



GUIDELINES FOR THE

HUMANE

TRANSPORTATION

OF RESEARCH ANIMALS



NATIONAL RESEARCH COUNCIL
OF THE NATIONAL ACADEMIES

GUIDELINES FOR THE **HUMANE** **TRANSPORTATION** **OF RESEARCH ANIMALS**

Committee on Guidelines for the
Humane Transportation of Laboratory Animals

Institute for Laboratory Animal Research

Division on Earth and Life Studies

NATIONAL RESEARCH COUNCIL
OF THE NATIONAL ACADEMIES

THE NATIONAL ACADEMIES PRESS
Washington, D.C.
www.nap.edu

THE NATIONAL ACADEMIES PRESS, 500 Fifth Street, N.W., Washington, DC 20001

NOTICE: The project that is the subject of this report was approved by the Governing Board of the National Research Council, whose members are drawn from the councils of the National Academy of Sciences, the National Academy of Engineering, and the Institute of Medicine. The members of the committee responsible for the report were chosen for their special competences and with regard for appropriate balance.

This study was supported by the Elizabeth R. Griffin Research Foundation, the National Center for Infectious Disease, and Contract No. N01-OD-4-2139, Task Order 118 between the National Institutes of Health and the National Academy of Sciences. Any opinions, findings, conclusions, or recommendations expressed in this publication are those of the authors and do not necessarily reflect the views of the organizations or agencies that provided support for the project. The content of this publication does not necessarily reflect the views or policies of the National Institutes of Health, nor does mention of trade names, commercial products, or organizations imply endorsement by the US government.

Library of Congress Cataloging-in-Publication Data

Guidelines for the humane transportation of research animals /
Committee on Guidelines for the Humane Transportation of
Laboratory Animals, Institute for Laboratory Animal Research,
Division on Earth and Life Studies.

p. ; cm.

Includes bibliographical references and index.

ISBN 0-309-10110-7 (pbk.)

1. Laboratory animals—Transportation. I. Institute for Laboratory
Animal Research (U.S.). Committee on Guidelines for the Humane
Transportation of Laboratory Animals.

[DNLM: 1. Animals, Laboratory—Guideline. 2. Transportation
—standards—Guideline. 3. Animal Welfare—standards—Guideline.

4. Laboratory Animal Science—standards—Guideline. 5. Safety
Management—standards—Guideline. QY 52 G946 2006]

SF406.7.G85 2006

636.088'5—dc22

2006010872

ISBN 0-309-65724-5 (PDF)

Additional copies of this report are available from the National Academies Press, 500 Fifth Street, NW, Lockbox 285, Washington, DC 20001; (800) 624-6242 or (202) 334-3313 (in the Washington metropolitan area); Internet, <http://www.nap.edu>

Disclaimer:

The Internet information and government forms referenced in this report were correct, to the best of our knowledge, at the time of publication. It is important to remember, however, the dynamic nature of the Internet. Resources that are free and publicly available one day may require a fee or restrict access the next, and the location of items may change as menus and homepages are reorganized.

Copyright 2006 by the National Academy of Sciences. All rights reserved.

Printed in the United States of America

THE NATIONAL ACADEMIES

Advisers to the Nation on Science, Engineering, and Medicine

The **National Academy of Sciences** is a private, nonprofit, self-perpetuating society of distinguished scholars engaged in scientific and engineering research, dedicated to the furtherance of science and technology and to their use for the general welfare. Upon the authority of the charter granted to it by the Congress in 1863, the Academy has a mandate that requires it to advise the federal government on scientific and technical matters. Dr. Ralph J. Cicerone is president of the National Academy of Sciences.

The **National Academy of Engineering** was established in 1964, under the charter of the National Academy of Sciences, as a parallel organization of outstanding engineers. It is autonomous in its administration and in the selection of its members, sharing with the National Academy of Sciences the responsibility for advising the federal government. The National Academy of Engineering also sponsors engineering programs aimed at meeting national needs, encourages education and research, and recognizes the superior achievements of engineers. Dr. Wm. A. Wulf is president of the National Academy of Engineering.

The **Institute of Medicine** was established in 1970 by the National Academy of Sciences to secure the services of eminent members of appropriate professions in the examination of policy matters pertaining to the health of the public. The Institute acts under the responsibility given to the National Academy of Sciences by its congressional charter to be an adviser to the federal government and, upon its own initiative, to identify issues of medical care, research, and education. Dr. Harvey V. Fineberg is president of the Institute of Medicine.

The **National Research Council** was organized by the National Academy of Sciences in 1916 to associate the broad community of science and technology with the Academy's purposes of furthering knowledge and advising the federal government. Functioning in accordance with general policies determined by the Academy, the Council has become the principal operating agency of both the National Academy of Sciences and the National Academy of Engineering in providing services to the government, the public, and the scientific and engineering communities. The Council is administered jointly by both Academies and the Institute of Medicine. Dr. Ralph J. Cicerone and Dr. Wm. A. Wulf are chair and vice chair, respectively, of the National Research Council.

www.national-academies.org

This report is respectfully dedicated to the memory of
CHARLES KEAN
March 25, 1942 – June 25, 2004
who dedicated his life to the care of humans and animals alike

**COMMITTEE ON GUIDELINES FOR THE HUMANE
TRANSPORTATION OF LABORATORY ANIMALS**

Ransom L. Baldwin (*Chair*), University of California, Davis, California

Chandra R. Bhat, University of Texas, Austin, Texas

Donald H. Bouyer, University of Texas, Galveston, Texas

Firdaus S. Dhabhar, Stanford University School of Medicine, Stanford,
California

Steven L. Leary, Washington University School of Medicine, St. Louis,
Missouri

John J. McGlone, Texas Tech University, Lubbock, Texas

Eric Raemdonck, International Air Transport Association, Montreal,
Quebec, Canada

Jennie L. Smith, Yale University, New Haven, Connecticut

Janice C. Swanson, Kansas State University, Manhattan, Kansas

Staff

Jennifer Obernier, Study Director

Marsha Barrett, Project Assistant

Kathleen Beil, Administrative Assistant

Kori Brabham, Intern

Norman Grossblatt, Senior Editor

Johnny Hernandez, Intern

John Horigan, Fellow

Susan Vaupel, Editor

INSTITUTE FOR LABORATORY ANIMAL RESEARCH COUNCIL

- Stephen W. Barthold** (*Chair*), University of California, Center for Comparative Medicine, Davis, California
- William C. Campbell**, Drew University, Madison, New Jersey
- Jeffrey I. Everitt**, GlaxoSmithKline Research and Development, Comparative Medicine and Investigator Support, Research Triangle Park, North Carolina
- Michael F. Festing**, Leicestershire, United Kingdom
- James G. Fox**, Massachusetts Institute of Technology, Division of Comparative Medicine, Cambridge, Massachusetts
- Estelle B. Gauda**, Johns Hopkins University School of Medicine, Johns Hopkins Hospital, Baltimore, Maryland
- Janet Gonder Garber**, Pinehurst, North Carolina
- Coenraad F.M. Hendriksen**, Netherlands Vaccine Institute, Bilthoven, The Netherlands
- Jon H. Kaas**, Vanderbilt University, Nashville, Tennessee
- Jay R. Kaplan**, Wake Forest University School of Medicine, Department of Pathology, Winston-Salem, North Carolina
- Joseph W. Kemnitz**, University of Wisconsin, Primate Research Center, Madison, Wisconsin
- Leticia V. Medina**, Abbott Laboratories, Abbott Park, Illinois
- Abigail L. Smith**, University of Pennsylvania, University Laboratory Animal Resources, Philadelphia, Pennsylvania
- Stephen A. Smith**, Virginia Polytechnic Institute and State University, Department of Biomedical Sciences and Pathobiology, Blacksburg, Virginia
- Peter Theran**, Massachusetts Society for the Prevention of Cruelty to Animals, Angell Animal Medical Center, Boston, Massachusetts

Staff

- Joanne Zurlo**, Director
- Kathleen Beil**, Administrative Assistant

Preface

This project was initiated in response to a letter from Charles Kean, an Associate Professor of Physiology and Pharmacology and Director of the Animal Care Facility at Loma Linda University, to the National Research Council's (NRC's) Institute for Laboratory Animal Research (ILAR), the Association for Assessment and Accreditation of Laboratory Animal Care (AAALAC) International, and the Office of Laboratory Animal Welfare (OLAW) of the National Institutes of Health (NIH) outlining the research animal care community's concerns about the safe and humane transportation of research animals. Dr. Kean requested that those organizations look into the transportation of research animals and into issues that were adversely affecting animal welfare. In response, ILAR hosted a meeting of various stakeholders to identify and discuss important issues in the transportation of research animals. The meeting was funded by NIH and included representatives of the scientific community, professional veterinary organizations, regulatory and accrediting agencies, animal breeders, and the transportation industry. Special thanks are due to the following for participating in the meeting, which took place December 4, 2001:

Kathryn Bayne, AAALAC International

Frank Black, Air Transportation Association of America, Inc.

Ralph Dell, ILAR

Nelson Garnett, OLAW

James Geistfeld, Taconic Farms, Inc.

Charles Kean, Loma Linda University

Carl Kole, United Airlines

J. Michael Krop, US Postal Service

Steven Leary, Washington University

Emilie Rissman, University of Virginia

Robert Russell, Harlan Sprague Dawley, Inc.

James Taylor, Office of Animal Care and Use, NIH

Richard Watkins, US Department of Agriculture (USDA) Animal and
Plant Health Inspection Service

William White, Charles River Laboratories

The meeting delineated the problems encountered during and resulting from air and ground transportation of live animals. The participants also focused on mechanisms to solve the problems, including the potential for a future ILAR study. As a result of this meeting, the Elizabeth R. Griffin Research Foundation, NIH, and the National Center for Infectious Diseases sponsored an ILAR committee to address problems associated with transportation of research animals and produce a report that includes recommendations intended for government agencies as well as for individual investigators/animal facility managers who may need to ship animals in the future.

Transportation of research animals may raise concerns related to the well-being of the animals and concerns about how animals are affected by general environmental conditions. These concerns often depend on the species being transported. Shipments from breeders to research institutions are generally well executed through the use of company-owned fleets of environmentally controlled vehicles, but arranging transport from vendors without established transport systems, or between research institutions, can be challenging. Animals may be shipped in vehicles without controlled environments and could be subjected to extreme temperatures. Specific requests for temperature-controlled vehicles may not be honored because the shipper may not have temperature-controlled vehicles available or the request may not have been passed on to a subcontractor hired by the shipper to transport the animals. The USDA has regulatory jurisdiction and inspection authority over transportation of animals through the Animal Welfare Act. However, most animals shipped are rats and mice, which are not covered under the act. The Public Health Service, whose oversight does include those species, does not inspect research animal transportation activities unless a complaint is filed.

The major problem in transporting nonhuman primates is that few airlines are willing to carry the animals. International shipment, the most common transportation of nonhuman primates, is often delayed by a cumbersome, multiagency permitting process involving the USDA Vet-

erinary Service, US Fish and Wildlife Service, the Centers for Disease Control and Prevention (CDC), and the Department of Transportation (DOT). Airlines have little incentive to carry the animals because it is not profitable and workers must wear protective clothing when handling them. The latter is disturbing both for workers and for travelers who see them. Finally, many animal rights activists have successfully lobbied the airlines to stop transporting nonhuman primates nationally and internationally.

Transportation of research animals is an essential component of the research enterprise. The integrity and well-being of the animals being transported are necessary for the quality of the research and the welfare of the animals. The lack of clear guidelines that cover all species can cause confusion for individuals without extensive experience in arranging transportation for research animals. In addition, investigators may find it difficult to identify a responsible shipper that will arrange for appropriate caging, inclusion of food and water, and other animal needs during transportation.

In the aftermath of the bioterror incidents involving anthrax in the fall of 2001, the possibility that research animals will be used to carry or disseminate bioterrorism agents must be considered. Breaches in good transportation practices, either purposeful or accidental, could result in the spread of infectious agents. In addition, new legislation (such as the Animal Health Protection Act of 2002) and several guidelines related to homeland security have the potential to complicate the importing, exporting, and transportation of animals and specimens for biomedical research.

The issues identified in the preceding statements led to appointment of the ILAR Committee on Guidelines for the Humane Transportation of Laboratory Animals. The committee held three meetings—in April, September, and December 2004. During the course of its deliberations, the committee sought assistance from many people, who gave generously of their time to provide valuable advice and information that were used in its deliberations. Special thanks are due to the following:

Richard Phelan, Taconic Farms, Inc.

Bonnie P. Dalton, Science Directorate, Ames Research Center,
National Aeronautics and Space Administration

Gale Galland, Division of Global Migration and Quarantine, National
Center for Infectious Diseases

Frank Kohn, FWS

John Monetti, World Courier

Erik Liebegott, Transportech, LLC

Robert Fernandez, Direct Services

William White, Charles River Laboratories

Carol Wigglesworth, OLAW
Bobby Brown, CDC
Carl Kole, Special Cargos, United Airlines
Charles Kean, Animal Research Facility, Loma Linda University
Barbara Kohn, Office of Animal Care, USDA
Eileen Edmonson, Office of Hazardous Materials Safety, DOT

The report has been reviewed in draft form by individuals chosen for their diverse perspective and technical expertise, in accordance with procedures approved by the NRC's Report Review Committee. The purpose of this independent review is to provide candid and critical comments that will assist the institution in making its published report as sound as possible and to ensure that the report meets institutional standards for objectivity, evidence, and responsiveness to the study charge. The review comments and draft manuscript remain confidential to protect the integrity of the deliberative process. We wish to thank the following individuals for their review of the report:

Susan Eicher, Purdue University, West Lafayette, IN
Steven Griffey, University of California, Davis, CA
Kathleen Hancock, Virginia Polytechnic University, Alexandria, VA
Barbara Hansen, All Children's Hospital, St. Petersburg, FL
Donald Lay, Purdue University, West Lafayette, IN
Tim Morris, GlaxoSmithKline, United Kingdom
William Morton, Paris NHP, Edmonds, WA
Barbara Orlans, Georgetown University, Washington, DC
Frankie Trull, National Association for Biomedical Research, Washington, DC
William White, Charles River Laboratories, Wilmington, MA
Walter Woolf, Air Animal, Tampa, FL

Although the reviewers listed above provided many constructive comments and suggestions, they were not asked to endorse the conclusions or recommendations, nor did they see the final draft of the report before its release. The review of the report was overseen by:

Johanna Dwyer, Tufts University, Boston, MA
Steven Pakes, University of Texas Southwestern Medical Center and VA North Texas Health Care System, Dallas, TX

Appointed by the NRC, they were responsible for making certain that an independent examination of the report was carried out in accordance

with institutional procedures and that all review comments were carefully considered. Responsibility for the final content of the report rests entirely with the authoring committee and the institution.

Ransom L. Baldwin, Chair
Committee on Guidelines for the
Humane Transportation of
Laboratory Animals

Contents

TABLES AND FIGURES	xvii
ABBREVIATIONS AND ACRONYMS	xix
SUMMARY	1
Major Recommendations, 3	
1 INTRODUCTION	7
2 REGULATIONS AND GUIDELINES FOR THE TRANSPORTATION OF RESEARCH ANIMALS	11
National Regulations and Guidelines, 11	
Department of Agriculture, 14	
US Fish and Wildlife Service, 14	
Centers for Disease Control and Prevention, 18	
US Department of Transportation, 20	
Public Health Service, 21	
Food and Drug Administration, 21	
State Health and Agricultural Regulations, 22	
International Regulations for Transporting Research Animals, 22	
CITES, 23	
International Civil Aviation Organization, 25	
International Air Transport Association, 27	
World Animal Health Organization, 28	
The European Union, 29	

3	GOOD PRACTICES IN THE TRANSPORTATION OF RESEARCH ANIMALS	33
	Stress During Transportation, 34	
	Allometric Scaling and Implication for Transportation Practices, 38	
	Thermal Environment, 39	
	Space Allocation, 52	
	Food and Water, 54	
	Social Interaction and Group Transportation, 59	
	Handling, 59	
	Monitoring Transportation, 60	
	Emergency Procedures, 61	
	Personnel Training, 61	
4	BIOSECURITY	65
	Protecting Public Health and Agricultural Resources, 66	
	Protecting the Biological Integrity of Research Animals and Colonies, 71	
5	RECOMMENDATIONS	81
	REFERENCES	89
	APPENDIXES	
A	SUMMARY OF THE ANIMAL WELFARE ACT REGULATIONS PERTAINING TO TRANSPORTATION	97
	Dogs and Cats (9 CFR 3.13 – 3.19), 97	
	Nonhuman Primates (9 CFR 3.86-3.92), 104	
	Guinea Pigs and Hamsters, Rabbits, and Other Animals (9 CFR 3.35-3.41, 9 CFR 3.60-3.65, 9 CFR 3.136 – 3.142), 110	
B	PATTERNS IN THE GROUND TRANSPORTATION OF RESEARCH ANIMALS IN THE UNITED STATES	117
	Background, 117	
	Data Preparation, 118	
	Quantitative Analysis, 119	
	Empirical Results, 120	
	Summary, 126	
	ABOUT THE AUTHORS	127
	INDEX	131

Tables and Figures

TABLE 1-1	Checklist of Issues to Consider When Arranging Transportation Between Research Facilities	10
TABLE 2-1	Federal Statutes/Programs Relevant to the Transportation of Vertebrate Research Animals and Products in the United States	12
TABLE 2-2	Designated Port for Importation or Exportation of Wildlife or Derivatives	16
TABLE 2-3	Endangered Species Act Listed Species of Nonhuman Primates	17
TABLE 2-4	Permitting Requirements Under CITES	25
TABLE 2-5	CITES Listed Species of Nonhuman Primates	26
TABLE 2-6	Checklist of Research Animal Regulations and Guidelines	31
TABLE 3-1	Thermoregulation Data on Common Research Animal Species	40
FIGURE 3-1	Graph representing relationship between metabolic rate and ambient temperature in homeotherms.	42
FIGURE 3-2	Changes in thermoneutral zone (range of ambient temperatures at which an animal's heat production is at a minimum) with age and size in chickens.	43
FIGURE 3-3	TNZ of various agricultural animals.	44
TABLE 3-2	Ambient Temperature Ranges for Safe Transportation of Common Adult Research Animals	48

TABLE 3-3	Effects of Various Factors on Effective Environmental Temperature and Relative Risk to Animal Health and Welfare	49
TABLE 3-4	Behavioral and Physiological Signs of Thermal Status	52
TABLE 3-5	Space Allowances for Group-Transported Animals	55
FIGURE 3-4	Space allowances during transportation.	57
TABLE 4-1	Agents and Toxins That Require Registration of the Facility with CDC	67
TABLE 4-2	Elements of an Emergency Plan	68
TABLE 4-3	Characteristics of a Good Shipper	68
TABLE 4-4	Examples of Zoonotic Diseases Transmissible from Research Animals to Humans	69
TABLE 4-5	Infectious Agents and the Susceptible Species of Research Animals	72
TABLE 4-6	Recommendations for Shipment of Research Animals Between Institutions	76
FIGURE 5-1	Locations of research facilities using nonhuman primates, major importation sites, and vendors of nonhuman primates in the United States.	83
FIGURE B-1	Candidate set for the facility-location problem.	121
FIGURE B-2	Solution set for the facility location problem for the NIH grants data set (rodents).	122
TABLE B-1	Total Weighted-System Travel-Distance Reduction with Increase in Supply Points for NIH Grants Data Set (Rodents)	123
TABLE B-2	Total Weighted-System Travel-Distance Reduction with Increase in Supply Points for USDA Cats Data Set	123
FIGURE B-3	Solution set for the facility location problem for the USDA cats data set.	124
FIGURE B-4	Solution set for the facility location problem for the USDA dogs data set.	125
TABLE B-3	Total Weighted-System Travel-Distance Reduction with Increase in Supply Points for USDA Dogs Data Set	126

Abbreviations and Acronyms

AAALAC	Association for Assessment and Accreditation of Laboratory Animal Care International
AATA	Animal Transportation Association
APHIS	Animal and Plant Health Inspection Service
AVMA	American Veterinary Medical Association
AWA	Animal Welfare Act
BMBL	Biosafety in Microbiological and Biomedical Laboratories
CDC	Centers for Disease Control and Prevention
CITES	Convention on International Trade in Endangered Species of Wild Fauna and Flora
DGMQ	Division on Global Migration and Quarantine
DRGs	Dangerous Goods Regulations
DOT	Department of Transportation
EAIIP	Etiologic Agent Import Permit Program
EDIM	Group A rotavirus
ESA	Endangered Species Act
EU	European Union
FASS	Federation of Animal Science Societies
FDA	Food and Drug Administration

FY	fiscal year
FWS	Fish and Wildlife Service
GD-VII	Theiler's murine encephalomyelitis virus strain
GIS	Geographic Information System
HANT	Hantaan
HEPA	high-efficiency particulate air (filters)
HMR	Hazardous Materials Regulation
HPA	hypothalamic pituitary adrenal axis
IATA	International Air Transport Association
ICAO	International Civil Aviation Organization
ILAR	Institute for Laboratory Animal Research
KRV	Kilham Rat virus
LARS	Live Animals Regulations
LCM	Lymphocytic choriomeningitis
LCMV	Lymphocytic choriomeningitis virus
LCT	lower critical temperature
MAD	Mouse adenovirus
MCMV	Murine cytomegalovirus
MHV	Mouse hepatitis virus
MPV (OPV)	Mouse parvovirus (Orphan parvovirus)
MTLV	Mouse thymic virus
MVM	Minute virus of mice
NAP	National Academies Press
NCRR	National Center for Research Resources
NIH	National Institutes of Health
NPRC	National Primate Research Center
NRC	National Research Council
OIE	Office International des Épizooties
OLAW	Office of Laboratory Animal Welfare
PhRMA	Pharmaceutical Research and Manufacturers of America
PHS	Public Health Service
PPE	personal protective equipment
PVM	pneumonia virus of mice

REO 3	Reovirus type 3
RHD	Rabbit haemorrhagic disease
RPV (OPV)	Rat parvovirus (Orphan parvovirus)
RRV	Ross River virus
SARS	Sudden Acute Respiratory Syndrome
SDA/RCV	Sialodacryoadentitis virus/Rat corona virus
SIV	Simian immunodeficiency virus
SPS Agreement	Agreement on the Application of Sanitary and Phytosanitary Measures
TNZ	thermoneutral zone
UCT	upper critical temperature
USDA	US Department of Agriculture
WTO	World Trade Organization

Summary

Transportation of research animals is an essential component of the biomedical research enterprise that can have substantial, although often little understood, effects on the physiological and psychological condition of the animals. Both environmental conditions and the novelty of the transportation experience can cause stress, which can cause short-term changes in physiology and thus affect subsequent research if the animals are utilized immediately after transportation. In addition, transportation stress in great intensity or duration can adversely affect animals' well-being.

Individuals at research facilities often find arranging transportation of animals a challenge. A confusing patchwork of local, national, and international regulations; a perceived lack of high-quality shipping services; a dearth of science-based good practices; and a lack of biosafety standards all represent difficulties with which people are confronted when attempting to move research animals to or from their institution.

In recognition of these challenges and the potential for transportation to affect subsequent research, the National Institutes of Health (NIH) and the National Center for Infectious Diseases asked the National Research Council to form a committee of experts to address current problems encountered in the transportation of research animals and to offer recommendations for rectifying the problems for the benefit of research animals and the research community.

Developing science-based good practices was the major focus of the committee's efforts, which was particularly challenging given the dearth

of literature available on transportation practices and the effect of transportation on the most common research animals. However, using the extensive body of literature that is available on agricultural animals, the committee was able to develop, in Chapter 3, a set of good practices based on some universal concepts of physiology and a scientific understanding of species-specific needs and differences. Good practices were developed to address thermal environment, space requirements, food and water requirements, social interaction and group transportation, handling, monitoring of transportation, emergency procedures, and personnel training. Although precise engineering standards are often preferred by human assessors, the scientific literature supports few engineering standards. Therefore, the good practices recommended by the committee were developed as “performance standards,” which define an outcome (such as animal well-being or safety) and provide criteria for assessing that outcome without limiting the methods by which to achieve that outcome. The use of performance standards allows institutions and organizations the flexibility to adjust their procedures to optimize animal welfare according to the species being transported, the mode of transportation, and local environmental conditions.

During public hearings, it was evident that the issue causing the greatest confusion among those attempting to ship research animals is the overlapping authority that multiple federal agencies have over the research animal transportation process. In Chapter 2, the committee identifies the federal agencies that may be involved in the process and summarizes their agencies’ statutory authority, the aspects of the transportation process that they regulate, and how they enforce their authority (inspection, permitting, and issuing standards). In addition, the committee identifies the major international treaties and agreements that may pertain to the importation or exportation of animals into or from the United States. At the end of the chapter is a checklist of questions that can be used to identify the standards that apply when transporting animals into, out of, or within the United States.

Several of the federal statutes were enacted to prevent the introduction, transmission, or spread of communicable disease in the United States, which could occur either intentionally or unintentionally through human exposure to animals. Infectious pathogens are not only a risk to public health, but can jeopardize animal health, research programs, and agricultural resources if introduced into animal colonies and laboratories. In Chapter 4, the committee identifies diseases of research animals that can be transmitted to humans, agricultural animals, and other research animals, and recommends good practices to avoid biosafety problems during transport and introduction at a new facility. Utilizing a good shipper is important for maintaining biosecurity during transport, as well

as for ensuring the safety and comfort of the animals. The committee identifies characteristics of a good shipper (Chapter 4) and also provides a checklist of factors to consider when arranging transportation of research animals between research facilities (Chapter 1).

The committee also examined the existing system for transporting research animals by truck in the United States (Appendix B). By combining information on the locations where research animals are utilized and the locations of the major research animal vendors and breeders in the United States, the committee was able to construct a geographic information system model. This hypothetical model provides a qualitative sense of the patterns of transportation of research animals. In addition, this section presents the results of a quantitative modeling effort to locate additional supply points “optimally” and an assessment of the potential benefit of the additional supply points.

MAJOR RECOMMENDATIONS

Declining Availability of Air Transportation for Nonhuman Primates

Over the last 10-15 years, most foreign and domestic airlines have implemented a ban on transporting nonhuman primates destined for research. Many factors may have contributed to this decline, including concerns about zoonoses, the high cost of training personnel and acquiring protective equipment, the negative reactions of airline passengers, pressure from animal rights activists, and the unprofitable nature of live-animal transportation. Currently, only one domestic airline and five foreign airlines will consistently transport nonhuman primates. In the committee’s judgment, the most promising solution for ensuring a stable means of transporting nonhuman primates into and within the United States is for NIH, through the National Center for Research Resources, to update and implement the National Primate Plan. This plan could include several different actions to be pursued, including developing national nonhuman primate resources and ensuring financial allowances for costs associated with chartering private airplane transportation.

The committee also recommends that the National Primate Research Centers (NPRCs) and research institutions utilizing nonhuman primates work together to encourage the development of reliable ground transportation for nonhuman primates. Although ground transport of nonhuman primates does occur, it is not widespread, possibly due to economic constraints and because most ground transportation companies are geared toward the transportation of rodents and other small mammals. However, NPRCs and researcher institutions could prepare for the possibility that domestic transportation on commercial airlines may one day become

unavailable by partnering to encourage reliable ground transport, perhaps through professional societies.

The committee also recommends that federal agencies that fund nonhuman-primate research and the commercial shipping community coordinate an initiative to develop a self-contained overshipper to ship nonhuman primates (and other animals) that pose a significant risk of zoonotic exposure. An overshipper is a closed, environmentally controlled container into which a standard primary enclosure is loaded in order to prevent a zoonotic exposure. The advantages in safety, security, and convenience could encourage more airlines to transport nonhuman primates. In addition, with increased research focus on potential agents of bioterrorism, an overshipper could increase the ability of the research community to access or exchange established animal models of infectious and zoonotic diseases.

Regulatory Burden

The complex and confusing regulatory environment surrounding the transportation of research animals led the committee to recommend the establishment of an interagency working group to coordinate all federal inspection and permitting activities related to the transportation of research animals and their products. Currently five federal agencies have oversight authority on various aspects of the transportation process. The resulting overlap of authority presents a significant regulatory burden to individual researchers and commercial shipping operations. Establishing a working group would centralize operations and communications, and would reduce regulatory burden by minimizing the number of inspections and permits that must be issued for each shipment. The committee also recommends that the various federal agencies work to clarify confusing and inconsistent regulations that pertain to transportation of research animals and their products, and in particular the Animal Welfare Act regulations, which are the federal regulations that establish standards for animal welfare that apply to most species of research animals during transportation.

Animal Welfare

Many issues must be considered in order to ensure the comfort, well-being, and safety of research animals during transportation. This can present a challenge to individuals at research institutions that have little previous experience with facilitating the transportation of research animals. These investigators may be unfamiliar with practices that address the welfare of animals, methods that minimize transit time, and charac-

teristics of an effective commercial shipper. Therefore, the committee recommends that research institutions, whether commercial or academic, designate a single individual to be responsible for ensuring the safe shipment and receipt of research animals. This individual would ensure appropriate registration and permitting, as well as the use of US Department of Agriculture-certified carriers, properly trained subcontractors, and appropriate transportation enclosures. It is expected that designation of a responsible individual will alleviate the burden on investigators of becoming familiar with the intricacies of federal regulations and resources such as commercial services, and will, most importantly, ensure the welfare of the animals involved.

Introduction

Over the last 10 years, the biomedical research enterprise has undergone tremendous growth. The amount of federal funding for biomedical research has more than doubled since 1995, and the pharmaceutical, biotechnology, and contract research sectors have all seen double-digit growth (PhRMA, 2005). That growth has been accompanied by parallel increases in research infrastructure, including an increase in the numbers of animals used in biomedical research. The humane transportation of research animals has been a priority, but there are concerns that the rapid increase in the numbers of animals transported, the increasing use of genetically modified animals that may have medical considerations, the complexity of permitting and inspection of research animals, and the dwindling availability of transportation services are adversely affecting the quality and ease of transportation in the United States.

Because of those concerns, the National Center for Infectious Diseases of the Centers for Disease Control and Prevention (CDC) and the National Institutes of Health asked the Institute for Laboratory Animal Research of the National Research Council to convene a committee to address problems associated with the transportation of research animals. The detailed charge to the committee is as follows:

A committee will be formed to address current problems encountered in the transportation of research animals and make recommendations to rectify these problems to the benefit of the research community and the

animals themselves. The committee will focus on all species used in biomedical research and all possible modes of transportation. Specifically, they will address: animal welfare concerns during transportation; availability of quality transportation services for animals, or lack thereof; overlaps or gaps in regulatory oversight; permitting issues; transportation of tissues/specimens; regulatory burden reduction; and potential biosecurity concerns.

The Committee on Guidelines for the Humane Transportation of Laboratory Animals, convened in April 2004, includes experts in veterinary medicine, biosecurity, stress and its psychophysiological effects, research animal logistics and regulatory issues, transportation modeling, research animal welfare during transportation, and the development of transportation guidelines. The committee met three times to deliberate and develop its report. During two of the meetings, the committee held workshops to solicit information from interested parties and the public. In addition, people could provide comments to the committee through the National Academies project website.

Transportation of research animals in the United States may be divided into two major categories: animals transported from a commercial breeder to a research facility, and animals transported between research facilities. It has been reported (White, 2004) that the large commercial rodent breeders transport in excess of 1.5 million containers of animals a year within the United States. Of those shipments, 45% go to for-profit customers and 55% to nonprofit customers. Most (about 92%) of the shipments are made by ground transportation and the remainder by air. The large commercial rodent breeders have established truck routes and either use an in-house fleet of environmentally controlled vehicles or have a standing relationship with shipping companies that specialize in research animal transportation. It is estimated that 70% of containers arrive at their destination in less than 24 hours, 16% in 24–48 hours, and 14% in more than 48 hours (White, 2004).

Commercial breeders' experience with transportation failures is relatively small. The large commercial rodent breeders estimate that only 0.035% of containers experience a problem during transportation, defined as a customer complaint or rejection of shipment: 0.03% of containers shipped by ground transportation and 0.04% of containers shipped by air transportation experience problems (White, 2004).

The importation and exportation of animals to this country are also of interest to this committee. Data on importation and exportation are not available for the majority of research animals, but the importation of non-human primates is tracked through the CDC Division of Global Migra-

tion and Quarantine because of the concern about zoonotic diseases (a detailed discussion of the Division of Global Migration and Quarantine is found in Chapter 2). The majority of nonhuman primates imported into the United States over the last 4 years were cynomolgus macaques. Currently, about half the shipments of nonhuman primates into the United States occur through Los Angeles; most of the rest go through San Francisco and Chicago (CDC, 2005c).

Many options are available for transportation of animals between research facilities. A single company might handle the door-to-door delivery of the animals. Most often, when a single company, known as a carrier, is used, the company will pick up a shipment of animals, consolidate the shipment with other shipments, fly or truck the consolidated shipment with its own fleet of vehicles along established shipping routes, and deliver the shipment to its destination. Occasionally, researchers will use a carrier to ship animals to another institution, unaware that, if the destination is not near an established shipping route or if there are not enough shipments to consolidate, the carrier might subcontract the delivery to a third-party carrier. It is also possible to have a specialty courier pick up, transport, and deliver a shipment of animals in a dedicated vehicle.

Sometimes it is necessary to use two or more transport companies. In that case, a company known as a freight forwarder or handler will pick up a shipment, deliver it to a third-party carrier, which may consolidate shipments and ship to an intermediary destination, and then pick up the shipment from the third-party carrier and deliver it to its destination.

Little information is available on the transportation of research animals between research institutions within the United States. Public records on such transportation are not maintained. Because of the lack of relevant data, the committee could not draw any conclusions about the quality of this type of transportation. The committee chose to identify some of the issues that an individual researcher should consider when making arrangements for the transportation of animals between research facilities (see Table 1-1). To further assist individuals, the committee also identified characteristics of good shippers (please refer to Table 4-3 in Chapter 4).

Many companies can coordinate or directly transport shipments of research animals in environmentally controlled trucks (AAALAC International, 2003). In addition, the US Department of Agriculture (USDA) maintains a list of registered carriers (trucking or airline companies that transport animals) and handlers (companies that pick up shipments and deliver to a third-party carrier for transportation) on its website at:

<http://www.aphis.usda.gov/ac/publications.html>

TABLE 1-1 Checklist of Issues to Consider When Arranging Transportation Between Research Facilities

Shipping Container	<ul style="list-style-type: none"> • Is the shipping container appropriate for the expected conditions? • Does the container comply with USDA standards set for warm-blooded vertebrates except rats, mice, and birds? • Does the container comply with International Air Transport Association standards if transport includes air travel?
Shipping Company	<ul style="list-style-type: none"> • Will the same company be transporting the animals during all legs of the journey, or will a third-party carrier or subcontractor be used for some legs?
Environmental Conditions	<ul style="list-style-type: none"> • Are environmentally controlled vehicles used for all segments of ground transportation? • If environmentally controlled vehicles will not be used, or air travel is involved, does the shipping company have contingency plans for maintenance of the animals if the ambient temperature is below or exceeds acceptable ranges? • Does the shipping company have standard operating procedures for ensuring that animals are not exposed to extreme environmental conditions during transfer between vehicles and at the end destination?
Training	<ul style="list-style-type: none"> • Does the company provide specialized training for all employees involved in transportation of animals? • If a third-party carrier or subcontractor is involved in the transportation, are they also trained?

Regulations and Guidelines for the Transportation of Research Animals

The purposes of this chapter are to identify national and international agencies responsible for the safe, humane, and expeditious transportation of live research animals and biological materials (tissues and specimens) derived from animals, to summarize and review regulations governing their transportation, and to identify overlaps and gaps among regulations and agency responsibilities insofar as they represent strengths and impediments (see Table 2-1). The first section of the chapter focuses on national and the second on international transportation regulatory agencies, their responsibilities and their regulations. Although the committee has attempted to provide a comprehensive summary of transportation regulations and requirements (see Table 2-6 at end of chapter), the shipper is advised to check with the appropriate agencies prior to shipment to ensure compliance with current regulations and requirements.

