# Zoophysiology Volume 37

Editors: S.D. Bradshaw W. Burggren H.C. Heller S. Ishii H. Langer G. Neuweiler D.J. Randall

Springer-Verlag Berlin Heidelberg GmbH

# Zoophysiology

Volumes already published in the series:

Volume 1: *P.J. Bentley* Endocrines and Osmoregulation (1971)

Volume 2: *L. Irving* Arctic Life of Birds and Mammals Including Man (1972)

Volume 3: A.E. Needham The Significance of Zoochromes (1974)

Volume 4/5: *A.C. Neville* Biology of the Arthropod Cuticle (1975)

Volume 6: K. Schmidt-Koenig Migration and Homing in Animals (1975)

Volume 7: *E. Curio* The Ethology of Predation (1976)

Volume 8: *W. Leuthold* African Ungulates (1977)

Volume 9: *E.B. Edney* Water Balance in Land Arthropods (1977)

Volume 10: *H.-U. Thiele* Carabid Beetles in Their Environments (1977)

Volume 11: M.H.A. Keenleyside Diversity and Adaptation in Fish Behaviour (1979)

Volume 12: *E. Skadhauge* Osmoregulation in Birds (1981)

Volume 13: S. Nilsson Autonomic Nerve Function in the Vertebrates (1983)

Volume 14: *A.D. Halsler* Olfactory Imprinting and Homing in Salmon (1983)

Volume 15: *T. Mann* Spermatophores (1984)

Volume 16: *P. Bouverot* Adaption of Altitude-Hypoxia in Vertebrates (1985)

Volume 17: *R.J.F. Smith* The Control of Fish Migration (1985)

Volume 18: *E. Gwinner* Circannual Rhythms (1986)

Volume 19: J.C. Rüegg Calcium and Muscle Activation (1986)

Volume 20: J.-R. Truchot Comparative Aspects of Extra-cellular Acid-Base Balance (1987) Volume 21: A. Epple and J.E. Brinn The Comparative Physiology of the Pancreatic Islets (1987)

Volume 22: *W.H. Dantzler* Comparative Physiology of the Vertebrate Kidney (1988)

Volume 23: *G.L. Kooyman* Diverse Divers (1989)

Volume 24: S.S. Guraya Ovariant Follicles in Reptiles and Birds (1989)

Volume 25: G.D. Pollak and J.H. Casseday The Neural Basis of Echolocation in Bats (1989)

Volume 26: *G.A. Manley* Peripheral Hearing Mechanisms in Reptiles and Brids (1989)

Volume 27: *U.M. Norberg* Vertebrate Flight (1990)

Volume 28: *M. Nikinmaa* Vertebrate Red Blood Cells (1990)

Volume 29: *B. Kramer* Electrocommunication in Teleost Fishes (1990)

Volume 30: W. Peters Peritrophic Membranes (1991)

Volume 31: *M.S. Kaulenas* Insect Accessory Reproductive Structures (1992)

Volume 32: A.L. Val and V.M.F. de Almeida-Val Fishes of the Amazon and their Environment (1995)

Volume 33: R. Wiltschko and W. Wiltschko Magnetic Orientation in Animals (1995)

Volume 34: S.B. Moffett Nervous System Regeneration in the Invertebrates (1996)

Volume 35: *H. Kobayashi* and *Y. Takei* The Renin-Angiotensin System (1996)

Volume 36: *L. Aitkin* Hearing – the Brain and Auditory Communication in Marsupials (1998)

Volume 37: J.N. Maina The Gas Exchangers: Structure, Function, and Evolution of the Respiratory Processes (1998)

# The Gas Exchangers

Structure, Function, and Evolution of the Respiratory Processes

With 108 Figures and 33 Tables



Prof. John N. Maina Faculty of Veterinary Medicine Dept. of Veterinary Anatomy University of Nairobi P.O. Box 30197 Nairobi, Kenya

Faculty of Health Sciences Dept. of Anatomical Sciences The University of Witwatersrand 7 York Road, Parktown Johannesburg 2193 South Africa

#### Cover illustration: A plastron of a crane fly larva Dicranomyia (Fig. 107)

ISBN 978-3-642-63756-8 ISSN 0720-1842

Library of Congress Cataloging-in-Publication Data

Maina, J.N.

The gas exchangers: structure, function, and evolution of the respiratory processes / J.N. Maina.
p. cm. - (Zoophysiology; v. 37)
Includes bibliographical references and index.
ISBN 978-3-642-63756-8 ISBN 978-3-642-58843-3 (eBook)
DOI 10.1007/978-3-642-58843-3
I. Respiration. 2. Physiology, Comparative. I. Title. II. Series.
QP121.M276 1998
573.2 — dc21

This work is subject to copyright. All rights are reserved, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilm or in any other way, and storage in data banks. Duplication of this publication or parts thereof is permitted only under the provisions of the German Copyright Law of September 9, 1965, in its current version, and permission for use must always be obtained from Springer-Verlag. Violations are liable for prosecution under the German Copyright Law.

© Springer-Verlag Berlin Heidelberg 1998

Originally published by Springer-Verlag Berlin Heidelberg New York in 1998

Softcover reprint of the hardcover 1st edition 1998

The use of general descriptive names, registered names, trademarks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

Cover design: Design & Production GmbH, Heidelberg Typesetting: Best-set Typesetter Ltd., Hong Kong

SPIN: 10521202 31/3137 - 5 4 3 2 1 0 - Printed on acid-free paper

"Biologists are seeking to integrate studies of morphology, development, physiology, ecology, systematics, and behaviour in order to understand how species and lineages deal with their environments and how they have diversified." Wake (1990)

# Dedication

This book would not have been written without the patience, understanding, and support of my wife, Wanjuku, and our children, Ndegwa, Wanjiru, and Kireru. I am particularly indebted to them for their forbearance when the work was a major preoccupation. They kept urging me on especially when the "spirit was willing but the body weak" by firmly and persistently inquiring when "the book" would be ready. I dedicate the work to them.

It has been both a challenge and pleasure writing this book. I hope that the reader will partake in some of it.

## Acknowledgement

I am immensely grateful to the numerous colleagues who have collaborated with me over the years and given me their ideas and time unreservedly. All cannot be mentioned here. The particularly noteworthy ones include Prof. A.S. King (Emeritus Prof.), University of Liverpool; Dr. M.A. Abdalla, King Saud University; Prof. G.M.O. Maloiy, formerly of the University of Nairobi, Prof. S.P. Thomas, Duquesne University, and Prof. C.M. Wood, McMaster University. I am indebted to Prof. D.J. Randall, Department of Zoology, University of British Columbia, the editor responsible for this Volume, for reading the manuscript and making most helpful suggestions. My most sincere thanks go to Dr. Dallas M. Hyde, Chair, Department of VM: Anatomy, Physiology and Cell Biology, University of California (Davis) for having me in his laboratory and putting all necessary facilities at my disposal during my 1996 year Fulbright sabbatical leave, when substantial organization of this work was carried out. I wish to record my gratitude to Messrs. M. Mwasela-Tangai and J. Gachoka for assistance with the preparation of the illustrations and Ms L. Mburu and J. Muhia for initial word processing of some parts of the manuscript. My past and present graduate students have been integral in shaping the thoughts expressed in this exposition. To all of them, I express my appreciation. Special thanks go to Profs. R.D. Farley (University of California - Berkeley) and D.O. Okello (University of Nairobi) for providing me with original copies of Figures 15 and 54, respectively, for reproduction and the following publishers for their generous permission to reproduce materials from their publications:

Academic Press Ltd. Annual Review Inc. Birkhahauser-Verlag Cambridge University Press Elsevier/North Holland Publishing Company Harvard University Press Kluwer Publications Ltd. Longman Group (UK) Ltd. McGraw-Hill Inc. McMillan Magazines Ltd. Oxford University Press Pergamon Press Springer-Verlag The Company of Biologists Ltd. Wiley/Liss Inc. I am indebted to the editorial staff at the Springer-Verlag office (Heidelberg) for their numerous courtesies.

Johannesburg, March 1998

J.N. Maina

## Preface

"Amongst animals, diversity of form and of environmental circumstances have given rise to a multitude of different adaptations subserving the relatively unified patterns of cellular metabolism. Nowhere else is this state of affairs better exemplified than in the realm of respiration". Jones (1972).

The field of comparative respiratory biology is expanding almost exponentially. With the ever-improving analytical tools and methods of experimentation, its scope is blossoming to fascinating horizons. The innovativeness and productivity in the area continue to confound students as well as specialists. The increasing wealth of data makes it possible to broaden the information base and meaningfully synthesize, rationalize, reconcile, redefine, consolidate, and offer empirical validation of some of the earlier anecdotal views and interpretations, helping resolve the issues into adequately realistic and easily perceptible models. Occasional reflections on the advances made, as well as on the yet unresolved problems, helps chart out new grounds, formulate new concepts, and stimulate inquiry. Moreover, timely assessments help minimize isolation among investigators, averting costly duplication of effort. This exposition focuses on the diversity of the design of the gas exchangers and gives a critical appraisal of the plausible factors that have motivated or constrained the evolvement of respiration. The cause-and-effect relationship between the phylogenetic, developmental, and environmental factors, conditions, and states which at various thresholds and under certain backgrounds conspired in molding the gas exchangers is argued. Convergence as well as divergence, retrogression and progression, parallel as well as serial developments have occurred in this stochastic process manifest with recurrent catastrophic crises. Gould (1994) asserts that "life's pathways are more contingent and chancy than predictable and directional". Such a caveat notwithstanding, with judicious use of data, an integrative multidisplinary approach into the evolvement of the gas exchangers should help develop a conceptual framework of the appropriation and synthesis of the myriad parameters and components involved in the assembly of the gas exchangers. The repertoire of the functional mechanisms conceived, the investments made to support such developments, the losses incurred, and the gains reaped in the course of the development of respiration can be grasped. The adaptive attributes common to or unique to broadly defined groups of organisms can be adduced. On that basis, the connectivity, the diversity, and the chronology of the changes that occurred during and after the incorporation of molecular O<sub>2</sub> into the respiratory process can be gauged. To place the historical developments of respiration in their proper perspective, an ecopaleobiological approach has been adopted. Borrowing heavily from works of others, pertinent past and present states, events, and circumstances have been associated. I have been consciously eclectic in order to remain both brief and focused on the subject matter. The embracing approach adopted provides instructive parametric insights into the permutations and spatial and temporal vectorial shifts of the causative factors during the genesis of the respiratory processes. This should explain how and why particular respiratory traits were acquired, some lost, and the means by which solutions to attendant challenges were found as the respiratory machines were forged. In evolutionary terms, contemporary animals are living edifices of past events and developments. They give us a narrow, nebulous window through which to espy and presage the assaults that life has endured and the changes which may have occurred. Kardong (1995) observes that "the architectural design of an organism expresses something about the processes that produced, it, the history out of which it came, and the functions its parts perform."

To present a truly comparative account, detailed considerations of individual animal species have been avoided. Instead, broad taxonomic groups are considered. The shared and dissimilar strategies, principles, mechanisms, and themes are instead rationalized. Data on individual species are given only to explain a feature, a process, a concept, or a theory. Effort has been made to balance out physiology with morphology, the latter having been largely relegated to the background in practically all earlier publications on the subject. This approach is adopted in the firm belief that morphology is not simply a synonym of anatomy, i.e., a description of the structure, topography, and composition of inert biological entities for their own sake. The discipline delves into the logical basis of form, casting back on the true spirit of the etymology of the word. Perceptively, Thompson (1959, p.14) expressed this notion as follows: "morphology is not only a study of material things and of the forms of material things, but has its dynamical aspect, under which we deal with the interpretation, in terms of force, of the operations of Energy."

This book was written with a broad readership in mind. Graduate students as well as established biologists, be they zoologists, physiologists, morphologists, paleontologists, or ecologists who work or may contemplate working on materials and aspects of respiration in whole organisms will find it useful. Scientists in Earth Sciences with an interest in the interactive developmental processes between the physical and biological realms will find the book of certain interest and perhaps stimulating. No apology is made that a rather lengthy section of the work has been devoted to the biophysical aspects of the respiratory media, habitats, and the accretion of molecular O2. The "history" of the evolution of the respiratory processes and that of gas exchangers in particular would be in great default without a clear grasp of the setting in which the changes occurred, the intractable challenges faced, and the successes and failures encountered as animals strived not only to survive but also to establish themselves and flourish in alien habitats. Some of the aspects covered here have taken me outside the limits of my personal experience and expertise. In such cases, I well realize that I may not have articulated the issues with the competence required by the specialists in those fields. Should any infelicities of judgment, inference, or omission have occurred, I beg the reader's indulgence, and would be most grateful if such aspects were pointed out to me. While consciously avoiding teleology, a rather difficult thing for one to do especially when rationalizing design, I have not hesitated to use explicative interpretations and suppositions where firm data are lacking. Fishman (1983) observed that "the understanding of biological systems rests on a combination of mechanisms, which deal with beginnings, and teleology, which deals with the concept of purpose and end", while Jones (1972) asserts that "frank and selfcontrolled (teleological) speculation is an essential part of the comparative approach". In such a long process, and one which has to be studied indirectly (due to lack of fossilized materials of soft tissues like the gas exchangers), it has to be conceded that "background noise", discernible as anomaly between the expected outcome and the reality, is to be anticipated. Evolutionary reconstruction of the history of a natural process is one of the most challenging problems that biologists have been called upon to solve. The skepticism with which such accounts are received was voiced by Rancour-Laferriere (1985), who stated that "anyone who attempts to narrate the evolution of something over geologic time is telling a story". Although, given the limitations imposed by the wide gaps in the fossil records, such skepticism is warranted, I consider such sentiments extreme and perhaps somewhat misplaced. When presenting a chronological account of a classical natural event, a measure of subjectivity should be allowed. More often than not, the best that can be accomplished is to build as complete a case as possible - based on intuitive sense, logical inferences, and known facts. A historically realistic model or a simulation of what could have happened should be fabricated and tested. This should show that, given specified conditions and circumstances, particular events were not only inevitable but most likely occurred. In the general purview of biology, Stebbins (1984) realistically observes that "the only law that holds without exception in biology is that exceptions exist for every law". I have explicitly stated where experimental facts are lacking and personal view has been proffered. I hope that such thoughts will provide areas for consideration and debate.

Due to limitation of space, it is not possible to give an encyclopedic review of the utterly immense aspects of the evolution and comparative respiratory morphology and physiology of the gas exchangers in a single work. There are excellent accounts, monographs, treatises, and perspectives on different areas of these aspects, especially on the vertebrates. The key publications are included in the reference list; they should be consulted where particular details are desired. While offering a synthesis of more recent findings, this account is meant to complement such comprehensive publications and not supersede them. I have purposely placed particular emphasis on the respiratory processes of the invertebrates, a group on which relatively less is known. In doing so, I hoped to avoid the conventional "prejudice" (against the group) and the parochial but somewhat legitimate interest and popular emphasis on the vertebrate aspects. In most people, mention of the word animal conjures up a vertebrate in their mind. With real setbacks, mammals in particular (a highly specialized taxon in most respects) have been used as the focal point (model group) in discussing comparative pulmonary structure and function. A prospective look at the design of the gas exchangers, i.e., from the lowest to the highest forms rather than retrospectively (i.e., back in time) has been made here in order to hopefully better fit the developments into the natural polarity of evolutionary change. This approach perhaps offers a more satisfactory causal explanation of the early developments in respiration. Moreover, reading evolution backwards has its intrinsic bias in that one inevitably sees an outcomes as the only possible result from a series of events. Nature is extremely complex and too unpredictable to be read easily. I hope that a satisfactory balance in the presentation has been achieved here. The designs of the gas exchangers in the so-called primitive animals (a very unfortunate term which has come to be equated with crude, mediocre, or imperfect, just as complex, superior, and advanced have come to be taken to mean better or efficient) are most instructive in understanding the structure and the operations of the more sophisticated processes of respiration. Tenney and Boggs (1985) observed that "the great events in respiratory system evolution can be appreciated best in lower classes". The diversity of the stratagems adopted for procuring molecular O<sub>2</sub> and eliminating CO<sub>2</sub> among animals having different lifestyles, occupying different habitats, and which have acquired remarkable morphological eccentricities are most evident when the underlying cost-benefit analysis and the compromises, the trade-offs (and even payoffs?), and the alternative solutions to the respiratory imperatives are comparatively examined. The protracted transactions and the transformations that went into the design and adoption of particular respiratory schemes and the perceivable limitations or capacities inherent in different gas exchangers can be recognized. A heuristic approach in biology reveals subtle areas of convergence, homology, analogy, and homeostasis, providing an explanation of the mechanisms which yielded particular states and phenomena. Nature, however, does not yield its secrets willingly. They have to be patiently gleaned and teased through protracted, well-planned inquiries and meticulous attention to details. A multidisplinary approach in such studies allows a broad understanding of a problem and offers robust answers.

The chapters in this account are sequenced to survey the respiratory events as they are conceived to have more or less chronologically unfolded. It is hoped that this approach offers a more logical understanding of the developments. Chapter 1 examines the fundamental attributes of life and its evolution on Earth, the accretion and fluctuations of O<sub>2</sub> and CO<sub>2</sub> in the biosphere, and the enigmatic nature of molecular O<sub>2</sub> in respiration and evolution of life itself and the intricate role it has played in the ecological adaptations of the animal life. Chapter 2 surveys the forms and designs of the gas exchangers, the underlying engineering principles which form the basis of their manifold plans and constructions, and argues out the factors which have imposed the different architectural contrivances. Chapter 3 considers the biophysical dictates of water and air as respiratory media and livable habitats, the adaptive respiratory needs for survival in unique environments and circumstances, the effects of the different respiratory milieus and habitats on the fabrication of the gas exchangers, the essence of the diverse respiratory stratagems adopted by different animal taxa, and the effect of gravity on the design and function of gas exchangers. Chapter 4 looks at the structure and function of the archetype aquatic gas exchanger - the gills - and considers the placenta as an ephemeral liquid-to-liquid gas exchanger. Chapter 5 examines the limitations and constraints which confronted animals during the switch from water- to air breathing and the pivotal import of transitional (bimodal) breathing in the traumatic process of terrestrial invasion. Chapter 6 outlines a cost-benefit analysis of what went into the utilization of the atmospheric O<sub>2</sub> and the structural and functional prescriptions in the evolved air-breathing organs and structures. I must regardless concede that in this treatise, not all aspects and issues pertaining to the evolution of respiration and the form and function of the gas exchangers have been covered and those which are may not have been exhausitively discussed and satisfactorily resolved. Based on personal preferences on the subject matter and keeping in perspective the restriction of space, many elisions were made. Only the weighed imperatives were included. My objective will have been more than adequately met if the discourse provides areas for reflection, points for discussion, or, better still, aspects for further investigations on the many yet gray areas.

# Contents

#### Chapter 1 Perspectives on Life and Respiration: How, When, and Wherefore

1.1	Life: Diversity, Complexity, and Uniformity Fabricated	
	on Simplicity	1
1.2	The Earth: a Highly Dynamic Planet	7
1.3	Factors that Encouraged the Evolution of Life on Earth	8
1.4	Oxygen: a Vital Molecular Resource for Life	10
1.5	Anaerobic Metabolism and Adaptive Success in Animals	12
1.6	Evolved Mechanisms and Strategies of Procuring Molecular O <sub>2</sub>	14
1.7	Explicating the Process of Evolution of Respiration: Limitations	20
1.8	Plans and Performance Measures of the Gas Exchangers	21
1.9	The Early Anoxic Earth and the Evolution of Life	25
1.10	Abundance of Molecular $O_2$ in the Earth's Biosphere	27
1.11	Shift from Anaerobiotic to Aerobiotic State in the Early Earth	27
1.12	Accretion of Molecular O <sub>2</sub>	33
1.13	CO <sub>2</sub> Pulses in the Biosphere	35
1.14	The Overt and Covert Roles of O <sub>2</sub> in Colonization	
	and Extinctions of Biota	38
1.15	Oxygen: a Paradoxical Molecule	40
1.16	The Rise of the Level of Molecular $O_2$ : a Curse or a Blessing?	41
	1.16.1 The Deleterious Reactive Radicals of Molecular O <sub>2</sub>	41
	1.16.2 Senescence: the Effects Molecular O <sub>2</sub>	43
	1.16.3 Biological Defenses Against O <sub>2</sub> Toxicity	45
1.17	The Evolution of Complex Metabolic Processes	47
1.18	Oxygen and CO <sub>2</sub> as Biochemical Factors in Respiration	48
1.19	Homeostasis: the Role of Respiration	49
Chap	pter 2 Essence of the Designs of Gas Exchangers –	
the I	Imperative Concepts	
2.1	Innovations and Maximization of Respiratory Efficiency	53
2.2	Safety Factors and Margins of Operation of Gas Exchangers	55
2.3	Engineering Principles in the Design of the Gas Exchangers	62
2.4	Scopes and Limitations in the Design and Refinement	
	of the Gas Exchangers	70
2.5	Optimal Designs in Biology and Gas Exchangers in Particular	73

2.5.1	Symmorphosis: the Debate	73
2.5.2	The Operative Strategies for Optimization	
	in the Gas Exchangers	76

	2.5.3	Symmorphosis and Optimization: are they Logical Outcomes	
	of Evolution?		
2.6	Fractal Geometry: a Novel Approach		
	for Discerning Biological Form		
2.7	From Diffusion, Perfusion, and Ventilation		
	to Res	piratory Pigments	87
	2.7.1	Diffusion	87
	2.7.2	Convective Flows	91
2.8	Blood	and the Respiratory Pigments	104
	2.8.1	Hemoglobinless Fish	113
2.9	Energ	etic Cost and Efficiency of Respiration	114
	2.9.1	The Requisites for Efficient Gas Exchange	121
	2.9.2	Efficient vs. Inefficient – Primitive vs. Advanced Gas	
• • •		Exchangers: the Contention	130
2.10	Model	ing: Utility in Study of Integrative Construction	
	of the	Gas Exchangers	132
	2.10.1	Evaluation of the Functional Efficiency	
		of the Gas Exchangers	134
	2.10.2	Modeling the Gas Exchangers	135
Chaj	oter 3	Gas Exchange Media, Respiratory States, and Environments	
3.1	Water	and Air as Respiratory Media: General Considerations	149
3.2	Physic	al Charateristics of Water and Air	151
3.3	The D	istribution of Water and Air on Earth	152
3.4	Water	a Respirable Medium and an Integral Molecule for Life	153
	3.4.1	Oxygen and CO <sub>2</sub> Content in Water: Effect on Respiration	155
	3.4.2	Density and Viscosity of Water	159
	3.4.3	Thermal Capacity and Conductivity of Water	160
	3.4.4	Derelict Waters: Respiratory Stress from Hypercapnia	
o -	-	and Hypoxia	161
3.5	Terres	trial Habitation and Utilization of Atmospheric $O_2$	165
3.6	Hydro	gen Sulfide Habitats. Tolerance and Utilization	166
3.7	The Po	prosphere and Fossorial Respiration	169
	3.7.1	Gaseous Composition in Burrows	170
• •	3.7.2	Burrowing Aquatic Annelids, Crustaceans, and Fish	172
3.8	Living	at High Altitude: Coping with Hypoxia and Hypobaria	173
	3.8.1	Tolerance of Arterial Hypocapnia in Birds	175
	3.8.2	Flying over Mt. Everest: the Bar Headed Goose,	
• •		Anser indicus	176
3.9	Gravit	y: Effects on Respiratory Form and Function	177
Chap	oter 4	Water Breathing: the Inaugural Respiratory Process	
4.1	The D	esign of the Gills	181
4.2	Adapt	ive Diversity and Heterogeneity of Gill Form	183
4.3	The Fu	unctional Innovations of the Gills for Aquatic Respiration	186
4.4	The Simple Gills		
	4.4.1	Morphological Characteristics	188

	4.4.2	Ventilation and Functional Capacities	189
	4.4.3	Gas Exchange Pathways and Mechanisms	190
4.5	The Co	omplex Gills	194
	4.5.1	Structure and Architectural Plans	194
4.6	The W	ater Lungs	198
4.7	The Pla	acenta: an Ephemeral Liquid to Liquid Gas Exchanger	204
	4.7.1	The Functional Reserves of the Placentae	209
Char	oter 5	Bimodal Breathing: Compromise Respiration	
5.1	The W	ater-Air Interface: an Abstract Respiratory Rubicon	217
5.2	Strateg	ies and Adaptive Convergence for Air Breathing	223
5.3	Risks,	Costs, and Benefits in the Change to Air Breathing	227
5.4	6.4 CO, Elimination: Impediment to Evolution of Air Breathing		
	and Te	rrestriality	231
5.5	Contro	and Coordination of the Bimodal Gas Exchange Process	237
	5.5.1	Ventilatory Modalities of the Gas Exchangers	
		in the Bimodal Breathers	240
	5.5.2	The Circulatory Patterns in the Gas Exchangers	
		of the Bimodal Breathers	242
5.6	Taxa w	with Notable Propensity for Bimodal Breathing	245
	5.6.1	Mollusks	245
	5.6.2	Crustaceans	248
	5.6.3	Fish	252
Chaj	pter 6	Air Breathing: the Elite Respiration	
<b>Chaj</b> 6.1	p <b>ter 6</b> Is the	Air Breathing: the Elite Respiration Surface of the Lung Dry, Moist, or Wet? Do Real Air	262
<b>Chaj</b> 6.1	p <b>ter 6</b> Is the Breath	Air Breathing: the Elite Respiration Surface of the Lung Dry, Moist, or Wet? Do Real Air ers Exist?	263
<b>Chaj</b> 6.1 6.2	p <b>ter 6</b> Is the Breath Lung a	Air Breathing: the Elite Respiration Surface of the Lung Dry, Moist, or Wet? Do Real Air ers Exist?	263
Chaj 6.1 6.2	pter 6 Is the Breath Lung a and fo	Air Breathing: the Elite Respiration Surface of the Lung Dry, Moist, or Wet? Do Real Air ers Exist? and Swim Bladder – Which Developed Earlier r What Purpose?	263 265
Chaj 6.1 6.2 6.3	oter 6 Is the Breath Lung a and fo Evolut	Air Breathing: the Elite Respiration Surface of the Lung Dry, Moist, or Wet? Do Real Air ers Exist?	263 265 271
Chaj 6.1 6.2 6.3 6.4	Is the Is the Breath Lung a and fo Evolut Aerial	Air Breathing: the Elite Respiration Surface of the Lung Dry, Moist, or Wet? Do Real Air ers Exist?	263 265 271 276
Chay 6.1 6.2 6.3 6.4 6.5	Is the Breath Lung a and fo Evolut Aerial The D	Air Breathing: the Elite Respiration Surface of the Lung Dry, Moist, or Wet? Do Real Air ers Exist?	263 265 271 276 277
Chay 6.1 6.2 6.3 6.4 6.5 6.6	Is the Breath Lung a and fo Evolut Aerial The D The M	Air Breathing: the Elite Respiration Surface of the Lung Dry, Moist, or Wet? Do Real Air ers Exist?	263 265 271 276 277 280
Chaj 6.1 6.2 6.3 6.4 6.5 6.6	Is the Breath Lung a and fo Evolut Aerial The D The M 6.6.1	Air Breathing: the Elite Respiration Surface of the Lung Dry, Moist, or Wet? Do Real Air ers Exist?	263 265 271 276 277 280 280
Chay 6.1 6.2 6.3 6.4 6.5 6.6 6.7	pter 6 Is the Breath Lung a and fo Evolut Aerial The D The M 6.6.1 The Co	Air Breathing: the Elite Respiration Surface of the Lung Dry, Moist, or Wet? Do Real Air ers Exist?	263 265 271 276 277 280 280 288
Chaj 6.1 6.2 6.3 6.4 6.5 6.6 6.7	pter 6 Is the Breath Lung a and fo Evolut Aerial The D The M 6.6.1 The Co 6.7.1	Air Breathing: the Elite Respiration Surface of the Lung Dry, Moist, or Wet? Do Real Air ers Exist?	263 265 271 276 277 280 280 288
Chaj 6.1 6.2 6.3 6.4 6.5 6.6 6.7	pter 6 Is the Breath Lung a and fo Evolut Aerial The D The M 6.6.1 The C 6.7.1	Air Breathing: the Elite Respiration Surface of the Lung Dry, Moist, or Wet? Do Real Air ers Exist?	263 265 271 276 277 280 280 288 288
Chaj 6.1 6.2 6.3 6.4 6.5 6.6 6.7	pter 6 Is the Breath Lung a and fo Evolut Aerial The D The M 6.6.1 The C 6.7.1 6.7.2	Air Breathing: the Elite Respiration Surface of the Lung Dry, Moist, or Wet? Do Real Air ers Exist?	263 271 276 277 280 280 288 288 288 298
Chaj 6.1 6.2 6.3 6.4 6.5 6.6 6.7	pter 6 Is the Breath Lung a and fo Evolut Aerial The D The M 6.6.1 The C 6.7.1 6.7.2 6.7.3	Air Breathing: the Elite RespirationSurface of the Lung Dry, Moist, or Wet? Do Real Airers Exist?ers Exist?and Swim Bladder – Which Developed Earlierr What Purpose?ion of Air Breathing and Terrestriality: the LimitationsGas Exchangers: Structural and Functional Diversityiffusive Type Gas Exchangersixed Type Gas ExchangersThe Insectan Tracheal Systemonvective Type Gas ExchangersVentilatory Mechanisms and Organizationof the Gas ExchangersThe Amphibian LungThe Reptilian Lung	263 271 276 277 280 280 288 288 298 304
Chaj 6.1 6.2 6.3 6.4 6.5 6.6 6.7	pter 6 Is the Breath Lung a and fo Evolut Aerial The D The M 6.6.1 The C 6.7.1 6.7.2 6.7.3 6.7.4	Air Breathing: the Elite RespirationSurface of the Lung Dry, Moist, or Wet? Do Real Airers Exist?und Swim Bladder – Which Developed Earlierr What Purpose?ion of Air Breathing and Terrestriality: the LimitationsGas Exchangers: Structural and Functional Diversityiffusive Type Gas Exchangersixed Type Gas ExchangersThe Insectan Tracheal Systemonvective Type Gas ExchangersVentilatory Mechanisms and Organizationof the Gas ExchangersThe Amphibian LungThe Mammalian Lung	263 271 276 277 280 280 288 288 298 304 308
Chaj 6.1 6.2 6.3 6.4 6.5 6.6 6.7	pter 6 Is the Breath Lung a and fo Evolut Aerial The D The M 6.6.1 The C 6.7.1 6.7.2 6.7.3 6.7.4 6.7.5	Air Breathing: the Elite RespirationSurface of the Lung Dry, Moist, or Wet? Do Real Airers Exist?und Swim Bladder – Which Developed Earlierr What Purpose?ion of Air Breathing and Terrestriality: the LimitationsGas Exchangers: Structural and Functional Diversityiffusive Type Gas Exchangersixed Type Gas ExchangersThe Insectan Tracheal Systemonvective Type Gas ExchangersVentilatory Mechanisms and Organizationof the Gas ExchangersThe Amphibian LungThe Nammalian LungThe Avian Lung	263 271 276 277 280 280 288 288 298 304 308 314
Chaj 6.1 6.2 6.3 6.4 6.5 6.6 6.7	pter 6 Is the Breath Lung a and fo Evolut Aerial The D The M 6.6.1 The C 6.7.1 6.7.2 6.7.3 6.7.4 6.7.5 The In	Air Breathing: the Elite RespirationSurface of the Lung Dry, Moist, or Wet? Do Real Airers Exist?und Swim Bladder – Which Developed Earlierr What Purpose?ion of Air Breathing and Terrestriality: the LimitationsGas Exchangers: Structural and Functional Diversityiffusive Type Gas Exchangersixed Type Gas ExchangersThe Insectan Tracheal Systemonvective Type Gas ExchangersVentilatory Mechanisms and Organizationof the Gas ExchangersThe Amphibian LungThe Reptilian LungThe Avian LungThe Avian Lungtternal Subdivision of the Lung: the Functional	263 271 276 277 280 280 288 288 298 304 308 314
Chaj 6.1 6.2 6.3 6.4 6.5 6.6 6.7	pter 6 Is the Breath Lung a and fo Evolut Aerial The D The M 6.6.1 The C 6.7.1 6.7.2 6.7.3 6.7.4 6.7.5 The Ir Implic	Air Breathing: the Elite RespirationSurface of the Lung Dry, Moist, or Wet? Do Real Airers Exist?und Swim Bladder – Which Developed Earlierr What Purpose?ion of Air Breathing and Terrestriality: the LimitationsGas Exchangers: Structural and Functional Diversityiffusive Type Gas Exchangersixed Type Gas ExchangersThe Insectan Tracheal Systemonvective Type Gas ExchangersVentilatory Mechanisms and Organizationof the Gas ExchangersThe Amphibian LungThe Reptilian LungThe Ammalian LungThe Avian LungAvian Lungations	263 265 271 276 277 280 280 288 288 298 304 308 314 323
Chaj 6.1 6.2 6.3 6.4 6.5 6.6 6.7 6.8 6.8 6.9	pter 6 Is the s Breath Lung a and fo Evolut Aerial The D The M 6.6.1 The C 6.7.1 6.7.2 6.7.3 6.7.4 6.7.5 The Ir Implic The Su	Air Breathing: the Elite RespirationSurface of the Lung Dry, Moist, or Wet? Do Real Airers Exist?und Swim Bladder – Which Developed Earlierr What Purpose?ion of Air Breathing and Terrestriality: the LimitationsGas Exchangers: Structural and Functional Diversityiffusive Type Gas Exchangersixed Type Gas ExchangersThe Insectan Tracheal Systemonvective Type Gas ExchangersVentilatory Mechanisms and Organizationof the Gas ExchangersThe Amphibian LungThe Reptilian LungThe Avian LungThe Avian Lungationsurfactant: a Versatile Surface Lining of the Gas Exchangers	263 265 271 276 277 280 280 288 288 298 304 308 314 323 330
Chaj 6.1 6.2 6.3 6.4 6.5 6.6 6.7 6.8 6.8 6.9 6.10	pter 6 Is the s Breath Lung a and fo Evolut Aerial The D The M 6.6.1 The C 6.7.1 6.7.2 6.7.3 6.7.4 6.7.5 The Ir Implic The Su Nonre	Air Breathing: the Elite RespirationSurface of the Lung Dry, Moist, or Wet? Do Real Airers Exist?and Swim Bladder – Which Developed Earlierr What Purpose?ion of Air Breathing and Terrestriality: the LimitationsGas Exchangers: Structural and Functional Diversityiffusive Type Gas Exchangersixed Type Gas Exchangersonvective Type Gas Exchangersventilatory Mechanisms and Organizationof the Gas ExchangersThe Amphibian LungThe Reptilian LungThe Avian LungThe Avian Lungationsatio	263 265 271 276 277 280 280 288 288 298 304 308 314 323 330 334
Chaj 6.1 6.2 6.3 6.4 6.5 6.6 6.7 6.8 6.9 6.10	pter 6 Is the J Breath Lung a and fo Evolut Aerial The D The M 6.6.1 The C 6.7.1 6.7.2 6.7.3 6.7.4 6.7.5 The Ir Implic The Su Nonre 6.10.1	Air Breathing: the Elite Respiration Surface of the Lung Dry, Moist, or Wet? Do Real Air ers Exist?	263 271 276 277 280 288 288 298 304 308 314 323 330 334 334

6.11 The Implications of Liquid Breathing in Air Breathers	341
6.12 Physical Gill and the Plastron:	
a Unique Underwater Respiratory Strategy	344
6.13 The Cleidoic Egg: a Fascinating Gas Exchanger	349
6.14 The Bottom Line	357
References	361
Subject Index	483

# Perspectives on Life and Respiration: How, When, and Wherefore

"A reconstruction of the remote past must necessarily be based upon inference, rarely from systematically collected data, more often from an inadequate number of facts which chance has placed in the way of competent investigators who can recognize their significance. As time goes on some gaps are filled, others remain forever empty, but the picture as a whole becomes progressively clearer." Beadle (1974).

#### 1.1 Life: Diversity, Complexity, and Uniformity Fabricated on Simplicity

Humankind has always been fascinated by the spectacle of extreme states and phenomena. The Guinness Book of Records, which after the Holy Bible is alleged to be the second most widely read book, is according to the publishers compiled "in hope of providing a means for peaceful setting of arguments about record performances". Though not given much attention outside the professional realms, the elegance and constellation of life on Earth is enchanting, bewildering, and intellectually intriguing. More than 284000 species of plants, 750000 species of insects, and 280000 species of other animals have been catalogued (e.g., Dixon 1994; Service 1997). Of this plethora, vertebrates represent only one phylum and a mere 50 000 species or so (e.g., Pough et al. 1989). Nature's fortitude for survival is remarkable. For example, albeit the tumultuous crises which preceded the Tertiary period when colossal population clashes occurred and many species were wiped out, by the end of the period, there were as many as 2500 families of animals (Benton 1995). From molecular sequence studies of different microcosms (e.g., Pace et al. 1985; Ward et al. 1990; Winker and Woese 1991; Olsen and Woese 1993), it is becoming unequivocally evident that, compared with the Metazoa and the Metaphyta (i.e., the visible world), the microbial domain (i.e., the microworld) presents a more complex biodiversity than was hitherto thought, and quantitatively remains largely unknown to us (e.g., Embley et al. 1994; Lovejoy 1994). The existing numerical data on the taxonomic diversity of animal life differ remarkably. It is envisaged that life's copious tree comprises between 5 and 50 million species of animals (e.g., May 1988, 1990, 1992; Hammond 1992). This hopelessly wide range owns up to the fact that we know very little of the actual richness and geographical patterning of contemporary animal life (e.g., Wilson 1992; Colwell and Coddington 1994). Since most of the crucial environmental processes are driven by the microbial activities which involve biochemical nutrient recycling operations such as nitrogen fixation, sulfur oxidation and reduction, ammonification, methanogenesis, and methane oxidation (e.g., Capone et al. 1997), it can legitimately be anticipated that the microorganismal biomass (which must form the base of life) should overwhelm that of plants and animals. The methanogenic bacteria which occur in the hindguts of many

arthropods contribute substantially to the loading of the atmosphere with methane (e.g., Hackstein and Stumm 1994). Fenchel (1992) estimated that a 1 cm<sup>3</sup> core of coastal marine sediment contains  $4 \times 10^{10}$  bacterial cells,  $10^4$  heterotrophic flagellates and amoebae, 10<sup>8</sup> chlorophyll a-containing microorganisms, and about 2000 ciliates. Expressing a personal view on the ambitious program named Systematics Agenda 2000, which envisages that all the Earth's species will be discovered, described, and classified in the next 25 years (Anonymous 1993), Wicksten (1994) doubts that "the world's organisms will be described within the next century - if ever"! The same may be said on the Fifty-Year Plan (e.g., Raven and Wilson 1992). Applying the lengthy traditional methods of identifying and cataloguing species, May (1990), Hawksworth (1991), and Hammond (1992) envisage that it will take a couple of centuries to gain an adequate understanding of species diversity on Earth. With speciation being a continuous process (e.g., Butlin and Tregenza 1997; Smith et al. 1997), there may not be an actual end to the task. From the microbes to the whales, living animals differ in mass by at least 21 orders of magnitude (McMahon and Bonner 1983; Schmidt-Nielsen 1984; Brown 1995). The staggering numerical density, specific diversity, and allometric disparity is a clear testimony of the tenacity, resiliency, richness, and innovativeness of nature's designs for survival. It remains a great challenge to the biologists to fully explain the factors which drive and determine specific patterning (Hutchison 1959; Brown 1981; Cracraft 1994; Butlin and Tregenza 1997).

Until the enunciation of the theory of evolution through natural selection (Darwin and Wallace 1858; Darwin 1859), the complexity of life on Earth was scientifically inexplicable. Since then, the kaleidoscopy has been perceived as integral to the dynamic process of evolution (e.g., Ruthen 1993), with the degree of profuseness and variability a mark of success and suitability to an environment. The spatial and temporal distribution of species is set by definite physical, biological, and environmental controls. Through the about 4 billion years (billion = 1 thousand million years =  $10^{9}$ ) of existence on Earth, living forms have adopted behavioral strategies, developed biochemical and functional capacities, and appropriated certain devices which enable them to occupy speciesspecific niches. Such stable states have been attained amidst profound ambient changes that have included variations in the rotation of the Earth, changes in the Earth's orbit, fluctuations in the average surface temperature, physical displacements of the continental plates, pulses in the O<sub>2</sub> and CO<sub>2</sub> levels, and fluctuations in the availability of important nutrients (e.g., Hayes et al. 1976; Hunt 1979; Ben-Avraham 1981; Boucot and Gray 1982; Walker et al. 1983; Bray 1985; Raymo and Ruddiman 1992). In their different ecological settings, the nature, dynamics, and heterogeneity of environments detail the dissimilitude of animal life: form and function are molded by the physical and biotic factors in the ecotopes. Based on the separate traits they have acquired through interfacing with environments, animal life can be grouped into aquatic, terrestrial, and aerial assemblages. Finer divisions place them into, e.g., nocturnal, diurnal, fossorial, pelagic, arboreal, benthonic types, etc. Since environments are dynamic spaces (e.g., Schaffer and Kot 1986; Doebeli 1993; Rand and Wilson 1993), organisms must equally be phenotypically fluid to continually adapt to the external cues (e.g., Stearns 1982; Prosser 1986). While freely interacting with the environment, organisms must not compromise their physical and biological integrity as self-sustaining, selfregulating dynamical entities. Energy must be obtained from the environment to regulate their internal states and defend homeostasis. To steady the environmental oscillations, strictly, organisms are a process in a nonsteady state (e.g., Levins and Lewontin 1985; Wainright 1988). As urged by Simpson (1953), when considering adaptation, "it is equally or more useful to focus neither on environment nor on organisms but on the complex interrelationship in which they are not really separable". The term comprehensive selective regime was used by Baum and Larson (1991) to define the combination of all the environmental and organismic factors which prescribe how natural selection acts on organismal variations. Chronobiology (circadian rhythmicity), a process which has evolved to sense and counterbalance the nuances of the environment, pervades all levels of biological organization from molecular, cellular, to organismal (e.g., Aschoff and Pohl 1970; Bünning 1973; Sweeney 1987; Ishii et al. 1989; Prinzinger and Hinninger 1992; Lloyd and Rossi 1993; Martin and Palumbi 1993; Page 1994). Through a mechanism termed endogenous clock or pacemaker (e.g., Sassone-Corsi 1996), circadian rhythmic activities occur even under constant environmental conditions (e.g., Aldrich and McMullan 1979; Prinzinger and Hinninger 1992). In such cases, it is thought that the property helps organisms anticipate the exigencies of life through programmed cyclic regulation of specific target genes. The common inhibitory neurotransmitter in the central nervous system, GABA (yaminobutyric acid), has been implicated in circadian rhythmicity (e.g., Wagner et al. 1997).

Life occurs in backgrounds which after cursory glance can resolutely be dismissed as being implicitly inhospitable. Nature, however, appears to abhor a vacuum. Practically every nook and cranny on Earth is filled with some kind of life. Animals which have adapted to extreme environmental circumstances have particularly intrigued biologists (e.g., Madigan and Marrs 1997). Habitats such as the subzero temperature glaciers (e.g., Arrigo et al. 1997), the fiercely hot bubbling hydrothermal (volcanic) vents in the deep seas (e.g., Meredith 1985; Jonnston et al. 1986), the highly desiccating tropical deserts, the fresh (nearly ionfree) water, the hypersaline lakes, the remote virtually anoxic reducing muds, and submarine environments where the hydrostatic pressures may exceed 1000 atmospheres have all been variably conquered and occupied. Life has been found at a depth of about 4 km below the surface of the Earth where the temperature is 75 °C (Ehrlich 1996; Frederickson and Onstott 1996) and in the ice-free, cold, dry valleys of the Antarctica, regions considered to be the closest terrestrial analogs of the Martian and other extraterrestrial planetary environments (Friedmann 1982; Friedmann and Ocampo 1976). While few, if any, multicellular organisms can tolerate a temperature above 50 °C (Huey and Kingsolver 1993), some microbes, i.e., the so-called hyperthermophiles, thrive at and above 100 °C (Madigan and Marrs 1997), the boiling point of water at sea level. In their quest to secure new habitats, organisms have devised design-specific solutions to the vast threats which have resolutely besieged them. These have ranged from momentous changes such as variations in solar insolation (e.g., Newman and Rood 1977; Frils-Christensen and Lassen 1991) and volcanic activity to minor spatial and temporal shifts in temperature and levels of respiratory gases in their immediate habitats. Such changes have had dramatic effects on the form, distribution, and lifestyles of animal life. Hippopotamuses are reported to have roamed the Yorkshire Dales some 125000 years before present (Shackleton 1993) and in northern Africa, the Sahara-Sahel boundary crept northwards by 10° of latitude between 18 and 8 thousand years before present (Petit-Maire 1991). The overall fitness of an organism in a particular environmental setting is an aggregate effect of the different adaptive strategies an animal has requisitioned during its evolutionary existence (Kozlowski 1993). This arsenal confers the adaptive capacity to withstand adverse changes in the environment and, if necessary, to actively carve out and exploit new, more hospitable and resourceful habitats.

From the smallest known entities, the fundamental subatomic particles such as quarks, to the observable universe (the largest entity of which humankind has knowledge), a scale which ranges from under  $10^{-15}$  to  $10^{27}$  (Ronan 1991), nature is governed by four fundamental forces. These are gravity, electromagnetism, the weak force (the force responsible for radioactive decay), and the strong nuclear force (the force that holds the nucleus of an atom together). Gravity, of which the carrier particle (the graviton) is still undiscovered, is the weakest of the forces (strength 10<sup>-38</sup>) but has an infinite range. The weak nuclear force, of which the carriers are the electrically charged  $W^+$  and the  $W^-$  and the neutral  $Z^\circ$  particles. has a range of  $10^{-15}$  and a strength of  $10^{-13}$ , the electromagnetic force, of which the carrier particles are photons, has an infinite range, a strength of  $10^{-2}$ , and operates between electrically charged particles, while the strong nuclear force which acts on the quarks is powerful (strength = 1) but has a range of only  $10^{-12}$ . By studying the universal properties of matter, science strives to understand and test how these characteristics contrived matter culminating in the phenomenon of life. It is hoped that the principles which govern organismal existence, design<sup>1</sup>, behavior, function, and life-style can be better grasped as a part of such a broad fundamental and integrative approach. The quest for the elusive grand unified theory (e.g., Hawking 1993; Weinberg 1994), an attempt to integrate a number of independent mathematical equations which seek to demonstrate that three of the fundamental forces of nature (electromagnetism, and the weak and strong nuclear forces) are essentially performances of the same superforce, continues with zeal and zest. After unification with gravity, the theoretical physicists contemplate advancing the "theory of everything", an encompassing principium which should explain the formation of the Universe at the Big Bang, the subsequent existence of the material world, and perhaps the development of life (Moore 1990a; Maynard-Smith and Szathmary 1996; Ronan 1991) and the end of it all at the Big Crunch. Though some measure of success has been achieved, especially in harmonizing the weak and electromagnetic forces, from what is

<sup>&</sup>lt;sup>1</sup> The term design, which is borrowed from engineering, is used in this book in a sense to mean "creative natural arrangement of parts (= components) in a device (= gas exchanger) for a particular purpose (respiration)". Vogel (1988) defined biological design as "functionally competent arrangement of parts resulting from natural selection".

known presently, the cosmos abounds with mystery and deliberately shuns order and conformity. While the Newtonian laws apply in the intermediate scales of biology, at subatomic (the so-called nanoworld) level, quantum mechanics reigns, and at the cosmological level, relativity holds true. Recently, a concept dubbed new physics was professed to be highly rewarding in analyzing the dynamics of the life processes (Davies 1989; Stonier 1990). The approach is based on arguments that: (1) life is transmitted through what are bits of information (algorithms) inscribed in the genome and (2) the frequency and amplitude of biological events are regulated through ultrafast integrated information processing. Applying an electronic analogy, Lloyd and Rossi (1993) asserted that "the living state is an ensemble of oscillators" and that information processing may be the "common denominator" (the missing link?) in the formulation of a "unifying theory". Inasmuch as our understanding of biological processes (especially neurobiology) continues to accumulate with the advances in computer technology, this supposition appears to be credible (e.g., Adams 1979; Kawasaki 1993). Other lines of inquiries attempting to simplify and explain biological order and diversity include the so-called life-history theory (e.g., Charnov 1993; Ruthen 1993) and the synthetic theory of biological organization (e.g., Eigen and Schuster 1979; Fontana and Buss 1993). According to these concepts, it is argued that for a given organism, if the fundamental parameters which govern and regulate resource procurement, utilization, regeneration, and self-perpetuation in a particular environment are known, it would be possible to model the life patterns which optimize fitness (Charnov 1997; Godfray 1997). Reductionism and all mechanistic approaches to biology endeavor to explain natural phenomena by manipulating fewer and simpler components that are responsive to exact simple physical laws (e.g., Popper 1968, 1969). However, when dealing with complex natural dynamical entities like organisms (e.g., Mann 1982; Brown 1993), it is not only difficult to correctly identify such components but also practically impossible to predict the outcomes of their myriad nonlinear interactions. Due to the hierarchical organization of biological systems from atomic, molecular, cellular, tissues, organs, organisms, through populations and communities to ecosystems, boundary conditions exist (Brown 1994): it is practically impossible to predict the outcomes from one level of organization to another (Polanyi 1968). Despite the caveats, the rationale behind the rather esoteric reductionistic investigations in biology includes among others the warranted recognition that although biological systems may evolve through means different from those that accrete the physical ones, their forms and states are fundamentally governed by the universal, permeative properties of matter and energy (e.g., Nagel 1961; Brooks 1994). The structure and the mechanistic chemical juxtaposition of organic molecules is a programmed process which generates exact structural configurations and arrangements which yield stable states. Life cannot violate the immutable laws of physics and chemistry during its development. Thompson (1959, p. 8) points out that "the forces which operate in the body are of the same character as are the inorganic forces". He envisages that mathematics and physics will greatly contribute in explaining biological phenomena even though they may not fully account for certain aspects that he consigns to the "soul". As cautioned by Giebisch et al. (1990) and Nurse (1997), when dealing with an intricate process such as life, reductionistic approaches may fail simply due to the fact that the underlying assumption that encompassing properties of an organism can be understood by studying its individual parts could be fatally flawed. The aggregate expression of the functional processes in an organism is not necessarily equal to that expressed by the intact animal. Thompson (1959, p. 41) observed that "the life of the body is more than the sum of the properties of the cells of which it is composed" and Hoagland and Dodson (1995) note that "an organism is greater than the sum of its parts". The total of the O<sub>2</sub> consumption of the individual tissues of the body, for example, may be lower (e.g., Weymouth et al. 1944; Itazawa and Oikawa 1983; Oikawa and Itazawa 1993) or higher (e.g., Terroine and Roche 1925; Crandall and Smith 1952; Vernberg 1954; Lilja 1997) than that of the whole animal. Based on a similar observation, Von Bertalanffy and Estwick (1953) proposed that the decrease in the mass-specific metabolic rate of an intact animal is regulated by "factors lying in the organism as a whole which do not appear in tissues excised from the intact animal". Brown (1994) contends that "while physical scientists seek precise answers to relatively simple problems, biologists on the whole seek approximate answers to very complex problems".

In what may be deemed convergence between natural and human engineering designs, modern research into natural configurations has led to the fascinating discipline of bionics (= biomechanics) (e.g., Nachtigall 1991; Witt and Lieckfeld 1991). Astonishing parallels in the "plans" and "constructions" of living organisms with technical principles abound. Though nature uses essentially the same structural materials as those used by human beings (i.e., materials found on Earth), while it is possible to mimic them, it is virtually impossible to exactly duplicate natural designs. During the long period of evolution, through progressive and vet recursive trial-and-error processes (e.g., Schaeffer 1965a), nature has honed and produced perfect or near-perfect innovations. Natural solutions for complex problems are often strikingly simple and fascinating (e.g., Hayes 1994): minimal resources are committed in configuring highly cost-effective structures. Amidst the remarkable diversity of form at the organismal level, however, the differences that distinguish the various kinds of life in the macroscopic and microscopic worlds disappear down the organizational cascade as similarities preponderate. From the perspective of structural chemistry, at the molecular and atomic levels, there are no differences between the living and nonliving worlds: organic molecules are made up of essentially the same elements (atoms) which comprise the inorganic ones but are arranged into complexes with unique properties. Life's diversity can be attributed to differences in the characters and arrangements of protein molecules which constitute more than half of the nonwater mass in a cell (Hoagland and Dodson 1995). Though the number of molecules which formed with the accrual of the Earth (inorganic evolution) and development of life (organic evolution) (Kirschner 1994; Weinberg 1994) is immense and to this bounty chemists keep on throwing in new ones, from this vast collection, life has been extremely selective on what it has appropriated. Of the about 8 million now known chemical compounds (e.g., Morgan 1995), only a very small number has been incorporated in the evolved biochemistry. Proteins, some of the largest and most complex molecules known and perhaps the most important organic factors, as they form enzymes which catalyze chemical reactions, are configured around only 20 different amino acids. From the infinite threedimensional possible dispositions, the forms and behaviors of proteins are limitless (e.g., Ronan 1991). Amazingly, the amino acids are produced through a code which is written in only four molecules (the nucleotide bases) which present 64 possible arrangements in triplets, the codons: 61 codons are distributed among 20 amino acids, the other 3 serving as stop codons. Proteins are intrisically dynamic molecules. Flauenfelder et al. (1991) pointed out that evolution occurs through changes in the primary sequence of proteins, a process which leads to changes in the structure and the conformational energy landscapes. From the well over 100 known elements, about 99% of the living matter is fundamentally made up of four elements, i.e., C, O<sub>2</sub>, N<sub>2</sub>, and H<sub>2</sub>. Of the 28 selected elements in the human body, H<sub>2</sub> and O<sub>2</sub> are the most abundant, respectively comprising 63 and 25.5% of it. Carbon is central to life. It forms chains and rings that can be elaborated into an immense number of complex compounds and makes up about 1% of the mass of the Earth. Carbonbased fuels contribute about 75% of the energy that is currently used on Earth. Methane is the most abundant organic molecule in the Universe (Ancilotto et al. 1997).

Succinctly put, biology uses a characteristic set of elements and compounds to carry out an infinite array of processes. It is bewildering that the most complex state of organization that matter has consummated has been fabricated through remarkable simplicity, essentially during a chemical circumstance which entailed microscopic architecture around the carbon atom. Though the chemical constituents of living organisms have been recognized and the biochemical processes which support life are now reasonably well understood, the actual origin of life remains a mystery (Szathmàry 1997). It is now known that organic molecules abound in the cosmos. Such molecules could have been seeded on Earth from space (e.g., Ronan 1991; Cohen 1995), landing at the right place at the right time. The recent report by NASA scientists on chemical fingerprints of extremely primitive life in a 4.5-billion-year-old piece of Martian rock (Kerr 1996a; McKay et al. 1996) may in future totally change our concept and the very definition of life.

#### 1.2 The Earth: a Highly Dynamic Planet

Though from space it looks serene and motionless, the Earth is a perpetually metamorphosing planet. Life, the most complex organization of matter, has astonishingly evolved against a highly dynamic setting. Geometrically about a sphere (but strictly an ellipse) some 12700 km in diameter, a mass of about  $6 \times 10^{21}$  tonnes and a volume of about  $1.1 \times 10^{12}$  km<sup>3</sup>, it spins round on an inclined axis (23.5° to the perpendicular of the plane of its orbit) at a speed of about 28 km/s (at the equator), moving around the sun (from a distance of 150 million km) along a slightly elliptical orbit about 300 million km in length at an incredible speed in excess of 1700 km/s. The rotation is completed in 23 h and 56 min and the revolution (around the sun) takes 365.25 days: the rotation causes

day and night while the revolution occasions seasons. Different places on the Earth's surface move at different speeds, the speed at the equator being the greatest. The spin greatly influences the shape (e.g., Dixon 1987), distorting the spherical figure to a slightly flattened shape and creating many irregularities on the surface: the polar diameter is about 43 km less than the equatorial one. In what has been called the Colioris Effect, due to the rotation, in the Northern Hemisphere a mass of air around a high pressure area is deflected in a clockwise direction and counterclockwise around a low pressure one: in the Southern Hemisphere there is an opposite effect. A centrifugal acceleration which tends to oppose gravity makes the value of gravity at the equator  $(9.780 \,\mathrm{m \, s^{-2}}) 0.35\%$  less than that of 9.832 m s<sup>-2</sup> at the poles: the maximum gravity  $(10.5 \text{ m s}^{-2})$  is reached at the boundary of the liquid core some 2900 km below the Earth's surface. Near the surface, gravity decreases by about  $0.003 \,\mathrm{m \, s^{-2} \, km^{-1}}$  distance above sea level. While the angular momentum in an elliptical orbit remains constant, according to Kepler's Second Law of Planetary Motion, at periastron (when the radius of the orbit is small) the speed is faster and at apastron (when the radius of the orbit is greater), the speed is slower. The giant outer gas planets, namely Jupiter, Saturn, Uranus, and Neptune (the Jovian planets), have solid cores surrounded by cold atmospheres of light gases such as methane, ammonia, helium, and hydrogen. The Earth, like the other three inner planets, i.e., Mercury, Venus, and Mars (terrestrial planets), is made up of a dense iron-nickel core (some 3400 km in radius at a temperature of 3700 °C), a rocky mantle 2900 km thick, and an outer shell, the crust (lithosphere) some 50 km thick. The light gases were lost from the terrestrial planets due to the fact that: (1) they are closer to the sun and hence received more heat that highly excited the gas molecules and (2) their smaller gravitational forces were not adequate to retain the fast-moving molecules. Compelling an escape velocity of 11.8 km/s, the Earth has been able to hold onto most of its gases except for the very light ones like hydrogen, neon, krypton, and argon. Hydrogen, produced by photodissociation of water vapor in the stratosphere, is presently estimated to be escaping from the Earth's atmosphere at a rate of 3  $\times$  $10^8$  atoms cm<sup>-2</sup> s<sup>-1</sup> (Donahue 1966; Joseph 1967; Hunten 1973; Hunten and Strobel 1974; Hunten and Donahue 1976). A planet or a satellite has to be more than 10% of the mass of the Earth to be able to hold its atmosphere. About two thirds of the Earth's surface is covered with water. The gravitational pulls of the Moon and the Sun cause tides and the movement of the air masses weather. Water (hydrosphere), air (atmosphere), and the superficial layer of the lithosphere (the soil) constitute the biosphere. They are the theaters in which life has experimented and developed.

#### **1.3 Factors that Encouraged the Evolution of Life on Earth**

Whether by default or design, the Earth is peculiarly well conditioned for habitation, at least by the kind of life which we know. As prescribed by nature, organisms adapt and evolve into environments: they do not carve them out to suit themselves. The fundamental factors which "allowed" life to develop on Earth included: (1) an O<sub>2</sub>-rich atmosphere with the moderate level being just appropriate for the respiratory needs for life (Urey 1959), (2) location at a right distance from the sun (orbiting between the terribly hot Venus and the hard frozen Mars) for the temperature to support presence of water in both liquid and vapor form, (3) the accretion of the giant planets like Jupiter, Saturn, Uranus, and Neptune drastically reduced the number of comets and other debris in orbit, minimizing the devastating effects of collision with the Earth (Weidenschilling and Marzari 1996), (4) the presence of atmospheric gases like CO<sub>2</sub> provided a mild greenhouse effect which kept the planet warm, (5) the mass of the Earth (and hence its gravity) is just right to hold and prevent loss of most atmospheric gases to the outer space without undue pressure on life, and (6) the Earth's magnetosphere is adequately strong to prevent sputtering of the atmospheric gases by the constant bombardment of energized ions. Jupiter's largest moon, Ganymede, with a magnetic field about one tenth that of Earth but greater than that of Mercury, Venus, and Earth's moon (Gurnett et al. 1996), has a thin atmosphere (Stevenson 1996) with  $O_2$  in a frozen state (Vidal et al. 1997). Though Mars, with its distant orbit which is 50% farther from the Sun than Earth, now presents a desolate, cold, and dry surface with a small ice cap especially at its north pole, in the past the planet appears to have experienced episodes during which an atmosphere may have existed to create a greenhouse effect adequate to generate ample liquid water on the surface (Kargel and Strom 1996): over time, the planet has lost large quantities of CO<sub>2</sub> as well as O<sub>2</sub> and H<sub>2</sub>, gases derived from breakdown of atmospheric water by sunlight, leaving a thin 7-km-thick gaseous envelope. The planet's atmosphere has been worn out (sputtered) by energetic O<sup>+</sup> ions created from escaping O<sub>2</sub> and hurled back to the atmosphere by the solar wind fields (Johnson and Liu 1996). The highly rarefied Martian atmosphere compares with that of Earth at an altitude of 50km, the atmospheric pressure being less than 1% of that on Earth. It cannot be completely ruled out that under similar or different circumstances a kind of life could have evolved elsewhere in the Universe (e.g., Powell 1993). Possible past occurrence of primitive life on Mars has been advanced (e.g., Kerr 1996b; McKay et al. 1996) and existence of life in other celestial bodies argued (e.g., Chyba 1997).

The Earth's atmosphere weighs about 500 million million tonnes. About 50% of it is in the lower layers about 5km from the Earth's surface. The troposphere, the part of the atmosphere where the air is well mixed, extends up to an altitude of about 10km above sea level and constitutes 80% of the total mass of the atmosphere. Barometric pressure is a consequence of the Earth's gravitational pull on the air which envelopes it. From the surface of the Earth, barometric pressure drops exponentially. However, at a given altitude, the actual pressure depends on factors such as latitude, season, and the prevailing weather conditions (e.g., Bouverot 1985). For every 5.5-km ascent from sea level (where the barometric pressure is 1013 mbar), the barometric pressure drops by a half and the temperature drops at a rate of 1 °C for every 150 m. Beyond 500 km, the atmosphere (exosphere) is highly rarefied and contains free atoms of O<sub>2</sub>, H<sub>2</sub>, and helium. The most important respiratory gases in air are O<sub>2</sub> and CO<sub>2</sub>. At normal pressures, N<sub>2</sub> is considered to be physiologically inert, but at high pressures it is harmful to life.

#### 1.4 Oxygen: a Vital Molecular Resource for Life

The Lord God formed the man from the dust of the ground and breathed into his nostrils the breath of life, and the man became a living being. (Genesis 2:7)

There are few, if any, processes in biology which are as encompassing and critical for life as respiration. For most animals, procuring O<sub>2</sub> from outside and delivering it to the tissues and voiding CO<sub>2</sub> produced from tissue oxidative metabolism are some of the main tasks of respiration. Laitman et al. (1996) assert that "the acquisition and processing of  $O_2$  and its by-products is the primary mission of any air-breathing vertebrate". Just now (as you read this line) you are breathing O2 and would die in a couple of minutes without it. Beyond about 3 min of cessation of breathing, irreparable damage, especially of the central nervous system, could occur even after successful resuscitation, and after about 6 min one would be declared brain dead. From a practical standpoint, this bespeaks the importance and urgency of procuring molecular O<sub>2</sub> at the right time and in the right quantities. In the course of evolution, preference would have been given to the organs and systems which support such crucial process. Though other cells such as the neurons (e.g., Schömig et al. 1987), endothelial cells (e.g., Mertens et al. 1990), and smooth muscle cells (e.g., Paul 1989) can cover energy deficits by anaerobic glycolysis, except for the hearts of the freeze-tolerant animals which stop at subzero temperatures (e.g., Storey and Storey 1986, 1988), the continuous mechanical performance of the myocardial cells is totally dependent on transient changes in cytosolic Ca<sup>2+</sup> (interacting with contractile proteins) and sustained oxidative production of energy in the mitochondria (e.g., Driedzic and Gesser 1994; Piper et al. 1994). While there are assertions that life can exist without  $O_{22}$ such states can only exist in the simplest forms of life (Hochachka et al. 1973; Herreid 1980; Fenchel and Finlay 1990a, 1991a, 1994). Intestinal parasites are alleged to live without molecular O2 and intertidal molluskan facultative anaerobes remain for days without it (Ghiretti 1966). In adverse conditions, adaptively, some animals enter latent (ametabolic) states where in cryptobiosis (Hochachka and Guppy 1987), the most extreme of such conditions, life virtually stops. However, even in such states, an infinitesimal amount of energy must be produced by the cells to sustain the crucial molecular processes of life such as protein turnover and ion flux. Even before the discovery of O<sub>2</sub> was made by Priestley and the composition of air demonstrated by Lavoisier (see Perkins 1964 for an account of these elegant discoveries), it was recognized that breathing, the mechanical pumping of air in and out of the body, a process which occurred in the majority of animals, was essential for life. For a long time and until recently, a test for death was the failure of breathing and the common method of killing was by strangulation. Presently, the phrase "the breath of life" is commonly used to indicate the need for continuous movement of air in and out of the body to support life. The comprehensive need for  $O_2$  for life was termed the call for oxygen by August Krogh (Krogh 1941).

Generation, storage, and utilization of energy are processes central to the activities and the very existence of living cells, just as they are relevant to the proper economic management of the Earth's resources. Excess energy is largely conserved in form of carbohydrate and lipid molecules to be utilized in event of deficit. The acquisition and utilization of energy in life occurs according to Maxwell's Laws of Thermodynamics. According to the first law, the amount of energy in the Universe is fixed: no more of it can be created nor can the existing amount be destroyed but can be converted from one form to another. In face of the steadily decreasing amount of freely usable (accessible) energy in the Universe (according to the Second Law of Thermodynamics), the natural state of matter is chaos (e.g., Prigogine and Stengers 1984). Since living systems constitute highly organized complex states of matter, from a casual glance, it would seem that life runs uphill in a downhill Universe, i.e., it proceeds counter to the natural dissipation of energy. In such a case, life would appear to negate the Second Law of Thermodynamics. This, however, is not the case. In fact, instead of threatening life, the Second Law of Thermodynamics actually guarantees it. Unlike the closed thermostatic state of the ordinary (controlled) chemical reactions, living systems are open thermodynamic processes which access energy from outside (especially from the sun) to steady entropy (chaos) through effervescent repair and rebuilding at the molecular level. Generally, organisms are efficient conduits of energy in the vastness of the Universe. Evolution by natural selection is thought to be driven by competition for the dwindling amount of energy in the Universe (e.g., Blum 1955). Much of the energy on Earth is conserved in the covalent bonds, especially of the large organic molecules such as glucose, fatty acids, starch, and glycogen. Intricate interdependency exists in nature where, directly or indirectly, living things rely on each other in appropriating raw materials and harnessing energy. Over time, evolution has modified morphological design and physiological processes to eliminate or reduce unnecessary expenditure of energy. More optimal states are established to better manage the finite resources. Superfluous structures are eliminated and hence support of unused or underutilized capacities is avoided.

No molecule has been as pervasive in its influence on life and paradoxical in its roles as O<sub>2</sub>. In all evolved complex animal life, O<sub>2</sub> is the most important molecular factor contracted from the ambient milieu. The metabolic rate of an organism correlates with the efficiency of procuring it. Nature has been particularly inventive in the development of gas exchangers and the respiratory processes. The many examples of convergence show that permeative forces have been involved in programming the design of the gas exchangers. Regarding the procurement and utilization of energy, living things are essentially open thermodynamic systems in a self-regulating steady state. A continuous influx and efflux of matter and energy occur as the necessary physiological and behavioral adjustments are made to maintain homeostasis. Such a dynamic state ensures that, though intimately relating to its immediate environment, an organism remains a viable, discrete entity. While life can be defined as a process of capturing and utilizing energy and raw materials, empirically, death is the cessation of all such activities, i.e., when energy production falls to zero. In such a state, the energy in a body is at equilibrium with that in the environment. For individual animals, the continuity of respiration is terminated at death, and for a species at extinction. Animals will generally live for weeks without food, days without water, but only minutes without  $O_2$ . Activities such as feeding, thermoregulation, locomotion, and even reproduction (e.g., Hurst and McVean 1996) can be adjusted, delayed, or abandoned altogether, depending on species and circumstances (McNamara and Houston 1996).

Energy is decisive in all biological events from molecular, biochemical, ecological to evolutionary levels (e.g., Bennett 1988). It is required for building, servicing, and maintaining the general infrastructural integrity of an organism as well as driving the physiological processes and fortifying homeostasis against external perturbations. The rate of respiration indicates the speed at which an animal uses its resources to meet the demands placed on it by the environment and the lifestyle it leads. Those species capable of maintaining a high rate of O<sub>2</sub> to CO<sub>2</sub> exchange ratio in relation to the volume and the complexity of the protoplasmic mass are able to establish stable tissue fluid gas concentrations under different environmental circumstances and metabolic states. It is unequivocally evident from the design of the gas exchangers that such specialized taxa are the most successful. From the earliest recorded fossils, which are 3.8 to 3.5 million years old (Schopf 1978, 1993; Mojzsis et al. 1996), to the first well-documented composite organisms, the ediacaran Metazoa which occurred more than 600 million years ago (e.g., Gould 1989; Knoll 1991; Levinton 1992; Runnegar 1992), for over 80% of its tenure on Earth, life remained exclusively unicellular (Gould 1994) and anaerobic. It is thought that lack of O<sub>2</sub> in most of the Precambrian may have been the main factor which repressed further progress (Knoll 1991): the so-called Cambrian explosion, when the biota underwent remarkable diversity, has been associated with the presence of and the increasing levels of molecular O<sub>2</sub>.

#### 1.5 Anaerobic Metabolism and Adaptive Success in Animals

The capacity to procure, transport, and utilize large amounts of O<sub>2</sub> has bestowed a monumental selective advantage on the evolution and adaptive radiation of the terrestrial vertebrate fauna. Predator avoidance correlates with the level of energy expenditure and the kind of food eaten (e.g., McNab 1966). Terrestrial species with low metabolic rates rely heavily on burrows or passive integumental structures such as shells, plates, and spines for protection. The evolution of efficiently ventilated and perfused gas exchangers and carrier-mediated O<sub>2</sub> transport systems appear to have been fundamental for supporting energetically demanding life-styles. Metabolic rate expresses the integral speed at which energy is mobilized, transformed, and utilized by an organism for biological activities (e.g., Kleiber 1965; Calder 1987; Brown et al. 1993; Lundberg and Persson 1993) and hence expresses the vitality of life (e.g., Zeuthen 1970; Calder 1984). In mammals, factors such as enzymatic activities (Emmett and Hochachka 1981), enzyme contents of tissues (Drabkin 1950), O<sub>2</sub> consumption (R.E. Smith 1956), and protein turnover (Munro and Downie 1964) reflect the effect of body size on metabolism. An inverse correlation between the specific metabolic rate (amount of O<sub>2</sub> consumed per gram body mass per unit time) of animal species and their life spans has been established (e.g., Adelman et al. 1988; Shigenaga et al. 1989). Dwarf mice live much longer than normal ones by as much as 350 days for males and 470 days for females (Brown-Borg et al. 1996).

Thompson (1959, p. 42) forthrightly stated that "size of body is no mere accident". Metabolic rate determines vital aspects such as life patterns, population fluctuations, behavioral ecology, and reproductive efficiency (e.g., Prothero 1986; Calder 1987; DeAngelis et al. 1991; Dunham 1993; McNamara and Houston 1996). In the modern ecosystems, the chance of extinction is directly proportional to body size (e.g., Carroll 1988). Diverse factors such as phylogeny, habitat, ambient temperature, O<sub>2</sub> consumption, food intake, latitude, climate, season, body size, shape, level of development, degree of activity, sex, and age to varying extents determine the metabolic rate (e.g., Zeuthen 1953; Else and Hubert 1985; Crews et al. 1987; Labra and Rosenmann 1994). Unlike metabolic substrates, e.g., carbohydrates and fats, which can be stored in large quantities in the body, except in a few heterothermic and anaerobic parasites (e.g., Ghiretti 1966), O<sub>2</sub> has to be derived from the external environment in the necessary measures. In the human being, about 120001 of air are filtered by the lung everyday (Burri 1985). The amount of O, dissolved in blood or plasma is insufficient for tissue requirements even at rest. The quantity falls far short of the amount which would be required to service physical activity when the uptake may increase by as much as 30 times during vigorous exercise, e.g., flight (Thomas 1987). A human being at rest requires 200 to 250 ml O<sub>2</sub> min<sup>-1</sup> but during maximal exercise the amount increases to about 5.51 min<sup>-1</sup> (Comroe 1974; Weibel et al. 1987a). A 70-kg human being has only 1.551 of O, in the body at any one moment, 370 ml being in the alveolar gas, about 280 in the arterial blood, about 600 in the capillary and venous blood, 60 ml dissolved in body tissues, and 240 ml bound to the muscle myoglobin (Farhi 1964): the total amount is adequate to support life for only 6 minutes, but irreparable damage starts to occur within about 3 min of cessation of breathing. Snyder (1983) observed that the amount of O<sub>2</sub> dissolved in tissue (about 0.8 ml/kg) is sufficient to support aerobic metabolism for only a few seconds. However, in the champion divers, e.g., the Weddell seal, Leptonychotes weddelli (Kooyman 1985), in a 450-kg animal, the  $O_2$  stored in the muscles can support aerobic metabolism at a rate of 4.2 ml O<sub>2</sub> kg<sup>-1</sup> min<sup>-1</sup> for about 15 min. At an estimated O<sub>2</sub> consumption rate of 1.6 ml O<sub>2</sub> per kg, a 20-tonne sperm whale, *Physeter catodon* can dive for 50 min while maintaining aerobic metabolism (Butler 1991a). The body stores of  $CO_2$  in solution and in form of  $HCO_3^-$  ions exceed those of  $O_2$  (Farhi and Rahn 1955). It is perhaps owing to its intrinsically great toxicity (e.g., Fenchel and Finlay 1990b) (Sect. 1.16.1) that animals have not evolved capacities of storing appreciable amounts of  $O_2$  in the body tissues and cavities: the rate of  $O_2$  uptake from the environment is approximately equal to its utilization. The infinitesimal amounts held in the bodies of most organisms, either chemically bound or in solution, are able to support aerobic requirements for only a short period of time. In some fish with physoclistic swim bladders, however, O<sub>2</sub> is held in the swim bladder at high pressures and concentrations (Saunders 1953). Such stores can be utilized during hypoxia (Randall and Daxboeck 1984) with adequate tissue oxygenation being sustained for several hours.

#### 1.6 Evolved Mechanisms and Strategies of Procuring Molecular 0<sub>2</sub>

Respiration has been pivotal in all the evolutionary and adaptive changes which have occurred in animal life. This is evinced by the fact that to a fair measure, the functional competencies of the gas exchangers correspond with the general phylogenetic statuses of animals. Respiration encompasses an impressive arsenal of biomechanical, physiological, and behavioral strategies and mechanisms that are involved in making available to an organism a sample of the external respiratory milieu from which molecular O2 is extracted and into which CO2 is voided. External respiration involves movement of two vectorial quantities in opposite directions, namely, influx of O<sub>2</sub> from the environment and efflux of CO<sub>2</sub> from the organism. Oxygen is delivered to the tissue cells across a panoply of structural compartments through self-regulating convective and diffusive processes. The operation starts with convective delivery of O<sub>2</sub> by a respiratory medium (water and/or air) to the gas-exchanging site, diffusion across the tissue barrier, binding to carrier pigments, convective transport by blood circulation, and ultimately diffusion from the blood into the cells (Figs. 1,2,3). In a steady nonlimiting state, servomechanically, the flow of O<sub>2</sub> from the environment across the steps to the mitochondria is constant (Weibel 1982; Wagner 1993). Mitochondria contain all enzymes associated with the processes of oxidative phosphorylation in their inner and cristae membranes and the enzymes of the TCA cycle in the mitochondrial matrix. The influx of  $O_2$  into the mitochondria, the terminal  $O_2$  sink which determines the flow of O<sub>2</sub> across the lung through the cardiovascular system (e.g.,



Fig. 1. Sites of convective and diffusive gas transfer in an aquatic breather. Water and blood spatially relate in a countercurrent manner at the gills. (After Satchell 1971)



Fig. 2. The intricate stratified arrangement of the gas exchange components in the diffusional pathway of  $O_2$  from the external milieu to the mitochondria. The partial pressure of  $O_2$  gradually decreases towards the tissue cells.  $CO_2$  is greatest in the tissues and is eliminated in the opposite direction to that of  $O_2$ . (After Hughes 1978)



Fig. 3. Cascade process of delivery of  $O_2$  from the ambient milieu in an air breather with a convective lung. The ventilatory and circulatory systems maintain a partial pressure gradient across the air/blood interface. The PO<sub>2</sub> decreases towards the tissue cells and the mitochondria. The utilization of  $O_2$  in the mitochondria maintains the flow. The components of the pathway are quantitatively sized to optimize flow. (Wood and Lenfant 1976)

Suarez 1992), is set by the phosphorylation potential of the tissue cells (Folkow and Neil 1971; Taylor et al. 1987a, 1989) especially in the skeletal muscle mass, which constitutes about 50% of the body mass in mammals and as much as 80% in fish (Goldspink 1985): the PO<sub>2</sub> drops from about 20kPa in water or air to almost zero in the vicinity of the mitochondria of the outlying tissues (Wittenberg and Wittenberg 1987, 1989; Graiger et al. 1995). Between the capillaries of the heart muscle and the mitochondria, the PO<sub>2</sub> drop is 2.7 kPa (Tamura et al. 1989) and that between the cytosol and the mitochondria is less than 0.03kPa (Wittenberg and Wittenberg 1987, 1989; Clark et al. 1987). In a rat (mean body mass 266 g), the total mitochondrial surface area in the liver, kidney, heart, brain, and lung is 460 m<sup>2</sup> and in a 1.8-kg bandicoot the value is 5520 m<sup>2</sup> for the same organs (Else and Hubert 1985). The drop in the PO<sub>2</sub> between the capillaries and the surrounding tissues is inversely proportional to the permeability of the tissue to  $O_2$  (Meng et al. 1992). In exercising human gastrocnemius muscle, the  $O_2$ tension decreases with the intensity of contraction, indicating that  $O_2$  may be a limiting factor for mitochondrial respiration (Fellenius et al. 1984): under such conditions, the PO<sub>2</sub> in the extracellular fluid may decrease by 70% and in the cells by 30% of the resting values. To increase the flow of O<sub>2</sub> from water to the tissues, the heart muscle of the hemoglobinless Antarctic icefish is profusely supplied with blood (Fitch et al. 1984; Johnston and Harrison 1985): the muscle mitochondrial volume density rivals that of the insect flight muscles (Londraville and Sidell 1990) but myoglobin is lacking in significant quantities (Douglas et al. 1985; Feller and Gerdy 1987; Sidell et al. 1987). The complexity of the O<sub>2</sub> conduction line differs between animals. It depends on factors such as body size, environment occupied, and lifestyle. In the Protozoa, O<sub>2</sub> diffuses across the cell membrane into the protoplasm. The simple invertebrates lack a circulatory system while the more complex ones and the vertebrates have a circulatory system which convectively transports O<sub>2</sub> from the respiratory site(s) to the body tissue cells. In insects (Sect. 6.6.1),  $O_2$  is delivered directly to the tissue cells by the trachea. Internal respiration entails utilization of  $O_2$  at cellular level to generate the high energy molecule adenosine triphosphate (ATP), with CO<sub>2</sub> and water as secondary products. Expulsion of  $CO_2$  occurs in the opposite direction to that of  $O_2$ , i.e., from the tissue cells to the gas exchanger, driven by the same mechanisms (Hills 1996) but carried through somewhat different processes (e.g., Davenport 1974; Heisler 1989):  $CO_2$  excretion occurs through a passive process along an electrochemical gradient from the site of production (Bidani and Crandall 1988; Nikinmaa 1990).

Depending on the function(s) they carry, organ systems have different needs for  $O_2$  (Else and Hubert 1983). By default, the gas exchangers are the only organs in which a conflict of interests can occur. For efficient uptake of  $O_2$ , the designs must effect transfer  $O_2$  with minimal utilization of it. Without compromising their functional integrity, as little tissue as possible must be committed in the construction of the gas exchangers. In the avian lung, the bodies of the extremely thin epithelial cells which line the air capillaries are confined to the atria and to a lesser extent in the infundibulae, which are non gas exchange sites (Figs. 88,90). In its thinnest sections, the blood-gas barrier of the avian lung consists of an endothelial, an epithelial, and a common basement membrane (Maina and King 1982a): interstitial spaces are largely lacking (Figs. 29,40). On average, the vertebrate lung is estimated to utilize as much as 10% of the total body  $O_2$  consumption (Slonim and Hamilton 1971).

While O<sub>2</sub> transfer in simple organisms occurs by simple diffusion across the entire body surface, in the more complex ones this takes place at specialized sites where tissue barriers are crossed. In spite of the intrinsic differences in the structural complexities, after about 2 billion years of evolution of aerobic metabolism, the transfer of molecular O<sub>2</sub> in all evolved gas exchangers still occurs by passive diffusion - only ways have changed but means have remained essentially the same. Aerobic metabolism must have evolved at a critical point when the ambient PO<sub>2</sub> was just adequate to drive the gas across the cell membranes of the amphiaerobes. The gradual increase of the PO<sub>2</sub> in the bioshere lead to reduction in the respiratory effort, supporting greater metabolic capacity. The delivery of  $O_2$ to the cells/tissues appears to have been machanistically optimized right from the point of incorporation of molecular O<sub>2</sub> into the aerobic processes. For a substance that is needed continuously throughout life, the alternative method of acquisition, i.e., by active means, as was envisaged to occur in the vertebrate lungs at the turn of this century (see, e.g., Haldane 1922), is improbable. To support respiration driven through active acquisition of O<sub>2</sub> would obligate an enormous investment in energy, rendering the entire process uneconomical and perhaps untenable within the present designs of the gas exchangers and the activities which animals perform. Gas exchangers are largely multifunctional organs. Many play nonrespiratory functions which are in some cases equal to, if not more important than, respiration (Sect. 6.10). Fish and crustacean gills, for example, serve indispensable osmoregulatory roles, are the principal pathway for ammonia and urea excretion, and are involved in the regulation of levels of some blood chemical factors, e.g., hormones (e.g., Zadunaisky 1984; Gilles and Pequeux 1985; C.M. Wood et al. 1989, 1994). The human lungs are important sites for elaboration, metabolism, and regulation of the concentrations of various active pharmacological agents in the body (e.g., Slonim and Hamilton 1971). The lung removes from circulation or destroys such chemical factors as prostaglandins, serotonin, and bradykinin, converts angiotensin I to angiotensin II, and synthesizes lipids such as the pulmonary surfactant (Sect. 6.10): the high  $O_2$  consumption of the vertebrate lung can be attributed to these metabolic processes. While the design requirements for gas exchange demand minimal tissue infrastructure, there must be a critical mass necessary to carry out the nonrespiratory roles. The definitive organization of the gas exchangers must integrate these rather constraining needs. The gas exchangers are unique in that there are no tissue cells which are ubiquitous to them as hepatocytes are to the liver, osteocytes to bone, or neurons to nervous tissue. A cell membrane with no distinct specializations (Figs. 4,5) as in the Protozoa and in many simple Metazoa is the most elementary gas exchanger (e.g., Mangum 1994). The unconventionalized designs of the gas exchangers can be ascribed to the fact that the simple passive process of diffusion of the respiratory gases does not oblige a specific structural plan. Based on the fundamental feature that at a respiratory site all that need exist is an O<sub>2</sub> concen-



Fig. 4A-F. Basic designs of gas exchangers. The least specialized gas exchanger (A) is found in the unicells where the transfer (shown with  $\rightarrow$ ) occurs by diffusion across a plain cell membrane. In the more complex animals, gas exchangers have formed either as evaginations (B) generally called gills and specialized for aquatic breathing, or invaginations (C-F) generally called lungs for aerial breathing. B An example of a unimodal aquatic breather. C A bimodal breather with an "unspecialized" accessory respiratory organ. D A bimodal breather with a "specialized" accessory respiratory organ. E A terrestrial air breather with the lung (an invagination) as the exclusive gas exchanger [in amphibians diffusion across the skin (shown with  $\rightarrow$ ) is an important respiratory pathway in appropriate environments]. F Insect tracheal system where air is delivered directly to the tissue cells across the air sacs, AS, especially in the larger species. (Maina 1994)

tration differential (a partial pressure gradient), exquisite experiments have been used to identify morphologically obscure respiratory sites by use of  $O_2$ -sensitive bioindicators. Suitable flagellates such as *Polytoma* (Thorpe 1932) and a protozoa such as *Bodo* (Fox 1921) have been utilized as markers of areas of rapid  $O_2$  influx in the *Simulium* and larval *Omorgus* (Ichneumonidae), respectively, and in *Cryptochaetum* (Agromyzidae). Terebellid worms (e.g., Weber 1978) and sea anamones, e.g., *Metridium senile* (Sassaman and Mangum 1972; Schick 1991) present some localized areas of the body where the thickness of the skin is drastically reduced. In the large scyphozoan, *Cyanea capillata*, and the tube anemone, *Ceriantheopsis americanus*, it is still debatable whether the skin over the body column plays any significant role in respiration (Sassaman and Mangum 1974; Mangum 1994). Up to now, gas exchangers still remain to be discovered!

animal body ambient medium	examples and remarks
integuments	eventually the surrounding medium may be air
gill	crustaceans fishes molluscs tadpoles also-book gill of limulus -podia of urchins
water lung	holothurians (sea cucumber) also – buccopharyngeal respiration – cloacal respiration
trachea	spiders insects also-book lung of scorpions tracheal lung of spiders
tracheal gill	aquatic larvae of insects eg mayflies
compressible gas gill	aquatic insects e.g. <u>Notonecta</u>
incompressible gas gill	aquatic insects e.g. <u>Aphelocheirus</u> syn.: plastron
air air lung	<ul> <li>vascularized cavities (land snail)</li> <li>air chamber of some air breathing fish</li> <li>simple air sac (sphenodon)</li> <li>alveolar lungs (reptiles and mammals)</li> <li>parabronchial lung of birds</li> </ul>

Fig. 5. Scheme showing the different anatomical designs of gas exchangers in water and air breathers and specialized modes of gas exchange. The organization of gas exchangers is mainly determined by the kind of respiratory medium utilized, habitat occupied, lifestyle, and the evolutionary level of development of a particular organism. (Dejours 1988)
#### 1.7 Explicating the Process of Evolution of Respiration: Limitations

Reconstructing the pathways and the stages through which gas exchangers have evolved is an undertaking beset with many difficulties. The main obstacle lies in the uncertainty of our grasping the nature, the severity, and the direction of the changes which occurred in what was a highly dynamic biosphere of the ancient Earth. There is a particularly glaring lack of fossilized materials of the gas exchangers except on one Devonian species, *Bothriolepis* (e.g., Denison 1941; Wells and Dorr 1985). Soft tissues are very seldom adequately geologically preserved. When it occurs, however, by their very nature of being delicate, the materials are reduced from their three-dimensional form to two-dimensional films through intense heat and compression (e.g., Behrensmeyer and Kidwell 1985), making their recognition and interpretation very difficult and unreliable.

Air breathing has only evolved in the lineages of the two osteichthvan (bony) fishes, the actinopterygian (ray-finned fishes), e.g., bichirs, gar, and bowfin and sarcopterygian (lobe-finned fishes), e.g., lungfishes (Romer 1972; Pough et al. 1989; Cloutier and Forey 1991). Though a very distant point in evolution, this, nevertheless, provides a focal point for seeking convergence and divergence of the animal groups from which air breathers evolved. The near total extinction of the crossopterygian fishes (the sole survivor being the coelacanth, Latimeria chalumnae), a group thought to be a direct progenitor of the tetrapods (e.g., Pough et al. 1989; Gorr et al. 1991), makes the discernment of the evolution of the respiratory processes much harder. More often than not, one has to more or less rely on circumstantial evidence. From molecular genetic studies, it has now been proved that the popular phrase that "ontogeny recapitulates phylogeny" is too simplistic for developmental biology (on its own) to be reliably meaningful in reconstructing phylogenies (e.g., Humburger 1980; Alberch 1985; De Queiroz 1985; Gans 1985; Northcutt 1990; Marshall and Schultze 1992). Neotenv and/or pedomorphy (e.g., Semlitsch and Wilbur 1989; Wake and Roth 1989) are but two characteristics which manifest nonconforming developments. Morphological, experimental embryological, physiological, biomechanical, paleontological, molecular, and biochemical investigations need to be integrated to effectively elucidate evolutionary transformations, mechanisms, and pathways. In amphibians in particular, dramatic changes in form, location, and function of the gas exchangers and the circulatory system occur during metamorphosis (e.g., Infantino et al. 1988; Hou and Burggren 1991; Burggren and Bemis 1992; Newman 1992; Burggren and Infantino 1994; Fig. 47). These entail radical modifications of the ventilatory mechanisms and transformations of the respiratory organs, changes which must be accompanied by appropriate neurophysiological reorganizations for proper motor functional control and coordination (Burggren et al. 1990).

Albeit the glaring lack of data, the evolutionary and adaptive developments of the gas exchangers can be patched together by collateral evidence gathered by studies of: (1) animals which have adapted to atypical habitats, (2) those that possess transitional respiratory features, (3) those at different phylogenetic levels of development, (4) those which have unique behavioral lifestyles, and (5) those which show peculiar developmental changes. Study of the ancient air-breathing fish such as the Dipnoi, Holostei, and Polypterida should provide a fertile ground for such inferential studies. For biologists, there is always some unique kind of satisfaction when a previously unknown phenomenon or state of natural history fits into a theoretical prediction. The excitement of discovering the so-called living fossils, rare animals which fill the evolutionary gaps, is great. The capture of the Carboniferous actinistian, the coelacanth Latimeria chalumnae in 1938 (see R.E. Smith 1956; Thomson 1986; Fricke 1988; Cloutier and Forey 1991; Bruton et al. 1992), a group thought to be long extinct, is a classical instance. It is now, however, profitable to recognize that since the first successful engineering of transgenic mice by Gordon et al. (1980), it is no longer necessary to formulate a scientific question to suit a biological system. These days, it is possible to specifically design an organism (e.g., a transgenic organism) to best answer a particular question (e.g., Cory and Adams 1988; Adams and Cory 1991; Taketo et al. 1991; Ho 1994). Such new experimental paradigms, which hitherto were not possible to configure, provide useful tools for analytical manipulations at molecular, organismal, and ecological levels. When adopted, they should contribute greatly in the advancement of comparative respiratory biology.

The diversity of the organization of the gas exchangers was highlighted in Maina (1994). In this account, the essence of the contrast is discussed with particular focus on: (1) the conditions under which the respiratory processes evolved, (2) the physical characteristics of the media in which these changes occurred, and (3) the different strategies which animals adopted to extract  $O_2$  from the external milieus. The simplest respiratory organs are generally found in the aquatic animals or those organisms which subsist in cryptozoic (humidic) habitats. In their rudimentary form, they occur in the form of permeable, moist, well-vascularized surface membranes, e.g., the integument of the invertebrates such as the earthworms and planaria and in vertebrates such as the eel and the frog, e.g., *Rana*. At the more advanced stages, cardiorespiratory coupling developed to enhance the transfer of  $O_2$  to the complex highly aerobic tissues. In those aquatic animals where the integument is the main respiratory pathway and in fish which have lost the bucco-pharyngeal ventilatory capacity of the gills (e.g., mackerel), locomotion provides an important respiratory activity.

#### 1.8 Plans and Performance Measures of the Gas Exchangers

Over and above the simple diffusive respiration of the unicellular organisms and the lower invertebrates, the gills and lungs are distinctively suited for respiration in water and air, respectively. Owing mainly to the remarkable differences between the two fluid respiratory media (Sect. 3.2), on very rare occasions the two organs are contrived to operate in both respiratory media with equal efficiency. With the progressive organizational complexity of animals, forms, and processes such as closed circulation, double circulation, convective movement of the respiratory media, and presence of respiratory pigments in the body fluids and erythrocytes evolved to match the intensifying demands. Such transformations invite interpretations and speculations. Compared with the more recent enhancements, the ancient elements of the respiratory improvements would have been optimized and conserved in due course. Both the physical characteristics of the respiratory media and the respiratory needs have determined the definitive functional designs of the gas exchangers. The various schemes of the respiratory systems, however, are not congruous with the classic concept of Darwinian radiative animal evolution, which is artistically presented as a branching tree with birds and mammals sitting somewhere at the top. Unlike the brain, which shows progressive development reaching the pinnacle in the human being, the gas exchangers of mammals do not present the ultimate pulmonary design. The structural and functional attributes of a gas exchanger cannot be easily predicted based on any single phylogenetic factor in a simple and direct way.

The importance of O<sub>2</sub> in the survival of organisms is reflected in the dramatic effect that hypoxia and hyperoxia have on the structure and function of the gas exchangers. Parameters such as blood O<sub>2</sub> carrying capacity, O<sub>2</sub> affinity, and myoglobin concentration in tissues can change within a matter of hours in response to aspects such as sojourn at high altitude or after being subjected to severe exercise. In the respiratory system, the working capacities at all steps, be they convective or diffusive, must be appropriately sized and regulated for optimal function. Decrease in size and increase in activity calls for more elaborate gas exchangers and more efficient means of O2 uptake and transport. Amidist these permutations, certain conflicts, compromises, and tradeoffs occur. For example, whereas intucking of the gas exchangers was essential for avoidance of desiccation on land, affording better protection against trauma, and achieving a more extensive respiratory surface area (Figs. 4,5), such organs could only be ventilated tidally, a pattern functionally inferior to the continuous unidirectional process which is possible in the evaginated gas exchangers (Figs. 6,18). In the erythrocytes, the main organic phosphate 2,3-diphosphoglycerate (2,3-DPG) and CO<sub>2</sub> combine with the same basic groups of the hemoglobin competing with each other (Davenport 1974). The effects of 2,3-DPG and CO<sub>2</sub> on the hemoglobin dissociation curve are not additive: the shift brought about by the two factors together is less than the sum of each separately. The avian trachea presents a good example illustrating the nature and extents of the compromises and structural adjustments effected to enhance the efficiency of the gas exchangers. To attain flight, birds totally committed the forelimbs for this singular function. Ipso facto (i.e., to substitute for the roles which the forelimbs played), birds evolved a long neck (and with it a long trachea) for defense, procuring food, construction of the nests, and preening. For animals of the same body mass, the avian trachea is three times longer than that of a mammal (Hinds and Calder 1971). In order to compensate for what may have led to a greater resistance to air flow (as resistance to air flow in a tube is directly proportional to the length but inversely proportional to the radius to the fourth power in laminar flow - and to the 4.75 power if the flow is turbulent), the avian trachea acquired a diameter 1.3 times greater than that of mammals (Hinds and Calder 1971): air flow in the trachea of the ostrich has been shown to be laminar (Schmidt-Nielsen et al. 1969). The net effect of these adjustments, i.e., increase in the diameter and the length of the trachea in birds, ensued in an overall resistance similar to that of the trachea of a mammal of the same size. In gaining a large tracheal volume and hence a large tracheal dead space (TDS) which is 4.5 times greater than in a mammal (Hinds and Calder 1971), it could



**Fig. 6a,b.** Spatial arrangement of the respiratory media in an invaginated gas exchanger (a) and an evaginated one (b). Except for the highly specialized bird lung, the gas exchange zone of the invaginated gas exchangers is ventilated tidally ( $\leftrightarrows$ , a) while the evaginated ones are ventilated unidirectionally ( $\rightarrow$ , b). *s* Secondary lamellae. In the fish gills (b), the direction of the flow of the blood in the secondary lamellae ( $\succ$ ) runs counter to that of the water in the interlamellar space ( $\rightarrow$ ). (Kylstra 1968)

conceivably be concluded that natural selection imparted an impediment on the function of the avian respiratory system. This, however, is not the case. The lower respiratory frequency (RF) of birds, which is 0.32 to 0.42 times that of a mammal of equal size, countered the limitations caused by the large TDS. Moreover, the lower RF has afforded room for remarkable configurations of the trachea (e.g., Forbes 1882). Extreme trachea lengths occur in birds such as the trumpet bird, *Phonygammus keraudrenii*, the magpie goose, *Anseranas semipalmata*, and the whooping crane, *Grus americana* (Clench 1978; McLelland 1989). Although

*P. keraudrenii* has the same body mass as a common flicker (*Colaptes auratus*), which has a trachea only 38 mm long, its trachea may be over 800 mm in length and compares with that of the much larger ostrich, *Struthio camelus* (Clench 1978). In *G. americana*, the overall tracheal length is 1.5 m (Welty 1979). Tracheal coiling has been taken to be an acoustic adaptation for lowering the pitch or amplifying sound (e.g., Greenewalt 1968; Gaunt et al. 1987). Furthermore, tracheal loops have been said to increase the tracheal respiratory surface area enhancing evaporative water loss during panting (e.g., Prange et al. 1985) without running a risk of respiratory alkalosis (Schmidt-Nielsen et al. 1969; Bech and Johansen 1980). Compared with birds with straight trachea, those with tracheal convolutions or other tracheal prolongations adaptively have relatively wider tracheal diameters (Hinds and Calder 1971). Perhaps to play similar roles, trachea diverticula have been reported in snakes (Young 1992).

Miscellaneous tissues and organs such as the cell membrane, skin, buccal cavity, gastrointestinal tract, gills, and lungs variably serve as respiratory sites. Because they characterize the more phylogenetically advanced animals, the airbreathing organs (lungs) are assumed to be the better (more efficient) gas exchangers. Except in the bimodal breathers (Chap. 5), gas exchangers are refined to operate best in only one respiratory medium. The human being at the epitome of evolution soon succumbs when the lungs are flooded with liquid (Sect. 6.11). In all respiratory organs, be they water or air breathing,  $O_2$  dissolves in a thin film of water as it traverses the tissue barrier (Sect. 6.1). The flux of the respiratory gases occurs under the prevailing partial pressure gradients across the water or airblood barrier and is maintained by utilization  $(O_2)$  and production  $(CO_2)$  in the tissue cells and the physical movements of the external and internal respiratory media. There has been protracted debate as to whether the diffusion of  $O_2$  across the cell membrane is entirely passive or is facilitated (e.g., Longmuir and Bourke 1959; Scholander 1960; Burns and Gurtner 1973; Wittenberg 1976). Hemoglobin, myoglobin, and a specific carrier (cytochrome  $P_{450}$ ) have been implicated in facilitated diffusion of  $O_2$  in tissues such as the lung, placenta, and the liver (Kreuzer 1970; Wittenberg and Wittenberg 1989). The significance of facilitated diffusion of  $O_2$  in tissues is not well known. The process may, however, be consequential in states of reduced O<sub>2</sub> flow across the blood-gas barrier, e.g., in cases of interstitial edema (Burns et al. 1975, 1976), and in hypoxic conditions (Longmuir 1976).

In both aquatic and terrestrial animals, the complexity of the gas exchangers correlates with the mode of life, habitat occupied, environment, and the general metabolic capacities (e.g., Hughes and Morgan 1973; Gehr et al. 1981; Maina et al. 1989a; Hughes 1995). In nature, the high metabolic needs of the endotherms have not been satisfied except by air breathing. The diffusion of  $O_2$  occurs at a rate of  $2.3 \times 10^{-5}$  cm<sup>2</sup> s<sup>-1</sup> (Grote 1967) across an extremely thin, expansive blood-gas barrier. The process is completed within 250 to 500 ms (West 1974). For the typical  $O_2$  uptake of 200 ml per min, a concentration gradient of only 0.057 kPa (a value which is negligible compared with the prevailing inspired air-arterial blood PO<sub>2</sub> gradient of about 6.7 kPa in the mammalian lung) is all that is necessary. The requisite structural and functional attributes of an efficient respiratory organ are an extensive surface area, a thin partitioning between the respiratory media, and efficient ventilatory and perfusive mechanisms to maintain the highest possible

pressure gradient across the barrier. In simple animals, the extensive surface area (per unit body volume) is more than adequate for gas transfer, while in the larger ones respiration is restricted to specialized sites. Such areas are brought about by outfolding (evagination) or intucking (invagination = cavitation = sacculation) of a part of the body surface (Figs. 4,5). The organs in the first category have been termed gills and those in the second lungs. The gills (Chap. 4) are the archetype aquatic respiratory organs while the lungs (Chap. 6) are the model ones for air breathing: the bimodal (transitional) breathers (Chap. 5) have evolved organs which are used to extract molecular O<sub>2</sub> from both water and air. In the multicellular organisms, the consequential features which must be presented either singly or in combination for an organ to be designated a gas exchanger include: (1) movement of the external respiratory medium, (2) the PO, must be lower and the CO<sub>2</sub> higher in the effluent respiratory medium than in the influent one (e.g., Qasim et al. 1960), and (3) perceptible structural modifications such as infolding or outfolding and internal subdivision of the respiratory surface. Vestiges of lungs which are used for water breathing and a number of gills modified for air breathing exist, but these are rare.

## 1.9 The Early Anoxic Earth and the Evolution of Life

Of all concepts which have been enunciated in biology, that of evolution is probably the most important and encompassing. Dobzhansky (1973) declares that "nothing in biology makes sense, except in right of evolution". Wainright (1988) asserts that "evolution is the single most important and inclusive concept in biology". Its practical utility in biology is summed up by Nelson (1978) as follows: "the concept of evolution is an extrapolation, or an interpretation, of the order liness of ontogeny". Although debate still continues even on the validity of the concept itself and the mechanisms through which it occurs, no other plausible principle can: (1) satisfactorily organize and explain the diversity of the existing life forms, (2) account for and align the preserved fossils and the extinct forms with the extant species, and (3) explain in the context of the contemporary species the paintings and sculptures made within recorded history by the early human beings. Fossils bespeak terminated (failed?) experiments in evolution. Living things have a shared biology. The theory of evolution is grounded on the fundamental belief that life has a common origin (e.g., Brown and Doolittle 1995): through natural selection, animals and plants have progressively developed and genomically diverged in the continuum of time. However unpalatable it may sound, from congruent evidence derived from multiple proteins (Baldauf and Palmer 1993), animals and fungi are sister groups with plants constituting a more distant evolutionary lineage! Cladistic classifications attempt to reconstruct the evolutionary histories and establish relationships between different taxa from study of states of shared derived characters (e.g., Benton 1995; Huelsenbeck and Rannala 1997). Molecular genetic sequences form the basis of many modern phylogenetic reconstructions (e.g., Stewart et al. 1987; Dean and Golding 1997). To delve into the origin of life and understand the subsequent inputs and changes

which culminated in the formation of the modern ecosystems for which utilization of molecular  $O_2$  was central, different scientific disciplines like biology, astronomy, atmospheric physics, geophysics, astrophysics, geochemistry, inorganic and organic chemistry, oceanography, and geology should be integrated. An interdisplinary approach better illumines the convergence of experimental and analytical data, connecting events across temporal and spatial scales. Scientific disciplines gradually diffuse into each other. The often aggressively defended boundaries are more apparent than real. They are often created for self-interests and preservation and have profoundly hindered advances in ratiocinative thought.

The age of the Universe is estimated at between 10 to 20 billion years (e.g., Schopf 1980; Peebles et al. 1994). For a long time after its accretion, some 4.5 billion years ago, the Earth was in a state of perpetual physical and chemical turbulence (Schopf 1978, 1993; Mojzsis et al. 1996). The surface temperature was in excess of 1500 K and the high pressure primary atmosphere consisted of water vapor (~8  $\times$  10<sup>22</sup> mol), CO<sub>2</sub> (~5  $\times$  10<sup>21</sup> mol), N<sub>2</sub> (~3  $\times$  10<sup>20</sup> mol), H<sub>2</sub>S  $(\sim 9 \times 10^{20} \text{ mol})$ , and SO<sub>2</sub>  $(\sim 7 \times 10^{19} \text{ mol})$  (Matsui and Abe 1986). With the cooling to below 650 K, the water vapor condensed, forming highly acidic primitive oceans. The minerals in the lithosphere soon neutralized the acids and the dissolved SO<sub>2</sub> formed sulfates and sulfides. Through outgassing (Allegre and Schneider 1994), the secondary atmosphere came to comprise mainly  $CO_2$ ,  $N_2$ , water vapor, and traces of CH<sub>4</sub>, NH<sub>3</sub>, and SO<sub>2</sub>. This composed the incipient neutral atmosphere which was essentially similar to the present one of Venus and Mars. Subsequently, the H<sub>2</sub>O vapor was photochemically dissociated into H<sub>2</sub> and H<sup>+</sup>, converting the secondary atmosphere into a reducing one. Some of the other most important changes to have occurred during the evolution of life on Earth have been: (1) variations in temperature and solar insolation (e.g., Foley et al. 1994; D'Hondt and Arthur 1997), (2) changes in the orbit (Imbrie et al. 1989), (3) plate tectonics (e.g., Raymo and Ruddiman 1992), (4) variations in the gaseous composition of the atmosphere (from a neutral, i.e., one where  $CO_2$  and  $N_2$ predominated to a highly reducing one where H<sub>2</sub> was the principal gas and finally an oxidizing one – with accretion of  $O_2$ ) (e.g., Tappan 1974; Chappellaz et al. 1992), (5) decrease in the rate of rotation (e.g., Scrutton 1978), and (6) small fluctuations of the atmospheric pressure (e.g., Hinton 1971) and gravitational forces (e.g., Carey 1976; McElhinny et al. 1978). It is widely postulated that the chemical evolution of life occurred by combination and transformation of a vast range of simple inorganic molecules such as carbon monoxide, CO<sub>2</sub>, N<sub>2</sub>, H<sub>2</sub>, and H<sub>2</sub>O into complex biologically relevant organic compounds. This process is envisaged as having been induced by enormous energy influx probably from solar radiation, heat, meteorite impact events, radioactive decay, electrical discharges, and thunder shock waves (Calvin 1956). Though organic molecules themselves may have been extraterrestrial in origin (e.g., Cohen 1995), life is thought to have been fabricated in a chemically reducing atmosphere (e.g., Chang et al. 1983; Cloud 1983a,b; Jenkins 1991), probably around geothermal springs (e.g., Stong 1979) or on the surface of catalytic iron sulfide crystals (Russell and Daniel 1992; Kaschke and Russell 1994; Russell et al. 1994). Through long intricate condensation, polymerization and oxidation-reduction reactions of organic molecules such as amino acids, sugars, and other suitable molecules (the so-called primordial broth, or organic soup), the high-energy phosphate bonds (for intracellular energy transfer), specificity of protein molecules as organic catalysts, genetic coding of the nucleotides, and membrane ionic transfer processes developed (e.g., Bar-Nun and Shaviv 1975). Biogenesis of self-repairing, self-constructing, highly dynamic molecules resulted in the first living entity called protobiont by Oparin (1953) and concept organism by Chapman and Ragan (1980). This microscopic unit is the simplest ancestral prokaryote which possessed the most basic requisites for life. Organic evolution had to await the development of genetic and protein-synthesizing pathways. Interestingly, Lee et al. (1996) have described a self-replicating peptide. Organic molecules like amino acids, protein-like polymers, and nucleic acid polymers have been synthesized in the laboratory by passing an electric are through a mixture of gases such as  $CH_4$ ,  $NH_3$ ,  $H_2$ , and  $H_2O$ vapor, i.e., by simulating what are thought to have been the atmospheric conditions and circumstances which existed in the primeval past (see e.g., Sagan 1994).

#### 1.10 Abundance of Molecular O<sub>2</sub> in the Earth's Biosphere

Of the nine solar system planets (eight – should Pluto finally turn out to be merely a piece space junk as recently suggested!), only the Earth has a veritable atmosphere suitable for life. While the atmosphere of Earth contains only tiny amounts of CO<sub>2</sub>, those of Venus and Mars contain 96.5 and 98% CO<sub>2</sub>, respectively. The atmospheres of Jupiter and Saturn are composed essentially of H<sub>2</sub> and helium. Mercury strictly lacks an atmosphere. The present atmosphere of Earth compares with that of Mars some 300 to 400 million years ago (Kargel and Strom 1996). Saturn's giant moon, Titan, has an atmosphere ten times larger than that of Earth and a surface pressure of 1.5 atm (Samuelson et al. 1981; Lorenz et al. 1997). The atmosphere is made up predominantly of molecular N<sub>2</sub> (82.2%), Ar (11.6%), 6%  $CH_4$ , and 0.2% H<sub>2</sub>. Like Earth, Titan has a greenhouse effect (McKay et al. 1991; Lorenz et al. 1997). The greenhouse effect on Venus, which is caused by  $CO_{2}$ , generates surface temperatures of around 455 °C. Life evolved on Earth nearly 4 billion years ago (Schopf 1980; Balter 1996). The first obligatory aerobic eukaryotic cells appeared between 2.0 to 1.5 billion years ago (e.g., Reader 1986; Schopf and Walter 1983) and the first multicellular organisms about 600 million years ago, i.e., at the beginning of the Cambrian period (Nursall 1959; Cloud 1983a). The tenure of life on Earth constitutes only about 15% of biogeologic history. The complex organisms have existed for an even shorter period, i.e., about 5% of it (Figs. 7,8).

#### 1.11 Shift from Anaerobiotic to Aerobiotic State in the Early Earth

The oxidative state of the Earth's biosphere has corresponded with the measure of the sources of  $O_2$  and the abundance of inorganic and organic reduced com-



**Fig. 7.** Scheme showing the relatively very short period over which terrestrial and in general airbreathing organisms have lived on Earth. The dates (in *parentheses*) are averages from different publications

pounds. The appearance of an oxygenic environment entailed a change from an inefficient to a more advanced energetically O2-dependent and metabolically highly versatile oxidizing ecosystems. Anaerobic fermentation is a highly inefficient source of energy as much of it is left locked in molecules such as alcohols and organic acids, end-products which must be removed before they accumulate to toxic levels. Fermentation of a molecule of glucose results in production of only 2 molecules of adenosine triphosphate (ATP) which contain only about 15 kcal of available energy, while in aerobic respiration 36 molecules of ATP equivalent to 263 kilocalories of utilizable energy are produced. Stated differently, to obtain the same amount of energy through fermentative respiration, a greater quantity of carbohydrate molecules must be utilized. The Pasteur effect, named in honor of Louis Pasteur (1822 to 1895), who first described the fermentation of yeast independent of O<sub>2</sub>, has been defined in the broadest terms as "the stimulation of carbohydrate consumption by reduced  $O_2$  tension" (Schmidt and Kamp 1996). The products of aerobic respiration are water and CO<sub>2</sub>, two innocuous molecules which are easily eliminated into any environment at minimal risk and cost. While the energy derived from fermentative processes is just adequate to support life, aerobic respiration provided excess energy, which organisms invested towards attaining greater structural and functional complexity, resulting in greater success compared with their predecessors. Without O2, life would probably not have developed above that of the unicellular fermentative prokaryotes.

# Evolution of the earth's earliest biosphere and biota

Biologic and Geologic



Fig. 8. Summary of the major biotic developments during the geological periods. (Schopf et al. 1983)

Oxygenic biochemistry has evolved regularly in the past in response to the changing levels of molecular  $O_2$  in the biosphere (e.g., Fox et al. 1980). These conditions have changed from neutral to reducing state and varied from anoxia, hypoxia, and hyperoxia (relative to the present) by no means chronologically in that order (e.g., Frakes 1979; Hendry 1993; Fig. 9). Changes in atmospheric  $O_2$  have paralleled biotic developments (Figs. 8,9). Until about 1 to 2 billion years ago, the atmosphere consisted essentially of carbon monoxide,  $NH_3$ ,  $CH_4$ ,  $H_2$ ,  $H_2$ O vapor, and other simple hydrocarbons (Schopf 1978, 1983; Owens et al. 1979; Chapman and Schopf 1983; Grieshaber et al. 1994). Chemical evolution of life occurred within such a reducing atmosphere after the surface of the Earth had cooled to a level compatible with synthesis of the labile organic molecules (Chang et al. 1983). It has been argued by, among others, Oparin (1938) that the present conditions on Earth are no longer suitable for compounding life from inorganic



Fig. 9. Changes in the levels of  $O_2$  and  $CO_2$  in the late Paleozoic. Oxygen levels have fluctuated from a low of about 15% (reached towards the end of the Paleozoic, i.e., 250 million years ago) peaking at 35% by the late Carboniferous (268 mya). The present atmospheric level (PAL) of 21% is shown with a *dotted line*.  $CO_2$  level was highest in the Ordovician-Silurian, dropped remarkably during the Devonian-Carboniferous, and increased in the late Permian. The PAL of  $CO_2$  of about 0.036% is shown with a *dotted line*. Note that the relative levels of the two gases fluctuate in an inverse manner. *C* Cambrian; *O* Ordovician; *S* Silurian; *D* Devonian; *C* Carboniferous; *P* Permian; *Tr* Triassic; *J* Jurassic; *K* Cretaceous; *T* Tertiary. (After Graham et al. 1995; reprinted by permission from *Nature*, Vol. 375, pp. 117–120, copyright 1995 Macmillan Magazines Ltd.)

matter because the atmosphere is too highly oxidizing. Anaerobic microorganisms flourished in water in excess of 500 million years before  $O_2$  production started (Fenchel and Finlay 1994). At the middle of the Precambrian era, a group of prokaryotes, the cyanobacteria (blue-green algae), evolved chlorophyll a, acquiring means of utilizing solar energy for the process of photosynthesis (e.g., Owens et al. 1979; DiMagno et al. 1995; Nisbet et al. 1995; Boussaad et al. 1997). Photosynthesis is the one large-scale process that abundantly converts simple inorganic compounds (CO<sub>2</sub>, H<sub>2</sub>O, and tiny amounts of minerals) into complex energy-rich organic carbohydrate (CH<sub>2</sub>O) molecules. It is the source of all living matter on Earth and in that case all biological energy (Rabinowitch and Govindjee 1965). It is interesting to note that the discovery of flourishing hydrothermal vent communities in the 1970s (e.g., Meredith 1985) demonstrated that life could exist on Earth totally independent of solar radiation. The accumulation of  $O_2$  (a product of the photosynthetic process) in the atmosphere resulted in the transformation of the Earth's nascent biotic ecosystems from an anaerobic to an aerobic state. This initiated decisive biological changes (Table 1). For successful progression of life from water to land, due to the harmful effects of the UV light, the presence of  $O_2$ , which generated a protective ozone layer in the atmosphere, was necessary. Compared with the present state, the solar UV light flux was more fierce and perhaps invariably lethal during the first 500 million years of the Earth's evolution (Gaustad and Vogel 1982).

Oxygen enrichment of the atmosphere resulted in an increase from 3 to 10% up to 100% of the present atmospheric level in the late Proterozoic and early Cambrian epochs (about 0.54 billion years ago) (Des Marais et al. 1992; Canfield and Teske 1996). It is believed that this led to the so-called Cambrian explosion, an event that was characterized by dramatic biotic developments which included: (1)

	21% O <sub>2</sub> present	35% O <sub>2</sub> 285 mya	15% O <sub>2</sub> 250 mya	
Respiratory gases				Biological significance
Oxygen				
O <sub>2</sub> partial pressure (kPa)	21.2	35.3	15.1	Respiration, lignin biosynthesis
Krogh's maximum radius (cm)	0.11	0.14	0.09	Size limit for diffusion dependence
Water $O_2$ content (ml <sup>-1</sup> )	6.9	7.4	4.9	Aquatic respiration
Carbon dioxide				
Carbon dioxide partial pressure (kPa)	0.03	0.03	0.09	Effects on photosynthesis, moisture content and
Water $CO_2$ content (ml <sup>-1</sup> )	0.31	0.31	0.31	Aquatic pH effects, acid- base balance and ion regulation
Air properties				-
Density $(kg m^{-3})$	1.29	1.56	1.12	Flight and respiratory mechanics, wind shear
Dynamic viscosity $(kgm^{-1}s^{-1})$	$18.2  imes 10^{-6}$	+	-	Boundary layer thickness
Specific heat (js <sup>-1</sup> deg <sup>-1</sup> )	1.006	+	_	Heat capacity and relative humidity
Thermal conductivity $(js^{-1}m^{-1}deg^{-1})$	$2.4 \times 10^{-2}$	+	_	Earth thermal budget, climate

**Table 1.** Comparison of physical properties of the present  $O_2$  atmosphere (21%  $O_2$ ) with those of the relatively hyperoxic late Carboniferous (35%  $O_2$ ) and relatively hypoxic end-Permian (15%  $O_2$ ). (Graham et al. 1995)

+ and – indicate increase and decrease, respectively, relative to the present 21% O<sub>2</sub> atmosphere; mya, million years ago.

the evolution of the multicellular life (Conway-Morris 1993), (2) synthesis of the structural protein collagen which is widely distributed in the metazoans (Towe 1970), and (3) remarkable adaptive radiation and ecological diversification of the animal life (Conway-Morris 1993; Canfield and Teske 1996; Knoll 1996). In general, episodes of rapid evolutionary change correspond with occurrences of speciation (e.g., Gould and Eldridge 1977; Stanley 1979). In water, the surge in the O<sub>2</sub> level accelerated the biodegradation of the dissolved iron and the organic (bacterial and algal) matter. Precipitation of the resultant complexes to the bottom increased the level of oxygenation of the surface waters (Logan et al. 1995), making them more habitable. Without the ancient cyanobacteria, the Earth would still be having little, if any, reactive molecular O2: like the atmospheres of Mars and Venus, CO<sub>2</sub> would still be the predominant atmospheric gas. In Rhodobacter sphaeroides, a metabolically versatile photosynthetic bacterium able to operate under a wide variety of environmental states, a decrease in O<sub>2</sub> availability leads to induction of the membranous photosynthetic apparatus (Yeliseev et al. 1997): the expression of gene-encoding components of the photosynthetic complexes, e.g., structural polypeptides, bacteriochlorophyll, and carotenoids, is closely directed by O<sub>2</sub> tension and light intensity. The momentous point at the end of the Early Proterozoic (some 2.0 to 1.5 billion years ago) (e.g., Kasting and Walker 1981), when the Earth changed from a mainly anoxic hydrosphere and atmosphere to an oxic one is marked by the time at which: (1) the production of banded iron abruptly stopped, (2) deposition of the highly oxidizable uraninite stopped, and (3) the first occurrence of blue-green algae of which the cells included thick-walled heterocysts which may have shielded the O<sub>2</sub>-sensitive nitrogenase enzymes, as the modern ones do. It was not until after the oxidization of the reducing gases and mineralogic factors, when photosynthetic O<sub>2</sub> discharge into the biosphere finally exceeded the turnover rate of the reduced matter, that O<sub>2</sub> became a vital and permanent factor in a stable aerobic atmosphere. The transition from reducing to oxidizing oceans and atmosphere may have been accelerated by a declining discharge of reducing gases and oxidizable substrates through less tectonic activity (Walker 1978). Depending on ecological settings, the transitional point may have differed profoundly in different parts of the Earth.

The extensive invasion of land by plants during the Devonian enhanced the rate of production of  $O_2$ . This shifted the base of photosynthesis from water to land (McLean 1978; Knoll 1979, 1991) with the productivity of  $O_2$  on land exceeding that of the oceans by a factor of 2 (Holland 1978). Practically all the molecular  $O_2$  which was produced during the Earth's history, much of which is now held in diverse organic sinks, arose from green plant type and blue-green algal cyanobacterial photosynthesis (van Valen 1971; Walker 1974). The  $O_2$  we respire today was de facto "excreted" by the cyanobacteria some 2 billion years ago during what is often called the age of the blue-green algae. As the level of  $O_2$  in water rose and by diffusion the gas was transferred to the atmosphere, the terrestrial obligate anaerobes of the time perished with only a few, e.g., tetanus bacteria, surviving until today. Some molecular  $O_2$  could, however, have been produced inorganically through UV light-induced photodissociation of water vapor in the primitive atmosphere after which  $H_2$  was lost into the interplanetary space. Such

a small quantity of O, would not have been of any biochemical consequence, as much of it would have been rapidly taken up by the unoxidized volcanically produced gases and mineralogic factors. However, a modicum level of photolytic (nonbiological) molecular  $O_2$  may have nurtured the evolution of biological aerotolerance to  $O_2$  in the elementary biota (Fay 1965; Holm-Hansen 1968) through development of specific biochemical pathways of mopping up and detoxifying intracellular O<sub>2</sub>. This would have imparted a selective advantage to such moderately adapted microorganisms (e.g., Schopf and Walter 1983). Based on 16S ribosomal RNA sequencing of the prokaryotes (Fox et al. 1980), it has been shown that under modicum level of O<sub>2</sub>, these fledgling life forms gave rise to the aerobic eukaryotes. The threshold for this transition (about 0.2% or 0.002 atm) has been incorrectly termed the Pasteur point as it resembles the Pasteur effect (e.g., Dejours 1975), the level of O<sub>2</sub> at which amphiaerobes change from anaerobic (fermentation) to aerobic metabolism. At that critical point, an organism converts from low efficiency fermentation to high efficiency aerobic energy-yielding catabolism. The actual point in time when this process occurred varied between organisms and environmental circumstances.

#### 1.12 Accretion of Molecular 0<sub>2</sub>

As a general rule, the ecological resources available to organisms in any environment are finite (Hutchison 1959; MacArthur and Levine 1967; Levine 1976; Brown 1981). Environments cannot endure if they are continuously depleted of resources. Life will last only if and as long as there is frugal utilization and coherent cycling and revitalization of materials and energy (Smil 1997). Hydrologic and atmospheric O<sub>2</sub>-CO<sub>2</sub> recycling are but two of the many global natural rotations of resources (e.g., McLean 1978). The changes in the partial pressures of O<sub>2</sub> and CO<sub>2</sub> in the biosphere are some of the most fascinating parameters which have influenced the direction and pace of development of life on Earth (Berkner and Marshall 1965; Rutten 1970). To stabilize and set optimal tolerable limits, at least since the Devonian, environmental PO<sub>2</sub> regulation was brought under direct biological control. This occurred through transfer between the biological sources (photosynthesis) and sinks (aerobic respiration) (Fig. 10). In effect, but not in mechanism, the processes of respiration and photosynthesis are diametrical: the former yields CO<sub>2</sub> and the latter O<sub>2</sub>. Plants (photosynthetic autotrophs) and animals (heterotrophs) are involved in a continuous, intricate process of resource recycling, maintaining constant levels of  $CO_2$  and  $O_2$  in the atmosphere (Fig. 10). Using sunlight, atmospheric  $O_2$  is continuously replenished by aquatic and terrestrial plant life and CO<sub>2</sub> and H<sub>2</sub>O are produced by the respiratory processes. The end products of the way of life of each group become food for the other. The sun is the decisive source of the energy which drives this global animal-plant continuum of resource recycling. Averaged globally, the Earth receives  $343 \text{ W m}^{-2}$  of energy from the sun in form of short wavelength radiation. About one third of it is reflected back into space by the atmosphere and the remainder  $(240 \text{ W/m}^2)$  is absorbed by the Earth's surface and the atmosphere. About one third  $(103 \text{ W/m}^2)$ 



Fig. 10. Scheme showing the processes which regulate the  $O_2$  levels in a closed habitat and the dynamics of gas transfer between water and air. The rather constant atmospheric  $O_2$  and  $CO_2$  levels are maintained by the cyclic balance between the photosynthetic and respiratory processes and supported by the fast diffusion rates of gases in air

of the net incoming solar radiation is reflected by the Earth's surface to the atmosphere (the Earth's albedo) in form of long wave-length radiation where it is absorbed by the greenhouse gases (e.g., water vapor, CO<sub>2</sub>, ozone, methane, and nitrous oxide) and the clouds maintaining the surface temperature about 33 °C warmer than it would otherwise be without them (= the natural greenhouse effect). Oxygen and CO<sub>2</sub> are exchanged in air and to an extent with water by diffusion. Without a self-regulating O2 and CO2 recycling mechanism, life on Earth would have been short-lived. In the modern atmosphere, nitrogen constitutes 78.09%, O<sub>2</sub> 20.95%, and CO<sub>2</sub> 0.03%, the rest being composed of rare gases such as argon, hydrogen, krypton, xenon, etc. The present so-called normoxic atmospheric level of  $O_2$  (21% by volume = about 0.2 atm) is strictly hyperoxic and far in excess of the optimum respiratory needs for life. Strictly, the modern terrestrial animals are exposed to an oxidative stress. According to the available data on amphiaerobic eukaryotic yeasts, systemic aerobic biochemistry can occur at values of 0.04% O<sub>2</sub> by volume (about 0.0004 atm) and organismic aerobiosis can occur at O<sub>2</sub> concentrations as low as 0.2 to 0.4% (i.e., 0.002 to 0.004 atm) (e.g., Rogers and Stewart 1973; Jahnke and Klein 1979). It is probably not coincidental that the Pasteur point is about 0.2% O, by volume, a value equal to the minimum O<sub>2</sub> level able to support organismal aerobiosis (Chapman and Schopf 1983). Adaptations to withstanding the harmful effects of the reactive factors of molecular  $O_2$  was of particular importance in those life forms which produced the gas itself or were immobile and hence unable to escape from microhabitats with high concentrations of it. Unlike animals which normally operate under rather constant and somehow manageable O<sub>2</sub> tensions, green plants which produce the molecule itself had to evolve a complex range of molecular factors for protection against oxidative attack. Chloroplasts are the main source of antioxidants which include vitamins C and E (Crawford et al. 1994). Furthermore, while the land plants are exposed to air (21% O<sub>2</sub> by volume), the roots are located in the soil which, depending on type and firmness, may be virtually anoxic at depth (Currie 1962, 1984). In the marine angiosperms (Teal and Kanwisher 1966; Armstrong 1970), O<sub>2</sub> is known to diffuse from the roots, creating aerobic zones in the immediate area. This provides a unique microhabitat for some marine creatures such as the eulamellibranch bivalve, Lucina floridana (Britton 1970).

By the start of the Paleozoic era (about 600 million years ago), the PO<sub>2</sub> in the water and air had risen to the modest level of 0.2 kPa, i.e., one hundredth (= 0.2%) O<sub>2</sub> by volume) of the modern sea level value. When the first vertebrates (ostracoderms) appeared some 550 million years ago (e.g., Forey and Janvier 1994), the PO<sub>2</sub> was only 0.9 kPa and by the Silurian-Permian periods, some 450 to 250 million years ago, the  $PO_2$  had risen to 4.7 kPa when the amphibians ventured onto land (McClanahan et al. 1994). The terrestrial arthropods and amphibians were well entrenched on land by the Devonian period, by which time the PO<sub>2</sub> had risen to 10.7 kPa. The critical environmental threshold (10% of the present-day O<sub>2</sub> level) was crossed late in the Proterozoic era (Canfield and Teske 1996). The present level of 21 kPa was not reached until the Carboniferous period (350 million years ago) when the first reptiles appeared on land (e.g., Carroll 1988). The level of atmospheric O<sub>2</sub> has fluctuated greatly in the Phanerozoic (e.g., Tappan 1974; Cloud 1983a; Berner and Canfield 1989; Graham et al. 1995; Fig. 9). During the late Paleozoic, over a period of about 120 million years, O<sub>2</sub> rose to a hyperoxic level of 35% (compared to the present atmospheric level of 21%) and then dropped precipitously to a hypoxic low of 15% (Berner and Canfield 1989; Landis and Snee 1991). These changes were duplicated in the water (e.g., Hosler 1977; Solem 1985; Dejours 1994) and had a dramatic influence on the aquatic life (Table 1), inducing relocation to land. The greater availability of  $O_2$  during the Mid-Devonian to Carboniferous hyperoxic episode would have made it possible for organisms, e.g., the arthropods, to attain larger body sizes (Graham 1994). Furthermore, the abundance of O<sub>2</sub> resulted in higher metabolic capacities and greater accessibility to resources instigating vast radiation of the animal life.

#### **1.13** CO<sub>2</sub> Pulses in the Biosphere

The atmospheric and aquatic levels of  $CO_2$  have undergone remarkable fluctuations in the past (e.g., Bender 1984; Shackleton and Pisias 1985; Walker 1985;

Barnola et al. 1987; Berger and Spitzy 1988; Jasper and Hayes 1990). The partial pressure of CO<sub>2</sub> is presumed to have been 100 to 1000 times more in the antediluvian Earth than now (Walker 1983). It is envisaged that  $CO_2$ ,  $CH_4$ , and NH<sub>3</sub> produced a greenhouse effect which sustained liquid water (e.g., Owens et al. 1979; Kasting 1997; Sagan and Chyba 1997). The solar luminosity during that time was 25 to 30% lower than at present (e.g., Newman and Rood 1977). Models of the early Earth after the end of the heavy bombardment suggest that the PCO<sub>2</sub> may have been as high as 10 bar (Walker 1977, 1983). From analysis of air trapped in the ice cores, Raynaud et al. (1993) observed that the atmospheric  $CO_2$  decreased from about 290 to 190 µmol mol<sup>-1</sup> over a period of about 10000 years during the last interglacial-glacial maxima. Whereas the atmospheric partial pressures of nitrogen and helium remained fairly constant across the Phanerozoic, those of neon, krypton, and argon may have increased through mantle and crustal degassing (Holland 1984; Warneck 1988). Carbon dioxide and O<sub>2</sub> oscillations have occurred in reverse manner (e.g., Delmas et al. 1980; Neftel et al. 1982; Graham et al. 1995; Fig. 9). This is due to the fact that over geological time, photosynthetic carbon fixation in the oceans has surpassed the respiratory oxidation of carbon (Holland 1984; Berner 1991; Walker 1987; Falkowski 1997): the difference between the two values has reflected the net increase in O<sub>2</sub> and reduction of CO<sub>2</sub> from the Earth's atmosphere. During the Devonian period (400 to 360 million years ago), the spread of rooted vascular plants to the elevated areas of the dry land may have enhanced chemical weathering leading to removal of CO<sub>2</sub> from the atmosphere (Berner 1997; Fig. 9). Mechanisms of phosphorus-mediated redox stabilization of the atmospheric and marine O2 levels (e.g., Redfield 1958; Broecker 1982; Cappellen and Ingall 1996) and nitrogen fixation and denitrification in sequestration of CO<sub>2</sub> in the oceans over geological time scales have been described by McElroy (1983), Codispoti and Christensen (1985), Shaffer (1990), and Falkowski (1997). During the past 2 million years, reduction in the atmospheric CO<sub>2</sub> level has correlated with increases in the deposition of organic carbon from the surface waters to the marine sediments (e.g., Sarnthein et al. 1988; Mix 1989; Hansell et al. 1997): carbon is traded between the atmosphere, the oceans, and the terrestrial biosphere, and in geological time scales between the sediments and the sedimentary rocks. The equatorial Pacific Ocean is the greatest oceanic source of CO<sub>2</sub> to the atmosphere and is also the main site of organic carbon discharge to the deep sea (Murray et al. 1994).

The highest levels of  $CO_2$  in the biosphere occurred in the Ordovician and Silurian, mainly owing to massive tectonic activities (Holland 1984). Between the late Miocene, Pliocene, and early Pleistocene (i.e., between 10 and 2 million years ago), the concentration of  $CO_2$  fluctuated within 280 and 370 ppm by volume (van der Burgh et al. 1993). By the Carboniferous, the level had dropped almost to the present one of 0.036% subsequently rising threefold by the end of the Permian (Graham et al. 1995). In the Archean,  $CO_2$  level may have been 100 times greater than it is today (Walker et al. 1983). In the geological recent past, the atmospheric concentration of  $O_2$  has been fairly constant but that of  $CO_2$  is estimated to have increased from 0.029 to 0.033% within a period of 50 years, i.e., between 1900 and 1950 (Callender 1940) owing to combustion of fossil fuels. From the start of this

century, anthropogenic emission of CO<sub>2</sub> (mainly from activities such as burning fossil fuel, cement production, and changes in land use) has led to its increase in mole fraction from 0.00030 to 0.00034 (Revelle 1982). Between the years 1980 and 1989, the average annual anthropogenic production of  $CO_2$  is estimated to have been 7.1 billion tonnes (Bolin et al. 1994). Since the industrial revolution, the concentration of  $CO_2$  has risen from 280 to 350 ppm by volume (ppmv), the highest value reached in the last 160 000 years (Bazzaz and Fajer 1992; Bolin et al. 1994). Estimations at the Mauna Loa observatory in Hawaii indicated about 20% real rise in CO<sub>2</sub> levels between the years 1957 and 1987 (Barnola et al. 1987). Owing to the greenhouse effect, the present global temperature change correlates with the logarithm of the atmospheric CO<sub>2</sub> concentration (Thomson 1995). It is projected that through anthropogenic emissions, global concentrations of CO<sub>2</sub> will double by the end of the 21st century, a process which may cause a temperature rise of about 3 °C, resulting in serious ecological consequences (e.g., Bazzaz and Fajer 1992; Azar and Rodh 1997). If CO<sub>2</sub> emission were held at the present level, this would lead to a nearly constant rate of increase in the atmospheric concentrations for at least two centuries, stabilizing at 500 ppmv by the end of the 21st century, i.e., about twice the level of the time before the industrial revolution, which was about 280 ppmv (Bolin et al. 1994). Photosynthesis can be stimulated by increased level of CO<sub>2</sub>: an inverse relationship between stomatal frequency in the leaves of C<sub>3</sub> plants and anthropogenic increase in atmospheric CO<sub>2</sub> concentration has been demonstrated (e.g., Wagner et al. 1996). Under optimal conditions of water and nutrient supply, there is a potential increase in photosynthesis by 20 to 40% when the level of CO<sub>2</sub> rises (Youvan and Marrs 1987). Undisturbed forests are important terrestrial sinks of CO<sub>2</sub> (Grace et al. 1995). The view that an atmosphere enriched with CO, will accelerate photosynthesis resulting in a "greener planet" and that the greenhouse effect will be brought under control by increased withdrawal of CO, from the atmosphere by the luxuriant plant growth has been deemed highly simplistic and short-sighted (e.g., Bazzaz and Fajer 1992). Presently, CO<sub>2</sub> is the most important gas in the causation of global warming by the greenhouse effect, a state which may lead to irreversible changes in plant physiology and pattern of the vegetation cover (Betts et al. 1997).

Although  $CO_2$  is well mixed in the atmosphere, variations in its concentration in air over land and that over the oceans and between the Northern and Southern Hemispheres are well recognized. In the more industrial Northern Hemisphere, the concentration of  $CO_2$  rises in winter and declines in summer mainly in response to seasonal growth in land vegetation (e.g., Chapin et al. 1996; Keeling et al. 1996): the seasonal cycle (peak to trough) is 15 to 20 ppmv in the far north. The equilibrium PCO<sub>2</sub> between air and seawater increases three times when temperature rises from 0 to 30 °C. The surface temperature of the sea modifies the  $CO_2$ content of the oceanic biosphere, making the cold polar air contain as much as 20 ppm less  $CO_2$  than the warmer continental and tropical air. In spite of such fluctuations, the composition of the atmosphere is considered to be reasonably homogenous. This is mainly attributable to the greater turbulence of air resulting from temperature differentials in various parts of the Earth and the high diffusivity of atmospheric gases at these temperatures. This accounts for the simplicity of the composition of the atmosphere and, for that matter, of any gas mixture. With as little information as temperature and barometric pressure, it is possible to accurately predict and estimate the changes in the tensions and concentrations which occur in a gas phase when  $O_2$  is consumed and  $CO_2$  released during respiration. Except for special microhabitats such as burrows and dens of mammals, reptiles, and birds, caves of freely roosting bats, within colonies of insects, and in pouches of marsupials (e.g., Mitchell 1964; Boggs et al. 1984; White et al. 1984), where relatively high levels of CO<sub>2</sub> and NH<sub>3</sub> and low concentrations of  $O_2$  may occur, deviations in the basic composition of atmospheric gases are only naturally encountered at high altitude due to changes in barometric pressure. The rise in the atmospheric O<sub>2</sub> and the drop in that of CO<sub>2</sub> were major factors in the development of the modern respiratory organs. In the derelict aquatic habitats, high concentrations of CO<sub>2</sub> in water constituted a decisive driving force for transition to air breathing (Chap. 5). High atmospheric levels of  $O_2$ led to a greater emphasis of the lungs for gas exchange, influenced the transition from buccal pumping to suctional breathing (Liem 1985; Brainerd 1994), and perhaps occasioned the change from O2- to a CO2-regulated respiratory control mechanism. The reduced water loss to O2 extraction ratio was an important benefit derived from occupying a normoxic atmosphere. The early Paleozoic aquatic animals and subsequently the early amphibians subsisted in hypoxic conditions similar to the inimical ones which presently occur in habitats such as in burrows, tropical swamps, ocean sediments, and high altitudes. The present-day bimodal breathers and the developing amphibians face challenges similar to those which confronted the pioneers of transition from water- to air breathing.

### 1.14 The Overt and Covert Roles of O<sub>2</sub> in Colonization and Extinctions of Biota

Except for the recent past when the Earth presented much the same kind of climate as it does now (e.g., Boucot and Gray 1982), the planet is replete with recurrent catastrophic crises of varying magnitudes, a number of which have more or less directly or indirectly corresponded with the levels of  $O_2$  in the biosphere. Five major episodes, in addition to numerous minor ones, have occurred. They took place at the end of the Ordovician, late Devonian, end of Permian, end of Triassic, and end of Cretaceous (e.g., Benton 1993; Weinberg 1994). Fairly fortuitously, life has navigated through these hazards, but at an enormous cost. About 99.99% of all animal species which have ever evolved on Earth are now extinct (Pough et al. 1989). During the mid-Paleozoic Mass Extinction, dubbed "the mother of all extinction" by Erwin (1993), an event which occurred towards the end of the Permian period (e.g., Robinson 1991; Allegre and Schneider 1994), adverse tectonic activity which lasted through the Triassic to the early Jurassic culminated in an abrupt temperature decline. Concurrently, a sudden drop in sea level occurred. The land submerged under water decreased from 30 to 5%, leading to exposure of expansive organic sediment (Erwin 1996). This resulted in increased utilization of O<sub>2</sub> in oxidative processes with an atten-

dant upsurge in the discharge of CO<sub>2</sub> into the atmosphere. The consequence was an extremely severe hypoxia which was exacerbated by hypercarbia (Benton 1995; Wignall and Twitchett 1996): a worldwide deep sea anoxic outcome which occurred across the Permo-Triassic (or Paleozoic and Mezozoic) boundary (~250 million years ago) which lasted for 20 million years has been described by Isozaki (1997). The mid-Paleozoic crisis led to a near-annihilation of the marine biota (e.g., Tappan 1974; Weinberg 1994). Even under such dire circumstances, the level of O<sub>2</sub> in air was greater than that in water, making air breathing an evolutionary advantage. Nearly 90% of the aquatic animals succumbed (e.g., McGhee 1989) and on land, more than two thirds of the reptilian and amphibian species perished (Erwin 1993, 1994). In the only mass extinction which insects have had to endure, a sign of the severity of the prevailing conditions, 30% of the orders died (Erwin 1996). In all taxa, the particularly vulnerable groups were those which had preadapted to the earlier hyperoxic milieu of the Carboniferous. It has, however, been argued, e.g., Graham et al. (1995), that lack of  $O_2$  per se was not the primary factor which precipitated this immense demise, since the drop in O<sub>2</sub> from the peak levels is envisaged to have been very gradual, occurring over a long period of time during which the animals should have adapted to the change. However, Erwin (1996) pointed out that the actual active period of extinction may have been as short 1 million years, if not less. In the late Triassic extinction (about 200 million years ago), 20% of the families of animals died out, eliminating some 50% of the species (Benton 1993). Animals such as the ammonoids and bivalves were severely decimated and the cocodonts disappeared. Contrary to expectation, no frequency or periodicity has been evident in the seven mass extinctions (i.e., in the Early Cambrian, Late Ordovician, Late Devonian, Late Permian, Early Triassic, Late Triassic, and End Cretaceous) which have occurred in the last 250 million years (Benton 1995): the episodes are separated by between 20 and 60 million years.

The ecological disaster which has caught most of the attention of scientists and the public, even though far less severe, is that of the dinosaurs and their contemporaries, the plesiosaurs and pterosaurs. This occurred between the Cretaceous and Tertiary (65 million years ago). The global faunal diversity was reduced by 60 to 80% (Raup and Jablonski 1993). Geological (terrestrial) as well as cosmic (extraterrestrial) events have been associated with the demise (e.g., Hallam 1987; Stanley 1987; Kerr 1988; Powell 1993). Based on a high concentration of iridium in some rock deposits at the end of the Cretaceous, a large asteroid or a comet about 10 km in diameter is thought to have impacted on Earth off the Yucatan Peninsula (Mexico) (e.g., Sheehan et al. 1991). The mass of dust thrown into the atmosphere blocked out the sun's rays, suppressed photosynthesis (perhaps causing a drop in the level of O<sub>2</sub> in both air and water), and occasioned severe changes in the Earth's climate. As in the "mid-Paleozoic Mass Extinction", sea level fluctuated by about 50 to 200 m over a period of 0.2 to 1 million years (Haq et al. 1987; Kerr 1996b; Stoll and Schrag 1996). The definite cause of the drop in sea level during the Cretaceous, a period during which the climate is predicted to have been fairly stable, where the equatorial temperatures were equal to the present ones (Herman and Spicer 1996), and the poles were ice-free (Barron et al. 1981), is still debatable.

#### 1.15 Oxygen: a Paradoxical Molecule

Most profoundly, molecular O<sub>2</sub> has influenced the geology of the Earth and pervasively directed the trajectory and forward momentum of evolution of life. Since the first appearance of the gas in the biosphere in appreciable quantity some 2 billion years ago (e.g., Owens et al. 1979), the history of life is literally inscribed on this single molecule. The buildup of O<sub>2</sub> and evolution of oxygenic respiration (following the radical conversion of the incipient high CO<sub>2</sub>-low O<sub>2</sub> atmosphere to low CO<sub>2</sub>-high O<sub>2</sub> atmosphere) led to the transformation of the early simple anaerobic cells to the versatile eukaryotic ones which subsequently accreted into the aerobic, multicellular organisms (Siever 1979; Allegre and Schneider 1994; Orgel 1994). The transformation enhanced the efficiency of carbon and nutrient recycling, leading to a climactic increase in the organic biomass. Margulis (1979) envisages that the toxicity of the O2 molecule enforced symbiotic associations on anaerobic bacteria, leading to development of eukaryotic cells. Symbiotic relationships are known to evolve under extreme circumstances (e.g., Childress et al. 1989; Rennie 1992). The presence and the resolutely increasing levels of O<sub>2</sub> may explain the short period (in evolutionary terms) of 700 million years which it took for the growth and change of the eukaryotic cells to multicellular organisms compared with the over 2 billion years which passed before the aerobic eukaryotic cells developed from the anaerobic prokaryotes (Gould 1994; Fig. 8).

In virtually all the solar system planets and their satellites, free atmospheric O<sub>2</sub> for which there are no known primary (geochemical) sources, is found in high concentration only on Earth. As a molecular factor, however, O<sub>2</sub> is not unique to our biosphere. Jupiter's moons Europa and lo, for example, have surface and atmospheric water, gaseous sodium, and small quantities of O<sub>2</sub> (Brown and Hill 1996; Kerr 1997), a state which is similar to that of Earth some 4 billion years ago. Due to its high reactivity with other elements at both the temperatures of the formation of the magma (500 to 1200  $^{\circ}$ C) and at ordinary surface temperatures, O<sub>2</sub> is the most abundant element of the average crustal rocks, followed by silicon, aluminum, and iron (Chapman and Schopf 1983). This notwithstanding,  $O_2$  is a somewhat alien factor to life. A reducing (nonoxidizing) environment was a prerequisite for the chemical evolution of life from the first organic molecules in the primitive atmosphere (e.g., Cloud 1974, 1988; Miller and Orgel 1974; Tappan 1974; Schidlowski 1975; Chang et al. 1983). It is an intellectually intriguing contradiction that life, which evolved in absence of  $O_2$ , is now tractably dependent on it. Oxygen is a necessary resource for body growth and development (e.g., Adelman and Smith 1970; Priede 1977; Armstrong et al. 1992). Extended exposure to hypoxia in newborn and young mammals causes a decrease in body growth (e.g., Timiras et al. 1957; LaManna et al. 1992). While animals that are well adapted to high altitude, like the Ilama and many small rodents and birds, are known to reproduce successfully up to an altitude of 5 km, human fetal growth is retarded at altitudes beyond 3km above sea level (e.g., Haas et al. 1980; Mayhew et al. 1990).

#### 1.16 The Rise of the Level of Molecular O<sub>2</sub>: a Curse or a Blessing?

#### 1.16.1 The Deleterious Reactive Radicals of Molecular 0<sub>2</sub>

Although the aerobic life-style confers great advantages, as metabolism using  $O_2$ yields 20 times more free metabolic energy than an anaerobic one, utilization of O<sub>2</sub> is accompanied by great danger (Pryor 1986; Joenje 1989; Sies 1991). Even under ordinary conditions, due to continuous formation of free radicals (chemically highly reactive molecules with an unpaired electron) which are intermediates of a number of biochemical reactions,  $O_2$  is a highly toxic substance. Its utilization by aerobes is harmful in both the short and long terms (e.g., Comroe et al. 1945; Clark and Lambertsen 1971; Fridovich 1978; Halliwell 1978; Hill 1978; Slater 1984; Sies and Cadenas 1985). Physical exercise increases formation of reactive O<sub>2</sub> species but in the long term, endurance training improves antioxidant handling (Sen 1995). Strictly, molecular  $O_2$  is not the toxic agent but rather its reactive derivatives (Cochrane 1991). Inflammatory cells (neutrophils, eosinophils, and macrophages), catalase negative bacteria, inhaled environmental pollutants (e.g., ozone and nitrous oxide), and even epithelial cells (e.g., intracellular production from mitochondrial respiration and xenobiotic drug metabolism) are potential sources of reactive O2 species (Bast et al. 1991; Kinnula et al. 1992; Cohn et al. 1994). The oxidants have been associated with pathogenesis of respiratory disorders such as adult respiratory distress syndrome, emphysema, asthma, and pollutant-precipitated diseases (Adler et al. 1990; Barnes 1990). During ordinary aerobic respiration, complete reduction of a molecule of O<sub>2</sub> to H<sub>2</sub>O requires four electrons which are sequentially utilized in the process:

$$e^ e^ e^ e^ e^-$$
  
 $O_2 \rightarrow O_2^- \rightarrow H_2O_2 \rightarrow OH^- \rightarrow H_2O_2$ 

Various active intermediates, highly reactive chemical species with one or more unpaired electrons, are produced. These include the superoxide anion radical  $(O_2^{-})$ , hydrogen peroxide  $(H_2O_2)$ , hydroxyl radical  $(OH^{-})$ , and singlet oxygen  $(^{1}O_{2})$ . The oxidant by products of metabolism cause extensive damage to DNA, proteins, and other macromolecules (Borteux 1993; Epe 1995). Degenerative diseases associated with aging such as arteriosclerosis, Parkinson's disease, diabetes, cancer, decline in the immune system, and senility have been associated with the oxidative damages caused by molecular O<sub>2</sub> (e.g., Fraga et al. 1990; Wagner et al. 1992; Ames et al. 1993; Gutteridge 1993; Halliwell 1994). The assault by the reactive  $O_2$  radicals on cell functional and structural integrity is intense. It is estimated that about 2 to 3% of the  $O_2$  taken up by aerobic cells results in production of  $O_2^$ radical and H<sub>2</sub>O<sub>2</sub> (Chance et al. 1979). Approximately 10<sup>12</sup> O<sub>2</sub> molecules are handled by a rat cell daily (Chance et al. 1979). This results in about  $2 \times 10^{10}$  (i.e., 2%)  $O_2^-$  and  $H_2O_2$  active (partially reduced) species. Frage et al. (1990) estimated that there are about  $9 \times 10^4$  attacks on the DNA per day per cell in a rat. Under a steady state, about 10% of protein molecules may undergo carbonyl modifications (Fridovich 1978; Ames et al. 1993; Orr and Sohal 1994). Ames et al. (1993) envisaged that  $O_2$  free radicals are responsible for 10000 or so DNA base modifications per cell per day. Such sustained attacks can easily overwhelm the cell's repair mechanisms. All cells have evolved a number of repair endonucleases which specifically recognize and repair the damages caused by the reactive species (Lindal 1990; Demple and Harrison 1994). When the biodegradative processes surpass the biosynthetic ones, the cumulative damages may result in significant loss in the cell functional capacity (Stadtman 1992). Oxygen free radical-mediated lipid peroxidation, for example, could easily lead to loss of membrane integrity and hence compromise the normal cellular activities. A substantial memory recovery was achieved by chronically treating old gerbils (15 to 18 months) with a free radical spin-trapping compound *N-tert*-butyl- $\beta$ -phenylnitrone (Carney et al. 1991). The process resulted in a substantial decrease in the amount of oxidized protein in the brain and an increase in the amounts of glutamine synthetase and neutral protease activities. Oxygen, and particularly the resultant OH<sup>-</sup> radical, increase the lethal effects of ionizing radiation (Von Sonntag 1987; Ames et al. 1993). Lipid peroxidation results in mutagenic factors (e.g., epoxides and alkoxyls) and <sup>1</sup>O<sub>2</sub>.

The formation of reactive O, species inside cells constitutes a serious threat to the functional and structural integrity of the cellular genome (Lindal 1990; Epe 1995). It is envisaged that control of O<sub>2</sub> toxicity could have necessitated the evolution of the nucleus and the nuclear membrane in the eukaryotic cells to minimize external affronts by molecular O<sub>2</sub> (e.g., Margulis 1981). The nucleus constitutes an anoxic and fairly safe location for the deoxyribonucleic acid (DNA). The DNA nearer to the nuclear membrane and that in the O<sub>2</sub>-rich cytoplasm are more susceptible to damage. The mitochondrial DNA (mtDNA) is particularly more exposed to O<sub>2</sub> toxicity compared with the nuclear DNA (nDNA) (Gupta et al. 1990; Dyer and Ober 1994). The mtDNA from the rat liver causes more than ten times the level of oxidative DNA damage than does nDNA from the same tissue (Richter et al. 1988). This may be due to factors such as lack of mtDNA repair enzymes, lack of histones protecting mtDNA, and the proximity of mtDNA to oxidants produced during oxidative phosphorylation. A high turnover of mitochondria ensures removal of the damaged organelles (which generate greater quantities of oxidants) but overall, oxidative lesions accumulate in the mtDNA at a greater rate than in the nDNA (Ames et al. 1993). Generally, mitochondria are protected from injury by: (1) the complex extracellular O<sub>2</sub> diffusion pathway where at the cellular level the PO<sub>2</sub> will have dropped to almost zero and (2) by clustering, a process which reduces the area available for influx of  $O_2$  into individual mitochondria (Gnaiger 1991). The PO, in blood capillaries of the cardiac muscle is ten times greater than in isolated mitochondria (Tamura et al. 1989). Clustering accounts for the difference in the  $O_2$  uptake capacities between isolated liver mitochondria and the intact ones in the hepatocytes (Jones 1986). Connett et al. (1985) estimated that the PO<sub>2</sub> in the red muscle cell is about 0.07 kPa. Mitochondria in living tissues are estimated to operate at low O<sub>2</sub> levels, frequently below 2% of air saturation of a  $PO_2$  of 0.5 kPa (Wittenberg and Wittenberg 1987, 1989; Graiger et al. 1995). Respiration in the mitochondria is not affected until the PO2 drops to below 0.01 to 0.1 kPa (Oshino et al. 1974; Sugano et al. 1974). Just as in high quantities, lack of  $O_2$  is equally injurious. In those tissues that are intolerant to hypoxia, e.g., the brain and the heart muscle, mitochondria undergo irreversible structural failures where the membrane potentials

decline, and the degree and capacity of molecular coupling is lowered (Zimmer et al. 1985).

#### 1.16.2 Senescence: the Effects of Molecular O<sub>2</sub>

Senescence is a progressive and irrevocable loss of functional capacity due to degeneration of somatic cells in the last part of life. It has been associated with the use of O<sub>2</sub> by the cells even under normal physiological conditions (e.g., Floyd 1991; Ames et al. 1993; Sohal and Weindruch 1996). As part of the metabolic processes, aerobic organisms generate potentially destructive O<sub>2</sub> species which cause serious oxidative damage to biological macromolecules (e.g., Fridovich 1978). The damage occurs in form of peroxidation of membrane polysaturated fatty acid chains, alteration in DNA protein bases configuration, and carbonylation and loss of sulfhydryls in proteins. Changes due to aging directly or indirectly affect the O<sub>2</sub> uptake process itself (Horvath and Borgia 1984). A senile lung is defined as one which presents dilation of the air spaces without tissue destruction (e.g., Hyde et al. 1977; Pinkerton et al. 1982; Snider et al. 1985; Dios-Escolar et al. 1994). Studies on the effects of age on lung structure have, however, been inconlusive (e.g., Thurlbeck 1980). The impedement has been due to the fact that the effects of environmental pollution and lifestyle have been difficult to fully assess and exclude from the various studies. No aging structural changes in the lung were observed in the growing specific-pathogen-free inbred male BALB/ cNNia mice between 38 days and 28 months by Masahiko et al. (1984). In adult humans, arterial O<sub>2</sub> tension decreases by an average of 0.28 to 0.54 kPa each decade while the alveolar PO<sub>2</sub> remains constant or increases slightly (Dill et al 1963; Sorbini et al. 1968). Pulmonary diffusing capacity decreases by 5 to 8% in each decade of life (Cohen 1964). With aging, the surface area of the erythrocytes decreases and the cells become more susceptible to osmotic changes and mechanical disruption while there is reduction in enzymatic activity, e.g., hexokinase, glucose-6-phosphate dehydrogenase, and lactate dehydrogenase (Prankerd 1961). Compared with young rats, old rats have a lower hind limb muscle respiratory capacity and whole body maximal O<sub>2</sub> consumption (Cartee and Farrar 1987). In both animals and plants, paraquat (methyl viologen), a commonly used herbicide, and antibiotics such as streptonigrin which enhance the rate of O<sub>2</sub> radical  $(O_2^{-})$  production, are more toxic under aerobic than anaerobic conditions. Longevity correlates inversely with the rate of mitochondrial production of the  $O_2^-$ ,  $H_2O_2$ , and  $OH^-$  species. In a number of mammals, the severity of oxidative DNA damage appears to correlate with the metabolic rate (Adelman et al. 1988; Shigenaga et al. 1989). Restriction of calorific intake, a process which lowers metabolic stress, reduces age-related changes and prolongs the life span of mammals (Sohal and Weindruch 1996). Overexpression of superoxide dismutase and catalase, antioxidative enzymes which respectively remove  $O_2^-$  and  $H_2O_2$ , extends the life span of Drosophila melanogaster (Orr and Sohal 1994). Reduction in production of active  $O_2$  species in transgenic Drosophila melanogaster leads to a 30% increase in the metabolic potential (Orr and Sohal 1994). In two human diseases associated with premature aging, Werner syndrome and progeria, oxidized protein residues increase at a faster rate than normally (Stadtman 1992). The gradual decay of the cell integrity with age is an indication that the antioxidant defenses of the aerobic cells are inefficient in face of sustained assault by the reactive  $O_2$  metabolites. Oxidative lesions on DNA increase with age (Ames et al. 1993): at the age of 2 years, a rat cell has about 2 million lesions per cell. This is about twice that of a young rat. Exogenous oxidant loading from activities such as smoking may deplete the endogenous antioxidant levels in the cells, such stress compromising the cell defenses (Csillag and Aldhous 1992).

Notwithstanding the deleterious effects of the endogenous reactive species, not all effects of the active species are harmful. Leukocytes and other phagocytic cells destroy bacteria or virus-infected cells by subjecting them to a lethal discharge of nitric oxide (NO), O<sub>2</sub><sup>-</sup>, H<sub>2</sub>O<sub>2</sub>, OH<sup>-</sup> and OCl<sup>-</sup>, a potent oxidant mixture (Baldridge and Gerard 1933; Seifert and Schultz 1991; Stamler et al. 1992). Like O<sub>2</sub>, among gases, NO (which, unlike O2, is elaborated in animal tissues but in very small quantities; De Belder et al. 1993; Ballingand et al. 1995) is an enigmatic molecule. In the atmosphere, NO is a toxic chemical but in small regulated quantities in the body, it plays vital physiological, pharmacological, and immunological roles. It participates in processes such as blood pressure control (through smooth muscle relaxation), platelet inhibition, neurotransmission, destruction of pathogens, penile erection, and has even been associated with learning and long-term memory (Culotta and Koshland 1992; Koshland 1992; Stamler et al. 1992; De Belder et al. 1993; Ballingand et al. 1995). Nitric oxide and, amazingly, carbon monoxide (one of the most feared gases) are the first gases known to physiologically regulate levels of guanosine 3,5-monophosphate and thus act as biological messengers and signaling molecules in mammals (Moncada et al. 1991; Toda and Okamura 1991; Galla 1993; Verma et al. 1993; Katušic and Cosentino 1994). Above certain thresholds, active radicals of NO such as nitrosonium cation (NO<sup>+</sup>), nitric oxide (NO<sup>-</sup>), and nitroxyl anion (NO<sup>-</sup>), similar to the redox states of O<sub>2</sub> (i.e., O<sub>2</sub><sup>-</sup>,  $H_2O_2$ , and  $OH^-$ ), form and oxidize biological molecules.

Evolution does not appear to have found any direct, enduring solution for neutralizing the harmful active species of molecular O<sub>2</sub>. However, to ameliorate the effects, cellular biochemistry has been configured such that the most important processes are the reductive ones: biological oxidations entail removal of H<sub>2</sub> rather than addition of O2. The increasing susceptibility of the cells to oxidative effects of the reactive O<sub>2</sub> radicals with age may be caused by several factors. These include: (1) an increase in the O<sub>2</sub> delivery rate to the cells as may occur with changes in the amounts of allosteric effectors which determine the O<sub>2</sub> binding capacity by the hemoglobin, (2) increased availability of the Fe(II) or Cu(II), metals which are catalytically involved in the production of the highly damaging O<sub>2</sub>-free species by changes in the efficacy of metal binding proteins and chelating agents, (3) an age-dependent increase in the production of the reactive  $O_2$  metabolites, (4) an intrinsic decline in the production of the endogenous antioxidant scavenger enzymes and metabolite defenses, (5) loss of capacity to biodegrade the products of cell oxidation, and (6) decrease in the capacity to mobilize and repair the damages caused by the active radicals. In the life cycles of practically all complex organisms, the integrity of the tissue cells appears to be guaranteed only

up to the useful reproductive period. It is possible that reproduction takes such a heavy toll of an animal's resources that the necessary amount needed for maintenance of the integrity of the somatic cells is irrevocably compromised.

#### 1.16.3 Biological Defenses Against O<sub>2</sub> Toxicity

Of the four fundamental elements of life, the so-called biogenetic elements, carbon, hydrogen, oxygen, and nitrogen, O, is a geochemical and biochemical anomaly. Although a structural component of biological tissues and an integral part of biochemical reactions, in combined as well as free diatomic state, depending on the organism and level of concentration, O<sub>2</sub> is a toxic factor. The accretion of molecular O<sub>2</sub> changed the entire global environment and dramatically influenced the nature and tempo of all subsequent evolutionary developments in life. Most animals die after extended exposure to 100%  $O_2$  (1 atm), the endothelial lining of the lung being the main site of injury (e.g., Matalon and Egan 1981; Crapo et al. 1984; Block et al. 1986; Crapo 1986). Rats die between 60 to 72 h after exposure to 100%  $O_2$  (Crapo 1987). Exposure of 21-day-old rats to >95%  $O_2$  for 8 days induces cholinergic hyperresponsiveness as well as hypertrophy of the airway epithelial and smooth muscle layers (Hershenson et al. 1994). More than 50% of 72-h-old chick embryos die on exposure to 3h of 5atm of  $O_2$  and 20 to 30%of those which hatch have deformities of the brain, eyes, upper jaw, legs, feet, and heart (Pizarello and Shircliffe 1967). In mammals, on exposure to hyperbaric O2 (partial pressure of  $O_2$  in the inspired air greater than 1 atm), nervous acute O<sub>2</sub> poisoning occurs. This is expressed in form of epileptiform convulsions (Barthelemy 1987). Nitrogenases, the enzymes necessary for the fixation of nitrogen (e.g., Leigh 1997), are inhibited by as little as 0.1% free O<sub>2</sub> (Postgate 1987). Many nitrogenous bacteria occupy anaerobic habitats, e.g., cells of leguminous plants under the soil. The reactivity of O<sub>2</sub> makes it a rather biologically enigmatic molecule (Cochrane 1991). It readily reacts with the reduced (H-rich) biochemical factors to produce energy (in aerobic respiration) and its highly reactive radicals oxidize and destroy enzymes. The reactive side products such as superoxides, peroxides, hydroxyl radicals, and  $^{1}O_{2}$  are biologically toxic to strict anaerobes and at even moderate concentrations are highly toxic to aerobes (Fridovich 1976, 1978; Halliwell 1978). Practically all organic compounds tend to be easily oxidized and are hence potentially unstable in the presence of molecular O<sub>2</sub> (Miller and Orgel 1974). Adaptively, the epithelial cells which line the respiratory organs contain antioxidant enzymes which prevent initiation of pulmonary pathology after contact with oxidants such as ozone, nitrous oxide, and oxidants produced by local inflammatory reactions (Kinnula et al. 1992; Cohn et al. 1994).

It is overtly anomalous that life started in an  $O_2$ -free environment and subsequently became so heavily dependent on it. The stubbornly increasing levels of  $O_2$ left the cells with no alternative but to somehow accommodate it or face certain annihilation. Its rather small molecular size and hence high intracellular diffusivity and a correct redox potential endeared  $O_2$  to utilization as an electron acceptor in the biochemical energy production of the tricarboxylic chain processes. Interestingly, in myocardial cells, when energy depletion in a cell, e.g., in the case of ischaemia or hypoxia, is extended beyond the stage when reoxygenation leads to spontaneous recovery, in what has been termed the oxygen paradox (e.g., Piper et al. 1994), reoxygenation may intensify tissue damage (Hearse et al. 1973; Ganote 1983; Gorge et al. 1991) presumably due to excessive production of active free radical (McCord 1988; Turrens et al. 1991): life's biochemistry has not achieved a capacity of totally taming molecular  $O_2$ .

As a part of antioxidant defense system, gas exchangers (e.g., Crapo and McCord 1976) and some organisms, e.g., bacterial aerobes like Escherichia coli (Gregory and Fridovich 1973) possess a battery of simple nonenzymatic molecules and complex enzymes which scavenge the oxidative O2 radicals (Forman and Fisher 1981; Freeman and Crapo 1982; Halliwell and Gutteridge 1985; Sies 1991). The former include glutathione, ascorbate, urate, bilirubin, ubiquinol,  $\beta$ carotene, and tacopherol, while the latter comprise superoxide dismutase (SD), catalase, and glutathione peroxidase. Superoxide dismutase converts the superoxide radical  $(O_2^{-})$  to  $H_2O_2$  plus  $O_2$  (e.g., Crapo and Tierney 1974; Fridovich 1975; Cassini et al. 1993) and catalases and peroxidases convert H<sub>2</sub>O<sub>2</sub> to H<sub>2</sub>O and O<sub>2</sub>. Experimentally, rats are protected from O<sub>2</sub> toxicity by intravenous injection of liposome-entrapped catalase and superoxide dismutase (Turrens et al. 1984). Hyperoxia increases release of reactive  $O_2$  species in the mitochondia (Turrens et al. 1982a,b). Adaptively, in old rats, even though glutathione synthesis is decreased, the tissue regeneration capacity appears to be increased to cope with oxidative stress (Ohkuwa et al. 1997). In rat lungs, on hyperoxic exposure, gene expression plays an important role in controlling manganese containing superoxide dismutase activity (Ho et al. 1996). Interestingly, molecular  $H_2$ , has been reported to destroy active O<sub>2</sub> radicals (Jones 1996). Aerobiosis could not have arisen directly from anaerobiosis. It was imperative that preadaptations for tolerating O<sub>2</sub> toxicity would have required time to evolve. It is speculated that photochemically produced O<sub>2</sub> at low manageable levels may have nurtured some degree of aerotolerance (Walker et al. 1983), a feature which would have imparted a selective advantage when a stable oxygenic atmosphere formed. The envisaged chronological order through which molecular O<sub>2</sub> was incorporated into the biochemical processes is: (1) development of defensive mechanisms (though it is difficult to explain how these could have been configured in absence of O<sub>2</sub> itself), (2) production of  $O_2$  by the cyanobacteria, and (3) incorporation and utilization of O<sub>2</sub> at the cellular level. The transition to an aerobic environment brought about and required evolution of higher redox potential biochemistry (e.g., Williams and Da Silva 1978). This necessitated evolution of new catalysts which were based on transitional metals. It has been estimated that the befitting redox potential of the primitive oceans may have been about  $-350 \,\mathrm{mV}$  (Osterberg 1974). Though O<sub>2</sub> is mainly utilized for energy production, many other nonrespiratory processes like collagen synthesis, oxidation of amino acids, and tanning of cuticle (in many insects) require molecular O<sub>2</sub>. In the facultative invertebrate anaerobes, growth is arrested with reduction in the level of O<sub>2</sub> (e.g., Hammen 1969, 1976). To avoid oxidation of nitrogenase enzymes, the nitrogen-fixing cyanobacteria have evolved a novel method of coexistence ensuring intracellular anoxia. Special cells called heterocytes which lack the full complement of photosynthetic pigments and are hence incapable of producing  $O_2$  have developed thick cell walls covered by a mucinous coat to limit  $O_2$  inflow: these cells are endowed with the appropriate enzymes for destroying the harmful  $O_2$  radicals.

#### 1.17 The Evolution of Complex Metabolic Processes

The evolution of the eukaryotes from prokaryotes (some 2 billion years ago), the attainment of capacity for sexual reproduction (some 1 billion years ago) and, subsequently, development of complex multicellular life about 600 million to 1 billion years ago (Schopf and Oehler 1976; Schopf 1978; Wray et al. 1996) were pivotal points in the development and proliferation of animal life (Romer 1967). It is conjectured that mitochondria, the power houses of cells, evolved from freeliving eubacteria-like endosymbionts which more than a billion years ago (e.g., Palmer 1997) invaded the eukaryotic cells (e.g., Margulis 1970, 1979; Vogel 1997). This is vindicated by the fact that organelles such as chloroplasts and mitochondria contain RNA and DNA which are different from those in the nuclei of the eukaryotic cells and in most ways resemble those of certain bacteria. The closest contemporary "relatives" of the mitochondria are the rickettsial group of the  $\alpha$ proteobacteria (Yang et al. 1985; Lang et al. 1997). The rat mitochondrial outer membrane localizes benzodiazepine receptor (MBR) which is expressed in wildtype and TspO<sup>-</sup> (tryptophan-rich sensory protein) strains of the facultative photoheterotroph, Rhodobacter sphaeroides (Yeliseev et al. 1997): functionally, MBR substitutes for TspO<sup>-</sup> and negatively regulates the expression of photosynthesis genes in response to  $O_2$ . This provides further evolutionary support for the origin of mammalian mitochondrion from a photosynthetic precursor. Since it was fully elucidated in the 1960s, the genetic code was thought to be universally identical as any mutations were deemed fatal - the frozen accident hypothesis of Crick (1966). Intellectually, this was a satisfying expectation as it supported the concept of a common origin of life and the parsimony of life in conservation of highly important factors. It is now known that the coding system in mitochondria in various mammalian phyla, certain bacteria, ciliated protozoa, mycoplasma, algae, and yeasts differs from the "universal code", in use of certain codons (e.g., Barrell et al. 1979; Anderson et al. 1981; Jukes 1985; Yamao et al. 1985; Jukes and Osawa 1993). It is envisaged that the universal code may be an evolutionary descendent of the mitochondrial-type code (Osawa et al. 1992) which shows extreme features of genomic economization perhaps to save space (Kurland 1992). By incorporating the endosymbionts, a process called secondary endosymbiosis, the eukaryotic cells did not, so to speak, have to reinvent respiration and photosynthesis through the long and costly trial-and-error process of genetic evolution. The origin of aerobic metabolism involved only extension and refinement of the preexisting anaerobic processes (the O2-independent primitive glycolytic fermentation) by development of catalysts (enzymes) (Keevil and Mason 1978; Gunsalus and Sligar 1978; White and Coon 1980) which acted on the new addition to the energetically more efficient citric acid (Krebs) cycle (Williams and Da Silva 1978; Chapman and Schopf 1983; Schopf 1989). In advanced eukaryotes (e.g., vertebrates and plants),  $O_2$  is involved in the sequence of chemical reactions only at the terminal stages, i.e., the most recently evolved sections of the pathway. The anaerobic bacteria are capable of effecting only the glycolytic pathway. With time, molecular  $O_2$  has been incorporated in the biochemical synthetic pathways of compounds such as phenols, polyunsaturated fatty acids, amino acids, cytochromes, and bile pigments (in vertebrates) and elsewhere only in the relatively advanced organisms of particular lineages. The  $O_2$ -dependent metabolites of the aerobes have amply been used only to refine in a biochemical, structural, and functional manner the earlier established anaerobic systems: no totally new molecular complexes have evolved (e.g., Rohmer et al. 1979; Chapman and Ragan 1980).

#### 1.18 Oxygen and CO<sub>2</sub> as Biochemical Factors in Respiration

The development of aerobic biochemistry is of fundamental interest to respiratory and evolutionary biologists. Compared with  $CO_2$ ,  $O_2$  is the more important factor in respiration. No permanent change especially of structural nature has been reported on exposure of living tissue to extended hypercapnia. In total anoxia, e.g., when breathing pure N2, a human being loses consciousness within 30 to 40 s and permanent brain damage may occur. On breathing 30%  $CO_2$  in  $O_2$ , a period of dyspnea occurs and loss of consciousness ensues. Complete resuscitation without any permanent damage can occur. Carbon dioxide-induced acidotic narcosis (Sieker and Hickam 1956) has been applied to treat some psychiatric disorders (e.g., Meduna 1950). Unequivocally, O<sub>2</sub> procurement is the primary goal of respiration and CO<sub>2</sub> elimination and hence acid-base regulation are secondary roles. A complex neurohormonal system has evolved especially in vertebrates (e.g., Fedde and Kuhlmann 1978; Ballintijn 1982) to monitor O<sub>2</sub> levels in blood. A reduction of the PO<sub>2</sub> in the ambient air results in a drop of that in blood. Below a certain threshold level, the arterial PO<sub>2</sub> chemoreceptors send impulses to the respiratory center in the brain stem from where ventilatory rate is adjusted. Although the brain constitutes one of the so-called noble organs (the others being the heart and the lung), which are structurally and functionally highly protected against O<sub>2</sub> fluctuations (e.g., Zapol et al. 1979; Freedman et al. 1980), the bloodbrain barrier is highly permeable to respiratory gases. The actual mechanism through which molecular CO<sub>2</sub> acts on the respiratory centers, if it does at all, has been highly debated since H<sup>+</sup> and HCO<sub>3</sub><sup>-</sup> ions exist in equilibrium with carbonic acid (H<sub>2</sub>CO<sub>3</sub>) (e.g., Crone and Lassen 1970; Bradbury 1979). Hyperoxia-induced hypercapnia, which may result in total cessation of ventilation in the trout (e.g., Dejours 1973), in the crayfish (Massabuau et al. 1984), and in the green crab (Jouve-Duhamel and Truchot 1983), shows that at least in water breathers, animals are insensitive to or are incapable of responding to elevated CO<sub>2</sub> in face of O<sub>2</sub> assault. Hypercapnea does not appear to affect ventilation in hyperoxic water breathers (Dejours 1988). However, in the only discordant view, Thomas et al. (1983) reported data to the contrary on the rainbow trout (Oncorhynchus mykiss): marked increase in ventilation was observed in specimens made hypercapnic even under hyperoxia. Except in the fossorial animals which adaptively tolerate high  $CO_2$  levels, whether in moderate hypoxia, normoxia, or hyperoxia, air breathers increase ventilatory rate after inhalation of  $CO_2$ -enriched air: the hypercapnia-induced hyperventilation enhances  $CO_2$  clearance, reduces arterial hypercapnia, and establishes normal pH. In a hypoxic environment, increased ventilation and heart rate, processes which are energetically costly to maintain, are not only unproductive but harmful. Faced with such a crises, reduction of metabolism is a beneficial process to overcome hypoxia (e.g., Hochachka 1988). Hypothermic hypometabolism in a squirrel results in an 88% saving on energy (Wang 1978).

#### 1.19 Homeostasis: the Role of Respiration

Respiration plays an integral part in regulation of the blood pH. To a large extent, respiratory activity is driven to meet the need. The PCO<sub>2</sub> of the arterial blood is the most important factor which governs respiration (Davenport 1974). The goal of regulating breathing is to minimize respiratory work while maintaining stable and optimal levels of respiratory gases and pH. The arterial blood O<sub>2</sub> concentrations are appropriately adjusted and the levels of CO<sub>2</sub> and H<sup>+</sup> ions in blood kept within the physiological range. At the normal steady-state respiratory rate, the alveolar and arterial  $PCO_2$  are adjusted around a value of 5.3 kPa. The respiratory system presents a metabolic servomechanism designed to match pulmonary and metabolic gas exchange rates without altering the internal chemical concentrations of the body fluids. The error-correcting feedback signals are provided by the concentrations of O<sub>2</sub>, CO<sub>2</sub>, and H<sup>+</sup> in the arterial blood. In the higher vertebrates, birds and mammals and to an extent terrestrial reptiles, pulmonary ventilation is used to regulate the rate of CO<sub>2</sub> elimination, a role played by the gills in aquatic breathers. The gas exchanger and the circulatory system effect these processes at rates corresponding with the prevailing metabolic demands. Uptake of O<sub>2</sub> affects body fluid homeostasis especially the acid-base status to a very small extent through release of  $H^+$  ions (after conversion of  $HCO_3^-$  ions to  $CO_2$  on the binding of  $O_2$  to hemoglobin – the Haldane effect) in the gas exchanger (e.g., Davenport 1974; Heisler 1989). In contrast, CO<sub>2</sub> is the main product of aerobic metabolism. It is involved in chemical reactions which affect the acid-base status of the body fluids. In a watery solution, on accumulation, CO<sub>2</sub> is a weak acid and has to be buffered to keep the body fluid pH relatively constant. Through carbonic anhydrase catalysis and chloride shift, the largest fraction of total CO<sub>2</sub> is carried in form of  $HCO_3^-$  ions in both blood plasma and the erythrocytes (Davenport 1974). This roughly constitutes about 90 to 95% of the total CO<sub>2</sub> in blood. At the gas exchanger, the HCO<sub>3</sub><sup>-</sup> ions are converted back into CO<sub>2</sub>, which diffuses out into the external medium (e.g., Perry and Laurent 1990). To ascertain CO<sub>2</sub> diffusion equilibrium, the amounts of carbonic anhydrase in the erythrocytes are much higher in the small than in the larger mammals up to an order of magnitude (Larimer and Schmidt-Nielsen 1960; Lindstedt 1984). Although hypoxia may be brought about by deficiency of O2 in the environment or excessive utilization of it

Table 2. Comparison of blood respiratory features of the trout (*Salmo gairdneri*), the tadpole and adult bullfrog (*Rana catesbeiana*), and the snapping turtle (*Chelydra serpentina*) to show the differences in water and air breathers

	Trout <sup>a</sup>	Bullfrog <sup>b</sup>		Bullfrog		Turtle <sup>c</sup>
		Tadpole	Adult	Tadpole	Adult	
T (°C) PCO <sub>2</sub> (mmHg) <sup>d</sup> pH $HCO_3^{-}$ (mEql <sup>-1</sup> )	20 2.42 7.80 4.63	20 1.95 7.83 4.0	20 13.4 7.90 32	23 4.36 7.80 8.0	23 18.9 7.70 27.5	20 25.2 7.76 49.0

<sup>a</sup> Trout – Randall and Cameron (1973).

<sup>b</sup> Tadpole and adult bullfrog – Erasmus et al. (1970/71) at 20 °C and Just et al. (1973) at 23 °C.

<sup>c</sup> Snapping turtle - Howell et al. (1970).

<sup>d</sup> To convert to kPa multiply by 0.133.

 Table 3. Some physiological characteristics of typical aquatic and terrestrial animals. (Dejours 1988)

Parameter	Aquatic	Terrestrial
Respiration	Skin and/or gills	Tracheae or lungs
PCO <sub>2</sub> /PO <sub>2</sub> <sup>a</sup>	Low ratio	High ratio
$(HCO_{3}^{-1})^{2}$	Low	High
N end products	Mainly ammonia	Mainly urea and/or uric acid and other purine derivatives
Water turnover	High or very high	Low or very low
Temperature	Poikilothermy (most of them)	Homeothermy (some of them)
Locomotion	Swimming	Running, flying

 $^{\rm a}$  PO\_ and PCO\_ designate the difference in PO\_ and PCO\_ values between body fluids and ambient milieu.

by an organism,  $O_2$  has no direct effect on the acid-base status. However, the increased ventilatory rate which befalls an animal in an attempt to supply the required amounts of  $O_2$  may lead to respiratory acidosis due to increased metabolic  $CO_2$  production or respiratory alkalosis due to excessive flushing out of  $CO_2$  from the gas exchanger. There is no systemic difference in pH among the water-, bimodal-, and air breathers. Since the PCO<sub>2</sub> is higher in air breathers than in water breathers, that of the bimodal breathers falling in between, the adjustments in the pH are made by body fluid bicarbonate concentrations which increase with the degree of air breathing (Tables 2,3).

A truly comparative account on respiration must not only endeavor to examine how different organs and organ systems have been refined and integrated for the purpose of exchange of  $O_2$  and  $CO_2$  but must also explore these states and phenomena outside the purview of the so-called model animals. In the class Agnatha, the most frequently studied species is the Atlantic hagfish, *Myxine* 

glutinosa (e.g., Strathmann 1963), in the elamobranchs the commonly studied species are a variety of dogfish, e.g., Scyliorhinus canicula, Saualus sucklevi, Squalus acanthias, and skates; among the bony fish (class: Pisces) studies have been made largely on the subclass Teleosti, the most highly studied species being the cod (Gadus morhua), eel (Anguilla anguilla), goldfish (Carassius auratus), trout (Onchorhynchus mykiss, formerly Salmo gairdneri), and the sea raven (Hemitripterus americanus). In amphibians, the common grass frog (Rana pipiens), European frog (Rana temporaria), and the marine toad (Bufo marinus), all of which are anurans, are taken to be representative of the diverse class Amphibia. Within the class Reptilia, particular interest has been shown in the painted turtles, which fall either into the genus Pseudemys or Chrysemys. The laboratory white rat (Rattus rattus) and the guinea pig (Carvia porcellus) have been used widely among mammals, while in birds, the domestic fowl (Gallus gallus variant domesticus), muscovy duck (Cairina moschata), and the guinea fowl (Numida meleagris) have been used extensively. These few animals, most of which have been selected more for convenience and availability than for any concrete morphological or physiological merit, are far from being genuinely representative of the many taxa in the Animal Kingdom. Most of the discrepancies in the conclusions and observations that abound in comparative biology have arisen from unwarranted extrapolations of observations based on a handful of unrepresentative animals.

# Essence of the Designs of Gas Exchangers – the Imperative Concepts

"To understand completely respiratory adaptations to the environment, it is an implicit but fundamental requirement that we understand how such adaptations evolved, not just how they operate in living animals." Burggren (1991)

# 2.1 Innovations and Maximization of Respiratory Efficiency

Gas exchangers have developed and tractably adapted with the respiratory requirements of whole organisms in different states and habitats. The environmental factors that have profoundly influenced the general phenotype have simultaneously shaped the designs of the gas exchangers. On that account, the functional constructs of the gas exchangers cannot be understood without recognizing both these drives as well as the underlying physical principles that govern organismal biology. Form is a gestalt of structure. The importance of morphology and physiology as investigative approaches towards conceptual understanding of comparative evolution by natural selection cannot be overstated (Cracraft 1983; Duncker 1985; Greenberg 1985; Huey 1987). It should, nonetheless, be cautioned that it is oftentimes possible to mislead these aspects (especially morphology) in accurate reconstruction of phylogeny. For example, the so-called cryptic species (e.g., Bruna et al. 1996) or sibling species (e.g., Mayr 1942) are morphologically identical but genetically different. Such mismatched animals can be utilized to investigate the ecological and evolutionary events and mechanisms which enforce congruent morphologies. Until recently, morphological characteristics were the primary and practically the only means of organizing and classifying animals (e.g., Eldredge 1993; Rieppel 1993). Molecular genetics now offers a powerful means of supplementing morphological observations and validating phylogenetic relationships between different animals (e.g., Sibley and Ahlquish 1990; Graur 1993; Larson and Chippindale 1993; Luckett and Hartenberger 1993; Blair 1994; Hedges and Sibley 1994; Janke et al. 1994; Averof and Akam 1995; Penny and Hasegwa 1997). Considering the remarkable diversity of animal life, the different habitats occupied, lifestyles led, and the disparate metabolic potentials, unless autamorphic features of respiration are emphasized, the uniformity of the gas exchange and transport mechanisms between species is astonishing. A reductionistic (mechanistic) perception of a gas exchanger is that of a construction where an external medium and an internal one are separated by a barrier and a concentration gradient of O2 and CO2 occurs between the two compartments. This overly simplistic concept provides a useful conceptual framework for understanding the fundamental comparative principles of respiratory biology. The ultimate design of the gas exchanger must present those useful features which natural selection has selected, rigorously tested, and genomically conserved.

Energy is integral for building, servicing, and supporting the tissue infrastructure of organisms. To maximize the finite quantity available to them, costeffective designs are essential. This calls for a logical plan of the constituent parts of the body. The most economic designs are those which demand least cost to construct, operate, and maintain while yielding the best possible results. Rosen (1967) deems optimization to be synonymous with "quest for minimum cost", Howell (1983) considers minimization of cost as "a pragmatic replacement for maximization of fitness", and Cannon (1939) termed optimal design simply "the wisdom of the body". Integrated arrangement occurs in the cardiopulmonary system of the fox (Longworth et al. 1989): though having among the highest massspecific  $O_2$  consumption in mammals (3.05 ml  $O_2$  per second per kg), the animal has only an ordinary mass-specific morphometric pulmonary diffusing capacity (Weibel et al. 1983). The high unit O<sub>2</sub> flux across the blood-gas barrier is achieved by a large  $PO_2$  gradient which drives  $O_2$  from the alveolus to the capillary blood. At  $VO_{2max}$  (3.6 ml  $O_2 s^{-1}$  per kg), the fox raises the alveolar PO<sub>2</sub> to 16 kPa by hyperventilating and maintains a low mean capillary PO<sub>2</sub> (12kPa) by having a short capillary transit time (0.31 of a second) due to a high mass specific cardiac output ( $25 \text{ ml s}^{-1} \text{kg}^{-1}$ ). In the horse lung, at VO<sub>2max</sub>, the capillary transit time is 0.4 to 0.5 of a second and the capillary blood is equilibrated with alveolar air after 75% of the transit time (Constantinopol et al. 1989). Among the evolved respiratory steps, the shape of the mammalian erythrocytes gives a good example of the multidimensionality in the enhancement of functional efficiency through morphological and biochemical refinements (Edsall 1972). With few exceptions, e.g., in the camel (Cohen 1978), where the cells are ellipsoidal, in mammals, the resting or minimum energy configuration is that of a biconcave disk (Nikinmaa 1990). The shape affords a high surface-to-volume ratio, with the definitive erythrocyte surface area of about 163 µm<sup>2</sup> being 70% larger than the surface of a spherical cell of equal volume. The camel's erythrocytes are subjected to considerable changes in the osmotic pressure of the plasma when the animal goes for 6 to 8 days without drinking water (Schmidt-Nielsen et al. 1957; Schmidt-Nielsen 1990).

Energy production by oxidative phosphorylation has been an irrevocable and continuous process since the evolution of the  $O_2$ -utilizing pathways in the primordial facultatively aerobic prokaryotic and eukaryotic unicells (Fig. 8). By way of binary division, unicellular organisms and sex cells of higher animals have been transmitted for millennia of generations. Though in finite amounts, these cells require  $O_2$  for subdivision. The  $O_2$  consumption of a single unfertilized mouse egg cell is  $0.37 \,\mu$ l  $O_2 \,mg^{-1}h^{-1}$ , a value which increases to 0.38 after fertilization (Mills and Brinster 1967). In the pea, *Pisum sativum*, mitosis is only completed above an  $O_2$  level of 0.004 of an atmosphere (Amoore 1961). Since the transition from anaerobiosis to aerobiosis, the function of the gas exchangers has remained essentially the same, i.e., taking up  $O_2$  from the external milieu and discharging metabolically produced  $CO_2$  into the same. The survival and adaptability of an organism are dependent on availability of the necessary resources and the capacity of the genotype to manipulate them (Phillipson 1981). In the Metazoa, molecular  $O_2$  is a critical factor in energy production. For such an

important procss, the strategies of procuring  $O_2$  should have been differently optimized very early after the inauguration of aerobic metabolism. The past history of an organism sets the boundaries and the scope of its future development and the present constraints define its prevailing operational latitudes. Working on genetic variation, natural selection shapes and hones biological structures, increasing their fitness. Excessive design and redundancy are expunged, eliminating superfluousity and hence avoiding the unnecessary cost of supporting underutilized capacities. Astute designs are particularly necessary in those organs like the respiratory ones which must remain stable in face of changing needs and circumstances. The cost effectiveness of a gas exchanger can be gauged from the difference between the energy expended to secure molecular  $O_2$  and that required by an organism for sustenance: optimization of the respiratory process endeavors to increase the net balance of  $O_2$ .

