FASTtrack

Pharmaceutics – Dosage Form and Design

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Dedicated to my mother May, my late father Fred,
my wife Linda and our children Dary and Holly.

Contents

Introduction to the
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FASTtrack is a new series of study guides created for undergraduate pharmacy students. The books are intended to be used in conjunction with textbooks and reference books as an aid to revision to help guide students through their exams. They provide essential information required in each particular subject area. The books will also be useful for pre-registration trainees preparing for the Royal Pharmaceutical Society of Great Britain's (RPSGB's) registration examination, and to practising pharmacists as a quick reference text.

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Preface

Pharmaceutical formulation involves the rational design and manufacture of dosage forms to ensure that the required biological and physical performances of the therapeutic agent are attained. The formulation scientist is therefore expected to have knowledge of several scientific disciplines, including physical pharmaceutics, pharmaceutical chemistry and biopharmaceutics. Pharmaceutical formulation may therefore be correctly viewed as an essential and unique component of the pharmaceutical sciences and, accordingly, the Pharmacy undergraduate degree. Due to the interdisciplinary nature, pharmaceutical formulation is often considered difficult by many students and challenging to all students. This textbook aims to ease the perceived difficulties of this subject and, hopefully, illustrate the significance of pharmaceutical formulation and the unique role of the pharmacist in the development of medicines.

The contents of this text have been designed to cover the vast majority of dosage forms that graduate pharmacists will encounter when they enter professional practice. In particular, the text aims to deliver the essential information concerning the formulation of these dosage forms in a format that will hopefully aid the understanding and hence remove the complexities of the various topics. The reader will observe that, in addition to the required theoretical aspects, each chapter contains information regarding the types (and physicochemical properties) of excipients that are used in the formulation of dosage forms. It is the author's opinion that this information is essential to ensure that the reader is able to traverse the information gap between the theory and practice of dosage form design. In understanding the physicochemical properties of excipients (and indeed therapeutic agents), the student will be able to optimise the prepared dosage form and, in addition, will be able to answer examination questions with the appropriate depth required to attain a high grade.

Each chapter describes a category of dosage form and, whilst the exact details change, it may be observed that the format of the chapters is similar. This will assist students in understanding and recalling the required information. The formulations of liquids and suspensions for oral administration, including the required theoretical aspects, are presented in Chapters 1 and 2. In Chapters 3 and 4 the theoretical aspects and formulations of products that contain disperse phases are described. Whilst these products are mostly administered topically, the formulation of emulsions for oral administration is also described. Chapter 5 describes parenteral formulations and, in particular, illustrates strategies for the successful formulation of parenteral solutions, suspensions and emulsions (including the considerations of osmolality and sterility and methods to attain sterility).

The formulation of ocular (and related) dosage forms is described in Chapter 6. In addition to the description of ocular formulations, this chapter concisely describes the biological demands of these formulations and, in so doing, highlights the effects of formulation type on the biological performance of the preparation. Chapter 7 examines the formulation of suppositories and pessaries, describing the various formulation strategies for these dosage forms and highlighting their biological demands. The formulation of respiratory dosage forms, namely metered-dose inhalers, dry-powder inhalers and solutions for nebulisation, is presented in Chapter 8. In addition to the description of the typical excipients, this chapter specifically highlights formulation factors that affect deposition of particles within the respiratory tract. It is worth noting that in Chapters 6–8 the biological aspects of drug delivery to the eye, nose, rectum, vagina and respiratory tract are described, as such information is required to ensure the design of formulations that provide optimal biological performance.

Finally, Chapters 9 and 10 describe the formulation of solid-dosage forms (tablets and capsules) with particular emphases on the excipients and methods used to prepare them, the different types of tablets and capsules, the tableting process, tableting defects and, finally, the film coating of tablets. The size of these chapters reflects the greater clinical usage of these dosage forms.

Writing this textbook presented many challenges to the author. Above all, I hope that, after completing this text, readers will gain a comprehensive understanding of pharmaceutical formulation that will enable them to formulate dosage forms and, importantly, give advice on this topic to other healthcare professionals, who are frequently unaware of the complexities of this topic.

It has been my great pleasure and privilege to have lectured on all of the topics described in this textbook to enthusiastic undergraduate and postgraduate students of pharmacy in both Northern Ireland and New Zealand. I am grateful to them, as their numerous questions and suggestions have informed the content of this textbook. I am also indebted to Professor Michael S. Roberts (University of Queensland) who, at the commencement of my academic career, inspired my enthusiasm for, and helped me navigate through, the scientific complexities of physical pharmaceutics and pharmaceutical formulation. His contribution to my career has been invaluable. My gratitude is also given to Dr Ryan Donnelly and Dr Gavin Andrews, both of the School of Pharmacy, Queen's University of Belfast, for the many hours that they spent proof-reading this text and for their comments on how its content and presentation could be improved. Finally I am sincerely grateful for the love, encouragement and support of my mother May, my late Father Fred, my wife Linda and our children, Dary and Holly, without which this academic journey would not have been possible. This book is dedicated to them.

David S. Jones
March 2008

About the author

DAVID JONES gained a BSc (first-class honours) in Pharmacy (1985), a PhD in Pharmaceutics (1988) and, in 2006, a DSc. He registered with the Pharmaceutical Society of Northern Ireland in 1989 and took up a lectureship in Pharmaceutics in the School of Pharmacy, University of Otago, New Zealand. In 1992 he was appointed as the Head of Formulations at Norbrook Laboratories. In 1994, he was appointed to a lectureship in Pharmaceutics and was subsequently promoted to a senior lectureship (1997) and to a personal Chair (in Biomaterial Science) in 1999. His research concerns the characterisation, formulation and engineering of pharmaceutical materials/dosage forms and biomedical devices.

He is the author of two textbooks, six patents and over 350 research papers/communications. Professor Jones is both a Chartered Engineer and a Chartered Chemist and is a Fellow of the Institute of Materials, Minerals and Mining, a Fellow of the Royal Statistical Society, a Fellow of the Royal Society of Chemistry and a Member of the Institute of Engineers in Ireland. He is the editor of the *Journal of Pharmacy and Pharmacology.*

chapter 1
Pharmaceutical solutions for oral administration

Overview

In this chapter we will:
- examine the types and uses of pharmaceutical solutions as oral drug delivery systems
- provide an overview of the advantages and disadvantages of pharmaceutical solutions as oral drug delivery systems
- describe the formulation considerations for orally administered pharmaceutical solutions.

General description

Pharmaceutical solutions may be generally defined as liquid preparations in which the therapeutic agent and the various excipients are dissolved in the chosen solvent system. Pharmaceutical solutions may contain a range of excipients, each with a defined pharmaceutical purpose. Examples of these include:
- the vehicle, usually purified water
- co-solvents, e.g. propylene glycol, glycerin, alcohol
- agents specifically to enhance the solubility of the therapeutic agent in the vehicle, e.g. surface-active agents
- preservatives, e.g. parahydroxybenzoate esters (methylhydroxy-benzoate and propylhydroxybenzoate), boric acid and borate salts, sorbic acid and sorbate salts, phenolics
- sweeteners, e.g. glucose, saccharin, aspartame
- rheology (viscosity) modifiers, e.g. hydrophilic polymers (cellulose derivatives, alginic acid, polyvinylpyrrolidone)
- antioxidants, e.g. sodium formaldehyde sulphoxylate, butylated hydroxyanisole, butylated hydroxytoluene
- colours
- flavours
- buffers to regulate the pH of the formulation, e.g. citrate buffer.

The specific roles of each of these formulation excipients will be described later in this chapter.

KeyPoints

- Pharmaceutical solutions are extensively used as dosage forms for the oral administration of therapeutic agents.
- Pharmaceutical solutions are homogeneous, i.e. the therapeutic agent(s) and excipients are dissolved in the vehicle.
- Pharmaceutical solutions for oral administration are non-sterile dosage forms.

Advantages and disadvantages of pharmaceutical solutions for oral administration

Advantages

- Therapeutic agents can easily be administered orally to individuals who have difficulty in swallowing, e.g. elderly patients, infants.
- The therapeutic agent is dissolved in the formulation and is therefore immediately available for absorption. Providing the drug does not precipitate within the gastrointestinal tract, the bioavailability of pharmaceutical solutions is greater than that of oral solid-dosage forms.
- Taste-masking of bitter therapeutic agents may be readily achieved.

Disadvantages

- Pharmaceutical solutions for oral administration are unsuitable for therapeutic agents that are chemically unstable in the presence of water.
- The poor solubility of certain therapeutic agents may prohibit their formulation as pharmaceutical solutions. The reader should note that certain techniques are available to enhance the solubility of poorly soluble drugs. These will be highlighted later in this chapter.
- Pharmaceutical solutions are expensive to ship and are bulky for the patient to carry due to the associated mass of the product.

Drug solubility

In pharmaceutical solutions both the therapeutic agent and the excipients are legally required to be present in solution over the shelf-life of the formulated product. As a result pharmaceutical solutions are termed homogeneous. One of the major challenges to the pharmaceutical scientist is the attainment of homogeneity in the formulation, due primarily to, in many cases, the limited aqueous solubility of the therapeutic agent. Initially there are possible scenarios regarding the formulation of pharmaceutical solutions of a therapeutic agent for oral administration:

- The aqueous solubility of the therapeutic agent is high at the selected pH of the formulation. Under these circumstances the therapeutic agent may be readily incorporated into the vehicle and formulated as an oral solution.
- The aqueous solubility of the therapeutic agent is moderate at the selected pH of the formulation, i.e. the aqueous solubility is less than the requested concentration of therapeutic agent. Under these circumstances the solubility of the therapeutic

agent in the formulation must be enhanced using co-solvents and related methods.
- The aqueous solubility of the therapeutic agent is low at the selected pH of the formulation. The difference between the aqueous solubility of the therapeutic agent and the required concentration is too great to be bridged by the use of co-solvents and related methods or the concentration of co-solvents or surfactants in the solubilised formulation may be toxic when administered orally. The drug may therefore be formulated as an alternative-dosage form, e.g. a suspension.

Prior to discussing the solubility of therapeutic agents and formulation strategies to modify this property, it is worth considering the process of drug dissolution. The dissolution of a therapeutic agent in water involves several key molecular steps: the removal of a molecule of the drug from the solid state, the formation of a cavity within the solvent and the accommodation of the drug molecule into the formed cavity. This process involves the breakage of solute–solute and solvent–solvent bonds (endothermic processes) and the formation of a bond between the solute and the solvent (with the subsequent liberation of energy). Dissolution occurs whenever the Gibb's free energy (ΔG) of the process is negative and involves a balance between the enthalpy of dissolution (ΔH) and the associated entropy (ΔS) at the temperature of dissolution (T), as defined below:

$$\Delta G = \Delta H - T\Delta S$$

Factors affecting the solubility of therapeutic agents

The solubility properties of drug molecules in a particular solvent system are sometimes difficult to predict and have been reported to be dependent, at least in part, on several physicochemical properties, including molecular weight, volume, radius of gyration, density, number of rotatable bonds, hydrogen bond donors and hydrogen bond acceptors. Furthermore, the properties of the solid state, e.g. crystal habit, crystalline/amorphous properties, will also affect the solubility of the therapeutic agent.

There are some empirical relationships between the physicochemical properties and the solubility of therapeutic agents that influence formulation strategies, as follows:
- The solubilities of a chemically related series of therapeutic agent are inversely related to their melting points. Therefore, as the melting point of the therapeutic agent is increased, the solubility would be expected to decrease.
- The solubility of a therapeutic agent is directly affected by both the type of chemical substituent groups and the substituent position. The solubility of therapeutic agents

containing hydrophilic groups (e.g. OH, COO⁻, ammonium ion) will accordingly be greater than those containing lipophilic substituent groups, e.g. methyl, ethyl, ethoxy or chlorine groups.

■ The solubilities of therapeutic agents that are either acids or bases (representing the vast majority of drug substances) are pH-dependent. The solubility of acids and bases increases as the degree of ionisation increases and may be easily calculated using the following equation (where S refers to the solubility of the drug and S_o is the intrinsic solubility, i.e. the solubility of the unionised form of the drug).

$$pK - pKa = \log\left(\frac{S - S_o}{S_o}\right) \qquad \text{for acids}$$

$$pH - pKa - \log\left(\frac{S_o}{S - S_o}\right) \qquad \text{for bases}$$

From these equations two invaluable conclusions may be drawn:

– At pH values *above* the pKa, the solubility of acidic drugs *increases.*
– At pH values *below* the pKa, the solubility of basic drugs *increases.*

In simple terms the solubility of acidic compounds increases as the pH of the solution is increased (above the pKa) and the solubility of basic compounds increases as the pH is lowered below the pKa.

Determination of the solubility properties of zwitterionic compounds, i.e. those that exhibit both acidic and basic properties, is more complicated than for simple acids or bases. However, in common with simple acids and bases, the solubility of zwitterionic therapeutic agents is affected by pH. At basic pH values the therapeutic agent behaves primarily as an acid whereas at low pH values the molecule behaves as a base. The pH range at which the therapeutic agent exhibits minimal solubility lies between the pKa values of the acidic and basic groups.

Formulation methods to enhance/optimise the solubility of therapeutic agents

The information described below may be employed to optimise the formulation of pharmaceutical solutions, remembering that the prerequisite for pharmaceutical solutions is the exclusive presence of dissolved therapeutic agent.

Appropriate selection of drug salt

The reader will be aware that the majority of therapeutic agents are commercially available to the pharmaceutical scientist in a range of salt forms, each form exhibiting a different aqueous solubility. The differences in solubility may be accredited, at least in part, to the crystal properties of the salt, which, in turn, affect the energy required to dissociate solute–solute bonds. Therefore, unless a specific salt form is specified or in the absence of a pharmaceutical approved salt of a therapeutic agent, the formulation scientist should select the salt that provides the required solubility in the dosage form.

Optimisation of the pH of the formulation

As mentioned above, the solubility of an ionised therapeutic agent is a function of both the pKa of the compound and the pH of the formulation. Importantly, the acceptable pH range of solutions for oral administration is large, ranging from circa 5 to 8 pH units. Therefore, a common formulation strategy involves the selection of a pH value for the formulation that optimises the ionisation and hence solubility of the therapeutic agent. Control of the pH in the formulation is achieved using a buffer that does not adversely affect the solubility of the therapeutic agent.

Use of co-solvents

Co-solvents are primarily liquid components that are incorporated into a formulation to enhance the solubility of poorly soluble drugs to the required level. In the formulation of pharmaceutical solutions for oral administration, aqueous solutions are preferred due to the lack of toxicity of water as the vehicle. However, if the solubility of the therapeutic agent renders this approach inappropriate, the incorporation of co-solvents within the formulation offers a pharmaceutically acceptable approach. Commonly employed co-solvents include glycerol, propylene glycol, ethanol and poly(ethylene glycol), details of which are provided in subsequent sections.

Prediction of the solubility of therapeutic agents in mixed solvent systems (the vehicle, water and the chosen co-solvent) is difficult, due to the effects of many variables on the solubility (as described previously). In practice the pharmaceutical scientist should measure the solubility of the chosen therapeutic agent in a series of mixed solvents to determine the most suitable solvent system for the given purpose. The final choice of the co-solvent system for a particular formulation involves consideration of the solubility of the therapeutic agent in the vehicle, the toxicity of the vehicle and the cost of the formulation. Indeed, it should be noted that the range of concentrations of each co-solvent used in oral formulations is primarily limited by concerns regarding toxicity.

Excipients used in pharmaceutical solutions for oral administration

Excipients in pharmaceutical formulations are physiologically inert compounds that are included in the formulation to facilitate the administration of the dosage form, e.g. pourability, palatability, to protect the formulation from issues regarding physical and chemical stability and to enhance the solubility of the therapeutic agent. Pharmaceutical solutions commonly contain a wide range of excipients, the details of which are provided below.

The vehicle

The preferred and most commonly used vehicle in solutions for oral administration is Purified Water USP, due to the low cost and low toxicity of this ingredient. Under normal circumstances tap (drinking) water should not be used due to the possibility of chemical imcompatibities within the formulation. The main features of Purified Water USP are as follows:

- It is prepared by distillation, ion exchange methods or by reverse osmosis.
- The solid residue (obtained after evaporation) is less than 1 mg per 100 ml of evaporated sample.
- It must not be used for the preparation of parenteral formulations.

In the case of parenteral formulations *Water for Injections BP* must be used, the specifications and use of which are described in Chapter 5.

Co-solvents

As defined previously, co-solvents are employed to increase the solubility of the therapeutic agent within the formulation. The main co-solvents that are used in the formulation of oral solutions are detailed below.

Glycerol

Glycerol (also termed glycerin) is an odorless, sweet liquid that is miscible with water and whose co-solvency properties are due to the presence of three hydroxyl groups (termed a triol) (Figure 1.1). It has similar co-solvency properties to ethanol.

Figure 1.1 Structural formula of glycerol.
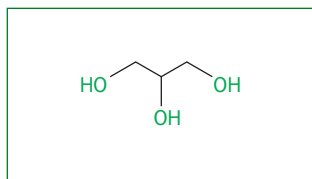
Alcohol USP (CH_3CH_2OH)

Alcohol USP contains between 94.9 and 96.0% v/v ethyl alcohol (ethanol) and is commonly used as a co-solvent, both as a single co-solvent and with other co-solvents, e.g. glycerol. The known pharmacological and toxicological effects of this co-solvent have compromised the use of alcohol in pharmaceutical preparations. As a result there are both labelling requirements for preparations that contain alcohol and upper limits with respect to the concentration of alcohol that may be used in formulations.

Propylene Glycol USP

Propylene Glycol USP is an odourless, colourless, viscous liquid diol that contains two hydroxyl groups (Figure 1.2). It is used in pharmaceutical preparations as a co-solvent, generally as a replacement for glycerin.
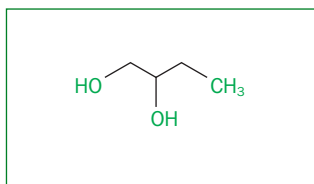
Figure 1.2 Structural formula of propylene glycol.

Poly(ethylene glycol) (PEG)

PEG (Figure 1.3) is a polymer composed of repeating units of the monomer ethylene oxide (in parenthesis). The physical state of the polymer is dependent on the number of repeat units (n) and hence on the molecular weight. Lower-molecular-weight grades (PEG 200, PEG 400) are preferred as co-solvents in pharmaceutical solutions.
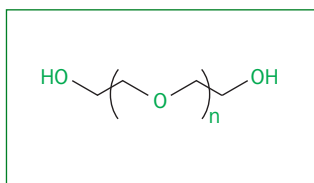
Figure 1.3 Structural formula of poly(ethylene glycol).

Miscellaneous agents used to enhance the solubility of therapeutic agents

In addition to the use of co-solvents, other pharmaceutical strategies are available to the pharmaceutical scientist to increase the solubility of therapeutic agents in the chosen vehicle. These include the use of surface-active agents and complexation, as detailed below.

Surface-active agents

Surface-active agents are chemicals that possess both hydrophilic (water-liking) and hydrophobic (water-disliking) regions. At dilute concentrations surface-active agents will orient at the interface between two phases (e.g. water/oil, water/air), with the hydrophilic and hydrophobic regions of the molecule being positioned to the hydrophilic and hydrophobic phases, respectively. As the concentration is increased, the interface will become saturated with surface-active agent and the molecules that are present in the bulk aqueous phase will orient themselves in an attempt to shield the hydrophobic regions of the surface-active agent. This orientation is referred to as a *micelle* and the concentration of surface-active agent at this occurs is termed the *critical micelle concentration* (CMC).

For further details regarding the physicochemical properties of surfactants, the reader should consult the companion text by David Attwood and Alexander T Florence (*FASTtrack: Physical Pharmacy* (London: Pharmaceutical Press; 2008). The use of surface-active agents for the solubilisation of poorly soluble drugs occurs exclusively in the presence of micelles and hence at concentrations of surface-active agents in excess of the CMC. In this the core of the micelle represents a hydrophobic region into which the poorly water-soluble drugs may partition. The location in the micelle is related to the chemical structure of the drug. For example, if the therapeutic agent is poorly soluble the molecule will locate exclusively within the micelle, whereas if the drug is water-insoluble but contains polar groups, the molecule will orient within the micelle, with the polar groups at the surface of the micelle and the hydrophobic region of the molecule located within the hydrophobic core of the micelle. In so doing the drug is solubilised within the colloidal micelles; due to their small size, the resulting solution appears homogeneous to the naked eye.

## Tip

As the reader will have observed, there are several methods that may be used for the solubilisation of therapeutic agents. The choice of method should involve consideration of the stability of the formed solution, the pharmaceutical acceptability of the solubilisation strategy and cost.

Complexation

Complexation refers to the interaction of a poorly soluble therapeutic agent with an organic molecule, e.g. surface-active agents, hydrophilic polymers to generate a soluble intermolecular complex. One particular concern regarding the use of solution of drug complexes is the ability of the complex to dissociate following administration. This is particularly important in situations where the complexing agent is a hydrophilic polymer, as the high molecular weight of the

drug–polymer complex would prevent drug absorption across biological membranes.

Common excipients in pharmaceutical solutions

There are several excipients that are commonly employed in the formulation of pharmaceutical solutions. These include: (1) buffers; (2) sweetening agents; and (3) viscosity-enhancing agents.

Buffers

Buffers are employed within pharmaceutical solutions to control the pH of the formulated product and, in so doing, optimise the physicochemical performance of the product. Typically pH control is performed:

- to maintain the solubility of the therapeutic agent in the formulated product. The solubility of the vast number of currently available drugs is pH-dependent and, therefore, the solubility of the therapeutic agent in the formulation may be compromised by small changes in pH
- to enhance the stability of products in which the chemical stability of the active agent is pH-dependent.

The concentration (and hence buffer capacity) of buffer salts employed in the formulation of oral solutions should be selected to offer sufficient control of the pH of the formulation but yet should be overcome by biological fluids following administration. This latter property is particularly appropriate for parenteral formulations to ensure that there is no irritation or damage following injection.

Examples of buffer salts used in pharmaceutical solutions include:

- acetates (acetic acid and sodium acetate): circa 1–2%
- citrates (citric acid and sodium citrate): circa 1–5%
- phosphates (sodium phosphate and disodium phosphate): circa 0.8–2%.

It must be remembered that the buffer system used in solution formulations should not adversely affect the solubility of the therapeutic agent, e.g. the solubility of drugs may be affected in the presence of phosphate salts.

Sweetening agents

Sweetening agents are employed in liquid formulations designed for oral administration specifically to increase the palatability of the therapeutic agent. The main sweetening agents employed in oral preparations are sucrose, liquid glucose, glycerol, sorbitol, saccharin sodium and aspartame. The use of artificial sweetening agents in formulations is increasing and, in many formulations,

saccharin sodium is used either as the sole sweetening agent or in combination with sugars or sorbitol to reduce the sugar concentration in the formulation. The use of sugars in oral formulations for children and patients with diabetes mellitus is to be avoided.

Viscosity-enhancing agents

The administration of oral solutions to patients is usually performed using a syringe, a small-metered cup or a traditional 5-ml spoon. The viscosity of the formulation must be sufficiently controlled in order to ensure the accurate measurement of the volume to be dispensed. Furthermore, increasing the viscosity of some formulations may increase the palatability. Accordingly there is a viscosity range that the formulation should exhibit to facilitate this operation. Certain liquid formulations do not require the specific addition of viscosity-enhancing agents, e.g. syrups, due to their inherent viscosity.

The viscosity of pharmaceutical solutions may be easily increased (and controlled) by the addition of non-ionic or ionic hydrophilic polymers. Examples of both of these categories are shown below:

- non-ionic (neutral) polymers
  - cellulose derivatives, e.g.:
    - methylcellulose
    - hydroxyethylcellulose
    - hydroxypropylcellulose
  - polyvinylpyrrolidone
- ionic polymers
  - sodium carboxymethylcellulose (anionic)
  - sodium alginate (anionic).

Full details of the physicochemical properties of these polymers are provided in later chapters.

Antioxidants

Antioxidants are included in pharmaceutical solutions to enhance the stability of therapeutic agents that are susceptible to chemical degradation by oxidation. Typically antioxidants are molecules that are redox systems which exhibit higher oxidative potential than the therapeutic agent or, alternatively, are compounds that inhibit free radical-induced drug decomposition. Typically in aqueous solution antioxidants are oxidised (and hence degraded) in preference to the therapeutic agent, thereby protecting the drug from decomposition. Both water-soluble and water-insoluble antioxidants are commercially available, the choice of these being made according to the nature of the formulation. Examples of antioxidants that are commonly used

for aqueous formulations include: sodium sulphite, sodium metabisulphite, sodium formaldehyde sulphoxylate and ascorbic acid. Examples of antioxidants that may be used in oil-based solutions include: butylated hydroxytoluene (BHT), butylated hydroxyanisole (BHA) and propyl gallate. Typically antioxidants are employed in low concentrations (< 0.2% w/w) and it is usual for the concentration of antioxidant in the finished product to be markedly less than the initial concentration, due to oxidative degradation during manufacture of the dosage form. Antioxidants may also be employed in conjunction with chelating agents, e.g. ethylenediamine tetraacetic acid, citric acid, that act to form complexes with heavy-metal ions, ions that are normally involved in oxidative degradation of therapeutic agents.

Preservatives

Preservatives are included in pharmaceutical solutions to control the microbial bioburden of the formulation. Ideally, preservatives should exhibit the following properties:

- possess a broad spectrum of antimicrobial activity encompassing Gram-positive and Gram-negative bacteria and fungi
- be chemically and physically stable over the shelf-life of the product
- have low toxicity.

A wide range of preservatives is available for use in pharmaceutical solutions for oral use, including the following (values in parentheses relate to the typical concentration range used in oral solutions):

- benzoic acid and salts (0.1–0.3%)
- sorbic acid and its salts (0.05–0.2%)
- alkyl esters of parahydroxybenzoic acid (0.001–0.2%). Usually a combination of two members of this series is employed in pharmaceutical solutions, typically methyl and propyl parahydroxybenzoates (in a ratio of 9:1). The combination of these two preservatives enhances the antimicrobial spectrum.

Factors affecting preservative efficacy in oral solutions

The activity of a preservative is dependent on the correct form of the preservative being available in the formulation at the required concentration to inhibit microbial growth (termed the minimum inhibitory concentration: MIC). Unfortunately, in many solution formulations, the concentration of preservative within the formulation may be affected by the presence of other excipients and by formulation pH. Factors that directly affect the efficacy of

preservatives in oral solutions include: (1) the pH of the formulation; (2) the presence of micelles; and (3) the presence of hydrophilic polymers.

The pH of the formulation

In some aqueous formulations the use of acidic preservatives, e.g. benzoic acid, sorbic acid, may be problematic.

Figure 1.4 Structural formula of (a) benzoic acid and (b) sorbic acid.
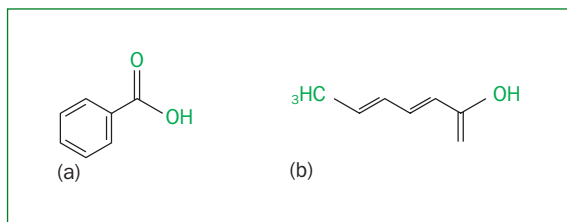
The antimicrobial properties are due to the unionised form of the preservative; the degree of ionisation being a function of the pH of the formulation. The activity of the unionised form of the acid in this respect is due to the ability of this form to diffuse across the outer membrane of the microorganism and eventually into the cytoplasm. The neutral conditions within the cytoplasm enable the preservative to dissociate, leading to acidification of the cytoplasm and inhibition of growth.

The fraction of acidic preservative at a particular pH may be calculated using a derived form of the Henderson–Hasselbalch equation, as follows:

$$Fraction = \left(\frac{1}{(1+10^{pH-pKa})} \right)$$

The use of this equation may be illustrated in the following example:

Worked example

Example 2.1

Assuming that the MIC for the unionised form of an acidic preservative (pKa 4.2) is 0.0185 mg/ml, calculate the required concentration to preserve an oral solution that has been buffered to pH 4.7.

The Henderson–Hasselbalch equation may be employed, as described above, to determine the fraction of unionised acid within the formulation.

$$Fraction = \left(\frac{1}{(1+10^{4.7-4.2})} \right) = 0.24$$

The required concentration is then calculated by dividing the MIC for the unionised form of the preservative by the fraction of unionised preservative present, i.e. $\left(\dfrac{0.0185}{0.24}\right) = 0.07$ mg/ml.

In practice an overage is added and therefore the actual concentration of preservative required would be 0.1–0.15 mg/ml.

As the reader will observe, the pKa of the preservative is a vital determinant within the above calculations. Organic acids, e.g. benzoic acid, sorbic acid, have pKa values that are circa 4.2 and therefore, in solution formulations whose pH is neutral, a high concentration of preservative will be required to ensure that the required concentration of the unionised species is obtained. If the above calculation is repeated for an oral solution at pH 7.2, the following result is obtained:

$$Fraction = \left(\frac{1}{(1+10^{7.2-4.2})}\right) = 0.00001$$

Therefore, the required preservative concentration is $\left(\dfrac{0.0185}{0.00001}\right) = 1850$ mg/ml.

Importantly, the preservative efficacies of parabens (alkyl esters of parahydroxybenozoic acid) and the phenolics are generally not affected by formulation pH (within a pH range between 4.0 and 8.0) due to the high pKa of the organic hydroxyl group. The structures of these preservatives are shown in Figure 1.5.
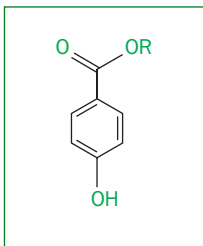
Figure 1.5 Structural formula for the parahydroxybenzoate esters (parabens, where R refers to an alkyl group).

The presence of micelles

The role of micelles for the solubilisation of lipophilic therapeutic agents was described above. If the preservative exhibits lipophilic properties (e.g. the unionised form of acidic preservatives, phenolics, parabens), then partition of these species into the micelle may occur, thereby decreasing the available (effective) concentration of preservative in solution. An equilibrium is established, as depicted in Figure 1.6.

Figure 1.6 Diagrammatic illustration of the equilibrium of an acidic preservative in the presence of micelles. HA and A$^-$ refer to the unionised and ionised states of the preservative.
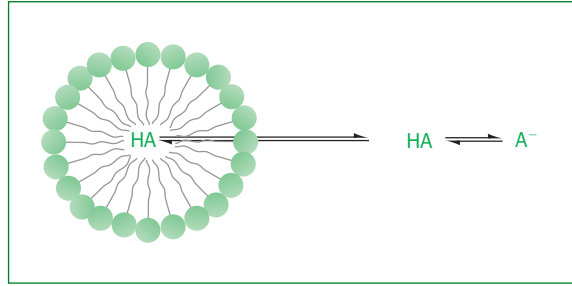
To correct this problem, the preservative concentration must be increased to ensure that the free concentration within the formulation is ≥ MIC of the preservative.

The presence of hydrophilic polymers
It has been shown that the free concentration of preservative in oral solution formulations is reduced in the presence of hydrophilic polymers, e.g. polyvinylpyrrolidone, methylcellulose. This is due to the ability of the preservative to interact chemically with the dissolved polymer. As described above, this problem is addressed by increasing the concentration of preservative in the formulation. In certain circumstances the preservative may be incompatible with hydrophilic polymers in the formulation due to an electrostatic interaction. Therefore, cationic hydrophilic polymers should not be used in conjunction with acidic preservatives in oral solution formulations.

Flavours and colourants
Unfortunately the vast majority of drugs in solution are unpalatable and, therefore, the addition of flavours is often required to mask the taste of the drug substance. Taste-masking using flavours is a difficult task; however, there are some empirical approaches that may be taken to produce a palatable formulation.

- The four basic taste sensations are salty, sweet, bitter and sour. It has been proposed that certain flavours should be used to mask these specific taste sensations. In particular:
  - Flavours that may be used to mask a salty taste include:
    - butterscotch
    - apricot
    - peach
    - vanilla
    - wintergreen mint.
  - Flavours that may be used to mask a bitter taste include:
    - cherry
    - mint
    - anise.

- Flavours that may be used to mask a sweet taste include:
  - vanilla
  - fruit and berry.
- Flavours that may be used to mask a sour taste include:
  - citrus flavours
  - raspberry.
- Usually a combination of flavours is used to achieve the optimal taste-masking property.
- Certain excipients may be added to oral solution formulations, referred to as *flavour adjuncts* (e.g. menthol, chloroform) that add flavour to the formulation but, in addition, act to desensitise the taste receptors. In so doing these agents augment the taste-masking properties of conventional flavours.

Colours are pharmaceutical ingredients that impart the preferred colour to the formulation. When used in combination with flavours, the selected colour should 'match' the flavour of the formulation, e.g. green with mint-flavoured solutions, red for strawberry-flavoured formulations. Although the inclusion of colours is not a prerequisite for all pharmaceutical solutions, certain categories of solution (e.g. mouthwashes/gargles) are normally coloured.

Types of pharmaceutical solutions

Pharmaceutical solutions for oral administration

There are three principal types of solution formulations that are administered orally: *oral solutions, oral syrups* and *oral elixirs*. In addition, other solution formulations are employed for a local effect, e.g. mouthwashes/gargles and enemas. Details of these are provided in the following sections.

Tips

The formulation of solutions for oral administration often requires the inclusion of several excipients.

It is important that each excipient included in the formulation is necessary and justified.

All excipients must be physically and chemically compatible (with each other and with the therapeutic agent).

Oral solutions

Oral solutions are administered to the gastrointestinal tract to provide systemic absorption of the therapeutic agent. Due to the resilience of the gastrointestinal environment, oral solutions may be formulated over a broad pH range. However, unless there are issues regarding the solubility or stability of the therapeutic agent, the usual pH of oral solutions is circa 7.0. Typically the following classes of excipients are used in the formulation of oral solutions:

- buffers (e.g. citrate, phosphate)
- preservatives (e.g. parabens, benzoic acid, sorbic acid)

- antioxidants (water-soluble antioxidants are used, e.g. sodium metabisulphite 0.01–1.0% w/w)
- flavours and colours (the colour should be selected to complement the flavour of the formulation)
- viscosity-modifying agents (to affect the pourability of the formulation. For this purpose hydrophilic polymers are used, e.g. sodium alginate, hydroxyethylcellulose).

The reader should note that, to be classified as a solution, all components of the formulation (including the therapeutic agent) should be soluble, with no evidence of precipitation.

Oral syrups

Syrups are highly concentrated, aqueous solutions of sugar or a sugar substitute that traditionally contain a flavouring agent, e.g. cherry syrup, cocoa syrup, orange syrup, raspberry syrup. An unflavoured syrup is available that is composed of an aqueous solution containing 85% sucrose. Therapeutic agents may either be directly incorporated into these systems or may be added as the syrup is being prepared. If the former method is employed, it is important to ensure that the therapeutic agent is soluble within the syrup base.

It should also be remembered that the choice of syrup vehicle must be performed with due consideration to the physicochemical properties of the therapeutic agent. For example, cherry syrup and orange syrup are acidic and therefore the solubility of acidic or some zwitterionic therapeutic agents may be lowered and may result in precipitation of the drug substance. Under these circumstances, the physical stability of the preparation will have been compromised and the shelf-life of the product will have been exceeded. The use of acidic syrups may additionally result in reduced chemical stability for acid-labile therapeutic agents.

The major components of syrups are as follows:

- *Purified water.*
- *Sugar (sucrose) or sugar substitutes (artificial sweeteners).* Traditionally syrups are composed of sucrose (usually between 60 and 80%) and purified water. Due to the inherent sweetness and moderately high viscosity of these systems, the addition of other sweetening agents and viscosity-modifying agents is not required. In addition, the high concentration of sucrose and associated unavailability of water (termed low water activity) ensures that the addition of preservatives is not required. As the concentration of sucrose is reduced from the upper limit (e.g. through dilution), the addition of preservatives may be required.

  In some formulations, other non-sucrose bases may replace traditional syrup. One of the most popular is Sorbitol

Solution USP, which contains 64% w/w sorbitol (a polyhydric alcohol: Figure 1.7), although other alternatives are available that are based on mixtures of sorbitol and glycerin. These non-sucrose bases may be mixed with traditional syrups, if required, in the formulation of oral syrups that possess a low concentration of sucrose in comparison to traditional syrups.
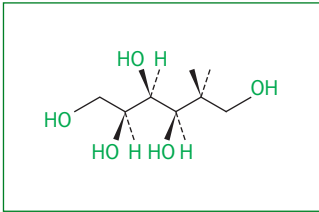
Figure 1.7 Structural formula of sorbitol.

More recently, many products have been formulated as medicated sugar-free syrups due to the glycogenetic and cariogenic properties of sucrose. For the afore-mentioned reasons, all medicinal products designed for administration to children and to diabetic patients must be sugar-free. Syrup substitutes must therefore provide an equivalent sweetness, viscosity and preservation to the original syrups. To achieve these properties artificial sweeteners (typically saccharin sodium, aspartame), non-glycogenetic viscosity modifiers (e.g. methylcellulose, hydroxyethylcellulose) and preservatives (e.g. sodium benzoate, benzoic acid and parahydroxybenzoate esters) are included.

■ *Preservatives.* As highlighted above, preservatives are not required in traditional syrups containing high concentrations of sucrose. Conversely, in sugar-free syrups, syrups in which sucrose has been substituted at least in part by polyhydric alcohol and in traditional syrups that contain lower concentrations of sucrose, the addition of preservatives is required. Typical examples of commonly used preservatives include:

– Mixtures of parahydroxybenzoate esters (usually methylhydroxybenzoate and propylhydroxybenzoate in a ratio of 9:1). The typical concentration range is 0.1–0.2% w/v. It is important to note that the preservative efficacy of these preservatives may be decreased in the presence of hydrophilic polymers (generally employed to enhance viscosity), due to an interaction of the preservative with the polymer. This effect is negated by increasing the overall preservative concentration. Other preservatives that are employed include benzoic acid (0.1–0.2%) or sodium benzoate (0.1–0.2%).

- *Flavours.* These are employed whenever the unpalatable taste of a therapeutic agent is apparent, even in the presence of the sweetening agents. The flavours may be of natural origin (e.g. peppermint, lemon, herbs and spices) and are available as oils, extracts, spirits or aqueous solutions. Alternatively, a wide range of synthetic flavours are available that offer advantages over their natural counterparts in terms of purity, availability, stability and solubility.

   Certain flavours are also associated with a (mild) therapeutic activity. For example, many antacids contain mint due to the carminative properties of this ingredient. Alternatively other flavours offer a taste-masking effect by eliciting a mild local anaesthetic effect on the taste receptors. Examples of flavours in this category include peppermint oil, chloroform and menthol.

   The concentration of flavour in oral syrups is that which is required to provide the required degree of taste-masking effectively.
- *Colours.* These are generally natural or synthetic water-soluble, photo-stable ingredients that are selected according to the flavour of the preparation. For example, mint-flavoured formulations are commonly a green colour, whereas in banana-flavoured solutions a yellow colour is commonly employed. Such ingredients must not chemically or physically interact with the other components of the formulation.

Oral elixirs

An elixir is a clear, hydroalcoholic solution that is formulated for oral use. The concentration of alcohol required in the elixir is unique to each formulation and is sufficient to ensure that all of the other components within the formulation remain in solution. For this purpose other polyol co-solvents may be incorporated into the formulation. The presence of alcohol in elixirs presents a possible problem in paediatric formulations and, indeed, for those adults who wish to avoid alcohol. The typical components of an elixir are as follows:

- *Purified water.*
- *Alcohol.* This is employed as a co-solvent to ensure solubility of all ingredients. As highlighted above, the concentration of alcohol varies depending on the formulation. Generally the concentration of alcohol is greater than 10% v/v; however, in some preparations, the concentration of alcohol may be greater than 40% v/v.
- *Polyol co-solvents.* Polyol co-solvents, e.g. propylene glycol, glycerol, may be employed in pharmaceutical elixirs to enhance the solubility of the therapeutic agent and associated excipients. The inclusion of these ingredients enables the

concentration of alcohol to be reduced. As before, the concentration of co-solvents employed is dependent on the concentration of alcohol present, the type of co-solvent used and the solubility of the other ingredients in the alcohol/co-solvent blend. The reader is directed to the pharmacopoeial monographs to observe the concentration of co-solvents in specific examples of the pharmaceutical elixirs. Two examples in the USP that illustrate the range of concentrations of co-solvents are Phenobarbital Elixir and Theophylline Elixir (Tables 1.1 and 1.2).

Table 1.1 Phenobarbital Elixir

Phenobarbital (therapeutic agent)	0.4% w/v
Orange oil (flavour)	0.025% v/v
Propylene glycol (co-solvent)	10% v/v
Alcohol	20% v/v
Sorbitol solution (sweetener)	60% v/v
Colour	As required
Purified water	ad 100%

Table 1.2 Theophylline Elixir

Theophylline (therapeutic agent)	0.53% w/v
Citric acid (pH regulation)	1.0% w/v
Liquid glucose (sweetening agent)	4.4% w/v
Syrup (sweetening agent)	13.2% v/v
Saccharin sodium (sweetening agent)	0.5% w/v
Glycerin (co-solvent)	5.0% v/v
Sorbitol solution (co-solvent)	32.4% v/v
Alcohol	20% v/v
Lemon oil (flavour)	0.01% w/v
FDC yellow no. 5 (colour)	0.01% w/v
Purified water	ad 100%

- *Sweetening agents.* The concentration of sucrose in elixirs is less than that in syrups and accordingly elixirs require the addition of sweetening agents. The types of sweetening agents used are similar to those used in syrups, namely syrup, sorbitol solution and artificial sweeteners such as saccharin sodium (Figure 1.8).

  It should be noted, however, that the high concentration of alcohol prohibits the incorporation of high concentrations of sucrose due to the limited solubility of this sweetening agent in the elixir vehicle. To obviate this problem, saccharin

Figure 1.8 Structural formula of saccharin sodium (dihydrate).
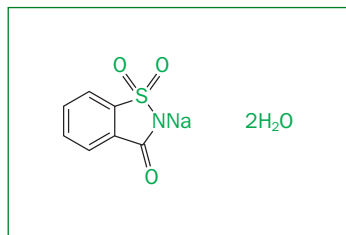
sodium, an agent which is used in small quantities and which exhibits the required solubility profile in the elixir, is employed.

- *Flavours and colours.* All pharmaceutical elixirs contain flavours and colours to increase the palatability and enhance the aesthetic qualities of the formulation. The presence of alcohol in the formulation allows the pharmaceutical scientist to use flavours and colours that may perhaps exhibit inappropriate solubility in aqueous solution. For example, it may be observed that in the two formulations cited above, essential oils were used as the flavouring agents. As before, the selected colour should optimally match the chosen flavour.
- *Ancillary comments*
- Preservatives are not required in pharmaceutical elixirs that contain greater than circa 12% v/v alcohol, due to the antimicrobial properties of this co-solvent.
- Due to the volatile nature of some of the components of elixirs, elixirs should be packaged in tight containers and not stored at high temperatures.
- The addition of viscosity-enhancing agents, e.g. hydrophilic polymers, may be required to optimise the rheological properties of elixirs.

Tips

The choice of liquid type for oral administration (solution, elixir or linctus) is often dependent on the physicochemical properties of the therapeutic agent. For example, if the drug has a bitter taste, linctuses are often used.

It should be noted that linctuses are now commonly formulated as sugar-free preparations.

The use of elixirs is not common.

Miscellaneous oral solutions

In addition to conventional solutions, syrups and elixirs, there are other solution-based dosage forms that are administered orally, in particular *linctuses* and *mouthwashes/ gargles*. These two subcategories are briefly described below.

Linctuses

Linctuses are viscous preparations that contain the therapeutic agent dissolved in a vehicle composed of a high percentage of sucrose and, if required, other sweetening agents. These formulations are administered

orally and are primarily employed for the treatment of cough, due to their soothing actions on the inflamed mucous membranes. Linctuses may also be formulated as sugar-free alternatives in which sucrose is replaced by sorbitol and the required concentration of sweetening agent.

Mouthwashes and gargles

Mouthwashes/gargles are designed for the treatment of infection and inflammation of the oral cavity. Formulations designed for this purpose employ water as the vehicle, although a co-solvent, e.g. alcohol, may be employed to solubilise the active agent. The use of alcohol as a co-solvent may act to enhance the antimicrobial properties of the therapeutic agent. Other formulation components are frequently required to enhance the palatability and acceptability of the preparation. These include preservatives, colours, flavouring agents and non-cariogenic sweetening agents.

Enemas

Enemas are pharmaceutical solutions that are administered rectally and are employed to ensure clearance of the bowel, usually by softening the faeces or by increasing the amount of water in the large bowel (osmotic laxatives). Enemas may be aqueous or oil-based solutions and, in some formulations, the vehicle is the agent that promotes bowel evacuation, e.g. arachis oil retention enema. Aqueous formulations usually contain salts (e.g. phosphates) to alter the osmolality within the rectum, thereby increasing the movement of fluid to the rectal contents. Viscosity-enhancing agents, e.g. glycerol, may be included to aid retention of the formulation within the rectum and to reduce the incidence of seepage.

Multiple choice questions

1. **Regarding weakly acidic drug molecules, which of the following statements are true?**
a. The solubility of weak acids increases as the pH is decreased.
b. The solubility of weak acids increases as the pH is increased.
c. The solubility of weak acids in pharmaceutical formulations may be affected by the presence of counterions.
d. All weakly acidic therapeutic agents exhibit an isoelectric point.

2. **Regarding weakly basic drug molecules, which of the following statements are true?**
a. The solubility of weak bases increases as the pH is decreased.

b. The solubility of weak bases increases as the pH is increased.
c. The solubility of weak bases in pharmaceutical formulations may be affected by the presence of counterions.
d. All weakly basic therapeutic agents exhibit an isoelectric point.

3. **Regarding the following drug substance, which of the following statements are true?**
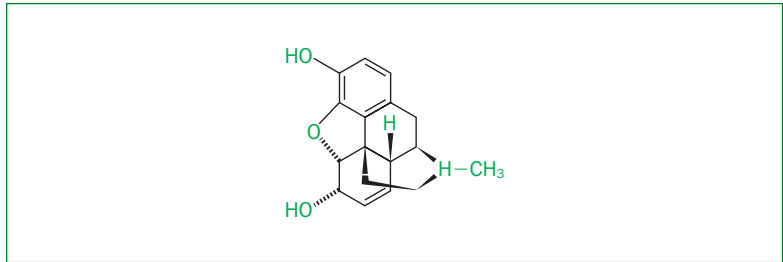
a. The solubility of the above drug increases as the pH is decreased from pH 9 to pH 4.
b. The solubility of the above drug increases as the pH is increased from 7 to 9.
c. The above drug may be susceptible to oxidation.
d. The above drug exhibits an isoelectric point.

4. **Regarding buffers for pharmaceutical solutions for oral administration, which of the following statements are true?**
a. Citrate buffer is commonly used as a buffer for pharmaceutical solutions.
b. Buffers are required solely to control the stability of therapeutic agents.
c. Buffer salts may affect the solubility of therapeutic agents.
d. The buffer capacity of a buffer system is increased as the concentration of buffer components is increased.

5. **Regarding the use of antioxidants in pharmaceutical solutions for oral administration, which of the following statements are true?**
a. Antioxidants are required in all solution formulations.
b. Antioxidants reduce the rate of oxidation of the therapeutic agent.
c. BHT and BHA are examples of antioxidants that are included in aqueous solutions.
d. The efficacy of antioxidants may be improved in the presence of ethylenediaminetetraacetic acid (EDTA).

6. **Regarding the use of co-solvents for the formulation of pharmaceutical solutions for oral administration, which of the following statements are true?**
a. Co-solvents are required in all pharmaceutical solution formulations.
b. Alcohols are commonly used as co-solvents in pharmaceutical solutions.
c. Glycerol may directly affect the pH of the formulation.
d. Co-solvents may affect the viscosity of the solution formulation.

7. **Regarding the use of preservatives in pharmaceutical solutions for oral administration, which of the following statements are true?**
a. Preservatives are required in all pharmaceutical solution formulations.
b. Preservatives are not required if the solution is manufactured under sterile conditions.
c. Esters of parahydroxybenzoic acid are used as preservatives for pharmaceutical solutions for oral administration.
d. Preservatives render pharmaceutical solutions for oral administration sterile.

8. **Regarding pharmaceutical elixirs, which of the following statements are true?**
a. Preservatives are required in all elixir formulations.
b. Elixirs generally require the addition of sweetening agents.
c. Elixirs generally contain < 10% alcohol USP.
d. Colours are required for all elixir formulations.

9. **Regarding pharmaceutical linctuses, which of the following statements are true?**
a. Preservatives are required in all linctus formulations.
b. Linctuses generally require the addition of synthetic sweetening agents.
c. Linctus formulations may contain high concentrations of sucrose.
d. Colours are required for all linctus formulations.

10. **Regarding oral syrups, which of the following statements are true?**
a. Preservatives are required in all oral syrups.
b. In certain syrups the concentration of sucrose may be ≤ 80% w/w.
c. Sugar-free syrups require the inclusion of a viscosity-modifying agent.
d. Colours are required for all oral syrups.

chapter 2

Pharmaceutical disperse systems 1: suspensions – general principles and the formulation of suspensions for oral administration

Overview

In this chapter the following points will be discussed:

- the physical stability of pharmaceutical suspensions
- the advantages and disadvantages of pharmaceutical suspensions
- formulation considerations for pharmaceutical suspensions
- considerations for the manufacture of pharmaceutical suspensions.

General description

Pharmaceutical suspensions are commonly referred to as dispersions in which the therapeutic agent is dispersed in the external phase (the vehicle). According to this definition the solubility of the therapeutic agent in the vehicle is low. The diameter of the disperse phase may range from circa 0.5 to 100 μm. Systems in which the particle size diameter falls below the above range are termed colloidal. In pharmaceutical dispersions (suspensions) the internal (drug) phase will separate upon storage; however, the main aim of the formulation scientist is to control the process of separation and, in so doing, optimise the stability of the formulation. A pharmaceutical suspension would be considered stable if, after agitation (shaking), the drug particles are homogeneously dispersed for a

KeyPoints

- Pharmaceutical suspensions are employed to deliver therapeutic agents of low (predominantly aqueous) solubility to the patient.
- Whereas pharmaceutical suspensions may be employed for the administration of drugs by many potential routes, this class of formulation is predominantly used for the delivery of drugs orally and parenterally (by injection) (see Chapter 5).
- Pharmaceutical suspensions are physically unstable; this instability is apparent by the presence of a solid cake and the resultant inability to redisperse the therapeutic agent. This leads to problems regarding the administration of the correct dosage of the therapeutic agent.
- In addition to enhancing the aesthetic properties, the excipients used in pharmaceutical suspensions are included to optimise the physical stability of the formulation.

sufficient time to ensure that an accurate dose is removed for administration to the patient.

The characteristics of an acceptable pharmaceutical suspension include the following:

- a low rate of sedimentation
- the disperse phase must be easily redispersed with gentle shaking
- the flow properties of the suspension should enable the formulation to be easily removed from the container (e.g. bottle, injection vial)
- aesthetically pleasing.

Advantages and disadvantages of pharmaceutical suspensions

Advantages

- Pharmaceutical suspensions are a useful drug delivery system for therapeutic agents that have a low solubility. Although low-solubility therapeutic agents may be solubilised and therefore administered as a solution, the volume of the solvent required to perform this may be large. In addition, formulations in which the drug has been solubilised using a co-solvent may exhibit precipitation issues upon storage.
- Pharmaceutical suspensions may be formulated to mask the taste of therapeutic agents.
- Pharmaceutical suspensions may be employed to administer drugs to patients who have difficulty swallowing solid-dosage forms.
- Pharmaceutical suspensions may be formulated to provide controlled drug delivery, e.g. as intramuscular injections (see Chapter 5).

Disadvantages

- Pharmaceutical suspensions are fundamentally unstable and therefore require formulation skill to ensure that the physical stability of the formulation is retained over the period of the shelf-life.
- The formulation of aesthetic suspension formulations is difficult.
- Suspension formulations may be bulky and therefore difficult for a patient to carry.

The physical stability of pharmaceutical suspensions

As detailed above, pharmaceutical suspensions are fundamentally unstable, leading to sedimentation, particle–particle interactions and, ultimately, caking (compaction). To gain an understanding of the physical stability of suspensions it is necessary to consider briefly two phenomena: the electrical properties of dispersed particles and the effect of distance of separation between particles on their subsequent interaction. It must be stressed that this is only a brief outline and the reader should consult the companion textbook in this series by David Attwood and Alexander T Florence (*FASTtrack: Physical Pharmacy*: London: Pharmaceutical Press; 2008) for a more comprehensive description of this topic.

Electrical properties of dispersed particles

Following dispersion within an aqueous medium, particles may acquire a charge due to either the ionisation of functional groups on the drug molecule and/or adsorption of ions to the surface of the particle. These are addressed independently below.

Ionisation of functional groups

Insoluble drug particles may possess groups at the surface that will ionise as a function of pH, e.g. COOH, NH_2. In this situation the degree of ionisation is dependent on the pKa of the molecule and the pH of the surrounding solution.

Adsorption of ions on to the surface of the particle

Following immersion in an aqueous solution containing electrolytes, ions may be adsorbed on to the surface of the particle. Furthermore, in the absence of added electrolytes, preferential adsorption of hydroxyl ions on to the surface of the particle will occur. Hydronium ions, by contrast, are more hydrated than hydroxyl ions and are therefore more likely to remain within the bulk medium. Following adsorption of ions on to the surface, a phenomenon referred to as the *electrical double layer* is established (Figure 2.1), the main features of which are as follows:

- Ions, e.g. cations, are adsorbed on to the surface of the particle, leaving the anions and remaining cations in solution. This generates a potential on the surface of the particle, termed the *Nearnst potential*. The ions responsible for this potential are termed *potential-determining ions*. Anions are then electrostatically attracted to the (positive) surface of the particle. The presence of these anions will repel the subsequent approach of further anions. This is referred to as

the first section of the double layer and is therefore composed of adsorbed ions on the surface, counterions and bound hydrated solvent molecules.

■ The boundary between the first and second layers of the electrical double layer is referred to as the *Stern plane*. The Stern plane is characterised by:

– the adsorbed ions on the surface of the particle
– the adsorbed counterions at the surface of the particle
– the Stern plane falls through the centre of the counterion layer
– normally the charge at the surface of the particle is greater than that at the Stern plane.

■ The second layer contains predominantly hydrated counterions that are loosely attracted to the surface of the particle, the features of which include:

– The boundary of this second layer will possess a potential, referred to as the *zeta potential.* The magnitude of this is generally less than that at the Stern plane.
– If the particle is rotated, this second layer forms the shear plane, i.e. the effective surface.

■ At a certain distance from the surface of the particle, electrical neutrality is restored.

Figure 2.1 Diagrammatic representation of the electrical double layer.
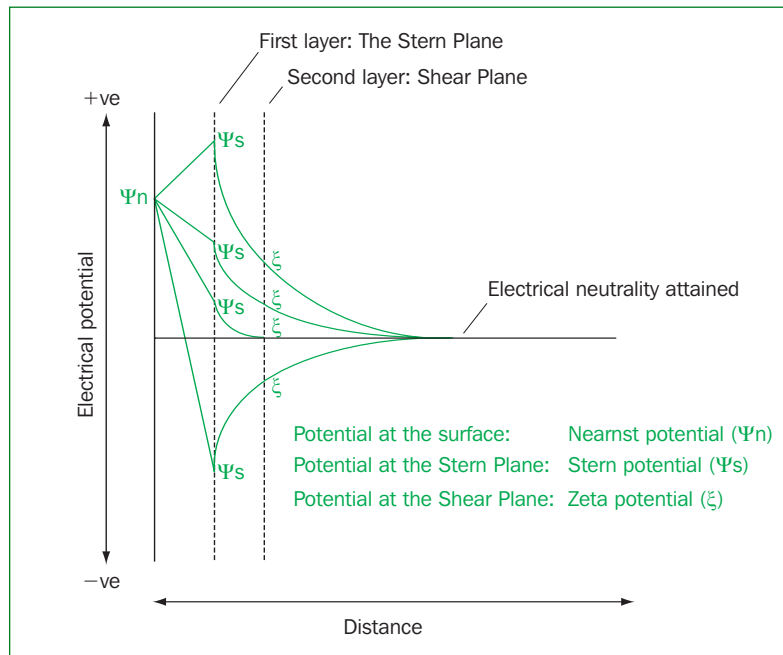
- A number of related alternative scenarios to that described above are possible:
- *Scenario 1.* Electrical neutrality may be achieved at the boundary of the second plane of the electrical double layer, i.e. the magnitude of the zeta potential is zero. In this the counterions are sufficiently present in this layer to neutralise the net positive charge on the surface of the particle.
- *Scenario 2.* If the number of counterions in the electrical double layer exceeds the adsorbed ions, the zeta potential will exhibit an opposite charge to that of the Nearnst potential. For example, if the Nearnst potential is positive, the zeta potential may be positive.
- *Scenario 3.* In certain circumstances molecules may interact with the charged particle surface via non-electrostatic mechanisms. For example, surface-active agents interact with surfaces via hydrophobic interactions. If the surface-active agents are charged, this will alter the Stern and zeta potentials. In this the magnitude of the Stern potential may be increased (i.e. exceeds the Nearnst potential) or may be totally reversed. There is a reduction in this charge at the shear plane (i.e. the zeta potential). Similarly, non-ionic surfactants may adsorb to the surface of the particle (again via hydrophobic interactions), thereby affecting both the Stern and zeta potentials.

The presence of electrolytes directly affects the above situations. As the concentration of electrolyte is increased there is a compression of the electrical double layer. The magnitude of the Stern potential is unaltered whereas the zeta potential decreases in magnitude. As the reader will discover, this approach may be used to stabilise pharmaceutical suspensions.

The relationship between distance of separation and the interaction between particles

The interaction between suspended particles in a liquid medium is related to the distance of separation between the particles. In principle, three states of interaction are possible:

1. No interaction, in which the particles are maintained sufficiently distant from one another. In the *absence* of sedimentation this is the thermodynamically stable state.
2. Coagulation (agglomeration), in which the particles form an intimate contact with each other. This results in the production of a pharmaceutically unacceptable formulation due to the inability to redisperse the particles upon shaking.
3. Loose aggregation (termed floccules), in which there is a loose reversible interaction between the particles, enabling the particles to be redispersed upon shaking.

The interaction (attraction/repulsion) between particles that have been dispersed in a liquid medium has been quantitatively described by Derjaguin, Landau, Verwey and Overbeek. In the simplest form the 'DLVO' theory assumed that, when dispersed in a liquid medium, particles will experience (electrical) repulsive forces and attractive (London/van der Waals) forces. The overall energy of interaction between the particles (V_t) can therefore be described as an addition of the energies of attraction (V_a) and repulsion (V_r), i.e. $V_t = V_a + V_r$.

■ The *energy of attraction* (V_a) is due to London/van der Waals forces and it is inversely proportional to the distance between particles. The attractive forces between particles tend to operate at greater distances than the repulsive forces.

■ The *energy of repulsion* (V_r) is due to the overlap of or interaction between the electrical double layers of each particle and operates over a distance of approximately the thickness of the double layer.

Figure 2.2 displays a diagrammatic representation of the relationship between the overall energy of interaction between two particles and their distance of separation. In this, three main regions may be observed:

Figure 2.2 Diagramatic representation of the overall interactive energy between two particles and their distance of separation.
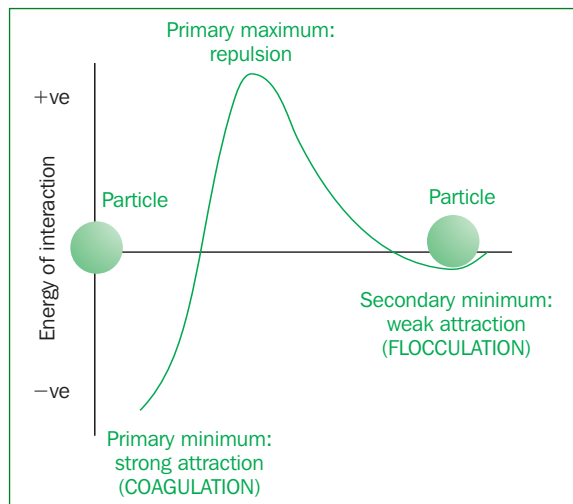
1. *The primary minimum.* This is a region of high attraction between particles. Particles that interact at distances corresponding to the primary minimum will irreversibly coagulate and the formulation so produced will be physically unstable.

2. *The primary maximum.* This region is responsible for the repulsion between particles, the magnitude of which is controlled by the zeta potential at the shear plane of the

particles. This region prevents the particles from interacting at close distances (the primary minimum).

The magnitude of the primary maximum is affected by the presence and concentration of electrolytes. As detailed previously, increasing the concentration of electrolyte decreases the thickness of the double layer, thereby reducing the zeta potential. This leads to a reduction in the magnitude of the primary maximum and increases the magnitude of the secondary maximum. This effect is also observed following the addition of ionic surface-active agents.

3. *The secondary minimum.* The secondary minimum is a region where attractive forces predominate; however, the magnitude of the attraction is less than that at the primary minimum. Particles located at the secondary minimum are termed floccules, this process being termed flocculation. This interaction increases the physical stability of the suspension by preventing the close approach to the primary minimum. Furthermore, the interaction between the particles may be temporarily broken by shaking, thereby enabling the removal of an accurate dose. The process by which particles are engineered to reside in the secondary minimum is referred to as *controlled flocculation.*

Sedimentation, controlled flocculation and the physical stability of suspensions

Thermodynamically a disperse system may be considered to be stable whenever there is no interaction between particles. However, in terms of pharmaceutical suspensions, this state is physically unstable. Particles in a suspension will sediment under the influence of gravity and settle at the bottom of the container, the larger particles reaching the bottom initially and the smaller particles occupying the space between the larger particles. The particles at the bottom of the container are gradually compressed by the weight of those above and, in so doing, sufficient energy is available to overcome the primary maximum (repulsive forces) and the particles become sufficiently close to form an irreversible interaction at the primary minimum. This is referred to as *caking*.

In flocculated systems the rate of sedimentation of the flocs is high and the volume of the sediment produced is large due to the large void volume within the floccule structure. Generally the size of the drug particles used in the formulation of suspensions is sufficiently large to exhibit a useful secondary minimum. In addition the irregular (generally non-spherical) nature of suspended drug particles will enhance this property. Finally, the ability to undergo flocculation will increase as the concentration of suspended particles increases, due to the greater probability of

particle–particle interactions. For particles in which the zeta potential (and hence the primary maximum) is high, manipulation of the magnitude of the secondary minimum is required (controlled flocculation). Generally this is performed by the addition of the required concentration of electrolyte or (ionic) surface-active agent. In performing controlled flocculation it must be remembered that if the reduction in the zeta potential (and hence the primary maximum) is too large, the resistance of particle–particle contact in the primary minimum is reduced. Therefore, irreversible coagulation of the particles may occur.

As caking in pharmaceutical suspensions is facilitated by sedimentation, it is of no surprise to note that controlling particle sedimentation may enhance the physical stability of pharmaceutical suspensions. The rate of sedimentation of particles (generally $< 2\%$ w/w) may be defined by Stokes' equation. Although many pharmaceutical suspensions are composed of more than 2% w/v solids in which the sedimentation of drug particles is influenced by other particles, the equation may be used to provide an indication of factors affecting sedimentation. The equation is as follows:

$$\frac{\delta v}{\delta t} = \frac{d^2}{18\eta_l} \frac{(\rho_s - \rho_t)g}{}$$

where: $\frac{\delta v}{\delta t}$ refers to the rate of sedimentation; d^2 refers to the average particle diameter; ρ_s and ρ_t refer to the densities of the solid particles and vehicle, respectively; η_l refers to the viscosity of the vehicle; and g refers to gravity.

Therefore, as may be observed from the above equation, the rate of sedimentation may be practically decreased by reducing the average particle diameter and increasing the viscosity of the vehicle. The former may be readily manipulated by milling the particles to the required size range whereas the latter may be increased by the inclusion of hydrophilic polymers. These points are addressed at a later stage.

Assessment of the sedimentation of drug suspensions is usually performed by measuring the *sedimentation volume* and/or the *degree of flocculation*.

■ *The sedimentation volume (F).* This is the ratio of the volume of the sediment (V_s) to the initial volume of the suspension (V_i):

$$F = \frac{V_s}{V_i}$$

The sedimentation volume may range from less than 1 to values that are greater than 1. The sedimentation volume of deflocculated suspensions is usually small, whereas the *F* value for flocculated systems is high (i.e. close to or greater than 1) due to the large volume occupied by the flocculated structure.

▪ *Degree of flocculation (β).* The degree of flocculation is defined as the ratio of the ultimate sedimentation volume of the flocculated suspension to the ultimate sedimentation volume of the deflocculated suspension. This is usually the preferred measurement as it provides a point of reference, i.e. the suspension before and after flocculation.

Formulation considerations for orally administered suspension formulations

The formulation of suspensions for oral administration requires consideration of both the physical properties of the therapeutic agent and the excipients required to ensure that the formulation is physically stable and suitable for administration to patients. General formulation considerations are as follows.