NATIONAL REGULATIONS AND GUIDELINES

Several federal agencies in the United States regulate the transport of research animals and their tissues specifically or of species that might be used for research. The agencies are vested with authority under various federal statutes. In this section, the regulations and guidelines to inform readers unfamiliar with the web of regulations are discussed. All of the regulations and guidelines are applicable to the transportation of research animals, but only two laws identify standards for humane care during

TABLE 2-1 Federal Statutes/Programs Relevant to the Transportation of Vertebrate Research Animals and Products in the United States^a

Agency	Agency Program or Federal Regulation	Nonhuman Primates		Cats and Dogs	
		Interstate	Import/export	Interstate	Import/export
USDA	Animal Welfare Act	X	X	X	X
FWS	CITES		X		++
	Lacey Act	X	X		
	Endangered Species Act	X	X		
CDC	Division on Global Migration and Quarantine (DGMQ)		X*		X*
	Etiologic Agent Import Permit Program (EAIPP)				
DOT	Infectious Substances Program				
	Radioactive/Poisonous Materials Program				
FDA	42 CFR 70.2 and 21 CFR 1240.30				

^a Multiple columns may be applicable to a single shipment of animals or animal products.
 * Regulates importation only.
 ** Regulates importation and interstate transportation only.
 *** Regulates interstate transportation only.
 + Animals that are known to harbor a human pathogen fall under the purview of DGMQ. Animals with human pathogens fall under the purview of EAIPP only when they are experimentally infected.
 ++ Wild-domestic hybrids, such as leopard cats, may be considered wildlife under CITES.
 +++ EAIPP regulates the importation and interstate transportation of all bats, regardless of disease status.

transportation: the Animal Welfare Act (AWA), which details specific standards for animal care during transportation, and the Lacey Act, which provides that wildlife be transported in accordance with the International Air Transport Association (IATA) Live Animals Regulations (LARs).

DEPARTMENT OF AGRICULTURE

The AWA provides standards for the humane handling, care, treatment, and transportation of animals (AWA, 7 USC 2131 et seq.). AWA regulatory authority is vested in the Secretary of Agriculture and implemented by the US Department of Agriculture (USDA) Animal and Plant Health Inspection Service (APHIS). With the exception of rats of the genus *Rattus* and mice of the genus *Mus*, bred specifically for research purposes, as well as all birds, and livestock or poultry used for improving animal nutrition, breeding, management, production efficiency, or food or fiber quality, the AWA regulates the transportation of all warm-blooded animals intended for use in research, teaching, or testing (9 CFR 1.1). That regulation applies to transportation of AWA-covered species within the United States, as well as their transportation on foreign air carriers traveling into, within, or from the United States, its territories or possessions, or the District of Columbia (Federal Register, Vol 69, No 66, pages 17899–17901).

With regard to the transportation of animals, the act contains standards for consignment (delivery of animals to an entity for transport), primary transportation enclosure, primary conveyance, food and watering requirements during transportation, terminal facilities, care in transit, and handling. The AWA contains standards for different groups of similar species, with separate rules for transporting dogs and cats, guinea pigs and hamsters, rabbits, nonhuman primates, marine mammals, and all other covered warm-blooded animals. The AWA standards are rather extensive and pertain directly to animal welfare. They are summarized in Appendix B for ease of use.

US FISH AND WILDLIFE SERVICE

The US Fish and Wildlife Service (FWS) regulates the importation, exportation, and interstate trade of dead and live wild animals and their tissues and products imported into or exported¹ from the United States

¹Wildlife (including parts and products) that are in transit through the United States from one foreign country to another foreign country are exempt provided that the wildlife stays in the United States only for the time needed to transfer the specimen to the mode of transportation used to continue to the final destination and remain under control of Customs and Border Protection. Wildlife that is listed as injurious (Part 16), endangered or threatened

for any purpose (including research). *Wildlife* means any wild animal, whether alive or dead, including any wild mammal, bird, reptile, amphibian, fish, mollusk (clam, snail, squid, or octopus), crustacean (crab, lobster, or crayfish), insect, sponges, corals, or other invertebrate, whether or not bred, hatched, or born in captivity and including any part, product (including manufactured products and processed food products), egg, or offspring thereof.

The FWS authority to regulate wildlife stems from the Fish and Wildlife Act of 1956 (16 USC §§ 742a-754j-2). All wildlife being imported into or exported from the United States must be declared to FWS through completion of Form 3-177 at the time of entry or exportation and is subject to inspection. Wildlife being transported through the United States to a final international destination does not require declaration to FWS, unless it is listed as an injurious (50 CFR 16), endangered, or threatened species (50 CFR 17 and 50 CFR 222-224); a marine mammal (50 CFR 18 and 50 CFR 216); a migratory bird (50 CFR 21); or a bald or golden eagle (50 CFR 22).

FWS requires that all wildlife be imported and exported through a series of designated ports. As shown in Table 2-2, some port locations are designated to allow the importation or exportation of any wildlife, and others are restricted to allow only particular species of wildlife.

Endangered Species Act

The purpose of the Endangered Species Act (ESA) of 1973 (16 USC §§ 1531-1544) is to conserve and recover species of fish, wildlife, and plants that are listed as threatened or endangered in the United States or elsewhere and to preserve the ecosystems upon which these species depend (ESA Section 2(b)). Criminal and civil penalties are designated under the ESA for violations. Nonhuman-primate species listed under the ESA are detailed in Table 2-3. Other species that are currently listed as endangered or threatened under the ESA can be found on line at:

<http://www.fws.gov/endangered/wildlife.html#Species>

FWS regulates species listed as endangered and threatened under the ESA through a system of permits. The permits are required to use these species for scientific purposes, to operate a captive-breeding program, and to transport these species through importation, exportation, or interstate

species (Parts 17 and 222-224), marine mammal (Parts 18, 216), migratory bird (Part 21), or a bald or golden eagle (Part 22) and is moving through the United States is considered an import and cannot be treated as in transit.

TABLE 2-3 Endangered Species Act Listed Species of Nonhuman Primates

<p>Endangered</p> <p><i>Allocebus</i> spp. (hairy-eared dwarf lemurs)</p> <p><i>Alouatta palliata</i> (mantled howler monkey)</p> <p><i>Ateles geoffroyi frontatus</i> (Central American spider monkey)</p> <p><i>Ateles geoffroyi panamensis</i> (Central American spider monkey)</p> <p><i>Avahi</i> spp. (Avahi, woolly lemurs)</p> <p><i>Brachyteles arachnoids</i> (woolly spider monkey)</p> <p><i>Bradypus torquatus</i> (Brazilian three-toed sloth)</p> <p><i>Cacajao</i> spp. (uakaris)</p> <p><i>Callimico goeldii</i> (Goeldi's marmoset)</p> <p><i>Callithrix aurita</i> (buffy tufted-ear, white-eared marmoset)</p> <p><i>Callithrix flaviceps</i> (buff-headed marmoset)</p> <p><i>Cercocebus galeritus galeritus</i> (Tana River mangabey)</p> <p><i>Cercocebus torquatus</i> (white-collared mangabey)</p> <p><i>Cercopithecus Diana</i> (Diana monkey)</p> <p><i>Cercopithecus erythrogaster</i> (red-bellied monkey)</p> <p><i>Cercopithecus erythrotis</i> (red-eared nose-spotted monkey)</p> <p><i>Cercopithecus lhoesti</i> (L'hoest's monkey)</p> <p><i>Cheirogaleus</i> spp. (dwarf lemurs)</p> <p><i>Chiropotes albinasus</i> (white-nosed saki)</p> <p><i>Chiropotes satanas satanas</i> (southern bearded saki)</p> <p><i>Colobus satanas</i> (black colobus monkey)</p> <p><i>Daubentonia madagascariensis</i> (Aye-aye)</p> <p><i>Gorilla gorilla</i> (gorilla)</p> <p><i>Hapalemur</i> spp. (bamboo lemurs and gentle lemurs)</p> <p><i>Hylobates</i> spp. (including <i>Nomascus</i>) (gibbons)</p> <p><i>Indri</i> spp. (indri)</p> <p><i>Lagothrix flavicauda</i> (yellow-tailed woolly monkey)</p> <p><i>Lemur</i> spp. (ring-tailed lemurs)</p> <p><i>Leontopithecus</i> spp. (golden-rumped tamarin)</p> <p><i>Lepilemur</i> spp. (sportive lemurs)</p> <p><i>Macaca silenus</i> (lion-tailed macaque)</p> <p><i>Mandrillus leucophaeus</i> (drill)</p> <p><i>Mandrillus sphinx</i> (mandrill)</p> <p><i>Microcebus</i> spp. (mouse lemurs)</p> <p>Monkey, black howler</p>	<p><i>Nasalis concolor</i> (Pagi Island langur)</p> <p><i>Nasalis larvatus</i> (proboscis monkey)</p> <p><i>Pan paniscus</i> (pygmy chimpanzee—wild)</p> <p><i>Pan troglodytes</i> (wild chimpanzee)</p> <p><i>Phaner</i> spp. (fork-marked lemurs)</p> <p><i>Planigale ingrami subtilissima</i> (little planigale)</p> <p><i>Pongo pygmaeus</i> (orangutan)</p> <p><i>Procolobus pennantii kirki</i> (Zanzibar red colobus monkey)</p> <p><i>Procolobus preussi</i> (Preuss' red colobus monkey)</p> <p><i>Procolobus rufomitratu</i>s (Tana River red colobus monkey)</p> <p><i>Propithecus</i> spp. (sifakas)</p> <p><i>Pygathrix nemaus</i> (douc langur)</p> <p><i>Rhinopithecus avunculus</i> (Tonkin snub-nosed monkey)</p> <p><i>Rhinopithecus bieti</i> (Yunnan snub-nosed monkey)</p> <p><i>Rhinopithecus brelichi</i> (guizhou snub-nosed monkey)</p> <p><i>Rhinopithecus roxellana</i> (Sichuan snub-nosed monkey)</p> <p><i>Saguinus bicolor</i> (pied tamarin)</p> <p><i>Saguinus oedipus</i> (cotton-top marmoset)</p> <p><i>Saimiri oerstedii</i> (red-backed squirrel monkey)</p> <p><i>Sennopithecus entellus</i> (gray, Hanuman langur)</p> <p>Tamarin, white-footed</p> <p><i>Trachypithecus francoisi</i> (Francois' langur)</p> <p><i>Trachypithecus geei</i> (golden langur)</p> <p><i>Trachypithecus pileata</i> (capped langurs)</p> <p><i>Varecia</i> spp. (ruffed lemur, variegated lemurs)</p> <p>Threatened</p> <p><i>Alouatta pigra</i> (black howler monkey)</p> <p><i>Macaca arctoides</i> (stump-tailed macaque)</p> <p><i>Macaca cyclops</i> (Formosan rock macaque)</p> <p><i>Macaca fuscata</i> (Japanese macaque)</p> <p><i>Macaca sinica</i> (Toque macaque)</p> <p><i>Nycticebus pygmaeus</i> (lesser slow lorise)</p> <p><i>Pan troglodytes</i> (chimpanzees—captive)</p> <p><i>Presbytis potenziiani</i> (long-tailed langur)</p> <p><i>Presbytis senex</i> (purple-faced langur)</p> <p><i>Saguinus leucopus</i> (white footed tamarin)</p> <p><i>Tarsius syrichta</i> (Philippine tarsier)</p> <p><i>Theropithecus gelada</i> (gelada baboon)</p>
--	---

Lacey Act

The FWS is also vested with federal regulatory authority from the Lacey Act of 1900 (18 USC 42). The Lacey Act is the oldest federal wildlife protection law in the United States and prohibits inhumane and unhealthful transportation conditions for any animal defined as “wildlife” (see above for definition). The regulations for transportation under the act (found in 50 CFR 14.101-14.172, which are specific for mammals and birds) are similar to the IATA-LARs. The Lacey Act also applies the guidelines provided in the LARs to nonairline methods of transportation (Kreger and Farris, 2003).

CENTERS FOR DISEASE CONTROL AND PREVENTION

The Centers for Disease Control and Prevention (CDC) is a component of the US Department of Health and Human Services and enforces regulations to prevent the introduction, transmission, or spread of communicable diseases in the United States. CDC regulates the importation of any animal or animal product capable of carrying a zoonotic disease. CDC regulates importation through permitting, registration, and quarantine; it does not regulate the welfare or care of the animals.

Importation of Live Animals

CDC regulates the importation of dogs, cats, turtles, nonhuman primates, and other animals. Dogs and cats are subject to inspection by CDC at the port of entry. Dogs must be accompanied by proof of vaccination against rabies at least 30 days before entry into the United States unless they were in areas considered rabies-free for at least 6 months before importation. Importation of dogs or cats into the United States does not require a permit or other paperwork from CDC. Dogs too young to be vaccinated or without a current rabies vaccination may be admitted into the United States if the owner signs a confinement agreement (Form 75.37), which can be accessed from the following website:

<http://www.cdc.gov/ncidod/dq/animal.htm>

To prevent infection of members of the public with Salmonella and Arizona bacteria, shipments containing more than six turtles with a carapace length of less than 4 in. or viable turtle eggs generally cannot be imported into the United States (42 CFR 71.52). However, shipments containing more than six turtles with a carapace length of less than 4 in. or viable turtle eggs or any combination of turtles and turtle eggs may be

imported into the United States for bona fide scientific, educational purposes, or for exhibition when accompanied by a permit issued by CDC, Division of Global Migration and Quarantine (DGMQ) (42 CFR 71.52(c)(2)).

CDC may also implement prohibitions on the importation of animals when new health risks to humans arise. As of June 2003, CDC had prohibited the importation of all African rodents into the United States because of concerns about monkeypox (42 CFR Part 71.56). In January 2004, CDC prohibited the importation of civets because of concerns about severe acute respiratory syndrome (CDC, 2004). However, importation of those animals for scientific, exhibition, or educational purposes is allowed with written permission from CDC (42 CFR 71.56; CDC, 2004). Since February 2004, CDC has prohibited the importation of Asian birds from selected countries due to highly pathogenic avian influenza A (H5N1). For more information about obtaining a permit or permission to import turtles, African rodents, or other prohibited animals, contact the CDC DGMQ at 404-498-1670 or online at:

<http://www.cdc.gov/ncidod/dq/nonhuman.htm>

Monkeys and other nonhuman primates cannot be imported as pets, but importation for scientific research, exhibition, or education is permitted. Under the Federal Quarantine Regulations (42 CFR 71.53), people importing nonhuman primates for research purposes must register with the CDC DGMQ and must

- Certify that imported nonhuman primates will be used only for *bona fide* science, education, or exhibition;
- Implement disease control measures to minimize human exposure to the animals during transportation, isolation, and quarantine;
- Isolate each shipment of nonhuman primates for 31 days, monitor the animals for illness, test for tuberculosis, maintain records regarding illness and death, and test for filovirus infection in shipments where illness or death occurs during the quarantine period;
- Report suspected zoonotic illness to CDC; and,
- Maintain records regarding the distribution of each shipment (NRC, 2003a).

Before registration and periodically thereafter, the CDC DGMQ inspects importer facilities and reviews their operations, including assessing transportation and disease-control measures and reviewing animal health records. For each shipment of nonhuman primates into the United States, CDC must review proposed plans for each shipment and monitor the handling of arriving shipments at the port of entry and at the quarantine facility (NRC, 2003a). To receive information on the importation of non-

human primates and to register as an importer of nonhuman primates, contact the CDC DGMQ, whose phone number and website are provided above.

Importation and Transportation of Etiologic Agents

The CDC Etiologic Agent Import Permit Program regulates the importation and subsequent transfer within the United States of live bats, as well as animals, insects, or animal tissues that contain etiologic agents. *Etiologic agents* are defined by CDC as a viable microorganism or its toxin which causes, or may cause, human disease; however, CDC does not specifically identify the microorganisms, viruses, or prions that it considers etiologic agents.

Contact information and permit applications for importation of etiologic agents or live bats (Office of Management and Budget form 0920-0199) are available at:

<http://www.cdc.gov/od/ohs/biosfty/impptper.htm>

US DEPARTMENT OF TRANSPORTATION

The US Department of Transportation (DOT), Pipeline and Hazardous Materials Safety Administration (PHMSA), Office of Hazardous Materials establishes regulations to govern the transportation of hazardous materials in interstate, intrastate, and foreign commerce under the Hazardous Materials Regulations (HMR; 49 CFR 171-180). Hazardous materials include materials that are poisonous, radioactive, and infectious—all of which may be contained in animals or animal products that may be transported in the course of biomedical research. An *infectious substance* is defined by DOT as a material known to contain or suspected of containing a pathogen, including a virus, microorganism, or prion that has the potential to cause disease in humans or animals. Live animals that contain an infectious substance are required to be transported under terms and conditions approved by PHMSA's Associate Administrator for Hazardous Materials Safety.

The HMR cover five areas of shipper responsibility:

- Determining the hazard class(es) of the material offered for transportation;
- Communicating the hazard(s) using shipping papers, labels, markings, placards, and emergency-response information, as required;
- Packaging requirements;
- Operational rules, including for transportation by air; and,
- Training and security.

The majority of these tasks are performed by the shipper. The carrier must also ensure that the materials received comply with and are transported in accordance with the HMR. The HMR also require people or institutions that are shipping certain types or amounts of hazardous materials, such as materials extremely toxic by inhalation, to register annually with DOT (49 CFR 107.601).

There are many exceptions, as some materials and situations that are common in biomedical research are not subject to the requirements of the HMR. The exceptions are too numerous to describe here, but persons interested in shipping hazardous materials, including infectious, poisonous, or radioactive animal products or live infectious animals, should contact PHMSA's Hazardous Materials Information Center to determine whether shipment would be subject to the requirements of the HMR or visit their website online at:

<http://hazmat.dot.gov/>

PUBLIC HEALTH SERVICE

Institutions that are funded by Public Health Service (PHS) funds are subject to the PHS Policy on the Humane Care and Use of Laboratory Animals (commonly referred to as the PHS Policy). Under the Health Research Extension Act of 1985, the PHS Policy mandates adherence to the guidelines in the *Guide for the Care and Use of Laboratory Animals* (the *Guide*) (NRC, 1996). The *Guide* provides performance standards on the transportation of research animals. It states that all transportation of animals should be planned to minimize transit time and the risk of zoonoses, protect against environmental extremes, avoid overcrowding, provide food and water when indicated, and protect against physical trauma.

The PHS Policy does not provide for regular inspections. Rather, institutions are expected to police themselves in order to ensure compliance with the principles of the PHS Policy and the *Guide*. The National Institutes of Health Office of Laboratory Animal Welfare will conduct investigations when it receives complaints.

FOOD AND DRUG ADMINISTRATION

Under 42 CFR 70.2 and 21 CFR 1240.30, authority is given to the commissioner of the Food and Drug Administration (FDA) to take actions believed "reasonably necessary to prevent" the spread of communicable disease when state and local actions are inadequate. By that authority, FDA prohibits the intrastate and interstate transportation—for the purposes of commerce, sale, or any other type of commercial or public distri-

bution—of psittacine birds, molluscan shellfish, turtles, and African rodents or other animals that may carry the monkeypox virus (refer to 21 CFR 1240.65, 21 CFR 1240.60, 21 CFR 1240.62, 21 CFR 1240.63, respectively). Essentially, CDC prohibits the importation of the animals, and FDA prohibits their domestic interstate and intrastate movement, with special procedures for exceptional circumstances.

Exceptions to the prohibitions are possible only by obtaining written permission from the FDA. A written request must be sent to the Listed Animal Permit Official at the FDA Center for Veterinary Medicine. The request must include the reasons why an exemption is needed and a description of the animals involved, how the animals will be transported, holding facilities, quarantine procedures, and veterinarian evaluation of the transportation. For more information, visit the following website:

<http://www.fda.gov/cvm/>

STATE HEALTH AND AGRICULTURAL REGULATIONS

All states have regulations that control the movement of animals into them, although not all of these regulations pertain to research animals. For example, the California Department of Health Services requires a permit to transport nonhuman primates into the state. Links to the regulations have been organized by the USDA APHIS Veterinary Services at:

<http://www.aphis.usda.gov/vs/sregs/>

State regulations that pertain specifically to wildlife are found at:

<http://offices.fws.gov/statelinks.html>

The committee recommends that people arranging transportation of research animals consult those websites, particularly when arranging for transportation of animals between research institutions.

INTERNATIONAL REGULATIONS FOR TRANSPORTING RESEARCH ANIMALS

Regulations pertaining to the international transportation of live animals, tissues, and specimens are intended to ensure the comfort of animals and the safety of animals and their handlers, and to minimize the biosecurity risks associated with the handling of live animals and of the colonies into which live animals are introduced. The international regula-

tory system is a patchwork of agreements and agencies that govern the following:

- Pest control and biosecurity;
- Research animals;
- Agriculture;
- Conservation of endangered species; and,
- Animals used in exhibits or events (shows or sports).

Because several agencies share responsibility for enforcing international transportation statutes and most countries have importation requirements, the regulatory climate is often complicated. The following is a discussion of the international treaties, agencies, or laws that provide regulation or guidance on the transportation of research animals. With the exception of CITES, enforced in 169 countries, each country has its own system of laws and guidance that may or may not draw on the treaties and laws discussed here. The Air Cargo Tariff book, published by the International Air Transport Association, is a source of information on international documentation and import requirements, though one should be aware that the information can change rapidly and requirements may be specific to a province or region. The committee suggests that the person(s) or institution(s) importing animals into the United States or exporting animals from the United States contact the consulate or website of the foreign country to determine which treaties the country enforces and the specific requirements for complying with local laws. In some cases, negotiations are necessary to address incompatibilities between US export and foreign import requirements. An export broker may be useful in assisting a shipper in fulfilling foreign importation requirements.

CITES

CITES, also called the Washington Convention, establishes a permit system for regulating the trade of plants and animals threatened by extinction and those that may be threatened by extinction if trade of that species is not controlled. In this context, *trade* refers to movement of a specimen across international borders for any purpose and includes commercial and noncommercial trade. It includes the importation, exportation, or re-exportation (exportation of a specimen that was imported) of live and dead plants and animals or parts or derivatives of them. At the time of publication, some 169 nations are parties to CITES. In the United States, the ESA of 1973 implements the international CITES treaty (50 CFR 23). FWS administers both the ESA and the CITES treaty and therefore is

the authority responsible for issuing importation and exportation permits in the United States.

Under CITES, animals covered by the treaty are listed in three appendixes:

- Appendix I lists species threatened with extinction. Trade in specimens of those species is permitted only in exceptional circumstances; commercial trade is prohibited.
- Appendix II lists species not necessarily threatened with extinction but in which trade must be controlled to avoid use incompatible with their survival.
- Appendix III lists species that are protected in at least one country that the animal is native to and that has asked other CITES parties for assistance in controlling the trade.

A database listing all of the CITES-listed species has been established. It can be accessed online at:

<http://www.cites.org/eng/resources/species.html>

The permitting requirements pertaining to the transportation of Appendix I, II, and III specimens are summarized in Table 2-4. In addition to requiring specific permits, CITES, under Resolution 10.21, endorses the IATA LARs for air and surface transportation. CITES-listed species can be imported or exported only through the following designated ports in the United States:

Anchorage, AK	Louisville, KY
Atlanta, GA	Memphis, TN
Baltimore, MD	Miami, FL
Boston, MA	Newark, NJ
Chicago, IL	New Orleans, LA
Dallas-Fort Worth, TX	New York, NY
Honolulu, HI	Portland, OR
Houston, TX	San Francisco, CA
Los Angeles, CA	Seattle, WA

Nonhuman primates are the group of CITES species most commonly used in biomedical research in the United States. As shown in Table 2-5, wild-caught species of nonhuman primates are in both Appendix I and Appendix II. However, animals in Appendix I that are bred in captivity for commercial purposes at a facility registered with the CITES secretariat

TABLE 2-4 Permitting Requirements Under CITES

Appendix I	Appendix II	Appendix III
Importation and exportation permits required	Exportation permit required if exporting from any country	Exportation permit required only if exporting from the nation that listed the species in Appendix III
Permits are limited to specimens that will be used in scientific research, education, or conservation		

are considered Appendix II specimens for the purposes of permitting (CITES Article VII.4). Furthermore, where a management authority of the state of export is satisfied that an animal was bred in captivity, a certificate by that management authority stating that the animal was bred in captivity *may* be accepted in lieu of any other required permits or certificates (CITES Article VII.5). This exemption also pertains to tissues derived from a captive-bred animal. However, the term 'bred in captivity' only applies when the animals in question are of a second or subsequent generation bred in captivity *and* there has been no introduction of specimens from the wild into the breeding population except to prevent or alleviate deleterious inbreeding (CITES Resolution Conf. 10.16-revised). As noted in Chapter 1, nonhuman primates are regularly imported into the United States for research purposes. Practically speaking, this means that, with rare exception, nonhuman primates bred for research in the United States cannot attain the designation of 'bred in captivity.'

INTERNATIONAL CIVIL AVIATION ORGANIZATION

The International Civil Aviation Organization (ICAO), an agency of the United Nations, publishes *Technical Instructions for the Safe Transport of Dangerous Goods by Air* biannually (currently the 2005-2006 edition). The *Technical Instructions* expand on the broad provisions governing the international transportation of dangerous goods by aircraft, which are in Annex 18 to the Convention on International Civil Aviation and are based on the United Nations recommendations for the Transport of Dangerous Goods. The *Technical Instructions* contain the detailed regulations necessary for the safe transportation of dangerous goods by aircraft and are for use by all parties involved in the transportation chain, such as shippers, carriers, and country authorities. They are recognized by the US DOT as an alternative to complying with the HMR.

TABLE 2-5 CITES Listed Species of Nonhuman Primates

Appendix I	Appendix II
<i>Alouatta coibensis</i> (Coiba Island howler monkey)	All primate species
<i>Alouatta palliata</i> (mantled howler monkey)	not listed in
<i>Alouatta pigra</i> (Mexican black howler monkey)	Appendix I
<i>Ateles geoffroyi frontatus</i> (black-foreheaded spider monkey)	
<i>Ateles geoffroyi panamensis</i> (Panamanian red spider monkey)	
<i>Brachyteles arachnoides</i> (woolly spider monkey, southern muriqui)	
<i>Cacajao</i> spp. (uakaris)	
<i>Callimico goeldii</i> (Goeldi's marmoset)	
<i>Callithrix aurita</i> (buffy tufted-ear marmoset)	
<i>Callithrix flaviceps</i> (buffy-headed marmoset)	
<i>Cercocebus galeritus galeritus</i> (tana crested mangabey)	
<i>Cercopithecus Diana</i> (Diana monkey)	
<i>Cheirogaleida</i> spp. (dwarf, lemurs)	
<i>Chiropotes albinus</i> (white-nosed bearded saki)	
<i>Daubentonia madagascariensis</i> (aye-aye)	
<i>Gorilla gorilla</i> (gorilla)	
<i>Hylobatidae</i> spp. (gibbons)	
<i>Indridae</i> spp. (indrides, woolly lemurs, avahs, sifakas)	
<i>Lagothrix flavicauda</i> (yellow-tailed woolly monkey)	
<i>Lemuridae</i> spp. (large lemurs)	
<i>Leontopithecus</i> spp. (lion tamarins)	
<i>Macaca silenus</i> (lion-tailed macaque)	
<i>Mandrillus leucophaeus</i> (drill)	
<i>Mandrillus sphinx</i> (mandrill)	
<i>Nasalis concolor</i> (pig-tailed langur)	
<i>Nasalis larvatus</i> (proboscis monkey)	
<i>Pan</i> spp. (chimpanzees)	
<i>Pongo pygmaeus</i> (orangutan)	
<i>Presbytis potenziani</i> (Mentawai Island leaf-monkey)	
<i>Procolobus pennantii kirkii</i> (Kirk's red colobus)	
<i>Procolobus rufomitratu</i> s (tana river red colobus)	
<i>Pygathrix</i> spp. (snub-nosed monkeys, douc langurs)	
<i>Saguinus bicolor</i> (Brazilian bare-faced tamarin)	
<i>Saguinus geoffroyi</i> (Geoffroy's tamarin)	
<i>Saguinus leucopus</i> (white-footed tamarin)	
<i>Saguinus oedipus</i> (cotton-top tamarin)	
<i>Saimiri oerstedii</i> (Central American squirrel monkey)	
<i>Semnopithecus entellus</i> (hanuman langur)	
<i>Trachypithecus geei</i> (golden langur)	
<i>Trachypithecus pileatus</i> (capped langur)	

INTERNATIONAL AIR TRANSPORT ASSOCIATION

The mission of the IATA is to “represent, lead, and serve the airline industry.” IATA is also a collective link between third parties and the airline industry. Working standards for the aviation industry are developed within IATA to foster safe and efficient air transportation and to serve the stated policies of most of the world’s governments.

Live Animals Regulations

The LARs are applicable to air transportation companies that are members of IATA. Persons or organizations that ship live animals by IATA-member airlines, whether as cargo or as baggage, must comply with the LARs in their entirety.

The IATA LARs, updated yearly, were adopted by CITES and the World Animal Health Organization (described below) as the guidelines for transportation of animals by air. Those regulations have been used by the Council of Europe as a basis for its code of conduct for the international transportation of farm animals. The European Union has adopted the LARs as the minimal standards for transporting animals in containers, pens, and stalls. The regulations are intended to prevent harm to these receptacles and to handling personnel when animals are being transported. Furthermore, they act to meet flight safety requirements for the benefit of the traveling public, the crew, and the airplane. The LARs consist of government regulations and specific variations filed by such agencies as USDA and FWS. For example, US variations USG-01 through USG-20 belong to USDA and contain more restrictive guidance than is required by the AWA. Similarly, USG-21 through USG-40 is issued by FWS. For more information about the IATA LARs, go online at:

<http://www.iata.org/ps/publications/9105.htm>

Dangerous Goods Regulations

The IATA Dangerous Goods Regulations (DGRs) are similar to the ICAO Technical Instructions for the Safe Transport of Dangerous Goods by Air. However, the DGRs include additional requirements that are more restrictive than the Technical Instructions, to reflect industry standard practices or operational considerations.

Dangerous goods are defined as goods that meet the classification criteria of one or more of nine United Nations hazard classes. These hazard classes are identical to those used by the US DOT’s Office of Hazardous Materials Standards, Pipeline and Hazardous Materials Safety

Administration, and include toxic, infectious, and radioactive animal tissues, and live infectious animals. The DGRs do not have official standing under the US DOT's HMR.

For more information on the IATA DGRs, go online at:

http://www.iata.org/whatwedo/dangerous_goods1

WORLD ANIMAL HEALTH ORGANIZATION

The primary functions of the World Animal Health Organization, also known as the Office International des Épizooties (OIE), are preventing and raising awareness of zoonoses worldwide. OIE was created in January 1995 as part of the World Trade Organization (WTO) Agreement on the Application of Sanitary and Phytosanitary Measures (SPS Agreement), which aims to harmonize animal health standards and thus reduce their dampening effect on international trade. The main goals of the OIE are:

- To ensure transparency in the global animal disease and zoonosis situation;
- To collect, analyze, and disseminate scientific veterinary information;
- To provide expertise and encourage international solidarity in the control of animal diseases;
- Within its mandate under the WTO SPS Agreement, to safeguard world trade by publishing health standards for international trade in animals and animal products;
- To improve the legal framework and resources of national veterinary services; and
- To provide a better guarantee of the safety of food of animal origin and to promote animal welfare through a science-based approach.

Among other tasks, the SPS Agreement charges OIE with developing international standards, guidelines, and recommendations for protecting animal health and preventing zoonoses. To that end, OIE has developed the *Terrestrial Animal Health Code* and the *Aquatic Animal Health Code*, which provide OIE member countries with standards, guidelines, and detailed recommendations for establishing their own regulations regarding the importation of animals, animal genetic material, and animal products.

OIE is also responsible for improving systems by which information on animal health is gathered and analyzed on a global basis. OIE manages the world animal health information system, which uses data submitted by member countries to help identify the diseases, including zoonoses, that present the most serious threats to animal and human health worldwide. During the recent outbreak of avian influenza in Asia, OIE played a

crucial role in alerting countries of the outbreak and recommending courses of action. As a result of OIE's activities, many countries temporarily halted importation from the affected areas.

Several years after OIE's founding, its member countries decided to give animal welfare high priority and to include it in their 2001-2005 strategic plan. Although OIE had not been specifically mandated by the WTO SPS to protect animal welfare, the OIE's status as the international authority on animal health and zoonoses and the member countries' strong need for guidelines to assist them in conducting bilateral negotiations led the member countries to the conclusion that OIE should provide international leadership in protecting the welfare of imported animals.

Member countries recognized that from a public-policy perspective, the protection of animals involved many scientific, ethical, economic, and political considerations. Therefore, they endeavored to develop a detailed vision and strategy that would incorporate and balance those considerations. Ultimately, the OIE International Committee decided that OIE would give high priority to animals used in agriculture and aquaculture and would address the issues of transportation, humane slaughter, and killing for disease-control purposes first, followed by housing and management. OIE would address other issues, such as research animals and wildlife, as resources permitted.

Member countries delegate responsibility for OIE reporting and participation to the directors of their veterinary services. For the United States, the USDA APHIS Associate Deputy Administrator for International Services assumes that role.

THE EUROPEAN UNION

The European Union (EU) laws regulating the transportation of research animals are enforced by the Health and Consumer Protection Directorate General. Research animal transportation is addressed through legislative and nonlegislative actions in three interrelated policy areas:

- Consumer policy (Treaty Articles 95 and 153);
- Public health (Treaty Articles 95, 152, and 300); and
- Food safety, animal health, animal welfare, and plant health (Treaty Articles 37, 95, and 152).

The first EU legislation on the protection of animals during transportation, Council Directive 77/489/EC, resulted from the 1968 Convention of the Council of Europe. It has since been replaced by the more detailed Council Directive 91/628/EC as amended by Directive 95/29/EC, which introduced important changes such as the approval of transporters, the

route plan, and loading densities and traveling-time limits. EU legislation on the transportation of live animals applies to all animals transported for commercial purposes. The present framework of general provisions has been introduced by Council Directive 91/628/EEC, which in Chapter I, Part E, provides that:

- Animals shall be transported in containers, pens, or stalls appropriate for the species, complying, at least, with the most recent IATA live-animal regulations.
- Precautions shall be taken to avoid extremely high or low temperatures on board, having regard for the species of animals. In addition, severe fluctuations of air pressure shall be avoided.
- In freight aircraft, a type of instrument approved by the competent authority shall be carried for slaughtering animals if necessary.

Council Directive 86/609/EEC was issued on the protection of animals used for experimental and other scientific purposes on November 24, 1986. It sets minimal standards for housing and care and for the training of personnel handling animals and supervising the experiments, but it does not provide specific standards on transportation of animals. Council Regulation 1/2005 was passed in January 2005 and will introduce new rules that will apply directly to each member state effect 2007. These rules introduce changes to improve animal welfare and enforcement, but do not change the maximum journey times that apply through the current Directive.

TABLE 2-6 Checklist of Research Animal Regulations and Guidelines

	Yes	No
Is the species to be transported a live, warm-blooded animal other than a bird or an animal of the genus <i>Rattus</i> or <i>Mus</i> ?	Comply with AWA regulations. Subject to inspection by USDA.	No action required
Is the species (or its tissues or products) to be imported or exported considered wildlife (including all CITES-listed species and nonhuman primates)?	Obtain FWS permits, consult: http://www.fws.gov/permits/applicationmain.shtml Comply with CDC Foreign Quarantine Regulations. Comply with IATA LARs. Subject to inspection by FWS.	No action required
Is the species (or its tissues or products) to be transported (exported, imported, or interstate trade) listed as threatened or endangered by FWS? Consult: http://www.fws.gov/angered .	Obtain FWS permits, consult: http://www.fws.gov/permits/applicationmain.shtml Subject to inspection by FWS. Comply with IATA LARs.	No action required
Is an animal or its products wild-caught, listed under CITES as an Appendix I species, and being transported internationally? Consult: http://www.cites.org/eng/resources/species.html .	Obtain importation and exportation permits from the appropriate countries. Obtain FWS permits, consult: http://www.fws.gov/permits/applicationmain.shtml Subject to inspection by FWS.	No action required
Is an animal or its products to be transported internationally and • listed under CITES as an Appendix II species OR • a captive-born animal and listed under CITES as an Appendix I species? Consult: http://www.cites.org/eng/resources/species.html .	Obtain export permit from country of export. Obtain FWS permits, consult: http://www.fws.gov/permits/applicationmain.shtml Subject to inspection by FWS.	No action required
Is the species to be imported a dog or cat?	Dogs require proof of rabies vaccination. Subject to inspection by CDC.	No action required

continued

TABLE 2-6 Continued

	Yes	No
Is the species to be imported a nonhuman primate, African rodent, civet, turtle of carapace length of less than 4 in., or other species prohibited by CDC? Consult http://www.cdc.gov/ncidod/dq/animal.htm .	Obtain permit, written permission, or registration from CDC Division of Global Migration and Quarantine. Subject to inspection by CDC.	No action required
Is the species to be imported a live bat or animal, insect, or animal tissue that contains an etiologic agent?	Submit OMB Form 0920-0199 to CDC's Etiologic Agent Import Permit Program.	No action required
Does the animal or animal product to be transported contain poisonous, radioactive, or infectious material?	Contact DOT's Hazardous Materials Information Center for further requirements. If shipped by air, comply with IATA DGRs.	No action required
Are the species to be transported live, vertebrate animals that were purchased with PHS funds?	Comply with PHS Policy and <i>Guide for the Care and Use of Laboratory Animals</i> .	No action required
Is the animal to be transported within the United States a nonhuman primate, African rodent, civet, turtle of carapace length of less than 4 in., or other species prohibited by FDA? Consult http://www.fda.gov/cvm/	Submit written request for permission from Listed Animal Permit Official at FDA's Center for Veterinary Medicine.	No action required
Is the animal being imported into the United States or traveling between states?	Comply with regulations from destination state available at: http://www.aphis.usda.gov/vs/sregs Comply with CDC Foreign Quarantine Regulations. Wildlife-specific state regulations are available at: http://offices.fws.gov/statelinks.html	No action required
Is the animal being transported by air?	Comply with IATA LARs (all species) Comply with 50 CFR 14 subpart J (AWA-covered species only)	No action required

Good Practices in the Transportation of Research Animals

Animals are transported to facilitate research, teaching, and training, and for breeding-colony establishment and maintenance. Stress during transportation is unavoidable and can affect the quality of ensuing research activities. However, when science-based good practices in animal handling and transport are identified and implemented, the transportation experience can be made less stressful.

