Rheumatic diseases

Immunological mechanisms and prospects for new therapies

Edited by J. S. Hill Gaston

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RHEUMATIC DISEASES

Immunological mechanisms and prospects for new therapies

This authoritative review volume provides a wide-ranging account of the immunological mechanisms that underlie many rheumatic diseases. Advances in our understanding of the immunopathology of diseases such as rheumatoid arthritis, ankylosing spondylitis and SLE are paving the way for the development of effective and rational new therapies. This exciting prospect is an important stimulus for groundbreaking research into these diseases and the investigation of new therapeutic options. As the first book to focus exclusively on this burgeoning area of clinical research, this is an invaluable and contemporary account for all rheumatologists, clinical immunologists and those seeking to develop effective new therapies to combat rheumatic diseases.

RHEUMATIC DISEASES

Immunological mechanisms and prospects for new therapies

Edited by

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1

Implications of advances in immunology for understanding the pathogenesis and treatment of rheumatic disease

J. S. H. GASTON

Advances in immunology

This book has been designed to meet the needs of those whose clinical or research interests are in rheumatic diseases. Within rheumatology, there has been a lively debate on the relevance or otherwise of advances in immunology to a better understanding of rheumatic diseases and their treatment. Until relatively recently, it would have to be conceded that treatment of a disease like rheumatoid arthritis (RA) has been based on empirical observation of the usefulness of certain drugs (usually tested originally on the basis of some wholly erroneous idea about pathogenesis); in short, rational therapies based on a new understanding of immunopathology were in short supply. This situation is now changing, with the successful application of 'biologic' treatments, such as antibodies to tumour necrosis factor alpha (TNFα), or recombinant interleukin 1 (IL-1) receptor antagonist in RA (Arend & Dayer, 1995; Maini et al., 1995), and the initial exploration of supplying such therapy by means of gene transfer (Evans & Robbins, 1996). It is very likely that much of the therapeutic effort of rheumatologists in the first part of the 21st century will be directed to defining the place of novel biological therapies in the management of rheumatological disease, while determining in more detail the mode of action of current empirical therapies so that they may be used to best advantage.

In order to participate in this process, it is necessary to have at least some working knowledge of the components of the immune system, and how they might be implicated in rheumatic disease. Although there may have been some dissatisfaction with the relatively small contribution of immunologists to the therapy of rheumatic disease since the late 1960s, it was clearly unrealistic to expect a major therapeutic impact on disease at a time when so little normal immune physiology had been defined. There has been an enormous explosion in knowledge since the late 1970s, and this process continues, fuelled by the power of current molecular techniques. In 1982, three interleukins had been characterized, whereas the number is now approaching 20. To these can be added those cytokines that for some reason do not qualify for interleukin status, such as TNF (three varieties), oncostatin-M, leukaemia inhibitory factor (LIF), and many others, most having names that completely fail to indicate the wide range of their involvement in inflammation and immunity – none of the three examples quoted has effects confined to neoplasia.

To these must be added a plethora of chemokines; around 40 are defined, but there are perhaps as many as 60 additional expressed-sequence tags (ESTs) that have features which make them putative chemokines (Schall & Bacon, 1994). The set of chemokines join IL-8, otherwise a somewhat anomalous member of the interleukin family. There are four major families of chemokines (CKs), classified according to the number and position of cysteine residues at their amino terminus. These are termed the C-X-C, C-C and C families (Baggiolini, Dewald & Moser, 1997), along with the recently described C-X₃-C chemokine (Bazan et al., 1997). The predominant actions of chemokines are on neutrophils (C-X-C family) or monocytes and lymphocytes (C-C, C and C-X₃-C families). An expanding family of receptors (CCRs and CXCRs) for these factors is also being defined. Faced with so many factors and receptors' it is tempting to assume substantial redundancy, and this is certainly the case for some functions (e.g. neutrophil chemotaxis), but recent investigations in HIV infection clearly show that polymorphisms in single chemokine receptors can have measurable effects on clinical outcome (Garred, 1998). For HIV, these effects probably reflect the fact that the virus hijacks the chemokine receptor to gain access to cells. Nevertheless, it is possible that associations between polymorphisms, in particular chemokine or chemokine receptor genes, will also be discovered with respect to rheumatic diseases when the genetic influences on disease incidence and characteristics are unravelled by current whole genome search strategies.

The most recent chemokines and chemokine receptors to be described are examples of 'reverse genetics', with the first inkling of their existence coming from DNA sequences rather than a biological activity (Forster *et al.*, 1996; Bazan *et al.*, 1997; Gunn *et al.*, 1998a,b). This effectively short-circuits the previous labour-intensive procedure of purifying, sequencing, cloning and expressing new biologically active factors. A similar route has also been used to define new cytokines such as IL-17 (Yao *et al.*, 1995a,b). Amongst newer chemokines are factors that have important roles in establishing the architecture of the immune system, by, for instance, attracting cells into germinal centres (Gunn *et al.*, 1998a). Since the rheumatoid arthritis synovium can take on the architectural appearance of a lymph node, including germinal centre formation, this is likely to reflect

chemokine production within the tissue, and the particular stimuli that give rise to this response in chronic inflammation will be of great interest.

Cell surface molecules on lymphoid and myeloid cells are now defined to the extent of more than 160 CD (cluster of differentiation) numbers, where 15 were required in 1982 (Shaw, Turni & Katz, 1998) (http://www.ncbi.nlm.nih.gov/prow/). Some of these can be cleaved from cell surfaces and then act as cytokines or can modulate cytokine action by binding them in solution. The precise function of many of these surface molecules has yet to be defined, but even at our current partial stage of understanding, the cells that make up the immune system are amongst the most thoroughly characterized in the whole body. It is evident that things have changed substantially from the classic description of the lymphocyte as a 'small round cell ... of which literally nothing of importance is known' (Gowans, 1996).

At one time, the interests of cellular immunologists were almost entirely focused on events at the cell surface, the cell itself being treated as something of a 'black box', but now many signalling pathways have been defined in great detail and shown to be very complex. Examples include signalling from T and B cell receptors, from co-stimulatory molecules and cytokine receptors, and the signalling mechanisms that mediate programmed cell death (apoptosis). Many of the components of the signalling pathways are not used uniquely by the immune system, and signalling pathways used by hormones and growth factors commonly intersect with those of lymphocytes. In addition, many of the transcription factors that are important in lymphocyte activation have a wide spectrum of activity in the activation of other genes (an example would be NFKB). However, signalling components and transcription factors that have a very particular role in the immune system have also been discovered; signalling through the IL-12 receptor is wholly dependent on the signal transducer and activator of transcription STAT-4 (Thierfelder et al., 1996), whereas STAT-6 is required for the actions of IL-4 (Shimoda et al., 1996; Takeda et al., 1996). Deficiency in a single tyrosine kinase, Btk, accounts for Bruton's X-linked agammaglobulinaemia (Vetrie et al., 1993). Increasing knowledge of signalling pathways is likely to be particularly important since it is often more realistic to manipulate the immune system by interfering with signalling pathways using conventional pharmacologic approaches, rather than using biological agents that act primarily on cellular interactions. Even where components of a signalling pathway are used for several biological functions, it is sometimes possible to obtain a useful therapeutic effect on one of the functions without necessarily producing the same phenotype as a genetic knock-out. A recent paper suggested that sulphasalazine, widely used in rheumatology with good efficacy and safety, has its effects by inhibiting the transcription factor NFKB (Wahl et al., 1998), even though NFKB has multiple

actions and genetic knock-out of certain NF κ B components can be highly deleterious (Baldwin, 1996).

These are just some examples of the exponential growth in knowledge of the immune system; equal attention could be paid to work on the multiple components of the major histocompatibility complex (MHC), the formation and structure of antigen-specific receptors, the components involved in the induction of apoptosis and its role in control of immune responses, and the mechanisms underlying lymphocyte homing and recirculation – to mention only a few areas that have undergone intense scrutiny and are highly likely to be relevant to the pathogenesis of rheumatic diseases.

Faced with the baroque complexity of the immune system, which now genuinely rivals the central nervous system as a finely tuned physiological mechanism, the rheumatologist who has not previously had to grapple with immunology might be tempted to despair. This book is designed to dispel such feelings. Although it would be foolhardy to imply that our current knowledge of immunology is other than partial, a large proportion of the immunological events that are likely to be responsible for rheumatic diseases have already been defined. This is not to say that we know the immunological basis of most rheumatic diseases - from it; rather we have a reasonably comprehensive list of the kinds of thing that might go wrong and can now determine those which actually do cause disease. To take a non-rheumatological example; prior to the discovery of the CD40-CD40-ligand interaction (Disanto et al., 1993), the mechanism of the sexlinked hyperIgM immunodeficiency (HIgM) syndrome was completely unknown. Although it is clearly possible, and indeed likely, that additional co-stimulatory molecules remain to be discovered, deficiencies in these will have some phenotypic similarities with the lack of CD40-ligand (CD40L), which is responsible for HIgM syndrome. Likewise, prior to the description of the cytokine IL-12, certain patients who had difficulty in combating infection by intracellular organisms such as salmonellae and atypical mycobacteria had an immunodeficiency that was unexplained. Recently, patients with just this clinical phenotype have been discovered to have abnormalities in IL-12 (Altare et al., 1998a) or its receptor (Altare et al., 1998b). In keeping with the physiological role of IL-12 in influencing interferon- γ (IFN- γ) production, a somewhat similar phenotype has been seen in patients with defective IFN-y receptors (Jouanguy et al., 1996). Thus, gradually, clinical phenotypes are being matched with abnormalities in defined components of the immune system. Although this is most easily done where there is the equivalent of a genetic knock-out in humans, more subtle defects that might contribute to the pathogenesis of rheumatic diseases are currently being sought.

Overview of this volume

The objective in each of the remaining chapters of this review is to describe current knowledge of the principal immunological mechanisms in order to provide a physiological 'map', which can then be used to indicate components that might 'go wrong' in rheumatic disease, or components that might in the future be useful targets of therapy. Examples of known defects, and therapies that have been found effective, are provided, but the up-to-date description of the immune system should provide a framework that will facilitate future thinking about the pathogenesis of rheumatological disorders, and an understanding of the rationale behind the novel therapies currently being dreamt up by biotechnology companies.

Specific immune responses begin with antigen recognition and the receptors that mediate this: surface immunoglobulin in the case of B cells, and the different forms of the T cell receptor for antigen. Autoantibodies, which are associated with rheumatic diseases and, in some cases, are directly pathogenic, have now been studied in great detail using molecular techniques, particularly from a structural point of view. The mechanisms whereby specific antibodies are generated, and the properties of particular autoantibodies, are described in Chapter 4.

T cell receptors differ radically from immunoglobulins in their inability to distinguish intact antigen; instead they recognize short peptides that result from processing and which are then presented by means of molecules encoded in the MHC (known as the human leukocyte antigen (HLA) system in humans). This difference has profound implications for our understanding of autoimmune T cell responses, and how these might arise. The concept of 'molecular mimicry' has been put forward for some time in relation to autoimmunity (Oldstone, 1990). Mimicry occurs when antigenic determinants on pathogens (e.g. viruses or bacteria) resemble a determinant on a self-protein, so that an immune response to one cross-reacts with the other. Recently, a first example of this postulated mechanism has been documented in the keratitis induced by infection with herpes simplex virus type 1 (Zhao et al., 1998). Initially, molecular mimicry was considered in relation to cross-reacting antibodies, where, for linear determinants that reflect a particular amino acid sequence, it is possible simply to compare the sequence of an autoantigen with that of a candidate mimic (e.g. a viral protein) to look for regions of sequence conservation. However, for a peptide to be recognized by a T cell there are only two requirements: it must bind to MHC and have appropriate amino acids to contact the T cell receptor. This means that there may be little or no linear sequence conservation between potential mimics, and they have to be sought in other ways. The necessary approaches are described in Chapter 2, having been pioneered by Wucherpfennig and Strominger (1995).

The prominent associations between MHC alleles and various rheumatic

disorders represent the most compelling evidence for the involvement of T lymphocytes in pathogenesis: the B27 – ankylosing spondylitis association is still, after 25 years, the strongest for any disease (Brewerton *et al.*, 1973). Despite the strength of these associations for both spondyloarthropathies and RA, and the intense scrutiny of HLA since the associations were first described, the mechanisms underlying them are frustratingly obscure and may well be different for different diseases. Hoyt Buckner and Nepom describe current understanding of the components of the MHC, how they function in immune responses and likely ways in which diseases might be associated with particular alleles (Chapter 3).

The idea that recognition of antigen alone would not be a sufficient signal for activation of B or T cells was predicted early on (Bretscher & Cohn, 1970) and has proved to be the case. Two chapters in this volume (Chapters 5 and 6) deal with important co-stimulatory receptor ligand pairs that have been identified in recent years and that are involved in T cell stimulation. The first of these involves, on the T cell, CD28 and the related molecule CTLA-4, both of which can bind to two other related ligands on antigen-presenting cells, B7.1 and B7.2. The second is CD40L on T cells, which binds to CD40 on antigen-presenting cells, particularly B cells. The importance of these interactions is underlined by the ability to induce allograft acceptance by blocking both pathways - a measure of the severe degree of immunosuppression that is produced (Larsen et al., 1996). However, although CD28 and CD40 were first described as important molecules for T cell and B cell co-stimulation, respectively (prior to the discovery of their ligands), the situation is inevitably more complex. Not only can T cells express B7.1/7.2 and CD40, and B cells CD40L, but in all cases the interaction of these receptor – ligand pairs results in a two-way conversation. Thus, there are important effects on both T and B cells in the CD40L-CD40 interaction, and the same ligand pair is involved in interactions between T cells and antigen-presenting cells, particularly dendritic cells. The same is true for CD28-B7.1, with additional complexity resulting from the inducible expression of CTLA-4 as an alternative ligand for B7.1/7.2 (Thompson & Allison, 1997). The interactions have implications for antibody production, the cytokine programme carried out by activated T cells, and immunity to intracellular pathogens; both co-stimulatory pathways are already being targeted in immunomodulation strategies (Durie et al., 1993; Webb, Walmsley & Feldmann, 1996).

Appropriate engagement of B and T cell receptors by antigen has profound consequences for the cell, such as entering the cell cycle, expressing activation markers on its surface and producing cytokines; in effect, an entirely new genetic programme is initiated and there are multiple differences in gene transcription (involving hundreds of genes) between activated and quiescent T or B cells. To achieve this, intracellular signalling mechanisms are required; in the first instance,

both B and T cells are dependent on components of the receptor complex other than those necessary for antigen recognition to initiate cell signalling. Other surface molecules, including the co-stimulatory molecules alluded to in the previous paragraph, can 'fine-tune' the response, again acting through their influence on components of the signalling pathway. Current understanding of these processes, and prospects for modulating intracellular signalling with drugs, are detailed in Chapter 7. It is clear that the notion that encounter with antigen would translate a T or B cell from an 'off' to an 'on' state in some all-or-nothing manner is quite inadequate. The effects of antigen on an antigen-specific cell will reflect the balance of positive and negative signals, in much the same way that a smooth motor action by a limb represents the graded activation of agonist and antagonist muscles rather than unopposed agonists. Indeed, there is an increasing need for such a balance in the interplay of antagonists and agonists in fine accurate movements. In addition to the signalling components involved in this process, the influence of the proportion of receptors on a cell that are activated, and the time for which they are activated, have recently been shown to be critical for the final outcome (Valitutti et al., 1995; Viola & Lanzavecchia, 1996; Iezzi, Karjalainen & Lanzavecchia, 1998).

None of the mechanisms for antigen recognition and the signalling of appropriate responses addresses the question of the geography of the immune system; immune responses take place at certain sites, both within the lymphoid system and in the tissues generally, and ways of ensuring that effector cells are delivered to appropriate locations are an essential component of the immune system. This has been a very active field of research, and an overview is provided by Pitzalis (Chapter 8). Lymphocyte trafficking to appropriate tissue requires expression of ligand on endothelial cells that can be recognized by leukocytes; in fact multiple ligands are involved in the arrest and eventual transmigration of cells across the endothelium into the tissues. More recently recognized components in the process are the chemokines produced by endothelial cells and recognized by specific receptors on the leukocytes (Gunn et al., 1998b); the chemokine-receptor interaction, in turn, modulates the affinity of the integrins required for firm adhesion of cells to the vessel wall. Again there are hopes that it may be possible to modulate leukocyte traffic therapeutically, and attempts to do this have already been made (Kavanaugh et al., 1994).

Having dealt with the principal mechanisms for recognizing and making appropriate responses to antigens, it is important to examine the downstream effects of the antigen-specific responses, since these are the processes that turn immunological recognition into disease. In relation to inflammatory arthritis, it is clear that the synovium is dominated by the presence of macrophage/monocyte-derived rather than T cell-derived cytokines, although the recent description of a novel cytokine, IL-17, exclusively produced by T cells but with monokine-like effects may alter this perception (Spriggs, 1997). However, there is good evidence that the monokines are directly responsible for joint destruction. Much attention has focused on the two principal monokines: TNF α and IL-1. The relative important of each and therapeutic possibilities of inhibiting either or both are discussed by van den Berg and van de Loo (Chapter 10). This review also emphasizes the possible roles of the more recently described proinflammatory monokines such as IL-12 and IL-15, and the production of regulatory monokines such as IL-10 and transforming growth factor beta (TGF β). It is clear that the effects of blocking one cytokine or adding another are not readily predictable in relation to arthritis, with differing effects depending on the clinical endpoint examined (e.g. joint swelling versus cartilage destruction), and depending on the phase in the evolution of arthritis at which they are applied.

Immune responses have to be controlled at the level of initiation by fine tuning their magnitude and ensuring that their consequences (production of a cytokine or immunoglobulin subclass) are appropriate to the context, e.g. IFN- γ to deal with mycobacterial infection, IgE production for an intestinal helminth. However, it is equally important to have mechanisms for ensuring that appropriate immune responses do not continue indefinitely, and the principal mechanism used by the immune system at all levels involves apoptosis. Apoptosis as a mechanism for the removal of immune or inflammatory effectors after they have performed their required actions is now realised to be of critical importance for normal immunological health. Strains of mice, which were already under scrutiny as examples of spontaneous systemic lupus erythematosus (SLE)-like illness, proved to lack molecules important in one of the major pathways for inducing apoptosis, Fas (in MRL lpr/lpr mice) or its ligand (FasL) (in C3H gld/gld mice). The mechanisms of apoptosis, the molecules involved (which have rapidly proliferated in recent years) and the regulation of the process are discussed in Chapter 9.

A further level of control in the immune system concerns the recognition of self-antigens. At one time, it was simply assumed that this was forbidden, and indeed mechanisms that allow the deletion of effector T and B cells with high-affinity receptors with self have been worked out in great detail (Kappler, Roehm & Marrack, 1987). However, it has also become apparent that potentially auto-immune effector cells are part of the normal B and T cell repertoire and that, in fact, the functional immune repertoire is selected on the basis of an ability to recognize self MHC–peptide complexes (Blackman *et al.*, 1986). In any case, a repertoire that deleted every cell that could recognize an epitope present on some self molecule would be fatally compromised, since there is considerable (possibly complete) overlap between the set of epitopes comprising self and those expressed by

pathogens. Accordingly, mechanisms for the control of potentially autoreactive cells in the periphery after thymic selection are required. These are only just beginning to be delineated; the potential for cells present in the normal T cell repertoire to mediate autoimmune disease is highlighted in Chapter 11, as is the existence in the same repertoire of T cell subsets that are able to control the autoimmune disease (diabetes, thyroiditis) and, more recently, in inflammatory bowel disease, where it may be necessary to tolerate normal intestinal organisms as well as self (Duchmann *et al.*, 1995; 1996). However it is probable that similar mechanisms protect against rheumatic disease or are at fault when it occurs.

The exquisite sensitivity of immune recognition by immunoglobulins and T cell receptors has long fascinated immunologists, and how this is accomplished is now understood in great detail. However, since the process rests fundamentally on chance recombination events to generate receptors of the required specificity, the experience of successfully generating an antibody or T cell to deal with an important pathogen cannot be passed on from generation to generation: the functional receptor is not encoded in the germ-line, only the component genes required for the recombination process. The acquired 'wisdom' of the immune response that is germ-line encoded comprises the innate immune system, which does not require the generation of novel, highly specific receptors (Fearon, 1997). For example, receptors like those that bind mannose are useful for interactions with many bacteria, as are components of the complement system. Having been previously dazzled by the sophistication of the adaptive immune response, immunologists are now paying much more attention to the innate immune system, and in particular to its interactions with the adaptive immune response, which produces the best of both worlds. A good example is the ability of C3d, when complexed to antigen, to boost the antibody response by several orders of magnitude, so that tiny quantities of antigen are rendered immunogenic (Dempsey et al., 1996). This effect depends on a complement receptor, CD21 or CR2; the family of complement receptors is reviewed in detail by Ahearn and Rosengard (Chapter 12), along with an account of how abnormalities in complement components and their receptors are likely to play a part in diseases such as SLE.

Concluding remarks

It can be argued that most rheumatic disorders occur in the context of an immune system that is generally competent – patients are able to protect themselves adequately from pathogens – but has some minor character defect that, as in all the best Greek tragedies, leads ultimately to disaster. These defects have not generally been defined but might include overexuberant responses to environmental

antigens in terms of the cytokines produced or the duration of response, or a crucial confusion between similar epitopes on foreign and self antigens. Our therapeutic response to these forgivable errors by the immune system has generally been to shut the whole system down to a substantial extent, with the inevitable consequence of vulnerability to the infectious agents. It is to be hoped that in future more subtle sanctions on the immune system, or a period of re-education, might have the desired effects on autoimmune/chronic inflammatory diseases, while maintaining intact defences against pathogens.

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2

The role of T cells in autoimmune disease R. J. MOOTS and K. W. WUCHERPFENNIG

Introduction

Knowledge about T lymphocyte function has expanded dramatically in the 1990s, resulting in many advances in the understanding of the mechanisms underlying autoimmune diseases. T lymphocytes, initially believed to play a small role, if any, in autoimmune disease, are now implicated even in 'antibody-mediated' conditions such as pemphigus vulgaris. The undoubted advances in understanding autoimmune diseases have not, however, developed at the same pace for all conditions. More is known about T cell behaviour in multiple sclerosis and myasthenia gravis than in rheumatological conditions such as Sjögren's syndrome, SLE and RA. Indeed there are still claims, with some evidence, that T cells do not play a significant role at all in RA (Fox, 1997). In this chapter, we shall present some of the evidence implicating a pathological role for T cells in autoimmune rheumatic diseases particularly RA and discuss this in the context of recent advances in knowledge of T cell function. We will focus on the potential mechanisms triggering the activation of T cells, and on effector mechanisms. Finally, we will discuss potential strategies for the modulation of the autoimmune response in therapy.

Are T cells involved in autoimmune disease?

Genetic predisposition

Many population, family and twin studies have clearly demonstrated that genetic factors exert a major influence on predisposition to autoimmune disease. Reviewed in detail elsewhere (Theofilopoulos, 1995), the best-defined association is with genes from the class II region of the MHC. These highly polymorphic genes encode cell surface glycoproteins, which present peptides to CD4+

T lymphocytes (Trowsdale, 1993). Potential mechanisms whereby MHC molecules may be involved in immune system dysfunction range from thymic selection, and generation of an immune repertoire, to presentation of disease-specific peptides to autoreactive T cells in the periphery.

Some of the clearest examples of HLA class II disease associations are seen with insulin-dependent diabetes mellitus (IDDM), pemphigus vulgaris and RA. Firstly, resistance to IDDM was found to correlate with the presence of the negatively charged amino acid aspartic acid at position 57 of the HLA-DQ β chain (Todd, Bell & McDevitt, 1987; Morel et al., 1988). HLA-DQ β chains with serine, valine or alanine at position 57 were then found to be associated with an increased susceptibility to IDDM (Nepom & Erlich, 1991). The profound effect of key residues within MHC alleles on susceptibility to autoimmune disease was confirmed by the discovery that patients with pemphigus vulgaris had a unique *HLA-DQB* allele, differing from closely related alleles at position 57 (Scharf *et* al., 1989). Subsequent structural analyses of MHC class II molecules by X-ray crystallography clearly illustrated the importance of position 57 in the class II β chain. This lies at one end of the peptide-binding groove, interacts intimately with the α chain and contributes to a peptide side-chain specificity pocket (Brown et al., 1993; Stern et al., 1994). Such a structure would limit potential autoantigens to those with appropriate size, shape, hydrophobicity and charge - implying that presentation of peptide to CD4+ T cells is important in the development of autoimmunity (Wucherpfennig & Strominger, 1995b).

The association between RA and class II MHC is also strong. Approximately 70% of patients with RA express HLA-DR4, increasing to over 90% in severe disease (Salmon et al., 1993), and nearly 100% in the subgroup Felty's syndrome. Furthermore, expression of HLA-DR4 (and other RA-related alleles) is a predictor of disease progression (Gough et al., 1994). A number of other HLA-DR alleles are also linked to RA. All of these share a common short sequence of amino acids (QKRAA), residues 67 to 71 in the polymorphic α helix of the HLA-DR β chain forming the third allelic hypervariable region or shared epitope (Gregersen, Silver & Winchester, 1987). Structural analysis of these disease-linked MHC molecules reveals that this region is intimately concerned with binding peptide and interacting with the antigen receptor on T cells (TCR) (Stern et al., 1994). In particular, the lysine at position 71 contributes a positive charge to the P4 specificity pocket. In vitro binding studies have defined properties characterizing peptides that are able to bind to RA-associated HLA-DR molecules (Hammer et al., 1994; Sinigaglia & Hammer, 1994; Woulfe et al., 1995). Such distinctions have allowed prediction of potential autoantigenic peptides, which include type II collagen (Fugger, Rothbard & Sonderstrup-McDevitt, 1996) and the 65 kDA heat shock protein (Hammer et al., 1995).

The significance of the P4 specificity pocket is underlined further in pemphigus vulgaris. In addition to HLA-DQ β , susceptibility to this disease is also associated with DRB1*0402, a rare subtype of HLA-DR4. This differs from the RA-associated DRB1*0404 at only three residues (DR β 67, 70 and 71), introducing a negative charge into the P4 pocket. Analysis of the self-peptide related to pemphigus vulgaris from the desmoglein 3 autoantigen (residues 190–204) reveals a complementary positive charge at position 4, appropriate for occupying the P4 pocket. Site-directed mutagenesis of DRB1*0402 confirmed that binding of desmoglein 3 (190–204) was indeed via the negatively charged residues of the P4 pocket (DR β 70 and 71) (Wucherpfennig *et al.*, 1995), highlighting the importance of the charge in the P4 pocket for binding of appropriate diseasespecific peptides to HLA-DR4.

Disease-associated MHC class II alleles may also contribute to the development of autoimmunity by mechanisms other than the presentation of pathogenic peptides to T cells. For example, such molecules may *fail* to present some peptide(s) that is important in *preventing* development of disease. This would predict, for example, that non-RA-associated *DRB1* alleles would be protective, and T cell activation occurs with other MHC molecules. Such a hypothesis has indeed been suggested. *HLA-DQ8* has been proposed to confer susceptibility to collagen-induced arthritis in transgenic mice, while certain *DRB1* alleles are proposed to confer protection (Zanelli, Gonzalez-Gay & David, 1995). Other explanations for links between the shared epitope and RA include sequence homology with bacterial proteins (Zanelli *et al.*, 1995), and direct binding of an endogenous 73 kDa heat shock protein to the QKRAA motif (Albani *et al.*, 1995). The latter may impede surface expression of the class II MHC molecule or may alter peptide-binding capability (Auger *et al.*, 1996).

T cells in lesions

If T cells are the culprits, are they found at the scene of the crime? This is indeed the case. Dense T cell infiltrates are characteristically seen in inflammatory plaques of demyelination in multiple sclerosis (Hauser *et al.*, 1986), in islet beta cells in IDDM and in the synovium in RA. T cell infiltrates in RA synovial tissue differ from normal or peripheral blood T cells in RA patients in a number of respects. They show many histological similarities to paracortical areas of lymph nodes, with occasional germinal centres. In addition the presence of specialized high endothelial venules (HEV) (intimately associated with T cell migration into tissues) suggests extensive T cell activity (Kurosaka & Ziff, 1983) and T cell infiltrates may sometimes be observed in clinically normal knees of patients with RA (Soden *et al.*, 1989).

The detailed characterization of such infiltrates is now possible by immunohistochemical staining and provides more clues as to what the T cells may be doing. In RA, the phenotype of synovial T cells differs significantly from that of the T cells in peripheral blood and in infiltrates in non-inflammatory joint lesions such as osteoarthrosis. The majority of infiltrating cells in the synovium express CD4, corresponding to the genetic MHC class II predisposition. They are highly differentiated, expressing the 'memory' CD45 RO⁺ marker in larger numbers than do peripheral blood cells from the same patients (reviewed in Salmon & Gaston, 1995). They also express markers of activation such as HLA-DR and the transferrin receptor (Smolen *et al.*, 1996). Other activation markers such as the IL-2 receptor (IL-2R) are lacking, or present on a smaller population of cells. However consideration of the dynamics of activation marker expression in response to mitogens *in vitro* (IL-2R, early and transient expression, HLA-DR later and prolonged expression), together with the expression of CD45 RO, suggests that they are in a stage of late activation.

Specificity of T cells

Detailed analysis of TCR usage has been performed in many conditions, but interpretation has been complicated by the fact that antigen-specific T cells represent only a small fraction of T cells in autoimmune lesions. A more convincing form of evidence for T cell-induced autoimmunity would be the characterization of self-proteins able to specifically activate T cells in patients. Furthermore, since autoreactive T cells may be isolated from normal individuals, autoaggressive T cells should occur with higher precursor frequency in patients. The search for candidate antigens for autoimmune responses started by focusing on proteins found in target organs, such as CNS myelin proteins in multiple sclerosis. Immunodominant myelin basic protein (MBP) peptide epitopes presented by disease-specific MHC molecules were characterized and T cells cloned. Such autoreactive MBP-specific T cells were found to be clonally expanded and persisted over time in patients with multiple sclerosis (Wucherpfenning et al., 1994b). Other myelin components including proteolipid protein (PLP) were subsequently found to contain T cell epitopes. Similarly, T cells reactive to islet proteins such as glutamic acid decarboxylase (GAD 65) have been isolated in IDDM (Nepom, 1995), and to components of the acetylcholine receptor protein in myasthenia gravis (Newsom-Davis et al., 1989).

Despite hard searching, no such autoantigen has yet been defined for RA, suggesting to some that T cells are not important in this disease. A number of candidates have, however, been proposed, including heat shock proteins, link protein and other proteoglycans. An important candidate is type II collagen (CII), expressed almost exclusively in synovial joints (Kuhn, 1987). An animal model, collagen-induced arthritis (CIA), has also been studied extensively. Here, an inflammatory erosive arthritis, with many similarities to RA, is induced by immunization with CII (Durie, Fava & Noelle, 1994) and mediated, at least in part, by CII-reactive T cells (Holmdahl *et al.*, 1989; Myers *et al.*, 1993). Reports of CII reactivity in humans, however, are limited to T cell lines, without characterization of peptide specificity. However, transgenic technology has lead to two recent reports describing T cell responses to CII restricted by HLA-DRI and HLA-DR4 in mice transgenic for human class II MHC and CD4 (Fugger *et al.*, 1996; Rosloniec *et al.*, 1997). In both cases, the immunodominant peptide, residues 259–271 of CII, was identical. The similarities in the peptide-binding site of these two MHC molecules support the hypothesis that presentation of a pathogenic 'arthritogenic' epitope to autoaggressive T cells is an important pathological mechanism.

Animal models

Many models for autoimmune disease exist in animals of many species, ranging from the tight-skinned chicken (systemic sclerosis) to the non-obese diabetic mouse (IDDM). These animal models have helped to define basic mechanisms of autoimmunity, although their relevance to the human diseases is not yet known. The major susceptibility factor in many of these models also lies within the MHC. The non-obese diabetic (NOD) mouse, for example, is spontaneously prone to develop an autoimmune insulitis, with an islet T cell infiltrate specific for islet proteins such as GAD 65 (Haskins & Wegmann, 1996). Susceptibility in NOD mice maps to the murine MHC class II molecule I-Ag⁷ and appears to be T cell mediated. Similarly, experimental allergic encephalomyelitis (EAE) in mice and rats is associated with a specific T cell response to CNS proteins including MBP, PLP and myelin oligodendrocyte glycoprotein (MOG). Myelin-specific T cell lines and clones have been produced, and specific peptides defined. T cell clones can transfer disease, even in the absence of antibodies or B lymphocytes (Steinman, 1996).

Collagen-induced arthritis is one of a number of animal models for RA, including adjuvant- and pristane-induced arthritis. It is characterized by an inflammatory polyarthritis sharing many features with RA. Disease develops in susceptible mouse or rat strains after immunization with CII and is characterized by specific T and B cell responses to CII (Staines & Wooley, 1994). It is interesting to note, however, that mice transgenic for a TCR reactive against a systemically expressed antigen were serendipitously observed to develop a localized joint disease (Kouskoff *et al.*, 1996). This suggests that the mechanisms underlying T cell autoimmunity may be considerably more complex than previously believed.

Clinical observations

The basic idea that pathological T cells cause autoimmune diseases is supported by clinical observations from different groups. An open pilot study of the use of IFN- γ therapy in multiple sclerosis was terminated soon after starting, because of an increased early relapse rate. This was associated with a high rate of spontaneous proliferation of peripheral blood lymphocytes and an increase in the specific response to MBP (Panitch *et al.*, 1987).

In RA, a variety of procedures designed to reduce T cell numbers or affect function have been used with some success. These include thoracic duct drainage (Vaughan *et al.*, 1984), total lymphoid irradiation (Helfgott, 1989), lymphapheresis (Wilder, Yarboro & Decker, 1982), and treatment with cyclosporin A (Forre, 1995). Recent clinical trials using a depleting anti-CD4⁺ monoclonal antibody did not produce clinical benefit, despite significant reduction in peripheral CD4⁺ cells (Weinblatt *et al.*, 1995). This has been cited as evidence against a significant role for T cells. However, measurement of peripheral cells may not accurately reflect an effect on T cells within the joint. Indeed, a different humanized *non-depleting* anti-CD4 monoclonal antibody appears to induce some clinical benefit (Panayi *et al.*, 1996).

What triggers autoreactivity in T cells?

The incomplete deletion of all potentially autoreactive T cells in the thymus may occur for a number of reasons. For example, not all self-proteins may be accessible to the thymus for presentation to maturing T cells. The resultant 'resting' autoreactive T cells in peripheral blood are considered a normal part of the immune repertoire and do not normally induce disease. Peripheral mechanisms hold these potentially pathogenic T cells in check. The key event in the induction of T cell-mediated autoimmunity would, therefore, need to overcome this and allow activation of autoreactive T cells. This is supported by data from animal models such as EAE, where disease can be adoptively transferred only with activated and not resting T cells (Zamvil & Steinman, 1990). In conditions such as multiple sclerosis and the murine model EAE, activation must occur outside the CNS and with a different protein, since the target proteins are not expressed outside the blood–brain barrier. A number of hypotheses have been proposed to explain this. The two major hypotheses currently are T cell activation by super-antigen, and molecular mimicry.

Both of these hypotheses implicate microbial pathogens. This is supported by both clinical and epidemiological evidence for potential infectious triggers of autoimmunity. For example, common viral infections such as measles and rubella may occasionally result in inflammatory CNS disease. In these conditions, there has been T cell reactivity to MBP, with no evidence of virus in the CNS (Johnson *et al.*, 1984). Viral infections have been reported to precede the development of IDDM and autoimmune myocarditis, and infection with a variety of pathogens has been thought for a long time to trigger RA. The complex epidemiology of all human autoimmune diseases indicates that a number of pathogens may be involved.

The potential role of superantigens in the triggering of autoreactive T cells

Superantigens are small proteins that are able to activate T cells by a mechanism bypassing the normal requirement for antigen processing (Marrack & Kappler, 1990). They bind to class II MHC molecules at conserved regions outside the peptide binding cleft and to TCR V β sequences away from the highly polymorphic CDR3 loops. Such an interaction with TCR is specific to particular V β families and, as such, superantigens are able to activate a large proportion of T cells. Indeed the term superantigen arose because of their potent T cell stimulatory capacity (in concentrations down to 10⁻¹³ M), leading to a marked skewing of the TCR V β repertoire. Many superantigens have been identified, including staphylococcal and streptococcal enterotoxins, a soluble mitogen from *Mycoplasma arthritidis* (MAM) and endogenous retroviral gene products.

Initially, superantigens were found to induce severe clinical conditions charactized by a massive secretion of cytokines, such as toxic shock syndrome (Jupin *et al.*, 1988). However, it soon became apparent that the binding of superantigen to MHC class II molecules and selected TCR V β gene products, coupled with their potency as T cell mitogens, could result in the activation of quiescent circulating T cells with a TCR specificity for self-protein. After clearance of the superantigen, most activated T cells would return to a resting state or be deleted. A small subset, however, of autoreactive T cells may home in to the site of expression of autoantigen and mediate disease (Fig. 2.1).

In the murine EAE model, MBP-specific I-A^u-restricted T cells using the V β 8.2 gene segment induce disease. The bacterial superantigen staphylococcal enterotoxin B (SEB) activates T cells expressing V β 8 and is able to induce clinical relapse or exacerbation in mice previously immunized with the MBP peptide (Brocke *et al.*, 1993). The murine V β 8 gene family is also utilized by pathogenic T cells in several other experimental models, including collagen-induced



Fig. 2.1. Superantigen-mediated activation of autoreactive T cells APC, antigenpresenting cell.

arthritis. Injection of MAM, a potent superantigen for cells with such TCRs, caused an exacerbation of arthritis in mice recovering from collagen-induced arthritis. Such a result is intriguing, as the organism from which MAM is derived is known itself to induce a chronic inflammatory erosive arthritis in rodents. Furthermore, while not a human pathogen, the major MAM-responsive human V β gene family is V β 17, reported to be overrepresented in synovial T cells in patients with RA (Zagon *et al.*, 1994). These observations suggest that superantigens could be significantly involved in the pathogenesis of chronic auto-immune disease, perhaps by inducing exacerbations and relapses. To test this hypothesis further, it will be important to compare the functional properties and fine specificity of, for example, V β 17-positive T cells in the blood and synovial fluid of RA patients. Such an involvement of superantigen in auto-immune disease would not only provide a link to infection – long postulated to be involved in the triggering of these disorders – but also offer the potential for novel therapy.



Fig. 2.2. Molecular mimicry. APC, antigen-presenting cell.

Molecular mimicry as trigger for autoreactive T cells

In many ways, the concept of mimicry, which depends upon a structural homology between self and foreign antigens, is intuitive. It would result in a cross-reactive T cell response if there were sufficient similarities between the two peptide epitopes. Microbial and host determinants would need to be similar enough to induce a cross-reacting immune response, yet different enough to break immunological tolerance (Fig. 2.2).

Molecular mimicry between microbes and antibodies that induce autoimmunity is well established. One of the earliest and best known is between group A streptococcal M antigens and myocardial and glomerular tissues, which occurs in rheumatic fever (Robinson & Kehoe, 1992). If such a phenomenon exists for T cells, it should be possible to demonstrate TCR cross-reactivity for selfpeptides and foreign peptides presented by appropriate (disease-associated) MHC molecules. Many workers have searched for this in vain, hampered by the far greater complexity of antigen recognition for T cells compared with antibodies. A cross-reactive T cell epitope must retain its ability to bind not only MHC but also a clonally distributed TCR. Another problem has been the equally complex biochemical identification of cross-reactive T cell epitopes from pathogens, requiring extensive biochemical fractionation before testing on T cell clones. In order to overcome these problems, many authors have used computer-based sequence homology searches to align linear sequences of similarity between self-proteins and foreign proteins. Predicted peptide epitopes were then synthesized and tested with specific T cell clones. Fujinami and Oldstone (1985) used this approach to immunize rabbits with a hepatitis B virus polymerase peptide, successfully inducing cross-reactive autoantibodies to MBP and hepatitis B virus polymerase peptide. Some cross-reactive T cells could be found, together with CNS perivascular infiltrates characteristic of EAE in four out of eleven rabbits. However, while EAE was induced normally by the MBP peptide, it was not possible to induce disease by immunization with the hepatitis B virus polymerase peptide.

This study demonstrated that immune-mediated injury may occur *after* the triggering immunogen had been cleared, a 'hit and run' event. It also made it much harder to link infection with autoimmunity clinically because the inducing agent was likely to have been cleared before the onset of autoimmune symptoms. If this time lag were a matter of years, it would be extremely difficult to prove an association between a common pathogen and autoimmune disease by epidemiological methods. Similar studies in animal models such as experimental autoimmune uveitis (hepatitis B virus DNA polymerase and S-antigen photoreceptor protein) (Singh *et al.*, 1990) and adjuvant arthritis (mycobacterial 65 kDa heat shock protein and joint proteoglycan) (van Eolen *et al.*, 1988) confirmed that different pathogens could induce autoimmune disease. While replicating infectious agents were not required to trigger molecular mimicry in those models, the triggering epitopes were derived from pathogens that did not normally affect the animal in question.

An early criticism of the T cell molecular mimicry hypothesis was that it appeared to counter the commonly held view that the TCR is exquisitely specific for a particular peptide–MHC complex. Unless the self and foreign sequences were identical (up to a 1 in 20⁶ chance), there would be no significant recognition. Over the last few years, however, it has become apparent that TCR recognition is more degenerate, and cross-reactivity at the T cell level may be more common than previously appreciated. In the mouse, a number of T cells clones have been reported that, while specific for MBP (Ac 1–9), were also able to recognize a dissimilar peptide presented by the same MHC class II molecule, I-A^u. As the two peptides had no obvious sequence similarity, the term 'space mimicry' was coined (Bhardwaj *et al.*, 1993).

Bearing in mind the evolving concept of degeneracy in TCR recognition, a

different strategy to investigate potential molecular mimicry in human autoimmune disease has been devised. Specific T cell clones reactive to human MBP were generated from patients with multiple sclerosis and used to characterize the structural requirements for recognition of the immunodominant MBP(85-99) peptide (Wucherpfennig et al., 1994a) in terms of MHC binding and TCR recognition. A series of amino acid substitutions were then selectively introduced at each critical position and the effects on T cell recognition analysed to define the sets of amino acids permitted at each critical residue. These structural data, together with the knowledge that amino acids side-chains required for binding to MHC molecules are degenerate, were used to search a protein sequence database of human pathogens. Candidate peptides were synthesized and tested for recognition by MBP-reactive human T cell clones derived from the peripheral blood of patients with multiple sclerosis. Seven viral and one bacterial peptide, from the 129 synthesized, were able to stimulate MBP-specific T cell clones efficiently, mimicking the immunodominant MBP(85–99) peptide (Wucherpfennig & Strominger, 1995a). Because of the degenerate MHC-binding motif, the viral/bacterial peptides had no obvious sequence homology with the MBP peptide. This indicated that some TCRs recognize not just a single peptide but rather a set of structurally related peptides derived from different antigens. A recent study that used combinatorial peptide libraries for definition of the T cell recognition motif has also detected cross-reactivity between dissimilar viral peptides and self-peptides for recognition by a MBP-specific T cell clone (Hemmer et al., 1997). The diverse nature of the viral peptides able to stimulate MBP-specific T cell clones in these data would suggest that more than one single pathogen is able to trigger autoimmunity. This may explain why it has been so difficult to link conclusively the pathogenesis of individual autoimmune diseases to particular pathogens, in the face of considerable evidence for association of disease with infection in general. Rather, it may be that a group of common microbial pathogens could be involved in the pathogenesis of autoimmune processes.

How do pathological T cells cause disease?

There is much evidence implicating T cells in the autoimmune process in humans. Moreover, T cells are able to transfer disease directly in animals. However, one basic problem remains. Many target cells, such as pancreatic beta cells and oligodendrocytes, do not express class II MHC. How, therefore, can CD4⁺ T cells mediate the damage? A number of mechanisms may explain this. Firstly, T cells secrete cytokines. Some induce expression of class II MHC, allowing T cell-induced damage to occur. Others recruit immune system cells, which damage target organs. Secondly, CD4⁺ T cells may kill target cells directly,

particularly utilizing the Fas/FasL pathway. These mechanisms will now be discussed in more detail.

Cytokine release

One consequence of CD4+ T cell activation is secretion of cytokines, in particular TNF α and TNF β , IFN- γ , and IL-2. These potent mediators have a variety of biological effects, potentially contributing to the autoimmune disease processes in several different ways (Cavallo, Pozzilli & Thorpe, 1994). IFN-y is a significant contributor to inflammation. It is able to recruit and activate a variety of inflammatory leukocytes, including macrophages. It also promotes upregulation of MHC class II molecules on antigen-presenting cells and induces their expression on non-professional antigen-presenting cells. Such a dysregulated expression of class II MHC molecules on target cells may amplify or facilitate an autoimmune response (Bottazzo et al., 1983) (Fig. 2.3). Indeed the aberrant expression of MHC class II molecules by non-immune cells has been described in a variety of conditions, such as thyrocytes in autoimmune thyroiditis (Hanafusa et al., 1983) and pancreatic cells in IDDM patients (Bottazzo et al., 1985; Foulis, Farquharson & Hardman, 1987). In both cases this would allow presentation of autoantigen to T cells. Cytokines that directly influence the immune system, such as IL-2, would have an obvious impact on an autoimmune response. Originally described as a T cell growth factor, IL-2 plays a critical role in the proliferation and differentiation of numerous classes of lymphocyte. Indeed, monoclonal anti-IL-2 antibodies may inhibit disease in animal models. Conversely, agents that stimulate the secretion of IL-2, such as IFN- α and IL-12, potentiate disease.

T helper cells (T_{H} 1 cells) secreting IL-2, IFN- γ and TNF can trigger EAE in rodent models and diabetes in the NOD mouse. TNF is one of the cytokines found in high concentration in rheumatoid joints. There is little doubt that this cytokine plays an important role in the pathophysiology of joint inflammation and destruction in both animals and humans (Feldmann, 1996; Feldmann, Brennan & Maini, 1996; Szekanecz, Szegedi & Koch, 1996). The biological effects include apoptosis, macrophage and polymorphonuclear leukocyte activation, B and T cell proliferation, secretion of proinflammatory cytokines (in a variety of cells), enhancement of fibroblast proliferation, nitric oxide release, secretion of collagenase and prostaglandin E_2 (PGE₂) by fibroblasts, and the resorption of bone and cartilage (Arend & Dayer, 1995; Lotz, 1996). Not surprisingly, there has been a considerable effort to modulate this cytokine as therapy for RA. Indeed, monoclonal anti-TNF antibody therapy induces dramatic early remissions in patients with RA (Elliott *et al.*, 1993; 1994), but, as yet, the effect is



Fig. 2.3. Effector mechanisms.

not long-lasting. Despite this, the full cytokine profile of synovial tissue and fluid is still being elucidated.

Fas-FasL interaction

Fas (APO-1, CD95) is a cell-surface receptor of the TNF receptor superfamily; it is able to induce apoptotic cell death when ligated by an appropriate antibody, or Fas ligand (FasL) (Nagata & Golstein, 1995). Indeed, membrane-bound TNF on T cells is also able to perform this and may synergize with the Fas/FasL pathway (Cleveland & Ihie, 1995). Although Fas signalling induces apoptosis, Fas ligation has also been shown to trigger cellular proliferation (Aggarwal *et al.*, 1995). FasL was originally identified on CD8⁺ T cells and can function as an effective perforin-independent mechanism whereby cytotoxic T cells (CTL) can induce target cell cytolysis (Lowin *et al.*, 1994). Subsequently, expression of FasL is observed on activated T cells, CD45 RO⁺ cells expressing very little until stimulated. Fas expression, in contrast, is generally found on peripheral blood T cells. The potential role for this molecule in regulating normal immune responses and maintaining self-tolerance became evident with the description of lymphoproliferative diseases in *lpr/lpr* and *gld/gld* mice, with defects in the genes encoding

Fas and FasL, respectively (reviewed in Lynch, Ramsdell & Alderson, 1995). These observations, together with the discovery that previously activated T cells expressing Fas may apoptose when stimulated through the TCR–CD3 complex (activation-induced cell death), suggested an intimate role for the Fas–FasL interactions in autoimmune disease (Kabelitz, Pohl & Pechhold, 1993; Crichfield *et al.*, 1994). Fas–FasL interaction occurring between T cells and other cells appear to result in clonal 'downsizing' of immune responses. If there was a dysfunction in this mechanism, then disordered immune responses and autoimmunity may result (Lynch *et al.*, 1995).

Recently, other observations have implicated an additional important role for Fas–FasL interactions: the direct killing of target cells by autoreactive CD4⁺ T cells. D'Souza *et al.* (1996) reported that CNS lesions in patients with multiple sclerosis demonstrated elevated levels of expression of Fas on oligodendrocytes, compared with tissue from normal subjects. In these lesions, microglia and infiltrating lymphocytes displayed an intense immunoreactivity for FasL. Rapid cell death in Fas-expressing oligodendrocytes could be induced by ligation with specific anti-Fas antibody or FasL, but, interestingly, apoptosis (as indicated by DNA fragmentation) did not occur.

Fas-mediated cell death appears to be a final common pathway in several T cell-mediated autoimmune diseases. The mechanism of beta cell loss in diabetes was investigated by crossing a transgenic mouse carrying a beta cell-specific TCR onto the NOD.scid background. These mice produced CD4⁺ T cells bearing transgenic TCR but were devoid of B cells or CD8⁺ cells. The mice developed an accelerated insulitis and overt disease. Pancreatic beta cell destruction was mediated by Fas-induced apoptosis (Chervonsky *et al.*, 1997; Kurrer *et al.*, 1997). Similarly, in a study by Giordano *et al.* (1997), thyrocytes from thyroid glands of patients with Hashimoto's thyroiditis, but not from non-auto-immune thyroids, expressed Fas. *In vitro* Fas expression was induced by IL-1 β , which is abundantly produced in the thyroid in Hashimoto's glands thyroiditis. Massive thyrocyte apoptosis was observed on cross-linking of Fas. Thyrocyte suicide or fratricide resulted because FasL was constitutively expressed both in normal and thyroiditis thyrocytes. Activated T cells that express FasL may also induce cross-linking of Fas.

Potential relevance to therapy

The goal behind so much of the work on T cell immunology in autoimmune disease has been to identify potential strategies for therapeutic intervention. The recent developments in the molecular and cellular immunology of T cells, discussed here, can now be used to develop alternative approaches to conventional

Process	Target
Antigen processing	Blockade of DM-mediated CLIP removal Selective inhibition of key proteases
Interference with target organ localization of T cells	Blockade of adhesion molecules/homing receptors Blockade of chemokines/chemoattractants
Blockade of T cell effector functions	Inhibition of T cell produced cytokines Inhibition of Fas–FasL interactions
Deletion/inactivation of autoaggressive T cells	Monoclonal antibody Mucosal or systemic administration of autoantigen/peptide

Table 2.1. Potential targets for treatment of autoimmune diseases

immunosuppression for the treatment of autoimmune diseases (Table 2.1). An ideal therapy would reverse established disease, or at least prevent further progression, by selectively inhibiting autoreactive cells while leaving the rest of the immune system intact.

As yet there appears to be a number of potential pitfalls with some of these approaches. A specific means of treating autoimmune disease would be to induce tolerance to key autoantigens. Perhaps the best (and oldest) known means of achieving this is by the administration of protein (or preferably peptide). It has long been realised that systemic or mucosal administration of protein or peptide in high doses can induce tolerance to the same protein. In animal models, these strategies have proved effective. Unfortunately their great potential has yet to be realised in humans. Early trials of oral MBP and CII in multiple sclerosis (Weiner et al., 1993) and RA (Trentham et al., 1993) have not provided any clinical benefit when subjected to larger formal study. However, systemic administration of specific peptides, or targeting to the mucosal immune system (such as with cholera toxin B subunit (Sun et al., 1996)), may improve the efficiency of tolerance induction and allow this to be of use in human disease. While we still rely on nonspecific drug therapy at present, the developments in understanding autoimmune disease discussed here suggest that there is considerable potential for major new developments.