Physical properties of the therapeutic agent
Particle size
As detailed previously in this chapter, the physical stability of suspensions may be enhanced by modifying (i.e. retarding) the rate of particle sedimentation. According to Stokes' law, the rate of sedimentation $\left(\frac{\delta v}{\delta t}\right)$ is directly proportional to the square of the average diameter of the particles (termed d^2). Therefore, as the average particle size of suspended particles is increased, there is a dramatic effect on the resultant rate of sedimentation, i.e. increasing the particle diameter twofold results in a fourfold increase in the rate of sedimentation. Therefore, the average particle diameter of therapeutic agent used in the formulation of suspensions has major implications in the physical stability of the formulation. To optimise the stability of the formulation, the particle size should be minimised. This may be performed by either chemical (controlled precipitation) or physical methods (e.g. milling). It should be remembered that the energy required to reduce the average particle diameter to less than 10 μm is

Tips

Pharmaceutical suspensions are unstable systems that require the addition of excipients and knowledge of the particle size of the dispersed phase to ensure that a stable suspension may be formulated.

Generally the alternative choice to formulating suspensions is the production of solutions using solubilising agents (see Chapter 1). Therefore, the formulation scientist must carefully consider both options before making the final formulation choice.

Two key parameters that must be controlled are the electrolyte concentration (and type of ions) and the viscosity.

significant and therefore a compromise may have to be made concerning the actual particle size of therapeutic product used.

A phenomenon that may affect pharmaceutical suspensions and which influences the average particle size is *crystal growth* (sometimes referred to as *Ostwald ripening*). Small particles have a greater solubility (dissolution rate) than larger particles when dispersed in an aqueous vehicle. If there is a change (slight increase) in the storage temperature, this may enable the smaller particles to dissolve in the vehicle. Crystallisation of the dissolved drug may then occur on the surface of the larger particles, thereby increasing the average diameter of the suspended drug particles. This may therefore have implications regarding the physical stability of the suspension. One method that may be employed to reduce crystal growth is the inclusion of hydrophilic polymers within the formulation. These adsorb on to the suspended drug particles and offer a protective effect. In light of the potential problems associated with crystal growth, it is customary in formulation development to expose the suspension formulation to temperature cycling (e.g. repeated freeze–thaw cycles) and monitor changes in the average particle diameter and physical stability.

Wetting properties of the therapeutic agent

Insoluble drug particles are hydrophobic and therefore may not be easily wetted, i.e. the vehicle will not readily form a layer around the suspended drug particle. To wet fully with an aqueous vehicle the contact angle (θ), i.e. the angle at which the liquid/vapour interface meets the surface of the solid, must be low. The contact angle may be defined in terms of the interfacial tensions between the three phases, i.e. solid (drug)/vapour ($\gamma_{s/v}$), liquid (vehicle)/vapour ($\gamma_{l/v}$) and solid (drug)/liquid (vehicle) ($\gamma_{s/l}$) in the Young equation:

$$\gamma_{l/v} \cos \theta = \gamma_{s/v} - \gamma_{s/l}$$

Therefore, decreasing the interfacial tensions between the vehicle and the vapour and between the solid and the vehicle may reduce the contact angle. In practice this is achieved by the incorporation of surface-active agents into the formulation. These agents will decrease the interfacial tension by adsorbing at the vehicle/vapour interface and at the solid/liquid interface.

It is important to ensure insoluble therapeutic agents are sufficiently wetted, as this will ensure that the particles are homogeneously distributed in the formulation and thereby enable the correct dosage of drug to be removed when required by the patient. Drug particles, if poorly wetted, will tend to aggregate spontaneously in an attempt to stabilise the system

thermodynamically (i.e. lower the Gibb's free energy), thereby resulting in problems regarding the physical stability of the formulation.

Excipients used in the formulation of suspensions for oral administration

As the reader will observe, there is a direct similarity between the types of excipients used for the formulation of suspensions and solutions for oral administration. The major difference between these two categories of formulations is the inclusion of excipients to physically stabilise suspensions. Many of the categories (and examples) of excipients used for suspensions are the same for suspensions as for solutions.

Vehicle

As in oral solutions (and related formulations), the most commonly used vehicle for the formulation of pharmaceutical suspensions for oral administration is Purified Water USP. The preparation and specifications of this have been previously detailed in Chapter 1.

In addition to purified water, the vehicle may contain buffers to control the pH of the formulation. Citric acid/sodium citrate is commonly used as a buffer system for oral suspension formulations.

Excipients to enhance the physical stability of suspensions

As detailed previously, pharmaceutical suspensions may be stabilised by controlled flocculation and by the control of the rate of particle/floccule sedimentation. Further details of these approaches are shown below.

Addition of electrolytes

Electrolytes may be employed to control flocculation by reducing the zeta potential and hence the electrical repulsion that exists between particles. In so doing the magnitude of the secondary minimum increases, thereby facilitating the interaction of particles at a defined distance. Buffers are electrolytes and may be used for this purpose; however, other salts can also be used.

To ascertain the correct ionic strength (μ), a series of formulations containing different concentrations of electrolyte are prepared and the sedimentation volume or degree of flocculation determined. In flocculated systems the sedimentation volume and degree of flocculation are high. The addition of either insufficient or excess electrolyte will produce physically unstable suspensions that exhibit caking.

Surface-active agents

Surface-active agents may influence the stability of pharmaceutical suspensions in several ways, as detailed in the previous sections. These are briefly summarised below.

Effect on wetting

Surface-active agents decrease the contact angle of insoluble particles, enabling greater wetting by the vehicle. This, in turn, assists product homogeneity and decreases aggregation.

Effect on flocculation

Surface-active agents, both ionic and non-ionic, can interact with the suspended particles and, in so doing, can affect the magnitude of the zeta potential. This may lead to the lowering of the primary maximum and an increase in the size of the secondary maximum, thereby facilitating flocculation. The correct concentration of surfactant required to stabilise a suspension may be experimentally obtained in a fashion similar to that described above for electrolytes.

For oral suspensions non-ionic surfactants are preferred, e.g. polyoxyethylene fatty acid sorbitan esters, sorbitan esters or lecithin, further details of which are provided in Chapter 3. The greater toxicity of ionic surfactants precludes their use in oral suspension formulations. The concentrations of surfactants required to stabilise pharmaceutical suspensions are dependent on the physical properties of the dispersed particles (e.g. zeta potential); however, concentrations less than 0.5% w/v are generally employed in the formulation of oral suspensions.

Hydrophilic polymers

Hydrophilic polymers are commonly used to enhance the physical stability and to affect the flow properties of oral suspensions. These two aspects are detailed below.

Effects on the physical stability of suspensions

Hydrophilic polymers may adsorb on to the surface of suspended drug particles in pharmaceutical suspensions. Due to their large molecular weight, one section of the polymer chain will adsorb on to the particles, leaving the remainder of the chain to extend into the aqueous vehicle. As the concentration of polymer in the formulation increases, the thickness of the adsorbed layer of polymer increases. As two (polymer-coated) particles approach each other, there will be stearic repulsion due to an overlap of the adsorbed polymer chains. This will prevent the particles coming into close contact (at the primary minimum). It is important to note that the ability of hydrophilic polymers to stabilise

suspensions stearically is dependent on several features, e.g. (1) the concentration of polymer; and (2) the type of polymer.

The concentration of polymer

The concentration of polymer affects the density of the adsorbed polymer layer on the surface of the particles. The required concentration of polymer should be that which enhances repulsion but does not prevent the interaction of the particles in the secondary minimum (flocculation). Generally flocculation occurs at a distance which is approximately twice the thickness of the adsorbed polymer layer.

The type of polymer

The type (and hence the chemistry) of polymer influences the stabilisation properties of hydrophilic polymers in two ways. Firstly, the chemical structure of the polymer will influence the nature of the adsorption on the surface of the drug particles which, in turn, influences the thickness and integrity of the adsorbed layer. Secondly, as the interaction between specific groups on adjacent polymer chains is responsible for the stearic stabilisation, the nature of the interacting groups on each chain is important. This ability to interact may effectively maintain the polymer-coated particles at a distance, resulting in the production of a structured floccule.

   Finally if ionic (e.g. anionic) polymers are added to the formulation in the presence of divalent ions (e.g. Mg^{2+}, Al^{3+}), the ions may act as a bridge by interacting both with the surface of the particle and with the charged moieties on the polymer. In this way the polymer is 'connected' to two particles and prevents the interaction of the particles at the primary minimum.

   Secondarily, the addition of hydrophilic polymers to the aqueous vehicle will increase the viscosity of the formulation. According to Stokes' equation (and remembering the limitations of the equation!), increasing the viscosity of an aqueous vehicle will reduce the rate of sedimentation, thereby increasing the physical stability of the formulation.

Effect on the rheological properties of oral suspension

As detailed in the previous paragraph, increasing the concentration of a hydrophilic polymer within an aqueous vehicle will alter the viscosity of the system. At very low polymer concentrations (frequently <0.01% for branched polymers), aqueous vehicles will behave as Newtonian systems, in which the shearing stress and rate of shear are proportional. However at higher polymer concentrations, typically of those used in oral suspensions, the flow properties are *pseudoplastic* (shear thinning). This is a useful property for suspensions as the

apparent viscosity (the reciprocal of the tangent of the flow curve) will be high under conditions of low shear stress (e.g. storage in the bottle) but under high shearing stress (e.g. those exerted whenever the contents of the bottle are shaken) the apparent viscosity will be low, thereby facilitating administration to the patient. In addition, pseudoplastic formulations may also exhibit *thixotropy*, a time-dependent recovery of the flow properties (illustrated in Figure 2.3 as the solid arrow). It is important to understand this property in particular, as this should ideally be minimised to enable the rapid recovery of the rheological properties of the formulation.

Figure 2.3 Ideal flow curves illustrating newtonian flow (dashed line) and pseudoplastic flow (with thixotropy). In the pseudoplastic flow curve, the direction of flow is indicated by arrows.
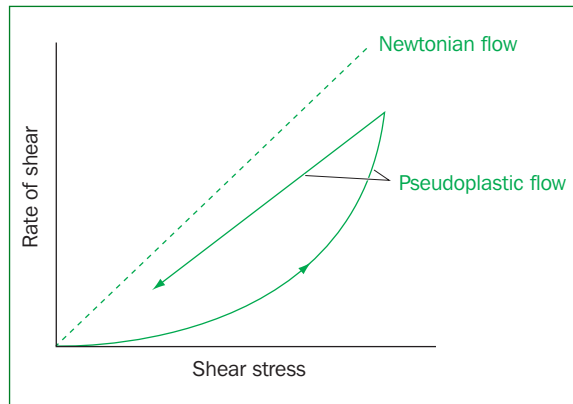
- Examples of polymers that are employed in oral suspensions to enhance the physical stability and to modify the flow (rheological) properties include:
  - Cellulose derivatives: these are branched polymers and therefore only low concentrations are required (<3% w/w). Specific examples that are commonly used include:
    - methylcellulose
    - hydroxyethylcellulose
    - hydroxypropylmethylcellulose
    - sodium carboxymethylcellulose (ionic).
  - Polyvinylpyrrolidone: this is a linear polymer and will therefore require a larger concentration to enhance the rheological properties of the formulation.
  - Sodium alginate (ionic).
  - Acacia, tragacanth and xanthan gum: these polymers are natural polysaccharide-based polymers that may show batch-to-batch variability in their effects on formulation viscosity/rheology. Only low concentrations of these are required to enhance the viscosity of the formulation.

- The concentration of each of these polymers will depend on the chemistry of the polymer, the molecular weight and, for ionic polymers, the pH of the formulation.
- Rheological structuring of oral suspensions may also be achieved using the hydrated silicates, e.g. magnesium aluminium silicate. These materials swell in water, resulting in a vehicle that exhibits plastic flow (with thixotropy). The concentration of magnesium aluminium silicate that is typically used is circa 5%. Caution should be issued when formulated in the presence of cationic compounds as this may induce flocculation of the dispersed, swollen silicates. Due to this susceptibility, hydrated silicates may be formulated along with hydrophilic polymers to produce formulations with the required consistency.

Preservatives

Oral suspensions are *non-sterile*; however there are restrictions on the number and type of microorganisms present in this type of dosage form. Whilst the presence of microorganisms within oral suspensions is allowed, it is essential that highly pathogenic microorganisms, e.g. *Escherichia coli,* are absent. Oral suspensions are multidose formulations and therefore inhibition of the growth/elimination of less pathogenic bacteria and fungi (the latter primarily causing spoilage) is required. Specifications are defined in the various pharmacopoeias regarding the number and type of microorganisms in oral products (solutions and suspensions). For example, the European Pharmacopoeia states that in oral products *E. coli* must be absent and, in addition, there should be not more than 1000 aerobic bacteria and not more than 100 fungi per gram or millilitre.

Examples of preservatives that are employed in oral suspensions include:

- Parabens (i.e. esters of parahydroxybenzoic acid, e.g. methyl and propyl parahydroxybenzoic acid are often used in combination in a ratio of 9:1 or 10:1. The concentration is usually 0.22% w/v (i.e. 0.2% w/v methylhydroxy benzoate and 0.02% w/v propylhydroxy benzoate).
- Organic acids, e.g. benzoic acid (circa 0.9% w/v).

When selecting the type and concentration of preservative for inclusion in an oral suspension formulation, the following points should be considered:

- To exert the antimicrobial effect, preservative must exist in solution within the formulation. Certain preservatives, e.g. the parahydroxybenzoate esters, will interact with hydrophilic polymers, e.g. polyvinylpyrrolidone, cellulose ethers and

surface-active agents (e.g. polysorbate 80), thereby reducing the concentration of 'free' preservative in the formulation. As a result the antimicrobial efficacy of the preservative is decreased. To overcome this problem it is customary to increase the initial concentration of preservative to ensure that, after adsorption to the dissolved polymer, the required free concentration of preservative is available.

- The selected preservative must not adversely affect the chemical and physical stability of the suspension product.

Sweetening agents/flavours

As is the case for oral solutions, sweetening agents (e.g. sucrose, liquid glucose, glycerol, sorbitol, saccharin sodium and aspartame) and flavours are used for taste-masking purposes. The type (particularly in the case of flavours) and the concentrations of these are selected, as before, to provide the necessary aesthetic properties. The details of these have been provided in Chapter 1.

Antioxidants

Antioxidants are required in certain pharmaceutical suspensions for oral usage to enhance the chemical stability of the therapeutic agent, where this may be compromised by oxidation. As before (Chapter 1), the chosen antioxidants are compounds that are redox systems which exhibit higher oxidative potential than the therapeutic agent or, alternatively, are compounds that inhibit free radical-induced drug decomposition. Commonly used examples for incorporation into oral suspensions (usually at concentrations less than 0.1% w/w) include:

- sodium sulphite
- sodium metabisulphite
- sodium formaldehyde sulphoxylate
- ascorbic acid.

As for solutions, chelating agents, e.g. ethylenediamine tetraacetic acid, citric acid, that form complexes with heavy metal ions which are normally involved in oxidative degradation of therapeutic agents, may be included.

Manufacture of suspensions for oral administration

Suspensions for oral administration are usually manufactured in one of two methods: (1) direct incorporation; and (2) precipitation method.

Tips

The formulation of suspension for oral administration will require the inclusion of several excipients.

All excipients must be physically and chemically compatible (with each other and with the therapeutic agent).

Possible interaction between components must be understood, e.g. preservatives and viscosity-enhancing agents.

Direct incorporation

- In this method the soluble components are normally dissolved in the appropriate volume of diluent (vehicle).
- The solid therapeutic agent is then dispersed into the vehicle with the aid of mixing, prior to correction for volume.
- The mixing rate employed during the addition is an important determinant in the manufacture of the formulation. If the suspension is flocculated, high-speed mixing may be employed as the flow properties of the system are pseudoplastic (shear thinning; Figure 2.3). However, if the formulation has been poorly designed and has poor flocculation properties, high-speed mixing will result in an increase in the viscosity of the product (termed *dilatant flow*). Ultimately this leads to issues regarding the quality of mixing as the increased viscosity may render the product difficult to mix homogeneously.
- The particle size of the suspended drug within the formulation may then be reduced using a ball mill. Alternatively, the particle size of the active ingredient may be optimised (by particle size reduction techniques) prior to incorporation into the vehicle.

Precipitation method

- In this method the drug is dissolved in the vehicle (or a portion of the available volume), prior to precipitation following the addition of a counterion; the salt formed is insoluble.
- Such systems are frequently deflocculated and are therefore mixed at low shear rates.
- The excipients are then dissolved in the vehicle, or dissolved in a portion of the vehicle, which is then added to the suspension of drug.
- At this stage the formulation may be exposed to high shearing rates to ensure homogeneity.
- The volume of the formulation is then corrected by adding the required mass of diluent.
- One potential problem with this technique is the production of ionic byproducts from the precipitation interaction. If the concentration of these is too high, then the precipitated therapeutic agent requires to be washed with an aqueous solvent.

Multiple choice questions

1. **Regarding the stability of pharmaceutical suspensions designed for oral administration, which of the following statements are true?**
   a. Suspensions are inherently pharmaceutically unstable.
   b. The stability of pharmaceutical suspensions is affected by the particle size of the dispersed drug.
   c. The stability of pharmaceutical suspensions is affected by the concentration of buffer salts used.
   d. The particle size range of dispersed solids affects the stability of pharmaceutical suspensions.

2. **Regarding the rate of sedimentation of pharmaceutical suspensions designed for oral administration, which of the following statements are true?**
   a. The rate of sedimentation is increased as the diameter of the dispersed drug particles is increased.
   b. The rate of sedimentation is increased as the viscosity of the continuous phase is increased.
   c. The rate of sedimentation is affected by the concentration of buffer salts.
   d. The rate of sedimentation may be increased by centrifugation.

3. **Regarding the electrical double layer, which of the following statements are true?**
   a. The zeta potential is principally due to ionisation of the drug particle.
   b. The zeta potential for insoluble basic drugs is always positive.
   c. Manipulation of the zeta potential may be used to enhance the physical stability of suspensions.
   d. Increasing the concentration of added electrolyte enhances the thickness of the electrical double layer.

4. **Regarding the DLVO theory, which of the following statements are true?**
   a. The zeta potential acts as a repulsion barrier.
   b. Particles residing within the primary minimum produce pharmaceutically acceptable suspensions.
   c. Alteration of the magnitude of the secondary minimum may be performed by increasing the concentration of electrolyte.
   d. Increasing the concentration of hydrophilic polymer in a suspension increases the stability of the suspension by increasing the magnitude of the primary maximum.

5. **Regarding the role of surfactants in pharmaceutical suspensions for oral administration, which of the following statements are true?**
a. Surfactants decrease the water contact angle of dispersed drug particles.
b. Surfactants promote flocculation.
c. Surfactants with low HLB are used to stabilise oral suspensions designed for oral administrations.
d. Surfactants increase the viscosity of the continuous phase of pharmaceutical suspensions.

6. **Regarding the use of hydrophilic polymers for the stabilisation of pharmaceutical suspensions for oral administration, which of the following statements are true?**
a. Hydrophilic polymers stabilise pharmaceutical suspensions by increasing the viscosity of the continuous phase and hence promoting the sedimentation of dispersed drug particles.
b. Hydrophilic polymers may affect the zeta potential of the dispersed drug.
c. The rheological properties of pharmaceutical suspensions containing hydrophilic polymers and 1–5% w/w dispersed drug may be described as dilatant.
d. Pharmaceutical suspensions may exhibit thixotropy.

7. **Flocculated suspensions exhibit the following properties:**
a. A low sedimentation volume.
b. A high degree of flocculation.
c. A high rate of sedimentation.
d. Homogeneity of drug concentration per unit dose.

8. **Concerning the manufacture of pharmaceutical suspensions designed for oral administration, which of the following statements are true?**
a. Suspensions for oral administration are prepared under aseptic conditions.
b. Suspensions for oral administration are frequently sterilised following manufacture.
c. High-speed mixing of concentrated suspensions may result in dilatant flow.
d. The particle size distribution of the dispersed drug may be reduced postmanufacture using a ball mill.

9. **Concerning the use of pharmaceutical suspensions designed for oral administration, which of the following statements are true?**
a. Suspensions for oral administration are primarily used for administration to children or the elderly.

b. Many antacid formulations are suspensions.

c. Drugs with high aqueous solubility are frequently formulated as suspensions designed for oral administration.

d. Pharmaceutical suspensions designed for oral administration must be coloured.

10. Concerning pharmaceutical suspensions for oral administration, which of the following statements are true?

a. Suspensions for oral administration may require the addition of sweetening agents.

b. Suspensions for oral administration may require the addition of flavours.

c. The pH of the continuous phase may affect the stability of aqueous suspensions designed for oral administration.

d. The particle size of dispersed drug in pharmaceutical suspensions designed for oral administration must be <10 μm.

chapter 3
Pharmaceutical disperse systems 2: emulsions and creams

Overview

In this chapter the following points will be discussed:

- the physical properties of disperse systems in which an insoluble liquid is dispersed in a second liquid phase (generically termed emulsions)
- types of emulsions and creams
- factors affecting the stability and methods that may be used to stabilise pharmaceutical (liquid) disperse systems (emulsions and creams)
- formulation strategies for emulsions and creams
- the advantages and disadvantages and uses of emulsions and creams
- considerations for the manufacture of pharmaceutical emulsions and creams.

General description

Pharmaceutical emulsions/creams are commonly used pharmaceutical products that are primarily prescribed for the treatment of external disorders. In addition to this use emulsions are clinically used for total parenteral nutrition (see Chapter 5), for the oral administration of therapeutic agents and for the rectal administration of antiepileptic agents. The terms emulsions and creams refer to disperse systems in which one insoluble phase is dispersed as droplets within a second liquid phase. The rheological properties (and hence the structure of the network within the formulation) of the two systems differ considerably. Creams are pseudoplastic systems with a greater consistency than, for example, oral or parenteral emulsions.

There are two principal types of emulsion/cream, termed oil in water (o/w) and water in oil (w/o). In the former system the oil (or internal) phase is dispersed as

KeyPoints

- Emulsions and creams are disperse systems in which an insoluble liquid phase is dispersed within a second liquid phase. Creams are emulsions that offer greater consistency (viscosity) and are applied topically.
- Emulsions and creams are termed either oil in water (o/w), in which oil is the disperse phase and water the external phase, or water in oil (w/o), in which water is the disperse phase and oil is the external phase.
- The major use of emulsions is as cream formulations (for external application); however, emulsions may also be administered intravenously (see Chapter 5), rectally or orally.
- Emulsions/creams are physically unstable: the various excipients in the formulation are present primarily to stabilise the physical properties of the system.

droplets through the external aqueous phase. Conversely, in w/o emulsions, the internal phase is composed of water droplets and the external phase is non-aqueous. In addition to the emulsion types described above there are further more structurally complex types, termed *multiple emulsions*. These are termed water in oil in water (w/o/w) and oil in water in oil (o/w/o) emulsions. However, the pharmaceutical uses of these are extremely limited due to their possible reversion to the parent primary emulsion. For example, an o/w/o emulsion may revert to a w/o emulsion. As the reader will observe later in this chapter, the nature of the excipients and the volume ratio of the two phases used in the formulation of these systems determine both the type and consistency of the emulsion.

Emulsions and creams, akin to pharmaceutical suspensions, are fundamentally unstable systems, which, in the absence of *emulsifying agents*, will separate into the two separate phases. The emulsifying agents used are principally surface-active agents. O/w emulsions may be administered topically or orally whereas the use of w/o creams is principally (but not exclusively) limited to formulations designed for topical application.

The characteristics of an acceptable pharmaceutical suspension include the following:

- Physical stability (no phase separation).
- The flow properties of the emulsion/cream should enable the formulation to be easily removed from the container. Furthermore, if the formulation is designed for external application to, for example, the skin, the formulation must be easily spread over the affected area.
- The formulation must be aesthetically and texturally pleasing. If the emulsion is designed for oral application, the flavour must be suitable whereas if emulsions are to be externally applied, they must have the correct 'feel' (termed texture).

Tips

Emulsions are physically unstable systems and indeed are more unstable than suspensions

The type of emulsion dictates the final use of the formulation

Oil in water emulsions (but not water in oil emulsions) may be administered orally

Both oil in water and water in oil creams are administered topically.

Advantages and disadvantages of pharmaceutical emulsions

Advantages

- Pharmaceutical emulsions may be used to deliver drugs that exhibit a low aqueous solubility. For example, in o/w emulsions the therapeutic agent is dissolved in the internal oil phase. Following oral administration the oil droplets (and hence the drug) may then be absorbed using the normal absorption mechanism for oils. Some drugs are more readily

absorbed when administered as an emulsion than as other oral comparator formulations.

■ Pharmaceutical emulsions may be used to mask the taste of therapeutic agents, in which the drug is dissolved in the internal phase of an o/w emulsion. The external phase may then be formulated to contain the appropriate sweetening and flavouring agents.

■ Emulsions may be commonly used to administer oils that may have a therapeutic effect. For example, the cathartic effect of oils, e.g. liquid paraffin, is enhanced following administration to the patient as droplets within an o/w emulsion. The taste of the oil may be masked using sweetening and flavouring agents.

■ If the therapeutic agent is irritant when applied topically, the irritancy may be reduced by formulation of the drug within the internal phase of an o/w emulsion.

■ Pharmaceutical emulsions may be employed to administer drugs to patients who have difficulty swallowing solid-dosage forms.

■ Emulsions are employed for total parenteral nutrition.

Disadvantages

■ Pharmaceutical emulsions are thermodynamically unstable and therefore must be formulated to stabilise the emulsion from separation of the two phases. This is by no means straightforward.

■ Pharmaceutical emulsions may be difficult to manufacture.

Emulsion instability and theories of emulsification

Emulsion instability and the role of surface-active agents

Emulsions are termed thermodynamically unstable systems. Following dispersion of an insoluble liquid, e.g. an oil into an aqueous phase, the oil phase will adopt a spherical (droplet) shape as this is the shape associated with the minimum surface area per unit volume. If the droplet contacts a second droplet, coalescence will occur to produce a single droplet of greater diameter and, in so doing, the surface area of the new droplet will be less than the surface areas of the two individual droplets prior to coalescence. This process will continue until there is complete phase separation, i.e. two liquid layers occur. An interfacial tension exists at the interface between the two phases due to the imbalance of forces at the interface. For example, at the interface between the two layers, there will be a net attractive force that is directed towards the bulk of each phase, due to the imbalance between the cohesive forces (oil–oil and water–water) within each

phase and the oil–water attractive forces at the interface. The interfacial tension therefore acts both to stabilise the system into two phases and to resist the dispersion of one phase as droplets within the other phase.

Thermodynamically, this situation may be described in terms of the change in the interfacial Gibb's free energy (ΔG), interfacial tension ($\gamma_{o/w}$) between the two phases and the change in surface area of the disperse phase when this is dispersed, albeit temporarily, as droplets within the external phase (ΔA) as follows:

$$\Delta G = \gamma_{o/w} \Delta A$$

The dispersion of one phase within the other will cause a dramatic increase in the surface area of the interface between the two phases which, in turn, renders the system unstable (due to the increase in the interfacial Gibb's free energy). The system will therefore attempt to correct this instability; the subsequent coalescence of the droplets reduces the surface area of the interface, thereby reducing ΔG. In this fashion the spontaneous coalescence of droplets of the internal phase may be explained. Accepting that a fundamental requirement for the formulation of pharmaceutical emulsions is the dispersal of one internal phase within a second external phase, this relationship provides an insight into one of the mechanisms of stabilisation of emulsions by emulsifying agents. As the reader will be aware, surface-active agents lower the interfacial tension and therefore, when present in emulsion systems, will partially negate the destabilising effects of the increase in surface area of the disperse phase. It is important to note that this is not the only mode of emulsification of these agents.

Classical studies on the stabilisation of emulsions have shown that the stability of the adsorbed layer was of primary importance. In these studies it was shown that whenever sodium cetyl sulphate (a hydrophilic surface-active agent) and cholesterol (a lipophilic surface-active agent) were employed as emulsifying agents, the two agents formed a stable film due to their interaction at the interface. The mechanical properties of this mixed surfactant film were sufficient to prevent disruption even when the shape of the droplets changed. Furthermore, the close-packed nature of the surface-active agents at the interface resulted in a greater lowering of the interfacial tension that could be achieved by either component when employed as a single emulsifying agent. The role of the interaction between surface-active agents (resulting in a mechanically robust interfacial film) was highlighted by replacing cholesterol with oleyl alcohol (a *cis* isomer), which resulted in a poor emulsion; however, the use of the *trans* isomer of oleyl alcohol, elaidyl alcohol, produced a

stable emulsion. Further studies have shown that interfacial surfactant films form three-dimensional liquid crystalline layers of defined mechanical structure.

In addition to the mechanical properties of the adsorbed interfacial (liquid crystal) film, the adsorbed layer may carry a charge which, depending on the magnitude, may offer electrical repulsion between adjacent droplets. This is frequently observed whenever the droplets have been stabilised using ionic surface-active agents. Interestingly, flocculation of droplets of the disperse phase may lead to physical instability (see later) and, therefore, controlled flocculation (see Chapter 2) is not performed.

Emulsion instability and the role of hydrophilic polymers

Hydrophilic polymers are frequently used as emulsion stabilisers in pharmaceutical emulsions. In contrast to surface-active agents, hydrophilic polymers do not exhibit marked effects on the interfacial tension. However, the stabilisation effect of these materials is due to their ability to adsorb at the interface between the disperse phase and the external phase to produce *multilayers* that are highly viscoelastic (gel-like) and can therefore withstand applied stresses without appreciable deformation. In so doing these polymers mechanically prevent coalescence. It should be noted that surface-active agents produce *monomolecular* not multimolecular films.

If the chosen hydrophilic polymer is ionic (e.g. gelatin, sodium alginate, sodium carboxymethylcellulose), then the multimolecular adsorbed film will be charged and therefore will exhibit a zeta potential. This may further protect the emulsion droplets from coalescence by offering an electrical repulsion, as described in the previous section (and in Chapter 2). Furthermore, it would be expected that stearic stabilisation of the droplets would occur due to the presence of the adsorbed polymeric layer. A similar phenomenon was described for suspensions in Chapter 2.

In addition, hydrophilic polymers will increase the viscosity of the external phase in an o/w emulsion and, in a similar fashion to suspensions, will affect the sedimentation rate of the droplets. This point is addressed in subsequent sections.

Emulsion instability and adsorbed particles

Emulsions may also be stabilised by the addition of finely divided solid particles, if the particles are sufficiently wetted by both the oil and water phases (but preferentially wetted by one of the phases). The particles will accumulate at the interface between the phases and, if the particles show high interparticulate adhesion (thereby ensuring mechanical robustness to the adsorbed layer), the stability of the emulsion will be greatly

enhanced. The type of emulsion produced by this method depends on the preference of the particles for each phase. For example, if the particles are wetted preferentially by the aqueous phase (i.e. the contact angle between the particle and water is less than 90°), an o/w emulsion will result. Conversely, if the finely divided solid is preferentially wetted by the oil phase, the resulting emulsion will be a w/o emulsion. Examples of finely divided solids that are employed in the formulation of o/w and w/o pharmaceutical emulsions are:

- o/w emulsions
  - aluminium hydroxide
  - magnesium hydroxide
  - bentonite
  - kaolin
- w/o emulsions
  - talc
  - carbon black.

Type of emulsion

In the preparation of an emulsion, oil, water and the specified emulsifying agents are mixed together, resulting in the formation of droplets of each phase. At this stage theoretically either an o/w or a w/o emulsion may form. The resultant emulsion type is defined by the stability of the droplet phase; the phase of lower stability (i.e. the greater rate of coalescence) coalesces to form the external (or continuous) phase. There are several determinants of the type of emulsion produced, including: (1) phase volume of the internal phase; (2) the chemical properties of the film surrounding the internal phase; and (3) viscosity of the internal and external phases.

Phase volume of the internal phase

Assuming that the internal phase is composed of spheres, it may be calculated that the maximum volume that may be occupied by the internal phase is 74%. This is termed the *critical value* and is dependent on the droplet size range and shape. Moreover, a large particle size range and irregular droplet shape may increase this value. In practice it is customary to use a phase volume ratio of 50% as this results in a stable emulsion (due to the loose packing of the internal phase). It should be remembered that the higher the phase volume of the internal phase, the greater the probability of droplet coalescence.

Interestingly, although the above description holds true for o/w emulsions, the critical value for w/o emulsions is markedly lower (circa 40%). This is due to the greater mechanical properties of hydrophilic polymer or polar surface-active agents (used to form o/w emulsions) than the hydrophobic groups that

stabilise w/o emulsions. This point is extended in the next
section.

The chemical properties of the film surrounding the internal phase

As the reader will now appreciate, the adsorption of a
mechanically robust film around the droplets of the internal
phase is important to prevent droplet coalescence. The chemical
composition of the surface-active agents (and hydrophilic
polymers) at the droplet/external phase interface will dictate
whether an o/w or w/o is formed. Typically oil droplets are
stabilised by an adsorbed film composed of non-ionic, and
especially ionic, surfactants or alternatively hydrated hydrophilic
polymer chains. The surface-active agents and polymers that are
responsible for this stabilisation are therefore predominantly (but
not exclusively; the reader should recall that surface-active agents
also possess hydrophobic groups) aqueous-soluble. Conversely, in
w/o emulsions, the droplet is stabilised by the non-polar portion
of the surface-active agent, which protrudes into the non-aqueous
external phase. Furthermore, the length of this non-polar section
plays an important role in the stabilisation of w/o emulsions,
enhancing the mechanical integrity and reducing the tendency for
the internal phase to coalesce.

The reader will therefore appreciate from this description
that the solubility characteristics of the emulsifying agent define
the type of emulsion that is formed. Therefore polymers and
surface-active agents that are predominantly hydrophilic will
form o/w emulsions, whereas predominantly hydrophobic
surfactants will form w/o emulsions. Surface-active agents
contain both hydrophilic and lipophilic groups and therefore it is
the relative contributions of these that determine whether the
agent is predominantly hydrophilic or lipophilic (hydrophobic).
The contribution of these to the overall solubility is commonly
referred to as the *hydrophile–lipophile balance* (HLB), a ratio
scale that assigns a number to a surface-active agent, based on the
contributions of the individual groupings on the molecule. This
number can then be used when selecting surface-active agents for
the formulation of either o/w or w/o emulsions.

The main features of the HLB scale are as follows:

- The HLB scale runs from circa 1 to 40; the water solubility
  of the surface-active agent increases as the HLB increases.
- Surface-active agents exhibiting an HLB between circa 3 and 6
  are used to produce w/o emulsions and are therefore termed
  w/o emulsifying agents. These agents form poor dispersions
  in water but are soluble in the oil phase. Examples include:
  - sorbitan sesquioleate (e.g. Arlacel 83): HLB 3.7
  - sorbitan monooleate (e.g. Span 80): HLB 4.3

- – sorbitan monostearate (e.g. Span 60): HLB 4.7
- – glyceryl monostearate: HLB 3.8.
- ▪ Surface-active agents that exhibit an HLB between circa 6 and 9 form non-stable milky dispersions in water. Examples include:
- – orbitan monopalmitate (e.g. Span 40): HLB 6.7
- – sorbitan monolaurate (e.g. Span 20): HLB 8.6.
- ▪ Surface-active agents exhibiting an HLB between circa 9 and 16 are used to produce o/w emulsions (termed o/w emulsifying agents). These agents form stable milky dispersions in water (HLB 9–10.5), translucent/clear dispersions in water (HLB 10.5–13) or clear solutions (HLB 13–16). Examples include:
- – polyoxyethylene sorbitan tristearate (e.g. Tween 65): HLB 10.5
- – polyoxyethylene sorbitan trioleate (e.g. Tween 85): HLB 11.0
- – polyoxyethylene sorbitan monostearate (e.g. Tween 60): HLB 10.5
- – polyoxyethylene sorbitan monooleate (e.g. Tween 80): HLB 15.0
- – polyoxyethylene sorbitan monopalmitate (e.g. Tween 40): HLB 15.6
- – polyoxyethylene sorbitan monolaurate (e.g. Tween 20): HLB 16.7.
- ▪ The HLB value of ionic surface-active agents is frequently greater than 16.

Viscosity of the internal and external phases

The type of emulsion produced is affected by the viscosity of both the internal and external phases. If the viscosity is high the diffusion of the surface-active agent to the droplet surface will be reduced, as viscosity is inversely proportional to the diffusion coefficient of the surface-active agents. Furthermore, the increased viscosity will affect the process of coalescence of the droplets of the external phase. In general, if the viscosity of one phase is preferentially increased, there is a greater chance of that phase being the external phase of the emulsion.

Tests to identify the type of emulsion

There are several tests that may be performed to identify the type of emulsion that has formed:

- ▪ *Electrical conductivity*: o/w emulsions conduct electric current whereas w/o emulsions do not.
- ▪ *Dilution with water*: o/w emulsions may be diluted with water (as this is the composition of the external phase) whereas w/o emulsions cannot be diluted with water.
- ▪ *Use of dyes*: oil-soluble dyes will stain the internal phase if the emulsion is an o/w emulsion whereas water-soluble dyes will dye the internal phase of a w/o emulsion.

Emulsion instability

One of the goals of the pharmaceutical scientist is to formulate an emulsion that is physically stable, i.e. where the droplets of the internal phase remain discrete, retain their diameter and are homogeneously dispersed throughout the formulation. Fundamental to achieving this goal is the presence of the interfacial film (monomolecular or multilayered) at the interface between the droplet and the external phase. Emulsion instability may be either reversible or irreversible and is manifest in the following ways: (1) cracking (irreversible instability); (2) flocculation; (3) creaming; and (4) phase inversion.

Cracking (irreversible instability)

Cracking refers to the complete coalescence of the internal phase, resulting in the separation of the emulsion into two layers, and occurs due to the destruction of the mono/multilayer film at the interface between the droplet and external phase. If an emulsion has cracked it cannot be recovered. This phenomenon may be due to:

- *Incorrect selection of emulsifying agents.* This results in the production of an interfacial film of insufficient mechanical properties. The role of complexation (interaction) between the surfactant molecules at the interface between the two phases has already been described earlier in this chapter.
- *Presence of incompatible excipients.* In the formulation of emulsions it is important that excipients do not interact with and destroy the interfacial film of surface-active agents. This will occur if, for example, a cationic surface-active agent (commonly used as a preservative in creams) is added to an emulsion in which the interfacial film of surface-active agents bears an anionic charge (e.g. due to sodium oleate, potassium oleate or sodium lauryl sulphate). Similarly, if a therapeutic agent or a divalent ion bears an opposite charge to that exhibited by the interfacial film, disruption of the film will occur due to this ionic interaction.
- *Temperature.* Emulsions are generally unstable at high and low storage temperatures.
- *Microbial spoilage.* Microbial growth generally leads to destabilisation of the emulsion and is thought to be due to the microorganisms being able to metabolise the surface-active agents.

Flocculation

The ability of emulsion droplets to flocculate has been introduced earlier in this chapter. In the flocculated state the secondary interactions (van der Waals forces) maintain the droplets at a defined distance of separation (within the secondary minimum). Application of a shearing stress to the formulation (e.g. shaking)

will redisperse these droplets to form a homogeneous formulation. Although flocculation may stabilise the formulation, there is also the possibility that the close location of the droplets (at the secondary minimum) would enable droplet coalescence to occur if the mechanical properties of the interfacial film are compromised.

Creaming

This phenomenon occurs primarily as a result of the density difference between the oil and water phases and involves either the sedimentation or elevation of the droplets of the internal phase, producing a layer of concentrated emulsion either at the top or bottom of the container. Creaming is predominantly an aesthetic problem as the resulting emulsion is rather unsightly; however, upon shaking the emulsion is rendered homogeneous. Patients often believe that an emulsion that shows evidence of creaming has exceeded its shelf-life.

It is therefore important to understand the physicochemical basis of creaming in emulsions and, in so doing, reduce the rate of or inhibit this phenomenon. The rate of creaming $\left(\frac{\delta v}{\delta t}\right)$ in an emulsion (in a similar fashion to suspensions) may be described by Stokes' equation:

$$\frac{\delta v}{\delta t} = \frac{2r^2(\rho_o - \rho_w)g}{9\eta}$$

where: r refers to the average radius of the droplets of the internal phase; $(\rho_o - \rho_w)$ refers to the density difference between the oil phase and the water phase; g refers to gravity (which is negative if upward creaming occurs); and η refers to the viscosity of the emulsion.

As may be observed, creaming may be prevented if the density difference between the two phases is zero. In practice, however, this cannot be easily achieved. Therefore, the most straightforward methods by which the rate of creaming may be reduced are:

- Reduce the average particle size of the disperse phase. This may be achieved by size reduction methods, e.g. the colloid mill.
- Increase the viscosity of the emulsion. This may be achieved by adding hydrophilic polymers to the external phase of o/w emulsions or by incorporating non-aqueous viscosity enhancers (e.g. aluminium stearate salts, Thixin®) into w/o emulsions.

Phase inversion

Phase inversion refers to the switching of an o/w emulsion to a w/o emulsion (or vice versa). This is a phenomenon that

frequently occurs whenever the critical value of the phase volume ratio has been exceeded. In o/w emulsions the frequently cited phase volume ratio (o:w) is 74:26 and for w/o emulsions this value is 40:60.

Formulation of pharmaceutical emulsions

In the formulation of pharmaceutical emulsions there are a number of questions that require to be initially addressed, including the type of emulsion required (o/w or w/o), the route of administration of the emulsion (e.g. oral or topical, the latter as a cream), the volume of the internal phase, the droplet size and the consistency required. These aspects are individually addressed below.

> **Tips**
>
> Emulsion instability may be classified as either reversible or irreversible.
>   Cracking is irreversible.
>   Phase inversion will result in the switching of the type of emulsion. Although the preparation may be stable, the nature (and hence the clinical performance) of the emulsion has changed and therefore this phenomenon should be avoided.
>   Creaming, whilst unsightly, is reversible upon shaking.

Type of emulsion

As the reader is now aware, there are two types of primary emulsions; however the clinical uses of these types differ. Emulsions that are designed for oral or intravenous administration are o/w, whereas emulsions for topical administration (creams) may be either o/w or w/o. O/w creams are generally (but not exclusively) used for the topical administration of water-soluble drugs to the skin to achieve a local effect (e.g. for the treatment of infection or inflammation). They are typically easily applied to the surface, are non-greasy and may be washed from the skin. Conversely, w/o emulsions are greasy in texture and, following application, will hydrate the skin. Most moisturising formulations are w/o emulsions.

Volume of the internal phase

The effect of the volume of the internal phase on emulsion stability has been addressed previously in this chapter. The ratio of the internal-to-external phase of o/w emulsions is typically 1:1; however, large oil-to-water ratios are theoretically possible. Usually the concentration of the internal phase is restricted to circa 60% to ensure stability. The maximum concentration of internal phase of w/o emulsions is 30–40%. Higher concentrations will result in phase inversion.

Droplet size

Previously it was shown that the rate of creaming of an emulsion may be reduced by reducing the average droplet size of the internal phase. In light of this it is customary when industrially

processing emulsions and creams to reduce the droplet size (and reduce the polydispersity of the size distribution) by passage through a colloid mill. The clinical importance of droplet size in parenteral emulsions, e.g. total parenteral nutrition, will be specifically addressed in Chapter 5.

Viscosity of the internal and external phases

One of the major differences between traditional emulsions for oral or parenteral administration and creams is the increased viscosity of the latter. The superior viscosity of these formulations facilitates the location and spreading of the formulation on the skin. In addition, the viscosity of emulsion/cream formulations also affects the stability, controlling the rate of upward/downward sedimentation (as described by Stokes' law).

Selection of type and concentration of emulsifying agents

All emulsion and cream formulations require the inclusion of emulsifying agents (principally surface-active agents) to ensure emulsion stability, the choice of which is determined by the type of emulsion required, clinical use and toxicity. For example, the use of anionic surfactants is restricted to external formulations. To determine the type of emulsifying agents used, reference is made to the HLB requirements of the internal phase of the formulation. If the HLB requirements are not known, it is common practice for the formulation scientist to prepare a series of emulsions using a mixture of surface-active agents that provides a range of HLB values using a weighted-mean approach. For example, an o/w emulsion may be prepared using a mixture of surface-active agents (1% w/w in total) that provides an overall HLB value of 10. A mixture of Span 60 (HLB 4.7) and Tween 80 (15.0) may be chosen for this purpose; the ratio of these two surfactants is calculated using the simple weighted-averages equation:

$$10 = 4.7x + (1-x)15$$

where x refers to the fraction of Span 60 and $(1 - x)$ is the fraction of Tween 80.

In this example the mixture of surface-active agents would be 0.485% w/w Span 60 and 0.515% w/w Tween 80. In practice a series of emulsions would be prepared, each differing in the HLB of the surfactant mixture but constant in terms of the overall concentration of surfactants. From this the most stable emulsion would be selected.

Alternatively, for certain non-aqueous components information is available regarding the required HLB to produce stable o/w or w/o emulsions. For example, to prepare a stable o/w

emulsion using cottonseed oil as the internal phase requires a mixture of surface-active agents that produces an HLB value of 10.0, whereas the formation of a w/o emulsion in which cottonseed oil is the external phase requires a mixture of surface-active agents that produces an HLB value of 5. If the lipophilic component of the formulation is composed of more than one excipient, then the combined HLB value for this phase should be calculated and the ratio of surface-active agents in the mixture selected to provide this HLB requirement. For example, consider an o/w cream in which the internal phase (of total weight 35% w/w) has the composition shown in Table 3.1.

Table 3.1 Calculation of required hydrophile–lipophile balance (HLB) values for an oil in water (o/w) cream

Components (% w/w of final formulation)	HLB requirement to produce an o/w emulsion	Composition (fraction of the oil phase)	Calculated HLB requirement
Cottonseed oil (30%)	10	$\left(\dfrac{30}{35}\right)=0.86$	$(0.86\times10)=8.6$
Stearyl alcohol (3%)	14	$\left(\dfrac{3}{35}\right)=0.09$	$(0.09\times14)=1.26$
Beeswax (2%)	12	$\left(\dfrac{2}{35}\right)=0.05$	$(0.05\times12)=0.6$
			Total: 10.46

The HLB requirement for the formulation of a stable emulsion is 10.46. For this example we will choose a mixture of two non-ionic surface-active agents – polyoxyethylene sorbitan monostearate (Tween 60: HLB 14.9) and sorbitan monostearate (Span 60: HLB 4.7). Allowing x to equal the fraction of sorbitan monostearate in the surfactant mixture and $(1 - x)$ to represent the fraction of the second surfactant, the following calculation may be performed:

$$10.46 = 4.7x + (1 - x)14.9$$

Therefore the ratio of Span 60 to Tween 60 is 0.44:0.56.
    Two further points should be noted regarding the use of surface-active agents for the stabilisation of pharmaceutical emulsions: (1) the choice of the mixture of surfactants and (2) the overall concentration of surfactant.

- In choosing the surfactant blend, the formulation scientist should examine the structure of the surfactants to ascertain whether the two components may interact at the interface between the internal and external phases. In addition, it is preferable that the surfactant blend used should not be composed of one surfactant with a low HLB and the second with a high HLB, as in this case emulsion stability may be problematic. However, the addition of a third surface-active agent of intermediate HLB may resolve this issue.
- The concentration of surfactant used should be the lowest concentration required to ensure stability.

Types of surface-active agents used as emulsifying agents

There are four categories of surface-active agents used to stabilise emulsion/cream formulations: (1) anionic; (2) cationic; (3) non-ionic; and (4) amphoteric. Examples of these are provided in the section below. It should be noted that the cited examples are representative of the surface-active agents within each of the four classes. Moreover, the details provided for each example/category of surfactant are designed to provide an insight into the types and uses of the various agents. For a more comprehensive understanding of these agents, the reader should consult the companion text in the FASTtrack series by David Attwood and Alexander T Florence (*FASTtrack: Physical Pharmacy*: London: Pharmaceutical Press, 2008).

Anionic surfactants

- These dissociate to produce negatively charged ions with surface-active activity.
- Whilst examples in this category are inexpensive, they are comparatively more toxic than for other categories of surface-active agent. This limits their use to external formulations, e.g. creams.
- Examples of anionic surfactants include:
- *Sodium/potassium salts of fatty acids*
  - Examples include sodium oleate (Figure 3.1), sodium stearate and ammonium oleate

Figure 3.1 Structural formula of sodium oleate.

$$H_3C(CH_2)_6 \qquad (CH_2)_6COO^-Na^+$$
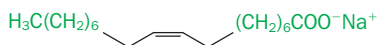
  - These agents produce o/w emulsions (usually in combination with a second surface-active agent to ensure

the formation of a mechanically robust film at the oil/water interface)
- Due to the effect of pH on the ionisation (and hence surfactant properties) of these molecules, the emulsifying properties are lost under acidic conditions. Similarly, the emulsifying properties are negated in the presence of di/trivalent cations.
- These surfactants may be formed in situ in the formulation by the co-addition of the fatty acid and a suitable counterion.

– *Calcium salts of fatty acids*
- These are generally formed in situ by the interaction of a calcium salt, e.g. calcium hydroxide, with a fatty acid, e.g. calcium oleate. This approach is used in Zinc Cream BP and in certain lotions.
- Calcium salts of fatty acids form w/o emulsions, due to their limited dissociation (and hence solubility: Figure 3.2).

$$\left[CH_3(CH_2)_{16}COO_-\right]_2 Ca^{2+}$$
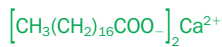
Figure 3.2 Structural formula of calcium stearate.

– *Amine salts of fatty acids*
- These are typically formed in situ in pharmaceutical emulsions, e.g. triethanolamine stearate.
- These surface-active agents form o/w emulsions.
- Akin to sodium/potassium salts of fatty acids, their emulgent properties are pH-dependent and may be negated in the presence of electrolytes.

– *Alkyl sulphates*
- These are used to produce o/w emulsions (in conjunction with a second non-ionic surfactant of low HLB, i.e. < 6). Fatty alcohols (e.g. cetyl, stearic alcohol) are frequently used for this purpose.
- Examples of these include sodium lauryl sulphate (Figure 3.3) and triethanolamine lauryl sulphate.

$$CH_3(CH_2)_{10}CH_2OSO_3^- Na^+$$

Figure 3.3 Structural formula of sodium lauryl sulphate.

Cationic surfactants
- These dissociate to produce positively charged ions with surface-active activity.

- They are primarily used pharmaceutically as preservatives of topical formulations; however, they may be used to form o/w emulsions (when combined with a second non-ionic surfactant of low HLB, i.e. < 6).
- The main example used in topical formulations is cetrimide, a mixture of trimethylammonium bromide, with smaller amounts of dodecyltrimethylammonium bromide and hexadecyltrimethylammonium bromide (Figure 3.4).

Figure 3.4 Structural formula of hexadecyltrimethylammonium bromide (cetrimide).

$$CH_3(CH_2)_{15} - \overset{\overset{\displaystyle CH_3}{|}}{\underset{\underset{\displaystyle CH_3}{|}}{\overset{H^+}{N}}} - CH_3Br^-$$
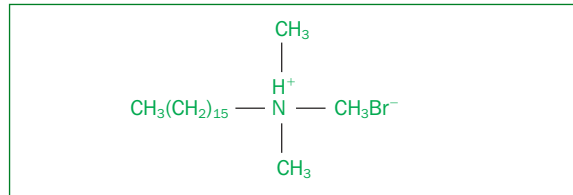
- Their emulgent properties are compromised in the presence of anionic agents (e.g. anionic surface-active agents, di/trivalent anions and polyelectrolytes, e.g. anionic polymers).

Non-ionic surfactants
- These are by far the most popular category of surface-active agents used for the formulation of pharmaceutical emulsions.
- They may be used to formulate both o/w and w/o emulsions.
- Generally combinations of two non-ionic surfactants (one water-soluble and the other oil-soluble) are employed to ensure the formation of a stable interfacial film around the surface of the droplets of the disperse phase. In certain circumstances a single non-ionic surfactant may be used that is of intermediate HLB value.
- Non-ionic surface-active agents are more stable than ionic surfactants in the presence of electrolyte and/or changes in pH.
- Generally the hydrophobic portion of the molecule is composed of a fatty acid or fatty alcohol whereas the hydrophilic portion is composed of an alcohol or ethylene glycol moieties.
- Examples of non-ionic surface-active agents include:
- *Sorbitan esters (e.g. Span series)*
   - This is a family of chemically related esters that are produced by esterifying a fatty acid to at least one of the hydroxyl groups of sorbitan.
   - Modification of the length of the fatty acid (denoted by the symbol R in Figure 3.5) will generate a range of surface-active agents with emulsifying properties (and low HLB values).

- By themselves sorbitan esters will form w/o emulsions; however, when combined with the polysorbates (see below), both o/w and w/o emulsions may be formulated.
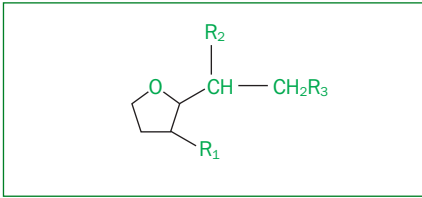
Figure 3.5 Generic structure of the sorbitan fatty acid esters. In sorbitan monoesters R_1 and R_2=OH, R_3 is the specific fatty acid derivative (e.g. lauric, stearate). R_1=OH and R_2 and R_3 refer to the specific fatty acid derivative for sorbitan diesters. In sorbitan trimesters R_1, R_2 and R_3 refer to the specific fatty acid derivative.

− Polyoxyethylene fatty acid derivatives of the sorbitan esters (e.g. Tween series)
  - This family of surface-active agents is prepared by forming polyoxyethylene esters of the sorbitan esters.
  - The emulsifying properties of the molecules in this series may be modified by altering the number of oxyethylene (OCH_2CH_2) groups and the type of fatty acid (denoted as R in Figure 3.6).
  - These surfactants are used to form o/w or w/o emulsions in combination with a second surface-active agent, e.g. sorbitan esters, cetyl alcohol, glyceryl monostearate, to ensure emulsion stability.
  - The emulsifying properties of this series are tolerant of changes in electrolyte concentration and pH.
  - Generally they are non-toxic and are used in both parenteral and non-parenteral emulsions.
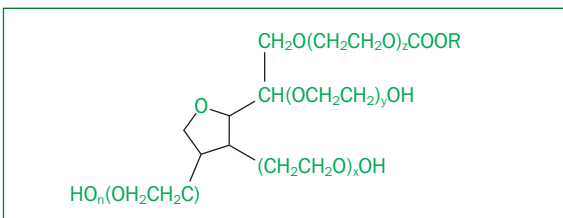
Figure 3.6 Generic structure of the polyoxyethylene sorbitan fatty acid monoesters. Di- and tri-esters may be formed by esterification of the terminal alcohol groups.

− Polyoxyethylene alkyl ethers (macrogols)
  - These are ethers formed between polyethylene glycol and a range of fatty alcohols (lauryl, oleyl, myristyl, cetyl, stearyl). Two commercial series of these compounds are Cremophor and Brij.
  - The physicochemical properties of these non-ionic surface-active agents may be modified by altering the length of the polyoxyethylene group and the length of the aliphatic chain (denoted as x and y in Figure 3.7).

Figure 3.7 Structural formula of the polyoxyethylene alkyl ethers.

$$CH_3(CH_2)_x(OCH_2CH_2)_yOH$$

- The macrogols are used as emulsifying agents for both o/w and w/o emulsions. Combinations of the more lipophilic and hydrophilic examples of this series are combined to produce stable emulsions.
- For example, cetomacrogol 1000 (Figure 3.8) is combined with cetostearyl alcohol to produce cream formulations.

Figure 3.8 Structural formula of cetomacrogol 1000.

$$CH_3(CH_2)_{15-17}(OCH_2CH_2)_{20-24}OH$$

- *Polyoxyethylene fatty acid esters*
    - These are a series of polyoxyethylene derivatives of fatty acids. The most commonly used derivatives are the stearate derivatives (the Myrj series). The surface-active properties of these compounds may be modified by varying the length of the oxyethylene substituent and, in addition, by mono- or diesterification of the acid, as shown in Figure 3.9.

Figure 3.9 Generic structures of polyoxyethylene monoester and poly(oxyethylene) ester. The number of repeating oxyethylene groups is denoted by n, whereas R refers to the chain length of the fatty acid.
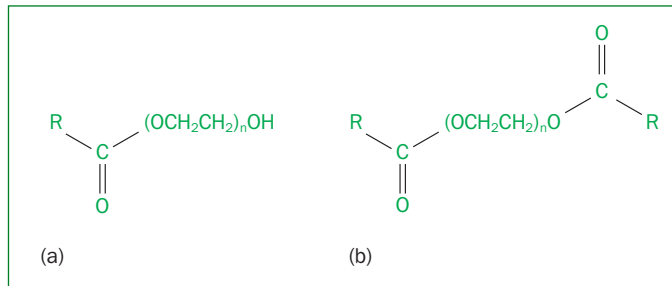
- Polyoxyethylene stearates (and related compounds) are non-ionic surfactants, offering a range of HLB values.
- They are frequently combined with stearyl alcohol (or related fatty alcohols) in the formulation of o/w emulsions.
- The emulsifying properties are tolerant of the presence of strong electrolytes.
- *Fatty alcohols*
    - Examples include cetyl alcohol and stearyl alcohol (Figure 3.10).

$$CH_3(CH_2)_{14}CH_2OH \qquad CH_3(CH_2)_{16}CH_2OH$$

(a)   (b)

Figure 3.10 Structural formula of (a) cetyl alcohol and (b) stearyl alcohol.

In addition, cetostearyl alcohol (a mixture of cetyl (20–35%) and stearyl (50–70%) alcohols, although other alcohols, e.g. myrisitc alcohol, are present) is available (see later in this chapter).

- Fatty alcohols are generally used in combination with more hydrophilic surfactants to produce stable o/w emulsions.
- When used alone, fatty alcohols act as w/o emulsifiers. Furthermore, the addition of these to a hydrophobic base will increase the water absorption properties of the formulation.
- In cream formulations excess fatty alcohols interact with the hydrophilic emulsifier to produce a viscoelastic external phase. In turn, this increases the viscosity of this phase (thereby decreasing upward/downward sedimentation), producing the consistency expected of cream formulations.
- Fatty alcohols may be used to enhance the viscosity of w/o creams.

- *Amphoteric surfactants*
  - These are compounds that possess both positively and negatively charged groups (cationic at low pH values and anionic at high pH values).
  - The emulsifying properties are reduced as the pH approaches the isoelectric point of the surface-active agent.
  - The most commonly used amphoteric surface-active agent is lecithin (Figure 3.11).
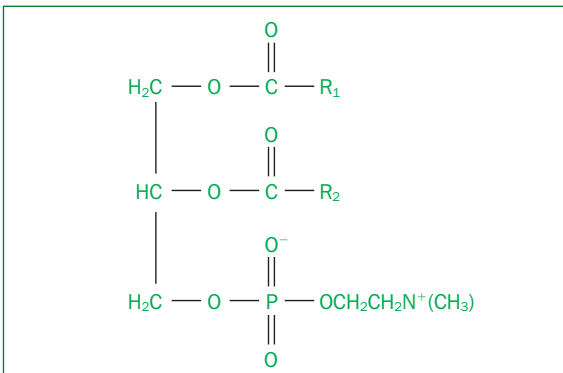
Figure 3.11 Structural formula for lecithin (R_1 and R_2 refer to either identical or different fatty acids).

- Lecithin is used in emulsions (for intravenous and intramuscular administration) and creams, in which it acts as an o/w emulsifying agent.

Miscellaneous surfactants that are used in emulsion and cream formulations

The following emulsifying agents are derived from natural sources and are composed of mixtures of compounds. They are used as emulsifying agents either alone, or, preferably, along with a second emulsifying agent in the production of o/w and w/o creams.

Lanolin (wool fat)

- Lanolin is a wax-like material that is derived from sheep's wool. It is a mixture of fatty alcohols, fatty acid esters of cholesterol and other sterols.
- On its own it may be used to produce w/o creams. Mixtures of this emulsifying agent with oils or soft paraffin produce emollient creams.
- Lanolin can absorb approximately twice its weight of water – this property is advantageous in the formulation of ointments.

Lanolin alcohols (wool alcohols)

- This is a mixture of steroid alcohols and triterpene alcohols (including cholesterol).
- Lanolin alcohol is an excellent emulsifying agent for w/o emulsions.
- It has excellent water-absorptive properties.
- It is prone to oxidation (and therefore requires an antioxidant).

Anionic emulsifying wax

- This is a mixture containing cetostearyl alcohol, water and sodium lauryl sulphate (or a related sulphated alcohol). The specific formula for anionic emulsifying wax in the British Pharmacoepeia (2004) is as follows:
  cetostearyl alcohol (90 g)
  sodium lauryl sulphate (10 g)
  purified water (4 ml).
- This mixture is used to produce o/w emulsions (using 2% w/w wax) or creams, at a higher wax concentration (10% w/w). However, the use of anionic emulsifying wax is predominantly for topical (cream) formulations.
- Due to the anionic nature of one of the ingredients, the emulgent properties of this wax are compromised by the presence of polyvalent metals and cations (e.g. quaternary ammonium compounds).

Non-ionic emulsifying wax

- Non-ionic emulsifying wax (also called cetomacrogol emulsifying wax) is composed of cetostearyl alcohol and cetomacrogol 1000.
- This wax is used as an emulsifier for the preparation of emulsions (at concentrations up to 5% w/w). However at higher concentrations this material additionally enhances the rheological structure of the preparation (15–25% w/w), thereby promoting stability. The primary use of non-ionic emulsifying wax is for the preparation of creams.
- The emulgent properties of non-ionic emulsifying wax are tolerant to the presence of electrolytes (and therapeutic agents).

Beeswax (white and yellow)

- There are two forms of beeswax – white and yellow. White beeswax is a bleached form of yellow beeswax.
- Beeswax is a mixture of esters of monohydric alcohols (C_{24}–C_{36}, in even numbers only) and straight-chain acids (even numbers of carbon atoms up to C_{36}), long-chain hydroxyacids (e.g. C_{18}). The principal component is myricyl palmitate.
- Beeswax is used as a w/o emulsifying agent for creams and is also used to enhance the consistency of creams.

Excipients used in pharmaceutical emulsions

One major category of excipients for pharmaceutical emulsions, namely surface-active agents, has been described above. However, as in other pharmaceutical formulations, other excipients are required to enhance the physical and chemical stability and to render the formulation aesthetically pleasing to the patient. These are described below.

Vehicle

There are two liquid phases in pharmaceutical emulsions: an aqueous phase and an oil phase, each of which is formulated separately. The vehicle in the aqueous phase for pharmaceutical emulsions designed for oral or topical administration is usually *purified water*, the details of which have been provided in Chapter 1. When formulated for intravenous administration, *sterile water for injections* is used as the external aqueous phase,

Tips

The type of surfactants used dictates the stability and type of the emulsion.

In the choice of surfactant type for an emulsion, due attention must be paid to the toxicity of the surfactant. For example, non-ionic, but not anionic, surfactants may be administered orally. Certain surfactants are only used in the formulations for topical administration. Cationic surfactants are principally used as preservatives and not for the stabilisation of emulsions.

Combinations of surfactants are used to stabilise emulsions. The elastic properties of the surfactant layer at the interface are important in the stabilisation of emulsions.

the details of which are provided in Chapter 5. If control of the pH of the aqueous external phase is required, buffers, e.g. citrate, phosphate, may be included in the aqueous vehicle. In light of the ability of electrolytes to compromise the emulsifying properties of surface-active agents (and certain hydrophilic polymers), the concentration and type of buffer should be carefully chosen.

The oil phase of pharmaceutical emulsions is typically composed of vegetable oils, e.g. cottonseed oil, arachis oil, almond oil (mono-, di- and triglycerides of mixtures of unsaturated and saturated fatty acids). However, pharmaceutical emulsions for topical application (creams) may be formulated using a greater range of non-aqueous components. Alternative non-aqueous phases used in the formulation of creams and ointments include: (1) petrolatum and mineral oil; (2) isopropyl myristate; (3) antioxidants; (4) flavours and sweeteners; (5) viscosity modifiers; (6) preservatives for emulsions and creams.

Petrolatum and mineral oil

Petrolatum and mineral oil are hydrophobic excipients that are derived from petrolatum. The former is a complex mixture of hydrocarbons (e.g. aliphatic, cyclic, saturated, unsaturated, branched hydrocarbons) that results in a wide range of chemical and physical specifications in the USP monograph. Mineral oil is a more purified fraction of petrolatum and is a mixture of aliphatic (C_{14}–C_{18}) and cyclic hydrocarbons.

Both materials are employed as the internal phase in o/w emulsions and as the external phase in w/o emulsions (usually in combination with a fatty alcohol as the emulsifying agent).

Isopropyl myristate (Figure 3.12)

Figure 3.12 Structural formula of isopropyl myristate.
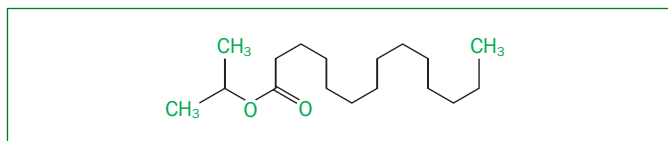
Isopropyl myristate is used as a non-aqueous component of cream formulations, either as the internal phase of o/w creams or as the external phase of w/o creams. More recently isopropyl myristate has been reported to enhance the permeation of drugs through the skin when applied topically.