There are a few publications and articles that discuss common practices for the transportation of research animals, including the *AATA Manual for the Transportation of Live Animals* (AATA, 2000), the *IATA Live Animals Regulations* (IATA, 2005), and a Report of the Transport Working Group Established by the Laboratory Animal Science Association (Swallow *et al.*, 2005). In addition, an extensive collection of scientific literature relating to the effects of transportation on agricultural animals has been produced, in part because the livestock industry often requires that animals be shipped to new locations during the production cycle, which involves social and economic pressures for the animals to arrive in optimal condition. Much of this literature is summarized in the *Guide for the Care and Use of Agricultural Animals in Agricultural Research and Teaching* (FASS, 1999).

Unfortunately, there is sparse scientific literature on the effects of transportation on most common research animals, but good practices for all research animals can be established by drawing some universal concepts from the available scientific literature and by understanding species-specific needs. Although precise engineering standards are often preferred

by human assessors, the scientific literature supports few engineering standards. This report emphasizes *science-based performance standards*, which define an outcome (such as animal well-being or safety) and provide criteria for assessing that outcome without limiting the methods by which to achieve that outcome (NRC, 1996). The use of performance standards allows researchers and shippers the flexibility to adjust their procedures to optimize animal welfare on the basis of the species being transported, the mode of transportation, and local environmental conditions.

STRESS DURING TRANSPORTATION

Although the word *stress* generally has adverse connotations, stress is a familiar aspect of life—a stimulant for some, a burden for others. Numerous definitions have been proposed for *stress*. Each definition focuses on aspects of an internal or external challenge, disturbance, or stimulus; on perception of a stimulus by an organism; or on a physiological response of the organism to the stimulus (Goldstein, 1995; Sapolsky, 1998; Selye, 1975). An integrated definition states that stress is a constellation of events including a stimulus (stressor), a reaction in the brain precipitated by the stimulus (stress perception), and an activation of the body's physiological fight or flight systems (stress response) (Dhabhar and McEwen, 1997). Transportation stressors can be physical (changes in temperature, humidity, or noise), physiological (limitation of access to food and water), and psychological (exposure to novel individuals or environments).

It is important to recognize that stress does not always have adverse consequences (Dhabhar and McEwen, 2001; Pekow, 2005), and it is often overlooked that a stress response has healthful and adaptive effects (Dhabhar and McEwen, 1996; McEwen, 2002).

Stress can be harmful when it is long-lasting and animals are unable to adapt successfully to it (Dhabhar and McEwen, 1997; Irwin, 1994; Kiecolt-Glaser *et al.*, 2002; McEwen, 2002); therefore, an important distinguishing characteristic of stress is its duration. Acute stress is defined as stress that lasts for minutes, hours, or a few days; and chronic stress as stress that persists for months or years (Dhabhar and McEwen, 1997; McEwen, 2002). Most transportation events last only a few days and are considered acute stress events. Even the transportation of animals from overseas does not take more than a few days, so there is little concern about chronic stress during transportation. However, care must be taken to minimize post-trip stress in order to ensure that animals are not chronically stressed.

Transportation of animals involves three phases or periods: pretrip, intermodal, and post-trip. During the intermodal period, trip time has a large effect on the stressfulness of the experience. Animals experience a

sudden and large stress response at the initiation of transportation. That response declines until a lower plateau is reached. Then, after a longer period, the stress of transportation gradually increases, especially if feed and water are not consumed. For small species in extreme thermal conditions, the length of time that an animal remains at a plateau of stress response before the stress of transportation begins to increase can be rather short (minutes). However, an animal with a large body store of nutrients in extreme thermal conditions may remain at that plateau of stress response for many days.

A main issue of concern during transportation is an animal's psychological experience. Normally, animals live in a uniform, familiar environment; during transport, almost every aspect of their environment changes. The transportation enclosure, motion, human handling, temperature, light, and perhaps social group mates, odors, sounds, floor surface, food and water availability, vibrations, unusual gravitational forces (such as during acceleration, braking, or turning of vehicles), and other factors all change from moment to moment. That change in multiple sensory experiences will be perceived as stressful, even under the best of conditions, for two major psychological reasons: the transportation experience is not part of the normal routine, and the animal has no control of the situation. Stress during transportation is unavoidable, so the optimal conditions for moving animals from one location to another would be those that minimize the intensity and duration of excessive stress. Reduction in the number of transportation experiences and in novelty are two ways to make transportation more predictable and to minimize stress; however, most animals will travel only once in their lifetime—from the location where they are bred to the research location. In that case, the goal is to make the single transportation experience as predictable as is practically possible, for example, by providing access to familiar bedding during transportation.

Efforts to minimize excessive stress should be implemented from the time animals are removed from their home cages in the shipping location to the time they are delivered to home cages in the receiving location. Minimizing the intensity and duration of stress in animal home cages is also important and is under the purview of animal caretakers at each institution where animals are housed. However, it must be recognized that even mild manipulations such as moving an animal from one room to another in the same animal facility have been shown to increase corticosterone levels and result in transient but marked changes in endocrine, serological, and hematological measures (Gartner *et al.*, 1980). Repeated transportation from one location to another in the same building was shown to increase numbers of sulfomucin-producing cells in mucosa of the descending colon of Sprague Dawley rats (Rubio and Huang, 1992). A simple rule of thumb for stress minimization during transportation

involves trying to mimic the animal's accustomed living conditions as closely as possible while recognizing that animals are resilient and can adapt to an array of conditions provided that their optimal living conditions are restored within a reasonable time frame.

Acute stress from successful transportation is not likely to affect the long-term health of an animal adversely, but it can substantially change important psychophysiological measures in ways that could alter the outcome of research if it is performed before these measures normalize. Most studies suggest that animal responses to transportation stress include activation of the brain, changes in behavior, neuroendocrine and peripheral endocrine responses, and activation of homeostatic mechanisms, but these responses can vary with age, species, and strain. They are generally of short duration. Some studies have attempted to define post-transportation recovery times that are required for normalization of specific measures after transport. Physiological changes due to transportation and recovery times are outlined below for the major species of research animals. Generally, physiological changes return to normal within a day or two of transportation. However, it is important to recognize that the sparse literature suggests that some psychophysiological measures may take longer to normalize after transportation and that the time until normalization can be influenced by the duration and intensity of the stress of transportation and the particular stress-responsivity characteristics of the species or strain being transported. In practice, many investigators allow 2 to 3 days to a week or more for animals to recover after transportation and to acclimate to their new environment.

Rodents

The level of plasma corticosterone, one of the principal stress hormones, increases substantially after transportation (Aguila *et al.*, 1988; Drozdowicz *et al.*, 1990; Landi *et al.*, 1982). The increase is accompanied by changes in immune characteristics, such as a decrease in splenic natural killer cell activity (Aguila *et al.*, 1988), total white cell numbers, lymphocyte counts, thymus weight (Drozdowicz *et al.*, 1990), and humoral immunity (Landi *et al.*, 1982). Body weight also decreases, even in rodents that have access to food and water during transportation (Dymsza *et al.*, 1963; Wallace, 1976; Weisbroth *et al.*, 1977). It has been suggested that normalization of most physiological changes (including corticosterone and body weight) occurs in 2 to 4 days (van Ruiven *et al.*, 1996). However, other measures may take several weeks to normalize. For example, in animals that experience a light-dark shift (as can occur during transportation between continents), the corticosterone circadian rhythm can take more than 2 weeks to resynchronize (Weinert *et al.*, 1994).

The strain of animal can also influence the magnitude of physiological changes caused by transportation. Reproduction in some strains of mice is adversely affected by transportation (Hayssen, 1998), and some murine strains (A/J, DBA/1, SWR, and other strains with different haplotypes of the H-2 histocompatibility complex) may show a greater incidence of shipping-associated development of isolated cleft palate when pregnant females are shipped during the 5 days of gestation before embryonic palate closure (Barlow *et al.*, 1975; Brown *et al.*, 1972; Gasser *et al.*, 1981). Studies have also shown that laboratory mice may be more resilient to transportation-associated stressors than wild-caught animals (Wallace, 1976).

Nonhuman Primates

There are only sparse data on the effects of transportation in nonhuman primates (Wolfensohn, 1997). One study involving owl monkeys documented the effect of international transportation on body weight (Malaga *et al.*, 1991). All animals in the study lost weight, but the amount of body weight lost was a function of age and not the length of transportation (3 to 14 days). Younger animals lost more weight than mature animals but regained more weight than adults during the 30 days after transportation, irrespective of the length of transportation.

Pregnancy outcome and reproduction rates after transportation have also been studied in nonhuman primates. Pregnancy outcomes of pig-tailed macaques, long-tailed macaques, and baboons were studied by Sackett (1981). He found that shipment during any trimester of pregnancy had no effect on the production of viable offspring versus unshipped controls. Rates of reproduction in the three species were also tracked over a period of 8 years. In general, numbers of offspring produced were unchanged after air transport and in some cases slightly greater. The only adverse effect, not present in all species, was increased latency in rebreeding after transportation.

Livestock

In cattle, as in other farm species, body temperature rises, heart and respiration rates increase, the hypothalamic pituitary adrenal axis (HPA) activates, and there is an increase in levels of nonesterified fatty acids, blood cortisol, glucocorticoid, and glucose after transportation (Marahrens *et al.*, 2003; Nyberg *et al.*, 1988; Warriss *et al.*, 1992). Creatine kinase, albumin, and total plasma protein concentrations also tend to increase with the duration of the journey (Warriss *et al.*, 1995).

In general, physiological changes are largely determined by the age of the animal. For example, transported calves that are less than 4 weeks old

do not appear to exhibit as large an HPA response as do mature cattle (Cole *et al.*, 1988; Mormede *et al.*, 1982). At 8 weeks, the response begins to change. Corticosteroids increase, but glucose is variable, either increasing or remaining unchanged (Crookshank *et al.*, 1979; Kent and Ewbank, 1983, 1986a, 1986b; Simensen *et al.*, 1980). In general, those changes return to baseline immediately after transportation (Knowles *et al.*, 1999; Warriss *et al.*, 1992), although some genotypes may have altered endocrine concentrations for months after transport (Nyberg *et al.*, 1988). Young pigs and calves have also been found to have an unstable metabolic rate after transportation, requiring 6 to 9 days to stabilize (del Barrio *et al.*, 1993; Heetkamp *et al.*, 2002; Schrama *et al.*, 1992).

The limited physiological responses observed in adult animals can be much more pronounced after extended periods of food and water deprivation. Livestock are often transported without access to food and water for safety reasons, and the longer the period of deprivation, the longer the time necessary for normalization. Extended periods (more than a day) without food and water may result in 5 days or more before normalization of physiological measures (Warriss *et al.*, 1995).

ALLOMETRIC SCALING AND IMPLICATION FOR TRANSPORTATION PRACTICES

Transported research animals vary greatly in size, from small rodents to very large sea mammals, and within each species animals can vary in size from neonates to adults. As the size of animals varies, so do the biological processes that affect transportation practices. However, variations in processes such as heat production, metabolic rate, and space requirement are not linear functions of animal size (Lindstedt and Schaeffer, 2002). In other words, an animal that is twice the size of another animal does not have twice its metabolic rate. Rather, the relationship is exponential. The term *allometric scaling* is used to describe methods of quantifying the dependence of biological processes on body mass (West *et al.*, 1997). Implicit in allometric scaling is the principle that small animals occupy more space per unit of body weight than larger animals. Small animals also produce more heat per unit of body weight than larger animals.

The relationships between surface area, metabolic rate, and space required by mammals are defined by the following allometric equations:

$$\begin{aligned} \text{Surface area (m}^2\text{)} &= 0.1 \times \text{weight}^{2/3} && \text{(Curtis, 1983)} \\ \text{Basal metabolic rate (kcal/hr)} &= 3.0 \times \text{weight}^{3/4} && \text{(Curtis, 1983)} \\ \text{Floor area (lateral recumbency, m}^2\text{)} &= 0.1 \times \text{weight}^{1/3} && \text{(Baxter, 1984)} \end{aligned}$$

The larger surface area and basal metabolic rates per unit of body weight of smaller animals means that they evaporate more water and lose more heat per unit of body weight than larger animals. The practical implication is that smaller animals are more susceptible to changes in temperature (cold or hot), wind speed, and humidity. The core temperatures of smaller animals can decrease in cold environments more quickly than those of larger animals.

Smaller animals also become dehydrated more quickly than larger animals and cannot live without water as long. That is because of the larger evaporative skin area and/or respiration rate of small animals. Some species have special adaptations to conserve water, but even among these species, the general relationships between young (smaller) and older (larger) animals apply.

Smaller animals generally have higher metabolic rates per unit of body weight than larger animals. That means that smaller animals can go without food for less time than larger animals, which, because they are larger, have relatively greater nutrient reserves.

THERMAL ENVIRONMENT

Provision of a proper thermal environment is the most important element of safe and humane animal transportation. Temperature has been implicated as the major factor in leading to animal mortality during transportation in many species (Abbott *et al.*, 1995; e.g., Bayliss and Hinton, 1990; Slanetz *et al.*, 1957). The principles of a safe thermal environment during transportation are not different from those in normal housing. The goal is to identify the range of ambient temperatures over which an animal is able to maintain a physiologically normal core body temperature. In this section, the basic principles of thermoregulation in warm-blooded animals are discussed to provide the scientific basis of the committee's recommendations and to inform the professional judgment of researchers, staff, and institutional animal care and use committees in meeting performance standards.

Principles of Thermoregulation

Warm-blooded animals are known as homeotherms because they maintain a constant body temperature through a high metabolic rate. That process keeps body temperature constant, independent of the ambient temperature. The average body temperatures of the most common research animal species are listed in Table 3-1.

The thermoneutral zone (TNZ) is the range of ambient temperatures within which an animal's metabolic rate is at a minimum and body tem-

TABLE 3-1 Thermoregulation Data on Common Research Animal Species

Species	Average Rectal or Intra-peritoneal Temperature (°C)	Reference	Thermoneutral Zone (°C) ^a	Reference
Mouse	36.5 ± 1.3	Herrington, 1939	26 to 34	Gordon, 1985; Herrington, 1940; Oufara <i>et al.</i> , 1987
Rat	36.7 ± .9	Herrington, 1940	26 to 33	Gordon, 1990; Gwosdow and Besch, 1985; Swift and Forbes, 1939; Szymusiak and Satinoff, 1981
Guinea pig	39.2 ± .7	Herrington, 1940	28 to 30	Fewell, Kang, and Eliason, 1997
Rabbit	39.5 (38.6 to 40.1)	Robertshaw, 2004	15 to 20	Brody, 1945
Hamster	36.8 ± .2	Jones <i>et al.</i> , 1976	28 to 32	Jones <i>et al.</i> , 1976
Rhesus macaque	39.1 (37.9 to 40.0)	Johnson and Elizondo, 1979	24.7 to 30.6	Johnson and Elizondo, 1979
Dog	38.9 (37.9 to 39.9)	Robertshaw, 2004	20 to 26	Brody, 1945
Pig	39.2 (38.7 to 39.8)	Robertshaw, 2004	16 to 23	Huynh <i>et al.</i> , 2005
Cat	38.6 (38.1 to 39.2)	Robertshaw, 2004	35 to 38	Adams <i>et al.</i> , 1970
Sheep	39.1 (38.3 to 39.9)	Robertshaw, 2004	21 to 25	Brody, 1945
Beef cow	38.3 (36.7 to 39.1)	Robertshaw, 2004	-18 to 23	Hahn, 1999
Dairy cow	38.6 (38.0 to 39.3)	Robertshaw, 2004	-15 to 26	Hahn, 1999
Stallion	37.6 (37.2 to 38.1)	Robertshaw, 2004	5 to 25	Morgan, 1998
Mare	37.8 (37.3 to 38.2)	Robertshaw, 2004	5 to 25	Morgan, 1998
Goat	39.1 (38.5 to 39.7)	Robertshaw, 2004	13 to 21	Brody, 1945

^aThermoneutral zones can vary by strain, age, and reproductive or health status. The measurement of an animal's thermoneutral zone may also be influenced by the room temperature and caging condition of the animal's regular housing.

^bThat results in no substantial change in core temperature over the time period indicated in parentheses. In some cases, lowest and highest tolerated ambient temperatures were determined in acclimated animals.

Lowest Tolerated Ambient Temperature ^b (°C)	Reference	Highest Tolerated Ambient Temperature ^b (°C)	Reference
-5 (3 hr)	Oufara <i>et al.</i> , 1987	34 (2 to 3hr)	Oufara <i>et al.</i> , 1987
-15 (3 hr)	Depocas <i>et al.</i> , 1957	34 (100 min)	Gordon, 1987
-20 (1.5 hr)	Huttunen, 1982	36 (30 min)	Fewell <i>et al.</i> , 1997
-10 (2 hr)	Harada and Kanno, 1975	32.2 (2 hr)	Besch and Brigmon, 1991
-30 (1hr)	Pohl, 1965	32 (60 to 80 min)	Jones <i>et al.</i> , 1976
15 (1 hr)	Johnson and Elizondo, 1979	40.0 (1 hr)	Johnson and Elizondo, 1979
-35 (30 min)	Good and Sellers, 1957	35.0 (2 hr)	Besch <i>et al.</i> , 1984
-20 (indefinitely)	FASS, 1999	35 (indefinitely)	FASS, 1999
-5 (1.5 hr)	Hensel and Banet, 1982	35 (1.5 hr)	Adams <i>et al.</i> , 1970
—	—	—	—
—	—	—	—
—	—	—	—
—	—	—	—
—	—	—	—
-13 (indefinitely)	Schaeffer <i>et al.</i> , 2001	—	—

perature is maintained solely through autonomic responses (piloerection and peripheral vasomotor tone) and behavioral responses (adjusting posture to bring limbs close to or away from body) (Bligh and Johnson, 1973). In Figure 3-1, the committee has modeled the relationship between metabolic rate and ambient temperature in homeotherms. The TNZ is represented in Figure 3-1 as the area between C, the lower critical temperature, and D, the upper critical temperature (UCT). This zone is narrow in some species, particularly the smaller research animals (see Table 3-1). The midpoint of the TNZ is usually 7 to 10°C below normal rectal temperature and 5°C below normal skin temperature (Brody, 1945).

The TNZs for rats and mice (Table 3-1) vary in the scientific literature because TNZ estimates depend on the environments in which the animals are housed and assessed. TNZs can also vary considerably with age and reproductive status, as illustrated for chickens in Figure 3-2 and several other livestock species in Figure 3-3. Comparable data on commonly used laboratory species are not available.

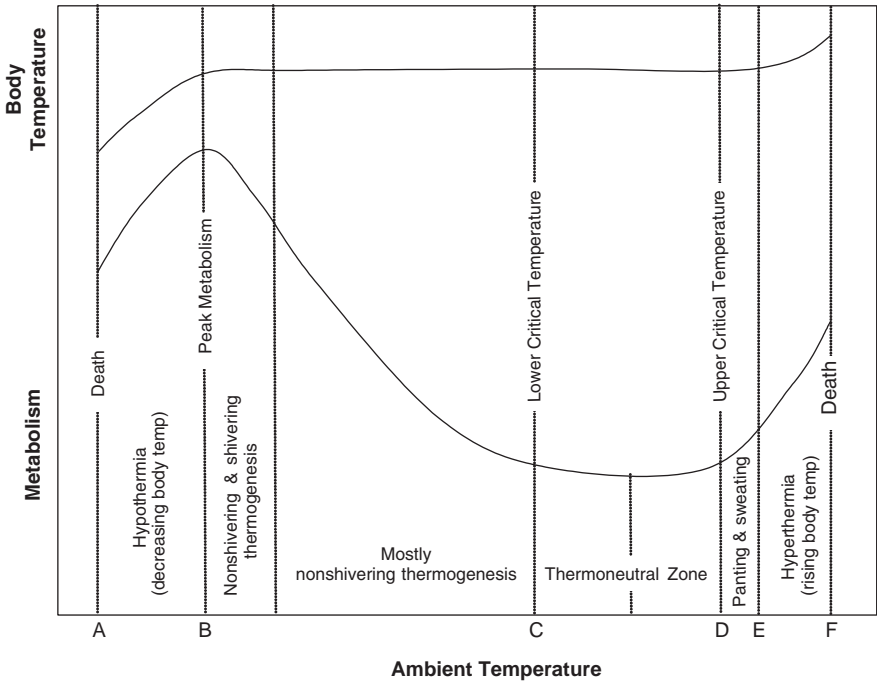


FIGURE 3-1 Graph representing relationship between metabolic rate and ambient temperature in homeotherms.

When the ambient temperature falls below the TNZ, physiological mechanisms collectively referred to as nonshivering thermogenesis (oxidation of fatty acids and brown adipose tissue) are initiated; they increase metabolic rate, balance heat production and loss, and maintain body temperature (Robertshaw, 2004). As ambient temperatures continue to fall, nonshivering thermogenesis is no longer adequate to offset heat loss and maintain body temperature. Shivering thermogenesis (involuntary contractions of skeletal muscles) then occurs and further increases heat production (Robertshaw, 2004). As ambient temperatures continue to decrease, heat production through nonshivering and shivering thermogenesis reaches the maximum rate that can be sustained over long periods. This point of maximal heat production is known as peak metabolism (B in Figure 3-1) (Robertshaw, 2004). The peak metabolic rate is 3 to 4 times the basal metabolic rate in most species (Brody, 1945). If the ambient

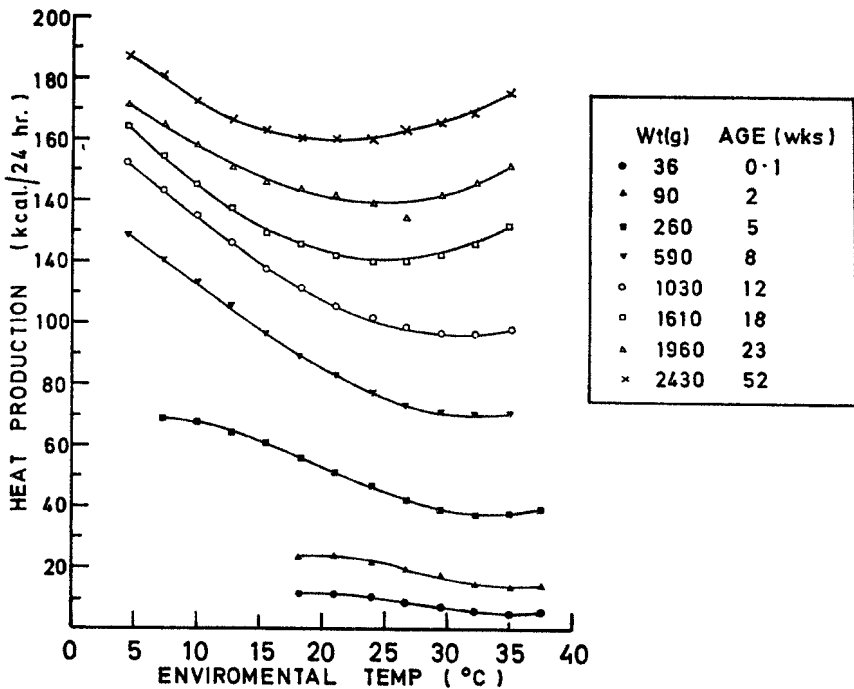


FIGURE 3-2 Changes in thermoneutral zone (range of ambient temperatures at which an animal's heat production is at a minimum) with age and size in chickens. TNZ of chickens shifts to lower temperatures as chickens grow and age. Reprinted from Fuller, 1969.

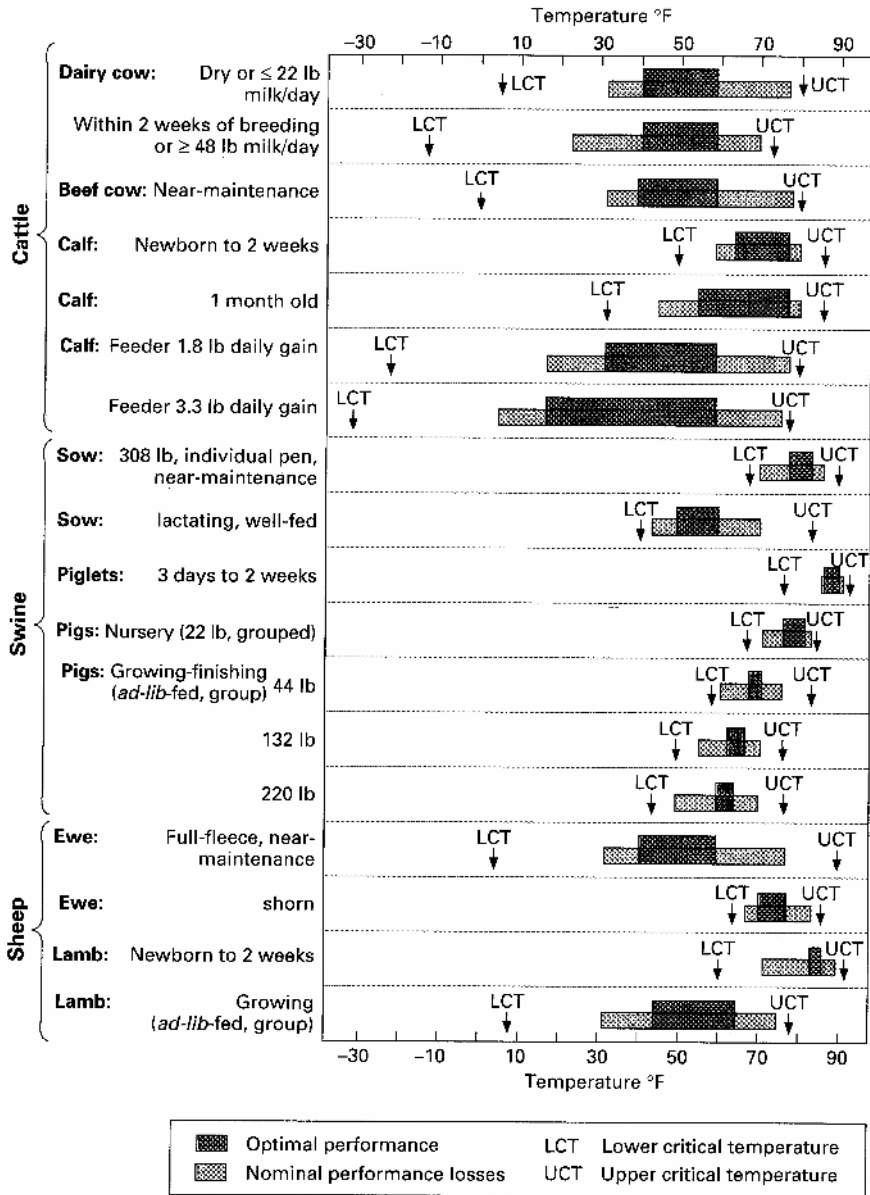


FIGURE 3-3 TNZ of various agricultural animals. Reprinted with permission from Taylor, R.W., and T.G. Field. 2004. *Scientific Farm Animal Production*, 8th ed. Upper Saddle River, NJ: Prentice Hall; p352. ©2004 Pearson Education, Inc.

temperature decreases further (below B in Figure 3-1), heat production decreases, body temperature decreases (hypothermia), and death eventually occurs. Animals can boost exertions at peak metabolic rates 10 to 15 times their basal metabolic rates during periods of high exertion (Robertshaw, 2004), as occurs in humans during a 100-m race. However, these peak metabolic rates cannot be sustained and are of little value in maintaining body temperature during prolonged cold exposure.

The range of ambient temperatures over which an animal can maintain its core body temperature is depicted in Figure 3-1 between B and E. The lowest ambient temperature at which an animal can maintain its body temperature is generally much lower than its TNZ (see Table 3-1), but the highest ambient temperature at which an animal can maintain its body temperature is generally close to the upper limit of its TNZ. This is because most research animals are nonsweating species (Brody, 1945) and have a limited capacity for dissipating heat. When the ambient temperature rises above the upper limit of the TNZ, heat is dissipated by increasing radiant heat loss (through increased peripheral vascular flow) and evaporative heat loss (through panting, sweating, and saliva spreading). When the ambient temperature is at body temperature, there is no radiant heat loss. Panting can still dissipate heat when ambient temperature reaches body temperature, but it has less capacity to dissipate heat because the energy expended (and heat produced) as panting increases eventually offsets the heat lost through panting (Brody, 1945).

Sweating provides the largest capacity for heat dissipation in homeotherms, so it is important to recognize that most common research animals are nonsweating species and do not have the same capacity to dissipate heat as profusely sweating species such as humans and horses. For example, at an ambient temperature of about 36°C, all of the heat produced by a human can be dissipated through sweating, but only 30 to 40% of the heat produced by nonsweating species—such as rodents, cats, dogs, rabbits, swine, sheep, and cattle—can be so dissipated (Brody, 1945). It should be noted that most nonhuman primates—rhesus macaques are an exception (Johnson and Elizondo, 1979)—have not been observed to sweat over their general body surface and should be considered nonsweating species for the purposes of determining safe ambient temperatures for transportation (Stitt and Hardy, 1971).

Animal handlers must be acutely aware that although they as humans may be able to remain reasonably comfortable because sweating reduces their heat load, the nonsweating animals they care for are not able to remain comfortable through sweating. That must be emphasized during the training of handlers, because temperatures well within human tolerance zones can be deadly to many species of research animals (Ohara *et al.*, 1975; Oufara *et al.*, 1987; Wright, 1976).

Safe Temperature Ranges During Transport

An animal must be transported in an environment in which it can maintain its body temperature, becoming neither hyperthermic nor hypothermic. Due to the many and often interacting physiological and environmental variables that can affect body temperature, it is impossible to specify exact ambient temperature ranges that guarantee that animals will remain homeothermic during transportation. The thermal requirements of animals vary widely depending on age, physiological state, and the environment in which they were previously housed and to which they have adapted. For example, newborns are less tolerant of thermal extremes than adults, and late-pregnancy and lactating animals are less tolerant of heat. In addition, animals previously maintained in cold environments with high feed intakes and high rates of resting heat production (Young, 1975) do not tolerate heat as well as animals previously maintained in a hot environment with low feed intakes and low metabolic rates (NRC, 1981). The effect of the higher rates of heat production associated with pregnancy, lactation, and cold acclimation is an increase in the amount of heat that animals must dissipate as environmental temperatures approach and exceed body temperature, that is, a reduction in heat tolerance.

It could be suggested that homeothermic animals be transported in ambient temperatures within their TNZ. The range of the TNZ for most agricultural animals is large, allowing for safe transportation over a relatively wide range of ambient temperatures (see Table 3-1). The Livestock Weather Safety Index was developed to guide decision making for transporting swine during weather extremes (Livestock Conservation Institute, 1970). More information about the Livestock Weather Index can be found at the National Institute for Animal Agriculture (formerly known as the Livestock Conservation Institute) website:

<http://animalagriculture.org/>

The TNZ for the more common research animals (rodents, cats and dogs, and nonhuman primates) is narrower than in livestock. However, upper limit of the safe range for transport of those animals is also based on the UCT of an animal's TNZ. Unlike humans, who can maintain their body temperatures above their TNZ through sweating, most research animals are nonsweating species and have a limited capacity for dissipating heat.

Though the highest ambient temperature at which those animals can maintain their body temperature is close to the upper limit of its TNZ, the lowest ambient temperature at which those animals can maintain their

body temperature is generally much lower than its TNZ. In order to determine the lower limit of the range of ambient temperatures for safe transportation, the committee turned to the acclimation literature. It is well established in the cold-acclimation literature that the more common research animals can be housed in conditions around 6°C for weeks or months with no ill effects (Besch *et al.*, 1984; Depocas *et al.*, 1957; Gordon, 1988; Whiting and Brandt, 2002). Therefore, the lower limit of the temperature range for safe transport was defined as the lowest temperature to which a species had been successfully acclimated (see Table 3-2).

The temperature ranges in Table 3-2 should be considered only as a general guideline. In many cases, air temperature alone is insufficient to determine whether a particular animal or species can be transported safely. Wind chill, sun exposure, and particularly humidity can greatly influence the temperature that an animal effectively experiences (see Table 3-3). For example, air feels colder with even a slight wind. In addition to environmental factors, physiological factors can affect the ambient temperature at which an animal can maintain its core body temperature. These factors include physiological states such as late pregnancy and lactation, previous acclimation, disease status, hydration state, feeding level, exercise, anesthesia, and body fat.

Nevertheless, short-term exposures to thermal extremes during transportation do not generally result in adverse physiological effects except when the short-term temperature exposures are particularly high. As documented in Table 3-1, most common research animal species are able to maintain their core temperature over the course of several hours at extreme temperatures. Even animals with narrow TNZs (e.g., Xin, 1997) can tolerate high temperatures and large fluctuations in temperature over extended periods if provided appropriate food and hydration. **The committee recommends that professional judgment be considered the final determinant of whether the ambient temperatures that animals will be exposed to during transportation are safe.** Many factors must be considered if professional judgment is utilized, including humidity, stocking density, the characteristics of the transportation cage, plumage and hair coat, previous adaptation, metabolic and behavioral characteristics, physiologic status, food and water consumption, trip length, and potential temperature extremes.

The effects of physiological and environmental factors on the effective temperature that an animal experiences during transportation have been well documented in birds and can be generalized to most other species. Birds with feather loss or poor feather condition lose insulating capability. This loss can be detrimental under cold conditions and beneficial in hot conditions. Likewise, thin birds are less able to maintain their body temperature in cold conditions than birds with more adequate body

TABLE 3-2 Ambient Temperature Range for Safe Transportation of Common Adult Research Animals^a

Species	Ambient Temperature Range for Safe Transportation ^b (°C)	References
Mouse	4 ^c to 34	Oufara <i>et al.</i> , 1987
Rat	6 ^c to 33	Depocas <i>et al.</i> , 1957
Guinea pig	4 ^c to 34	Himms-Hagen <i>et al.</i> , 1995
Rabbit	4 ^c to 33 ^c	Cooper <i>et al.</i> , 1980; Honda <i>et al.</i> , 1962
Hamster	6 ^c to 34 ^c	Jones <i>et al.</i> , 1976; Pohl, 1965
<i>Macaca mulatta</i>	6 ^c to 35	Oddershede and Elizondo, 1980, 1982
Dog	-10 ^c to 28	Nagasaka and Carlson, 1965
Pig	-20 to 35 ^d	FASS, 1999
Cat	5 ^c to 30 ^c	Adams, 1963; Hensel and Banet, 1982
Sheep	-12 ^c to 25	Horton, 1981
Beef cow	-18 to 23	
Dairy cow	-15 to 26	
Stallion	5 to 25	
Mare	5 to 25	
Goat	-13 ^c to 21	Schaeffer <i>et al.</i> , 2001

^aHumidity, wind chill, sun exposure, hydration state, physiological state, age, acclimation, and so on, can greatly influence these ranges (see Table 3-3). Professional judgment must be used in determining safe transportation of research animals.