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3

The role of MHC antigens in autoimmunity J. HOYT BUCKNER and G. T. NEPOM

Introduction

The immunological basis for autoimmune disease is complex, and autoimmune diseases themselves are diverse in character. The pathogenesis of specific autoimmune diseases may well be different, involving both genetic and environmental factors. However, many autoimmune diseases share an association with the HLA locus and, more specifically, the highly polymorphic class I and class II alleles found in this region. Recent advances in the understanding of the structure and function of HLA class I and class II molecules have made it possible to begin to decipher mechanisms by which these genes may be contributing to the development of autoimmunity. In this chapter, we will address the evidence that the HLA locus is associated with autoimmunity, the possible mechanisms by which HLA genes could contribute to disease, and the implication that this information will have on future assessment and treatment of patients with autoimmune diseases.

The HLA locus

The HLA locus is a cluster of genes found on chromosome 6 (6p21.3). This region includes the genes encoding HLA class I and class II proteins, complement and other factors important in the generation of the immune response. Figure 3.1 demonstrates the distribution of genes across this region. The HLA class I genes are located at the telomeric end of the human MHC. The *HLA-A*, *HLA-B* and *HLA-C* loci are referred to as class I genes and encode the principle transplantation antigens, which are expressed in all nucleated cells. The class I genes each encode a single polypeptide, the HLA class I heavy chain. The class I heavy chain forms a complex with β_2 -microglobulin, a protein encoded outside of the MHC region, to form the functional HLA class I molecule. The HLA class II complex is also shown in Fig. 3.1. Each class II haplotype may contain up to 14



Fig. 3.1. A representation of the HLA locus on chromosome 6 emphasizing the genes discussed in the text.

different class II loci, clustered into three subregions, termed *HLA-DR*, *HLA-DQ* and *HLA-DP*. Each subregion contains at least one functional beta (B) locus and one functional alpha (A) locus. The *HLA-DRA* and *HLA-DRB1* loci encode the α and β chains, respectively, which together form a mature class II HLA-DR molecule. Similarly, the products of the *HLA-DQA1* and *HLA-DQB1* loci encode the DQ molecule, and the *HLA-DPA1* and *HLA-DPB1* loci encode the DP molecule. A second DR molecule is encoded on most HLA haplotypes by the *HLA-DRA* and *HLA-DRB3*, *HLA-DRB4* or *HLA-DRB5* loci. Thus, most HLA haplotypes encode four distinct expressed class II molecules. The HLA

class I and class II molecules produced by this genetically complex system are highly polymorphic in the population, leading to a source of interindividual differences (Dupont, 1989).

The class I and class II genes mentioned above represent only about half of the known genes lying within the HLA region (Ragoussis et al., 1991; Kelly & Trowsdale, 1994). Among the remaining genes, several have interesting immunological functions that could be important in pathogenic events relating to HLAassociated autoimmune disease. One such region is the HLA class III region, a cluster of genes between the class I and class II complexes (Fig. 3.1). This region includes the genes for 21-hydroxylase and complement components C2, C4 and Bf (Carroll et al., 1984; White, 1989). Within the class II complex between the DQ and DP clusters lies a group of genes involved in antigen processing. The genes for a transporter, TAP, and for low molecular weight protein (LMP) are within this region. These polypeptides are implicated in HLA class I antigen processing and peptide transport (Spies et al., 1989; 1990; Deverson et al., 1990; Trowsdale et al., 1990). The DMA and DMB genes found in this region have a similar involvement in the class II antigen-processing pathway (Mellins et al., 1990). Other genes of particular interest include a collagen gene centromeric of the class II region, an hsp 70 gene found near the HLA class I complex and the cytokine genes for lymphotoxin, TNF β and TNF α , centromeric of the HLA-B locus (Carroll et al., 1987; Hanson et al., 1989; Sargent et al., 1989; Trowsdale, Ragoussis & Campbell, 1991).

The HLA region is, therefore, densely packed with genes important in the immune response. The polymorphic nature of these genes may contribute to differences in the immune response between individuals, and probably accounts for the HLA genetic linkage to a predisposition to autoimmunity. The structure and function of the proteins that these genes encode have been elucidated by crystallography and cellular biology. This information has deepened our understanding of how the HLA locus is associated with autoimmunity and the possible mechanisms by which this region contributes to the development of autoimmunity.

The structure of HLA molecules and the trimolecular complex

The crystal structure of both class I and class II molecules has been solved, allowing us a look at the structure of these HLA proteins. Both molecules are heterodimers with a peptide-binding site defined by an eight-stranded β sheet and two α helixes. For class I, a constant chain β_2 -microglobulin combines with either HLA-A, HLA-B, or HLA-C heavy chain to form a binding pocket in which peptides of only 8–10 amino acid residues in length are able to bind (Fig. 3.2*a*) (Madden *et al.*, 1991; Madden, Garboczi & Wiley, 1993). The class II proteins



Fig. 3.2. Computer-generated models of HLA molecules based on known crystal structures (Kraulis, 1991). (*a*) A model of the HLA class I heavy chain molecule. Two views of the HLA-A*0201/TAX peptide complex are shown (Garboczi *et al.*, 1996). The top view has the α_2 helix on the bottom and the α_1 helix on the top with peptide amino terminus on the left end of the binding pocket. The bottom view includes the α_2 helix on the right and the α_1 helix on the left, with the peptide amino terminus coming out of the page. (*b*) A model of HLA class II molecule. Two views of the DR1/HA peptide are shown (Stern *et al.*, 1994). In the top view, the DR α chain forms the top α helix, and the DR β chain forms the bottom α helix; the peptide is bound with the amino terminus on the left. In the bottom view, the DR α chain is shown on the left, the DR β chain on the right and the peptide amino terminus is coming out of the page. The peptide extends beyond the groove, unlike the class I molecule.

include α and β chains, which combine to form a more open binding groove, allowing peptides of greater length to bind in the groove (Fig. 3.2*b*) (Stern *et al.*, 1994). The crystal structure of class I and class II molecules has also demonstrated that within the binding groove there are pockets which act as anchors for the peptide; these pockets favour interactions with specific amino acids based on size and polarity. Interestingly, those pockets that are essential for peptide-binding specificity are the regions where the most polymorphic residues are found.



Fig. 3.3. Model of the trimolecular complex based on the crystal structure of HLA-A2–TAX–TCR A6 (Garboczi *et al.*, 1996). This model demonstrates the interaction of the TCR with both the polymorphic regions of the HLA molecule and the peptide.

The HLA class I and class II molecules are found on the surface of cells and interact with CD8⁺ and CD4⁺ T cells, respectively, via the T cell receptor (TCR). The complex formed in this interaction – HLA molecule, bound peptide in the groove and the TCR – is referred to as the trimolecular complex. Here the crystal structure is enlightening as well. The crystal structure of the class I molecule HLA-A2 has been solved with a peptide TAX and the TCR A6 (Garboczi *et al.*, 1996). This structure demonstrates that the TCR has a relatively flat surface that interacts with the class I molecule and peptide in such a way as to contact both peptide and HLA molecule (Fig. 3.3). Of importance is the finding that half of the contact sites are with peptide residues, while the other contact sites are with the class I molecule and include polymorphic residues. This demonstrates that a TCR is not only peptide specific but is also specific for self-MHC. Indeed, the peptide may be altered as long as the residues that contact the TCR are not altered, allowing for the possibility of several different peptides being able to interact with the TCR in the context of one HLA molecule.

These studies of the structure of HLA class I and class II molecules

demonstrate that the polymorphisms found in these genes encode the region important for T cell contact and for the peptide bound in the groove, leading to a unique surface appearance to the TCR for each HLA allele. At the level of the gene, the importance of these polymorphisms is demonstrated by the fact that the HLA class II genes share sequence over large regions, intermixed with regions of polymorphism, that correlate with T cell contact or with the binding pockets. These regions of polymorphism seem to be like 'cassettes' that are shuffled between alleles. An example of this concept can be found in the *DRB* locus, as shown in Fig. 3.4. Not only are these cassettes found among the *DRB* loci of humans, but they can be seen in other primates, suggesting that they play an important role in the immune response and have been conserved through evolution (Erlich & Gyllensten, 1991; Gaur *et al.*, 1997). Of further significance is the fact that these cassettes have been linked to disease susceptibility (see below).

Function of class I and class II genes

The function of the HLA class I and class II alleles are similar. Both molecules form heterodimers on the cell surface with peptide bound in the binding groove which forms a recognition signal for the TCR. Binding and activation of an antigen-specific T cell requires both HLA molecule and bound peptide. Class I and class II molecules differ in several ways. The HLA class I molecule contains a binding site for recognition by the T cell co-receptor known as CD8 (Rosenstein et al., 1989) and the class II molecule contains a binding site preferential for the T-cell differentiation antigen CD4 (Mittler et al., 1989). Binding of CD8 to the class I molecule achieves at least two things: it increases the affinity of interaction between T cell and antigen-presenting cell, which facilitates activation, and it provides for specificity by focusing the attention of CD8+ T cells on antigens presented via the class I pathway (Garcia et al., 1996; Gao et al., 1997). The binding of the CD4 molecule to the class II molecule acts in a similar manner by increasing the affinity of interaction between T cell and the class II-peptide complex and it facilitates activation (Mittler et al., 1989). Therefore, the engagement of the HLA-peptide complex with the TCR leads to the formation of the trimolecular complex and to the generation of the T cell response. The specificity of this interaction is dependent on the recognition of the self-HLA molecule and a specific peptide bound in the cleft.

The primary function of HLA molecules in the immune activation pathway is to bind antigenic peptides for presentation to a T cell. Therefore, the ability of an antigenic peptide to be bound by an HLA molecule is a primary determinant of whether or not an immune response to that peptide can be generated. The question of whether or not a peptide will satisfactorily bind to an HLA molecule is

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HLA DRB1*	6 0										7 0								7 8
0101	Y	w	N	s	Q	K	D	L	L	E	Q	R	R	А	A	V	D	Т	Y
0401												К					150		
0404										1		100			1	3			
0405											10								
1402									6										
0402								1			D	Е							
1301								14		0	D	E				200			
0801								F			D		2		L				
1101								F	1		D								
0701	S							I			D		G	Q			-		
0301	S										10	K	G	Q			N		
1501								1				А							

Fig. 3.4. The single letter amino acid code for residues 60–78 is given for several *HLA-DRB1* alleles. The shaded region is the third hypervariable region of the class II molecule. Those alleles producing similar sequences are grouped together. Notably these patterns fall into groups; the sequence shared by *0101, *0401, *0404, *0405 and *1402 is found only in alleles associated with RA and is referred to as 'the shared epitope'.

determined by the amino acid residues on the peptide in combination with the amino acid residues of the binding pockets of the HLA molecule (Buus, Sette & Grey, 1987; Krieger *et al.*, 1991). This structural interaction between an antigenic peptide and the HLA peptide-binding groove is one of the key ways in which HLA genes control the immune response. Only a few peptides have the potential to be immunogenic, by virtue of their ability to bind to an HLA molecule. The genetic control in this system is achieved by allelic variation. The principle polymorphic regions of HLA molecules include residues in the peptide-binding groove. Therefore, individual variation in the HLA molecule caused by genetic polymorphism determine which peptides are antigenic in one individual but are not in another individual.

The structural interaction between a peptide and the HLA peptide-binding groove determines which peptides are allowed to bind but it does not determine which peptides are actually available to bind. This aspect of peptide binding is determined by the intracellular events of antigen processing. Antigen processing takes place within a series of specialized compartments in the cell. Within this compartmentalized pathway, HLA molecules assemble, bind peptide and move to the cell surface where interactions with selected T cells occur (reviewed in Braciale & Braciale (1991) and Neefjes & Ploegh (1992)). Peptide comes to be bound by class I or II molecules via distinct processing pathways.

After synthesis, the class I heavy chain is transported to the endoplasmic reticulum (ER). In the ER, class I heavy chain forms a heterodimer with β_2 -microglobulin and binds peptides derived from cytosolic proteins. These proteins have been partially degraded by proteasomes (which utilize the MHC-encoded *LMP* gene products) and make their way as proteolytically cleaved peptides into the ER in a form suitable for binding to class I molecules (Belich & Trowsdale, 1995). This movement of peptides into the ER is accomplished by a transporter molecule, TAP, which is itself encoded by genes within the MHC complex (Spies & de Mars, 1991). The interaction of the class I heavy chain and peptide in the ER results in transport of the class I-peptide complex into the Golgi and post-Golgi compartments on the way to the plasma membrane. In general, as a result of these features of the class I pathway, the peptides bound by class I molecules predominantly derive from self-peptides, viral or other endogenously synthesized proteins.

MHC class II molecules are synthesized in the ER, where the invariant (Ii) chain then aids in the assembly and transport of the nascent class II into the lysosome-like MIIC compartment and other endosomal compartments (Peters et al., 1991). During this process the Ii is cleaved, leaving a peptide, CLIP, in the binding cleft. Proteins enter the cell via pinocytosis or receptor-mediated endocytosis. They then enter the acidic environment of the endosomes where proteases digest them into peptides. These peptides and the class II heterodimers both localize to the MIIC where CLIP is replaced by exogenous peptide in the MHC class II binding groove (Pieters, 1997). The non-classical class II molecule DM is thought to facilitate the removal of CLIP and the binding of peptides in this compartment (Sanderson et al., 1994; Kropshofer et al., 1997). The newly formed HLA-peptide complex then moves to the cell surface, possibly via endosomal fusion, where it becomes a mediator of T cell activation. As a result of these features of class II antigen processing, exogenous peptides are bound by class II molecules, although it has been demonstrated that class II molecules do present endogenous peptides as well (Nuchtern et al., 1990; Chicz et al., 1992).

Epidemiology of disease association

The association of HLA with several of the rheumatic diseases has been well recognized for some time (Tiwari & Terasaki, 1985a). This evidence has come from family, twin and population studies. Initially this work correlated serological specificities associated with HLA polymorphisms with patients and with con-

Disease	HLA-associate	ed Race	Relat	ive References
	allele		risk	
Ankylosing spondylitis	B27		90	Khan and Kellner (1992)
Reiter's syndrome	B27		41	Khan and Kellner (1992)
Inflammatory bowel disease	B27		10	Khan and Kellner (1992)
Psoriatic arthritis	<i>B</i> 27		10	Khan and Kellner (1992)
Rheumatoid arthritis	DRB1*0101	Israeli Jews	6	Gao et al. (1991)
	DRB1*0401	Caucasians	6	Nepom et al. (1989)
	DRB1*0404	Caucasians	5	
	DRB1*0405	Japanese	3.5	Ohta et al. (1982);
	DRB1*1402	Yakima Indians	3.3	Willkens et al. (1991)
Systemic lupus	DR2	Caucasians	3	Arnett and Reveille
	DR3		3	(1992)
Sjögren's syndrome	DR3		6	Arnett et al. (1989)
Pauciarticular	DR8		5	Nepom (1991)
juvenile rheumatoid	DR5		4.5	
arthritis	DP2.1		4	
Polyarticular	DRB1*0401		7	Nepom et al. (1984)
juvenile rheumatoid arthritis	DRBI*0404			
Juvenile	DR3		6	-
dermatomyositis				

Table 3.1. Some autoimmune diseases which have HLA associations and the relative risk conferred by the associated HLA gene

trols. More recent studies have used molecular techniques to identify specific haplotypes and, in some cases, specific genes that account for these associations. Table 3.1 lists the best characterized examples of rheumatic disorders associated with HLA genes. The strength of the association of HLA and disease is reflected in the term 'relative risk', which is an odds ratio representing the risk of disease in an individual carrying a particular genetic marker compared with the risk in individuals in that population without that marker.

In many rheumatic diseases there is an increased frequency of the disease found among family members of affected individuals, suggesting a genetic link. This has been seen in SLE, where a patient with SLE has a 10-fold increased likelihood of having a first-degree relative with SLE (Arnett & Reveille, 1992); similar results are seen in RA (Deighton & Walker, 1991) and

multiple sclerosis (Ebers et al., 1986). Studies of monozygotic and dizygotic twins also support a strong genetic component to these diseases. The concordance rate for monozygotic twins in SLE is 24-69% compared with dizygotic twins, where the rate is 2-9% (Deapen et al., 1992). In RA, twin studies have shown that 30-50% of identical twin pairs will be concordant for RA, compared with fraternal twins, which have 3-5% concordance rate (Winchester, 1981). These findings confirm a strong genetic basis for these diseases and suggest that the genetic contribution to these diseases is polygenic in nature, and not a simple single gene trait. However, a lack of complete penetrance in the genetically identical siblings also implicates other factors (Winchester et al., 1992). These factors could include environmental exposures, both infectious and chemical, and stochastic events. The 30-50% concordance rate in identical twins is found in many autoimmune diseases. This finding may occur because, although identical twins have identical germ-line genes, they do not have identical immune systems since much of the immune system is developed along stochastic lines. Therefore, recombination events such as the formation of the TCR and immunoglobulins may be implicated as factors in the development of autoimmunity.

Population studies have been a further source for evaluation of the link between the HLA locus and autoimmunity. These studies have involved HLA typing patients with disease and controls taken from the population. When an allele is found to be more frequent in the disease population it is thought to be associated with the disease. Population studies allow the study of large groups of individuals and have broad applicability of findings. However, interpretation of findings may differ between ethnic groups. The strength of an association may differ based on the disease frequency and the baseline frequency of the susceptibility allele in the population.

An example of this is the *HLA-DR4* association with RA. In Caucasians, there is a frequency of disease that reaches 1%, and the presence of *DR4* in the general population is 23%. Of those affected by RA in this population 70% are *DR4*, thus demonstrating a clear association (Nepom & Nepom, 1992). However, in the African American population of the USA, the frequency of RA is similar to whites, but the presence of *DR4* in this population is lower, at 14%. In this population, *DR4* is found in 45% of patients with RA. This is considerably lower than the number of *DR4*-positive Caucasian RA patients; however, when the low overall frequency of *DR4* in the population is taken into account the association between disease and *DR4* is still present (Karr *et al.*, 1980).

Interpretation of the epidemiological data is complicated by several phenomenon that may lead to a demonstrable HLA association but also lead to differing interpretations of the data. These include variation in ethnic groups (as discussed above), allelic differences, linkage disequilibrium and *trans* complementing dimers.

Most HLA alleles within a single locus are highly related. Therefore, when a disease is associated with a specific allele within a single locus, differences between alleles may be quite small, as little as one nucleotide difference leading to an alteration in one amino acid residue on the entire molecule. As was discussed above, these polymorphisms play an important part in the structure at the site of T cell interaction and peptide binding. Therefore, it is important to identify differences between alleles because they have direct implications for the structure and function of the HLA molecules. An example of this is the association of both RA and pemphigus vulgaris with HLA-DR4. DR4 is marker for a family of alleles that are quite similar; however, if the association of DR4 with these two diseases is looked at at the allelic level important differences are found. RA is associated with DRB1*alleles 0401, 0404, 0405 and 0408 (Nepom & Nepom, 1992), whereas pemphigus is found to be associated with DRB1*0402 (Ahmed et al., 1990; Scharf et al., 1988a; Scharf, Long & Erlich, 1988b). Table 3.2 demonstrates the significant polymorphism between these alleles. The RA-associated alleles share a strong similarity at the β 67, β 70, β 71 residues, which differ in size and charge on *DRB1*0402*. The aspartic acid and glutamic acid at positions 70 and 71, respectively, both carry a negative charge, unlike those positions in the RA-associated allelic products, which include a glutamine at β 70 and a positive residue at β 71, either a lysine or an arginine. These polymorphisms are structurally and functionally significant and link the genetic variability with disease-specific properties.

Genetic association must also be interpreted with the understanding that HLA genes are physically linked to one another. A high degree of linkage disequilibrium found among HLA loci means that different alleles at the different HLA loci do not randomly assort in each generation but rather exist on haplotypes that are relatively fixed in the population. Therefore, inheritance of a specific allele at one locus is often accompanied by the inheritance of a specific allele at a nearby linked locus. This means that when a polymorphic HLA marker is noted to be associated with a particular disease, the actual susceptibility gene may correspond to a linked gene elsewhere on the haplotype.

This may be the case with the HLA association of multiple sclerosis. Multiple sclerosis has been associated with the DR2 haplotype in Caucasians (Tiwari & Terasaki, 1985b). This haplotype includes DRB1*1501, DRB5*0101, DQA1*0102 and DQB1*0602. These genes are usually found as linked alleles in patients with multiple sclerosis (Allen *et al.*, 1994). Therefore, the high degree of linkage disequilibrium does not allow a distinction to be made between the contribution of individual genes in the DRB1 and that of the DQB1 region to the risk for multiple sclerosis.

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DRB1* alleles	Residue position							
-	57	67	70	71	86			
0401	D	L	Q	K	G			
0402	D	Ι	D	Ε	V			
0404	D	L	Q	R	V			
0405	S	L	Q	R	G			
0408	D	L	Q	R	G			

Table 3.2. Comparison of amino acid residues produced by DR4 alleles

Another phenomenon that must be taken into consideration is that of transcomplementary dimers. Most individuals in the population are heterozygotes, and, therefore, an individual's HLA phenotype is determined by two HLA haplotypes. Most individuals express six class I molecules and at least eight class II molecules. A further source of diversity in individuals heterozygous for class II haplotypes is provided by *trans*-complementation. *Trans*-complementation occurs when the HLA class II molecule α - β heterodimer is formed from an α chain encoded on one haplotype and a β chain encoded on the other haplotype, creating novel dimers in heterozygotes. For example, an individual heterozygous for the DQA1*0501 and DQB1*0201 genes on one haplotype, and the DQA1*0301 and DQB1*0302 genes on the other haplotype, will actually express four, not two, different DQ heterodimers resulting from the combinatorial association of each of the α and β chains. *Trans*-complementing dimers cannot occur with all combinations of α and β chains in the class II complex, so the extent of the additional diversity provided by this mechanism depends on the specific haplotypes in each individual (Kwok et al., 1988).

One such example is coeliac disease. This disease has been associated with HLA-DR3 and to a lesser extent with HLA-DR7 (Kagnoff *et al.*, 1989). The DR3 haplotype is relatively fixed as DQB1*0201 DQA1*0501 DRB1*0301, while the DR7 haplotype shares the DQB1*0201 allele (in common with DR3) but not the DQA1*0501 allele. However, a high frequency of patients who are not DR3 carry both DR7 and DR5. The DR5 haplotype includes DQ1*0301 and DQA1*0501; as a result when the DR7 haplotype is also present a *trans*-complementing DQ heterodimer DQA1*0501 DQB1*0201 can be formed in DR5/DR7 heterozygotes (Sollid *et al.*, 1989). When population studies are evaluated in this light, 90% of patients with coeliac disease have DOA1*0501 DOB1*0201, demonstrating a clear association (Fig. 3.5).

When these factors are considered, many clear associations with the HLA locus



Fig. 3.5. *Trans*-complementing dimers in the susceptibility to coeliac disease. The DQ heterodimer DQB*0201 DQA*0501 is associated with coeliac disease. In individuals heterozygous for *DR7* and *DR5*, three combinations of *DQB* and *DQA* alleles are possible through *trans*-complementation, one of which is the heterodimer DQB*0201 DQA*0501.

and in some cases specific genes can be shown. With this knowledge the mechanisms by which these genes confer increased risk can be investigated.

Mechanisms by which HLA may contribute to autoimmunity

Genetic associations with autoimmunity can be applied to clinical use for prediction and prognostic purposes and can also be used as a means to define the mechanism by which these genes lead to autoimmunity. The structure of the HLA molecule and its role in the formation of the trimolecular complex give us clues to follow. In the remaining part of this chapter we will discuss the hypotheses that have been generated based on this information and summarize data that support these hypotheses.

Disease-associated alleles give rise to unique peptide binding

As discussed above, the function of HLA class I and class II heterodimers is to bind peptides in their binding groove and interact with the TCR. This then leads to signalling within the T cell and activation of the immune response. Since this is the primary immunological function of these molecules, it is reasonable to consider the ability of specific allelic products to present a pathogenic peptide as the basis by which they contribute to autoimmunity. This hypothesis would require evidence that allelic differences in HLA molecules lead to differences in the peptides which are bound, and the identification of pathogenic peptides.

The ability of a peptide to be bound in the groove of an HLA molecule is determined by the residues lining the pockets of the binding groove (as demonstrated by the crystal structure). These pockets determine a 'motif' for the amino acid residues needed or tolerated within the groove. Allelic polymorphisms occur at these sites and, therefore, alter the binding pockets and the binding motifs. Investigation into binding motifs has been carried out for many of the class I and class II alleles (Rammensee, Friede & Stevanoviic, 1995). These studies have shown that allelic differences do alter the peptide-binding motifs. An example of these differences can be demonstrated with the motif produced by the DR4alleles 0401, 0402 and 0404. As described above, these DR4 molecules are similar except at several residues located on the β chain (Table 3.2 above). These differences are most profound at residues 70 and 71, where 0401 and 0404 both have a glutamine at 70 and a positive residue at position 71. These residues line pocket 4, and motif studies have demonstrated that pocket 4 in these alleles favours a negatively charged residue. In contrast, the 0402 molecule carries a negative charge at both position 70 and 71, leading to an alteration in motif, with a preference for a positive charge at pocket 4 (Table 3.3). In this way, these alleles favour binding of different peptides and, therefore, generate different antigen-specific responses. In this instance, the allelic polymorphism, which is disease-associated, demonstrates a functional difference which can be associated with disease (Wucherpfennig & Strominger, 1995).

If disease is caused by the binding of a pathogenic peptide, then understanding of these motifs, in theory, could lead to the definition of important autoantigens. In diseases with known HLA associations, suspect autoantigens can be screened for peptides compatible with the motif of disease-associated allelic products.

This type of work has been done in the setting of pemphigus vulgaris and the

DRB1*	DRb71	Pocket 1 binding preference	Pocket 4 binding preference	Pocket 6 binding preference	Disease association
0401	K (positive)	V, L, I, M, F	D, E, (negative)	S, T, N, V	Rheumatoid arthritis
0402	E (negative)	V, L, I, M, F	K, R (positive)	S, T, N, V	Pemphigus vulgaris
0404	R (positive)	V, L, I, M, F	D, E, (negative)	S, T, N, V	Rheumatoid arthritis

Table 3.3. Binding motifs formed by DR4 alleles

*DRB1*0402* allele. Pemphigus is an immune-mediated, blistering disease of the skin. Patients with phemphigus vulgaris frequently carry the *DRB1*0402* allele. These patients have a specific antibody response to desmoglian, a skin protein, and this antibody response is directly implicated in the pathogenesis of the disease (Merlob *et al.*, 1986). Using the predicted binding motif of 0402 to scan the protein desmoglian for a peptide that could bind 0402, a peptide from this protein was identified that had the predicted binding motif for 0402, and in four patients T cells responsive to this peptide have been demonstrated. This finding suggests that an immune response to this self-peptide may be a component of the autoimmune phenomenon leading to disease, and that this occurs in 0402 individuals (Wucherpfennig *et al.*, 1995).

As described above, the motif is important in determining what peptides are allowed to bind, but which peptides are actually available to bind is determined by antigen processing. The mechanisms by which antigens are processed are being elucidated and at this time are not completely understood. However, it is clear that antigen processing may be important in determining the peptides available for binding, particularly since aspects of these processes may have allelic specificity. Interestingly, many of the proteins involved in processing are encoded by genes on the HLA locus. Several factors important in processing have been shown to be allele specific, including the effect of different intracellular environments such as pH, affinity for the chaperone CLIP, and the stability of the heterodimer. A specific example of this type has been suggested by Auger and Roudier. They have shown that a member of the heat shock protein family HSP73 binds DR4 molecules and may act as a molecular chaperone. This may lead to an alteration in the processing pathway for DR4 molecules with changes in the set of peptides accessible to this group of alleles (Auger *et al.*, 1996).

These concepts also suggest another method by which differences between alleles in the ability to bind a peptide may influence autoimmunity. In these scenarios instead of a pathogenic peptide being bound in the disease-associated allele, a protective epitope may be bound and presented in other alleles, leaving individuals without the protective allele at risk.

Unique MHC-TCR interactions

The crystal structure of the trimolecular complex discussed above demonstrates that the surface of the HLA molecule–peptide complex seen by the TCR is relatively flat. Contact with the TCR occurs via both the residues of the peptide and the components of the α helix of the HLA molecule. Therefore, class I and class II molecules may influence the immune response via direct contact between the TCR and HLA residues. One such mechanism would require direct interaction between the polymorphic residues on the HLA molecule with the TCR, leading to selection and activation of autoreactive T cells. The crystal structure of TCR–peptide–class I demonstrates interaction between the polymorphic sites on the class I protein and the TCR, and although a ternary complex has not yet been crystallized for class II it is likely from what is known of the structure that polymorphic residues will be in contact with the TCR in this setting as well. This has been demonstrated with alloclones, which specifically 'see' polymorphic residues on DRB1*0404, this recognition is lost on DRB1 mutant proteins, which differ only by one residue (Hiraiwa *et al.*, 1990; Penzotti *et al.*, 1996).

Another mechanism by which a direct interaction between the TCR and HLA molecule might occur is in the presence of a bacterial superantigen. Superantigens are molecules that are able to interact with the MHC and the TCR outside of the peptide-binding groove, usually using a common structural component from the TCR V β region (Dellabona *et al.*, 1990). These molecules then lead to an immune response that is not antigen specific. If a superantigen was specific for a class I or class II allele, it could lead to an amplification of autoreactive T cells in those patients with the disease-associated HLA type and initiation of autoimmunity. Such a mechanism might help explain the link between the genetic and environmental influences in autoimmunity.

Yet another hypothesis to be considered when thinking of HLA–TCR interactions is the ability of two closely related alleles to produce proteins that interact with the same TCR. Many HLA alleles are closely related and their products may be able to bind the same peptides. The surface of these molecules may be similar enough to be recognized by a TCR yet include polymorphisms on the HLA molecule that could subtly alter the interaction, leading to differences in signalling and possibly the immune outcome of the response (Madrenas & Germain, 1996). This type of mechanism could come into play in individuals who are heterozygous for two closely related HLA alleles, such as 0401 and 0404. A T cell may be positively selected in the thymus on 0401, but recognition of 0404 with a self-peptide may lead to activation and an autoimmune response. In this model, heterozygosity is important, as has been demonstrated in RA where most severe disease is found in individuals who are DRB1*0401/DRB1*0404 (Weyand et al., 1992).

Molecular mimicry

A model referred to as molecular mimicry has been suggested to explain the association of HLA with some autoimmune diseases. This model suggests that the contribution to the disease-associated region of the HLA complex is not functioning as a class I or class II molecule but is itself presented as a peptide on class II. In the example of RA, this would mean that the *HLA-DR*-encoded 'shared epitope' would be the peptide bound by other class II molecules and would lead to recognition by an autoreactive T cell. Although direct evidence of this phenomenon has not been demonstrated in disease, several lines of evidence suggest that it may be possible. Elution of peptides from HLA class II molecules have revealed that, although an important role of HLA class II is to present exogenous peptides to CD4⁺ T cells, the majority of peptides found bound to class II are derived from (membrane-bound) endogenous proteins, and some of these peptides are from the polymorphic regions of the MHC molecules (Chicz *et al.*, 1993; Rammensee *et al.*, 1995).

Evidence that self-peptides derived from the polymorphic regions of HLA molecules generate an immune response in humans has been demonstrated (Liu et al., 1992; Salva et al., 1994). These endogenous peptides may well play a role in the immune response, either at the level of T cell repertoire selection or later in immune modulation. In these cases, of course, studies would show linkage or association with the MHC. One trigger for mimicry could be an infection. Searches for infectious agents that have proteins homologous to these regions have been done. In the case of HLA B27, several proteins derived from *Klebsiella* pneumoniae and a Shigella sp. plasmid have been described (Schwimmbeck, Yu & Oldstone, 1987; Stieglitz, Fosmire & Lipsky, 1989) and the 'shared epitope' of RA has been shown to be present both in an Epstein-Barr virus protein and in a protein found in Escherichia coli (La Cava et al., 1995). In this setting, a patient's ability to continue presenting a peptide similar to that of the infectious agent may lead to persistence of inflammation. These data raise the possibility that the polymorphic regions of self-proteins may act as peptides that are a factor in the protection from, or the initiation and propagation of, autoimmunity.

HLA genes other than those for class I and II may contribute to disease association

Other genes on the MHC certainly may contribute to autoimmunity. As discussed above, only half of the known genes found within the HLA locus encode HLA class I and II. Several of these other genes are important in immunological function, such as the cytokine genes for TNF β and TNF α , genes involved in processing and genes of the class II region, which includes the complement components. In these cases, an HLA association may result from linkage with nearby non-HLA genes. Some forms of SLE may represent this type of disease association (Arnett et al., 1984). Both HLA-DR2 and HLA-DR3 haplotypes are associated with SLE in Caucasians, and the predominant HLA-DR3+ haplotype in Caucasians has very tight linkage with other genes within the MHC, notably Al, B8, Cw7 and C4A. This last gene, a structural gene for the C4 complement component, is a silent allele on this DR3 haplotype. The C4A 'null' alleles are significantly increased in patients with SLE, even on non-DR3 haplotypes, and particularly when the patient is also HLA-DR2+ (Fielder et al., 1983; Christiansen et al., 1991). This relationship may indicate that it is the presence of a C4A silent allele, rather than the expression of any particular class II gene, which is the primary predisposing genetic element in these patients.

Case studies in rheumatological disease

To demonstrate the concepts discussed above, we will discuss two rheumatic diseases with well known HLA associations. The possible mechanisms of disease can be explored for each of these diseases, although the relationship to HLA is different for each.

HLA-B27 and spondyloarthropathies

The association of HLA-B27 with the spondyloarthropathies is well established. Initial population studies in patients with ankylosing spondylitis demonstrated a marked predominance of HLA-B27 in the population with disease (90%) compared with the control population (10% Caucasians) (Benjamin & Parham, 1990). This association has been confirmed in different racial and ethnic groups (Khan & Kellner, 1992). To further establish the importance of the gene in development of this disease, a transgenic animal model has been developed in which the HLA-B27 molecule is expressed in the rat (Hammer *et al.*, 1990). In this model, the rats which express HLA-B27 develop a disease that bears striking similarity to spondyloarthropathies, directly validating the role of HLA-B27 in development of disease.

Another finding in the rat model confirms data found in humans. This is the finding that disease develops only in the rats that are exposed to infections (Taurog *et al.*, 1994), similar to reactive arthritis in humans (a form of spondylo-arthropathy), which has been clearly associated with preceding infectious diarrhoea (Hahn, 1993). These findings support the concept that both an environmental agent and HLA are important in this disease. However, the pathogenic mechanism by which B27 triggers disease is not yet known. Possible mechanisms include (i) the binding of a pathogenic peptide to HLA-B27, which leads to a CD8⁺ CTL response that causes joint inflammation; (ii) molecular mimicry, in which a bacterial peptide similar to a peptide found on the HLA-B27 heavy chain drives an immune response that persists after infection because of the persistent presence of the peptide from the HLA-B27 molecule itself; and (iii) HLA-B27 having a unique immunological role that is not yet defined.

Evidence to support the idea that the HLA-B27 molecule can bind a unique pathogenic peptide is supported by the finding that several subtypes of HLA-B27 exist (Khan & Kellner, 1992). These subtypes are all quite similar and all are associated with disease. These class I alleles share strong similarity in their peptide-binding pockets, particularly P2 (arginine site) (Khan & Kellner, 1992). An exception to this is the allele B*2703, which is found in West Africans and African Americans. Spondyloarthropathies are rare among these groups and this rarity may result from the differences found on the HLA-B*2703 allele, which differs from the most common allele B*2705 in giving a single residue change at position 59 (tyrosine to histidine) (Hill et al., 1991). Molecular mimicry models for HLA-B27 are supported by the finding that several pathogens share similar amino acid sequences with HLA-B27 AA71-77. A more novel mechanism involving a less well-defined function of the HLA B27 heavy chain is suggested by the finding that HLA-B27 transgenic mice, which do not express the β_2 microglobulin chain, still develop disease (Khare et al., 1995). The mice are unable to form stable HLA-B27 heterodimers on the cell surface yet still get disease, suggesting a role that may not require the β_2 -microglobulin chain for this molecule, such as HLA-B27 acting as a port of entry into the cell for pathogens or as a unique receptor on the cell surface.

RA and the shared epitope

Unlike the spondyloarthropathies, RA is a disease in which the HLA association has been mapped not to one allele but to a specific amino acid sequence found in several DR allelic forms. This sequence is referred to as the shared epitope. Population studies have demonstrated an association with *DRB1*0401* and *DRB1*0404* in the Caucasian population. Studies in several ethnic groups that have a low prevalence of *DR4* have shown other *DR* associations including: *DRB1*0101* in Israel, *DRB1*0405* among Japanese and *DRB1*1402* in the people of the Yakima Nation (Willkens *et al.*, 1991; Ollier & Thomson, 1992). Remarkably, each of these HLA alleles have in common the β 67–74 region. As shown in Fig. 3.4, this region is not found on any other haplotype and although all these RA-associated alleles carry the same shared epitope, they differ in other regions of the HLA molecule. Therefore, it appears that it is the shared epitope itself, rather than any specific single HLA allele, that is the principle determinant of the genetic susceptibility for RA.

As discussed for HLA-B27, similar theories for the role of the associated HLA molecules in RA have been proposed: the ability of these molecules to bind a unique arthritogenic peptide, the shared epitope presenting itself as a peptide, leading to molecular mimicry, and a unique immunological interaction with the T cell by the disease-associated molecules. However, in RA these ideas need to be viewed differently. In RA, the idea that a unique peptide is bound in the RA-associated alleles can be supported by some similarities in the binding motifs of the molecules in pocket 4 (Woulfe et al., 1995). However, unlike the HLA-B27 alleles, the similarity between the associated molecules is not over the entire binding groove but is in one region of the class II β chain only (Fig. 3.6). This region includes the α helix. Other regions of the binding groove are not conserved between the associated alleles; therefore differences in binding motif occur in the disease-associated alleles. The molecular mimicry hypothesis, as in HLA-B27, is supported by the finding that there are bacterial sequences which are similar to the amino acid sequence found in the region of the shared epitope. The position of the shared epitope on the α helix also suggests that it may lead to unique interactions with the T cell, as discussed above. Hopefully, as our understanding of the importance of allelic differences on the immune response broadens, we can clarify the underlying mechanisms involved with the HLA molecule and diseases such as RA and the spondyloarthropathies.

Future directions in research and therapy

Understanding the association of HLA and autoimmune disease has led to new areas of investigation into the pathogenesis of these diseases. It has also had an impact on clinical practice. The use of these genetic markers to identify members of the general population at risk is not feasible, since these HLA types are found in many more unaffected individuals than those affected by autoimmunity. However, use of these genetic markers is already helpful in assessing patients with arthritis. In the case of a patient presenting with signs and symptoms that



Fig. 3.6. Location of the shared epitope. The backbone trace of the DR1 variable domain is shown with the amino acid residues 67–74 highlighted as a ribbon. This region is the location of the shared epitope on DR molecules.

may be consistent with spondyloarthropathy, information regarding the presence of HLA-B27 can help determine the likelihood of the diagnosis of spondyloarthropathy. This must always be done with the knowledge of the prevalence of the allele in the general population. In RA, the role of HLA typing is not helpful for diagnosis but may be useful for prognostic purposes and therapeutic decisions. It has been suggested that the presence of the shared epitope is associated with severity of disease, particularly having both the 0401 and 0404 alleles (Weyand et al., 1992). Knowledge that the patient may expect more severe disease, particularly joint destruction, may lead to more aggressive early treatment. Response to therapy may also be influenced by a patient's HLA type. In a study comparing the response to methotrexate or combination therapy, patients carrying the shared epitope did better on combination therapy (O'Dell et al., 1998). In this setting, knowledge of a patient's HLA type may help in the choice of initial therapies. Several new therapies are being developed to take advantage of our understanding of the structure and function of class II alleles; peptide-based therapies are being studied in multiple sclerosis and RA with the hope of interfering in the abnormal immune response to self by altering the peptides presented via the HLA molecules. Understanding the specific genes and the mechanisms by which they predispose to disease, and the environmental factors that influence disease, will allow us to target diagnostics and therapeutics more specifically in the future.

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B cells: formation and structure of autoantibodies

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Introduction

Many autoimmune diseases are characterized by production of autoantibodies, and detection and measurement of these antibodies in patients' sera is used in diagnosis and monitoring of disease. For some autoantibodies, such as antibodies to double-stranded DNA (dsDNA) in SLE, or anti-acetylcholine receptor antibodies in myasthenia gravis, the autoantibodies appear to be involved in pathogenesis. Until recently, investigation in patients has been restricted largely to the mixed antibody populations in serum. However, using hybridoma and molecular biological technology, it is now possible to dissect out the individual antibodies from patients' sera and analyse specificity and molecular structure. By combining cloned and sequenced antibodies with the increasingly defined autoantigens, we are gaining insight into the features of autoantibodies that are responsible for pathogenicity. This review will describe our current understanding of antibody structure and the immunoglobulin (Ig) genes that encode the binding sites of autoantibodies. Knowledge at the genetic level may offer therapeutic opportunities based on rational design in order to block pathogenic interactions.

Variable region genes

V gene recombination

The events occurring in immunoglobulin genes during maturation of a B cell from a pro-B cell to a fully differentiated plasma cell are shown in Fig. 4.1. The genes encoding the heavy chains of immunoglobulin are highly unusual in having three separate genetic elements that must be combined prior to transcription (Tonegawa, 1983). Recombination takes place in two steps and is initiated by the lymphoid-specific RAG1 and RAG2 proteins (Gellert, 1992). Patients with



Fig 4.1. Changes in immunoglobulin occurring during normal B cell development. As B cells mature from the pluripotential stem cell to the fully differentiated plasma cell, immunoglobulin genes are recombined and modified. Following recombination, the pre-B cell expresses immunoglobulin μ chains with a surrogate light chain. Subsequently, light chains recombine and naive B cells express both IgM and IgD. Binding of antigen may then initiate further processes of somatic mutation and isotype switching, leading to plasma cells and memory cells.

certain rare forms of severe combined immunodeficiency (SCID) have defects in these proteins and consequently fail to generate immunoglobulin or T cell receptors (Kamachi *et al.*, 1993). Recombination begins at the pro-B cell level with the joining of a $D_{\rm H}$ segment gene to one of the six potentially functional $J_{\rm H}$ genes. Only a limited level of transcription occurs at this point until one of the available $V_{\rm H}$ genes combines to generate $V_{\rm H}$ - $D_{\rm H}$ - $J_{\rm H}$. Transcription is then activated by the proximity of the promoter upstream of $V_{\rm H}$ to the immunoglobulin enhancer sequence, and the cell at this pre-B stage can synthesize μ heavy chains. As maturation proceeds, a similar recombination event takes place for the light chain to create $V_{\rm L}$ - $J_{\rm L}$, and IgM is expressed (Fig. 4.1) at the immature B cell stage. Subsequent events following recognition of antigen are described below.

One feature of the joining process is that it may be imprecise, with gain or loss of nucleotides at the junctions, and in $V_{\rm H}$ the *D* segment gene may be read in different reading frames. Complexity at this site may be increased by *D–D* gene fusion or inversion, leading to a virtually unique sequence in the third complementarity-determining region (CDR3), which becomes the 'clonal signature' of a B cell. This sequence is maintained when a B cell clone undergoes expansion or neoplastic transformation and is proving useful in tracking tumour cells in patients after therapy (Steward, Potter & Oakhill, 1992). For normal B cells, the consequence of this potentially hazardous rearrangement of DNA is generation

of a wide range of sequences, with CDR3 positioned at the centre of the antibody combining site (Kirkham & Schroeder, 1994). In some cases, the rearrangement will produce non-functional sequences, and if the second chance provided by the allelic chromosome also fails, the cell will die.

V_H gene usage

The choice of $V_{\rm H}$ gene for recombination is from a repertoire of approximately 51 potentially functional genes available in unrearranged germ-line DNA (Cook & Tomlinson, 1995). These genes can be divided into seven families, $V_{\rm H}1-V_{\rm H}7$, with members of each family having >80% sequence similarity (Table 4.1). Size of family varies from a single gene segment ($V_{\rm H}6$) to 22 gene segments ($V_{\rm H}3$) (Cook & Tomlinson, 1995). The V κ and λ light chains have a similar family structure, with 32 (Schable & Zachau, 1993) and 24 (Williams & Winter, 1993) potentially functional genes identified, respectively. The nature of the V gene segment dictates the basic sequence of $V_{\rm H}$ and $V_{\rm L}$, although further diversification of sequence can occur by somatic hypermutation (see below). However, the sequence of the framework regions (FWs) tends to be conserved, and replaced amino acids often cluster in the CDRs, which are the known contact points for antigen (Fig. 4.2).

It has been difficult to estimate the relative use of different $V_{\rm H}$ genes by the normal expressed antibody repertoire, partly because of possible perturbation of gene usage by encounter with environmental antigens. In fact, studies using complementary DNA (cDNA) libraries (Logtenberg et al., 1992), or in situ hybridization (Guigou et al., 1990), have demonstrated that the expression of each $V_{\rm H}$ family is approximately equivalent to that expected from the available repertoire. However, recent analysis of single B cells from the blood of two normal subjects has shown that use of individual $V_{\rm H}$ genes varies, with the V_{3-23} gene (DP-47) being apparently overrepresented (Brezinschek et al., 1997). One explanation may be that this gene is duplicated in certain haplotypes (Sasso, Buckner & Suzuki, 1995). Increased usage of particular $V_{\rm H}$ genes also occurs as a result of stimulation by B cell superantigens (Goodglick & Braun, 1994). Examples of B cell superantigens are found among common pathogens, such as staphylococcal protein A, which binds to immunoglobulin encoded by genes from the $V_{\rm H}3$ family (Goodglick & Braun, 1994). Superantigen binding is to the FWs of the $V_{\rm H}$ sequence, rather than to the conventional sites in CDR1, CDR2 and CDR3 (Fig. 4.2). By this means, superantigens can stimulate a wide range of B cells, perhaps generating an early low-affinity IgM antibody response against invading pathogens.
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$V_{\rm H}$ family	Number of gene segments	
V _H 1	11	
$V_{\rm H}^2$	3	
V _H 3	22	
$V_{\rm H}4$	11	
$V_{\rm H}5$	2	
$V_{\rm H}6$	1	
$V_{\rm H}^{}$ 7	1	
Total	51	

Table 4.1. $V_{\rm H}$ content of the functional locus on chromosome 14_a32.3

Cook & Tomlinson, 1995



Fig. 4.2. Expanded view of the immunoglobulin heavy chain variable region (V_H) indicating the complementarity-determining regions (CDRs), known to be involved in high-affinity binding to conventional antigen. Also shown are the framework (FW) regions, which provide structural support for V_H , but which also may bind to certain 'superantigens'.

Somatic hypermutation

The naive mature IgM⁺ IgD⁺ B cell repertoire may undergo negative selection in the bone marrow environment, with removal of cells expressing immunoglobulin recognizing local autoantigens (Ghia *et al.*, 1995). This could be a crucial step in avoiding subsequent production of autoantibodies. Circulating B cells then can encounter antigen and enter the germinal centre of the lymphoid tissue (Fig. 4.1). In this site, the available range of V region sequence is extended by somatic hypermutation, which introduces mutations across $V_{\rm H}$ and $V_{\rm L}$ (Berek & Milstein, 1987). If the mutations generate replacement amino acids, they can be selected by limiting antigen, leading to a concentration of optimal amino acids in the contact points in the CDRs (Berek & Milstein, 1987). If mutations are deleterious, or if the cells are not selected, they will die by apoptosis (Liu *et al.*, 1989). Antigen-selected B cells will proliferate and, in the presence of CD4⁺ T cells and appropriate cytokines, undergo isotype switching (Berek & Milstein, 1987). Memory B cells and plasma cells are the final products of this maturation process.

Clearly, at this stage, where a new repertoire is being developed, there is a possibility of producing autoantibodies, some of which could be of high affinity. To avoid this, a second process of negative selection must operate. One of the mechanisms could be counter-selection against sequences containing mutations in CDRs that confer increased affinity for autoantigen. Intriguingly, this appears to occur in rheumatoid factors produced by normal individuals, but not in patients with RA (Borretzen et al., 1994). Studies in transgenic mouse models have shown that there are several alternative mechanisms that can operate to suppress autoantibody production, including inactivation or deletion of B cells (Shokat & Goodnow, 1995). At the levels of V genes, a further process, termed 'receptor editing', has been described, whereby a transgenic heavy and light chain pair from an autoanti-DNA antibody is rendered non-reactive by substitution of a rearranged endogenous V₁ sequence (Tiegs, Russell & Nemazee, 1993). Finally, high levels of soluble protein antigen can induce apoptosis in specific B cells (Shokat & Goodnow, 1995). Evidently there are many potential checks and balances to prevent autoimmunity, and these shape the B cell repertoire. However, the normal shaped repertoire includes B cells capable of secreting IgM autoantibodies. These B cells can produce significant levels of these antibodies under certain conditions, such as following infection with Epstein–Barr virus (EBV) (Chapman et al., 1993). One possible check point is at isotype switch, since these B cells may either fail to undergo isotype switch or lose autoantibody activity in the switched variant.

Isotype switching

The process of isotype switching is likely to take place close to the site of somatic mutation in the germinal centre. Isotype switching results from a deletional DNA recombination event in which the $C\mu$ constant region, initially located downstream of the *VDJ* region, is replaced by a $C\gamma$, $C\alpha$ or $C\varepsilon$ constant region

(Coffman, Lebman & Rothman, 1993). The deletional endpoints are within or close to repetitive switch region sequences situated upstream of each constant region gene (Coffman *et al.*, 1993). DNA between the switch sites is excised and can form a released 'switch circle'. Switching is influenced by cytokines, with IL-4, IFN- γ , and TGF β being particularly important. Patients with X-linked hyper-IgM syndrome, who lack CD40 ligand (CD40L) normally expressed by T cells in the germinal centre, make IgM but fail to produce germinal centres and to generate significant amounts of IgG and IgA. They also show minimal levels of somatic mutations, implicating CD40L⁺ T cells in both somatic mutation and isotype switching events (Di Santo *et al.*, 1993).

It appears that a considerable amount of somatic mutation can occur prior to isotype switch, and mutated IgM⁺ B cells can enter the blood (Klein, Kuppers & Rajewsky, 1994). However, there may be further accumulation of mutations after switching, and the change in the Fc region of the protein will influence the pathogenicity of autoantibodies. In fact, in autoimmune diseases such as SLE, pathogenic high-affinity antibodies tend to be of the IgG isotype. This points to a major role for CD4⁺ T cells in autoimmune disease, and there has been immense effort in investigating the T cell populations in various autoimmune diseases. In SLE, CD4⁺ T cell clones have been isolated that appear to recognize peptides from nucleosomes, and these clones are able to help B cells to produce anti-DNA antibodies (Mao *et al.*, 1994).

Autoantibodies in rheumatic diseases

The knowledge of B cell biology described in the first part of this chapter can be applied to the study of potentially pathogenic autoantibodies found in rheumatic disorders. In this section, we will discuss this with reference to two examples, anti-dsDNA and anti-phospholipid (aPL) antibodies.

Anti-dsDNA antibodies in SLE

Although many different autoantibodies can be found in the serum of patients with SLE, those binding to dsDNA are most closely linked with tissue damage, particularly glomerulonephritis (reviewed in Isenberg *et al.*, 1997). There is a considerable body of evidence to support this statement. While anti-dsDNA antibodies are found in approximately 75% of patients with SLE (Cervera *et al.* 1993), they are almost never seen in those with other diseases or in healthy individuals, including the normal relatives of patients with lupus (Christian & Elkon 1980; Isenberg *et al.*, 1985). The levels of these antibodies tend to be high when the disease is at its most active (Ter Borg *et al.*, 1990; Cervera *et al.*, 1993) and

they are deposited in the inflamed glomeruli of patients with lupus nephritis (Koffler, Schur & Kunkel, 1967).