The modern gas exchangers are recent products of long-standing evolutionary developments in the ancestral animals. They give us an opening to conceive what may have evolved in the past. The environment has directed and regulated the amplitude and frequency of the adaptive changes which have occurred in the respiratory organs. Environments are highly dynamical systems (Peitgen and Richter 1986; Wainright 1988) which affect the development of organisms in complex and multidimensional ways. Even clonal populations show phenotypic variability. Dubbed Dollo's Law of irreversibility of Evolution (see Meyer 1988), it is considered statistically improbably that an organism can follow exactly the same evolutionary pathway in either direction, i.e., during progressive and retrogressive transformations. This is because environments "evolve" moment by moment. As soon as they are vacated, they are immediately taken over by other animals (e.g., Harvey 1993). The hypothesis frequently called Gause's Principle (see Moore 1990a) asserts that no two species can occupy the same niche. The pneumonate gastropods, a group which displays remarkable subtlety in respiratory strategies (Sect. 5.6.1), constitute an excellent prototype showing the effect environment has on the design of the gas exchangers. With realization or air breathing, the mantle cavity was transformed into a lung (Fig. 59) as the gills (ctenidia) gradually regressed: some species in the group have readopted water breathing through regrowth of similar but not identical structures (Cheatum 1934; Yonge 1952).

#### 2.2 Safety Factors and Margins of Operation of Gas Exchangers

While biological systems largely function at a steady (unstressed) state, occasionally momentary severe conditions call for large adjustments in the level of operation. Highly trained human athletes and some elite animal species such as the horse and the dog (Snow 1985) can increase their  $O_2$  consumption above rest 20to 30-fold (e.g., Seeherman et al. 1981; Jones et al. 1989). To accommodate such adjustments, the relevant biological systems are intrinsically malleably designed and constructed (e.g., di Prampero 1985). In the case of the gas exchangers, maximal gas transfer is effected at the maximum  $O_2$  consumption (VO<sub>2max</sub>) when further increase in exercise does not result in a corresponding increase in O<sub>2</sub> uptake across the gas exchanger (e.g., Taylor et al. 1981, 1987a; Weibel et al. 1987b): supplementary energy is supplied anaerobically, with the accumulating lactic acid eventually inhibiting exercise (Margaria 1976; Taylor et al. 1981, 1989).  $Vo_{2max}$  increases with increasing PO<sub>2</sub> in the inspired air (e.g., Margaria et al. 1972; Welch and Pedersen 1981), transfusion of erythrocytes (e.g., Ekblom et al. 1975; Buick et al. 1984), and endurance exercise training (Saltin and Gollnick 1983; Saltin 1985). The scale of adjustment which enables a biological system to cope with functional loads is said to constitute a reserve capacity (=safety factor). Such capacities could strictly be viewed as "excessive constructions" over and above those necessary for minimum operation (Gans 1979). Biological designs appear to be configured for the worst-case scenarios and when these extremes are exceeded, death or irreparable damage occurs. In the Australian agamid lizard, Amphibolurus nuchalis, intense endurance exercise results in a decrease (rather than an increase) in the maximal O<sub>2</sub> consumption by a factor of 18% and pathological changes in the muscles and joints (Garland et al. 1987). Muscle fiber necrosis has been reported in human marathon runners (Hikida et al. 1983).

In engineering schemes (e.g., Gordon 1978; Petroski 1985), a safety factor is defined as the ratio between the load that just causes failure of a device (i.e., the component's maximal capacity (=strength = performance) to the maximum load that the device is anticipated to bear during operation. Within certain extents, biological systems change harmonically with the fluctuating strains and stresses to which they are subjected (e.g., Gilbert 1988). In composite systems (as are biological tissues), theoretically there should be room for infinite design creativity. Physical (constructional) and biological (phylogenetic, developmental, functional, and ecological) constraints, however, limit the number of possible outcomes (phenotypes) (Thompson 1959; Alberch 1980; Alexander 1985) delimiting the most optimal ultimate configuration(s). Increasing the operational safety margin calls for commitment of greater resources for construction and maintenance. From a perspective of cost-benefit analysis, through refinement brought about by natural selection acting on the phenotype, the measure of the safety factors contrived into different biological systems is aligned with specified needs (Diamond and Hammond 1992; Weibel et al. 1998). Excess functional capacities and extravagant structures precipitate unnecessary costs in form of energy required for maintenance of the infrastructure and operation. For an organ such as the brain, which consumes as much as 25% of the total resting O<sub>2</sub> consumption (Dejours 1990), optimal structure and function with inbuilt malleability is critical. Three-week exposure of rats to hypobaric hypoxia (0.5 of an atmosphere) causes an increase in the brain blood flow by 71% and microvessel density in the frontopolar cerebral cortex by 76% (LaManna et al. 1992). Lack of space in the body may be an important factor in determining the location and the definitive sizes of different organs (e.g., Diamond 1998). The unilateral development of normally paired organs such as the lung in, e.g., snakes and caecilians (e.g., Renous and Gasc 1989), animals with long cylindrical bodies, may be a consequence of a 'crowding-out effect. In bats (Maina et al. 1982a), the capacity to procure the large amounts of O<sub>2</sub> needed for flight has mainly been attained by
development of remarkably large lungs (Maina and King 1984; Maina et al. 1991): compared with the thoracic cavity, the abdominal cavity is remarkably small. The gastrointestinal (GIT) system is simple (Makanya and Maina 1994; Makanya et al. 1995), and the transit time of the ingesta through the GIT is short (Klite 1965; Morrison 1980). Animals evolve just enough of what they need to overcome natural loads (Gans 1988): perfection and elegance are not pursued for their own sake. In what Taylor et al. (1987b) termed shared adaptive effort, unless there are certain underlying constraints, animals prefer to use compensatory combinations of multiple factors to achieve a greater broad-based safety margin of operation rather than few thinly stretched ones (e.g., Maina 1998). Gans (1985) envisaged that "it is the rule rather than the exception that each role utilizes multiple structures and that each structure inevitably supports multiple roles". In different species, biological functions are carried out in a variety of different ways (Brown 1994), indicating that "successful adaptations to particular conditions differ from species to species and from one group to another". This phenomenon has been called multiple realisability by Kitcher (1984) and Brooks (1994). Unless the "coupling", the "working", and the environment in which a gas exchanger operates are well understood (e.g., Connett et al. 1990; Wasserman 1994), respiratory safety margins may be perceived as overadaptations or even redundancies (e.g., Cannon 1939). By way of illustration, in mammals, the morphometric (DLo<sub>2</sub>m) and the physiological (DLo<sub>2</sub>p) diffusing capacities of the lung for  $O_2$  differ by a factor of 2 (Gehr et al. 1978; Weibel et al. 1983). While the difference between the two values may be attributed to intrinsic deficiencies and assumptions in the model used to estimate the  $DLo_2m$ , an aspect discussed by Crapo et al. (1986, 1988) and recently critically reviewed by Weibel et al. (1993), it is thought that the disparity may register a fabricated functional reserve which is utilized during extreme circumstances (Weibel 1984a, 1990; Weibel and Taylor 1986). In a similar study which suggested the existence of a safety factor in the design of gas exchangers, Weibel and Taylor (1986) and Karas et al. (1987a) observed that the athletic animals (dog and pony) had a 2.5-fold greater maximum O<sub>2</sub> consumption  $(Vo_{2max})$  compared with the less athletic ones (goat and calf): the latter group utilized one third of their pulmonary diffusing capacity while the former used three quarters of it at Vo<sub>2max</sub>. The estimations made by Wagner and West (1972) and Hill et al. (1973) indicated that the blood-gas barrier of the human lung, which is normally 0.62 µm thick (Gehr et al. 1978), would have to be increased four to ten times before it became a limiting factor for end capillary  $PO_{2}$  equilibration.

Among animals, different structures, organs, and organ systems possess different functional reserves and redundancies (Alexander 1981, 1982b, 1996; Gans 1985, 1988; Karasov and Diamond 1985; Diamond et al. 1986; Karasov et al. 1986; Toloza et al. 1991; Diamond 1998). The human intestine has a factor of 2 to 2.5, leg bones of mammals 3, breast of most mammals 2, shell of a squid 1.3 to 1.4, and lung of a small dog 1.25. Calculations indicate that at VO<sub>2max</sub>, in contrast to other animals like the pony, dog, calf, and goat (Karas et al. 1987b; Taylor et al. 1987b), the fox uses almost all the capillary transit time for O<sub>2</sub> equilibration, i.e., under extreme effort the fox virtually exhausts the capillary length in O<sub>2</sub> transfer. In the normal placenta at full term, for example, about 21% of the total volume comprises nonparenchymal tissue which is not involved in either gas exchange or metabolite transfer (Aherne and Dunnill 1966). The lung of the shrew, the smallest extant and most highly metabolically active mammal (Morrison et al. 1959; Fons and Sicart 1976; Gehr et al. 1980; Sparti 1992), is elegantly refined for gas exchange. The alveoli are as small as 30 µm in diameter (Tenney and Remmers 1963), the alveolar surface area density is  $2800 \text{ cm}^2 \text{ cm}^{-3}$ , and the harmonic mean thickness of the blood-gas barrier is only 0.25 µm. This gives a mass pulmonary morphometric diffusing capacity of  $0.143 \text{ ml O}_2 \text{ s}^{-1} \text{ mbar}^{-1} \text{ g}^{-1}$  compared with the values of 0.08 and 0.05 in the rat and man, respectively. Based on the geometry and profusity of the pulmonary blood capillaries in the alveolar septa of the lung of the shrew, Weibel (1979, 1984a) observed a resemblance between the honeycomb arrangement of the alveoli of the shrew with the intimately intertwined airand blood capillaries of the bird lung (Maina 1982a, 1988a; Figs. 88, 89). The physiological diffusing capacities of the chorioallatois estimated by Piiper et al. (1980) of  $8.7 \times 10^{-5}$  ml O<sub>2</sub>s<sup>-1</sup> mbar<sup>-1</sup> and that of  $7.5 \times 10^{-5}$  ml O<sub>2</sub>s<sup>-1</sup> mbar<sup>-1</sup> by Tazawa and Mochizuki (1976) compare with the morphometric diffusing capacities of O<sub>2</sub> of a 16-day-old chicken egg of  $8.5 \times 10^{-5}$  ml O<sub>2</sub> s<sup>-1</sup> mbar<sup>-1</sup> estimated by Wangensteen and Weibel (1982). Wangensteen and Weibel (1982) and Weibel (1984a) interpreted this similarity to bespeak the underlying optimization of gas transfer capacity across the chicken egg shell. While this may apply to individual eggs, egg shell conductance differs remarkably between and within species (Tazawa 1987). In the African parrot, Enicognathus ferrugineus, in a single clutch, the shell conductances may differ by a factor of 7 (Bucher and Barnhart 1984). Similar variations have been reported in the turkey, Melleagris gallopavo (Rahn et al. 1981; Tullet 1981), and in the chicken, Gallus domesticus (Tullet and Deeming 1982; Tazawa et al. 1983a; Visschedijk et al. 1985), eggs. Although large differences in the air cell PO<sub>2</sub> and PCO<sub>2</sub> must occur between the low and high conductance eggs, the O<sub>2</sub> consumption during the last stages of incubation are the same in both kinds of eggs (Tazawa 1987). This must allude to occurrence of either underutilized capacity (functional reserve) in the high conductance eggs or possible compensatory adjustments in the blood-gas uptake and transport variables to promote O<sub>2</sub> availability in the low conductance eggs. High conductance eggs should withstand environmental changes in O<sub>2</sub> levels better than the low conductance ones, improving chances of survival in hypoxic environments. The morphometric diffusing capacity of the human placenta (DPo<sub>2</sub>m) was estimated to range between 0.05 to 0.08 ml O<sub>2</sub> per s per mbar (Mayhew et al. 1984). Based on estimations of O<sub>2</sub> consumption in the pregnant human uterus and the PO<sub>2</sub> in the maternal and fetal blood streams, Metcalfe et al. (1967) estimated the physiological diffusing capacity of the placenta (DPo2p) to lie between 0.014 to 0.018 mlO<sub>2</sub> s<sup>-1</sup> mbar<sup>-1</sup>. However, calculations based on diffusing capacities of CO<sub>2</sub> in pregnant women (Forster 1973) yielded a higher value of 0.025 ml  $O_2 s^{-1} mbar^{-1}$ . Based on a mathematical model which simulated the effects of uterine contractions on placental O<sub>2</sub> exchange, Longo et al. (1969) estimated a  $DPo_2p$  of 0.038 ml  $O_2s^{-1}$  mbar<sup>-1</sup>. The present data indicate that the DPo<sub>3</sub>p of the human placenta lies between 0.025 to 0.038 ml O<sub>2</sub>s<sup>-1</sup>mbar<sup>-1</sup>, a value which is lower than the DPo<sub>2</sub>m by a factor of about 2. As may apply to the mammalian

lung, the difference between the DPo<sub>2</sub>m and the DPo<sub>2</sub>p may constitute a reserve which is exploited by the placenta during extreme circumstances (Mayhew et al. 1984). The reserve is brought about by factors such as vascular shunts, placental O<sub>2</sub> consumption, and regional inequalities of perfusion (Metcalfe et al. 1967; Mayhew et al. 1984, 1990). Lack of uniformity in the thickness of the villous membrane may lead to local inhomogeneities of diffusional resistances across the sporadically attenuated barrier (Mayhew et al. 1984; Jackson et al. 1985). Like other gas exchangers, the placenta is a multifunctional organ which must present a compromise design (Sect. 4.7). Though fundamentally constructed for gas exchange, the organ plays important endocrine roles and constitutes an important barrier which protects the fetus from harmful agents and factors in the maternal blood. In some situations, fetotropic viruses such as rubella virus, cytomegalovirus, hepatitis B virus, human immunodeficiency virus, enterovirus, and Theiler's murine encephalomyelities virus are prevented from affecting the fetus (e.g., Alford et al. 1964; Hayes and Gibas 1971; Maury et al. 1989; Garcia et al. 1991; Abzug 1994). It is material to note that the maximum morphometric pulmonary diffusing capacity of the lung for  $O_2$  (DLO<sub>2</sub>m) of 2.38 ml  $O_2 s^{-1} mbar^{-1}$  (Gehr et al. 1978) is about 30 times greater than the morphometric diffusing capacity of the placenta (DPo,m) of 0.08 ml O, s<sup>-1</sup> mbar<sup>-1</sup>. This should constitute an enormous functional reserve which guarantees O2 supply to the fetus under different respiratory conditions and circumstances.

The trade-offs necessary for the development of optimal designs and functional reserves are manifest in many gas exchanges. For example, the completely aquatic lobster, Homarus vulgaris, which shows neither evidence for terrestrial adaptations nor propensity for air breathing when stranded on the beach (a real hazard for intertidal animals) will breath air, maintaining its O2 uptake at nearaquatic levels (Thomas 1954). Facultative air breathing appears to be an acquired adaptive feature which allows solely aquatic animals to withstand a transient, stressful physiological condition. In eggs, high porosity compromises fitness by increasing water loss, but the same process improves it by enhancing the conductance of the egg shell to respiratory gases. In the mammalian lung, while intense subdivision of the pulmonary parenchyma provides a greater respiratory surface area, according to the Young-Laplace relationship  $(P = T r^{-1})$ , where P is the recoil pressure, T surface tension, and r the radius of curvature at the air-liquid interface, the decrease in the diameter (with the resultant increase in the surface radius of curvature of the alveolus) engenders greater surface tension. At an invariable recoil pressure, this increases the disposition of the alveoli to collapse as well as the energy needed to inflate them with air during inspiration (e.g., Wilson 1981; Wilson and Bachofen 1982). On expiration, the alveolar surface tension reaches a value close to zero (Schürch et al. 1985). The design of the lung of the shrew, e.g., Microsorex hoyi which weighs as little as 2.3g (Lasiewski 1963a,b), has been greatly improved and probably driven to the very limit of a functionally operable mammalian lung. On the lung of the minute shrew, Weibel (1979) pointed out that "it may well be that a limit of bioengineering feasibility has been reached at all levels of the respiratory system". The same may apply for the lungs of the 2-g Cuban bee humming bird, and the Thai bumblebee bat (Suarez 1992). To maintain extremely small sizes especially during the larval stages of development, endothermy, a high-cost approach to life, is out of reach of the ectothermic air breathing vertebrates such as amphibians and reptiles. The smallest amphibian is the arrow-poison frog, Sminthilus limbatus, which in adulthood measures only 11 mm from nose to anus. In biological systems, through compromises, concessionary states are established by harmonizing the composite fitness components with the limiting ones. In the fish gills, the two main parameters that can be adjusted to increase transbranchial O<sub>2</sub> diffusion are respiratory surface area and the partial pressure gradient of O<sub>2</sub>: the former can be increased through lamellar recruitment (e.g., Booth 1978) and the latter by increasing ventilatory and perfusion rates. Adjustments of these parameters leads to what has been called osmorespiratory compromise. While improving gas exchange, increasing the gill surface area results in osmoregulatory problems engendered by increased ionic loss and influx of water in the freshwater teleosts or water loss and ionic loading in the marine teleosts (Perry and Laurent 1993). It has been demonstrated by, e.g., Randall et al. (1972), Wood and Randall (1973), and Gonzalez and McDonald (1992), that increased O<sub>2</sub> consumption is accompanied by increase in Na<sup>+</sup> flux, a process that calls for increased energy expenditure. It was envisaged by, e.g., Satchell (1984) and Nilsson (1986), that fish oblige the osmorespiratory compromise by limiting the surface area and increasing the partial pressure gradient of O<sub>2</sub>, a process which increases O<sub>2</sub> flux without provoking problems of ionic transfer. Part et al. (1984) demonstrated nonrespiratory areas in perfused gills of the rainbow trout, Oncorhynchus mykiss, supporting the long-held proposition that fish are able to regulate the surface area of the gills for the purposes of gaseous and osmotic exchange (e.g., Randall 1982; Butler and Metcalfe 1983). To enhance gas exchange, the more energetic species of fish appear to rely more on increasing the respiratory surface area while the less energetic ones utilize hemodynamic adjustment (Perry and McDonald 1993). In the rainbow trout, at rest, only 60% of the gill lamellae are perfused (Booth 1978). During exercise, the blood perfusing the gills is shunted from the less wellventilated basal channels to the more central ones (Nilsson 1986). The complex anatomy of the circulatory system of the gills where the vascular arrangement has been differentiated into respiratory and nonrespiratory pathways (e.g., Gannon et al. 1973; Dunel-Erb and Laurent 1980a; Butler and Metcalfe 1983; De Vries and De Jager 1984) has been associated with the plasticity of the gills for varying the exposure of blood to water. In the European eel, Anguilla anguilla, the volume of the blood in the nonrespiratory vasculature of the gills comprises 5% of the total blood volume in the body (Bennett 1988). Interestingly, though the blood of the dogfish, Scyliorhinus canicula, like that of the other elasmobranchs, is almost isosmotic to seawater (Burger and Bradley 1951) and hence gill perfusion does not affect ionic flux across the water-blood barrier significantly, both kinds of vascular circuits were reported (Metcalfe and Butler 1986). Unlike the teleosts (Pettersson and Nilsson 1979), the gills of the dogfish, S. canicula, lack nervous control of blood flow across the gills (Metcalfe and Butler 1984).

The existence of safety margins in biology is well shown in some physiological processes. Oxygen regulators, for example, operate between two levels, a critical and a limiting one. In the lugworm, *Arenicola marina*, the critical PO<sub>2</sub> is near air saturation at 16 kPa (Toulmond 1975) but the anaerobic processes do not start

until the PO<sub>2</sub> drops to 6kPa (Schöttler et al. 1983). Between the two levels, an organism can maintain aerobic metabolism by rearranging and/or mobilizing different physiological factors. In extreme circumstances, the metabolic level of activity is reduced (Hochachka 1988). In principle, the function of a biological system is facilitated by a structure that is correct for the settings where function occurs. As animals establish themselves in relatively more stable environments, adopt more successful designs and acquire more efficient behavioral, physiological, and biochemical responses to external perturbations, flexibility for genetic change and transformation is gradually blunted. While prokaryotic organisms exhibit remarkable capacity to change metabolism in response to changes in substrate and environmental conditions, cells and tissues of eukaryotic organisms, particularly the higher animals, are considered less responsive since they live in more stable environments (e.g., Golspink 1985). Whether a catastrophic event occurs or not, in all species, the capacity to survive inevitably decays with geological time. Schopf (1984) estimated that the average species' longevity may be as short as 200 000 years. No vertebrate species has avoided extinction for more than a few million years (Carroll 1988). Effective adaptive changes demand selection and refinement of only those features which are favorable in a particular milieu. Apparently, not all features presented by an organism are intrinsically adaptive (e.g, Futuyuma 1986): most organisms present anachronistic features. These features may arise when a particular selective pressure drive stops midstream (e.g., if an environmental stress factor abates) or if a certain structure is diverted to configure a totally different one from that initiated. The common perception that "biological innovations tend to appear soon after environmental conditions become favorable to them" (e.g., Cloud 1974) is a gross oversimplification of the evolutionary process. Strictly, organisms are not passive participants totally subservient to the drifts of the environment: they identify the evolutionary pathways they wish to follow, set their own selective pressures, and actively engage the environment in determining the direction, rate of progress, and nature of change (e.g., Gillis 1991). Only the traits that require a driving force (e.g., natural selection) to establish and impart a performance advantage are adaptations (e.g., Baum and Larson 1991). For example, at the various stages of their evolution, invagination, compartmentalization, and ventilation of the gas exchangers were necessary improvements for respiratory efficiency (e.g., Brainerd et al. 1993). In organisms that were not adequately inventive, these requirements constituted limitations which in some cases stopped any further evolutionary progress. Retaining the buccal force pump and particularly the skin as a gas exchanger apparently consigned the amphibians to water or humidic habitats. The evolution of homologous structures in biology illustrates convergence in solutions to common problems (Gould 1966). In the reptilian lung, to increase the respiratory surface area, the subdivision of the lung was achieved through an inward growth of trabeculae towards the central air space, and in the mammalian lung, the process took place by outward projection of the alveolar septae from the bronchial system. In the latter, while an extensive respiratory surface was achieved, an efficient costodiaphragmatic ventilatory system was necessary to ventilate the more profusely compartmentalized lung.

## 2.3 Engineering Principles in the Design of the Gas Exchangers

Biological processes are intrinsically finite in magnitude and frequency. The boundaries within which operations occur are set by determinate physical constraints and regulated by evolved biological feedback mechanisms. For about 2 billion years, limitations set by diffusion firmly confined animal life to the protozoan domain. This grip was loosened only when  $O_2$  rose above nascent levels and convective processes of transporting respiratory gases from the outside to the proximity of the gas exchangers onward to the tissue cells developed. A wide spectrum of respiratory plans, structures, and strategies based reasonably on common engineering plans has evolved in animals. With certain modifications, the sheet-flow (e.g., Fung and Sobin 1969; Tenney 1979; Farrell et al. 1980; Fung 1993) or tubular design (e.g., Guntheroth et al. 1982), depending on how one visualizes the thin blood conduits, occurs in practically all evolved gas exchangers, e.g., in fish (Figs. 11,49) and crustacean gills (Fig. 13) and in the reptilian



Fig. 11. Cast of the gills of a tilapiine fish, *Oreochromis alcalicus grahami*, showing a gill filament artery, *f*, giving rise to afferent lamellar arterioles, *a*, which supply blood to secondary lamellae, *s*. The constriction of the afferent lamellar arterioles,  $\star$ , are thought to be valves which regulate lamellar perfusion. \* marginal channel;  $\succ$  pillar cells which subdivide the lamellar plate into spaces through which blood percolates. *Bar* 5 µm. (Maina 1994)



Fig. 12. Sheet blood flow pattern in a mammalian lung. *Top inset* The alveoli, *a*, separated in a honeycomb manner by interalveolar septa. *Main figure* Closeup of the interalveolar septum which is formed by two parallel epithelial cell layers,  $\succ$ , between which the blood capillaries are contained. *Bottom inset* Parallel epithelial cell layers,  $\succ$ ; *e* erythrocytes; *a* alveolus; *x* endothelial cells. *Top inset bar* 21 µm; *main figure*, 3 µm; *bottom inset* 3 µm



**Fig. 13.** Sheet blood flow pattern in a gill lamellar of a crab showing epithelial cells, \*, which line the blood space, v. The high energetic demands for ionic exchange are evinced by the abundance of mitochondria in the epithelial cells, p, and the highly amplified basal infoldings,  $\Box$ .  $\rightarrow$ , cuticular lining;  $\succ$ , points where the epithelial cells closely approximate to regulate the rate of blood flow. *Bar* 3.5 µm. (Maina 1990b)

(Fig. 14), mammalian (Fig. 12), and avian (Fig. 29) lungs. The constructional plan comprises thin parallel epithelial cell layers which are joined by connective tissue or pillar-like cell struts. This architectural configuration produces fine channels through which blood flows, spreading out into an extremely thin film over an



Fig. 14. a Semimacerated latex rubber cast of the gas exchange air spaces, *s*, of the lung of the snake, the black mamba (*Dendroapis polylepis*) showing the blood capillary bearing septae,  $\succ$ , **b** A completely macerated double latex cast of the gas exchange region of the snake lung showing the sheets of septal blood capillaries,  $\succ$ , which surround the air spaces, *s. Inset* Critical point dried material showing a septum,  $\succ$ , lining the air spaces. **a** *Bar* 100 µm; **b** 50 µm; *insert* 25 µm. (Maina 1989e)

extensive respiratory surface. This appears to be the only plan which adequately meets the structural requisites for efficient gas exchange. It provides optimal exposure of hemoglobin to the external milieu for maximal gas transfer. The external and internal gas exchange media are brought into as close proximity as possible, interfacing over an extensive surface area in a highly dynamic organ. In the human lung, about 213 cm<sup>3</sup> of the blood in the capillaries is spread over a respiratory surface area of 143 m<sup>2</sup> (Gehr et al. 1978; Weibel 1989), i.e., about 1.5 cm<sup>3</sup> of capillary blood per m<sup>2</sup>, generating an extremely thin film (a sheet) of blood. While the mammalian fetal pulmonary circulation comprises less than 10% of the biventricular cardiac output, pulmonary vascular resistance is higher than the systemic and the PO<sub>2</sub> of blood is low. Soon before birth, the blood flow increases eight to ten times and a transition to low resistance circulation occurs (e.g., Dawes et al. 1953; Cassin et al. 1964; West 1974; Teitel et al. 1987). These changes appear to be influenced by the changes in the PO<sub>2</sub> in blood both in utero and after birth, the process being mediated by changes in the K<sup>+</sup> and Ca<sup>2+</sup> channels in the smooth muscles of the pulmonary artery (e.g., Lewis et al. 1976; Sheldon et al. 1978; Accurso et al. 1986; Cornfield et al. 1994). Pulmonary vascular resistance is a product of pulmonary blood flow and the difference between the pulmonary artery pressure (average 2kPa) and the left atrial pressure (about 0.7 kPa). Even at rest, all the capillaries of the lung are perfused with plasma within 2 min of blood leaving the heart (König et al. 1993). Failure in reduction of the pulmonary circulatory resistance after birth is a consequential clinical problem sometimes termed persistent pulmonary hypertension of the newborn and leads to various neonatal respiratory disturbances (Heymann and Hoffman 1984). In the fish gills, blood flow is slowed down as it percolates through the narrow vascular channels formed by the pillar cells (Fig. 15) improving  $O_2$  uptake by the erythrocytes (e.g., Hughes and Wright 1970; Soivio and Hughes 1978; Farrell et al. 1980; Nilsson et al. 1995). While in transit, the erythrocytes are greatly compressed (Fig. 86) and exposed to the external respiratory medium on all sides across a thin tissue barrier. In the mammalian lung, it takes less than 1s (West 1974; Lindstedt 1984; Swenson 1990) for the erythrocytes to pass through the alveolar blood capillaries, a period within which the erythrocytes are fully saturated with O<sub>2</sub> (Weibel 1984a). Because of the two-phase nature of blood, the erythrocytes and the plasma may take different paths through the capillary network (Okada et al. 1992; König et al. 1993) presumably depending on the resistances offered across different capillary segments (Okada et al. 1992) and the preponderance of leukocyte sequestration in the lumen of the blood capillaries (Perlo et al. 1975; Lien et al. 1987; Hogg et al. 1988; Yoder et al. 1990). In the skeletal muscle, 1 ml of mitochondria relates to about 14 km capillary length and 0.22 cm<sup>3</sup> of capillary blood (Conley et al. 1987). At term, the human placenta contains 45 cm<sup>3</sup> of capillary blood which is spread over an area of about 11 m<sup>2</sup> (Aherne and Dunnill 1966), generating a film of blood 0.41 um thick. In the book lungs of arthropods (Fig. 15), air rather than blood flows through the thin external conduits.

The similarity in the design and construction of the gas exchangers could be attributed to the plausibility that gas exchangers arose from a common ancient



Fig. 15. a Book lungs of the desert scorpion, *Paruroctonus mesaensis*. The lamellae, *x*, are kept apart by vertical struts, **\***. **b** A higher view of the lamellae, *x*, and vertical struts, **\***. **a** *Bar* 40  $\mu$ m; **b** *bar* 13  $\mu$ m. (Farley 1990)

structure, whose basic plan has developmentally been highly conserved. Alternatively, as a process, gas exchange may enforced some basic, invariable structural attributes which all organs had to meet. Biological evolution does not occur by attrition but by parsimonious remodeling of common ancestral plans (Meyer 1988; Lauder and Liem 1989; Atchley and Hall 1991). In the ostariophysans (Order

Siluriformes), the accessory respiratory organs of the air-breathing fish developed directly from the branchial tissue in the immediate concavities above the gills (Hughes and Munshi 1968). In the African catfish, Clarias mossambicus, the suprabranchial chamber membrane is well vascularized (Fig. 66) and, where necessary, the labyrinthine organs developed as outgrowths from the gill arches (Maina and Maloiy 1986; Fig. 65c). Most bimodal-breathing fish have utilized default (already existing) organs like the stomach, intestines, and the anus. Only one siruriid, Pangasius, has evolved a gas bladder primarily for respiration (Browman and Kramer 1985). In principle, whether at molecular, cellular, or organismic level, when change must be made, existing structures are overhauled and improved. Rarely do totally new structures have to develop. For example, by evolving the appropriate enzymes, the evolution of aerobic respiration in the eukaryotes entailed addition of the citric acid (Krebs) cycle onto the original anaerobic (glycolytic) pathway of the anaerobic prokaryotes (Chapman and Schopf 1983). Homologous structures (similar constructional plans enforced by natural selection) illustrate the conservativeness of evolution. Investigating the basis of the different aerobic capacities of the athletic and nonathletic animals, Hoppeler et al. (1987) observed that in all species, the maximal average O<sub>2</sub> consumption of the mitochondria was the same  $(3.4 \text{ to } 4.6 \text{ ml } O_2 \text{ min}^{-1} \text{ ml}^{-1})$  independent of the aerobic capacity of the species: the greater oxidative capacity of the athletic species was brought about by "building more mitochondria of similar kind, rather than by modification of the metabolic rate of the mitochondria". It is, however, recognized that at a certain level of mitochondria-muscle fiber ratio, further increases in the mitochondrial volume density may not only be futile but may compromise muscle function (e.g., Pennycuick and Rezende 1984; Weibel 1985a; Hochachka 1987; Hochachka et al. 1988; Suarez 1992). Once established, morphologic characteristics appear to be retained for indefinite periods of time until there are imperatives for change. For example, mammalian orders have retained a basically similar body form for 50 to 60 million years (e.g., Eldredge and Gould 1972; Carroll 1988). Compared with the total longevity of a species, however, changes take a much shorter period to be effected (Carroll 1988). Only a very small proportion of the genome appears to be directed towards morphological restructuring. The amino acid sequence of 12 varied proteins differs by only 1% between humans and chimpanzees (King and Wilson 1975).

Though argued to the contrary (e.g., Rose and Bown 1984; MacFadden 1985; Chaline and Laurine 1986), evolutionary changes are not progressive nor are they necessarily gradual improvements on earlier designs (e.g., Carroll 1988; Gould 1994; Kardong 1995). The apparent disparity of form between the various gas exchangers has resulted from the singular fact that animals at different phylogenetic levels of development have had to respond to common selective pressures (e.g., Murdock and Currey 1978), in such circumstances, individual solutions being found using different strategies and resources. To a certain extent, phylogenetic plasticity will allow correspondent structures to develop, but in most cases, alternative solutions are pursued. For example, long lungs (which are difficult to ventilate) have evolved in the thin cylindrical animals such as the snakes and the caecilians (Renous and Gasc 1989). The ventilation of such lungs is greatly hindered by locomotion. In lizards, breathing has been uncoupled from locomotion

(Carrier 1984, 1991). On the other hand, in an energy-saving strategy, bats have adopted a 1:1 ratio between wing beat and breathing (Thomas 1987). Clearly, what may constitute a constraint in one animal may be beneficial in another. Amidst these shifts, some structures which may appear to be of no evident biological value to organisms may evolve (e.g., Bock and von Wahlert 1965; Kimura 1983; Pierce and Crawford 1997).

In biological systems, structure encompasses the qualitative and quantitative characteristics of the constitutive components and their geometric features and arrangements. By altering the proportions, positions, and configurations, new polarities and states are created and different functional states are established. The components of complex structures constitute an integrated pattern of upand downregulation of diverse functional capacities. For example, the arrangement of the mineral crystals of CaCO<sub>3</sub> in the eggshell determines the porosity of the shell and hence its diffusing capacity for O<sub>2</sub>, a feature which, in turn, ensures proper development of the embryo. The permeability of the fish's swim bladder to gases depends on the orientation of guanine crystals in the bladder wall (Lapennas and Schmidt-Nielsen 1977). Swim bladders of fish which operate at depths greater than 1000 m have a greater concentration of guanine per unit area of the wall (Denton et al. 1970). The principles of homology and analogy are fundamental to understanding the correlation between structure and function as modified by natural selection and effected by the process of adaptation. Those structures which undergo irreversible deconstruction become vestigial and eventually disappear while some may be commissioned to perform roles different from those for which they were initially configured. Such adaptive traits were called exaptations by Baum and Larson (1991). For example, the surfactant evolved in the ancestral piscine lungs (e.g., Todd 1980) mainly to protect the epithelial surface (Liem 1987a). However, with the development of the more complex lungs in the tetrapods (where the buccal force pump was no longer adequate to ventilate such lungs), compartmentalization of the gas exchanger was necessary to enhance respiratory efficiency. With this modification, the surfactant (Sect. 6.9), by increasing the lateral stability of the phospholipid layer (Cochrane and Revak 1991), assumed the important role of reducing surface tension (e.g., Wilson 1981; Golde et al. 1994). In the anaconda, Eunectes murinus, the surfactant lining occurs in the alveolated and the succular parts of the lung (Phleger et al. 1978). While in the protochordates the pharygeal region serves as a filter-feeding apparatus (Sect. 6.10.1), in the chordates, with the addition of a respiratory role, the trophic one was phased out. The complex relocation, erosion, and eventual disintegration of some of the aortic arch blood vessels (which serviced the gills) on the formation of the lungs, is another example of such drastic transformations. Among vertebrates, the pulmonary arteries make their first entry in the Dipnoi (lungfishes) as branches of the sixth pair of aortic arches. Early in the development, the blood vessels supplied the swim bladder which is thought to have given rise to the lungs (Sect. 6.2). Only the necessary early structures (e.g., some of the blood vessels of the branchial arches) were retained as the primal gill arch blood vessels were reconfigured. The degeneration of one lung in, e.g., snakes (Ophidia) and the caecilians (Gymnophiona), is thought to have been one of the sacrifices they had to make to develop thin, limbless, cylindrical bodies which are important for slithering through confined spaces.

Unlike human-made machines, which are configured to carry out specified functions, biological structures are dynamic, multifunctional, composite entities designed to continuously absorb and respond to the fluidity of the external pressures of natural selection. In what de Beers (1951) called mosaic evolution, the different parts of an animal are variably affected by natural selection. Needs, to a greater extent than phylogenetic level of development, dictate the direction, nature, and magnitude of adaptive change. For example, the similarity of the lung-air sac system of birds (Sect. 6.7.5) and the tracheal-air sac system of insects (Sect. 6.6.1), animals separated by over 200 million years of evolution, indicates a morphological convergence for flight. Furthermore, compared with the respiratory system of birds and that of insects, the lung of the human being at the acme of evolutionary development is not as efficient. In the benign evolution of the Homo sapiens, natural selection appears to have targeted the nervous and musculoskeletal systems, leading to development of complex mental capacity, bipedal locomotion, opposable thumb, articulated sound (speech), etc. In an adult human being, the brain utilizes 25% of the overall resting O<sub>2</sub> consumption. While it would be anticipated that these developments would impart particular specializations, especially with respect to the mechanisms and control of breathing, the pulmonary system appears to have been disregarded during these transformations as long as it was adequate to support the ongoing changes. Except for the Hering-Breuer reflex, which appears to be much less well developed, the human respiratory characteristics are similar to those of all other mammals (Dejours 1990).

# 2.4 Scopes and Limitations in the Design and Refinement of the Gas Exchangers

Animals are aphoristically said to be structurally and functionally well constructed to meet the adversities of life (e.g., Olson and Miller 1958; Frazzetta 1975). Though easily conceivable, dedicated biological engineering is not easy to experimentally test, empirically prove, and convincingly demonstrate. This is largely due to our anthropocentric approach and misconception that evolutionary change is determined and driven by the same rules (to serve the same purposes) as human technological innovations (e.g., Basalla 1989). Whereas for human insight advancement means improvement and improvement means more sophisticated products, in nature, changes are generally highly resisted. If they occur, they are strictly survival-oriented and are configured specifically around the existing structures and prevailing conditions to engage known loads. Paleontological studies do not support the popular belief that through the evolutionary continuum, organismal design and function have undergone appreciable refinement and complexity (e.g., Rudwick 1964; Hickman 1987; McShea 1991): the ancient organisms were no less exquisitely designed than the modern ones. Indeed, some ancient structures of now extinct animals seem to have dealt with

complex problems that are unsolved in the present animals. Stipulation for originality in technical inventions is purely a property of human ambition and competitiveness: evolution advances without purpose or direction, resulting in unpredictable changes. Adaptation entails cumulative selection of innovations that build on top of primeval ones. Mosaic evolution, i.e., selective degeneration combined with progressive specialization, e.g., in the acoustic adaptations of the fossorial rodents (e.g., Nevo et al. 1982; Heth et al. 1985) is an occurrence which shows nature's conservativeness and yet quest for optimization. It has been suggested that human technological progress may be proceeding 10 million times faster than natural evolution (Arthur 1997). Experiments on guppies, Poecilia reticulata (Reznick et al. 1997), however, demonstrated that evolution can occur very rapidly: in a 4-year observation, the rate of change of certain features was some 10000 to 1 million times faster than the average rates estimated from fossil record. Thompson (1911) pointed out (but see counterviews by, e.g., Cody 1974 and Howell 1983) that "biology does not necessarily make progress toward perfection by mechanical analysis of changes that go on in living bodies". In nature, even where radical changes such as mutations occur, designs are altered through reconfiguring existing structures or enforcing new roles. Figuratively speaking, animals appear constantly to "reinvent the wheel" (though they have yet to evolve a real one!) as they look for the most relevant and least traumatic solutions to the demands prevailing in their ecological settings. Future developments are not anticipated. By combining the different assets they have gathered along the way, new "adaptive functional complexes" of high selective value are configured, conferring greater survival potential to an organism. S.A. Kauffman (cited in Ruthen 1993) asserts that "by selecting an appropriate strategy, organisms tune their coupling to their environment to whatever value fits them best". Comparative biology reveals the parameters in the "primitive" life forms which have been conserved during the evolvement of the complexity that characterizes the most "advanced" kinds (Fishman 1983), defining in broad terms the pathways followed and the strategies adopted in the quest for survival and self-perpetuation.

Establishment of optimum states calls for sound analysis of the alternative strategies, evaluation of the costs incurred, the benefits which accrue, and the difference between the level of operation with the theoretical maxima. In bats, for example, a typical mammalian lung has been structurally and functionally refined to supply the enormous amounts of O<sub>2</sub> needed for flight (Thomas 1987; Maina et al. 1991); (Sect. 6.7.4.1). Unequivocally, this shows that the lung-air sac system of birds and the tracheal-air sac system of insects, the gas exchangers which have evolved in the only other two volant taxa, are not prerequisite respiratory designs for flight. While the structural parameters are fixed, the functional ones are more flexible. Bats have shrewdly utilized combinations of these parameters and promoted the efficiency of a plainly inferior gas exchanger to rival and in a manner equal that of the distinctly superior bird lung (Maina 1998). After sojourn at high altitude (e.g., Bard et al. 1978; Heath et al. 1984; Durmowicz et al. 1993), factors such as ventilation, hematocrit, hemoglobin concentration, erythrocyte count, and blunted hypoxic pulmonary vasoconstrictor response change (some within a matter of hours or days) to avoid cellular hypoxia and to correct for adverse changes in the blood pH. The structural parameters, e.g., the thickness of the



Fig. 16. Schematic drawing showing the resistance barriers across which  $O_2$  diffuses in a water breather and the principal structural features, namely the barrier thickness, *t*, and the respiratory surface area, *s*, which influence the diffusion process. The partial pressure gradient of  $O_2$  ( $\Delta PO_2$ ) decreases with the diffusion distance

blood-gas barrier and respiratory surface area, take months to change (e.g., Weibel 1984a). Whether in a water- or an air breather,  $O_2$  procurement depends on parameters such as the diffusional distance, respiratory surface area, physical permeative properties of the respiratory barriers, and functional properties such as ventilation and perfusion states (Fig. 16).

For optimal energy production, in organisms, gas transfer is aligned with the metabolic needs. The correlation between an animal's environment and its respiratory needs is a very intimate one. Whereas O<sub>2</sub> need at rest is moderate, during exercise, e.g., flight, it increases tremendously (Weis-Fogh 1967; Tucker 1972; Thomas 1987). When exposed to a hypoxic environment, to support their normal metabolic activities, organisms still need to extract and transfer to the tissues the same quantity of  $O_2$  as in a normoxic one. The respiratory system must be designed with adequate flexibility to accommodate the extreme demands which may occasionally be loaded on the organism (e.g., Maina 1998). Coordination of external factors (i.e., those involved in the procurement) and the internal ones (i.e., those involved in the uptake, transport, and distribution) ensures a satisfactory supply of  $O_2$  to the tissues. In the event of hypoxia, it is the external factors rather than internal ones which become limiting. The high metabolic capacities, especially in the endothermic homeotherms, were achieved through evolution of more efficient means of O<sub>2</sub> procurement. This entailed development of features such as double circulation, greater blood pressure, higher tissue capillarity, higher mitochondrial volume density, and larger hemoglobin and myoglobin concentrations.

#### 2.5 Optimal Designs in Biology and Gas Exchangers in Particular

#### 2.5.1 Symmorphosis: the Debate

Form and function are inextricably interrelated. Indeed, it is axiomatic that animals should be constructed reasonably in order to be able to efficiently carry out the essential activities of life. The perception of a form-function correlation alludes to some advantages for the animals that possess the attribute (Gans 1988). Vogel and Wainright (1969) put it that "function without structure is a ghost; structure without function is a corpse" and Trivers (1985) bluntly stated that "organisms are designed to do something". The concept of optimal design of biological structures is based on the recognition that natural selection continuously regulates every aspect of structure and function. Though perceptively simple to conceive, the actual manner in which structure and function actuate each other is not that simple to read. This is particularly well known from the disappointing attempts to conceive form from phylogenetic reconstruction based on fossils (e.g., Cutler 1995). A correlation between structure and function can only be meaningfully made if both structure and function are simultaneously observed and quantified.

Based on their "firm belief that animals are built reasonably" and their confidence that "structural design is optimized for the role it plays", the rationale of structure-function interdependence for adept performance of the lung was formulated by Taylor and Weibel (1981) and Taylor et al. (1987b) and termed symmorphosis. Symmorphosis was defined as a state of structural design commensurate to functional needs resulting from regulated morphogenensis, whereby the formation of structural elements is regulated to satisfy but not exceed the requirements of the functional system. Direct morphological and physiological data on the mammalian lung and the skeletal muscle mitochondria, the primary  $O_2$  sink, were adduced to show that the gas exchanger was reasonably well constructed to meet the metabolic needs. However, scaling the morphometric diffusing capacity (DLo<sub>2</sub>m) and maximum O<sub>2</sub> consumption (VO<sub>2max</sub>) with body mass indicated a paradox (Gehr et al. 1981; Weibel et al. 1983; Weibel 1989): VO<sub>2max</sub> and DLo<sub>2</sub>m scaled differently with body mass - VO<sub>2max</sub> with a scaling factor (mass exponent b) of 0.8 and DLo<sub>2</sub>m with that of unity. This suggested that large animals have a DLo<sub>2</sub>m far in excess of their metabolic requirements and called in question the validity of what was initially conceived to be an encompassing concept (Taylor et al. 1987b; Weibel et al. 1991). Considering these observations, Taylor et al. (1987b) concluded that the principle of symmorphosis is only partially satisfied in the lung. This departure from the expected perhaps serves as a timely reminder of the intrinsic complexities of the biological systems. Such entities are multidimensionally regulated and when integrated, singular normally inconsequential events are greatly accentuated. Mathematical models with their intrinsic assumptions cannot possibly inclusively describe the arrangements and define the capacities of such complex matter. In complete departure from the Newtonian predictability in physical sciences, the cause and event interactions in complex systems are not necessarily linear and the output may not be anticipated. In fact, the predictive power in the physical sciences is only good for simple and

single events operating under very narrow circumstances. Where multiple and possibly conflicting parameters interact and compromises and trade-offs permutate in space and time with unpredictable probability, except for within very broad limits, there may not be a single best solution to a problem. Maynard-Smith (1968) observed that "we rarely know enough about laws governing the components of biological systems to be able to write down the appropriate equation with any confidence". Wilkie (1977) pointed out that "biological systems are to some degree mixed regimes whose behavior is affected by physical quantities other than those assumed to be dominant". Compared with the physical structures, biological structures are complex and possess intrinsic hierarchical stratifications with certain resolutely restricting boundary conditions (Bartholomew 1982a; Brown 1994). Living entities are more difficult to conceive mentally while physical ones are generally simple enough to be adequately expressed in mathematical terms. Despite the limitations of applications in biology, mathematical reasoning is fundamental in defining the logical consequences of a situation at intellectual depths and expressing and understanding what is theoretically contemplated or observed.

Symmorphosis is not a totally new concept. Actually, the idea is as old as science itself: the great Greek philosopher Aristotle (3 B.C.) observed that 'nature does nothing to no purpose". The concept of symmorphosis corresponds with the theory of optimality (e.g., Rosen 1967; Maynard-Smith 1978; Alexander 1982b; 1996; Kramer 1987) and the principle of minimum work (e.g., Murry 1926; Mandelbrot 1983; Rossitti and Löfgren 1993a) which contend that, operating on finite resources, self-regulating systems strive to optimize their operations. Through cost-benefit trade-offs, some factors are subordinated while others are augmented as an optimal match between structural design and functional performance is established (e.g., Dullemeijer 1974; Wainright et al. 1976; Barel et al. 1989; Barel 1993). Remarking on the variability of the thermostability of proteins, an observation which may apply to all levels of biology, Wedler (1987) observed that "there is no single or universal mechanism (in nature)... it appears that nature has utilized every imaginable means, in different combinations, to achieve the same end". Evolution by natural selection is a continuous process of cultivating optimization: ways and means of enhancing fitness are aggressively pursued. Of all forms of life which have evolved during the about 4 billion years life has existed on Earth, about 99.99% are now extinct (e.g., Pough et al. 1989). Extant organisms represent a very narrow range of the designs which have developed during the long period of trial-and-error evolutionary experimentation and even less of the infinite theoretical possibilities. The history of life occurs in form of a complex, sporadic pattern of past evolutionary expansions and contractions as assemblages of organisms in a particular time frame respond to the prevailing conditions (Gans 1988): useful functions are intensified and improved while the less beneficial ones are relegated or even eliminated. For instance, the evolution of the complex, more efficient tetrapod lung entailed suppression and eventual elimination of the hydrostatic role of the ancestral piscine lung (Joss et al. 1991), creating a favorable situation for transition from aquatic to terrestrial life (Liem 1987a).

The structures which comprise the gas exchange pathway, namely the lung, heart, blood vessels, and mitochondria, have determinate functional capacities (Weibel 1984a, 1987). Pulmonary design is coadjusted with functional needs to supply O<sub>2</sub> at rates consonant with prevailing demands. Since animals have a definable capacity to bear natural loads (be they from within or without), overdesign and redundancy of biological systems is costly and untenable. An optimum structural design is one which requires the least metabolic (energetic) cost to work and sustain (e.g., Thompson 1959; Rosen 1967). Such a design may operate at an optimum level, though it may not necessarily be optimally adapted. While endeavoring to optimize their operations, biological systems appear never actually to attain an optimum condition (e.g., Gans 1983; Lindstedt and Jones 1988). Optimization is a process which appears to be continuously aspired for. There are, however, assertions that such states exist. For example, in their geometry, the cerebral arteries (Rossitti and Löfgren 1993b) and the medium pulmonary airways (Hammersley and Olson 1992) are thought to be optimized in respect to the ratio respectively between blood/air flow and blood vessel/airway radius minimizing resistance. Intuitively, a definitive optimal state should never be consummated. Considering the dynamical nature of environments and the plasticity of biological systems, optimum states cannot possibly be satisfied. As a matter of fact, it would be imprudent to achieve such a condition since an organism's capacity to respond to further changes would be undermined. Joyce (1997) asserts that "evolutionary search is a parellel process in which every individual in a population has an opportunity to give rise to novel variants with increased fitness": this is thought necessary in order to minimize the real possibility of the population "getting trapped in an evolutionary blind alley from which further improvements in the fitness are precluded". In constant pursuit of optimization, the best level of "workmanship" possible under certain constraints and opportunities is established, affording the best level of fitness. In this context, Gans (1983) and Lindstedt and Jones (1988) define optimization as "the best level of improvement possible under the evolutionary circumstances". Williston's law, which asserts that serially repeated structures are reduced to fewer sets of undifferentiated organs, illustrates an evolutionary process of optimization where through a rigorous process of trial and error superfluous features are pruned, leaving the bare essentials for life. The quest for optimization is a universal process which encompasses all self-organizing structures. Howell (1983) pointed out that "optimization does not belong exclusively to biologists, but to any discipline where the subject has some effect on fitness".

As a philosophical definition of the correlation between structure and function in the design of biological systems, the concept of symmorphosis is heuristically useful and as a working hypothesis implicitly valuable. It constitutes a useful tool for bridging the different disciplines of biology. The breakdown of the concept at some levels and in certain cases (e.g., Gehr et al. 1981; Weibel et al. 1983) is a clear manifestation of the vicissitudes of evolution. Paradigms of pulmonary structural-functional reciprocity occur in the nonmammalian gas exchangers but refinements to the extent envisaged in the dictum of symmorphosis cannot be proved owing to paucity of data. Birds, in particular, exhibit remarkable speciesspecific variations in the degrees of pulmonary structural refinements. The lungs of the nonflying species, e.g., the domestic fowl, Gallus gallus variant domesticus (Abdalla et al. 1982), emu Dromaius novaehollandiae (Maina and King 1989), and the penguin, Spheniscus humboldti (Maina and King 1987), have subordinate parameters compared with those of the more energetic ones, e.g., the passeriforms (Maina 1984) and the hummingbirds (Dubach 1981) (Sect. 6.7.5). Bats, the only volant mammals (Thewissen and Babcock 1992), have remarkably highly specialized lungs (Maina et al. 1982a; Maina et al. 1991) which enable them to provide the large amounts of O, needed for flight. Natural selection does not seek efficiency for its own sake but only in so far as it can improve fitness for survival and ensure self-perpetuation. Interestingly, in the construction of animal bodies, gravity has spared the very small. The structural integrity of a body which takes a jump from a height of 10 m is not threatened until the animal attains a size of a puppy or is larger (Went 1968). Interspecific deviations of the aspects which are envisaged to constitute safety factors, redundancies, or limitations exist: they may explain why different biological systems manifest different physiological threshold pressures.

#### 2.5.2 The Operative Strategies for Optimization in the Gas Exchangers

The ervthrocytes are packages which are said to provide an optimum environment for the function of the hemoglobin (Horvath and Borgia 1984). They present an excellent example of the "multiple tradeoffs" or "shared adaptive effort" as termed by Taylor et al. (1987b) of the morphohological, physiological, and biochemical processes involved in enhancing O2 uptake and transfer (e.g., Edsall 1972). In vertebrates, the hemoglobin concentration of about 5 mM is close to the saturation mark (e.g., Riggs 1976). Acclimatization to hypoxia, e.g., at high altitude (Petchow et al. 1977; Wood and Lenfant 1979), and increase in aerobic capacity (Carpenter 1975; Balasch et al. 1976) are largely accompanied by an increase in the hemoglobin concentration so as to increase the O<sub>2</sub> carrying capacity of blood. In an exercising horse and steer, as O<sub>2</sub> consumption increases, circulating hemoglobin concentration rises, thereby enhancing the delivery of O<sub>2</sub> to the tissues through the circulatory system (Jones et al. 1989). While compared with the little auk, Plautus alle, the Arctic tern, Stema paradisaea has a lower hematocrit due to smaller red blood cells and hence a lower total repiratory surface area of the RBCs, the hemoglobin content per unit area of the RBCs in the two species is similar (Kostelecka-Mycha 1987). Upward regulation of the hematocrit (Hct) in particular serves a useful purpose only up to a certain point, when the returns start to diminish. The flow of blood is exponentially related to the blood viscosity, which in turn relates with the hematocrit. Change in the blood O<sub>2</sub> carrying capacity through increase in the Hct contracts extra cost of pumping work on the heart muscle. With a Hct of 65%, the blood viscosity of the elephant seals is three times that of the rabbit, which has a Hct of 35% (Hedrick and Duffield 1986). Birds with higher Hct such as the pigeon (Hct = 52%) adaptively have low plasma protein concentration in blood to maintain the viscosity of blood

at a level similar to that of the birds with lower Hct, e.g., 32% in the domestic fowl (Viscor et al. 1984). There are diverse adaptations involved in optimizing gas exchange in blood. While the hemoglobin-hematocrit-blood viscosity intercourse appears to suggest an exhaustive process of refinement (e.g., Clarke and Nicol 1993), other evolved innovations continue to be discovered. For example, during hypoxia and/or hypercarbia, fish erythrocytes swell, resulting in intracytoplasmic dilution of the hemoglobin and nucleoside triphosphates, increasing the O<sub>2</sub> affinity (Lykkebone and Weber 1978). The rheology of the erythrocytes (Merrill 1969; Schmid-Schönbein 1975) and the hemodynamics of the blood flow in the blood capillaries can improve the delivery of O<sub>2</sub> to the tissues (Zander and Schmid-Schönbein 1973; Kon et al. 1983; Nilsson et al. 1995). This mainly occurs through diminution of the diffusion boundary layer of blood plasma around the erythrocytes and possible intracellular physical agitation of the hemoglobin molecules (Skalak and Branemark 1969; Secomb 1991; Maeda and Shiga 1994). The nucleated erythrocytes, e.g., those of birds, are more resistant to shear deformation than the nonnucleated mammalian ones (Gaehtgens et al. 1981; Nikinmaa and Huestis 1984; Nikinmaa 1990). The shear modulus of the membrane of the nucleated erythrocytes, which have a better-developed cytoskeletal system, is 5 to 15 times higher than that of the nonnucleated mammalian erythrocytes (Waugh and Evans 1976; Chien 1985). Decreased deformability causes the erythrocytes to be trapped in organs like the spleen, lung, and liver (Groom 1987; Simchon et al. 1987), reducing the microcirculatory blood flow in the tissues. Although experimentally more resistant to deformation than the nonnucleated erythrocytes, the nucleated erythrocytes are more deformed as they pass the blood capillaries (e.g., Chien et al. 1971; Akester 1974). Despite having a diameter 30% greater than the human erythrocytes, the fish erythrocytes are as deformable as the human ones (Hughes et al. 1982; Hughes and Kikuchi 1984). Deformation of the erythrocytes reduces the apparent viscosity of the blood (Chien 1970) and provokes convective mixing of blood (Bloch 1962). Contrary to anticipation, the size of the erythrocytes in mammals does not affect the lung diffusing capacity for O<sub>2</sub> (Betticher et al. 1991): small erythrocytes have a greater surface-to-volume ratio, thinner plasma boundary layer (Vandegriff and Olson 1984), and shorter intracellular diffusion distance (Yamaguchi et al. 1988), factors which would be expected to favor  $O_2$  transfer. However, since a sphere is not deformable unless it is squeezed, the less spherical an erythrocyte, the more malleable it is without changing the surface area. Betticher et al. (1991), conceptualized that as the small erythrocytes are more spherical, the plasma around them is less well mixed than in larger cells, thus decreasing O<sub>2</sub> transfer into the cell. The elephant seals are thought to increase O<sub>2</sub> storage at the expense of aerobic scope which results from viscositylimited perfusion efficiency (Hedrick and Duffield 1986). The normal hemoglobin concentration of 150 g/l of the mammalian blood is the value at which maximum amount of  $O_2$  (210 cm<sup>3</sup>), equivalent to the relative atmospheric concentration of the gas (Davenport 1974), is transported with least circulatory work (Schmidt-Nielsen 1984). Adaptive increase in the hemoglobin concentration at high altitude and in small animals such as shrews (Ulrich and Bartels 1963) and bats (Jürgens et al. 1981), values which may respectively be as high as 170 and 244 g of hemoglobin per l may set the operational limit to which hemoglobin and hematocrit levels may be usefully applied to improve respiratory efficiency. In the tracheolar system of insects (Sect. 6.6.1), the mean free path of  $O_2$  molecules in air (i.e., the average distance a molecule travels in air before colliding with another) which is about  $0.008 \mu m$  (Pickard 1974) sets the limit of the smallest tracheolar diameter. Due to the remarkable variability and functional lability of the respiratory pigment system (Sect. 2.8), among the evolved sections of the integrated gas exchange system, the respiratory pigments appear to be the most recent addition and hence the least conserved parts. As opined by Jones (1972), the more ancient factors such as the morphological features of the gas exchanger and the vascular system, which are less flexible, would to a greater extent constitute limiting factors in respiration.

Metabolically, animals operate at two extremes, a steady-state resting condition and under maximal stress. Contrived reserves enable them to harmonically perform under the two different sets of conditions. To ensure structural and functional integrity, the body systems must be designed to withstand the maximal stresses they are likely to be subjected to. Digressing a little from the gas exchangers, in all evolved biological structures, the spider's dragline perhaps best illustrates the process of optimization in biology. A spider's dragline, a tool which determines survival by providing means for procurement of food and escape from predators (a lifeline in the true sense of the word!) is a multiphase material which consists of double filaments (Vollrath 1992). Individually the filaments can support the weight of the animal (if one is accidentally cut). A single line will break at a stress equivalent to that generated by about six times the spider's weight (Osaki 1996). Activities such as movement, jumping, and rapid descent and ascent (when greater stress is exerted on the dragline) must be accommodated in the design. The elastic-limit of a dragline gives the maximum safety (a safety coefficient being the ratio between the mechanical strength of a dragline and a spider's weight) for supporting a spider's weight. The mechanical properties of the spiders' draglines have been refined over their 400 million years of evolution to this level of sophistication. Turning to the respiratory organs, in the rainbow trout and the lingcod, only about two thirds of the secondary lamellae are perfused at rest (Booth 1978; Farrell et al. 1979; Nilsson et al. 1995) but the fish can increase O2 consumption during exercise eight to ten times (Jones and Randall 1978). Hypoxia, e.g., in warm, bottom, and standing waters and during low tide (in intertidal animals), e.g., in the blue crab, Callinectes sapidus is avoided by increased ventilation and perfusion of the gills (Tuurala et al. 1984; DeFur and Pease 1988), lamellar recruitment (Booth 1979; Farrell et al. 1980), shortening of the thickness of the water-blood barrier (Farrell et al. 1980; Soivio and Tuurala 1981), and effecting changes in the blood O<sub>2</sub> binding characteristics (Jensen and Weber 1985). In extreme circumstances, especially where hypoxia is accompanied by high temperatures, the metabolic rate may drop to conserve O<sub>2</sub> and/or the fish may relocate to less poorly oxygenated waters (Jones 1952; Whitmore et al. 1960). Cephalopods, e.g., Loligo and Octopus, can increase their O<sub>2</sub> consumption by a factor of 2 to 3 from rest (O'Dor 1982; Wells et al. 1983). Squids which live in cold, deep seawaters have a large gill surface area and a thin water-blood barrier compared with those which live in shallow waters (Roper 1969; Madan and Wells 1996). The large gill surface area in the pelagic cephalo-

pods may enable them to cope with the hypoxia prevalent in their habitat and could give them a competitive edge over fish (Madan and Wells 1996). This view was, however, disputed by Seibel and Childress (1996) on the basis of the fact that many fish sympatrically coexist with the squids and in some cases may even displace them. The cephalopod heart mainly works aerobically, relying largely on amino acids as substrates for oxidative metabolism (Hoeger and Mommsen 1985). By retracting into the shell, the bivalve mollusk, Pholas dactylus, can effect a complete shutdown of the posterior parts of the gills and, at maximal extension, the gills may be three times as long as the shell itself (Knight and Knight 1986). Bats and birds increase their O<sub>2</sub> uptake from rest to flight by a factor of 10 to 20 times (e.g., Thomas 1987; Butler 1991b) and insects by as much as 120 to 400 times (Weis-Fogh 1967). In gills, ion pumping and gas transfer can be regulated by modulating gill ventilation (Randall et al. 1972; Wood and Randall 1973). The mammalian lung maintains a remarkable excess of diffusing capacity up to a factor of 2 (Weibel 1984a). Even under hypoxic conditions, goats attain maximum O<sub>2</sub> consumption and only the smallest mammals use all of their diffusing capacity under such conditions (Taylor et al. 1987a).

Those animals which have relatively greater  $O_2$  demands during exercise, e.g., birds and insects, are in most respects endowed with more efficient respiratory designs than the human alveolar lung. In a radical departure from the norm, in some respects, it could be argued that, compared with the other organ systems, the respiratory processes have been less sensitive to phylogenetic changes. Adequacy for specific metabolic needs in a given habitat appears to be the primary factor which has determined the refinement and construction of the gas exchangers. Fluid designs have made animals at completely different levels of phylogenetic development coexist in the same general habitats by varying their metabolic demands to meet individual requirements. From the perspective of respiration, animals have had few choices. There are only two naturally occurring respirable fluids – water and air – and three livable spaces, namely the aerosphere, the hydrosphere, and the lithosphere, i.e., the livable superficial part of the Earth's crust generally called soil.

### 2.5.3 Symmorphosis and Optimization: are they Logical Outcomes of Evolution?

The concepts of optimization and symmorphosis may have been accepted by biologists basically for their instinctive appeal to simple intuitive logic. Recently, the concepts have been ardently debated in depth (see Diamond 1992; Weibel et al. 1998). Conceived as perfect matching between structure and function, symmorphosis has been questioned from phylogenetic and developmental considerations (e.g., Gould and Lewontin 1979; Lewontin 1979; Garland and Huey 1987; Gans 1988). Optimization for particular conditions and circumstances, though logically desirable, curtails the potential and the range of adjustments which organisms can mobilize to counter external assaults. As a practical rule, every adaptive refinement for a particular circumstance that an organism or organ system undergoes leads to exclusion of capacity to take on others. Moreover, what may be an optimal solution for a particular situation at a particular time may be a limiting factor in another. Moore (1990a) asserts that "organisms cannot be complete specialists and complete generalists at the same time". Natural selection refines each and every lineage towards effective utilization of a specific quota of the available resources. Faunal structuring, partitioning of dietary and habitat resources, and the different morphological specializations acquired for procuring them minimize competition, providing optimal survival conditions. Animals which face similar ecological and developmental constraints evolve similar features, as demonstrated by the phenomena of convergent evolution (e.g., Lauder 1981; Lauder and Liem 1989; Wagner 1989). In the process of perfecting for a particular lifestyle in a given environment, each incremental increase in fitness is accompanied by progressively narrower scope of structural and functional adaptive flexibility. This predisposes a species to collapse and extinction through what has been termed overadaptive meltdown (e.g., Minkoff 1983; Stuart 1991). Borrowing from the comment made by Wells (1962) on the cephalopods, remarking on the extreme marine predatory capacity of the elasmobranchs, Tota and Hamlett (1989) observed that "in doing so (adapting) they have become specialized to the point where their own structure and physiology will preclude further adaptive radiation". Gans (1985) contends that "we should not be surprised to find that no structure is perfect and that few structures are optimized to any particular role". Through intense genetic breeding, a process termed directed evolution by (Joyce 1992), the horse and the domestic fowl, Gallus gallus variant domesticus, have respectively been exceedingly manipulated for speed and weight gain. While better feed and feeding regimen may contribute, in 1960 it took 70 days for a table bird to reach a live weight of 1.8 kg and in 1985 it took only 40 days (Smith 1985). In the course of this enforced productivity, the functional integrity of some organ systems appears to have been severely compromised. Death due to aortic rupture (e.g., Carlson 1960) and vascular pathology (e.g., Julian et al. 1984) is a common problem in the turkey industry. A worldwide increase in occurrence of ascites has been reported in young broilers by Julian and Wilson (1986), Maxwell et al. (1986a,b) and Julian (1987). The syndrome was associated with right ventricular hypertrophy (Huchzermeyer and De Ruyk 1986; Julian and Wilson 1986). In an attempt to explain the pathogenesis of the condition, Huchzermeyer (1986) and Huchzermeyer et al. (1988) contended that hypoxia may cause pulmonary vasoconstriction with consequent pulmonary hypertension, resulting in a right ventricular hypertrophy. In the mammalian fetal lung, vasoconstriction due to hypoxia is thought to restrict blood flow to the developing lung in utero (Morin and Egan 1992). Environmental factors such as cold and altitude as well as nature of food have been associated with ascites in birds (Maxwell et al. 1986a,b; Julian 1987). Respiratory inadequacies in the domesticated birds may result from the poor pulmonary morphometric parameters which generally characterize the group (Abdalla and Maina 1981; Abdalla et al. 1982; Maina and King 1982a; Vidyadaran et al. 1987, 1988, 1990). The growth of the lung of a highly selected line of turkey did not match the increase in body mass (Timwood and Julian 1983; Timwood et al. 1987). The free-ranging village chickens are not susceptible to ascites (Pizarro et al. 1970). For similar reasons, i.e., intense selection for productivity in total disregard of the necessity for commensurate adaptation of the supporting organ systems, domestic fowls (particularly the males) are totally incapable of attaining  $VO_{2max}$  on treadmill exercise (Brackenbury and Avery 1980; Brackenbury et al. 1981; Brackenbury 1984). In the horse (an animal which has been fiercely genetically manipulated for speed performance), the pulmonary capillary blood pressure increases from 2.4 kPa to about 6 kPa during exercise (Sinha et al. 1996). Exercise-induced pulmonary hemorrhage has been reported to affect more than 40 to 80% of horses during high intensity exercise (Mason et al. 1983; Burrell 1985; West et al. 1991, 1993). Experimentally, the capillary transmural blood pressure which causes stress failure in the horse's pulmonary blood capillaries ranges between 10 and 13 kPa (Birks et al. 1997). Even at maximum exercise, a safety margin appears to exist in the lung's capacity to tolerate transmural mechanical stress at the alveolar level.

Based on the fact that many complex designs and patterns can be generated by a few simple natural algorithms, an almost infinite number of different designs of gas exchangers should perchance have evolved in animals. Certain constraints, however, must have enforced convergence and adoption of similar designs, even in animals of remarkably different ancestry. Employing different strategies and resources, such animals have found similar solutions to common challenges. Like most organs and organ systems, the gas exchangers are known to carry out multiple functions which include feeding, sexual display (inflation of the lung, e.g., in frogs), osmoregulation, and secretion and metabolism of certain pharmacological factors (Sects. 6.10.1 and 6.10.2). Interspecific differences in the roles which similar structures play and the many roles carried out by the same structure in an individual animal occur (e.g., Gans 1985, 1988; Bennett 1988). The conflict between the roles may keep some parameters from optimizing. Moss (1962) called the aggregate number of roles an individual structure plays a functional matrix. For performance of multiple functions, such organs have integrated (compromised) designs. Over time, the individual structural components have been refined to carry out a particular role best. Uncoupling the roles that certain structures play eliminates the constraints placed on their improvement and hence imparts opportunity for greater diversity and refinement in form and function (Liem and Wake 1985). To illustrate this point, the emergence of suctional breathing from the buccal force pump dissociated the feeding from the breathing functions, leading to diversification and greater specialization of the two processes in reptiles. Biological structures evolve to satisfy the immediate needs. They do not have to be optimal to be conserved by natural selection. The difference between the cost of operation of an organ during unstressed state and under severe stress can be considered to be its adaptive phenotypic plasticity. Unless the maximal loads a system can bear are well known, the available scope of operation can be read as an overdesign, overconstruction, or even redundancy. It is envisaged that overdesign results from a genetic programming which is based on an unpredictable environment (Gans 1988). Though normally operating at a lower scope (e.g., Bennett 1988), organisms appear to preadapt for the worst-case scenario (Gans 1979). The apparent paradox in the scaling of the morphometric pulmonary diffusing capacity (DLo<sub>2</sub>m) of mammalian lung and the Vo<sub>2max</sub> with body mass (Gehr et al. 1981) suggested that, compared with the small animals, large ones have overconstructed lungs, with DLo<sub>2</sub>m being 10 to 20 times that which they actually need. A number of explanations have been offered to explain this discordance. Federspiel (1989) suggested that erythrocyte redistribution in the pulmonary capillaries during exercise may bring the physiological diffusing capacity closer to the morphometric one. Heusner (1983) asserted that redundancy in the gas exchangers increases with body size. It is plausible that, compared with the small ones, the large animals may possess a greater capacity to generate multifarious solutions to different functional needs. Alternatively, rather than being a redundancy, this feature may be indicative of a safety margin of operation in the lungs of the large mammals in order to support their higher mass-specific expenditure of energy in locomotion (e.g., Hill 1950). Dejours (1990) contended that time-dependent events such as the duration of exercise performed to determine  $VO_{2max}$  are longer in the larger than in the small animals. If this were so, the larger animals would need greater gas exchange potential than the small animals to support aerobic metabolism over the longer duration. At sea level, Wagner (1993) concluded that the parameters which are involved in O<sub>2</sub> uptake and delivery to the tissues (e.g., ventilation, hemoglobin concentration, cardiac output, lung, etc.) have been optimized (i.e., there is very little reserve in them) such that further adjustments do not affect VO<sub>2max</sub>: under such circumstances, the most important parameter which determines the VO<sub>2max</sub> is the cardiac output - not the lung. Pennycuick (1992) suggested that the surface area fractal dimension of the mammalian lung, which may be as high as 2.5, differs between the lungs of the small and large animals, a feature that may impart a better gas exchange capacity in the lungs of the large mammals. The extinction episodes that have befallen multicellular animal life indicate that in cases of sudden catastrophic events, the existing safety reserves are easily overwhelmed. Animals have no inbuilt contingencies for such rare and sudden events of great magnitude. Raup and Sepkoski (1984) argued that major extinction events recur with a periodicity of 26 million years. Rather than pay the cost of supporting superfluous structures which may not be utilized for many generations, it would seem that animals have gambled away their survival and invested on features such as diversity and numerical density in hope that these would see them through such occurrences. It is possible that during the long periods of relative stability, new species with better fitness characteristics would have arisen from ill-prepared ancestral ones. Gould (1994) envisages that the reason that mammals survived the dinosaur demise of the end-Cretaceous (though the two groups had coexisted for over 100 million years) is not, as has been widely argued, that the mammals had evolved special adaptations – as animals cannot possibly anticipate and prepare for future events. Their relatively small size (size of a rat or smaller) enabled mammals to fit into less hostile ecological niches which were out of reach to the more robust reptiles. Janis (1993) appropriately calls the succession of mammals victory by default. Like mammals, birds appear to have endured the pressure very well: at least 20 mammalian groups and 22 avian lineages predate the Cretaceous-Tertiary catastrophe (Coope and Penny 1997).

The thickness of the blood-gas barrier and the diameter of the erythrocytes are two factors in the gas exchangers which have been pushed close to optimization if this has not already occurred. While mammals span the enormous range of body mass from the 2.5-g shrew to the about 150-t whale, a factorial difference of 60 million, the average thickness of the blood-gas barrier of the lung of the shrew (Gehr et al. 1980) of 0.334 um is comparable to that of 0.350 um of the lung of the whale, Balaena mysticetus (Henk and Haldman 1990). Although no estimations of stress failure of the pulmonary capillary blood vessels are available on these two animals, the alveolar wall in the lung of the rabbit, Oryctolagus cuniculus, fails above a transmural blood pressure of 3.3 kPa (Tsukimoto et al. 1991; Costello et al. 1992), a value which is astounding since the harmonic mean thickness of the blood-gas barrier is only 0.50 µm (Weibel 1973). The pulmonary capillary pressure in the rabbit (Maarek and Grimbert 1994), dog (Okada et al. 1992), and human being (Hellems et al. 1949; Comroe 1974), animals of notably different body sizes, is about 1.1 kPa, giving a safety factor of 3. In the dog, Okada et al. (1992) observed that the resistance in a segment of an alveolar blood capillary remains stable even after large changes in the transmural pressure. In the human being, pulmonary capillary blood pressure elevation has been associated with rupture of the alveolar blood-capillary barrier (Severinghaus 1971) and presence of erythrocytes on the alveolar surface in cases of high altitude pulmonary edema (Schoene et al. 1988; Heath and Williams 1989). The pulmonary blood flow is pulsatile (e.g., Wiener et al. 1966; Milnor 1982; Maarek and Chang 1991). It is envisaged that dampening of the pressure wave occurs in the capillary system (Wiener et al. 1966). The pulsatility of the pulmonary microvascular pressure may influence filtration of fluid across the capillary wall and gas exchange at the alveolar level (Maarek and Chang 1991). In mammals, the size of the erythrocytes presents a good example of structural optimization. The erythrocytes of the shrew, the smallest extant mammal, have a diameter of 7.5µm and compare with those of the humpback whale of 8.2µm (Altman and Dittmer 1961). Since the smallest blood capillaries should set the limit for the ervthrocyte diameter (though the cells fold greatly as they traverse these narrow conduits), the diameters of the blood capillaries in the smallest and largest mammal appear to be comparable. This suggests an optimal setting of the capillary diameter for satisfactory supply of  $O_2$  to the tissues.

The criteria of categorizing a biological structure as inconsequential, superfluous, excessive, subservient, nonfunctional, or constituting a controlling, regulatory, limiting, constraining, or even being a triggering factor is oftentimes highly subjective. In some cases, by acting differently, opposing selective pressures greatly alter the physiological profiles of the functional components, whereby some processes may be suppressed, some may synergize, while others may become totally obliterated or remain functionally neutral. The operational definition of what is optimal is oftentimes biased and misleading. The interpretations may depend on circumstances and even on anthropocentric perceptions and personal preference of what may be deemed beneficial. For example, while intrapulmonary air and lighter bones may be considered to be useful in increasing the buoyancy of a flying animal, in an aquatic one, both parameters are a vulnerability rather than a benefit. Though the lung is considered to be specifically designed in view of efficient uptake of  $O_2$  and discharge of  $CO_2$  (e.g., Weibel 1983a, 1985b), the organ plays important defensive, pharmacological, and endocrine functions (Sect. 6.10.2). To yet unknown extents, such roles must be accommodated in the overall design of the organ. The composite nature of biological tissues and the multiplicative effects between the various integral components may explain why the sum of the individual functional processes often exceeds the aggregate value expressed by the whole organism (Hoagland and Dodson 1995). For example, while the full energy budget of an organism in normoxia is totally aerobic, even the well-oxygenated mammalian cell cultures invariably manifest a partly anaerobic state (e.g., Gnaiger 1991).

#### 2.6 Fractal Geometry: a Novel Approach for Discerning Biological Form

Geometry, as advanced by Euclid (300 B.C.) and Pythagoras (6 B.C.), uses straight lines and smooth and regular curves to make flat shapes and figures such as squares, triangles, and circles to model structures. When applied to natural things, however, it idealizes form in inherently chaotic structures (e.g., Olsen and Degn 1985; West 1990). The classical geometry defines space in terms of discrete dimensions, e.g., a point has no (zero) dimensions, a line has one dimension, a plane (area) has two, and a solid (volume) has three dimensions. These integer dimensions are unsatisfactory in describing the highly complex natural forms and the dynamical physiological processes that do not have specific scales of length and time. To adequately define the topological characteristics of dynamical structures and processes, fractional power dimensions (fractals) are necessary. The development of biological states and events are regulated by nonlinear, iterative algorithms which program morphologies and physiologies that lack absolute spatial and temporal boundaries (e.g., West 1985, 1987; Nonnenmacher 1987; Voss 1988; Nelson et al. 1990; Bassingthwaighte et al. 1994). The designs and operations present scale-invariant properties and self-similarity (Horsfield and Woldenberg 1986; Barnsley et al. 1987; Bassingthwaighte 1988; Giaver and Keese 1989; Goldberger et al. 1990). Discovered by Mandelbrot (1977, 1983), fractal geometry provides a powerful tool for rationally investigating form and function (e.g., Barnsley et al. 1987; Goldberger and West 1987; Tsonis and Tsonis 1987; West and Goldberger 1987; Glenny et al. 1991). Fractal characteristics have been reported in viruses (e.g., Briggs 1992) and tissue cells (e.g., Nonnenmacher 1988; Smith et al. 1989; Losa et al. 1992). They are integral to the functions of organs like the heart (Goldberger et al. 1985; van Beek et al. 1989; Goldberger 1991) and the gastrointestinal system (Pennycuick 1992). Structures such as the bronchial tree of the lung (e.g., Mandelbrot 1983; West et al. 1986; Nelson et al. 1990; Weibel 1991, 1994; Bates 1993, but see a dissenting view by Phillips et al. (1994) and the pulmonary arterial tree (Krenz et al. 1992) present fractal attributes. Blood flow in the lung (Glenny and Robertson 1990; Caruthers and Harris 1994), in the middle cerebral artery (Rossitti and Stephenson 1994), and in the myocardium of humans (Bassingthwaighte et al. 1989) has fractal properties.

Fractal geometry provides means for realistically analyzing dynamic forms and processes (e.g., Nelson and Manchester 1988) and studying physiological properties (Goldberger and West 1987). A corrugated structure in threedimensional space is physically in transition between a smooth surface and a volume. Depending on the degree of amplification, such structures should have a

fractal dimension between 2 and 3. A fractal dimension defines to what extent the topological details of an object fit between the Euclidean dimensions. The surface fractal value of the mammalian lung is about 2.5 (Weibel 1991). Mandelbrot (1977, 1983) observed that both the fractal dimension, D, and the diameter component,  $\Delta$ , of the bronchial tree were about 3. Such high values indicate that the surface area of the lung has been sufficiently highly folded and the peripheral airways sufficiently regularly branched as to nearly fill a three-dimensional space. The two-dimensional surface area of the human vascular system is so highly folded that it has an effective fractal dimension of 3, with the arteries alone having a value of 2.7 (Briggs 1992). It has been argued by Goldberger et al. (1990), Nonnenmacher (1989), Weibel (1991, 1994), and Rossitti and Stephensen (1994) that fractal designs should permit biological systems to operate over a wide range of perturbations without failure. Fluidity in the configuration imparts greater error tolerance (i.e., safety margin) to a biological structure and may be decisive in the trial-and-error process of evolution (West 1987). The inbuilt fractal algorithm reduces the probability of error during morphogenesis since it is dependent on a well-tested self-similar, repeating (iterative) process. Functionally, a fractal attribute eliminates the need for drastic constructional overhauls in a system operating within reasonable boundaries and exposed to moderate assaults. Pennycuick (1992) envisages that the use of fractal structures can allow a particular plan to be scaled over a wide range of sizes without consequential allometric change. Functional plasticity should make adaptation and the more or less trialand-error evolutionary process by genetic programming much easier to effect, against stochastic settings (e.g., Doebeli et al. 1997).

Unique to most organ systems, and perhaps an indication of the importance of respiration for survival, the gas exchangers have been configured by essentially molding together three characteristically fractal entities, namely the pulmonary arterial system (e.g., Lefevre 1983; Glenny and Robertson 1991a; Krenz et al. 1992), pulmonary venous system, and the bronchial-alveolar system (Weibel 1986). Though it may appear passive, the lung is intrinsically a highly dynamic organ. It is subjected to continuous cardiovascular hemodynamic changes as well as biomechanical ventilatory rhythms. It acts as an interface between blood and air, media which are physically remarkably different. The lung is the only organ in the body which transmits the entire volume of blood in the systemic circulation. The multifunctionality of the lung was pointed out by Bakhle (1975), who declared that "the lung should now be considered not merely as an apparatus for gas exchange or mechanical filtration of blood but also that of providing an essential control of the blood levels of many biologically active substances". The fractal characteristics of the surface of the lung allow the large internal surface area to be homogeneously ventilated and perfused at low energy cost (Weibel 1983a, 1991, 1996) and the circulatory system to be highly distensible (Caro and Saffman 1965; Yen 1989a,b; Bshouty and Younes 1990) to contend with the fluctuating hemodynamic blood pressures (Maloney et al. 1970; Zhuang et al. 1983; Al-Tinawi et al. 1991). Although only about 9% of the total blood volume is contained in the heart (Dock et al. 1961; Milner 1980), Hainsworth (1986) observed that the "distensibility of the pulmonary circulation is of particular importance as it permits transient imbalance between the outputs of the left and right hearts". Pulmonary blood flow is pulsatile from the entrance of the pulmonary circulation to its outlet in the left atrium (Morkin et al. 1965; Wasserman et al. 1966; Milnor 1982; Maarek and Chang 1991). The fractal dimensions of the diameter element of the arterial and the venous tree in the human lung are 2.71 and 2.64, respectively, while equivalent values for the length element are 2.97 and 2.86, respectively (Huang et al. 1996). In the dog lung, the fractal dimension of the blood flow is 1.22 (Barman et al. 1966). Pennycuick (1992) suggested that the morphological complexity of the avian pulmonary system (Sect. 6.7.5) may be due to a high fractal surface (with a dimension of about 2.5), enabling birds to achieve exercise capacities such as sustained flights at altitude without the need to vary the general plan of the respiratory system. The morphological features of the avian respiratory system bear this suggestion out: except for differences in the sizes and locations of the air sacs, pneumatization of the long bones, and development of the parabronchial systems (Sect. 6.7.5), the configuration of the lung-air sac system in birds is uncommonly uniform in such a numerically large and ecologically diverse group. A small fractal dimension in the design of the characteristically mammalian bat lung (Sect. 6.7.4.1) may have called for multiple extrapulmonary compensatory changes (Maina 1998) in order to provide the large volume of O<sub>2</sub> needed for flight. This line of reasoning is supported by a number of observations which include: (1) bats have enormous lungs which occupy much of the celomic cavity (e.g., Maina and King 1984), and (2) the mass of the heart and parameters such as the hematocrit and hemoglobin concentration are some of the highest values among mammals (Jürgens et al. 1981). The high demands imposed by flight on a gas exchanger of moderate efficiency may explain why the heaviest bats, the flying foxes (pteropodids), weigh only about 1.5 kg, a value which is an order of magnitude smaller than the weight of the heaviest flying bird (about 15 kg). For gas exchange, birds operate on what was termed a broad-based low-keyed strategy where many moderately refined parameters are variably utilized in combination, affording a large functional reserve (Maina 1998). The highly efficient tracheal system of insects (Sect. 6.6.1) is thought to have fractal surfaces of high dimension (Pennycuick 1992). The fractal dimension of 2 suggested for the fish gills by Pennycuick (1992) may be due to the assumption that the secondary lamellae of fish gills are smooth. Lamellar microridges (Fig. 50b) are characteristic of secondary lamellae of most fish gills, even those of the most ancient fish such as the coelacanth, Latimeria chalumnae (Hughes 1995), and the sturgeon, Acipenser transmontanus (Burggren et al. 1979). Microridges are, however, poorly developed in the gills of Trachurus mediterraneus (Hughes and Mondofino 1983) (a carangid fish) and appear to be lacking in the secondary lamellae of the hill-stream fish such as Danio dangila (Ojha and Singh 1986). It would seem that generally, like other gas exchangers, the gill surface is a fractal construction. Microridges increase the respiratory surface area of the gills, provide anchorage for the mucous lining, generate turbulence, and reduce drag forces at the water-gill interface (e.g., Sperry and Wassersug 1976; Hughes and Mondolfino 1983; Hughes 1979, 1984). Recently, applying fractal geometry to the allometric scaling of animal size, West et al. (1997) concluded that the enormous size disparity in the evolved animal life has been possible due to the intrinsic fractal nature of tissues and organ systems. Cope's rule (see Cope 1896, generally considered a pervasive evolutionary pattern, but see dissenting views by Fenchel 1993 and Jablonski 1997), asserts that phylogenetically, animal lineages tend to evolve larger body size for reasons that they achieve greater mating success, better defense capacity, predatory ability, and resistance to environmental extremes (Bonner 1988). In biology, time-dependent events, e.g., ventilatory cycle, incubation and gestation periods, and duration of exercise needed to reach maximal  $O_2$  consumption are longer in large than in small animals (Dejours 1990). The mass specific cost of transport is lower in larger animals than in smaller ones (Boulière 1975; Taylor 1977).

#### 2.7 From Diffusion, Perfusion, and Ventilation to Respiratory Pigments

#### 2.7.1 Diffusion

The evolution of respiration unfolds critical moments in the past when at certain times momentous developments occurred. Some of the most profound changes took place during the transformation of the anaerobiotic prokaryotes to the aerobiotic eukaryotes (the inquisition of molecular  $O_2$  into respiration), accretion of cells into the Metazoa, shift from water- to air breathing, transition from water to land, and change from ectothermic-heterothermy to endothermic-homeothermy. At each of these stages, the metabolic needs of organisms would have exceeded those that could be serviced by the default gas exchanger. Changes which resulted in greater diversity and specializations of the gas exchangers were thereby instituted. In the theory of chaos (e.g., Stewart 1990) such phenomenal moments when conditions occur to create dynamical, self-driving structures by amplifying and locking the system's feedback together are called bifurcation points (Briggs 1992).

A distinct respiratory system cannot be clearly delineated below the level of mollusks and arthropods. The most primitive gas exchanger in such simple metazoans is an unspecialized epidermis. Gas transfer occurs by diffusion across the general body surface and in some cases through inbuilt modifications (e.g., Mangum 1994). Diffusion is a natural phenomenon which dictates spontaneous flow of molecules from places of high to low concentrations free of energy, just as water flows downhill. This must have been the earliest mode of gas transfer which was initiated by the increase of molecular  $O_2$  in the primeval biosphere, resulting in the evolution of aerobic processes. When it is the only means of supplying O<sub>2</sub> to the cells, diffusion determines the shape and size of organisms (e.g., Burggren and Roberts 1991). A small and flat body form enhances gas exchange by diffusion (e.g., Jell 1978; Runnegar 1982). When O<sub>2</sub> needs are high, at 1 atm, diffusion can only be adequate in organisms up to about 1 mm in diameter and in animal tissues 2 to 5 mm thick (e.g., Comroe 1966; Schmidt-Nielsen 1990). Larger animal forms must possess a low rate of metabolism if they are to rely wholly on diffusion as the only means of securing O<sub>2</sub> and voiding CO<sub>2</sub>. The evolution of complex energetic animals obliged the development of diverse specialized systems to deliver O<sub>2</sub> from the external environment to the tissue cells (Comroe

1966). As a spherical body provides the lowest surface to volume ratio, to overcome diffusional limitations, animals have deviated from this design by evolving long, highly attenuated bodies. Factors such as the habitat occupied and the mode of life determine the definitive shape and size of animals (Gould 1966) as well as the type of a gas exchanger needed to effectively service the total cytoplasmic mass. The innermost part of the developing embryos in an egg mass of the sand snail, Polinices sordidus, which weighs as much as 210g (Shepherd and Thomas 1989) and has a radius of as much as 40 mm, experiences extreme hypoxia ( $PO_2 < 1 \text{ kPa}$ ) though the PO<sub>2</sub> in the outer layer of water may be moderately high at  $PO_2 > 10$  kPa (Booth 1995). Water inside a spawn of *Rana temporaria* was found to be only 3 to 16% saturated while the saturation about 50 cm away from the egg cluster was 136% (Savage 1935). By adopting appropriate morphological designs and subsisting in a well-oxygenated medium, an organism can attain greater body size while relying on diffusion only. Through an assemblage of irregular shapes which increase the surface area, the diffusional distances can be reduced. In the sponges, where there is no internal perfusive mechanism, the body has flagellated cells (choanocytes) which move water (by the beating of cilia) through numerous incurrent pores or ostia into ramified water channels of about 1 mm diameter. This brings the tissue cells close to water enhancing gas exchange and nutrient uptake (Fig. 17). As much as 90% of O<sub>2</sub> is extracted from the water passing through the pores of sponges during maximal activity (Hazelhoff 1939). The largest known sponge is the barrel-shaped loggerhead sponge (Spheciospongia vesparium) which is found in the West Indies and in the waters off the coast of Florida: it stands at a height of 1.5 cm and is 9 cm in diameter. Sponges have been recovered from depths up to 5.6km. In the coelenterates, a group which has a remarkably low metabolic rate, a steady water flow by ciliary movements across the gastrovascular canals is maintained. Pelagia,



Fig. 17. A External morphology and sectional profiles B and C of an asconoid sponge, *Leucosolenia* showing the canals (**\**, B) through which water passes into the body cavity. D, E, F, G, and H show the various types of cells which comprise the body wall. (Jessop 1995)

which can grow to a diameter of about 6 cm (Henze 1910), and the coelenterate *Cynea*, which can grow to a diameter of 2 cm, are said to rely sorely on diffusion for their O<sub>2</sub> needs (Krogh 1941). The tropical earthworms, animals which are as large as 1 kg in body mass, e.g., Rhinodrilus fafner and Megascolides australis have been recorded to attain lengths of 2.2 m (but have a diameter of about 24mm), rely entirely on cutaneous diffusion for gas exchange (Stephenson 1930). In helminths, Fry and Jenkins (1984) estimated that to depend entirely on diffusion to maintain aerobic respiration, the critical thickness is 0.4mm for the nematodes and 0.75 mm for the cestodes. The interstitial animals (animals which live in the water-filled spaces between the aquatic sediments) like the polychaete Stygocapitella subterranea and oligochaete Marionina achaeta do not have specialized respiratory surfaces. They acquire their  $O_2$  needs entirely by diffusion through their great surface-to-volume ratio achieved from their thin, cylindrical body configurations. In an adult gastrotrich 500 µm long, Colacino and Kraus (1984) estimated that the O<sub>2</sub> transfer across the body surface was three times greater than its O<sub>2</sub> consumption when the PO<sub>2</sub> between the water and the mitochondria was only 0.13 kPa. Due to the much greater rate of O<sub>2</sub> diffusion in air than in water (Table 4) and the more constant level of O<sub>2</sub> in air, for the same concentration difference, the mass transfer attained over 1 µm in water can occur over a much longer distance of 1 cm in air. The PO, in the hemolymph of the gooseneck barnacle, Pollicipes, is elevated in air (e.g., Petersen et al. 1974). The air can support a spherical animal 100 times greater than water at the same rate of metabolism. However, such animals do not exist due to the parallel complications which would arise in air such as risk of desiccation and lack of mechanical support. In simple organisms, there exists a delicate compromise between the

Parameter	Unit	Water	Air	Water/air	Air/water
Density	$gml^{-1}$	1.000	0.0012	833	_
Viscosity	Centipoises	1.00	0.02	50	-
O <sub>2</sub> Content	$ml 100 ml^{-1}$	0.66	20.95	_	32
CO <sub>2</sub> Content	$ml \ 100 \ ml^{-1}$	0.03	0.033	0.91	-
Thermal capacity	Cal ml <sup>-1</sup> °C	1.00	0.0003	3333	-
Diffusion coefficients	$\mathrm{cm}^2\mathrm{s}^{-1}$				
0,		0.000025	0.198	-	8000
CO <sub>2</sub>		0.00008	0.155	_	2000
Capacitance coefficients	$nMolml^{-1}mmHg^{-1}$				
0,		1.82	54.74	-	30
CŌ,		51.89	54.73	-	1
Krogh's constants	$nMol cm^{-1}s^{-1}$ mmHg <sup>-1</sup>				
0,	0	0.000046	10.84	_	20000
CO <sub>2</sub>		0.00093	8.46	-	9000

Table 4. Some physicochemical properties of water and air<sup>a</sup>

<sup>a</sup> Measurements made at 20 °C and at 1 atmosphere pressure.

need to have an extensive and thin respiratory barrier or a thick and less waterpermeable one. By living in nondesiccating (cryptozoic = humidic) habitats, risk of desiccation is minimized and a thin barrier can prevail. In most organisms, exceeding a diameter of about 1 cm, a circulatory system becomes a necessity. From hypothetical mathematical computations (e.g., Harvey 1928; Rashevsky 1960), in a normobaric environment and at moderate O<sub>2</sub> consumption, the maximum radius of a spherical cell, where anoxic state would not occur except at the center (i.e., the farthest point from the surface), was estimated to be 0.5 mm. Krogh (1941) calculated that for a homogenous spherical organism of a radius of 1 cm, at an O<sub>2</sub> consumption of 100 ml kg<sup>-1</sup> h<sup>-1</sup> (about half of the O<sub>2</sub> consumption of resting man) an external PO<sub>2</sub> of 25 atm (2533 kPa) would be required for O<sub>2</sub> to diffuse to all parts of the organism. He calculated that an organism cannot have a radius of more than 0.5mm if it were to rely on diffusion alone, even assuming that it lives in water almost fully saturated with air at 1 atm of pressure. Where the external medium is separated by a 50-µm barrier from an internal circulating medium, a PO<sub>2</sub> of one quarter of an atmosphere would be required for satisfactory diffusion. Relying entirely on diffusion, a paramecium (volume 0.0006 cm<sup>3</sup>, O<sub>2</sub> uptake 1.3 ml per g per h and diameter 0.11 cm) would need a PO<sub>2</sub> of 0.73 of an atmosphere (Prosser and Brown 1962). Clearly, such high PO<sub>2</sub> rarely occurs in natural environments. Denney (1993) estimated that at the normal metabolic rate of a typical protozoan of about  $0.1 \,\mathrm{mol}\,\mathrm{m}^{-3}\mathrm{s}^{-1}$ , a cell would have to be almost 7.5 cm in radius to experience hypoxia. The disparities in the estimated theoretical maxima of organisms indicate likely flaws in the idealized models used to calculate the largest possible sizes that microorganisms can attain. Except probably for eggs, there are no organisms which are absolutely spherical. At the organismal level, scaling with size is nonisometric (Schmidt-Nielsen 1984). The largest protozoans that have ever lived were the now extinct nummulites which had a diameter of about 2.4 cm and the largest extant protozoan is *Pelomyxa* palustris, which can attain a length of up to 1.5 cm. This size difference may be accounted for by the emerging possibility that the PO<sub>2</sub> in the biosphere at certain time(s) in the evolutionary past was higher than the present level (e.g., Graham et al. 1995). One such case occurred in the late Paleozoic (between the Carboniferous and the Permian), when for about 120 million years, O<sub>2</sub> level was 1.7 times greater than in the present atmosphere (Berner and Canfield 1989; Landis and Snee 1991; Graham et al. 1995; Fig. 9). Extraordinarily, the endothermic plants, e.g., Philodendron selloum, which produce heat at a rate surpassing that of the insect flight muscles and can maintain a temperature gradient of 30 °C with the ambient (Seymour 1997; Koch et al. 1983) rely entirely on diffusion for supply of the  $O_2$ needed for generation of energy. In a single floret of the inflorescence of P. selloum, O, diffuses across a distance of about 1.2 mm through about 170 stomates (Koch et al. 1984): the average diffusional length from the surface to the individual cells, which is less than 0.75 mm, compares with that of the trachea of most small diffusion-dependent insects (Sects. 6.5 and 6.6.1). This reveals nature's amazing congruent solutions to similar needs!

Diffusion over an undifferentiated surface is the method of respiration in the simple organisms, e.g., Protozoa, Rotatoria, Planaria, Nematoda, eggs and young embryos, copepods and ostracodes, while the more complex ones (which utilize this process) include Spongia, Cerripedia, Coelenterata, arthropods like Tardigrada and Pauropoda, eggs, and early developing embryos (e.g., McMahon and Wilkens 1983). Larvae of many insects rely on diffusion across the integument (Fraenkel and Herford 1938) with the flux of gases essentially being regulated by O<sub>2</sub> consumption and CO<sub>2</sub> production within the organism. The tracheal system of the Cossus larvae delivers adequate O2 entirely by diffusion (Krogh 1920a). The occurrence of air in the tracheoles of the young adult which emerges after the larval gas-filled tracheoles have been shed (there having been no previous contact with air) is probably due to passive diffusion of  $O_2$  (Keilin 1924; Buck and Keister 1955). Surface-to-volume ratio decreases with body size since volume increases as the cube while surface area increases as the square of the radius. Assuming that the metabolic rate remains constant, O<sub>2</sub> transfer by diffusion should decrease with size. Since some organisms (including the amphibian eggs) develop to sizes greater than those theoretically predictable and the PO<sub>2</sub> inside the cell exceeds that which would be expected from diffusion alone, it was conceived that another process must promote gas transfer (e.g., Longmuir and Bourke 1960; McDougal and McCobe 1967). In the protozoa, protoplasmic streaming, a normal circulation-like process in living cells, enhances intracellular gas transfer (Seifriz 1943; Andrews 1955). Mechanical vibrations appear to intensity permeability of tissues to gases (Longmuir and Bourke 1960). Dynamic organs such as the heart, lung, and diaphragm, as well as activities such as change in muscle tone and physical interaction between the erythrocytes and the endothelial wall of the blood vessels may to an unknown extent influence gas transfer at the tissue level.

#### 2.7.2 Convective Flows

#### 2.7.2.1 Perfusion

The inadequacy of diffusion as a means of gas exchange necessitated development of auxiliary respiratory processes as organisms became larger, more complex, and their O<sub>2</sub> needs increased. A progressive development of a circulatory system occurred, promoting the efficiency of the respiratory processes (Figs. 18,19,20). Mature animals having an elementary circulatory system close to the surface of the body, e.g., earthworms and echinoderms, where the blood is moved by a heart and not cilia, can grow to a body mass of a few grams and a length of 30 to 40 cm without calling for development of special respiratory organs as long as they subsist in water or in a humidic environment. In such animals, a directional flow of the lymph may not exist and hence a circulatory system strictly does not exist. Churning of the fluid underlying the respiratory surface through contraction of body muscles should enhance the flux of the respiratory gases. In organisms such as Chaetopoda, Synapta, and Pantopoda, there is no regular circulation but the coelomic fluid is kept in motion by cilia (Lindroth 1939). The blood of the earthworms has a high  $O_2$  affinity ( $P_{50} = 0.3$  to 1.1 kPa), is highly sensitive to temperature (Laverack 1963) and has a high O<sub>2</sub> carrying capacity of 8 to 12 vol % (Haughton et al. 1958). The blood of the giant earthworm



Fig. 18a-e. Development of the respiratory processes from unicellular to multicellular organisms. Diffusion (a) is the main process of gas exchange in simple animals with external (b) or internal convection (c). More complex animals combine perfusion and ventilation (d and e). While the gills are configured for continuous and unidirectional ventilation (d), the lungs and their derivatives are tidally ventilated (e). (Burggren and Pinder 1991)

(Glossoscolex giganteus), which can attain a body mass of 600g, a length of 120 cm and a width of 2 to 3 cm, has a P<sub>50</sub> of 0.9 kPa (at 20 °C and pH of 7.5) and a small Bohr shift (Johansen and Martin 1966). By regulating cutaneous perfusion (Burggren and Feder 1985) and surface area (Noble 1925), the hairy frog, Astylosternus robustus, can adjust gas exchange across the skin. In the higher vertebrates, the skin has been rendered virtually impermeable to O<sub>2</sub> and in the human being only 0.2% of the total O<sub>2</sub> need is acquired through it (Krogh 1941). The plethodontid salamanders (Collazo 1993; Ruben et al. 1993; Tilley and Bernado 1993; Wake and Marks 1993), which live in cold, well-oxygenated habitats and have adopted long and cylindrical body forms, rely entirely on the skin for gas exchange: this group, which is extremely successful (Pough et al. 1989), is presumed to have originated from torrential mountain streams (Beachy and Bruce 1992). In the freeze-tolerant frogs which can endure temperatures of between -3 and -7 °C, e.g., Hyla versicolor, Rana sylvatica, Hyla cricifer, and Pesudacris triseriata macurata, the heart does not beat in the frozen state (Lotshaw 1977; Storey and Storey 1988). In some species, intracellular glucose level increase and appear to serve both as a cryoprotectant and a metabolic fuel (Storey and Storey 1986). At natural wintering temperatures of 3 °C, freshwater turtles can remain submerged without O2 for months (Carr 1952; Ultsch and Jackson 1982). Nemertines (ribbon worms) have only two main longitudinal blood vessels and in the species which have hemoglobin, reversal of blood flow is common (Hyman 1951). The bootlace worm, Lineus longissimus, which is found in the North Sea, can reach a length of 55 m. In some annelids, a closed circuit


Fig. 19a–f. Mechanisms of gas exchange. The most basic gas exchange design is that which occurs across a cell membrane, e.g., in protozoa or across an unperfused skin (a). Perfusion of a respiratory site improves gas transfer by maintaining a concentration gradient (b). In the tracheal gills, e.g., of the mayfly, air rather than blood is contained in the gills (c). In the tracheal system of insects, air is delivered directly to the tissue cells by diffusion (d) and in the larger species by convection through abdominal pumping. In the conventional gills, e.g., of fish, the organs are well perfused with blood (e) and ventilated with water. The elaborate vertebrate gas exchangers combine tidal ventilation with perfusion (f). (Jessop 1995)

with well-developed blood vessels and pulsatile ancestral "hearts" exist in form of modifications along the blood vessels. In certain teleosts and elasmobranchs (sharks, skates, and stingrays), caudal hearts, which are located near the tail and powered by skeletal muscle, aid in venous return (e.g., Satchell 1992). Auxiliary hearts occur in the circulatory system of decapod crustaceans (Steinacker 1975) and locomotor movements generate large pressure differentials which promote the flow of the hemolymph (Belman 1975). The oscillations in the dorsal aortic blood pressure in the Atlantic hagfish, *Myxine glutinosa*, are associated with contractions of the gill musculature, a process which may be involved in propulsion of blood (Johansen 1960; Strathmann 1963). Such a process, however, does not seem to occur in the gills of the Pacific hagfish, *Eptatretus stoutii* (Chapman et al. 1963). At 10 °C, *M. glutinosa* has a heart rate of about 22 beats min<sup>-1</sup>, a mean ventral aortic blood pressure of 1 kPa and a cardiac output of 9 ml per min per g



**Fig. 20.** Comparative schematic illustration of the stages at which cardiovascular and respiratory systems develop in relation to life histories of different vertebrate taxa. Some stages may fail to develop in some groups. For example, not all fish attain capacity for air breathing and not all amphibians develop lungs. Variably, the sequence in the increasing complexity and efficiency of the gas exchangers appears to progress from the simple membrane gas exchanger, a perfused one, a carrier-supported one, and finally to a ventilated one. Of note is the fact that hemoglobin (*Hb*) develops before respiration starts. (After Burggren and Pinder 1991, with the processes in birds and mammals added)

heart mass (Forster et al. 1988, 1991; Axelsson et al. 1990). While buried under the soft mud, Myxine glutinosa can remain without O<sub>2</sub> for at least 1 h (Strathmann 1963) and feed inside the body cavity of dead animals for a long period of time (Hardisty 1979): at 5°C, the hagfish can survive in anoxic water for at least 20h (Hansen and Sidell 1983). In most gastropods, to ensure forward blood flow, passive valves exist throughout the body (Jones 1983). In the open circulatory system, a large quantity of fluid is located in the intercellular space, providing mechanical support for locomotory activity and feeding movements (e.g., Jones 1983; Russell and Evans 1989). A fast circulatory return in a closed system with a smaller blood volume should be a more efficient design compared with a sluggish one with a large volume. Open circulations without hearts suffice where the respiratory demands are not high and where the diffusional distances are not great (Farrell 1991a). Heart rate in the terrestrial slug, Deroceras reticulatum, increases during feeding, a feature attributable to the need for substantial hemocoelic pressure required to protrude the odontophore (Duval 1983). Respiratory gas exchange across the body surface coupled with a simple circulatory

system occurs in leeches, all oligochaetes, and some polychaete annelids. The organization is more complex in the more advanced animals (Fig. 19). In some fish embryos and larvae, circulation develops before the respiratory organs. Blood pressures as high as 2.5 kPa and fairly fast flow velocities have been recorded in some large earthworms (Johansen and Martin 1966). Ventricular pressure measurements in the black-lip abalone, *Haliotis ruber* (Russell and Evans 1989) ranges from 0.4 to 1.2 kPa and heart rate increases with water temperature, the maximum rate being reached at 22 °C. In teleosts and elasbobranchs, typical heart rates are 10 to 60 beats min<sup>-1</sup>, cardiac outputs are 6 to 40 ml min<sup>-1</sup>kg<sup>-1</sup> body mass and the mean ventral aortic pressure ranges between 3 and 6kPa (Farrell 1984, 1991b; Lai et al. 1990; Axelsson et al. 1992).

The gastropod (Jones 1983; Andrews and Taylor 1988) and cephalopod mollusks (Wells 1983) were the first taxons to acquire a distinct circulatory system. The heart is well organized internally for directional blood propulsion and has regular beats. In the pneumonate gastropod mollusks, the systolic pressure is as high as 4 kPa (Jones 1983; McMahon and Wilkens 1983). In some terrestrial crabs, the systolic peak blood pressure can be as high as 6.7 kPa (Cameron and Mecklenburg 1973), gradually dissipating to zero after the gills which contribute 40% of the peripheral resistance (Bourne and Redmond 1977). The closed circulation in the cephalopods constitutes the threshold towards circulatory adaptation for more efficient gas transfer. With certain exceptions, the most common response to hypoxia in gastropods and bivalves (DeFur and Mangum 1979; Russell and Evans 1989) is bradycardia. In fish, a similar response occurs (Randall and Shelton 1963; Farrell 1982a; Gehrke and Fielder 1988; Fritsche 1990); the heart rate may drop by as much as 50%. Interestingly, hypoxic bradycardia is weak or does not occur in fish such as the sea raven, Hemitripterus americanus (Saunders and Sutterlin 1971), winter flounder, Pseudopleuronectes americanus (Cech et al. 1977), and the rockpool fish, Gobius cobitis (Berschick et al. 1987). While these differences may be genuine interspecific adaptive responses to hypoxia, it cannot be ruled out that the experimental approaches and the depth of hypoxia at which the tests are being carried out may contribute. Compared with other teleosts, e.g., the eels and the goldfish, which can withstand hypoxia for hours at elevated temperatures (e.g., Walker and Johansen 1977; Waarde et al. 1983), the salmonids have a particularly limited capacity of coping with hypoxia (Doudoroff and Shumway 1970). In aquatic amphibians where cutaneous respiration is particularly important (Johansen and Burggren 1980), a marked bradycardia, accompanied by decreased blood pressure and cardiac output, occurs with submergence (Shelton and Jones 1965). Cardiac output may be reduced to 5% of the predive levels and heart rate may drop to as low as 4 to 8 beats min<sup>-1</sup> in turtles (White and Ross 1966; Penney 1977; Herbert and Jackson 1985). The mass specific volume of blood is greater in animals with an open circulatory system compared with those with closed ones (Prosser 1961). In the former, the peripheral resistance and blood pressures are generally low (Jones 1983). In crustaceans, blood volume constitutes about 30% of the body weight (Prosser 1973), in gastropod molluscs 25 to 30%, in the bivalves as much as 60%, and in the dog only 8.3%. The relative blood volume decreases with body size in mammals (Gregersen and Rawson 1959). In most insects (Sect. 6.6.1.), the circulatory system serves no consequen-



**Fig. 21A–D.** Arrangement of the vascular system relative to the accessory respiratory organs. In the single arch of a typical fish (A), the entire cardiac output is directed towards the gills. In various air-breathing fish, e.g., *Synbranchus* (B), *Hoplosternum* (C), and lungfish (D), to varying extents mixing occurs. The process is, however, minimal in the dipnoans (lungfish). (Johansen 1968)

tial role in gas exchange. In the lungless salamanders, respiration is entirely cutaneous. In the group, the left auricle is lacking. Invertebrates such as the crustaceans and mollusks have an open circulation where a capillary system between the arteries and veins is largely lacking and the blood returns to the heart more or less at random through a system of tissue spaces unbounded by endothelial cells (McMahon and Wilkens 1983; Burggren and McMahon 1988a).

The pinnacle of development of circulation, the double circulatory system took nearly 300 million years to configure from a single circulation. The lungfishes (Dipnoi), where a pulmonary vein and a partly divided heart are first encountered (Fig. 21), present a vital point in the evolution of the double circulation (Bugge 1960; Satchell 1976). All amphibians with a lung have a pulmonary vein and a complete or partial septum which separates the right and left atria. In airbreathing toads, at 22 °C, heart rate is 26 beats min<sup>-1</sup> with an arterial pressure of 3 kPa and a systemic arch blood flow of 36 ml min<sup>-1</sup> kg<sup>-1</sup> body mass (Withers et al. 1988): the total cardiac output is about 30 ml per min per g heart mass (Driedzic and Gesser 1994). The crocodiles, a relatively remarkably advanced reptilian group (Densmore and Owen 1989; Norell 1989; Tarsitano et al. 1989), are the only ectothermic group which has virtually advanced to the stage of a four-chambered heart. In air-breathing turtles at 20 to 25 °C, the heart rate is 30 to 40 beats min<sup>-1</sup>, cardiac output about 50 mlmin<sup>-1</sup>g<sup>-1</sup> heart mass and the aortic pressure about 3kPa (Driedzic and Gesser 1994). At 35 °C, both the savanna monitors (Varanus exanthematicus) and the green iguana (Iguana iguana) have a resting heart rate of 40 beats min<sup>-1</sup> with cardiac output of about 40 ml min<sup>-1</sup> per g heart mass in V. exanthematicus and  $70 \text{ mlmin}^{-1}$  per g heart mass in I. iguana (Gleeson et al. 1980). In a resting V. exanthematicus, the systemic blood pressure may approach 9kPa (Burggren and Johansen 1982). It is only in the postembryonic endotherms, mammals and birds, where the heart is completely divided and the pulmonary and systemic circuits are anatomically distinct. The resting heart rate of a 500-g ectothermic vertebrate at 15 to 20 °C under normoxia is 20 to 60 beats min<sup>-1</sup>, the arterial blood pressure ranges from 3 to 5kPa and the cardiac output is in the order of 10 to  $50 \text{ ml min}^{-1} \text{ kg}^{-1}$  body mass (Driedzic and Gesser 1994): a mammal of similar size at 37 °C has a heart rate of 280 beats min<sup>-1</sup>, a left ventricular output of 125 ml min<sup>-1</sup>g<sup>-1</sup> and an aortic blood pressure of 13 kPa. The pulmonary circuit, which handles the entire systemic venous blood, is a low resistance circulation where the pressures on average are 1.7 kPa (Rushner 1965; West 1974). In the human lung, the resistance to blood flow across the blood capillaries of the lung (diameter 10 to  $14 \mu m$ ) which comprise a surface area of nearly  $150 m^2$  (Gehr et al. 1978) is so low that 5 to 10l of blood can flow through the lung each minute with a pressure of less than 1.3 kPa (Comroe 1974). In fish, gill vascular resistance is one half to one third that of the systemic circuit (Cameron et al. 1977), with the difference being much greater (about ten times) in mammals and birds (Langille and Jones 1975). In the amphibious ghost crab, Ocypode saratan, the gills are perfused both in submerged and air-breathing crabs. The lungs are preferentially perfused in air but not during submergence (Al-Wassia et al. 1989).

### 2.7.2.2 Ventilation

The need to reduce cutaneous water loss, especially with advent of terrestrial habitation, necessitated development of an impermeable skin. This rendered much of the body surface nonrespiratory. Specialized respiratory sites, where soft and well-vascularized parts could be exposed to the ambient respiratory medium, formed. For such areas to be effective, convective movements of air or water through mechanical effort were necessary in order to create and sustain a satisfactory partial pressure to maintain O<sub>2</sub> influx. Ventilation entails mass renewal of the environmental medium in the immediate proximity of a respiratory surface. Except for the freshwater limpets, Ancylus fluvialis and Acroloxus lacustris (Berg 1951), invertebrates which subsist in running water (i.e., are passively ventilated) show a higher metabolic rate than those from stagnant water (Fox et al. 1935; Walshe 1948). The convective systems which have evolved differ remarkably and reflect heavily on the restrictions imposed by the physical characteristics of the respiratory media on the design of the exchangers (Fig. 18). In simple aquatic animals, e.g., the bivalve mollusks and amphioxus (e.g., Baskin and Detmers 1976), and ascidians, the movement of water across the gills is effected by cilia. The much more sophisticated branchial pumps move the water across the gills in the advanced fish. Tubiculous polychaetes rely on ciliary currents (e.g., Nephtys),

peristalsis (e.g., Arenicola) or undulating movements of the body (e.g., Chaetopterus). In mollusks, where the gills are largely used both for feeding and respiration, water is moved across the gills by the beating of cilia which are located on the gills (Borradaille et al. 1963). The echinoderms respire through movable tubes (podia) which extend from basal dilatations (ampullae), structures which protrude through openings in the calcaneous outer covering of the body (Hyman 1955; Steen 1965). The hemolymph, which contains no respiratory pigments, is moved through ciliary action into the microcirculatory units. In the sea urchin, Strongylocentrotus droebachiensis, the podial respiratory surface area becomes limiting to O<sub>2</sub> consumption only at higher temperatures (Steen 1965): at 19°C, O<sub>2</sub> consumption (2mlh<sup>-1</sup>, 70g body mass) is directly proportional to the available respiratory area, while at 6 °C, only 20% of the available surface is utilized to transfer O<sub>2</sub>. In S. droebachiensis, a 70-g specimen has some 100 podia which are about 20 mm long and 0.4 mm in diameter. The overall respiratory surface area is about  $250 \,\mathrm{cm}^2$  and the thickness of the diffusional pathway is about  $15 \mu m$ . The soft-bodied cucumbers (Holothuroidea) use an internal respiratory tree-like organ which they rhythmically ventilate through muscular contractions. Holothuria tubulosa renews the water of the respiratory tree every 1 to 4 min, with the expelled water having an O<sub>2</sub> content of about 50 to 80% of that of the surrounding water (Hazelhoff 1939). The marine annelid, Chaetopterus variopedatus, a burrow-dwelling filter feeder, has a high mass-specific ventilatory rate of the burrows of about  $110 \text{ ml g}^{-1} \text{h}^{-1}$  at 15 °C, a low O<sub>2</sub> extraction coefficient (the ratio of the amount of O<sub>2</sub> taken up to that available in the inspired medium) of about 30% and an  $O_2$  consumption of 11  $\mu$ mol g<sup>-1</sup> h<sup>-1</sup> in an actively ventilating organism of 4g wet weight (Dales 1969). Fish embryos develop a rhythmic contraction of the tail muscles before the respiratory movements begin and show motor response to hypoxia (Polimanti 1912). To a slight extent, beating of cilia moves water currents into the molluskan mantle cavity. Some burrowing annelids, e.g., the marine echiuran worm, Urechis caupo (Wells 1949; Mangum 1985), generate a water current over their bodies and in the tubes by waving their bodies in water through peristaltic contractions of their muscular body wall. The mud shrimp, Callianassa truncata, a species which inhabits sand sediments in the Mediterranean Sea makes burrows which may be as much as 8 m deep (Ziebis et al. 1996): at a depth of 48 cm, the shrimp can maintain burrow O<sub>2</sub> concentration at 3 to 12% of air saturation by generating a water current of 10m per second. Rhythmic movements of the external gills in the urodele, Necturus, renew the water on the surface of the gills. Increases in the PO<sub>2</sub> of the water in the burrows were recorded after a short period of irrigation (mean duration 21s) produced by body undulations of the snake blenny, Lumpenus lampretaeformis (Atkinson et al. 1987): a flow rate of  $40 \text{ ml min}^{-1}$  in a burrow of a diameter of 2 cm was produced. Depending on the PO<sub>2</sub> in the water, the red band fish, Cepola rubescens, irrigates its burrow by body movements (Pullin et al. 1980) creating a water flow of  $10 \text{ mlmin}^{-1}$  in a burrow of a diameter of 5 cm. In areas where the water flow is fast, the design of the burrows and the locations of the opening(s) relative to the direction of water flow may create pressure gradients which may passively suck water into the burrow (Vogel 1977). Passive ventilation is thought to occur in the burrows of the tile fish, *Lopholatilus chamaeleonticeps* (Grimes et al. 1986), and the mud-shrimp, Callianassa truncata (Ziebis et al. 1996). At a PO<sub>2</sub> of about 5kPa, the lug worm, Arenicola marina stops ventilating the burrows (Toulmond 1991). Below the critical  $PO_{22}$  it is clearly no longer cost-effective to expend energy ventilating the burrows. In the shelled Nautilus (a paleontological relic of more than 2000 extinct genera of nautiloids and ammonoids) where mantle movement is not possible, the ventilatory stream is generated by movements of fused collar and funnel folds, the "wings", which create small pressure gradients of the order of 0.1 kPa (Wells and Wells 1985). The ventilatory frequency, which is 35 times min<sup>-1</sup> at 16°C, increases with temperature and the stroke volume ranges from 5 to 22 ml for a 395-g animal. The volume of the mantle cavity in a fixed specimen of Nautilus (470 g) is 75 ml (Packard 1972). In Octopus vulgaris, the number of respiratory movements per minute decrease with increasing body mass (Polimanti 1913). Through a process called apneic oxygenation (Malan 1982; Szewczak and Jackson 1992), O2 moves from the atmosphere down the trachea to the lung by diffusion or by bulk convection. If the respiratory quotient is greater than 1, in some hibernating animals with a low metabolic rate and long apneic periods, a significant amount of the resting O<sub>2</sub> needs can be met by diffusion down the respiratory tract through an open glottis during nonrespiratory periods. Ventilation by oscillatory movements of special appendages occurs in some polychaetes (e.g., Chaetopterus), amphipods (e.g., Gammarus), isopods (Idotea), and crustaceans, e.g., Cancer pagurus (Bradford and Taylor 1982). Such currents may deliver food, e.g., in Chirocephalus, Artemia, and Daphnia. In animals which live in torrential, air-saturated waters (rheophilic species) where the  $PO_2$  of the water next to the gas exchanger is equal or almost equal to the atmospheric one, e.g., the hill-stream fish like Danio dangila which subsists in hyperoxic water with a concentration of  $O_2$  of  $9 \text{ mg } O_2 l^{-1}$  (Ojha and Singh 1986), the gills and skin are passively ventilated with water in an energysaving process. In the oegopsid cranchid squids, which are known to store ammonia in the enlarged coelomes to regulate buoyancy (Denton et al. 1958), the mantle contractions do not participate in respiration. The flow of water over the gills is effected by movements of the coelom (Clarke 1962). The urodele salamanders (Plethodontidae), which lack lungs or gills, rely entirely on cutaneous respiration. Movements which stir the external respiratory medium and subcutaneous perfusion act as the only aids to the diffusive conductance across the skin (e.g., Gatz et al. 1974; Piiper et al. 1976).

The convection requirements are high in water breathers compared with air breathers. The requirements correlate inversely with the concentration of the molecular  $CO_2$  of the medium. Due to the relatively low concentration of  $CO_2$  in water, aquatic animals exhibit ventilatory rates 10 to 30 times those of the air breathers (White 1978). Increased ventilation in response to declining environmental  $O_2$  has been observed in marine and freshwater bivalves (Zinkler 1966; McMahon 1988), crustaceans (Hughes et al. 1969a; Taylor 1982), polychaetes and oligochaetes (Mangum 1963), and holothurians (Newell and Courtney 1965). In cephalopods, hypoxia results in reduction of respiratory movements (Fredericq 1878) while activity (Ghiretti 1966) and hypercapnia (Winterstein 1925) elevate it. In some crustaceans, e.g., the crayfish, *Astacus leptodactylus* (Angersbach and Dekker 1978), and the crabs, e.g., *Cancer productus* (McMahon and Wilkens 1977)



Fig. 22. (a) Gills of the freshwater crab, *Potamon niloticus* (g), showing a scaphognathite, p, the afferent gill artery,  $\rightarrow$ , and the efferent artery,  $\succ$ ; c skeletal mass. (b) A closeup of a gill arch showing gill lamellae, s, and afferent, a, and efferent, e, blood vessels. (a) *Bar* 500 µm; (b) 200 µm. (Maina 1990b)

and Cancer pagurus (Bradford and Taylor 1982), special mouth part appendages (the scaphognathites - flattened exopodities of the maxillae) ventilate the gills which are covered by lateral extensions of the carapace lined by a membrane, the branchiostegite (Lockwood 1968; Burggren et al. 1974; Fig. 22). The flow of the water through the branchial chamber is adequately effective to maintain a PO<sub>2</sub> of 15 to 20 kPa at the respiratory surface of the gills in normoxia (Butler et al. 1978; Wheatly and Taylor 1981). The PO<sub>2</sub> in the arterialized haemolymph is as high as 10 to 13 kPa (Butler et al. 1978; McMahon and Wilkens 1983). The direction of air flow can be reversed in some species of crabs, e.g., Carcinus maenas and C. guanhumi (Taylor and Butler 1978; Burggren et al. 1985a). It has been established that in some species of crabs (e.g., Yonge 1947; Hughes et al. 1969a), and the larvae of air-breathing fish (Liem 1981) the interaction between water flow and that of blood (in the gill lamellae) is countercurrent. In the species which burrow into the substratum, the direction of the ventilatory current is reversed when the animal is buried in the sediment (Arudpragasm and Naylor 1964a,b; Dyer and Uglow 1978). Except in Holthuisana transversa (Greenaway and Taylor 1976; Taylor and Whitley 1979), where tidal ventilation is achieved by movements of the membranous thoracic wall, the scaphognathites are effective in ventilation of the gills and the lungs of the land crabs while in air (Taylor and Butler 1978; Burggren et al. 1985a; Al-Wassia et al. 1989) especially during hypercapnia. Depending on ambient temperature (Taylor and Wheatly 1979), the shore crab,

Carcinus maenas, occasionally partially emerges from water to bubble air through the branchial chamber, aerating the water it holds in it (Taylor and Butler 1973; Taylor et al. 1973; Taylor and Whitley 1979). The movements of the scaphognathites of Coenobita clypeatus in air create pressure wave forms corresponding to those generated by the scaphognathites of water breathers (McMahon and Burggren 1979). The ventilatory mechanism in H. transversa (Taylor and Greenaway 1979) is unique among land crabs. Its efficiency (of which the operational definition is mechanical work output per metabolic work input) approximates to that of vertebrate respiration of about 10%. In Ocypode saratans, the scaphognathites beat at a rate of 53 times min<sup>-1</sup> in a submerged crab, 218 times min<sup>-1</sup> when active in air, 43 times min<sup>-1</sup> when inactive in air, and 235 times  $\min^{-1}$  when exposed to hypercapnia (Al-Wassia et al. 1989). The intermittent beating of the scaphognathites is neurogenically synchronized with that of the heart (Young 1978; Young and Cover 1979). By changing the dimensions of the gill lamellar blood vessels, scaphognathite movements which generate pressures of -0.53 to 0.93 kPa may play a significant role in the perfusion of the gills and the lung (Blatchford 1971). In the crab, Carcinus maenas (Taylor et al. 1973; Wheatly and Taylor 1979) and the crayfish, Orconectes rusticus (McMahon and Wilkes 1983), air is bubbled through water held in the branchial cavity by the scaphognathites beating in the reverse direction. In the amphibious ghost crab, Ocypode saratan, heart rate varies with ventilation (Al-Wassia et al. 1989): the lungs are not perfused while in water, but in air the lungs are perfused at four times the rate of the gills. The routine breathing frequencies in the juvenile and adult fish range from 30 to 70 times per minute (Roberts 1975) and ventilatory flows range from 100 to 300 mlkg<sup>-1</sup>min<sup>-1</sup> (Wood et al. 1970; Johansen 1982). Functional coupling between respiration and locomotion occurs in some tetrapods. This may be a means of improving gas exchange efficiency or a saving on respiratory work. When a lizard runs, the left and right lungs are alternately compressed, pumping the air between the two lungs (Carrier 1987a,b). However, no significant movements of air from outside occur. In bats, during flight, synchronization between wing beat, respiratory rate, and heart rate has been reported (e.g., Thomas 1987) but in birds, this occurs in only a few species (Torre-Bueno 1985). In the horse and many mammals, breathing is closely coupled with locomotion (Bramble and Carrier 1983).

Gas exchangers have evolved in form of invaginations or evaginations from the body surface (Figs. 4,5). The former are generally categorized as gills and are largely used in aquatic respiration while the later are termed lungs and are used for aerial respiration. Gills are unidirectionally ventilated while tidal ventilation occurs in the lungs (Fig. 6,18). Compared with other vertebrates, fish exhibit diverse ventilatory mechanisms. These range from active and passive (ram) ventilation, continuous or intermittent unidirectional flow of water across the gills, to tidal process in the air-breathing organs of the bimodal breathers. In a four-phase serial pressure suctional buccal force pump, the gills are ventilated with a constant flow of water which is taken into the mouth and forced out through the opercular flap (Liem 1985; Brainerd 1994). Pressure differences of about 0.4kPa fill the mouth and those of 0.7 to 1.3kPa move the water across the gills (Ogden 1945; Hughes and Shelton 1958). Fish which swim strongly and much of the time such as the mackerel, the tuna, and some sharks move with their mouths open. Such fish have lost the capacity for mechanical gill ventilation and rely on ram effect (passive ventilation of water powered by swimming) for ventilating their gills. Because of the atrophy of the brachiomeric muscles, active ventilatory rate is very low in the group. Such fish cannot maintain an optimal level of oxygenation of the blood if they are held in restricted enclosures where forward movement is prohibited. The bimodal breathing fish use a slightly altered buccal force pump (analogous to that utilized on the gills) to ventilate their accessory respiratory organs. Air is forced into the organ(s) and exhalation is thought to be a passive process, especially in those air-breathing fish which have a fixed air space (e.g., a suprabranchial chamber) or through pulmonary elastic tissue recoil (DeLaney and Fishman 1977; Farrell and Randall 1978; Liem 1980). In some airbreathing fish, e.g., Anabas testudineus and probably in Clarias mossambicus (Maina and Maloiy 1986), where the inhalant and exhalant apertures are respectively contained in the pharyngeal and opercular cavities, the buccal and opercular pumps effect a unidirectional air flow across the labyrinthine organs which are found in the suprabranchial chamber (Peters 1978; Liem 1980). The cephalopods and mollusks have evolved a throughflow ventilatory mechanism which falls between the bidirectional one of the air breathers and the unidirectional one of fish: the inspired and expired streams flow through different openings over the gills, with some mixing probably occurring in the mantle cavity (e.g., Packard 1972; Gosline et al. 1983; Bone et al. 1994). With an O<sub>2</sub> extraction factor of 33 to 72%, e.g., in the scallops (van Dam 1954), and a possibility that the arterial  $PO_2$ may in some cases be higher than that in the effluent water in the octopus (Johansen and Lenfant 1966), the throughflow ventilatory mechanism is exceptionally efficient. In *Nautilus*, there are two pairs of gills instead of one and the funnel is the main contractile structure (Ghiretti 1966). In the holothurians, e.g., sea cucumber and the cephalopods, the cloaca pumps water tidally across the ramified diverticula of the hind gut which forms the respiratory tree. In the sea cucumber, Holothuria forksali, 60% of the total O<sub>2</sub> needs is taken from O<sub>2</sub>saturated water across the cloaca, the remainder passing over the skin (Newell and Courtney 1965): O<sub>2</sub> consumption decreases with arrest of cloacal respiration. About ten successive cloacal contractions, each of which transfers 1 ml of water into the animal, are followed by body contractions which inject it out. Below an ambient PO<sub>2</sub> of 12kPa, the organisms respond by respiratory inhibition and relocation to better oxygenated sites (Lutz 1930). Tidal breathing of water across the hind gut also occurs in annelids, e.g., Urechis caupo (Pritchard and White 1981; Menon and Arp 1992a), and in insects, e.g., dragon-fly nymph Aeshna (Fig. 23), via the branchial openings in lamprey eels (Johansen 1971) and through the mouth in some amphibians and reptiles, e.g., the soft-shelled turtle, Amyda. Such rather simple organs, of which the primary functions are olfactory and/or gustatory, are collectively called water-lungs (Sect. 4.6). Contrary to what one would expect from morphological evidence, a countercurrent exchange may occur in the lamprey during slow inspiratory phase (Johansen 1971). At normoxia, hind gut ventilation with seawater in U. caupo is about 0.7 ml per g body mass and at a PO<sub>2</sub> of 4kPa rises to about 2ml of seawater  $g^{-1}$  body mass (Julian and Arp 1992; Menon and Arp 1992a). The mucosa of the hind gut contains collagenous and



Fig. 23. a Rectal tracheal gills of the dragonfly. b Enlargement of a gas exchange site and c the gas exchange mechanism between water and air in the trachea. The rectum is tidally ventilated, ↔. (Hughes 1982)

Ventilatory process	Respiratory milieu	Vertebrate taxon	Environment
Buccal and opercular pumps with unidirectional flow of the medium	Water Air	Fish (gills) Some air-breathing	Aquatic Aquatic
Modified buccal force pump with tidal flow of medium	Air	Most air-breathing fish Dipnoi Amphibians	Aquatic
Aspiration pump with tidal flow of medium	Air	<i>Arapaima</i> Reptiles Birds Mammals	Aquatic Majority terrestrial, but some aquatic forms

**Table 5.** Distribution of the ventilatory mechanisms in vertebrates and the respiratory media inwhich they occur. (Randall et al. 1981)

elastic fibers which may allow greater stretching during filling under hypoxic conditions (Menon and Arp 1992a). A similar rectal-gill mechanism exists in the echiuran worm, *Arhynchite pugettensis* where extensive cloacal diverticula occur (Manwell 1960). In the diving turtle, *Stemothaerus minor*, 30% of the O<sub>2</sub> need is met by rhythmic gular movements which maintain movement of water in and out of the bucco-pharyngeal cavity (Belkin 1968).

In aquatic animals, buccal pumping is the ancestral mode of ventilation of the gills (e.g., Brainerd et al. 1993; Brainerd 1994). It may initially have been used to ventilate the accessory respiratory organs on attainment of utilization of atmo-

spheric  $O_2$  (Randall et al. 1981). Aspirational (suctional) breathing characterizes air breathers while buccal pumping occurs in both water and air breathers (Table 5). Buccal and aspirational breathing coexist in the obligate air breather *Arapiama gigas*, where the gas bladder is ventilated by aspiration and the gills by a characteristic teleostean buccal force pump (Farrell and Randall 1978; Randall et al. 1978a). During buccal pumping in the frog *Rana pipiens*, work is done by the floor of the mouth (against the elastic forces of the lung) at an average efficiency of about 8%. The  $O_2$  cost of breathing constitutes about 5% of the total energy budget (West and Jones 1975). In *Rana*, the cost of respiration is about two times that in the human being but falls within the same range of fish.

## 2.8 Blood and the Respiratory Pigments

The increase in the complexity and efficiency of the gas exchangers has been a gradual process which has developed in response to specific needs that have called for greater and more efficient means of O<sub>2</sub> uptake and delivery to the tissues to support higher aerobic capacities. As in practically all transformations in biology, improvements were made on former simpler designs (e.g., Schaeffer 1965a,b; Riggs 1976). With the evolution of a circulatory system followed by a ventilatory mechanism, O2 was initially carried in solution in the body fluids. In the ensuing period, demands must have exceeded supply, rendering the process inadequate. Respiratory pigments have evolved widely in the Animal Kingdom (e.g., Antonini 1967). They reversibly bind, store, and transport  $O_2$  (and to an extent  $CO_2$ ), increasing the  $O_2$  carrying capacity of blood (Bauer 1974; Jensen 1991). The pigments are found dissolved in the plasma or in a two-phase system where the carrier is packaged in corpuscles, especially in those animals which require efficient internal fluid transport of O<sub>2</sub> to the body tissues (Lamy et al. 1985; Burggren et al. 1991). The primitive type of blood or hemolymph which contains no respiratory pigments has essentially the same respiratory capacity as salt water and can carry only about 0.2 ml O<sub>2</sub> per 100 ml water while vertebrate blood can carry 5 to  $45 \text{ ml O}_2$  per 100 ml blood. In fish, the physically dissolved O<sub>2</sub> usually constitutes less than 5% of the total O<sub>2</sub> carried in blood (Boutilier et al. 1984). The development of hemoglobin increased the O<sub>2</sub> carrying capacity of blood by about 100 times. In the crustaceans, the presence of hemocyanin increases the O<sub>2</sub> carrying capacity of the hemolymph above the dissolved levels by a factor of 2 to 4 (Taylor 1982; Mangum 1980, 1983a,b; Shiga 1994). The arterial PO<sub>2</sub> (PaO<sub>2</sub>) in resting crustaceans is reported to be generally low (1 to 3kPa at 13 to 15°C) independent of the blood pigment concentration (Forgue et al. 1992a,b). This baseline value corresponds with that of the mussel Anodonta cygnea which lacks blood pigment (Massabuau et al. 1991). Compared with the high values of PaO<sub>2</sub> (range from 2 to 13kPa) that have been reported by Shelton et al. (1986) and McMahon and Wilkens (1983) in the water breathers, hemocyanin gives crustaceans a large functional reserve for uptake and transfer of O<sub>2</sub>. Theoretically, in the human being, if O<sub>2</sub> was carried in physical solution in blood instead of by the hemoglobin, the circulatory rate would have to be 30 times more to meet the metabolic demands. Interestingly, the  $O_2$  content of blood is equal to that of air, i.e., 20 ml  $O_2$  per 100 ml (Davenport 1974; West 1974). This suggests a possible optimization of chemical binding and transfer of  $O_2$  by the blood.

The development and refinement of the respiratory carriers added a significant factor to the gas exchange capabilities in animals, increasing the adaptability of the respiratory system to different environments. This entailed impressive molecular creativity directed at meeting the special needs of organisms (Perutz 1970, 1990a; Manning et al. 1990). For example, in the dimeric hemoglobin of the blood clam, Sapharca inaequivalvis, the hemes and the heme-linked helices E and F of adjacent subunits are in contact instead of facing outwards as is the case in the vertebrate hemoglobins (Royer et al. 1985). Certain invertebrate species have hemoglobins in which each polypeptide relates with multiple heme binding domains. Polymeric globin is thought to have occurred through fusion of multiple monomeric globin transcriptional units at the gene level in organisms ancestral to the invertebrate lineage some 200 to 500 million years ago (Manning et al. 1990). The tetrameric hemoglobin of the fat inn-keeper worm, Urechis caupo, shows neither cooperativity of O<sub>2</sub> binding nor a Bohr effect, while the hemoglobin of the brine shrimp, Artemia, the blood clam, Sapharca, and the earthworm, Lumbricus, displays both properties (see Perutz 1990a,b). Every vertebrate hemoglobin is an oligometric protein comprising four polypeptide subunits (protomers), each of which possesses an O<sub>2</sub> binding protoheme (Bauer 1974). While the fish hemoglobins are  $\alpha_2\beta_2$  tetramers with a tertiary and quaternary structure similar to that of other vertebrate hemoglobins, among the teleosts, the amino acid sequences differ greatly between different groups (Kleinschmidt and Sgouros 1987). Many of the differences are functionally neutral (Perutz 1983).

Physiologically, the most important properties of hemoglobin are the cooperativity of  $O_2$  binding and the effect of  $H^+$ ,  $CO_2$ , and organic phosphate components on the affinity of hemoglobin for  $O_2$ . Cooperativity arises from the change of hemoglobin from low - to high-affinity forms with the binding of O<sub>2</sub> (e.g., Hewitt et al. 1972; Ten Eyck 1972; Perutz 1979). The limit to O<sub>2</sub> storage capacity in blood is set by the product of the respiratory pigment concentration and the blood volume (e.g., Davenport 1974). In general, diving animals have a larger blood volume than the nondivers (e.g., Bond and Gilbert 1958; Butler 1991a). They have a greater  $O_2$  storage capacity of blood (Ridgway and Johnston 1966; Lenfant et al. 1969; Hedrick and Duffield 1986). During submersion, the concentration of the hemoglobin increases by a factor of 60 to 70% in the blood of the Weddell seal (Qvist et al. 1986). In the winter months, when the muskrat has to dive under water to look for food, its hemoglobin concentration in blood is at the high of 20g per 100 ml of blood, while in summer, when the animal stops diving, the value drops to 14g per 100 ml (Aleksiuk and Frohlinger 1971). In the aquatic pneumonate gastropod, *Planorbis corneus*, and several species of *Daph*nia, a low PO, initiates almost instantaneous synthesis and increase in the concentration of the hemoglobin (Fox 1955), a feature which has great survival value. Hemoglobin plays an important role in O<sub>2</sub> transport especially in hypoxic habitats (e.g., Johnson 1942; Cosgrove and Schwartz 1965). The lugworm, Arenicola marina, which has adapted to the anoxic intertidal sediment mainly because of its efficient anaerobic metabolic pathways, has a high O<sub>2</sub> affinity hemoglobin (Toulmond 1985) and two kinds of body wall myoglobins of very high  $O_2$  affinity ( $P_{50}$ , 0.1 and 0.2 kPa, at 20 °C) (Weber and Pauptit 1972). Presence of a hemoglobin with very high  $O_2$  affinity ( $P_{50}$  about 0.13 kPa) was reported in specialized cells of gastrotrichs by Colacino and Kraus (1984). The alvinellids, polychaete annelids which live in the deep-sea hypoxic hydrothermal vents, have complex morphometrically well-adapted gills (Jouin and Toulmond 1989; Toulmond 1991). *Alvinella pompejana* has a closed vascular system (Toulmond 1991) and a high molecular mass extracellular hemoglobin (Terwilliger and Terwilliger 1984) with a high  $O_2$  affinity ( $P_{50}$ : 0.02 to 0.3 kPa – measured at 20 °C, 1 atm pressure and pH 7.6 to 6.6; Toulmond 1991). Complex molecular mechanisms which included the effect of pH,  $CO_2$ , organic phosphates, and temperature developed to regulate  $O_2$  uptake and transport by the hemoglobin (Nikinmaa 1990). Of the total proteins in the vertebrate erythrocytes, hemoglobin constitutes 95% (Antonini 1967).

The evolution of carrier pigments in blood (Fig. 24) constituted a significant improvement on the respiratory function and is probably one of the most recent innovations in the complex gas exchange adaptive strategies. The presence of blood pigments enhanced the rate of diffusion of O<sub>2</sub> even at low PO<sub>2</sub>s (e.g., Hemmingsen 1963; Moll 1966). In the mammalian skeletal muscle (Wittenberg and Wittenberg 1989) and the heart muscle (Braulin et al. 1986), myoglobin facilitates diffusion of O<sub>2</sub> and in the fish cardiac muscle, it enables the hearts of some fish to extract O<sub>2</sub> at lower levels of ambient PO<sub>2</sub> than would otherwise be possible (Driedzic and Gesser 1994). Polar icefish (e.g., Douglas et al. 1985) and interestingly even nonpolar fish such as lumpfish (Cyclopterus lumpus), monkfish (Lophius piscatorius), and ocean pout (Macrozoarces americanus) have insignificant myoglobin content in their tissues (Driedzic and Stewart 1982; Sidell et al. 1987). On the other hand, some species of fish such as tuna (Thunnus thynnus), mackerel (S. scombrus), and the carp (Cyprinus carpio) have remarkably high levels of myoglobin in their hearts, respectively 580, 332, and 488 nmol per g wet heart mass (Giovane et al. 1980; Sidell et al. 1987). High myocardial myoblobin characterizes fish that have high swimming capacities and those tolerant to hypoxia (Giovane et al. 1980; Driedzic 1988; Driedzic and Gesser 1994). Presence of myoglobin enables the heart muscle to function at a lower level of extracellular PO<sub>2</sub> than would otherwise be possible (Braulin et al. 1986; Taylor et al. 1986).

Bohr and Root effects of the hemoglobin (decrease in  $O_2$  affinity of blood) boost diffusion by increasing the blood-to-tissue  $O_2$  gradient at the sites of  $CO_2$ production. Blood hemoglobin concentration is increased in hypoxic mammals (Lenfant 1973). In fishes, both hypoxia and exercise highly elevate the blood hemoglobin level (Kiceniuk and Jones 1977; Weber and Jensen 1988). A high hemoglobin concentration in blood achieves a high blood  $O_2$  capacitance coefficient, decreasing the pumping requirement of the heart (Jensen 1991). In the diving animals, myoglobin enhances the amount of  $O_2$  which can be stored in muscle tissue. The concentrations increase with the increasing diving ability (Butler 1991a). In the physoclistic swim bladders, secretion of lactic acid from glucose metabolism at the gas-gland of the rete mirabile (Pelster and Scheid 1991, 1993) forces  $O_2$  out of the hemoglobin (and  $N_2$  out of solution) compressing them into the bladder (e.g., Pelster and Scheid 1992a,b). Through still unknown mechanisms, the Root effect (pH-dependent release of  $O_2$  from the hemoglobin) is



Pigment	Environment	Occurrence	Comments
Hemoglobin	Intracellular (RBCs)	Nemertines, annelids, mollusks (bivalves), Phoronida, Echiurida, Echinoderms, Hemichordates, Chordates	Small: monomers to octamers
	Extracellular	Annelids, mollusks, arthropods	Highly variable
Chlorocruorin	Extracellular	Annelids	Formyl substitution on protoporphyrin
Hemerythrin	Intracellular (RBCs)	Brachiopods, annelids, sipunculids, priapulids	Nonporphyrin iron
Hemocyanin	Extracellular	Mollusks, arthropods	Probably of different origin in the two taxa

Table 6. Distribution of oxygen-carrying pigments in different groups of animals. (Cameron 1989)

controlled by proton-induced allosteric conformational changes in fish hemoglobins (Howlett 1966). Hemoglobin may have evolved from the ubiquitous cytochrome molecule, a complex multicatalyst (Bernhardt 1995), with which it shares a common porphyrin nucleus and which has been implicated with facilitated diffusion of  $O_2$  in some tissues (e.g., Scholander 1960; Burns and Gurtner 1973; Longmuir 1976). The incorporation of hemoglobin in the convective transport of blood in the vertebrates (Fig. 20) fully exploited the O2 delivery role of the molecule. The large interspecific differences in the blood O<sub>2</sub> capacity, hemoglobin affinity, the extent of Bohr shift and erythrocyte morphometry and morphology (e.g., Riggs 1979; Dejours 1988), features which do not strictly correlate with phylogeny, environment, and mode of respiration (Fig. 24, Table 6) are strongly indicative of an ongoing intricate optimizing process. The carriers as well as other molecular factors are probably still being refined and integrated into the cardiovascular and respiratory systems for  $O_2$  uptake and transfer. For example, as a means of ascertaining efficient  $O_2$  delivery to the tissues by regulating peripheral blood pressure, while the erythrocytes are being oxygenated in the lung, the hemoglobin is S-nitrosylated to form S-nitrosohemoglobin (Jia et al. 1996; Perutz 1996). Subsequently, nitric oxide (NO), a recently recognized potent endothelium-derived relaxing factor (e.g., Palmer et al. 1987; Koshland 1992; Nathan 1992) is released during arterial-venous transit, causing vasodilation. The hemebound NO [Hb(Fell)NO] is detectable in the venous blood when animals are subjected to oxidative stress (Kagan et al. 1996). Nitric oxide has, moreover, been reported to increase glucose transport in skeletal muscle (Balon and Nadler 1997). By increasing blood flow, nitric oxide ameliorates the effect of carbon dioxide on the brain (Meilin et al. 1996). Since the reaction between hemoglobin and  $O_2$  is exothermic, increase in temperature results in a corresponding decrease in  $O_2$ affinity, an adaptive feature especially in the homeotherms where high temperatures prevail in exercising tissues (e.g., skeletal muscle). A decrease in O<sub>2</sub> affinity promotes O<sub>2</sub> unloading. The hemoglobin in the tuna, *Thunnus*, a fish with exceptionally well-developed myogenic endothermia (e.g., Carey and Teal 1966) and high body temperature differentials, is insensitive to heat (Johansen and Lenfant 1972). Chum salmon have multiple hemoglobins, one group with a low Bohr effect and low temperature sensitivity and the other with normal proton and temperature sensitivity (Hashimoto et al. 1960). These forms are adaptively mobilized depending on the ambient temperature fluctuations. In facultative air breathers such as Hypostomus species and Pterygoplichthys, O<sub>2</sub> binding properties depend on the mode of breathing (Weber et al. 1979). While breathing air, the blood  $O_2$  affinity increases and the Bohr effect decreases mainly owing to a drop in the erythrocytes organic phosphate concentration. Reptiles, a group which typically experience sudden body temperature changes especially as they bask in and withdraw from the sun, have a somewhat temperature-insensitive hemoglobin (Sullivan and Riggs 1967; Wood and Moberly 1970).

The effect of body size on the magnitude of Bohr effect and hemoglobin-O<sub>2</sub> affinity has been highly debated. Conflicting results ranging from a direct relationship (e.g., Schmidt-Nielsen and Larimer 1958), an inverse one (e.g., Clausen and Ersland 1968), and a weak or no correlation (e.g., Hilpert et al. 1963; Lahiri 1975) have been reported. The divergence may be characteristic of a nonoptimized state. The amphibians, a group which is highly instructive in the study of evolution of respiratory processes, show dramatic ontogenetic transformations in hemoglobin function (e.g., Gahlenbeck and Bartels 1970; Wood 1971). Changes in O<sub>2</sub> availability accompany the metamorphosis of a water-breathing tadpole to the adult air breather (Wood 1971; Broyles 1981). A notable decrease in O<sub>2</sub> affinity, increase in O<sub>2</sub> carrying capacity of blood, and metabolic rate occur with changes from larval to adult stages. With some exceptions, e.g., in the tuna and the lugworm, Arenicola, the concentration of the pigments is higher in the air breathers than in water breathers (Toulmond 1975; Dejours 1988). The O<sub>2</sub> affinity of the hemoglobin of the aquatic breathers which live in well-oxygenated water, e.g., the mackerel and salmon, are similar to those of air-breathing forms in the same habitat. This indicates that availability of O<sub>2</sub> rather than the nature of the respiratory medium influences O<sub>2</sub> affinity. Air breathers generally have a larger blood O<sub>2</sub> capacity, lower hemoglobin-O<sub>2</sub> affinity, and a larger Bohr shift than water breathers, at least in closely related species (Johansen and Lenfant 1966; Johansen et al. 1978; Fig. 63). Based on a wide cross section of species from the two taxa, a discordant view has, however, been expressed by Powers et al. (1979). Environmental PCO<sub>2</sub> and pH appear to determine the blood O<sub>2</sub> capacity of particular fish. At the physiological blood pH of 7.8 to 8.4, the Bohr effect is totally nonexistent in the blood of *Tilapia grahami*, which lives in the alkaline (pH 9.5 to 10.5) Lake Magadi of Kenya (e.g., Lykkeboe and Johansen 1975). In comparison with terrestrial mammals, the hemoglobin in diving animals has a low O<sub>2</sub> affinity, a high  $O_2$  carrying capacity, and a large Bohr effect (Andersen 1966; Lenfant et al. 1970a; Wood and Johansen 1974). In the tench, *Tinca tinca*, hypoxia-hypercapnia causes an increase in hematocrit (due to an increase in the erythrocyte volume) associated with reduced intracellular concentration of the hemoglobin (Jensen and Weber 1985).

Increased capillarization of the tissues, a process which minimized the transcapillary-tissue diffusional distance and development of tissue-based high affinity O2-storing noncirculating hemoglobin factor, myoglobin (found in some tissues) (e.g., Manwell 1963; Kreuzer 1970), were important innovations which enhanced delivery of O<sub>2</sub> to the cells. In the chiton, *Chryptochiton stelleri*, myoglobin has a lower  $P_{50}$  (0.4 kPa) than circulating hemocyanin, 2.7 kPa (Manwell 1958). In the buccal muscle of the mollusk, Aplysia deplians, the hemoglobin content is 6 mg% (Rossi-Fanelli and Antonini 1957) and in the human muscle tissue, myoglobin comprises about 2.5%. In his bucket-bridge model, Scholander (1960) envisaged that O<sub>2</sub> passes from one hemoglobin molecule to another. In this way, the O<sub>2</sub> flux can be increased eight times. It is, however, interesting that intracellular hemoglobin does not exist in the tissue barriers of the lung and fish gills. The role of hemoglobin in O<sub>2</sub>-facilitated transfer has been questioned by Hemmingsen (1965) and remains an unsettled issue. The early proposition that the  $O_2$  affinity of bird blood is lower than that of mammals (e.g., Jones 1972; Prosser 1973; but see dissenting views, e.g., Scheid and Kawashiro 1975 and Baumann and Baumann 1977) has been attributed to methodological error due to the time lapse between the collection of blood and analysis (Lutz et al. 1973, 1974; Holle et al. 1977). This may arise from the high metabolic rate of the nucleated avian ervthrocytes. Though the erythrocytes in 90% of the vertebrate species are nucleated (e.g., Nikinmaa 1990), annucleation is thought to be the more evolved feature. Nucleated erythrocytes generate energy aerobically and their transmembrane pathways are more diverse than those of the nonnucleated mammalian cells. Nonnucleated erythrocytes have been described in some fish (Hansen and Wingstrand 1960). Compared with annucleate erythrocytes, nucleated erythrocytes have lower deformability, greater orientation instability, and show a greater propensity to interact with each other during steady capillary blood flow (Gaehtgens 1990). In these respects, nonnucleated erythrocytes which encounter less resistance due to low viscosity provide a distinct advantage in O<sub>2</sub> transfer.

Paradoxically, in some organisms, the contribution of the respiratory pigments in gas exchange is highly questionable. Some choronomid larvae and insects lack an  $O_2$  carrier. Pigments appear to be of significance for life only during circumstances of high  $O_2$  demand. Goldfish behave normally in water equilibrated with 80% carbon monoxide and 20%  $O_2$  at temperatures below 20 °C (Anthony 1961). In the abalone, *Haliotes corrugata*, the concentration of the hemocyanin in different specimens was reported to differ by a factor of 900 times (Pilson 1965). In water equilibrated with air containing enough carbon monoxide to make hemoglobin totally inefficient as an  $O_2$  carrier, the eel, the carp, and the pike will live for hours (Nicloux 1923). Extended exposure to reduced  $PO_2$  results in manufacture of respiratory pigments in some animals but not in others (Fox 1955). In *Artemia*, hemoglobin synthesis is stimulated by high external salinity (Gilchrist 1954). The respiratory properties of the copper-containing hemocyanin

are highly labile (Mangum 1980a) with the blood of the cephalopods being very sensitive to pH changes (i.e., having a marked Bohr shift), temperature, ionic, and osmotic composition of blood (Houlihan et al. 1982). By increasing hemocyanin-O<sub>2</sub> affinity, Octopus vulgaris can cope with hypoxia even if 30 to 50% of the gill surface area is surgically removed (Wells and Wells 1984). In Sepia officinalis, the  $P_{50}$  is 0.4 kPa at a pH of 7.97 but is 9.3 kPa at a pH of 7.24 (Wolvekamp et al. (1942). Although a notable degree of evolutionary refinement of the hemocyanin-O<sub>2</sub> transport has occurred in the cephalopods (Mangum 1980b, 1990), O<sub>2</sub> transport in blood is the major limiting factor for power output in the taxon (Brix et al. 1989; O'Dor and Webber 1991). The O<sub>2</sub> carrying capacity of the blood in Loligo pealei is only one-half that of the hemoglobin-based bloods of vertebrates (Mangum 1990). The O<sub>2</sub> affinity of the hemocyanins in the crabs, Cancer magister (Terwilliger and Brown 1993) and Cancer productus (Wache et al. 1988), and the lobster, Momarus americanus (Olson et al. 1988, 1990b), changes with the stage of development. Hemolymph inorganic ions particularly divalent cations change with development (Brown and Terwilliger 1992) and influence the O<sub>2</sub> affinity and cooperativity of decapod hemocyanins (van Holde and Miller 1982).

The evolutionary biology of the respiratory pigments is an unsettled subject (e.g., Wells 1990; Nikinmaa 1990; Mangum 1992; Weber 1992). Their intertaxonomic distribution and functional diversity are intriguing (Jensen 1991). The best-known metalloprotein O2 carriers are hemoglobin, hemocyanins, chlorocruorins, and hemerythrins (Fig. 24, Table 6). With some exceptions, chlorocruorins are found in some polychaetes, hemocyanins predominate in the mollusks but not bivalves (Ghiretti 1966; Mangum 1980b) and crustaceans (McMahon 1985; Lallier and Truchot 1989) while hemerythrins are restricted in distribution, occurring in unrelated taxa such as in the polchaete, Magelona, most sipunculids, some brachiopods, and some priapulids. Some mollusks have hemoglobin while some have both hemoglobin and hemocyanin. Polychaetes have either chlorocruorin or hemoglobin while yet others, e.g., Serpula, have both types of pigments (Jones 1972). Among the gastropods, in contrast to a large number of Prosobranchia and Pulmonata, Ophisthobranchia do not seem to have a respiratory pigment (Ghiretti 1966). Lack or presence of a particular pigment does not appear to correlate with either the type of respiratory medium a species utilizes or its phylogenetic level of development. It is conjectured that the evolution of the respiratory pigments is polyphyletic (Mangum 1985). Depending on species, hemoglobin is found in blood corpuscles or dissolved in plasma, chlorocruorin and hemocyanins are found only in solution, and hemerythrin is found in cells (Ghiretti 1966; Mangum 1980a; Cameron 1989). The occurrence of pigment carriers in cells, an ubiquitous feature in the higher vertebrates, has been interpreted to be an evolutionary innovation enabling more efficient O<sub>2</sub> uptake and transfer to support the higher aerobic states. The intracellular location of the carriers is thought to maintain an optimal colloidal osmotic pressure and viscosity of plasma for an equivalent amount of protein (e.g., Snyder 1977), provide a more efficient intracellular control of allosteric modulators of the pigment's O<sub>2</sub> affinity (Gillen and Riggs 1973), and avoid loss of the small hemoglobin molecule through renal filtration. Mangum (1985) asserted that the erythrocyte evolved several millions of years before it became associated with the roles used to explain its derivation. Some animal groups have even lost the erythrocyte in preference for dissolved extracellular pigments (Cameron 1989). As argued elsewhere in this account, from their remarkable plasticity (a feature characteristic of least conserved parameters), the carrier pigments and especially the erythrocytes appear to be the terminal parts of the respiratory evolutionary chain. In the trial-anderror process of refining O<sub>2</sub> uptake and transfer, the erythrocytes may have been suddenly mobilized to contribute to the respiratory process. This apparently was not without certain costs. Schmidt-Nielsen and Taylor (1968; but see contrary conclusions by Snyder 1973 and Gaehtgens 1990) observed that at equal O<sub>2</sub> carrying capacity, the viscosity of the red cell suspension is higher than that of the hemoglobin solutions. The diffusing capacity of the rabbit lung for  $O_2$  is higher when the lung is perfused with a hemoglobin solution compared with when perfused with a washed human red cell suspension (Geiser and Betticher 1989). This is attributable to an extraerythrocytic diffusion resistance which arises from an unstirred plasma boundary layer (Coin and Olson 1979; Huxley and Kutchai 1983; Vandegriff and Olsen 1984; Yamaguchi et al. 1985). The resistance depends on the hydrodynamic conditions of the erythrocyte flow (Rice 1980) and the physical resistance offered by the lipid cell membrane and the hemoglobin (Fischkoff and Vanderkooi 1975; Kon et al. 1980). The stagnant plasma boundary layer of blood is as thick as 4.2 µm (Weingarden et al. 1982). In a trade-off process, in animals with particulate blood, i.e., where the hemoglobin is contained in cells, gas exchange efficiency may have been sacrificed for proper osmoregulation of the blood. Optimal O<sub>2</sub> uptake and transport in blood is established through compromises between factors such as hematocrit, hemoglobin concentration, body temperature, metabolic state, and erythrocyte morphology (Guard and Murrish 1975; Weathers 1976; Shepherd and Riedel 1982; Hedrick et al. 1986). The carriers which are found in cells have low molecular weights whereas the extracellular ones have high ones. The O<sub>2</sub> transporting function of respiratory pigments is dependent on interactions with organic and inorganic cofactors (e.g., Mangum and Lykkeboe 1979). The O<sub>2</sub> affinity of the vertebrate hemoglobins is decreased by the erythrocytes' organic phosphates (e.g., Weber and Jensen 1988). In the crustacean hemocyanin, L-lactate (Truchot 1980; Bridges and Morris 1986) and urate (Morris and Bridges 1986) increase O<sub>2</sub> affinity. In a self-regulating process, Lacetate and urate, products of anaerobiosis in decapod crustaceans (e.g., Bouchet and Truchot 1985; Czietrich et al. 1987), sustain O2 transport in conditions of hypoxia or intense activity. Through resisting Bohr shift with pH change (Truchot 1987), the arterial blood remains highly oxygenated (Mangum 1983; Lallier and Truchot 1989): over some range of pH, especially in the marine gastropods, a similar phenomenon, i.e., a negative or reversed Bohr effect, occurs. During hypoxia, the Bohr effect improves O<sub>2</sub> loading and transport since hypoxia induces acidosis and hemocyanin-O<sub>2</sub> affinity increases with decrease in pH (e.g., Brix 1982).

When integrated with the circulatory system, the presence of blood pigments facilitates greater uptake, transport, and delivery of  $O_2$  between the gas exchanger and the tissue cells, supporting high metabolic rates. A complex chain of respiratory adaptations was observed by Green et al. (1973) in the volcanic crater Lake Borambi Mbo in the Cameroon (West Africa), where one of the endemic species

of cichlid fish, Konia dikume, which feeds on the larvae of Chaoborus, migrates to the anoxic layer to feed. A high mean concentration of the hemoglobin of 16g per 100 ml was far above the range of the values of 5.4 to 8.7 g per 100 ml in the other ten sympatric cichlids. Fish from high latitudes generally have low concentrations of hemoglobin compared with tropical ones (e.g., Everson and Ralph 1968). Many fish possess multiple hemoglobins which in some cases (e.g., in the carp) are functionally similar (Tan et al. 1972; Weber and Lykkeboe 1978). The more energetic species, however, show functional differences in their O<sub>2</sub> binding capacities, allowing O<sub>2</sub> transport under different circumstances (Powers 1972; Brunori 1975; Weber et al. 1976). The hearts of some species of fish, e.g., the tuna, Thunnus thynnus, mackerel, S. scombrus, and carp, Cyprinus carpio, have high levels of myoglobin, respectively of about 580, 332, and 488 nmol per g tissue wet weight (Sidell et al. 1987). The maximum rates of O<sub>2</sub> consumption in active, freeswimming skipjack tuna (2.5 mg  $O_2 g^{-1} h^{-1}$ ) is more than two times the values reported for other teleosts (Stevens and Carey 1981). Rainbow trout can survive and even swim up to 70% of their maximal capacity with the coronary vasculature ligated (Farrell 1993). The microaerophilic parasitic nematode, Ascaris, has a muscle myoglobin with high O<sub>2</sub> affinity and the perienteric fluid has an abundant hemoglobin that binds O<sub>2</sub> 25000 times more tightly than its mammalian homologue (Sherman et al. 1992).

## 2.8.1. Hemoglobinless Fish

Although hemoglobin is widely distributed in vertebrates, a few animals exist without it (Ruud 1954). The well-known group is that of the Antarctic icefish of the family Chaenichthyidae, e.g., Chaenocephalus aceratus and Pseudochaenichthys georgianus (Rudd 1965; Steen and Berg 1966; Jakubowski et al. 1969) which lack it. Furthermore, the fish generally lack myoglobin, except small amounts in the ventricles of the heart (e.g., Feller and Gerdy 1987; Sidell et al. 1997). The Perciform (suborder: Notothenioidei) which includes the icefish evolved around the Antarctica within the last 25 million years (Eastman 1993), a time during which the continent became separated on the opening of the Drake Passage and formation of circumpolar currents which produced rapid cooling of the Southern Ocean. In the icefish, O<sub>2</sub> is carried essentially physically dissolved in plasma: the O<sub>2</sub> capacity of blood in the icefish (0.7 vol%) is no higher than that of seawater (Ruud 1954) and is substantially lower (onetenth) that of about 8% by volume of the fish having hemoglobin (Holeton 1970). Icefish have evolved in well-oxygenated waters whose temperature fluctuates between +0.3 and -1.87 °C (Eastman 1991). To survive in such habitats, the fish have evolved manifold adaptive features. Special blood serum glycoproteins which lower their freezing temperature below that of the near or below zero sea temperature protect them from imminent death (DeVries 1971; Ahlgren et al. 1988). In C. aceratus, a relatively high arterial PO<sub>2</sub> of 16 kPa exists (Holeton 1970): since the water has a high concentration of O<sub>2</sub>, less of it is passed over the gills in order to secure the required amount of O<sub>2</sub>. This and the absence of erythrocytes, a feature which

lowers the blood viscosity, reduces the cardiac work of pumping blood. The  $O_2$  consumption of *C. aceratus* compares with that of other Antarctic fish (Ralph and Everson 1968; Holeton 1970).

Under similar conditions and circumstances, in absolute terms, the Antarctic icefish display higher rates of O<sub>2</sub> consumption than temperate ones by about two fold (Holeton 1974; Somero 1991). Delivery of O<sub>2</sub> is effected through immense cardiac outputs (per unit body mass) facilitated by particularly large hearts (0.3% body mass), and large blood volumes even though heart rate (14 times  $\min^{-1}$ ) is particularly low (Holeton 1970; Feller and Gerdy 1987; Harrison et al. 1991; Tota et al. 1991). In C. aceratus, at 1 °C, heart rate is about 16 beats min<sup>-1</sup>, the mean ventral aortic pressure is about 2 kPa and the cardiac output is 20 to 40 ml min<sup>-1</sup> per g heart mass (Holeton 1970; Hemmingsen et al. 1972). The large cardiac output may be a compensation for the relatively small gill surface area in the species (Hughes 1972a), a favorable feature which results in low resistance to branchial water flow. In order to enhance the flow of O<sub>2</sub> from the water to the tissues, the hemoglobinless Antarctic icefish have highly aerobic muscles (Johnston et al. 1983; Johnston and Harrison 1985; Harrison et al. 1991), remarkably profuse muscle blood capillary supply (Fitch et al. 1984), and the muscle mitochondrial volume density compares with that of the flight muscles of insects (e.g., Elder 1975; Ready 1983; Londraville and Sidell 1990). The activities of the mitochondrial enzymes that are essential for aerobic fatty acid catabolism are significantly greater in the Antarctic icefish compared with other ectotherms (Sidell et al. 1987). Although the percentage utilization of  $O_2$  in water is relatively low, the efficiency of oxygenation of blood is very high in the icefish gills (Hughes 1972a) and the group survives very well in absence of hemoglobin (Andriashev 1962). The icefish, however, are not able to tolerate hypoxia as well as the redblooded fish (Holeton 1970). C. aceratus succumbs when the PO<sub>2</sub> falls to below 6.7 kPa while the sympatric red-blooded species, e.g., Notothenia neglecta, and N. gibberiforms can extract  $O_2$  down to a PO<sub>2</sub> of 2kPa.

# 2.9 Energetic Cost and Efficiency of Respiration

In biology,  $O_2$  is an essential resource which must be procured from outside at a cost. In bimodal breathers, the expense of procurement of  $O_2$  from the surface can be gauged from the duration of stay at the surface, the distance traveled, and the intervals between surfacing. The proportion of  $O_2$  acquired from aerial respiration decreases as the cost of traveling to the surface increases (e.g., Kramer 1988; Shannon and Kramer 1988). Bimodal breathers can regulate the cost of procuring  $O_2$  to match the level of metabolism and the availability of  $O_2$  from the environment (e.g., Anderson 1978). The respiratory patterns are determined by factors such as the efficiency of the water-breathing organ, the ambient temperature, and economical utilization of  $O_2$  during submergence. Insects exhibit what has been termed cyclic  $CO_2$  release (e.g., Buck 1962; Kanwisher 1966), a process which entails irregular discharge of  $CO_2$ . The frequency of spiracular opening corresponds with the energetic demands, with the bursts being more frequent during

activity. Although no ventilatory movements accompany such events, as gas transfer takes place essentially by diffusion across the spiracles (at least in the small species), some degree of energy saving must be gained when the intervals of spiracular opening are widely spaced. At an ambient temperature of below 10 °C, when  $O_2$  diffusion across the closed or fluttering spiracles is adequate to support the low rate of metabolism, in *Cecropia*, bursts stop altogether (Kanwisher 1966).

Respiratory efficiency is a measure of the performance of a gas exchanger and should express the ratio of gas procurement and transfer of O<sub>2</sub> against that of the energy expended in the process. The amount of O<sub>2</sub> consumed during respiration is required in two main areas: (1) to physically move the respiratory medium over the respiratory site(s) and (2) to power the contractions of the muscles which drive the process. Energy is needed for both ventilation and perfusion of the gas exchanger(s), complex processes of which the absolute costs remain largely unknown (e.g., Scheid 1987). In fish, the cost of breathing water (per O<sub>2</sub> uptake) has been reported to range from a mere 0.5 to 10% during routine ventilation to 70% of the overall O<sub>2</sub> consumption during exercise (e.g., Schumann and Piiper 1966; Alexander 1967; Edwards 1971; Jones and Schwarzfeld 1974; Jones and Randall 1978; Steffensen and Lomholt 1983; Milsom 1989; Rantin et al. 1992). The very high values in some species may be due to technical and experimental problems such as strenuous experimental conditions entailing physical restraint and effect of anethesia (e.g., Cameron and Cech 1970). Compared with those animals which breath air, e.g., in the healthy human being where the value is 1 to 2% (Dejours 1975), the high cost of breathing water is attributable to the greater density (about 800 times that of air), viscosity of water (50 times air) (Table 4) and the low solubility of O<sub>2</sub> in the medium: more work is needed for pumping water. Under normal circumstances, fish faced with hypoxia move from it (Dandy 1970) or reduce their O<sub>2</sub> consumption to conform with the availability (Hughes 1981). Positional changes of the gill filaments during water breathing and strict coupling of the buccal and opercular pumps, factors which improve the efficiency of the hydrodynamic flow of water, reduce ventilatory work (Ballintijn 1972). Faced with hypoxia, as an energy-saving strategy, most fish (e.g., Smith and Jones 1982; Fernandes and Rantin 1989; Rantin et al. 1992) increase gill ventilation by augmenting tidal volume and not ventilatory rate. A mechanical ventilator working at three times per minute was used to ventilate the human lung with hyperoxygenated saline (Kylstra and Schoenfisch 1972). It is probably due to the enormous cost of reversing water flow that tidal ventilation in aquatic respiration is a rare process in the evolved animals. It occurs naturally only in a few primitive organisms. In experimental liquid breathing, water has to be mechanically moved through the air passages (e.g., Kylstra et al. 1966; Sect. 6.11). The mean efficiency of the buccal force pump as it inflates the anuran lung is 8%, the O<sub>2</sub> consumption per 100g of the respiratory muscle is 0.89 mlO<sub>2</sub> per min, and the O<sub>2</sub> cost of breathing at rest is about 5% of the total resting O<sub>2</sub> consumption (West and Jones 1975). The piston pump ventilatory mechanism of the lugworm, Arenicola marina, which occurs by means of peristaltic movements of its dorsal body wall musculature (Wells 1966) takes about 40% of the total O2 uptake in animals ventilating normoxic or slightly hypoxic seawater (Toulmond 1975; Toulmond and Tchernigovtzeff 1984). At a hypoxia below 5.3kPa, the worm is unable to

pump water at an adequately high rate to overcome the reduced water O<sub>2</sub> content (Toulmond et al. 1984) and can withstand several days of experimental anoxia (Schöttler et al. 1984). During routine activity, in normoxic water, the gill convection requirement is 200 to 400 ml H<sub>2</sub>O per ml O<sub>2</sub> (Johansen 1982). The cephalopod mollusk, Nautilus, which lives at depths of 100 to 300 m and regularly encounters areas of low O<sub>2</sub> concentrations, overcomes such episodic occurrences of hypoxia by remarkable suppression of aerobic metabolism to as low as 4 to 8% of that at the normoxic level (Boutilier et al. 1996), a value comparable to that of wellknown facultative anaerobes (Hochachka and Guppy 1987). A hypometabolichypoxic animal conserves energy by decreasing the level of activity and by extending ventilatory and circulatory pauses. Water breathers have to irrigate their gills with a 40 times larger volume of water than the air breathers (with air) to extract an equivalent amount of O<sub>2</sub> (Block 1991a). Since water contains substantially less dissolved O<sub>2</sub> than air, to extract the same quantity of O<sub>2</sub>, the ventilation-perfusion ratio in aquatic breathers is ten times or more that for air breathers. In mammals, the ratio is about 1. Deviations from unity result in dramatic impairment of O<sub>2</sub> diffusion (e.g., Escourrou et al. 1993). Ventilatory requirements for fish are four to eight times higher than those of terrestrial ectotherms with similar metabolic demands (Milsom 1989). In active species, ventilatory requirements can increase 10 to 15 times above resting while O<sub>2</sub> consumption rarely increases by a factor greater than 5 (Brett 1972). In juvenile to adult fish (body mass <100 g), ventilatory flow rates range from 100 to 300 ml per kg per minute (Johansen 1982). Indirect estimation of the cost of water breathing in fish as a factor of the drop in O<sub>2</sub> consumption when the fish changes from active to passive (ram) ventilation indicates that in the trout, O<sub>2</sub> consumption falls by more than 10% and in the sharksucker about 5% (Steffensen and Lomholt 1983). During emergence into air, the crayfish, Austropotamobius pallipes, ventilates its branchial chambers at very low rates (5 ml per kg per min; Taylor and Wheatly 1980).

To minimize the energy expended on ventilatory work, fish have adopted different strategies of optimizing gas exchange to fit particular circumstances as well as to relate the respiratory process to activities such as osmoregulation. In the flounder, *Platichthys fleus*, about 7 ml per kg per h of O<sub>2</sub> and in the rainbow trout, Oncorhynchus mykiss,  $6 \text{ ml kg}^{-1} h^{-1}$  of O<sub>2</sub> are utilized for osmoregulation, values which constitute 10 to 15% of their standard metabolism (Kirschner 1993). In absolutely resting fish, ventilation can be intermittent (Perry and McDonald 1993). Benthic fish such as the bullhead catfish, Ictalurus nebulosus, exhibit apneic periods which may last for as long as 1 min, whereas others show periods of very shallow breathing alternating with strong ones (Roberts and Rowell 1988). Ventilatory pauses may be as long as 30 min in some crabs (Taylor 1984). In the intermittent ventilatory pattern of the reptiles as well as during hibernation and sleep in endotherms, i.e., where the ventilatory periods alternate with breath-holding ones, the nonventilatory period may constitute an energy-saving strategy for minimizing ventilatory cost (e.g., Glass and Wood 1983; Milsom 1991; Wood and Glass 1991; West et al. 1992) and for reducing convective respiratory water loss (Innes et al. 1986). Pulmonary blood flow increases during the breathing phase (e.g., Shelton and Burggren 1976; Burggren 1977; Burggren et al. 1977).

The nonventilatory period in the crocodile, Alligator sinensis, may last from a few seconds to 20 to 30 min (Zhao-Xian et al. 1991): the pattern of "discontinuous" or "intermittent" ventilation has been reported in the air breathing brachyurans, e.g., Cancer paguras (Burnett and Bridges 1981), Pseudothelphusa garhami garhami (Innes et al. 1986), Cardisoma (Wood and Randall (1981) and Holthuisana (Greenaway et al. 1983), terrestrial insects (Schneiderman 1960), and air-exposed bivalves (Jokumsen and Fyhn 1982). In such animals, to optimize gas transfer during the ventilatory period, heart rate, cardiac output, and pulmonary perfusion are synchronized with ventilation (e.g., Shelton and Burggren 1976; White 1978; Zhao-Xiao et al. 1991). In a quiescent, unstressed state, the crab, Cancer magister, utilizes unilateral ventilation whereby only one scaphognathite beats (McDonald et al. 1977). During such a state, variable levels of oxygenation of the postbranchial blood are attained (McMahon 1985). In the active state, the combined ventilatory volume is 288 ml per kg per min and the PO<sub>2</sub> in the hemolymph is 10.8 kPa while in a quiescent one the values are 50% less and the O<sub>2</sub> consumption is reduced by 30% (McDonald et al. 1977). The displacement of the limb girdles into the rib cage and fusion of the skeletal parts into the protective armor prohibits turtles from utilizing abdominal or thoracic movements to ventilate the lungs. In the group, respiration entails rotation of the limb girdles and hence expiration and inspiration are active processes (Gans 1976). By estimating the O<sub>2</sub> consumption during artificial ventilation and normal ventilation, at 22 °C, the energetic cost of moving air in the lung of the turtle Pseudemys *floridana* was found to be  $0.0047 \text{ ml } O_2 \text{ ml}^{-1}$  of gas ventilated (Kinney and White 1977).

In general, fish respond to hypoxia by increasing gill ventilation, reducing heart rate, and increasing the stroke volume. The cardiac output is held constant or is even elevated (Randall 1970). The two main costs of gill ventilation are energy loss due to the resistance offered by the gills to water flow and the loss of energy from cyclic acceleration and deceleration of water as it is pumped through the branchial chambers. Gill resistance to water flow occurs both during branchial and ram ventilation while loss of kinetic energy is abolished during ram ventilation. Frequently, ram ventilation occurs in the large pelagic predatory fishes, e.g., sharks, tuna, striped bass, and mackered which swim constantly (e.g., Roberts and Rowell 1988; Burggren and Bemis 1992). In very active fish, e.g., the mackered, the bucco-pharyngeal movements are not sufficient to provide the required volume of water for adequate O<sub>2</sub> to be extracted. The fish has to be continuously in motion to enhance ventilation. In the juvenile paddlefish, Polyodon spathula, as swimming speed increases, buccal ventilation becomes intermittent and continuous ram ventilation occurs above a speed of 0.6 to 0.8 body lengths  $s^{-1}$  (Burggren and Bemis 1992). In the torrential hillstreams, a fish need only physically place itself along a moving water current for the water to flow over the gills. In most fish, active buccal ventilation changes to ram ventilation at swimming speeds of 20 to  $60 \text{ cm s}^{-1}$ , with the actual transitional velocity depending on factors such as environmental PO<sub>2</sub> and temperature (e.g., Roberts and Rowell 1988). A transition speed between the two ventilatory processes is well marked in fishes such as bluefish (Pomatomus saltatrix), striped bass (Morone saxatilis), and rainbow trout (Oncorhynchus mykiss) (Freadman 1981; Steffensen and Lomholt 1983). During locomotory ram ventilation, the work of respiration is essentially transferred from the buccal and opercular muscles to the swimming muscles of the trunk (e.g., Burggren and Bemis 1992). In the bluefish and striped bass, the transition to ram ventilation is accompanied by as much as 50% increase in swimming speed without an increase in metabolic rate (Freadman 1981). Although constant swimming is energetically expensive, the energy conserved appears to justify adoption of ram ventilation. Polyodon spathula and fish such as anchovies and menhaden combine raw ventilation with filter feeding (James and Probyn 1989; Burggren and Bemis 1992), a process which results in energy saving on the cost of gill ventilation over the combined one which would be required for buccal pumping and foraging. Exercising fish increase their O<sub>2</sub> consumption five times and the ventilation of the gills about 15 times, raising the  $O_2$  consumption of respiratory muscles to 15% of the total O<sub>2</sub> consumption (Hughes and Shelton 1958). The response to hypercapnia entails an increase in gill ventilation (Jansen and Randall 1975) but hyperoxia can alleviate or even abolish the ventilatory response (Randall and Jones 1973). In contrast to air breathers, which utilize a medium with high O<sub>2</sub> concentration, fish have lower heart rates than respiratory rates, a higher ventilatory requirement being necessary in a medium of low O<sub>2</sub> content. The ventilation-perfusion ratios in fish are greater than unity. In aquatic breathers, the ventilatory response is mainly directed to O<sub>2</sub> availability rather than to elimination of CO<sub>2</sub> (Randall and Cameron 1973). The bimodal breathers perceive and react to both hypoxia and hypercapnia in water and air to similar levels of sensitivity (Johansen 1970), with the degree of perfusion and ventilation depending on the actual role of a particular structure in CO<sub>2</sub> elimination or O<sub>2</sub> uptake. Some air-breathing fish, e.g., Trichogaster, however, seem to be incapable of distinguishing O<sub>2</sub> levels in the inspired air and water (Burggren 1979). In the continuous air breathers, where the lung is ventilated by a single medium (air) and the intrapulmonary CO<sub>2</sub> and O<sub>2</sub> levels are somewhat fixed, compared with the bimodal breathers, respiratory control is a rather simple and direct process. Peripheral as well as pulmonary afferent inputs are integrated in the brain to make the necessary cardiovascular and pulmonary adjustments in response to O<sub>2</sub> needs and CO<sub>2</sub> levels in the body. Fetal respiratory movements (e.g., Cooke and Berger 1990) are well known and their significance in lung developments has been suggested (e.g., Alcorn et al. 1980; Liggins et al. 1981). Fetal breathing is said to be energetically expensive, constituting as much as 30% of the overall fetal O<sub>2</sub> consumption (Rurak and Gruber 1983). This may explain the reduction in "respiratory" frequency towards term (Berger et al. 1986), the long periods of fetal apnea in utero, and the total apnea during labor when O<sub>2</sub> is conserved for utilization by the contracting uterine muscles of the mother.

In assessing and comparing the efficiencies of biological systems, it is important to estimate the extreme performances since biological systems have different inbuilt safety margins (e.g., Alexander 1981, 1996; Currey 1967; 1984; Wainright et al. 1976; Vogel 1988). In the case of gas exchangers, at the maximum  $O_2$  consumption, all the reserve is exhausted as the functional and structural parameters are fully committed in procuring and delivering molecular  $O_2$  to the tissue cells (e.g., Gehr et al. 1981; Weibel and Taylor 1986). The cost and efficiency of respiration will depend on the rate and nature of propulsion of the external respiratory medium to the respiratory site as well as the geometrical pattern of presentation of the internal and external gas exchange media. Using different means, the gas exchangers have been variably optimized. For example, the rectification of the air flow in the bird lung (whereby the highly efficacious unidirectional and continuous flow of air in the paleopulmonic parabronchi is generated) is passively achieved – there being no evident anatomical valves (Jones et al. 1981). The geometry of the secondary bronchi (especially the mediodorsal ones) relative to the primary bronchus and a recently described constriction of the primary bronchus (just before the origin of the first mediodorsal secondary bronchus) called segmentum accelerans (Banzett et al. 1991; Wang et al. 1992) are associated with the throughflow of air in the bird lung. Inspiratory aerodynamic valving has been shown to be dependent on factors such as gas density and the convective inertial forces generated by the air in motion (Banzett et al. 1987, 1991; Butler et al. 1988; Kuethe 1988; Wang et al. 1988).

It is now well recognized that in biological systems, geometric configuration and spatial disposition of the constitutive components can in some cases constitute limiting factors in function (e.g., West et al. 1986). Due to the great diversity in design of the gas exchangers, as well as the complexity of the respiratory process itself, the factors required to estimate the cost of respiratory work are presently uncertain and imprecise. Assumptions have to be made even on some fundamental aspects. Parameters such as  $O_2$  extraction from the environment and the arterio-venous difference in  $O_2$  content are used as approximate measures of



Fig. 25. Work of perfusing the systemic and gas exchange circulations in some vertebrate animals. The low work entailed in perfusing gas exchangers compared with the systemic circuit constitutes a substantial energy saving. (Johansen 1972)



**Fig. 26.** Perfusion pressures and vascular resistance (VR) of the gas exchange and systemic circulations in some vertebrates. The *bar widths* show VR. Compared with water breathers, VR is very low in the circulatory system of the gas exchangers. This constitutes immense energy saving on respiratory work in the air breathers. *Arrows* show the scope of increased vascular resistance; *PRU* physiological resistance unit – a relative measure of resistance in the different animal groups. (Johansen 1972)

respiratory efficiency. In general, the greater specific gravity, viscosity, and lower O<sub>2</sub> content of water compared with air make the cost of aquatic respiration per unit volume of  $O_2$  extracted greater than the aerial one (Johansen 1972). To optimize the gas transfer process, the vascular resistance of the gas exchangers is particularly low compared with the systemic one, especially in the air breathers (Figs. 25,26). In water, the ventilatory rate of aquatic crabs is three times that in air and the ventilatory cost is 30% that of O<sub>2</sub> consumption (Herreid and Full 1988). In the eel, a drop in the concentration of  $O_2$  from saturation to  $4 \text{ mll}^{-1}$ results in a 40% increase in  $O_2$  uptake (=30% of the total metabolism) largely due to a five fold increase in ventilatory rate resulting from greater activity of the branchial muscles (Jones 1972). The energetic cost of aquatic ventilation in the crab, Cancer, may be as high as 76% (McMahon and Burggren 1979). The cost of breathing in the human being has been determined by increase in O<sub>2</sub> consumption due to isocapnic voluntary hyperventilation (e.g., Cournand et al. 1954) or by increased hyperventilation consequent to additional instrumental dead spaces, e.g., Milic-Emili (1991). At rest, the cost ranges from 1 to 2% of the total  $O_2$ consumption but increases in hyperbaria because of increase in gas density and during exercise due to the work needed to overcome inertia and gas turbulence. Deep sea divers need to substitute the lighter helium for nitrogen at depths in excess of 50 m in order to reduce respiratory work (Lanphier 1969). In fish, the respiratory cost, which ranges between 10 and 25%, is appreciably higher than in the human being (e.g., Hughes 1965; Dejours et al. 1970). The ratio of ventilatory requirement per unit  $O_2$  consumption in the human being is ten times lower than in a goldfish (Dejours et al. 1970). In a hypocapnic turtle, *Pseudemys floridana*, the cost of breathing ranges from 10% (at 37 °C) to 40% (at 10 °C) of total  $O_2$  consumption (Kinney and White 1977). The inverse relationship between the ventilatory cost and the body temperature (White 1978) indicates that respiration becomes more efficient with increasing metabolic capacity.

Air breathing has the salient advantage of energy saving on respiratory work and gives flexibility in the adjustment of respiratory rate without undue increase in energetic demands. The energy thus saved can be utilized for growth and development and to secure newer ecological opportunities. In air breathers, the cost of breathing is a fixed fraction of the total O<sub>2</sub> consumption and is in general about 5% or slightly less (Tenney 1979). The respiratory cost in the diving marine mammals, e.g., the cetaceans where the lungs are capable of emptying to 10 to 15% of total lung capacity (compared with only 20 to 40% in man) in as short a time as 0.3s (1.5 to 2.0s in man) have not been estimated (Olsen et al. 1969; Kooyman and Sinnett 1979; Kooyman and Cornell 1981). In fish, myocardial power output is a useful indicator of the O<sub>2</sub> cost of the cardiac pumping (Farrell 1993). Myocardial  $O_2$  consumption is about  $0.3 \text{ ml s}^{-1}$  per mW<sup>-1</sup> of cardiac pumping in various species of fish (Davie and Farrell 1991). The cost of cardiac pumping at rest constitutes 0.6 to 4.6% of resting O<sub>2</sub> consumption but in the hemoglobin-free Antarctic fish, Chaenocephalus aceratus, the cost may be as high as 23% (Farrell and Jones 1992).