^bThe maximum and minimum temperatures of the range were derived from the upper limit and lower limit, respectively, of that species thermoneutral zone as described in Table 3-1, unless otherwise noted. Most larger mammals can be transported when the temperature is below freezing as long as the temperature inside the transport compartment does not cause frostbite or other signs of extreme cold. Conditions inside transport compartments, especially warm, deep bedding will allow animals to establish a microenvironment that is comfortable. Professional judgment should be used to assess risks to animal welfare when animals are preconditioned or not preconditioned for transport.

^cTemperature derived from the lowest or highest temperature to which that species has been acclimated. It is possible that animals could be safely transported at more extreme temperatures; however, the literature neither supports nor negates the possibility.

^dRecommended thermal conditions for swine.

conditions (Schrama *et al.*, 1996). The age of the animal also influences the thermal conditions that are suitable for safe transport. As shown in Figure 3-2, young birds have a narrower TNZ than mature birds. Thus the temperature at which thermogenesis begins is higher in younger birds, making these animals more susceptible to cold.

The phenotypes of some animal strains and transgenic animals must also be considered when the animals have abnormal metabolic characteristics. Some pigs carry a mutation in a gene that causes malignant hyper-

TABLE 3-3 Effects of Various Factors on Effective Environmental Temperature and Relative Risk to Animal Health and Welfare

Factor	Very Cold Outside Temperature	Risk	Very Hot Outside Temperatures	Risk
High relative humidity	Little effect	Low	Much warmer	Very high
High air velocity	Much colder	High	Feels colder as long as air temperature is below animal core temperature	Low
Food deprivation	Much colder	High	Little effect	Low
Water deprivation	Colder	Medium	Much warmer	High
High stocking density	Warmer	Zero or positive effect	Much warmer	Very high
Illness (fever)	Colder	Medium	Warmer	Medium
Fat (high subcutaneous insulation)	Warmer (protective)	Low	Warmer	High
Stress susceptible genotypes ^a	Colder	High	Warmer	Very high
Surface cover (plumage and hair coat)	Warmer	Protective	Warmer	High
Previous experience or adaptation to cold temperatures	Warmer	Zero or positive effect	Warmer	High
Previous experience or adaptation to hot temperatures	Colder	High	Colder	Zero or positive effect

^aMay include malignant hyperthermia or transgenic animals that have thermoregulatory or physiological dysfunction.

SOURCE: Table adapted from Schrama *et al.*, 1996.

thermia when stress is experienced (McGlone and Pond, 2003). These animals become warm and red-skinned during stress and develop muscle tremors and an inability to walk. In some instances, animals may die as a result of malignant hyperthermia.

Behavior can affect the thermal experience. The stocking density of birds within the transportation crate may inhibit or encourage thermal regulating behaviors, such as stretching and fanning wings during heat episodes or huddling close to other birds under cold conditions. Posthatch-fasted male chicks held in shipping containers exhibited dispersal behaviors at 35°C (Xin and Harmon, 1996), and birds transported at high densities during hot weather are more prone to heat stress (Schrama *et al.*, 1996). As chicks were subjected to lower temperatures, huddling became more evident, and the most huddling occurred at 20°C (Xin and Harmon, 1996). Those behaviors also altered moisture production within the container; there was more moisture loss from animals as they spread out at higher temperatures.

Safe temperature ranges for transportation are more difficult to establish for poikilotherms such as reptiles and amphibians. In poikilotherms, a decrease or increase in body temperature of a few degrees is not a cause for concern. Animal activity and alertness may be better indicators of comfort temperature than body temperature (which varies considerably with air temperature).

Effect of Transportation Caging on Thermal Environment

A major factor that can influence an animal's effective environmental temperature is the nature of its transportation cage. Transportation containers for small research animals (such as rodents and chicks) are almost always stacked. That feature potentially restricts air flow into or around the containers and can increase the temperature in a container to exceed that of the surrounding environment. For example, when a commonly used commercial chick container was stacked six high in four stacks with 2.5 cm of vertical distance between containers and 5.1 cm between the stacks, the temperature inside the containers was about 5.5 to 10°C above the ambient temperature (Tanaka and Xin, 1997). The IATA Live Animals Regulations (Container Standard #84) and the Animal Welfare Act (9 CFR 3.14(c)(2), 9 CFR 3.36(a)(6), 9 CFR 3.61(a)(5), 9 CFR 3.87(c)(2), 9 CFR 3.137(a)(5)) provide guidance on using spacers or projecting rims in order to prevent obstruction of the ventilation openings and provide space for air circulation.

Transportation containers with biocontainment filters can similarly restrict air flow into a container, increasing the container temperature. In some facilities, rodent transportation caging is autoclaved for reuse. It has

been suggested that such autoclaving may increase the air resistance of the biocontainment filter and restrict air flow (White, 2004). Until it can be established that reautoclaving biocontainment filters does not restrict air flow below acceptable levels, the committee suggests that the prudent course of action is to avoid reautoclaving and reuse of the containers.

Behavioral Monitoring of Thermal Environment

Ideally, all animals should be transported in environmentally controlled vehicles. That method would eliminate concerns about thermal stress. However, such transportation is often unavailable, particularly for nonrodent species, and other precautions and procedures, such as monitoring animal behavior, must be used to ensure the animals' welfare.

As discussed above, when ambient temperatures change, animals use both physiological and behavioral mechanisms either to increase heat production or to promote heat loss and maintain a homeothermic state. Evidence of those mechanisms does *not* necessarily indicate that an animal has become hypothermic or hyperthermic. However, prolonged display of the behaviors listed in Table 3-4 is an indication that the animal is stressed and may not remain homeothermic. Although the temperature ranges in Table 3-2 can generally be considered safe for the transportation of research animals, many factors can affect the effective ambient temperature (see Table 3-3). **Therefore, when animals cannot be transported in environmentally controlled vehicles, the committee recommends frequent visual inspection of the animals when practical, as signs of thermogenesis or heat loss may indicate that the animal's thermal environment should be adjusted.** Exposure to unaccustomed temperature extremes, particularly high temperature, can be more stressful, harmful, and deadly than suspected by researchers and staff who are accustomed to handling and observing animals in controlled housing conditions.

Thermal Acclimation

When it is anticipated that an animal will encounter extreme temperatures during transportation, as may occur during loading of animals onto airplanes during winter and summer in some parts of the United States, prior acclimation may be appropriate, if practical. During acclimation, prolonged exposure to a single component of the environment—in this case either heat or cold—results in physiological changes that allow an animal to respond more effectively to that component (Robertshaw, 2004). For example, rats acclimatized to 6°C can maintain their body temperature when exposed to -15°C for a period of at least 3 hr (Gordon, 1990).

TABLE 3-4 Behavioral and Physiological Signs of Thermal Status

Species	Signs of Thermogenesis	Signs of Heat Loss
Rodents	Piloerection Cutaneous vasoconstriction (paleness of skin of ears or feet) Shivering Drawing limbs close to body (curling up)	Saliva spreading Cutaneous vasodilation (redness of skin of the ears or feet) Closed-mouth panting (increased respiratory frequency)
Dogs and cats	Piloerection Cutaneous vasoconstriction (paleness of skin of ears or feet) Shivering Drawing limbs close to body (curling up)	Open-mouth panting Extending limbs (maximizing surface area) Cutaneous vasodilation (redness of skin of ears or feet)
Agricultural animals (cows, sheep, goats, and horses)	Piloerection Cutaneous vasoconstriction (paleness of skin of ears or feet) Shivering Drawing limbs close to body (curling up)	Sweating Closed-mouth panting (increased respiratory frequency)
Birds	Piloerection Cutaneous vasoconstriction (paleness of skin of ears or feet) Shivering Drawing limbs close to body (curling up)	Open-mouth panting

SOURCE: Adapted from Robertshaw, 2004.

SPACE ALLOCATION

The need for space allocations during transportation of research animals is the subject of public concern and regulations—but few scientific studies. The space needs of animals during transportation are different from their space needs in resident housing. Space needs also vary with animal temperament, social relationships, thermal environment, and species-specific behavioral requirements.

It is clear that the floor space required during transportation is different from the floor space required during long-term housing. In conventional housing, there are often few adverse effects of providing too much space, but this is not the case during transportation. If transported animals have too much space, they can fall, injure themselves, or even be killed.

Animals should never be in a situation in which they may come into contact with container walls with force or roll around. At the other end of the spectrum, animals should never have so little space that they pile on top of one another; this situation can cause animal injury and may potentially lead to suffocation.

Although smaller animals occupy less space than larger animals, they occupy more space per unit of body weight. Thus, 10 1-kg animals require more space than one 10-kg animal if other considerations are equal. However, the space that animals occupy depends on their posture. All species commonly adopt distinct postures during transportation. For example, horses and adult cattle stand during the entire trip, but rodents, pigs, young calves, dogs, and cats lie down during stable parts of the trip. The order of space occupancy from greatest to least is lateral recumbency, sternal recumbency, sitting, and standing. The space needs of individual animals also depend on whether they are transported alone or in groups and on whether they normally stand or lie down during transportation. The personnel responsible for placing animals in transportation caging must be familiar with the normal behavior of each species to assess the adequacy of the floor space provided.

Determining the appropriate density of animals in a transportation cage or vehicle must take into account weather conditions, the physical characteristics of the species (such as horns, pilage condition), and the preferred posture, if any, adopted during transportation. Simply providing transported animals large amounts of space may not be conducive to their welfare in all instances. For example, low stocking densities can present problems with balance during transportation. Many species, including cattle, rarely adopt a vulnerable posture such as lateral recumbency. If they remain standing, those animals may fall down as the transportation vessel experiences movement in different directions. Even if they eventually adopt a sternal recumbent posture, they can fall or roll in the transportation compartment and be injured. Cattle tend to prefer standing, so it is imperative that they can adjust footing or brace against other cattle to prevent slipping and falling.

Some people have suggested that a high stocking density is preferable for horses and cattle because the animals "hold each other up," preventing injuries due to falls or rolls (Friend, 2001). However, studies of stocking density indicate that high stocking densities are associated with a higher rate of injury (Friend, 2001). Pushing and mounting behaviors tend to increase with stocking density (Tarrant *et al.*, 1992), which can lead to injury. Also, the ability of animals to rise after a fall is hampered at high stocking densities, leading to more injuries and a greater severity of injury (Collins *et al.*, 2000). In addition, high stocking densities decrease social interactions among animals and may prevent them from assuming a

preferred orientation (parallel or perpendicular to motion) during travel (Eldridge *et al.*, 1988; Kenny and Tarrant, 1987; Lambooy and Hulsegge, 1988; Tarrant *et al.*, 1992). Based on the literature, a moderate stocking density for cattle and horses maximizes animal welfare (Swanson and Morrow-Tesch, 2001; Tarrant and Grandin, 2000).

There is abundant literature on the space requirements or stocking densities necessary to optimize the welfare of agricultural animals during transportation, but none on the most common species of research animals—rats and mice. Rodent vendors have developed space allowances for rodents on the basis of practical experience. When a regression of the space allowances for agricultural animals and rodents was performed (Table 3-5), the trend line (Figure 3-4) had a very high coefficient of determination ($r^2 = 0.9915$). That value suggests that there is a mathematical algorithm that describes the transportation space required to maximize the well-being of group-transported animals, and it provides information on the transportation space required for unusual research animals for which space requirements are unknown. Transportation space requirements for guinea pigs and hamsters mandated in the Animal Welfare Act also follow the trend line; although they are not obviously based on empirical data, those space requirements might be appropriate. The algorithm would be useful to people who are attempting to determine the transportation space needs for an uncommon species of research animal for whose transportation there are neither guidelines nor much practical experience.

FOOD AND WATER

Most animals react to the experience of being transported by becoming anorexic and adipsic. The stressful experiences of a novel environment, movement of the transportation vehicle, and food and water sources that differ from those in the animal's previous environment for logistical reasons inhibit food and water consumption. However, animals lose weight more rapidly when transported than they would normally during the same period without feed and water. That consequence implies that transportation is stressful for reasons beyond the lack of feed and water.

Provision of feed or water during transportation can be problematic because of food spoilage and water spillage; wetting of the floor by spilled water, which results in chilling, slipping, and injuries; animals' lack of ability to eat or drink while in motion; motion sickness; and lack of motivation to eat or drink during the trip. Thus, providing food or water may not be of any benefit during short trips because of lack of motivation to consume food and water. Provision of feed and water during very long trips requires special attention, especially if the vehicle stops or has periods of stability during which animals may seek food and water. In

TABLE 3-5 Space Allowances for Group-Transported Animals^a

Species	(lb)	(kg)	(ft ²)	(m ²)	Source
Mice	0.053	0.024	0.09	0.008	Harlan ^b
Mice	0.055	0.025	0.04	0.004	Jackson Laboratories ^b
Mice	0.075	0.034	0.10	0.009	Harlan ^b
Mice	0.077	0.035	0.07	0.006	Charles River ^b
Gerbils	0.077	0.035	0.08	0.007	Charles River ^b
Gerbils	0.110	0.050	0.11	0.010	Charles River ^b
Gerbils	0.132	0.060	0.13	0.012	Charles River ^b
Gerbils	0.154	0.070	0.18	0.017	Charles River ^b
Rats	0.110	0.050	0.13	0.012	Charles River ^b
Rats	0.110	0.050	0.10	0.009	Taconic ^b
Rats	0.163	0.074	0.16	0.015	Harlan ^b
Rats	0.165	0.075	0.16	0.015	Charles River ^b
Rats	0.165	0.075	0.11	0.010	Taconic ^b
Rats	0.218	0.099	0.19	0.018	Harlan ^b
Rats	0.220	0.100	0.21	0.019	Charles River ^b
Rats	0.220	0.100	0.12	0.011	Taconic ^b
Rats	0.273	0.124	0.22	0.020	Harlan ^b
Rats	0.276	0.125	0.27	0.025	Charles River ^b
Rats	0.276	0.125	0.13	0.012	Taconic ^b
Rats	0.328	0.149	0.25	0.023	Harlan ^b
Rats	0.331	0.150	0.30	0.028	Charles River ^b
Rats	0.331	0.150	0.15	0.014	Taconic ^b
Rats	0.384	0.174	0.29	0.027	Harlan ^b
Rats	0.386	0.175	0.18	0.017	Taconic ^b
Rats	0.441	0.200	0.33	0.031	Charles River ^b
Rats	0.494	0.224	0.35	0.032	Harlan ^b
Rats	0.505	0.229	0.43	0.040	Harlan ^b
Rats	0.551	0.250	0.44	0.041	Charles River ^b
Rats	0.551	0.250	0.24	0.023	Taconic ^b
Rats	0.606	0.275	0.29	0.027	Taconic ^b
Rats	0.661	0.300	0.53	0.050	Charles River ^b
Rats	0.717	0.325	0.36	0.034	Taconic ^b
Rats	0.882	0.400	0.67	0.062	Charles River ^b
Rats	0.882	0.400	0.49	0.045	Taconic ^b
Rats	0.992	0.450	0.89	0.083	Charles River ^b
Hamsters	0.110	0.050	0.11	0.010	Charles River ^b
Hamsters	0.132	0.060	0.13	0.012	Harlan ^b
Hamsters	0.176	0.080	0.13	0.012	Charles River ^b
Hamsters	0.220	0.100	0.15	0.014	Harlan ^b
Hamsters	0.287	0.130	0.17	0.015	Harlan ^b
Guinea Pigs	0.549	0.249	0.27	0.025	Harlan ^b
Guinea Pigs	0.769	0.349	0.33	0.031	Harlan ^b
Guinea Pigs	0.772	0.350	0.27	0.025	Charles River ^b
Guinea Pigs	1.210	0.549	0.44	0.041	Harlan ^b
Guinea Pigs	1.323	0.600	0.44	0.041	Charles River ^b
Guinea Pigs	1.764	0.800	0.53	0.050	Charles River ^b

continued

TABLE 3-5 Continued

Species	(lb)	(kg)	(ft ²)	(m ²)	Source
Rabbits	7.915	3.59	1.44	0.134	Harlan ^b
Swine	10.00	4.54	0.70	0.065	Whiting and Brandt, 2002
Swine	20.00	9.07	0.90	0.084	Whiting and Brandt, 2002
Swine	30.00	13.60	1.00	0.093	Whiting and Brandt, 2002
Swine	50.00	22.70	1.50	0.139	Whiting and Brandt, 2002
Swine	60.00	27.20	1.70	0.158	Whiting and Brandt, 2002
Swine	70.00	31.20	1.80	0.167	Whiting and Brandt, 2002
Swine	80.00	36.30	1.90	0.177	Whiting and Brandt, 2002
Swine	90.00	40.80	2.10	0.195	Whiting and Brandt, 2002
Swine	100.00	45.40	2.20	0.204	Whiting and Brandt, 2002
Swine	110.00	49.90	2.30	0.214	Whiting and Brandt, 2002
Swine	120.00	54.40	2.50	0.232	Whiting and Brandt, 2002
Swine	130.00	59.00	2.60	0.242	Whiting and Brandt, 2002
Swine	140.00	63.50	2.80	0.260	Whiting and Brandt, 2002
Swine	150.00	68.00	2.90	0.269	Whiting and Brandt, 2002
Sheep (Full Fleece)	60.00	27.00	2.20	0.210	FASS, 1999
Sheep (Full Fleece)	80.00	36.00	2.60	0.240	FASS, 1999
Sheep (Full Fleece)	100.00	45.00	3.00	0.270	FASS, 1999
Sheep (Full Fleece)	120.00	55.00	3.40	0.310	FASS, 1999
Calves	200.00	91.00	3.50	0.320	FASS, 1999
Calves	300.00	136.00	4.80	0.460	FASS, 1999
Calves	400.00	182.00	6.40	0.570	FASS, 1999
Calves	600.00	273.00	8.80	0.800	FASS, 1999
Cattle (Horned)	800.00	364.00	10.90	1.000	FASS, 1999
Cattle (Horned)	1,000.00	455.00	12.80	1.200	FASS, 1999
Cattle (Horned)	1,200.00	545.00	15.30	1.400	FASS, 1999
Cattle (Horned)	1,400.00	636.00	19.00	1.800	FASS, 1999

^aMore space may be given during transportation than is listed, but more floor space increases the risk of animal injury. More space per animal is needed in warm weather and during long trips (over 48 hr; FASS, 1999). Space allowances are to be tempered with professional judgment to accommodate strains, species, thermal conditions, special models, and protocol requirements.

^bSpace allowances calculated from caging density and cage specification data available in corporate literature.

cases where an animal may refuse food because it is presented in a novel form or source, animals should be adapted to the travel and post-travel diets and to feed and water dispensers before travel. Exposure to the food forms and water sources that will be used during travel before the trip may help to reduce dehydration and weight losses during transportation.

Small animals (young animals or small animal species of any age) can survive less time without food and water than larger animals. Water is the

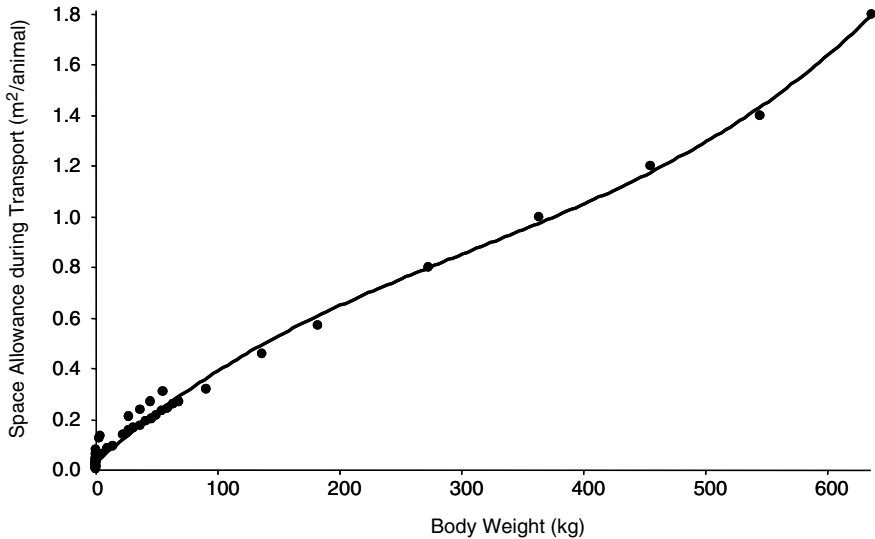


FIGURE 3-4 Space allowances during transportation. Based on transportation space allowances in Table 3-5. Second-order polynomial regression resulted in trend line ($y = 8^{-9}x^3 - 8^{-6}x^2 + 0.0043x + 0.0302$) with a coefficient of determination (r^2) of 0.9929.

most important consideration for trips of intermediate length for most species. Small animals lose more heat, require more calories per unit of body mass, and become dehydrated more quickly than larger animals. Schlenker and Muller (1997) identified the duration of water and food deprivation as factors in the high mortality of air-shipped chicks. Post-hatching metabolic changes and physical development of chicks exacerbate the development of pathological conditions.

In most cases, small animals (less than 1 kg) will require a source of food and water during transportation that lasts more than a few hours. Several commercially available gel moisture sources have been developed to provide an alternative to the use of water bottles during transportation (Maher and Schub, 2004). These gel moisture sources provide uniform, spill-proof, and contamination-free hydration for rodents; however, they are not nutritionally complete, and a food source should also be utilized during transportation. Xin and Lee (1996) found that the provision of water (or a substitute) and feed were also important for sustaining male day-old chicks during long trips (experiments were conducted under simulated conditions for a duration of 72 hr).

Larger animals can go longer without food or water without ill effects. Studies indicate that only after 24 hr of road transportation does a lack of water and physical fatigue become detrimental to cattle welfare (Knowles *et al.*, 1997; Tarrant and Grandin, 2000). Cattle are typically fasted for 6 to 12 hr before transportation (Lapworth, 2004), and this state must be considered when assessing the physical condition of cattle during transportation. The primary reason for fasting is to limit manure accumulation in the trailer and thus prevent slipping and falling.

Horses can also experience dehydration after 24 hr of transportation (Friend, 2000; Friend *et al.*, 1998; Stull and Rodiek, 2000). Friend *et al.* (1998) found that horses transported for long distances during hot weather drank less water (20.9 L) than horses penned under similar conditions (38.2 L). In a later study, Friend (2000) found that respiration, heart rate, blood sodium, osmolality, and chloride were significantly higher in nonwatered horses after 30 hours of transportation in hot conditions (indicating dehydration), than in horses that had received water during similar transport. However, offering of water to horses transported under cool conditions appeared to result in no added benefit to their well-being. Stull and Rodiek (2000) assessed the condition of show horses transported in a commercial van during the summer. They, too, concluded that after 24 hr of transportation, horses begin to show changes in physiological markers of hydration. In general, it appears that horses in good physical condition can be safely transported in hot weather for at least 24 hr, when provided with water.

Estimating the amounts of food and water that should be placed in the enclosure during transportation is relatively simple. Initially, the caloric and water requirements of the species must be determined. That information can be found in the *Nutrient Requirements of Domestic Animals* series, a group of reports from the National Academies that cover farm animals, laboratory species, wildlife, and companion animals. When the minimal requirements have been determined, several other factors must be considered, including (Wallace, 1976):

- Expected duration of the journey;
- Initial weight and life stage of the animal (for example, caloric and water requirements vary with age);
- Special requirements of the species or strain of animal (for example, some transgenic animals may have altered nutritional or caloric requirements); and
- Expected environmental conditions (for example, animals may consume more water in low-humidity environments).

SOCIAL INTERACTION AND GROUP TRANSPORTATION

Animals can be transported in individual or group enclosures (caging or vehicles). Isolation, such as during transportation, can minimize social stress in solitary animals and species, but isolation can induce stress in social species (Tamashiro *et al.*, 2005). New social groups of nonsocial animals constitute a stressor, and these animals should be transported individually. Socially dominant pigs are less adversely influenced by the stress of transportation than are socially intermediate or submissive pigs (McGlone *et al.*, 1993). Some animals, particularly large ones, are aggressive and are best transported alone or with conspecifics in sensory but not physical contact. Baldock and Sibly (1990) found that spatial isolation (4 to 90 m) alone did not have a substantial effect on heart rate in sheep, but that visual isolation produced a substantial increase in heart rate, vocalization, and activity within the first 5 min of treatment.

For prey species such as sheep, being shipped near a predator species such as a dog is especially stressful and should be avoided. In contrast, having familiar conspecifics in the same compartment reduces the stress of a new experience. Most laboratory and farm animals are social animals, and they are often housed in compatible social groups at the site of trip origin. If social groups are transported, it is recommended that the groups be established before transportation where appropriate so that dominance orders will not need to be established during or after transportation. However, it has been found that rats adapt quickly to unfamiliar social environments (Sharp *et al.*, 2005) and unfamiliar social environments have no negative effect when chickens travel together in the same shipping enclosure (Knowles and Broom, 1990). The performance standard for social interaction is the lack of social aggression and injury resulting from aggressive social interactions.

HANDLING

An animal's experience can greatly affect its response to the transportation environment. Animals can be preconditioned to transportation by being exposed to the transportation container and the food or water that will be available during transportation. In addition, frequent human handling before the handling associated with transportation will help animals to respond better to the transportation experience. Animals that have been socialized with people and have been handled respond more favorably to the handling associated with transportation than those not similarly exposed. In many cases, preconditioning animals to handling already occurs as part of routine husbandry procedures. For example, rodents are often handled on a daily or weekly basis in breeding and

research facilities; additional handling to precondition the animals to transportation handling is probably not necessary.

Many species of research animals are typically handled and then caged for transportation, and this practice can produce an additive stress effect of both the handling and the novel enclosure. Although an animal's stress response to human handling associated with transportation may not be completely ameliorated, the method of handling can reduce or exacerbate the stress response. Kannan and Mench (1996) demonstrated substantial physiological response differences between methods of handling of laying hens. Either birds were captured and then held and carried inverted (single carry or multi-bird carry) or single birds were captured and then held upright and carried gently. When compared with unhandled controls, both methods produced an alarm response. However, gentle upright handling yielded a lower response than inverted handling. Kannan and Mench (1996) also found that caging of the birds produced a powerful fear response. Capture, carrying, and caging were found to be less stressful to chickens when conducted under low light (Knowles and Broom, 1990).

The activity of horses during road transportation can contribute to the increased incidence of injury or stress. The physiological responses of horses to head restraint (cross-tied vs. loose) after 24 hr of transportation were measured by Stull and Rodiek (2000). Cross-tied horses had higher blood glucose and cortisol concentrations, neutrophil-to-lymphocyte ratios, and white blood cell counts than horses traveling loose in small compartments. The authors recommended that horses be allowed to travel loose during long periods of transportation.

MONITORING TRANSPORTATION

For facilities or people that transport large numbers of animals, the quality of transportation can be monitored by tracking mortality, morbidity, and injury during transportation and comparing these measures with published data. For instance, Malaga et al. (1991) reported that in-transit mortality for air transport of owl monkeys (*Aotus nancymai*) was 0.67% and total mortality at the end of a 30-day observation period was 2.44%. When mortality, morbidity, or injury exceed published norms (for instance, exceeding 2 standard deviations from the mean), action should be taken to adjust protocols or provide training. If small numbers of animals are occasionally transported, careful attention should be paid to ensuring that a reputable shipper is used and that the entire trip is adequately planned to transfer the animals smoothly from consignor to carrier, shipper, and consignee.

EMERGENCY PROCEDURES

Emergencies may occur during any phase of the shipping process. During the peritrip period, emergencies encountered have included extended delays before the start of long trips, exposure to extreme temperatures, animal escapes, and mechanical problems with transportation vehicles.

To ensure animal comfort and safety, all plans for animal shipments must include instructions for emergency responses in accordance with the mode of transportation used. For example, Appendix B of the International Air Transport Association Live Animals Regulations contains a section covering emergency responses. The section provides a summary of actions appropriate to emergency situations, including delays, container damage, escapes, illness, and segregation. It is important that when an emergency occurs, those directly involved with the transport of the animals (the shipper and the organization and individual(s) providing transport) need to be able to contact each other and the means of contact be established prior to transport. Planning must also include procedures to follow in the event of an emergency. Both a primary plan and a backup plan should be available for each phase of the trip. For example, if animals are to be transported by plane or truck and a mechanical problem causes a long delay, animal needs must be accommodated to avoid tragedy. Animals should not remain unprotected from extreme weather for more than a few minutes, and comfortable accommodations should be available.

In rare circumstances, a situation may arise in which it must be determined whether euthanasia of an animal is necessary. For example, an animal might become moribund during transportation, or might endanger the safety of the human handlers, as can happen if a horse becomes uncontrollable during a flight and kicks at the aircraft's doors. A part of the emergency procedure plan should document specifics that identify which persons are trained and qualified to make and carry out decisions (usually a veterinarian) and the methods and equipment to administer anesthesia or perform euthanasia safely in the transportation situation.

PERSONNEL TRAINING

Personnel who handle animals must be properly trained in routine and emergency procedures for the species they handle. Training should include procedures applicable to the mode of transportation and should cover at least

- Shipper and carrier responsibilities;
- Inspection of primary enclosures;

- Documentation;
- Acceptance, handling, and delivery;
- Loading and off-loading procedures and precautions;
- Operator and government regulations; and
- Emergency procedures.

Personnel must also be trained in species-specific husbandry and environmental requirements of animals. They can be deemed competent when they possess, as appropriate to the species and mode of transportation, the following:

- Ability to recognize when an animal becomes ill or unfit for transport;
 - Ability to recognize signs of stress and alleviate the cause, if possible;
 - Knowledge of how to contact and interact with local emergency personnel, including veterinarians who have skills in the treatment of injuries; and
 - Knowledge of the administration of veterinary drugs and methods of euthanasia.

Personnel must also be trained to recognize physiological signs that a problem is developing in a particular animal or group of animals. The signs may include

- Increased respiratory rate (in warm weather);
- Excessive sweating (in species that sweat during warm weather);
- Excessive shivering or huddling (in cool weather);
- Aggressive interactions and injuries associated with fighting;¹
- Excessive weight loss;¹ and
- Dehydration.¹

At least one person associated with each segment of the trip should be fully trained. Employers should provide training (initial and recurrent) for employees with respect to transportation of animals so that their employees will be able to ensure the safety of animals and of their own equipment and can explain to shippers the conditions under which animals are transported. Personnel that simply move containers into, out of, or between conveyances also must have at least minimal training to recognize potentially unsafe conditions (for the animal or the handler) and to know whom to contact in case of questions or problems.

¹Normative benchmarks should be known for the species in a given transportation protocol.

An American Veterinary Medical Association animal air-transportation study group evaluated, on a national basis, the adequacy of employee training as related to the protection of dogs and cats in air transportation (AVMA, 2002). The group found that although initial training was adequate for all of the airlines, continuing education and education of contractors were inconsistent. The group recommended the establishment of a formal training program that would incorporate:

- A time line for recurrent training;
- A consistent standard and frequency of training for ground handling staff, especially for outside contractors that are used more frequently by smaller airports; and
- A standard training program to minimize the amount of informal on-the-job training and thereby avoid omission of important considerations for safe animal care and transportation; this would also minimize delays in training during staff turnovers.

Biosecurity

Biosecurity, for the purposes of this report, is defined as the policies and measures taken to minimize the risk of introducing an infectious pathogen into the human, agricultural animal, and research animal populations. Animals have long been recognized as hosts of zoonoses (infectious diseases that can be transmitted to humans or other species of animals). Biosecurity should be a consideration when transporting research animals because of the close contact that can occur between research animals and human handlers or other transported animals. This creates the potential for unintentional or intentional transmission of a zoonosis into the human or agricultural animal populations. Many zoonoses, including potential (agro)bioterrorism agents, are difficult to detect in an infected animal because they cause asymptomatic disease in the host species. However, the effect of zoonoses can be significant in humans and agricultural animals, causing severe disability or death and negatively affecting the capacity of the agricultural sector.

Another biosecurity concern is transmission of an infectious pathogen to a research animal during transportation and introduction of the pathogen into the colony that receives the animal. Infectious pathogens can negatively affect the health of the research animal and colonies, confounding research utilizing the infected animals.

PROTECTING PUBLIC HEALTH AND AGRICULTURAL RESOURCES

Minimizing Risks Associated with Transporting Research Animals with Experimentally Introduced Zoonoses

Increased efforts to improve the biosecurity of human populations and the agricultural sector have resulted from passage of the USA Patriot Act (2001), the Bioterrorism Preparedness and Response Act (2002), and enforcement of three parts of the *Code of Federal Regulations* (42 CFR 73, 7 CFR 331, and 9 CFR 121). These regulations establish lists of agents and toxins that have been deemed threats to humans, animals, and plants (see Table 4-1). The regulations require research laboratories that possess any of the aforementioned agents to register its facility with the Centers for Disease Control and Prevention (CDC), designate a responsible official, perform background checks of persons who have access to the agents (conducted by the Department of Justice), and have a security plan for containment of the infectious agent. When infected animals must be transported, a plan for secure transportation must be in place. That plan would normally require:

- close communication between shipper and recipient;
- presence of responsible officials at the originating and receiving institutions;
- transfer of health records and assurances;
- identification of a carrier registered by the US Department of Agriculture;
- documentation of safety and security training of animal care personnel;
- notification of the appropriate institutional or CDC officials in case of emergency, loss, or theft;
- existence of emergency procedures (see Table 4-2); and
- good record maintenance.

Institutions are also required to have the appropriate level of laboratory biocontainment as outlined by CDC and the National Institutes of Health in the *Biosafety in Microbial and Biomedical Laboratories Manual* (BMBL). Although many shippers meet some of the requirements for laboratory biocontainment, not all meet all of the requirements, the result of which is a lack of uniformity in biosecurity during transportation. The characteristics of a good shipper are outlined in Table 4-3. Further, biocontainment requirements for transportation of infected animals (Appendix C, BMBL) are not as clearly defined as laboratory biocontainment

TABLE 4-1 Agents and Toxins That Require Registration of the Facility with CDC

Bacteria	Viruses (continued)
<i>Bacillus anthracis</i>	Cercopithecine herpesvirus 1 (Herpes B virus)
Botulinum neurotoxin producing species of <i>Clostridium</i>	Classical swine fever virus
<i>Brucella abortus</i> , <i>Brucella melitensis</i> , <i>Brucella suis</i>	Crimean-Congo haemorrhagic fever virus
<i>Burkholderia mallei</i> , <i>Burkholderia pseudomallei</i>	Eastern Equine encephalitis virus
<i>Cowdria ruminantium</i> (Heartwater)	Ebola virus
<i>Coxiella burnetii</i>	Foot-and-mouth disease virus
<i>Francisella tularensis</i>	Goat pox virus
<i>Mycoplasma capricolum</i> /M.F38/M. <i>mucoides capri</i> (contagious caprine pleuropneumonia), <i>Mycoplasma mycoides mycoides</i> (contagious bovine pleuropneumonia)	Hendra virus
<i>Rickettsia prowazekii</i> , <i>Rickettsia rickettsii</i>	Influenza virus (reconstructed replication competent forms of the 1918 pandemic influenza virus containing any portion of the coding regions of all eight gene segments)
<i>Yersinia pestis</i>	Japanese encephalitis virus
Fungi	Lassa virus
<i>Coccidioides immitis</i> , <i>Coccidioides posadasii</i>	Lumpy skin disease virus
Toxins	Malignant catarrhal fever virus (Alcelaphine herpesvirus type 1)
Abrin	Marburg virus
Botulinum neurotoxins	Menangle virus
<i>Clostridium perfringens</i> epsilon toxin	Monkeypox virus
Conotoxins	Newcastle disease virus (velogenic)
Diacetoxyscirpenol	Nipah virus
Ricin	Peste des petits ruminants virus
Saxitoxin	Rift Valley fever virus
Shigatoxin and Shiga-like ribosome inactivating proteins	Rinderpest virus
Staphylococcal enterotoxins	Sheep pox virus
Tetrodotoxin	South American haemorrhagic fever viruses (Junin, Machupo, Sabia, Flexal, Guanarito)
T-2 toxin	Swine vesicular disease virus
Prions	Tick-borne encephalitis (flavi) viruses (Central European tick-borne encephalitis, Far Eastern tick-borne encephalitis [Russian spring and summer encephalitis], Kyasanur Forest disease, Omsk hemorrhagic fever)
Bovine spongiform encephalopathy agent	Variola major virus (Smallpox virus)
Viruses	Variola minor virus (Alastrim)
African horse sickness virus	Venezuelan equine encephalitis virus
African swine fever virus	Vesicular stomatitis viruses (exotic)
Akabane virus	
Avian influenza virus (highly pathogenic)	
Bluetongue virus (exotic)	
Camel pox virus	

TABLE 4-2 Elements of an Emergency Plan

-
- Containers must be labeled clearly with the appropriate biohazard labels and identification of contents to inform transportation workers of package contents.
 - Package documentation must identify potential biohazards in the package and 24-hr contact information for a responsible official from the consignor institution.
 - The responsible official must be knowledgeable about the contents of the shipment and the potential hazards that it may pose and monitor the shipment in transit until receipt and confirmation by the consignee.
 - The responsible official will assist in the coordination of responses of emergency officials or transportation-worker safety officers in case of accident or theft.
-

TABLE 4-3 Characteristics of a Good Shipper^a

A good shipper of research animals:

- finalizes the method of shipment, route, and special care required for a shipment before accepting it;
 - obtains all required federal, state, and regulatory-agency permits and documents;
 - ensures that the carrier is US Department of Agriculture (USDA)-certified for live-animal shipments or meets International Air Transport Association (IATA) and Department of Transportation (DOT) regulations for dangerous goods;
 - provides the carrier with information about the shipment, including type of animal (scientific and common name), sex, physical conditions, number of animals per container, medication given, and whether the shipment is a dangerous good;
 - provides containers that adhere to current IATA regulations for live animals or dangerous goods;
 - provides suitable bedding and food for the animals;
 - obtains all documentation and correct information required by IATA, by national and carrier regulations, and for the shipper's certification;
 - provides special feeding and watering instructions on the outside of each container and provides the carrier with a respective copy;
 - develops an emergency plan and a 24-hr contact number for a responsible official to coordinate responses; and
 - maintains records for each shipment—including such information as the species and number of animals, dates of shipment and receipt, carrier, and name and address of consignee—for at least 3 years.
-

^aAdapted from Section 1.2 of the Live Animals Regulations (IATA, 2005).

requirements. However, Appendix C of the BMBL, as well as Chapter 2 of this report, contains comprehensive information on which agencies must be contacted when transporting animals with human and agricultural animal zoonoses and can provide assistance in development of appropriate biocontainment plans for transportation.