In a number of animal models, it has been shown that monoclonal mouse or human anti-dsDNA antibodies can deposit in glomeruli to give proteinuria and nephritis (Madaio *et al.*, 1987; Raz *et al.*, 1989; Ehrenstein *et al.*, 1995). However, it is important to note that not all anti-dsDNA antibodies are equally pathogenic in such models. This correlates with the clinical data showing that a subset of human anti-dsDNA is particularly closely involved in renal damage in lupus. This subset consists primarily of IgG antibodies that bind specifically to dsDNA with high affinity. Okamura *et al.* (1993) demonstrated that in 40 patients with untreated lupus nephritis the degree of renal damage was related to the serum level of IgG anti-dsDNA but not to that of IgM anti-dsDNA or to anti-single stranded (ss)DNA of either isotype.

It is important to recognize that the high-affinity antibodies associated with tissue damage in a single patient arise from a number of different B cell clones, and that the study of any single clone will not necessarily give a true picture of the whole disease process. However, in order to draw conclusions about antibody sequence, genetic origin and fine structure, it is usually necessary to study monoclonal antibodies. These can be produced from peripheral blood lymphocytes or spleen cells by making hybridomas or by EBV transformation of cells.

Attempts have been made to define pathogenically important antibodies by means of idiotypes. An idiotype is a determinant in the variable region of an antibody molecule that is definable by a polyclonal antiserum. An idiotope is a variable region determinant defined by a monoclonal antibody. Some idiotypes are present on antibodies of several different specificities. These *public* idiotypes are generally believed to be encoded by elements in the FWs. Other idiotypes are more restricted and such *private* idiotypes are encoded primarily by the CDRs. Where it can be shown that the level of a particular idiotype is high in patients with SLE and that antibodies carrying it are detectable in damaged tissues (Isenberg & Collins, 1985; Kalunian *et al.*, 1989), it suggests that monoclonal anti-dsDNA antibodies carrying the same idiotype may be representative of the pathogenetically important clones in such patients.

In rare cases, it has been possible to localize an idiotope accurately. For example the idiotope 9G4 is encoded by the amino acid residues AVY at positions 23–25 in FW1 of the germ-line gene V_{4-34} (Potter *et al.*, 1993) and, therefore, serves as a marker for antibodies where the heavy chain is encoded by this gene. More recently, it has been shown that the idiotope actually comprises an interaction between these three amino acids and a tryptophan at position 7 in FW1 (Mockridge *et al.*, 1996). It has been estimated that 45% of patients with SLE may possess raised levels of 9G4 idiotope-positive antibodies in their serum and

that in 3 out of 11 patients these antibodies were deposited in the kidneys (Isenberg *et al.*, 1993). Studies of monoclonal anti-dsDNA antibodies encoded by V_{4-34} may, therefore, be relevant to the pathogenesis of lupus nephritis.

Evidence from sequence analysis of monoclonal anti-dsDNA antibodies

By cloning and sequencing cDNA encoding the V_H and V_L regions of a monoclonal antibody, it is possible to discover a great deal about the history of the B cell clone which produced it. Firstly, the identities of the V and J genes that have been rearranged functionally to produce the heavy and light chains of this antibody can be deduced. It may be possible to identify the D genes, though this is more difficult because of the complexity of recombination in this area. The cDNA sequence can then be compared with the sequence of the corresponding germline gene, ideally by sequencing non-rearranged genomic DNA from the individual from whose cells the monoclonal antibody was derived. Differences between the two sequences are likely to result from somatic mutation.

Many monoclonal anti-dsDNA antibodies have been derived from mouse models of SLE. Certain genes seem to be used preferentially to encode such antibodies. For example, antibodies from three different mouse models all use the same $V_{\rm H}$ genes, which are members of the J558 gene family (Krishnan & Marion, 1993). The relevance of mouse autoantibody data to human disease is not entirely clear, however. The mouse $V_{\rm H}$ gene repertoire has not been fully sequenced and is more complicated than that of humans. Estimates of the number of functional mouse $V_{\rm H}$ genes vary from 100 to 1000 (Kofler *et al.*, 1992), compared with 51 in humans. In addition, expressed mouse V genes show a generally lower degree of complexity in CDR3 sequences, and of somatic mutation, than human V genes. These facts suggest that choice of $V_{\rm H}$ gene may play a more important role in generation of antibody diversity in the mouse than in humans. Nevertheless, certain features, such as the role of basic amino acids in binding to DNA, seem to be common to both species (see below).

In humans, cold agglutinins (which bind to the I/i antigen on the surface of erythrocytes) all use the V_{4-34} gene. Although this shows that gene preference can be involved in determining antigen specificity in humans, the available evidence from human monoclonal anti-dsDNA antibodies shows no definite preference for any particular family or for particular V genes (Isenberg *et al.*, 1997). However, the numbers of antibodies studied are small, especially those of IgG isotype. This conclusion must 'therefore' be guarded.

By studying IgM and IgG cDNA libraries from unstimulated human peripheral blood lymphocytes, Huang and Stollar (1993) demonstrated that IgG gener-

ally tend to carry more somatic mutations. This is also true of human monoclonal anti-dsDNA antibodies. IgM monoclonal antibodies with relatively low affinity and low specificity for DNA have been isolated from healthy subjects as well as from patients with SLE. These antibodies have often shown cDNA sequences almost identical to those of the germ-line genes (reviewed in Isenberg et al., 1994), although some mutated IgM have been reported (Rioux et al., 1995). In contrast, it has proved very difficult to produce human IgG monoclonal antidsDNA antibodies, even from patients with active disease. The few IgG molecules that have been reported tend to have numerous somatic mutations, particularly in the V_u regions (van Es et al., 1991; Winkler, Fehr & Kalden, 1992; Stevenson et al., 1993; Ehrenstein et al., 1994). The replacement mutations are not evenly distributed, being clustered in the CDRs. This is often expressed in the form of the ratio of replacement to silent mutations (R:S) in each region of the sequence. Where R:S is higher in the CDRs than the FWs, this finding suggests that the B cell clone producing the monoclonal antibody has been subject to antigen-driven clonal expansion. This observation can be expressed statistically by using the binomial theorem to calculate the probability that a particular distribution of R and S mutations could have arisen by chance without requiring antigen-mediated selection (Chang & Casali, 1994).

Antigen promotes accumulation of sequence motifs associated with high-affinity binding

The theory that a particular clone of antibody-producing cells is stimulated to expand by antigen and that later generations of cells accumulate R mutations in the CDRs predicts that it should be possible to produce distinct but clonally related hybridomas from an individual patient with SLE. Such clonally related hybridomas have been reported in studies of both rheumatoid factors (Schlomchik *et al.*, 1987) and anti-DNA antibodies (Schlomchik *et al.*, 1990) in mice. The clonal relationship is deduced from the fact that the antibodies share the same 'clonal signature' in V_H CDR3. Dersimonian *et al.* (1987) reported three IgM monoclonal anti-DNA antibodies derived from a single patient that showed very few differences from the germ-line $V_{\rm H}26$ (now called V_{3-23}). Two of these showed exactly the same V_H sequence, including the clonal signature. Therefore, even where extensive mutation has not occurred, clonal expansion may contribute to the production of anti-DNA antibodies.

In patients with SLE and in murine models of the disease, therefore, certain clones of autoreactive B cells seem to escape the normal control mechanisms. Under the influence of appropriate cytokines, antigen drives these clones to expand and to produce high-affinity 'pathogenic' antibodies. As noted above, this

may be associated temporally with isotype switching. By comparing the sequences of many such antibodies, it has been possible to suggest which particular mutations contribute most to this high affinity. Much of this work has been done in mice, simply because of the large number of monoclonal antibodies available. The most striking feature is that the number of basic or positively charged amino acids in the sequence is often increased in high-affinity anti-dsDNA antibodies, especially in V_HCDR3, where they may arise from junctional diversity or N base addition as well as from mutation (Krishnan & Marion, 1993). It has been postulated that these changes enhance binding to negatively charged DNA. Computer programs can use the variable region sequence of such an antibody to construct a three-dimensional model of the antibody-DNA complex. In an extensive review of murine anti-DNA antibodies, Radic and Weigert (1994) showed that such models often demonstrate an important role for basic amino acids such as arginine (R), asparagine (N) and lysine (K) at the site of binding to the DNA double helix. These amino acids could be situated in any of a number of different CDRs, particularly V_H CDR1, V_H CDR2, V_H CDR3 and V_L CDR1.

In human IgG anti-DNA antibodies, similar patterns have been found. As in mice, basic or positively charged residues are prominent especially, but not exclusively, in V_H CDR3 (Winkler *et al.*, 1992; Ehrenstein *et al.*, 1994). The mere presence of such residues is not sufficient to confer high affinity for dsDNA. Some aPL, for example, have basic CDRs but no DNA-binding activity (Rahman *et al.*, 1996). This may be because of the different spatial arrangement of charged groups in DNA and in anionic phospholipids, which are the main targets of these aPL. The repeating double helical structure of DNA can provide a regular array of negative charge whereas this is not true of phospholipids.

Nevertheless, studies of anti-DNA antibodies encoded by V_{4-34} support the importance of basic residues in $V_{\rm H}$ CDR3. D5 and RT 79 are monoclonal anti-DNA antibodies produced in the same laboratory, that both use this $V_{\rm H}$ gene. Their heavy chain sequences have many basic residues in $V_{\rm H}$ CDR3, a feature not seen in monoclonal cold agglutinins encoded by V_{4-34} that do not bind to DNA (Stevenson *et al.*, 1993). Conversely, some groups have described sequence motifs that occur commonly in anti-dsDNA antibodies but do not contain positive residues. An example is the tetrapeptide YYGS, which has been reported to occur in several such antibodies and to be derived from the germ-line D segment DXP'1 (Cairns *et al.*, 1989).

There is, therefore, no simple relationship between the absolute number of basic amino acids in the CDRs of a monoclonal antibody and its affinity for dsDNA. It is the actual position of these residues that is critical. It is possible to use computer-generated models of the type mentioned above to hypothesize

that particular residues are crucial in the binding site (Radic *et al.*, 1993; Kalsi *et al.*, 1996). To test such a hypothesis, it is necessary to compare the binding properties of the original antibody with those of a slightly modified molecule that differs only at the residues in question. Whereas such modifications can be readily achieved by site-directed mutagenesis, it is more problematic to develop systems for expressing the mutant and wild-type anti-DNA antibodies.

Use of expression systems to investigate autoantibody function

To be useful scientifically, the expressed product must possess a complete antigen-binding site, and expression of both $V_{\rm H}$ and $V_{\rm L}$ cDNA in the same cell is, therefore, required. Some workers have sought to express the cloned $V_{\rm H}$ and $V_{\rm L}$ genes of previously characterized monoclonal antibodies for which the binding properties are known. This has been achieved in a number of ways: as whole antibodies in mammalian cells (Radic et al., 1993; Pewzner-Jung, Simon & Eilat, 1996), as single chain Fv fragments (Polymenis & Stollar, 1994) or as phagebound Fab fragments (Mockridge et al., 1996). Phage-bound Fab can also be used in a different way by creating heavy and light chain cDNA libraries, allowing a large number of different heavy and light chain combinations to be expressed on the surface of phage molecules and testing for DNA-binding affinity (Barbas et al., 1995). In general, the results of experiments using all these techniques have tended to confirm the predictions of sequencing and modelling studies. For example, Radic et al. (1993) showed that removal of arginine residues from sites believed to be important in the heavy chain of the murine monoclonal antibody 3H9 did result in reduced DNA-binding affinity. The sequential addition of arginine residues increased affinity, but not in a linear fashion, and some sites appeared more critical than others. Using a similar method, Katz, Limpanasithikul & Diamond (1994) found that DNA-binding affinity could be perceptibly altered by single amino acid residue changes in the $V_{\rm H}$ sequence of another murine antibody, R4A. However, some changes that increased positive charge actually reduced affinity for DNA, and when the mutant antibodies were introduced into SCID mice, the ability to cause glomerulonephritis did not always vary in parallel with the ability to bind dsDNA.

Much less work has involved mutagenesis of human monoclonal anti-dsDNA. However, in a recent study of the V_{4-34} -encoded antibody D5, Mockridge *et al.* (1996) demonstrated that basic residues in V_H CDR3 were important in binding dsDNA, as were changes caused by somatic mutations in the light chain but not the heavy chain. Intriguingly, binding to the 9G4 idiotope carried on this antibody was independent of changes in V_H CDR3 but completely abolished by specific mutations in the gene for V_H FW1. This framework region is also known to be important for the binding of the red cell antigen I/i, so that in V_{4-34} -encoded antibodies, it seems clear that sites for two different antigens can be encoded by different parts of the $V_{\rm H}$ sequence. As noted previously, binding to the framework regions is characteristic of B cell superantigens (Fig. 4.2), and the fact that the red cell binding site is in FW1 may result from molecular mimicry between the I/i antigen and such a superantigen.

Overall, the method of reasoning described above can show how high-affinity anti-dsDNA antibodies might develop and can help to build up a picture of the antibody-DNA interaction. It does not tell us the identity of the stimulating antigen, or exactly how the antibodies cause tissue damage. Although there are small amounts of DNA present in the bloodstream, mammalian DNA is a poor immunogen and it seems likely that a low-affinity autoantibody response is initially stimulated by a DNA - protein complex such as the nucleosome (Rumore & Steinman, 1990). Antibodies more specific for dsDNA might then develop after clonal expansion, antigen-driven mutation and isotype switching. A number of different mechanisms have been suggested to explain how these antibodies could cause glomerulonephritis. For example, they may cross-react with cell surface proteins (Raz et al., 1993) or antibody - DNA - histone complexes might interact with heparan sulphate in the glomerular basement membrane (Brinkman et al., 1990). The more it becomes possible to distinguish the fine structure of those antibodies which do cause such clinical effects from those which do not, the closer we will be to resolving these issues.

Anti-phospholipid antibodies

Antibodies to phospholipid occur in 1.5 to 5% of the population (Harris & Spinnato, 1991) but are present at much higher frequency (up to 25–40%) in patients with SLE (Harris, Gharavi & Hughes, 1985). These antibodies bind to a variety of neutral or negatively charged phospholipids. aPL can occur in patients with a variety of infectious or neoplastic conditions but under these circumstances they appear to bind cardiolipin without the need for a cofactor and are not associated with an increased risk of thrombosis (Hunt *et al.*, 1992). aPL found in patients with autoimmune disorders appear to recognize the antigen in association with a serum cofactor known as β_2 -glycoprotein I (β_2 -GPI) and are, in some cases, associated with increased risk of thrombosis, leading to the clinical features of the anti-phospholipid antibody syndrome (APS) (Hunt *et al.*, 1992). These features include venous thromboses, increased prevalence of stroke, skin rash, thrombocytopenia and recurrent spontaneous miscarriages (Asherson *et al.*, 1989). As in the case of anti-dsDNA antibodies described above, it has been found that high-affinity aPL of the IgG isotype are more closely associated with

the presence of disease features than IgM antibodies of lower affinity and specificity (Alarçon-Segovia *et al.*, 1989).

As has been described, the methods of molecular biology and immunology have been used extensively to study the origin and properties of anti-dsDNA antibodies. It is becoming clear that very similar reasoning can be applied to the study of other potentially pathogenic antibodies. aPL antibodies provide a good example as they share many characteristics with anti-DNA antibodies. A number of low-affinity monoclonal IgM antibodies bind both these antigens, and it has been known since the early 1980s that a single replacement mutation in a murine antibody can be enough to convert it from an anti-DNA into an aPL antibody (Diamond & Scharff, 1984).

Just as for anti-DNA antibodies, human IgG monoclonal aPL antibodies are commonly more mutated than IgM and bind with higher affinity and specificity (van Es *et al.*, 1992; Menon *et al.*, 1997). This suggests a role for antigen-driven clonal expansion in the development of pathogenic, high-affinity IgG aPL. Further evidence supporting this is the isolation of two clonally related monoclonal aPL antibodies from a single patient with features of APS (Menon *et al.*, 1997). Attempts to identify sequence motifs conferring enhanced binding properties are not as far advanced as in anti-DNA antibodies. However, aPL antibodies found in patients with clinical features of APS bind primarily to negatively charged phospholipids. It is 'therefore' striking that monoclonal aPL antibodies that show specific binding or are derived from patients with active APS tend to have more basic residues in their CDRs than less-specific aPL antibodies from healthy volunteers (Rahman *et al.*, 1996). The next step will be to carry out modelling and mutagenesis experiments to investigate the antibody – phospholipid interface.

Summary

The complex processes whereby the immune system uses a relatively small number of gene segments to produce an antibody repertoire capable of defending the body against a vast number of potential antigenic targets have been described. A possible side effect of these mechanisms for generating antibody diversity is the production of dangerous autoantibodies. In some rheumatic diseases, clones producing such antibodies escape the normal regulatory systems and the autoantibodies produced can contribute to tissue damage. Knowledge of B cell biology can be used to study the sequence and properties of such antibodies so that we may eventually be able to prevent their formation or block their adverse effects.

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The role of CD40 in immune responses P. LANE

Introduction

CD40 and its ligand (CD154: CD40L) play a central role in initiating and maintaining both T and B cell immune responses and are crucial in activating other effector cells such as macrophages. CD40 is far more widely expressed than previously thought, and recent data showing expression of this molecule on endothelium have implicated CD40 in the recruitment of cells at sites of inflammation. This review will focus on the experimental evidence in mouse and human that has contributed to our understanding of the normal function of this molecule *in vivo*. This will be viewed in the context of how CD40 contributes to disease processes, particularly autoimmune diseases, and how targeting of this model may offer new therapeutic possibilities for modifying disease activity. Although there is now some evidence for intracellular signalling pathways via CD40, these are covered elsewhere (Cheng *et al.*, 1995; Hanissian & Geha, 1997).

Structure of CD40 family members and their ligands

CD40 is a member of an ever increasing family of receptors for which the TNF receptor is the prototype (Smith, Farrah & Goodwin, 1994). The family includes the TNF receptors, CD30, CD27, OX40, the 4-IBB antigen, Fas, nerve growth factor receptor, several viral genome products and the newly described TRAIL molecule (Pan *et al.*, 1997). The extracellular portion of members of the TNF family share significant homology mainly because of conservation of the basic extracellular binding domain, which is composed of cysteine-rich pseudo-repeats, each containing about six cysteine motifs and 40 amino acid residues. Some molecules in the family encode soluble forms by alternative splicing. Intracellular domains, in contrast, vary considerably, indicating a likely diversity in intracellular signalling. CD40 has a cytoplasmic tail of 62 amino acid residues, which

is required for intracellular signalling. Mutation resulting in a change at position 234 of a threonine residue to serine abolishes intracellular signalling (Clark & Lane, 1991).

The ligands for the TNF family of receptors are type II membrane proteins, and their homology resides in the carboxyl terminal 150 residues. CD40L has a relatively long stretch of amino acids that link the carboxy terminal region to the 22 residue cytoplasmic tail. Like some of the receptors, there is potential for alternatively spliced forms of the ligands' which may be secreted as soluble factors. The ligand can certainly be shed from expressing cells after engagement of CD40, and there is some evidence that the molecule is downregulated after engagement (Yellin *et al.*, 1994).

The crystal structures of TNF and its receptor have given important insights into the mechanisms of action of the receptor/ligand family (Banner *et al.*, 1993). The ligands form trimeric structures that may dimerize or trimerize the receptors. Evidence from monoclonal antibody (mAb) binding studies suggests that dimerization is sufficient for activation. Conservation of the cytoplasmic domain of CD40L between mouse and humans indicates an important role, possibly related to the regulation of expression of the molecule or to regulation of the endocytosis that occurs following engagement of CD40 (Yellin *et al.*, 1994). Additionally, there is potential for signalling through the cytoplasmic domain in cells that express it, blurring the distinction between receptor and ligand (Smith *et al.*, 1994).

Expression of CD40 and its ligand

CD40 is expressed widely (Banchereau *et al.*, 1994). Virtually all cells of bone marrow origin express CD40; these include B cells from the earliest precursors until late plasmablasts (CD40 is lost on terminal differentiation to plasma cells), macrophages, monocytes and dendritic cells, CD34⁺ stem cell precursors and (most recently identified) T cells (Attrep *et al.*, 1996). Cells of other origins expressing CD40 include the basement layer of epithelial cells, many carcinoma cell lines (constitutively expressed), thymic epithelial cells in the cortex and medulla, and also endothelium. The function of the CD40 molecule in this other group of cell types is poorly defined.

Originally, the ligand for this molecule was found to be expressed on activated T cells (Armitage *et al.*, 1992; Lane *et al.*, 1992). Its expression has also been described at low levels on basophils and mast cells (Gauchat *et al.*, 1993) and also, surprisingly, on activated B cells (Grammer *et al.*, 1995). Initially there was some controversy with regard to the sites of expression of CD40L in lymphoid tissue (Foy *et al.*, 1994); now it is clear that activated T cells in the T cell areas

and within germinal centres express CD40L (Casamayor-Palleja, Khan & MacLennan, 1995). It is not clear by immunohistology which, if any, B cells physiologically express the molecule. It is thought that CD40L expressed on mast cells may play a local role in switching B cells to IgE synthesis (Gauchat *et al.*, 1993).

Ontogeny of immune responses

Because CD40 plays such a pivotal role in both T and B cell immune responses, its function needs to be understood in the context of the normal sequence of events that leads to T and B cell priming.

Priming to T cells

T cell priming occurs in the T cell areas associated with specialized antigenpresenting cells called interdigitating dendritic cells (IDCs) (Inaba *et al.*, 1983). These cells are derived from Langerhans cells, which are phenotypically immature antigen-presenting cells that constitutively pick up soluble and particulate antigen via pinocytosis (Sallusto & Lanzavecchia, 1994). Subsequently, these cells migrate from the tissues via afferent lymphatics to lymph nodes, stop processing antigen and present peptide fragments derived from these proteins as they upregulate expression of class I and class II molecules as well as other co-stimulatory molecules. The migration of Langerhans cells from the tissue to central tissues probably occurs at a basal 'tickover' rate, but this is greatly augmented by inflammation and inflammatory cytokines like TNF (Austyn, 1996).

Resting T cells migrate from blood into lymphatic tissue via specialized postcapillary venules called high endothelial venules (HEV). Most evidence suggests that the T cells 'crawl' over antigen-presenting IDCs searching for antigen, which if encountered leads to T cell activation, proliferation and 'priming'. Primed antigen-specific T cells migrate to the outer T zone where they can interact with antigen-specific B cells. They also leave via efferent lymphatics. Activated T cells, through differential expression of adhesion receptors (Mackay, Marston & Dudler, 1991), can migrate out into inflamed tissues where they activate local inflammatory cells or are specifically cytotoxic against infected cells.

Primary B cell activation

By identifying antigen-specific B and T cells, it has been possible to study the primary interactions between B (Jacob, Kassir & Kelsoe, 1991; Liu *et al.*, 1991) and T cells (Gulbranson-Judge & MacLennan, 1996; Luther *et al.*, 1997).

Antigen-specific B cells carrying receptors that have been triggered by binding antigen get trapped in the T cell areas 'looking' for primed antigen-specific T cells (Cyster, Hartley & Goodnow, 1994). B cell presentation to unprimed T cells is poor (Lassila, Vainio & Matzinger, 1988; Fuchs *et al.*, 1992). There is an initial burst of B cell proliferation in the T cell areas. Many of these B cells differentiate locally to short-lived plasma cells in the red pulp of spleen, the medulla of lymph nodes and the lamina propria of the gut and respiratory mucosa.

Some B cells do not differentiate but migrate into follicles, where they proliferate and form germinal centres (MacLennan *et al.*, 1990); it appears that these B cells are clonally related to those B cells proliferating in the T cell areas (Jacob & Kelsoe, 1992). Germinal centres are sites where antigen-specific B cells are expanded, somatic mutation is switched on and mutants are selected. This environment allows affinity maturation to take place. B cell mutants that bind antigen better than other B cell clones are selected in germinal centres. Some progeny differentiate to produce plasma cells, which have most often switched to production of a different class of immunoglobulin, and memory B cells. Others continue the cycle of somatic mutation and selection.

Location of memory B cells and maintenance of antibody responses

Memory B cells that leave the germinal centre reaction enter the marginal zone in the spleen, where they exit the cell cycle (Liu, Oldfield & MacLennan, 1988). In lymph nodes, there is a comparable B cell area just underneath the capsule and adjacent to the lymphatics (Lagresle, Bella & Defrance, 1993). Both marginal zone and subepithelial B cells lie in antigen-traffic zones so are ideally situated to re-encounter antigen. Many of these memory B cells are long lived (MacLennan *et al.*, 1990), especially those that have switched to IgG production (Schittek & Rajewsky, 1990). Specific antibody responses are maintained by chronic activation of antigen-specific B cells within B cell follicles long after the germinal centre reaction has died down (MacLennan *et al.*, 1990).

Secondary B and T cell responses

Memory B cells activated through their antigen-specific receptors are rapidly recruited into immune responses and migrate again to the T cell areas. There is intense proliferation at this site, where the majority of proliferating cells differentiate to plasmablasts, which migrate to the bone marrow (Benner, Hijmans & Haaijman, 1981) and become long-lived plasma cells (Ho *et al.*, 1986). The observed cellular reaction is accompanied by elevated switched specific antibody responses in the sera.

Memory T cells are activated by antigen presented by IDCs, but they can also be restimulated by other antigen-presenting cells such as B cells because their stimulatory requirements are less stringent than those of unprimed cells.

Immunodeficiency resulting from CD40L and CD40 deficiency

The key insights into the function of CD40 and its ligand have come from studies in genetically deficient mice and humans. Deficient expression of CD40L was reported by several different groups more or less at the same time (reviewed in Callard *et al.*, 1993).

Humans

Deficiency of CD40L is associated in humans with the sex-linked hyper-IgM syndrome, where affected males have very low levels of IgG and IgA and often have elevated levels of IgM. The elevated levels of IgM are driven by infections, as IgM levels often return to normal with immunoglobulin replacement. In addition to their antibody defect, affected patients have problems dealing with infections with the intracellular fungal parasite *Pneumocystis carinii*. Variable neutropenia is also present.

Mice

Mice deficient in both CD40 and its ligand have been created using gene technology to make knock-out mice (Kawabe *et al.*, 1994; Xu *et al.*, 1994). The murine phenotype is similar to that seen in humans, indeed mice even get *P. carinii* infections. Normally they do not have elevated levels of IgM, but this probably reflects the fact that mice tend to be reared in specific pathogen-free conditions. Detailed analyses of these mice have contributed greatly to our understanding of the role of CD40 in T and B cell priming, although several questions remain unanswered. In general a few points need to be clarified.

Antibody responses that do not depend on T cells, so called thymus-independent responses to polysaccharide-based antigens, appear to be normal. Furthermore, B cells are able to switch to IgG_3 , the IgG isotype associated with polysaccharide responses. It is not absolutely clear whether this is also true in humans where antibody responses may be partially dependent on T cells (Lane, 1996).

Primary antibody responses to conventional protein-based antigens occur but are restricted to the IgM isotype. This may partially reflect the relative T cell independence of B cells in primary immune responses.

Role of CD40 in primary immune responses

Effect on antigen-presenting cells: IDCs

The earliest event in T cell priming involves recognition of antigen presented by IDCs. It seems probable that antigen-specific triggering of T cells is sufficient to upregulate the expression of CD40L (Foy *et al.*, 1996), although others have found that expression is strongly upregulated by co-stimulation through CD28 (de Boer *et al.*, 1993; Klaus *et al.*, 1994). Whatever, the initial sequence of events, CD40 activation of IDCs leads to upregulation of the CD28 co-stimulatory B7 ligands (Chapter 6; Ranheim & Kipps, 1993) and HLA molecules and potently induces the expression of IL-12 (Cella *et al.*, 1996). This sequence of events clearly leads not only to more effective antigen presentation but also to co-stimulation of T cells.

T cell priming

The major influence of CD40 in T cell priming is probably through the effects of CD40L expressed by T cells on antigen presentation by IDCs. This conclusion is supported by observations of patients with CD40L deficiency, who have problems eradicating intracellular infections but are not profoundly T cell immunodeficient. Experimental studies in genetically deficient mice have given us a more detailed understanding of CD40's role. Using CD40L-deficient mice, Flavell's group have found T cell priming was impaired although not absent (Grewal, Xu & Flavell, 1995). In this system, CD40L-deficient T cells failed to proliferate in vivo in response to antigen. However, if IDCs were stimulated to upregulate B7 ligands by CD40-independent signals, then CD40L-deficient T cells were able to proliferate. This indicated that the dependence of T cell priming on CD40 was principally through upregulation of co-stimulation on antigenpresenting cells. Recently, this result has been repeated in a different way. CD40L-deficient mice cannot be primed to get encephalomyelitis, but priming can be restored by immunization with B7.1 expressing antigen-presenting cells, where upregulation of B7 ligands through CD40 has been bypassed (Grewal et al., 1996).

The one caveat to the above experimental conclusions is that activated T cells can express CD40 (Attrep *et al.*, 1996) and CD40L will co-stimulate T cells directly (Armitage *et al.*, 1993; Fanslow *et al.*, 1994). Therefore, one might expect co-stimulatory effects between activated T cells mediated by CD40 to play some role in the expansion of T cell clones.

Role of CD40 in the effector phase of the immune response

Cytotoxic T cell priming

Humans with deficient CD40L expression do not, in general, have problems with viral infections. This is largely confirmed in mice. CD40-deficient mice make normal CD4⁺ T cell responses to infectious viruses (Oxenius *et al.*, 1996) although antibody responses are grossly impaired. Primary cytotoxic responses are also normal although memory responses are impaired, for reasons that are not clear (Borrow *et al.*, 1996).

Macrophage activation

Humans clearly have problems dealing with intracellular infections like that with *P. carinii*. CD40 activation of macrophages leads not only to IL-12 production but also to production of other proinflammatory cytokines like TNF and IFN- γ (reviewed in Grewal & Flavell (1996)). CD40L-negative mice are susceptible to leishmaniasis (Soong *et al.*, 1996), and the defect can be corrected by IL-12, which plays a key role in switching the T cell immune response to produce T_H^1 cytokines (Trinchieri, 1995), which are instrumental in activating macrophages to kill intracellular organisms.

B cell priming

CD40L-deficient humans have impaired immunoglobulin class switching, no germinal centre formation and impaired affinity maturation of antibody responses because of a lack of somatic mutation. This phenotype is confirmed in CD40-(Kawabe *et al.*, 1994) and CD40L-deficient mice (Xu *et al.*, 1994).

Normal B cell responses depend on T cell priming and the appropriate helper signals, so it is appropriate to try to dissect the role of CD40 at different stages of this process. As stated above, T cell priming is impaired but not absent in mice deficient in CD40 signalling. Whether such T cells can efficiently deliver help for B cells is controversial. Two groups have taken T cells primed in the absence of CD40 and examined whether they can help normal B cells make antibody. Unfortunately, opposite answers were obtained: one group found T cells helped B cells (Oxenius *et al.*, 1996), the other did not (van Essen, Kikutani & Gray, 1995). The reasons for this discrepancy are not apparent and may be related to the degree of T cell priming.

B cell activation to plasma cells

B cell differentiation to plasma cells following cognate interactions between antigen-specific B and T cells in the T zones is relatively CD40 independent. Primary antibody responses are fairly normal and this is associated with the local differentiation of activated B cells to plasma cells. There is independent confirmation of this finding from *in vitro* studies: CD40L-deficient T cells will induce plasma cell differentiation in resting B cells (Lane *et al.*, 1995), and blocking CD40 induces plasma cell differentiation in activated B cells (Arpin *et al.*, 1995).

B cell proliferation

CD40 activation of B cells is a potent proliferation signal especially in conjunction with signals through the immunoglobulin receptor. Humans and mice deficient in this signalling pathway lack germinal centres in B cell follicles. Germinal centres are sites in the immune system that serve at least two functions.

- 1. Clonal amplification of high-affinity B cells
- 2. Somatic mutation and selection.

CD40 plays a clear role in B cell proliferation and selection (Liu *et al.*, 1989). Its role in somatic mutation is less clear because it is difficult to amplify and select somatic mutants and, therefore, they are difficult to detect. There is some published evidence that signalling through CD40 and surface immunoglobulin is important for maintaining somatic mutation in B cells *in vitro* (Källberg *et al.*, 1996), but this does not mean CD40 switches on mutation in B cells. There are some theoretical reasons to suppose that the mechanisms that regulate mutation and proliferation should be kept separate (Lane, 1997).

Class switching

Mice deficient in CD40 signalling have very low levels of IgA and IgG although levels of IgG_3 , the isotype associated with thymus-independent responses, are normal. CD40 signalling induces sterile transcription of downstream IgG loci (Jumper *et al.*, 1994), indicating a significant contribution to successful class switching, although CD40 signalling alone seems to be insufficient.

Role of CD40 in B cell established immune responses

Because CD40 plays such a crucial role at so many different locations during immune responses, blocking CD40 interactions during established immune responses has profound immunosuppressive effects (Foy *et al.*, 1996). Its effect

on established B cell immune responses where there is germinal centre formation is to cause dissolution of germinal centres and differentiation to plasma cells (Han *et al.*, 1995). This result is similar to that found *in vitro*, where CD40Ldeficient T cell help leads to plasma cell formation (Arpin *et al.*, 1995; Lane *et al.*, 1995). The effects of blocking CD40 on long-term memory B cells are unknown.

Effects of CD40 in established T cell immune responses

Some effects of blocking CD40L are complicated by the fact that anti-CD40L antibodies may in some circumstances delete T cells expressing CD40L. In addition, it is very difficult to distinguish the role of CD40L-blocking events at distinct locations within the immune system because CD40 is so widely expressed. Nevertheless, its is clear that blocking CD40L *in vivo* exerts potent immuno-suppressive effects. In mice prone to autoimmunity (Early, Zhao & Burns, 1996), anti-CD40L antibodies abrogate disease, as they do for graft-versus-host disease (Blazar *et al.*, 1997). Furthermore, in a transplantation model, antibodies to CD40L were potent enhancers of graft acceptance, especially in conjunction with blocking CD28 co-stimulation (Larsen *et al.*, 1996).

Tolerance induction

Although CD40L-deficient mice have relatively poor T cell priming (Grewal & Flavell, 1996), there is little evidence to suggest that T cells are tolerized by the lack of CD28 co-stimulation. Prior donor blood transfusion has been known for a long time to have a beneficial effect on the subsequent graft outcome, and it has been speculated that donor B cells induce tolerance to allogeneic antigens (Lassila *et al.*, 1988; Eynon & Parker, 1992; Fuchs *et al.*, 1992). Recent evidence suggests that CD40-deficient B cells are even more potent at inducing tolerance (Hollander *et al.*, 1996), and this is also true with blocking antibodies to CD40L (Buhlmann *et al.*, 1995). The exact mechanism by which B cells are tolerogenic is still not completely clear.

Summary

CD40 is widely expressed and plays a key role both in the afferent limb of the immune response in the priming of antigen-specific B and T cells and in the development of effector function, including macrophage activation, the development of effector T cells and in the clonal expansion and affinity maturation of B cells. Blocking CD40 with antagonists has the potential to block many different

aspects of immune responses and exerts potent immunosuppressive effects that may be exploited in the treatment of autoimmune disease, transplant rejection and tolerance induction.

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6

Manipulation of the T cell immune system via CD28 and CTLA-4

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Introduction

In order to provide protection from a vast array of infectious agents, the immune system has evolved a series of defences based on specialized immune cells. One component of this system, the T lymphocyte, is critical in organizing and effecting cellular responses by providing helper functions for B cells, as well as by generating direct cytotoxic actions. The major challenge faced in controlling T cell responses is how to generate a sufficiently large immune repertoire capable of recognizing any possible foreign antigen while at the same time maintaining T cells in an unresponsive state towards an equally large array of self-antigens. Clearly any breakdown in the barriers that prevent recognition and activation of T cells by self-antigens allows the possibility of developing autoimmune conditions, which include RA and SLE. In order to gain an understanding of potential disease mechanisms and to provide initiatives for novel therapeutic strategies, it is necessary to understand these mechanisms of T cell tolerance.

Since the late 1970s, substantial progress has been made in our understanding of the molecular basis of antigen recognition by T cells. Based on the observations of Zinkernagel and Doherty, who demonstrated that T cell recognition of foreign antigens requires appropriate (self) MHC antigens, it is now well established that the function of MHC class I and class II antigens is to bind and display both self and non-self peptide fragments on the cell surface (Zinkernagel & Doherty, 1974; 1975; Brown *et al.*, 1993). In the case of MHC class II molecules (HLA-DR, HLA-DQ and HLA-DP), expression is generally restricted to professional antigen-presenting cells, such as dendritic cells, monocytes, macrophages and activated B cells. MHC-displayed peptides thus form the central focus of T cell antigen recognition and it is, therefore, significant in the context of autoimmune conditions that diseases such as RA show strong selection for particular *HLA-DR* alleles such as *DR1* and *DR4* (Stastny, 1978;

Gregerson, Silver & Winchester, 1987) suggesting a possible T cell-driven aetiology.

Since antigens to both 'self' and 'non-self' are normally available for T cell recognition, T cell activation must be tightly controlled by self-tolerance, which involves both 'central' and 'peripheral' mechanisms. The central mechanism involves thymic selection, which has traditionally been seen as the major source of self-tolerance whereby the majority of T cells expressing clonally rearranged antigen receptors undergo apoptotic death (deletion) as a result of their expression of 'inappropriate' receptors (Kappler, Roehm & Marrack, 1987; Pullen, Kappler & Marrack, 1989; MacDonald & Lees, 1990). The details of this process are beyond the scope of this review; however, T cells that survive thymic selection emigrate from the thymus and then form the basis of the peripheral T cell pool. Despite the removal of large numbers of immature T cells during development in the thymus, there is increasing evidence that thymic selection is insufficient to completely account for self-tolerance and that other mechanisms must exist. Indeed evidence from mice that are defective in the control of peripheral tolerance mechanisms suggest these may be more important in preventing autoimmunity (Bluestone, 1997).

Our understanding of the mechanisms of peripheral self-tolerance was considerably advanced in the late 1980s when several lines of evidence began to show definitively that despite the expression of self-antigens and the presence of specific antigen-reactive T cells for these antigens immune activation did not necessarily ensue (Lo *et al.*, 1988; Markmann *et al.*, 1988; Ramensee, Kroschewski & Frangoulis, 1989; Schwartz, 1990). This underlined the existence of peripheral tolerance mechanisms and indicated that factors other than antigen recognition could determine the outcome of T cell receptor engagement. This review will focus on two molecules, CD28 and CTLA-4 (CD152), which are now known to play a key role in determining the fate of antigenically challenged T cells. Recent evidence suggests that while both CD28 and CTLA-4 interact with the same ligands (CD80 and CD86), CD28 and CTLA-4 function differentially to act as 'go' and 'stop' signals, respectively, for T cell activation and proliferation.

A general scheme for the interactions between these receptors and their ligands is shown in Fig. 6.1.

CD28 and CTLA-4

Structure of CD28 and CTLA-4

CD28 is a cell surface glycoprotein found on the majority of peripheral blood T cells, with preferential expression on CD4⁺ cells (95% positive) compared with



Fig. 6.1. Schematic representation of CD28 and CTLA-4 interactions. CD28 expressed on T cells initially interacts with the more abundantly expressed CD86 molecule on antigen-presenting cells. This provides positive 'go' signals for T cell proliferation and cytokine production in the context of antigenic stimulation via the MHC–TCR interaction. CD80 is induced upon activation of antigen-presenting cells and provides a second ligand capable of CD28 co-stimulation. In contrast, the higher-affinity CTLA-4 receptor is expressed intracellularly or in low abundance at the cell surface. Interaction of CTLA-4 with either CD80 or CD86 molecules initiates negative 'stop' signals' causing T cell cycle arrest and inhibition of T cell responses to antigen.

CD8⁺ cells (50% positive) (Linsley & Ledbetter, 1993; June *et al.*, 1994). The gene for CD28 is a member of the immunoglobulin gene superfamily and expresses a protein containing a single extracellular 'V'-like domain. An initial polypeptide of M_r 23 000 is heavily glycosylated to give a mature glycoprotein of approximately 44 000, ultimately expressed at the cell surface as a 90 000 homodimer. The mature CD28 protein consists of 202 amino acid residues giving a single 134 residue extracellular domain, a transmembrane section and a 41 residue cytoplasmic tail responsible for signal transduction. This cytoplasmic

domain contains a number of consensus amino acid motifs, including a YMNM motif, which are presumed to mediate interactions with signalling molecules inside the cell.

The CTLA-4 receptor, which is related to CD28, was initially isolated from a subtractive cDNA library screen for molecules expressed in activated T cells (Brunet et al., 1987). The CTLA-4 gene is a member of the immunoglobulin superfamily encoding a single extracellular V-like domain polypeptide (approximately 20 kDa), which is reported to be expressed as a 40 kDa monomer at the cell surface (Lindsten et al., 1993). Despite the fact that CD28 and CTLA-4 share only 30% amino acid identity, they are both located on chromosome 2 (2q33-34) in humans (Dariavach et al., 1988) and share a similar intron/exon structure, suggesting that the two arose from a common ancestor (Harper et al., 1991). Like CD28, CTLA-4 also contains a short cytoplasmic tail of some 36 amino acid residues, with a YVKM motif; this may be involved in its signalling functions and/or in control of cell surface CTLA-4 expression. Strikingly, the CTLA-4 cytoplasmic domain displays 100% evolutionary conservation in several species (June et al., 1994; Harlan, Abe & June, 1995), indicating a highly conserved and essential function. Primary sequence comparisons and mutagenesis studies of the extracellular domains of CD28 and CTLA-4 have identified a conserved amino acid sequence (MYPPPY) present in the extracellular domains of both CTLA-4 and CD28 that appears to be required for their interactions with ligands (Peach et al., 1994; Fargeas et al., 1995; Morton et al., 1996); however evidence suggests that CD80 and CD86 bind to discrete but overlapping sites.

Expression of CD28 and CTLA-4

The expression patterns of CD28 and CTLA-4 display marked differences. CD28 is constitutively expressed and is readily detected on the surface of the majority of resting T cells by flow cytometric analysis (FACS). In response to T cell activation, CD28 surface expression is subsequently modulated, during which both decreases and increases in expression can be observed. Currently, evidence suggests that initially CD28 is downregulated upon interaction with its ligand (Linsley *et al.*, 1993b). This downregulation occurs within a few hours of ligand engagement but is followed by re-expression during the following 24–48 hours. It has been suggested that the signalling capacity of CD28 may be compromised following re-expression, although this has not been confirmed. Our own observations indicate that while CD28 downregulation can be induced by ligand binding alone, re-expression of CD28 is contingent on TCR activation signals. Furthermore, in some circumstances, upon re-expression the levels of CD28 may be substantially higher, especially in cells that have encountered both antigen and

CD28 co-stimulation. This indicates that CD28 upregulation may be a consequence of correct activation and may possibly serve to adjust the balance between CD28 and CTLA-4 usage in activated T cells.

Expression data for CTLA-4 are more limited, since the protein is much less abundant than CD28 and, until recently, antibodies have not been widely available. Information from mRNA analysis surprisingly indicates similar levels of message for both CD28 and CTLA-4 in activated T cells yet CTLA-4 is found at much lower levels at the cell surface (Freeman et al., 1992; Linsley et al., 1993b). However, resting T cells do not express a detectable message for CTLA-4, in contrast to CD28 (Lindsten et al., 1993). Activation signals via TCR and CD28 are also thought to enhance the surface expression of CTLA-4, which is reported to be maximal at 48-72 hours (Walunas et al., 1994; Alegre et al., 1996). However, it should be noted that even at maximal expression the surface levels of CTLA-4 are substantially lower than those of CD28. In line with these observations, current data indicate that CTLA-4 may be predominantly an intracellular protein that translocates to the surface following T cell activation (Alegre et al., 1996; Wang et al., 1996). This expression appears to be focally localized towards the contact site between the T cell and its target and is possibly regulated by a calcium-dependent mechanism (Linsley et al., 1996). Recent studies have revealed that CTLA-4 is retained inside the cell via an association with the AP50 component of the clathrin-associated adaptor AP-2 (Chuang et al., 1997; Shiratori et al., 1997). This association is prevented by phosphorylation of the CTLA-4 YVKM motif, which thus allows surface CTLA-4 expression (Shiratori et al., 1997). This observation is in agreement with mutagenesis studies on CTLA-4, which demonstrated that replacing tyrosine 164 in the YVKM motif with a phenylalanine residue also results in increased cell surface expression (Leung et al., 1995).

Overall, the expression levels and mechanisms of expression of CD28 and CTLA-4 differ considerably, with much higher levels of CD28 being observed compared with CTLA-4. Both molecules are responsive to activation events and this may provide a way of altering the balance between these two receptors in order to maintain control of T cell activity. Given that both known ligands appear capable of binding either CTLA-4 or CD28, it seems likely that some mechanism must be required to allow interactions with either CTLA-4 or CD28 to predominate under appropriate circumstances. One possibility is that the mechanism which promotes CD28 downregulation enhances CTLA-4 expression. While there is little direct evidence for such a mechanism, the kinetics of CD28 downregulation and CTLA-4 expression as well as changes in mRNA levels (Lindsten *et al.*, 1993) are consistent with such a possibility.

CD80 and CD86

Currently, two ligands for CD28 and CTLA-4 have been cloned from humans and characterized (Freeman et al., 1989; 1993; Azuma et al., 1993a); both of which bind to CTLA-4 with higher affinity than to CD28. The first cloned ligand was originally termed B7 and renamed B7.1 when a second ligand B7.2/B70 was identified. Subsequently, both B7.1 and B7.2/ B70 have been renamed CD80 and CD86, respectively. Like CD28 and CTLA-4, both CD80 and CD86 molecules are cell surface glycoprotein members of the immunoglobulin superfamily, which expresses two disulphide-linked extracellular domains. Similarly to CD28 and CTLA-4, the genes for both CD80 and CD86 are located to the same chromosomal region (3 Q13-Q23) in humans (Fernandez-Ruiz et al., 1995), suggesting that they have also arisen via a gene duplication. The CD80 protein is a 262 amino acid residue, 30 kDa polypeptide that has a mature glycosylated mass of approximately 60 kDa. The CD86 molecule is slightly larger (34 kDa; 323 amino acid residues) because of an extended cytoplasmic tail, which is glycosylated to approximately 70 kDa, as determined by SDS-PAGE analysis (Azuma et al., 1993a).

Despite only 23% sequence identity, both CD80 and CD86 were initially reported to have similar affinities for CD28 and CTLA-4 (Linsley et al., 1994). However, more recent studies using surface plasmon resonance analysis have suggested that the affinity differences between CD80 and CD86 binding may be as large as 10-fold (Greene et al., 1996). In addition, there are differences in the kinetics of binding to and dissociation from CTLA-4 (Linsley et al., 1994; van der Merwe et al., 1997). However, despite this, it has been difficult to demonstrate discrete functions for CD80 and CD86, as both are capable of effectively co-stimulating via CD28 (Lanier et al., 1995). To date, the major differences between these two ligands appear to be mainly at the level and kinetics of surface expression. In this regard, CD86 is the most widely expressed and is found constitutively on professional antigen-presenting cells such as resting monocytes and dendritic cells (Azuma et al., 1993a; Fleischer et al., 1996). In addition, CD86 is upregulated by cytokines, such as IFN- γ upon activation, where increased levels are seen on monocytes and induction is found on activated T and B cells (Azuma et al., 1993a; Stack et al., 1994). Some reports have also suggested that CD86 may be expressed on resting T cells and that CD80 and CD86 may be reciprocally regulated following activation (Prabhu Das et al., 1995). In contrast, CD80 has a much more limited expression profile, being virtually undetectable on peripheral blood mononuclear cells in the absence of activation (Koulova & Dupont, 1991; Azuma et al., 1993a; Fleischer et al., 1996). Following activation, CD80 is found on activated monocytes, albeit at lower levels than CD86, and is found on activated B cells and many B cell lines (Yokochi, Holly & Clark, 1982). Varying levels of CD80 are also found on T cells, depending on their state of activation (Sansom & Hall, 1993; Azuma *et al.*, 1993b). Therefore, in terms of providing initial CD28 ligation, CD86 appears to be the primary ligand, a view supported by the fact that anti-CD86 antibodies are more efficient inhibitors of T cell activation than anti-CD80. Furthermore, mice that are deficient for CD86 have a more severe immunodeficient phenotype than do mice which lack CD80 (Borriello *et al.*, 1997).

Given the interest in the CD28 pathway in controlling T cell activation, numerous studies have investigated the expression of these molecules in clinical conditions, including RA and SLE. Much of this work is still at the descriptive stage and there is not yet a consensus as to whether there are significant alterations in expression of these molecules in rheumatic diseases (Sfikakis *et al.*, 1995; Summers *et al.*, 1995; 1996; Balsa *et al.*, 1996; Garciacozar *et al.*, 1996; Schmidt, Goronzy & Weyand, 1996; Folzenlogen *et al.*, 1997; Sfikakis & Via, 1997). Accordingly, both increased expression and lack of expression has been reported for CD80/CD86 and CD28 molecules and the role of these proteins in causing or sustaining disease needs a more detailed functional analysis.

Functional aspects of CD28 and CTLA-4

CD28 co-stimulates T cell activation

During the late 1980s the two-signal model of T cell activation as proposed by Bretscher and Cohen received substantial experimental support from a number of sources, including a series of experiments by Jenkins et al. (Jenkins & Schwartz, 1987; Jenkins, Aswell & Schwartz, 1988; Mueller, Jenkins & Schwartz, 1989). These studies mainly involved the use of modified antigen-presenting cells and demonstrated that, by presenting antigen to T cells after fixation of the antigen-presenting cells, T cells could be made specifically unresponsive (anergic) to the presented peptide whereas normal antigen-presenting cells induced a proliferative response. The missing component from the fixed cells was identified as a cell surface molecule found on 'non-T' spleen cells, which could rescue from anergy if provided at the same time as TCR engagement. Similarly, studies using transgenic mice to study the autoimmune effects of ectopic expression of MHC class II molecules on pancreatic cells concluded that, instead of the predicted autoimmune attack on the class II-positive beta cells, T cells were instead made tolerant to the class II antigen (Lo et al., 1988; 1989; Markmann et al., 1988). Therefore, it emerged that both a TCR and a 'co-stimulatory' signal were required for T cell activation, and that in the absence of co-stimulatory signals T cells
became unresponsive to antigen. The characteristics of this co-stimulatory signal were that it (i) was dependent on a cell surface molecule found on antigenpresenting cells; (ii) was required at the same time as TCR engagement; (iii) could be delivered by a cell distinct from the cell expressing the antigen target; and (iv) resulted in IL-2 production from the T cells.

Independently of these studies, cDNA cloning experiments had identified the genes for the B cell surface antigen B7 (Freeman *et al.*, 1989) and the gene for CD28 (Aruffo & Seed, 1987), culminating in the formal identification of B7 as a ligand for CD28 (Linsley *et al.*, 1991a) and the demonstration that this pathway fulfilled the above co-stimulatory criteria (Gimmi *et al.*, 1991; Jenkins *et al.*, 1991; Linsley *et al.*, 1991a; Razi-Wolf *et al.*, 1992; Sansom *et al.*, 1993). In particular, CD28 was found to be important in enhancement of cytokine production by T cells, an effect mediated by both increased transcription and stabilization of a number of cytokine mRNAs including those for IL-2, IL-4, IL-8, IL-13 and TNF γ (Lindsten *et al.*, 1989; Thompson *et al.*, 1993; June *et al.*, 1994). Furthermore, CD28 engagement was also shown to be capable of preventing T cell anergy, thus identifying it as a key second signal for T cell activation (Jenkins *et al.*, 1991; Harding *et al.*, 1992).

These experiments have resulted in a model of T cell activation where CD28 signals are thought to provide a check on T cell activation and only encounters with antigen in the context of CD80–CD86 interactions on professional antigenpresenting cells result in proliferation and cytokine production. Conversely, a lack of co-stimulatory signals is predicted to lead to an antigen-specific inactivation of the T cell (anergy), thereby providing a mechanism of peripheral self-tolerance. Whilst this view is most likely to be an oversimplification of the requirements of T cell activation, support for this model has been derived from a number of experimental systems.