Antioxidants

As described in Chapter 1, antioxidants are included within pharmaceutical formulations to enhance the stability of

drugs/components to oxidation. In emulsions and creams the two major components that may be liable to oxidise are the therapeutic agent and the oil selected for the oil phase, vegetable oils. Therefore the inclusion of lipophilic antioxidants within the oil phase may be required, e.g. butylated hydroxyanisole (circa 0.02–0.5% w/w), butylated hydroxytoluene (circa 0.02–0.5% w/w) and propyl gallate (< 0.1% w/v) (Figure 3.13).
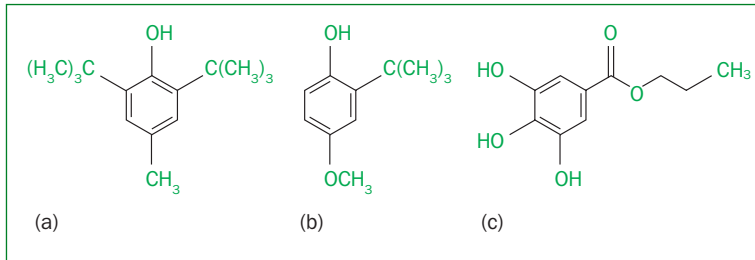
Figure 3.13 Structural formula of (a) butylated hydroxytoluene; (b) butylated hydroxyanisole; and (c) propyl gallate.

If the antioxidant is required in the aqueous phase of an emulsion or cream then a water-soluble example should be used, e.g. sodium metabisulphite (0.01–1.0% w/v) or sodium sulphite (0.1% w/v).

Flavours and sweeteners
Flavours and sweeteners are commonly included in emulsions for oral administration to mask the unpalatable taste of the therapeutic agent or the internal oil phase. Suitable examples of these have been given in Chapter 1.

Viscosity modifiers
The viscosity of emulsions and creams has been previously described to influence the physical stability of emulsions by decreasing the rate of creaming and therefore viscosity control within a formulation is an important attribute. The inclusion of hydrophilic polymers, e.g. methylcellulose, hydroxyethylcellulose, polyacrylic acid and sodium carboxymethylcellulose, to increase the viscosity of aqueous systems has been described in Chapters 1 and 2 and the same principles exist for emulsion formulations. It must be remembered that, as the viscosity of formulations increases, so does the difficulty in administration and, therefore, this must be borne in mind when the final viscosity of the o/w emulsion is selected. Furthermore, the ability of hydrophilic polymers to form a multimolecular layer around the surface of the dispersed-oil droplet is an important function of polymers within emulsion formulations.

Preservatives for emulsions and creams

The concept of preservation of pharmaceutical systems, i.e. solutions and suspensions, has been discussed in previous chapters. In particular the effects of pH and the presence of hydrophilic polymers, dispersed particles and surfactant micelles on the available preservative concentration were highlighted. The preservation of o/w emulsions and creams becomes a challenging task to the pharmaceutical scientist due to the possible co-requirement for pH control of the external phase and the inclusion of hydrophilic polymers. However, the complexity of this issue is enhanced due to the presence of a dispersed-oil phase into which the antimicrobial active form of the preservative may partition and hence be unavailable to exert its antimicrobial effect. An equilibrium is therefore established, as depicted in Figure 3.14.

Figure 3.14 Diagrammatic representation of the equilibrium that exists between a dispersed-oil phase, the external aqueous phase, micelles of surface-active agent, a hydrophilic polymer (e.g. methylcellulose, polyvinylpyrrolidone) and the antimicrobially active form of an organic acid preservative (denoted as HA).
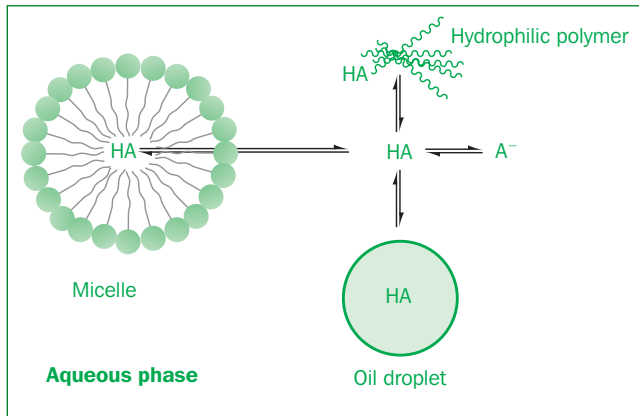
Figure 3.14 illustrates the partitioning of the unionised form of a weak acid preservative into micelles and oil droplets. The term HA may be replaced by a non-ionisable (or minimally ionisable) preservative, e.g. chlorocresol. As may be observed in Figure 3.14, the available (active) concentration is decreased by these various partitioning phenomena and therefore the concentration of preservative must be increased to ensure that required concentration of free preservative is obtained. The concentration of preservative required to inhibit microbial growth in emulsions/creams may be estimated using the following formula:

$$C_w = C\left(\frac{\emptyset+1}{K_w^o\emptyset+R}\right)$$

where: C_w refers to the concentration of 'free' preservative in the aqueous phase; C refers to total concentration of preservative; ø refers to the ratio of oil (internal phase) to water in the emulsion/cream; K_w^o refers to the partition coefficient of the preservative between the oil and water phases; and R is the ratio of the total preservative to the free preservative.

With knowledge of the above parameters, the free concentration of preservative may be calculated, as illustrated in the example below.

Worked example

Example 3.1

Calculate the free concentration of chlorocresol in an emulsion in which the oil:water partition coefficient of the preservative is 1.5, the phase ratio in the emulsion is 1:1, the ratio of total to free preservative is 4, the pH is 7.2 and the initial concentration of preservative is 0.3% w/v.

According to the above equation the concentration of free preservative is:

$$C_w = 0.3 \left(\frac{1+1}{1.5 \times 1+4} \right) = 0.1\%$$

Therefore, 33.3% of the preservative is available to the formulation to exert its antimicrobial effect within the formulation.

This situation becomes further complicated if the preservative ionises as a function of the pH of the formulation. To accommodate this, the degree of ionisation must be calculated (again using the Henderson–Hasselbalch equation). The pKa for chlorocresol is 9.2.

$$Fraction = \frac{1}{(1+10^{pH-pKa})} = \frac{1}{(1+10^{-2})} = 0.99$$

It can therefore be seen that at pH 7.2 chlorocresol is essentially unionised and no modification of the concentration is required. Conversely, if an organic acid is used (pKa 4.2), the fraction unionised will be 0.001 and therefore this must be considered in the calculation of the required concentration.

From the discussions to date, the role of the oil:water partition coefficient in the calculation of the free concentration of preservative is apparent. Therefore, to optimise preservative efficacy, the oil used as the internal phase should have low oil:water partition coefficient for the selected preservative. For

example, the partition coefficients of methyl parahydroxybenzoic acid in mineral oil and vegetable oil are 0.02 and 7.5. Therefore an obvious method of minimising the concentration of preservative in the formulation (whilst retaining the required antimicrobial activity) would be to select mineral oil as the internal phase. Alternatively, if an internal phase is specified, the total concentration of preservative may be minimised by selecting an alternative preservative with a low oil:water partition coefficient.

Manufacture of emulsions

Generically the manufacture of emulsions involves the following steps:

1. dissolution of the oil-soluble components in the oil vehicle and the (separate) dissolution of the water-soluble components in the aqueous phase
2. mixing of the two phases under turbulent mixing conditions to ensure the dispersion of the two phases into droplets.

At the laboratory the manufacture of emulsions usually involves the use of a mechanical stirrer whereas the manufacture of creams involves mixing the two (heated) phases using a mortar and pestle. The emulsification of production-scale batches is normally performed using mechanical stirrers, homogenisers, ultrasonifers or colloid mills. The use of colloid mills is usually reserved for formulations of higher viscosity, e.g. creams, due to the high running cost and slow production rate of this apparatus.

Tips

The formulation of emulsions requires the incorporation of several excipients to maintain stability and to ensure product performance.

As in other dosage forms, all excipients must be physically and chemically compatible (with each other and with the therapeutic agent).

The choice of preservative for an oil in water emulsion requires consideration of both the pH of the aqueous phase (if the preservative is acidic) and the solubility within the oil phase. Significant solubility of the preservative in the oil phase (and/or excessive ionisation of the preservative) lowers the preservative efficacy.

Multiple choice questions

1. **Regarding the stability of pharmaceutical emulsions, which of the following statements are true?**
a. Emulsions are inherently pharmaceutically unstable.
b. The stability of pharmaceutical emulsions is affected by the size of the dispersed phase.
c. The stability of pharmaceutical suspensions is affected by the concentration of dispersed phase.

d. Phase volume of the internal phase directly affects the
    stability of pharmaceutical emulsions.

2. **Regarding the rate of creaming of pharmaceutical emulsions,
    which of the following statements are true?**
a. The rate of creaming is increased as the diameter of the
    internal phase is increased.
b. The rate of creaming is increased as the viscosity of the
    continuous phase is increased.
c. The rate of creaming is affected by the concentration and type
    of incorporated surfactants.
d. The rate of creaming is decreased by centrifugation.

3. **Regarding emulsions, which of the following statements are
    true?**
a. Multiple emulsions are more stable than primary emulsions.
b. Water in oil emulsions are commonly administered orally.
c. Oil in water emulsions are stable following dilution with
    water.
d. Dispersed globules of the internal phase do not possess a zeta
    potential.

4. **Regarding the role of surfactants in the formulation of
    emulsions, which of the following statements are true?**
a. Oil in water emulsions are promoted by the presence of
    surfactants with HLB values > 8.
b. The elastic properties of the surfactant layer at the interface
    between the internal and external phases are important in the
    stabilisation of the emulsion.
c. Anionic surfactants may be used for the stabilisation of
    emulsions designed for oral administration.
d. Surfactants stabilise emulsions by increasing interfacial Gibb's
    free energy of the emulsion.

5. **Regarding the role of adsorbed particles in the stabilisation
    of pharmaceutical emulsions, which of the following
    statements are true?**
a. Adsorbed particles promote the formation of oil in water
    emulsions.
b. Examples of adsorbed particles that are employed
    pharmaceutically include kaolin.
c. The elastic properties of the adsorbed layer are primarily
    responsible for the stabilisation of emulsions.
d. The particle size of the adsorbed particles directly affects
    emulsion stability.

6. **Regarding the use of hydrophilic polymers for the stabilisation of pharmaceutical emulsions, which of the following statements are true?**
   a. Hydrophilic polymers stabilise pharmaceutical emulsions by increasing the viscosity of the continuous phase.
   b. Hydrophobic polymers may be employed to stabilise oil in water emulsions.
   c. Hydrophilic polymers form a stable monomolecular layer at the interface between the internal and external phases.
   d. Pharmaceutical emulsions may exhibit thixotropy.

7. **Regarding the preservation of pharmaceutical suspensions for oral administration, which of the following statements are true?**
   a. Hydrophilic polymers that are present in pharmaceutical emulsions may reduce preservative efficacy.
   b. Preservative efficacy may be reduced by partitioning of the preservative from the oil phase to the water phase of water in oil emulsions.
   c. Parabens are commonly used as preservatives for emulsions.
   d. The preservative efficacy of organic acids in oil in water emulsions may be affected by the type of oil used as the internal phase.

8. **Concerning the physical properties of emulsions, which of the following statements are true?**
   a. Water in oil emulsions conduct electrical current.
   b. Water in oil emulsions are more stable than oil in water emulsions.
   c. High-speed mixing of water in oil emulsions results in dilatant flow.
   d. The particle size distribution of the internal phase may be reduced postmanufacture using a colloid mill.

9. **Concerning the use of oil in water emulsions, which of the following statements are true?**
   a. Oil in water emulsions may be used for the oral administration of therapeutic agents.
   b. Oil in water emulsions may be formulated for topical administration.
   c. Drugs with high aqueous solubility are frequently formulated as oil in water emulsions.
   d. Oil in water emulsions designed for oral administration must be coloured.

10. Concerning the formulation of pharmaceutical emulsions, which of the following statements are true?

a.   Emulsions for oral administration may require the addition of sweetening agents.

b.   Emulsions for oral administration may require the addition of flavours.

c.   Emulsions for oral administration may require the addition of antioxidants.

d.   The optimum phase–volume ratio for emulsions is 50:50.

chapter 4

Pharmaceutical disperse systems 3: ointments, pastes, lotions, gels and related formulations

Overview

In this chapter the following points will be discussed:
- an overview/description of the physical properties and uses of ointments, pastes, lotions, liniments, collodions and gels
- formulation strategies for ointments, pastes, lotions, liniments, collodions and gels, including consideration of the excipients used
- the advantages and disadvantages and uses of ointments, pastes, lotions, liniments, collodions and gels
- considerations for the manufacture of ointments, pastes, lotions, liniments, collodions and gels.

Introduction

This chapter deals with the formulation of several types of *disperse* systems, namely ointments, pastes, lotions, liniments, collodions and gels. In the vast majority of cases these formulations contain a therapeutic agent and are designed for the localised treatment of a designated area, e.g. haemorrhoids, infection, inflammation. Whilst the above formulation types may be classified as disperse systems, there are distinct differences in their uses and the strategies utilised in their successful formulation. Accordingly, in this chapter, each dosage form is discussed separately.

KeyPoints

- Ointments, pastes, lotions, liniments, collodions and gels are further examples of disperse systems into which a therapeutic agent may be incorporated.
- Ointments, pastes, lotions, liniments, collodions and gels are topical formulations, being applied externally or into accessible body cavities (e.g. mouth, rectum, vagina).
- These formulations exhibit similar concerns as other disperse systems regarding physical stability.

Advantages and disadvantages of pharmaceutical ointments, pastes, lotions, liniments, collodions and gels

Advantages

- Pharmaceutical ointments may be easily spread on skin, being retained at the site of application as an occlusive layer, thereby preventing moisture loss from the skin. This is particularly useful whenever restoration of the physical characteristics of the skin is required (e.g. due to inflammation).
- Pharmaceutical ointments are associated with lubricating/emollient properties, properties that may be employed to reduce trauma of an affected site upon spreading.
- In general, pharmaceutical ointments persist at the site of application, enabling the duration of drug release to be greater than for many other topical dosage forms. The increased viscosity of pharmaceutical pastes ensures that a thick film of the dosage form is applied to the site of action, which shows excellent persistence. This property is particularly useful if protection of an inflamed site is required, e.g. in eczema, psoriasis.
- The hydrophobicity and retention of pharmaceutical ointments are useful attributes whenever applied to mucosa, e.g. inflamed haemorrhoids, eyelids, where fluid flow/ inflammation at these sites would normally serve to remove other formulations (e.g. oil in water creams) by dilution. It should be noted, however, that spreading of ointments on to moist surfaces may be difficult due to the hydrophobic properties of most ointments.
- Due to the high solids content, pharmaceutical pastes are often porous, allowing moisture loss from the applied site. Furthermore, pastes may act to absorb moisture and chemicals within the exudates.
- The opaque nature of pastes (due to the high solids content) enables this formulation to be used as a sunblock.
- The chemical stability of therapeutic agents that are prone to hydrolysis will be dramatically enhanced by formulation within pharmaceutical ointments and pastes.
- Pharmaceutical gels may be formulated to provide excellent spreading properties and will provide a cooling effect due to solvent evaporation. Similarly solvent evaporation from liniments will provide a cooling effect.

Disadvantages

- Pharmaceutical ointments are generally greasy and difficult to remove (and are therefore often cosmetically unacceptable). Similarly, liniments and lotions may also be cosmetically unacceptable to the patient and difficult to use.
- Pharmaceutical pastes are generally applied as a thick layer at the required site and are therefore considered to be cosmetically unacceptable.
- Staining of clothes is often associated with the use of pharmaceutical pastes and ointments.
- The viscosity of pharmaceutical ointments, and in particular pastes, may be problematic in ensuring spreading of the dosage form over the affected site. Conversely, the low viscosity of liniments and lotions may result in application difficulties.
- Pharmaceutical ointments may not be applied to exuding sites (however, please note that this does not hold for pastes). Liniments may not be applied to broken skin.
- Problems concerning drug release from pharmaceutical ointments may occur if the drug has limited solubility in the ointment base.
- Pharmaceutical pastes are generally not applied to the hair due to difficulties associated with removal.
- Therapeutic agents that are prone to hydrolysis should not be formulated into aqueous gels.

Pharmaceutical ointments and pastes

General description

Pharmaceutical ointments (termed *unguents*) are semisolid systems that are applied externally, primarily to the skin and also to mucous membranes, e.g. the rectum, the vagina/vulva, the eye. Typically, medicated ointments are used for the treatment of infection, inflammation and pruritus. However, non-medicated ointments are commonly used due to their emollient/lubricating properties. Pharmaceutical pastes are generally composed of ointment bases that contain a high concentration (frequently > 50% w/w) of dispersed drug. The viscosity of pharmaceutical pastes is greater than that of pharmaceutical ointments.

Introduction

The formulation of ointments and pastes involves the dispersal or dissolution of the selected therapeutic agent into an *ointment base* and, therefore, in addition to the physical properties of the dispersed/dissolved drug, the physicochemical properties of the ointment base are fundamental to the clinical and non-clinical performance of this type of dosage form. The choice of ointment

base is dependent on several factors, including: (1) the site of application; (2) the required rate of drug release; (3) the chemical stability of the drug; and (4) the effect of the therapeutic agent on formulation viscosity.

The site of application
In certain clinical conditions the site to which the ointment will be applied may be dry, e.g. psoriasis, or moist. If the area is dry, ointments are often used to occlude the site, thereby retaining moisture. Indeed, this effect is considered to play an important role in the treatment of certain clinical conditions. Conversely, occlusive ointment bases are not applied to sites in which there is fluid exudate.

The required rate of drug release
Following application, the therapeutic agent must be released to exert its pharmacological effect, either locally or, after absorption, systemically. Drug release from the ointment base requires solubility (albeit partial) of the therapeutic agent within the formulation. This will allow diffusion of the therapeutic agent (a molecular process) through the ointment base until it reaches the biological substrate. Therefore the choice of the ointment base is partially dictated by the physicochemical properties (and in particular the solubility) of the therapeutic agent.

The chemical stability of the drug
If a therapeutic agent is prone to hydrolysis, incorporation into a water-based formulation, e.g. oil in water creams, may lead to drug degradation and hence a shortened shelf-life. This problem may be obviated by incorporating the drug into a hydrophobic ointment base. For example, the shelf-life of hydrocortisone is markedly greater in an ointment formulation than in an oil in water cream formulation.

The effect of the therapeutic agent on formulation viscosity
The effect of the physical incorporation of a therapeutic agent into an ointment base on the rheological properties of the formulated product will be dependent on the required drug concentration, the physical properties of the therapeutic agent (e.g. particle size, shape) and the chemical composition and viscosity of the ointment base. Therefore, it is important that an ointment base is selected that will produce a product that may be readily applied to the required site. In light of the high drug content, this point is particularly important in the formulation of pastes.

Types of base for ointments and pastes

There are four types of base that are used to formulate pharmaceutical ointments and pastes: (1) hydrocarbon; (2) absorption; (3) water-miscible/removable; and (4) water-soluble.

Hydrocarbon bases

Hydrocarbon bases are non-aqueous formulations, based on various paraffins, that have the following properties:

- emollient, thereby restricting water loss from the site of application due to the formation of an occlusive film
- excellent retention on the skin
- predominantly hydrophobic, and therefore difficult to remove from the skin by washing and difficult to apply to (spread over) wet surfaces (e.g. mucous membranes, wet skin)
- only a low concentration ($< 5\%$) of water may be incorporated into hydrocarbon bases (with careful mixing)
- chemically inert.

Hydrocarbon bases frequently contain the following components: (1) hard paraffin; (2) white/yellow soft paraffin; (3) liquid paraffin (mineral oil); and (4) microcrystalline wax.

Hard paraffin

This is a mixture of solid saturated hydrocarbons that are derived from petroleum or shale oil. Hard paraffin is a colourless or white wax-like material that is physically composed of a mixture of microcrystals. The melting temperature of hard paraffin is between 47 and 65°C and, when solid, it is used to enhance the rheological properties of ointment bases.

White/yellow soft paraffin

This is a purified mixture of semisolid hydrocarbons (containing branched, linear and cyclic chains) that are derived from petroleum. White/yellow soft paraffin consists of microcrystals embedded in a gel composed of liquid and amorphous hydrocarbons that are themselves dispersed in a gel phase containing liquid and amorphous hydrocarbons. The melting range of the soft paraffins is between 38 and 60°C. White soft paraffin and yellow soft paraffin (the former being a bleached form of yellow soft paraffin) may be used as an ointment base without the need for additional components, although it may be combined with liquid paraffin (see below)

Liquid paraffin (mineral oil)

This is a mixture of saturated aliphatic (C_{14}–C_{18}) and cyclic hydrocarbons that have been refined from petroleum. It is usually formulated with white/yellow soft paraffin to achieve the required viscosity for application to the required site.

Formulations containing liquid paraffin require the incorporation of an antioxidant due to the ability of this material to undergo oxidation.

Microcrystalline wax

This is a solid mixture of saturated alkanes (both linear and branched) with a defined range of carbon chain lengths (C_{41}–C_{57}). This excipient is used to enhance the viscosity of ointments (and creams). One of the advantages of microcrystalline wax is the greater physical stability provided to formulations containing liquid paraffin (reduced bleeding of the liquid component).

Absorption bases

Unlike hydrocarbon bases, absorption bases may be formulated to contain significant amounts of an aqueous phase. These may be either non-aqueous formulations to which an aqueous phase may be added to produce a water in oil emulsion (termed *non-emulsified bases*) or *water in oil emulsions* that can facilitate the incorporation of an aqueous phase (without phase inversion or cracking). Although absorption bases can accommodate a larger volume of aqueous phase than hydrophobic bases, they are still difficult to remove from the site of application by washing. This is due to the predominantly hydrophobic properties of this formulation class.

The key properties of both non-emulsified bases and water in oil emulsions that are relevant to the formulation of ointments and pastes are detailed below.

Non-emulsified bases

These are hydrophobic formulations to which water may be added. Following application, a film is formed that offers occlusion (and hence emollient properties); however, the extent of occlusion is less than for hydrocarbon bases. The spreading properties of these formulations are more favourable than for hydrocarbon bases.

Typically non-emulsified bases are commonly composed of: (1) one or more paraffins (see previous section) and (2) a sterol-based emulsifying agent. Examples of the types of emulsifying agents used in absorption bases include: (1) lanolin (wool fat); (2) lanolin alcohols (wool alcohols); and (3) beeswax (white or yellow).

Lanolin (wool fat)

Lanolin is a wax-like material that is derived from sheep's wool. It is available in two forms, termed lanolin (wool fat) and hydrous lanolin (wool alcohols). Lanolin is typically mixed with vegetable oils or paraffins to produce an ointment base that can absorb

approximately twice its own weight of water to produce water in oil emulsions. The usual concentrations of lanolin used in ointments (e.g. Simple Ointment BP) range from 5 to 10% w/w.

Lanolin alcohols (wool alcohols)

As detailed in Chapter 3, wool alcohol is a crude mixture of sterols and triterpene alcohols and contains at least 30% cholesterol and 10–13% isocholesterol. This is added to mixtures of paraffins (hard, so white/yellow soft or liquid) to produce the required consistency. The inclusion of wool alcohols (5% w/w) results in a 300% increase in the concentration of water that may be incorporated into paraffin bases.

Beeswax (white or yellow)

Beeswax is a wax that consists of esters of aliphatic alcohols (C_{24}–C_{36} even numbers) and linear aliphatic fatty acids (up to C_{36}, even numbers) that is combined with paraffins to produce non-emulsified bases. White beeswax is the bleached form of yellow beeswax.

Water in oil emulsions

Ointment bases in this category can accommodate a greater concentration of water but yet can still provide similar performance to that provided by non-emulsified bases with respect to, e.g. occlusion, spreading properties. A common excipient that is employed in the formulation of this type of ointment base is *hydrous lanolin*, which is a mixture of lanolin and circa 25–30% water. It is incorporated into paraffins and oils to produce a base that can incorporate the subsequent addition of an aqueous phase. The water content of bases that have been formulated using hydrous lanolin is significant, e.g. Oily Cream BP is a water in oil emulsion ointment base that is composed of wool alcohols (50% w/w) and water (50% w/w).

Water-miscible/removable bases

These are water-miscible bases that are used to form oil in water emulsions for topical applications. The use of these bases offers a number of advantages, including:

- They are able to accommodate large volumes of water, e.g. aqueous solutions of drug, excess moisture at the site of application, e.g. exudate from abrasions and wounds.
- They are not occlusive.
- They may be easily washed from the skin and from clothing. Furthermore, they may be readily applied to (and removed from) hair.
- They are aesthetically pleasing.

The British Pharmacopoeia describes three water-miscible/removable bases:

1. emulsifying ointment
2. cetrimide emulsifying ointment
3. cetomacrogol emulsifying ointment.

Each of these contains:
liquid paraffin 20% w/w
white soft paraffin 50% w/w
anionic, cationic or non-ionic emulsifying wax 30% w/w.
As may be observed, an important component of this ointment base is emulsifying wax, of which there are three types: (1) anionic; (2) non-ionic; and (3) cationic. The important properties of these waxes are as follows:

Anionic emulsifying wax

- This is a waxy solid that, when incorporated into a paraffin base, may be used to produce an oil in water emulsion, e.g. Aqueous Cream BP (which contains 10% w/w anionic emulsifying wax).
- Anionic emulsifying wax is composed of:
  cetostearyl alcohol 90 g
  sodium lauryl sulphate 10 g
  purified water 4 ml.

Non-ionic emulsifying wax

- This is also referred to as *Cetomacrogol Emulsifying Wax BP* and is composed of:
  cetostearyl alcohol 800 g
  cetomacrogol 1000 (macrogol cetostearyl ether 22) 200 g.

Cationic emulsifying wax

- This is also referred to as *Cetrimide Emulsifying Wax BP.*
- Cationic Emulsifying Wax BP is composed of:
  cetostearyl alcohol 900 g
  cetrimide 100 g.

Water-soluble bases

The reader will have observed that the three previous ointment bases are predominantly hydrophobic, are hydrophobic with added surface-active agents or are water-miscible, containing both water-soluble and insoluble components. In contrast, water-soluble bases are composed entirely of water-soluble ingredients.

The advantages of the use of these bases include:

- They are non-greasy and may be easily removed by washing.
- They are miscible with exudates from inflamed sites.
- They are generally compatible with the vast majority of therapeutic agents.

Water-soluble bases are predominantly prepared using mixtures of different molecular weights of polyethylene glycol (Figure 4.1) to produce the required ointment consistency. Lower average molecular weights of this polymer (200, 400 and 600 g/mol) are liquids. As the average molecular weight increases, the consistency of this polymer changes from a liquid to a waxy solid ($\geq$ 1000 g/mol).
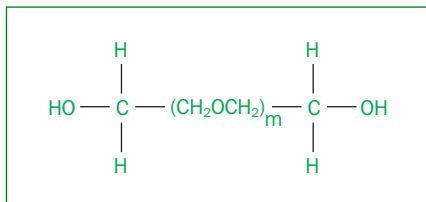
Figure 4.1 Structural formula of polyethylene glycol (m refers to the number of repeating ethylene oxide groups).

Blends of 60% w/w polyethylene glycol 400 (a liquid) and 40% w/w polyethylene glycol 4000 (a solid) have been used as a water-soluble ointment base. If required, the consistency may be increased by lowering the ratio of polyethylene glycol 400 to polyethylene glycol 4000 in the ointment base. Blending the two polyethylene glycol fractions is performed by heating the mixture followed by cooling of the homogeneous liquid at a controlled rate.

The main disadvantage associated with water-soluble bases is their inability to incorporate large volumes of aqueous solutions as these will soften and, if the concentration of water is large enough (> 5% w/w), dissolve the ointment base. Therefore the use of these bases is usually reserved for the incorporation of solid therapeutic agents. However, these bases may incorporate up to 25% of an aqueous solution if a portion of the lower-molecular-weight polyethylene glycol is replaced with stearyl alcohol. This will enhance the mechanical properties of the ointment.

Miscellaneous excipients used in the formulation of ointments and pastes

This chapter has described various strategies for the formulation of bases for ointment and paste formulations. In these the therapeutic agent may be directly incorporated as a solid component or, in the case of the absorption and water-miscible bases, the addition may be in the form of a solution. This solution may be aqueous, alcoholic (e.g. propylene glycol, glycerol) or hydroalcoholic and must not adversely affect the physical stability and/or appearance of the formulated product.

Tips

The choice of ointment base employed in the formulation of ointments is dependent on the proposed use of the dosage form and other formulation factors, e.g. stability of the therapeutic agent and the capacity of the formulation for water.

The properties of ointment bases range from highly hydrophobic (e.g. hydrocarbon bases) to water-miscible systems.

Hydrophobic ointment bases should not be applied to exuding areas due to poor water uptake capacity of these dosage forms. Conversely, ointments prepared using water-miscible bases may be applied to such sites.

Other excipients may be included in ointments and pastes, including: (1) additional/alternative solvents; (2) preservatives; and (3) antioxidants.

Additional/alternative solvents

These are hydrophobic liquid components that may be added to ointment bases (predominantly hydrophobic or absorption bases). Examples of these include: (1) liquid silicone; (2) vegetable oils; and (3) organic esters.

Liquid silicone (polydimethylsiloxane)

This may be used in barrier ointments due to the water-repellent properties of this component.

Vegetable oils

Vegetable oils may be used either to replace mineral oils or, alternatively, may be added to hydrophobic or absorption bases to increase the emollient properties of the formulated product. Examples of oils that are used for this purpose are coconut oil and arachis oil.

Organic esters

These may be used partly to replace a mineral oil to enhance the spreadability and to enhance drug dissolution within the ointment base. One of the most commonly used examples is isopropyl myristate.

Preservatives

Topically applied ointments and pastes are not sterile products; however, they are manufactured under clean conditions to minimise the microbial bioburden within the formulated product. Ointments/pastes that do not contain water do not usually require the addition of a preservative (due to the low water activity in the formulation). However if the product contains water, then a preservative will be required. Preservatives that may be used in formulations designed for external use include:

- phenolics: phenol (0.2–0.5%), chlorocresol (0.075–0.12%)
- benzoic acid and salts (0.1–0.3%)
- methylparabens (methylparahydroxybenzoic acid) (0.02–0.3%)
- propylparabens (methylparahydroxybenzoic acid) (0.02–0.3%) (and their mixtures)
- benzyl alcohol (≤ 3.0%)
- phenoxyethanol (0.5–1.0%) (Figure 4.2).
- bronopol (0.01–0.1%, usually 0.02%) (Figure 4.3).
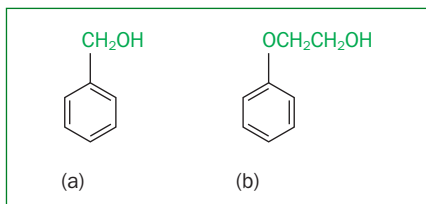
Figure 4.2 Structural formulae of (a) benzyl alcohol and (b) phenoxyethanol.
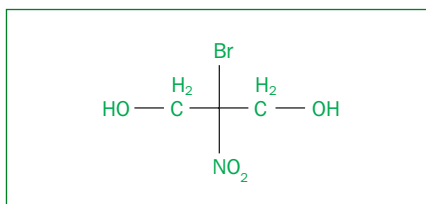
Figure 4.3 Structural formula of bronopol.

In the preservation of ointments, the same physicochemical and microbiological principles exist and therefore partitioning of the preservative from the aqueous to the non-aqueous phase may occur. Under these circumstances it is important to ensure that the required concentration ($\geq$ minimum inhibitory concentration) of the antimicrobial species is present within the aqueous phase.

Antioxidants

The use of antioxidants has been described in previous chapters. In pharmaceutical ointments antioxidants are employed to prevent or reduce oxidation of either the non-aqueous components of the ointment base (e.g. mineral/vegetable oils) and/or the therapeutic agent. The types of preservatives used for this purpose include:

- lipophilic antioxidants (to be dissolved within the non-aqueous vehicle), e.g. butylated hydroxyanisole (0.005–0.02%), butylated hydroxytoluene (0.007–0.1%), propyl gallate ($\leq$ 1%)
- hydrophilic antioxidants (to be dissolved in the aqueous phase), e.g. sodium metabisulphate (0.01–0.1%), sodium sulphite (0.1%).

Tips

Ointments, similar to emulsions and creams, contain a range of excipients that are required to stabilise the formulation.

The formulator must ensure that no interactions occur between the excipients.

The use of pastes is generally reserved for certain topical conditions, e.g. the treatment of warts.

Manufacture of ointments and pastes

The manufacture of ointments and pastes is similar to that described for emulsions and creams. The most straightforward example involves the dispersal of the powdered therapeutic agent into the preheated hydrocarbon base using a mechanical mixer.

Heat is required to lower the viscosity of the base, thereby facilitating the mixing of the solid drug.

If the therapeutic agent is incorporated into the ointment base as a separate liquid phase, the hydrophobic components and hydrophilic components are separately dissolved in the lipophilic and hydrophilic liquid phases, respectively (again with the aid of heating and mechanical mixing). In general (following dissolution of the various components), the two phases are maintained at circa 70°C and then mixed together (with stirring). The mixing of the two phases may be performed by:

- mixing the two phases simultaneously
- adding the aqueous phase to the non-aqueous phase.

Following complete mixing, the temperature of the formulation is gradually reduced to room temperature.

Pharmaceutical lotions, liniments, collodions and paints

General description

Pharmaceutical lotions, liniments, collodions and paints are external, liquid-based formulations that are applied externally for the treatment of local conditions, e.g. inflammation, acne, infection (bacterial, fungal, viral and parasitic). Although the clinical use of these classes of formulation is relatively minor, they still represent a formulation option to the pharmaceutical scientist.

Lotions

Lotions are formulated either as solutions or suspensions and, in addition to the therapeutic agent(s), may contain:

- *alcohol*: this acts as a coolant (due to evaporation following application) and as a co-solvent.
- *humectants*: these act to retain moisture on the skin after application. The most commonly used example is glycerol.
- *vehicle*: lotions are aqueous formulations and therefore will contain *purified water* (with or without the addition of buffer salts).
- *preservatives*: examples of these have been described previously in this chapter.
- *components to stabilise the suspended therapeutic agent*: if the lotion has been formulated as a suspension, agents are required to maintain the physical stability of the formulation (see Chapter 2).

Liniments

Liniments are alcohol- or oil-based solutions that are applied externally to unbroken skin with gentle rubbing. There are two

types of formulation bases that are used in the formulation of liniments: (1) alcohol-based liniments; and (2) oil-based liniments. Alcohol-based liniments act as counterirritants and rubefacients (causing reddening of the skin) and may act to increase the penetration of the drug through the skin. In addition, these formulations will provide a cooling effect due to evaporation of the alcohol base. Conversely, oil-based liniments are employed for conditions in which a massage effect is required. Typical oils used for this purpose are arachis oil and cottonseed oil.

Liniments are normally employed for the treatment of inflammatory conditions, e.g. sciatica, fibrositis and neuralgia. Examples of oil-based liniments include Camphor Liniment BP and Methyl Salicylate Liniment BP. Soap Liniment BPC is an example of an alcohol-based liniment.

In general, no other excipients are used in the formulation of liniments.

Collodions

Collodions are solutions of pyroxylin (a nitrated cellulose, predominantly cellulose tetranitrate, that is obtained following the treatment of defatted cellulose with nitric and sulphuric acids), castor oil and colophony dissolved in an organic solvent (composed of alcohol and ether). These are normally applied to dry skin using a brush applicator and, following the evaporation of the solvent, will form an occlusive film. Collodions may contain therapeutic agents, e.g. colliodion and salicylic acid collodion.

Collodion

This is a solution of pyroxylin in a solvent composed of ether (3 parts) and alcohol (1 part). This forms an inflexible, mechanically strong film on the skin and is normally used to seal abrasions. The film may be rendered more flexible by adding 2% camphor and 3% castor oil to the above formulation (termed flexible collodion). The oil acts as a plasticiser (thereby facilitating the use of the product over flexible areas) whereas the presence of camphor renders the films waterproof.

Salicylic acid collodion

This is a solution of salicylic acid (10%) in flexible collodion that is used for the treatment of warts.

Paints

These are aqueous, hydroalcoholic, alcoholic or organic solutions of a therapeutic agent that are applied topically. The use of paints nowadays is limited due to the emergence of more elegant dosage forms.

Pharmaceutical gels

General description

Pharmaceutical gels are semisolid systems in which there is interaction (either physical or covalent) between colloidal particles within a liquid vehicle. The vehicle is continuous and interacts with the colloidal particles within the three-dimensional network that is formed by the bonds formed between adjacent particles. The vehicle may be aqueous, hydroalcoholic, alcohol-based or non-aqueous. The colloidal particles may be dispersed solids, e.g. kaolin, bentonite or, alternatively, dispersed polymers. *Xerogels* are gels in which the vehicle has been removed, leaving a polymer network, e.g. polymer films.

There are two main categories of pharmaceutical gels, based on the nature of the three-dimensional network of particles: (1) dispersed solids and (2) hydrophilic polymers.

Gels based on dispersed solids

As discussed in Chapter 2, under certain conditions dispersed solids will undergo flocculation. If flocculation extends throughout the system a continuous solid particle network is established, with the liquid vehicle dispersed in the void volume between the particles. The nature of the interaction between the particles in the network may be van der Waals interactions (at the secondary minimum), e.g. *Aluminium Hydroxide Gel USP*. However, for certain dispersed solids the nature of the interaction is electrostatic bonding. Examples of the particles that exhibit this type of interaction include kaolin, bentonite and aluminium magnesium silicate. The particles exhibit a plate-like crystal structure in which there are electronegative regions along the flat face of the crystal (due to O^-) and electropositive regions (due to the ionised aluminium and magnesium ions) at the edges of the plates. The interaction of these two regions facilitates the establishment of a structured 'house of cards'-type particle network. The bond strength between the particles is weak: interparticle bonds are broken by the application of relatively low shearing stresses (such as those that occur whenever the product is shaken), thereby liberating the individual particles. Following removal of the stress the bonds between the particles will reform and hence the rheological structure of these systems is recovered. This time-dependent recovery of the rheological structure (that was lost upon shaking) is termed *thixotropy*.

Gels based on hydrophilic polymers

Pharmaceutical gels are most commonly (but not exclusively) manufactured by dispersing hydrophilic polymers within an appropriate aqueous vehicle. When dissolved within an aqueous

phase, hydrophilic polymers behave as lyophilic colloids and their unique physical properties result from the self-association of the dissolved polymer and its interaction with the aqueous medium. There are two types of self-association (termed irreversible and reversible) that may be demonstrated by lyophilic colloids and this allows gels that are manufactured from lyophilic colloids to be classified as either type 1 or type 2 gels.

Type 1 gels

In type 1 gels (often termed *hydrogels*) the interaction between the polymer chains is covalent and is mediated by molecules that cross-link the adjacent chains (termed cross-linkers). A diagrammatic representation of the covalent interactions within a type 1 (chemical) is shown in Figure 4.4.

Figure 4.4 Diagrammatic representation of polymer–polymer interactions within a type 1 (chemical) gel.
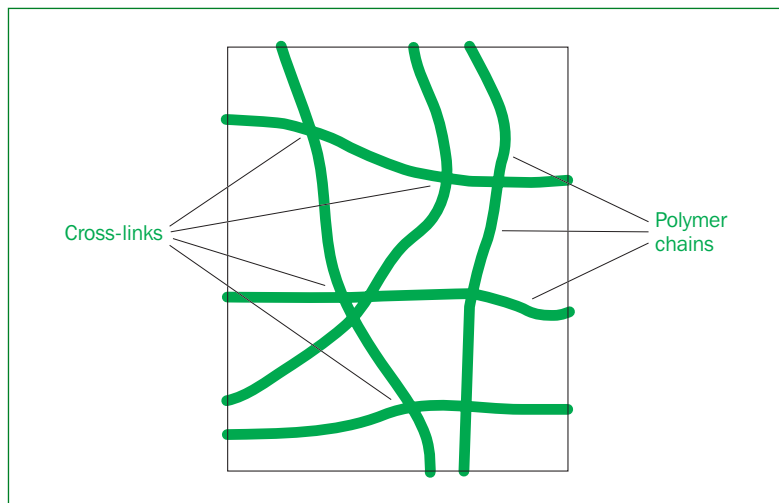
An example of the chemical structures of a cross-linked hydrogel and the monomer (hydroxyethylmethacrylate) and cross-linker (ethyleneglycol dimethacrylate) used in the synthesis of the hydrogel is provided in Figure 4.5 below. In this diagram the points of intersection of the polymer chains are covalent cross-links.

These gels exhibit unique physicochemical properties, including:

- The ability to absorb a considerable mass of aqueous fluid (often 100 times the original mass) whilst still retaining a three-dimensional structure.
- Hydrogels exhibit robust mechanical properties, being resistant to fracture following exposure to stresses frequently up to 1 kPa. Moreover, hydrogels exhibit excellent flexibility.

Figure 4.5 Structural formula of (a) hydroxyethylmethacrylate, (b) ethyleneglycol dimethacrylate and (c) poly(hydroxyethylmethacrylate) that has been cross-linked using ethyleneglycol dimethacrylate.
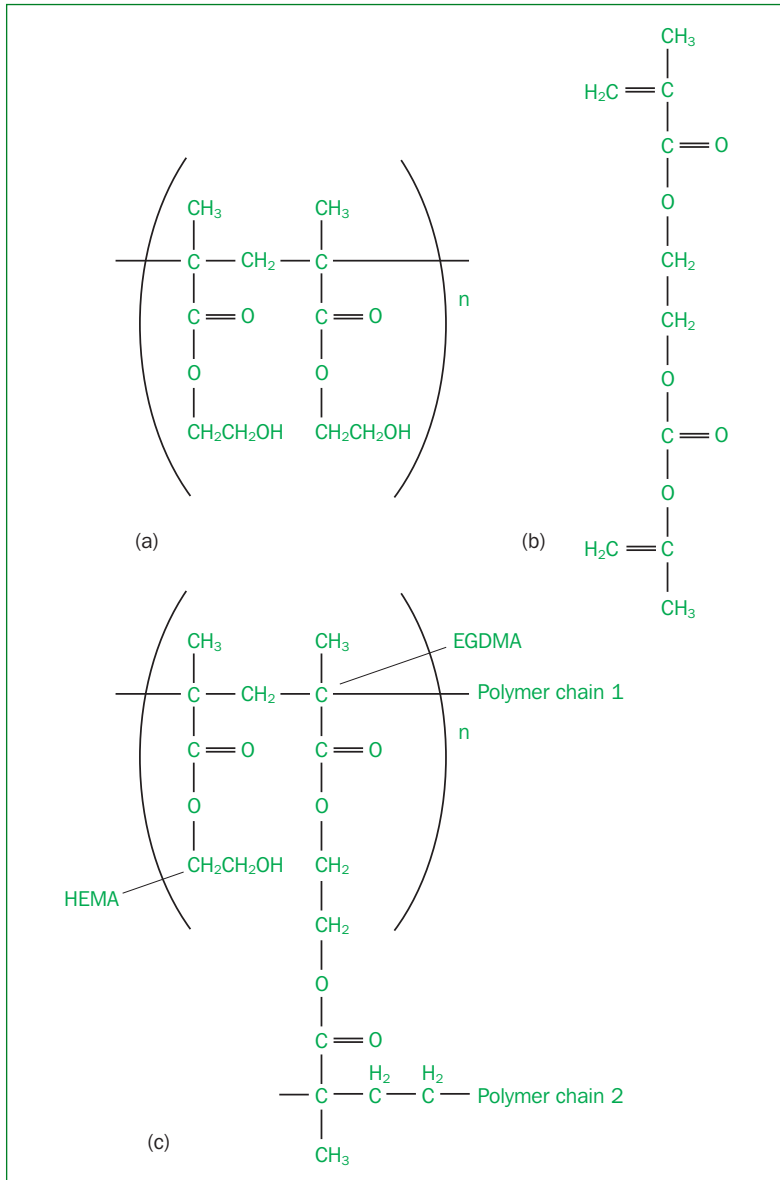
Conversely, xerogels (hydrogels from which the aqueous phase has been removed by drying) are brittle. In this case the absorbed solvent acts as a plasticiser. Unlike type 2 gels (see below), type 1 gels do not exhibit flow when exposed to an applied stress due to the inability of the stress to overcome

(destroy) the covalent bonds. Under these conditions, the elastic properties of type 1 gels enable the applied energy to be stored and utilised (after the stress is removed) to return the polymer chains to their equilibrium position.

Due to this ability to absorb a large mass of fluid (whilst retaining their mechanical properties), hydrogels are clinically used as wound dressings, as lubricious coatings on urethral catheters and as soft contact lenses. In addition, hydrogels may be used for the controlled delivery of therapeutic agents at the site of implantation.

Type 2 gels

In type 2 gels the interactions between the polymer chains are reversible and are facilitated by weaker bonds, e.g. hydrogen bonding, ionic association or van der Waals interactions. The application of stresses to type 2 gels will end in the temporary destruction of these bonds, thereby enabling the formulation to flow. As a result, type 2 gels are rheologically referred to as *pseudoplastic (shear-thinning)* systems. Following the removal of the stress, the intermacromolecular bonds are reformed and the viscosity of the formulation returns to its equilibrium value.

A diagrammatic representation of the interactions that occur in type 2 (physical) gels is shown in Figure 4.6. As may be observed, the areas where adjacent polymer chains interact are referred to as junction zones and, in practice, a substantial fraction of the polymer is involved in polymer–polymer interactions at these zones.

Figure 4.6 Diagrammatic representation of the interactions between polymer chains in a type 2 gel.
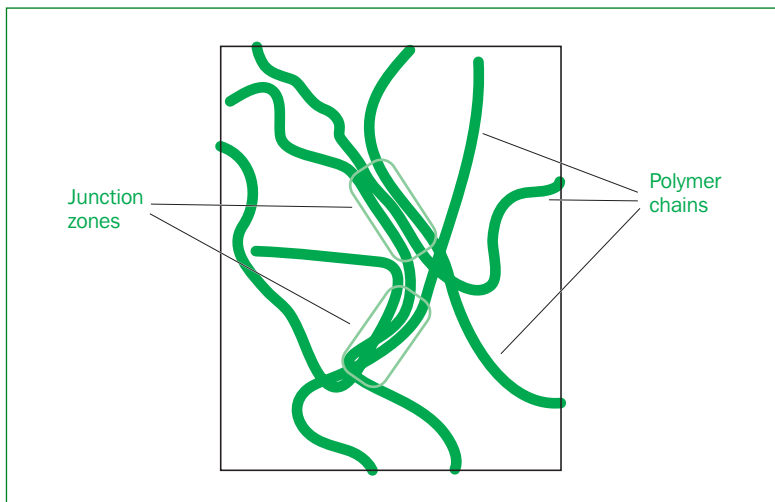
The overwhelming majority of pharmaceutical gels are type 2 gels and typically the following polymers are employed in the formulation of these systems: (1) cellulose derivatives; (2) polysaccharides derived from natural sources; and (3) polyacrylic acid.

Cellulose derivatives
The cellulose derivatives represent a family of chemically related polysaccharides that are structurally derived from cellulose (following the appropriate chemical substitution). The most commonly used examples from this series that are used to formulate pharmaceutical gels include:
- methylcellulose
- hydroxyethylcellulose
- hydroxypropylcellulose
- sodium carboxymethylcellulose.

The structural formulae of these polymers are presented in Figure 4.7.

Polysaccharides derived from natural sources
Polysaccharides that have been derived from natural sources are commonly used as the basis for pharmaceutical gels. Examples of these include: (1) carrageenan; and (2) alginic acid/sodium alginate.

Carrageenan
This is a family of polysaccharides that is derived from red seaweed. There are three chemically related carrageenans, termed lambda, iota and kappa which differ according to the location of sulphate groups and the presence or absence of anhydrogalactose. Kappa carrageenan exhibits excellent gelling properties (due to the presence of a tertiary helical structure); iota carrageenan (but not lambda carrageenan) also displays gelling (albeit weaker) properties. The typical range of concentrations of kappa carrageenan used to form pharmaceutical gels is 0.3–1.0% w/w.

Alginic acid/sodium alginate
Alginic acid is a polysaccharide that is derived from algae (Phaecophyceae family). Addition of calcium ions to a solution of alginic acid will result in an electrostatic interaction, producing a viscous gel at low concentrations of calcium and a cross-linked polymer at higher concentrations. Alginic acid is incompatible with basic drug molecules.

Poly(acrylic acid)
Poly(acrylic acid) (Figure 4.8) is a synthetic polymer that is produced following the polymerisation of acrylic acid and cross-linking with either allyl sucrose or allyl ethers of pentaerythritol.

Figure 4.7 Structural formulae of cellulose derivatives commonly used to formulate pharmaceutical gels. (a) Methylcellulose; (b) hydroxyethylcellulose; (c) hydroxypropylcellulose; (d) sodium carboxymethylcellulose. Typically concentrations within the range of 1–10% w/w are required to produce pharmaceutical gels.

(a)

(b)

(c)

(d)
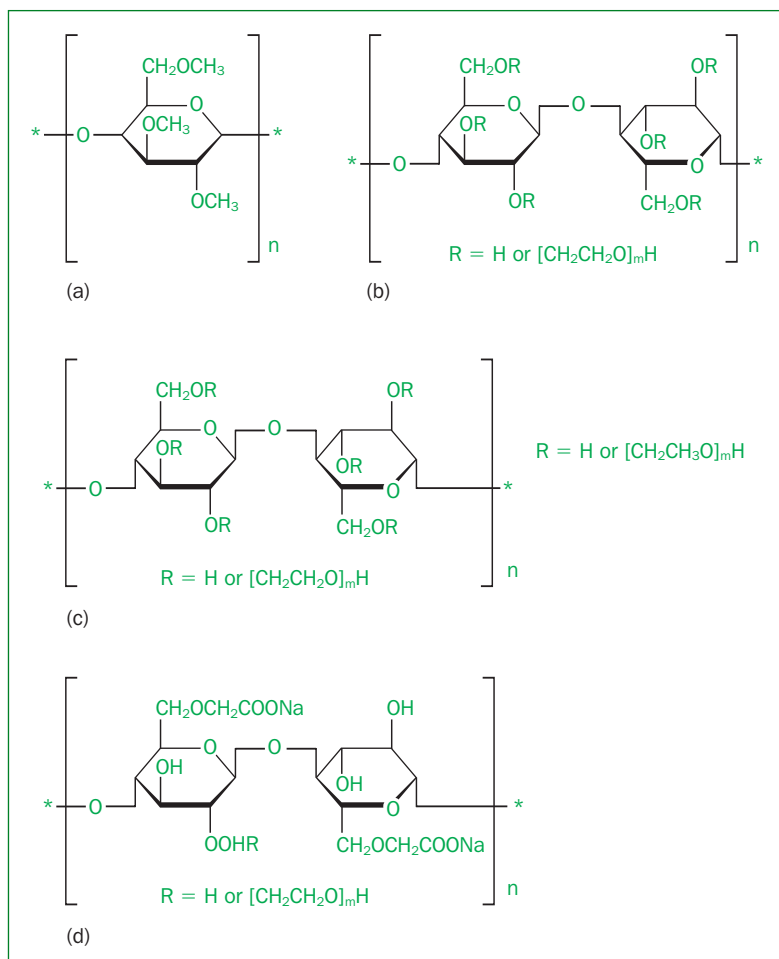
In water polyacrylic acid exists as aggregated (coiled) colloidal particles of minimal viscosity (pH circa 3). However, if the pH of the system is neutralised by the addition of an appropriate base, e.g. triethanolamine, triethylamine or sodium hydroxide, the pendant carboxyl groups will ionise, resulting in expansion of the polymer chains due to repulsion of the adjacent ionised groups. In so doing the viscosity of the formulation is dramatically increased. Typically pharmaceutical gels are produced using 0.5–2.0% w/w poly(acrylic acid) that has been neutralised with an appropriate base. Incompatibilities exist between poly(acrylic acid) and basic therapeutic agents.

Figure 4.8 Structural formula of poly(acrylic acid). R refers to allylsucrose or pentaerythritol. The subscripts refer to a number of repeating units of acrylic acid that reside between the cross-links.
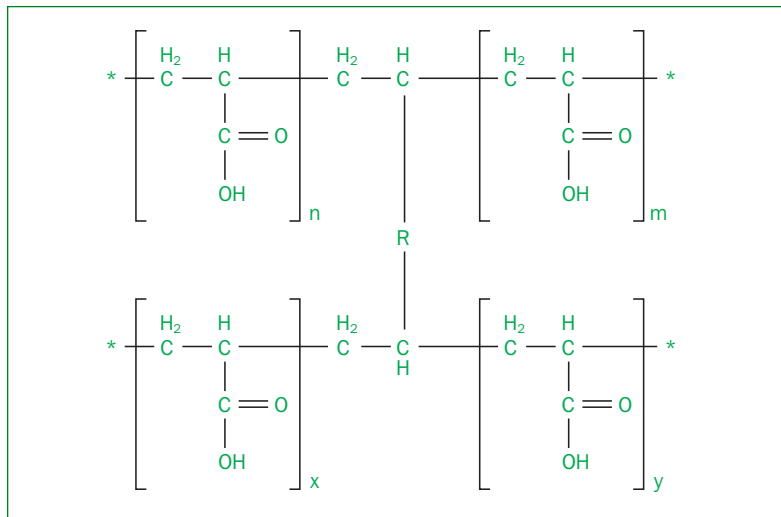
Furthermore, the viscosity of gels prepared using poly(acrylic acid) is adversely affected by medium/high concentrations of electrolytes.

Factors affecting gelation of type 2 gels

Gelation in type 2 gels occurs whenever a sufficient number of polymer–polymer interactions (junction zones) occur. However, both the mechanism of gelation and the number (frequency) of interactions are affected by physicochemical and environmental factors, as outlined below.

Concentration of hydrophilic polymer

At low concentrations, solutions of hydrophilic polymers exhibit Newtonian flow due to the limited number of polymer–polymer interactions. As the concentration of polymer increases, the number of polymer–polymer interactions increases and eventually, at a defined polymer concentration, the flow properties of these systems become non-Newtonian (termed the gel point). Further increases in the concentration of polymer lead to an increase in the number of junction zones and hence the resistance to deformation from an applied stress (the viscosity) increases. Therefore, the physicochemical and rheological properties of a pharmaceutical gel may be readily manipulated by altering the concentration of hydrophilic polymer.

Molecular weight of the polymer

As the molecular weight of the hydrophilic polymer increases (at a defined concentration of polymer), there are a greater number of available sites on the polymer chains that may engage in polymer–polymer interactions. As a result the viscosity of the formulation increases.

Nature of the solvent

In solvents that are described as 'good solvents', the chains of a polymer will exist in the expanded state. Conversely, in the presence of a poor solvent, the polymer chains will exist in a non-expanded (coiled) state. The viscosity of a polymer solution is dependent on the expansion of the polymer chains. Therefore, the concentration of polymer that results in gel formation and the physicochemical (rheological) properties of the gel are dependent on the solvent system into which the hydrophilic polymer is dissolved. In poor solvents gelation will not occur.

pH of the solvent

As previously discussed in this chapter, the pH of the solvent directly affects the ionisation of acidic or basic polymers which, in turn, affects the conformation (expansion) of the polymer chains. In the non-ionised state acidic and basic polymers exist in a coiled (non-expanded) state and gelation does not occur. The rheological properties of ionic polymers are optimal with a range of pH values at which maximum expansion of the polymer chains occurs. The rheological properties of non-ionic polymers are unaffected by the pH of the solvent, usually over a large pH range (circa 4–10).

Ionic strength of the solvent phase

The rheological properties of both non-ionic and (in particular) ionic polymers are affected by the ionic strength of the solvent. At high concentrations of electrolytes (and hence large ionic strength), non-ionic polymers may be 'salted out' of solution due to desolvation of the polymer chains. Conversely, at lower concentrations of electrolyte, shielding of the charge on the pendant groups of the ionic polymer by a counterion will occur. This will therefore reduce the capacity of the polymer to interact with the solvent and hence the rheological properties of the gel will be compromised. If the concentration of electrolyte is sufficiently large, salting out of the ionic polymer will result.

Temperature

Certain hydrophilic polymers may undergo a thermally induced transition that results in an increase in the rheological properties. Two examples of this are solutions of methylcellulose and

hydroxypropylcellulose which have been reported to undergo gelation at elevated temperatures (circa 50–60°C). Whilst this transition has limited biological relevance, one polymer system, poly(oxyethylene)-poly(oxypropylene) block co-polymers (the Pluronic or Synperonic series) undergoes a thermal transition within a biologically useful temperature range ($< 37°C$). At temperatures below this (sol–gel) transition temperature ($T_{sol/gel}$), solutions of this polymer exhibit Newtonian flow and low viscosity (the sol state). Conversely, above $T_{sol/gel}$ the polymer sol is converted into a gel with pronounced elasticity and viscosity. In solution at temperatures below $T_{sol/gel}$ and above the critical micelle concentration, the polymer exists in the micellar state. Elevation of the temperature (to above the $T_{sol/gel}$) results in the further production of micelles and (close) intermicellar aggregation. This results in a gel of pronounced rheological structure. Lowering the temperature of the system to below the $T_{sol/gel}$ will result in deaggregation of the micelles and the re-emergence of the sol (low-viscosity) state. The ability to modulate the rheological structure of these gels in the manner described has led to an interest in their use as drug delivery systems within the oral cavity and rectum.

Ionic gelation

Certain hydrophilic polymers may undergo gelation in the presence of inorganic metal ions. Examples of these include:

- The gelation of polyhydroxypolymers, e.g. poly(vinyl alcohol) may occur in the presence of suitable anions, e.g. borate, permanganate. Poly(vinyl alcohol) is known to form structured gels in the presence of borate anions. The mechanism of the interaction between the polymer and borate anions is shown in Figure 4.9. The gels formed by this mechanism exhibit excellent mechanical strength, due to the borate anion-mediated cross-links. A non-pharmaceutical application of this interaction is the children's toy Kids Slime.
- As highlighted previously, gelation of alginic acid occurs in the presence of positively charged di/trivalent ions, e.g. Mg^{2+}, Ca^{2+}, Al^{3+}.

Formulation considerations for pharmaceutical gels

There are several formulation considerations open to the pharmaceutical scientist concerning the formulation of pharmaceutical gels. These include: (1) the choice of vehicle; (2) the inclusion of buffers; (3) preservatives; (4) antioxidants; (5) flavours/sweetening agents; and (6) colours.
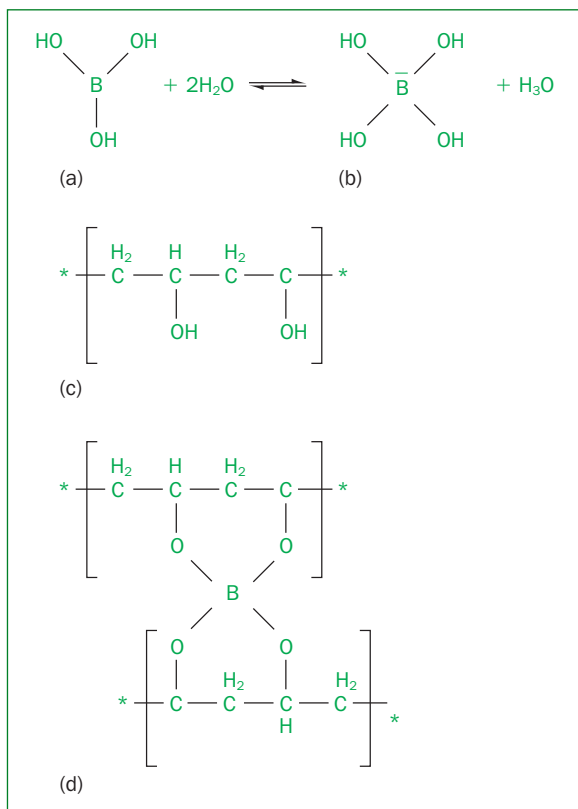
Figure 4.9 Diagrammatic representation of the interaction between borate anions and poly(vinyl alcohol) that leads to gel formation. (a) Boric acid; (b) borate anion; (c) poly(vinyl alcohol); (d) the interaction between poly(vinyl alcohol) and borate anions.

The choice of vehicle

Purified water is the normal solvent/vehicle used in the formulation of pharmaceutical gels. However, co-solvents may be used, e.g. alcohol, propylene glycol, glycerol, polyethylene glycol (usually polyethylene glycol 400) to enhance the solubility of the therapeutic agent in the dosage form and/or (in the case of ethanol) to enhance drug permeation across the skin.

If the drug has poor chemical stability and/or poor solubility in water or water-based vehicles, pharmaceutical gels may be formulated using polyhydroxy solvents, e.g. propylene glycol, glycerol, polyethylene glycol 400 and polyacidic polymers, e.g. poly(acrylic acid). In these systems gelation is facilitated by hydrogen bonding between the hydroxyl and carboxylic acid groups and this results in: (1) expansion of the pendant groups on the polymer chain and (2) non-covalent cross-linking of adjacent polymer chains.

Tips

The main difference between type 1 and type 2 gels is the nature of the cross-links between adjacent polymer chains. In type 1 gels the cross-links are covalent, whereas in type 2 gels the cross-links are non-covalent (secondary).

The vast majority of gels used as dosage forms are type 2. The use of type 1 gels is reserved for wound dressings.

The commonly used polymers for the formulation of type 2 gels are cellulose derivatives, alginates and poly(acrylic acid). Gels are normally formed by increasing polymer concentration and, if the polymer is ionic, by altering the pH.

The inclusion of buffers

As in other pharmaceutical formulations, buffers (e.g. phosphate, citrate) may be included in aqueous and hydroalcoholic-based gels to control the pH of the formulation. It should be noted that the solubility of buffer salts is decreased in hydroalcoholic-based vehicles.

Preservatives

Pharmaceutical gels require the inclusion of preservatives and, in general, the choice of preservatives is similar to that described for ointments and pastes in the early sections of this chapter. It should be remembered that certain preservatives, e.g. parabens, phenolics, interact with the hydrophilic polymers used to prepare gels, thereby reducing the concentration of free (antimicrobially active) preservative in the formulation. Therefore, to compensate for this, the initial concentration of these preservatives should be increased.

Antioxidants

As in other formulations, antioxidants may be included in the formulation to increase the chemical stability of therapeutic agents that are prone to oxidative degradation. The choice of antioxidants is based on the nature of the vehicle used to prepare the pharmaceutical gel. Therefore, as the majority of pharmaceutical gels are aqueous-based, water-soluble antioxidants, e.g. sodium metabisulphite, sodium formaldehyde sulphoxylate, are commonly used.

Flavours/sweetening agents

Flavours and sweetening agents are only included in pharmaceutical gels that are designed for administration into the oral cavity, e.g. for the treatment of infection, inflammation or ulceration. As before, the choice of sweetener/flavouring agents is dependent on the required taste, the type and concentration selected to mask the taste of the drug substance efficiently. Examples of flavours/sweetening agents used for this purpose have been detailed in Chapter 1.

Colours

Colours, e.g. those described in Chapter 1, may be (but are not usually) added into pharmaceutical gels.

Manufacture of pharmaceutical gels

In the manufacture of pharmaceutical gels, generally the water-soluble components/excipients are initially dissolved in the vehicle in a mixing vessel with mechanical stirring. The hydrophilic polymer must be added to the stirred mixture slowly

to prevent aggregation and stirring is continued until dissolution of the polymer has occurred. It should be noted that excessive stirring of pharmaceutical gels results in entrapment of air. Therefore, to prevent this the mixing rate must not be excessive or a mixing vessel may be used to which a vacuum may be pulled, thereby removing air.

Multiple choice questions

1. **With respect to pharmaceutical gels, which of the following are true?**
a. Pharmaceutical gels reflect light, causing the product to have a 'creamy white' appearance.
b. The structural properties of the gel are specifically due to the high water content of the formulation.
c. Pharmaceutical gels are frequently formulated using long-chain hydrocarbons.
d. They may contain surface-active agents to enhance the solubility of the therapeutic agent.

2. **With respect to ointments, which of the following are true?**
a. The bases of hydrocarbon ointments are typically derived from petroleum.
b. Hydrophobic ointment bases are unsuitable for exuding lesions.
c. An ointment may be formed by fusion of silicone oil and a wax.
d. Drug solubility in an ointment base may be modified by the inclusion of a co-solvent such as propylene glycol.

3. **Concerning pharmaceutical ointments, which of the following are true?**
a. Ointment may stain patients' clothes.
b. Ointments are generally highly viscous formulations.
c. Medicated ointments may be used to treat hamorrhoids.
d. Drugs that are prone to hydrolysis should not be formulated as ointments.

4. **With respect to ointment formulations, which of the following are true?**
a. Ointment formulations may be water-miscible.
b. Ointments prepared using hydrocarbon bases require the inclusion of preservatives.
c. Ointments may be formulated as emulsions.
d. Ointments may require the addition of antioxidants to enhance the stability of the therapeutic agent.

5. Concerning ointments, which of the following are true?

a. Ointments are structurally similar to creams.

b. An ointment base consists of two components, which are liquids at room temperature.

c. Hydrocarbon ointments are mixtures of C_{30}–C_{50} hydrocarbons and C_{16}–C_{30} hydrocarbons.

d. Cosmetically, ointments are unsatisfactory, being greasy and difficult to remove.

6. With respect to paste formulations, which of the following are true?

a. Pastes contain high drug loadings.

b. Pastes may not be applied to exuding wounds.

c. Pastes are opaque and therefore used as sunblock formulations.

d. Pastes are cosmetically acceptable formulations.