### 2.9.1 The Requisites for Efficient Gas Exchange

The process of gas exchange is governed by various structural elements and functional events. These include: (1) the partial pressure gradient prevailing across the tissue barrier separating the blood and/or body fluids from the external milieu (Fig. 27), (2) the thickness of the blood-gas barrier, (3) the surface area available for gas transfer (Fig. 16), (4) the material properties (i.e., the intrinsic biophysical attributes) of the barrier tissue, properties which determine the permeability of the respiratory gases (Roughton and Forster 1957), (5) capillary transit times across the gas exchanger, (6) O<sub>2</sub>-hemoglobin binding kinetics, (7) the physical properties of the external respiratory medium, e.g., air, water or artificial atmosphere or liquid, (8) an efficient neuroregulatory mechanism for coordination of ventilation and perfusion of the gas exchanger to establish optimal gas fluxes, and (9) dynamics and geometric pattern of presentation of the internal and external respiratory media. The limitations inherent in a particular respiratory medium and the nature of transfer of  $O_2$  can be best assessed on the basis of the differences in the PO<sub>2</sub> and PCO<sub>2</sub> across a gas exchange surface. Oxygen and CO<sub>2</sub> molecules are convectively transported at equal rates whereas diffusion



Fig. 27. a Schematic diagram showing the diffusion of  $O_2$  in an alveolus across the blood-gas barrier,  $\downarrow$ , and the oxygenation profile of pulmonary capillary blood,  $\checkmark$ , in transit through the mammalian lung. The PO<sub>2</sub> in the alveolus, PAO<sub>2</sub>, decreases as O<sub>2</sub> diffuses into the capillary blood, **b**. The pulmonary capillary transit time is longer during rest than during exercise when the capillary length is almost fully utilized. In normal cases, capillary transit times are adequate for complete arterialization of blood both during exercise and rest. PIO<sub>2</sub>, partial pressure of O<sub>2</sub> in the inspired air; PaO<sub>2</sub>, partial pressure of O<sub>2</sub> in the arterial blood; PCO<sub>2</sub>, average capillary partial pressure of O<sub>2</sub>; PvO<sub>2</sub>, partial pressure of O<sub>2</sub> in the venous blood;  $\triangle PO$ , partial pressure of O<sub>2</sub>. (Weibel 1984a; reprinted by permission of Harvard University Press; copyright 1984 by the President and Fellows of Harvard College)

transport favors  $O_2$  over the larger  $CO_2$  molecule both in gaseous and aqueous media (e.g., Rahn et al. 1971). Since the ratio of the diffusion rate of  $CO_2$  to  $O_2$  is 0.78, for the same  $CO_2$  tension, the  $O_2$  gradient at a respiratory surface will always be higher (Wangensteen et al. 1971). This difference is particularly important in some organisms such as insects at various stages of development and avian, reptilian, and insect eggs, which depend wholly or partly on the diffusion of  $O_2$ and  $CO_2$  across the gas exchange surface. The extant amphibian eggs are restricted to a diameter of 9 mm (Carroll 1970) whereas in the much large eggs of reptiles and birds, where diffusion would not suffice in transfer of  $O_2$ , adaptations such as development of a well-vascularized chorioallantoic membrane which promotes uptake and transfer have evolved (e.g., Luckett 1976; Sect. 6.13). For historical interest now, at the beginning of this century, it was believed by physiologists as eminent as Christian Bohr and J.S. Haldane (see Haldane 1992), apparently from erroneous measurements which indicated a higher PO<sub>2</sub> in the pulmonary capillary arterial blood than in the alveolar air, that gas exchange in the vertebrate lung occurred by an active process. It was envisaged that the lung was able to "absorb" O<sub>2</sub> from the air spaces and "secrete" it into the blood. This was thought to occur particularly during exercise when the O<sub>2</sub> demand was high and during adaptation to hypoxia when there was deficiency of the same. The irreconcilable morphological observations of the day did not help to resolve the matter. Until the application of electron microscopy, debate ranged as to whether an epithelial lining covered the lung. Among others, Albert Policard (e.g., Policard 1929) championed the "naked alveolar capillaries" concept asserting that the respiratory surface of the lung was "like the flesh of an open wound" and that the inhaled air came into direct contact with blood. More accurate estimations of the PO<sub>2</sub> and PCO<sub>2</sub> in the alveolar and pulmonary capillary arterial blood (Krogh 1910) and better instrumentation and tissue processing when complete alveolar epithelium was discovered (Low 1953) dispelled these notions. In different animals, the alveolar PO<sub>2</sub> is consistently higher than the mean capillary blood PO<sub>2</sub> by an appreciable margin (e.g., Lindstedt 1984; Karas et al. 1987b; Constantinopol et al. 1989). In the lung of the Arctic fox, Alopex lagopus (Longworth et al. 1989), the pressure head (the pressure gradient = the driving force) for diffusion of  $O_2$  ( $\Delta PO_2$ ) across the blood-gas barrier was estimated to be about 5kPa, a value greater than that in the lungs of other highly aerobic animals, e.g., the pony (3.4kPa) and the dog (4.6kPa), the goat (1.9kPa), and the calf (2.4kPa) (Weibel and Taylor 1986; Karas et al. 1987b). In the horse,  $\Delta PO_2$  is 2.6 kPa and in the steer 2.1 kPa at VO<sub>2max</sub> (Contantinopol et al. 1989). In muscles, during severe hypoxia, a condition when the  $\Delta PO_2$  between the blood capillaries and the mitochondria is drastically reduced, a diffusion limitation for O<sub>2</sub> occurs in animals with a normal hemoglobin O<sub>2</sub> affinity (Stein and Ellsworth 1993): left-shift of the oxyhemoglobin dissociation curve provides the means for the tissue to preserve its level of O2 extraction within the blood capillary network. It has been shown that VO2max can be increased by raised O<sub>2</sub> supply to the body through increased PO<sub>2</sub> in the inspired air (e.g., Welch 1987; Knight et al. 1992; Gonzalez et al. 1993). Hemoglobin concentration affects  $VO_{2max}$ , with changes as little as 1 to  $2gdl^{-1}$  being significant (Woodson 1984). Infusion of autologous frozen erythrocytes, a process which resulted in elevation of hemoglobin concentration from 15 to 16.3 g dl<sup>-1</sup>, produced a 5% increase of  $VO_{2max}$  24h later and 4% 7 days later in the human being (Buick et al. 1984). In a similar procedure, Williams et al. (1981) reported an increased running capacity on a treadmill while Robertson et al. (1982) observed a 13% increased of  $VO_{2max}$  and 16% of endurance time in human subjects injected with 375 ml of frozen autologous erythrocytes on 2 successive days when hemoglobin concentration rose from 13.8 to 17.6 g dl<sup>-1</sup> and the hematocrit from 43 to 55%. Carrier-mediated transfer of  $O_2$  in the lung (Burns et al. 1975) and the placenta (Burns and Gurtner 1973) are known to occur, but play an insignificant role in the overall gas exchange process. Diffusion and convection are the only known processes involved in the transport of  $O_2$  into the respiratory organs and within organisms. It is only in rare, specialized organs such as the swim bladder (e.g., Fänge 1966; Sect. 6.2) and the choroid rete of the eye of teleosts

(Wittenberg and Wittenberg 1962), where  $O_2$  is known to be secreted against a concentration gradient. The respiratory roles of such organs, if any, is secondary. Since the solubility of CO<sub>2</sub> in water and tissues is much greater than that of O<sub>2</sub>, its elimination and transportation in solution presents no problems to an aquatic organism and the need for its active secretion is even less necessary. The transfer of gases from the ambient milieu to the tissue cells takes place essentially by convection (the mass transfer of  $O_2$  by the moving air in which  $O_2$  is contained) and by diffusion, the molecular flux of O<sub>2</sub> through air or liquid in a manner essentially related to the continuous random Brownian motion. High rates of O<sub>2</sub> uptake and CO<sub>2</sub> elimination are germane to the high metabolic rate of the endothermic terrestrial vertebrate endotherms. These processes are only rivaled in rate and magnitude by the transfer of water vapor between an animal and its immediate environment. The efficiency and plasticity of the gas exchange processes is underscored by the fact that during exercise or exertion, the levels of flux of respiratory gases may increase more than ten times above the resting level.

Gas exchange is constrained by temporal limitations consequent to finite rates of convection, diffusion, and reaction kinetics of CO<sub>2</sub> and O<sub>2</sub> with the carrier pigments. Inert gases, i.e., those gases which dissolve in blood without undergoing any chemical reaction, equilibrate much faster than do  $CO_2$  and  $O_2$ , which are chemically bound to the hemoglobin and other carriers. Such gases require a longer transit time at the respiratory site for full equilibration to occur. Respectively, O<sub>2</sub> and CO<sub>2</sub> require 430 and 210 ms to reach 99% equilibration while inert gases of equivalent molecular weights require only 15 to 20ms (Wagner 1977). Capillary transit times depend on factors such as tissue, species, metabolic rate, temperature, blood flow rates, pressure, viscosity, and capillary architecture, i.e., density, number, length, and geometric configuration (Karas et al. 1987b; Fig. 27). At rest, capillary transit times (in s) scale disproportionately to body mass (kg) to a power of 0.20 in the mammalian lungs (Lindstedt 1984). In mammals, capillary transit times are shorter in small animals than in the larger ones (Swenson 1990; Table 7). Oxygen binding to hemoglobin may limit maximum O<sub>2</sub> consumption in the smallest mammals (Lindstedt 1984). Birds have capillary transit times always less than 1 s. Under experimental conditions, transit times as short as 0.3 s have been reported (Henry and Fedde 1970). Adversely short transit times are prevented by capillary recruitment (e.g., Malvin 1988) and opening of arterio-venous anastomoses. Since on average the transit times are longer than the saturation times and the pulmonary capillary distance necessary for exposure and full saturation of blood is more than adequate (Karas et al. 1987b), it is conceivable that the inert and the respiratory gases are fully equilibrated as the blood traverses the pulmonary capillaries, at least under resting normoxic state. The pulmonary capillary transit time of about 0.5 to 3.0 of a second in the fish gills (Perry and McDonald 1993) is the same order of magnitude as in mammals, i.e., 0.75 of a second (Roughton 1945; Constantinopol et al. 1989) but in birds (e.g., chicken) the average transit time appears to be much shorter (e.g., Burton and Smith 1967; Henry and Fedde 1970), a value as short as 0.31 of a second having been estimated by Henry and Fedde (1970) in the chicken. This may explain the lower arterial O<sub>2</sub> tensions in birds compared with mammals (Jones and Johansen 1972). Relative to

Species	Organ	Condition	Average transit time (s)
Human	Lung	Rest Exercise Rest-hypoxia	1.8 0.4 0.3
Cow	Lung	Rest Exercise	4.6 0.9
Dog	Lung	Rest Exercise	1.8 0.3
	Muscle	Rest Exercise	0.9 0.13
_	Heart	Exercise	0.6
Fox	Lung	Exercise	0.12
Mouse	Lung	Rest Exercise	0.6 0.12
Frog	Lung Skin	Rest Rest	1.3 1.0
Fish	Gill	Rest	1.0
Bird	Lung	Rest Exercise	0.8 0.5

 Table 7. Capillary transit times in various vertebrate species and organs. (After Swenson 1990)

mammals, birds have relatively larger hearts (e.g., Hartman 1954, 1955; Tucker 1968) and faster heart rates and hence greater cardiac outputs and stroke volumes off (Lasiewski and Calder 1971; Grubb 1983). A flying budgerigar, Mellopsitacus undulatus, has a cardiac output of about 41kg<sup>-1</sup>min<sup>-1</sup>, a value which is seven times greater than that of a mammal of equivalent body mass (Tucker 1968) and a heart rate as fast as 1020 beats min<sup>-1</sup> has been reported in the giant hummingbird, Patagona gigas (Lasiewski et al. 1967). The cardiac blood pressure in birds is very high. Systolic pressures as high as 53kPa have been determined in the domestic turkey, Melleagris gallopavo (Speckman and Ringer 1963). In contrast, among the living vertebrate ectotherms (i.e., fish, amphibians, and reptiles), systolic blood pressure rarely exceeds 7kPa (Lindsay et al. 1971). Poikilothermic vertebrates store lipids primarily as tricylglycerols (Sheridan 1994). In birds, occasional deaths commonly attributed to fright are largely due to hemorrhage consequent to arterial rupture resulting from sudden excessive hypertension arising from such encounters (Walkinshaw 1945; Carlson 1960; Hamlin and Kondrich 1969). In Nautilus, during the hypometabolic state, i.e., after the PO<sub>2</sub> in the arterial blood has fallen to below 0.7kPa, a reversal of the normal arterialvenous blood PO<sub>2</sub> gradient occurs with the venous PO<sub>2</sub> exceeding that of the arterial blood (Boutilier et al. 1996). This is thought to be brought about by an extremely slow intermittent blood flow during severe hypoxia when the large vena cava which is located on the roof of the mantle cavity has time to equilibrate with the ambient PO<sub>2</sub>. Most interestingly, during such episodes, the vena cava serves as a gas exchanger, transferring respiratory gases between the blood and the air in the mantle cavity!

Morphometric studies have shown that the erythrocytes offer the greatest resistance to O<sub>2</sub> diffusion in the gas exchangers (e.g., Hallam et al. 1989; Maina et al. 1989a). Due to lack of carbonic anhydrase in the capillary endothelial cells of the gills (e.g., Perry and Laurent 1990; Lessard et al. 1995), unlike in the mammalian lung, in fish, the erythrocytes appear to be the only site for HCO<sub>3</sub><sup>-</sup> ion dehydration (Perry and McDonald 1993). The relatively long transit time of the blood in the gills, the high capacitance of water for CO<sub>2</sub>, and the efficacious countercurrent system of the gills may account for the effectiveness of the gills in CO<sub>2</sub> elimination. The factors which may inhibit end capillary PO<sub>2</sub> equilibration include: (1) an overly high cardiac output, e.g., during exercise and disease conditions factors which cause loss of the functional capillary bed, resulting in shorter transit times (Wagner et al. 1986), (2) thicker tissue diffusional barriers, e.g., in pathological conditions such as edema (Staub et al. 1967; Staub 1974), (3) severe hypoxia (reduction in O<sub>2</sub> driving gradient), e.g., during high altitude exercise (Wagner et al. 1986), (4) large erythrocytes which are associated with greater unstirred layers, and (5) pathological conditions which are accompanied by reduction of the concentration of the hemoglobin or those characterized by reduced erythrocytes count, a factor which increases the thickness of the unstirred layer resulting in reduction of O<sub>2</sub> binding velocity (e.g., Nguyen-Phu et al. 1986). The erythrocytes of the bullfrog, which have a volume of  $700 \,\mu\text{m}^3$ , take up O<sub>2</sub> at a rate five times slower than the smaller ones of 20 µm<sup>3</sup> of the goat (Holland and Forster 1966). Diffusional resistance during severe exercise may inhibit end capillary  $O_2$ equilibration as the capillary transit times fall to as low as 200 ms (Groebe and Thews 1987). Owing to the shape of the haemoglobin-O<sub>2</sub> dissociation curve which causes large  $O_2$  loading during the early part of intracapillary transit (see. e.g., Comroe 1974; Shiga 1994), even when the end capillary complete equilibration is not attained, the amount of O<sub>2</sub> taken up is adequate for the respiratory needs. The differences in the equilibration time courses in the inert gases and in the biochemically active gases such as CO<sub>2</sub> and O<sub>2</sub> can be attributed to the ratio of the solubility of the gas in the membrane barrier to that in blood. In inert gases, the ratio is unity while the value is several orders of magnitude less for CO<sub>2</sub> and O<sub>2</sub> (Piiper and Scheid 1980). The greater solubility of O<sub>2</sub> and CO<sub>3</sub> in blood relative to the membrane is due to the reversible chemical binding and reaction with hemoglobin and other proteins in the erythrocytes and plasma (Shiga 1994). The rate at which the partial pressure of a particular gas builds up is exclusively dependent on its solubility in blood. The more soluble a gas, the slower will be its rate of partial pressure change for a given quantity of gas transferred. The rate of equilibration of CO<sub>2</sub> in any solution is strongly dependent on the buffering capacity. The rate-limiting steps in O<sub>2</sub> transfer are ascribed both to diffusional resistances and chemical reactions in an inhomogeneous medium. Unlike in CO<sub>2</sub> exchange and in the Bohr-Haldane effects, these fast processes do not require enzymes or membrane transport carriers. Even during exercise, in mammals, O<sub>2</sub> transfer and equilibration with the capillary blood are usually complete by the end of the transit time (e.g., Karas et al. 1987b; Longworth et al. 1989; Fig. 27).

Ventilatory and perfusive processes are particularly important in gas transfer during episodes of extreme exertion, hypoxia, and respiratory acidosis. In the course of its passage from the external environment to the mitochondria, O<sub>2</sub> travels across several anatomical spaces which offer different degrees of resistance (Figs. 2,3). According to Fick's Law, the diffusing capacity of a gas through a barrier depends on surface area, the concentration gradient, the thickness of a barrier (the diffusional distance), and the diffusion (permeation) coefficient across the barrier (Figs. 16,28,29). Among air breathers, the thickness of the blood-gas barrier of the gas exchangers decreases from the amphibians, reptiles, mammals, to birds (e.g., Maina 1994). The water-blood barrier in the aquatic gas exchangers is in general thicker than the blood-gas barrier of the lung (e.g., Hughes and Morgan 1973). The diffusion rate in the mammalian lung is very efficient. Determinations by Wagner and West (1972) and Hill et al. (1973) showed that in the human alveolar-capillary barrier whose normal thickness is  $0.62\,\mu m$  (Gehr et al. 1978), the thickness would have to be increased four to ten times before it becomes a limiting factor for end capillary PO<sub>2</sub> equilibration. From theoretical considerations (e.g., Scheid 1978) and experimental work by Powell and Wagner (1982a), in the avian lung the diffusion of  $O_2$  in the gas phase of the air capillaries which radiate from the parabronchial lumen (Figs. 30,31) is not thought to constitute a limiting factor at rest (Powell 1982; Powell and Scheid 1989). Owing to the long diffusional distance (20 to 50 µm), cutaneous respiration is accompanied by limitations of diffusion (e.g., Malvin 1988) but movements of air over the skin may enhance the diffusional process. Increased perfusion may be utilized to enhance O<sub>2</sub> uptake particularly during hypoxic episodes (Malvin and Hlastala 1986). Movements by skin-breathing aquatic organisms may reduce



Fig. 28. Schematic view of the oxygen diffusional pathway in an aquatic breather. The diffusing capacity of the gas exchanger depends on the thickness and surface area of the respiratory barrier. In the water breathers, the unstirred layer of water over the gills may constitute a significant part of the resistance to  $O_2$  diffusion and may need to be considered in modeling the diffusion capacity of the gills



**Fig. 29. a** Blood capillary in the lung of the domestic fowl, *Gallus domesticus*, showing a file of erythrocytes in a blood capillary. **\***, blood-gas barrier. **b** The extremely thin blood-gas barrier, **\***, shows sporadic attenuations. *e* erythrocytes. **a** *Bar* 0.6μm; **b** 0.6μm. (Maina 1993)

diffusional limitations in the layer of water next to the skin (Feder and Burggren 1985a,b). In fish, factors such as low body temperature, thick water-blood barrier, and presence of an unstirred water layer over the secondary lamellae reduce  $O_2$  transfer. They may cause significant diffusion limitations and curtail end-capillary equilibration particularly during resting normoxic conditions (Randall and Daxboeck 1984). At a simulated altitude of about 12 km, the resting barheaded goose, *Anser indicus*, tolerated inspired PO<sub>2</sub> of about 3 kPa and no difference was observed between the PO<sub>2</sub> in inspired air and that in the arterial blood (Black and Tenney 1980): the arterial PCO<sub>2</sub> was as low as 1.1 kPa. Due to the large arteriovenous shunts which cause notable arterial  $O_2$  desaturation in the amphibian and reptilian lungs (Glass and Wood 1983; Wood and Glass 1991), diffusional limitations are masked.


Fig. 30. a Lung of the domestic fowl, *Gallus domesticus* showing parabronchi, p, surrounded by gas exchange tissue, e, and blood vessels,  $\succ$ , located in the interparabronchial septa. b Closeup of a parabronchial lumen showing atria, a, and infundibulae,  $\succ$ , which channel air to the exchange tissue (e). a *Bar* 70 µm; b 20 µm



**Fig. 31.** Latex cast of the airways of a bird lung, *Gallus domesticus*. This closeup shows an atrium arising from a parabronchus, p, giving rise to an infundibulum, i, terminating in an air capillary, a. The perpendicular to the arrows,  $\rightarrow$ , converging onto the star, \*, illustrate the distance  $O_2$  has to diffuse from the parabronchus to the air capillary.  $\succ$ , spaces occupied by blood capillaries. *Bar* 40 µm

# 2.9.2 Efficient vs. Inefficient – Primitive vs. Advanced Gas Exchangers: the Contention

From cladistic principles, the morphological, physiological, and biological character states of organisms are considered either to be primitive or derived. Commonly, these interpretations are subjective. For example, some organisms (e.g., bacteria) which are notably speciose and highly adapted to particular habitats where they thrive and even outcompete supposedly better-adapted forms are deemed primitive. Strictly, in a particular animal lineage, the primeval (plesiomorphic) features should be taken to be those which characterize the ancestral forms. The derived (apomorphic) features typify the more recent animal groups. A character which is unique to an animal (an autamorphic mark) indicates a unique adaptive state. The primitive features are assumed to be inefficient while the advanced (= derived) ones are taken to be adaptive (= more efficient). The fact that the so-called primitive features have been saved for millennia amidst intense selective pressures emphasizes either their neutral selective value or the exceptional advantage(s) they confer for survival in a particular habitat. Gould (1994) opines that "our impression that life evolves towards greater complexity is probably only a bias inspired by parochial focus on ourselves and consequent over attention to complexifying creatures while we ignore just as many lineages adapting equally well by becoming simpler in form". Along the same argument, Thompson (1959, p. 41) points out that "however important and advantageous the subdivision of the tissues into cells may be from the constructional, or from the dynamic, point of view, the phenomenon has less fundamental importance than was once, and is often still, assigned to it".

Primitive and derived features exquisitely allow different kinds of animals to subsist in the same habitat. The different resource procuring capacities enable the congeners to fill species-specific spatial and/or temporal ecological niches without undue competition. Special environments are characterized by low biodiversity though the numerical densities of the occupying species may be very high. Pough et al. (1989) assert that "the primitive character state is not inferior or necessarily less adaptive, it is simply older". These sentiments are echoed by Kardong (1995), who observes that "the notion of better does not apply to biological changes: primitive and advanced species represent different ways of surviving not better ways of surviving". From these sentiments, the terms primitive and derived should be used to define only the temporal appearance, i.e., the chronological advent of certain attributes in the phylogentic history of a particular animal lineage and not the polarity of a change. The primitive features are those that occur in the ancestral members of the group while the derived ones are found in the modern forms and differ in notable morphological and physiological ways from the primordial condition. Gas exchangers are designed to service given cytoplasmic masses operating within certain metabolic boundaries. Overdesign is costly and wasteful. In the context of functional capacities, strictly, primitive gas exchangers do not exist.

The simplest design and apparently the most efficient evolved gas exchanger has occurred in form of the insectan tracheal system (Sect. 6.6.1; Figs. 74,75). Few animals have successfully emulated it. Gas exchangers present certain properties which are fundamental in explaining the remarkable diversity of design. These are: (1) the transfer of gases from the external milieu to the body and vice versa occurs by the physical process of diffusion, (2) there is no ubiquitous structural plan: one or more pathways may be utilized simultaneously or in phase, (3) there is only a weak correlation between the constructional complexity, efficiency, and the phylogenetic level of development, and (4) the diffusion of gases across the tissue barriers occurs in solution. To enhance the diffusing capacity, efficient gas exchangers must have an extensive respiratory surface area, a thin blood-gas barrier, and must be well perfused with blood and ventilated with the ambient respiratory medium. Organs as diverse as the anus, stomach, mouth, and skin are used as gas exchangers. The advanced animals, e.g., mammals, do not necessarily have the most efficient gas exchangers. Invagination of the gas exchangers not only afforded a greater respiratory surface area but also provided better control and regulation of the physical characteristics of the inhaled air which must be cleansed and particularly moistened to curtail water loss from the alveolar surface.

# 2.10 Modeling: Utility in Study of Integrative Construction of the Gas Exchangers

Gas exchangers are complex, dynamic, high fidelity systems. Respiration entails intricate interplay between physiological features such as convection, diffusion, and chemical reactions (e.g., Piper et al. 1971; Wagner 1977) and anatomical factors such as the architecture and geometry of the airways and the respiratory barriers. Morphological observations and quantification of the components of gas exchangers makes it possible to construct realistic models and also advance understanding of the evolutionary processes that have precipitated the diversity in form and function in the process of adaptation to environments and adoption of different lifestyles. Through selective pressure acting on the genotype, the gas exchangers have been honed to meet species-specific, design-oriented needs. The final constructional plans present rational, multifaceted, integrated engineering (Beament 1960; Dullemeijer 1974; Gutman 1977). An intractable number of variables (of which the individual functional capacities may not be sufficiently known) contribute to the ultimate fabrications of biological systems. Biology has long been mathematically emancipated and somewhat changed from how it was once perhaps legitimately described, as a descriptive (inexact) science (compared with physical science). In the gas exchangers, the pulmonary diffusing capacity, defined as the flow rate of O<sub>2</sub> from air to blood under partial pressure gradient between alveolar and mean O<sub>2</sub> tension in the alveolar capillary blood (Bohr 1909; Roughton and Forster 1957; Forster 1964; Weibel 1970/71) is the outcome of the integration of the functional and structural parameters. The functional anatomies of composite structures (e.g., the gas exchangers) can be adequately described through appropriate modeling, i.e., by integrating the relevant parameters central to an organ system, e.g., the respiratory (Singh et al. 1980; Federspiel and Popel 1986; Bozinovic 1993) and the cardiovascular systems (Zagzoule and Marc-Vergnes 1986; Wang et al. 1989; Melchior et al. 1992). Gutman and Bonik (1981) defines a model as "an abstraction of a real situation which describes only the essential aspects of the situation". Implicit if not explicit assumptions underlying various parameters and properties are often made. A mathematical model aims to "dismember" a structure into the formidable number of individual components, isolates those components that are relevant in answering the question at hand, and then reassembles it. The model must endeavor to be accurately descriptive of the complex reality of nature, must be simple and easy to conceptualize, and must be theoretically and practically testable. By applying appropriate physical and physiological constants, a static structural model can mathematically be converted to an acceptably dynamic one. The net worth of a model can be judged from its predictive potential. In modeling highly dynamic systems, e.g., the wing beat kinematics of an insect in flight (Weis-Fogh 1973; Ellington 1981, 1984; Wilkin and Williams 1993) and atmospheric turbulence (e.g., Hollinger et al. 1994), a defined state (e.g., a steady- or a quasisteady state) has to be assumed. Complex systems are intrinsically chaotic and are hence unpredictable (Gleick 1987). Experimental manipulation of a functional model gives an insight into the cause-and-effect relations between the individual and the covariant factors that drive a system. Outcomes can be evaluated by altering one or more factors or

conditions while holding others constant. Constraining, potentiating, and superfluous components can be identified and the relative performances determined. Based on structural-functional integrative studies, encompassing observations can be made on the gas exchangers. Depending perhaps on the approach adopted and the emphasis laid on the different parameters which are considered, in some cases the conclusions have turned out different. Assessing the factors which determine O<sub>2</sub> flow from the lung to the muscle tissue during exercise, Weibel et al. (1987a) and Karas et al. (1987a) concluded that "cardiac output is the most important variable controlling flow of O<sub>2</sub> through all steps of the respiratory system". However, evaluating the factors which limit the delivery of O, to the skeletal muscles in an exercising human being, Saltin (1985) resolved that "pulmonary diffusing capacity is the ultimate limiting factor" during extreme conditions. In a theoretical model, Wagner (1993) concluded that at sea level, cardiac output has the greatest effect on VO<sub>2max</sub> while at altitude, muscle diffusing capacity is the most important parameter. Explaining the 2.5-fold higher maximum O<sub>2</sub> consumption in the athletic species (pony and dog) over that of the less athletic ones (calf and goat), Karas et al. (1987b) attributed one half of it to the higher PO, at the blood-gas barrier (1.7-fold greater in the athletic species) and the other half to the diffusing capacity which was 1.5-fold larger. It is most instructive to be able to accurately predict, infer the consequences of, or explain how one or a group of respiratory variables affects the others. Physiologic and morphometric estimations of diffusing capacities of different gas exchangers, e.g., fish gill (Scheid and Piiper 1976; Piiper et al. 1977, 1986; Hughes et al. 1986b), mammalian lung (Gehr et al. 1978; Weibel et al. 1983), and bird lung (Maina et al. 1989a; Vidyadaran et al. 1988; Burger et al. 1979) are now available. In all cases, the morphometric value exceeds the physiological one. This has been interpreted to indicate a safety margin of operation. In estimating the morphometric diffusing capacity, an ideal state is assumed, i.e., that O<sub>2</sub> flux occurs across the entire water-air /blood barrier under a ventilation-perfusion ratio of unity. This state is only approached at maximum O<sub>2</sub> consumption (e.g., Powell and Wagner 1982a,b). Gans (1985) holds that "animals cannot afford one-function/one-structure designs "and" each activity tends to involve multiple aspects of the phenotype and each aspect of the phenoytpe may be involved in multiple activities" (Gans 1988). Empirical prototypes in form of biological models provide useful insights (even though in theoretical way) into the underlying control mechanisms, the multidimensional utilization of the phylogenetic characteristics, and the effect altered parameters have on the entire system (Cameron 1989; Anker and Dullemeijer 1996). Simplification, abstraction, and generalization of biological processes and concepts have contributed greatly to the advancement of scientific thought (Homberger 1988; Vogel 1988).

Gas exchangers, like most organs, present a multilevel organizational character. The various parts of a structure are arranged in discrete functional units which are, in turn, intricately connected to ensure integrity in the overall performance. For optimal transfer of gases at the exchange site, the convective (ventilatory and perfusive) pumps must be coherently linked so that the gas delivered across the respiratory barrier is immediately removed to maintain a concentration gradient. Overperfusion relative to ventilation or vice versa incurs unnecessary metabolic cost. The actual quantity of respiratory gases transferred across the barrier in such episodes is lower than that prevailing during the optimal and less expensive conditions. The ideal ventilation-perfusion ratio in mammals is about 1:1 (West 1977a) and in the chelonian reptiles it is 2:1-5:1 (Burggren et al. 1977). In aquatic fish, the equivalent value is 9:1-35:1 (Randall 1970). By refining and multiplicatively integrating the structural and physiological parameters such as lung volume, cardiac output, hemoglobin concentration, muscle mass, and mitochondrial number, land vertebrates such as the pronghorn antelope, Antilocapra americana (Lindstedt et al. 1991), and the cheetah, Acionyx jubatus (Hildebrand 1959, 1961), can attain and sustain speeds of 75 to 100km per h. Lindstedt et al. (1991) estimated that the pronghorn antelope can take up  $O_2$  at a late of 3.2 to 5.1 ml  $O_2$  kg<sup>-1</sup>s<sup>-1</sup>, a value 3.3 times greater than that predicted for a typical mammal  $(1.5 \text{ ml O}_2 \text{ kg}^{-1} \text{ s}^{-1})$  of a similar body mass of 32 kg. On the other hand, the naked mole rat, Heterocephalus glaber, a highly inbred (Reeve et al. 1990), eusocial (Burda 1993), heterothermic, low metabolism animal (McNab 1966) which lives in a stable environment (Cossins 1991) has neotenic lungs (Maina et al. 1992). The pulmonary morphometric diffusing capacity of the lung of the mole rat, Spalax ehrenbergi was found to be 40% greater than that of an equivalently sized surface-dwelling white rat (Widmer et al. 1997).

## 2.10.1 Evaluation of the Functional Efficiency of the Gas Exchangers

Analysis of the performance of a gas exchanger entails physiological measurements of O<sub>2</sub> tensions prevailing in the gas exchange media or that between the arterial and venous blood. This is done by taking cognizance of the inherent construction of the exchanger and the fluid flow dynamics within and without the gas exchanger. Respiratory efficiency can also be gauged from the quantity of O<sub>2</sub> transferred against the energy expended during the process (e.g., Piiper and Scheid 1992; Shelton 1992). By varying or holding certain factors constant while integrating or suppressing others, the contributions of the various variables to the overall function can be evaluated and the limiting or potentiating factors identified. The efficiencies of different or same gas exchanger(s) under different operating conditions can be compared. Though gross mathematical abstractions of a complex biological system which is regulated by an infinite number of variables, all of which are difficult to individually assess, manipulatable theoretical analogs are highly instructive in comparative experimental and analytical studies: they generate "artificial" didactic situations which afford a perception of a complex outlay. Physiological phenomena are cumulative effects of interactions between numerous minuscule, individually noneventful, nonlinear, but coupled local unit processes which interact at various levels with a potentiating or even a depreciative effect. Though with intrinsic limitations, functional and morphological models are extremely valuable in the understanding of the complex relations in multidimensional systems (e.g., Riggs 1963; Penry and Jumars 1987; Usry et al. 1991; Horn and Messer 1992). From the results of functional and structural analyses, a piecemeal synthesis of an organ system can be made. The efficiency of



**Fig. 32a-c.** Schematic drawing showing the spatial relationship between air and blood flow,  $\rightarrow$ , in the fish gills (a) and bird (b) and mammalian (c) lungs. In fish, air and blood flowing in a countercurrent manner. In birds, the flow between the parabronchial blood (*small arrows*) and the air flow in the parabronchial lumen is crosscurrent. In the mammalian lung, the flow is described as uniform pool. *e* Parabronchial gas exchange tissue and air flow in the lumen,  $\uparrow$ ;  $\nu$  venous blood; *a* (in b) arterial blood; c *a* alveolar air;  $\downarrow$ , diffusion of  $O_2$  across the blood-gas barrier:  $\rightarrow$ , direction of alveolar capillary blood flow;  $\leftrightarrows$ , tidal ventilation of the mammalian lung. To a large extent, the efficiency of the gas exchangers depends on the presentations between the gas exchange media

a gas exchangers is largely determined by the nature of presentation of the respiratory media (Fig. 32) which is dependent on the intrinsic refinements and the geometric arrangements of the structural components (Figs. 29,33,34). Capillary geometry and tortuosity has a profound effect on the efficiency of  $O_2$  transfer in muscles (e.g., Ellis et al. 1983; Groom et al. 1984a,b; Mathieu-Costello 1990; Mathieu-Costello et al. 1992). Biologists can appropriate the mathematical and conceptual contrivances of engineers to discern how animals function and why they have acquired certain morphologies.

#### 2.10.2 Modeling the Gas Exchangers

In its most simplistic form, the respiratory system can be conceptualized as a set of compartments interposed between the body tissue cells and the environment. Across the cascade, which comprises the gas exchanger (ventilatory convection), water/air-blood barrier (diffusion), blood (circulatory convection), systemic capillary bend, interstitial fluid, and cells (diffusion),  $O_2$  and  $CO_2$  flux at intensities essentially determined by the metabolic states and environmental circumstances. It is mainly in the chordates where adequate structural and functional data have



Fig. 33. Schematic view of a pulmonary blood capillary showing the in-series components of the air-hemoglobin pathway, namely the blood-gas (tissue) barrier (which comprises a surface lining, an epithelial cell, a basement membrane, and an endothelial cell), plasma layer, and the erythrocytes.  $O_2$  uptake by the erythrocytes constitutes the greatest point of resistance to flow. (Maina 1994)

been gathered to warrant satisfactory modeling of the gas exchangers. The general model relating diffusion of gases across the blood-gas barriers and the water-blood barriers to ventilation and perfusion (e.g., Piiper and Scheid 1975) has been extensively used in different gas exchangers (e.g., White and Bickler 1987). Morphometric pulmonary modeling after Weibel (1970/71) has been carried out in many gas exchangers, e.g., fish gills (Hughes 1972b), reptilian (e.g., Perry 1983), avian (e.g., Maina 1989a), and mammalian lungs (e.g., Gehr et al. 1981).

When the gas exchange media, i.e., the external (the ventilatory) and the internal (the perfusive) media run parallel to each other and in the same direction, the design is termed concurrent. If the media run in opposite directions (e.g., water and blood in the fish gills), it is called countercurrent and when the media run at right angles to each other, e.g., the blood capillaries of the parabronchial exchange tissue relative to the flow of air in the parabronchial lumen of the bird lung), the arrangement is designated crosscurrent. When the external medium is held constant against a gas exchanger (e.g., skin) or is ventilated with a medium of which the gaseous partial pressures are fairly uniform (e.g., mammalian lung where the alveolar gas exerts a steady-state concentration)



**Fig. 34.** Model for estimating pulmonary diffusing capacity or conductance of a lung for  $O_2$  (DLo<sub>2</sub>). The principal barriers which influence  $Do_2$  are the tissue barrier [epithelium + basal lamina (interstitium in some cases) + endothelium], the plasma layer, and the erythrocytes. DLo<sub>2</sub> is directly proportional to the surface area and the permeation constant and inversely proportional to thickness. Respectively:  $Dto_2$ ,  $Dpo_2$  and  $Deo_2$  diffusing capacities of the tissue barrier, plasma layer, and erythrocytes; St and Sp surface areas of the tissue barrier and plasma layer; Vc volume of the pulmonary capillary blood;  $Kto_2$  and  $Kpo_2$  Krogh's oxygen permeation coefficients of the tissue barrier and plasma layer;  $o_2 - O_2$  uptake coefficient by the whole blood. The products of St and  $Kto_2$  and  $Kpo_2$  must be divided (not shown on the figure) by the harmonic mean thickness (tht) of the respective barriers. (Maina 1993 courtesy Pergamon Press). For a detailed account on the model as applied to the bird lung, see Maina et al. (1989a)

(Milhorn et al. 1965), respectively, the configurations are termed infinite pool and uniform pool (White 1978; Figs. 30,32,35,36). In the small insects,  $O_2$  is conducted directly to the tissues by diffusion through trachea (Maina 1989b; Figs. 36,37) and by convection through the air sacs in the larger ones (Fig. 38). Delivery of  $O_2$ directly to the tissue cells is a more efficient means of gas transfer (Sect. 6.6.1) compared with the conventional one where the molecule is convectively transported through a circulatory system (Levi 1967; Bromhall 1987).

By fitting or approximating a gas exchanger to any of the above main models, the efficiency and the limitations (be they developmental, structural, or functional) inherent in a particular gas exchanger under different circumstances can be gauged (Hills 1972; Piiper and Scheid 1972, 1975; Sikand et al. 1976). The relative efficiencies between different gas exchangers can be assessed. In a concurrent design, the gas tension in the internal medium leaving the exchanger cannot possibly exceed that of the external medium in normal and about equal fluid flows. However, under special conditions, e.g., when large quantities of water flow very fast over a small volume of slowly moving film of blood, a high degree of oxygenation with low utilization can be realized (Hughes 1963) but after



Fig. 35. Countercurrent system. The exchange media which contain  $O_2$  at different concentrations flow in opposite directions. A diffusional gradient prevails over the duration and length of contact. A countercurrent arrangement occurs in the fish gills where  $O_2$  extraction has been reported to be as high as 96%. Numbers Percentage concentrations of  $O_2$  in the respiratory media;  $\prec O_2$  flows down a concentration gradient; T tissue barrier; W water; E erythrocytes; P plasma; B blood. (Maina 1994)

great energy expenditure on ventilatory work. In such cases, the gain in real terms is very small. In a countercurrent system, since the efferent blood has been exposed to the inhalant water, the PO<sub>2</sub> between the two media is high. Unlike in the mammalian lung, where theoretically ventilation-perfusion homogeneity may allow alveolar  $PO_2$  to equilibrate with the arterial  $PO_2$ , in the fish gills arterial blood and inspired water never reach equilibrium (Randall 1970). This may be attributed to factors such as the thick water-blood barrier (e.g., Hughes and Morgan 1973), the large ventilation-perfusion inequalities (Booth 1978), and added diffusion limitations resulting from the boundary layer of water close to the surface of the secondary lamellae (Hills 1972). The ventilated uniform-pool design in most vertebrate lungs is more efficient than the nonventilated one (e.g., the skin and the buccal cavity). However, behavioral changes such as exposure to air currents and movement may initiate passive ventilation enhancing gas exchange. In the human lung, due to the tidal nature of air renewal, only about 12% of the intrapulmonary air volume is exchanged at the alveolar level per inspiratory cycle. Due to the low efficiency of the concurrent system, the design has only rarely been invoked in evolutionary biology. In contrast, the countercurrent system is widely encountered in biology (Scholander 1958) especially in form of heat exchangers, e.g., cephalic and ophthalmic rete (Jackson and Schmidt-Nielsen 1964; Kilgore et al. 1979; Baker 1982; Midtgärd 1983, 1984; Kamau et al. 1984;



Fig. 36A-F. Schematic illustrations showing the geometric interactions between the respiratory media in different gas exchangers. The respiratory efficiencies are shown by the PO<sub>2</sub> and PCO<sub>2</sub> profiles in the inspired air, *I*, expired air, *E*, venous blood, *V*, and arterial blood, *A*. A Cocurrent system: gas exchange media flow in the same direction and O<sub>2</sub> flux is very low; **B** The skin: the gas exchanger is perfused but not ventilated. **C** Countercurrent arrangement: the PO<sub>2</sub> in the arterial blood exceeds that in the end expired air. **D** Uniform-pool: the influx of O<sub>2</sub> depends on ventilation and perfusion inequalities. **E** Crosscurrent system in the bird lung: through the multicapillary serial arterializational arrangement, the PO<sub>2</sub> in the arterial blood may exceed that in the expired airs. **D** using the direction of the tissue cells (by the tracheoles, *T*) with the PO<sub>2</sub> at the cell level being only slightly lower than that in the ambient. The PO<sub>2</sub> in the arterial blood may exceed that in the expired respiratory medium only in models C, **E**, and F. *m* Respiratory medium which depending on the type of animal could be air or water: *p* parabronchus; *c* cell; *v* blood (venous); *w* water. In the schematic drawings, the *single arrows* show the directions of flow of respiratory media and the diffusion of O<sub>2</sub>; = *arrows* show the gas exchangers which are tidally ventilated. (Maina 1994)

Pinshaw et al. 1985; Block 1987), salt-concentrating systems such as kidneys and salt glands of birds (Kokko and Tisher 1976; Schmidt-Nielsen 1990), and in gas secretion organs, e.g., the choroid rete (Wittenberg and Wittenberg 1962) and the rete mirabile of the swim (air) bladder of fish (Fig. 39; e.g., Kuhn et al. 1963; Kobayashi et al. 1989a,b; Pelster and Scheid 1992a). Chemical (e.g., Coulson and Richardson 1965) and mechanical engineers (e.g., Carslaw and Jaeger 1959) exploit countercurrent systems to enhance concentration gradients in order to improve mass transfer. The functional efficiency of the countercurrent system depends on factors such as a thin and extensive surface area for interaction of the gas exchange media, optimum flow rates and physical states and characteristics.



Fig. 37. a Tracheal system of a grasshopper, *Chrotogonus senegalensis*. The primary trachea which start near the spiracles,  $\succ$ , are connected through anastomotic chains, *t. m*, Malpighian tubules. b Closeup of a primary trachea, *t*, showing the spiral taenidia which maintain patency.  $\triangleright$ , secondary trachea. (Maina 1989b.) a *Bar* 700 µm. b 80 µm



Fig. 38a,b. Air sacs of the grasshopper, *Chrotogonus senegalensis*. The air sacs occur in pairs (a), singly (b) or in clusters. They are characteristic of the large and the energetic insects which use abdominal pumping to enhance movement of air along the tracheal system. >, efferent trachea;  $\nu$ , possible location of a valvular apparatus which maintains a unidirectional flow of air in the tracheal system; *m*, Malpighian tubules. Like the trachea, the air sacs are made up of helical taenia, indicating that they are simply dilatations of the trachea. (Maina 1989b) a *Bar* 700 µm; b 220 µm



Fig. 39. The gas gland cell, g (top inset) and the rete mirabile of the swim bladder of a tilapiine fish, Oreochromis alcalicus grahami (main figure and bottom inset). The bottom inset is a latex cast SEM preparation of the arterial vasculature at the rete. The venules (v in the main figure and arrow in the bottom inset) and arterioles, a (main figure) run parallel to each other. Secretion of the acidic metabolites produced by the gas gland cells into the arterial blood and subsequent transfer into the venous blood causes release of blood gases into the bladder.  $\leftarrow$  (main figure) shows an extremely thin area of separation between a venule and an arteriole, sites which may promote the transfer of the acidic metabolites released by the gas gland cells. r Red blood cell. Bars top inset,  $0.25 \mu$ m; main figure  $0.8 \mu$ m; bottom inset,  $16 \mu$ m. (Maina et al. 1996a)

In biology, however, countercurrent heat exchangers cannot simultaneously serve as efficient gas exchangers. To enhance heat transfer by bulk flow, the blood vessels of the heat exchanger have a diameter in excess of 1 mm. Gas exchange by diffusion can only efficiently occur at the capillary level where the walls are adequately thin. In fact, if O<sub>2</sub> were to diffuse equally well in the heat exchangers, very little, if any, O<sub>2</sub> would reach the peripheral tissue. Such arrangements (countercurrent heat exchangers) would have to be very sparingly used (if at all) in biology. The low O<sub>2</sub> tension found in the kidney tissue (e.g., Landes et al. (1964), an organ supplied with a large volume of blood, may be due to diffusion of O<sub>2</sub> between the blood capillaries supplying the renal papilla and those leaving it, i.e., the descending and ascending versa recta. In birds, interestingly, while the ophthalmic rete is considered a countercurrent exchanger (e.g., Kilgore et al. 1979; Midgard 1983), it has been shown to supplement O<sub>2</sub> supply to the brain during high altitude flight (Bernstein 1989, 1990; Bernstein et al. 1984): the arterial blood passing through the rete en route to the brain exchanges O<sub>2</sub> with the venous blood, which has a higher PO<sub>2</sub> after draining the highly vascular eye, nasal cavity, and upper respiratory passages. Pigeons have been reported to increase cerebral blood flow during hypoxia (Pavlov et al. 1987). In mammals, direct estimations of the PO<sub>2</sub> in the arterial vessels of the carotid rete and systemic arterial blood indicate that a gas exchange process does not occur in the organ (Duling et al. 1979). The great efficiency of the countercurrent system of the fish gills (Fig. 36) may have been necessary for survival in a medium usually deficient in O<sub>2</sub> and where the levels may fluctuate dramatically spatially and temporarily. In birds, the interaction of the pulmonary capillary blood in the parabronchial gas exchange tissue (Maina et al. 1982b) occurs across a thin blood-gas barrier and an extensive surface area (Maina 1984, 1987a; Fig. 40). A continuous unidirectional air flow enhances the efficiency of the gas exchange process (Figs. 32,36). Through the multicapillary additive serial arterialization system which is built in the crosscurrent geometric arrangement between the parabronchial air flow (the air in the parabronchial lumen) and the pulmonary venous blood (Figs. 41,42), in some conditions (e.g., hypoxia and exercise), O<sub>2</sub> level in the arterial blood may exceed that in the end expired air (Scheid and Piiper 1972, 1989; Fig. 36). The avian lung-air sac system is more efficient in arterializing blood than all the other evolved air breathing organs of the vertebrates (Powell 1990). The arterial PCO, in birds may be as low as 0.93 kPa (equivalent value in mammals = 5.6 kPa) compared with that in the expired air of 5.3 kPa (Scheid and Piiper 1972; Scheid 1990). The  $O_2$  extraction ratio in fish and cephalopods ranges between 50 and 80%, in polychaetes 30 and 70%, in crustacea 43 and 76%, sponges 6 and 40%, lamellibranchs 3 and 10%, and ascidians 4 and 7% (Benedict 1938; Hazelhoff 1939; Randall et al. 1967; Hanson and Johansen 1970). A remarkably high value of 92% has been reported in the triggerfish, Balistes capriscus, by Hughes (1967). In fish inhabiting well aerated water, the arterial PO2 is generally high and usually exceeds the values needed to totally saturate the blood (Stevens and Randall 1967). Expectedly, the ventilation-perfusion ratios in fish are very high and range from 9 in the shark, Scyliorhinus (Baumgarten-Schumann and Piiper 1968), to as high as 70 in the trout (Randall et al. 1967): the markedly high value in the trout is due to a high ventilatory rate occurring with low O<sub>2</sub> extraction from water.



Fig. 40a,b

Oxygen extraction decreases with increasing ventilation-perfusion ratio in the dogfish, *Squalus suckleyi* (Hanson and Johansen 1970), and the water boundary layer provides about 80% of the overall resistance to  $O_2$  exchange (Hills and Hughes 1970). The low  $O_2$  extraction values in the gills with a trophic role (Table 8) may be due to the very high rate of flow of water necessary to procure food. In the human lung, where the concentration of  $O_2$  in the inhaled air is 20.9% and in the expired air 16.4%, the  $O_2$  extraction factor is about 22% while in birds (where the concentration of  $O_2$  in expired air 14.5%), the value is 31% (Sturkie



Fig. 41. Double latex cast preparation, i.e., simultaneous injection of cast material into the vasculature and into the airways of the lung of the domestic fowl, *Gallus domesticus* showing the crosscurrent relationship between the blood flow in the intraparabronchial arteries, x, and the airflow in the parabronchi of which the locations and the directions are shown by *arrow-heads*. Through this arrangement, gas exchange is effected along the parabronchial lengths, the concentration of O<sub>2</sub> in the pulmonary vein being an aggregate effect of that effected at many sites. The efficiency of the bird lung is largely attributable to this design. *it* Interparabronchial artery; *a* atria. *Bar* 100µm. (Maina 1988a)

**Fig. 40.** a Blood-gas barrier of the lung of a bird, the house sparrow (*Passer domesticus*) showing the extremely thin epithelial cells,  $\succ$ , and a much thicker endothelial one, *r*; *p* plasma layer; *e* erythrocyte. **b** View of the gas exchange region of the lung of the domestic fowl, *Gallus domesticus* showing the interdigitation of the blood, *c*, and air capillaries, *a*; *e*, erythrocytes;  $\succ$ , blood-gas barrier. **a** *Bar* 10.3 µm; **d** 2 µm. (**a** Maina and King 1982; **b** Maina 1982)



Fig. 42. Closeup view of a double latex cast preparation, i.e., injection of cast material into the vasculature and into the airways of the lung of the domestic fowl, *Gallus domesticus*, showing the crosscurrent relationship between the blood flow in the intraparabronchial arteries, *i*, and the air flow in a parabronchus (direction shown by *black arrowheads outlined in white*) which are studded by atria which terminate in air capillaries, *a*. The relationship between the intraparabronchial arteries with the parabronchi is crosscurrent, i.e., is perpendicular, while that at the gas exchange site (*large black arrowheads*,  $\succ$ ) is countercurrent. The efficiency of the bird lung is largely attributable to the multicapillary serial arterialization system integral to the crosscurrent arrangement. *white arrowheads*, interatrial septae. *Bar* 20µm. (Maina 1988a)

Filter-feeding animals		Non-filter-feeding animals	
	FIO <sub>2</sub>		FIO,
Sponges	$0.19(0.06-0.57)^{a}$	Annelids	0.41 (0.30-0.66)
Lamellibranchs	0.07 (0.05-0.09)	Crustaceans	0.49 (0.29-0.76)
Ascidians	0.06 (0.04-0.07)	Gastropods	0.66 (0.38-0.70)
		Echinoderms	0.53 (0.49-0.55)
		Fish	0.62(0.46-0.82)
Mean	0.11		0.55

Table 8. Coefficient of oxygen extraction  $(FIO_2)$  from water in filter-feeding and non-filter-feeding animals. (Hazelhoff 1939)

<sup>a</sup> Values in parentheses show the range of the values.

1954). These values are on the lower side of the maximal ones of the water breathers given above. Although a countercurrent flow was established in the gills of the shore crab, Carcinus maenas, the O2 extraction factor of 7 to 23% is relatively very low (Hughes et al. 1969a) compared with the values reported by Hazelhoff (1939) of 60 to 90% in Caloppa granulata. This is probably due to factors such as presence of a large branchial dead space, low permeability of the lamella cuticular lining, and inadequate lamellar perfusion in Carcinus. The importance of the countercurrent arrangement in the efficiency of gill function is evident from the fact that if the direction of flow of either of the respiratory media is reversed (i.e., to establish a concurrent system), the O<sub>2</sub> extraction ratio falls to below 10% (Hughes 1963). In the crosscurrent system, however, a similar procedure has no appreciable effect on the O<sub>2</sub> extraction ratio (Scheid and Piiper 1972). As expected from the geometric configuration of the parabrochial vasculature and blood flow to the exchange tissue relative to the air flow (Figs. 41,42), only the sequence of capillary arterialization is changed. The degree of arterialization remains the same.

# Gas Exchange Media, Respiratory States, and Environments

"The interaction between organisms and their environment is an old but very important problem to biologists. Organisms respond to environmental change in different ways according to the time during which the environmental change persists and according to the magnitude of the stress". Prosser (1958)

# 3.1 Water and Air as Respiratory Media: General Considerations

Regarding the part of the biosphere they occupy, animal life is classified into aquatic, terrestrial, and aerial groups. Among vertebrates, fish are predominantly aquatic, amphibians are transitional, and reptiles, birds, and mammals are fundamentally terrestrial. Overlaps in occupation of various ecosystems occur. Among mammals, the cetaceans have reinvaded water while some amphibians live in highly desiccating deserts (McClanahan et al. 1994). The insects, the now extinct pterosaurs, the birds, and the bats, chronologically in that order, are the only groups which have evolved powered flight. The assortment of animals such as the flying squirrels, lemurs, snakes, lizards, and flying fish which can momentarily remain air-borne are essentially gliders. They use a part of their body to delay the fall and did not have to grapple with the aerodynamic and aerobic challenges which beset the active flyers.

The biophysical properties of water and air have profoundly influenced life patterns and body forms of animals (e.g., Alexander 1990; Giorgio 1990; Strathmann 1990). These features are fundamental to understanding the evolution of form, function, and the divergence between terrestrial and aquatic animals (e.g., Bliss 1979; Graham 1990; Table 9). In particular, the differences between water and air have so greatly influenced the respiratory processes that the mechanisms for obtaining  $O_2$  and for eliminating  $CO_2$  which are efficient in water often fail in air. Water has been abundantly and intricately incorporated in the composition of the living tissues and cells. It is required for life by practically all living things. This ubiquitous molecule makes up as much as 90% of the total protoplasmic mass. It is unequivocally the sine qua non of life. It has been worshipped (e.g., River Ganges in India), wars have been fought over it, and civilizations have risen from it or collapsed after losing or mismanaging it (e.g., Leopold and Davies 1968). While some very simple organisms can live without O2, none can grow without it. The fluctuations in the levels of O<sub>2</sub> and CO<sub>2</sub> in water as well as in air have greatly determined the biomass and the species composition and distribution of animal communities (Davies 1975; Graham 1990).

Among animals, remarkable differences in the tolerance, response, and susceptibility to low  $O_2$  and high concentrations of  $CO_2$  exist. The primary factors which govern respiratory adaptation in both air and water include: (1) the

Parameter	Water	Air	Air/Water	Consequences
$O_2$ and $CO_2$ diffusivity	+	++++	≈8000	O <sub>2</sub> and CO <sub>2</sub> tensions; acid-base balance
O <sub>2</sub> capacitance	+	++	30	
$CO_2$ capacitance	++	++	1	
NH <sub>3</sub> capacitance	++++	++	1/700	N end products
Viscosity	++	+	1/60	Work of breathing
Density	+++	+	1/800	Circulation, skeleton, locomotion
Kinematic viscosity	+	++	13	Buoyancy, gravity
Water availability	+++	Very variable		Water turnover
Ionic environment	Very variable	-	_	Osmoregulation, ionoregulation
Sound velocity	++	+	1/4	Audition
Sound absorption	++	+	1/4	
Light refractive index	1.33	1	0.75	Vision
Light absorption	++	+	1/12	
Dielectric constant	+++	+	80	Electroreception
Solubility of molecules	Variable	-	_	Distance
Volatility of molecules	_	+		Chemoreception
Diffusivity of molecules	+	++++		
Heat capacity	++++	+	1/3500	Heat dissipation
Heat conductivity	++	+	1/24	Body temperature
Heat evaporation	$\approx$ 2450 kJl <sup>-1</sup>			, <u>1</u>

Table 9. Main differences between water and air and their physiological consequences. (Dejours 1988)

molecular characteristics of the respiratory medium, (2) the solubility of the respiratory gases in a medium, and (3) the mode of transfer of the respiratory gases in a medium. Since other than subsistence in the different fluid media (water and air) there have been no physical barriers in the evolution of the possible gas exchangers, as envisaged by Perry (1989), water gills, air gills, water lungs, and air lungs should have evolved to the same extent. However, owing to the relatively low solubility of O<sub>2</sub> in water (0.031 mlO<sub>2</sub> per ml water), high viscosity of water, and low vapor pressure in air, water lungs and air gills have only rarely evolved. The difference in the physicochemical attributes of water and air prohibited direct conversion of the gills to lungs: a transitory gas exchanger which had to function equally well in both media was necessary. Air gills and water lungs occur (but rarely) in simple organisms, e.g., in the respiratory pleopods of the terrestrial isopods (e.g., Marsh and Branch 1979; Hoese 1983), in arachnids (Kaestner 1929), and in cases of retrogression from air- to water breathing, e.g., in the aquatic pneumonate gastropods. The gills nurtured the development of the air-breathing organs. In-series vascular connection followed. There was subsequently a gradual deemphasis of aquatic respiration, leading to subsidence of the gills. The gills were directly replaced by the air-breathing organs only where the skin served as a bridging organ.

# 3.2 Physical Characteristics of Water and Air

For utilization as respiratory media, only fluids are configured to offer the necessary convective transport of the respiratory gases. Water, a liquid over the biological range of temperature and pressure, and air, a gas under similar conditions, are the only two naturally occurring respirable fluids. The structural and functional consequences of the interactions between the respiratory media with the gas exchangers have depended on the magnitudes of the changes in the levels of the respiratory gases and the time scale over which the interfacing has occurred (Table 9). Water is an exacting environment to survive in. In saturated water, at 20 °C, 1 ml of O<sub>2</sub> is contained in 200 g of water while 1 ml of O<sub>2</sub> is present in 5 ml of air (weight, 7 g). The rate of diffusion of O<sub>2</sub> in water of  $3.3 \times 10^{-5}$  cm<sup>2</sup> s<sup>-1</sup> (Grote 1967) is lower by a factor of 10<sup>5</sup> compared with that in air of  $1.98 \times 10^{-1}$  cm<sup>2</sup> s<sup>-1</sup> (Reid and Sherwood 1966) while the capacitance coefficient, i.e., increment of concentration per increment in partial pressure of O<sub>2</sub>, in water is only  $1.82 \text{ nmol min}^{-1}$  per 0.133 kPa (Dejours 1988; Table 4).

As a respiratory medium, the general properties of air are more obliging than those of water. Convection requirements are high in water breathers compared with the air breathers. An octopus ventilates 17l of water for each mmol of O<sub>2</sub> consumed (Dejours et al. 1970). In air, all other factors held constant, diffusion facilitates supply of greater quantities of O2. Less energy is expended in the convective transfer of the gas to the respiratory site. Aquatic organisms have evolved within the constraints of an O<sub>2</sub>-deficient environment. Zaccone et al. (1995) contend that the lungs phylogenetically evolved as adaptations to the hypoxic conditions in the aquatic medium in which fish lived. Due to the high solubility of CO<sub>2</sub> in water, the molar concentration of the free gas is about equal to that in air while the concentration of O<sub>2</sub> in water is only about 5% of that in air. Diffusion rather than concentration reduces the rate of transfer of CO<sub>2</sub> in water. Both decreased diffusion and low concentration slow down the transfer of  $O_2$  in water. Many accounts dealing with adaptations of organisms to dissolved gas levels in water address themselves to O<sub>2</sub> availability rather than the concentration of  $CO_2$ . Owing to the high  $CO_2$ :  $O_2$  solubility ratio, if originally normoxic water was to be rendered anoxic from aerobic metabolism alone, the PCO<sub>2</sub> would only increase by about 0.9 kPa (Rahn 1966). Owing to the high  $PO_2$  in air, diffusion across the blood-gas barrier is highly efficient. The arterial partial pressures of O<sub>2</sub> reasonably approach those prevailing in the respiratory medium. While the O<sub>2</sub> extraction factor in the water breathers is high, e.g., 90% in sponges, 60 to 90% in the crab (Caloppa granulata), 33 to 70% in the octopus, 85% in the eel (van Dam 1938, 1954), and on average 85% in fishes (Steen and Kruysse 1964), in the air breathers, the O<sub>2</sub> extraction factor very rarely exceeds 25%.

# 3.3 The Distribution of Water and Air on Earth

There is ample evidence from geological records that water was available on Earth in abundant quantities as early as 3.8 to 3.5 billion years ago (Schopf and Walter 1983). The hydrosphere, which presently covers about 72% of the Earth, was much more extensive during the early times (e.g., Handerson 1913), reaching the greatest extent in the early Paleozoic (Bray 1985). The Earth has a geometrical surface area of about  $5 \times 10^8$  km<sup>2</sup> and the seas and oceans cover  $3.6 \pm 10^8$  km<sup>2</sup> of this area and contain  $1.3 \times 10^9 \text{km}^3$  of water. Practically all the Earth's water is contained in the oceans, with the water in the lakes and rivers constituting only 0.01% of that which is available to life. Freshwater constitutes about 2.5% of the total volume of the Earth's water, with nearly 75% of it locked in a frozen state in the polar ice caps and in glaciers while about 0.5% of it is held in aquifers as underground water (Gross 1990; Shiklomanov 1993). Rivers and lakes are complex, biologically highly productive habitats with short histories on geological time scale. For example, Death Valley in California, the hottest and driest place in the United States, was covered by a lake some 60 m deep only about 20000 years ago (Leopold and Davies 1968). About one third of all bony fish have evolved and live in freshwater. While it is possible to accurately determine the limits of the aquasphere, the dimensions of the aerosphere are more difficult to define. The air gradually rarefies with altitude, approaching the total vacuum of the vast outer space at an altitude of between about 100 to 1000km (e.g., Denney 1993). From the highest altitude at which aerial animals, especially birds, operate, which with some exceptions is about 10 km, the biologically utilizable fraction of the atmosphere is  $5 \times 10^9$  km<sup>3</sup> in volume. This is nearly four times the volume of the oceans. The physical characteristics of the ocean and seawater are fairly constant while those of the freshwater lakes and rivers are more variable (e.g., Clarke 1991). Mass movement of air by convection and diffusion equalizes the gas tensions within and between habitats. In the open seas, circadian variations in the respiratory gas tensions and concentrations are common in the superficial layers of the water (Riley and Skirrow 1975) but are much less than in ponds or rock pools.

By definition, a normoxic medium is one in which the  $PO_2$  at sea level is about 20 to 21.3 kPa. For any  $PO_2$ , air contains more  $O_2$  than water. The difference between  $O_2$  production through photosynthesis and utilization in respiration determines the net levels and changes in the concentration of  $O_2$  while  $CO_2$ uptake by plants (during photosynthesis) and production during respiration by both plants and animals regulate its level (Fig. 10). Ensuing from the high diffusivity of  $CO_2$  in air, increases in the  $PCO_2$  to a level that would cause respiratory stress in the free atmosphere rarely occur except in certain microhabitats, e.g., in the waterlogged soils where local hyperbaria may occur. In free air, hyperoxia does not arise. However, in ponds or seawater pools, it may develop as a localized phenomenon. Peak levels as high as 80kPa have been reported (Truchot and Jouve-Duhamel 1980; Heisler 1982a). In our presently stable aerial environment, hyperoxia is only studied as a means of understanding the respiratory control mechanisms and management of some clinical problems.

Although O<sub>2</sub> has been produced by photosynthetic organisms for the last 3.5 billion years (Chapman and Schopf 1983; Schopf et al. 1983; Fig. 8), anaerobic habitats have existed continuously during the entire Earth's history. The marine detrital sediments constitute the most extensive global continuum of an anaerobic niche. Oxygen penetrates the sediments to only a few millimeter (Revsbech et al. 1980a). The widespread distribution of black shales is a firm indicator of the anoxic conditions in the seas at particular depths in the early periods of the Earth's geomorphosis (Berry and Wilde 1978). Much of the CO<sub>2</sub> in the natural waters is derived from carbonates in solution and very little from the atmosphere. Acidity increases the PCO<sub>2</sub> in water while alkalinity, even in form of carbonates, lowers it. Even in carbonate-free water, the PCO<sub>2</sub> cannot be very much due to the high solubility of the gas in water, a factor which reduces its occurrence in a free molecular state to almost zero. It is presumably due to this singular fact that CO<sub>2</sub> does not constitute a regular respiratory stimulant in gill and other water breathers. In some habitats, putrefactive processes which entail bacterial anaerobic breakdown of organic matter may produce enormous quantities of CO<sub>2</sub>, which may result in adverse tensions of it. In water which is free of or contains scarce aquatic plant life, dissolved O2 is the most important respiratory factor. At a critical level, an organism may be unable to procure adequate amounts of O<sub>2</sub> for aerobic metabolism. In such cases, it has to evoke certain behavioral, physiological, morphological, and biochemical measures (essentially in that order) (e.g., Bartholomew 1988; Carroll 1988; Gans 1988). These may include relocation to more favorable habitats, increase in O<sub>2</sub> uptake from the inimical environment through physiological adjustments, and reduction in O2 need by entering an ametabolic state. In long-standing cases of hypoxia, the animal is driven to evolve a capacity to procure O<sub>2</sub> directly from the atmosphere. In some organisms, extreme hypoxia results in reversible cessation of respiration especially in those animals like the lugworm, Arenicola marina (Toulmond and Tchernigovtzeff 1984), and the prawn, Palaemon elegans (Morris and Taylor 1985), which experience large diurnal fluctuations in O2 levels in the rockpools. Hypoxia increases ventilation in all animals which have been studied and variably, hyperoxia results in hypoventilation inducing hypercapnia (Dejours 1988).

# 3.4 Water: a Respirable Medium and an Integral Molecule for Life

It is believed (e.g., Jervis 1995) that the quantity of water presently found on Earth already existed when the planet was formed some 5 billion years ago, but in the vapor form. With the gradual cooling of the planet to below  $100 \,^{\circ}$ C, liquid water was formed. In biological systems, the fundamental life processes like ionic and gas fluxes take place in aqueous solution. The suitability and fitness of the water molecule as the habitat in which life evolved on Earth was well affirmed by Handerson (1913). Over millions of years water has shaped and continues to shape the Earth. As a chemical compound, water (H<sub>2</sub>O) is unique and paradoxical in many ways. It is a remarkably stable odorless, colorless and tasteless liquid with powerful solvent properties. Until 1783 (some 200 years ago) when Henry

Cavendish synthesized the water molecule by igniting hydrogen and oxygen, it was believed that water was an indestructible element rather than a chemical compound. The basis of the unique chemical and physical properties of water (e.g., powerful solvency power, high capillarity, expansion of liquid water between 0 and 4 °C, high thermal capacity) is the covalent bonding between O<sub>2</sub> and H<sub>2</sub> producing a dipolar molecule. Water boils at about 162 °C higher than its analog hydrogen sulfide. Its density at 4 °C is 1 g cm<sup>-3</sup>. At 1 atm pressure, water melts at 0 °C and boils at 100 °C (the thermometric fixed points). It is a good ionizing solvent, a property connected with its high dielectric constant and its ability to donate and share electrons. Water acts as a catalyst, reactant as well as a solvent. For example, in some reactions dry ammonia will not react with hydrogen chloride and neither will dry carbon monoxide and dry O<sub>2</sub>. Under geologic conditions of time, heat, and pressure, water has an important role in the conversion of plant and animal matter into organic fuels. At high temperatures, water carries out condensation, cleavage, and hydrolysis reactions (Siskin and Katritzky 1991). Ethers and esters, compounds which are not susceptible to heat alone, suffer facile cleavage and hydrolysis respectively in water at 250 and 350 °C. As the temperature of the water rises from 25 to 300 °C, some of its physical characteristics change dramatically: the density decreases from 0.997 to 0.731 g per ml, the dielectric constant decreases from 78.85 to 19.66, and the solubility parameter from 23.4 to 14.5 cal ml<sup>-1</sup>. Between 250 and 350 °C, the water solvent properties approach those of polar organic solvents at room temperature.

Among the nine planets in our solar system, the Earth is endowed with large quantities of water naturally occurring in the three fundamental states of matter, i.e., liquid, solid, and vapor. It is debatable how liquid water was maintained on early Earth and Mars (e.g., Carr 1996; Sagan and Chyba 1997) despite the solar luminosity being 25 to 30% lower than at present (e.g., Newman and Rood 1977). It is popularly believed (e.g., Owens et al. 1979) that high levels of CO<sub>2</sub> produced by action of the carbonate-silicate cycle provided a greenhouse effect adequate to warm the early Earth. Though now cold and dry and having a thin atmosphere from constant loss of water and CO<sub>2</sub> into space, from study of the surface topography and the geometry of outflow channels, Mars appears to have experienced episodes of massive flooding (Baker et al. 1991). As recently as 300 million years ago, the conditions on Mars appear to have been amenable to life (Kargel and Strom 1996), at least as we know it. Compared with air, water is a more dynamic habitat. Aquatic animals may face wide extremes in O2 availability due to factors such as ice cover, plant respiration, animals burrowing into the substratum for food or protection, and high environmental CO<sub>2</sub> and hence low pH, usually during nighttime plant metabolism.

Some of the unique physical features of water which are fundamental to animal and plant physiology include: (1) a high specific heat (0.9988 calories per g per °C) which affords a stable temperature, (2) relatively weak intermolecular forces which enable heat to be efficiently transferred by convection (thermal conductivity, 5.14 cal  $h^{-1}$  cm<sup>-1</sup> K<sup>-1</sup> at 20 °C) within the medium, (3) high surface tension which at 72.8 dynes cm<sup>-1</sup> is one of the highest among liquids, (4) high wetting property and capillarity, features which are crucial in protoplasmic-cell organelle interfacing, (5) neutral pH which promotes fast reaction kinetics in most

biochemical processes, (6) great solvency power and high dielectric constant (80.1 in pure form) which enable it to accommodate different molecular and ionic factors, (7) maximum density (at 4 °C) which makes it possible for water to sink to the bottom while still in a liquid form. The top layer, which cools to below 0 °C, expands and floats to the top, providing a surface cover of ice which prevents the entire water mass from freezing and killing most of the organisms (which take refuge at the bottom), and (8) in vapor form, water has high diffusivity and is thus important for thermoregulatory processes. The presence of dissolved salts, e.g., in seawater, lowers the temperature at which the maximum density is attained. In brackish water (salinity 18 ppt), the maximum density occurs at 0 °C and in the ocean water (35g of salt per kg of liquid), it occurs at -3.5 °C. Within ordinary conditions of life, water is incompressible. The intermolecular distance is about ten times less than the average value for most gases. Owing to the much weaker cohesive forces (van der Waal's forces), CO<sub>2</sub> and O<sub>2</sub> are gaseous at the physiological range of temperature while in water, where the forces are much stronger, the gas/liquid transition (when the cohesive forces are overcome by the kinetic energy of the molecules) takes place at the relatively high temperature of 100 °C (373 K) at 1 atm (101.3 kPa) pressure. It is only at extreme pressures, e.g., at great depths of the seas and oceans, that compression of water becomes significant in biology (Somero 1992). At a depth of 10km, the PO<sub>2</sub> is 0.8 atm (Enns et al. 1965). Gases such as CH<sub>4</sub>, H<sub>2</sub>S, N<sub>2</sub>, NH<sub>3</sub>, SO<sub>2</sub>, H<sub>2</sub>, and CO<sub>2</sub> are found dissolved in some aquatic habitats. Many flatfishes burrow into the bottom mud, a process likely to reduce the flow of water across the gills impeding excretion of CO<sub>2</sub> and O<sub>2</sub> uptake (Lennard and Huddart 1989). A concentration of H<sub>2</sub>S of 12 mg per l was reported in a lake in Cyrenaica by Smith (1952). Despite its abundance in the atmosphere,  $N_{2}$ , which constitutes about 78.08% by volume at normal pressures, plays no known respiratory role. At high pressures, it may be lethal by inducing narcosis.

## 3.4.1 Oxygen and CO<sub>2</sub> Content in Water: Effect on Respiration

In natural circumstances,  $O_2$  and  $CO_2$  are the only gases of biological interest in water. The concentrations of these gases have dramatically fluctuated during the past geological epochs (Graham et al. 1995; Fig. 9). While  $CO_2$  tension decreased from a peak values of about 100 times of the Precambrian period to the present low level,  $O_2$  has exhibited remarkable changes (e.g., Kasting et al. 1979). Mainly owing to the complex interrelationship between the total concentration of  $CO_2$ and its partial pressure in water, particularly if water contains  $CO_2$  fixing (buffering) factors such as carbonates, in many freshwaters and to a slight extent in marine water, it is difficult to accurately predict the amount of  $CO_2$  in a given kind of water. The levels of  $CO_2$  in waters (except in distilled water, which is devoid of any buffer system and hence  $CO_2$  exists in a dissolved form) differ remarkably as opposed to its rather stable level in the atmosphere. Large quantities of  $CO_2$  may exist at very low tensions due to formation of bicarbonates. Changes in  $CO_2$  levels greatly affect its excretion and hence pH regulation in water breathers. In the ocean water, which has a high environmental buffering capacity, the PCO<sub>2</sub> does not vary much from the value of 0.03 kPa which is the equilibrium point with the atmosphere. When exposed to water, the atmospheric gases are taken up at the surface by diffusion and convection until an equilibrium ( $PO_2 = 20$  to 21.3 kPa) is reached: the enrichment is effected mainly by the currents which are produced by the winds as well as by the variations in the temperatures which influence the specific gravity of the water. Over a distance of a few mm, diffusion is a very slow process. Krogh (1941) estimated that it would take as long as 42 years for an O<sub>2</sub> molecule to reach a depth of 250 m of water. In oceanic inlets, notable vertical stratifications in O<sub>2</sub> levels attributable to changes in temperature and salinity, horizontal variation associated with pockets of photosynthesis, and respiration and decay of organic materials occur (e.g., Platt and Irwin 1972): while the surface water is close to saturation, the bottom one is virtually anoxic, with the  $O_1$  levels generally being below 1 mll<sup>-1</sup> at depths exceeding 100 m. In seas and oceans, as temperature and salinity increase, O<sub>2</sub>, CO<sub>2</sub>, and NH<sub>3</sub> content decrease (Gameson and Robertson 1955). In most cases, anoxia is accompanied by increased concentrations of CO<sub>2</sub> and H<sub>2</sub>S (Powell et al. 1979). With increased temperature, O<sub>2</sub> content drops due to reduced solubility while the PO<sub>2</sub> drops only moderately owing to compensatory increase in the kinetic molecular diffusivity. The effect of temperature on the solubilities of CO<sub>2</sub> and NH<sub>3</sub> is rather complex owing to the chemical reactions with the water molecule. It is not as easily predictable as is the case for the relatively inert  $O_2$ .

Dependent on factors such as temperature, barometric pressure, photosynthetic activity of the plant matter, respiratory processes of the microorganisms, circulatory and mixing processes, and concentration of dissolved solids, at sea level, a liter of distilled and surface sea water contains only 6.34 and 5.11 cm<sup>3</sup> of O<sub>2</sub> respectively. These values are equivalent to the O<sub>2</sub> content of the rarefied air at an altitude of over 20km. At 15°C, 11 of pure water dissolves 11 of CO<sub>2</sub> at 1 atm pressure. The great solubility of CO<sub>2</sub> in water compared with air suggests that the greatest challenge towards the evolution of air breathing was not that of acquisition of O<sub>2</sub> but rather that of elimination of CO<sub>2</sub> into the air (Sect. 1.18 and 5.4). The respiratory epithelia of the air breathing organs, e.g., gills in fish (Randall et al. 1981), skin and lungs of amphibians (Toews et al. 1978), and lungs of most higher vertebrates (e.g., Bidani and Crandall 1988) contain appreciable quantities of carbonic anhydrase which accelerates the dehydration of HCO<sub>3</sub><sup>-</sup> ions in blood, enhancing CO<sub>2</sub> clearance across the gas exchanger (Burnett et al. 1981; Gros 1991). In torrential rivers and streams, as well as the surface waters of lakes, seas, and oceans, the PO<sub>2</sub> and PCO<sub>2</sub> are at or near equilibrium with the atmosphere due to turbulence and agitation, which enhances the solubility of air (Piiper et al. 1962). However, in a variety of closed or stagnant collections of water such as the Black Sea, (Sverdrup et al. 1949), the inner continental shelves like the Gulf of Mexico (Leming and Stuntz 1984), bottom or even surface waters of tropical marshes (Carter and Beadle 1931; Jones 1961), small ponds and intertidal pools (Sverdrup et al. 1949; Truchot and Jouve-Duhamel 1980), interstitial water held between sediments (Brafield 1964; Revsbech et al. 1980b; Reimers et al. 1986), and the hydrothermal vents of the midocean ridges (Johnston et al. 1986), the water is hypoxic and even virtually anoxic. In such waters, the depletion of O<sub>2</sub> usually correlates with elevated CO<sub>2</sub> concentration and low pH status especially at night

Т		Distilled water		Seawater <sup>a</sup>		Air
°C	$\beta WO_2$ $\mu mol l^{-1} mmHg$	βWCO <sub>2</sub>	$\beta WO_2/\beta WCO_2$	$\beta WO_2$ µmol l <sup>-1</sup> mmHg	βWCO <sub>2</sub>	βg
40	1.35	31.33	0.0431	1.10	29.40	51.23
37	1.40	33.52	0.0418	1.17	30.40	51.73
35	1.43	34.99	0.0409	1.21	31.30	52.06
30	1.53	39.30	0.0389	1.32	34.47	52.92
20	1.66	44.86	0.0370	1.43	39.01	53.81
15	2.01	60.23	0.0334	1.67	51.58	55.68
10	2.23	70.57	0.0316	1.83	60.00	56.66
5	2.52	84.17	0.0299	2.03	70.79	57.68
0	2.87	101.25	0.0283	2.28	85.00	58.74

Table 10. Oxygen and carbon dioxide capacitances in distilled water, in seawater, and air at various temperatures. (Dejours 1981)

 $^{\rm a}$  Data on seawater based on chlorinity of 19 ppt; in air, the capacitance of  $O_2$  and  $CO_2$  are identical.

when photosynthesis stops and respiration continues (Heisler et al. 1982; Truchot and Jouve-Duhamel 1980). Water PCO<sub>2</sub> values as high as 8 kPa (Heisler et al. 1982) and PO<sub>2</sub> values beyond 67 kPa (Dejours et al. 1977) have been recorded in natural waters. By suppressing gas ventilation and hence CO<sub>2</sub> elimination, hyperoxia may elevate PCO<sub>2</sub> in blood and precipitate acidosis. Respiratory gases are less soluble in salt solutions (Table 10). Whereas pure water at 0 °C contains 10.29 mlO<sub>2</sub> per l, at a salinity of 10 ppt, it contains  $9.13 \text{ mlO}_2$  per l and at 20 ppt,  $7.97 \text{ mlO}_2$  per l. At 30 °C, the quantities of dissolved  $O_2$  are 5.57 mlO<sub>2</sub> per l (pure water), 5.01 mlO<sub>2</sub> per l (10 ppt salinity) and 4.46 mlO<sub>2</sub> per l (20 ppt salinity). Contingent on location, size of the water mass, and physical mixing, convective movements occur up to a particular depth, the thermocline. Below the critical level, the water is stagnant and is held at a relatively low temperature. In oceans, convective currents equilibrate the respiratory gas tensions to an appreciable depth but at 100 to 1000 m, O<sub>2</sub>-poor areas exist (Harvey 1957). In physically isolated water masses such as the Black Sea and the Gulf of Panama, where conditions for circulation are restricted, hypoxia may exist at all levels. Due to the relatively slower rate of diffusion and convective movement of the respiratory gases, compared with air, aquatic environments are subject to greater spatial and temporal variations in O<sub>2</sub> and CO<sub>2</sub> levels resulting from biological activities of plant and animal life (Garey and Rahn 1970). In isolated water basins and fjords, convective mixing may be prohibited by flow and thermal differentials and/or salinity-induced layering of water leading to hypoxic and even anoxic conditions (e.g., Seliger et al. 1985). In some coastal estuarine areas, due to its lower density, the riverine water floats on top of the heavier seawater (as a result of its less salinity and higher temperature) causing differences in  $O_2$  and  $CO_2$  tensions between surface and deep waters (e.g., Gnaiger 1983; Officer et al. 1984). The diversity of benthic species decreases with depth and varies seasonally with the available  $O_2$  (Hoss 1973).

Fluctuations in the levels O<sub>2</sub> and CO<sub>2</sub> are common in water. The frequency and amplitude are determined by factors such as depth and rate of water circulation, transparency and presence of dissolved and particulate substances, temperature, density of flora and fauna, eutrophication of organic matter, and presence of ice or plant cover (e.g., Carter 1955). The penetration of light through water is an important factor which determines productivity and hence availability of food to aquatic life. It depends on factors such as surface cover, turbidity from presence of suspended mineral particles, organic matter and microorganisms, and the wavelength of light. The blue-green light travels to a depth of 50 m and the red light is least penetrative (Harvey 1957). Solar light is entirely extinguished at a depth of 1000 m in the clearest of the oceans (Pough et al. 1989). In the turbid water of Lake Victoria, long wavelengths penetrate into the water layers more than the shorter ones (van Oijen et al. 1981; de Beer 1989): during the day, at a depth of 2.5 m, the light is about 5% of the intensity at the surface. During the day, owing to photosynthetic activity of algae and other simple plants, O<sub>2</sub> production may exceed the respiratory demands with the concentration increasing to very high levels, while during hours of darkness the water may be rendered totally anoxic from the ensuing respiration (Ultsch 1973; Kramer et al. 1978; Fig. 43). Compared with freshwaters,  $O_2$  saturation levels are generally more stable in marine waters where, due to turbulent mixing with air, surface water is often at equilibrium with the atmospheric gases. In the sea, O<sub>2</sub> levels in the photic zone occasionally rise to 130% saturation or above owing to photosynthetic activities



**Fig. 43.** Diurnal fluctuations of the partial pressure of  $O_2$  and that of temperature in a pool of water containing aquatic plants. The water is saturated with  $O_2$  at about midday and is virtually anoxic after midnight. *Dashed line* indicates the PO<sub>2</sub> in air. (After Dejours 1988)

(Fairbridge 1966). At the bottom, most water masses are asphyxic. In the brackish Japanese Lake Nakanoumi, at the surface, owing to algal photosynthetic activity, the water is 100% saturated with O<sub>2</sub> while at the bottom (depth 6.5 m), the concentration is almost zero due to the biodegradation of organic matter (Kimoto and Fujinaga 1990). In the marine intertidal rockpools, the PO<sub>2</sub> ranges from 0.27 kPa at night to more than 66.7 kPa during the day while the PCO<sub>2</sub> and the pH respectively range from 0.36 to  $1.3 \times 10^{-4}$  kPa and 7.3 to 10.2 (e.g., Truchot and Jouve-Duhamel 1980). Resulting from photosynthetic activity, over a 24-h period, in rockpools, the temperature reaches a high of 24 °C and a minimum of 14 °C. During the day, the concentration of  $O_2$  is in excess of 20 mg per l (more than 300% saturation) dropping to less than 1 mg per l (about 3% saturation) during the night (Daniel and Boyden 1975): the concentration of  $CO_2$  is reduced from 100 mg per l to about half of the value while the pH drops from a night time value of 7.5 to a daytime maximum of 9.5. In warm sunny days, the shallow ditches of the Dutch polder lands present a diurnal PO<sub>2</sub> peak level of 66.7 kPa dropping to 2.7 kPa at night (Jones 1961). Supersaturation with  $O_2$  as high as 364.5% in the upper portion of a Wisconsin lake was attributed to the photosynthesis of the algal growth (Welch 1952). In Lake Waubesa (Wisconsin, USA), a concentration of O<sub>2</sub> as high as 30 mg per l which resulted in sudden fish mortalities was reported by Woodbury (1942). Similar observations were made by Wiebe (1933). Seasonal low levels of O<sub>2</sub> in freshwater masses can lead to massive deaths of fish (winter kills) which result from decay of organic materials especially where atmospheric O<sub>2</sub> recharge is prohibited by surface ice cover (Gnaiger and Forstner 1983). In such cases, aquatic hypoxia is exacerbated by reduced level of photosynthesis due to the decrease of the solar (shortwave) radiation available to the plant life. Atlantic lobster kills are sometimes attributed to low dissolved O<sub>2</sub> (Young 1973). Where light can get through, however, the bubbles which are released by underwater plants and accumulate under the ice may contain as much as 45% O<sub>2</sub> by concentration. The air may be utilized by aquatic organisms (Krogh 1941). In most air-breathing vertebrates, hypoxia is the main drive for respiration, but at sea level CO<sub>2</sub> and H<sup>+</sup> are the basic biochemical regulators. Such animals maintain CO<sub>2</sub> in a steady state leading to an average arterial PCO<sub>2</sub> of about 5.3 kPa. In the aquatic animals, PCO<sub>2</sub> levels are very low. The importance of CO<sub>2</sub> in the regulation of breathing increases with dependence on air breathing. Ventilatory activity in fish which breath water is driven by  $O_2$  levels and is virtually insensitive to CO<sub>2</sub>. Diving (e.g., Andersen 1966) and fossorial (e.g., Augee et al. 1970/71) airbreathing vertebrates (birds and mammals) have blunted sensitivity to CO<sub>2</sub>.

#### 3.4.2 Density and Viscosity of Water

Water is about 1000 times denser than air. The molecular diameter of water is about 2 Å while the intermolecular distance is about 3.1 Å. The average intermolecular distance in gases is about 33 Å while the molecular diameters of most gases ranges from 2 to 5 Å, i.e., about one tenth of their intermolecular spacing. Water, a medium with a high specific gravity and viscosity, provided the necessary support which promoted the development and evolution of the delicate invertebrate life to amazing sizes: a specimen of the Atlantic giant squid (*Architeuthis* sp.) weighing 2t has been captured. However, the metabolic cost of convective transfer of water in the gas exchangers as well as the restrictive physical movement in it sets a limit to the exercise and metabolic capacities of aquatic organisms. The fastest fish, the tuna, can only briefly attain and sustain a maximal speed of about  $20 \text{ m s}^{-1}$  (for 10 to 20 s) while a bird, e.g., the swift will attain and sustain a speed of 40 m per s (Gray 1968). The cost of aquatic respiration is increased by the fact that although the PO<sub>2</sub> may be similar in water and air, water contains 30 times less O<sub>2</sub>. To extract an equivalent quantity of O<sub>2</sub>, a water breather has to expend much more energy than does an air breather (Sect. 2.9).

### 3.4.3 Thermal Capacity and Conductivity of Water

The heat transfer properties, caloric capacity, and conductivity of water is about 3 orders of magnitude greater than that of air (Tables 4,9). With exception of liquid ammonia, water has the highest specific heat  $(4200 \, J \, kg^{-1})$  of any substance in liquid form at room temperature. The thermal conductivity and capacity of water are respectively 24 and 3000 times greater than that of air. These features present constraints for survival and respiration in water. In aquatic breathers, the metabolic heat carried by the blood to the gills is soon lost to the environment (Carey 1973; Carey and Lawson 1973). The temperature of the body tissues is within 1 °C of the ambient water temperature (Carey et al. 1971; Reynolds et al. 1976). The limitations precipitated by the water with respect to heat conservation and body temperature regulation obliged terrestrial location for evolution of endothermic-homeothermy to develop. For similar reasons, except for the endothermic fish, the extant members of the vertebrate classes Agnatha, Chondrichthyes, and Osteichthyes are obligatively poikilothermic (Hazel 1993).

For the period over which the multicellular organisms have been on Earth (Schopf et al. 1983; Fig. 8), the average temperature of the tropical surface waters has varied by only about 5 °C while that of air has changed by perhaps 15 °C (e.g., Cloud 1988). Surface temperatures may change dramatically within short intervals and distances. Annual temperature fluctuations of as much as 60 to 70 °C on land are occasionally recorded. The annual latitudinal fluctuations in temperatures throughout all temperate and subtropical seas range between 0 to 28 °C and at no place in the open sea is the annual range of temperature more than 10 °C (Nicol 1960). In the North Atlantic Ocean, the surface temperature varies by only 8°C while at the equator, the mean yearly temperature variation is only 0.5°C (Sverdrup et al. 1949): deeper waters exhibit less variations in temperature. The mean annual temperature of the ocean at the equator is about 27 °C, at 30°S latitude it is 20 °C, and at 30 °N it is about 17.5 °C (Wüst et al. 1954). The narrow range of temperatures found in the ocean is attributable to constant convective circulation (F.G.W. Smith 1957) and the high specific heat of water (Dorsey 1940). Freshwater lakes exhibit greater stratification and fluctuation of temperature than oceans, the gradient depending on factors such as drainage, mass of water,

latitudinal location, surrounding terrain, and depth (Beadle 1974; Hutchison 1975). The temperature characteristics of a lake particularly in the warmer parts of the year are more complicated than those in the seas and oceans (Beadle 1974): the temperature from the surface to a depth of about 6 m is fairly stable at about 20 °C, from 6 to 10 m it drops suddenly to about 5 °C (the thermocline) and stabilizes below that depth. Very small masses of water may show a range of temperature of 0 to 42 °C (Young and Zimmerman 1956). In the salt water pools in southeast England, Mardsen (1976) noted that in summer, diurnal temperature dropped from 24 to 14 °C over a 24-h period and in winter the temperature ranged from a minimum of -0.5 to 5 °C. Many aquatic animals are able to regulate their body temperatures within a very narrow range by ascending or descending in a water column or moving to shallower or deeper waters (e.g., Feder et al. 1982). As the O<sub>2</sub> content of water varies inversely with the water temperature, a compromise has to be established between the needs for O<sub>2</sub> and the preferred body temperature.

# 3.4.4 Derelict Waters: Respiratory Stress from Hypercapnia and Hypoxia

Derelict waters reproduce the inimical environmental conditions in which air breathing evolved. The adaptations by which animals are able to survive in hypoxic and hypercapnic waters are hence of relevant scientific interest. They demonstrate the factors which enforced and the strategies which animals adopted for transition from water- to air breathing. Current geoclimatic data suggest that the environment during the Devonian period, a time during which most of the bony fish first evolved lungs (Pough et al. 1989), was in all likelihood similar to a tropical swamp. It was characteristically anoxic, reducing, and CO<sub>2</sub> occurred at high concentrations (e.g., Valentine and Moores 1976). During the continental formative years, the drainage systems of most tropical swamps particularly the Central ones were not well formed (Bishop and Trendall 1967). Especially in the major land mass located at the equator, which is often called the Old Red Continent (e.g., Livermore et al. 1985), intense putrefaction of the plant matter resulted from the prevalent high temperatures. Pressure and the need to evolve potential for air breathing and even relocation to land was very intense for survival in such pernicious and noxious habitats. While the quantity and the PCO<sub>2</sub> in air is generally low, in some waters, a fairly large amount of CO<sub>2</sub> occurs at low tension but in a few cases it may be found in such a high tension as to suppress respiration (Table 11). Such conditions are common in the warm waters of the tropical regions of the world (Beadle 1974; Munshi and Hughes 1992). Apart from the ephemeral ones which exist only in wet seasons, there are few inland water masses which have not been radically changed in form and drainage pattern during the past million years. Such habitats are associated with abundant and distinct animal and plant communities. The presence of amphibious vertebrates in a swamp corresponds with a radical transformation of one ecosystem to another, a process which may be enhanced by natural and human factors such as drought, siltation, excessive infestation with aquatic flora, drainage, and land reclamation.

Swamp	Area	Т	pН	cO <sub>2</sub>	CO <sub>2</sub>	Alkalinity	Conductivity
Nile swamps							
Swamp rivers	Bahr el Gabel	25	7.5	0.66-6.2	3-18	-	112-550
	Bahr el Ghazal	25	7.4	1.3-8.0	-	-	40-370
	Bahr el	25	7.3	3.4-6.2	-	15-50	110-280
	Bahr el Zeraf	25	7.8	2.2-5.9	-	92–116	245-370
Standing waters	Shambe Lagoon	25	7.5	2.4-6.0	-	32	·_
	Lagoon RP 12	25	7.8	6.1–102	-	21-31	200-250
	Lake	25	6.8	8.0	-	25	40-55
	Khar Perboi	25	-	1.8–7.2	-	-	-
	Khar Atar	25	8.3	6.9–12	-	29	-
Guiana swamps	Grass Swamp A	28	4.5	0.22-1.2	8-9	0	-
	Other Swamps	28	4.5	0.65	14	0	-
	Pool B	28	4.1	0.6-1.5	5.0	0	-
East African swamps	Lake Naivasha (Papyrus swamp)	25	7.8	2.5	-	-	-
	Kazinga Channel (papyrus swamp)	27	5.9	0	-	7.5	-
	Kitoma (papyrus swamp)	-	6.7	0	-	72	
Indian swamps	Polluted swamp	26	8.8	6.5-8.2	16-20	148 ppm	-
	Semi- senescent swamp	26	8.3	0.6-3.2	20	143 ppm	-
	Senescent swamp	31	8.0	1.5-2.7	12	94-162	-

Table 11. Environmental features of some representative swamps of the tropical world. (Beadle 1932; Carter 1935; Rzoska 1974; Dehadrai and Tripathi 1976)

Symbols and units: T – temperature (°C);  $cO_2$  – dissolved  $O_2 mgl^{-1}$ ;  $CO_2$  – free  $CO_2$ ,  $mgl^{-1}$ ; alkalinity – total alkalinity,  $10^{-4}$  N; conductivity,  $\mu$  mho cm<sup>-1</sup> (20°C).



**Fig. 44.** Cross-section of the edge of a Paraguayan swamp showing the effect of aquatic plant growth on the  $O_2$  levels in water. The  $O_2$  tensions both in the open pools and the weeds decrease with depth but even close to the surface, the levels are very low. The commonest fish are *Hoplosternum litorale* and *Erythrinus unitaeniatus*. Both are air breathers. (Carter and Beadle 1931)

The tropical derelict masses of water which are shallow and stagnant or slow moving present adverse respiratory conditions. In standing waters, mixing of surface and deeper waters is inhibited by thermal stratification. The bottom water may be virtually anoxic while the top layer is saturated with  $O_2$ . In a Danish lake, Nielsen and Gargas (1984) observed that O2 saturation was as low as 5% in the near-bottom water layers. In the tropical Paraguayan swamps, the Chaco, which are dominated by emergent macrophytes, Thalia and Typha, both of which grow to a height of 5 m and floating vegetation such as Pistia, Azolla, and Aichhornia, the surface O<sub>2</sub> levels are seldom above 50% saturation and the bottom ones are perpetually anoxic (Carter and Beadle 1931; Fig. 44). In some swamps, the dense plant canopy reduces the photosynthetic production of O<sub>2</sub> by algal activity and prohibits water stirring, factors which compounded by the intense utilization of O<sub>2</sub> in the putrefactive processes make the water anoxic within a fraction of a centimeter of the surface (Carter 1955; Beadle 1957). In the vegetation-covered Floridan swamps, the PO<sub>2</sub> is less than 0.67 kPa, PCO<sub>2</sub> = 8.3 kPa and the pH is 5.6 (Heisler et al. 1982). The waterlogged mat and the bottom peat are not only totally devoid of  $O_2$  but are also highly reducing. Redox potentials (Eh) of -100 mV have been recorded within 30 cm of the water surface (Beadle 1957) and in most such cases, no measurable O2 was detected within 2 cm of the surface. Due to the fact that the decomposing organic matter in the Lake Victoria basin swamps (largely covered by papyrus overgrowth) is mainly carbohydrate, the gaseous end product is mainly CH<sub>4</sub> (60%) with CO<sub>2</sub> consitituting only 30% and the remaining 10%

being made up of H<sub>2</sub>, carbon monoxide, and ethylene (Visser 1963; Dehadrai and Tripathi 1976). In the extensive Sudd swamps of the Upper Nile (Rzoska 1974), a high concentration of H<sub>2</sub>S has been reported (Talling 1957), a feature associated with the much greater reducing conditions where the O<sub>2</sub> levels deep inside the vegetation are only about 10% saturation (Rzoska 1974). The peculiar aspects of the swamps (scarcity of O<sub>2</sub>, highly reducing environment and high CO<sub>2</sub> levels; Table 11) determine the variety, nature, and the biomass and productivity in such habitats (Carter and Beadle 1931). To complicate matters, toxic gases such as H<sub>2</sub>S occur in high concentrations (Somero et al. 1989). In an ecosystem in which the conditions are highly adverse to life, adaptation for air breathing is intense. Such a change confers great selective advantage (Carter and Beadle 1931; Dehadrai and Tripathi 1976). In the Ugandan swamps, aquatic insects (Hemiptera, Coleoptera, mosquito larvae), pulmonate gastropods (e.g., Biomphalaria sudanica), oligochaete worms (e.g., Alma emini), and fishes (e.g., Protopterus aethiopicus, Polypterus bichir, Clarias lazera, Ctenopoma muriei, Gymnarchus niloticus), all of which are air breathers coexist (Beadle 1974; David et al. 1974). The swamps are characterized by a rapid growth of macrophyte cover and subsequent decomposition of the luxuriant organic matter leading to intense putrefaction and anaerobic decomposition of the overgrowth. These processes generally result in a hypoxic and hypercarbic habitat (Nassar and Munshi 1971; Dehadrai and Tripathi 1976; Ultsch 1976), except during episodes of peak photosynthesis.

The tropical swamps shrink during the hot spells or become muddy to the extent that the gills become unsuitable for gas exchange due to clogging by masses of floating and suspended detritus. Such waters are slightly acidic (with a pH of 6 to 6.5) mainly from the high levels of dissolved  $CO_2$ . The Guiana swamps have very low pH ranging from 4.3 to 4.4 (Carter 1935) while those in India are alkaline (pH 8 to 10; Dehadrai and Tripathi 1976; Munshi and Hughes 1992). Dissolved and bound  $CO_2$  in the papyrus Ugandan swamps was found to be 148 mg per l at the end of a rainless season (Milburn and Beadle 1960). Up to 30 ppm free CO<sub>2</sub> was recorded in some Indonesian swamps by van Vass and Vaas (1960). During summer, in the water hyacinth-infested Floridan and Louisianian swamps, Lynch et al. (1947) measured a concentration of dissolved CO<sub>2</sub> as high as 80 ppm (about 213.3 kPa). A zero concentration of  $O_2$  at a depth of about 30 cm, 0.6 ppm (about 1.2 kPa) at a depth of 5 cm and dissolved CO<sub>2</sub> level as high as 8 kPa were determined in a hyacinth infested pond in Gainesville, Florida (Ultsch 1976). In the tropical swamps, the CO<sub>2</sub> levels can rise to a level which would adversely affect the oxygenation of the hemoglobin and disrupt acid-base balance. In such desolate aggregations of water, in circumstances where the dissolved O<sub>2</sub> is low, that of CO<sub>2</sub> is usually high (Lynch et al. 1947). High concentration of O<sub>2</sub> alleviate the effects of high CO<sub>2</sub> levels: a 230-g salamander, S. lacertina, can tolerate a PCO<sub>2</sub> of 28 kPa for as long as 70 h when held in water in which the PO<sub>2</sub> ranges from 17.9 to 37.3 kPa (Ultsch 1976). Owing to their low metabolic rate (Whitford and Hutchison 1967) and capacity to tolerate high concentration of CO<sub>2</sub> (Ultsch 1976), the sirenids are well adapted to subsist in the hyacinth-infested waters where they constitute the dominant vertebrate fauna (Ultsch 1973). In the vegetation-covered lakes of the southeastern United States inhabited by two species of salamanders and the Congo eel, Heisler et al. (1982) reported a PO<sub>2</sub> of less than 0.67 kPa, a PCO<sub>2</sub> of 8.3 kPa, and a pH of 5.6. In a hyacinth (*Eichhomia crassipes*)-infested swamp, zero concentration of  $O_2$  and a high one of  $CO_2$  of 80 ppm were recorded (Lynch et al. 1947; Ultsch 1973). The temperate tidal pools examined by Truchot and Jouve-Duhamel (1980) had a temperature which ranged from 12 to 24 °C and the  $O_2$  content amounted from virtual anoxia to hyperoxia within a 24-h cycle.

High concentration of CO<sub>2</sub> in water leads to certain physiological stesses on the aquatic animal life which call for specific adaptations for subsistence in such a habitats (e.g., Dubale 1959). These include: (1) low O<sub>2</sub> affinity of the blood (the Bohr effect), (2) decrease in O<sub>2</sub> capacity of blood (Root effect), (3) acid-base imbalance due to the decrease in pH of water consequent to an increase in PCO<sub>22</sub> and (4) reduced capacity to discharge  $CO_2$  into the surrounding water owing to a reduction of the PCO, between the blood and the surrounding water. In Anabas testudineus, pH influences the pathway adopted for respiration (Hughes and Singh 1970s). Both aquatic and aerial respiration occur in neutral waters (pH 8 to 6.85) while aerial respiration predominates and aquatic respiration is depressed between pH 6.5 and 6.25. Gill ventilation completely stops (and is replaced by aerial respiration) in very highly acidic water (pH < 6.25). Similar behavior was observed in the vallow (Erythrinus erythrinus), an air-breathing fish, where gill ventilation virtually stopped and was replaced by aerial respiration in hypercarbic waters with a concentration of  $CO_2$  above 39 ml per l (Willmer 1934). Such a response may be a safeguard against excessive transfer of  $CO_2$  from water into blood across the gills, avoiding possible respiratory acidemia (Singh 1976). By switching to air breathing, the fish can tolerate high concentrations of CO<sub>2</sub> without adverse effects. Air-breathing fish, most of which subsist in hypoxic waters, are confronted with a real problem of losing O<sub>2</sub> through the gills into the surrounding hypoxic water (e.g., Smith and Gannon 1978). This is, however, minimized or avoided by vascular reflexes which lead to shunting of blood from the gills with a momentary elimination of branchial ventilation making the accessory respiratory organ(s) the only pathway(s) for transfer of O<sub>2</sub> to the body. Airbreathing fish, e.g., Anabas (Munshi 1968), Channa (Wu and Chang 1947; Hakim et al. 1978; Wu 1993), Amphipnous (Munshi and Singh 1968). and Monopterus (Liem 1961), have very poorly developed gills, perhaps to curtail O<sub>2</sub> loss through the gills during hypoxia. In the lungfish, Protopterus, the arteries of the embryonic 3rd and 4th branchial arches are devoid of gill filaments and form shunt vessels which correspond with the carotid and systematic arches of the Amniota (e.g., Wood and Lenfant 1976).

# 3.5 Terrestrial Habitation and Utilization of Atmospheric O<sub>2</sub>

The degree of specialization and the survival strategies of most animals are determined by the efficiency with which they can in the first instance procure  $O_2$  and secondly eliminate  $CO_2$ . In open aerial environments, production and utilization of  $CO_2$  and  $O_2$  are relatively small compared within the enormous volumes and high capacitances of the gases in the atmosphere (Dejours 1988). The constant level of  $O_2$  and  $CO_2$  in the atmosphere is of biological importance as major
deviations from the tolerable range have harmful effects on unacclimatized animal life. With respect to composition and availability of respiratory gases, air is a much simpler medium to handle than water. Biological, physical, and chemical processes greatly influence the  $O_2$  content of water. In stagnant waters and especially those covered by plant matter, there is reciprocity of environmental  $O_2$  and  $CO_2$  levels. As a result of organic respiratory processes, hypoxia will invariably be associated with some degree of hypercapnia. Except for fossorial habitats, terrestrial habitats are not normally liable to hypoxia or hypercapnia due to the great diffusivity of the gases in air.

#### 3.6 Hydrogen Sulfide Habitats: Tolerance and Utilization

The energy-rich hydrogen sulfide  $(H_2S)$  is common in the hypoxic marine sediments and around hot springs at concentrations of 1 to 300 ppm (e.g., Berner 1963). Hydrogen sulfide is highly toxic even at very low molar concentrations. A concentration greater than 1 ppm is lethal to most organisms (Oseid and Smith 1974; Smith et al. 1976). The gas inhibits several heavy metal containing enzymes especially by binding to the heme of mitochondrial cytochrome c oxidase much as cyanide does. This prevents O<sub>2</sub> transport by the hemoglobin, arresting aerobic respiration (e.g., Somero et al. 1989). About 90% of cytochrome c oxidase is inhibited by a 5-molar sulfide solution (e.g., Julian and Arp 1992) and a sulfide concentration of 17 mol initiates maximal O<sub>2</sub> consumption (Eaton and Arp 1993). At moderately low concentrations (3.63 mol per l), specimens of Rivulus marmoratus were observed to leap from H<sub>2</sub>S-contaminated water (Abel et al. 1987). Some animals, however, have acquired a capacity to tolerate H<sub>2</sub>S and even utilize it for metabolic processes producing water and various sulfates (e.g., Felbeck et al. 1981). Others, e.g., in the Phylum Gnathostomulida (Farris 1976) and the turbellarian families Solenofilomorphidae (Crezee 1976) and Retronectidae (Sterrer and Rieger 1974), have adapted so well that they are virtually confined to the extreme anoxic H<sub>2</sub>S-rich habitats (Fenchel and Riedl 1970). They survive by adopting different strategies which include: (1) physical exclusion of sulfide from the body, (2) possession of sulfide-insensitive cytochrome c oxidase, (3) direct detoxification whereby sulfide ( $H_2S$ ,  $HS^-$  or  $S^{-2}$ ) is chemically converted to less toxic products such as sulfate  $(SO_4^{-2})$ , sulfite  $(SO_3^{-2})$ , and thiosulfite  $(S_2O_3^{-3})$  at the superficial tissue layers of the body or in certain specialized organs (Vetter et al. 1987; Menon and Arp 1992b), (4) strategic coexistence with chemoautotrophic endosymbiotic bacteria which break down H<sub>2</sub>S, utilizing the energy thus acquired for production of ATP (e.g., Felbeck 1983; Fisher and Hand 1985; Powell and Somero 1985; Belkin et al. 1986; Firsher 1990), and (5) dependence on anaerobic metabolism. The marine echiuran worm, Urechis caupo, subsists in U-shaped burrows in intertidal mud flats where the concentration of  $H_2S$  may be as high as  $66 \mu M$  and the concentration of  $O_2$  as low as 3.3 kPa(Eaton and Arp 1993). Hydrogen sulfide passes through the body wall and the hind gut (Julian and Arp 1992), where hematin, which is contained in coelomic fluids and in the coelomocytes (Arp 1991), catalyzes its oxidation to nontoxic sulfur compounds as  $O_2$  continues to be utilized for aerobic respiration (Powell and Arp 1989; Eaton and Arp 1993). A correlation between hematin concentration in the coelomic fluid and sulfide oxidizing activity was reported in *U. caupo* by Powell and Arp (1989). *U. caupo* has a very high tolerance for  $H_2S$  (Somero et al. 1989; Julian et al. 1991). The worm remains aerobic at concentrations above those which suppress the process in most aerobic organisms. In the intertidal lugworm, *Arenicola marina*, a specialized heme compound called brown pigment is thought to catalyze sulfide oxidation (Patel and Spencer 1963). The hemoglobins of the lugworm and the deep-sea hydrothermal vent-living polychaete, *Alvinella pompejana* (Desbruyères and Laubier 1986), are highly resistant to oxidation (Toulmond et al. 1988).

Extreme physiological conditions have been described in the submarine geothermal springs which have been discovered in the Galapagos Rift and East Pacific Rise fracture zones (e.g., Childress et al. 1989; Nisbet 1988). The water emerging from the fissures is as hot as 200 °C, is anoxic, and contains a high concentration of H<sub>2</sub>S (Desbruyères et al. 1982; Johnston et al. 1988). The hot plume of water interfaces with the remarkably different sea-water at a temperature of about 2 °C, with a PO<sub>2</sub> 6.7 kPa, pH about 7.5, and is totally devoid of H<sub>2</sub>S (Johnston et al. 1986). This creates the greatest temperature gradient in any known environment occupied by animal life. Simple photosynthetic life is thought to have evolved in the submarine hydrothermal vents where chemotrophic organisms acquired the capacity to detect light (e.g., Russell et al. 1994; Nisbet et al. 1995). The interface between the hydrothermal plume and the bottom seawater constitutes a highly dynamic and complex ecosystem with fluctuating PO<sub>2</sub> (Johnston et al. 1986). A large number of different organisms such as sulfur oxidizing bacteria and animal species such as giant pogonophoran tube worms, crabs, shrimps, giant crams, fishes, and mussels flourish in complete darkness some 3km below the suface (Grassle 1985; Fustec et al. 1987). Some of these species were unknown to science more than 20 years ago! The animals which stray from the vent risk starvation while those which get too close die of poisoning or heat. The hydrothermal plume crab, Bythograea thermydron, which has no bacterial symbionts, can tolerate high concentration of H<sub>2</sub>S through having a particularly efficient sulfide oxidation capacity in the hepatocytes (Vetter et al. 1987) while Riftia pachyptila and Solemva velum possesses hemoglobins which hold and transport  $H_2S$  preventing toxicity (e.g., Doeller et al. 1988). In the deep sea hydrothermal vent cram, Calyptogena pacifica, which has symbiotic bacteria in the gills, O<sub>2</sub> consumption increases on exposure to concentrations of H<sub>2</sub>S as high as 130 to 160µM (Childress and Mickel 1982). The endosymbiotic bacteria in the gutless cram, Solemya reidi, which live in H<sub>2</sub>S-rich sewage outflows, are able to maintain aerobic metabolism in the tissues through oxidation of H<sub>2</sub>S to thiosulfate in presence of a concentration of  $H_2S$  of 100  $\mu$ M (Anderson et al. 1990) and a much higher one of 500µM in Solemya velum (Chen et al. 1987). The granules in the cytoplasm of the gill cells in Solemya reidi (Powell and Somero 1985; Powell and Arp 1989) and the osmiophillic electron dense organelles in the epithelial cells of the hind gut of U. caupo (Menon and Arp 1992b) are associated with H<sub>2</sub>S detoxification. Similar granules have been observed in the epithelial cells of the "lung" groove (Fig. 45) of the oligochaete swamp worm, Alma emini (Maina et al. 1998)



**Fig. 45.** Dorsal view of the temporary respiratory groove, g, of the oligochaete worm, *Alma emini*, which lives in putrefying plant matter of the East and Central African tropical swamps. The worm occasionally surfaces, forms a lung, and exchanges gases with the atmosphere. \* cloaca. *Bar* 0.95mm. From Maina et al. (1998).

which lives in waterlogged soils where intense putrefaction of plant matter with possible release of H<sub>2</sub>S occurs.

### 3.7 The Porosphere and Fossorial Respiration

In addition to the aerosphere and the hydrosphere, soil, the thin skin of the lithosphere, offers an important natural habitat to many animals. About 150 mammalian species temporarily or permanently live underground (e.g., Nevo 1970). The respiratory properties of the soil are very fluid. Its characteristics fluctuate between those of water (when the soil is wet or waterlogged) to those of air when it is dry. In three-dimensional space, the porosphere can be conceptualized as an intricate maze of fine air conduits between the soil particles. The air spaces open to the free atmosphere. Gas diffusivity in the soil may be up to one third that of free air (Ar 1987) but moist compact soil may offer a severe hindrance to gas diffusion (Currie 1984). Depending on the global temperature changes, soils offer an important source or sink of  $CO_2$  (Susan et al. 1996). The principal factors which determine  $O_2$  and  $CO_2$  content of the porosphere are: (1) the intensity of respiration of the organic matter in the soil (Vannier 1983), (2) moisture content of the soil (e.g., Wilson and Kilgore 1978), and (3) the chemistry of the soil (Verdier 1975).

In general, the underground microenvironments are characterized by a high temperature, reduced light intensity, and in most cases perpetual darkness, low PO<sub>2</sub>, high PCO<sub>2</sub>, and frequently high humidity (e.g., Kennerly 1964; McNab 1966; Arieli 1979). These variables are translations of the surface (free atmospheric) factors except for CO<sub>2</sub>, which is intrinsically produced by the soil fauna. The subsurface microclimates are more stable compared with the more variable and cyclic characteristics above. However, subterranean features such as O<sub>2</sub> and CO<sub>2</sub> levels and, in some cases, moisture content show greater range and oftentimes faster shifts than the corresponding surface ones. The stimulus of light as a physiological phenomenon, which to varying extents influences many of the surface-dwelling animals, is virtually eliminated under the soil. Since the gas requirements of an animal living in an underground burrow must in the first instance be derived from the free atmosphere by diffusion through the soil, many underground conditions, the physical features of the soil especially its porosity, are important in the distribution, abundance, and respiratory activity of fossorial animals (Kennerly 1964).

While many animals spend only part of their time underground in open burrows, only a few of them permanently live underground in closed burrows. In some habitats, subterranean (fossorial) animals constitute an important part of the total fauna. Burrows provide fossorial animals with protection against environmental extremes and predators and enable them to gain access to subterranean parts of plants particularly geophytes (roots, tubers, bulbs, and corms) and soil invertebrates. However, fossoriality imposes certain constraints on the animals which face huge energetic costs in subterranean excavation during foraging (Vleck 1979). Depending on soil hardness and burrow diameter, the cost of burrowing may be 360 to 3400 times greater than that of traveling the same distance on the surface (Vleck 1981). Pocket gophers can dig well over 200 m in 48 h (Hill 1944). The burrows of the naked mole rat (*Heterocephalus glaber*) may be as long as 3 km (Brett 1986). Wet soils constitute a limitation for thermoregulation in fossorial animals (Wierenga et al. 1969). As a consequence of their having had to adapt to a similar mode of life, remarkable convergence of form and function (e.g., Jarvis and Bennett 1990) has occurred in a remarkably phylogenetically diverse group.

The rate of diffusion of a gas or vapor through the soil is determined by the porosity, type of gas, and the prevailing concentration gradient between the air in the burrow and the free atmosphere (Penman (1940a,b). High concentration of CO<sub>2</sub> correlates with the soil moisture content (Kennerly 1964). When the content is high, water rather than air occupies the interstitial spaces between the soil particles thereby impeding the diffusion of gases. Respectively, moisture contents as high as 13 and 18% have been reported in fields occupied by the pocket gopher, Geomys bursarius (Kennerly 1964) and Thomomys bottae (Miller 1948). After heavy rain, 30 cm below the surface, Ege (1916) observed that the PCO<sub>2</sub> increased from 1.5 to 6.1 kPa and PO<sub>2</sub> dropped from 20.4 to 8.5 kPa. When the soil is waterlogged, gas diffusion may become a limiting factor, the soil becoming virtually anoxic (Currie 1962). Some animals living underground (e.g., the earthworms) may succumb to hypoxia or react to it by surfacing. The distribution of the pocket gopher, Geomys pinetis, correlated with that of the soils with a high water-holding capacity (McNab 1966). Acidic soils (podzols) are unable to chemically fix CO<sub>2</sub> while calcareous ones (e.g., clay soil) are able to buffer some CO<sub>2</sub>. Due to the greater solubility and capacitance coefficient of  $CO_2$  in water,  $O_2$  rather than  $CO_2$ diffusion should be the limiting factor in moist soils. Kennerly (1964), however, observed that in soils with different moisture contents, O<sub>2</sub> and CO<sub>2</sub> diffused through the soil at different rates, with O<sub>2</sub> reaching equilibrium much faster than  $CO_2$ . While  $CO_2$  is organically produced within the soil, there is no subterranean source of  $O_2$ , which must be derived by diffusion from the surface. With the  $PO_2$ falling to between 16kPa in Caretta and 12kPa in Chelonia and PCO<sub>2</sub>, respectively, at 3.3 and 4 kPa in the nests, sea turtles hatch into a subterranean hypoxic and hypercapnic environment (Maloney et al. 1990): within 3 to 5 days, they must reach the surface or die of suffocation.

### 3.7.1 Gaseous Composition in Burrows

In addition to soil chemistry and porosity, the other factors which determine the composition of the air in the burrows include the size, length, location, and geometry of the burrow (Wilson and Kilgore 1978), and the numerical density of the burrow congeners. The burrows of the pocket gophers tend to be deeper in summer when the porosity and hence the diffusion capacity of the soil for  $O_2$  is higher (Kennerly 1964). In most soils, the sum of the concentrations of  $O_2$  and  $CO_2$  remains almost constant and for soils with air porosities greater than 10% by volume, the rate of exchange of  $O_2$  and  $CO_2$  with the surface comes to equilibrium

up to depths of 30 cm within a period of 1 h (Collins-George 1959). In the burrows of the pocket gopher, Geomys bursarius, burrows up to 30 cm in depth permit diffusion of  $O_2$  from the surface at rates adequate for the respiratory needs of the gopher. Thermal equilibrium occurs up to a depth of 60 cm (Kennerly 1964). In a forest, 30 cm below the surface, Ege (1916) reported a CO<sub>2</sub> concentration of 0.2% (0.2 kPa) and one of O<sub>2</sub> of 20.6% (20.3 kPa). Aeration of the burrow must be most critical after rains when diffusion of O<sub>2</sub> and CO<sub>2</sub> between the burrow and the ground surface is greatly hindered. Under such circumstances, a net O<sub>2</sub> deficit in the burrow might occur. In the pocket gophers, Thomomys (Miller 1948) and G. bursarius (Kennerly 1964), burrows are opened in the early morning and late evening or night, a time when the cooler laver of air next to the warm ground enhances the diffusion of the air into the extensive burrow systems which range from 60 to 200 m in length. With an average diameter of 7.5 cm, a total burrow roof exposure area of about 60 to 200 m<sup>2</sup> is attained. This was considered by Kennerly (1964) to be more than adequate for the transfer of the respiratory gases while providing access to sufficient vegetation cover with subterranean plant parts, insects, and worms for food.

Concentrations of CO<sub>2</sub> as high 10% and O<sub>2</sub> levels as low as 10% have been reported in burrows (Boggs et al. 1984). The burrows of the pocket gopher (Thomomys bottae) had a concentration of  $O_2$  as low as 6% and  $CO_2$  as high as 3.8% (McNab 1966; Darden 1970; Chapman and Bennett 1975). Concentrations of O<sub>2</sub> as high as 20% and as low as 12.1% have been reported in the burrows of birds, with corresponding values for CO<sub>2</sub> ranging from 1.2 to 9% (e.g., Wickler and Marsh 1981). In ant hills, in summer, Portier and Duval (1929) found a PCO<sub>2</sub> of up to 1.9kPa while Ege (1916) observed that while the PO<sub>2</sub> in the field was 20.4kPa, in an ant hill it was 19.3 kPa and in a decaying beech trunk the value was 17.7 kPa. Concentrations of  $O_2$  of 10 to 15% have been reported in some mammalian burrows by Kennerly (1964). In the burrows of five fossorial rodent species, the concentration O<sub>2</sub> ranged from 15 to 20% and CO<sub>2</sub> from 0.5 to 2.0% (McNab 1966). Concentrations of  $CO_2$  as high as 2.3% and  $O_2$  levels as low as 6% were estimated in the burrows of the pocket gopher, Geomys bursarius (Kennerly 1964), and in dens of hibernating mammals, concentrations of  $CO_2$  as high as 13.5% and  $O_2$  as low as 4% were reported by Williams and Rausch (1973). Soil flora (especially algae and bacteria) is thought to be the main source of CO<sub>2</sub> in the burrows (Kennerly 1964) though the intensity of activity, especially during mound building, must greatly elevate it. The gas concentrations in the burrows are remarkably different from those which can be tolerated by nonburrowing mammals. Extreme hypoxia as well as hypercapnia affect cardiac function in most mammals (e.g., Tucker et al. 1976), induce artificial hypothermia and in some cases torpor (Hyden and Lindberg 1970), have a general depressing effect on growth (Xu and Mortola 1989), and lower the ventilatory rate (Arieli and Ar 1979). The hypoxia that the mole rat, Spalax ehrenbergi, can withstand is comparable to an altitude of more than 9km (Arieli et al. 1977). The respiratory physiology for subterranean subsistence is outlined by, e.g., Arieli and Ar (1979), Boggs et al. (1984), and Webb and Milsom (1994). Insensitivity of the pulmonary vasculature to hypoxia (Walker et al. 1982, 1984) may reduce resistance and ensure adequate pulmonary perfusion. Compared with those of the adult mammals, the lungs of the eusocial naked mole rat, *Heterocephalus glaber* show a remarkable degree of pedomorphy/ neoteny (Maina et al. 1992). Perhaps what appears like a primitive (ancestral = plesiomorphic) state constitutes what has been called an evolutionary stable strategy by, e.g., Maynard-Smith (1996) and McNamara and Houston (1996) where the phenotype has been specially selected for the particular habitat the mole rat lives in. A highly refined resource use and niche occupation may reduce or even eliminate sympatric competition, decreasing fitness cost (e.g., Martin 1996).

#### 3.7.2 Burrowing Aquatic Annelids, Crustaceans, and Fish

For reasons basically similar to those of the fossorial mammals, i.e., escape from predators and occupation of a more stable micromileu, a number of annelids (Mangum 1964; Myers 1972; Toulmond 1991), crustaceans (Little 1983; Atkinson and Taylor 1988), and fish (Pelster et al. 1988a,b; Taylor and Atkinson 1991) have adopted a subaquatic burrowing life-style. Among crustaceans, burrow construction is commonest in members of the infraorders Astacidea, Thalassinidea, and Brachyura. Fish such as the tile fish (Branchiostegidae) (Able et al. 1982) and the garden eels (Anguilliformes) construct mucus-lined tubular burrows (Taylor and Atkinson 1991). The marine polychaete, Chaetopterus variopedatus, lives in elaborate U-shaped tubes which are said to be impermeable to water and gases (Fauvel 1927; Dales 1969; Brown and McGee-Russel 1971) while the lugworm, Arenicola marina, an intertidal polychaete, lives in an L-shaped mucus-lined burrow system (Wells 1966). The mud shrimp, Mediterranean shrimp, Callianassa truncata, builds extensive burrows in the sediments to a depth of 48 cm (Ziebis et al. 1996): the shrimp can maintain burrow O<sub>2</sub> concentration at 3 to 12% of air saturation by generating water currents of a flow rate of  $10 \,\mathrm{m\,s}^{-1}$ .

Like the surface-based fossorial mammals, the subaquatic burrowing animals face unique respiratory challenges that call for special adaptive strategies to overcome hypoxia and hypercapnia. High concentrations of H<sub>2</sub>S are a common occurrence (Bridges 1987; Atkinson and Taylor 1988; Toulmond 1991). Burrowing fish such as *Periophthalmus cantonensis* may encounter extreme hypoxia (PO<sub>2</sub>, 0.7 to 3kPa) at a depth of 5 to 15 cm (Gordon et al. 1978) and may face total anoxia at a depth of about 1 m (Gordon et al. 1985). In the burrows of the snake blenny, *Lumpenus lampretaeformis*, the PCO<sub>2</sub> of the water in the burrow is above 0.2kPa compared with that at the surface of the mud (Atkinson et al. 1987). To alleviate adverse respiratory stress, the crabs use their pleopods (e.g., Farley and Case 1968; Hill 1981) and to a small extent their scaphognathites (Taylor and Atkinson 1991) to intermittently ventilate their burrows and their gas exchangers with better oxygenated water. In most species, e.g., *Callianassa californiensis*, the beating of the pleopods increases with the level of hypoxia (e.g., Felder 1979).

Certain physiological adaptations have been reported in the aquatic burrowing crustaceans and fish. Greater  $O_2$  affinity and high Bohr values, favorable parameters for  $O_2$  uptake in hypoxia, have been reported (e.g., Innes 1985; Brigdes 1986; Pelster et al. 1988a,b). These features, however, do not appear to be specific to the

burrowing mode of life but rather to the habitat they occupy (Taylor and Atkinson 1991). The greatest adaptive strategy that the burrowing crabs and fish possess is their overt capacity to tolerate hypoxia (e.g., Hagerman and Uglow 1985; Swain et al. 1987) and even withstand anoxia (Hill 1981; Mukai and Koike 1984). The red band fish, *Cepola rubescens*, and *Lumpenus lampretaeformis* can maintain their normal aerobic metabolism constant down to a PO<sub>2</sub> of 7 to 9kPa (Pullin et al. 1980; Pelster et al. 1988a,b). Hypoxia causes increased opercular ventilation in the burrowing decapods and fish (Pelster 1985; Bridges 1987).

## 3.8 Living at High Altitude: Coping with Hypoxia and Hypobaria

With some exceptions, the majority of living organisms have evolved at or close to sea level. They have hence adapted to an O<sub>2</sub>-enriched environment with an ambient atmospheric pressure of 1 atmosphere (101.3 kPa; Table 12). Including other factors such as cold, rarefied atmosphere, and harmful cosmic radiation, hypoxia is the most significant factor which limits survival at high altitude. Since the demands for  $O_2$ , whether at sea level or at altitude, are essentially the same, all other factors held constant, movement to a hypoxic environment imposes great metabolic demands on an animal and calls for definite physiological adaptations for procurement of the necessary amounts of O2. Acute exposure to severe hypobaric-hypoxia equivalent to that at the top of Mt. Everest (altitude 8848m, barometric pressure 33.7 kPa), where only 30% of the initial maximum aerobic capacity (maximum  $O_2$  consumption) at sea level is lost, would be fatal to most unacclimatized human beings without an auxiliary source of O<sub>2</sub>. With adequate adjustments, however, fit human subjects performing at the limits of their aerobic capacity have reached some of the highest peaks (Dejours 1982; West 1983; Houston et al. 1987). Animals living at sea level benefit from a considerable head

T (°C)	Alt. (m)	BP (mmHg <sup>a</sup> )	K1	K2	FIO <sub>2</sub>	CIO₂ ml STPD/ml BTPS	CIO <sub>2</sub> mmol/lBPTS
41	150	747	0.787	35.13	0.2095	0.165	7.37
15	150	747	0.915	40.83	0.2095	0.192	8.55
37	0(SL)	760	0.826	36.84	0.2095	0.173	7.72
37	3400	500	0.525	23.40	0.2095	0.110	4.91
37	7200	300	0.293	0.293	0.2095	0.061	2.74

 Table 12. Oxygen concentration in inspired air in animals of various body temperatures at different barometric pressures. (After Dejours 1981)

T, body temperature; BP, barometric pressure; K1, reduction factor from values BTPS to volumes STPD; K2, reduction factor from 1 li of volume BTPS to the dry quality of substance in mMol; FIO<sub>2</sub>, fractional concentration of O<sub>2</sub> in normal dry air; CIO<sub>2</sub>, concentration of O<sub>2</sub> in inspired air; BTPS, body temperature, pressure, saturated with water; STPD, standard temperature, pressure, dry air (°C, 1 atmosphere pressure).

<sup>a</sup> To convert to kPa multiply by 0.133.

pressure of  $O_2$  in the atmospheric air ( $PO_2 = 21.2$  kPa). At the respiratory site, the pressure drives  $O_2$  in adequate amounts past the tissue barriers into the blood and finally to the mitochondria where the prevailing  $PO_2$  is between 0.1 to 0.3 kPa (Figs. 2,3). Diffusion of  $O_2$  across the blood-gas barrier is the limiting factor for survival at high altitude (West and Wagner 1980; Piiper and Scheid 1981).

About 30% of the Earth's surface lies above 1 km altitude. There are many areas with an elevation in excess of 2.5km (Webber 1979). Natural acclimation to moderate hypoxia is thus a common feature to many animals. Above an altitude of 3km, most unacclimatized subjects will show overt signs of high altitude respiratory distress such as shortness of breath and increased pulse rate (e.g., Heath and Williams 1981). Physiological disturbances such as reduced aerobic capacity (Squires and Buskirk 1982) or night eversion (McFarland and Evans 1939) may start at the much lower altitudes of 1.2 to 1.5 km. At 11 km, an auxiliary source of O<sub>2</sub> will be needed while at about 19km, without compensatory hyperventilation, little if any O<sub>2</sub> reaches the alveoli even when pure O<sub>2</sub> is breathed. This is because at that altitude the total barometric pressure of 11.6 kPa equals the sum of the partial pressure of the water vapor of 6.3 kPa (at  $37 \,^{\circ}$ C) and that of CO<sub>2</sub> of 5.3 kPa. Above this altitude, when the external vapor pressure falls below that of the body fluids at body temperature, in homeotherms, evaporation which may be accompanied by life-threatening formation of vapor bubbles (boiling) in the tissues, blood vessels, and body cavities occurs (e.g., Armstrong 1952). Just as the rarefaction of the atmosphere with altitude has a significant effect on the aerodynamics of flight, it has considerable influence on the tidal movement of air in the respiratory tract and the forces generated in the mechanical ventilation of the lung (e.g., Luft 1965). Since the resistance to turbulent flow scales with the square of the flow velocity, the effect of rarefaction of air with altitude is more predominant at high flow rates, e.g., during hyperpnea, which is one of the initial responses to high altitude hypoxia. The lungs and the chest, which function as pneumatic pumps, operate at a disadvantage at altitude while the driving force of the respiratory muscles is somewhat fixed (Johnson 1964). The overall respiratory work at altitude is, however, less than that at sea level mainly owing to the reduction of turbulent flow in favor of laminar one (Ulvedal et al. 1963). This may account for the fact that at 6.1 km, in acclimatized men, spontaneous pulmonary ventilation may exceed 2001 per minute during strenuous exercise (Pugh 1962). Though the fractional concentration of O<sub>2</sub> in the dry atmosphere stays constant at 20.93%, at least until an altitude up to 110 km, due to the compressible nature of gases, at sea level, the PO<sub>2</sub> is greater. With ascent to high elevations, the molar concentration of O<sub>2</sub> in air decreases in proportion to the decrease of PO<sub>2</sub>. Longterm and immediate human physiological adjustments at high altitude are now well known. The subject has recently been reviewed by West (1991). High altitude hypoxia does not appear to restrict the distribution of reptiles and amphibians (Hock 1964). Eleven species of frogs live between a height of 3.7 and 5.2 km. The highest living known reptile, Leiolopisma ladacense, is found at an altitude of 5.5km. Telmatobius culeus, a frog found in Lake Titicaca (altitude 3.8km) is adapted to low aquatic PO, by combining behavioral, physiological, and morphological adaptations to the cool 10 °C O2-saturated (13 kPa) water (Parker 1940; Monge and Monge 1968; Hutchison et al. 1976). Its skin is well vascularized and

folded (to increase the surface area) and the blood has small, numerous erythrocytes (the highest count among amphibians), high hemoglobin concentration, and high hematocrit. These factors give rise to a high  $O_2$  capacity. When swimming, the frog increases the convective movement of water over the skin through what has been described as bobbing behavior by Hutchison et al. (1976), a violent locomotory agitation of the water. Since the inhaled air in transit through the respiratory passages is warmed up to the body temperature and is maximally humidified, the PO<sub>2</sub> in the moisture saturated air is about 20% lower than that of the ambient dry air. The possible respiratory advantages of a lowered partial pressure of the water vapor in the ectotherms, where due to the lower body temperature the "alveolar" PO<sub>2</sub> should be higher compared with that in the endotherms, has not been fully investigated. The water vapor partial pressure is 6.2 kPa at the body temperature of 37 °C in mammals and is even higher in birds (7.3 to 8.3 kPa, Tb = 40 to 42 °C).

#### 3.8.1 Tolerance of Arterial Hypocapnia in Birds

Migrating birds have been tracked at altitudes of over 6km (e.g., Richardson 1976). The highest authenticated record of a flying bird is that of a Ruppell's griffon vulture, Gyps rueppellii, which was sucked into the engine of a jet craft at an altitude of 11.3 km over Abidjan (Côte d'Ivoire, West Africa; Laybourne 1974). At that altitude, the barometric pressure is about 24kPa (i.e., 20% of that at sea level), the PO, in the expired air is less than 5.3 kPa (closer to 2.7 kPa if hyperventilation could bring the PCO<sub>2</sub> to about 0.67 kPa) and the ambient temperature is about  $-60\,^{\circ}$ F (Torre-Bueno 1985). The capacity of birds to survive, let alone exercise, under such circumstances is unmatched among animals. Flapping flight is energetically a very expensive mode of exercise (e.g., Tucker 1972). An actively flying animal consumes O2 at two times the rate of a ground-dwelling one at maximum exercise (e.g., Thomas 1987). Hummingbirds hover with a muscle power output of nearly 100 to 120 W kg<sup>-1</sup> at a 9 to 11% mechanical energy (Wells 1993a). In the housefly, a sure sign of approaching death is that of inability to fly (Sohal and Weindruch 1996). In a rarefied atmosphere, as occurs at high altitude, for a particular speed, the cost of flight is much greater than at sea level (Torre-Bueno 1985).

A fundamental respiratory physiological difference between birds and mammals is that birds can withstand greater hypocapnia where the arterial PCO<sub>2</sub> may drop to below 0.8kPa (as the pH rises to 7.96) during panting (e.g., Faraci and Fedde 1986). Part of the respiratory efficiency and tolerance to hypoxia at altitude in birds must indirectly be related to this particular attribute which enables birds to hyperventilate and thus acquire sufficient amounts of O<sub>2</sub> without respiratory complications. A house sparrow at a simulated altitude of 6.1 km and ambient temperature of 5 °C has a respiratory frequency 38% above the sea level value and the ventilation of the parabronchi increases by 75% (Bernstein 1990). Birds can withstand the high level of arterial hypoxemia because brain blood flow is not affected by arterial hypocapnia (e.g., Faraci 1990). In fact, in species such as pigeons, the bar-headed geese, Anser indicus, and the duck, Anas platyhynchos, blood flow to the brain is not affected or actually starts to increase when the arterial blood PO<sub>2</sub> drops to below 10kPa: the flow can increase sixfold (Grubb et al. 1978; Faraci and Fedde 1986; Pavlov et al. 1987). In the human being, lowering the arterial PCO<sub>2</sub> to 1.3 kPa results in hypocapnic cerebral vasoconstriction, causing a reduction of the flow of blood to the brain by about 50% (Wollman et al. 1968). In the bar-headed goose, Anser indicus, cerebral blood flow is not affected by reduction of the arterial PCO<sub>2</sub> to 0.9 kPa (Faraci and Fedde 1986). The remarkable efficiency of the bird lung particularly during hypoxia is brought about by a multiplicity of cardiovascular and pulmonary factors; the most significant advantage is imparted by the crosscurrent arrangement between the parabronchial air and blood flows (Fedde et al. 1985; Maina 1994, 1996, 1998). At altitude flight, birds can hyperventilate without the risk of experiencing respiratory alkalosis and cerebral vasoconstriction from the resultant hypocapnia. Using measurements and estimates of man at the altitude of Mt. Everest (Dejours 1982; West 1983), Scheid (1985) calculated that if the human alveolar lung were replaced by the avian parabronchial one, for the same arterial blood gases, the person would ascend 780 m higher in altitude.

#### 3.8.2 Flying over Mt. Everest: the Bar-Headed Goose, Anser indicus

Perhaps the most astounding high altitude flight behavior in birds is that exhibited by the bar-headed goose (Anser indicus) during its annual trans-Himalayan migration from the wintering grounds of the Indian subcontinent to the breeding grounds around the large lakes in the south-central regions of Asia which are at elevations of about 5.5 km (Swan 1970). The birds take off from virtually sea level and cross the Himalayan mountains almost directly, reaching an altitude of about 10 km over the summits of Mt. Averest and Annapurna 1 (Swan 1961, 1970; Black et al. 1978; Black and Tenney 1980). The barometric pressure at these altitudes is about 31 kPa and the PO<sub>2</sub> in dry air is 6.5 kPa (West et al. 1983). Assuming that during these excursions the geese maintain a constant body temperature of 41 °C and the inhaled air is warmed to that of the body and is fully saturated with moisture, the PO<sub>2</sub> in the humid inhaled air which arrives at the gas exchange surface of the lung would barely exceed 4.9kPa. Experimentally, the bar-headed goose withstands hypoxia at a simulated altitude of 11 km (Black and Tenney 1980) and up to an altitude of 6.1 km, it maintains normal O<sub>2</sub> consumption without need to hyperventilate. On ascending to 11 km, where the concentration of  $O_2$  is only 1.4 mmoll<sup>-1</sup>, the bird takes in adequate  $O_2$  to necessitate only a minimal increase in ventilation. At 39 °C and pH 7.4, the blood of the bar-headed goose has a much higher  $O_2$  affinity ( $P_{50} = 3.9$  kPa) than that of the greylag goose (Anser anser) ( $P_{50} = 5.3 \text{ kPa}$ ), a close relative which subsists at lower altitudes (Petschow et al. 1977; Black et al. 1978).

Extreme high altitude-adapted animals show a left shift of the  $O_2$ -hemoglobin curve (Perutz 1990b; Weber et al. 1993). In *A. indicus* (Weber et al. 1993) and the Andean goose, *Cleophaga melanoptera* (Hiebl et al. 1987), where unusually high

blood  $O_2$  affinity is crucial for survival, the amino acid residues  $\alpha$ -119 and  $\beta$ -55, which form an  $\alpha_1\beta_1$  contact in human hemoglobin, are respectively altered in these two birds. The loss of contact appears to increase  $O_2$  affinity (Weber et al. 1993). In A. indicus, proline is replaced by shorter amino acid alanine (Weber et al. 1993) and in Cleophaga, leucine is replaced by short-chained serine, resulting in the loss of a single intramolecular contact (Hiebl et al. 1987). These observations suggest that adaptive changes in protein function evolved by substitution of one or a number of amino acids at specific sites, but a large proportion of such mutations are selectively neutral (Perutz 1983). The similarities in the strategies adopted by the bar-headed and the Andean geese, i.e., two single-point amino acid mutations that alter intramolecular contact enhancing hemoglobin-O2 affinity in two geographically separated species (Weber et al. 1993) is a classic case of convergent evolution at the molecular level. In the bar-headed goose, cardiopulmonary parameters indicate that muscle blood supply and O<sub>2</sub> loading from the blood capillaries rather than ventilation or pulmonary gas transfer are the limiting steps in the supply of O<sub>2</sub> to the contracting flight muscles under hypoxia (Fedde et al. 1989). This conforms with the observation made by Black and Tenney (1980) that the PO<sub>2</sub> in the arterial blood at a simulated altitude of 11.6 km is only 0.13 kPa less than that in the exhaled air. Weinstein et al. (1985) observed that "the evolution of hypoxia tolerance in birds may have developed secondary to that of the effective gas exchange and transport features needed for flight".

## 3.9 Gravity: Effects on Respiratory Form and Function

Land animals evolved from aquatic forms which subsisted in virtual weightlessness. Within historical times, humans have experienced the almost constant gravity of about 9.81 N kg<sup>-1</sup> on the Earth's surface. Weightlessness or zero gravity (0-G) presents a completely new dimension in biology. Compared with the other three forces that govern matter in the Universe, i.e., electromagnetism and the weak and strong nuclear forces, gravity is unique in many ways: it only pulls but does not push and is a relatively very weak force that has an infinite range. Thompson (1959, p 32) observed that "gravity not only controls the actions but also influences the forms of all save the least of organisms" and humorously cites "sagging wrinkles", "hanging breasts" and "many other signs of old age" as part of gravity's slow, remorseless creations. Due to its assumed invariability since the formation of the Earth (but see different views from geological, e.g., Carey 1976 and paleontological, e.g., Holden 1993, studies that argue that the parameter has varied in the past) and lack of easy means and ways of manipulating it for purposes of testing, gravity has largely been considered to be of little consequence in biology. Experimentally, increase in gravitational field has been shown to increase the metabolic rate (Smith 1976, 1978; Economos 1979; Pace and Smith 1981). Like the other features which influence aspects such as body size, shape, and locomotion, and hence prescribe the metabolic scope of an animal, gravity must have greatly influenced the definitive designs, the allometric scaling, and the structural and functional parameters in all evolved life forms. The lung and chest, which on the ground have to be elastic to be able to carry out ventilatory activity, are easily deformed by their own weight just as the gills collapse out of water. Hypogravity greatly modifies the pleural pressure, shape, and regional distribution of air and blood in the respiratory system (e.g., Engel 1991). Both the structural and functional features of the pulmonary system such as alveolar and blood capillary size and volume (Glazier et al. 1967; Hogg and Nepszy 1969; Gehr and Erni 1980), ventilatory distribution (Michels and West 1978), and blood perfusion (West 1977a), factors which cause deformation of the parenchyma (e.g., West and Mathews 1978), are variably affected by gravity. The lungs of a variety of mammals and reptiles show remarkable vertical stratification of blood flow with the "highest" regions of the lung receiving substantially less blood than the dependent ones (e.g., West 1977b; Seymour et al. 1981). In the horse and the dog, there is a fourfold difference in the volume of alveoli from the apex to the base of the lung (Glazier et al. 1967; Hogg and Nepszy 1969; Gehr and Erni 1980). Recent studies, e.g., those by Glenny and Robertson (1991b) and Hlastala et al. (1996), however, indicate that the effect of gravity on the pattern of pulmonary perfusion is less important than was earlier thought. In the dog, topographical differences in pleural pressure exist (e.g., Proctor et al. 1968) and the intrapleural pressure is more negative on the dorsal than on the ventral aspect. The dependent regions of the lung have a small resting volume but a large increase in the inspiratory volume (Milic-Emili et al. 1966). The spatial characteristics of the pulmonary arterial tree play an important part in the perfusion of the lung (Glenny 1992). In the land crab, Holthuisana transversa, stratification of the hemolymph in the branchiostegal circulation occurs, with the ventral parts containing more hemolymph than the dorsal ones (Taylor and Greenaway 1984). Though the concentration of the vascular units in the apical and basal regions of the lung is the same (McGrath and Thomson 1959), hydrostatic differences lead to a decrease of blood flow from the gravity dependent parts of the lung to the apical parts (in an upright human lung), there being a nine times difference between the two regions of the lung (West et al. 1964). Ventilation decreases from the lower to the upper lobes of the lung though the decrease is not as much as that of the blood (e.g., West and Dollery 1965). The ventilation-perfusion ratio increases five times from the "bottom" to the "top" of the upright human lung, leading to regional differences in gas exchange efficiency (West and Jones 1965; Wilson and Beck 1992). Gravity-independent ventilatory inhomogeneities are as large as gravitydependent ones (Verbanck et al. 1996). Such differences are to be expected in an organ that separates blood from air, fluids of very different densities, over a thin barrier.

Practically all the regional ventilation-perfusion inequalities which are observed at 1-G state are significantly reduced at 0-G (e.g., Michels and West 1978) or in a lung filled with and immersed in a fluid of which the specific gravity is the same as that of blood (West et al. 1965). At 0-G, increased redistribution of pulmonary perfusion to the upper parts of the lung occurs (e.g., Michels et al. 1979), increasing the diffusing capacity of the lung (Prisk et al. 1993; Vaïda et al. 1997). Acceleration (i.e., increased G), a process which is accompanied by compressive stresses, results in increased regional differences in lung expansion in the dog (Glazier et al. 1967) and in the human being (Crosfill and Widdicombe 1961).

Parameter	Ground			Inflight		
	Supine (+1 Gx)	Upright (+1 Gz)	Supine (1 Gz)	Upright sitting (+1Gz)	0G	Upright
CVP, mmHg HR, beats min <sup>-1</sup>	7.7 -	1.9 -	5.0 65	2.6 70	68 79	2.8 80

Table 13. Effects of weightlessness during parabolic flight and of +2Gz on central venous pressure and heart rate in humans. (After Norsk et al. 1987)

CVP, central venous pressure; HR, heart rate; +Gz, headward acceleration with a head to foot direction of resultant inertial force; +Gx, forward acceleration with a chest to back direction of resultant inertial force.

Redistribution of the blood and extracellular fluid due to removal of the hydrostatic pressure gradient coupled with normal tissue elasticity and muscle tone in the lower limbs is an important response to acute weightlessness which induces a significant increase in the central venous pressure (CVP) in humans compared with that in the supine and upright sitting positions at 1-G (e.g., Pendergast et al. 1987; Engel 1991; Table 13). In the human being, the venous hydrostatic indifference point is located below the heart (Blomqvist and Stone 1983). The increase in the CVP leads to congestion of the intrathoracic circulation, including that of the respiratory system due to the negative intrathoracic pressure as the blood is removed from peripheral circulation and concentrated in the central systemic circulation. At 0-G, the CVP is 0.9kPa compared with a value of 0.3kPa on the ground (1-G, sitting position) and 0.35 kPa in flight (1-G, upright sitting position) (Norsk et al. 1987). The blood volume shift decreases the functional residual capacity while leading to an increase in the thoraco-abdominal volume (e.g., Kimball et al. 1976): the changes in blood volume cause reciprocal changes in lung volume and the chest wall. Increased intrathoracic pressure may lead to congestion and subsequent engorgement of the pulmonary vasculature, a feature which, all factors being normal or nearly so, would lead to increased pulmonary diffusing capacity of  $O_2$  due to the greater capillary surface area. However, elevated pulmonary blood volume results in increased microvascular pressure which is likely to cause interstitial edema of the blood-gas barrier. This causes a reduction in the diffusing capacity of the lung especially during exercise, when pulmonary arterial and venous pressures rise in correspondence with the increase in pulmonary blood flow. Interstitial edema resulting from increased pulmonary capillary blood pressure due to redistribution of blood to the central circulation in microgravity leads to rapid shallow breathing as a result of stimulation of the Jreceptors (Engel 1991) and may be life-threatening. Excesssive transmural pressure (TMP) may lead to stress failure of the blood-gas barrier, causing disruption of capillary endothelial cells, alveolar epithelial cells, or both (Tsukimoto et al. 1991; West et al. 1991; Costello et al. 1992). The blood-gas barrier is designed to withstand high intramural stress by having a thin layer of the strong collagen-IV in the extracellular matrix (Costello et al. 1992). Stress failure of the pulmonary capillaries leads to some pathophysiological conditions such as edema, exerciseinduced pulmonary hemorrhage, and emphysema in horses (e.g., West et al. 1991). Above a TMP of 3.3 kPa, stress failure in the alveolar wall in the rabbit lung enhances exudation onto the alveolar surface (Costello et al. 1992). In the horse, the capillary transmural blood pressure which causes stress failure in the pulmonary capillaries ranges between 10 and 13 kPa (Birks et al. 1997).

# Water Breathing: the Inaugural Respiratory Process

"Of all the substances that are necessary to life as we know it on Earth, water is by far the most important, the most familiar, and the most wonderful; yet most people know very little about it." Thomson (1961)

# 4.1 The Design of the Gills

By way of the hydrologic cycle, water on Earth is believed to have remained unchanged in amount and character for about 3000 million of years (Leopold and Davis 1968). From the current concepts of paleobiology, it is popularly considered that life started in water (e.g., Thompson 1980; Selden and Edwards 1989). Currently, as many as 21 000 species of fishes (e.g., Nelson 1976; Gilbert 1993), the largest extant vertebrate taxon, live in it. More than half of the living vertebrates have arisen from evolutionary lineages which still inhabit water (Pough et al. 1989). For the first 150 to 200 million years of life on Earth, owing to the harmful effects of the UV light, life was consigned to water. The earliest complete fish fossils, members of the long extinct group named ostracoderms, date to at least 425 million years ago (Repetski 1978). As an ecosystem, water presents greater microhabitat diversity than air and land. Some extreme aquatic habitats include the hot geothermal springs at the floor of the deep oceans 3km from the surface (e.g., Childress et al. 1989) and the volcanic, hot, alkaline lakes, e.g., Lake Magadi of the Kenyan Rift Valley where the osmolarity of the water is  $600 \text{ mOsm} \text{l}^{-1}$ , pH 9.6-10.5,  $O_2$  level 2.2 mgl<sup>-1</sup> and temperature about 43 °C (Reite et al. 1974; Johansen et al. 1975). During the millions of years that they have lived in water, fish have adapted very well. Presently, they have a cosmopolitan distribution, occupying diverse ecological niches.

Compared with air breathing, water breathing is the more ancient mode of respiration (e.g., Kämpfe 1980). Pharyngeal gills are a characteristic feature of the phylum Chordata (e.g., Gutman and Bonik 1981). Except for detailed morphology, for such a heterogenous taxon, the basic structure of the gills in the agnathan and gnathostomatous fishes is strikingly similar. The gills combine simplicity of design with functional complexity. Interestingly, some species such as *Hypopomus*, a tropical swamp fish from Paraguay, and *Gillichthys mirabilis* take in air bubbles and hold them over their conventional gills (Carter and Beadle 1930, 1931; Todd and Ebeling 1966; Gans 1971). This may have been the first but futile attempt to extract  $O_2$  directly from air before respiratory concavities and pouches which subsequently evolved into lungs developed. The open aquatic environment contains respiratory gases in equilibrium with the aerosphere. Evaporative water loss as a factor in respiration does not arise. While the gills



**Fig. 46.** Gill filaments (f) of a tilapiine fish, *Oreochromis alcalicus grahami*, showing the bilaterally located secondary lamellae (s). The secondary lamellae are an efficient site of gas exchange where the blood and water flow in a countercurrent manner. *Bar* 80 µm

are highly efficient in O<sub>2</sub> uptake in water, except in a few specially adapted air-breathing fish, the lamellae, the numerous closely packed leaf-like plates which are arranged around the gill filaments (Fig. 46), after drying out in air, become impermeable to gases. Furthermore, out of water, the lamellae cohere due to surface tension and collapse under the force of gravity, in the manner of shore weeds at low tide. This reduces the respiratory surface area, creates large diffusional dead air spaces, and increases branchial vascular resistance, ultimately drastically lowering the gas exchange capacity of the gills (Burnett and McMahon 1987; Graham 1990). The animal becomes anoxic, hypercapnic, and eventually succumbs to asphyxia though exposed to a medium rich in O<sub>2</sub>. Moreover, during exposure to air, CO<sub>2</sub> levels in blood increase, precipitating a hypercapnic and subsequently an acidotic state due to the inefficiency of the gills in discharging  $CO_2$  into the air. The transfer of  $O_2$  in the gills and lungs of the coconut crab, Birgus latro, decreases 30-fold if the chitin is allowed to dry up (Harms 1932). The decline of O<sub>2</sub> consumption of the intertidal limpet, Patella granularis, when held in air is due to a functional impairment of the external pallial gills which dry up after evaporative water loss (Marshall and McQuaid 1992). When out of water, the gas exchange capacity of the gills of aquatic crustaceans is severely reduced by a factor of 3 in Callinectes sapidus (e.g., O'Mahoney and Full 1984) and up to five times in Cancer productus (DeFur and McMahon 1978). After adaptations such as greater rigidity of the gills through increased sclerotization and wider spacing of the secondary lamellae, features which minimize adhesion and collapse, Carcinus maenas can use its gills in air (Taylor and Butler 1978) and may tolerate air exposure for several days (Truchot 1975). In the terrestrial crabs, e.g., Geograpsus gravi, G. crinipes, Cardisoma hirtipes, and Gecarcoidea natalis, and in fish like mudskippers, e.g., Periophthalmus, which live in shallow mudflats (Gordon et al. 1969, 1985; Mutsaddi and Bal 1969; Tytler and Vaughan 1983; Clayton and Vaughan 1986), the gills are diversely adapted through wider gill filament spacing, gill lamellae stiffening by thicker chitinous deposition and presence of nodules which physically keep the lamellae apart (e.g., Low et al. 1990; Farrelly and Greenaway 1992) enabling the gills to be used in air (Tamura and Moriyama 1976; Tamura et al. 1976). Terrestrial vertebrates which can permanently use gills in air have not evolved and probably never will since, in addition to the gas exchange role, the gills serve other important functions such as ionic regulation and ammonia excretion (e.g., Wendelaar-Bonga and Meis 1981; Laurent and Perry 1991) which can only be effectively carried out in water. Over 90% of the NH<sub>3</sub> excreted in aquatic fish takes place across the gills (Smith 1929). The development of accessory respiratory organs was utilized to circumvent this evolutionary dead end. Some fish which are well adapted to terrestriality, e.g., the mudskipper, Periophthalmus (Tytler and Vaughan 1983), can convert ammonia to the less toxic urea (Gregory 1977) and thus extend their survival out of water. Interestingly, insectan wings are thought to have evolved from ancestral gills which were used by aquatic insects for ventilation and swimming (Marden and Kramer 1994).

# 4.2 Adaptive Diversity and Heterogeneity of Gill Form

Developmentally, gills are gas permeable evaginated outgrowths from the body of aquatic animals (Figs. 4,5). They constitute an interface between two compartments filled with aqueous solutions, an external medium (water) and internal extracellular fluid (hemolymph or blood). Gills occur in different sizes, forms, and locations. They range in complexity from the simple tegmental evaginations like the tube feet and pupullae of some classes of echinoderms which provide only a minor supplement to gas transfer (much of which takes place across the skin), the external gills of the annelid polychaetes, some molluskan nudibranchs, larvae of teleosts (e.g., Gymnarchus), and the tadpoles to the more elaborate multifunctional internal ones contained in the branchial chambers, e.g., in crustaceans and teleosts or accommodated in a mantle cavity, e.g., in mollusks (e.g., Laurent 1982; Hughes 1984). The fragile and yet elegant structure of the gills (Ojha and Singh 1986; Olson 1996) has fascinated biologists for a long time. They constitute a paradigm of an efficient external (evaginated) aquatic gas exchanger and provide a fundamental model for the study of the manifold respiratory processes and mechanisms adopted by the higher vertebrates. Gills carry out diverse seemingly unrelated functions which include respiration, osmoregulation, acid-based balance, ammonia excretion, regulation of circulating hormones like catecholamines and angiotensin, and detoxification of plasma-borne harmful substances (e.g., Neckvasil and Olson 1986), locomotion (e.g., Septibranchs), and feeding. The definitive design of the gills must be a compromise between all the functional requirements (McDonald et al. 1991). The trophic (filter-feeding) role of the gills is exhibited by the simple chordates in the subphyla Tunicata and Cephalochordata, e.g., Amphioxus and Petromyzon (e.g., Baskin and Detmers 1976; Youlson and Freeman 1976) and actinopterygian fish, Polypterus (Hughes 1980), bivalve mollusks, in Ascidia, in ammocoete larva of lampreys (Youlson and Freeman 1976), and in embryos and larvae of amphibians (Billet and Courtenay 1973). Divergent views as to whether the gills initially evolved as trophic or respiratory organs persist (e.g., Willmer 1970; Gutman and Bonik 1981; Hickman 1984). In the lamellibranch mollusks, the gills have shifted from the primary respiratory function to filter feeding (Hazelhoff 1930; Jorgensen 1952). In those animals where the gills serve a dual function of respiration and filter feeding, ventilation is generated by cilia. Movement of the water in contact with the surface of the gills is an important part of the gas exchanging process and may occur in simple animals in form of ciliary action, e.g., on the branchial crown of sabellids, on the parapodial gills of polychaetes, e.g., Nephtys, and in the ctenidia of aquatic mollusks and ascidians. While enhancing procurement of food, the fast flow of water necessary for food uptake reduces the O<sub>2</sub> extraction capacity due to the short transbranchial transit time of the water (Table 8). Larval caecilians use gills for respiration in the egg but are lost soon after hatching (Welsch 1981). Some invertebrate organisms such as the terebellid worms (e.g., Weber 1978) present some localized areas of the body where the thickness is drastically reduced, gas exchange occurring to the same degree of efficiency as in the gills proper.

Vertebrate gills are categorized into external gills, i.e., those which dangle freely into the water, e.g., in the larval lungfishes and many amphibians and internal ones, i.e., those which are covered by various forms of cutaneous modifications, e.g., the opercular flap of the teleosts or are contained deep in the mesenchymal tissue mass, e.g., in sharks and the cyclostomes (e.g., Emery and Szczepanski 1986). Except for the cuticular gills of some insect larvae (Wigglesworth 1950), the external gills are highly susceptible to physical and chemical damage. Furthermore, they physically restrict locomotion in water. Typically, teleost fish have four pairs of gill arches. Gills have to be selectively permeable and well tuned to control and regulate transepithelial flux of ions and water between the internal and external milleu to curtail excessive loss or overload (Kirschner 1982). A conflict in the design of the gills is brought about by the need to optimize gas exchange by fully exposing extremely delicate organs to the elements. Any protective cover in form of a corneum cuticle or mucus would drastically limit the permeability and hence the transfer, especially of gases across the gills. In many animals with gills, however, the organs are housed in enclosures such as a mantle cavity, a shell (e.g., mollusks), outgrowths of the carapace (e.g. arthropods) and crustaceans, or are contained within a branchial cavity enclosed by the integument, e.g., in teleost fishes. While providing protection to the internal gills, this modification impedes water flow over the gills. This limitation was circumvented by evolution of an energetically expensive buccal force pump for moving a viscous O<sub>2</sub>-deficient medium, through narrow spaces. In the larval amphibians, external and/or internal gills together with the general body surface

play a major role in respiratory gas exchange. Except in two groups of animals, holothurians and cephalopods, aquatic respiratory ventilation is largely circulatory rather than tidal (Figs. 6,18). However, in sea cucumbers, the cloaca pumps water tidally whereas in the octopus and squid, the mantle serves as both a locomotory device and a respiratory one by pumping water into and from the mantle cavity. Tidal breathing of water across the anus occurs among annelids (e.g., Urechis caupo) and insects (e.g., dragon fly nymph, Aeshna; Fig. 23), via the branchial pores in lamprey eels and via the mouth in some amphibians and reptiles, e.g., the soft-shelled turtle, Amyda (McCutcheon 1954). The energetic cost of oscillatory ventilation in water explains why tidal ventilation rarely evolved in the active water breathers where throughflow designs predominate. Contraction of the gills, ciliary movements and mechanical ventilation (e.g., in the decapods and fish), placing the body in a moving current of water or active body movement maintain a PO<sub>2</sub> gradient across the respiratory surface. Fish which move constantly like the mackerel and the tunas ventilate their gills passively simply by keeping their mouths open. They have lost the power to mechanically pump water across the gills and hence must swim continuously. To avert physical damage during the fast forward movements, the gill filaments of such fish are fused to each other (Muir and Kendall 1968). By hitching a ride on a shark, the gills of Remora remora are passively ventilated but at rest, the fish utilizes active respiratory movements (Muir and Buckley 1967). In the siren, Necturus maculosus, the muscles associated with the gills are well developed and play an efficient respiratory role. Waving the external gills (in water) generates convective movement of the water. The process is stimulated both by increased water temperature and reduced O<sub>2</sub> content (Guimond and Hutchison 1972). The development of the external gills of larval amphibia depends on O<sub>2</sub> tensions in water (e.g., Foxon 1964). When placed in hypoxic water, the gills of Rana temporaria hypertrophy while in well-oxygenated water they atrophy. The external gills of the larvae of the salamanders which live in well-oxygenated waters are less well-developed, i.e., the respiratory surface area is small and the water-blood barrier is thicker than in those which live in O<sub>2</sub>-poor waters (Dratisch 1925; Bond 1960). In fish living in water with a PO<sub>2</sub> of 10.7 kPa, the gills are much larger than in those at 100 kPa (Dratisch 1925). In Xyelacyba myersi, a fish which lives in the deep water of the Pacific Ocean (at depths of 1400 m), the gills are poorly developed (Hughes and lwai 1978). Unlike in some fish, amphibian gills are not specialized for exchange of gases in air. They never constitute the exclusive respiratory organs at any one stage of development. The contribution of the gills to the overall respiratory process in the amphibians differs from within and between species, depending on the habitat occupied and the level of development (e.g., Shield and Bently 1973a, b; Burggren et al. 1983; Malvin 1989). In the early stages of development, e.g., in the neotenic urodele larvae, the skin, lungs, and buccopharyngeal cavity function as adjunct respiratory organs. When exposed to low  $O_2$  tensions (2 mll<sup>-1</sup>), ventilation in *Nereis virens* may become continuous and at very low values (0.6 ml per l) stop altogether (Lindroth 1938a). In the octopus, Octopus vulgaris, the O<sub>2</sub> extraction factor may be as high as 80% (Winterstein 1925), with the ventilatory rate of the gills increasing ten times above normal during hypoxia.

# 4.3 The Functional Innovations of the Gills for Aquatic Respiration

In all evolved gas exchangers, the structural adaptations which favor  $O_2$  transfer are a thin barrier between the respiratory media and extensive surface area. In many aquatic animals, these features engender new problems of maintaining ionic and osmotic steady states in the body fluids. Opercular breathing movements in fish have been reported to occur before hatching. They appear to assist in the hatching process itself (S. Smith 1957). Except for the external gills of the neotenic urodeles, the amphibian gills (Fig. 47) can be considered to be transient or disposable gas exchange organs. Vascular shunt (anastomotic) vessels between the afferent (dorsal segment of the aortic arch) and efferent (ventral segment of



**Fig. 47A–C.** Respiratory organs of amphibians at different stages of development. A External gills of the larvae enlarged in the inset. **B** Internal gills of a bullfrog tadpole. **C** Lungs of an adult frog showing the large peripherally located air cells. (Jessop 1995)

the aortic arch) gill arteries which bypass the external gills occur in both *Rana temporaria* and *Bufo bufo* (De Saint-Aubain and Wingstrand 1981) and in the gills of the urodele amphibians, e.g., *Amblystoma tigranum* (Baker 1949). The shunts are presumed to sustain circulation after metamorphosis when the external gills atrophy on changeover to pulmonary respiration.

Although aquatic animals procure O<sub>2</sub> through the gills, the majority of them take up a substantial fraction across the skin and lose a significant amount of CO<sub>2</sub> through it. In normoxic diving, the soft-shelled turtles, Trionyx spiniferus asperus, and musk turtles, Stemotherus odoratus, can remain submerged under water for 100 days and maintain normal acid-base status (Belkin 1968). Although the skin of fish has been considered to be impermeable to gases, measurements made on the eel, trout, and tench indicate that substantial uptake of  $O_2$  occurs transcutaneously (Kirsch and Nonnotte 1977). Much of the  $O_2$  is, however, used by the skin itself. The partial pressure of  $CO_2$  in the blood of the lungfish, Protopterus declines after vasodilation of the cutaneous vasculature owing to enhanced loss of CO<sub>2</sub> (DeLaney et al. 1974). Nonpulmonary CO<sub>2</sub> elimination is substantial in many aquatic reptiles but only a small fraction of the total O<sub>2</sub> need is acquired in that way. Of the total CO<sub>2</sub> output in the aquatic turtle, Trionyx mucita (Jackson et al. 1976), 65% and as much as 94% in the sea snake, Pelamis platurus (Graham 1974), is lost cutaneously. In the aquatic snake, Acrochordus javanicus, 8% of the total O2 uptake and as much as 33% of CO2 excretion occurs through the skin (Standaert and Johansen 1974). Invertebrates such as the oligochaetes, platyhelminthes, mollusks such as Scaphoda, some Aplacophora, and some Gastropoda which have no distinct respiratory organs rely entirely on the skin for O<sub>2</sub> uptake. Such animals have a large surface-to-volume ratio which allows enough O<sub>2</sub> to be taken up by diffusion. Blood capillaries arranged in form of loops are present in the outer (circular) muscle layers in the gill-less maldanids (Pilgrim 1966), terebellid worms (e.g., Weber 1978), and on the parapodia of the annelid sandworm, Nereis succinea (Mangum 1982a). In such areas, the diffusional distance is thin enough to facilitate satisfactory transcutaneous movement of the respiratory gases. In N. succinea, the diffusional distance from the medium to the blood ranges from only 1.5 to 2.3 µm (Mangum 1982a). The naked gastropods of the genera Lymnea and Helicostoma can stay under water for a long period acquiring O<sub>2</sub> through the skin (Cheatum 1934). In the limpet, Siphonaria zelandica, 25% of gaseous exchange occurs across the side of the foot (Innes et al. 1984). Some salamanders such as Desmognathus quadramaculatus (Gatz et al. 1974) rely entirely on the skin for gas exchange. Oxygenation of the tissues which lie beyond the limiting diffusional distance must be effected by the coelomic fluid though in most species the fluid has a very low O2 carrying capacity. In Arenicola cristata, at the microenvironmental  $PO_2$ , the  $PO_2$  in the coelomic fluid is less than 6.7 kPa (Mangum 1982a). The metabolic rates of the deeply located and poorly vascularized tissues is relatively low and is finely tuned to O<sub>2</sub> availability. Respiration in Arenicola is intermittent with pauses of about 20 min. During the activity periods which last for about 10 min (at 20 °C), about 90 ml of water is passed across the body with an O<sub>2</sub> extraction factor of 50% (van Dam 1935, 1938). In amphibians, those species with smooth, poorly vascularized internal surface have a thinner epidermis and a much denser capillary network on the skin than those with well-developed lungs (Foxon 1964; Czopek 1965). The relative length of the cutaneous capillaries expressed as percentage of the total length of respiratory surface-associated capillaries is about 50 to 80% (Czopek 1965).

# 4.4 The Simple Gills

# 4.4.1 Morphological Characteristics

The general features of both the simple and the complex gills have been described by Kennedy (1979). Not all gills or gill modifications play a significant role in gas transfer (e.g., Fox 1921; Thorpe 1930). Certain hair-like or plate-like external outgrowths from the body which have been called gills play no determinate respiratory role. Such forms are found externally in echinoderms, mollusks (e.g., Nudibranchia), arthropods (e.g., Brachiopoda) (Kikuchi 1992), fishes (e.g., adult lungfish, Lepidosiren), blood gills of Chironomous, and in some amphibians (e.g., adult salamanders). In the larval stages, such gills may function as appendages and/or filter feeders with respiration being a secondary process. The freshwater brachiopod, Branchinella kugenumaensis, has ten pairs of gills (metepipodite segments of thoracic legs) and the neck organ (located on the cephalothorax) is an important respiratory organ (Kikuchi 1992). The anal gills of Diptera larvae are largely salt-absorbing organs (Koch 1938). Among the Cirripedia, the extensions attached to the cirri in the four genera of Lepadidae, though described as gills by Darwin (1851), appear to play no role in respiration. This was, however, refuted by Kaestner (1970) and Burnett (1972): a large number of species in the group flourish without them. On the other hand, some structures generally considered to be feeding organs such as the tentacles of the sea anemones may play a notable role as gas exchangers (Sassaman and Mangum 1973; Mangum 1994). The so-called ventral gills or blood gills of the aquatic Chironomid larvae take up a smaller quantity of  $O_2$  from the water than the skin (Harnisch 1937). The annelids are among the simplest animals which have an organ morphologically recognizable as a gill. In the ancestral members of this taxon, the gill is a simple, smooth tubular evagination of the body wall (e.g., Nakao 1974; Mangum 1976a) where, e.g., in Glycera and in Nereis succinea the water outside and in the coelomic fluid is kept in motion by cilia, a closed circulatory system lacking. In others such as the oligochaete, Alma nilotica, the comparably simple gills (Gresson 1927; Khalaf EI Duweini 1957) are perfused through a closed circulatory system (Stephenson 1930). The parapodia in many polychaetes, e.g., Nereis, Arenicola, and Dasybranchus, are well vascularized and serve as gills (Nicoll 1954). Relatively simple gills which are located inside the branchial skeleton rather than outside (Randall 1972), as is the case in the gnathostomatous fishes, are found in the lower animal forms, e.g., the Cyclostomata, i.e., the hagfish (Jensen 1966; Bardack 1991) and the lampreys (Youlson and Freeman 1976; Lewis and Potter 1982; Mallatt and Paulsen 1986). The larval forms of the viviparous aquatic caecilian, Typhlonectes, exchanges gases by apposition of the fetal gills to the highly vascularized wall of the oviduct (Wake 1977): the gills are absorbed before birth.

## 4.4.2 Ventilation and Functional Capacities

The advantage of having respiratory organs on locomotory parts of an aquatic animal, e.g., in some crabs (Fig. 48) is great (Maitland 1986). Mechanical displacements of the appendages confer a ventilatory advantage. This may be of particular importance especially in standing hypoxic water. The development of gill modifications on the locomotor parapodia in polychaetes (e.g., Arenicola and Dasybranchus) guarantees efficient ventilation. The primary role of respiratory pumping in the marine snail, Aplysia californica, has been presumed to be that of enhancing O<sub>2</sub> uptake into the hemolymph through the gill epithelium (Kanz and Quast 1992). A negative correlation between respiratory pumping and O<sub>2</sub> consumption under hypoxic conditions has, however, been reported by Levy et al. (1989) in related species, i.e., A. depilans and A. fasciata. Gills without any specific ventilatory adjuncts are encountered in echinoderms where cilia produce fluid movement, e.g., in mollusks (Nudibranchia), arthropods (Brachiopoda and insect larvae), fishes (e.g., adult lungfish Lepidosiren - in the male during breeding and in larvae of teleosts and elasmobranchs), and amphibians (e.g., mud puppy, Necturus, and a number of other salamander adults and in numerous larval and tadpole stages of frogs). In Necturus maculosus, when the animal is stationary,



Fig. 48. Sand-bubbler crab, *Scopimera inflata*, has pairs of membranous disks (gas windows) in each leg meral segment,  $\star$ , which are utilized for gas exchange during low tide. The gas windows are mechanically ventilated during locomotion, enhancing the gas exchange process. (Maitland 1986; reprinted by permission from *Nature*, vol. 319, pp. 493–495; copyright 1986; Macmillan Magazines Ltd.)

muscles at the base move the gills especially when the animal is at rest (Guimond and Hutchison 1972) but many other urodeles, though possessing similar gills, are not known to execute such maneuvers. The gills hang out in water, where they are passively ventilated by the physical movement of the animal in water or when the gills are placed in a water current. Agitation of the immediate (boundary) water layer over the gills or the skin renews the water, thereupon increasing the PO<sub>2</sub> (Feder and Burggren 1985a, b; Feder and Pinder 1988). Consequently, diffusion of O<sub>2</sub> across the tissue barrier is enhanced. Some species, e.g., Dasybranchus, exhibit sporadic gill contractions presumably effected by movements of the coelomic fluid (Mangum et al. 1975). Ventilatory activities can be effected by ciliary movements (e.g., in nudibranchs), by physical movement (e.g., in Phyllopoda), muscular contractions of the branchial outgrowths (e.g., in Amphibia and Ephemend larvae) and in others, e.g., crustaceans, by the beating of special appendages (e.g., pleopods) which maintain a water current across the gills. The high shore littoral crustacean, Ligia, obtains about 50% of its O<sub>2</sub> needs through the pleopods and the rest through the ventral surface of the abdomen (Edney and Spencer 1955). Terrestrial woodlice breath through the pleopods, with Porcellio scaber obtaining as much as 70% of their O<sub>2</sub> needs through them. The book gills of the chelicerate arthropod, Limulus polyphemus, are borne on five pairs of modified appendages which are located on the ventral surface of the opisthosoma (Mangum 1982a). Each modified appendage consists of about 150 rounded lamellae which in a 1-kg specimen are 3 to 4 cm wide. The thickness of the water-blood barrier is on average 5.6µm thick. When Limulus is in hypoxic water, the frequency of the ventilatory movements of the opisthosomal appendages increases. On exposure to extremely hypoxic or hypercarbic water, the totally aquatic and solely cutaneous breathing amphibians, Cryptobranchus alleganiensis and Telmatobius culeus, rock or sway their bodies back and forth in water (Hutchison et al. 1976; Boutilier and Toews 1981), passively ventilating their skin, In *Necturus*, at 25 °C, through agitation of the gills in water, 60% of the O<sub>2</sub> needs can be transferred across the gills (Guimond and Hutchison 1976) while at the same temperature, in the inactive gills of Siren lacertina, the gills meet less than 5% of the O<sub>2</sub> needs (Guimond and Hutchison 1973a, 1976). Necturus is found in diverse habitats which range from clear waters of lakes, streams, weed-choked canals, and drainage ditches (Bishop 1943). Adaptive features such as physical movement of gills, capacity to undergo long periods of anaerobiosis, and lowering metabolic rate some 30 to 40% below normal are some of the adaptive features which enable animals to withstand hypoxia.

### 4.4.3 Gas Exchange Pathways and Mechanisms

Amphibians possess gills during their larval stages of development. Caudata and caecilians (Gymnophiona = Apoda), have external gills, while in Silentia they are internal. In the neotenic forms, e.g., *Necturus* and *Ambystoma*, the gills persist throughout life. Many aquatic salamanders utilize a trimodal gas exchange strategy in which pulmonary, branchial, and cutaneous surfaces variably contribute to

gas exchange. In the Siren, of the total amount of  $O_2$  taken up by the animal (at 25 °C), the gills account for only 2.5% while they eliminate 12% of the total  $CO_2$ (Guimond and Hutchison 1972). A 42% reduction in the total gas exchange capacity occurs on ligation of the gills of Necturus at 22 °C (Shield and Bentley 1973a,b). The external gills contribute as much as 54% of the total  $O_2$  need and void 61% of the total  $CO_2$  at 5 °C, with the  $O_2$  consumption through the gills increasing to 60% at 25 °C (Guimond and Hutchison 1972). Necturus quickly lowers the aquatic PO<sub>2</sub> from 20 to 5.3 kPa (Guimond and Hutchison 1972). An arterial PO<sub>2</sub> of about 4.7 kPa was reported by Lenfant and Johansen (1967) in the blood of Necturus, a value indicative of a high efficiency of the gills in O<sub>2</sub> extraction at very low ambient PO<sub>2</sub>. Fanelli and Goldstein (1964), however, observed that the gills of Necturus are of no consequence in respiration. This may, nevertheless, only be true in an inactive animal at low ambient temperatures in O2 saturated water. In cool (5 to 15 °C) aerated water (PO<sub>2</sub> = 17 to 20 kPa), in undisturbed animals, the gills do not move and are retracted and held to the side of the head. However, as temperature increases, the animal becomes restless and the gills oscillate (Guimond and Hutchison 1976). When removed from water, at 20 °C, the gills of Necturus collapse (Lenfant and Johansen 1967). This is followed by a dramatic rise in the arterial  $PCO_2$  and a marked reduction in arterial  $PO_2$  in spite of evident pulmonary ventilatory and gulping movements. The gills of Siren are less involved in respiration than those of *Necturus* and are notwithstanding more efficient in  $CO_2$  elimination than  $O_2$  uptake (Shield and Bentley 1973a,b; Bentley and Shield 1973). Features such as a thick branchial epithelium (e.g., Cope 1885), abundant arterial venous shunts which bypass the gill filaments (e.g., Darnell 1949), small size, structural simplicity, and immobility of the gills (Guimond and Hutchison 1976) explain the respiratory inefficiency of the gills of the Siren. The branchial beating frequency in Necturus depends on factors such as metabolic rate and environmental hypoxia or hypercapnia. The rate increases from about 10 times per min to more than 50 times per min (with temperature increasing from 10 to 25 °C (Guimond and Hutchison 1972, 1973a). In insect and amphibian larvae, extended hypoxia induces gill growth and cutaneous vascularization (Bond 1960). The role of the external gills of the larval anuran amphibians in gas exchange is not known (Boutilier 1990) but the internal ones of the lungless tadpoles of Rana catesbeiana and R. berlandieri account for as much as 40% of the total O2 and CO2 exchange at 20 °C (e.g., West and Burggren 1982). In some anuran species, ventilatory rates as high as 90 times per min at 20 °C have been reported by Burggren and West (1982) and in some, ventilatory frequency appears to be synchronized with the heart rate (Wasserzug et al. 1981).

In the different amphibian species, the levels of development of the external gills correspond with the availability of  $O_2$  in the environment in which they subsist and the functional needs (Noble 1931). During the development of the lungs, the unidirectional ventilation of the internal gills with water through bucco-pharyngeal muscular activity decreases by as much as 50% as the skin assumes a prominent respiratory role (Burggren and West 1982). *Siren*, which is endowed with capacity for efficient cutaneous and pulmonary respiration, has moderately developed gills while *Necturus*, which relies on pulmonary exchange,

has more elaborate gills to supplement cutaneous gas exchange (Guimond and Hutchison 1976). Gills of *Necturus* kept in cool well-oxygenated water atrophy compared with those kept in warmer nonaerated water (Guimond and Hutchison 1976). After being forcefully held under water for over 2 weeks, the gills of Necturus enlarge. The lugworm, Arenicola cristata which lives in burrows, a fairly anoxic habitat, has 11 pairs of gills which are located in the midregion of the body. The main trunks of the gills branch four times, the terminal branches which account for much of the respiratory surface being 0.6mm long and 0.07 to 0.10 mm in diameter while the diffusional distance is 2 to 4 um thick (Mangum 1982a). Features such as high hemoglobin-O<sub>2</sub> affinity and great cooperativity of O<sub>2</sub> binding (Mangum 1976b) may enable such species to subsist in a hypoxic environment. In the polychaete families (e.g., Wells et al. 1980) and in the filamentous gills of other taxa such as crustaceans (McLaughlin 1983), the gills are organized in the same general pattern as in Arenicola. The gills of the members of the family Sabellidae, which inhabit vertical tubes either within soft sediment or, as in the case of Eudistylia vancouver, attached to firm substratum, are considered to be more advanced (in some respects) than those of the lugworms (Mangum 1982a). The gill, a pinnate structure known as a branchial crown, is not metamerically arranged but is confined to the anterior end where the respiratory gases can be easily exchanged with water: the gill serves as a filter-feeding organ while the organism is still confined to the tube. In E. vancouveri, the gill consists of two principal trunks or branchioles which divide at the first level to give rise to numerous long ciliated pinnae giving rise to a double row of filaments called pinnules at the second level of branching (Vogel 1980). A single branchial crown consists of about 54000 pinnules which provide a surface area accounting for more than 70% of the surface area of the body. This translates into a mass-specific surface area of  $30 \text{ cm}^2 \text{l}^{-1}$  (wet wt.). Individual branchioles are associated with a single blood vessel which divides in the same bifurcating plan, terminating blindly at the pinnules. Owing mainly to the absence of the afferent and efferent blood vessels, the blood flow pattern in the branchioles is believed to be tidal in nature (Fox 1938; Ewer 1941). In the cephalochordates and tunicates, the blood flow in the gills is irregular. In the tunicates, the flow reverses in direction (Remane et al. 1980). In both groups, hemoglobin is lacking but the tunicates have a vanadium-based pigment which may carry O2. The respiratory differences between the sabellids and the lugworms include: (1) the ventilatory rate in the sabellids is three times greater than in arenicolids (Dales 1961; Mangum 1976b), (2) the O<sub>2</sub> extraction factor from water is only 6 to 10% in sabellids and 30 to 60% in arenicolids (Dales 1961; Mangum 1976b), and (3) the affinity of the sabellid respiratory pigment (chlorocruorin) is generally low, ranging from 5.9 kPa in Myxicola (Wells and Dales 1975) to 6.9 kPa in Spirographis (Antonini et al. 1962) at a pH of 7.4 and a temperature of 15 °C. Of all respiratory organs which have been studied, in E. vancouveri, blood may pass in and out of the gill a number of times before reaching the ventral blood vessel which distributes it to the body tissues (Ewer 1941). The sebellid gills present a unique model of a microcirculatory configuration in the animal kingdom in which multiple cycling of the blood occurs (within a gas exchanger) to maximize O2 uptake by the blood.

In modern tadpole larvae of Anura, internal gills (Fig. 47) from rows of branched lamellae supported by gill bars which are separated by four gill slits (e.g., Uchiyama et al. 1990). With the onset of pulmonary respiration, the gill slits close up except in some Urodela which have readapted to aquatic life where some slits remain open throughout life. External gills are rare in adult fish but play an important respiratory role in amphibian larval forms of the tadpoles and neotenic larvae (e.g., in Amblyostoma - Axolotl), in Discoglossus, and in water-breathing adults (e.g., Necturus maculatus). In the larval forms of elasmobranchs and some larvae of Chondrostei and Teleosti, external gills form as long filaments floating in the albuminous fluid within the egg case. True external gills occur in the larval forms of fishes, e.g., Polypteridae and Dipnoi, and in amphibians in form of threads or fine feathers (Dunel-Erb and Laurent 1980b). In the less advanced invertebrate gill-breathing life forms, the circulatory system is not very well connected to the branchial respiration: the gills are located on the venous side of the heart which receives arterial blood. In most Gastropoda, Cephalopoda, advanced Crustacea, and fishes, the venous blood is collected and passed to the gills and then distributed to the rest of the body. The amphibian external gills differ remarkably from those of fish in that macroscopically they form arborescent organs and are not arranged in a refined hierarchical order.

In many lower forms of animals, the regulation of branchial respiration is absent or very poorly developed. In Ligia oceanica, the beating of the pleopods, which produce water currents across the gills, does not change with O<sub>2</sub> tension in water except at very high or low concentrations. In Gammarus locusta, the pleopods stop beating at a concentration of O<sub>2</sub> above 5.6 ml per l while a drop causes rapid beating (Fox 1921). In the aquatic insect larvae, Cryptochaetum iceryae (Diptera, Agromyzidae) and Icerya purchasi (Coccidae, Monophlebini), Thorpe (1932) observed that CO<sub>2</sub> efflux occurred at specific areas of the body. In the fish larvae, before the gills develop, the body surface serves as the only gas exchange pathway. To maintain a high PO<sub>2</sub> in the layer of water next to the skin, the larvae cause convective movement of the surrounding water either by positioning themselves in moving water currents or by executing physical movements (Liem 1981). The external gills of newly hatched larvae of Protopterus have cilia which move water across the gills before the ventilatory muscles develop (Whiting and Bone 1980). During the postlarval development, in *Protopterus ampibius*, the fraction of the total  $O_2$  which is acquired through the external gills and the skin declines with the development of the lung (Johansen et al. 1976). In the bimodally breathing teleost Monopterus, the large muscular fins generate a posteriorly directed water current which interacts with a well-vascularized region below the epithelial surface of the fins and the yolk, creating a highly efficient countercurrent gas exchange system (Liem 1981). This adaptive property may enable the fish to subsist in hypoxic water. The tuft-like structure on the pelvic fins of the male Lepidosiren paradoxa which is highly vascularized has been considered to be important in the parental rearing of the eggs and larval forms in burrows constructed for that purpose (Kerr 1898; Agar 1908): the gill-like organ has been assumed to be involved in O<sub>2</sub> uptake from water. This prospect may be supported by the fact that during the husbandry period, the fish does not surface as frequently as it normally does to exchange gases in air. It is thought that the pelvic fin respiratory elaboration may be utilized for transferring  $O_2$  from the blood of the male fish to the immediate vicinity of the eggs and the developing larvae in the frequently hypoxic tropical swamps in which the fish lives (Kerr 1898; Cunningham and Reid 1932). On hatching, the young larvae of *Lepidosiren* respire through the external gills which start to atrophy after 45 days of life (Carter and Beadle 1930; Krogh 1941). At about the same time, the pelvic gills of the male lungfish begin to regress as the larvae surface to breath air (Agar 1908).

# 4.5 The Complex Gills

## 4.5.1 Structure and Architectural Plans

The structure and arrangement of the internal gills varies remarkably. The most complex ones are found in the bony fish (class: Pisces) which typically have four pairs of gill arches. In fish, a single ventricle receives venous blood that is subsequently pumped to the gills for the uptake of O<sub>2</sub>. The gills of the elasmobranchs lack a skeletal support. In adult cyclostomes, the design of the gills fundamentally differs from that of fish: the main skeletal mass lies external to the gill epithelium (Jarvik 1968; Youlson and Freeman 1976; Lewis 1980; Lewis and Potter 1982). In the hagfish, coronary arteries are lacking: the heart, which is thin-walled, receives nourishment from the venous blood it pumps. Gills are generally poorly developed in air-breathing teleosts (e.g., Munshi 1976), mudskippers (e.g., Low et al. 1988, 1990), and the lungfishes, Dipnoi (Laurent 1982). The bimodal breathing fish face the singular problem of losing  $O_2$  (acquired during air breathing) to the surrounding hypoxic water as the blood traverses the gills. Shunting of the blood away from the gills (Fig. 21) and reduction in the gill respiratory surface area avert such losses. In the most terrestrial of these fish, the gills are retained mainly for elimination of CO<sub>2</sub> which is more readily discharged into the water. The gills of the lungfishes are remarkably different from those of other classes of fish (Laurent 1982, 1996). They do not form regular arrays of primary and secondary lamellae but look like the external arborescent gills of a tadpole rather than the gills of Teleosti and Chondrichthyes (Laurent et al. 1978). Moreover, pillar cells (Fig. 49) are lacking in the dipnoan gills (Laurent et al. 1978; Laurent 1982). The gills of the hemoglobinless Antarctic ice fish, Chaenocephalus aceratus and Chamsocecephalus esox (Steen and Berg 1966), and Chaenichthys rugosus (Jakubowski et al. 1969), much as they are structurally similar to those of other fish which have hemoglobin, have fewer secondary lamellae and the skin is very well vascularized. Fast-swimming fish, e.g., the tuna, show certain modifications of the gills which include presence of calcified flattened gill rays which offer the filaments better physical support (Iwai and Nakamura 1964) and fusion of the gill filaments to the lamellae, preventing lamellar deformation during high speed cruising especially during passive or ram ventilation (Muir and Kendall 1968). The bowfin, Amia calva, a freshwater fish which differs remarkably in habit and habitat from the marine fish, has independently arrived at lamelar fusion (Bevelander 1934). It has been speculated that the attribute supports gas exchange



**Fig. 49.** Gills of a tilapiine fish, *Oreochromis alcalicus grahami* showing a gill filament, *g*, and secondary lamellae separated by an interlamellar space, *x*; *p* pillar cells; *c* vascular channels; *e* erythrocytes; *w* white blood cell;  $\succ$  intercellular junction of epithelial cells. Note the extremely thin water-blood barrier over the vascular channels. *Bar* 17 µm. (Maina 1990)

in the  $O_2$ -deficient turbid water and prevents gill collapse during air breathing. Cutaneous gas exchange contributes about 40% of the total  $O_2$  uptake in these fish (Hemmingsen and Douglas 1970). The high efficiency of the skin in  $O_2$ transfer compared with that of the gills may be a means of economizing on respiratory work through conserving the activities of the bucco-pharyngeal pump. In the cold polar water where  $O_2$  levels are normally high, to increase gas exchange, a fish only needs to physically move to sufficiently "ventilate" the skin. The resting  $O_2$  consumption of the ice fish is one half to one third that of hemoglobin-carrying fish of similar size found in the same habitat (Hemmingsen and Douglas 1970). Further compensations for the low  $O_2$  capacity of the ice fish include a high blood volume (7.5% of the body weight) compared with 2 to 3% in normal fish and a reduced viscosity of blood due to absence of erythrocytes, factors which enhance  $O_2$  transport by blood. In fish, during growth, the number of gill filaments increases more rapidly than that of the secondary lamellae (Hughes 1982).