In particular, considerable progress has been made using a recombinant chimaeric molecule (CTLA-4-Ig) generated by fusing the extracellular domain of CTLA-4 to an immunoglobulin heavy chain constant region (Linsley *et al.*, 1991b). The resulting protein is a soluble high-affinity antagonist of both CD80 and CD86. Results using this protein have demonstrated the tolerogenic potential of blocking CD28 interactions both *in vitro* (Tan *et al.*, 1993) and *in vivo* (Lenschow *et al.*, 1992). In particular, *in vivo* data have yielded impressive results in transplantation models, where CTLA-4-Ig treatment resulted not only in prevention of graft rejection but also in induced tolerance to subsequent grafts (Lenschow *et al.*, 1992; Blazar *et al.*, 1994; Pearson *et al.*, 1994; Ibrahim *et al.*, 1997). This suggests that blocking CD28 interactions is not only immunosuppressive but also that T cells manipulated in this way subsequently undergo a form of anergy. Clearly this mode of action has considerable therapeutic potential, and CTLA-4-Ig is currently under investigation as an immunotherapeutic agent.

CD28 promotes T cell survival

In addition to co-stimulating antigen-induced proliferation and stimulating cytokine production, CD28 signalling has also been increasingly associated with promoting T cell survival. Studies on CD28-deficient T cells have demonstrated that while proliferation can be initiated in the absence of CD28-derived signals the response cannot be sustained (Lucas et al., 1995), which may be associated with decreased cell viability at late time points. Likewise in vitro studies indicate substantial T cell apoptosis following anti-CD3 engagement alone, which is prevented by co-stimulation with anti-CD28 (Boise et al., 1995; Radvanyi et al., 1996; Sperling et al., 1996). Consistent with an anti-apoptotic role for CD28, studies in our own laboratory have analysed T cells that have received co-stimulatory signals and compared their survival with bystander-activated cells. This revealed a striking resistance to Fas-mediated apoptosis in those cells receiving co-stimulatory signals (McLeod et al., 1998). The mechanisms that underlie CD28-mediated survival effects have not yet been clearly defined but two potential candidates are the production of anti-apoptotic lymphokines such as IL-2 and the induction of the antiapoptotic protein Bcl-X₁. Whilst the role of CD28 enhancement of cytokine production by T cells is well established (Fraser et al., 1991; Fraser & Weiss, 1992; Lindsten et al., 1989; Thompson et al., 1993), the role of IL-2 in survival is becoming less clear. The demonstration that IL-2-receptor knock-out mice exhibit lymphoproliferation and autoimmunity (Suzuki et al., 1995) and a defect in Fas-mediated apoptosis indicates that IL-2 signalling may be involved in the maintenance of self-tolerance by facilitating programmed cell death.

The long splice variant of the *bcl-X* gene (*bcl-X_L*) (Boise *et al.*, 1993) has been found to prevent apoptosis induced by a diverse array of stimuli (Grillot, Merino & Nunez, 1995). Since CD28 stimulation upregulates *bcl-X_L*, this is potentially a major route for CD28 survival signals and has been observed in a number of studies (Boise *et al.*, 1995; Noel *et al.*, 1996; Radvanyi *et al.*, 1996; Sperling *et al.*, 1996). Therefore, there is considerable evidence that CD28 co-stimulation protects T cells from apoptosis and that this may involve Bcl-X_L; however it is likely that other routes also exist.

CTLA-4 acts as an inhibitor of T cell activation

While studies of CD28 interactions provide a clear picture of the co-stimulatory role of CD28, the role of CTLA-4 has been more difficult to elucidate. Our

understanding is based on the use of anti-CTLA-4 monoclonal antibodies, and in spite of the fact that CTLA-4 is the higher affinity receptor for both ligands there are no functional data using ligands to stimulate CTLA-4. The interpretation of anti-CTLA-4 antibody data is somewhat complicated, since the various antibodies can be seen as (i) delivering a negative signal (decreasing T cell responses); (ii) blocking a negative signal (increasing responses); (iii) delivering a co-stimulatory signal (increasing responses); or (iv) blocking a co-stimulatory signal (decreasing responses). Interpretation of these experiments relies on the fact that intact antibodies generally deliver operative signals whereas using Fab fragments inhibits CTLA-4 interactions.

While initial studies utilizing CTLA-4 monoclonal antibodies appeared to indicate that CTLA-4 might play a role similar to CD28 in enhancing T cell activation (Linsley et al., 1993a), more recent studies have indicated that CTLA-4 plays a role in inhibiting T cell activation. These experiments showed that blocking CTLA-4 enhances T cell proliferation whereas cross-linking agonistic antibodies to CTLA-4 revealed potent immunosuppression of T cell proliferation (Walunas et al., 1994; Krummel & Allison, 1995; 1996; Walunas, Bakker & Bluestone, 1996). Most critically, CTLA-4 knock-out mice develop fatal lymphoproliferative disease (see below), indicating the importance of CTLA-4 in maintaining self-tolerance. The nature of this inhibitory pathway is, as yet, unidentified; however, data from two laboratories (Krummel & Allison, 1996; Walunas et al., 1996) indicate that CTLA-4 can block T cell function at a relatively early stage (within 48 hours), preventing upregulation of activation markers, entry into the cell cycle and the generation of IL-2. Strikingly, these effects are seen when surface levels of CTLA-4 are undetectable. One further recent study has also suggested that CTLA-4 may be required for the induction of anergy (Perez et al., 1997), again consistent with a negative role for CTLA-4 but suggesting that the induction of T cell unresponsiveness requires CTLA-4. This result is difficult to reconcile with the view that lack of CD80/CD86 during TCR engagement in vitro can also induce anergy, and it may reflect an additional mechanism for anergy induction.

The above data provide a convincing picture of CTLA-4 as an inhibitor of T cell activation; however, there are still studies that do not fit this general picture (Gribben *et al.*, 1995; Wu *et al.*, 1997). Furthermore, at present, there is little direct evidence of the ability of natural ligands to provide inhibitory signals in a similar manner to the anti-CTLA-4 antibodies and, most obviously, nearly all the inhibitory data are based on murine studies. While it is likely that natural ligands do stimulate CTLA-4 function, the circumstances under which CTLA-4 predominates have yet to be established. Likewise, the kinetics with which CTLA-4 is utilized by its natural ligands also remains unclear. To date, most

T cell experiments using transfected ligands indicate that engagement of CD80/CD86 in the presence of anti-CD3 appears effectively to deliver proliferative signals via CD28, with little evidence for CTLA-4 function under these circumstances (Linsley *et al.*, 1991a; Gjorloff Wingren *et al.*, 1993). Therefore, while the role of CTLA-4 as an inhibitory molecule appears to be of fundamental importance to the control of self-tolerance, a considerable amount remains to be learnt about how and when CTLA-4 is utilized *in vivo*.

Studies in knock-out mice

The use of transgenic and knock-out technologies has now been applied with considerable effect to the study of CD28/CTLA-4 interactions. Knock-out approaches have been successfully performed to generate CD28, CTLA-4, CD80, CD86 and CD80/CD86 double knock-out mice (Shahinian *et al.*, 1993; Green *et al.*, 1994; Tivol *et al.*, 1995; Waterhouse *et al.*, 1995; Borriello *et al.*, 1997). In general, results from these mice provide support for the conclusions reached from the above *in vitro* and *in vivo* studies.

CD28-deficient mice

Studies on mice deficient for CD28 reiterate the positive role played by CD28 in T cell activation. These mice have impaired responses to mitogens and superantigens (Shahinian *et al.*, 1993); although proliferative T cell responses are seen in these mice, these responses appear to be of much lower magnitude and shorter duration (Lucas *et al.*, 1995). However, while CD28 clearly enhances and prolongs T cell responses, it does not appear to be strictly essential for all forms of T cell activation, providing support for the view that further important co-stimulatory ligands probably exist. For example, T cells from CD28-deficient mice display a number of surprisingly intact T cell responses, including the ability to reject skin allografts (Kawai *et al.*, 1996) and generate graft-versus-host reactions (Speiser *et al.*, 1997).

In addition to influencing whether or not a proliferative response is obtained, there is also evidence which suggests that CD28 may qualitatively affect the outcome of TCR engagement. In particular, CD28 co-stimulation may be involved in the ability to make T_H^2 responses (Rulifson *et al.*, 1997). However, this may depend on the nature of the antigenic challenge (Gause *et al.*, 1997a,b).

CD28-deficient mice display profound deficiencies in antibody production in addition to their T cell defects. These mice lack germinal centres (Ferguson *et al.*, 1996) and have strongly impaired ability to generate class-switched antibody isotypes, showing similar defects as mice treated with CTLA-4-Ig (Linsley *et*

al., 1992). It would appear that one of the major functions of CD28 *in vivo* is to provide T cell help for antibody production, a function that most likely also involves the interaction between CD40 and its T cell ligand (CD40L), which has been shown to be dependent on CD28 for expression (Klaus *et al.*, 1994; Somoza & Lanier, 1995; Yang & Wilson, 1996). These data appear to confirm the roles of CD28 in initiating and sustaining T cell responses, protecting from T cell apoptosis and facilitating efficient B cell help.

CD80- and CD86-deficient mice

Perhaps unsurprisingly, mice deficient in CD80 or CD86 ligands have a phenotype that is highly reminiscent of the CD28 knock-out. For example, mice that lack CD80 or CD86 either singly or in combination demonstrate defects in immunoglobulin class switching and germinal centre formation. This is interpreted as highlighting the requirement for CD28-dependent T cell help in many of these responses. In addition, there is a spectrum of severity in these knockout mice. Mice lacking CD80 have a relatively mild phenotype, consistent with the concept that CD86 is the major primary ligand, whereas mice deficient for CD86 have a more severe phenotype, again affecting antibody class switching. This is especially noticeable when immunizations occur in the absence of adjuvant, suggesting that CD80 can compensate when induced by inflammation. However, mice lacking both CD80 and CD86 have the most severe phenotype, which is very similar to that found in mice defective for CD28 in that they lack germinal centres and have highly defective T cell-dependent antibody responses as well as defects in T cell activation (Sharpe, 1995; Borriello et al., 1997; Schweitzer et al., 1997). Interestingly, these mice are not specifically defective in either $T_{H}1$ or $T_{H}2$ responses, suggesting no obligatory role for either CD80 or CD86 in these responses.

CTLA-4-deficient mice

The most revealing in this series of knock-out mice have been mice deficient for CTLA-4. These animals provide powerful evidence that CTLA-4 is a negative regulator of T cell function since they develop a spectacular lymphoproliferative disease that proves fatal a few weeks after birth (Tivol *et al.*, 1995; Waterhouse *et al.*, 1995). Analysis of the T cell compartment reveals a dramatic expansion of mature CD25⁺ activated T cells, consistent with an unchecked expansion of activated T cells. One possibility is that these mice fail to downregulate TCR and/or CD28 activation signals following antigenic stimulation. This hypothesis is supported by the fact that these mice can be effectively treated with CTLA-

4-Ig, suggesting that the lymphoproliferation is CD80/CD86 co-stimulation dependent (Tivol *et al.*, 1997). This implies that CTLA-4 can negatively regulate either or both the TCR and CD28 pathways. Biochemical analysis of these mice is still in its early stages but hyperactivation of *src* kinases has been suggested as one possible mechanism (Marengere *et al.*, 1996). However, given that this study investigated T cells that were clearly activated, this may be an effect of uncontrolled T cell expansion rather than the cause. This study also indicated that CTLA-4 may interact with the Syp phosphatase, and that this may be involved in inhibition of activation (Marengere *et al.*, 1996); however, this work has yet to be confirmed. Therefore, despite significant gaps in our understanding of the mechanism of CTLA-4 function, the importance of this receptor in negatively regulating T cells is beyond doubt. Clearly, given the essential role of CTLA-4, a complete understanding of its functions will provide significant opportunities for immune intervention and therapies for autoimmune disease.

CD28/CTLA-4 signalling mechanisms

A detailed understanding of the signalling mechanisms utilized by CD28 and CTLA-4 is still being elucidated and has been the subject of several extensive reviews (June *et al.*, 1994; Ward, June & Olive, 1996; Sansom *et al.*, 1997). To date, there is little information on the mechanisms used by CTLA-4 and, therefore, only the salient features of CD28 signalling will be reviewed here.

The cytoplasmic domains of CD28 and CTLA-4 are 41 and 36 amino acids residues long, respectively, and, in common with many receptors of the immune system, do not contain any intrinsic enzymatic activity. The most obvious and intensively studied region of interest is the YMNM motif based around tyrosine (Y) 173 in CD28. This motif has been identified as a binding site for protein SH-2 domains, which interact with phosphorylated tyrosine residues. In particular, this motif effectively recruits the p85 subunit of the enzyme phosphatidylinositol 3-kinase (PI₃K) and it has been established that CD28 recruits and activates this lipid kinase (Ward et al., 1993; 1995; Pages et al., 1994; Prasad et al., 1994; Cai et al., 1995) upon ligand binding. The role of PI₃K in the downstream events of CD28 signalling is still controversial, in particular with respect to its requirement in IL-2 responses (Crooks et al., 1995). Nonetheless, given the increasing evidence that CD28 is required for cell survival and proliferation, it is likely that these may provide important targets for PI₃K. This possibility is strengthened by the fact that recently elucidated downstream targets include protein kinase B and p70 S6kinase, which are essential for cell proliferation (Burgering & Coffer, 1995; Downward, 1995; Franke et al., 1995). Given that the YMNM motif requires tyrosine phosphorylation before it can interact with PI_3K , there have been a number of efforts to identify the kinase responsible. These have largely concentrated on *src*-coded kinases, such as Lck and Fyn as well as Tec family kinases such as Itk. At present, the best evidence suggests that Lck is capable of phosphorylating CD28; however, it is not yet clear whether this occurs *in vivo* (Raab *et al.*, 1995). Interestingly, while Itk has been shown to be phosphorylated following CD28 activation (August *et al.*, 1994), recent data suggest that Itk performs a negative regulatory role in T cell proliferation and is, therefore, not a likely candidate for CD28 co-stimulatory signals (Liao *et al.*, 1997).

Further downstream targets for CD28 signals include the activation of Jun amino terminal kinase, and it has been suggested that activation of this kinase may be a point of integration between signals from the TCR and CD28 (Su *et al.*, 1994). Other studies of CD28 have suggested a role for the enzyme acidic sphingomyelinase, which results in the subsequent production of ceramide (Boucher *et al.*, 1995). This pathway is a candidate for transmitting signal to Jun kinase as it is clear that other potent activators of Jun kinase such as TNF α also trigger sphingomyelinase activation (Wiegmann *et al.*, 1994).

Since a major effect of CD28 signalling is on cytokine production, transcription factor targets that bind to cytokine promoters have been sought which are responsive to CD28 signals. In this respect, a number of transcription factors have been observed to be regulated by CD28, including a putative CD28-response element (CD28RE) as well as other targets such as AP-1 and NF κ B (Fraser *et al.*, 1991; Verweij, Geerts & Aarden, 1991; Fraser & Weiss, 1992; Edmead *et al.*, 1996). Interestingly, one of the major features of CD28 signalling is its resistance to the drug cyclosporin A, which is a potent inhibitor of the transcription factor NFAT. The fact that CD28 can still induce proliferation in the presence of cyclosporin suggests that CD28 signals may provide an alternative pathway to NFAT activation (Ghosh *et al.*, 1996).

Relevance of CD28 and CTLA-4 to rheumatic diseases

The role of T cells in RA

Given that a major hypothesis of the aetiology of many autoimmune conditions invokes a breakdown in T cell self-tolerance provoked by an environmental antigenic stimulus, then manipulation of the CD28/CTLA-4 system is an attractive target for therapeutic intervention. While there is a solid basis suggesting an autoimmune component in the development of RA, the aetiology of this disorder is largely unknown, and the relative contribution of T lymphocytes to disease has been controversial (Firestein & Zvaifler, 1990). In particular, the evidence that inflammatory cytokines such as TNF α and IL-1 are clearly involved in joint inflammation, and the relative lack of T cell cytokines such as IL-2, has prompted questions about the role of T cells (Feldman, Brennan & Maini, 1996). To some extent, this may result from problems of timing, since analysis of patients with classical RA may occur well after incitement of disease and observations will predominantly reflect inflammatory sequelae. Alternatively, it may be that expectations of T cell infiltration by a limited number of causative T cell clones in affected joints is over optimistic. Nonetheless, it is evident that in virtually all models where joint disease initiation can be studied, T lymphocytes play a central, causative role (Kadowaki *et al.*, 1994; Mima *et al.*, 1995; Sakata *et al.*, 1996; Tada *et al.*, 1996; Stasiuk *et al.*, 1997).

This evidence is strengthened by the classical observation that the major genetic predisposition to RA development maps to the HLA region (Stastny, 1978; Sansom *et al.*, 1987; 1989; Lanchbury *et al.*, 1991). Detailed studies of the protein sequences of the HLA susceptibility alleles gave rise to the 'shared epitope theory' proposed by Gregersen and colleagues (Gregerson *et al.*, 1987) in which the protein sequence of the polymorphic third hypervariable region (HVR) of the HLA-DR1 and HLA-DR4 appears to predispose to disease. Since the only known function of HLA-DR molecules is the presentation of peptide fragments to T cells, this has been used to support a central role for antigen-specific T cell activation in RA aetiology.

The recent identification by Mathis and co-workers of a novel animal model of spontaneous arthritis has provided a new perspective on the initiation and progression of autoimmune conditions (Kouskoff *et al.*, 1996). In this model, a novel mouse strain was generated by breeding a TCR transgenic line (KRN) onto the NOD background. This strain spontaneously develops a symmetrical erosive arthritis that is characterized by features highly reminiscent of RA. Analysis of this strain indicated a mechanism involving the recognition of a specific mouse class II molecule and yet, significantly, despite having a definitive T cell-MHC allele-dependent disease, T cells from diseased animals displayed poor proliferative responses and were difficult to detect in affected joints. Therefore, the lack of obvious T cell activity in the RA synovial compartment does not preclude a causative role for T cells in RA. Accordingly, there is still considerable scope for therapeutic approaches based on manipulating the CD28/CTLA-4 system.

Manipulation of CD28 interactions in autoimmune disease

A significant number of studies have now been carried out to investigate the effects of CD28/CTLA-4 manipulations in disease situations, including several autoimmune models as well as transplantation and tumour therapy settings. Initial

successes using CTLA-4-Ig in transplant models, suggested that blocking both CD80 and CD86 could be used as an effective means to prevent graft rejection as well as to generate subsequent tolerance (Lenschow et al., 1992; Linsley et al., 1992). Subsequently, CTLA-4-Ig has been utilized in a number of autoimmune conditions. Particularly striking has been the treatment of lupus-prone NZB/NZW (F1) mice with soluble murine CTLA-Ig. This effectively inhibited T-dependent primary immune responses, leading to suppression of autoantibody production, decreased renal disease and enhanced survival rates (Finck, Linsley & Wofsy, 1994). Encouragingly, initiation of CTLA4-Ig therapy late on in disease progression (when lupus-associated mortality had reached 40%) still inhibited further production of autoantibodies and markedly prolonged life (Finck et al., 1994), indicating significant efficacy even in the context of a well-established immune response. CTLA-4-Ig has also been utilized successfully in the amelioration of collagen-induced arthritis (Webb, Walmsley & Feldmann, 1996). Here again, administration at the time of antigen challenge was effective at preventing arthritis and effects were apparent even when CTLA-4-Ig was administered later in disease progression.

While studies in models of rheumatic diseases are still limited, more detailed work has been carried out in the context of other autoimmune conditions. In general, studies in experimental allergic encephalomyelitis (EAE) have shown similar results in that early administration of CTLA-4-Ig prevented disease onset (Miller *et al.*, 1995; Perrin *et al.*, 1995; 1996; Racke *et al.*, 1995). Therefore, blocking CD80 and CD86 interactions necessary for the activation of T cells, and consequently B cells, can establish effective immunosuppressive regimens. Similarly, initial experiments in the NOD mouse have established that administration of CTLA-4-Ig blocked the establishment of full blown diabetes (Lenschow *et al.*, 1995). Therefore, CTLA-4-Ig has been generally found to be efficient at suppressing and treating a variety of T cell-driven autoimmune diseases.

However, the NOD model has also revealed unexpected complexity in manipulating the CD28/CTLA-4 system. In particular, breeding of the NOD mouse onto a CD28 knock-out mouse resulted in increased incidence and severity of disease instead of the predicted amelioration (Lenschow *et al.*, 1996). Strikingly, the CD28-negative background allowed 80% of the male mice, which are normally largely unaffected, to become diseased. Furthermore, the use of anti-CD80/CD86 antibodies has also had mixed results, including both suppression and exacerbation of disease (Lenschow *et al.*, 1995; 1996). One hypothesis is that this relates to the requirements for CD80 and CD86 to influence differentially $T_H 1$ and $T_H 2$ cell development. In some diseases $T_H 2$ responses may be protective, and, therefore, blocking CD28-related $T_H 2$ effects may exacerbate

disease. While a differential signalling role of CD80 versus CD86 is unlikely in view of the data from CD80 and CD86 knock-out mice (Borriello *et al.*, 1997; Schweitzer *et al.*, 1997), an alternative view is that T_H^2 development is dependent on the 'strength of signal' (Sperling & Bluestone, 1996). Accordingly, T cells that receive CD28 co-stimulation either via CD80 or CD86 have a greater propensity to develop T_H^2 responses, as indicated by increased IL-4 production. This hypothesis has received some experimental support, which indicates that co-stimulation may indeed be influential in development of T_H^2 responses (Seder *et al.*, 1994; Freeman *et al.*, 1995; Thompson, 1995; Gause *et al.*, 1997b; Rulifson *et al.*, 1997). However, this finding is not universal and may depend on the nature of the antigenic challenge (Brown *et al.*, 1996; Gause *et al.*, 1997a; Schweitzer *et al.*, 1997).

At first sight, it is difficult to reconcile the preventative effects of CTLA-4-Ig treatment with the exacerbation in NOD × CD28 knock-out mice. However, reevaluation of the treatment of NOD mice with CTLA-4-Ig has revealed that the timing of CTLA-4-Ig administration has a critical influence on disease outcome and that CD28 functions may differ at different times of treatment (Sperling & Bluestone, 1996).

In principle, the molecule with the most obvious therapeutic potential in autoimmune disease is CTLA-4. Since this molecule is a potent 'off switch' for T cells, CTLA-4 agonists may prove very useful for downregulating autoimmune responses. As yet, however, there are few data studying the effects of CTLA-4 agonists in autoimmune disease models. Encouragingly, however, where antagonists of CTLA-4 have been used, they have had spectacular and predicted results. Most striking is the enhancement of anti-tumour immunity following the blockade of CTLA-4. Here, mice rejected unmanipulated tumour cells when treated with anti-CTLA-4, whereas control mice universally succumbed to tumours (Leach, Krummel & Allison, 1996). More recently, other studies have shown that blocking CTLA-4 exacerbates autoimmune EAE models, again suggesting CTLA-4 is involved in downregulating autoimmune responses (Perrin et al., 1996; Hurwitz et al., 1997). Finally perhaps the most vivid demonstration that CTLA-4 may normally be involved in preventing autoimmunity is the fatal disease seen in CTLA-4 knock-out mice in which destructive myocarditis, pancreatitis and infiltration of lungs and liver have been described.

The notable successes of manipulating the CD28/CTLA-4 system in many disease models indicate that this pathway has excellent potential as a route for controlling autoimmune disease. However, the fact that some manipulations lead to disease exacerbation and the fact that CD80/CD86 ligands operate both inhibitory and stimulatory controls over T cells suggests the need for a cautious approach driven by a more detailed understanding of the system. Nonetheless,

the benefits offered in terms of potential long-term tolerance, as well as the ability to manipulate only activated T cells, make this system an extremely promising area for developing future therapy.

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7

Lymphocyte antigen receptor signal transduction D. R. ALEXANDER

Introduction

Lymphocytes can be separated into thymus-derived cells (T cells) that are specialized for protecting the body against intracellular pathogens (cell-mediated immunity) and the bone-marrow derived cells (B cells, originally described in the bursa of Fabricius in the chicken) that protect against extracellular pathogens (humoral immunity). The specificity of these responses is achieved by the expression of clonotypic B and T cell antigen-specific receptors on the lymphocyte surface, known as the BCR and TCR, respectively. The BCR is specialized for the recognition, capture and internalization of antigens for processing and presentation on the cell surface in association with MHC class II molecule. The TCR is specialized for the recognition of small peptides presented on antigen-presenting cells in association with MHC class I or class II molecules. The peptides are typically 8–10 amino acid residues long when in association with class I molecules in association with class II molecules.

The outcome of BCR or TCR engagement by ligands depends on the avidity of the interaction, the length of time for which ligands bind, the lymphocyte differentiation state and the nature of other co-stimulatory signals being received by the cell. A fascinating aspect of antigen receptor signalling is that the same receptor can mediate signals resulting in apoptosis, activation or cellular non-responsiveness ('anergy') depending on the interplay between these four variables. During T cell development in the thymus, for example, selection events occur at the CD4+CD8+ stage of differentiation that define which T cells will mature and eventually exit to the periphery as CD4+ and CD8+ cells. High-avidity interactions between MHC–peptides and the TCR expressed on CD4+CD8+ thymocytes cause their deletion by apoptosis. When ligand–receptor interactions of equivalent avidity occur on mature peripheral T cells, however, the result is

the activation and clonal expansion of the T cell population. In the absence of the correct cohort of co-stimulatory signals, the same ligand–TCR engagement could result in a state of peripheral T cell non-responsiveness. A similar diversity of outcomes characterizes BCR engagement. For example, when cell-bound polyvalent antigens bind to the BCR on immature B cells, then the cells die by apoptosis, whereas the same stimulus leads to a proliferative response in mature B cells.

The differences in the signalling pathways that mediate these radically different outcomes still remain poorly understood. Elucidation of these differences is critical for our understanding of tolerance, autoimmunity and immunosuppression, not least in the context of lymphocyte pathology in the inflamed synovium. From the practical point of view, it is also important to remember when assessing the rapidly growing literature on lymphocyte signalling pathways that the BCR and TCR do not necessarily mediate identical signals in any particular B or T cell. These will depend on the variables already listed above. Relevant questions to ask about any study of lymphocyte signalling, therefore, include the following: 'Are the lymphocytes being used primary cells or transformed cell lines?'; 'What is the differentiation state of the cells?'; 'Are the lymphocytes being triggered with antibodies or with physiological ligands?'; and 'Are co-receptors being stimulated at the same time as engagement of antigen receptors?'.

This chapter will focus mainly on the signals mediated by antigen receptors expressed on mature lymphocytes and on the regulation of receptor thresholds by CD45. The role of co-receptors such as CD4/CD8 on T cells, and CD22 and CD19/CD21 on B cells, which are intimately involved in antigen receptor signal transduction, will also be briefly summarized. Extensive reviews are available describing signalling pathways in T cells (Chan, Desai & Weiss, 1994a; Cantrell, 1996; Wange & Samelson, 1996; Alexander, 1997; Berridge, 1997; Frearson & Alexander, 1997) and in B cells (Cambier, Pleiman & Clark, 1994a; Pleiman, Dambrosio & Cambier, 1994; DeFranco, 1997; Kurosaki, 1997; O'Rourke, Tooze & Fearon, 1997).

It should be noted that at the level of lymphocyte cell biology the immediate consequences of ligand binding to the BCR and TCR are distinct. Thus, engagement of the BCR expressed on mature quiescent B cells by multimeric antigens results directly in cell proliferation in a T cell-independent manner. Engagement of the TCR on mature quiescent T cells, however, does not provide a proliferative signal, *per se*, but rather promotes the entry of the cells into the cell cycle, known as the G_0 to G_1 transition (Fig. 7.1). During this transition TCR-mediated signals cause the induction of IL-2 and IL-2 receptor- α (CD25) genes with the consequent secretion of IL-2 and expression of high-affinity IL-2 receptors on the cell surface. The subsequent autocrine or paracrine binding of IL-2 to IL-



Fig. 7.1. The two major phases of T cell activation.

2 receptors then induces a panoply of signals that cause DNA synthesis and T cell proliferation. Many of these signals are quite distinct from those mediated by the TCR. Thus B cell activation is a one-phase process whereby engagement of the BCR by appropriate antigens triggers the full gamut of proliferative signals, whereas T cell activation is a two-phase process comprising antigen-driven entry into the cell cycle followed by cytokine-induced proliferation.

The structure of antigen receptors

Characterization of the molecular structures of the BCR and TCR have provided critical insights into the way these receptors couple to intracellular signalling pathways. The TCR comprises the polymorphic α and β subunits, which recognize MHC-associated peptides, in non-covalent association with the invariant transmembrane CD3 γ , δ and ε chains and the TCR- ς chain homodimer (Fig. 7.2*a*) (Weiss, 1993). These invariant polypeptides are responsible for stabilizing the assembly of the receptor to enable its expression at the cell surface, and for coupling the TCR to intracellular signalling pathways. The equivalent antigenrecognizing component of the BCR is surface immunoglobulin (sIg), a tetrameric complex of immunoglobulin heavy and light chains (Fig. 7.2*b*) that are extensively



Fig. 7.2. The structures of the TCR and BCR.

homologous to the immunoglobulins which are secreted by differentiated daughter cells (Cambier et al., 1993). These polymorphic chains are non-covalently associated with an invariant disulphide-linked transmembrane heterodimer of Ig- α (CD79a) and Ig- β (CD79b) polypeptides, which are encoded by the immunoglobulin superfamily genes *mb-1* and *B29*, respectively. The same $Ig-\alpha/Ig-\beta$ heterodimer is associated with all of the five different heavy (H) chain classes of sIg molecules (Venkitaraman et al., 1991), and plays an analogous role to that of the CD3/TCR- ς chains in both receptor assembly and signal transduction. In addition, the cytoplasmic tails of Ig- α and Ig- β are involved in the internalization of proteins bound to the BCR and their subsequent targeting to endosomal compartments for processing (Bonnerot et al., 1995). Interestingly, however, the short cytoplasmic tails of sIg H chains, which vary in length from 3 to 28 amino acid residues, are also involved in targeting receptor-bound antigens to endosomes (Weiser et al., 1997), and, in some cases, antigen internalization and presentation in memory B cells is largely independent of Ig- α and Ig- β expression (Tarlinton, 1997). It is, therefore, possible that sIg exists in two pools, either with or without association with Ig- α /Ig- β . Upon sIg ligation by antigen, both receptor pools would mediate antigen internalization and processing, whereas only the pool containing $Ig-\alpha/Ig-\beta$ would induce intracellular signals.

The invariant chains of the TCR and BCR contain 'immunoreceptor tyrosine-based activation motifs' (ITAMs) comprising the sequence

E/DX₂YX₂L/IX₇YX₂L/I (single-letter code for amino acids where X represents any amino acid). The TCR- ς chain contains three ITAMs, whereas the CD3, Ig- α and Ig- β chains each contain one ITAM. Upon binding of ligands to the TCR or BCR, the tyrosine residues of the ITAMs are phosphorylated by tyrosine kinases, so forming high-affinity binding sites for signalling proteins, which engage the phosphorylated sequences by means of their Src homology type 2 (SH2) domains. In this way, protein complexes are recruited to the receptors, which, in turn, activate intracellular signalling pathways involved in transcription factor regulation. It is not yet clear whether each phosphorylated ITAM *in vivo* has a selective binding capacity for different proteins or whether the ITAMs of a receptor bind the same proteins, thereby providing a signal amplification system. Evidence is available for both scenarios.

Engagement of the TCR or BCR by ligands does not necessarily induce optimal ITAM phosphorylation. For example, the binding of altered peptide ligands (APLs) to the TCR expressed on T cell clones can cause the generation of abnormal phosphoisomers of TCR- ς , which are probably incompletely phosphorylated (Sloan-Lancaster *et al.*, 1994; Madrenas *et al.*, 1995). Such signals correlate with the induction of states of cellular non-responsiveness in which cells do not proliferate or secrete IL-2 upon subsequent challenge with an MHC-peptide (Sloan-Lancaster & Allen, 1996). Abnormal ITAM phosphorylation states may, therefore, be involved in the induction of cellular non-responsiveness.

Tyrosine kinases and antigen receptors

In contrast to growth factor receptors, such as the epidermal growth factor receptor, in which tyrosine kinase activity is expressed within the receptor cytoplasmic tail, antigen receptor polypeptides do not possess any kinase activity. Instead they utilize cytosolic tyrosine kinases for phosphorylating their ITAM motifs and for triggering a cascade of phosphorylation events that involve the regulation of serine/threonine protein kinases.

Tyrosine kinases and TCR signalling

The role of tyrosine kinases in TCR signalling is more clearly defined than for BCR signalling. There are four families of tyrosine kinases now known to be involved in TCR signal transduction (Chan *et al.*, 1994a; Qian & Weiss, 1997). The first comprises the p59^{fyn} and p56^{lck} members of the Src family of tyrosine kinases. As Fig. 7.3 illustrates, these kinases share the same domain structure, with myristoylated and palmitylated amino terminal domains which tether the kinases to the plasma membrane, and which also bear receptor-binding sequences.

The SH3 domain interacts with proteins carrying proline-rich motifs and the SH2 domain interacts by an intramolecular association with a regulatory phosphorylated tyrosine residue in the carboxy-terminus of the kinase. Upon dephosphorylation of this tyrosine residue, the SH2 domain is then available to engage with other tyrosine phosphorylated proteins. The p59fyn kinase associates directly with rather weak affinity and low stoichiometry with the TCR (Samelson et al., 1990). The p56^{lck} kinase associates with the CD4 and CD8 co-receptors with a considerably higher affinity and stoichiometry (Rudd et al., 1988). The expression of CD4 is characteristic of helper T cells whereas CD8 expression is a marker of cytotoxic T cells. It is thought that when an MHC-peptide engages the TCR, the class I or class II molecule simultaneously binds to CD8 or CD4, respectively, thereby bringing the associated p56^{lck} kinase into association with the CD3/TCR-c ITAMs, so causing their phosphorylation. The marked reduction in TCR- ς and CD3- ϵ phosphorylation observed in mice lacking p56lck expression is consistent with such a model (van Oers, Killeen & Weiss, 1996a). Furthermore, there is a correlation between the ability of TCR antibodies to stimulate T cell activation and their efficiency at promoting CD4-TCR association and TCR-c phosphorylation (Janeway, 1992). In contrast, in mice lacking p59fyn, TCR-c phosphorylation is normal (van Oers et al., 1996b), suggesting that the role of p56^{lck} is more critical than that of p59^{fyn} in regulating ITAM phosphorylation. This conclusion is supported by the severe defects in thymic development observed in Lck-/- mice, which contrast with the normal thymic development that occurs in Fyn-/- mice (Molina et al., 1992). Interestingly, however, in mice lacking both p56lck and p59fyn the block in T cell development is more severe than in Lck-/- mice (van Oers et al., 1996b), suggesting that there could be some overlapping functions between these two kinases. Furthermore, the proliferation of mature T cells is not completely ablated in Lck-/- mice, suggesting that p56lck is not the only kinase involved in coupling the TCR to activation pathways. Therefore, a role for p59^{fyn} in phosphorylating ITAMs in certain contexts cannot yet be excluded.

The second family of kinases involved in TCR signal transduction is represented by Csk (C-Src kinase). As shown in Fig. 7.3, this kinase phosphorylates the regulatory carboxy terminal tyrosine residues in Src kinase family members, thereby inhibiting their ability to phosphorylate relevant substrates.

The third family of kinases is represented by ZAP-70 (ς -associated protein-70) and its homologue Syk (Fig. 7.4.). ZAP-70 is more important than Syk in TCR signalling, whereas ZAP-70 is not expressed in B cells. The critical role of ZAP-70 in TCR signalling is illustrated by patients with severe combined immunodeficiency (SCID) carrying mutations of ZAP-70 resulting in deficient ZAP-70 protein levels or loss of its kinase activity (Arpaia *et al.*, 1994). Whereas



Fig. 7.3. The structure and regulation of the p56^{lck} and p59^{fyn} tyrosine kinases. CS kinase, C-Src kinase. The numbers refer to amino acids that demarcate domains or which are phosphorylated.

CD4+CD8+ thymocytes are present in these patients, only CD4+ and not CD8+ T cells emerge to the periphery. The peripheral CD4+ cells do not proliferate in response to TCR stimulation, fail to secrete IL-2 and have markedly reduced TCR-induced protein tyrosine phosphorylation, suggesting that ZAP-70 plays a central role in mediating T cell activation. Mice deficient in ZAP-70 expression have an even more severe phenotype in which there is a failure of both positive and negative selection events so that neither CD4+ nor CD8+ T cells exit to the periphery (Negishi *et al.*, 1995). The residual ZAP-70 function that may remain in SCID patients could explain why their phenotype is less severe than that of ZAP-70^{-/-} mice, and the bias to maturation of CD4+ T cells in these patients most likely results from the greater affinity of p56^{lck} binding to CD4 than to CD8. The secondary importance of Syk for T cell signal transduction is illustrated by the normal T cell development which occurs in Syk^{-/-} mice (Turner *et al.*, 1995).

ZAP-70 and Syk both possess two SH2 domains but lack the carboxy terminal regulatory tyrosine residues that are characteristic of the Src kinases. Upon TCR stimulation, ZAP-70 and Syk engage via both their SH2 domains with doubly phosphorylated TCR- ζ ITAMs (Iwashima *et al.*, 1994; Isakov *et al.*, 1995), thereby facilitating their phosphorylation by kinases such as p56^{lck} and



Fig. 7.4. The structure and regulation of the ZAP-70 tyrosine kinase. The major sites of tyrosine phosphorylation are indicated.

p59^{fyn}. The crystal structure of two ZAP-70 SH2 domains bound to a phosphorylated ITAM motif has revealed the importance of this interaction for the conformation of the kinase (Hatada *et al.*, 1995). Thus, the binding pocket for the amino terminal phosphorylated tyrosine in the ITAM is formed by the carboxy terminal SH2 domain alone of ZAP-70. In contrast, the formation of the binding pocket for the carboxy terminal phosphorylated tyrosine in the ITAM requires residues provided by both SH2 domains, in the process facilitating a coiled-coil structure in the interdomain region between the two SH2 domains. The dual interaction between the two SH2 domains and the two phosphorylated ITAM tyrosine residues helps to explain the high affinity of interaction between ZAP-70 and TCR- ς .

The binding of ZAP-70 to phosphorylated ITAM motifs *per se* is insufficient to cause kinase activation. This has been confirmed both by *in vitro* peptidebinding studies and by the observation that in murine thymocytes ZAP-70 is constitutively associated with the ς -chain without activation (van Oers *et al.*, 1994). In fact activation of ZAP-70 requires phosphorylation by p56^{lck} at Tyr-493 on the putative regulatory loop of its kinase domain (Chan *et al.*, 1995), which is followed by further phosphorylation of the kinase (Fig. 7.4.), involving both positive and negative regulatory sites, some of which results from transphosphorylation ('autophosphorylation') between the activated ZAP-70 molecules (Wange *et al.*, 1995). An actual increase in ZAP-70 kinase activity is critical for coupling the TCR to intracellular signals (Wiest *et al.*, 1997). There is some evidence that several ZAP-70 molecules need to be juxtaposed on adjacent phosphorylated ITAM motifs (Fig. 7.5), in order for effective ZAP-70 trans-



Fig. 7.5. The aggregation model of ZAP-70 regulation. (*a*) Some evidence suggests that when several ZAP-70 molecules are bound in tandem to the three phosphorylated ITAMs present in TCR- ζ then ZAP-70 autophosphorylation (more strictly transphosphorylation) proceeds more efficiently, so generating binding sites for SH2-domain-containing proteins (Neumeister *et al.*, 1995). (*b*) In contrast, when only a single ZAP-70 molecule is bound to one phosphorylated ITAM, ZAP-70 autophosphorylation is inefficient.

phosphorylation to occur (Neumeister *et al.*, 1995). Phosphorylation of ZAP-70 results in the generation of binding sites for SH2 domain-containing proteins, leading to the further recruitment of signalling proteins (Neumeister *et al.*, 1995). It has also been suggested that ZAP-70 phosphorylation results in the recruitment of p56^{lck} via its SH2 domain, so providing a mechanism whereby the association of CD4/CD8-p56^{lck} with the TCR could be stabilized (Duplay *et al.*, 1994). Overall, therefore, ZAP-70 is critical for coupling the TCR to intracellular signalling pathways. When ZAP-70 is absent, Syk is able to substitute for its functions to a certain extent, suggesting some redundancy between these two kinases (Gelfand *et al.*, 1995). In normal T cells, however, it is not yet clear whether Syk has any actions in TCR signalling distinct from those of ZAP-70. Interestingly, Syk expression is relatively high in thymocytes, whereas it is expressed at low levels in mature peripheral T cells, pointing to a possible selective function in thymocytes (Chan *et al.*, 1994).

The fourth family of tyrosine kinases implicated in T cell signalling is the Tec family, and the member of this kinase preferentially expressed in T cells is Itk (Siliciano, Morrow & Desiderio, 1992). The Tec family of kinases share some homology with the Src kinase family but lack the amino-terminal myristoylation consensus sequence and carboxy terminal tyrosine residues that characterize the Src kinases. The Tec family of kinases are cytosolic but their pleckstrin homology (PH) domains may be involved in targeting them to membranes and/or in mediating their regulation by phospholipids. Mice lacking Itk have a reduced number of mature thymocytes, implicating Itk in T cell development, and their mature T cells proliferated poorly in response to antigenic stimulation, suggesting that Itk is involved in TCR signal transduction (Liao & Littman, 1995). The molecular mechanism of action of Itk in this context remains to be elucidated.

Tyrosine kinases and BCR signalling

The same four families of tyrosine kinases involved in TCR signalling have also been implicated in BCR signalling (Bolen, 1995; Kurosaki, 1997). The Src family members Blk, Lyn, Fgr and Fyn have been reported to associate with the BCR and to be activated following sIg cross-linking (Yamanashi *et al.*, 1991; Cambier *et al.*, 1993). Binding studies *in vitro* suggest that these kinases bind to resting receptors through an association of their amino terminal residues with the Ig- α chain (Clark *et al.*, 1992), an association that does not appear to require ITAM phosphorylation (Clark, Johnson & Cambier, 1994). Upon BCR ligation, the ITAMs within the Ig- α and Ig- β chains become phosphorylated and this is thought to result in further recruitment of Src family kinases via their SH2 domains, so promoting kinase activation (Clark *et al.*, 1994). An attractive model, therefore, involves the relatively weak association of Src family kinases with the BCR in resting B cells, which then become activated upon engagement of their SH2 domains with phosphorylated ITAMs (Pleiman *et al.*, 1994). Whether the Ig- α and Ig- β ITAMs are initially phosphorylated by a member of the Src family or by Syk is an issue that awaits a clear resolution. Of the various Src kinase family members implicated in BCR signalling, p59^{lyn} has received the most attention since peripheral B cells appear to be hyperresponsive in Lyn^{-/-} mice, which develop circulating autoantibodies despite apparently normal B cell development (Hibbs *et al.*, 1995). One possible explanation for this finding is that the dominant role of p59^{lyn} is to phosphorylate a tyrosine residue in the cytoplasmic tail of CD22, which normally exerts a negative action on BCR signalling (see below). In the absence of this phosphorylation event, BCR signalling might then be amplified (Kurosaki, 1997). Such an interpretation does not exclude a role for p59^{lyn} in phosphorylating Ig- α or Ig- β ITAMs, or indeed other substrates.

The role of Csk in suppressing the activity of Src kinases is the same in B cells as in T cells. As far as the third family of tyrosine kinases is concerned, Syk is clearly the key player in BCR signalling. In mice deficient for Syk, there is a developmental block in the pro-B to pre-B cell transition and, despite the production of small numbers of immature B cells, mature B cells fail to accumulate, suggesting a role for Syk in the production or maintenance of mature B cells (Turner *et al.*, 1995). Overall BCR signal transduction appears to be as dependent upon the actions of Syk as, for example, TCR signalling is dependent upon the actions of ZAP-70.

The way in which Syk is activated also appears to be different from the mechanism of ZAP-70 activation. Engagement of the two SH2 domains of Syk by phosphorylated ITAM peptides causes a change in conformation of Syk and kinase activation, but under these conditions ZAP-70 activation does not occur (Shiue, Zoller & Brugge, 1995; Kimura *et al.*, 1996). Upon BCR stimulation *in situ*, Syk is, therefore, recruited to phosphorylated Ig- α /Ig- β ITAMs, which causes its activation and transphosphorylation together with further phosphorylation by a Src family tyrosine kinase (Kurosaki *et al.*, 1994; Rowley *et al.*, 1995). Tyrosine-phosphorylated Syk then dissociates from the BCR by a process that is itself regulated by a tyrosine phosphorylation event within the Syk molecule (Keshvara *et al.*, 1997). As with ZAP-70, the phosphorylation of Syk leads to the formation of signal transduction complexes with other SH2 domain-containing proteins.

The fourth tyrosine kinase family is well illustrated in B cells by the important role played by Btk (Bruton's tyrosine kinase). Defects in the *btk* gene are associated with X-linked agammaglobulinaemia (XLA), the first primary immunodeficiency to be identified (by Bruton). Affected males have decreased levels of circulating immunoglobulins and a severe deficit in the number of B cells; however, the patients have normal myeloid and T cell function as well as intact cellular immunity, consistent with the lack of Btk expression in T cells. Ligation of the BCR in Btk-deficient splenic B cells results in severely defective proliferation when compared with wild-type cells (Khan *et al.*, 1995). The Src family kinases appear to be necessary for the BCR-mediated regulation of Btk activation (Afar *et al.*, 1996; Rawlings *et al.*, 1996) and Btk interacts via a proline-rich motif within its Tec homology domain with the SH3 domains of Src family kinases (Yang *et al.*, 1995). Analysis of signal transduction pathways in a B cell line lacking Btk has implicated the kinase in the coupling of the BCR to increased phosphatidylinositol hydrolysis and calcium signalling (see below) (Takata & Kurosaki, 1996). The downstream signals regulated by Btk in primary cells remain the focus of active research (Kurosaki, 1997).

The regulation of tyrosine kinases by the CD45 phosphotyrosine phosphatase

CD45 is a transmembrane phosphotyrosine phosphatase that regulates the threshold of TCR and BCR signal transduction (Fig. 7.6) (Frearson & Alexander, 1996; Alexander, 1997). CD45 has a large ectodomain with a receptor-like structure, although physiologically relevant ligand(s) remain to be identified. Studies utilizing mutant cell lines and transgenic mice have established that CD45 exerts a positive regulatory role on both TCR- and BCR-mediated signals (Pingel & Thomas, 1989; Koretzky et al., 1990; Justement et al., 1991; Kishihara et al., 1993). In CD45 mice there are severe defects in T cell development, particularly in the positive selection of CD4+CD8+ thymocytes (Byth et al., 1996). The few mature T cells that exit to the periphery in these mice do not proliferate upon TCR stimulation (Stone et al., 1997). Studies using mutant cell lines and CD45-/mice have established that in T cells p56lck and p59fyn are substrates for CD45 (Ostergaard et al., 1989; Shiroo et al., 1992; Stone et al., 1997). In the absence of CD45, these kinases are hyperphosphorylated at their regulatory carboxy terminal tyrosine residues and are in their closed 'inactive' conformations (Sieh, Bolen & Weiss, 1993; Stone et al., 1997). As a consequence, there are extensive defects in TCR signal transduction coupling. For example, TCR ς chain phosphorylation is much reduced in the absence of CD45, consistent with an important role for CD45-activated p56^{lck} in phosphorylating the TCR ζ chain. Furthermore, upon TCR ligation an abnormally phosphorylated form of the ζ chain is generated (Stone et al., 1997), which is reminiscent of the partial signals induced upon engagement of the TCR with altered peptide ligands (Sloan-Lancaster et al., 1994; Madrenas et al., 1995). As a result, ZAP-70 fails to be



Fig. 7.6. Regulation of p56^{lck} by the CD45 phosphotyrosine phosphatase is critical for phosphorylation of TCR- ζ and CD3- ε ITAMs and subsequent recruitment of signal transduction complexes.

recruited efficiently to TCR-ς. Interestingly, however, TCR-mediated Syk phosphorylation is normal or even increased in the absence of CD45, underlining the relative independence of Syk regulation from regulation by the Src family kinases (Chu *et al.*, 1996). Not surprisingly in light of the early point in the signal transduction cascade at which CD45 acts, there are multiple downstream signalling defects in cells lacking CD45 (Shiroo *et al.*, 1992). However, it should be noted that in primary CD45^{-/-} thymocytes, some TCR-mediated signals still occur, and at high ligand concentrations calcium signals, for example, are only partially defective (Stone *et al.*, 1997). Therefore, in T cells, CD45 appears to act like a 'gatekeeper' to modulate the intensity of signals mediated by the TCR by regulating the actions of Src family kinases (Frearson & Alexander, 1997).

The role of CD45 in B cells appears less stringent than in T cells, perhaps reflecting the importance of CD45-regulated p56^{lck} in T cell development and activation, in contrast to any comparable role for a single CD45-dependent kinase in B cells. However, B cell maturation is defective in CD45^{-/-} mice, leading to a relative deficit of IgM-low, IgD-high B cells and an accumulation of the more

immature IgM-high, IgD-low B cells. Furthermore, splenic B cells fail to proliferate in response to sIg cross-linking in the absence of CD45 (Byth *et al.*, 1996). The defects in BCR signalling observed in primary CD45^{-/-} B cells are quite subtle. Whereas major changes in BCR-triggered protein tyrosine phosphorylation have not been observed in CD45^{-/-} spleen cells, the BCR is selectively uncoupled from activation of the MAP kinase pathway (see below for more details of this pathway) (Cyster *et al.*, 1996) and from calcium influx (Benatar *et al.*, 1996). Such defects are likely to explain the increased threshold for BCR signal transduction that has been noted in CD45^{-/-} B cells (Cyster *et al.*, 1996; Cyster, 1997).

Other tyrosine phosphatases are also involved in lymphocyte signal transduction. Some of these, like the SH2-domain-containing SHP-2 phosphatase, appear to exert a positive regulatory action on antigen receptor-mediated signals, whereas others, such as SHP-1, exert negative effects (Frearson & Alexander, 1997). The early events of antigen receptor signalling are, therefore, regulated by a fine balance between the actions of tyrosine kinases and phosphatases.

Antigen receptors and signal transduction complexes

As already noted, ligation of the TCR and BCR causes the recruitment of receptor-associated 'signal transduction complexes' that couple the receptors to signalling pathways. As the kinase activation cascade continues, cytosolic 'docking proteins' also become tyrosine phosphorylated, recruiting proteins in the process via SH2 domains and other types of protein–protein interaction to form signal transduction complexes that are not necessarily associated with receptors (Wange & Samelson 1996; DeFranco, 1997). One function of such docking proteins is to activate or inhibit associated enzymes or exchange factors, and to target them to their relevant substrates.