Minimizing Risks Associated with Transporting Research Animals with Unknown Zoonoses

Though research animals may be experimentally infected with agents, many zoonoses of concern are endemic in research animals or may be naturally acquired. A list of zoonotic diseases communicable from research animals to humans is presented in Table 4-4. Because many of those zoonoses cause asymptomatic disease in animals, it may not be apparent that the animal is infected. The challenge is to identify the zoonoses that animals potentially harbor, consider the likelihood that the animal is a carrier and potential for exposure, evaluate the likely severity of an adverse event, and take steps to mitigate the risk.

TABLE 4-4 Examples of Zoonotic Diseases Transmissible from Research Animals to Humans

Disease	Potential Animal Vectors	Potential Route of Transmission
<i>Bacterial</i>		
Anthrax	Contaminated herbivores	Cutaneous, inhalation
Brucellosis	Cattle, goats, swine, dogs	Inhalation, ingestion, direct contact
Leptospirosis	Cattle, dogs, horses, swine, rodents, reptiles, amphibians	Inhalation of contaminated fluids, direct contact
Salmonellosis	Birds, swine, reptiles, turtles, tortoises	Direct contact, fecal-oral
Tuberculosis	Domestic and wild animals	Droplets
Q fever	Cattle, sheep, goats	Inhalation, direct contact with infected animals, their birth products, or infected materials such as bedding
<i>Viral</i>		
Influenza	Birds, horses, swine	Aerosol, physical contact
Hantaviruses	Rodents	Aerosol, direct contact with mucous membranes, animal bites
Ebola	Unknown	Direct contact with infected materials, possibly droplets
Monkey B virus	Old World monkeys	Animal bites, direct contact with mucus membranes
Monkeypox	Ground squirrels, gambian rats	Droplets
Rabies	Dogs, cats, wild carnivores, bats, foxes, raccoons	Animal bite, possibly airborne
<i>Fungal</i>		
Ringworm	Bovine, birds	Direct contact

There are reports of zoonotic disease transmission from pet hamsters, rabbits, and rodents to humans (CDC, 2001, 2005a, 2005b). Since companion animals are often transported in unfiltered containers and are transported along with research animals (particularly during air transport), the potential for cross contamination during transport must also be considered.

In general, contact between animals and people during transportation should be restricted to prevent exposure to or transfer of zoonoses. When possible, human contact should be limited to trained animal handlers who are knowledgeable of good sanitation practices, biosafety and biocontainment, and precautions for protection against zoonoses.

Special Considerations When Transporting Nonhuman Primates

The transportation of nonhuman primates requires special consideration because the risk of zoonotic disease transmission is greater with nonhuman primates than any other species of research animal due to the close phylogenetic relationship between humans and nonhuman primates (NRC, 2003b). Macaques imported for research have been implicated in the transmission of B virus and Ebola virus to laboratory workers, both potentially fatal diseases in humans (Cohen *et al.*, 2002; Palmer, 1987). B virus (also known as *Herpesvirus simiae*) is of particular concern as it is endemic in some populations of macaques and infected animals are generally asymptomatic. B virus and Ebola virus can be transmitted through aerosols, animal bites, scratches, contact with body fluids or tissue material, or equipment that has been contaminated with body fluids (NRC, 2003b).

Due to the risks associated with zoonotic diseases transmitted from nonhuman primates, a common standard for personal protective equipment (PPE) has been established for workers who come into contact with nonhuman primates or equipment that has been exposed to nonhuman primates (NRC, 2003b). This standard recommends that dedicated clothing, gloves, and masks be utilized when in contact with nonhuman primates and that eye and face protection be mandatory for individuals who come into contact with macaques. Eye and face protection are also highly recommended for individuals who come into contact with other Old World monkeys.

Ensuring public safety and maintaining public confidence in the shipping process should be concerns of both regulatory agencies and carriers. Public confidence is difficult to maintain when airline passengers observe transportation workers wearing PPE boarding their plane. However, the development and use of overshippers (a closed, environmentally controlled container into which a standard primary enclosure would be

loaded in order to prevent a zoonotic exposure) would mitigate the need for some types of PPE (please refer to Chapter 5 for further discussion).

Special Considerations When Transporting Specimens and Tissues

Diagnostic specimens and tissues that are used for research are usually isolated from animals that are suspected of having an infectious disease or that are from an endemic area, and they should be treated as potentially infectious and hazardous materials. Those materials should be handled according to guidelines in BMBL (5th edition, Section VI and Appendix C). All diagnostic and tissue samples should be packaged according to IATA regulations as dangerous goods (Chapter 2). Depending on the suspect sample, it may be necessary for only persons who have IATA training to handle the sample. Access to the packages should be limited. If the sample potentially contains a select agent, both the shipper and the recipient must have all pertinent clearances and permits required by CDC and USDA and must have notified all appropriate agencies.

PROTECTING THE BIOLOGICAL INTEGRITY OF RESEARCH ANIMALS AND COLONIES

In recent years, greater attention has been paid to maintaining the microbial status of research animals and animal colonies. Scientists have rapidly expanded the use of immunocompromised rodents, such as nude mice and transgenic animals with immune deficits. Preventing exposure to infectious agents is necessary to maintain the health of these animals. In addition, scientists have discovered infectious agents, such as mouse and rat parvoviruses and *Helicobacter* species, that cause subclinical infections but can significantly alter research results (Jacoby and Lindsey, 1998). A list of viral, bacterial, and parasitic organisms found in commonly used species of research animals is presented in Table 4-5. Many of these organisms can infect multiple species, increasing the potential for intra-species and interspecies disease transfer.

The most common routes of disease transmission between animals are infectious aerosols, close contact, and fomite (an inanimate object, such as clothing, capable of transmitting infectious organisms) transmission. Each of these routes poses a risk during shipping; however, there are methods to prevent the transmission of diseases among research animals, including barrier containment, specific-pathogen diagnosis, disinfection of vehicles and shipping containers, use of personal protective equipment (PPE), and segregation of animals. Though research animal vendors generally have well-established procedures to minimize biosecurity concerns, the typical researcher may need guidance in addressing biosecurity concerns

TABLE 4-5 Infectious Agents and the Susceptible Species of Research Animals

	Mice	Rats
Viruses		
Sendai	X	X
PVM (Pneumonia virus of mice)	X	X
MHV (Mouse hepatitis virus)	X	
MVM (Minute virus of mice)	X	
GD-VII (Theiler's murine encephalomyelitis virus strain)	X	
REO 3 (Reovirus type 3)	X	X
EDIM (Group A rotavirus)	X	
Lymphocytic choriomeningitis vVirus	X	X
Polyoma	X	
MCMV (Murine cytomegalovirus)	X	
Ectromelia	X	
MPV (OPV) (Mouse parvovirus – Orphan parvovirus)	X	
MAD (Mouse adenovirus)	X	X
K virus	X	
MTLV (Mouse thymic virus)	X	
Hantavirus		
Adenovirus		
Parainfluenza		
Rotavirus		
PI-1 (Parainfluenza-1)		
PI-2 (Parainfluenza-2)		
RHD (Rabbit haemorrhagic disease)		
LCM (Lymphocytic choriomeningitis)		
H1 (Toolan's H1 virus)		X
KRV (Kilham Rat virus)		X
SDA/RCV (Sialodacryoadentitis virus/Rat corona virus)		X
HANT (Hantaan)		X
RRV (Ross River virus)		X
Foamy virus		
Dengue		
Yellow fever		
Pox viruses		
Ebola		
SIV (Simian immunodeficiency virus)		
B virus		
Rat picornavirus		X
RPV (OPV) (Rat parvo virus – Orphan parvo virus)		X

Guinea Pigs	Gerbils	Rabbits	Hamsters	Nonhuman Primates
X			X	
X			X	
X		X	X	
X	X			
X				
X				
		X		
		X		
		X		
		X		
			X	
				X
				X
				X
				X
				X
				X
				X
				X

continued

TABLE 4-5 Continued

	Mice	Rats
Bacteria		
<i>Streptococcus zooepidemicus</i>		
<i>Pasteurella multocida</i>		
<i>Pasteurella</i> spp.		
<i>Treponema cuniculi</i>		
<i>Bordetella bronchiseptica</i>	X	X
<i>Citrobacter rodentium</i>	X	
<i>Corynebacterium kutscheri</i>	X	X
<i>Clostridium piliforme</i>	X	X
<i>Salmonella</i> spp.	X	X
<i>Mycoplasma pulmonis</i>	X	X
<i>Streptobacillus moniliformis</i>	X	X
<i>Helicobacter hepaticus</i>	X	X
<i>Campylobacter</i> spp.		
<i>Yersinia</i> spp.		
<i>Mycobacterium</i> spp.		
<i>Burkholdria</i>		
CAR bacillus	X	X
Parasites		
Ectoparasites	X	X
Gastrointestinal helminths	X	X
Gastrointestinal protozoa and sporozoans	X	X
<i>Encephalitozoon cuniculi</i>	X	X
Hepatic coccidia		
Metazoa		
Intestinal coccidia		
Other protozoa		
Demodex		

Guinea Pigs	Gerbils	Rabbits	Hamsters	Nonhuman Primates
X				
		X		
		X		
		X		
X	X	X	X	
	X	X		
X	X	X	X	X
X	X		X	
X				
				X
				X
				X
		X		
		X		
X	X	X	X	X
X	X		X	X
X	X		X	X
X		X	X	
		X		
		X		X
		X		X
		X		X
			X	

TABLE 4-6 Recommendations for Shipment of Research Animals Between Institutions

-
- Shipments of research animals between institutions should be coordinated between responsible persons at the sending and receiving institutions. They should ensure that all documentation is in order, including federal (CDC, USDA, and DOT), state, and local permits.
 - Shipments should be prepared by persons who have documented training in animal handling procedures and the proper use of PPE.
 - Animals should be placed in shipping containers that will provide protection to them and the receiving colony from microbial contamination.
 - Animals should be packaged according to regulatory-agency (IATA, USDA, and DOT) standards.
 - Shippers should provide documentation and assurances to recipients that the animals are healthy. If the recipient requires a more extensive health report, including testing for specific pathogens, the costs of the diagnostic tests should be covered by the consignee.
 - Only USDA-certified carriers should be used for transportation of research animals.
 - On receipt, the consignee should place the research animals at the appropriate level of biocontainment and quarantine before introduction into the laboratory colony.
-

when shipping animals to a colleague. Recommendations for shipment of animals between research institutions can be found in Table 4-6.

Barrier Containment

Most small-animal vendors have designed shipping containers that incorporate spun polypropylene filters to provide a physical barrier to the transfer of microbial contaminants into or out of each container, thus protecting research animals, colonies, and animal handlers from pathogen exposure during transportation. For gnotobiotic animals (animals whose microfauna and microflora are known in their entirety) and immunocompromised animals, microisolation shipping containers are also available. Although the sturdy construction of the vendor containers may tempt researchers to reuse them to transfer research animals to other researchers, this practice is not recommended. Most vendors sterilize or disinfect the animal containers, food, and water before loading animals. Once a container has been opened at the recipient's facility, its sterility has been compromised. Some facilities autoclave shipping containers for reuse; however, this may increase the air resistance of the polypropylene filters, restricting air flow (White, 2004). Until it can be established that autoclaving does not restrict air flow below acceptable levels, the committee suggests that the prudent course of action is to avoid autoclaving shipping containers for reuse. To ensure the biosafety of their animals,

individual researchers who wish to transfer animals to other researchers should purchase presterilized shipping containers from animal vendors.

Specific Pathogen Diagnosis

Normally, when major vendors communicate with clients, they identify the pathogens for which their animals have been tested to assure recipients that the animals sent to them are free of pathogens that could disrupt their colonies or experiments (for example, introduction of a respiratory infection would disrupt and invalidate an experiment on respiration). People shipping a few animals to colleagues do not always know the pathogen status of their animals and therefore cannot assure colony supervisors that the imported animals can be introduced safely into their colonies. To avoid inadvertent introductions of diseases, some colony supervisors quarantine incoming animals until they can gather the data required to ensure the health and safety of incoming animals (Otto and Tolwani, 2002). If facilities do not have the space and testing facilities required for that precaution, then it may be necessary for testing to be conducted at the institution of origin. Unfortunately, testing in such instances does not monitor for disease transmission during transport. Arrangements for testing and provision of assurances before introduction of shipped animals into a new colony require communication between responsible and knowledgeable officials of the institutions involved. Such officials can help to arrange safe and secure shipment of animals and arrange for the most efficient assurance of colony security.

Disinfection

Transportation protocols should have standardized procedures for disinfection of animal cages, transportation vehicles, and holding areas that conform to IATA, USDA, DOT, and CDC standards. Disinfection prevents transmission of pathogens from one shipment of animals to the next shipment transported in the same vehicle. Bedding, food, and water may be sterilized by autoclaving or gamma irradiation before and after shipment to prevent contamination of research animals and the receiving colony. Disinfection by sterilization or irradiation is not feasible for such items as transfer-vehicle cargo holds, large cages, and transportation-company holding areas. In these cases, however, chemical disinfection should be conducted after each transfer event. To ensure maximal efficiency of the disinfection process, disinfection should be applied using concentrations of chemical disinfectants and application times should be optimized according to manufacturers' instructions.

Disinfection of the outside of the shipping containers should also be considered. As discussed above, companion animals are often transported in unfiltered containers and may be transported along with research animals. Therefore, the potential for cross contamination of shipping containers is present and must be considered. Another situation that may result in cross contamination is the entry of wild mice infected with lymphocytic choriomeningitis virus or mouse hepatitis virus into an animal holding area along the transportation route, such as an airport or cargo transfer station. The infected wild mouse can shed virus, thus contaminating the outside of the shipping container. Transmission of pathogens to research animals or colonies can then occur if the container is brought into a facility or animals are removed from the container without disinfection of the outside surfaces. Since such infections occur during transportation, diagnostic testing by the source provider does not ensure the biosecurity of either the animals or the receiving colony.

Personal Protective Equipment

The appropriate use of PPE can also protect research animals from human pathogens and cross contamination from other animals. For example, macaques are susceptible to human infections such as measles and tuberculosis. The use of PPE will not only prevent the transmission of B virus from a macaque to a human, but also can prevent the inadvertent transmission of measles or tuberculosis to the macaque. People who handle animals should cover their street clothing and exposed body surfaces with PPE to reduce the risk of pathogen introduction through direct contact or aerosol. In some instances, it may be appropriate to provide handlers with a shower-based entry system. The appropriate disposal of PPE is also necessary so that the PPE does not act as a fomite for transmitting pathogens. For example, if PPE is worn while disinfecting incoming shipping containers, the PPE should be disposed of before moving on to other tasks.

Segregation of Animals

Separation of different shipments of animals is also a method for preventing intra- and interspecies transmission of pathogens presented in Table 4-4. The committee suggests close adherence to the recommendations in the LARs regarding segregation of species and separation of animals of the same species of different origins (LARs Sections 5.3 and 10.3.2). Briefly, these regulations state that:

1. animals in quarantine must be segregated from those which are not;
2. animals known to be for laboratory use must not be stored adjacent to other animals in order to reduce any risk of cross-infection or contamination;
3. nonhuman primates from different continents must be isolated from each other in aircraft holds, airport cargo warehouses, animal holding facilities, and during all phases of ground transportation; and
4. animals that are natural enemies, e.g., cats and dogs, may be loaded in the same hold provided they are not in sight of one another.

Situations in which some aspects of these recommendations are not feasible may arise. For example, an airport may not have containment facilities to separate nonhuman primate species. In these cases, other measures must be employed to prevent disease transmission. An effective means of overcoming this problem would be the development of self-contained overshippers, as recommended in Chapter 5.

Recommendations

Preceding chapters have provided background regarding the laws, regulations, and regulatory agencies responsible for animal transportation (Chapter 2); principles that underlie good practices of animal handling, management, and care essential to maintaining animal comfort, health, and well-being during and immediately after transportation (Chapter 3); and concepts and principles related to preventing exposure of the general public, transported animals, animal handlers, and animal colonies to infectious organisms (Chapter 4). These factors provided the foundation for the following four recommendations.

Recommendation 1: The National Institutes of Health (NIH), through the National Center for Research Resources, should update and reimplement the National Primate Plan to ensure a stable means for transporting nonhuman primates into and within the United States. In addition, research institutions that use nonhuman primates should encourage the development of reliable ground transport for nonhuman primates to protect against the possibility that domestic transportation of nonhuman primates on commercial airlines may one day become unavailable.

For US research institutions, there are several sources for obtaining nonhuman primates for use in biomedical research. Sources include overseas breeding colonies, National Primate Research Center (NPRC) breeding colonies, breeding colonies at academic institutions, and US commercial breeding colonies, with overseas breeding colonies supplying the majority

of nonhuman primates (Robinson and Beattie, 2003). Therefore, the majority of nonhuman primate resources in the United States must be transported by air, both into the United States and generally to their final destination (see Figure 5-1 for the locations of research facilities, importation sites, and vendors of nonhuman primates). Currently, few foreign airlines will consistently transport nonhuman primates into the United States, and no US domestic airlines will transport nonhuman primates into the United States. For the purpose of transporting nonhuman primates once they are already in the United States, research institutions may use the one US domestic airline that currently transports nonhuman primates nationally (Kemnitz, J. personal communication, August 4, 2005).

The paucity of carriers that transport live animals is due, in part, to the often unprofitable nature of live-animal transportation in the air-transportation industry. For typical passenger/cargo-configured network carriers, such as the major airlines present in the US market, cargo contributes only 2 to 5% of total operating revenues. The contribution of live-animal transportation is even less significant and involves costs for personnel training and environmental controls to comply with federal regulation. In addition to the economic disincentives discussed above, several other reasons have been identified for the declining number of commercial airlines that will transport nonhuman primates, including the following:

- concern regarding zoonoses between nonhuman primates and humans;
- the high cost of training personnel and acquiring protective equipment;
- potentially higher insurance rates due to liability issues;
- required disinfection of cargo areas after holding nonhuman primates;
- pressure from animal rights activists; and
- the potential danger associated with escapes (DePoyster, 2003).

There is no clear solution to the foreign commercial airline situation. Some organizations have started chartering private jets to import shipment of nonhuman primates to the United States (DeMarcus, 2003). This option increases the cost of transportation (DeMarcus, 2003), and private chartering companies are not immune to the pressures that led the commercial airlines to refuse to transport nonhuman primates. Though corporate research institutions have some ability to cover the cost of private charters, academic researchers utilizing nonhuman primates have no mechanism for absorbing the increased cost of transportation by private charters. In the committee's judgment, the practical approach is to pre-



FIGURE 5-1 Locations of research facilities using nonhuman primates, major importation sites, and vendors of nonhuman primates in the United States.

pare for the eventuality that all nonhuman primates may have to be transported into the US by private charter. To prevent the financial impact of private charters from restricting research involving nonhuman primates, it is imperative for agencies that fund research using nonhuman primates to ensure that allowances are made for the increased costs associated with private chartering.

In some ways, the domestic air transportation system is more likely to reach a crisis point because currently only one domestic airline will transport nonhuman primates within the United States. A similar situation arose several years ago, when US airlines refused to carry day-old chicks and adult avian species via airmail transport. As a result, Congress enacted Public Law 107-67, the Treasury and General Government Appropriations Act of 2002. Section 651 of this law amended Section 5402(c) of Title 39 so that "the Postal Service may require any air carrier to accept as mail shipments of day-old poultry and such other live animals as postal regulations allow to be transmitted as mail matter." While legislative action by Congress would virtually eliminate the domestic air transportation problem, that solution is perhaps the least likely to occur, and the committee felt it imperative to provide other pathways to improve the transportation of nonhuman primates.

Some individuals in the academic research community believe that the NPRC breeding colonies will eventually be able to meet the domestic need for nonhuman primates, eliminating the need to transport nonhuman primates into the United States in large numbers. The source of this assumption is the National Primate Plan (DHEW, 1978). The National Primate Plan was published in 1978 by the Interagency Primate Steering Committee, which included representatives from the National Science Foundation, Department of Defense, Environmental Protection Agency, Veterans Administration, and Department of Health, Education, and Welfare. The National Primate Plan was developed to ensure adequate supplies of primates to meet research needs by coordinating the various federal program activities. One of the recommendations of the National Primate Plan was that domestic primate production be sufficiently expanded to ensure a continuous, stable, and long-term supply of primates and that domestic production must provide for all of the nation's need for commonly used species, specifically rhesus and cynomolgus macaques. At the time, the Plan's authors estimated that domestic production was fulfilling about 50% of the domestic need. Since 1980, imported nonhuman primates have comprised, on average, 26% of the nonhuman primates used in research. However, the percentage of nonhuman primates used in research that are imported has steadily increased over the last seven years and, in 2004, imported nonhuman primates comprised 35% of nonhuman primates used in

research.¹ This is a cause for concern, as the NPRC's facilities are limited and being utilized to the maximum (Robinson and Beattie, 2003).

Considering that reliance on imported nonhuman primates is increasing and that the NPRCs are functioning at maximum capacity, it is unlikely that the US research community's demand for imported nonhuman primates will decrease. Moreover, the mission of the NPRCs is to provide researchers with access to nonhuman primates, not to supply nonhuman primates directly to researchers. Any research using NPRC animals must be conducted at an NPRC, and NIH-funded research projects take precedence over activities funded by other resources (NIH, 2002). The NPRC breeding colonies are not intended to supply nonhuman primates to other academic or corporate research institutions and there is no indication that their mission will be revised. In the committee's judgment, the most promising solution for permanently addressing the declining availability of transport services is to update and reimplement the National Primate Plan. *An updated National Primate Plan could ensure allowances for increased costs associated with domestic private chartering where necessary and relaunch the nation's efforts to develop domestic production of the most commonly used nonhuman primates to meet national needs.*

Recommendation 2: NPRCs and research institutions utilizing non-human primates should work jointly, perhaps through professional societies, to encourage the development of reliable ground transportation for nonhuman primates.

Most producers of small research animals have reacted to the declining availability of economical domestic airline services by developing systems for ground transport of their research animals either by utilizing their own personnel and vehicles or by contracting with independent ground transport companies. Ground transport of nonhuman primates does seem to occur, but is not a widespread occurrence, possibly due to economic constraints and because most ground transport companies are geared toward the transportation of rodents and other small mammals. It would be prudent for the NPRCs and individual research institutions to work jointly, perhaps through professional societies, to encourage the development of reliable ground transport for nonhuman primates to protect against the possibility that domestic transportation of nonhuman primates on commercial airlines may one day become unavailable.

¹Estimates of the percentage of imported nonhuman primates used in biomedical research in the United States were generated from US Department of Agriculture (USDA) data on the number of animals used in research (USDA, 2004) and data from the US Fish and Wildlife Service and Centers for Disease Control and Prevention (CDC) on the number of nonhuman primates imported into the United States for research purposes (DeMarcus, 2003).

Recommendation 3: Federal agencies that fund nonhuman-primate research and the commercial shipping community should coordinate an initiative to develop a self-contained overshipper to ship non-human primates and other animals that pose a significant risk of zoonotic exposure.

Recommendation 3 is directed to the possibility of cost-effectively shipping nonhuman primates and other infectious animals in standard primary enclosures that would be loaded into closed devices (overshippers). Overshippers could be built in such a fashion as to incorporate HEPA filters, temperature and humidity controls, viewing windows, and outside access. An overshipper with these characteristics would minimize or eliminate exposure to or contact between humans and animals, as well as between animals in the same conveyance, while providing an escape-proof and safe environment. Government agencies, shippers, inspectors, handlers, and carriers alike might prefer the limited human intervention, reduced zoonotic exposure, and convenience this method would provide. Because it is unlikely that the airline industry itself would be in a position to pay for the development of such units, the cost must be borne by the shipping and animal research communities at large. The resulting improvements in terms of safety, security, and convenience may encourage more airlines to transport nonhuman primates and other species that pose similar problems.

Recommendation 4: An interagency working group should be established to coordinate all federal inspection and permitting activities related to transportation of animals and animal products under one entity. In addition, the individual agencies should move to clarify the language of federal regulations or issue guidance documents to increase the readability and understanding of those regulations.

As discussed in Chapter 2 (and diagrammed in Table 2-1), five different federal agencies oversee various aspects of the transportation of research animals and their products. The USDA provides inspection oversight to ensure the welfare of animals during transportation, the Fish and Wildlife Service issues permits and inspects shipments to control the movement of wildlife and conserved species, the CDC registers and inspects shipments to control the introduction of zoonotic disease through imports, the Food and Drug Administration (FDA) performs similar functions to control the intrastate and interstate spread of zoonotic diseases, and the Department of Transportation issues permits to control hazards presented by the transportation of materials such as infectious live animals or radioactive animal products. Due to overlapping regulatory authority, it is possible for one shipment of animals to be inspected by two different

agencies and be required to register or obtain permits from three or more agencies. This overlap presents a significant regulatory burden to individual investigators trying to navigate this complex network of regulatory authority. Establishing one federal entity that can be consulted would greatly reduce regulatory burden for shippers, whether they are individual investigators or large commercial operations. Such a working group also has the potential to centralize and reduce the number of redundant inspections and permits that must be issued for each shipment.

There is also substantial room for improving the comprehensibility of the various federal regulations. The committee was particularly concerned with areas of inconsistency and obscurity in the Animal Welfare Act (AWA) regulations, which are the federal regulations that establish standards for animal welfare that apply to most species of research animals during transportation. The AWA regulations are organized by groups of common species: 9 CFR 3 subpart A details standards for the transportation of dogs and cats, subpart B covers guinea pigs and hamsters, subpart C covers rabbits, subpart D covers nonhuman primates, etc. In this way, species with similar physiology and behavioral characteristics are regulated by the same standards. While differences between species account for the majority of differences in standards between species groupings, there are several cases of inconsistencies that have no apparent underlying scientific reason. For example, the regulations pertaining to dogs and cats state that the cargo area must be pressurized when flying above 8,000 ft (9 CFR 3.15(d)). There is no similar requirement for nonhuman primates, rabbits, guinea pigs, or hamsters. Another example involves acclimation of guinea pigs. For all species besides guinea pigs, there is a provision in the AWA regulations for transporting animals in temperature conditions that fall below the minimum allowed temperature if the animal has been acclimated to lower temperatures as certified by a USDA-accredited veterinarian (e.g., 9 CFR 3.13(e)). However, no such provision exists for guinea pigs, though there is literature suggesting that guinea pigs can be successfully cold-acclimated (Sobel *et al.*, 1960, 1965; Vapaatalo *et al.*, 1984).

There are also a number of inconsistencies in standards that apply to the same animal. For example, for nonhuman primates, the temperature in a terminal facility cannot fall below 45°F for more than *four consecutive hours* (9 CFR 3.91 (d)), but the temperature during handling (movement into or out of a terminal facility or conveyance) cannot fall below 45°F for more than *45 minutes* (9 CFR 3.92(a)(3)). During air transport, the ambient temperature inside the plane must be maintained at a level that ensures the welfare of the nonhuman primate (9 CFR 3.88(d)); however, during surface transportation, the temperature must be maintained between 45°F and 85°F (9 CFR 3.88(e)).

There are also instances in the Animal Welfare Regulations where there are inconsistencies in reporting and documentation. For example, the certification that must accompany dogs, cats, and nonhuman primates must include the name, address, and telephone number of the consignee (9 CFR 3.13(b) and 9 CFR 3.86(b)); however, this information does not have to be included in the certification that must accompany rabbits, guinea pigs, or hamsters. While inconsistencies in reporting and documentation are unlikely to affect the welfare of the animal directly, they make compliance with AWA regulations onerous, thus diminishing their effectiveness.

Though the committee had greater concerns about the AWA regulations because they directly impact the welfare of the animals, the committee found that all of the federal regulatory language lacked clarity. This makes it difficult to determine which federal agencies would oversee a particular transportation situation. In particular, it is exceedingly difficult to determine which federal regulations apply to the transportation of animals or animal products that contain a known or suspected zoonotic/infectious agent, whether naturally occurring or experimentally induced (see Table 2-1). Federal agencies should work to clarify these issues, either through the development of an interagency working group or the issuance of guidance documents by the individual agencies.

Recommendation 5: Shipments of research animals between institutions should be coordinated between responsible individuals at both the sending and receiving institutions.

In the committee's judgment, instances of mortality, morbidity, and adverse effects on animal health occur most frequently when individual investigators unfamiliar with the vagaries of the research animal transportation system ship animals to investigators at other institutions. Such problems can be avoided if each institution designates a single individual responsible for ensuring the safe shipment and receipt of research animals. This designated individual would ensure that communication between institutions is effective at all stages of the transport process; that all documentation including health certifications required by the receiving institution and all federal, state, and local permits are in order; and that animals are placed in shipping containers according to the appropriate standard. This designated individual would further ensure that USDA-certified carriers are utilized and that, upon receipt, the consignee is notified and places the animals in the appropriate level of biocontainment/quarantine before introduction into the laboratory colony. Though institutional animal care and use committees are not required to directly review each instance of animal transport, transport activities should be reviewed during the semiannual program evaluation to ensure that appropriate procedures are being followed (Silverman *et al.*, 2000).

References

- AAALAC International. 2003. Overcoming the challenges of animal transport. *AAALAC International Connection*.
- AATA. 2000. *AATA Manual for the Transportation of Live Animals*, 2 ed. Surrey, England: Harris Associates Limited.
- Abbott, T.A., H.J. Guise, E.J. Hunter, R.H.C. Penny, P.J. Baynes, and C. Easby. 1995. Factors influencing pig deaths during transit: an analysis of drivers' reports. *Animal Welfare* 4:29-40.
- Adams, T. 1963. Mechanisms of cold acclimatization in the cat. *Journal of Applied Physiology* 18:778-80.
- Adams, T., M.L. Morgan, W.S. Hunter, and K.R. Holmes. 1970. Temperature regulation of the unanesthetized cat during mild cold and severe heat stress. *Journal of Applied Physiology* 29(6):852-8.
- Aguila, H.N., S.P. Pakes, W.C. Lai, and Y.S. Lu. 1988. The effect of transportation stress on splenic natural killer cell activity in C57bl/6J mice. *Laboratory Animal Science* 38(2):148-51.
- American Veterinary Medical Association (AVMA). 2002. *A Report from the American Veterinary Medical Association Animal Air Transportation Study Group*.
- Baldock, N.M., and R.M. Sibly. 1990. Effects of handling and transportation on the heart rate and behaviour of sheep. *Applied Animal Behaviour Science* 28:15-39.
- Barlow, S.M., P.R. McElhatton, and F.M. Sullivan. 1975. The relation between maternal restraint and food deprivation, plasma corticosterone, and induction of cleft palate in the offspring of mice. *Teratology* 12(2):97-103.
- Baxter, S. 1984. *Intensive Pig Production*. London: Granada Press.
- Bayliss, P.A., and M.H. Hinton. 1990. Transportation of broilers with special reference to mortality rates. *Applied Animal Behaviour Science* 28:93-118.
- Besch, E.L., and R.L. Brigmon. 1991. Adrenal and body temperature changes in rabbits exposed to varying effective temperatures. *Laboratory Animal Science* 41(1):31-4.
- Besch, E.L., H. Kadono, and R.L. Brigmon. 1984. Body temperature changes in dogs exposed to varying effective temperatures. *Laboratory Animal Science* 34(2):177-80.

- Bligh, J., and K.G. Johnson. 1973. Glossary of terms for thermal physiology. *Journal of Applied Physiology* 35(6):941-61.
- Brody. 1945. *Bioenergetics and Growth*. New York: Reinhold Publishing Corporation.
- Brown, K.S., M.C. Johnston, and J.D. Niswander. 1972. Isolated cleft palate in mice after transportation during gestation. *Teratology* 5:119-24.
- Centers for Disease Control and Prevention (CDC). 2001. Tularemia—Oklahoma, 2000. *MMWR* 50(33):704-6.
- Centers for Disease Control and Prevention (CDC). 2004. Order of the Centers for Disease Control and Prevention, Department of Health and Human Services. [Online.] Available: http://www.cdc.gov/ncidod/sars/pdf/order_civet_ban_011304.pdf. [accessed September 2005].
- Centers for Disease Control and Prevention (CDC). 2005a. Outbreak of multidrug-resistant *salmonella typhimurium* associated with rodents purchased at retail pet stores—United States, December 2003-October 2004. *MMWR* 54(17):429-33.
- Centers for Disease Control and Prevention (CDC). 2005b. Interim guidance for minimizing risk for human lymphocytic choriomeningitis virus infection associated with rodents. *MMWR* 54(30):747-9.
- Centers for Disease Control and Prevention (CDC). 2005c. Nonhuman Primate Imports FY 2001-2004. Unpublished raw data.
- Cohen, J.I., D.S. Davenport, J.A. Stewart, S. Deitchman, J.K. Hilliard, L.E. Chapman, and B Virus Working Group. 2002. Recommendations for prevention of and therapy for exposure to B virus (Cercopithecine Herpesvirus 1). *Clinical Infectious Diseases* 35(10):1191-203.
- Cole, N., T.H. Camp, L.D. Rowe, D.G. Stevens, and D.P. Hutcheson. 1988. Effect of transport on feeder calves. *American Journal of Veterinary Research* 49:178-83.
- Collins, M.N., T.H. Friend, F.D. Jousan, and S.C. Chen. 2000. Effects of density on displacement, falls, injuries and orientation during horse transportation. *Applied Animal Behaviour Science* 67:169-79.
- Cooper, K.E., A.V. Ferguson, and W.L. Veale. 1980. Modification of thermoregulatory responses in rabbits reared at elevated environmental temperatures. *Journal of Physiology* 303:165-72.
- Crookshank, H.R., M.H. Elissalde, R.G. White, D.C. Clanton, and H.E. Smalley. 1979. Effect of transportation and handling of calves upon blood serum composition. *Journal of Animal Science* 48:430-5.
- Curtis, S.E. 1983. *Environmental Management in Animal Agriculture*. Ames, Iowa: Iowa State University Press.
- del Barrio, A.S., J.W. Schrama, W. van der Hel, H.M. Beltman, and M.W.A. Verstegen. 1993. Energy metabolism of growing pigs after transportation, regrouping, and exposure to new housing conditions as affected by feeding level. *Journal of Animal Science* 71:1754-60.
- DeMarcus, T. 2003. Nonhuman primate importation and quarantine: United States, 1981-2001. In: *International Perspectives: The future of nonhuman primate resources*. NRC. Washington, DC: The National Academies Press.
- Department of Health, Education, and Welfare (DHEW). 1978. *National Primate Plan*. Washington, DC: DHEW Publication No. (NIH) 80-1520.
- Depocas, F., J.S. Hart, and O. Heroux. 1957. Energy metabolism of the white rat after acclimation to warm and cold environments. *Journal of Applied Physiology* 10(3):393-7.
- DePoyster, J. 2003. Transportation of primates and the animal welfare act. Pp. 183-6. In: *International Perspectives: The future of nonhuman primate resources*. NRC. Washington, DC: The National Academies Press.
- Dhabhar, F.S., and B.S. McEwen. 1996. Stress-induced enhancement of antigen-specific cell-mediated immunity. *Journal of Immunology* 156(7):2608-15.