7. With respect to pharmaceutical lotions, which of the following are true?

a. Pharmaceutical lotions may be formulated as solutions or suspensions, with solutions being preferred.

b. Pharmaceutical lotions are typically used for the treatment of local conditions.

c. Pharmaceutical lotions are principally aqueous formulations.

d. Pharmaceutical lotions do not require the inclusion of preservatives.

8. With respect to pharmaceutical liniments, which of the following are true?

a. Pharmaceutical liniments are formulated as solutions or suspensions.

b. Pharmaceutical liniments are typically used for the treatment of local conditions.

c. Pharmaceutical lotions are aqueous formulations.

d. Pharmaceutical lotions may be rubefacient, thereby increasing drug penetration across the skin.

9. Which of the following statements are true concerning type 1 pharmaceutical gels?

a. The mechanical properties of the gels are primarily due to chemical cross-linking.

b. Type 1 gels may be formulated to exhibit large swelling ratios in biological fluids.

c. Type 1 gels are easily dispensed from an ointment tube.

d. Type 1 gels are plasticised by water.

10. Which of the following statements are true concerning type 2 pharmaceutical gels?

a.   Type 2 pharmaceutical gels are pseudoplastic in nature.

b.   Type 2 gels require the inclusion of preservatives.

c.   Gelation in type 2 gels may be performed by increasing the concentration of hydrophilic polymer within the aqueous vehicle.

d.   Type 2 gels may only be formulated using water as the vehicle.

chapter 5
Parenteral formulations

Overview

In this chapter we will:
- examine the types of parenteral formulations
- provide an overview of the advantages and disadvantages of parenteral formulations
- describe the formulation considerations for parenteral formulations
- briefly describe manufacturing consideration of parenteral formulations.

General description

Parenteral administration of drugs involves the injection of therapeutic agents, in the form of solutions, suspensions or emulsions, into the body. In so doing, one of the major barriers to drug entry (the skin) is breeched. Parenteral formulations have been officially recognised since the mid 19th century when morphine solution appeared in the 1874 addendum to the British Pharmacopoeia (1867). Currently many classes of drug are formulated as parenteral dosage forms and, indeed, the control of certain disease states is dependent on parenteral administration, e.g. type 1 diabetes mellitus. Parenteral products are therefore essential components of modern medicine.

Routes of parenteral administration

There are several different routes by which parenteral products may be administered and indeed there are very few, if any, organs into which parenteral dosage forms may not be injected. However, there are three routes by which parenterals are most frequently administered: (1) intravenous (IV); (2) intramuscular (IM); and (3) subcutaneous (SC). These sites are located beneath the epidermis/dermis within the skin.

KeyPoints

- Parenteral formulations are extensively used in the treatment and control of numerous disease states, e.g. diabetes, infection, pain.
- Parenteral formulations can be subdivided into three categories, namely solutions, suspensions and emulsions. Furthermore parenteral formulations may be defined as either large-volume or small-volume parenterals.
- Parenteral formulations are principally administered by three routes (intravenous, intramuscular and subcutaneous).
- The volume of injection and the requirements for onset of action primarily define the type of parenteral formulation (solution, suspension, emulsion) required.
- Parenteral formulations are *sterile* and therefore require specialist (and more expensive) manufacturing processes.
- Parenteral formulations are *pyrogen-free*.

Intravenous route

- This involves administration of the parenteral formulation into a vein, usually a large proximal vein. The veins are located beneath the subcutaneous tissue, embedded within the muscle.
- IV administration achieves a rapid and predictable response.
- It ensures 100% drug bioavailability.
- Both large- (up to 500 ml) and small- (up to 10 ml) volume formulations may be administered intravenously. Large volumes are infused into the vein at a controlled rate, e.g. total parenteral nutrition, infusion of solutions of electrolytes/nutrients either containing or devoid of drugs.
- Formulations are usually solutions or emulsions (in which the size of the disperse phase is small, <1 μm). Suspensions (or solutions that precipitate within the blood stream) must not be administered IV due to disruption of blood flow.
- Due to the subsequent dilution of the injected dose and the relative insensitivity of venous walls of veins, IV administration may be employed for the administration of drugs that would normally be too irritating if administered by other routes.
- Care must be taken regarding the rate of administration of the parenteral formulation. If the administration is performed too quickly, an excessive concentration of drug at the target organ may result, leading to drug-induced shock.
- Training is required to ensure that the dosage form is actually administered to the vein and that puncture of the vein is avoided.

Intramuscular route

- This involves administration into a muscle, usually the gluteal (buttocks), vastus lateralis (lateral thigh) or deltoid (upper arm) muscles. The musculature resides below the subcutaneous tissue (which itself lies beneath the epidermis and the dermis).
- The volume of injection is small, usually 1–3 ml or up to 10 ml in divided doses.
- Faulty injection technique may lead to local muscle damage.
- IM injection results in relatively rapid absorption, second only to IV with respect to the time taken for the onset of action. The nature of the formulation directly affects the rate of absorption of drugs administered by the IM route. Drug absorption from aqueous solutions is greater than from aqueous suspension or non-aqueous (oil-based) solutions of drugs.
- IM injections are usually used for controlled-release formulations.

Subcutaneous route

- This involves administration into the subcutaneous tissue, a layer of fat located below the dermis.
- There is a slower onset of action and sometimes less total absorption of therapeutic agents when compared to the IV or IM routes of administration. As before, the nature of the formulation directly affects the rate of drug absorption from this site: oily solutions or aqueous suspension of therapeutic agents exhibit slower drug absorption.
- The volume of injection is typically circa 1 ml; however, large-volume parenteral solutions (electrolyte or dextrose, up to 1000 ml) may be infused subcutaneously. This technique is termed hypodermoclysis and is only employed when there is difficulty in accessing a vein. On some occasions hyaluronidase may be administered to increase the available volume at the site by catalysing the temporary breakdown of the connective tissue at the site (hyaluronic acid).
- Viscous formulations are not generally administered subcutaneously.
- Typical sites of SC injection include the arms, legs and abdomen.
- SC administration is the route of choice for the administration of insulin.

Miscellaneous routes

In addition to the primary routes of administration cited above, there are other parenteral routes of administration that are not used as frequently as the IM, IV and SC routes but do play an important role in medicine. These include: (1) intradermal (ID); (2) intra-arterial (IA); (3) intrathecal (IT); (4) intradural and extradural; (5) and intracardiac (IC) routes.

Intradermal route

- This is the injection into the dermal layer of the skin.
- The ID route is generally used for diagnostic purposes, e.g. for the diagnosis of allergy and for the tuberculin test.
- Despite being a vascular site, absorption is slow and limited from this region.
- Only small volumes may be injected, circa 0.1 ml.

Intra-arterial route

- The parenteral formulation is injected into an accessible artery.
- This route requires specialist training to administer therapeutic agents as if the artery is missed, possible damage to adjacent nerves may result.

- The IA route is used to administer radiopaque media to visualise organs, e.g. heart, kidney.
- It is used to administer anticancer drugs to ensure that the highest possible concentration of drug reaches the target organ.

Intrathecal (IT) route
- This route is used to administer therapeutic agents to the cerebrospinal fluid to ensure that the appropriate concentration of drug is obtained at this site (e.g. for the treatment of infection).

Intradural and extradural routes
- Intradural and extradural administration is employed to achieve spinal anaesthesia.
- Intradural administration involves injection of the therapeutic agent within the dural membrane surrounding the spinal cord.
- Extradural administration involves injection of the therapeutic agent outside the dural membrane and within the spinal caudal canals.

Intracardiac (IC) route
- The intracardiac route involves injection of the formulation directly into the muscles of the heart.
- This route is normally used whenever there is a cardiac emergency.

Tip

The route of administration of the parenteral product directly affects the pharmacokinetic profile of the dosage form. Typically the intravenous route is employed for rapid onset of action whereas controlled-release formulations are administered by the intramuscular or subcutaneous routes.

Other more specialised routes are employed for the delivery of the therapeutic agent directly to the target site/region.

Advantages and disadvantages of parenteral formulations

Advantages
- An immediate physiological response may be achieved (usually by the IV route). This is important in acute medical situations, e.g. cardiac arrest, anaphylactic shock, asthma.
- Parenteral formulations are essential for drugs that offer poor bioavailability or those that are rapidly degraded within the gastrointestinal tract (e.g. insulin and other peptides).
- They offer a method to administer drugs to patients who are unconscious or uncooperative or for patients with nausea and vomiting (and additionally dysphagia).

- As trained medical staff primarily administer parenteral formulations, there is control of both the dosage and frequency of administration. The main exception to this is the administration of insulin, which, in the absence of complications (e.g. ketoacidosis), is performed exclusively by patients.
- Local effects may be achieved using parenteral formulations, e.g. local anaesthesia.
- Parenteral formulations provide a means by which serious imbalances in electrolytes may be corrected (using infusion solutions).
- Parenteral formulations may be readily formulated to offer a wide range of drug release profiles, including:
- rapidly acting formulations (generally drug solutions that are administered IV)
- long-acting formulations (generally drug suspensions, or solutions in which the drug is precipitated out of solution at the injection site, administered by the IM or SC routes). Examples of these include intermediate/long-acting insulin formulations and steroid injections.
- In patients who cannot consume food, total parenteral nutrition offers a means by which nutrition may be provided using specially formulated solutions that are infused into the patient.

Disadvantages

- The manufacturing process is more complicated than for other formulations due to the requirement for aseptic technique. The level of training of staff involved in the manufacture of parenteral formulations is high and often specialist equipment is required to ensure that the finished product specification is achieved.
- Skill of administration is required to ensure that the dosage form is administered by the correct route. If a parenteral suspension, which is designed for administration by the IM or SC route, is incorrectly administered by the IV route, a pulmonary microcapillary blockage may occur leading to a blockage in the flow of blood at that site.
- Parenteral formulations are associated with pain on administration.
- If the patient is allergic to the formulation (the therapeutic agents and/or the excipients), parenteral administration will result in both rapid and intense allergic reactions.
- It is difficult to reverse the effects of drugs that have been administered parenterally, even immediately after administration. This is not strictly the case with other routes of administration, e.g. oral, transdermal.

Formulation considerations for parenteral formulations

Parenteral formulations may be categorised as solutions (aqueous or oil-based), suspensions (aqueous or oil-based) or emulsions. For the most part, these formulations use similar (or indeed identical) excipients but, as may be expected, there are certain excipients that are unique to each category. Details of the key formulation excipients for each formulation type are provided below. In addition, the reader will observe a similarity in the excipients used for the formulation of solutions, suspensions and emulsions for parenteral administration and those used for non-parenteral use. The main formulation considerations for parenteral formulations are described below.

Type of preparation

The initial choice that must be made concerns the type of parenteral formulation required, a choice that is informed by the physicochemical properties of the therapeutic agent, the intended route of administration of the formulation, the volume to be administered and the general preference for a particular formulation based on the perceived pharmacological effect or onset of pharmacological effect. In some cases, the formulation scientist may be asked to formulate a specific preparation, e.g. an aqueous suspension for IM administration. However, in the absence of this information, several issues, detailed below, must be addressed.

Solubility of the therapeutic agent

With respect to the formulation of pharmaceutical products, therapeutic agents may be categorised into three groups:
1. Good solubility, in which the therapeutic agent is freely soluble in the chosen solvent (either aqueous or oil-based) at the concentration required in the parenteral product. In this case a parenteral solution is a possible formulation option.
2. Moderate solubility but insufficient to produce a solution in conventional solvents (e.g. water, oil). In this scenario, the use of co-solvents may sufficiently increase the solubility of the therapeutic agent in the vehicle to produce a parenteral solution containing the required drug concentration. This is generally the preferred strategy for therapeutic agents of moderate solubility. However, if required, the therapeutic agent may be formulated as a suspension, although one cautionary note regarding this approach is the potential recrystallisation of soluble drug during storage, a phenomenon that may affect the physical stability of the preparation.

3. Low solubility in the chosen vehicle. The preferred choice for therapeutic agents that exhibit low solubility in the chosen vehicle is a parenteral suspension formulation.

Preferred route of administration

If there is a preferred route of administration for the parenteral product, this will directly influence the nature of the parenteral product. In particular:

- IV products must be *aqueous solutions* and, furthermore, must not precipitate in the blood stream following administration. Emulsions may also be administered by this route provided the particle size of the internal phase is sufficiently small.
- Parenteral suspensions (aqueous or oil-based) and oil-based parenteral solutions must be administered either subcutaneously or intramuscularly. Aqueous solutions may also be administered intramuscularly or subcutaneously.
- There are other restrictions for those less commonly used routes of parenteral administration that are specific for each route.

Volume of dose to be administered

The volume of product to be parenterally administered will directly affect the type of formulated product.

- Large-volume parenterals (up to 500 ml) are administered intravenously (although the SC route of administration is infrequently used for this purpose).
- Small-volume parenterals may be administered by all routes (bearing in mind the restrictions of oil-based and suspension formulations).

Onset/duration of action

A wide range of predictable plasma drug concentration–time profiles may be obtained with parenteral formulations that are dependent on both the type of formulation and route of administration. Indeed, this is one of the major rationales for the use of parenteral administration of therapeutic agents. Whilst the pharmacokinetics of parenteral dosage forms is not under direct consideration in this book, it is important to summarise the effects of both formulation type and route of administration on such properties.

- Formulations administered intravenously will have an immediate pharmacological effect. The rates of drug absorption from the other main routes of administration (SC and IM) are slower.
- The absorption of therapeutic agents from aqueous solutions when administered by the IM or SC routes is faster than from

oil-based solutions, oil-based suspensions and aqueous suspensions. As a result, the slower absorption of therapeutic agents from oil-based solutions/suspensions and aqueous suspensions enables these formulations to offer a prolonged clinical effect whenever administered by the IM or SC routes. This may be illustrated by the following examples:

– When injected subcutaneously, the onset of action of soluble insulin (aqueous solution) is rapid (circa 30 minutes), peaks between 2 and 4 hours and has a duration of action of up to 8 hours. Conversely, intermediate/long-acting insulins (aqueous suspensions), when administered subcutaneously, have an onset of action of 1–2 hours, a peak action between 4 and 12 hours and a duration between 16 and 35 hours.

– Triamcinolone acetonide is administered intramuscularly or intra-articularly as an aqueous suspension to suppress inflammation. The duration of action of a single dose is approximately 21 days.

Physicochemical properties of the therapeutic agent

The physicochemical properties are important determinants of the stability and absorption of the therapeutic agents when formulated as parenteral suspensions (aqueous or oil-based). Conversely, when formulated as a parenteral solution the effects of the physicochemical properties of the therapeutic agent on the above properties are limited. In particular, the following properties directly affect the rate of dissolution (and hence the rate of absorption) of poorly soluble therapeutic agents following IM or SC administration.

Solid-state properties

In the solid state therapeutic agents may exist in either crystalline or amorphous states. In *crystalline* compounds the molecules are packed (bonded) in a defined, repeating order. Crystalline compounds exhibit a defined melting point, which occurs whenever there is sufficient energy applied to the crystal to overcome the attractive forces between the molecules in the lattice. As the strength of the interactive forces increases, so does the melting point of the crystal. *Polymorphism* refers to the ability of molecules to exist in more than one crystalline form. Usually one crystalline form is the stable form and the other(s) are less stable, being referred to as metastable. Over time the metastable form(s) will revert to the stable form. Importantly, different polymorphic forms of a particular therapeutic agent will exhibit different melting points and, as a result, will exhibit different dissolution rates. Polymorphic forms of a particular therapeutic agent will possess the same chemical structure; however, the different solubilities of the different polymorphs will lead to

different rates of dissolution after IM or SC administration and hence different rates of drug absorption.

Solubilities of insoluble salt forms

The reader will be aware that different salt forms of a particular therapeutic agent exhibit different aqueous solubilities. The rate of dissolution of poorly soluble therapeutic agents is directly proportional to the saturated solubility of the compound, as defined in the Noyes–Whitney equation:

$$\frac{\delta M}{\delta t} = \frac{DAC_s}{h}$$

where: $\frac{\delta M}{\delta t}$ refers to the rate of dissolution of the therapeutic agent; h refers to the thickness of the unstirred diffusion layer that surrounds each particle that is undergoing dissolution; D is the diffusion coefficient of the dissolved drug molecule through the unstirred diffusion layer; A is the surface area of the particle undergoing dissolution; C_s is the saturated solubility of the drug, i.e. the concentration that exists in solution adjacent to the dissolving particle.

Therefore, by altering the solubility of the salt form, the rate of dissolution of the drug particles following administration (IM or SC) may be modified. This approach has been successfully used commercially in the formulation of intermediate and long-acting insulin preparations. For example, the solubility of protamine insulin is dramatically lower than soluble insulin and this is reflected by the greater duration of action of the former system. The duration of action of insulin may be further enhanced by forming a salt with zinc or with protamine and zinc, which has lower solubility and hence a lower rate of dissolution.

Particle size

Particle size is a fundamental property that directly controls both the rate of dissolution and physical stability of parenteral suspensions. Referring to the Noyes–Whitney equation, it may be observed that the rate of dissolution of a poorly soluble drug increases as the surface area of the particle increases. In practice the surface area of the drug particle increases as the mean diameter of the particles decreases. Therefore, decreasing the particle size may increase the rate of dissolution of an insoluble therapeutic agent.

The role of particle size on the absorption of an insoluble drug may be illustrated in the following example, which relates to an aqueous testosterone suspension.

- A suspension of testosterone propionate (particle size range 40–100 µm) exhibits a duration of action of *8* days following IM administration.
- A suspension of testosterone propionate (particle size range 50–200 µm) exhibits a duration of action of *12* days following IM administration. Therefore, in this formulation the area of drug particle in contact with the biological fluids is less than in the previous example and, in accordance with the Noyes–Whitney equation, the longer duration may be explained by the slower rate of drug dissolution.
- Interestingly, a suspension of testosterone isobutyrate (particle size range 50–200 µm) exhibited a duration of action of *20* days following IM administration. The greater lipophilicity, and hence lower aqueous saturated solubility, of this testosterone ester would result in a slower rate of dissolution than for the more hydrophilic propionate ester of this drug.

In addition to the effect of particle size on the solubility and hence rate of dissolution of poorly soluble drugs, particle size plays an important role in the physical stability of parenteral suspensions. The reader will recall the Stokes' equation, in which the rate of particle sedimentation is related to particle size as follows:

$$\frac{\delta v}{\delta t} = \frac{2r^2 \, (\rho_s - \rho_l)g}{9\eta_l}$$

where: $\frac{\delta v}{\delta t}$ refers to the rate of particle sedimentation; r refers to the radius of the dispersed particles; $(\rho_s - \rho_l)$ refers to the density difference between the solid phase and the liquid phase; g refers to gravity; and η_l refers to the viscosity of the liquid phase.

It is accepted that reducing the rate of sedimentation of the dispersed drug particles will enhance the physical stability of suspensions. One method by which this may be achieved is to reduce the particle size of the drug particles. Increasing the particle size of the dispersed drug will both increase the rate of sedimentation (and possibly decrease the physical stability of the formulation) and decrease the rate of dissolution of the drug, the latter leading to a slower onset of activity but a prolonged duration of action following IM administration. The interplay between particle size and both the rate of dissolution (and hence absorption) and the physical stability of suspensions should be fully appreciated by students.

Vehicle

All parenteral formulations may be
formulated using an aqueous vehicle, an oil
vehicle or a hydroalcoholic vehicle, the
choice being determined (in part) by the
required solubility of the active agent in
the formulation and the desired type of
formulation.

Aqueous vehicles

Water for injection is the major vehicle of
choice for:

- freely soluble therapeutic agents (for the preparation of
  parenteral solutions)
- therapeutic agents of low aqueous solubility (for the
  preparation of parenteral suspensions)
- the external phase of parenteral emulsions.

Water for injection has specifications set regarding:

- appearance (clear, odourless and within a defined pH range,
  5–7)
- purity (limits on the mass of ions, heavy metals and
  oxidisable compounds and also on the total amount of
  dissolved solids, <10 ppm)
- sterility:
- Water for Injections USP is non-sterile and is used in the
  preparation of parenterals that will be terminally sterilised
  (i.e. during or after the manufacture process).
- Sterile Water for Injections USP is available. This is water for
  injections that has been sterilised and which has been
  packaged in single units (1 litre in volume). It may contain a
  greater mass of dissolved solids due to leaching of solid
  matter from the containers during sterilisation. It is intended
  to be used as a vehicle for products that have been packaged
  and sterilised, e.g. for the reconstitution of antibiotic powders
  as solutions or suspensions.
- pyrogens:
  Water for injection must be free of pyrogens (fever-producing
  compounds) that are primarily associated with Gram-negative
  bacteria. It is important to have knowledge of the
  physicochemical properties of pyrogens as these properties
  directly influence the choice of methods that may be used to
  ensure removal of these compounds. In particular pyrogens
  are:
- thermostable, thereby invalidating their removal using simple
  heating cycles

- water-soluble, thereby invalidating their removal using conventional filtration techniques
- unaffected by bactericides.

In light of the above, pyrogens are effectively removed from water using either distillation or reverse osmosis. Following treatment, water for injection must be stored in *pyrogen-free containers* at a defined temperature (either 5°C or 60–90°C) if the period of storage exceeds 24 hours. Removal of pyrogens from the storage containers is typically performed by heating the container at either 250°C for 30–45 minutes or at 180°C for 3–4 hours.

As a variation on the above, *Bacteriostatic Water for Injections USP* is also available. Similar to Water for Injections USP, bacteriostatic water for injection is sterile and devoid of pyrogens. It additionally contains an antimicrobial agent, e.g. 0.9% w/v benzyl alcohol (a bacteriostatic preservative) and is commonly supplied in a multidose container (≤30 ml). Samples from this container may be repeatedly removed and used to dissolve or dilute therapeutic agents prior to injection. To prevent potential toxicity, Bacteriostatic Water for Injections USP is only used whenever the volume of formulation to be administered is less than 5 ml. Care must be given in the use of Bacteriostatic Water for Injections USP to ensure that the antimicrobial agent does not deleteriously interact with the therapeutic agent.

Non-aqueous vehicles

- ▪ Non-aqueous vehicles are employed for the production of:
- – non-aqueous parenteral solutions of therapeutic agents that are water-insoluble
- – non-aqueous parenteral suspensions of therapeutic agents that are water-soluble and/or exhibit aqueous instability
- – the internal phase of parenteral emulsions.
- ▪ Fixed oils are predominantly used as non-aqueous vehicles (e.g. corn oil, cottonseed oil, peanut oil, sesame oil); however, non-aqueous esters may be used, e.g. ethyl ethanoate:
- – Sesame oil is generally the oil of choice as it is more stable.
- – Oils must be free from rancidity and must not contain mineral oils or solid paraffins.
- ▪ Two major problems associated with the use of non-aqueous pharmaceutical solutions are:
- – Pain/irritation on injection. It should be noted that the viscosity of fixed oils increases at lower storage temperatures. This will, in turn, affect the ease of administration by injection and the pain/irritation at the site of injection. It is essential to ensure that the viscosities of oil-based solutions and suspensions are minimised both to reduce pain on injection and to enhance the ease of administration (injection).

- Patients may exhibit sensitivity to the oils and therefore the oil used in the formulation must be explicitly stated on the label/patient information.
■ In some oily formulations, agents may be added to enhance the solubility of the therapeutic agent in the oil vehicle. Benzyl benzoate (itself a non-aqueous vehicle) may be used for this purpose.

Inclusion of co-solvents

■ As highlighted in Chapter 1, co-solvents are employed whenever the solubility of the drug in water (or occasionally in oil-based vehicles) alone is insufficient for the required application. The types and choices of co-solvent that may be employed in parenteral formulations are generally similar to those used to formulate pharmaceutical solutions; however, when used in parenteral formulations, the potentially greater toxicity of these agents when administered parenterally should be carefully considered. Furthermore, the toxicity of co-solvents is dependent on the route of administration; toxicity is greater whenever administered by the IV in comparison to the IM and SC routes.
■ Examples of co-solvents used in parenteral formulations include:
- glycerol
- ethanol (high concentrations of ethanol are known to produce pain on injection)
- propylene glycol
- polyethylene glycol 400.
  In veterinary formulations other co-solvents may be used, including 2-pyrrolidone and dimethyacetamide; however these co-solvents are not registered for use in parenteral formulations for humans.
■ The concentration of co-solvent used should be sufficient to render the drug soluble within the formulation (over the shelf-life of the product) but should not be irritant or toxic to the patient.

Surface-active agents

The use of surface-active agents in solutions and suspensions designed for oral administration has been addressed in Chapters 1 and 2. In general the incorporation of surface-active agents within parenteral formulations is conceptually identical. In this section some of the basic concepts are revisited within the context of the specific use of these agents in parenteral formulations. In particular:

- Surface-active agents may be incorporated into parenteral solutions to enhance the solubility of the therapeutic agent to the required concentration. In this scenario, the concentration of surface-active agent employed will exceed the critical micelle concentration (CMC) of the surface-active agent. Surface-active agents may be incorporated into aqueous or non-aqueous (oil-based) vehicles for this purpose.
- When included in parenteral suspension formulations, surface-active agents act to enhance the physical stability of the formulation by adsorbing to the surface of the dispersed therapeutic agent and preventing caking of the solid particles. For this purpose concentrations of surface-active agents that are below the CMC may be used. It is extremely common for non-ionic surface-active agents to be used in this manner. Examples include:
  - polyoxyethylene sorbitan fatty acid esters (Tween series), within the concentration range of 0.1–0.5% w/v
  - poly(oxyethylene)-poly(oxypropylene) block co-polymers (Poloxamers), within the concentration range 0.01–5% w/v
  - lecithin, within the concentration range of 0.5–2.0% w/w.
- The choices of surface-active agent and the concentration to be used are dependent on the nature of the vehicle and the type of parenteral formulation (i.e. solution or suspension). Accordingly surfactants with low and high hydrophile–lipophile balance values will be used to stabilise oil-based and aqueous drug suspensions, respectively. Similarly, these surfactants are used to solubilise drugs in oil-based and aqueous vehicles, respectively; in this scenario higher concentrations (> CMC) are required.
- The use of surface-active agents to solubilise therapeutic agents is commercially employed. Examples of these formulated systems include:
  - Steroids have been solubilised for parenteral use using combinations of non-ionic surfactants, e.g. polyoxyethylene sorbitan fatty acid esters (Tween series) and sorbitan esters (Span series).
  - The poorly water-soluble vitamins (A, D, E and K) may be solubilised using surface-active agents as parenteral solution formulations. For example, phytomenandione is formulated as a colloidal solution in a mixed-micelles vehicle and is designed for administration either by slow IV injection or by IV infusion (after incorporation within a 5% glucose solution).
  - The poorly water-soluble antifungal agent amphotericin B is commercially available as a complex with sodium deoxycholate (Fungizone), L-α-dimyristoylphosphatidylcholine and

L-α-dimyristoylphosphatidylglycerol (Abelcet), sodium
cholesteryl sulphate (Amphocil). These powders are
constituted with water for injection to produce colloidal
solutions prior to use as IV infusions.

Buffers

As highlighted in Chapter 1, buffers are
commonly included in parenteral
formulations to control the pH of the
formulation. This is similarly the case for
parenteral formulations where control of the
pH of the formulation may:

- Maintain the solubility of the drug in the
  vehicle over the shelf-life of the
  preparation. Importantly, if there is
  precipitation of the therapeutic agent within a parenteral
  solution during storage, the preparation can no longer be
  referred to as a solution and thus the shelf-life of the product
  has been reached. Furthermore, the IV administration of a
  parenteral solution in which there is precipitated drug may
  lead to a blockage within the capillaries, with the associated
  deleterious effects on the organ to which the blood would
  normally be transported.
- Enhance the chemical stability of the therapeutic agent by
  maintaining the pH of the formulation within the range of
  optimum chemical stability of the therapeutic agent.

Examples of commonly used buffers include acetic acid/sodium
acetate, citric acid/sodium citrate and sodium
phosphate/disodium phosphate (see Chapter 1).

Polymers to modify formulation viscosity and/or drug solubility

The inclusion of polymers within parenteral suspensions occurs
more frequently than in parenteral solutions. In parenteral
solutions the inclusion of hydrophilic polymers will increase the
viscosity of the formulations, which may in turn result in
difficulties in administration. It must be remembered that during
administration the formulation must flow through the narrow
bore of the injection needle. Under these circumstances small
changes in formulation viscosity will be amplified during the
passage through the needle. In addition increased formulation
viscosity may result in pain at the injection site. Whilst
hydrophilic polymers may be added to solutions designed for oral
administration to increase the viscosity, e.g. to aid the accurate
measurement of a dose on the measuring spoon/cup, this
requirement is unnecessary for parenteral solution formulations.
Therefore, the inclusion of hydrophilic polymers in parenteral

solution formulations is restricted to aqueous solutions to enhance the solubility of the therapeutic agent by complexation. Poly(vinylpyrrolidone) (PVP) is an example of a polymer that may be used for this purpose; it is present in aqueous tetracycline and aqueous oxytetracycline injections for veterinary applications. Importantly, the molecular weight of PVP that is employed in the formulation of parenteral solutions is low (circa 12 000) and this, in conjunction with the linear nature of this polymer in aqueous solutions, ensures that large concentrations (up to 18% w/v) may be used without the viscosity of the resultant formulation being excessive for clinical use.

Lipophilic polymers are rarely, if ever, used to solubilise therapeutic agent in oil-based vehicles for parenteral administration. As stated previously, if required the solubility of drugs in the oil-based vehicle may be enhanced by the incorporation of surfactants (e.g. sorbitan esters, Span) or co-solvents, e.g. benzyl benzoate.

When used in aqueous suspensions, hydrophilic polymers maintain the physical stability of the formulation by a number of mechanisms.

Stearic stabilisation

In this the polymer chains adsorb on to the surface of the dispersed drug particles and, in so doing, the close approach of two particles is stearically inhibited. In terms of the DLVO theory (see Chapter 2), the presence of the adsorbed hydrophilic polymer is sufficient to prevent the two particles interacting at the primary minimum. In addition, if the polymer is a polyelectrolyte (e.g. sodium alginate), the polymer chains may effectively form a bridge between two particles when in the presence of an oppositely charged divalent or trivalent ion. The ion provides an effective charge on the surface of the particles, with which the charged polymer chain interacts. In so doing the two particles are maintained/stabilised at distances greater than the primary minimum.

Enhancement of the viscosity of the formulation

As previously defined, the physical stability of suspensions is dependent, at least in part, on the rate of sedimentation of the suspended particles (Stokes' equation). In addition the rate of sedimentation is inversely related to the viscosity of the formulation. The presence of hydrophilic polymers in parenteral suspensions will increase the viscosity of the formulation and will therefore act to stabilise the formulation. However it must be remembered that, as the viscosity of the formulation is increased, the ease of administration decreases and likelihood of pain upon injection increases. Therefore, the concentration of hydrophilic

polymer that may be used for this purpose (and hence the viscosity achieved) is limited by these potential clinical limitations.

The physical stabilisation of aqueous parenteral drug suspensions is usually achieved by the incorporation of surface-active agents and/or hydrophilic polymers. It should be noted that the range of surfactants and hydrophilic polymers that may be used in this fashion is lower than for comparator preparations designed for oral administration. This is primarily due to the potentially greater toxicity of excipients following parenteral administration.

The stabilisation of oil-based suspensions for parenteral administration is generally not performed using (lipophilic) polymers due to their limited availability. Instead the physical stability of oil-based systems may be effectively enhanced using salts of fatty acids or fatty acid esters, which primarily increase the viscosity of the formulation. Examples of these include:

- Aluminium salts of stearic acid (e.g. aluminium stearate, aluminium distearate, aluminium tristearate). These are normally prepared by dissolving the required concentration of aluminium salt (up to 5%, depending on the salt type) into the oil vehicle at high temperatures (circa 165°C). Upon cooling the drug may be dispersed into the rheologically structured vehicle.
- Trihydroxystearin (Thixcin). This may be dissolved in the oil-based vehicle without heating and, in a similar fashion to the example above, produces a rheologically structured vehicle into which the therapeutic agent may be dispersed.

Preservatives

Preservatives are incorporated into parenteral formulations whenever:

- The product is a multidose preparation. In this, several separate doses will be removed from the same container; the inclusion of preservatives is necessary to control microbial growth due to microbial introduction into the product.
- The product has not been terminally sterilised, e.g. by irradiation or heat. In this situation the preservative is required to guard against any possible breakdown in the aseptic manufacturing process.

In all other situations the presence of a preservative is deemed unnecessary.

Examples of preservatives employed in parenteral formulations include:

- esters of parahydroxybenzoic acid, e.g. methyl and propyl parahydroxybenzoic acid are often used in combination in a ratio of 9:1. The concentration is usually circa 0.2% w/v

- phenolic compounds, e.g. phenol (0.25–0.5% w/v) or chlorocresol (0.1–0.3% w/v).

Formulation considerations for the inclusion of preservatives into parenteral formulations include the following:

- In aqueous parenteral suspensions and in some aqueous parenteral solutions hydrophilic surfactants (included to enhance/maintain the solubility of the therapeutic agent or to ensure the physical stability of the formulation over the proposed shelf-life) may interact with esters of parahydroxybenzoate. In so doing the effective (free, unbound) concentration of preservative and hence the preservative efficacy are decreased. This problem is resolved by increasing the concentration of preservative (generally up to 0.25% w/v).
- Preservatives may similarly interact with the container and closure of the parenteral product, necessitating an increase in the concentration of preservative required or, preferably, a change in the type of container closure. For example, phenol has been shown to interact with rubber closures. In this situation, rather than increasing the concentration of phenol added, the rubber closure may be exchanged with a suitable replacement, e.g. nitrile closures.
- It is essential that the preservative does not adversely affect the chemical and physical stability of the parenteral product. For example, as insulin formulations are usually multidose preparations, preservatives are required to inhibit microbial contamination of the product. The physical stability of zinc insulin is compromised in the presence of phenol (but not methylparahydroxybenzoic acid).
- In the preservation of parenteral emulsions, the formulation scientist must be aware of the ability of the preservative to distribute between the inner oil phase and the outer aqueous phase. The preservative is required in the aqueous phase of the emulsion to exert the antimicrobial effect. Distribution between the two phases will therefore decrease the concentration of preservative in the aqueous phase and, accordingly, reduce the preservative efficacy. To overcome this, the concentration of preservative in the dosage form should be increased. This may be easily calculated by consideration of the partition coefficient of the preservative between the oil and aqueous phases and the solubility of the preservative in the two phases (Chapter 3).
- Oil-based parenteral products (solutions and suspensions) do not generally require the inclusion of a preservative due to the low water activity of this medium.

- The potential toxicity of preservatives must be considered when formulating parenteral products in light of the greater potential toxicity of preservatives when administered parenterally. It is therefore desirable to avoid the inclusion of preservatives whenever possible.

Agents to modify the osmolarity of parenteral products

Osmolarity refers to the mass of solute that, when dissolved in 1 litre of solution, will produce an osmotic pressure equivalent to that produced by a one-molar (1 mol) solution of ideal unionised substance. The units for osmolarity that are used in conjunction with parenteral preparations are mosmol/kg. An *isotonic* solution is one that exhibits the same effective osmotic pressure as blood serum, whereas hypotonic and hypertonic solutions refer to solutions in which the osmotic pressure exerted by the solution is less than and greater than blood serum, respectively. The tonicity of parenteral formulations is an important design criterion. In the presence of a hypotonic solution, red blood cells will swell (due to water entry into the cell) and eventually burst (termed haemolysis) whereas, in the presence of a hypertonic solution, water will leave the red blood cells, leading to crenation.

Ideally the administration of parenterals and in particular the IV administration of parenteral should be isotonic (circa 291 mosmol/l) to avoid potential damage. However, many parenteral products designed for IV administration are hypertonic. Products within the osmolarity range of 300–500 mosmol/l may be administered by the IV route rapidly (with care being taken with formulations at the higher extremes of this range to ensure that administration is slower). IV fluids may be profoundly hypertonic, e.g. the tonicity of Hyperamine 30 and Sodium Bicarbonate Intravenous Infusion BP (4.2% and 8.4% w/v) is 1450, 1000 and 2000 mosmol/l, respectively. Clinically this does not present any problems as these fluids will be administered to patients by a central venous line in which the infusion rate is slow and the infusion is rapidly diluted within the blood stream.

It is recommended that parenteral formulations designed for IV administration should not be hypotonic. Therefore, hypotonic solutions should be rendered isotonic by the addition of compounds that will increase the osmotic pressure of the solution. Typically sodium chloride or dextrose is used for this purpose. There are two methods by which the mass of these compounds required to render the solution isotonic may be calculated: (1) consideration of the gram-molecular concentration; and (2) consideration of the freezing-point depression of the solution. These are individually addressed below.

Gram-molecular concentration

- The gram-molecular concentration refers to the number of moles of substance in 100 grams of solvent.
- For example if 1 gram molecule (mole) of a non-ionic compound is dissolved in 100 grams of water, the gram-molecular concentration is 1%.
- As osmotic pressure is a colligative property, the number of moles of substance in solution is important. Therefore if 1 mol of NaCl is dissolved in water, 2 mol of ions are produced.
- A solution is isotonic whenever the gram-molecular concentration is 0.03%.
- The following three examples show how this may be used to produce isotonic solutions.

Worked examples

Example 5.1

Calculate the concentration (% w/v) of dextrose (molecular weight 180 g/mol) that should be added to water to produce an isotonic solution.

Dextrose is non-ionic and therefore 1 mol of dextrose when added to 100 grams of water will produce a gram-molecular concentration of 1%. Therefore:

$$0.03 \times 180 = 5.4\% \ w/v$$

Example 5.2

Calculate the concentration (% w/v) of sodium chloride (molecular weight 58.5 g/mol) that should be added to water to produce an isotonic solution.

Sodium chloride is ionic and therefore 1 mol of NaCl, when added to 100 grams of water, will produce a gram-molecular concentration of 2%. Therefore:

$$\left(\frac{0.03 \times 58.5}{2} \right) = 0.9\% \ w/v$$

Example 5.3

Calculate the concentration of sodium chloride that must be added to a 1% solution of lidocaine hydrochloride (molecular weight 270 g/mol) to render this isotonic.

Initially the gram-molecular concentration of the drug solution must be calculated (remembering that 1 mol of lidocaine hydrochloride dissociates to produce 2 mol of ions). The above

equations must be rearranged in terms of the gram-molecular concentration. Thus:

$$\left(\frac{2 \times 1}{270}\right) = 0.007\%$$

As this is less than 0.03%, the reader will observe that a simple solution of lidocaine hydrochloride (1% w/v) is hypotonic.

The gram-molecular percentage that must be added to correct this imbalance is therefore:

$$0.03 - 0.007 = 0.023\%$$

Therefore, the concentration of sodium chloride required to render this solution isotonic is:

$$\left(\frac{0.023 \times 58.5}{2}\right) = 0.67\% \ w/v$$

Freezing-point depression
▪ The inclusion of ions in a solvent will lower the freezing point of that solvent: the extent of the depression of the freezing point is dependent on the number of ions in solution. This is a basic colligative property.
▪ An isotonic solution exhibits a freezing-point depression of 0.52°C.
▪ Therefore the solution of drug should be adjusted to produce a freezing-point depression of 0.52°C to render the solution isotonic.
▪ Tables are available which provide the freezing-point depressions for various compounds.
▪ The following are two examples of the use of this technique.

Example 5.4
Calculate the concentration (% w/v) of sodium chloride that should be added to water to produce an isotonic solution.

The freezing-point depression of a 1% solution of NaCl is 0.576°C (derived from tables).

Therefore, the concentration of sodium chloride required to render a solution isotonic is:

$$\left(\frac{0.52}{0.576}\right) \times 1 = 0.9\% \ w/v$$

Example 5.5
Calculate the concentration of sodium chloride that must be added to a 1% solution of lidocaine hydrochloride (freezing-point depression of a 1% solution is 0.130°C) to render this isotonic.

Initially the freezing-point depression for the 1% solution of drug is calculated:

$$1 \times 0.130 = 0.130°C$$

Therefore the difference in freezing-point depressions of an isotonic solution and the described drug solution is:

$$0.52 - 0.13 = 0.39°C$$

The freezing-point depression of a 1% w/v solution of sodium chloride is 0.576°C. This allows the calculation of the required mass of sodium chloride to be added:

$$\left(\frac{0.39}{0.576}\right) = 0.677g$$

Antioxidants

As detailed in Chapter 1, many drugs are susceptible to degradation by oxidation, a process involving the addition of an electronegative atom or radical or the removal of an electropositive atom, radical or electron. Oxidation may occur due to the action of molecular oxygen; however, this is a slow process, especially in aqueous solution in which the concentration of dissolved oxygen is low. Alternatively oxidation may be facilitated by free radicals, with breakdown occurring via a chain reaction process. Radicals are formed due to the action of light, heat or transition metals (e.g. iron, copper) that are present in the formulation. Several important classes of therapeutic agents may undergo oxidative degradation, including phenothiazines, polyene antimicrobial agents, steroids, morphine and tetracyclines.

Antioxidants are included in parenteral formulations to slow down or inhibit oxidative degradation of therapeutic agents. These agents either act to prevent the formation of free radicals (e.g. butylated hydroxyanisole, butylated hydroxytoluene) or alternatively are strong reducing agents and are therefore oxidised in preference to the therapeutic agent (e.g. sodium metabisulphite, sodium formaldehyde sulphoxylate). Furthermore, chelating agents, e.g. ethylenediamine diacetic acid, may be added to extract dissolved transition metals, thereby reducing their ability to generate free radicals or to be involved in electron transfer reactions.

A further strategy that may be used to enhance the stability of the therapeutic agent is to flush the injection container/vial with nitrogen prior to closure. In so doing oxygen is removed from the headspace within the packaged product. Although successful, the

limitation of this approach is the availability of specialised filling equipment that will provide satisfactory gas purging prior to closure of the product.

Parenteral emulsions

The reader will be aware that the primary focus of this chapter has been the formulation of parenteral solutions and suspensions, due to the overwhelming majority of parenteral formulations being formulated as these dosage form types. However, under certain circumstances, emulsions are employed as parenteral formulations. As the reader will be aware, emulsions are disperse systems in which one immiscible liquid is dispersed in another liquid. Whilst two forms of emulsion exist, water in oil (w/o, in which water droplets are dispersed in an oil phase) and oil in water (o/w, in which oil droplets are dispersed in an aqueous phase), it is the latter type (o/w) that is usually administered parenterally. Examples of the parenteral use (past and present) of emulsions include:

- The subcutaneous administration of allergenic compounds in a w/o emulsion was performed by Freund & McDermott (1942) to enhance the resultant antibody response.
- The IM administration of o/w emulsions to provide controlled drug release.
- The IV administration of o/w emulsions as total parenteral nutrition emulsions. In these, 10–20% oil is emulsified within an aqueous phase using phospholipids and lecithin surfactants to stabilise the emulsion. The emulsion is rendered isotonic by the addition of glycerol and glucose. The oils are subsequently broken down to triglycerides that provide essential fatty acids and act as a source of calories for patients who cannot consume food orally.

There are several problems associated with the use of parenteral emulsions that have restricted their pharmaceutical use:

- There is a limited list of surface-active agents that may be employed to stabilise parenteral emulsions (due to toxicity concerns).
- When administered intravenously it is essential that the droplet size is less than 1 μm to prevent blockage of blood flow within the capillaries. The physical instability of emulsions, which normally causes the droplets of the internal

Tips

The control of the osmotic properties of parenteral formulations for human use is an important formulation consideration, which should not be overlooked by the formulation scientist.

Parenteral formulations are sterile and therefore preservatives are only usually required whenever the preparation is a multidose formulation, e.g. veterinary parenterals.

Possible interactions between the preservative and viscosity-modifying agent must be considered in the selection of the type and concentration of preservative for parenteral formulations.

phase to coalesce, is therefore a potential dangerous consequence of a poorly formulated emulsion.

■ Emulsions are difficult to sterilise. Normal sterilisation methods, e.g. heat and filtration, are generally inappropriate.

Manufacture of parenteral formulations

As detailed previously, two essential requirements of parenteral formulations are sterility and the absence of pyrogens. It is of no surprise to the reader that these two requirements directly influence the methods by which parenteral formulations are manufactured. All raw materials must be of sufficient (injectable) grade and therefore are assured to be pyrogen-free. Furthermore, the methods used to remove pyrogens from both equipment (storage and manufacture) and water when used as a vehicle for parenterals have been detailed previously.

Before describing different strategies for the manufacture of sterile parenteral formulations, it is worth while briefly discussing the concept of sterility and the different methods by which the sterilisation of parenteral formulations may be achieved. Knowledge of these processes is important in the understanding of sterile product manufacture.

Sterilisation may be defined as the absence of viable microorganisms (either through the destruction of all living microorganisms or by their removal) in pharmaceutical preparations. There are five established methods by which pharmaceutical raw materials and pharmaceutical preparations may be sterilised: (1) moist-heat sterilisation; (2) dry-heat sterilisation; (3) filtration sterilisation; (4) sterilisation by exposure to ionising radiation; and (5) gas sterilisation.

Moist-heat sterilisation

■ performed in an autoclave and employs steam under pressure

■ offers efficient sterilisation at lower temperatures than dry-heat sterilisation, due to the presence of moisture

■ mode of action is thought to be due to the denaturation/coagulation of microbial proteins

■ at normal pressure the temperature of water cannot exceed 100°C. In moist-heat sterilisation the pressure in the autoclave is increased to enable an increase in the processing temperature

■ exposure of the pharmaceutical product at the required temperature for the required time will result in efficient sterilisation. For example, at 103.4 kPa (i.e. 15 pounds per square inch) and 121°C sterilisation is achieved in 20 minutes whereas at 68.91 kPa (10 pounds per square inch) sterilisation

is achieved in 30 minutes. It must be remembered that the time of sterilisation must include a lag period, i.e. the temperature within the interior of the product
■ moist-heat sterilisation is principally used to sterilise materials that are both thermostable (within the conditions of the sterilisation cycle) and through which moisture can perfuse. These include:
– glassware
– dressings
– closures
– aqueous solutions: the aqueous nature of these systems ensures that moist-heat sterilisation is an ideal method for terminal sterilisation (assuming that the therapeutic agent is thermostable). In this process the parenteral solution is either presented to the autoclave sealed in vials or sealed using a closure and aluminium cap. The temperature within the container is elevated to the designated sterilisation temperature at which it is held for the appropriate period.

Dry-heat sterilisation
■ In this process microorganisms are destroyed following cellular dehydration and then pyrolysis/oxidation.
■ Dry-heat sterilisation is performed in ovens.
■ Due to the lower microbicidal efficiency of dry-heat sterilisation (in comparison to moist-heat sterilisation), dry-heat sterilisation is performed at higher temperatures and requires longer times of exposure of the microorganism to this temperature. Examples of dry-heat sterilisation cycles include:
– 170°C for 1 hour
– 160°C for 2 hours
– 140°C for 4 hours.
■ As with moist-heat sterilisation, the described sterilisation conditions refer to the time of residence of the product/article in the oven following attainment of the defined temperature. A lag time is therefore required to ensure that the article/container has achieved this temperature.
■ Dry-heat sterilisation is employed to sterilise materials/products that cannot be readily sterilised by moist heat (and which are thermostable following exposure to the sterilisation cycle), e.g.:
– oils and other aqueous vehicles (e.g. glycerin, propylene glycol)
– heat-stable therapeutic agents/excipients
– glassware (e.g. bottles).

Filtration sterilisation

- In this method microorganisms are removed (not destroyed) from solutions using sterilising filters of pore diameter 0.22 μm. This pore diameter is sufficient to entrap/retain bacteria and fungi. Following use, the filters (containing the entrapped/retained microorganisms) are then safely discarded.
- To maximise the lifetime of the filter (i.e. the volume that may be passed through the filter with blockage of fluid flow), the solution is passed through a series of clarifying filters of defined diameter, e.g. 1 μm, 0.45 μm, prior to passage through the sterilising filter.
- Filtration sterilisation is used to sterilise solutions of therapeutic agents that are thermolabile.
- This is an efficient, inexpensive technique.
- As the product is filtered prior to filling into the final container (e.g. vial/bottle), one concern with filtration sterilisation is the possible concern of the product being non-sterile due to a flaw in the manufacture (and hence function) of the sterilisation filter.

Sterilisation by exposure to ionising radiation

- involves exposure of the raw material/product to a defined dose of ionising radiation. Typically gamma radiation is employed, sterilisation occurring following exposure to 25–40 kGy
- requires specialist equipment and is therefore expensive for routine use
- used to sterilise therapeutic agents/excipients or the production of parenteral formulations that are manufactured and packaged under aseptic conditions but are neither terminally sterilised nor sterilised by filtration
- it must be noted that certain therapeutic agents and excipients are unstable in the presence of sterilising doses of ionising radiation. Therefore the effects of ionising radiation on the stability of formulation ingredients must be individually examined.

Gas sterilisation

- Gas sterilisation involves exposure of materials/products to mixtures of ethylene oxide or propylene oxide and an inert gas, e.g. carbon dioxide within a specially designed apparatus.
- Sterilisation efficiency increases in the presence of moisture (up to 60%) and elevated temperature (circa 55°C).
  A reduction in the operating temperature will result in an increased time required for sterilisation.

- Due to the highly penetrative nature of the gas medium, this technique is frequently used to sterilise medical devices (e.g. packaged catheters) and porous surgical accessories (e.g. blankets). However, this technique may also be employed for the sterilisation of therapeutic agents/excipients.
- Due to the toxicity of the gas mixture, sufficient time must be allowed after sterilisation to enable the sterilising gas to be desorbed from the product/ingredient.

Specific manufacturing requirements for parenteral products

The manufacture of parenteral products occurs under aseptic conditions, in an area that is only used for the preparation of sterile products. The large-scale manufacturing equipment within the facility must be capable of sterilisation in situ. For example, the manufacturing vessels in which the solutions are mixed are generally jacketed and may be sealed. Temperature can then be applied to these to render them sterile. In addition small-scale equipment (which has been previously sterilised and sealed) and raw materials (including the therapeutic agent) are entered into the aseptic manufacturing area through a special portal (with a positive airflow to prevent the ingress of microorganisms). The air supply to the manufacturing area is filtered. Operators within the manufacturing area must wear special (sterile) work clothing, ensuring that there is no operator contamination of the product and environment.

Whilst the manufacture of parenteral formulations is similar to that of non-sterile comparator formulations, the main difference (in addition to the points described in the paragraph above) is the means by which the product is rendered sterile. In this context the manufacture of parenteral solutions (aqueous and oil-based) and parenteral suspensions (aqueous and oil-based) will be briefly discussed separately.

Aqueous parenteral solutions

The main steps in the manufacture of aqueous parenteral solutions are as follows:

- The formulation components (including the therapeutic agent) are dissolved in the main mixing vessel within the manufacturing suite.
- If the active ingredient is thermostable, the formulation is filled into the final containers and sealed. Sterilisation is then performed using moist-heat sterilisation. Unless the preparation is a multidose product, the inclusion of a preservative is unnecessary.
- If the active ingredient is thermolabile, the product is sterilised using filtration and collected into a second mixing

vessel, from which filling into the final container is performed. In some cases it may be possible to fill the product into the final container immediately after filtration, obviating the need for a second mixing vessel. In this case preservatives are normally included.

Oil-based parenteral solutions

Moist-heat sterilisation cannot be used to sterilise oil-based solutions and therefore the manufacturing procedure must accommodate this limitation.

The main steps involved in the manufacture of oil-based parenteral solutions are as follows:

- Sterilisation of the oil-based vehicle (containing the various soluble excipients) within the mixing vessel in the aseptic manufacturing suite (performed using dry-heat sterilisation). If the therapeutic agent is stable under the conditions of dry-heat sterilisation, the drug may be incorporated (dissolved) at this stage. This product is then ready for filling into the final container (which is then sealed). Alternatively the product may be manufactured and terminally sterilised using dry-heat sterilisation.
- If the therapeutic agent is not stable under the above sterilisation conditions, sterile drug should be dissolved in the sterile oil-based vehicle using normal mixing facilities. This product is then ready for filling into the final container, which is subsequently sealed.

Aqueous parenteral suspensions

As suspensions contain drug that has been dispersed in the chosen vehicle, filtration cannot be performed to render the product sterile. Therefore, sterile drug must be added to the chosen vehicle and suitably dispersed under aseptic conditions. Typically, the main steps in the manufacture of aqueous parenteral suspensions are:

- dissolution of the formulation excipients in an aqueous (or hydroalcoholic) vehicle within the main vessel
- sterilisation of the vehicle by filtration and collection into a second vehicle, under aseptic conditions
- dispersal of the (sterile) therapeutic agent into the sterile vehicle. If required the particle size of the disperse phase may be reduced by, for example, passage through a ball mill (whose contents and packing material are both sterile)
- the formulation may then be filled into the final container, followed by sealing
- drug suspensions are often physically unstable if exposed to moist-heat sterilisation; however, if the suspension is stable under these conditions, then the formulation may be

manufactured and filled into the final container, followed by terminal sterilisation.

Oil-based parenteral solutions

The manufacture of oil-based suspensions combines the manufacturing considerations of both oil-based solutions and aqueous suspensions and includes:

- sterilisation of the oil-based vehicle (containing the various soluble excipients) within the mixing vessel in the aseptic manufacturing suite (performed using dry-heat sterilisation)
- mixing of the sterile therapeutic agent into the sterile vehicle, prior to filling into and sealing the final container
- if the formulation can physically and chemically withstand the conditions for dry-heat sterilisation, the product may be manufactured and filled in the final container. This may then be sterilised using dry heat.

Further reading

Freund J, McDermott K (1942) Sensitization to horse serum by means of adjuvants. *Proc Soc Exp Biol Med* 49: 548–553.

Multiple choice questions

1. **Regarding parenteral formulations, which of the following are true?**
a. Parenteral formulations are sterile products.
b. Parenteral formulations must be pyrogen-free.
c. The majority of parenteral formulations for human administration are multidose formulations.
d. The vehicle of choice for human parenteral formulations is water.

2. **Regarding the intravenous route of parenteral administration, which of the following are true?**
a. Large-volume parenteral formulations may be administered intravenously.
b. Intravenous administration results in a rapid onset of drug action.
c. Emulsion parenteral formulations must never be administered intravenously.
d. The intravenous route of parenteral administration provides limited drug bioavailability.

3. **Regarding the intramuscular route of parenteral administration, which of the following are true?**
a. Large-volume parenteral formulations are routinely administered intravenously.

b. Solution parenteral formulations may be administered intramuscularly.

c. Emulsion parenteral formulations may be administered intramuscularly.

d. Intramuscular administration of parenteral formulations provides a rapid onset of drug action.

4. **With respect to parenteral formulations, which of the following are true?**

a. The particle size of the suspended therapeutic agent affects the subsequent onset of drug action.

b. The majority of veterinary parenteral formulations are formulated as suspensions.

c. The majority of insulin formulations are administered intramuscularly.

d. Increasing the particle size of the suspended therapeutic agent will decrease the duration of action following intramuscular administration.

5. **Regarding the osmotic pressure of parenteral solutions, which of the following are true?**

a. A solution which has a molecular concentration of 0.9% is isotonic with blood plasma.

b. 1 mol of a substance in 100 g water has a gram-molecular concentration of 1%.

c. Sodium chloride ionises in solution to yield 2 gram ions.

d. Dextrose, boric acid and phenol are regarded as practically non-ionising in aqueous solutions.

6. **Regarding parenteral solutions, which of the following are true?**

a. Parenteral solutions must contain a preservative.

b. All parenteral solutions must be buffered.

c. Glycerol is frequently chosen as a co-solvent in small-volume parenteral solutions.

d. Parenteral solutions may be administered by the intradermal route of administration.

7. **Regarding the use of surface-active agents in parenteral formulations, which of the following are true?**

a. Surface-active agents are employed to lower the interfacial free energy of water, thereby adding injection.

b. Cationic surface-active agents are commonly used in parenteral solution formulations to enhance the solubility of therapeutic agents.

c. Surface-active agents are used to enhance the solubility of therapeutic agents at concentrations lower than the critical micelle concentration.
d. Lecithin is commonly used to stabilise emulsions designed for parenteral administration.

8. **Concerning Water for Injections, which of the following are true?**
a. Water for Injections must be sterile.
b. Water for Injections has defined limits on the total amount of dissolved solids.
c. Water for Injections is pyrogen-free.
d. Water for Injections contains buffers to control the pH.

9. **Regarding oily solution parenterals, which of the following are true?**
a. The viscosity of the formulation may increase as the storage temperature is decreased.
b. Examples of oils that may be used as the vehicle include arachis oil and liquid paraffin.
c. The rheological properties of oily injections may be modified by the addition of aluminium salts of stearic acid.
d. Oily injections require the inclusion of a preservative.

10. **Regarding the manufacture of parenteral formulations, which of the following are true?**
a. Parenteral suspensions may be terminally sterilised by filtration.
b. Commonly used sterilisation cycles destroy pyrogens.
c. Aqueous parenterals may be sterilised using dry-heat sterilisation.
d. Oil-based parenterals are commonly sterilised using moist-heat sterilisation.

chapter 6
Ocular, nasal and otic dosage forms

Overview

In this chapter the following points will be discussed:
- an overview/description of the uses of ocular dosage forms
- formulation strategies for ointments
- formulation strategies for ocular dosage forms, including consideration of the excipients used
- the advantages and disadvantages and uses of ocular dosage forms
- considerations for the manufacture of ocular dosage forms
- an introduction into the formulation of nasal and otic dosage forms.

Introduction

This chapter predominantly deals with the formulation of dosage forms that are designed to be administered topically to the eye (principally the conjunctiva or the eyelid) for the treatment of primarily local disorders: glaucoma, a disease of the interior of the eye, may also be treated using these dosage forms. There are three main types of dosage form that are clinically used: (1) solutions; (2) suspensions; and (3) ointments. The administration of these to the eye is usually performed using a dropper (or a container with a dropper nozzle) or a tube with a nozzle. Access of principally solution or suspension formulations to the remote anatomical regions of the eye may also be achieved parenterally (termed intraocular injections) to the anterior, posterior and vitreous chambers (Figure 6.1). Ocular formulations (akin to parenteral formulations) must be sterile. Formulation considerations for nasal and otic formulations will be discussed later in this chapter.

KeyPoints

- Ocular dosage forms are commonly used to treat local ocular disorders, e.g. infection and inflammation; however, intraocular disorders, notably glaucoma, may also be successfully treated.
- Ocular dosage forms are principally solutions, ointments and suspensions. Intraocular injections are available for the treatment of more serious disorders.
- These formulations exhibit similar concerns as comparator formulations regarding physical and chemical stability.
- Ocular dosage forms must be sterile.
- Nasal and otic dosage forms are non-sterile dosage forms that are inserted into the nasal cavity and ear canal (respectively) for the treatment of local disorders.
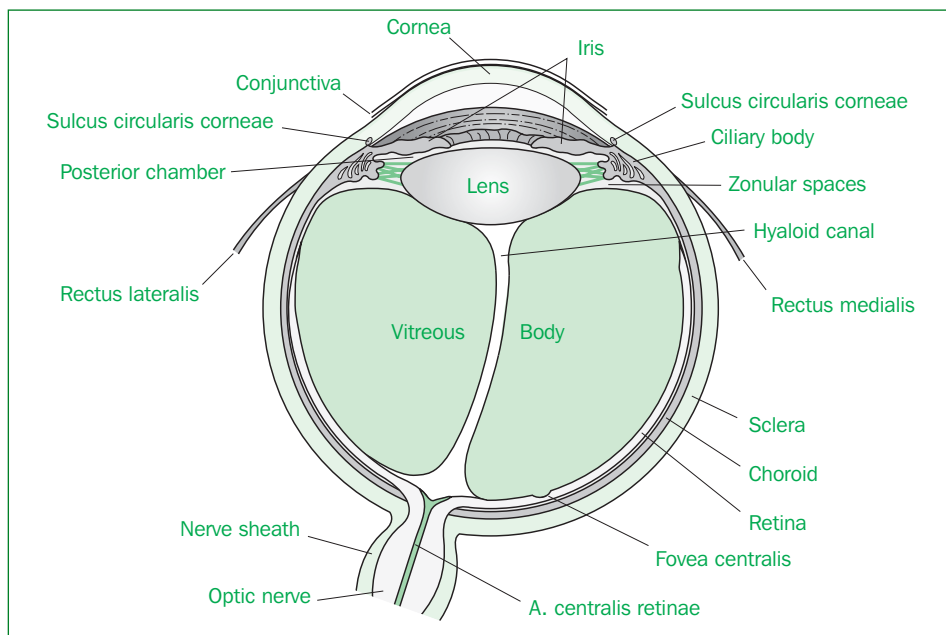
Advantages and disadvantages of ocular dosage forms

Advantages
- The application of the therapeutic agents directly to the site of action ensures that the therapeutic agent is available at higher concentrations than may be achieved following oral administration.
- Administration of the therapeutic agent locally ensures that the incidence of side-effects is minimised.
- Following training, the administration of the dosage form locally to the eye may be easily performed by the patient.

Disadvantages
- The retention of the drug at the site of action is relatively poor, due principally to the low tear volume (7 μl for the blinking eye, 30 μl for the non-blinking eye). The typical volume of two drops of a solution formulation is circa 100 μl and therefore the majority of the applied dose is lost either through spillage on to the face or via the lacrimal duct.
- In addition, the retention time of applied solutions on the surface of the eye is poor. For example, it has been reported that, following the administration of a pilocarpine solution to the eye, removal of the solution from the precorneal region of

the eye occurs in less than 2 minutes, resulting in absorption of approximately 1% of the originally applied dose of drug.

- Therefore, to overcome these deficiencies in practice, the patient is required to administer the ocular solution formulations (containing high concentrations of therapeutic agent) frequently, which is inconvenient and may lead to patient non-compliance. These inadequacies have inspired pharmaceutical scientists to devise strategies by which the retention of the drug within the precorneal region may be enhanced.
- Ocular formulations are sterile and therefore specialist facilities are required for the manufacture of these dosage forms.
- Local side-effects may be experienced to ocular dosage forms (to either the high concentration of therapeutic agent ($\leq 5\%$ w/w) or excipients used in the formulation). Typically pain and irritation are the major side-effects encountered by patients.
- The application of ointment formulations to the eye may result in a temporary blurring of vision.

Administration of therapeutic agents to the eye

The main anatomical features of the eye are illustrated in Figure 6.1.

The main features of the eye that are of interest for ocular drug delivery and hence the formulation of ocular dosage forms are: (1) conjunctiva; (2) cornea; and (3) lacrimal fluid.

Conjunctiva
- The conjunctiva is located at the side of the eye and joins on to the cornea and eyelids.
- The surface area of the conjunctiva is large (circa 18 cm^2).
- The conjunctiva helps produce and maintain the tear film.
- The permeability of the conjunctiva to the diffusion of therapeutic agents is greater than that of the cornea.

Cornea
- The cornea is composed of three layers:
- epithelium (adjacent to the conjunctiva): a multilayered epithelium that is rich in lipids
- stroma (central region): this is an aqueous matrix composed of collagen and keratocytes
- endothelium: a lipid-rich, single-cellular epithelium that maintains corneal hydration.
- The diffusion of drugs into the inner chambers of the eye is controlled by the cornea; diffusion occurs via paracellular routes.

- The lipid outer and inner layers (epithelium/endothelium) of the cornea and the predominantly aqueous stroma control drug diffusion into the internal regions of the eye. To be effectively absorbed, therapeutic agents must exhibit intermediate solubility in both these lipid and aqueous phases and must be of low molecular weight.
- The cornea is non-vascular and negatively charged.

Lacrimal fluid
- Lacrimal fluid is secreted from glands and is located on the surface of the eye.
- The pH of the lacrimal fluid is 7.4 and this fluid possesses a good buffer capacity (due to the presence of carbonic acid, weak organic acids and protein), being able to neutralise *unbuffered* formulations effectively over a wide range of pH values (3.5–10.0).
- Lacrimal fluid is isotonic with blood. Typically ocular aqueous dosage forms are not specifically formulated to be isotonic (0.9% w/w NaCl equivalent) and may be formulated within a range of tonicity values equivalent to between 0.7 and 1.5% w/w NaCl.
- The rate of turnover of lacrimal fluid is approximately 1 μl/min and the blinking frequency in humans is circa 15–20 times per minute. These physiological functions act to remove the therapeutic agent/formulation from the surface of the eye.

Formulation considerations for aqueous ocular dosage forms
There are two categories of aqueous ocular dosage forms: (1) ocular solutions; and (2) ocular suspensions. Whilst the vast majority of aqueous ocular dosage forms are solutions, suspensions may be required whenever the therapeutic agent exhibits problems regarding chemical stability or, in the case of steroids (e.g. dexamethasone, prednisolone), the potency of the lipophilic drugs is greater than that of the water-soluble salts. The majority of formulations considerations for ocular dosage forms are similar to those described for pharmaceutical solutions in general. Accordingly the main considerations for the formulation of ocular solutions and suspensions are as follows.

