48	0.51	0.42	M71250
<i>Ef1a100E</i>	elongation factor 1- $\alpha$ F1	3-[102]	463	0.76	0.44	0.76	X06869
<i>Ef1a48D</i>	elongation factor 1- $\alpha$ F2	2-[64]	462	0.79	0.44	0.71	X06870
<i>Ef2b</i>	elongation factor 2	2-[54.6]	844	0.66		0.64	X15805
<i>Egon</i>	embryonic-gonad: Zn finger protein	3-[47]	373	0.68		0.54	X16631
<i>Eip71CD</i>	ecdysone-induced protein	3-[42]	255	0.64	0.36	0.55	X04024
<i>Eip74EF</i>	ecdysone-induced protein	3-[45]	883	0.72		0.56	X15087
<i>Eip75B</i>	ecdysone-induced protein	3-[45]	1,443	0.73		0.59	X15586
<i>elav</i>	embryonic lethal, abnormal vision	1-[0.0]	483	0.68		0.56	M21152
<i>emc</i>	extramacrochaetae protein	3-0.0	199	0.75		0.63	M31902
<i>en</i>	engrailed: homeodomain EF	2-62.0	60*	0.88	0.43	0.71	X01765
<i>Eno</i>	enolase phosphoglycerate hydrolase	2-[3]	433	0.87		0.86	X17034
<i>esg</i>	escargot: Zn finger protein	2-[51]	470	0.66		0.56	M83207
<i>E(spl)</i>	enhancer of split	3-89.1	186	0.70		0.66	X16553
<i>Est6</i>	esterase 6	3-35.9	544	0.50	0.25	0.42	J04167
<i>EstP</i>	esterase P	3-35.9	544	0.42	0.32	0.35	M33780
<i>Ets2</i>	ets-oncogene analog	2-[100]	159*	0.60	0.22	0.51	M20408
<i>eve</i>	even-skipped: homeodomain TF	2-[59]	376	0.78	0.41	0.68	M14767
<i>Fas1</i>	fasciclin I	3-[59]	652	0.69	0.41	0.61	M32311
<i>Fas2</i>	fasciclin II	1-[6]	811	0.62		0.55	M77165
<i>Fas3</i>	fasciclin III	2-[53]	508	0.70		0.61	M27813
<i>Fcp3C</i>	vitelline membrane protein 3C-1	1-[1.5]	210	0.58	0.45	0.42	M18281

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APPENDIX 37.A. *Continued.*

<i>Gene</i>	<i>Function/Product</i>	<i>Map</i>	<i>AA</i>	<i>GC<sub>S</sub></i>	<i>GC<sub>I</sub></i>	<i>F<sub>op</sub></i>	<i>Acc.#</i>
<i>fkh</i>	fork head: DNA-binding protein	3-95	510	0.78		0.57	J03177
<i>Fmrf</i>	FMRFamide polyprotein	2-[59]	342	0.74		0.62	J03232
<i>Fps85D</i>	fps-oncogene analog: P Tyr kinase	3-[49]	803	0.69		0.61	X52844
<i>fs(1)h</i>	FS: bromodomain membrane protein	1-21	2,038	0.64		0.52	M23221
<i>fs(1)K10</i>	FS: DNA-binding protein	1-0.5	463	0.64	0.40	0.53	X12836
<i>fs(1)Ya</i>	FS: nuclear envelope protein	1-[1.5]	708	0.75		0.62	M38442
<i>ft</i>	fat: cadherin-like protein	2-12.0	5,147	0.58		0.49	M80537
<i>ftz</i>	fushi tarazu: homeodomain TF	3-47.5	413	0.77	0.29	0.67	X00854
<i>ftz-f1</i>	ftz transcription factor 1	3-[45]	1,043	0.64		0.52	M63711
<i>Fur1</i>	furin-1: serine protease	—	899	0.66		0.54	X59384
<i>fz</i>	frizzled: transmembrane protein	3-41.7	581	0.66		0.53	X54646
<i>Gapdh1</i>	glyceraldehyde-3-phosphate DH 1	2-[57]	332	0.83		0.80	M11254
<i>Gapdh2</i>	glyceraldehyde-3-phosphate DH 2	1-[51]	332	0.75		0.72	M11255
<i>Gb13F</i>	G protein b subunit	1-[51]	340	0.66		0.58	M22567
<i>Gld</i>	glucose dehydrogenase	3-48	612	0.67	0.40	0.58	M29298
<i>Glu-R11</i>	glutamate receptor II	2-[17]	906	0.68		0.59	M73271
<i>G-<math>\alpha</math>47A</i>	G-protein $\alpha$ subunit	2-[60]	354	0.64		0.57	M86660
<i>Gpdh</i>	glycerol-3-phosphate dehydrogenase	2-17.8	362*	0.75	0.39	0.69	X61224
<i>Gprk1</i>	G-protein coupled receptor kinase 1	2-[55.1]	700	0.31		0.23	M80493
<i>Gprk2</i>	G-protein coupled receptor kinase 2	3-[102]	427	0.74		0.65	M80494
<i>grh</i>	grainy head: AHLH TF	2-86	1,063	0.69		0.57	X15657
<i>gro</i>	groucho: G-protein b-subunit-like	3-89.1	719	0.67		0.57	M20571
<i>G-sa60A</i>	G-protein Sa-60A	2-[106]	385	0.45	0.42	0.37	M33998
<i>Gst</i>	glutathione S-transferase 1-1	3-[51]	209	0.88		0.83	X14233
<i>gt</i>	giant: AHLH (Leu zipper)	1-1.0	448	0.70	0.49	0.59	X61148
<i>h</i>	hairy: AHLH	3-26.5	337	0.76		0.67	X15905
<i>H2.0</i>	homeodomain P 2.0 TF	2-[20]	410	0.68		0.57	Y00843
<i>hb</i>	hunchback: Zn finger protein	3-48.3	758	0.71	0.40	0.60	Y00274
<i>His1</i>	histone H1	2-[54.6]	255	0.48		0.41	X14215
<i>His2A</i>	histone H2A	2-[54.6]	124	0.54		0.54	X14215
<i>His2AvD</i>	histone H2A variant	3-[91]	134*	0.44		0.34	X07485
<i>His2B</i>	histone H2B	2-[54.6]	123	0.63		0.53	X14215
<i>His3</i>	histone H3	2-[54.6]	136	0.57		0.55	X14215
<i>His4</i>	histone H4	2-[54.6]	103	0.55		0.54	X14215
<i>HmG-CoAR</i>	3-OH-3-Methylglutaryl CoA reductase	3-[81]	916	0.62		0.55	M21329
<i>HmgD</i>	high mobility group protein D	2-[99]	112	0.74		0.65	M77023
<i>Hrb87Fa</i>	RNA-binding protein	3-[54]	386	0.64	0.33	0.60	X59691
<i>Hsc70-1</i>	heat-shock protein cognate 1	3-[41]	68*	0.67	0.42	0.58	J01085
<i>Hsc70-2</i>	heat shock protein cognate 2	3-[52]	68*	0.78	0.29	0.66	K01297
<i>Hsc70-4</i>	heat shock protein cognate 4	3-[57]	651	0.79		0.78	M36114
<i>Hsp22</i>	heat shock protein 22 kD	3-[28]	174	0.77		0.68	J01098
<i>Hsp23</i>	heat shock protein 23 kD	3-[28]	186	0.75		0.69	J01100
<i>Hsp26</i>	heat shock protein 26 kD	3-[28]	208*	0.75		0.69	J01099
<i>Hsp27</i>	heat shock protein 27 kD	3-[28]	213	0.72		0.64	J01101
<i>Hsp67Ba</i>	heat shock protein	3-[28]	238	0.71		0.59	M26267