- Dhabhar, F.S., and B.S. McEwen. 1997. Acute stress enhances while chronic stress suppresses cell-mediated immunity in vivo: a potential role for leukocyte trafficking. *Brain, Behavior, & Immunity* 11(4):286-306.
- Dhabhar, F.S., and B.S. McEwen. 2001. Bidirectional effects of stress & glucocorticoid hormones on immune function: possible explanations for paradoxical observations. In: *Psychoneuroimmunology*, 3rd edition. R. Ader, D. L. Felten, and N. Cohen, eds. San Diego: Academic Press.
- Drozdzowicz, C.K., T.A. Bowman, M.L. Webb, and C.M. Lang. 1990. Effect of in-house transport on murine plasma corticosterone concentration and blood lymphocyte populations. *American Journal of Veterinary Research* 51(11):1841-6.
- Dymsha, H.A., S.A. Miller, J.F. Maloney, and H.L. Foster. 1963. Equilibration of the laboratory rat following exposure to shipping stresses. *Laboratory Animal Care* 13(1):60-5.
- Eldridge, G.A., C.G. Winfield, and D.J. Cahill. 1988. Responses of cattle to different space allowances, pen sizes and road conditions during transport. *Australian Journal of Experimental Agriculture* 28:155.
- Federation of Animal Science Societies (FASS). 1999. *Guide for the Care and Use of Agricultural Animals in Agricultural Research and Teaching*, 1st revised ed. Savoy, IL: FASS.
- Fewell, J.E., M. Kang, and H.L. Eliason. 1997. Autonomic and behavioral thermoregulation in guinea pigs during postnatal maturation. *Journal of Applied Physiology* 83(3):830-6.
- Friend, T.H. 2000. Dehydration, stress, and water consumption of horses during long-distance commercial transport. *Journal of Animal Science* 78:2568-80.
- Friend, T.H. 2001. A review of recent research on the transportation of horses. *Journal of Animal Science* 79(Suppl. E):E32-40.
- Friend, T.H., M.T. Martin, D.D. Householder, and D.M. Bushong. 1998. Stress responses or horses during a long period of transport in a commercial truck. *Journal of the American Medical Association* 212:838-44.
- Fuller, M.F. 1969. Climate and growth. In: *Animal Growth and Nutrition*. E.S.E. Hafez and I.A. Dyer, eds. Philadelphia, PA: Lea & Febiger.
- Gartner, K., D. Buttner, K. Dohler, R. Friedel, J. Lindena, and I. Trautschold. 1980. Stress response of rats to handling and experimental procedures. *Laboratory Animals* 14(3):267-74.
- Gasser, D.L., L. Mele, D.D. Lees, and A.S. Goldman. 1981. Genes in mice that affect susceptibility to cortison-induced cleft palate are closely linked to Ir genes on chromosomes 2 and 17. *Proceedings of the National Academy of Sciences USA* 78(5):3147-50.
- Goldstein, D.S. 1995. *Stress, Catecholamines, and Cardiovascular Disease*. New York: Oxford University Press.
- Good, A.L., and A.F. Sellers. 1957. Effects of carbon dioxide, epinephrine and ilidar on skin, blood and rectal temperatures of unanesthetized dogs exposed to extreme cold. *American Journal of Physiology* 188(3):451-5.
- Gordon, C. 1985. Relationship between autonomic and behavioral thermoregulation in the mouse. *Physiology & Behavior* 34(5):687-90.
- Gordon, C.J. 1987. Relationship between preferred ambient temperature and autonomic thermoregulator function in rat. *American Journal of Physiology* 252(21):R1130-7.
- Gordon, C.J. 1988. Simultaneous measurement of preferred ambient temperature and metabolism in rats. *American Journal of Physiology* 254(23):R229-34.
- Gordon, C.J. 1990. Thermal biology of the laboratory rat. *Physiology & Behavior* 47(5):963-91.
- Gwosdow, A.R., and E.L. Besch. 1985. Effect of thermal history on the rat's response to varying environmental temperature. *Journal of Applied Physiology* 59(2):413-9.
- Hahn, G.L. 1999. Dynamic responses of cattle to thermal heatloads. *Journal of Animal Science* 77(Suppl. 2):10-20.

- Harada, E., and T. Kanno. 1975. Rabbit's ear in cold acclimation studied on the change in ear temperature. *Journal of Applied Physiology* 38(3):389-94.
- Hayssen, V. 1998. Effect of transatlantic transport on reproduction of agouti and nonagouti deer mice, *Peromyscus maniculatus*. *Laboratory Animals* 32(1):55-64.
- Heetkamp, M.J., J.W. Schrama, W.G. Schouten, and J.W. Swinkels. 2002. Energy metabolism in young pigs as affected by establishment of new groups prior to transport. *Journal of Animal Physiology & Animal Nutrition-Zeitschrift Fur Tierphysiologie Tierernahrung Und Futtermittelkunde* 86(5-6):144-52.
- Hensel, H., and M. Banet. 1982. Adaptive changes in cats after long-term exposure to various temperatures. *Journal of Applied Physiology: Respiratory, Environmental & Exercise Physiology* 52(4):1008-12.
- Herrington, L.P. 1939. *The Heat Regulation of Small Laboratory Animals at Various Environmental Temperatures*. John B. Pierce Laboratory, Connecticut.
- Herrington, L. 1940. The heat regulation of small laboratory animals at various environmental temperatures. *American Journal of Physiology* 129:123-39.
- Himms-Hagen, J., J. Triandafillou, N. Begin-Heick, M. Ghorbani, and A.L. Kates. 1995. Apparent lack of beta 3-adrenoceptors and of insulin regulation of glucose transport in brown adipose tissue of guinea pigs. *American Journal of Physiology* 268(1 Pt 2):R98-104.
- Honda, N., W.V. Judy, and L.D. Carlson. 1962. Effects of adrenaline and noradrenaline on ear vessels in cold- and warm-adapted rabbits. *Journal of Applied Physiology* 17:754-8.
- Horton, G.M. 1981. Responses of shorn and full-fleeced lambs given two levels of feed intake and exposed to warm and cold temperatures. *American Journal of Veterinary Research* 42(12):2151-4.
- Huttunen, P. 1982. Hypothalamic catecholamines and tolerance for severe cold in ethanol-treated guinea-pigs. *British Journal of Pharmacology* 75(4):613-6.
- Huynh, T.T.T., A.J.A. Aarnink, M.W.A. Verstegen, W.J.J. Gerrits, M.J.W. Heetkamp, B. Kemp, and T.T. Canh. 2005. Effects of increasing temperatures on physiological changes in pigs at different relative humidities. *Journal of Animal Science* 83:1385-96.
- International Air Transport Association (IATA). 2005. *Live Animals Regulations*, 32 ed. IATA.
- Irwin, M. 1994. Stress-induced immune suppression: role of brain corticotropin releasing hormone and autonomic nervous system mechanisms. *Advances in Neuroimmunology* 4(1):29-47.
- Jacoby, R.O., and R. Lindsey. 1998. Risks of infection among laboratory rats and mice at major biomedical research institutions. *ILAR Journal* 39(4):266-71.
- Johnson, G.S., and R.S. Elizondo. 1979. Thermoregulation in macaca mulatta: a thermal balance study. *Journal of Applied Physiology: Respiratory, Environmental & Exercise Physiology* 46(2):268-77.
- Jones, S.B., X.J. Musacchia, and G.E. Temple. 1976. Mechanisms of temperature regulation in heat-acclimated hamsters. *American Journal of Physiology* 231(3):707-12.
- Kannan, G., and J.A. Mench. 1996. Influence of different handling methods and crating periods on plasma corticosterone concentrations in broilers. *British Poultry Science* 37:21-31.
- Kenny, F.J., and P.V. Tarrant. 1987. The physiological and behavioral responses of cross-bred Friesen steers to short-haul transport by road. *Livestock Production Science* 17:63-75.
- Kent, J.E., and R. Ewbank. 1983. The effect of road transportation on the blood constituents and behavior of calves. I. Six months old. *The British Veterinary Journal* 139:228-35.
- Kent, J.E., and R. Ewbank. 1986a. The effect of road transportation on the blood constituents and behavior of calves. I. Three months old. *The British Veterinary Journal* 142:326-35.
- Kent, J.E., and R. Ewbank. 1986b. The effect of road transportation on the blood constituents and behavior of calves. II. One to three weeks old. *The British Veterinary Journal* 142:131-40.

- Kiecolt-Glaser, J.K., L. McGuire, T.F. Robles, and R. Glaser. 2002. Emotions, morbidity, and mortality: new perspectives from psychoneuroimmunology. *Annual Review of Psychology* 53:83-107.
- Knowles, T.G., and D.M. Broom. 1990. The handling and transport of broilers and spent hens. *Applied Animal Behaviour Science* 28:75-91.
- Knowles, T.G., P.D. Warriss, S.N. Brown, and J.E. Edwards. 1999. Effects on cattle of transportation by road for up to 31 hours. *The Veterinary Record* 145:575-82.
- Knowles, T.G., S.N. Warriss, S.N. Brown, J.E. Edwards, P.E. Watkins, and A.J. Phillips. 1997. Effects on calves less than one month old of feeding or not feeding them during road transport of up to 24 hours. *The Veterinary Record* 140:116-24.
- Kreger, M., and M. Farris. 2003. International transportation of nonhuman primates: US fish and wildlife service perspective. In: *International Perspectives: The Future of Nonhuman Primate Resources*. NRC. Washington, DC: The National Academies Press.
- Lambooy, E., and B. Hulsegge. 1988. Long-distance transport of pregnant heifers by truck. *Applied Animal Behaviour Science* 20:249-58.
- Landi, M.S., J.W. Kreider, C.M. Lang, and L.P. Bullock. 1982. Effects of shipping on the immune function in mice. *American Journal of Veterinary Research* 43(9):1654-7.
- Lapworth, J. 2004. *Beef cattle transport by road: pre transport management*. Department of Primary Industries and Fisheries. Queensland Government, Brisbane Australia.
- Lindstedt, L., and P.J. Schaeffer. 2002. Use of allometry in predicting anatomical and physiological parameters of mammals. *Laboratory Animals* 36(1):1-19.
- Livestock Conservation Institute. 1970. *Patterns of Transit Losses*. Omaha, NE: Livestock Conservation, Inc.
- Maher, J.A., and T. Schub. 2004. Laboratory rodent transportation supplies. *Lab Animal* 33(8):29-32.
- Malaga, C.A., R.E. Weller, E. Montoya, J. Moro, and R.L. Buschbom. 1991. Mortality and body weight changes in *Aotus nancymai* shipped from Iquitos, Peru to Richland, Washington. *Journal of Medical Primatology* 20(1):6-11.
- Marahrens, M., I. Von Richthofen, S. Schmeiduch, and J. Hartung. 2003. Special problems of long-distance road transports of cattle. *Deutsche Tierärztliche Wochenschrift* 110(3):120-5.
- McEwen, B.S. 2002. *The End of Stress As We Know It*. Washington, DC: Dana Press/Joseph Henry Press.
- McGlone, J., and W.G. Pond. 2003. *Pig Production: Biological Principles and Applications*. Clifton Park, NY: Delmar Publishers.
- McGlone, J.J., J.L. Salak, E.A. Lumpkin, R.I. Nicholson, M. Gibson, and R.L. Norman. 1993. Shipping stress and social status effects on pig performance, plasma cortisol, natural killer cell activity, and leukocyte numbers. *Journal of Animal Science* 71(4):888-96.
- Morgan, K. 1998. Thermoneutral zone and critical temperatures of horses. *Journal of Thermal Biology* 23(1):59-61.
- Mormede, P., J. Soissons, R. Bluthe, J. Raoult, G. Legarff, D. Levieux, and R. Dantzer. 1982. Effect of transportation on blood serum composition, disease incidence, and production traits in young calves. Influence of the journey duration. *Annales De Recherches Veterinaires* 13(4):369-84.
- Nagasaka, T., and L.D. Carlson. 1965. Responses of cold- and warm-adapted dogs to infused norepinephrine and acute body cooling. *American Journal of Physiology* 209(1):227-30.
- National Institutes of Health (NIH). 2002. *NCRR Fact Sheet*. Bethesda, MD: NIH.
- National Research Council (NRC). 1981. *Effect of Environment on Nutrient Requirements of Domestic Animals*. Washington, DC: National Academy Press.
- National Research Council (NRC). 1996. *Guide for the Care and Use of Laboratory Animals*. Washington, DC: National Academy Press.

- National Research Council (NRC). 2003a. *Guidelines for the Care and Use of Mammals in Neuroscience and Behavioral Research*. Washington, DC: The National Academies Press.
- National Research Council (NRC). 2003b. *Occupational Health and Safety in the Care and Use of Nonhuman Primates*. Washington, DC: The National Academies Press.
- Nyberg, L., K. Lundstrom, I. Edfors-Lilja, and M. Rundgren. 1988. Effects of transport stress on concentrations of cortisol, corticosteroid-binding globulin and glucocorticoid receptors in pigs with different halothane genotypes. *Journal of Animal Science* 66(5):1201-11.
- Oddershede, I.R., and R.S. Elizondo. 1980. Body fluid and hematologic adjustments during resting heat acclimation in rhesus monkey. *Journal of Applied Physiology: Respiratory, Environmental & Exercise Physiology* 49(3):431-7.
- Oddershede, I.R., and R.S. Elizondo. 1982. Body fluid and hematologic adjustments during resting cold acclimation in rhesus monkey. *Journal of Applied Physiology: Respiratory, Environmental & Exercise Physiology* 52(4):1024-9.
- Ohara, K., F. Furuyama, and Y. Isobe. 1975. Prediction of survival time of rats in severe heat. *Journal of Applied Physiology* 38(4):724-9.
- Otto, G., and R.J. Tolwani. 2002. Use of microisolator caging in a risk-based mouse import and quarantine program: a retrospective study. *Contemporary Topics in Laboratory Animal Science* 41(1):20-7.
- Oufara, S., H. Barre, J.L. Rouanet, and J. Chatonnet. 1987. Adaptation to extreme ambient temperatures in cold-acclimated gerbils and mice. *American Journal of Physiology* 253(1 Pt 2):R39-45.
- Palmer, A.E. 1987. B virus, herpesvirus simiae: historical perspective. *Journal of Medical Primatology* 16(2):99-130.
- Pekow, C. 2005. Defining, measuring, and interpreting stress in laboratory animals. *Contemporary Topics in Laboratory Animal Science* 44(2):41-5.
- Pharmaceutical Research and Manufacturers of America (PhRMA). 2005. *PhRMA Annual Report 2004-1005*.
- Pohl, H. 1965. Temperature regulation and cold acclimation in the golden hamster. *Journal of Applied Physiology* 20:405-10.
- Robertshaw, D. 2004. Temperature regulation and the thermal environment. In: *Dukes' Physiology of Domestic Animals*, 12th ed. W.O. Reece, ed. Ithaca and London: Cornell University Press.
- Robinson, J., and G. Beattie. 2003. Nonhuman primate resource needs: a moving target. In: *International Perspectives: The future of nonhuman primate resources*. NRC. Washington, DC: The National Academies Press.
- Rubio, C.A., and C.B. Huang. 1992. Quantification of the sulphomucin-producing cell population of the colonic mucosa during protracted stress in rats. *In Vivo* 6(1):81-4.
- Sackett, G.P. 1981. Pregnancy outcome following jet transport stress in nonhuman primates. *Journal of Medical Primatology* 10(2-3):149-54.
- Sapolsky, R.M. 1998. *Why Zebras Don't Get Ulcers: A guide to stress, stress-related disease, and coping*. W.H. Freeman and Company.
- Schaeffer, P.J., J.F. Hokanson, D.J. Wells, and S.L. Lindstedt. 2001. Cold exposure increases running VO_{2max} and cost of transport in goats. *American Journal of Physiology—Regulatory Integrative & Comparative Physiology* 280(1):R42-7.
- Schlenker, G.R., and W. Muller. 1997. Air transportation of day-old chicks with regard to animal welfare. *Berliner Und Munchener Tierarztliche Wochenschrift* 110(9):315-19.
- Schrama, J.W., W. van der Hel, A. Arieli, and M.W. Verstegen. 1992. Alteration of energy metabolism of calves fed below maintenance during 6 to 14 days of age. *Journal of Animal Science* 70(8):2527-32.

- Schrama, J.W., W. van der Hel, J. Gorssen, A.M. Henken, M.W. Verstegen, and J.P. Noordhuizen. 1996. Required thermal thresholds during transport of animals. *Veterinary Quarterly* 18(3):90-5.
- Selye, H. 1975. Stress and distress. *Comprehensive Therapy* 1:9-13.
- Sharp, J., T. Azar, and D. Lawson. 2005. Selective adaptation of male rats to repeated social encounters and experimental manipulations. *Contemporary Topics in Laboratory Animal Science* 44(2):28-31.
- Silverman, J., M.A. Suckow, and S. Murthy, eds. 2000. *The IACUC Handbook*. Boca Raton, FL: CRC Press.
- Simensen, E.B., B. Laksesvela, A.K. Blom, and O.V. Sjaastad. 1980. Effects of transportation, a high lactose diet and ACTH injections on the white blood cell count, serum cortisol and immunoglobulin G in young calves. *Acta Veterinaria Scandinavica* 21:278-90.
- Slanetz, C.A., I. Fratta, C.W. Clouse, and S.C. Jones. 1957. Stress and transportation of animals. *Proceedings of the Animal Care Panel* 7:278-89.
- Sobel, H., G.C. Haberfelde, and A.E. Reeves. 1965. Reversibility of endocrine changes produced in guinea pigs by exposure to cold. *American Journal of Physiology* 208(1):115-17.
- Sobel, H., M. Sideman, and R. Arce. 1960. Steroid excretion by guinea pigs exposed to cold for prolonged periods. *American Journal of Physiology* 198(5):1107-10.
- Stitt, J.T., and J.D. Hardy. 1971. Thermoregulation in the squirrel monkey (*Saimiri sciureus*). *Journal of Applied Physiology* 31(1):48-54.
- Stull, C.L., and A.V. Rodiek. 2000. Physiological responses of horses to 24 h of road transportation using a commercial van during summer conditions. *Journal of Animal Science* 78:1458-66.
- Swallow, J., D. Anderson, A. Buckwell, T. Harris, P. Hawkins, J. Kirkwood, M. Lomas, S. Meacham, A. Peters, M. Prescott, S. Owen, R. Quest, R. Sutcliffe, and K. Thompson. 2005. Guidance on the transport of laboratory animals. *Laboratory Animals* 39(1):1-39.
- Swanson, J.C., and J. Morrow-Tesch. 2001. Cattle transport: historical, research and future perspectives. *Journal of Animal Science* 79(Suppl. E):E102-9.
- Swift, R., and R.M. Forbes. 1939. The heat production of the fasting rat in relation to the environmental temperature. *Journal of Nutrition* 18:307-18.
- Szymusiak, R., and E. Satinoff. 1981. Maximal REM sleep time defines a narrower thermoneutral zone than does minimal metabolic rate. *Physiology & Behavior* 26(4):687-90.
- Tamashiro, K.L., M.M. Nguyen, and R.R. Sakai. 2005. Social stress: from rodents to primates. *Frontiers in Neuroendocrinology* 26(1):27-40.
- Tanaka, A., and H. Xin. 1997. Effects of structural and stacking configuration of containers for transporting chicks in their microenvironment. *Transactions of the ASAE* 40(3):777-82.
- Tarrant, P.V., F.J. Kenny, D. Harrington, and M. Murphy. 1992. Long distance transportation of steers to slaughter: effect of stocking density on physiology, behavior and carcass quality. *Livestock Production Science* 30:223-38.
- Tarrant, V., and T. Grandin. 2000. Cattle transport. In: *Livestock Handling and Transport*. New York: CABI Publishing.
- Trull, F.L., and B.A. Rich. 1999. More regulation of rodents. *Science* 284(5419):1463.
- US Department of Agriculture (USDA). 2004. *Animal Welfare Report: Fiscal Year 2004*. Washington, DC: USDA.
- van Ruyven, F., G.W. Meijer, L.F.M. van Zutphen, and J. Ritskes-Hoitinga. 1996. Adaptation period of laboratory animals after transport: a review. *Scandinavian Journal of Laboratory Animal Science* 23(4):185-90.
- Vapaatalo, H., J. Hirvonen, and P. Huttunen. 1984. Lowered cold tolerance in cold-acclimated and non-acclimated guinea pigs treated with diazepam. *Zeitschrift Fur Rechtsmedizin—Journal of Legal Medicine* 91(4):279-86.

- Wallace, M.E. 1976. Effects of stress due to deprivation and transport in different genotypes of house mouse. *Laboratory Animals* 10(3):335-47.
- Warriss, P., S. Brown, T. Knowles, S. Kestin, J. Edwards, S. Dolan, and A.J. Phillips. 1995. Effects on cattle of transport by road for up to 15 hours. *The Veterinary Record* 136(13):319-23.
- Warriss, P.D., S.N. Brown, J.E. Edwards, M.H. Anil, and D.P. Fordham. 1992. Time in lairage needed by pigs to recover from the stress of transport. *Veterinary Record* 131(9):194-6.
- Weinert, D., H. Eimert, H.G. Erkert, and U. Schneyer. 1994. Resynchronization of the circadian corticosterone rhythm after a light/dark shift in juvenile and adult mice. *Chronobiology International* 11(4):222-31.
- Weisbroth, S.H., R.G. Paganelli, and M. Salvia. 1977. Evaluation of a disposable water system during shipment of laboratory rats and mice. *Laboratory Animal Science* 27(2):186-94.
- West, G.B., J.H. Brown, and B.J. Enquist. 1997. A general model for the origin of allometric scaling laws in biology. *Science* 276(5309):122-6.
- White, W. 2004. *Shipment of Animals by Breeders: Presentation at the NRC Workshop on Guidelines for the Humane Transport of Laboratory Animals*. Washington, DC: NRC.
- Whiting, T.L., and S. Brandt. 2002. Minimum space allowance for transportation of swine by road. *Canadian Veterinary Journal* 43(3):207-12.
- Wolfensohn, S.E. 1997. Brief review of scientific studies of the welfare implications of transporting primates. *Laboratory Animals* 31(4):303-5.
- Wright, G.L. 1976. Critical thermal maximum in mice. *Journal of Applied Physiology* 40(5):683-7.
- Xin, H. 1997. Mortality and body weight of breeder chicks as influenced by air temperature fluctuations. *Journal of Applied Poultry Research* 6:199-204.
- Xin, H., and J.D. Harmon. 1996. Responses of group-housed neonatal chicks to posthatch holding environment. *Transactions of the ASAE* 39(6):2249-54.
- Xin, H., and K. Lee. 1996. Use of Aqua-Jel® and feed for nutrient supply during long journey air transport of baby chicks. *Transactions of the ASAE* 39(3):1123-6.
- Young, B.A. 1975. Effects of winter acclimatization on resting metabolism of beef cows. *Canadian Journal of Animal Science* 55:619.

Appendix A

Summary of the Animal Welfare Act Regulations Pertaining to Transportation

This summary of Animal Welfare Act (AWA) standards is divided into three sections. The first and second cover regulations that pertain to the transportation of dogs and cats and of nonhuman primates, respectively. The third section covers AWA regulations that pertain to the transportation of guinea pigs, hamsters, rabbits, and animals defined as other animals by the US Department of Agriculture. *Other animals* would include any species of warm-blooded animal other than dog, cat, guinea pig, hamster, rabbit, nonhuman primate, marine mammal, or bird. AWA regulations pertaining to marine mammals are not included in this summary but can be found in 9 CFR 3.112-3.118. This summary is not meant to be exhaustive. For example, specific directions on water temperatures and detergents that should be used to sanitize transportation enclosures are not included. However, each section refers to the original AWA regulation if further specifics are required.

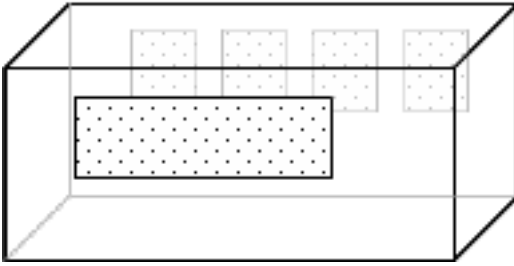
DOGS AND CATS (9 CFR 3.13 – 3.19)

Enclosure

1. The primary enclosure must be strong enough to contain the animals securely and comfortably and to withstand the normal rigors of transportation.
2. Enclosures must be large enough that each animal has enough space to turn around normally while standing, to sit and stand, and to lie in a natural position.

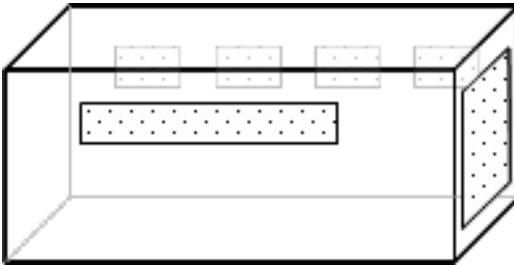
3. Interior has no sharp points, edges, or protrusions that could injure an animal.
4. Each animal is securely contained in the enclosure and cannot put any part of its body outside the enclosure in a way that could result in injury to itself, handlers, or other persons or animals.
5. The openings of the enclosure are easily accessible at all times for emergency removal of the animals.
6. Unless the enclosure is permanently affixed to the conveyance, adequate handles or handholds should be present so that it can be lifted without tilting and ensure that anyone handling the enclosure will not come into physical contact with an animal that is inside.
7. Unless the enclosure is permanently affixed to the conveyance, labeling on the top and at least one side with the words "Live Animals" in letters at least 1 in. high and arrows indicating the correct upright position of the enclosure must be present.
8. Any material or treatment in or on the enclosure must be nontoxic.
9. A solid, leakproof bottom or removable leakproof tray under a slatted or mesh floor that prevents seepage of waste products from the enclosure is required. The slatted or mesh floor must be constructed so an animal cannot put any part of its body through the raised floor. Unless it is on a raised floor, the enclosure must contain enough unused, nontoxic litter to absorb and cover excreta.
10. Food and water receptacles must be attached inside the enclosure and placed so that they can be filled from outside the enclosure without opening the door.
11. The primary enclosure must be cleaned and sanitized before each use. If an animal is in transit for more than 24 hr, the enclosure must be cleaned and litter replaced or the animal moved to another clean and sanitized enclosure. If moving an animal from the enclosure is necessary, this procedure must be completed in a way that safeguards the animal from injury and prevents escape.
12. Projecting rims must be on the exterior of walls that contain ventilation openings and provide a minimal air circulation space of 0.75 in.
13. The size and arrangement of ventilations openings must comply with one of the following options:

Option A: Two ventilation openings



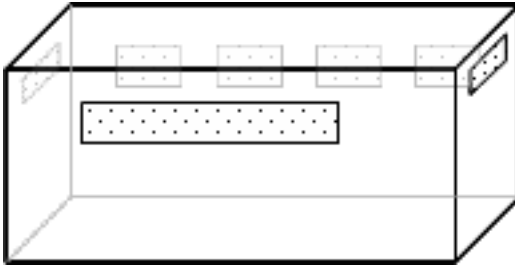
Surface area of each opening is at least 16% of wall surface area.
 Total surface area of openings is at least 14% of total surface area of all walls.
 One-third of surface area of openings must be in upper half of enclosure.

Option B: Three ventilation openings



Total surface area of openings on opposing walls is at least 8% of those walls' total surface area.
 Surface area of openings on third wall is at least 50% of surface area of that wall.
 Total surface area of openings is at least 14% of total surface area of all walls.
 One-third of surface area of openings must be in upper half of enclosure.

Option C: Four ventilation openings



Surface area of each opening is at least 8% of surface area of that wall.
 Total surface area of openings is at least 14% of total surface area of all walls.
 One-third of surface area of the openings must be in upper half of enclosure.

14. If the enclosure is permanently affixed to the conveyance so that there is only a front ventilation opening, the opening cannot be blocked, must open directly to an unobstructed aisle, must be at least 90% of the total surface area of the front wall, and must be covered with bars, wire mesh, or smooth expanded metal that has air spaces.

Compatibility

1. Dogs or cats transported in the same enclosure must be of a single species and be maintained in compatible groups.
2. Puppies or kittens 4 months old or younger cannot be transported in an enclosure with adult animals other than their dams.
3. Animals that are overaggressive or exhibit a vicious disposition must be transported individually.
4. A female in estrus may not be transported in an enclosure with a male animal.
5. When transportation is by air,
 - a. No more than one animal 6 months old or older or weighing over 20 lb may be transported in the same enclosure.
 - b. No more than two puppies or kittens 8 weeks to 6 months old, of comparable size, and weighing 20 lb or less each may be transported in the same enclosure.
 - c. Weaned animals less than 8 weeks old must be of comparable size or must be littermates and be accompanied by their dam.

6. When transportation is on the ground or by privately owned aircraft,
 - a. No more than 4 animals 8 weeks old or older and of comparable size may be transported in the same enclosure.
 - b. Weaned animals less than 8 weeks old and of comparable size or animals less than 8 weeks old that are littermates and are accompanied by their dam may be transported in the same enclosure.

Food and Water

1. Each animal 16 weeks old or older must be offered food at least once every 24 hr.
2. Animals less than 16 weeks old must be offered food at least once every 12 hr.
3. Each animal must be offered water at least once every 12 hr.
4. Those periods start when an animal was last offered food and water before transportation began.
5. Each animal must be offered food and water within 4 hr before being transported or within 4 hr before delivery to a carrier or intermediate handler.
6. It is not acceptable to withhold food or water unless withholding is directed by a veterinarian.

Documentation

1. Certifications must be securely attached to the outside of the primary enclosure in a manner that makes them easily noticed and read.
2. For surface transportation only, the operator of the conveyance may hold documents. Certification must include
 - a. Consignor's name, address, and dated signature.
 - b. Consignee's name, address, and telephone number.
 - c. Each animal's tattoo or tag number.
 - d. Time and date when each animal was last fed and watered (not to occur more than 4 hr before delivery to carrier or intermediate handler).
 - e. Instructions for satisfying in-transit food and water requirements for a 24-hr period, administration of drugs or medication, or other special care.
3. Certification of acclimation (required only if temperature of animal holding area is less than 45°F) must include
 - a. Consignor's name and address.
 - b. Each animal's tattoo or tag number.
 - c. Signed statement from veterinarian, dated no more than 10 days before delivery, that each animal is acclimated to temperatures

lower than 50°F, and the specific minimal temperature that the animal is acclimated to.

Delivery to Carriers and Handlers

1. Carriers and handlers cannot accept an animal for transportation more than 4 hr before departure time; this can be extended by 2 hr by agreement of the carrier and consignor.
2. Carriers and handlers cannot accept an animal for transportation unless the enclosure meets the requirements described above, the enclosure is free of obvious defects, the appropriate written certifications listed above are attached to the enclosure, and the carriers' holding area meets the minimal temperature requirements described below under "Terminal Facilities" unless the animal is accompanied by a certificate of acclimation (see "Documentation" above).

Terminal Facilities

1. Shipments of animals cannot be comingled with inanimate cargo in animal holding areas of terminal facilities.
2. All holding areas must be cleaned and sanitized as often as necessary to prevent an accumulation of debris and to minimize disease hazards; an effective program must be implemented for the control of insects, ectoparasites and birds and mammals considered pests.
3. Ventilation must be provided to minimize drafts, odors, and moisture condensation; auxiliary ventilation must be used when the ambient temperature is 85°F or higher.
4. Temperature must not be below 45°F or above 85°F for more than 4 consecutive hours; temperature must be measured at a point not more than 3 ft away from the outside of the enclosure and halfway up the side of the enclosure.
5. The facility must provide shelter from sunlight, extreme heat, rain, and snow.
6. Upon arrival at a facility, if a consignee cannot be notified with 24 hr of arrival, the handler must return the animal to the consignor; if the consignee is notified but does not accept delivery within 48 hr of arrival, the handler must return the animal to the consignor.

Handling

1. When an animal is moved to or from a facility or conveyance, it must be sheltered from sunlight, extreme heat, rain, and snow.

2. The animal cannot be exposed to temperature above 85°F for more than 45 min.
3. Transporting devices must be covered when the temperature falls below 50°F; the animal cannot be exposed to temperatures below 45°F for more than 45 min unless it is accompanied by a certificate of acclimation; temperature is measured from a point not more than 3 ft from the outside of the enclosure and halfway up the side of the enclosure.
4. Handlers must avoid causing physical harm or distress to the animal.
5. The enclosure cannot be placed on an unattended conveyor belt or on an elevated conveyor belt or tossed, dropped, needlessly tilted, or stacked in a manner that results in its falling.
6. The enclosure must be handled in a manner consistent with written instructions attached to it.

Primary Conveyance

1. The cargo space of conveyances to transport animals must be designed, constructed, and maintained in a manner that protects the health and well-being of each animal and ensures its safety and comfort.
2. The cargo space must have a supply of air that is sufficient for normal breathing of all animals, and entry of engine exhaust must be prevented.
3. Enclosures must be positioned in the cargo space in a manner that provides protection from the elements, allows each animal enough air for normal breathing, and allows for the removal of the animals in the event of an emergency.
4. During surface transportation, auxiliary ventilation must be used in the cargo space when the temperature reaches 85°F. The temperature may not exceed 85°F or fall below 45°F for a period of more than 4 hr.
5. During air transportation, cargo areas must be heated or cooled as necessary to maintain a temperature that ensures the health and well-being of each animal. The cargo area must be pressurized in flights above 8,000 ft.
6. The cargo space must be kept clean and animals may not be transported with any items that may be expected to cause harm to them.

Care in Transit

1. During surface transportation, the operator of the conveyance must observe the animals at least once every 4 hr to ensure sufficient air and acceptable ambient temperatures. If an animal is in obvious physical distress, the operator must obtain appropriate veterinary care at the closest available facility.

2. During air transport, animals must be observed at least once every 4 hr unless the cargo area is not accessible during flight, in which case the animals must be observed whenever they are loaded or unloaded to ensure sufficient air and acceptable ambient temperatures. If an animal is in obvious physical distress, veterinary care must be provided as soon as possible.
3. During transportation, animals shall not be removed from their enclosure unless placed in another enclosure or facility that conforms to the appropriate AWA regulations.

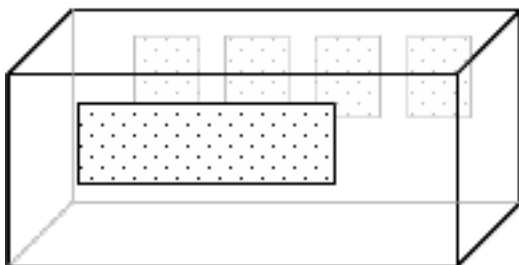
NONHUMAN PRIMATES (9 CFR 3.86-3.92)

Enclosure

1. The primary enclosure must be strong enough to contain the animals securely and comfortably and to withstand the normal rigors of transportation.
2. The enclosure must be large enough that each animal has enough space to turn around normally and to sit without its head touching the top of the enclosure. Larger species may be restricted in their movements when freedom of movement would be dangerous to the animals or people.
3. The enclosure interior should not have sharp points, edges, or protrusions that could injure the animal.
4. The animal must be securely contained in the enclosure and unable to put any part of its body outside the enclosure in a way that could result in injury to it, handlers, or other persons or animals.
5. Openings of enclosures must be easily accessible at all times for emergency removal of animals.
6. Openings of enclosure must be secure with animalproof devices that prevent accidental opening, including opening by nonhuman primates.
7. Unless the enclosure is permanently affixed to the conveyance, adequate handles or handholds must be provided so that it can be lifted without tilting and to ensure that anyone handling it will not come into physical contact with the animal in it.
8. Ventilation openings should be covered with bars, wire mesh, or smooth expanded metal having air spaces.