The filament is the functional unit of a branchial arch system (e.g., Olson 1996). Gas exchange, however, occurs across the secondary lamellae which are thin semicircular flaps which are bilaterally set on the filament perpendicular to its longitudinal axis (Figs. 11,46). An elaborate epithelium (the primary epithelium) covers the gill filament while a less complex one, the secondary epithelium, lies over the secondary lamellae (Laurent and Dunel-Erb 1980; Laurent 1984; Maina 1990a, 1991). Three types of cells, the pavement cells, the chloride (= mitochondria-rich = ionocytes) cells, and the mucous cells are most prevalent on both types of epithelia. The different functions of the gills, i.e., osmoregulation (e.g., Leatherland et al. 1974; Eddy et al. 1981), acid-base balance (e.g., Heisler 1984), elimination of products of nitrogen metabolism (Goldstein 1982; Randall et al. 1989), and respiration appear to occur at specific sites of this highly wellorganized and differentiated epithelium. The first two processes take place in the composite primary epithelium while the last two occur in the thin, less elaborate secondary epithelium. Like the vertebrate lung (Sect. 6.10.2), the gills perform certain metabolic functions whereby they modify the plasma hormones in the arterial blood before they pass to the systemic circuit (Neckvasil and Olson 1986; Olson et al. 1986a). The pavement cells are simple squamous broad cells which are characterized by surface microridges (e.g., Olson and Fromm 1973; Kendall and Dale 1979; Hughes 1979; Hughes and Umezawa 1983; Hossler et al. 1986): the patterns of the microridges differ between species. Lamellar epithelia of pelagic fish such as bluefish, Pomatomus saltatrix, Atlantic mackerel, Scomber scombrus, and Atlantic bonito, Sarda sarda (Olson 1996) and the hill stream fish, Danio dangila (Ojha and Singh 1986), are virtually devoid of microridges. The microridges decrease in size and frequency from the gill arch, the gill filament to the secondary lamellae (Dunel-Erb and Laurent 1980a; Karlsson 1983), and have been associated with diverse roles which include trapping and holding mucus, providing structural integrity to the gill epithelium, and increasing the surface area at the water-epithelial surface interface (Sperry and Wassersug 1976). Olson (1996) contemplated that microridges generate an unstirred boundary layer of water over the gill epithelium hindering gas transfer. The presence of a mucous cover, which streamlines the gill surface by evening out the crypts between the microridges, should reduce the resistance of the flow of water across the gills (Daniel 1981; Hughes and Mondolfino 1983). The presence of microridges on the nonrespiratory surfaces of the accessory respiratory organs (AROs) in fish such as the climbing perch, Anabas testudineus (Munshi and Hughes 1991; Wu 1993), and the snake-head fish, Channa striata (Hughes and Munshi 1986), supports the suggestion that developmentally, the AROs arise from in situ modifications of the gills. Microridges have been said to enable the pavement cells to greatly distend without engendering mechanical disruption (Knutton et al. 1976). This may be crucial for the integrity of cells which are exposed to a medium whose ionic composition and osmotic pressures may change over short distances. The size and shape of microridges are thought to be influenced by electrolytes, salinity, hormones, and hydrodynamic flow of the water over the gills (Hughes 1979; Schwerdtfeger 1979; Wendelaar-Bonga and Meiss 1981). The mucus covering of the gills has been associated with numerous functions which include protection from mechanical damage and invasion by pathogens, absorption and expropriation of toxic heavy metal ions (e.g., Varansi et al. 1975; Varansi and Markey 1978), and regulation of O<sub>2</sub> (Ultsch and Gros 1979), and electrolyte (Handy 1989) transfer across the epithelial lining of the gills. From morphological characteristics and location within the epithelium, factors which may depend on the stage of cell maturity, two types of chloride cells have been described in fish gills (e.g., Laurent 1984; Maina 1991). An accessory chloride cell has been described in the gills of the saltwater fish (Dunel-Erb and Laurent 1980a; Laurent et al. 1995). The epithelial cells of the gills, especially the chloride cells, are highly sensitive to changes in ambient conditions. Movement of euryhaline fish from freshwater and seawater generates reversible changes in chloride cell morphology, location, and numerical density (e.g., Hossler 1980; Laurent and Hebibi 1989; Laurent and Perry 1991). Hyperoxic-effected hypercapnic acidosis causes an increase in the apical surface area of the chloride cells (Laurent and Perry 1991) while ambient hypercapnia increases the chloride cell number in the catfish, Ictalurus punctatus (Cameron and Iwama 1987). In what was construed to be an adaptive process, injections with cortisol for 10 days increased the number of chloride cells by a factor of 3 (Laurent and Perry 1990). The organization of the gill microcirculatory pathways in fish gills is far from settled (e.g., Steen and Kruysse 1964; Boland and Olson 1979; Dunel-Erb and Laurent 1980b; Olson 1996). The lamellae are divided into vascular channels which are lined by polygonal endothelial pillar cells (Figs. 11, 49). In the gills of some land crabs like Carcinus maenas (e.g., Farrelly and Greenaway 1992) and some teleosts, e.g., Oreochromis alcalicus grahami (Fig. 11), the pillar cells which contain the contractile microfilamental actomyosin elements and collagen (Bettex-Galland and Hughes 1973; Youlson and Freeman 1976) may play an important role in regulating translamellar blood flow. No shunts that bypass the secondary lamellae en route from the ventral to the dorsal aorta occur in the common water breathing teleosts (Olson 1996). In the gills of the European eel, Anguilla anguilla, the principal sites of gill vascular resistance are at the level of the afferent lamellar arterioles and the secondary lamellae (Bennett 1988).

The epithelial cell laver, basement membranes, interstitial space and endothelial cell (Fig. 49) constitute the water-blood barrier. Though typically thick in most fish (Hughes and Morgan 1973), the barrier may be as thin as 0.2 µm in some regions of the secondary lamellae of some species (Maina 1990a, 1991; Laurent et al. 1995). The morphometric features of the gills, especially the respiratory surface area, correlate with the metabolic demands of fish as well as the environment in which they live (e.g., Hughes and Morgan 1973; Maina et al. 1996a). The pillar cells are arranged as struts which span the width of the space between the two parallel epithelial sheets. They maintain the structural integrity of the secondary lamellae and the blood spaces by preventing overdistension under undue intramural blood pressures which may be as high as 12kPa (Bettex-Galland and Hughes 1973; Hughes 1976). Fish can regulate the surface area of their gills as well as the exposure of the chloride cells to water (Fig. 50). In that way, gas exchange and ionic regulation can be optimized to suit the prevailing needs and circumstances (e.g., Randall 1982; Butler and Metcalfe 1983). The position of the gill filaments can be varied by contraction of smooth muscles which are enervated by adrenergic nerves (e.g., Nilsson 1985). In a single gill filament, the blood flow across the lamellae at the tip is less than in those at the base (Hughes 1980) and can be regulated by certain pharmocological agents, e.g., serotonin, adrenaline, and noradrenaline (Östlund and Fänge 1962; Nilsson 1986; Fritsche et al. 1993; Sundin et al. 1995). In an individual secondary lamella, by contraction or relaxation of the pillar cells, the blood/hemolyphatic flow may be shifted to and from the larger marginal channels (Fig. 11). In the gills of the mudskipper, Boleophthalmus boddarti, a species which lives on the surface of the mudflats of the Arabian Gulf where it makes U-shaped burrows which may be as deep as 1 m (Clayton and Vaughan 1986), the water-blood barrier is thinner around the marginal channels (Hughes and Al-Kadhomiy 1996). Blood remains in contact with water for about 0.5 s, a duration considered to be adequate for complete oxygenation of the ervthrocytes (Hughes et al. 1981).

## 4.6 The Water Lungs

Gas transfer from water by ventilated lung-like (invaginated) structures is utilized by only a small group of animals. It presents a very simple mode of respiration. In the relatively more advanced organisms, it illustrates a retrogressive use of an earlier air breathing organ. In the marine snail, *Aplysia califormica*, respiratory pumping (e.g., Kandel 1979) or interneuron II response as it was called by, e.g., Eberly et al. (1981), is an activity which entails synchronized contraction of the gills, parapodia, and siphon. The seawater is taken in and injected from the mantle cavity. A more complex respiratory pattern which is depressed by longterm (24h) exposure to hypoxia (about  $2 \text{ mlO}_2$  per l of water) occurs in *A. califomica* (Kanz and Quast 1990). Hypercapnia (Croll 1985), hypoxia, or anoxia (Levy et al. 1989) increase the respiratory frequency in *A. califormica*. Aquatic animals like the sea cucumber (holothurians) have water lungs which occur in form of branched thin-walled tree-like perivisceral tubes that stretch from the



Fig. 50. a Chloride cell on the gills of a tilapiine fish, *Oreochromis alcalicus grahami*. The cells are involved in ionic exchange between the blood and water. > mitochondria; s supporting cells; p pavement cells; \* pore; b basement cell. b A surface view showing closed chloride cells (c) and an open mucus cell (m). a Bar 1  $\mu$ m; b 6  $\mu$ m. (a Maina 1990a)

cloaca, running through the whole length of the body. Expulsion of the seawater is produced by irregular contractions and relaxations of the body wall muscles and inspiration by the pumping action of the cloaca (Newell and Courtney 1965): gas exchange takes place between the inspired sea water and the hemoglobin containing coelomic fluid, a process enhanced by the stirring action of the peristaltic waves. In Holothuria forksali, cloacal O<sub>2</sub> uptake contributes about 60% of the total need. Gas exchange is reduced by 50% when cloacal ventilation is arrested. Although circulation of the body fluids in the Holothuria has been described (e.g., Kawamoto 1927), it appears to occur to a very small extent. The ventilatory activity of the lung is a more significant factor in effecting movement and mixing of body fluids. In a number of annelids, respiration through the walls of the gut has been suspected though the actual mechanism is not known (Krogh 1941). In a large number of Oligochaetes and polychaetes, antiperistaltic and ciliary movements in the anus are considered to be respiratory (Stephenson 1930). With the exception of Owenia, however, Lindroth (1938b) refuted that any significant per rectal movement of water occurs in the polychaetes to warrant the gastrointestinal system being considered to be of any functional consequence in respiration. Stephenson (1930) described a water-swallowing process and expulsion through the anus in the large worm, Aphrodite aculeata. In the small threadlike freshwater tubiform worms, e.g., Tubifex and Limnodrilus, antiperistatic movements of the hind gut occur (Alsterberg 1922). The worm, Limnodrilus, builds tubes in the very soft organic mud (which is virtually free of O<sub>2</sub>) and stretches its hind parts further into the surrounding water and makes strong undulating movements. When the O<sub>2</sub> tension drops to below 0.08 ml per l, the worms become immobile. In well-aerated water, Tubifex tubifex completely retracts into its burrow and, in a hypoxic condition, the worm waves its tail freely in the water (Alsterberg 1922; Palmer 1968). High O<sub>2</sub> concentration is said to be toxic to T. tubifex (Fox and Taylor 1954; Walker 1970). The worm has been shown to be highly tolerant to H<sub>2</sub>S toxicity (Degan and Kristensen 1981). The tracheal gills on the abdomen of nymphal Plecoptera and Ephemeroptera and larval Trichoptera consist of a panoply of tracheae which are ventilated by rhythmic movements in water while in some larval Odonata (e.g., Aeschna), similar structures are found in the hind gut where they are ventilated by contractions of the muscles of the body wall (Fig. 23).

The freshwater pulmonate gastropods of the Order Basommatophora after evolving air breathing have readapted remarkably well to aquatic life (Macchin 1974). This is demonstrated by the fact that the group lacks ctenidia and the mantle cavity has been totally converted to a water lung (Hunter 1953). In *Planorbis corneus*, the mantle is regularly extended into the water to serve as an accessory gill while in *Lymnaea pereger*, air is taken in as the animal dives. This presents a gas-gill function. In specimens living at considerable depths, the mantle cavity functions as a water lung (Ghiretti and Ghiretti 1975). In most pulmonate gastropods, a substantial quantity of O<sub>2</sub> is normally absorbed through the skin. The lung can remain closed for a long period of time and some of the aquatic species, e.g., *Limnea* and *Helicosoma* can remain indefinitely submerged at considerable depths at low temperatures (Cheatum 1934). If the O<sub>2</sub> drops below a critical level, the animal surfaces to breath. When exposed to a hypoxic environ-

ment or as the tide recedes, the bivalve Pholas dactylus collects water in the aperture on the inhalant siphon from which it draws 47% of its O<sub>2</sub> needs (Knight 1984; Knight and Knight 1986). The lungs of aquatic pulmonate gastropods, Lymnaea pereger and Physa fontinalis, which live away from water, may be filled with water throughout life (Hunter 1953). Arhynchite pugettensis has extensive cloacal diverticula (Manwell 1960) which may serve a similar role. The gephyrean worm, Urechis caupo, lives in U-shaped burrows in the intertidal flats along the Pacific coast of America. The levels of H<sub>2</sub>S may be as high as 25 to 30µl and the water becomes hypoxic during low tide, with  $O_2$  levels dropping to 46% of the air saturation. Through muscular contraction of the body wall, the worm ventilates its hind gut (which serves as the primary site for gas exchange) with seawater (e.g., Julian and Arp 1992): the maximum inflation of the hind gut is 2 ml of water per g body weight. At normoxia, the mean hind gut ventilatory volume is 0.70 ml of water per gram body mass and increases to  $1.4 \text{ mlg}^{-1}$  at 4kPa, the critical PO<sub>2</sub> at which O<sub>2</sub> uptake ceases (e.g., Eaton and Arp 1990). At maximal inflation, the mucoserosal thickness at the respiratory hind gut decreases from its resting size of 283 to 21 µm (Menon and Arp 1992a), a change which reduces the diffusional resistance to  $O_2$ , enhancing gas transfer but adversely increasing the permeability of H<sub>2</sub>S into the tissues (Julian and Arp 1992). A similar rectal gill mechanism exists in the echiuran worm, Arhynchite pugettensis, which has an extensive cloacal diverticulum (Manwell 1960). The skin of the echiuroid worm is remarkably thick to reduce O<sub>2</sub> loss to the ambient hypoxic water. To compensate for this, the hind gut is thin-walled and extends along the whole length of the body (e.g., Redfield and Florkin 1931). In a 60-g animal, the total ventilation of the burrow is 29 ml per min when the animal is feeding, and when it is not, it is 13 ml per min (Hall 1931): one half of the water is taken into the hind gut where one third of O<sub>2</sub> is utilized. Over and above causing mixing of water in the hind gut and hence ensuring more efficient delivery of O<sub>2</sub> into the tissues, the peristaltic waves also lead to movement of the coelomic fluid (which contains a large number of corpuscles which have hemoglobin), further promoting O<sub>2</sub> transfer to the tissue cells. The soft-shelled river turtles of the family Trionychidae, namely Amyda mutica and Aspidonotus spirifer, ventilate their mouths and pharynx with water at a rate of 16 times min<sup>-1</sup> when submerged and absorb O<sub>2</sub> through villus-like highly vascularized processes which cover the mucous membrane of the pharynx (Simons and Sussana 1886). The well-vascularized mouth of the cuchia eel, Amphinous cuchia, is utilized for gas exchange in both air and water (Singh et al. 1984). Through rhythmic ventilation of the cloacal bursae of the Amazon turtle, Podocnemys, 90% of the O<sub>2</sub> needs are met (Steen 1971). Due to the very small quantities of water taken up by the peristaltic processes, doubts have been expressed on the effectiveness of the rectal and intestinal gas exchange process particularly in hypoxic water (Krogh 1941). In such organisms, the skin is utilized for respiration to a greater extent.

A number of air-breathing aquatic snails successfully subsist in the tropical African swamps. The best-known one is the large amphibious *Pila ovata* (Ampullariidae), of which the mantle cavity is divided into water- and air-breathing chambers, and *Biomphalaria sudanica* (Planorbidae), an entirely aerial pulmonate gastropod. The European snails, *Planorbis* (*Biomphalaria*) corneus


**Fig. 51A–F.** Respiratory mechanisms and structure of the "lung" of the swampworm, *Alma emini*. A Surfacing from the soil. **B** Formation of a respiratory groove. **C**, **D**, **E** Closing up the respiratory groove. **F** Retracting into the mud. (Beadle 1957)

and P. ovata (Jones 1964), are, however, only marginally adapted for survival in a hypoxic environment by having a slightly higher blood O<sub>2</sub> affinity. Compared with the well-established fish, the tenuous hold of the snails and the insects on the anoxic tropical swamps is a reflection of their much recent reinvasion of this habitat, conceivably after earlier perfection for life on dry land. The swamp worm, Alma emini, presents a particularly fascinating adaptation for respiration in the tropical African swamps. Its ecological success in the habitat is reflected in its numerical abundance in the floating mats of papyrus swamps (Stephenson 1930; Beadle 1974). Like most African species of the genus Alma, the worm can lead a successful amphibious existence (Beadle 1957; Wasawo and Visser 1959). It can extract O<sub>2</sub> from both air and water: in water, the respiratory groove functions as a water lung. Alma subsists in a habitat which is both anoxic and highly reducing. The worm can survive for at least 2 days in total absence of  $O_2$  (Beadle 1957). The dorsal surface which is highly vascularized is spread out on emerging from the soil (Fig. 51) to form a temporary tubular "lung" (Figs. 45,52) through which gas exchange occurs (Maina et al. 1998). Air bubbles are thought to be trapped in the lung and drawn down into the soil for use during the subterranean sojoum. Mangum et al. (1975), however, noted that much of the air captured by the lung at the surface is lost during descent but some bubbles may be held by the hydrophobic cuticle, leaving a volume of about  $0.2 \mu$ l in an average specimen. The rate of O<sub>2</sub> uptake by the lung is 50 to 60% of that which occurs across the total



Fig. 52. Schematic view of the respiratory groove of the swampworm, *Alma emini*, which subsists in waterlogged hypoxic soil with abundant putrefying plant matter. The capacity to form a temporary lung for exchange of gases with air enables the worm to live in a hypoxic and hypercarbic habitat

body surface while in water or in air though the lung constitutes only 1.5% of the surface area of the body (Mangum et al. 1975): a 0.75-g worm was reported to have a respiratory surface of 11.4 mm<sup>2</sup>. Greater values (lung volume 67 cm<sup>3</sup>) and respiratory surface area 245 mm<sup>2</sup>) were estimated by Maina et al. (1998). The blood of Alma contains an extracellular high molecular weight hemoglobin with a remarkably high O<sub>2</sub> affinity (Mangum et al. 1975). The hemoglobin is fully oxygenated at a PO<sub>2</sub> of 0.3 kPa in absence of CO<sub>2</sub> and at less than 1.3 kPa at very high PCO<sub>2</sub> (27 kPa) (Beadle 1957). The high mortality rate when the worms are prohibited from gaining access to air (Beadle 1957) indicates that O<sub>2</sub> uptake occurs exclusively through the "lung". Metabolism in Alma may be mainly anaerobic with aerobic gas exchange serving only to neutralize the toxic end products of glycolysis. The biochemistry of aerobic metabolism in Alma is essentially similar to that of other multicellular animals (Beadle 1957; Coles 1970). Coles (1970) estimated that O<sub>2</sub> consumption in air is only  $10.8 \mu l g^{-1} h^{-1}$  at 23 °C but much higher values were reported in both water  $(123 \mu l g^{-1} h^{-1})$  and air  $(230 \mu l g^{-1} h^{-1})$  by Mangum et al. (1975). These observations correspond with those made by Laverack (1963) on related aquatic tubificid worms. In the juvenile stages, the suprabranchial chamber membranes of the climbing perch, Anabas testudineus, are used for aquatic respiration, i.e., as water lungs (Munshi and Hughes 1986), a process which persists in adult anabantoids (e.g., Peters 1978).

# 4.7 The Placenta: an Ephemeral Liquid to Liquid Gas Exchanger

Though ubiquitous among the metatherian and eutherian mammals, in lineages where viviparity is ancient and appears to have evolved only once from a common inceptive ancestor and was from then conserved (e.g., Guillette and Hotton 1986; Packard et al. 1989), the placenta has developed in practically all vertebrate groups except in the agnathan and avian species. In some vertebrate classes, viviparity has evolved repeatedly (Hamlett 1986, 1989; Wourms and Callard 1992). For example, among reptiles, in the squamates, e.g., the lizards Sphenomorphorus quoyii (Grigg and Harlow 1981) and Niveoscincus metallicus (Stewart and Thompson 1994), and the snakes, e.g., the adder, Vipera berus (Bellairs et al. 1995), and the garter snake, Thamnophis sirtalis (Hoffman 1970), the process has evolved many times (Weekes 1935; Blackburn 1993). In the sphenodontids, crocodilians, and turtles, however, it does not appear to have ever evolved (e.g., Blackburn 1982; Shine 1985). The lizard, Sceloporus aeneus, which lives at high altitudes in Mexico, exhibits a bimodal mode of reproduction, i.e., both viviparity and oviparity can occur (Guillette 1982; Guillette and Jones 1985). Among amphibians, a few anurans, 15% of the urodeles, and 50% of the caecilians (e.g., Typhlonectes compressicauda; Garlick et al. 1979) exhibit viviparity (Wake 1989, 1993). Fish like the teleost, Zoarces viviparous (Weber and Hartvig 1984; Hartvig and Weber 1984), sharks, e.g., Scyliohinus settaris (Wourms et al. 1988; Hamlett 1989; Wourms 1993), and insects (e.g., the tse-tse fly) have functional placentae. Viviparity affords protection of the embryo from adverse environmental conditions and predation (Blackburn 1982; Shine 1983; Shine and Guillette 1988). Extended internal fetal nutrition supports development to a more advanced state (e.g., Lillegraven et al. 1987; Shine 1989; Guillette 1993), improving chances of survival. Oviparity is thought to be ancestral to viviparity (Hamlett 1989). It is envisaged that reduction of the thickness of the eggshell, e.g., in the lizard, Sphenomorphus fragilis (Greer and Parker 1979) where the thickness of the shell is only 10 µm, the egg membrane in sharks (Hamlett 1987, 1989) and reduction of the number of eggshell glands, and increase of oviductal vascularity, e.g., in the shark, Squalus acanthias (Jollie and Jollie 1967) may be the initial morphological preparations that preceded the evolution of viviparity (Guillette 1989, 1991). Decrease in the eggshell thickness brings the embryonic and maternal circulations closer, predisposing implantation, egg retention, and formation of the placenta, an organ that serves both nutritive and respiratory roles. The transition from oviparity to viviparity is a gradual process which entails both morphological and endocrine changes (Guillette 1991; Hamlett 1989).

Like the gills, the placenta is a liquid-to-liquid gas exchanger. Unique to practically all evolved gas exchangers, the placenta is a secondary gas exchanger, in that it relies entirely on another organ (gill or lung) for gas transfer. While the placenta performs other important roles which include synthesis of hormones, transfer of ions and metabolites (e.g., Faber 1993), and protection of the fetus from adverse ambient pertubations (Laburn et al. 1994), its primary function is unequivocally that of gas exchange (Mayhew 1992). As in other multifunctional organs, the ultimate design of the placenta must accommodate all the constitutive roles. Faber et al. (1992) observed that, despite the purpose of the placenta in all species that have evolved it being essentially the same, "there is no other mammalian organ whose structure and functions are so species-diverse". The respiratory challenges faced by the mammalian fetus are to an extent similar to and in some ways different from those of the bird embryo (Sect. 6.13): both operate within a limited and fixed space. Compared with the eggshell, however, the placenta is a dynamic organ which grows and changes with gestation to meet the increasing fetal demands for O<sub>2</sub>. The placental function is determined by the growth and development of the terminal villi, the sites where materno-fetal exchange of respiratory gases and metabolites occurs (e.g., Teasdale 1980; Fox 1986; Mayhew et al. 1986; Stoz et al. 1988; Jackson et al. 1992). The human placenta has been conceived as a spongy medium whose porosity is set by the spatial interdigitation and configuration of the maternal and fetal villous systems (Schmid-Schönbein 1988), giving a villous surface area of about 11 m<sup>2</sup> contained in an average volume of about 500 cm<sup>3</sup> (Aherne and Dunnill 1966). The geometry of the maternal and fetal vasculature determines the perfusive (hemodynamic) characteristics and the diffusive capacities of the placenta to the respiratory gases (e.g., Lee and Mayhew 1995). With a few falling in between, dependent on factors such as molecular weight, electrical charge, vascular geometry, and concentration gradient, the transfer of substances across the placenta is either entirely flow-limited or diffusion-limited (Faber et al. 1992). During normal human placental development, villous maturation is characterized by increased tissue and blood volume. The cross-sectional surface area of the terminal villi increases gradually stabilizing at 2000  $\mu$ m<sup>2</sup> between the 28th and 36th week of gestation (Ruckhäberle et al. 1977; Teasdale 1980; Stoz et al. 1988; Karsdorp et al. 1996). As in all gas exchangers, a short diffusional distance and an extensive surface area are the structural features for efficient gas exchange (Figs. 53,54). In the placenta, the distance between the two blood streams, i.e., the materno-fetal placental barrier, ranges between 2 to 6µm in man, 6 to 8µm in the cat, and 1.5 to 3µm in rodents and



**Fig. 53.** Schematic drawing of the maternal-fetal gas exchange pathway. The maternal placental circulation is separated from the fetal one by a tissue barrier, *vm*, composed mainly of trophoblast, *t*, syncytiotrophoblast, *s*, and endothelial cells, *e*. The placental barrier is highly attenuated to enhance gas exchange; *mp* maternal plasma; *fp* fetal plasma; *me* maternal erythrocytes; *fe* fetal erythrocytes



Fig. 54. Maternal-fetal barrier,  $\succ$ , of the placenta of the spotted hyena, *Crocuta crocuta*, showing the fetal capillaries, *f*, and the maternal blood spaces, *m*; *e* erythrocytes; *t* trophoblast. The *white spaces* in the barrier are lipid aggregations which have been removed after tissue processing. *Bar* 3 µm. (Courtesy Prof. D.O. Okello, Department of Veterinary Anatomy, University of Nairobi, Kenya)

Leporidae (Bartels 1970). The transplacental O<sub>2</sub> transfer is effected by diffusion driven by prevalent partial pressure gradient between the maternal and fetal blood streams. A human placenta with a surface area of 15 m<sup>2</sup> and a thickness of 5 µm will transfer 0.113 ml O<sub>2</sub> s<sup>-1</sup> mbar<sup>-1</sup> (Bartels 1970). The microsomal membrane carrier, cytochrome P<sub>450</sub> which reversibly binds with O<sub>2</sub> has been implicated

in promoting  $O_2$  transfer across the placenta (e.g., Burns and Gurtner 1973). The importance of  $P_{450}$  in the flux of  $O_2$  across the materno-fetal placental barrier, however, appears to be very small. No increase in the concentration of  $P_{450}$  occurs during hypoxia (Gilbert et al. 1979). In dog and sheep lungs (Burns et al. 1975, 1976), in the liver (Rosen and Stier 1973), and in body tissues in general (Longmuir and Sun 1970),  $P_{450}$  has also been incriminated with  $O_2$  transfer. There is presently no hard evidence to support occurrence of active transport of  $O_2$  in any evolved gas exchanger (Dawes 1965). Present experimental evidence strongly indicates that the process occurs entirely by simple physical diffusion.

The placental diffusional pathway is complicated, even in the much simpler hemochorial human placenta. The maternal-fetal placental barrier is neither uniform in thickness nor is it homogenous in composition (e.g., Jackson et al. 1985). Oversimplified physiological models fail to take into account factors such as tissue inhomogeneity (e.g., Laga et al. 1974), maternal-fetal placental perfusion inhomogeneities (Bøe 1954), and placental O<sub>2</sub> consumption and vascular shunts (Barcroft and Barron 1946). About 40% of the human placenta consists of maternal blood (hematocrit, 36%) which is three to five times greater in volume than that of the fetal blood (Ht, 50%) (Mayhew et al. 1984): the mean harmonic mean diffusional distances of the maternal blood plasma, the villous membrane, and the fetal plasma are respectively 0.92, 4.08, and 1.88 µm, giving an overall placental thickness of 6.8µm. Aherne and Dunnill (1966) reported a thinner maternalfetal barrier of 3.5µm. The villous membrane exhibits remarkable sporadic attenuation (Aherne and Dunnill 1966; Jackson et al. 1985), a property similar to that presented by the blood-gas barrier of the mammalian (Gehr et al. 1981) and avian lungs (Maina and King 1982a; Figs. 29b,49a), where it is said to enhance O<sub>2</sub> transfer by generating an overall thin boundary without compromising the mechanical integrity of the gas exchanger. In the human placenta, the maternal-fetal barrier, which constitutes as much as 58 cm<sup>3</sup> of the trophoblast (Aherne and Dunnill 1966), may constitute a significant sink for  $O_2$ .

The fundamental structural features of any gas exchanger include an extensive surface area and a thin barrier between the respiratory media. These features are achieved in the placenta through different processes which include: (1) plasmodial activity which results in nuclear aggregation of the trophoblastic cell masses at syncytial knots leading to attenuation in some regions, especially those overlying the blood capillaries - such sites from the extremely thin vasculosyncytial membranes which promote gas and nutrient transfer (Bender 1974; Jones and Fox 1977; Heijden 1981), (2) through a mechanistic process which causes distension and margination of fetal capillaries and their relocation to the overlying trophoblast (Amaladoss and Burton 1985; Jackson et al. 1988a,b; Mayhew and Wadrop 1994), and (3) enlargement of the surface area through increase in the number of microvilli (Firth and Farr 1977; Heijden 1981). Attenuation of the trophoblast occurs in the guineapig placenta (Bacon et al. 1984) and in the human cultured one after long-term maternal exposure to hypoxia (Burton et al. 1989). Some parts of the placenta may be concerned with gas exchange while others are involved in processes which require greater tissue density such as hormonal synthesis and nutrient transfer (e.g., Bartels and Metcalfe 1965) as well as mechanical support. Interestingly, a similar engineering process appears to occur in the bird lung where the cell bodies of the pneumocytes are largely confined to the penultimate gas exchange sites, the atria and infundibulae: the extremely thin blood-gas barrier is virtually lined by the attenuated cytoplasmic extensions (Maina and King 1982a). In the mammalian lung, thin and thick sides of the interalveolar septae occur (Fig. 86). The thin parts are utilized for gas transfer while the thick ones render mechanical support and provide pathways for lymphatic drainage (Fishman 1972).

It is widely assumed that the structural complexity of a placenta, i.e, the number of tissue layers comprising it, determines the thickness of the maternofetal placental barrier and hence the diffusing capacity of the organ for O<sub>2</sub>. The less elaborate hemochorial placentae are hence thought to be more efficient than the more elaborate epitheliochorial ones. There are, however, no reliable data to indicate that the simple placentae are in a way more efficient in gas exchange and in promoting fetal growth and development (e.g., Dempsey 1960). Furthermore, it is known that in certain epitheliochorial placentae, the overall barrier thickness is smaller than in some hemochorial ones, which specific areas of the barrier being only about 1 µm thick (Ludwig 1965). The maternal-fetal placental barrier thickness varies greatly in different species and even in the same species during different stages of gestation. Moreover, it has been shown that placental forms may be mixed, i.e., epitheliochorial and hemochorial parts may coexist in the same placenta (e.g., Starck 1959). The diffusing capacity of the placenta depends not only on the path length of the barrier and permeability but also on factors such as the mean diffusion gradient and the placental surface area, features which can be altered according to needs. The transplacental  $O_2$  gradient in the Ilama (about 2 kPa) is remarkably low, a state perhaps compensated for by the extensive placental gas exchange surface (Barron et al. 1964). The mean diffusional gradient of O<sub>2</sub> in the hemochorial placenta of the rabbit at 27 to 30 days gestation (term about 31 days) ranges from 1.2 to 1.9kPa and in the syndesmochorial placenta of sheep between 126 and 137 days of gestation (term about 147 days) ranges between 4.3 and 6.4 kPa (Barron and Meschia 1954). From estimates of O<sub>2</sub> consumption of the pregnant human uterus and PO<sub>2</sub> in the maternal and fetal blood streams, Metcalfe et al. (1967) calculated that the physiological diffusing capacity of the human placenta ranges from 0.014 to  $0.018 \text{ ml } O_2 \text{ s}^{-1} \text{mbar}^{-1}$ . Calculations based on the diffusing capacity of CO<sub>2</sub> in pregnant women (Forster 1973) gave higher value of  $0.025 \text{ ml } O_2 \text{ s}^{-1} \text{ mbar}^{-1}$ . On average, data indicate that the physiological diffusing capacity of the human placenta lies between 0.013 and 0.038 ml  $O_2 s^{-1} mbar^{-1}$  and is appreciably lower than the morphometric one, which ranges from 0.055 to 0.072 ml O<sub>2</sub>s<sup>-1</sup> mbar<sup>-1</sup> (Mayhew et al. 1984). The placental membrane of the physiologist (= villous membrane + serial blood plasmas) accounts for 86 to 94% of the total placental resistance with only minor contributions made by the O<sub>2</sub>-hemoglobin interactions with the erythrocytes (Mayhew et al. 1984, 1986). In the human lung (Gehr et al. 1978), the physiological and the morphometric diffusing capacities of the lung differ by a factor of 2. This is taken to constitute a functional reserve (Weibel 1984a). The value of the physiological diffusing capacity approaches the morphometric one at the maximum O<sub>2</sub> consumption (Vo<sub>2max</sub>). The difference between the morphometric and the physiological diffusing capacity of the placenta may functionally be accounted for by vascular shunts, placental  $O_2$  consumption, and regional inequalities of perfusion (Metcalfe et al. 1967; Mayhew et al. 1984, 1990), as well as the lack of uniformity of the thickness of the villous membrane. The latter feature may lead to local inhomogeneities of diffusion resistances across the sporadically attenuated barrier (Mayhew et al. 1984; Jackson et al. 1985). In the vertebrate lung, anatomical and functional shunts and regional inhomogeneities in gaseous diffusion and vascular perfusion contribute to the discordance between the physiological and morphometric diffusing capacities (e.g., Crapo and Crapo 1983). Aherne and Dunnill (1966) envisaged that the mass of the placenta and hence the development of the chorionic villous area (transfer area) correlate directly with the total fetal metabolism.

While the lung is subjected to sudden increases in functional demands from rest to maximal O<sub>2</sub> consumption during exercise, the needs of a fetus tend to be fairly stable and gradually increase with gestation. In the human being, during the 3rd month of fetal life, the O<sub>2</sub> capacity is 12 ml O<sub>2</sub> per 100 ml blood, increasing to 20 to 22 ml  $O_2$  per-100 ml blood during the next 5 months: the maximum  $O_2$ capacity is reached at 6 to 8 weeks (e.g., Betke 1958). From the evident progressive degenerative changes such as infarcts, fibrinoid deposition, thickening of the trophoblastic basement membrane, partial obliteration of decidual arteries, and endothelial cell proliferation and calcification (Mayhew et al. 1984), the morphometric diffusing capacity of the placenta would be expected to remarkably increase towards term. This deterioration is, however, counteracted by a gradual decrease in the thickness of the villous membrane which occurs with gestation when the remarkably attenuated vasculosyncytial regions become more pronounced (Fox 1964a; Jackson et al. 1988b). The arithmetic mean thickness of the trophoblast in the human placenta decreases from about 11 um at 12 weeks to  $5 \mu m$  at 38 weeks (Jackson et al. 1988b), a change brought about by displacement of the trophoblastic cell masses, leading to better exposure of the fetal capillaries to maternal blood (Teasdale 1978; Jackson et al. 1988a,b).

#### 4.7.1 The Functional Reserves of the Placentae

The placenta is widely taken to be a progressively aging organ of which the functional capacity declines gradually towards term (Winick et al. 1967). This process is oddly accompanied by increased fetal demands which occur with growth and development. Towards term, fetal requirements come close to totally eroding the functional reserves of the placenta, in some cases leading to intrauter-ine growth retardation (Hellman et al. 1970; Garrow and Hawes 1971). Winick et al. (1967) found that no further increase in placental DNA content occurred once the fetus reached a body mass of 2.4 kg or the placenta reached a weight of 300 g. According to Rolschau (1978; but see a contrary observation by Sands and Dobbing 1985), growth of the human placenta levels up at 35 weeks of gestation. At altitudes above 3 km above sea level, due to hypobaric hypoxia (Mayhew 1991), human fetal growth is retarded (e.g., Haas 1976). Increased fetal erythropoiesis expressed as high hematocrit, hemoglobin concentration, and proportion of

hemoglobin-F suggest that the fetus experiences hypoxia during the last stages of pregnancy (Ballew and Haas 1986). Maternal adjustments, among others hyperventilation and elevated hematocrit, during high altitude pregnancy (Moore et al. 1982), appear to fall short of providing the necessary driving pressure of  $O_2$  across the placenta. Mayhew (1991) observed that with altitude, while adaptive diffusive changes occur on the maternal side of the placenta, fetal conductances especially of the erythrocytes and plasma do not adjust to the same degree, leading to low fetal birth weights at altitude (Mayhew et al. 1990). The difference between the maternal and fetal adjustments for high altitude hypoxia is greatest at birth weights greater than 3 to 3.3 kg, which is about the average birth weight at altitude (Mayhew 1991).

The O<sub>2</sub> transferred by the placenta and made available to the fetus must initially be procured by the maternal lung. The fetus is essentially a temporary addition to the maternal gas transfer cascade (Figs. 2,3). On this account, it is plausible that at some critical point, the fetus would be indirectly affected by extreme states and circumstances which may occasion inadequacy in maternal pulmonary gas flux and cardiovascular transfer. These may result from exposure to hypoxia, i.e., reduction of the driving pressure of gases across the lung and the placenta or may be due to pathological conditions at either or both sites. An increase in the fetal O, saturation is generated on maternal exposure to high PO, (e.g., Dawes and Mott 1962; Cassin et al. 1964; Assali et al. 1968). Overdesign and/ or plasticity to respond to circumstances when the functional capacity of the maternal gas exchanger may become insufficient is necessary (e.g., Becker 1963; Bender 1974). Such instances include maternal exposure to acute life-threatening hypoxia and pathological conditions such as edema, atelectasis, and pulmonary infarction (e.g., Staub et al. 1967; Staub 1974; Heijden 1981), changes which constrain gas exchange. Perfectly normal reproduction can occur when the maternal blood PO<sub>2</sub> is subnormal, e.g., in women with congenital heart disease (Bartels 1970): in most cases, the consequences largely affect the mother more than the fetus (Burwell and Metcalfe 1958). The Korean Ama (sea women) who voluntarily dive to harvest food at the bottom of the ocean (at depths as much as 30 m) work up to the last day of pregnancy and after giving birth nurse the babies between shifts (Hong and Rahn 1967). Maternal diabetes mellitus does not appear to affect placental development (Teasdale and Jean-Jacques 1986). Oxygen tension in the umbilical vessels was found to be similar in sheep fetuses at sea level and those of ewes living at high altitude (3.5 to 4.0 km) (Metcalfe et al. 1962), indicating that the mother offers efficient protection against changes in blood PO<sub>2</sub>. During the birth process, the fetus is protected from hyperthermia: in sheep, during parturition, the maternal body temperature rose at 0.70 °Ch<sup>-1</sup> in the final stages of labor but the fetal one rose at a significantly lower rate of 0.45 °Ch<sup>-1</sup> (Laburn et al. 1994). In normally developing human pregnancies, a drop in the peripheral vascular resistance in the placenta occurs after a gestational age of 16 weeks. This results in positive end diastolic flow velocity waveform in the umbilical artery (van Zalen et al. 1994). In pregnancies complicated by hypertension and/or intrauterine growth retardation, however, placental flow resistance is elevated. This may lead to absent or even reversed (negative) end diastolic flow velocities in the umbilical arteries during the second and third trimester of pregnancy. Such a condition may result in preterm delivery, neonatal death, and lower birth and placental weights (e.g., Aherne and Dunnill 1966; Trudinger et al. 1991; Pattinson et al. 1993; Karsdorp et al. 1994, 1996).

Like the avian and mammalian lungs, the placentae possess a substantial functional reserve. Large fetal lambs (a few days to term) can maintain an O<sub>2</sub> consumption of 5 to 6 ml kg<sup>-1</sup> min<sup>-1</sup> (values within the normal range) even when the umbilical venous and arterial saturations are respectively reduced to 54 and 35% at an umbilical flow rate of  $180 \,\mathrm{ml\,kg^{-1}\,min^{-1}}$  (Dawes et al. 1953). Perhaps it is as a part of an inbuilt safety margin of operation that the morphometric diffusing capcity of the human lung ( $2.47 \text{ ml} \ \hat{O}_2 \text{ s}^{-1} \text{ mbar}^{-1}$ ; Gehr et al. 1978) is about 33 times greater than that of the placenta of  $0.075 \text{ ml O}_2 \text{ s}^{-1} \text{ mbar}^{-1}$  (Mayhew et al. 1984). Structurally, the placenta is a highly adaptable organ with a considerable functional reserve capacity (e.g., Karsdorp et al. 1996). Perfusion of the fresh human placenta with varying concentrations of O2 causes obvious thinning of the trophoblast from 0.44 to  $3.3 \,\mu m$  in a matter of 6 h and when the O<sub>2</sub> levels are brought back to normal, the dimensions are reversed (Tominage and Page 1966). The proliferation of the cytotrophoblast appears to be sensitive to the prevailing O, levels (Fox 1964b; Kaufmann 1972): cytotrophoblast decreases when oxygenation is good and increases in conditions associated with intrauterine hypoxia (Fox 1964b, 1970; Kaufmann et al. 1977). Chronic maternal exposure to hypoxia in guinea pigs leads to thinning of the trophoblast (Bacon et al. 1984). The rate of flow of the placental blood at term, which is 500 to  $600 \,\mathrm{ml\,min^{-1}}$ , is in excess of that of 300 to 400 ml required to supply 16 ml of O<sub>2</sub> to the human fetus per minute (Gahlenbeck et al. 1968). A maternal-fetal PO<sub>2</sub> of 0.3 to 0.4 kPa is adequate to supply the required amount of  $O_2$  to the fetal tissues (Bartels 1970) but a much higher gradient of 2 to 4kPa has been determined in the larger blood vessels (Metcalfe et al. 1967). Infections, underlying pathological conditions, and exposure to severe conditions such as extreme hypoxia appear to accelerate placental development (Jackson et al. 1995; Lee and Mayhew 1995; Karsdorp et al. 1996). Terminal villi of placentae, of which the pregnancy is accompanied by absent or reversed end diastolic blood flow in the umbilical artery, show a more homogeneous pattern of small villi (Karsdorp et al. 1996). Hitschold et al. (1992) suggested that the accelerated development of the terminal villi may be a compensatory mechanism but the potential advantages gained from it may be curtailed by the concomitant reduction in the blood flow rate in the umbilical artery (Erskine and Ritchie 1985; Karsdorp et al. 1996). The harmonic mean thickness of the placentae of women living at high altitude (average thickness  $4.5\,\mu m$ ) was 8 to 19% thinner than that of those living in the lowlands (average thickness 5.2 $\mu$ m) and the morphometric diffusing capacity of O<sub>2</sub> in the former was higher than in the later (Mayhew et al. 1984; Table 14). Elevated fetal hematocrit, high hemoglobin concentration, low O<sub>2</sub> affinity, and high O<sub>2</sub> carrying capacity of blood (e.g., Ballew and Haas 1986; Tables 15,16) are vital physiological adjustments in highland pregnancy and increase the diffusive conductance of the placenta in a hypobaric and hypoxic circumstance. The most critical point of the development of the human placenta when there is substantial increase in volume,

Variable	Low altitude	High altitude		
Dme	1050	1700		
Dmp	620	1190		
Dtr	28.6	28.7		
Dst	49.8	70.8		
Dfp	726	666		
Dfc	290	237		
Overall Dp	15.7	17.5		
Specific Dp	4.7	5.79		

**Table 14.** Diffusive conductances in low and high altitude placentae.(After Mayhew et al. 1990)

Units:  $mlO_2min^{-1}kPa^{-1}$ ; specific value,  $mlO_2min^{-1}kPa^{-1}kg^{-1}$ .

Symbols: me, maternal erythrocytes; mp, maternal plasma; tr, trophoblast; st, stroma; fp, foetal plasma; fe, foetal erythrocytes; Dp, diffusing capacity of the placenta.

Table 15. Oxygen affinity ( $P_{50}$ ) and oxygen carrying capacity of maternal and fetal blood of various species (After Novy and Parer 1969)

Species	P <sub>50</sub> at pH 7.40 (n	nmHg)	$O_2$ capacity mlO <sub>2</sub> 100 ml <sup>-1</sup>		
	Maternal <sup>a</sup>	Fetal <sup>a</sup>	Maternal	Fetal	
Man	26	22	15	22	
Rhesus monkey	32	19	15	18	
Rabbit	31	27	15	14	
Sheep	34	17	15	17	
Goat	30	19	13	12	
Pig	33	22	13	13	
Elephant	24	21	20	17	
Camel	20	17	15	17	
Llama	21	18	14	19	
Cat	36	36	12	16	

<sup>a</sup> To convert to kPa multiply by 0.133.

surface area, length of villi, and overall thinning of the trophoblast lies between 17 to 21 and 22 to 26 weeks of gestation (Jackson et al. 1992; Simpson et al. 1992; Mayhew and Simpson 1994).

The performance of the placenta falls between that of the lung, which on average operates well below its maximal capacity but can respond to sudden demands placed on it, and the eggshell, in which the structural parameters are firmly incorporated. There are fundamental limitations intrinsic to the placenta as a gas exchanger: the O<sub>2</sub> consumption of the human hemochorial placenta (2 to  $10 \text{ ml O}_2 \text{ kg}^{-1}$  wet mass min<sup>-1</sup>) at term indicates that placental tissue utilizes as much O<sub>2</sub> as the fetus itself (e.g., Nyberg and West 1957). The placental O<sub>2</sub> con-

Species	O <sub>2</sub> Capacity ml O <sub>2</sub> 100 ml bl	$ood^{-1}$	Half saturation pressure (mmHg)		
	Maternal	Fetal	Maternal <sup>a</sup>	Fetal <sup>a</sup>	
Man	15	22	26	22	
M. mulatta	15	18	-	-	
Rabbit	15	14	31	27	
Sheep	15	17	34	17	
Goat	13	12	30	19	
Guineapig	16	16	30	19	
Elephant	20	17	23	17	
Camel	15	17	21	16	
Llama	14	19	21	18	
Cow	15	12	21	22	
Chicken	14	12	49	34	
Seal	32	28	29	21	

Table 16. Maternal and fetal oxygen capacities of a number of spacies. (After Bartels 1970)

<sup>a</sup> To convert to kPa multiply by 0.133.

sumption in the more complex placentae like the epitheliochorial ones which have a greater tissue density would be expected to have an even greater O<sub>2</sub> consumption. In sheep, the placenta and fetal membranes consume as much as one third of the fetal  $O_1$  uptake (e.g., Longo et al. 1973). This suggests that in cases of anoxia, the fetus may be critically deprived of its normal O<sub>2</sub> needs. In the cow, the physiological diffusing capacity between the 5th and 9th month of gestation is  $0.0016 \text{ ml O}_{2} \text{ s}^{-1} \text{ mbar}^{-1} \text{ kg}^{-1}$  (Gahlenbeck et al. 1968) but the placenta increases in mass three to ten times during the same period. Experimental Swiss mice at a simulated altitude of 4.3 to 6.1 km show normal mating behavior, fertility, and reproduction, but early embryo resorption occurs at a greater frequency than in the control (sea level) group (Baird and Cook 1962). The resorption takes place at an embryo size of about 7 mm crown rump length, presumably due to deprivation of O<sub>2</sub> (by the placenta) at a critical stage of development. In an attempt to overcome these limitations, the epitheliochorial placentae of sheep and goat and the hemochorial ones of rabbit and guinea pig have developed the efficient countercurrent arrangement between fetal and maternal blood flows (e.g., Kaufmann and Davidoff 1977; Faber et al. 1992). In the rabbit and the guinea pig, the PO<sub>2</sub> in the umbilical vein exceeds that in the uterine vein. The placentae of the rabbit and the guinea pig are categorized among the efficient group of exchangers with the hemochorial placentae of the rhesus monkey and human being including the epitheliochorial placentae of goat and sheep being placed in the inefficient category (Faber et al. 1992; Table 17). From studies of compensatory mechanisms of the injured guinea pig placenta, Heijden (1981) observed that the organ has a very small functional reserve capacity and its potential to respond to reduced capacity is very poor. The countercurrent system of the placenta in sheep and goat is notably inefficient as evinced by the lower physiological diffusing capacity of the organ in the two species (Metcalfe et al. 1967). Having an efficient placenta has its

Species	Placental type	Vascular geometry	Vo <sub>2</sub>	PO <sub>2</sub> (mmHg) <sup>a</sup>	Dpo <sub>2</sub> <sup>b</sup>
Human	Hemochorial	Multivillous	7.4	23	0.32
M. mulatta	Hemochorial	Multivillous	10.0	33	0.30
Cow	Epitheliochorial	Countercurrent	9	38	0.24
Sheep	Epitheliochorial	Countercurrent	9	40	0.22
Goat	Epitheliochorial	Countercurrent	12	40	0.30
Rabbit	Hemochorial	Countercurrent	7.7	10	0.70
Guineapig	Hemochorial	Countercurrent	7.0	10	0.70

**Table 17.** Placental type, vascular geometry, foetal oxygen consumption  $(Vo_2)$ , oxygen pressure gradient between maternal and fetal blood  $(PO_2)$  and diffusion capacity  $(Dpo_2)$  of placentas of several species. (After Bartels 1970)

<sup>a</sup> To convert to kPa multiply by 0.133.

<sup>b</sup> Dpo<sub>2</sub> calculated using the carbon monoxide method of Longo et al. (1967).

Units:  $Vo_2$ , ml  $O_2$  min<sup>-1</sup> kg fetus<sup>-1</sup>;  $PO_2$ , mmHg;  $Dpo_2$ , ml  $O_2$  min<sup>-1</sup> mmHg<sup>-1</sup> kg<sup>-1</sup> fetus.

Table 18. Oxygen partial pressure in maternal arterial and uterine vein blood as well as in venous and arterial fetal blood for a number of species. (After Bartels 1970)

Species	PO <sub>2</sub> <sup>a</sup>			Placenta type	Flow pattern	
	Maternal		Fetal			
	Art.	Ven. Ut.	Ven. Umb.	Art. Umb.		
Cow	70	34	21	12	Epitheliochorial	Countercurrent
Sheep	86	52	29	18	Epitheliochorial	Countercurrent
Goat	84	46	33	14	Epitheliochorial	Countercurrent
Man	100	33	29	17	Hemochorial	Multivillous
M. mulatta	88	30	16	12	Hemochorial	Multivillous
Rabbit	80	25	46	17	Hemochorial	Countercurrent
Guinea pig	92	20	30	8	Hemochorial	Countercurrent

<sup>a</sup> Units: mmHg – to convert to kPa multiply by 0.133.

Art., arterial; Ven., venous; Ut., uterine; Umb., umbilical.

price and risk. The rate of flow-limited transfer diminishes much more steeply in the countercurrent exchanger than in the concurrent one when the flow rate of one of the blood streams decreases (Faber et al. 1992). After a flow reduction of 50%, gas exchange in the countercurrent system falls by as much as one half but in the concurrent system this drops by only one third. In this respect, the rabbit and guinea pig placentae possess a lower safety margin or operation, as has been noted by workers who have observed the remarkable fragility of the rabbit and guinea pig placentae, compared with the more resilient ones of the sheep. Despite the intrinsic structural differences, the mature hemochorial placentae show notable similarity in diffusional permeabilities regardless of whether they belong to the hemomonchorial (guinea pig and human), hemodichorial (rabbit), or hemotrichorial (rat) groups (e.g., Metcalfe et al. 1967; Štulc 1989; Tables 17,18). Dawes (1965) observes that "there is a rugged quality about the way these machines (placentae) are put together which appears to give a wider safety margin than is usually supposed". Placental morphology very poorly reflects the phylogenetic and systematic affinities, especially in mammals (Mossiman 1987; Luckett 1993). The morphological disparity of the placenta defies simple logic. It is a showpiece of the remarkable intrinsic plasticity of biological entities for solving different challenges. Different animal groups have achieved viviparity through manifold strategies and with it design-specific placentae. In a transient organ on which enormous resources are invested to ensure proper growth and development of the fetus, perhaps need to evolve a common architectural plan, a process which would be limiting to some animals, has not arisen. As for the evolution of the blood pigment carriers (Sect. 2.8), those aspects of biology which show great diversity of form and function may be symptomatic of conditions and processes which are far from attaining optimal designs and states. In such cases, encompassing solutions are untenable due to immutable phylogenetic constraints.

# **Bimodal Breathing: Compromise Respiration**

"For animal lines moving from water to land, the changes in physical and chemical characteristics of the environment are immense. These changes affect all possible life processes, from respiration and excretion to methods of movement, the functioning of the sense organs and reproductive mechanisms. The transition must therefore have been made very gradually, and some of the transition stages are repeated by present day groups." Little (1990)

#### 5.1 The Water-Air Interface: an Abstract Respiratory Rubicon

The division of the Animal Kingdom into aquatic and terrestrial life is ancient: it is still relevant to contemporary life. This distinction is ascribed to the different structural and functional attributes which have been imposed by the physical characteristics of the two different fluid media. While looking at this broad picture, it is too often forgotten than a rich assortment of animals regularly commutes between water and air. While some largely subsist in water and extract their  $O_2$  needs from the air above, some start their development in water and end up on land at maturity. The mutual assemblage and the distribution of the animals that share this complex lifestyle demonstrates the highly pragmatic strategies which animals have adopted to overcome the ceaseless selective pressures that have beset them (Fig. 55). While hypoxic conditions are a rare occurrence in terrestrial environments except at high altitude and in deep, compact wet soil, many marine and freshwater habitats are characteristically hypoxic and/or hyperbaric (Carter and Beadle 1931; Carter 1935, 1955; Hora 1935; Saxena 1963; Rhoads and Morse 1971). Depending on nature and rate of organic putrefaction of plant and animal matter, such habitats may contain noxious gases such as H<sub>2</sub>S, a potent inhibitor of cytochrome c oxidase systems, and ammonia (e.g., Theede et al. 1969).

One of the pivotal developments which promoted the adaptive radiation in the Animal Kingdom was that of realization of capacity to procure molecular  $O_2$ directly from air in the Paleozoic fish (e.g., Romer 1967; Johansen 1968; Packard 1974; Dejours 1994; Smatresk 1994). Once having attained air breathing, the aquatic animals seized the opportunity for terrestrial habitation. In evolutionary terms, such milestone events have been called evolutionary novelties (e.g., Miller 1949; Mayr 1960; Riedl 1978), key innovations (Lauder and Liem 1989), or broad adaptations (e.g., Schaeffer 1965a). Baum and Larson (1991) defined such monumental events as "derived traits instrumental in acquiring entry into a novel adaptive zone featuring novel selective regimes". Those innovative animals that underwent the transition to air breathing and life on land are by inference thought to have formed the stem reptiles and are all now certainly extinct. The ones caught at the water-air interface include the extant air-breathing amphibi-



Fig. 55. Schematic view showing the evolution of air breathing and transition from sea to land. Also shown are some of the benefits gained and challenges faced. Representative taxa at various levels of respiratory development are given. Air breathing evolved in hypoxic stagnant waters which were claimed by plants to form swamps. The amphibians still rely on water for many of their physiological processes such as respiration and reproduction while reptiles, mammals, birds, and insects, after developing a waterproof cover, rely entirely on internalized gas exchangers for O<sub>2</sub> procurement. Hypoxia was the main driving pressure which prompted sojourn on land

ous forms and the aquatic air breathers (Rahn and Howell 1976). The contemporary transitional (= amphibious = intermediate = bimodal = dual) breathers, animals which are able to exchange gases in both air and water (using the same or multiple organs), however, do not constitute the direct progenitors, i.e., the bridging animals, between the gill breathers and animals with a modern lung (Rahn and Howell 1976). The early stem forms of animals with a derived (modern) lung would have been the Devonian amphibians (Romer 1972). They had a simple lung capable of taking up ample amounts of O<sub>2</sub> and eliminating a substantial measure of metabolic CO<sub>2</sub>, as the gills gradually regressed. The surviving bimodal breathers constitute a contemporary paradigm of the processes and pathways through which air breathing and terrestriality evolved: they occupy a central position in the ecology and physiology of the evolved animal life. There is no physiological or paleontological evidence to indicate that direct passage from water to land has ever occurred (Dejours 1994). The contemporary transitional vertebrate breathers, in particular the lungfishes, comprise a provocative group that occupies a pivotal point in the evolution of the terrestrial tetrapods (Joss et al. 1991; Meyer and Dolven 1992). The adaptive diversities and the extents to which animals went to attain air breathing suggest that the selective pressures that launched the change were extremely severe and pervasive (Carter 1957, 1967). The great flexibility required to adapt to the transitional habitats may explain the dearth of intermediate breathers in many taxa. On exposure to a new habitat, adaptive measures are instituted in the order of behavioral, physiological, and structural shifts, transformations, and modifications. The intensity and the frequency of the changes in the environment and the capacity to adapt to the variations may determine the difference between survival and extinction.

The subject of the evolution of air breathing is of particular interest to both physiologists and morphologists. It presents a model for understanding some fundamental changes in the structure and function of one of the central organ systems in an animal's body. Air breathing was a monumental event in the sequence of different preadaptations for terrestrial habitation, a process which culminated in the emancipation of animals from water (Das 1940; Dejours 1989; Moore 1990b). The archaic fish, e.g., lungfishes (Dipnoi) and the bichirs and three quarters of the modern amphibious fish that inhabit the tropical and subtropical regions (Table 19) breath air (Burggren et al. 1985a; Munshi and Hughes 1992). This suggests that the factors which inspired air breathing may have been most severe in such regions. Fish in the Amazonian basin, the largest freshwater equatorial basin, have been widely studied (e.g., Carter and Beadle 1931; Junqueira et al. 1967; Johansen 1968; Kramer et al. 1978; Stevens and Holeton 1978a; Cala 1987): the majority of them have evolved air breathing. The induction of air breathing in the aquatic breathers is widely ascribed to intrinsic changes in the aquatic environment, especially hypoxia and, by extension, ambient temperature (e.g., Townsend and Earnest 1940; Davis et al. 1963; Tulkki 1965; Moshiri et al. 1979; Kutty and Saunders 1973). High metabolic demands for O<sub>2</sub> and intense putrefactive processes precipitated a hypoxic crisis which was exacerbated by hypercapnia (Johansen 1968; Table 11). This combination of events appears to have constituted a decisive force which prompted search for an alternative source

Species	AROsª	Family	Habitat
Electrophorus electricus	BPA	Electrophoridae	Rivers and swamps – S. America
Opicephalus (=Channa) punctatus, marulius, striatus, gachua	PL	Ophicephalidae	Tropical ponds and rivers – Asia and Africa
Amphipnous cuchia	PL	Ophicephalidae	Tropical ponds and rivers – Asia & Africa
Hvpopomus brevicostris	OC	Sternarchidae	Swamps – S. America
Symbranchus marmoratus	OP	Symbranchidae	Swamps – S. America
Monopterus javanensis	OP	Symbranchidae	Freshwaters – Southern Asia
Pseudapocryptes lanceolatus	OP	Gobiidae	Pools and swamps – S. Asia
Heteropneustes (=Saccobranchus) fossilis	OL	Saccobranchidae	Ponds and swamps – Sri Lanka, India, Burma, Laos, Thailand, and Vietnam
Clarias (C. lazera, C. magur and C. mossambicus)	OL	Clariidae	Ponds and swamps – Africa, S. and W. Asia
Macropodus cupanus	OL	Anabantidae	Tropical ponds – Asia
Colisa fasciata	OL	Anabantidae	Freshwater – S. Asia
Betta	OL	Anabantidae	Freshwater – S. Asia
Osphronemus gorami	OL	Anabantidae	Freshwater – S. Asia
Anabas testudineus	OL	Anabantidae	S. Asia, IndoMalaysian Archipelago, Tropical and S. Africa
Ancistrus (A. anisitsi, A. chagresi)	SM	-	Swamps – S. America
Plecostomus plecostomus	SM	Loricariidae	Swamps – S. America
Misgurnus fossilis	Ι	Cobitidae	Rivers and pools – Europe and Asia
Lepidocephalichthys guntea	Ι	Cobitidae	Freshwaters – Asia
Doras	Ι	Doradidae	Rivers and swamps – S. America
Hoplosternum litorale	Ι	-	Swamps – S. America
Arapaima gigas	SB	Arapaimidae	Swamps and R. Amazon – S. America
Gymnarchus	SB	Gymnarchidae	Swamps and rivers – S. Africa
Ervthrinus unitaeniatus	SB	Characinidae	Swamps – S. America
Umbra	SB	Umbridae	Stagnant waters – Europe and N. America
Notopterus (N. notopterus, N. chitala)	SB	Notopteridae	Freshwater – Asia
Phractolaemus ansorgii	SB	Phractolaemidae	Tropical – W. Africa
Anguilla (A. anguilla, A. bengalensis, A. japonicus)	SK	Anguillidae	Rivers – Europe, Asia, Africa and N. America

**Table 19.** Systematic position, habitat, and types of accessory respiratory organs (AROs) of airbreathing teleosts. (After Dehadrai and Tripathi 1976)

<sup>a</sup> BPA, buccopharyngeal apparatus; PL, pharyngeal lung; OC, opercular chamber; OL, opercular lung; SC, stomach; I, intestine; SB, swimbladder; SK, skin.

of molecular O<sub>2</sub> (e.g., Das 1927, 1940; Saxena 1960; Johansen 1968; Kramer et al. 1978; Kramer 1980; Randall et al. 1981; Davenport 1985). Dramatic spatial and temporal variations in aquatic hypoxia in local inland water masses, especially those formed after flooding, have been a common feature of the Earth's surface since the Cambrian period (e.g., Barrell 1916; Fish 1956; Street and Grove 1976; Jenkyns 1980; Bray 1985; Little 1990). At various geological times, increases in environmental temperatures resulted in low solubility of O<sub>2</sub> especially in the tropical freshwater ponds, prompting the quest for an alternative source of molecular O<sub>2</sub> (e.g., Graham 1949; Grigg 1969; Graham et al. 1978a,b). Drying up of the shallow and extensive continental shelves not only aggravated the respiratory conditions but caused overcrowding and competition for finite resources. Physicochemical changes such as increase in salinity and turbidity must have acted as additional stimuli for abandoning water for land (Sayer and Davenport 1991). Evidently, severe respiratory episodes have occurred frequently in the past. In the Silurian-Devonian periods (e.g., Inger 1957; Berkner and Marshall 1965; Thompson 1971), O<sub>2</sub> levels are thought to have dropped to about 10% compared with the present-day levels (Fig. 9): arising from microbial as well as animal respiratory processes, a reciprocality of environmental O<sub>2</sub> and CO<sub>2</sub> levels occured. Invariably, environmental hypoxia is associated with some degree of hypercapnia especially in standing, plant-infested waters. Hypoxia, especially when accompanied by hypercapnia, constitutes a powerful force which induces air breathing. In the tench, Tinca tinca, hypoxia-hypercapnia reduces routine O<sub>2</sub> consumption and causes a swelling of the erythrocytes (Soivio and Nikinmaa 1981; Jensen and Weber 1985). Though morphologically similar (Godoy 1975), the respiratory physiology of the two ecologically distinct erythrinid fish, Hoplias malabaricus and *H. lacerdae*, which occur in the South American tropical and subtropical shallow waters and streams, is remarkably different (Rantin et al. 1992, 1993): compared with H. lacerdae which lives in well-aerated streams, H. malabaricus, which inhabits stagnant O<sub>2</sub>-deficient environments, is characterized by low metabolism, higher O2 extraction, tolerance to low O2 tensions, low gill ventilation (Rantin and Johansen 1984; Rantin et al. 1992), and large respiratory surface area (Fernandes et al. 1984; Fernandes and Rantin 1985) and high aerobic capacity (Driedzic et al. 1978; Hochachka et al. 1978). The critical O<sub>2</sub> tensions for H. malabaricus is 2.7 kPa and for H. lacerdae 4.7 kPa (Rantin et al. 1992). Around the Gulf of Mexico, during the summer months when the eastwards winds push the surface waters offshore and make the hypoxic deeper water flow inshore (May 1973), the marine crab, Callinectes sapidus, emerges into the air (Loesch 1960). An El Niño effect, a major oceanographic change related to shift in global weather patterns that originated from the eastern Pacific during the winter of 1982-1983 and the spring and summer of 1983, caused massive extensive upwelling of apparently hypoxic warm tropical equatorial water along the shores of South and North America (Cane 1983; Philander 1983) resulting in massive mortality of the Oregon's coho (Oncorhynchus kisutch) and chinook (O. tshawytscha) salmon (Johnson 1988): El Niños have been associated with larval dispersal of and southward displacement of tropical species (DeVries et al. 1997). When exposed to hypoxic water (4 to 5.3 kPa), the freshwater crayfishes, Austropotamobius pallipes (Taylor and Wheatly 1980) and Orconectes rusticus (McMahon and Wilkes 1983),

surface to ventilate their (branchial chambers) "lungs". The air-breathing fish Erpetoichthys calabaricus senses and deliberately avoids hypoxic areas of the water in which it lives (Beitinger and Pettit 1984). No examples of air breathing have evolved in animals which subsist in well-aerated waters such as fast-flowing streams where the  $O_2$  levels are perpetually at or near saturation. Krogh (1941) considered the transition to air breathing to have been stimulated by "emergency respiration" resulting from withdrawal of water (from which animals extracted  $O_2$ ) from the intertidal animals. From the morphological similarity and close topographical relationships between the gills and the accessory respiratory organs (both are located around or open in the pharynx), the evolution of air breathing appears to have been a carefully crafted slow process. It did not involve an ovehaul of the gills but a gradual phasing out. In the mud-eel Amphipnous cuchia, for example, the air sac which develops from the gill arches 2 to 5 and the ectoderm cells (derived from the integument bordering the pharyngeal openings) receives venous blood through the afferent branchial vessels of the second and third gill arches and the vascular papillae which develop in the buccal cavity and in the air sacs exchange gases with air (Singh et al. 1984).

The exigency to procure O<sub>2</sub> directly from air has evolved many times in response to different environmental pressures (e.g., Randall et al. 1981; Graham 1994). In *Polpterus bichir*, the gas bladder functions as an accessory respiratory organ only when the O<sub>2</sub> content of the water is low (Budgett 1900). A few airbreathing fish, however, show anomalous air-breathing behavior. About 40 marine species in 6 families regularly breath air in a habitat which is not characterized by hypoxia or hypercarbia (Graham 1976). In some species of aquatic bimodal breathers, e.g., young tarpons, Megalops atlantica, air breathing is socially regulated (Böhlke and Chapline 1968; Kramer and Graham 1976). Interestingly, not all the present-day bimodal air breathers are phylogenetically ancient forms which have survived. This indicates that the quest for air breathing is an ongoing covert process which is being aggressively pursued by some of the contemporary aquatic animals especially those which experience hypoxia or sudden fluctuations of  $O_2$  in their habitats. Oxygen is both an important factor for aerobic metabolism and a necessary resource in growth and development. Bader (1937) demonstrated that normal development of the accessory respiratory organs of Macropodus (Belontiidae) was hindered if the fish was refused access to air. It has been interestingly postulated that by reducing the risk of desiccation through respiratory water loss, atmospheric hyperoxia, as occurred during the mid-Devonian and Carboniferous periods (Fig. 9), may have enhanced terrestrial habitation: breathing hyperoxic air reduces respiratory frequency and hence lowers respiratory water loss (Withers 1992). The evolution of air breathing, however, did not have a direct causal relationship with terrestrial colonization (e.g., Carter and Beadle 1931). Occupation of land happened to offer one of the many solutions to the prevalent respiratory stress in water. This is evinced by, among others, the air-breathing fish which even after acquiring a significant preadaptation for terrestriality (i.e., capacity to breathe air) permanently live in water. In water, a highly dynamic habitat, animals are behaviorally and physiologically adapted to cope with sudden changes in O<sub>2</sub> availability, temperature, and salinity (Horn and Gibson 1989): at high tide they take O<sub>2</sub> from the water and during low tide from air. The dragon-fly larva surfaces to breath when the PO<sub>2</sub> in water falls below 7.3 kPa (Wallengren 1914). Similar behavior occurs in the fish, *Leuciscus erythrophthalamus*, when the PO<sub>2</sub> falls to below 2 kPa (Winterstein 1908). Behavioral, functional, structural, biochemical, and molecular changes, plausibly in that order, were utilized to accommodate aquatic hypoxia as animals switched from water to air breathing (Hiebl et al. 1987; Weber et al. 1993) and then gravitated towards land. The gills, the archetype aquatic gas exchangers honed for respiration in water proved deficient in air, a physically remarkably different medium (Tables 4,9). The gills were gradually phased out as extraction of O<sub>2</sub> from water was deemphasized and air breathing consolidated. In terrestrial crabs (Sect. 5.6.2), a great deal of this change entailed expansion of the branchial cavity, modification of the branchial epithelium, and mechanical ventilation of the same. In the pneumonate gastropods (Sect. 5.6.1), the ctenidia in the mantle cavity was replaced by a lung.

### 5.2 Strategies and Adaptive Convergence for Air Breathing

The intermediate breathers use the accessory respiratory organs or lungs to procure about two-thirds of their O<sub>2</sub> needs and eliminate only about one quarter of the metabolic CO<sub>2</sub>. The nonpulmonary gas exchanger(s) void roughly three fourths of the metabolic CO<sub>2</sub> irrespective of whether the animal lives in water or air (Rahn and Howell 1976). Some of the primary attributes of an accessory respiratory organ are that: (1) it must possess regular or irregular means for renewal of air, (2) the gas voided from the organ must contain less  $O_1$  and more  $CO_2$  compared with the inspired (atmospheric) air, and (3) the epithelial surface must (in most cases) present conspicuous morphological modifications such as good vascularization and/or surface amplifications. An inverse correlation occurs between the level of commitment of an accessory respiratory organ to air breathing and the degree of regression of the alternative respiratory sites such as the skin, buccal cavity, and gills. The capacity of air breathing in many animal groups and the remarkable uniformity of the morphological, physiological, and biochemical features inaugurated in so phylogenetically different taxa is a model case of convergent evolution. Those features which are common to a wide crosssection of animals contributed the foundations to the comprehensive gas exchange process and those traits unique to a particular group present a solution to a specific problem. By identifying the pressures to which the animals were subjected, by analogy, reconstruction of the events which initiated and sustained the momentous change can be made. There is now ample evidence indicating that the need to breath air evolved essentially because the O<sub>2</sub> levels in water were unstable and largely low (e.g., Carter and Beadle 1931; Schmalhausen 1968; Randall et al. 1981). Due to the remarkable differences in the physiochemical properties water and air (Sect. 3.2), the shift drastically affected the form and function of practically all the biological processes (e.g., Edney 1960; Young 1972; Mangum 1982b; Greenaway et al. 1983; Bridges 1988; Val et al. 1990; Morris 1991; Morris and Bridges 1994). These included locomotion, respiration, reproduction, excretion,

and sensory perception of external stimuli. The impact was, however, greatly minimized since need for direct switch from water (a fairly stable environment) to air (land), a highly fluid one, rarely arose. The aquatic and terrestrial habitats intergraded extensively especially during the formative years of the continental land masses when massive uplifting, subsidence, and displacement of the plates caused dramatic shifts in the sea level and flooding of land (Takeuchi et al. 1970; Ben-Avraham 1981; Fig. 55). The intermediate zone had adequate water vapor pressure stability to sustain the development of the accessory respiratory organs without the risk of desiccation in predominantly aquatic animals. In such humidic habitats, the animals adapted to hypoxia tolerance in water and instituted measures for air breathing and terrestrial existence. Among the erythrinids (Order: Cypriniformes) which inhabit shallow O<sub>2</sub>-poor freshwaters of the tropical and subtropical regions of South America (Dickson and Graham 1986), Hoplias malabaricus, which lives in stagnant hypoxic water, is more tolerant to hypoxia (threshold PO<sub>2</sub> for onset of bradycardia = 2.6 kPa) than *H. lacerdae*, which inhabits well-oxygenated rivers (threshold  $PO_2$  for onset of bradycardia = 4.7 kPa; Rantin et al. 1993). Factors such as the larger respiratory surface area (320 mm<sup>2</sup> perg; Fernandes et al. 1984) and high O<sub>2</sub> affinity of blood (Wood and Lenfant 1979) in Hoplias malabaricus compared with H. lacerdae may explain the differences in hypoxia tolerance in the two species. Some erythrinid fish, e.g., Hoplererythrinus unitaeniatus and Erythrinus erythrinus (Randall et al. 1981), have adopted air breathing. Surface skimming for air and/or well-oxygenated top water layer is a common strategy of overcoming hypoxia in the tropical freshwaters (Kramer and McClure 1982). Gulping air at the surface elevates O<sub>2</sub> transport during aquatic hypoxia in the goldfish, Carassius auratus (Burggren 1982a). Of the 20000 or so species of fish, only a relatively small number has evolved the capacity for air breathing (e.g., Dehadrai and Tripathi 1976; Sayer and Davenport 1991; Graham 1994; Table 19). An even much smaller number has adopted terrestrialness. Clearly, air breathing and subsequently shift to land were attempted at the extremes of circumstances, e.g., at the threshold of failure of normal O<sub>2</sub> transfer and/or when there were particular benefits to be derived from the shift. The least drastic and most economical solution to air breathing in the Devonian fish would have been to utilize the gills, the highly engineered aquatic gas exchangers, for procuring O2. The probable sequence of events utilized for adaptation to hypoxia should have entailed: (1) physical avoidance of it by relocating to more hospitable areas (Whitmore et al. 1960; Cook and Boyd 1965; Costa 1967; Gamble 1971), and (2) skimming the top 1 to 2mm surface of water which is richer in O<sub>2</sub> (Burggren 1982a; Kramer and McClure 1982; Barton and Elkins 1988; Horn and Gibson 1989), gulping in air, e.g., in the gobies (Gee 1976; Graham 1976; Gee and Gee 1995) and holding it in vascularized buccal and/or pharyngeal cavities (Coutant 1987; Kramer 1987). In the interim, long-term physiological tolerance to hypoxia would have become established. In the bivalve mollusk, Pholas dactylus, air gaping occurs in the laboratory after a drop in PO<sub>2</sub> in the surrounding water and in nature during low tide (Knight 1984). The Amazonian freshwater ray (Paratrygon sp.) surfaces when the PO<sub>2</sub> drops to below 2kPa and utilizes the O<sub>2</sub>-rich surface water (Steen 1971). Increased CO<sub>2</sub> in water depresses branchial respiration and stimulates air breathing. Like the fossorial mammals and birds (Sect. 3.7), fish which live in derelict waters are less sensitive to  $CO_2$  than those from well-aerated ones (Hughes 1963). It has been argued that it was not lack of molecular  $O_2$  per se but factors such as availability of new food sources, reproductive needs, escape from predators (Little 1990; Sayer and Davenport 1991), and the enormous energetic advantages derived from the switch (Bennett 1978; Fig. 56) which led to the development of air breathing, especially in the shore tidal areas. The gastropod mollusk, particularly those of the family Ampullariidae, provide excellent examples of the extents to which animals went to attain and maintain an air-breathing capacity. Some, e.g., the amphibious prosobranch, *Pomacea depressa*, which lives in the Everglades swamps in Florida, a rather extensive stagnant mass of warm water where intense organic putrefaction of the massive vegetational growth occurs, have evolved a divided mantle cavity with a gill in one half, the other half acting as a lung (McClary 1964; Little 1990). Such animals are able to breathe air and water simultaneously or switch from one medium to another depending on needs and circumstances.

Water characteristically constitutes an environment where O<sub>2</sub> is scarce (Table 4). In some habitats, survival is compounded by the presence of a high concentration of CO<sub>2</sub> and other gases such as H<sub>2</sub>S and NH<sub>3</sub>. Owing to the effect of CO<sub>2</sub> on the central control, O<sub>2</sub> consumption falls drastically with rising PCO<sub>2</sub> particularly when the  $PO_2$  is low (Saxena 1962; Tenney 1979). When exposed to hypoxic water, Gillichthys gulps air as its oxidative metabolism decreases (Todd 1971). In Pseudapocryptes (Das 1934), hypoxia induces air breathing and in Tomicodon (Eger 1971), stagnation of water elicits air breathing. When held in hypoxic water, Clarias and Heteropneustes show metabolic rates which are 60% below normal (Hughes and Singh 1970b; Singh and Hughes 1973). Depending on the ambient temperature and level of activity, the bowfin, Amia calva, an ancestral halecomorph North American actinopterygean fish which is an active swimmer and subsists in an environment which is cold and covered with ice in winter and warm in summer, relies on the gills and a vascularized gas bladder for gas exchange (Johansen et al. 1970a; Liem 1987a; Hedrick and Jones 1993): at temperatures between 10 and 30 °C, O<sub>2</sub> consumption is shared equally between the gills and the air bladder, at above 30 °C the air bladder accounts for more than two thirds of the  $O_2$  consumption while at 10 °C and below, the gills meet all the  $O_2$ needs. Breath holding can last from 5 to 55 min depending on temperature, amount of light, and the O<sub>2</sub> concentration in the water (Liem 1987a). Ambient temperature (Burggren et al. 1983) and PO<sub>2</sub> (Burggren and Mwalukoma 1983; Burggren and West 1982) influences the pathway utilized for procuring O<sub>2</sub> in the larval Rana berlandieri and R. catesbeiana.

The development of the amphibians from eggs through tadpoles to airbreathing adult forms is accompanied by dramatic changes in the respiratory strategies (Fig. 47) and offers a highly instructive model in understanding the change from water to air breathing and transition from water to land. The amphibian eggs acquire  $O_2$  entirely by diffusion across the surface, the tadpoles initially have external gills which are followed by internal ones, and later in life the adults develop fully functional lungs. Metamorphosis starts in water, a medium from which  $O_2$  is less available, with the animal proceeding to the much better oxygenated aerial habitat. The access to a more  $O_2$ -rich medium is accompanied by a



certain costs such as adaptations for reproduction independent of water, more efficient capacity for osmoregulation, and excretion of products of and intertidal pools). While the prompting factors differed from habitat to habitat, the pervasive features which prompted air breathing and shift to land included decreasing O<sub>2</sub> and food availability in isolated, shrinking aquatic habitats. In this complex and highly dynamic process, other factors included increasing temperatures, CO<sub>2</sub>, and pH. While definite rewards were reaped such as access to abundant O<sub>2</sub> and newer resources, Fig. 56. Principal biotic and abiotic factors which elicited emergence behavior from water by amphibious fish in closed systems (e.g., freshwater nitrogen metabolism, e.g., urea and uric acid, were called for. (Sayer and Davenport 1991 $\overline{)}$  decrease in the  $O_2$  affinity on metamorphosis from a tadpole to an adult amphibian (Broyles 1981; Burggren and Wood 1981). The  $P_{50}$  of the blood of the tadpole of a bullfrog, *Rana catesbeiana*, is 0.7 kPa (McCutcheon 1936), a PO<sub>2</sub> at which in the adult frog the blood is only 5% saturated. After giving rise to the successful amniotes, the amphibians have literally lingered on with one foot in water and the other on land. Though the first animals to conquer terra firma, owing to their reliance on water for crucial physiological processes such as reproduction, excretion of waste products, osmoregulation, and gas exchange, in general, the amphibians, a group recalcitrant to change, are a defeated group. They constitute an inconspicuous taxon among the extant tetrapods. The contemporary amphibians can be looked on as a relic of an evolutionary stage between air-breathing fish on one hand and reptiles, the first animals with a true lung, on the other.

### 5.3 Risks, Costs, and Benefits in the Change to Air Breathing

In changing residence from water to land, animals faced fundamentally different environmental and physiological challenges (Fig. 7). They had to procure O<sub>2</sub> from air and void CO<sub>2</sub> into the same, avoid desiccation, store or excrete different toxic products of nitrogen metabolism, somewhat regulate body temperature, and avoid or confront completely new predators. Whereas aquatic habitats offer rather stable and predictable "climatic" features, terrestrial ecosystems show greater spatial and temporal diversity. The terrestrial macrophytes, particularly gymnosperms and angiosperms, generate remarkably different microclimates over short distances (e.g., Geiger 1965). In order to derive the best of two worlds, the aquatic (water-residing) air breathers have chosen to physically remain in water and periodically surface to extract O<sub>2</sub> from the air. Such animals lack specializations like means for aerial vision, terrestrial locomotion, adaptations to curtail cutaneous water loss, and specific means for elimination of products of nitrogen metabolism in form of urea and uric acid, as occurs in amphibious fish (e.g., Gordon et al. 1978; Table 9). Although most aquatic animals will cope with hypoxia by appropriate microhabitat selection, this strategy is not very effective in dealing with long-term (i.e., diurnal and seasonal) fluctuations in O2 levels. In such cases, permanent solutions are obligatory. Aquatic animals, especially those which live in highly dynamic microhabitats such as tidal pools, thermoclines, or shallow coastal waters display different mechanisms for coping with the extremes of hypoxia. Though indirect, the effect of temperature on respiration is farreaching. Elevated temperature leads to reduced solubility of O2 in water and increased metabolic rate. In a hypoxic condition, raised temperature makes life highly precarious, and access to the atmosphere, where O<sub>2</sub> is available in large quantities, becomes a necessity (Serfaty and Gueutal 1943). In the freshwater fish, temperature preference increases with exposure to hypoxia (Bryan et al. 1984; Schurmann et al. 1991). By selecting a lower temperature, the animal takes advantage of reduced metabolism and increased blood O2 affinity (Schurmann and Steffensen 1992). Similar strategies have been adopted by the reptiles (Hicks and Wood 1985) and salamanders and crayfish (Dupré and Wood 1988). Owing to the

more stable  $PO_2$  in air, habitat relocation in a heterothermal environment with changing  $O_2$  saturations is particularly important in aquatic ectotherms, especially in coastal and standing waters. When the circulation of water is limited or respiratory demands of aquatic organisms are high, anaerobiosis may result. In the bowfin, *Amia calva*, air-breathing frequency increases with aquatic hypoxia (Johansen et al. 1970a). This also occurs in most other air-breathing fish (e.g., Shelton et al. 1986). Due to factors such as different capacities to tolerate hypoxia and the variability in the levels of hypoxia in different habitats, the physiological traits acquired during evolution of air breathing appear to have followed different and independent pathways which were dictated entirely by need. The designs of the contemporary gas exchangers and the existing respiratory adaptations cannot be accurately used to discern the systematic affinities between taxa.

The transition to air breathing and, subsequently, terrestrialness called for profound changes in the respiratory strategies. Animals aggressively mobilized resources and changed their habits and habitats with the specific goal of acquiring O<sub>2</sub> from "above". Switching from one respiratory medium to another and from one gas exchange pathway to another enabled the transitional breathers to utilize the most convenient and efficient method(s) for extracting O<sub>2</sub> from the alternative respiratory media. This provided the necessary flexibility to optimally meet the changing conditions. Plainly, the evolution of air breathing was not ipso facto the attainment of terrestriality. Procurement of molecular O, directly from air, a medium eloquent of the gas, was simply the immediate, most convenient, and permanent solution for surmounting the critical problem of hypoxia prevailing in the aquatic biotope. Residence on terra firma, a venture first attempted by the now extinct rhipidistian crossopterygians some 350 million years ago (in the Upper Devon) (Pough et al. 1989), was a costly, risky process which was approached parsimoniously using different strategies (Figs. 57,58). It had to await decisive preadaptions which included: (1) redesigning of the existing gas exchangers, (2) development of an impermeable surface cover, (3) solution of problems of acid-base and osmotic balance, and (4) development of more appropriate ways and means of eliminating products of nitrogen metabolism (e.g., Little 1990). Furthermore, animals had to cope with factors such as thermal instability, exposure to new predators, and locomotory problems resulting from the greater gravitational effect on the body. Air breathing and subsequently relocation to land arose when and only if there was absolute need for it or where tangible advantages and rewards, e.g., acquisition of more livable space and greater ecological opportunities, were to be reaped to offset the enormous risk and cost. Due to the abundance of  $O_2$  in the free air, when expressed in terms of ventilatory requirement per unit of O2 consumption, an air breather expends much less energy to extract an equivalent volume of O<sub>2</sub> compared with an aquatic one (Dejours et al. 1970; Sect. 2.9). When subjected to intra- or interspecific competition, amphibious fish, e.g., the climbing perch, Anabas testudineus and Monopterus albus, will embark on overland excursions (Liem 1987b). The highly aerial behavior of the pearl blenny, Entomacrodus nigricans, is thought to have arisen as a result of competition amongst the intertidal fish (Graham et al. 1978a). Air breathing and subsequent transfer to land opened new ecological opportunities which resulted in remarkable adaptive radiation. Among the marine teleost fish, the family Gobiidae, which has extensively evolved air breathing (Lewis 1970), contains the largest number of the present-day species in this taxon. Generally, the obligate air-breathing fish, which, due to greater access to  $O_2$ , are apparently more agile and can hence acquire more nutriments, are larger (25 to 30 cm) than the facultative ones (at 7 to 15 cm) (Munshi and Srivastava 1988).

The extant bimodal breathers provide modern analogs of the transitional animal forms in evolution of air breathing. In the lungfish, contact with air is a more powerful stimulus than a tactile or painful stimulus (Johansen and Lenfant 1968). The intertidal clingfish, *Sicyases sanguineus*, increases the number of exposures to air as the dissolved concentration of  $O_2$  decreases (Ebeling et al. 1970). As the PO<sub>2</sub> in water drops, *Mnierpes macrocephalus* spends 92% of the time out of water, returning briefly at regular intervals (Graham 1970). The shanny, *Blennius pholis*, totally avoids water at low partial pressures of  $O_2$  (Davenport and Woolmington 1981). The catfish, *Eremophilus mutisii*, uses its accessory respiratory organ (the stomach) in both normoxic and hypoxic water (Cala 1987) by periodically dashing to the surface to swallow air. In fish which surface more often in normoxic than hypoxic water, air breathing may be utilized for buoyancy control rather than gas exchange (e.g., Gee 1976; Gee and Graham 1978). Since the water breathers have evolved definite mechanisms for regulating hydrogen ions in face of high



Fig. 57. Change from gill system in fish to the modern lung in the higher vertebrates. The transitional animal forms went through stages of bimodal gas exchange with a large gas exchange ratio initially in the gill-skin system and a low ratio initially in the primitive lung (air organ). During this stage, the skin was naked and acted as a "bridging" respiratory organ. Such animals were highly susceptible to dehydration on land. The development of surface covers like scales, feathers, and hair had to await the development of the modern lung. In essence, the switch over from water to air breathing was made very slowly and cautiously. *R.Q.* Respiratory quotient. (Rahn and Howell 1976)



Fig. 58. Suggested pathways for evolution of air breathing, the increasing organizational complexities of organisms, and the corresponding sophistications of the gas exchangers. Endothermy, as evolved in birds and mammals, could only be supported by ventilated and perfused aerial gas exchangers. (Perry 1989)

ambient concentration of CO<sub>2</sub>, deficiency of O<sub>2</sub> rather than elevation of CO<sub>2</sub> appears to have been the most important stressor in the evolution of air breathing. Changing pH levels and/or raising CO<sub>2</sub> levels do not elicit emergence from water in Boleophthalmus pholis (Davenport and Woolmington 1981). Low ambient PO<sub>2</sub> does not cause Monopterus albus and Anabas testudineus (Liem 1987b) or the mangrove forest fish, Rivulus marmoratus (Abel et al. 1987), to immerse their bodies into water. In the bimodally breathing fish, the accessory respiratory organs are located away from contact with water. In this way, such organs can be utilized simultaneously with the gills. The bimodal breathers avoid desiccation and CO<sub>2</sub> accumulation by subsisting in water while exploiting air as a source of O<sub>2</sub>: they hence enjoy the benefits which accrue from the two habitats. While movement to land may be induced by a definite need to feed or may simply be behavioral and purposeless (e.g., Gordon et al. 1978; Sacca and Burggren 1982), abiotic factors such as high  $CO_2$  and  $NH_3$  levels and biotic ones such as predation and interspecific aggression may, to varying extents, prompt migrations to land (Fig. 8). Compared with the many complex integrated processes which were essential for terrestrial habitation (Little 1990), though an important initial step, the attainment of air breathing was a rather simple and direct affair.

# 5.4 CO<sub>2</sub> Elimination: Impediment to Evolution of Air Breathing and Terrestriality

Oxygenation is the primary purpose of respiration and in case of conflict, CO<sub>2</sub> clearance is unequivocally subordinate (e.g., Dejours 1988). Adaptations favoring O<sub>2</sub> uptake are automatically adopted in preference to those aiding CO<sub>2</sub> elimination. In this sense,  $O_2$  plays a more central role in the process of gas exchange. In the bimodal breathers, while the accessory respiratory organs supply O2 to the body, they are unable to eliminate  $CO_2$  as efficiently (e.g., Randall et al. 1978a). Indeed, this particular limitation appears to have been a fundamental obstacle in the transition from water- to air breathing as the modern lung evolved (e.g., Gans 1970, 1971; Withers 1992; Olson 1994; Fig. 57). Even after some bimodal breathers could secure as much as 96% of their O, needs from the air, the skin and/or the gills continued to be the major organ(s) for CO<sub>2</sub> elimination (Tables 20,21). This restricted such species to water. In the totally land-based solely air-breathing crustaceans, appropriation of the enzyme carbonic anhydrase into the membrane fraction of the branchiostegites may have been one of the vital molecular events which enabled pulmonary CO<sub>2</sub> elimination into the air (Morris and Greenaway 1990; Henry 1994). During activity, in the highly terrestrial coconut crab, Birgus latro, CO<sub>2</sub> accumulates in the hemolymph if the gills are surgically removed (Smatresk 1979). Since  $CO_2$  is not a highly toxic molecule, at least not as much as O<sub>2</sub> (Sect. 1.18), and concentrations can moderately rise without causing irreparable physical damage, regulation of the concentration of CO<sub>2</sub> in the body is utilized strictly to adjust body pH and not CO<sub>2</sub> levels. Unlike the charged molecules such as  $HCO_3^-$  and  $H^+$  ions, which are transferred through ionic exchange for Cl<sup>-</sup> and Na<sup>+</sup> ions, respectively, the uncharged molecular CO<sub>2</sub> easily moves

Taxon	Species	T	Lung (%)	Lung (%)		Skin (%)	
		(°C)	VCO <sub>2</sub>	VO <sub>2</sub>	VCO <sub>2</sub>	VO <sub>2</sub>	
Urodela Anura	Ambystoma maculatum	25	29	69	71	31	
	Taricha granulosa	25	32	68	68	32	
Anura	Rana clamitans	25	24	74	76	26	
	Rana pipiens	25	43	68	57	32	
	Hyla gratiosa	25	19	68	81	32	
	Hyla versicolor	25	16	67	84	33	
	Bufo americanus	25	17	66	83	34	
	Bufo boreas	25	31	69	69	31	
	Bufo cognatus	25	30	44	70	56	
	Bufo marinus	25	24	50	76	50	
	Bufo terrestris	25	25	78	75	22	
Mean			26	65	74	35	

Table 20. Partitioning of gas exchange between the lung and the skin system in terrestrial amphibia. (After Rahn and Howell 1976)

**Table 21.** Gas exchange partitioning between water-breathing organs (gill, G and skin, S) and air-breathing organs of some air-breathing fish. (After Singh 1976)

T (°C)	Species	VO <sub>2</sub> (%)			VCO <sub>2</sub> (%)		
		From water	Organ	From air	Into water	Organ	Into air
20	Amia calva	63	G	37	75	G	25
25	Heteropneustes						
	fossilis	59	G,S	41	94	G,S	6
25	Anabas testudineus	46	G	54	91	G	9
25	Clarias batrachus	42	G,S	58	94	G,S	6
22	Lepisosteus osseus	27	G	73	92	G	8
23	Monopterus albus	25	G,S	75	-	-	-
26	Electrophorus						
	electricus	22	S	78	81	S	19
25	Cobitis fossilis	20	-	80	34	-	66
20	Protopterus						
	aethiopicus	11	G	89	70	G	30
24	Protopterus						
	aethiopicus	10	G	90	68	G	32
20	Lepidosiren (Juvenile)	64	G,S	36	76	G,S	24
20	Lepidosiren (Adult)	4	G,S	96	41	G,S	59

across the cell membranes. During the transition from water to land, the skin, though often considered evolutionarily a dead end or a failed experiment, functionally acted as an important respiratory/acid-base bridging organ during the development of air breathing and subsequently terrestrial habitation (Fig. 57).

With the gradual involution of the gills, ionic exchange and CO<sub>2</sub> clearance were translocated to the skin (Rahn 1967; Randall et al. 1981). In amphibians, blood flow to the mid-dorsal skin is 1.8 times that to the ventral thoracic skin (Moalli et al. 1980). This indicates that the former site may be more important in CO<sub>2</sub> excretion than the latter (Talbot and Feder 1992). In the contemporary bimodal breathers, the gill-skin system removes about 76% and the lung 24% of CO<sub>2</sub> while O<sub>2</sub> uptake varies with the species, the gas exchanger utilized, and habitat occupied (Emilio et al. 1970; Rahn and Howell 1976). In aquatic amphibian species, Siren *lacertina* and *Amphiuma means*, the lung takes up 65% of the O<sub>2</sub> needs but the gills and/or skin eliminate nearly 75% of the CO<sub>2</sub> (Guimond and Hutchison 1973a, 1976). The utilization of the skin for gas exchange in many animals in different habitats attests to its great importance of having served as a bridging organ during the development of the aerial gas exchanges. To compensate for its inherent limitations as a gas exchanger, cutaneous respiration is much less energetically expensive (Feder and Burggren 1985a,b). Interestingly, though the marine air-breathing fish do not have very well-developed accessory respiratory organs, they appear to have developed the capacity to release  $CO_2$  into the air so efficiently that most of them will maintain the same level of metabolism when exposed to air (Graham 1976).

Relatively, the absolute values of  $PCO_2$  and  $HCO_3^-$  in blood and tissues are lower in the water breathers than in the air breathers. Weighted against the air breathers, generally, challenges for acid-base balance are less severe in water breathers (e.g., Driedzic and Gesser 1994). In the plasma of fishes, PCO<sub>2</sub> ranges between 0.1 to 0.5 kPa and  $HCO_3^-$  from <5 to 15 mM (Heisler 1984). Fish gills contain a high concentration of carbonic anhydrase which catalyzes breakdown of plasma HCO<sub>3</sub><sup>-</sup> ions to CO<sub>2</sub> (e.g., Haswell and Randall 1978) with a small amount being voided across the kidneys (e.g., Wood and Cadwell 1978). About 90% of the total CO<sub>2</sub> in Anabas, Clarias, and Heteropneustes is voided through the gills (Hughes and Singh 1970a,b, 1971): the ratio of aquatic to aerial CO<sub>2</sub> removal is 10:1. Characteristically, the drop in pH induced by hypercapnia is corrected by elevation of HCO<sub>3</sub><sup>-</sup>. Exposure of Conger conger and Scyliorhinus stellaris to an environmental PCO, of 1 kPa lowers plasma pH by about 0.4 units as plasma PCO, rises from about 0.25 to about 1.3 kPa (Heisler 1982b): over a period of 5 to 10h, the pH was partially corrected by an elevation of  $HCO_3^-$  from <10 to about 20 mM. In exercising S. stellaris, H<sup>+</sup> released from skeletal muscle caused the arterial pH to drop from 7.8 to 7.2, the PCO<sub>2</sub> to rise from about 0.25 to 0.67 kPa, and the HCO<sub>3</sub><sup>-</sup> to drop from 7 to 3mM (Holeton and Heisler 1983). In the facultative air-breathing fish, Symbranchus marmoratus, change from water to air breathing alters PCO<sub>2</sub> from about 0.75 to 3.5 kPa with an accompanying drop in pH of about 0.6 units and a fourfold increase in intracellular HCO<sub>3</sub><sup>-</sup> (Heisler 1982b). Vertebrate air breathers excrete much of their CO<sub>2</sub> in molecular form across the lung although a little of it is removed as HCO<sub>3</sub><sup>-</sup> ions through the kidneys (Boutilier et al. 1979a,b). In fish, the processes of CO<sub>2</sub> elimination in pH regulation are different from those of mammals: HCO<sub>3</sub><sup>-</sup> ions regulation, rather than molecular CO<sub>2</sub>, is the more important factor in the process (e.g., Cameron 1978). Unlike in mammals, in fish, ventilation affects the  $CO_2$  concentration of blood and hence pH only in the extremes of circumstances, e.g., during hyperoxia when ventilation is suppressed (Randall and Jones 1973) or during extreme hypoxia (Dejours 1973). Fish have evolved in a demanding habitat naturally deficient in  $O_2$ . Ventilation has been configured solely for delivery of  $O_2$  to the gills, rendering  $CO_2$  clearance somewhat of secondary importance.

The transition to air breathing and residence on land presented a challenge towards acid-base balance. It necessitated an increase in blood PCO<sub>2</sub> and a corresponding increase in plasma HCO<sub>3</sub><sup>-</sup> ions for maintenance of normal pH (e.g., Rahn 1966; Hughes 1966; Howell et al. 1970; Lenfant and Johansen 1972; Tables 2,22). Plasma PCO<sub>2</sub> tends to increase as an animal shifts from water- to air breathing (Driedzic and Gesser 1994) though there is no clear correlation between the mode of breathing and the total CO<sub>2</sub> (Toews and Boutilier 1986). The high capacitance coefficient of CO, in water compared with O, curtails the ability of aquatic animals to regulate internal acid-base status by ventilatory adjustments of blood PCO<sub>2</sub> (e.g., Cameron 1979). In aquatic breathers, pH control is largely effected by exchange of acidic or basic equivalents between the extracellular fluid and the environment. This leads to changes in plasma HCO<sub>3</sub><sup>-</sup> ion levels. In transitional breathers, ventilation does not correspond with CO<sub>2</sub> levels in blood as in the air breathers. The significance of ventilation in pH regulation is of little importance as the gills and the skin are the main pathways for CO<sub>2</sub> excretion. Fish appear to possess a mechanism for enzymatically curtailing CO<sub>2</sub> and H<sup>+</sup> ion loss across the gill epithelium (e.g., Haswell and Randall 1978) with the respiratory rate being mainly responsive to the PO<sub>2</sub> in the arterial blood (e.g., Randall et al. 1981). On the other hand, in the air breathers, CO<sub>2</sub> has to be voided before it accumulates to the level where the lowered pH interferes with the O<sub>2</sub> binding properties of the hemoglobin. Elimination of CO<sub>2</sub> across the vertebrate lung and hence the significance of ventilation in pH regulation was enforced onto the lung with the decrease in the CO<sub>2</sub> diffusing capacity of the gills. This resulted in high levels of CO<sub>2</sub> in blood (Randall et al. 1981). The increase in the permeability of the erythrocytes to HCO<sub>3</sub><sup>-</sup> ions comprised a further means of enhancement of CO<sub>2</sub> clearance through the lungs.

Taxon	T (°C)	PIO <sub>2</sub> (mmHg) <sup>a</sup>	PEO <sub>2</sub> (mmHg) <sup>a</sup>	PbO <sub>2</sub> (mmHg) <sup>a</sup>	PbCO <sub>2</sub> (mmHg)ª	pHb	$(HCO_{3}^{-1})b$ (meq l <sup>-1</sup> )
Water breathers		,					
Lugworm Arenicola marina	19	160	35	_	1.7	7.3	1.6
Octopus dofleini	11	127	94	78	3.1	7.1	-
Crab Carcinus maenas	10	156	-	-	2.3	7.8	7.2
Cancer magister	8	141	97	75	1.7	7.9	-
Dogfish Scyliorhinus stellaris	16	149	56	49	2.0	7.8	-

**Table 22.** Some respiratory variables in some water and air breathers. (After Dejours 1988, from where individual sources of data should be consulted)

Taxon	T (°C)	PIO <sub>2</sub> (mmHg) <sup>a</sup>	PEO <sub>2</sub> (mmHg) <sup>a</sup>	PbO <sub>2</sub> (mmHg) <sup>a</sup>	PbCO <sub>2</sub> (mmHg) <sup>a</sup>	pHb	$(HCO_{3}^{-1})b$ (meq l <sup>-1</sup> )
Transitional breather	:s						
Frog							
Rana catesbeiana	22	150			4.2	7.0	6.0
A dult	25 23	150	-	_	4.3 17.2	7.8 7.8	0.8 29
Salamanders	20	150			17.2	,	2,
Cryptobranchus alleganiensis	25	154	-	27	6.2	7.8	-
Desmognathus fuscus	19	150	-	40	7.9	7.4	9.2
Lungfish							
Neoceratodus	18	131	-	40	4.7	7.6	5.3
Protopterus aethiopicus	25	130	-	27	26	-	-
Gar-fish	25				10	- 4	10.2
Lepisosteus osseus	25	155	_	-	13	7.4	10.2
Electric eel Electrophorus electricus	27	150	-	12	27	7.6	30
Amphipnous cuchia	25	155	-	-	12	7.5	-
Air breathers							
Snail							
Otala lactea	15	154	144	144	8.6	8.1	-
Crab							
Gecarcinus lateralis	21	155	-	-	8.9	7.5	11.4
Uca pugilator	20	155	-	-	10.6	7.7	21.2
Birgus latro	29	150	-	27	9	7.7	12.7
Turtle Pseudemys scripta	25	155	-	-	27.4	7.6	33.0
Pekin duck	41	144	10.8	91	33	7.5	23
Toad (Adult)	-		-			-	
Bufo marinus	25	155	-	80	11.1	7.8	21.4
Human being	22	149	119	93	20.4	7.7	-

## Table 22. Continued

Symbols: PIO<sub>2</sub> and PEO<sub>2</sub> partial pressure of O<sub>2</sub> in the inspired and expired air/water; PbO<sub>2</sub> and PbCO<sub>2</sub>, partial pressures of O<sub>2</sub> and CO<sub>2</sub> in the arterial blood; pHb, acidity of blood;  $(HCO_3^{-1})$  bicarbonate concentration in blood.

<sup>a</sup> To convert ot kPa multiply by 0.133.

In order to disengage the primary functions of the gills (respiration, ionic exchange, and pH regulation), a transitional respiratory stage was necessary (Fig. 57). The accessory respiratory organ was charged with  $O_2$  uptake and the gills or any other respiratory surface, e.g., the skin and the buccal cavity with CO<sub>2</sub> removal and hence pH regulation and ionic homeostasis. A relocation of CO<sub>2</sub> excretion to the accessory respiratory organs had to await development of adequately efficient gas exchangers and ventilatory mechanisms. Osmoregulation and excretion of nitrogenous waste products was consigned to the kidneys (Schwartz 1976), the urinary bladder (Sachs 1977), and to a lesser extent to the skin (Ehrenfeld and Garcia-Romeu 1977). In the higher vertebrates, the lungs play an important part in acid-base regulation by tuning PCO<sub>2</sub> levels in blood with ventilation and, to a smaller extent, through elimination of plasma HCO<sub>3</sub><sup>-</sup> ions by the kidneys (Davenport 1974; Burg and Green 1977). Such animals can dissociate themselves from water for ion and pH regulation. The highly terrestrial Trinidad mountain crab, Pseudothelphusa garhami, which has evolved very efficient lungs (Innes and Taylor 1986a,b), has very a low PCO<sub>2</sub> in the hemolymph. The passive buffering capacity of blood with respect to both HCO<sub>3</sub><sup>-</sup> and non- HCO<sub>3</sub><sup>-</sup> ions is five to six times smaller in water breathers than in mammals and birds, with the buffer value of the skeletal muscle tissue being only 50 to 70% that of mammals (e.g., Heisler 1984). The arterial PCO<sub>2</sub> in the rainbow trout (Oncorhynchus *mykiss*) is about 0.3 kPa, in the facultative air-breathing jeju (Hoplerythrinus unitaeniatus) it is 1.6kPa, and in the obligate air breather piracucu, Arapaima gigas, it is 3.7 kPa (Randall et al. 1978a,b). The absolute values of PCO<sub>2</sub> and HCO<sub>3</sub><sup>-</sup> ions in blood and tissues are lower in water breathers than in air breathers (Tables 2,22). The PCO<sub>2</sub> characteristically ranges between 0.1 and 0.5 kPa and HCO<sub>3</sub><sup>-</sup> ions from 5 to 15 mM in the plasma of fishes (Heisler 1984): a drop in pH induced by hypercapnia is counteracted by elevation of HCO<sub>3</sub><sup>-</sup> ions. Exposure of the eel, Conger conger, to an environmental PCO<sub>2</sub> of 1 kPa lowered plasma pH by about 0.4 units as plasma PCO<sub>2</sub> rose from 0.25 to 1.3 kPa (Heisler 1982b): over a duration of 5 to 10h, the concentration of  $HCO_3^-$  ions increased from under 10 to about 20 mM, partly adjusting the pH. In the facultative air breather, Synbranchus marmoratus, movement from water to air changes plasma PCO<sub>2</sub> from about 0.75 to 3.5 kPa: pH falls by about 0.6 units and the intracellular concentration of  $HCO_3^-$  ions rises by a factor of 4 (Heisler 1982b).

The changes in the blood PCO<sub>2</sub> which occurred in animals moving from water to air are well demonstrated by the contemporary amphibians which start their life (larval stage) in water where they are obligate water breathers and metamorphose to become perfect air breathers. In *Rana catesbeiana*, at the tadpole stage, where the gills and the skin are the main gas exchangers, the blood PCO<sub>2</sub> is about 0.3 kPa while during and after metamorphosis, when the gills disappear and the lung assumes a prominent respiratory role, the blood PCO<sub>2</sub> increases notably (Erasmus et al. 1970/1971; Just et al. 1973; Burggren and Wood 1981; Tables 2,22). In the garfish, *Lepisosteus osseus*, the blood PCO<sub>2</sub> fluctuates with the ambient concentration of O<sub>2</sub> and the use of the gills or lungs for respiration (Rahn et al. 1971). During the summer months when the concentration of O<sub>2</sub> decreases due to the elevated temperature and its demands are high owing to increased metabolism, the blood PCO<sub>2</sub> increases. In winter (when the dissolved concentration of O<sub>2</sub> is high and metabolic rate lower) the gills are largely utilized and the blood  $PCO_2$  drops. At such times, the fish rarely comes to the surface to breath and the diving durations are longer (e.g., Kruhoffer et al. 1987). In the bimodal air breathers, long respiratory intervals are possible because the greatest porportion of  $CO_2$  is eliminated through the skin. The lungfish, e.g., *Protopterus amphibius*, and the bowfin, *Amia calva* (Dence 1933), regularly estivate during unpropitious circumstances, with the former staying in this state for as long as 5 years. When in water, the fish loses  $CO_2$  and  $NH_3$  through the gills and on land (during estivation),  $CO_2$  accumulates internally and  $NH_3$  is converted to urea and stored. This strategy allows the fish to survive during a time when water breathing is impossible.

#### 5.5 Control and Coordination of the Bimodal Gas Exchange Process

In both air- and water breathers, ventilation is generally a constant and rhythmic process. Endogenous respiratory rhythm in mammals might emanate from specialized pacemaker cells located in the respiratory neural system (e.g., Onimaru and Homma 1987; Smatresk 1990; Smith et al. 1991). Response to hypoxia in fish and mammals is mediated by peripheral chemoreceptors (Sinclair 1987; Burleson and Milsom 1990; Smatresk 1994). In the air breathers, the central ventilatory control is more responsive to PCO<sub>2</sub> and/or pH while in the water breathers there does not appear to be a direct correlation between PCO<sub>2</sub> and ventilation (e.g., Batterton and Cameron 1978; Hedrick et al. 1991). Only after successfully switching the major CO<sub>2</sub> clearance role to the lung could a bimodal breather dispense with the need to use the skin for respiration and develop an impermeable surface cover to avoid dehydration on land. Such a change, however, had to await development of a more efficient lung and respiratory ventilatory mechanism to clear CO<sub>2</sub>. Aspiratory breathing (a costal suction pump effected by action of the ribs and the intercostal muscles) first evolved in reptiles (Hughes 1963). The modern lung (first encountered in reptiles) has adopted the dual role of O<sub>2</sub> and CO<sub>2</sub> exchange and acid-base regulation. The cost and risk in air breathing was that of increased pulmonary water loss, a process accentuated by the fact that the air breathers relied exclusively on respiratory ventilation for O<sub>2</sub> acquisition and CO<sub>2</sub> clearance. Furthermore, except in reptiles, respiration in mammals and birds is regular and continuous. Exposure of the eel, Anguilla, to air causes the blood  $CO_2$  level to rise and the arterial  $PO_2$  and the  $O_2$  consumption to decrease (Berg and Steen 1965). In Synbranchus, aerial respiration inhibits CO<sub>2</sub> elimination leading to increase in the blood PCO<sub>2</sub> (Johansen 1966). The inability to lose  $CO_2$  into the air through either the gills or simple lungs is one of the main factors which constrained emergence from water (Hughes 1966; Thompson 1969). This explains why some modern air breathers with lungs and/or special respiratory organs inhabit water for the singular purpose of ease of CO<sub>2</sub> elimination. The air-breathing crabs which cannot eliminate CO<sub>2</sub> across the hard carapace have retained the gills (Innes and Taylor 1986a,b). This, while enabling them to maintain a low arterial PCO<sub>2</sub>, confines them to habitats where water is easily accessible. In the robber crab, *Birgus latro*, much of the  $CO_2$  is eliminated across the gills during rest, but during activity about one half of it is voided via the lungs (Greenaway et al. 1988).

On adopting air breathing, the bimodal breathers were not only faced with the problem of detecting respiratory stimuli from both internal and external milieus but also that of controlling physiologically distinct modes of respiration (e.g., West and Burggren 1983; Shelton and Croghan 1988; Smatresk 1988; Boutilier 1990). The respiratory control changed from a centrally modulated process to a diffusely coordinated one: the central control on the branchial ventilation was attenuated and the aquatic and air breathing integrated. The complexity of the plan depended on the relative importance of each mode of breathing to the overall respiratory needs and on factors such as the degree of adaptation to aquatic or terrestrial subsistence, the level of activity, and external factors such as hypoxia, temperature, and humidity. Fast, multiple respiratory responses are essential for animals subsisting in highly dynamic habitats characterized by precipitate fluctuations in O<sub>2</sub> and CO<sub>2</sub> levels. In the unimodal air breathers, e.g., birds and mammals, the control of ventilation is mediated centrally with the peripheral inputs simply modulating centrally produced impulses. However, in the intermittent air breathers, the ventilatory effort and pulmonary blood flow appear to be mainly regulated and synchronized through chemical and/or mechanical information initiated by the prevailing needs for  $O_2$  (e.g., Johansen et al. 1997; Milsom 1990; West et al. 1992). Mechanoreceptors, which detect the gradual decrease in lung inflation (as O<sub>2</sub> is used up to some baseline value) have been described in the lungs of the bowfin (Amia calva) (Milsom and Jones 1985) and in the lungfish (Protopterus aethiopicus) (DeLaney et al. 1983). In Amia, aquatic hypercapnia at a concentration of CO<sub>2</sub> of up to 3% increases branchial ventilation (Johansen 1970). The absence of ventilatory or cardiovascular responses after intracranial perfusions of hypercapnic and low pH solutions shows that a central chemoreceptor involvement of O<sub>2</sub> or CO<sub>2</sub>/pH in ventilatory regulation is lacking in A. calva (Hedrick et al. 1991). In terrestrial vertebrates, central chemoreceptive loci sensitive to CO<sub>2</sub> levels and/or pH changes are known to include the ventral region of the medulla oblongata (e.g., Hitzig and Jackson 1978). Internal O<sub>2</sub> chemoreceptors which stimulate air breathing have been described in the "lung" of the gar-fish (Smatresk et al. 1986) and in P. aethiopicus (Lahiri et al. 1970). The characteristic response of bimodal breathers to hypoxia is a notable increase in the ventilation of the accessory respiratory organs and a reduction in gill ventilation (Willmer 1934; Johansen et al. 1970b): in the gar-fish, a ten-times increase in ventilatory volume occurs consequent to increases in the ventilatory frequency and tidal volume (Smatresk et al. 1986). In Protopterus, respiratory responses appear to be highly sensitive to arterial gas tensions or pH (Lenfant and Johansen 1968; Johansen and Lenfant 1968). In Neoceratodus forsteri, the Australian lungfish, branchial ventilation increases remarkably as the PO<sub>2</sub> in the "inhaled" water falls below 10kPa while aerial respiration is induced at a water PO<sub>2</sub> level of 11.3 kPa (Johansen et al. 1967): the chemoreceptors responsible for these respiratory responses are apparently external (branchial) as injection of urea into the lung does not elicit any compensatory response. In adult Protopterus aethiopicus, water PO<sub>2</sub> does not affect branchial or aerial breathing (Johansen and Lenfant
1968) but reduced lung PO<sub>2</sub> and systemic PO<sub>2</sub> stimulate the frequency of air breathing (Johansen and Lenfant 1968; Lahiri et al. 1970). Sectioning the nerves to the branchial arches reduces the responsiveness of *P. aethiopicus* to hypoxia (Lahiri et al. 1970), suggesting that chemoreceptive area(s) may exist in the branchial arches of the species. Such site(s) have, however, not been morphologically identified (Fishman et al. 1989). Using chemical probes such as sodium cyanide for O<sub>2</sub>-sensitive elements (Lahiri et al. 1970) or nicotine (Johansen 1970), the existence of intravascular chemoreceptors in *Protopterus* has been demonstrated. Juvenile *Protopterus* rely more on aquatic respiration and respond to low water PO<sub>2</sub> (8% O<sub>2</sub> by volume) with a 50% increase in branchial and 300% increase in aerial breathing.

In the air-breathing fish, the ambient and internal CO<sub>2</sub> tensions are important regulatory factors of aquatic and aerial respiration. There are indications that water facing chemoreceptors occur in the gills of the air-breathing fish, e.g., P. aethiopicus (Johansen and Lenfant 1968) and the gar-fish, Lepisosteus (Smatresk et al. 1986; Smatresk and Azizi 1987). In *Protopterus*, the CO<sub>2</sub> receptors are located in the branchial region (or arterial side of the branchial circulation) (Johansen and Lenfant 1968), a feature which shows that increased gill PCO<sub>2</sub> inhibits branchial breathing before the arterial PCO<sub>2</sub> increases (Jesse et al. 1967). Externally situated O<sub>2</sub> receptors are important in monitoring the ambient PO<sub>2</sub> while the internal ones monitor the O<sub>2</sub> levels in blood. Switching to the most economical mode of gas exchange, i.e., from aquatic to aerial modes and vice versa, optimizes O<sub>2</sub> consumption to suit different metabolic needs and circumstances. Air-breathing fish are particularly sensitive to CO<sub>2</sub>. In Neoceratodus, increased gill concentration of CO<sub>2</sub> reflexively inhibits branchial respiration while stimulating air breathing (Johansen 1966). A biphasic response to CO<sub>2</sub> similar to that in the terrestrial vertebrates was observed in Protopterus by Johansen and Lenfant (1968) and Jesse et al. (1967): at physiological levels of water concentration of CO<sub>2</sub> of 0.5% by volume or lower, both gill and lung ventilation were stimulated and at higher levels of  $CO_2$  (1 to 5%), gill ventilation declined steadily. In both air and water breathers, the main response to hyperoxia is reduction of ventilation in correspondence to increased PO<sub>2</sub> in blood. The ventilatory inhibition in air breathers leads to accumulation of CO<sub>2</sub>, resulting in respiratory acidosis which stimulates ventilation to correct the acid-base status. In many surviving airbreathing fish, maximal heart rates occur soon after the ventilation of the aerial gas exchanger (Singh and Hughes 1973). This optimizes transfer and utilization of the inhaled air. Due to the apparent ventilatory insensitivity of the water breathers to PCO,, a persistent hypercapnic acidosis develops (Heisler 1982b). Mechanical inflation of the accessory respiratory organ in many air-breathing fish and amphibians often causes depression of the air-breathing activity (Lenfant et al. 1970b; Pack et al. 1984, 1992). In Amphiuma, inflation of the lung with pure nitrogen delays the onset of breathing, suggesting that mechanical distension may be an important factor in regulating  $O_2$  ventilatory patterns (Toews 1971). Following an interbreath period of 4 to 5 min, in Protopterus (Lenfant and Johansen 1968), soon after inhalation, cardiac output is increased four times over the apneic level and almost three quarters of the injected blood is directed to the lungs (Johansen et al. 1968a). In the frogs, Xenopus laevis (Shelton 1970) and *Rana pipiens* (Jones and Shelton 1972), the relative flow of blood to the gas exchangers depends on the breathing pattern. As in lungfishes and other airbreathing fish, apneustic breathing in amphibians determines the blood flow through the different circuits. During sustained breathing, the pulmocutaneous blood flow exceeds the systemic one, a condition reversed during apnea (Shelton 1976).

In Bufo marinus (e.g., West and Burggren 1984) and the lungfish, P. aethiopicus (DeLaney et al. 1983), mechanoreceptor elements are an integral factor in regulating and effecting ventilation-perfusion equality through regulated vasodilatation and constriction of the pulmonary vasculature. During air breathing, in lungfishes, vasomotor activity maintains a lower pulmonary than systemic vascular resistance, enhancing lung perfusion. In the most aquatic of all lungfishes, the Australian Neoceratodus forsteri, branchial vascular resistance is highest in the taxon and compares with that in the elasmobranchs and teleosts (Johansen et al. 1968a). Branchial vascular resistance in Protopterus as well as in Lepidosiren is very low due to the presence of low resistance vascular shunts which allow much of the cardiac output to bypass the branchial arches which have virtually lost all the gill filaments (Johansen and Reite 1968; Laurent et al. 1978). The same regulatory mechanism prevails in the clawed toad, Xenopus laevis, where pulmocutaneous blood flow increases two to four times over the prebreath level compared with the systemic flow which almost remains constant (Shelton 1970). In the snakehead fish, *Channa argus*, aerial hypoxia (a rather rare occurrence) results in a five-times increase in "lung" ventilation as the PO2 decreases from 20.7 to 5.3 kPa, but hypercapnia produces little change in lung ventilation (Glass et al. 1986). In intermittently breathing animals, e.g., Xenopus (Emilio and Shelton 1974), the diving turtles, Pelomedusa subflava (Glass and Wood 1983), Chrysemys (=Pseudemsy) scripta and Testudo graeca (Burggren and Shelton 1979), Chelonia mydas (West et al. 1992), and the garter snake, Thamnophis (Burggren 1977), O2 exchange is greatest soon after inspiration when the blood flow is highest. In some air-breathing fish, e.g., the gourami, Trichogaster (Burggren 1979), aquatic hypercapnia results in increase in ventilation but in the spotted gar-fish no such a significant response seems to occur (Smatresk and Cameron 1982a,b). While in air, Trichogaster can efficiently eliminate CO<sub>2</sub> across the accessory respiratory organ (the labyrinthine organ) because the epithelial lining which is derived from gill tissue contains a high concentration of carbonic anhydrase (Randall et al. 1981).

### 5.5.1 Ventilatory Modalities of the Gas Exchangers in the Bimodal Breathers

During transition to air breathing, the general pattern of breathing changed from regular transfer of water across the gills to the episodic breathing pattern which typifies a large number of air-breathing fish, amphibians, and reptiles. A determinate breathing pattern (of air) was thereafter reverted to (under resting conditions) in mammals and birds (Table 5). In the Dipnoi (McMahon 1969; Burggren and Johansen 1986) and the amphibians (Burggren and Doyle 1986), the air-

breathing mechanism was derived from the aquatic mechanism with inspiration occurring by a buccal force pump and expiration by elastic recoil of the lung. In the bimodally breathing teleosts, air ventilation is less well understood (Peters 1978; Kramer 1978; Liem 1980; Hellin and Chardon 1981; Ishmatsu and Itazawa 1981. In Channa, air breathing is complex and differs remarkably from water breathing (Liem 1984). In what was described as a cough mechanism, convergently, phylogenetically different air-breathing teleosts, the Channidae (Liem 1984), Anabantoidae (Peters 1978), and Clariidae (Hellin and Chardon 1981) have adopted a primeval aquatic process used to clear debris from the gills to ventilate the air-breathing organ(s). Since coughing is a water-dependent mechanism, fish utilizing the process only to ventilate their accessory respiratory organs are restricted in the period during which they can stay out of water (Liem 1984). The pressures which induced selection for interrupted ventilation and hence a lower respiratory frequency in bimodally breathing fish include: (1) need to reduce the number of vertical migrations to breath at the air-water interface, providing a substantial energy saving (e.g., Kramer and McClure 1981; Kramer 1983), and (2) avoidance of risks of predation during travels to the surface. When breathing air, Gillichthys becomes positively buoyant and has to hold onto substrate with its sucker-like pelvic fin (Todd and Ebeling 1966) while *Pseudapocryptes* is reported to float on the surface of the water with the opercular chambers inflated (Das 1934). During a dive, turtles, e.g., Chrysemys can breathhold for as long as 1 to 2h even when they have access to air: active breathing takes only 2 to 15% of the total activity when undisturbed (Belkin 1964; Burggren 1975). Intermittent breathing occurs in the bimodal breathers due to the large  $O_2$ stores in their body and the fact that during the breath-holds, gas exchange can be effected through secondary respiratory site(s) (e.g., Farrell and Randall 1978). Animals which do not have these features, e.g., fish and vertebrates like birds and mammals, though with large O<sub>2</sub> stores, have a high O<sub>2</sub> consumption and must continuously ventilate their gas exchangers. Those animals whose O<sub>2</sub> needs are low, e.g., amphibians, reptiles, and air-breathing fish, can: (1) acquire supplemental  $O_2$  through multiple pathways, (2) reduce their  $O_2$  consumption by adopting anaerobic metabolism, or (3) reduce their overall energy needs, e.g., by estivating or displaying rhythmic breathing. Since they contain O<sub>2</sub>-rich air, the accessory respiratory organs of the bimodal breathers increase body O<sub>2</sub> stores by an order of magnitude. This lowers the ratio of  $O_2$  consumption to  $O_2$  store in the body. Episodic respiration in bimodal breathers rarely influences CO<sub>2</sub> and pH levels in the body as  $CO_2$  is eliminated through pathways such as the skin and the gills. However, the intermittent air breathers have higher body CO<sub>2</sub> stores than the continuously breathing aquatic animals. Most bimodal breathers increase the ventilation of the accessory respiratory organ when the arterial or mixed venous blood PO<sub>2</sub> falls. This occurs, e.g., after exposure to hypoxia, when the ambient temperature increases beyond a critical point or when the activity level is raised. In the larva of the dragonfly (Aeschna), when the O<sub>2</sub> content is reduced to 2.5 ml per liter ( $PO_2 = 7.3$  kPa), the organisms move to the surface of the water. Such a reflex is not initiated in the fish, Leuciscus erythrophthalmus, until the concentration of  $O_2$  drops to a low of 0.6 mll<sup>-1</sup> (Winterstein 1908). In *Protopterus*, exposure to 8%  $O_2$  increases lung ventilation three times but gill ventilation is raised by a

factor of only one half. This suggests that different receptors are involved in the control and coupling of respiration in the air breather (Jesse et al. 1967). The buccal force pump used to ventilate the gills of the water breathers has been carried over to the air-breathing fish and the amphibians to ventilate the accessory respiratory organs. The pump in *Rana catesbeiana* is particularly efficient since remarkably high arterial PO<sub>2</sub> levels (values as high as 12.7 kPa) are attained (Lenfant and Johansen 1967). In fish, it is only in the Amazonian osteoglossid, *Arapaima gigas* (Farrell and Randall 1978), and the lungfish, *Protopterus amphibius*, during estivation (Lomholt et al. 1975) that aspirational breathing has been alleged. Suctional breathing may have evolved in water but is clearly a design characteristic of air breathers (e.g., Brainerd 1994). The respiratory frequency in air-breathing fish can vary from 1 breath min<sup>-1</sup> to less than 1 breath h<sup>-1</sup> (Johansen et al. 1971a; Smatresk and Cameron 1982a,b). It has been suggested that the buccal force pump in modern amphibians may have evolved secondarily after loss of ribs (Foxon 1964).

#### 5.5.2 The Circulatory Patterns in the Gas Exchangers of the Bimodal Breathers

The transition from water to air breathing entailed drastic reorganization of the cardiovascular system. An elaborate mechanism of regulating the perfusion of the aerial gas exchanger(s), especially where they were arranged in parallel with the systemic circuit, was formed (e.g., Olson 1994). The occurrence of anatomical gill shunts is unique to the bimodal breathers (Laurent 1985). They have not been reported in the exclusively water-breathing fish (Laurent 1984). A number of attendant physiological problems arose from the superimposition of the accessory respiratory organ(s) on the branchial and systemic vascular circuits. These changes are most evident and better understood in the vertebrate transitional breathers. In the single arc circulatory pattern of the ancestral water-breathing fish, the entire cardiac output is directed to and perfuses the gills. The oxygenated blood passes to the dorsal aorta for distribution to the rest of the body. Attainment of air breathing necessitated integration of a separate circulatory network onto the original single circuit (Fig. 21). After the relocation of the ionic and gas exchange from the gills to the kidneys and the lungs, respectively, the aquatic vertebrates (which by now had developed into the terrestrial tetrapods) reverted to the original plan where all the cardiac output was driven through the only gas exchanger, the lung. The parallel arrangement in the bimodal breathers was converted into two circuits, pulmonary and systemic circuits.

The attendant gas exchange complications which arose with the evolution of bimodal breathing included: (1) undersaturation of arterial blood resulting from mixing of deoxygenated systemic blood with the efferent (oxygenated) blood from the accessory respiratory organ, (2) an apparent drop in respiratory efficiency consequent to perfusion of a gas exchanger with blood which had already passed through the gills, (3) shunting of the efferent oxygenated blood from the accessory respiratory organ(s) resulting in admixture with the systemic venous blood, (4) risk of transbranchial efflux of  $O_2$  from the oxygenated blood if the gills

were ventilated with hypoxic ambient water, and (5) undue tissue blood pressure arising from the addition of extravascular resistance pathways. These limitations were overcome or minimized by: (1) a drastic modification of the vascular layout, (2) axial blood flow separation, and (3) development of vascular shunts. In the lungfishes (Dipnoi), which have gills and lungs, the blood pumped from the ventricle is separated into two streams. One stream perfuses the anterior gill arches which are devoid of gill filaments (Parker 1892) and the other passes through the posterior gills and then onto the pulmonary artery (Johansen and Lenfant 1967; Burggren and Johansen 1987; Fig. 21). The latter pathway is accentuated when the ambient  $PO_2$  is low (Johansen et al. 1968a). Laurent et al. (1978) described a vascular shunt in the posterior gill arches (IV and VI) of P. aethiopicus which bypassed the secondary lamellae. The arrangement corresponds with that of the external gills of the amphibians (e.g., Fige 1936) and Lepidosiren paradoxa (Robertson 1913). As the blood returns to the heart, mixing of the pulmonary arterial blood with the systemic deoxygenated blood is avoided by the presence of a spiral valve in the conus arteriosus. By establishing and maintaining laminar flow, the valve effectively separates the two blood streams in the heart. The oxygenated blood passes through the gill arches to the body while the deoxygenated blood passes to the lungs (Johansen and Hol 1968; Satchell 1976) for oxygenation. In the lungfishes, the degree of intracardiac separation of the arterial and venous blood depends on the degree of reliance on air for  $O_{2}$ , with the process being least effective in the obligate water breather, Neoceratodus, and best developed in the obligate air breathers, Lepidosiren and Protopterus (Johansen and Lenfant 1967; Johansen and Hanson 1968; Lenfant and Johansen 1968). The partial separation between the pulmonary and systemic circuits similarly occurs in amphibians although no distinct intracardiac anatomical modifications are evident (DeJong 1962; Johansen and Hanson 1968). Unlike in fish, in amphibians, the cutaneous circulation is arranged in parallel with the systemic and the pulmonary circuits and can be greatly varied without affecting the other two pathways (Johansen 1979). In the air-breathing teleosts such as Channa (Ishmatsu et al. 1979; Munshi et al. 1994; Olson et al. 1994), Anabas testudineus (Munshi et al. 1986a; Olson et al. 1986b), Monopterus cuchia (Munshi et al. 1989, 1990), Heteropneustes fossilis (Munshi et al. 1986b; Olson et al. 1990a; Hughes et al. 1992), and the lungfishes (Fishman et al. 1989), cardiovascular remodeling occurs as a part of a gross reorganization of the vascular connections between the gills, the accessory respiratory organs, and the systemic circuit. To a less conspicuous but significant extent, modifications in the form of vascular shunts (preferential perfusion channels), changes in microcirculation, and vascular endothelium (Hughes and Munshi 1979) develop.

Hemodynamic considerations and physiological functions are the main factors which initiated, regulated, and determined the changes in the circulatory patterns in bimodal breathers (Olson et al. 1994). In *Channa*, the vascular organization allows partial separation of blood flow to the gills, accessory respiratory organs, and systemic tissues (Munshi et al. 1994) resulting in basically two functional circuits: (1) heart  $\rightarrow$  gills (1st and 2nd arches)  $\rightarrow$  the accessory respiratory organ(s)  $\rightarrow$  heart, and (2) heart  $\rightarrow$  gills (3rd and 4th arches)  $\rightarrow$  systemic tissue  $\rightarrow$  heart. The accessory respiratory organs are essentially in series with the branchial

vasculature and in parallel with the systemic circuit. The total cardiac output must hence first pass through the gills before perfusing the accessory respiratory organ or systemic tissues and both the deoxygenated systemic venous blood and oxygenated blood from the accessory respiratory organs returns to the heart. The first pathway (at the gill level) performs functions like osmoregulation, acid-base balance, CO<sub>2</sub> clearance, and metabolic N<sub>2</sub> excretion: Oxygen is subsequently taken up at the level of the accessory respiratory organ. The second pathway serves the role of shunting blood past the gills and delivering oxygenated blood directly to the systemic tissues: this curtails  $O_2$  loss to a hypoxic aquatic medium. In the holostean, Amia calva, to minimize O<sub>2</sub> efflux from the gill blood, the blood flow in the gills is altered so that much of it is shunted to the nonrespiratory parts of the gills (Johansen et al. 1970a). In having elaborate gill filaments and secondary lamellae with an adequate number of chloride cells (ionocytes), the 1st and 2nd pairs of gill arches of the bimodal breathers retain features characteristic of those of the water-breathing teleosts and the well-developed gills in air-breathing species. The 3rd and 4th arches are less elaborate (e.g., Munshi et al. 1990; Olson et al. 1990a; Olson 1991). In most species, e.g., in the cuchia eel Amphipnous cuchia (Lomholt and Johansen 1976), and the electric eel, Electrophorus electricus (Johansen et al. 1968b), the 3rd and 4th gill arches are used as a bypass of the vestigial gills to avoid loss of O2. Vascular and intracardiac modifications close to the heart and gills reduce the mixing of the arterial and venous streams of blood. The deoxygenated systemic venous blood and the oxygenated blood from the accessory respiratory organ returns to the heart, facilitating delivery of the deoxygenated fraction to the accessory respiratory organs. The systemic tissues are perfused with the oxygenated fraction (Ishmatsu et al. 1979; Ishmatsu and Itazawa 1983; Munshi et al. 1994).

The intracardiac physical separation of the venous and arterial streams of blood was unequivocally demonstrated by Ishmatus and Itazawa (1983): in Channa argus, it was found that the PO<sub>2</sub> in the anterior ventral aortic blood is lower than that in the posterior ventral aortic blood. The overall efficiency of the gas exchange process is to a great extent dependent on the effectiveness of minimization of the mixing of these two blood streams in the heart. In air-breathing fish, the perfusion of the accessory respiratory organ is well matched to the  $O_2$ available for aerial gas exchange. In the cuchia eel, Amphipnous cuchia (Lomholt and Johansen 1976), and electric eel, Electrophorus electricus (Johansen et al. 1968b), when breathing air, 75% of the cardiac output, respectively 80 and  $70 \,\mathrm{ml\,kg^{-1}\,min^{-1}}$ , is directed to the accessory respiratory organ: during apnea, the perfusion of the organ drops to 20%. This marks an incipient stage in the evolution of synchronization of ventilation and perfusion and functional separation of pulmonary and systemic perfusion. The process is better refined in the lungfishes, where a partial separation has been attained (Johansen at al. 1968a; Fig. 21). Total separation is reached in the higher vertebrates, birds and mammals. The low pressure pulmonary circuit (Figs. 25,26) which appears to have evolved very early, e.g., in the Dipnoi (Johansen et al. 1968a) enables extreme refinements of the structural parameters of the gas exchangers accommodating the development of a very thin blood-gas barrier in a highly dynamic organ.

## 5.6 Taxa with Notable Propensity for Bimodal Breathing

# 5.6.1 Mollusks

The Phylum Mollusca, with about 100000 species, is one of the largest and most successful in the Animal Kingdom (e.g., Jones 1983; Seed 1983). It is exceeded in specific diversity and numerical density only by Arthropoda and perhaps by Protozoa and Nematoda. Ecologically, the mollusks have penetrated a wider range of habitats than any other animal group (Ghiretti 1966). The gastropods, which constitute about 80% of all living mollusks, are the only group in the taxon which has colonized land. After the arthropods, the slugs are probably the most successful land invertebrates (Jones 1983). The success of the pulmonate gastropods is largely attributable to the development of certain physiological and behavioral adaptations which have enabled them to cope with problems of water balance, temperature, osmotic and ionic regulation, and gas exchange (Riddle 1983). Mollusks, which have a respiratory pigment with low O<sub>2</sub> unloading tension, can subsist in hypoxic environments and to a certain point may show metabolic independence to external O<sub>2</sub> levels (Borden 1931; Chaetum 1934).

Air breathing in pulmonate gastropods probably arose in O<sub>2</sub>-deficient waters such as estuaries, swamps, and muddy rivers (Seed 1983). In the prosobranches. e.g., Littorina rudis, there is a tendency towards formation of a lung on exposure to air. In the intertidal pulmonate limpets constituting the Siphonariidae, on return to the sea, secondary gills may appear within the former lung (Yonge 1952). The related Trimusculus (Gadinia) continues to breath air while residing in water (Yonge 1958). In the pulmonate gastropods, the mantle cavity has been converted into a highly vascularized lung (Maina 1989c; Figs. 59,60) where a large volume of "blood" is brought into close proximity to a respiratory medium for gas exchange in air or secondarily in water. Pulmonate gastropods are structurally the most conservative gastropods and their evolutionary advances have been committed essentially to face the challenges of terrestrial colonization. In a notable deviation from the normal pattern, some pulmonates, e.g., the siphonarid limpets, possess secondary gills which are housed in a dorsal mantle cavity (e.g., McMahon 1983; DeVilliers and Hodgson 1987). The gills of the pulmonate, Siphonaria capensis, bear ciliary tufts (about 400 cilia per mm<sup>2</sup>). The surface is covered by a single layer of epithelial cells with interspersed mucus-secreting cells (DeVilliers and Hodgson 1987). From the ciliated gills and other anatomical features, Siphonaria is believed to be one of the least developed pulmonates (Marcus and Marcus 1960; Hyman 1967). While Marcus and Marcus (1960) contend that the secondary gills have always been a feature of siphonariids, it has been suggested by, e.g., Yonge (1952) and Morton (1979) that the gills were initially lost as the taxon evolved terrestriality but redeveloped when it reverted to water. In the freshwater pulmonate gastropods, Lymnaea stagnalis and Planorbis corneus, which have secondarily taken to water after the extirpation of the gill-like ctenidia, the mantle cavity was converted into a well-vascularized water lung which is rhythmically filled with water (Precht 1939; Hunter 1953). As the ambient PO<sub>2</sub> drops below a critical level, the snail comes to the surface to breath air



Fig. 59. Surface of the lung of a pneumonate gastropod, *Trichotoxon copleyi*, showing the vascular network of blood capillaries (*inset*). The *main figure* shows one of the blood capillaries and clusters of mucus cell,  $\blacklozenge$ , which lie on the epithelial surface. *Bar* 10µm; *inset* 90µm. (Maina 1989c)

(Cheatum 1934). When access to air is prohibited, Lymnaea responds by elevating the concentration of hemocyanin in blood to enhance extraction of O<sub>2</sub> from the lung (Jones 1972). The opening of the pneumostome is regulated to optimize gas exchange while limiting water loss. Low ambient concentration of O<sub>2</sub> stimulates the opening of the pneumostome, which may remain permanently open at concentrations below 10% (Ghiretti 1966). The opening of the pneumostome in the terrestrial gastropods Limax, Arion, and Helix is influenced by the level of CO<sub>2</sub>. It remains permanently open at a concentration of CO<sub>2</sub> of 3 to 5% (Dahr 1927; Prosser 1961). In Helix pomatia, at 20% O<sub>2</sub>, the pneumostome remains closed and only opens when O<sub>2</sub> drops to 10% or less (Ghiretti 1966). Other factors which regulate pneumostomal size include temperature and humidity (Ysseling 1930; Wit 1932; Mass 1939). In the inactive slug, Limax maximus, the lung contributes 20% of the  $O_2$  consumption, the value rising to 50% during activity (Prior et al. 1983). In all mollusks, even where specific respiratory organs exist, gas exchange through the body surface occurs to varying extents. Transcutaneous gas exchange may be of critical importance under certain circumstances. In the land snail, Otala lactea, a significant amount of CO<sub>2</sub> is eliminated across the skin (Barnhart 1986a). In the aquatic air-breathing snails, P. corneus and L. stagnalis, where CO<sub>2</sub> is soon lost into the water through the skin, CO<sub>2</sub> does not act as a respiratory



**Fig. 60.** Blood capillaries, *c*, on the surface of the lung of a pneumonate gastropod *Trichotoxon copleyi*; *e* epithelial cell; *h* hemocyte;  $\rightarrow$  microvilli on the epithelial cell surface. *Bar* 2 µm. (Maina 1989c)

stimulus. Factors such as  $PO_2$  and loss of buoyancy regulate respiration (Jones 1961). In the Athoracophoridae, the lung is small. Many fine tubules lead from it to the roof of a blood sinus in a manner resembling the trachea of insects (Runham and Hunter 1970). Accordingly, the taxon has been termed the Tracheopulmonata.

The respiratory surface area of the lung in pulmonate gastropods ranges from 7 to  $13.5 \text{ cm}^2 \text{g}^{-1}$  (wet body mass ) (Yonge 1947). The dimensions compare and exceed those of some vertebrate air breathers, e.g., the lizard, *Tupinambis nigropunctus*  $3.2 \text{ cm}^2 \text{ perg}$  (Perry 1983) and the domestic fowl, *Gallus domesticus*,  $10 \text{ cm}^2 \text{g}^{-1}$  (Abdalla et al. 1982). Perhaps due to the large respiratory surface area of the lung and the thin blood-gas barrier (Maina 1989a), diffusion is adequate for gas exchange even at the very low PO<sub>2</sub> of 0.3 kPa characteristic of the habitats in which the slugs live (Ghiretti 1966). Depending on temperature, slugs can increase their O<sub>2</sub> consumption by as much as 400 to 500% (Mackay and Gelperin 1972; Prior et al. 1983). In *O. cygnea*, in the process of repaying the O<sub>2</sub> debt after activity, maximum O<sub>2</sub> consumption is reached after 1 h (Hers 1943). In active snails, PO<sub>2</sub> and PCO<sub>2</sub> in the lung of the land snail, *Otala lactea*, differed by less than 2 kPa from air, the arterial PCO<sub>2</sub> was similar to that in the gas in the lung but

the arterial PO<sub>2</sub> was 8kPa lower than that in the lung (Barnhart 1986b). This bespeaks a strong diffusional limitation of O<sub>2</sub> transfer across the blood-gas barrier. Only about 10% of the hemocyanin-O2 carrying capacity is utilized at 5 °C but rises above 70% at 25 °C in the active land snail O. lactea (Barnhart 1986c). In H. pomatia, O<sub>2</sub> affinity is 1.5 to 2 kPa at 20 °C and pH 7.6 to 8.2 (e.g., Konings et al. 1969), in Arion ater it is 2.2 kPa at 20 °C, pH 7.94 (Wells and Weber 1982) and in O. lactea 5.3 kPa at 25 °C, pH 7.9 (Barnhart 1986b). In Trichotoxon copleyi (Maina 1989c) and H. pomatia (Pohunkova 1967), the lung has developed in the form of a modified mantle cavity. The roof, which is highly vascularized, is lined by a respiratory epithelium which is made up of squamous cells bearing stubby microvilli. The blood vessels protrude into the air space (Figs. 59,60). Scattered among the squamous cells are goblet mucus cells (Fig. 59). Contrary to the observations made by Pohunkova (1967) on H. pomatia, the air-blood pathway lacks a continuous endothelial lining (Maina 1989c). In the pneumonate gastropods in general, the thin blood-gas barrier ranges in thickness from 6 to 10µm (Runham and Hunter 1970). In Trichotoxon (Maina 1989c), some parts of the barrier are as thin as 0.2 µm. The diffusion potential for O2 in the lung of Agriolimax agrestis is six times greater than the animal's total O<sub>2</sub> requirement (Runham and Hunter 1970). As in the lung-breathing exothermic vertebrates (e.g., Jackson 1978), in the arterial blood of O. lactea, PCO, increases and pH and PO<sub>2</sub> decrease with temperature, presumably due to a hypoventilatory response (Barnhart 1986b).

### 5.6.2 Crustaceans

The crustaceans have exploited habitats ranging from totally terrestrial ones to the deep sea (Bliss 1979; Hartnoll 1988; Henry 1994). The decapod crustaceans present remarkable respiratory diversity with evident progressive sophistication occurring from aquatic through to air breathing and terrestrial species (e.g., Bliss 1979; Cameron 1981; McMahon and Wilkens 1983; Innes and Taylor 1986a,b; Burggren and McMahon 1988b; Henry 1994). Land crabs are members of the Anomura and Brachyura which can to varying extents survive on land. The majority belong to the families Coenobitidae, Gecarcinidae, Grapsidae, Potamoidea, and Ocypodidae (Hartnoll 1988). Among the terrestrial species, respiratory efficiencies correlate with the O<sub>2</sub> demands which are imposed by factors such as habitat and level of activity (Johnson and Rees 1988). Most of the non-malacostracan crustaceans have no specialized respiratory organs: gas exchange occurs across the integumentary surfaces (Wolvekamp and Waterman 1960; McLaughlin 1983). In brachiopods and phyllopods, it occurs through appendages and their modifications such as the gills which come in remarkably different forms (Eriksen and Brown 1980a,b; McLaughlin 1983). The internal surfaces of the carapace are well adapted for gas exchange in the Brachyura (Cameron 1981; McLaughlin 1983).

Crustaceans developed in the sea and have evolved air breathing and invaded land severally (e.g., Bliss and Mantel 1968; Bliss 1979; Cameron 1981). The notably

terrestrial species come from the groups Amphipoda, Isopoda, and Decapoda (Bliss and Mantel 1968; Powers and Bliss 1983). Land crabs are of particular interest in comparative respiratory physiology as they display relevant adaptations associated with transition from water- to air breathing and terrestrial colonization (Innes and Taylor 1986a,b; Taylor and Innes 1988). Air-breathing crabs have complex lungs which are contained in the branchial chambers formed from branchiostegal and sometimes thoracic walls (e.g., Farrelly and Greenaway 1987, 1992; Maitland 1987; Fig. 61). In crustaceans, terrestrialness and air-breathing efficiency correlate with the elaboration of the lung (e.g., Diaz and Rodriguez 1977). Though restricted to burrows and demonstrating nocturnal activity, isopods inhabit some of the most xerix habitats such as the Sahara desert (Cloudsley-Thompson 1977). The notably terrestrial species of crabs such as Birgus latro, Holthuisana (=Austrothelphusa) transversa, Pseudothelphusa garhami, and Ocypode saratan have evolved an elaborate well-perfused, highly amplified epibranchial chamber lining which constitutes for all practical purposes a lung (e.g., Farrelly and Greenaway 1987, 1992; Maitland 1987; Al-Wassia et al. 1989; Fig. 62). Such species can stay away from water for a long time. The anomuran coconut crab, Birgus latro, can survive in air with its gills removed (Harms 1932; Smatresk 1979) and, like Ocypode, will drown if forcefully submerged in water. The respiratory physiology of Birgus differs from that of aquatic crabs in certain ways (Cameron and Mecklenburg 1973). As in the vertebrate lung breathers, the acid-base balance of its hemolymph is effected by respiratory exchange rather than by ionic transfer mechanisms. The blood pressure in most terrestrial crabs is two to four times greater (up to 6.6 kPa) than aquatic crabs and ventilation of the branchial cavity is efficient and continuous, a mechanism which, though the O<sub>2</sub> extraction factor of the lung is only 2 to 8%, promotes gas exchange rate greatly (McMahon and Burggren 1988; Fig. 63). The PCO<sub>2</sub> of the venous blood of Birgus latro, which is as low as 1.2 kPa (Burggren and McMahon 1981), may be explained by high ventilatory rates (Dejours and Truchot 1988). During severe hypoxia or hypercapnia, continuous scaphognathite beating (at a

**Fig. 61.** Transverse section of the body of a soldier crab, *Mictyris longicarpus*, through the branchial chambers which are divided into an inner gill space and an outer space which has been converted into a lung. (Farrelly and Greenaway 1987)





**Fig. 62.** Schematic diagram of the vascularization of the epibranchial membrane of the lungs of the soldier crab, *Mictyris longicarpus*. The lung is very elaborate, providing a very highly efficient gas exchange capacity and hence ability to subsist over long periods on land. *DAV* Dorsal arterial vessel; *MAV* middle arterial vessel. (Farrelly and Greenaway 1987)

rate of 50 to 300 beats min<sup>-1</sup>) produces a ventilatory volume of  $100 \text{ cm}^3 \text{ min}^{-1}$  in a 1- to 2-kg specimen of *Birgus iatro*. In such circumstances, O<sub>2</sub> extraction factor may increase to 20% (Maitland 1990a). The gills provide  $29 \,\mu \text{l O}_2 \text{ g}^{-1} \text{ h}^{-1}$  while the balance of about 60 to  $110 \,\mu \text{l O}_2 \text{ g}^{-1} \text{ h}^{-1}$  in a routinely active land crab (at 25 °C) is transferred across the lungs.

As in the air-breathing fish (Hughes and Morgan 1973; Pelzenberger and Pohla 1992), in crabs, the evolution of alternative air-breathing sites is associated with reduction in the gill surface area (Table 23). While the active aquatic crabs have a respiratory surface area of about 900 to 1400 mm<sup>2</sup>g<sup>-1</sup> (Gray 1954, 1957; Veerannan 1974, Hawkins and Jones 1982; Johnson and Rees 1988; Henry et al. 1990), that in the intertidal ones ranges from 500 to  $900 \,\mathrm{mm^2 g^{-1}}$  (Hawkins and Jones 1982; Rabalais and Cameron 1985; Santos et al. 1987) and in the fully terrestrial species the values range from 12 to 500 mm<sup>2</sup>g<sup>-1</sup> (Cameron 1981; Farrelly and Greenaway 1992). The branchial chambers of the bimodal breathing crabs are larger than those of the aquatic ones (Henry 1994). Internal modifications in form of intense foldings and invaginations (e.g., Innes and Taylor 1986a; Farrelly and Greenaway 1987; Maitland 1987; Fig. 62) provide a large respiratory surface area (Greenaway and Taylor 1976; Diaz and Rodriguez 1977). Combined with a remarkably thin blood-gas barrier of 0.2 to  $0.4 \mu m$  (Farrelly and Greenaway 1992; Table 23), a high diffusing capacity of the lung for  $O_2$  is generated in the terrestrial crabs. Air channels in P. garhami run from the lung to the air sacs beneath the carapace and are actively ventilated in a throughflow manner (Diaz and Rodriguez 1977; El Haj et al. 1986) giving a high arterial PO<sub>2</sub> (16 to 18.7 kPa)



**Fig. 63.** Correlation between O<sub>2</sub> affinity of blood of air-breathing crabs and mode of gas exchange. In an aquatic breather, e.g., *Carcinus*, O<sub>2</sub> uptake through the gills is aided by the high affinity of the hemocyanin to O<sub>2</sub> (i.e., values of  $P_{50}$  – the PO<sub>2</sub> at which 50% of the Hc is saturated – are low). In terrestrial crabs with well-vascularized gill chambers, e.g., *Cardisoma* and species such as *Pseudothelphusa* which can efficiently ventilate their lungs, a high arterial O<sub>2</sub> tension can be attained. In such species,  $P_{50}$  is much greater. (Innes and Taylor 1986a)

and a low PCO2: gas exchange does not seem to be diffusion-limited (Innes and Taylor 1986a,b). The arterial PO<sub>2</sub> in the soldier crab, Mictyris longicarpus, is 12.8 kPa (Farrelly and Greenaway 1987) and in the shore crab, Carcinus maenas, is 14.2 kPa (Lallier and Truchot 1989). Many semaphore and grapsid intertidal and supratidal crabs carry some water on the lower parts of the branchial chambers (e.g., Burnett and McMahon 1987; Maitland 1990b). On average, crabs carry 0.35 to 0.5 ml of water per 7-g live body mass. The pumping action of the scaphognathites (aided by setae) circulates 0.2 ml of it (per 3-g body mass) around the body across the branchial chambers while on excursion on land (e.g., Hawkins and Jones 1982; Felgenhauer and Abele 1983). The water keeps the gills moist (avoiding desiccation and enabling aquatic respiration in air), functions as a sink for the excreted CO<sub>2</sub> (Burnett and McMahon 1987), and moistens the food during eating (Fielder 1970; Maitland 1990b). In some species, air is bubbled through the water in the branchial cavity to oxygenate it (McMahon and Burggren 1988). In the fully terrestrial land crabs, e.g., Birgus latro, CO2 is excreted through the gills (Greenaway et al. 1988).

Although the factors which have limited the terrestrial radiation of the land crabs are not well known (e.g., Hartnoll 1988), compared with the most successful arthropod groups, e.g., Arachnida, Chilopoda, Diplopoda, Insecta, and terrestrial vertebrates like birds and mammals, as a group, the crustaceans display relatively weak adaptation for air breathing and terrestrialness. They manifest an extremely

Crab	Diffusion distance		$VO_2$	PaO <sub>2</sub>	PVO <sub>2</sub>
	Gills (µm)	Lung (µm)	(µmolg 'h ')	(mmoll <sup>*</sup> )	(mmoll ')
Terrestrial					
Holothuisana transversa	5-8	0.2-0.3	0.80	_	-
Birgus latro	-	0.5-1.2	1.41	27	13
Pseudothelphusa garhami	5	0.4-1.0	1.51	124–140	-
Mictyris longicarpus	3.5-7.0	1.2-3.6	-	79–95	20-27
Ocypode ceratophthalmus	2.9-3.5	0.25-0.3	-	-	-
Cardisoma hirtipes	5-12	2.5-4.5	2.29	-	-
Aquatic					
Procambarus clarkii	3.15-8.7	_	_	_	_
Astacus pallipes	1.4-5.7	-	-	-	-

 Table 23. Gas diffusional distances and respiratory parameters in some terrestrial and aquatic crabs

Respiratory parameters from McMahon and Burggren (1988) and thicknesses from Burggren and McMahon (1988b).

tenuous hold on land. In contemporary biology, the crustaceans are a living model for studying the prerequisites for air breathing, terrestrial colonization, and occupation of new habitats. It is not surprising that in the taxon, potential respiratory organs/sites are still being discovered and debated (e.g., Mangum 1994).

# 5.6.3 Fish

The common expression "like a fish out of water" conveys the general inability of most fish and other aquatic life forms to survive and effectively exchange gases in air. Phylogenetically, the lungs are thought to have evolved as adaptations in hypoxic or anoxic conditions in the aquatic medium in which the ancestors of the modern fish lived. A small number of the contemporary descendants of the ancient fish, especially those which continued to subsist in derelict hypoxic warm tropical freshwaters, have retained air breathing (Burggren et al. 1985b). In the entire ichthyoid fauna, only the Holostei and the tropical freshwater teleosts can efficiently breathe air (e.g., Munshi and Hughes 1992). The process of air breathing has become so perfected in some fish that they succumb on extended forced submersion. Fish, the most successful vertebrate taxon with 20000 or so species,

have lived in water for over 350 million years (Gilbert 1993). Only a handful, the majority of which are teleosts, have evolved air breathing (Bertin 1958; Fänge 1976). As many as 60 (Sayer and Davenport 1991) and, according to Graham (1994), as many as 370 species of fish are known to breath air to varying extents (Table 19). More than any other animal group, fish present the broadest perspective of the evolution of air breathing (Graham 1994) and, to an extent, terrestriality.

The extant teleosts are far from being ancestral to the terrestrial air-breathing vertebrates as they only arose from the holosteans during the late Triassic period (e.g., Jarman 1970). They are thus far removed from the mainstream progenitors of the land vertebrates. The air-breathing teleostean fishes present remarkable morphological, physiological, biochemical, and behavioral respiratory diversity. However, considering their very different systematic backgrounds, the similarity in the preadaptations and the strategies adopted to achieve air breathing are astonishing and overwhelm the differences. In general, larval and juvenile bimodal breathing fish, e.g., Clarias, Colisa, and Anabas, derive much of their O<sub>2</sub> needs from water utilizing the gill/skin system. Their dependence on water for O<sub>2</sub> decreases with age (Hughes et al. 1973; Prasad 1988). These fishes acquire the airbreathing capacity at about 8 to 20 days after hatching (Bruton 1979; Singh and Mishra 1980; Singh et al. 1982). In Anabas, the labyrinthine organs start developing at the 5th day of hatching but air breathing does not occur until the 13th to 14th day (Hughes et al. 1986a). Obligate air breathers, e.g., adult piracucu, A. gigas, die if refused access to the atmospheric air even when kept in welloxygenated water (Stevens and Holeton 1978b). The gar-fish, Lepisosteus, is a facultative air breather at low temperatures but an obligate one at higher ones (Rahn et al. 1971). At high PO<sub>2</sub>, in water, Piabucina derives 10% of its O<sub>2</sub> need from air but at a lower water PO<sub>2</sub> of 4.7 kPa, it acquires 70% from air (Graham et al. 1977, 1987). In the snake-headed fish, Channa punctatus, and the climbing perch, Anabas testudineus, the accessory respiratory organs develop during larval and juvenile stages. They leave water only when the development is complete (Singh et al. 1982; Hughes et al. 1986). Many air-breathing fish possess reduced scales and have a well-vascularized skin through which substantial gas exchange occurs (Lenfant and Johansen 1972; Romer 1972). At a temperature of 7 °C, buccal and cutaneous breathing in the common eel, Anguilla vulgaris, supports the metabolic needs (Faber and Rahn 1970). The buccal cavity of the eel is both profusely diverticulated and highly vascularized. It is ventilated every 2 to 5 min with air (Johansen et al. 1968b). At 26 °C, the O<sub>2</sub> uptake rate of 30 ml per kg per h, which constitutes 80% of the overall needs, is procured from the air across the buccal cavity and the rest across the skin. The highly atrophied gills contribute an insignificant amount of O<sub>2</sub>. The gills are, however, responsible for elimination of as much as 94% of  $CO_2$  (Table 21). The accessory respiratory organs in fish are diverse and are variably ventilated. They include the skin (e.g., Amphipnous cuchia), bucco-pharyngeal membrane (e.g., Electrophorus electricus), suprabranchial chamber membranes, and labyrinthine organs (e.g., Clarias mossambicus), gastrointestinal system (e.g., Ancistrus anisitsi and Plecostomus plecostomus), and the air bladders (e.g., Arapima gigas and Amphipnous cuchia). In Plecostomus and Ancistrus (both tropical Siluroidae), the stomach is a respira-

tory organ into which air is swallowed and regurgitated (Carter and Beadle 1931; Carter 1935). In the pond loach, Cobitis (=Misgurnus), the middle and distal parts of the gastrointestinal system (GIT) are respiratory (McMahon and Burggren 1987) and the residual air is passed out through the anus (Krogh 1941; Jeuken 1957). The blood-gas barrier in the respiratory sites of the GIT is remarkably thin (Jasinski 1973). Respiration using the gastrointestinal system may have been a forerunner to the development of the lung by evagination of the foregut. Morphometrically, the air sac of Heteropneustes fossilis is superior to that of Aphipnous cuchia, similar to that of Lepidosiren, but less than that of Protopterus (Hughes et al. 1992): the respiratory surface area of a 40-g specimen was reported to be  $24 \text{ cm}^2$ , the harmonic mean thickness of the blood-gas barrier 0.342  $\mu$ m, and the morphometric diffusing capacity  $3.2 \times 10^{-5}$  ml O<sub>2</sub> s<sup>-1</sup> mbar<sup>-1</sup>. In those airbreathing fish which use isolatable diverticula such as the stomach, intestine, and gas bladder, the gills can simultaneously be ventilated to supplement  $O_2$  uptake and carry out processes such CO<sub>2</sub> elimination, ionic transfer, and clearance of nitrogenous waste products. This cannot occur in those fish, e.g., the electric eel (Electrophorus electricus) and the knifefish (Hypopomus) which use the buccal cavity as an accessory respiratory organ, a time when aquatic respiration is impeded and hence bimodal respiration momentarily eliminated (e.g., Graham et al. 1987). Reckoning from the spatial and temporal diversity of the evolved accessory respiratory organs, air breathing is an independently evolved attribute. Different lineages endeavored to look for solution to common selective pressures, especially hypoxia. Bimodal breathing is estimated to have evolved at least 67 separate times (Graham 1994). Regression of the accessory respiratory organs has occurred in many fish which have radiated into habitats where need for bimodal breathing is not intense. This has occurred in fish which live in stable, welloxygenated waters, e.g., in the swift mountain streams, expansive freshwater lakes, or at great depths (Greenwood 1961; Graham 1994). Together with the ancestral crossopterygians which gave rise to the primitive amphibians, the dipnoans are the closest extant relatives of the modern tetrapods (e.g., Rosen et al. 1981; Duellman and Traub 1986; Panchen and Smithson 1988; Pough et al. 1989; Joss et al. 1991; Meyer and Dolven 1992). For this reason, the group continues to arouse substantial interest in biology. Respiratory modifications for air breathing were extensively found in most or all of the late Paleozoic fishes (Romer 1972) when the O<sub>2</sub> content of the water, e.g, in the Silurian period, was only 1.9 kPa. The earliest functional lungs are presently found in only the three genera (and six living Southern Hemisphere freshwater species) of lungfishes (Dipnoi), namely Lepidosiren, Protopterus, and Neoceratodus, in the chondrosteans (Polypterus and Calamoichthys), and the holosteans (Amia and Lepisosteus) (e.g., Burggren et al. 1985b; Sacca and Burggren 1982). Developmentally, these fish have been extremely conservative. They have changed very little since they attained the capacity for breathing air and hence serve as living examples for understanding the adaptive strategies and the backgrounds against which air breathing evolved. The South American Lepidosiren and the African Protopterus, genera which evolved in separate continents after the splitting of the Gondwana Land in the early Mezozoic but continued to live in similar habitats (respectively in the poorly oxygenated waters of the Amazonian basin and the derelict inland freshwater

masses of the continental Africa) have paired, symmetrical internally sudivided lungs (Fig. 64; Klika and Lelek 1967; Hughes 1973; Hughes and Weibel 1976; Maina and Maloiy 1985; Kimura et al. 1987; Maina 1987a) and vestigial gills (Laurent et al. 1978; Laurent 1996). Both are obligate air breathers (Table 25) and have a better CO<sub>2</sub> buffering capacity of blood than the Western Australian Neoceratodus forsteri, a facultative air breather. Protopterus aethiopicus acquires 89% of its O<sub>2</sub> needs and eliminates 40% of CO<sub>2</sub> through the lungs (Johansen and Lenfant 1968; Lenfant and Johansen 1968; Table 24). Neoceratodus has a single, unpaired, thin-walled but internally subdivided lung (Grigg 1965) and possesses fully functional gills (Johansen and Lenfant 1967). The diameter of the pulmonary blood capillaries of *Neoceratodus* is 20 µm, a value which corresponds with the large ( $40 \times 30 \mu m$ ) oval erythrocytes (Gannon et al. 1983). In *Protopterus*, the posterior branchial arches carry a notable number of lamellae while the anterior ones, which provide shunt pathways for systemic circulation, are vestigial (Laurent 1996). To support CO<sub>2</sub> clearance, the functional branchial arches are continuously perfused (Lenfant and Johansen 1968; Laurent et al. 1978) even though pulmonary ventilation is intermittent. Encased in a cocoon of hard soil as the water dries up, Protopterus amphibius has been reported to survive in a semianimated (estivated) state for months to years as it episodically breathes air through a "snorkel" which connects the cocoon with the surface (DeLaney et al. 1974). The harmonic mean thickness of the blood-gas barrier in Neoceratodus ranges from 1.5 (Hughes 1973) to 2.5 µm (Gannon et al 1983), that of Lepidosiren is 0.85 um (Hughes and Weibel 1976), Protopterus 0.37 (Maina and Maloiy 1985) and  $0.85 \mu m$  (Hughes and Weibel 1976), and in *Polypterus* 1.22  $\mu m$  (Zaccone et al. 1995).

To varying extents, the air-breathing fish rely on air or water for their  $O_2$  needs (Table 25). The more terrestrial species are obligate air breathers. Among the actinopterygian fishes (Lauder and Liem 1983), only the polypterids have devel-

Species/T (°C)	Total VO <sub>2</sub> (mlmin <sup>-1</sup> kg <sup>-1</sup> )	Lung $VO_2$ (% of total)	Lung VCO <sub>2</sub> (% of total)	Tidal vol. (% of lung vol.)
Neoceratodus				
(18-20)	0.25	0	0	40-60
Protopterus		-	-	
(20)	0.3	89	40	60-80
Lepidosiren				
(18–20)	0.37	96	60	≤100
(25–28)	-	-	-	-
Lepisosteus				
(22)	0.65	73	40	40
(10)	0.30	0	-	-
Amia				
(10)	0.30	0	0	-
(20)	1.5	35	25	≤100
(30)	2.0	75	40	-

Table 24. Gas exchange in the fish lungs. (After Wood and Lenfant 1976)



**Fig. 64.** a Cross section of the lung of the African lungfish, *Protopterus aethiopicus*, showing the eccentrically located air duct, d, and the gas exchange cells, \*. b Closeup of the gas exchange air cells, c, of the lung of *Protopterus* which are formed by hierarchically arranged septa, i.e., the primary septa, p, secondary septa, s, and tertiary septa, t. The septa support the blood capillaries and contain the contractile and supporting tissue elements like collagen, elastic tissue, and smooth muscles. **a** *Bar* 0.67 mm; **b** 0.28 mm. (Maina 1987a)

T (°C)	Species	O <sub>2</sub> uptake (	Aquatic/aerial		
		Aquatic	Aerial	Total	
25	Anabas testudineus	52.62 (46.40)	60.80 (53.60)	113.42	0.865
25	Clarias batrachus	38.85 (41.60)	54.54 (58.40)	93.39	0.712
25	Heteropneustes (=Saccobranchus) fossilis	50.10 (59.25)	34.45 (40.75)	84.55	1.454
26	Electrophorus electricus	6.92 (23.07)	23.08 (76.93)	30.00	0.300
20	Amia calva	66.00 (68.75)	30.00 (31.25)	96.00	2.20
30	A. calva	30.00 (26.32)	84.00 (73.68)	114.00	0.357
22	Lepisosteus osseus	14.40 (24.66)	39.00 (75.34)	58.40	0.369
25	Cobitis fossilis	14.40 (19.51)	59.40 (80.49)	73.80	0.242
20	Protopterus aethiopicus	1.26	10.14 (88.95)	11.40	0.123
24	P. aethiopicus	5.80	56.70 (90.72)	62.50	0.102
18	Lepidosiren (Iuvenile 150g)	54.00 (63.83)	30.60 (36.17)	84.60	1.764
20	Lepidosiren (adult)	1.4 (4.36)	30.70 (95.64)	32.10	0.045

**Table 25.** Oxygen uptake in some air-breathing fishes. The proportion of  $Vo_2$  from water and air and the quotient between aquatic and aerial  $Vo_2$  are also given. Numbers in parentheses are % proportions. (After Singh 1976)

oped conspicuous lungs. The lungs are, however, relatively more primitive than those of the Dipnoi. The lungs of Polypterus are slender and lack discernible internal compartmentation (Klika and Lelek 1967; Hughes and Pohunkova 1980) with the right lung being better developed than the left. Hoplostemum thoracatum from the Paraguayan swamps, which travels overland across the grass from one pool to another (as they dry), swallows air continuously:  $O_2$  is absorbed from the posterior part of the intestine which is profusely supplied with blood (Carter and Beadle 1931; Huebner and Chee 1978). The Indian catfish, Clarias batrachus, which subsists in shallow derelict waters at night emerges from the water to feed on earthworms (Dehadrai and Tripathi 1976). During such a time, it uses the suprabranchial chamber membranes and the labyrinthine organs for respiration. The organs are similar to those of Clarias mossambicus, an African catfish (Figs. 65,66; Maina and Maloiy 1986). Other Indian species such as the climbing perch, Anabas testudineus, and the Cuchia eel, Amphipnous cuchia, are highly terrestrial, spending much of their time out of water (Munshi and Hughes 1992). Anabas (weighing 29 to 51 g) acquires about 53.6% of its  $O_2$  need from air (Hughes and Singh 1970b). Due to the lower metabolic rate and the relatively greater



**Fig. 65.** a Respiratory organs of the catfish, *Clarias mossambicus*, showing the gills, g, labyrinthine organ (o) and suprabranchial chamber membrane, s. b A gill arch showing gill filaments, f. c Tree-like labyrinthine organ which terminates in rounded knobs,  $\succ$ . d Suprabranchial chamber membrane which has many blood vessels,  $\blacktriangleright$ . a *Bar* 0.2 mm; b 0.1 mm; c 0.15 mm; d 0.15 mm. (Maina and Maloiy 1986)

amount of dissolved O<sub>2</sub>, at lower temperatures (17 to 20 °C), specimens of *Anabas* weighing up to 30g survive well for a few months without air breathing but at higher temperatures (30 to 31 °C) even smaller fish (10 to 15g) succumb within 24 h (Dube 1972). *Amphipnous* has a particularly high O<sub>2</sub> capacity exceeding 20 vol% and high hemoglobin-O<sub>2</sub> affinity (Lomholt and Johansen 1976): after 30 min



**Fig. 66.** Views of the surface of the labyrinthine organ of the African catfish, *Clarias mossambicus*, showing the intense vascularization in form of surface transverse capillaries (*c*) which are separated by bare tracts,  $\succ$ , and  $\diamond$  (*inset*), areas that contain mucus cells, *g* (*inset*).  $\rightarrow$  (*inset*), transverse capillaries, *e*, erythrocytes contained in the transverse capillaries; *w*, white blood cell. *Bar* 13 µm; *inset* 10 µm

of apnea, only a modest drop in arterial saturation (90 to 60%) occurs in the fish. The secondary reduction of the suprabranchial chambers and the increase in the gill respiratory surface area in some species of *Dinotopterus* (Clariidae) in the deep Lake Malawi and that in *Sandelia capensis*, an anabantoid fish in the South



Fig. 67. Plot between total respiratory surface area and body mass in different species of fish. Energetics, habitat, and mode of respiration determine the development of the gas exchangers. The tuna, a highly energetic fish, has remarkably extensive gill surface area while in the airbreathing fish, gill development is generally inhibited. In the air-breathing fish, e.g., *Anabas testudineus* and *Saccobranchus fossilis*, the accessory respiratory organs have a high diffusing capacity of  $O_2$ . This compensates for the reduction in the development of the gills. Data on *Oreochromis* from Maina et al. (1996a) and those of the other species from Hughes and Morgan (1973) and other publications

African Cape region (Barnard 1943), may be viewed as a reversion to totally aquatic respiration in fish formerly adapted to aerial breathing. A greater spacing of secondary lamellae, an adaptation which prevents gill filament adherence and collapse in air, characterizes most air-breathing fish (e.g., Todd and Ebeling 1966; Hughes and Morgan 1973). In marine air-breathing fish, this situation occurs in Sicvases (Ebeling et al. 1970) and in the mudskipper, Periophthalmus vulganris (Singh and Munshi 1968). In Periophthalmus schlosseri (Schöttle 1932), the first gill arch is replaced by a well-vascularized epithelium and the fish has only three pairs of functional gill arches. In Gillichthys (Todd and Ebeling 1966), the gills are short and the total respiratory surface area is less than in those species which do not breathe air. The more amphibious fish, e.g., Mnierpes (Graham 1973), have stronger gills (with a smaller surface area) which are less susceptible to collapsing out of water than in aquatic ones, e.g., Blennius pholis (Milton 1971). A smaller gill surface area reduces the rate of loss of O, through the gills into the hypoxic external medium. Shunting of blood within the gill system reduces the energetic cost of branchial perfusion (Satchell 1976; Randall et al. 1981; Smatresk and Cameron 1982a,b). The bimodal breathers in general have smaller respiratory surface area than the entirely water-breathing types (Figs. 67,68, Table 26). The extremely thin blood-gas barriers of the accessory respiratory organs provide a greater diffusing capacity than the gills (Table 26; Maina and Maloiy 1986). The



**Fig. 68.** Correlation between total gill surface area and body mass in fish living in different waters and exhibiting different modes of respiration. Marine endothermic fish which include the tuna have relatively high values while air breathers, especially the obligate ones, have low values. (Pelzenberger and Pohla 1992)

Species/organ	Body wt. (g)	Thickness of tissue barrier (µ)	Surface area (mm <sup>2</sup> g <sup>-1</sup> )	Dt $(ml O_2 min^{-1}mmHgkg^{-1})$
Air-breathing fish				
Anabas testudineus	100			
All gills		10.00	47.2	0.0071
Labyrinthine organ		0.21	32.0	0.2286
Heteropneustes fossilis	100			
All gills		3.58	57.7	0.0242
Air sac		1.60	30.7	0.0288
Skin		98.00	200.0	0.0031
Clarias mossambicus	458			
All gills		1.97	19.30	0.0213
Suprabranchial chamber		0.313	7.79	0.050
Labyrinthine organ		0.287	4.65	0.070
Channa punctatus	_			
All gills		2.033	71.88	0.0530
Suprabranchial chamber		0.780	39.17	0.0753
Labyrinthine organ		-	-	-
Amphipnous cuchia	-			
Áir sac		0.435	4.84	0.0165
Skin		0.44	227.5	-
Channa striatus				
Suprabranchial chamber	-	1.359	-	-
Clarias batrachus	-			
Labyrinthine organs		0.55	-	-
Water-breathing fish				
Tinca tinca	141	2.47	228.0	0.1493
Salmo gairdneri	35	4.30	260.0	0.1180
Tuna	100	0.50	2000.0	6.000
Opsanus tau	100	5.00	210.0	0.0630
Latimeria chalumnae	10 000	5.00	18.9	0.0057
Invertebrates				
Carcinus maena	100	5.00	744.0	0.0540
37				

**Table 26.** Morphometric pulmonary diffusing capacity of the tissue barrier (Dt) and measurements for respiratory surfaces of the gills and air-breathing organs of some air-breathing fish. Data on some water-breathing fish and invertebrates are included for comparison

Sources of data: Anabas, Hughes et al. (1973); Heteropneustes, Hughes et al. (1974); Clarias mossambicus, Maina and Maloiy (1986); Carcinus, Scammell and Hughes (1981); Nautilus, Perseneer (1935); rest Hughes and Morgan (1973).

fact that the gills have been widely retained in virtually all transitional breathing fish shows their importance for  $CO_2$  clearance in water. In amphibians, the change to terrestriality and air breathing is accompanied by greater dependence on the lungs for gas exchange (Lenfant and Johansen 1967).

# Air Breathing: the Elite Respiration

"The external gas exchangers that have evolved in higher organisms – fish gills, bird lungs, and alveolar lungs of amphibia, reptiles, and mammals have some basic features in common, irrespective of the different principles that determine their functioning in detail." Weibel (1984a)

# 6.1 Is the Surface of the Lung Dry, Moist, or Wet? Do Real Air Breathers Exist?

Water forms an important structural and functional constituent of the intercellular and intracelular lung tissue (e.g., Bastacky et al. 1987). Furthermore, a hydrated layer lines the air spaces of the lung (e.g., Fishman et al. 1957; Cantin et al. 1987; Chinard 1992). In the larger air spaces, the aqueous layer is comprised mostly of mucus, a glycoprotein-containing phase which is about 98% water (e.g., Sturgess 1979). The mucus forms an important source of moisture which humidifies the inhaled air, traps solid particles, and protects the ciliated epithelium. At the alveolar level, the hydrated layer occurs in form of an aqueous subphase in which proteins, carbohydrates, ions, and surfactant are dissolved. In the vertebrate lungs, where detailed investigations have been carried out, gas exchange occurs across an extracellular alveolar fluid film which lines the surface. The lining has been lucidly demonstrated by Finley et al. (1968), Weibel and Gil (1968), Kikkawa (1970), Bastacky et al. (1987, 1993), and Hook et al. (1987). In the airways, the thickness of the surface liquid lining is 20 to 150µm (Widdicombe 1997) while on the alveolar surface, the thickness ranges from 0.1 to  $0.24 \mu m$ (Weibel and Gil 1968; Bastacky et al. 1993, 1995; Stephens et al. 1996). In the human lung, it has been estimated physiologically (e.g., Rennard et al. 1986) that the epithelial lining fluid (ELF) makes up 20 to 40 ml while through morphometric techniques (e.g., Untersee et al. 1971; Gorin and Steward 1979), the ELF was estimated to range from 15 to 70 ml. The alveolar fluid layer contributes significantly to the gas exchange function of the lung. During strenuous exercise, accumulation of extracellular fluid on the surface of the lung is thought to cause a transient decrease in the membrane-diffusing capacity of the lung (Manier et al. 1991). In a delicate process which includes regulation of hydrostatic and colloidal osmotic forces across the capillary wall, the pulmonary surface is kept moist but not flooded (Levine et al. 1965; Fishman 1972). In the mammalian lung, 1 to 4% of the blood fluids lost to the surrounding tissue is efficiently carried away by the lymphatic vessels (Comroe 1974). When introduced into the alveoli, water quickly passes into the pulmonary blood capillaries (Effos and Mason 1983; Jones et al. 1983; Effros et al. 1992; Grimme et al. 1997). This is because the pulmonary blood capillary (microvascular hydrostatic) pressure of about 1.1 to 1.2kPa (which is inclined to filter blood into the alveoli) is always well below the colloidal

(osmotic) pressure of the plasma proteins of about 3.3 to 4kPa which draws fluid from the alveoli into the blood (Comroe 1974). Normally, the lung would be expected to be dry and to absorb but not filter fluid in its circulation.

Water is not a foreign factor in the lungs of the air-breathing vertebrates. Due to their extensive surface area, during prenatal development, the lungs play an important role in the production of the amniotic fluid (Setnikar et al. 1959). During fetal life, the lung is filled with liquid that flows into the developing air spaces in response to Cl<sup>-</sup> secretion across the epithelium of the respiratory tract (Olver and Strang 1974). In sheep, the rate of production may be as high as about 2 mlkg<sup>-1</sup>h<sup>-1</sup> (Normand et al. 1971) and at birth the total pulmonary fluid is estimated to be as much as 30 mlkg<sup>-1</sup> (Normand et al. 1971). Experimental or congenital obstruction of the trachea or the bronchi leads to intrapulmonary accumulation of fluid and overdistension of the lung (Potter and Bohlender 1941). In some animals, e.g., the turtles, which have relatively high arterial pulmonary blood pressures and low osmotic pressure of the plasma proteins (White and Ross 1965; Shelton Burggren 1976), water may constitute a significant portion of the  $O_2$  diffusional pathway. In *Pseudemys scripta*, at a blood flow rate of 12 to 14 mlkg<sup>-1</sup> min<sup>-1</sup>, during ventilation, 20 to 40% of the fluid may be left behind in the nonvascular part of the lung tissue (Burggren 1982b): the net loss of plasma into the tissues in the ventilated reptile lung is 10 to 20 times greater than in the mammalian lung. Well-developed intercellular junctions in the air-blood barrier were demonstrated in the lung of the turtle, Pseudemys by Bartels and Welsch (1983) by freeze fracture electron microscopy: they may account for the remarkable "leakiness" of the pulmonary capillaries of the turtle's lung. It is interesting to note that in insects, the terminal tracheoles contain fluid which is osmotically absorbed into the surrounding cells or released in the air tubules depending on the metabolic state of the tissues (Wigglesworth 1953). In mammals, about 2 to 3 days before birth (Kitterman et al. 1979; Dickson et al. 1986) or during labor (Bland et al. 1982; Brown et al. 1983), pulmonary filtration stops and absorption of the intrapulmonary fluid starts. Both processes are influenced by the level of circulating epinephrine or isoproterenol (e.g., Walters and Olver 1978), adrenaline (Brown et al. 1983), or Na<sup>+</sup> flux out of the lung lumen (Cotton et al. 1983; Bland 1990; Chapman et al. 1994). In the newborn calf, the mean pulmonary arterial pressure drops from 8.7 to 4kPa in the first 6h of birth (Reeves and Leathers 1964). The intrapulmonary fluid is physically expelled through: (1) the upper respiratory airways due to pressure exerted on the fetal thorax during transit through the pelvic canal (Borell and Fernstrom 1962; Karlberg et al. 1962); (2) absorption into the lymphatics (Humpreys et al. 1967; Strang 1967; Gonzalez-Crussi and Boston 1972); and (3) transfer by the pulmonary capillaries (Egan et al. 1975; Hutchison et al. 1985). The fluid remaining in the lung after delivery is cleared from the lung within a couple of hours (Humpreys et al. 1967; Fletcher et al. 1970; Adams et al. 1971; Bland et al. 1980).

The general perception of the surfaces of the internalized gas exchangers of the terrestrial air breathers as dry organs needs to be reconsidered. It is more realistic to consider them as moistened but not flooded. Oxygen does not diffuse efficiently across dry tissue barriers. In the toad (Dupre et al. 1991), dehydration reduces cutaneous gas diffusion capacity. Owing to a dependence on water for

efficient respiration across the lungs, it could be argued that air breathing has not yet strictly evolved. However, since terrestrial animals seek and take up  $O_2$  from air and discharge  $CO_2$  into the same, according to the general definition, such animals are recognized as air breathers. This also includes those insects which have secondarily invaded water and extract  $O_2$  from air bubbles regularly ferried from the surface (Sect. 6.12). It is only if an air breather was to "aspirate" water into its lungs, a rather rare retrogressive move which has, nevertheless, developed in some simple animals, e.g., aquatic pulmonate gastropods (Sect. 5.6.1) after the ctenidia (gills) were lost and the mantle cavity converted into a "water lung", that an animal is categorized a water breather.

About 1 week before hatching, in the loggerhead turtle, *Caretta caretta* (Perry et al. 1989a), and during stage one of development of *Salamandra salamandra* (Goniakowska-Witalinska 1982) the lungs are filled with water. The intrapulmonary fluid is removed soon after hatching (Gatzy 1975) and in sheep soon after birth (Strang 1977; Bland 1990). It has been postulated that the flooding of the lungs with fluid provides biomechanical support necessary for proper development of the air passages and spaces (Alcorn et al. 1977; Maloney 1984). It is envisaged that after the contraction of the smooth muscles, the pressure in the fluid-filled lung increases, leading to an outward formation of the air cells (e.g., Marcus 1937). In deviation from the moist surface requirement, some animals such as the minute aerial arthropods are known to exchange respiratory gases through a dry cuticle (Krogh 1941). The same process probably occurs in the book lungs of some spiders (Zoond 1931; Herreid et al. 1981; Strazny and Perry 1984). The low O<sub>2</sub> demands in such animals may account for the adequacy of O<sub>2</sub> transfer across a dry surface.

# 6.2 Lung and Swim Bladder – Which Developed Earlier and for What Purpose?

Swim bladders and lungs are said to be homologous (Fänge 1983). Lungs are envisaged to have evolved in the Paleozoic freshwater vertebrates as a means for adaptation to hypoxic stress (e.g., Romer 1972; Graham et al. 1978a,b). All bony fish (Osteichthyes) have a swim (air) bladder at least at one stage of their development (e.g., Marshall 1960). In most contemporary fish, the swim bladders are hydrostatic with no discernible respiratory role while some teleosts breath air by a lung or a swim bladder (Fänge 1983; Alexander 1993). The bladder is historically interesting for the following reasons: (1) in some fish, the organ serves as an auxiliary respiratory organ and hence its biology may shed some light on the evolution of the respiratory organs and processes, (2) its role as a hydrodynamic organ helps to explain some of the adaptive processes which were essential for subsistence in water (a medium of relatively higher viscosity) and the necessary subsequent postural and locomotory biomechanical adjustments for living on land, and (3) the capacity of the organ to passively secrete and concentrate gases to several hundred atmospheres of pressure far in excess of those in blood and in the external environment (e.g., Scholander 1954; Marshall 1960; Fänge 1976; Kobayashi et al. 1989a,b) is a most intriguing and unique biophysical phenomenon. Recent works and reviews on this aspect include those of Gerth and Hemmingsen (1982), Fänge (1983), Kobayashi et al. (1989a,b), and Pelster and Scheid (1992a,b). It is now generally thought that a multiplicative salting-out effect in the countercurrent system of the rete mirabile (e.g., Pelster et al. 1988c); (Fig. 39) explains the passive secretion of gases into the swim bladder. The lactate acid which is produced by glucose metabolism in the gas-gland epithelium (Pelster and Scheid 1991, 1993; Figs. 39,70) is secreted into the blood (D'Aoust 1970; Kobayashi et al. 1989a,b; Pelster et al. 1989), where it acidifies the blood, increasing the blood PO<sub>2</sub> by Bohr and Root effects, PCO<sub>2</sub> by conversion of HCO<sub>2</sub> into  $CO_2$ , and the inert gases by reducing their physical solubility (= salting-out effect) (Pelster et al. 1988c). Back diffusion of CO<sub>2</sub> in the rete mirabile increases the blood pH, enhancing the secretion of  $O_2$  into the swim bladder (Kobayashi et al. 1990). Oxygen constitutes the greater proportion up to 95% of the gas mixture secreted into the swim bladder (Wittenberg 1965) with  $N_2$ , CO<sub>2</sub>, and rare gases like argon constituting lesser proportions (Tait 1956). The PO<sub>2</sub> in the swim bladders of fish which live in great depths may exceed 150 atm (Scholander and van Dam 1953). In such cases, the PO<sub>2</sub> in the swim bladder may be as much as  $10\,000$  times that in the surrounding water (Kanwisher and Ebeling 1957). Considering that all the gases must in the first instance be derived from water, to generate such a pressure differential calls for a remarkably efficient concentrating process. In the European eel, Anguilla anguilla, under hypoxic conditions, gas deposition and blood flow through the rete mirabile were significantly reduced (Pelster and Scheid 1992c). Fish which live at depth may required a substantial amount of work to secrete air into a swim bladder against partial pressure gradients (Alexander 1982). In fish such as the swordfish, Xiphius, which have been recorded to surface from a depth of 100 m in less than 5 min (Carey and Robinson 1981) and the lantern fishes which are recorded to commute between 300 and 50 m at rates in order of  $160 \text{ km h}^{-1}$  (Barham 1966), the process must be remarkably efficient.