An interesting example of such a 'signal transduction complex' is provided by the interaction that occurs in T cells between molecules known as Grb-2, Vav, SLP-76 (SH2 domain-containing leukocyte protein of 76 kDa) and SLAP-130 (also called Fyb), as illustrated in Fig. 7.7. Grb-2 is an adaptor protein containing two SH3 domains and one SH2 domain. The role of Grb-2 is to bind to proteins bearing proline-rich motifs via its two SH3 domains and to bring these proteins into association with tyrosine-phosphorylated proteins via its SH2 domain. Vav is a docking protein that contains a single SH2 and two SH3 domains, as well as a PH domain, which together mediate its interactions with other proteins (Bustelo, Ledbetter & Barbacid, 1992). Furthermore, Vav contains an enzymatic domain which catalyses guanine nucleotide exchange on the Rho/Rac/CDC42 family of low-molecular-weight Ras-like GTP-binding pro-



Fig. 7.7. The Vav–SLP-76–Grb-2 signal transduction complex. Note that only certain domains of the molecules are shown to illustrate the way in which multimolecular complexes are assembled upon TCR engagement. The p36 molecule is membrane associated and upon tyrosine phosphorylation engages with the adaptor protein Grb-2, which in turn is associated with SLP-76 by an interaction that does not require tyrosine phosphorylation. Tyrosine phosphorylation of SLP-76 leads to its interaction with Vav; another tyrosine phosphorylated protein, SLAP-130 (Fyb), also binds to SLP-76. Such complexes provide a way in which several molecules may be brought to the plasma membrane in a functionally active form.

teins. Mature T and B cells lacking Vav proliferate poorly and, in the case of T cells, produce little IL-2 in response to TCR stimulation (Tarakhovsky *et al.*, 1995). Vav is rapidly phosphorylated by tyrosine kinases upon TCR engagement and then acts to promote Rac-1 to the active GTP-bound state (Crespo *et al.*, 1997). Another protein called SLP-76 is also tyrosine phosphorylated, probably by ZAP-70 (Wardenburg *et al.*, 1996) and binds to Vav via its SH2 domain, an interaction that appears to be critical for signals leading to IL-2 gene induction (Wu *et al.*, 1996; Raab *et al.*, 1997). SLP-76 also contains an SH2 domain and in turn binds via this domain to a further tyrosine-phosphorylated protein called SLAP-130 (also called Fyb), which may be a negative regulator of signal transduction (Musci *et al.*, 1997). There is evidence that this Grb-2/Vav/SLP-76/SLAP-130 complex is involved in coupling ZAP-70 activation to the Ras and calcium-signalling pathways (described further below). The rate of dissociation of such complexes is probably as important as the rate at which they

associate, since such rates presumably determine how long specific signalling pathways continue to be activated. It is, therefore, of interest that therapeutic reagents which suppress T cell activation may act by preventing the formation of such signal transduction complexes (Jabado *et al.*, 1997).

Signal transduction pathways induced by antigen receptors

Lymphocyte activation involves three major signalling pathways that lead from antigen receptors to the regulation of nuclear events and which are now beginning to be understood in some detail. These pathways involve the elevation of intracellular calcium, the activation of protein kinase C (PKC) and the activation of the Ras/MAP kinase pathway and are illustrated for T cells in Fig. 7.8. The same pathways are also activated in B cells, but results from T cells will be used to describe these pathways and the way in which they integrate to regulate the IL-2 gene by means of transcription factors such as AP-1, NF κ B and nuclear factor of activated T cells (NFAT).

The calcium-signalling pathway

TCR stimulation causes the tyrosine phosphorylation and activation of phospholipase-Cyl (PLCyl) (Secrist, Karnitz & Abraham, 1991) followed by its recruitment to the plasma membrane, possibly by means of an associated tyrosine-phosphorylated 36 kDa molecule (Motto et al., 1996). Activated PLCyl then hydrolyses its substrate phosphatidylinositol 4,5-bisphosphate, which is located in the membrane, to generate diacylglycerol (DAG) and inositol 1,4,5trisphosphate (IP₃). DAG activates PKC whereas IP₃ binds to IP₃ receptors, calcium-release channels on the endoplasmic reticulum that open to cause the release of calcium from intracellular stores (Berridge, 1993). TCR stimulation causes the tyrosine phosphorylation of IP₃ receptors by a process involving the p59^{fyn} tyrosine kinase (Jayaraman et al., 1996). The depletion of these stores in turn triggers the opening of calcium channels in the plasma membrane, known as store operated channels (SOCs), causing an influx of calcium ions from outside the cell (Zweifach & Lewis, 1993; Berridge, 1995). The opening of the SOCs depends upon the membrane potential, which is kept hyperpolarized by both voltage-gated and calcium-sensitive potassium channels (Lewis & Cahalan, 1995). If the potassium channels are blocked using inhibitors, then the membrane is depolarized, calcium influx no longer occurs and T cell proliferation is inhibited. For full activation and proliferation to occur, the prolonged elevation of intracellular calcium ions that results is essential (Goldsmith & Weiss, 1988; Wacholtz & Lipsky, 1993). For example, a T cell line lacking type 1 IP₃ receptors was unable to increase the



Fig. 7.8. The integration of TCR and CD28-mediated signal transduction pathways in T cells. CsA, cyclosporin A; I κ B, inhibitor- κ B; NFAT, nuclear factor of activated T cells; SRF, serum response factor; JNK, Jun kinase.

intracellular calcium concentration or produce IL-2 following TCR stimulation, demonstrating the critical role of this pathway in T cell activation (Jayaraman *et al.*, 1995). A patient with a severe immunodeficiency has been described with defective T cell SOCs (Ledeist *et al.*, 1995) and in T cell-lines expressing various SOC mutations a close correlation was found between the level of calcium influx and calcium-dependent gene transcription (Fanger *et al.*, 1995).
An important consequence of the increased intracellular calcium ion concentration is the activation of a calcium-dependent phosphoserine/threonine phosphatase called calcineurin. Calcineurin activation results in the rapid translocation (within minutes) to the nucleus of NFAT proteins, which are located in the cytosol in primary resting T cells (Luo et al., 1996). There they combine with AP-1 to form a functional transcriptional factor complex (Rao, Luo & Hogan, 1997). Translocation requires two nuclear localization motifs found in the NFAT molecule, one of which is in an intramolecular association with phosphoserines located at its amino terminus. Dephosphorylation by calcineurin exposes these motifs so that NFAT moves into the nucleus (Beals et al., 1997a). In the nucleus, NFAT is then rephosphorylated by the serine/threonine kinase glycogen synthase kinase-3, so promoting its return to the cytsolic compartment after several hours, providing that the calcium ion level has declined to resting levels (Loh et al., 1996; Beals et al., 1997b). Therefore, just as the earliest events of antigen receptor-signal transduction coupling are regulated by the balance between the actions of tyrosine kinases and phosphatases, so downstream signalling pathways are controlled by the balance between serine kinases and phosphatases. The antigen receptor-calcium-calcineurin-NFAT pathway also provides a good example of the way in which tyrosine kinases 'translate' the effects of receptor occupation into protein serine phosphorylation/dephosphorylation, so in turn regulating nuclear events.

The immunosuppressive drugs cyclosporin A (CsA) and FK506 (tacrolimus) exert their inhibitory effects by binding to immunophilins known as cyclophilin and the FK506-binding protein (FKBP), respectively (Liu *et al.*, 1991; Fruman *et al.*, 1992). The CsA–cyclophilin and FK506–FKBP complexes then bind in a calcium-dependent manner to calcineurin, so inhibiting its phosphatase activity. Thus, the addition of these drugs to lymphocytes blocks NFAT dephosphorylation and translocation to the nucleus, thereby preventing IL-2 gene induction by inhibiting the formation of functional transcription factor complexes (Rao *et al.*, 1997).

Other consequences of increased intracellular calcium ions are mediated by the binding of calcium to a binding protein called calmodulin, which leads to the activation of calcium-sensitive serine/threonine kinases such as calmodulin-activated kinase IV (CaM kinase IV) (Park & Soderling, 1995). CaM kinase IV has been implicated in regulation of the AP-1 gene (Ho, Gullberg & Chatila, 1996) and in cell-cycle regulation (Larsson *et al.*, 1995).

Protein kinase C activation

It has long been known that a full T cell activation programme can be induced by the addition of phorbol esters, which activate PKC, and calcium ionophores, which greatly increase the concentration of intracellular calcium, so bypassing the early events of TCR signal transduction coupling (Trunch et al., 1985). It is now realised, however, that the pharmacological activation of PKC has multiple effects, including the activation of the Ras/MAP kinase pathway (see below), whereas the TCR in situ does not couple to the Ras pathway using PKC (Cantrell, 1996). The fact that phorbol esters have effects on certain pathways does not mean that the same TCR-stimulated pathways are mediated by PKC in vivo. Furthermore, multiple PKC isoforms are present in lymphocytes, some of which are activated by the combination of calcium ions plus DAG, whereas the regulation of others is calcium independent. One characteristic of PKC enzymes is that they are translocated to membranes upon activation, so bringing them into contact with their membrane-localized substrates. Interestingly, although six different PKC isoforms were detected in antigen-specific T cell clones, only one of these, PKC θ , was found to translocate to the site of cell contact between antigen-presenting cells and T cells; the increased activity of PKC θ correlated well with T cell activation (Monks et al., 1997). Thus specific PKC isoforms may have highly selective effects during lymphocyte activation; in fact, active forms of PKC isoforms transfected into T cell lines demonstrate different effects on gene regulation (Genot, Parker & Cantrell, 1995). The selectivity of PKC isoform actions is also illustrated by the phenotype of mice lacking PKC β , in which T cell development and TCR-triggered proliferation was normal whereas B cell development was perturbed and the proliferation of B cells in response to BCR ligation was much reduced (Leitges et al., 1996). Furthermore, another PKC isoform, PKCµ, has been found to associate with the BCR and is activated upon BCR engagement (Sidorenko et al., 1996).

The function of PKC in T cell signal transduction pathways has been investigated by the use of inhibitors. A selective PKC inhibitor inhibits IL-2 secretion and T cell proliferation (Birchall *et al.*, 1994), but only the TCR stimulation of NF κ B induction is inhibited, not that of NFAT or AP-1 (Williams *et al.*, 1995). Since the binding of NF κ B to a site on the IL-2 gene promoter is necessary for IL-2 gene induction (Fig. 7.8), these results point to a role for PKC in regulating IL-2 secretion via NF κ B. Like NFAT, NF κ B is present in the cytosol in an inactive complex. NF κ B consists of a heterodimer of p50 (NF κ B-1) and p65 (RelA) polypeptides in complex with an inhibitory subunit (I κ B), which, when proteolytically degraded, unmasks a nuclear localization motif on RelA, so promoting the translocation of NF κ B to the nucleus (Baldwin, 1996). The important role of NF κ B for T cell activation is well illustrated by the actions of immunosuppressive glucocorticoids, which act by inhibiting NF κ B induction, possibly by inducing the synthesis of I κ B (Auphan *et al.*, 1995). Signals from the CD28 receptor also appear to be involved in NF κ B activation (Lai & Tan, 1994). The precise role of TCR/CD28 stimulation and PKC activation in the regulation of NF κ B awaits further elucidation.

The Ras/MAP kinase pathway

Ras is a low-molecular-weight (21 000) guanine nucleotide-binding protein localized in the plasma membrane that can exist in an active GTP-bound form or an inactive GDP-bound form. In resting T cells, Ras exists in its inactive GDPbound form but is rapidly converted to its active form upon TCR stimulation (Downward *et al.*, 1990). The equilibrium between these two forms involves the action of exchange factors such as Sos and C3G, which catalyse the conversion of Ras-GDP to Ras-GTP, and the activation of the GTPase activity of Ras by the Ras-GTPase activating protein (Ras-GAP), which hydrolyses Ras-bound GTP so converting Ras back to its inactive form. The exact way in which TCR stimulation causes Ras activation is not yet understood but appears to involve both inhibition of Ras-GAP (Downward et al., 1990) and the recruitment of Grb2-Sos and Crk-C3G complexes to the membrane, possibly by means of a membrane-associated p36 tyrosine-phosphorylated protein (Buday et al., 1994; Pastor, Reif & Cantrell, 1995; Sawasdikosol et al., 1995) (the same p36 protein as illustrated in Fig. 7.7). Like Grb-2, Crk is an adaptor protein and contains three SH3 domains and a single SH2 domain. Similar mechanisms couple the BCR to Ras activation, although in this context the Shc adaptor protein is thought to be involved in bringing Sos into association with its Ras-GDP substrate (DeFranco, 1997).

An important function of Ras is to couple the TCR to the mitogen-activated protein kinase (MAP kinase) pathway (also known as the extracellular signal regulated kinase pathway, ERK), which comprises a cascade of kinases leading from the plasma membrane to the nucleus (Fig. 7.8). The MAP kinase kinase kinase (MAPKKK) Raf-1 binds directly to Ras-GTP, which thereby recruits Raf-1 to the membrane where it is activated by a complex series of events (Morrison & Cutler, 1997). Activated Raf-1 in turn activates a MAP kinase kinase (MAPKK or MEK), which then phosphorylates and stimulates the MAP kinases Erk1 and Erk2, which translocate to the nucleus where they phosphorylate transcription factors (Robinson & Cobb, 1997). An important substrate for Erk2 is the transcription factor Elk1, which forms a ternary complex with the transcriptional activator serum response factor, so playing an important role in expression of fos (Hunter & Karin, 1992). Fos and Jun together form complexes (AP-1) that bind to sites in the IL-2 gene promoter. The multimolecular AP-1/NFAT complex ('NFAT') also binds to a further site in this promoter. Studies using both active and inactive mutants of Ras have shown that Ras does indeed synergize

with calcium to induce active NFAT complexes (Cantrell, 1996), although the MAP kinase cascade is probably not the only pathway whereby Ras regulates NFAT (Genot *et al.*, 1996). Furthermore, the CD28 signalling pathway synergizes with TCR-mediated signals in the activation of the Jun kinase (JNK) member of the MAP kinase family, which is activated by a cascade analogous to that which regulates Erk1/2 (Su *et al.*, 1994). JNK is thought to phosphorylate Jun and thereby promote the formation of active Fos–Jun (AP-1) complexes, illustrating the way in which the signals from the TCR and CD28 can integrate at the transcription factor level. In contrast to the TCR, physiological ligands for CD28 do not appear to activate Ras (Nunes *et al.*, 1994).

An important issue that is not yet fully resolved is the extent to which Erk1/2 activation is important in mature peripheral T cells. It should be noted that many of the data available on Ras function in T cells have been generated using cell lines as well as transgenic mice. The expression of dominant-negative mutant forms of Ras and of MEK-1 in transgenic mice has unambiguously demonstrated that the Ras/MAP kinase pathway is essential for the positive selection but not the negative selection of CD4+CD8+ thymocytes (Alberolaila *et al.*, 1995; 1996; Swan *et al.*, 1995). However, whereas the dominant-negative Ras completely inhibits proliferative responses to TCR stimulation (Swan *et al.*, 1995), the over-expression of dominant-negative MEK-1 does not perturb TCR-stimulated proliferation of, or IL-2 secretion from, mature splenic T cells, despite the inability of the TCR to trigger Erk1/2 activation under such conditions (Alberolaila *et al.*, 1995). Therefore, the possibility remains that Ras activation is essential for T cell proliferation but the critical Ras-mediated pathway that mediates T cell proliferation does not involve Erk1/2 activation in mature T cells.

The role of co-receptors in antigen receptor signalling

The term 'co-receptor' is sometimes restricted in use to molecules such as CD4/CD8, which are intimately involved in TCR functions, but in other contexts the word is used more widely to refer to any receptor that either upmodulates or downmodulates antigen receptor signal transduction. The term here is used in this second more general sense. The important role of CD4 and CD8 in TCR signalling has already been summarized, and the signalling functions of CD28 and CTLA-4 expressed on T cells are described in Chapter 6. On B cells, there are two important co-receptors that either upregulate (CD19) or downregulate (CD22) BCR-mediated signals, thereby playing important roles in establishing signalling thresholds (Fig. 7.9) (O'Rourke *et al.*, 1997).

CD19 is a member of the immunoglobulin superfamily that is expressed on B cells in association with CD21 and CD81 (Tedder, Inaoki & Sato, 1997a). CD21



Enhancement of BCR-mediated signals. Co-ligation of CD19/CD21/CD81 complex to mlg by C3d-antigen recruits positive signal transduction effectors that augment B cell activation. Ligation of CD22 by glycoconjugates in lymphoid organs releases Fig. 7.9. The role of CD19 and CD22 in regulating BCR signal transduction. (a) Suppression of BCR-mediated signals. Crosslinking of mlg induces tyrosine phosphorylation of CD22, recruitment of SHP-1 and downregulation of signal transduction. (b) mlg and prevents the negative effects of CD22. (From O'Rourke et al., 1997.) is the complement receptor type 2 and binds the cleavage products of the third complement component (C3). If an antigen has covalently bound C3 molecules, it will bind to the BCR and simultaneously engage the CD19/CD21 complex, so promoting an association between CD19 and the BCR. For example, hen egg lysozyme becomes 10 000-fold more immunogenic when it is attached to the C3d complement protein (Dempsey et al., 1996). This amplification system is mediated by synergistic interactions occurring between CD19 and the BCR. CD19 is a signalling molecule with a long cytoplasmic tail, containing nine conserved tyrosine residues that become phosphorylated upon engagement of CD19 with activating mAbs or ligation of the BCR. If CD19 is cross-linked with the BCR, then the signals induced are greater than those triggered by either receptor alone (Carter & Fearon, 1992), and BCR-associated tyrosine kinases such as Syk may be responsible for CD19 phosphorylation (Carter et al., 1997). Upon phosphorylation, a number of SH2 domain-containing proteins are recruited to the CD19 cytoplasmic tail, including p59^{fyn}, p56^{1ck}, p59^{1yn}, Vav and the p85 regulatory subunit of phosphoinositide (PI) 3'-kinase, which generates the important second messenger phosphoinositide trisphosphate (Tedder et al., 1997a). p59fyn binds CD19 at a different site from PI 3'-kinase and so both molecules are probably bound simultaneously. This may allow the involvement of the p59fyn SH3 domain in activating PI 3'-kinase. A CD19-recruited complex involving Vav and PI 3'kinase is, therefore, likely to be important in triggering intracellular pathways, although the downstream consequences are not yet well understood.

The nature of CD19 as an amplification system for BCR signal transduction has been confirmed by the study of CD19-deficient mice in which there is a severe deficit in mature B cells (B-1), the cells which normally have the highest levels of cell-surface CD19 expression (Rickert, Rajewsky & Roes, 1995). In these mice, B cells no longer proliferate upon BCR engagement, whereas in mice overexpressing CD19, B cells demonstrate increased proliferative responses, underlining the key role that CD19 plays in altering BCR signalling thresholds.

In contrast to CD19, CD22 is thought to exert a negative action on BCR signal transduction (O'Rourke *et al.*, 1997; Tedder *et al.*, 1997b). CD22 becomes associated with the BCR upon BCR cross-linking and is phosphorylated on tyrosine residues within its cytoplasmic tail (Schulte *et al.*, 1992). Three of the six tyrosine residues located in the tail are found within immunoreceptor tyrosinebased inhibitory motifs (ITIMs), which have the sequence V/I-X-Y-X-X-L. These motifs have also been found in the Fc γ Rllb receptor, which likewise has a negative regulatory role in BCR signalling (Muta *et al.*, 1994). Upon phosphorylation, an SH2 domain-containing tyrosine phosphatase called SHP-1 is recruited to an ITIM within the CD22 tail, where it is activated (Doody *et al.*, 1995). The negative effects of SHP-1 in lymphocyte signalling are well established (Frearson & Alexander, 1997). The phosphatase may exert its effects by dephosphorylating one or more tyrosine-phosphorylated proteins required for coupling the BCR to the calcium-signalling pathway, although the uncoupling effects of other molecules recruited to CD22 cannot yet be excluded. This model is supported by results obtained from CD22-deficient mice in which B cell development is normal but splenic B cells were found to be hyperresponsive to BCR signalling, with increased and prolonged calcium signals and proliferative responses (O'Keefe et al., 1996; Nitschke et al., 1997). Interestingly, B cells from 'motheaten mice', which have defects in SHP-1, display a similar phenotype to CD22-/- mice and likewise have elevated calcium responses to B cell antigens (Cyster & Goodnow, 1995). In neither CD22-/- nor in motheaten mice, however, are there global defects in the coupling of the BCR to intracellular protein tyrosine-phosphorylation events (Nitschke et al., 1997). It seems likely, therefore, that the negative actions of the CD22 co-receptor result from the highly selective effects of SHP-1 and perhaps other CD22-recruited molecules on the BCR signalling apparatus.

The balance between the positive and negative effects of CD28 and CTLA-4 on T cell function, and the parallel types of effect of CD19 and CD22 on B cell function, illustrate the complex way in which the responses of antigen receptor-mediated signals are 'fine tuned' to respond appropriately to a very broad spectrum of antigens encountered within a wide range of cellular contexts.

Pharmaceutical intervention in lymphocyte signalling pathways

The present available range of immunosuppressive drugs are of great value in the clinic but suffer from various harmful side effects. Cyclosporin A and FK506, for example, have the disadvantage that their target (calcineurin) is widely expressed in non-haematopoietic tissues, thereby facilitating deleterious effects of these drugs in organs such as the kidney and heart. An enormous amount of research is, therefore, being carried out by pharmaceutical companies to target molecules in lymphocyte activation pathways that would provide a more selective means of immunosuppression. Several of the key enzymes already identified, such as ZAP-70, p56lck, p56lck and CD45, are largely restricted in their expression to haematopoietic cells, making such molecules attractive targets for pharmaceutical intervention. Monoclonal antibodies against cell-surface antigens such as CD4 have also been used with some success to suppress the inappropriate activation of T cells in autoimmune diseases such as RA. By humanizing such antibodies, that is by engineering the antibody to mimic human immunoglobulin as much as possible, the risk of the recipient undergoing an immune response to the therapeutic reagent is lessened.

The ability of receptors such as CD28 and CTLA-4 to induce signals that modulate the proliferative and survival responses of T cells has also aroused great interest in the pharmaceutical industry, since hopes have been raised that in the future it will prove possible using appropriate drugs to raise and lower T cell activation levels at will. The temporary upregulation of T cell responses could be invaluable in promoting, for example, the cytotoxic actions of tumourinfiltrating lymphocytes, or in boosting the immune system to attack residual malignant cells following surgical removal of cancerous tissue. The temporary suppression of T cell responses is clearly important in the context of organ transplantation, autoimmunity and inflammation.

Such pharmaceutical hopes lie largely in the future. Nevertheless, it is very likely that the basic research carried out on lymphocyte signal transduction pathways in the late twentieth century will pave the way for a rational approach to drug-induced manipulation of the immune system that will reach its fruition during the course of the twenty-first century.

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8

The role of adhesion mechanisms in inflammation

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Introduction

Adhesion mechanisms play extraordinarily diverse roles in many biological phenomena. These range from physiological events in early life, when they are crucial for the development of the embryo, to pathological processes, including tumour growth and metastasis. In some cases, adhesion molecules act as receptors for infective agents such as viruses and parasites. From the immunological point of view, they are involved in virtually every process involving cell contact from thymic selection to antigen processing, from antigen priming to cell activation, from cytotoxicity to lymphocyte recirculation. The last process is part of a sophisticated system that allows efficient surveillance of the various tissues in the body for the presence of infectious pathogens and 'dangerous' exogenous or endogenous antigens. However, when the immune response goes astray, these same mechanisms are often responsible for the perpetuation of inflammation. A classical example is provided by RA and the other chronic inflammatory arthropathies, where the persistent synovial inflammation is maintained, among other factors, by the continuous migration of inflammatory cells from the bloodstream into the joint. Adhesion mechanisms further contribute to the perpetuation of the inflammatory response by the retention of these cells within the tissues, through interactions with extracellular matrix components, and by facilitating contact-dependent immunological events.

In this review, I will concentrate firstly on the general mechanisms that regulate the process of leukocyte extravasation from the blood into the tissues. Secondly, I will discuss the additional mechanisms that finely tune this process to allow the selective migration of distinct leukocyte populations in specific conditions and into different organs. Thirdly, I will consider some of the mechanisms involved in cell migration, spatial orientation and retention within the tissues. Finally, I will focus on the current therapeutic developments that target adhesion mechanisms.





Regulation of leukocyte migration: general mechanisms

The initial stage in the process of cell extravasation into the tissues is the interaction of circulating leukocytes with vascular endothelium. As initially shown by Gowans and co-workers (Gowans & Knight, 1964; Marchesi & Gowans, 1964), and subsequently confirmed by others (Anderson & Anderson, 1976; Smith & Ford, 1983), cell migration into lymphoid tissues occurs at specialized postcapillary vascular sites called high endothelial venules (HEV) because of the particular cuboidal morphology of the endothelial cells. In non-lymphoid tissues, migration takes place also in postcapillary venules, where blood flow velocity is reduced, but, unlike lymphoid HEV, the vessels are lined by flat endothelium. Interestingly, in cronically inflamed tissues such as the rheumatoid synovium, HEV-like vessels are also formed (Freemont et al., 1983; Freemont, 1988; Yanni et al., 1993). Here, HEV are localized in areas where the infiltrating lymphocytes are organized in lymphoid follicle-like structures, in contrast to areas with a diffuse lymphocytic infiltrate, where the endothelium remains flat (Yanni et al., 1993; van Dinther-Janssen et al., 1990). This suggests that HEV formation can be induced outside the lymphoid tissues in response to local factors such as inflammatory cytokines and in association with an increase in lymphocyte traffic. Similar factors also appear to be important for the maintenance of HEV in lymphoid tissues, as lymph node HEV convert from a high to a flat endothelial morphology following ligation of the afferent lymphatics (Mebius et al., 1991).

Our understanding of the actual process of adhesion to and migration through the endothelium has grown considerably since the mid-1980s mainly following studies investigating leukocyte adhesion under conditions of flow both in vitro and in vivo (Lawrence & Springer, 1991; Ley & Gaehtgens, 1991; Ley et al., 1991; von Andrian et al., 1991). It has become apparent that the way in which leukocytes overcome the shear forces associated with blood flow is through a series of co-ordinated events mediated by specific adhesion molecules. For example, it was observed that some of the leukocytes come into brief contact with the endothelial ligands, slow their movement and start rolling gently. Some cells then disengage and are carried on by the flow, whereas others come to a complete halt. These latter cells change shape, acquiring a flattened morphology and, within a few minutes, actively migrate between the endothelial cells. The cascade of molecular events that regulate these processes has been clarified by a number of studies. These have given rise to a consensus model of four sequential steps: (i) tethering/rolling, (ii) triggering/activation, (iii) strong adhesion, and (iv) transendothelial migration (Butcher, 1991; Adams & Shaw, 1994; Springer, 1994). These four phases, shown schematically in Fig. 8.1, will now be described in more detail.

Tethering/rolling

The first step, tethering/rolling, is transient, activation independent and mediated by inducible or constitutively expressed selectin molecules and their cognate oligosaccharide ligands (Table 8.1). The selectin family (CD62) consists of three similar single-chain membrane glycoproteins (L-selectin, E-selectin and Pselectin) each of which comprises a Ca²⁺-dependent amino terminal lectin domain, a proximal epidermal growth factor-like motif and a series of complement regulatory protein-like units of approximately 60 amino acid residues each (Bevilacqua & Nelson, 1993; Lasky, 1995a). L-selectin is constitutively expressed by the majority of neutrophils and monocytes and by approximately half of the lymphocytes. E-selectin is expressed only by endothelial cells, mainly following activation by, for example, endotoxin, IL-1 or TNF α . Its expression requires de novo mRNA and protein synthesis and it is maximally expressed at 4 to 6 hours, with a decline to baseline level by 24 hours. P-selectin is found in synthesized form in the α granules of platelets and the Weibel–Palade bodies of the endothelium. P-selectin is rapidly mobilized, both in platelets and endothelial cells, to the cell surface after stimulation by several mediators including thrombin, histamine and terminal complement components (Bevilacqua & Nelson, 1993; Lasky, 1995a).

Selectins are long molecules, which makes them ideally designed to protrude from the cell surface and to interact with appropriate receptors in conditions of flow (Lasky, 1992). All selectins appear to bind to sialylated and fucosylated carbohydrate determinants on their counter-receptors. E-selectin and P-selectin recognize carbohydrate structures closely related to the tetrasaccharide sialyl Lewis^x and its isomer sialyl Lewis^a (Lasky, 1995a). In addition, P-selectin binds to a heavily glycosylated 120 kDa glycoprotein named P-selectin glycoprotein ligand 1 (PSGL-1). This is a member of the sialomucin family of adhesion proteins, which contain numerous O-linked carbohydrate side-chains that are attached to serine- and threonine-rich domains (Lasky, 1995b). Two other important members of this family function as ligands for L-selectin: glycosylation-dependent cell adhesion molecule-1 (GlyCAM-1) and CD34. GlyCAM-1 is highly expressed by peripheral lymph node HEV and, for this reason, it is believed to act as an addressin molecule for lymphocyte homing to peripheral lymph nodes (see below). Protein sequence analysis revealed that GlyCAM-1 does not contain a transmembrane anchoring motif, which suggests that either it binds indirectly to the luminal surface of HEV or that it functions as a soluble molecule. In contrast, CD34 contains a classical transmembrane domain that stabilizes the molecule within the endothelial membrane. Another important difference is that CD34 is globally expressed at various endothelial sites. However, the glycosy-

Endothelial distribution	Endothelial receptor/ligand	Leukocyte receptor/ligand	Leukocyte distribution
HEV, activated endothelium	GlyCAM-1, CD34	L-selectin/CD62L	Lymphocyte subset, monocytes, neutro- phils, eosinophils
Endothelial Weibel–Palade granules, platelet α granules	P-selectin/ CD62PP	P-selectin glycoprotein ligand 1 (PSGL-1), sialyl Lewis ^{x/a}	Neutrophils, mono- cytes, lymphocyte subset, natural killer cells
Activated endothelium	E-selectin/CD62E	Sialyl Lewis ^{x/a}	Neutrophils, mono- cytes, eosinophils, basophils, lympho- cyte subset, natural killer cells

Table 8.1. Selectins and their ligands in leukocyte-endothelial interactions

lation of CD34 differs according to the tissue in which it is expressed. Notably, the CD34 expressed by the HEV of peripheral lymph nodes contains the sulphated carbohydrate ligand for L-selectin while the CD34 expressed at other sites does not contain this unique oligosaccharide (Baumhueter et al., 1994). Therefore, it is likely that the binding specificity of sialomucin is dependent on the types of carbohydrate modification that occur during its transit through the secretory pathway to the cell surface. The rigid rod-like structure of these molecules, however, is likely to produce the extended matrix upon which the various oligosaccharide ligands are presented above the cellular glycocalyx. Here they will be recognized by selectin molecules expressed on specific microvillar cell surface projections of leukocytes flowing nearby. This interaction, although not very strong, is sufficient to slow down the leukocytes, make them roll on the endothelium and facilitate their sampling the local microenvironment for the presence of migratory signals such as inflammatory products. If these are present, the adhesion cascade will be triggered to proceed towards the subsequent steps. If not, the transient nature of selectin binding allows leukocytes to disengage and to be carried on by the flow.

The functional role of selectin molecules *in vivo* is further illustrated by selective gene deletion in animals. Lymphocytes from L-selectin-deficient mice do not bind to peripheral lymph node HEV and show a significant defect in their rolling and migration capacity (Arbones *et al.*, 1994; Tedder, Steeber & Pizcueta, 1995). In E- and P-selectin double knock-out animals, leukocytes are virtually unable to extravasate from the circulation (Frenette *et al.*, 1996). As a consequence inflammatory/immune responses, including delayed-type hypersensitivity, are grossly impaired (Staite *et al.*, 1996). A human disease called leukocyte adhesion deficiency 2 (LAD-2) illustrates the importance of the carbohydrate ligands to which selectin molecules bind (Etzioni *et al.*, 1993). In this condition, an unknown metabolic defect results in a complete lack of fucose production, which, in turn, leads to the absence of surface-expressed fucosylated selectin ligands. Neutrophils from these patients are unable to bind to immobilized E-selectin *in vitro*, and, not surprisingly, these patients cannot mount an appropriate inflammatory response to external pathogens.

Triggering/activation

The second step, triggering/activation is necessary for the activation of integrin molecules. This leads to an increase in their avidity, which renders them capable of mediating the third phase: strong leukocyte-endothelial adhesion. Several chemotactic factors have been described as having triggering/activating properties, including bacterial wall components and complement products (Table 8.2). The most important molecules, however, appear to be a series of chemoattractant cytokines (chemokines) of which more than 40 have been described (Schall & Bacon, 1994; Baggiolini et al., 1997). Chemokines (CKs) are small-molecularweight molecules (68 to 120 amino acid residues in size) that share structural similarities, including four conserved cysteine residues, which form disulphide bonds in the tertiary structure of the proteins. The majority can be incorporated into two large subfamilies: C-X-C (where X is any amino acid) and C-C chemokines, according to whether an intervening residue spaces the first two cysteines in the motif or they remain adjacent. A third subfamily, which so far has only one member, lymphotactin, has been described (Kennedy et al. 1995). Lymphotactin has a high degree of homology with the other two subfamilies but lacks the first and third cysteine. Finally, a fourth molecule (fractalkine), characterized by an interposition of three amino acids between the first two cysteines (C-X₃-C), has recently been added to the list (Bazan et al., 1997). Interestingly, unlike other CK types, the polypetide chain of fractalkine is predicted to be part of a target 373 amino acid residue protein with a CK domain and an extended mucin-like stalk. This would allow one end of the molecule to anchor to the surface of the endothelium while the other end, carrying on its top the CK motif, would protrude deeply in the vascular lumen. Similarly, it has been suggested that other CKs can be immobilized in a solid phase on the endothelial surface

Chemoattractant	Origin	Responding cells
Classical chemoattractants <i>N</i> -formyl peptides	Bacterial protein processing	Monocytes, neutrophils, eosinophils, basophils
C5a	Complement activation	Monocytes, neutrophils, eosinophils, basophils
Leukotriene B4 Platelet-activating factor	Arachidonate metabolism Phosphatidylcholine metabolism	Monocytes, neutrophils Monocytes, neutrophils, eosinophils
C-X-C chemokines SDF-1, IL-8; GROα, GROβ, GROγ; NAP-2; ENA-78; GCP2; PF4, Ip10; Mig; CKα1; CKα2	T lymphocytes, monocytes, endothelial cells, fibroblasts, keratinocytes, chondrocytes, mesothelial cells	Neutrophils, basophils, fibroblasts
C-C chemokines MCP -1, -2, -3, -4; eotaxin; MIP-1α, MIP-1β; I–309; CK β4, β6, β7, β8, β9, β11, β12, β13; RANTES; HCC-1, -2, -3; TARC	T lymphocytes, monocytes, fibroblasts, endothelial cells, smooth muscle cells, platelets, mast cells	Monocytes, T lymphocyte subpopulation, eosinophils, basophils

Table 8.2 Leukocyte chemoattractants

via electrostatic interactions with negatively charged proteoglycans and CD44 molecules expressed by endothelial cells (Tanaka *et al.*, 1993a; Tanaka, Adams & Shaw, 1993b). This hypothesis provides a valid model of how, *in vivo*, a high concentration of chemotactic factors is maintained at the luminal surface of blood vessels adjacent to areas of inflammation, preventing them from being washed away by the flow.

CK are thought to mediate their effects via interactions with seven membranespanning domain receptors (CKR) that form a distinct group of structurally related proteins within the superfamily of receptors that signal through heterotrimeric GTP-binding proteins (Kelvin *et al.*, 1993; Neote *et al.*, 1993; Premack & Schall, 1996). CKR also have two conserved cysteines (one in the amino terminal domain and the other in the third extracellular loop), which are assumed to form a disulphide bond critical for the conformation of the ligand-binding pocket. Engagement of CKR results in activation of phospholipases,

Receptors	New nomenclature	Old nomenclature	Ligands
C-X-C family	CXCR1	IL-8R1 (type A)	IL-8
	CXCR2	IL-8R2 (type B)	IL-8, GROα,
			GROβ, GROγ,
			NAP2, ENA78,
			GCP-2
	CXCR3	IP10/MigR	IP10, Mig
	CXCR4	LESTR, HUMSTR	SDF-1
C-C family	CCR1	RANTES, MIP-1\alpha R	RANTES, MIP-1α,
			MCP-2, MCP-3
	CCR2a/b	MCP-1RA/B	MCP-1, MCP-2,
			MCP-3, MCP-4
	CCR3	EotaxinR, C-C	Eotaxin, RANTES,
		CKR3	MCP-3, MCP-4
	CCR4	C-C CKR4	RANTES, MIP-1α,
			MCP-1
	CCR5	C-C CKR5	RANTES, MIP-1α,
			MIP-1β

Table 8.3 Ligand selectivity of chemokine (CK) receptors

with downstream generation of inositol triphosphate and intracellular mobilization of Ca²⁺ and of small GTP-binding proteins such as the Ras, Rac and Rho families. The Rho family, in particular, seems to be instrumental in relaying the signal from the CKR to cell-surface integrins (Laudanna, Campbell & Butcher, 1996). For each CK class several CKR have been identified (Table 8.3) (Premack & Schall, 1996; Baggiolini *et al.*, 1997). The majority of CKR interact with more than one CK (shared specificity within class), except CXCR1 and CXCR4, which are specific for IL-8 and SDF-1, respectively.

Besides the functions described, the interaction of CKs with their receptors appears to serve another very important role, which is the selective chemoattraction of different leukocyte populations. It is well established that the C-X-C family act mainly on neutrophils while C-C members attract principally lymphocytes, monocytes, eosinophils and natural killer cells (Schall & Bacon, 1994; Schall *et al.* 1993b). Lymphotactin is uniquely specific for lymphocytes (Kennedy *et al.*, 1995) while, within the C-C family, eotaxin specifically attracts eosinophils (Kitaura *et al.*, 1996). CK selectivity is partially owing to a quantitative and/or qualitative variation of CKR expression by the responsive cells. For example, eotaxin specificity is strengthened by the high expression of the eotaxin receptor (CCR3) by eosinophils (Ponath *et al.*, 1996). Therefore, the combination of varied CK production and differential receptor expression may play an important role in regulating the diversity of leukocyte migration (see below). The report that, at least *in vitro*, some C-C CK (RANTES, MIP-1 α , MIP-1 β , MCP-1) favour chemoattraction, and transendothelial migration of specific lymphocyte subsets lends support to this hypothesis (Schall *et al.*, 1993a; Roth, Carr & Springer, 1995). Another factor to bear in mind is that, in response to inflammatory stimuli, several chemoattractant molecules are produced by the endothelial cells themselves (Table 8.2), which, therefore, actively contribute to recruiting leukocytes at sites of inflammation (Huber *et al.*, 1991; Zimmerman, Prescott & McIntyre, 1992).

Strong adhesion

The third stage is strong adhesion of the leukocytes. During this step leukocytes stop rolling and firmly adhere to the vascular endothelium before transmigrating between the endothelial cells. The third phase is primarily mediated by activated β_2 and α_4 integrins (Hogg & Landis, 1993; Hogg *et al.*, 1993; Springer, 1994) and their counter-receptors belonging to the immunoglobulin superfamily: ICAM-1, ICAM-2, VCAM-1 and MADCAM-1 (Table 8.4). Integrins are a large family of heterodimeric glycoproteins formed by two non-covalently linked subunits, a larger α chain (120–180 kDa) and smaller β chain (90–110 kDa). Both subunits are integral membrane proteins with a small carboxy terminal cytoplasmic domain and a large amino-terminal extracellular domain. At present there are at least 20 α chains and eight β chains, which can be found in many but not all of the possible combinations (Larson & Springer, 1990; Ruoslahti, 1991; Hynes, 1992). The heterodimeric structure of β_2 integrins comprises an α chain linked to a common β chain (Table 8.4). Three members belong to this subfamily: LFA-1 ($\alpha_{I}\beta_{2}$; CD11a/CD18), Mac-1 ($\alpha_{M}\beta_{2}$; CD11b/CD18), and p150,95 $(\alpha_{v}\beta_{2}; CD11c/CD18)$. LFA-1 is present constitutively on the surface of virtually all circulating leukocytes while Mac-1 and p150,95 have a more limited distribution on neutrophils, monocytes and natural killer cells. Apart from their key role in mediating adhesion to endothelium, β_2 integrins are also involved in most immune functions requiring homo- and heterotypic cell contact. In comparison to the β_2 integrins, α_4 integrins have a common α chain associated with a variable β chain (Table 8.4). The $\alpha_4\beta_1$ molecule belongs to the β_1 integrin subfamily. This group is also known as VLA molecules and its main function is to act as receptors for extracellular matrix components (see below). However, in contrast to other members of this family, $\alpha_{a}\beta_{1}$ integrins can also mediate binding to the endothelial ligand VCAM-1 (see below). Therefore, $\alpha_4\beta_1$ integrin can, through different binding sites, mediate leukocyte interactions with both the endothelium

Subunit	Alternative name	Distribution	Ligand
β2-integrins			
$\alpha_L^{}\beta_2^{}$	LFA-1, CD11a/CD18	B and T lymphocytes, monocytes, neutrophils	ICAM-1, ICAM-2, ICAM-3
$\alpha_m \beta_2$	Mac-1, CR3, CD11b/CD18	Monocytes, neutrophils, natural killer cells	ICAM-1, iC3b, fibrinogen, factor X
$\alpha_X\beta_2$	p150,95, CD11c/CD18	Monocytes, neutrophils, natural killer cells	iC3b, fibrinogen
α4-Integrins			
$\alpha_4\beta_1$	VLA-4, CD49d/CD29	B and T lymphocytes, monocytes, neural crest-derived cells, fibroblasts, muscle cells	VCAM-1, fibronectin
$\alpha_4\beta_7$	LPAM-1, CD49d/CD	B and T lymphocyte subpopulations	MadCAM-1, VCAM-1, fibronectin

Table 8.4. Integrins and their ligands in leukocyte-endothelial interactions

cells and the extracellular matrix (Pulido *et al.*, 1991; Masumoto & Hemler, 1993; Humphries *et al.*, 1995). This can facilitate not only the process of cell extravasation but also cell orientation and migration within the tissues. The α_4 integrin chain can also associate with the β_7 chain to form $\alpha_4\beta_7$ which, besides mediating adhesion to VCAM-1 and fibronectin, is involved in lymphocyte homing to mucosal associated lymphoid tissue and, for this reason, will be discussed below (Andrew *et al.*, 1994).

The immunoglobulin supergene family is a large group of cell membrane glycoproteins that are typified by a common structure similar in its domain organization to immunoglobulins. Their structure consists of a single chain that contains a series of C_2 -like heavy chain immunoglobulin domains of approximately 100–110 amino acid residues. ICAM-1 (CD54) is a glycosylated cell surface protein, of about 90 kDa, containing five tandem extracellular immunoglobulin-like domains (Simmons, 1995; Etzioni, 1996). In contrast to LFA-1, which is restricted to leukocytes, ICAM-1 is expressed on a variety of other cell types including thymic epithelial cells, fibroblasts, epidermal keratinocytes and endothelial cells. On resting endothelium, ICAM-1 is present only at a low level, but its expression is rapidly upregulated following stimulation by several inflammatory mediators such as lipopolysaccharide, IFN- γ , IL-1 and TNF α . Enhanced surface expression of ICAM-1 is first seen at 4 hours, is usually maximal at 24 hours and can persist for up to 48 hours or longer (Dustin, Staunton & Springer, 1988). In contrast to ICAM-1, ICAM-2 has only two immunoglobulin-like domains, which show a 35% homology with the two amino terminal domains of ICAM-1 (Staunton, Dustin & Springer 1989). Furthermore, ICAM-2 is highly expressed on resting endothelium and cannot be upregulated by inflammatory mediators. Therefore, it would appear that ICAM-1 has a more prominent role in activation-dependent adhesion and migration, while ICAM-2 is likely to be more important in regulating basal physiological cell migration and cyto-adhesiveness. A third ICAM (ICAM-3) is also a ligand for LFA-1 but is not expressed by endothelial cells and will be not discussed further.

VCAM-1 has a molecular weight of approximately 110 000 and consists of six or seven extracellular immunoglobulin-like domains generated by alternative splicing of the same gene located on chromosome 1 (Cybulsky et al., 1991a,b). Both forms are fully functional and, as is the case for ICAM-1 (Staunton et al., 1990), only the first two domains are required for its adhesion function (Osborn et al., 1994). VCAM-1 is not expressed on resting endothelial cells but is upregulated with the same kinetics as ICAM-1 after cytokine or endotoxin stimulation of the endothelium. This suggests that VCAM-1 is important in regulating cell migration in response to inflammation. Support for this hypothesis came from several studies, which confirmed the notable contribution of VLA-4dependent pathways in lymphocyte adhesion and migration in inflammatory conditions (Elices et al., 1993; Yang et al., 1993; Rabb et al., 1994). However, since the expression of VCAM-1 in the synovium has been shown to be confined mainly to the synovial lining and is expressed only weakly by the endothelium (Morales-Ducret et al., 1992; Wilkinson et al., 1993), some doubts have been expressed as to the real importance of the VLA-4/VCAM-1 interaction in leukocyte extravasation to the joint. The report of the presence of the CS-1 peptide of fibronectin on the luminal surface of synovial endothelial cells has prompted the attractive proposal that, in such an environment, the preferred ligand for VLA-4 could be the CS-1 fibronectin peptide rather than VCAM-1 (van Dinther-Janssen et al., 1993; Elices et al., 1994). The other important member of the immunoglobulin superfamily is the mucosal cell adhesion molecule 1 (MAdCAM-1). MAdCAM-1 is a hybrid molecule composed of three immunoglobulin-like domains and a mucin-like region interposed between domains 2 and 3 (Briskin et al., 1993; Sampaio et al., 1995). Given its predominant expression on the intestinal endothelium and in mucosal-associated lymphoid tissue (MALT), hence its name, MAdCAM-1 is instrumental in lymphocyte homing to mucosal organs and will be discussed in greater detail below.

As mentioned above, integrin activation is an essential component in the increased molecular adhesiveness that allows leukocytes to bind strongly to

endothelial ligands and overcome the hydrodynamic shear forces in the circulation. Integrin-binding avidity has been shown to be rapidly modulated (within seconds) by divalent cations, such as Mn²⁺, N-formylated peptide and chemokines (Dransfield et al., 1992; Hogg & Landis, 1993). The mechanism by which this occurs is not completely clear, but surface redistribution leading to integrin clustering and/or conformational changes in the tertiary molecular structure are thought to be the most important (Kornberg et al., 1992; Hogg & Landis, 1993; Hogg et al., 1993). Conformational changes in LFA-1 and Mac-1 are suggested by studies using monoclonal antibodies (mAbs) that react with these integrins after cellular activation but not in resting conditions (Diamond & Springer, 1993; Landis, Bennett & Hogg, 1993). For example, saturation binding studies have shown that, following neutrophil activation with chemoattractants, 10% of surface Mac-1 molecules express an activation epitope. Furthermore, blockade of this epitope inhibits neutrophil binding to purified ICAM-1 (Diamond & Springer, 1993). Finally, measurements of the affinity of cell surface LFA-1 for soluble monomeric ICAM-1 have directly demonstrated an increase of approximately 200-fold in a subpopulation of LFA-1 molecules after cellular activation (Lollo et al., 1993). The rapid ability to modulate the adhesion/de-adhesion integrin status is a crucial mechanism to prevent random adhesion in the bloodstream, to respond swiftly to microenvironmental signals and to facilitate the process of diapedesis (see below).

As for the selectins, much information on the importance of β_2 and α_4 integrins is gained from selective gene deletion experiments in animals (Sharpe, 1995; Arroyo *et al.*, 1996; Shier *et al.*, 1996) and from the human syndrome of leukocyte adhesion deficiency 1 (LAD-1) (Anderson & Springer, 1987; Fischer *et al.*, 1988). This condition is caused by a mutation in the gene of the common β_2 subunit, which leads to deficient expression of mature integrins on the cell surface. Neutrophils from these patients fail to migrate in response to chemoattractants and are unable to bind to and cross the endothelium at sites of infection. Patients suffer from recurrent bacterial infections that are often fatal in childhood. Of course, the severity of the syndrome is also influenced by the fact that integrin molecules function as accessory molecules in leukocyte interactions with other cells such as antigen-presenting cells, stromal cells or infected cells and are, therefore, important for many other immune functions including antigen, presentation/response and cytotoxicity (Harlan, 1993).

Transendothelial migration

Of the four steps involved in leukocyte extravasation, transendothelial migration is the least characterized. To complete the process of transendothelial migration, or diapedesis, leukocytes that are strongly attached to the endothelium have to crawl between the endothelial cells and cross the basal membrane. Initiation of cell locomotion involves a directional protrusion of the leading edge, presumably via actin polymerization, to form a lamellipodium, which attaches to the substratum. The adhesive strength at the cell front must be sufficient to generate force and traction to pull the cell forward. In contrast, the adhesive strength at the cell rear must be weaker to allow the cell to detach and retract forward. Although our understanding of the molecular mechanisms governing these processes is limited, activated integrins seem to play an important role (Hogg & Landis, 1993; Springer, 1994). Integrins connect the external substratum with the cytoskeleton. The integrin cytoplasmic domain, at least in vitro, has been shown to associate with structural proteins present in focal adhesion such as talin and α-actin (Pavalko et al., 1991; Otev, Pavalko & Burridge, 1990). The integrin-receptor complex serves not only as a structural link but also as a signalling unit, since, in addition to the above structural proteins, regulatory proteins such as focal adhesion kinase (FAK) are found in association with it (Kornberg et al., 1992; Parsons et al., 1994). Furthermore, phospholipase Cy (PLC γ) and phosphoinositide (PI) 3-kinase also appear to associate with integrins as a necessary interaction for motogenesis (Clark & Brugge, 1995). Finally, the Rho subfamily is also involved in the signalling cascade that leads to the formation of lamellipodia, focal adhesion and stress fibres (Hotchin & Hall, 1996; Tapon & Hall, 1997). In addition, regulation of Rho is implicated in adhesive release, as when Rho is inactivated the actin cytoskeleton collapses and the cells adopt a rounded morphology (Miura et al., 1993). Even less is known concerning the mechanisms involved in crossing the basal membrane; however, surfaceassociated proteases seem to be important.

In conclusion, although there are still more questions than answers, some progress has been made in defining the relevant molecules that regulate this last process of cell extravasation.

Diversity of leukocyte migration: specific mechanisms

The mechanisms discussed up to now apply generally to all leukocytes. However, the migration of various leukocyte populations is not a random process but has been fine tuned to allow for diversity and specialization in response to different conditions. These include, among others, the stage and type of inflammation, the prevalent pattern of cytokines/chemokines produced in various tissues and the state of lymphocyte differentiation (Pitzalis, 1993). It is well known, for example, that during an acute inflammatory response neutrophils are the prevalent cells, while during the chronic phase mononuclear cells predominate. In allergic

reactions or in responses to parasites, by comparison, eosinophils come prominently into play. This is mainly a result of the increased production of IL-5, which stimulates eosinophil differentiation in the bone marrow and, as discussed above, of the local production of eotaxin, which favours eosinophil extravasation into the allergic tissues (Rothenberg et al., 1996). Finally the state of leukocyte differentiation can greatly influence their trafficking pattern. The most striking example is the different recirculatory pattern shown by naive/resting (CD45RA⁺) versus memory/activated (CD45R0+) lymphocyte populations. Naive lymphocytes recirculate mainly through secondary lymphoid organs (peripheral lymph nodes, Peyer's patches, tonsils and spleen), where they are primed to various antigens drained from different tissues. Memory/activated lymphocytes, although capable of accessing lymphoid organs, primarily recirculate to peripheral inflamed tissues, where they can exert their effector function (Mackay et al., 1992a). Evidence for this dichotomy in vivo comes from animal work demonstrating that immunoblasts from the gut and skin, as well as from peripheral lymph nodes, preferentially home to the same type of tissue from which they were isolated (Gowans & Knight, 1964; Rose, Parrott & Bruce, 1978; Issekutz, Chin & Hay, 1982; Mackay et al., 1992b). However, in humans the evidence is mainly indirect. In vitro studies, using a lymphocyte-binding assay with frozen tissue sections, have demonstrated a differential ability of various lymphoblastoid lines to bind to peripheral lymph node tissue and synovial microvascular endothelium (Jalkanen et al., 1986). Using the same assay, it has been also shown that T cell lines derived from peripheral lymph node, gut and synovium preferentially adhere to homotypic tissue sections (Salmi et al., 1992). On the basis of these and many other studies, it has been proposed that entry into different tissue sites is regulated by specific receptor/counter-receptor pairs of adhesion molecules expressed by migrating lymphocytes and by the microvascular endothelium of a given organ. The lymphocyte-associated molecules, bearing the organ selective interaction with the microvascular endothelium, are termed homing receptors (HR), while the cognate microvascular endothelial ligands are defined as vascular addressins (VA) (Butcher, 1992; Picker, 1992; Butcher & Picker, 1996).