APPENDIX 37.A. *Continued.*

<i>Gene</i>	<i>Function/Product</i>	<i>Map</i>	<i>AA</i>	<i>GC<sub>S</sub></i>	<i>GC<sub>I</sub></i>	<i>F<sub>op</sub></i>	<i>Acc.#</i>
<i>Hsp67Bb</i>	heat shock protein	3-[28]	111	0.55	0.30	0.45	X07311
<i>Hsp67Bc</i>	heat shock protein	3-[28]	169	0.53		0.44	X06542
<i>Hsp70A</i>	heat shock protein 70 kD	3-[51]	643	0.75		0.68	J01103
<i>Hsp70B</i>	heat shock protein 70 kD	3-[51]	641	0.73		0.66	J01104
<i>Hsp83</i>	heat shock protein 83 kD	3-[5]	375*	0.77	0.35	0.76	K01685
<i>Ide</i>	insulin-degrading enzyme	—	990	0.63		0.54	M58465
<i>ImpE2</i>	ecdysone inducible gene E2	3-[6]	466	0.57		0.52	M55099
<i>inaC</i>	protein kinase C	2-82	700	0.53		0.48	J04845
<i>Inr</i>	insulin-like receptor b subunit	3-[70]	300*	0.56		0.46	M13568
<i>Jra</i>	jun-related AHLH (Leu zipper)	2-[59]	289	0.72		0.64	M36181
<i>Kin</i>	kinesin heavy chain	2-[76]	975	0.74		0.66	M24441
<i>Klp54D</i>	kinesin-like protein (KLP1)	2-[80]	133*	0.65		0.50	M74427
<i>Klp61F</i>	kinesin-like protein (KLP2)	3-[0]	130*	0.55		0.49	M74428
<i>Klp64D</i>	kinesin-like protein (KLP4)	3-[19]	129*	0.55		0.48	M74430
<i>Klp67A</i>	kinesin-like protein (KLP3)	3-[27]	118*	0.69		0.56	M74429
<i>Klp68D</i>	kinesin-like protein (KLP5)	3-[36]	123*	0.63		0.50	M74431
<i>Klp98A</i>	kinesin-like protein (KLP6)	3-[98]	95*	0.66		0.53	M74432
<i>kni</i>	knirps: steroid receptor P family	3-[46]	429	0.75	0.42	0.62	X13331
<i>knrl</i>	knirps-related protein	3-[46]	647	0.58		0.49	X14153
<i>Kr</i>	Krüppel: Zn finger protein	2-107.6	467	0.54	0.33	0.44	X03414
<i>Kr-h</i>	Kr homolog: Zn finger protein	2-[20]	79*	0.83		0.74	M14940
<i>l(1)sc</i>	lethal at scute: T3 AHLH protein	1-0.0	257	0.59		0.51	X12549
<i>l(2)37Cc</i>	mitochondrial protein	2-53.9	203	0.73	0.42	0.61	X04227
<i>l(2)gl</i>	lethal giant larvae: transmembrane P	2-0.0	1,160	0.36		0.28	X05426
<i>lab</i>	labial: homeodomain TF	3-[47.5]	495*	0.71		0.57	X13103
<i>Lam</i>	nuclear lamin	2-[17]	621	0.79		0.73	X07278
<i>LanA</i>	laminin A chain	3-[21]	1,951*	0.60		0.52	M75882
<i>LanB1</i>	laminin B1 chain	2-[24]	1,787	0.61		0.53	M19525
<i>LanB2</i>	laminin B2 chain	3-[28]	1,639	0.68		0.61	M25063
<i>Lcp1</i>	cuticle protein I	2-[58]	130	0.70	0.47	0.69	J01080
<i>Lcp2</i>	cuticle protein II	2-[58]	126	0.66	0.43	0.65	J01081
<i>Lcp3</i>	cuticle protein III	2-[58]	112	0.77	0.45	0.73	J01081
<i>Lcp4</i>	cuticle protein IV	2-[58]	111	0.74	0.40	0.72	J01081
<i>lds</i>	lodestar: DEAH-family NTP-binding	3-47.8	974	0.59		0.49	X62629
<i>Lsp1-a</i>	α larval serum protein	1-39.5	70*	0.83	0.48	0.76	X03872
<i>Lsp1-b</i>	β larval serum protein	2-1.9	100*	0.91	0.47	0.88	X03873
<i>Lsp1-g</i>	gamma larval serum protein	3-[0]	105*	0.69	0.42	0.64	X03874
<i>LvpD</i>	larval visceral protein	2-[58]	508	0.62		0.55	V00204
<i>LvpH</i>	larval visceral protein	2-[58]	521	0.70	0.31	0.65	V00204
<i>LvpL</i>	larval visceral protein	2-[58]	505	0.71	0.45	0.65	V00204
<i>LysD</i>	lysozyme	3-[0]	140	0.76		0.72	X58382
<i>LysP</i>	lysozyme	3-[0]	141	0.73		0.65	X58382
<i>M(2)21C</i>	ribosomal protein 21C	2-0.0	112	0.82		0.81	Y00504
<i>M(3)67C</i>	ribosomal protein S17	3-28.9	131	0.84	0.37	0.84	M22142
<i>M(3)99D</i>	ribosomal protein rp49	3-[101]	133	0.78	0.47	0.72	X00848
<i>mam</i>	mastermind: neurogenic protein	2-70.3	1,596	0.67		0.58	X54251
<i>Map205</i>	microtubule-associated 205 kD	3-[105]	1,163	0.45		0.37	X54061

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APPENDIX 37.A. *Continued.*

<i>Gene</i>	<i>Function/Product</i>	<i>Map</i>	<i>AA</i>	<i>GC<sub>s</sub></i>	<i>GC<sub>t</sub></i>	<i>F<sub>op</sub></i>	<i>Acc.#</i>
<i>Mdr49</i>	P-glycoprotein (drug resistance)	2-[67]	1,302	0.67		0.60	M59076
<i>Mdr65</i>	P-glycoprotein (drug resistance)	3-[21]	1,302	0.51		0.43	M59077
<i>me31B</i>	maternal expression: DEAD-helicase	2-[37]	459	0.46		0.41	M59926
<i>mex1</i>	midgut expression 1	3-[42]	83	0.79	0.33	0.70	M63626
<i>Mhc</i>	myosin heavy chain	2-52.2	1,962	0.77	0.40	0.76	M61229
<i>Mlc1</i>	myosin light chain 1	3-[98]	155	0.75		0.70	K01567
<i>Mlc2</i>	myosin light chain 2	3-[101]	222	0.73	0.45	0.70	M11947
<i>mle</i>	male-less: DEAH-family helicase	2-55.2	1,293	0.53		0.46	M74121
<i>mod</i>	modulo: DNA-binding protein	3-[102]	544	0.49		0.40	X15702
<i>Mov34</i>	Mov34	2-[106]	338	0.78		0.70	M64643
<i>Mp20</i>	muscle-specific protein 20	2-[68]	184	0.82	0.34	0.83	Y00795
<i>msh1</i>	muscle homeodomain 1	3-[100]	61*	0.43		0.31	M38582
<i>Mst26Aa</i>	male accessory gland	2-[20]	264	0.41	0.32	0.33	Y00219
<i>Mst26Ab</i>	male accessory gland	2-[20]	90	0.51	0.36	0.43	Y00219
<i>Mst87F</i>	sperm protein	3-[45]	56	0.47	0.29	0.45	Y00831
<i>Mst95E</i>	male-specific protein msp316	3-[81]	52	0.39	0.31	0.29	M32022
<i>mys</i>	myospheroid: integrin b-subunit	1-[21]	846	0.70		0.61	J03251
<i>N</i>	notch: transmembrane protein	1-3.0	2,703	0.63	0.42	0.52	M16152
<i>nau</i>	nautilus: AHLH protein	3-[81]	332	0.62		0.50	X56161
<i>ncd</i>	non-claret disjunctional	3-100.7	700	0.72	0.39	0.63	X52814
<i>ninaA</i>	ninaA: transmembrane protein	2-1.4	237	0.80	0.39	0.68	M22851
<i>ninaC</i>	ninaC: protein kinase	2-[22]	1,501	0.62	0.32	0.54	J03131
<i>ninaE</i>	opsin-R1/R6	3-66.4	373	0.79	0.33	0.71	K02315
<i>NK1</i>	NK-1 homeodomain TF	3-[72]	659	0.70		0.54	X55393
<i>NK2</i>	NK-2 homeodomain TF	1-[0.0]	158*	0.66		0.55	M27290
<i>NK3</i>	NK-3 homeodomain TF	3-[72]	194*	0.77	0.38	0.64	M27291
<i>nod</i>	kinesin-like protein	1-36	666	0.64		0.52	M36195
<i>nonA</i>	RNA-binding protein	1-52.3	700	0.55		0.48	X55902
<i>norpA</i>	phospholipase C-b-type	1-6.5	1,095	0.67		0.59	J03138
<i>nos</i>	nanos: posterior determinant	3-66.2	401	0.68	0.37	0.55	M72421
<i>Nrq</i>	neuroglian Ig-like	1-23.6	1,239	0.65		0.57	M28231
<i>Nrt</i>	neurotactin: Ser protease-like TMP	3-[44]	846	0.64		0.55	X53837
<i>numb</i>	numb	2-[35]	556	0.66		0.55	M27815
<i>oc</i>	ocelliless: homeodomain TF	1-23.1	671*	0.65		0.47	X58983
<i>Ocr</i>	octopamine receptor	3-[100]	601	0.80		0.69	M60789
<i>ogre</i>	optic ganglion reduced	1-18.8	362	0.79		0.71	X61180
<i>omb</i>	optomotor-blind	1-7.5	974	0.63		0.48	M81796
<i>osk</i>	oskar: maternal effect	3-48.4	606	0.63	0.27	0.51	M65178
<i>Ote</i>	otefin: nuclear envelope protein	2-[86]	406	0.62		0.51	X17495
<i>otu</i>	ovarian tumors	1-22.7	811	0.55		0.46	X13693
<i>pAbp</i>	poly(A)-binding protein	2-[80]	574	0.70		0.65	M38019
<i>Pah</i>	phenylalanine-4-hydroxylase	—	453	0.61		0.52	M32802
<i>para</i>	paralytic: Na-channel $\alpha$ subunit	1-52.1	1,820*	0.56		0.49	M32078
<i>Pcna</i>	proliferating cell nuclear antigen	2-[88]	260	0.79	0.30	0.72	M33950
<i>Pcp</i>	pupal cuticle protein	2-[22]	184	0.70	0.51	0.60	J02527
<i>pcx</i>	pecanex transmembrane protein	1-0.9	2,483	0.56	0.50	0.45	M74329
<i>Pep</i>	protein on ecdysone puffs: Zn finger	3-[45]	716	0.67		0.64	X56689