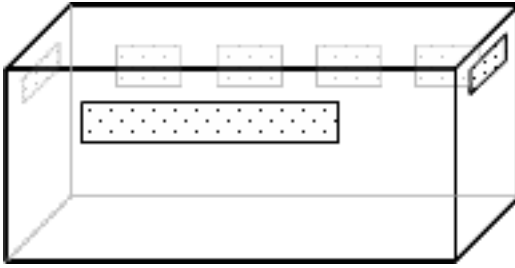
9. All enclosures must be marked on the top and at least one side with the words "Live Animals" or "Wild Animals" in letters at least 1 in. high and with arrows indicating the correct upright position of the enclosure.
10. Any material or treatment in or on the enclosure should be nontoxic.
11. A solid, leakproof bottom or removable leakproof tray under a slatted or mesh floor that prevents seepage of waste products outside of the enclosure is required. The slatted or mesh floor must be constructed so that the animal cannot put any part of its body through the raised floor. The enclosure must contain enough unused, nontoxic litter to absorb and cover excreta.
12. Food and water receptacles must be attached inside the enclosure and placed so that they can be filled from outside the enclosure without opening the door. Food and water receptacles must be designed and installed so that a nonhuman primate cannot leave the primary enclosure through the food or water opening.
13. The primary enclosure must be cleaned and sanitized before each use.
14. Projecting rims must be on the exterior of walls that contain ventilation openings and provide a minimal air circulation space of 0.75 in.
15. The size and arrangement of ventilations openings must comply with one of the following options:

Option A: Two ventilation openings



Surface area of each opening is at least 16% of wall surface area.
Each opening must be above midline of enclosure.

Option B: Four ventilation openings



Surface area of each opening is at least 8% of surface area of that wall.
Each opening must be above midline of enclosure.

16. If the enclosure is permanently affixed to the conveyance so that there is only a front ventilation opening, this opening cannot be blocked, must open directly to an unobstructed aisle, must be at least 90% of the total surface area of the front wall, and must be covered with bars, wire mesh, or smooth expanded metal having air spaces.

Compatibility

1. Only one animal may be transported in a primary enclosure, except for
 - a. A mother and her nursing infant.
 - b. An established male-female pair or family group (but a female in estrus cannot be transported with a male nonhuman primate).
 - c. A compatible pair of juveniles of the same species that have not reached puberty.
2. Nonhuman primates of different species cannot be transported in adjacent or connecting enclosures.

Food and Water

1. Each animal 1 year old or older must be offered food at least once every 24 hr and water at least once every 12 hr.
2. Animals less than 1 year old must be offered food and water at least once every 12 hr.
3. Those periods start when an animal was last offered food and water before transportation began.
4. Each animal must be offered food and water within 4 hr before being transported or within 4 hr before delivery to a carrier or intermediate handler.

Documentation

1. Certifications must be securely attached to the outside of the primary enclosure in a manner that makes them easily noticed and read.
2. For surface transportation only, documents may be held by the operator of the conveyance. Certification must include
 - a. Consignor's name, address, and dated signature.
 - b. Consignee's name, address, and telephone number.
 - c. Species of nonhuman primate.
 - d. Time and date when animal was last fed and watered (not to occur more than 4 hr before delivery to carrier or intermediate handler).
 - e. Instructions for in-transit food and water requirements for a 24-hr period, administration of drugs or medication, or other special care required.
3. Certification of acclimation (required only if the temperature of the animal holding area is less than 45°F) must include
 - a. Consignor's name and address.
 - b. Species of nonhuman primate.
 - c. Signed statement from veterinarian, dated no more than 10 days before delivery, that the animal is acclimated to temperatures lower than 50°F and the specific minimal temperature the animal is acclimated to.

Delivery to Carriers and Handlers

1. Carriers and handlers cannot accept an animal for transportation more than 4 hr before departure time. This can be extended by 2 hr by agreement of the carrier and consignor.
2. Carriers and handlers cannot accept an animal for transportation unless the enclosure meets the requirements described above, the enclosure is free of obvious defects, the appropriate written certifications listed above are attached to the enclosure, and the carriers' holding areas meet the minimal temperature requirements described below under "Terminal Facilities" unless the animal is accompanied by a certificate of acclimation (see "Documentation" above).

Terminal Facilities

1. Shipments of animals cannot be comingled with inanimate cargo in animal holding areas of terminal facilities. Nonhuman primates must not be placed near any other animals, including other species of nonhuman primates, and must not be able to touch or see any other animals, including other species of nonhuman primates.

2. All holding areas must be cleaned and sanitized as often as necessary to prevent an accumulation of debris and to minimize disease hazards. An effective program must be implemented for the control of insects, ectoparasites, and birds and mammals considered pests.
3. Ventilation must be provided to minimize drafts, odors, and moisture condensation. Auxiliary ventilation must be used when the ambient temperature is 85°F or higher.
4. The temperature must not fall below 45°F or rise above 85°F for more than 4 consecutive hr. Temperature must be measured at a point not more than 3 ft away from the outside of the enclosure and halfway up the side of the enclosure.
5. The facility must provide shelter from sunlight, extreme heat, rain, and snow.
6. Upon arrival at a facility, the carrier or handler must attempt to notify the consignee immediately and once every 6 hr thereafter. The time, date, and method of attempted notification, actual notification of consignee, and name of the person who notified or attempted to notify the consignee must be written either on the carrier's copy of the shipping document or on the copy that accompanies the primary enclosure. If a consignee cannot be notified with 24 hr of arrival, the handler must return the animal to the consignor. If the consignee is notified but does not accept delivery within 48 hr of arrival, the handler must return the animal to the consignor.

Handling

1. When an animal is moved to or from a facility or conveyance, it must be sheltered from sunlight, extreme heat, rain, and snow.
2. The animal cannot be exposed to a temperature above 85°F for more than 45 min.
3. Transporting devices must be covered when the temperature falls below 45°F; the animal cannot be exposed to temperatures below 45°F for more than 45 min unless it is accompanied by a certificate of acclimation; temperature is measured from a point not more than 3 ft from the outside of the enclosure and halfway up the side of the enclosure.
4. Handlers must avoid causing physical harm or distress to the animal.
5. The enclosure cannot be placed on an unattended conveyor belt or on an elevated conveyor belt or tossed, dropped, needlessly tilted, or stacked in a manner that results in its falling.
6. The enclosure must be handled in a manner consistent with written instructions attached to it.

Primary Conveyance

1. The cargo space of conveyances to transport animals must be designed, constructed, and maintained in a manner that protects the health and well-being of each animal and ensures its safety and comfort.
2. The cargo space must have a supply of air that is sufficient for normal breathing of all animals, and entry of engine exhaust must be prevented.
3. Enclosures must be positioned in the cargo space in a manner that provides protection from the elements, allows each animal enough air for normal breathing, and allows for the removal of the animals in the event of an emergency.
4. During surface transportation, temperatures inside cargo areas must be kept at between 45°F to 85°F at all times.
5. During air transportation, cargo areas must be maintained at a level that ensures the health and well-being of each animal at all times.
6. The cargo space must be kept clean, and animals may not be transported with any items that may be expected to cause harm to them.
7. Enclosures containing nonhuman primates must be placed far enough away from animals that are predators or natural enemies. Regardless of whether the other animals are in enclosures, the nonhuman primates should not be able to touch or see them.

Care in Transit

1. During surface transportation, the operator of the conveyance must observe the animals at least once every 4 hr to ensure sufficient air and acceptable ambient temperatures. If an animal is in obvious physical distress, the operator must obtain appropriate veterinary care at the closest available facility.
2. During air transportation, animals must be observed at least once every 4 hr unless the cargo area is not accessible during flight, in which case animals must be observed whenever they are loaded or unloaded to ensure sufficient air and acceptable ambient temperatures. If an animal is in obvious physical distress, veterinary care must be provided as soon as possible.
3. During transportation, animals shall not be removed from their enclosure unless it is to be placed in another enclosure or facility that conforms to the appropriate AWA regulations. Only persons who are experienced and authorized by the shipper, consignor, or consignee may remove a nonhuman primate from its enclosure during transportation, unless it is required for the health or well-being of the animal.

GUINEA PIGS AND HAMSTERS, RABBITS, AND OTHER ANIMALS
(9 CFR 3.35-3.41, 9 CFR 3.60-3.65, 9 CFR 3.136-3.142)

Enclosure

1. The primary enclosure must be strong enough to contain the animals securely and comfortably and to withstand the normal rigors of transportation.
2. The enclosure must be large enough that each animal has enough space to turn around normally while standing and to stand and lie in a natural position. Some species may be restricted in their movements when freedom of movement would constitute a danger to the animals or people.
3. Enclosure interiors must have no protrusions that could injure the animals.
4. For guinea pigs and hamsters, inner surfaces of fiberboard, cardboard, or plastic enclosures must be covered or laminated with wire mesh or screen where necessary to prevent escape.
5. Openings of the enclosures must be easily accessible at all times for emergency removal of animals.
6. Unless the enclosure is permanently affixed to the conveyance, adequate handles or handholds must be provided so that the enclosure can be lifted without tilting and to ensure that anyone handling it will not come into physical contact with animals in it.
7. Projecting rims must be on the exterior of walls that contain ventilation openings and provide a minimal air circulation space of 0.75 in.
8. Unless the enclosure is permanently affixed to the conveyance, it must be marked on top and on at least one side with the words "Live Animals" or "Wild Animals" in letters at least 1 in high and with arrows indicating the correct upright position of the enclosure.
9. A solid, leakproof bottom that prevents seepage of waste products outside the enclosure is required. Unless it is on a wire or other nonsolid floor, the enclosure must contain enough unused, nontoxic litter to absorb and cover excreta.
10. The primary enclosure must be cleaned and sanitized before each use.
11. The interior height and space per animal of each enclosure must comply with the following minimal standards for guinea pigs and hamsters (there are no minimal requirements for rabbits and other animals):

Guinea Pigs

<i>Weight</i>	<i>Minimal Interior Height of Enclosure</i>
<500 g	15.2 cm
>500 g	17.8 cm

<i>Weight</i>	<i>Minimal Enclosure Space per Animal</i>
<350 g	193.6 cm ²
350 to 600 g	290.3 cm ²
>600 g	354.8 cm ²

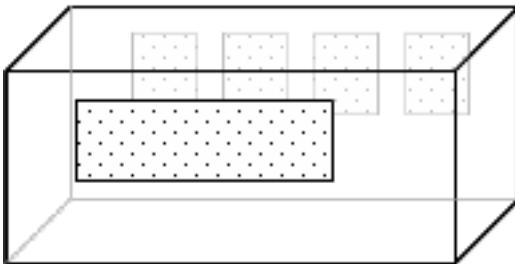
Hamsters

	<i>Minimal Interior Height of Enclosure</i>
Dwarf Hamsters	12.7 cm
All Other Hamsters	15.2 cm

	<i>Minimal Enclosure Space per Animal</i>	
<i>Age</i>	<i>Dwarf Hamsters</i>	<i>All Other Hamsters</i>
Weaning-5 weeks	32.2 cm ²	45.2 cm ²
5 to 10 weeks	48.3 cm ²	71.0 cm ²
>10 weeks	58.1 cm ²	96.8 cm ²

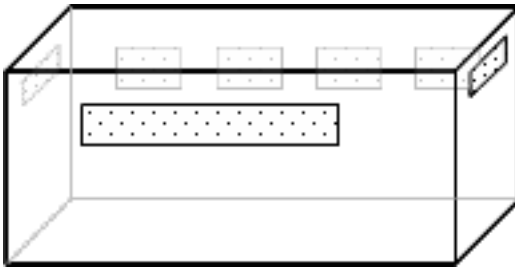
12. The size and arrangement of ventilations openings must comply with one of the following options:

Option A: Two ventilation openings



Surface area of each opening is at least 16% of surface area of that wall. One-third of surface area of openings must be in lower half of enclosure, and at least one-third of surface area of openings must be on upper half of enclosure.

Option B: Four ventilation openings



Surface area of each opening is at least 8% of surface area of that wall. One-third of surface area of openings must be in lower half of enclosure, and at least one-third of surface area of openings must be on upper half of enclosure.

13. If the enclosure is permanently affixed to the conveyance so that there is only a front ventilation opening, this opening cannot be blocked, must open directly to an unobstructed aisle, must be at least 90% of the total surface area of the front wall, and must be covered with bars, wire mesh, or smooth expanded metal.

Compatibility

1. Animals transported in the same enclosure must be of the same species and be maintained in compatible groups.
2. No more than 15 rabbits, 15 guinea pigs, or 50 hamsters can be transported in the same enclosure. For all other animals, animals that have not reached puberty shall not be transported in the same enclosure with adult animals other than their dams; socially dependent animals (such as sibling, dam, and other members of a family group) must be allowed visual and olfactory contact. A female animal in estrus shall not be transported in the same enclosure with a male animal.

Food and Water

1. For guinea pigs, hamsters, and rabbits when transported for a period of more than 6 hr, food and water or a type of food that satisfies the requirement for food and water must be provided in the primary enclosure.
2. For all other animals, water must be offered within 4 hr before transportation and at least once every 12 hr after initiation of transporta-

tion. Consigned animals must be provided with water at least once every 12 hr after acceptance for transportation. Exceptions are allowed for animals that require water more frequently, are hibernating, or are under veterinary treatment.

3. Each animal must be fed at least once every 24 hr; exceptions are allowed for animals that require food more frequently, are hibernating, or are under veterinary treatment.

Documentation

1. Certifications must be securely attached to the outside of the primary enclosure in a manner that makes them easily noticed and read.
2. For surface transport only, documents may be held by the operator of the conveyance. Certification must include
 - a. Consignor's name, address, and dated signature.
 - b. Number of animals in enclosure.
 - c. A statement that enclosure complies with US Department of Agriculture standards.
 - d. 24 hr feeding instructions for animals other than guinea pigs, hamsters, and rabbits (whose enclosures should already contain sufficient food and water for the duration of transportation).
3. Certification of acclimation (required only if the temperatures of the animal holding area is less than 45°F) must include (it is important to note that the AWA does not recognize that guinea pigs can be acclimated to temperatures below than 45°F):
 - Consignor's name and address.
 - Number of animals in shipment.
 - Signed statement from veterinarian, dated no more than 10 days before delivery, that the rabbit or hamster is acclimated to temperatures lower than 45°F and the specific minimal temperature the animal is acclimated to.

Delivery to Carrier and Handlers

1. Carriers and handlers cannot accept an animal for transportation more than 4 hr before departure time. This can be extended by 2 hr by agreement of the carrier and consignor.
2. Carriers and handlers cannot accept an animal for transportation unless the enclosure meets the requirements described above or the consignor furnishes the carrier with a certificate stating that the enclosure complies with the standards set forth in the AWA, the enclosure is free from obvious defects, the appropriate written certifications listed above are attached to the enclosure, and the carriers' holding

areas meet the minimal temperature requirements described below under “Terminal Facilities” unless the animal is accompanied by a certificate of acclimation (see “Documentation” above).

Terminal Facilities

1. Shipments of animals cannot be commingled with inanimate cargo in animal holding areas of terminal facilities.
2. All holding areas must be cleaned and sanitized as often as necessary to prevent an accumulation of debris and to minimize disease hazards. An effective program must be implemented for the control of insects, ectoparasites, and birds and mammals considered pests.
3. Ventilation must be provided to minimize drafts, odors, and moisture condensation. Auxiliary ventilation must be used when the ambient temperature is 75°F or higher.
4. The temperature must not fall below 45°F or rise above 85°F ever or for more than 4 consecutive hr. Temperature must be measured at a point not more than 3 ft away from the outside of the enclosure and halfway up the side of the enclosure.
5. On arrival at a facility, the carrier or handler must attempt to notify the consignee at least once every 6 hr following arrival. The time, date, and method of each attempt at notification, final notification, and name of the person notifying the consignee shall be recorded on the copy of the shipping document retained by the carrier or intermediate handler and the copy accompanying the animal shipment.

Handling

1. When an animal is moved to or from a facility or conveyance, it must be sheltered from sunlight, extreme heat, rain, and snow.
2. The animal cannot be exposed to a temperature above 85°F for more than 45 min.
3. Transporting devices must be covered when the temperature falls below 50°F; the animal cannot be exposed to temperatures below 45°F for more than 45 min. Temperature must be measured from a point not more than 3 ft from the outside of the enclosure and halfway up the side of the enclosure.
4. Handlers must avoid causing physical harm or distress to the animals.
5. The enclosure cannot be tossed, dropped, needlessly tilted, or stacked in a manner that results in its falling.

Primary Conveyance

1. The cargo space of conveyances for transport of animals must be designed, constructed, and maintained in a manner that protects the health and well-being of each animal and ensures its safety and comfort.
2. The cargo space must have supply of air that is sufficient for normal breathing of all animals, and entry of engine exhaust must be prevented.
3. Enclosures must be positioned in the cargo space in a manner that provides protection from the elements, allows each animal enough air for normal breathing, and allows for the removal of animals in the event of an emergency.
4. During transport, auxiliary ventilation must be used in the cargo space when the temperature reaches 75°F. The temperature may not exceed 85°F or fall below 45°F except when rabbits are being transported and a certificate of acclimation to lower temperatures has been provided.
5. The cargo space must be kept clean, and animals may not be transported with any items that may be expected to cause harm to them.

Care in Transit

1. During surface transportation, animals must be observed at least once every 4 hr to ensure sufficient air and acceptable ambient temperatures. If an animal is in obvious physical distress, the operator must obtain appropriate veterinary care as soon as possible.
2. During air transportation, animals must be observed at least once every 4 hr unless the cargo area is not accessible during flight, in which case animals must be observed whenever they are loaded and unloaded to ensure sufficient air and acceptable ambient temperatures. If an animal is in obvious physical distress, veterinary care must be provided as soon as possible.
3. During transportation, animals shall not be removed from their enclosure unless it is to be placed in another enclosure or facility that conforms to the applicable AWA regulations.

Appendix B

Patterns in the Ground Transportation of Research Animals in the United States

BACKGROUND

This appendix focuses on inventorying the locations of institutions that use animals for research and the locations of the vendors that breed research animals for research in order to visualize the movement of research animals in the United States.¹ In addition, the appendix presents the results of a quantitative modeling effort directed towards locating additional supply points “optimally” and assessing the potential benefit of the additional supply points. All of the analyses in this appendix are focused on the transportation of research animals by truck from US breeding vendors to US research institutions; they do not consider transportation of research animals by other modes (air, sea, etc.) and transportation of research animals between research institutions or from abroad.

The next section discusses data-preparation efforts. The following two sections present a brief overview of the quantitative analysis and the empirical results of the analysis.

¹The transportation model was limited to those species for which the vendors/breeders supplying research animals were clearly identifiable. Livestock and some other species were excluded because animals of these species used in research can be obtained from a number of sources, some of which the committee could not identify.

DATA PREPARATION

Data Sources

The three primary data sets used in this analysis are the National Institutes of Health (NIH) academic institution data from 2003, the US Department of Agriculture (USDA) species and facilities data from 2003, and data on research animal vendors. The first set, which is referred to as the NIH grants data, consists of 1020 research institutions (academic and nonacademic) in the United States that receive grant awards for research utilizing research animals. For each institution, the data include the location (city and state), the number of NIH grant awards that utilize animals in 2003, and the award amount received in 2003. This data set was used as a proxy for rodent (rats and mice) use in the United States. Unlike other species of research animals, rodent use is not reported to any federal agency, so there is no census of rodents used in biomedical research. However, it has been estimated that rodents comprise approximately 95% of the animals utilized in research (Trull and Rich, 1999). Therefore, the total dollar amount of the NIH grants awarded to each institution was used as a proxy for the relative magnitude of rodent use at that institution.

The second data set, which is referred to as the USDA data,² contains information on 985 research institutions (academic and nonacademic), their locations, and the numbers of animals utilized in research or breeding programs, by species (cats, dogs, rabbits, guinea pigs, hamsters, pigs, sheep, other farm animals, and nonhuman primates). Those two sets of data include all warm-blooded vertebrates (with the exception of rats, mice, and birds) used in research in the United States, and those vertebrates (including rats, mice, and birds) utilized in research supported by NIH funding. The one subgroup of research animals not described in those data sets are rats, mice, and birds utilized at research institutions not supported by NIH funding. This would include most commercial research institutions, such as pharmaceutical and biotech companies. Rodent vendors have estimated that 45% of shipments are to for-profit customers, suggesting that a large portion of the facilities that utilize rodents are not represented in the data sets used for this model.

The third data set, which is referred to as the vendor data, includes the major vendors that supply research animals in the United States, the locations of production facilities, and the species they supply. Of the 45 vendors in this data set, four vendors supply cats, 16 supply dogs, 20 supply rodents (rats and mice); some vendors supply multiple species.

²This data was obtained from the USDA through a Freedom of Information Act request.

In addition to the three primary data sources listed above, the US Cities Geographic Information System (GIS) file provided by TransCAD® (Caliper, Inc.) was used in the analysis. The US Cities GIS file locates US cities on the basis of latitude and longitude.

Geocoding Procedure

Geocoding of research institution and vendor locations from the primary data sources was accomplished using TransCAD®. Specifically, data on the location of the institutions were matched with the city and state data fields in the US Cities GIS file to determine the geographic coordinates (latitude and longitude) of the institution or vendor locations. A TransCAD® macro was written to automate the process. Some 80% of the research institutions and all of the vendor locations were geocoded using the automated procedure. However, the US cities GIS file did not have city data fields corresponding to the remaining 20% of research institutions. For these, we used internet websites to manually determine geographic coordinates of the cities in which the research institutions were located.

Computation of Distances Between Utilization Points and Supply Locations

The geocoding of the locations of utilization points and supply locations was followed by an overlay procedure in TransCAD® in which the geocoded locations were overlaid on the TransCAD® highway file and mapped to the nearest highway network node. A TransCAD® shortest-path routine was then run to obtain distances between utilization points and supply locations.

QUANTITATIVE ANALYSIS

The objective of the quantitative analysis was to locate additional supply points optimally, with an assessment of the potential benefit of doing so. The fundamental motivation was to answer the following questions: If one additional breeding facility is being considered in the United States, where should it be to make the new total distance in the animal transportation system lowest? By how much is the new total transportation distance shorter than the former total distance? The questions could then be extended to more than one additional breeding facility. The facility-location model is thus a minimization model that considers various alternative location possibilities and determines the ones that provide the shortest transportation distance.

An important issue is that the solution space for identifying the best locations for new facilities is the entire United States. However, it is computationally infeasible to enumerate the objective function over all possible locations in the United States. Furthermore, for the scale of the transportation problem under study, there is no need to consider two closely spaced cities (such as Dallas and Fort Worth) as separate candidate locations. In the current analysis, the United States is divided into 40 grids, and the centroids of the grids are considered as the candidate solutions for the facility-location problem (see Figure B-1). The distances from all of the research centers to those potential facility locations are calculated with the procedure discussed above.

The minimization function in the facility-location problem is weighted to ensure that a new facility is located closer to high-utilization demand points. In the NIH grants file, the weighting was based on the total dollar amount of NIH grants. In the USDA file, the weighting for each species category was based on the total number of each species used at each utilization point.

A couple of points should be noted about the results of the quantitative modeling effort presented in the next section. First, because a spatial system that identifies 40 grids of the entire United States is used to identify candidate location sites, the city locations identified in the next section as optimal location points for additional facilities must not be taken literally. The spatial resolution adopted for the quantitative analysis implies that any location within 100-200 miles of the city locations identified in the next section would be appropriate and reasonable. Second, the locations of additional supply points are based solely on the criterion of reducing total transportation distance. Clearly, there are other factors that this model does not take into consideration, e.g., all stocks and strains of any given species are not maintained at each production facility and some vendors have established truck routes that determine which production facility will supply certain research institutions. Therefore, the locations suggested by the model in the next section should be considered in light of these complexities.

EMPIRICAL RESULTS

Three facility location models were estimated, one each for the NIH grants data set (rodents), the USDA cats data set, and the USDA dogs data set. For each of the three analyses, the optimal locations for one and two additional supply points were determined. In the analyses, the solutions for additional supply points were incremental and are not included here.

The results from the NIH grants data set (rodents) are provided in Figure B-2 and Table B-1. The current supply points are represented in the

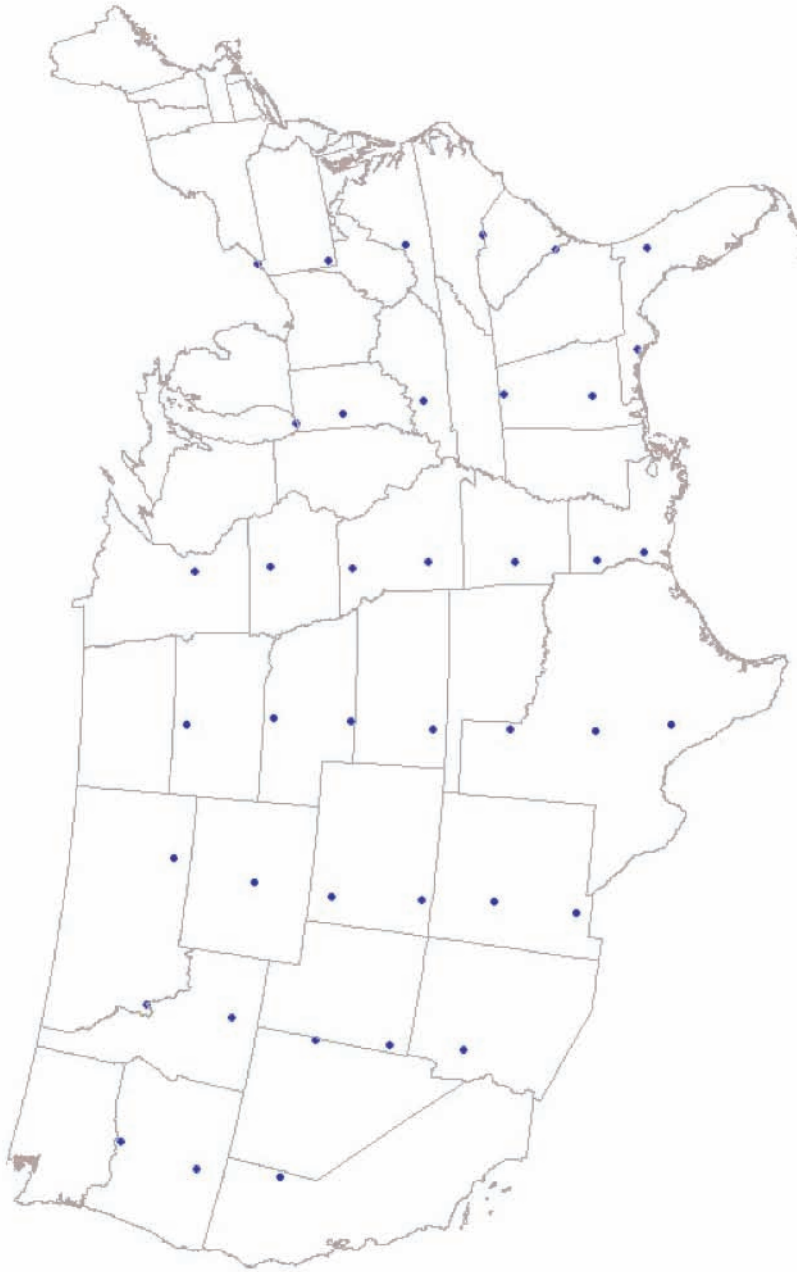


FIGURE B-1 Candidate set for the facility-location problem. Dots indicate the centroids for the 40 grid segments used in the analysis.

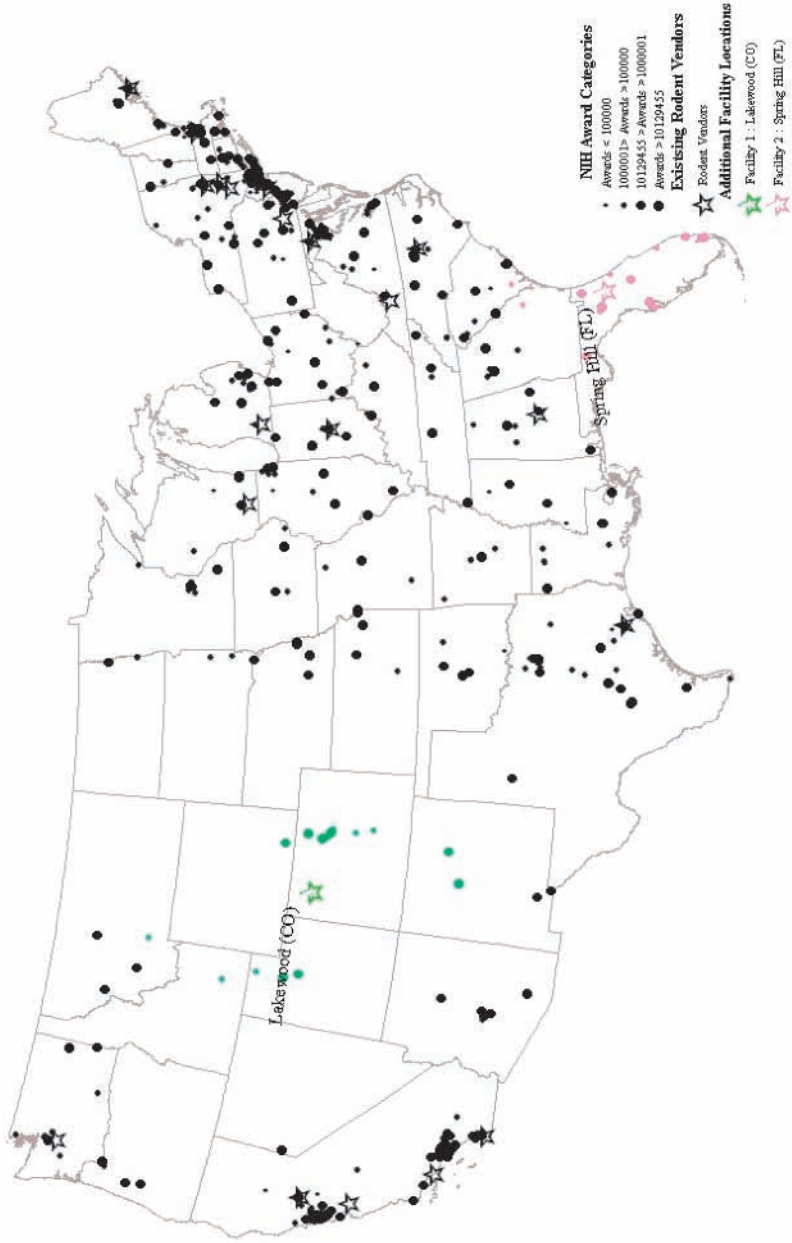


FIGURE B-2 Solution set for the facility location problem for the NIH grants data set (rodents).

shape of stars in Figure B-2, and the current demand points (i.e., research institutions) are represented by dots, with the diameter of the dot indicating the total amount of funding received by the research institution in 2003. The “new” supply points are identified by a colored star and labeled. The figure indicates that the optimal location for one additional supply point would be Lakewood, CO, and for two additional supply points would be Lakewood, CO, and Spring Hill, FL. Table B-1 indicates the estimated reduction in total weighted travel distance in the research animal transportation system due to the addition of supply points. The reductions are substantial and show, as expected, the decreasing marginal returns with the number of additional supply points.

The corresponding results for the USDA cats data set are provided in Figure B-3 and Table B-2, and for the USDA dogs data set in Figure B-4 and Table B-3. In the figures, the diameters of the dots representing research institutions indicate the number of animals utilized at the research institution. As can be observed from Tables B-1 through B-3, the reductions in total weighted travel distance are substantial for each species.

TABLE B-1 Total Weighted-System Travel-Distance Reduction with Increase in Supply Points for NIH Grants Data Set (Rodents)

No. of Facilities	Optimal Supply-Point Locations	Total Weighted-System Travel Distance (miles)	Incremental Percentage Change
0 (base case)	—	13,651,385	—
1	Lakewood, CO	12,198,700	10.64
2	Lakewood, CO and Spring Hill, FL	11,714,939	3.96

TABLE B-2 Total Weighted-System Travel-Distance Reduction with Increase in Supply Points for USDA Cats Data Set

No. of Facilities	Optimal Supply-Point Locations	Total Weighted-System Travel Distance (miles)	Incremental Percentage Change
0 (base case)	—	13,101,225	—
1	Chico, CA	9,108,974	30.47
2	Chico, CA and Spring Hill, FL	6,915,744	24.08

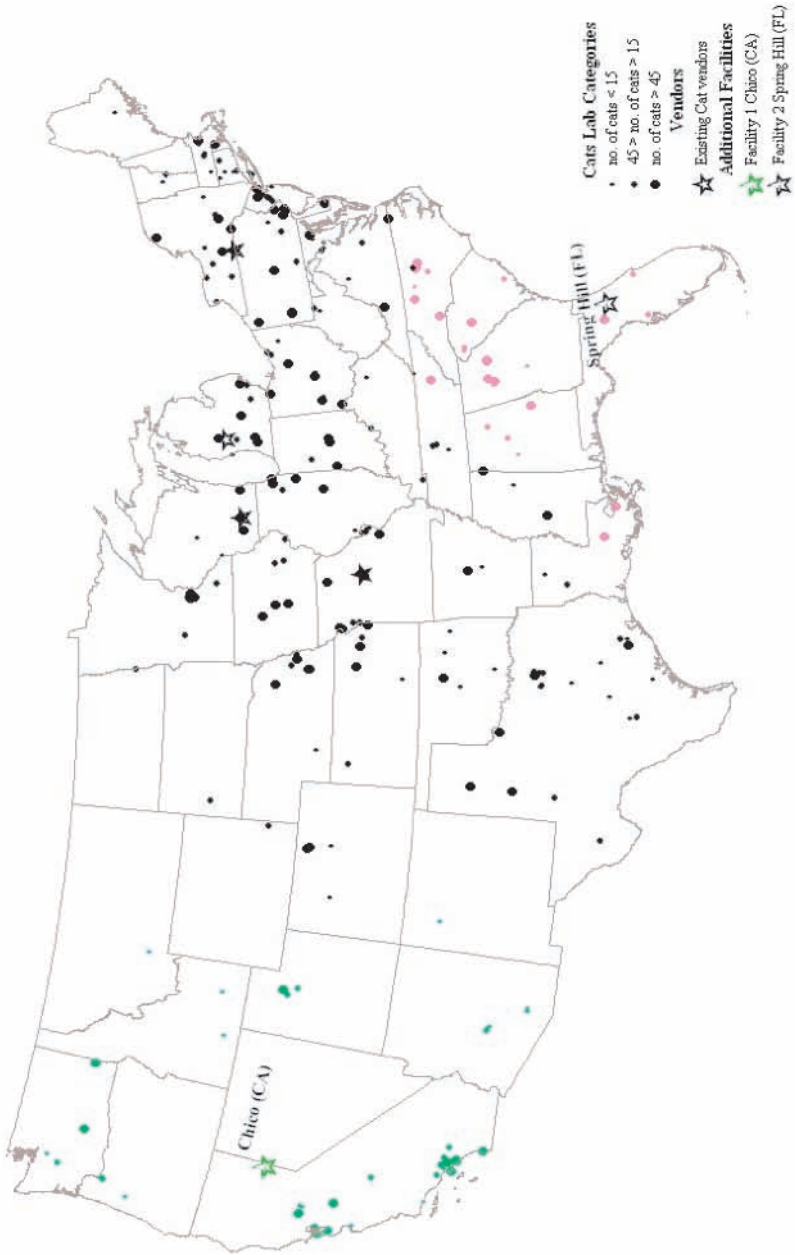


FIGURE B-3 Solution set for the facility location problem for the USDA cats data set.