The chronology of the evolvement of the lung and the swim bladder and whether the two organs are homologous is an ongoing debate (e.g., Romer 1972; Fänge 1976; Liem 1987a). While the early evolution in potentially O<sub>2</sub>-deficient waters may be a pointer to an essential initial respiratory role (e.g., Liem 1991), on the contrary, subsistence in the pelagic marine well-oxygenated waters emphasizes a predominant hydrostatic function (Liem 1987a). It is conceivable that swim bladders may have been necessary to offset the increase in weight with the development of the bony skeleton in fish (Alexander 1993). However, the hydrodynamic and respiratory roles of the swim bladder are by no means mutually exclusive and neither can the possibility that the two organs evolved independently of each other be discounted based on the presently available data (e.g., Liem 1989; Hedrick and Jones 1993; Alexander 1993). It is plausible that an initial simple saccular air-containing organ may have served both roles, as evident in some of the Central American catfishes (e.g., Gee 1976, 1981) and the subsequent functional and structural differences may have arisen to meet specific environmental demands. In Hoploerythrinus unitaeniatus, a small tropical fish, during the periods between the respiratory cycles, the concentration of  $O_2$  in the swim bladder is reduced to 8%, indicating a very high degree of extraction of molecular  $O_2$  from the organ (Carter and Beadle 1931). The absorption rate of  $O_2$  from the swim bladder of the mudminnow, Umbra limi, is very high (Safford-Black 1944). Some teleosts in which the swim bladder functions as a lung include Arapaima (Osteoglossidae), Gymnarchus (Mormyridae), Erythrinus (Characinidae), Umbra (Esocidae), Notopterus (Notopteridae), and Lepisosteus (Potter 1927; Dehadrai 1962; Fänge 1976). Gillichthys consumes O<sub>2</sub> from its physoclistous (closed) bladder to support metabolism during the short transitional period (average 17 min) from aquatic to aerial respiration (Todd and Ebeling 1966). Through a buccal force pump mechanism, physostomatous fish (i.e., those with open bladders) which live close to the surface of the water inflate their bladders with air gulped from the surface of the water (Fänge 1976). Some physoclistic fish are known to use a pneumatic duct during their larval stage for the first filling of their bladders (e.g., Jacobs 1938). In Erythrinus unitaeniatus, the middle region of the physostomatous bladder is well vascularized and is used for gas exchange (Carter and Beadle 1931). The tarpon, *Megalops*, regularly ventilates its physostomatous swim bladder even in normoxic water (Böhlke and Chaplin 1968). The tarpon's swim bladder has been described as being lung-like since it has alveolar-like tissue (Shlaifer and Breder 1940): the swim bladder facilitates aerial respiration, a process which augments the tarpon's aquatic respiration and enhances the fish's metabolic scope of activity (Johansen 1970). In the holostean air-breathing fish, Amia calva, and the teleost, Arapaima gigas, gas exchange is effected through a physostomatous swim bladder which is supplied with blood from the dorsal aorta, coeliac, or mesenteric arteries. While not directly involved in gas exchange, the physoclistous swim bladder of Dormitator in hypoxic water generates positive buoyancy, which enables the fish to expose its vascularized frontal skin to air (Todd 1973). The swim bladder of the teleost, Oreochromis alcalicus grahami, a small cichlid fish which lives in the alkaline lagoons of the volcanic Lake Magadi of Kenya (Wood et al. 1994; Maina et al. 1996b), has a well-vascularized swim bladder (Figs. 69,70); (Maina et al. 1996b). The bladder is probably used for gas exchange during the night when the water is virtually anoxic (Narahara et al. 1996): on exertion, e.g., after being chased with a net, the fish are observed to skim the surface of the water, probably taking air into the bladder or ventilating the gills with the better oxygenated top layer of water. In the physostomatous bladders, the neuromuscular mechanisms involved in the intake and expulsion of air are not well known. Inspiration may be effected by the buccal force pump while expiration may be brought about by the activity of the smooth muscles of the pneumatic duct of the bladder. In *Hoploerythrinus*, the pneumatic duct is notably wide and muscular (Lüling 1964) and in the eel, Anguilla anguilla, the duct is adapted for gas exchange by diffusion and has a distinct circulatory system (Fänge 1953). In most physostomatous fish, e.g., in the salmonids (Fahlén 1971), cyprinids (Plattner 1941), the electric eel (Evans 1929), and notopterids (Müller 1950), the entrance of the pneumatic duct to the esophagus is guarded by smooth and striated muscle sphincters. Dependent on species, gas is released through the pneumostome by deflation. This occurs by a reflex action, the so-called gas spitting reflex, which is initiated by a reduction of the external pressure and/or by nervous excitation (Fänge 1976). The elasticity of the bladder wall due to contraction of the well-developed smooth muscles (Maina et al. 1996b) and contraction



Fig. 69a-c

of body wall muscles (Evans and Damant 1928) may help in the release of gases. In an aquatic animal, the use of a gas exchanger for hydrostatic control is inefficient since when  $O_2$  is removed and little if any  $CO_2$  is secreted back, the animal becomes less buoyant and sinks (Gee 1976; Gee and Graham 1978). The animal then has to surface to refill the lung. Oxygen consumption (i.e., removal of O<sub>2</sub> from the lung) has to be synchronized both with hydrostatic requirements and respiratory frequency. During hypoxic episodes, the cephalopod mollusk, Nautilus (often referred to as a living fossil due to its multichambered shell which resembles that of extinct animal forms from the early Paleozoic) draws O<sub>2</sub> from the buoyancy chambers of the shell (Boutilier et al. 1996): in a 459-g (fresh weight) Nautilus at 18 °C, the shell volume is  $100 \text{ cm}^3$ , the O<sub>2</sub> store in the shell is  $6.9 \text{ cm}^3$ and a partial pressure gradient of O<sub>2</sub> of 7.3 kPa exists. In a hypometabolic state, it is estimated that the O<sub>2</sub> stores in the shell should support metabolic rate for as long as 6h. In the European eel, Anguilla anguilla, 83% of O<sub>2</sub> removed from the blood was secreted into the swim bladder and only 17% was used for metabolic purposes (Pelster and Scheid 1992c).

The swim bladder of the teleosts may have evolved independently from the gas chambers of Polypterus and the lungfishes (Dipnoi), a respiratory organ having been more urgent for gas exchange in the early hypoxic aquatic medium. On the other hand, it has been surmised that the swim bladder may have evolved entirely for buoyancy control and in some cases, e.g., in the Dipnoi, secondarily acquired a respiratory role. The lungs of the Dipnoi and the ancestral actinopterygian fish such as Amia, Polypterus, and Lepisosteus are considered to be homologous with the swim bladders of fish (Packard 1974). In teleosts, Dipnoi and Polypteridae, the swim bladder and the lungs originate as outgrowths from the wall of the foregut, the primodial air-sac/lung becoming evident very early in development (Packard 1974). Both organs are enervated by branches of the vagus and sympathetic nerves. However, this is as far as the similarity between the two organs goes. The swim bladder: (1) arises from the dorsal or lateral walls of the foregut (e.g., Goodrich 1930) and the lungs from the floor (ventral aspect) of the pharynx, (2) the lungs receive left and right pulmonary arteries which originate symmetrically from the last pair of epibranchial arteries but the swim bladder is supplied with arterial blood from the aorta (Goodrich 1930), (3) the lungs remain connected to the pharynx while physoclistic swim bladders totally lose connection with it, and (4) the lungs are generally lined by a surface active factor, a complex mixture of phospholipids, neutral lipids, and proteins. Due to their ventral location and the inherent problems of balance which would accompany such an organ, the lungs may have all along been respiratory while the dorsally located swim bladder could combine hydrostatic and respiratory roles. In teleosts, the swim bladder is mainly

**Fig. 69.** a Oreochromis alcalicus grahami, a small tilapiine cichlid fish which lives in the highly alkaline Lake Magadi of Kenya where there are dramatic diurnal fluctuations of levels of  $O_2$  in water. **b** The swim bladder, *x*, and the gills, *g*, are very well developed. **c** Latex rubber cast of the gills, *g*, and the swim bladder, *b*, showing the air way, marked by a cannula, *r*, and vascular connections,  $\succ$ , between the two organs. **a** Bar 7 mm; **b** 5 mm; **c** 3 mm. (Maina et al. 1996b)



**Fig. 70a,b.** Views of the surface of the swim bladder of *Oreochromis alcalicus grahami*, showing the profuse vascularization, v; g gas-gland cells. It is probable that the physostomatous swim bladder in this species may be utilized for gas exchange during extreme hypoxia **a** Bar 20 $\mu$ m; **b** 17 $\mu$ m. (Maina et al. 1996b)

a hydrostatic organ and is thus structurally much less complex than the Dipnoan (Fig. 64) and Polypteridae lungs which are essentially respiratory (Klika and Lelek 1967). The nonseptate saccular lungs of the urodele amphibians, the salamanders, e.g., *Cryptobranchus* and *Necturus*, appear to be largely hydrostatic (Noble 1931; Guimond and Hutchison 1973b). However, the well-developed finely subdivided lungs of some aquatic frogs, e.g., *Xenopus laevis* and *Pipa pipa* (Czopek 1962a; Goniakowska-Witalinska 1995), which inhabit hypoxic waters and the salamanders, *Amphiuma tridactylum* (Stark-Vancs et al. 1984) and *Salamandra salamandra* (Goniakowska-Witalinska 1978; Meban 1979), which have an extensive surface area, are predominantly respiratory.

## 6.3 Evolution of Air Breathing and Terrestriality: the Limitations

In virtually all terrestrial habitats, the atmospheric air is never 100% saturated with moisture. On this account, terrestrial habitation is accompanied by an enduring conflict between the need to procure  $O_2$  and the necessity to preserve water. Extended emergence from water was impossible until the two processes could be harmonized. Conservation of water entailed development of an impermeable surface cover and invagination of the respiratory organs (e.g., Hadley 1980; Loveridge 1980; Ouinlan and Hadley 1993). Terrestrial air breathers are in constant danger of desiccation especially through respiratory water loss. No other internal organ is in more intimate contact with the external environment as the lung. Everyday, about 12000l of air are filtered by the human lung and 6000l of blood perfuse it (Burri 1985). The interaction occurs over a surface area of nearly  $150 \text{ m}^2$  across a blood-gas barrier  $0.6 \mu \text{m}$  thick (Gehr et al. 1978). In most respects, air breathing is a less complicated process than water breathing. Whereas in air breathing only O<sub>2</sub> and CO<sub>2</sub> are exchanged, in water, superimposed on the gas exchange process is ionic exchange and elimination of end products of nitrogen metabolism (e.g., Goldstein 1982; Zadunaisky 1984). Even if all other factors were held constant, gill respiration should energetically be more demanding than air breathing owing to the fact that except for the animals which are iso-osmotic to seawater, energy has to be expended to selectively regulate ionic flux. A large number of mitochondria rich cells (= ionocytes = chloride cells), occur, e.g., in fish (Fig. 50; Maina 1990a, 1991) and the crab gills (Maina 1990b; Figs. 13,71). Aquatic animals are exposed to a medium which has the same heat capacity as blood compared with the air breathers where the heat capacity of air is 3000 times lower than that of blood (Table 9). Since for equivalent volumes water contains about one thirtieth of the volume of  $O_2$  in air, to extract an equal amount of  $O_2$ , aquatic breathers expose their blood to a heat sink which has a heat transfer capacity  $9 \times 10^4$  times greater than air (Steen 1971). This may explain why only some degree of endothermy has evolved in a few aquatic animals and in most cases only in specific tissues. In fish, endothermy occurs in a few fish, e.g., the swimming muscles of the tuna and some sharks (e.g., Carey and Teal 1966, 1969; Block 1991b). Endothermic-homeothermy, an evolutionary step which delinks an animal's physiological processes from the environmental thermogenic



Fig. 71a-d. Gills of the freshwater crab, *Potamon niloticus*, showing: a the basal aspect of the epithelial cells with numerous membrane infoldings which attach onto the surface cuticle (*c*); mitochondria, *m*, and a micropinocytotic vesicles,  $\succ$ . b A closeup of the basal aspect of an epithelial cell showing the attachment of the basal infoldings,  $\rightarrow$ , to the cuticle, *c*. c A view of the mitochondria, *m*, in close proximity to the micropinocytotic vesicles,  $\succ$ . d The numerous mitochondria, *m*, with profuse cristae in close proximity to micropinocytotic vesicles,  $\succ$ . The general structural features of the gills of the crab illustrate a compromise design between a thin water-blood (hemolymph) barrier and osmoregulatory exchange. a *Bar* 0.75 µm; b 0.5 µm; c 0.3 µm; d 0.3 µm. (Maina 1990b)

fluctuations, has been exclusive to the elite air-breathing animals, namely mammals and birds (Romer 1967; Crompton et al. 1978; Bennett and Ruben 1979; Hochachka 1979). At the extreme small body sizes, e.g., in the about 2-g Cuban bee hummingbird, the Etruscan shrew, and the Thai bumblebee bat, endothermic homeothermy is extremely expensive to maintain. Hummingbirds hover more than 100 times a day as they feed on nectar (Krebs and Harvey 1986), each bout lasts about a minute, and 20% of the daylight hours are spent feeding (Diamond et al. 1986). At night, some but not all hummingbirds undergo torpor to conserve energy (Lasiewski 1963b; Bucher and Chappell 1989). Though energetically expensive (Else and Hubert 1981, 1985), regulation of body temperature at a higher level imparts certain biological advantages. The biochemical reactions are faster, the rate of information processing is rapid (through quicker nervous coordination and response to environmental changes), and because diffusion is a physical process, gas transfer across the respiratory organs is faster. It is envisaged (e.g., Randall et al. 1981) that endothermy in terrestrial vertebrates evolved in temperate regions where the low ambient temperature may have called for dissociation of the body temperature from the ambient. In whichever way it evolved, the origin of endothermy had to await development of more efficient respiratory organs and processes to provide the large amounts of O<sub>2</sub> necessary to support a high metabolic rate (Wood and Lenfant 1979; Duncker 1991). Amazingly, for somewhat different purposes, some plants, e.g., aroid plants such as Philodendron selloum and Arum maculatum (Nagy et al. 1972; Seymour 1997) and the lotus plant (Seymour and Schultze-Motel 1996) have evolved endothermy. Heat production in the two aroid plants rivals the high levels generated by the flight muscles of bees (Seymour 1997)! In P. selloum, a 125-g spadix produces about 9W of heat to maintain a temperature of 40 °C against an ambient one of 10 °C (Koch et al. 1983, 1984; Seymour 1991), surpassing heat output in some thermoregulating animals (Seymour 1997).

In abandoning liquid breathing and adjusting to air breathing, the chances of survival of animals on land depended on the level of structural development and functional adequacy of the gas exchangers as well as that of the necessary homeostatic changes, especially those relating to acid-base balance (e.g., Robin et al. 1969; Rahn 1974; Reeves 1977). Among the different animal lines, air breathing has evolved independently many times, following different pathways (e.g., Truchot 1990; Graham 1994). The possession of lung-like structures by nonteleostean groups of ray-finned fish is paleontologically well documented (e.g., Romer 1972; Jarvik 1980). Primeval lungs which developed from the floor of the junction of the esophagus and the pharynx are recognized in the ancestral armored fossil placoderm fish, Bothriolepis (Denison 1941), and among some of the oldest fishes, e.g., the bichirs, Polypterus (Lauder and Liem 1983; Burggren et al. 1985a). The elaborate dipnoan lungs (Fig. 64) are acclaimed to be the possible precursors of those of the tetrapods (e.g., Inger 1957; Romer 1967; Løvtrup 1977; Gardner 1980). The earliest definite lungs in the chordates occurred in the ancestral fishes. Lungs are present in all extant holosteans, dipnoans, and predominate in the chondrosteans. The evolution of the lungs enabled animals to fully emerge from water and enjoy the physical advantages of occupying an O,-potent environment and an almost limitless habitat. So perfected has the process of air breathing become that most of such animals soon succumb on submersion in water irrespective of the concentration of  $O_2$ . Inundation of the pulmonary airway tree prohibits gas exchange in the lung, producing obstructive asphyxia. Physiologically, this is characterized by hypoxia, hypercapnia, and acidosis. Depending on the nature of the aspirated liquid, ionic imbalance also occurs (Moritz 1944) leading to complications such as hydremia, hemoconcentration, hemolysis of the erythrocytes, ventricular fibrillation, cardiac arrest, and death.

The colonization of land by the tetrapods at about the Early Devonian and that of the higher plants in the Upper Silurian or a little earlier (e.g., Romer 1967; Gray 1985a,b) formed important steps in the evolution of the major ecosystems on Earth. The problems and challenges which accompanied air breathing and transition to land included: (1) postural problems as a result of increased effect of gravity on the body as animals lost the buoyancy provided by the water; (2) potentially large ecological variability of water vapor pressure and hence risk of desiccation; (3) thermal instability due to the low thermal capacity of air; and (4) problems of reproduction and fertilization (in water the gametes could simply be released into the water). Evidence of a causal relationship between hypoxia and evolution of air breathing is overwhelming (Sects. 5.1 and 6.3). Air breathing in general and terrestrial habitation in particular entailed a conflict between  $O_2$ uptake, water conservation, and acid-base balance. To effect the change, certain trade-offs and compromises were necessary. It is probably in an attempt to maximize on the advantages while giving up very little that a substantial number of animals after having evolved the capacity for aerial respiration, a major preadaptive step towards land habitation, opted to subsist in water. Indeed, it is only in arthropods and vertebrates where extensive evolutionary adaptation to air breathing has occurred. The main problem which faced the pioneering colonizers of the aerial biotope was elimination of CO<sub>2</sub>. The challenges in acid-base balance are apparently less in water than in air due to the high solubility of  $CO_2$  in water. The absolute values of PCO<sub>2</sub> and HCO<sub>3</sub><sup>-</sup> ions in blood and tissues are lower in water breathers than in air breathers (Tables 2,22). In reptiles, the first exclusively lung breathers (except for the aquatic species which significantly use the skin), the PCO<sub>2</sub> and the concentration of HCO<sub>3</sub><sup>-</sup> ions vary remarkably depending on ecological adaptations, morphological design, and lifestyle (Jackson 1986). In Squamata and Crocodilia, the mean concentration of  $HCO_3^-$  ions is  $15 \text{ mMl}^{-1}$  at a PCO<sub>2</sub> of 2 kPa and in Chelonia, the concentration of  $HCO_3^{-1}$  ions is 39 mM  $l^{-1}$  and the PCO<sub>2</sub> 4.5 kPa (Howell 1969). The PCO<sub>2</sub> ranges between 0.1 and 0.5 kPa and the concentration of HCO<sub>3</sub><sup>-</sup> ions from 5 to 15 mMl<sup>-1</sup> in the plasma of fishes (Heisler 1984). Owing to the greater amount of  $O_2$  in air, to maintain a PO<sub>2</sub> of 13.3 kPa in the alveolar air (at a respiratory quotient of 1), an air breather need only move 17 ml air min<sup>-1</sup> ml<sup>-1</sup>  $O_2$  compared with an aquatic animal which must move 480 ml water min<sup>-1</sup> ml<sup>-1</sup>O<sub>2</sub> to maintain an equivalent PO<sub>2</sub> in the gill water (Rahn 1967; Howell 1969, 1970): the alveolar PCO<sub>2</sub> in the air breather would be 6.7 kPa while in a water breather the PCO<sub>2</sub> in the gill effluent would be only 0.24 kPa. The ventilatory rate of an aquatic animal at 20 °C is 28 times (at that temperature the solubility of CO<sub>2</sub> is about 28 times greater than the solubility of O<sub>2</sub> in water!) that of an air breather: the PCO<sub>2</sub> of the blood of a fish is one-twenty eighth that of an
air breather. Thus, animals which endeavored and succeeded in attaining air breathing could drastically reduce ventilatory rate but would be faced with a profound increase in arterial PCO<sub>2</sub> which in terms of acid-base balance results in a respiratory acidosis. The alternative strategies which were used to surmount these obstacles included: (1) renal mechanisms which resulted in increased blood concentration of  $HCO_3^-$  ions thus maintaining a constant  $OH^-/H^+$  ratio and/or, (2) evolution of an alternative pathway other than the lung, e.g., the skin and the buccal cavity, for CO<sub>2</sub> clearance during the traumatic switchover of respiration from the gills to the modern lung (Fig. 57). Conceivably, the transition was interfaced by a relatively simple precursor of the modern lung which must have been, from perspectives of design and mechanical ventilation, relatively inefficient. The forfeiture of the skin, after the integument was covered with scales or armor to reduce water loss, in reptiles (Mertens 1960) was the initial step in the transfer of gill to lung respiration (Hughes 1963). During this time, the lung and respiratory mechanics developed to a level where the air-breathing organ could eliminate the entire load of CO<sub>2</sub> produced in the body. The role of acid-base balance was wholly shifted to the modern lung and, to a smaller extent, the kidney. At 20 °C, only about 3% of the CO<sub>2</sub> output is cutaneous in the tortoise, Testudo dendriculata (Jackson et al. 1976) and in the desert lizard, the chuckawalla (Sauromalus obesus) about 4% of CO<sub>2</sub> output and less than 2% of O<sub>2</sub> uptake occurs through the skin at 25 °C (Crawford and Schultetus 1970). In the landdwelling box turtle, Terrapene ornata, a large proportion of CO<sub>2</sub> is voided through the skin at the low hibernating temperatures (Glass et al. 1976). Gans (1970) observed that cutaneous respiration is an acquired mode of respiration unique to the modern amphibians without any relationship to the Devonian ancestral lineage of the extant tetrapods. Romer (1972) and Colbert and Morales (1991) have espoused the theory that the Carboniferous amphibians were completely heavily armored. If this is correct, the highly gas- and water-permeable skin characteristic of the extant amphibian species is an adaptive secondary condition. Clearly, in amphibious animals the skin was not as efficient as the gills in CO<sub>2</sub> elimination, as demonstrated by the fact that, despite the role the integumentary system played in gas exchange, it was still necessary for the level of the plasma HCO<sub>3</sub><sup>-</sup> ions to be elevated in the air breathers. The increase in the arterial PCO<sub>2</sub> and the concentration of the HCO<sub>3</sub><sup>-</sup> ions occurred as animals became more dependent on air for their O<sub>2</sub> needs, starting from the water-breathing fish through the various forms of bimodal breathers to the air breathers. Despite the differences in the PCO<sub>2</sub> and the concentration of the HCO<sub>3</sub><sup>-</sup> ions in the arterial blood of the water- and air breathers (e.g., in the carp and the bullfrog), the pH (about 7.9) of these animals at the same temperature (20 °C) is essentially the same (Howell et al. 1970; Table 2). The pH of the water-breathing tadpole at 20 °C is the same as that of the air-breathing frog at the same temperature despite the fact that the arterial PCO<sub>2</sub> and the concentration of the HCO<sub>3</sub><sup>-</sup> ions of the bullfrog are about five times greater than that of the tadpole (Erasmus et al. (1970/71). Conceivably, since the bimodal breathing occurred in O<sub>2</sub>-deficient waters, the O<sub>2</sub> affinity of blood of such animal forms should be lower, i.e., P<sub>50</sub> should be higher, the more aerial-breathing an animal is. The  $P_{50}$  of the blood of the air-breathing fish ranges from 2 to 2.7 kPa at 20 °C, values which are higher than those of water

breathers in similar habitats. In the amphibians, the most aquatic species have a  $P_{50}$  about one half that of the completely aquatic aerial species (Lenfant and Johansen 1967). The differences are accounted by the fact that air is a respiratory medium much richer in  $O_2$ .

### 6.4 Aerial Gas Exchangers: Structural and Functional Diversity

The air-breathing organs are thought to have evolved as special adaptations to the prevalent hypoxic conditions, presumably those which prevailed in the warm tropical waters of the late Devonian (e.g., Zaccone et al. 1995). Confinement of gas exchange to specific site(s) of the body where the process could be well regulated, water loss restricted, the inhaled air "cleaned" and physically modified, and better protection from toxicants and trauma afforded were prerequisites for efficient aerial respiration and successful terrestrial habitation. If the human lungs were like the external gills of fish, i.e., were evaginated and exposed to air, even in a moderately desiccating environment, the water loss would be about 5001 day<sup>-1</sup>. This value is about 1000 times greater than the normal loss (McCutcheon 1964). In arthropods, removing the cuticle (by scratching) increases cuticular water loss much more than it affects gas exchange (Ito 1953; Richards 1957). The reduction of the respiratory rate in the air breathers (owing to accessibility to a greater O<sub>2</sub> concentration) constituted an important step in the reduction of respiratory water loss. Most animals die if water loss exceeds 20 to 50% of their body mass (Adolph 1943). The majority of terrestrial vertebrates cannot tolerate a body temperature of 45 °C or more but, when provided with adequate water for evaporative cooling, they will withstand even higher temperatures (Calder and King 1974). The development of internalized gas exchangers (Figs. 4,5), however, was accompanied by certain functional limitations. While the gills, e.g., in crustaceans, mollusks, agnathans, and fish by virtue of their external location can be ventilated unidirectionally and continuously, the "dead-ended" lungs of the vertebrate air breathers can only be ventilated periodically and bidirectionally (i.e., tidally) through narrow opening(s) (Figs. 6,18). This constituted a major trade-off in the design of the aerial gas exchangers. While in the gills the configuration was compatible with the highly efficacious countercurrent system, as a consequence of their invagination, only the relatively inefficient uniform pool arrangement was tenable in the aerial gas exchangers (Figs. 6,35,36; Sects. 6.2 and 6.4). The internalized gas exchangers fail to maximally exploit the high ambient PO<sub>2</sub>: the inspired air is greatly diluted by the residual air, reducing the head pressure from 21 kPa to about 13 kPa, a loss of about one third of the potential partial pressure gradient. The arterial PO<sub>2</sub> in Amia calva, Synbranchus, and Neoceratodus (an obligate water-breathing lungfish) are below 1.3 kPa in water but are higher when held in air (Lenfant et al. 1966; Johansen et al. 1970a). In a resting human being, where the dead space is about 140 cm<sup>3</sup>, about 28% of the 500 cm<sup>3</sup> of the inhaled air does not reach the gas exchange region. Compensatory advantages such as low cost of ventilation in air, greater O<sub>2</sub> loading and transport due to presence of hemoglobin, and better ventilation-perfusion controls help overcome the appar-

ent limitations intrinsic to the design. On the plus side, tidal ventilation makes it possible for a much more stable and well-controlled local respiratory condition to be established. The alveolar  $PO_2$  is lower and the  $PCO_2$  higher than in the ambient air, the elevated alveolar PCO<sub>2</sub> in the vertebrate lung being used in the  $HCO_3^{-1}$  ion buffer system for pH regulation. Such microenvironments are impossible to create in those gas exchangers which are unidirectionally ventilated or directly exposed to the external respiratory medium. Mechanical ventilation is necessary for mass renewal of the air in contact with the respiratory surface(s) to maintain the partial pressure gradient essential for O<sub>2</sub> influx. The most elaborate of the respiratory processes are restricted to the large and phylogenetically more advanced animals. A highly developed nervous system and proper nervous integration were necessary to operate these complex organs and functions. Shrewdly, by isolating the lung from the air sacs, birds have developed a unidirectional continuous air flow in the parabronchial system of the lung within an invaginated, tidally ventilated organ! The tracheal system of insects has attained a remarkable state of efficiency through synchronization of spiracular opening with the ventilation of the air sacs (Weis-Fogh 1967). Intermittent breathing in reptiles (e.g., Glass and Wood 1983; Wood and Glass 1991), amphibians (De Jong and Gans 1969), and lungfishes (Lenfant and Johansen 1968) is possible because of their low O, needs and the great O, stores in their lungs. Episodic respiratory pattern constitutes an energy-saving strategy compared with the continuous mode of breathing in birds and mammals (Milsom and Jones 1979). Although incidental breathing is a common feature of the ectothermic vertebrates (Milsom 1988; Shelton and Croghan 1988), it occurs in hibernating nondiving mammals such as hedgehog, dormouse, and the little brown bat (e.g., D.W. Thomas et al. 1990) and in the ground squirrel, Spermophilus lateralis, during deep hypothermia (e.g., Garland and Milsom 1994). When in such a state, large fluctuations in lung and blood PO2 and PCO2 occur (e.g., Musacchia and Volkert 1971). Functionally, gas exchangers in terrestrial air breathers fall into three general categories. In the diffusive type which, e.g., occur in most small insects, pneumonate gastropods, and some terrestrial crabs, no ventilatory movements occur: diffusion is adequate to supply O<sub>2</sub> and remove CO<sub>2</sub>. In the mixed type, as found in the large insects and terrestrial crustaceans, mechanical pumping aids convective movement of air at least in the principal respiratory pathways. Mass transport of air to the pulmonary surface is necessary to produce efficient transfer of respiratory gases in the convective (ventilatory) type gas exchangers. These are found in the air-breathing fish, amphibians, reptiles, mammals, and birds.

# 6.5 The Diffusive Type Gas Exchangers

As a means of delivering  $O_2$  to a respiratory site, diffusion is only efficient over short distances. The process can hence only effectively service the extremely small, low-metabolism animals. Diffusion lungs occur in most pulmonate gastropods (Ghiretti 1966). A diffusive tracheal system has evolved in the terrestrial arthropods – in the uniramians (e.g., insects, centipedes, and millipedes), in chelicerates (e.g., scorpions and spiders), in isopods particularly the Porcellionidae (e.g., sow bugs) (Paul et al. 1987; Fincke and Paul 1989), and even in plants (Walsby 1972). The simplest lungs are found in snails and slugs where the mantle cavity has been converted into well-vascularized internal respiratory spaces (Fig. 59). Through muscular contractions when the pneumostome is closed, the intrapulmonary pressure increases to about 0.2 to 0.3 kPa. The periodic compression of the air inside the lung is thought to enhance the diffusion of O<sub>2</sub> into the blood (Ghiretti 1966). According to Dahr (1924, 1927), a very small PO<sub>2</sub> of 0.3 kPa is adequate to provide the animal with sufficient  $O_{2}$ . In Arion and Helix, at a PO<sub>2</sub> of 2kPa, the pneumostome opens 15 to 30 times in 30 min, remaining open for 7 min (Dahr 1924). The opening of the pneumostome in the gastropods is determined by factors such as levels of CO<sub>2</sub>, O<sub>2</sub>, humidity, and temperature (Mass 1939). The arachnids are among the earliest animals to occupy land and acquire the capacity to breathe air (Paul 1992). Their respiratory organs, the book lungs, have literally been frozen in time. In having changed very little since the Devonian, they are highly instructive in the understanding of the design of the gas exchangers in the original air breathers. The book lungs of the spiders and the scorpions (Figs. 15,72) consist of stacks of parallel blood-filled lamellae or plates contained in invagination(s) of the abdominal wall (Pohunkova 1969; Moore 1976; Vyas and Laliwala 1976; Herreid et al. 1981; Strazny and Perry 1987). The spiracular muscles, which respond to CO<sub>2</sub> (or low pH) and PO<sub>2</sub> (Fincke and Paul 1989) regulate the size of the opening into the atrium, expelling  $CO_{22}$  letting in  $O_{23}$ and regulating water loss and pH (Angersbach 1978; Fincke and Paul 1989). Gas



Fig. 72. Schematic view of the diffusive, sheet flow book lungs of a spider showing stacks of lamellae through which hemolymph flows,  $\rightarrow$ , and spikes which keep the lamellae apart

exchange is effected with the hemolymph, which is pumped across the hollow chitinous lamellae (Anderson and Prestwich 1982; Fincke and Paul 1989; Farley 1990; Fig. 72): in Eurypelma, when the spiracles are open, arterial PO, rapidly rises from 3.7 to 9.9kPa and hemocyanin is fully saturated (Angersbach 1978). The absence of ventilatory activity and control of the spiracle entrance area in the diffusion lungs minimizes water loss. It is compatible with low aerobic metabolic rate reflected in the low O<sub>2</sub> uptake especially in arachnids (Anderson 1970). Peristaltic movements in the book lung lamellae of some spiders have been reported (Moore 1976). The actual significance of these movements in respiration is, however, not well known (Paul et al. 1987). Anaerobic metabolism plays an important part in the activities of arachnids (Prestwich 1983). Their locomotory muscles lack mitochondria (Linzen and Gallowitz 1975). Only the heart, CNS, Malpighian tubules, and midgut glands work aerobically (Paul et al. 1987). The largest known spider is the bird-eating spider, Theraphosa blondi, of northern South America, which attains a body mass of about 55g (Dresco-Derouet 1974).



Fig. 73a,b. The well-developed tracheal system in the opisthosoma and prosoma (a) and first legs (b) of the web monitoring spider, *Uloborus glomosus*; ► book lungs. The trachea in the highly active legs are well developed. (Opell 1987)

Variably, spiders possess two kinds of respiratory organs, the book lungs and the trachea. The so-called pulmotracheates possess both organs (Fig. 73). The more metabolically active spiders and scorpions have book lungs and a tracheal system, while the ancestral ones (e.g., the orthognaths) have book lungs only (Paul et al. 1987; Strazny and Perry 1987). In some families of spiders, e.g., Caponiidae and Symphytognathidae, only the trachea exist (Paul 1992) but unlike in insects, the trachea do not directly contact the tissue cells. Hemocyanin is used to carry  $O_2$  over the remaining distance (Foelix 1982). Some pulmonate snails of the family Athoracophoridae (e.g., tropical snails, Janellidae) and some other unrelated animals, e.g., the sow bug (an isopod) Porcellio, the house centipede (chilopods), and Scutigera (Krogh 1941) present a fairly complicated respiratory system which combines the features of gastropod lungs with the tracheal system in insects: the pneumostome/spiracles open into a common vestibulum from which an array of fine diverticula radiate into a blood sinus providing a large respiratory surface area. A pneumotracheal organization provides a more efficient gas exchange capacity compared with simple smooth respiratory invaginations. In analogy, the design is closer to the mammalian tracheobronchial lung than to the tracheal system of insects. The largest diffusive lungs are probably those of the African pulmonate snails, Achatina and Bulimus, which attain a volume of up to 500 ml (Krogh 1941). In the freshwater pulmonates, Planorbis corneus and Lymnea stagnalis, despite lack of ventilatory activity, the PO<sub>2</sub> rises to 18 kPa before the closure of the pneumostome (a value higher than the alveolar level of the mammalian lung of about 13kPa. The PO<sub>2</sub> may drop to as low as 2.7 kPa before the pneumostome reopens (Precht 1939; Jones 1961). There is no respiratory system in the vertebrates which is adapted to diffusive respiration alone. In states such as hibernation (e.g., in the lungfish) and conditions such as exposure to high humidity (e.g., in the bullfrog), the skin may meet most, if not all, the O<sub>2</sub> requirements. Dogs have been experimentally kept alive for an hour on diffusive respiration alone. This, however, occurs only under conditions of minimal O<sub>2</sub> demands such as in states of anesthesia and while the animal is breathing pure O<sub>2</sub> (Lambertsen 1961).

# 6.6 The Mixed Type Gas Exchangers

### 6.6.1 The Insectan Tracheal System

Among the air-breathing animals, the insectan tracheal respiration is unique. In many ways, it is astonishing both for its intuitive architectural simplicity and functional efficiency. In a degree of refinement almost past belief, the insects have disengaged the circulatory and respiratory systems, totally relegating the former from any meaningful role in gas exchange. This is a radical deviation from the prevalent plan in the vertebrate and the invertebrate (nontracheate) air and water breathers where a circulatory system is intercalated between the gas exchanger and the body tissues (Figs. 1,2,3). In insects,  $O_2$  is delivered by the trachea directly to the body tissues (Figs. 75,76). The trachea, the portals of entry of air to the body



Fig. 74a-c. Schematic diagram of the air flow pathway in the tracheal system of insects. The vulvular spiracles (a) open to the outside while the tracheoles terminate deep in the animal's body (b) in some cases indenting some tissue cells (c)

(Figs. 37,74), form as ectodermal invaginations. With minimal drop in PO<sub>2</sub> along the way, virtually every individual cell in the body is served by a tracheole(s), structures which are analogous to the vertebrate blood capillaries (Fig. 77). The PO<sub>2</sub> between the tracheoles and the metabolizing tissue cells in insects is about 5.3 kPa (Weis-Fogh 1964a, 1967) compared with that of less than 0.3 kPa in the mammalian tissues. In adult Aphelocheirus, between the spiracles and the tracheoles, the PO<sub>2</sub> drops by only 0.3 kPa (Thorpe and Crisp 1941). Generally, the concentration of  $O_2$  drops by only 1% from the spiracle to the tissues (Buck 1962). The tracheal system can supply ten times more  $O_2$  g<sup>-1</sup> tissue than the blood capillary system (Steen 1971). With the spiracular valve serving as a carburetor, in mechanical terms, the trachea operate simultaneously both as a compressor and an exhaust pipe, presenting the epitome in the design of the gas exchangers. Contrary to the cases in the branching tubular structures like the bronchial system of the mammalian lung (e.g., Horsfield 1981), where between the principal and the terminal bronchi the flow velocity decreases by a factor of 700 due to increase in the cross-sectional area (Horsfield and Thurlbeck 1981; Horsfield and Woldenberg 1986), the insect trachea are the only system of air conduits known where the cumulative cross-sectional area and hence the flow velocity remain constant with length (Krogh 1941). In the endothemic insects (Heinrich 1992), thermoregulation of the thorax during flight enables them to attain some of the highest known muscle power outputs in the Animal Kingdom (Harrison et al. 1996). The aerobic rates of the flight muscles are some of the highest reported for any tissue (Kammer and Heinrich 1978), values which approach those of pure microbial cultures (e.g., Hughes and Wimpenny 1969). Insect flight muscles do



Fig. 75a,b. Air supply to the flight muscles of a grasshopper, *Chrotogonus senegalensis* showing: a secondary trachea, s, approaching the flight muscle, m, and tertiary trachea,  $\rightarrow$ , indenting the muscle. The trachea are supported by the spiral taenidia ( $\succ$  *inset*) which keep them open. The main figure is an enlargement of the enclosed region in the inset. b Closeup of a tertiary trachea with distinct taenidia,  $\rightarrow$ , about to indent the flight muscle, m. *Inset* \*, taenidia. a *Bar* 20µm; *inset* 80µm; b 45µm; *inset* 30µm. (Maina 1989b)



Fig. 76. Flight muscles, *m*, of the desert locust, *Locusta migratoria migratoria* showing secondary trachea, *p*, on the surface of the muscle and tertiary trachea, *e*, indenting the muscle. The terminal tracheal,  $\rightarrow$ , lie in very close proximity to the muscle fibers and the mitochondria,  $\succ$ . Bar 3 µm



Fig. 77. Secondary trachea, t, giving rise to tertiary trachea,  $\checkmark$ , indenting the abdominal muscles of the grasshopper, *Chrotogonus senegalensis*. *Inset* View of a tertiary trachea in the flight muscle, m. Bar 185 µm; inset 0.6 µm. (Maina 1989b)

not contain myoglobin (Elder 1975). In some small insects the flight muscles may attain a wing contraction frequency of up to 1000 Hz (Sotavalta 1947), a process amazingly powered entirely by aerobic metabolic pathways. Such high metabolic rates are possible because of the direct delivery of  $O_2$  to the tissues and the remarkably high volume density of the mitochondria (Weis-Fogh 1964a; Elder 1975; Ready 1983). In the process of attaining an optimal respiratory design, by exposing their body tissues to the ambient environment through a myriad of air conduits, the tracheates, and in particular insects, have had to pay a great price. They are particularly vulnerable to invasion by pathogens (e.g., Engelhard et al. 1994) and aerosol-based toxicants which pass the spiracles. It is common knowledge that some of the insecticides which are commercially advertised as to knock – down – insects – dead do exactly that! While vertebrate muscles can continue contracting by anaerobic metabolism for a period of time after  $O_2$  supply stops, since insect muscles lack lactic dehydrogenase, to function, they have to have a continuous supply of  $O_2$  (Pringle 1983).

Although best studied in insects, tracheal respiration has evolved in animal groups such as Onychophora (Peripatus), Solifugae, Phalangidae, some Acarina, Myriapoda, and Chilopoda. The bodies of the tracheates are pervaded by fine airfilled tubes, the trachea, which are simple noncollapsible hollow airways strengthened by endocuticular spiral or annular chitinous thickenings, the taenidia (Figs. 37,78). In some of the most metabolically active tissues, the finest branches of the trachea (the tracheoles) as they approach the tissue cells may be as narrow as 0.2 µm in diameter and may indent the cells in the manner of a finger poked into a balloon (Steen 1971; Fig. 74). In the case of the flight muscles, in some insects, the very narrow unventilated tracheoles, which are about 1 mm long, must supply  $O_2$  at a tremendous rate of 6.5 mol per m<sup>3</sup> of tissue per s (Weis-Fogh 1964a). In the highly metabolically active tissue such as the flight muscles, the terminal tracheoles are never more than 0.2 to 0.5 µm from a mitochondrion (Fig. 76) and in some tissues they may be as close as 0.005 µm (Wigglesworth and Lee 1982; Maina 1989b). The mitochondria cluster around the terminal tracheoles forming what has been termed mitochondrial continuum (Edwards et al. 1958). In the flight



Fig. 78. Trachea, *e*, in the abdominal muscles of the desert locust, *Locusta migratoria migratoria*, surrounded by tracheoblasts, *x*, cells that are thought to be involved in laying down trachea.  $\succ$ , taenidia. *Bar* 0.3 µm

muscle, the tracheoles may invest single muscle fibrils (Krogh 1941). The tracheoles terminate blindly (Richards and Korda 1950) though possible anastomoses have been reported (Buck 1948). Estimations made on the tracheal system of the giant lepidopteran, *Cossus cossus*, larva (mass 3.4g, length 60 mm) gave a total cross-sectional surface area of all trachea supplying the tissues of  $6.7 \text{ mm}^2$  with an average length of 6 mm: O<sub>2</sub> diffuses at a rate of  $0.3 \text{ mm}^3 \text{ s}^{-1}$  at a pressure head of 1.5 kPa, a value which is more than adequate even during muscular exertion (Krogh 1920b). Depending on factors such as age and stage of development, the tracheal system constitutes 5 to 50% of the volume of an insect. In the silkworm, *Bombyx mori*, the tracheoles are  $1.5 \text{ m} \log (\text{Buck 1962})$  and the volume which for a 5.7-g worm is  $49 \,\mu l \,g^{-1}$  makes about 5% of the body volume (Bridges et al. 1980). In the adult cockchafer, *Melolotha*, the trachea constitute a volume of  $585 \,\mu l g^{-1}$  (Demoll 1927). The tracheal volume of a 5-g *Cecropia* pupae is about 250 mm<sup>3</sup> (Kanwisher 1966).

The terminal tracheoles are filled with fluid, the degree of filling depending on the level of activity (e.g., Wigglesworth and Lee 1982). The endotracheal fluid is removed osmotically by the increased concentration of the end products of metabolism in the interstitial cell spaces during times of increased metabolic demands and exposure to hypoxia. The air/fluid interface is brought closer to the tissue cells as the interstitial fluid is drawn upstream of the peripheral tracheoles and into the cytoplasm of the surrounding cells (Wigglesworth 1953, 1965). The process shortens the diffusional pathway of the respiratory gases. When the acidic metabolites are eliminated and better aerobic conditions preponderate, the action is reversed. Such a sequence of events is not unique to insects as it is utilized by many other animals to open up capillary beds in specific body tissues during conditions of high O<sub>2</sub> demand. Evidently, the diffusion-based insectan tracheal system evolved entirely for aerial respiration. Because the diffusion of O<sub>2</sub> in water is  $10^3$  lower than in air, the tracheoles would need to be as small as  $12 \mu m$  to adequately supply O<sub>2</sub> to tissues respiring at the same rate as the insect flight muscle (Denney 1993). Theoretically, aquatic insects relying on delivery of O<sub>2</sub> entirely by diffusion would need to greatly increase tracheolar density, lower their metabolic rate, and drastically reduce the distance between the body tissue cells and the surrounding water. This would call for extremely thin, minuscule, intensely tracheated bodies. Such requirements may not be compatible with the essential insectan morphology. Those insects which have reverted to living in water have retained air-filled trachea (the gas gills) which essentially act as internal plastrons (Sects. 6.6.1 and 6.12): the  $O_2$  taken up from the surrounding water is transferred along a gas phase within the tracheal system in much the same way as in a surface-dwelling insect.

The insectan tracheal system provides a unique and perhaps the most costeffective design for supplying body tissue cells with  $O_2$ . However, the limitation of diffusion and the large mechanical ventilatory forces which have to be generated to move air at extremely high rates in the countless fine conduits have consigned the insects to small body sizes. This may explain why such an efficient scheme was never adopted in the larger animals. The heaviest living insects are the Goliath beetles (family: Scarbaeidae) of the Equatorial Africa: *Goliathus goliathus* weighs between 70 and 100g. The hairly-winged beetles of the family Ptiliidae (=

Trichopterygidae) and the battledore-wing fairly flies (parasitic wasps) of the family Myrmaridae which measure only 0.2 mm in length are the smallest insects: the smallest insects are smaller than some of the largest protozoa! The average tracheolar length for optimal diffusion appears to be 5 to 10mm and the minimum diameter 0.2 um (Krogh 1920a,b; Weis-Fogh 1964a). The tracheates which utilize diffusion as the main mode of gas exchange include the Onychophora (Peripatus), the tracheate Arachnoidea, Myriapoda, and Chilopoda, almost all terrestrial insect larvae, all pupae, and most of the small imagines. This is made possible by the relatively fast diffusion of  $O_2$  in air compared with that in water. The largest ever known insect was the dragonfly-like Meganeura of the Carboniferous, which reached a length of 60 cm and was 3 cm in width (Krogh 1941). It is well known that the levels of atmospheric O, have greatly fluctuated over geological times (Sect. 1.11). A greater PO<sub>2</sub> in air as occurred in the Carboniferous period (Graham et al. 1995; Fig. 9) may have allowed the development of the giant insects. Presently, the largest extant insects are the tropical beetles which reach a length of 15 cm. Flying insects range in body mass from 1 ug to 20 g (Norberg 1990). The stick insects best demonstrate the compromises between size and shape which have occurred in insects, features which have been modulated for optimal tracheolar ventilation and diffusion. Some of the longest insects in the world are the tropical stick insect, Phamacia serratipes, and the Central and South American dragonfly, Megaloprepes caeruleata, which have very narrow bodies which are respectively 33 and 12 cm long. While a housefly, which weighs about 15 to 20 mg, does not need to ventilate the tracheal system, a bee, which is more energetic and weighs about 100 mg, does so regularly. In insects such as locusts, dragonflies, and cockroaches, at rest, well-synchronized abdominal and, to a smaller extent, thoracic ventilation occurs (Brocher 1931). Although during rest no ventilatory movements take place in the cockroaches, Peripaneta and Blatella, during flight when O<sub>2</sub> consumption increases 10 to 100 times, wing beats compress the thorax ventilating the trachea and the air sacs (Brocher 1920; Portier 1933). Ventilation in flight may be aided by direct inflow of ambient air at the ventral surface and the slightly reduced pressure over the abdominal spiracles due to a Bernoulli-Venturi effect. During steady flight in the desert locust, about  $3201 \text{kg}^{-1} \text{h}^{-1}$  of air with an average tidal volume of 167 cm<sup>3</sup> and frequencies of 30 to 60 times min<sup>-1</sup> is ventilated into the tracheal system by abdominal and thoracic pumping. The intratracheal pressure increases to 0.9 to 3.3kPa at the peak of an abdominal contraction (Miller 1960; Weis-Fogh 1967). The giant beetle, *Petrognatha gigas*, has a ventilatory rate of about  $2000 \text{ kg}^{-1} \text{ h}^{-1}$  (Miller 1966).

Through synchronized action of the spiracles, the trachea are ventilated continuously and unidirectionally, particularly among the Orthoptera (e.g., Fraenkel 1932; Weis-Fogh 1964a, 1967). In the honeybee, the flow is unidirectional during flight (Bailey 1954) and in *Sphodromantis*, 95% of the inhaled air passes unidirectionally while only 5% passes tidally (Miller 1974). Among the cockroaches, *Periplaneta* and *Blatella*, tidal ventilation only occurs during stress while in other roaches, *Byrsotria*, *Blaberus*, and *Nyctobra*, anteroposterior ventilation occurs during rest (Buck 1962). The unidirectional and continuous ventilation, as occurs in the parabronchial bird lung (e.g., Scheid 1979), minimizes or abolishes dead space air, ascertaining that the gas exchange site is supplied with air at the highest possible PO<sub>2</sub>. Abdominal pumping is inadequate in supplying O<sub>2</sub> to the long muscles of the legs in some large insects. In the grasshopper, the concentration of O<sub>2</sub> in the tibial tracheae is fairly high (16%) in the resting state but drops to 5% during physical exertion (Krogh 1913). Special spiracles have developed on the legs of the harvestmen (Opiliones) apparently to overcome the diffusive and convective limitations (Hansen 1893). The smallest tracheoles in insects which measure about  $0.2\,\mu\text{m}$  are close to the mean free path (MFP) of O<sub>2</sub> molecules in air (the average distance a molecule travels in air before colliding with another) which is about  $0.008\,\mu\text{m}$  (Pickard 1974). Below the MFP, the effective diffusion coefficient is reduced, lowering the rate at which O<sub>2</sub> is delivered to the tissues. The minimum tracheolar diameter in insects appears to have been set by the MFP of molecules in air. In the most energetic species, this parameter seems to have been optimized in the most highly metabolically active tissues such as the flight muscles.

The development of the tracheal system appears to be partly determined by certain intrinsic factors in the target tissues (Locke 1958a,b,c) while the actual distribution to specific organs is determined by the local aerobic conditions (Edwards et al. 1958). In much the same way as occurs between the capillarization of the tissues and metabolic activity in vertebrates, in insects, tracheolar density is dependent on the metabolic activities and PO<sub>2</sub> levels in particular organs and parts of the body (Edwards et al. 1958; Wigglesworth 1965; Steen 1971). The trachea are particularly well developed in the legs, which are actively used for web-monitoring activity in the spiders of the family Uloboridae (Opell 1987). In larval meal-worms, Tenebrio molitor, hypoxia influences tracheal growth. Development at an ambient  $PO_2$  of about 10 kPa leads to wider trachea (London 1989). In the wing muscle of the locust, between  $10^{-1}$  and  $10^{-3}$  (volume of the trachea per volume of muscle) is taken up by the tracheal system (Weis-Fogh 1967). The entire respiratory system in insects may form as much as 50% of the entire body volume (Steen 1971). In the small and relatively inactive insects and arachnids, the tracheal system may be simple but in larger and more energetic species (e.g., wasps and bees), it may be complex with the system comprising an intricate maze of longitudinal and transverse branches (Fig. 37) connected to air sacs (Fig. 38). The air sacs increase the tidal volume by as much as 70% of the total air capacity affordable by the trachea alone (Bursell 1970) and reduce the longitudinal diffusion gradient for  $O_2$  along the gas exchange pathway. They are well developed in Diptera and Hymenoptera but are absent in the subclass Apterygota. In cicada, Fidicina monnifera, the air sacs together with the tracheal system constitute 45% of the body volume (Bartholomew and Barnhart 1984).

# 6.7 The Convective Type Gas Exchangers

#### 6.7.1 Ventilatory Mechanisms and Organization of the Gas Exchangers

The ventilatory lungs have evolved only in the vertebrates. They were a major factor which provided the means for realization of large and complex body sizes

and forms as well as high metabolic lifestyles. Different ventilatory processes have evolved in the air-breathing vertebrates. The bucco-pharvngeal pump, where air is literally swallowed, occurs in the amphibians and the dipnoans (Brainerd et al. 1993). The mechanism appears to have arisen as a compromise between respiration and feeding. The energy which operates the force pump arises from contraction of the muscles of the mouth (De Jong and Gans 1969; West and Jones 1975; Liem 1987a). The amount of air which can be transferred during a single breath is a function of the pressure differential between the lung and the buccal cavity and the potential change in the volume of the buccal cavity (Liem 1987a). In the aquatic habitat, during feeding, pressure changes far exceed those recorded during air ventilation (Lauder 1980; Bemis and Lauder 1986). The buccal force pump operates below its maximum potential during air breathing and is hence an overdesign for the role it plays in the medium. The tidal volume acquired by the buccal force pump is limited. The buccal force pump is ineffective for filling long narrow lungs like those of snakes (Gans 1971; Guimond and Hutchison 1976) and is a rather inefficient method which cannot support high levels of metabolism. The ventilatory inefficiency of the buccal force pump in the terrestrial settings is thought to have constituted a major obstacle in the evolutionary progression of the amphibians (Gans 1970; Liem 1987a). Suctional (aspirational) breathing occurs in most reptiles and all mammals and birds (Bainerd 1994), animals which operate at a higher level of metabolism and have long complex lungs which cannot be effectively serviced by a buccal force pump. The evolution of suctional breathing dissociated the feeding apparatus from the breathing one. It comprised a milestone in the development of more complex, efficiently ventilated gas exchangers which were necessary on transition to land and the resultant higher metabolic requirements. While suctional breathing occurs in various reptiles, positive-pressure inflation through the buccal force pump preponderates in the taxon (Gans and Hughes 1967; Gans 1971): breathing is exclusively effected by movement of the rib cage and complex changes in the volume of the visceral cavity, as a diaphragm is lacking (e.g., Perry and Duncker 1980; Gaunt and Gans 1969; Gans and Clark 1976, 1978). Modem amphibians have vestigial ribs. It is, however conjectured that the larger now extinct forms had functional ribs which they utilized to draw air into their lungs. In amphibians and reptiles, postural changes and presumably contraction of the pulmonary smooth muscles (Figs. 79,80; Stark-Vancs et al. 1984; Maina 1989b) and elastic tissue (Fig. 80) as well as hydrostatic forces (when the animal is immersed in water) may aid in expiration or air. The smooth muscles which line the central cavity of the lung of the tegu lizard, Tupinambis nigropunctus (Hlastala et al. 1985), as well as snake lungs (Maina 1989e; Fig. 80) have been associated with changing the shape of the lung and assisting in the convective mixing of the intrapulmonary air. In birds, the ventilation of the lung is unique among vertebrates. The virtually rigid lungs are ventilated by a synchronized activity of the air sacs which are interspersed in the abdominal cavity, pneumatizing some of the adjacent bones. The pressure changes in the air sacs are generated by contraction of muscles which attach to the thoracic wall adjusting the thoracoabdominal space. A partition between the thorax and the abdomen is lacking: the liver and not the lung (as is the case in mammals) surrounds the heart. The oblique and the horizontal septa (see King



Fig. 79. Smooth muscles, m, and collagen, c, in the lung of the tree frog, *Chiromantis petersi*. p pericyte; n capillary endothelial cell; e erythrocyte. Contraction of the smooth muscle may assist in the expiration. *Bar* 1  $\mu$ m. (Maina 1989d)

and McLelland 1975) are thought to be homologous to the mammalian diaphragm but fall far too short of anatomically dividing the coelomic cavity. The mammalian lung can be divided in two zones: the nonparenchyma (the conducting zone) of branching air conduits (the bronchi) and the parenchyma (the respiratory zone), made up of millions of alveoli. Together with the intercostal



**Fig. 80**. Smooth muscles, *m*, in the lung of the black mamba, *Dendroapis polylepis*. *x* Elastic tissue; *c* collagen; *p* pericyte; *e* erythrocyte in a blood capillary. The smooth muscles may assist in intrapulmonary movement of air. *Bar* 1  $\mu$ m. (Maina 1989e)

muscles, the diaphragm is an important respiratory muscle. By changing the volume of the thoracic cavity, air is moved into and out of the lung. The complete separation of the coelomic cavity into the thoracic and the abdominal cavities by the diaphragm was a key evolutionary development for efficient respiration in

mammals. The evolved ventilatory mechanisms show varied levels of refinement which correspond with the phylogenetic levels of progression. The lungs in amphibians are filled with air under pressure from the bucco-pharyngeal cavity. In some reptiles (e.g., *Chamaeleo*), this process has been conserved and is used only in emergency cases to inflate the lungs. The various forms of suctional breathing which have developed in both terrestrial and aquatic animals allow greater flexibility in the tidal volume and have facilitated the development of the respiratory organs independent of the ventilatory mechanisms. Gas exchange has been totally disengaged from feeding. In general, those vertebrates which are adapted to respiration in xeric environments have the more specialized lungs (Fig. 81) and are capable of the highest sustained rate of gas exchange. This supports a high level of metabolic activity, enabling a greater degree of organizational and ecological progress. Such animals are found in only a few taxa which include insects,



**Fig. 81.** Scheme of the structural complexities of the vertebrate lungs showing that a parameter such as an extensive respiratory surface area can be attained both in homogenous and heterogenous lungs through different modifications. For example, birds have isolated the ventilatory and the gas exchange parts of the lung and through intense subdivision of the lung achieved an extensive respiratory surface area in a relatively small lung. Pulmonary design corresponds with the metabolic needs and phylogenetic statuses of individual animal groups. Animals shown *on a black background* are now extinct. (Perry 1989)

reptiles, birds, and mammals. In these animals the integument is almost totally impermeable to water, O2, and CO2. Among vertebrates, the simplest forms of lungs are the smooth-walled saccular types, e.g., in Proteus (Hughes 1970), which are poorly vascularized. Though such lungs develop at the early larval stages, they play very little, if any, role in respiration until the midlarval stage (Goniakowska-Witalinska 1995). In the aquatic or amphibious groups, the lungs play diverse roles such as hydrostatic (Milsom and Johansen 1975; Pohunkova and Hughes 1985a), sexual display, and sensory perception (Duykers and Percy 1978; Schmidt 1982; Ehret et al. 1990). Internal subdivision of the lung imparts a greater respiratory surface area (Maina et al. 1989b; Fig. 82), increases the volume of blood in the lung, provides better exposure of blood to air (Hughes 1978), and ensures mechanical integrity of the organ. In most lungs, the exposure of the pulmonary capillary blood to air is promoted by a construction which entails not only increase in vascularity but also distension of the blood vessels over the epithelial surface, e.g., in the pneumonate gastropods (Maina 1989c; Figs. 59,60) and the mammalian lung (Fig. 87). In the double capillary system of the amphibians (Maina 1989d; Fig. 84) and most reptilian lungs (Maina 1989e; Figs. 83,85), a parallel row of blood capillaries which are exposed to air on only one side occur. A single capillary plan, where a sheet of blood capillaries which are supported by



**Fig. 82a–d.** Subdivisions of the parenchyma of the vertebrate lungs. The simple unicameral lungs (a) have large air spaces, the ediculae; the faveolar lungs have layered air spaces (b), the faveoli; the highly specialized mammalian (c) and avian lungs (d), respectively, have alveoli and air capillaries as the terminal gas exchange components. Evidently, a basic design appears to have given rise to all forms of lungs, the particular design and complexity depending on the level of development and the metabolic needs of different animals. *T* Trabeculae; *A* artery; *V* vein; *E* ediculae; *L* lumen; *P* pneumocytes; *F* faveolus; *A* alveolus; *Br* bronchus; *At* atrium; *M* exchange tissue; *Ac* air capillary; *I* infundibulum. (Perry 1989)



Fig. 83. Double capillary arrangement in the snake lung, *Dendroapis polylepis*. This design is limiting in that only one side of the blood capillary can be used for gas exchange. An attempt is made to increase the available surface area by the blood capillaries protruding over the surface of the lung. *c* Blood capillary; *e* erythrocytes; *s* interfaveolar septum; *m* white blood cell. *Bar*  $5\mu$ m. (Maina 1989e)

a thin septum, exists in the highly refined mammalian lungs (e.g., Alcorn et al. 1980; Burri 1984a; Figs. 86,87). In such cases, the capillary blood is essentially suspended in a three-dimensional space. The arrangement provides better exposure of blood to air compared with the double capillary one where the capillary loading, the ratio between the blood volume and the surface area available for gas exchange, is very high (Perry 1983). A notable exception to the general plan in the mammalian lung is that found in the rather placid herbivorous marine mammals, the sirenians, i.e., manatees and dugongs. Abundant connective tissue in the interalveolar septa, large alveoli (Tenney and Remmers 1963), and a welldeveloped double capillary system (e.g., Belanger 1940; Wislocki and Belanger 1940) occur. The pattern is similar to that found during the embryonic stage of development of the mammalian lung (e.g., Pinkerton et al. 1982). The preponderance of collagen and other supporting tissue elements in the lung (Laurent 1986) may provide the biomechanical support necessary to overcome the hydrostatic forces during deep dives. The exposure of the pulmonary capillary blood to air in the lungs of birds occurs in form of a diffuse arrangement between the air and blood capillaries (Maina 1982a, 1988a; Figs. 88,89) which intimately interdigitate with each other maximizing the respiratory surface area (Dubach 1981; Maina et



**Fig. 84a-d.** Structrue of the lung of the tree frog, *Chiromantis petersi.* **a** Blood capillaries, *c*, which contain erythrocytes, *e*, exposed to air across a thin blood-gas barrier,  $\succ$ . **b** Blood capillaries, *c*, bulging into an air space; \*, granular pneumocyte. **c** Granular pneumocyte containing lamellated osmiophilic bodies,  $\succ$ . **d** *p*, nucleus of a Type I pneumocyte overlying a blood capillary. **a** *Bar* 8µm; **b** 4µm; **c** 3µm; **d** 0.2µm. (Maina 1989d)



**Fig. 85a–d.** Structure of the lung of the black mamba, *Dendroapis polylepis*, showing blood capillaries bulging into the air spaces. Reptilian lungs generally manifest a better exposure of blood to air than the amphibian ones. **a** *c* blood capillaries. **b** \* granular pneumocyte (type II cell) located between blood capillaries; *x* interstitial macrophange. **c** *v* a blood capillary bulging into an air space;  $\succ$  smooth muscle. **d** \* nonmyelinated axons in the lung;  $\succ$  squamous type I cell; *e* erythrocytes. **a** Bar 10µm; **b** 5µm; **c** 5µm; **d** 1µm. (Maina 1989e)



Fig. 86. Lung of a bat, *Epomophorus wahlbergi*, showing alveoli, *a*, and blood capillaries, *c*, containing erythrocytes, *e*. The interalveolar septum contains smooth muscle,  $\succ$ , and elastic tissue elements,  $\backsim$ . The pulmonary capillary blood in the mammalian lung is better exposed to air than in amphibian (cf. Fig. 84) and reptilian (Fig. 85) lungs but thick and thin parts of the blood-gas barrier are evident.  $\bigstar$ , detached surfactant lining. *Bar* 1.8µm. (Maina et al. 1982a)



Fig. 87. An alveolus of the lung of the lesser bushbaby, *Galago senegalenis*, showing the blood capillaries, *c*, bulging into the air space. > junctions of type I cells; *s* interalveolar septum. *Bar* 120  $\mu$ m. (Maina 1990c)



Fig. 88. Exchange tissue of the lung of the house sparrow, *Passer domesticus*, showing the air capillaries, *a*, and blood capillaries, *c*, which interdigitate very profusely. The blood capillaries are exposed to air on all sides giving rise to an extensive respiratory surface area per unit volume of the exchange tissue.  $\succ$  blood-gas barrier; *e* erythrocytes. Bar  $3\mu m$ 

al. 1989a) and providing an extremely thin blood-gas barrier (Figs. 40a,90). These features generate a remarkably high pulmonary diffusing capacity for  $O_2$  (Maina 1989a, 1993; Maina et al. 1989a). In the lungfish, *Lepidosiren paradoxa*, the pulmonary capillary blood volume constitutes 3.5% of the total lung volume, in the rat lung the value is 14%, and in the bird lung 25% (Maina et al. 1989a).

# 6.7.2 The Amphibian Lung

The actual evolutionary origin of the amphibians is not well known (e.g., Szarski 1962; Schaeffer 1965b; Løvtrup 1977; Milner 1988). The osteolepiform fish (which are thought to have evolved a primal lung) or the lungfishes (Dipnoi) are conjectured to have given rise to the tetrapods (Romer 1946, 1967, 1972; Pough et al. 1989; Meyer and Dolven 1992). The transition to land by the vertebrates was inaugurated by the amphibians in the Devonian and concluded by the reptiles in



**Fig. 89. a** Cast preparations of the air capillaries, *A*, and **b** blood capillaries, *C*, of the lung of the domestic fowl, *Gallus domesticus*. **a** \* spaces occupied by blood vessels. **b** \* spaces occupied by the air capillaries. The two gas exchange units intertwine intimately increasing the surface area available for gas exchange. **a** *Bar* 10 $\mu$ m; **b** 3 $\mu$ m. (Maina 1982)

the Paleozoic era. For a good part of their evolution (about 70 million years), the early amphibians were strictly aquatic bimodal breathers (Thompson 1991). The contemporary amphibians occupy a focal point in the study of some fundamental biological processes such as the realization of air breathing and terrestrial colonization (Szarslo 1977; Smits and Flanagin 1994). Dual subsistence in water and land has obligated development of unique physiological and morphological respiratory adaptations in the taxon (Lenfant and Johansen 1967; Duellman and Trueb 1986; Jackson 1987; Burggren 1989). The unique nature of development (metamorphosis from aquatic to air breathing), the diversity of the habitats the taxon occupies, and the multifunctional nature of the lungs (respiration, vocalization, buoyancy control, and defense) explain both the heterogeneity of the lung morphology and the multiple pathways utilized for gas exchange (Guimond and Hutchison 1972, 1973a,b; Rahn and Howell 1976; Burggren and Wood 1981). Amphibians largely live in water and humidic habitats but a few species have adapted to the highly desiccating deserts (e.g., McClanahan et al. 1994). Terrestrial anurans such as *Chiromantis xerampelina*, which have developed an impermeable skin (Stinner and Shoemaker 1987) and ureotelism (Shoemaker et al. 1972), have acquired remarkable tolerance to desiccation. The two species can withstand a water loss in excess of 60% of their body mass (Loveridge 1970). The highly xeric African tree frog, *Chiromantis petersi*, leads a characteristically unamphibian lifestyle: it prefers direct solar insolation and temperatures of 40 to  $42 \,^{\circ}$ C (Loveridge 1970, 1976). The need to balance water conservation with gas exchange may explain why there are no large existing amphibians.

During the larval stages, amphibians have transient external and internal gills. Subsequently, the lungs and the bucco-pharngeal cavity become highly vascularized and assume an important respiratory role in air. The skin is a dual respiratory organ, exchanging gases in both water and air. The neotenic amphibians retain the gills throughout life. In the egg, larval caecilians use gills for respiration. The gill are lost soon after hatching (Welsch 1981). Using gills, the larvae of the viviparous caecilian, Typhlonectes, exchange gases and nutrients with the lining of the oviduct (Wake 1977): the gills are lost before birth. The neotenic larva of Amblyostoma (Axolotl) has perennial external gills. The flow of blood to the various respiratory organs is controlled mainly by ventilatory rate and the levels of ambient O<sub>2</sub> (Wood and Glass 1991). Hypoxia affects the flow of blood to the gills, skin, and the lungs (Shelton et al. 1986; Malvin and Heisler 1988). A drop in the PO<sub>2</sub> in the lung increases the perfusion of the skin (Boutilier et al. 1986) and vice versa. In aerated water, the skin receives 20% of the pulmocutaneous blood flow, the amount decreasing to an insignificant amount when the water is hypoxic (Wood and Glass 1991). In the lungless salamanders (Plethodontidae), gas exchange which occurs entirely across the skin and the buccal cavity (Gatz et al. 1974; Gratz et al. 1974) is diffusion-limited (Piiper et al. 1976). Such animals have acquired long, cylindrical bodies to secure a greater surface-to-volume ratio, a feature which promotes gas exchange across the skin. The lungless salamanders normally live in cold, well-aerated waters. In the aquatic amphibians, locomotion generates passive ventilation of the skin, a process which can increase O2 uptake markedly (Feder 1985; Full 1985).

Living amphibians exist in three orders, namely the Gymnophiona (= Apoda = caecilians), Salentia (= Anura), and Caudata (= Urodela). Biologically, the caecilians are the least known group (Stiffler et al. 1990; Smit and Flanagin 1994). They are an elusive, vermiform tropical aquatic, semiterrestrial, or subterranean group (Nieden 1913; Taylor 1968; Renous and Gasc 1989). Evolutionarily, the caecilians are a monophyletic distant group (Milner 1988): they have been isolated from the other living amphibian orders for at least 70 million years. As characteristic of other similar animals with thin cylindrical bodies, e.g., the snakes (Ophidia) (e.g., George and Shah 1956), the caecilians possess long, tubular lungs (Fig. 99; Wake 1974; Maina and Maloiy 1988; Renous and Gasc 1989). In some species, the left lung is remarkably reduced or totally missing (e.g., Wiedersheim 1879; Pattle et al. 1977; Maina and Maloiy 1988; Burggren 1989; Fig. 98). In the aquatic *Typhlonectes compressicauda* (Sawaya 1947), however, as many as three lungs occurs (Toews and MacIntyre 1978): the anterior (tracheal) lung is located between the buccal cavity and the heart, while the other two run from the buccal cavity to the cloaca (Fuhrmann 1914; Toews and MacIntyre 1978). The lungs of the caecilians are internally subdivided to form air spaces by a single row of septa which are mechanically reinforced by two diametrically opposite trabeculae (Wiedersheim 1879; Wake 1974; Welsch 1981; Maina and Maloiy 1988; Fig. 99). Marcus (1928, 1937) observed that the structure of the caecilian lung is particularly primitive. The organization of the lung compares with that of the almost limbless, large aquatic salamanders like Amphiuma and Siren (Bell and Stark-Vancs 1983; Martin and Hutchison 1979). A relatively elaborate lung as in Boulengerula (Maina and Maloiy 1988) and Geotrypetes (Bennett and Wake 1974) may enable the caecilians to procure adequate amounts of O<sub>2</sub> from the hypoxic fossorial environment in which they live. Adaptively, the hematocrit and the hemoglobin concentration respectively, of the aquatic Typhlonectes compressicauda (38% and 11.3 gHb per 100 cm<sup>3</sup> blood; Toews and MacIntyre 1977, 1978) and the terrestrial Boulengerula taitanus (Wood et al. 1975; 40% and 10.3 gHb per 100 cm<sup>3</sup> blood) are some of the highest values reported in amphibians. The large blood volume (24 to 26% of the body mass; Toews and MacIntyre 1978) and low P<sub>50</sub> of the blood hemoglobin, e.g., in *Typhlonectes* 3 kPa (Toews and MacIntyre 1978) and 3.7 kPa in Boulengerula (Wood et al. 1975), provide an efficient mechanism for uptake and storage of O<sub>2</sub> in the blood. While the caecilians have a lower resting rate of metabolism than the anurans and the urodeles, the aerobic capacity during exercise exceeds that of the two other groups (Smits and Flanagin 1994). The elongated nature of the lung of the caecilians may introduce ventilatory limitations during locomotion as a result of compression of the lung by the trunk muscles. A temporal dissociation between breathing and locomotion has been reported in the running lizard (Carrier 1991), an animal with a similarly long lung and general body form. In many ways, the development of the gas exchangers in the amphibians identifies with that of the lungfishes (Dipnoi) (Wassnetzov 1932; see Fig. 64a,b). Physiological adaptations such as high hemoglobin concentration, small numerous erythrocytes, and large blood volume have been reported in the caecilians which live in hypoxic environments (Wood et al. 1975; Toews and MacIntyre 1978). At 25 °C, O<sub>2</sub> uptake in the fossorial caecilian, Boulengerula taitanus, which lives in hypoxic-hypercarbic habitats, is equal to that of other amphibians (Hutchison 1968; Wood et al. 1975). The Caudata, which are mostly aquatic, e.g., the newts, have poorly vascularized lungs with a smooth internal surface (Hightower et al. 1975; Meban 1977; Goniakowska-Witalinska 1980a,b). Such animals mainly use the skin for gas exchange (Noble 1925,1929). The characteristically low metabolic rate newt, Triturus alpestris, has smooth-surfaced lungs (e.g., Claussen and Hue 1987) with 569 capillary meshes cm<sup>-2</sup> while the metabolically more active tree frog, *Hyla arborea* (Goniakowska-Witalinska 1986), has more elaborate lungs with 652 capillary meshes per cm<sup>2</sup> (Czopek 1965). Plethodontidae constitute the largest family among the Caudata (Feder 1976; Ruben et al. 1993; Wake and Marks 1993). They acquire all their O<sub>2</sub> needs from the cold, well-oxygenated water in which they live across their highly vascularized skin. The length of the skin capillaries constitutes 90% of all blood vessels associated with the respiratory surfaces, with the other 10% being in the buccal cavity (Czopek 1965). The epithelial lining of the buccal cavity is very thin (Noble 1931; Czopek 1965). In caudates such as Salamandra, Amphiuma,

Megalobatrachus, and Siren, species which predominantly utilize the lung for gas exchange, the internal surface of the lung is well subdivided (e.g., Goniakowska-Witalinska 1978; Meban 1979; Hashimoto et al. 1983; Matsumura and Setoguti 1984). The skin is poorly vascularized, with the epidermis being very thick (47 to 110µm) (Czopek 1965). The lungs of most amphibian species such as Amphiuma and the toad, Bufo marinus, have a preponderance of smooth muscle tissue (Czopek 1962b; Smith and Rapson 1977; Martin and Hutchison 1979; Goldie et al. 1983; Stark-Vancs et al. 1984; Maina and Maloiy 1988; Maina 1989d), which may account for the great compliance of the lungs (Hughes and Vergara 1978). In Amphiuma, during expiration, the lung virtually collapses, producing an almost 100% turnover of air (Stark-Vancs et al. 1984). The amphibian lungs are best developed in the Salentia where septa intensely subdivide the lung, converting the large central air space into small stratified air cells (e.g., Okada et al. 1962; Smith and Rapson 1977; Goniakowska-Witalinska 1986). The internal morphology of these relatively elaborate lungs is similar to that of the lungs of the lungfishes (Dipnoi) (e.g., de Groodt et al. 1960; Klika and Lelek 1967; Gannon et al. 1983; Maina 1987a; Fig. 64). The lungs of Pipa pipa (Marcus 1937) and Xenopus laevis (Goniakowska-Witalinska 1995) are reinforced with septal cartilages to ensure patency of the air passages. In Salentia, the skin contributes very little towards gas exchange. The length of the skin capillaries constitutes only 30% of the total length of the blood capillaries located in the respiratory surfaces (Czopek 1965). However, in two species of Salentia which live in well-oxygenated high mountain lakes, e.g., Telmatobius and Batrachophrynus, the lungs are very small, the body very well vascularized, and the epidermis very thin (Muratori et al. 1976; Czopek and Szarski 1989). Well-differentiated pneumocytes (Fig. 103) as well as dust cells (free phagoctes; Fig. 102) have occasionally been observed on the surface of the amphibian lung (Welsch 1983; Maina 1989d).

The morphological heterogeneity of the amphibian gas exchangers and the lungs in particular correspond with the remarkable diversity of the environments they occupy, the mode of life they lead, and their property of interrupted development. Though amphibians have multiple options for gas exchange, only one pathway is best developed for optimal performance in a particular environment (Guimond and Hutchison 1976). Pulmonary vascularization correlates with the degree of terrestriality, behavior, and tolerance to dehydration. The skin is the main pathway for gas transfer in the predominantly aquatic species, while in the more terrestrial ones it has been relegated or rendered totally redundant as the lung has assumed a central position in gas exchange. Necturus experimentally held in cool, well-aerated water has better-developed gills than those of animals kept in warmer, poorly aerated water (Guimond and Hutchison 1976). Despite its strong reliance on water, the lung of the anuran clawed toad, Xenopus laevis, is very well developed for aerial respiration (Smith and Rapson 1977; Pohunkova and Hughes 1985a): 80% of its O<sub>2</sub> needs are transferred across the skin (Emilio and Shelton 1974). The perfusion of the skin is regulated by dilation and constriction of the cutaneous vasculature (Poczopko 1959). In Bufo marinus and Rana catesbeiana (Hillman 1987a,b), dehydration leads to increased vascular resistance due to a hemoconcentration effect on the blood viscosity. Dehydration in the terrestrial frog, *Eleutherodactylus coqui* (Pough et al. 1983), and the aquatic Xenopus laevis (Hillman 1987a,b) lowers the capacity to utilize aerobic metabolism during activity. Systemic  $O_2$  transport may be the limiting factor to aerobic capacity in the air-breathing amphibians (Hillman et al. 1985). In the xerophilous anuran, *Phyllomedusa sauvagei* and *Chiromantis xerampelina*, at the resting evaporative water loss (EWL), cutaneous respiration is insignificant, but with increased water loss, fractional cutaneous gas exchange correlates with the cutaneous EWL (Stinner and Shoemaker 1987). Adaptively, the blood of the highly xeric *Chiromantis petersi* has a low  $O_2$  affinity and a temperature-insensitive  $O_2$ hemoglobin binding capacity (Johansen et al. 1980).

Compared with the other air-breathing vertebrates, the amphibians have some of the simplest lungs, generally with low diffusing capacities for O<sub>2</sub> (Glass et al. 1981a). The lungs of Necturus and Cryptobranchus are thin-walled, transparent, poorly vascularized, and nonsepted (Guimond and Hutchison 1976): hydrostatic control may have the more significant role in such simple lungs. Regarding morphological and morphometric characteristics, generally, the lungs of the anurans and the apodans are more advanced than those of the urodeles (Meban 1980): on average, the arithmetic mean thickness of the blood-gas barrier in the urodeles is 2.59 µm, 2.35 µm in the Apoda, and 1.89 µm in the Anura. Some areas of the blood-gas barrier of the lung of the caecilians Chthonerpoton indistinctum and Ichthyophis paucesulcus (Welsch 1981) may be only 1 µm thick, while in the tree frog, Hyla arborea, the barrier may be as thin as 0.6µm (Goniakowska-Witaliniska 1986): H. arborea has a relatively high metabolic rate (Goniakowska-Witaliniska 1973) and the skin is richly vascularized (Czopek 1965). The lungs of the terrestrial species, e.g., the toad, Bufo marinus (Smith and Rapson 1977), the tree frogs, Hyra arborea (Goniakowska-Witalinska 1986) and Chiromantis petersi (Maina 1989d; Fig. 84), are eminently elaborate, having a series of hierarchical septa which delineate the air cells, which range in diameter from 1.45 mm in Rana pipiens to 2.3 mm in Bufo marinus and Rana catesbeiana (Tenney and Tenney 1970). The respiratory surface area in the lungs of the more terrestrial species is higher than that in the lungs of the aquatic ones (Tenney and Tenney 1970). Elementary lungs are adequate in amphibians, a group which characteristically has low aerobic metabolism (Whitford and Hutchison 1967; Goniakowska 1973; Goniakowska-Witalinska 1974; Feder 1976; Guimond and Hutchison 1976). Together with the Dipnoi, the amphibians have the largest tissue cells among vertebrates (Wintrobe 1934; Misiek and Szarski 1978; Szarski 1983). Amphiuma and Necturus exhibit great tolerance to anaerobiosis, withstanding 6h of total anoxia (Rose and Zambernard 1966). When exposed to hypoxia and hypercapnia, the amphibians utilize behavioral hypothermia to reduce their O2 need (Glass et al. 1983; Riedel and Wood 1988; Wood and Glass 1991).

Though the first vertebrates to invade land some 300 million years ago, by erecting a strong dependence on water for essential processes like aquatic oviparous reproduction, in regard to specific and numerical abundance and ecological distribution, modern amphibians constitute only a rather obscure vertebrate taxon. Their geographical distribution corresponds with freshwater, wet, high humidity, and high rainfall areas, feature which reflect strongly on their physiology. Cutaneous respiration (CR) presents very few possibilities for innovative designs and is generally considered an evolutionary dead end. Gans (1970), however, argued that CR may be a secondary condition in amphibians. This observation was supported by the suggestion by Romer (1972) that the Carboniferous amphibians may have been well scaled and thus had a water-impermeable skin cover. For all it is worth, the water-permeable skin of modern amphibians may represent a specialized secondary condition!

#### 6.7.3 The Reptilian Lung

The reptiles were the first vertebrates to be adequately adapted for terrestrial habitation and pulmonary respiration. By evolving a cleidoic egg and an impermeable surface cover, parameters above those achievable by amphibians from which they evolved (e.g., Olmo 1991), they were able to delink their physiology from water. The Mesozoic era, which lasted for nearly 200 million years, is often called the age of the reptiles, as the taxon dominated the Earth. Reptiles are exclusively lung breathers. Like the amphibians, reptiles display remarkable pulmonary structural diversity which to an extent can be correlated with the diverse habitats occupied and lifestyles led. There is no single model for the reptilian lungs. Based largely on the nature of internal organization (e.g., Milani 1894; Marcus 1937; Baudrimont 1955; Duncker 1978a, 1979; Perry 1983, 1992a; Hlastala et al. 1985), different morphological classifications of the lungs have been attempted. The lungs range in complexity from the profusely compartmentarized (multicameral) ones of the turtles, monitor lizard, crocodiles, and snakes (Perry 1978, 1988; Perry and Duncker 1978, 1980; Maina et al. 1989b; Maina 1989e; see Figs. 92,97) through the less elaborate (paucicameral) ones of the chameleons (Fig. 96) and the iguanids to the simple, saccular, smooth-walled, transparent, (unicameral) ones of, e.g., the teju lizard, Tupinambis nigropunctatus (Klemm et al. 1979; Perry 1983). This classification is overly simplistic as transitional forms and gradations occur. The simplest lungs, which correspond in development to the amphibian lung, occur in the Sphenodontia. Such lungs have a central air duct and peripherally situated, shallow air cells, which give a low surface-to-volume ratio.

The brochoalverolar lung of mammals and parabronchial lung of birds are thought to have evolved from transformation of a multicameral lung (e.g., George and Shah 1956, 1965; Duncker 1978a; Klaver 1981; Perry 1983, 1989; Becker et al. 1989). With an elaborate anterior space in which much of the gas exchange occurs, and a simple posterior one analogous to the air sacs of the bird lung, the design of the avian lung-air sac system is more closely related to the reptilian lungs (Brackenbury 1987). While the reptilian lungs fill up from the peripheral walls into the axial air space (centripetal = centralizing = compacting growth), the air conduits constituting the unfilled gaps, the mammalian and avian lungs fill from inside (centrifugal = radiative = diffusive growth), i.e., from outward bifurcation of the central airways. These developmental differences may account for the fact that the volume density of the parenchyma in the reptilian lung is only 25.2% in the tegu lizard, *Tupinambis nigropunctus*, 32.1% in the monitor lizard, *Varanus exanthematicus* (Perry 1981), and 25% in the Nile crocodile, *Crocodylus*  niloticus (Perry 1988). In the avian lung the average value is 50% (e.g., Maina et al. 1989a) and in mammals the value is as high as 90% (e.g., Gehr et al. 1981; Maina and King 1984). The faveolar air spaces in the parenchyma constitute 40% of the intrapulmonary air in the crocodile lung (Perry 1988), a value lower than that in the mammalian lung (about 56%; Maina and King 1984) and 53% in the bird lung (Maina et al. 1989a). In the more advanced snakes, e.g., Colubridae, Viperidae, and Elapidae, the left lung is greatly reduced or is totally lacking. In the primitive species, e.g., the boas and the pythons (Cope 1894; Verde 1951; Luchtel and Kardong 1981; Pohunkova and Hughes 1985b; Maina 1989e; Pastor 1995), the left lung occurs. The right lung is atrophied in the Amphisbenia (Gibe 1970). In the order Squamata, single-chambered lungs predominate especially in the families Teiidae (Klemm et al. 1979), Scindae (Gibe 1970), Lacertidae (Meban 1978a), and Gekkonidae (Perry et al. 1989b). Similarly, simple lungs occur in the family Angioidea (Meban 1978b). The land-based chelonians have paucicameral lungs, i.e., lungs with two or three peripheral compartments, which open into a central air space and lack an intrapulmonary bronchus. The marine species have multichambered bronchiolated lungs (Solomon and Purton 1984; Pastor et al. 1989). The elongated lungs of the ophidia and the amphisbaenids are divided into two functional zones, an anterior respiratory region which is well vascularized and a posterior one which is saccular and avascular (Kardong 1972; Klemm et al. 1979; Stinner 1982; Maina 1989e; Pastor 1995). In the crocodile lung, most of the parenchyma is located in the anterior two thirds of the lung where the blood makes 38 to 50% of the total volume (Perry 1988). The posterior part of the lung is thought to store air (e.g., Heatwole 1981), serve a hydrostatic role (Graham et al. 1975), and mechanically ventilate the anteriorly located exchange tissue in the manner of air sacs in birds: the arrangement may enhance the efficiency of gas exchange (Gratz et al. 1981; Stinner 1987; Vitalis et al. 1988). In heterogenous lungs, the design should convey a distinct functional advantage since the lungs of the more primitive reptiles are more homogenous (e.g., Luchtel and Kardong 1981). The varanids present the greatest level of pulmonary complexity in the suborder Sauria. Varanus exanthematicus and the pancake tortoise, Malacochersus tornieri, have multichambered lungs with a bifurcated intrapulmonary bronchi and profuse internal subdivision (Perry and Duncker 1978; Maina et al. 1989b; Fig. 93). The single-chambered lungs with an edicular parenchyma are thought to require low energy for convective ventilation: they occur in animals with low metabolic rates (Pastor 1995). The reptilian lung constitutes 5% of the body weight (Tenney and Tenney 1970). In animals of similar body mass, a reptile has a lung volume which is seven times greater than that of a mammal (Crawford et al. 1976; Glass and Johansen 1981) but the diffusing capacity for O<sub>2</sub> is relatively low (Crawford et al. 1976). The aerobic capacity of reptiles is remarkably lower than that of mammals. At a temperature of 37 °C, a 1kg lizard consumes  $122 \text{ ml } O_2 \text{ h}^{-1}$ , a value which constitutes 18% of the O<sub>2</sub> consumption of an equivalent-sized mammal (Bennett and Dawson 1976). The muscle capillary surface per unit muscle mass of a reptile is about 20% the value of a mammal of similar size (Pough 1980). At maximal exercise, anaerobic metabolism provides 86% of the total energy consumption in the water snake, Natrix rhombifera (Gratz and Hutchison 1977). At a body temperature of 20 to 23 °C, the

physiological diffusing capacity of the reptilian lung (DLo<sub>2</sub>p) is an order of magnitude smaller than that of a mammal of the same size (Crawford et al. 1976; Glass and Johansen 1981). The DLo<sub>2</sub>p of the lung is similar in reptiles and amphibians (Glass et al. 1981a,b; Gatz et al. 1987; Lutcavage et al. 1987). Compared with mammals (e.g., Baldwin and Winder 1977; Scheuer and Tipton 1977; Dudley et al. 1982), where endurance exercise precipitates changes such as increases in tissue oxidative capacities and O<sub>2</sub> consumption, the adaptive response of lizards, *Amphibolurus nuchalis*, to endurance exercise is different (Gleeson 1979; Garland et al. 1987): trained lizards exhibit decreased heart and muscle masses but increases in liver mass, hematocrit, liver pyruvate kinase, and heart citrate synthetase activities. Interestingly, physical training enhances the swimming performance of the African clawed frog, *Xenopus laevis* (Miller and Camilliere 1981).

The epithelial cells lining the respiratory surface of the reptilian lung, are completely differentiated into types I and II cells (e.g., Okada et al. 1962; Nagaishi et al. 1964; Luchtel and Kardong 1981; Daniels et al. 1990; Maina 1989d.e; Perry et al. 1989b) and type III cells - the brush cells (Gomi 1982). A rare mitochondriarich cell has been described in the lung of the turtle, *Pseudemys scripta* (Bartels and Welsch 1984). The type I cells are squamous and have remarkably thin, long cytoplasmic extensions. The much smaller, rather cuboidal surfactant-secreting type II cells are scattered between the type I cells (Figs. 84,85,87,90). In general, the pneumocytes in the lungs of the amphibians (Goniakowska-Witalinska 1995) and the lungfishes (Hughes and Weibel 1976; Maina 1987a) are undifferentiated while those in the reptilian, mammalian, and avian ones are. The differentiation of the pneumocytes in the higher vertebrates may be an adaptive strategy for greater functional efficiency (Maina 1994): reducing the numerical density of the more metabolically active surfactant-producing type II cells to a minimum and the type I cells adopting an extremely thin form lowered the overall O<sub>2</sub> consumption by the tissues of the gas exchanger. Furthermore, the design generated a thin blood-gas barrier, an important structural feature which enhanced the diffusing capacity of the gas exchanger for O<sub>2</sub> (Liem 1987a; Maina 1987a). Sporadic attenuation of the endothelium (Figs. 40,90), where extremely thin areas of the bloodgas barrier are generated without jeopardizing the mechanical integrity of the lung (Figs. 29b,40a,90) is a common scheme evoked to maximize gas transfer in the lungs of the higher vertebrates (e.g., Weibel 1973; Maina and King 1982a). Dust cells (macrophages) have been described in some reptilian lungs, e.g. in the turtle, Testudo graeca (Pastor et al. 1989). Unlike in the mammalian (Fig. 86) and bird lungs (Figs. 88,90), where owing to a single capillary system arrangement the exposure of the pulmonary capillary blood to air is very efficient, in the reptilian lung, depending on the species and the particular areas of the lung, a double capillary system (Fig. 85) commonly occurs. Reptilian lungs have a prevalence of smooth muscle tissue (Fig. 80). In the tegu and the monitor lizards, respectively, smooth muscle tissue constitutes 7.4 and 1.3% of the nontrabecular tissue (Perry 1981; Perry et al. 1989b,c). The smooth muscle tissue has been associated with intrapulmonary convective movement of air (Klemm et al. 1979; Tenney et al. 1984; Carrier 1988). The compliance of the lung of the garter snake, Thamnophis sirtalis of 0.042 ml per cm H<sub>2</sub>O per g (Bartlett et al. 1986) is 50 times that of the lung of a mouse, a mammal of about the same body mass (Bennett and Tenney



Fig. 90. High power view of the pulmonary blood capillaries, c, of the lung of the domestic fowl, *Gallus domesticus*, showing the blood-gas barrier,  $\succ$ , which is characterized by sporadic attenuations especially of the endothelial cell.  $\rightarrow$ , endothelial cell junctions; r red blood cells; e nucleus of the red blood cell; a air capillary. *Bar* 0.5 µm

1982). At the peak of an expiratory phase, the residual volume of air  $(18 \text{ mlkg}^{-1})$  in the crocodile lung is only 13% of the maximal lung volume (Perry 1988). The compliance of the lung of the crocodile  $(0.7 \text{ ml cm}^{-1} \text{ H}_2\text{O g}^{-1})$  is over four times that of the body wall (Perry 1988). The overall compliance of the reptilian lung is determined by the contractile elements of the lung, i.e., smooth muscle and elastic tissue, the saccular nature of the lung (Craig 1975), and the very effective pulmonary surfactant (Perry and Duncker 1978). Coupled with the irregular pattern of breathing, the properties may provide an energy-saving system on the respiratory work (Milsom 1984). The volume-specific lung compliance of the multicameral lung of the crocodile is similar to that of the much simpler lung of the gecko (Perry 1988; Perry et al. 1989b), suggesting that lung compliance in the reptilian lung may be an attribute of the parenchymal structure and not of lung type. The reptilian lungs may serve as air stores during apnea and in the aquatic species may support extended dives (Ackerman and White 1979). In the alligator, the pulmonary O<sub>2</sub> store constitutes 85% of the total lung volume (Andersen 1961).

Turtles can withstand complete anoxia for days or months, in the process accumulating lactic acid levels up to 200 mmoll<sup>-1</sup>, surviving a decrease in brain pH to 6.4 (Glass and Wood 1983). During the episodic apneic periods, which may last for a few minutes to hours (Glass and Wood 1983), the perfusion of the gas exchanger is reduced through a decrease in heart rate, stroke volume, and/or by the blood being shunted away from the lung. Ventilation-perfusion matching is necessary for efficient gas exchange (Burggren et al. 1977; Wood et al. 1978). In the Chelonia, the difference between the  $PO_2$  in air spaces and that in the arterial blood (a factor determined by central cardiovascular admixture of the systemic venous blood into the systemic arteries and shunting of the blood within the pulmonary circulation; Seymour 1978) may be as high as 6 to 6.7 kPa (Burggren and Shelton 1979), in sea snakes it may exceed 8kPa (Seymour and Webster 1975), and in resting lizards it ranges from 2.7 to 13 kPa (Mitchell et al. 1981). The respiratory parameters such as ventilation-perfusion ratio, respiratory frequency, and tidal volume are difficult to characterize in the periodic breathers (Glass et al. 1979). It has been hypothesized that structural limitations in the design of the reptilian lungs prevented the reptiles from attaining endothermic-homeothermy (Perry 1992a), relegating their aerobic capacities behind birds and mammals. In the green turtle, Chelonia mydas, during swimming, intermittent breathing gives way to a continuous breathing pattern (Butler et al. 1984). Uncharacteristic of reptiles, green turtles can endure sustained exercise: they make long migrations between their feeding grounds and breeding beaches, covering distances of as much as 4800 km, most of the time continuously swimming in the open sea at speeds of 2 m s<sup>-1</sup> (Carr and Goodman 1970; Carr et al. 1974). They can increase their O<sub>2</sub> consumption by a factor of 3 (Prange 1976; Butler et al. 1984) without resulting in a significant level of anaerobic metabolism as most reptiles do during long bouts of activity (e.g., Bennet 1982). Hypoxia (10% O<sub>2</sub>) caused continuous ventilation in resting green turtles and pulmonary blood flow was elevated and sustained (West et al. 1992): during swimming, increased heart rate was accompanied by a sevenfold increase in the ventilatory rate. Interestingly, physiological conditions of periodic ventilatory flow pattern do constitute a significant impairment in the parabronchial gas exchange efficiency in the avian lung (Scheid et al. 1977). This may indicate the remarkable functional reserves inherent in the bird lung (Sect. 6.7.5) where adjustments can be made to overcome certain limitations.

### 6.7.4 The Mammalian Lung

The beginning of the Tertiary saw the mammals take over and eclipse the reptiles as the dominant terrestrial vertebrate group (Romer 1967). The appearance of the placental mammals (subclass: Eutheria) formed the pinnacle in the development of the taxon. Among vertebrates, the mammalian lung, especially the human one, has been best studied both structurally and functionally. Recent excellent integrative reviews on these aspects include those by Weibel (1984a,b, 1986) and Burri (1984a,b). Owing to the abundance of literature in the area and need for brevity

here, in-depth discussion of the mammalian lung (except for that of bats, which are less well known) will not be made. Suffice it to say now that the blind-ending, tidally ventilated mammalian lung falls far short of the level of efficiency attained by other gas exchangers, e.g., the insectan tracheal system and the avian lung-air sac system. With respect to O<sub>2</sub> consumption in standard conditions as well as maximal activity, Dejours (1990) observed that "compared with other mammals, the human exhibits no species-specific characteristics". Among the air-breathing vertebrates, the heterogeneity of the design of the lung decreases from the ectotherms - the Dipnoi (lungfishes), amphibians, and reptiles to the endotherms mammals and birds. Morphological homogeneity appears to be an essential attribute for optimal design in gas exchangers. Structurally, the lungs of mammals are similar except for fine differences like bronchiolar bifurcation, lobulation, and topographic relationships between the airways and the vasculature. The need to optimize respiratory function must have become critical with changes to lifestyles which called for increased metabolic demands for O<sub>2</sub>. Over and above the phylogenetic level of development, the organization complexity of the gas exchangers and respiratory stratagems adopted by an animal are greatly determined by the needs placed on them and the habitat occupied. For example, in the lungs of the nonhuman primates, e.g., the baboon, Papio anubis (Maina 1987b), and the vervet monkey, Cercopithecus aethiops (Maina 1988b), the lungs of the small, supposedly primitive prosimians, e.g., the lesser bushbaby, Galago senegalensis (Maina 1990c), are better from a morphometric perspective. The naked mole rat, Heterocephalus glaber, a small, eusocial poikilothermic fossorial bathvergid rodent which lives in thermostable burrows, has remarkably neotenic lungs (Maina et al. 1992). The diffusing capacity of the lung of the Japanese waltzing mouse, Mus wagneri, a pathologically hyperactive animal, is 55% greater than that of a normal one (Geelhaar and Weibel 1971). Lungs of high altituderaised rats have smaller alveoli and a greater respiratory surface area than sea level dwellers (Pearson and Pearson 1976). Experimental lobectomy results in a compensatory overgrowth (Rannels and Rannels 1988), the remaining lobes in young rats attaining the same diffusing capacity as the control animals (Burri and Sehovic 1979).

### 6.7.4.1 Lungs of Bats

Bats (order: Chiroptera) are unique among mammals by their ability to fly (e.g., Thewissen and Babcock 1992). Flight has enabled remarkable adaptive radiation to occur in bats. Of all known mammalian species, one in five is a bat. About 800 species of bats have been recognized, the number falling between that of Rodentia (1600) and Insectivora (400). After the human being, *Myotis* (family: Vespertilionidae) is reputed to be the most widely spread naturally occurring mammalian genus on Earth (Yalden and Morris 1975). Despite the abundance and the wide geographical distribution, perhaps due to their elusive nocturnal lifestyle to which they were relegated by the birds, the only other older apparently well-established volant vertebrate, bats remain animals of curiosity, myth, and prejudice. The phylogenetic affinity between bats and the mainstream mammals is not clear (e.g., Novacek 1980, 1982; Scholey 1986). The groups which have evolutionary been closely associated with bats include Scandentia (Tupaiidae or tree shrews), primates, and the Dermoptera (e.g., van Valen 1979; Padian 1982; Pettigrew et al. 1989). Bats comprise the Suborders Megachiroptera and Microchiroptera and are considered by some investigators, e.g. van Valen (1979) and Novacek (1982), to be monophyletic, while others, e.g., Jepsen (1970), Smith (1977), Scholey (1986) and Pettigrew et al. (1989), consider them diphyletic. The earliest reliably known fossil record of a bat is that of *Icaronycteris index* of the Eocene (50 million years ago) which morphologically resembles the modern Microchiroptera (Jepsen 1970): the protobats may have been treedwelling omnivores which started to glide between trees while foraging (Norberg 1981, 1986). The general anatomy of bats is mammalian (Yalden and Morris 1975).

Powered flight is defined as capacity to produce lift, accelerate, and maneuver at various speeds (e.g., Pennycuick 1975; Norberg 1976a,b; Rayner 1986). The large number of animals which are said to fly, for example the freshwater butterfly fish, Pantodon buchholzii of the West African rivers, the parachuting frog of Borneo, Rhacophorus dulitensis, the flying snakes of the jungles of Borneo, Chrysopelea sp., the flying squirrel of North America, Glaucomys volans, the flying lemur, Cyanocephalus volans, and the East Indian gliding lizard, Draco volans, are essentially acrobatic passive gliders or parachutists which use part of their body to slow down a fall by using drag and lift (see Scholey 1986; Davenport 1994). Energetically, powered flight is a highly demanding mode of locomotion which has evolved in only a few elite animals (Tucker 1972; Berger and Hart 1974; Carpenter 1975; Thomas 1975), the insects, pterosaurs, birds, and bats - chronologically in that order. The mass-specific aerobic capacities of flying bats are essentially the same as those of forward-flapping birds but are 2.5 to 3 time those of running mammals of the same size (Thomas and Suthers 1972; Carpenter 1975; Thomas 1987). Bats can increase their  $O_2$  consumption during sustained flight by a factor of 20 to 30 times (Bartholomew et al. 1964; Thomas and Suthers 1972). At an ambient temperature of 20 °C, a 12-g bat, Myotis velifer, is reported to increase its O<sub>2</sub> consumption by an astounding factor of 130 (Riedesel and Williams 1976). By avian standards, bats are excellent fliers in terms of speed, distance, and maneuverability (Vaughan 1966; Griffin 1970; Norberg 1976a,b; Fenton et al. 1985; Norberg and Rayner 1987). Speeds of 16km h<sup>-1</sup> in Pipistrellus pipistrellus (Jones and Rayner 1989), 30 to 50 km h<sup>-1</sup> in Myotis (Hayward and Davis 1964) and 64 km h<sup>-1</sup> in *Eptesicus fuscus* (Petterson and Hardin 1969) have been estimated. Migratory distances of about 1000 km have been reported in Lasiurus borealis, L. cinereus, Lasionycteris noctivagans, Nyctalus noctula, and Tadarida brasiliensis (Baker 1978; Thomas 1983). As they forage, Epomophorus wahlbergi and Scotophilus viridis are known to cover distances of about 500 km in a night (Fenton et al. 1985). Flight style and optimum speed in bats depend on, among other factors, choice of food, foraging behavior, and habitat selection (Norberg 1981). The small bats show a greater scope for flight with respect to agility and are even able to momentarily hover (Norberg 1976a,b). However, unlike birds, which commonly use energy-saving modes of flight like gliding and soaring (Rayner 1985), bats only rarely do so. *Pipistrellus pipistrellus* adopts gliding flight for only
13.4% of its flight time and the glides last for only 0.1 to 0.3 s (A.L.R. Thomas et al. 1990).

Despite the basic limitations intrinsic to the design of their characteristically mammalian lungs (Yalden and Morris 1975; Maina 1985), bat lungs have been structurally highly refined (e.g., Maina and Nicholson 1982; Maina et al. 1982a, 1991; Maina and King 1984; Maina 1986). These features have been functionally closely integrated with other anatomical and physiological aspects, enhancing the uptake and transport of the necessary large amounts of O<sub>2</sub> for flight. The strategies necessary to afford flight in bats evolved within the constraints presented by an inferior lung. This is a classic case of the innate plasticity of biological systems. In bats, a typical mammalian lung was exquisitely modified to exchange respiratory gases during flight at rates equal to those of the seemingly better-designed bird lungs. The most important parameters in this portfolio included (1) development of relatively large hearts with a huge cardiac output (Hartman 1963; Snyder 1976; Jürgens et al 1981), (2) high hematocrit, hemoglobin concentration, and O<sub>2</sub> carrying capacity of blood (e.g., Riedesel 1977; Wolk and Bodgdanowicz 1987), and exceptional pulmonary structural parameters (Maina et al. 1982a, 1991; Lechner 1984; Jürgens et al. 1981; Maina and King 1984). Phyllostomus hastatus can maintain the same high lung  $O_2$  extraction factor of 20%, a value comparable to that of bird during the metabolic stress of flight (Thomas 1981; Thomas et al. 1984) or when at rest during exposure to a severe hypoxic stress (Farabaugh et al. 1985). The mean resting  $O_2$  extraction factor at thermoneutral range in *Noctilio* albiventris of 18.3% lies between that of a bird (20.8%) and a nonflying mammal (16.6%) of the same body mass (Chappell and Roverud 1990). It increases from 35 to 40% at low ambient temperature. These values surpass those of most birds under similar conditions (Bucher 1985). One of the highest venous hematocrits in the vertebrates (75%) has been reported in a specimen of a 13-g bat, Tadarida mexicanobrasiliensis (Black and Wiederhielm 1976). In the five species of bats examined by Jürgens et al. (1981), venous hematocrits ranged from 51 to 63% and hemoglobin concentration and erythrocytes numbers were respectively 24.4 g dl<sup>-1</sup> and 26.2 million  $l^{-1}$ . While high hematocrit may enhance O<sub>2</sub> uptake, the advantages conferred are soon compromised by the increased viscosity of blood (Stone et al. 1968; Hedrick et al. 1986; Hedrick and Duffield 1991). On account of the different effects of hematocrit on blood  $O_3$  capacity and viscosity, where  $O_3$  capacity increases linearly with the hematocrit while viscosity increases exponentially, an optimal O<sub>2</sub> transport level is established (Crosswell and Smith 1967; Shepherd and Riedel 1982; Kiel and Shepherd 1989). Increase in the hematocrit beyond optimal level causes a reduction in cardiac output, maximum O<sub>2</sub> consumption, and aerobic scope (Hillman et al. 1985; Tipton 1986).

In all bats which have been studied, a 1:1 synchronization between wing beat and breathing cycles has been observed (Suthers et al. 1972; Thomas 1981, 1987; Carpenter 1985, 1986): in birds, this occurs in only a few species (e.g., Tomlinson 1963; Berger et al. 1970; Berger and Hart 1974; Butler and Woakes 1980). It has been suggested (e.g., Bramble and Carrier 1983) that locomotory activity provides mechanical assistance to respiratory muscles. In birds, though flight muscles attach on the sternum, there appears to be very little effect on the actual pulmonary respiratory air flow and volume. Bernstein (1987), however, suggests that the

disparity between the wing beat to breathing rates may be caused by the complex nature of the air flow in the avian parabronchial lung. In the European starling, Sternus vulgaris, the ventilatory volume change associated with wing beat ranges from 3 to 11% at most (Banzett et al. 1992). To a vet undetermined degree, the synchronization of wing beat with breathing cycles must enable bats to ventilate their lungs at a lower cost and probably more efficiently. In the bats Antrozous pallidus (Basset and Wiederhielm 1984) and Myotis daubentoni (Krátký 1981; Lundberg et al. 1983), the maturation of the  $O_2$  transport system (i.e., blood  $O_2$ capacity) to the adult status closely coincides with the start of flight behavior. Bats have exceptionally large lungs which occupy a large proportion of the coelomic cavity (e.g. Maina and King 1984; Maina et al. 1991). The gastrointestinal system is small and morphologically poorly differentiated (Makanya and Maina 1994; Makanya et al. 1995). Compared with nonflying mammals, bats have relatively large respiratory surface areas. The mass-specific respiratory surface area of  $138 \text{ cm}^2 \text{g}^{-1}$  reported by Maina et al. (1982a) in the epauletted fruit bat, Epomophorus wahlbergi, is the highest value so far reported in a vertebrate. Compared with nonflying mammals, bats have relatively thin blood-gas barriers. The thinnest blood-gas barrier so far reported among mammals is that of 0.1204 µm in Phyllostomus hastatus (Maina et al. 1991), a bat in which the O<sub>2</sub> extraction factor equals that of an energetic bird of comparable size (Thomas 1987) and which has a venous hematocrit of 60% (Jürgens et al. 1981). Pulmonary respiratory surface area can be increased by an overall enlargement of the lung, as occurred in bats, and/or increased subdivision of the gas exchange tissue, as in birds. In the compliant mammalian lung, increased subdivision of the parenchyma, however, generates smaller alveoli which are not only highly susceptible to collapse (due to large surface tensional forces at the air-tissue interface) but obligate greater amount of energy to inflate.

Unlike birds, which have dispersed widely and penetrated the remote cold regions of the world such as Antarctica, bats are largely tropical and neotropical in distribution (Wimsatt 1970; Yalden and Morris 1975; Carpenter 1985). This may probably be due to the need for reliable food sources in order to procure adequate metabolic substrates for production of the large amounts of energy for flight. Furthermore, due to the relatively poor insulation of the bat wings and presence of skeletal muscles on the wings (compared with the feathered bird wings which do not have muscles on the wing surface), excessive convective heat loss to the cold air at extremely low temperatures may occur in bats. The wing muscles may be cooled to a critical temperature where the proper coordination which is necessary for efficient flight is curtailed. On isolated forearm muscles from a number of species of temperate zone bats, Nelson et al. (1977) observed that the duration of contraction of forearm muscles at temperatures below 8°C were five to ten times longer than those at 32 °C. However, some bats have been reported to fly in ambient temperatures as low as -5 °C (O'Farrell and Bradely 1977). Temperate-zone bats are known to hibernate or migrate to warmer regions of the world during winter (Kulzer 1965; A.L.R. Thomas et al. 1990). In hibernating Myotis lucifugus, O<sub>2</sub> consumption is 1.5% of their respective normothermic resting rates at or about thermoneutrality (Hock 1951): at 5°C, breathing is arrhythmic in character, and apneic phases last for as long as 48 min.

In principal, animals use different strategies to attain optimal states (Howell 1983). Due to their remarkably different evolutionary backgrounds, different genetic resources, designs, and strategies were utilized by bats and birds to independently meet the aerodynamic and energetic requirements of flight. In pursuit of optimization of respiration, bats and birds evoked remarkably different paradigms: structural and functional parameters were variably integrated to promote gas exchange efficiency (Maina 1998). From modeling anatomical and physiological data, it appears that in bats, the process occurred through synergism of performances of relatively fewer, highly refined parameters which operate at or close to their maximum capacity. We have called this the narrow-based - highkeyed scheme (NB-HKS). Birds, on the other hand, have incorporated a wide spectrum of parameters in their gas exchange arsenal (and conspicuously conserved some) in what has been termed the broad-based - low-keyed scheme (BB-LKS): similar to this scheme, Heinrich (1983) has described a program of integrated suboptimal parts for a better whole in foraging bumblebees. Inevitably, consequent to the different respiratory contrivances in birds and bats, different functional reserves have evolved. In bats, the NB-HKS leaves a very limited margin of operation while the BB-LKS of birds affords an enormous reserve. This model-based inference (Maina 1998) is supported by the observation made by Chappell and Roverud (1990) that whereas with changing ambient temperature birds and nonflying mammals adjust respiratory frequency and tidal volume to meet changing O<sub>2</sub> demands, O<sub>2</sub> extraction remains fairly constant and in some species actually decreases (Casey et al. 1979; Withers et al. 1979; Bucher 1985; Chappell 1985; Kaiser and Bucher 1985; Chappell and Souza 1988; Bucher et al. 1990), bats (at least the lesser bulldog bat, Noctilio albiventris, on which data are available) accommodate varying thermogenetic O<sub>2</sub> consumption by simultaneously changing all three factors, namely respiratory frequency, tidal volume, and O<sub>2</sub> extraction, which may increase by a factor of as much as 2. The margin of operation in bats depends on factors such as the magnitude, nature, and duration of exposure to stress. Some degree of respiratory functional reserve must, however, exist in bats since they are able to absorb changes which call for moderate increases in O<sub>2</sub> demand such as flight during pregnancy, flight after the premigratory weight gain, and thermogenetic heat production associated with low ambient temperatures. For example, a female red bat, Lasiurus borealis, reported to weigh 12.9 g and presumed to have a wing loading of  $0.09 \,\mathrm{g \, cm^{-2}}$  is reported to have carried four young ones whose total mass was 23.4 g, i.e., 181% of her body mass (Staines 1965). In experimental tests to assess weight-lifting capacities, Davies and Cockrum (1964) observed that a female long-eared bat, *Plecotus townsendii*, with a body mass of 10.1 g, could lift 73.3% of her own mass, but Tadarida brasiliensis could manage only 9.3%. No bat is truly herbivorous as rodents are. Fermentative microbial digestion of cellulose calls for a voluminous gastrointestinal system (GIT) (e.g., Chivers and Hladik 1980; Warner 1981) and a long transit time for the ingesta (e.g., Balch and Campling 1965). Such an increase in mass for low energy yield per unit weight of the ingesta perhaps would be untenable for flight: bats have simple GITs contained in restrictive abdominal cavities (Forman 1972; Makanya and Maina 1994; Makanya et al. 1995) and the transit times of the ingesta must be relatively short. Bats of the temperate regions are exclusively insectivorous but tropical regions support both fruit-eating and insectivorous bats (Wilson 1973; Yalden and Morris 1975). While there are numerous herbivorous birds, e.g., the hoatzin, *Opisthocomus hoazin* (Dominguez-Bello et al. 1993), such birds are fairly large and have poor, if any, flight capacity.

From differentiation and integration the factors involved in gas exchange (Maina 1998), birds appear to have refined the structural parameters in preference to the physiological ones in contrast to bats, which favored the physiological ones: about 60% of the gas exchange capacity in birds can be attributed to the structural parameters and 40% to functional ones while in bats, 61% of the capacity can be attributed to physiological factors and 39% to the structural ones. Being generally small in size, the heaviest bat, Pteropus edulis, being about 1.5 kg (Yalden and Morris 1975; Carpenter 1985; Pough et al. 1989), leading a particularly energetic lifestyle, and operating from a level of a rather inferior lung, bats appear to have had very few choices but to fully refine and maximally exploit practically all the resources available to them. An "optimized" species is threatened with extinction if and when it is faced with increased demands to which it cannot respond (e.g., Minkoff 1983). Appropriate behavior and niche selection help minimize ambient pressures. In bats, we may unconsciously be witnessing a taxon living within very narrow tolerance limits of the environmental factors. The nocturnal lifestyle led by the bats may be explained by their more recent evolution of flight long after the birds had firmly occupied the diurnal niche for about 100 million years. The rather mundane respiratory physiological specializations for flight apparent in bats ruled out any possibility of successfully competing with birds.

### 6.7.5 The Avian Lung

It has been speculated by, e.g., Duncker (1978b) and Perry and Duncker (1980), that the avian lung-air sac system had its origin in the multicameral reptilian lungs like those found in the monitor lizards. Such lungs have a large ventilatable surface area (Maina et al. 1989a). The close phylogenetic affinity between reptiles and birds (e.g., de Beers 1954; Ostrom 1975; Jones et al. 1993) accounts for the anatomical similarities which abound between the two taxa. The chameleons possesses extensions from the lung similar to the air sacs of birds (Grassé 1970; Patt and Patt 1969; Klaver 1973, 1981; Fig. 96). Having arisen from reptiles much later than mammals, birds are phylogenetically more primitive than mammals (Romer 1967). From the arguments that some degree of endothermy may have developed in the Mesozoic and even Paleozoic reptiles (e.g., Romer 1967; Bakker 1975) and from the presence of a robust pectoral girdle, extensively ossified sternum, and expansive deltopectoral crest of the humerus (Romer 1966) (features which indicate well-developed flight muscles, excellent capacity to fly, and hence a high aerobic capacity), it has been conjectured that the direct progenitors of birds, the pterosaurs (order: Archosauria) of the Jurassic and Cretaceous, e.g., the 250-kg *Quetzalcoatlus northropi* (Langston 1981; Paul 1990, 1991) and *Pteranodon* (Bramwell 1971; Padian 1983) had developed complex, multichambered lungs. We shall, however, never know for certain about the structure and function of the lungs in these interesting now extinct reptiles. Fundamental differences in the design of the lungs with respect to respiratory gas flow pattern and overall efficiency occur between reptiles, birds, and mammals (Scheid and Piiper 1987).

Unique to the other vertebrate air-breathing groups, in birds, the lung has been uncoupled from the compliant bellows-like ventilatory air sacs (e.g., Fedde 1980; Maina 1983, 1996): the avian lungs are compact and virtually nonexpansile (Fig. 91). They are continuously ventilated by a synchronized action of the totally avascular air sacs (Lucas and Denington 1961; Marin-Girón et al. 1975) which play no role in gas exchange (Magnussen et al. 1976). During respiration, the volume of the lungs changes by a mere 1.4% (Jones et al. 1985). Experimental compression of the lung does not result in a substantial collapse of the air capillaries (Macklem et al. 1979). Although birds have smaller lungs per unit body mass than mammals (Maina et al. 1982b, 1983, 1989a; Maina 1989a; Maina and Settle 1982), the virtual rigidity of the avian lung has resulted in a substantial increase in the surface area per unit volume of the lung. The respiratory surface exceeds that of mammals of similar body mass (Maina et al. 1989a). The "insertion" of the bird



**Fig. 91.** Cast of the lung of the domestic fowl, *Gallus domesticus*, showing compact nature. The air conduits include the primary bronchus, p, the secondary bronchi, e, and the parabronchi, x. The  $\succ$  show the anastomoses of the parabronchial system. *Bar* 8 mm

lung to the ribs followed by the relegation of the ventilatory compliance to the air sacs meant that surface tension was no longer a restricting factor in the extent of internal subdivision of the parenchyma. This resulted in extremely narrow terminal gas exchange components, the air capillaries, which gave rise to an extensive respiratory surface area and a thin blood-gas barrier (Maina 1981a; Maina and King 1982a). Considering the large number of species of birds (about 9000 species; e.g., Morony et al. 1975; Gruson 1976) and the remarkably diverse habitats they live in, the bird lung is morphologically remarkably homogenous. Subtle differences such as the degree of development of the parabrochi, spatial arrangement of the secondary bronchi, and size and location of air sacs, however, occur. The congruency in the morphological configuration of the bird lung may perhaps arise from the fact that all birds evolved from a common volant lineage (e.g., Cracraft 1986) and hence at one stage, they had to grapple with the demands for flight which some groups abandoned late. A fascinating difference in the morphologies of the bird lung is, however, that of the spatial arrangement, the degree of development, and the location of the tertiary bronchi (parabronchi). Two sets of parabrochial systems, the paleopulmo and neopulmo have been described (e.g., King 1966; Duncker 1974). The main differences between the papeopulmonic and the neopulmonic zones of the lung are: (1) the paleopulmonary parabronchi are located on the dorsocranial region of the lung and constitute about two thirds of the lung volume, while the neopulmonic set is located ventrocaudally and comprises about one third of the lung volume; (2) the paleopulmonic parabronchi are arranged in parallel stacks while the neopulmonic ones are irregularly arranged and anastomose profusely (e.g., López 1995); (3) while the air flow in the paleopulmo is continuous and unidirectional, that in the neopulmo changes with the phase of respiration (e.g., Fedde 1976); and (4) both sets of parabronchi start to develop at different embryonic stages (Romanoff 1960). The lungs of the primitive birds such as the kiwi and the penguin only have the paleopulmonic parabronchi while the neopulmo is variably well developed in the less conserved species, reaching maximal development in the relatively highly evolved passerines (King 1966; Duncker 1974). The substantive functional implication of the presence and absence of the development of the paleo- and neopulmo is still unclear. There are no morphometric differences in the structure of the gas exchange components in the two regions of the lung (Maina 1982b; Maina et al. 1982c). It is plausible that the bidirectionally ventilated neopulmonic parabrochi may provide a site for CO<sub>2</sub> cycling to alleviate occurrence of respiratory alkalosis due to excessive washout of CO2 across the undirectionally ventilated paleopulmonic parabrochi especially during panting in thermal stressed birds (e.g., Jones 1982). The ostrich can pant continuously for as long as 8h without experiencing acid-base imbalance (Schmidt-Nielsen et al. 1969). The avian lung is satisfactorily described by the crosscurrent model which considers that blood and parabronchial air interact at right angles (Fig. 32). Unlike in the mammalian alveolar system, under most conditions, the PCO<sub>2</sub> in the arterial blood (PaCO<sub>2</sub>) and the PCO<sub>2</sub> in the end-expired air (PECO<sub>2</sub>) are different, with the PaCO<sub>2</sub> being as much as 0.8 kPa less than PECO<sub>2</sub> (Scheid and Piiper 1970; Meyer et al. 1976; Hastings and Powell 1986). High-frequency respiration reduces the respiratory efficiency of the avian lung (Hastings and Powell 1987).

#### 6.7.5.1 The Energetic Demands for Endurance and High Altitude Flight in Birds

Flight is the most energetically demanding form of locomotion that has evolved in animals. It places substantial metabolic demands on an animal. Skeletal muscle accounts for 96% of a flying animal's total energy consumption during flight, as it does in a human being at maximum exercise (e.g., Folkow and Neil 1971). In practically all active vertebrate groups, locomotion exerts the greatest demands on the respiratory system (Banzett et al. 1992). A significant metabolic barrier differentiates volant vertebrates from nonvolant ones. The fact that birds and bats are the only extant vertebrates capable of powered flight attests to the extreme selective pressure these taxa have endured during the course of evolution of this mode of locomotion. Wind tunnel experiments show that the energetic demands of flight are beyond those attainable by nonflying vertebrates (e.g., Tucker 1968; Carpenter 1975; Thomas 1975). In turbulent air or when ascending, a bird can increase O<sub>2</sub> consumption for brief periods by about 20 to 30 times whereas even a good human athlete can attain such an increase for only a few minutes (Tucker 1970). The O<sub>2</sub> consumption in the pigeon while running was estimated to be 27.4 ml per min and while flying at 19m per s was 77.8 ml per min, a factorial difference of 2.8 (Butler et al. 1977; Grubb 1982). Although expensive in its absolute demands for energy, active flight is a highly efficient form of locomotion, as at fast speeds the distance covered per unit of energy expended is less than in most other forms of locomotion (Tucker 1970; Thomas 1975; Rayner 1981). In the bats, Phyllostomus hastatus and Pteropus gouldii, the energy required to cover a given distance is only one sixth and one fourth, respectively, of that needed by the same-sized nonflying mammal (Thomas 1975). At their optimal speeds, the minimum cost of flying for a 380-g bird is about 30% of the energetic cost of a 380-g mammalian runner (Hainsworth 1981). After evolving independently from reptiles, birds and small mammals acquired aerobic metabolic scopes between resting and maximal rates of exercise or cold-induced thermogenesis which are 4 to 15 times greater than those of their progenitors at the same body temperature (Bennett and Dawson 1976; Bartholomew 1982b; Dawson and Dawson 1982).

Whereas most birds fly at moderate speeds with the smaller and more agile passerines attaining speeds of 15 to 40kph, the swifts (Apodidae), the loons (Gaviidae), and the pigeons (Columbidae) have been reported to reach speeds of between 90 and 150kph, while the peregrine falcon (*Falco peregrinus*) has been reported to dive on its prey at a speed in excess of 180kph (Welty 1964): a yet unsubstantiated speed of 360kph has been reported in a diving male falcon. When the avian flight speeds, which may appear mediocre by general standards, are normalized with body lengths covered, they turn out to be remarkably fast. The small passeriform birds, e.g., swifts, starlings, and chaffinches, flying at a moderate speed of 40kph, cover about 100 body lengths per s (Alerstam 1982; Kuethe 1975) compared with only 5 in a highly athletic human being and only 18 body lengths per s in the cheetah, one of the fastest land mammal (Hildebrand 1959, 1961). In its annual migration, the Arctic tern (*Sterna paradisea*) flies from pole to pole, a distance of 35000km between breeding seasons (Berger 1961; Salomonsen 1967) while the American golden plover (*Pluvialis dominica*) flies

3300km nonstop from Aleutian Islands to Hawaii in only 35h (Johnston and McFarlane 1967). Collision between a vulture with a jet craft at an altitude of 11 km was reported by Laybourne (1974) and a flock of swans (probably whooper, Cygnus cygnus) was observed by radar at an altitude of 8.5km (Stewart 1978; Elkins 1983). Amazingly, the 3-g ruby-throated hummingbird (Archilochus colubris) flies nonstop for nearly 1000km across the Gulf of Mexico from the Eastern United States, a distance which may require about 20h to cover (Lasiewski 1962, 1963a,b). A 3- to 4-g rufous hummingbird (Selasphorus rufus) has about 1 g of flight muscles which consume  $O_2$  at a rate of 82 µmol per minute (Suarez et al. 1990). Many passerine species are known to fly continuously for 50 to 60h on the Europe to Africa trans-Saharan route (Berger 1961). The swifts (Apodidae) are said to fly continuously, day and night, sleeping, eating, drinking, and mating on the wing (Lockely 1970): they only come to land when nesting or drop down when they die! The same has been said of the wandering albatross (Jameson 1958). The alpine chough (Pyrrhocorax graculus), which is reported to nest above 6.5 km (Swan 1961), not only faces hypoxia ( $PO_2 = 9 kPa$ ) but also low temperatures  $(-27 \,^{\circ}\text{C})$  and a desiccating atmosphere. In premigrating hummingbirds, body mass may increase by up to 60% with fat stores (Odum and Connell 1956; Norris et al. 1957). Carpenter et al. (1983) and Carpenter and Hixon (1988) observed that during the period before migration, the rofous hummingbirds gain weight at an average rate of 0.23 to 0.30g per day (about 8-10% of lean body mass). Brain hypoxia at altitude inhibits the central nervous stimulus for shivering thermogenesis and may set the limit for high altitude residence (Bernstein 1990). Due to the lower density and viscosity of air at altitude, though the aerodynamic drag is reduced, a bird has to do more work to generate the required thrust for level flight (Tucker 1974).

### 6.7.5.2 Functional and Structural Respiratory Adaptations for Flight in Birds

The efficiency of the lung-air sac system of birds is remarkable. Its capacity to provide the large amounts of O<sub>2</sub> needed for flight at high speeds, across huge distances, and at high altitude is exceptional by mammalian standards. Unequivocally, the main adaptive feature which enables birds to fly in hypobaric hypoxia is their unrivaled tolerance to hypocapnia (Bouverot et al. 1976; Black et al. 1978; Black and Tenney 1980). The increased ventilatory rate under strenuous activity of flight is not accompanied by an increase in the tidal volume and excessive CO<sub>2</sub> washout (Bernstein 1987). By reducing the PO<sub>2</sub> gradient between the arterial blood and that in the inhaled air (Shams and Scheid 1987) and by evoking a Bohr effect which raises the O<sub>2</sub> content of blood (Grubb et al. 1979), the hyperventilatory response during high altitude hypoxia enhances O<sub>2</sub> uptake. The champion high altitude flyer, the bar-headed goose, Anser indicus (Sect. 3.8.2) exhibits unique adaptations for coping with hypoxia. Experimentally, the goose is able to withstand hypoxia to a simulated altitude of 11 km (Black and Tenney 1980) and cerebral O<sub>2</sub> flow is not limiting (Faraci et al. 1984): up to an altitude of 6.1 km, the bird maintains normal gas exchange without hyperventilating, and at about 11 km, when the  $O_2$  concentration is only 1.4 mmoll<sup>-1</sup>, it extracts adequate amounts of it to necessitate only a minimal increase in ventilation (Black and Tenney 1980). Fedde et al. (1989) observed that muscle blood supply and  $O_2$  loading from the muscle capillaries rather than ventilation or pulmonary gas transfer are the limiting steps in the contraction of the flight muscles of the barheaded goose under hypoxia. Black and Tenney (1980) observed that the PO<sub>2</sub> in the arterial blood at a simulated altitude of 11.6 km is only 0.13 kPa less than that in the inhaled air, indicating a very high  $O_2$  extraction efficiency of the lung.

The remarkable functional efficiency of the avian lung is a product of a synergism of various structural and functional parameters and processes. The principal ones are: (1) the crosscurrent arrangement between the parabronchial air and the pulmonary venous blood (Figs. 36,41,42); (2) to a vet undetermined extent the countercurrent disposition between the air capillaries and the blood capillaries (Maina 1988a; Fig. 42); (3) a large tidal volume; (4) a large cardiac output; (5) a continuous and highly efficacious unidirectional ventilation of the parabronchial gas exchange tissue (Colacino et al. 1977; Scheid 1979); (6) a short but adequate pulmonary circulatory time (Burton and Smith 1967); and (7) superior morphometric parameters which provide a high diffusing capacity of the lung (Maina 1989a; Maina et al. 1989a). The total volume of the pulmonary system in birds (i.e., the volume of the lungs, air sacs, and pneumatic spaces) on average constitutes about 20% of the total body volume, with the value being as high as 34% in the mute swan, Cvenus olor (Duncker and Guntert 1985a,b). The volume of the entire avian respiratory system (lung + air sacs) is three to five times larger than in mammals and two times larger than in reptiles. The total volume of blood in the bird lung comprises as much as 36% of the lung volume, with 58 to 80% of it being held in the capillaries (Duncker and Guntert 1985a,b; Maina et al. 1989a). The pulmonary capillary blood volume in birds is 2.5 to 3 times greater than in the mammalian lung, where only 20% of it is found in the alveolar capillaries (Weibel 1963). The gas exchange tissue (the parenchyma) of the bird lung on average constitutes only about 46% of the lung volume (Maina et al. 1982b) while in the mammalian lung the parameter constitutes a greater proportion (about 90%) of the lung volume (Gehr et al. 1981; Maina and King 1984; Maina et al. 1991). The large surface area per unit volume of the parenchyma in the avian lung (Maina 1989a; Maina et al. 1989a) is achieved through intense subdivision of the parenchyma into the remarkably small terminal gas exchange components, the air capillaries. The air capillaries generally range in diameter from 5 to 10 µm (Maina et al. 1981; Maina 1982a,b; Duncker 1974). In comparison, the smallest alveoli in the mammalian lung are the 30-um diameter ones of the lung of the shrew (Tenney and Remmers 1963), the smallest extant mammal with the highest massspecific metabolic rate (Fons and Sicart 1976). The surface density of the bloodgas barrier (the surface area per unit volume of parenchyma) in birds ranges from  $172 \text{ mm}^2 \text{ mm}^{-3}$  in the domestic fowl (Gallus domesticus) Abdalla et al. 1982) to 389 mm<sup>2</sup> mm<sup>-3</sup> in the hummingbird, Colibri coruscans (Dubach 1981; Duncker and Guntert 1985a,b): in the mammalian lung, the values are about one tenth those of birds (Gehr et al. 1981). It should be empasized that the intense internal subdivision of the gas exchange tissue in the bird lung occurs within the confines of a proportionately much smaller lung which has a smaller parenchymal volume proportion (Maina 1989a; Maina et al. 1989a). This division is so intense that the

epithelial surface area of the air capillaries is essentially equal to that of the capillary endothelium. In the process of optimizing the gas transfer surface area, the terminal gas exchange components interdigitate closely with each other, nearly constituting mirror images. The capillary loading, i.e., the ratio of the pulmonary capillary blood volume to the respiratory surface area, a parameter which indicates the degree of exposure of the capillary blood to air, is near unity (Maina 1989a; Maina et al. 1989a). The rigidity of the avian lung (Macklem et al. 1979), of which the change in volume has been estimated not to exceed 1.4% during inspiration (Jones et al. 1985), has facilitated the intense subdivision of the bird lung, providing an extensive and thin blood-gas barrier with a high diffusing capacity for O<sub>2</sub> (Maina and King 1982a; Maina et al. 1989a). While in the mammals the compliance of the respiratory system (excepting the thoracic walls) is determined by the terminal parts of the respiratory tree (mainly the alveolar spaces), in the avian system, compliance is confined to the air sacs (Piiper and Scheid 1989). In the domestic fowl, the maximum compliance of the respiratory system was reported to be  $9.6 \text{ ml cm}^{-1} \text{ H}_2\text{O}$  (Scheid and Piiper 1969), in the duck  $30 \text{ ml cm}^{-1} \text{ H}_2\text{O}$  (Gillespie et al. 1982) and in the anesthetized pigeon 2.8 ml cm $^{-1}$ H<sub>2</sub>O (Kampe and Crawford 1973).

### 6.7.5.3 Functional Reserves of the Avian Respiratory System

The lungs of birds maintain a large functional reserve. In the pigeon, Columba livia, Bech et al. (1985) and Koteja (1986) observed that cold exposure did not result in any perceivable respiratory distress, and Brackenbury et al. (1989) and Brackenbury and Amaku (1990) noted that experimental isolation of the thoracic and abdominal air sacs, a surgical procedure which renders about 70% of the total air capacity nonfunctional, had no detrimental effect on the gas exchange efficiency of the respiratory system of the domestic fowl: the birds could still run at three times the preexercise metabolic rate. From these experiments, Brackenburry (1991) observed that "there is considerable redundancy within the lung-air sac system in terms of its ability to adapt to removal of functional capacity, both at rest and during exercise". Investigations by other researchers corroborate this conclusion. The lung O<sub>2</sub> extraction was the same in exercising Pekin ducks in both normoxic and hypoxic conditions (Kiley et al. 1985). During flight, at ambient temperatures between 12 and 22 °C, lung O<sub>2</sub> extraction remains unchanged in the crows (Bernstein and Schmidt-Nielsen 1976) and starlings (Torre-Bueno 1978). The mute swan, Cygnus olor (Bech and Johansen 1980), has a lung O<sub>2</sub> extraction factor of 33% and the ventilation-perfusion ratio is near unity (Hastings and Powell 1986). Lung O<sub>2</sub> extraction values as high as 60 to 70% have been reported in some species of birds (Brent et al. 1984; Stahel and Nicol 1988). They exceed those of 40 to 45% reported in bats by Chappell and Roverud (1990). In the bar-headed goose, A. indicus, neither ventilation nor pulmonary gas transfer were limiting in a bird experimentally exercising in a hypoxic environment (7% oxygen) (Fedde et al. 1989). Hummingbirds, e.g., Selasphorus platycerus, can tolerate additional loading up to a maximum of 29.4% (Wells 1993b): a 10% increase in load called for a 5.7% increase in mass-specific O<sub>2</sub> consumption. The rufous-tailed hummingbird, Amazilia tzacatl has been reported to lift a mass about 80% of its body mass (Marden 1987). With high operational reserves, figuratively speaking, birds have not found it necessary to refine and/or utilize some of the structural and functional parameters commonly used by other animals. For example, the respiratory rate in birds is generally lower than in mammals (Calder 1968; Lasiewski and Calder 1971; Schmidt-Nielsen 1975) and the hemoglobin concentration and O<sub>2</sub> affinity of blood on average are essentially similar in the two groups (Lutz et al. 1973, 1974; Palomeque et al. 1980). In the mute swan, Cygnus olor, the respiratory rate is as low as three times per minute (Bech and Johansen 1980). Shams and Scheid (1989) and Scheid (1990) have suggested that the great endurance of birds to hypoxia is not due to the higher efficiency of the parabronchial lung compared with the broncho-alveolar one but rather in their ability to withstand lower arterial PCO<sub>2</sub> levels than mammals. In the resting domestic fowl, the arterial PCO<sub>2</sub> was estimated to be 4.4kPa (Kawashiro and Scheid 1975) and in the exercising emu, Dromaius novaehollandiae, a value of 4kPa, which is much lower than the average one of 5.3 kPa in the mammalian lung, was measured (Jones et al. 1983). In the mute swan, a pronounced positive PCO<sub>2</sub> difference between end-tidal gas (5.6 kPa) and mixed venous blood (4.3kPa) averaging 1.3kPa was reported by Bech and Johansen (1980). Experimentally, birds have been shown to tolerate arterial PCO<sub>2</sub> below 0.9 kPa (Shams and Scheid 1987; Faraci and Fedde 1986; Scheid 1990). Even after an increase of ventilatory rate by about 78% at a simulated altitude of 6.1 km, house sparrows, Passer domesticus, do not incur respiratory alkalosis (Tucker 1968; Lutz and Schmidt-Nielsen 1977). Unlike mammals, birds can withstand the high level of arterial hypoxemia because through a yet unknown mechanism, brain blood flow is not affected by arterial hypocapnia (Grubb et al. 1977; Wolfenson et al. 1982; Faraci and Fedde 1986). Paradoxically, in some species such as pigeons and bar-headed geese, the flow increases during such circumstances (Grubb et al. 1978; Faraci et al. 1984; Pavlov et al. 1987). Grubb et al. (1979) observed that by evoking a Bohr effect, hypocaphic birds reduced arterial PCO<sub>2</sub>, raising blood O<sub>2</sub> content for any particular PO<sub>2</sub>. This illustrates that in hypoxia, a hypocaphic bird enjoys better cerebral O<sub>2</sub> delivery than a normoxic one! In the human being, a reduction of the arterial PCO<sub>2</sub> to 1.3kPa results in cerebral vasoconstriction, leading to a reduction of the flow of blood to the brain by about 50% (Wollman et al. 1968). In an apparent reassessment of their earlier view which recognized the superior design of the avian lung as a contributive factor in its exceptional efficiency (e.g., Scheid and Piiper 1987, pp. 123-124), based on small differences of the arterial PO<sub>2</sub> during gas exchange between birds and mammals, Scheid (1979, 1990) and Shams and Scheid (1989) cast doubt on the importance of the structure of the parabronchial lung (compared with the broncho-alveolar one of mammals) during exposure to extreme hypoxia. In Scheid (1990, p. 6), he expresses his sentiments as follows: "we have then to admit that we cannot decide whether the lung structure of birds has evolved out of functional needs or simply out of structural constraints with no significance for the higher efficiency". These sentiments warrant further debate. It is highly unlikely that the evolution of such an intricate and efficient system as the avian-lung air sac system could have been coincidental.

The lungs of the small and the metabolically active species of birds show distinct morphometric specializations (Maina 1993; Duncker and Guntert 1985a,b). The generally small passerine species which have a higher metabolic rate and operate at a higher body temperature (see King and Farner 1969) have lungs superior to those of the nonpasserine ones (Maina 1981b, 1982c,d, 1984). The small highly energetic violet-eared hummingbird, Colibri coruscans (Dubach 1981), has remarkably highly refined lungs while the gliding and soaring birds which expend less energy for flight, e.g., gulls (see Norberg 1985), have relatively inferior lungs (Maina and King 1982b; Maina 1987c). Among birds, hummingbirds have the highest hemoglobin content, O<sub>2</sub> carrying capacity, erythrocyte counts, and large hearts (e.g., Carpenter 1975; Johansen et al. 1987). By developing a very small body size, the hummingbirds have occupied an ecological niche used earlier only by the insects (Suarez 1992). Hummingbirds have a heart rate as high as 1300 times min<sup>-1</sup> during hovering flight (Lasiewski 1964), a wing beat frequency of up to 80 times  $s^{-1}$  (Greenewalt 1960), a heart size about twice that of a mammal of equivalent body mass (Schmidt-Nielsen 1984), and whole body circulatory time of about 1 s (Johansen 1987). During hovering, the  $O_2$  consumption is 40 ml  $O_2$  g<sup>-1</sup> per h (Lasiewski 1963a,b; Epting 1980; Bartholomew and Lighton 1986; Suarez et al. 1990). To support hovering, the most energetical mode of flight when lift is generated both at up- and downstrokes to overcome gravity (Weis-Fogh 1972; Epting 1980), the flight muscles (Pectoralis and supracoracoideus) constitute about 30% of the body mass (Hartman 1961; Suarez et al. 1990). The mitochondrial volume density (the fraction of the muscle fiber volume occupied by the mitochondria) in the flight muscles of hummingbirds is about 35% (Suarez et al. 1991). The highest mass-specific respiratory surface area of about  $90 \text{ cm}^2 \text{g}^{-1}$  has been reported in the small and highly energetic violet-eared hummingbird (Dubach 1981) and the African rock martin, Hirundo fuligula (Maina 1984). The value is substantially greater than that of  $43 \text{ cm}^2 \text{g}^{-1}$  in the shrews, Crocidura flavescens and Sorex sp. (Gehr et al. 1980). The very high value of the repiratory surface area of  $800 \text{ cm}^2 \text{g}^{-1}$  reported in a hummingbird by Stanislaus (1937) must be treated with caution. An extremely thin bloodgas barrier (harmonic mean thickness) of 0.090 um has been reported in the rock martin (Maina 1984) and the violet-eared humming bird (Dubach 1981): the thickness of the blood-gas barrier in the shrew is 0.334 µm (Gehr et al. 1981). The indolent galliform species, e.g. the domestic fowl, Gallus gallus variant domesticus (Abdalla et al. 1982), a bird which has been domesticated for over 5000 years (West and Zhou 1988) and the guinea fowl, Numida meleagris (Abdalla and Maina 1981), have low pulmonary diffusing capacities. Among birds, the lowest pulmonary morphometric values have been reported in the emu, Dromaius novaehollandiae (Maina and King 1989), a huge bird which in nature is exposed to few predators. The Humboldt penguin, Spheniscus humboldti, a good diver (Butler and Woakes 1984), has a remarkably thick blood-gas barrier (Maina and King 1987), a feature presumed to enable the lung to resist collapse under hydrostatic pressure (Welsch and Aschauer 1986).

### 6.7.5.5 The Amazing Design Similarities Between the Avian and the Insectan Respiratory Systems

The resemblance between the structure (presence of air sacs and air conduits) and function (unidirectional and continuous air flow) between the avian and insectan respiratory sytems (Sects. 6.7.5 and 6.6.1) is an astonishing example of morphological convergence (Weis-Fogh 1964b; Maina 1989b). The insects evolved at the middle of the Paleozoic era some 400 million years ago (Callahan 1972) and the birds about 150 million years ago (de Beers 1954). The congruent morphology in such evolutionary distant animals presents a historic masterpiece of convergence which occurred as the two groups of animals grappled with the challenges involved in acquiring the large amounts of O<sub>2</sub> needed for volancy. This entailed radical transformations of their ancient gas exchangers. While bats retained but highly refined the structure of the mammalian lung, the insects honed the book lungs into the exceptionally efficient tracheal system, pterodactyls (of which the design of the respiratory system is unknown), and birds modified the ancestral multicameral reptilian lung. Clearly, there is no immutable design of a gas exchanger which is a prerequisite for flight. Subject to other features like aerodynamic configuration of the body (e.g., development of wings) being met, theoretically, at a cost, the mammalian lung can be refined for flight. Except in insects with the highly efficient tracheolar system (Sect. 6.6.1), endothermy appears to be a fundamental requirement for flight. Interestingly, momentary (preflight) endothermy has evolved in many insects (e.g., Heinrich 1992). The different strategies adopted by distant animals to solve common problems of uptake and delivery of O<sub>2</sub> to the tissues during strenuous activities such as flight depend on the different resources available to them to make the necessary adjustments. Those animals which accomplished flight reaped substantial benefits which included occupation of the almost limitless aerial biotope, adoption of a more economical mode of foraging, escape from ground predators, and dispersal to diverse habitats. Such taxa have undergone remarkable adaptive radiation. In the Animal Kingdom, the insects are the most populous taxon with about 750000 species (Wigglesworth 1972). Among the terrestrial vertebrates, birds constitute the most speciose group with about 9000 species (e.g., Morony et al. 1975; Gruson 1976) compared with 4200 in mammals. Among mammals, the bats, of which there are about 1000 species (Wimsatt 1970; Yalden and Morris 1975), constitute 25% of all mammalian species. The now extinct flying reptiles which included Pteranodon, the largest flying animal that ever evolved, displayed exceptional diversity of body form (Norberg 1990).

## 6.8 The Internal Subdivision of the Lung: the Functional Implications

In the higher vertebrates, an extensive respiratory surface area per unit volume is achieved by internal partitioning of a finite volume of parenchyma, e.g., in reptiles (Figs. 92,93), mammals (Fig. 94), and in birds (Fig. 95). Among the air breathers, except for the avian lung and the insectan tracheal system where the terminal gas exchange components, i.e., the air capillaries and the tracheoles are



**Fig. 92.** Cast of the lung of the monitor lizard, *Varanus exanthmaticus*, showing the remarkable differences in the sizes of the air spaces in different regions of the lung,  $\succ$ . Unlike the lung of birds (Fig. 91), this lung presents some degree of lobulation, *x*; *t* trachea; *b* principal bronchi. *Bar* 13 mm

tubular structures, the amphibian, reptilian, and mammalian lungs are made up of numerous foam-like air bubbles surrounded by a moist surfactant-lined epithelium. Dilatation of these spaces requires energy. According to Laplace's Law, the pressure required to inflate a sphere is proportional to the surface tension and inversely proportional to the radius of a bubble. In practical terms, it requires more energy to inflate a lung with small alveoli than that with larger ones. It hence follows that while an intensely subdivided lung provides a more extensive respiratory surface area, more energy is needed to ventilate it. There must be an intrinsic compromise between the endeavor to establish an optimal pulmonary design and the cost of operating it. The low metabolism ectotherms have evolved lungs with large air spaces, mammals have developed small alveoli and an efficient ventilatory mechanism, while birds have evolved a rigid, noncompliant lung with remarkably small terminal gas exchange components. Initially, the surfactant developed in the archaic piscine lungs mainly to provide a protective continuous barrier over the epithelium (Liem 1987a). With the increasing internal complexity of the lung, the surfactant was commissioned to serve as a surface tension-reducing agent in order to prevent the collapse of the small terminal gas exchange components (Wilson and Bachofen 1982). In snakes (Ophidia), there is a tendency for a single lung to occur and in chameleons, extrapulmonary saccular extensions are characteristic of the lung (Fig. 96). The general pulmonary design in such reptiles approximates that of birds, where the gas exchange tissue (the



**Fig. 93a–d.** The respiratory surface area of the lung is increased by internal subdivisions. This process is shown in the reptilian lung. **a** Cast and **b** critical point dried preparation showing spaces, *A*, of the lung of the pancake tortoise, *Malacochersus tornieri*, formed by septation of the lung, \*. **c** Lung of a snake, *Dendropis polylepi*, showing air spaces, *f*, separated by septae, >. **d** Cast preparation of the lung of the monitor lizard, *Varanus exanthematicus*, showing the various levels of septations; *p* primary septum; *s* secondary septum; > tertiary septa; *x* terminal air spaces-definition of symbols also apply to fig. b. **a** *Bar* 300 µm; **b** 400 µm; **c** 500 µm; **d** 360 µm. (**a**, **b**, **d** Maina et al. 1989b; **c** Maina 1989e)

respiratory zone) is separated from the avascular (mechanical) region. Gas exchange tissue is located at the cranial part of the lung (Fig. 97) with the caudal saccular part ventilating it. The air passes twice through the gas exchange tissue, i.e., during inspiration and expiration, possibly enhancing  $O_2$  extraction. A



**Fig. 94.** Lung of the lesser bushbaby, *Galago senegalensis*, showing alveoli, *a*, the terminal gas exchange components.  $\succ$  interalveolar septa; \* alveolar duct; *b* blood vessel. *Bar* 133 µm. (Maina 1990c)



Fig. 95. Exchange region of the lung of the emu, *Dromaius novaehollandiae*, showing an intraparabronchial arteriole, x, giving rise to blood capillaries, c, which interdigitate with air capillaries, a; e erythrocytes. *Bar* 2.5 µm. (Maina 1994)



Fig. 96. Cast preparation of the lung of the chameleon, *Chamaeleo chameleo*, showing the notably large terminal gas exchange spaces and the size differences between the dorsal and ventral aspects of the lung,  $\succ$ . *Arrows* show the posterior saccular extensions similar to the air sacs of the lungs of birds. Some degree of lobulation of the lung, *x*, is evident. *Bar* 6 mm

similar organizational plan occurs in the lungs of the serpentine caecilians (Apoda), e.g., *Bourengerula taitanus* (Maina and Maloiy 1988). The left lung is very small and the tubular right lung is well subdivided in the cranial region with the caudal part being smooth (Figs. 98,99). Stratified septa provide mechanical support, which is necessary to avoid lung collapse. The smooth muscle and elastic tissue (Stark-Vancs et al. 1984; Goniakowska-Witalinska 1986) impart the tractability essential for pulmonary ventilation in air (Smith and Campbell 1976; Toews and MacIntyre 1978). In two genera of Salentia (Pipidae), the lung is internally supported by cartilaginous plates which are located in the 1st order of the septal walls in *Pipa pipa* (Marcus 1937) while in two Gymnophiona species, *Chthonerpeton indistinctum* and *Ichthyophis paucisulcus*, tiny aggregates of cartilage cells occur in the proximal part of the lung (Welsch 1981).

Internal partitioning of the lung increases its respiratory surface area. The intensity of the internal partitioning positively correlates with factors such as the metabolic rate, the lifestyle, and the environment in which an animal lives. While a sphere of a volume of 1 cm<sup>3</sup> has a surface area of  $4.8 \text{ cm}^2$ , 1 cm<sup>3</sup> of the lung of the shrew, *Sorex minutus*, has an alveolar surface area of  $2100 \text{ cm}^2$  (Gehr et al. 1980). In the human lung, there are about 300 million alveoli of an average diameter of  $250 \mu \text{m}$  (Weibel 1963), giving an overall surface area of nearly  $150 \text{ m}^2$  and a thickness of the blood-gas barrier of  $0.65 \mu \text{m}$  (Gehr et al. 1978). The respiratory surface area is about 100 times the total surface area of the body (Comroe 1974). Such an extensive surface area over which the respiratory media, air and



**Fig. 97**. Semimacerated double cast of the gas exchange region of the lung of the black mamba, *Dendroapis polylepis*, showing the air spaces, *f*, which radiate from the central air duct, \*; *p* dorsal aorta; > pulmonary artery. *Bar* 2 mm



**Fig. 98.** Cast of a lung of a caecilian, *Boulengerula (Afrocaecicilia) taitanus*, showing the vestigial left lung, x, and the pulmonary artery,  $\succ$ , which runs three quarters of the length of the right lung. The posterior part of the lung, s, is smooth and saccular while the gas exchange region, r, is found in the anterior part of the lung. *Bar* 5 mm

blood, are separated by a thin barrier has called for development of various lines of defense to avoid damage by toxic substances and infection by pathogens. These include presence of mucus and ciliated cells on the upper respiratory passages (Figs. 100,101) and alveolar macrophages in the dependent regions (e.g., Brain



**Fig. 99.** View of the right lung of the caecilian, *Boulengerula (Afrocaecicilia) taitanus*, showing the central air duct, *a*, and peripheral gas exchange spaces, \*. The rather simple lung is supported by two diametrically located trabeculae, *t. Bar* 115 µm. (Maina and Maloiy 1988)



Fig. 100. a, b, Epithelial lining of the trachea of the respiratory system of the lung of the black mamba, *Dendroapis polylepis*, showing ciliated cells, \*, and mucus cells, *m*. a *Bar* 2µm; b 6µm

1985; Fig. 102). In spite of the fact that the respiratory mucosa presents the largest interface between the external environment and the internal milieu, in absence of pulmonary disease, below the larynx, the respiratory system is virtually sterile (Skerret 1994; Agostini et al. 1995). In the amphibians (Welsch 1983; Maina 1989d; Goniakowska-Witalinska 1995) and probably in ectothems in general, lung macrophages are rare. The number of pulmonary macrophages appears to



Fig. 101. Epithelial lining of the air passages of the lung of the vervet monkey, *Cercopithecus aethiops:* \* (*main figure*) ciliated cells;  $\star$  mucus cells; *m* mucus debri; > secretory pore of a mucus cell. *Top inset* Clara cells, *c. Bottom inset* a mucus cell with secretory granules, *s*, next to a ciliated cell, \*. *Bar* (*main fig.*) 3.6 µm; *top inset* 0.5 µm; *bottom inset* 2 µm. (Maina 1988b)

correlate with the metabolic capacities of animals and the environment in which they live.

# 6.9 The Surfactant: a Versatile Surface Lining of the Gas Exchangers

While the insectan trachea, which are supported by helical cuticular taenidia, and the fish gills, whose filaments are physically separated by water, have little need for stabilization against surface forces, practically all evolved air-breathing



**Fig. 102.** Alveolar macrophages (*m*). *Main figure* (rat lung), \* a particle about to be ingested;  $\succ$  filopodia;  $\star$  interalveolar pore; *c* blood capillary; *e* erythrocytes. The macrophage shown in the *inset* is from a lung of a tree frog, *Chiromantis petersi*:  $\succ$  filopodia. *Bar* 0.6 µm; *inset* 6 µm. (*Inset* Maina 1989d) organs require presence of an active surface film for stability (Stratton 1984). The process of increasing surface area by subdivision of the lung engenders an increase in the surface tension in the terminal respiratory units. The energy necessary to dilate the gas exchanger during inspiration (an important factor in the cost of breathing) rises. High surface tension affects fluid balance across the blood-gas barrier which coupled with the capillary blood pressure enhances filtration of fluids from the capillaries. The surfactant, a phospholipid material (dipalmitoylphosphatidylcholine) which lines the respiratory surfaces in all gas exchangers evolutionary derived from the lung, reduces the deleterious effects of surface tension at the air-water interface by spreading on the respiratory surface as a monomolecular phospholipid film (Cochrane and Revak 1991). By lowering the surface tension, the surfactant, at least in the mammalian lungs, stabilizes the extremely small terminal gas exchange spaces. In human premature newborn, inadequacy of the surfactant due to lack of type II cells leads to the respiratory distress syndrome which may be fatal. In the vertebrate lungs, the surfactant serves to smooth the alveolar air-liquid interface (Bastacky et al. 1995) and to promote the displacement of deposited particles into the aqueous subphase where they are cleared by the pulmonary macrophages and the mucociliary carpet (Schürch et al. 1990).

In various forms and quantities, the surfactant is widely distributed in the vertebrate lungs (e.g., Clements et al. 1970; Hughes 1973; Pattle 1976; Dierichs and Dosche 1982; Hills 1988; McGregor et al. 1993). Lamellar bodies have been described in the lungs of the lungfishes (Hughes and Weibel 1976; Maina 1987a), those of fish such as bichirs, Polypterus delhezi and P. ornatipinnis (Zaccone et al. 1989), the gar-fish (Lepisosteus osseous) and the bowfin, Amia calva (Hughes 1973), in the epithelial lining of the fish swim bladder (Copeland 1969; Brooks 1970) and in the distal (respiratory) part of the intestine of the pond roach, Misgumus fossilis (Jasinski 1973). In the lungs of the higher vertebrates, it is secreted by the granular pneumocytes, the type II cells (Fig. 103). The composition of the surfactant and probably its function (at least in the human lung) may be regulated by the O<sub>2</sub> tension in the air spaces (Acarregui et al. 1995). The amphibian pulmonary type surfactant may or may not be discharged into the air space as tubular myelin type (Goniakowska-Witalinska 1980a,b; Bell and Stark-Vancs 1983), is mainly composed of phosphatidylcholine (e.g., Vergara and Hughes 1981), occurs in large quantities (Clements et al. 1970), and is not particularly surface-active like that of the higher vertebrates (e.g., Hughes and Vergara 1978; Daniels et al. 1989). The role of the surfactant, especially in the amphibian lung, is enigmatic, as even the most elaborate lungs have wide air cells which are not susceptible to collapse. Pattle et al. (1977) showed that in the lung of Triturus vulgaris, the surfactant only partly reduces the surface tension to 0.2 mNm<sup>-1</sup> compared with the more efficient mammalian one which reduces it to 0.1 mN m<sup>-1</sup> or less. In addition to the better-known roles of preventing atelectasis, edema, reduction of respiratory work, and stabilization of the terminal gas exchange components (e.g., Fishman 1972; Farrell 1982b), the surfactant plays several roles such as prevention of transendothelial transudation of substances across the blood-gas barrier, immune suppression, chemotaxis of macrophages (e.g., Daniels et al. 1993), and antioxidant function (Brooks 1970). The



**Fig. 103.** A Type II cell from the lung of the tree frog, *Chiromantis petersi*, showing mitochondria, *m*, Golgi bodies, *g*, and osmiophilic lamellated bodies, *o*. > microvilli. *Inset* a type II cell from the lung of a caecilian, *Boulengerula (Afrocaecicilia) taitanus*, showing secretion of the surfactant onto the surface of the lung. *Bar* 0.7  $\mu$ m; *inset* 0.3  $\mu$ m. (Main figure Maina 1989e; *inset* Maina and Maloiy 1988)

multifunctionality of the surfactant may help explain why the simple lungs, e.g., fish and amphibian lungs which have large air cells, possess the lining. In such animals, surface tension reduction may not be the primary role of the lining. In agamid lizard, *Ctenophorus nuchalis* (Daniels et al. 1990), the salamander,

Amphiuma tridactylum (Stark-Vancs et al. 1984), and Siren intermedia (Guimond and Hutchison 1976; Martin and Hutchison 1979), the surfactant-like lipids are envisaged to act as an antiglue preventing epithelial adhesion after near total pulmonary collapse during apnea (Frappell and Daniels 1991; Daniels et al. 1993; McGregor et al. 1993). In aquatic amphibians this occurs during a dive when the hydrostatic pressure may lead to virtual pulmonary collapse (Daniels et al. 1994) and in mammals in cases of alveolar collapse at different points in a ventilatory cycle (Hills 1971; Weibel et al. 1973; Sanderson et al. 1976). In the Weddell seal, Leptonychotes weddelli, the alveoli are totally collapsed at depths of 30 to 35 m (Falke et al. 1985) but the lung of the dolphin, Tursiops truncatus, does not collapse, at least up to a depth of 70 m (Ridgway and Howard 1979). In the agamid lizard, the lung contains 70 times more surfactant per respiratory surface area than those of a comparable-sized mammal (Daniels et al. 1989). The antiglue role of the surfactant may be important in the lungs of those animals which lack well-configured intrapulmonary conducting airways and a diaphragm (e.g., amphibians, reptiles, and birds), where the lung is mechanically more susceptible to the displacements of the organs in the coelomic cavity (Daniels et al. 1994). The presence of the surfactant on the air capillaries of the bird lung (Fig. 40a), structures which are somewhat fixed in size, is most intriguing. While it may simply be a phylogenetic carryover from a primordial reptilian lung, especially the crocodile one (e.g., Perry 1992b), it may as well have evolved as a means of curtailing transendothelial filtration of fluids from the blood capillaries in order to prevent flooding of the air capillaries. It is interesting that there is no surfactant either in the fish gills or in the accessory respiratory organs of the bimodal breathing fish. The surfactant appears to be an innovation of the lung and its most immediate derivatives. As evident in most facets of respiration, exceptions pervade the field. Some air-breathing fish such as the gourami, which construct bubble nests, secrete a surface-active (surpellic) substance to reduce surface tension at the liquidair interface (Phleger and Saunders 1978).

# 6.10 Nonrespiratory Roles of the Gas Exchangers

## 6.10.1 Trophic, Sensory, and Locomotory Functions

Owing to the fact that the respiratory system shares a common passage, the pharynx, with the digestive system, activities such as change in posture, eating, swallowing, drinking, and rumination affect ventilatory activity, but only momentarily. This results in moderate but noneventful asphyxia. In some animals, however, predominant activities such as feeding (in filter feeders), husbandry of young (e.g., in the mouth-brooding fish), and phonation greatly conflict with the gas transfer function. In some cases, respiration may be relegated in preference for a nonrespiratory role. Compared with *Chaetopterus variopedatus*, a filter-feeding marine annelid which ventilates its burrow continuously, the lugworm, *Arenicola marina*, a deposit feeder which swallows the detritus and regurgitates sand after extracting organic matter (Jacobsen 1967) has a ventilatory rate (at

15 °C) about ten times lower (Toulmond 1991): the O<sub>2</sub> extraction coefficient in C. variopedatus is 30%, while in A. marina the value is 80%. Debate has persisted (e.g., Willmer 1970) as to whether the initial task of the archetype respiratory organ, the gill, was respiratory or trophic. Filter feeders like sponges, ascidians, and lamellibranchs move water over their body surfaces, establishing a PO<sub>2</sub> gradient between the surrounding water and the body cells. In the process, microscopic unicellular organisms are entrapped (Hazelhoff 1939; Jorgensen 1955). Hazelhoff (1939) observed that at the same PO<sub>2</sub> in water, the O<sub>2</sub> extraction coefficient in the filter feeders is lower than that of the nonfilter feeders. This is presumably due to the necessary high flow rate of water needed to procure an adequate quantity of food. Jorgensen (1975) suggested that the rate of water flow over the gills is well synchronized with the O<sub>2</sub> and food concentration in a particular environment. In the polychaete worms, the irrigation of the branchial crown is 70 ml  $g^{-1}h^{-1}$  while that of the tube is only 12 ml  $g^{-1}h^{-1}$  (Dales 1961): O<sub>2</sub> utilization by the crown is about two times that of the skin. The marine polychaete, Chaetopterus variopedatus, a suspension feeder, continuously ventilates its burrow by body movements (Brown 1975, 1977), extracting O<sub>2</sub> and nutrients from the water current (McGinitie 1939). In organisms which use the same site for feeding and respiration, synchronization between gas transfer and feeding is most critical. The larva of the insect Chironomus plumosus exhibits cyclic behavior which changes from filter feeding to rest and from rest to respiration (water ventilation). Normally, the rest intervals are brief, with the two other episodes being of about the same duration: ventilation increases in a hypoxic condition at the expense of the feeding interval, the limiting external PO<sub>2</sub> for filter feeding being 2kPa (Walshe 1950; Ewer 1959; Rubenstein and Koehl 1977). In aquatic animals such as the sea urchins, the respiratory organs (podia or tube feet) are utilized both for locomotion and sensory perception - in addition to their gas exchange role (Steen 1965). In the sabellid polychaete, S. spallanzani, a 60% drop in O<sub>2</sub> uptake was recorded by Fox (1938) after amputation of the branchial crown, a very active ciliary feeding structure and evidently an important respiratory one. In the bivalve mollusks, except for some Protobranchia, the ctenidia are greatly hypertrophied for their added role in extraction and transportation of food (e.g., Atkins 1936). In Pholas dactylus, particulate food is filtered from the large volumes of water (about 1001) which pass through them per day (Knight 1984). The current of water created is far in excess of that needed for respiratory purposes in these rather sluggish, often sessile animals. The cephalopods, a toxon which traces its evolutionary lineage some 500 million years back, have conservatively utilized jet propulsion as the basic locomotor mechanism (e.g., Teichert 1988; Chamberlain 1990; O'Dor and Webber 1991). The ventilatory currents serve a locomotory role (Yonge 1947; Wells 1952). Oxygen extraction in cephalopods is variable, depending on the degree to which the animal uses jet propulsion for locomotion. In *Nautilus*, when in motion,  $O_2$  extraction drops to as low as 4% and while at rest it is on average 5 to 10% (Wells and Wells 1985). Sepia and Octopus, which can uncouple locomotion from respiration, have O<sub>2</sub> extraction rates which are 35 to 45% or even much higher (O'Dor and Webber 1991). The volume of the water across the gills of swimming squids is so large that  $O_2$  extraction is only 5 to 10% and on exposure to hypoxia and extreme exercise rises to a maximum of 17% (Wells et al. 1988; Shadwick et al. 1990). The most aerobic muscle fibers in squids, which are located in the inner and outer surfaces of the mantle, contain as much as 50% mitochondria by volume (Bone et al. 1981; Mommsen et al. 1981), a value which compares with that of the insect flight muscle (Elder 1975; Ready 1983).

As a general rule, multifunctionality of a gas exchanger is an indication of primitiveness, especially where the roles greatly conflict, e.g., in designs for such processes as feeding, sensation, and respiration. Such exchangers must inherently be designed to accommodate remarkably different needs which cannot be compromised. Filter feeding occurs in fish such as the paddle fish, Polyodon spathula, anchovies, and menhaden (Durbin et al. 1981; Lazzaro 1987; James and Probyn 1989; Burggren and Bemis 1992) which utilize ram ventilation as they swim continuously. In the sabellid, Schizobranchia insignis, extirpation of the crown resulted in a 75% drop in  $O_2$  consumption (Vo<sub>2</sub>) while those worms which retained the crown but were prohibited from extending it into the water showed a 60% drop in Vo<sub>2</sub> (Dales 1961). The importance of the ciliary organs in respiratory function in polchaetes is remarkable. Assuming that the surgical procedure does not greatly interfere with the metabolic level, the total O<sub>2</sub> uptake by the two halves of a transversely bisected S. pavolina is not significantly lower than that of the whole worm but in *Myxicola infundibulum*, a dramatic drop in total O, uptake occurs on bisection. The posterior body part gives lower values compared with those of S. pavolina (Wells 1952). The differences show that in S. pavolina, which irrigates its tube through body movements, the ciliary crown does not contribute to the O<sub>2</sub> requirements of the rest of the well-ventilated body. However, in Myxicola, where the tube is not irrigated, dependence on the crown is much greater. The role of the respiratory organs in phonation and the physiological consequences on respiration and acid-base status of blood has been outlined by Dejours et al. (1967) and Bouhuys (1974), while the thermoregulatory aspects have been reported in mammals by Cunningham and O'Riordan (1957) and in birds by Calder and Schmidt-Nielsen (1968) and Lasiewski (1972).

### 6.10.2 Metabolic and Pharmacokinetic Functions of the Lung

The lung is best known for its role as the oxygenator of the blood and the provider of  $O_2$  to the body tissues (e.g., West 1974; Weibel 1984a). This role is axiomatic from its design, which incorporates a large surface area and thin blood-gas barrier. The facts that the lung is mechanically ventilated with air by the ribs, perfused with blood by the heart, and gas exchange occurs by the passive process of diffusion is suggestive of an inert organ. For these reasons, the biochemical, pharmacological, and metabolic roles of the lungs are not as well recognized as the respiratory one (Heinemann and Fishman 1969; Vane 1969; Tierney 1974). This is despite the fact that as far back as over 70 years ago, Starling and Verney (1925) observed that isolated kidneys perfused with defibrinated blood quickly caused vasoconstriction but if the blood was passed through a heart-lung pump preparation, the vasoconstrictor response was absent. They concluded that the blood was "detoxified" in the lungs. Twenty five years later, Rappaport et al. (1948) identified the serum vasoconstrictor substance as 5-hydroxytryptamine (5-HT) and demonstrated that isolated lung preparations inactivated 5-HT. The processes of substrate handling by the lung was well documented by Eiseman et al. (1964) and the subject reviewed by, among others, Bakhle (1975). Though the complete spectrum of the roles the lung plays in physiological and pathological states remains largely unappreciated, it is now well recognized that the organ performs various important metabolic tasks. It is said to be "a peculiar immunological organ which can operate independently and synchronously with the general immune apparatus" (Agostini et al. 1995). Certain tasks are as important, if not more important, than the respiratory one. Bakhle (1975) asserts that "instead of referring to pharmacokinetics as one of the functions of the lung, we ought to refer to gas exchange as one of the nonpharmacokinetic functions of the lung". In some instances, the lung performs the nonrespiratory functions more efficiently than other organs which are considered more specialized (Table 27). The lung is an important source of biologically active agents, constitutes an important organ for defense, and, paradoxically, may initiate, actively participate in, or even sustain pathological processes such as allergy (Nicolet et al. 1975), emphysema (Kimbel and Weinbaum 1975), shock (Bleyl and Büsing 1971), pulmonary cancer (Cohen 1975), pulmonary necrosis (Reid et al. 1973), and hyaline membrane disease (Avery and Mead 1959). To carry out the roles which range from defense, clearance of mucus, and electrolyte transport, to the pharmacological ones, the lung has a large number of different cell types (e.g., Breeze and Wheeldon 1977; Andrews 1981; Weibel 1984b; Welsh 1987).

Anatomically, the lung is strategically located in the general circulation dividing the venous and the arterial circulations. Owing to its vast vascular transit distance, it contains the highest volume of blood per unit time of any other organ in the body. During the transit of blood through the lung, a fine balance between providing the essential metabolic nutriments to the tissue cells and limiting transport of salts and water into and out of vascular space must be maintained (Welsh 1987). More than in other tissues, the lung must make allowance for unhindered passage of some hormones and biologically active substances from the arterial to the venous side of circulation (Ryan and Ryan 1975). The adjust-

Substrate	Lung	Liver	
	nmol HCOH formed $mg^{-1}$ microsomal protein min <sup>-1</sup>	nmol HCOH formed mg <sup>-1</sup> microsomal protein min <sup>-1</sup>	
Aminopyrine	5.2	15	
Ethylmorphine	2.2	3.5	
N-Methyl-alanine	3.0	3.0	
N-Methyl-p-chloroaniline	7.2	4.8	
(+) – Benzphetamine	8.8	4.7	

 Table 27. N-demethylation rate of several drugs in rabbit microsomes prepared either from lung or liver. (Remmer 1975)

ments which take place after a substrate passes through the pulmonary circulation can be categorized into intrinsic ones, i.e., those which affect the activity of the substrate itself and the extrinsic ones, i.e., those which entail in situ production of biologically active factors (Bakhle 1975): the latter constitutes an endocrine function. The active factors are handled differently by the endothelial cells of the pulmonary vasculature particularly at the capillary level (e.g., Ryan and Rvan 1975). The endothelial cells of the lung have abundant caveolae cellulares (micropinocytotic vesicles; Figs. 29b,40a), most of which are in direct contact with the circulating blood and are irrefutably the sites of degradation, transformation, interaction, and biosynthesis of the macromolecules for which the lung has affinity and transendothelial transfer of the same to the alveolar surface (e.g., Bignon 1975). The lung has a very high pharmacokinetic specificity for the circulating biologically active enzymatic complexes. For example, the pulmonary circulation acts as a filter, letting adrenaline or histamine pass through but inactivating 5-HT, bradykinin, and synthesizing angiotensin II from angiotensin I (e.g., Bakhle 1975; Ryan and Ryan 1975; Nossaman et al. 1994) (Tables 28,29,30). A well-known example of the endocrine role of the lung is that of appearance of histamine in the lung perfusate when an antigen is passed through the pulmonary circulation of the isolated lungs from sensitized guinea pig (Bartosch et al. 1933).

tryptamine (5-HT). (Junod 1975)				
Species	Concentration of 5-HT	Extraction (%)		
Man	Bolus injection of 7.6 $\mu$ g ml <sup>-1</sup>			
Rat	$5-100 \mathrm{ng}\mathrm{ml}^{-1}$	92		
	$0.02 \mu g \mathrm{ml}^{-1}$	20-58		
Dog	$10-20\mu gml^{-1}$	80-98		
Rabbit	$250 \mathrm{ng}\mathrm{ml}^{-1}$	52.6		
	$500 \mathrm{ng}\mathrm{ml}^{-1}$	56		
Guinea pig	$1 - 1000 \mathrm{ng}\mathrm{ml}^{-1}$	48-60		

Table 28. Pulmonary extraction of circulating 5-hydroxy-tryptamine (5-HT). (Junod 1975)

Table 29. Pulmonary inactivation of bradykinin. (Friedli et al. 1973)

Group	Age of Gestation (Days)	Inactivation (% mean and standard deviation)
Ewes n = 5	Adult	93.4 ± SD 1.20
Lambs $(n = 5)$	Immature (12 Days)	$68.0 \pm SD 5.28$
Fetal lambs at term $(n = 4)$	Term (144 Days)	46.5 ± SD 4.80
Premature fetal lambs $(n = 4)$	Immature (118 Days)	-

It is now well known that different pharmacologically active substances are released during anaphylaxis and that other stimuli are capable of causing the release of active substances from unsensitized lungs (e.g., Piper 1975). The lung secretes numerous active biochemical substances such as histamine (Bakhle 1975), slow reacting substance of anaphylaxis (Brocklehurst 1960), dipeptidyl carboxypeptidase (Roth and Depierre 1975), bradykininase, angiotensin I converting factor (Bakhle 1975), and prostaglandins (e.g., Änggård 1975; Piper 1973) in response to different stimuli which include anaphylaxis, physical deformation, biogenic amines, and peptides. Other than in anaphylaxis, the relevance of these experimental findings on isolated organs of laboratory animals to in vivo situations and particularly extrapolation to the human being is still a matter of speculation (Berry et al. 1971). More recently, however, the use of cardiopulmonary bypass as an investigative procedure (e.g., Parker et al. 1975) has corroborated some of the earlier assumptions. It is now well established that the metabolic and the pharmacokinetic functions of the lung are important for some crucial processes such as blood coagulation and inactivation and destruction of hormones and other biologically active factors. It is plausible that these processes may serve as an alternative safety system should other integral organs like the liver, or the kidneys be rendered ineffective. The nonrespiratory functions of the

Class of substance	Activation	Inactivation	No change in activity
Biogenic amines	_	5-Hydroxytryptamine Tryptamine Noradrenaline Acetylcholine	α-Methyl 5- hydroxytryptamine Adrenaline Isoprenaline Histamine
_	Angiotensin I Reno-active peptide	Bradykinin Reno-active peptide	Angiotensin Eledoisin Physalaemine Oxytocin Vasopressin
Prostaglandins	Arachidonic acid Other PG precursors	Prostaglandins of E and F type	Prostaglandins
Nucleotides	-	Adenosine monophosphate (AMP) Adenosine triphosphate (ATP)	-
Basic drugs	-	By absorption: Imipramine chlorocyclizine aphetamine By metabolism: methadone	-

**Table 30.** Effect of passage through the pulmonary circulation of biological activity. (Bakhle1975)

lung are affected by factors such as age (Melmon et al. 1968), pregnancy (Bedwani and Morley 1974), and exposure to gaseous anesthetics (Naito and Gillis 1973).

The diversity of the functions carried out by the lung compares with the wellestablished multifunctional ones of the fish gills which, in addition to respiration, include osmoregulation, acid-base balance, nitrogen excretion, and modification and conditioning of plasma hormones before perfusion of the systemic circuit (e.g., Olson 1996). The lung is also known to greatly modify biological activity of many substances endogenous and exogenous to the pulmonary circulation (Alabaster and Bakhle 1972; Remmer 1975). Smooth muscle contractility in the femoral arterial blood of cats was much less (after infusion of bradykinin) when the chemical was infused into the right ventricle than when the infusion was made into the aorta just distal to the aortic valve (Alabaster and Bakhle 1972; Levine et al. 1973): an inactivation factor of bradykinin ranging from 75 to 99.9% during the passage from the right ventricle to the heart was determined. The degradation of bradykinin in the lung was shown by Ryan et al. (1970) to be very fast. A peptidase from the pig lung has been shown to split the COOH-terminal dipeptide from bradykinin (Igic et al. 1972; Dorer et al. 1974). The ability of the lungs to inactivate bradykinin, a hypotensive factor, while converting (activating) angiotensin I to angiotensin II, a potent hypertensive agent (Nossaman et al. 1994; Table 30) indicates the central role of the lung in blood pressure regulation. Thomas and Vane (1967) showed that 98% of the biological activity of 5-HT was lost during its passage through the pulmonary circulation (Table 28). The disappearance of 5-HT from the circulation results from its uptake by the lung tissue which is followed by fast oxidative deamination yielding 5-OH indole acetic acid (Junod 1975). The extraction process seems to be: (1) saturable when the concentration of the substrate is increased (Junod 1972; Iwasawa et al. 1973); (2) limited by transport rather than metabolism (Junod 1972); and (3) an active temperaturedependent process (Junod 1975). 5-HT, which is exclusively found in endothelial cells of the pulmonary circulation from large vessels to capillaries (Strum and Junod 1972; Iwasawa et al. 1973) and norepinephrine which, compared with 5-HT, is extracted to a lesser extent (40%) (Hughes et al. 1969b; Iwasawa and Gillis 1974) are the only naturally occurring amines affected to a significant extent by the lung and show similar uptake behavior. Histamine, dopamine, 1-dopa, and norepinephrine are not significantly retained or degraded (Vane 1969). The level of  $P_{450}$  in the sheep lung is 0.092 nmol per mg protein (Burns et al. 1975). Besides other possible functions,  $P_{450}$  has been implicated with  $O_2$  transfer in the lung (Burns and Gurtner 1973; Burns et al. 1975) and the placenta (Burns and Gurtner 1973; Gurtner et al. 1982).

In summary, the central position of the lung in the circulatory system and the fact that among all organs in the body it contains the largest number of endothelial cells (Junod 1975; Ryan and Ryan 1975) as well as the greatest diversity of constituent cells (Ballard and Ballard 1974), of which as many as 20 types are in place as early as the 14th week of gestation (Avery 1968), to a large extent enables the pulmonary system to protect itself as well as the arterial circulation from the influences of locally produced and exogenous biologically active molecular factors. The large number of different cells provides the necessary receptor sites over which the different functions of the lung are carried out (Conolly and Greenacre 1975). The pulmonary endothelial cells are known to be the active sites of metabolic and biosynthetic functions of the lung (Ryan et al. 1968; Smith and Ryan 1970). It is possible that the number of endothelial cells exposed to the circulating blood can be varied during different states (Ryan and Ryan 1975). Prostaglandins are released from isolated perfused lungs of several species by diverse stimuli which may be immunological, chemical, or mechanical (Saeed and Roy 1972; Änggård 1975), the possible sites of production being the perivascular cells, type II alveolar cells, and macrophages (Smith and Ryan 1973a,b). Under experimental and pathological conditions such as septicemia, improper transfusion of blood, trauma, or poisoning, the lung is highly susceptible to disseminated extraneous intravascular coagulation. Blockage of blood flow may be a secondary cause of death.

## 6.11 The Implications of Liquid Breathing in Air Breathers

The commonality of the mechanisms of gas exchange by diffusion across the tissue barriers and the ventilatory and circulatory adjustments evoked to meet the metabolic needs independent of the milieu an animal lives in raises a fundamental theoretical and practical question as to whether air breathers can survive by breathing water or any other liquid, especially if such fluid is charged with O<sub>2</sub> (Kylstra 1968, 1969). This would hopefully reveal why the air- and waterbreathing organs are structurally so different in spite of the fact that their basic functions are the same. It is now well recognized that in utero, the lungs are not collapsed but are distended by aspirated and secreted fluid measuring at about 30 ml per kg in the human fetal lung. Presence of an appropriate volume of liquid is thought to be necessary for normal intrauterine lung growth and development (Moessinger et al. 1990). Immediately before birth (e.g., Dickson et al. 1986), during labor, and/or soon after (e.g., Brown et al. 1983), pulmonary filtration ceases (as the respiratory epithelium stops secreting Cl<sup>-</sup> ions and starts to absorb Na<sup>+</sup> from the lumen of the lung) (e.g., Bland 1990; Chapman et al. 1994). The two process are regulated by levels of circulating catecholamines (e.g., Mortola 1987). Efficient removal of liquid from the air spaces during and after birth is vital for normal switching from placental to pulmonary gas exchange. Liquid is thus not an entirely alien factor to the respiratory surfaces of aerial gas exchangers, at least during the early stages of development. It is of profound interest to know to what extent the structural and functional modifications of the aerial gas exchangers, the lungs, have lost the capacity of utilizing the primordial respiratory medium - water.

In an applied sense, liquid breathing, if and when perfected, will overcome the serious obstacles which human beings would experience for successful survival in deep subaquatic habitats and in the outer space. If a diver breathes a suitable liquid enriched with  $O_2$  instead of air, since the liquid in the lungs would resist the external pressure without significantly changing in volume, it would be possible to descend to depths and ascend to the surface rapidly without the risk of decom-

pression sickness (bends) (Paulev 1965). The ultimate obstacle in the conquest of speed apparently will not be technology but the fragility of the human body to withstand the stresses and strains that have to be endured during such states. Only a few people have "gone supersonic" without the aid of an air craft and survived (Cameron 1990). In future space travels enormous accelerations will be necessary to escape from the gravitational pulls of the much larger celestial bodies. Though constant displacement (speed), irrespective of how fast, produces little stress on the human body, accelerations, decelerations, and sudden maneuvers exert strong forces on the body. Such stresses and strains may not be tolerable to the delicate parts of the human body such as the lungs (e.g., Bullard 1972). The destructive effect could be minimized or even totally eliminated if a whole animal is externally supported by a respirable liquid of the same specific gravity as the body fluids (instead of air). Furthermore, fluid-filled lungs should be able to bear much greater stresses and strains (e.g., Margaria et al. 1958). Absence of gas in the body would equalize the density throughout the body, preventing relative inertial movements of the heart, lungs and other visceral organs while pulmonary arterio-venous shunting would be prevented (Sass et al. 1972). Experimentally, dependent pulmonary atelectasis, arterio-venous shunting and downward displacement of the heart brought about by gravitational-inertial force exposure were prevented in dogs breathing oxygenated liquid fluorocarbon in a whole-body immersed respirator (Sass et al. 1972). Liquid lavage has been used successfully in treating pulmonary conditions such as asthma, bronchiectasis, and mucoviscidosis (Kylstra et al. 1971), in treatment of acute lung injury (e.g., Richman et al. 1993), and management of respiratory failure (Rogers et al. 1972). The pioneering experimentation on breathing O<sub>2</sub>-saturated liquids was carried out by, among others, Kylstra and Tissing (1963), Clark and Gollan (1966) and Kylstra et al. (1966).

Due to the remarkably different physicochemical properties of water and air (Tables 4,9), water breathing poses distinct problems to an air breather. The primary limitations are that: (1) water under normal atmospheric pressure contains too little dissolved O<sub>2</sub>, and (2) the differences in the ionic composition of water compared with that of blood upsets ionic equilibrium with the body fluids (e.g., Lowe et al. 1979). Owing to its greater viscosity, the maximum flow rate of water from isolated dog lungs is much lower than the maximum expiratory flow rate of air at equal lung volumes (Leith and Mead 1966). Furthermore, the driving pressure affordable for maximum expiratory flow, which is limited by the lung's static recoil pressure in liquid filled lungs (Mead et al. 1967) is less than in an airfilled one (Wood and Bryan 1971). Aquatic organisms have evolved within the physical constraints of their relatively O<sub>2</sub>-deficient medium. Most of them possess highly specialized gas exchange systems (gills) which promote extraction of the available O2. When saline is inhaled into a lung, it physically destroys the delicate terminal gas exchange components (e.g., Curtis et al. 1993), dissolves and mechanically displaces the surfactant (Lewis et al. 1993), osmotically interferes with the composition of the body fluids, causes pathological changes such as interstitial edema, produces intrapulmonary froth and atelectasis (Blenkarn and Hayes 1970), causes loss of macrophages and rupture of the alveolar cell and basement membrane (Huber and Finley 1965), and upsets the integrity of the blood-gas

barrier (Reidbord 1967). Direct interference with the surfactant production reduces or even totally eliminates surface tension. This lowers the driving pressure available for maximum expiratory flow to a value no greater than that produced by the elastic recoil of the lung tissue, which is only a small portion of the total static recoil pressure available during air breathing (Kylstra and Schoenfisch 1972). Presence of liquid in the airways increases airway constriction (Yager et al. 1989). With the dissolving of the surfactant and other alveolar hypophase liquid materials, the viscosity and density of the saline increases tremendously, elevating the cost of the ventilatory effort (Blenkarn and Hayes 1970). Addition of exogenous surfactant to liquid ventilated lungs reduces maximal inflatory pressures (Tarczy et al. 1996).

By using isotonic solution supercharged with O<sub>2</sub> to a pressure equivalent to that of air at sea level, it has been possible to keep rats (Pegg et al. 1963), mice (Kylstra 1962), and dogs (Kylstra et al. 1966) alive breathing water for hours. The eventual survival times have depended on factors such as the experimental temperature and the chemical composition of the fluid used. Death during liquid ventilation in a normothermic cat (Clark and Gollan 1966) and the dog (Kylstra et al. 1966) did not result from anoxia but from difficulties in removing CO<sub>2</sub> at the necessary rate. This limitation resulted in acidosis, thought to be secondary to a high arterial PCO<sub>2</sub> (e.g., Shaffer et al. 1976). It was not until mechanically assisted liquid breathing systems were designed (e.g., Saga et al. 1973; Shaffer and Moskowitz 1974) and the effect of CO<sub>2</sub> overloading minimized that the significance of acidosis was fully appreciated as a particular metabolic complication during liquid breathing: unequivocally,  $O_2$  delivery to the tissues during liquid breathing is more than adequate (Shaffer and Moskowitz 1974; Shaffer et al. 1976). The exhaled breath of a mammal on average contains 50 ml of  $CO_2 l^{-1}$  while at the same temperature and  $PCO_2$ , a solution with the same salt concentration as blood contains only about  $30 \text{ ml l}^{-1}$ . This indicates that to remove as much CO<sub>2</sub> through the water as that eliminated via the air, a water-breathing mammal would have to exhale about twice as large a volume of water as of air (Kylstra and Tissing 1963). This is practically made difficult by the narrowness of the bronchial passages and the greater viscosity of water which requires about 36 times more power to move than air. In a state of laminar flow, a process which appeared to occur in saline breathing dogs (Kylstra et al. 1966), a liquid breather would have to expend 60 times the energy required in breathing air. The maximum expiratory flow of a liquid has been estimated to be 40 to 100 times lower than air ventilation (Leith and Mead 1966). Liquid-breathing mice died as a result of exhaustion of the respiratory muscles and accumulation of CO<sub>2</sub> to toxic levels (Kylstra 1962), a complication which has been reported in other animals (e.g., Clark and Gollan 1966; Modell et al. 1970).

To demonstrate the inadequacy of  $CO_2$  elimination during liquid breathing, the survival time of mice was increased to 18h by addition of tris(hydroxymethyl) amino methane (a substance that minimizes the harmful effects of  $CO_2$  accumulation) to the experimental solution (Kylstra 1962; Nahas 1962). An anesthetized dog cooled to 32 °C was kept alive for 24 min and resuscitated, and normothermic dogs were able to breath liquid for 45 min, 40% of them surviving the exposure well (Kylstra et al. 1966; Mathews et al. 1978): during the experiment, the blood pressure was lower than normal, the heart and respiratory rates were below normal but regular, the arterial blood was fully saturated with  $O_2$  (arterial PO<sub>2</sub>, 6.8 to 39kPa) but the  $CO_2$  content of the arterial blood increased gradually from 5.7 to 10.7 kPa, indicating that the dog's respiratory efforts were not enough to eliminate adequate amounts of  $CO_2$  from the body. West et al. (1965) and Kylstra et al. (1966) observed that the overall pulmonary gas exchange in liquidfilled lungs is diffusion-limited and remarkably large gas tensional gradients occur within the liquid-filled terminal gas exchange components due to the slow rate of diffusion in water. In dogs subjected to lavage with hyperbarically oxygenated saline (PO<sub>2</sub> in the inspired air, 387 kPa), arterial oxygenation was 32 kPa after 15 min, but a severe respiratory acidosis (arterial PCO<sub>2</sub>, 9.3 kPa; pH, 7.2) developed.

## 6.12 Physical Gill and the Plastron: a Unique Underwater Respiratory Strategy

The insectan tracheal system (Sect. 6.6.1) evolved fundamentally as a means for air breathing and hence as an adaptation for terrestrial habitation. A large number of species in the taxon have, however, many times successfully secondarily invaded wet and even aquatic environments. The retention of atmospheric respiration in such cases is a clear manifestation of the advantages derived from air breathing in the taxon and animals in general. In being able to carry air with them in form of gas bubbles (Fig. 104), the insects have retained the major advantage of extracting  $O_2$  from air while subsisting in water. They have gained access to new resources in water and escaped from surface predators. The simplest mode of aerial respiration in aquatic insects is the snorkel one utilized by many larval forms, e.g., the mosquito larva *Culex*, where a breathing tube opening to a spiracle placed on one end of the body is brought into contact with the surface



**Fig. 104a–c.** Underwater air-breathing in insects. **a** An air bubble entrapped underneath a wing of a beetle. **b** A physical gill. **c** Gas exchange in a plastron. (Hughes 1982)

Fig. 105. Schematic diagram of a compressible (physical) gas gill. Oxygen diffuses into the bubble from the surrounding water () to replace the amount consumed in the body. The  $CO_2$  released from the body dissolves into the surrounding water. The partial pressure of nitrogen decreases, leading to eventual collapse of the bubble



of the water. The air is transferred to the body by diffusion, the surface film being utilized for support using hydrofuge hairs. Amazingly, the underwater gas bubble utilized by the insect is comparable to the air held in the lung of a pneumonate gastropod (Hunter 1953), providing a micromileu which facilitates exchange of  $O_2$  and  $CO_2$  across the water-gas interface (Wolvekämp 1955; Rahn and Paganelli 1968).