APPENDIX 37.A. *Continued.*

<i>Gene</i>	<i>Function/Product</i>	<i>Map</i>	<i>AA</i>	<i>GC<sub>S</sub></i>	<i>GC<sub>I</sub></i>	<i>F<sub>op</sub></i>	<i>Acc.#</i>
<i>Pepck</i>	phosphoenolpyruvate carboxykinase	—	647	0.74		0.69	Y00402
<i>per</i>	period: biological clock protein	1-1.2	1,218	0.79	0.48	0.62	M30114
<i>Pgd</i>	6-phosphogluconate dehydrogenase	1-0.5	481	0.80	0.47	0.73	M80598
<i>phl</i>	pole-hole: raf-oncogene analog	1-[1]	666	0.64	0.31	0.51	X07181
<i>Pig1</i>	pre-intermoult gene 1	1-[3]	187	0.47		0.38	X15760
<i>Pka-C1</i>	cAMP-dependent protein kinase A	2-[34]	353	0.83		0.74	M18655
<i>Pka-C2</i>	cAMP-dependent protein kinase-B	3-[102]	354	0.72	0.46	0.65	X16960
<i>Pka-C3</i>	cAMP-dependent protein kinase-related	3-[43]	502	0.61		0.49	X16961
<i>Pkc53E</i>	protein kinase C 53E	2-[78]	639	0.61	0.26	0.55	X05283
<i>Pkc98E</i>	protein kinase C 98E	3-[99]	634	0.78		0.69	J04848
<i>Pkg24A</i>	cGMP-dependent protein kinase 24A	2-[9]	894	0.69		0.59	M30147
<i>Plc21C</i>	phospholipase C	2-[0.1]	1,312	0.63		0.52	M60453
<i>polo</i>	protein Ser/Thr kinase	3-46	576	0.74		0.65	X63361
<i>Ppl-87B</i>	protein-Ser/Thr phosphatase 1 $\alpha$	3-[51]	302	0.77		0.71	X15583
<i>PpY-55A</i>	protein Ser/Thr phosphatase Y	2-[83]	314	0.53		0.46	Y07510
<i>prd</i>	paired: homeodomain TF	2-45	613	0.66	0.47	0.56	M14548
<i>Prm</i>	paramyosin	3-[26]	477	0.88		0.84	X62591
<i>pros</i>	prospero: homeodomain	3-[51]	1,407	0.71		0.60	M81389
<i>Pros28</i>	proteasome 28 kD subunit	—	249	0.72		0.70	M57712
<i>Pros35</i>	proteasome 35 kD subunit	3-[59]	279	0.63		0.57	X15497
<i>Psc</i>	posterior sex combs: Zn finger	2-67	1,603	0.56		0.45	X59275
<i>Ptp</i>	protein Tyr phosphatase	—	1,462	0.49		0.42	M27699
<i>Ptp10D</i>	protein Tyr phosphatase 10D	1-[36]	1,558	0.68		0.57	M80538
<i>Ptp99A</i>	protein Tyr phosphatase 99A	3-[100]	1,301	0.68		0.59	M81795
<i>pum</i>	pumilio	3-48.5	1,533	0.64		0.53	X62589
<i>R</i>	roughened: ras analog	3-1.4	184	0.84		0.75	M80535
<i>r</i>	rudimentary: dihydroorotase	1-54.5	2,236	0.64		0.54	X04813
<i>Rab3</i>	ras-related GTP-binding protein	2-[60]	220	0.74		0.64	M64621
<i>Ras64B</i>	GTPase ras-analog 2	3-[15]	187	0.79		0.71	K01962
<i>Ras85D</i>	GTPase ras-analog 1	3-[49]	189	0.72		0.66	K01960
<i>Rdl</i>	GABA-A receptor	3-[27]	606	0.54		0.46	M69057
<i>ref(2)P</i>	male fertility (Zn finger)	2-54.0	599	0.58	0.34	0.50	X16993
<i>Rh2</i>	rhodopsin-2	3-[65]	381	0.65	0.33	0.54	M12896
<i>Rh3</i>	rhodopsin-3	3-[67]	383	0.72		0.62	M17718
<i>Rh4</i>	rhodopsin-4	3-[44]	378	0.74		0.61	M17730
<i>Rm62</i>	DEAD-family helicase	3-[47.4]	575	0.72		0.68	X52846
<i>RpA1</i>	ribosomal protein A1	2-[78]	113	0.80		0.78	X05016
<i>RpI135</i>	RNA polymerase I 135 kD subunit	2-[0.1]	1,129	0.53	0.32	0.44	X17298
<i>RpII140</i>	RNA polymerase II 140 kD subunit	3-54	1,123	0.58	0.26	0.55	X05709
<i>RpII215</i>	RNA polymerase II 215 kD subunit	1-35.7	1,896	0.66	0.34	0.58	M27431
<i>RpIII128</i>	RNA polymerase III 128 kD subunit	2-	1,135	0.64		0.56	X58826

(continued)