Choice of drug salt for use in ocular solutions
One of the main determinants surrounding the choice of salt type for inclusion in solution formulations is the solubility – the salt form being chosen to achieved the required solubility. To compensate for the poor retention of therapeutic agents within the precorneal region (and therefore to achieve the maximum pharmacological effect), the concentration of therapeutic agents in

ocular solutions is relatively high. Under these circumstances the choice of drug salt is important as the local ocular application of certain drug salts results in greater pain/irritation (e.g. stinging). This may be exemplified by considering three salt forms of adrenaline (epinephrine) (the hydrochloride, borate and bitartrate salts), their physicochemical properties in solution and the associated pain when applied to the eye. Adrenaline bitartrate is the most acidic salt in solution (pH 3–4) and, due to the strong buffer capacity, has been reported to produce moderate to severe stinging following application. Conversely, only mild stinging is associated with the topical application of the borate salt due to the low buffer capacity and lower acidity of this salt (pH 5.5–7.5). Adrenaline hydrochloride, although relatively acidic (pH 2.5–4.5), exhibits medium buffer capacity and is therefore more effectively neutralised than the bitartrate salt. This results in reduced stinging in comparison to the bitartrate salt. Importantly, as the discomfort of the salt form increases, there will be a greater tendency for the drug to be washed off the surface of the eye due to the production of lacrimal fluid.

Physical properties of the dispersed therapeutic agent

The physical properties of the therapeutic agent are important design features of ophthalmic suspensions and indeed in all types of suspension dosage forms. In addition to the physical properties previously outlined in Chapter 2, the particle size used in ocular suspensions is of primary importance due to the relationship between particle size and ocular irritation. Typically 95% of dispersed particles should have an average particle diameter that is less than 10 μm.

Solution pH/inclusion of buffers

The pH and the control of the pH of ocular formulations are important determinants of the stability of the therapeutic agent, the ocular acceptability of the formulation and the absorption of the drug across the cornea. Ideally the pH of the formulation should be that which maximises the chemical stability (and, if required, absorption) of the formulation. This issue is particularly important due to the effect of pH on the stability of alkaloid drugs, e.g. atropine, pilocarpine, carbochol. As highlighted in the previous section, the pH and the buffer capacity directly affect the subsequent discomfort of the formulation.

Ideally the pH of the ocular solution should be controlled at 7.4 as this is the pH of tear fluid. However, the choice of pH of the formulation is also dictated by the stability of the therapeutic agent at that pH (which in turn serves to define the shelf-life of the formulation) and whether (or not) absorption of the active agent across the cornea is required.

Chemical stability

pH is an important contributor to the degradation of therapeutic agents that undergo hydrolysis, e.g. alkaloids. The effect of pH on the stability of two alkaloids at two different temperatures is shown in Table 6.1.

Table 6.1 The effect of temperature and pH on the stability (half-life) of pilocarpine and atropine

Drug	Temperature (°C)	pH	Half-life
Atropine	121	6.8	1 hour
	121	5	60 hours
	25	6.8	2 years
	25	5	130 years
Pilocarpine	121	6.8	34 minutes
	121	5	24 hours
	25	6.8	66 days
	25	5	Several years

As may be observed, the stability of the two alkaloids at room temperature increases as the pH is increased. Therefore, solutions of these alkaloids are prepared at the lower pH to ensure that the product has a commercially viable shelf-life. Interestingly, exposure to high temperatures, such as those encountered during moist-heat sterilisation, also reduces the stability of these therapeutic agents (although this is significantly less within a formulation of lower pH). Therefore, the choice of pH may also affect the method by which the formulation is sterilised. Importantly, if the pH of alkaloidal solutions is buffered to pH 6.8 (e.g. using a phosphate buffer), the formulations must not be sterilised using thermal methods. For this purpose aseptic preparation involving sterile filtration is used.

Drug absorption across the cornea

The successful treatment of glaucoma using ocular formulations requires that there is sufficient drug absorption across the cornea. To be effectively absorbed the drug must exhibit differential solubility, i.e. the ionised and non-ionised forms coexist. The effects of pH and ionisation on the absorption of drugs across the cornea are represented in Figure 6.2.

As may be observed, sufficient concentration of the non-ionised form is required to partition into and diffuse across the lipid-rich outer layer of the cornea (the epithelium). The inner layer of the cornea (the stroma) is predominantly aqueous and therefore ionisation of the drug must occur to enable partitioning into this phase. Following diffusion to the interface of between the stroma and the endothelial (lipid-rich) layer, absorption of the non-ionised (but not the ionised) form occurs. The non-ionised
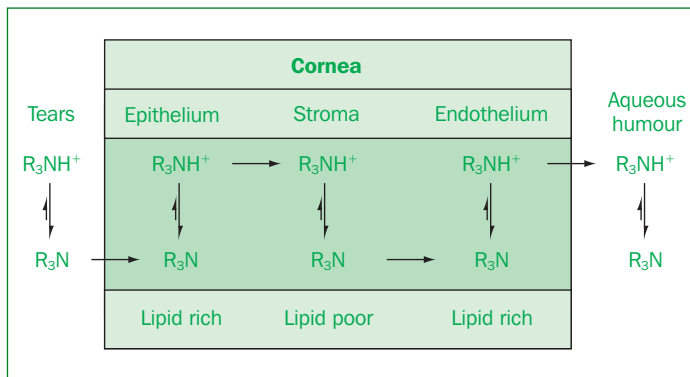
Figure 6.2
Diagrammatic
representation of the
effect of ionisation (and
hence pH) on drug
absorption across the
cornea.

drug then diffuses to the endothelium/aqueous humour interface where ionisation and dissolution into the aqueous humour occur. The pKa of the therapeutic agent determines the ionisation of the therapeutic agent at defined pH values. The role of pH on the stability of alkaloids was highlighted in the previous section. Therefore, whilst the chemical stability of alkaloids is enhanced in formulations of lower pH (5), under these conditions the therapeutic agent will be predominantly ionised (circa 99%) and drug absorption will be poor. Increasing the pH of the formulation will decrease the percentage ionisation of the therapeutic agent (and hence increase absorption); however, the chemical stability may be compromised.

To overcome this problem alkaloidal drugs are frequently used in the form of the acid salt, where the pH of the solution will be acidic and stability is optimised. No buffers are added and therefore whenever the formulation is instilled into the eye, the lacrimal fluid adjusts the pH to physiological conditions, thereby facilitating absorption.

If pH control is required, e.g. for acidic drugs, a borate buffer may be used to produce solutions of acidic pH (circa 5). Stabilisation of this pH within the lacrimal fluid will, if required, enhance the absorption of acidic drugs. Phosphate buffers may be used to control the pH of eye drops (at physiological pH). Finally it should be remembered that control of the pH is often used to prevent precipitation of the therapeutic agent in the formulation.

Tip

Most eye drop formulations are solutions due to the less complex nature of the formulation (as compared to suspensions) and greater comfort in use.

Vehicle

The vehicle that is predominantly used for the formulation of aqueous ocular dosage forms is Purified Water USP. Water for injections is not a specific formulation requirement. Occasionally oil is used if the therapeutic agent is profoundly unstable within

an aqueous vehicle. The choice of oils for ocular use is similar to that for parenteral use.

Viscosity-modifying agents

Viscosity-modifying (enhancing) agents are hydrophilic polymers that are added to ocular solutions for two main reasons: (1) to control the rate at which the drop flows out of the container (and thus enhance ease of application); and, more importantly, (2) to control the residence time of the solution within the precorneal environment. For example, it has been shown that the retention of an aqueous solution within the precorneal region is short (frequently less than 1 minute); however, if the viscosity is increased, the retention may be enhanced. Furthermore, it has been reported that there is a critical formulation viscosity threshold (circa 55 mPa/s) above which no further increase in contact time between the dosage form and the eye occurs. It must be remembered that there is an upper limit below which the viscosity of eye drop formulations must be maintained as this may lead to blockage of the lacrimal ducts. The viscosities of commercially available products are frequently less than 30 mPa/s. The enhancement of the viscosity of ocular suspensions will serve to enhance the physical stability of ocular suspensions.

Ideally the viscosity-modifying agent should exhibit the following properties:

- *Easily filtered*: all eye drop solutions are filtered during the manufacturing process. This may be simply to remove any particles (e.g. clarification using a 0.8-μm filter) or clarification in conjunction with filtration sterilisation.
- *Easily sterilised*: sterilisation of eye drop solution is usually performed by filtration (see above) or by heat. If heat sterilisation is performed, the viscosity-modifying agent must be chemically and physically stable under these conditions.
- *Compatible with other components*: the interaction of hydrophilic polymers with certain preservatives is a well-known phenomenon and is usually resolved by increasing the concentration of preservative. However, the pharmaceutical scientist must ensure that there is no or only limited interaction between the therapeutic agent and the viscosity-modifying agent, e.g. basic therapeutic agents and polyacidic polymers.

Examples of polymers that are used as viscosity-modifying agents include:

- *Hydroxypropylmethylcellulose (Hypromellose USP).* Hydroxypropylmethylcellulose (HPMC) is a partially methylated and *O*-(2-hydroxypropylated) cellulose derivative (Figure 6.3).
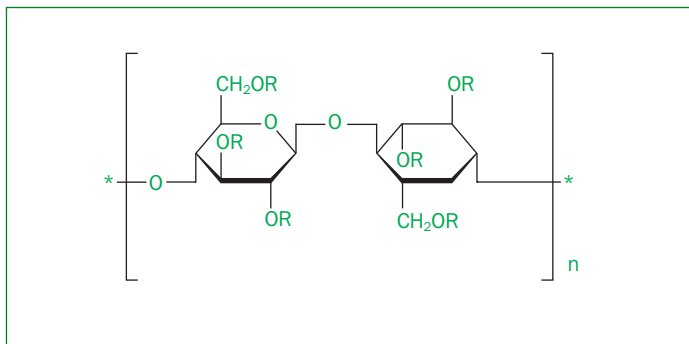
In aqueous ocular formulations HPMC is used in the concentration of 0.45–1.0% w/w.

■ *Poly(vinyl alcohol) (Figure 6.4).* This is a water-soluble vinyl polymer that is available in three grades: (1) high-viscosity (average molecular weight 200 000 g/mol); (2) medium-viscosity (average molecular weight 130 000 g/mol); and (3) low-viscosity (average molecular weight 20 000 g/mol). It is used to enhance the viscosity of ocular formulations in concentrations ranging from 0.25 to 3.00% w/w (the actual concentration being dependent on the molecular weight of polymer used).
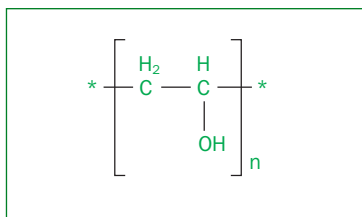
Figure 6.4 Structural formula of poly(vinyl alcohol).

Poly(acrylic acid)

This is a water-soluble acrylate polymer that is cross-linked with either allyl sucrose or allyl ethers of pentaerythritol. It is predominantly used in ocular aqueous formulations for the treatment of dry-eye syndrome. However, it may be used to increase the viscosity of ocular formulations that contain a therapeutic agent.

Preservatives

Ocular formulations are sterile; however, as they are designed as multidose systems, the addition of a preservative is required. The main features of preservatives and preservation in general have been described in previous chapters and therefore this section will specifically address issues regarding the preservation of ophthalmic formulations, including a description of the key

preservatives that are used in this class of pharmaceutical product.

Cationic preservatives

The two main cationic preservatives that are commonly used in ocular solutions and suspensions are *benzalkonium chloride* and *benzethonium chloride*. It should be noted that cationic preservatives are incompatible with anionic therapeutic agents, e.g. pilocarpine nitrate, physostigmine, used for the control or diagnosis of ocular conditions, e.g. sodium fluorescein. Furthermore, interaction of the these compounds may occur with non-ionic hydrophilic polymers (used to modify the viscosity of ocular formulations).

Benzalkonium chloride

This is a mixture of alkylbenzyldimethylammonium chlorides that is used in ocular solutions/suspensions at a concentration between 0.002 and 0.02% w/v (typically 0.01% w/v). The resistance of certain microorganisms that are ocular pathogens to benzalkonium chloride (most notably *Pseudomonas aeruginosa*) has been observed. Therefore, it is customary to include 0.1% w/v *disodium edetate* in ocular formulations in which benzalkonium chloride is used. This agent acts to enhance the antimicrobial activity of benzalkonium chloride by chelating divalent cations in the outer membrane of the bacterial cell, thereby rendering the bacteria more permeable to the diffusion of the antimicrobial agent. The antimicrobial properties of benzalkonium chloride decrease whenever the pH of the formulation falls below 5.

Benzethonium chloride

Benzethonium chloride (unlike benzalkonium chloride) is a pure compound (and not a mixture of compounds) (Figure 6.5).

Figure 6.5 Structural formula of benzethonium chloride. It is commonly used in ophthalmic formulations within the concentration range of 0.01–0.02% w/v (although it has been reported to exhibit lower antimicrobial activity than benzalkonium chloride).
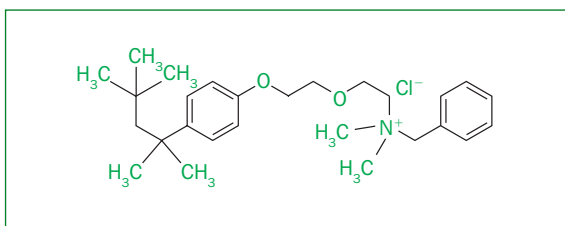
Esters of parahydroxybenzoates (parabens)

Mixtures of methyl and propyl esters of parahydroxybenzoic acid are used in ophthalmic formulations (typically at a combined concentration of 0.2% w/w). There is a concern regarding the ocular irritancy of the parabens, which limits their use in

ophthalmic preparations. This problem is augmented by the need to increase the concentration of parabens in ocular formulations that contain hydrophilic polymers (due to the interaction between these two species).

Organic mercurial compounds

These are antimicrobial agents that contain mercury and, due to environmental and toxicity concerns, are not commonly used in ocular formulations nowadays. The main examples of these compounds that have found pharmaceutical use are phenylmercuric acetate (Figure 6.6a), phenylmercuric nitrate (Figure 6.6b) (which is sometimes supplied as a mixture with phenylmercuric hydroxide, Figure 6.6c) and thimerosal (Figure 6.6d). The typical concentration ranges of the antimicrobial agents used in ocular formulations are: 0.001–0.002% w/v for phenylmercuric acetate, 0.002% w/v for phenylmercuric nitrate and 0.001–0.15% w/v and 0.001–0.004% w/v thimerosal (when used in ophthalmic solutions and suspensions, respectively).
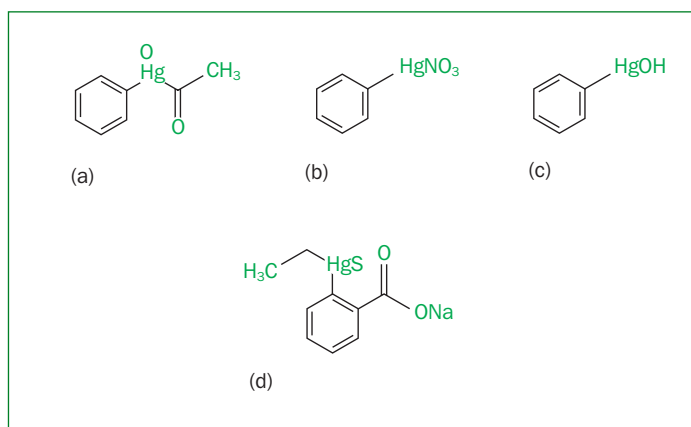
Figure 6.6 Structural formulae of: (a) phenylmercuric acetate; (b) phenylmercuric nitrate; (c) phenylmercuric hydroxide; and (d) thimerosal.

The use of the phenylmercuric salts has been reported to be deposited in the lens of the eye (termed mercurialentis) when formulated in preparations that are designed for chronic usage, e.g. for the treatment of glaucoma. Thimerosal is not associated with this problem; however, it has been associated with ocular sensitisation. As a result, these preservatives are only used in ocular formulations whenever there is no suitable option.

Organic alcohols

Two main organic alcohols are used as preservatives for ophthalmic formulations: (1) chlorobutanol; and (2) phenylethylalcohol. The main features of these are described below.

Chlorobutanol

Chlorobutanol (Figure 6.7) is used in ocular formulations at a concentration of circa 0.5% w/v. Under alkaline conditions hydrolysis of chlorobutanol occurs, liberating HCl as a byproduct (the rate of reaction increases with increasing temperature, e.g. during autoclaving).

Figure 6.7 Structural formula of chlorobutanol.
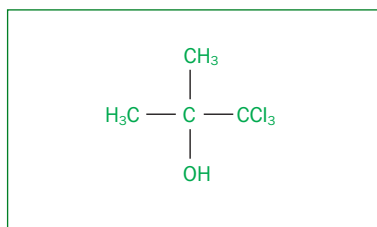
Therefore, the use of chlorobutanol is reserved for acidic ophthalmic preparations. In addition, chlorobutanol is volatile and lost from solution if stored in polyolefin containers (due to partitioning). Accordingly, preparations that employ this preservative must be stored in glass containers. One final problem associated with the use of chlorobutanol is its limited solubility. The concentration of chlorobutanol used in ophthalmic preparations (0.5% w/v) is close to the saturation solubility (0.7% w/v at room temperature). Therefore, if the formulation is stored below room temperature, precipitation of the preservative may occur in situ.

Phenylethylalcohol

Phenylethylalcohol (Figure 6.8) has similar properties to chlorobutanol and indeed shares similar problems, e.g. poor solubility, volatile and partitioning into plastic containers. The typical concentration used in ophthalmic preparations is 0.25–0.50% v/v.

Figure 6.8 Structural formula of phenylethylalcohol.
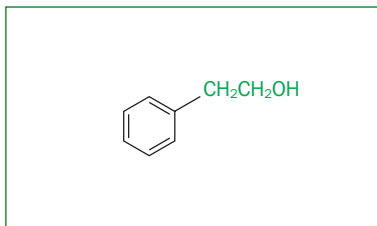
Antioxidants

As in other formulations, antioxidants may be added to ocular solutions/suspensions to optimise the stability of therapeutic agents that degrade by oxidation. Sodium metabisulphite (circa 0.3%) is an example of an antioxidant that is commonly used for this purpose. Other examples have been cited in previous chapters.

Surface-active agents

Surface-active agents are predominantly employed in aqueous suspension to enhance the physical stability of the dispersed particles; however, there have been reports concerning the use of these agents to solubilise therapeutic agents in aqueous ocular solutions. One of the primary concerns regarding the use of surface-active agents in ocular dosage forms is the potential toxicity/irritancy. Accordingly, non-ionic surfactants are preferentially (and predominantly) used whereas the use of anionic surfactants on ocular solution/suspension dosage forms is avoided.

Antioxidants

As before, antioxidants may be added to aqueous ocular formulations to enhance the stability of therapeutic agents that are prone to oxidative degradation (e.g. adrenaline). For this purpose, sodium metabisulphite and related sulphite antioxidants may be used.

Tips

The use of mercuric preservatives in eye drop formulations is less common than for other preservatives due to possible concerns regarding toxicity.

Cationic surfactants are commonly used as preservatives in eye drop formulations.

Eye drop formulations are sterile. The inclusion of preservatives is required due to the multidose nature of this type of dosage form.

Tip

The formulation of eye drop dosage forms involves the use of similar components to those used in the formulation of conventional solutions; however, it should be remembered that ocular dosage forms are sterile.

Manufacture of aqueous ophthalmic formulations

The manufacture of aqueous ophthalmic solutions/suspensions is similar to that described for the manufacture of parenteral solutions and suspension (Chapter 5). Akin to parenteral formulations, ophthalmic solutions/suspensions must be sterile. The manufacture of these systems may involve at least one of the following steps.

Production under clean conditions followed by sterilisation by autoclaving

Typically ophthalmic solutions may be manufactured and packaged in the final container under clean conditions. Sterilisation may then be performed using moist-heat sterilisation (assuming the therapeutic agent is chemically stabile under these conditions).

Production under clean or aseptic conditions followed by aseptic sterilisation by filtration

If the therapeutic agent is thermolabile then sterilisation by heat should be avoided. The production of ocular solutions is then performed by initially manufacturing the dosage form under clean or aseptic conditions. Clarification of the solution is performed by filtration through an appropriate filter (circa 1 µm), followed by sterilisation filtration (0.22-µm filter) and packaging (both under aseptic conditions).

Production under aseptic conditions

Ophthalmic suspensions may not be sterilised by filtration. Therefore, the manufacture of these systems usually involves the dispersion of the sterile therapeutic agent into the sterile vehicle (with added excipients) and subsequent packaging, both under aseptic conditions.

Formulation considerations for aqueous ocular dosage forms

Ophthalmic ointments are commonly used for the treatment of ocular infection and/or inflammation. One major disadvantage associated with the use of ophthalmic ointments is their general greasiness and the associated (temporary) blurring of vision. For these reasons patients may prefer aqueous ocular formulations or, alternatively, may prefer to reserve the use of ophthalmic ointments to night-time application. Due to their greater retention within the precorneal region (approximately 2–4 times greater than for aqueous solutions), the frequency of administration of ointments is generally lower than their aqueous-solution counterparts.

As is the case for all ointment formulations, ophthalmic ointments are prepared by dispersion of the therapeutic agent in the pre-prepared ointment base. The physicochemical properties and pharmaceutical strategies for the formulation of ointment bases have been detailed in Chapter 4 and, accordingly, these details will not be repeated in this chapter. This chapter will, however, define the particular formulation requirements of ophthalmic ointments and redirect the reader to the afore-mentioned chapter for further details.

Ointment bases for ocular administration

Various types of ointment bases may be employed for ocular administration, as follows.

Hydrocarbon bases

Hydrocarbon bases are widely used in ophthalmic ointments and are generally composed of a mixture of paraffins (to achieve the correct viscosity). In the formulation of these systems, yellow soft

paraffin is preferred to white soft paraffin: white soft paraffin is a bleached form of yellow soft paraffin, thereby being associated with a lower potential for ocular irritancy. Examples and properties of the various excipients that are used to formulate these systems have been provided in Chapter 4.

Non-emulsified absorption bases

As detailed in Chapter 4, these bases are composed of one or more paraffins (e.g. liquid and yellow soft paraffin) and a sterol containing emulsifying agent (e.g. lanolin derivatives). These bases are less greasy than those prepared using hydrocarbons alone and, in addition, aqueous solutions of drug may be incorporated into non-emulsified absorption bases.

Water-soluble bases/aqueous gels

Water-soluble bases may be successfully administered to the eye and are associated with a lower incidence of blurred vision (due to their water-miscibility). These bases are primarily formulated using polyethylene glycols. More recently aqueous gels containing a dissolved or dispersed therapeutic agent have been successfully used for the treatment of infection and/or inflammation. Aqueous gels composed of poly(acrylic acid) are commonly used for this purpose and, in addition, have been successfully employed for the treatment of glaucoma (requiring fewer administrations per day in comparison to aqueous solutions containing this drug). Furthermore, poly(acrylic acid) gels (devoid of therapeutic agent) have also been used for the treatment of keratoconjunctivitis sicca (dry-eye syndrome).

Excipients in ocular ointments

The nature of the excipients required in ophthalmic ointments is dependent on the nature of the ointment base used in the formulation. Typically hydrocarbon-based ointment bases and bases that are either devoid of or contain lower concentrations of water require no further excipients. Exceptionally the inclusion of non-aqueous antioxidants, e.g. butylated hydroxytoluene, butylated hydroxyanisole, may be required if the therapeutic agent is prone to degradation by oxidation. Conversely, aqueous systems, e.g. poly(acrylic acid) gels (which may contain > 95% w/w water), will require the same types of excipients that are commonly employed in aqueous solutions, e.g. preservatives, buffers and water-soluble antioxidants.

Manufacture of ophthalmic ointments

The manufacture of ophthalmic ointments is very similar to that used for the formulation of ointment formulations; however, one important difference is the requirement that ophthalmic

Tips

The formulation of ocular ointments involves the use of similar components to those used in the formulation of conventional ointments; however, it should be remembered that ocular dosage forms are sterile.

Eye drop formulations are generally preferred to ocular ointments by patients due to the ease of administration and non-blurring of vision associated with eye drops.

ointments must be sterile. Due to their relatively high viscosity, terminal sterilisation of ointments is a difficult operation and may only be performed (in limited examples) at elevated processing temperatures. Therefore, ointments are typically manufactured and packaged under aseptic conditions. The components of the base are customarily prepared in an enclosed mixing vat to which heat may be applied both to aid dissolution and mixing of the ingredients and, importantly, to sterilise the ointment base. The sterile therapeutic agent may then be added to the sterile base and mixed until homogeneous. Filling of the ointment into the final container is performed under aseptic conditions.

Nasal formulations

Introduction
Nasal formulations are aqueous-based systems that are instilled within or sprayed into the nasal cavity and are predominantly employed for the treatment of congestion (ephedrine hydrochloride, pseudoephedrine hydrochloride), allergic rhinitis (e.g. beclomethasone dipropionate, sodium cromoglicate, levocabastine) and infection (mupirocin, chlorhexidine). Importantly, under the appropriate conditions, the nasal route of administration provides a rapid systemic delivery of therapeutic agents (whilst avoiding first-pass metabolism). Examples of this approach include the treatment of migraine (ergot alkaloids, sumatriptin), diabetes insipidus (desmopressin), prostate cancer/endometriosis (gonadotrophin analogues) and smoking cessation (nicotine).

Formulation considerations for nasal preparations
In addition to providing the required pharmacological activity, nasal formulations must not interfere with the cleansing action of the cilia on the nasal mucosa. This requirement has led to the avoidance of the use of non-aqueous vehicles for nasal formulations. Many of the formulation requirements of nasal solutions/sprays are based on the preservation of ciliary function and include the following.

Control of the pH of the formulation

Nasal formulations are usually buffered within the pH range of 5.5–6.5 to preserve nasal function. For this purpose buffers (e.g. citrate, phosphate) are included in these formulations. The reader should recall that the pH of the formulation affects the ionisation of therapeutic agents and this will, in turn, affect the rate of systemic absorption of the therapeutic agent following nasal absorption. This phenomenon does not affect the activity of drugs within nasal formulations that act locally within the nasal cavity, e.g. antimicrobial agents.

Tonicity of the formulation

To maintain ciliary function nasal formulations are formulated to be isotonic. The approach taken to achieve this is identical to that described for parenteral formulations, i.e. the inclusion of a suitable salt (e.g. sodium chloride).

Choice of the vehicle

As detailed above, nasal formulations are aqueous and therefore use purified water as the vehicle. Non-aqueous solvents are not used to formulate nasal dosage forms as these may interfere with ciliary function. Small concentrations of co-solvents (e.g. glycerol, polyethylene glycol, propylene glycol) may be employed to enhance the solubility of the therapeutic agent in the formulation. In addition glycerol acts as a humectant and may therefore reduce or minimise irritation to the nasal mucosa.

Addition of viscosity-modifying agents

As with other formulation types, control of the viscosity of nasal formulations is important as it directly affects both the ease of administration to, and the retention of, the formulation on the nasal mucosa. The viscosity of nasal formulations may be easily modified by the inclusion of a suitable hydrophilic polymer, e.g. methylcellulose, hydroxyethylcellulose, sodium carboxymethyl-cellulose, poly(acrylic acid). The viscosity of the formulation is usually modified to be similar to that of nasal mucus (to preserve nasal ciliary function). It must be remembered that the viscosity of the formulation must be suitable to the method of application. For example, the viscosity of nasal formulations delivered using a spray application is lower than those delivered using a bottle/dropper, due to the requirement of the former system for the formulation to be aerosolised as fine droplets.

Preservatives

Similar to ocular formulations, nasal formulations are multidose dosage forms and therefore require the addition of a preservative. The preservatives used for this purpose include:

- chlorobutanol (0.5% w/w)
- combinations of parabens (0.2% w/w total)
- benzalkonium chloride (0.002–0.02% w/v)
- thimerosal (0.002–0.005% w/v), frequently in combination with benzalkonium chloride.

The reader should be aware that there have been reports that the presence of preservatives in nasal decongestant formulations may be irritant to the nasal mucosa, resulting in rebound nasal congestion. In spite of these concerns preservatives are necessary and are therefore included in nasal formulations.

Antioxidants

These are included to enhance the chemical stability of therapeutic agents that are prone to oxidative degradation, e.g. adrenaline. As before, water-soluble antioxidants are preferred, e.g. sodium metabisulphite, sodium sulphite.

Tip

Nasal preparations must be formulated to ensure that there are no detrimental effects on ciliary function.

Manufacture of nasal preparations

The manufacture of nasal formulations is similar to that for other non-sterile solution dosage forms. The reader should consult earlier chapters, in particular Chapter 1, to acquire further details on this process.

Otic formulations

Introduction

Otic dosage forms are applied to the ear canal and are used for the treatment of infection, inflammation and/or pain. Whilst suspensions and ointments may be applied to the ear canal, the vast majority of nasal dosage forms are solutions and therefore only this category of dosage form will be described. Again, to prevent repetition, only the main formulation considerations that are directly relevant for otic formulations will be described and, when a similarity exists with previously discussed formulation types, reference will be made to the appropriate chapter(s). It will be of interest for the reader to note that the considerations for the formulation of otic ointments and suspensions are similar to those previously described in Chapters 2 and 4, respectively.

Formulation considerations for otic preparations

In common with other pharmaceutical formulations, otic formulations may be formulated as aqueous or non-aqueous solutions and contain one (or more than one) category of therapeutic agent, e.g. local anaesthetics (benzocaine), steroids (prednisolone sodium phosphate, hydrocortisone), antimicrobial agents (polymixin B sulphate, chloramphenicol), in addition to

agents that soften ear wax (e.g. carbamide peroxide). The following factors should be considered in the formulation of solutions for application to the ear.

Vehicle
The vehicle in otic formulations may be:
- aqueous, e.g. purified water
- non-aqueous, e.g. mineral oil (e.g. liquid paraffin), vegetable oil (e.g. arachis oil)
- non-aqueous but miscible with water, e.g. glycols (propylene glycol, glycerol).

Non-aqueous vehicles are predominantly used as vehicles in preparations that are used to remove ear wax, as their lipophilic character assists in the solubilisation of the wax. The choice of aqueous solvent or non-aqueous solvent as the vehicle is primarily dependent on the solubility of the therapeutic agent(s) in each of these. Accordingly, preparations containing water-soluble therapeutic agent(s), e.g. prednisolone sodium phosphate, may be prepared using water as the vehicle, whereas drugs that exhibit lower aqueous solubility may be formulated using mixtures of water and glycols, e.g. hydrocortisone or, alternatively, using the glycol solvent alone, e.g. chloramphenicol. In some cases, the drug may be formulated as a suspension using the same solvents/solvent mixtures as the vehicle. Interestingly, the hygroscopic nature of glycerol and propylene glycol is employed in otic formulations to reduce the swelling of, and remove exudate from, inflamed tissues.

Preservatives
Otic formulations are multidose and frequently aqueous systems and therefore require the inclusion of a preservative to inhibit microbial growth. The preservatives used in otic formulations are similar to those used in ocular/nasal formulations and include:
- chlorobutanol (0.5% w/w)
- combinations of parabens (0.2% w/w total)
- benzalkonium chloride (0.002–0.02% w/v)
- thimerosal (0.002–0.005% w/v).

Benzalkonium chloride and thimerosal are frequently combined in otic preparations.

Preservatives are not required in otic formulations of low water content.

Viscosity-modifying agents
As in other topical formulations, the control of product viscosity is important in the design of otic formulations, affecting both the ease of administration to, and retention at, the site of application. Modification of the viscosity of aqueous-based formulations may

be achieved using hydrophilic polymers (as described previously in this chapter). The viscosity of glycol-based (and especially glycerol-based) formulations will be greater than their aqueous counterparts and therefore viscosity modification by the inclusion of polymeric components may not be necessary.

Antioxidants

Antioxidants may be included in otic formulations to improve the stability of drugs that are prone to oxidative degradation. The choice of antioxidant is based on the solubility characteristics of the formulation.

Multiple choice questions

1. **Regarding ocular dosage forms, which of the following statements are true?**
   a. Ocular dosage forms are always sterile.
   b. Ocular dosage forms are principally employed to treat local conditions, e.g. infection, inflammation.
   c. Ocular dosage forms may be employed to treat intraocular disorders.
   d. Ocular dosage forms may be used to deliver therapeutic agents systemically.

2. **Regarding nasal dosage forms, which of the following statements are true?**
   a. Nasal dosage forms are always sterile.
   b. Nasal dosage forms are principally employed to treat local conditions, e.g. infection, inflammation.
   c. Nasal dosage forms are principally aqueous formulations.
   d. Nasal dosage forms may be used to deliver therapeutic agents systemically.

3. **Concerning ocular dosage forms, which of the following statements are true?**
   a. The retention of eye drop formulations at the site of application is poor.
   b. Elimination of eye drop formulations from the eye principally occurs *via* blinking and drainage into the lacrimal duct.
   c. The retention of ointments at the site of application is similar to that of eye drop formulations.
   d. Drug absorption following the administration of eye drop formulations frequently exceeds 5% of the originally applied dose.

4. **Regarding the cornea, conjunctiva and lacrimal fluid, which of the following statements are true?**
a. The permeability of the conjunctiva to the diffusion of therapeutic agents is similar to that of the cornea.
b. The cornea is non-vascular.
c. The cornea is multilayered in structure.
d. The lacrimal fluid is isotonic with blood.

5. **The following statements concern important factors when developing medicines for use in the eye. Indicate if each is true or false.**
a. Lacrimal fluid is usually normal in the pH range 6.0–7.4.
b. Drugs in ocular formulations are mainly absorbed through the cornea.
c. Without blinking, the eye can typically hold a volume of approximately 7 µl.
d. Lipophilic drugs are typically more readily absorbed than hydrophilic ones.

6. **Concerning the absorption of drugs following administration, which of the following statements are true?**
a. The choice of drug salt influences the subsequent absorption.
b. The pH of the formulation influences drug absorption.
c. Increasing the viscosity of the formulation results in a greater extent of drug absorption from eye drop formulations.
d. The rate of drug absorption from ointment formulations is greater than from eye drop formulations.

7. **Concerning viscosity modifiers for eye drop formulations, which of the following statements are true?**
a. Increasing the viscosity of eye drop formulations increases their retention at the site of application.
b. The viscosity of commercially available eye drop formulations is typically greater than 100 mPa/s.
c. The incorporation of viscosity-modifying agents may adversely affect the efficacy of preservatives.
d. Examples of polymers used to enhance the viscosity of eye drop formulations include poly(acrylic acid) and ethylcellulose.

8. **Concerning the use of preservatives in eye drop formulations, which of the following statements are true?**
a. Preservatives are not required in single-use eye drop formulations.
b. Anionic surfactants are commonly used as preservatives in eye drop formulations.

c. Cationic surfactants are commonly used as preservatives in eye drop formulations.

d. The use of organic mercurial preservatives may result in the deposition of the preservative within the lens of the eye.

9. With respect to the formulation of ophthalmic ointments, which of the following statements are true?

a. The therapeutic agent must be dissolved in the ophthalmic ointment formulation.

b. Ophthalmic ointments prepared using hydrocarbon bases require the addition of preservatives.

c. Ophthalmic ointments prepared using hydrocarbon bases require the addition of buffer salts to control the pH of the formulation.

d. The use of ophthalmic ointments may result in temporary blurring of vision.

10. With respect to the formulation of nasal dosage forms, which of the following statements are true?

a. Nasal dosage forms are isotonic.

b. Nasal dosage forms require the addition of preservatives.

c. Co-solvents, e.g. propylene glycol, may be added to nasal formulations to enhance the solubility of the therapeutic agent.

d. The pH of nasal formulations is typically alkaline (> pH 7).

Vaginal and rectal dosage forms

Overview

In this chapter the following points will be discussed:
- an overview/description of rectal and vaginal dosage forms and the rationale for their use
- formulation strategies for suppositories, pessaries and related products that are specifically designed for administration to the rectum or vagina
- the advantages and disadvantages of rectal and vaginal dosage forms
- considerations for the manufacture of rectal and vaginal dosage forms.

Introduction

This chapter is divided into two sections that individually describe the formulation of rectal and vaginal products. The first of these sections describes the general physiology of the rectum, the advantages and disadvantages of rectal drug delivery and strategies for the formulation of rectal dosage forms; the main emphasis is on the formulation of suppositories. In the second section the general physiology of the vagina, the advantages and disadvantages of vaginal drug delivery and strategies for the formulation of vaginal dosage forms are addressed. In a similar fashion to the section on rectal dosage forms, an emphasis is placed on pessaries, although other unique dosage forms are described, albeit briefly.

Rectal dosage forms

Advantages and disadvantages of rectal dosage forms

Advantages
- Rectal dosage forms may be successfully employed to provide a local effect for the treatment of infection and inflammation, e.g. haemorrhoids, proctitis.

KeyPoints

- Administration of drugs to the rectum and vagina may be performed to treat local disorders, e.g. infection and inflammation, or to achieve systemic absorption of the therapeutic agent in situations where alternative routes of drug administration are inappropriate.
- Suppositories are the main class of rectal dosage forms; however, creams, gels and foams are also used for this purpose.
- Pessaries, creams and gels are commonly used for the delivery of therapeutic agents to the vagina.
- The strategies for the formulation of gels and creams for rectal or vaginal administration are identical to those described in previous chapters.
- Suppositories and pessaries are semisolid or solid dosage forms that are inserted into the rectum and vagina, respectively. Advice must be given to patients regarding their administration.
- Rectal and vaginal dosage forms are non-sterile formulations.

- Rectal dosage forms are used to promote evacuation of the bowel (by irritating the rectum), to relieve constipation or to cleanse the bowel prior to surgery.
- Rectal dosage forms may be employed to provide systemic drug absorption in situations where oral drug absorption is not recommended. Examples of such applications include:
- patients who are unconscious, e.g. in intensive care or who are postoperative
- patients who are vomiting, e.g. gastrointestinal infection, migraine
- gastroirritant drugs, e.g. non-steroidal anti-inflammatory agents, particularly in chronic usage
- drugs that are prone to degradation in the stomach
- drugs that are erratically absorbed from the upper gastrointestinal tract
- administration of drugs that are extensively first-pass metabolised. If administered correctly, the therapeutic agent is absorbed directly into the systemic circulation, thereby avoiding direct entry into the liver.

  It should be noted that any drug that is absorbed following oral administration would be systemically absorbed when administered rectally.
- Rectal dosage forms may be employed to provide local treatment of diseases of the colon, e.g. Crohn's disease, ulcerative colitis.
- Following advice from the pharmacist, the administration of the rectal and vaginal dosage forms may be easily performed by the patient.

Disadvantages
- In certain countries, especially the USA and the UK, the rectal dosage forms are generally unpopular, especially for systemic administration of therapeutic agents, whereas the opposite is true in European countries.
- Specialist advice is required concerning the administration of dosage forms.
- The absorption of therapeutic agents from the rectum is slow and prone to large intrasubject and intersubject variability. The presence of faeces within the rectum considerably affects both the rate and extent of drug absorption.
- Rectal administration of therapeutic agents may result in the development of local side-effects, in particular proctitis.
- The industrial manufacture of suppositories is more difficult than for other common dosage forms.

Physiology of the rectum

A diagrammatic representation of the gastrointestinal tract, featuring the rectum, is shown in Figure 7.1.

Figure 7.1 Diagrammatic representation of the gastrointestinal tract, with particular emphasis on the rectum (see insert). (Adapted from Human Physiology en.wikibooks.org.)
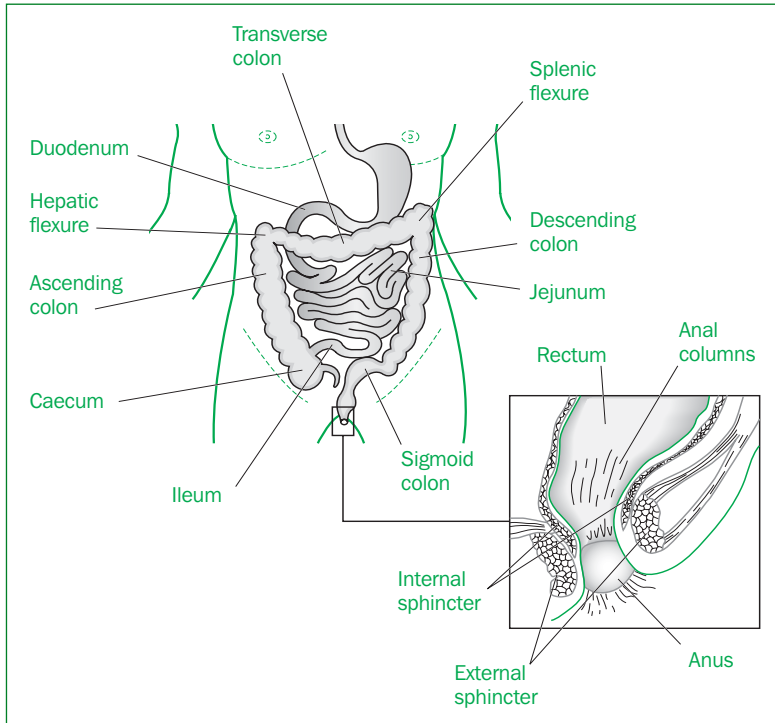
The main physiological features of the rectum that are related to drug delivery and hence to the formulation of rectal products are as follows:

- The length of the rectum is circa 15–20 cm. The rectum is joined to the sigmoid colon at the top and to the anus.
- The rectum is divided into two sections: (1) the anal canal; and (2) the ampulla. The ampulla is the larger of the two sections (approximately four times larger than the anal canal). Faeces are stored in the ampulla and excreted through the anus (a circular muscle) via the anal canal.
- Following absorption from the rectum, the therapeutic agent enters the haemorrhoidal veins. Blood from the upper haemorrhoidal vein enters the portal vein, which flows into the liver, where drug metabolism occurs. Conversely, blood in the middle and lower haemorrhoidal veins enters the general circulation (Figure 7.2).

Figure 7.2 Diagrammatic representation of blood flow into and from the rectum. (Adapted from http://www.instantanatomy.net/diagrams/AB073.jpg.)
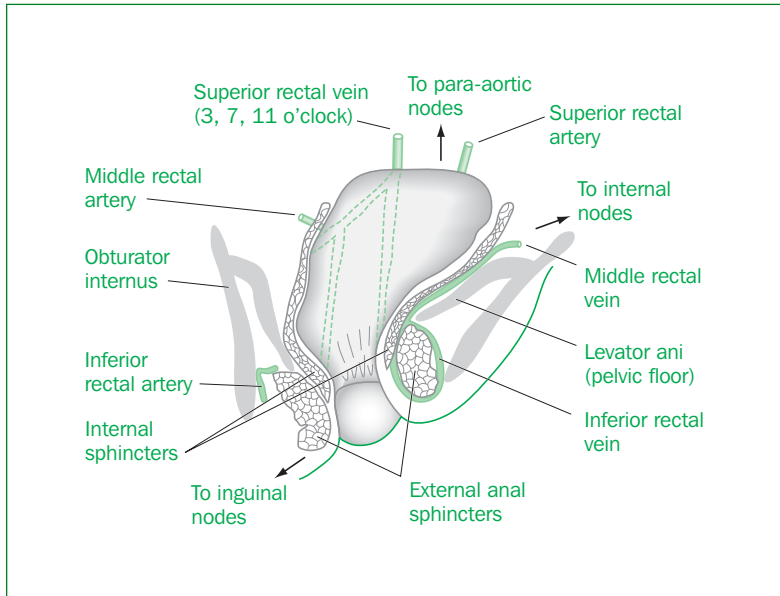
- The wall of the rectum is composed of an epithelial layer that is one cell thick. Two cellular types exist: (1) cylindrical cells; and (2) goblet cells – the latter are responsible for the secretion of mucus. There are no villi (or microvilli).
- When empty the rectum contains circa 3 ml of mucus, spread over a rectal surface area of approximately 300 cm^2.
- The pH within the rectum is essentially neutral with minimal buffering capacity. Therefore, due to the inability of the fluids within the rectum to alter the degree of ionisation, the salt form of the drug is an important determinant of the resulting local efficacy and/or systemic absorption. The presence of faecal matter will markedly affect both the dissolution of the drug in the rectal fluids and the subsequent absorption of the drug into the systemic circulation.
- As detailed above, the fate of the absorbed drug is dependent on the area of the rectum from which absorption has occurred. Drugs that are absorbed into the *inferior* and *middle haemorrhoidal veins* will enter the circulation via the *inferior vena cava* and will subsequently avoid direct exposure of the drug to, and hence metabolism by, the liver. Absorption into the *upper (superior) haemorrhoidal vein* will result in entry into the liver (and subsequent metabolism) via the portal vein.
- Examples of the classes of drugs that are administered by the rectal route for systemic absorption include:

- – antiemetics (e.g. prochlorperazine)
- – analgesics (e.g. oxymorphone hydrochloride)
- – anti-inflammatory agents (e.g. indometacin)
- – ergot alkaloids.
- ▪ There are no esterases or peptidases in the rectal fluid.
- ▪ Local muscle activity within the rectal wall may influence the rate of dissolution of solid dosage forms within the rectum, i.e. suppositories.

Factors affecting the rectal absorption of therapeutic agents

The process by which drugs are absorbed into the systemic circulation involves the dissolution of the drug in the rectal fluids (which is often preceded by the dissolution/melting of the dosage form), diffusion of the drug through the rectal fluids to the rectal mucosa and then absorption. There are various factors that affect the rate and extent of drug absorption, as follows.

Site of absorption within the rectum

As highlighted previously, the site of absorption will affect the fate of the therapeutic agent within the blood stream. Therefore the metabolism of therapeutic agents will depend on the location of the dosage form. It should be noted that it is difficult to ensure that drug absorption occurs exclusively via the inferior and middle haemorrhoidal veins and, therefore, first-pass metabolism of therapeutic agents will occur following rectal administration (although the extent of this is markedly lower when compared to drug absorption following oral administration).

The partition coefficient and degree of ionisation of the therapeutic agent

Some rectal products are lipophilic (see later in this chapter) and therefore if the therapeutic agent is also lipophilic, the release of the drug will be slow and the solubility of the drug in the rectal fluids will be low (a factor that is compounded by the low volume of the latter). Conversely, the use of (dispersed) hydrophilic drugs in lipophilic bases is preferred, as these would exhibit a greater thermodynamic tendency to dissolve in the rectal fluid. It should be noted that the limited volume of fluid in the rectum affects both the rate and extent of drug dissolution and, therefore, increasing the drug loading in suppositories does not enhance drug absorption.

   The reader will be aware that the degree of ionisation of therapeutic agents affects the resultant absorption by biological membranes, as absorption requires the presence of the unionised form of the drug. Commonly the pH and buffer capacity of

biological fluids will control the degree of ionisation and hence the absorption of the therapeutic agent. However, the poor buffer capacity of the rectal fluid results in the inability of this fluid to control the pH and hence the ionisation. Therefore, the salt form of the therapeutic agent effectively controls the degree of ionisation.

The particle size of the dispersed active agent

If the formulation is composed of an active agent that has been dispersed in the appropriate formulation base/vehicle, e.g. a hydrophilic drug dispersed in a lipophilic base or vice versa, the rate of dissolution of the drug is inversely proportional to the particle size of the dispersed active agent. A reduction in particle size of the dispersed therapeutic agent will also affect the physical stability of the formulation, as previously discussed for suspensions (Chapter 2).

The physicochemical properties of the formulation base

One of the main criteria for rectal formulations is the ability to release the drug. Any interactions between the formulation and dosage form effectively delay release and hence local action/absorption. This interaction may be specific, e.g. resulting from a specific interaction between the drug and the vehicle, or may be non-specific, in which lipophilic drugs are dissolved in lipophilic bases. The latter concern may be obviated by choice of drug salt. In addition, it is important to ensure that the formulation is non-irritant to the rectal mucosa as irritation frequently results in defecation and hence removal of the dosage form before the required mass of drug has been released.

Formulation of rectal dosage forms

There are several formulation types that are used rectally, including creams, ointments, gels, solutions and suppositories. The formulation considerations for semisolid preparations (creams, ointments, gels) and solutions that are designed for rectal use are similar to those that are formulated for non-rectal use. Therefore, for more information on these aspects the reader should consult the earlier chapters. As a result the remainder of this section will address the formulation of suppositories.

Formulation of suppositories

Suppositories are solid-dosage forms that are inserted into the rectum where they undergo softening, melting or dissolution to liberate

Tips

The location of the suppository within the rectum directly affects the route of entry of drug into the systemic circulation and hence the extent of metabolism of the drug. This is particularly relevant whenever suppositories are employed for the systemic delivery of therapeutic agents.

The formulation of the suppository directly affects the resultant onset and duration of action.

the therapeutic agent. Suppositories may also be inserted into the vagina and other accessible body cavities, e.g. the urethra and, as such, are available in a wide range of sizes and shapes, as depicted in Figure 7.3.
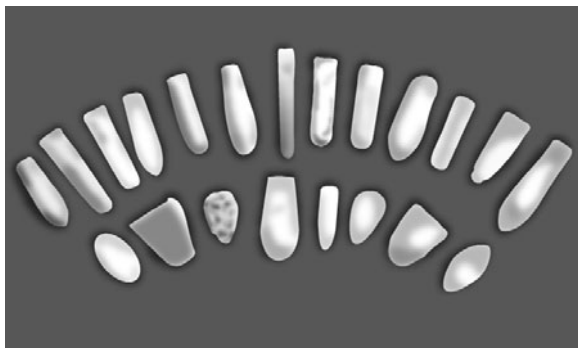
Figure 7.3 Examples of the different shapes and sizes of suppositories. Taken from Allen L V Jr (2008) *Suppositories*. London: Pharmaceutical Press.

The typical weight range for suppositories is 1–4 grams, with the 2-gram suppository being the commonly used size. The smallest suppositories are mainly reserved for use in children, whereas the largest size may be administered to adults, e.g. glycerin suppositories that are used to relieve constipation in adults. Suppositories are tapered at one end (to aid insertion) and are frequently wider in the middle before tapering towards the other end (thereby aiding retention in the rectum and enabling the suppository to be pressed forward by the anal sphincter). The drug loading of suppositories ranges from circa 0.1 to 40% w/w. In general, suppositories are composed of an inert base into which the therapeutic agent is incorporated (dissolved/dispersed). The physicochemical properties of both the therapeutic agent and suppository base are important determinants of the clinical and non-clinical performance of suppositories. The role of the solubility, salt type and particle size on the dissolution and hence pharmacological activity of rectal dosage forms, e.g. suppositories, has been addressed earlier in this chapter. The next subsection describes the types and physicochemical properties of suppository bases.

Suppository bases

There are a number of key properties that an ideal suppository base should exhibit. These are as follows:

- The base should be solid at the storage temperature of the formulation but should soften, melt or dissolve in the rectal

fluid following insertion into the rectum, thereby releasing the therapeutic agent.

- The base should be non-irritant to the rectal mucosa.
- The base should not chemically interact with the therapeutic agent.
- The base should be chemically and physically stable over the period of storage (shelf-life).

There are two main categories of suppository base: (1) fatty (oleaginous) bases; and (2) water-miscible bases. A third (sub-) category may also be used that consists of lipophilic and hydrophilic excipients and therefore shares properties common to both fatty and water-miscible bases.

Fatty (oleaginous) bases

Fatty bases are predominantly composed of naturally occurring or semisynthetic/synthetic fatty acid esters of glycerol. These systems are designed to melt within the rectum thereby facilitating drug release and subsequent dissolution. There are several types of fatty base.

Cocoa butter (theobroma oil)

This is a natural material that consists of a mixture of fatty acid esters of glycerol, e.g. stearic, palmitic and oleic, predominantly triesters, e.g. glyceryl tripalmitate. The presence of unsaturated (e.g. oleic acid) esters contributes to the low melting point of cocoa butter (30–36°C), thereby facilitating cocoa butter melting following insertion within the rectum. The incorporation of lipophilic drugs into cocoa butter has been reported to lower the melting-point range of suppositories produced using this base, which may lead to stability problems and may result in suppositories that are too soft to insert. To overcome this problem, beeswax (circa 4% w/w) or cetyl esters wax (circa 20% w/w) may be added to the suppository (thereby increasing the melting temperature). Cocoa butter is safe, non-toxic and non-irritating.

One major problem with the use of cocoa butter as a base for suppositories is polymorphism, i.e. the ability of this material to exist in different crystalline forms; this is accredited to the high content of triglycerides. Cocoa butter exists in four polymorphic forms: (1) alpha (α, melting point circa 20°C); (2) beta (β, melting point circa 34–35°C); (3) beta prime (β′, melting point circa 28°C); and (4) gamma (γ, melting point circa 34–35°C). If cocoa butter is melted at circa 36°C and allowed to solidify slowly, the stable (β) polymorph will form. However, if the temperature is markedly elevated above 36°C and allowed to cool, the other polymorphic forms will be produced. Being unstable, the αβ and β′ polymorphs will slowly revert to the stable β polymorph.

However, as this conversion may require several days, there may be associated stability issues (notably poor setting properties or remelting of the suppositories following manufacture).

It should be noted that the melting/softening point of cocoa butter may be inappropriate (too low) for storage at room temperature and therefore either storage under controlled temperature or, as detailed previously, the inclusion of excipients that raise the melting/softening temperature of cocoa butter (e.g. beeswax, cetyl esters wax) may be required.

Semisynthetic/synthetic triglycerides (Hard Fat BP, US Pharmacopeia National Formulary (USPNF) or Adeps solidus PhEur (European Pharmacopeia)

The main problems associated with cocoa butter (polymorphism and variation in composition) led to the development and use of suppository bases that were based on synthetic/semisynthetic glycerides. Typically these systems are composed of mixtures of triglycerides of higher *saturated* fatty acids (ranging from $C_8H_{17}COOH$ to $C_{18}H_{37}COOH$) and di/monoglycerides. Semisynthetic/synthetic triglyceride bases are safe, non-toxic and non-irritating and, unlike cocoa butter, can be reheated during processing (whilst not affecting the solidification temperature) and exhibit low batch-to-batch variability.

Choice of synthetic/semisynthetic base for use in the formulation of suppositories

There are several different grades of synthetic/semisynthetic suppository bases that are commercially available and are currently used in the formulation of suppositories. These bases differ in a number of physicochemical properties, including the following.

Hydroxyl number

This refers to the presence of hydroxyl groups, which in turn is a measure of the presence of mono- and diglycerides in the suppository base. The available suppository bases range in hydroxyl numbers from circa 5 to 15, e.g. Witespol H15, H175, H185, E75, E85 to > 55, e.g. Witespol S51 (55–70), S58 (60–70). The use of bases exhibiting high hydroxyl numbers should be reserved for therapeutic agents that exhibit low reactivity towards this chemical group. It should also be noted that as the hydroxyl number of the base increases so does the relative hydrophilicity. This may affect drug release and subsequent absorption.

Melting properties

There is a wide range in the melting points of commercially available synthetic/semisynthetic suppository bases. For example, Suppocire AIP and Witespol H32 exhibit low melting points

(30–33°C and 31–33°C, respectively). Conversely, the melting points of other bases in these product ranges may exceed 40°C, e.g. Suppocire D, ND and DM (42–45°C) and Witespol E85 (42–44°C). Low melting bases (< 37°C) are generally used for the formulation of suppositories in which systemic absorption of the therapeutic agent is desired, whereas bases of higher melting points are frequently used in formulations in which a local effect is desired.

The melting properties of these bases and, in addition, the rheological properties of the molten bases are important considerations for the successful formulation of suppositories. The importance of each of these topics is presented below.

Melting point

In the formulation of suppositories that are intended to provide systemic administration of the therapeutic agent, the selected base should melt to facilitate drug release and hence absorption. On the other hand, the bases of suppository formulations that are designed to provide a local effect do not necessarily have to melt: it is sufficient for them to soften for the intended use.

The melting properties of the base should be considered in conjunction with the solubility of the chosen therapeutic agent. If the active agent is soluble in the base, this will lead to a reduction in the melting point of the base. Under these circumstances a suppository base may be used that offers a higher melting point, e.g. Witespol E75 or E85.

Consideration of the solubility of the therapeutic agent within both the molten and cooled base is important. If the drug is more soluble in the base during the manufacturing process (in which heat is applied to melt the base), the drug may precipitate out following cooling (particularly during storage). This may produce different polymorphic forms of the drug or crystals of a different size fraction, both of which will result in a change in the performance of the product.

Viscosity of the melted base

The viscosity of the melted base can affect the performance of suppositories in two ways:
1. During the manufacture of suppositories the base is melted and into this the drug may be dispersed. The molten base (containing the dispersed drug) is then poured into the appropriate mould prior to cooling. The viscosity of the base will therefore affect both the mixing of the drug with the molten base and, in addition, the flow of the molten dispersion into the moulds. Furthermore, the dispersion of solid drug particles into the molten base will dramatically

affect the viscosity of the product. Therefore, in selecting a
suppository base, the viscosity of the molten base is a major
consideration.

2. The viscosity of the melted base also affects the spreading of
    the formulation on the rectal mucosa and the subsequent drug
    release. Bases with high viscosity will exhibit poorer
    spreading properties and slower rates of drug release when
    compared to bases of lower melt viscosity.

Water-soluble and water-miscible bases

There are two main categories of suppository base in this
classification: (1) glycerol–gelatin base (which dissolves in the
rectal fluids); and (2) water-miscible bases, composed of
polyethylene glycols (PEGs).

Glycerol–gelatin

This suppository base is prepared by dissolving gelatin (circa
20% w/w) in glycerol (70% w/w) with the aid of heating (circa
100°C). The required drug is generally dissolved/dispersed in an
aqueous phase (< 10% w/w) and then combined with the
glycerol phase with stirring prior to pouring into the suppository
mould. As an alternative method the gelatin may be dissolved in
the heated aqueous phase, into which the drug is dispersed/
dissolved and then added to the heated glycerol. The mechanical
properties of the formed suppository may be manipulated by
increasing or decreasing the mass of added gelatin. For example,
as the mass of added aqueous phase is increased, the
suppositories will become softer, thereby leading to potential
problems regarding their insertion by the patient. Increasing the
concentration of gelatin will negate this effect, e.g. in Gelato-
glycero suppositories BPC the concentrations of aqueous phase,
gelatin and glycerol are 27.5% w/w, gelatin 32.5% w/w and 40%
w/w, respectively. In urethral suppositories the ratios of gelatin to
glycerol to water (containing the active agent) is 3:1:1: the
enhanced mechanical properties facilitate insertion into the
urethra.

Due to the presence of water in glycerol–gelatin suppositories
it is necessary to include preservatives, e.g. mixtures of methyl
and propylhydroxyparahydroxybenzoic acid (total concentration
0.2% w/w). These are dissolved within the aqueous phase during
the manufacturing process.

Glycerol–gelatin bases may be used for the formulation of
suppositories that contain a water-soluble therapeutic agent.
However, the use of this type of base is restricted by several
disadvantages, including:

■   *An associated physiological effect.* Glycerol–gelatin
    suppositories will induce defecation and, indeed, are used for

this purpose to relieve constipation or to facilitate bowel evacuation prior to surgery.

- *Difficult to manufacture.* Suppositories prepared using glycerol–gelatin bases are generally more difficult to manufacture than other types of base, e.g. triglyceride systems.
- *Hygroscopic.* Glycerol–gelatin bases will absorb moisture from the atmosphere and therefore must be carefully packaged to prevent moisture uptake and, ultimately, to maintain both the shape and mechanical properties of the suppository. This ability of glycerol–gelatin bases to absorb water will also occur within the rectum, leading to dehydration and irritation of the rectal mucosa. This action prompts bowel evacuation. To minimise this phenomenon, the suppository may be moistened with water prior to insertion.
- *Potential interactions with therapeutic agents.* Gelatin is a protein that is obtained following partial acid hydrolysis (termed type A) or partial alkaline hydrolysis (termed type B) of animal collagen. The isoelectric point of type A ranges between 7 and 9 whereas the isoelectric point of type B lies between 4.7 and 5.3. Accordingly the pH of aqueous solutions of each gelatin type will affect ionisation; type A acts as a base if formulated at a pH below 4.7 whereas type B will act as an acid if the pH is raised above 5.3. As a result of this a possibility arises for gelatin to interact with therapeutic agents of opposite charge in a typical acid–base interaction. This will lower the solubility of gelatin and may lead to precipitation. Therefore it is important to select the pH of the aqueous phase and gelatin type carefully when formulating suppositories containing acidic or basic drugs.

Water-miscible bases

Water-miscible bases are composed of PEGs possessing a molecular weight greater than 1000 g/mol. The melting point of these higher grades of PEGs increases as the molecular weight increases, e.g. the melting points of PEG 1000 and PEG 8000 are 37–40°C and 60–63°C, respectively. Typically the melting point of PEG suppository bases is circa 42°C; this is generally achieved and controlled using the appropriate mixtures of grade of this polymer. The higher melting point of these systems obviates the need for storage under cold conditions. In addition to controlling the melting point, different molecular weights of this polymer may also be blended to control the mechanical properties of PEG-based suppositories. In this scenario, the lower-molecular-weight PEGs, e.g. PEG 400, will act to reduce the brittle behaviour of these suppository bases.

Following insertion into the rectum, these suppositories will not melt but, due to their hygroscopic properties, will instead gradually dissolve (the volume of rectal fluid is too small to allow rapid dissolution) and, in so doing, will enable drug dissolution to occur. This ability to absorb moisture may lead to patient discomfort due to the extraction of water from the rectal mucosa into the suppository; however, this may be minimised by the inclusion of water (> 20% w/w) and by moistening the suppository prior to insertion. As before, PEG-based suppositories will require storage in moisture-resistant packaging. There are two final concerns regarding the use of PEG-based suppositories. PEG is known to enhance the solubility of therapeutic agents and therefore this interaction between the drug and polymer may affect the subsequent release of the drug from the liquefied base. Secondly, the solubility of the drug in the solid base may change as functions of both storage conditions and time and this may result in crystal growth within the suppository. This in turn may affect the mechanical properties of, and drug release from, the suppository.

> **Tip**
>
> Semisynthetic suppository bases are generally used for the formulation of suppositories.

Formulation excipients that may be used in the formulation of suppositories

As with other formulation types, the formulation of successful suppositories may necessitate the inclusion of other pharmaceutical excipients. The list of possible excipients is more limited for suppositories than for other dosage form types that have been discussed in this textbook to date. Examples include: (1) surface-active agents; (2) agents to reduce hygroscopicity; and (3) agents to control the melting point of the base.

Surface-active agents

These are included to enhance the wetting properties of the suppository base with the rectal fluid. This in turn will enhance drug release/dissolution. The use of surfactants is mainly reserved for formulations composed of a lipophilic suppository base and/or a lipophilic drug. Examples of excipients that are included in suppository formulations include sorbitan esters and polyoxyethylene sorbitan fatty acid esters.

Agents to reduce hygroscopicity

Agents that reduce hygroscopicity, e.g. colloidal silicon dioxide, may be included in fatty suppository bases to reduce the uptake of water from the atmosphere during storage and, in so doing, the physical and chemical stability of the dosage form/therapeutic agent may be enhanced.

The hygroscopicity of water-soluble or water-miscible bases is large and is indeed partly responsible for the in vivo performance of formulations base. Water uptake during storage will result in changes to the mechanical properties (softening) and shape of these dosage forms. Accordingly, protection against water uptake during storage is afforded by the use of moisture-resistant packaging.

Agents to control the melting point of the base

As previously mentioned, the melting point of the base may be manipulated to enhance the mechanical properties and physical stability of the suppository in response to the deleterious effects of storage at higher temperature and/or the presence of a therapeutic agent that is soluble in the suppository base. Examples of excipients that are commonly used to increase the melting point of suppositories prepared using fatty bases include:

- beeswax (white or yellow wax)
- cetyl esters wax USPNF
- stearic acid
- atearic alcohol
- aluminium mono- or distearate
- colloidal silicon dioxide
- magnesium stearate
- bentonite.

Conversely there may be a requirement to reduce the melting point of the fatty suppository base, e.g. to enable melting within the rectum. Examples of excipients that may be used for this purpose include:

- glyceryl monostearate
- myristyl alcohol
- polysorbate 80
- propylene glycol.

As detailed previously, the melting point of PEG-based suppositories may be controlled by altering the grade of PEG used. In this approach the melting point may be increased by the inclusion of higher-molecular-weight PEG. Conversely, the melting point of water-miscible suppository bases may be lowered by incorporating the required concentration of low-molecular-weight PEG (PEG 400).

Manufacture of suppositories

Suppositories are manufactured principally using a moulding method, i.e. a method in which the base is heated to above the melting temperature, the drug dispersed (or dissolved) in the heated liquid prior to dispensing the molten product into suppository moulds (Figure 7.4).

Figure 7.4 Suppository mould used in the manufacture of suppositories. (a) Armstrong suppository mold. Photo courtesy of Professional Compounding Centers of America, Houston, TX. Taken from Allen L V Jr (2008) *Suppositories*. London: Pharmaceutical Press. (b) Plastic mould filled with the suppository formulation. (Courtesy of Dr Ryan Donnelly, School of Pharmacy, Queen's University of Belfast.) (c) Plastic-filled suppository mould and formed suppositories. (Courtesy of Dr Ryan Donnelly, School of Pharmacy, Queen's University of Belfast.)