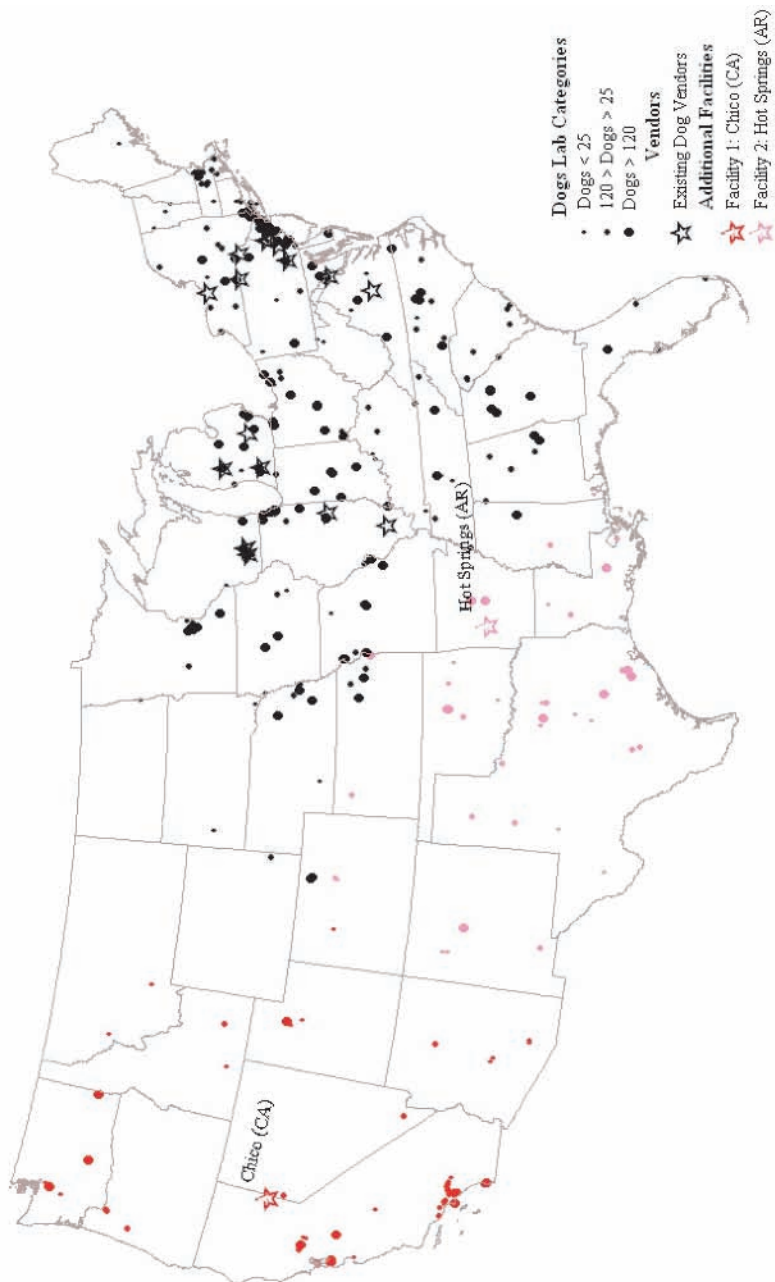


FIGURE B-4 Solution set for the facility location problem for the USDA dogs data set.

TABLE B-3 Total Weighted-System Travel-Distance Reduction with Increase in Supply Points for USDA Dogs Data Set

No. of Facilities	Optimal Supply-Point Locations	Total Weighted-System Travel Distance (miles)	Incremental Percentage Change
0 (base case)	—	25,387,410	—
1	Chico, CA	18,110,828	28.66
2	Chico, CA and Hot Springs, AR	14,765,449	18.47

SUMMARY

This chapter has described the committee's efforts to inventory and visualize the "utilization" points of research animals (the locations of institutions that use research animals) and the "supply" points of research animals (the locations of the vendors that breed research animals). Such an effort provides a qualitative sense of the patterns of transportation of research animals. In addition, the chapter presents the results of a quantitative modeling effort to locate additional supply points "optimally" and an assessment of the potential benefit of the additional supply points.

It is important to again emphasize that the locations for additional supply points developed from the quantitative analysis are based solely on the criterion of reducing total weighted travel distance in the research animal transportation system. Furthermore, because a grid system was used to identify candidate location sites and because of the scale of the research animal transportation system, any location within 100 to 200 miles of the locations identified in the analyses would be reasonable.

About the Authors

Ransom L. Baldwin, PhD (Chair), is professor and session chair of the Department of Animal Science of the University of California, Davis. He is known for his research on nutrient use and energetics in ruminant animals. Dr. Baldwin has extensive National Research Council committee experience, including involvement on the Committee to Revise the Guide for the Care and Use of Laboratory Animals (1996) and his role as chair of the Subcommittee on Input-Output Relationships in Animal Production. He is a member of the National Academy of Sciences.

Chandra R. Bhat, PhD, is an associate professor of civil engineering at the University of Texas at Austin. He is also the associate chairman of the Civil Engineering Department and the Fluor Centennial Teaching Fellow in Engineering. He has expertise in transportation-system analysis and travel-demand modeling. Dr. Bhat has done work for a number of metropolitan planning organizations on improvements in their travel-modeling procedures, including Boston, Dallas-Fort Worth, Houston-Galveston, and Seattle. He is chair of the National Research Council Committee on Passenger Travel Demand Forecasting and is a member of the National Research Council Committee for Review of Travel Demand Modeling by the Metropolitan Washington Council of Governments.

Donald H. Bouyer, PhD, is an assistant professor of pathology at the University of Texas Medical Branch at Galveston. His current research interests include host-parasite mechanisms of rickettsial diseases. He has

expertise in zoonotic diseases. Dr. Bouyer is a project leader at the Western Regional Center of Excellence for Biodefense and Emerging Infectious Diseases Research. The regional centers were created and funded by the National Institute for Allergy and Infectious Disease to develop and conduct programs to promote scientific discovery and translational research capacity to create the next generation of therapeutics for select agents and to provide facilities and support to first-line responders in the event of a national biodefense emergency.

Firdaus S. Dhabhar, PhD, is an associate professor of psychiatry and behavioral sciences at the Stanford University School of Medicine. He has expertise in stress and its effects on the endocrine and immune systems. Using rodent and human models, Dr. Dhabhar has elucidated critical psychophysiological, cellular, and molecular mechanisms by which stress may exert enhancing and suppressive effects on immune function in vivo. A major part of his effort is focused on examining the novel and unexpected immunoenhancing effects of mild stressors on innate, adaptive, and antitumor immune responses in the skin. For this work, Dr. Dhabhar received the PsychoNeuroImmunology Research Society Young Investigator Award in 2000, the Stazen Research Excellence Award in 2002, and the Fields Award for Excellence in Research and Teaching in 2003. Dr. Dhabhar served as a member of the Institute of Medicine committees on Gulf War and Health (Phase 1): Health Effects Associated with Exposure during the Persian Gulf War and Assessing Interactions among Social, Behavioral, and Genetic Factors in Health. He has also served as a grant reviewer for the National Institutes of Health and as a member of the Scientific Advisory Committee for the Chronic Fatigue and Immune Dysfunction Syndrome Association.

Steven L. Leary, DVM, is the assistant vice chancellor for veterinary affairs, director of the Division of Comparative Medicine, and research associate professor in the Department of Pathology at Washington University School of Medicine. He has a long history with laboratory animal transportation issues. Dr. Leary has previously served on an American Veterinary Medical Association committee to address concerns about the air transportation of companion animals. He has served as a member of the Association for Assessment and Accreditation of Laboratory Animal Care International. He is also a member of other research animal welfare organizations, including the American Association for Laboratory Animal Science and the American College of Laboratory Animal Medicine. Dr. Leary has served on the Editorial Board of *Comparative Medicine* and *Laboratory Animal Science* and on the Scientific Review Board of *Laboratory Animals*.

John J. McGlone, PhD, is a professor of animal science and cell biology and biochemistry in a joint appointment with Texas Tech University and Texas Tech University Health Sciences Center. He is also director of the Pork Industry Institute for Research and Education at Texas Tech. He has done extensive research on the behavior, welfare, and stress physiology of pigs and other research animals, including rodents and macaques. Dr. McGlone served in several capacities on the Council on Accreditation of the Association for Assessment and Accreditation of Laboratory Animal Care International. He is also a member of several societies, including the American Association for Laboratory Animal Science, the American Society of Animal Science, the Animal Behavior Society, and the International Society for Applied Ethology. He has served as the section editor for the Environment and Behavior Section of the *Journal of Animal Science*, the Federation of Animal Science Societies animal care committee, and the Food and Drug Administration Center for Veterinary Medicine advisory committee.

Eric Raemdonck is the manager of live animals and perishable goods in the International Air Transport Association (IATA). He is in charge of the IATA Live Animals Regulations manual, which contains guidelines for shipping live animals by air. With over 15 years of multimodal cargo transportation experience, he has vast knowledge of and experience with regulatory issues pertaining to the transportation of live animals (particularly primates).

Jennie L. Smith is the coordinator of the Yale Animal Resources Center, a centralized animal facility at Yale University. Ms. Smith oversees animal procurement and receiving for the university. She is also in charge of domestic and international importation and exportation of animals for the university. In addition to her knowledge and understanding of regulatory procedures and policies, Ms. Smith has experience in the ground transportation of rodents and other research animals.

Janice C. Swanson, PhD, is a professor of animal sciences and industry at Kansas State University. She is involved in research and activities addressing animal welfare concerns associated with farm animals exposed to intensive conditions, including confinement and transportation. Before her appointment at Kansas State University, Dr. Swanson worked for the Animal Welfare Information Center at the US Department of Agriculture. She is a member of numerous professional societies, including the Animal Behavior Society, the American Society of Animal Science, and the International Society for Applied Ethology.

Index

A

- AAALAC. *See* Association for Assessment and Accreditation of Laboratory Animal Care International
- AATA. *See* Animal Transportation Association
- Academic researchers. *See also* Responsible individuals
utilizing nonhuman primates, 82
- Acclimation. *See* Thermal environment in the transportation of research animals
- Adipsia, 54
- Agencies
checking with beforehand, 11
that fund research, 84
- Agents, that require registration of a facility with CDC, 67
- Agreement on the Application of Sanitary and Phytosanitary Measures (SPS Agreement), 28
- (Agro)bioterrorism agents, 65
- Air Cargo Tariff book, 23
- Airlines, 4
commercial, 3
- Allometric scaling, and implications for the transportation of research animals, 38–39
- Ambient temperature, ranges for safe transportation of common adult research animals, 48
- American Veterinary Medical Association (AVMA), 63
- Anesthesia, 61
- Animal and Plant Health Inspection Service (APHIS), 14, 22
- Animal rights activists, pressure from, 82
- Animal Transportation Association (ATA), *Manual for the Transportation of Live Animals*, 33
- Animal welfare, 4–5
investigators of, 4–5
- Animal Welfare Act (AWA), 12–14
dogs and cats, 97–104
guinea pigs and hamsters, rabbits, and other animals, 110–115
nonhuman primates, 104–110
regulations pertaining to transportation, 50, 54, 87, 97–115
- Animal Welfare Regulations, 88
- Animals
companion, 70, 78
cross-tying, 60
gnotobiotic, 76
immunocompromised, 76
lactating, 46
large, 38–39, 58

- late-pregnancy, 37, 46
- live, CDC rules on the importation of, 18–20
- natural enemies, 79
- preconditioning, 59
- segregation of, 71, 78–79
- small, 38–39, 42, 56
- Animals/animal products
 - with radioactive/poisonous material, federal statutes/programs relevant to the transportation of in the United States, 12–13
 - with zoonotic/infectious diseases, federal statutes/programs relevant to the transportation of in the United States, 12–13
- Anorexia, 54
- APHIS. *See* Animal and Plant Health Inspection Service
- Aquatic Animal Health Code*, 28
- Association for Assessment and Accreditation of Laboratory Animal Care International (AAALAC), 9
- Asymptomatic disease, 65, 69
- Autoclaving, 77
 - avoiding, 76
- Avian influenza A (H5N1), 19, 28–29
- Avian species, 84
- AVMA. *See* American Veterinary Medical Association
- AWA. *See* Animal Welfare Act

B

- B virus, 70
- Bacteria
 - requiring facility registration with CDC, 67
 - and the susceptible species of research animals, 74–75
 - zoonotic diseases transmissible from research animals to humans, 69
- Barrier containment, 71, 76–77
- Bats, live, 20
- Behavioral and physiological signs of thermal status, 52
- Behavioral monitoring of thermal environment, 51
- Biocontainment, 88
- Biomedical research
 - federal funding for, 7, 20
 - use of nonhuman primates in, 24

- Biomedical research enterprise, 1
- Biosafety in Microbial and Biomedical Laboratories Manual* (BMBL), 66, 71
- Biosecurity, 65–79
 - protecting public health and agricultural resources, 66–71
 - protecting the biological integrity of research animals and colonies, 71–79
- Bioterrorism agents, 65
- Bioterrorism Preparedness and Response Act, 66
- Bird flu, 19
- BMBL. *See* *Biosafety in Microbial and Biomedical Laboratories Manual*
- Breeders, commercial, 7–8

C

- California Department of Health Services, 22
- Candidate set for the facility-location problem, 121
- Care in transit
 - for dogs and cats, 103–104
 - for guinea pigs and hamsters, rabbits, and other animals, 115
 - for nonhuman primates, 109
- Carriers, third-party, 9
- Cats. *See* Dogs and cats
- CDC. *See* Centers for Disease Control and Prevention
- Center for Veterinary Medicine, 22
- Centers for Disease Control and Prevention (CDC), 7, 18–20, 66, 71, 85n, 86
 - agents and toxins that require registration of a facility with, 67
 - Division of Global Migration and Quarantine, 8–9, 12–13, 19
 - Etiologic Agent Import Permit Program, 12–13, 20
- Centers for Disease Control and Prevention (CDC) regulations and guidelines for the transportation of research animals, 18–20
 - importation and transportation of etiological agents, 20
 - importation of live animals, 18–20
- Certifications, 101, 107
- CFR. *See* *Code of Federal Regulations*
- Checklist, of research animal regulations and guidelines, 31–32
- Chicago, IL, 9

Chico, CA, 123–126
 CITES. *See* Convention on International Trade in Endangered Species of Wild Fauna and Flora
 Cleaning holding areas, 102, 108
Code of Federal Regulations (CFR), 66
 Colony security, 76
 Commercial airlines, 3
 Commercial breeders, 7–8
 Committee on Guidelines for the Humane Transportation of Laboratory Animals, 6, 51, 87
 Companion animals, 70, 78
 Compatibility
 for dogs and cats, 100–101
 for guinea pigs and hamsters, rabbits, and other animals, 112
 for nonhuman primates, 106
 Computation of distances, between utilization points and supply locations, 119
 Convention of the Council of Europe, 29
 Convention on International Civil Aviation, 25
 Convention on International Trade in Endangered Species of Wild Fauna and Flora (CITES), 12–13, 16, 23–25
 listed species of nonhuman primates, 26
 Resolution Conf., 25
 Conveyor belts, 103
 Cool-acclimation, 47
 Cool conditions, 58
 Core body temperature, 45
 Corporate research institutions, 82
 Council of Europe, 27
 Council Directives, 29–30
 Courier pick ups, 9
 Cross contamination, 70, 78
 Cross-infection, 79
 Cross-tying, 60

D

Dangerous goods, defining, 37
 Dangerous Goods Regulations (DGRs), IATA rules on, 27–28, 68
 Data preparation, 118–119
 computation of distances between utilization points and supply locations, 119
 data sources, 118–119
 geocoding procedure, 119

Data sources, 118–119
 Dehydration, 58
 Delays, 61
 Delivery to carriers and handlers
 for dogs and cats, 102
 for guinea pigs and hamsters, rabbits, and other animals, 113–114
 for nonhuman primates, 107
 Designated ports for importation or exportation of wildlife or derivatives, 16
 DGMQ. *See* Division on Global Migration and Quarantine
 DGRs. *See* Dangerous Goods Regulations
 Disease, asymptomatic, 65, 69
 Disinfection, 71, 77–78
 chemical, 71
 Division of Global Migration and Quarantine (DGMQ), 8–9, 9, 12–13, 19
 Documentation. *See also* Certifications
 for dogs and cats, 101–102
 for guinea pigs and hamsters, rabbits, and other animals, 113
 inconsistencies in, 88
 for nonhuman primates, 107
 of training of animal care personnel, 66
 Dogs and cats (9 CFR 3.13-3.19), 87–88, 97–104
 care in transit, 103–104
 compatibility, 100–101
 delivery to carriers and handlers, 102
 documentation, 101–102
 enclosure, 97–100
 federal statutes/programs relevant to the transportation of in the United States, 12–13
 food and water, 101
 handling, 102–103
 importing, 18
 primary conveyance, 103
 terminal facilities, 102
 DOT. *See* US Department of Transportation

E

EAIPP. *See* Etiologic Agent Import Permit Program
 Ebola virus, 70
 EDIM. *See* Group A rotavirus
 Emergency procedures
 elements of a plan for, 68

in the transportation of research animals, 61

Enclosures, 110–112
for dogs and cats, 97–100
for guinea pigs, 111
for guinea pigs and hamsters, rabbits, and other animals, 110–112
for hamsters, 111
for nonhuman primates, 104–106

Endangered Species Act (ESA) of 1973, 12–13, 15–17, 23
listed species of nonhuman primates, 17

Endangered/threatened wildlife, federal statutes/programs relevant to the transportation of in the United States, 13

Environmental conditions, 10

Environmental extremes, protecting against, 21

Environmental Protection Agency (EPA), 84

Environmentally controlled trucks, 8–9

EPA. *See* Environmental Protection Agency

ESA. *See* Endangered Species Act of 1973

Escapes, dealing with, 61, 82

Etiologic Agent Import Permit Program (EAIPP), 12–13, 20

Etiological agents, CDC rules on the importation and transportation of, 20

European Union (EU), Health and Consumer Protection Directorate General, 27, 29–30

Euthanasia, 61

Evaluation, 88

Exemptions, 22

Export brokers, 23

Extreme temperatures, exposure to, 61

F

Facility location problem
candidate set for, 121
models for, 120–122
for the NIH grants data set (rodents), 122
for the USDA cats data set, 124
for the USDA dogs data set, 125

Fasting, 58

FDA. *See* Food and Drug Administration

Federal Quarantine Regulations, regarding nonhuman primates, 19

Federal statutes/programs relevant to the transportation of vertebrate research animals and products in the United States, 12–13

Food and Drug Administration (FDA), 12–13, 21–22, 86
Center for Veterinary Medicine, 22

Food and water
for dogs and cats, 101
for guinea pigs and hamsters, rabbits, and other animals, 112–113
for nonhuman primates, 106
providing when indicated, 21
in the transportation of research animals, 54–58

Freedom of Information Act, 118n

Funding, for biomedical research, federal, 7

Fungi, requiring facility registration with CDC, 67

FWS. *See* US Fish and Wildlife Service

G

Gamma irradiation, 77

GD-VII. *See* Theiler's murine encephalomyelitis virus strain

Gel moisture sources, 57

Geocoding procedure, 119

Geographic information system (GIS), 119

Gnotobiotic animals, 71

Good practices in the transportation of research animals, 33–63
allometric scaling and implication for transportation practices, 38–39
emergency procedures, 61
food and water, 54–58
handling, 59–60
monitoring transportation, 60
personnel training, 10, 61–63
social interaction and group transportation, 59
space allocation, 52–54
stress during transportation, 34–38
thermal environment, 39–52

Good shippers, characteristics of, 9

Ground transportation, 3–4, 85

Group A rotavirus (EDIM), 72–73

Guide for the Care and Use of Agricultural Animals in Agricultural Research and Teaching, 33

Guide for the Care and Use of Laboratory Animals, 21

Guidelines. *See* Regulations and guidelines for the transportation of research animals

Guinea pigs and hamsters, rabbits, and other animals (9 CFR 3.35-3.41, 9 CFR 3.60-3.65, 9 CFR 3.136-3.142), 87, 110-115

care in transit, 115

compatibility, 112

delivery to carriers and handlers, 113-114

documentation, 113

enclosure, 110-112

food and water, 112-113

handling, 114

primary conveyance, 115

terminal facilities, 114

H

Hamsters, 70, 111. *See also* Guinea pigs and hamsters, rabbits, and other animals

Handler training. *See* Personnel training

Handling

- dogs and cats, 102-103
- guinea pigs and hamsters, rabbits, and other animals, 114
- nonhuman primates, 108
- preconditioning animals to, 59
- in the transportation of research animals, 59-60

Hantaan (HANT) virus, 72-73

Hazardous Materials Information Center, 21

Hazardous Materials Regulation (HMR), 20-21, 25

Health Research Extension Act of 1985, 21

Helicobacter species, 71

High-efficiency particulate air (HEPA) filters, 86

HMR. *See* Hazardous Materials Regulation

Holding areas, cleaning and sanitizing, 102, 108

Homeotherms, 45-46, 51

- graph representing relationship between metabolic rate and ambient temperature in, 42

Host species, 65

Hot conditions, 58. *See also* Thermal environment in the transportation of research animals

Hot Springs, AR, 125-126

HPA. *See* Hypothalamic pituitary adrenal axis response

Husbandry, species-specific, 62

Hydration, 57

Hypothalamic pituitary adrenal axis (HPA) response, 37-38

I

IATA. *See* International Air Transport Association

ICAO. *See* International Civil Aviation Organization

ILAR. *See* Institute for Laboratory Animal Research

Immunocompromised animals, 76

Importation sites, major, 9, 83

Incompatibilities, among export and import requirements, 23

Inconsistencies, in documentation and reporting requirements, 87-88

Individuals at institutions, taking responsibility, 5, 88

Infectious agents and the susceptible species of research animals, 72-75, 88

- bacteria, 74-75
- parasites, 74-75
- viruses, 72-73

Infectious pathogens, risk of introducing, 65

Infectious substances, 20

Infectious Substances Program, 12-13

Institute for Laboratory Animal Research (ILAR), 7

Institutions, individuals at taking responsibility, 5, 88

Insurance rates, 82

Interagency Primate Steering Committee, 84

Interagency working group, needed to coordinate all federal inspection and permitting of animals, 51, 86-88

International Air Transport Association (IATA), 14

International Air Transport Association (IATA) regulations and guidelines for the transportation of research animals, 23-24, 27-28, 30, 50, 71

Dangerous Goods Regulations, 27-28, 68

Live Animals Regulations, 14, 24, 27, 61, 78-79

International Civil Aviation Organization (ICAO), 25–27
 International regulations for transporting research animals, 22–23
 Irradiation, 77
 Isolation, 59

K

Kilham Rat virus (KRV), 72–73

L

Laboratory Animal Science Association, Report of the Transport Working Group, 33
 Lacey Act, 12–14, 18
 Lactating animals, 46
 Lakewood, CO, 123
 LARs. *See Live Animals Regulations*
 LCM. *See Lymphocytic choriomeningitis*
 LCT. *See Lower critical temperature*
 Listed Animal Permit Officials, 22
Live Animals Regulations (LARs), IATA rules on, 14, 24, 27, 61, 78–79
 Livestock, stress experienced during transportation, 37–38
 Livestock industry, 33
 Livestock Weather Safety Index, 46
 Long-term housing, 52
 Los Angeles, CA, 9
 Lower critical temperature (LCT), 44
 Lymphocytic choriomeningitis (LCM), 72–73

M

MAD. *See Mouse adenovirus*
 Malign hyperthermia, 48, 50
Manual for the Transportation of Live Animals, 33
 Maps
 candidate set for the facility-location problem, 121
 locations of research facilities using nonhuman primates, major importation sites, and vendors of nonhuman primates in the United States, 83

 solution set for the facility location problem for the NIH grants data set (rodents), 122
 solution set for the facility location problem for the USDA cats data set, 124
 solution set for the facility location problem for the USDA dogs data set, 125
 MCMV. *See Murine cytomegalovirus*
 MHV. *See Mouse hepatitis virus*
 Mice, wild, 78
 Microisolation shipping containers, 76
 Minimizing risks associated with transporting research animals with experimentally introduced zoonoses, 66–68
 Minimizing risks associated with transporting research animals with unknown zoonoses, 69–71
 special considerations when transporting nonhuman primates, 70–71
 special considerations when transporting specimens and tissues, 71
 Minute virus of mice (MVM), 72–73
 Monitoring
 behavioral, of thermal environment, 51
 of the transportation of research animals, 60
 Mouse adenovirus (MAD), 72–73
 Mouse hepatitis virus (MHV), 72–73
 Mouse parvovirus (Orphan parvovirus) (MPV [OPV]), 71
 Mouse thymic virus (MTLV), 72–73
 MPV (OPV). *See Mouse parvovirus* (Orphan parvovirus)
 MTLV. *See Mouse thymic virus*
 Murine cytomegalovirus (MCMV), 72–73
 MVM. *See Minute virus of mice*

N

National Center for Infectious Diseases, 1, 7
 National Center for Research Resources (NCRR), 3, 81
 National Institutes of Health (NIH), 1, 3, 7, 21, 66, 81
 data from, 118
 research projects funded by, 85

National Primate Plan, 3, 81, 84
 National Primate Research Center (NPRC),
 3, 81, 84–85
 National Science Foundation, 84
 NCRR. *See* National Center for Research
 Resources
 Newborns, 46
 NIH. *See* National Institutes of Health
 NIH grants data set (rodents), 120
 solution sets for the facility location
 problem for, 122
 total weighted-system travel-distance
 reduction with increase in supply
 points for, 123
 Nonhuman primates (9 CFR 3.86-3.92), 15,
 87–88, 104–110
 academic researchers utilizing, 82
 care in transit, 109
 compatibility, 106
 delivery to carriers and handlers,
 107
 documentation of, 107
 enclosures for, 104–106
 encouraging more airlines to transport,
 4
 Federal Quarantine Regulations
 regarding, 19
 federal statutes/programs relevant to
 the transportation of in the United
 States, 12
 food and water, 106
 handling, 108
 isolating, 79
 locations of research facilities using,
 83
 options available for transporting
 between research facilities, 9
 ports involved, 9
 primary conveyance, 109
 registering importers of, 20
 special considerations when
 transporting, 70–71
 stress experienced during
 transportation, 37
 terminal facilities, 107–108
 use in biomedical research, 24
 NPRC. *See* National Primate Research
 Center
Nutrient Requirements of Domestic Animals,
 58

O

Office International des Épipzooties (OIE),
 28–29
 International Committee, 29
 Office of Hazardous Materials, 20, 27
 Office of Laboratory Animal Welfare
 (OLAW), 21
 OIE. *See* Office International des Épipzooties
 OLAW. *See* Office of Laboratory Animal
 Welfare
 Orientation, preferred, 54
 Overcrowding, avoiding, 21
 Overshippers. *See* Self-contained
 overshippers

P

Parasites, and the susceptible species of
 research animals, 74–75
 Patterns in the ground transportation of
 research animals in the United
 States, 117–126
 background, 117
 data preparation, 118–119
 empirical results, 120–126
 quantitative analysis, 119–120
 Performance standards, science-based, 34
 Permitting Requirements under CITES, 25
 Personal protective equipment (PPEs), 70–
 71, 76, 78
 Personnel training, 10
 documenting, 66, 76
 high cost of, 3, 82
 standard program for, 63
 in the transportation of research
 animals, 61–63
 Pharmaceutical Research and
 Manufacturers of America
 (PhRMA), 7
 PHMSA. *See* Pipeline and Hazardous
 Materials Safety Administration
 PhRMA. *See* Pharmaceutical Research and
 Manufacturers of America
 PHS. *See* Public Health Service
 PHS Policy. *See* Policy on the Humane Care
 and Use of Laboratory Animals
 Physical distress
 observing, 15, 103–104, 109
 protecting against, 21

Pipeline and Hazardous Materials Safety
 Administration (PHMSA), 20, 27
 Associate Administrator for Hazardous
 Materials Safety, 20
 Hazardous Materials Information
 Center, 21
 Office of Hazardous Materials, 20, 27
 Pneumonia virus of mice (PVM), 72–73
 Poikilotherms, 50
 Policy on the Humane Care and Use of
 Laboratory Animals, 21
 Post-transportation recovery times, 36
 PPEs. *See* Personal protective equipment
 Preconditioning animals, 59
 Predators of wildlife, Federal statutes/
 programs relevant to the
 transportation of in the United
 States, 13
 Pregnant animals, 37, 46
 Primary conveyance
 for dogs and cats, 103
 for guinea pigs and hamsters, rabbits,
 and other animals, 115
 for nonhuman primates, 109
 Principles of thermoregulation, 39–45
 Prions, requiring facility registration with
 CDC, 67
 Private charters, covering the cost of, 82
 Professional judgment, 47
 Professional societies, 4
 Program evaluation, 88
 Protecting public health and agricultural
 resources, 66–71
 minimizing risks associated with
 transporting research animals with
 experimentally introduced zoonoses,
 66–68
 minimizing risks associated with
 transporting research animals with
 unknown zoonoses, 69–71
 Protecting the biological integrity of
 research animals and colonies, 71–79
 barrier containment, 76–77
 disinfection, 77–78
 personal protective equipment, 78
 segregation of animals, 78–79
 specific pathogen diagnosis, 77
 Protective equipment. *See* Personal
 protective equipment
 Public Health Service (PHS), Policy on the
 Humane Care and Use of
 Laboratory Animals, 21
 PVM. *See* Pneumonia virus of mice

Q

Quantitative analysis, 119–120
 Quarantine procedures, 22, 77, 79, 88

R

Rabbit haemorrhagic disease (RHD), 72–73
 Rabbits, 70. *See also* Guinea pigs and
 hamsters, rabbits, and other animals
 Rabies, 18
 Radioactive/Poisonous Materials Program,
 12–13
 Rat parvovirus (Orphan parvovirus) (RPV
 [OPV]), 71
 Rats, federal statutes/programs relevant to
 the transportation of in the United
 States, 13
 Recommendations, 3–5, 81–88
 animal welfare, 4–5
 declining availability of air
 transportation for nonhuman
 primates, 3–4
 interagency working group needed to
 coordinate all federal inspection and
 permitting of animals, 51, 86–88
 National Primate Plan, updating and
 reimplementing, 3, 81–85
 regulatory burden, 4
 reliable ground transportation for
 nonhuman primates, collaboration
 needed to establish, 4, 85
 self-contained overshippers to ship
 zoonotic animals, need to develop, 4,
 86
 shipments of research animals between
 institutions, ensuring coordination
 between responsible individuals at
 both institutions, 5, 76, 88
 Registered carriers, 9
 Regulations and guidelines for the
 transportation of research animals,
 11–32
 complex and confusing, 4, 86–87
 compliance within, 11
 international, 22–23
 national, 11–14
 Reovirus type 3 (REO 3), 72–73
 Report of the Transport Working Group
 Established by the Laboratory
 Animal Science Association, 33

Reporting requirements, inconsistencies in, 87–88

Researchers. *See also* Responsible individuals
utilizing nonhuman primates, 82

Responsible individuals, at institutions, 5, 88

RHD. *See* Rabbit haemorrhagic disease

Rodents, 3, 70, 118
African, ban on importation of, 19
stress experienced during transportation, 36–37

Ross River virus (RRV), 72–73

RPV (OPV). *See* Rat parvovirus (Orphan parvovirus)

RRV. *See* Ross River virus

S

Safe temperature ranges during transport, 46–50

San Francisco, CA, 9

Sanitizing holding areas, 102, 108

Science-based performance standards, 34

SDA/RCV. *See* Sialodacryoadentitis virus/Rat corona virus

Self-contained overshippers, to ship zoonotic animals, need to develop, 4, 70–71, 79, 86

Shipping companies, 10. *See also* Carriers

Shipping company, characteristics of a good, 68

Shipping containers, 10. *See also* Self-contained overshippers

Shipping routes and destinations, 9, 120

Short-term exposures, 47

Sialodacryoadentitis virus/Rat corona virus (SDA/RCV), 72–73

Simian immunodeficiency virus (SIV), 72–73

Social animals and species, 59

Social interaction and group transportation, in the transportation of research animals, 59

Solitary animals and species, 59

Solution sets for the facility location problem
for the NIH grants data set (rodents), 122
for the USDA cats data set, 124
for the USDA dogs data set, 125

Space allocation in the transportation of research animals, 52–54, 57
for group-transported animals, 55–56

Spatial isolation, 59

Species-specific husbandry, 62

Specific-diagnostic diagnosis, 71

Specific pathogen diagnosis, 77

Specimens, special considerations when transporting, 71

Spring Hill, FL, 123–124

SPS Agreement. *See* Agreement on the Application of Sanitary and Phytosanitary Measures

Staff turnovers, 63

State health and agricultural regulations, 22

Sterilization, 76–77

Stocking density, 53

Stress during transportation, 34–38
acute, 34, 36
chronic, 34
livestock, 37–38
nonhuman primates, 37
rodents, 36–37

Supply locations, 119

Sweating species, vs. nonsweating, 45

T

Technical Instructions for the Safe Transport of Dangerous Goods by Air, 25, 27

Temperatures. *See also* Ambient temperature
core body, 45
exposure to extreme, 61
safe ranges during transport, 46–50

Terminal facilities
for dogs and cats, 102
for guinea pigs and hamsters, rabbits, and other animals, 114
for nonhuman primates, 107–108
temperatures in, 87

Terrestrial Animal Health Code, 28

Theiler's murine encephalomyelitis virus strain (GD-VII), 72–73

Thermal environment in the transportation of research animals, 39–52
behavioral monitoring of, 51
effect of transportation caging on, 50–51
exposure to extreme temperatures, 61
principles of thermoregulation, 39–45

- safe temperature ranges during transport, 46–50
- thermal acclimation, 47, 51–52, 102–103
- Thermogenesis, 48
 - nonshivering vs. shivering, 43
- Thermoneutral zone (TNZ), 39, 42–48
 - changes in (range of ambient is at a minimum) with age and size in chickens, 43
 - of various agricultural animals, 44
- Thermoregulation data, on common research animal species, 40–41
- Third party carriers, 9
- Tissues, special considerations when transporting, 71
- TNZ. *See* Thermoneutral zone
- Total weighted-system travel-distance reduction with increase in supply points
 - for NIH grants data set (rodents), 123
 - for USDA cats data set, 123
 - for USDA dogs data set, 126
- Toxins that require registration of a facility with CDC, 67
- Trade, defining, 23
- Training. *See* Personnel training
- TransCAD®, 119
- Transport of Dangerous Goods, United Nations recommendations for, 25
- Transportation of animals
 - phases of, 34
 - between research facilities, checklist of issues to consider when arranging, 10
 - stressors in research animals, 34–38
- Treasury and General Government Appropriations Act of 2002, 84
- Truck routes, 120
- Turtles, importing, 18–19
- US Department of Agriculture (USDA), 14, 71, 85n, 86, 88, 97, 113
 - Animal and Plant Health Inspection Service, 14, 22
 - Animal Welfare Act, 12–13
 - data from, 118
- US Department of Defense, 84
- US Department of Health, Education, and Welfare, 84
- US Department of Justice, 66
- US Department of Transportation (DOT), 20–21, 25, 68, 86
 - Infectious Substances Program, 12–13
 - Pipeline and Hazardous Materials Safety Administration, 20, 27
 - Radioactive/Poisonous Materials Program, 12–13
- US Fish and Wildlife Service (FWS), 14–18, 27, 85n, 86
 - CITES, 12–13, 16, 23
 - Endangered Species Act, 12–13, 15–17, 23
 - Lacey Act, 12–14, 18
 - Permit Website, 16
- USA Patriot Act, 66
- USDA. *See* US Department of Agriculture
- USDA cats data set
 - solution sets for the facility location problem for, 124
 - total weighted-system travel-distance reduction with increase in supply points for, 123
- USDA dogs data set
 - solution sets for the facility location problem for, 125
 - total weighted-system travel-distance reduction with increase in supply points for, 126
- Utilization points, 119

U

- UCT. *See* Upper critical temperature
- United Nations, recommendations for the Transport of Dangerous Goods, 25
- United States, vendors of nonhuman primates in, 83
- Upper critical temperature (UCT), 42, 44, 46
- US Cities Geographic Information System, 119

V

- Vendors of nonhuman primates in the United States, 83
 - data on, 118
- Ventilation, 102, 108
- Vertebrates, warm-blooded, federal statutes/programs relevant to the transportation of in the United States, 13

Veterans Administration, 84
Veterinary care, 61, 87, 101, 103–104, 109, 115
Viruses
 B, 70
 Ebola, 70
 monkeypox, 22
 requiring facility registration with CDC, 67
 and the susceptible species of research animals, 72–73
 zoonotic diseases transmissible from research animals to humans, 69
Visual isolation, 59

W

Warm-blooded vertebrates, federal statutes/programs relevant to the transportation of in the United States, 13
Washington Convention. *See* Convention on International Trade in Endangered Species of Wild Fauna and Flora

Water bottles, 57
Wild mice, infected, 78
Wildlife
 defining, 15
 federal statutes/programs relevant to the transportation of in the United States, 13
Working groups. *See* Interagency working group; Report of the Transport Working Group Established by the Laboratory Animal Science Association
World Animal Health Organization, 28–29

Z

Zoonotic diseases, 3, 9, 28
 controlling, 86
 risk of, 21, 82
 suspected, 88
 transmissible from research animals to humans, 69