APPENDIX 37.A. *Continued.*

<i>Gene</i>	<i>Function/Product</i>	<i>Map</i>	<i>AA</i>	<i>GC<sub>s</sub></i>	<i>GC<sub>t</sub></i>	<i>F<sub>op</sub></i>	<i>Acc.#</i>
<i>RpL1</i>	ribosomal protein L1	3-[98]	407	0.79		0.79	X13382
<i>RpS14A</i>	ribosomal protein S14 A	1-[21]	151	0.70	0.42	0.72	M21045
<i>RpS14B</i>	ribosomal protein S14 B	1-[21]	151	0.69	0.39	0.71	M21045
<i>Rrp1</i>	recombination repair protein	2-[6]	679	0.56		0.48	M62472
<i>run</i>	runt: ATP-binding protein	1-65	509	0.80		0.65	X56432
<i>rut</i>	rutabaga: adenylyl cyclase	1-46	2,248	0.70		0.59	M81887
<i>ry</i>	rosy: xanthine dehydrogenase	3-[52]	1,335	0.64	0.37	0.55	Y00308
<i>sala</i>	spalt accessory	2-44	142	0.28	0.25	0.26	X57474
<i>sas</i>	stranded at second	3-[47.5]	1,348	0.64		0.52	M68866
<i>sc</i>	scute: AHLH protein	1-0.0	345	0.51		0.43	M17119
<i>sca</i>	scabrous: fibrinogen-like	2-66.7	774	0.73		0.61	M60065
<i>Scr</i>	sex combs reduced: homeodomain TF	3-47.5	73*	0.36		0.22	X05228
<i>sd</i>	scalloped: DNA-binding protein	1-51.5	440	0.56		0.45	M83787
<i>Sd</i>	segregation distorter: Leu zipper	2-54	363	0.59		0.51	X60218
<i>Ser99Da</i>	serine protease 1	3-[101]	265	0.82		0.78	M24379
<i>Ser99Db</i>	serine protease 2	3-[101]	265	0.82		0.77	M24379
<i>Ser99Dc</i>	serine protease 3	3-[101]	61	0.54		0.51	M24380
<i>sev</i>	sevenless: protein Tyr kinase	1-33.4	2,554	0.66	0.37	0.54	J03158
<i>sgg</i>	shaggy: Ser/Thr kinase	1-1.3	514	0.57		0.49	X53332
<i>Sgs4</i>	salivary gland secretion	1-[3]	182*	0.47		0.37	X06565
<i>Sgs5</i>	salivary gland secretion	3-[60]	163	0.54	0.25	0.46	X04269
<i>Sh</i>	shaker K <sup>+</sup> -channel	1-57.6	643	0.48		0.38	X07132
<i>Shab</i>	shaker cognate b	3-[3]	924	0.63		0.54	M32659
<i>Shal</i>	shaker cognate l	3-[46]	490	0.79		0.66	M32660
<i>Shaw</i>	shaker cognate w	2-[10]	498	0.71		0.62	M32661
<i>shi</i>	shjbire: dynamin	1-51.5	836	0.56		0.50	X59448
<i>sim</i>	single-minded: AHLH protein	3-52.2	655*	0.73		0.61	M19020
<i>sina</i>	seven in absentia: nuclear protein	3-[44]	314	0.80		0.69	M38384
<i>sli</i>	slit: transmembrane protein	2-77	1,480	0.73		0.64	X53959
<i>slo</i>	slowpoke: Ca-activated K <sup>+</sup> -channel	3-86	1,184*	0.59		0.50	M69053
<i>sn</i>	singed	1-21.0	512	0.75	0.37	0.65	X17549
<i>sna</i>	snail: Zn finger protein	2-51	390	0.73		0.64	Y00288
<i>snk</i>	snake: serine protease	3-52.1	435	0.67		0.59	X04513
<i>snRNP27D</i>	sn-ribonucleoprotein 70 kD	2-[21]	448	0.76	0.47	0.65	M31162
<i>Sod</i>	Cu-Zn superoxide dismutase	3-[34]	153	0.74		0.67	Y00367
<i>sol</i>	small optic lobes: Zn finger	1-[65]	1,597	0.73		0.60	M64084
<i>Sos</i>	son of sevenless: G-exchange	2-[48]	1,595	0.68		0.56	M83931
<i>Spec-a</i>	$\alpha$ -spectrin	3-[1.5]	2,415	0.76		0.72	M26400
<i>SR55</i>	Ser-Arg RNA-binding protein	3-[53]	350	0.66		0.65	X58720
<i>Src29A</i>	src-oncogene analog	2-[24]	590	0.63		0.55	M16599
<i>Src64B</i>	src-oncogene analog	3-[15]	552	0.74		0.65	M11917
<i>Sry-a</i>	serendipity $\alpha$	3-[101]	530	0.70		0.60	X03121
<i>Sry-b</i>	serendipity $\beta$ : Zn finger protein	3-[101]	351	0.88		0.78	X03121
<i>Sry-d</i>	serendipity $\delta$ : Zn finger protein	3-[101]	430	0.88	0.53	0.80	X03121
<i>stau</i>	staufen	2-83.5	1,026	0.61		0.47	M69111
<i>Ste</i>	stellate: casein kinase II-b-like	1-45.7	172	0.74	0.35	0.61	X15899
<i>stg</i>	string: Tyr phosphatase	3-[100]	479	0.78		0.70	M24909
<i>su(f)</i>	suppressor of forked	1-65.9	733	0.59	0.34	0.52	X62679

APPENDIX 37.A. *Continued.*

<i>Gene</i>	<i>Function/Product</i>	<i>Map</i>	<i>AA</i>	<i>GC<sub>S</sub></i>	<i>GC<sub>I</sub></i>	<i>F<sub>op</sub></i>	<i>Acc.#</i>
<i>Su(H)</i>	suppressor of Hairless: DNA-binding	2-50.5	550	0.65		0.56	X58393
<i>su(Hw)</i>	suppressor of Hairy wing	3-54.8	944	0.60	0.33	0.54	Y00228
<i>su(s)</i>	suppressor of sable: RNA-binding	1-0.0	1,334	0.59	0.38	0.49	M57889
<i>Su(var)20</i>	suppressor of variegation: DNA-binding	2-31.1	206	0.64	0.37	0.56	M57574
<i>Su(z)2</i>	suppressor of zeste-2: Zn finger	2-[67]	1,364	0.58	0.39	0.44	X56798
<i>sup</i>	seven-up: steroid receptor	3-[51]	543	0.75		0.65	M28863
<i>Sx1</i>	Sex-lethal: RNA-binding protein	1-19.2	366	0.58		0.52	M59448
<i>Syt</i>	synaptotagmin-p65	2-[7]	474	0.72		0.64	M55048
<i>Takr86C</i>	tachykinin-like receptor	3-[50]	504	0.71		0.60	M77168
<i>Takr99D</i>	tachykinin-like receptor	3-[101]	519	0.80		0.64	X62711
<i>T-cpl</i>	T complex protein 1 analog	3-[76]	557	0.72		0.69	M21159
<i>term</i>	terminus: Zn finger protein	3-[45]	428	0.82		0.73	M19140
<i>TfIID</i>	transcription factor IID	2-[99]	353	0.67		0.56	M38082
<i>Tgfb-60A</i>	TGF-b-like	2-[106]	455	0.86		0.75	M84795
<i>tin</i>	tinman: homeodomain TF	3-[72]	150*	0.74	0.29	0.65	M27292
<i>tko</i>	technical knockout: mt RP S12	1-1.0	140	0.79		0.68	M19494
<i>Tl</i>	Toll: transmembrane protein	3-91	1,097	0.66		0.53	M19969
<i>tld</i>	tollid: bone morphogenetic P-1-like	3-85	1,057	0.61		0.53	M76976
<i>tll</i>	tailless: steroid receptor	3-102	452	0.73		0.64	M34639
<i>Tm1</i>	tropomyosin I	3-[55]	284	0.84	0.44	0.85	K02623
<i>Tm2</i>	tropomyosin II/troponin H	3-[55]	285	0.71		0.72	M15466
<i>top</i>	torpedo: protein Tyr kinase	2-[97]	174*	0.70		0.61	K03417
<i>Top2</i>	type II DNA topoisomerase	2-[54]	1,447	0.63	0.37	0.57	X61209
<i>tor</i>	torso: receptor Tyr kinase	2-57	923	0.60	0.33	0.51	X15150
<i>tra2</i>	transformer-2: RNA-binding protein	2-[71]	264	0.54		0.49	M23633
<i>trp</i>	serine protease	3-[100]	1,275	0.68	0.38	0.59	M34394
<i>trx</i>	trithorax: Zn finger protein	3-54.2	3,759	0.50		0.41	M31617
<i>Try</i>	trypsin-like Ser protease	2-[60]	256	0.73		0.67	X02989
<i>tsh</i>	teashirt: Zn finger protein	2-[54.8]	993	0.49		0.39	M57496
<i>ttk</i>	tramtrack: Zn finger protein	3-[102]	641	0.69		0.55	X17121
<i>tub</i>	tube: (dorso-ventral polarity)	3-[47.1]	462	0.59		0.44	M59501
<i>Tuba67C</i>	$\alpha$ -4 tubulin	3-[28]	462	0.72	0.30	0.64	M14646
<i>Tuba84B</i>	$\alpha$ -1 tubulin	3-[47.5]	450	0.79	0.39	0.79	M14643
<i>Tuba84D</i>	$\alpha$ -3 tubulin	3-[48]	450	0.79		0.79	M14645
<i>Tuba85E</i>	$\alpha$ -2 tubulin	3-[49]	449	0.74	0.32	0.69	M14644
<i>Tubb60D</i>	$\beta$ -3 tubulin	2-[107]	454	0.88	0.42	0.80	M22335
<i>Tubb85D</i>	$\beta$ -2 tubulin	3-48.5	446	0.72		0.66	M20420
<i>Tubb97EF</i>	$\beta$ -1 tubulin	3-[92]	447	0.81		0.79	M20419
<i>Tubg</i>	gamma-tubulin	2-[6]	475	0.66		0.58	M61765
<i>tud</i>	tudor protein	2-97	2,515	0.54		0.47	X62420
<i>tuf</i>	tufted: transmembrane protein	2-59	1,286	0.78	0.35	0.65	M28999
<i>twi</i>	twist: AHLH protein	2-[102]	490	0.81	0.25	0.71	X12506
<i>twn</i>	twain: homeodomain protein	2-[46]	601	0.69		0.59	M65015
<i>UbcD6</i>	ubiquitin conjugating enzyme	3-[47.1]	151	0.43		0.38	M63792
<i>Ubi-f</i>	ubiquitin-RP hybrid	1-[17]	128	0.80		0.80	X53059