The first characterized example of HR–VA interaction was the HR expressed by lymphocytes homing to peripheral lymph nodes. This HR, initially defined in the mouse by the mAb MEL-14 (Gallatin, Weissman & Butcher, 1983) and in human by the DREG series of mAbs (Kishimoto, Jutila & Butcher, 1990), which recognizes L-selectin, specifically mediates binding to endothelial VA of the peripheral lymph nodes (Imai *et al.*, 1991). The carbohydrate-based VA in the peripheral lymph nodes was initially identified in mice using the mAb MECA-79 (Streeter, Rouse & Butcher, 1988b). This was subsequently found to be the equivalent of human GlyCAM-1 (Michie *et al.*, 1993). Another mAb from

the MECA-series (MECA-367) (Streeter et al., 1988a) was instrumental in identifying the mucosa-associated VA. While MECA-79 differentially stains and blocks the adhesive function of VA in peripheral lymph nodes, MECA-367 shows the converse pattern of reactivity, staining the microvascular endothelium of Peyer's patches. This led to the definition of a second pair of HR/VA molecules in which the $\alpha_4\beta_7$ integrin was shown to selectively bind to mucosal VA, now called MAdCAM-1 (Berlin et al., 1993; Briskin et al., 1993). The functional role of L-selectin and $\alpha_{4}\beta_{7}$ integrin in mediating specific lymphocyte homing has been recently demonstrated by specific gene ablation in mice. As expected, L-selectin knock-out mice have a severe reduction in the size of their peripheral lymph nodes (Arbones *et al.*, 1994) while β_7 integrin-deficient animals show an underdeveloped MALT (Wagner et al., 1996). Lymphocytes derived from these animals display a decreased capacity to bind to peripheral lymph nodes and mucosal HEV, respectively. Moreover, in humans, a particular type of non-Hodgkin's lymphoma characterized by multifocal infiltration of the intestinal tract (malignant lymphomatous polyposis) expresses $\alpha_4\beta_7$ integrin, unlike the nodal variety of the disease (Pals et al., 1994). This suggests that HR are involved not only in normal lymphocyte recirculation but also in determining the pattern of dissemination of malignant cells. Further support for this hypothesis comes from the fact that, in cutaneous T cell lymphoma, most lymphocytes infiltrating the skin express the cutaneous lymphocyte antigen (CLA) (Picker et al., 1990). CLA is a cell-surface glycosylated protein that contains a CD15 (Lewis^x) carbohydrate backbone and binds strongly to E-selectin (Koszik et al., 1994). This has led to the proposal of CLA and E-selectin as the HR-VA specific for the skin (Picker et al., 1990; Berg et al., 1991; Picker, 1992).

Although other homing specificities to organs such as the lung (Picker *et al.*, 1994) and the synovium (Jalkanen *et al.*, 1986) have been described, the idea of selective migration to different peripheral tissues remains controversial. Whereas there is little doubt of the critical importance of HR–VA adhesive interactions, it has become clear that the molecular control of lymphocyte homing is a flexible process that can adapt to different pathophysiological situations. For example, while selective lymphocyte homing is of major importance during the 'basal patrolling' of various organs, it becomes much less so during acute inflammation. For instance, in basal conditions, the traffic of memory cells through lymph nodes is low while this increases dramatically in those lymph nodes draining an inflammatory focus (reactive lymphadenopathy) (Mackay, Marston & Dudler, 1992a). It has also been recognized that a model postulating that a single tissue specificity depends on a single homing receptor–addressin molecule interaction is too simplistic (Picker, 1994; Butcher & Picker, 1996). Some HR, for example, facilitate lymphocyte adhesion to more than one tissue and some of the

proposed 'specific' VA molecules have a widespread distribution in different organs (Picker, 1994; Butcher & Picker, 1996). For example, as mentioned above, it has been suggested that the accumulation of memory T cells to the skin is dependent on the interaction of CLA with E-selectin (Picker et al., 1990; Berg et al., 1991). However, E-selectin is highly expressed on the MVE of various inflamed tissues, including the skin and synovium in psoriatic arthritis, but CLApositive lymphocytes are found only in the skin and not the joint (Pitzalis *et al.*, 1996). Therefore, the expression of E-selectin per se is not sufficient to favour the migration of CLA-positive cells to tissues other than the skin. Conversely, a single molecule found mainly on one tissue can bind more than one subset of T cells. For example, a variant of the gut addressin MAdCAM-1, when expressing a carbohydrate determinant that ligates L-selectin, binds both $\alpha_{4}\beta_{7}$ -positive/Lselectin-positive (naive) and $\alpha_{4}\beta_{7}$ -positive/L-selectin-negative (memory) lymphocytes (Sampaio et al., 1995). Therefore, it is likely that the extravasation of different lymphocyte subsets into various tissues is the result of the interaction of multiple adhesion molecule pairs acting in combination, 'telephone area code model' (Springer, 1994). Quantitative as well as qualitative differences in the expression of HR/VA molecules also appear to be important (Butcher & Picker, 1996).

In addition, it is now clear that the original description of the multistep model as a hierarchical process, in which each step was mediated in sequence by different adhesion molecules, was too inflexible. Subsequent studies have, in fact, demonstrated that the same molecule can be involved in more than one step of the adhesion cascade and that some adhesion proteins can be equally involved in general migration and organ-specific homing. For example, α_4 integrins can mediate both secondary and primary adhesion, as indicated by the fact that $\alpha_4\beta_1$ and $\alpha_{A}\beta_{7}$ integrins can mediate both firm adhesion and tethering/rolling to purified VCAM-1 and MAdCAM-1, respectively, under conditions of flow (Alon et al., 1995; Berlin et al., 1995). L-selectin, however, is involved in primary adhesion as well as in naive lymphocyte homing to peripheral lymph nodes. Moreover, it is also apparent that selective leukocyte extravasation is dependent not only on ligand-receptor adhesive interactions but also on specific activation signals. For example, it would appear that GlyCAM-1 mediates the migration of naive lymphocytes to the peripheral lymph nodes, not only because of its preferential expression on HEV in that tissue and its interaction with L-selectin, but also because of its capacity to activate CD45 RA lymphocytes specifically (Hwang et al., 1996). Finally, the capacity of CKs to activate selectively different leukocyte populations can further contribute to the development of migration specificity and diversity. In summary, therefore, evolution has allowed provisions for a combinatorial adhesion and signalling system in which general

mechanisms of migration integrate and overlap with organ-specific homing, depending on the various requirements associated with different pathophysio-logical situations.

Mechanisms of cell orientation, migration and retention within tissues

Once recruited into tissues, leukocytes follow different fates. Neutrophils tend to die while mononuclear cells may return to the circulation via the lymphatics or may be retained within the tissue, where they tend to segregate into specialized microenvironments. The best example of this 'microenvironmental homing' is represented by the specific localization of B and T cells in particular areas of lymphoid tissue. In chronic inflammatory lesions, such as the RA synovium, mononuclear cells can be found either in a diffuse distribution, scattered throughout the synovium, or in large focal aggregates, constituting follicle-like structures similar to those found in lymphoid organs. The molecular mechanisms controlling cell locomotion and retention within tissues are complex, once again, and not completely understood. However, adhesion to extracellular matrix constituents such as collagen, proteoglycans, laminin and fibronectin plays a major part. Lymphocytes interact with the extracellular matrix by means of specific adhesion receptors mainly belonging to the β_1 integrin subfamily (also known as VLA molecules). Each VLA integrin has been shown to mediate adhesion to at least one of the three major extracellular matrix glycoproteins. In addition, each ligand is recognized by multiple VLA integrins. Lymphocytes adhere to fibronectin mainly via VLA-4 and VLA-5 integrin receptors, which recognize two different binding sites on the fibronectin molecule. VLA-4 binds to the third connecting segment (IIICS) region (Wayner et al., 1989; Mould et al., 1990), whilst VLA-5 recognizes the key short peptide sequence RGDS within the central cell-binding domain (Pierschbacher, Hayman & Ruoslahti, 1985; Ruoslahti & Pierschbacher, 1987). The collagen receptor has been identified as VLA-1, VLA-2 and VLA-3. Interestingly, VLA-1 and VLA-2 are not present on resting T cells, but they are expressed after long-term activation in vitro and in approximately 60% of synovial T cell in RA. The VLA-6 integrin, which was first identified as a laminin receptor on platelets, has recently also been shown to be the laminin receptor on lymphocytes. As a result, the extracellular matrix can provide the supporting physical environment in which lymphocytes can come into close contact with other cells such as antigen-presenting cells for the initiation of the immune response. Furthermore, proteins in the extracellular matrix have multiple adhesive domains that facilitate, in return, a multiplicity of interactions between cells and the matrix; these provide co-stimulatory signals for the amplification of such responses (Matsuyama et al., 1989; Shimizu et al., 1990; Yamada et al., 1991).

In addition to these general mechanisms, specific mechanisms must exist to provide directionality and selective interactions with specialized stromal cells such as the follicular dendritic cells of B cell follicles and the interdigitating cells of T cell zones. In this respect, chemotactic factors and hapoptactic signals are thought to be important. In chemotaxis, cells migrate in the direction of the highest concentration of a chemoattractant (typically a soluble molecule). In hapoptaxis, cells move toward the region of highest adhesiveness (increased expression of adhesion ligands). Several of the molecules already discussed appear to be important in providing spatial orientation signals. CK and CKR may play a critical role in microenvironmental homing, as suggested by the lack of germinal centre formation in BLR-1 (CKR homologue)-deficient animals, which arises because of the inability of B cells from these animals to migrate from the T cell to the B cell areas (Forster et al., 1996). Cytokines also seem important. For example, focal lymphoid aggregate formation in inflamed tissues appears to be dependent on the local production of TNF α as suggested by the fact that TNF α deficient animals lack proper germinal centres (Pasparakis et al., 1996). By comparison, transgenic animals overexpressing the human $TNF\alpha$ gene have increased formation of focal lymphoid aggregates and develop a chronic arthritis very similar to RA (Keffer et al., 1991). In summary, cell migration and retention within the tissues is probably regulated by overlapping and combinatorially determined adhesion and activation signals in a similar manner to that seen in the events that control cell extravasation from the blood into the tissues.

Adhesion mechanisms as therapeutic targets

Studies in the 1990s have illustrated the paramount importance of adhesion mechanisms in the pathogenesis of inflammation. Not surprisingly, therefore, there has been a great deal of interest in targeting these mechanisms to treat inflammatory conditions such as rheumatic diseases. Moreover, it has become clear that some of the commonly used therapeutic agents in rheumatology have the capacity of modulating these mechanisms. For example, corticosteroids are known to induce leukocytosis and a concomitant decrease in leukocytes in the RA synovium by mobilizing the marginated pool and inhibiting the migration of leukocytes to the joint (Smith *et al.*, 1988; Youssef *et al.*, 1996). Studies *in vitro* have confirmed that corticosteroids can directly inhibit the expression of endothelial (Eselectin and ICAM-1) and leukocyte (LFA-1 and CD2) adhesion molecules (Cronstein *et al.*, 1992; Pitzalis *et al.*, 1997). Methotrexate has been shown to decrease *in vivo* leukocyte adhesion and migration in rat mesenteric venules (Asako, Wolf & Granger, 1993). This effect is related to the ability of methotrexate to enhance adenosine release, which in turn inhibits the production of the superoxide normally responsible for increasing leukocyte adhesion to endothelial cells (Cronstein *et al.*, 1991). Sulphasalazine, but not sulphapyridine (inactive moiety), has been shown to inhibit the activation-dependent upregulation of CD11b/CD18 by granulocytes and monocytes but not from lymphocytes (Greenfield *et al.*, 1993). Gold treatment decreases the expression of E-selectin by synovial endothelial cells in patients with RA (Corkill *et al.*, 1991). Finally, in patients with psoriasis, cyclosporin A has been shown to reduce dramatically the number of T cells infiltrating the skin and the expression of ICAM-1 on keratinocytes, although, interestingly, not on endothelial cells (Horrocks *et al.*, 1991).

In addition to the 'old drugs', new therapeutic modalities specifically designed to modulate adhesion mechanisms have been developed. The therapeutic potential of mAbs against adhesion molecules has been demonstrated in a variety of animal models, including allograft rejection, cardiac reperfusion injury and experimental autoimmune encephalomyelitis (Isobe et al., 1992; Ma et al., 1992; Yednock et al., 1992). As far as arthritis is concerned, blocking either the LFA-1 or the VLA-4 pathways has been shown to inhibit disease in animals (Jasin et al., 1992; Barbadillo et al., 1995). As a result of these studies, a trial using anti-ICAM-1 mAb has been conducted in RA (Kavanaugh et al., 1994). The patients showed a transient improvement that correlated with the development of a peripheral blood lymphocytosis, suggesting that inhibition of lymphocyte migration into inflammatory sites was occurring. However, the therapeutic effect could also have occurred by inhibiting other adhesion-dependent immune functions in which ICAM-1 plays a major role. Other attempts to block ICAM-1 have been made using soluble and recombinant ICAM-1 constructs (Martin et al., 1993). A new and very exiting therapeutic modality, presented by W. R. Shanahan at the Fourth Symposium on Immunotherapy in Cyprus (May, 1997), is the usage of an ICAM-1 antisense oligodeoxynucleotide product. This preparation hybridizes with a 20 base sequence of the 3'-untranslated region of human ICAM-1 mRNA and inhibits ICAM-1 expression in vivo as well as in vitro. The drug was shown to prolong cardiac allograft survival and to inhibit dextran sulphate-induced colitis and collagen-induced arthritis in mice. The results of a double-blind placebocontrolled trial in patients with steroid-dependent Crohn's disease were also very encouraging. Despite a very short half-life of approximately 60 minutes, the drug produced a significant steroid-sparing effect and durable remission (mean approximately 5 months) in 47% of the patients. Other ways of modulating cell adhesion involve the use of selectin blockers. Several animal studies have demonstrated the efficacy of this therapeutic modality either using mAbs or blocking
oligosaccharides (Mulligan et al., 1993; Nelson et al., 1993; Buerke et al., 1994; Ward & Mulligan, 1994).

However, although therapies designed to block general mechanisms of adhesion might be feasible and effective in acute inflammation or during flares, their usage in chronic diseases requires some caution. In these conditions, the prolonged adhesion blockade, which is likely to be required, may lead to unacceptable side effects similar to those seen in the leukocyte adhesion deficiency syndromes. The alternative strategy would be to target molecules responsible for selective leukocyte migration, for example the inhibition of eotaxin in allergic inflammation. Blocking molecules involved in specific homing may also be a fruitful option. For example, the blockade of the $\alpha_4\beta_7$ integrin has been very effective in an animal model of inflammatory bowel disease (Hesterberg *et al.*, 1996). The discovery of joint-specific mechanisms of migration might open this therapeutic avenue for the treatment of rheumatic diseases.

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Regulation of apoptosis in the rheumatic disorders

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Introduction

Apoptosis is the name used to describe the morphology of cells that die by a variety of insults as well as during normal physiological processes such as embryogenesis and metamorphosis (Kerr, Wyllie & Currie, 1972). The characteristic features of apoptosis include membrane ruffling, cytoplasmic and organelle contraction, nuclear condensation and DNA cleavage. The resulting cellular fragments or apoptotic bodies, are subject to rapid ingestion by neighbouring cells and resident tissue phagocytes. Cell death associated with apoptosis, programmed cell death, has been contrasted with necrotic cell death, which occurs as the result of external damage and is characterized by cell expansion and apparently random disintegration of cellular constituents. More recently, apoptosis and necrosis have been shown not to be two entirely different modes of death but rather the consequence of the nature and strength of the inductive stimulus coupled with the availability of energy sources such as ATP (Leist et al., 1997). A detailed description of apoptosis and its relevance to biomedical science is beyond the scope of this review and the reader is referred to a series of reviews in Science, 10 March, 1995 and 28 August, 1998.

Biochemical regulation of apoptosis

There are two aspects of apoptotic death that are of particular relevance to immune responses.

1. Apoptotic death is non-inflammatory. Cell fragments are phagocytosed by neighbouring cells and/or professional phagocytes and degraded. Most evidence indicates that phagocytes are not activated following ingestion of these bodies (Savill *et al.*, 1993). The receptors and ligands responsible for uptake of apoptotic cells in mammalian cells are not yet defined.

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2. Apoptosis occurs through ordered biochemical pathways, and because of this is often referred to as programmed cell death (PCD). The pathways are regulated by a variety of stimuli related to immune cell function. These biochemical pathways have been intensely studied in the 1990s. They are schematically illustrated in Figs. 9.1–9.3 and discussed below.



Fig. 9.1. Cell death pathways: inducers and early signal transduction pathways of the major known apoptosis pathways. Interaction of the ligand with the receptor causes receptor clustering and recruitment of the adaptor molecules. These, in turn, facilitate enzymatic activity of the caspases. Note that TNF α may induce a death signal through TRADD or stimulate cell activation through the NF κ B pathway. These pathways are modulated by the Bcl-2 family of proteins (see Figs. 9.2 and 9.3) and by other downstream signal transduction pathways (see text). Dissolution of the cell is orchestrated by caspases and nucleases. DD, death domain; DED, death effector domain; CED3, *C. elegans* 3 homologous domain.



Fig. 9.2. Inhibitors of apoptosis. Endogenous and virus-encoded (Crm A, p35) inhibitors of apoptosis block the biochemical pathway at different stages. See text for discussion and references.



Fig. 9.3. Mechanisms of apoptosis regulation by the Bcl family. Most of the Bcl family are located at the outer mitochondrial membrane. Unapposed, proteins such as Bcl-2 and Bcl- X_L promote cell survival whereas Bax and Bad promote cell death. Bcl-2 and Bcl- X_L may act by forming heterodimers with the Bax or Bad and preventing the latter molecules from forming pores in the membrane (left side of figure). Pore formation is thought to allow the release of Apafs (apoptotic protease activating factors), which include cytochrome *c*. Another way in which survival is regulated is through phosphorylation of Bad (see right side of figure), which, in turn, results in the dissociation of Bad from Bcl-2/Bcl- X_I .

Induction and early signal transduction

Fas ligand (FasL) and TNF α are the prototypic inducers of apoptosis (Nagata & Golstein, 1995). These ligands induce clustering of their cognate receptors (Fas, TNFR1 or TNFR2), which leads to recruitment of the early signal transducing molecules (Fig. 9.1) (reviewed in Fraser & Evan, 1996). TNF α may induce either cell death or activation depending on cell type, the receptor (TNFR1 or TNFR2) engaged and the action of intracellular regulators. While FasL and TNF α are the best characterized, a number of other apoptotic ligands (TRAIL (APO-2L) and AIM-2) and receptors (DR3 (also called APO-3/TRAMP/wsl-1/LARD), DR4 and DR5) have recently been identified. The receptors share a domain of 60–80 amino acid residues in the intracytoplasmic portion of the protein, called the 'death domain'. TRAIL is the ligand for DR4, DR5 and DcR1, a 'decoy

receptor' that shares the same ligand binding as DR4 and DR5 but lacks a cytoplasmic signalling domain (see Gura, 1997) (Fig. 9.1). Since the roles of the newly described receptors have not yet been studied in rheumatic diseases, they will not be considered further in this review.

Fas and TNFR belong to a superfamily of molecules that now includes almost 20 members. The majority of the family members (e.g. CD40, OX-40, CD30, 4-1BB) promote cell survival. The complex interplay between different members of this family are exemplified by CD40 and Fas. Whereas activation of B cells by CD40 upregulates Fas expression and enhances susceptibility of B cells to Fas-mediated apoptosis, co-ligation of the B cell receptor inhibits apoptosis (Elkon & Marshak-Rothstein, 1996).

Intracellular regulators

Regulators of different stages of the apoptotic pathway have been identified and are shown schematically in Fig. 9.2. FLIPs (FLICE inhibitory proteins) regulate the most proximal part of the apoptotic pathway by competing for binding of FADD and FLICE (caspase 8) (Irmler *et al.*, 1997). Crm A and p35 are virusencoded products that inhibit the function of specific caspases. IAPs (inhibitors of apoptosis) are thought to bind to TRAFs (Uren *et al.*, 1996) – although the precise way in which they promote cell survival remains to be determined.

Mitochondria have become a major focus for studies of apoptosis regulation. Kroemer, Zamzami & Susin (1997) have proposed that opening of mitochondrial megachannels associated with a 'permeability transition' (PT) is a necessary step for the release of soluble factor(s) required for the execution of apoptosis. Bcl-2 family members (Reed, 1994) are localized to the mitochondrial and nuclear membranes. Bcl-2 and Bcl-X₁ are prototypic cell survival proteins whereas Bax and Bad induce cell death. These molecules are thought to regulate cell fate by differential pairing (Oltvai & Korsmeyer, 1994; Yin, Oltvai & Korsmeyer, 1994). For example, heterodimers of Bax and Bcl-2 repress cell death, whereas if most Bax protein is in a homodimeric form, apoptosis is induced (Fig. 9.3, left side). Since the Bcl-2 family share sequence and structural homology to bacterial poreforming proteins (Muchmore et al., 1996), one scenario is that induction of pores by Bax homodimers allows the leakage of cytochrome c and Apafs (apoptotic protease-activating factors) from the mitochondria (Golstein, 1997). Very recently, another regulatory pathway influencing Bcl pairing has been described (Franke & Cantley, 1997). Survival signals activate a PI₃-kinase/Akt signalling cascade leading to phosphorylation of Bad (Fig. 9.3, right side). Phosphorylation of Bad results in dissociation of this molecule from Bcl-2 or Bcl-X₁, allowing homodimers to suppress release of Apafs.

In lymphocytes, many survival signals are thought to act via modulation of expression of Bcl-2 family members; for example, CD28 co-stimulation of T cells or IL-4 stimulation of B cells enhance expression of the anti-apoptotic protein Bcl- X_{I} .

Proteolytic cascade

The major family of proteases involved in apoptosis of mammalian cells comprises cysteine proteases (previously called the ICE family, now renamed caspases: cysteine aspartate-specific proteases) that are homologous of the C. elegans CED-3 (Fraser & Evan, 1996). The first protease identified was an ICE (interleukin-1β-converting enzyme) homologue, a cysteine protease that cleaves pro-IL-1 β . More recently, a number of ICE homologues, caspases 2–9, have been identified (Alnemri et al., 1996). Caspases are cysteine-dependent proteases that have an unusual substrate specificity for peptidyl sequences with a P1 aspartate residue. They can be grouped into ICE-like (caspases 1, 4 and 5) and CED-3like (caspases 3, 6, 7, 8, 9 and 10) proteases based on sequence similarity and their relative susceptibility to the tetrapeptide inhibitors zYVAD and DEVDfmk, respectively. Most evidence suggests that, analogous to the clotting and complement pathways, many caspases activate each other in a cascade that leads to amplification of the death signal. The exact sequence of events in this cascade remain to be determined, although recent in vitro studies indicate that FLICE (Fig. 9.1) is able to process/activate all known ICE/CED-3-like caspases (Margolin et al., 1997). Transcriptional alterations in caspase levels have been detected under some circumstances but by far the most important mode of caspase regulation is post-translational and occurs by protein-protein interaction and proteolytic cleavage (reviewed in Kumar, 1995). All of the caspases are expressed constitutively as proenzymes that are cleaved to generate two subunits of ~ 20 and 10 kDa; these combine to form the catalytically active $\alpha_2\beta_2$ tetramer. The specificity of caspases for aspartate residues is similar to that of granzyme B, one of the proteolytic enzymes released by cytotoxic T cells, suggesting that similar substrates are cleaved by quite different inducers of apoptosis.

Apoptosis modifiers

Susceptibility to apoptosis is affected by the stage of ontogeny and the activation of the cell. Pro-proliferative cytokines such as IL-4, L-7 and IL-15 that signal through the common γ chain of the IL-2 receptor are generally anti-apoptotic for T (Akbar *et al.*, 1996) and B (Elkon & Marshak-Rothstein, 1996) cells and are associated with upregulation of Bcl-2 and/or Bcl-X_I. Cytokines such as IL-2 and IL-10 play more complex roles that can promote or inhibit apoptosis. Other signal transduction pathways impact on the final decision of the cell, including ceramide, the balance between the stress kinases (JNK and p38) and MAP kinases, and the *Ras* pathway of cellular activation.

Viruses regulate apoptosis

Viruses have often been implicated as aetiological agents in autoimmune diseases, including Sjögren's syndrome (see below). A number of viruses express apoptotic inhibitory proteins (reviewed in Thompson, 1995) and it is, therefore, worth considering their role in disease in this context. Two examples discussed are Epstein-Barr Virus (EBV) and herpesviruses, which produce v-FLIPs (see also Fig. 9.2). EBV expresses a number of anti-apoptotic proteins: LMP1, a transmembrane protein, recruits TRAF-like molecules and results in NFkB activation, cell proliferation and induction of Bcl-2. EBV also expresses a homologue of Bcl-2 (BHRF1) as well as a homologue of IL-10; these homologues protect human B cells from apoptosis. EBV, therefore, employs a number of antiapoptotic strategies that may lead to persistent viral infection. Viruses of the γ herpesviruses family and molluscipoxvirus express viral homologues of FLIP, an endogenous protein known to inhibit Fas-mediated apoptosis of naive lymphocytes. The viral inhibitors of apoptosis (v-FLIPs) act as FLICE antagonists (Fig. 9.2) and are thought to block the early signalling events of several other death receptors (DR-3, DR-4 and TNFR1).

Apoptosis in the immune system

Cell turnover

Cells of the immune system include T and B lymphocytes, monocytes, dendritic cells, natural killer cells and neutrophils. The enormous (billions) daily production of these cells is balanced by their removal. There is abundant evidence that these cells die by apoptosis and are removed by non-inflammatory mechanisms. A potential explanation for this observation in T cells could be that a Kruppel-like zinc finger transcription factor, lung Kruppel-like factor (LKLF), is transcribed in resting cells and maintains their viability (Kuo, Veselits & Leiden, 1997).

Central tolerance

Deletion of high-affinity, self-reactive lymphocytes in the thymus is one of the cardinal mechanisms that helps to avoid autoimmunity. Immature T (thymocytes)

and B (early B cells in the bone marrow) lymphocytes are more sensitive to ligation of their antigen receptors than are mature lymphocytes, ensuring that the high-affinity cells are deleted. Although very large numbers of these immature lymphocytes die by apoptosis, the precise signalling pathways have not yet been defined. Abnormalities that influence thresholds for apoptosis may have major impact on susceptibility to autoimmunity (see below).

Peripheral tolerance

Since T cells are positively selected on self MHC (perhaps + peptide) in the thymus, T cells react with low affinity to self MHC. Recent studies, in fact, suggest that this interaction is necessary for the short-term survival of T cells in the peripheral immune system (Benoist & Mathis, 1997). Autoreactivity is generally avoided by the induction of anergy when the T cell receptor (TCR) is engaged but CD28, the major co-receptor, is not (Schwartz, 1996). However, at sites of inflammation, co-receptor ligands such as B7–1 and B7–2 are highly expressed by antigen-presenting cells. Furthermore, these cells present self-peptides in the MHC grooves. This situation provides a potent setting for the initiation of autoimmune responses. It is counteracted by the removal of potentially autoreactive cells by activation-induced cell death (AICD).

Antigen-specific T cell activation and proliferation is associated with upregulation of Fas and FasL and, after several days, acquired susceptibility to FasL–Fas mediated apoptosis. Since Fas and FasL may be expressed on the same cell, activated clones may die by fratricide or suicide (Nagata & Golstein, 1995). The Fas pathway is probably one of many pathways to terminate T cell proliferation. AICD, therefore, limits unwanted expansion of T cell clones reacting against foreign antigens beyond the course of infection, as well as of clones that crossreact with self-antigens or have been activated by autoantigens during an inflammatory event.

Autoreactive B cell clones are subject to several levels of regulation (Nossal, 1996). To generate high-affinity IgG antibodies, B cells also require at least two signals: one generated through the B cell receptor and others derived from T cells (CD40L and cytokines). B cells that chronically encounter low levels of antigen downregulate their antigen-receptor signalling apparatus such that autoantigen recognition does not lead to clonal activation and proliferation (Goodnow, 1992). These anergized clones are rendered susceptible to FasL-mediated apoptosis in lymph nodes by a mechanism involving CD40-mediated upregulation of Fas on the B cell (Rathmell *et al.*, 1996).

Apoptosis regulation in autoimmune disorders

Defects in apoptosis are a cause of systemic autoimmunity

SLE is a systemic autoimmune disease characterized by the presence of autoantibodies directed against a range of intracellular nucleoprotein targets. The role of lymphocyte apoptosis in both central and peripheral tolerance has suggested that self-reactivity may result from abnormal regulation of apoptosis (Fig. 9.4*a*).





Fig. 9.4. Possible role of apoptotic pathways in autoimmune disorders. (a) Defects in apoptotic pathways promote the survival of potentially autoreactive, proinflammatory cells. The defect may arise through failure of a suicide mechanism, as shown on the left, or a failure to 'murder' activated target cells of the T, B or macrophage lineages (right). Failure to eliminate activated cells can result in prolonged effector functions such as CD40L 'help' to B cells, inappropriate survival of primed autoantibody-producing B cells or cytokine release by macrophages. (b) Apoptosis as a source of immunogens. In view of the prominence of nucleosomes and negatively charged phospholipids as antigens in diseases such as SLE, it has been speculated that highly accelerated rates and/or abnormal sites or abnormal processing of apoptotic cells could lead to autoantibody production. (c) Organ suicide. Cytokines could provoke the destruction of cells by turning on a cell death pathway. It has been reported that thyrocytes constitutively express FasL and that IL1-B induces the expression of Fas resulting in apoptotic death of the gland (Giordano et al., 1997). A similar co-expression of Fas and FasL, coupled with high expression of Bax, but not Bcl-2, in acini, may account for loss of glandular tissue in Siögren's syndrome (Kong et al., 1997) and, possibly, in insulin-dependent diabetes mellitus (Chervonsky et al., 1997). (d) Tissue injury in organ-specific autoimmune diseases. As shown in (a), defects in apoptosis may allow persistence of activated effector cells. The effector cell may be resistant to apoptosis but can itself induce tissue injury by apoptosis of specialized cells within the organ or by proinflammatory effects (b). Examples may include graft-versus-host disease, multiple sclerosis and, possibly, RA and Sjögren's syndrome. Most organ-specific autoimmune diseases show evidence of fragmented DNA at sites of tissue injury. This is not always proportional to the degree of inflammation, as macrophages are remarkably efficient at ingesting apoptotic cells.

Murine models of autoimmunity

Three different mouse models of lupus-like autoimmunity with lymphadenopathy and mutations in Fas or FasL have been described (reviewed in Nagata & Golstein, 1995). The severity of the lymphadenopathy and autoimmunity varies among the strains, which either fail to express Fas (MRL*lpr/lpr*), produce a dysfunctional Fas molecule (CBA*lpr*^{cg}/*lpr*^{cg}) or a mutant FasL (C3H*gld/gld*). Analysis of these mice has led to several important conclusions. All three strains exhibit diminished Fas-mediated lymphocyte apoptosis, resulting in lymphocyte accumulation and lymphadenopathy. Lymphocyte apoptosis is probably normal in the thymus and bone marrow but clearly is abnormal in the periphery, resulting in the failure to delete autoreactive T and B cells (defective AICD) (Russell *et al.*, 1993; Singer & Abbas, 1994). The mutations amplify any pre-existing tendency to autoimmunity, such as those that exist in the MRL (Watson *et al.*, 1992) but not the C57BL/6 strains.

Other lupus-prone strains have also been reported to have abnormalities in apoptosis. In these cases, the mutations probably interfere with the threshold required for apoptosis induction rather than defects in apoptosis receptors *per se*. In motheaten mice, deficiency of SHP-1 (PTP1c), a phosphatase that downregulates antigen receptor-mediated signalling in T and B cells and macrophages, leads to enhanced survival of autoreactive lymphocytes, autoantibody formation and an aggressive multisystem autoimmune syndrome (Tsui *et al.*, 1993; Pani *et al.*, 1995). NZB mice have reduced B cell apoptosis induced by cross-linking of surface IgM receptors (Kozono, Kotzin & Holers, 1996), suggesting that a defect in B cell signalling may explain the survival of autoreactive B cells.

The idea that defects in apoptosis induce systemic autoimmune disease is strongly supported by transgenic and knock-out studies in mice, as illustrated in the following examples. Bcl-2 transgenic mice with B cell-specific overexpression of Bcl–2 produced anti-DNA, anti-Sm autoantibodies and developed glomerulonephritis (Strasser *et al.*, 1991). IL-2 and IL-2 receptor knock-out mice developed lymphoproliferation, polyclonal T and B cell activation and evidence of systemic autoimmunity, with anti-red cell and anti-colon antibodies (Sadlack *et al.*, 1993; Suzuki *et al.*, 1995; Willerford *et al.*, 1995). Rather than the expected immunodeficiency state, defects in the IL-2 pathway led to reduced sensitization to Fas-mediated AICD (Fournel *et al.*, 1996). Widespread autoimmunity was reported in CTLA-4-deficient mice, consistent with the observation that CTLA-4 (the inhibitory homologue of the CD28 T cell co-stimulatory molecule) down-regulates antigen-specific responses and may directly lead to T cell apoptosis (Tivol *et al.*, 1995; Waterhouse *et al.*, 1995). Deficiency of Lyn, a *src* family tyrosine kinase that is part of the B cell antigen-receptor signalling cascade, causes

impaired B cell apoptosis associated with anti-dsDNA antibodies and glomerulonephritis (Hibbs *et al.*, 1995; Nishizumi *et al.*, 1995).

Human lupus and related systemic syndromes

Inherited Fas deficiency in humans (Canale–Smith syndrome (Drappa *et al.*, 1996), ALPS (Fisher *et al.*, 1995) HLPS (Rieux-Laucat *et al.*, 1995)) is also associated with lymphadenopathy, hepatosplenomegaly and systemic autoimmunity: most commonly antibodies against red cells and platelets. As in *1pr* mice, the phenotype associated with human *fas* mutations depends upon predisposing background factors (genetic or environmental). Although there are obvious differences between SLE and the Canale–Smith syndrome, it should be noted that one SLE patient with a *fasL* mutation has been reported (Wu *et al.*, 1996) and we have recently encountered a young girl with a heterozygous *fas* mutation, autoantibodies and lupus-like disease involving the central nervous system and kidneys. A small subgroup of SLE patients may, therefore, have structural Fas or FasL defects. Perhaps the most interesting possibility is that, as in the mice, *fas* mutations amplify autoimmunity traits in the affected families.

Most patients with SLE do not appear to have a defect in Fas expression or function (Mysler *et al.*, 1994; Ohsako *et al.*, 1994). Soluble Fas (sFas), a secreted variant with the potential to block FasL, can be detected in the serum of 10–50% of SLE patients. The levels of sFas, however, are only twice those of normals and similar findings have been observed in many other systemic autoimmune disorders (Mountz *et al.*, 1995). Detailed studies of FasL function and soluble FasL (sFasL) are yet to be published. One study has reported reduced anti-CD3-mediated T cell apoptosis in T cell lines derived from SLE patients (Kovacs *et al.*, 1996).

Observations in murine models of systemic autoimmunity and in the Canale–Smith syndrome suggest that regulation of lymphocyte apoptosis is crucial to the maintenance of peripheral tolerance. The only pathway that has been extensively analysed in human lupus so far is Fas/FasL. Other cell death molecules have now been identified (see the machinery of apoptosis, above) and it will be important to consider alternative pathways that contribute to the elimination of activated T and B cells. For example, TNF α is a potent inducer of apoptosis and some RA patients treated with anti-TNF α antibodies develop antidsDNA antibodies, which disappeared on cessation of treatment (Elliot *et al.*, 1993).

Apart from its role in immune tolerance, apoptosis may play a direct role in antigen selection and, possibly, even the autoimmune response in SLE (Fig. 9.4*b*). It has recently been shown that the translocation of nuclear antigens to near the

surface of ultraviolet-irradiated keratinocytes (LeFeber *et al.*, 1984; Golan *et al.*, 1992) can be explained by apoptosis (Casciola-Rosen, Anhalt & Rosen, 1994). Membrane alterations during apoptosis cause externalization of phosphatidylserine, of potential importance in the generation of anti-phospholipid antibodies. Furthermore, SLE patients frequently have lymphopenia and accelerated AICD *in vitro* (Emlen, Niebur & Kadera, 1994). On the basis of these findings, it has been suggested that apoptotic bodies contain biochemically modified self-antigens, which could be immunogenic (Casciola-Rosen *et al.*, 1994). However, Tan and colleagues (Casiano *et al.*, 1996) have demonstrated that a minority of lupus autoantigens are cleaved during apoptosis, and, conversely, many proteins that are not common antigens in SLE *are* cleaved.

In preliminary studies, Mevorach, Zhou & Elkon (1996) reported that large amounts of apoptotic material administered by the intravenous route can, albeit transiently, induce the production of certain autoantibodies in mice. In most individuals, however, extensive apoptosis occurs in the central lymphoid organs without evidence of autoimmunity, so an understanding of the qualitative and quantitative aspects of apoptosis, as well as the disposal mechanisms of apoptotic bodies may be informative.

Rheumatoid arthritis

In RA, an abnormal accumulation of inflammatory cells in the synovium leads to pannus formation and destruction of cartilage and bone. The mechanism responsible for synovial hyperplasia is not well understood and may result from an increase in the number of and/or reduced apoptosis of synoviocytes and infiltrating cells. Although evidence of apoptosis has been detected in RA synoviocytes (Firestein, Yeo & Zvaifler, 1995; Nakajima et al., 1995), it has been suggested that the extent of synoviocyte apoptosis in vivo is inadequate to counteract ongoing proliferation. This imbalance may be explained by the production of cytokines such as TGF β_1 and IL-1 β , which favour synoviocyte proliferation and inhibit susceptibility to apoptosis (Kawakami et al., 1996; Tsuboi et al., 1996). These and other signals could reduce apoptosis of synoviocytes, possibly through increased expression of the Bcl-2 family of proteins (Firestein et al., 1995; Sugiyami *et al.*, 1996). TNF α is a pleiotropic cytokine that appears to be acting as a proinflammatory molecule in the RA synovium (Paleolog et al., 1996). Since TNF α most likely signals through TNFR2 (see Fig. 9.1), NF κ B is activated, which, in turn, inhibits apoptosis (Beg & Baltimore, 1996). Alternatively, inflammatory changes such as oxidation may result in upregulation (Sugiyami et al., 1996) and mutations (Firestein et al., 1996) of the p53 protein. It remains to be determined whether there is a loss of function of this important growth suppressor.

Synoviocytes can be triggered to undergo apoptosis via a Fas-mediated pathway. Whereas anti-Fas-mediated apoptosis of synoviocytes from patients with and osteoarthritis was equivalent *in vitro* (Firestein *et al.*, 1995; Nakajima *et al.*, 1995), an increase in sFas has been detected in RA synovial fluid (Hasunuma *et al.*, 1997). At present it is unclear whether sFas is functional and whether sFasL is also produced in the joint.

The inflammatory infiltrate in RA comprises T cells, plasma cells, macrophages and neutrophils. Despite expression of Fas and detection of FasL mRNA (Sumida et al., 1997a) in infiltrating T cells, in situ observations of synovial lymphoid aggregates suggest low levels of apoptosis ((Firestein et al., 1995; Sugiyami et al., 1996). This may be because of high Bcl-2 expression by the T cells (Firestein et al., 1995; Sugiyami et al., 1996) or the production of an anti-apoptotic factor(s) by stromal cells (Salmon et al., 1997). Of interest, Sumida et al. (1997a) report that there are two distinct T cell populations in the rheumatoid synovium, one susceptible to Fas-mediated apoptosis and the other resistant. Fas-sensitive T cell clones had conserved amino acid residues in the CDR3 domain of their T cell receptor, consistent with an *in situ* antigen-driven response. The same group has reported that local administration of an anti-Fas mAb to HTLV-1 tax transgenic mice (a mouse model of RA) led to an improvement in arthritis within 48 hours which was sustained for 2 weeks (Fujisawa et al., 1996). Immunohistochemistry showed that 35% of synovial fibroblasts, 75% of mononuclear cells and some of the infiltrating neutrophils underwent apoptosis. It will be important to determine whether the induction of apoptosis in the rheumatoid joint can be used to therapeutic advantage.

Neutrophils isolated from RA joints spontaneously undergo programmed cell death *in vitro* with the same kinetics as those from peripheral blood (Bell *et al.*, 1995) and evidence of phagocytosis by macrophages has been reported (Savill *et al.*, 1989; Jones *et al.*, 1993). It is unknown whether macrophage phagocytosis of apoptotic neutrophils is sufficient to remove all of these cells and whether there are any pathological consequences of this process.

Sjögren's syndrome

Sjögren's syndrome is associated with intense lymphocytic infiltration and destruction of the salivary and lacrimal glands. Several reports have implicated apoptosis in the glandular destruction, and the high expression of the proapoptotic Bax protein in acini (Krajewski *et al.*, 1994; Kong *et al.*, 1996) suggests that they may be especially vulnerable to induction of apoptosis.

Acinar destruction may occur secondary to abnormal co-expression of Fas and FasL on the epithelial cells. Talal and co-workers have reported that whereas Fas

is only expressed on ductal epithelium and FasL is not expressed in normal glands both molecules are expressed on the acini and ducts in Sjögren's syndrome (Kong *et al.*, 1997). This abnormal co-expression could lead to apoptosis by a suicidal mechanism (Fig. 9.4*c*). In Hashimoto's thyroiditis, a similar mechanism, triggered by IL-1 β , has been proposed (Giordano *et al.*, 1997). IL-1 β leads to upregulation of Fas, engagement of constitutively expressed FasL and apoptosis of thyrocytes.

Since the majority of infiltrating mononuclear cells are CD4⁺, and FasL mRNA has been detected in lesional tissue (Sumida *et al.*, 1997b), an alternative mechanism of glandular injury has been proposed (Matsumura *et al.*, 1996; Sumida *et al.*, 1997b). In this model (Fig. 9.4*d*), cytotoxic T cells infiltrating the salivary glands induce apoptosis of the acinii.

Although Kong *et al.* (1997) have detected increased expression of Bcl-2 in the infiltrating mononuclear cells by immunohistochemistry, 40% of the T cells isolated from the glands of patients with Sjögren's syndrome are susceptible to Fas-mediated death *in vitro* (Sumida *et al.*, 1997b). The susceptibility of those T cells with more conserved TCR amino acid motifs to anti-Fas-mediated apoptosis led the authors to suggest that the anti-Fas-sensitive cells are the activated, effector T cell population. In addition to the acini and T cells, it will be important to examine the susceptibility of B cells to apoptosis since the 'pseudolymphomas' that occur in Sjögren's syndrome are derived from B lymphocytes.

Conclusions

Human diseases can be classified in terms of life and death problems of specialized cells (Carson & Ribeiro, 1993; Thompson, 1995). In this context, many autoimmune rheumatic diseases are characterized by immune cell infiltration resulting in the destruction of local specialized cells. Other autoimmune diseases are associated with inflammatory infiltrates and the accumulation of cells that fail to die. The first phase of investigative studies in these human diseases has focused on the necessary baseline information regarding which cells show the hallmarks of apoptosis and which cells express high levels of inducers or inhibitors of apoptosis. These studies, however, provide limited understanding of apoptosis regulation *in vivo*. Fortunately, in many cases, it has been possible to obtain cells from diseased tissues and examine their effector status and susceptibility to apoptosis *in vitro*. These functional studies, coupled with phenotypic or other markers of pathogenic effector cells, are necessary to distinguish whether the effectors are sensitive or resistant to apoptosis. This crucial distinction has important therapeutic implications.

In view of the major role of cytokines in mediating inflammation, it will also

be important to integrate the results of the studies on apoptosis with known abnormalities of cytokine production in each disease. For example, cytokines that promote proliferation (see above) are often associated with expression of survival molecules such as Bcl-2 and Bcl- X_L and will be expected to block many apoptotic pathways. Therefore, high Bcl-2/Bcl- X_L expression and relative resistance to apoptosis induction should not be interpreted as a primary abnormality of an apoptotic programme under these circumstances. Removal of cells from their *in vivo* milieu may artefactually effect their survival through cytokine/growth factor withdrawal.

The elucidation of the biochemical pathways and specific proteins that regulate apoptosis provide a remarkable opportunity to manipulate the life and death decisions of the cell. For example, terapeptide inhibitors can selectively block different caspases and arrest apoptosis (Rodriguez *et al.*, 1996). Alternatively, agonists of apoptosis such as FasL, TNF- α and even soluble autoantigens (Critchfield *et al.*, 1994) can be harnessed to destroy autoreactive cells or cells that have escaped from the normal regulation of cell death programme. The basic understanding and therapeutic manipulation of apoptosis will have far reaching implications for the future health of patients with rheumatic disease.

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The role of monokines in arthritis W. B. VAN DEN BERG and F. A. J. VAN DE LOO

Introduction

Rheumatoid arthritis (RA) is generally considered an autoimmune disease, with its main manifestation a chronic inflammatory process in the synovial tissues of multiple joints and concomitant local destruction of cartilage and bone. RA synovial fluid is predominantly enriched with neutrophils, whereas the most abundant cells in the synovial tissue are macrophages and T cells, with foci of plasma cells and variable degrees of fibroblast proliferation. Another characteristic feature is the considerable thickening of the synovial lining layer, comprising macrophage-like type A cells and fibroblast-like type B cells. A major site of cartilage and bone destruction originates from overgrowth of activated synovial tissue at the margins, the so-called pannus tissue, although cartilage destruction away from overgrowth is noted as well. A key ongoing issue of debate in this destructive process is whether it is mainly linked to the inflammatory infiltrate in the synovial tissue or whether autonomous activation of synovial tissue fibroblasts and macrophages is a major event. There is at least suggestive evidence from transfer studies in SCID mice that RA synovial tissue cells are potentially destructive and may cause substantial cartilage damage in the absence of T cells. Apart from mechanism of the destructive process, there is also a longstanding debate on chronic synovitis: is it a dominant T cell-dependent immune inflammation or does it reflect perpetuated activation of synovial fibroblasts and macrophages, with secondary but minor involvement of T cells. Arguments for the latter hypothesis are based on the detection of an abundance of macrophagederived cytokines (monokines) in the RA synovial tissue and the relative paucity of T cell factors (Firestein & Zvaifler, 1990; Firestein, Alvaro-Garcia & Maki, 1990). However, low amounts of T_{H} 1-derived cytokines such as IFN- γ and IL-2 may be sufficient to maintain T cell-driven synovitis, since similar difficulties were encountered in detecting T cell cytokines in proven, T cell-dependent delayed-type hypersensitivity reactions. There is no doubt that arthritis in animal models is more chronic and more destructive in the presence of T cell reactivity, either directed against retained antigens in joint tissues or cartilage-specific autoantigens such as collagen type II and proteoglycans. However, in the absence of a defined autoantigen in RA, antigen-specific immunomodulation is not a therapeutic option. In this chapter, we will focus on the role of the monokines IL-1, TNF, IL-6, IL-10, IL-12 and IL-15 in arthritic processes, with an emphasis on findings in animal models as well as human RA and identification of potential therapeutic targets.

Pathogenesis of arthritis

A central problem in chronic arthritis is the nature of the stimuli that drive the synovial inflammation. Theoretically, this may be provided by persistant exogenous stimuli such as bacteria or viruses that localize to joint tissues, either causing direct activation of synovial macrophages and fibroblasts or driving this pathway through T cell triggering. In addition, cartilage autoantigens may be involved, initiating the process or playing a major part at later stages as a consequence of marked tissue destruction and autoantigen release in the presence of loss of tolerance. Articular cartilage must somehow be involved, retaining exogenous antigens, releasing autoantigens or amplifying the synovial activation, since it is a long recognized finding in RA that arthritis wanes in a particular joint when the cartilage is fully destroyed or removed at joint replacement. A final option emerged from careful analysis of RA synovial cells, which demonstrate tumour-like behaviour upon culture in vitro and do not need additional stimuli to continue to mediator release and proliferation for several cell passages. Whether this transformed phenotype is cause or consequence of the chronic synovitis remains to be established.

As a reflection of T cell-driven or direct activation of macrophages and fibroblasts, large amounts of cytokines, chemokines and enzymes are produced. Proinflammatory cytokines and chemokines will then amplify the synovitis through stimulation of neighbouring cells, upregulation of adhesion molecule expression on endothelial cells, facilitating influx of more leukocytes from the blood and promoting enzyme release, which contributes to cartilage damage. Cytokines detected in vast amounts include TNF α , IL-1, GM-CSF and other growth factors, IL-6 and, more recently, IL-15 (Firestein *et al.*, 1990; Feldmann, Brennan & Maini, 1996). In addition, two modulatory cytokines are also abundant, IL-10 and TGF β , which possess immunoregulatory as well as anti-inflammatory activity, inhibiting IL-1/TNF production but also inducing enzyme inhibitors such as TIMP, the natural inhibitor of metalloproteinases (Fig. 10.1). IL-4 is virtually absent in the RA synovium, in line with T_H1 dominance.



Fig. 10.1. Interplay of cytokines in the synovial tissue. The regulatory cytokines IL-4, IL-10 and TGF β may interfere at various levels, their actions including skewing of T cell balances, inhibition of cytokine production by macrophages (Macr) and synovial fibroblasts (Fibro), and direct protective effects on the chondrocytes (Chondr) in the articular cartilage. APC, antigen-presenting cell.

Cytokine hierarchy, dominant role of TNF/IL-1

Given the vast abundance of a whole range of inflammatory mediators in the RA synovial tissue, it is encouraging to note that there seems to be substantial hierarchy. Evidence has emerged that TNF and IL-1 function as so-called master cytokines, orchestrating the synovitis (Arend & Dayer, 1995; Feldmann *et al.*, 1996). IL-1 and TNF protein were detected in the synovial fluid and in the synovial tissue; these mediators have been demonstrated at the mRNA level by Nothern blotting and *in situ* hybridization. Immunolocalization has identified predominant expression in macrophages. In addition, culture studies of synovial membranes or isolated cells further confirmed the enhanced production in RA synovia of these cytokines and it was claimed that TNF drives most of the IL-1 production (Feldmann *et al.*, 1996). It has to be noted that the synovial tissue is usually obtained at joint replacement surgery, at late stages of the disease. Samples from early stages are now becoming available through arthroscopic biopsies or blind small needle biopsies. Although some studies confirm the earlier findings, TNF and IL-1 are not always found and uncoupled mRNA expression of these cytokines may occur as well (Deleuran, 1996; Tak *et al.*, 1997; Wagner *et al.*, 1997). It is our impression from recent analysis of multiple biopsies that large variations exist among differing areas in the arthritic synovial tissue and clearly also among RA patients. TNF and IL-1 are not always seen in such samples.

Clinical trials with TNF/IL-1 inhibitors

Apart from detection of the presence of a cytokine, their roles can only be validated in vivo, by using specific antagonists such as neutralizing antibodies, soluble receptors or, in the case of IL-1, a natural inhibitor, IL-1ra. First studies done with the Centocor cA2 anti-TNF antibody led to rapid improvement in all RA patients studied (Elliott et al., 1993) and the magnitude of the clinical response and reduction in CRP was reproduced in a randomized, double-blind, multicentre placebo-controlled trial. Subsequent studies with other anti-TNF antibodies and engineered soluble TNF receptors linked to the Fc portion of IgG confirmed the prominent role of TNF in RA, although the degree of suppression was variable. Second and third injections were often less efficacious, and precise use and limitations in individual patients must await further development. First studies into the mechanism of action of anti-TNF therapy points to reduction of expression of vascular adhesion molecules and lower cell numbers in the synovial tissue (Paleolog et al., 1996). Recent X-ray analysis of a 6-month followup study did not provide evidence for protection against ongoing joint destruction (unpublished observations) and it is tempting to speculate that TNF is mainly involved in symptomatic aspects and pain in RA and less so in the joint destructive process. This would fit with a more dominant role of IL-1 in the latter process, which has emerged from studies in experimental arthritis models (see below).