Two modes of underwater air breathing have evolved in insects especially in the Hemiptera and Coleoptera. In the "compressible gas-gill" (Rahn and Paganelli 1968; Fig. 105), gas pockets or bubbles of air adherent to the body parts (Fig. 104) are utilized as sources of O2. Groups such as the diving beetles (Dytiscids), the black swimmers (Notonectids), and water boatman (Corixa) (Ege 1918; Crisp 1964) utilize this mode of gas exchange where gas transfer is effected between the trachea, the gas-gill, and the water. In a compressible gas-gill, the pressure of the gases is dependent on the prevailing hydrostatic pressure, i.e., the depth at which the insect operates. Depth shortens the duration of the dive by increasing the rate of  $O_2$  loss into the water due to the increased  $PO_2$  (Rahn and Paganelli 1968): a dive of 2 m increases the PO<sub>2</sub> in the gas-gill to 25 kPa, creating a partial pressure gradient of 4 kPa in favor of loss of O<sub>2</sub> from the gas-gill. Oxygen is extracted by the trachea through the spiracles which open into the bubble. The discharged  $CO_2$  rapidly dissolves into the surrounding water. As the  $PO_2$  in the bubble decreases, in well-oxygenated waters, O<sub>2</sub> diffuses inwards from the surrounding water. Due to the increase of the partial pressure of nitrogen in the bubble, indirectly through loss of CO<sub>2</sub> into the surrounding water and directly through the hydrostatic pressure which increases at a rate of 1 atm per 10 m,  $N_2$  is slowly lost. This results in a steady decrease in the size of the bubble and its eventual collapse at a critical diameter (Rahn and Paganelli 1968; Liew 1970). To increase the rate of diffusion of O<sub>2</sub> into the bubble, some insects actively ventilate the gas-gill by moving water currents around it. The frequency at which an insect makes periodic visits to the surface to pick up fresh air is dependent on: (1) the pressure prevailing in the gas bubble, which in turn is determined by the working depth (the PN, between the gill and water increases at a rate 9.9 kPa m<sup>-1</sup>); (2) O<sub>2</sub> uptake by the tissues which indirectly leads to an in situ elevation of the concentration of N<sub>2</sub> enhancing its efflux; and (3) the PO<sub>2</sub> in the surrounding water and hence the rate of O<sub>2</sub> recharge of the gas gill from the surrounding water or is even lost from the air bubble when the animal is in hypoxic water. Before the collapse of the gas gill, however, the insect will have extracted a volume of O<sub>2</sub> from the water which is 7 to 13 times greater than that in the bubble at the beginning of the dive. This extends the dive duration 8 to 13 times (Ege 1918; Rahn and Pagannelli 1968). In a Corixa, Ege (1918) observed that the insect obtains enough O<sub>2</sub> out of the water by diffusion into the air bubbles covering its body, a supply which extends its underwater stay 10 to 30 times longer than would be possible if it were to rely only on the initial amount of  $O_2$  in the gas bubble(s) at submergence. The spider, Argyroneta, which always carries a layer of air around the entire abdomen and part of the thorax, is highly adapted for underwater gas gill respiration, with the air sometimes lasting for several days in summer (Braun 1931). Interestingly, an insect can stay underwater longer when the gas gill is filled with air (in airsaturated water) than when the gill is filled with pure O2. A backswimmer survives for only 35 min in O<sub>2</sub> saturated water but for as long as 6 h in air-saturated water. When a Notonecta was put in O<sub>2</sub>-saturated water and also made to breathe from a pure O<sub>2</sub> bubble, the animal became heavier than water in 14 min and succumbed in 35 min. When breathing ordinary air (at the same temperature), however, it could survive for 6h without surfacing (Ege 1918). This is due to the fact that the presence of N<sub>2</sub> enables the gas gill to function as a physical gill, i.e., it allows O<sub>2</sub> and CO<sub>2</sub> transfer between the water and the gas gill. During winter when the surface of water is covered with ice, a number of species of the Dytiscidae family are highly active, catching air bubbles arising from aquatic plants or from the mud (Krogh 1941). Although such bubbles may contain relatively little  $O_2$ , they should replenish the quantity of N<sub>2</sub> in the air bubbles carried by the insect and hence prolong the survival of the gas gill. While the spiracles and hydrophobic hairs may act as barriers, it is intriguing how the insects totally keep the trachea from flooding with water through capillarity. If the trachea are filled with water, the transfer of O2 would decrease 10000-fold and the insect would almost certainly drown. This has found a practical application in destruction of mosquito larvae by applying oil on the surface of standing water, a process which eliminates the supporting surface tension. Denney (1993) contemplates that the trachea are lined by a waxy substance which repels water.

In insects which possess "incompressible gas gill" (Fig. 106), a layer of gas is held firmly (over the epicuticle which covers those parts of the body onto which the spiracles open) by stiff hydrophobic hairs or a cuticular meshwork of hairs, the plastron (e.g., Hinton 1966) (Fig. 107). The hairs are about  $5\mu m$  long. In the bug, *Aphelocheirus*, the hairs number about  $2.5 \times 10^6$  mm<sup>-2</sup>, are about  $0.5\mu m$  apart, have a diameter of about  $0.2\mu m$ , and are strong enough and adequately hydrophobic to withstand 4 to 5 atm of external pressures before the wetting of the cuticle can occur (Thorpe 1950; Crisp 1964). The surface tensional forces
Fig. 106. Schematic diagram of an incompressible gas gill (plastron). The gas air bubble is firmly attached to the body by hydrofuge hairs. As  $O_2$  is consumed and  $PO_2$  drops in the bubble, the gas diffuses inwards from the surrounding water (). Carbon dioxide dissolves into the surrounding water. Owing to the firm physical support of the bubble, the pressure of nitrogen in the bubble is not a factor in the longevity of a plastron. Theoretically, the bubble may last indefinitely





Fig. 107. Schematic drawing of the hairs of a plastron of a cranefly larva, *Dicranomyia*, that consists of narrow spaces delineated by a roof made up of a thin holey cuticle, n, and thick basal cuticle, f. Vertical struts,  $\succ$ , connect the roof to the floor. The air contained in the interstices is not directly subjected to hydrostatic pressures at natural depths. (Hinton 1976)

augmented by the support provided by the hairs counteract the hydrostatic pressure preventing compression of the gas phase, making the volume of the plastron remain fairly constant (Rahn and Paganelli 1968): the  $PN_2$  in the bubble is equal to that in the water and the rate of  $O_2$  uptake by the insects is the same as that diffusing in from the water. Consequently, the loss of the inert gases, mainly  $N_2$ , which sets the limit of the gas-gill existence, is avoided. Theoretically, this should allow an insect to remain submerged indefinitely (Thorpe and Crisp 1949; Hinton 1976; Rahn and Paganelli 1968). Strictly, the insects endowed with "compressible" (physical gas gill) or "incompressible" gas bubbles are essentially air breathers as they actually exchange  $O_2$  and  $CO_2$  with a gas phase which, in turn, exchanges gases with the surrounding water. The empiric changes of the gas profiles within the tracheal system and air sacs of an insect utilizing such devices are fundamentally similar to those in an insect breathing free air.

The compressible gas-gill is suitable for insects which operate close to the water surface. In such cases, large energetic costs are not involved for surfacing and diving. Incompressible gas gills, within limits, act well at depths and serve those insects which stay under the water for long periods of time well. While the compressible gas gill requires no definite structural adaptations, plastron respiration demands a greater degree of anatomical specializations in form of hydrophobic hairs and/or other cuticular modifications. The typical environment inhabited by insects with a plastron is that which has well-aerated water and one which dries up at times, exposing the insect to the atmosphere. Plastrons are also quite common on insect eggs which are liable to being covered by water when it rains (Hinton 1953, 1966; Crisp 1964). Such eggs have areas which consist of an intricate meshwork of air channels lined by hydrophobic substances.

A plastron is a highly versatile respiratory organ which functions as an efficient underwater gas exchanger and when the insect is exposed to air provides a satisfactory pathway for gas transfer while minimizing risk of desiccation (Hinton 1966). The insectan plastron corresponds with the fine hydrophobic air channels of many aquatic plants which prevent entrance of water while allowing in air. This makes such plants relatively more buoyant. Of particular interest is the mode of respiration in the larva of the bot-fly, Gastrophilus intestinalis which subsists in the virtually anoxic stomach of the horse. In a more or less plastron manner,  $O_2$ is extracted from the air bubbles (which are swallowed during feeding) through a special organ made up of large cells around an array of trachea (Krogh 1941). A totally different mechanism of maintaining a permanent gas-gill has evolved in the volant aquatic elmid beetle: before diving, the beetle carries a large bubble of air which extends from the first femora back over and past the dorsum. In calm water, the beetle has to surface from time to time to replenish the air but in the torrential mountain stream waters, where the currents are faster than  $70 \,\mathrm{cm\,s^{-1}}$ , the Bernoulli-Venturi effect generated by the flow of water across the convex face of the bubble causes the pressure in the bubble to fall below the atmospheric. This effects an inward flux of O<sub>2</sub>. The tracheal (air) gills which comprise of an air-filled tracheal system sealed off from water, e.g., the highly tracheated flaps or filamental outgrowths from the thoracic and/or abdominal segments of the body in nymphal Plecoptera and Ephemeroptera and larval Trichoptera constitute an

extreme retrogressive transformation which allows permanent aquatic habitation:  $O_2$  diffuses from the surrounding water into the trachea which serve as internal plastrons and  $CO_2$  is discharged and dissolved by the surrounding water.

Remarkable behavioral, morphological, and physiological convergencies in underwater respiration occur. Some high-latitude winter-diving mammals are known to use gas pockets trapped between the water-ice interface (e.g., Hart and Fisher 1964; Mitchell and Reeves 1981; MacArthur 1992). Such air pockets may arise from air entering through natural fissures in the ice, from exhaled gases released by the diving animals (e.g., Harrison et al. 1972), from the hair, or from photosynthetic activity of the aquatic plants. In nature, diving muskrats, Ondatra zibethicus (Errington 1963), and experimentally submerged gray seals, Halichoerus grypus (Harrison et al. 1972), are reported to breath in air in these gas pockets. Since CO<sub>2</sub> is more soluble (30 times) in water than O<sub>2</sub>, the PCO<sub>2</sub> in the air in the bubbles discharged from the lung or from the surface of the body should drop with time. Depending on the PO<sub>2</sub> in the surrounding water, the bubbles trapped under ice may be charged with  $O_2$ . A high concentration of  $O_2$  (26.67 to 29.04%) was determined in the air bubbles collected under ice by MacArthur (1992). The investigator, however, attributed it to photosynthetic activity of submerged macrophytes. In addition to the O<sub>2</sub> contained in the intrapulmonary air and the body tissues, the air bubbles in the water/ice interface could provide an important auxiliary source of O<sub>2</sub> during prolonged dives. In the muskrat, O. zibethicus, dive duration was increased by 31% when the animals were allowed access to underwater gas pockets (MacArthur 1992).

## 6.13 The Cleidoic Egg: a Fascinating Gas Exchanger

The evolution of the cleidoic (self-contained) egg in reptiles and birds was a monumental advance in the attainment of terrestriality. For once, the land invaders were able to totally dissociate themselves from water for fundamental activities such as reproduction. In the about 9000 species of birds, in addition to the presence of the ubiquitous feather, oviparity, the mode of reproductive which involves formation of fertilized eggs encased in an egg shell, is a unifying feature. While viviparity has variably evolved in most other vertebrate groups, e.g., amphibians like the caecilian Typhlonectes compressicauda (Garlick et al. 1979), reptiles like the lizard, Sphenomorphorus quoyii (Grigg and Harlow 1981), and teleost fish like Zoarces viviparous (Hartvig and Weber 1984; Weber and Hartvig 1984), Embiotoca lateralis (Ingermann and Terwilliger 1981), and Clinus superciliosus (Veith 1980), birds constitute the only vertebrate class which is exclusively oviparous. Debate ranges on the reasons for retention of this ancestral reproductive condition (e.g., Blackburn and Evans 1986; Anderson et al. 1987; Lewin 1988): it has been suggested that viviparity in birds would inhibit flight due to the increased pay load, during pregnancy. Such an argument is clearly untenable if it is recalled that bats have evolved both flight and viviparity and of the at least 15 families of birds which have given up flight, none has evolved viviparity. Simple cost-benefit analysis may explain the case. Being ectothermic, birds can incubate their eggs outside the body – they do not need to retain them in the body for development. Since the eggs are formed at different times (one every 25 h or so), if retained in the oviduct and/or uterus (space allowing), it would mean that embryos would have to be delivered at different times (even when the bird is still laying!) depending on the incubation period and the period of laying (clutch size). The benefits of retaining oviparity perhaps outweigh the costs and risks. Due to its intrinsic self-sufficiency, some authorities (e.g., Bender 1992) consider the egg to be a "cell". In birds, the 1.5-kg ostrich (*Struthio camelus*) egg is hence the largest extant cell while the dinosaur eggs, e.g., those of the sauropod, *Hypselosaurus priscus* (Kerourio 1981), which had a volume of about 21 (about 40 times the volume of a chicken egg) and the 10-kg ones of the now extinct 500 to 1000-kg elephant bird, *Aepyornis* (Amadon 1947, Rahn et al. 1975; Feducia 1980), are some of the largest cells which ever formed.

Both theoretically and practically, the egg is a marvel of morphogenetic engineering (e.g., Ar et al. 1974). The microcosmos contains all the necessary factors needed for embryonic development such as nutriments, minerals, and water: the only factor missing is O<sub>2</sub>, a resource which must be procured from outside. The products of metabolism, CO<sub>2</sub> and water, must be removed while the nongaseous ones (e.g., products of nitrogen metabolism) are stored in the allantois. The shell must be strong enough to mechanically and physically protect the developing chick from trauma during brooding and from toxic, infectious, and parasitic agents: O<sub>2</sub> must be allowed in and CO<sub>2</sub> let out, excessive water loss must be avoided, while the shell must be adequately weak for the chick to break out at the end of incubation (Ar et al. 1979; Schmidt-Nielsen 1984). The eggshell is made up of calcium carbonate (CaCO<sub>3</sub>) arranged in orderly crystalline arrays embedded in a protein matrix separated by fine air spaces, the pores. The differences in the pattern of deposition of CaCO<sub>3</sub> crystals accounts for the disparities in the eggshell conductances of newly laid eggs for O<sub>2</sub> (Tazawa 1987). The conductances of the eggshell of the African parrot, Enicognathus ferrugineus, differs by a factor of 7 (Bucher and Barnhart 1984) and in the turkey, Meleagris gallopavo, the number of pores in the eggshells changes during a laying cycle, increasing during the late stages of laving (Rahn et al. 1981). The shell of a 60-g chicken egg has a surface area of about  $70 \text{ cm}^2$  and about 10000 pores which are 17 um in diameter and 0.35 mm in length (Wangensteen et al. 1971; Tazawa 1987). Much as it is a rigid structure, the shell does not completely isolate the developing chick from the external environment. The embryo receives information in form of sound or mechanical movements from the parent(s) and the adjacent eggs, cues used to synchronize hatching in a clutch of eggs (Drent 1975). The need to reconcile remarkably different requirements has led to what appears like near-optimization of the gas transfer capacity of the avian eggshell. Wangensteen and Weibel (1982) observed that the diffusing capacity of the chorioallantois of a 16-day-old egg was equal to the physiological diffusing capacity. Weibel (1984a) attributed the optimal state of the avian eggshell to its transient nature since it does not add immense cost to support and maintain. It is instructive to note that the placenta (Sect. 4.7), an equally ephemeral organ, maintains a substantial functional reserve during most of the gestation period (Karsdorp et al. 1996).

Since the capacity to conduct the respiratory gases is somewhat fixed at the formative stage of the eggshell, i.e., before incubation starts (Wangensteen et al. 1971; Kayar et al. 1981; but see dissenting views e.g., Kutchai and Steen 1971; Lomholt 1976a; Tullet and Board 1976), the design must somewhat adaptively preempt the maximum  $O_2$  and  $CO_2$  flux at the peak of embryonic development. Unique to all gas exchangers, where both  $CO_2$  and  $O_2$  are transported with balanced facility, in the chorioallantois of the bird egg, gases are transported at different rates corresponding with their molecular weights. Unique to all other gas exchangers, each gas has its own diffusion coefficient. On this basis, Rahn and Paganelli (1982) suggested that while the convective transport of the lung can be described as an egalitarian transport, that in the avian egg should be considered an elite transport system. Throughout the incubation period, a time when  $O_2$  consumption increases by a factor of about 800 times (Kutchai and Steen 1971; Rahn et al. 1974; Table 32), the bird embryo is encased in a rigid shell of invariable area and thickness (Table 31). Kutchai and Steen (1971) estimated that  $O_2$  and  $O_2$ 

Species	Body wt. (approx – kg)	Egg wt. (g)	Shell thickness (mm)	Mean pore diameter (mm)	Thickness of the membranes (mm)
Aepvornis	500	12000	4.40	-	_
African ostrich	150	1400	1.95	0.035	0.200
Australian swan	17	700	0.69	0.34	0.165
Holland turkey	8	80	0.41	0.040	-
Chickens	3.3	53.7	0.31	0.018	0.065
Pheasant	3.0	32	0.26	0.012	-
Ouail	2.0	9	0.13	-	0.067
Finch	0.01	1	0.09	-	0.005
Hummingbird	0.008	0.5	0.06	-	-
Auk	_	-	-	0.041	-
Duck	-	-	-	0.023	-
Gull	-	-	-	0.013	-

Table 31. Morphometric parameters of the eggs of several species of birds. (Romanoff and Romanoff 1949)

**Table 32.** Day of incubation (DI), cardiac output (CO), oxygen consumption ( $Vo_2$ ), hemoglobin content (cHb), oxygen capacity ( $O_2C$ ), transport capacity (TC), and oxygen utilization coefficient ( $O_2UC$ ) of the chicken during development. (Romanoff 1967; Bartels et al. 1996)

DI	CO	Vo <sub>2</sub>	cHb	O <sub>2</sub> C	TC	O <sub>2</sub> UC
3	20	3.8	0.015	0.0201	0.4	950
5	120	9.4	0.028	0.0375	4.5	210
12	4800	121	0.069	0.0926	445	27
17	6800	305	0.098	0.1310	826	37
17	6800	305	-	0.1100	750	41

permeabilities of the shell and the membranes of a newly laid egg are not adequate to support gas exchange needs during the last stages of the development of the embryo. The megapodes, birds of the Southern-Pacific area, e.g., the Australian malee fowl, Leipoa ocellata, and the brush turkey, Alectura lathami (Brom and Dekker 1992; Dekker and Brom 1992; Jones and Birks 1992), deposit their eggs deep in the soil among putrefying organic matter and on rare occasions in geothermal heat to provide natural warmth for incubation (e.g., Fleav 1937; Frith 1956). In the egg mound, the eggs must face critical levels of hypoxia and hypercapnia especially during the last stages of incubation. Furthermore, the developing embryos are exposed to a moisture-saturated environment where there is minimal water loss. In most birds, water loss, which may constitute as much as 20% of the initial mass of the egg, is essential for proper development of the eggs (Rahn and Ar 1974; Ar and Rahn 1980; Simkiss 1980). Within a short period, the hatchlings must find their way to the surface or face suffocation. The shells of the eggs of the megapodes are remarkably thin, allowing rapid outward diffusion of metabolic water, out flux of CO<sub>2</sub>, and influx of O<sub>2</sub> (Drent 1975). The eggs of the painted turtle, Chrysemys picta (Emydidae), present another interesting mode of development: though the eggs hatch in late summer or early autumn, the neonates do not emerge from their subterranean nests until the following spring, i.e., 6 to 9 months after hatching (e.g., Linderman 1991; Constanzo et al. 1995). Though the great mortality of the neonates has been associated with hypothermia (Packard and Packard 1997), hypoxia and hypercapnia (especially when the ground is covered with ice) may be significant contributing factors to the fatalities. Whereas turning of the eggs during incubation is critical for proper embryonic development (e.g., Romanoff 1960; Metcalfe et al. 1979), the eggs of the megapodes develop normally without it (Seymour and Rahn 1978; Seymour and Ackerman 1980). In some reptiles, the sex of the embryo is determined by the ambient temperature (e.g., Ferguson 1992). In fixed incubating eggs, concentration of the albumen occurs arresting the development of the chorioallantoic vessels (Romanoff 1960) and that of the albumin sac (Randles and Romanoff 1950). The normal clutch of about 100 eggs laid at a depth of about 1 m by the green turtle, Chelonia myda, face extreme hypoxia and hypercapnia at the end of incubation. The O<sub>2</sub> level falls to 12% and that of CO<sub>2</sub> rises to above 2.24% (Prange and Ackerman 1974). The dinosaurs, e.g., the sauropod, Hypselosaurus priscus, are thought to have deposited their eggs in small groups (e.g., Kerourio 1981) instead of about 50 in one clutch, as such a mass of eggs would have consumed O<sub>2</sub> faster than it could diffuse through the walls of the nest (Seymour 1979).

Birds have been able to adapt to a wide variety of nestling conditions by adjusting the eggshell microarchitecture to provide optimal gas exchange for the special needs in their habitats. The incubation period of the bird eggs is inversely related to metabolic rate and the eggshell conductance (Rahn et al. 1974). Pathological conditions which affect the shells of the bird eggs occur frequently especially in cases of malnutrition, e.g., when laying birds experience calcium deficiency. The shells are poorly formed and fragile. A recent environmental problem which has lead to extensive eggshell deformation in birds has resulted from use of pesticides such as DDT (Cooke 1976; Fox 1976; Risenbrough 1986). During the 21-day incubation period, a 60-g chicken egg will take 61 (8.6 g) of  $O_2$ , give off 4.51 (8.8g) of CO<sub>2</sub>, lose 9g in weight (=111 of water vapor) and by the end of incubation, about 30 kcal of the initial 100 kcal energy in the egg will have been consumed to form the 39g of the chick (e.g., Carey et al. 1980; Rahn and Ar 1980). In the species which nest at high elevations (see Rahn 1977), the respiratory demands of the eggs are daunting. Not only do the eggs of such species have to cope with the prevailing hypoxia but must also experience low ambient temperatures and excessive water loss due to the low vapor pressure prevalent at elevation (Rahn et al. 1976). Hatchability of chicken and turkey eggs decreases with altitude (Weiss 1978), the mortality rate being particularly more pronounced during the second week of incubation. Incubated under the same conditions, the eggs of species from wet habitats lose mass at a higher rate than those from drier habitats (Lomholt 1976b). Eggs laid by chickens acclimatized to an altitude of 3.8km showed a reduction in the total pore area (Wangensteen et al. 1974; Packard et al. 1977; Carey 1980a,b), a feature which should curtail water loss (Tazawa 1987). In eggs that are laid at higher altitude, the number of pores in the shell is less, the shells are thicker, and the water content is higher (Rahn et al. 1977; Carey 1980a,b). The eggs of the birds which nest over water, e.g., the pied-billed grebe, Podilymbus podiceps, have a high density of pores which allows them to lose water to the relatively humid air around them (Drent 1975). The hatchability of the eggs laid by high-altitude (3.8 km) hens improves with the generations of residence at elevation (e.g., Carey et al. 1982). The total effective pore surface area of eggshells decreases with the barometric pressure (Packard et al. 1977). Increased diffusivity of gases through the eggshell due to enhanced gas-phase diffusion compensates for the prevailing hypoxia but results in hypocapnia due to increased CO<sub>2</sub> efflux (Rahn and Ar 1974). The PCO<sub>2</sub> in the air cell drops from 4.3 to 2.9 kPa when sea level eggs are exposed to 0.5 atm of a gas mixture containing 40% O<sub>2</sub> in N<sub>2</sub>, to prevent hypoxia. Hyperoxia has a damaging effect on the development of the chick embryo. Exposure of O<sub>2</sub> at 5 atm for 3 h on 72-h-old chick causes a more than 50% mortality and 20 to 30% of those chicks which hatch have deformities of the brain, eyes, upper jaw, legs, feet, and heart (Pizarello and Shircliffe 1967). Due to the gas diffusion changes which occur at altitude and the fact that the measure of egg shell conductance is somewhat fixed once an egg is laid, fertile eggs laid at sea level but incubated at altitude require specific changes in the composition of gases such as enrichment with O<sub>2</sub> and CO<sub>2</sub> in the incubator for normal hatching to occur (Visschedjik and Rahn 1981). Incubation of eggs laid at sea level at an altitude of between 3.1 and 3.8 km results in reduced metabolic rates, prolonged incubation periods, low body mass at hatching, and unpredictable hatchability (Beattie and Smith 1975). To achieve the same conductance, the shells of the large eggs are relatively thicker than those of small eggs but are much more porous (e.g., Ar et al. 1974). The pores occupy 0.02% of the area of a chicken's egg shell but 0.2% of the area of that of an ostrich.

Gas exchange in the avian egg from the ambient air to the chorioallantoic capillary blood takes place by diffusion in the gas phase. The pathway through which  $O_2$  diffuses from outside comprises of: (1) a variably thick cuticle (a noncellular mucinous coat closely applied on the surface of the eggshell); (2) a shell (a hard calcaneous and porous structure); (3) outer and inner shell membranes; and

(4) allantoic capillary blood (Fig. 108). The thickness of the eggshell (0.3 mm in the domestic fowl) varies among species (Table 31), depending on factors such as body and egg size and nutrition and nutritional status (Romanoff and Romanoff 1949). The inner shell membrane surrounds the albumin and the outer one is cemented to the shell. While the inner shell membrane has been presumed to be "wet" and hence presents significant resistance to O<sub>2</sub> transfer (e.g., Kutchai and Steen 1971; Wangsteen 1972; Tullet and Board 1976; Lomholt 1976a), morphometric determination of the diffusing capacity of the egg by Wangensteen and Weibel (1982) indicated that the membrane is "dry" and offers negligible resistance to O<sub>2</sub> flux. Of the three components of the chorioallantoic blood-gas barrier, the blood-gas barrier itself of which the harmonic mean thickness is 0.40 µm offers 10%, the plasma layer about 2%, and the O<sub>2</sub> binding rate 88% of the total resistance to O<sub>2</sub> flow (Wangensteen and Weibel 1982). Feeding acidic salts such as ammonium chloride (Hunt and Aitken 1962) and exposure to high concentrations of CO<sub>2</sub> for half a day for about 2 days (Helbacka et al. 1963; Hunt and Simkiss 1967), factors which lead to metabolic acidosis, decrease shell thickness. Pore geometry and the diffusion coefficient of a particular gas determine the permeability of the eggshell. With the onset of incubation when the embryonic heart starts beating, the circulation of the blood through the capillaries of the embryo



**Fig. 108.** Cross-section of an avian egg at an advanced stage of development. The dimensions of the main components which form the gas exchange pathway are shown. (Wangensteen et al. 1971)

add a convective process to the earlier entirely diffusive one. The formation of blood starts about 25h after the start of incubation and hemoglobin containing erythrocytes appear by the 2nd day (Romanoff 1960). As occurs in the lungs of birds (Maina et al. 1989a), the greatest impedance to O<sub>2</sub> uptake in the egg lies in the reaction between O<sub>2</sub> and the hemoglobin. The process contributes 88% of the overall resistance, the plasma only 2%, and the remarkably thin blood-gas barrier 10% (Wangensteen and Weibel 1982). The concentration of the hemoglobin does not change significantly between the 12th day of incubation and hatching, when 90% of it is converted to the adult form (Sandreuter 1951). Similarly, over the greater part of the period, the  $O_2$ -carrying capacity of blood does not change considerably (Bartels et al. 1966). Erythropoiesis in the developing chick is suppressed by hyperoxia and enhanced by hypoxia (Jalavisto et al. 1965). In complete contrast to the mammalian placenta (Sect. 4.7) which adjusts its gas exchange capacity with gestation, i.e., as the metabolic requirements of the fetus increase, the gas exchange machinery of the egg especially regarding pore number, size, and geometry are somewhat fixed despite the gradual increases in O<sub>2</sub> demands with incubation (Wangensteen et al. 1971). Romijn (1950) and Kutchai and Steen (1971), however, observed that eggshell permeability increases with incubation and estimated that at day 20 of incubation, 575 ml O<sub>2</sub> diffuse into the egg per day. Like the mammalian embryo, the avian one subsists under perpetual hypoxia. Towards the end of incubation, high  $PCO_2$  and low  $PO_2$  in the air space (Romijn 1948) may prompt the chick to break through the shell. A change in the  $O_2$ dissociation curve of the blood of 20-day-old embryos was observed by Bartels et al. (1966): the  $P_{50}$  was only 4 kPa in the embryonic blood compared with 6.7 kPa after a few days of hatching. In the egg, the  $PO_2$  and  $PCO_2$  of the air cell is set by the O<sub>2</sub> uptake and CO<sub>2</sub> release (metabolism) of the embryo and the conductances of the egg shell (Wangensteen and Rahn 1970; Paganelli 1980). Throughout incubation, CO<sub>2</sub> permeability of the eggshell and membranes is two to three times higher than that of O<sub>2</sub>, showing that diffusion occurs mainly through air-filled rather than fluid-filled pores (Kutchai and Steen 1971). Given the relatively low diffusion coefficient of O<sub>2</sub> in water, eggs with solid shells would theoretically have to be very small and possess a very high density of pores and low rate of O<sub>2</sub> consumption to survive in water. Such eggs occur in skates and rays (Denney 1993). In a water-saturated environment, the embryos in the larger avian eggs soon drown. The chorioallantoic membrane, a very well-vascularized organ which is closely applied to the inner shell membrane, is the only adaptable structural factor in the egg. It develops steadily and only completely underlies the whole shell towards the beginning of last half of the incubation period (Wangensteen and Rahn 1970; Wangensteen 1972). The degree of vascularization of the outer allantoic surface is increased when the eggs are incubated in a lower PO<sub>2</sub> (Remotti 1933).

The chorioallantoic membrane is considered to be a homolog of the mammalian placenta (Metcalfe and Stock 1993). The diffusing capacity of the chorioallantoic membrane for  $O_2$  in chicks increases sixfold between days 10 and 18 of incubation. During late incubation, the mass-specific diffusing capacity and the capillary blood volume are similar to those of the human lung (Tazawa and Mochizuki 1977). Compared with the viviparous species, where the change from

embryonic respiration is abrupt on the breaking of the umbilical cord, in the oviparous species, the change is a gradual process. Mainly owing to the fact that the metabolic rate of the developing embryo increases steadily while the permeability of the shell arguably remains almost constant (Kutchai and Steen 1971; Rahn et al. 1974), in bird eggs, a critical hypoxic phase occurs towards the end of embryogenesis (Freeman and Misson 1970; Girard and Muffat-Joly 1971; Wangensteen 1972). As incubation progresses, the PO<sub>2</sub> in the air cell decreases from 18.7 to 13.3 kPa and that of CO<sub>2</sub> increases from 0.67 to 5.3 kPa (Wangensteen 1972; Table 33). At about day 19, the concentration of O<sub>2</sub> is about 10 to 12% and that of CO<sub>2</sub> lies between 6 and 8% (Tazawa et al. 1983b). As the embryo uses the O<sub>2</sub> inside the shell's air space (Fig. 108), the PO<sub>2</sub> decreases. This increases the partial pressure gradient of O<sub>2</sub> between the ambient air and that inside the egg, enhancing the O<sub>2</sub> influx. Until the 8th day of incubation, the partial pressure gradient of  $O_2$  between the outside of the shell and the air space is about 1.3 kPa, at the 16th day it rises to 6.0, and rises to 10kPa by the 18th day (Metcalfe 1967). This indicates that as hatching advances, the PO<sub>2</sub> in the allantoic arterial blood must fall to very low levels, increasing the danger of the chick succumbing to hypoxia. Bartels et al. (1966) observed that after day 17, compared with the state after hatching, the chick embryo showed signs of respiratory acidosis: the CO<sub>2</sub> content of blood at a PCO<sub>2</sub> of 5.3 kPa is 54 ml per 100 ml of blood at the 17th day and decreases to about 36 ml after hatching. The autoregulating process of the PO<sub>2</sub> in and outside the developing egg ascertains that the embryo is well supplied with  $O_2$  by diffusion. In regulating  $O_2$  flux, the embryo is not a passive participant. The hypoxia which occurs during the late embryogenesis induces increase in the level of the catecholamines which improve the blood gas status (Wittman and Prechtl 1991). The permeability of the eggshell must adaptively be designed to provide the maximum amount of O<sub>2</sub> required by the embryo, regulating it around the

Embryo age (days)	PAO <sub>2</sub>	PACO <sub>2</sub>	
7	141.6	13.6	
8	140.0	12.7	
9	138.1	16.2	
10	137.5	16.0	
11	134.7	18.3	
12	131.3	19.4	
13	129.2	21.2	
14	124.5	24.8	
15	117.9	30.8	
16	111.9	36.6	
17	106.4	41.8	
18	109.9	38.7	
19	102.3	43.3	

Table 33. Measured air cell gas tensions  $(mmHg)^a$  of an incubated chicken egg. (Wangensteen and Rahn 1970)

<sup>a</sup> To convert to kPa multiply by 0.013.

critical minimum arterial PO<sub>2</sub> needed for survival. Reciprocatively, the O<sub>2</sub> consumption of an embryo is determined by the conductance of the shell and the lowest tolerable arterial PO<sub>2</sub> (Wangensteen and Rahn 1970). The acid-base status of the embryo is regulated by increases in plasma concentration of the HCO<sub>3</sub><sup>-</sup> ions in direct proportion with the changes in the PCO<sub>2</sub>. This maintains a constant pH of about 7.45 (Erasmus et al. 1970/71). During incubation, the arterial PCO<sub>2</sub> reaches a value of about 5.3 kPa, which is about the same level as that in the lungs of the hatched chicks. Wangensteen and Rahn (1970) considered this to be a preparation for smooth transition to air breathing when the gas exchange process changes from diffusion respiration of the chorioallantois to the convective one of the lungs. The changeover from total chollioallantoic respiration to pulmonary respiration takes 22 to 30h in the chicken (Visschedijk 1968a; Vince 1973). The time of pipping corresponds with increased PCO<sub>2</sub> or decreased PO<sub>2</sub> in the egg air space (Vince et al. 1975), the stimulating effect of CO<sub>2</sub> being two times more potent than that of O<sub>2</sub> (Visschedijk 1968b).

## 6.14 The Bottom Line

In nearly all evolved life forms, throughout their existence, molecular O<sub>2</sub> to varying extents has been in constant demand. Respiration has hence been integral in molding the complex morphologies in biology. As a terminal hydrogen acceptor in the ubiquitous scheme of intracellular oxidation by electron transfer,  $O_2$  is involved in the vital energy-producing processes which sustain life. The gas exchangers have evolved and adapted in tandem with the respiratory requirements of whole organisms in different states, environments, and habitats. Once they were genomically inaugurated, the designs were continually fashioned by needs and circumstances. The shared morphological contrivances evident in the constructional plans of the gas exchangers indicate that similar pressures have inspired the congruous designs. The aphorism that necessity is the mother of invention is as relevant to evolution and adaptation to novel designs in biology as it is to human activities and needs in the contemporary world. Change in size and activity and subsistence in unique habitats has called for appropriate adjustments for efficient O<sub>2</sub> uptake, transfer, and utilization. Among many animals, solutions to these needs have differed only in details, and not in generality. For example, sheet flow of blood at the gas exchanger, closed circulation, double circulation, presence of respiratory pigments in blood and muscle tissue, etc., are some of such prevalent devices. There are no rules in the construction and working of the gas exchangers: means justify ends! Contingent upon the available resources and the specific requirements for survival, the final solutions are arrived at independently. For instance, in insects, the lengthy and costly developments that would have entailed formation of, e.g., a circulatory system, blood, carrier pigments, and blood cells were ingeniously circumvented simply by elaboration of a tracheal system which delivered  $O_2$  directly to the tissue cells from the atmosphere. An animal's success in life is determined by the arsenal of vital adaptations it has creatively appropriated, devised, and harnessed along the way. In the formidable continuous forward momentum enforced by natural selection, befitting improvements in the respiratory processes are pivotal for survival. Our response to the question asked by Weibel (1983b) – is the lung built reasonably? – is yes, and to that by Dempsey (1986) – is the lung built for exercise? – is no. We hasten to qualify the latter response as follows: exercise is an extreme costly state of operation which in virtually all animals is endured only momentarily. Gas exchangers are designed to operate at the most economical levels but have inbuilt plasticity of making functional adjustments in response to moderate pressures.

As established in the laws of thermodynamics, a fundamental aspect integral to life's existence on Earth is that it survives on finite, diminishing resources. This rather fatalistic state of affairs calls for efficient means of procurement and economical utilization of the limited resources. It obliges optimal designs of biological systems. Strategies for thrifty use and maximization of the energy reserves and supply developed very early in evolution (e.g., Priede 1977; Szarski 1983). Through painstaking cost-benefit analysis of causes and effects and rationalization and appraisal of the utilities of the attributes that they are endowed with, animals are innately engineering and carefully crafted by natural selection to a high fidelity state from which they are able to ameliorate or overcome the external pressures which continually besiege them. The practical consequence of evolution and adaptation has been to fashion optimal systems where form and function associate harmonically. Nowhere else has this need been of more paramount importance than in the design of the gas exchangers. On a low-energy life-style animal, the snail, Ramsay (1968) observed that "the more one looks into the snail's way of life (plant food is abundant in near vicinity and when threatened it takes refuge in its shell) the more one sees that it has nothing to gain by stepping up its activity, and for its low level of activity a respiratory organ and a circulatory system of moderate efficiency are adequate". In extreme circumstances and when need has justified it, animals have adopted extreme measures of reducing their metabolic requirements, e.g., by going into programmed low energy retreats like estivation and cryptobiosis (e.g., Storey 1988, 1989; Storey and Storey 1990). When warranted, some animals have resolutely deconstructed organs and organ systems and, where need has justified it, readopted past simpler designs and more economic strategies of operation. In many cases, if not in most, when all else has failed, two or more animals (and even animals and plants!) have abandoned their genetically programmed evolutionary trajectories and adopted beneficial symbiotic (= cooperative) associations (e.g., Margulis 1979; Trench 1979; May 1981; Childress et al. 1989; Fenchel and Finlay 1991a,b; McFall-Ngai and Ruby 1991; Finlay and Fenchel 1993; Rennie 1992; Vogel 1997). By pooling their capabilities, the partnership ensured access to resources which otherwise would be unreachable to the individuals alone. Identifying and reconciling the factors that enforce the different or similar morphologies and lifestyles on organisms remains an interesting and challenging aspect in biology. It calls for a vast synthesis of different disciplines of science. Cognizance of the effects past events have engendered on present life is both theoretically engaging and practically useful. It is hopefully by understanding how we got here that we may be able to better anticipate where we are going. The very survival of the humankind may rest

squarely on understanding and considerate management of our single most important resource – biodiversity. The opinion offered by Thomas Thomson in 1802 is as valid now as it was some two centuries ago. It is an appropriate closing introspection.

"As soon as man begins to think and to reason, the different objects which surround him on all sides naturally engage his attention. He cannot fail to be struck with their number, diversity, and beauty; and naturally feels a desire to be better acquainted with their properties and uses. If he reflect also, that he himself is altogether dependent upon these objects, not merely for his pleasures and comforts, but for his very existence, this desire must become irresistible. Hence (it is) that curiosity, that eager thirst for knowledge, which animates and distinguishes generous minds." A system of chemistry, by Thomas Thomson, 1802 – quoted in Holmyard and Palmer (1952)

## References

Abdalla MA, Maina JN (1981) Quantitative analysis of the exchange tissue of the avian lung (Galliformes). J Anat 134:677–680

Abdalla MA, Maina JN, King AS, King DZ, Henry J (1982) Morphometrics of the avian lung. 1. The domestic fowl (*Gallus gallus variant domesticus*). Respir Physiol 47:267–278

- Abel DC, Koenig SS, Davis WP (1987) Immersion in the mangrove forest fish, *Rivulus marmoratus*: a unique response to hydrogen sulfide. Envion Biol Fishes 18:67-72
- Able KW, Twichell DC, Grimes CB, Jones RS (1987) Tile fishes of the genus *Caulolatilus* construct burrows in the sea floor. Bull Mar Sci 40:1-10
- Abzug MJ (1994) Characterization of the placental barrier to murine enterovirus. Placenta 16:207-219
- Acarregui MJ, Brown JJ, Mallampalli RK (1995) Oxygen modulates surfactant protein mRNA expression and phospholipid production in human foetal lung in vitro. Am J Physiol 268:L818–L825
- Accurso FJ, Alpert B, Wilkening RB, Petersen RG, Meschia G (1986) Time dependent response of foetal pulmonary blood flow to an increase in foetal oxygen tension. Respir Physiol 63:43– 52
- Ackerman RA, White FN (1979) Cyclic carbon dioxide exchange in the turtle *Pseusemys scripta*. Physiol Zool 52:378–389
- Adams D (1979) The hitchhiker's guide to the galaxy. Pocket Books, New York
- Adams FH, Yanagisawa M, Kuzela D, Martinek H (1971) The disappearance of foetal lung fluid following birth. J Paediatr 63:881-888
- Adams JM, Cory S (1991) Transgenic models for haemopoietic malignancies. Biochim Biophys Acta 1072:9–31
- Adelman IR, Smith LL (1970) Effects of oxygen on growth and food conversion efficiency of northern pike. Prog Fish Cult 32:93–96
- Adelman R, Saul RL, Ames BN (1988) Oxidative damage to DNA: relation to species metabolic rate and life span. Proc Natl Acad Sci USA 85:2706–2708
- Adler K, Stauffer-Holden W, Repine J (1990) Oxygen metabolites stimulate release of high molecular weight glycoconjugates by cell and organ cultures of rodent respiratory epithelium via an arachidonic acid-dependent mechanism. J Clin Invest 85:75-85
- Adolph EF (1943) On the appearance of vascular filaments on the pectoral fin of *Lepidosiren* paradoxa. Anat Anz 33:27–30
- Agar WE (1908) On the appearance of vascular filaments on the pectoral fin of Lepidosiren paradoxa. Anat Anz 33:27-30
- Agostini C, Cipriani A, Cadrobbi P, Semenzato G (1995) The pulmonary immune system and HIV infection. Eur Respir Monogr 2:89–124
- Aherne W, Dunnill MS (1966) Morphometry of the human placenta. Br Med Bull 22:5-8

 Ahlgren JA, Cheng CC, Shrag JD, De Vries AL (1988) Freezing avoidance and the distribution of antifreeze glycoproteins in body fluids and tissues of Antarctic fish. J Exp Biol 137:549-563
Akester AR (1974) Deformation of red blood cells in avian blood capillaries. J Anat 117:658

Alabaster VA, Bakhle YS (1972) The inactivation of bradykinin in the pulmonary circulation of isolated lungs. Br J Pharmacol 45:299–309

Alberch P (1980) Ontogenesis and morphological diversification. Am Zool 20:653-667

Alberch P (1985) Problems with the interpretation of developmental sequences. Syst Zool 34:46– 58

- Alcorn D, Alexander IGS, Adamson TM, Maloney JE, Ritchie BC, Robinson PM (1977) Morphological effects of chronic tracheal ligation and drainage in the foetal lamb lungs. J Anat 123:649–660
- Alcorn D, Adamson TM, Maloney JE, Robinson PM (1980) Morphological effects of either chronic bilateral phrenectomy or vagotomy in the foetal lamb lung. J Anat 130:683–695
- Aldrich JC, McMullan PM (1979) Observations on non-locomotory manifestations of biological rhythmns and exitement in the oxygen consumption rates of crabs. Comp Biochem Physiol 62:707–709
- Aleksiuk M, Frohlinger A (1971) Seasonal metabolic organization in the muskrat (*Ondatza zibethica*). I. Changes in growth, thyroid activity, brown adipose tissue, and organ weights in nature. Can J Zool 49:1143–1154
- Alerstam T (1982) Fagelflyttning (bird migration). Signum, Lund
- Alexander RMcN (1967) Functional design in fishes. Hutchison, London
- Alexander RMcN (1981) Factors of safety in the structure of animals. Sci Prog Oxf 67:109-130
- Alexander RMcN (1982a) The energetics of vertical migration by fishes. Symp Soc Exp Biol 26:273-294
- Alexander RMcN (1982b) Optima for animals. Edward Amold, London
- Alexander RMcN (1985) The ideal and the feasible: physical constraints on evolution. Biol J Linn Soc 26:345–358
- Alexander RMcN (1990) Size speed and buoyancy adaptations in aquatic animals. Am Zool 30:189–196
- Alexander RMcN (1993) Buoyancy. In: Evans DH (ed) The physiology of fishes. CRC Press, Boca Raton, pp 75–97
- Alexander RMcN (1996) Optima for animals. Princeton University Press, Princeton
- Alford CA, Neva FA, Weller TH (1964) Virologic and serologic studies on human products of conception after maternal rubella. N Engl J Med 271:1275–1281
- Allegre CJ, Schneider SH (1994) The evolution of the earth. Sci Am 271:144-151
- Alsterberg G (1922) Die respiratorischen Mechanismen der Tubificiden. Acta Univ Lund NF Avd 2 Bd 18:1–176
- Al-Tinawi A, Madden JA, Dawson CA, Linehan JH, Harder DR et al. (1991) Distensibility of small arteries of the dog lung. J Appl Physiol 71:1714–1722
- Altman PL, Dittmer DS (1961) Blood and other body fluids. Fed Am Soc Exp Biol, Washington, DC
- Al-Wassia AH, Innes AJ, Whiteley NH, Taylor EW (1989) Aerial and aquatic respiration in the ghost crab *Ocypode saratan*. I. Fine structure of respiratory surfaces, their ventilation and perfusion, oxygen consumption, and carbon dioxide production. Comp Biochem Physiol 94A:755–764
- Amadon D (1947) An estimated weight of the largest known bird. Condor 49:159-164
- Amaladoss ASP, Burton GJ (1985) Organ culture of human placental villi in hypoxic and hyperoxic conditions: a morphometric study. J Dev Physiol 7:113-118
- Ames BN, Shigenaga MK, Hagen TM (1993) Oxidants, antioxidants, and the degenerative diseases of aging. Proc Natl Acad Sci USA 90:7915-7922
- Amoore JE (1961) Dependence of mitosis and respiration in roots upon O<sub>2</sub> tension. Proc R Soc Lond 154B:109–129
- Ancilotto F, Chiarotti GL, Scandolo S, Tosatti E (1997) Dissociation of methane into hydrocarbons at extreme planetary pressure and temperature. Science 275:1288–1290
- Andersen HT (1961) Physiological adjustments to prolonged diving in the American alligator, Alligator mississippiensis. Acta Physiol Scand 53:23-45
- Andersen HT (1966) Physiological adaptations in diving vertebrates. Physiol Rev 46:212-243
- Anderson AE, Felbeck H, Childress JJ (1990) Aerobic metabolism is maintained in animal tissue during rapid sulfide oxidation in the symbiont-containing clam Solemya reidi. J Exp Zool 256:130–134
- Anderson DJ, Stoyan NC, Ricklefs RE (1987) Why are there no vivaparous birds? A comment. Am Nat 130:941–947
- Anderson JF (1970) Metabolic rates of spiders. Comp Biochem Physiol 33:51-72
- Anderson JF, Prestwich KN (1982) Respiratory gas exchange in spiders. Physiol Zool 55:72-90
- Anderson M (1978) Optimal foraging area: size and allocation of search effort. Theor Popul Biol 13:397–409

Anderson SA, Bankier AI, Barrell BG, de Brujin MHL et al. (1981) sequence and organization of the human mitochondrial genome. Nature (Lond) 290:457–465

- Andrews EA (1955) Some minute movements in protoplasm. Biol Bull 108:121-124
- Andrews EB, Taylor PM (1988) Fine structure, mechanism of heart function and hemodynamics
- in the prosobranch gastropod mollusc *Littorina littorea* (L.). J Comp Physiol B 158:247-262 Andrews P (1981) Characteristic surface topographies of cells lining the respiratory tract.
  - Biomed Res 2:281–288
- Andriashev AP (1962) A general view of the Antarctic fish fauna. In: Oye V, Mieghem J (eds) Biogeography and ecology in Antarctica. Dr W Junk, The Hague, pp 491–550
- Angersbach D (1978) Oxygen transport in the blood of the tarantula *Eurypelma californicum*: PO<sub>2</sub> and pH during rest, activity and recovery. J Comp Physiol 123:113-125
- Angersbach D, Decker (1978) Oxygen transport in crayfish blood: effect of thermal acclimation and short-term fluctuations related to ventilation and cardiac performance. J Comp Physiol 123B:105–112
- Änggård E (1975) Biosynthesis and metabolism of prostaglandins in the lung. In: Junod AE, Haller R (eds) Lung metabolism. Academic Press, New York, pp 301-311
- Anker GC, Dullemeijer P (1996) Transformation morphology on structures in the head of cichlid fishes. In: Datta-Munshi JSD, Dutta HM (eds) Fish morphology: horizon of new research. Science Publishers, Lebanon, New Hampshire, pp 1–20
- Anonymous (1993) Systematics agenda 2000: charting the biosphere. Department of Ornithology, American Museum of Natural History, New York, 10pp
- Antonini E (1967) Haemoglobin and its reaction with ligands. Science 158:1417-1425
- Antonini E, Rossi-Farelli A, Caputo A (1962) Studies on chlorocruorin. I. The oxygen equilibrium of *Spirographis chlorocruorin*. Arch Biochem Biophys 97:336–342
- Antony EH (1961) Survival of the goldfish in presence of carbon monoxide. J Exp Biol 38:109– 125
- Ar A (1987) Physiological adaptations to underground life in mammals. In: Dejours P (ed) Comparative physiology of environmental adaptations, vol 2. Karger, Basel, pp 208–221
- Ar A, Rahn H (1980) Water in the avian egg: overall budget of incubation. Am Zool 20:373–395
- Ar A, Paganelli CV, Reeves RB, Greene DG, Rahn H (1974) The avian egg: water vapour conductance, shell thickness, and functional pore area. Condor 76:153–158
- Ar A, Rahn H, Paganelli CV (1979) The avian egg: mass and strength. Condor 81:331-337
- Arieli R (1979) The atmospheric environment of the fossorial mole rat (*Spalax ehrenbergi*): effects of season, soil texture, rain, temperature and activity. Comp Biochem Physiol 63A:569–575
- Arieli R, Ar A (1979) Ventilation of a fossorial mammal Spalax ehrenbergi in hypoxic and hypercapnic conditions. J Appl Physiol 47:1011-1017
- Arieli R, Ar A, Shkolnik A (1977) Metabolic responses of fossorial rodent (*Spalax ehrenbergi*) to simulated burrow conditions. Physiol Zool 50:61–75
- Armstrong HG (1952) Boiling of body fluids. In: Dill DB, Adolph EF, Wilber CG (eds) Principles and practices of aviation medicine, 3rd edn. Williams and Wilkins, Baltimore
- Armstrong JD, Priede IG, Lucas MC (1992) The link between respiratory capacity and changing metabolic demands during growth of northern pike, *Esox lucius*. J Fish Biol 41:65–75
- Armstrong W (1970) Rhizosphere oxidation in rice and other species: a mathematical model based on the oxygen flux component. Physiol Plant 23:623-630
- Arp AJ (1991) The role of heme compounds in sulfide tolerance in the echurian worm *Urechis caupo*. In: Vinogradov D, Kapp C (eds) Structure and function of invertebrate oxygen proteins. Springer, Berlin Heidelberg New York, pp 337-346
- Arrigo KR, Worthen DL, Lizotte MP, Dixon P, Dieckmann G (1997) Primary production in Antarctic ice. Science 276:394–397
- Arthur WB (1997) Wonders: how fast is technology evolving? Sci Am 276:89-91
- Arudpragasm KD, Naylor E (1964a) Gill ventilation and the role of reversed respiratory currents in *Carcinus maenas* (L). J Exp Biol 41:299–307
- Arudpragasm KD, Naylor E (1964b) Gill ventilation volumes, oxygen consumption and respiratory rhythms in *Carcinus maenas* (L). J Exp Biol 41:2307–2321
- Aschoff J, Pohl H (1970) Rhythmic variations in energy metabolism. Fed Proc Am Soc Exp Biol 29:1541–1552

- Assali NS, Kirschbaum TH, Dilts PV (1968) The effects of hyperbaric oxygen on uteroplacental and foetal circulation. Circ Res 22:573–588
- Atchley WR, Hall BK (1991) A model for development and evolution of complex morphological structures. Biol Rev 66:101–157
- Atkins D (1936) On the ciliary mechanism and interrelationship of Lamellibranches. Part I. New observations on sorting mechanisms. Q J Microsc Sci 79:181–309
- Atkinson RJA, Taylor AC (1988) Physiological ecology of burrowing decapods. In: Fincham A, Rainbow PS (eds) Aspects of the biology of decapod Crustacea. Oxford University Press, Oxford, pp 201-226
- Atkinson RJA, Pelster B, Bridges CR, Taylor AC, Morris S (1987) Behavioural and physiological adaptations to a burrowing life-style in the snake blenny, *Lumpenus lampretaeformis*, and the red band fish, *Cepola rubescens*. J Fish Biol 31:639–659
- Augee ML, Elsner RW, Gooden BA, Wilson PR (1970/71) Respiratory and cardiac responses of a burrowing animal, the echidna. Respir Physiol 11:327–334
- Averof, Akam M (1995) Insect-crustacean relationships: insights from comparative developmental and molecular studies. Philos Trans R Soc Lond B 347:293-303
- Avery ME (1968) The lung and its disorders in the newborn infant, 1st edn. WB Saunders, Philadelphia
- Avery ME, Mead J (1959) Surface properties in relation to atelectiasis and hyaline membrane disease. Am J Dis Child 97:517–523
- Axelsson M, Farrell AP, Nilsson S (1990) Effects of hypoxia and drugs on the cardiovascular dynamics of the Antlantic hagfish *Myxine glutinosa*. J Exp Biol 151:297–316
- Axelsson M, Davison W, Forster ME, Farrell AP (1992) Cardiovascular responses of the redblooded Antarctic fishes, *Pagothenia bernachii* and *P. borchgrevinki*. J Exp Biol 167:179-201
- Azar C, Rodh N (1997) Targets for stabilization of atmospheric CO2. Science 276:1818-1819
- Bacon BJ, Gilbert RG, Kaufmann P (1984) Placental anatomy and diffusing capacity in guinea pigs following long term maternal hypoxia. Placenta 5:475-488
- Bader R (1937) Bau, Entwicklung und Funktion des akzessorischen Atmungsorgans der Labyrinthfische. Z Wiss Zool 149:323-401
- Bailey L (1954) The respiratory currents in the tracheal system of the adult bee. J Exp Biol 31:589-595
- Baird B, Cook SF (1962) Hypoxia and reproduction in Swiss mice. Am J Physiol 202:611-615
- Bainerd EL (1994) The evolution of lung-gill bimodal breathing and the homology of vertebrate respiratory pumps. Am Zool 34:289–299
- Baker CL (1949) Comparative anatomy of the aortic arches of the urodeles and their relation to respiration and degree of metamorphosis. J Tenn Acad Sci 24:12–40
- Baker MA (1982) Brain cooling in endotherms in heat and exercise. Annu Rev Physiol 44:85–96 Baker RR (1978) The evolutionary ecology of animal migration. Hodder, London
- Baker VR, Strom RG, Gulick VC, Kargel JS, Komatsu G, Kale VS (1991) Ancient oceans, ice sheets and hydrological cycles on Mars. Nature (Lond) 352:589–594
- Bakhle YS (1975) Pharmocokinetic function of the lung. In: Junod AF, Haller R (eds) Lung metabolism. Academic Press, New York, pp 293–299
- Bakker RT (1975) Dinosaur renaissance. Sci Am 232:58-78
- Balasch J, Musqueras S, Palacios L et al. (1976) Comparative haematology of some Falconiformes. Condor 78:258–273
- Balch CC, Campling RC (1965) Rate of passage of digesta through the ruminant digestive tract. In: Dougherty RW (ed) Physiology of digestion in the ruminant. Butterworths, Washington, DC, pp 108–146
- Baldauf SL, Palmer JD (1993) Animals and fungi are each other's closest relatives: congruent evidence from multiple proteins. Proc Natl Acad Sci USA 90:11558–11562

Baldridge GW, Gerard RW (1933) Extra respiration of phagocytosis. Am J Physiol 103:235-236

- Baldwin KM, Winder WW (1977) Adaptive responses in different types of muscle fibers to endurance exercise. Ann NY Acad Sci 301:411–423
- Ballard PL, Ballard RA (1974) Cytoplasmic receptor for glucocoticoids in lung of the human foetus and neonate. J Clin Invest 53:477–486
- Ballew C, Haas JD (1986) Hematologic evidence of fetal hypoxia among newborn infants at high altitude in Bolivia. Am J Obstet Gynecol 155:166–169

- Ballingand JL, Kobzik L, Han X, Kaye DM et al. (1995) Nitric oxide dependent parasymphathetic signaling is due to activation of constitutive endothelial (type III) nitric oxide synthetase in cardiac myocytes. J Biol Chem 270:14582–14586
- Ballintijn CM (1972) Efficiency, mechanics, and motor control of fish respiration. Respir Physiol 14:124–141
- Ballintijn CM (1982) Neural control of respiration in fishes and mammals. In: Adink ADF, Spronk N (eds) 3rd congress of ESCPB, vol I. Pergamon Press, Oxford, pp 127–140
- Balon TW, Nadler JL (1997) Evidence that nitric oxide increases glucose transport in skeletal muscle. J Appl Physiol 82:359-363
- Balter M (1996) Looking for clues to the mystery of life on Earth. Science 273:870-872
- Banzett RB, Butler PJ, Nations CS, Barnas GM, Lehr JL, Jones JH (1987) Inspiratory aerodynamic valving in goose lungs depends on gas density and velocity. Respir Physiol 70:287-300
- Banzett RB, Nations CS, Wang N, Fredberg JJ, Butler JP (1991) Pressure profiles show features essential to aerodynamic valving in geese. Respir Physiol 84:295–309
- Banzett RB, Nations CS, Wang N, Butler PJ, Lehr JL (1992) Mechanical interdependence of wing beat and breathing in starlings. Respir Physiol 89:27-36
- Barcroft J, Barron DH (1946) Observations upon the form and relations of the maternal and foetal vessels in the placenta of sheep. Anat Rec 94:569–595
- Bard H, Fouton JC, Robillard JE, Cornet A, Soukini MA (1978) Red cell oxygen affinity in foetal sheep: role of 2,3-DPG and adult haemoglobin. J Appl Physiol 45:7-10
- Bardack D (1991) First fossil hagfish (Myxinoidea): a record from the Pennsylvanian of Illinois. Science 254:701–703
- Barham EG (1966) Deep-scattering layer migration and composition: observations from a diving saucer. Science 151:1399–1416
- Barel CDN (1993) Concepts of an architectonic approach to transformation morphology. Acta Biotheor 41:345–381
- Barel CDN, Anker GC, Witte F, Hoogerhoud RJC, Goldschmidt T (1989) Constructional constraint and its ecomorphological implications. Acta Morphol Neerl Scand 27:83-109
- Barman SA, McCloud LL, Catravas JD, Ehrhart IC (1996) Measurement of pulmonary blood flow by fractal analysis of flow heterogeneity in isolated canine lungs. J Appl Physiol 81:2039–2045
- Barnard KH (1943) Revision of the indigenous fresh water of the S.W. Cape region. Ann S Afr Mus 36:101-262
- Barnes P (1990) Reactive oxygen species and airway inflammation. Free Radical Biol Med 9:235–243
- Barnhart MC (1986a) Respiratory gas tensions and gas exchange in active and dormant land snails, *Otala lactea*. Physiol Zool 59:733–745
- Barnhart MC (1986b) Control of acid base status in active and dormant land snail, Otala lactea (Pulmonata, Helicidae). J Comp Physiol B156:347–354
- Barnhart MC (1986c) Hemocyanin function in active and dormant land snails, Otala lactea. Physiol Zool 59:725-732
- Barnola JM, Raynaud D, Korotkevich YS, Lorius C (1987) Vostok ice core provides 160 000-year record of atmospheric CO<sub>2</sub>. Nature (Lond) 329:408–414
- Barnsley MF, Massopust P, Strickland H, Sloan AD (1987) Fractal modelling of biological structure and fuction. Ann NY Acad Sci 504:179–194
- Bar-Nun A, Shaviv A (1975) Dynamics of the chemical evolution of the Earth's primitive atmosphere. Icarus 24:197–210
- Barrell J (1916) Influence of Silurian-Devonian climates on the rise of air breathing vertebrates. Bull Geol Soc Am 27:387–436
- Barrell BG, Bankier AT, Drouin J (1979) A different genetic code in human mitochondria. Nature (Lond) 282:189–194
- Barron DH, Meschia G (1954) A comparative study of the exchange of the respiratory gases across the placenta. Cold Spring Harbour symposium on quantitative biology, vol XIX, the mammalian fetus: physiological aspects of development. Long Island Biological Association, New York, pp 93–101
- Barron DH, Metcalfe J, Meschia G, Huckabee W, Hellegers A, Prystowsky H (1964) Adaptations of pregnant ewes and their fetuses to high altitude. In: Weihe WH (ed) The physiological effects of high altitude. Pergamon Press, Oxford, pp 115–129

- Barron ET, Thompson SL, Schneider SH (1981) An ice-free Cretaceous? results from climate model simulations. Science 212:501–514
- Bartels H (1970) The diffusion capacity of the placenta. In: Neuberger A, Tatum EL (eds) Frontiers of biology, vol 17, prenatal respiration. North Holland, London, pp 61–67
- Bartels H, Metcalfe J (1965) Some aspects of the comparative physiology of placental gas exchange. Int Union Physiol Sci 4:34–52
- Bartels H, Welsch U (1983) Freeze-fracture study of the turtle lung. 1. Intercellular junctions in the air-blood barrier of *Pseudemys scripta*. Cell Tissue Res 231:157–172
- Bartels H, Welsch U (1984) Freeze-fracture study of the turtle lung. 2. Rod-shaped particles in the plasma membrane of a mitochondria-rich pneumocyte in *Pseudemys (Chrysemys) scripta*. Cell Tissue Res 236:453–467
- Bartels H, Haller G, Reinhardt W (1966) Oxygen affinity of chicken blood before and after hatching. Respir Physiol 1:345–356
- Barthelemy L (1987) Oxygen poisoning. In: Dejours P (ed) Comparative physiology of environmental physiology, vol 2. Karger, Basel, pp 152–162
- Bartholomew GA (1982a) Scientific innovation and creativity: a zoologists point of view. Am Zool 22:227–235
- Bartholomew GA (1982b) Energy metabolism. In: Gordon MS (ed) Animal physiology: principles and adaptations. Macmillan, New York, pp 57–110
- Bartholomew GA (1988) Interspecific comparison as a tool for ecological physiology. In: Feder ME, Bennet AF, Burggren WW, Huey RB (eds) New directions in ecological physiology. Cambridge University Press, Cambridge, pp 11–37
- Bartholomew GA, Barnhart CM (1984) Tracheal gases, respiratory gas exchange, body temperature and flight in some tropical cicadas. J Exp Biol 111:131–144
- Bartholomew GA, Lighton JRB (1986) Oxygen consumption during hover-feeding in freeranging Anna hummingbirds. J Exp Biol 123:191–199
- Bartholomew GA, Leitner P, Nelson JE (1964) Body temperature, oxygen consumption, and heart rate in three species of Australian flying foxes. Physiol Zool 37:179–198
- Bartlett D, Mortola JP, Doll EJ (1986) Respiratory mechanics and control of the ventilatory cycle in the garter snake. Respir Physiol 64:13–27
- Barton M, Elkins K (1988) Significance of aquatic surface respiration in the comparative adaptation of two species of fishes (*Notropis chrysocephalus* and *Fundulus catenus*) to headwater environments. Trans Ky Acad Sci 49:69–73
- Bartosch R, Feldberg W, Nagel E (1933) Weitere Versuche über Freiwerden eines histaminähnlichen Stoffes aus der durchströmten Lunge sensibilisierter Meerschweinchen beim Auslösen einer anaphylaktischen Lungenstarre. Pfluegers Archiv Gesamte Physiol Henschen Tiere 231:616–629
- Basalla G (1989) The evolution of technology. Cambridge University Press, New York
- Baskin DG, Detmers PA (1976) Electron microscopic study of the gill bars of *Amphioxus* (*Brachiostoma californiense*) with special reference to neurocilliary control. Cell Tissue Res 166:167–178
- Basset JE, Wiederhielm CA (1984) Postnatal changes in hematology of the bat Antrozous pallidus. Comp Biochem Physiol 78A:737-742
- Bassingthwaighte JB (1988) Physiological heterogeneity: fractals link determinism and randomness in structures and functions. News Physiol Sci 3:5–9
- Bassingthwaighte JB, King RB, Roger SA (1989) Fractal nature of regional myocardial blood flow heterogeneity. Circ Res 65:578–590
- Bassingthwaighte JB, Liebovitch LS, West BJ (1994) Fractal physiology. Oxford University Press, New York
- Bast A, Haenen G, Doelman C (1991) Oxidants and antioxidants: state of the art. Am J Med 91:S2–S13
- Bastacky J, Hook GR, Finch GL, Goerke J et al. (1987) Low temperature scanning electron microscopy of frozen hydrated mouse lung. Scanning 9:57–70
- Bastacky J, Goerke J, Lee CYC, Yager D et al. (1993) Alveolar lining liquid layer is thin and continuous: low temperature SEM of normal rat lung. Am Rev Respir Dis 147: 148

- Bastacky J, Lee CYC, Goerke J, Koushafar H et al. (1995) Alveolar lining layer is thin and continuous: low temperature scanning electron microscopy of rat lung. J Appl Physiol 79:1615–1628
- Bates JHT (1993) Stochastic model of the pulmonary airway tree and its implications for bronchial responsiveness. J Appl Physiol 75:2493-2499
- Batterton CV, Cameron JN (1978) Characteristics of resting ventilation and response to hypoxia, hypercapnia and emersion in the blue crab *Callinectes sapidus* (Rathbun). J Exp Zool 203:403-418
- Baudrimont A (1955) Organization générale du poumon et structure des alvéoles pulmonaires des vertébrés (amphibiéns, reptiliens, mammiféres) considérés dans leurs rapports avec la mécanique respiratoire, la circulation pulmonaire fonctionelle et l'activité métabolique de ces animaux. Arch Anat Histol Embryol 38:99–136
- Bauer C (1974) On the respiratory function of haemoglobin. Rev Physiol Biochem Pharmacol 70:1-31
- Baum DA, Larson A (1991) Adaptation reviewed: a phylogenetic methodology for studying character macroevolution. Syst Zool 40:1–18
- Baumann FH, Baumann R (1977) A comparative study of the respiratory properties of bird blood. Respir Physiol 31:333-343
- Baumgarten-Schumann D, Piiper J (1968) Gas exchange in the gills of resting unanaethetized dogfish, *Scyliorhinus*. Respir Physiol 5:317-325
- Bazzaz FA, Fajer ED (1992) Plant life in a CO<sub>2</sub>-rich world. Sci Am 266:18–24
- Beachy CK, Bruce RC (1992) Lunglessness in plethodontid salamanders is consistent with the hypothesis of a mountain stream origin: a response to Ruben and Boucot. Am Nat 139:839–847
- Beadle LC (1932) Scientific results of the Cambridge Expedition to the East African Lakes 1930– 31. 4. The waters of some East African Lakes in relation to their fauna and flora. J Linn Soc (Zool) 38:157–211
- Beadle LC (1957) Respiration in the African swampworm Alma emini Mich. J Exp Biol 34:1-10
- Beadle LC (1974) The inland waters of tropical Africa: an introduction to tropical limnology. Longman, London
- Beament JWL (1960) Physical models in biology. In: Beament JWL (ed) Models and analogues in biology. Symp Soc Exp Biol No 14. Academic Press, New York, pp 66–123
- Beattie J, Smith AH (1975) Metabolic adapatations of the chick embryo to chronic hypoxia. Am J Physiol 228:1346–1350
- Bech C, Johansen K (1980) Ventilation and gas exchange in the mute swan, *Cygnus olor*. Respir Physiol 39:285–295
- Bech C, Rautenberg W, May B (1985) Ventilatory O<sub>2</sub> extraction during cold exposure in the pigeon (*Columba livia*). J Exp Biol 116:499–502
- Becker HO, Bohme W, Perry SF (1989) The lung morphology of lizards (Reptilia:Varaniidae) and its taxonomic-phylogenetic meaning. Bonn Zool Beitr 40:27–56
- Becker V (1963) Funktionelle Morphologie der Placenta. Arch Gynaekol 198:3-28
- Bedwani JR, Morley PB (1974) Increased inactivation of prostaglandin E<sub>2</sub> by the rabbit lung during pregnancy. Br J Pharmacol 50:459–460P

Behrensmeyer AK, Kidwell SM (1985) Taphonomy and paleobiology. Paleobiology 11:105-119

- Beitinger TL, Pettit MJ (1984) Comparison of low oxygen avoidance in a bimodal breather, *Erpetoichthys calabaricus* and an obligate water breather, *Percina caprodes*. Environ Biol Fishes 11:235–240
- Belanger LF (1940) A study of the histological structure of the respiratory portion of the lungs of aquatic mammals. Am J Anat 67:437–461
- Belkin DA (1964) Variation in heart rate during voluntary diving in the turtle *Pseudemys* concinna. Copeia 2:321-330
- Belkin DA (1968) Aquatic respiration and underwater survival of two fresh water turtle species. Respir Physiol 4:1–14
- Belkin S, Douglas CN, Jannasch HW (1986) Symbiotic assimilation of CO<sub>2</sub> in two hydrothermal vent animals, the mussel *Bathymodiolus thermophilus* and the tube worm *Riftia pachyptila*. Biol Bull 170:110–121

- Bell PB, Stark-Vancs VI (1983) SEM study of the microarchitecture of the lung of the giant salamnder *Amphiuma tridactylum*. Scanning Electron Microsc 1983:449–456
- Bellairs R, Griffiths I, Bellairs AA (1955) Placentation in the adder, *Vipera berus*. Nature (Lond) 176:657–658
- Belman BW (1975) Some aspects of the circulatory physiology of the spiny lobster *Panulirus interruptus*. Mar Biol 29:295–305
- Bemis WE, Lauder GV (1986) Morphology and function of the feeding apparatus of the lungfish, Lepidosiren paradoxa (Dipnoi). J Morphol 187:81–108
- Ben-Avraham Z (1981) The movement of the continents. Am Sci 69:291-299
- Bender HG (1974) Placenta-Insuffizienz. Morphometrische Untersuchungen am Modell der Rhesus-Placenta. Arch Gynaekol 216:289–300
- Bender L (1992) The human body: its mysteries and marvels. Bramley Books, Godalming, Surrey
- Bender ML (1984) On the relationship between ocean chemistry and atmospheric PCO<sub>2</sub> during the Cenozoic. In: Climate processes and climate sensitivity. Geophys Monogr 29:352–359
- Benedict FG (1938) Vital energetics: a study in comparative basal metabolism. Carnegie Institute, Washington, DC, Publ No 503
- Bennett AF (1978) Activity metabolism in the lower vertebrates. Annu Rev Physiol 400:447–469 Bennett AF (1982) The energetics of reptilian activity. In: Gans C, Pough FH (eds) Biology of the

Reptilia, physiological ecology. Academic Press, New York, pp 155–199

- Bennett AF (1988) Structural and functional determinates of metabolic rate. Am Zool 28:699-708
- Bennett AF, Dawson WR (1976) Metabolism. In: Gans C, Dawson WR (eds) Biology of Reptilia, vol 5. Academic Press, New York, pp 127–223
- Bennett AF, Ruben JA (1979) Endothermy and activity in vertebrates. Science 206:649-655
- Bennett AF, Tenney SM (1982) Comparative mechanics of mammalian respiratory system. Respir Physiol 49:131–140
- Bennett AF, Wake MH (1974) Metabolic correlates of activity in the caecilian, *Geotrypetes* seraphini. Copeia 1874:764-769
- Bennett MB (1988) Morphometric analysis of the gills of the European eel, Anguilla anguilla. J Zool (Lond) 215:549–560
- Bentley PJ, Shield JW (1973) Ventilation of toad lungs in absence of the buccopharyngeal pump. Nature (Lond) 243:538–539
- Benton MJ (1993) Late Triassic extinctions and the origin of the dinosaurs. Science 260:769–770 Benton MJ (1995) Diversification and extinction in the history of life. Science 268:52–58
- Berg K (1951) On the respiration of some molluscs from running and stagnant water. Ann Biol 33:561–567
- Berg T, Steen JB (1965) Physiological mechanisms for aerial respiration in the eel. Comp Biochem Physiol 15:469-484
- Berger AJ (1961) Bird study, 1st edn. Dover Publications, New York
- Berger M, Hart JS (1974) Physiology and energetics of flight. In: Farner DS, King JR (eds) Avian biology, vol 4. Academic Press, New York, pp 415–477
- Berger M, Roy OZ, Hart JS (1970) The coordination between respiration and wing beats in birds. Z Vergl Physiol 66:190–200
- Berger PJ, Walker AM, Horne R, Brodecky V, Wikinson MH et al. (1986) Phasic respiratory activity in the foetal lamb during late gestation and labour. Respir Physiol 65:55-68
- Berger WH, Spitzy A (1988) History of atmospheric CO<sub>2</sub>: constraints from the deep-sea records. Paleoceanography 3:401–411
- Berker LV, Marshall LC (1965) The origin and rise of oxygen concentration in the Earth's atmosphere. J Atmos Sci 22:225-261
- Berkner LV, Marshall LC (1965) On the origin and rise of oxygen concentration in the Earth's atmosphere. J Atmos Sci 22:225–261
- Berner RA (1963) Electrodes studies of hydrogen sulfide in marine sediments. Geochim Cosmochim Acta 27:563-575
- Berner RA (1991) Model for atmospheric CO<sub>2</sub> over the Phanerozoic. Am J Sci 291:339–376
- Berner RA (1997) The rise of plants and their effect on weathering and atmospheric CO<sub>2</sub>. Science 276:544–545

- Berner RA, Canfield DE (1989) A new model for atmospheric oxygen over Phanerozoic time. Am J Sci 289:333–361
- Bernhardt R (1995) Cytochrome P<sub>450</sub>: structure, function, and generation of reactive oxygen species. Rev Physiol Biochem Pharmacol 127:137–221
- Bernstein MH (1987) Respiration in flying birds. In: Seller TJ (ed) Bird respiration, vol II. CRC Press, Boca Raton, Florida, pp 43-73
- Bernstein MH (1989) Temperature and oxygen supply in the avian brain. In: Wood SC (ed) Comparative pulmonary physiology: current concepts. Marcel Dekker, New York, pp 343– 368
- Bernstein MH (1990) Avian respiration and high altitude tolerance. In: Sutton JR, Coates GC, Remmers JE (eds) Hypoxia: the adaptations. BC Decker, Burlington, Ontario, pp 30-40
- Bernstein MH, Schmidt-Nielsen K (1976) Vascular responses and foot temperature in pigeons. Am J Physiol 226:1350–1355
- Bernstein MH, Duran HL, Pinshow B (1984) Extrapulmonary gas exchange enhances brain oxygen in pigeons. Science 226:564–566
- Berry EM, Edmonds JF, Wyllie JH (1971) Release of prostaglandin  $E_2$  and unidentified factors from ventilated lungs. Br J Surg 58:189–192
- Berry WBN, Wilde P (1978) Progressive ventilation of the oceans: an explanation for the distribution of the lower Paleozoic black shales. Am J Sci 278:257-275
- Berschick P, Bridges CR, Grieshaber MK (1987) The influence of hyperoxia, hypoxia and temperature on the respiratory physiology of the intertidal rockpool fish, *Gobius cobitis pallas*. J Exp Biol 130:369–387
- Bertin L (1958) Organes de la respiration aérienne. Grassé PG (ed) Traite de zoologie, vol 13, pt 2. Massonet, Paris, pp 1363–1398
- Betke K (1958) Hämatologie der ersten Lebenszeit. Ergeb Inn Med Kinderheilkd 9:437-509
- Bettex-Galland M, Hughes GM (1973) Contractile filamentous material in the pillar cells of fish gills. J Cell Sci 13:359–366
- Betticher DC, Geiser J, Tempini A (1991) Lung diffusing capacity and red blood cell volume. Respir Physiol 85:271–278
- Betts RA, Cox PM, Lee SE, Woodward FI (1997) Contrasting physiological and structural vegetation feedbacks in climate change simulations. Nature (Lond) 387:196-799
- Bevelander G (1934) The gills of *Amia calva* specialized for respiration in an oxygen deficient habitat. Copeia 3:123–127
- Bidani A, Crandall ED (1988) Velocity of CO<sub>2</sub> exchanges in the lungs. Annu Rev Physiol 50:639–652
- Bignon J (1975) Several plasma proteins demonstrated in lung tissue by immuno-electron microscopy using the peroxidase coupling technique. In: Junod AF, Haller R (eds) Lung metabolism. Academic Press, New York, pp 425-431
- Billet FS, Courtenay TH (1973) A stereoscan study of the origin of ciliated cells in the embryonic epidermis of *Amblyostoma mexicanum*. J Embryol Exp Morphol 29:549–558
- Birks EK, Mathieu-Costello O, Fu Z, Tyler WS, West JB (1997) Very high pressures are required to cause stress failure of pulmonary capillaries in thoroughbred racehorses. J Appl Physiol 82:1584–1592
- Bishop SC (1943) Handbook of salamanders. Comstock Publications, Ithaca
- Bishop WW, Trendall AF (1967) Erosion surfaces, tectonics, and volcanic activity in Uganda. Q J Geol Soc Lond 122:385–420
- Black CP, Tenney SM (1980) Oxygen transport during progressive hypoxia in bar-headed geese (Anser indicus) acclimatized to sea-level and 5600 m. In: Piiper J (eds) Respiratory function in birds, adult and embryonic . Springer, Berlin Heidelberg New York, pp 79–83
- Black CP, Tenney SM (1980) Oxygen transport during progressive hypoxia in high altitude and sea level water-fowl. Respir Physiol 39:217–239
- Black CP, Tenney SM, Kroonenburg MV (1978) Oxygen transport during progressive hypoxia in bar-headed geese (Anser anser) acclimated to sea level and 5600 m. In: Piiper J (ed) Respiratory function in birds adult and embryonic. Springer, Berlin Heidelberg New York, pp 79–83
- Black LL, Wiederhielm CA (1976) Plasma oncotic pressures and hematocrit in the intact, anaesthetized bat. Microvasc Res 12:55–58

- Blackburn DG (1982) Evolutionary origins of viviparity in the Reptilia. 1. Sauria. Amphibia-Reptilia 3:185–205
- Blackburn DG (1993) Chorioallantoic placentation in squamate reptiles: structure, function, development, and evolution. J Exp Zool 266:414-430
- Blackburn DG, Evans HE (1986) Why are there no viviparous birds? Am Nat 128:165-190
- Blair HS (1994) Molecular evidence for the origin of birds. Proc Natl Acad Sci USA 91:2621-2624
- Bland RD (1990) Lung epithelial ion transport and fluid movement during the perinatal period. Am J Physiol 259:L30–L37
- Bland RD, McMillan DD, Bressack MA, Dong L (1980) Clearance of liquid from lungs of newborn rabbits. J Appl Physiol 49:171–177
- Bland RD, Hansen TN, Haberkern CM, Bresack MA et al. (1982) Lung fluid balance in lambs before and after birth. J Appl Physiol 53:992-1004
- Blatchford JG (1971) Hemodynamics of *Carcinus maenas* (L.). Comp Biochem Physiol 9A:193-202
- Blenkarn GD, Hayes JA (1970) Bilateral lung lavage with hyperbarically oxygenated saline in dogs. J Appl Physiol 29:786–793
- Bleyl U, Büsing CM (1971) Perpetuation des Shocks durch die Schocklunge. Z Prakt Anaesth Wiederbeleb 6:249–262
- Bliss DE (1979) From sea to tree: saga of a land crab. Am Zool 19:385-410
- Bliss DE, Mantel LH (1968) Adaptations of crustaceans to land: a summary analysis of new findings. Am Zool 8:673-685
- Bloch EH (1962) A quantitative study of the hemodynamics in the living microvascular system. J Anat 110:125–145
- Block BA (1987) Strategies for regulating brain and eye temperatures: a thermogenic tissue in fish. In: Dejours P, Bolis L, Taylor CR, Weibel ER (eds) Comparative physiology: life in water and on land. Fidia Research Series, vol 9. Liviana Press, New York, pp 401–420
- Block BA (1991a) Endothermy in fish: thermogenesis, ecology and evolution. In: Hochachka PW, Mommsen T (eds) Elsevier, Amsterdam, pp 269–298
- Block BA (1991b) Evolutionary novelties: how fish have built a heater out of muscle. Am Zool 31:726-742
- Block ER, Patel JM, Angelides KJ, Sherdan NP, Garg C (1986) Hyperoxia reduces plasma membrane fluidity: a mechanism for endothelial cell dysfunction. J Appl Physiol 60:826– 835
- Blomqvist CG, Stone HL (1983) Cardiovascular adjustments to gravitational stress. In: Fenn WO, Rahn H (eds) Handbook of physiology: the cardiovascular system, peripheral circulation and organ blood flow, sect 2, vol III, Part 2. Am Physiol Soc, Bethesda pp 1025–1063

Blum HF (1955) Time's arrow and evolution. Princeton University Press, Princeton

- Bock WJ, von Wahlert G (1965) Adaptation and the form-function complex. Evolution 19: 269–299
- Bøe F (1954) Vascular morphology of the human placenta cold Spring Harbour Symposium. Q J Biol 19:29–35
- Boggs DF, Kilgore DL, Birchard GF (1984) Respiratory physiology of burrowing mammals and birds. Comp Biochem Physiol 77A:1-7
- Böhlke JE, Chaplin CCG (1968) Fishes of the Bahamas and adjacent tropical waters. Academy of Natural Sciences, Philadelphia
- Bohr C (1909) Ueber die spezifische Tätigkeit der Lungen bei der respiratorischen Gasaufnahme. Scand Arch Physiol 22:221–280
- Boland EJ, Olson KR (1979) Vascular organization of the catfish gill filament. Cell Tissue Res 198:487–500
- Bolin B, Houghton J, Filho LGM (1994) Radiative forcing of climatic change: the 1994 report of the Scientific Assessment Working Group of Intergovernmental Panel on Climatic Change (IPCC). WHO and UNEP, Meteorological Office Marketing Cummunications Studio, Washington, DC, 48pp
- Bond AN (1960) An analysis of the response of salamander gills to changes in the oxygen concentration of the medium. Dev Biol 2:1-20
- Bond CF, Gilbert PW (1958) Comparative study of blood volume in representative aquatic and nonaquatic birds. Am J Physiol 194:519–521

Bone Q, Pulsford A, Chubb AD (1981) Squid mantle muscle. J Mar Biol Assoc UK 61:327-342

- Bone Q, Brown ER, Travers G (1994) On the respiratory flow in the cuttlefish, *Sepia officinalis*. J Exp Biol 194:153–165
- Bonner JT (1988) The evolution of complexity. Princeton University Press, Princeton
- Booth DT (1995) Oxygen availability and embryonic development in sand snail (*Polinices sordidus*) egg masses. J Exp Biol 198:241-247
- Booth JH (1978) The distribution of blood flow in the gills of fish: application of a new technique to rainbow trout (*Salmo gairdnen*). J Exp Biol 73:119-129
- Booth JH (1979) The effect of oxygen supply, epinephrine and acetycholine on the distribution of blood flow in trout gills. J Exp Biol 83:31–39
- Borden MA (1931) A study of the respiration and the function of haemoglobin in *Planorbis* corneus and Arenicola marina. J Mar Biol Assoc UK 17:709-738
- Borell U, Fernstrom I (1962) The shape of the foetal chest during its passage through the birth canal: a radiolographic study. Acta Obstet Gynecol Scand 41:213-222
- Borradaille LA, Potts FA, Eastham LES, Saunders JT (1963) Invertebrata, 4th edn (Revised by GA Kerkut). Cambridge University Press, Cambridge
- Borteux S (1993) Properties and biological functions of the NTH and FPG proteins of *Escherichia coli*: two DNA glycosylases that repair oxidative damage in DNA. Photochem Photobiol B 19:87-96
- Bouchet JY, Truchot JP (1985) Effects of hypoxia and L-lactate on the hemocyanin oxygen affinity of the lobster, *Homarus vulgaris*. Comp Biochem Physiol 80A:69-73
- Boucot AJ, Gray J (1982) Paleozoic data of climatological significance and their use for interpreting Silurian-Devonian climate. Geophysics Study Commitee, National Academy Press, Washington, DC, pp 189–198
- Bouhuys A (1974) Breathing: physiology, environment and lung disease. Grune and Stratton, New York
- Boulière F (1975) Mammals, small and large: the ecological implication of size. In: Golley FB, Petrusewicz K (eds) Small mammals: their productivity and population dynamics. Cambridge University Press, Cambridge, pp 1–8
- Bourne GB, Redmond JR (1977) Hemodynamics in the pink albarone, *Haliotis corrugata*. I. Pressure relations and pressure gradients in intact animals. J Exp Zool 200:9–16
- Boussaad S, Tazi A, Leblac RM (1997) Chlorophyll *a* dimer: a possible primary electron donor for the photosynthesis II. Proc Natl Acad Sci USA 94:3404-3506
- Boutilier RG (1990) Respiratory gas tensions in the environment. In: Boutilier RG (ed) Advances in comparative and environmental physiology, vol 6: Vertebrate gas exchange from environment to cell. Springer, Berlin Heidelberg New York, pp 1–13
- Boutilier RG, Toews DP (1981) Respiratory, circulatory and acid-base, adjustments to hypercapnia in a strictly aquatic and predominantly skin breathing urodele *Cryptobranchus alleganiensis*. Respir Physiol 46:177–192
- Boutilier RG, Randall DJ, Shelton G, Toews DP (1979a) Acid-base relationships in the blood of the toad *Bufo marinus*. I. The efects of environmental CO<sub>2</sub>. J Exp Biol 84:289-302
- Boutilier RG, Randall DJ, Shelton G, Toews DP (1979b) Acid-base relationships in the blood of the toad *Bufo marinus*. III. The effects of burrowing. J Exp Biol 82:357-365
- Boutilier RG, Heming TA, Iwama GK (1984) Physicochemical parameters for use in fish respiratory physiology. In: Hoar WS, Randall DJ (eds) Fish physiology, vol 10A. Academic Press, New York, pp 403–456
- Boutilier RG, Glass ML, Heisler N (1986) The relative distribution of pulmocutaneous blood flow in *Rana catesbeiana*: effects of pulmonary or cutaneous hypoxia. J Exp Biol 126:33–39
- Boutilier RG, West TG, Pogson GH, Mesa KA, Wells J, Wells MJ (1996) Nautilus and the art of metabolic maintenance. Nature (Lond) 382:535
- Bouverot P (1985) Adaptation to altitude-hypoxia in vertebrates. Springer, Berlin Heidelberg New York
- Bouverot P, Hildewein G, Oulhen P (1976) Ventilatory and circulatory O<sub>2</sub> convecion at 400 m in pigeons at neutral or cold temperature. Respir Physiol 28:371–385
- Bozinovic F (1993) Scaling basal and maximal metabolic rate in rodents and the aerobic capacity model for the evolution of endothermy. Physiol Zool 65:921–932
- Bradbury M (1979) The concept of blood-brain barrier. John Wiley, New York

- Bradford SM, Taylor AC (1982) The respiration of *Cancer pagurus* under normoxic and hypoxic conditions. J Exp Biol 97:273–288
- Brafield AE (1964) The oxygen content of interstitial water in sandy shores. J Anim Ecol 33:97– 116
- Brain JD (1985) Macrophages in the respiratory tract. In: Fishman AP, Fisher AB (eds) Handbook of physiology, vol 1. Circulation and nonrespiratory functions. American Physiological Society, Bethesda, pp 447–471
- Brainerd EL (1994) Lung ventilation in fishes and amphibians: the evolution of vertebrate airbreahing mechanisms. Am Zool 34:289–299
- Brainerd EL, Ditelberg JS, Bramble DM (1993) Lung ventilation in salamanders and the evolution of vertebrate air-breathing mechanisms. Biol J Linn Soc 49:163–183
- Brakenbury JH (1984) Physiological responses of birds to flight and running. Biol Rev 59:559– 575
- Brakenbury JH (1987) Ventilation of the lung-air sac system. In: Seller TJ (ed) Bird respiration vol. I. CRC Press, Boca Raton, pp 39–69
- Brakenbury JH (1991) Ventilation, gas exchange and oxygen delivery in flying and flightless bird. In: Woakes AJ, Grieshaber MK, Bridges CR (eds) Physiological strategies for gas exchange and metabolism. Cambridge University Press, Cambridge, pp 125–147
- Brakenbury JH, Amaku J (1990) Effects of combined abdominal and thoracic air-sac occlusion on respiration in domestic fowl. J Exp Biol 152:93–100
- Brakenbury JH, Avery P (1980) Energy consumption and ventilatory mechanisms in the exercising fowl. Comp Biochem Physiol 66A:439–445
- Brakenbury JH, Avery P, Gleeson M (1981) Respiration in exercising fowl. I. Oxygen consumption, respiratory rate, and respired gases. J Exp Biol 93:317–325
- Brakenbury JH, Darby C, El-Sayed MS (1989) Respiratory function in exercising fowl following occlusion of the thoracic air sacs. J Exp Biol 145:227–237
- Bramble DM, Carrier DR (1983) Running and breathing in mammals. Science 219:251-256
- Bramwell CD (1971) Aerodynamics of Pteranodon. J Linn Soc Biol 3:313-328
- Braulin EA, Wahler GM, Śwayze CR, Lucas RV, Fox IJ (1986) Myoglobin facilitated oxygen diffusion maintains mechanical function of mammalian cardiac muscle. Cardiovasc Res 20:627–636
- Braun F (1931) Beiträge zur Biologie und Atmungsphysiologie der Argyroneta aquatica. Zool Jahrb Abt Syst 62:175–262
- Bray AA (1985) The evolution of the terrestrial vertebrates: environmental and physiological considerations. Philos Trans R Soc Lond 309B:289–322
- Breeze RG, Wheeldon EB (1977) The cells of the pulmonary airways. Am Rev Respir Dis 116:705-777
- Brent R, Pedersen PF, Bech C, Johansen K (1984) Lung ventilation and temperature regulation in the European coot (*Fulica atra*). Physiol Zool 57:19–25
- Brett JR (1972) The metabolic demand for oxygen in fish, particularly salmonids and a comparison with other vertebrates. Respir Physiol 14:151–174
- Brett RA (1986) The ecology and behaviour of the naked mole rat (*Heterocephalus glaber* Ruppell) (Rodentia: Bathyergidae). PhD Diss, University of London, London
- Brigdes CR (1986) A comparative study of the respiratory properties and physiological function of haemocyanin in two burrowing and two non-burrowing crustaceans. Comp Biochem Physiol 83A:261–270
- Brigdes CR (1987) Environmental extremes the respiratory physiology of intertidal rockpool fish and sublittoral burrowing fish. Zool Beitr 30:65–84
- Bridges CR (1988) Respiratory adaptations in intertidal fish. Am Zool 18:79-96
- Bridges CR, Morris S (1986) Modulation of hemocyanin oxygen affinity by L-lactate; a role for other cofactors. In: Lizen B (ed) Invertebrate oxygen carriers. Springer, Berlin Heidelberg New York, pp 341–352
- Bridges CR, Kester P, Scheid P (1980) Tracheal volume in the pupa of the saturniid moth *Hyalophora cecropia* determined with inert gases. Respir Physiol 40:281–291
- Briggs J (1992) Fractals: The pattern of chaos. Simon and Schuster, New York
- Britton JC (1970) The Lucinidae (Mollusca: Bivalvia) of the western Antlantic Ocean. PhD Diss, The George Washington University, Washington, DC

- Brix O (1982) The adaptive significance of the reversed Bohr and Root shifts in blood from the marine gastropod, *Buccinum undulum*. J Exp Zool 221:27–36
- Brix O, Bardgard A, Cau A, Colosimo A, Condo SG, Giardina B (1989) Oxygen-binding properties of cephalopod blood with special reference to environmental temperatures and ecological distribution. J Exp Zool 252:34-42
- Brocher F (1920) Étude expérimentale sur le fonctionnement du vaisseau dorsal et sur la circulation du sang chez le Insectes. III. Le Sphinx convolvuli. Arch Zool Exp Gén 60:1-45
- Brocher F (1931) Le mécanisme de la respiration et celui de la circulation du sang chez les insectes. Arch Zool Exp Gén 74:25-32
- Brocklehurst WE (1960) The release of histamine and formation of slow-reacting substance (SRS-A) during anaphylactic shock. J Physiol (Lond) 151:416-425
- Broecker WS (1982) Glacial to interglacial changes in ocean chemistry. Prog Oceanogr 11:151– 197
- Brom TG, Dekker RWRJ (1992) Current studies of megapode phylogeny. Zool Verh (Leiden) 278:7–17
- Bromhall C (1987) Spider heart rates and locomotion. J Comp Physiol B 157:451-460
- Brooks D (1994) How to perform reduction. Philos Phenomen Res 54:84-99
- Brooks RE (1970) Ultrastructure of the physostomatous swim bladder of rainbow trout Salmo gairdneri. Z Zellforsch Mikrosk Anat 106:473-483
- Browman MW, Kramer DL (1985) *Pangasius sutchi* (Pangassidae), an air-breathing catfish that uses the swim bladder as an accessory respiratory organ. Copeia 1985:994–998
- Brown AC (1993) Variability in biological systems. S Afr J Sci 89:308-309
- Brown AC (1994) Is biology science? Trans R Soc S Afr 49:141-146
- Brown AC, Terwilliger NB (1992) Developmental changes in ionic and osmotic regulation in the Dungeness crab, *Cancer magister*. Biol Bull Mar Biol Lab Woods Hole 182:270–277
- Brown J (1995) Macroecology. University of Chicago Press, Chicago
- Brown JH (1981) Two decades of homage to Santa Rosalia: toward a general theory of diversity. Am Zool 21:877–888
- Brown JH, Marquet PA, Taper ML (1993) Evolution of body size: consequences of an energetic definition of fitness. Am Nat 142:573–584
- Brown JR, Doolittle WF (1995) Root of universal tree of life based on ancient aminoacyl-tRNA synthetase gene duplications. Proc Natl Acad Sci USA 92:2441–2445
- Brown ME, Hill RE (1996) Discovery of an extended atmosphere around Europa. Nature (Lond) 380:229–231
- Brown MJ, Olver RE, Ramsden CA, Strang LB, Walters DV (1983) Effects of adrenaline and of spontaneous labour on the secretion and absorption of lung liquid in foetal lamb. J Physiol (Lond) 344:137–152
- Brown SC (1975) Biomechanics of water pumping by *Chaetopterus variopedatus* Renier: Skeletomusculature and kinematics. Biol Bull 149:136–150
- Brown SC (1977) Biomechanics of water pumping by *Chaetopterus variopedatus* Renier: kinematics and hydrodynamics. Biol Bull 153:121-132
- Brown SC, McGee-Russel S (1971) Chaetopterus tubes: ultrastructural architecture. Tissue Cell 3:65-70
- Brown-Borg HM, Borg KE, Meliska CJ, Bartke A (1996) Dwarf mice and ageing process. Nature (Lond) 384:33
- Broyles RH (1981) Changes in the blood during amphibian metamorphosis. In: Gilbert Ll, Frieden E (eds) Metamorphosis: a problem in developmental biology, 2nd edn. Plenum Press, New York, pp 461–490
- Bruna EM, Fisher RN, Case TJ (1996) Morphological and genetic evolution appear decoupled in Pacific skinks (Squamata: Scincidae: Emoia). Proc R Soc Lond 263 B:681–688
- Brunori M (1975) Molecular adaptation to physiological requirements: the hemoblobin systems of trout. Curr Top Cell Regul 9:1–39
- Bruton MN (1979) The breeding biology and early development of *Clarias gariepinus* (Pisces: Clariidae) in Lake Sibaya, South Africa, with a review of breeding in species of subgenus *Clarias*. Trans Zool Soc Lond 35:1-45
- Bruton MN, Cabral AJP, Fricke H (1992) First capture of a coelacanth, *Latimeria chalumnae* (Pisces, latimeriidae) off Mozambique. S Afr J Sci 48:225–227

Bryan JD, Hill LG, Neill WH (1984) Interdependence of acute temperature preference and respiration in the plains minnows. Trans Am Fish Soc 113:557-562

Bshouty Z, Younes M (1990) Distensibility and pressure-flow relationship of the pulmonary circulation. II. Multibranched model. J Appl Physiol 68:1514–1527

- Bucher TL (1985) Ventilation and oxygen consumption in *Amazona viridigenalis*: a reappraisal of resting respiratory parameters in birds. J Comp Physiol B 155:269–276
- Bucher TL, Barnhart MC (1984) Varied egg gas conductance, air cell gas tensions and development in Agapornis roseicollis. Respir Physiol 55:277-294
- Bucher TL, Chappell MA (1989) Energy metabolism and patterns of ventilation in euthermic and torpid hummingbirds. In: Bech C, Reinertsen RE (eds) Physiology of cold adaptation in birds. Plenum Press, New York, pp 187–195
- Bucher TL, Chappell MA, Morgan KR (1990) The ontogeny of oxygen consumption and ventilation in the Adélie penguin, *Pygoscelis adelie*. Respir Physiol 82:269–388
- Buck JB (1948) The anatomy and physiology of the light organs in fire flies. Ann NY Acad Sci 49:397-482
- Buck JB (1962) Some physical aspects of insect respiration. Annu Rev Entomol 7:27-56
- Buck JB, Keister M (1955) Further studies of gas filling in the insect tracheal system. J Exp Biol 32:681–691
- Budgett JS (1900) Observations on *Polypterus* and *Protopterus*. Proc Cambr Philos Soc 10:236–240
- Bugge J (1960) The heart of the African lungfish, *Protopterus*. Vidensk Medd Dan Naturhist Foren 123:193–210
- Buick FJ, Gledhill N, Froese AB, Spriet L, Meyers EC (1984) Effect of induced erythrocythemia on aerobic work capacity. J Appl Physiol 48:636–642
- Bullard RW (1972) Physiological problems of space travel. Annu Rev Physiol 34:205-234
- Bünning E (1973) The physiological clock, 3rd edn. Springer, Berlin Heidelberg New York
- Burda H (1993) Evolution of eusociality in Bathyergidae: the case of the giant mole rats (*Cryptomys mechowi*). Naturwissenschaften 80:235–237
- Burg M, Green N (1977) Bicarbonate transport by isolated perfused rabbit proximal convoluted tubules. Am J Physiol 233:F307–F314
- Burger PJ, Bradley SE (1951) The general form of the circulation of the dogfish (Squalus acanthias). J Cell Comp Physiol 37:389-402
- Burger RE, Meyer M, Werner G, Scheid P (1979) Gas exchange in the parabronchial lung of birds: experiments in unidirectionally ventilated ducks. Respir Physiol 36:19–37
- Burggren WW (1975) A quantitative analysis of ventilation tachycardia and its control in two chelonians *Pseudemys scripta* and *Testudo graeca*. J Exp Biol 63:367–380
- Burggren WW (1977) Circulation during intermittent lung ventilation in the garter snake, Thamnophis. Can J Zool 55:1720-1725
- Burggren WW (1979) Bimodal gas exchange during variation in environmental oxygen and carbon dioxide in the air-breathing fish *Trichogaster trichopterus*. J Exp Biol 82:197–213
- Burggren WW (1982a) "Air gulping" improves blood oxygen transport during aquatic hypoxia in the goldfish *Carassius auratus*. Physiol Zool 55:327–334
- Burggren WW (1982b) Pulmonary blood plasma filtration in reptiles: a wet vertebrate lung. Science 215:77–78
- Burggren WW (1989) Lung structure and function: amphibians. In: Wood SC (ed) Comparative pulmonary physiology: current concepts. Marcel Dekker, New York, pp 153–192
- Burggren WW (1991) Does comparative respiratory physiology have a role in evolutionary biology (and vice versa)? In: Woakes AJ, Grieshaber MK, Bridges CR (eds) Physiological strategies for gas exchange and metabolism. Cambridge University Press, Cambridge, pp 1–13
- Burggren WW, Bemis WE (1992) Metabolism and ram gill ventilation in juvenile paddlefish, *Polyodon spathula* (Chondrostei: Polodontidae). Physiol Zool 65:515-539
- Burggren WW, Doyle M (1986) Ontogeny of regulation of gill and lung ventilation in the bullfrog, *Rana catesbeiana*. Respir Physiol 66:279–291
- Burggren WW, Feder ME (1985) Skin breathing in vertebrates. Sci Am 253:106-118
- Burggren WW, Infantino RL (1994) The respiratory transition from water- to air breathing during amphibian metamoprphosis. Am Zool 34:238-246

- Burggren WW, Johansen K (1982) Ventricular hemodynamics in the monitor lizard Varanus exanthematicus: pulmonary and systemic pressure separation. J Exp Biol 96:343-354
- Burggren WW, Johansen K (1986) Circulation and respiration in lungfishes (Dipnoi). J Morphol Suppl 1:217-236
- Burggren WW, Johansen K (1987) Circulation and respiration in lungfishes (Dipnoi). In: Bemis WE, Burggren WW, Kemp NE (eds) The biology and evolution of lungfishes. Alan R Liss, New York, pp 217–236
- Burggren WW, McMahon BR (1981) Hemolymph oxygen transport, acid-base status, and hydromineral regulation during dehydration in three crabs, *Cardisoma, Birgus* and *Coenobita*. J Exp Zool 218:53-64
- Burggren WW, McMahon BR (1988a) Circulation. In: Burggren BB, McMahon BR (eds) Biology of the crabs. Cambridge University Press, Cambridge, pp 298-332
- Burggren WW, McMahon BR (1988b) Biology of the land crabs: an introduction. In: Burggren BB, McMahon BR (eds) Biology of the crabs. Cambridge University Press, Cambridge, pp 1–5
- Burggren WW, Mwalukoma A (1983) Respiration during chronic hypoxia and hyperoxia in larval and adult bullfrogs (*Rana catesbeiana*). I. Morphological responses of lungs, skin and gills. J Exp Biol 105:191–203
- Burggren WW, Pinder AW (1991) Ontogeny of cardiovascular and respiratory physiology in lower vertebrates. Annu Rev Physiol 53:107–135
- Burggren WW, Roberts J (1991) Respiration and metabolism. In: Prosser CL (ed) Environmental and metabolic animal physiology. Wiley/Liss, New York, pp 353-435
- Burggren WW, Shelton G (1979) Gas exchange and transport during intermittent breathing in chelonian reptiles. J Exp Biol 82:75–92
- Burggren WW, West NH (1982) Changing importance of gills, lungs and skin during metamorphosis in the bullfrog *Rana catesbeiana*. Respir Physiol 47:151–164
- Burggren WW, Wood SC (1981) Respiration and acid-base balance in the salamander, Ambystoma tigrum: influence of temperature acclimation and metamorphosis. J Comp Physiol 144:241-246
- Burggren WW, McMahon BR, Costerton JW (1974) Branchial water- and blood-flow patterns and the structure of the gills of the crayfish *Procambarus clarkii*. Can J Zool 52:1511– 1518
- Burggren WW, Glass ML, Johansen K (1977) Pulmonary ventilation/perfusion relationships in terrestrial and aquatic chelonian reptiles. Can J Zool 55:2024–2034
- Burggren WW, Dunn J, Barnard K (1979) Branchial circulation and gill morphometrics in the sturgeon, Acipenser transmontanus. Can J Zool 57:2160-2170
- Burggren WW, Feder ME, Pinder AW (1983) Temperature and the balance between aerial and aquatic respiration in larvae of *Rana berlandieri* and *Rana catesbeiana*. Physiol Zool 56:263–273
- Burggren WW, Pinder AW, McMahon BR, Wheatly M, Doyle M (1985a) Ventilation, circulation and their interactions in the land crabs, *Cardisoma guanhumi*. J Exp Biol 117:133–154
- Burggren WW, Johansen K, McMahon B (1985b) Respiration in phyletically ancient fish. In: Foreman RE, Gorbman A, Dodd JM, Olson R (eds) Evolutionary biology of primitive fishes. Plenum Press, New York, pp 217–252
- Burggren WW, Infantino L, Townsend DS (1990) Developmental changes in cardiac and metabolic physiology of the direct developing tropical frog. *Eleutherodactylus coqui*. J Exp Biol 152:129–147
- Burggren WW, McMahon BR, Powers D (1991) Respiratory functions of blood. In: Prosser CL (ed) Environmental and metabolic animal physiology. Wiley/Liss, New York, pp 445–508
- Burggren WW, Bicudo JE, Glass ML, Abe AS (1992) Development of blood pressure and cardiac reflexes in the frog, *Pseudis paradoxus*. Am J Physiol 263:R602-R608
- Burleson ML, Milsom WK (1990) Propranolol inhibits oxygen sensitive chemoreceptor activity in trout gills. Am J Physiol 258:R1089–R1091
- Burnett BR (1972) Aspects of the circulatory system of *Pollicipes polymerus* (Cirripedia: Thoracica). J Morphol 136:79-108
- Burnett LE, Bridges CR (1981) The physiological properties and function of ventilatory pauses in the crab, *Cancer paguras*. J Comp Physiol 145:81-88

- Burnett LE, McMahon B (1987) Gas exchange hemolymph acid-base status and the role of branchial water stores during air exposure in three littoral crab species. Physiol Zool 60:27–36
- Burnett LE, DeFur PL, Jorgesen DD (1981) Application of the thermal dilution technique measuring cardiac output and assessing stroke volume in crabs. J Exp Zool 218:165-173
- Burns B, Gurtner GH (1973) A specific carrier for O<sub>2</sub> and CO<sub>2</sub> in the lung and placenta. Drug Metab Dispos 1:374–379
- Burns B, Gurtner GH, Peavy H, Cha YN (1975) A specific carrier for oxygen and carbon dioxide. In: Junod AF, Haller R (eds) Lung metabolism. Academic Press, New York, pp 159–184
- Burns B, Young-Nam C, Purcell JM (1976) A specific carrier for  $O_2$  and  $CO_2$  in the lung: effects of volatile anaesthetics on gas transfer and drug metabolism. Chest 69:316–321
- Burrell MH (1985) Endoscopic and virological observations of respiratory disease in a group of young thoroughbred horses in training. Equine Vet J 17:99–103
- Burri PH (1984a) Foetal and postnatal development of the lung. Annu Rev Physiol 46:617-628
- Burri PH (1984b) Lung development and histogenesis. In: Fishman AP, Fischer AB (eds) handbook of physiology; respiration, vol 4. American Physiological Society Press, New York, pp 1–46
- Burri PH (1985) Morphology and respiratory function of the alveolar unit. Int Arch Allergy Appl Immunol 76:2–12
- Burrin PH, Schovic S (1979) The adaptive responses of the rat lung after bilobectomy. Am Rev Respir Dis 119:769–777
- Bursell E (1970) An introduction to insect physiology. Academic Press, London
- Burton GJ, Mayhew TM, Robertson LA (1989) Stereologic re-examination of the effects of varying oxygen tensions on human placental villi maintained in organ culture for up to 12 hr. Placenta 10:263–273
- Burton RR, Smith AH (1967) Blood and air volumes in the avian lung. Poult Sci 47:85-91
- Burwell CS, Metacalfe J (1958) Heart disease and pregnancy; physiology and management. Little Brown and Company, Boston
- Butler J, Woakes AJ (1980) Heart rate, respiratory frequency and wing beat frequency of freeflying barnacle geese, *Branta leucopsis*. J Exp Biol 85:213–226
- Butler J, Woakes AJ (1984) Heart rate and earobic metabolism in Humboldt penguins, Spheniscus humboldti, during voluntary dives. J Exp Biol 108:419-428
- Butler PJ (1991a) Respiratory adaptations to limited oxygen supply during diving in birds and mammals. In: Woakes AJ, Grieshaber MK, Bridges CR (eds) Physiological strategies for gas exchange and metabolism. Cambridge University Press, Cambridge, pp 235–257

Butler PJ (1991b) Exercise in birds. J Exp Biol 160:233-262

- Butler PJ, Metcalfe JD (1983) Control of respiration and circulation. In: Rankin JC, Pitcher TJ, Duggan R (eds) Control processes in fish physiology. Croom Helm, Beckenham, pp 41–65
- Butler PJ, West NH, Jones DR (1977) Respiratory and cardiovascular responses of the pigeon to sustained level flight in a wind tunnel. J Exp Biol 71:7–26
- Butler PJ, Taylor EW, McMahon BR (1978) Respiratory and circulatory changes in the lobster (*Homarus vulgaris*) during long-term exposure to moderate hypoxia. J Exp Biol 73:131– 146
- Butler PJ, Milsom WK, Woakes AJ (1984) Respiratory cardiovascular and metabolic adjustments during steady state swimming in the green trutle, *Chelonis mydas*. J Comp Physiol 154B:167-174
- Butler PJ, Banzett RB, Fredberg JJ (1988) Inspiratory valving in avian bronchi: aerodynamic consideration. Respir Physiol 72:241–256
- Butlin RK, Tregenza T (1997) Is speciation no accident?. Nature (Lond) 387:551-553
- Cala P (1987) Aerial respiration in the catfish, *Eremophilus mutisii* (Trichomycteridae, Siluriformes), in the Rio Bogota Basin, Colombia. J Fish Biol 31:301-303
- Calder WA (1968) Respiratory and heart rates of birds at rest. Condor 70:358-365
- Calder WA (1984) Size, function, and life history. Harvard University Press, Cambridge
- Calder WA (1987) Scaling energetics of homeothermic vertebrates. An operational allometry. Annu Rev Physiol 49:107–120
- Calder WA, King JR (1974) Thermal and caloric relations of birds. In: Farner DS, King JR (eds) Avian biology, vol 4. Academic Press, London, pp 259–413

- Calder WA, Schmidt-Nielsen K (1968) Panting and blood carbon dioxide in birds. Am J Physiol 215:477–482
- Callahan PS (1972) The evolution of insects. Holliday House, New York
- Callender GS (1940) Variations of the amounts of CO<sub>2</sub> in different air currents. Q J R Soc 66:395–400
- Calvin M (1956) Chemical evolution and the origin of life. Am Sci 44:248-263
- Cameron E (ed) (1990) Flight: How things work. The Time Inc Book Company, Alexandria, Virginia
- Cameron JC (1989) The respiratory physiology of animals. Oxford University Press, Oxford

Cameron JN (1978) Regulation of blood pH in teleost fish. Respir Physiol 33:129-144

- Cameron JN (1979) Excretion of CO<sub>2</sub> in water breathing animals: a short review. Mar Biol Lett 1:3–13
- Cameron JN (1981) Brief introduction to the land crabs of the Palau Islands: stages in the transition to air breathing. J Exp Zool 218:1-5
- Cameron JN, Cech JJ (1970) Notes on the energy cost of gill ventilation in teleosts. Comp Biochem Physiol 34:447-455
- Cameron JN, Iwama GK (1987) Compensation of progressive hypercapnia in channel catfish and blue crabs. J Exp Biol 133:183–197
- Cameron JN, Mecklenburg TA (1973) Aerial gas exchange in the coconut crab, *Birgus latro*, with some notes on *Gecarcoidea Ialandii*. Respir Physiol 19:245–261
- Cameron JN, Randall DJ, Davis JC (1977) Regulation of the ventilation-perfusion ratio in gills of Dasyatis sabina and Squalus suckleyi. Comp Biochem Physiol 39A:505-519
- Cane MA (1983) Oceanographic events during EI Niño. Science 222:1189-1195
- Canfield DE, Teske A (1996) Late Proterozoic rise in atmospheric oxygen concentration inferred from phylogenetic and sulphur-isotope studies. Nature (Lond) 382:127–132
- Cannon WB (1939) The wisdom of the body. Norton, New York
- Cantin AM, North SL, Hubbard RC, Crystal RC (1987) Normal alveolar epithelial lining fluid contains high levels of glutathione. J Appl Physiol 63:152–157
- Capone DG, Zehr JP, Paerl HW, Bergman B, Carpenter EJ (1997) *Trichodesmium*, a globally significant marine cyanobacterium. Science 276:1221–1229
- Cappellen PV, Ingall ED (1996) Redox stabilization of the atmosphere and oceans by phosphorus-limited marine productivity. Science 271:493-496
- Carey C (1980a) Physiology of the avian egg. Am Zool 20:325-489
- Carey C (1980b) Adaptation of the avian egg to high altitude. Am Zool 20:449-464
- Carey C, Rahn H, Parisi P (1980) Calories, water, lipid and yolk in avian eggs. Condor 82:335–345

Carey FG (1973) Fishes with warm bodies. Sci Am 228:36-54

- Carey FG, Lawson KD (1973) Temperature regulation in free-swimming bluefin tuna. Comp Biochem Physiol 44A:375-386
- Carey FG, Robinson BH (1981) Daily patterns in the activities of swordfish, *Xiphius gladius*, observed by acoustic telemetry. Fish Bull US 79:277-289
- Carey FG, Teal JM (1966) Heat conservation in tuna fish muscle. Proc Natl Acad Sci (USA) 56:1464–1469
- Carey FG, Teal JM (1969) Mako and porbeagle: warm-bodied sharks. Comp Biochem Physiol 28:199–204
- Carey FG, Teal JM, Kanwisher JW, Lawson KD, Beckett JS (1971) Warm bodied fish. Am Zool 11:137–158
- Carey FG, Kanwisher JW, Brazier O, Gabrielson G, Casey JG, Pratt HW (1982) Temperature and activities of a white shark. Copeia 1982:254–260
- Carey SW (1976) The expanding Earth. Elsevier, Amsterdam
- Carlson CW (1960) Aortic rupture. Turkey Producer. January Issue
- Carney JM, Starke-Reed PE, Oliver CN, Landum RW, Cheng MS, Wu JF, Floyd RA (1991) Reversal of age-related increase in brain protein oxidation, decrease in enzyme activity, and loss in temporal and spatial memory by chronic administration of the spin-trapping compound N-tert-butyl-alpha-phenylnitrone. Proc natl Acad Sci USA 88:3633-3636
- Caro CG, Saffman PG (1965) Extensibility of blood vessels in isolated rabbit lungs. J Physiol (Lond) 178:193-210

- Carpenter FL (1975) Bird hematocrits: effects of high altitude and strength of flight. Comp Biochem Physiol 50A:415–417
- Carpenter FL, Hixon MA (1988) A new function for torpor: fat conservation in a wild migrant hummingbird. Condor 90:373–378
- Carpenter FL, Paton DC, Hixon MA (1983) Weight gain and adjustment of feeding territory size in migrant Rufous hummingbirds. Proc Natl Acad Sci USA 80:7263
- Carpenter RE (1975) Flight metabolism in flying foxes. In: Wu TYT, Brokaw CJ, Brennen C (eds) Swimming and flying in nature, vol 2. Plenum Press, New York, pp 883–890
- Carpenter RE (1985) Flight physiology of flying foxes, Pteropus poliocephalus. J Exp Biol 114:619-747
- Carpenter RE (1986) Flight physiology of intermediate sized fruit-bats (Family: Pteropodidae). J Exp Biol 120:79–103
- Carr AF (1952) Handbook of turtles. Cornell University, Ithaca
- Carr AF, Goodman D (1970) Ecological implications of size and growth in *Chelonia*. Copeia 1970:783-786
- Carr AF, Ross P, Carr S (1974) Interesting behaviour of the green turtle, *Chelonia mydas*, at a mid-ocean island breeding colony ground. Copeia 1974:702–706
- Carr MH (1996) Water on Mars. Oxford University Press, Oxford
- Carrier DR (1984) Lung ventilation in running lizards. Am Zool 24:84A
- Carrier DR (1987a) The evolution of locomotor stamina in tetrapods: circumventing a mechanical contraint. Paleobiology 13:326-341
- Carrier DR (1987b) Lung ventilation during walking and running in four species of lizards. Exp Biol 47:33–42
- Carrier DR (1988) Ventilation in iguana iguana: an action of tonic muscle. Am Zool 28:197A
- Carrier DR (1991) Conflict in the hypaxial musculoskeletal system: documenting an evolutionary constraint. Am Zool 31:644-654
- Carroll RL (1970) Quantitative aspects of the amphibian-reptilian transition. Forma Functio 3:165–178
- Carroll RL (1988) Vertebrate palaeontology and evolution. WH Freeman, New York
- Carslaw HS, Jaeger JC (1959) Conduction of heat in solids. Oxford University Press, Oxford
- Cartee GD, Farrar RP (1987) Muscle respiratory capacity and VO<sub>2max</sub> in identically trained young and old rats. J Appl Physiol 63:257–261
- Carter GS (1935) The fresh waters of the rain forest areas of British Guiana. J Linn Soc Lond Zool 39:147–193
- Carter GS (1955) The papyrus swamps of Uganda. W Heffer, Cambridge
- Carter GS (1957) Air breathing. In: Brown ME (ed) The physiology of fishes. Academic Press, London, pp 65–79
- Carter GS (1967) Structure and habitat in vertebrate evolution. Washington University Press, Seattle
- Carter GS, Beadle LC (1930) Notes on the habits and development of *Lepidosiren paradoxa*. J Linn Soc Zool 37:327–368
- Carter GS, Beadle LC (1931) The fauna of the swamps of the Paraguayan Chaco in relation to its environment: respiratory adaptations in fishes. J Linn Soc Zool 37:205–258
- Caruthers SD, Harris TR (1994) Effects of pulmonary blood flow on the fractal nature of flow heterogeneity in sheep lungs. J Appl Physiol 77:1474-1479
- Casey TM, Withers PC, Casey KK (1979) Matabolic and respiratory responses of Arctic mammals to ambient temperature during summer. Comp Biochem Physiol 155B:751-758
- Cassin S, Dawes GS, Mott JC, Ross BB, Strang LB (1964) The vascular resistance of the foetal and newly ventilated lung of the lamb. J Physiol (Lond) 171:61–79
- Cassini A, Favero M, Albergoni V (1993) Comparative studies in red-blooded and whiteblooded Antarctic teleost fish, *Pagothenia bemacchii* and *Chionodraco hamatus*. Comp Biochem Physiol 106C:333-336
- Cech JJ, Rowell DM, Glasgow JS (1977) Cardiovascular responses of the winter flounder *Pseudopleuronectes americanus* to hypoxia. Comp Biochem Physiol A 53:123-125
- Chabotarev DF, Korlsushko OV, Ivanov LA (1974) Mechanisms of hypoxia in the elderly. J Gerontol 29:393-400

- Chaline J, Laurine B (1986) Phyletic gradualism in a European Plio-Pleistocene *Mimomys* lineage (Arvicolidae, Rodentia). Paleobiology 12:203-216
- Chamberlain JA (1990) Jet propulsion of Nautilus: a surviving example of early Paleozoic cephalopod locomotor design. Can J Zool 68:806-814
- Chance B, Sies H, Boveris A (1979) Hydroperoxide metabolism in mammalian organs. Physiol Rev 59:527–605
- Chang DM, Mark R, Miller SL, Strathearn GE (1983) Prebiotic organic syntheses and the origin of life. In: Schopf JW (ed) Earth's earliest biosphere: its origin and evolution. Princeton University Press, Princeton, pp 53–92
- Chapin FS, Zimov SA, Shaver GR, Hobble SE (1996) CO<sub>2</sub> fluctuations at high latitudes. Nature (Lond) 383:585–586
- Chapman CB, Jensen D, Wildenthal K (1963) On circulatory control mechanisms in the Pacific hagfish. Circ Res 12:427–440
- Chapman DJ, Ragan MA (1980) Evolution of biochemical pathways: evidence from comparative biochemistry. Annu Rev Plant Physiol 31:639–678
- Chapman DJ, Schopf JW (1983) Biological and biochemical effects of the development of an aerobic environment. In: Schopf JW (ed) Earth's earliest atmosphere: its origin and evolution. Princeton University Press, Princeton, pp 302-320
- Chapman DL, Carlton DP, Nielson DW, Cummings JJ, Poulain FR, Bland RD (1994) Changes in lung liquid during spontaneous labour in foetal sheep. J Appl Physiol 76:523–530
- Chapman RC, Bennett AF (1975) Physiological correlates of burrowing in rodents. Comp Biochem Physiol 51A:599-603
- Chappell MA (1985) Effects of ambient temperature and altitude on ventilation and gas exchange in deer mice (*Peromyscus maniculatus*). J Comp Physiol 155B:751-758
- Chappell MA, Roverud RC (1990) Temperature effects on metabolism, ventilation and oxygen extraction in a Neotropical bat. Respir Physiol 81:401-412
- Chappell MA, Souza SL (1988) Thermoregulation, gas exchange and ventilation in Adélie penguins (*Pygoscelis adelie*). J Comp Physiol 157B:783–790
- Chappellaz J, Barnola JM, Raynaud D, Korotkevich YS, Lorius C (1992) Ice-core record of atmospheric methane over the past 160 000 years. Nature (Lond) 345:127-131
- Charnov EI (1993) Life history invariants, Oxford University Press, Oxford
- Charnov EI (1997) Trade-off-invariant rules for evolutionary stable state life histories. Nature (Lond) 387:393–394
- Cheatum EP (1934) Limnological investigations on respiration, annual migratory cycle and other related phenomena in fresh water pulmonate snails. Trans Am Microsc Soc 53:348–407
- Chen C, Rabourdin B, Hammen CS (1987) The effect of hydrogen sulfide on the metabolism of *Solemya velum* and enzymes of sulfide oxidation in gill tissue. Comp Biochem Physiol 88B:949-952
- Chien S (1970) Shear dependence of effective cell volume as a determinant of blood viscosity. Science 168:977-979
- Chien S (1985) Role of blood cells in microcirculatory regulation. Microvasc Res 29:129-151
- Chien S, Usami S, Dellenback RJ, Bryant CA (1971) Comparative hemorheology hematological implications of species differences in blood viscosity. Biorheology 8:35–57
- Childress JJ, Mickel TJ (1982) Oxygen and sulfide consumption rates of the vent cram Calyptogena pacifica. Mar Biol Lett 3:73-79
- Childress JJ, Felbeck H, Somero GN (1989) Symbiosis in the deep sea. Sci Am 256:107-112
- Chinard FP (1992) Quantitative assessment of epithelial fluid in the lung. Am J Physiol 263:L617-L618
- Chivers DJ, Hladik CM (1980) Morphology of the gastrointestinal tract in primates: comparison with other animals in relation to diet. J Morphol 166:337–386
- Chyba CF (1997) Life on other planets. Nature (Lond) 385:201
- Clark A, Clark PAA, Connett RJ, Gayaski TEJ, Honig CR (1987) How large is the drop in PO<sub>2</sub> between the cytosol and mitochondrion? Am J Physiol 252:C583–587

Clark JM, Lambertsen CJ (1971) Pulmonary oxygen toxicity: a review. Pharmacol Rev 23:37–98 Clark LC, Gollan F (1966) Survival of mammals breathing organic liquids equilibrated with

oxygen at atmospheric pressure. Science 152:1755–1756

Clarke FW (1911) The data of geochemistry. US Geol Surv Bull 491, pp 43-158

- Clarke MR (1962) Respiratory and swimming movements in the cephalopod, *Crachia scabra*. Nature (Lond) 196:351-352
- Clarke J, Nicol S (1993) Blood viscosity of little penguin, *Eudyptula minor*, and the Adélie penguin, *Pygoscelis adeliae*: effect of temperature and shear rate. Physiol Zool 66:720–731
- Clausen G, Ersland A (1968) The respiratory properties of the blood of two diving rodents, the beaver and the water vole. Respir Physiol 5:350–359
- Claussen CP, Hue A (1987) Light- and electronmicroscopic studies of the lung of Triturus alpestris (Laurenti) (Amphibia). Zool Anz 218:115-128
- Clayton DA, Vaughan TC (1986) Territorial acquisition in the mudskipper *Boleophthalmus* boddarti (Teleosti, Gobiidae) on the mudflats of Kuwaiti. J Zool (Lond) 209:501–519
- Clements JA, Nellenbogen J, Trahan HJ (1970) Pulmonary surfactant and evolution of the lungs. Science 169:603–604
- Clench M (1978) Tracheal elongation in birds-of-paradise. Condor 80:423-430
- Cloud P (1973) Paleoecological significance of the banded iron-formation. Econ Geol 68:1135– 1143
- Cloud P (1974) Evolution of ecosystems. Am Sci 62:54-66
- Cloud P (1988) Oasis in space. WW Norton, New York
- Cloud P (1983a) The biosphere. Sci Am 249:176-189
- Cloud P (1983b) Early biogeologic history: the emergence of a paradigm. In: Schopf JW (ed) Earth's earliest biosphere: its origin and evolution. Princeton University Press, Princeton, pp 14-31
- Cloudsley-Thompson JL (1977) The water and temperature relations of woodlice. Meadowfield Press, Durham, England
- Cloutier R, Forey PL (1991) Diversity of extinct and living actinistian fishes (Sarcopterygii). Environ Biol Fishes 32:59-74
- Cochrane CG (1991) Cellular injury by oxidants. Am J Med 91:23S-30S
- Cochrane CG, Revak SD (1991) Pulmonary surfactant protein B (SP-B):structure-function relationships. Science 254:566–568
- Codispoti LA, Christensen JP (1985) Nitrification, denitrification and nitrous oxide cycling in the eastern tropical south Pacific Ocean. Mar Chem 16:277–300
- Cody ML (1974) Optimization in ecology. Science 183:1156-1164
- Cohen GM (1975) Pulmonary metabolism of inhaled substances and possible relationship with carcinogenicity and toxicity. In: Junod AF, Haller R (eds) Lung metabolism. Academic Press, New York, pp 185–200
- Cohen J (1995) Getting all tunned around over the origins of life on Earth. Science 267:1265-1266
- Cohen JE (1964) Age and the pulmonary diffusing capacity. In: Cander L, Moyer J (eds) Aging of the lung. Grune and Stratton, New York, pp 163–172
- Cohen WD (1978) On erythrocyte morphology. Blood Cells 4:449-451
- Cohn LE, Kinnula VL, Adler KB (1994) Antioxidant properties of guinea pig tracheal epithelial cells in vitro. Am J Physiol 266:L397–L404
- Coin JT, Olson JS (1979) The rate of oxygen uptake by human erythrocytes. J Biol Chem 254:1178-1190
- Colacino JM, Kraus DW (1984) Haemoglobin-containing cells of *Neodasys* (Gastrotricha, Chaetonotida). II. Respiratory significance. Comp Biochem Physiol 79A:363–369
- Colacino JM, Hector DH, Schmidt-Nielsen K (1977) Respiratory responses of ducks to simulated altitude. Respir Physiol 29:265–281
- Colbert EH, Morales M (1991) Evolution of the vertebrates, 4th edn. Wiley-Liss, New York
- Coles GC (1970) Some biochemical adaptations of the swampworm *Alma emini* to low oxygen levels in tropical swamps. Comp Biochem Physiol 34:481-489
- Collazo A (1993) Evolutionary correlations between early development and life history in plethodontid salamanders and teleost fishes. Am Zool 33:60A
- Collins-George N (1959) The physical environment of soil animals. Ecology 40:550-557
- Colwell RK, Coddington JA (1994) Estimating terrestrial biodiversity through extrapolation. Philos Trans R Soc Lond 345B:101–118

Comroe JH (1966) The lung. Sci Am 57-68

- Comroe JH (1974) Physiology of respiration: an introductory text. Year Book Medical Publishers, Chicago
- Comroe JH, Dripps RD, Dumke PR, Deming M (1945) Oxygen toxicity: the effect of inhalation of high concentrations of oxygen for 24 hours on normal men at sea level and at simulated altitude of 18000 ft. J Am Med Assoc 128:710-725
- Conley KE, Kayar SR, Rösler K, Hoppeler H, Weibel ER, Taylor CR (1987) Adaptive variation in the mammalian respiratory system in relation to energetic demand: IV. Capillaries and their relationship to oxidative capacity. Respir Physiol 69:47–64
- Connett RJ, Gayeski TEJ, Honig CR (1985) An upper bound on the minimum PO<sub>2</sub> for O<sub>2</sub> consumption in red muscle. In: Kreuzer F, Cain SM, Turek Z, Goldstick TK (eds) Oxygen transport to tissue, vol 7. Plenum Publishing Corporation, New York, pp 291–300
- Connett RJ, Honig CR, Gayeski TEJ, Brooks GA (1990) Defining hypoxia: a systems view of O<sub>2</sub>, glycolysis, energetics, and intracellular PO<sub>2</sub>. J Appl Physiol 68:833–842
- Conolly ME, Greenacre JK (1975) Hormone receptors in the lung. In: Junod AF, Haller R (eds) Lung metabolism. Academic Press, New York, pp 233–249
- Constantinopol M, Jones JH, Weibel ER, Hoppelar H, Lidholm A, Karas RH (1989) Oxygen transport during exercise in large mammals. II. Oxygen uptake by the pulmonary gas exchanger. J Appl Physiol 67:871-878
- Constanzo JP, Iverson JB, Wright MF, Lee RE (1995) Cold hardiness and overwintering strategies of hatchilings in an assemblage of northern turtles. Ecology 76:1772-1785
- Conway-Morris S (1993) The fossil record and the early evolution of Metazoa. Nature (Lond) 361:219-225
- Cook RH, Boyd S (1965) The avoidance by *Gammarus aceanicus* Segerstrale (Amphiopoda: Crustacea) of anoxic regions. Can J Zool 43:971-975
- Cooke AS (1976) Changes in egg shell characteristics of the sparrow hawk (*Accipeter nisus*) and peregrine (*Falco peregrinus*) associated with exposure to pollutants during recent decades. J Zool (Lond) 187:245–263
- Cooke IRC, Berger PJ (1990) Precursor of respiratory pattern in the early gestation mammalian fetus. Brain Res 522:330–336
- Coope A, Penny D (1997) Mass survival across the Cretaceous-Tertiary boundary: molecular evidence. Science 275:1109-1113
- Cope ED (1885) The retrograde metamorphosis of Siren. Am Nat 19:1226-1227
- Cope ED (1894) On the lungs of Ophidia. Am Philos Soc 33:217-224
- Cope ED (1896) The primary factor of organic evolution. Open Court, Chicago
- Copeland DE (1969) Fine structural study of gas secretion in the physoclistous swim bladder of *Fundulus heteroclitus* and *Gadus callarias* and in the euphysoclistous swim bladder of *Opsanus tau.* Z Zellforsch 93:305-331
- Cornfield DN, Stevens T, McMurtry IF, Abman SH, Rodman DM (1994) Acute hypoxia causes membrane depolarization and calcium influx in fetal pulmonary artery smooth muscle cells. Am J Physiol 266:L469–L475
- Cory S, Adams JM (1988) Transgenic mice and oncogenesis. Annu Rev Immunol 6:25-48
- Cosgrove WB, Schwartz JB (1965) The properties and function of the blood pigment of the earthworm, *Lumbricus terrestris*. Physiol Zool 38:206-212
- Cossins AR (1991) Cold facts and naked truth. Nature (Lond) 353:699
- Costa HH (1967) Responses of *Gammarus pulex* (L) to modified environment. III. Reactions to low oxygen tensions. Crustaceana 13:175-189
- Costello ML, Mathieu-Costello O, West JB (1992) Stress failure of alveolar epithelial cells studied by scanning electron microscopy. Am Rev Respir Dis 145:1446–1455
- Cotton CU, Lawson EE, Boucher RC, Gatzy JT (1983) Bioelectric properties and ion transport of airways excised from adult and fetal sheep. J Appl Physiol 55:1542–1549
- Coulson JM, Richardson JF (1965) Chemical engineering. Pergamon Press, Oxford
- Cournand A, Richards DW, Bader RA, Bader ME, Fishman AP (1954) The oxygen cost of breathing. Trans Assoc Phys 67:162-173
- Coutant CC (1987) Thermal preference: when does an asset become a liability? Env Biol 18:161– 172

- Cracraft JA (1983) The significance of phylogenetic classification for systematic and evolutionary biology. In: Felsenstein J (ed) Numerical taxonomy. Springer Heidelberg, Berlin New York, pp 1–17
- Cracraft JA (1986) The origin and early diversification of birds. Paleobiology 12:383-399
- Cracraft JA (1994) Species diversity, biogeography, and the evolution of biotas. Am Zool 34:33– 47
- Craig P (1975) Respiration and body weight in the reptilian genus *Lacerta*: a physiological, anatomical and morphometric study. PhD Thesis, University of Bristol, England
- Crandall RR, Smith AH (1952) Tissue metabolism in growing birds. Proc Soc Exp Biol Med 79:345-346
- Crapo JD (1986) Morphologic changes in pulmonary oxygen toxicity. Annu Rev Physiol 48:721– 731
- Crapo JD (1987) Hyperoxia: lung injury and localization of antioxidant defenses. In: Dejours P (ed) Comparative physiology of environmental adaptations, vol 2. Karger, Basel, pp 163– 176
- Crapo JD, Crapo RO (1983) Comparison of total lung diffusion capacity and the membrane component of diffusion capacity as determined by physiologic and morphometric techniques. Respir Physiol 51:221-280
- Crapo JD, McCord JM (1976) Oxygen-induced changes in pulmonary superoxide dismutase, assayed by antibody titrations. Am J Physiol 231:1196–1203
- Crapo JD, Tierney DF (1974) Superoxide dismutase and pulmonary oxygen toxicity. Am J Physiol 226:1401–1407
- Crapo JD, Barry BE, Chang LY, Mercer RR (1984) Alterations in lung structure by inhalation of oxidants. J Toxicol Environ Health 13:301-321
- Crapo JD, Crapo RO, Jensen RL, Mercer RR, Weibel ER (1988) Evaluation of lung diffusing capacity by physiological and morphometric techniques. J Appl Physiol 64:2083– 2091
- Crapo RO, Crapo JD, Weibel ER (1986) Comparison of physiologic and morphometric estimations of the components of CO diffusing capacity. Prog Respir Res 21:88–91
- Crawford EC, Schultetus RR (1970) Cutaneous gas exchange in the lizard, *Sauromalus obesus*. Copeia 1970:179–180
- Crawford EC, Gatz RN, Magnussen H, Perry SF, Piiper J (1976) Lung volumes, pulmonary blood flow and carbon monoxide diffusing capacity of turtles. J Comp Physiol 107:169–178
- Crawford RMM, Hendry GAF, Goodman BA (eds) (1994) Oxygen and environmental stress in plants. Proc R Soc Edinb 102B:1–549
- Crezee M (1976) Solenofilomorphidae (Acoela), major component of a new turbellarian association in sulfide system. Int Rev Gesamten Hydrobiol Hydrogr 61:105–129
- Crews D, Grassman M, Garstka WR, Halpert A, Camazine B (1987) Sex and seasonal differences in metabolism in the red-sided garter snake, *Thamnophis sirtalis parietalis*. Can J Zool 65:2362-2368
- Crick FH (1966) Codon-anticodon pairing: the wobble hypothesis. J Mol Biol 38:367-379
- Crisp DJ (1964) Plastron respiration. In: Danielli JF, Pankhurst KGA, Riddiford AC (eds) Recent advances in surface science. Academic Press, London, pp 377–425
- Croll RP (1985) Sensory control of respiratory pumping in *Aplysia californica*. J Exp Biol 117:15–27
- Crompton AW, Taylor CR, Jagger JA (1978) Evolution in homeothermy in mammals. Nature (Lond) 272:333-336
- Crone C, Lassen NA (1970) Capillary permeability. The transfer of molecules and ions between capillary blood and tissue. Munksgaard, Copenhagen
- Crossfill ML, Widdicombe JG (1961) Physical characteristics of the chest and lungs and the work of breathing in different mammalian species. J Physiol (Lond) 158:1–14
- Crosswell JW, Smith EE (1967) Determination of optimal hematocrit. J Appl Physiol 22:930–933 Csillag C, Aldhous P (1992) Signs of damage by radicals. Science 258:1875–1876
- Culotta E, Koshland DE (1992) Molecule of the year: no news is good news. Science 258:1862-1865
- Cunningham DJ, O'Riordan JLH (1957) The effect of rise in temperature of the body on the respiratory response to carbon dioxide. Q J Exp Physiol 42:329–345

- Cunningham JT, Reid DM (1932) Experimental researches on the emission of oxygen by the pelvic filaments of the male *Lepidosiren* with some experiments on *Synbranchus* marmoratus. Proc R Soc Lond 110B:234-248
- Currie JA (1962) The importance of aeration in providing the right conditions for plant growth. J Sci Food Agric 13:380–385
- Currie JA (1984) Gas diffusion through soil crumbs: the effects of compaction and wetting. J Soil Sci 35:1–10
- Currey JD (1967) The failure of exoskeletons and endoskeletons. J Morphol 123:1-16
- Currey JD (1984) The mechanical adaptations of bones. Princeton University Press, Princeton Curtis SE, Peek JT, Kelly DR (1993) Partial liquid breathing with perfluocarbon improves arterial oxygenation in acute canine lung injury. J Appl Physiol 75:2696–2702
- Cutler A (1995) Notes from the underground. The Sciences (Public NY Acad Sci) Jan/Feb Issue: 36–40
- Czietrich HM, Bridges CR, Grieshaber MK (1987) Purine metabolism of the crayfish Astacus leptodactylus. Verh Dtsch Zool Ges 80:207
- Czopek J (1962a) Vascularization of respiratory surfaces in some caudata. Copeia 1962:576-587

Czopek J (1962b) Smooth muscle in the lungs of some urodeles. Nature (Lond) 193:798-811

- Czopek J (1965) Quantitative studies of the morphology of respiratory surfaces in amphibians. Acta Anat 62:296–323
- Czopek J, Szarski H (1989) Morphological adaptations to water movements in the skin of anuran amphibians. Acta Biol Cracov 31:81–96
- Dahr E (1924) Die Atmungsbewegungen der Landpulmonaten. Lund Univ Aarsskr NF Avd 20:1– 19
- Dahr E (1927) Studien uber die Respiration der Landpulmonaten. Lund Univ Aarsskr NF Avd 23:1–118
- Dales RP (1961) Observations on the respiration of the sabellid polychaete Schizobranchia insignis. Biol Bull Mar Biol Lab Woods Hole 121:82–91
- Dales RP (1969) Respiration and energy metabolism in annelids. In: Florkin M, Stoltz EH (eds) Chemical zoology, vol 4. Academic Press, New York, pp 93–109
- Dandy JWT (1970) Activity response to oxygen in brook trout, *Salvelinus fontinalis* (Mitchill). Can J Zool 48:1067–1078
- Daniel TL (1981) Fish mucus. In situ measurements of polymer drag reduction. Biol Bull 60:376– 382
- Daniel MJ, Boyden CR (1975) Diurnal variations in physicochemical conditions within rockpools. Field Stud 4:161–176
- Daniels CB, Barr HA, Nicholas TE (1989) A comparison of the surfactant associated lipids derived from reptilian and mammalian lungs. Respir Physiol 75:335–348
- Daniels CB, Barr HA, Power JHT, Nicholas TE (1990) Body temperature alters the lipid compo-
- sition of pulmonary surfactant in the lizard *Ctenophorus nuchalis*. Exp Lung Res 16:435–449 Daniels CB, Eskandari-Marandi BD, Nicholas TE (1993) The role of surfactant in the static lung mechanics of the lizard *Ctenophorus nuchalis*. Respir Physiol 94:11–23
- Daniels CB, Orgeig S, Wilsen J, Nicholas TE (1994) Pulmonary type surfactants in the lungs of terrestrial and aquatic amphibians. Respir Physiol 95:249-258
- D'Aoust G (1970) The role of lactic acid in gas secretion in the teleost swim bladder. Comp Biochem Physiol 32:637-668
- Darden TR (1970) Respiratory adaptations of a fossorial mammal, the pocket gopher (*Thomomys bottae*). PhD Diss, University of California, Davis
- Darnell RM (1949) The aortic arches and associated arteries of caudate amphibians. Copeia 1949:18-31
- Darwin CR (1851) A monograph of the subclass Cirripedia. The Lepadidae. John Murray, London
- Darwin CR (1859) On the origin of species by means of natural selection, or the preservation of favoured races in the struggle for life. John Murray, London
- Darwin CR, Wallace AR (1958) On the tendency of species to form varieties and on the perpetuation of varieties and species by natural means of selection. J Linn Soc Zool 3:45–62
- Das BK (1927) The bionomics of certain air-breathing fishes of India together with an account of the development of the air-breathing organs. Philos Trans R Soc Lond 216B:183–219
- Das BK (1934) The habits and structure of *Pseudapocryptes lanceolatus*, a fish in the first stages of structural adaptation to aerial respiration. Proc R Soc Lond 115B:422-435
- Das BK (1940) Nature and causes of evolution and adaptation of the air-breathing fishes. Proc 27th Indian Sci Congr 2:215–260
- Davenport HW (1974) The ABC of acid-base chemistry, 6th edn. The University of Chicago Press, Chicago
- Davenport J (1985) Environmental stress and behavioural adaptation. Croom Helm, London

Davenport J (1994) How and why do flying fish fly? Rev Fish Biol Fisheries 4:184-214

- Davenport J, Woolmington AD (1981) Behavioural responses of some rocky shore fish exposed to adverse environmental conditions. Mar Behav Physiol 8:1–12
- David A, Rao NGS, Ray P (1974) Tank fishery resources of Karnataka. Bull Cent Ini Fish Res Inst Barrackpore 20:1–87
- Davie P, Farrell AP (1991) Cardiac performance of an isolated heart preparation from the dogfish (*Squalus acanthias*): the effects of hypoxia and coronary artery perfusion. Can J Zool (Lond) 69:1822–1828
- Davies JC (1975) Minimal dissolved oxygen requirements of aquatic life with emphasis on Canadian species: a review. J Fish Res Board Can 32:2295-2395
- Davies P (ed) (1989) The new physica. Cambridge University Press, Cambridge
- Davies R, Cockrum EL (1964) Experimentally determined weight-lifting capacity of five species of western bats. J Mammal 45:643–644
- Davis GE, Foster J, Warren CE, Daudoroff P (1963) The influence of oxygen concentration on the swimming performance of juvenile Pacific salmon at various temperatures. Trans Am Fish Soc 92:111–124
- Dawes GS (1965) Oxygen supply and consumption in late foetal life and the onset of breathing at birth. In: Fenn WO, Rahn H (eds) Handbook of physiology, sect 3, respiration, vol II, American Physiolgical Society, Washington, DC, pp 1313–1328
- Dawes GS, Mott JC (1962) The vascular tone of the foetal lung. J Physiol (Lond) 164:465-477
- Dawes GS, Mott JC, Widdicombe JG (1953) Changes in the lungs of the newborn lamb. J Physiol (Lond) 121:141–162
- Dawes GS, Mott JC, Widdicombe JG (1954) The foetal circulation in the lamb. J Physiol (Lond) 126:563–587
- Dawson TJ, Dawson WR (1982) Metabolic scope and conductance in response to cold of some dasyurid marsupials and Australian rodents. Comp Biochem Physiol 71A:59-64
- Dean AM, Golding GB (1997) Protein engineering reveals ancient adaptive replacements in isocitrate dehydrogenase. Proc Natl Acad Sci USA 94:3104-3109
- DeAngelis DL, Godbout L, Shuter BJ (1991) An individual based approach to predicting density dependent dynamics in smallmouth bass populations. Ecol Model 57:91–115
- de Beer M (1989) Light measurements in Lake Victoria, Tanzania. Ann Mus R Afr Cent Ser Quarto Zool 257:57-60
- de Beers G (1951) Embryos and ancestors. Oxford University Press, Oxford
- de Beers G (1954) Archeopteryx lithographica. Br Mus Nat Hist Lond
- De Belder AJ, Radomski MW, Why HJF, Richardson CA et al. (1993) Nitric oxide synthase activities in human myocardium. Lancet 341:84–85
- DeFur PL, Mangum CP (1979) The effects of environmental variables on the heart rate of invertebrates. Comp Biochem Physiol A 62:283–294
- DeFur PL, McMahon BR (1978) Respiratory responses of *Cancer productus* to air exposure. Am Zool 18:605A
- DeFur PL, Pease AL (1988) Metabolic and Respiratory compensation during long term hypoxia in blue crabs, *Callinectes sapidus*. Understanding the Estuary, Advances in Chesapeake Bay Research, Chesapeake Res Consortium, Publication No 129
- Degan H, Kristensen B (1981) Low sensitivity of *Tubifex* sp. respiration to hydrogen sulfide inhibitors. Comp Biochem Physiol 69B:809-817
- De Groodt M, Lagasse A, Sebruyns M (1960) Electronenmikroskopische Morphologie der lungenalveolen des *Protopterus* und *Amblystoma*. Proc Int Congr Electron Microscopy, Springer, Berlin Heidelberg New York, 418 pp
- Dehadrai PV (1962) Respiratory function of the swim bladder of *Notopterus* (Lacepede). Nature (Lond) 185:929

- Dehadrai PV, Tripathi SD (1976) Environment and ecology of fresh water air-breathing teleosts. In: Hughes GM (ed) Respiration of amphibious vertebrates. Academic Press, London, pp 39– 72
- De Jong HJ (1962) Activity of the body wall musculature of the African clawed toad, *Xenopus laevis* (Daudin) during diving and respiration. Zool Med 47:135-144
- De Jong HJ, Gans C (1969) On the mechanism of respiration in the bullfrog, *Rana catesbeiana*: reassessment. J Morphol 127:259–290
- Dejours P (1973) Problems of control of breathing in fishes. In: Bolis L, Schmidt-Nielsen K, Madrell SHP (eds) Comparative physiology. Elsevier/North Holland, Amsterdam, pp 117-133
- Dejours P (1975) Principles of comparative respiratory physiology, 1st edn. Elsevier-North Holland, Amsterdam
- Dejours P (1981) Principles of comparative respiratory physiology, 2nd edn. Elsevier-North Holland, Amsterdam
- Dejours P (1982) Mount Everest and beyond: breathing air. In: Taylor CR, Johansen K, Bolis L (eds) A companion to animal physiology. Cambridge University Press, Cambridge, pp 17–27
- Dejours P (1988) Respiration in water and air: adaptations, regulation and evolution. Elsevier, Amsterdam
- Dejours P (1989) From comparative physiology of respiration to several problems of environmental adoptations and to evolution. J Physiol (Lond) 410:1–19
- Dejours P (1990) Comparative aspects of maximal oxygen consumption. Respir Physiol 80:155– 162
- Dejours P (1994) Environmental factors as determinats in bimodal breathing: an introductory overview. Am Zool 34:178–183
- Dejours P, Truchot JP (1988) Respiration of the immersed shore crab at variable ambient oxygenation. J Comp Physiol 158B:387-391
- Dejours P, Wagner S, Dejager M, Vichon MJ (1967) Ventilation et gaz alvéolaire pendant langage parlé. J Physiol (Paris) 59:386
- Dejours P, Garey WF, Rahn H (1970) Comparison of ventilatory and circulatory flow rates between animals in various physiological conditions. Respir Physiol 9:108-117
- Dejours P, Toulmond A, Truchot JP (1977) The effect of hyperoxia on breathing of marine fishes. Comp Biochem Physiol 58A:409-411
- Dekker RWRJ, Brom TG (1992) Megapode phylogeny and the interpretation of incubation strategies. Zool Verh Leiden 278:9-31
- DeLaney RG, Fishman AP (1977) Analysis of lung ventilation in the aestivating lungfish Protopterus aethiopicus. Am J Physiol 233:R181-187
- DeLaney RG, Lahiri GS, Fishman AP (1974) Aestivation of the African lungfish *Protopterus* aethiopicus: cardiovascular and respiratory functions. J Exp Biol 61:111-118
- DeLaney RG, Laurent P, Galante R, Pack AI, Fishman AP (1983) Pulmonary mechanoreceptors in the dipnoan lungfish *Protopterus* and *Lepidosiren*. Am J Physiol 244:R418-R428
- Delmas RJ, Ascencio JM, Legrand M (1980) Polar ice evidence that the atmospheric  $CO_2 20\,000\,yr$ BP was 50% of the present. Nature (Lond) 284:155–157
- Demoll R (1927) Untersuchungen über die Atmung der Insekten. Z Biol 87:8-22
- Demple B, Harrison L (1994) Repair of oxidative damage to DNA enzymology and biology. Annu Rev Biochem 63:915-948
- Dempsey EW (1960) Histophysical considerations: In: Villee CA (ed) The placenta and foetal membranes. Williams and Wilkins, Baltimore, pp 29–35
- Dempsey JA (1986) Is the lung built for exercise? Med Sci Sports Exercise 18:143-155
- Dence WA (1933) Notes on large bowfin (*Amia calva*) living in a mud puddle. Copeia, Ichtyol Notes 1:35
- Denison RH (1941) The soft anatomy of Bothriolepis. J Paleontol 15:553-561
- Denney MW (1993) Air and water: the biology and physics of life's media. Princeton University Press, Princeton
- Densmore LD, Owen RD (1989) Molecular systematics of the order Crocodilia. Am Zool 29:181A Denton EJ, Shaw TI, Gilpin-Brown JB (1958) Bathyscaphoid squid. Nature (Lond) 182:1810
- Denton EJ, Liddicoat JD, Taylor DW (1970) Impermeable "silvery" layers in fishes. J Physiol (Lond) 207:64P

- De Queiroz K (1985) The ontogenetic method for determining character polarity and its relevance to phylogenetic systematics. Syst Zool 34:280-299
- De Saint-Aubain ML, Wingstrand K (1981) A sphincter in the pulmonary artery of the frog *Rana* temporaria and its influence on blood flow in skin and lungs. Acta Zool (Stockh) 60:163–172
- Desbruyères D, Laubier B (1986) Les Alvineliidae, une famille nouvelle d'annélides polychètes inféodées aux sources hydrothermales sous-marines: systématique, biologie et écologie. Can J Zool 64:2227-2245
- Desbruyères D, Crassous P, Grassle J, Khripounoff A, Reyss D, Rio M, van Praet M (1982) Données écologiques sur un nouveau site d'hydrothermalisme actif de la ride du Pacifique oriental. C R Hebd Séances Acad Sci Paris Ser III 295:489–494
- Des Marais DJ, Strauss H, Simmons RE, Hayes JM (1992) Carbon isotope evidence for the stepwise oxidation of the Proterozoic environment. Nature (Lond) 359:605–609
- De Villiers CJ, Hodgson AN (1987) The structure of the secondary gills of Siphonaria capensis (Gastropoda; Pulmonata). J Molluscan Stud 53:129–138
- De Vries AL (1971) Glycoproteins as biological antifreeze agents in Antarctic fishes. Science 172:1152–1155
- De Vries R, De Jager (1984) The gill in the spiny dogfish, Squalus acanthias: respiratory and nonrespiratory function. Am J Anat 169:1–29
- De Vries TJ, Ortlieb L, Diaz A (1997) Determining the early history of El Niño. Science 276:965–966
- D'Hondt S, Arthur MA (1997) Late Cretaceous oceans and the cool tropical paradox. Science 271:1838–1841
- Diamond JM (1992) The red flag of optimality. Nature (Lond) 355:204-205
- Diamond JM (1998) Evolution of biological safety factors: a cost benefit analysis. In: Weibel ER, Taylor CR, Bolis L (eds) Principles of animal design: the optimization and symmorphosis debate. Cambridge University Press, London, pp 21–27
- Diamond JM, Hammond KA (1992) The matches, achieved by natural selection, between biological capacities and their natural loads. Experientia 48:551–557
- Diamond JM, Karasov WH, Phan D, Carpenter FL (1986) Digestive physiology is a determinant of foraging bout frequency in hummingbirds. Nature (Lond) 320:62-63
- Diaz H, Rodriguez G (1977) The branchial chamber in terrestrial crabs: a comparative study. Biol Bull 153:485-504
- Dickson KA, Graham JB (1986) Adaptations to hypoxic environments in the erythrinid fish, *Hoplias microlepis*. Environ Biol Fish 15:301–308
- Dickson KA, Maloney JE, Berger PJ (1986) Decline in liquid volume before labour in foetal lambs. J Appl Physiol 61:2266-2272
- Dierichs R, Dosche C (1982) The alveolar lining layer in the lung of the axolotol, *Ambryostoma mexicanum*. Cell Tissue Res 222:677–686
- Dill DB, Graybill A, Hurtado A, Taguini AC (1963) Gaseous exchange in the lungs in old age. J Am Geriatr Soc 11:1063–1076
- DiMagno L, Chan CK, Jia Y, Lang MJ, Newman JR, Mets L, Fleming GR, Haselkorn R (1995) Energy transfer and trapping in photosystem reaction centers from cyanobacteria. Proc Natl Acad Sci USA 92:2715–2719
- Dios-Escolar J, Gallego B, Tejero C, Escolar MA (1994) Changes occurring with increasing age in the rat lung: morphometrical study. Anat Rec 239:287–306
- Di Prampero PE (1985) Metabolic and circulatory limitations to Vo $_{\rm 2max}$  at the whole animal level. J Exp Biol 115:319–331
- Dixon B (1994) Power unseen: how microbes rule the world. WH Freeman, New York
- Dixon D (1987) The planet Earth. World Book, Chicago
- Dobzhansky T (1973) Nothing in biology makes sense except in the right of evolution. Am Biol Teach 35:125–129
- Dock DS, Kraus WL, McGuire LB, Hyland JW, Haynes FW, Dexter L (1961) The pulmonary blood volume in man. J Clin Invest 40:317–328
- Doebeli M (1993) The evolutionary advantage of controlled chaos. Proc R Soc Lond 254B:281– 285
- Doebeli M, Blarer A, Ackerman M (1997) Population dynamics, demographic stochasticity, and the evolution of cooperation. Proc Natl Acad Sci USA 94:5167–5171

Doeller JE, Kraus DW, Colacino JM, Wittenberg JB (1988) Gill haemoglobin may deliver sulfide to bacterial symbionts of *Solemya velum* (Bivalvia, Mollusca). Biol Bull 175:388–396

- Donahue TM (1966) The problem of atomic hydrogen. Ann Geophys 22:175-188
- Dominguez-Bello MG, Ruiz MC, Michelangeli F (1993) Evolutionary significance of fore gut fermentation in the hoatzin (*Opisthocomus hoazin*) Aves:Opisthocomidae. J Comp Physiol 163:594–601
- Dorer FE, Kahn JR, Lentz KE, Levine M, Skeggs WT (1974) Hydrolysis of bradykinin by angiotensin-converting enzyme. Circ Res 34:824-827
- Dorsey NE (1940) Properties of ordinary water substance. Reinhold, New York
- Doudoroff P, Shumway DL (1970) Dissolved oxygen requirements of freshwater fishes. FAO Fish Tech Pap 86:291
- Douglas EL, Peterson KS, Gysi JR, Chapman DJ (1985) Myoglobin in the heart tissue of fishes lacking haemoglobin. Comp Biochem Physiol 81:885–888
- Drabkin DL (1950) The distribution of the chromoproteins, myoglobin, and cytochrome *c*, in the tissues of different species, and the relationship of the content of each chromoprotein to body mass. J Biol Chem 182:317–333
- Dratisch L (1925) Über das Leben der Salamandra-Larven bei hohem und niedrigem Sauerstoffpartialdruck. Z Vergl Physiol 2:632–657
- Drent R (1975) Incubation. In: Farner DS, King JR (eds) Avian biology, vol 5. Academic Press, New York, pp 333-419
- Dresco-Derouet L (1974) Étude des mygales II. Premier résultats sur la biologie et le métabolisme respiratoire de différentes espèces tropicales en captivité. Bull Mus Natl Hist Nat 42:1054–1062
- Driedzic WR (1988) Matching cardiac oxygen delivery and fuel supply to energy demand in teleosts and cephalopods. Can J Zool 66:1078-1083
- Driedzic WR, Gesser H (1994) Energy metabolism and contractility in ectothermic vertebrate hearts: hypoxia, acidosis, and low temperature. Physiol Rev 74:221–258
- Driedzic WR, Stewart JM (1982) Myoglobin content and the activities of enzymes of energy metabolism in red and white fish hearts. J Comp Physiol 149:67–73
- Driedzic WR, Phleger CF, Fields JH, French C (1978) Alterations in energy metabolism associated with transitions from water to air breathing fish. Can J Zool 56:730–735
- Dubach M (1981) Quantitative analysis of the respiratory system of the house sparrow, budgerigar, and violet-eared hummingbird. Respir Physiol 46:43–60
- Dubale MS (1959) A comparative study of the oxygen carrying capacity of the blood in water and air-breathing teleosts. J Anim Morphol Physiol 6:48–54
- Dube SC (1972) Investigations on the functional capacity of respiratory organs of certain fresh water teleostean fishes. PhD Thesis, Banaras University, India
- Dudley GA, Abraham WM, Terjung RL (1982) Influence of exercise intensity and duration on biochemical adaptations in skeletal muscle. J Appl Physiol 53:844–850
- Duellman WE, Trueb L (1986) Biology of amphibians. McGraw-Hill, New York
- Duling BR, Kuschinsky W, Wahl M (1979) Measurements of the perivascular PO<sub>2</sub> in the vicinity of the pial vessels of the cat. Pfluegers Arch 383:29–34
- Dullemeijer P (1974) Concepts and approaches in animal morphology. Van Corkum, Assen
- Duncker H-R (1974) Structure of the avian respiratory tract. Respir Physiol 22:1-34
- Duncker H-R (1978a) General morphological principles of amniotic lungs. In: Piiper J (ed) Respiratory function in birds, adult and embryonic. Springer, Berlin Heidelberg New York, pp 1–15
- Duncker H-R (1978b) Development of the avian respiratory system. In: Piiper J (ed) Respiration in birds, adult and embryonic. Springer, Berlin Heidelberg New York, pp 260–273
- Duncker H-R (1979) General morphological principles of amniotic lungs. In: Piiper J (ed) Respiratory function in birds, adult and embryonic. Springer, Berlin Heidelberg New York, pp 1–15
- Duncker H-R (1985) The present situation of morphology and its importance for biological and medical sciences. In: Duncker H, Fisher W (eds) Vertebrate morphology. Fischer, Stuttgart, p 9
- Duncker H-R (1991) The evolutionary biology of homoithermic vertebrates: the analysis of complexity as a specific task of morphology. Verh Dtsch Zool Ges 84:39–60

- Duncker HR, Guntert M (1985a) The quantitative design of the avian respiratory system: from hummingbird to the mute swan. In: Nachtigall W (ed) BIONA report No 3. Gustav Fischer, Stuttgart, pp 361–378
- Duncker HR, Guntert M (1985b) Morphometric analysis of the avian respiratory system. In: Duncker HR, Fleischer G (eds) Vertebrate morphology. Gustav Fischer, Stuttgart, pp 383– 387
- Dunel-Erb S, Laurent P (1980a) Ultrastructure of marine teleost gill epithelia: SEM and TEM study of the chloride cell apical membrane. J Morphol 165:175–186
- Dunel-Erb S, Laurent P (1980b) Functional organization of the gill vasculature in different classes of fish. In: Lahlou B (ed) Epithelial transport in the lower vertebrates. Cambridge University Press, Cambridge, pp 37–58
- Dunham AE (1993) Population responses to environmental change: physiologically structured models, operative environments, and population dynamics. In: Karieva PM, Kingsolver JS, Huey RB (eds) Biotic interactions and global change. Sinauer, Sunderland, pp 196–275
- Dupré RK, Wood SC (1988) Behavioural temperature regulation by aquatic ectotherms during hypoxia. Can J Zool 66:2649–2652
- Dupré RK, Burggren WW, Vitalis TZ (1991) Dehydration decreases cutaneous gas diffusing capacity in the toad, *Bufo woodhouseii*. Am Zool 31:75A
- Durbin AG, Durbin G, Verity PG, Smayda TJ (1981) Voluntary swimming speeds and respiration rates of a filter-feeding planktivore, the Atlantic menhaden, *Brevoortia tyranuus* (Pisces: Clupeidae). Fish Bull 78:877–886
- Durmowicz AG, Hofmeister S, Kadyraliev TK, Aldashev AA, Stenmark KR (1993) Functional and structural adaptation of the yak pulmonary circulation to residence at high altitude. J Appl Physiol 74:2276–2285
- Duval A (1983) Heartbeat and blood pressure in terrestrial slugs. Can J Zool 61:987-992
- Duykers LRB, Percy JL (1978) Lung resonance characteristics of submerged mammals. J Acoust Soc Am 64:S97
- Dyer BD, Ober RA (1994) Tracing the history of the eukryotic cells. Columbia University Press, New York
- Dyer MF, Uglow RF (1978) Gill chamber ventilation and scaphognathite movements in *Crangon crangon* (L). J Exp Mar Biol Ecol 31:195–207
- Eastman JT (1991) Evolution and diversification of Antarctic notothenioid fishes. Am Zool 31:93-109
- Eastman JT (1993) Antarctic fish biology: evolution in a unique environment. Academic Press, San Diego
- Eaton R, Arp AJ (1990) The effect of sulfide on oxygen consumption rate of *Urechis caupo*. Am Zool 30:69A
- Eaton R, Arp AJ (1993) Aerobic respiration during sulfide exposure in the marine echiuran worm *Urechis caupo*. Physiol Zool 66:1–19
- Ebeling AW, Bernal P, Zuleta A (1970) Emersion of the amphibious Chilean clingfish Sicyases sanguineus. Biol Bull Mar Biol Lab woods Hole 139:115–137
- Eberly LB, Kanz JE, Taylor C, Pinsker H (1981) Environmental modulation of a central pattern generator in freely behaving *Aplysia*. Behav Neural Biol 32:21–34
- Economos AC (1979) Gravity, metabolic rate, and body size of mammals. Physiologist 22:S71
- Eddy FB, Bamford OS, Maloiy GMO (1981) Na<sup>+</sup> and Cl<sup>-</sup> effluxes and ionic regulation in *Tilapia* grahami, a fish living in conditions of extreme alkalinity. J Exp Biol 91:349-353
- Edney EB (1960) Terrestrial adaptations. In: Waterman TH (ed) The physiology of Crustacea, vol I. Academic Press, New York, pp 367-393
- Edney EB, Spencer J (1955) Cutaneous respiration in woodlice. J Exp Biol 32:256-269
- Edsall JT (1972) Blood and haemoglobin: the evolution of knowledge of functional adaptation in a biochemical system. J Hist Biol 5:205–257
- Edwards GA, Ruska H, Harven de E (1958) The fine structure of insect tracheoblasts, tracheae and tracheoles. Arch Biol 69:351–369
- Edwards RRC (1971) An assessment of the energy cost of gill ventilation in the plaice (*Pleuronectes platessa* L). Comp Biochem Physiol 40:391–398
- Effros RM, Mason GR (1983) Measurements of pulmonary epithelial permeability in vivo. Am Rev Respir Dis 127:S59–S65

- Effros RM, Murphy C, Ozker K, Hacker A (1992) Kinetics of urea exchange in air- filled and fluid-filled lungs. Am J Physiol 263:L619–L626
- Egan EA, Olver RE, Strand LB (1975) Changes in non-electrolyte permeability of alveoli and the absorption of lung liquid at the start of breathing in the lamb. J Physiol (Lond) 244:161–179
- Ege R (1916) Less known respiratory media. Vidensk Medd Dan Naturhist Foren 67:14-16
- Ege R (1918) On the respiratory function of the air stores carried by some aquatic insects (Corixidae, Dytiscidae, and Notonecta). Z Vergl Physiol 17:81–124
- Eger WH (1971) Ecological and physiological adaptations of intertidal clingfishes (Teleostei; Gobisocidae) in the Gulf of California. PhD Dissertation, Department of Biological Sciences, University of Arizona, Tucson
- Ehrenfeld J, Garcia-Romeu F (1977) Active hydrogen excretion and sodium absorption through isolated frog skin. Am J Physiol 233:F46–F54
- Ehret G, Tautz J, Schmitz B (1990) Hearing through the lungs: lung-eardrum transmission of sound in the frog *Eleutherodactylus coqui*. Naturwissenschaften 77:192–194
- Ehrlich HL (1996) Geomicrobiology, 3rd edn. Marcel Dekker, New York
- Eigen M, Schuster P (1979) The hypercycle. Springer, Berlin Heidelberg New York
- Eiseman B, Bryant B, Waltuch T (1964) Metabolism of vasomotor agents by the isolated perfused lung. J Thorac Cardiovasc Surg 48:798-806
- Ekblom B, Wilson G, Åstrand PO (1975) Central circulation during exercise after venesection and reinfusion of red blood cells. J Appl Physiol 40:379–383
- Elder HY (1975) Muscle structure. In: Usherwood PNR (ed) Insect muscle. Academic Press, New York, pp 1–74
- Eldredge N (1993) History, function, and evolutionary biology. Evol Biol 27:33-50
- Eldredge N, Gould SJ (1972) Punctuated equilibria: an alternative to phyletic gradualism. In: Schopf TJM (ed) Models in paleobiology. Freeman Cooper, San Francisco, pp 82–115
- El Haj AJ, Innes AJ, Taylor EW (1986) Ultrastructure of the pulmonary, cutaneous and branchial gas exchange organs of the Trinidad mountain crab. J Physiol (Lond) 373:84P
- Elkins N (1983) Weather and bird behaviour. T and AD Poyser, Calton, Stoke on Trent
- Ellington CP (1981) The aerodynamics of flapping animal flight. Am Zool 24:95–105
- Ellington CP (1984) The aerodynamics of hovering insect flight. VI. Life and power requirements. Philos Trans R Soc Lond 305B:145–181
- Ellis CG, Potter RF, Groom AC (1983) The Krogh cylinder geometry is not appropriate for modelling O<sub>2</sub> transport in contracted skeletal muscle. Adv Exp Med Biol 159:253–268
- Else PL, Hubert AJ (1981) Comparison of the "mammal machine" and the "reptilian machine" energy production. Am J Physiol 240:R3–R9
- Else PL, Hubert AJ (1983) A comparative study of the metabolic capacity of hearts from reptiles and mammals. Comp Biochem Physiol 76A:553–557
- Else PL, Hubert AJ (1985) Mammals: an allometric study of metabolism at tissue and mitochondrial level. Am J Physiol 248:R415–R421
- Embley MT, Hirt RP, Williams DM (1994) Biodiversity at the molecular level: the domains, kingdoms and phyla of life. Philos Trans R Soc Lond 345B:21-33
- Emery SH, Szczepanski A (1986) Gill dimensions in pelagic elasmobranch fishes. Biol Bull 171:441-449
- Emilio MG, Shelton G (1974) Gas exchange and its effect on blood-gas concentrations in the amphibian, *Xenopus laevis*. J Exp Biol 60:567–579
- Emilio MG, Machado MM, Menano HP (1970) The production of hydrogen ion gradient across the isolated frog skin: quantitative aspects and the effects of acetazolamide. Biochem Biophys Acta 203:394–409
- Emmett B, Hochachka PW (1981) Scaling of oxidative and glycolytic enzymes in mammals. Respir Physiol 45:261–272
- Engel LA (1991) Effect of microgravity on the respiratory system. J Appl Physiol 70:1907–1911
- Engelhard EK, Kam-Morgan LNW, Washburn JO, Volkman LE (1994) The insect tracheal system: a conduit for the systemic spread of *Autographa californica* nuclear polyhedrosis virus. Proc Natl Acad Sci USA 91:3224–3227
- Enns T, Scholander PF, Bradstreet ED (1965) Effect of hydrostatic pressure on gases dissolved in water. J Phys Chem 69:389–391

- Epe B (1995) DNA damage profiles induced by oxidizing agents. Rev Physiol Biochem Pharmacol 127:223-249
- Epting RJ (1980) Functional dependence of the power for hovering on wing disc loading in hummingbirds. Physiol Zool 53:347-352
- Erasmus B, Howell B, Rahn H (1970/71) The ontogeny of acid-base balance in the bullfrog and chicken. Respir Physiol 11:46–53
- Eriksen CH, Brown RJ (1980a) Comparative respiratory physiology and ecology of phyllopod Crustacea. I. Conchostraca. Crustaceana 39:1–10
- Eriksen CH, Brown RJ (1980b) Comparative respiratory physiology and ecology of phyllopod Crustacea. II. Anostraca. Crustaceana 39:11–21
- Errington PL (1963) Muskrat populations. Iowa State University, Iowa
- Erskine RLA, Ritchie JWK (1985) Umbilical artery blood flow characteristics in normal and growth retarded fetuses. Br J Obstet Gynecol 5:605-610
- Erwin DH (1993) The great Paleozoic crisis: life and death in the Permian. Columbia University Press, New York
- Erwin DH (1994) The Permo-Triassic extinction. Nature (Lond) 367:231-236
- Erwin DH (1996) The mother of mass extinctions. Sci Am 275:56-62
- Escourrou P, Qi X, Weiss M, Mazmanian GM, Gaultier C, Hervé P (1993) Influence of pulmonary blood flow on gas exchange in piglets. J Appl Physiol 75:2478–2483
- Evans HM (1929) Some notes on the anatomy of the electric eel, *Gymnotus electrophorus*, with special reference to a mouth-breathing organ and the swim bladder. Proc Zool Soc Lond 57:17–23
- Evans HM, Damant GCC (1928) Observations on the physiology of the swim bladder of cyprinoid fishes. J Exp Biol 6:42-55
- Everson I, Ralph R (1968) Blood analyses of some Antarctic fish. Br Antarct Surv Bull 15:59-62
- Ewer DW (1941) The blood systems of Sabella and Spirographis. Q J Microsc Soc 82:587-620

Ewer DW (1959) A toad (Xenopus leavis) without haemoglobin. Nature (Lond) 183:271

- Faber JJ (1993) Diffusion permeability of the immature placenta of the rabbit embryo in inert hydrophilic molecules. J Appl Physiol 265:H1804–H1808
- Faber JJ, Rahn H (1970) Gas exchange between air and water and the ventilation pattern of the electric eel. Respir Physiol 9:151–161
- Faber JJ, Thornburg KL, Binder NC (1992) Physiology of placental transfer in mammals. Am Zool 32:343–354
- Fahlén G (1971) The functional morphology of the gas bladder of the genus *Salmo*. Acta Anat 78:161–184
- Fairbridge RW (1966) The encyclopedia of oceanography. Reinhold Publishing, New York
- Falke KJ, Hill RD, Schneider RC, Guppy M, Liggins GC et al. (1985) Seal lungs collapse during free diving: evidence from arterial nitrogen tensions. Science 229:556–567
- Falkowski PG (1997) Evolution of the nitrogen cycle and its influence on the biological sequestration of  $CO_2$  in the ocean. Nature (Lond) 387:272–275
- Fanelli GM, Goldstein L (1964) Ammonia excretion in the neotenous newt *Necturus maculosus* (Raf). Comp Biochem Physiol 13:193–204
- Fänge R (1953) The mechanisms of gas transport in the euphysoclist swim bladder. Acta Physiol Scand 30:1–133
- Fänge R (1966) Physiology of the swim bladder. Physiol Rev 46:299-322
- Fänge R (1976) Gas exchange in the swim bladder. In: Hughes GM (ed) Respiration in amphibious vertebrates. Academic Press, London, pp 189–211
- Fänge R (1983) Gas exchange in the fish swim bladder. Rev Physiol Biochem Pharmacol 97:111– 158
- Farabaugh AT, Thomas DB, Thomas SP (1985) Ventilation and body temperature of the bat *Phyllostomus hastatus* during flight at different air temperatures. Physiologist 28:272
- Faraci MF (1990) Cerebral circulation during hypoxia: is a bird brain better? In: Sutton JR, Coates G, Remmers JE (eds) Hypoxia: the adaptations. BC Decker, Burlington, Ontario, pp 26–29
- Faraci MF, Fedde MR (1986) Regional circulatory responses to hypocapnia and hypercapnia in bar-headed geese. Am J Physiol 250:R499–R504

Faraci MF, Kilgore DL, Fedde MR (1984) Oxygen delivery to the heart and brain during hypoxia: Pekin duck vs bar-headed geese. Am J Physiol 247:R69–R75

Farhi L (1964) Gas sores of the body. In: Fenn WO, Rahn H (eds) Handbook of physiology, sect 3. respiration, vol I. Am Physiol Soc, Washington, DC, pp 873–924

Farhi L, Rahn H (1955) Gas stores of the body and the steady state. J Appl Physiol 7:472-484

Farley RD (1990) Regulation of air and blood flow through the book lungs of the desert scorpion, *Paruroctonus mesaensis*. Tissue Cell 22:547–569

- Farley RD, Case JF (1968) Perception of external oxygen by the burrowing shrimp, *Callianassa californiensis* Dan and *C. affinis* Dana. Biol Bull Mar Biol Lab Woods Hole 134:261–265
- Farrell AP (1982a) Cardiovascular changes in the unanethetized lingcod (*Ophiodon elongatus*) during short-term, progressive hypoxia and spontaneous activity. Can J Zool 60:933–941

Farrell AP (1982b) Lung development: biological and clinical perspectives. Academic Press, London

Farrell AP (1984) A review of cardiac performance in the teleost heart: intrinsic and humoral regulation. Can J Zool 62:523-536

Farrell AP (1991a) Circulation of body fluids. In: Prosser CL (ed) Environmental and metabolic animal physiology. Wiley/Liss, New York, pp 509–558

Farrell AP (1991b) Cardiac scope in lower vertebrates. Can J Zool 69:1981-1984

- Farrell AP (1993) Cardiovascular system. In: Evans DH (ed) The physiology of fishes. CRC Press, Boca Raton, pp 219–250
- Farrell AP, Jones DR (1992) The heart. In: Hoar WS, Randall DJ, Farrell AP (eds) Fish physiology: cardiovascular systems, vol 12. Academic Press, New York, pp 1–87
- Farrell AP, Randall DJ (1978) Air-breathing mechanisms in two Amazonian teleosts, Arapaima gigas and Hoplerythrinus unitaeniatus. Can J Zool 56:939–945
- Farrell AP, Daxboeck C, Randall DJ (1979) The effect of input pressure and flow on the pattern and resistance to flow in isolated perfused gills. J Comp Physiol 133:233-248
- Farrell AP, Sobin SS, Randall DJ, Crosby S (1980) Sheet blood flow in the secondary lamellae of teleost gills. Am J Physiol 239:R428–436

Farrelly CA, Greenaway P (1987) The morphology of and vasculature of the lungs and gills of the soldier crab *Mictyris longicarpus*. J Morphol 193:285–304

- Farrelly CA, Greenaway P (1992) Morphology and ultrastructure of the gills of terrestrial crabs (Crustacae, Gecarcinidae and Grapsidae): adaptations for air-breathing. Zoomorphology 112:39–49
- Farris R (1976) Systematics and ecology of Gnathostomulida from North Carolina and Bermuda. PhD Diss, University of North Carolina, Chapel Hill
- Fauvel P (1927) Polychaètes sédentaires. In: Office Central de Faunistique (ed) Faune de France, vol 16. Lechevalier, Paris, pp 1–494
- Fay P (1965) Heterotrophy and nitrogen fixation in *Chlorogloea fritschii*. J Gen Microbiol 39:11–20
- Fedde MR (1976) Respiration. In: Sturkie PD (ed) Avian physiology, 3rd edn. Springer, Berlin Heidelberg New York, pp 122–145
- Fedde MR (1980) The structure and gas-flow pattern in the avian respiratory system. Poult Sci 59:2642–2653
- Fedde MR, Kuhlmann WD (1978) Intrapulmonary carbon dioxide sensitive receptors: amphibians to mammals. In: Piiper J (ed) Respiratory function in birds, adult and embryonic. Springer, Berlin Heidelberg New York, pp 33–50
- Fedde MR, Faraci FM, Kilgore DL, Cardinet GH, Chatterjee A (1985) Cardiopulmonary adaptations in birds for exercise at high altitude. In: Giles R (ed) Circulation, respiration, and metabolism. Springer, Berlin Heidelberg New York, pp 149–163
- Fedde MR, Orr JA, Shams H, Scheid P (1989) Cardiopulmonary function in exercising barheaded geese during normoxia and hypoxia. Respir Physiol 77:239–262
- Feder ME (1976) Lungless, body size, and metabolic rate in salamanders. Physiol Zool 49:398–418
- Feder ME (1985) Effects of thermal acclimation on locomotor energetics and locomotor performance in a lungless salamander. Physiologist 28:342
- Feder ME, Burggren WW (1985a) Cutaneous gas exchange in vertebrates: design, patterns, control, and implications. Biol Rev 60:1-45

- Feder ME, Burggren WW (1985b) The regulation of cutaneous gas exchange in vertebrates. In: Giles R (ed) Circulation, respiration, and metabolism. Springer, Berlin Heidelberg New York, pp 101–112
- Feder ME, Pinder AW (1988) Ventilation and its effect on "infinite pool" exchangers. Am Zool 28:973-983
- Feder ME, Lynch JF, Shaffer HB, Wake DB (1982) Field body temperatures of tropical and temperate zone salamanders. Smithson Herp Inf Service 52:1-23
- Federspiel WJ (1989) Pulmonary diffusing capacity: implications of two-phase blood flow in capillaries. Respir Physiol 77:119–134
- Federspiel WJ, Popel AS (1986) A theoretical analysis of the effect of the particulate nature of blood on oxygen release in capillaries. Microvasc Res 32:164–189
- Feducia A (1980) The age of birds. Harvard University Press, Cambridge
- Felbeck H (1983) Sulfide oxidation and carbon fixation by the gutless clam Solemya reidi: an animal-bacteria symbiosis. J Comp Physiol 152:3-11
- Felbeck H, Childress JJ, Somero GN (1981) Calvin-Benson cycle and sulphide oxidation enzymes in animals from sulphide-rich habitats. Nature (Lond) 293:291–293
- Felder DL (1979) Respiratory adaptations of the estuarine mud shrimp, *Callianassa jamaicense* (Schmitt 1935) (Crustacea, Decapoda, Thalassinidea). Biol Bull Mar Biol Lab Woods Hole 157:125-138
- Felgenhauer BE, Abele LG (1983) Branchial water movement in the grapsid crab Sesarma reticulatum Say. J Crustacean Biol 3:187–195
- Fellenius ACB, Idstrom JP, Holm S (1984) Muscle respiration during exercise. Am Rev Respir Dis 129:S10–S12
- Feller G, Gerdy C (1987) Metabolic pattern of the heart of a haemoglobin-free Antarctic fish Channichyths rhinoceratus. Polar Biol 7:225-229
- Fenchel T (1992) What can ecologists learn from microbes: life beneath a square centimetre of sediment surface. Funct Ecol 6:499–507
- Fenchel T (1993) There are more small than large species? Oikos 68:375-378
- Fenchel T, Finlay BJ (1990a) Anaerobic free-living protozoa: growth efficiencies and the structure of anaerobic communities. FEMS Microbiol Ecol 74:269–276
- Fenchel T, Finlay BJ (1990b) Oxygen toxicity, respiration and behavioural responses to oxygen in free-living anaerobic ciliates. J Gen Microbiol 136:1953–1959
- Fenchel T, Finlay BJ (1991a) The biology of free-living anaerobic ciliates. Eur J Prostist 26:201–215
- Fenchel T, Finlay BJ (1991b) Endosymbiotic metanogenic bacteria in anerobic ciliates: significance of the growth efficiency of the host. J Protoz 38:18-32
- Fenchel T, Finlay BJ (1994) The evolution of life without oxygen. Am Nat 82:22-29
- Fenchel T, Riedl (1970) The sulfide system: a new biotic community underneath the oxidised layer of marine sand bottoms. Mar Biol 7:255–268
- Fenton MB, Bringham RM, Mills AM, Rautenbach IL (1985) The roosting and foraging areas of *Epomophorus wahlbergi* (Pteropodida) and *Scotophilus viridis* (Vespertilionide) in Kruger National Park, South Africa. J Mammal 66:461–468
- Ferguson MWJ (1992) Temperature-dependent sex determination in alligators: phenomenon, mechanism and evolution. Biol J Linn Soc 46:49–58
- Fernandes MN, Rantin FT (1985) Gill morphometry of the teleost *Hoplias malabaricus* (Bloch). Bol Fisiol Anim Univ São Paulo 9:57–65
- Fernandes MN, Rantin FT (1989) Respiratory responses of *Oreochromis niloticus* (Pisces, Cichlidae) to environmental hypoxia under different thermal conditions. J Fish Biol 35:509–519
- Fernandes MN, Rantin FT, Losa LL (1984) Estudo morfo-functional comparativo das brânquias de três espêcies fa familia Erythrinidae (Pisces; Teleosti): Hoplias malabaricus, Hoplias lacerdae e Hoplererythrinus unitaeniatus – implicaçõoes ecológicas. Cie Cult (Suppl) 36:608–609
- Fielder DR (1970) The feeding behaviour of the sand crab Scopimera inflata (Decapoda, Ocypodidae). J Zool (Lond) 160:35-49
- Fige FHJ (1936) The differential reaction of the blood vessels of a branchial arch of *Amblystoma tigrum* (Colorado Axolotl). I. The reaction of adrenaline, oxygen and carbon dioxide. Physiol Zool 9:79–101

- Fincke T, Paul R (1989) Booklung function in arachnids. III. The function and control of the spiracles. J Comp Physiol 159B:433-441
- Finlay BJ, Fenchel T (1993) Methanogens and other bacteria as symbionts of free-living anaerobic ciliates. Symbiosis 14:375–390
- Finley TN, Pratt SA, Ladman AJ, Brewer L, McKay MB (1968) Morphological and lipid analysis of the alveolar lining material in dog lung. J Lipid Res 9:351–365
- Firth JA, Farr A (1977) Structural features and quantitative age-dependent changes in the intervascular barrier of the guinea pig haemochorial placenta. Cell Tissue Res 184:507–516
- Fischkoff S, Vanderkooi JM (1975) Oxygen diffusion in the biological and artificial membrane determined by fluorochrome pyrene. J Gen Physiol 65:663–676
- Fish GR (1956) Soma aspects of the respiration of six species of fish from Uganda. J Exp Biol 33:186–195
- Fisher CR (1990) Chemoautotrophic and methanotrophic symbioses in marine invertebrates. Rev Aquatic Sci 2:399–436
- Fisher MR, Hand SC (1985) Chemoautotrophic symbionts in the bivalve *Lucida floridana* from seagrass beds. Biol Bull 167:445–459
- Fishman AP (1972) Pulmonary edema: the water-exchange function of the lung. Circulation XLVI:390-408
- Fishman AP (1983) Comparative biology of the lung. Am Rev Respir Dis 128:S90-S91
- Fishman AP, Becker EL, Fritts HW, Heinemann HO (1957) Apparent volumes of distribution of water, electrolytes and hemoglobin within the lung. Am J Physiol 188:95–101
- Fishman AP, Galante RJ, Pack AI (1989) Diving physiology: lungfish. In: Wood SC (ed) Comparative pulmonary physiology: current concepts, vol 39: Lung biology in health and disease. Marcel Dekker, New York, pp 645–676
- Fitch NA, Johnston IA, Wood RE (1984) Skeletal muscle capillary supply in fish that lacks respiratory pigments. Respir Physiol 57:201–211
- Flauenfelder H, Shigar SG, Wolynes PG (1991) The energy landscapes and motion of proteins. Science 254:1598–1602
- Fleay DH (1937) Nesting habits of the brush turkey. Emu 36:153-163
- Fletcher BD, Sachs BF, Kotas RV (1970) Radiologic demonstration of postnatal liquid in the lungs of newborn lambs. Paediatrics 46:252–258
- Floyd RA (1991) Oxidative damage to behaviour during aging. Science 254:1597
- Foelix RF (1982) Biology of spiders. Harvard University Press, Cambridge
- Foley JA, Kutzbach JE, Coe MT, Levis S (1994) Feedbacks between climate and boreal forests during the Halocene epoch. Nature (Lond) 371:52–54
- Folkow B, Neil E (1971) Circulation. Oxford University Press, New York
- Fons R, Sicart R (1976) Contribution à la connaissance du métabolisme énergétique chez deux Crocidurinae: *Suncus etruscus* (Savi 1822) et *Crocidura russula* (Herman 1780), Insectivora, Soricidae. mammalia. Mammalia 40:229–311
- Fontana W, Buss LW (1993) The arrival of the fitness: toward a theory of biological organization. Technical Report No SFI 93-09-055, Santa Fe Institute, Santa Fe, pp 1–71
- Forbes WA (1882) On the convoluted trachea of two species of manucode (*Manucodia atra*) and (*Phonygama gould*); with remarks on similar structures on other birds. Proc Zool Soc Lond 1882:347–353
- Forey P, Janvier P (1994) Evolution of the early vertebrates. Am Sci 82:554-565
- Forgue J, Massabuau JC, Truchot JP (1992a) When are resting water breathers lacking  $O_2$ ? Arterial  $PO_2$  at the anaerobic threshold in crab. Respir Physiol 88:247–256
- Forgue J, Truchot JP, Massabuau JC (1992b) Low arterial PO<sub>2</sub> in resting crustaceans is independent of blood oxygen-affinity. J Exp Biol 170:257–264
- Forman GL (1972) Comparative morphological and histochemical studies of stomachs of selected American bats. Univ Kans Sci Bull 49:591–729
- Forman HJ, Fisher AB (1981) Antioxidant defense. In: Glber DL (ed) Oxygen and living processes: an interdisplinary approach. Springer, Berlin Heidelberg New York, pp 235–249
- Forster ME, Davie PS, Davison W, Satchell GH, Wells RMG (1988) Blood pressures and heart rates in swimming hagfish. Comp Biochem Physiol 89:247–250
- Forster ME, Axelsson M, Farrell AP, Nilsson S (1991) Cardiac function and circulation in hagfishes. Can J Zool 69:1985–1992