(continued)

APPENDIX 37.A. *Continued.*

<i>Gene</i>	<i>Function/Product</i>	<i>Map</i>	<i>AA</i>	<i>GC<sub>s</sub></i>	<i>GC<sub>i</sub></i>	<i>F<sub>op</sub></i>	<i>Acc.#</i>
<i>Ubi-m</i>	ubiquitin-RP S27A hybrid	—	156	0.71		0.70	M22536
<i>Ubi-p</i>	poly-ubiquitin protein	3-[6]	231	0.68		0.69	M22428
<i>Ubx</i>	Ultrabithorax: homeodomain TF	3-58.8	246	0.70	0.54	0.59	M24608
<i>up</i>	upheld: troponin-T	1-41.0	396	0.76		0.74	X54504
<i>Uro</i>	urate oxidase	2-[24]	352	0.70	0.22	0.62	X51940
<i>usp</i>	ultraspiracle: chorion 1 TF	1-[0.5]	508	0.77		0.65	X53417
<i>uzip</i>	unzipped	2-107.6	500	0.53		0.44	X07450
<i>v</i>	vermillion: tryptophan oxidase	1-33.0	379	0.68	0.39	0.61	M34147
<i>vas</i>	vasa: DEAD-family helicase	2-51	661	0.47	0.32	0.41	X12946
<i>Vha</i>	vacuolar H <sup>+</sup> -ATPase 16 kD subunit	—	159	0.63		0.55	X55979
<i>Vm26Aa</i>	vitelline membrane protein 26Aa	2-[20]	168	0.74		0.72	M20936
<i>Vm26Ab</i>	vitelline membrane protein 26Ab	2-[20]	141	0.82		0.79	M18280
<i>Vm32Ec</i>	vitelline membrane protein 32Ec	2-[44]	116	0.59		0.49	M27647
<i>Vm34Ca</i>	vitelline membrane protein 34Ca	2-[47]	96*	0.72		0.65	X01802
<i>w</i>	white eye	1-1.5	687	0.71		0.58	X51749
<i>wg</i>	wingless: int1-oncogene analog	2-[22]	468	0.74		0.63	M17230
<i>y</i>	yellow body	1-0.0	541	0.46	0.33	0.35	X04427
<i>yema</i>	nuclein a DNA-binding protein	3-[99]	1,022	0.68	0.35	0.55	X63503
<i>Yp1</i>	yolk protein 1	1-30	442	0.80	0.38	0.76	X01524
<i>Yp2</i>	yolk protein 2	1-30	439	0.80	0.25	0.75	X01524
<i>Yp3</i>	yolk protein 3	1-44	420	0.80	0.37	0.75	M15898
<i>z</i>	zeste	1-1.0	575	0.68	0.38	0.58	Y00049
<i>Z600</i>	histone-like protein	3-[42]	90	0.68		0.63	X58286
<i>zfh1</i>	Zn-finger homeodomain protein 1	3-[102]	1,060	0.76		0.65	M63449
<i>zfh2</i>	Zn-finger homeodomain protein 2	4-[1]	3,005	0.43		0.35	M63450
<i>zip</i>	zipper: myosin heavy chain	2-[108]	1,972	0.63		0.56	M35012
—	65 kD protein phosphatase	—	591	0.67		0.62	M86442
—	retinal specific G- $\alpha$ protein	—	353	0.52	0.37	0.46	M58016
—	fushi tarazu repressor	—	641	0.69		0.56	M62856
—	Glu-tRNA aminoacyl synthetase	—	1,475	0.57		0.51	M74104
—	DNA polymerase	—	1,505	0.55	0.29	0.47	D90310
—	laminin receptor	—	253	0.88		0.83	M77133

Genes are presented in alphabetical order; gene names follow FlyBase (Ashburner 1992). Map is the genetically defined map location. AA is the length of the gene in codons, \* indicates a partial gene sequence. GC<sub>s</sub> is the G + C content at silent third positions of codons (i.e., excluding Trp, Met and stop codons); GC<sub>i</sub> is the G + C content in introns. F<sub>op</sub> is the frequency of optimal codons (see text for definition). Acc.# indicates the accession number allowing retrieval of the sequence from the GenBank/EMBL/DDBJ DNA sequence data library. Abbreviations: AHLH = amphipathic helix-loop-helix; DH = dehydrogenase; FS = female sterile; G = guanine; mt = mitochondrial; P = protein; RP = ribosomal protein; TF = transcription factor; TGF = transforming growth factor; TMP = transmembrane protein.

## Appendix 37.B. Codon Usage Bias in *D. melanogaster* Transposable Elements

<i>Family</i>	<i>Element</i>	<i>Gene</i>	<i>AA</i>	<i>GC<sub>S</sub></i>	<i>F<sub>op</sub></i>	<i>Acc.#</i>
LINE-like:	F	NA binding	122	0.50	0.43	M17214
		RT	858	0.45	0.38	
	I	NA binding	429	0.38	0.36	M14954
		RT	1,086	0.41	0.37	
	Jockey	NA binding	583	0.38	0.32	M22874
		RT	916	0.46	0.37	
	DOC	NA binding	565	0.42	0.37	X17551
		RT	888	0.41	0.35	
	R1Dm	orf1	471	0.63	0.50	X51968
		RT	1,021	0.56	0.45	
R2Dm	RT	1,057	0.46	0.36	X51967	
Ty-like	Copia 1,731		1,409	0.28	0.23	X02599
		gag	273	0.49	0.35	X07656
		pol	982	0.50	0.39	
Retrovirus-like:	17.6	gag	445	0.30	0.27	X01472
		pol	1,058	0.33	0.28	
		env	472	0.28	0.26	
	297	gag	424	0.31	0.26	X03431
		pol	1,059	0.26	0.23	
		env	471	0.29	0.26	
	Gypsy	gag	451	0.52	0.43	M12927
		pol	1,035	0.54	0.45	
		env	509	0.51	0.44	
	412	gag	444	0.36	0.32	X04132
pol		1,219	0.27	0.22		
Foldback:	FB4	orf	148	0.35	0.30	J01084
	FBw <sup>c</sup>	orf1	633	0.39	0.29	X15469
		orf2	403	0.37	0.29	
P-like:	P element		751	0.38	0.31	V01520
	HOBO		644	0.32	0.26	M69216

Abbreviations: NA = nucleic acid; RT = reverse transcriptase. See also the footnote to Appendix 37.A.