Clinical trials with anti-IL-1 therapy are less well advanced, probably because of several factors, including the dogma that TNF blocking will eliminate most of the IL-1 effect, as well, the absence of eminent humanized anti-IL-1 antibodies, problems encountered with proper, scavenging soluble IL-1 receptors (sIL-1R) and the poor pharmacokinetics of IL-1 receptor antagonist (IL-1ra). When soluble IL-1 receptor was administered daily for 28 days and significant improvement was noted. Later on it became apparent that two types of IL-1 receptor exist (Collota *et al.*, 1993), the type I being the signalling receptor and the type II being a decoy receptor, having a function in scavenging. Unfortunately, the type I receptor was used in the clinical studies; it has poor affinity for IL-1ra, and high affinity for IL-1ra, in this respect interfering with the natural inhibitors. Recent studies with sustained dosing of IL-1ra yielded promising results. The impact on clinical parameters was less impressive compared with the anti-TNF data, but the treatment appeared to reduce joint erosions (Breshnihan *et al.*, 1996; Campion *et al.*, 1996). Given the huge dosages needed to control experimental arthritis with IL-1Ra, it is possible that optimal dosing has simply not been achieved yet. However, the relatively minor effect on joint inflammation but consistent effect on joint destruction fits with observations of IL-1 blocking in certain animal models of arthritis (see below).

Studies in animal models

While no animal model of arthritis fully mimics human RA, such models can be used to study certain aspects of the disease and comparison of findings in various models may provide insight into general pathogenic mechanisms. Further belief in an initiating role of TNF in arthritis has emerged from elegant studies in TNF transgenic mice. Overexpression of human TNF leads to chronic, erosive arthritis, and the disease can be prevented with anti-human TNF antibodies (Keffer *et al.*, 1991). It is not yet understood why arthritis is the major pathology in this mouse. Interestingly, the arthritis was completely abolished upon treatment with antibodies to the IL-1R, excluding the direct action of TNF. This is in line with the major arthritogenic potential of IL-1 (Probert *et al.*, 1995) and fits with earlier studies of direct injections of cytokines into knee joints of mice or rabbits. These studies showed major synovitis and characteristic cartilage damage with IL-1, either alone or in combination with TNF, whereas single injections of TNF alone were only marginally effective (Henderson & Pettipher, 1989; van de Loo & van den Berg, 1990).

Although the above data illustrate the potency of TNF and IL-1, it remains to be proved that these cytokines are also of crucial importance in established models of arthritis. The model most widely studied is the autoimmune collageninduced arthritis (CIA) in mice, which is based on immunization with foreign collagen type II(CII) in genetically susceptible mice, mainly DBA/1j or B10RIII mice. The arthritis expression is dependent on a mixture of anti-collagen type II antibodies and anti-CII T_H1 cells, and arthritis expression can be facilitated by injection of TNF/IL-1 or generation of non-specific inflammatory events with concomitant cytokine release at the time of expected onset (Killar & Dunn, 1989; Joosten, Helsen & van den Berg, 1994). When anti-TNF antibodies or other TNF inhibitors are given before or shortly after onset of disease, marked amelioration of arthritis was demonstrated by various groups (Williams, Feldmann & Maini, 1992; Wooley et al., 1993). We have recently compared the efficacy of TNF and IL-1 blocking in this model, including a kinetic study starting anti-cytokine treatment at day 0, 3 or 7 after onset of arthritis. It confirmed that TNF blocking is mainly effective in early stages and less so at established disease, whereas the


Fig. 10.2. Treatment of collagen-induced arthritis with anti-TNF α (*a*) or anti-IL-1 α , β (*b*). A single injection of antibodies was given intraperitoneally, at various time points after disease onset (arrows). Rat anti-TNF α was marginally effective at late stages. In contrast, combinations of anti-IL-1 α /anti-IL-1 β showed prevention at the beginnning and impressive reduction when given in fully established disease (Joosten *et al.*, 1996).

suppressive effect of IL-1 elimination was more pronounced and also still prominent in established disease (Fig. 10.2). The role of IL-1 was substantiated with IL-1Ra treatment, which, however, appeared only effective when supplied with osmotic minipumps and high, sustained dosing (van den Berg et al., 1994; Joosten et al., 1996). Recently, we also showed efficacy of local IL-1Ra gene transfer in the knee joint of mice with CIA (Bakker et al., 1997). The strong IL-1 dependence of this model is further substantiated by reduced expression in IL-1 β converting enzyme (ICE) knock-out mice and efficacy of ICE inhibitors in CIA in normal mice (Ku et al., 1996). Moreover, full prevention of CIA is seen in IL-1ß knock-out mice (unpublished observations). Similar approaches using immune-driven models in TNF-deficient mice are hampered by the fact that these mice show major developmental abnormalities in lymphoid tissue organization and generation of immune responses (Pasparakis et al., 1996). An elegant alternative is provided by the recent studies in TNF-receptor 1 (TNFR1)-deficient mice, crossed to DBA/1. Upon immunization with CII, these mice developed CIA with a low incidence and in a milder form. However, once a joint was afflicted, the arthritis progressed in that joint to the same end stage as observed in the wild-type mice (Mori et al., 1996). This again argues that TNF is important at onset but that progression is TNF independent. It could also suggest that anti-TNF treatment in RA patients may be beneficial, when the chronicity of the disease is, in fact, based on repeated flares, with each flare displaying TNF dependency.

In earlier studies in the mouse, it was shown that a chronic synovitis can be caused to flare with tiny amounts of antigen, when the joint displays local hyperreactivity by retained T cells in the chronic infiltrate (Lens *et al.*, 1986). Anti-IL-1 treatment appeared effective in these flares (van de Loo *et al.*, 1995a). However, such a joint is also highly sensitive to flares induced by cytokines such as TNF or IL-1 and, in fact, it suggests that any non-specific inflammatory insult, with concomitant release of such proinflammatory cytokines, may exacerbate a chronic arthritis through reactivation of retained macrophages (van de Loo, Arntz & van den Berg, 1992a).

In other models of arthritis we have analysed the relative impact of TNF α and IL-1 in inflammation and cartilage destruction. In a simple passive immune complex model in mice, we found strong IL-1 dependence (van Lent *et al.*, 1994), in line with the observations in the collagen arthritis and perhaps suggesting that collagen arthritis is to a large extent an immune complex-driven disease. In murine antigen-induced arthritis (a model based on a T cell-driven reaction to locally injected and retained antigen in the knee joint of preimmunized animals, the initial inflammation (i.e. joint swelling and early cell influx) was barely dependent on TNF and IL-1, suggesting substantial overkill by other inflammatory

mediators. However, late cellular infiltrate in the synovium and cartilage destruction was markedly reduced with anti-IL-1 treatment, showing high IL-1 dependence of these aspects of inflammation. In subsequent studies in non-immune arthritis, induced with zymosan or streptococcal cell wall (SCW) fragments, the IL-1 dependence of cartilage destruction was further substantiated (van de Loo *et al.*, 1992b; 1995b). Although IL-1 is not always the dominant cytokine in early joint swelling, cell influx and enzymatic proteoglycan degradation of cartilage, in all models tested so far it is the pivotal cytokine in cartilage proteoglycan synthesis inhibition. Normalization of this function by IL-1 neutralization always results in markedly reduced cartilage damage in late stages of the disease.

The TNF-IL-1 cascade

Returning to the issue of a TNF-IL-1 cytokine cascade in arthritis, we have carefully analysed the relative role of TNF and IL-1 in arthritis induced with SCW, allowing for proper measurement of inflammation, cartilage destruction and TNF/IL-1 levels in the inflamed synovium. In this model, we see pronounced uncoupling of joint swelling and cartilage proteoglycan synthesis inhibition. When TNF was blocked, using either anti-TNF antibodies or TNF binding protein (TNFbp: an engineered soluble receptor from Amgen), marked reduction in joint swelling was found, whereas inhibition of chondrocyte proteoglycan synthesis remained unchanged. In sharp contrast, IL-1 blocking with neutralizing antibodies or IL-1ra did not reduce joint swelling but fully normalized the chondrocyte synthetic function (Fig. 10.3). Intriguingly, TNF blocking in vivo did not reduce IL-1 levels in the arthritic synovial tissue. These findings indicate that TNF and IL-1 have separate functions. Moreover, a stringent TNF-IL-1 cascade is not seen in this model, and, in fact, potential uncoupling was already suggested by the greater impact of anti-IL-1 treatment compared with anti-TNF treatment in CIA. Potential uncoupling was further supported by unabated IL-1 production in SCW-induced arthritis in TNF-deficient mice. Swelling was much reduced but late arthritis was still prominent (unpublished observations). Although the triggering process in the human synovial tissue of RA patients may be different to that in SCW-induced arthritis or CIA, the claim that TNF is the key therapeutic target must be treated cautiously, especially with respect to the pivotal role of IL-1 in amplifying late synovial infiltrate and cartilage destruction.

A final remark should be made regarding the role of soluble TNF versus membrane-bound TNF. Although aspects of CIA and SCW-induced arthritis could be blocked with TNF inhibitors, pointing to TNF involvement, it appeared difficult to detect substantial levels of TNF in tissue washouts apart from the first hours after induction of arthritis. Recently, it was shown that arthritis could also



Fig. 10.3. Differential effect of anti-TNF α and anti-IL-1 α , β on joint swelling and inhibition of chondrocyte proteoglycan synthesis in the early stage (day 2) of SCW-induced arthritis. The latter in the articular cartilage is only normalized with anti-IL-1. IL-10 displays an activity pattern similar to anti-TNF α treatment, consistent with dominant suppression of TNF α levels. Similar TNF/IL-1 specific results were obtained using TNF-binding proteins and IL-1ra as scavenging molecules (data not shown).

be induced by transgenic overexpression of transmembrane TNF (Georgopoulos, Plows & Kollias, 1996), elegantly demonstrating that soluble TNF is not needed to get full expression of arthritis. First studies with SCW-induced arthritis in TNF-deficient mice, 'knocked in' with transmembrane TNF (in collaboration with the group of George Kollias do suggest restoration of the arthritis. It remains to be seen whether anti-TNF treatment with antibodies or binding proteins is sufficiently effective in such mice.

Role of IL-6

IL-6 is a glycosylated polypeptide of around 26 kDa mainly produced by monocytes, T cells and fibroblasts. It belongs to a family of proteins with overlapping functions: leukemia inhibitory factor (LIF), oncostatin-M, ciliary neurotrophic factor and IL-11. The IL-6 receptor consists of two subunits: the α chain, with the IL-6 binding site (gp80) and the signal-transducing β chain (gp130). The

gp130 receptor is shared by all members of the IL-6 family (Kishimoto, Akira & Taga, 1992). Both TNF and IL-1 can induce IL-6 in numerous cell types, and of the growing list of cytokines found in RA synovial fluid the concentration of IL-6 is high, many times greater than IL-1 and TNF levels (Houssiau, 1995). The levels of IL-6 in paired samples of synovial fluid and blood correlated in patients with RA. The higher levels found in synovial fluid of affected joints makes a plasma IL-6 flow into these joints unlikely, pointing to local production as the main source. In rats and mice, increased serum IL-6 activity followed closely the kinetics of development of collagen-induced and adjuvant arthritis (Sugita et al., 1993). Indeed, human synovial tissue and the articular chondrocytes can produce huge amounts of IL-6 (Guerne et al., 1989; Guerne, Carson & Lotz, 1990), and IL-1 and TNF synergistically increase the production of IL-6 in both tissues (Guerne et al., 1990) The expression of mRNA and protein in the inflamed RA synovium showed that IL-6 was detected in cells present in lymphocyte-rich aggregates, and these cells were often in contact with CD14+ tissue macrophages (Wood et al., 1992). It was shown that CD13+ synoviocytes can be stimulated to produce IL-6 by macrophages via CD14-mediated cell-cell contact (Chomarat et al, 1995a). In RA patients, the synoviocytes also expressed mRNA and protein of the other members of the IL-6-family, IL-11 and LIF. Oncostatin M was, however, exclusively expressed in the synovial tissue macrophages (Okamoto et al., 1997). The physiological relevance of this discrepancy has to be elucidated.

Although IL-6 levels reflect the local inflammatory activity, radiological changes in the joint did not correlate with IL-6 (van Leeuwen et al., 1995). However, IL-6 has recently been detected in complex with the shed form of the IL-6 receptor (sIL-6R; pg80) and, in contrast to IL-6, the local levels of IL-6/sIL-6R complex did correlate with the radiographic grades of joint destruction in patients with RA (Kotake et al., 1996). Both sIL-6R and 'neutralizing' anti-IL-6 antibodies can chaperone circulating IL-6, and in high-IL-6 producers, monomeric immune complexes are formed, causing accumulation of IL-6 and preserving its bioactivity (May et al., 1993). In the open study of Wendling, Racadot & Wijdenes (1993), murine anti-human IL-6 mAb treatment of patients with RA markedly improved ESR (eosinophil sedimentation rate) and the Ritchie's articular index. Indeed this treatment raised the serum IL-6 levels, suggesting a dominant carrier instead of scavenging function for the antibodies, resulting in enhanced IL-6 activity and, thus, supporting a beneficial role of IL-6 under these conditions. The recent development of IL-6R mutants with clean antagonistic activity (Romani et al., 1997) will provide new tools to study the role of IL-6 in human diseases.

The general impression generated by all of the activities linked to IL-6 is that

this molecule may have inflammatory as well as protective activity in the RA process (Fig. 10.4). A prominent activity, associated with its former name BSF-2 (B cell stimulatory factor) is stimulation of antibody production. Marked reduction in antibody levels was found after T cell-dependent antigen exposure in IL-6-deficient mice, whereas the T cell-independent IgM responses were not reduced. This suggests that IL-6 might have a prominent role in autoimmune rheumatoid factor (RF) production, as far as it reflects a T cell-dependent triggering. Although the role of RF is still not clear, its presence correlates with more severe and more destructive disease. A second aspect of proinflammatory activity is linked to its role in the bone marrow. IL-6, either injected or transgenically expressed in mice, resulted in increased numbers of megakaryocytes in bone marrow (Ishibashi et al., 1989). Enhanced maturation of macrophages and granulocytes and subsequent influx in inflamed joints might contribute to severity. It is also claimed that IL-6 can induce chemokines. Intriguingly, endothelial cells do express gp130 but lack the IL-6R. It was elegantly shown that IL-6 alone was ineffective but that IL-6 together with sIL-6R clearly induced chemokines (Romani et al., 1997). The relevance for leukocyte traffic of this IL-6 activity remains to be seen, since in the same study IL-1 and TNF α were shown to be much more potent inducers. A final activity to be mentioned is the stimulating role of IL-6 in bone resorption. IL-6 is produced by osteoblasts and exerts its activity through osteoclasts. Oestrogen-induced bone loss is reduced in IL-6-deficient mice, but no information is yet available on bone loss in arthritis.

Apart from these potentially proinflammatory activities, there is no doubt that IL-6 also induces protective effects. Induction of acute-phase proteins in hepatocytes includes enzyme inhibitors such as α_1 -antitrypsin. Recently, it was shown that soluble IL-1 receptor antagonist (sIL-1ra) is also an acute-phase protein and levels of both IL-6 and sIL-1ra paralleled the fever spikes in systemic juvenile chronic arthritis (Prieur, Roux-Lombard & Dayer, 1996; Gabay et al., 1997). Moreover, IL-1 and TNF are potent inducers of IL-6; however, in marked contrast, IL-6 does not induce destructive proteases in macrophages or fibroblasts but instead upregulates inhibitors, including TIMP as well as IL-1ra and TNF soluble receptors (Ito et al., 1992; Tilg et al., 1994) This suggests that IL-6 is not a downstream mediator of TNF/IL-1 activity but more a protein exerting feedback control. Cartilage cells make large amounts of IL-6 upon stimulation with IL-1, but studies in IL-6-deficient mice excluded a cofactor role of IL-6 in the cartilage destructive activity of IL-1 (van de Loo et al., 1997). In addition, there is ample evidence of the suppressive effect of IL-6 on IL-1 and TNF production.

Since it is difficult to block the high levels of IL-6 satisfactorily with



Fig. 10.4. Pleiotropic activity of IL-6, reflecting both inflammatory and protective actions.

antibodies or receptor blockers, the best approach in animal models is the use of IL-6-deficient mice. Given the role of IL-6 in antibody responses, immune models based on active immunization should be excluded and it comes as no surprise that CIA is markedly reduced in such a background, these mice displaying markedly lower anti-CII antibody levels. We have chosen to analyse non-immuno-logically mediated acute joint inflammation by intra-articular injection of zymosan in IL-6-deficient mice. No differences were observed in local mediator release of IL-1 or nitric oxide, or in systemic corticosterone concentrations compared with wildtype mice (van de Loo *et al.*, 1997). However, although the cellular infiltrate in the synovium was slightly reduced, cartilage destruction was higher, suggesting that IL-6 normally exerts a major protective role in cartilage damage.

IL-10 and TGF β , regulation of arthritis

Additional cytokines with immunoregulatory and anti-inflammatory properties include IL-4, IL-10, IL-13 and TGF β . These cytokines may derive from T_H² cells or T3 cells (TGF β), and their function in regulation of T_H1 responses and its consequence for chronic synovitis will be addressed elsewhere in this book. Apart from immunomodulation, these mediators also suppress IL-1 and TNF production and have a direct impact on the chondrocytes in the articular cartilage. Moreover, activated macrophages involved in control of inflammation will be an important source of IL-10 and TGF β in the inflamed synovium. At the local level in the inflamed joint, this might be a more important contribution in regulation of the arthritis than T_H² or T3 cells. T_H² cells are virtually absent in RA synovia as is IL-4 (Deleuran, 1996; Feldmann *et al.*, 1996). IL-10 is abundant in RA synovial tissue as detected by reverse transcriptase–polymerase chain reaction (RT–PCR) and immunostaining and also in supernatant of synovial cell cultures (Katsikis *et al.*, 1994). The endogenous IL-10 is functional, since neutralization enhanced the TNF α and IL-1 production in RA synovial cultures. However, the control is not maximal, since addition of recombinant IL-10 further inhibited TNF/IL-1 production. In similar studies with intact synovial tissue culture, it was found that exogenous IL-10 again inhibited IL-1 β production, but IL-4 appeared more potent and also induced the inhibitor IL-1ra. IL-10 appears to upregulate soluble TNF receptors. The sensitivity to IL-10 regulation is dependent on the differentiation stage of the synovial cells (Chomarat, Banchereau & Miossec, 1995b; Dechanet *et al.*, 1995a), and in that sense the use of isolated cell cultures may give a skewed impression of *in vivo* events.

Studies in arthritis models demonstrated that systemic anti-IL-10 treatment shortly before onset of CIA markedly enhanced the expression and severity of the disease (Kasama et al., 1995) and this is probably linked to increased expression of chemokines as well as enhanced IL-1/TNF α levels. We found that the best expression of CIA was achieved with a combination of anti-IL-10 and anti-IL-4 antibodies (Joosten et al., 1997a). Moreover, suppression of established CIA was shown with IL-10 treatment (Walmsley et al., 1996), but a much more pronounced suppression, including protection against cartilage destruction, was achieved with the combination of IL-10 and IL-4, although IL-4 alone was marginally effective (Joosten et al., 1997a). RT-PCR revealed suppressed TNF/IL-1 levels and upregulated IL-1Ra/IL-1 balance in synovial tissue and arthritic cartilage. Further studies in SCW-induced arthritis confirmed that pronounced suppression of TNF as well as IL-1 was only achieved with the combination treatment. Again, IL-10 was a dominant endogenous regulator, since anti-IL-10 treatment prolonged the chronicity of the synovitis and enhanced the cartilage damage (Lubberts et al., 1998).

Although IL-4/IL-10 treatment may seem an intriguing option compared with plain inhibition of TNF/IL-1, with additional upregulation of inhibitors, one should be aware of potential side effects. IL-10 is a potent B cell stimulator and is suggested to be involved in RF production in RA synovia (Dechanet *et al.*, 1995b). IL-4 has a positive effect on chondrocytes, preincubation reducing subsequent destructive effects of IL-1. However, IL-4 can also stimulate fibroblast proliferation and enzyme release, and analysis of safe dosing regimens needs proper attention.

TGF β is perhaps even more pleiotropic, showing strong immunosuppressive activity but also chemotactic potential. When given systemically it reduced CIA, but when injected in a normal joint, it induced recruitment of leukocytes and

stimulated local fibroblast proliferation, mimicking tissue fibrosis. When injected in an inflamed joint, it enhanced the cell mass in the synovial tissue yet counteracted the proteoglycan depletion in the articular cartilage, probably by stimulation of the chondrocyte proteoglycan synthesis and through its ability to inhibit production of metalloproteinases and to induce TIMP. This protective effect on the articular cartilage has been demonstrated by co-injection of TGF β , both in an IL-1-driven arthritis (van Beuningen *et al.*, 1994a; Glansbeek *et al.*, 1997) and in zymosan-induced arthritis. In addition, TGF β induced characteristic osteophytes at the joint margins, through its strong activation of periosteal cells and cartilage-inductive potential (van Beuningen *et al.*, 1994b). High levels of TGF β are found in RA synovial fluid and expression in synovial tissue has been noted, but interpretation of its role is complicated by difficulties in discriminating between the inactive precursor form, complexed with LAP (latency associated peptide) and the active TGF β .

In general, the destructive character of an arthritis probably depends more on the balance of proinflammatory and modulatory cytokines than on the actual levels of the former. Some models of arthritis are more aggressive than others, and it is tempting to speculate that this is linked to different cytokine balances, as a result of various arthritogenic stimuli or more T cell- or macrophage-driven pathogenesis. Progression of RA in various patients is also variable, perhaps related to different underlying processes or linked to differences in genetic regulation of proinflammatory and modulatory cytokines. There is no doubt that factors such as IL-10, IL-4 and TGF β have a protective potential, controlling the impact of proinflammatory and destructive cytokines such as TNF and IL-1. However, given the risk of side effects, it seems more safe just to block the proinflammatory cytokines. Further studies are needed to examine whether protective effects and side effects of modulatory cytokines are seen at similar or different concentration ranges or whether they vary with differing routes of administration (including targeted gene transfer).

IL-12, unmasking or controlling autoimmune reactivity

IL-12 is a heterodimeric cytokine, mainly produced by activated macrophages. It stimulates the development of naive T cells into $T_{\rm H}1$ cells and stimulates IFN γ secretion by such cells. Efficient stimuli to induce IL-12 in macrophages include lipopolysaccharides (LPS), bacteria and intracellular parasites, and this production is enhanced by IFN γ and suppressed by IL-4 and IL-10. In general, IL-12 is considered as a principal protective factor in bacterial infections, where it bridges innate resistance and antigen-specific immunity. However, IL-12 may unmask $T_{\rm H}1$ -dependent autoimmune reactions by its skewing of the $T_{\rm H}1/T_{\rm H}2$

ratios and may be a crucial intermediate in the often suggested link between bacterial infections and expression of autoimmune diseases. There are few data on production of IL-12 in RA synovial tissue, but in view of the predominance of T_{μ} cells in this compartment, its involvement is not unlikely. First studies in murine CIA have shown that early treatment with IL-12 during immunization promotes severe arthritis in disease-prone DBA/1 mice; in fact, IL-12 can replace the mycobacterial adjuvant normally needed to induce anti-collagen type II autoimmunity (German et al., 1995). However, IL-12 failed to promote CIA in C57B1 mice: it provoked CII-specific IFNy-producing T cells but failed to induce the arthritogenic anti-CII-specific antibody subclasses needed to bind to cartilage surface and to initiate arthritis by release of cartilage-specific T cell epitopes (Szeliga et al., 1996). Follow-up studies in DBA/1 mice showed that daily high-(1 µg) instead of low-dose IL-12 treatment suppressed CIA, and this was linked to reduced antibody responses (Hess et al., 1996). It is now apparent that high IL-12 dosing has profound side effects on lymphoid organs and bone marrow. Apart from the role of IL-12 in skewing immune responses, we have studied the role of IL-12 at later stages of the development of arthritis. When IL-12 is given at the time of onset of CIA, it markedly enhanced disease expression and severity, whereas anti-IL-12 antibodies prevented spontaneous and LPS-triggered expression of CIA. This makes IL-12 a challenging therapeutic target. However, in established disease, anti-IL-12 treatment is poorly suppressive; in contrast, late daily IL-12 treatment (0.1 µg) significantly suppressed instead of enhanced CIA (Joosten et al., 1997b). It was also apparent that IL-12 is a potent inducer of IL-10 production by macrophages and T cells in vitro, providing an important negative feedback mechanism that could prevent excessive activation and tissue pathology (Meyaard et al., 1996). In fact, we did observe extremely high IFNy and IL-10 levels after late IL-12 treatment, and the suppression of CIA by IL-12 could be abolished with concomitant anti-IL-10 antibody treatment (Joosten et al., 1997b). This suggests that IL-12 promotes arthritis at the onset whereas it is a suppressive factor in late-stage disease. This bimodal activity will seriously complicate IL-12-directed therapy in human arthritis.

IL-15, an alternative to IL-2 in T cell stimulation

IL-15 is an IL-2 homologue that, in contrast to IL-2 itself, is abundantly present in RA synovia. Many cell types including macrophages and fibroblasts, with the notable exception of normal T cells, can produce IL-15, but the trigger inducing IL-15 production is not known. Its potent T cell-activating activity and the subsequent triggering of macrophage TNF production by these IL-15 activated T cells makes it a pivotal candidate in the RA process. IL-15 potently attracts memory T cells, and much of the T cell chemotactic activity present in RA synovial fluid appears to be attributable to IL-15 (McInnes *et al.*, 1996; 1997). The induction of macrophage TNF production by IL-15-activated T cells appeared to be dependent on cell–cell contact and was linked to CD69 expression. Interestingly, CD69 is also pivotal in the cell–cell interaction of T cells and synoviocytes, resulting in cartilage-degrading protease release (Lacraz *et al.*, 1994). This new pathway of T cell-driven TNF production sheds new light on the critical T cell involvement in the RA process, which was long debated because of low IL-2 and IFN γ levels. Future studies with anti-IL-15 antibodies in RA patients and animals models are needed to show the relevance of this cytokine, which may potentially be higher in the hierarchy than TNF/IL-1.

Final considerations

This discussion of the role of the various monokines in the arthritic process surveys recent developments and provides some insight into the contribution of the more recently discovered cytokines with the higher code numbers. Although some of these display intriguing activities, it might still be therapeutically attractive to focus on the straightforward, arthritogenic cytokines TNF and IL-1. Given the separate activities regarding inflammation and cartilage destruction and the clear evidence from experimental models that IL-1-driven processes may occur uncoupled from TNF, it may be advisable to go for combination therapy, perhaps also including a protease inhibitor to control for cytokine-independent enzyme activities in the synovial tissue. As a basic approach, we have, in recent studies, selectively eliminated the synovial lining macrophages from the murine knee joint, using local application of phagocytosable toxic liposomes, before arthritis induction. This treatment almost completely reduced the onset of arthritis in the joint but did not fully control the destructive process (van Lent et al., 1996; van den Berg & van Lent, 1996). The synovial lining cells are highly reactive to TNF and IL-1 and are a major source of chemokines. However, our treatment probably also eliminated lining macrophages involved in production of controlling cytokines. It would appear that, in line with the various subsets of lymphocytes, there exists macrophage subset with more destructive or protective character. If such cells can be more properly defined, for instance with regard to CD markers, this may offer novel therapeutic targets in the near future.

This approach does not encompass arthritogenic antigens. If these exist in RA and are not different in every patient, they would provide an even better therapeutic target, provided that we could achieve in antigen-specific immunomodulation in established arthritis. At present, we are left with manipulation of key mediators. This might include the 'immunologic' approach of local generation of suppressive cytokines/monokines from T cell/macrophage origin, such as IL-4 and TGF β by so-called bystander suppression. This refers to skewing towards protective T_H2 or T_H3 responses against endogenous proteins abundantly present in the inflamed joint, such as heat-shock proteins or cartilaginous components. Another intriguing variant might be the focused introduction of 'planted antigens' in the joint by direct injection, in the presence of a manipulated, protective T cell response against these proteins. Further knowledge is needed on the true protective character of mediators such as IL-4, IL-10, IL-13 and TGF β in arthritis before such approaches can be safely applied to human arthritides.

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11

T lymphocyte subsets in relation to autoimmune disease

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Introduction

The immune system has evolved a wide variety of potent mechanisms for the elimination of pathogens. As these mechanisms are potentially damaging to the host, an essential feature of the adaptive immune response is that it should be able to distinguish self from non-self and respond, when appropriate, only to the latter. The failure of the immune system to make this distinction can result in autoreactive cells mounting immune responses against self tissues, causing life-threatening pathology. There are now an increasing number of diseases in which an autoimmune process has been implicated in the pathogenesis, including RA, insulin-dependent diabetes mellitus (IDDM), multiple sclerosis, thyrotoxicosis, systemic lupus erythematosus (SLE), autoimmune haemolytic anaemia and myasthenia gravis to name a few. In order to avoid the damaging reactions that can cause this type of disease, the immune system has evolved a number of strategies for preventing autoreactive cells from making responses against host tissues, thus maintaining a state of self-tolerance.

In broad terms, mechanisms of self-tolerance can be categorized into two different classes. In the first, tolerance is maintained by both intra- and extrathymic mechanisms. Autoreactive T cells that encounter self-antigen within the thymus are clonally deleted (Kappler, Roehm & Marrack, 1987) while autoreactive cells that escape this process are either rendered anergic (Mueller, Jenkins & Schwartz, 1989), a state thought to result from the non-immunogenic presentation of selfantigens, or simply fail to respond to self-antigens, a state known as clonal indifference (Ohashi *et al.*, 1991). These mechanisms may be termed 'passive' since they rely upon the functional absence of autoreactive cells or the failure of the self-antigens to be presented in an immunogenic form. In recent years, it has become evident that these mechanisms are not sufficient to account for tolerance to all self-antigens. There is now compelling evidence for the existence of a second class of mechanism for self-tolerance, in which autoimmunity is prevented by specific regulatory T cells. These mechanisms are termed 'active' since they rely upon the action of a regulatory population controlling autoreactive cells.

This chapter will discuss how T cell subsets have been implicated both in the cause and prevention of autoimmunity. T cell subsets will be defined by the pattern of cytokine secretion and their cell surface phenotype.

$T_{H}1$ and $T_{H}2$ subsets of peripheral T cells

Mouse CD4+ T cells

Although some CD4⁺ T cells can be cytotoxic, it seems that cells of this phenotype regulate the immune response predominantly through the production of cytokines. Long-term mouse CD4⁺ T cells clones can be divided on the basis of cytokine synthesis into two mutually exclusive subsets: T_{H1} cells, secreting IFN- γ , IL-2 and TNF, and T_{H2} cells, which secrete IL-4, IL-5, IL-6, IL-10 and IL-13 (Mosmann & Coffman, 1989). These cytokines activate different effector mechanisms on different target cells. For instance, IFN- γ is the major macrophage-activating factor and induces IgG2a antibody responses (Coffman *et al.*, 1988); IL-4 induces switching of B cells to IgE and IgG₁ production, and IL-5 is the principal eosinophil-differentiation factor. The activity of the cytokines produced make T_{H1} cells primarily mediators of cell-mediated responses whereas T_{H2} are preferentially mediators of humoral responses (Mosmann & Coffman, 1989). The existence of two different T helper populations that regulate independently cell-mediated and humoral immunities provides an explanation for the observed reciprocal regulation of these two responses *in vivo* (Parish, 1972).

The activation of murine T_H^1 and T_H^2 T cell subsets appears to be mutually antagonistic not only because the cytokines secreted by these subsets have distinct functional properties but also because they are often mutually inhibitory. For example, the synthesis and activation of IFN- γ is inhibited by the T_H^2 cytokines IL-4 and IL-10 (Fiorentino, Bond & Mosmann, 1989; Powrie & Coffman, 1993). Conversely, IFN- γ inhibits both the action of IL-4 on IgE and IgG₁ synthesis and the proliferation of T_H² cells (Fernandez-Botran *et al.*, 1988; Gajewski & Fitch, 1988).

Murine CD4 T cell clones have been isolated that do not fall into the $T_H 1/T_H 2$ classification but secrete both types of cytokine and are, therefore, described as $T_H 0$ (Firestein *et al.*, 1989). The existence of these $T_H 0$ clones suggests that CD4+T cells pass through an intermediate stage of differentiation and these uncommitted cells can be induced to secrete either $T_H 1$ or $T_H 2$ cytokines on subsequent stimulations depending on the environment in which they were primarily

activated. In strong immune responses characterized by repetitive stimulation, the $T_{H}1$ and $T_{H}2$ phenotypes may become dominant. Several factors have been shown to regulate the balance between $T_{H}1$ and $T_{H}2$ responses. The route of administration and physical state of the antigen (Asherson & Stone, 1965), dose of antigen (Hosken et al., 1995), MHC (Murray et al., 1989), antigen-presenting cell involved in the T cell activation (Chang et al., 1990; Gajewski et al., 1991; Stockinger et al., 1996) and hormones present at the time of activation (Daynes & Araneo, 1989; Ramírez et al., 1996) have all been shown to influence this balance in different experimental systems. However, the most important factors in determining the pattern of cytokines produced by T cells are the cytokines they themselves produce and those produced by the antigen-presenting cells. Early studies showed that IL-4 supports the development of $T_{\rm H}^2$ effector cells (Le-Gros et al., 1990) whilst IFN- γ , IL-12 and TGF β are strong inductors of T_H1 cell development (Swain, Weinberg & English, 1990; Hsieh et al., 1993). It is widely accepted that macrophages and dendritic cells when activated by bacterial components produce IL-12, and this cytokine directs the development of $T_{\rm H}1$ cells (Macatonia et al., 1993). However, the in vivo source of IL-4 is a matter of controversy and several cell types have been proposed as candidates in different experimental systems: NK1.1+ CD4+ T cells (Yoshimoto & Paul, 1994), recent thymic emigrants (Bendelac & Schwartz, 1991), eosinophils/mast cells (Paul, Seder & Plaut, 1993; Sabin & Pearce, 1995) and CD4+ T cells (Launois et al., 1997).

Human CD4+ T cells

Earlier studies analysing cytokine production by short-term human T cell clones failed to demonstrate segregation into T_H^1 and T_H^2 cells similar to that seen in murine T cell clones. The majority of these clones displayed a T_H^0 phenotype (Maggi *et al.*, 1988). However, when CD4⁺ T cell clones were obtained from individuals with chronic inflammatory or allergic diseases and they were activated with the disease-inducing antigen, the division between T_H^1 and T_H^2 clones was observed. T cell clones from these patients probably represent T cells that have been repetitively stimulated *in vivo*, suggesting that the chronic antigen exposure leads to a more stable polarization than does short-term culture. A T_H^2 pattern of cytokine production is observed in allergen-specific cells from atopic patients (Wierenga *et al.*, 1990). In contrast, T_H^1 cytokines are synthesized by T cell clones specific for *Borrelia burdorgferi* antigens obtained from patients with Lyme arthritis and by *Mycobacterium leprae*-reactive T cell clones obtained from individuals with tuberculoid leprosy (Haanen *et al.*, 1991; Yssel *et al.*, 1991).

CD8+ T cells

Whereas CD8⁺ mouse T cells stimulated *in vitro* usually secrete a T_H^1 pattern of cytokines (Fong & Mosmann, 1990), there is now increasing evidence that a T_H^2 -like pattern of cytokines is also inducible in these cells by IL-4 addition (Seder *et al.*, 1992). Recent studies have shown that alloantigen-activated mouse CD8⁺ T cells can be treated in a similar way to CD4⁺ T cells to generate T_H^1 or T_H^2 responses, the so-called T_C^1 and T_C^2 cells (Sad, Marcotte & Mosmann, 1995). Some human CD8⁺ T cell clones have been found to secrete a T_H^0 pattern of cytokines (Paliard *et al.*, 1988).

$T_{\rm H} 1$ and $T_{\rm H} 2$ cells and autoimmune diseases

As discussed, T_H^1 responses are characterized by the activation of the cellular component of the immune system and the synthesis of inflammatory cytokines. In contrast, some of the T_H^2 cytokines display anti-inflammatory effects and inhibit the development of T_H^1 responses, suggesting that T_H^2 cells could be the physiological regulators of T_H^1 cells. According to this hypothesis, the T_H^1/T_H^2 balance could be important in the induction and regulation of autoimmunity. T_H^1 responses against self-antigens would provoke the autoimmune disease and T_H^2 cells would have the potential to downregulate this autoimmune attack. This concept has been explored in several experimental autoimmune models, the best characterized being experimental allergic encephalomyelitis (EAE) and the spontaneous development of diabetes by the non-obese diabetic (NOD) mouse. It is also important to point out that some experimental autoimmune diseases are characterized by a dysregulated T_H^2 response (Prigent *et al.*, 1995).

Experimental allergic encephalomyelitis

EAE is an experimental paralysing disease that is studied as a model for multiple sclerosis (Zamvil & Steinman, 1990). The disease is induced in the appropriate animal strains after immunization with CNS antigens emulsified in complete Freund's adjuvant (CFA). This procedure is termed 'active EAE' in contrast to the induction of 'passive EAE' by the injection into naive animals of CNS antigen-specific lymphocytes. Most rat and mouse strains are resistant to the induction of active EAE, but this disease can be induced in Lewis rats after immunization with myelin-basic protein (MBP) in CFA (Hughes & Stedronska, 1973) and in SJL/J mice after immunization with pertussis toxin (Levine & Sowinski, 1973).

Lewis rats are susceptible to the induction of many experimental autoimmune

and inflammatory diseases such as EAE, mycoplasmosis (Davis *et al.*, 1982), experimental autoimmune myasthenia gravis (Biesecker & Koffler, 1988) and arthritis (Griffiths & DeWitt, 1984). The high susceptibility in Lewis rats has been related to the defective regulation of glucocorticoid hormone production. During immune or inflammatory responses, blood glucocorticoid concentration increases as a result of activation of the hypothalamic–pituitary–adrenal axis by inflammatory and immune mediators such as IL-1, IL-6 and TNF (Besedovsky *et al.*, 1986; Naitoh *et al.*, 1988; Warren *et al.*, 1988). The glucocorticoid hormones produced in this way have an inhibitory effect on the synthesis of these cytokines and, as a result, the amounts produced are regulated to avoid an excessive and dangerous immune and inflammatory response (Munck, Guyre & Holbrook, 1984). Lewis rats manifest a deficiency in this regulatory interaction between the immune and neuroendocrine systems (Sternberg *et al.*, 1989) and there is also evidence that patients with RA show a similar defective regulation of corticosteroid production (Chikanza *et al.*, 1992).

Similar to multiple sclerosis, certain forms of EAE in mice and rats are characterized by relapsing paralysis (Lublin et al., 1981; Feurer, Prentice & Cammisuli, 1985), but EAE in Lewis rats is characterized by a single episode of paralysis followed by spontaneous recovery. The animals develop a transient paralysis caused by the action of CD4+ T lymphocytes that infiltrate the CNS (Sedgwick, Brostoff & Mason, 1987). Animals recover spontaneously and become refractory to attempts to induce further episodes of disease (MacPhee & Mason, 1990). Several mechanisms responsible for the spontaneous recovery from EAE and the maintenance of the refractory phase have been proposed, including different regulatory leukocyte populations and several cytokines and soluble factors (Lider et al., 1988; Kennedy et al., 1992; Santambrogio et al., 1993). A role has been also suggested for neuroendocrine-mediated immunoregulation (Levine, Sowinski & Steinetz, 1980; MacPhee, Antoni & Mason, 1989). Blood corticosteroid concentration increases before the onset of paralysis, with its peak at the time of maximum clinical score. The animals start to recover from paralysis after the peak in serum corticosterone and when they recover fully the glucocorticoid levels fall to their normal values. The role of endogenously produced corticosterone was analysed by evaluating the effect of adrenalectomy in Lewis rats subjected to EAE. It was observed that adrenalectomized animals developed an unremitting and progressive paralysis with a fatal outcome. If these adrenalectomized rats were given corticosterone-replacement therapy at the onset of paralysis, they recovered within a few days and developed the refractory state like non-adrenalectomized animals (MacPhee et al., 1989). These observations showed that endogenously produced corticosterone plays an essential role in the recovery of rats from EAE. The interpretation of these observations is that the spontaneous recovery occurs because corticosteroids acutely depress the autoimmune response through their immunosuppressive and anti-inflammatory effects. Once animals have recovered from EAE, adrenalectomy has no adverse effect and the refractory phase develops as it does in non-adrenalectomized controls. Evidently corticosteroids are not directly responsible for the maintenance of the refractory phase of the disease, but these observations do not exclude the possibility that the transiently elevated levels of glucocorticoid hormones during paralysis have long-term effects on the immune response to MBP.

The analysis of cytokines synthesized by animals with EAE during the different phases of the disease show a predominant production of Th1 cytokines previous to the onset of disease and predominance of IL-10 and TGF- β during and after the recovery from paralysis (Kennedy et al., 1992; Issazadeh et al., 1995). The studies performed employing MBP-specific lines has shown that EAE in Lewis rats is mediated by CD4+ T cells that produce IL-2 and IFN-y (Sedgwick, MacPhee & Puklavec, 1989) whilst IL-4-producing lines are not able to induce passive EAE (F. Ramírez, unpublished data). Similar findings have been described in the mouse EAE model: MBP-specific T_H1 clones can induce disease but T_H2 clones do not and, in some circumstances, can inhibit the development of EAE (Chen et al., 1994; Kuchroo et al., 1995). However, other authors have obtained encephalitogenic $T_{H}1$ lines that were not suppressed by $T_{H}2$ lines (Khoruts, Miller & Jenkins, 1995). Our experience with the Lewis rat model confirms this observation: rat $T_{H}1$ lines are encephalitogenic and $T_{H}2$ lines do not transfer disease; however, we do not observe any protective effect from the T_{μ}^{2} lines when these are injected simultaneously with T_H1 lines (F. Ramírez, unpublished data).

From these observations it seems that EAE is provoked by a T_H^{1-} or cellmediated response but that after recovery the T_H^{1} reactivity may become dominated by T_H^{2} cytokines with the ability to downregulate cell-mediated responses. If glucocorticoids can tip the $T_H^{1/T_H^{2}}$ balance, favouring the latter, as suggested by the *in vitro* experiments (Daynes & Araneo, 1989; Ramírez *et al.*, 1996), then the refractory phase of EAE in Lewis rats can, in principle, be explained by endogenously produced glucocorticoids inducing a switch from a T_H^{1} to a T_H^{2} response to MBP (Mason, 1991). This hypothesis provides an explanation for the recovery and refractory periods, although a more detailed analysis is necessary to verify it.

NOD mice

NOD mice spontaneously develop insulitis that progresses into diabetes with a higher incidence amongst females. Studies have shown some evidence for a path-

ogenic role for T_H1 cells and a protective function for T_H2 cells in diabetes. The *in vivo* administration of antibodies against IFN-γ prevents the onset of diabetes in NOD mice (Debray-Sachs *et al.*, 1991) and systemic administration of IL-4 prevents the disease (Rapoport *et al.*, 1993). T cell clones with a T_H1 phenotype are able to accelerate the onset of disease and T_H2 T cell clones are innocuous but cannot prevent the damaging effects of the T_H1 clones (Katz, Benoist & Mathis, 1995). Recent data have indicated that the situation is very complex. For instance, IFN-γ knock-out NOD mice develop diabetes like normal NOD mice, indicating that this cytokine is not necessary for the induction of diabetes (Hultgren *et al.*, 1996). Another paradox is that IL-10 transgenes expressed in the pancreas of NOD mice rather that inhibiting actually promote the development of diabetes (Lee *et al.*, 1996). This result shows that in different *in vivo* systems, T_H2 cytokines can inhibit or promote the development of disease. It is apparent that a more detailed analysis is required to understand the aetiology of diabetes in NOD mice.

Subsets of peripheral CD4⁺ T cells identified by the expression of different CD45 isoforms

In the preceding discussion, subsets of T cells were characterized according to cytokine production following activation. An alternative approach is to examine the expression of cell-surface antigens by T lymphocytes. One marker that has proved useful in defining functionally distinct subsets of CD4⁺ T cells is the leukocyte common antigen (LCA or CD45). CD45 is expressed in a number of different isoforms, generated by the differential use of exons A, B and C at the amino terminus of the extracellular end of the molecule (Thomas, 1989). Any combination of these three exons may be used, resulting in theoretically eight different forms, including CD45 molecules lacking all three exons, denoted as CD45RO. The functional significance of the range of isoforms is unknown. However, the analysis of subsets of CD4⁺ peripheral T cells defined by the expression of different CD45 isoforms has provided clear evidence for functional specialization amongst these cells.

In the rat, a mouse mAb specific for the C exon of CD45 reveals two subsets of CD4⁺ T cells: one third CD45RC^{low} and two thirds CD45RC^{high}. Studies of these subsets reveal extensive functional differences. Responses to OVA-DNP made by nude rats reconstituted with different subsets of CD4⁺ T cells revealed that it was CD4⁺CD45RC^{high} cells that provided helper activity for primary antibody responses (Powrie & Mason, 1989). In contrast CD4⁺CD45RC^{low} cells were more potent at providing B cells with help for secondary antibody responses (Spickett *et al.*, 1983), suggesting that CD45RC^{high} cells are the precursors of

CD45RC^{low} memory cells. Further, CD45RC^{high} cells were found to mediate lethal graft-versus-host (GVH) disease or local GVH responses in the popliteal lymph node assay (Spickett *et al.*, 1983).

In vitro experiments with these subsets both confirmed and extended the data from *in vivo* studies. CD45RC^{low} but not CD45RC^{high} cells were found to mediate secondary anti-hapten antibody responses when cultured with B cells. Activation of these subsets also revealed differences in the patterns of secreted cytokines: levels of IL-2 and IFN- γ were much higher in cultures of activated CD45RC^{high} cells than in similar cultures of CD45RC^{low} cells, but the latter expressed higher levels of IL-4 mRNA (Arthur & Mason, 1986; McKnight, Barclay & Mason, 1991). The repertoire of cytokines synthesized by these subsets *in vitro* is in broad agreement with their behaviour *in vivo*. However, while there are considerable functional differences between these subsets, neither is homogeneous and other markers subdivide them (Fowell & Mason, 1993). Further, there is evidence that the CD4+CD45RC^{high} subset contains long-term memory cells (Bunce & Bell, 1997).

Subsets of mouse CD4⁺ lymphocytes based on the differential expression of the isoform CD45RB have been identified, and it is apparent that there are considerable similarities in the behaviour of these subsets with those of rat subsets defined by CD45RC expression (Bottomly *et al.*, 1989). In humans, there are two non-overlapping T cell subsets defined by the expression of the isoforms containing the CD45RA exon and an epitope characteristic of the CD45RO isoform, which are thought to correspond to the naive and memory T cell pool, respectively (Akbar *et al.*, 1988). However this conclusion has been reconsidered (Pilling *et al.*, 1996.)

Evidence for the presence of autoreactive cells in the T cell repertoire of normal rodents

There is now much evidence that overtly autoreactive cells are present in the normal repertoire of T cells and that they are prevented from mediating pathological responses to self-antigens through active regulation by other T cells. In particular, studies in which congenitally athymic (nude) rodents are reconstituted with congenic T cells or in which the T cell pool of normal rodents is perturbed by protocols rendering the host lymphopenic have provided convincing support.

Reconstitution of nude rodents with subsets of peripheral CD4+ lymphocytes

Previous studies have examined the behaviour of different T cell subsets following their adoptive transfer into nude rats. Nude rats receiving CD4+CD45RC^{low} cells isolated from congenic donors remained healthy and were more resistant to infection than unreconstituted rats. In contrast, rats injected with CD4+CD45RC^{high} cells from congenic donors developed a wasting disease with a fatal outcome. Examination of tissues from these rats revealed mononuclear cellular infiltrates in organs including liver, pancreas, stomach and thyroid. The development of the multiorgan pathology mediated by CD4+CD45RC^{high} cells could be completely prevented by the co-transfer of CD4+CD45RC^{low} cells from normal animals (Powrie & Mason, 1990).

Similar studies in the mouse have shown that reconstitution of C. B-17 *scid* mice with CD4+CD45RB^{high} lymphocytes from B*alb/c* mice caused the mice to develop a lethal wasting disease. Mice were found to be suffering from an inflammatory bowel disease (IBD) characterized by profound mononuclear infiltrates of the colon. Mice receiving injections of CD45RB^{low}, unfractionated CD4+ lymphocytes or co-transfer of CD4+CD45RB^{low} cells with CD4+CD45RB^{high} cells at a ratio of 1 : 2, respectively, remained healthy (Powrie *et al.*, 1993). Further studies of the mechanisms of pathology and disease control showed that *in vivo* blockade of IFN- γ by neutralising mAb and systemic administration of IL-10 but not IL-4 prevented colitis development in recipients of CD45RB^{high} cells (Powrie *et al.*, 1994) whilst anti-TGF β blocking mAb prevented control of colitis in recipients receiving both CD45RB^{high} and CD45RB^{low} cells (Powrie *et al.*, 1996).

In both these sets of studies, a subset of T cells capable of causing lethal inflammatory responses was demonstrable amongst CD4⁺ lymphocytes of normal rodents and their pathological activity was controlled by a distinct subset of lymphocytes, providing clear evidence of regulatory interactions between these subsets.

Cyclophosphamide-induced lymphopenia in NOD mice

The effect of cyclophosphamide, a cytotoxic chemotherapeutic agent known to induce lymphopenia, on the development of diabetes in NOD mice has been the subject of several studies. NOD mice treated with cyclophosphamide developed diabetes at an accelerated rate and the drug also increased the incidence of disease amongst male mice (Yasunami & Bach, 1988). Adoptive transfer of mono-nuclear cells from non-diabetic NOD mice prevented the accelerated disease

development. Cyclophosphamide was shown to be non-diabetogenic in other mouse strains and it was, therefore, concluded that it was not cytotoxic towards beta cells of the pancreas. The drug probably has an immunomodulatory action, allowing diabetogenic cells to act on beta cells (Charlton *et al.*, 1989). Studies of lymphocyte population dynamics of NOD mice treated with cyclophosphamide revealed an initial drop in cell numbers in all lymphoid organs followed by an excessive repopulation of the pancreatic lymph node coincident with development of insulitis (Zhang, Georgiou & Mandel, 1993). These data suggest that the immune response that results in the destruction of the beta cells of the pancreas may be initiated in the pancreatic lymph node, which may also be the site of regulation of diabetogenic T cells.

Autoimmune-like pathology in mice thymectomized neonatally

In particular mouse strains, thymectomy of 3-day-old mice results, several months later, in organ-specific cell-mediated immune responses. The organs affected depend on the mouse strain; BALB/c mice predominantly develop an autoimmune gastritis (AIG) while other strains have been shown to develop oophritis (Taguchi et al., 1980), thyroiditis (Kojima et al., 1976), orchitis (Taguchi & Nishizuka, 1981), prostatitis (Taguchi, Kojima & Nishizuka, 1985) and pancreatitis (Bonomo, Kehn & Shevach, 1995). Development of disease is highly dependent on thymectomy at 2-4 days, since mice thymectomized at birth or at 7 days are unaffected. The data suggest that the numbers of T lymphocytes exported in the first 3 days of life are not sufficient to allow regulatory cells to prevent autoimmune responses. An alternative explanation is that autoreactive cells that initially escape deletion in the neonatal thymus can be deleted when they recirculate to the thymus, a phenomenon described elsewhere (Surh, Sprent & Webb, 1993). However, this explanation is not compatible with studies on adult thymectomized rats (described below). Direct evidence that disease induced by neonatal thymectomy results from ineffective regulation comes from studies in which AIG transferred to nude recipients from neonatally thymectomized mice can be controlled by co-transfer of splenocytes from normal syngeneic euthymic donors (Nishio et al., 1995). The protective population is contained in a subset of peripheral CD4+ T cells that express IL-2 receptor (Asano et al., 1996). These CD4+IL-2R+ cells appear in the first days of life in normal animals but their number in the periphery decreases after thymectomy. Under normal circumstances, these cells express *in vivo* mRNAs for IL-4, IL-10 and TGFβ.

Autoimmunity in high-dose irradiated mice

Total lymphoid irradiation of mice causes various organ-specific autoimmune diseases, such as gastritis, thyroiditis and orchitis, depending on the irradiation regimen and the mouse strain. Radiation-induced tissue damage does not seem to be the cause of disease. Irradiated animals reconstituted with thymocytes, splenocytes or bone marrow cells from normal syngeneic animals do not develop the autoimmune syndrome. CD4⁺ T cells from irradiated animals induce autoimmunity in syngeneic nude mice and CD4⁺ T cells from normal animals prevent the development of autoimmunity (Sakaguchi, Miyai & Sakaguchi, 1994). These results suggest that irradiation perturbs the balance between an autoreactive T cell population and a regulatory one that normally prevents autoimmunity.