(a)
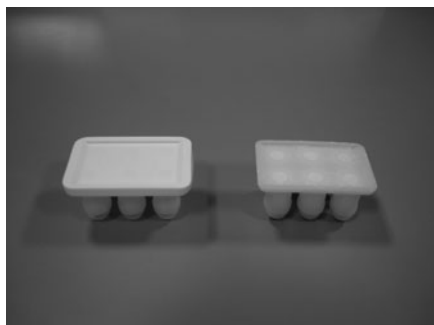
(b)
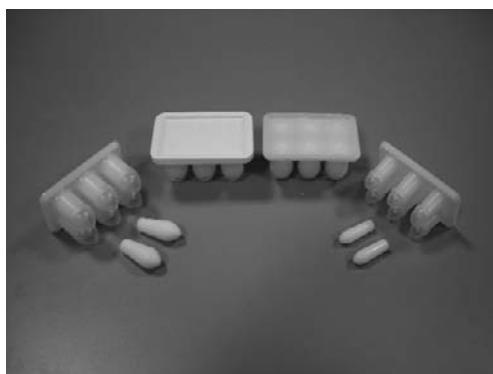
(c)

In the laboratory the suppository formulation is heated in a small vessel (typically an evaporating basin) and then manually poured into the mould. This is then allowed to cool (usually in a refrigerator) prior to removal and packaging. Suppositories prepared using cocoa butter or glycerol–gelatin bases are prone to sticking in the metallic suppository mould after cooling. To prevent this, the moulds are usually precoated (e.g. arachis oil may be used for glycerol–gelatin suppositories whereas an alcohol/surfactant mixture is commonly used for suppositories prepared using cocoa butter). The composition of the lubricant must differ from that of the base to prevent any interaction.

Alternatively, plastic moulds may be used that do not require lubrication (Figure 7.4 b, c). In large-scale manufacture, the preparation is heated within a vessel and the molten dosage form is automatically dosed into the mould, cooled and mechanically removed. The suppositories are then automatically packaged into the final container. Alternatively the molten formulation may be injected directly into the final packaging (termed mould strips), which are then sealed.

Calculation of the mass of base required

One concern regarding the manufacture of suppositories is the calculation of the mass of base that is required. The volume of each suppository mould is known (and has been calibrated) and therefore the molten formulations are dispensed into the moulds according to volume. However, if the drug is *dispersed* in the molten formulation, the volume of the formulation will be dependent on the mass of drug present (remembering that solids displace an equal volume of base). To ensure that the correct volume of base is used, a calculation is performed based on the *displacement value*, i.e. the ratio of the weight of the drug to the weight of base displaced by the drug. The displacement factor may be visualised as the weight of drug required to displace unit weight of base. For example, the displacement value of cinchocaine hydrochloride is 1.5 for cocoa butter (and indeed other triglyceride bases). Therefore 1.5 grams of this therapeutic agent will displace 1 gram of cocoa butter.

In practice the displacement value is calculated as follows:

- The average weight of the suppository mould is calculated using the blank suppository base (the molten base is added to the correct volume, allowed to cool and then weighed).
- The weight of drug needed for the total number of suppositories is calculated (weight of drug per suppository × number of suppositories manufactured).
- Suppositories are then prepared by adding the mass of drug to the notional mass of suppository base, melting and then dispensing into the suppository moulds. The weight of the cooled suppositories is then determined.

Examples of pertinent calculations are shown below.

Worked examples

Example 7.1

Determine the displacement value for a therapeutic agent in a triglyceride base. The concentration of drug in each suppository should be 10%.

Step 1
Prepare (e.g. six) blank suppositories and weigh.
Total weight determined as 6 grams (1 gram per suppository).

Step 2
Prepare six suppositories containing 10% drug in each
suppository (based on the volume of the suppository mould) and
weigh.
Total weight determined as 6.11 grams.

Step 3
Calculate the masses of base and drug in the six suppositories
(remembering that the drug loading of the suppositories is 10%).

$$Base = \frac{90}{100} \times 6.11 = 5.50g$$

$$Drug = \frac{10}{100} \times 6.11 = 0.61g$$

Step 4
Calculate the mass of base displaced by the 0.61 gram of drug.

$$6.0 - 5.5 = 0.5g$$

Step 5
Calculate the displacement value for the drug.

$$DV = \frac{0.61}{0.50} = 1.22$$

A list of displacement values for a range of drugs is available and
these are used to calculate the required weight of base. The use of
these values is illustrated in the next example.

Example 7.2
You have been asked to prepare six (2-gram) suppositories
containing 5% cinchocaine hydrochloride (displacement value of
1.5). Calculate the final formulae for the requested dosage form.
    As this is a small batch, an overage must be included in the
calculation. Therefore, the weights required for nine suppositories
will be determined:
▪   Calculate the weight of drug required for nine suppositories:

$$Wt(g) = \left(\frac{5}{100} \times 2\right) \times 9 = 0.90g$$

- Calculate the weight of base displaced by the desired weight of drug (900 mg):

$$DV = \frac{Wt_{(Drug)}}{Wt_{(BaseDisplaced)}} = 1.5 = \frac{0.90}{Wt_{(BaseDisplaced)}}$$

$$Wt_{(BaseDisplaced)} = \frac{0.9}{1.5} = 0.60g$$

- Determine the amount of base required:
  Weight of base required = (theoretical weight – weight of base displaced)

$$Wt_{(Required)} = 18.0g - 0.6g = 17.4g$$

- Define final formulation (Table 7.1).

Table 7.1 Calculated composition of the suppository formulation described in example 2

Formulation component	Weight (g) for nine suppositories
Cinchocaine hydrochloride	0.90
Suppository base	17.40

Note: the final weight of the formulation does not necessarily add up to 9 grams. Thus, in the above formulation the weight of each suppository is 2.03 g, i.e.: $\left(\frac{17.40 + 0.90}{9}\right)$.

Although the above calculations are primarily directed to therapeutic agents that are dispersed within suppositories prepared using fatty bases (e.g. cocoa butter, semisynthetic/synthetic triglycerides), this argument may be extended to water-soluble and water-miscible systems in which the drug has been dispersed. Furthermore, the density of glycerol–gelatin suppositories is 1.2 g/cm³, and therefore, after consideration of the displacement values (as described earlier in this section), the calculated amount of base is increased by a factor of 1.2. Displacement values are not considered in formulations in which the drug is soluble in the base.

Vaginal dosage forms

The administration of drugs to the vagina is principally performed to achieve a local effect, e.g. for the treatment of infection. However, it is also known that, due to the highly vascular nature of the vaginal wall, systemic absorption of therapeutic agents may occur following administration by this route. Furthermore, absorption into the systemic circulation by this route avoids first-pass metabolism in the liver. Akin to the

rectum, the fluid content within the vagina is low under normal physiological conditions.

Advantages and disadvantages of vaginal dosage forms

Advantages

- Vaginal dosage forms may be successfully employed to provide a local effect for the treatment of infection (fungal and bacterial) and for the treatment of hormone deficiency (topical hormone replacement therapy for the treatment of vaginal atrophy).
- Vaginal dosage forms may be employed to provide systemic drug absorption of certain therapeutic agents. Examples of these applications include:
- steroid hormones (e.g. oestradiol acetate) for the treatment of the systemic symptoms of the menopause and for long-term contraception
- prostaglandins (for cervical ripening).

Figure 7.5 Diagram of the vaginal physiology. (Adapted from Hyde J S, DeLamater J (2003) *Understanding Human Sexuality*, 8th edn. Boston: McGraw-Hill.)
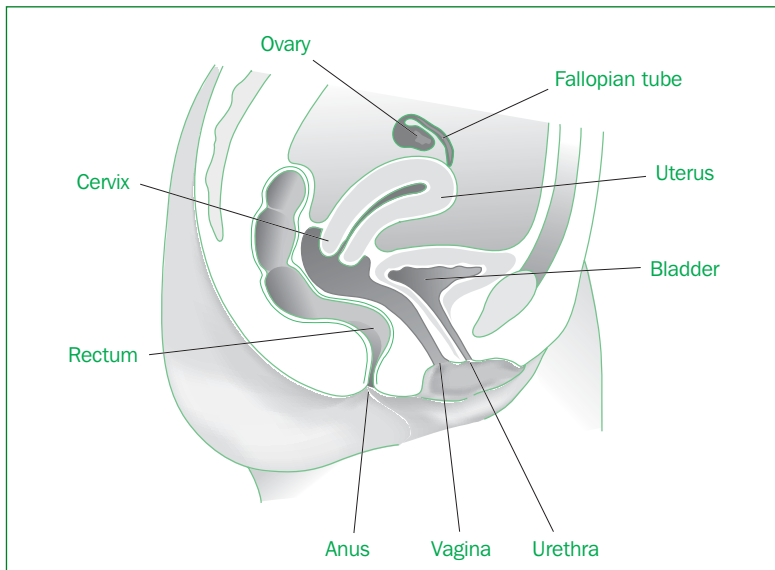
Disadvantages

- Whilst the use of topical preparations for the treatment of local disorders is accepted, the use of vaginal dosage forms for systemic drug administration is not as popular.
- Specialist advice is required concerning the administration of dosage forms to ensure correct placement and hence retention in the vagina. This may often require the use of special applicators.

- The industrial manufacture of vaginal pessaries (suppositories) is more difficult than for other common vaginal dosage forms.

Vaginal physiology

The key aspects of vaginal physiology (Figure 7.5) that are related to drug delivery (and hence formulation) are as follows:

- There are ridges (folds) in the vagina, termed vaginal rugae, that increase the surface area of, and hence drug absorption from, the vagina. In addition the vaginal rugae will enhance the retention of the dosage form in the vagina (in particular solid and semisolid systems), thereby facilitating prolonged drug release to this site.
- The vaginal epithelium is composed of stratified squamous cells. The upper section of the vagina is a mucous membrane. Vaginal mucus is produced within the vagina, principally by the cervix, which, under normal physiological conditions, results in a limited volume of fluid within the vagina. Both the volume and consistency of vaginal mucus changes during the menstrual cycle. The limited volume of fluid within the vagina will limit the dissolution of moderate to highly lipophilic drugs.
- The vagina is highly vascular; a plexus of arteries and veins is located round the vagina. The venous blood supply from the vagina does not enter the portal system and therefore first-pass metabolism does not occur. This renders the vagina a useful site for the systemic administration of therapeutic agent.

Types of vaginal dosage forms

There are several types of dosage forms that are administered to the vagina. The formulation considerations for many of these types are similar to those described previously (and subsequently in more specialised chapters). Therefore only a brief overview of the specific formulation considerations that are relevant to vaginal dosage forms will be considered in this chapter. The main types of dosage forms that are administered via the vagina to achieve a local or systemic effect are: (1) semisolid formulations; (2) tablets and capsules; (3) pessaries (vaginal suppositories); and (4) vaginal implants.

Semisolid formulations

Semisolid formulations (creams, ointments and gels) are commonly administered to the vagina (and vulva) and typically contain antimicrobial agents, hormones or contraceptive agents. The administration of these to the vagina may require a special applicator (nozzle). The formulation of creams, ointments and gels for vaginal administration is generally the same as previously

described; however, in vaginal products the pH may be modified to match that of the vaginal secretions (circa 4.5). In addition, foams may be used as a drug delivery system for the vagina. These are aerosol products based on oil in water creams (aerosol products will be discussed in Chapter 8).

Tablets and capsules

Vaginal tablets and capsules are commonly used dosage forms for the delivery of antimicrobial agents and steroid hormones. Due to the relatively low content of aqueous fluid in the vagina under normal physiological conditions, disintegrants may be incorporated into tablets to assist in tablet disintegration and subsequent drug dissolution. The formulation of tablets and capsules is described in Chapters 9 and 10. The administration of tablets and capsules to the vagina requires the use of an applicator. The patient should be advised by the pharmacist on the correct method to insert these dosage forms into the vagina (usually towards the back of the vagina).

Pessaries (vaginal suppositories)

Pessaries (often termed vaginal suppositories) are visibly similar to rectal suppositories and are used for the treatment of infection (yeast and bacterial), vaginal atrophy and for contraceptive purposes. In addition, pessaries may be used to achieve systemic absorption of therapeutic agents. Pessaries are formulated using glycerol–gelatin or PEG bases, into which the drug is incorporated (as previously described in this chapter). The pH of pessaries that contain an aqueous phase may be buffered to pH 4.5. The administration of these dosage forms generally requires an applicator and training (akin to tablets and capsules).

Vaginal implants

Vaginal implants are solid formulations that provide a controlled release of therapeutic agent into the vaginal fluids and may be used to achieve either a local effect or systemic absorption. One example of this in Femring, a doughnut-shaped implant that is located adjacent to the cervix and which provides controlled systemic absorption of oestradiol. The mechanical properties of the product ensure excellent retention of the dosage form at the site of application.

Multiple choice questions

1. **Regarding suppositories, which of the following statements are true?**
a. Suppositories are employed for the treatment of local disorders, e.g. haemorrhoids.

b. Suppositories may be used to deliver therapeutic agents systemically.

c. Suppositories are sterile products.

d. Suppositories are normally formulated with the inclusion of colourants.

2. **Regarding pessaries, which of the following statements are true?**

a. Pessaries are employed for the treatment of local disorders, e.g. infection.

b. Pessaries may be used to deliver therapeutic agents systemically.

c. Pessaries are sterile products.

d. Pessaries are normally formulated with the inclusion of preservatives.

3. **Regarding the physiology of the rectum, which of the following statements are true?**

a. The rectum is joined to the bottom of the sigmoid colon.

b. The rectum is divided into two sections termed the anal canal and the ampulla – the ampulla is the smaller of the two sections.

c. The rectal surface area is relatively small, i.e. < 100 cm².

d. Following absorption, drugs enter the systemic circulation via the haemorrhoidal vein.

4. **Factors affecting drug absorption from the rectum include:**

a. The pH of the rectal fluid.

b. The presence of faecal matter.

c. The presence of esterases in the rectal fluid.

d. The location of the suppository within the rectum.

5. **Regarding the formulation of suppositories, which of the following statements are true?**

a. Suppositories should melt or dissolve within the rectum.

b. The typical weight range for suppositories is between 1 and 4 grams.

c. The concentration of drug within suppositories typically ranges from 0.1 to 40% w/w.

d. Suppositories formulated using oleaginous bases require the addition of preservatives.

6. **Concerning the use of theobroma oil in the formulation of suppositories, which of the following statements are true?**

a. It is a natural product consisting of a mixture of fatty acid esters of glycerol.

b. It is a pure substance.

c. It exists in different polymorphic states, the number and type of which are dependent on the processing temperature.
d. The melting temperature of suppositories that have been formulated using theobroma oil may be enhanced by the addition of beeswax.

7. **In the formulation of suppositories the following excipients may be used for the following reasons:**
a. Surface-active agents – to facilitate manufacture of suppositories.
b. Propylene glycol – to reduce the melting point of suppository bases.
c. Colloidal silicon dioxide – to reduce the hygroscopicity of fatty suppository bases.
d. Ethanol – to enhance the solubility of the therapeutic agent within the suppository base.

8. **Concerning vaginal physiology, which of the following statements are true?**
a. The vagina is highly vascular.
b. Following absorption from the vagina, the drug enters the portal system and hence first-pass metabolism occurs.
c. There is a limited volume of fluid within the vagina into which drug dissolution occurs.
d. There is a high available surface area with the vagina for drug absorption.

9. **Concerning vaginal dosage forms, which of the following statements are true?**
a. Creams and ointments may be used for vaginal administration.
b. Pessaries are frequently formulated using oleaginous bases.
c. Vaginal tablets are generally formulated to include disintegrants.
d. Type 1 gels are commonly used as vaginal dosage forms.

10. **The following agents may be used as the basis of suppository formulations:**
a. Triglycerides of higher saturated fatty acids.
b. Poly(acrylic acid).
c. Poly(ethylene) glycols.
d. Glucose.

chapter 8
Respiratory dosage forms

Overview

In this chapter the following points will be discussed:
- an overview/description of respiratory dosage forms and the rationale for their use
- formulation strategies for aerosols and related products that are specifically designed for administration to the respiratory tract
- the advantages and disadvantages of respiratory dosage forms
- considerations for the manufacture of respiratory dosage forms.

Introduction

Drug delivery to the respiratory tract is principally performed for the treatment of local disorders, e.g. asthma and cystic fibrosis. However, due to the excellent blood perfusion of this organ, large surface area of the alveoli and the thin barrier for absorption, this route of administration may be employed to achieve drug absorption into the systemic circulation without direct passage to the liver. These features have been commercially exploited for the systemic delivery of insulin, ergotamine and other drugs for which oral absorption is inappropriate. There are several methods by which therapeutic agents may be successfully delivered to the lungs, including the use of aerosols, dry-powder inhalers (DPIs) and nebulisers. Uniquely, the performance of the respiratory drug delivery system is dependent on both the formulation and the nature/properties of the delivery system (i.e. the inhalation system).

Advantages and disadvantages of respiratory drug delivery

Advantages
- Respiratory drug delivery ensures that the required dose of drug is delivered to the site of action

KeyPoints

- Administration of dosage forms to the respiratory tract is performed for the treatment of localised disorders, e.g. asthma (β_2-agonists, corticosteroids), infections (gentamicin, ciprofloxacin). However, due to the extensive blood supply to the alveoli, this route of administration may be used to gain systemic absorption (without first-pass metabolism). The size of the inhaled solution/particles is a key determinant of the resulting clinical activity.
- The main types of dosage form that are administered by the respiratory route are solutions (for nebulisation) and particles (the latter either as a drug powder or as an aerosol). The control of particle size of solutions is performed by the nebuliser whereas the control of the particle size for dry powders/aerosols is defined by the size of the particle used in the formulation and the possible aggregation either in the solid or suspended state.

(where local effect is required). In so doing the incidence of side-effects is minimised.

▪ Due to the large surface area of the lungs (i.e. the alveoli), the excellent blood supply and the thin nature of the barrier between the lung and the systemic circulation, the lungs act as a portal for systemic drug absorption. Therefore, respiratory drug delivery systems offer a viable alternative to parenteral medications.

▪ There is a rapid onset of action following respiratory drug delivery. This is particularly beneficial for the treatment of asthma and, more recently, for the lowering of postprandial blood glucose levels.

▪ The delivery system used in respiratory dosage forms is, in most instances, portable and is therefore convenient for the patient to carry.

Disadvantages

▪ In some dosage forms (e.g. conventional aerosols) coordination is required between activating the inhaler and inspiration. Failure to do so results in the deposition of the drug in the upper airways. There are respiratory delivery systems that do not require this coordination; however, many of these are bulky and therefore conspicuous in use.

▪ Deposition of drug to the lower airways may be impeded in the presence of high volumes of mucus (e.g. due to an infection).

▪ The physical stability of pharmaceutical aerosols may be problematic.

Physiology of the respiratory tract

The key physiological aspects of the respiratory tract that are pertinent to respiratory drug delivery are as follows:

▪ The physiological role of the respiratory tract is in the transfer of oxygen into the blood from inspired air and the removal of carbon dioxide from the blood into the expired air.

▪ The respiratory tract (Figure 8.1) may be subdivided into several regions:

– The upper respiratory tract, comprising:
  ● nose
  ● throat
  ● pharynx
  ● larynx.

Figure 8.1 Diagrammatic representation of the respiratory tract. (Adapted from http://faculty.ircc.edu/faculty/tfischer/micro%20resources.htm.)
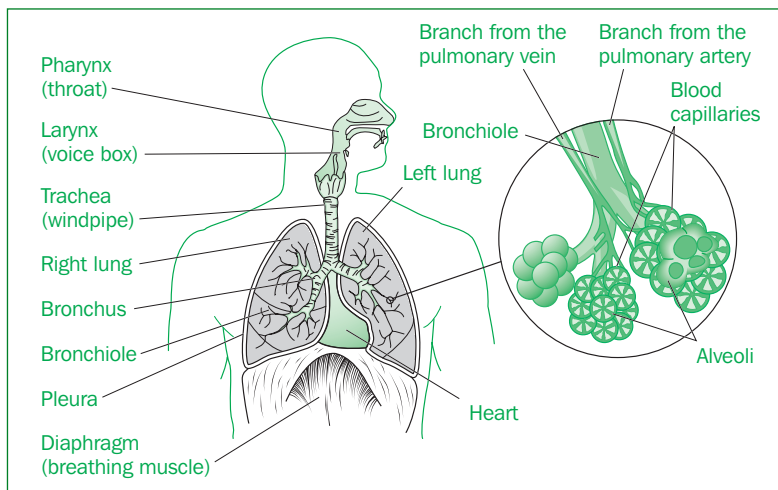
- The lower respiratory tract, comprising:
  - trachea, dividing into:
  - bronchi, dividing into:
  - bronchioles, dividing into:
  - alveoli.
- The role of the upper respiratory tract and the trachea, bronchi and bronchioles is in the conductance of air to and from the alveoli. The role of the alveoli is in the transfer of respiratory gases.
- The diameters of the conducting airways sequentially decrease towards the alveoli. This has a profound effect on the deposition of particles/droplets within the respiratory tract.
- The bronchioles divide into a series of alveolar sacs (Figure 8.1), each of which contains circa $2-6 \times 10^8$ alveoli. An extensive capillary network is associated with these alveolar sacs and, furthermore, the diffusion barrier between the blood supply and the alveoli is relatively thin, enabling rapid drug diffusion and absorption into the blood stream. The combined surface area of the alveoli is large ($70-80$ m^2), which, in addition to the points raised previously, makes the respiratory tract particularly suitable for the systemic absorption of therapeutic agents.
- The epithelial cells in the conducting airways are ciliated and are responsible for the transport of foreign particles upwards towards the upper respiratory tract for subsequent elimination. Furthermore, goblet cells produce mucus which acts to trap foreign particles (prior to removal by the ciliated epithelia).

Factors affecting the deposition of particles/droplets within the respiratory tract

All respiratory dosage forms deliver the drug to the respiratory tract as either particles or droplets. It is therefore essential that these particles/droplets are deposited to the required site of pharmacological action, e.g. the bronchioles for asthma treatment or the alveoli for systemic absorption. To achieve the optimum pharmacological effect, the pharmaceutical scientist must fully understand and embrace the factors that affect the deposition of the drug within the respiratory tract as these will influence the formulation of respiratory dosage forms. These factors are as follows: (1) the size of the inspired particles/droplets; and (2) the effect of humidity on particle size.

The size of the inspired particles/droplets

The diameters of the various airways leading to the alveoli sequentially decrease and therefore the location of the deposited particle/droplet is dependent on particle size. The distribution of diameters of particles in an aerosol is non-normal and, as a result, these are normally plotted as log-normal distributions. Due to this heterogeneous distribution of particles, conventional parametric statistics may not be used to summarise the central tendency and variance of the distribution. The size of particles designed for pulmonary administration is defined using a parameter that accounts for this limitation and relates the diameter of the particles to the particular appplication, namely, the *aerodynamic diameter*. This is the diameter of a spherical particle of unit density (1 g/cm³) that possesses the same gravitational settling velocity as the particle under examination and, in the case of spherical particles, may be expressed as follows:

$$d_s = \sqrt{\frac{\rho}{\rho_o}} \times d$$

where: d_s is the aerodynamic diameter; ρ is the density of the particle under examination; ρ_o is the density of the spherical particle (i.e. 1 g/cm³); and d is the diameter of the particle under examination.

Typically the aerodynamic diameter is proportional to the velocity of settling of the particle within the respiratory tract.

In addition, the aerodynamic diameter may be expressed in terms of the *mass median aerodynamic diameter* (MMAD), i.e. the aerodynamic diameter that divides the distribution equally in two parts in terms of the weight of particles.

The variance (and hence the dispersity) of the log-normal distribution of particles in therapeutic aerosols may be expressed

in terms of the *geometric standard deviation* (s_g). The log-normal distribution will be normally distributed and the geometric standard deviation is therefore the standard deviation of the distribution following the log-normal transformation.

Following inhalation, the deposition of the particles within the respiratory tract is dependent on the size of the inhaled particles. Typically particles with a mass median aerodynamic diameter greater than 10 µm will be trapped in the trachea, whereas particles exhibiting MMADs of circa 5 and 2 µm will be deposited within the bronchioles and alveoli. Particles that are trapped in the higher regions of the respiratory tract will be removed from the lung by muciliary clearance (entrapment in the mucus and movement towards the mouth by the action of the cilia). Particles with an MMAD diameter of <1 µm are inhaled to the lowest sections of the lung but are then exhaled, thereby providing no pharmacological effect.

The effect of humidity on particle size

Following entry within the respiratory tract, the particle will be exposed to a highly humid environment (circa 99%) and this results in the deposition of a layer of moisture on the surface of the particle. The effect of this deposited layer on particle size is dependent on the hydrophilicity/lipophilicity of the particle:

- *Lipophilic inhaled particles.* The adsorbed layer of moisture is negligible and therefore does not affect the MMAD of the inhaled particle.
- *Hydrophilic inhaled particles.* The adsorbed layer of moisture results in the dissolution of the hydrophilic particle. Due to the differences in the vapour pressures of water (moisture) and the solution of therapeutic agent that has formed on the surface of the hydrophilic particle (the latter being lower), further moisture is adsorbed on to the surface of the particle until the vapour pressures of the two systems are similar. The continuing adsorption of moisture on to the surface of the particle results in further dissolution of drug with the attendant increase in effective particle size and possible deposition within the higher regions of the respiratory tract.

Mechanisms of particle deposition within the respiratory tract

The previous section has outlined the effect of particle size on the deposition of particles within the respiratory tract (and hence on the clinical efficacy of aerosol products). Particle size additionally affects the mechanisms by which the particles are deposited within the respiratory tract. There are four main mechanisms that operate in aerosol products: (1) inertial impaction;

(2) gravitational sedimentation; (3) Brownian diffusion; and (4) electrostatic precipitation.

Inertial impaction

Inertia may be defined as the property of a particle that enables it either to remain at rest or to remain in uniform motion until exposed to an external force. Following inhalation, aerosol particles must (frequently) change direction to ensure deposition at the required site within the respiratory tract. Initially the particles will move horizontally towards the back of the throat; however, the flow of air will change direction both into the trachea and then at the other main tributaries along the respiratory tract until the alveoli are reached. The inertial effect resists the change in direction of flow and, as a result, particles with sufficient momentum (the product of the velocity and the mass) will attempt to maintain their initial path of flow. This resistance results in impaction of the particle with the respiratory tract at sites prior to the bronchioles and alveoli.

The probability of impaction ($Pr_{impaction}$) may be mathematically defined as:

$$Pr_{impaction} = \frac{U_t U \sin\theta}{rg}$$

where: U_t refers to the terminal settling velocity (i.e. the velocity of motion whenever the force of the falling particle downwards is equal to the drag force acting in the opposite direction; this results is a net zero force and a constant particle velocity); U refers to the velocity of the air stream following inhalation; θ refers to the angle of the airflow change; g refers to gravity; and r is the radius of the airway.

Accordingly, the probability of the occurrence of inertial impaction increases as the angle and velocity of airflow increase, whereas the probability decreases in airways of larger radius. Inertial impaction is relevant for particles of larger MMAD (>5 μm) within the larger airways, e.g. nose, mouth, pharynx and larynx. As the airflow velocity decreases within the lower sections of the respiratory tract (due to the effects of branching of the airways providing resistance to the flow of air), the contribution of inertial impaction on the deposition of particles within the deeper regions of the respiratory tract is lower.

Gravitational sedimentation

As the term implies, gravitational sedimentation refers to the downward movement of particles under the action of gravity. Gravitational sedimentation is an important mechanism for the deposition of particles of small MMAD (typically 1.0–5.0 μm)

within the bronchioles and alveoli. Furthermore, the probability of impaction by gravitational sedimentation within these sites may be enhanced by either a steady rate of breathing or by holding the breath. This is particularly relevant for the deposition of particles with an MMAD of circa 1.0 µm.

Brownian diffusion

Brownian motion refers to the random movement of particles within a fluid (liquid or air), which, within the respiratory tract, enables small particles (<0.5 µm) to move towards and be deposited on the walls of the various sections of the respiratory tract. This is not a significant contributor for the deposition of particles from therapeutic aerosols.

Electrostatic precipitation

This is a phenomenon in which the charge on the surface of a particle may affect the resultant deposition, i.e. a charged particle interacts with a site within the respiratory tract that possesses an opposite charge. Typically, this mechanism is not important for particles of MMAD that are greater than circa 4 µm. Furthermore, the charge on particles may induce an interaction with the plastic surfaces of containers/spacers/inhalers and may therefore compromise the effective delivery of the particles to the required site within the respiratory tract.

Tip

The region of deposition of the inhaled particles/droplets within the respiratory tract directly affects the clinical performance of the dosage form. Therefore, it is important that the formulation scientist designs the dosage form to ensure that, following inhalation, the particle/droplet size is of the correct MMAD to reach the target site within the respiratory tract.

Formulation of respiratory dosage forms

There are three main types of delivery system for respiratory dosage forms: (1) metered-dose inhalers (MDIs); (2) dry-powder inhalers; and (3) nebulisers. The formulation and design of these systems are described in this section.

Metered-dose inhalers

MDIs are a commonly used system – in use for over 50 years – for the delivery of therapeutic agents to the respiratory tract. The advantages and disadvantages associated with the use of MDIs are detailed below.

Advantages

- portable (which is particularly useful for the treatment of acute respiratory conditions, e.g. breathlessness in asthma patients)
- low-cost dosage form

■ hermetically sealed and therefore oxidation of the therapeutic agent and microbial contamination are minimised
■ effective treatment of respiratory disorders.

Disadvantages
■ MDI most frequently contain *dispersed* drug and therefore problems may arise concerning the physical stability of the formulation.
■ Frequently there is ineffective use of drug. Typically the majority of drug that is emitted from the aerosol fails to reach the proposed site of pharmacological action. This point is further addressed later in this chapter.
■ Clinical efficacy is often dependent on the ability of the patient to use the MDI correctly: efficacy is lowered by the inability of the patient to synchronise inhalation and actuation of the dosage form.

The main components of MDIs are the container, the metered valve and the formulation (within the container), as illustrated in Figure 8.2.

Figure 8.2 Illustrated cross-section through a metered-dose inhaler showing the location of the formulation and the design of the container. (Adapted from www.solvay-fluor.com.)

- Aerosol container
- Active ingredient suspension or solution with propellant
- Seal
- Valve
- Mouthpiece
- Spray mist
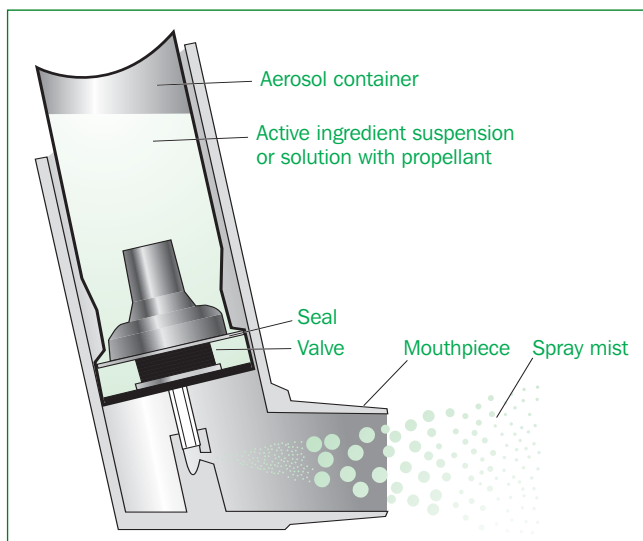
Formulation and operation of an MDI
The formulation component of MDIs comprises two parts: the propellant and the therapeutic agent (which may be present as a solution or, most commonly, as a suspension). Following actuation, a mixture of the propellant and the therapeutic agent is released into the oral cavity of the patient in the form of droplets (circa 40 µm MMAD). The volume of product (typically 25–100 µm) dispensed from the MDI during the actuation process is controlled by a *metering valve*. In this, the depression of the

stem of the canister (containing the formulation) facilitates the release of a volume of product into the mouthpiece of the device. Release of the downward pressure on the stem of the canister allows the free volume within the metering valve to be replenished. Removal of liquid propellant following actuation results in a momentary change in the vapour pressure within the canister. To correct this, liquid propellant evaporates to occupy the free volume within the canister, thereby maintaining a constant vapour pressure and hence operating pressure.

When formulated correctly the propellant will rapidly evaporate following exposure to atmospheric conditions and the droplets are reduced to solid particles of therapeutic agent, which may then be deposited at the required site in the respiratory tract. The reader will be aware of the particle size requirements for penetration into the lower regions of the respiratory tract and therefore to ensure that the therapeutic agent reaches the required site. There are three prerequisites for this process to occur successfully:

1. The particle size of the therapeutic agent that has been incorporated into the formulation must exhibit the required diameter and polydispersity. Reduction in the particle size to achieve the required diameter is most commonly performed using milling techniques. It is more difficult to achieve the required polydispersity.

2. Following shaking of the MDI (prior to use), the drug particles must be readily resuspended. Aggregation of drug particles will increase their effective size and will lead to deposition at the incorrect region of the respiratory tract and, if the size of the aggregate is sufficiently large, clogging of the nozzle of the MDI.

3. As highlighted previously, the formulation exits the MDI as a droplet composed of the dispersed therapeutic agent with the liquid propellant. The purpose of the propellant is therefore to provide sufficient pressure to deliver the dose of drug from the MDI to the upper respiratory tract. However, it is essential that evaporation of the propellant occurs rapidly to ensure that the solid drug particle is free to travel to the lower regions of the lung.

Based on these three requirements, the formulation of MDI involves consideration of the properties of both the propellant and the therapeutic agent.

■ *Propellants used in MDI.* The purpose of the propellant is twofold: to provide the driving pressure to force the therapeutic agent from the MDI to the upper respiratory tract and, secondly, to exhibit the required evaporation rate to facilitate particle delivery to the required site within the

respiratory tract. Both the pressure within the metal canister of the MDI and the subsequent rate of evaporation of the propellant are functions of the partial vapour pressure of the propellants within the canister. To achieve these required properties mixtures of propellants are used. Currently there are two main classifications of propellants used, as follows: (1) chlorofluorocarbons (CFCs) and (2) hydrofluorocarbons.

– *CFCs.* CFCs have been used as propellants for many years; however, due to concerns with their contribution to the depletion of the ozone layer, their use has been prohibited in all aerosols, with the exception of MDIs (under the proviso that no suitable alternatives are available).
The main examples of these that have been or are being used as propellants in MDI, including their physicochemical properties, are shown in Table 8.1 and Figure 8.3.

Figure 8.3 Structural formulae of: (a) dichlorodifluoromethane (propellant 12); (b) dichlorotetrafluoroethane (propellant 114); and (c) trichloromonofluoromethane (propellant 11).
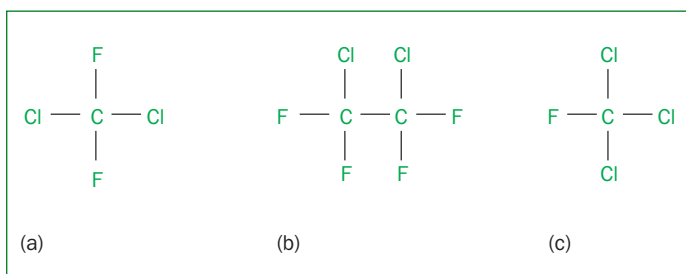
Table 8.1 The physicochemical properties of propellants 11 (trichloromonofluoromethane), 12 (dichlorodifluoromethane) and 114 (dichlorotetrafluoroethane)

Propellant	Boiling point	Vapour pressure at 20°C	Freezing point
12	−29.8°C	568 kPa	−158°C
114	3.6°C	183 kPa	−94°C
11	23.7°C	89 kPa	−111°C

Propellant 12 may be used as the sole propellant in MDI formulations; however, propellants 11 and 114 are generally used in combinations with other propellants. For example, blends of propellants 11 and 12 or propellants 11, 114 and 12 are used in MDI to provide the correct vapour range of vapour pressures (103–484 kPa) at room temperature and hence provide the required expulsion pressure, droplet size distribution and evaporation rate to ensure optimal clinical performance. Furthermore, mixtures of propellants may be required to ensure the surfactants that are used to stabilise the dispersed drug particles are soluble in the propellant. This is

an important consideration in the formulation of a stable MDI product.

Calculation of the vapour pressure of propellant mixtures is performed using *Raoult's law*, which states that the vapour pressure of a mixture of propellants (p_{total}) is simply the sum of their partial vapour pressures (e.g. p_1 and p_2 for propellants 1 and 2, respectively).

$$P_{total} = p_1 + p_2$$

The partial vapour pressure of each component is the product of the mole fraction of that component (χ) and the partial vapour pressure of the propellant (p^0_1):

$$p_1 = \chi_1 \times p^0_1$$

This equation can be used to calculate the total vapour pressure of a mixture of propellant 11 and propellant 12 (60:40 ratio), as follows:

● *Determination of the mole fraction of each propellant in the mixture*

Propellant 11 (molecular weight 137.50 g/mol)

$$n_{11} = \frac{60.00}{137.50} = 0.44$$

Propellant 12 (molecular weight 121.00 g/mol)

$$n_{12} = \frac{40.00}{121.00} = 0.33$$

Total number of moles of propellant

$$n_{total} = n_{11} + n_{12} = 0.44 + 0.33 = 0.77$$

Mole fraction of propellant 11

$$\chi_{11} = \frac{n_{11}}{(n_{11} + n_{12})} = \frac{0.44}{(0.44 + 0.33)} = 0.57$$

Mole fraction of propellant 12

$$\chi_{12} = \frac{n_{12}}{(n_{11} + n_{12})} = \frac{0.33}{(0.44 + 0.33)} = 0.43$$

- *Determination of the partial vapour pressures of each component in the mixture*
  Partial vapour pressure of propellant 11

  $$P_{11} = \chi_{11} \times p^0_{11} = 0.57 \times 89.00 = 50.86 \text{ kPa}$$

  Partial vapour pressure of propellant 12

  $$P_{12} = \chi_{12} \times p^0_{12} = 0.43 \times 568.00 = 243.43 \text{ kPa}$$

- *Calculation of the total vapour pressure within the canister*

  $$P_{total} = P_{11} + P_{12} = 50.86 + 243.43 = 294.29 \text{ kPa}$$

As the reader will observe, the partial pressure of the propellant mixture may be readily manipulated by altering the ratio of the individual propellants. The partial vapour pressure of the mixture is an important consideration in the formulation of aerosols. The vapour pressure is responsible for the expulsion of the defined volume of product from the aerosol. If the pressure is too high, impaction of the expelled droplet/particle on to the surfaces of the upper respiratory tract will occur, thereby diminishing the clinical performance of the MDI. Reduction of the partial vapour pressure of the system may be achieved by increasing the mole fraction of a less volatile propellant (e.g. propellant 11) into the formulation. However, as the mole fraction of a less volatile propellant is increased, the rate of evaporation of the propellant following actuation decreases and, hence, this may reduce the percentage of the actuated dose of the therapeutic agent that reaches the lower airways.

- *Hydrofluorocarbons (HFCs).* HFCs were developed to address the deleterious effects of CFC propellants on the ozone layer. Unfortunately, although the effects on the environment are lower than for CFCs, these propellants are also damaging to the ozone layer. In spite of this HFCs represent a significant evolution in the development of propellants that are ozone-friendly. Two examples of these that are employed in MDIs are heptafluoropropane and tetrafluoroethane (Figure 8.4).
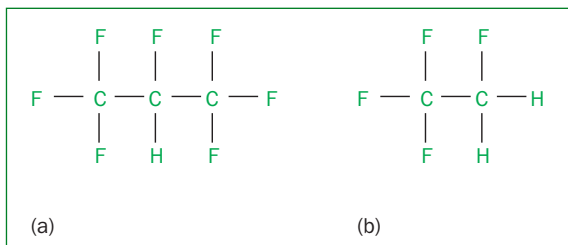
Figure 8.4 Structural formulae of selected hydrofluorocarbons used as propellants in metered-dose inhalers: (a) heptafluoropropane; (b) tetrafluoroethane.

Both heptafluoropropane and tetrafluoroethane may be employed as propellants for MDIs. One potential problem with their use is their extremely hydrophobic properties. As a result, the solubility of the commonly used surfactants (sorbitan trioleate, oleic acid, sorbitan sesquioleate) may not be sufficiently high to stabilise the formulation.

The physicochemical properties of the HFC propellants are similar to CFC, reflecting their applicability as propellants for MDIs. These are summarised in Table 8.2. As before, modification of the vapour pressure of the propellant system may be achieved by blending the HFC propellants (according to Raoult's law).

Table 8.2 The physicochemical properties of selected hydrofluorocarbon propellants

Propellant	Boiling point	Vapour pressure at 20°C	Freezing point
Tetrafluororethane	−26.2°C	569 kPa	−108°C
Heptafluoropropane	−16.5°C	398 kPa	−131°C

There are other formulation considerations for MDI. These are summarised below.

The physicochemical properties of the therapeutic agent
There are two key physicochemical properties that affect the stability and clinical performance of MDI – the solubility of the therapeutic agent in the propellant system and the MMAD (and polydispersity) of the dispersed particles. If the drug shows limited solubility in the propellant that is susceptible to changes in storage conditions, this may result in fluctuations in the solubility of the drug in the propellant that may, in turn, result in recrystallisation of the drug. This will result in the generation of larger particles. It is additionally important that the MMAD (and polydispersity) of the therapeutic agent that is to be incorporated into the propellant system is within the inspirable size range (remembering that the MMAD of particles that will be deposited in the lower airways is circa 1–5 µm). The drug particle size should be manipulated to the required size fraction by milling techniques.

Maintenance of the physical stability of the MDI

Most commonly the therapeutic agent is present in the MDI as dispersed particles within the liquid propellant and therefore there are similar concerns regarding the physical stability of the formulation. Irreversible aggregation of the particles (caking) will result in problems regarding both the successful deposition of drug within the respiratory tract and the successful discharge of the particles through the orifice of the MDI. To obviate these problems surface-active agents of low hydrophile–lipophile balance are incorporated into the liquid propellant which will adsorb to the dispersed particles and, in so doing, will stabilise the drug suspension (see Chapter 3 for a full description of the mechanism of stabilisation). Examples of surface-active agents that are used for this purpose (generally in concentrations ranging from 0.1 to 3.0% w/w) include:

- sorbitan trioleate
- sorbitan sesquioleate
- oleic acid.

Unlike suspensions designed for oral application, enhancement of suspension stability by modification of the viscosity of the liquid propellant phase is not performed for MDI.

Enhancement of drug solubility in the propellant

In some aerosol products, e.g. those for topical administration, the pharmaceutical scientist may wish to dissolve the therapeutic agent in the propellant mixture. For this purpose ethanol is frequently added as a co-solvent to facilitate this demand. This approach is infrequently used for MDI as the presence of ethanol will result in the generation of droplets after actuation that requires a longer period to evaporate (due to the vapour pressure of ethanol being substantially lower than that of the propellant mixture). This may affect the clinical performance of the MDI.

Spacer devices for MDIs

Previously in this section, the importance of the evaporation rate of the propellant after actuation to the subsequent deposition of the therapeutic agent within the respiratory tract was detailed. Furthermore, the reliance of conventional MDI on the ability of the patient to coordinate actuation and inhalation was discussed. One method by which these two problems may be minimised is the use of spacer devices (Figure 8.5).

Tip

Formulations that are delivered to the respiratory tract using metered-dose inhalers are generally composed of the drug (dispersed or dissolved), a surfactants(s), the propellant(s) and, occasionally, a co-solvent (to enhance drug solubility within the propellant).
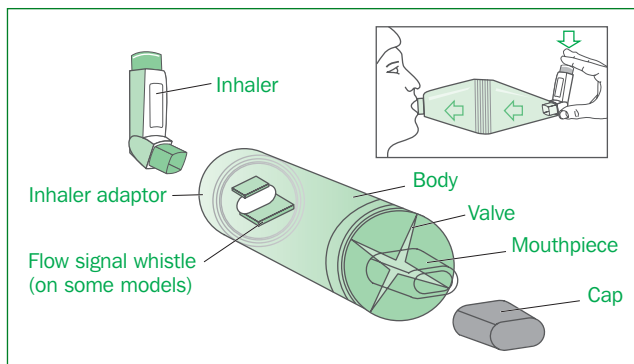
Figure 8.5 Diagrammatic representation of a spacer device, showing the clinical use of the spacer with the metered-dose inhaler. (Adapted from http://www.asthma.ca/images/adults/treatment/spacer.gif.)

Following actuation the propellant droplets (containing suspended particles of therapeutic agent) will reside within the spacer device, thereby facilitating evaporation of the propellant and allowing the patient to inspire the dose of drug without the need for coordination between actuation of the MDI and inhalation. One disadvantage of spacer systems is their size (rendering them conspicuous in use and relatively non-portable).

Manufacture of MDIs

There are two methods by which MDIs are manufactured: (1) pressure filling; and (2) cold filling.

Pressure filling

In this method the therapeutic agent (and excipients) is mixed with a portion of a propellant which possesses a relatively high boiling point and low vapour pressure, e.g. trichloromonofluoromethane (23.7°C and 89 kPa, respectively) at or below 20°C. This drug/propellant slurry is then filled into the canister and the canister is then crimped with the attached valve. The second more volatile propellant is then filled into the canister through the value under pressure until the correct propellant blend is attained. Although this technique is straightforward, it is dependent on the use of a less volatile propellant whose boiling point is greater than that of the temperature in the mixing area.

Cold filling

In cold filling the drug (and excipients) and propellants are mixed under cold conditions (circa −30°C) and filled into the canister (which is subsequently crimped with the attached valve). Propellant is then added into the canister through the valve at this low temperature until the correct mass has been added.

Dry-powder inhalers

DPIs are respiratory dosage forms in which a powder (containing the therapeutic agent) is inhaled into the respiratory tract. The flow of the powder is activated whenever the patient inhales and therefore no propellant is required. It has been reported that MDIs are more efficient respiratory delivery systems than their dry-powder comparators.

There are a number of designs of these systems:

- *Inhalers in which the drug (and excipients) are present within a hard gelatin capsule.* In these the hard gelatin capsule (containing the therapeutic agent) is located inside the device. Prior to use the gelatin capsule is pierced in two locations to enable airflow through the capsule. As the patient inhales, a rotor is activated, resulting in turbulent airflow (>35 l/min), which carries the powder to the patient.
- *Inhalers in which the drug (and excipients) are present within a blister pack.* In these systems, a multidose unit comprising four or eight individual doses is located within a circular device containing a mouthpiece (Figure 8.6). After the device has been assembled the lid is elevated and the individual disc containing the therapeutic agent (and excipients) is pierced. Inhalation by the patient results in the passage of air through the device, which delivers the bioactive powder to the patient.

Figure 8.6 Diagrammatic representation of a diskhaler device, illustrating the individual components and assembly. (Adapted from http://www.asthma.ca/adults/treatment/diskhaler.php.)
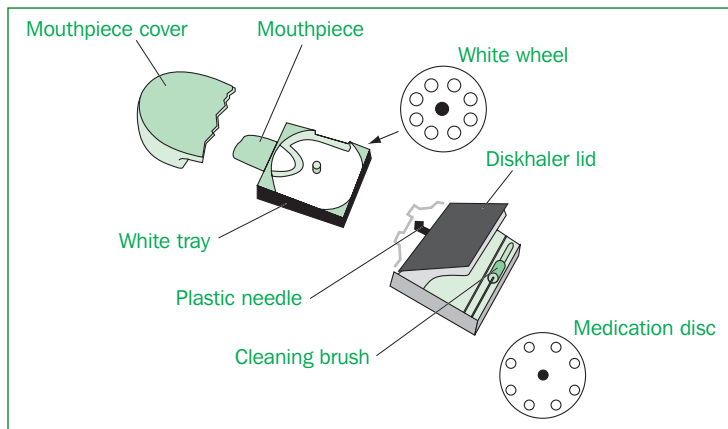
- *Inhalers in which the drug (and excipients) are present within the inhaler.* In these systems the inhaler is preloaded with multiple doses of drug. As before, the packaging containing each dose is broken and the therapeutic agent (and excipients) are carried to the patients during inhalation. These systems may be replenished with drug by incorporating a replacement reservoir. In an alternative strategy the drug (*without*

excipient) is located within a reservoir (not as individual units). The device is activated (e.g. by twisting the base), resulting in the passage of drug into a chamber. As before, when the patient inhales, air is drawn through the device and the powdered drug will be delivered to the patient.

Formulation considerations for DPIs

The formulation of DPIs is relatively straightforward, particularly when compared to MDIs. The major formulation concerns for DPIs are as follows:

> **Tip**
>
> DPIs do not use a liquid propellant. The powder enters the respiratory tract as the patient inhales air through the device.

■ *Therapeutic agent.* As before, the particle size (MMAD) of the therapeutic agent should be less than 5 μm.

■ *Excipients.* Excipients are used in the majority of DPIs for two main reasons:

– *To facilitate production.* As the dose of therapeutic agent in the DPI is generally low, the addition of an inert excipient may be used to enhance the quality of the filling process.

– *To improve the flow properties of the therapeutic agent during inhalation.* To ensure optimum clinical efficacy, it is essential that following inhalation the flow properties of the therapeutic agent are sufficient to enable the dose of drug to be efficiently removed from the device. If the drug particles are aggregated or if there is an interaction between the drug and the plastic interior of the device, this will result in both a variable and inaccurate dose of drug being delivered to the patient. Therefore, to address this problem the therapeutic agent is frequently mixed with an inert excipient, most commonly lactose (MMAD 30–60 μm). The larger particle size of the excipient ensures that it does not enter into the lower airways. Within the device the drug particles are adsorbed on to the larger lactose particles and, in so doing, their flow properties are improved. It is essential, however, that dissociation of the drug/carrier interaction occurs prior to impaction of the lactose carrier within the oral cavity/pharynx.

The interaction between the therapeutic agent and the carrier is dependent on several factors, including:

● the surface areas of the particles
● the surface energies of the particles
● the morphologies of the particles.

These factors are dependent (to a large degree) on the techniques used to process the powders, in particular, the milling conditions.

Solutions for nebulisation

Nebulisation involves the application of energy (either a high-velocity gas or by the use of ultrasonic systems) to a solution of a therapeutic agent and results in the formation of droplets of solution, which are then inspired by the patient through a facemask. The energy source is provided by a nebuliser (most commonly a jet or ultrasonic nebuliser). The use of nebulisers is generally reserved for the treatment of acute conditions (e.g. acute asthma, respiratory infection) or in those patients who have difficulties using other respiratory dosage forms.

Formulation of solutions for nebulisation

The general formulation considerations for solutions for nebulisation are similar to those described in Chapter 1 and therefore in this section only the particular formulation requirements for solutions designed for administration to the respiratory tract will be discussed.

Vehicle

The vehicle for solutions designed for nebulisation is water for injections (nebuliser solutions are sterile). Due to the possibility of acidic solutions (pH <5) causing bronchoconstriction, the pH of nebuliser solutions is greater than 5. This may be modified via the use of buffers (e.g. citrate, phosphate).

Co-solvents

Co-solvents may be used in nebuliser solutions to increase the solubility of the drug. However, as in other applications, care should be taken when selecting the concentration of these to ensure that there is no toxicity to the respiratory epithelia. Examples of co-solvents that may be used include:
- propylene glycol
- glycerol
- ethanol.

Osmolality-modifying agents

The use of hypo- and hyperosmotic solutions within the respiratory tract has been linked to bronchoconstriction and therefore solutions designed for nebulisation should be formulated to be iso-osmotic. As described previously (Chapter 5), modification of the osmolality of nebuliser solutions may be performed by adding the appropriate ionic concentration (e.g. sodium chloride, potassium chloride, mannitol).

Miscellaneous agents

Other components that may be present in nebuliser solutions include:

- antioxidants (water-soluble examples; however, it should be noted that sulphites may cause brochospasm)
- preservatives: some multidose nebuliser solutions contain preservatives; however, the vast majority of formulations do not contain these excipients. Generally nebuliser solutions are packaged as single-dose vials/ampoules and, as such, do not require the addition of preservatives.

Multiple choice questions

1. **Concerning respiratory drug delivery, which of the following statements are true?**
   a. Delivery of therapeutic agents to the respiratory tract is performed primarily for the treatment of local conditions, e.g. asthma, infection.
   b. The treatment of asthma involves the deposition of the therapeutic agent at the alveoli.
   c. Systemic absorption of certain therapeutic agents occurs following respiratory delivery.
   d. Respiratory drug delivery is associated with a slow onset of drug action.

2. **Regarding the physiology of the respiratory tract, which of the following statements are true?**
   a. The diameters of the conducting airways decrease towards the alveoli.
   b. Drug absorption from the respiratory tract occurs at the alveoli.
   c. The surface area of the alveoli is >100 m^2.
   d. The primary mechanism of elimination of particles >5 μm in diameter is expiration.

3. **Which of the following factors influence the deposition of particles within the respiratory tract?**
   a. The size of the inspired droplets: increasing particle/droplet size results in greater deposition within the lower airways.
   b. Humidity.
   c. The vapour pressure of the propellant.
   d. The patient's inhaler technique.

4. **Concerning the formulation and use of metered-dose inhalers, which of the following statements are true?**
   a. Oxidation of drugs is minimised.

b. Metered-dose inhalers are usually formulated to ensure that the drug is soluble within the propellant system.

c. Metered-dose inhalers require the inclusion of a preservative.

d. Metered-dose inhalers may be easily formulated to contain water.

5. **Concerning propellants for metered-dose inhalers, which of the following statements are true?**

a. The vapour pressure remains constant throughout the lifetime of use of the inhaler.

b. The use of chlorofluorocarbons has increased in recent years.

c. The volume of propellant released upon actuation is controlled by a metering valve.

d. All metered-dose inhalers are filled under atmospheric conditions.

6. **The following excipients are included in metered-dose inhalers for the following reasons:**

a. Ethanol – to increase the vapour pressure of the propellant.

b. Tween 80 – to enhance the solubility of the therapeutic agent within the propellant.

c. Oleic acid – to stabilise the suspended drug in the propellant.

d. Antioxidants – to inhibit oxidative degradation of the therapeutic agent.

7. **Concerning the formulation of dry-powder inhalers, which of the following statements are true?**

a. Liquid propellants are not required.

b. The Mass Median Aerodynamic Diameter of the therapeutic agent should be less than 5 mm.

c. Lactose is commonly used to improve the flow properties of the powdered drug.

d. Following inspiration, both lactose and the powdered drug reach the site of action.

8. **The advantages of metered-dose inhalers include:**

a. Administration of high doses of therapeutic agent.

b. Convenience for the patient.

c. Requirement for patient's ability to coordinate actuation and inhalation.

d. Greater efficiency than nebulisers.

9. **Concerning nebulisers, which of the following statements are true?**

a. Nebulisers require either a compressed-gas source or an ultrasonic device.

b. Nebulsiers are portable and convenient to use.

c. They require patient coordination.
d. They are suitable for the delivery of all drugs.

10. Regarding the formulation of solutions for nebulisation, which of the following statements are true?
a. The pH of nebuliser solutions is always greater than 5.
b. Propylene glycol may be employed as a co-solvent in nebuliser solutions.
c. Nebuliser solutions should be hypertonic.
d. In general, preservatives are not required in nebuliser solutions.

chapter 9
Solid-dosage forms 1: tablets

Overview

In this chapter the following points will be discussed/described:

- an overview/description of the various types of solid-dosage forms and the rationale for their use
- formulation strategies for solid-dosage forms
- an overview/description of the methods of manufacture of solid-dosage forms, including post-manufacture processes (e.g. coating)
- the advantages and disadvantages of the different methods that are used to manufacture solid-dosage forms
- a detailed description of the excipients used in the formulation and manufacture of solid-dosage forms.

Introduction

Solid-dosage forms broadly encompass two types of formulation, namely tablets and capsules (capsules are described in Chapter 10). It has been estimated that solid-dosage forms constitute circa 90% of all dosage forms used to provide systemic administration of therapeutic agents. This highlights the importance of these dosage forms in the treatment and management of disease states. The widespread use of tablets has been achieved as a result of their convenience and also the diversity of tablet types. Examples of tablet types available include: (1) conventional compressed tablets; (2) multiple compressed tablets; (3) enteric-coated tablets; (4) sugar-coated tablets; (5) film-coated tablets; (6) chewable tablets; (7) effervescent tablets; (8) buccal and sublingual tablets; and (9) vaginal tablets.

KeyPoints

- Solid-dosage forms encompasses the largest category of dosage forms that are clinically used.
- There are several types of tablet solid-dosage form that are designed to optimise the absorption rate of the drug, increase the ease of administration by the patient, control the rate and site of drug absorption and mask the taste of a therapeutic agent.
- The formulation of tablets involves the use of several components, each of which is present to facilitate the manufacture or to control the biological performance of the dosage form.
- There are four main methods by which tablets may be manufactured: (1) wet granulation; (2) dry granulation (slugging); (3) direct compression; and (4) roller compaction. There are advantages and disadvantages associated with the use of these methods.

Conventional compressed tablets

These tablets are designed to provide rapid disintegration and hence rapid drug release, and represent a significant proportion of tablets that are clinically used. The manufacture of these tablets involves the compression of granules or powders (both containing drug) into the required geometry. Following ingestion, the tablets will disintegrate within the gastrointestinal tract (stomach), allowing the drug to dissolve in the gastric fluid and, ultimately, be absorbed systemically.

Multiple compressed tablets

These are tablets that are composed of at least two layers. Typically there are two designs of multiple compressed tablets: (1) multiple-layered; and (2) compression coated. In the former design the first layer is formed by a relatively light compression of the drug containing powder mix/granules. The next layer is then formed by compression of the powder/granule mix (containing drug) on top of the lightly compressed first layer. Additional layers are formed in a similar fashion. In the second approach the initial layer is prepared by light compression (as described above), removed and located in a second tablet press. The granules/powders of the second coat are fed into the press and allowed to form a constant mass around the surface (and edges) of the pressed tablet prior to compression to form the finished product. It is, of course, possible to prepare tablets containing more than two layers although, in so doing, the complexity of the manufacturing process is dramatically increased.

There are several applications of the use of multiple compressed tablets, including:

- the separation of drugs into separate layers that may be incompatible when formulated as a (single-layer) conventional tablet
- the delivery of therapeutic agents at different rates or to different sites within the gastrointestinal tract from a single tablet. The dissolution of the layers of the tablet or, indeed, the dissolution/diffusion of the drug from the layer may be controlled by the inclusion of polymeric excipients
- the production of tablets that are coated. This is important in cases where the drug has a bitter taste or where the drug is irritant to the stomach (e.g. non-steroidal anti-inflammatory drugs) or is chemically unstable under acidic conditions.

Enteric-coated tablets

These are tablets that are coated with a polymer that does not dissolve under acidic conditions (i.e. the stomach) but does dissolve under the more alkaline conditions of the small intestine

(i.e. pH > 4). Enteric polymers are primarily employed as coatings of conventional tablet dosage forms and, by inhibiting the dissolution of the therapeutic agent within the stomach, offer protection against possible drug degradation (e.g. erythromycin) or irritation of the gastric mucosa (e.g. non-steroidal anti-inflammatory drugs). Following dissolution of the coating, the tablet will disintegrate and the drug will dissolve in the gastric fluids (thereby facilitating absorption).

Examples of polymers that are used for this purpose include: (1) cellulose actetate phthalate/cellulose acetate butyrate; (2) hydroxypropylmethyl cellulose succinate; and (3) methacrylic acid co-polymers (Eudragit).

Cellulose acetate phthalate/cellulose acetate butyrate
These are cellulose derivatives in which a proportion of the hydroxyl groups have been esterified with either phthalic acid or butyric acid. Dissolution of the polymer occurs in solutions where the pH > 6.

Hydroxypropylmethyl cellulose succinate
This is a cellulose derivative in which a proportion of the hydroxyl groups on hydroxypropylmethyl cellulose have been esterified with either acetic acid or succinic acid. Dissolution of this polymer occurs within the intestine, as it is insoluble in the stomach.

Methacrylic acid co-polymers (Eudragit®)
These are a range of co-polymers based on methacrylic acid in which the pendant carboxylic acid groups have been esterified to form ethyl methacrylate, dimethylaminomethacrylate, butylmethacrylate and methyl methacrylate. The presence of the wide range of functional groups enables the production of co-polymers that exhibit a range of solubilities. One example of the Eudragit family that is employed as an enteric coating is poly(methacrylic acid-co-methylmethacrylate) (Figure 9.1).
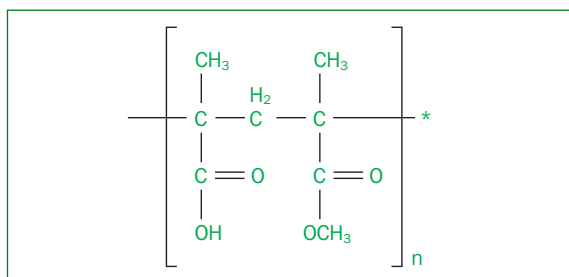
Figure 9.1 Structural formula of poly(methacrylic acid-co-methylmethacrylate).

Two commercial co-polymers are available which resist dissolution in the stomach but dissolve at higher pH, namely Eudragit L100 (poly(methacrylic acid-co-methylmethacrylate, 1:1 molar ratio of monomers), which is soluble in the intestinal fluid from pH 5.5, and Eudragit S100 (poly(methacrylic acid-co-methylmethacrylate, 1:2 molar ratio of monomers), which is soluble in the intestinal fluid from pH 7.

Sugar-coated tablets

These are conventional tablets that have been coated with a concentrated sugar solution to improve the appearance of the formulation and/or to mask the bitter taste of the therapeutic agent. The use of sugar coatings has dramatically decreased due to the advent of film-coated tablets (as a result of the improved mechanical properties of the latter coating).

Film-coated tablets

These are conventional tablets that have been coated with a polymer or a mixture of polymers (and, when required, a plasticiser to render the coating flexible). Film coatings show improved mechanical properties when compared to sugar coatings and, furthermore, film coatings may be deposited over embossed markings on the tablet surface. Film coatings are generally less elegant than sugar coatings. Examples of polymers that are used to film-coat tablets (and which dissolve in the stomach to enable tablet disintegration and drug dissolution) include:

- hydroxypropylmethylcelluose
- hydroxypropylcellulose
- Eudragit E100 (a co-polymer of butylmethacrylate, 2-dimethyl-aminoethylmethacrylate and methylmethacrylate, 1:2:1)

In addition to improving the appearance of conventional tablets, film coatings are employed to control the rate and duration of drug release or to target drug release to certain regions of the gastrointestinal tract, e.g. the colon. If the film coating is insoluble, the tablet will retain its shape during transit along the gastrointestinal tract. Drug release occurs by diffusion through the insoluble coating and subsequent partitioning into the gastrointestinal fluids. Examples of polymers that may be used for this purpose include: (1) ethylcellulose; and (2) Eudragit RS and RL.

Ethylcellulose

Ethylcellulose is an ethyl ether of cellulose in which there is complete substitution of the hydroxyl groups with ethyloxy groups (Figure 9.2). It is insoluble in aqueous solutions at all pH values. When used as a coating, a plasticiser (e.g. tributyl citrate,

triethyl citrate) is required to lower the glass transition temperature of the polymer and hence to enhance the flexibility of the coating.
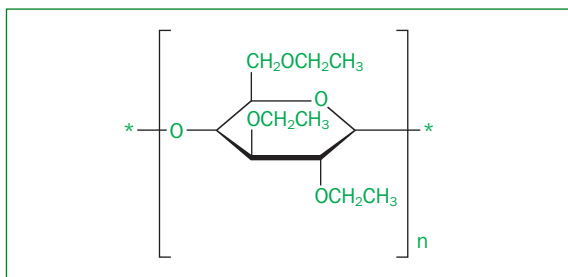
Figure 9.2 Structural formula of ethylcellulose.

Eudragit RS and RL

Eudragit RS and RL are methacrylate co-polymers that are insoluble, with the RL grade being more permeable than the RS grade. Chemically the RL grade is composed of poly(ethyl acrylate-co-methyl methacrylate-co-triethylammonioethyl methacrylate chloride), the ratios of the three monomers being 1:2:0.2, respectively. Similarly, the RS grade is a co-polymer of the above three monomers, with a different ratio of monomers (1:2:0.1, respectively).

The use of film coatings to target drug release within the gastrointestinal tract requires the use of polymers that dissolve within certain pH ranges. For example, enteric coatings offer drug targeting to regions of the gastrointestinal tract in which the pH is greater than 5.5 (e.g. Eudragit L-100). Targeting drug release to the colon involves the use of polymer coatings that dissolve at higher pH values (> 7), e.g. Eudragit S-100.

Chewable tablets

As indicated by the name, these tablets are chewed within the buccal cavity prior to swallowing. The main applications for this dosage form are:

- for administration to children and adults who have difficulty in swallowing conventional tablets
- antacid formulations in which the size of the tablet is normally large and the neutralisation efficacy of the tablet is related to particle size within the stomach.

Conversely, chewable tablets are not conventionally used if the drug has issues regarding taste acceptability.

Effervescent tablets

Effervescent tablets are added to aqueous solutions where they will rapidly disintegrate and produce either a drug suspension or an aqueous solution. The disintegration of the tablet is due to a

chemical interaction that occurs between two components: (1) an organic acid (e.g. citric acid); and (2) sodium bicarbonate in the presence of water. The evolution of carbon dioxide from this reaction results in tablet disintegration. The patient then consumes the solution/suspension. The main advantage of the use of effervescent tablets is the production of a dosage form from which the therapeutic agent is more rapidly absorbed than from alternative solid-dosage forms (e.g. conventional tablets). Conversely, the main disadvantages of this type of dosage form are the possible (un)availability of water and the need to package these tablets in moisture-impermeable packaging (typically aluminium foil), to inhibit the interaction between the acid and sodium bicarbonate due to the presence of environmental moisture.