## APPENDIX

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### Early Stages of Embryonic Development

Many of the genes treated in Part I are expressed in early embryos. Four figures that summarize different aspects of the processes involved are presented in this appendix.

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FIG. A.1. "Schematic drawing of the embryonic stages leading up to gastrulation in *D. melanogaster*" from Foe and Alberts (1983). "This figure is modified from Zalokar & Erk (1976) to show the correct times of appearance of pole and somatic buds and to indicate the cessation of division of the yolk nuclei. The number beside each embryo, which denotes its developmental stage, corresponds to the total number of nuclear division cycles undergone by the almost synchronously dividing embryonic nuclei. A stage begins with the start of interphase and ends with the conclusion of mitosis. Stage 1 is the fertilized zygote during its first interphase and mitosis. The subsequent stages, each of which corresponds to one complete nuclear division cycle (interphase plus mitosis), are numbered consecutively. Embryos are shown in longitudinal section and with their anterior ends at the top. They are depicted without vitelline membranes to emphasize the changes in surface morphology of the plasma membrane that surrounds the syncytial embryo. Solid black circles represent nuclei, stippled regions denote yolk, and non-textured regions denote the yolk-free regions of cytoplasm. As shown, when development begins there is a thin layer of yolk-free cytoplasm at the egg periphery (the 'periplasm'), and a yolk-free region of cytoplasm surrounding each nucleus (the 'protoplasmic islands'). For stages 1–5 all nuclei are indicated, even though they would not all normally be in the same plane. For stages 6–14, only a fraction of the embryonic nuclei is shown."

"Stages 1–7: The nuclei multiply exponentially in the central region of the egg."

"Stage 8: The majority of the still dividing nuclei, with their enveloping protoplasmic islands, have started their migration outwards, leaving the future yolk nuclei behind. These yolk nuclei will divide in approximate synchrony with the remaining nuclei in cycles 8–10, and thereafter cease dividing and become polyploid."

"Stage 9: Early in their 9th interphase, a few migrating nuclei appear in the posterior periplasm, creating there the posterior cytoplasmic protuberances called pole buds. At the end of this stage, these nuclei (like all others in the syncytium) enter into mitosis, thus doubling the number of pole buds." (continued)

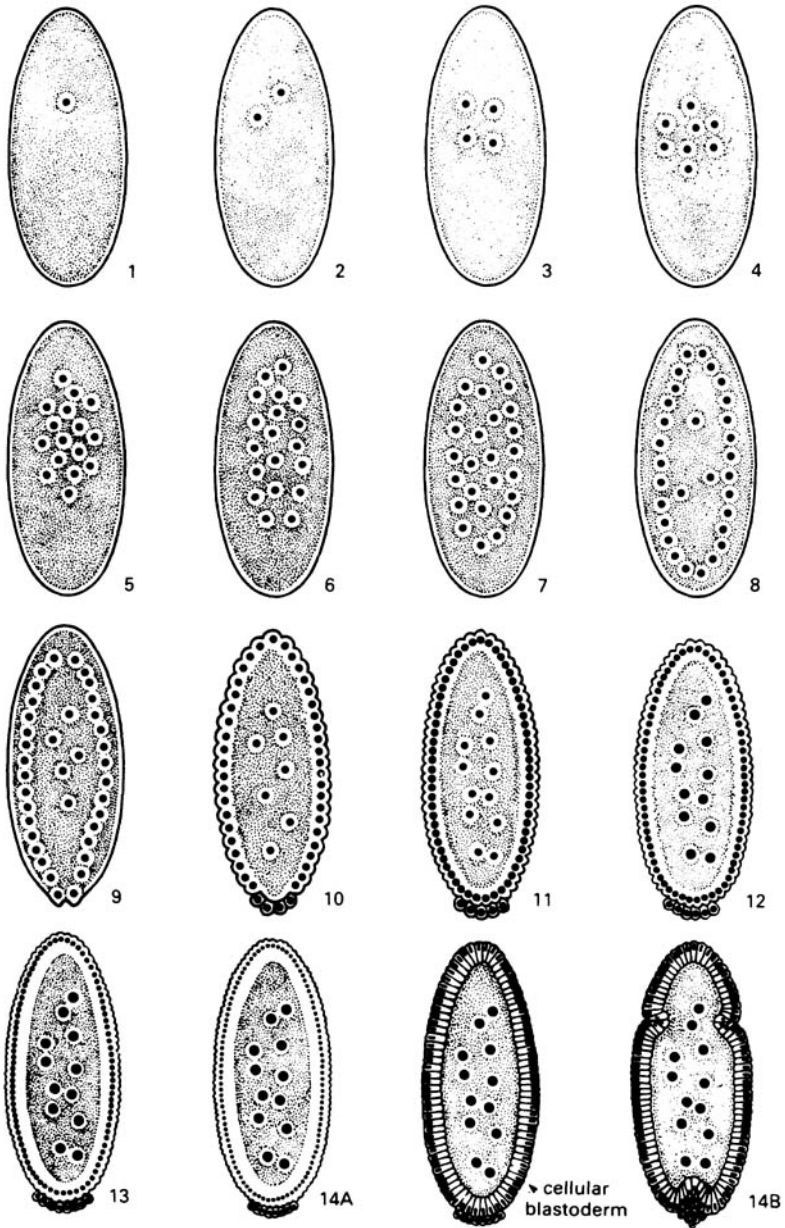


FIG. A.1 *continued*. “Stage 10: The remainder of the migrating nuclei appear in the periplasm at the beginning of their 10th interphase, organizing somatic buds over the entire embryonic surface. During mitosis of this cycle, the pole buds divide again and, nearly simultaneously, are pinched off from the syncytial embryonic mass to produce the pole cells; after this stage these cells, which are the potential germ cell progenitors, will continue to divide, but they lose mitotic synchrony with the embryonic syncytium.”

“Stages 10–13: The syncytial nuclei in their somatic buds at the embryonic periphery divide with near synchrony.

(*continued*)



FIG. A.1 *continued*. During cycle 13, the depth of the yolk-free periplasm increases dramatically at the expense of the central yolk region."

"Stage 14A: Plasma membrane formation occurs synchronously between all of the peripheral nuclei to generate separate cells. During this process, the nuclei elongate, matching the shape of the elongated blastodermal cells that are forming. Stage 14A is depicted at both early (no cell membranes evident) and late (cellularization just completed) times. The cells that form at this time are the progenitors of the somatic tissues."

"Stage 14B: Immediately following cellularization, gastrulation movements begin. The infolding of cells depicted about one-third of the distance down from the anterior pole is a section of the cephalic furrow (also called the anterior oblique cleft), and the invagination of the posterior pole is part of the posterior midgut furrow (all called the amnioproctodaeal invagination) into which the pole cells move. Not knowing when nuclear division occurs during stage 14, Zalokar & Erk (1976) designated the early gastrula as stage 15, rather than as stage 14B. The cells do not begin the mitosis of cycle 14 synchronously, but rather enter mitosis in a consistent region-specific sequence beginning 15 min after the start of gastrulation. Note also that a true 'cellular blastoderm' stage hardly exists in *Drosophila*, since gastrulation begins as soon as cells have formed."