Autoimmunity in adult thymectomized irradiated rats

Autoimmunity can be induced in normal PVG rats by a protocol of thymectomy at a few weeks of age followed by four doses of γ -irradiation, rendering the rats lymphopenic. A significant proportion of rats treated in this way developed thyroiditis, with mononuclear infiltrates of the thyroid gland and high titres of antithyroglobulin antibody in their sera (Penhale *et al.*, 1973). Disease could be prevented by reconstituting treatment of the rats following their last irradiation with lymph node cells or splenocytes from congenic donors (Penhale *et al.*, 1976). A small proportion of the thyroiditis-prone rats also developed insulin-dependent diabetes (Penhale *et al.*, 1990).

Development of diabetes in thymectomized and irradiated rats

The same protocol of adult thymectomy and γ -irradiation induces a very high incidence of diabetes in PVG.RT1^u rats. Three weeks after the final dose of irradiation, these animals start to develop the disease and by 12 weeks 70% of females and practically all male rats develop diabetes with selective destruction of the beta cells of the pancreas (Fowell & Mason, 1993). Disease development requires CD8⁺ cells, as is the case in the spontaneously diabetic NOD mouse (Bendelac *et al.*, 1987).

Reversal of the lymphopenia in prediabetic PVG.RT1^u rats by their reconstitution with unfractionated lymphocytes prevented disease in approximately 50% of recipients, and this protective capacity was shown to result entirely from the CD4⁺ cells. Significantly development of disease could be prevented in all recipients by their reconstitution with CD4⁺CD45RC^{low} cells (Fowell & Mason, 1993). The protective lymphocytes are a long-lived population of cells and not recent thymic emigrants, since prediabetic rats could also be protected by reconstitution with CD4+CD45RC^{low} cells from long-term thymectomized congenic donors. Further studies of phenotype revealed that the protective population was amongst CD4+CD45RC^{low} TCR $\alpha\beta$ +RT6+ Thy-1- cells, which is also the phenotype of a memory cell. CD4+CD45RC^{low} cells have been shown to be a potent source of IL-4 but not IFN- γ following activation and, therefore, resemble T_H2-like T cells (McKnight *et al.*, 1991). However, the importance of IL-4 in the protection of prediabetic rats from development of diabetes by CD4+CD45RC^{low} lymphocytes has yet to be determined.

These studies in adult thymectomized irradiated rats demonstrate firstly that normal animals contain cells with the potential to cause autoimmunity and, secondly, that these cells are prevented from fulfilling their pathological potential by regulatory T cells. However, maintenance of self-tolerance by regulatory T cells can be compromised either genetically or by disruption of the peripheral T cell pool, resulting in development of autoimmunity.

CD4+CD8- thymocytes are a potent source of cells that prevent development of autoimmune diabetes in adult thymectomized irradiated rats

Since the phenotype of the regulatory population of peripheral T cells is also the phenotype of a long-lived memory cell, it was assumed that this regulatory population was generated from naive precursors following encounter with an unknown antigen in the periphery. It was, therefore, a surprise to find that low doses of CD4+CD8- thymocytes purified from normal donors, a naive population of cells, could also prevent development of diabetes, albeit in only a proportion of recipients. Comparisons of the potency of peripheral cells and thymocytes reveal that the latter population is far more potent at preventing diabetes. Furthermore, the potency of thymocytes upon adoptive transfer appears to be restricted to their ability to control autoimmune responses. When assayed for their ability to provide primed B cells with help for secondary antibody responses upon adoptive transfer, CD4+CD8- thymocytes are far less able to support such a humoral response than either antigen-primed or unprimed peripheral CD4+ cells (Saoudi *et al.*, 1996).

Significantly, even at the highest dose of CD4+CD8- thymocytes used, only a proportion of rats are protected from development of diabetes. This inability to protect all recipients resembles the protection observed in prediabetic rats reconstituted with unfractionated CD4+ peripheral cells. In this instance, the incomplete protection was attributed to an antagonism of the CD45RC^{high} cells upon the protective CD45RC^{low} constituents of the inoculum. If this argument is applied to the protection by CD4+CD8- thymocytes, it implies that these cells, like peripheral CD4+ cells, are a functionally heterogeneous population containing both cells capable of promoting autoimmune responses and cells that regu-

late such responses. However, attempts to fractionate these cells into functionally distinct subsets phenotypically have so far been unsuccessful (Seddon *et al.*, 1996).

Conclusion

There is now strong support for the view that normal animals contain cells with the potential to cause autoimmunity and that these cells are under the control of regulatory T cells. The mechanism that underlies this phenomenon is mostly unknown and is the object of intense research. The $T_H 1/T_H 2$ paradigm does not satisfactorily account for the activity of regulatory cells since self-tolerance is associated with an absence of both cell-mediated ($T_H 1$) and humoral ($T_H 2$) autoreactive responses. It may be that the T cells that prevent autoimmunity in some of the systems described in this chapter produce a number of the cytokines characteristic of $T_H 2$ cells, but if $T_H 2$ cells are identified as those that promote humoral immune responses then regulatory T cells and $T_H 2$ cells are not identical populations.

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Complement receptors

J. M. AHEARN and A. M. ROSENGARD

Complement receptor type 1

The primary function of complement receptor (CR) type 1 (CR1, CD35) is phagocytosis and clearance of immune complexes, which is mediated by its capacity to bind complement ligands C3b, iC3b, C4b and C1q and its capacity to inactivate the C3 and C5 convertases of the alternative and classical complement pathways through decay-accelerating and cofactor functions (Fearon, 1979; Iidia & Nussenzweig, 1981; Medof *et al.*, 1982; Klickstein *et al.*, 1997). CR1 is expressed primarily by haematopoietic cells, including erythrocytes, mononuclear phagocytes, eosinophils, B lymphocytes and T lymphocytes. It is also present on glomerular podocytes and follicular dendritic cells (Fearon, 1980; Wilson, Tedder & Fearon, 1983; Gelfand, Frank & Green, 1975; Kazatchkine *et al.*, 1982; Reynes *et al.*, 1985).

Structurally, CR1 is a type I transmembrane glycoprotein (Fig. 12.1). Four CR1 allotypes have been identified (Dykman *et al.*, 1983a,b; 1985; Wong, Wilson & Fearon, 1983; Dykman, Hatch & Atkinson, 1984), with M_r under reducing/non-reducing conditions of ~250 000/190 000 (A, F), 290 000/220 000 (B, S), 210 000/160 000 (C, F') and >290 000/250 000 (D) (Wong & Fearon, 1987). All CR1 allotypes share the same transmembrane domain of 25 amino acid residues and a 43 residue cytoplasmic tail. All allotypes have extracytoplasmic domains composed entirely of structural motifs, referred to as short consensus repeats (SCR), or complement control protein (CCP) modules, arranged in tandem. The A(F) allotype comprises 2039 residues, including a 41 residue signal peptide and a 1930 residue extracellular domain composed entirely of 30 SCRs (Klickstein *et al.*, 1988; Hourcade *et al.*, 1988). The amino terminal 28 SCRs are arranged in four long homologous repeats (LHRs) (A–D) of seven SCRs each; two additional SCRs link LHR-D with a 25 residue transmembrane region and a 43 residue cytoplasmic domain (Klickstein *et al.*, 1988).



Fig. 12.1. Structure and function of complement receptor type 1 (CR1, CD35).

The other CR1 allotypes differ from one another structurally by an integral number of LHRs (Klickstein *et al.*, 1987). Allotypes A(F), B(S), and C(F') have extracytoplasmic domains that consist of four, five and three LHRs, respectively, as well as two additional SCRs that separate the last LHR from the transmembrane domain in each allotype. The D allotype has not yet been cloned and characterized, but it presumably consists of six LHRs. The frequencies of the A(F), B(S), C(F') and D allotypes are 0.82, 0.18, <0.01 and <0.01, respectively (Dykman *et al.*, 1984).

A single copy gene encoding CR1 maps to band q32 of chromosome 1, within the 750 kb regulators of complement activation (RCA) gene cluster, which also includes genes encoding CR type 2 (CR2, CD21), decay accelerating factor (DAF, CD55), membrane cofactor protein (MCP, CD46), factor H and C4-binding protein (Weis *et al.*, 1987; Rodriguez *et al.*, 1985; Lublin *et al.*, 1988; Bora *et al.*, 1989; Carroll *et al.*, 1988; Rey-Campos, Rubinstein & Rodriguez de Cordoba, 1988). The S allele of CR1 spans 158 kb and contains 47 exons (Vik & Wong, 1993; Wong *et al.*, 1989). The F allele, which does not contain the genomic region of the S allele that encodes LHR-B/A, spans 133 kb and is composed of 39 exons (Vik & Wong, 1993). The F' allele differs from the F allele in that it lacks LHR-B (Wong & Farrell, 1991).

The tandem alignment of 30 SCRs in the F allotype of CR1 creates a flexible, filamentous structure 80–90 nm in length, as determined by electron microscopic analysis (Weisman *et al.*, 1990).

Amino acid sequence homologies between SCRs that occupy the same relative positions in two different LHRs range from 56 to 100% identity (Klickstein *et al.*, 1987; 1988). The sequence of SCR-3 through SCR-7 of LHR-A differs from the sequence of SCR-10 through SCR-14 of LHR-B by only 1 of 327 residues. SCR-8 through SCR-11 of LHR-B and SCR-15 through SCR 18 of LHR-C differ at 3 of 253 positions.

There are 25 potential N-linked glycosylation sites in the F allotype (Klickstein *et al.*, 1988), of which six to eight appear to be modified by tri- and tetra-antennary complex-type oligosaccharides (Sim, 1985; Lublin, Griffith & Atkinson, 1986). Variable glycosylation is responsible for differences in CR1 size observed between erythrocytes and polymorphonuclear leukocytes in the same individual (Sim, 1985). A naturally occurring soluble form of CR1 has been demonstrated in serum and plasma of normal individuals (17.8–55.7 μ g/l) (Yoon & Fearon, 1985; Pascual *et al.*, 1993), and serum levels have been demonstrated to fluctuate widely in a variety of pathological states (Pascual *et al.*, 1993); however, it has not been determined whether levels correlate with *in vivo* complement activation. It has also not been determined whether this soluble form of the receptor is secreted or shed from plasma membranes. The ligand-binding domains of

CR1 have been mapped through deletion mutagenesis (Klickstein *et al.*, 1988), substitution mutagenesis (Krych, Hourcade & Atkinson, 1991; Krych *et al.*, 1994) and creation and characterization of chimaeric receptors (Kalli *et al.*, 1991b). Together, these studies have identified three distinct and independent ligand-binding domains within the F allotype of CR1, each of which is capable of binding both C3 and C4 fragments (Klickstein *et al.*, 1988). SCR-1 and SCR-2 of LHR-A form a primary C4-binding site and a secondary C3-binding site, and the amino terminal pair of SCRs within both LHR-B and LHR-C are primary C3-binding sites and secondary C4-binding sites. One report was unable to detect binding of C3b dimers to receptors containing SCR1–4 of LHR-A, suggesting that this secondary affinity may be considerably lower than that of the primary recognition sites (Makrides *et al.*, 1992). The nine SCRs common to all CR1 allotypes lack an intact C3/C4-binding domain.

Although the amino terminal pair of SCRs within LHR-A-C were initially shown to determine ligand specificity, it was subsequently recognized that the first four SCRs of an LHR are required to bind C3b with an affinity indistinguishable from that of the wild-type receptor (Kalli *et al.*, 1991b). The dissociation constants for polymeric C3b of wild-type CR1 and recombinant chimaeric receptors bearing LHR-BD and LRH-CD were 1.0–2.7 nM, and chimaeric receptors that contained SCRs 1–4, 1–3 or 2–4 of LHR-B or LHR-C had K_d of 1.8–2.4, 6–9 and 22–36 nM, respectively (Kalli *et al.*, 1991b; Seya, Holers & Atkinson, 1985).

The entire C4b ligand-binding domain of CR1 also consists of four SCRs. SCRs 1–4 of LHR-A bind C4b dimers with an affinity ($K_d \sim 4 \times 10^{-7}$ M) similar to wild-type receptor, whereas SCRs 1–2 alone bind C4b dimers with lower affinity ($K_d = 1.4 \times 10^{-6}$ M) (Reilly *et al.*, 1994). The first four SCRs of LHR-C (SCRs 15–18) are less effective in binding C4b dimers ($K_d = 1.2 \times 10^{-6}$ M) compared with the first four SCRs of LHR-A (SCRs 1–4).

Potential functional differences among the CR1 allotypes have been investigated through studies of soluble, recombinant forms of the F, S and F' allotypes (Wong & Farrell, 1991). All three soluble forms of the receptor were shown to be equally capable of binding monomeric C3b and serving as cofactors for factor-I-mediated conversion of C3b to iC3b and C3dg (Wong & Farrell, 1991). In contrast to these observations using monomeric C3b, the three variant forms of CR1 (which contain zero, one, and two LHR-B domains, respectively) differed considerably in their capacities to bind dimeric C3b (Wong & Farrell, 1991) and in their capacities to inhibit the alternative pathway C3 and C5 convertases, with the ABBCD and the ABCD forms being 30-fold more effective than the ACD form of soluble CR1 (Wong & Farrell, 1991). Together, these investigations have led to a model in which tandem ligand-binding domains contained within adjacent LHRs of a single CR1 molecule promote multivalent receptor-ligand interactions (Wong *et al.*, 1985).

Current efforts are focused upon identification of amino acids within these four SCR domains that determine ligand specificity. One strategy has involved substitution of residues from C3-binding sites for those in analogous positions of C4-binding sites and vice versa (Krych et al., 1991). The most interesting and informative results of these studies to date involved characterization of a mutant in which five amino acid residues at the carboxyl terminus of SCR-2 (DNET-PICD) were replaced by the five corresponding residues of SCR-9 (STKP-PICO). These substitutions resulted in the acquisition of iC3 binding without alteration of C4b binding. This is one of only two model systems in which the sequences of individual SCRs have been manipulated to result in a gain of function, and the results suggest that these residues are critical in determining the C3binding specificity of LHR-B and LHR-C. These amino acid positions in general may be critical in ligand discrimination. Similar studies have demonstrated the potential to enhance the ligand-binding capacity of CR1 beyond that of the endogenous receptor (Subramanian et al., 1996). In these investigations, transfer of four amino acids - glycine from SCR-1 and arginine, asparagine and tyrosine from SCR-2 - to corresponding locations in SCR-8 and SCR-9 created a ligandbinding domain with greater iC3- and C4b-binding activity than any wild-type site and enhanced cofactor activity for both ligands. Furthermore, replacement of serine-37 of SCR-1 with tyrosine from the homologous position of SCR-9 and replacement of glycine-79 of SCR-2 with aspartate from the corresponding position of SCR-16 resulted in acquisition of C3b binding by the C4b-binding site of CR1, whereas neither substitution alone had this effect.

Most recently, it has been reported that CR1 binds specifically to complement component C1q (Klickstein *et al.*, 1997). This interaction involves the collagenlike tail domain of C1q and LHR-D of CR1. Therefore, CR1 is capable of specifically binding to all opsonic ligands of the complement system, namely C3derived ligands, C4-derived ligands and C1q. The functional consequences of C1q binding to CR1 remain to be determined.

Complement receptor type 2

The type 2 receptor (CR2, CD21) is expressed on B lymphocytes and cell lines (Ross *et al.*, 1973; Eden, Miller & Nussenzweig, 1973; Bhan *et al.*, 1981; Nadler *et al.*, 1981; Iida, Nadler & Nussenzweig, 1983; Tedder, Clement & Cooper, 1984; Weis, Tedder & Fearon, 1984), some T cell lines (Fingeroth, Clabby & Strominger, 1988; Tsoukas & Lambris, 1993; Sinha *et al.*, 1993), thymocytes (Tsoukas & Lambris, 1988), peripheral blood T cells (Fischer, Delibrias &



Fig. 12.2. Structure and function of complement receptor type 2 (CR2, CD21).

Kazatchkine, 1991), and follicular dendritic cells (Reynes *et al.*, 1985; Liu *et al.*, 1997). Structurally, CR2 is a 140 kDa type I transmembrane glycoprotein (Fig. 12.2). Short (CD21_s) and long (CD21_L) forms of CR2 share the same 24 residue transmembrane region and 34 residue cytoplasmic tail; they differ only in their extracellular domains (Liu *et al.*, 1997; Weis *et al.*, 1986; 1988; Moore *et al.*, 1987). The extracellular domains are composed entirely of either 15 (CD21S) or 16 (CD21L) SCR tandem repeats of approximately 60 amino acid residues. Functionally, CR2 is a receptor for the C3b, iC3b and C3dg cleavage fragments of C3 (Ross *et al.*, 1973; Eden *et al.*, 1973; Ross & Polley, 1975; Lambris, Dobson & Ross, 1975; Weis *et al.*, 1984; Frade *et al.*, 1985b; Mold, Cooper & Nemerow, 1986), the B lymphocyte receptor for Epstein–Barr virus (EBV) (Jondal & Klein, 1973; Fingeroth *et al.*, 1984; Frade *et al.*, 1985a; Mold *et al.*, 1986) and a recep-

tor for CD23 (Aubry *et al.*, 1992; 1994; Bonnefoy *et al.*, 1993). Interferon α (Delcayre *et al.*, 1991) has also been reported to be a ligand for CR2.

A single copy gene encoding CR2 spans 35 kb (Yang, Behrens & Weis, 1991) within the 750 kb RCA gene cluster (see above). Restriction fragment length polymorphisms (RFLPs) of the gene have been identified for *TaqI* and *HaeIII* (Fujisaku *et al.*, 1989). Two *TaqI* allelic fragments have frequencies of 0.7 and 0.3 in normal Caucasians (Fujisaku *et al.*, 1989), and three *HaeIII* allelic fragments have frequencies of 0.93, 0.05, and 0.02 in normal Caucasians (Rey-Campos *et al.*, 1988).

Structurally, the short form of CR2 consists of a 20 amino acid residue signal peptide, a 954 residue extracellular domain organized into 15 SCRs of 57 to 74 residues each, a 24 residue transmembrane region and a 34 residue cytoplasmic tail (Weis et al., 1988). The long form of CR2 also contains a 16th SCR referred to as 10a because of its location between SCR-10 and SCR-11 of the 15 SCR form of CR2 (Weis et al., 1988). The 15 SCR (CD21,) and 16 SCR (CD21₁) forms of CR2 are products of alternative splicing (Toothaker, Henjes & Weis, 1989). Functionally, the 16th SCR (10a) does not participate in binding ligands iC3b or gp350/220 (Kalli, Ahearn & Fearon, 1991a). However, it has recently been demonstrated that follicular dendritic cells selectively express the 16 SCR form while B cells selectively express the 15 SCR form of CR2 (Liu et al., 1997), making CR2 the first human molecule specific to follicular dendritic cells to be characterized. The functional significance of this interesting and unexpected observation has not been determined. Hydrodynamic and electron microscopic studies of CR2 have demonstrated that the receptor is a highly extended, highly flexible molecule (Moore et al., 1989).

The cytoplasmic domain of CR2 contains four tyrosine, four serine and two threonine residues (Moore *et al.*, 1987; Weis *et al.*, 1988). CR2 is phosphorylated in tonsillar B cells and in Raji cells following exposure to PMA (Changelian & Fearon, 1986). Glycosylation is required for CR2 stability but not for the receptor to bind C3–Sepharose (Weis & Fearon, 1985). There are 11 potential N-linked glycosylation sites in the 15 SCR form of CR2 and two additional sites in SCR 10a (Weis *et al.*, 1988; Moore *et al.*, 1987). Of these sites, 8–11 bear oligo-saccharides on mature CR2 polypeptides in B lymphoblastoid cells (Weis & Fearon, 1985).

Soluble forms of CR2 have been identified in culture supernatants of Raji B lymphoblastoid cells, HPB-ALL acute leukemia T cells, and in normal human serum (Myones & Pross, 1987; Fremeaux-Bacchi *et al.*, 1996).

Four specific functions have been mapped to different structural domains of CR2. First, SCR-1 and SCR-2 are both necessary (Lowell *et al.*, 1989) and together sufficient (Lowell *et al.*, 1989; Carel *et al.*, 1990) for binding ligands

C3b, iC3b and C3dg. Second, SCR-1 and SCR-2 are both necessary (Lowell et al., 1989) and together sufficient (Lowell et al., 1989; Carel et al., 1990) for binding and EBV ligand gp350/220. Soluble recombinant full-length CR2 (Nemerow et al., 1990), as well as soluble, recombinant forms of the ligand-binding domain of CD21 (Hebell, Ahearn & Fearon, 1991; Moore et al., 1991) block binding of gp350/220 to the receptor and the infection of B cells by EBV. Consistent with these observations, it has been determined that SCR-1 and SCR-2 are both necessary (Lowell et al., 1989) and together sufficient (Lowell et al., 1989; Carel et al., 1990) for binding mAb OKB7 (Rao et al., 1985), which blocks binding of CR2 to C3d and to EBV (Nemerow et al., 1985). Anti-CR2 mAb HB-5, which does not block binding of either ligand to the receptor, binds to an epitope within SCR-3 and SCR-4. Third, CD23 (FceR2), a low-affinity receptor for IgE, binds to CD21 (Aubry et al., 1992); SCRs 5-8 of CR2 may be involved in this interaction, and N-linked oligosaccharides within this domain may participate (Aubry et al., 1994). This interaction between CR2 and CD23 may influence the survival of B cells in germinal centres. Fourth, CD21 and CD19 participate in a multimolecular B cell membrane complex (Matsumoto et al., 1991) in which CD21 interacts with CD19 on the B cell membrane via its transmembrane and extracellular domains (Matsumoto et al., 1993). CR2 also forms a bimolecular complex with CD35 on B lymphocytes (Tuveson et al., 1991) via the extracellular domains of the two receptors (Matsumoto et al., 1993).

Although SCR-1 and SCR-2 of CR2 contain the binding sites for natural ligands derived from C3, the viral ligand gp350 and the mAb OKB7, the requirements for binding C3dg versus gp350 versus OKB7 are actually distinct. Studies that led to this conclusion took advantage of the differential capacities of murine CD21 and human CD21 to bind EBV. Murine CR2 and human CR2 are both composed of 15 SCRs, and the ligand-binding domains of the two receptors (SCR-1 and SCR-2) share 61% amino acid identity (Molina et al., 1990); both receptors bind human C3dg (Molina et al., 1991). However, murine CD21 and human CD21 are functionally distinct in that only the human receptor is capable of binding EBV. Characterization of the ligand-binding capacities of a panel of 24 human-murine CD21 chimaeric receptors demonstrated that preferential binding of EBV to human CR2 occurs because of the distinct conformation of the human receptor, which can be achieved in murine CR2 with single amino acid substitutions in two discontinuous regions of the primary structure: replacement of proline at position 15 with the corresponding serine from human CR2, and elimination of a potential N-linked glycosylation site between SCR-1 and SCR-2 (Martin et al., 1991).

Finally, the role of CR2 in determining tropism of EBV for B lymphoblastoid cells appears to be limited to capture of virions at the cell surface, after which cofactors not associated with CR2-mediated postbinding events (Martin, Marlowe & Ahearn, 1994). This conclusion is based upon studies in which the ligand-binding domain of the rhinovirus receptor, ICAM-1 (CD54), was replaced with SCR-1 and SCR-2, the EBV gp350/220-binding domain of CR2. This CR2–ICAM-1 chimaeric receptor mediated EBV infection of three B lymphoblastoid cell lines, demonstrating that the extracytoplasmic, transmembrane and cytoplasmic non-ligand-binding domains of CR2 are not required for EBV infection of these B lymphoblastoid cells. This conclusion has not yet been extended to include primary B lymphocytes.

Complement receptor type 3

Complement receptor type 3 (CR3, CD11b/CD18, Mac-1, Mo1, OKM-1 α_m/β_2), an α/β heterodimeric adhesion molecule, is a member of the leukocyte-restricted integrin family (Springer *et al.*, 1979). CR3 shares a common 95 kDa β subunit with CD11a/CD18 (LFA-1, α_1/β_2) and CD11c/CD18 (CR4, p150, 95, α_x/β_2). It is the α subunits that distinguish the three heterodimers from one another. CR3 is expressed primarily by mononuclear phagocytes and natural killer (NK) cells, and expression of CR3 on activated granulocytes is considerably higher than on either of the other two leukocyte-restricted heterodimers. Expression of CR3 is greater than expression of CR4 in resting monocytes, whereas in tissue macrophages the opposite is true (Miller, Schwarting & Springer, 1986; Freyer *et al.*, 1988).

Functionally, CR3 is the most versatile complement receptor. Although iC3b is the only complement ligand for CR3 (Beller, Springer & Schreiber, 1982; Arnaout, Todd & Dana et al., 1983; Micklem & Sim, 1985), CR3 is also a receptor for coagulation factor X (Altieri & Edgington, 1988a), fibrinogen (Wright et al., 1988; Altieri et al., 1988; Trezzini et al., 1988), lipopolysaccharide (Wright & Jong, 1986), zymosan (Ross, Cain & Lachmann, 1985), soluble Fcy receptor type III (FcyRIII, CD16) (Galon et al., 1996), the counter-receptor ICAM-1 (CD54) (Smith et al., 1989; Diamond et al., 1990), filamentous haemagglutinin of Bordetella pertussis (Relman et al., 1990), Leishmania promastigote surface glycoprotein gp63 (Russell & Wright, 1988) and for the acute phase protein haptoglobin (El Ghmati et al., 1996). CR3 participates in a variety of cellular functions including adhesion of mononuclear phagocytes to endothelial cells (Anderson et al., 1986; Arnaout, Lanier & Faller, 1988b; Lo et al., 1989; Smith et al., 1989), cellular extravasation and chemotaxis to sites of inflammation (Smith et al., 1989), phagocytosis of CR3-ligand opsonized particles (Ezekowitz et al., 1983) and enhancement of NK cell activity for C3-coated targets (Ramos et al., 1988).

The single copy CD11b gene spans 55 kb on chromosome 16, band p11–p11.2 (Corbi *et al.*, 1988b; Arnaout *et al.*, 1988c), clustered with the other CD11 α subunits. Structurally, CR3 is a type I transmembrane glycoprotein with a 1 : 1 stoichiometry of α (CD11b) (165 000 M_r) and β (CD18) (95 000 M_r) subunits that associate non-covalently. Structurally, CD11b consists of a 16 residue signal peptide, a 1092 (or 1091) residue extracellular domain, a 26 residue transmembrane region and a 19 residue cytoplasmic tail (Arnaout *et al.*, 1988a; Corbi *et al.*, 1988a; Hickstein *et al.*, 1989). The one residue difference in the extracellular domain is a glutamine, located between two conserved cysteines at 478 and 489, which is of unknown functional significance. The single serine residue present in the cytoplasmic domain is constitutively phosphorylated in resting human leukocytes (Chatila, Geha & Arnaout, 1989; Buyon *et al.*, 1990).

The extracellular domain of CD11b contains seven homologous tandem repeats of ~60 residues. Repeats V through VII contain the consensus sequence DXDXDGXXDXXE, characteristic of EF-loop metal-binding proteins. Repeats I–IV lack this consensus sequence but contain the sequences YFGAS/AL and LVTVGAP, which are conserved in other integrins (Corbi *et al.*, 1988a). An I domain, consisting of 187 residues (150–338), inserted between repeats II and III, is homologous to the A domains of von Willebrand factor, factor B and C2.

There are 19 potential N-linked glycosylation sites in the CD11b sequence. The mature $\alpha_{\rm M}$ subunit of ~167 000 is partially sensitive to Endo H ($M_{\rm r}$ ~153 000) and completely susceptible to N-glycanase ($M_{\rm r}$ ~137 000) (Miller & Springer, 1987).

CD11b shares approximately 25% amino acid identity (Corbi *et al.*, 1988a) with the α subunits of platelet glycoprotein IIb/IIIa (Poncz *et al.*, 1987), vitronectin receptor (Suzuki *et al.*, 1987) and fibronectin receptor (Argraves *et al.*, 1987). CD11b is specifically distinguished structurally from the last three receptors by the absence of a cleavage site that is involved in proteolytic processing of the α subunits into light chains, which are membrane anchored, and heavy chains, which are entirely extracellular and linked by disulphide bond to the α chains. The amino acid sequences of the transmembrane domains of CD11b and CD11c are 88% identical (Corbi *et al.*, 1987; 1988a). This observation suggests that these hydrophobic sequences may serve an important functional role in addition to simply providing a membrane anchor for the proteins.

CD18 is a type I transmembrane protein consisting of a 22 residue signal peptide, an extracellular domain of 678 residues, a 23 residue transmembrane domain and a 46 residue cytoplasmic tail (Kishimoto *et al.*, 1987; Law *et al.*, 1987). The cytoplasmic domain contains one tyrosine, three threonine and four serine residues. Induction with either PMA or FMLP generates primarily phosphoserine, with small amounts of phosphothreonine and phosphotyrosine (Chatila *et al.*, 1989; Buyon *et al.*, 1990). The extracellular domain of CD18 contains six potential N-linked glycosylation sites and 56 cysteine residues. One cysteine-rich (20%) region, extending from residues 445 to 631, contains four tandem repeats of an eight cysteine residue motif that is thought to be involved in extensive intrachain disulphide bond formation (Kishimoto *et al.*, 1987).

Binding sites for the numerous CR3 ligands have been partially characterized as follows. ICAM-1 (CD54), a counter-receptor for both LFA-1 (CD11a/CD18) (Marlin & Springer, 1987) and CR3 (CD11b/CD18) (Diamond *et al.*, 1990), has an extracellular region that consists of five tandem immunoglobulin-like domains (Simmons, Makgoba & Seed, 1988; Staunton *et al.*, 1988), and CR3 binds to the third immunoglobulin-like domain (Diamond *et al.*, 1991). Competitive inhibition studies have suggested that the binding sites for fibrinogen, iC3b, factor X and gp63 on CR3 are overlapping, if not identical (Altieri & Edgington, 1988a,b; Wright *et al.*, 1988).

Homologous sequences in human C3 (LEICTRYRGD) (1386-1395), fibrinogen (LGGAKQAGD) (402-410), leishmania gp63 (367-376) and filamentous haemagglutinin of Bordetella pertussis (TVGRGDPHQ) (1094–1102) were initially thought to mediate these CR3 – ligand interactions, based upon the capacity of peptides containing these sequences to inhibit binding of the respective ligands to CR3 (Wright et al., 1987; 1988; Russell & Wright, 1988; Relman et al., 1990). However, subsequent studies suggested that these RGD peptides indirectly affect CR3 function by binding to leukocyte response integrin (LRI), a β_{2} integrin that is known to enhance the phagocytic capacity of polymorphonuclear cells (Brown & Goodwin, 1988; Gresham et al., 1989; Altieri et al., 1990; Taniguchi-Sidle & Isenman, 1992; van Strijp et al., 1993). CR3 appears to be expressed in an inactive state on resting leukocytes, and high-avidity ligand binding by CR3 occurs only following stimulation of cells via heterologous receptors such as LRI (Altieri & Edgington, 1988b; Buyon et al., 1988; Diamond & Springer, 1993; Stockl et al., 1995). Additional structure-function studies based upon CR3/CR4 chimaeric receptors have demonstrated that the I domain is a major recognition site for iC3b, ICAM-1 and fibrinogen (Diamond et al., 1993). In contrast, sCD16 is thought to bind outside the I domain, near the lectin-like region of CR3 (Galon et al., 1996). The binding site for lipopolysaccharide (Wright & Jong, 1986) is also distinct from the iC3b/fibrinogen/gp63-binding domain (Wright et al., 1989).

The capacity of CR3 to bind β glucan is controversial (van Strijp *et al.*, 1993); however, one study has suggested that CR3 is probably the only leukocyte β glucan receptor. CR3 has only one type of lectin site; this ligand-binding domain has broad sugar specificity, is cation independent and one or two such sites are located at the carboxy terminal area to the I domain (Thornton *et al.*, 1996). CR3 requires calcium or magnesium for ligand binding. A novel divalent cationbinding site has been identified in the A (I) domain of CR3, suggesting that the capacity of CD11b to bind iC3b, and perhaps other ligands, is dependent upon divalent cation binding to the A domain (Michishita, Videm & Arnaout, 1993). Determination of the secondary structure and the crystal structure of the A domain has confirmed and extended these observations (Perkins *et al.*, 1994; Lee *et al.*, 1995). Crystallographic studies have demonstrated that the A domain consists of one antiparallel and five parallel β strands that form a central sheet, surrounded by seven α helices, a classic 'Rossmann' fold (Lee *et al.*, 1995). A single metalbinding site, termed the metal ion-dependent adhesion site (MIDAS) motif (Lee *et al.*, 1995) sits at the top of the β sheet, on the surface of the A domain. The MIDAS consensus sequence is DXSXS (residues 140–144 in CD11b), and T²⁰⁹ and D²⁴² from discontinuous regions of the polypeptide.

Recent data suggest that CR3 may form a membrane complex with glycophosphatidylinositol (GPI)-anchored FcγRIIIB (CD16) on human granulocytes (Stockl *et al.*, 1995). The carboxy terminal domain (which contains 15 of the 19 potential N-linked glycosylation sites in CD11b) has been implicated in this interaction, and a lectin-like interaction between these two molecules has been proposed (Zhou *et al.*, 1993).

Complement receptor type 4

Complement receptor type 4 (CR4; CD11c/CD18; p150,95; α_x/β_2) α/β heterodimeric adhesion molecule is a member of the leukocyte-restricted integrin family (Sanchez-Madrid *et al.*, 1983; Lanier *et al.*, 1985; Springer, Miller & Anderson, 1986). CR4 shares a common 95 kDa β subunit with CD11a/CD18 (LFA α_1/β_2) and CD11b/CD18 (Mac-1, α_M/β_2), whereas the α subunit distinguishes the three heterodimers from one another. CR4 expression is restricted to leukocytes, including myeloid cells (Miller *et al.*, 1986), dendritic cells (Freudenthal & Steinman, 1990), NK cells (Werfel, Witter & Gotze, 1991), activated B lymphocytes (Postigo *et al.*, 1991), some cytotoxic T cells (Miller *et al.*, 1986; Keizer *et al.*, 1987) and platelets (Vik & Fearon, 1987). Functionally, iC3b is the primary complement ligand for CR4. Although it is not reported to be as versatile as CR3, CR4 also serves as a receptor for fibrinogen (Loike *et al.*, 1991), lipopolysaccharide (Wright & Jong, 1986), soluble FcγRIII (Galon *et al.*, 1996) and CD23 (Lecoanet-Henchoz *et al.*, 1995).

The single copy gene encoding CD11c is located on the short arm of chromosome 16, between 16p11 and 16p13.1, together with the genes encoding CD11a and CD11b (Corbi *et al.*, 1988b; Marlin *et al.*, 1986). The CD18 gene is located on chromosome 21, band q22.3 (Corbi *et al.*, 1988b; Marlin *et al.*, 1986). Structurally, CD11c is a type I transmembrane glycoprotein, consisting of a 1081 residue extracellular domain, a 26 residue transmembrane region and a 30 residue cytoplasmic tail (Corbi *et al.*, 1987). The extracellular domain contains three tandem repeated sequences of 58, 70 and 68 residues, respectively. Centred in each of the repeats is a putative divalent cation-binding motif that has been determined to form a helix-loop-helix conformation, referred to as an 'EF hand' (Kretsinger & Nockolds, 1973). In this configuration, the calcium-binding domain resembles a right hand with the thumb (helix f) and the forefinger (helix E) extended, and fingers three through five clenched. The middle finger represents the calcium-binding loop.

Residues 125 to 318 of CD11c form an I domain, as described above for CR3, and there are 10 N-X-S/T sites (Corbi *et al.*, 1987), of which five or six appear to be glycosylated (Miller & Springer, 1987).

CD11c shares 35 and 67% amino acid identity with CD11a and CD11b, respectively (Corbi *et al.*, 1987), and approximately 25% amino acid identity with the α subunits of platelet glycoprotein IIb/IIIa (Poncz *et al.*, 1987), vitronectin receptor (Suzuki *et al.*, 1987) and fibronectin receptor (Argraves *et al.*, 1987). As described above for CD11b, the structure of CD11c is distinguished from the structures of these three RGD receptors by the absence of the cleavage sites that are involved in the proteolytic processing of each of the other three integrin α subunits (into a light chain that is membrane anchored and a heavy chain that is entirely extracellular, linked by disulphide bond to the α chain).

The structure of CD18 has been described above for CR3. There is no homology between CD11c and CD18.

Functionally, CR4 shares with CD11a/CD18 and CD11b/CD18 the capacity to mediate adhesion of leukocytes to endothelium (Anderson *et al.*, 1986; Stacker & Springer, 1991). CR4 has a counter-receptor on human umbilical vein endothelial cells, and the capacity of mAbs that recognize different regions of CR4 to inhibit the interaction between leukocytes and endothelial cells suggests that more than one ligand mediates the interaction (Stacker & Springer, 1991).

The role of CR4 as a complement receptor has been somewhat controversial. Early studies suggested that CR4 had the capacity to bind iC3b (Micklem & Sim, 1985; Anderson *et al.*, 1986; Malhotra, Hogg & Sim, 1986; Myones *et al.*, 1988). CD11c/CD18 was actually identified via iC3b–Sepharose chromatography (Micklem & Sim, 1985). However, cells expressing recombinant CD11c/CD18 failed to bind iC3b (van Strijp *et al.*, 1993). Subsequent studies suggested that the I domain of the α subunit, CD11c, must adopt a particular conformation to acquire the capacity to bind iC3b (Bilsland, Diamond & Springer, 1994).

Soluble Fc γ RIII, a putative ligand for CR4, appears to bind outside of the I domain (Galon *et al.*, 1996). CR4 is also reported to serve as a monocyte

receptor for CD23 (Lecoanet-Henchoz et al., 1995), and ligation of CR4 activates neutrophil spreading and neutrophil respiratory burst (Berton et al., 1989).

Complement receptor C1qR

Activation of the classical pathway of complement begins with C1, which is a multimolecular complex in which one C1q subunit is complexed with two C1r and two C1s molecules (C1qC1r₂C1s₂). The single C1q subunit, comprising 18 polypeptides, has been described structurally as a bouquet of six 'tulips' in which each of the six stalks comprises three distinct polypeptide chains that are arranged in a collagen-like triple helix and in which each 'flower' is formed when the carboxyl terminal portions of these three associated strands expand into globular domains. The first stage of the classical pathway begins when the globular domain of C1q becomes bound to the Fc portion of an antibody that has attached to antigen. These antigen-antibody complexes that activate the classical pathway via Clq may be soluble, bound to the surface of cells or trapped in interstitial spaces. This activation process is tightly regulated to ensure that it is set into motion only under certain appropriate conditions. Once C1q has attached to an immune complex via its globular (head) domains, the collagen-like (tail) domains of C1q stimulate a variety of cellular responses through interaction with numerous cell types including mononuclear phagocytes, lymphocytes, fibroblasts and endothelial cells.

The first phagocytic cell surface protein conclusively shown to modulate C1qtriggered responses was C1qRp (Nepomuceno et al., 1997). The mature receptor consists of a 559 residue extracellular domain, a 25 residue transmembrane region and a 47 residue cytoplasmic tail that contains the sequence RAMENQY (638-644), which represents a tyrosine kinase recognition motif. The amino terminus of the extracellular region contains a 156 residue sequence that resembles a calcium-dependent carbohydrate recognition domain (CRD) (Nepomuceno et al., 1997; Drickamer, 1987; 1988). This domain contains the sequence FWIGLQREK, which is similar to sequences present in membrane receptors believed to modulate endocytosis, including the human mannose macrophage receptor (Nepomuceno et al., 1997; Ezekowitz et al., 1990). Carboxyl terminal of the CRD are five epidermal growth factor (EGF)-like domains (Cooke et al., 1987). Within each of the third, fourth and fifth EGF-like domains is an asparagine hydroxylation motif: D/N-X-D/N-Q/E-C-X₄₋₇-C-X₃-C-X-D*/N*-X₄-Y/F, in which the asterisks represent hydroxylated residues. Similar EGF-like domains are present in a variety of extracellular proteins, such as the vitamin Kdependent plasma proteins factors IX and X, protein C and protein S, the connective tissue component fibrillin-1, and the complement proteins C1r and C1s (Rees *et al.*, 1988). X-ray crystallographic studies of a calcium-binding EGF-like domain from factor IX have suggested that calcium bound to EGF-like domains functions both to stabilize the structure of the domain and to mediate protein–protein interactions directly. These interactions can be between EGF-like domains or between EGF domains and other domain types, which many be diverse among this structurally related but functionally diverse group of proteins (Rao *et al.*, 1995). Based upon these data, the multiple tandem EGF-like domains of C1qRp are unlikely to function independently of one another (Rao *et al.*, 1995).

Modification of the single potential N-linked glycosylation site at position 325 as well as extensive O-linked glycosylation appear to be responsible for the discrepancy between the predicted molecular mass (66 495 Da) of C1qR_p and the apparent molecular mass of the receptor as determined by SDS–PAGE (126 000 Da) (Nepomuceno *et al.*, 1997). C1qR_p is a recently characterized membrane receptor with an extracellular domain characterized by a calcium-dependent carbohydrate recognition domain, five EGF domains, of which three are likely to bind calcium, and a cytoplasmic domain that contains a tyrosine phosphorylation motif. Demonstration of the capacity of C1qR_p to enhance phagocytosis following cellular capture of C1q-containing complexes, in addition to the recent demonstration of the capacity of CR1 to bind C1q (Klickstein *et al.*, 1997), will undoubtedly generate additional important and unanticipated insights into multiple inflammatory processes.

Participation of the complement system in rheumatic diseases

The complement system plays a fundamental role during inflammatory and innate immune responses. As such, complement ligands and complement receptors are likely to participate in the pathogenesis of most, if not all, rheumatic diseases. For example, complement activation has been demonstrated in arthritides, immune complex vasculitides, glomerulonephritides, demyelinating disorders, coagulopathies, immune thrombocytopenias and cutaneous inflammation manifested by a variety of rashes. The role of complement in the pathogenesis of specific rheumatic disease has been preliminarily investigated in disorders such as RA, dermatomyositis, systemic lupus erythematosus (SLE), Henoch-Schonlein purpura, Goodpasture's syndrome, idiopathic thrombocytopenic purpura, autoimmune cutaneous diseases such as pemphigus and bullous pemphigoid, and multiple sclerosis. For most of these conditions, studies have focused on determining whether the complement system is activated during the disease process and which complement ligands are deposited in affected tissues. In general, these studies have indicated that the fundamental molecular mechanisms responsible for initiating rheumatic diseases do not involve the complement system. Rather,

complement activation appears to participate downstream of causative molecular events. Therefore, the complement system serves to amplify inflammation, coagulation and immune responses in these diseases, and thereby contribute to disease pathogenesis. A detailed review of these studies is beyond the scope of this chapter, and recent excellent reviews of this subject are available (e.g. Holers, 1996).

Although complement activation appears to play a secondary role in the pathogenesis of most rheumatic diseases, there is one exception. The complement system serves a critical role in maintaining immune tolerance and protecting against the development of SLE. Specifically, complement ligands generated by the classical pathway of complement, and CR1 (CD35), appear to protect against the development of SLE. This observation is the most intriguing relationship between complement receptors and rheumatic diseases described to date. Clues to the molecular mechanism(s) by which the complement system protects against the development of SLE will be the focus of the remainder of this chapter.

The clinical phenotype of SLE is generally considered to represent several diseases that result from multiple genetic factors interacting with one or more environmental agents. Some of the genetic risk factors that have been defined influence the disease phenotype rather than actually cause the disease. For example, class II alleles of the MHC have been shown to be closely associated with production of specific autoantibodies by patients with SLE, but these loci do not appear to actually increase the risk of developing lupus itself (Arnett, 1994). In contrast, studies of monozygotic twins, in which the concordance rate for SLE ranges between 24% and 69%, suggest that there are hereditary factors that are actually causative (Arnett & Reveille, 1992). The strongest genetic risk factor for the development of human SLE is complete deficiency of one of the components of the classical pathway of complement (C1q, C1r, C1s, C4 and C2) (Nishino et al., 1981; Meyer et al., 1985; Agnello, 1986; Arnett & Moulds, 1991; Colten & Rosen, 1992; Suzuki et al., 1992; Bowness et al., 1994; Morgan & Walport, 1994). In fact, an individual with complete deficiency of one of these proteins is nearly guaranteed to develop SLE. For example, of 32 patients with homozygous genetic deficiency of C1q reported in the literature, 30 have developed SLE, and one has developed discoid lupus (Shibuya & Nishida et al., 1981; Suxuki et al., 1992; Bowness et al., 1994; Nishino et al., 1981; Slingsby et al., 1996). An individual with C1q deficiency has a greater risk of developing lupus than an individual who has a monozygotic twin affected by the disease. This suggests that in those patients with complement-deficient lupus, no other genetic factor is involved. This is supported by the nearly complete penetrance of the defective gene together with the observation that such individuals have been reported throughout the world from different ethnic and genetic backgrounds. There is no reason to suspect other shared genetic defects among this group of patients. Therefore, although the aetiology and immunopathogenesis of SLE in general is most likely multifactorial, lupus that results from complement deficiency is most likely a single disease. It either results from the single genetic defect alone or a combination with one or more relatively ubiquitous environment agents such as ultraviolet light or a common virus. The classical pathway of complement plays a fundamental role during immune complex solubilization and clearance, primarily via CR1. Therefore, it has been generally presumed that complete deficiency of one of the classical pathway components causes lupus because of an impaired capacity to clear aggregates of antibody and antigen. However, this hypothesis has not been proved and there are surprisingly few data to support it. Alternatively, there are several reasons for suspecting that complement deficiency causes lupus through a molecular mechanism other than failure to clear antigen–antibody complexes.

First, patients who develop SLE as a result of complement deficiency have a relatively homogeneous, characteristic and specific syndrome. The nearly invariant feature of the disease is a severe photosensitive skin rash, which is frequently accompanied by high titres of anti-Ro(SSA) antibody (Meyer *et al.*, 1985). Patients tend to develop the disease at a younger age than other forms of lupus, and there is no female predominance, again supporting the idea that no other genetic factors are involved. There is no reason to believe that a general failure to clear immune complexes should lead to development of a consistent and specific disease phenotype compared with any other autoimmune disease. Why do patients with complement deficiency usually develop lupus with cutaneous photosensitivity and anti-Ro antibody instead of RA, Sjögren's syndrome, etc.?

Second, patients who develop SLE as a result of complement deficiency do not have greater difficulties with immune complex deposition in tissues as compared with SLE patients in general. If anything, this is less a feature of their clinical syndrome. The 'lupus band test', which many consider to represent immune complex deposition in the skin, is often negative in these patients.

Third, most antigen–antibody complexes can be activated and cleared by the alternative pathway, which is intact in all of these patients.

Fourth, patients with complete deficiency of C2 develop lupus even though they are capable of activating the classical pathway and coating circulating antigen–antibody complexes with C4b, a ligand that is readily recognized and cleared by CR1-bearing erythrocytes.

Fifth, numerous non-immunoglobulin activators of C1q *in vitro* have been described, including cardiolipin, DNA, RNA, cytoskeletal intermediate filaments, decorin, phosphatidylglycerol and C-reactive protein (Cooper, 1985; Krumdieck *et al.*, 1992). Most of these have a relatively high density of negative

charges carried by phosphate or sulphate groups, and there are some suggestions that an appropriate mix of anions and cations will activate C1q. Although the *in vivo* relevance of these interactions has not been elucidated, these observations suggest that the role of C1q *in vivo* may not be limited to interaction with the Fc domain of specific immunoglobulin isotypes. Primarily for these reasons, alternative explanations for the molecular immunopathogenesis of complement-deficient lupus are being explored. One alternative hypothesis with supporting evidence will be presented here.

As discussed above, a common feature of SLE in patients with normal complement levels, but particularly in those with complement deficiency, is cutaneous photosensitivity. Recent observations suggest that the skin may not only be a primary immune target in SLE but it may also be the site at which tolerance is initially broken. Keratinocytes exposed to ultraviolet light undergo apoptosis, a programmed form of cell death in which cytoplasmic and nuclear condensation and blebbing are prominent features. Cytoplasmic and nuclear blebs that form on the surfaces of apoptotic cells have recently been shown to contain several autoantigens that are common targets of autoantibodies in lupus, including DNA, Ro, La and snRNP (Casciola-Rosen, Anhalt & Rosen, 1994). These packages not only contain higher concentrations of autoantigens that might be encountered elsewhere in vivo, but there is also evidence that the self-antigens within the blebs may be cleaved by intracellular proteases specific to the apoptotic process, thereby revealing cryptic epitopes (Casciola-Rosen, Anhalt & Rosen, 1995; Casiano et al., 1996). In addition, it has been demonstrated that when apoptosis of human keratinocytes is induced by virus infection, viral antigens and autoantigens cocluster in specific subsets within surface blebs and may present a novel challenge to self-tolerance if not cleared and processed properly (Rosen, Casciola-Rosen & Ahearn, 1995). It has also been demonstrated that C1q, the first component of the classical complement pathway, specifically and directly binds to blebs generated from human keratinocytes (Korb & Ahearn, 1997). This capacity to bind C1q appears to occur in focal patches on the plasma membrane early during the course of apoptosis, and it does not appear to involve nuclear constituents. Later during the course of apoptosis, these membrane patches surround packages of cytoplasm, and eventually these blebs are released. In vivo, these blebs are likely to consist of high concentrations of self-antigen that may be packaged with viral antigen or may bear cryptic epitopes generated by novel apoptotic proteases. C1q and the classical pathway may be required for proper clearance of these blebs from the skin to maintain immune tolerance (Fig. 12.3). A complete deficiency of C1q may lead to immunization with autoantigens of cutaneous origin. This scenario may also explain the observation that individuals with lower levels of CR1 expression are at greater risk for developing SLE. As described earlier in



Fig. 12.3. Opsonization of apoptotic blebs by complement ligands. Shown is a cell undergoing apoptosis as a result of exposure to ultraviolet light (UV) or virus (V). Apoptosis leads to generation of nuclear (solid circles) and cytoplasmic (clear circles) blebs. One subset of blebs contains self-antigen (S). One subset of blebs contains self-antigen (S). One subset of blebs contains self-antigen that has been modified to reveal immunocryptic epitopes, as might occur from proteolysis (S*). One subset of blebs contains self in a novel context, as might occur following viral infection and co-clustering of self-antigen and viral antigen (SV). Direct binding of C1q to the bleb surfaces may be the initial event in a molecular pathway by which the classical pathway of complement and receptors such as CR1 or C1qR_p maintain immune tolerance.

this chapter, CR1 has traditionally been recognized as a primary receptor for C3- and C4-derived ligands. The recent demonstration that CR1 also serves as a receptor for C1q suggests that CR1 can bind all opsonic ligands generated by the classical pathway of complement (Klickstein *et al.*, 1997). It remains to be determined if CR1 may participate in the clearance of apoptotic blebs derived from keratinocytes or other tissue sources.

Other complement receptors, such as CR2, CR3, CR4, and C1qR must also be considered as potential participants in a molecular mechanism responsible for clearance of apoptotic blebs containing autoimmunogens. Recent observations that complement ligands and receptors can influence cytokine production and $T_H 1-T_H 2$ polarization (Karp, Wysocka & Wahl, 1996) have provided further incentive to revisit the relationship between the complement-deficient state and the development of SLE.

In summary, studies of the participation of the complement system in rheumatic diseases have primarily focused on secondary events that lead to deposition of complement ligands in affected tissues and organs. The one exception to this pattern has involved investigation of the role of complement deficiency in the aetiology and pathogenesis of SLE. Although it has been presumed that this is associated with an impaired capacity to clear immune complexes, evidence in support of this hypothesis has not accumulated and clues to alternative explanations have mounted. The complexity of the complement system suggests that many of its functions remain to be discovered. In parallel, the role of complement receptors in the aetiology and pathogenesis of rheumatic diseases is just beginning to surface.

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