Buccal and sublingual tablets

Buccal and sublingual tablets are dosage forms that are held within the oral cavity and slowly dissolve; the drug is absorbed across the buccal mucosa to produce a systemic effect. The type of tablet dictates the location within the oral cavity. Accordingly buccal tablets are positioned between the cheek and the gingiva whereas sublingual tablets are positioned underneath the tongue. These tablets are employed to achieve either rapid absorption into the systemic circulation (e.g. glyceryl trinitrate sublingual tablets) or, alternatively, to enable systemic drug absorption in situations where oral drug delivery is inappropriate, e.g. nausea. Drug absorption across the buccal mucosa avoids first-pass metabolism. Typically buccal and sublingual tablets should be formulated to dissolve slowly in vivo (and not disintegrate) and to be retained at the site of application and should not contain components that stimulate the production of saliva.

Tips

The different types of solid-dosage forms are designed to provide flexible platforms for the delivery of therapeutic agents to the gastrointestinal tract.

Coating tablets with polymers offers opportunities for the release of the therapeutic agent at particular regions within the gastrointestinal tract, e.g. the small intestine, colon.

Effervescent tablets are used to enhance the absorption of poorly soluble drugs.

The use of multilayered tablets has decreased in recent years.

Vaginal tablets

These are ovoid-shaped tablets that are inserted into the vagina (using a special inserter). Following insertion, retention and slow dissolution of the tablet occur, releasing the therapeutic agent to provide the local pharmacological effect (e.g. for the treatment of bacterial or fungal infection). Vaginal tablets may also be used to provide systemic absorption of therapeutic agents. In a similar fashion to buccal/sublingual tablets, it is important that dissolution, and not disintegration, of the tablet occurs in vivo, as disintegration will reduce tablet retention within the vagina.

Advantages and disadvantages of tablets as dosage forms

As stated in the introduction, tablets are the most popular dosage form used today and therefore there are several advantages associated with their use. However it is also important to highlight the disadvantages associated with their use.

Advantages
- Tablets are convenient to use and are an elegant dosage form.
- A wide range of tablet types is available, offering a range of drug release rates and durations of clinical effect. Tablets may be formulated to offer rapid drug release or controlled drug release, the latter reducing the number of daily doses required (and in so doing increasing patient compliance).
- Tablets may be formulated to release the therapeutic agent at a particular site within the gastrointestinal tract to reduce side-effects, promote absorption at that site and provide a local effect (e.g. ulcerative colitis). This may not be easily achieved by other dosage forms that are administered orally.
- Tablets may be formulated to contain more than one therapeutic agent (even if there is a physical or chemical incompatibility between each active agent). Moreover, the release of each therapeutic agent may be effectively controlled by the tablet formulation and design.
- With the exception of proteins, all classes of therapeutic agents may be administered orally in the form of tablets.
- It is easier to mask the taste of bitter drugs using tablets than for other dosage forms, e.g. liquids.
- Tablets are generally an inexpensive dosage form.
- Tablets may be easily manufactured to show product identification, e.g. exhibiting the required markings on the surface.
- The chemical, physical and microbiological stability of tablet dosage forms is superior to other dosage forms.

Disadvantages
- The manufacture of tablets requires a series of unit operations and therefore there is an increased level of product loss at each stage in the manufacturing process.
- The absorption of therapeutic agents from tablets is dependent on physiological factors, e.g. gastric emptying rate, and shows interpatient variation.
- The compression properties of certain therapeutic agents are poor and may present problems in their subsequent formulation and manufacture as tablets.
- The administration of tablets to certain groups, e.g. children

and the elderly, may be problematic due to difficulties in swallowing. These problems may be overcome by using effervescent tablet dosage forms.

Manufacture of tablets

There are four main methods by which tablets are manufactured: (1) wet granulation; (2) dry granulation; (3) direct compression; and (4) roller compaction (chilsonisation).

The choice of manufacturing process employed is dependent on several factors, including the compression properties of the therapeutic agent, the particle size of the therapeutic agent and excipients and the chemical stability of the therapeutic agent during the manufacturing process. Generically, the manufacture of tablets may be considered to consist of a series of steps:

- mixing of the therapeutic agents with the excipients
- granulation of the mixed powders (note: this is not performed in direct compression)
- mixing of the powders or granules with other excipients (most notably lubricants)
- compression into tablets.

The details of each of these steps will vary depending on the manufacturing method used.

Excipients used in the manufacture of tablets

The choice of excipients required for tablet manufacture is dependent on the process employed. Typically the following excipients are used in the manufacture of conventional tablets: (1) diluents/fillers; (2) binders; (3) disintegrants; (4) lubricants; (5) glidants; and (6) miscellaneous.

Diluents/fillers

Diluents are employed in the formulation of tablets (by all methods) to increase the mass of the tablets that contain a low concentration of therapeutic agent and thereby render the manufacturing process more reliable and reproducible. Diluents must exhibit good compression properties and be inexpensive. Examples of diluents for tablets are: (1) anhydrous lactose; (2) lactose monohydrate; (3) spray-dried lactose; (4) starch; (5) dibasic calcium phosphate; (6) microcrystalline cellulose (MCC); and (7) mannitol.

Anhydrous lactose

Anhydrous lactose is either pure anhydrous β lactose (*O*-β-D-galactopyranosyl-(1-4)-β-D-glucopyranose) or a mixture of anhydrous β lactose (70–80%) and 20–30% anhydrous α-lactose (*O*-β-D-galactopyranosyl-(1-4)-α-D-glucopyranose) (Figure 9.3).
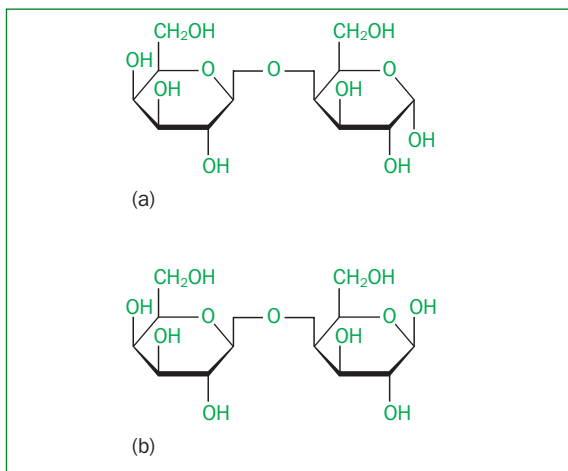
Figure 9.3 Structural formulae of: (a) anhydrous α-lactose and (b) anhydrous β-lactose.

Anhydrous lactose is available in a range of particle sizes and is commonly used as a diluent in wet granulation and dry granulation processes. It is a crystalline material.

Lactose monohydrate

Lactose monohydrate is composed of the monohydrate of α-lactose monohydrate and, whilst crystalline in nature, there may be varying proportions of the amorphous form. Lactose monohydrate is available in a wide range of grades that offer differing physical properties, e.g. particle size distribution/polydispersity and density (bulk and tapped).

Spray-dried lactose

Spray-dried lactose is a mixture of crystalline α-lactose monohydrate (80–90%) and 10–20% amorphous lactose. It is prepared by spray-drying a suspension of α-lactose monohydrate. The suspension is composed of circa 80–90% suspended α-lactose monohydrate, the remainder being in solution (which subsequently forms the amorphous portion of the spray-dried material). The specific use of spray-dried lactose for the manufacture of tablets by direct compression is described at a later stage in this chapter.

Starch

Starch is a polysaccharide composed of amylose and amylopectin, which is used as a diluent (and also as a binder and disintegrant) in tablet formulations. A pregelatinised grade is also available in which the granules of starch have been physically and chemically modified to produce a free-flowing powder. An example of a commercially available pregelatinised starch is Starch 1500LM (which has a low moisture content).

Dibasic calcium phosphate

This is a commonly used diluent in tablet formulations and is available as different hydrate forms and in a range of particle sizes. It is a basic excipient and may therefore chemically interact with acidic components in the presence of moisture. It has excellent flow and compression properties.

Microcrystalline cellulose (MCC)

MCC is a crystalline powder that is prepared by the controlled acid hydrolysis of cellulose. Several grades of MCC are commercially available that differ in their physicochemical properties, e.g. density, flow properties and particle size distribution. In addition to its use as a diluent, MCC may also be used as a binder in wet granulation (see below) and a disintegrating agent. Two commonly used grades of MCC are Avicel pH-101 (powder) and pH-102 (granules).

Mannitol

Mannitol is a polyol that is commonly employed as a diluent in tablets, particularly chewable tablets, due to the inherent sweetness and negative heat of solution. It exhibits excellent flow properties (Figure 9.4).

Figure 9.4 Structural formula of mannitol.
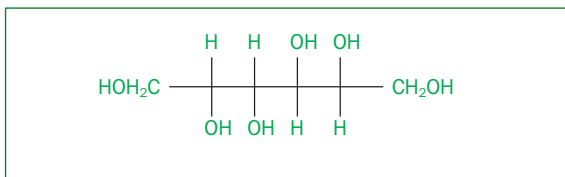
Binders

Binders are predominantly (but not exclusively) polymeric components that are employed in the production of tablets by the wet granulation method of manufacture. In this role, binders are either added as a solution or as a solid into the powder mix (following which the granulating fluid, typically water, is added). This process is described in the next section. The properties of commonly used binders are summarised in Table 9.1.

Disintegrants

Disintegrants are employed in tablet formulations to facilitate the breakdown of the tablet into granules upon entry into the stomach. If the formulated tablet is hydrophobic and/or it has been manufactured using a high compression force, the rate of water uptake into, and hence disintegration of, the tablet will be unacceptably low. In these situations disintegrants are an essential formulation component, enabling tablet disintegration to

Table 9.1 Examples and properties of binders commonly used in the manufacture of tablets by wet granulation

Binder	Concentration	General comments
Hydroxypropylmethylcellulose	2–5% w/w	The required concentration is dependent on molecular weight (grade) used
Polyvinylpyrrollidone	0.5–5% w/w	The required concentration is dependent on molecular weight (grade) used
Hydroxypropylcellulose	2–6% w/w	The required concentration is dependent on molecular weight (grade) used
Sucrose	50–67% w/w	Added as a syrup. Produces hard tablets whose properties are affected by moisture
Microcrystalline cellulose	20–90% w/w	Often used as a binder and diluent
Acacia	1–5% w/w	Produces hard tablets

occur within the specifications defined in the various pharmacopoeias (typically disintegration of conventional tablets must occur within 15 minutes). There are several mechanisms by which disintegrants elicit their effect:

■ Disintegrants may *increase* the *porosity* and *wettability* of the compressed tablet matrix. In so doing gastrointestinal fluids may readily penetrate the tablet matrix and thereby enable tablet breakdown to occur. In this approach it is necessary for the disintegrant to be homogeneously dispersed throughout the tablet matrix (requiring concentrations between 5 and 20% w/w). Examples of disintegrants that operate in this fashion include:

– starch
– MCC
– sodium starch glycolate (Figure 9.5).
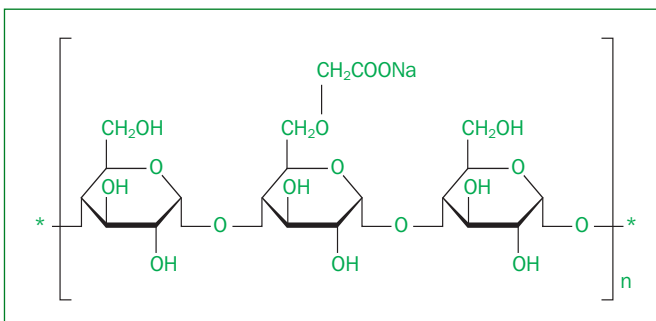
Figure 9.5 Structural formula of sodium starch glycolate.

Sodium starch glycolate is the sodium salt of the carboxymethyl ether of starch and is employed at circa 5% w/w as a disintegrant within tablet formulations.

■ Disintegrants may operate by *swelling* in the presence of aqueous fluids, thereby expediting tablet disintegration due to the increase in the internal pressure within the tablet matrix. Disintegrants in this category are typically hydrophilic polymers. To operate effectively it is important that the incorporated disintegrant does not cause a significant increase in viscosity, thereby decreasing the diffusion of gastrointestinal fluids into the tablet matrix. Examples of excipients that facilitate tablet disintegration by swelling in the presence of aqueous fluids include:

– sodium starch glycolate
– croscarmellose sodium (a cross-linked sodium carboxymethyl-cellulose) (usual working range 0.5–5.0% w/w)
– crospovidone (a cross-linked polyvinylpyrollidone) (usual working range 2–5% w/w)
– pregelatinised starch (usual working concentration of 5% w/w).

■ Tablet disintegration may also be mediated by the production of gas whenever the tablet contacts aqueous fluids. This is the mechanism of disintegration of effervescent tablets.

Lubricants

During compression lubricants act at the interface between the face of the die and the surface of the tablet and act to reduce the friction at this interface during ejection of the tablet from the tablet press. Inadequate lubrication of this interface results in the production of tablets with a pitted surface and is due to the inability of the tablet surface to detach from the surface of the tablet die. The appearance of the tablet is an important pharmacopoeial (and consumer) requirement and, therefore, inadequate lubrication will lead to rejection of the tablet batch.

There are two main categories of lubricant: (1) insoluble; and (2) soluble.

Insoluble lubricants

Insoluble lubricants are added to the final mixing stage prior to tablet compression. The concentration of lubricant used is an important consideration in the subsequent disintegration and dissolution of the drug from the tablet: high lubricant concentrations result in reduced rates of disintegration and dissolution and possible failure of the quality control tests for these parameters. Conversely insufficient concentrations of disintegrant, as stated above, may lead to tableting defects. In

addition to concentration, the time of mixing of the lubricant with the granules/powders and the particle size of the lubricant will affect the performance of the lubricant. Overmixing (in terms of both shearing stress and time) may adversely affect tablet disintegration and drug dissolution. In particular mixing of the disintegrant and the insoluble lubricant together should be avoided as this leads to the formation of a film of lubricant on the surface of the disintegrant, which subsequently reduces the wettability of, and water uptake by, the disintegrant, leading to compromised disintegration. Furthermore, it has been shown that the efficacy of the lubricant is enhanced if the surface area is increased (i.e. the particle size of the lubricant is decreased). Examples of insoluble lubricants that are commonly used are as follows:

- *Magnesium stearate.* Magnesium stearate is not a pure compound, being a mixture of magnesium salts of a range of fatty acids, predominantly (but not exclusively) stearic and palmitic acid. The lubricant properties of magnesium stearate are particularly susceptible to variations in the physical properties, e.g. specific surface area/particle size, crystalline structure, moisture content and the composition of fatty acids. Information regarding the chemical properties and physical composition of the batch of magnesium stearate used must therefore be available to the formulator prior to use as a tablet lubricant. Bearing these factors in mind, the concentration range of magnesium stearate employed is 0.25–0.50% w/w.

- *Stearic acid.* Similar to magnesium stearate, stearic acid is a mixture of stearic (> 40% w/w) and palmitic acids, the sum of the two acid components being greater than 90% w/w. As above, there is a wide range of grades of stearic acid that differ in their physical and chemical properties, e.g. melting range, acid, iodine and saponification values. The concentration range of stearic acid employed as a lubricant is 1.0–3.0% w/w.

- *Glyceryl behenate.* Glyceryl behenate is a mixture of glyceryl fatty acid esters, in which the concentration of monoesters (glyceryl monodocosanoic acid) should be between 12% and 18% w/w. The remaining concentration is composed of glyceryl didocosanoic acid and glyceryl tridocosanoic. It is employed as a tablet lubricant within the concentration of 1.0–3.0% w/w.

- *Glyceryl palmitostearate.* Glyceryl palmitostearate is a mixture of mono-, di- and triglycerides of fatty acids containing between 16 and 18 carbon atoms. It is used as a lubricant in tablet manufacture at concentrations between 1.0 and 3.0% w/w.

Soluble lubricants

Soluble lubricants are principally employed to overcome the possible deleterious effects of their insoluble counterparts on the time required for tablet disintegration and hence drug dissolution. The use of soluble lubricants does overcome these issues; however, for the most part, the efficacy of insoluble lubricants is superior to soluble lubricants. Examples of soluble lubricants include:

- *Polyethylene glycol (PEG).* Typically, PEGs, e.g. PEG 4000, 6000 and 8000, that exhibit large average molecular weights are used as lubricants in the manufacture of tablets. The efficacy as a lubricant is less than that of magnesium stearate.
- *Polyoxyethylene stearates.* The polyoxyethylene stearates belong to a group of polyethyoxylated derivatives of stearic acid that differ in the number of oxyethylene repeating groups along the polymer chain and whether present as a mono- or distearate. All members of this group are non-ionic surfactants that may be employed as tablet lubricants within the concentration range of 1.0–2.0% w/w.
- *Lauryl sulphate salts.* Salts of lauryl sulphate (e.g. sodium, magnesium) are anionic surfactants that may be employed as lubricants within the concentration range of 1.0–2.0% w/w. In addition to their action as lubricants, the dissolution of poorly soluble drugs from tablets will be enhanced due to the surface-active activity of these agents.

Glidants

Glidants act to enhance the flow properties of the powders within the hopper and into the tablet die in the tablet press. The reduced friction between the powders/granules and the surfaces of the hopper and dies has been suggested to be due to the ability of the particles of the glidants to locate within the spaces between the particles/granules. To achieve this effect it is therefore necessary for the glidant particles to be firstly small and, secondly, to be arranged at the surface of the particles/granules. Glidants are typically hydrophobic and therefore care should be taken to ensure that the concentration of glidants used in the formulation does not (in a similar fashion to lubricants) adversely affect tablet disintegration and drug dissolution. Examples of glidants used in tablet manufacture include: (1) talc; and (2) colloidal silicon dioxide.

Talc

Chemically talc is hydrated magnesium silicate (typically $Mg_6(SiO_2)_4(OH)_4$) and is a crystalline material of small particle size. It is typically used as a glidant in tablet formulations at concentrations between 5.0 and 30.0% w/w. Whilst talc is non-

toxic whenever ingested orally, inhalation of large masses of talc may result in the formation of granulomas. This has restricted the use of talc in many pharmaceutical formulations.

Colloidal silicon dioxide

Colloidal silicon dioxide is commonly used in tablet formulations as a glidant (0.1–0.5% w/w) due to a combination of its hydrophobic properties and low (colloidal) particle size (typically < 15 nm).

Miscellaneous excipients used in the formulation of tablets

In addition to the principal tablet components described above, the successful formulation of tablets may require the incorporation of other classes of excipients. These include: (1) adsorbents; (2) sweetening agents/flavours; (3) colours; and (4) surface-active agents.

Adsorbents

Adsorbents are used whenever it is required to include a liquid or semisolid component, e.g. a drug or a flavour, within the tablet formulation. As the production of tablets requires solid components, the liquid/semisolid constituent is adsorbed on to a solid component which, in many cases, may be one of the other components in the tablet formulation (e.g. diluent) during mixing. If this approach is not possible, an adsorbent is specifically included in the formulation. Examples of these include: (1) magnesium oxide/carbonate; and (2) kaolin/bentonite.

Magnesium oxide/carbonate

Magnesium oxide is a solid adsorbent that is commercially available in two forms termed light (less dense, 5 grams, occupying circa 40–50 ml) and heavy (more dense, occupying 10–20 ml). Magnesium carbonate consists of several forms (hydrate, the basic form and anhydrous). Both salts are employed as adsorbents in tablets within the concentration range of 0.5–1.0% w/w. In addition, magnesium oxide may also be used as a glidant whereas magnesium carbonate is used as a diluent. Both salts are basic and therefore may interact with acidic drugs.

Kaolin/bentonite

These are natural materials that are composed of hydrate aluminium silicate. Unlike kaolin, bentonite is a colloidal material and therefore the particle size of this excipient is lower than for kaolin. These excipients are employed as adsorbents within the concentration range of 1.0–2.0% w/w.

Sweetening agents/flavours

Sweetening agents and flavours (in accordance with other dosage forms) are employed to control the taste and hence the acceptability of tablets. These agents are of particular importance if the conventional tablet contains a bitter drug or, more importantly, if the tablet is a chewable tablet. The excipients that are used to increase the sweetness and improve the flavour of tablets have been described in previous chapters.

Colours

Coloured tablets are generally formulated either to improve the appearance or to identify the finished product uniquely. In some formulations the drug is coloured and, when manufactured, may result in the tablet exhibiting a speckled, and hence, inelegant appearance. To obviate this problem, an appropriate colouring agent is included in the tablet formulation. It is important that the colour is distributed equally throughout the tablet and this is normally achieved by adding a water-soluble colour to the granulation liquid in the wet granulation method of manufacturing tablets.

Surface-active agents

Surface-active agents may be incorporated into tablets to improve the wetting properties of hydrophobic tablets and hence increase the rate of tablet disintegration. In addition, surface-active agents may increase the aqueous solubility of poorly soluble drugs in the gastrointestinal tract and, as a result, the rate of dissolution of the active agent will increase. One of the most popular choices of surface-active agent for this purpose is sodium lauryl sulphate. However, it should be noted that the surface-active agent should not interact with the therapeutic agent as this may affect the dissolution rate of the drug.

Tips

Tablets will generally contain several excipients that are required to ensure that the tablets may be successfully manufactured and will provide the required biological performance.

Certain excipients are common to the vast majority of tablets, e.g. lactose, stearates, microcrystalline cellulose.

Methods used for the manufacture of tablets

Tablets are commonly manufactured by one of the following manufacturing processes:

- wet granulation
- dry granulation (slugging or roller compaction)
- direct compression.

The above methods may be subdivided into two distinct classes depending on the requirement for a liquid in the manufacturing process. Consequently, in wet granulation a liquid is required in the manufacture of the granules (which are then compressed into

tablets), whereas in the other methods no liquids are required. In general, the choice of method for the manufacture of tablets is dependent on a number of factors, including:

■ the physical and chemical stability of the therapeutic agent during the manufacturing process
■ the availability of the necessary processing equipment
■ the cost of the manufacturing process
■ the excipients used to formulate the product.

The key aspects of each manufacturing method are summarised in the following section.

Manufacture of tablets by wet granulation

A stepwise summary of the manufacturing steps used in the manufacture of tablets by the wet granulation method is detailed below.

Stage 1: mixing of the therapeutic agent with the powdered excipients (excluding the lubricant)

This step involves the introduction of the powdered excipients and drug (excluding the lubricant) into a powder mixer. The mixing speed and time must be sufficient to ensure that a homogeneous mixture is produced, i.e. in which the concentration of each component in each region of the mixer/sample is identical. The efficiency of mixing is enhanced by the use of powders that have similar average particle size/distribution, although this is often not the case in many mixing operations. Generally the shear rate required to mix pharmaceutical powders is low. Examples of mixers that are used within the industry include:

■ *Planetary bowl mixer.* The planetary bowl mixer is a vertical mixer in which the powders are mixed in a bowl using a rotating mixing attachment. The mixing arm is connected to the mixer by means of a shaft whose movement is offset. In addition, the mixing shaft rotates around the mixing bowl to ensure that the contents of the bowl are thoroughly mixed. The movement of the mixing shaft and mixing arm is termed planetary due to the similarity with the movement of the planets around the sun.
■ *Rotating drum mixers.* In rotating drum mixers, the drum is attached to a drive shaft, which is then attached to a motor. The drive shaft rotates and, in so doing, the drum – and hence the powdered contents – is rotated. There are a number of available mixing geometries.
■ *High-speed mixers.* High-speed mixers are commonly employed in the pharmaceutical industry due to their excellent powder-mixing properties and their combined use as a mixer and wet granulator. The powders are contained

within a stainless-steel mixing bowl, on the bottom of which is a horizontal three-bladed mixing head (impeller) that is capable of high-speed rotation. Additional mixing of the powder bed is performed through the (horizontal or vertical) introduction of a second impeller into the powder bed. The combined actions of the two impellers offer excellent mixing properties. Following mixing the powder mixture is discharged through a screen.

- *Ribbon/trough mixers.* In ribbon/trough mixers, mixing is performed via the rotation of blades that are housed within a central drive shaft. Efficient mixing may be achieved in short mixing times using, if required, high shear rates. Ribbon and trough mixers differ in the nature of the mixing geometries (as shown in Figure 9.6).

Figure 9.6 Commercially available (a) ribbon and (b) trough mixers, illustrating the mechanism of mixing associated with the different mixing geometries used in each mixer: (a) refers to ribbon mixers whereas (b) illustrates a typical geometry that is used in trough mixers. (With permission from http://www.kemutec.com/htm/products/gardner/mixers/ ribbon.htm and http://www.kemutec.com/htm/products/gardner/mixers/plough.htm.)

(a)

(b)
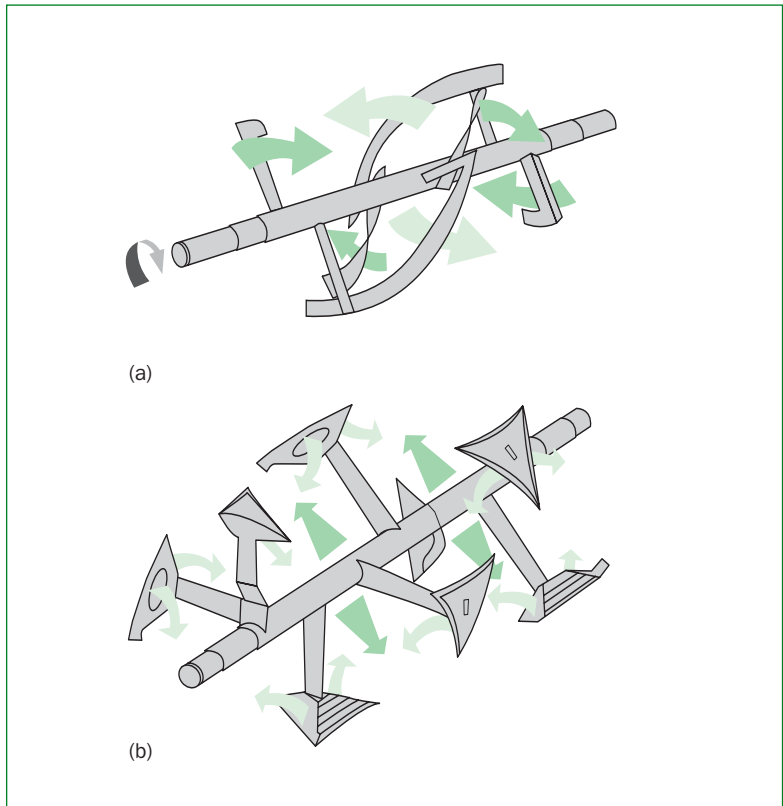
Stage 2: wet granulation of the powder mix

Granulation is a unit operation in which mixed powders are aggregated into and retained as larger particles (circa 0.2–4.0 mm in diameter). Ideally the chemical composition of the granules should be homogeneous. There are several advantages for the use of granules in tablet manufacture, as follows:

- *Prevention of segregation of powder components during the tableting process or during storage.* Following mixing, the individual particulate components of powders will tend to segregate due to differences in the particle size and density of each component; the degree of segregation increases as the time of storage increases. If the segregated powder is then compressed into tablets, the intrabatch chemical composition of the dosage form will vary, resulting in issues regarding the quality of the product. This problem is obviated if the powders are granulated, as there is no component segregation within the granule.
- *Enhancement of the flow properties.* Due to differences in particle size/shape and cohesive/electrostatic properties, the flow properties of mixed powders from the tablet hopper into the tablet die may be inappropriate and irregular, once again leading to issues regarding the intrabatch variability of tablet dosage forms. The flow properties of granules are both improved and more reproducible than those of mixed powder systems and are relatively unaffected by the adsorption of low amounts of moisture (during storage). Conversely, the adsorption of moisture on to the surface of powders may result in caking, which may ultimately adversely affect powder flow.
- *Enhancement of the compaction properties.* In comparison to powder mixes, the compaction properties of granules are improved due to the presence of the binder component on the surface of the granules, leading to greater intergranule adhesive interactions.
- *Lower incidence of dust production.* One problem associated with the processing of powders into tablets is possible dust production. This problem is obviated with the use of granules.

Description of wet granulation

Granulation is a unit operation in which mixed powders are simultaneously mixed with a suitable fluid, e.g. water, isopropanol or ethanol (or mixtures thereof). To achieve cohesion between the powders, it is necessary to include a binding agent within the formulation, either in the solid state within the powder mix or dissolved in the binding fluid. Powder mixing, in conjunction with the cohesive properties of the binder, enables

the formation of granules. A description of commonly employed strategies for wet granulation is outlined below.

Oscillating granulator

In this strategy, the mixing of the formulation components is performed using a relatively low shear mixer, e.g. a planetary bowl mixer. The granulating fluid (which may contain the appropriate concentration of binder) is then sprayed on to the powder mass whilst maintaining mixing. The wetted powder mass is then passed into an oscillating granulator, which forces the powder mass through a metal screen under the action of an oscillatory stress. This process is illustrated in Figure 9.7.

Figure 9.7 Diagrammatic representation of (a) the oscillatory action and (b) the orientation of sieves in an oscillatory granulator. (Adapted from www.granulatorsindia.com.)
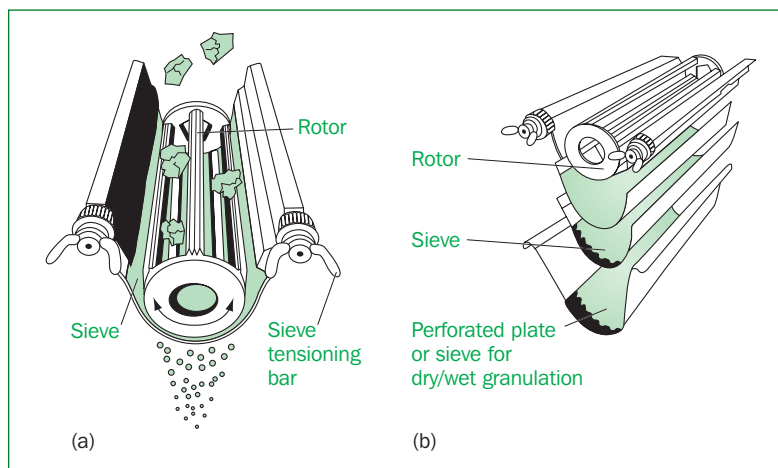
Providing the correct mass of granulation fluid has been employed, granules are produced following pass through the oscillatory granulator. The size of the granules produced using this method is dependent on the size of the metal screen used within the granulator. It is worth noting that the powdered binder may either be added to the powder mix or dissolved within the granulation fluid and added to the powder mix. In the former scenario the granulation fluid is composed only of the chosen solvent. Due to the complete dissolution within the granulation fluid prior to addition to the powder mix, the cohesive properties of granules produced by the latter method are greater.

High-speed mixers/granulators

More recently, it has become common practice to perform the powder-mixing and wet granulation steps within a single

operation. Typically, high-speed mixers/granulators are employed,
e.g.:
- the Diosna mixer/granulator
- Collette-Gral mixer/granulator.

The design and operation of these have been described previously
in this chapter.

Fluidised-bed granulation

In this technique, the mixed powders are fluidised within a
chamber by the action of a positive (temperature-controlled)
stream of air. Whilst in this dispersed state, the particles are
sprayed with the granulation fluid (usually containing dissolved
binder), causing the particles to interact with (adhere to) each
other. The fluidised-bed granulator contains filters to prevent the
escape of fluidised particles into the environment, thereby
enabling these to be recycled into the fluidised granules.
Following completion of the spraying step, fluidisation of the
granules is maintained to enable evaporation of the solvent used
in the granulation fluid. A schematic representation of fluid-bed
granulation is presented in Figure 9.8.

Figure 9.8 Diagrammatic representation of the operation of fluid-bed granulators.
(a) The powders are suspended by a vertical flow of air from the bottom of the
granulator. The granulation fluid is sprayed on to the powders from the top of the
granulator. (b) The powders are similarly suspended by a vertical upward flow of air
but the airflow is tangential, enabling flow in a circular fashion. The granulation fluid
is sprayed into this tangential airflow. (Adapted from http://www.glatt.com/e/01_
technologien/01_04_09.htm.)

(a)   (b)
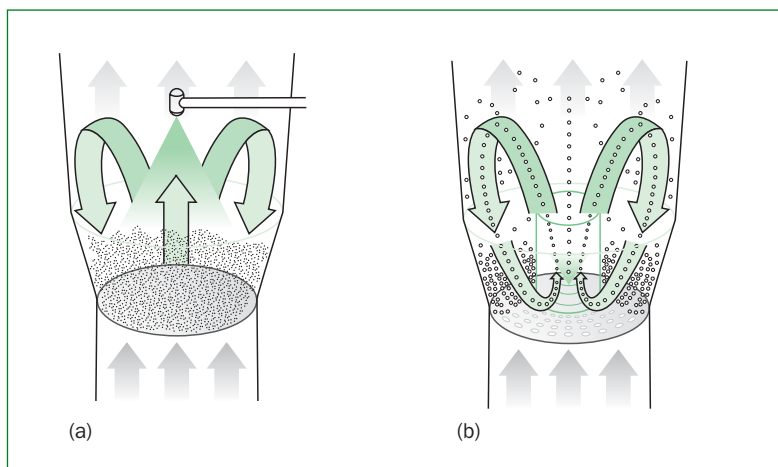
Extrusion

Extrusion is a technique in which a premixed powder mass
(containing the required formulation components and to which
the required mass of granulation fluid has been added) is placed

into the barrel of the extruder via a hopper. In the barrel, the wet mass is moved horizontally from the end adjacent to the hopper by the turning motion of either a single-screw or a twin-screw system. This action also serves to enhance the mixing of the granulation fluid within the wet mass. Following passage through the barrel of the extruder, the wet mass is passed through a perforated plate (termed the die) into lengths. To obtain granules successfully, it is important that:

- there is good dispersion of the granulation fluid through the wet mass
- the mechanical properties of the wet mass (and hence extrudate) should facilitate deformation
- the formulation should be sufficiently cohesive to form granules but not adhesive (to prevent aggregation of formed granules). When formulated and processed correctly, the extruded strands should break to produce granules of uniform particle size.

Stage 3: processing granules into tablets

Following the production of (drug-containing) granules, further unit operations are required for the successful production of tablets. These are as follows:

- *Drying of the granules.* Following production, granules are then dried to facilitate further processing. There are several types of drier that are used within the pharmaceutical industry for this purpose, including:
  - *The shelf or tray drier.* The shelf or tray drier is similar in design to a conventional oven. In this the wet granules are uniformly located on a series of horizontally arranged, shallow trays. Air enters the drier where it is warmed by heaters prior to being passed across the surface of the granules. The moisture is then passed from the drier through an outlet. An example of an industrial tray drier is shown in Figure 9.9.

    The drying efficiency of this type of system may be enhanced by the use of a vacuum. In this design there is no inlet or outlet for air and, following the introduction of the granules into the shelves, the door is closed and a vacuum created using a vacuum pump. The removed moisture is condensed into a liquid using a condenser and collected prior to disposal. The use of the vacuum design lowers both the time and temperature of the drying process.
  - *The fluidised-bed drier.* The design and operation of the fluidised-bed drier for the production of granules have been addressed in the previous section. When used as a drier, the granules are suspended in the fluidised warmed air, enabling relatively rapid drying. There are several advantages of the

use of fluidised-bed driers for the drying of granules that have rendered this technique popular within the pharmaceutical industry, including:

- Excellent heat transfer to and mass (moisture) transfer from the granules, due primarily to the suspension in, and drying of, individual granules in the warmed airflow. This enables granules to be rapidly and reproducibly dried.
- Accurate control of the drying conditions may be achieved using a fluidised-bed drier, e.g. airflow, temperature.

Conversely, limitations of this technique include:

- Attrition of granules whilst suspended in the warmed airflow.
- Loss of particles from the drier. Particles are normally prevented from escaping by means of a bag filter.
- Generation of static electricity within the fluid-bed drier (due to the rapid movement of particles), which may lead to explosions (particularly if a volatile solvent has been used as the granulation fluid).

■ *Milling of the granules (reduction in the granule size).* Following drying, the next stage in the production of tablets by wet granulation usually involves milling of the granules to

the required particle size. This step is employed for two main reasons:

– to control the particle size (and particle size distribution), which, in turn, improves both the flow of the granules into the tablet die and the fill of the granules within the die
– the choice of the granule size is determined by the size of the die (and hence the final tablet size). In general the granule size decreases as the tablet size decreases.

Reduction in the size of granules may be performed using a range of particle size reduction methods, including:

■ *Oscillating granulator.* In this method the dried granules are forced through a screen of defined mesh size, as described previously. The mesh size of the screen is usually smaller than that used for granulation.
■ *Quadro Comil.* In the Quadro Comil (Figure 9.10) the dried granules are placed inside a conical chamber (containing a screen of defined mesh size). The granules are then passed through the screen in a centrifugal manner by the action of a rotating impeller, after which the granules are collected.

Figure 9.10 The laboratory Quadro Comil. (a) The encased drive unit, the conical chamber and the powder collection zone. (b) The conical chamber, the screen and the rotating impeller. (Courtesy of Dr Brendan Muldoon, Warner Chilcott PLC.)

(a)

(b)

■ *Mixing of the granules with lubricant.* The penultimate stage in the manufacture of tablets using wet granulation involves a final mixing of the dried granules with the lubricant. This step is normally conducted using the same equipment that was employed to mix the powders at the start of the process. It is important to note that both the time and shear rate of mixing in this step will affect the final performance of the tablet, in terms of both the quality of the tablet produced and the subsequent drug release.

■ *Compression of the formulation into tablets.* This unit
   operation will be described at a later section in this chapter.

Stages of wet granulation

During wet granulation particles interact with each other to form
aggregates (granules); the extent of this interaction is a major
contributor to the performance of the finished dosage form. There
are two primary stages involved in particle–particle interactions
during wet granulation, as follows:

■ *Particle–particle interactions facilitated by the formation of
   liquid bridges.* As may be expected, the presence and,
   importantly, volume of the granulation medium contribute
   greatly to the granulation process. The interactions between
   particles are dependent on the mass of free water available
   and these are characterised into four categories (as depicted in
   Figure 9.11):

– *Pendular state.* The pendular state (Figure 9.11a) is
   established at lower moisture levels and relates to the
   formation of liquid bridges (connections) between adjacent
   particles. The attractive forces between the particles are due
   to both interfacial and hydrostatic effects.

– *Funicular state.* As the mass of water increases, the mass of
   the liquid bridges – and, accordingly, the adhesive forces –
   increases between the particles (Figure 9.11b).

– *Capillary state.* Further increases in the mass of available
   water result in the displacement of air from the free volume
   between the particles (Figure 9.11c). The adhesive interaction
   between the particles is now principally due to a capillary
   effect at the liquid/air interface (i.e. at the surface of the
   growing granule).

(a)   (b)   (c)   (d)
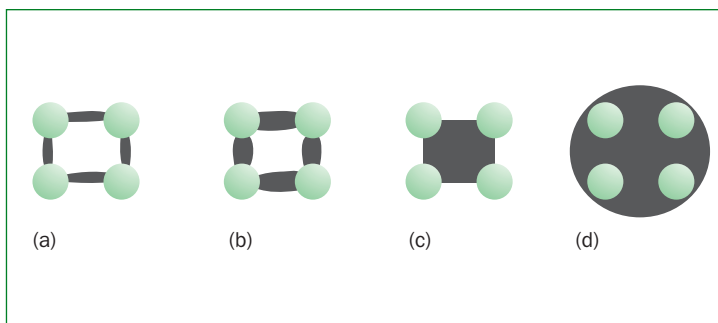
Figure 9.11
Diagrammatic
representation of
particle–particle
interactions during
wet granulation.
(a) Pendular;
(b) funicular; and
(c) capillary states.
(d) Overwetted
(undesirable) state.

– The final state (Figure 9.11d) is the overwetted state and this
   is pharmaceutically unacceptable due to the associated
   processing (mixing and drying) problems. Following drying,
   individual granules will not be produced and, in addition,
   due to the excessive mass of binder associated with these

particles, the disintegration and dissolution properties of the dosage form may be unduly extended. It should be noted that, if an insufficient mass of granulating fluid is used in wet granulation (i.e. underwetted), the granules will disintegrate during the subsequent processing steps and, if processed into tablets, the tablets will exhibit poor mechanical strength.

■ *Particle–particle interactions facilitated by the formation of solid bridges.* Solid bridges, consisting predominantly of the polymeric binder, are formed following drying. As drying progresses a polymeric network (resembling the pendular structure) is formed, which largely contributes to the mechanical properties of the resultant granule. Overdrying of the granule results in destruction of these bridges (due to the compromised effects on their mechanical properties). This is the main mechanism of action of polymeric binders. Under certain conditions, crystallisation of the binder on to the surface of the dispersed particles may occur on drying (e.g. sugar), leading to a hardening effect. If the drug is soluble in the granulation fluid, crystallisation may additionally occur, which, in turn, may influence the physicochemical properties of the formulation, e.g. by affecting the hardness of the formed tablet or by affecting drug dissolution.

Advantages and disadvantages of wet granulation

Wet granulation is a popular technique within the pharmaceutical industry for the manufacture of tablets. There are several advantages and disadvantages associated with this technique.

Advantages

■ Reduced segregation of formulation components during storage and/or processing, leading to reduced intra- and interbatch variability
■ useful technique for the manufacture of tablets containing low concentrations of therapeutic agent
■ employs conventional excipients and therefore is not dependent on the inclusion of special grades of excipient (cf. the requirement for spray-dried excipients in the direct compression method of tablet manufacture)
■ most manufacturing plants are built around wet granulation tablet manufacture
■ tablets produced by wet granulation are amenable to post-processing unit operations, e.g. tablet-coating techniques.

Disadvantages

■ Often several processing steps are required.
■ Solvents are required in the process: this leads to a number of concerns, e.g.:

- Drug degradation may occur in the presence of the solvent. This is particularly relevant if water is used as the granulation medium due to the susceptibility of some drugs to hydrolysis. To overcome this concern, a hydroalcoholic (water/alcohol) or an alcohol (ethanol or isopropanol) granulation medium should be used.
- The drug may be soluble in the granulation fluid. During the drying process the drug will then precipitate/crystallise, resulting in possible changes in the polymorphic form. If the drug and some excipients are soluble in the granulation medium, subsequent drying will result in deposition of these components on the surface of the insoluble particles and, in so doing, this may enhance the hardness of the granule (and hence both the tablet and biological properties).
- Heat is required to remove the solvent. This may result in the degradation of thermally labile therapeutic agents. In addition, drying is a costly operation and, furthermore, if alcohols are used as the granulation medium, there are issues regarding solvent recovery and flammability.

Manufacture of tablets by dry granulation

Similar to wet granulation, the manufacture of tablets by dry granulation involves the production and subsequent compression of granules into the final dosage form. In dry granulation no solvent is required; the aggregation of particles into granules is facilitated by the application of high stresses to the mixed powders. There are two methods by which dry granules are formed: (1) slugging; and (2) roller compaction.

Slugging

In this technique the powders are mixed (as described previously) and then compressed into a primordial *oversized* tablet using a tableting press that is capable of applying a high stress (to ensure that aggregation of the particles and then aggregation of granules occur during compaction). Following this the tablet is milled to produce granules of the required size (using conventional milling equipment, as described previously).

Tips

Wet granulation is the most commonly used method for the manufacture of tablets.

As water is frequently used as the granulation fluid (and heat is employed to dry the formed granules), it is important to ensure that the therapeutic agent is chemically stable during the granulation process.

Tablets manufactured by wet granulation exhibit sufficient mechanical properties to be subsequently exposed to other unit operations, e.g. film coating.

Granule (and hence tablet) quality is directly affected by the choice and concentration of binder and by the type and volume of granulation fluid employed.

Due to the number of unit operations required, the manufacture of tablets by wet granulation is not as efficient as other methods, e.g. direct compression.

Roller compaction

In roller compaction the formulation ingredients are mixed (as described previously) and are then compressed using a roller compactor. In this the powders are fed from a hopper on to a moving belt and then transported to, and compressed by, the passage between the narrow gap between two (oppositely) rotating rollers to produce a sheet/film of compressed material. The compressed sheet is then milled to produce granules of the required size (again using conventional milling equipment, as described previously).

Excipients used in dry granulation

The types of excipients used to prepare tablets (and the rationale for their use) using dry granulation methods are similar to those described for wet granulation. Typically, the following excipients are required:

- *diluent/filler*, e.g.:
  - anhydrous lactose or lactose monohydrate
  - starch
  - dibasic calcium phosphate
  - MCC.
- *disintegrants*, e.g.:
  - starch
  - MCC
  - sodium starch glycolate
  - croscarmellose sodium
  - crospovidone.
- *lubricants,* e.g.:
  - stearates (magnesium stearate, stearic acid)
  - glyceryl fatty acid esters (glyceryl behenate, glyceryl palmitostearate)
  - PEG
  - polyoxyethylene stearates
  - sodium lauryl sulphate.
- *glidants*, e.g.:
  - talc
  - colloidal silicon dioxide
- miscellaneous excipients (colours, sweetening agents, etc.).

Stages of dry granulation

In dry granulation, the aggregation of particles is facilitated by the application of high compression stresses and therefore the stages/mechanisms of granule formation are different from those described for wet granulation. In dry granulation particle–particle interactions occur due to: (1) electrostatic forces; (2) van der Waals interactions; and (3) melting of components within the powder mix.

Electrostatic forces

These are thought to play a role in the initial cohesive interaction between particles; however, they do not contribute greatly to the granule strength

Van der Waals interactions

Van der Waals interactions play a major role in the interactions between particles in the dry state. As the reader will be aware (see Chapter 2), van der Waals forces increase in magnitude as the distance between particles decreases. Under normal circumstances, particles are unable to interact due to the inability of the particles to locate at close distances to one another. However, this problem is obviated in dry granulation by the application of high stresses to the powder mix.

Melting of components within the powder mix

During dry granulation processing the powders will be exposed to shear stresses that may result in the partial melting of excipients (of low melting point). Upon cooling solidification of the excipients will occur, resulting in increased interactions between adjacent particles. This mechanism does not strongly contribute to particle–particle interactions in dry granulation.

Following dry granulation, the granules are processed using a series of unit operations (identical to those used in wet granulation). These are as follows:

- milling of the granules (reduction in the granule size)
- mixing of the granules with lubricant
- compression of the formulation into tablets.

This unit operation will be described later in this chapter.

Advantages and disadvantages of dry granulation

The popularity of dry granulation for the manufacture of tablets has decreased in recent years, having been superseded by direct compression (see following section). However, both slugging and roller compaction are still employed in tablet manufacture. As with wet granulation, there are several advantages and disadvantages associated with these techniques.

Advantages of dry granulation

- Both roller compaction and slugging require conventional (i.e. non-specialist) grades of excipients.
- These methods are not generally associated with alterations in drug morphology during processing.
- No heat or solvents are required.

Disadvantages of dry granulation

- Specialist equipment is required for granulation by roller compaction.
- Segregation of components may occur postmixing.
- There may be issues regarding powder flow.
- The final tablets produced by dry granulation tend to be softer than those produced by wet granulation, rendering them more difficult to process using post-tableting techniques, e.g. film coating.
- Slugging and roller compaction lead to the generation of considerable dust. Therefore, containment measures are required. Furthermore, there may be a reduction in the yield of tablets.

> **Tip**
>
> Dry granulation is an older method of manufacture of tablets. Its use has been superseded by other methods, most notably direct compression.

Manufacture of tablets by direct compression

The manufacture of tablets using wet granulation or dry granulation methods is, in light of the requirement for a series of unit operations, both time-consuming and potentially costly. A potentially more attractive option for the manufacture of tablets involves powder mixing and subsequent compression of the powder mix, thereby obviating the need for granulation (and related unit operations). This process is termed direct compression. The mechanisms of particle–particle interactions in tablets produced by direct compression are similar to those operative in tablets produced by dry granulation and roller compaction.

Stages in the manufacture of tablets by direct compression

A summary of the various steps used in the manufacture of tablets by direct compression is detailed below.

Premilling of formulation components

In the preparation of tablets by direct compression, both the particle size and particle size distribution of the therapeutic agent and the excipients are important determinants of the compression properties of the powder mix. Whilst the excipients may be purchased to a particular particle size specification, frequently the particle size properties of the drug may require to be modified by milling, using, for example, the Quadro Comil or a high-energy mill, the Fitzmill. The operation of the Quadro Comil has been previously outlined. The operation of the Fitzmill is illustrated in Figure 9.12.

In this process the powder is fed into a chamber where it is exposed to a rotating-blade system. The particle size is reduced by the cutting action of the blades or the impaction of the

Figure 9.12 The operation of the Fitzmill. (Adapted from http://www.fitzmill.com/ pharmaceutical/size_reduction/theory_applications/theory_applications_pD37F.html.)

The feed throat introduces material on a tangential path to the comminuting chamber

Blade profile Helps determine degree of reduction based on material being processed

Screen type Helps regulate particle output within a specified size range

Rotor speed Helps regulate particle output within the specified size range
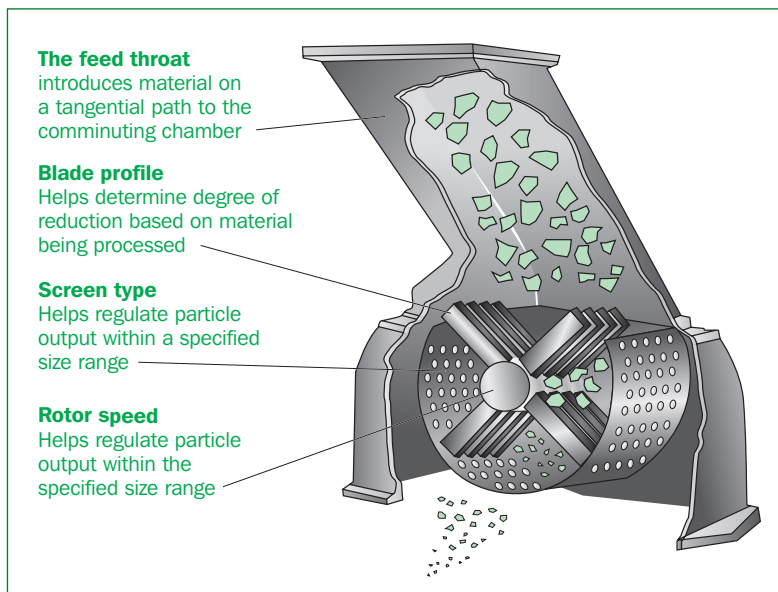
particles by the blades and the screen on the periphery of the chamber. The milled particles are then discharged through the screen. The particle size/particle size distribution may be controlled by altering the rotor (and hence blade) speed, the morphology of the blades and the mesh size of the screen.

Mixing of the therapeutic agent with the powdered excipients (including the lubricant)

This step involves introducing all the powdered excipients and drug (including the lubricant) into a powder mixer. The types of mixers used for this process are identical to those that have been previously described in this chapter and include:

- planetary bowl mixer
- rotating drum mixers, e.g. the Y-cone, cube or double-cone mixers
- high-speed mixers, e.g the Diosna mixer.

Compression of the mixed powders into tablets

This unit operation will be described at a later section in this chapter.

Excipients used in the manufacture of tablets by direct compression

In a similar fashion to the other methods of tablet manufacture, the production of tablets necessitates the inclusion of several

types of excipients. The rationale for the inclusion of these excipients is identical to that explained previously. However, and most importantly, to achieve the correct powder flow and compression properties, certain grades of excipients are specifically employed for the manufacture of tablets by direct compression. These grades have typically been prepared by specific methods (e.g. spray drying) to achieve the correct physicochemical properties (e.g. particle size/distribution and flow properties). Examples of excipients that are employed for the formulation and manufacture of tablets by direct compression include: (1) diluent; (2) compression aid; (3) distintegrant; and (4) lubricants and glidants.

Diluent
Examples of commonly used diluents include:
- spray-dried lactose (e.g. Lactopress Spray-Dried, Lactopress Spray-Dried 250, Pharmatose DCL 11, Pharmatose DCL 14)
- dicalcium phosphate (e.g. Encompress grades)
- mannitol (granular or spray-dried grades, e.g. Pearlitol)
- sorbitol
- MCC (e.g. Avicel pH-102).

Compression aid
Examples of commonly used compression aids include MCC (e.g. Avicel pH-102).

Disintegrant
Examples of commonly used disintegrants include:
- pregelatinised starch (e.g. Starch 1500)
- sodium starch glycolate (e.g. Explotab, Primojel)
- croscarmellose sodium (e.g. Ac-Di-Sol, Explocel)
- crospovidone (e.g. Polyplasdone XL, Polyplasdone XL-10, Kollidon CL, Kollidon CL-M).

Lubricants and glidants
The types of lubricants and glidants that are employed for the manufacture of tablets by direct compression are similar to those used in other tablet manufacture methods and include:
- lubricants (e.g. magnesium stearate, stearic acid, sodium stearyl fumarate)
- glidants (e.g. talc, colloidal silicon dioxide).

Advantages and disadvantages of tablet manufacture by direct compression
The advantages of the direct compression method may at this stage be obvious; however, it is important to note that this method does suffer from several disadvantages. As for other methods, this

section details the main advantages and disadvantages of this manufacturing method.

Advantages of direct compression
- There are fewer processing steps (unit operations) and therefore the method is potentially more cost-effective than other methods.
- Direct compression does not require the use of water or other solvents. This therefore negates potential problems regarding the stability of therapeutic agents in the presence of the solvents. In addition, heating (a costly unit operation) is not required in direct compression.
- Lubrication is performed in the same vessel as powder mixing, thereby reducing both transfer losses and contamination of equipment.

Disadvantages of direct compression
- Specialist (and more expensive) excipients are required. The excipients that are used for direct compression processing are typically specially processed (e.g. spray-dried) for this application to achieve the correct physicochemical properties.
- The quality of the final dosage form is dependent on the powders being easily mixed and remaining homogeneously mixed. Segregation of the mixed components is minimised by ensuring that the excipients and the dosage forms exhibit similar morphologies, densities and particle size/distribution properties.
- There may be issues regarding powder flow into the tableting machine.
- The final tablets produced by direct compression tend to be softer than those produced by wet granulation, rendering them more difficult to process using post-tableting techniques, e.g. film coating.
- If the loading of the therapeutic agent in the final formulation is high (> circa 10% w/w), the compression properties of the powder mix are significantly affected by the compression properties of the therapeutic agent. In some cases the physical properties of the chosen drug may be inappropriate for compression using this technique.
- Direct compression is not used if a colourant is required in the formulation due to the mottled appearance of the resulting dosage form.

Tips

Direct compression is of interest to the pharmaceutical industry for the manufacture of tablets due to the low number of processing steps and the absence of liquid.

Tablets produced by direct compression are often softer than their counterparts that have been produced by wet granulation and therefore they may be difficult to film-coat.

The cost of excipients for the manufacture of tablets by direct compression is greater than the cost of those used in other methods of tablet manufacture.

The compression process in tableting

The final stage of tableting involves the compression of the granules/powders between the punch and die of the tablet press. There are several identifiable stages within the compression process, which are individually described below.

Stage 1: the filling of the die with the granules/powders

The formulated powders or granules are fed (usually by gravitational flow) into the die section, i.e. in the space between the lower and upper punches, from the hopper on the tablet press. The volume of space occupied by the granules/powders is defined by the position of the lower punch and the die plate (also where the feed shoe of the hopper resides). Accordingly, the mass of solid material within this space (and hence the size of the tablet) is altered by increasing/decreasing the position of the lower punch.

Stage 1 is illustrated in Figure 9.13.

Figure 9.13 A diagrammatic representation of the initial stage in tablet formation.
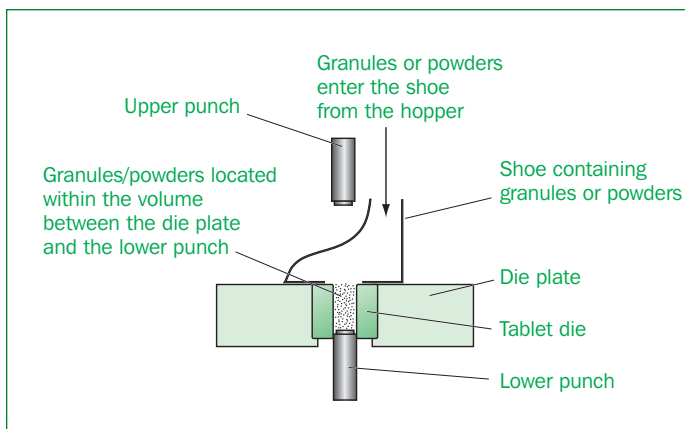
Stage 2: compression of the powder/granule bed

This stage commences with the retraction of the shoe to facilitate access of the upper punch to the powder/granule bed. The upper punch then descends to the powder/granule bed, at which point a stress is applied, thereby compressing the powder/granule bed into a tablet. In general the lower punch remains static during this stage; however, the manufacturing cycle may be designed to enable the lower punch to move vertically upwards during the compression phase. In addition to the choice and concentration of formulation components, changing the stress applied to the powder/granule bed by the upper punch alters the tablet hardness.

Stage 2 is illustrated in Figure 9.14.
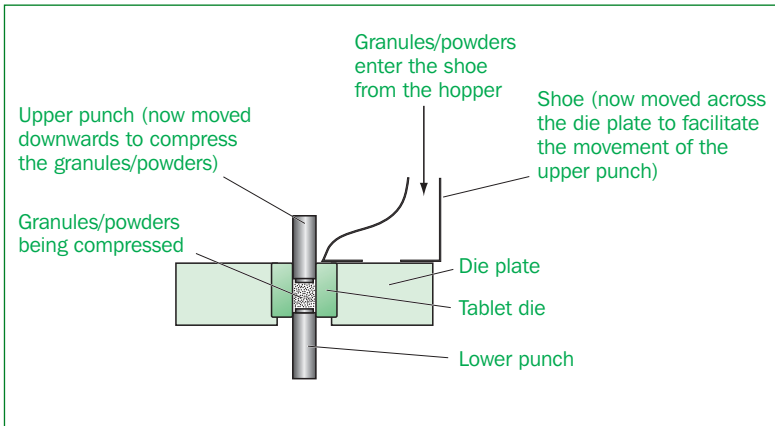
Figure 9.14
Diagrammatic
representation
of the
compression
stage in tablet
formation.

Stage 3: tablet ejection

Following compression, the upper punch is elevated to its original position. At this stage the lower punch moves upwards until it is flush with the die plate. The shoe of the hopper is then moved across the die plate where it pushes the tablet from the lower press. Simultaneously the lower punch descends to its starting position and granules/powders fill the volume between the shoe and the lower plate. At this point the manufacturing sequence returns to stage 1 once more.

   This final stage is illustrated in Figure 9.15.
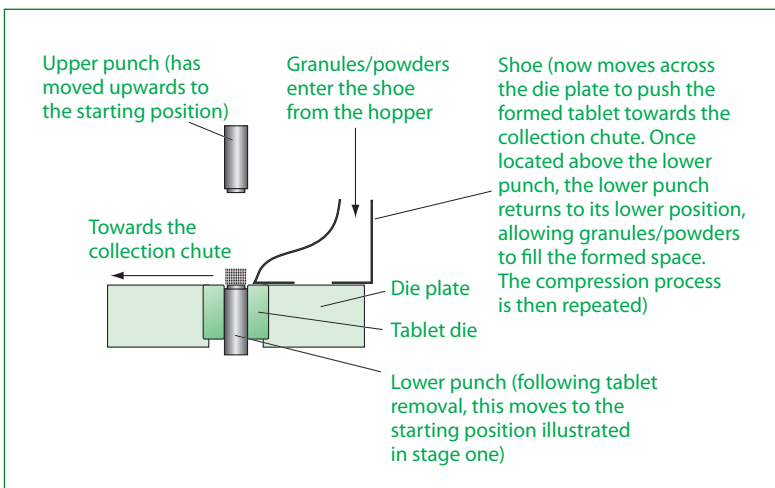
Figure 9.15
Diagrammatic
representation
of the final
stage in tablet
formation.

Types of tablet presses

There are two basic designs of tablet presses used in the pharmaceutical industry: (1) single-punch presses and (2) rotary presses.

Single-punch presses

The operation of the single-punch press has been described in the previous section. This tablet press is composed of one set of punches and a die (Figure 9.16).

Figure 9.16 Examples of punches and dies used in a single-punch tablet press. The tablets produced using these punches will be (a) circular and (b) oval, respectively. (Courtesy of Dr Brendan Muldoon, Warner-Chilcott PLC.)

(a)   (b)

Typically the output from a single-punch tablet press is approximately 200 tablets per minute. The single-punch tablet press is frequently employed for pilot-scale manufacture of tablets and, in addition, it is used for the preparation of primordial tablets in dry granulation (slugging).

Rotary tablet presses

Rotary tablet presses are employed for the large-scale manufacture of tablets, often producing up to 10 000 tablets per minute. As with the single-punch tablet press, the powder/granule bed is compressed between two punches; however, the mechanism by which this process is performed is different. Rotary tablet presses are composed of a series of upper and lower punches (up to 60 per machine) that are housed within a circular die table that rotates in a circular motion. Both punches (upper and lower) are lowered and elevated by the action of an upper and lower roller. The powders/granules are fed from the hopper on to the upper surface of the die table. These are then transported by a feed frame into the die, where they are subsequently compressed by the simultaneous movement of the upper and lower punches. As before, the tablets are removed from the rotating die table into a chute, from which they are collected. A diagrammatic representation of the rotary tablet press is shown in Figure 9.17.
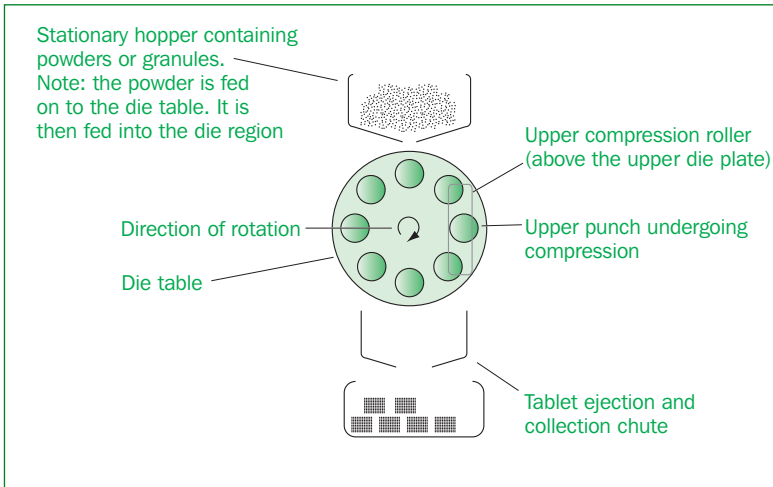
Figure 9.17
Diagrammatic
representation
of the
operation of
the rotary
tablet press.

Compression of powders and granules

Ultimately the preparation of tablets requires the compression of, and subsequent cohesion between, the powders/granules. It is therefore of interest to consider both the various processes that occur within the powder/granule bed during compression and, additionally, the nature of the interactions between the compressed particles. As may be expected, the fate of a powder bed under compression will differ from a granule bed under compression and, accordingly, these two scenarios will be discussed separately. Following the application of a stress, solid particles will undergo deformation, the nature of which is dependent on the magnitude of the applied stress and the time of loading. A generic relationship between applied stress and the strain (proportional elongation) is illustrated in Figure 9.18.
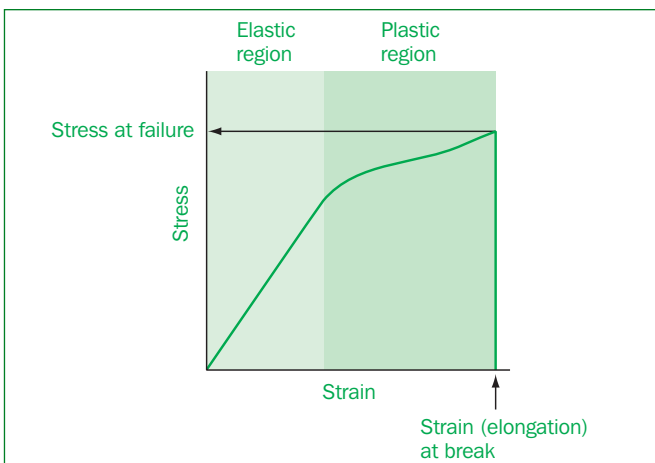
Figure 9.18 The effect of stress on the resultant strain of solid particles.

Certain regions may be identified in this figure: (1) elastic region; (2) plastic region; and (3) fragmentation.

Elastic region

This is the initial *linear* region of the relationship between stress and strain during which the particles undergo elastic deformation. The observed deformation is due to compression of the molecular structure (either crystalline or amorphous) of the particles. The release of the stress within this region will result in the return of the structure of the solid to the equilibrium position. If the compression properties of powders/granules are predominantly elastic, this will result in delamination and subsequent tablet failure. For similar reasons the stresses employed for the manufacture of tablets are most commonly greater than those required for elastic deformation.

Plastic region

This is the second *non-linear* region of the relationship between stress and strain where the particles undergo plastic deformation. Irreversible deformation occurs within this range of applied stresses and is due to the movement of molecules in the direction of the stress. The successful compression of particles requires the inclusion of components that undergo plastic deformation, as there is limited structural recovery of such materials within the plastic region, resulting in good cohesion between compressed particles. Whilst plastic deformation is often employed for the successful compression of particles into tablet, the quality of the compressed tablet may be affected by the inclusion of excipients that do not exhibit this type of deformation

Fragmentation

Fragmentation of particles occurs following the application of a defined stress (termed the ultimate tensile stress) that results in the destruction of intraparticle bonds. In so doing the particle is fractured into a number of smaller particles. Continued application of the applied stress will result in further fragmentation. The surface area of the powder is thus increased, thereby increasing the possible sites for particle–particle interactions. Tablets formed by fragmentation of powders are relatively insensitive to the effects of other excipients; however, the strength of the tablet formed under these increased stresses is often dependent on the nature of fragmentation (e.g. particle size range).