"The average time required for stages (nuclear cycles) 1-9 is 8 min at 25°C. Stages 10, 11, 12, 13 and 14 occupy about 9, 10, 12, 21, and more than 65 min, respectively." From Foe and Albert (1983); reproduced by permission.

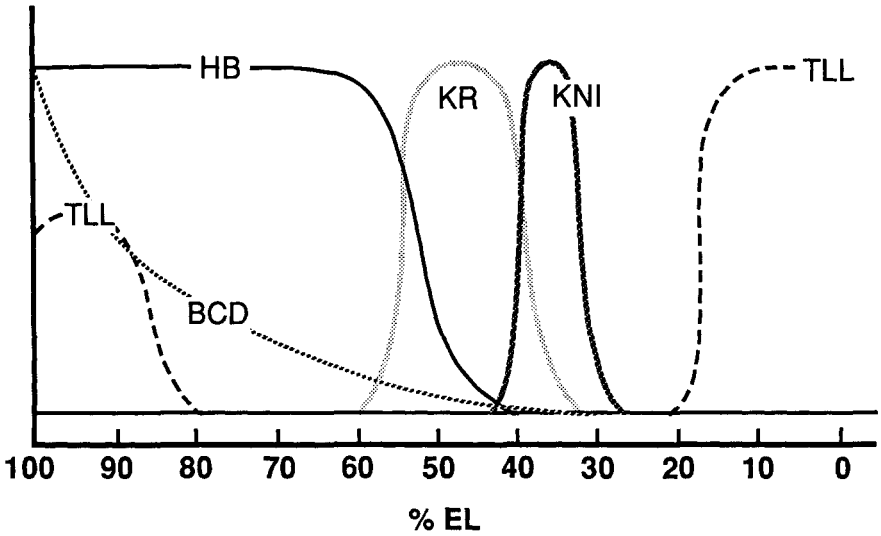


FIG. A.2. Main zones of expression of several gap genes along the antero-posterior axis of the egg (%EL) during blastoderm stage (modified from Hülskamp et al. 1990). 0% egg length = posterior pole to the right; the scale on the vertical axis is arbitrary and cannot be used to compare levels of gene products to each other. The horizontal line near the bottom of the graph represents the threshold of detection; it is meant to indicate that the presence, and effect, of some of these products may extend beyond the region of the embryo where they are detected. Localization of these products occurs at the mRNA level and it is due, at least in part, to the following interactions.

Maternal *bcd* RNA is anchored at the anterior pole by cytoskeletal elements. BCD stimulates transcription of *hb* thus limiting this RNA to the anterior half of the embryo. Low concentrations of BCD and HB stimulate transcription of *Kr* in the middle section of the embryo, while high concentrations repress it (thus defining the anterior border of the KR band). KR in turn represses *hb* thus defining this gene posterior border of expression, and it activates *kni* in a band immediately posterior to its own. Low to moderate concentrations of the HB and TLL repress *kni* thus defining the anterior and posterior border of the KNI band.

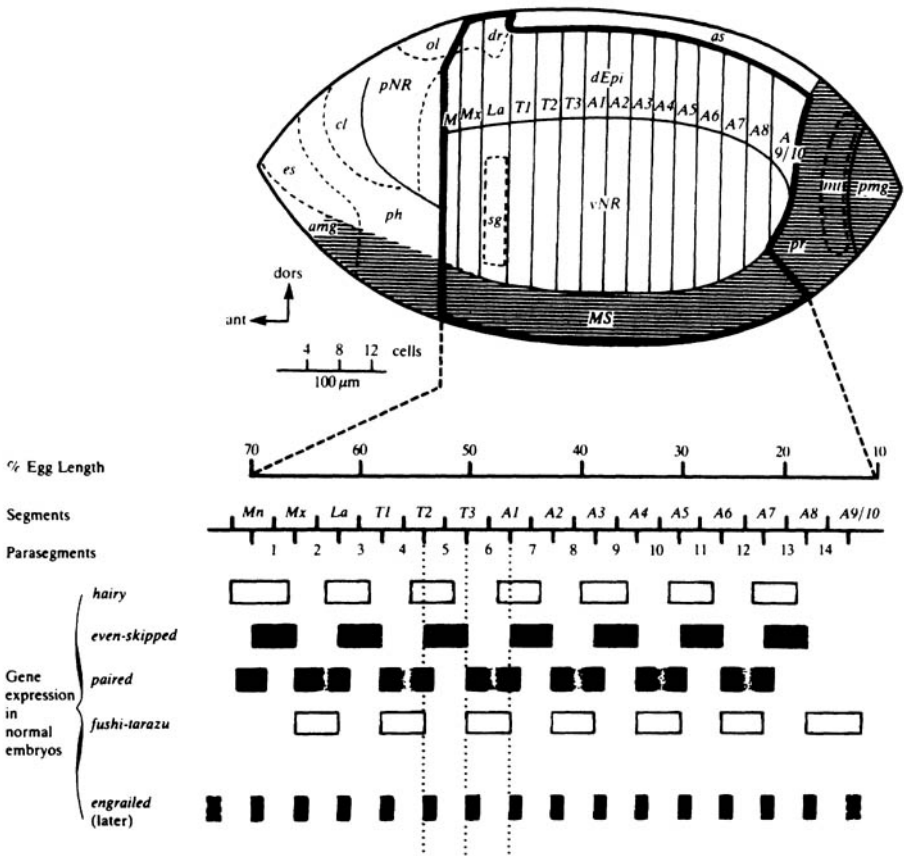


FIG. A.3. Top. "A fate map of the *Drosophila* blastoderm (from Campos-Ortega and Hartenstein, 1985)" as modified by Akam (1987). "The shape is a planimetric reconstruction of the blastoderm surface. All parts of the egg surface contribute to the embryo proper, except the narrow dorsal primordium for the amnioserosa (*as*). Hatched areas will invaginate at gastrulation. Cells that will generate metameric structures are enclosed by a thick line. Abbreviations: *amg*, anterior midgut; *ant*, anterior; *as*, amnioserosa; *cl*, clypeolabrum; *dEpi*, dorsal epidermis; *dors*, dorsal; *dr*, dorsal ridge; *es*, oesophagus; *mt*, Malpighian tubules; *MS*, mesoderm; *ol*, optic lobe; *ph*, pharynx; *pmg*, posterior midgut; *pNR*, procephalic neurogenic region; *pr*, proctodeum; *sg*, salivary gland; *vNR*, ventral neurogenic region; *M*" or *Mn*, "mandibular segment; *Mx*, maxillary segment; *La*, labial segment; *T1-T3*, thoracic segments; *A1-A10* abdominal segments."

Bottom. "Expression of segmentation genes in the *Drosophila* blastoderm: approximate registration of pair-rule, *engrailed* expression and metameric units." Each segment is divided, by the parasegment line, into an anterior (A) and a posterior (P) compartment. "The patterns of expression are shown for four of the pair-rule genes at about cleavage stage 14A/B. The later patterns of *engrailed* expression have been projected onto the same diagram, even though at this mid-blastoderm stage hybridization reveals only a single well-defined *engrailed* stripe (stripe 2)."