- Forster RE (1964) Diffusion of gases. In: Fenn WO, Rahn H (eds) Handbook of physiology, sect 3. respiration, vol I. Amer Physiol Soc, Washington, DC, pp 839–872
- Forster RE (1973) Some principles governing maternal-foetal transfer in the placenta. In: Comline KS, Cross KW, Dawes GS, Nathanielz PW (eds) Foetal and neonatal physiology, Proc of the Sir Henry Barcroft Centenary Symposium. Cambridge University Press, Cambridge, pp 223-237
- Fox GA (1976) Eggshell: its ecological and physiological significance in a DDT contaminated tern population. Wilson Bull 88:459–477
- Fox H (1964a) The pattern of villous variability in the normal placenta. J Obstet Gynecol 71:749– 758
- Fox H (1964b) The villous trophoblast as an index of placental ischaemia. J Obstet Gynecol 71:885–983
- Fox H (1986) Pathology of the placenta. Clinical Obstet Gynecol Lond 13:501-509
- Fox HM (1921) Methods of studying the respiratory exchange in small aquatic organisms with particular reference to the use of flagellates as an indicator for oxygen consumption. J Gen Physiol 3:565–573
- Fox HM (1938) On blood circulation and metabolism of sabellids. Proc R Soc Lond 125B:554– 569
- Fox HM (1955) The effect of oxygen on the concentration of haem in invertebrates. Proc R Soc Lond 143B:203–214
- Fox HM, Taylor AER (1954) The tolerance of oxygen by aquatic invertebrates. Proc R Soc Lond 143B:214–225
- Fox HM, Simmonds BG, Washbourn R (1935) Metabolic rates of ephemerid nymphs from swiftly flowing and from still waters. J Exp Biol 12:179–184
- Fox GE, Stackebrandt E, Hespell RB, Gibson J, Maniloff J et al. (1980) The phylogeny of procaryocytes. Science 209:457–463
- Foxon GEH (1964) Blood and respiration. In: Moore JA (ed) Physiology of Amphibia. Academic Press, New York, pp 151–209
- Fraenkel G (1932) Der Atmungsmechanismus des Skorpions. Z Vergl Physiol 11:656-661
- Fraenkel G, Herford GVB (1938) The respiration of insects through the skin. J Exp Biol 15:266–280
- Fraga CG, Shigenaga MK, Park JW, Degan P, Ames BN (1990) Oxidative damage to DNA during aging: 8-hydroxy-2'-deoxyguanosine in rat organ DNA and urine. Proc Natl Acad Sci USA 87:4533-4537
- Frakes LA (1979) Climates throughout geologic time. Elsevier, Amsterdam
- Frappell P, Daniels CB (1991) Temperature effects on ventilation and metabolism in the lizard *Ctenophorus nuchalis*. Respir Physiol 86:257–270
- Frazzetta TH (1975) Complex adaptations in evolving populations. Sinauer, Sunderland
- Freadman MA (1981) Swimming energetics of striped bass (Morone saxatilis) and bluefish (Pomatomus saltatrix): hydrodynamic correlates of locomotion and gill ventilation. J Exp Biol 90:253-265
- Frederickson JK, Onstott TC (1996) Microbes deep inside the Earth. Sci Am 275:42-47
- Fredericq L (1878) Recherches sur la physiologie de poulpe commun. Arch Zool Exp Gen 7:535–583
- Freedman LS, Samuels I, Fish SA, Schwartz B, Lange M, Morgano L (1980) Sparing of the brain in neonatal undernutrition: amino acid transport and incorporation into brain and muscle. Science 207:902–904
- Freeman BA, Crapo JD (1982) Free radicals and tissue injury. Lab Invest 47:412-426
- Freeman BM, Misson BH (1970) pH, PO<sub>2</sub> and PCO<sub>2</sub> of blood from foetus and neonate of *Gallus domesticus*. Comp Biochem Physiol 33:763–772
- Fricke H (1988) Coelacanths, the fish that time forgot. Natl Geogr 173:824-838
- Fridovich I (1975) Superoxide dismutases. Am Rev Biochem 44:147-159
- Fridovich I (1976) Oxygen radicals, hydrogen peroxide, and oxygen toxicity. In: Pryor WG (ed) Free radicals in biology, vol 1. Academic Press, New York, pp 239–277
- Fridovich I (1978) Biology of oxygen radicals. Science 201:875-879
- Friedli B, Kent G, Olley P (1973) Inactivation of bradykinin in the pulmonary vascular bed of newborn and foetal lambs. Circ Res 33:421–427

- Friedmann EI (1982) Endolithic microorganisms in the Antarctic cold desert. Science 215:1045– 1053
- Friedmann EI, Ocampo R (1976) Endolithic blue algae in the dry valleys: primary producers in the Antarctic desert ecosystems. Science 193:1247–1249
- Frils-Christensen E, Lassen K (1991) Length of the solar cycle: an indicator of solar activity closely associated with climate. Science 254:698-700
- Frith HJ (1956) Breeding habits in the family Magapodiidae. Ibis 98:620-640
- Fritsche R (1990) Effects of hypoxia on blood pressure and heart rate in three marine teleosts. Fish Physiol Biochem 8:85–92
- Fritsche R, Thomas S, Perry SF (1993) Effects of serotonin on circulation and respiration in the rainbow trout *Oncorhynchus maykiss*. J Exp Biol 178:191-204
- Fry M, Jenkins DC (1984) Nematoda: aerobic respiratory pathways of adult parasitic species. Exp Parasitol 57:86-92
- Full RJ (1985) Exercising without lungs: energetics and endurance in a lungless salamander, *Plethodon jordani*. Physiologist 28:342
- Fung YB (1993) Biomechanics: mechanical properties of living tissues, 2nd edn. Springer, Berlin Heidelberg New York
- Fung YB, Sobin SS (1969) Theory of sheet flow in lung alveoli. J Appl Physiol 26:472-488
- Fuhrmann O (1914) Le genre Typhlonectes. Neuchatel Mem Soc Sci Nat 5:112-123
- Fustec A, Desbruyères D, Juniper SK (1987) Deep-sea hydrothermal vent communities at 13°N on the East Pacific rise: microdistributions and temporal variations. Biol Oceanogr 4:121– 164
- Futuyuma DJ (1986) Evolutionary biology, 2nd edn. Sinauer, Sunderland
- Gaehtgens P (1990) Avian versus mammalian blood: experimental observations and physiologic relevance. In: Sutton JR, Coates GC, Remmers JE (eds) Hypoxia: the adaptations. BC Decker, Toronto, pp 20–25
- Gaehtgens P, Will G, Schmidt F (1981) Comparative rheology of nucleated and non-nucleated red blood cells. II. Rheological properties of avian red cell suspensions in narrow capillaries. Pfluegers Arch Ges Physiol 390:283–287
- Gahlenbeck H, Bartels H (1970) Blood gas transport properties in gill and lung forms of the axolotol (*Ambryostoma mexicanum*). Respir Physiol 9:175–182
- Gahlenbeck H, Frerking H, Rathschlag-Schaefer AM, Bartels H (1968) Oxygen and carbon dioxide exchange across the cow placenta during the second part of pregnancy. Respir Physiol 4:119–131

Galla HJ (1993) Nitric oxide: an intracellular messanger. Angew Chem Int Ed Engl 32:378-380

- Gamble JC (1971) The responses of the marine amphipods *Corphium arenarium* and *C. volutator* to gradients and to choices of different oxygen concentrations. J Exp Biol 54:275–290
- Gameson ALH, Robertson KG (1955) The solubility of oxygen in pure water and sea water. J Appl Chem 5:502-523
- Gennon BJ, Campbell G, Randall DJ (1973) Scanning electron microscopy of the vascular casts for the study of vessel connections in a complex vascular bed – the trout gill. Proc Elect Microsc Soc Am 31:442-443
- Gannon BJ, Randall DJ, Browning J, Lester RJG, Rogers LJ (1983) The microvascular organization of the gas exchange organs of the Australian lungfish, *Neoceratodus forsteri* (Krefft). Aust J Zool 31:651–673
- Ganote CE (1983) Contraction band necrosis and irrevasible myocardial injury. J Mol Cell Cardiol 15:67-73

Gans C (1970) Respiration in the early tetrapods: the frog is a red herring. Evolution 24:740-751

- Gans C (1971) Strategy and sequence in the evolution of the external gas exchangers of ectothermal vertebrates. Forma Functio 3:66-104
- Gans C (1976) Ventilatory mechanisms and problems in some amphibious aspiration breathers (*Chelydra*, Caiman- Reptilia). In: Hughes GM (ed) Respiration of amphibious vertebrates. Academic Press, London, pp 357-374
- Gans C (1979) Momentarily excessive construction as the basis for protoadaptation. Evolution 331:227–233

- Gans C (1983) On the fallacy of perfection. In: Fay RR, Gourevitch G (eds) Perspectives on modern auditory research: papers in honour of EG Wever. Amphora Press, Groton, pp 101–114
- Gans C (1985) Vertebrate morphology: tale of a phoenix. Am Zool 25:689-694
- Gans C (1988) Adaptation and the form-function relation. Am Zool 28:681-697
- Gans C, Clark B (1976) Studies on ventilation of *Caiman crocodilus* (Crocodilia, Reptilia). Respir Physiol 26:285–301
- Gans C, Clark B (1978) Air flow in reptilian ventilation. Comp Biochem Physiol 60A;453-457
- Gans C, Hughes GM (1967) The mechanism of lung ventilation in the tortoise *Testudo graeca* Linne. J Exp Biol 47:1–20
- Garcia AGP, Basso NGD, Fonseca MEF, Zuardi JAT, Outanni HN (1991) Enterovirus associated placental morphology: a light virological, electron microscopic and immunologcic study. Placenta 12:533–547
- Gardner BG (1980) Tetrapod ancestry: a reappraisal. In: Panchen AL (ed) The terrestrial environment and the origin of land vertebrates. Academic Press, London, pp 135–207
- Garey WF, Rahn H (1970) Normal arterial gas tensions and pH and the breathing frequency of the electric eel. Respir Physiol 9:141–150
- Garland RJ, Milson WK (1994) End tidal gas composition is not correlated with episodic breathing in hibernating ground squirrels. Can J Zool 72:1141-1148
- Garland T, Huey RB (1987) Testing symmorphosis: does structure match functional requirements? Evolution 41:1404–1409
- Garland T, Else PL, Hulbert AJ, Tap P (1987) Effects of endurance training on activity metabolism of lizards. Am J Physiol 21:R450–R456
- Garlick RL, Davis BJ, Farmer M, Fyhn UEH, Noble RW et al. (1979) A foetal maternal shift in the oxygen equilibrium of haemoglobin from the viviparous caecilian, *Typhlonectes compressicauda*. Comp Biochem Physiol 62A:239-244
- Garrow JS, Hawes SF (1971) The relationship of the size and composition of the human placenta to its functional capacity. J Obstet Gynecol Br Commw 78:22–28
- Gatz RN, Crawford EC, Piiper J (1974) Respiratory properties of the blood of lungless and gillless salamander, *Desmognathus fuscus*. Respir Physiol 20:33-41
- Gatz RN, Glass ML, Wood SC (1987) Pulmonary function of the green sea turtle, *Chelonia* mydas. J Appl Physiol 62:459-463
- Gatzy JT (1975) Ion transport across the excised bullfrog lung. Am J Physiol 228:1162-1171
- Gaunt AS, Gans C (1969) Mechanics of resiration in the snapping turtle, *Chelydra serpentina* (Linne). J Morphol 128:195–228
- Gaunt AS, Gaunt SLL, Prange HD, Wasser JS (1987) The effects of tracheal coiling on the vocalization of the cranes (Aves: Gruidae), J Comp Physiol 161A:43-58
- Gaustad JE, Vogel SN (1982) High energy solar radiation and the origin of life. Origins Life 12:3– 8
- Gee JH (1976) Buoyancy and aerial respiration: factors influencing the evolution of reduced swim bladder volume of some Central American catfishes (Trichomycteridae, Callichthyidae, Loricariidae, Astroblepidae). Can J Zool 54:1030–1037
- Gee JH (1981) Coordination of respiratory and hydrostatic functions of the swim bladder in the Central American mudminnow, *Umbra limi*. J Exp Biol 92:37–53
- Gee JH, Gee PA (1995) Aquatic surface respiration, buoyancy control and the evolution of airbreathing gobies (Gobiidae: Pisces). J Exp Biol 198:79–89
- Gee JH, Graham JP (1978) Respiratory and hydrostatic functions of the intestine of the catfishes Hoplosternum thoracatum and Brochis splendens (Callichthydidae). J Exp Biol 74:1-16
- Geelhaar A, Weibel ER (1971) Morphometric estimation of pulmonary diffusing capacity. III. The effect of increased oxygen consumption in Japanese waltzing mice. Respir Physiol 11:354-366
- Gehr P, Erni H (1980) Morphometric estimation of pulmonary diffusion capacity in two horse lungs. Respir Physiol 41:199–210
- Gehr P, Bachofen M, Weibel ER (1978) The normal human lung: ultrastructure and morphometric estimation of diffusion capacity. Respir Physiol 32:121–140
- Gehr P, Schovic S, Burri PH, Claasen H, Weibel ER (1980) The lung of shrews: morphometric estimation of diffusion capacity. Respir Physiol 44:61-86

- Gehr P, Mwangi DK, Amman A, Maloiy GMO, Taylor CR, Weibel ER (1981) Design of the mammalian respiratory system: V. Scaling morphometric diffusing capacity to body mass: wild and domesic animals. Respir Physiol 44:61-86
- Gehrke PC, Fielder DR (1988) Effects of emperature and dissolved oxygen on heart rate, ventilation rate and oxygen consumption of spangled perch, *Leiopotherapon unicolor* (Günther 1859) (Percoidei, Teraponidae). J Comp Physiol 157:771–782
- Geiger R (1965) Das Klima der bodennahen Luftschicht. F Viewig, Brunswick, Germany. Translated as The climate near the ground. Harvard University Press, Cambridge
- Geiser J, Betticher DC (1989) Gas transfer in isolated lungs perfused with red cell suspension or haemoglobin solution. Respir Physiol 77:31-40
- George JC, Shah RV (1956) Comparative morphology of the lung in snakes with remarks on the evolution of the lung in reptiles. J Anim Morphol Physiol 3:1–7
- George JC, Shah RV (1965) Evolution of air sacs in Sauropodia. J Anim Morphol Physiol (Bombay) 12:255-263
- Gerth WA, Hemmingsen FA (1982) Limits of gas secretion by salting-out effect in the fish swim bladder rete. J Comp Physiol 146B:129–136
- Ghiretti F (1966) Respiration. In: Wilbur KM, Yonge CM (eds) Physiology of Mollusca. Academic Press, London, pp 175–298
- Ghiretti F, Ghiretti M (1975) Respiration. In: Fretter VV, Peake J (eds) Pulmonates, vol V. Academic Press, London, pp 33-52
- Giaver J, Keese CR (1989) Fractal motion of mammalian cells. Physica D 38:128-133
- Gibe J (1970) L'appareil respiratoire. In: Grasse PP (ed) Traité de zoologie, tome XIV, fascicule III. Masson, Paris, pp 499–520
- Giebisch GH, Granger JP, Greenleaf JE, Lydie RB, Mitchell RH et al. (1990) What's past is prologue. Physiologist 33:161-180
- Gibert CR (1993) Evolution and phylogeny. In: Evans DH (ed) The physiology of fishes. CRC Press, Boca Raton, pp 1–45
- Gilbert LI (1988) Developmental biology. Sinauer, Sutherland
- Gilbert RD, Cummings LA, Juchau MR, Longo LD (1979) Placental diffusing capacity and foetal development in exercising or hypoxic guinea pigs. J Appl Physiol 46:828–834
- Gilchrist BM (1954) Haemoglobin in Artemia. Proc R Soc Lond 143B:136-146
- Gillen RC, Riggs A (1973) Structure and function of the isolated haemoglobins of the American eel, *Anguilla rostrata*. J Biochem 248:1961–1969
- Gilles R, Pequeux A (1985) Ion transport in crustacean gills: physiological and ultrastructural approaches. In: Gilles R, Gilles-Baillien M (eds) Transport process, iono- and ultrastructural approaches. Springer, Berlin Heidelberg New York, pp 136–158
- Gillespie JR, Sagot JC, Gendner JP, Bouverot P (1982) Impedance of the lower respiratory system in ducks under four conditions: pressure breathing, anaesthesia, paralysis or breathing CO<sub>2</sub>enriched gas. Respir Physiol 47:177–191
- Gillis AM (1991) Can organisms direct their evolution? BioScience 41:202-205
- Giorgio P (1990) Adaptation to water and the evolution of echolocation in the cetacea. Ethol Ecol Evol 2:135–163
- Giovane A, Greco G, Maresca A, Tota B (1980) Myoglobin in the heart ventricle of tuna and other fishes. Experientia 36:219–220
- Girard H, Muffat-Joly M (1971) Evolution de la pression partielle d'oxygène et du pH sanguins chez l'embryon de poulet au cours de la croissance. Pfluegers Arch Ges Physiol 328:21– 35
- Glass ML, Johansen K (1981) Pulmonary diffusing capacity in reptiles (relations to temperature and oxygen uptake). J Comp Physiol 107:169–178
- Glass ML, Wood SC (1983) Gas exchange and control of breathing in reptiles. Physiol Rev 63:232-260
- Glass ML, Hicks JW, Riedesel ML (1976) Respiratory responses to long-term temperature exposure in the box turtle, *Terrapene ornata*. J Comp Physiol 131:353–359
- Glass ML, Wood SC, Hoyt RW, Johansen K (1979) Chemical control of breathing in the lizard *Varanus exanthematicus*. Comp Biochem Physiol 62A:999–1003
- Glass ML, Abe AS, Johansen K (1981a) Pulmonary diffusing capacity in reptiles: relations to temperature and  $O_2$ -uptake. J Comp Physiol 142:509–514

- Glass ML, Burggren WW, Johansen K (1981b) Pulmonary diffusing capacity of the bullfrog (*Rana catesbeiana*). Acta Physiol Scand 113:485–490
- Glass ML, Boutilier RG, Heisler N (1983) Ventilatory control of arterial PO<sub>2</sub> in the turtle, *Chrysemys picta bellii*: effects of temperature and hypoxia. J Comp Physiol 151:145–153
- Glass ML, Ishmatsu A, Johansen K (1986) Responses of aerial ventilation to hypoxia and hypercapnia in *Channa argus*, an air-breathing fish. J Comp Physiol B 156:425-430
- Glazier JB, Hughes JMB, Maloney JE, West JB (1967) Vertical gradient of alveolar size in lungs and of dogs frozen intact. J Appl Physiol 23:694–705
- Gleeson TT (1979) The effects of training and captivity on the metabolic capacity of the lizard *Sceloporus occidentalis*. J Comp Physiol 129:123–128
- Gleeson TT, Mitchell GS, Bennett AF (1980) Cardiovascular responses to graded activity in the lizards *Varanus* and *lguana*. Am J Physiol 239:R174–R179
- Gleick J (1987) Chaos. Sphere Books, New York
- Glenny RW (1992) Spatial correlation of regional pulmonary perfusion. J Appl Physiol 72:2378–2386
- Glenny RW, Robertson HT (1990) Fractal properties of pulmonary blood flow characterization of spatial heterogeneity. J Appl Physiol 69:532–545
- Glenny RW, Robertson HT (1991a) Fractal modeling of pulmonary blood flow heterogeneity. J Appl Physiol 70:1024–1030
- Glenny RW, Robertson HT (1991b) Gravity is a minor determinant of pulmonary blood flow distribution. J Appl Physiol 71:620–629
- Glenny RW, Robertson HT, Yamashiro S, Bassinthwaighte JB (1991) Applications of fractal analysis to physiology. J Appl Physiol 70:2351–2367
- Gnaiger E (1983) In situ measurement of oxygen profiles in lakes: microstratifications, oscillations, and the limits of comparison with chemical methods. In: Gnaiger E, Forstner H (eds) Polarographic oxygen sensors. Springer, Berlin Heidelberg New York, pp 245-264
- Gnaiger E (1991) Animal energetics at very low oxygen: information from calorimetry and respirometry. In: Waokes AJ, Grieshaber MK, Bridges CR (eds) Physiological strategies for gas exchange and metabolism. Cambridge University Press, Cambridge, pp 149–171
- Gnaiger E, Forstner H (1983) Polarographic oxygen sensors. Springer, Berlin Heidelberg New York
- Godfray HCJ (1997) Making life simpler. Nature (Lond) 387:351–352
- Godoy MP (1975) Familia Erythrinidae. In: Godoy MP (ed) Peixes do Brasil, suborden Characoidei, vol 3. Editora Fransiscana, Piracicaba, SP, pp 23-57
- Goldberger AL (1991) Is the normal heart beat chaotic or homeostatic? News Physiol Sci 6:87-91
- Goldberger AL, West BJ (1987) Fractals in physiology and medicine. Yale J Biol Med 60:421-435
- Goldberger AL, Bhargava V, West BJ, Mandell AJ (1985) On a mechanism of cardial electrical stability. Biophys J 48:525–528
- Goldberger AL, Rigney DR, West BJ (1990) Chaos and fractal in human physiology. Sci Am 262:34-41
- Golde LMG, Batenburg JJ, Robertson B (1994) The pulmonary surfactant system. News Physiol Sci 9:13–20
- Goldie RG, Bertram JF, Warton A, Papadimitriou JM, Paterson JW (1983) Pharmacological and ultrastructural study of alveolar contractile tissue in toad (*Bufo marinus*) lung. Comp Biochem Physiol 75:343-349
- Goldspink G (1985) Malleability of the motor system: a comparative approach. J Exp Biol 115:375-391
- Goldstein L (1982) Gill nitrogen excretion. In: Houlihan DF, Rankin JC, Shuttleworth HR (eds) Gills. Cambridge University Press. Cambridge, pp 93–206
- Gomi T (1982) Electron microscopic studies of the alveolar brush cell of the striped snake (Elaphe quadrivirgata). J Med Soc Toho Univ 29:481-102
- Goniakowska-Witalinska L (1973) Metabolism, resistance to hypotonic solutions and ultrastructure of erythrocytes of five amphibian species. Acta Biol Cracov 16:114–123
- Goniakowska-Witalinska L (1974) Respiration, resistance to hypotonic solutions and ultrastructure of erythrocytes of Salamandra salamandra. Bull Acad Pol Sci 22:59–75

- Goniakowska-Witalinska L (1978) Ultrastructure and morphometric study of the lung of the European salamander, Salamandra salamandra L. Cell Tissue Res 191:343-356
- Goniakowska-Witalinska L (1980a) Ultrastructural and morphometric changes in the lung of newt, *Triturus cistatus camifex* (Laur.) during ontogeny. J Anat 130:571-583
- Goniakowska-Witalinska L (1980b) Scanning and transmission electron microscopic study of the lung of the newt, *Triturus alpestris* Laur. Cell Tissue Res 205:133-145
- Goniakowska-Witalinska L (1982) Development of the larval lung of Salamandra salamandra L. Anat Embryol 164:113–137
- Goniakowska-Witalinska L (1986) Lung of the tree frog, *Hyla arborea*: a scanning and transmission electron microscope study. Anat Embryol 174:379–389
- Goniakowska-Witalinska L (1995) The histology and ultrastructure of the amphibian lung. In: Pastor LM (ed) Histology, ultrastructure and immunohistochemistry of the respiratory organs in non-mammalian vertebrates. Publicaciones Universidad de Murcia 1995, Murcia, Spain, pp 77–112
- Gonzalez NC, Clancy RL, Wagner PD (1993) Determinants of maximal oxygen uptake in rats acclimated to simulated altitude. J Appl Physiol 75:1608–1614
- Gonzalez RJ, McDonald DG (1992) The relationships between oxygen consumption and ion loss in fresh water fish. J Exp Biol 163:317-326
- Gonzalez-Grussi F, Boston RW (1972) The absorptive function of the neonatal lung. Ultrastructural study of horseradish peroxidase uptake at the onset of ventilation. Lab Invest 26:114– 121
- Goodrich ES (1930) Air bladder and lungs. In: Studies on the structure and development of vertebrates (Chap XI). Macmillan, London
- Gordon JE (1978) Structure; or, why things don't fall down. Plenum Press, New York
- Gordon MS, Boetius I, Evans DH, McCarthey R, Oglesby LC (1969) Aspects of the physiology of terrestrial life in amphibious fishes. I. The mudskipper *Periophthalmus sobrinus*. J Exp Biol 50:141–149
- Gordon MS, Ng WW, Yip AY (1978) Aspects of terrestrial life in amphibious fishes. III. The Chinese mudskipper, *Periophthalmus cantonensis*. J Exp Biol 72:57–75
- Gordon MS, Gabaldon DJ, Yip AY (1985) Exploratory observations on microhabitat selection within the intertidal zone by the chinese mudskipper fish *Periophthalmus cantonensis*. Mar Biol 85:209–215
- Gordon JW, Scangos GA, Plotkin DJ, Barbosa JA, Ruddle FH (1980) Genetic transformation of mouse embryos by microinjecion of purified DNA. Proc Natl Acad Sci USA 77:7380– 7384
- Gorge G, Chatelain P, Schaper J, Lerch R (1991) Effects of increasing degrees of ischemic injury on myocardial oxidative metabolism early after reperfusion in isolated rat hearts. Circ Res 68:1681–1692
- Gorin AB, Steward PA (1979) Differential permeability of endothelial and epithelial barriers to albumin flux. J Appl Physiol 47:1315–1324
- Gorr T, Kleinschmidt T, Fricke H (1991) Close tetrapod relationships of the coelacanth Latimeria indicated by heamoglobin sequences. Nature (Lond) 351:394-397
- Gosline JM, Steeves JD, Harman AD, deMont ME (1983) Patterns of circular and radial muscle activity in respiration and jetting of the squid *Loligo opalescens*. J Exp Biol 104:97–109
- Gould SJ (1966) Allometry and size in ontogeny and phylogeny. Biol Rev 41:587-640
- Gould SJ (1989) Wonderful world. Penguin Books, London
- Gould SJ (1994) The evolution of life on Earth. Sci Am 271:63-69
- Gould SJ, Eldridge N (1977) Punctuated equilibria: the tempo and mode of evolution reconsidered. Paleobiology 3:115–151
- Gould SJ, Lewontin RC (1979) The spandrels of San Macro and Panglossian paradigm: a critique of adaptationist programme. Proc R Soc Lond 205B:581–598
- Grace J, Lloyd J, McIntyre J, Miranda AC et al. (1995) Carbon dioxide uptake by undisturbed tropical rain forest in southwest Amazonia, 1992 to 1993. Nature 375:811-819
- Graham JB (1970) Preliminary studies on the biology of the amphibious clinid Mnierpes macrocephalus. Mar Biol 5:136-140
- Graham JB (1973) Terrestrial life of the amphibious fish *Mnierpes macrocephalus*. Mar Biol XXX 83–91

Graham JB (1974) Aquatic respiration of the seasnake, *Pelamis platurus*. Respir Physiol 21:1–7 Graham JB (1976) Respiratory adaptations of marine air-breathing fishes. In: Hughes GM (ed) Respiration of amphibious vertebrates. Academic Press, New York, pp 165–187

- Graham JB (1990) Ecological, evolutionary, and physical factors influencing aquatic animal respiration. Am Zool 30:137-146
- Graham JB (1994) An evolutionary perspective for biomdal respiration: a biological synthesis of fish air breathing. Am Zool 34:229–237
- Graham JB, Gee JH, Robinson FS (1975) Hydrostatic and gas exchange functions of the lung of the sea snake, *Pelamis platurus*. Comp Biochem Physiol 50:477–482
- Graham JB, Kramer DL, Pineda E (1977) Respiration of the air-breathing fish *Piabucina festae*. J Comp Physiol 122B:295–310
- Graham JB, Kramer DL, Pineda E (1978a) Comparative respiration of an air-breathing and nonair-breathing characoid fish and the evolution of aerial respiration in characins. Physiol Zool 51:279–288
- Graham JB, Rosenblatt RH, Gans C (1978b) Vertebrate air breathing arose in fresh waters and not in the ocean. Evolution 32:459–463
- Graham JB, Baird TA, Stockmann W (1987) The transition to air-breathing in fishes. IV. Impact of branchial specializations for air-breathing on the aquatic respiratory mechanisms and ventilatory costs of the swamp eel *Synbranchus marmoratus*. J Exp Biol 129:83–106
- Graham JB, Dudley R, Agullar NM, Gans C (1995) Implications of the late Paleozoic oxygen pulse of physiology and evolution. Nature (Lond) 375:117-120
- Graham JM (1949) Some effects of temperature and oxygen pressure on the metabolism and activity of the speckled trout, *Salvelunus fontinalis*. Can J Res 27:270–288
- Graiger E, Steinlechner P, Maran R, Méndez G, Eberl T, Margreiter R (1995) Control of mitochondrial and cellular respiration by oxygen. J Bioenerg Biomembr 27:583–596
- Grassé PP (1970) Traité de zoologie, vol XIV, Reptilia. Masson, Paris
- Grassle JF (1985) Hydrothermal vent animals: distribution and biology. Science 229:713-717
- Gratz RK, Ar A, Geiser J (1981) Gas tension profile of the lung of the viper, *Vipera xanthina palestinae*. Respir Physiol 44:165–171
- Gratz RK, Hutchison H (1977) Energetics for activity in the diamondback water snake, *Natrix rhombifera*. Physiol Zool 50:99–114
- Gratz RK, Crawford EC, Piiper J (1974) Metabolic and heart rate response of the plethodontid salamander *Desmognathus fuscus* to hypoxia. Respir Physiol 20:43–49
- Graur D (1993) Molecular phylogeny and the higher classification of eutherian mammals. Trends Ecol Evol 8:141–147
- Gray IE (1954) Comparative study of the gill area of marine fish. Biol Bull Mar Biol Lab Woods Hole 107:219–225
- Gray IE (1957) Comparative study of the gill area of crabs. Biol Bull Mar Biol Lab Woods Hole 112:34–42
- Gray J (1968) Animal locomotion. Weidenfeld and Nicolson, London
- Gray J (1985a) The microfossil record of early land plants: advances in understanding of early terrestriality, 1970–1984. Philos Trans R Soc Lond 309B:167–195
- Gray J (1985b) Ordovician-Silurian land plants: the interdependence of ecology and evolution. Spec Pap Palaeontol 32:281–295
- Gray MW (1992) The endosymbiont hypothesis revisited. Int Rev Cytol 141:233-357
- Green J, Corbet SA, Betney E (1973) Ecological studies on crater lakes in West Cameron. The blood of endemic cichlids in Barombi Mbo in relation to stratification and vertical distribution of the zooplankton. J Zool (Lond) 170:30–67
- Greenaway P, Taylor HH (1976) Aerial gas exchange in Australian arid-zone crab, Parathelphusa transversa, von Mertens. Nature (Lond) 262:711-713
- Greenaway P, Taylor HH, Bonaventura J (1983) Aquatic gas exchange in Australian fresh water/ land crabs of the genus *Holthuisana*. J Exp Biol 103:237–251
- Greenaway P, Morris S, MacMahon BR (1988) Adaptations to a terrestrial existence by the robber crab *Birgus latro*. II. *In vivo* respiratory gas exchange and transport. J Exp Biol 140:493-509
- Greenberg MJ (1985) Ex Bouillabaisse Lux: the charm of comparative physiology and biochemistry. Am Zool 25:737-749

Greenewalt CH (1960) Hummingbirds. Doubleday, New York

- Greenewalt CH (1968) Bird song: acoustics and physiology. Smithsonian Institution Press, Washington, DC
- Greenwood PH (1961) A revision of the genus *Dinotopterus* Blgr (Pisces, Clariidae) with notes on the comparative anatomy of the suprabranchial organs in the Clariidae. Bull Br Mus Nat Hist 7:217-241
- Greer AC, Parker F (1979) On the density of the New Guinea scincid lizard *Lygosoma fragile* Macleay 1877, with notes on its natural history. J Herpetol 13:221–225
- Gregersen MI, Rawson RA (1959) Blood volume. Physiol Rev 39:307-342
- Gregory RB (1977) Synthesis and total excretion of waste nitrogen by fish of the *Periophthalmus* (mudskipper) and Scartelaos families. Comp Biochem Physiol 57A:33-36
- Gregory EM, Fridovich I (1973) Induction of superoxide dismutase by molecular oxygen. J Bacteriol 114:543-548
- Gresson R (1927) On the structure of the branchiae of the gilled Oligochaete Alma nilotica. Ann Mag Nat Hist 19:348
- Grieshaber MK, Harding I, Kreutzer U, Pörtner HO (1994) Physiological and metabolic responses to hypoxia in invertebrates. Rev Physiol Biochem Pharmacol 125:44-144
- Griffin DR (1970) Migrations and homing of bats. In: Wimsatt WA (ed) Biology of bats, vol 1. Academic Press, London, pp 233-264
- Grigg GC (1965) Studies on the Queensland lungfish, *Neoceratodus forsteri* (Krefft). Aust J Zool 13:243–257
- Grigg GC (1969) The failure of oxygen transport in a fish at low levels of ambient oxygen. Comp Biochem Physiol 29:1253–1257
- Grigg GC, Harlow P (1981) A foetal-maternal shift of blood oxygen affinity in an Australian viviparous lizard, *Sphenomorphus quoyii* (Reptilia, Scincidae). J Comp Physiol B 142:495–499
- Grimes CB, Able KW, Jones RS (1986) Tilefish, Lopholatilus chamaeoleonticeps, habitat, behaviour and community structure in mid-Atlantic and southern New England waters. Environ Biol Fishes 15:273-292
- Grimme JD, Lane SM, Maron MB (1997) Alveolar liquid clearance in multiple nonperfused canine lung lobes. J Appl Physiol 82:348–353
- Groebe K, Thews G (1987) Time courses of erythrocytic oxygenation in capillaries of the lung: lower and upper bounds on red cell transit times. Adv Exp Med Biol 215:165–169
- Groom AC (1987) The microcirculatory society Eugene M. Landis award lecture. Microcirculation of the spleen: new concepts, new challenges. Microvasc Res 34:269–289
- Groom AC, Ellis CG, Potter RF (1984a) Microvascular architecture and red cell perfusion in skeletal muscle. Prog Appl Microcirc 5:64-83
- Groom AC, Ellis CG, Potter RF (1984b) Microvascular geometry in relation to modeling oxygen transport in contracted skeletal muscle. Am Rev Respir Dis 129:S6–S9
- Gros G (1991) The role of carbonic anhydrase within the tissues, with special reference to mammalian striated muscle. In: Woakes AJ, Grieshaber MK, Bridges CR (eds) Physiological strategies for gas exchange and metabolism. Cambridge University Press, Cambridge, pp 35–54
- Gross MG (1990) Oceanography: a view of the Earth, 5th edn. Prentice Hall, Englewood Cliffs
- Grote J (1967) Die Sauerstoffdiffusionkonstanten in Lungengewebe und Wasser und ihre Temperaturabhängigkeit. Pfuegers Arch Gesamte Physiol Menschen Tiere 295:245-254
- Grubb BR (1982) Cardiac output and stroke volume in exercising ducks and pigeons. J Appl Physiol 53:203-211
- Grubb BR (1983) Allometric relations of cardiovascular function in birds. Am J Physiol 14:H567-H572
- Grubb BR, Mills CD, Colacino JM, Schmidt-Nielsen K (1977) Effect of arterial carbon dioxide on cerebral blood flow in ducks. Am J Physiol 232:H596–H601
- Grubb BR, Colacino JM, Schmidt-Nielsen K (1978) Cerebral blood flow in birds: effects of hypoxia. Am J Physiol 234:H230-H243
- Grubb BR, Jones JH, Schmidt-Nielsen K (1979) Avian cerebral blood flow: influence of the Bohr effect on oxygen supply. Am J Physiol 236:H744–H753
- Gruson ES (1976) Checklist of birds of the world. William Collins, London

Guard CL, Murrish DE (1975) Effects of temperature on the viscous behaviour of blood from Antarctic birds and mammals. Comp Biochem Physiol 52A:287–290

- Guillette LJ (1982) The evolution of viviparity and placentation in the high elevation Mexican lizard, *Sceloporus aeneus*. Herpetologia 38:94–103
- Guillette LJ (1989) The evolution of vertebrate viviparity: morphological modifications and endocrine control. In: Wake D, Roth G (eds) Complex organismal functions: integration and evolution in vertebrates. J Wiley (Dahlem Workshop Report), Chichester, pp 219–233
- Guillette LJ (1991) The evolution of viviparity in amniote vertebrates: new insights, new questions. J Zool (Lond) 521-526

Guillette LJ (1993) The evolution of viviparity in lizards. BioScience 43:742-750

- Guillette LJ, Hotton N (1986) The evolution of mammalian reproductive characteristics in therapsid reptiles. In: Hotton N, MacLean PD, Roth JJ, Roth EC (eds) The ecology and biology of mammal-like reptiles. Smithsonian Institution, Washington, DC, pp 239-250
- Guillette LJ, Jones RE (1985) Ovarian, oviductal and placental morphology of the reproductively bimodal lizard species, *Sceloporus aeneus*. J Morphol 84:85–98
- Guimond RW, Hutchison HV (1972) Pulmonary branchial and cutaneous gas exchange in the mudpuppy, *Necturus maculosus maculosus* (Ranfinesque). Comp Biochem Physiol 42A:367–393
- Guimond RW, Hutchison HV (1973a) Trimodal gas exchange in the large aquatic salamander Siren lacertina. Comp Biochem Physiol 46A:249–268
- Guimond RW, Hutchison HV (1973b) Aquatic respiration: an unusual strategy in the hellbender, *Cryptobranchus alleganiensis alleganiensis* (Dudin). Science 182:1263-1265
- Guimond RW, Hutchison VH (1976) Gas exchange of the giant salamanders of North America. In: Hughes GM (ed) Respiration of amphibious vertebrates. Academic Press, New York, pp 313-338
- Gunsalus IC, Sligar SG (1978) Oxidation reduction by the  $P_{450}$  monoxygenase system. Adv Enzymol 47:1-44
- Guntheroth WG, Luchtel DL, Kawabori I (1982) Pulmonary microcirculation: tubules rather than sheet and post. J Appl Physiol 153:510-515
- Gupta KP, van Golen KL, Randerath E, Randerath K (1990) Age dependent covalent DNA alterations (I-compounds) in rat mitochondrial DNA. Mutat Res 237:17–27
- Gumett DA, Kurth WS, Roux A, Bolton SJ, Kennel CF (1996) Evidence for a magnetosphere at Ganymede from plasma-wave observations by Galileo spacecraft. Nature (Lond) 384:535– 537
- Gurtner GH, Traystman RJ, Bums B (1982) Interactions between placental O<sub>2</sub> and CO transfer. J Appl Physiol 52:479–487
- Gutman WF (1977) Phylogenetic reconstruction: theory, methodology, and application to chordate evolution. In: Hecht MK, Goody PC, Hecht BM (eds) Major patterns of vertebrate evolution. NATO Adv Study Inst Series, vol 14. Plenum Press, New York, pp 45–96
- Gutman WF, Bonik K (1981) Kritische Evolutionstheorie. Gerstenberg, Hildesheim
- Gutteridge JMC (1993) Free radicals in disease processes: a complication of cause and consequences. Free Radic Res Commun 19:141–158
- Guyer C, Slowinski JB (1993) Adaptive radiation and the tolology of large phylogenies. Evolution 47:253–263
- Haas JD (1976) Prenatal and infant growth and development. In: Baker PT, Little MA (ed) Man in the Andes: a multidisplinary study of high altitude Quechua. Hutchinson Ross, Stroudsburg, Pennsylvania, pp 161–179
- Haas JD, Frongillo EA, Stepick CD (1980) Altitude, ethnic and sex differences in birth weight in Bolivia. Hum Biol 52:459–477
- Hackstein JHP, Stumm CK (1994) Methane production in terrestrial arthropods. Proc Natl Acad Sci USA 91:5441–5445
- Hadley NF (1980) Surface waxes and integumentary permeability. Am Sci 68:546-553
- Hagerman L, Uglow RF (1985) Effects of hypoxia on the respiratory and circulatory regulation of *Nephros norvegicus*. Mar Biol 87:273-278
- Hainsworth FW (1981) Locomotion. In: Hainsworth FW (ed) Animal physiology: adaptation in function. Addison-Wesley, Reading, pp 259–292

- Hainsworth R (1986) Vascular capacitance: its control and importance. Rev Physiol Biochem Pharmacol 105:101-173
- Hakim A, Munshi JSD, Hughes GM (1978) Morphometrics of the respiratory organs of the Indian green snakehead fish, *Channa punctata*. J Zool (Lond) 184:519–543
- Haldane JS (1922) Respiration. Yale University Press, New Haven
- Hall VE (1931) The muscular activity and oxygen consumption of Urechis caupo. Biol Bull 61:400-416
- Hallam A (1987) End-Cretaceous mass extinction event: argument for terrestrial causation. Science 238:1237-1242
- Hallam JF, Dawson TJ, Holland RAB (1989) Gas exchange in the lung of a dasyurid marsupial: morphometric estimation of diffusion capacity and blood oxygen uptake kinetics. Respir Physiol 77:309-322
- Halliwell B (1978) Biochemical mechanisms accounting for the toxic action of oxygen on living organisms. The key role of superoxide dismutase. Cell Biol Int Rep 2:113–129
- Halliwell B (1994) Free radicals, antioxidants and human disease: curiosity, cause or consequence? Lancet 344:721-724
- Halliwell B, Gutteridge JMC (1985) Free radicals in biology and medicine. Clarendon, Oxford
- Hamlett WC (1986) Prenatal nutruent absorptive structures in selachains. In: Uyeno T, Arai T, Taniuchi T, Matsuura K (eds) Indo-Pacific fish biology. Ichtyol Soc Japan, Tokyo, pp 333– 344
- Hamlett WC (1987) Comparative morphology of the elasmobranch placental barrier. Arch Biol Brux 98:135–162
- Hamlett WC (1989) Evolution and morphogenesis of the placenta in sharks. J Exp Zool (Suppl) 2:35–52
- Hamlin RL, Kondrich RM (1969) Hypertension, regulation of heart rate, and possible mechanism contributing to aortic rupture in turkeys. Proc Fed Am Soc Exp Biol 28:451–456
- Hammen CS (1969) Metabolism of the oyster, Crassostrea virginica. Am Zool 9:309-318
- Hammen CS (1976) Respiratory adaptations: invertebrates. In: Wiley M (ed) Estuarine processes. Academic Press, New York, pp 347-355
- Hammersley JR, Olson DE (1992) Physical models of the smaller pulmonary airways. J Appl Physiol 72:2402-2414
- Hammond P (1992) Species inventory. In: Groombridge B (ed) Global biodiversity: status of the Earth's living resources. Chapman and Hall, London, pp 17-39
- Handerson LJ (1913) The fitness of the environment. Macmillan, New York
- Handy RD (1989) The ionic composition of rainbow trout body mucus. Comp Biochem Physiol 93A:571–575
- Hansell DA, Bates NR, Carson CA (1997) Predominance of vertical loss of carbon from the surface waters of the equatorial Pacific Ocean. Nature (Lond) 386:59–61
- Hansen CA, Sidell BD (1983) Atlantic hagfish cardiac muscle: metabolic basis of tolerance to anoxia. Am J Physiol 244:R356-R362
- Hansen HJ (1893) Organs and characters in different orders of Arachnida. Entomol Medd 4:135-144
- Hansen VK, Wingstrand KG (1960) Further studies on the nonnucleated erythrocytes of *Maurolieus mulleri* and comparison with blood cells of related fish. AF Host, Dana Report No 54, Copenhagen
- Hanson D, Johansen K (1970) Relationship of gill ventilation and perfusion in dogfish, Squalus suckleyi. J Fish Res Board Can 27:551-564
- Haq BU, Hardenbol J, Vail PR (1987) Chronology of fluctuating sea levels since the Triassic. Science 235:1156-1167
- Hardisty MW (1979) Biology of cyclostomes. Chapman and Hall, London
- Harms JW (1932) Die Realisation von Genen und die konsekutive Adaptation. II. *Birgus latro* L. als Landkrebs und seine Beziehungen zu den Coenabiten. Z Wiss Zool 140:167–190
- Harnisch O (1937) Primäre und sekundäre Oxybiose wirbelloser Tiere. Verh Dtsch Zool Ges 1937:129–136
- Harrison JF, Fewell JH, Roberts SP, Hall HG (1996) Achievement of thermal stability by varying metabolic heat production in flying honeybees. Science 274:88–90

- Harrison P, Zummo G, Farina F, Tota B, Johnson IA (1991) Gross anatomy, myoarchitecture, and ultrastructure of the heart ventricle in hemoblobinless icefish *Chaenocephalus aceratus*. Can J Zool 69:1339–1347
- Harrison RJ, Ridway SH, Joyce PL (1972) Telemetry of heart rate in diving seals. Nature (Lond) 238:280
- Hart JS, Fisher HD (1964) The question of adaptations to polar environments in marine mammals. Fed Proc 23:1207-1214
- Hartman FA (1954) Cardiac and pectoral muscles of trochlids. Auk 71:467-469
- Hartman FA (1955) Heart weight in birds. Condor 57:221-238
- Hartman FA (1961) Locomotor mechanisms of birds. Smithson Misc Collect 143:1-91
- Hartman FA (1963) Some flight mechanisms of bats. Ohio J Sci 63:59-65
- Hartnoll RG (1988) Evolution, systematics, and geographical distribution. In: Burggren WW, McMahon BR (eds) Biology of the land crabs. Cambridge University Press, Cambridge, pp 6– 54
- Hartvig M, Weber RE (1984) Blood adaptations for maternal-foetal oxygen transfer in the viviparous teleost, *Zoarces viviparous* L. In: Seymour RS (ed) Respiration and metabolism of embryonic vertebrates. Junk, Dordrecht, pp 17–30
- Harvey EN (1928) The oxygen consumption of luminous bacteria. J Comp Physiol 11:469-475
- Harvey HW (1957) The chemistry and fertility of sea waters. Cambridge University Press, London
- Harvey PH (1993) The ecology of evolutionary succession. Curr Biol 3:106-108
- Hashimoto K, Yamaguchi Y, Matsura F (1960) Comparative studies on the haemoglobins of salmon. IV. Oxygen dissociation curve. Bull Jpn Soc Sci Fish 26:827-834
- Hashimoto T, Gomi T, Kimura A, Tsuchiya H (1983) Light electron microscopic study of the lung of the giant salamander *Megalobateracus japonicus davidanus*. J Med Soc Toho Univ 29:52–69
- Hastings RH, Powell FL (1986) Physiological dead space and effective parabronchial ventilation in ducks. J Appl Physiol 60:85–91
- Hastings RH, Powell FL (1987) High frequency ventilation of ducks and geese. J Appl Physiol 63:413-417
- Haswell MS, Randall DJ (1978) The pattern of carbon dioxide excretion in the rainbow trout Salmo gairdneri. J Exp Biol 72:17-24
- Haughton TM, Kerkut GA, Munday KA (1958) The oxygen dissociation and alkaline denaturation of haemoglobin from two species of earthworms. J Exp Biol 35:360–368
- Hawking S (1993) Black holes and baby universes and other essays. Bantam Books, New York
- Hawkins AJS, Jones MB (1982) Gill area and ventilation in two mud crabs, *Helice crassa* Dana (Grapsidae) and *Macrophthalmus hirtipes* (Jacquinot) (Ocypodidae). J Exp Mar Biol Ecol 60:103-118
- Hawksworth DL (1991) The fungal dimension of biodiversity: magnitude, significance, and conservation. Mycol Res 95:641–655
- Hayes B (1994) Nature's algorithms. Am Sci 82:206-210
- Hayes JD, Imbrie J, Shackleton NJ (1976) Variations in the Earth's orbit: pacemaker of the ice ages. Science 194:1121-1131
- Hayes K, Gibas H (1971) Placental cytomegalovirus infection without foetal involvement following primary infection in pregnancy. J Paediatr 79:401-405
- Hayward B, Davis R (1964) Flight speeds in western bats. J Mammal 45:236-242
- Hazel JR (1993) Thermal biology. In: Evans DH (ed) The physiology of fishes. CRC Press, Boca Raton, pp 427–467
- Hazelhoff EH (1939) Über die Ausnützung des Sauerstoffs bei verschiedenen Wassertieren. Z Vergl Physiol 26:306–327
- Hearse DJ, Humprey SM, Chain EB (1973) Abrupt reoxygenation of the anoxic potassiumarrested perfused rat heart: a study of myocardial enzyme release. J Mol Cell Cardiol 5:395– 407
- Heath D, Williams DR (1981) Man at high altitude: the pathophysiology of acclimatization and adaptation. Churchill Livingstone, Edinburgh
- Heath D, Williams DR (1989) High altitude pulmonary oedema. High altitude medicine and pathology. Butterworths, London, pp 132–148

- Heath D, Williams DR, Dickson J (1984) The pulmonary arteries of the yak. Cardiovasc Res 18:133-139
- Heatwole H (1981) Role of the saccular lung in the diving of the sea krait, *Laticuda colubrina* (Serpentes: Laticaudidae). Aust J Herpetol 1:11-16
- Hedges SB, Sibley CG (1994) Molecules vs. morphology in avian evolution: the case of the "pelecaniform" birds. Proc Natl Acad Sci USA 91:9861–9865
- Hedrick MS, Duffield DA (1986) Blood viscosity and optimal hematocrit in deep-diving mammal, the northern elephant seal (*Mirounga angustirostris*). Can J Zool 64:2081–2085
- Hedrick MS, Duffield DA (1991) Hematological and rheological characteristics of blood in seven marine mammal species: physiological implications for diving behaviour. J Zool (Lond) 225:273-283
- Hedrick MS, Jones DR (1993) The effects of altered aquatic and aerial respiratory gas concentrations on air-breathing patterns in a primitive fish *Amia calva*. J Exp Biol 181:81–94
- Hedrick MS, Duffield DA, Cornell LH (1986) Blood viscosity and optimal hematocrit in a deepdiving mammal, the northern elephant seal (*Mirounga angustirostris*). Can J Zool 64:2081– 2085
- Hedrick MS, Burleson ML, Jones DR, Milsom WK (1991) An examination of central chemosensitivity in an air-breathing fish (*Amia calva*). J Exp Biol 155:165–174
- Heijden FL (1981) Compensation mechanisms for experimental reduction of the functional capacity of the guinea pig placenta. Acta Anat 111:359–366
- Heinemann HO, Fishman AP (1969) Nonrespiratory functions of mammalian lung. Physiol Rev 49:1–61
- Heinrich B (1983) Do bumblebees forage optimally, and does it matter? Am Zool 23:273-281
- Heinrich B (1992) The hot-blooded insects. Harvard University Press, Cambridge
- Heisler N (1982a) Intracellular and extracellular acid-base regulation in the tropical fresh water teleosts *Synbranchus marmoratus* in response to the transition from water-breathing to airbreathing. J Exp Biol 99:9–28
- Heisler N (1982b) Transepithelial ion transfer processes as mechanisms for fish acid-base regulation in hypercapnia and lactacidosis. Can J Zool 60:1108-1122
- Heisler N (1984) Acid-base regulation in fishes. In: Hoar WS, Randall DJ (eds) Fish physiology vol XA. Academic Press, New York, pp 315–401
- Heisler N (1989) Interactions between gas exchange, metabolism and ion transport in animals: an overview. Can J Zool 67:2923–2935
- Heisler N, Forcht G, Ultsch GR, Anderson JF (1982) Acid-base regulation in response to environmental hypercapnia in two aquatic salamanders, *Siren lacertina* and *Amphiuma means*. Respir Physiol 49:141–158
- Helbacka NV, Caserline JL, Smith CJ (1963) The effect of high CO<sub>2</sub> atmosphere on the laying hen. Poult Sci 42:1082–1097
- Hellems HK, Haynes FW, Dexter L (1949) Pulmonary capillary pressure in man. J Appl Physiol 2:24–52
- Hellin B, Chardon M (1981) Observations sur le trajet de l'air durant la respiration aérienne chez *Clarias lazera* Cuvier et Valenciennes, 1840. Ann Soc R Zool Belg 113:97–106
- Hellman LM, Kobayashi M, Tolles WE, Cromb E (1970) Ultrasonic studies of the volumetric growth of the human placenta. Am J Obstet Gynecol 108:740–750
- Hemmingsen EA (1963) Enhancement of oxygen transport by myoglobin. Comp Biochem Physiol 10:239-244
- Hemmingsen EA (1965) Transfer of oxygen through solutions of heme pigments. Acta Physiol Scand Suppl 246
- Hemmingsen EA, Douglas EL (1970) Respiratory characteristics of the haemoglobin-free fish, Chaenocephalus aceratus. Comp Biochem Physiol 32:733-744
- Hemmingsen EA, Douglas EL, Johansen K, Millard RW (1972) Aortic blood flow and cardiac output in the haemoglobin-free fish, *Chaenocephalus aceratus*. Comp Biochem Physiol A 43:1045–1051
- Hendry GAF (1993) Oxygen, free radicals processes and seed longevity. Seed Sci Res 3:141-153
- Henk WG, Haldiman JT (1990) Microanatomy of the lung of the bowhead whale, *Balaena mysticetus*. Anat Rec 226:187-197

Henry JD, Fedde MR (1970) Pulmonary circulatory time in the chicken. Poult Sci 49:1286-1293

Henry RP (1994) Morphological, behavioural, and physiological characterization of bimodal breathing crustaceans. Am Zool 34:205-215

Henry RP, Perry HM, Trigg CB, Handley HL, Krarup A (1990) Physiology of two species of deep water crabs, *Chaceon fenneri* and *C. quiquedens*: gill morphology, and haemolymph ionic and nitrogen concentrations. J Crust Biol 10:375–381

- Henze M (1910) Ueber den Einfluss des Sauerstoffdrucks auf den Gaswechsel einiger Meerrestiere. Biochem Z 26:255–278
- Herbert CV, Jackson DC (1985) Temperature effects on the responses to prolonged submergence in the turtle *Chrysemys picta belii*. II. Metabolic rate, blood acid-base and ionic changes, and cardiovascular function in aerated and anoxic water. Physiol Zool 58:670– 681
- Herman AB, Spicer RA (1996) Paleobotanical evidence for a warm Cretaceous Arctic ocean. Nature (Lond) 380:330-333
- Herreid CF (1980) Hypoxia in invertebrates. Comp Biochem Physiol 67A:311-320
- Herreid CF, Full RJ (1988) Energetics of locomotion. In: Burggren WW, McMahon BR (eds) Biology of the land crabs. Cambridge University Press, Cambridge, pp 333–377
- Herreid CF, Lee LW, Spampata R (1981) How do spiders breathe? Am Zool 21:917
- Hers MJ (1943) Relation entre respiration et circulation chez *Anodonta cygnea* L. Ann Soc R Zool Belg 74:45–54
- Hershenson MB, Abe MK, Kelleher MD, Naureckas ET, Garland A et al. (1994) Recovery of airway structure and function after hyperoxic exposure in immature rats. Am J Respir Crit Care Med 149:1663-1669
- Heth G, Frankenberg E, Nevo E (1985) Adaptive optimal sound for vocal communication in tunnels of a subterranean mammal (*Spalax ehrenbergi*): a physcial analysis. Experientia 42:1287–1289
- Heusner AA (1983) Body size, energy metabolism and the lung. J Appl Physiol 54:867-873
- Hewitt JA, Kilmartin JV, Ten Eyck LF, Perutz MF (1972) Noncooperativty of the αβ-dimer in the reaction of haemoglobin with oxygen. Proc Natl Acad Sci USA 69:302–307
- Heymann MA, Hoffman JIE (1984) Pulmonary hypertension in the newborn. In: Weir EK, Reeves JT (eds) Pulmonary hypertension. Futura, New York, pp 45–71
- Hickman CS (1984) Form, function, and evolution of gastropod filters. Am Malacol Bull 3:95

Hickman CS (1987) Analysis of form and function in fossils. Am Zool 28:775-793

- Hicks JW, Wood SC (1985) Temperature regulation in lizards: effects of hypoxia. Am J Physiol 248:R595–R600
- Hiebl I, Braunitzer G, Schneeganss D (1987) The primary structures of the major and minor haemoglobin-components of adult Andean goose (*Chleophaga melanoptera*, anatidae) the mutation  $\alpha$ -Leu-Ser in position 55 of the  $\beta$ -chains. Biol Chem 368:1559–1569
- Hightower A, Burke JD, Haar JL (1975) A light microscopic study of the respiratory epithelium of the adult newt, *Notophthalmus viridescens*. Can J Zool 53:465–478
- Hikida RS, Staron RS, Hagerman FC, Sherman WM, Costill DL (1983) Muscle fiber necrosis associated with human marathon runners. J Neurol Sci 59:185–203
- Hildebrand M (1959) Motions of running cheetah and horse. J Mammal 40:481-495
- Hildebrand M (1961) Further studies on the motion of the cheetah. J Mammal 42:84-91
- Hill AV (1950) The dimensions of animals and their muscular dynamics. Sci Prog 38:209-230
- Hill B (1981) Respiratory adaptations of three species of Upogebia (Thalassinidea, Crustacea)
- with reference to low tide periods. Biol Bull Mar Biol Lab Woods Hole 160:272-279
- Hill EP, Power GG, Longo LD (1973) Mathematical simulation of pulmonary  $\rm O_2$  and  $\rm CO_2$  exchange. Am J Physiol 224:909–917
- Hill HAO (1978) The superoxide iron and the toxicity of molecular oxygen. In: Williams RJP, Da Silva JRRF (eds) New trends in bioinorganic chemistry. Academic Press, New York, pp 173– 208
- Hill JE (1944) Rodent miners. Nat Hist 53:21
- Hillman SS (1987a) Dehydrational effects of cardiovascular and metabolic capacity of two amphibians. Physiol Zool 60:608-613
- Hillman SS (1987b) The roles of oxygen delivery and electrolyte levels in the dehydrational death of *Xenopus laevis*. J Comp Physiol 128:169–178

- Hillman SS, Withers PC, Hedrick MS, Kimmel PB (1985) The effects of erythrocythemia on blood viscosity, maximal systemic oxygen transport capacity and maximal rates of oxygen consumption in an amphibian. J Comp Physiol 155B:577–581
- Hills BA (1971) Geometric irreversibility and compliance hysteresis in the lung. Respir Physiol 13:50–61
- Hills BA (1972) Diffusion and convection in lungs and gills. Respir Physiol 14:105-114
- Hills BA (1988) The biology of the surfactant. Cambridge University Press, Cambridge
- Hills BA (1996) Effects of gill dimensions on respiration. In: Munshi JSD, Dutta HM (eds) Fish morphology: horizon of new reseach. Science Publishers, Lebanon NH, pp 235–247
- Hills BÅ, Hughes (1970) A dimensional analysis of oxygen transfer in the fish gill. Respir Physiol 9:126–140
- Hilpert P, Fleischmann RG, Kempe D, Bartels H (1963) The Bohr effect related to blood and erythrocytes pH. Am J Physiol 205:337–340
- Hinds DS, Calder WA (1971) Tracheal dead space in the respiration of birds. Evolution 25:429–440
- Hinton HE (1953) Some adaptations of insects to environments that are alternately dry and flooded, with some notes on the habits of the Stratiomyidae. Trans Soc Br Entomol 11:209–227
- Hinton HE (1966) Plastron respiration. Nature (Lond) 209:220-221
- Hinton HE (1971) Reversible suspension of metabolism. In: Marois M (ed) From theoretical physics to biology. Editions du Centre National de la Recherche Scientifique, Paris, pp 332–357
- Hinton HE (1976) Plastron respiration in bugs and beetles. J Insect Physiol 22:1529-1550
- Hitschold T, Beck T, Muntefering H, Berle P (1992) Plazentamorphometrie bei diastolischem Null- und Negativeflow der Nabelarterien. Geburtsh Frauenheilkd 52:270–274
- Hitzig BM, Jackson DC (1978) Central chemical control of ventilation in the unanaesthetized turtle. Am J Physiol 235:R257-R264
- Hlastala MP, Standaert TA, Pierson DJ, Luchtel DL (1985) The matching of ventilation and perfusion in the lung of the tegu, *Tupinambis nigropunctus*. Respir Physiol 60:277– 294
- Hlastala MP, Bernard SL, Erickson H et al. (1996) Pulmonary blood flow distribution in standing horses is not dominated by gravity. J Appl Physiol 81:1051–1061
- Ho YS (1994) Transgenic models for the study of lung biology and disease. Am J Physiol 266:L319-L353
- Ho YS, Dey MS, Crapo JD (1996) Antioxidant enzyme expression in rat lungs during hyperoxia. Am J Physiol 270:L810–L818
- Hoagland M, Dodson B (1995) The way life works. Ebury Press, London
- Hochachka PW (1979) Cell metabolism, air breathing, and the origins of endothermy. In: Wood SC, Lenfant C (eds) Evolution of respiratory processes: a comparative approach. Marcel Dekker, New York, pp 253–288
- Hochachka PW (1987) Limits: how fast and how slow muscle metabolism can go. In: Benzi G (ed) Advances in myochemistry, vol 1. John Libbey, Eurotext, New York, pp 3–12
- Hochachka PW (1988) Metabolic suppression and O2 availability. Can J Zool 66:152-158
- Hochachka PW, Guppy M (1987) Metabolic arrest and the control of biological time. Harvard University Press, Cambridge
- Hochachka PW, Fields J, Mustafa T (1973) Animal life without oxygen: basic biochemical mechanisms. Am Zool 13:543-555
- Hochachka PW, Guppy M, Goderley HE, Storey KB, Hulbert WC (1978) Metabolic biochemistry of water- vs. air-breathing fishes: muscle enzymes and ultrastructure. Can J Zool 56:736– 750
- Hochachka PW, Emmett B, Suarez RK (1988) Limits and contraints in the scaling of oxidative and glycolytic enzymes in homeotherms. Can J Zool 66:1128–1138
- Hock RJ (1951) The metabolic rates an body temperatures of bats. Biol Bull Mar Biol Lab Woods Hole 101:289–299
- Hock RJ (1964) Animals in high altitudes: reptiles and amphibians. In: Dill DB, Adolph EF, Wilber CG (eds) Handbook of physiology, sect 4, adaptation to environment. American Physiological Society, Washington DC, pp 841–842

- Hoeger U, Mommsen TP (1985) Role of free amino acids in the oxidative metabolism of cephalopod hearts. Circulation, respiration, and metabolism. Springer, Berlin Heidelberg New York, pp 367-376
- Hoese B (1983) Struktur und Entwicklung der Lungen der Tylidae (Crustacea, Isopoda, Oniscoidea). Zool Jb Anat 109:487-501
- Hoffman LH (1970) Placentation in the garter snake, Thamnophis sirtalis. J Morphol 131:57-88
- Hogg JC, Nepszy S (1969) Regional lung volume and pleural pressure gradient estimated from lung density in dogs. J Appl Physiol 27:198–203
- Hogg JC, McLean T, Martin BA, Wiggs B (1988) Erythrocyte transit and neutrophil concentration in the dog lung. J Appl Physiol 65:1217–1225
- Holden T (1993) Giants in the Earth. Velikovskian 1(4):7-17
- Holeton GF (1970) Oxygen uptake and circulation by a haemoglobinless Antarctic fish (*Chaenocephalus aceratus*, Lonnberg) compared with three red-blooded Antarctic fish. Comp Biochem Physiol 34:457–471
- Holeton GF (1974) Metabolic cold adaptation of polar fish: fact or artifact? Physiol Zool 47:137–154
- Holeton GF, Heisler N (1983) Contribution of net ion transfer mechanisms to acid-base regulation after exhausting activity in the larger spotted dogfish (*Scyliorhinus stellaria*). J Exp Biol 103:31–46
- Holland HD (1978) The chemistry of the atmosphere and the oceans. Wiley, New York
- Holland JB (1984) The chemical evolution of the atmosphere and oceans. Princeton University Press, Princeton
- Holland RAB, Forster RE (1966) The effect of size of red cells on the kinetics of their oxygen uptake. J Gen Physiol 49:727–742
- Holle JP, Meyer M, Scheid P (1977) Oxygen affinity of duck blood determined by in vivo and in vitro techniques. Respir Physiol 29:355–361
- Hollinger DY, Kellihaer FM, Schulze ED, Köstner MM (1994) Coupling of tree transpiration to turbulence. Nature (Lond) 371:60–62
- Holm-Hansen O (1968) Ecology, physiology, and biochemistry of blue-green algae. Annu Rev Microbiol 22:47–70
- Holmyard EJ, Palmer WG (1952) A higher school inorganic chemistry. JM Dent, London
- Homberger DG (1988) Models and tests in functional morphology: the significance of description and integration. Am Zool 28:217–229
- Hong SK, Rahn H (1967) The diving women of Korea and Japan. Sci Am 216:34-43
- Hook GR, Bastacky J, Conhaim RL, Staub NC, Hayes TL (1987) A new method for pulmonary edema research: scanning electron microscopy of frozen hydrated edematous lung. Scanning 9:71–79
- Hoppeler H, Kayar SR, Claassen H, Uhlman E, Karas RH (1987) Adaptive variation in the mammalian respiratory system in relation to energetic demand: III. Skeletal muscles: setting the demand for oxygen. Respir Physiol 69:27–46
- Horn MH, Gibson RN (1989) Intertidal fishes. In: Gould JL, Gould CG (eds) Life at the edge: readings from Scientific American. WH Freeman, New York, pp 59–67

Horn MH, Messer KS (1992) Fish guts as chemical reactors: a model of the alimentary canals of marine herbivorous fishes. Mar Biol 113:527–535

- Hora SL (1935) Physiology, bionomics and evolution of the air-breathing fishes of India. Trans Natl Inst Sci India 1:1–16
- Horsfield K (1981) The science of branching systems. In: Scadding JG, Cumming G (eds) Scientific foundation of respiratory medicine. Heinemann, London, pp 45–54
- Horsfield K, Thurlbeck A (1981) Relation between diameter and and flow in the bronchial tree. Bull Math Biol 43:681–691
- Horsfield K, Woldenberg MJ (1986) Branching ratio and growth of tree-like structures. Respir Physiol 63:97–107
- Horvath SM, Borgia JF (1984) Cardiopulmonary gas transport and aging. Am Rev Respir Dis 129:S68–S71
- Hosler P (1977) Castrophic chemical events in the history of the oceans. Nature (Lond) 267:403–408

- Hoss LM (1973) A study of the benthos of an anoxic marine basin and factors affecting its distribution. MSc Thesis, Dalhouse Univ, Nova Scotia
- Hossler FE (1980) Gill arch of the mullet, *Mugil cephalus*. III. Rate of response to salinity change. Am J Physiol Regul Integr Comp Physiol 238:379–398
- Hossler FE, Harpole JH, King JA (1986) The gill arch of the striped bass, *Morone saxatilis*. I. Surface ultrastructure. J Submicrosc Cytol 18:519–528
- Hou PCL, Burggren WW (1991) Hemodynamic development in anuran larvae *Xenopus laevis*. Am Zool 31:76A
- Houde P (1986) Ostrich ancestors found in the Northern Hemisphere suggest new hypothesis of ratite origin. Nature (Lond) 324:563–565
- Houlihan DF, Innes AJ, Wells MJ, Wells J (1982) Oxygen consumption and blood gases. J Comp Physiol B 148:35-40
- Houston CS, Sutton JR, Cymerman A, Reeves JT (1987) Operation Everest II. Man at extreme attitude. J Appl Physiol 63:877–882
- Howell BJ (1969) Acid-base balance in the transition from water-breathing to air-breathing. In: Farhi LE, Rahn H (eds) Studies in pulmonary physiology, mechanics, chemistry, and circulation. USAF School of Aerospace Medicine, Aerospace Medical Division (AFSC), Brooks Airforce Base, Texas, pp 270–275
- Howell BJ (1970) Acid-base balance in transition from water to air breathing. Fed Proc 29:1130– 1134
- Howell BJ, Baumgardner FW, Bondi K, Rahn H (1970) Acid-base balance in cold-blooded vertebrates as a function of body temperature. Am J Physiol 218:600–606
- Howell DJ (1983) Optimization of behaviour: introduction and overview. Am Zool 23:257-260
- Howlett R (1966) Root cause of fish buoyancy. Nature (Lond) 380:203
- Huang W, Ten RT, McLaurie M, Bledsoe G (1996) Morphometry of the human pulmonary vasculature. J Appl Physiol 81:2123–2133
- Huber GL, Finley TN (1965) Effect of isotonic saline on the alveolar architecture. Anaesthesiology 26:252-253
- Huchzermeyer FW (1986) Causes and prevention of broiler ascites. SAPA Poult Bull 346
- Huchzermeyer FW, de Ruyk AMC (1986) Pulmonary hypertension syndrome associated with ascites in broilers. Vet Rec 119:94
- Huchzermeyer FW, de Ruyk AMC, van Ark H (1988) Broiler pulmonary hypertension syndrome. III. Commercial broiler strains differ in their susceptibility. Onderstepoort J Vet Res 55:5-9
- Huebner E, Chee G (1978) Histological and ultrastructural specialization of the digestive tract of the intestinal air breather *Hoplostemum thoracatum* (Teleost). J Morphoe 157:301-325
- Huelsenbeck JP, Rannala B (1997) Phylogenetic methods come of age: testing hypotheses in an evolutionary context. Science 276:227–232
- Huey RB (1987) Phylogeny, history and comparative method. In: Burggren WW, Huey RB (eds) New directions in ecological physiology. Cambridge University Press, Cambridge, pp 77–98
- Huey RB, Kingsolver JG (1993) Evolution of resistance to high temperature in ectotherms. Am Nat 142:S21–S46
- Hughes DM, Wimpenny JWT (1969) Oxygen metabolism by microorganisms. In: Rose AH, Wilkinson JF (eds) Advances in microbial physiology, vol 3. Academic Press, London, pp 197–231
- Hughes GM (1963) Comparative physiology of vertebrate respiration, 1st edn. Heineman, London
- Hughes GM (1965) Comparative physiology of vertebrate respiration, 2nd edn. Heineman, London
- Hughes GM (1966) Evolution between air and water. In: de Reuck AVS, Porter R (eds) Development of the lung. Churchill, London, pp 64–80
- Hughes GM (1967) Experiments on the respiration of the trigger fish (Balistes capriscus). Experientia 23:1077
- Hughes GM (1970) Ultrastructure of the air-breathing organs of some lower vertebrates. 7th Int Congr Electron Microsc, Grenoble 7:599–600

- Hughes GM (1972a) Distribution of oxygen tension in the blood and water along the secondary lamella of the icefish gill. J Exp Biol 56:481–492
- Hughes GM (1972b) Morphometrics of the fish gills. Respir Physiol 14:1-25
- Hughes GM (1973) Ultrastructure of the lung of *Neoceratodus* and *Lepidosiren* in relation to the lungs of other vertebrates. Folia Morphol (Pragne) 2:155–161
- Hughes GM (1976) Fish respiratory physiology. In: Spencer-Davies P (ed) Perspectives in environmental biology. Pergamon, Oxford, pp 235–245
- Hughes GM (1978) Some features of gas transfer in fish. Bull Inst Math Appl 14:39-43
- Hughes GM (1979) Scanning electron microscopy of the respiratory surfaces of trout gills. J Zool Lond 188:553–453
- Hughes GM (1980) Functional morphology of fish gills. In: Lahlou B (ed) Epithelial transport in lower vertebrates. Cambridge University Press, Cambridge, pp 15–36
- Hughes GM (1981) The effects of low oxygen and pollution on the respiratory systems of fish. In: Pickering AD (ed) Stress in fish. Academic Press, New York, pp 121–146
- Hughes GM (1982) An introduction to the study of gills. In: Houlihan DF, Rankin JC, Shuttleworth TJ (eds) Gills. Cambridge University Press, Cambridge, pp 1–24
- Hughes GM (1984) General anatomy of the gills. In: Randall DJ (ed) Fish physiology, vol XA. Academic Press, London, pp 1–72
- Hughes GM (1995) The gills of the coelacanth, *Latimeria chalumnae*, a study in relation to body size. Philos Trans R Soc Lond 347B:427–438
- Hughes GM, Al-Kadhomiy NK (1986) Gill morphometry of the mudskipper, *Boleophthalmus boddarti*. J Mar Biol Assoc UK 66:671–682
- Hughes GM, Iwai T (1978) A morphometric study of the gills in some Pacific deep-sea fishes. J Zool (Lond) 184:155–170
- Hughes GM, Kikuchi Y (1984) Effects of in vivo and in vitro changes in PO<sub>2</sub> on the deformability of erythrocytes of rainbow trout (*Salmo gairdnen*). J Exp Biol 111:253–257
- Hughes GM, Mondolfino RM (1983) Scanning electron microscopy of the gills of *Trachurus* mediterraneus. Experientia 39:518–519
- Hughes GM, Morgan M (1973) The structure of fish gills in relation to their respiratory function. Biol Rev 48:419–475
- Hughes GM, Munshi JSD (1968) Fine structure of the respiratory surfaces of an air-breathing fish, the climbing perch, *Anabas testudineus*. Nature (Lond) 219:1382–1384
- Hughes GM, Munshi JSD (1979) Fine structure of the gills of some Indian air-breathing fishes. J Morphol 160:169–194
- Hughes GM, Munshi JSD (1986) Scanning electron microscopy of the accessory respiratory organs the snake-head fish, *Channa strata* (Bloch) (Channidae, Channiformes). J Zool (Lond) 209:305–317
- Hughes GM, Pohunkova H (1980) Scanning and transmission electron microscopy of the lungs of *Polypterus senegalensis*. Folia Morphol 110:112–123
- Hughes GM, Shelton G (1958) The mechanism of gill ventilation in three fresh water teleosts. J Exp Biol 35:807–823
- Hughes GM, Singh BN (1970a) Respiration in air-breathing fish, the climbing perch, Anabas testudineus II. Respiratory patterns and the control of breathing. J Exp Biol 53:281–298
- Hughes GM, Singh BN (1970b) Respiration in air-breathing fish, the climbing perch, *Anabas testudineus* Bloch. I. Oxygen uptake and carbon dioxide release into air and water. J Exp Biol 53:265–280
- Hughes GM, Singh BN (1971) Gas exchange with air and water in an air-breathing catfish, Saccobranchus (Heteropneustes) fossilis. J Exp Biol 55:667–682
- Hughes GM, Umezawa SI (1983) Gill structure of the yellowtail and frogfish. Jpn J Ichthyol 30:176–183
- Hughes GM, Vergara GA (1978) Static pressure-volume curves for the lung of the frog (*Rana pipiens*). J Exp Biol 76:140–165
- Hughes GM, Weibel ER (1976) Morphometry of fish lungs. In: Hughes GM (ed) Respiration of amphibious vertebrates. Academic Press, London, pp 213–232
- Hughes GM, Wright DE (1970) A comparative study of the ultrastructure of the water-blood pathways in the secondary lamellae of teleost and elasmobranch fishes benthic forms. Z Zellforsch 104:478-493

- Hughes GM, Knight B, Scammel CA (1969a) The distribution of PO<sub>2</sub> and hydrostatic pressure changes within the branchial chambers in relation to gill ventilation in the shore crab *Carcinus maenas* L. J Exp Biol 51:203–220
- Hughes J, Gillis CN, Bloom FE (1969b) The uptake and disposition of dl-norepinephrine in perfused rat lung. J Pharmacol Exp Ther 169:237-248
- Hughes GM, Dube SC, Munshi JSD (1973) Surface area of the respiratory organs of the climbing perch, *Anabas testudineus* (Pisces: Anabantidae). J Zool (Lond) 170:227-243
- Hughes GM, Singh BR, Guha G, Dube SC, Munshi JSD (1974) Respiratory surface areas of an airbreathing siluroid fish *Saccobranchus (Heteropneustes) fossilis* in relation to body size. J Zool (Lond) 172:215–232
- Hughes GM, Horimoto M, Kikuchi Y, Kakiuchi Y, Koyama T (1981) Blood flow velocity in microvessels of the gill filaments of goldfish (*Carassius auratus L.*). J Exp Biol 90:327– 331
- Hughes GM, Kikuchi Y, Watari H (1982) A study of the deformability of erythrocytes of a teleost fish, the yellow tail (*Seriola quinqueradiata*) and a comparison with human erythrocytes. J Exp Biol 96:209–220
- Hughes GM, Munshi JSD, Ojha J (1986a) Post-embryonic development of water- and airbreathing organs of Anabas testudineus (Bloch). J Fish Biol 29:443-450
- Hughes GM, Perry SF, Piiper J (1986b) Morphometry of the gills of the elasmobranch *Scyliohinus stellaris* in relation to body size. J Exp Biol 121:27-42
- Hughes GM, Roy PK, Munshi JSD (1992) Morphometric estimation of oxygen-diffusing capacity for the air sac in the catfish *Heteropneustes fossilis*. J Zool (Lond) 227:193–209
- Hulbert AJ, Else PL (1983) Oxygen demand of organ systems. Proc Physiol Soc NZ 3:95-101
- Humburger V (1980) Embryology and the modern synthesis in evolutionary theory. In: Mayr E, Provone WB (eds) The evolutionary synthesis: perspectives on the unification of biology. Harvard University Press, Cambridge, pp 97–111
- Humpreys PW, Normand ICS, Reynolds EOR, Strang LB (1967) Pulmonary lymph flow and the uptake of liquid from the lungs of the lamb at the start of breathing. J Physiol (Lond) 193:1–29
- Hunt BG (1979) The effects of past variations of the Earth's rotation rate on climate. Nature (Lond) 281:188–191
- Hunt JR, Aitken JR (1962) The effect of ammonium and chloride ions in the diet of hens on egg shell quality. Poult Sci 41:343–357
- Hunt JR, Simkiss K (1967) Acute respiratory acidosis in the domestic fowl. Comp Biochem Physiol 21:223-237
- Hunten DM (1973) The escape of light gases from planetary atmospheres. J Atmos Sci 30:1481– 1494
- Hunten DM, Donahue TM (1976) Hydrogen loss from the terrestrial planets. Annu Rev Earth Planet Sci 4:265–292
- Hunten DM, Strobel DF (1974) Production and escape of terrestrial hydrogen. J Atmos Sci 31:305-317
- Hunter WR (1953) The condition of mantle cavity in two pulmonate snails living in Loch Lomond. Proc R Soc Edinb 65B:143-165
- Hurst LD, McVean G (1996) Evolutionary genetics and scandalous symbionts. Nature (Lond) 381:650–651
- Hutchison AA, McNicol KJ, Loughlin GM (1985) The effect of age on pulmonary epithelial permeability in unanaethetized lambs. Pediatr Pulmonol 1:53-58
- Hutchison GE (1959) Homage to Santa Rosalia, or why are there so many kinds of animals? Am Nat 93:145–159
- Hutchison GE (1975) A treatise on limnology, vol I: geography, physics and chemistry. John Wiley, London
- Hutchison VH (1968) Relation of body size and surface area to gas exchange in anurans. Physiol Zool 41:65–85
- Hutchison VH, Haines HB, Engbretson G (1976) Aquatic life at high altitude: respiratory adaptations in the Lake Titicaca frog, *Telmatobius culeus*. Respir Physiol 27:115-129
- Huxley VH, Kutchai H (1983) Effect of diffusion boundary layers on the initial uptake of O<sub>2</sub> by red cells. Theory versus experiment. Microvasc Res 26:89–107

- Hyde DM, Robinson NE, Gillespie JR, Tyler WS (1977) Morphometry of the distal air spaces in lungs of aging dogs. J Appl Physiol 43:86–91
- Hyden P, Lindberg R (1970) Hypoxia induced torpor in pocket mice (genus *Perognathus*). Comp Biochem Physiol 33A:167–179
- Hyman LH (1951) The invertebrates: Platyhelminthes and Rynchocoela. The Acoelomate Bilateria, vol II. McGraw-Hill, New York
- Hyman LH (1955) The invertebrates, vol IV. Echinodermata, water vascular system. McGraw-Hill, London
- Hyman LH (1967) The invertebrates VI: Mollusca I. McGraw-Hill, New York
- Igic R, Erdös EG, Yeh HSJ, Sorrells K, Nakajima Y (1972) Angiotensin I converting enzyme of the lung. Circ Res 30 & 31, Suppl II:51–61
- Imbrie J, McIntyre A, Mix A (1989) Oceanic response to orbital forcing in the late Quaternary: observational and experimental strategies. In: Berger A (ed) Climate and geosciences. Yale University Press, New Haven, pp 121–164
- Infantino RL, Burggren WW, Twonsend DS (1988) Physiology of direct development in the Puerto Rican frog *Eleutherodactylus coqui*. Am Zool 28:23A
- Inger RF (1957) Ecological aspects of the origin of the tetrapods. Evolution 11:373-376
- Ingermann RL, Terwilliger RC (1981) Oxygen affinities of foetal and maternal haemoglobins of the viviparous seaperch, *Embiotoca lateralis*. J Comp Physiol 142:523-531
- Innes AJ (1985) Aerobic scope for activity of burrowing mantis shrimp, *Heterosquilla tricarinata*, at low environmental oxygen tensions. Comp Biochem Physiol 81:827-832
- Innes AJ, El Haj AJ, Gobin JF (1986) Scaling of the respiratory, cardiovascular and skeletal muscle systems of the fresh water/terrestrial mountain crab, *Pseudothelphusa garhami* garhami. J Zool (Lond) 209:595–606
- Innes AJ, Mardsen ID, Wong PPS (1984) Bimodal respiration of intertidal pulmonates. Comp Biochem Physiol 77A:441–445
- Innes AJ, Taylor EW (1986a) The evolution of air breathing in crustaceans: a functional analysis of branchial, cutaneous and pulmonary gas exchange. Comp Biochem Physiol 85A:621–637
- Innes AJ, Taylor EW (1986b) Air breathing crabs of Trinidad: adaptive radiation into the terrestrial environment. I. Aerobic metabolism and habitat. Comp Biochem Physiol 85A:373-382
- Ishii Y, Hasegawas S, Uchiyama Y (1989) Twenty four-hour variations in subcellular structures of the rat type II alveolar epithelial cell: a morphometric study at the electron microscopic level. Cell Tissue Res 256:347–353
- Ishmatsu A, Itazawa Y (1981) Ventilation of the air-breathing organ in the snakehead *Channa* argus. Jpn J Ichthyol 28:276–282
- Ishmatsu A, Itazawa Y (1983) Difference in blood oxygen levels in the outflow vessels of the heart of an air-breathing fish, *Channa argus*. Do separate blood streams exist in a teleost heart? J Comp Physiol B 149:435-440
- Ishmatsu A, Itazawa Y, Takeda T (1979) On the circulatory systems of the snakeheads *Channa* maculata and *C. argus* with reference to bimodal breathing. Jpn J Ichthyol 26:167–180
- Isozaki Y (1997) Permo-Triassic boundary superanoxia and stratified superocean: records from lost deep sea. Science 276:235–238
- Itazawa Y, Oikawa S (1983) Metabolic rates in excised tissues of the carp. Experientia 39:160-161
- Ito T (1953) The permeability of the integument to oxygen and carbon dioxide in vivo. Biol Bull 105:308–315
- Iwai T, Nakamura I (1964) Branchial skeleton of the bluefin tuna, with special reference to the gill rays. Bull Misaki Mar Biol Inst Kyoto Univ 6:21–25
- Iwasawa Y, Gillis CN (1974) Pharmacological analysis of norepinephrine and 5hydroxytryptamine removal from the pulmonary circulation: differentiation of uptake sites for each amine. J Pharmacol Exp Ther 188:386–393
- Iwasawa Y, Gillis CN, Aghajanian G (1973) Hypothermic inhibition of 5-hydroxytryptamine and norepinephrine uptake by lung: cellular location of amines after uptake. J Pharmacol Exp Ther 186:498–507
- Jablonski D (1997) Body size evolution in Cretaceous molluscs and the status of Cope's rule. Nature (Lond) 385:250–252

- Jackson DC (1978) Respiratory control in air-breathing ectotherms. In: Davies DG, Barnes CD (eds) Regulation of ventilation and gas exchange. Academic Press, London, pp 93–130
- Jackson DC (1986) Acid-base regulation of reptiles. In: Heisler N (ed) Acid-base regulation in animals. Elsevier, Amsterdam, pp 235–263
- Jackson DC (1987) How do amphibians breathe both water and air. In: Dejours P, Bolis L, Taylor CR, Weibel ER (eds) Comparative physiology: life in water and on land. Springer, Berlin Heidelbery New York, pp 49–58
- Jackson DC, Schmidt-Nielsen K (1964) Counter-current heat exchange in the respiratory gases. Proc Natl Acad Sci USA 51:1192–1197
- Jackson DC, Allen J, Strupp PK (1976) The contribution of the nonrespiratory surfaces to CO<sub>2</sub> loss in six species of turtle at 20 °C. Comp Biochem Physiol 55A:243–246
- Jackson MR, Joy CF, Mayhew TM, Haas JD (1985) Stereological studies on the true thickness of the villous membrane in human term placentae: a study of placentae from high altitude pregnancies. Placenta 6:249–258
- Jackson MR, Mayhew TM, Haas JD (1988a) On the factors which contribute to the thinning of the villous membrane in human placentae at high altitude. I. Thinning and regional variation in thickness of trophoblast. Placenta 9:1–8
- Jackson MR, Mayhew TM, Haas JD (1988b) On the factors which contribute to thinning of the villous membrane in human placentae at high altitude. II. An increase in the degree of peripherization of foetal capillaries. Placenta 9:9–18
- Jackson MR, Mayhew TM, Boyd PA (1992) Quantitative description of the elaboration and maturation of villi from ten weeks of gestation to term. Placenta 13:357–370
- Jackson MR, Walsh AJ, Morrow RJ, Brendan J, Mullen M et al. (1995) Reduced placental villous tree elaboration in small-for-gestational-age pregnancies: relationship with umbilical artery Doppler waveforms. Am J Obstet Gynecol 172:518–525
- Jacobs W (1938) Untersuchungen zur Physiologie der Schwimmblase der Fische. IV. Die erste Gasfüllung der Schwimmblase bei jungen Seepferdchen. Z Vergl Physiol 25:379–388
- Jacobsen VH (1967) The feeding of the lugworm (*Arenicola marina*) (L.). Quntitative studies. Ophelia 4:91-109
- Jahnke L, Klein HP (1979) Oxygen as a factor in eukaryocyte evolution: some effects of low levels of oxygen on *Saccharomyces cerevisiae*. Origins Life 9:329–334
- Jakubowski M, Byczkowska-Smyk W, Mikhalev Y (1969) Vascularization and size of the respiratory surfaces in the antarctic white-blooded fish *Chaenichthys rugosus* Regan (Percoidei, Chaenichthydea). Zool Pol 19:303–317
- Jalavisto E, Kuorinka J, Kyllästinen M (1965) Responsiveness of the erythron to variations of oxygen tension in the chick embryo and young chicken. Acta Physiol Scand 53:479-486
- James AG, Probyn T (1989) The relationship between respiration rate, swimming speed and feeding behaviour in the Cape anchovy *Engraulis capensis* Gilchrist. J Exp Mar Biol Ecol 131:81–100
- Jameson W (1958) The wandering albatross. Hart-Davis, London
- Janis C (1993) Victory by default: the mammalian succession. In: Gould SJ (ed) The book of life. Ebury-Hutchison, London, pp 169–217
- Janke A, Feldmaier-Fuchs G, Thomas WK, Haeseler AV, Paabo S (1994) The marsupial mitochondrial genome and the evolution of placental mammals. Genetics 137:243–256
- Jansen RG, Randall DJ (1975) The effects of changes in pH and PCO<sub>2</sub> in blood and water on breathing in rainbow trout, *Salmo gairdneri*. Respir Physiol 25:235–245
- Jarman C (1970) Evolution of life. Bantam Books, Toronto
- Jarvik E (1968) Aspects of vertebrate phylogeny. In: Orvig T (ed) Current problems of lower vertebrate phylogeny. Interscience, New York, pp 497–527
- Jarvik E (1980) Basic structure and evolution of vertebrates, vol 2. Academic Press, London
- Jarvis JUM, Bennet NC (1990) The evolutionary history, population biology and social structure of African mole rats: family Bathyergidae. In: Nevo E, Reig OA (eds) Evolution of subterranean mammals at the organismal and molecular levels. Wiley-Liss, New York, pp 97–128
- Jasinski A (1973) Air-blood barrier in the respiratory intestine of the pond roach, *Misgumus fossilis* L. Acta Anat 86:376-391
- Jasper JP, Hayes JM (1990) A carbon isotope record of  $CO_2$  levels during the late Quartenary. Nature (Lond) 347:462–464

Jell PA (1978) Trilobite respiration and genal caeca. Alcheringa 2:251-260

Jenkins RJF (1991) The early environment. In: Bryant CH (ed) Metazoan life without oxygen. Chapman and Hall, London, pp 38-64

Jenkyns HC (1980) Cretaceous anoxic events: from continents to oceans. J Geol Soc (Lond) 137:171-188

Jensen D (1966) The hagfish. Sci Am 214:82-90

- Jensen FB (1991) Multiple strategies in oxygen and carbon dioxide transport by haemoglobin. In: Woakes AJ, Grieshaber MK, Bridges CR (eds) Physiological strategies for gas exchange and metabolism. Cambridge University Press, Cambridge, pp 55–78
- Jensen FB, Weber RE (1985) Kinetics of the acclimational responses of tench to combined hypoxia and hypercapnia. I. Respiratory responses. J Comp Physiol B 156:197–203
- Jepsen GL (1970) Bat origins and evolution. In: Wimsatt WA (ed) The biology of bats, vol I. Academic Press, New York, pp 1-64
- Jervis P (1995) Water: vital resource in home for agriculture and for industry. Franklin Watts, London
- Jesse MJ, Shub C, Fishman AP (1967) Lung and gill ventilation of the African lungfish. Respir Physiol 3:267–287
- Jessop NM (1995) General zoology, 6th edn. McGraw-Hill, New York
- Jeuken M (1957) A study of the respiration of *Misgurnus fossilis* (L.), the pond loach. Thesis, University of Leiden, Leiden
- Jia L, Bonaventura C, Bonaventura J, Stamler J (1996) S-Nitrosohaemoglobin: a dynamic activity of blood involved in vascular control. Nature (Lond) 380:221–226
- Joenje H (1989) Genetic toxicology of oxygen. Mutat Res 219:193–208
- Johansen K (1960) Circulation in the hagfish, Myxine glutinosa L. Biol Bull 118:289–295
- Johansen K (1966) Air-breathing in the teleost Synbranchus marmoratus. Comp Biochem Physiol 18:383-395
- Johansen K (1968) Air-breathing fishes. Sci Am 219:102-111
- Johansen K (1970) Air-breathing in fish. In: Hoar WS, Randall DJ (eds) Fish physiology, vol 4. Academic Press, London, pp 361–411
- Johansen K (1971) Comparative physiology: gas exchange and circulation in fishes. Annu Rev Physiol 33:569–612
- Johansen K (1972) Heart and circulation in gill, skin and lung breathing. Respir Physiol 14:193–210
- Johansen K (1979) Cardiovascular support of metabolic functions invertebrates. In: Wood SC, Lenfant C (eds) Evolution of respiratory processes: a comparative approach. Marcel Dekker, New York, pp 107–192
- Johansen K (1982) Respiratory gas exchange of vertebrate gills. In: Houlihan DF, Rankin JC, Shuttleworth TJ (eds) Gills. Cambridge University Press, Cambridge, pp 99-128
- Johansen K (1987) The world as a laboratory: physiological insights from Nature's experiments. In: McLennan H, Ledsome JR, McIntosh CHS, Jones DR (ed) Advances in physiological research. Plenum Press, New York, pp 377–396
- Johansen K, Burggren WW (1980) Cardiovascular function in lower vertebrates. In: Bourne GH (ed) Hearts and heart-like organs. Academic Press, New York, pp 61–117
- Johansen K, Hanson D (1968) Functional anatomy of the hearts of lungfishes and amphibians. Am Zool 8:191-210
- Johansen K, Hol R (1968) A radiological study of the central circulation in the lungfish *Protopterus aethiopicus*. J Morphoe 126:333-348
- Johansen K, Lenfant C (1966) Gas exchange in the Cephalopod, *Octopus dofleini*. Am J Physiol 210:910–918
- Johansen K, Lenfant C (1967) Respiratory function in the South American lungfish. J Exp Biol 46:205–218
- Johansen K, Lenfant C (1968) Respiration in the African lungfish, *Protopterus aethiopicus*. II. Control of breathing. J Exp Biol 49:453-468
- Johansen K, Lenfant C (1972) A comparative approach to the adaptability of O<sub>2</sub>-Hb affinity. In: Roth M, Astrup P (eds) Oxygen affinity of haemoglobin and red cell acid-base status. Munksgaard, Copenhagen, pp 750–780

Johansen K, Martin AW (1966) Circulation in the giant earthworm, *Glossoscolex giganteus*. I. Contractile processes and pressure gradients in the large blood vessels. J Exp Biol 43:333–347

- Johansen K, Reite OB (1968) Effects of acetlycholine and biogenic amines on pulmonary smooth muscle in the African lungfish *Protopterus aethiopicus*. Acta Physiol Scand 71:465–471
- Johansen K, Lenfant C, Grigg GF (1967) Respiratory control in the lungfish, Neoceratodus forsteri (Krefft). Comp Biochem Physiol 20:835-854
- Johansen K, Lenfant C, Hanson D (1968a) Cardiovascular dynamics in the lungfish. Z Vergl Physiol 59:157–186
- Johansen K, Lenfant C, Schmidt-Nielsen K, Petersen JA (1968b) Gas exchange and control of breathing in the electric eel *Electrophorus electricus*. Z Vergl Physiol 61:137–163
- Johansen K, Lenfant C, Hanson D (1970a) Respiration in a primitive air-breather, Amia calva. Respir Physiol 9:162–174
- Johansen K, Lenfant C, Hanson D (1970b) Phylogenetic development of pulmonary circulation. Fed Proc 29:1135–1140
- Johansen K, Maloiy GMO, Lykkeboe G (1975) A fish in extreme alkalinity. Respir Physiol 24:159-162
- Johansen K, Lomholt JP, Maloiy GMO (1976) Importance of air and water breathing in relation to size of the African lungfish *Protopterus amphibius*, Peters. J Exp Biol 65:395–399
- Johansen K, Burggren WW, Glass M (1977) Pulmonary stretch receptors regulate heart rate and pulmonary blood flow in the turtle, *Pseudemys scripta*. Comp Biochem Physiol 58:185–191
- Johansen K, Mangum CP, Lykkeboe G (1978) Respiratory properties of the blood of Amazon fishes. Can J Zool 56:898-906
- Johansen K, Lykkeboe G, Kornerup S, Maloiy GMO (1980) Temperature-insensitive oxygen binding in blood of the tree frog *Chiromantis petersi*. J Comp Physiol 136:71-76
- Johansen K, Berger M, Bicudo JEPW, Ruschi A, De Almeida PJ (1987) Respiratory properties of blood and myoglobin in hummingbirds. Physiol Zool 60:269–278
- Johnson LF (1964) The effects of decreased barometric pressure on maximum pressure-volume relationships of the human respiratory system. J Aerosp Med 35:637–642
- Johnson L, Rees CJ (1988) Oxygen consumption all gill surface area in relation to habitat and lifestyle of four crab species. Comp Biochem Physiol 89A:243-246
- Johnson ML (1942) The respiratory function of the haemoglobin of the earthworm. J Exp Biol 18:266–277
- Johnson RE, Liu M (1996) The loss of atmosphere from mass. Science 274:1932
- Johnson SL (1988) The effects of the 1983 El Niño on Oregon's coho (*Oncorhynchus kitutch*) and chinook (*O. tshawytscha*) salmon. Fish Res 6:105–123
- Johnston AI, Harrison P (1985) Contractile and metabolic characteristics of muscle fibers from Antarctic fish. J Exp Biol 116:223-237
- Johnston AI, Fitch N, Zummo G, Wood RE, Harrison P, Tota B (1983) Morphometric and ultrastructural features of the ventricular mrocardium of the haemoglobin-less icefish *Chaenocephalus aceratus*. Comp Biochem Physiol 76:475-480
- Johnston JD, Mcforlane RW (1967) Migration and bioenergeties of flight in the Pacific golden prover. Condor 69:156–168
- Johnston KS, Beehler CL, Sakamoto-Arnold CM, Childress JJ (1986) In situ measurements of chemical distributions in a deep-sea hydrothermal vent field. Science 231:1139-1141
- Johnston KS, Childress JJ, Beehler CL (1988) Short term temperature variability in the Rose Garden hydrothermal vent field: an unstable deep-sea environment. Deep Sea Res 35:1711– 1721
- Jokumsen A, Fyhn HJ (1982) The influence of aerial exposure upon respiratory and osmotic properties of haemolymph from intertidal mussels, *Mytilus edulis* L and *Modiolus modiolus*. J Exp Mar Biol Ecol 61:189-203
- Jollie WP, Jollie LG (1967) Electron microscopic observations on accommodations to pregnancy in the uterus of the spiny dogfish, *Squalus acanthias*. J Ultrastruct Res 20:161–178
- Jones C, Fox H (1977) Syncytial knots and intervillous bridges in the human placenta: an ultrastructural study. J Anat 124:275–286
- Jones D (1996) Hydrogen for ever. Nature (Lond) 383:766
- Jones D, Birks S (1992) Megapodes: recent ideas on origins, adaptations and reproduction. Trends Ecol Evol 7:88-91

Jones DP (1986) Intracellular diffusion gradients of O<sub>2</sub> and ATP. Am J Physiol 250:C663-675

- Jones DR, Johansen K (1972) The blood vascular system of birds. In: Farner DS, King JR (eds) Avian biology, vol II. Academic Press, New York, pp 157–285
- Jones DR, Randall DJ (1978) The respiratory and circulatory systems during exercise. In: Hoar WS, Randall DJ (eds) Fish physiology, vol 7. Academic Press, New York, pp 425-501
- Jones DR, Schwarzfeld T (1974) The oxygen cost to the metabolism and efficiency of breathing in trout (*Salmo gairdnen*). Respir Physiol 21:1992–1999
- Jones DR, Shelton G (1972) Factors affecting diastolic blood pressures in the systemic and pulmocutaneous arches of anuran amphibia. J Exp Biol 57:789-803
- Jones G, Rayner JMV (1989) Optimal flight speed pipistrelle bats (*Pipistrellus pipistrellus*). In: Hanak V, Horaceck I, Gaisler J (eds) European bat research 1987: Proc 4th Europ Bat Res Symp. Charles University Press, Praha, pp 87–103
- Jones HD (1983) Circulatory systems of gastropods and bivalves. In: Saleuddin ASM, Wilbur MW (eds) The Mollusca, vol 5, physiology, part 2. Academic Press, London, pp 189-238
- Jones JD (1961) Aspects of respiration in *Planorbis corneus* L. and *Lymnaea stagnalis* L. (Gastropoda: Pulmonata). Comp Biochem Physiol 4:1-29
- Jones JD (1964) Respiratory gas exchange in the aquatic pulmonate *Biompalaria sudanica*. Comp Biochem Physiol 12:297-310
- Jones JD (1972) Comparative physiology of respiration. Edward Amold, London
- Jones JG, Roystom D, Minty BD (1983) Changes in alveolar-capillary barrier function in animals and humans. Am Rev Respir Dis 127:S51–S59
- Jones JH (1982) Pulmonary blood flow in distribution in panting ostriches. J Appl Physiol 53:1411-1417
- Jones JH, Effmann EL, Schmidt-Nielsen K (1981) Control of air flow in bird lungs: radiographic studies. Respir Physiol 45:121–131
- Jones JH, Grubb B, Schmidt-Nielsen K (1983) Panting in the emu causes arterial hypoxemia. Respir Physiol 54:189–195
- Jones JH, Effmann EL, Schmidt-Nielsen K (1985) Lung volume changes in during respiration in ducks. Respir Physiol 59:15–25
- Jones JH, Longworth KE, Lindholm A, Conley KE, Karas RH et al. (1989) Oxygen transport during exercise in large mammals. I. Adaptive variation in oxygen demand. J Appl Physiol 67:862–870
- Jones JRE (1952) The reaction of fish to water of low oxygen concentration. J Exp Biol 29:403–415
- Jones RE, Smith HM, Bock CE (1993) Reptilian and avian ovarian cycles and evolutionary origin of volant birds. J Comp Physiol 163:594–601
- Jorgensen CB (1952) On the relationship between water transport and food requirements in some marine filter feeding invertebrates. Biol Bull Mar Biol Lab Woods Hole 103:356–363
- Jorgensen CB (1955) Quantitative aspects of filter feeding in invertebrates. Biol Rev 30:391-453
- Jorgensen CB (1975) Comparative physiology of suspension feeding. Annu Rev Physiol 37:57-70
- Joseph JH (1967) Diurnal and solar variations of neutral hydrogen in the atmosphere. Ann Geophys 23:365–373
- Joss JM, Cramp N, Baverstock PR, Johnson AM (1991) A phylogenetic comparison of 18S ribosomal RNA sequences of lungfish with those of other chordates. Aust J Zool 39:509-518
- Jouin C, Toulmond A (1989) The ultrastructure of the gill of the lugworm, Arenicola marina. Acta zool 70:121-129
- Jouve-Duhamel A, Truchot JP (1983) Ventilation in the shore crab *Carcinus maenas* (L) as a function of ambient oxygen and carbon dioxide field and laboratory studies. J Exp Mar Biol Ecol 70:281–296
- Joyce GF (1992) Directed molecular evolution. Sci Am 267:48-55
- Joyce GF (1997) Evolutionary chemistry: getting from there from here. Science 276:1658–1657
- Jukes TH (1985) A change in the genetic code in *Mycoplasma capricolum*. J Mol Evol 22:361–362
- Jukes TH, Osawa S (1993) Evolutionary changes in the genetic code. Comp Biochem Physiol 106B:489-494
- Julian D, Arp AJ (1992) Sulfide perrmeability in the marine invertebrates *Urechis caupo*. J Comp Physiol 162B:59–67

- Julian D, Menon JG, Arp AJ (1991) Structural and functional adaptations to hypoxia and sulfide in *Urechis caupo*. Am Zool 31:73A
- Julian RJ (1987) The effects of increased sodium in the drinking water on right ventricular failure and ascites in broiler chickens. Avian Pathol 16:61–71
- Julian RJ, Wilson JB (1986) Right ventricular failure as a cause of ascites in broiler and roaster chickens. Proc IVth Int Symp Vet Lab Diag 1986:608–611
- Julian RJ, Moran ET, Revinton W, Hunter DB (1984) Acute hypertensive angiopathy as a cause of sudden death in turkeys. Proc Am Vet Med Assoc 117
- Junod AF (1972) Uptake, metabolism and efflux of <sup>14</sup>C-5-hydroxytryptamine in isolated, perfused rat lungs. J Pharmacol Exp Ther 183:431-355
- Junod AF (1975) Mechanism of uptake of biogenic amines in the pulmonary circulation. In: Junod AF, Haller R (eds) Lung metabolism. Academic Press, New York, pp 387-396
- Junqueira IC, Steen JB, Tincoce RM (1967) The respiratory area of the fishes of teleosts from Rio Negro and Rio Branco area. Research Papers from the Alpha Helix Amazon Expedition, pp B20–21
- Jürgens JD, Bartels H, Bartels R (1981) Blood oxygen transport and organ weight of small bats and small nonflying mammals. Respir Physiol 45:243-260
- Just JJ, Gatz RN, Crawford EC (1973) Changes in respiratory functions during metamorphosis of the bullfrog, *Rana catesbeiana*. Respir Physiol 17:276–282
- Kaestner A (1929) Bau und Funktion der Fächertracheen einiger Spinnen. Z Morphol Ökol Tiere 13:463–558
- Kaestner A (1970) Invertebrate zoology, vol 3 (transl by Levi HW and Levi LR). Wiley, New York
- Kagan VE, Day BW, Elsayed NM, Gorbunov NV (1996) Dynamics of haemoglobin. Nature (Lond) 383:30-31
- Kaiser T, Bucher TL (1985) The consequences of reverse sexual size dimorphism for oxygen consumption, ventilation and water loss in relation to ambient temperature in the prairie falcon, *Falco mexicanus*. Physiol Zool 58:748–758
- Kamau JMZ, Maina JN, Maloiy GMO (1984) The design and the role of the nasal passages in temperature regulation in the dik-dik antelopes, *Rhynchotragus kirkii* with observations on the carotid rete. Respir Physiol 56:183–194
- Kammer AE, Heinrich B (1978) Insect flight muscle metabolism. Adv Insect Physiol 13:133-228
- Kampe G, Crawford EC (1973) Oscillatory mechanics of the respiratory system of pigeons. Respir Physiol 18:188–193
- Kämpfe L (1980) Evolution und Stammesgeschichte der Organismen. Fischer, Stuttgart
- Kandel ER (1979) Behavioural biology of Aplysia. WH Freeman, San Francisco
- Kanwisher JW (1966) Tracheal gas dynamics in pupae of the *Cecropia* silkworm. Biol Bull 130:96–105
- Kanwisher JW, Ebeling A (1957) Composition of the swim bladder gas in bathypelagic fishes. Deep Sea Res 2:211-223
- Kanz JE, Quast WD (1990) Respiratory pumping seizure: a newly discovered spontaneous stereotyped behaviour pattern in the opisthobranch mollusc Aplysia californica. J Comp Physiol 166A:619-627
- Kanz JE, Quast WD (1992) Respiratory pumping behaviour in the marine snail Aplysia califomica as a function of ambient hypoxia. Physiol Zool 65:35-54
- Karas RH, Taylor CR, Jones JH, Linstedt SL, Reeves RB, Weibel ER (1987a) Adaptive variation in the mammalian respiratory system in relation to energetic demand. VII. Flow of oxygen across the pulmonary gas exchanger. Respir Physiol 69:101–115
- Karas RH, Taylor CR, Rösler K, Hoppeler H (1987b) Adaptive variation in the mammalian respiratory system in relation to energetic demand: V. Limits to oxygen transport by circulation. Respir Physiol 69:65–79
- Karasov WH, Diamond JH (1985) Digestive adaptations for fuelling the cost of endothermy. Science 228:202-204
- Karasov WH, Phan D, Diamond JH, Carpenter FL (1986) Food passage and intestinal nutrient absorption in hummingbirds. Auk 103:453–464
- Kardong KV (1972) Morphology of the respiratory system and its musculature in different snake genera (part I), *Crotalus* and *Elaphe*. Gegenbaurs Morphol Jahrb 117:285-302

Kardong KV (1995) Vertebrate: comparative anatomy, function and evolution. Wm C Brown, Dubuque, lowa

Kargel JS, Strom RG (1996) Global climatic change on Mars. Sci Am 275:60-68

Karlberg P, Adams FH, Geubelle F, Wallgren G (1962) Alterations of the infant's thorax during vaginal delivery. Acta Obstet Gynecol Scand 41:223–229

- Karlsson L (1983) Gill morphology in the zebra fish, *Brachydanio rerio* (Hamilton-Buchanan). J Fish Biol 23:511–524
- Karsdorp VHM, van Vugt JMG, van Geijn HP, Kostense PJ, Arduini D et al. (1994) Clinical significance of absent or reversed end diastolic velocity waveforms in umbilical artery. Lancet 344:1664–1668
- Karsdorp VHM, Dirks BK, van der Linden JC, van Vugt JMG, Baak JPA, van Geijn HP (1996) Placenta morphology and absent or reversed end diastolic flow velocities in the umbilical artery: a clinical and morphometrical study. Placenta 17:393–399
- Kaschke M, Russell MJ (1994) [FeS/FeS<sub>2</sub>]: a redox system for the origin of life. Origins Life Evol Biosphere 24:43–56
- Kasting JF (1997) Warming early Earth and Mars. Science 276:1213-1215
- Kasting JF, Walker JCG (1981) Limits on oxygen concentration in prebiological atmosphere and the rate of abiotic fixation of nitrogen. J Geophys Res 86:1147–1158
- Kasting JF, Liu SC, Donahue TM (1979) Oxygen levels in the prebiological atmosphere. J Geophys Res 84:3097-3107
- Katušic ZS, Cosentino F (1994) Nitric oxide synthetase: from molecular biology to cerebrovascular physiology. News Physiol Sci 9:64–66
- Kaufmann P (1972) Untersuchungen uber die Langhanszellen in der menschlichen Placenta. Z Zellforsch 128:283–302

Kaufmann P, Davidoff M (1977) The guinea-pig placenta. Adv Anat Embryol Cell Biol 53:1-90

- Kaufmann P, Gentzen DM, Davidoff M (1977) Die Utrastruktur von Langhanszellen in pathologischen menschlichen Placenten. Arch Gynecol 22:319–332
- Kawamoto N (1927) The anatomy of *Caudina chilensis* (J Müller) with special reference to the perivisceral cavity, the blood and the water vascular systems in their relation to the blood circulation. Sci Rep Tôhoku Univ IV Biol 2:239–264
- Kawasaki M (1993) Independently evolved jamming avoidance responses employ identical computational algorithms: a behavioural study of the African electric fish, *Gymnarchus niloticus*. J Comp Physiol A 173:9–22
- Kawashiro T, Scheid P (1975) Arterial blood gases in undisturbed gases in undisturbed resting brids: measurements in chicken and duck. Respir Physiol 23:337–342
- Kayar SR, Snyder GK, Birchard GF, Black CP (1981) Oxygen permeability of the shell and membranes of chicken eggs during development. Respir Physiol 11:16-34
- Keeling CD, Chin JFS, Whorf TP (1996) Increased activity of northern vegetation inferred from atmospheric CO<sub>2</sub> measurements. Nature (Lond) 382:146–149
- Keevil T, Mason HS (1978) Molecular oxygen in biological oxidations an overview. In: Fleischer S, Paker L (eds) Biomembranes: methods in enzymology, vol 3. Academic Press, New York, pp 3–40
- Keilin D (1924) On appearance of gas in the trachea of insects. Proc Camb Philos Soc Biol Sci 1:63–70
- Kendal MW, Dale JE (1979) Scanning and transmission electron microscopic observations of rainbow trout (*Salmo gairdnen*) gill. J Fish Res Board Can 36:1072-1079
- Kennedy B (1979) Blood circulation in polychaete gills. Am Zool 19:868
- Kennerly TE (1964) Microenvironmental conditions of the pocket gopher burrow. Tex J Sci 16:395-441
- Kerourio P (1981) Nouvelles observations sur la mode de nidification et de ponte chez les dinosauriens du Cretace terminal du Midi de la France. C R Somm Séances Soc Geol Fr 1:25–28
- Kerr RA (1988) Was there a prelude to the dinosaurs demise? Science 239:729-730
- Kerr RA (1996a) Ancient life on Mars? Science 273:864-866
- Kerr RA (1996b) Did a plate tectonic surge flood earth? Science 274:1611
- Kerr RA (1997) An ocean emerges on Europa. Science 276:355
- Kerr RJ (1898) The dry-season habits of Lepidosiren. Proc Zool Soc Lond 1898:41-44

- Khalaf El Duweini A (1957) On the gills and respiration in *Alma nilotica* Grube. Publ 2nd Sci Arab Congr (1955) Cairo 833-838
- Kiceniuk JW, Jones DR (1977) The oxygen transport system in trout (Salmo gairdnen) during sustained exercise. J Exp Biol 69:247–260
- Kiel JW, Shepherd AP (1989) Optimal hematocrit for canine gastric oxygenation. Am J Physiol 256:H472-H477
- Kikkawa Y (1970) Morphology of alveolar lining layer. Anat Rec 167:389-400
- Kikuchi S (1992) Histological and fine structural evidence of the ion-transporting role of the gill and the neck organ of a fresh water brachiopod, *Branchinella kugenumaensis*. Annu Rep Iwate Med U Sch Lib Arts Sci 27:13-28
- Kiley JP, Faraci FM, Fedde MR (1985) Gas exchange during exercise in hypoxic ducks. Respir Physiol 59:105–115
- Kilgore DL, Boggs DF, Birchard GF (1979) Role of the rete mirabile ophthalmicum in maintaining body-to-brain temperature difference in pigeons. J Comp Physiol 129:119–122
- Kimball RE, Reddy K, Peirce TH, Schwartz LW, Mustafa MG, Cross CE (1976) Oxygen toxicity: augmentation of antioxidant defense mechanisms in rat lung. Am J Physiol 230:1425–1431
- Kimbel P, Weinbaum G (1975) Role of leucoproteases in the genesis of emphysema. In: Junod AF, Haller R (eds) Lung metabolism. Academic Press, New York, pp 25-37
- Kimoto T, Fujinaga T (1990) Non-biotic synthesis of organic polymers on H<sub>2</sub>S-rich sea floor: a possible reaction in the origin of life. Mar Chem 30:179–192
- Kimura M (1983) The neutral theory of molecular evolution. Cambridge University Press, Cambridge
- Kimura A, Gomi T, Kikuchi Y, Hashimoto T (1987) Anatomical studies of the lung of air breathing fish. I. Gross anatomical and light microscopic observations of the lungs of the African lungfish *Protopterus aethiopicus*. J Med Soc Toho Univ 34:1-18
- King AS (1966) Structural and functional aspects of the avian lung and its air sacs. Int Rev Gen Exp Zool 2:171–267
- King AS, McLelland J (1975) Outlines of avian anatomy. Bailliéle Tindall, London
- King MC, Wilson AC (1975) Evolution at two levels in human and chimpanzees. Science 188:107-116
- King JR, Farner DA (1969) Energy metabolism, thermoregulation, and body temperature. In: Marshall AJ (ed) Biology and comparative physiology of birds, vol 2. Academic Press, London, pp 215–279
- Kinney JL, White FN (1977) Oxidative ventilation in a turtle, Pseudemys floridana. Respir Physiol 31:327-332
- Kinnula VL, Adler K, Akley N, Crapo J (1992) Release of reactive oxygen species by guinea pig tracheal epithelial cells *in vitro*. Am J Physiol 262:L708–L712
- Kirsch R, Nonnotte G (1977) Cutaneous respiration in there fresh water teleosts. Respir Physiol 29:339–354
- Kirschner LB (1982) Physical basis of solute and water transfer across gills. In: Houlihan DF, Rankin JC, Shuttleworth TJ (eds) Gills. Cambridge University Press, Cambridge, pp 63–76
- Kirschner LB (1993) The energetics of osmotic regulation in ureotelic and hypoosmotic fishes. J Exp Zool 267:19-26
- Kirschner RP (1994) The Earth's elements. Sci Am 271:37-43
- Kitcher P (1984) 1953 and all that: a tale of two sciences. Philos Rev 93:102-124
- Kitterman JA, Ballard PL, Clements JA, Mescher EJ Tooley WH (1979) Tracheal fluid in fetal lambs: spontaneous decrease prior to birth. J Appl Physiol 47:985–989
- Klaver CJJ (1973) Lung anatomy: aid in chameleon taxonomy. Beaufortia 20:155-177
- Klaver CJJ (1981) Lung morphology in the chamaeleonidae (Sauria) and its bearing upon phylogeny, systematics and Zoogeography. Z Zool Syst Evolutionsforsch 19:36–58
- Kleiber M (1965) Respiratory exchange and metabolic rate. In: Fenn WO, Rahn H (eds) handbook of physiology, sect 3, respiration, vol II. American Physiological Society, Washington, DC, pp 927–938
- Kleinschmidt T, Sgouros JG (1987) Haemoglobin sequences. Biol Chem Hoppe-Seyler 368:579– 615
- Klemm RD, Gatz RN, Westfall JA, Fedde MR (1979) Microanatomy of the lung parenchyma of a tegu lizard *Tupinambis nigropunctatus*. J Morphol 161:257–280
- Klika E, Lelek A (1967) A contribution to the study of the lungs of *Protopterus annectens* and *Polypterus segegalensis*. Folia Morphol 15:168–175
- Klite PD (1965) Intestinal bacterial flora and transit time in three neotropical bat species. J Bacteriol 90:375–379
- Knight DR, Schaffartzik W, Poole DC, Hogan MC, Bebout DE, Wagner PD (1992) Hyperoxia improves leg VO<sub>2max</sub>. FASEB J 139:A1466
- Knight J (1984) Studies on the biology and biochemistry of *Pholas dactylus* L., PhD Thesis, University of London, London
- Knight J, Knight R (1986) The blood vascular system of the gills of *Pholas dactylus* L. (Mollusca, Bivalvia, Eulamellibranchia). Philos Trans R Soc Lond 313B:509–523
- Knoll AH (1979) Archean photoautotrophy: some alternatives and limits. Origins Life 9:313–327 Knoll AH (1991) The end of the Proterozoic Eon. Science 265:64–73
- Knoll AH (1996) Breathing room for early animals. Nature (Lond) 382:111-112
- Knutton S, Jackson D, Graham JM, Micklem KJ, Pastemak CA (1976) Microvilli and cell swelling. Nature (Lond) 262:52–54
- Kobayashi H, Pelster B, Scheid P (1989a) Solute back-diffusion in counter-current flow. Respir Physiol 78:59–71
- Kobayashi H, Pelster B, Scheid P (1989b) Water and lactate movement in the swim bladder of the eel, *Anguilla anguilla*. Respir Physiol 78:45–57
- Kobayashi H, Pelster B, Scheid P (1990) Carbon dioxide back-diffusion in the rete aids O<sub>2</sub> secretion in the swim bladder of the eel. Respir Physiol 79:231–242
- Koch H (1938) The absorption of chloride ions by the anal papillae of diptera larvae. J Exp Biol 15:152–160
- Koch R, Seymour S, Bartholomew GA, Barnhart MC (1983) Respiration and heat production by inflorescence of *Philodendron selloum*. Planta 157:336-343
- Koch R, Seymour S, Barnhart MC, Bartholomew GA (1984) Respiratory gas exchange during thermogenesis in *Philodendron selloum*. Planta 161:229–232
- Kokko JP, Tisher CC (1976) Water movement across nephron segments involved with the counter-current multiplication system. Kidney Int 10:64–81
- Kon K, Maeda N, Sekiya M, Shiga T, Suda T (1980) A method for studying oxygen diffusion barrier in erythrocytes: effects of haemoglobin content and membrane cholestrol. J Physiol (Lond) 309:569–590
- Kon K, Maeda N, Shiga T (1983) The influence of deformation of transformed erythrocytes during flow on the rate of oxygen release. J Physiol (Lond) 339:573–584
- König MF, Lucocq JM, Weibel ER (1993) Demonstration of pulmonary vascular perfusion by electron and light microscopy. J Appl Physiol 75:1877–1883
- Konings WN, van Driel R, van Bruggen EF, Gruber M (1969) Structure and properties of hemocyanins. V. Binding oxygen and copper in *Helix pomatia* hemocyanin. Biochim Biophys Acta 194:55–66
- Kooyman GL (1985) Physiology without restraint in diving mammals. Mar Mammal Sci 1:166– 178
- Kooyman GL, Cornell LH (1981) Flow properties of expiration and insiration in a trained bottlenosed porpoise. Physiol Zool 54:55-61
- Kooyman GL, Sinnett EE (1979) Pulmonary shunts in harbour seals and sea lions during simulated dives to depth. Physiol Zool 55:105-111
- Koshland DE (1992) The molecule of the year. Science 258:1861
- Kostelecka-Mycha A (1987) Respiratory function of a unit of blood volume in the little auk
- (*Plautus alle*) and the Arctic tern (*Stema prardisaea*). Comp Biochem Physiol 86A:117-120 Koteja P (1986) Maximum cold-induced oxygen consumption in the house sparrow, *Passer domesticus* (L.). Physiol Zool 59:43-48
- Kozlowski J (1993) Measuring fitness in life history studies. Trends Ecol Evol 8:84-85
- Kramer DL (1978) Ventilation of the respiratory gas bladder in *Hoplerythrinus unitaeniatus* (Pisces, Characoidei, Erythrinidae). Can J Zool 56:931–938
- Kramer DL (1980) A comparative test of the adaptive significance of aquatic surface respiration in fishes. Am Zool 20:742
- Kramer DL (1983) The evolutionary ecology of respiratory mode in fishes: an analysis based on the costs of breathing. Environ Biol Fish 9:145–158

Kramer DL (1987) Dissolved oxygen and fish behaviour. Environ Biol Fish 18:91-92

Kramer DL (1988) The behavioural ecology of air breathing by aquatic animals. Can J Zool 66:89-94

- Kramer DL, Graham JB (1976) Synchronus air breathing, a social component of respiration in fishes. Copeia 1976:689–697
- Kramer DL, McClure M (1981) The transit cost of aerial respiration in the catfish, *Corydoras* aeneus (Callichthyidae). Physiol Zool 54:189–194
- Kramer DL, McClure M (1982) Aquatic surface respiration, widespread adaption to hypoxia in tropical fresh water fishes. Environ Biol Fish 7:47–55
- Kramer DL, Lindsey CC, Moodie GEE, Stevens ED (1978) The fishes and the aquatic environment of the Central Amazon Basin, with particular reference to respiratory patterns. Can J Zool 56:717–729
- Kràtký J (1981) Postnatale Entwicklung der Wasserfledermaus, Myotis daubentoni Kuhl, 1981 und bisherige Kenntnis dieser Problematik im Rahmen der Unterordnung Microchiroptera (Mammalia: Chiroptera). Fol Mus Rer Natur Bohem Occident Zool 16:1– 34
- Krebs JR, Harvey PH (1986) Busy doing nothing efficiently. Nature (Lond) 320:18-19
- Krenz GS, Linehan JH, Dawson CA (1992) A fractal continuum model of the pulmonary arterial tree. J Appl Physiol 72:2225–2237
- Kreuzer F (1970) Facilitated diffusion of oxygen and its possible significance: a review. Respir Physiol 9:1-30
- Krogh A (1910) On the mechanism of the gas exchange in the lungs. Skand Arch Physiol 23:248– 278
- Krogh A (1913) On the composition of the air in the tracheal system of some insects. Skand Arch Physiol 29:29–36
- Krogh A (1920a) Studien über Tracheen Respiration. II. Über Gasdiffusion in den Tracheen. Pfluegers Arch Gesamte Physiol Menschen Tiere 179:95–112
- Krogh A (1920b) Studien über Tracheen Respiration. III. Die Kombination von mechanischer Ventilation mit Gasdiffusion nach Versuchen an Dytiscuslarven. Pfluegers Arch Gesamte Physiol Menschen Tiere 179:113–120
- Krogh A (1941) The comparative physiology of respiratory mechanisms. University of Pennsylvania Press, Philadelphia
- Kruhoffer M, ML, Abe AS, Johansen K (1987) Control of breathing in an amphibian Bufo paracnemius: effects of temperature and hypoxia. Respir Physiol 69:267-275
- Kuethe AM (1975) Prototypes in nature. The carry-over into technology. TechniUM, Spring Issue 1975:3-20
- Kuethe DO (1988) Fluid mechanical valving of air flow in bird lungs. J Exp Biol 136:1-12
- Kuhn W, Ramel A, Kuhn H, Marti E (1963) The filling mechanism of the swim bladder. Generation of high gas pressures through hairpin counter-current multiplication. Experientia 19:497-511
- Kulzer E (1965) Temperaturregulation bei Fledermäusen (Chiroptera) aus verschiedenen Klimazonen. Z Vergl Physiol 50:1–34
- Kurland CG (1992) Evolution of mitochondria genomes and the genetic code. BioAssays 14:709–714
- Kutchai H, Steen JB (1971) Permeability of the shell and shell membranes of hen's eggs during development. Respir Physiol 11:265-278
- Kutty MN, Saunders RL (1973) Swimming performance of young Atlantic salmon (Salmon salar) as affected by reduced ambient oxygen concentration. J Fish Res Board Can 30:223–227
- Kylstra JA (1962) Drowning: the role of salts in the drowning fluid. Acta Physiol Pharmacol Neerl 10:327-334
- Kylstra JA (1968) Experiments in water-breathing. Sci Am 219:66-74
- Kylstra JA (1969) The feasibility of liquid breathing and artificial gills. In: Bennet PB, Elliot DH (eds) The physiology and medicine of diving and compressed air work. Bailleire, London, pp 193–212
- Kylstra JA, Schoenfisch WH (1972) Alveolar surface tension in fluorocarbon-filled lungs. J Appl Physiol 33:32–35

- Kylstra JA, Tissing MO (1963) Fluid breathing. In: Clinical applications of hyperbaric oxygen. Proc 1st Int Congr, Amsterdam, pp 371–379
- Kylstra JA, Paganelli CV, Lanphier EH (1966) Pulmonary gas exchange in dogs ventilated with hyperbarically oxygenated liquid. J Appl Physiol 21:177–184
- Kylstra JA, Rausch DC, Hall KD, Spock A (1971) Volume-controlled lung lavage in the treatment of asthma, bronchiectasis, and mucoviscidosis. Am Rev Respir Dis 103:651–665
- Labra MA, Rosenmann M (1994) Energy metabolism and evaporative water loss of *Pristidactylus* lizards. Comp Biochem Physiol A 109:369–376
- Laburn HP, Goelst K, Mitchell D (1994) Body temperatures of lambs and their mothers measured by radio-telemetry during parturition. Experientia 50:708–711
- Laga EM, Driscoll SG, Munro HN (1974) Human placental structure: relationship to foetal nutrition. In: Josmovich JB, Reynods M, Cobo E (eds) Problems of human reproduction, vol 2, lactogenic hormones, foetal nutrition and nutrition. John Wiley, London, pp 143–181
- Lahiri S (1975) Blood oxygen affinity and alveolar ventilation in relation to body weight in mammals. Am J Physiol 229:529-536
- Lahiri S, Szidon JP, Fishman AP (1970) Potential respiratory and circulatory adjustments to hypoxia in the African lungfish. Fed Proc 29:1141-1148
- Lai NC, Graham JB, Burmett L (1990) Blood respiratory properties and the effect of swimming on blood gas transport in the leopard shark *Triakis semifasciata*. J Exp Biol 151:161–173
- Laitman TJ, Reidenberg JS, Marquez S, Gannon PJ (1996) What the nose knows: new understandings of the Neanderthal upper respiratory tract. Proc Natl Acad Sci USA 93:10543– 10545
- Lallier F, Truchot JP (1989) Hemolymph oxygen transport during environmental hypoxia in the shore crab, *Carcinus maenas*. Respir Physiol 77:323–336
- LaManna JC, Vendel LM, Farrell RM (1992) Brain adaptation to chronic hypobaric hypoxia in rats. J Appl Physiol 72:2238–2243
- Lambertsen CJ (1961) Respiration. In: Bard P (ed) Medical physiology. Mosby, St Louis, pp 32– 98
- Lamy J, Truchot JP, Giles R (1985) (eds) Respiratory pigments in animals. Springer, Berlin Heidelberg New York
- Landes RR, Leohardt KO, Duruman N (1964) A clinical study of the oxygen tension of the urine and renal structures. II. J Urol 92:171–178
- Landis GP, Snee LW (1991) 40Ar/39r systematics and argon diffusion in amber: implicitons for ancient Earth atmosphere. Paleogeogr Paleoclimatol Paleoecol 97:63–67
- Lang BF, Burger G, O'Kelly CJ, Cedergren R et al. (1997) An ancestral mitochondrial DNA resembling a eubacterial genome in miniature. Nature (Lond) 387:493-496
- Langille BL, Jones (1975) Central cardiovascular dynamics of ducks. Am J Physiol 228:1856– 1861
- Langston W (1981) Pterosaurs. Sci Am 244:92-102
- Lanphier EJ (1969) Pulmonary function. In: Bennett PB, Elliot DH (eds) The physiology and medicine of diving and compressed-air work. Bailere, Tindall and Cassell, London, pp 58–112
- Lapennas GN, Schimdt-Nielsen K (1977) Swim bladder permeability to oxygen. J Exp Biol 67:175-196
- Larimer JL, Schimdt-Nielsen K (1960) A comparison of blood carbonic anhydrase of various mammals. Comp Biochem Physiol 1:19-23
- Larson A, Chippindale P (1993) Molecular approaches to the evolutionary biology of plethodontid salamanders. Herpetologia 49:204–215
- Lasiewski RC (1962) The energetics of migrating hummingbirds. Condor 64:324
- Lasiewski RC (1963a) Oxygen consumption of torpid, resting, active, and evaporative water loss in hummingbirds. Physiol Zool 36:122-140
- Lasiewski RC (1963b) The energetic cost of small size hummingbirds. Proc XIII Int Congr 1095– 1103
- Lasiewski RC (1964) Body temperatures, heart and breathing rate, and evaporative water loss in hummingbirds. Physiol Zool 37:212–223
- Lasiewski RC (1972) Respiration function in birds. In: Famer DS, King JR, Parkes KC (eds) Avain biology, vol II. Academic Press, New York, pp 271-342

- Lasiewski RC, Calder WR (1971) A preliminary allometric analysis of respiratory variables in resting birds. Respir Physiol 11:152–166
- Lasiewski RC, Weathers WW, Bernstein MV (1967) Physiological responses of the giant hummingbird, *Patagona gigas*. Comp Biochem Physiol 23:797–813
- Lauder GV (1980) Evolution of feeding mechanisms in primitive actinopterygian fishes: A functional anatomical analysis of *Polypterus*, *Lepisosteus* and *Amia*. J Morphol 163:283-317
- Lauder GV (1981) Form and function: structural analysis in evolutionary morphology. Paleobiology 7:430-442
- Lauder GV, Liem KF (1983) The evolution and interrelationships of actinopterygian fishes. Bull Mus Comp Zool 150:95–197
- Lauder GV, Liem KF (1989) The role of historical factors in the evolution of complex organismal functions. In: Wake DB, Roth G (eds) Complex organismal functions: integration and evolution in vertebrates. John Wiley, London, pp 63–78
- Laurent GJ (1986) Lung collagen: more than scaffoding. Thorax 41:418-428
- Laurent P (1982) Structure of vertebrate gills. In: Houlihan DF, Rankin JC, Shuttleworth TJ (eds) Gills. Cambridge University Press, Cambridge, pp 25–43
- Laurent P (1984) Gill internal morphology. In: Hoar WS, Randall DJ (eds) Fish physiology, vol A. Academic Press, New York, pp 73–183
- Laurent P (1985) Organization and control of the respiratory vasculature in lower vertebrates: are there anatomical gill shunts? In: Johansen K, Burrgren WW (eds) Cardiovascular shunts. Alfred Benzon Symposium 21, Munksgaard, Copenhagen, pp 57–70
- Laurent P (1996) Vascular organization of lungfish, a landmark in ontogeny and phylogeny of air-breathers. In: Munshi JSD, Dutta HM (eds) Fish morphology: horizon of new research. Science Publishers, Lebanon, New Hampshire, pp 47–58
- Laurent P, Dunel-Erb S (1980) Morphology of gill epithelia. Am J Physiol 238:R147-R159
- Laurent P, Hebibi (1989) Gill morphometry and fish osmoregulation. Can J Zool 67:3055-3063
- Laurent P, Perry SF (1990) Effects of cortisol on gill chloride cell morphology and ionic uptake in the fresh water salmonid fish. Cell Tissue Res 259:429–442
- Laurent P, Perry SF (1991) Environmental effects on gill morphology. Physiol Zool 69:2-25
- Laurent P, Delaney RG, Fishman AP (1978) The vasculature of the gills in the aquatic and aestivating lungfish (*Protopterus aethiopicus*). J Morphol 156:173-208
- Laurent P, Maina JN, Bergman HL, Narahara A, Walsh PJ, Wood CM (1995) Gill structure of a fish from an alkaline lake: effect of short-term exposure to neutral conditions. Can J Zool 73:1170–1181
- Laverack MS (1963) The physiology of earthworms. Pergamon Press, Oxford
- Laybourne RC (1974) Collision between a vulture and an aircraft at an altitude of 37 000 ft. Wilson Bull 86:461–462
- Lazzaro X (1987) A review of planktivorous fishes: their evolution, feeding behaviours, selectivities, and impacts. Hydrobiologia 146:97–167
- Leatherland JF, Hyder M, Ensor DM (1974) Regulation of Na<sup>+</sup> and K<sup>+</sup> concentrations in five African species of *Tilapia* fishes. Comp Biochem Physiol 48A:699-710
- Lechner AJ (1984) Pulmonary design in microchiropteran lung (*Pipistrellus subflavus*) during hibernation. Respir Physiol 59:301–312
- Lee DH, Granja JR, Martinez JA, Severin K, Ghadiri MR (1996) A self-replicating peptide. Nature (Lond) 382:525–531
- Lee R, Mayhew TM (1995) Volumes of villi and intervillous pores in placentae from low and high altitude pregnancies. J Anat 186:349–355
- Lefevre J (1983) Teleonomical optimization of a fractal model of the pulmonary arterial bed. J Theor Biol 102:225–248
- Leigh GJ (1997) Biological nitrogen fixation and model chemistry. Science 275:1442
- Leith DE, Mead J (1966) Maximum expiratory flow in liquid filled lungs. Fed Proc 25:506
- Leming TD, Stuntz WE (1984) Zones of coastal hypoxia revealed by satellite scanning have implications for strategic fishing. Nature (Lond) 310:136–138
- Lenfant C (1973) High altitude adaptation in mammals. Am Zool 13:447-456
- Lenfant C, Johansen K (1967) Respiratory adaptations in selected amphibians. Respir Physiol 2:247-260

- Lenfant C, Johansen K (1968) Respiration in an African lungfish, *Protopterus aethiopicus*: respiratory properties of blood and normal patterns of breathing and gas exchange. J Exp Biol 49:437-452
- Lenfant C, Johansen K (1972) Gas exchange in gill, skin and lung breathing. Respir Physiol 14:211-218
- Lenfant C, Johansen K, Grigg GC (1966) Respiratory properties of blood and pattern of gas exchange in the lungfish *Neocertodus forsteri* (Kreffti). Respir Physiol 2:1–21
- Lenfant C, Elsner R, Kooyman GL, Drabek CM (1969) Respiratory function of blood of adult and fetus Weddell seal *Leptonychotes weddelli*. Am J Physiol 216:1595–1597
- Lenfant C, Johansen K, Torrance JD (1970a) Gas transport and oxygen storage capacity in some pinnipeds and the sea otter. Respir Physiol 9:277–286
- Lenfant C, Johansen K, Hanson D (1970b) Bimodal gas exchange and ventilation-perfusion relationship in lower vertebrates. Fed Proc 29:1124–1129
- Lennard R, Huddart H (1989) Electrophysiology of the flounder heart (*Platichthys flesus*) the effect of agents which modify transmembrane ion transport. Comp Biochem Physiol 93C:499–509
- Leopold LB, Davies KS (1968) Water. Time-Life Books, Amsterdam
- Lessard J, Val AL, Aota S, Randall DJ (1995) Why is there no carbonic anhydrase activity available to fish plasma? J Exp Biol 1995:31-38
- Levi HW (1967) Adaptations of respiratory systems of spiders. Evolution 21:571
- Levine BW, Talamo RC, Kazemi H (1973) Action and metabolism of bradykinin in dog lung. J Appl Physiol 34:821–826
- Levine OR, Mellins RB, Fishman AP (1965) Quantitative assessment of pulmonary oedema. Circ Res 17:414–423
- Levine S (1976) Competitive interactions in ecosystems. Am Nat 110:903-910
- Levins R, Lewontin R (1985) The dialetical biologist. Harvard University Press, Cambridge
- Levinton JS (1992) The big bang of animal evolution. Sci Am 267:52-59
- Levy M, Achituv Y, Susswein AJ (1989) Relationship between respiratory pumping and oxygen consumption in *Aplysia depilans* and *A. fasciata*. J Exp Biol 141:389–405
- Lewin R (1988) Egg laying in birds remains a hot issue. Science 233:465
- Lewis AB, Heymann MA, Rudolph AM (1976) Gestational changes in pulmonary vascular responses in foetal lambs *in utero*. Circ Res 39:536-541
- Lewis JF, Tabor B, lkegami M, Jobe AH, Joseph M, Absolom D (1993) Lung function and surfactant distribution in saline-lavaged sheep given instilled vs. nebulized surfactant. J Appl Physiol 74:1256–1264
- Lewis RW (1970) The densities of three classes of marine lipids in relation to their possible role as hydrostatic agents. Lipids 5:151–165
- Lewis SV (1980) Respiration in lampreys. Can J Fish Aquatic Sci 37:1711-1722
- Lewis SV, Potter IC (1982) A light and electron microscope study of the gills of the lampreys (*Geotria australis*) with particular reference to the water-blood pathway. J Zool (Lond) 198:157–176
- Lewontin RG (1979) Fitness, survival and optimality. In: Horn DH, Mitchell R, Stairs GR (eds) Analysis of ecological systems. Ohio State University, Columbus, pp 3–21
- Liem KF (1961) Tetrapod parallelisms and other features in the functional morphology of the blood vascular system of *Fluta alba* Zuiew (Pisces: Teleostei). J Morphol 108:131–143
- Liem KF (1980) Air ventilation in advanced teleosts: biomechanical and evolutionary aspects. In: Ali MA (ed) Environmental physiology of fishes. Plenum Press, New York, pp 57–91
- Liem KF (1981) Larvae of air-breathing fishes as counter-current flow devices in hypoxic environments. Science 211:1177–1179
- Liem KF (1984) The muscular basis of aquatic and aerial ventilation in the air breathing teleost fish *Channa*. J Exp Biol 113:1–18
- Liem KF (1985) Ventilation. In: Hildebrand M, Bramble DM, Liem KF, Wake DB (eds) Functional vertebrate morphology. Harvard University Press, Cambridge, pp 185–209
- Liem KF (1987a) Form and function of lungs: the evolution of air-breathing mechanisms. Am Zool 28:739–759
- Liem KF (1987b) Functional design of the air ventilation apparatus and overland excursions by teleosts. Fieldiana Zool New Ser 37:1–29

- Liem KF (1989) Respiratory gas bladders in teleosts: functional conservatism and morphological diversity. Am Zool 29:333-352
- Liem KF (1991) Towards a new morphology: pluralism in research and education. Am Zool 31:759–767
- Liem KF, Wake DB (1985) Morphology: current approaches and concepts. In: Hildebrand M (ed) Functional vertebrate morphology. Harvard University Press, Cambridge, pp 269– 305
- Lien DC, Wagner WW, Capen C, Haslett WL et al. (1987) Physiological neutrophil sequestration in the lung: visual evidence for localization in capillaries. J Appl Physiol 62:1236–1243
- Liew HD (1970) Maintenance of underwater gas bubbles: how sea weed solves the problem. In: Farhi LE, Rahn H (eds) Studies in pulmonary physiology: mechanics, chemistry, and circulation of the lung. USAF School of Aerospace Medicine, Aerospace Medical Division (AFSC), Brooks Air Force Base, Houston Texas, pp 237–253
- Liggins GC, Vilos GA, Campos GA, Kitterman JA, Lee CH (1981) The effect of spinal cord transection on lung development in foetal sheep. J Dev Physiol 3:267-274
- Lilja C (1997) Oxygen consumption and vital organ masses in young growing quail Coturnix coturnix japonica. Acta Physiol Scand 160:113–114
- Lillegraven JA, Thompson SD, McNab BK, Patton JL (1987) The origin of the Eutherian mammals. Biol J Linn Soc 32:281-336
- Lindal T (1990) Repair of intrisic DNA lesions, Mut at Res 238:305-311
- Linderman PV (1991) Survivorship of overwintering painted turtles, *Chrysemys picta* in Northern Idaho. Can Field Nat 105:263–266
- Lindroth A (1938a) Studien über die respiratorischen Mechanismen von *Nereis virens* Sars. Zool Bidr Upps 17:367–497
- Lindroth A (1938b) Gibt es bei den Polychäten intestinale Atmung per anum? Z Vergl Physiol 25:283–292
- Lindroth A (1939) Beobachtungen an Capitelliden, besonders hinsichtlich ihrer Respiration. Zool Anz 127:285–297
- Lindsay HA, Friex ED, Hoversland AS et al. (1971) Respiration and circulation. In: Altiman PL, Dittmer DS (eds) Handbook of physiology: respiration and circulation. Am Soc Exp Biol, Bethesda, pp 438–543
- Lindstedt SL (1984) Pulmonary transit time and diffusing capacity in mammals. Am J Physiol 246:R384-R388
- Lindstedt SL, Jones JH (1988) Symmorphosis: The concept of optimal design. In: Feder ME, Bennet AF, Burggren WW, Huey RB (eds) New directions in ecological physiology. Cambridge University Press, Cambridge, pp 290–310
- Lindstedt SL, Hokanson JF, Wells DJ, Swain SD, Hoppelar H, Navarro V (1991) Running energetics in the pronghom antelope. Nature (Lond) 353:748
- Linzen B, Gallowitz P (1975) Enzyme activity patterns in muscles of the lycosid spider, *Cupiennius salei*. J Comp Physiol 96:101-109
- Little C (1983) The colonization of land. Origins and adaptations of terrestrial animals. Cambrigde University Press, Cambridge
- Little C (1990) The terrestrial invasion: an ecophysiological approach to the origins of land animals. Cambridge University Press, Cambridge
- Liversmore RA, Smith AG, Briden JC (1985) Palaeomagnetic constraints on the distribution of continents in the rate Silurian and early Devonian. Philos Trans R Soc Lond 309B:29–56
- Lloyd D, Rossi EL (1993) Biological rhythms as organization and information. Biol Rev 68:563– 577
- Locke MJ (1958a) The structure of insect tracheae. Q J Microsc Sci 98:487-492
- Locke MJ (1958b) Coordination of growth in the tracheal system of insects. Q J Microsc Sci 99:373-391
- Locke MJ (1958c) The formation of trachea and tracheoles in *Rhodinius prolixus*. Q J Microsc Sci 99:29–46
- Lockely Rm (1970) The most aerial bird in the world. Animals 13:4-7
- Lockwood APM (1968) Aspects of physiology of Crustacea. Oliver Boyd, Edinburgh
- Loesch H (1960) Sporadic mass shoreward migration of demersal fish and crustaceans in Moblie Bay, Alabama. Ecology 41:291–298

- Logan GA, Hayes JM, Hieshima GB, Summon RE (1995) Terminal Proterozoic reorganization of biogeochemical cycles. Nature (Lond) 376:53–56
- Lomholt JP (1976a) The development and of the oxygen permeability of the avian egg shell during incubation. J Exp Zool 198:177-203
- Lomholt JP (1976b) Relationship of weight loss to ambient humidity of birds eggs during incubation. J Comp Physiol 105:189-196
- Lomholt JP, Johansen K (1976) Gas exchange in the amphibious fish, Amphipnous cuchia. J Comp Physiol 107:141-157
- Lomholt JP, Johansen K, Maloiy GMO (1975) Is aestivating lungfish the first vertebrate with suctional breathing? Nature (Lond) 257:787-788
- Londraville RL, Sidell BD (1990) Ultrastructure of aerobic muscles in Antarctic fishes may contribute to maintenance of diffusive fluxes. J Exp Biol 150:205-220
- Longmuir IS (1976) Search for new tissue oxygen carriers. In: Oxygen and physiological function. Professional Information Library, Dallas
- Longmuir IS, Bourke A (1959) Application of Warburg's equation to tissue slices. Nature (Lond) 184:634–635
- Longmuir IS, Bourke A (1960) The measurement of the diffusion of oxygen through respiring tissue. Biochem J 76:225–229
- Longmuir IS, Sun S (1970) A hypothetical tissue oxygen carrier. Microvasc Res 2:287-293
- Longo LD, Power GG, Forster RE (1967) Respiratory function in the placenta as determined by carbon monoxide in sheep and dogs. J Clin Invest 46P:812-828
- Longo LD, Power GG, Forster RE (1969) Placental diffusing capacity for carbon monoxide at varying aprtial pressures of oxygen. J Appl Physiol 26:360-370
- Longo LD, Hill EP, Power GG (1973) Factors affecting placental oxygen transfer. Chemical Engineering in Medicine. Adv Chem Ser 118:88–129
- Longworth KE, Jones JH, Bicudo JEP, Taylor CR, Weibel ER (1989) High rate of  $O_2$  consumption in exercising foxes: large  $PO_2$  difference drives diffusion across the lung. Respir Physiol 77:263–276
- López J (1995) Anatomy and histology of the lung and air sacs of birds. In: Pastor LM (ed) Histology, ultrastructure and immunohistochemistry of the respiratory organs in nonmammalian vertebrates. Publicaciones de la Universitat de Murcia, University of Murcia, Murcia, Spain, pp 179-233
- Lorenz RD, McKay CP, Lunine JI (1997) Photochemically driven collapse of Titan's atmosphere. Science 275:642–643
- Losa GA, Baumann G, Nonnenmacher TF (1992) Fractal dimension of pericellular membranes in human lymphocytes and lymphoblastic leukemia cells. Pathol Res Pract 188:680–686
- Lotshaw DP (1977) Temperature adaptation and effects of thermal acclimation in Rana sylvatica and Rana catesbeiana. Comp Biochem Physiol 56A:287-294
- Loudon C (1989) Tracheal hypertrophy in mealworms: design and plasticity in oxygen supply systems. J Exp Biol 147:217-235
- Lovejoy TE (1994) The quantification of biodiversity: an esoteric quest or a vital component of sustainable development? Philos Trans R Soc Lond 345B:81–87
- Loveridge JP (1970) Observations on nitrogenous excretion and water relations of *Chiromantis xerapherina* (Amphibia, Anura). Arnoldia 5:1-6
- Loveridge JP (1976) Strategies of water conservation in the Southern Africa frogs. Zool Afr 11:319-333
- Loveridge JP (1980) Cuticle water relations. In: Miller TA (ed) Cuticle techniques in arthropods. Springer, Berlin Heidelberg New York, pp 301–366
- Løvtrup S (1977) The phylogeny of vertebrata. Wiley, London
- Low FN (1953) The pulmonary alveolar epithelium of laboratory animals and man. Anat Rec 117:241-263
- Low WP, Lane DJ, Ip YK (1988) A comparative study of terrestrial adaptations of the gills in three mudskippers – Periophthalmus chrysospolos, Boleophthalmus boddaerti, and Periophthalmus schlosseri. Biol Bullmar Biol Woods Hole 175:334-438
- Low WP, Ip YK, Lane DJ (1990) A comparative study of the gill morphology of the mudskippers, *Periophthalmus chrysospilos, Boleophthalmus boddaerti* and *Periophthalmus schlosseri*. Zool Sci 7:29–38

- Lowe CA, Tuma RF, Sivieri EM, Shaffer TH (1979) Liquid ventilation: cardiovascular adjustments with secondary hyperlactatemia and acidosis. J Appl Physiol 47:1051–1057
- Lucas AM, Denington EM (1961) A brief report on anatomy, histology and nomenclature of air sacs in the fowl. Avian Dis 5:460–481
- Luchtel DL, Kardong KV (1981) Ultrastructure of the lung of the rattlesnake, Crotalus viridis oreganus. J Morphol 169:29-47
- Luckett WP (1976) Ontogeny of amniote foetal membranes and their applications to phylogeny. In: Hecht MK, Goody PC, Hecht BM (eds) The major patterns in vertebrate evolution. Plenum Press, London, pp 439–516
- Luckett WP (1993) Uses and limitations of mammalian fetal membranes and placenta for phylogenetic reconstruction. J Exp Zool 266:514-527
- Luckett WP, Hartenberger JL (1993) Monophyly or polyphyly of the order rodentia: possible conflict between morphological and molecular interpretations. J Mammal Evol 1:127-147
- Ludwig KS (1965) Zur Feinstruktur der materno-foetalen Verbindung im Placenta des Schafes (Ovis ovis). Experientia 18:212-213
- Luft UC (1965) Aviation physiology. In: Fenn WO, Rahn H (eds) Handbook of physiology, sect 3, respiration vol II. American Physiological Society, Washinton, DC, pp 1099–1147
- Lüling KH (1964) Über die Atmung des Hoploerythrinus unitaeniatus (Pisces, Erythrinidae). Bonn Zool Beitr 15:90-102
- Lundberg K, Almgren B, Odelberg C (1983) Nagot om vattenfladdermusens (*Myotis dubenonoi*) ekologi. Fauna Flora (Stockh) 78:237–242
- Lundberg S, Persson L (1993) Optimal body size and resource density. J Theor Biol 164:163-180
- Lutcavage ME, Lutz PL, Baier H (1987) Gas exchange in the loggerhead sea turtle, *Caretta*. J Exp Biol 131:365–372
- Lutz BR (1930) The effect of low oxygen tension on the pulsations of the isolated holothurian cloaca. Biol Bull 58:74-84
- Lutz PL, Schmidt-Nielsen K (1977) Effects of simulated altitude on blood gas transport in the pigeon. Respir Physiol 30:383–395
- Lutz PL, Longmuir IS, Turtle JV, Schmidt-Nielsen K (1973) Dissociation curve of bird blood and effect of red cell oxygen consumption. Respir Physiol 17:269–275
- Lutz PL, Longmuir IS, Schmidt-Nielsen K (1974) Oxygen affinity of bird blood. Respir Physiol 20:325-330
- Lykkeboe G, Johansen K (1975) Functional properties of haemoglobins in the teleost, *Tilapia graphami*. J Comp Physiol 104:1-11
- Lykkeboe G, Weber RE (1978) Changes in the respiratory properties of the blood in the carp, *Cyprinus carpio*, induced by diurnal variation in ambient oxygen tension. J Comp Physiol 128:117-125
- Lynch JJ, King JE, Chamberlain TK, Smith AL (1947) Effects of aquatic weed infestations on the fish and wildlife of the Gulf States. US Dept Int Spec Sci Rep 39:1–71
- Maarek JMI, Chang HK (1991) Pulsatile pulmonary microvascular pressure measured with vascular occlusion techniques. J Appl Physiol 70:998–1005
- Maarek JMI, Grimbert F (1994) Segmental pulmonary vascular resistances during oleic acid lung injury in rabbits. Respir Physiol 98:179–191
- MacArthur RA (1992) Gas bubble release by muskrats diving under ice: lost gas or potential oxygen pool? J Zool Lond 226:151–164
- MacArthur RH, Levine R (1967) The limiting similarity, convergence, and divergence of coexisting species. Am Nat 101:377–395
- Macchin J (1974) Water relations. In: Fretter VV, Peake J (eds) Pulmonates, vol V. Academic Press, London, pp 105-163
- MacFadden BJ (1985) Patterns of phylogeny and rates of evolution in fossil horses: Hipparions from the Miocene and Pliocene of North America. Paleobiology 11:245-257
- Mackay AR, Gelperin A (1972) Pharmacology and reflex responsiveness of the heart in the giant garden slug *Limax maximus*. Comp Biochem Physiol 43A:877–896
- Macklem PT, Bouverot P, Scheid P (1979) Measurement of the distensibility of the parabronchi in duck lungs. Respir Physiol 38:23-35
- Madan JJ, Wells MJ (1996) Why squid can breathe easy. Nature (Lond) 380:590
- Madigan MT, Marrs BL (1997) Extremophiles. Sci Am 276:66-71

- Maeda N, Shiga T (1994) Velocity of oxygen transfer and erythrocytes rheology. News Physiol Sci 9:22-27
- Magnussen H, Willmer H, Scheid P (1976) Gas exchange in the air sacs: contribution to respiratory gas exchange in ducks. Respir Physiol 26:129–146
- Maina JN (1981a) Morphometric study of the blood-gas barrier of the avian lung. J Anat 130
- Maina JN (1981b) Morphometry of the passerine and non-passerine lungs. Zentralbl Veterinaermed Reihe C Anat Histol Embryol 9:366–367
- Maina JN (1982a) A scanning electron microscopic study of the air and blood capillaries of the lung of the domestic fowl (*Gallus domesticus*). Experientia 35:614–616
- Maina JN (1982b) A stereological analysis of the paleopulmo and neopulmo respiratory regions of the avian lung (*Streptopelia decaocto*). Int Res Comm Syst (Biochem) 10:328
- Maina JN (1982c) Relationship between pulmonary morphometric characteristics and energetic requirements in four avian orders. J Anat 135:825
- Maina JN (1982d) A morphometric comparison of the lungs of two species of birds of different exercise capacities. J Anat 134:604–605
- Maina JN (1983) The bird lung: how is it made and how does it work? East African Natural History Bulletin, Nairobi, Kenya, July/August Issue, pp 51–54
- Maina JN (1984) Morphometrics of the avian lung. 3. The structural design of the passerine lung. Respir Physiol 55:291–309
- Maina JN (1985) A scanning and transmission electron microscopic study of the bat lung. J Zool (Lond) 205B:19–27
- Maina JN (1986) The structural design of the bat lung. Myotis 23:71-77
- Maina JN (1987a) The morphology of the lung of the African lungfish, *Protopterus aethiopicus*: a scanning electron microscopic study. Cell Tissue Res 250:191–196
- Maina JN (1987b) The morphology and morphometry of the adult normal baboon lung, *Papio anubis*. J Anat 150:229–245
- Maina JN (1987c) Morphometrics of the avian lung. 4. The structural design of the charadriiform lung. Respir Physiol 68:99–119
- Maina JN (1988a) Scanning electron microscopic study of the spatial organization of the air- and blood conducting components of the avian lung. Anat Rec 222:145–153
- Maina JN (1988b) The morphology and morphometry of the normal lung of the adult vervet monkey *Cercopithecus aethiops*. Am J Anat 183:258–267
- Maina JN (1989a) Morphometrics of the avian lung. In: King AS, McLelland J (eds) Form and function in birds, vol 4. Academic Press, London, pp 307-368
- Maina JN (1989b) A scanning and transmission electron microscopic study of the tracheal airsac system in a grasshopper (*Chrotogonus senegalensis*, Kraus) (Orthoptera: Acrididae: Pygomorphinae). Anat Rec 223:393-405
- Maina JN (1989c) The morphology of the lung of a tropical terrestrial slug, *Trichotoxon copleyi* (Mollusca: Gastropoda; Pulmonata): a scanning and transmission electron microscopic study. J Zool (Lond) 217:335–366
- Maina JN (1989d) The morphology of the lung of the East African tree frog *Chiromantis petersi* with observations on the skin and the buccal cavity as secondary gas exchange organs: a TEM and SEM study. J Anat 165:29–43
- Maina JN (1989e) The morphology of the lung of the black mamba *Dendroaspis polylepis* (Reptilia: Ophidia: Elapidae): a scanning and transmission electron microscopic study. J Anat 167:31-46
- Maina JN (1990a) A study of the morphology of of the gills of an extreme alkalinity and hyperosmotic adapted teleost *Oreochromis alcalicus grahami* (Boulenger) with particular emphasis on the ultrastructure of the chloride cells and their modifications with water dilution. Anat Embryol 181:83–98
- Maina JN (1990b) The morphology of the gills of the African fresh water crab *Potamon niloticus* (Ortmann-Crustacea-Brachyura-Potamonidae): a scanning and transmission electron microscopic study. J Zool (Lond) 221:499–515
- Maina JN (1990c) The morphology and morphometry of the lung of the lesser bushbaby *Galago* senegalensis. J Anat 172:129–144

- Maina JN (1991) A morphometric analysis of chloride cells in the gills of the teleosts Oreochromis alcalicus and Oreochromis niloticus and a description of presumptive urea excreting cells in Oreochromis alcalicus. J Anat 175:131-145
- Maina JN (1993) Morphometries of the avian lung: the structural-functional correlations in the design of the lungs of birds. Comp Biochem Physiol 105:397–410
- Maina JN (1994) Comparative pulmonary morphology and morphometry: The functional design of respiratory systems. In: Gilles R (ed) Advances in comparative and environmental physiology, vol 20. Springer, Berlin Heidelberg New York, pp 111–232
- Maina JN (1996) Perspectives on the structure and function in birds. In: Rosskoff E (ed) Diseases of cage and aviary birds. Williams and Wilkins, Baltimore, pp 163–256
- Maina JN (1998) The lungs of the volant vertebrates birds and bats: how are they relatively structurally optimized for this elite mode of locomotion? In: Weibel ER, Taylor CR, Bolis L (eds) Principles of animal design: the optimization and symmorphosis debate. Cambridge University Press, London, pp 177–185
- Maina JN, King AS (1982a) The thickness of the avian blood-gas barrier: qualitative and quantitative observations. J Anat 134:553-562
- Maina JN, King AS (1982b) Morphometrics of the avian lung. 2. The wild mallard (Anas platyrhynchos) and the greylag goose (Anser anser). Respir Physiol 50:299-313
- Maina JN, King AS (1984) The structural functional correlation in the design of the bat lung. A morphometric study. J Exp Biol 111:43–63
- Maina JN, King AS (1987) A morphometric study of the lung of the Humboldt penguin (Spheniscus humboldti). Zentralbl Veterinaermed Reihe C Anat Histol Embryol 16:293– 297
- Maina JN, King AS (1989) The lung of the emu, *Dromaius novaehollandiae*: a microscopic and morphometric study. J Anat 163:67–74
- Maina JN, Maloiy GMO (1985) The morphometry of the lung of the lungfish (*Protopterus aethiopicus*): its structural-functional correlations. Proc R Soc Lond 244B:399-420
- Maina JN, Maloiy GMO (1986) The morphology of the respiratory organs of the African airbreathing catfish (*Clarias mossambicus*): a light, and electron microscopic study, with morphometric observations. J Zool (Lond) 209:421-445
- Maina JN, Maloiy GMO (1988) A scanning and transmission electron microscopic study of the lung of a caecilian *Boulengerula taitanus*. J Zool (Lond) 215:739–751
- Maina JN, Nicholson T (1982) The morphometric diffusing capacity of a bat *Epomophorus* wahlbergi. J Physiol (Lond) 325:36-37
- Maina JN, Settle JG (1982) Allometric comparison of some morphometric parameters of avian and mammalian lungs. J Physiol (Lond) 330:28P
- Maina JN, Howard CV, Scales L (1981) The determination of the length densities and size distribution of blood and and capillaries in the avian lung involving log normal fitting procedure. Stereol Yugosl 3:673–678
- Maina JN, King AS, King DZ (1982a) A morphometric analysis of the lungs of a species of bat. Respir Physiol 50:1–11
- Maina JN, Abdalla MA, King AS (1982b) Light microscopic morphometry of the lungs of 19 avian species. Acta Anat 112:264–270
- Maina JN, Howard CV, Scales L (1982c) The length densities and size distributions of the air and blood capillaries of the paleopulmo and neopulmo regions of the avian lung. Acta Stereol 2:101–107
- Maina JN, King AS, King DZ (1983) Lung volume-body weight correlation in birds and mammals. Zentralbl Veterinaermed Reihe C Anat Histol Embryol 11:362
- Maina JN, King AS, Settle G (1989a) An allometric study of the pulmonary morphometric parameters in birds, with mammalian comparison. Philos Trans R Soc Lond 326B:1–57
- Maina JN, Maloiy GMO, Warui CN, Njogu EK, Kokwaro ED (1989b) A scanning electron microscope study of the reptilian lungs: the savanna monitor lizard (*Varanus exanthematicus*) and the pancake tortoise (*Malacochersus tornieri*). Anat Rec 224:514–522
- Maina JN, Thomas SP, Dallas DM (1991) A morphometric study of bats of different size: correlations between structure and function of the chiropteran lung. Philos Trans R Soc Lond 333B:31–50

- Maina JN, Maloiy GMO, Makanya AN (1992) Morphology and morphometry of the lungs of two East African mole rats, *Tachyoryctes splendens* and *Heterocephalus glaber* (Mammalia, Rodentia). Zoomorphology 112:167–179
- Maina JN, Kisia SM, Wood CM, Narahara AB, Bergman HL, Laurent P, Walsh PJ (1996a) A comparative allometric study of the morphometry of the gills of an alkalinity-adapted cichlid fish, *Oreochromis alcalicus grahami*, of Lake Magadi, Kenya. Int J Salt Lake Res 5:131–156
- Maina JN, Wood CM, Narahara A, Bergman HL, Laurent P, Walsh P (1996b) Morphology of the swim (air) bladder of a cichlid teleost: *Oreochromis alcalicus grahami* (Trewavas, 1983), a fish adapted to a hyperosmotic, alkaline and hypoxic environment: a brief outline of the structure and function of the swim bladder. In: Dutta HH, Munshi JSD (eds) Fish morphology: horizon of new research. Science Publishers, Lebanon, New Hampshire, pp 179–192
- Maina JN, Maloiy GMO, Wood CM (1998) Respiratory stratagems, mechanisms, and morphology of the "lung" up a tropical swampworm, *Alma emini* Mich. (Oligochaeta: Glossoscolecidae): A transmission and scanning electron microscopic study, with field and laboratory observations. J Zool Lond, in press
- Maitland DP (1986) Crabs that breathe air with their legs Scopimera and Dotilla. Nature (Lond) 319:493-495
- Maitland DP (1987) A highly complex invertebrate lung: the gill chambers of the soldier crab Mictyris longicarpus. Naturwissenschaften 74:293–295
- Maitland DP (1990a) Aerial respiration in the semiphore crab, *Holoecius cordiformis*, with or without branchial water. Comp Biochem Physiol 95A:267–274
- Maitland DP (1990b) Carapace and branchial water circulation, and water-related behaviours in the semiphore crab *Heloecius cordiformis* (Decapoda: Brachyura: Ocypode). Mar Biol 105:275–286
- Makanya AN, Maina JN (1994) Comparative morphology of the gastrointestinal tract of fruit and insect-eating bats. Afr J Ecol 32:158–168
- Makanya AN, Mayhew TM, Maina JN (1995) Morphometry of the gastrointestinal system of insectivorous and frugivorous bats: analysis of an anisotropic tissue. J Anat 187:361–368
- Malan A (1982) Respiration and acid-base state in hibernation. In: Layman CP, Willis JS, Malan A, Wang LCH (eds) Hibernation and torpor in mammals and birds. Academic Press, New York, pp 237–282
- Mallat J, Paulsen C (1986) Gill ultrastructure of the Pacific hagfish *Eptatretus stouti*. Am J Anat 177:243–269
- Maloney JE (1984) The development of the respiratory system in placental mammals. In: Seymour R (ed) Respiration and metabolism in embryonic vertebrates. Dr W Junk, Dordrecht, pp 57-109
- Maloney JE, Rooholamini SA, Wexler L (1970) Pressure-diameter relations of small blood vessels in isolated dog lung. Microvasc Res 2:1-12
- Maloney JE, Drian-Smith C, Takahashi V, Limpus CJ (1990) The environment for development of the embryonic loggerhead turtle (*Caretta caretta*) in Queensland. Copeia 1990:2
- Malvin GM (1988) Microvascular regulation of cutaneous gas exchange in amphibians. Am Zool 28:999–1007
- Malvin GM (1989) Gill structure and function: amphibian larvae. In: Wood SC (ed) Comparative pulmonary physiology: current concepts, vol 39. Lung biology in health and disease. Marcel Dekker, New York, pp 121–151
- Malvin GM, Heisler N (1988) Blood flow patterns in the salamander, *Ambryostoma tigrinum*, before, during and after metamorphosis. J Exp Biol 137:53-74
- Malvin GM, Hlastala MP (1986) Regulation of cutaneous gas exchange by environmental  $O_2$  and  $CO_2$  in the frog. Respir Physiol 65:99–111
- Mandelbrot BB (1977) Form, chance, and dimension. Freeman, New York
- Mandelbrot BB (1983) The fractal geometry of nature. Freeman, New York
- Mangum CP (1963) Oxygen consumption in different species of polchaete worms. Comp Biochem Physiol 10:335–349
- Mangum CP (1964) Activity patterns in metabolism and ecology of polychaetes. Comp Biochem Physiol 11:239–250
- Mangum CP (1976a) Primitive adaptations. In: Newell RC (ed) Adaptations to the environment: essays on the physiology of marine animals. Butterworth, London, pp 191–278

Mangum CP (1976b) The oxygenation of haemoglobin in lugworms. Physiol Zool 49:85–99 Mangum CP (1980a) Respiratory function of the hemocyanins. Am Zool 20:19–38

- Mangum CP (1980b) Distribution of the respiratory pigments and the role of anaerobic metabolism in the lamellibranch molluscs. In: Gilles R (ed) Animals and environmental fitness. Pergamon, Oxford, pp 171–184
- Mangum CP (1982a) The functions of gills in several groups of invertebrate animals. In: Houlihan DF, Rankin JC, Shuttleworth TJ (eds) Gills. Cambridge University Press, Cambridge, pp 77–97
- Mangum CP (1982b) On the relationship between  $P_{50}$  and the mode of gas exchange in tropical crustaceans. Pac Sci 36:403–410
- Mangum CP (1983) Oxygen transport in blood. In: Mantel LH (ed) The physiology of Crustacea, vol 5. Academic Press, London, pp 373–429
- Mangum CP (1985) Oxygen transport in invertebrates. Am J Physiol 248:R505-R514
- Mangum CP (1990) Gas transport in the blood. In: Gilbert DL, Adelman WJ, Arnold JM (eds) Squid as experimental animals. Plenum Press, New York, pp 443-468
- Mangum CP (1992) Physiological adaptation of crustacean hemocyanins: an extended investigation of the blue crab, *Callinectes sapidus*. In: Wood SC, Weber RE, Hargens AR, Millard RW (eds) Physiological adaptations in vertebrates: respiration, circulation, and metabolism. Marcel Dekker, New York, pp 279–293
- Mangum CP (1994) Multiple sites of gas exchange. Am Zool 34:184–193
- Mangum CP, Lykkeboe G (1979) The influence of inorganic ions and pH on oxygenation properties of the blood in the gastropod mollusc *Busycon canaliculatum*. J Exp Zool 207:417–430
- Mangum CP, Lykkeboe G, Johansen K (1975) Oxygen uptake and the role of haemoglobin in the East African swampworm *Alma emini*. Comp Biochem Physiol 52A:477–482
- Manier G, Moinard J, Téchoueyres P, Varène N, Guénard H (1991) Pulmonary diffusion limitation after strenous exercise. Respir Physiol 83:143–154
- Mann KH (1982) Ecology of coastal waters: a system approach. Blackwell, Oxford
- Manning AM, Trotman CNA, Tate WP (1990) Evolution of a polymeric globin in the brine shrimp *Artemia*. Science 248:653-656
- Manwell C (1958) The oxygen respiratory pigment equilibrium of the hemocyanin and myoglobin of the amphineuron mollusc *Cryptochiton stelleri*. J Comp Physiol 52:341–353
- Manwell C (1960) Histological specifity of respiratory pigments. I. Comparisons of the coelom and muscle haemoglobins of the polychaete worm *Travisia pupa* and the echiuroid worm *Arhynchite pugettensis*. Comp Biochem Physiol 1:267–276
- Manwell C (1963) The chemistry and biology of haemoglobin in some marine clams. I. Distribution of the pigment and the properties of the oxygen equilibrium. Comp Biochem Physiol 8:209-218
- Marcus H (1928) Lungenstudien III und IV. Gegenbaurs Morphol Jahrb 59:287-342
- Marcus H (1937) Lungen. In: Bolk L, Goppert E, Kallius E, Lubosch W (eds) Handbuch der vergleichenden Anatomie der Wirbeltiere, III. Urban and Schwarzenberg, Berlin, pp 909–1018
- Marcus E, Marcus C (1960) On Siphonaria hispida. Bull Fac Filos Ciencias e letras, Univ de Sao Paulo 23:107–140
- Marden JH (1987) Maximum lift production during take-off in flying animals. J Exp Biol 130:235-258
- Marden JH, Kramer MG (1994) Surface-skimming stoneflies: a possible intermediate stage in insect flight evolution. Science 266:427-430
- Mardsen ID (1976) Effect of temperature on the microdistribution of the isopod Sphaeroma rugicauda from a salt marsh habitat. Mar Biol 38:117-128
- Margaria R (1976) Biomechanics and energetics of muscular exercise. Clarendon Press, Oxford
- Margaria R, Gualtierotti T, Spinelli D (1958) Protection against acceleration forces in animals by immersion in water. J Aviat Med 29:433–437
- Margaria R, Caproresi E, Aghemo P, Sassi G (1972) The effect of  $O_2$  breathing on maximal aerobic power. Pfluegers Archir Gesamte Physiol Menschen Tiere 336:225–235
- Margulis L (1970) Origin of eukaryotic cells. Yale University Press, New Haven

- Margulis L (1979) Symbiosis and evolution. In: Life origin and evolution: readings from Scientific American, WH Freeman, San Francisco, pp 101–110
- Margulis L (1981) Symbiosis in cell evolution. WH Freeman, New York
- Marin-Girón F, Cedres T, Otero A (1975) Contribución al estudio microscópico de la histología de los sacos aéreos. Bol R Soc Española Hist Nat (Biol) 73:275–296
- Marsh BA, Branch GM (1979) Circadian and circatidal rhythms of oxygen consumption in sandy-beach isopod *Tylos granulatus* Kraus. J Exp Mar Biol Ecol 37:77–89
- Marshall C, Schultze HP (1992) Relative importance of molecular, neontological, and paleontological data in understanding the biology of the vertebrate invasion of land. J Mol Evol 35:93– 101
- Marshall DJ, McQuaid CD (1992) Comparative aerial metabolism and water relations of the intertidal limpets, *Patella granularis* L. (Mollusca: Prosobranchia) and *Siphonaria oculus* Kr. (Mollusca: Pulmonata. Physiol Zool 65:1040–1056
- Marshall NB (1960) Swim bladder structure of deep sea fishes in relation to their systematics and biology. Discovery Rep 31:1–122
- Martin AP, Palumbi SR (1993) Body size, metabolic rate, generation time, and the molecular clock. Proc Natl Acad Sci USA 90:4097–4091
- Martin KM, Hutchison VH (1979) Ventilatory activity in Amphiuma tridactylum and Siren lacertina (Amphibia, Caudata). J Herpatol 13:427-434
- Martin TE (1996) Fitness cost of resource overlap among coexisting bird species. Nature (Lond) 380:338–340
- Masahiko K, Paul JL, Thurlbeck WM (1984) The effect of age on lung structure in male BALB/ cNNia. Am J Anat 170:1–21
- Mason DK, Collins AE, Watkins KL (1983) Exercise induced pulmonary haemorrhage in horses. In: Snow DH, Persson SGB, Rose RJ (eds) Equine exercise physiology. Granta Editions, Cambridge, pp 57–63
- Mass JA (1939) Über die Atmung von Helix pomatia, Z Vergl Physiol 26:605-610
- Massabuau JC, Dejours P, Sakakibara Y (1984) Ventilatory CO<sub>2</sub> drive in the crayfish: influence of oxygen consumption level and water oxygenation. J Comp Physiol 154B:65–72
- Massabuau JC, Burtin B, Wheatly M (1991) How is O<sub>2</sub> consumption maintained independent of ambient oxygen in mussel *Anodonta cygnea*? Respir Physiol 83:103–114
- Matalon S, Egan EA (1981) Effects of 100% oxygen breathing on permeability of alveolar epithelium to solute. J Appl Physiol 50:859–863
- Mathieu-Costello O (1990) Histology of flight: tissue and muscle gas exchange. In: Sutton JR, Coates G, Remmers JE (eds) Hypoxia: the adaptations. BC Dekker, Toronto, pp 13–19
- Mathieu-Costello O, Szewczak JM, Logemann RB, Agey PJ (1992) Geometry of blood-tissue exchange in bat flight muscle compared with bat hindlimb and rat soleus muscle. Am J Physiol 262:R955-R965
- Matsui T, Abe Y (1986) Evolution of an impact-induced atmosphere and magma ocean on the accreting Earth. Nature (Lond) 319:303-305
- Matsumura H, Setoguti T (1984) Electron microscopic studies of the lung of the salamander, *Hynobius nebulosus*. I. A scanning and transmission microscopic observation. Okajimas Folia Anat Jpn 61:15–25
- Matthews WH, Balzer RH, Shelburne JD, Pratt PC, Kylstra JA (1978) Steady-state gas exchange in normothermic, anaethetized, liquid-ventilated dogs. Undersea Biomed Res 5:341–354
- Maury W, Potts BJ, Rabson AB (1989) HIV 1 infection of first trimester and term human placental tissue: a possible mode of maternal-foetal transmission. J Infect Dis 160:15-23
- Maxwell MH, Robertson GW, Spence S (1986a) Studies on an ascitic syndrome in young broilers. I. Haematology and pathology. Avian Pathol 1986:15:511–524
- Maxwell MH, Robertson GW, Spence S (1986b) Studies on an ascitic syndrome in young broilers. II. Ultrastructure. Avian Pathol 1986:15:525-538
- May EB (1973) Extensive oxygen depletion in Mobile Bay, Alabama. Limnol Oceanogr 18:353– 366
- May RM (1981) The evolution of cooperation. Nature (Lond) 292:291-292
- May RM (1988) How many species are there on Earth? Science 241:1441-1449
- May RM (1990) How many species? Philos Trans R Soc Lond 330B:293-316
- May RM (1992) How many species inhabit Earth? Sci Am October Issue:18-24

Mayhew TM (1991) Scaling placental oxygen diffusion to birthweight: studies on placentae from low- and high-altitude pregnancies. J Anat 175:187–194

Mayhew TM (1992) The structural basis of oxygen diffusion in the human placenta. In: Egginton S, Ross HF (eds) Oxygen transport in biological tissue systems: modelling pathways from environment to cell. Cambridge University Press, Cambridge, pp 79–101

Mayhew TM, Simpson RA (1994) Quantitative evidence for the spatial dispersal of trophoblast nuclei in human placental villi during gestation. Placenta 15:837–844

Mayhew TM, Wadrop E (1994) Placental morphogenesis and the star volumes of villous trees and intervillous pores. Placenta 15:209-217

Mayhew TM, Joy CF, Haas JD (1984) Structure-function correlation in the human placenta: the morphometric diffusing capacity for oxygen at term. J Anat 139:691–708

Mayhew TM, Jackson MR, Haas JD (1986) Microscopical morphology of the human placenta and its effects on oxygen diffusion: a morphometric model. Placenta 7:121-131

Mayhew TM, Jackson MR, Haas JD (1990) Oxygen diffusive conductances of human placentae from term pregnancies at low and high altitudes. Placenta 11:493–503

Maynard-Smith J (1968) Mathematical ideas in biology. Cambridge University Press, Cambridge

Maynard-Smith J (1978) Optimization theory in evolution. Annu Rev Ecol Syst 9:31-56

Maynard-Smith J (1996) The games lizards play. Nature (Lond) 380:198-199

Maynard-Smith J, Szathmary E (1996) On the likelihood of habitable worlds. Nature (Lond) 384:107

Mayr E (1942) Systematics and the origin of species from the viewpoint of a zoologist. Columbia University Press, New York

Mayr E (1960) The emergence of evolutionary novelties. In: Tax S (ed) Evolution after Darwin, vol 2. University of Chicago Press, Chicago, pp 349–380

McClanahan LL, Rodolfo R, Shoemaker VH (1994) Frogs and toads in deserts. Sci Am 273:82-88

McClary A (1964) Surface inspiration and cilliary feeding in *Pomacea paludosa* (Prosobranchia: Mesogastropoda: Ampullaridae). Malacologia 2:87–104

McCord JM (1988) Free radicals and myocardial ischaemia: overview and outlook. Free Radic Biol Med 4:9–14

McCutcheon FH (1936) Haemoglobin function during the life history of the bullfrog. J Cellular Comp Physiol 8:63–81

McCutcheon FH (1954) Phylogenetic aspects of respiratory function. Evolution 8:181–191

McCutcheon FH (1964) Organ systems in adaptation: the respiratory system. In: Dill DB, Adolph EF, Wilber CG (eds) Handbook of physiology, sect 4, adaptation to the environment. American Physiological Society, Washington, DC, pp 167–191

McDonald DG, McMahon BR, Wood CM (1977) Patterns of heart and scaphognathite activity in the crab *Cancer magister*. J Exp Zool 202:33–44

McDonald DG, Caudek V, Ellis R (1991) Gill design in fresh water fishes: interrelationships among gas exchange, ion regulation, and acid base regulation. Physiol Zool 64:103-123

McDougall JDB, McCobe M (1967) Diffusion coefficent of oxygen through tissues. Nature (Lond) 215:1173-1174

McElhinny MW, Taylor SR, Stevenson DJ (1978) Limits to the expansion of the Earth, Moon, Mars and Mercury and to changes in the gravitational constant. Nature (Lond) 271:316– 321

McElroy MB (1983) Marine biological controls on atmospheric CO<sub>2</sub> and climate. Nature (Lond) 302:328–329

McFall-Ngai MJ, Ruby EG (1991) Symbiont recognition and subsequent morphology as early events in animal-bacterial mutualism. Science 254:1491–1494

McFarland RA, Evans JN (1939) Alterations in dark adaptation under reduced oxygen tensions. Am J Physiol 127:37–50

McGhee GR (1989) Frasian-Famennian extinction event. In: Briggs DEG, Crowther PR (eds) Paleobiology – a synthesis. Blackwell, Oxford, pp 97–176

McGinitie GE (1939) The method of feeding in Chaetopterus. Biol Bull 77:115-118

McGrath MW, Thomson ML (1959) The use of helium and sulphur hexafluoride for assessing diffusive mixing in the lung. J Physiol (Lond) 148:72-73

- McGregor LK, Daniels CB, Nicholas TE (1993) Lung ultrastructure and surfactant-like system of the central Australian netted dragon, *Ctenophoruos nuchalis*. Copeia 1993:326–333
- McKay C (1997) Organic synthesis in experimental impact shocks. Science 276:390-392
- McKay CP, Pollack JB, Courtin R (1991) The greenhouse and antigreenhouse effects on Titan. Science 253:1118-1121
- McKay DS, Gibson EK, Thomas-Kerpta KL, Vali H, Romanek CS et al. (1996) Search for pas tlife on Mars: possible relic biogenic activity in martian meteorite ALH 84001. Science 273:924– 930
- McLaughlin PA (1983) Internal anatomy. In: Mantel LH (ed) The biology of Crustacea, vol 5. Academic Press, New York, pp 1–53
- McLean DM (1978) Land floras: the major late Proterozoic atmospheric carbon dioxide/oxygen control. Science 200:1060–1062
- McLelland J (1989) Larynx and trachea. In: King AS, McLelland J (eds) Form and function in birds, vol 4. Academic Press, London, pp 69–103
- McMahon BR (1969) A functional anlysis of the aquatic and aerial respiratory movements of an African lungfish, *Protopterus aethiopicus*, with refrence to the evolution of the lung ventilation mechanism in vertebrates. J Exp Biol 51:407–430
- McMahon BR (1988) Physiological responses to periodic emergency in intertidal molluscs. Am Zool 28:97–114
- McMahon BR, Burggren WW (1979) Respiration and adaptation to the terrestrial habitat in the land hermit crab, *Coenobita clypeatus*. J Exp Biol 79:265–281
- McMahon BR, Burggren WW (1987) Respiratory physiology of intestinal air breathing in the teleost fish, *Misgurnus anguillicaudatus*. J Exp Biol 133:371–393
- McMahon BR, Burggren WW (1988) Respiration. In: Burggren WW, McMahon BR (eds) Biology of the land crabs. Cambridge University Press, Cambridge, pp 249–297
- McMahon BR, Wilkens JL (1977) Periodic respiratory and circulatory performance in the red rock crab, *Cancer productus*. J Exp Zool 202:363–374
- McMahon BR, Wilkens JL (1983) Ventilation, perfusion and oxygen uptake. In: Mantel LH (ed) The physiology of Crustacea, vol 5. Academic Press, London, pp 289-372
- McMahon BR, Wilkes PRH (1983) Emergence response and aerial ventilation in normoxic and hypoxic crayfish, Orconectes rusticus. Physiol Zool 51:133-141
- McMahon RF (1983) Physiological ecology of fresh water pulmonates. In: Russell-Hunter WD (ed) The Mollusca, vol 6. Academic Press, Orlando, pp 359–430
- McMahon RF (1985) Functions and functioning of crustacean hemocyanin. In: Lamy J, Truchot JP, Gilles R (eds) Respiratory pigments in animals. Springer, Berlin Heidelberg New York, pp 35–58
- McMahon TA, Bonner JT (1983) On size and life. Scientific American Library, New York
- McNab B (1966) The metabolism of fossorial rodents: a study of convergence. Ecology 47:712–733
- McNamara JM, Houston AI (1996) State-dependent life histories. Nature (Lond) 380:215-221

McShea DW (1991) Complexity and evolution: what everbody knows. Biol Philos 6:303-324

- Mead J, Turner JM, Macklem PT, Little JB (1967) Significance of the relationship between lung recoil and maximum expiratory flow. J Appl Physiol 22:95–108
- Meban C (1977) Ultrastructure of the respiratory epithelium in the lungs of the newt *Triturus* cristatus. Acta Zool Stockh 58:151–167
- Meban C (1978a) Functional anatomy of the lungs of the green turtle, *Lacerta viridis*. J Anat 125:421-436
- Meban C (1978b) The respiratory epithelium in the lungs of the slow-worm, *Anguilis fragilis*. Cell Tissue Res 190:337–354
- Meban C (1979) An electron microscopy study of the respiratory epithelium in the lungs of the fire salamander (*Salamandra salamandra*). J Anat 128:215–221
- Meban C (1980) Thicknesses of the air-blood barriers in vertebrate lungs. J Anat 131:299–307 Meduna JL (1950) Carbon dioxide therapy. CC Thomas, Springfield, Illinois
- Meilin S, Rogatsky GG, Thoms R, Zarchin N et al. (1996) Effect of carbon monoxide on brain may be mediated by nitric oxide. J Appl Physiol 81:1078-1083
- Melchior FM, Srinivasan RS, Charles JB (1992) Mathematical modeling of human cardiovascular system for simulation of orthostatic response. Am J Physiol 262:H1920–H1933

- Melmon KL, Cline MJ, Hughes T, Nies AS (1968) Kinins; possible mediators of neonatal circulatory changes in man. J Clin Invest 47:1279–1302
- Melsom MN, Johansen JK, Flatebø T, Müller C (1997) Distribution of pulmonary ventilation and perfusion measured simultaneously in awake goats. Acta Physiol Scand 159:199–220
- Meng H, Bentley TB, Pittman RN (1992) Oxygen diffusion in hamster striated muscle: comparison of in vitro and near in vitro conditions. Am J Physiol 263:H35–H39
- Menon JG, Arp AJ (1992a) Morphological adaptations of the respiratory hind gut of a marine echiurian worm. J Morphol 214:131-138
- Menon JG, Arp AJ (1992b) Symbiotic bacteria may prevent sulfide poisoning of the body wall of Urechis caupo. Am Zool 32:59A
- Meredith JL (ed) (1985) Hydrothermal vents of the eastern Pacific: An overview. Bull Biol Soc Washington, December Issue
- Merrill EW (1969) Rheology of blood. Physiol Rev 19:863-888
- Mertens R (1960) The world of amphibians and reptiles. McGraw-Hill, New York
- Mertens S, Noll T, Spahr R, Krützfeldt A, Piper HM (1990) Energetic response of coronary endothelial cells to hypoxia. Am J Physiol 258:H689-H694
- Metcalfe J, Stock MK (1993) Oxygen uptake in the chorioallantoic membrane, avian homologue of the mammalian placenta. Placenta 14:605–614
- Metcalfe JD, Butler PJ (1984) On the nervous regulation of gill blood flow in the dogfish (Scyliorhinus canicula). J Exp Biol 113:253-267
- Metcalfe JD, Butler PJ (1986) The functional anatomy of the gills of the dogfish (*Scyliorhinus canicula*). J Zool (Lond) 208:519-630
- Metcalfe J, Meschia G, Hellegers A, Prystowsky H, Huckabee W, Barron DH (1962) Observations on the placental exchange of the respiratory gases in pregnant ewe at high altitude. Q J Exp Physiol 47:74–92
- Metcalfe J, Bartels H, Moll W (1967) Gas exchange in the pregnant uterus. Physiol Rev 47:782– 838
- Metcalfe J, Bissonnette JM, Bowles RE, Matsumoto JA, Dunham SJ (1979) Hen's eggs with retarded gas exchange. I. Chorioallantoic capillary growth. Respir Physiol 36:97-108
- Metcalfe R (1967) The oxygen supply of the foetus. In: Reuck AVS, Porter R (eds) Development of the lung. Churchill, London, pp 37–63
- Meyer A, Dolven SI (1992) Molecules, fossils, and the origin of tetrapods. J Mol Evol 35:102-113
- Meyer M, Worth H, Scheid P (1976) Gas-blood CO<sub>2</sub> equilibration in parabronchial lungs of birds. J App. Physiol 41:302–309
- Meyer WV (1988) The role of form and function in the collegiate biology curriculum. Am Zool 28:619–664
- Michels DB, West JB (1978) Distribution of pulmonary ventilation and perfusion during short periods of weightlessness. J Appl Physiol 45:987–998
- Michels MD, Friedman PJ, West JB (1979) Radiographic comparison of human lung shape during normal gravity and weightlessness. J Appl Physiol 47:851-885
- Midtgärd U (1983) Scaling of the brain and eye cooling system in birds: a morphometric analysis of the rete ophthalmicum. J Exp Zool 225:197–207
- Midtgärd U (1984) Blood vessels and the occurrence of arteriovenous anastomoses in the cephalic heat loss areas of the mallards, *Anas platyhynchos* (Aves). Zoomorphology 104:323–335
- Milani A (1894) Beiträge zur Kenntnis der Reptilienlunge. II. Zool Jahrb Abt Anat Ontog Tiere 7:545–592
- Milburn TR, Beadle LC (1960) The determination of total carbon dioxide. J Exp Biol 37:444-460
- Milhorn HT, Benton R, Ross R, Guyton AC (1965) A mathematical model of the human respiratory control system. Biophys J 5:27-46
- Milic-Emili J (1991) Work of breathing. In: West JB, Crystal RG (eds) The lung: scientific foundations. Raven Press, New York, pp 1065–1075
- Milic-Emili J, Henderson JAM, Kaneko K (1966) Regional distribution of inspired gas in the lung. J Appl Physiol 21:749-759
- Miller AH (1949) Some ecological and morphologic considerations in the evolution of higher taxonomic categories. In: Mayr E, Schuz E (eds) Ornithologie als biologische Wissenschaft. Carl Winter Universitätsverlag, Heidelberg, pp 84–88