To consider the effects of compression on tablet formation, it is convenient to discuss the fate of powders and granules independently.

Compression of a powder bed during tableting

This may be defined by a number of stages.

- *Stage 1: rearrangement of the powder bed upon the application of a stress.* Following the application of the initial stress, the particles in the powder bed will undergo rearrangement to minimise the free space between particles. The extent of this rearrangement is dictated by both the size of the particles and frictional forces that operate between the particles.

- *Stage 2: deformation of the powders under the applied stress.* During this stage the powders will undergo deformation (elastic, plastic or fragmentation) as a result of exposure to the applied stress. The physicochemical properties of the powders will affect the nature of the predominant deformation type.

  It is important to note that the two main mechanisms of tablet formation in powder systems are plastic deformation and fragmentation. As illustrated in Figure 9.18, the nature of powder deformation is dependent on the magnitude of the applied stress and, in addition, the physicochemical properties of the materials under compression.

- *Stage 3: bonding of the compressed powders.* Following the application of the required stress, interparticle bonding occurs, resulting in the production of a tablet. There are two predominant bonding mechanisms in tablets prepared by direct compression: (1) adsorption and (2) diffusion.

- *Bonding by adsorption.* Following application of the required stress, the repulsive forces between adjacent powder particles will be overcome and hence the particles will be brought into close contact. Akin to caked suspensions, the interaction between particles at close distances of separation is due to van der Waals forces. This is the predominant mechanism of interparticle interactions in tablets formed by the compression of powders.

- *Bonding by diffusion.* Bonding by diffusion occurs whenever there is molecular mobility at the surface of a particle due to, for example, melting or the particle exhibiting rubbery characteristics under the conditions of compression. This increased mobility enables molecular diffusion into and interaction between particles. The prerequisite for this type of interaction is molecular mobility and therefore this is an important transition in tablets containing excipients that exhibit a stress-induced relaxation in the surface mechanical properties.

Compression of a granule bed during tableting

As before, the events that surround the compression of granules into tablets may be described as a series of stages.

- *Stage 1: rearrangement of granule structure.* Following exposure to stress by the movement of the upper punch, the distribution of the granules is rearranged within the space between the two punches. The extent of this movement is small due to the course and regular nature of the granules.
- *Stage 2: granule deformation and bond formation.* As the loading is increased, the granules will undergo initially elastic and then plastic deformation. The granule structure is retained (i.e. the granules are reduced to individual particles), but the stress acts to change both granule shape and porosity due to movement of the individual particles within the granule.
- *Stage 3: formation of intergranule bonds.* As before, the application of the required stress results in the formation of intergranule bonds and hence tablets. Similar to compressed powders, there are two main mechanisms responsible for intergranule bonding, as follows:
  - *Bonding by adsorption.* As the reader will be aware, granules may be formed by wet granulation (involving the presence of a binder) or dry granulation methods. The interaction of granules produced by dry granulation methods (roller compaction or slugging) involves adsorption bonds that are formed between close contact of the granule surfaces; van der Waals forces are responsible for bond formation. This type of interaction is referred to as substrate–substrate interactions. In granules formed by wet granulation, theoretically all the granules have been coated with the polymeric binder. Therefore, in this scenario, adsorption bonds may be formed in a similar fashion. However a range of surface interactions is possible:
    - binder-coated granules with binder-coated granules
    - binder-coated granules with uncoated (substrate) granules
    - uncoated (substrate) granules with uncoated (substrate) granules.
  - *Bonding by diffusion.* In the previous section it was noted that bonding of particles by diffusion requires molecular mobility at the surface of the particle. For uncoated granules this interaction is mostly facilitated by the melting (and solidification) of certain components under the effect of applied stress. However, the interaction of binder-coated particles involves the interaction (by diffusion) of the binder molecules on the surfaces of interacting granules. As before, this increased molecular mobility is a result

Tip

To ensure the production of tablets with optimal mechanical properties, it is important that tablet compression is facilitated by both elastic and plastic deformation.

of the effects of the applied stress on the viscoelastic
properties of the binder.

Tablet defects

Typically there are three main types of tablet defect observed: (1)
pitting; (2) capping; and (3) lamination. *Pitting* refers to the
production of pit marks on the surface of the tablet and is
accredited to insufficient lubricant at the tablet/punch interface or
punches with a roughened surface. This defect may be corrected
by increasing the concentration of lubricant in the formulation or
by altering the mixing conditions (time and rate of mixing),
whereas punches should be polished regularly to prevent such
adhesions. *Capping* and *lamination* (illustrated in Figure 9.19)
refer to mechanical splitting of the tablet. In the former scenario,
the top (cap) of the tablet is fractured whereas in lamination the
fracture may occur within the main body of the tablet.
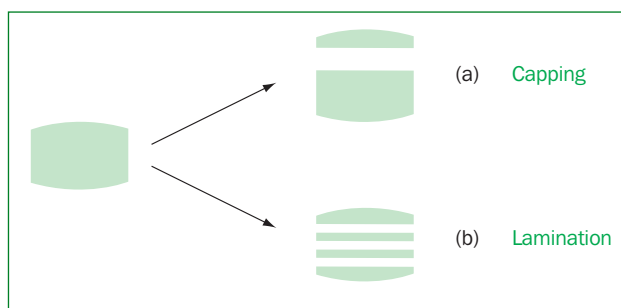
Figure 9.19 Diagram illustrating capping and lamination in tablets.

(a) Capping

(b) Lamination

These two tablet defects occur during the ejection stage of the
manufacturing process and are accredited to stress-induced
fracture. Alteration in the formulation/manufacture to increase
the strength of the tablets may resolve these problems. For
example, the type and concentration of the binder may be altered,
whereas capping and lamination of tablets may occur if the
geometry of the punches is intricate. Whilst increasing the
compression pressure is known to increase the mechanical
strength of tablets, it should be noted that the application of
excessive stresses often results in a reduction in the tablet
strength, which may result in capping and lamination.

Postprocessing effects on tablet properties

Tablets are known to undergo postprocessing alterations in their
physicochemical properties, particularly under defined
conditions of humidity and temperature. Storage may affect the
mechanical properties of the tablet due to the following
processes:

- Adsorption of water on to the tablet surface and within the pores of the tablet may lead to dissolution and then crystallisation of tablet components, which in turn act as solid bridges and hence increase the powder/powder or granule/granule interactions.
- Adsorption of water on to the tablet surface and within the pores of the tablet may induce crystallisation of amorphous excipients or change the polymorphic state of components within the tablet.

Conversely, in the presence of moisture, tablet strength may be decreased due to:
- disruption of the granule/granule or powder/powder bonds due to dissolution of formulation components
- stress relaxation of viscoelastic components. Polymers are viscoelastic and, as such, will undergo stress relaxation as a function of time following removal of the compression stress. This relaxation may be visualised as movement/rearrangement of the polymeric chains, which may, in turn, compromise the mechanical properties of the tablet.

Coating of tablets

At the start of this chapter a description of the various types of commercially available tablets was provided. The reader will have observed that tablets are often formulated with an external coating. A summary of the rationale for tablet coatings is provided below:
- to protect the drug from degradation in the stomach (an enteric coating)
- to prevent drug-induced irritation at a specific site within the gastrointestinal tract, e.g. the stomach for non-steroidal anti-inflammatory drugs
- to provide controlled release of the drug throughout the gastrointestinal tract
- to target drug release to a specific site in the gastrointestinal tract, e.g. the delivery of drug to the colon for the treatment of inflammatory conditions
- to mask the taste of drugs
- to improve the appearance of the tablet.

Examples of the materials that are commonly used as tablet coatings have been described in a earlier section within this chapter.

General description of tablet coating
The main steps involved in the coating of tablets are as follows:
- The tablets (or granules) are placed within the coating apparatus and agitated.

- The coating solution is sprayed on to the surface of the tablets.
- Warm air is passed over the tablets to facilitate removal of the solvent from the adsorbed layer of coating solution on the surface of the tablets.
- When the solvent has evaporated, the tablets will be coated with the solid component of the original coating solution.

Coating formation

Coating solutions are available in two main formulation types: (1) solutions; and (2) emulsions.

Coating solutions

Coating solutions contain the coating material (polymers or sugar), the coating solvent and other excipients that are required to improve the formulation or performance of the tablet coating, e.g. colourants/opacifiers, plasticisers (to render the film flexible). The choice of the solvent/solvent blend is generally performed according to the physicochemical properties of the coating material (i.e. the compatibility of the material with the solvent); however, other considerations include the volatility and the flammability of the solvent. The concentration of the coating material within the solution is also a consideration. Increasing the concentration of coating material within the solvent will reduce the processing time; however, by increasing the concentration of material, the viscosity of the solution may be unacceptably high to achieve the correct spray properties during coating.

Coating emulsions

More recently emulsions have been developed as tablet-coating systems. In these the polymer is dissolved in a volatile organic phase (with plasticiser and colourants/opacifiers, as required) and this is emulsified within an external aqueous phase. The use of coating emulsions has been favoured in recent years due to environmental concerns associated with conventional organic coating solutions.

The initial stage in the coating process involves the deposition and subsequent spreading of the atomised coating solution/emulsion on the surface of the tablet (or granule). To achieve a uniform surface distribution of the coating solution/emulsion on the tablet, consideration of the wetting properties of the solution/emulsion on the surface of the tablet is required. Furthermore the surface rugosity (roughness) and porosity contribute to the tenacity of the interaction initially between the coating solution/emulsion and the tablet but also influence the bond strength between the formed coating and the tablet surface. Following spreading, evaporation of the solvent

initially enables coalescence of the organic droplets, and hence initial film formation on the surface of the tablet. As drying continues, the saturation solubility of the coating material in the solvent is exceeded and the solid coating is formed on the surface of the tablet. It should be noted that contact, spreading, droplet coalescence and solvent evaporation occur almost instantaneously.

Tablet coating in practice

There are several designs of systems that are used in industrial practice to coat tablets (or granules). Examples of these systems are described below.

Pan coater

The pan coating system is generically composed of a metal pan (drum) into which the tablets are placed and that may be rotated at a range of speeds. The coating solution is sprayed on to the surface of the tablets within the pan whilst the drum is rotated. Simultaneously warm air is passed over the surface of the tablets to facilitate the evaporation of the solvent in which the coating material has been dissolved. A schematic diagram of the operation of a basic pan coater is shown in Figure 9.20.

Figure 9.20 Diagram of the operation of the standard pan coater.
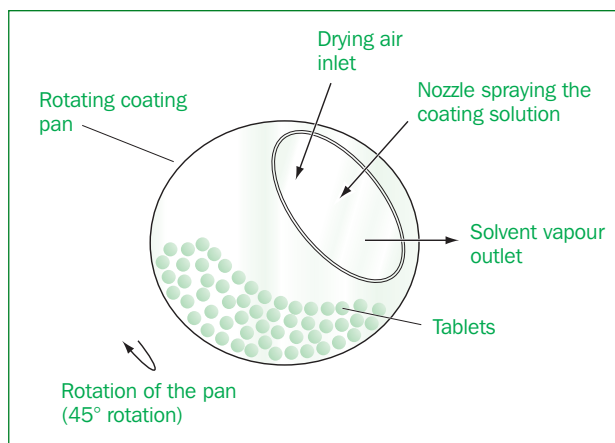
Control of the coating process is obtained by modifying the following parameters:

- rotation rate of the drum/pan
- airflow rate
- temperature of the air
- concentration of sugar/polymer within the coating solution/emulsion.

More recently, pan coaters have been developed in which the pan is perforated (e.g. the Accela-Cota and Hi-Coater systems). In these systems the warmed air is passed into the drum and through the tablet bed before being exhausted (with the solvent from the coating solution) via the perforated drum. In the Driacoater system, the drum is composed of a serious of perforated fins (typically 8 per drum) from which the warmed air is provided. As the drum rotates, the tablets in the tablet bed are mixed by and collected on the fins before being suspended in the warmed air. The tablets are then dropped into the tablet bed and the process is repeated. The warmed air is then exited from the rear of the pan.

Air suspension coaters

Air suspension coaters are highly efficient coating systems in which the coating solution is sprayed on to tablets (or granules) that have been suspended in a positive (warmed) airflow. This ability simultaneously to suspend and coat tablets leads to high coating efficiency. Typically the tablets are initially suspended in the centre of the chamber and then move to the periphery of the chamber before falling to the bottom, at which stage the process is continuously repeated. The coating solution is fed into the fluidisation chamber (usually at the bottom of the chamber) as an atomised spray that has been generated either by passage of the coating solution through a nozzle under high pressure or by passage of the coating solution through a nozzle at low pressure, at which point the solution comes into contact with two high-pressure air streams. A diagrammatic representation of the fluidised-bed coating process is shown in Figure 9.21.
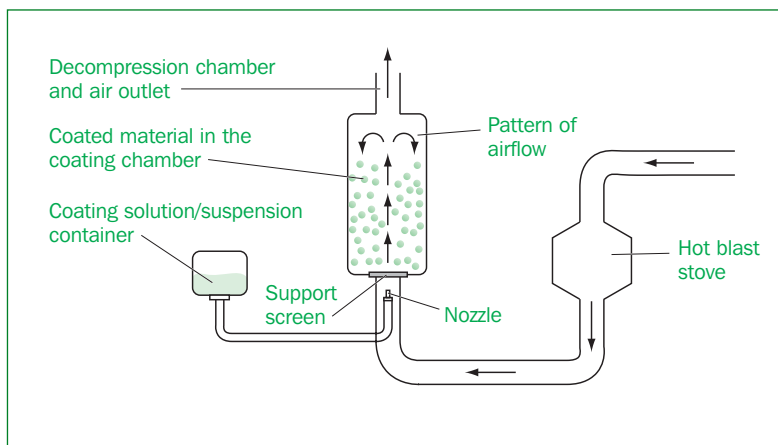
Figure 9.21 Diagrammatic representation of fluidised-bed coating.

Process variables in fluidised air coating

There are several process factors that control both the efficiency of the coating process and the quality of the formed coat. These are: (1) evaporation rate of the solvent; (2) fluidised air volume; (3) specific humidity; and (4) coating spray rate and duration.

Evaporation rate of the solvent

The rate of evaporation of the solvent directly affects both the quality (in particular the mechanical properties) of the tablet coating and the time required to form tablet coating. Whilst it is important to process the coating in the minimum time, increasing the evaporation rate of the solvent decreases the time available for polymer–polymer interactions to occur. Therefore, if solvent evaporation rate is too rapid, the mechanical properties of the films will be compromised due to the adverse effects on polymer interactions. Both the solvent vapour pressure and the process temperature affect the rate of evaporation of a solvent. Therefore, a low process temperature is normally employed for coating solutions/solvents containing a solvent of high vapour pressure, e.g. dichloromethane. Small fluctuations in processing temperature will have greater effects on the quality of the tablet coatings prepared using organic solvents than when processed using an aqueous coating solution.

Fluidised air volume

The fluidised air volume will affect both the velocity of the droplets of coating solutions/emulsions and their fluidised pattern within the coating chamber.

Specific humidity

It is important to control the specific humidity within the warmed air and hence in the coating chamber to ensure that the quality of the tablet coating is optimised. If the relative humidity in the coating chamber is high, evaporative cooling by the solvent may occur. This will, in turn, lower the temperature of the air to below the dew point, resulting in the condensation of water on to the tablet surface. This will interfere with the coating process, resulting in poor adhesion of hydrophobic coatings to the tablet surface and visual imperfections in the formed coating. Therefore, control (but not elimination) of the relative humidity within the coating process is required. The presence of humidity within the coating chamber may be useful in dispelling static electricity that may occur after the coating process has been completed.

Coating spray rate and duration

The coating spray rate is controlled within the coating process and is selected according to the solubility of the coating solvent

in the air volume and the viscosity of the atomised droplets. In addition, it is important to ensure droplet integrity (i.e. minimise droplet aggregation) during the process. It should be noted that excessive spray rates will produce coatings that exhibit poor adhesion to the tablet surface.

Typically the coating process will involve several passes through the coating apparatus. Therefore one method by which the thickness of the coating on the tablet may be modified is to increase the time spent within the coating chamber. Alternatively, the concentration of coating material may be increased within the coating solution. The viscosity of the solution must be considered to ensure that the increased viscosity does not compromise the atomisation process, and specifically the droplet size.

Problems associated with tablet coatings

There are several problems associated with tablet coatings, including: (1) poor adhesion of the coating to the tablet; (2) tablet abrasion; (3) filling tablet markings; (4) rough surface; (5) formation of cracks in the coating; and (6) variations in the colour of the coating.

Poor adhesion of the coating to the tablet

This phenomenon may be due to:

- high relative humidity within the coating chamber when coating tablets using an organic solvent system
- high coating spray rate
- concentration of polymer in the coating solution/emulsion is too low
- temperature of air is too low, resulting in a slow rate of solvent evaporation (particularly valid for coating systems that employ solvents of low vapour pressure, e.g. water)
- air fluidisation rate or pan rotation rate is too slow
- the tablet substrate has minimal curvature. Typically curved surfaces are easier to coat than flat surfaces.

Tablet abrasion

The coating process involves exposing the tablets to shearing stresses that are generated as a result of collisions with other tablets and also with the walls of the coating chamber. This may result in damage to the tablet surface. This problem may occur due to: (1) inappropriate tablet hardness; (2) irregular tablet shape; (3) tablet bed is too heavy during coating; and (4) the speed of rotation of the pan or the air fluidisation rate is excessive.

Inappropriate tablet hardness

Tablet hardness may be improved by increasing the compaction pressure or binder concentration (in wet granulation). Generally

the hardness of tablets produced by wet granulation is greater
than by other methods and therefore the tablets produced by this
method are generally suitable for coating

Irregular tablet shape
Irregular tablet shapes are more prone to abrasion than regular
tablet shapes. This problem may therefore be overcome by
changing the tablet shape.

Tablet bed is too heavy during coating
This leads to increased tablet–tablet contact. To correct this
problem the loading of tablets within the coating chamber is
reduced.

The speed of rotation of the pan or the air fluidisation rate is excessive
By reducing the speed of pan rotation or air fluidisation rate,
tablet–tablet contact is decreased.

Filling tablet markings
Manufacturers may wish to identify their product with a
particular mark/name (performed by using a tablet punch that has
been embossed with the specified mark). If the coating conditions
are unsuitable, the coating will excessively deposit within the
mark/name and, in so doing, the marking will be partially
obscured. This may occur due to:
- the use of deep markings
- use of an excessive volume of coating solution
- air temperature is too low
- pan rotation speed/fluidisation flow rate is too low.

Rough surface
One of the major problems of tablet coating is the production of
tablets that exhibit a rough surface. This phenomenon is often
associated with drying of the coating droplets prior to reaching
the surface of the tablet. To correct this problem the spray rate
may be increased and the inlet air temperature decreased.

Formation of cracks in the coating
The formation of cracks in tablet coatings is principally due to the
use of an inappropriate coating formulation. The reader is aware
that plasticisers are employed to lower the glass transition
temperature of polymer coatings. This in turn renders the film
more flexible and less brittle. Therefore cracking in polymer
coatings may indicate that either the plasticiser concentration
should be increased or, alternatively, a different plasticiser that is
more compatible with the polymer chosen for the coating should

be considered. In certain situations cracking of polymer coats may occur due to the use of a polymer that has a low stress resistance and is therefore prone to stress failure. To rectify this situation either the molecular weight of the polymer under investigation should be increased or, alternatively, a different polymer should be used that has a greater resistance to the applied stress (i.e. an increased ultimate tensile strength).

Variations in the colour of the coating

Tablets that have been coated with a polymer containing a colourant should show uniform colour. Variations in the colour of a tablet coating may be due to:

- improper mixing of the colour within the coating formulation
- uneven coating process, resulting in regional differences in the thickness of the applied coating
- migration of coloured components within the tablet core into the coating. This may be resolved by the use of a coloured coating that will mask the effects of the migration or by the use of a coating in which the components within the table core are insoluble.

Multiple choice questions

1. **Concerning roller compaction as a method for tablet manufacture, which of the following statements are true?**
   a. Roller compactions requires the application of heat.
   b. Roller compaction is suitable for the manufacture of tablets containing high-potency (i.e. low-dose) drugs.
   c. Roller compaction produces soft tablets.
   d. Roller compaction requires only conventional excipients.

2. **Concerning direct compression as a method for tablet manufacture, which of the following statements are true?**
   a. It is a simple process, involving a lower number of unit operations in comparison to wet granulation.
   b. The morphology of particles is not important in direct compression.
   c. Direct compression employs spray-dried excipients only.
   d. Coloured tablets may be easily produced by direct compression.

3. **Concerning the pan coating of tablets, which of the following statements are true?**
   a. The coating solution may be aqueous or organic in nature.
   b. Hydrophilic polymers may be used to coat tablets using this technique.

c. The mass of tablets in the pan affects the quality of the coating.
d. The speed of rotation of the pan does not affect the quality of the coating.

4. **Concerning wet granulation, which of the following statements are true?**
a. Alcohol may be employed as a granulating fluid.
b. Only spray-dried excipients may be used.
c. Wet granulation may be unsuitable for thermolabile drugs or drugs that are prone to hydrolysis.
d. Tablets produced by this method are prone to capping.

5. **Concerning pharmaceutical tablets, which of the following statements are true?**
a. Pharmaceutical tablets may include a disintegrant to improve tablet strength.
b. Pharmaceutical tablets may include magnesium stearate to prevent tablet adhesion to punches.
c. Pharmaceutical tablets include diluents such as colloidal silicone dioxide.
d. Pharmaceutical tablets may involve a granulation stage in order to prevent segregation of the powder mix.

6. **The dissolution rate of drugs from tablets may be increased by:**
a. Increasing the compression stress during tableting.
b. Increasing the concentration of lubricant.
c. Increasing the concentration of disintegrant.
d. Film-coating tablets with ethylcellulose.

7. **Concerning film coatings, which of the following statements are true?**
a. Film coatings may enhance the dissolution rate of therapeutic agents in the gastrointestinal tract.
b. Enteric film coatings should dissolve in the stomach.
c. Aqueous film coatings, e.g. hydroxypropylcellulose, may be employed to target drug release at the colon.
d. Film coatings may be employed to mask the taste of unpalatable therapeutic agents.

8. **Concerning pan coating of tablets, which of the following statements are true?**
a. Only organic polymer solutions may be employed to coat tablets using the pan coating method.
b. Pan coating is a more effective method than fluidised bed coating.

c. The quality of the tablet coating is improved by increasing the temperature of the inlet air.

d. The quality of the tablet coating is affected by the rotation speed of the pan.

9. **The following excipients are employed in tablets for the stated reasons:**

a. Lactose – as a diluent.

b. Sucrose – as a binder.

c. Hydroxypropylmethylcellulose – as an enteric coating.

d. Polyethylene glycol – as a lubricant.

10. **Regarding the various types of tablets, which of the following statements are true?**

a. Enteric-coated tablets release the therapeutic agent within the small intestine but not in the stomach.

b. Effervescent tablets are used to enhance the dissolution rate and hence absorption rate of poorly soluble drugs.

c. Sugar-coated tablets are employed to reduce drug degradation within the stomach.

d. Film-coated tablets may be used to target drug release to the colon.

chapter 10
Solid-dosage forms 2: capsules

Overview

In this chapter the following points will be discussed:
- an overview/description of the various types of capsules, their manufacture and the rationale for their use
- formulation strategies for hard and soft capsules
- an overview/description of the methods of manufacture of hard and soft capsules
- the advantages and disadvantages of the use of capsules
- a detailed description of the excipients used in the formulation and manufacture of capsules.

Introduction

Capsules are solid-dosage forms that are most commonly composed of gelatin and are designed to contain a drug-containing formulation. Two types of capsule are available – hard and soft gelatin capsules. These differ in both their mechanical properties and in capsule design. Hard gelatin capsules are less flexible and are composed of two pieces, termed the capsule and the body, whereas soft gelatin capsules are more flexible and are composed of a one-piece capsule shell. A wide range of formulation types may be included within the interior of the capsule. For example, powders, tablets, semisolids and non-aqueous liquids/gels may be filled into *hard* capsules, with powders being the most common formulation option. Soft gelatin capsules are usually filled with non-aqueous liquids containing the therapeutic agent either dispersed or dissolved within this carrier. Capsules offer the pharmaceutical scientist with an alternative method for the formulation of solid-dosage forms.

KeyPoints

- Capsules are solid-dosage forms that are available in two types – hard (two-piece) or soft (one-piece). The major component of the capsule shell is gelatin, although other polymers have been investigated as capsules (e.g. starch, hydroxypropylmethyl-cellulose).
- Capsules may be filled with a range of formulation types, including powders, tablets, semisolids (hard capsules) and liquids (soft capsules).
- Akin to tablets, the formulation that is included within the capsule contains several components, each of which is present to facilitate the manufacture or to control the biological performance of the dosage form.

Advantages and disadvantages of capsule formulations

Advantages

The formulation of capsules may be preferred for several reasons:

- The use of capsules avoids many unit operations that are associated with the manufacture of tablets, e.g. compression, granulation, drying.
- Capsules (generally soft gelatin capsules) may be formulated to increase the oral bioavailability of poorly soluble therapeutic agents. This is particularly the case when formulated as a liquid-filled hard gelatin or soft gelatin capsule.
- Capsules are a convenient method by which liquids may be orally administered to patients as a unit dosage form.
- Capsules are difficult to counterfeit.
- The stability of therapeutic agents may be improved in a capsule formulation.
- Capsules are a convenient means of formulating substances of abuse, e.g. temazepam.

Disadvantages

The disadvantages of capsule formulations include:

- the requirement for specialised manufacturing equipment
- potential stability problems associated with capsules containing liquid fills
- problems regarding the homogeneity of fill weight and content may be associated with capsule formulations.

Materials and manufacture of capsules

Capsules are primarily (but not exclusively) manufactured using gelatin; however, the suitability of other materials, e.g. hydroxy-propylmethylcellulose and starch, has been investigated as suitable replacements. Gelatin is a mixture of proteins that is extracted from animal collagen (derived from animal skins, sinews and/or bovine bones) by either partial acid or partial alkaline hydrolysis. From these processes two types of gelatin are obtained, termed type A and type B. Type A is obtained by an acid treatment of pig skin (HCl, H_2SO_4, H_2SO_3 or H_3PO_4, pH 1–3 for approximately 1 day) whereas type B is obtained using an alkaline treatment of demineralised bones (immersion in a calcium hydroxide slurry for 1–3 months), following which gelatin is extracted using a series of hot-water washes (of successive increasing temperatures). The gelatin solutions are then cooled to form a gel; subsequent evaporation of water results in the production of dried gelatin. The isoelectric points

of type A and type B gelatin differ (between 7 and 9 and between 4.7 and 5.3, respectively), resulting in differing solubilities as a function of pH. Typically the molecular weight range for gelatin is 15 000–250 000. Whilst the two types of gelatin are often used independently, mixtures of the two types are commercially available. The manufacture of the two types of gelatin is summarised in Figure 10.1.

Figure 10.1 Schematic for the production of gelatin. (Adapted from www.geafiltration.com.)
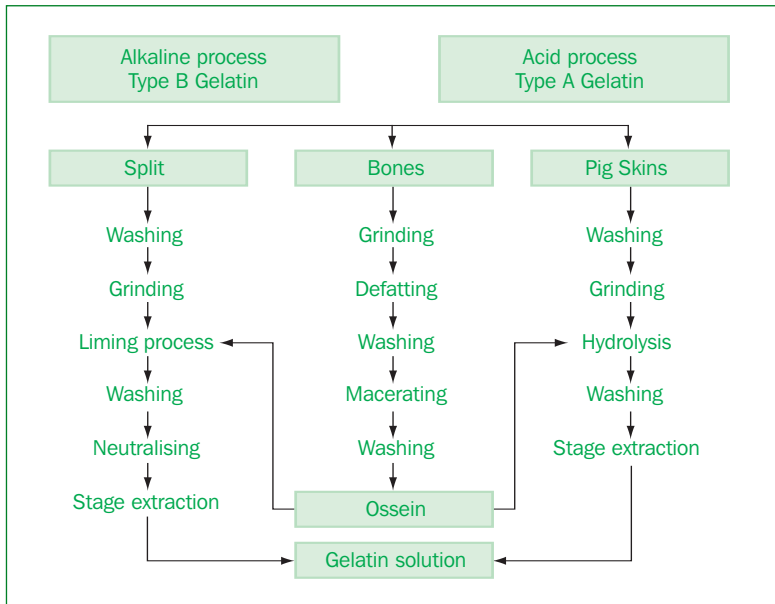
The grade of gelatin is defined by the *bloom strength*, which is defined as the weight (in grams) required to depress a plunger (of defined diameter, 12.7 mm) to a defined depth (4 mm) within an aged gelatin gel (6.66% w/w in water).

The use of gelatin as a capsule material is principally due to the excellent physicochemical and biological properties of this material, including:

- non-toxic material, being used widely as a component of foods. More recently, the production of gelatin using bovine sources has received considerable attention due to the possible transmission of bovine spongiform encephalopathy (BSE). Therefore, if gelatin has been manufactured from bovine sources, it is important that this has been produced from countries in which the incidence of BSE is low – termed grades 1 (highly unlikely) or 2 (unlikely but not excluded)

- soluble in biological fluids at room temperature (note: gelatin capsules do not dissolve but swell when immersed in an aqueous solution < 30°C)
- excellent mechanical properties, most notably exhibiting good film, and hence capsule-forming properties
- excellent rheological properties at elevated temperatures. At 50°C, gelatin acts as a mobile liquid (termed a sol), thereby enabling the production of capsules by dip processing (see later)
- undergoes a sol–gel transition at relatively low temperatures. Therefore, gelatin is readily converted to the rigid (gel) state by allowing warmed solutions of this material to cool.

Description and manufacture of hard gelatin capsules

Hard gelatin capsules are composed of two halves, termed the cap and the body. During manufacture of the dosage form, the formulation is filled into the body (using a range of different mechanical techniques) and the cap is pushed into place. The two halves of the capsule are joined, the cap overlapping with the body. Due to the tight fit between the two halves, separation of the cap and body does not normally occur under normal storage conditions or in clinical use. However, other capsule designs are available in which the two halves form a mechanical seal via indentations on the outside of the body and/or inside the cap (Figure 10.2).

Figure 10.2 Diagram of the Conisnap capsule, illustrating the locking mechanism between the two capsule halves. (Adapted from http://www.capsugel.com/products.)
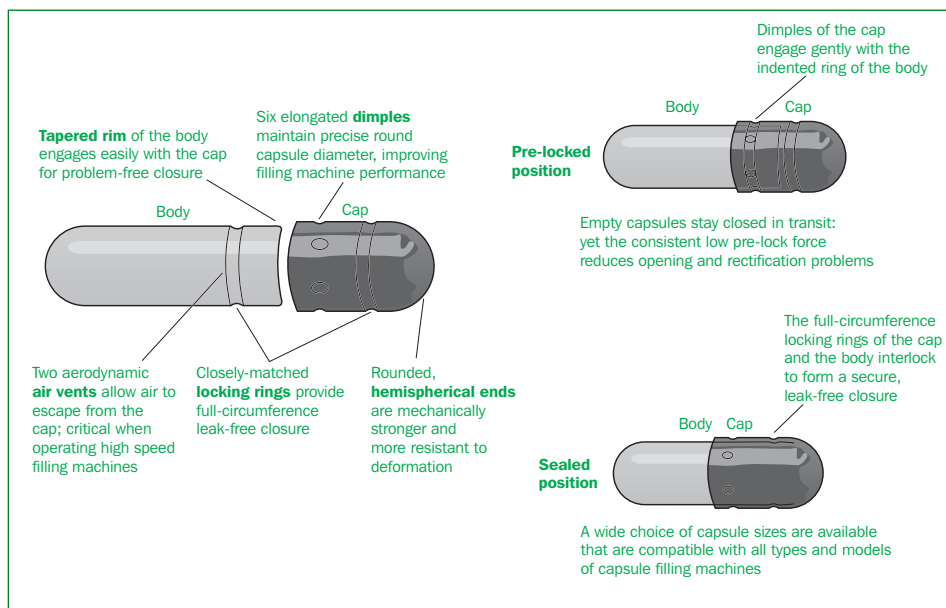
Sealing of hard gelatin capsules may also be performed using two further methods:

1. *Gelatin band sealing.* In this method a dilute solution of gelatin is applied to the centre of the capsule (between the two halves) which, once dried, produces a hermetic seal.

2. *Hydroalcoholic solvent seal.* A hydroalcoholic solution (1:1 water/ethanol) is applied to the centre of the capsule (between the two halves). This softens the capsule and, following heating to 45°C, the interface fuses to produce a seal.

The manufacture of hard gelatin capsules is performed using a dip-coating method. The various stages of this are as follows.

Preparation of the gelatin solution

Initially a concentrated solution of gelatin is prepared (circa 35–40% w/w) in demineralised hot water with stirring. Following dissolution of the polymer a vacuum is then applied to the mixing vessel to remove entrapped air.

Incorporation of other constituents of the capsule

Other excipients may be included within the heated gelatin solution, e.g.:

- *Colourants.* Most frequently hard gelatin capsules are coloured to enhance the aesthetic properties and also to act as a means of identifying the product. For this purpose the chosen dye is added at the required concentration into the heated gelatin solution.

- *Wetting agents/lubricants.* It is permissible to formulate hard gelatin capsules with sodium lauryl sulphate ($\leq 0.15\%$ w/w) as an included component to increase the wetting properties of the capsule shell following contact with an aqueous solution (and hence enhance the dissolution properties of the formulation contained within the capsule). Furthermore, the presence of sodium lauryl sulphate in the heated gelatin solution will also enhance the wetting of the (hydrophilic) gelatin solution on the metal pins during the manufacturing process. This leads to the production of gelatin capsules of uniform thickness.

Control of the viscosity of the gelatin solution

Following the inclusion of all components within the heated gelatin solution, the viscosity of the solution is then modified (reduced). Control of the viscosity is important as this regulates the thickness of the capsule (generally circa 100 μm). As the viscosity is lowered the capsule thickness will decrease.

Dip-coating the gelatin solution on to metal pins (moulds)

The machine used to manufacture capsules consists of two sets of bars, each containing a series of pins (aligned in columnar formation) that have been lubricated prior to use. The pins (one set for the production of the cap and one for the body of the capsule) are dipped into a pan that contains the heated gelatin solution (maintained at 35–45°C). Following adsorption of the gelatin solution on to the surface of the pins, the bar containing the pins is removed and rotated. The reduction in the temperature of the gelatin and the rotating action cause the gelatin to gel on the surface of the pins in a uniform manner. The pins are then advanced through a series of air driers in which air of the required humidity is passed across the surface of the gelatin film. Following this, the (hardened) capsules are removed from the pins and cut to the appropriate size prior to joining the two halves of the final capsule.

Properties of the final capsule

The final capsules should exhibit a water content of 13–16% w/w. This is an important consideration in light of the effects of water content on the mechanical (and hence in-use) properties of the capsules. Water acts as a plasticiser for gelatin to ensure that the mechanical properties of the capsule are sufficiently robust so that the capsule does not either crack or permanently deform during manufacture, handling or storage. If the water content is lower than the above specification, the capsule shell will become brittle and will crack when exposed to the appropriate stress. Conversely, if the water content is excessive, the capsule will undergo plastic flow upon exposure to stress and will lose shape. It is therefore important that the desired water concentration in the capsule is achieved within the drying phase of the manufacturing process. It is additionally preferable that capsules should be stored under conditions that do not adversely affect this parameter.

A wide range of capsule sizes is available that can accommodate fill volumes between 0.20 and 0.67 ml.

The production of coloured capsules requires the addition of the appropriate colour and opacifying agent (e.g. titanium dioxide) in the heated gelatin solution during the capsule-manufacturing process.

Formulation considerations for hard gelatin capsules

The fill for hard gelatin capsules may be formulated as either a powder (or granule) containing the required drug or as a liquid

into which the drug is either dispersed or dissolved. The formulation considerations for both of these strategies are individually described below.

Solid capsule fills

As the reader will observe, many of the excipients used for the formulation of solid capsule fills for capsules are commonly used in tablet formulations for similar purposes. Therefore, to prevent repetition, only the basic details of the excipients (including their properties and rationale for use) will be described.

General properties of solid fills

There are several general properties of powders that should be recognised when formulating these as capsule fills, as follows:

- The particle size distributions of the various components of the powder mix (including the therapeutic agent) should be similar both to ensure homogeneous mixing and to minimise segregation.
- It is preferable that the particle size distribution of the powder blend is both monomodal and exhibits low polydispersity (low standard deviation) to ensure predictable and reproducible flow during the filling process. Conversely, multimodal and polydisperse powder mixes will exhibit a tendency to segregate, with resultant problems associated with the homogeneity of the mix and a gradual increase in fill mass as a function of filling time.
- Problems may occur during the filling of particles with an irregular shape (e.g. needle shape).

Filling of hard gelatin capsules

There are two main methods by which powders are filled into hard gelatin capsules, termed *dependent* and *independent* methods. The design and operation of these are described below:

- *Dependent method (dosing system).* In this method the lower half of the capsule is placed into slots that are located within a revolving turntable. The upper part of the capsule is also housed in a similar turntable. The turntable containing the lower half of the capsule (which may be rotated at a range of speeds) is rotated under a hopper that contains the powder formulation and, as a result, the powder falls into the capsule. The flow of the powder through the hopper and the homogeneity of the powder mix are maintained by the circular movement of an auger. At the end of the operation the two capsule halves are brought together to form the finished dosage form (Figure 10.3).

    The mass of powder that is dispensed into each capsule is dependent on the length of time that the hopper spends above

Figure 10.3 Diagrammatic representations of the design and operation of the dependent dosing system for filling hard gelatin capsules with powders.

Dependent dosing system (side view)

Rotating turntable (vertical view)

Auger

Hopper

Spaces for location of the lower half of the capsule shell

Rotating turntable (dark shading represents powder-filled capsules)

(a)   (b)
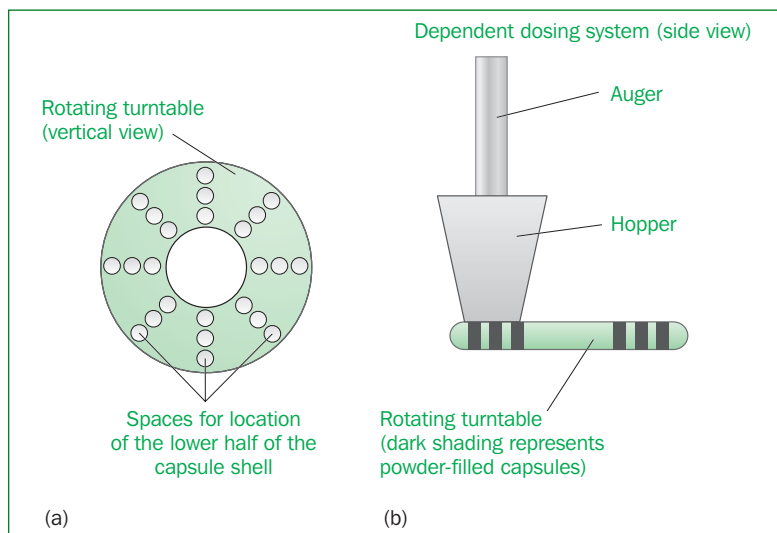
the capsule (which is itself dependent on the speed of rotation of the turntable). At the end of the filling process the filled capsules are removed from the turntable.

- *Independent method (dosing system).* The independent method of capsule-filling involves the physical transfer of a plug of powder from the mixed powder into the capsule. In this method a tube, which contains a spring-loaded piston, is depressed into a powder bed enabling a volume (plug) of powder to enter the tube. The settings of the spring-loaded piston control the volume of powder that enters the tube. If required, the bonding between the particles within the plug may be enhanced by the application of a compression pressure. The tube (containing the plug of powder) is then elevated out of the powder bed, rotated and located above the lower half of the capsule and the plug of powder is dispensed into the capsule by the depression of the piston.

The filling of liquids/semisolid formulations into hard gelatin capsules is most frequently performed using a volumetric dosing system. If the rheological properties of the fill are particularly responsive to temperature, filling is performed at temperatures in which the fill is in the liquid state. It should be noted that hard gelatin capsules containing liquid/semisolid fills may be prone to leakage and, therefore, treatment of the capsule shell at the join between the body and the cap is required to prevent this problem.

Formulation of the fill of hard gelatin capsules

As detailed above, there are three main classes of fill that are employed in hard gelatin capsules, namely powders and liquids/semisolids. The formulation considerations for these categories are considered separately in the following sections.

Powders

In general, powder formulations for inclusion within hard gelatin capsules should exhibit the following properties:

- *Homogeneity of mix* (described above).
- *Good flow properties.* Filling of the capsules requires the reproducible flow of the powder from the powder bed, through the filling apparatus and into the capsule. As the filling is performed according to volume, it is important that the packing of the particles is also reproducible, as variations in this property will result in differences in the mass of powder filled into each capsule. Acceptable powder flow also requires that clumping of the powders does not occur. Finally, as the filling process may require plug formation (within the dossator), the powder bed should be compressible. Typically the flow properties and the packing properties of the powders are assessed using the following techniques:
- *Angle of repose (f).* In this method powder is passed through a funnel until the angle of inclination of the powder is too small to overcome cohesive forces between the particles. The angle of repose is the angle that the powder makes with the horizontal plane. The technique provides an indication of the flow properties of the powder and, specifically, is a measure of cohesion within the powder mass. The tangent of the angle of repose is frequently referred to as a measure of the internal friction of the powder bed.

  If the measured angle exceeds 50°, the flow properties of the powder are poor. Typically an angle of circa 25° is indicative of a powder that would be expected to exhibit suitable flow for manufacturing process. Powder that exhibit high angles of repose will require the addition of a glidant to reduce particle–particle cohesion.
- *Torque rheometry.* Torque rheometry is a rheological technique in which a stress is applied to the powder bed (by means of a mixing head) and the subsequent deformation (rate of shear) of the powder bed is determined. Powder beds that should demonstrate high cohesion will require greater shearing stresses to initiate and maintain flow.

  This technique is often applied to the characterisation of the wet granulation process and, in particular, to examine the mixing requirements and end-point of the granulation process.

- *Tap density.* Tap density measures the volume occupied by the powder bed, both before (r_o) and following (r_f) the application of a consolidation stress (generally shaking at a defined rate and for a defined period). The ratio of the density of the powder bed before to that after shaking is referred to as a Hausner ratio. Generally a Hausner ratio of circa 1.2 is acceptable whereas, when the Hausner ratio exceeds 1.6, the powder may be problematic to fill into capsules due to the unnecessarily high cohesive interactions between the particles. These cohesive properties may result in erratic filling due to arching and related phenomena.

■ *Compatibility between the formulation components and between the formulation components and the capsule.* The excipients that are employed in the formulation of powder fills for hard gelatin capsules are similar to those employed for the formulation and manufacture of tablets. Accordingly the main physicochemical properties of these excipients have been described in Chapter 9. In particular the following excipients are used for the formulation of powder fills:

– *Diluents.* Diluents are employed to increase the working mass of the powder bed and thereby enhance the reproducibility of the filling process. In addition the diluents may offer additional properties, most notably their flow properties and their ability to undergo compression. Examples of excipients that are employed in the formulation of powder fills include:
  - lactose (monohydrate)
  - maize starch
  - microcrystalline cellulose.

– *Lubricants/glidants.* Lubricants are used to reduce the interaction of the powders with the metal dossator and/or other metal components of the filling machine, whereas glidants are used to lower the interparticle attraction, thereby reducing clumping and aiding powder flow. The types of lubricants and glidants used in the formulation of powder fill capsules are identical to those employed for tablet manufacture and include:
  - magnesium stearate (and other stearates) as a lubricant
  - colloidal silicon dioxide as a glidant.

– *Disintegrants.* Disintegrants are employed (as before) to break up the powder mass following release into the stomach. Examples that are used for this purpose include:
  - maize starch
  - microcrystalline cellulose
  - sodium starch glycolate

- crospovidone
- croscarmellose.
– *Surface-active agents.* Surface-active agents are employed within powder fill hard gelatin capsules to increase the wetting properties of the powder bed following release within the gastrointestinal tract. Their inclusion is particularly important if the formulation contains significant concentrations of hydrophobic components, e.g. lubricants and glidants. To achieve rapid drug release the powder fill should be readily dispersed within the gastrointestinal contents, a feature that is enhanced by the presence of surface-active agents of high hydrophile–lipophile balance, e.g. sodium lauryl sulphate. In addition, the presence of sodium lauryl sulphate within the hard gelatin capsule shell is allowed and acts similarly to enhance the uptake of fluid into the capsule shell.

Liquids/semisolid fills for hard gelatin capsules
Liquid/semisolid fills for hard gelatin capsules may be subdivided into various categories:
- *Lipophilic liquids/oils containing dissolved or dispersed therapeutic agent.* Examples of the types of liquids that are commonly used in this category include:
– vegetable oils (e.g. sunflower, arachis, olive)
– fatty acid esters (e.g. glyceryl monostearate).
- *Water-miscible liquids containing dissolved/dispersed therapeutic agent.* Examples of the types of liquids that are commonly used in this category include:
– polyethylene glycols (PEGs) that are solid at room temperature but will liquefy upon heating (e.g. higher-molecular-weight PEGs)
– liquid polyoxyethylene-polyoxypropylene block co-polymers (Pluronics).

A major concern in the choice and hence formulation of solvents for liquid fill formulations is the effect of the formulation on the stability of the capsule. As stated previously, the equilibrium moisture content of hard gelatin capsules should be 13–16% to ensure that the capsule exhibits the optimal mechanical properties. Solvents that are hygroscopic when filled into hard gelatin capsules will enhance moisture uptake into and result in splitting of the capsule shell. The moisture uptake of lipophilic solvents, solid PEGs and Pluronics is low and therefore these solvents are preferred for liquid fill formulations for hard gelatin capsules.

To stabilise the liquid fill formulations for hard gelatin capsules, other excipients will be required, e.g.

- *Surface-active agents.* These are included in liquid fills for hard gelatin capsules to:
  - solubilise the therapeutic agent within the solvent
  - stabilise the suspended therapeutic agent
  - enhance the dissolution of poorly water-soluble therapeutic agents within the gastrointestinal tract.
- *Viscosity-modifying agents.* These are included in liquid fills for hard gelatin capsules to:
  - stabilise the suspended therapeutic agent
  - modify the viscosity of the formulation to optimise filling of the capsule. Typically, viscosity values within the range of 0.1–25 Pa/s are considered to be acceptable as liquid fills for hard gelatin capsules. If the viscosity is lower than this range, there will be a loss in the capsule contents due to splashing of the liquid from the capsule during filling. Conversely, liquids of viscosities greater than 25 Pa/s may cause filling problems due to the inability of pumps to fill liquids of this (high) viscosity reproducibly. The coefficient of variation in the volume of liquid fills of high viscosity (> 25 Pa/s) will exceed 0.03.
- stabilisers (e.g. antioxidants, colours)

As the reader will observe, the formulation of the above categories has been addressed in previous chapters. Therefore, for the purpose of this chapter, only the points relevant to the formulation of hard gelatin capsules have been considered.

Tips

Many of the excipients that are used in the formulation of the powder fill of hard gelatin capsules are also used in the formulation of tablets.

Water cannot be used as a liquid fill for hard gelatin capsules. Fill materials for hard gelatin capsules are non-aqueous in nature.

Soft gelatin capsules

Soft gelatin capsules are capsules in which the mechanical properties of gelatin have been manipulated by the addition of a plasticiser (most notably glycerol or other polyhydric alcohols, e.g. sorbitol), resulting in a more flexible capsule. The popularity of soft gelatin capsules has increased in recent years due to the ability to formulate liquid-based formulations that provide a greater C_{max} than tablet formulations. This has particular applications in the treatment of acute conditions, e.g. pain.

A wide range of shapes of soft gelatin capsule is available, including round, oval, oblong, tubular and various other miscellaneous shapes. The various components of the soft gelatin capsule shell are as follows: (1) gelatin; (2) plasticising agents; (3) water; and (4) miscellaneous excipients.

Gelatin

Typically type B (alkali-processed) gelatin is used; however, type A (acid-processed) may also be employed for the manufacture of soft gelatin capsules.

Plasticising agents

As detailed previously, the mechanical properties of the soft gelatin capsule are controlled by the inclusion (and concentration) of plasticisers. For this purpose polyhydric alcohols, principally glycerol or sorbitol or mixtures of these, are used. The concentration of plasticiser is generally 20–30% w/w of the wet mass (refer to the manufacturing method below). This concentration range is an important determinant of the mechanical properties of the soft gelatin capsule. Concentrations in excess of 30% w/w will result in the capsule being too flexible and tacky whereas capsules in which the concentration of plasticiser is below 20% w/w will be too brittle. In both scenarios splitting of the capsule shell may occur either on storage or during use. The hardness of the finished (dried) soft gelatin capsule is specifically defined by the ratio of the plasticiser to gelatin, with ratios of 0.4:1.0 and 0.8:1.0 offering extremes of the typically mechanical properties (hard and soft, respectively).

Water

Water is required both during the manufacturing process (to facilitate manufacture) and in the finished product to ensure that the capsule is flexible. Initially during manufacture of the soft gelatin capsules the concentration of water is between 30 and 40% w/w of the wet mass; however, following formation of the capsule, the product is exposed to controlled drying, resulting in a final capsule water content of 5–8% w/w. If the capsule is overdried a brittle product will result.

Miscellaneous excipients

As is the case for hard gelatin capsules, soft gelatin capsules may be coloured or opaque, the chosen colour(s)/opacifiers being added during the manufacturing process. Titanium dioxide is primarily used as an opacifier for capsules. In addition, if required, flavouring agents may be added to the capsule shell.

Manufacture of soft gelatin capsules

The original manufacturing method for soft gelatin capsules was originally patented by RP Scherer in 1933. Whilst variations of this patent have been published, the main principles of the original Scherer method are still employed for the manufacture of soft gelatin capsules. The main features of the method are illustrated in Figure 10.4.

Figure 10.4 Illustration of the manufacturing method for soft gelatin capsules. (Adapted from http://www.madehow.com/.)
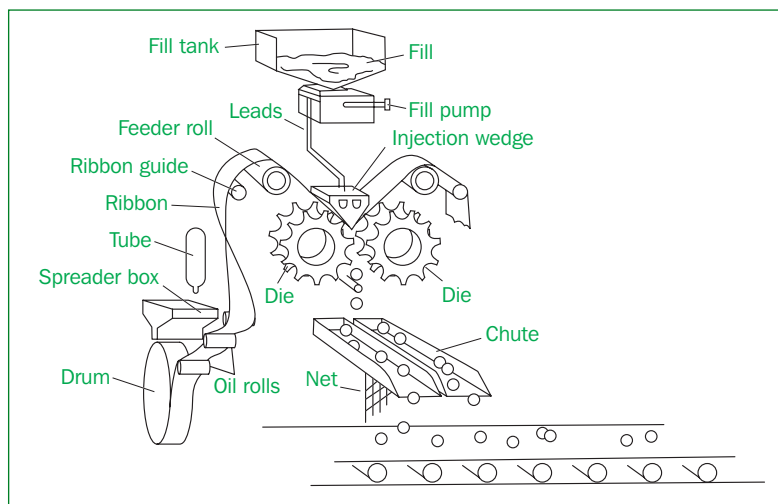
In this method the wet mass formulation is initially prepared containing gelatin, plasticiser(s), water and other excipients, as required. Following this the gelatin solution is fed on to two drums via a spreader box, at which stage ribbons of gelatin are produced. The two sets of ribbons are then fed between two rotary dies (generally lubricated with mineral oil) to form pockets whilst, simultaneously, a metered volume of the capsule fill material is dispensed into the forming pocket. The two halves of capsule (containing the fill material) are sealed by the application of heat (37–40°C) and pressure, detached from the gelatin ribbon and collected. Following collection, the capsules are washed to remove any lubricant (mineral oil) from the surface of the capsule, dried to remove circa 60–70% w/w water before being allowed to equilibrate under defined conditions of humidity (20–30% relative humidity) and temperature (21–24°C). Following equilibration, the water content within the capsule shell will be at the required level. It should be noted that other related methods (e.g. the reciprocating die process) are available for the production of soft gelatin capsules; however, a full description of these is beyond the scope of this text.

Formulation of the fill of soft gelatin capsules

As the reader will have observed from the description of the manufacture of soft gelatin capsules, the fill material for soft gelatin capsules is primarily liquid-based (although powder filling equipment is available). The therapeutic agent may be dissolved or dispersed within the fill material. There are several categories of fill material: (1) lipophilic liquids; (2) self-emulsifying systems; and (3) water-miscible liquids.

Lipophilic liquids

Lipophilic liquids are commonly used as fill materials for soft gelatin capsules and incorporate both vegetable oils (e.g. soyabean oil) and fatty acid esters. Only a limited number of therapeutic agents are soluble in these materials and therefore, when used as a fill material, the formulation will either require the inclusion of co-solvents and/or surface-active agents (see below) or, alternatively, the fill may be formulated as a suspension (requiring the inclusion of viscosity-modifying agents and/or surfactants).

Self-emulsifying systems

Self-emulsifying systems are lipophilic liquids that contain a non-ionic emulsifying agent (e.g. the Tween series). Following release into the gastrointestinal tract, the fill material rapidly emulsifies into small droplets (with high surface area) and, in so doing, enhances the dissolution and hence absorption of the therapeutic agent.

Water-miscible liquids

Liquids in this category include high-molecular-weight alcohols, e.g. PEG 400, PEG 600, non-ionic surface-active agents (e.g. Tweens) and Pluronics (polyoxyethylene-polyoxypropylene block co-polymers). Small concentrations of ethanol and water (generally less than 10%) may be tolerated within soft gelatin capsules. As before, the selected therapeutic agent may be dissolved or, if the solubility is exceeded, dispersed within the solvent, the latter necessitating the addition of surface-active or viscosity-modifying agents to stabilise the formulation.

As required, additional formulation excipients will be required in the above liquid systems: the details of these have been outlined in previous chapters. Examples of the categories of excipient include:

Tips

Liquids are used as the fill materials for soft gelatin capsules; however, water is not employed as a fill material due to the detrimental effects on the physical stability of the capsule.

Soft gelatin capsules may be formulated to enhance the rate of dissolution of poorly soluble therapeutic agents within the gastrointestinal tract.

Drugs of abuse are often formulated as soft gelatin capsules.

- viscosity-modifying agents
- surface-active agents
- colours
- co-solvents (and other solubilising agents).

Multiple choice questions

1. **Regarding capsules, which of the following statements are true?**
a. Capsules are more straightforward to manufacture than tablets.
b. Capsules may be formulated to increase the bioavailability of poorly soluble drugs.
c. Capsules may be coloured to aid identification.
d. The stability of therapeutic agents when formulated as capsules is always greater than that of tablets.

2. **Concerning hard gelatin capsules, which of the following statements are true?**
a. Gelatin is a polysaccharide that is derived from animal sources.
b. There are two types of gelatin, each exhibiting different isoelectric points.
c. The grade of gelatin is defined by the bloom strength.
d. Gelatin is freely soluble in water at room temperature.

3. **Regarding hard gelatin capsules, which of the following statements are true?**
a. Hard gelatin capsules are composed of two halves that are joined following filling.
b. Hard gelatin capsules may be filled with aqueous liquids.
c. Hard gelatin capsules may be filled with non-aqueous liquids.
d. Hard gelatin capsules may be filled with aqueous drug suspensions.

4. **Concerning soft gelatin capsules, which of the following statements are true?**
a. Soft gelatin capsules generally contain a disintegrant.
b. Soft gelatin capsules may be formulated with a powder fill.
c. Soft gelatin capsules often contain emulsions.
d. Soft gelatin capsules may exhibit favourable drug bioavailability due to lipid breakdown.

5. **Concerning the composition and manufacture of hard gelatin capsule shells, which of the following statements are true?**
a. Hard gelatin capsule shells contain water.
b. Hard gelatin capsule shells may contain colouring agents.

c. Hard gelatin capsule shells always contain low concentrations
    of surfactants.
d. Hard gelatin capsule shells are prepared using a
    polymerisation process.

6. **Solid fills for hard gelatin capsules frequently contain which
    of the following excipients for the designated purpose?**
a. Glidants – to improve powder flow.
b. Lactose – as a diluent.
c. Magnesium stearate – as a lubricant.
d. Starch – as a disintegrant.

7. **Ideally, solid fills for hard gelatin capsules should exhibit
    which of the following properties?**
a. Homogeneity of mix.
b. A low angle of repose.
c. A high Hausner ratio (> 1.6).
d. Low shearing stresses during powder flow.

8. **Which of the following are suitable as liquid fills for hard
    gelatin capsules?**
a. Polyethylene glycols.
b. Arachis oil.
c. Fatty acid esters.
d. Purified water.

9. **Regarding soft gelatin capsules, which of the following
    statements are true?**
a. The capsule shell contains glycerol as a plasticising agent.
b. The capsule shell contains approximately 20% water.
c. The capsule shell is preferentially composed of type A
    gelatin.
d. The capsule shell may contain colours or opacifiers.

10. **Which of the following excipients are common components
    of the fill of soft gelatin capsules?**
a. Alcohol.
b. Lipophilic liquids, e.g. soyabean oil.
c. Water-miscible liquids, e.g. surface-active agents.
d. Lubricants, e.g. magnesium stearate.

Multiple choice answers

Chapter 1

1.	**a.** False.	**b.** True.	**c.** True.	**d.** False.
2.	**a.** True.	**b.** False.	**c.** True.	**d.** False.
3.	**a.** True.	**b.** False.	**c.** True.	**d.** True.
4.	**a.** True.	**b.** False.	**c.** True.	**d.** True.
5.	**a.** False.	**b.** True.	**c.** False.	**d.** True.
6.	**a.** False.	**b.** True.	**c.** False.	**d.** True.
7.	**a.** True.	**b.** False.	**c.** True.	**d.** False.
8.	**a.** False.	**b.** True.	**c.** False.	**d.** False.
9.	**a.** False.	**b.** False.	**c.** True.	**d.** False.
10.	**a.** False.	**b.** True.	**c.** True.	**d.** False.

Chapter 2

1.	**a.** True.	**b.** True.	**c.** True.	**d.** True.
2.	**a.** True.	**b.** False.	**c.** True.	**d.** True.
3.	**a.** False.	**b.** False.	**c.** True.	**d.** False.
4.	**a.** True.	**b.** False.	**c.** True.	**d.** False.
5.	**a.** True.	**b.** True.	**c.** False.	**d.** False.
6.	**a.** False.	**b.** True.	**c.** False.	**d.** True.
7.	**a.** False.	**b.** True.	**c.** True.	**d.** True.
8.	**a.** False.	**b.** False.	**c.** True.	**d.** True.
9.	**a.** True.	**b.** True.	**c.** False.	**d.** False.
10.	**a.** True.	**b.** True.	**c.** True.	**d.** False.

Chapter 3

1.	**a.** True.	**b.** True.	**c.** True.	**d.** True.
2.	**a.** True.	**b.** False.	**c.** True.	**d.** False.
3.	**a.** False.	**b.** False.	**c.** True.	**d.** False.
4.	**a.** True.	**b.** True.	**c.** False.	**d.** False.
5.	**a.** False.	**b.** True.	**c.** True.	**d.** True.
6.	**a.** True.	**b.** False.	**c.** False.	**d.** True.
7.	**a.** True.	**b.** False.	**c.** True.	**d.** True.
8.	**a.** False.	**b.** False.	**c.** False.	**d.** True.
9.	**a.** True.	**b.** True.	**c.** False.	**d.** False.
10.	**a.** True.	**b.** True.	**c.** True.	**d.** True.

Chapter 4

1.	**a.** False.	**b.** False.	**c.** False.	**d.** True.
2.	**a.** True.	**b.** True.	**c.** True.	**d.** True.
3.	**a.** True.	**b.** True.	**c.** True.	**d.** False.
4.	**a.** True.	**b.** False.	**c.** True.	**d.** True.
5.	**a.** False.	**b.** False.	**c.** True.	**d.** True.
6.	**a.** True.	**b.** False.	**c.** True.	**d.** False.
7.	**a.** False.	**b.** True.	**c.** True.	**d.** False.
8.	**a.** False.	**b.** True.	**c.** False.	**d.** True.
9.	**a.** True.	**b.** True.	**c.** False.	**d.** True.
10.	**a.** True.	**b.** True.	**c.** True.	**d.** False.

Chapter 5

1.	**a.** True.	**b.** True.	**c.** False.	**d.** True.
2.	**a.** True.	**b.** True.	**c.** False.	**d.** False.
3.	**a.** True.	**b.** True.	**c.** True.	**d.** False.
4.	**a.** True.	**b.** True.	**c.** False.	**d.** False.
5.	**a.** False.	**b.** True.	**c.** True.	**d.** True.
6.	**a.** False.	**b.** False.	**c.** True.	**d.** True.
7.	**a.** False.	**b.** False.	**c.** False.	**d.** True.
8.	**a.** False.	**b.** True.	**c.** True.	**d.** False.
9.	**a.** True.	**b.** False.	**c.** True.	**d.** False.
10.	**a.** False.	**b.** False.	**c.** False.	**d.** False.

Chapter 6

1.	**a.** True.	**b.** True.	**c.** True.	**d.** False.
2.	**a.** False.	**b.** True.	**c.** True.	**d.** True.
3.	**a.** True.	**b.** True.	**c.** False.	**d.** False.
4.	**a.** False.	**b.** True.	**c.** True.	**d.** True.
5.	**a.** False.	**b.** True.	**c.** False.	**d.** False.
6.	**a.** True.	**b.** True.	**c.** True.	**d.** False.
7.	**a.** True.	**b.** False.	**c.** True.	**d.** False.
8.	**a.** True.	**b.** False.	**c.** True.	**d.** True.
9.	**a.** True.	**b.** False.	**c.** False.	**d.** True.
10.	**a.** True.	**b.** True.	**c.** True.	**d.** False.

Chapter 7

1.	**a.** True.	**b.** True.	**c.** False.	**d.** False.
2.	**a.** True.	**b.** True.	**c.** False.	**d.** True.
3.	**a.** True.	**b.** False.	**c.** False.	**d.** True.
4.	**a.** False.	**b.** True.	**c.** False.	**d.** True.
5.	**a.** True.	**b.** True.	**c.** True.	**d.** False.
6.	**a.** True.	**b.** False.	**c.** True.	**d.** True.

7.	**a.** False.	**b.** True.	**c.** True.	**d.** False.
8.	**a.** True.	**b.** False.	**c.** True.	**d.** True.
9.	**a.** True.	**b.** False.	**c.** True.	**d.** True.
10.	**a.** True.	**b.** False.	**c.** True.	**d.** False.

Chapter 8

1.	**a.** True.	**b.** False.	**c.** True.	**d.** False.
2.	**a.** True.	**b.** True.	**c.** False.	**d.** False.
3.	**a.** False.	**b.** True.	**c.** True.	**d.** True.
4.	**a.** True.	**b.** False.	**c.** False.	**d.** False.
5.	**a.** True.	**b.** False.	**c.** True.	**d.** False.
6.	**a.** False.	**b.** False.	**c.** True.	**d.** False.
7.	**a.** True.	**b.** True.	**c.** True.	**d.** False.
8.	**a.** False.	**b.** True.	**c.** True.	**d.** True.
9.	**a.** True.	**b.** False.	**c.** False.	**d.** False.
10.	**a.** True.	**b.** True.	**c.** False.	**d.** True.

Chapter 9

1.	**a.** False.	**b.** False.	**c.** True.	**d.** True.
2.	**a.** True.	**b.** False.	**c.** True.	**d.** False.
3.	**a.** True.	**b.** True.	**c.** True.	**d.** False.
4.	**a.** True.	**b.** False.	**c.** True.	**d.** False.
5.	**a.** False.	**b.** True.	**c.** False.	**d.** True.
6.	**a.** False.	**b.** False.	**c.** True.	**d.** False.
7.	**a.** False.	**b.** False.	**c.** False.	**d.** True.
8.	**a.** False.	**b.** False.	**c.** False.	**d.** True.
9.	**a.** True.	**b.** True.	**c.** False.	**d.** True.
10.	**a.** True.	**b.** True.	**c.** False.	**d.** True.

Chapter 10

1.	**a.** True.	**b.** True.	**c.** True.	**d.** False.
2.	**a.** False.	**b.** True.	**c.** True.	**d.** False.
3.	**a.** True.	**b.** False.	**c.** True.	**d.** False.
4.	**a.** False.	**b.** False.	**c.** False.	**d.** True.
5.	**a.** True.	**b.** True.	**c.** False.	**d.** False.
6.	**a.** True.	**b.** True.	**c.** True.	**d.** True.
7.	**a.** True.	**b.** True.	**c.** False.	**d.** True.
8.	**a.** True.	**b.** True.	**c.** True.	**d.** False.
9.	**a.** True.	**b.** False.	**c.** False.	**d.** True.
10.	**a.** False.	**b.** True.	**c.** True.	**d.** False.

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