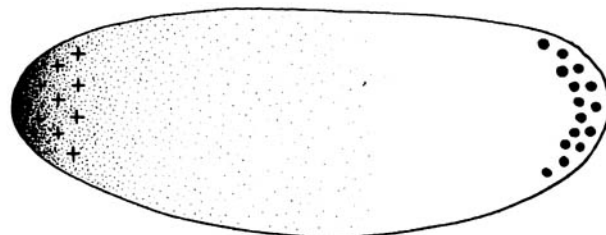
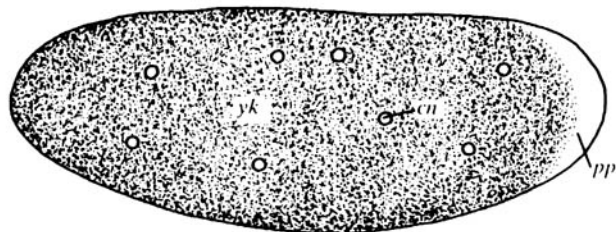
"The bands of *engrailed* expression define P compartments and so lie at the anterior margin of each parasegment. (continued)

FIGURE A.3 *continued*. Stripes of *even-skipped* and *fushi-tarazu* expression are each approximately four cells wide at mid-blastoderm, and appear to lie out of phase with each other. Double-labelling experiments in later embryos suggest that the anterior margins of both *ftz* and *eve* stripes coincide precisely with the *engrailed* stripes, and hence define parasegment boundaries (Lawrence et al. 1987). *hairy* stripes are about the same width, but are displaced slightly with respect to parasegments and overlap those of *ftz*. *paired* stripes are broader than a single metamer repeat, but the seven stripes split into fourteen before gastrulation." (This figure combines elements from Figs 1 and 4B from Akam (1987); reproduced by permission.) For a review and discussion see also Carroll (1990).

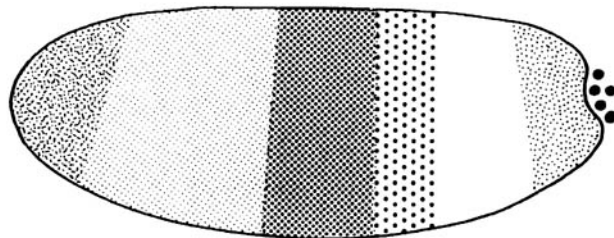
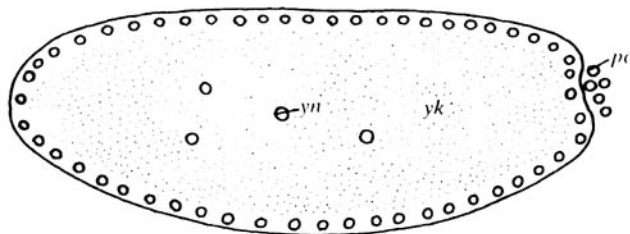
*Morphology*

*Gene activity*

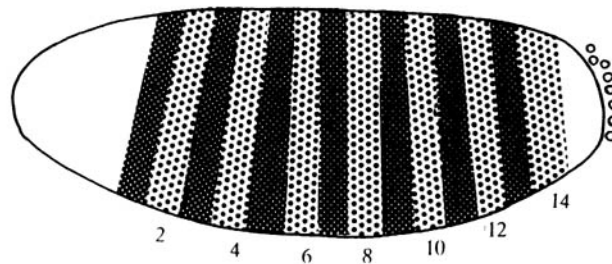
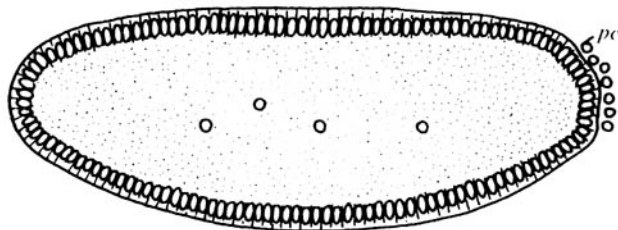
A. Nuclear migration (1-25 h, ~128 nuclei)



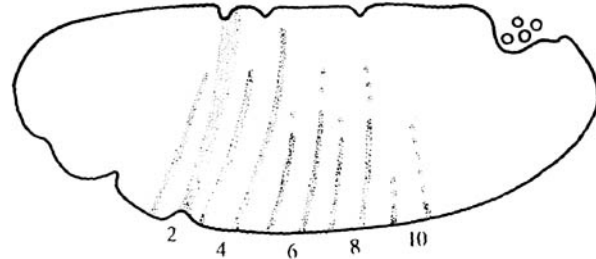
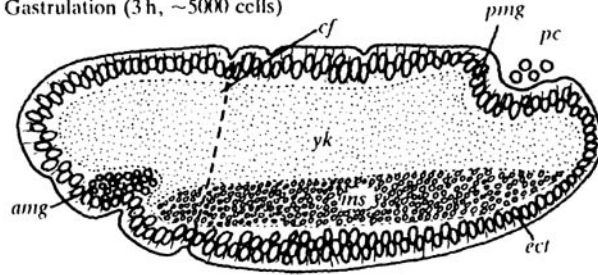
B. Syncytial blastoderm (2h, ~1500 nuclei)



C. Cellular blastoderm (2.5h, ~5000 cells)



D. Gastrulation (3 h, ~5000 cells)



E. Extended germ band (4.5 h, >5000 cells).

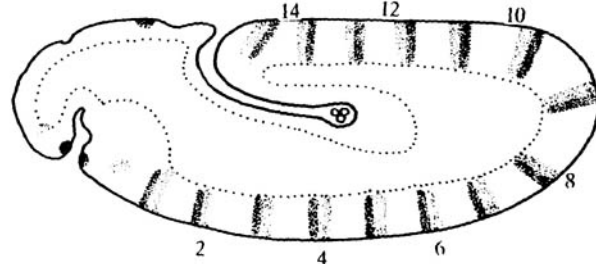
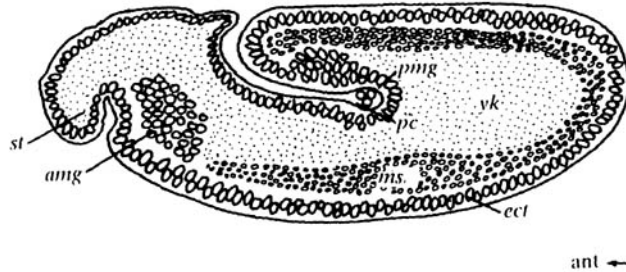


FIG. A.4. “Patterns of gene activity during early *Drosophila* development . . . Diagrams on the left show the morphology of stages during early embryogenesis. Corresponding panels on the right show patterns of gene activity established at the corresponding stages: A. Localized maternal determinants: *bicoid* RNA (crosses); polar granules (dots). The *bicoid* protein gradient is shown by shading. B. Gap gene expression: *hunchback*, *Krüppel*, *knirps* (shading, zones from anterior to posterior); *tailless* (stipple at both ends). C. Pair rule stripes: *even-skipped* (dark) and *fushi-tarazu* (light). D, E. Evolving pattern of segment polarity gene expression: *wingless* (dark) and *engrailed* (light).” By M. Akam, from *The Encyclopaedia of Molecular Biology* (Oxford: Blackwell Scientific Publications), reproduced by permission.

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*Gene expression (cont.)*

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Hemolymph protein

CEC 51

YP 305

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## Intron, alternative splicing

*bcd* 64*Dac* 111*otu* 232*Ubx* 278

## Intronless gene

*ac* 6*AmyA, AmyB* 46*ase* 12*Hsp67B* 171*Hsp70* 192*lsc* 12*sc* 10*Sryα* 262*Sryδ* 268*Vm26Aa* 291*Vm26Ab* 293*Vm32E* 294*Vm34C* 295*janA, janB* 201, 261*janus*, see *janA, janB**kni* 160, 167, 205, 216*knirps*, see *kni**knirps-related*, see *knrl**knrl* 214*Kr* 60, 140, 160, 162, 209, 212, 215*Krüppel*, see *Kr*Larval cuticle proteins genes (*Lcp1–4*) 88*Lcp1–4* 88–93

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multiple

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*hb* 168

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*Act5C* distal 20

*Act79B* 29

*Act87E* 31

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*Adh* distal 38

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*Anp* 53

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*Yp2* 314

with TATA box

*ac* 6

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*Act79B* 29

*Act87E* 31

*Act88F* 34

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*Adh* proximal 38

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*ase* 12

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*Cp15* 81

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*Pcp* 93  
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*Cp18* 85  
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EVE 135  
 KR 215  
 UBX 270

## ATP-binding site HSP70 189

## basic helix-loop-helix (bHLH)

AS-SC family 3  
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## degradation signal

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## denatured protein binding site HSP70 189

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## OPA repeat

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