- Miller K, Camilliere JJ (1981) Physical training improves swimming performance of the African clawed frog, *Xenopus laevis*. Herpetologica 37:1–10
- Miller MA (1948) Seasonal trends in burrowing of pocket gophers (*Thomomys*). J Mammal 29:38-44
- Miller PL (1960) Respiration in the desert locust. III. Ventilation and spiracles during flight. J Exp Biol 37:264–278
- Miller PL (1966) The supply of oxygen to the active flight muscles of some large beetles. J Exp Biol 45:285-304
- Miller PL (1974) Respiration aerial gas transport. In: Rockstein M (ed) The physiology of insects, 2nd edn. Academic Press, New York, pp 346-402
- Miller SL, Orgel LE (1974) The origins of life on Earth. Prentice-Hall, Englewood Cliffs, New Jersey
- Mills RM, Brinster RL (1967) Oxygen consumption of preimplantation mouse embryos. Exp Cell Res 47:337–344
- Milner AR (1988) The relationships and origins of living amphibians. In: Benton MJ (ed) The phylogeny and classification of tetrapods, vol 1, Amphibians, reptiles and birds. Clarendon Press, Oxford, pp 59–102
- Milner WR (1980) Pulmonary circulation. In: Mountcastle VB (ed) Medical physiology, vol 2. Mosby, St Louis, pp 1108–1117
- Milnor WR (1982) Hemodynamics. Williams and Williams, Baltimore
- Milsom WK (1984) The interrelationship between pulmonary mechanics and spontaneous breathing pattern in the Tokay lizard *Gekko gecko*. J Exp Biol 113:203–214
- Milsom WK (1988) Control of arrythmic breathing in aerial breathers. Can J Zool 66:99-108
- Milsom WK (1989) Mechanisms of ventilation in lower vertebrates: adaptations to respiratory and non-respiratory constraints. Can J Zool 67:2943–2963
- Milsom WK (1990) Control and co-ordination of gas exchange in air breathers. In: Boutilier RG (ed) Advances in comparative environmnetal physiology, vol 6. Vertebrate gas exchange from environment to cell. Springer, Berlin Heidelberg New York, pp 374–400
- Milsom WK (1991) Intermittent breathing in vertebrates. Annu Rev Physiol 53:87-105
- Milsom WK, Johansen K (1975) The effect of buoyancy-induced lung volume changes on respiratory frequency in a chelonian (*Caretta caretta*). J Comp Physiol 98:157–160
- Milsom WK, Jones DR (1979) The role of pulmonary afferent information and hypercapnia in the control of the breathing pattern in chelonia. Respir Physiol 37:101–107
- Milsom WK, Jones DR (1985) Characteristics of mechanoreceptors in the air breathing organ of the holostean fish, *Amia calva*. J Exp Biol 117:389–399
- Milton P (1971) Oxygen consumption and osmoregulation in the shanny, *Blennius pholis*. J Mar Biol Assoc UK 51:247–265
- Minkoff EE (1983) Evolutionary biology. Addison-Wesley, Reading, Massachusetts
- Misiek L, Szaski H (1978) Dimensions of cells in some tissues of six amphibian species. Acta Biol Cracov Ser Zool 21:127–136
- Mitchell E, Reeves RR (1981) Catch history and cumulative catch estimates of initial population size of cetaceans in the eastern Canadian Arctic. Rep Int Whale Comm 31:645–682
- Mitchell GS, Gleeson TT, Bennett AF (1981) Pulmonary oxygen transport during activity in lizards. Respir Physiol 43:365–375
- Mitchell HA (1964) Investigation of the cave atmosphere of a Mexican bat colony. J Mammal 45:568–577
- Mix AC (1989) Influence of productivity variations on long-term atmospheric CO<sub>2</sub>. Nature (Lond) 337:541–544
- Moalli R, Meyers RS, Jackson DC, Millard RW (1980) Skin circulation of the frog, Rana catesbeiana: distribution and dynamics. Respir Physiol 40:137-148
- Modell JH, Gollan F, Giammona ST, Parker D (1970) Effect of fluorocarbon liquid on surface tension properties of pulmonary surfactant. Chest 57:263–265
- Moessinger AC, Harding R, Adamson TM, Singh M, Kiu GT (1990) Role of lung fluid volume in growth and maturation of the foetal sheep lung. J Clin Invest 86:1270–1277
- Mojzsis SJ, Arrhenius G, McKeegan KD, Harrison TM, Nutman AP, Friends CRL (1996) Evidence of life on Earth before 3800 million years ago. Nature (Lond) 384:55–59
- Moll W (1966) The diffusion coefficient of haemoglobin. Respir Physiol 1:357-365

- Mommsen TP, Ballantyne J, MacDonald D, Gosline J, Hochachka PW (1981) Analogous of red and white muscle in squid mantle. Proc Natl Acad Sci USA 78:3274–3278
- Moncada S, Palmer RMJ, Higgs EA (1991) Nitric oxide: physiology, pathophysiology, and pharmacology. Pharmacol Rev 43:109–142
- Monge MC, Monge CC (1968) Adaptation to high altitude. In: Hafez ESE (ed) Adaptation of domestic animals. Lea and Febiger, Philadelphia, pp 194–201
- Moore JA (1990a) Science as a way of knowing form VII: a conceptual framework for biology part III. Am Zool 30:1–123
- Moore JA (1990b) The ability to live on dry land, rather than in water, required major adjustments in structure and physiology. Am Zool 30:847–849
- Moore LD, Jahnigen D, Rounds SS, Reeves JT, Grover RF (1982) Maternal hyperventilation helps preserve arterial oxygenation during high altitude pregnancy. J Appl Physiol 52:690–694
- Moore SJ (1976) Some spider organs as seen by the scanning electron microscope, with special reference to the booklung. Bull Br Arachnol Soc 3:177–187
- Morgan N (1995) Chemistry in action: the molecules of everyday life. Oxford University Press, New York
- Morin FC, Egan EA (1992) Pulmonary hemodynamics in fetal lambs duuring development at normal and increased oxygen tension. J Appl Physiol 73:213–218
- Moritz AR (1944) Chemical methods for the determination of drowning. Physiol Rev 24:70-88
- Morkin E, Collins JA, Goldman HS, Fishman P (1965) Pattern of blood flow in the pulmonary veins of the dog. J Appl Physiol 20:1118–1128
- Morony JJ, Bock WJ, Farrand J (1975) Reference list of the birds of the world. Department of Ornithology, American Museum of Natural History, New York
- Morris S (1991) Respiratory gas exchange and transport in crustaceans: ecological determinats. Mem Queense Mus 31:241–261
- Morris S, Bridges CR (1986) Novel non-lactate cofactors of hemocyanin oxygen affinity in crustacean. In: Linzen B (ed) Invertebrate oxygen carriers. Springer Berlin Heidelberg New York, pp 353–356
- Morris S, Bridges CR (1994) Properties of respiratory pigments in bimodal breathing animals: air- and water-breathing by fish and crustaceans. Am Zool 34:216–228
- Morris S, Greenaway P (1990) Adaptations to a terrestrial existence in the robber crab, *Birgus latro* L.V. Preliminary investigations of carbonic anhydrase activities. J comp Physiol 160B:217-221
- Morris S, Taylor AC (1985) The respiratory response of the intertidal prawn *Palaemon elegans* (Rathke) to hypoxia and hyperoxia. Comp Biochem Physiol 81A:633–639
- Morrison DW (1980) Efficiency of food utilization by fruit bats. Oecologia 45:270-273
- Morrison PR, Ryser FA, Dawe AR (1959) Studies on the physiology of the masked shrew, *Sorex cinereus*. Physiol Zool 32:256–271
- Mortola JP (1987) Dynamics of breathing in newborn mammals. Physiol Rev 67:187-243
- Morton JE (1979) Molluscs. Hutchison, London
- Moshiri GA, Goldman CR, Godshalk GL, Mull DR (1970) The effects of variations in oxygen tension on certain aspects of respiratory metabolism in *Pacifastacus leniusculus* (Dana) (Crustacea: Decapoda). Physiol Zool 43:23–29
- Moss ML (1962) The functional matrix. In: Kraus B, Reidel R (eds) Vistas in orthodontics. Lea and Febiger, Philadelphia, pp 85–98
- Mossiman HW (1987) Vertebrate foetal membranes. Rutgers University Press, New Brunswick, New Jersey
- Muir BS, Buckley RM (1967) Gill ventilation in Remora remora. Copeia 1967:581-586
- Muir BS, Kendall JI (1968) Structural modifications in the gills of tunas and some other oceanic fishes. Copeia 1968:388–398
- Mukai H, Koike I (1984) Behaviour and respiration of burrowing shrimps Upogebia major (de Haan) and Callianassa japonica (de Haan). J Crust Biol 4:191-200
- Müller W (1950) Die Mündung des Luftganges beim Messerfisch und ihre konstruktive Gestaltung. Zool Anz Ergänz 145:635–642
- Munro HN, Downie ED (1964) Relationship of liver composition to intensity of protein metabolism in different mammals. Nature (Lond) 203:603–604

- Munshi JSD (1968) The accessory respiratory organs of *Anabis testudineus* (Bloch) (Anabantidae, Pisces). Proc Linn Soc Lond 179:107–126
- Munshi JSD (1976) Gross and fine structure of the respiratory organs of air-breathing fishes. In: Hughes GM (ed) Respiration of amphibious vertebrates. Academic Press, London, pp 73–102
- Munshi JSD, Hughes GM (1986) Scanning electron microscopy of the respiratory organs of juvenile and adult climbing perch, *Anabas testudineus*. Jpn J Ichthyol 33:39–45
- Munshi JSD, Hughes GM (1991) Structure of the respiratory islets of accessory respiratory organs and their relationship with the gills in the climbing perch. Anabas testudineus (Teleostei, Perciformes). J Morphol 209:241-256
- Munshi JSD, Hughes GM (1992) Air breathing fishes of India: their structure, function and life history. AA Balkema Uitgevers BV, Rotterdam
- Munshi JSD, Singh BN (1968) On the respiratory organs of Aphipnous cuchia (Ham. Buch). J Morphol 124:423-444
- Munshi JSD, Srivastava MP (1988) Natural history of fishes and systematics of fresh water fishes of India. Narendra Publishing House, New Delhi
- Munshi JSD, Olson KR, Ojha J, Ghosh TK (1986a) Morphology and vascular anatomy of the accessory respiratory organs of air-breathing climbing perch, *Anabas testudineus* (Bloch). Am J Anat 176:321–331
- Munshi JSD, Weibel ER, Gehr P, Hughes GM (1986b) Structure of the respiratory air sac of *Heteropneustes fossilis* (Bloch) (Heteropneustidae, Pisces) an electron microscope study. Proc Indian Nat Sci Acad 52B:703–713
- Munshi JSD, Hughes GM, Gehr P, Weibel ER (1989) Structure of the air-breathing organs of the swamp mud eel, *Monopterus cuchia*. Jpn J Ichthyol 35:453–465
- Munshi JSD, Olson KR, Ghosh TK(1990) Vasculature of the head and respiratory organs in an obligate air-breathing fish, the swamp eel *Monopterus* (=*Amphipnous*) *cuchia*. J Morphol 203:181-201
- Munshi JSD, Roy PK, Ghosh TK, Olson KR (1994) Cephalic circulation in the air-breathing snakehead fish, *Channa punctata*, *C. gashua*, and *C. marulius* (Ophiocephalidae, Ophiocephaliformes). Anat Rec 238:77-91
- Muratori RA, Falugi C, Colosi R (1976) Osservazioni su alcuni aspetti dell'anatomia del *Telmatobius culeus* (Gorman 1875) visti come addattamento al particolare ambiente del lago Titicaca. Atti Accad Naz Lincei 61:508–519
- Murdock GR, Currey JD (1978) Strength and design of the two ecologically distinct barnacles, Balanus balanus and Semibalanus balanoides. Biol Bull 155:169-192
- Murray JW, Barber RT, Roman MR, Bacon MD, Feely RA (1994) Physical and biological controls on carbon cycling in the equatorial Pacific Ocean. Science 266:58–65
- Murry CD (1926) The physiological principle of minimum work. I. The vascular system and the cost of blood volume. Proc Natl Acad Sic USA 12:207–214
- Musacchia XK, Volkert WA (1971) Blood gases in hibernating and active ground squirrels: HbO<sub>2</sub> affinity at 6 and 38 °C. Am J Physiol 221:128–130
- Mutsaddi KB, Bal DV (1969) Some observations on habits and habitat of *Boeophthalmus* dussumierei (Cuv. and Val.). J Univ Bombay 65:33-41
- Myers AC (1972) Tube worm sediment relationships of *Diopatra cuprea* (Polychaeta: Onuphidae). Mar Biol 17:350–356
- Nachtigall W (1991) Flight. In: Witt R, Lieckfeld CP (eds) Bionics: nature's patents. Pro Futura, Munich, pp 7–31
- Nagaishi Ch, Okada Y, Ishiko S, Daido S (1964) Electron microscopic observations of the pulmonary alveoli. Exp Med Surg 22:81–97
- Nagel E (1961) The structure of science. Routledge and Kegan Paul, London
- Nagy JA, Odell DK, Seymour RS (1972) Temperature regulation by the inflorescence of Philodendron. Science 178:1195–1197
- Nahas GG (1962) The pharmacology of *tris* (hydroxymethyl) methane (THAM). Pharmacol Rev 14:447–472
- Naito H, Gillis CN (1973) Effects of halothane and nitrous oxide on removal of norepinephrine from the pulmonary circulation. Anaesthesiology 30:575–580
- Nakao T (1974) The fine structure and innervation of gill lamellae in Anodonta. Cell Tissue Res 157:239–254

Narahara AB, Bergman HL, Maina JN, Lauren P, Walsh PJ, Wood CM (1996) Respiratory physiology of the Lake Magadi Tilapia (*Oreochromis alcalicus grahami*), a fish adapted to a hot, alkaline, and frequently hypoxic environment. Physiol Zool. 69(5):1114-1136

Nassar SAK, Munshi JSD (1971) Studies on the macrophytic biomass production and fish population in an abandoned pond at Bhagalpur, Bihar. J Sci Bihar University, India IV:8-16

Nathan C (1992) Nitric oxide as a secretory product of mammalian cells. FASEB J 6:3051–3064 Neckvasil NP, Olson KR (1986) Extraction and metabolism of circulating catecholamines by the

trout gill. Am J Physiol 19:R5276–5287

Neftel A, Oeschger H, Schwander J, Stauffer B, Zunbrunn R (1982) Ice core sample meaurements give atmospheric CO, content during the past 40000 yrs. Nature (Lond) 295:220–223

Nelson G (1978) Ontogeny, phylogeny, paleontology and biogenic law. Syst Zool 27:324–345 Nelson JS (1976) Fishes of the world. Wiley, New York

Nelson TR, Manchester DK (1988) Modeling of lung morphogenesis using fractal geometries. IEEE Trans Med Imaging 7:321-327

Nelson TR, West BJ, Goldberger AL (1990) The fractal lung: universal and species related scaling patterns. Experientia 46:251–254

Nelson ZC, Hirshfield D, Schreiweis DO, O'Farrell MJ (1977) Flight muscle contraction in relation to ambient temperature in some species of desert bats. Comp Biochem Physiol 56A:31-36

Nevo E (1979) Adaptive convergence and divergence of subterranean mammals. Annu Rev Ecol Syst 10:269–308

Nevo E, Guttman R, Haber M, Erez E (1982) Activity patterns of evolving mole rats. J Mammal 63:453–463

Newell RC, Courtney WAM (1965) Respiratory movements in *Holothuria forskali*. J Exp Biol 42:45-57

Newman RA (1992) Adaptive plasticity in amphibian metamorphosis. BioScience 42:671-678

Newman MJ, Rood RT (1977) Implications of solar evolution for Earth's atmosphere. Science 198:1035–1037

Nguyen Phu D, Yamaguchi K, Scheid P, Piiper J (1986) Kinetics of oxygen uptake and release by erythrocytes of the chicken. J Exp Biol 125:15–27

Nicloux M (1923) Action de l'oxyde de carbone sur les poissons et capacité respiratoire du sang de ces animaux. Cr Blanc Soc Biol 89:1328–1331

Nicol JAC (1960) The biology of marine animals. Interscience, New York

Nicolet J, Bannerman ESN, Haller R (1975) Mycotic proteolytic enzymes. In: Junod AF, Haller R (eds) Lung metabolism. Academic Press, New York, pp 57–65

Nicoll PA (1954) The anatomy and behaviour of the vascular systems in *Nereis virens* and *Nereis limbata*. Biol Bull Mar Lab Woods Hole 106:69–82

Nieden F (1913) Gymnophiona (Amphibia: Apoda). Tierreich 37:1-31

Nielsen EG, Gargas E (1984) Oxygen nutrients and primary production in the open Danish waters. Limnologica 15:303-310

Nikinmaa M (1990) Vertebrate erythrocytes: adaptation of function to respiratory requirements. Springer, Berlin Heidelberg New York

Nikinmaa M, Huestis WH (1984) Shape changes in goose erythrocytes. Biochim Biophys Acta 773:317-320

Niles E (1993) History, function, and evolutionary biology. Evol Biol 27:33-50

Nilsson GR, Löfman CO, Block M (1995) Extensive erythrocytes deformation in fish gills observed by in vivo microscopy: apparent adaptations for enhancing oxygen uptake. J Exp Biol 198:1151–1156

Nilsson S (1985) Filament position in fish gills is influenced by a smooth muscle enervated by adrenergic nerves. J Exp Biol 118:433-437

Nilsson S (1986) Control of gill blood flow. In: Nilsson S, Holmgren S (eds) Fish physiology: recent advances. Croom Helm, Dover HH, pp 86–101

Nisbet EG (1988) The young Earth. Allen and Unwin, London

Nisbet UG, Cann JR, Dover van CL (1995) Origins of photosynthesis. Nature (Lond) 373:479-480

Noble GK (1925) Integumentary, pulmonary, and cardiac modifications correlated with increased cutaneous respiration in the amphibian: a solution to the "hairy frog" problem, J Morphol Physiol 40:341-416

- Noble GK (1929) The adaptive modifications of the arboreal tadpoles of *Hoplophryne* and torrent tadpoles of *Staurois*. Bull Am Mus Nat Hist 58:291–334
- Noble GK (1931) The biology of Amphibia. McGraw-Hill, New York
- Nonnenmacher TF (1987) A scaling model for dichotomous branching processes. Biol Cybern 56:155–157
- Nonnenmacher TF (1988) Fractal shapes of cell membranes and pattern formation by dichotomous branching processes. In: Lamprecht I, Zotin Al (eds) Thermodynamics and pattern formation in biology. Walter de Gruyter, Berlin, pp 371-394
- Nonnenmacher TF (1989) Fractal scaling mechanisms in biomembranes: oscillations in the lateral diffusion coefficient. Eur Biophys J 16:375-379
- Norberg UM (1976a) Aerodynamics of hovering flight in long-eared bat *Plecotus auritus*. J Exp Biol 65:459–470
- Norberg UM (1976b) Kinematics, aerodynamics, and energetics of horizontal flapping flight in the long-eared bat *Plecotus auritus*. J Exp Biol 65:179–212
- Norberg UM (1981) Why foraging birds in trees should climb and hop upwards rather than downwards. Ibis 123:281-288
- Norberg UM (1985) Flying, gliding, and soaring. In: Hildebrand M (ed) Functional vertebrate morphology. Harvard University Press, Cambridge, pp 366–377
- Norberg UM (1986) On the evolution of flight and wing forms in bats. In: Nachtigall W (ed) Bat flight. BIONA report 5, Gustav Fischer, Stuttgart, pp 13–26
- Norberg UM (1990) Vertebrate flight: mechanics, physiology, morphology, ecology and evolution. Springer, Berlin Heidelberg New York
- Norberg UM, Rayner JMV (1987) Ecological morphology and flight in bats (Mammalia: Chiroptera): wing adaptations, flight performance, foraging strategy and echolocation. Philos Trans R Soc Lond 316B:335-427
- Norell MA (1989) The higher level relationships of the extant crocodilia. J Herpetol 23:325-335
- Normand ICS, Olver RE, Reynolds EOR, Strang LB (1971) Permeability of lung capillaries and alveoli to nonelectrolytes in the foetal lamb. J Physiol Lond 219:303–330
- Norris RA, Connell CE, Johnston DW (1957) Notes on fall plumages, weights and fat condition in the ruby-throated hummingbird. Wilson Bull 69:155–163
- Norsk P, Foldager N, Bonde-Petersen F, Elmann-Larsen B, Johansen TS (1987) Central venous pressure in humans during short periods of weightlessness. J Appl Physiol 63:2433-2437
- Northcutt RG (1990) Ontogeny and phylogeny: a reevaluation of conceptual relationships and some applications. Brain Behav Evol 36:116–140
- Nossaman BD, Feng CJ, Wang J, Kadowitz PJ (1994) Analysis of angiotensins I, II, III in pulmonary vascular bed of the rat. Am J Physiol 266:L389–L396
- Novacek MJ (1980) Cranioskeletal features in tupaiids and selected *Eutheria* as phylogenetic evidence. In: Luckett WP (ed) Comparative biology and evolutionary relationships of tree shrews. Plenum, New York, pp 35–67
- Novacek MJ (1982) Information for molecular studies from anatomical and fossil evidence on higher eutherian phylogeny. In: Goodman M (ed) Macromolecular sequences in systematic and evolutionary biology. Plenum Press, New York, pp 3–41
- Novy MJ, Parer JT (1969) Absence of high blood oxygen affinity in the foetal cat blood. Respir Physiol 6:144–151
- Nursall JR (1959) Oxygen as a prerequisite to the origin of the Metazoa. Nature (Lond) 183:1170-1172
- Nurse P (1997) The ends of understanding. Nature (Lond) 387:657
- Nyberg R, West B (1957) The influence of oxygen tension and some drugs on human blood vessels. Acta Physiol Scand 39:216–227
- O'Dor RK (1982) Respiratory metabolism and swimming performance of the squid, *Loligo* opalescens. Can J Fish Aquat Sci 39:580–587
- O'Dor RK, Webber DM (1991) Invertebrate athletes: trade-offs between transport efficiency and power density in cephalopod evolution. J Exp Biol 160:93–112
- Odum EP, Connell CE (1956) Lipid levels in migrating birds. Science 123:892-894
- O'Farrell MJ, Bradley WG (1977) Comparative thermal relationships of flight for some bats in southwestern United States. Comp Biochem Physiol 58A:223–227

Officer CB, Biggs RB, Taft JL, Cronin LE, Tyler MA, Boynton WR (1984) Chesapeake Bay anoxia: origin, development and significance. Science 223:22–27

Ogden E (1945) Respiratory flow in Mustelus. Am J Physiol 145:134-139

- Ohkuwa T, Sato Y, Naoi M Chang LY (1997) Glutathione status and reactive oxygen generation in tissues of young and old exercised rats. Acta Physiol Scand 159:237–244
- Oikawa S, Itazawa Y (1993) Allometric relationship between tissue respiration and body mass in a marine teleost, pogy *Pagrus major*. Comp Biochem Physiol 195:129–133
- Ojha J, Singh SK (1986) Scanning electron microscopy of the gills of a hill-stream fish, Danio dangila (Ham.). Arch Biol Bruxelles 97:455-467
- Okada O, Presson RG, Kirk KR, Godbey PS, Capen RL, Wagner WW (1992) Capillary perfusion patterns in single alveolar walls. J Appl Physiol 72:1838–1844
- Okada Y, Ishiko S, Daido S, Kim J, Ikeda S (1962) Comparative morphology of the lung with special reference to the alveolar epithelial cells. I. Lungs of Amphibia. Acta Tuberc Jpn 11:63–87
- Olmo E (1991) Genome variations in the transition from amphibians to reptiles. J Mol Evol 33:68-75
- Olsen CR, Hale FC, Elsner R (1969) Mechanics of ventilation in the pilot whale. Respir Physiol 7:137–149
- Olsen GJ, Woese CR (1993) Ribosomal RNA: a key to phylogeny. FASEB J 7:113-123
- Olsen LF, Degn H (1985) Chaos in biological systems. Q Rev Biophys 18:165-225
- Olson E, Miller R (1958) Morphological integration. Univ Chicago Press, Chicago
- Olson KR (1991) Vasculature of the fish gills: anatomical correlates of physiological function. J Electron Microsc Tech 2:217–228
- Olson KR (1994) Circulation in bimodally breathing fish. Am Zool 34:280-288
- Olson KR (1996) Scanning electron microscopiy of the fish gill. In: Munshi JSD, Dutta HM (eds) Fish morphology: horizon of new research. Science Publishers, Lebanon, New Hampshire, pp 31–45
- Olson KR, Fromm PO (1973) A scanning electron microscope study of secondary lamellae and chloride cells of rainbow trout (*Salmo gairdnen*). Z Zellforsch Mikrosk Anat 143:439-449
- Olson KR, Kullman DK, Narkates AJ, Oparil S (1986a) Angiotensin extraction by trout tissues *in vivo* and metabolism by perfused gill. Am J Physiol 250:R532–541
- Olson KR, Munshi JSD, Ghosh TK, Ojha J (1986b) Gill microcirculation of the air-breathing climbing perch, *Anabas testudineus* (Bloch): relationships with the accessory respiratory and systemic circulation. Am J Anat 176:305–320
- Olson KR, Terwilliger N, Capuzzo MJ (1988) Structure of hemocyanin in larval and adult American lobsters. Am Zool 28:47A
- Olson KR, Munshi JSD, Ghosh TK (1990a) Vascular organization of the head and respiratory organs of the air-breathing catfish, *Heteropneustes fossilis*. J Morphol 203:165-179
- Olson KR, Taylor A, Capuzzo MJ (1990b) Correlation between hemocyanin structure and function in American lobsters. Am Zool 30:94A
- Olson KR, Roy PK, Ghosh TK, Munshi JSD (1994) Microcirculation of gills and accessory respiratory organs from the air-breathing snakehead fish, *Channa punctata, C. gachua*, and *C. marulius*. Anat Rec 238:92–107
- Olver RE, Strang LB (1974) Ion fluxes across the pulmonary epithelium and the secretion of lung liquid in the fetal lamb. J Physiol (Lond) 241:327–357
- O'Mahoney P, Full RJ (1984) Respiration of crabs in air and water. Comp Biochem Physiol 79A:275-282
- Onimaru H, Homma I (1987) Respiratory rhythm generator neurons in medulla of brain stemspinal cord prepartion from new born rat. Brain Res 403:380–384
- Oparin AI (1938) The origin of life. Macmillan, New York
- Oparin AI (1953) The origin of life. Dover Publications, London
- Opell BD (1987) The influence of web monitoring tactics of the tracheal systems of spiders in the family Uloboridae (Arachnida, Areneida). Zoomorphology 107:255–259
- Orgel LE (1994) The origin of life on the earth. Sci Am 271:53-61
- Orr WC, Sohal RS (1994) Extension of lifespan by overexpression of superoxide dismutase and catalase in *Drosophila melanogaster*. Science 263:1128–1130
- Osaki S (1996) Spider silk as mechanical lifeline. Nature (Lond) 384:419

- Osawa S, Jukes TH, Watanabe K, Muto A (1992) Recent evidence for evolution of the genetic code. Microbiol Rev 56:229–264
- Oseid D, Smith L (1974) Chronic toxicity of hydrogen sulfide to Gammarus pseudolimnaeus. Trans Am Fish Soc 103:819-822

Oshino N, Sugano T, Oshino R, Chance B (1974) Mitochondrial function under hypoxic conditions: The steady states of cytochrome  $a + a^3$  and their relation to mitochondrial energy state. Biochim Biophys Acta 368:298–310

Osterberg R (1974) Origins of metal ions in biology. Nature (Lond) 249:382-383

Östlund E, Fänge R (1962) Dasodilation by adrenaline and noradrenaline and the effects of some other substances on perfused fish gills. Comp Biochem Physiol 97:292–303

Ostrom JH (1975) The origin of birds. Annu Rev Earth Planet Sci 3:55-77

- Owens T, Cess RD, Ramanathan V (1979) Enhanced carbon dioxide greehouse to compensate for reduced solar luminosity on early Earth. Nature (Lond) 277:640–641
- Pace N, Smith AH (1981) Gravity, metabolic rate effects in mammals. Physiologist 24:S37-40
- Pace NR, Stahl DA, Lane DJ, Olsen GJ (1985) Analyzing natural microbial populations by rRNA sequences. Am Soc Microbiol News 51:4–12
- Pack AI, Galante R, Fishman AP (1984) Breuer-Hering reflexes in the African lungfish (Protopreus annectens). Fed Proc 43:A433
- Pack AI, Galante R, Fishman AP (1992) Role of lung inflation in control of air breath duration in African lungfish (*Protopterus annectens*). Am J Physiol 262:879–884
- Packard A (1972) Cephalopods and fish: the limits of convergence. Biol Rev 47:241-307
- Packard CC, Sotherland PR, Packard MJ (1977) Adaptive reduction in permeability of avian eggshells to water vapour at high altitudes. Nature (Lond) 266:252

Packard GC (1974) The evolution of air-breathing in Paleozoic gnathostome fishes. Evolution 28:320–325

Packard GC, Packard MJ (1997) Type of soil affects survival by overwintering hatchilings of the painted turtle. J Therm Biol 22:53–58

Packard GC, Sotherland PR, Packard MJ (1997) Adaptive reduction in permeability of avian egg shells to water vapour at high altitudes. Nature (Lond) 266:255–256

- Packard GC, Elinson RP, Gavaud J, Guillette L, Lombardi J et al. (1989) How are reproductive systems integrated and how has viviparity evolved? In: Wake D, Roth G (eds) Complex organismal functions: integration and evolution in vertebrates. John Wiley (Dahlem Workshop Report), Chichester, pp 281–293
- Padian K (1982) Macroevolution and the origin of major adaptations: vertebrate flight as a paradigm for the analysis of pattern. Proc 3rd N Am Paleontol Convec, pp 387–392
- Padian K (1983) A functional analysis of flying and walking in pterosaurs. Paleobiology 9:218–239

Paganelli CV (1980) The physics of gas exchange across the avian egg shell. Am Zool 20:329-338

- Page TL (1994) Time is the essence: molecular analysis of the biological clock. Science 263:1570– 1572
- Palmer JD (1997) The mitochondrion time forgot. Nature (Lond) 387:454-455
- Palmer MF (1968) Aspects of the respiratory Physiology of *Tubifex tubifex* in relation to its ecology. J Zool (Lond) 154:463–473
- Palmer RMJ, Rerrige AC, Moncada S (1987) Nitric oxide release accounts for the biological activity oe endothelium derived relaxing factor. Nature (Lond) 327:524–526
- Palomeque J, Rodriguez JD, Placios L, Planas J (1980) Blood respiratory properties of swifts. Comp Biochem Physiol 67A:91-95
- Panchen AL, Smithson TR (1988) The relationships of the earliest tetrapods. In: Benton MJ (ed) The phylogeny and classification of the tetrapods, vol 1: Amphibians, reptiles birds. Clarendon Press, Oxford, pp 1–32
- Parker DJ, Cook S, Warwick MT (1975) Serum complement studies during and following cardiopulmonary bypass. In: Junod AF, Haller R (eds) Lung metabolism. Academic Press, New York, pp 481-487
- Parker HW (1940) The Percy Sladen Trust Expedition to Lake Titicaca, Amphibia ser 3. Trans Linn Soc Lond 12:203–216
- Parker WN (1892) On the anatomy and physiology of *Protopterus annectens*. Trans R Irish Acad 30:111–230

- Part P, Tuurala H, Nikinmaa M, Kiessling A (1984) Evidence for nonrespiratory intralamellar shunt in perfused rainbow trout gills. Comp Biochem Physiol 79A:29–34
- Pastor LM (1995) The histology of the reptilian lungs. In: Pastor LM (ed) Histology, ultrastructure and immunohistochemistry of respiratory organs in non-mammalian vertebrates. Secretariado de Oublicaciones de la Universidad de Murcia. Spain, pp 131-153
- Pastor LM, Ballesta J, Castells MT, Perez-Tomas R, Marin JA, Madrid JF (1989) A microscopic study of the lung of *Testudo graeca*. J Anat 162:19-33
- Patel S, Spencer (1963) Studies on the haemoglobin of Arenicola marina. Comp Biochem Physiol 8:65-82
- Patt DI, Patt GR (1969) The respiratory system. In: Comparative vertebrate histology. Harper and Row, New York
- Pattinson RC, Odendall HJ, Kirsten G (1993) The relationship between absent end-diastolic velocities of the umbilical artery and perinatal mortality and morbidity. Early Human Dev 33:61–69
- Pattle RE (1976) The lung surfactant in the evolutionary tree. In: Hughes GM (ed) Respiration of amphibious vertebrates. Academic Press, London, pp 233–255
- Pattle RE, Schock C, Creasey JM, Hughes GM (1977) Surpellic films, lung surfactant, and their cellular origin in newt, caecilian and frog. J Zool (Lond) 182:125-136
- Paul GS (1990) A re-evaluation of the mass of and flight of giant pterosaurs. J Vertebr Paleontol 10:37
- Paul GS (1991) The many myths, some old, some new, of dinosaurology. Mod Geol 16:69– 99
- Paul R, Fincke T, Linzen B (1987) Respiration in the tarantula Eurypelma califormicum: evidence of diffusion lungs. J Comp Physiol B 157:209-217
- Paul RJ (1989) Smooth muscle energetics. Annu Rev Physiol 51:331-349
- Paul RJ (1992) Gas exchange, circulation and energy metabolism in spiders. In: Wood SC, Weber RE, Hargens AR, Millard RW (eds) Physiological adaptations in vertebrates: respiration, circulation, and metabolism. Marcel Dekker, New York, pp 169–197
- Paulev P (1965) Decompression sickness following repeated breathhold dives. J Appl Physiol 20:1028-1031
- Pavlov NA, Krivchenko AT, Cherepivskaya EN, Zagvazdin YS, Zayat ND (1987) Reactivity of cerebral vessels in the pigeon, *Columba livia*. J Evol Biochem Physiol 23:447-451
- Pearson OP, Pearson AK (1976) A streological analysis of the ultrastructure of the lungs of wild mice living at low and high altitude. J Morphol 150:359–368
- Peebles PJE, Schramm DN, Turner EL, Kron RG (1994) The evolution of the Universe. Sci Am 271:29–33
- Pegg JH, Horner TL, Wahrenbrock EA (1963) Breathing of pressure-oxygenated liquids. Proc 2nd Symp Underwater Physiol, Natl Acad Sci Natl Res Council Publ 1191, pp 166– 170
- Peitgen HO, Richter PH (1986) The beauty of fractals: images of complex dynamical systems. Springer, Berlin Heidelberg New York
- Pelseneer P (1935) Essai d'éthologie zoologique. Acad R Belgique Classe des Sciences, Publ Found Brussels, Agathon Potter, Paris
- Pelster B (1985) Mechanismen der Anpassung an das Leben in extremen Biotopen: vergleichede Studien zur Atmungsphysiologie bei *Lumpenus lampretaeformis* und *Bleennius pholis*. PhD Thesis, University of Düsseldorf
- Pelster B, Scheid P (1991) Activities of enzymes for glucose catabolism in the swim bladder of the European eel Anguilla anguilla. J Exp Biol 156:207–213
- Pelster B, Scheid P (1992a) Counter current concentration and gas secretion in the fish swim bladder. Physiol Zool 65:1-16
- Pelster B, Scheid P (1992b) Metabolism of the swim bladder epithelium and the single concentrating effect. Comp Biochem Physiol 105A:383–388
- Pelster B, Scheid P (1992c) The influence of gas gland metabolism and blood flow on gas deposition into the swim bladder of the European eel Anguilla anguilla. J Exp Biol 173:205–216
- Pelster B, Scheid P (1993) Glucose metabolism of the swim bladder tissue of the European eel, Anguilla anguilla. J Exp Biol 185:169–178

- Pelster B, Bridges CR, Grieshaber MK (1988a) Respiratory adaptations of the burrowing marine teleost *Lumpenus lampretaeformis*. (Walbaum). II. Metabolic adaptations. J Exp Mar Biol Ecol 124:43–55
- Pelster B, Bridges CR, Taylor AC, Morris S, Artikinson RJA (1988b) Respiratory adaptations of the burrowing marine teleost *Lumpenius lampretaeformis* (Walbaum). I.  $O_2$  and  $CO_2$  transport, acid-base balance: a comparison with *Depola rubescens*. J Exp Mar Biol Ecol 124:31-42
- Pelster B, Kobayashi H, Scheid P (1988c) Solubility of nitrogen and argon in eel whole blood and its relation to pH. J Exp Biol 135:243–252
- Pelster B, Kobayashi H, Scheid P (1989) Metabolism of the perfused swim bladder of the European eel: oxygen, carbon dioxide, glucose and lactate balance. J Exp Biol 144:495-506
- Pelzenberger M, Pohla H (1992) Gill surface area of water and air breathing fish. Rev Fish Biol Fisheries 2:187–216
- Pendergast DR, Olszowska AJ, Rokitka MA, Farhi LE (1987) Gravitational force and the cardiovascular system. In: Dejours P (ed) Comparative physiology of environmental adaptations, vol 2. Karger, Basel, pp 15–26
- Penman HL (1940a) Gas and vapour movements in the soil. I. The diffusion of vapours through porous solids. J Agric Sci 30:437-462
- Penman HL (1940b) Gas and vapour movements in the soil. II. The diffusion of carbon dioxide through porous solids. J Agric Sci 30:570–581
- Penney DG (1977) Effects of prolonged diving anoxia on the turtle, *Pseudemys scripta elegans*. Comp Biochem Physiol A 47:933–941
- Penny DG, Hasegawa M (1997) The platypus put in its place. Nature (Lond) 387:549-550
- Pennycuick CJ (1975) Mechanics of flight. In: Famer DS, King JR (eds) Avian biology, vol 5. Academic Press, New York, pp 1–75
- Pennycuick CJ (1992) Newtonian rules in biology. Oxford University Press, New York
- Pennycuick CJ, Rezende MA (1984) The specific power output of aerobic muscle, related to the power density of mitochondria. J Exp Biol 108:377–392
- Penry DL, Jumars PA (1987) Modelling animal guts as chemical reactors. Am Nat 129:69-96
- Perkins JR (1964) Historical development of respiratory physiology. In: Handbook of physiology. Respiration, sect 3, vol I, Am Physiol Soc, Washington, DC, pp 1–62
- Perlo S, Jalowayski AA, Durand CM, West JB (1975) Distribution of red and white blood cells in alveolar walls. J Appl Physiol 38:117–124
- Perry SF (1978) Quantitative anatomy of the lungs of the red-eared turtle. *Pseudemys scripta elegans*. Respir Physiol 35:245-262
- Perry SF (1981) Morphometric analysis of pulmonary structure: methods for evaluation of unicameral lungs. Microscopie 38:278-293
- Perry SF (1983) Reptilian lungs: functional anatomy and evolution. Adv Anat Embryol Cell Biol 79:1–81
- Perry SF (1988) Functional morphology of the lungs of the Nile crocodile *Crocodylus niloticus*: non-respiratory parameters. J Exp Biol 143:99–117
- Perry SF (1989) Mainstreams in the evolution of vertebrate respiratory structures. In: King AS, McLelland J (eds) Form and function in birds, vol V, Academic Press, London, pp 1-67
- Perry SF (1992a) Evolution of the lung and its diffusing capacity. In: Bicudo JPW (ed) Vertebrate gas transport cascade adaptations to environment and mode of life. CRC press, Boca Raton, pp 142–153
- Perry SF (1992b) Gas exchange strategies in reptiles and the origin of the avian lung. In: Wood SC, Weber RE, Hargens AR, Millard RW (eds) Physiological adaptations in vertebrates: respiration, circulation, and metabolism. Marcel Dekker, New York, pp 149–167
- Perry SF, Duncker HR (1978) Lung architecture, volume and static mechanics in five species of lizards. Respir Physiol 34:61–81
- Perry SF, Duncker HR (1980) Interrelationship of static mechanical factors and anatomical structure in lung ventilation. J Comp Physiol 138:321–334
- Perry SF, Laurent P (1990) The role of carbonic anhydrase in carbon dioxide excretion, acid base balance and ionic regulation in aquatic gill breathers. In: Truchot JP, Lahlou B (eds) Transport, respiration and excretion: comparative and environmental aspects. Karger, Basel, pp 39–67

- Perry SF, McDonald G (1993) Gas exchange. In: Evans DH (ed) The physiology of fishes. CRC Press, Boca Raton, pp 251–278
- Perry SF, Darian-Smith C, Alston D, Limpus CJ, Maloney JE (1989a) Histological structure of the lungs of the loggerhead turtle, *Caretta caretta*, before and after hatching. Copeia 1989:1000–1010
- Perry SF, Bauer AM, Russell AP, Alston JT, Maloney JE (1989b) Lungs of the geccko *Rhacodactylus leachianus* (Reptilia: Gekkonidae): a correlative gross anatomical and light and electron microscopic study. J Morphol 199:23–46
- Perry SF, Auman U, Maloney JE (1989c) Intrinsic lung musculature and associated ganglion cells in a teiid lizard, *Tupinambis nigropunctatus spix*. Herpetologica 45:217–227
- Perutz MF (1970) Stereochemistry of cooperative effects in haemoglobin. Haem-haem interaction and problem of allostery. The Bohr effect and combination with organic phosphates. Nature (Lond) 228:726-733
- Perutz MF (1979) Regulation of oxygen affinity of haemoglobin: influence of structure of the globin on the haeme iron. Annu Rev Biochem 48:327–386
- Perutz MF (1983) Species adaptation in a protein molecule. Mol Biol Evol 1:1-28
- Perutz MF (1990a) Molecular inventiveness. Nature (Lond) 348:583-584
- Perutz MF (1990b) Mechanisms regulating the reactions of human haemoglobin with oxygen and carbon monoxide. Annu Rev Physiol 52:1–25
- Perutz MF (1996) Taking the pressure off. Nature (Lond) 380:205-206
- Peters HM (1978) On the mechanism of air ventilation in anabantoids (Pisces, Teleostei). Zoomorphologie 89:93-123
- Petersen JA, Fyhn HJ, Johansen K (1974) Ecophysiological studies of an intertidal crustacean, *Pollicipes polymerus* (Cirripeda, Lepadomorpha): Aquatic and aerial respiration. J Exp Biol 61:309-320
- Petit-Maire N (1991) Paléoenvironments du Sahara. CNRS, Paris
- Petroski H (1985) To engineer is human. St Martin's Press, New York
- Petschow D, Würdinger, I, Baumann R, Duhm J, Braunitzer G, Bauer C (1977) Causes of high blood oxygen affinity of animals living at high altitude. J Appl Physiol 42:139–143
- Petterson A, Hardin J (1969) Flight speeds of five species of vespertilionid bats. J Mammal 50:152-153
- Pettersson K, Nilsson S (1979) Nervous control of the branchial vascular resistance of the Atlantic cod, *Gadus morhua*. J Comp Physiol 129:179–183
- Pettigrew JD, Jamieson BGM, Robson SK, Hall LS, McNally KI, Cooper HM (1989) Phylogenetic relations between microbats, megabats and primates (Mammalia: Chiroptera and Primates). Philos Trans R Soc Lond 325B:489–559
- Philander SGH (1983) El Niño southern oscillation phenomena. Nature (Lond) 302:295-301
- Phillips CG, Kaye SR, Schroter RC (1994) A diameter-based reconstruction of the branching pattern of the human bronchial tree part I. Description and application. Respir Physiol 98:193-217
- Phillipson J (1981) Bioenergetic options and phylogeny. In: Townsend CR, Calow P (eds) Physiological ecology: an evolutionary approach to resource use. Blackwell, London, pp 20– 45
- Phleger CF, Saunders BS (1978) Swim bladder surfactants of Amazon air breathing fishes. Can J Zool 56:946–952
- Phleger CF, Smith DG, Macintyre DH, Saunders BS (1978) Alveolar and saccular lung phospholipids of the anaconda, *Eunectes murinus*. Can J Zool 56:1009–1013
- Pickard WF (1974) Transition regime diffusion and the structure of the insect tracheolar system. J Insect Physiol 20:947–956
- Pierce VA, Crawford DL (1997) Phylogenetic analysis of glycolytic enzyme expression. Science 276:256–259
- Piiper J, Scheid P (1972) Maximum gas transfer efficacy of models for fish gills, avian lungs and mammalian lungs. Respir Physiol 14:115–124
- Piiper J, Scheid P (1975) Gas transfer efficacy of gills, lungs and skin: theory and experimental data. Respir Physiol 23:209–221
- Piiper J, Scheid P (1980) Blood-gas equilibration in lungs. In: West JB (ed) Pulmonary gas exchange, vol I. Academic Press, New York, pp 121–171

- Piiper J, Scheid P (1981) Model for capillary-alveolar equilibration with special reference to O<sub>2</sub> uptake in hypoxia. Respir Physiol 46:193–205
- Piiper J, Scheid P (1989) Respiratory mechanics and air flow in birds. In: King AS, McLelland J. Form and function in birds, vol. 4. Academic Press, London, pp 369–391
- Piiper J, Scheid p (1992) Modeling of gas exchange in vertebrate lungs, gills, and skin. In: Wood SC, Weber RE, Hargens AR, Millard RW (eds) Physiological adaptations in vertebrates: respiration, circulation, and metabolism. Marcel Dekker, New York, pp 69–95
- Piiper J, Humphrey HT, Rahn H (1962) Gas composition of pressurized, perfused gas pockets and the fish swim bladder. J Appl Physiol 17:275–282
- Piiper J, Dejours P, Haab P, Rahn H (1971) Concepts and basic quantities in gas exchange physiology. Respir Physiol 13:292-304
- Piiper J, Gatz RN, Crawford EC (1976) Gas transport characteristics in an exclusively skin breathing salamander, *Desmognathus fuscus* (Plethodontidae). In: Hughes GM (ed) Respiration in amphibious vertebrates. Academic Press, London, pp 339–356
- Piiper J, Meyer M, Worth H, Willmer H (1977) Respiration and circulation during swimming activity in the dogfish, *Scyliorhinus stellaris*. Respir Physiol 30:338–349
- Piiper J, Tazawa H, Ar A, Rahn H (1980) Analysis of chorioallantoic gas exchange in the chick embryo. Respir Physiol 39:273–287
- Piiper J, Scheid P, Perry SF, Hughes GM (1986) Effective and morphometric oxygen diffusing capacity of the gills of the elasmobranch *Scyliorhinus stellaris*. J Exp Biol 123:27–41
- Pilgrim M (1966) The anatomy and histology of the blood system of the maldanid polychaetes *Clymenella torquata* and *Euclymene oerstedi*. J Zool (Lond) 149:261
- Pilson ME (1965) Variation of hemocyanin concentration in the blood of four species of haliotis. Biol Bull 128:459–472
- Pinkerton KE, Barry BE, O'Neil JJ, Raus JA, Pratt PC, Crapo JD (1982) Morphologic changes in the lung during the lifespan of Fischer 344 rats. Am J Anat 164:155–174
- Pinshaw B, Bernstein MH, Arad Z (1985) Effects of temperature and PCO<sub>2</sub> on O<sub>2</sub> affinity of pigeon blood: implications for brain O<sub>2</sub> supply. Am J Physiol 249:R759–R764
- Piper HM, Noll T, Siegmund B (1994) Mitochondrial function in the oxygen depleted and reoxygenated myocardial cell. Cardiovasc Res 28:1-15
- Piper PJ (1973) Distribution and metabolism. In: Cuthbert MF (ed) The prostaglandins. Pharmacological and therapeutic aspects. Heineman, London, pp 125-150
- Piper PJ (1975) Conditions of release of prostaglandins from the lung. In: Junod Af, Haller R (eds) Lung metabolism. Academic Press, New York, pp 315–319
- Pizarro B, Salas A, Paredes J (1970) Mal de altura en aves. Inst Vet Invest Trop Altura Cuarto Boletin Extraord 1970:147–151
- Pizarello DJ, Shircliffe AC (1967) Hyperbaric oxygen: toxic effects in chick embryos. Am Surg 33:958–957
- Platt T, Irwin B (1972) Phytoplankton productivity and nutrient measurements in Petpeswick Inlet, 1971–1972. Fish Res Board Can Tech Rep 314
- Plattner W (1941) Etudes sur la fonction hydrostatique de la vessie natatoire des poissons. Rev Suisse Zool 48:201–338
- Poczopko P (1959) Changes in blood circulation in *Rana escutenta* L. while diving. Zool Pol 10:29-43
- Pohunkova H (1967) The ultrastructure of the lung of the snail *Helix pomatia*. Folia Morphol 15:250–257
- Pohunkova H (1969) Lung ultrastructure of the Arachnida-Arachinoidea. Folia Morphol Prague 17:309–361
- Pohunkova H, Hughes GM (1985a) Structure of the lung of the clawed toad (*Xenopus laevis* Daudin). Folia Morphol XXXIII:385-390
- Pohunkova H, Hughes GM (1985b) Ultrastructure of the lungs of the garter snake. Folia Morphol Prague 23:254–258
- Polanyi M (1968) Life's irreducible structure. Science 160:1308-1312
- Policard A (1929) Les nouvelles idées sur la disposition de la surface respiratoire pulmonaire. Presse Med 80:1–20
- Polimanti O (1912) Über den Beginn der Atmung bei Embryonen von Scyllium. Z Biol 57:237– 272

- Polimanti O (1913) Sui rapporti fra peso del corpo e ritmo respiratoria di Octopus vulgaris Lam. Z Allg Physiol 15:449–455
- Popper K (1968) The logic of scientific discovery. Hutchinson, London

Popper K (1969) Conjectures and refutations. Routledge and Kegan Paul, London

- Portier P (1933) Locomotion aérienne et respiration des lépidoptères, un nouveau rôle physiologique des ailes et des écailles. Trav V Congr Int Ent Paris 2:25-31
- Portier P, Duval M (1929) Recherches physiologiques sur la teneur en gaz carbonique de l'atmosphére interne des fourmilieres. CR Soc Biol 102:906–908
- Postgate JR (1987) Nitrogen fixation. Edward Arnold, London
- Potter EL, Bohlender GP (1941) Intrauterine respiration in relation to development of foetal lung with report of 2 unusual anomalies of respiratory system. Am J Ogstet Gynecol 42:14– 22
- Potter GE (1927) Respiratory function of swim bladder in Lepidosteus. J Exp Zool 49:45-52
- Pough FH (1980) Blood oxygen transport and delivery in reptiles. Am Zool 31:455-456
- Pough FH, Taigen TL, Stawart MM, Brussard PF (1983) Behavioural modification of evaporative water loss by a Puerto Rican frog. Ecology 60:608–613
- Pough FH, Heiser JB, McFarland WN (1989) Vertebrate life, 3rd edn. Macmillan, New York
- Powell CS (1993) Livable planets. Sci Am 268:7-8
- Powell FL (1982) Diffusion in avian lungs. Fed Proc 41:53-55
- Powell FL (1990) Acclimatization to high altitude. In: Sutton JR, Coates GC, Remmers JE (eds) Hypoxia: the adaptations. BC Decker, Toronto, pp 41–44
- Powell FL, Scheid P (1989) Physiology of gas exchange in the avian respiratory system. In: King AS, McLelland J (eds) Form and function in birds, vol 4. Academic Press, London, pp 393–437
- Powell FL, Wagner PD (1982a) Ventilation-perfusion inequality in the avian lungs. Respir Physiol 38:233-241
- Powell FL, Wagner PD (1982b) Measurement of continuous distributions of ventilationperfusion in non-alveolar lungs. Respir Physiol 48:219–232
- Powell MA, Arp AJ (1989) Hydrogen sulfide oxidation by abundant nonhaemoglobin heme compounds to marine invertebrates from sulfide rich habitats. J Exp Zool 249:121– 132
- Powell MA, Somero GN (1985) Sulfide oxidation occurs in the animal tissue of the gutless clam, Solemya reidi. Biol Bull 169:164–181
- Powell MA, Crenshaw MA, Rieger RN (1979) Adaptations to sulfide in the meiofauna of the sulfide system. 1<sup>35</sup> S-sulfide accumulation and the presence of a sulfide detoxification system. J Exp Mar Biol Ecol 37:57–76
- Powers DA (1972) Haemoglobin adaptation for fast and slow water habitats in sympatric catostomid fishes. Science 177:360-362
- Powers LW, Bliss DE (1983) Terrestrial adaptations. In: Vernberg FJ, Vernberg WB (eds) Environmental adaptations: biology of Crustacea, vol 8. Academic Press, New York, pp 271-333
- Powers DA, Fyhn HJ, Fyhn UEH, Martin JP, Garlick RL, Wood SC (1979) A comparative study of the oxygen equilibria of blood from 40 genera of Amazonian fishes. Comp Biochem Physiol 62A:67-86
- Prange HD (1976) Energetics of swimming in a sea turtle. J Exp Biol 64:1-12
- Prange HD, Ackerman RA (1974) Oxygen consumption and mechanisms of gas exchange of green turtle (*Chelonia mydas*) eggs and hatchlings. Copeia 3:758–763
- Prange HD, Wasser JS, Gaunt AS, Gaunt SLL (1985) Respiratory responses to acute heat stress in cranes (Gruidae): the effects of tracheal coiling. Respir Physiol 62:95–103
- Prankerd TAJ (1961) The red cell: an account of its chemical physiology and pathology. Blackwell, Oxford
- Prasad MS (1988) Morphometrics of gills during growth and development of air-breathing habit in *Colisa fasciatus* (Bloch and Schneider). J Fish Biol 32:367–381
- Precht H (1939) Die Lungenatmung der Süsswasserpulmonaten. Z Vergl Physiol 26:696-738
- Prestwich KN (1983) The roles of aerobic and anerobic metabolism in active spiders. Physiol Zool 56:122–132
- Priede IG (1977) Natural selection for energetic efficiency and relationship between activity level and mortality. Nature (Lond) 267:610

Prigogine I, Stengers I (1984) Order of chaos: man's new dialogue with nature. Heineman, London

Pringle JWS (1983) Insect flight. Carolina Biology Readers, Burlington, North Carolina

Prinzinger R, Hinninger CH (1992) Endogenous? Diurnal rhythmn in the energy metabolism of pigeon embryos. Naturwissenschaften 79:278–279

- Prior DJ, Hume M, Varga D, Hess SD (1983) Physiological and behavioural aspects of water balance and respiratory function in the terrestrial slug, *Limax maximus*. J Exp Biol 104:111– 127
- Prisk GK, Guy HJB, Eliot AB, Deutschman RA, West JB (1993) Pulmonary diffusing capacity, capillary blood volume and cardiac output during sustained microgravity. J Appl Physiol 75:15-26
- Pritchard A, White GN (1981) Metabolism and oxygen transport in the innkeeper worm *Urechis caupo*. Physiol Zool 54:44–54
- Proctor DF, Caldini P, Permutt S (1968) The pressure surrounding the lungs. Respir Physiol 5:130-144
- Prosser CL (1958) The nature of physiological adaptations. In: Prosser DL (ed) Physiological adaptations. American Physiological Society, Washington, DC, pp 167–180
- Prosser CL (1961) Oxygen: respiration and metabolism. In: Prosser CL, Brown FA (eds) Comparative animal physiology. Saunders, Philadelphia, pp 198-287
- Prosser CL (1973) Comparative animal physiology, 3rd edn. Saunders, Philadelphia
- Prosser CL (1986) Adaptational biology: molecules to organisms. John Wiley, New York
- Prosser CL, Brown FA (1962) Comparative animal physiology, 2nd edn. WB Saunders, London
- Prothero J (1986) Scaling of energy metabolism in unicellular organisms: a re-analysis. Comp Biochem Physiol 83A:2243–248
- Pryor WA (1986) Oxy-radicals and related species: their formation, life times, and reactions. Annu Rev Physiol 48:657–667
- Pugh LGCE (1962) Physiological and medical aspects of the Himalayan scientific and mountaineering expedition, 1960–61, Br Med J 2:621–626
- Pullin RŠV, Morris DJ, Bridges CR, Atkinson RJA (1980) Aspects of the respiratory physiology of the burrowing fish *Cepola rubescens* L. Comp Biochem Physiol 66A:35-42
- Qasim SZ, Qayyum A, Garg KK (1960) The measurement of carbon dioxide produced by airbreathing fishes and evidence of respiratory function of accessory respiratory organs. Proc Indian Acad Sci 52:19–26
- Quinlan MC, Hadley NF (1993) Gas exchange, ventilatory patterns, and water loss in two lubber grasshoppers: quantifying cuticular water and respiratory transpiration. Physiol Zool 66:628-642
- Quist J, Hill RD, Schneider RC, Falke KJ, Liggins GC et al. (1986) Hemoglobin concentrations and blood gas tensions of free-diving Weddell seals. J Appl Physiol 61:1560–1569
- Rabalais NN, Cameron JN (1985) Physiological and morphological adaptations of adult Uca subcylindrica to semi-arid environments. Biol Bull 168:135-146
- Rabinowitch E, Govindjee I (1965) The role of chlorophyll in photosynthesis. Sci Am 213:74-82
- Rahn H (1966) Aquatic gas exchange: theory. Respir Physiol 1:1-12
- Rahn H (1967) Gas transport from the external environment to the cell. In: de Reuck AVS, Porter R (eds) Development of the lung: a CIBA Foundation Symposium. Churchill, London, pp 3–23
- Rahn H (1974) PCO<sub>2</sub>, pH and body temperature. In: Nahas G, Smith KE (eds) Carbon dioxide and metabolic regulation. Springer, Berlin Heidelberg New York, pp 752–761
- Rahn H (1977) Adaptations of the avian embryo to high altitude. In: Paintal AS, Gill-Kumar P (eds) Respiratory adaptations, capillary exchange and reflex mechanisms. Proc Krogh Centenary Symp, New Delhi University, India, pp 94–105
- Rahn H, Ar A (1974) The avian egg: incubation time and water loss. Condor 76:147-152
- Rahn H, Ar A (1980) Gas exchange of the avian egg: time, structure and function. Am Zool 20:477-484
- Rahn H, Howell BJ (1976) Bimodal gas exchange. In: Hughes GM (ed) Respiration of amphibious vertebrates. Academic Press, London, pp 271–285

Rahn H, Paganelli CV (1968) Gas exchange in gas gills of diving insects. Respir Physiol 5:145-164

Rahn H, Paganelli CV (1982) Role of diffusion in gas exchange of the avian egg. Fed Proc 41:2134-2146

- Rahn H, Rahn KB, Howell BJ, Gans C, Tenney SM (1971) Air-breathing of the garfish, Lepisosteus osseus. Respir Physiol 11:285-307
- Rahn H, Paganelli CV, Ar A (1974) The avian egg: air-cell gas tension, metabolism and incubation time. Respir Physiol 22:297–309
- Rahn H, Paganelli CV, Ar A (1975) Relation of avian egg weight to body weight. Auk 92:750-765
- Rahn H, Paganelli CV, Nisbet ICT, Whittow GC (1976) Regulation of incubation water loss in eggs of seven species of terns. Physiol Zool 49:245–259
- Rahn H, Carey C, Balmas K, Bhatia B, Paganelli C (1977) Reduction of pore area of the avian eggshell as an adaptation to altitude. Proc Natl Acad Sci USA 74:3095–3098
- Rahn H, Christensen VL, Edens FW (1981) Changes in shell conductans, pores, and physical dimensions of egg and shell during the first breeding cycle of turkey hens. Poult Sci 60:2536–2557
- Ralph R, Everson I (1968) The respiratory metabolism of some Antarctic fish. Comp Biochem Physiol 27:299–307
- Ramsay JA (1968) Physiological approach to the lower animals, 2nd edn. Cambridge University Press, Cambridge
- Rancour-Laferriére D (1985) Signs of the fresh: an essay on the evolution of hominid sexuality. Mouton de Gluyter, Berlin
- Rand DA, Wilson HB (1993) Evolutionary catastrophes, punctuated equilibria and gradualism in ecosystem evolution. Proc R Soc Lond 253B:137-141
- Randall DJ (1970) Gas exchange in fish. In: Hoar WS, Randall DJ (eds) Fish physiology, vol 4. Academic Press, London, pp 252–292
- Randall DJ (1972) Respiration. In: Hardisty MW, Potter IC (eds) The biology of lampreys, vol 2. Academic Press, London, pp 287–306
- Randall DJ (1982) The control of respiration and circulation in fish during exercise and hypoxia. J Exp Biol 100:275–288
- Randall DJ, Cameron JN (1973) Respiratory control of arterial pH as temperature changes in rainbow trout, *Salmo gairdneri*. Am J Physiol 225:997–1002
- Randall DJ, Daxboeck C (1984) Oxygen and carbon dioxide transfer across fish gills. In: Hoar WS, Randall DJ (eds) Fish physiology, vol 10, part A. Academic Press, New York, London, pp 263–314
- Randall DJ, Jones DR (1973) The effect of deafferentation of the pseudobranch on the respiratory response to hypoxia and hyperoxia in the trout (Salmo gairdneri). Respir Physiol 17:291–301
- Randall DJ, Shelton G (1963) The effects of changes in the environmental gas concentrations on the breathing and heart rate of a teleost fish. Comp Biochem Physiol 9:229–239
- Randall DJ, Holeton GF, Stevens ED (1967) The exchange of oxygen and carbon dioxide across the gills of the rainbow trout. J Exp Biol 46:339–348
- Randall DJ, Baumgarten D, Malyusz M (1972) The relationship between gas and ion transfer across the gills of fishes. Comp Biochem Physiol 41A:629–637
- Randall DJ, Farrell AP, Haswell MS (1978a) Carbon dioxide excretion in the jeju, *Hoplerythrinus unitaeniatus*, a facultative air-breathing teleost. Can J Zool 56:970–973
- Randall DJ, Farrell AP, Haswell MS (1978b) Carbon dioxide excretion in the piracucu (*Arapiama gigas*), an obligate air-breathing fish. Can J Zool 56:977–982
- Randall DJ, Burggren WW, Farrell AP, Haswell MS (1981) The evolution of air-breathing in vertebrates. Cambridge University Press, Cambridge
- Randall DJ, Wood CM, Perry SF, Bergman H, Maloiy GMO, Mommsen TP, Wright PA (1989) Ureotelism in a completely aquatic teleost fish: a strategy for survival in an extremely alkaline environment. Nature (Lond) 337:165-166
- Randles CA, Romanoff AL (1950) Some physical aspects of the amnion and allantois of the developing chick embryo. J Exp Zool 114:87-105
- Rannels DE, Rannels SR (1988) Compensatory growth of the lung following partial pneumonectomy. Exp Lung Res 14:157–182

- Rantin FT, Johansen K (1984) Responses of the teleost *Hoplias malabaricus* to hypoxia. Environ Biol Fish 11:221–228
- Rantin FT, Kalinin AL, Glass ML, Fernandes MN (1992) Respiratory responses to hypoxia in relation to mode of life of two erythrinid species (*Hoplias malabaricus* and *Hoplias lacerdae*). J Fish Biol 41:805–812
- Rantin FT, Glass ML, Kalinin AL, Verzola RMM, Fernandes MN (1993) Cardio-respiratory responses in two ecologically distinct erythrinids (*Hoplias malabaricus* and *Hoplias lacerdae*) exposed to graded environmental hypoxia. Environ Biol Fish 36:93–97
- Rappaport MM, Greene AA, Page IH (1948) Serum vasomotor (serotonin). J Biol Chem 176:1243-1251
- Rashevsky N (1960) Mathematical biophysics: physico-mathematical foundations of biology. Dover, New York
- Raup DM, Jablonski D (1993) Geography of end-Cretaceous marine bivalve extinctions. Science 260:971–973
- Raup DM, Sepkoski JJ (1984) Periodicity of extinctions in the geologic past. Proc Natl Acad Sci USA 81:801–805
- Raven PH, Wilson EO (1992) A fifty-year plan for biodiversity surveys. Science 258:1099-1100
- Raymo M, Ruddiman WF (1992) Tectonic forcing of late Cenozoic climate. Nature (Lond) 359:117-122
- Raynaud D, Jouzel J, Barnola JM, Chapellaz J, Delmas RJ, Lorius C (1993) The ice record of greenhouse gases. Science 259:926–934
- Rayner JWV (1981) Flight adaptations in vertebrates. Symp Zool Soc (Lond) 48:137-172
- Rayner JMV (1985) Bounding and undulating flight in birds. J Theor Biol 117:47-77
- Rayner JMV (1986) Vertebrate flapping flight mechanics and aerodynamics, and the evolution of flight in bats. In: Nachtigall W (ed) BIONA report No 5, Bat flight Fledermausflug. Gustav Fischer, Stuttgart, pp 27–74
- Reader J (1986) The rise of life: the first 3.5 billion years. Alfred A Knopf, New York
- Ready NE (1983) Wing development in hemimetabolous insects. PhD Thesis, University of California, Irvine
- Redfield AC (1958) The biological control of chemical factors in the environment. Sci Am 46:205-221
- Redfield AC, Florkin M (1931) The respiratory function of the blood of *Urechis carpo*. Biol Bull 61:185–210
- Reeve HK, Westneat DF, Noon WA, Sherman PW, Aquadro F (1990) DNA fingerprinting reveals high levels of inbreeding in colonies of the eusocial naked mole-rat. Proc Natl Acad Sci USA 87:2496–2500
- Reeves JT, Leathers JE (1964) Circulatory changes following birth of the calf and the effect of hypoxia. Circ Res 15:343–354
- Reeves RB (1977) The interaction of body temperatures and acid-base balance in ectothermic vertebrates. Annu Rev Physiol 40:559–586
- Reid WD, Ilett KF, Glick JM, Krishna G (1973) Metabolism and binding of aromatic hydrocarbons in the lung: relationship to experimental bronchiolar necrosis. Am Rev Respir Dis 107:539–551
- Reid RC, Sherwood TK (1966) The properties of gases and liquids, 2nd edn. McGraw-Hill, New York
- Reidbord HE (1967) An electronmicroscopic study of the alveolar capillary wall following intratracheal administration of saline and water. Am J Pathol 50:275–283
- Reimers CE, Fischer KM, Merewether R, Smith KL, Jahnke RA (1986) Oxygen microporfiles measured in situ in deep ocean sediments. Nature (Lond) 320:741-744
- Reite OB, Maloiy GMO, Aasenhaug B (1974) pH, salinity and temperature tolerance of Lake Magadi, Kenya. Nature (Lond) 247:315
- Remane A, Storch V, Welsch U (1980) Systematische Zoologie. Fischer, Stuttgart
- Remmer H (1975) Pulmonary drug-metabolizing enzymes. In: Junod AF, Haller R (eds) Lung metabolism. Academic Press, New York, pp 133–158
- Remotti E (1933) Development of allantoic circulation in response to external variations in gas content. Bull Mus Lab Zool Anat Comp, Univ Genova 13:1–19

Rennard SL, Basset G, Lecossier K, O'Donnell KM et al. (1986) Estimation of epithelial lining fluid recovered by lavage using urea as a marker for dilution. J Appl Physiol 60:532–538

Rennie J (1992) Living together. Sci Am 266:104-113

- Renous S, Gasc JP (1989) Body and vertebral proportions in Gymnophiona (Amphibia): diversity of morphological types. Copeia 1989:837–847
- Repetski JE (1978) A fish from Upper Cambrian of North America. Science 200:529
- Revelle R (1982) Carbon dioxide and world climate. Sci Am 247:33-41
- Revsbech NP, Jørgensen BB, Blackburn PH (1980a) Oxygen in the sea bottom measured with microelectrode. Science 207:1355–1356
- Revsbech NP, Sørensen J, Blakburn TH, Lomholt JP (1980b) Distribution of oxygen in marine sediments measured with microelectrodes. Limnol Oceanogr 25:403-411
- Reynolds WW, McCauley RW, Casterlin ME, Crawsha LI (1976) Body temperature of behaviourally thermoregulating largemouth bass, *Micropterus salmoides*. Comp Biochem Physiol 59A:461-475
- Reznick DN, Shaw FH, Rodd FH, Shaw RG (1997) Evaluation of the rate of evolution in natural populations of guppies (*Poecilia reticulata*). Science 275:1934–1936
- Rhoads DC, Morse PW (1971) Evolutionary and ecologic singificance of oxygen-deficient marine basins. Letharia 4:413-428
- Rice SA (1980) Hydrodynamic and diffusion considerations of rapid-mix experiments with erythrocytes. Biophys J 29:65-78
- Richards AB (1957) Studies on arthropod cuticle. XIII. The penetration of dissolved oxygen and electrolytes in relation to the multiple barriers of the epicuticle. J Insect Physiol 1:23–29
- Richards AG, Korda FH (1950) Studies on arthropod cuticle. IV. An electron microscope survey of the intima of arthropod tracheae. Ann Entomol Soc Am 43:49–71
- Richardson J (1976) Autumnal migration over Puerto Rico and the Western Atlantic: a radar study. Ibis 118:309–332
- Richman PS, Wolfson MR, Shaffer TH (1993) Lung lavage with oxygenated perfluorochemical liquid in acute lung injury. Crit Care Med 21:768–774
- Richter C, Park JW, Ames BN (1988) Normal oxidative damage to mitochondrial and nuclear DNA is extensive. Proc Natl Acad Sci USA 85:6465–6467
- Riddle WA (1983) Physiological ecology of land snails and slugs. In: Russell-Hunter WD (ed) The Mollusca. Academic Press, New York, pp 431–448
- Ridgway SH, Howard R (1979) Dolphin lung collapse and intramuscular circulation during free diving: evidence from nitrogen washout. Science 206:1182–1183
- Ridgway SH, Johnston DG (1966) Blood oxygen and ecology of porpoises of three genera. Science 151:456-467
- Riedel C, Wood SC (1988) Effects of hypercapnia and hypoxia on temperature selection of the toad, *Bufo marinus*. Fed Proc 2:500A
- Riedesel ML (1977) Blood physiology. In: Wimsatt WA (ed) Biology of bats, vol II. Academic Press, New York, pp 485-517
- Riedesel ML, Williams BA (1976) Continuous 24-hr oxygen consumption studies of *Myotis* velifer. Comp Biochem Physiol 54A:95–99
- Riedl R (1978) Order in living organisms. John Wiley, New York
- Rieppel O (1993) The conceptual relation of ontogeny, phylogeny, and classification: the taxic approach. Evol Biol 27:1-32
- Riggs A (1976) Factors in the evolution of haemoglobin function. Fed Proc 35:2115-2118
- Riggs A (1979) Studies of the haemoglobins of Amazonian fishes: A review. Comp Biochem Physiol 62A:257–271
- Riggs DS (1963) The mathematical approach to physiological problems. Williams and Wilkins, Baltimore
- Riley JP, Skirrow G (1975) Chemical oceanography, 2nd edn, vol 2. Academic Press, London, pp 134–198
- Risenbrough RW (1986) Pesticides and bird population. In: Johnston RF (ed) Current ornithology, vol 3. Plenum Press, New York, pp 397–427
- Roberts JL (1975) Respiratory adaptations of aquatic animals. In: Vernberg FJ (ed) Physiological adaptation to the environment. Intext Educational Publishers, New York pp 395–435

Roberts JL, Rowell DM (1988) Periodic respiration of gill breathing fishes. Can J Zool 66:182–192

- Robertson JI (1913) The development of the heart and vascular system of *Lepidosiren paradoxa*. Q J Microsc Sci Lond 59:53–132
- Robertson RJ, Gilcher R, Metz KF et al. (1982) Effect of induced erythrocythemia on hypoxia tolerance during physical exercise. J Appl Physiol 53:490-495
- Robin ED, Bromberg PA, Cross CE (1969) Some aspects of the evolution of vertebrate acid-base regulation. Yale J Biol Med 42:448–476
- Robinson JM (1991) Global planetary change. Paleobiology 97:51-62
- Rogers PJ, Stewart PR (1973) Respiratory development in *Saccharomyces cerevisiae* grown at controlled oxygen tension. J Bacteriol 115:88–97
- Rogers RM, Braunstein MS, Shuman JF (1972) Role of bronchopulmonary lavage in the treatment of respiratory failure: a review. Chest 62:958–1058
- Rohmer M, Bouvier P, Ourisson G (1979) Molecular evolution of biomembranes: structural equivalents and phylogenetic precusors of sterols. Proc Natl Acad USA 76:847-851
- Rolschau J (1978) A prospective study of the placental weight and content of protein, RNA and DNA. Acta Obstet Gynecol Scand (Suppl) 72:28-43
- Romanoff AL (1960) The avian embryo. Macmillan, New York
- Romanoff AL (1967) Biochemistry of the avian embryo. John Wiley, New York
- Romanoff AL, Romanoff AJ (1949) The avian egg. John Wiley, New York
- Romer AS (1946) The early evolution of fishes. Q Rev Biol 21:33-69
- Romer AS (1966) Vertebrate paleontology, 3rd edn. University of Chicago Press, Chicago
- Romer AS (1967) Major steps in vertebrate evolution. Science 158:1629-1637
- Romer AS (1972) Skin breathing primary or secondary? Respir Physiol 14:183-192
- Romijn C (1948) Respiratory movements of the chicken during the parafoetal period. Physiol Comp Oecol 1:24-48
- Romijn C (1950) Foetal respiration in the hen: gas diffusion through the egg shell. Poult Sci 29:42-51
- Ronan CA (1991) The natural history of the Universe. Doubleday, London
- Roper CFE (1969) Systematics and zoogeography of the worldwide bathypelagic squid Bathyteuthis (Cephalopoda: Oegopsida). Smithsonian Inst Press, Washington, DC, pp 1-210
- Rose FL, Zambernard J (1966) Cardiac glycogen depletion in Amphiuma means during induced anoxia. J Morphol 120:391-396
- Rose KD, Bown TM (1984) Gradual phyletic evolution at the generic level in early Eocene omomyid primates. Nature (Lond) 309:250-252
- Rosen R (1967) Optimality principles in biology. Plenum Press, New York
- Rosen DE, Forey PL, Gardiner BG, Petterson C (1981) Lungfishes, tetrapods paleontology and plesiomorphology. Bull Am Mus Nat Hist 167:159–276
- Rosen P, Stier A (1973) Kinetics of  $CO_2$  and  $O_2$  complexes of rabbit liver microsomal cytochrome  $P_{450}$ . Biochem Biophys Res Commun 51:603–611
- Rossi-Fanelli A, Antonini E (1957) A new type of myoglobin isolated and crystallized from muscles of Aplysiae. Biochemistry (USSR) 22:312–321
- Rossitti S, Löfgren J (1993a) Vascular dimensions of the cerebral arteries follow the principle of minimum work. Stroke 24:371–377
- Rossitti S, Löfgren J (1993b) Optimality principles and flow oderliness at the branching points of cerebral arteries. Stroke 24:1029–1032
- Rossitti S, Stephensen H (1994) Temporal heterogeneity of the blood flow velocity at the middle cerebral artery in the normal human characterized by fractal analysis. Acta Physiol Scand 1511:191–198
- Roth M, Depierre D (1975) Dipeptidyl carboxypeptidase in lung and blood plasma. In: Junod AE, Haller R (ed) Lung metabolism. Academic Press, New York, pp 337-345
- Roughton FJW (1945) The average time spent by the blood in the human lung capillary and its relation to the rate of  $CO_2$  uptake and elimination. Am J Physiol 143:621
- Roughton FJW, Forster RE (1957) Relative importance of diffusion and chemical reaction rates in determining rate of  $O_2$  exchange of pulmonary membrane and volume of blood in lung capillaries. J Appl Physiol 11:290–302
- Royer WE, Love WE, Fenderson FF (1985) Cooperative dimeric and tetrameric clam haemoglobins are novel assemblages of myoglobin folds. Nature (Lond) 316:277-280

- Ruben JA, Reagan NL, Verrell PA, Boucot AJ (1993) Plethodontid salamabder origins: a response to Beachy and Bruce. Am Nat 142:1038-1051
- Rubenstein DL, Koehl MAR (1977) The mechanism of filter-feeding: some theoretical consideration. Am Nat 111:981–994
- Ruckhäberle KE, Franke J, Viehweg B, Gerl D (1977) Quantitative Veränderungen an Resorptionszotten normaler menschlicher Plazenten im Verlauf der Gestation. Zentralbl Gynaekol 99:1313–1322

Rudwick MJS (1964) The inference of function from structure in fossils. Br J Philos Sci 15:27–40 Runham NW, Hunter PJ (1970) Terrestrial slugs. Hutchison, London

- Runnegar B (1982) Oxygen requirements, biology and phylogenetic significance of the late Precamrian worm Dickinsonia, and the evolution of the burrowing habit. Alcheringa 6:223– 239
- Runnegar B (1992) Evolution of the earliest animals. In: Schopf JW (ed) Major events of the history of life. Johns and Bartlett, Boston, pp 65–95
- Rurak DW, Gruber NC (1983) Increased oxygen consumption associated with breathing activity in foetal lambs. J Appl Physiol 54:701–707
- Rushner RF (1965) General characteristics of the cardiovascular system. In: Ruch TC, Patton HD (eds) Physiology and biophysics, 19th edn. Saunders, Philadelphia, pp 543–549
- Russell CW, Evans BK (1989) Cardiovascular anatomy and physiology of the black-lip abalone, Haliotis ruber. J Exp Zool 252:105–117
- Russell MJ, Daniel RM (1992) Emergence of life via catalytic hydrothermal colloidal iron sulphide membranes. Ann Geophys Suppl III 10:506
- Russell MJ, Daniel RM, Hall AJ, Sherringham JA (1994) A hydrothermally precipitated catalytic iron sulphide membrane as a first step toward life. J Mol Evol 39:231–243
- Ruthen R (1993) Adapting to complexity. Sci Am 268:130-140
- Rutten MG (1970) The history of atmospheric oxygen. Space Life Sci 2:5-17
- Ruud JT (1954) Vertebrates without erythrocytes and blood pigment. Nature (Lond) 173:848-850
- Ruud JT (1965) The ice fish. Sci Am 213:108-114
- Ryan JW, Ryan US (1975) Metabolic activities of plasma membrane and caveolae of pulmonary endothelial cells, with a note on pulmonary prostaglandin synthetase. In: Junod AE, Haller R (eds) Lung metabolism. New York, Academic Press, pp 399–424
- Ryan JW, Roblero J, Stewart JM (1968) Inactivation of bradykinin in the pulmonary circulation. Biochem J 110:795–797
- Ryan JW, Roblero J, Stewart JM (1970) Inactivation of bradykinin in rat lung. Adv Exp Med Biol 8:263–274
- Rzoska J (1974) The Upper Nile swamps: a tropical wetland study. Freshwater Biol 4:1-30

Sacca R, Burggren WW (1982) Oxygen uptake in water and air in the air-breathing reedfish *Calamoichthys calabaricus* role of skin, gills and lungs. J Exp Biol 97:179–186

- Sachs G (1977) Ion pumps in the renal tubule. Am J Physiol 233:F359-365
- Saeed SA, Roy AC (1972) Purification of 15-hydroxyprostaglandin dehydrogenase from bovine lung. Biochem Res Commun 47:96–107
- Safford-Black V (1944) Gas exchange in the simbladder of the mudminnow, Umbra limi (Kirtland). Proc Nova Scotia Inst Sci 21:1–22
- Saga S, Modell HJ, Calderwood HW, Lucas AJ, Tham ML, Swenson EW (1973) Pulmonary function after ventilation with fluorocarbon liquid P-12F (caroxin-F). J Appl Physiol 34:160–164
- Sagan C (1994) The search for extraterrestrial life. Sci Am 271:71-77
- Sagan C, Chyba C (1997) The early faint sun paradox: organic shielding of ultraviolet-labile greenhouse gases. Science 276:1217–1221
- Salomonsen F (1967) Migratory movements of the Arctic tern (*Sterna paradisea pontoppidan*) in the Southern Ocean. Det Kgl Dan Videns Selsk Biol Med 24:1–37
- Saltin B (1985) Malleability of the system in overcoming limitations: functional elements. J Exp Biol 115:345–354
- Saltin B, Gollnick PD (1983) Skeletal muscle adaptability: singificance for metabolism and performance. In: Peachy LD, Adrian RH, Geiger SR (eds) Handbook of physiology: skeletal muscle. Williams and Wilkinson, Baltimore, pp 555–631

- Samuelson RE, Hanel RA, Kunde VG, Maguine WC (1981) Mean molecular weight and hydrogen abundance of Titan's atmosphere. Nature (Lond) 292:688-698
- Sanderson RJ, Paul CW, Vatter AE, Filley GF (1976) Morphological and physical basis for lung surfactant action. Respir Physiol 27:379–392
- Sandreuter A (1951) The structure and function of the avian egg. Acta Anat II, Suppl 14, pp 1– 72. Quoted from Romanoff (1960)
- Santos EA, Baldisseroto B, Biachini A, Colares EP, Nery LEM, Manzoni GC (1987) Respiratory mechanisms and metabolic adaptations of an intertidal crab, *Chasmagnathus graulata* (Dana, 1851). Comp Biochem Physiol 88A:21-25
- Sarnthein M, Winn K, Duplessy JC, Fontugne MR (1988) Global variations of surface ocean primary productivity in low and mid latitudes: influence on CO<sub>2</sub> reservoirs of the deep ocean and atmosphere during the last 21 000 years. Paleoceanography 3:361–399
- Sass DJ, Ritman EL, Caskey PE, Banchero N, Wood EH (1972) Liquid breathing: prevention of pulmonary arterial-venous shunting during accelaration. J Appl Physiol 32:451–455
- Sassaman C, Mangum CP (1972) Adaptations to environmental oxygen levels in infaunal and epifaunal sea anemones. Biol Bull 143:657-678
- Sassaman C, Mangum CP (1973) Adaptations to environmental oxygen levels in infaunal sea anemones. Biol Bull Mar Biol Lab Woods Hole 143:657–678
- Sassaman C, Mangum CP (1974) Gas exchange in a cerianthid. J Exp Zool 188:297-305
- Sassone-Corsi P (1996) Same clock, different works. Nature (Lond) 384:613-614
- Satchell GH (1971) Circulation in fishes. Cambridge University Press, Cambridge
- Satchell GH (1976) The circulatory system of air-breathing fish. In: Hughes GM (ed) Respiration of amphibious vertebrates. Academic Press, New York, pp 105–124
- Satchell GH (1984) Respiratory toxicology of fishes. In: Weber LJ (ed) Aquatic toxicology, vol 2. Raven Press, New York, pp 1–35
- Satchell GH (1992) The venous system. In: Hoar WS, Randall DJ, Farrell AP (eds) Fish physiology, vol 12. Academic Press, New York, pp 141-234
- Saunders RL (1953) The swim bladder gas content of some fresh water fish with particular reference to the physostomes. Can J Zool 31:547–560
- Saunders RL, Sutterlin AM (1971) Cardiac and respiratory responses to hypoxia in the sea raven, *Hemitripterus americanus*, and an investigation of possible control mechanisms. J Fish Res Board Can 28:491–503
- Savage RM (1935) The ecology of young tadpoles, with special reference to some adaptations to the habitat of mass spawning in *Rana temporaria* L. Proc Zool Soc (Lond) 605-610
- Sawaya P (1947) Metabolisms respiratoria de Amphibio Gymnophiona, *Typhylonectes compressicauda* (Dum et Bibr). Biol Fac Fil Cien Let U Sao Paulo 1947:51-56
- Saxena DB (1960) On the asphyxiation and influence of CO<sub>2</sub> on respiration of air-breathing fish, Heteropneustes fossilis and Clarias batrachus. J Zool (Lond) 12:114–124
- Saxena DB (1962) Studies on the physiology of respiration in fishes: V. Comparative study of the gill area in the fresh water fishes *Labeo rohita*, *Ophicephalus* (= *Channa*) striatus and *Anabas testudineus*. Ichthyologica 1:59–70
- Saxena DB (1963) A review of ecological studies and their importance in the physiology of air breathing fish. Ichthyologica 2:116–128
- Sayer MDJ, Davenport J (1991) Amphibious fish: why do they leave water? Rev Fish Biol Fisheries 1:159-181
- Scammell CA, Hughes GM (1981) Comparative study of the functional anatomy of the gills and ventilatory currents in some British decapod crustaceans. Biol Bull 156:35–47
- Schaeffer B (1965a) The role of experimentation in the origin of higher levels of organization. Syst Zool 14:318–336
- Schaeffer B (1965b) The rhipidistian-amphibian transition. Am Zool 5:267-276
- Schaffer WM, Kot M (1986) Chaos in ecological systems: the coals that Newcastle forgot. Trends Ecol Evol 1:58–63
- Scheid P (1978) Analysis of gas exchange between air capillaries and blood capillaries in the avian lung. Respir Physiol 32:27–49
- Scheid P (1979) Mechanisms of gas exchange in bird lungs. Rev Physiol Biochem Pharmacol 86:137-186

- Scheid P (1985) Significance of lung structure for performance at high altitude. In: Ilyicher VD, Gavrilov VM (eds) Acta XVIII Int Congr of Ornithology, vol III. Nauka, Moscow, 976pp
- Scheid P (1987) Cost of breathing in water- and air-breathers. In: Dejours P, Taylor CR, Weibel ER (eds) Comparative physiology: life on land and water, Fidia Res Series, vol 9. Liviana Press, Padova, pp 83–92
- Scheid P (1990) Avian respiratory system and gas exchange. In: Sutton JR, Coates G, Remmers JE (eds) Hypoxia: the adaptations. BC Decker, Burlington, Ontario, pp 4–7
- Scheid P, Kawashiro T (1975) Metabolic changes in avian blood and their effects on determination of blood gases and pH. Respir Physiol 23:291–300
- Scheid P, Piiper J (1969) Volume, ventilation and compliance of the respiratory system in the domestic fowl. Respir Physiol 6:298–308
- Scheid P, Piiper J (1970) Analysis of gas exchange in the avian lung: theory and experiments in the domestic fowl. Respir Physiol 9:246–262
- Scheid P, Piiper J (1972) Cross-currrent gas exchange in the avian lungs: effects of reversed parabronchial air flow in ducks. Respir Physiol 16:304–312
- Scheid P, Piiper J (1976) Quantitative functional analysis of branchial gas transfer: theory and application to Scyliorhinus stellaris (Elasmobrachii). In: Hughes GM (ed) Respiration of amphibious vertebrates. Academic Press, New York, pp 17–38
- Scheid P, Piiper J (1987) Gas exchange and transport. In: Seller TJ (ed) Bird respiration, vol I. CRC Press, Boca Raton, pp 97–129
- Scheid P, Piiper J (1989) Respiratory mechanics and air flow in birds. In: King AS, McLelland J (eds) Form and function in birds, vol 4. Academic Press, London, pp 369–391
- Scheid P, Worth H, Holle JP, Meyer M (1977) Effects of oscillating and intermittent ventilatory flow on efficacy of pulmonary O<sub>2</sub> transfer in the duck. Respir Physiol 31:251–258
- Scheuer J, Tipton CM (1977) Cardiovascular adaptations to physical training. Annu Rev Physiol 39:221–251
- Schick JM (1991) A functional biology of sea anemones. Chapman and Hall, London
- Schidlowski M (1975) Archean atmosphere and evolution of the terrestrial  $O_2$  budget. In: Windley BF (ed) The early history of the Earth. John Wiley, London, pp 125–201
- Schmalhausen II (1968) The origin of terrestrial vertebrates. Academic Press, Lonon
- Schmid-Schönbein H (1975) Erythrocytes rheology and the optimization of mass transport in the circulation. Blood Cells 1:285-306
- Schmid-Schönbein H (1988) Conceptual proposition for a specific microcirculatory problem: maternal blood flow in hemochorial multivillous placentae as percolation of a "porous medium". Trophoblast Res 3:17-38
- Schmidt H, Kamp G (1996) The "Pasteur effect" in facultative anaerobic metazoa. Experientia 52:440–448
- Schmidt-Nielsen K (1975) Recent advances in avian respiration. In: Peaker M (ed) Avian physiology. Academic Press, London, pp 33–47
- Schmidt-Nielsen K (1984) Scaling: why is animal size so important? Cambridge University Press, Cambridge
- Schmidt-Nielsen K (1990) Animal physiology: adaptation and environment, 4th edn. Cambridge University Press, Cambridge
- Schmidt-Nielsen K, Larimer JL (1958) Oxygen dissociation curves of mammalian blood in relation to body size. Am J Physiol 195:424-428
- Schmidt-Nielsen K, Taylor CR (1968) Red blood cells: why or why not? Science 162:274-275
- Schmidt-Nielsen K, Schmidt-Nielsen B, Jarnum SA, Houpt TR (1957) Body temperature of the camel and its relation to water economy. Am J Physiol 188:103–112
- Schmidt-Nielsen K, Kanwisher J, Lasiewski RC, Cohn JE, Bretz WL (1969) Temperature regulation and respiration in the ostrich. Condor 71:341–352
- Schmidt RS (1982) Possible importance of lung inflation related sensory input to frog calling circuits. Copeia 1982:196–198
- Schneiderman HA (1960) Discontinuous respiration in insects; role of the spiracles. Biol Bull Mar Biol Lab Woods Hole 119:494–528
- Schoene RB, Swenson ER, Pizzo CJ, Hackett PH, Roach RC et al. (1988) The lung at high altitude: bronchoalveolar lavage in acute mountain sickness, and pulmonary oedema. J Appl Physiol 64:2605–2613
- Scholander PF (1954) Secretion of gases against high pressures in the swim bladder of deep sea fishes. II. The rete mirabile. Biol Bull Mar Biol Lab Woods Hole 107:247–259
- Scholander PF (1958) Counter-current exchange. A principle in biology. Hvalrådets Skr 44:1–24 Scholander PF (1960) Oxygen transport through haemoglobin solutions. How does the presence
- of haemoglobin in a wet membrane mediate an eightfold increase in oxygen passage? Science 131:585-590
- Scholander PF, van Dam L (1953) Composition of the swim bladder gas in deep sea fishes. Biol Bull 104:75–97
- Scholey K (1986) The evolution of flight in bats. In: Nachtigall W (ed) BIONA report no 5. Gustav Fischer, Stuttgart, pp 1–12
- Schömig A, Fischer S, Kurz Th, Richardt G, Schömig E (1987) Nonexcytotic release of endogenous noradrenaline in the ischemic and anoxic heart: mechanism and metabolic requirements. Circ Res 60:194-205
- Schopf JW (1978) The evolution of the earliest cells. Sci Am 239:110–138
- Schopf JW (1980) Paleoceanography. Harvard University Press, Cambridge
- Schopf JW (1983) Earth's earliest biosphere: its origins and evolution. Princeton University Press, Princeton
- Schopf JW (1984) Rates of evolution and notion of living fossils. Annu Rev Earth Planet Sci 12:245-292
- Schopf JW (1989) The evolution of the earliest cells. In: Gould JL, Gould CG (eds) Life at the edge: readings from the Scientific American Magazine. WH Freeman, New York, pp 7–23
- Schopf JW (1993) Microfossils of the early Archean Apex chert: new evidence of antiquity of life. Science 260:640–646
- Schopf JW, Oehler DZ (1976) How old are the eukaryotes? Science 193:47-49
- Schopf JW, Walter MR (1983) Archean microfossils: new evidence of ancient microbes. In: Schopf JW (ed) Earth's earliest biosphere: its origin and evolution. Princeton University Press, Princeton, pp 214-239
- Schopf JW, Hayes JM, Walter MR (1983) Evolution of Earth's earliest ecosystems: recent progress and unresolved problems. In: Schopf JW (ed) Earth's earliest biosphere: its origin and evolution. Princeton University Press, Princeton, pp 361-384
- Schöttle E (1932) Morphologie und Physiologie der Atmung bei wasserschlamm- und landlebenden Gobiiformes. Z Wiss Zool 140:1-114
- Schöttler U, Wienhausen G, Zebe E (1983) The mode of energy production in the lugworm *Arenicola marina* at different oxygen concentrations. J Comp Physiol 149:547–555
- Schöttler U, Wienhausen G, Werterman J (1984) Anaerobic metabolism in the lugworm *Arenicola marina* L.: the transition from aerobic to anaerobic metabolism. Comp Biochem Physiol 79B:93-103
- Schumann D, Piiper J (1966) Der Sauerstoffbedarf der Atmung bei Fischen nach Messungen an der narkotisierten Schleie (*Tinca tinca*). Arch Gesamte Physiol Mens Tiere (Pfluegers) 288:14–26
- Schürch S, Bachofen H, Weibel ER (1985) Alveolar surface tension in exercised rabbit lungs: effect of temperature. Respir Physiol 62:31–45
- Schürch S, Gehr P, Hof VI, Geiser M, Green F (1990) Surfactant displaces particles toward the epithelium in airways and alveoli. Respir Physiol 80:17–32
- Schurmann H, Steffensen JF (1992) Lethal oxygen levels at different temperature and the preferred temperature during hypoxia in the Atlantic cod, *Gadus morhua* L. J Fish Biol 41:927– 934
- Schurmann H, Steffensen JF, Lomholt JP (1991) The influence of hypoxia on the preferred temperature of rainbow trout *Oncorhynchus mykiss*. J Exp Biol 157:75-86
- Schwartz JH (1976)  $H^+$  current response to  $CO_2$  and carbonic anhydrase inhibition in turtle bladder. Am J Physiol 231:565–572
- Schwerdtfeger WK (1979) Morphometrical studies of the ultrastructure of the epidermis of the guppy, *Poecilia reticulata* Peters, following adaptation to sea water and treatment with prolactin. Gen Comp Endocrinol 38:476–483
- Scrutton CT (1978) Periodic growth features in fossil organisms and the length of the day and month. In: Brosche P, Sunderman (eds) Tidal friction and the Earth's rotation. Springer Berlin Heidelberg, New York, pp 87-169

- Secomb TW (1991) Erythrocytes mechanics and capillary blood rheology. Cell Biophys 18:231–251
- Seed R (1983) Structural organization, adaptive radiation and classification of molluscs. In: Hochachka PW (ed) The Mollusca, vol I. Academic Press, New York, pp 1–54
- Seeherman HJ, Taylor CR, Maloiy GMO, Armstrong RB (1981) Design of the mammalian respiratory system. II. Measuring maximal aerobic capacity. Respir Physiol 44:11–23
- Seibel BA, Childress JJ (1996) Deep sea breathing cephalopods? Nature (Lond) 384:421
- Seifert R, Schultz C (1991) The superoxide-forming NADPH oxidase of phagocytes: an enzyme system regulated by multiple mechanisms. Rev Physiol Biochem Pharmacol 117:1-338
- Seifriz W (1943) Protoplasmic streaming. Biol Rev 9:49-123
- Selden P, Edwards D (1989) Colonization of the land. In: Allen KC, Briggs DEG (eds) Evolution and ecology. Pinter, London, pp 67-127
- Seliger HH, Boggs JA, Biggley WH (1985) Catastrophic anoxia in the Chesapaeke Bay in 1984. Science 228:70–73
- Semlitsch RD, Wilbur HM (1989) Artificial selection for paedomorphosis in the salamander, Ambystoma talpoideum. Evolution 43:105-112
- Sen CK (1995) Oxidants and antioxidants in excercise. J Appl Physiol 79:675-686
- Serfaty A, Gueutal J (1943) La résistance de la grenouille à l'asphyxie lors d'une immersion prolongée. CR Soc Biol 137:154-156
- Service RF (1997) Microbiologists explore life's rich, hidden kingdoms. Science 275:1740-1742
- Setnikar IE, Agostini E, Taglietti A (1959) The foetal lung, a source of amniotic fluid. Proc Soc Exp Biol Med 101:842–845
- Severinghaus JW (1971) Transarterial leakage: a possible mechanism of high altitude pulmonary oedema. In: Porter R, Knight J (eds) High altitude physiology: cardiac and respiratory aspects. Churchill-Livingstone, Edinburgh, pp 61–77
- Seymour RS (1978) Gas tensions and blood distribution in sea snakes at surface pressure and at simulated depth. Physiol Zool 51:388–407
- Seymour RS (1979) Dinosaur eggs: gas conductance through the shell, water loss during incubation and clutch size. Paleobiology 5:1–11
- Seymour RS (1991) Analysis of heat production in a thermogenic arum lily, Philodendron *selloum* by three calorimetric methods. Thermochim Acta 193:91–97
- Seymour RS (1997) Plants that warm themselves. Sci Am 276:90-95
- Seymour RS, Ackerman RA (1980) Adaptations to underground nesting in birds and reptiles. Am Zool 20:437–457
- Seymour RS, Rahn H (1978) Gas conductance in the eggshell of mound building brush turkey. In: Piiper J (ed) Respiratory function in birds, adult and embryonic. Springer, Berlin Heidelberg New York, pp 243–262
- Seymour RS, Schultze-Motel P (1996) Thermoregulating lotus flowers. Nature (Lond) 383:305
- Seymour RS, Webster MED (1975) Gas transport and blood acid-base balance in diving sea snakes. J Exp Zool 191:169-181
- Seymour RS, Spragg RG, Hartman MT (1981) Distribution of ventilation and perfusion in the sea snake, *Pelamis platurus*. J Comp Physiol 60A;145:109–115
- Shackleton NJ (1993) The climate system in the recent geological past. Philos Trans R Soc Lond 341B:209–213
- Shackleton NJ, Pisias NG (1985) Atmospheric carbon dioxide, orbital forcing and climate. In: Sundquist ET, Broecker WS (eds) The carbon cycle and atmospheric CO<sub>2</sub>: natural variations Archean to present. Geophys Monogr 32:303–317
- Shadwick RE, O'Dor RK, Gosline JM (1990) Respiratory and cardiac function during exercise in squid. Can J Zool 68:792–798
- Shaffer G (1990) A non-linear climate oscillator controlled by biochemical cycling in the ocean: an alternative model for Quatenary ice ages cycles. Clim Dyn 4:127–143
- Shaffer TH, Moskowitz GD (1974) Demand-controlled liquid ventilation of the lungs. J Appl Physiol 36:208–215
- Shaffer TH, Rubenstein D, Moskowitz GD, Delivoria-Papadopoulos M (1976) Gaseous exchange and acid-base balance in premature lambs during liquid ventilation since birth. Pediatr Res 10:227–231

- Shams H, Scheid P (1987) Respiration and blood gases in the duck exposed to normocapnic and hypercapnic hypoxia. Respir Physiol 67:1–12
- Shams H, Scheid P (1989) Efficiency of the parabronchial gas exchange in deep hypoxia measurements in the resting duck. Respir Physiol 77:135–146
- Shannon P, Kramer DL (1988) Water depth alters respiratory behaviour of *Xenopus laevis*. J Exp Biol 137:597–602
- Sheehan PM, Fastovsky DE, Hoffman RG, Berghaus CB, Gabriel DL (1991) Sudden extinction of the dinosaurs: latest Cretaceous, Upper Great Plains, USA. Science 254–835–838
- Sheldon RE, Peeters LLH, Jones MD, Makowski EL, Meschia G (1978) Redistribution of the cardiac output and oxygen delivery in the hypoxemic foetal lamb. Am J Obstet Gynecol 135:1071–1078
- Shelton G (1970) The effect of lung ventilation on blood flow to the lungs and body of the amphibian, *Xenopus laevis*. Respir Physiol 9:183–196
- Shelton G (1976) Gas exchange, pulmonary blood supply and the partially divided amphibian heart. In: Spencer-Davies P (ed) Perspectives in environmental biology. Pergamon Press, Oxford, pp 247–259
- Shelton G (1992) Model applications in respiratory physiology. In: Egginton S, Ross HF (eds) Oxygen transport in biological systems: modelling of pathways. Cambridge University Press, Cambridge, pp 1–44
- Shelton G, Burggren WW (1976) Cardiovascular dynamics of the chelonia during apnea and lung ventilation. J Exp Biol 64:323–343
- Shelton G, Croghan PC (1988) Gas exchange and its control in non-steady state systems: the consequences of evolution from water to air breathing in vertebrates. Can J Zool 66:109–123
- Sheltor G, Jones DR (1965) Central blood pressure and heart output in surfaced and submerged frogs. J Exp Biol 42:339–357
- Shelton G, Jones DR, Milsom WK (1986) Control of breathing in ectothermic vertebrates. In: Cherniack NS, Widdicombe JG (eds) Handbook of physiology, sect 3. Respiratory system, vol 2. Control of breathing. American Physiological Society, Bethesda, pp 857–909
- Shepherd AP, Riedel GL (1982) Optimal hematocrit for oxygenation of canine intestine. Circ Res 51:233–240
- Shepherd SA, Thomas IM (1989) Marine invertebrates of Southern Australia. South Australian Govt Press, Adelaide
- Sheridan MA (1994) Regulation of lipid metabolism in poikilothermic vertebrates. Comp Biochem Physiol 107B:495–508
- Sherman DR, Guinn B, Perdok MM, Goldberg DE (1992) Components of sterol biosynthesis assembled on the oxygen-avid haemoglobin of *Ascaris*. Science 258:1930-1932
- Shield JW, Bentley PJ (1973a) Respiration of some urodele and anuran Amphibia in water. I. Role of the skin and the gills. Comp Biochem Physiol 46A:17–28
- Shield JW, Bentley PJ (1973b) Respiration of some urodele and anuran Amphibia in air. II. Role of the skin and lungs. Comp Biochem Physiol 46A:29–38
- Shiga T (1994) Oxygen transport in microcirculation. Jpn J Physiol 44:19-34
- Shigenaga MK, Gimeno CJ, Ames BN (1989) Urinary 8-Hydroxy-2'-deoxyguanosine as a biological marker of in vivo oxidative DNA damage. Proc Natl Acad Sci USA 86:9697–9701
- Shiklomanov IA (1993) Water in crisis: a guide to the worlds fresh water resources. In: Gleick PH (ed) Land and water use. Oxford University Press, New York, pp 13–24
- Shine R (1983) Reptilian viviparity in cold climates: testing the assumptions of an evolutionary hypothesis. Oecologia 57:397–405
- Shine R (1985) The evolution of viviparity in reptiles: an ecological analysis. In: Gans C, Billett F (eds) Biology of Reptilia, vol 15. John Wiley, New York, pp 604–694
- Shine R (1989) Ecological influences on the evolution of vertebrate viviparity. In: Wake D, Roth G (eds) Complex organismal functions: integration and evolution in vertebrates. (Dahlem Workshop Report), John Wiley, Chichester, pp 263–278
- Shine R, Guillette LJ (1988) The evolution of viviparity in reptiles: a physiological model and its ecological consequences. J Theor Biol 132:43–50
- Shlaifer A, Breder CM (1940) Social and respiratory behaviour of small tarpon. Zoologica 25:493-512

- Shoemaker VH, Balding D, Raubal R, McClanahan LL (1972) Uricotelism and low evaporative water loss in a South American frog. Science 175:1018–1020
- Sibley CG, Ahlquist JE (1990) Phylogeny and classification of birds: a study in molecular evolution. Yale University Press, New Haven
- Sidell BD, Driedzic WR, Stowe DB, Johnston IA (1987) Biochemical correlations of power development and metabolic fuel preferanda in fish hearts. Physiol Zool 60:221-232
- Sidell BD, Vayda ME, Small DJ, Moylan TJ, Londraville RL et al. (1997) Variable expression of myoglobin among the haemoglobinless Antarctic icefishes. Proc Natl Acad Sci USA 94:3420–3424
- Sieker HO, Hickam JB (1956) Carbon dioxide intoxication: the clinical syndrome, its aetiology and management with particular reference to the use of mechanical respirators. Medicine 35:389-408
- Sies H (1991) Oxidative stress: oxidants and antioxidants. Academic Press, Orlando
- Sies H, Cadenas E (1985) Oxidative stress: damage to intact cells and organs. Philos Trans R Soc Lond 311B:617–631
- Siever R (1979) The Earth readings from Scientific American: life origin and evolution. WH Freeman, San Francisco, pp 25–31
- Sikand RS, Magnussen H, Scheid P, Piiper J (1976) Convection and diffusive gas mixing in human lungs: Experiments and model analysis. J Appl Physiol 40:362–371
- Simchon S, Jan KM, Chien S (1987) Influences of reduced red cell deformability on regional blood flow. Am J Physiol 253:H898-H903
- Simkiss K (1980) Eggshell porosity and the water metabolism of the chick embryo. J Zool (Lond) 192:1–18
- Simons GH, Sussana PG (1886) Aquatic respiration in soft-shelled turtles: a contribution to the physiology of respiration in vertebrates. Am Nat 20:233–236
- Simpson GG (1953) The major features of evolution. Columbia University Press, New York
- Simpson RA, Mayhew TM, Barnes PR (1992) From 13 weeks to term, the trophoblast of human placenta grows by the continuous recruitment of new proliferative units: a study of nuclear number using the disector. Placenta 13:501–512
- Sinclair JD (1987) Respiratory drive in hypoxia: carotid body and other mechanisms compared. News Physiol Sci 2:57–69
- Singh BN (1976) Balance between aquatic and aerial respiration. In: Hughes GM (ed) Respiration of amphibious vertebrates. Academic Press, New York, pp 125–164
- Singh BN, Hughes GM (1973) Respiration of an air-breathing catfish *Clarias batrachus* (Linn). J Exp Biol 55:421–434
- Singh BN, Munshi JSD (1968) On the respiratory organs and mechanics of breathing in *Periophthlmus vulgaris*. Zool Anz 183:92-110
- Singh BR, Mishra AP (1980) Development of air-breathing organs in Anabas testudineus (Bloch). Zool Anz 205:359-370
- Singh BR, Mishra AP, Singh RP (1982) Development of the air-breathing organ in the snakeheaded fish, *Channa punctatus*. Zool Anz 208:428-439
- Singh BR, Mishra AP, Singh I (1984) Development of the air-breathing organ in the mud-eel, Amphipnous cuchia (Ham.). Zool Anz 213:395–407
- Singh MP, Khetarpal K, Sharan MA (1980) A theoretical model for studying the rate of oxygenation of blood in pulmonary capillaries. J Math Biol 9:305–330
- Sinha AK, Gleed RD, Hakim TS, Dobson A, Shannon J (1996) Pulmonary capillary pressure during exercise in horses. J Appl Physiol 80:1792–1798
- Siskin M, Katritzky AR (1991) Reactivity of organic compounds in hot water: geochemical and technological implications. Science 254:231–327
- Skalak R, Branemark PI (1969) Deformation of erythrocytes in capillaries. Science 164:717–719 Skerret SJ (1994) Host defenses against respiratory infection. Med Clin North Am 78:941–965 Slater TF (1984) Free radical mechanisms in tissue injury. Biochem J 222:1–15
- Slonim NB, Hamilton LH (1971) Respiratory physiology, 2nd edn. CV Mosby Company, St Louis Smatresk NJ (1979) Scaphognathite activity, heart rate and acid base balance after exercise in the
- coconut crab Birgus latro. Alpha Helix Rep, Scripps Oceanogr Inst, La Jolla, pp 108-119
- Smatresk NJ (1988) Control of the respiratory mode of air-breathing fishes. Can J Zool 148:88– 152

Smatresk NJ (1990) Chemoreceptor modulation of the endogenous respiratory rhythm in vertebrates. Am J Physiol 259:R887–897

- Smatresk NJ (1994) Respiratory control in the transition from water breathing to air breathing in vertebrates. Am Zool 34:264–279
- Smatresk NJ, Azizi SQ (1987) Characteristics of lung mechanoreceptors in spotted gar, a bimodal breather, *Lepisosteus oculatus*. Am J Physiol 252:R1066-R1072
- Smatresk NJ, Cameron JN (1982a) Respiration and acid-base physiology of the spotted gar, a bimodal breather. I. Normal values and the response to severe hypoxia. J Exp Biol 96:263–280
- Smatresk NJ, Cameron JN (1982b) Respiration and acid-base physiology of the spotted gar, a bimodal breather. II. Responses to temperature change and hypercapnia. J Exp Biol 96:281–293
- Smatresk NJ, Burleson ML, Azizi SQ (1986) Chemoreflexive responses to hypoxia and NaCN in longnose gar: evidence for two chemoreceptor loci. Am J Physiol 251:R116-R125
- Smil V (1997) Cycles of life: civilization and the biosphere. Scientific American Library. WH Freeman, New York
- Smits AW, Flanagin JI (1994) Bimodal respiration in aquatic and terrestrial apodan amphibians. Am Zool 34:247–263
- Smith AH (1976) Physiological changes associated with-long term increases in acceleration. In: Sneath PHA (ed) COSPAR: life sciences and space research 14. Akademie-Verlag, Berlin, pp 91–100
- Smith AH (1978) The role of body mass and gravity in determining the energy requirements of homoitherms. In: Holmquist R, Stickland AC (eds) COSPAR: life sciences and space research 16. Pergamon Press, Oxford, pp 83–88
- Smith DG, Campbell G (1976) The anatomy of the pulmonary vascular bed in the toad, Bufo marinus and Xenopus laevis. Cell Tissue Res 178:1-14
- Smith DG, Gannon BJ (1978) Selective control of branchial arch perfusion in an air-breathing Amazonian fish *Hoplerythrinus unitaeniatus*. Can J Zool 56:959–964
- Smith DG, Rapson L (1977) Differences in pulmonary microvascular anatomy between Bufo marinus and Xenopus laevis. Cell Tissue Res 178:1-15
- Smith FGW (1957) Rivers in the sea. Smithson Inst Annu Rep 1956:431-441
- Smith FM, Jones DR (1982) The effect of changes in blood oxygen carrying capacity on ventilation volume in the rainbow trout (*Salmo gairdnen*). J Exp Biol 97:325–334
- Smith HW (1929) The excretion of ammonia and urea by the gills of fish. J Biol Chem 81:727-742
- Smith JH (1985) Breeders must respond to market trends. Poultry-Misset Int 34: January Issue Smith JLB (1956) Old four legs. Longman, London
- Smith JD (1977) Comments on flight and the evolution of bats. In: Hecht MK, Goody PC, Hecht M (eds) Major problems in vertebrate evolution. Plenum Press, New York, pp 427–437
- Smith JC, Ellenberger HH, Ballanyi K, Richter DW, Feldman JL (1991) Pre-Botzinger complex: a brainstem region that may generate respiratory rhythm in mammals. Science 254:726–729
- Smith L, Oseid D, Olson L (1976) Acute and chronic toxicity of hydrogen sulfide to the fathead minnow *Pimephales promelas*. Environ Sci Technol 10:565–568
- Smith RE (1952) Cyprinodont fishes from a sulphur-producing lake in Cyrenaica. Ann Mag Nat Hist Ser 125:888–892
- Smith RE (1956) Quantitative relations between liver mitochondria metabolism and total body weight in mammals. Ann NY Acad Sci 62:403–422
- Smith S (1957) Early development and hatching. In: Brown ME (ed) The physiology of fishes, vol 1. Academic Press, New York, pp 1–57
- Smith TB, Wayne RK, Girman DJ, Bruford MW (1997) A role for ecotones in generating rainforest biodiversity. Science 276:1855–1857
- Smith TG, Marks WB, Lange GD, Sheriff WH, Neale EA (1989) A fractal analysis of cell images. J Neurosci Methods 27:173–180
- Smith U, Ryan JW (1970) An electron Microscopic study of the vascular endothelium as a site for bradykinin and ATP inactivation in the rat lung. Adv Exp Med Biol 8:249–262
- Smith U, Ryan JW (1973a) Electron microscopy of endothelial cells collected on cellulose acetate paper. Tissue Cell 5:333–336
- Smith U, Ryan JW (1973b) Electron microscopy of endothelial and epithelial components of the lungs: correlations of structure and function. Fed Proc 32:1957–1966

Snider GL, Kleinerman J, Thurlbeck WM, Bengali ZH (1985) The definition of emphysema. Report of a National Heart, Lung, and Blood Institute, division of lung disease workshop. Am Rev Respir Dis 132:182–185

Snow DH (1985) The horde and the dog, elite athletes – why and how. Proc Nutr Soc 44:267–272

- Snyder GK (1973) Erythrocyte evolution: the significance of the Fahraeus-Lindqvist phenomenon. Respir Physiol 19:271-278
- Snyder GK (1976) Respiratory characteristics of whole blood and selected aspects of circulatory physiology in the common short-nosed fruit bat, *Cynopterus brachyotes*. Respir Physiol 28:239–247
- Snyder GK (1977) Blood corpuscles and blood haemoglobin: a possible example of coevolution. Science 195:412–421
- Snyder GK (1983) Respiratory adaptations in diving mammals. Respir Physiol 54:269-294
- Sohal RS, Weindruch R (1996) Oxidative stress, calorific restriction, and aging. Science 273:59–63
- Soivio A, Hughes GM (1978) Circulatory changes in secondary lamellae of *Salmo gairdneri*. Ann Zool Fenn 15:221–225
- Soivio A, Nikinmaa M (1981) The swelling of erythrocytes in relation to the oxygen affinity of the blood of the rainbow trout, *Salmo gairdneri* Richardson. In: Pickering AD (ed) Stress and fish. Academic Press, London, pp 103–119
- Soivio A, Tuurala H (1981) Structural and circulatory responses to hypoxia in the secondary lamellae of *Salmo gairdneri* gills at two temperatures. J Comp Physiol 145:37–43
- Solem A (1985) Origin and diversification of pulmonate land snails. In: Trueman ER, Clarke MR (eds) The Mollusca, vol 10, Evolution. Academic Press, London, pp 269–293
- Solomon SE, Purton M (1984) The respiratory epithelium of the lung in the green turtle (Chelonia mydas L). J Anat 139:353-361
- Somero GN (1991) Biochemical mechanisms of cold adaptation and stenothermality in Antarctic fish. In: di Prisco G, Maresca B, Tota B (eds) Biology of Antarctic fish. Springer, Berlin Heidelberg New York, pp 231–287
- Somero GN (1992) Adaptations to high hydrostatic pressure. Annu Rev Physiol 54:557-577
- Somero GN, Childress JJ, Anderson AE (1989) Transport, metabolism, and detoxification of hydrogen sulfide in animals from sulfide-rich marine environments. CRC Crit Rev Mar Aquat Sci 1:591–614
- Sorbini CH, Grassi V, Solinas E, Muiesan G (1968) Arterial oxygen tesion in relation to age in healthy subjects. Respiration 25:3–13
- Sotavalta O (1947) The flight tone (wing stroke frequency) of insects. Acta Entomol Fenn 4:1– 117
- Sparti A (1992) Thermogenic capacity of shrews (Mammalia, Soricidae) and its relationship with basal rate of metabolism. Physiol Zool 65:77–96
- Speckmann EW, Ringer RK (1963) The cardiac output and carotid and tibial blood pressure of the turkey. Can J Biochem Physiol 41:2337–2354
- Sperry DG, Wassersug RJ (1976) A proposd function for microridges on epithelial cells. Anat Rec 185:253–258
- Squires RW, Buskirk ER (1982) Aerobic capacity during acute exposure to simulated altitude, 914 to 2286 m. Med Sci Sports Exercise 14:36–40
- Stadtman ER (1992) Protein oxidation and aging. Science 257:1220-1224
- Stahel CD, Nicol SC (1988) Ventilation and oxygen extraction in the little penguin (*Eudyptula minor*), at different temperatures in air and water. Respir Physiol 71:387–398
- Staines HJ (1965) Female red bat carrying four young. J Mammal 46:333
- Stamler JS, Singel DJ, Loscalzo J (1992) Biochemistry of nitric oxide and its redox-activated forms. Science 258:1898–1902
- Standaert T, Johansen K (1974) Cutaneous gas exchange in snakes. J Comp Physiol 89:313– 320
- Stanislaus M (1937) Untersuchungen an der Kolibrilunge. Z Morphol Oekol Tiere 33:261–289
- Stanley SM (1979) Macroevolution; pattern and process. WH Freeman, San Francisco
- Stanley SM (1987) Extinction: a scientific American book. WH Freeman, San Francisco
- Starck D (1959) Ontogenie und Entwicklungsphysiologie der Säugetiere. Handbuch der Zoologie, 8(1). De Gruyter, Berlin

- Stark-Vancs V, Bell PB, Hutchison VH (1984) Morphological and pharmacological basis for pulmonary ventilation in Amphiuma tridactylum: an ultrastructural study. Cell Tissue Res 238:1-12
- Starling EH, Verney EB (1925) The secretion of urine as studied on the isolated kidney. Proc R Soc Lond 97B:321-363

Staub NC (1974) Pulmonary oedema. Physiol Rev 54:678-811

Staub NC, Nagano H, Paerce NL (1967) Pulmonary oedema in dogs, especially the sequence of fluid accumulation in lungs. J Appl Physiol 22:227–240

- Stearns SC (1982) The role of development in the evolution of life histories. In: Bonner JT (ed) Evolution and development. Springer, Berlin Heidelberg New York, pp 237–258
- Stebbins GL (1984) [Quoted in) Fremontia. January 1984, p 16
- Steen JB (1965) Comparative aspects of the respiratory gas exchange of sea urchins. Acta Physiol Scand 63:164–170
- Steen JB (1971) Comparative physiology of respiratory mechanisms. Academic Press, London
- Steen JB, Berg T (1966) The gills of two species of haemoglobin-free fishes compared to those of other teleosts, with a note on severe anemia in an eel. Comp Biochem Physiol 18:517–526
- Steen JB, Kruysse A (1964) The respiratory function of teleostan gills. Comp Biochem Physiol 12:127-142
- Steffensen JF, Lomholt JP (1983) Energetic cost of active branchial ventilation in the sharksucker *Echeneis naucrates*. J Exp Biol 103:185-192
- Stein JC, Ellsworth ML (1993) Capillary oxygen transport during severe hypoxia: role of haemoglobin oxygen affinity. J Appl Physiol 75:1601–1607
- Steinacker A (1975) Perfusion of the central nervous system of decapod crustaceans. Comp Biochem Physiol 52A:103-104
- Stephens RH, Benjamin AR, Walters DV (1996) Volume and protein concentration of epithelial lining liquid in perfused in situ postnatal sheep lungs. J Appl Physiol 80:1911–1920
- Stephenson J (1930) The Oligochaeta. Clarendon Press, Oxford
- Sterrer W, Rieger R (1974) Retronectidae: a new cosmopolitan marine family of Catenulida (Turbellaria). In: Riser N, Morse M (eds) The biology of Turbellaria. McGraw-Hill, New York, pp 108–147
- Stevens ED, Carey FG (1981) On why of warm-blooded fish. Am J Physiol 240:R151-R175
- Stevens ED, Holeton GF (1978a) The aprtitionin of oxygen uptake from air and from water by erythrinids. Can J Zool 56:965–969
- Stevens ED, Holeton GF (1978b) The partitioning of oxygen uptake from air and from water by the large obligate air breathing teleost, pirarucu (*Arapaima gigas*). Can J Zool 56:974–976
- Stevens ED, Randall DJ (1967) Changes in gas concentrations in blood and water during moderate swimming activity in rainbow trout. J Exp Biol 46:329–337
- Stevenson DJ (1996) When Galileo met Ganymede. Nature (Lond) 384:511-512
- Stewart AG (1978) Swans flying at 8000 m. Brit. Birds 71:459-460
- Stewart CB, Schilling JW, Wilson AC (1987) Adaptive evolution in the stomach lysosomes of foregut fermenters. Nature (Lond) 330:401-404
- Stewart I (1990) Does God play dice?: the mathematics of chaos. Blackwell, Cambridge
- Stewart JR, Thompson MB (1994) Placental structure of the Australian lizard, *Niveoscincus* metallicus (Squamata: Scincidae). J Morphol 220:223-236
- Stiffler DF, DeRuyter ML, Talbot CR (1990) Osmotic and ionic regulation in the aquatic caecilian *Typhlonectes compressicauda* and the terrestrial caecial *Ichthyophis kohtaoensis*. Physiol Zool 63:649–668
- Stinner JN (1982) Functional anatomy of the lung of the snake, *Pituophis melanoleucus*. Am J Physiol 243:R251–257
- Stinner JN (1987) Gas exchange and air flow in the lung of the snake, *Pituophis melanoleucus*. J Comp Physiol 157:307–314
- Stinner JN, Shoemaker VH (1987) Cutaneous gas exchange and low evaporative water loss in the frogs *Phyllomedusa sauvagei* and *Chiromantis xeraphelina*. J Comp Physiol 157B:423-427
- Stoll HM, Schrag DP (1996) Evidence for glacial control of rapid sea level changes in early Cretaceous Arctic Ocean. Science 272:1771–1774
- Stone HO, Thomson HK, Schmidt-Nielsen K (1968) Influence of erythrocytes on blood viscosity. Am J Physiol 214:913–918

- Stong CL (1979) The amateur scientist. Life: origin and evolution: readings from Scientific American. WH Freeman, San Francisco, pp 57–62
- Stonier T (1990) Information and the internal structure of the Universe. Springer, Berlin Heidelberg New York
- Storey KB (1988) Suspended animation: the molecular basis of metabolic depression. Can J Zool (Lond) 66:124–132
- Storey KB (1989) Integrated control of metabolic rate depression via reversible phosphorylation of enzymes in hibernating mammals. In: Malan A, Canguilhem B (eds) Living in cold II. John Libbey Eurotext, Montrouge, pp 309–319
- Storey KB, Storey JM (1986) Freeze-tolerant frogs: cryoprotectants and tissue metabolism during freeze-thaw cycles. Can J Zool 64:49–56
- Storey KB, Storey JM (1988) Freeze tolerance in animals. Physiol Zool 68:27-84
- Storey KB, Storey JM (1990) Facultitative matabolic rate depression: molecular regulation and biomedical adaptation in anerobiosis, hibernation, and estivation. Q Rev Biol 65:145–174
- Stoz F, Schuhmann RA, Schebesta B (1988) The development of the placental villus during normal pregnancy: morphometric data base. Arch Gynecol Obstet 244:23–32
- Strang LB (1967) Uptake of liquid from the lungs at the start of breathing. In: de Reuck AVS, Porter R (eds) Development of the lung. Little Brown, Boston, pp 348–361
- Strang LB (1977) Growth and development of the lung: foetal and postnatal. Annu Rev Physiol 39:253–276
- Strathmann RR (1963) The behaviour of myxine and other Myxinoids. In: Brodal A, Fänge R (eds) The biology of myxine. Universitesforlaget, Oslo, pp 22–32
- Strathmann RR (1990) Why life histories evolve differently in the sea. Am Zool 30:197-207
- Stratton CJ (1984) Morphology of surfactant producing cells and of the alveolar lining layer. In: Robertson B, van Golde LMG, Batenburg JJ (eds) Pulmonary surfactant. Elsevier, Amsterdam, pp 67–118
- Strazny F, Perry SF (1984) Morphometric diffusion capacity and functional anatomy of the book lungs in the spider, *Tegenaria* sp. (Agelenidae). J Morphol 182:339–354
- Strazny F, Perry SF (1987) Respiratory system: structure and function. In: Netwig W (ed) Ecophysiology of the spiders. Springer, Berlin Heidelberg New York, pp 78–94
- Street FA, Grove AT (1976) Late Quartenary lake level fluctuations in Africa: environmental and climatic implications. Nature (Lond) 261:385–390
- Strum J, Junod AF (1972) Autoradiographic demonstration of <sup>3</sup>H-5-hydroxytryptamine uptake by pulmonary endothelial cells. J Cell Biol 54:456–467
- Stuart AJ (1991) Mammalian extinctions in the late Pleistocene of Northern Eurasia and North America. Biol Rev 66:453–562
- Stulc J (1989) Extracellular transport pathways in the haemochorial placenta. Placenta 10:113–119
- Sturgess JM (1979) Mucous secretions in the respiratory tract. Pediatr Clin N Am 26:481-501
- Sturkie PD (1954) Avian physiology. Comstock, Ithaca
- Suarez RK (1992) Hummingbird flight: sustaining the highest mass-specific metabolic rates among vertebrates. Experientia 48:565–570
- Suarez PK, Lighton JRB, Moyes CD, Brown GS et al. (1990) Fuel selection in hummingbirds: ecological implications of metabolic biochemistry. Proc Natl Acad Sci USA 87:9207-9210
- Suarez RK, Lighton JRB, Brown GS, Mathieu-Costello O (1991) Mitochondrial respiration in hummingbird flight muscles. Proc Natl Acad Sci USA 88:4870-4873
- Sugano T, Oshino N, Chance B (1974) Mitochondrial functions under hypoxic conditions. The steady states of cytochrome c reduction and energy metabolism. Biochim Biophys Acta 347:340-358
- Sullivan B, Riggs A (1967) Structure, function, and evolution of turtle haemoglobins III. Oxygenation properties. Comp Biochem Physiol 23:459–474
- Sundin L, Nilsson GE, Block M, Löfman CO (1995) Control of gill filament flow by serotonin in the rainbow trout, *Oncorhynchus mykiss*. Am J Physiol 268:R1224–R1229
- Susan ET, Chadwick OA, Amundson R (1996) Rapid exchange between soil carbon and atmospheric carbon dioxide driven by temperature change. Science 272:393-396
- Suthers RA, Thomas SP, Suthers BJ (1972) Respiration, wing-beat and ultrasonic pulse emission in an echolocating bat. J Exp Biol 56:37–48

- Sverdrup HU, Johnson MW, Fleming RH (1949) The oceans: their physics, chemistry and general biology. Prentice-Hall, New York
- Swain R, Marker PF, Richardson AMM (1987) Respiratory responses to hypoxia in streamdwelling (Astacopsis franklinii) and burrowing (Parastacoides tasmanicus) parastacid crayfish. Comp Biochem Physiol 87A:813–817
- Swan LW (1961) The ecology of the high Himalayas. Sic Am 205:67–78
- Swan LW (1970) Goose of the Himalayas. Nat Hist 79:68-75
- Sweeney BM (1987) Rhythmic phenomena in plants. Academic Press, San Diego

Swenson ER (1990) Kinetics of oxygen and carbon dioxide exchange. In: Boutilier RG (ed) Advances in comparative and environmental physiology, vol 6: Vertebrate gas exchange from environment to cell. Springer, Berlin Heidelberg New York, pp 163–210

- Szarski H (1962) The origin of Amphibia. Q Rev Biol 37:189-241
- Szarski H (1983) Cell size and the concept of wasteful and frugal evolutionary strategies. J Theor Biol 105:201–243
- Szarslo H (1977) Sarcopterygii and the origin of the tetrapods. In: Hecht MK, Goody PC, Hecht BM (eds) Major patterns of vertebrate evolution. Plenum Press, New York, pp 517– 544
- Szathmàry E (1997) The first two billion years. Nature (Lond) 387:662-663
- Szewczak JM, Jackson DC (1992) Apneic oxygen uptake in the torpid bat, *Eptesicus fuscus*. J Exp Biol 173:217–227
- Tait JS (1956) Nitrogen and argon in salmonoid swim bladders. Can J Zool 34:58-62
- Taketo M, Schroeder AC, Mobraaten LE, Gunning KB, Hanten G et al. (1991) FVB/N: an inbred mouse strain preferable for transgenic analyses. Proc Natl Acad Sci USA 88:2065–2069
- Takeuchi H, Uyeda S, Kanamori H (1970) Debate about the Earth. Freeman, Cooper, San Francisco
- Talbot CR, Feder ME (1992) Relationships among cutaneous surface area, cutaneous mass and body mass in frogs: a reappraisal. Physiol Zool 65:1135–1147
- Talling JF (1957) The longitudinal succession of water characteristics in the White Nile. Hydrobiologia 11:73-89
- Tamura M, Hazeki O, Nioka S, Chance B (1989) In vivo study of tissue oxygen metabolism using optical and nuclear magnetic resonance spectroscopies. Annu Rev Physiol 51:813– 834
- Tamura O, Moriyama T (1976) On the morphological feature of the gill of amphibious and airbreathing fishes. Bull Fac Fish, Nagasaki Univ 41:1-8
- Tamura O, Morii H, Yazuriha M (1976) Respiration of amphibious fishes *Periophthalmus* cantonensis and Boleophthalmus chinensis in water and land. J Exp Biol 65:97-107
- Tan AL, De Young A, Noble RW (1972) The pH dependence of the affinity, kinetics and cooperativity of ligand binding to carp haemoglobin, *Cyprinus carpio*. J Biol Chem 247:2493–2498
- Tappan H (1974) Molecular evolution. In: Hayaishi O (ed) Molecular oxygen in biology. Elsevier-North Holland, Amsterdam, pp 81–135
- Tarczy-Hornoch P, Hildebrandt J, Mates EA, Standaert TA, Lamm WJE et al. (1996) Effects of exogenous surfactant on lung pressure-volume characteristics during liquid ventilation. J Appl Physiol 80:1764–1777
- Tarsitano SF, Frey E, Riess J (1989) The evolution of the crocodilia: a conflict between morphological and biochemical data. Am Zool 29:843–856
- Taylor AC (1984) Branchial ventilation in the burrowing crab, *Atelecyclus rotundatus*. J Mar Biol Assoc UK 64:7–20
- Taylor AC, Atkinson RJA (1991) Respiratory adaptations of aquatic decapod crustaceans and fish to a burrowing mode of life. In: Woakes AJ, Grieshaber MK, Bridges CR (eds) Physiological strategies for gas exchange and metabolism. Cambridge University Press, Cambridge, pp 211–234
- Taylor CR (1977) Why large animals? Cornell Vet 67:155–175
- Taylor CR (1987) Structural and functional limits to oxidative metabolism: insights from scaling. Annu Rev Physiol 49:135–146
- Taylor CR, Weibel ER (1981) Design of the mammalian respiratory system. Respir Physiol 11:1– 10

- Taylor CR, Maloiy GMO, Weibel ER, Langman VA, Kamau JMZ et al. (1981) Design of the mammalian respiratory system. III. Scaling maximum aerobic capacity to body mass: wild and domestic mammals. Respir Physiol 44:25–37
- Taylor CR, Karas RH, Weibel ER, Hoppeler H (1987a) Adaptive variation in the mammalian respiratory system in relation to energetic demand: II. Reaching limits to oxygen flow. Respir Physiol 69:7–26
- Taylor CR, Weibel ER, Karas RH, Hoppelar H (1987b) Adaptive variation in the mammalian respiratory system in relation to energetic demand: VIII. Structural and functional design principles determining the limits to oxidative metabolism. Respir Physiol 69:117-127
- Taylor CR, Weibel ER, Karas RH, Hoppelar H (1989) Matching structures and functions in the respiratory system: allometric and adaptive variations in energy demand. In: Wood SC (ed) Comparative pulmonary physiology: current concents. Marcel Dekker, New York, pp 27-65
- Taylor DJ, Mathews PM, Radda GK (1986) Myoglobin-dependent oxidative metabolism in hypoxic rat heart. Respir Physiol 63:275–283
- Taylor EH (1968) The caecilians of the world. University of Kansas Press, Lawrence
- Taylor EW (1982) Control and co-ordination of ventilation and circulation in crustaceans: responses to hypoxia and exercise. J Exp Biol 100:289-319
- Taylor EW, Butler PJ (1973) The behaviour and physiological responses of the shore crab Carcinus maenus during changes in environmental oxygen tension. Neth J Sea Res 7:496-505
- Taylor EW, Butler PJ (1978) Aquatic and aerial respiration in the shore crab, *Carcinus maenas* (L.), acclimated to 15 °C. J Comp Physiol 127:315–323
- Taylor EW, Innes AJ (1988) A functional analysis of the shift from gill- to lung breathing during the evolution of land crabs (Crustacea, Decapoda). Biol J Linn Soc 34:309–316
- Taylor EW, Wheatly MG (1980) Ventilation, heart rate and respiratory gas exchange in the crayfish *Austropotamobius pallipes* (Lereboullet) submerged in normoxic water and following 3-hour exposure in air at 15 °C. J Comp Physiol 138:67–78
- Taylor EW, Butler PJ, Sherlock PJ (1973) The respiratory and cardiovascular changes associated with the emersion response of *Carcinus maenas* (L.) during environmental hypoxia, at three different temperatures. J Comp Physiol 86:95–116
- Taylor HH, Greenaway P (1979) The structure of the gills and lungs of the arid zone crab, *Holthuisana (Austrothelphusa) transversa* Morgens (Sundathelphusidae: Brachyura) including observations on arterial vessels within the gills. J Zool (Lond) 189:359–384
- Taylor HH, Greenaway P (1984) The role of the gills and branchiostegites in gas exchange in a bimodally breathing crab, *Holthuisana transversa*: evidence for a facultative change in the distribution of respiratory circulation. J Exp Biol 11:103–122
- Taylor HH, Wheatly MG (1979) The behaviour and respiratory physiology of the shore crab, *Carcinus maenus* (L.) at moderately high temperatures. J Comp Physiol 130:309–316
- Tazawa H (1987) Embryonic respiration. In: Seller TJ (ed) Bird respiration. CRC Press, Boca Raton, pp 3-41
- Tazawa H, Mochizuki M (1976) Estimation of contact time and diffusing capacity of oxygen in the chollioallantoic vascular plexus. Respir Physiol 28:119–128
- Tazawa H, Mochizuki M (1977) Oxygen analyses of chicken embryo blood. Respir Physiol 31:203-216
- Tazawa H, Visschedijk AHJ, Piiper J (1983a) Blood gases and acid-base status in chicken embryos with naturally varying egg shell conductance. Respir Physiol 53:137–158
- Tazawa H, Visschedijk AHJ, Wittmann J, Piiper J (1983b) Gas exchange, blood gases and acid base status in the chick before, during and after hatching. Respir Physiol 53:173–185
- Teal JM, Kanwisher JW (1966) Gas transport in the marsh grass, Spartina alterniflora. J Exp Bot 17:355–361
- Teasdale F (1978) Functional significance of the zonal morphologic differences in the normal human placenta: a morphometric study. Am J Obstet Gynecol 130:773–781
- Teasdale F (1980) Gestational changes in the functional structure of the human placenta in relation to foetal growth: a morphometric study. J Obstet Gynecol 137:560–568
- Teasdale F, Jean-Jacques G (1986) Morphometry of the microvillous membrane of the human placenta in maternal diabetes mellitus. Placenta 7:81–88

- Teichert C (1988) Main features of cephalopod evolution. In: Clarke MR, Trueman ER (eds) The Mollusca, vol 12, Paleontology and neontology of cephalopods. Academic Press, London, pp 11–79
- Teitel DF, Iwamoto HS, Rudolph AM (1987) Effects of birth-related events on central blood flow patterns. Pediatr Res 22:557–566
- Ten Eyck LF (1972) Stereochemistry of haemoglobin. In: Rorth M, Astrup P (eds) Oxygen affinity of haemoglobin and red cell acid-status. Alfred Benzon Symposium IV, Munksgaard, Copenhagen, pp 19–31
- Tenney SM (1979) A synopsis of breathing mechanisms. In: Wood SC, Lenfant C (eds) Evolution of respiratory processes. A comparative approach. Marcel Dekker, New York, pp 51–106
- Tenney SM (1980) Avian physiology and performance at altitude. In: Sutton JR, Coates GC, Remmers JE (eds) Hypoxia: the adaptations. BC Decker, Toronto, pp 1–3
- Tenney SM, Boggs DF (1985) Comparative mammalian respiratory control. In: Fishman AP (ed) Handbook of physiology: the respiratory system II, sect 3. American Physiological Society, Bethesda, pp 833–855
- Tenney SM, Remmers JE (1963) Comparative morphology of the lung: diffusing area. Nature (Lond) 197:54–56
- Tenney SM, Tenney JB (1970) Quantitative morphology of cold blooded lungs: Amphibia and Reptilia. Respir Physiol 9:197–215
- Tenney SM, Bartlett D, Farber JP, Remmers JE (1984) Mechanics of the respiratory cycle in the green turtle (*Chelonia mydas*). Respir Physiol 22:361–368
- Terroine EF, Roche J (1925) Production calorique et respiration des tissus *in vitro* chez les homèothermes. CR Acad Sci 180:225–227
- Terwilliger NB, Brown C (1993) Ontogeny of hemocyanin function in the Dungeness crab, *Cancer magister*: the interactive effects of developmental stage and divalent cations on hemocyanin oxygenation properties. J Exp Biol 183:1-13
- Terwilliger NB, Terwilliger RC (1984) Haemoglobin from "Pompeii worm", Alvinella pompejana, an annelid from deep sea hot hydrothermal vent environment. Mar Biol Lett 5:191-201
- Theede J, Ponat A, Hiroki K, Schlieper C (1969) Studies on the resistance of marine bottom invertebrates to oxygen deficiency and hydrogen sulfide. Mar Biol 2:325–337
- Thewissen JMG, Babcock SK (1992) The origin of flight in bats: to go where no mammal has gone before. BioScience 42:340–345
- Thomas ALR, Jones G, Rayner JMV, Hughes PM (1990) Intermittent gliding flight in the pipistrelle bat (*Pipistrellus pipistrellus*) (Chiroptera: Vespertilionidae). J Exp Biol 149:407-416
- Thomas DP, Vane JR (1967) 5-hydroxytryptamine in the circulation of the dog. Nature (Lond) 216:335–338
- Thomas DW (1983) The annual migrations of three species of West African fruit bats (Chiroptera: Pteropodidae). Can J Zool 61:2266-2272
- Thomas DW, Cloutier D, Gagne D (1990) Arrythmic breathing, apnea and non-steady-state oxygen uptake in hibernating little brown bats (*Myotis lucifugus*). J Exp Biol 149:395–406
- Thomas HJ (1954) The oxygen uptake of the lobster (*Homarus vulgaris* Edw). J Exp Biol 31:228–251
- Thomas S, Fievet B, Barthelemy L, Peyraud C (1983) Comparisons of exogenous and endogenous hypercapnia on ventilation and oxygen uptake in the rainbow trout (*Salmo gairdneri* R). J Comp Physiol 151B:185–190
- Thomas SP (1975) Metabolism during flight in two species of bats, *Phyllostomus hastatus* and *Pteropus gouldii*. J Exp Biol 63:273-293
- Thomas SP (1981) Ventilation and oxygen extraction in the bat, *Pteropus gouldii*, during rest and during steady flight. J Exp Biol 94:231–250
- Thomas SP (1987) The physiology of bat flight. In: Fenton MB, Racey P, Rayner JMV (eds) Recent advances in the study of bats. Cambridge University Press, Cambridge, pp 75–99
- Thomas SP, Suthers R (1972) The physiology and energetics of bat flight. J Exp Biol 57:317-335 Thomas SP, Lust MR, van Riper HJ (1984) Ventilation and oxygen extraction in the bat
- Phyllostomus hastatus during rest and steady flight. Physiol Zool 57:237-250 Thompson AJ (1911) Introduction to science. Henry Holt, New York

- Thompson D'AW (1959) On growth and form, 2nd edn. Cambridge University Press, Cambridge
- Thompson KS (1969) The environment and distribution of Paleozoic sarcopterygian fishes. Am J Sci 267:457–464
- Thompson KS (1971) The adaptation and evolution of early fishes. Q Rev Biol 46:139-166
- Thompson KS (1980) The ecology of the Devonian lobe-finned fish. In: Panchen AL (ed) The terrestrial environment and the origin of land vertebrates, Systematics Association, Spec Vol 15. Academic Press, London, pp 45–156
- Thompson KS (1991) Where did tetrapods come from? Sci Am 79:488-490
- Thompson DJ (1995) The seasons, global temperature and procession. Science 268:59-68
- Thomson KS (1961) Water miracle of nature. Collier-Macmillan, London
- Thomson KS (1986) Marginalia: a fishy story. Sci Am 74:169-171
- Thorpe WH (1930) The biology, post-embryonic development, and economic importance of *Cryptochaetum iceryae* (Diptera, Agromyzidae) parasitic on *Icerya purchasi* (Coccidae, Monophlebini). Proc Zool Soc Lond 60:929-971
- Thorpe WH (1932) Experiments upon respiration in the larvae of certain parasitic Hymenoptera. Proc R Soc Lond 109B:450-471
- Thorpe WH (1950) Plastron respiration in aquatic insects. Biol Rev 25:344-391
- Thorpe WH, Crisp DJ (1941) Studies on plastron respiration. II. The respiratory efficiency of the plastron in *Amphelocheirus*. J Exp Biol 24:270–303
- Thorpe WH, Crisp DJ (1949) Studies on plastron respiration. IV. Plastron respiration in the Coleoptera. J Exp Biol 26:219–260
- Thurlbeck WM (1980) The effect of age on the lung. In: Dietz AA (ed) Aging its chemistry. American Association for Clinical Chemistry, Washington, DC, pp 114–131
- Tierney DF (1974) Lung metabolism and biochemistry. Annu Rev Physiol 36:209-234
- Tilley SG, Bernado J (1993) Life history evolution in plethodontid salamanders. Herpetologia 49:154–163
- Timiras PS, Krum AA, Pace N (1957) Body and organ weights of rats during acclimatization to an altitude of 12,470 feet. Am J Physiol 191:598–604
- Timwood KT, Julian LM (1983) Early lung growth in the turkey. Proc 32 Western Poult Dis Conf, Davis, pp 21–23
- Timwood KT, Hyde DM, Plopper CG (1987) Lung growth of the turkey, *Meleagris gallopavo*: II. Comparison of two genetic lines. Am J Anat 178:158–169
- Tipton CM (1986) Determinants of VO<sub>2max</sub>: insights gained from non-human species. Acta Physiol Scand 128:33-43
- Toda N, Okamura T (1991) Role of nitric oxide in neurally induced cerebroarterial relaxation. J Pharmacol Exp Ther 258:1027–1032
- Todd ES (1971) Respiratory control in the longjaw, *Gillichthys mirabilis*. Comp Biochem Physiol 39A:147–164
- Todd ES (1973) Positive buoyancy and air-breathing: a new piscine gas bladder function. Copeia (1973):461–464
- Todd ES, Ebeling AW (1966) Aerial respiration in the longjaw mudsucker *Gillichthys mirabilis* (Teleostei: Gobiidae). Biol Bull Mar Biol Lab Woods Hole 130:265–288
- Todd GT (1980) Evolution of the lung and the origin of bony fishes a casual relationship? Am Zool 20:757A
- Toews DP (1971) Factors affecting the onset and termination of respiration in the salamander, Amphiuma tridactylum. Can J Zool 49:1231–1237
- Toews DP, Boutilier RG (1986) Acid-base regulation in the amphibia. In: Heisler N (ed) Acidbase regulation in animals. Elsevier, Amsterdam, pp 266–308
- Toews DP, MacIntyre D (1977) Blood respiratory properties of a viviparous amphibian. Nature (Lond) 266:464–465
- Toews DP, MacIntyre D (1978) Respirattion and circulation in an apodan amphibian. Can J Zool 56:199–214
- Toews DP, Boutilier RG, Todd L, Fuller N (1978) Carbonic anhydrase in the amphibia. Comp Biochem Physiol 59A:211–213
- Toloza EM, Lam M, Diamond JM (1991) Nutrient extraction by cold exposed mice: a test of digestive safety margins. Am J Physiol 261:G608-G620

- Tominaga T, Page EW (1966) Accommodation of the human placenta to hypoxia. J Obstet Gynecol 94:679-691
- Tomlinson JT (1963) Breathing of birds in flight. Condor 514-523
- Torre-Bueno JR (1978) Evaporative cooling and water balance during flight in birds. J Exp Biol 75:231–236
- Torre-Bueno JR (1985) The energetics of avian flight at altitude. In: Nachtigall W (ed) Bird flight, BIONA report 3. Gustav Fischer, Stuttgart, pp 45–87
- Tota B, Hamlett WC (1989) Epilogue: evolutionary and contemporary biology of elasmobranchs. J Exp Zool 2:193-196
- Tota B, Aciero R, Agnisola C (1991) Mechanical performance of the isolated and perfused heart of the haemoglobinless Antarctic icefish *Chionodraco hamatus*. Philos Trans R Soc Lond 332B:191–198
- Toulmond A (1975) Blood oxygen transport and metabolism of the confined *Iugworm Arenicola marina* (L). J Exp Biol 63:647–660
- Toulmond A (1985) Circulating respiratory pigments in marine animals. In: Laverack MS (ed) Physiological adaptations of marine animals. Cambridge University Press, Cambridge, pp 163–206
- Toulmond A (1991) Respiratory and metabolic adaptations of aquatic annelids to low environmental oxygen tensions. In: Woakes AJ, Grieshaber MK, Bridges CR (eds) Physiological strategies for gas exchage and metabolism. Cambridge University Press, Cambridge, pp 191– 210
- Toulmond A, Tchernigovtzeff C (1984) Ventilation and respiratory gas exchanges of the lugworm *Arenicola marina* (L.) as functions of ambient PO<sub>2</sub> (20–700 torr). Respir Physiol 57:349–363
- Toulmond A, Tchernigovtzeff C, Greber P, Jouin C (1984) Epidermal sensitivity to hypoxia in the lugworm. Experientia 40:541–543
- Toulmond A, de Frescheville J, Frisch MH, Jouin C (1988) Les pigments respiratoires de la faune inféodée à l'hydrothermalisme profond. Oceanol Acta 8:195–201
- Towe KM (1970) O<sub>2</sub> collagen priority and early Metazoan fossil record. Proc Natl Acad Sci USA 65:781–788
- Townsend LP, Earnest D (1940) The effects of low oxygen and other extreme conditions on salmonod fish. Proc 6th Pax Sci Congr 3:345–351
- Trench RK (1979) The cell-biology of plant-animal symbiosis. Annu Rev Plant Physiol 30:485– 531
- Trivers R (1985) Social evolution. Benjamin/Cummings, Menlo Park
- Truchot JP (1975) Blood acid-base changes during experimental emersion and reimmersion of the intertidal crab *Carcinus maenas* (L). Respir Physiol 23:351–360
- Truchot JP (1980) Lactate increases the oxygen affinity of crab hemocyanin. J Exp Zool 214:205–208
- Truchot JP (1987) Comparative aspects of extracellular acid-base balance. Springer, Berlin Heidelberg New York
- Truchot JP (1990) Respiratory and ionic regulation in the invertebrates exposed to both water and air. Annu Rev Physiol 52:61–76
- Truchot JP, Jouve-Duhamel A (1980) Oxygen and carbon dioxide in the marine intertidal environment: diurnal and tidal changes in rockpools. Respir Physiol 39:241–254
- Trudinger BJ, Cook CM, Giles WB, Fong E, Connelley A, Wilcox W (1991) Foetal umbilical artery velocity waveforms and subsequent neonatal outcome. Br J Obstet Gynecol 98:378–384
- Tsonis AA, Tsonis PA (1987) Fractals: a new look biological shape and patterning. Perspect Biol Med 30:355–361
- Tsukimoto K, Mathieu-Costello O, Prediletto R, Elliot AR, West JB (1991) Ultrastructural appearances of pulmonary capillaries at high transmural pressures. J Appl Physiol 71:573–582
- Tucker V (1968) Respiratory physiology of house sparrows in relation to high altitude flight. J Exp Biol 48:55-66
- Tucker V (1970) Energetic cost of locomotion in mammals. Comp Biochem Physiol 34:841–846 Tucker VA (1972) Respiration during flight in birds. Respir Physiol 14:75–82

- Tucker VA (1974) Energetics of natural avian flight. In: Paynter RA (ed.) Avian energetics. Nuttal Ornithological Club, Cambridge, pp 298–333
- Tucker CE, James WE, Berry MA, Johnstone CJ, Glover RF (1976) Depressed myocardial function in the goat at high altitude. J Appl Physiol 41:356-361
- Tulkki P (1965) Dissapearance of the benthic fauna from the basin of Bronholm (Southern Baltic) due to oxygen deficiency. Can Biol Mar 6:455–463
- Tullett SG (1981) Theorerical and practical aspects of eggshell porosity. Turkeys 29:24-58
- Tullett SG, Board RG (1976) Oxygen flux across the integument of the avian egg during incubation. Br Poult Sci 17:441–450
- Tullett SG, Deeming DC (1982) The relationship between eggshell porosity and oxygen consumption of the embryo in the domestic fowl. Comp Biochem Physiol 72A:529–540
- Turrens JF, Crapo JD, Freeman BA (1984) Protection against oxygen toxicity by intravenous injection of liposome-entrapped catalase and superoxide dismutase. J Clin Invest 73:87-95
- Turrens JF, Freeman BA, Crapo JD (1982a) Hyperoxia increases H<sub>2</sub>O<sub>2</sub> release by lung mitochondria and microsomes. Arch Biochem Biophys 217:411-421
- Turrens JF, Freeman BA, Levitt JG, Crapo JD (1982b) The effect of hyperoxia on superoxide production by lung submitochondrial particles. Arch Biochem Biophys 217:401–410
- Turrens JF, Beconi M, Barilla J, Chavez UB, McCord JM (1991) Mitochondrial generation of oxygen radicals during reoxygenation of ischaemic tissues. Free Radic Res Commun 12– 13:681–689
- Tuurala H, Part P, Nikinmaa M, Soivio A (1984) The basal channels of secondary lamellae in *Salmo gairdneri* gills a nonrespiratory shunt. Comp Biochem Physiol 79A:35–39
- Tytler P, Vaughan TC (1983) Thermal ecology of the mudskippers, *Periophthalmus koelreuteri* (Pallas) and *Boleophthalmus boddarti* (Pallas), of Kuwaiti bay. J Fish Biol 23:327–337
- Uchiyama M, Yoshizawa H, Wakasugi C, Oguro C (1990) Structure of the internal gills in tadpoles of the crab-eating frog, *Rana cancrivora*. Biol Sci 7:623-630
- Ulrich SPH, Bartels H (1963) Über die Atmungsfunktion des Blutes von Spitzmäusen, weissen Mäusen und syrischen Goldhamstern. Pfluegers Archiv Gesamte Physiol Menschen Tiere 277:150–165
- Ultsch GR (1973) The effects of water hyacinth (*Eichhornia crassipes*) on the microenvironment of aquatic communities. Arch Hydrobiol 4:460–473
- Ultsch GR (1976) Eco-physiological studies of some metabolic and respiratory adaptations of sirenid salamanders. In: Hughes GM (ed) Respiration of amphibious vertebrates. Academic Press, New York, pp 287-312
- Ultsch GR, Gros G (1979) Mucus, a diffusion barrier to oxygen: possible role in O<sub>2</sub> uptake at low pH in carp (*Cyprinus carpio*) gills. Comp Biochem Physiol 62A:685–689
- Ultsch GR, Jackson DC (1982) Longterm submergence at 3 °C of the turtle, *Chrysemys picta belii*, in normoxic and severely hypoxic water. I. Survival, gas exchange and acid-base status. J Exp Biol 96:11–28
- Ulvedal F, Morgan TE, Cutler RG, Welch BE (1963) Respiratory function studies during prolonged exposure to simulated altitude without hypoxia. USAF School of Aerospace Medicine, Rept No SAM-TDR-63-31, Brooks AFB, Texas
- Untersee P, Gil J, Weibel ER (1971) Visualization of extracellular lining layer of lung alveoli by freeze etching. Respir Physiol 13:171–185
- Urey HC (1959) The atmosphere of the planets, Pt II, The Earth's atmosphere. In: Flügge S (ed) Encyclopedia of physics, LII, Astrophysics III, The solar system. Springer, Berlin Heidelberg New York, pp 366–383
- Usry JL, Turner LW, Stahly TS, Bridges TC, Gates RS (1991) Gl tract simulation model of the growing pig. Trans Am Soc Agric Eng 34:1879–1890
- Vaïda P, Kays CK, Rivière D, Tèchoueyres P, Lachaud JL (1997) Pulmonary diffusing capacity and pulmonary capillary blood volume during parabolic flights. J Appl Physiol 82:1091– 1097
- Val AL, Fonseca VM, Affonso EG (1990) Adaptive features of Amazon fishes: haemoglobins, hematology, intraerythrocytic phosphates and whole blood Bohr effect of *Pterygoplichthys multiradiatus* (Siluriformes). Comp Biochem Physiol 97B:435–440
- Valentine JW, Moores EM (1976) Plate tectonics and history of life in the oceans. In: Wilson IT (ed) Continents adrift and continents aground. WH Freeman, San Fransisco, pp 196–205

- van Beek JHGM, Roger SA, Bassingthwaighte JB (1989) Regional myocardial flow heterogenenity explained with fractal networks. Am J Physiol 257:H1670–1680
- van Dam L (1935) On the utilization of oxygen by Mya arenaria. J Exp Biol 12:86-94
- van Dam L (1938) On the utilization of oxygen and regulation of breathing in some aquatic animals. Diss, Groningen
- van Dam L (1954) On the respiration in scallops (Lamellibranchia). Biol Bull Mar Biol Lab Woods Hole 107:192–202
- van der Burgh J, Visscher H, Dilcher DL, Kurschner WM (1993) Paleoatmospheric signatures on Neogene fossil leaves. Science 260:1788–1790
- Vandergriff KD, Olson JS (1984) Morphological and physiological factors affecting oxygen uptake and release by red blood cells. J Biol Chem 259:12619–12627
- Vane JR (1969) The release and fate of vasoactive hormones in pulmonary circulation. Br J Pharmacol 35:209–242
- van Holde KE, Miller KI (1982) Hemocyanins. Q Rev Biophys 15:1-129
- Vannier G (1983) The importance of ecophysiology for both biotic and abiotic studies of take in the rainbow trout (Sads in soil biology). Ottignies-Louvain-La-Neuve, Dieu-Brichart, pp 289-314
- van Oijen MJP, Witte F, Witte-Mass ELM (1981) Introduction to ecology and taxonomic investigations on the haplochromine cichlids from the Mwanza Gulf of Lake Victoria. Neth J Zool 31:149–174
- van Valen L (1971) The history and stability of atmospheric oxygen. Science 171:439-443
- van Valen L (1979) The evolution of bats. Evol Theory 4:103-121
- van Vass O, Vass KF (1960) Appraisal of inland fishery resources. Biological appraisal. In: Lectures presented at the 3rd Int Training Congr. FAO, Bogor, Indonesia (1955). 1:199
- van Zalen MM, van Vugt JMG, Colebrander GJ, van Geijn HP (1994) First-trimester uteroplacental and foetal blood flow velocity waveforms in normally developing fetuses: a longitudinal study. Ultrasound Obstet Gynecol 4:284–288
- Varansi U, Markey D (1978) Uptake and release of lead and cadmium in skin and mucus of coho salmon (*Oncorhynchus kitutch*). Comp Biochem Physiol 60C:187–191
- Varansi U, Robisch PA, Malins DC (1975) Structural alterations in fish epidermal mucus produced by water-borne lead and mercury. Nature (Lond) 258:431-432
- Vaughan TA (1966) Morphology and flight characteristics of mollosid bats. J Mammal 47:75-82
- Veerannan KM (1974) Respiratory metabolism of crabs from marine and estuarine habitats: an interspecific comparison. Mar Biol 26:35–43
- Veith WJ (1980) Viviparity and embryonic adaptations in the teleost *Clinus superciliosus*. Can J Zool 58:1–12
- Verbanck S, Linnarsson D, Prisk GM, Paiva M (1996) Specific ventilation distribution in microgravity. J Appl Physiol 180:1458–1465
- Verde MR (1951) The morphology and histology of the lung in snakes. J Univ Bombay 19:79-89
- Verdier B (1975) Etude de l'atmosphère du sol. Eléments de comparaison et signification écologique de l'atmosphère d'un sol brun calcaire et d'on sol lessivé podzolique. Rev Ecol Biol Sol 12:591-626
- Vergara AG, Hughes GM (1981) Phospholipids in washings from the lung of the frog (*Rana pipiens*). J Comp Physiol 27:117–120
- Verma A, Hirsch DJ, Glatt CE, Ronnett GV, Snyder SH (1993) Carbon monoxide: a putative neural messanger. Science 259:381–384
- Vernberg FJ (1954) The respiratory metabolism of tissues of tissues of marine teleosts in relation to activity and body size. Biol Bull 106:360–370
- Vetter RD, Wells ME, Kurtsman AL, Somero GN (1987) Sulfide detoxification by the hydrothermal vent crab *Bythograea thermydron* and other decapod crustaceans. Physiol Zool 60:121– 137
- Vidal RA, Bahr D, Baragiola RA, Peters M (1997) Oxygen on Ganymede: laboratory studies. Science 276:1839–1842
- Vidyadaran MK, King AS, Kassim H (1987) Deficient anatomical capacity for oxygen uptake of the developing lung of the female domestic fowl when compared with red-jungle fowl. Schweiz Arch Tierheilkd 129:225–237

- Vidyadaran MK, King AS, Kassim H (1988) Quantitative studies of the lung of the domestic fowl, Gallus gallus var domesticus. Partanika 11:229–238
- Vidyadaran MK, King AS, Kassim H (1990) Quantitative comparisons of lung structure of adult domestic fowl and the red-jungle fowl, with reference to broiler ascites. Avian Pathol 19:51–58
- Vince M (1973) Effects of external stimulation on the onset of lung ventilation and the time of hatching in the fowl, duck and goose. Br Poult Sci 14:389–405
- Vince M, Misson BH, Freeman BM (1975) Blood gas partial pressures and the onset of lung ventilation in the chick embryo. Comp Biochem Physiol 51A:457–469
- Viscor G, Fuentes J, Palomeque J (1984) Blood rheology in the pigeon (*Columba livia*), hen (*Gallus gallus domesticus*), and black-headed gull (*Larus ridibundus*). Can J Zool 62:2150– 2156
- Visschedijk AHJ (1968a) The air space and embryonic respiration. II. The times of pipping and hatching as influenced by artificially changed permeability of the shell over the air space. Br Poult Sci 9:185–196
- Visschedijk AHJ (1968b) The air space and embryonic respiration. III. The balance between oxygen and and carbon dioxide in the air space of incubating chicken egg and its role in stimulating pipping. Br Poult Sci 9:197–213
- Visschedijk AHJ, Rahn H (1981) Incubation of chicken eggs at altitude: theoretical consideration of optimal gas composition. Br Poult Sci 22:451–460
- Visschedijk AHJ, Tazawa H, Piiper J (1985) Variability of shell conductance and gas exchange of chicken eggs. Respir Physiol 59:339–354
- Visser SA (1963) Gas production in the decomposition of Cyperus papyrus. J Water Pollut Control Fed 35:973-788
- Vitalis TZ, Furilla RA, Burggren WW (1988) Ventilation and gas exchange in the snake, *Thamnophis elegans*. Am Zool 28:47A
- Vleck D (1979) The energy cost of burrowing by the pocket gopher *Thomomys bottae*. Physiol Zool 52:122–125
- Vleck D (1981) Burrow structure and foraging costs in the fossorial rodent, *Thomomys bottae*. Oecologia (Berl) 49:391–396
- Vogel G (1997) Parasite shed light on cellular evolution. Science 275:1422
- Vogel GM (1980) Oxygen uptake and transport in the sabellid polychaete *Eudistylia vancouveri* (Kinburg). MA Thesis, College of William and Mary, Williamsburg, Virginia
- Vogel S (1977) Flows in organisms induced by movement of the external medium. In: Pedley TJ (ed) Scale effects in animal locomotion. Academic Press, London, pp 285–297
- Vogel S (1988) Life's devices: the physical world of animals and plants. Princeton University Press, Princeton
- Vogel S, Wainright SA (1969) A functional bestiary. Addison-Wesley, Reading

Vollrath F (1992) Spider webs and silks. Sci Am 266:70-76

- Von Bertalanffy L, Estwick RR (1953) Tissue respiration of musculature in relation to body size. Am J Physiol 173:58–60
- Von Sonntag C (1987) The chemical basis of radiation biology. Taylor and Francis, London
- Voss RF (1988) Fractals in nature: from characterization to simulation. In: Petgen HO, Saupe D (eds) The science of fractal images. Springer, New York, pp 21–770
- Vyas AB, Laliwala SM (1976) Anatomical studies on the book lungs of the scorpion *Buthus tamulus* with a note on the respiratory mechanism. J Anim Morphol Physiol 23:3–7
- Waarde V, Thillart VD, Kesbeke F (1983) Anaerobic energy metabolism of the European eel, Anguilla anguilla L. J Comp Physiol 149:469–475
- Wache S, Terwilliger NB, Terwilliger RC (1988) Hemocyanin structure changes during early development of the crab *Cancer productus*. J Exp Biol 247:23–32
- Wagner F, Below R, Klerk PM, Dilcher DL, Joosten H, Kürschner WM, Visscher H (1996) A natural experiment on plant acclimation: lifetime stomatal atmospheric carbon dioxide increase. Pro Natl Acad Sci USA 93:11705-11708
- Wagner GP (1989) The origin of morphological characters and the biological basis of homology. Evolution 43:1157–1171
- Wagner JR, Hu CC, Ames BN (1992) Endogenous oxidative damage of deoxycytidine in DNA. Proc Natl Acad Sci USA 89:3380-3384

Wagner PD (1977) Diffusion and chemical reaction of pulmonary gas exchange. Physiol Rev 57:257-312

Wagner PD (1993) Algebraic analysis of the determinants of VO<sub>2max</sub>. Respir Physiol 93:221–237

Wagner PD, West JB (1972) Effects of diffusion impairment on O<sub>2</sub> and CO time courses in pulmonary capillaries. J Appl Physiol 33:62–71

Wagner PD, Gale GE, Moon RE, Torre-Bueno JR, Stolp BW, Saltman HA (1986) Pulmonary gas exchange in humans exercising at sea level and simulated altitude. J Appl Physiol 61:260–270

Wagner S, Castel M, Gainer H, Yarom Y (1997) GABA in the mammalian suprachiasmic nucleus and its role in diurnal rhythmicity. Nature (Lond) 387:598–603

Wainright SA (1988) Form and function in organisms. Am Zool 28:671-680

Wainright SA, Biggs WD, Currey JD, Gosline JM (1976) Mechanical design in organisms. Wiley, New York

Wake MH (1974) The comparative morphology of the caecilian lung. Anat Rec 178:483

Wake MH (1977) The reproductive biology of caecilians: an evolutionary perspective. In: Taylor DH, Guttman SI (eds) The reproductive biology of amphibians. Plenum Press, New York, pp 73–101

 Wake MH (1989) Phylogenesis of direct development and viviparity in vertebrates. In: Wake D, Roth G (eds) Complex organismal functions: integration and evolution in vertebrates. (Dahlem Workshop Report), John Wiley Chichester, pp 235–250

Wake MH (1990) The evolution of integration of biological systems: an evolutionary perspective through studies on cells, tissues, and organs. Am Zool 30:897–906

Wake MH (1993) Evolution of oviductal gestation in amphibians. J Exp Zool 266:394-413

Wake MH, Marks SB (1993) Development and evolution of plethodontid slalmanders: a review of prior studies and a prospectus for future research. Herpetologia 49:194–203

Wake MH, Roth G (1989) Paedomorphosis: new evidence for its importance in salamander evolution. Am Zool 29:134A

Walker BR, Voelkel NF, McMurtry IF, Adams EM (1982) Evidence for diminished sensitivity of the hamster pulmonary vasculature to hypoxia. J Appl Physiol 52:1571–1574

Walker BR, Berend N, Voelkel NF (1984) Comparison of muscular pulmonary arteries in low and high altitude hamsters and rats. Respir Physiol 56:45–50

Walker JCG (1974) Stability of the atmospheric oxygen. Am J Sci 274:193-214

Walker JCG (1977) Evolution of the atmosphere. Macmillan, New York

Walker JCG (1978) Oxygen and hydrogen in the primitive atmosphere. Pure Appl Geophys 116:222-231

- Walker JCG (1983) Possible limits on the composition of the Archean Ocean. Nature (Lond) 302:518–520
- Walker JCG (1985) Carbon dioxide on the early earth. Origin Life Evol Biosphere 16:117-127

Walker JCG (1987) Was the Archean biosphere upside down? Nature (Lond) 329:710-712

Walker JCG, Klein C, Schidlowski M, Schopf JW, Stevenson DJ, Walter MR (1983) Environmental evolution of the Archean-early Paleozoic Earth. In: Schopf W (ed) The Earth's earliest biosphere: its origin and evolution. Princeton University Press, Princeton, pp 260–290

Walker JG (1970) Oxygen poisoning in the annelid *Tubifex tubifex*. I. Response to oxygen exposure. Biol Bull 138:235-244

Walker RM, Johansen PH (1977) Anaerobic metabolism in the goldfish (*Carassius auratus*). Can J Zool 55:1304–1331

Walkinshaw LH (1945) Aortic rupture in field sparrow due to fright. Auk 62:141

Wallengren H (1914) Physiolog.-Biolog. Studien über die Atmung bei den Arthropoden. III. Die Atmung der Aeschnalarven. Lunds Univ Aasskr NF Avd 10:1–28

Walsby AE (1972) Gas-filled structures providing buoyancy in photosynthetic organisms. In: Soc exp biol symp No 26, The effects of pressure on organisms. The Company of Biologists, Cambridge, pp 233–250

Walshe BM (1948) The oxygen requirements and thermal resistance of chironomid larvae from flowing and from still waters. J Exp Biol 25:35–44

Walshe BM (1950) The function of haemoglobin in *Chironomus plumosus* under natural conditions. J Exp Biol 27:73–95

Walters DV, Olver RE (1978) The role of catecholamines in lung liquid absorption at birth. Pediatr Res 12:239–242

- Wang J, Welkowitz TW, Kostis J, Semmlow J (1989) Incremental network analogue model of the coronary artery. Med Biol Eng Comput 27:416–422
- Wang LCH (1978) Energetic and field aspects of mammalian torpor: the Richardson's ground squirrel. In: Wang LCH, Hudson JW (eds) Strategies in cold: natural torpidity and thermogenesis. Academic Press, New York, pp 109–145
- Wang N, Banzett RB, Butler JP, Fredberg JJ (1988) Bird lung models show that convective inertia effects inspiratory aerodynamic valving. Respir Physiol 73:109–124
- Wang N, Banzett RB, Nations CS, Jenkins F (1992) An aerodynamic valve in the avian primary bronchus. J Exp Zool 262:441-445
- Wangensteen OD (1972) Gas exchange by a bird's embryo. Respir Physiol 14:64-74
- Wangensteen OD, Rahn H (1970) Respiratory gas exchange by the avian embryo. Respir Physiol 11:31–45
- Wangensteen OD, Weibel ER (1982) Morphometric evaluation of chorioallantoic oxygen transport in the chick embryo. Respir Physiol 47:1–20
- Wangensteen OD, Wilson D, Rahn H (1971) Diffusion across the shell of the hen's egg. Respir Physiol 11:16-30
- Wangensteen OD, Rahn H, Burton RR, Smith AH (1974) Respiratory gas exchange of high altitude-adapted chick embryos. Respir Physiol 21:61-70
- Ward DM, Weller R, Bateson MM (1990) 16S rRNAsequences reveal numerous uncultured inhabitants in a natural community. Nature (Lond) 345:63-65
- Warneck P (1988) Chemistry of the natural atmosphere. Academic Press, San Diego
- Warner ACI (1981) Rate of passage of digesta through the gut of mammals and birds. Nutr Abstr Rev 51B:789–820
- Wasawo DPS, Visser SA (1959) Swamp worms and tussock mounds in the swamps of Teso, Uganda. E Afr Agric J 25:86–90
- Wasserman K (1994) Coupling of external to cellular respiration during exercise: the wisdom of body revisited. Am J Physiol 29:E159–E539
- Wasserman K, Butler J, Kessel VA (1966) Factors affecting the capillary blood blood flow pulse in amn. J Appl Physiol 21:890-900
- Wasserzug RJ, Paul RD, Feder ME (1981) Cardiorespiratory synchrony in anuran larvae (Xenopus laevis, Pachymedusa dacnicolor, and Rana berlandien). Comp Biochem Physiol 70A:329-334
- Wassnetzov W (1932) Über die Morphologie der Schwimmblase. Zool Jahrb Abt Anat Ont Tiere 56:1–36
- Waugh RE, Evans EA (1976) Viscoelastic properties of erythrocyte membranes of different vertebrate animals. Micovasc Res 12:291-304
- Weathers WW (1976) Influence of temperature on the optimal hematocrit of the bullfrog, *Rana catesbeiana*. J Comp Physiol 105:173–184
- Webb CL, Milsom WK (1994) Ventilatory responses to acute and chronic hypoxic hypercapnia in ground squirrel. Respir Physiol 98:137–152
- Webber PJ (ed) (1979) High altitude geoecology. Westview, Boulder
- Weber RE (1978) Respiration. In: Mill JP (ed) Physiology of annelids. Academic Press, London, pp 369–392
- Weber RE (1992) Molecular strategies in the adaptation of vertebrate haemoglobin function. In:
   Wood SC, Weber RE, Hargens AR, Millard RW (eds) Physiological adaptations in vertebrates: respiration, circulation, and metabolism. Marcel Dekker, New York, pp 257–277
- Weber RE, Hartvig M (1984) Specific foetal haemoglobin underlies the foetal-maternal shift in blood oxygen affinity in a viviparous teleost. Mol Physiol 6:27-32
- Weber RE, Jesnen FB (1988) Functional adaptations in haemoglobins from ectothermic vertebrates. Annu Rev Physiol 50:161–179
- Weber RE, Lykkeboe G (1978) Respiratory adaptations in carp blood. Influences of hypoxia, red cell organic phosphates, divalent cations and CO<sub>2</sub> on haemoglobin-oxygen affinity. J Comp Physiol 128:127–137
- Weber RE, Pauptit E (1972) Molecular and functional heterogeneity in myoglobin from the polchaete *Arenicola marina* (L). Arch Biochem Biophys 148:322–324
- Weber RE, Wood SC, Lomholt JP (1976) Temperature acclimation and oxygen binding properties of blood and multiple haemoglobins of rainbow trout. J Exp Biol 65:333-345

- Weber RE, Wood SC, Davis BJ (1979) Acclimation to hypoxic water in facultative airbreathing fish: blood oxygen affinity and allosteric effectors. Comp Biochem Physiol 62A:125-129
- Weber RE, Jesnen TH, Malte H, Tame J (1993) Mutant haemoglobin ( $\alpha$ -Ala and  $\beta$ -Ser): functions related to high-altitude respiration in geese. J Appl Physiol 75:2646–2655
- Wedler FC (1987) Determination of molecular heat stability. In: Henle KJ (ed) Thermotolerance, vol II: Mechanisms of heat tolerance. CRC Press, Boca Raton, pp 1–18
- Weekes HC (1935) Review of placentation among reptiles with particular regard to the function and evolution of the placenta. Proc Zool Soc (Lond) 2:625–645
- Weibel ER (1963) Morphometry of the human lung. Springer, Berlin Heidelberg New York
- Weiber ER (1970/71) Morphometric estimation of pulmonary diffusion capacity. I. Model and method. Respir Physiol 11:54–75
- Weibel ER (1973) Morphological basis of the alveolar-capillary gas exchange. Physiol Rev 53:419-495
- Weibel ER (1979) Oxygen demand and size of respiratory structures in mammals. In: Wood SC, Lenfant C (eds) Evolution of respiratory processes: a comparative approach. Marcel Dekker, New York, pp 289–346
- Weibel ER (1982) The pathway for oxygen: lung to mitochondria. In: Taylor CR, Johansen K, Bolis L (eds) A companion to animal physiology. Cambridge University Press, Cambridge, pp 31–48
- Weibel ER (1983a) How does lung structure affect gas exchange? Chest 83:657-665
- Weibel ER (1983b) Is the lung built reasonably? Am J Respir Dis 128:752-760
- Weibel ER (1984a) The pathways for oxygen. Harvard University Press, Harvard
- Weibel ER (1984b) Lung cell biology. In: Fishman AP, Fisher AB (eds) Handbook of physiology: respiration, vol 4. American Physiological Society, Washington, DC, pp 47–91
- Weibel ER (1985a) Design performance of muscular systems: an overview. J Exp Biol 115:405-412
- Weibel ER (1985b) Lung cell biology. In: Fishman AP (ed) Handbook of physiology, vol III, sect 2. American Physiological Society, Bethesda, pp 47–91
- Weibel ER (1986) Functional morphology of lung parenchyma. In: Handbook of physiology. The respiratory system. Mechanics of breathing, sect 3, vol III, chapt 8. Am Physiol Soc, Bethesda, pp 89–111
- Weibel ER (1987) Scaling of structural and functional variables in the respiratory system. Ann Rev Physiol 49:147–159
- Weibel ER (1989) Lung morphometry and models in respiratory physiology. In: Chang HK, Paiva M (eds) Respiratory physiology: an analytical approach. Marcel Dekker, New York, pp 1–56
- Weibel ER (1990) Morphometry: stereological theory and practical methods. In: Gill J (ed) Models of lung disease: microscopy and structural methods. Marcel Dekker, New York, pp 199–251
- Weibel ER (1991) Fractal geometry: a design principle for living organisms. Am J Physiol 261:L361-L369
- Weibel ER (1994) Design of biological organisms and fractal geometry. In: Nonnenmacher T, Losa GA, Weibel ER (eds) Fractals in biology and medicine. Birkhäuser, Basel, pp 68–85
- Weibel ER (1996) The structural basis of lung function. In: West JB (ed) Respiratory physiology: people and ideas. Oxford University Press, New York, pp 3–46
- Weibel ER, Gil J (1968) Electron microscopic demonstration of an extracellular duplex lining layer of alveoli. Respir Physiol 4:42–57
- Weibel ER, Taylor CR (1986) Morphometric modelling of pulmonary diffusing capacity. Prog Respir Res 21:52–55
- Weibel ER, Untersee P, Gil J, Zulauf M (1973) Morphometric estimation of pulmonary diffusion capacity. VI. Effect of varying positive pressure inflation of air spaces. Respir Physiol 18:285–308
- Weibel ER, Taylor CR, O'Neil JJ, Leith DE, Gehr P, Hoppeler H, Langman V, Baudinette RV (1983) Maximal oxygen consumption and pulmonary diffusing capacity: a direct comparison of physiologic and morphometric measurements in canids. Respir Physiol 54:173–188

- Weibel ER, Taylor CR, Hoppeler H, Karas RH (1987a) Adaptive variation in the mammalian respiratory system in relation to energetic demand: I. Introduction to problem and strategy. Respir Physiol 68:1-6
- Weibel ER, Marques LB, Constantinopol F, Doffey F, Gehr P, Taylor CR (1987b) Adaptive variation in the mammalian respiratory system in relation to energetic demand: VI. The pulmonary gas exchanger. Respir Physiol 69:81–100
- Weibel ER, Taylor CR, Hoppeler H (1991) The concept of symmorphosis: A testable hypothesis of structure-function relationship. Proc Natl Acad Sci 88:10357-10361
- Weibel ER, Federspiel WJ, Doffey FF, Hsia CCW, König M, Navarro VS, Vock R (1993) Morphometric model for pulmonary diffusing capacity. I. Membrane diffusing capacity. Respir Physiol 93:125-149
- Weibel ER, Taylor CR, Bolis L (eds) (1998) Principles of animal design: the optimization and symmorphosis debate. Cambridge University Press, London, pp 1–314
- Weidenschilling SJ, Marzari F (1996) Gravitational scattering as a possible origin for giant planets at small stellar distances. Nature (Lond) 384:619-620
- Weinberg S (1994) Life in the Universe. Sci Am 271:22-27
- Weingarden M, Mizukami H, Rice SA (1982) Factors defining the rate of oxygen uptake by the erythrocytes. Bull Math Biol 44:135–147
- Weinstein Y, Bernstein MH, Bickler PE, Gonzales DV, Samaniego FC, Escobedo MA (1985) Blood respiratory properties in pigeons at high altitudes: effects of acclimation. Am J Physiol 249:R765–R776
- Weis-Fogh T (1964a) Diffusion in insect flight muscle, the most active tissue known. J Exp Biol 41:229–256
- Weis-Fogh T (1964b) Functional design of the tracheal system of flying insects as compared with the avian lung. J Exp Biol 41:207–228
- Weis-Fogh T (1967) Respiration and tracheal ventilation in locusts and other flying insects. J Exp Biol 47:561-587
- Weis-Fogh T (1972) Energetics of hovering flight in hummingbirds and in *Drosophila*. J Exp Biol 56:79–104
- Weis-Fogh T (1973) Quick estimates of flight fitness in hovering animals, including novel mechanisms for lift production. J Exp Biol 59:169–230
- Weiss HS (1978) The role of shell diffusion area in incubating eggs at simulated altitude. J Appl Physiol 45:551–556
- Welch HG (1987) Effects of hypoxia and hyperoxia on human performance. Exercise Sport Sci Rev 15:191–221
- Welch HG, Pedersen PK (1981) Measurement of metabolic rate in hyperoxia. J Appl Physiol 51:725-731
- Welch PS (1952) Limnology. McGraw Hill, New York
- Wells DJ (1993a) Muscle performance in hovering hummingbirds. J Exp Biol 178:39-57
- Wells DJ (1993b) Ecological correlates of hovering flight of hummingbirds. J Exp Biol 178:59-70
- Wells GP (1949) Respiratory movements of *Arenicola marina* L. Intermittent irrigation of the tube and intermittent aerial respiration. J Mar Biol Assoc UK 28:447-464
- Wells GP (1952) The respiratory significance of the crown in the polychaete worms *Sabella* and *Myxicola*. Proc R Soc Lond 138B:278–299
- Wells GP (1966) The lugworm (Arenicola): a study in adaptation. Neth J Sea Res 3:294-313

Wells MJ (1962) Brain and behaviour in cephalopods. Heineman, London

- Wells MJ (1983) Circulation in the cephalopods. In: Saleuddin ASM, Wilbur KM (eds) The Mollusca, vol 5: Physiology, part 2. Academic Press, New York, pp 239–290
- Wells MJ, Wells J (1984) The effects of reducing gill area on the capacity to regulate oxygen uptake and on metabolic scope in a cephalopod. J Exp Biol 108:393-401
- Wells MJ, Wells J (1985) Ventilation and oxygen uptake by Nautilus. J Exp Biol 118:297-312
- Wells MJ, O'Dor RK, Mangold K, Wells J (1983) Oxygen consumption in movement by Octopus. Mar Behav Physiol 9:289–303
- Wells MJ, Hanlon RT, Lee PG, DiMarco FP (1988) Respiratory and cardiac performance in *Loliguncula brevis* Blainville, 1823 (Cephalopoda: Myopsida); the effects of activity, temperature and hypoxia. J Exp Biol 138:17–36

- Wells NA, Dorr JA (1985) From and function in the fish *Bothriopepis* (Devonian: Placodermi, Antiarchi): the first terrestrial vertebrate? Mich Acad 17:157-173
- Wells RMG (1990) Haemoglobin physiology in vertebrate animals: a cautionary approach to adaptationists thinking. In: Boutilier RG (ed) Advances in comparative and environmental physiology, vol 6: Vertebrate gas exchange from environment to cell. Springer, Berlin Heidelberg New York, pp 143–162
- Wells RMG, Dales RP (1975) Haemoglobin function in *Terebella lapidaria* L., an intertidal terebellid polychaete. J Mar Biol Assoc UK 55:419-495
- Wells RMG, Weber RE (1982) The Bohr effect of the hemocyanin-containing blood from the terrestrial slug, *Arion ater*. Mol Physiol 2:149–159
- Wells RMG, Jarvis PJ, Shumway SE (1980) Oxygen uptake, the circulatory system, and haemoglobin function in the intertidal polychaete *Terebella haplochaeta* (Ehlers). J Exp Mar Biol Ecol 46:255–277
- Welsch U (1981) Fine structure and enzyme histochemical observations on the respiratory epithelium of the caecelian lungs and gills. A contribution to the understanding of the evolution of the vertebrate respiratory epithelium. Arch Histol Jpn Okayama 44:117–133
- Welsch U (1983) Phagocytosis in the amphibian lung. Anat Anz 154:323-327
- Welsch U, Aschauer B (1986) Ultrastructural observations on the lung of the Emperor penguin (Apternodytes forsteri). Cell Tissue Res 243:137-144
- Welsh MJ (1987) Electrolyte transport by airway epithelia. Physiol Rev 67:1143-1184
- Welty JC (1964) The life of birds, 1st edn. Constable, London
- Welty JC (1979) The life of birds, 2nd edn. Saunders, Philadelphia
- Wendelaar-Bonga SE, Meis S (1981) Effects of external osmolarity, calcium and prolactin on growth and differentiation of the epidermal cells of the cichlid teleost *Sarotheradon mossambicus*. Cell Tissue Res 221:109–123
- Went FW (1968) The size of man. Sci Am 56:400-413
- West B, Zhou BX (1988) Did chickens go north? New evidence for domestication. J Archaeol Sci 15:515–533
- West BJ (1985) An essay on the importance of being nonlinear lecture notes in biomaterials, 62. Springer, Berlin Heidelberg New York
- West BJ (1987) Fractals, intermittency and morphogenesis. In: Degn H, Holden AV, Olsen LF (eds) Chaos in biological systems. Plenum Press, New York, pp 305-314
- West BJ (1990) Fractal physiology and chaos in medicine. World Scientific, Singapore
- West BJ, Goldberger AL (1987) Physiology in fractal dimensions. Am Sci 75:354-365
- West BJ, Bhargava V, Goldberger AL (1986) Beyond the principle of similitude in the bronchial tree. J Appl Physiol 60:1089–1097
- West BJ, Brown JH, Enquist BJ (1997) A general model for the origin of allometric scaling laws in biology. Science 276:122–126
- West JB (1974) Respiratory physiology: the essentials. Williams and Wilkins, Baltimore
- West JB (1977a) Ventilation/blood flow and gas exchange, 3rd edn. Blackwell, Oxford
- West JB (1977b) Regional differences in the lung. Academic Press, New York
- West JB (1983) Climbing Mt Everest without oxygen: an analysis of maximal exercise during extreme hypoxia. Respir Physiol 52:265-274
- West JB (1991) High altitude. In: Crystal RG, West JB (eds) The lung: scientific foundations. Raven Press, New York, pp 2093–2107
- West JB, Dollery CT (1965) Distribution of blood flow and ventilation-perfusion ratio in the lung, measured with radioactive CO<sub>2</sub>. J Appl Physiol 15:405-410
- West JB, Jones NL (1965) Effects of changes in topographical distribution of lung blood flow on gas exchange. J Appl Physiol 20:825–835
- West JB, Matthews FL (1978) Stresses, strains, and surface pressures in the lung caused by its weight. J Appl Physiol 32:332–345
- West JB, Wagner PD (1980) Predicted gas exchange on the summit of Mt Everest. Respir Physiol 42:1–11
- West JB, Dollery CT, Naimark A (1964) Distribution of blood flow in isolated lung: relation to vascular and alveolar pressures. J Appl Physiol 19:713–724
- West JB, Dollery CT, Matthews CME, Zardini P (1965) Effects of aerolized artificial surfactant on repeated oleic acid injury in sheep. Am Rev Respir Dis 141:1014–1019

- West JB, Lahiri S, Maret KH, Peters RM, Pizzo CJ (1983) Barometric pressure at extreme altutudes on Mt Everest: Physiological significance. J Appl Physiol 54:1188–1194
- West JB, Tsukimoto K, Mathieu-Costello O, Prediletto R (1991) Stress failure in pulmonary capillaries. J Appl Physiol 70:1731–1742
- West JB, Mathieu-Costello O, Jones JH, Birks EK, Logerman RB, Pascoe JR, Tyler WS (1993) Stress failure in pulmonary capillaries in race horses with excercise-induced pulmonary haemorrhage. J Appl Physiol 75:1097–1109
- West NH, Burggren WW (1982) Gill and lung ventilatory responses to steady-state aquatic hypoxia and hyperoxia in the bullfrog tadpole. Respir Physiol 47:165-176
- West NH, Burggren WW (1983) Reflex interactions between aerial and aquatic gas exchange organs in larval bullfrogs. Am J Physiol 244:R770–R777
- West NH, Burggren WW (1984) Control of pulmonary and cutaneous blood flow in the toad, Bufo marinus. Am J Physiol 247:R884-R894
- West NH, Jones DR (1975) Breathing movements in the frog *Rana pipiens*. I. The mechanical events associated with lung and buccal ventilation. Can J Zool 53:332–344
- West NH, Butler PJ, Bevan RM (1992) Pulmonary blood flow at rest and during swimming in the green turtle, *Chelonia mydas*. Physiol Zool 65:287–310
- Weymouth FW, Crimson JM, Hall VE, Belding HS, Field II (1944) Total and tissue respiration in relation to body weight. A comparison of the kelp crab with other crustaceans and with mammals. Physiol Zool 17:50-71
- Wheatly MG, Taylor EW (1979) Oxygen levels, acid-base status and heart rate during emersion of the shore crab, *Carcinus maenas* (L) into air. J Comp Physiol 132B:305-311
- Wheatly MG, Taylor EW (1981) The effect of progressive hypoxia on heart rates, ventilation, gas exchange and acid-base status in the crayfish, *Austropotamobius pallipes*. J Exp Biol 92:125–141
- White FN (1978) Comparative aspects of vertebrate cardiorespiratory physiology. Annu Rev Physiol 40:471–499
- White FN, Bickler PE (1987) Cardiopulmonary gas exchange in the turtle: a model analysis. Am Zool 27:31–40
- White FN, Ross G (1965) Blood flow in turtles. Nature (Lond) 208:759-760
- White FN, Ross G (1966) Circulatory changes during experimental diving in the turtle. Am J Physiol 211:15-18
- White FN, Kinney J, Siegfried WR, Kemp AC (1984) Thermal and gaseous conditions of hornbill nests. Natl Geogr Res Rep 17:931–936
- White RE, Coon MJ (1980) Öxygen activation by cytochrome P<sub>450</sub>. Annu Rev Biochem 49:315– 356
- Whitford RW, Hutchison VH (1967) Body size and metabolic rate in salamanders. Physiol Zool 40:127–133
- Whiting HP, Bone Q (1980) Ciliary cells in the epidermis of the larval Australian dipnoan, *Neoceratodus*. J Linn Soc Zool Lond 68:125-137
- Whitmore CM, Warren CE, Doudoroff P (1960) Avoidance reactions of salmonid and centrachid fishes to low oxygen concentrations. Trans Am Fish Soc 89:17–26
- Wickler SJ, Marsh RL (1981) Effects of nestling age and burrow depth on CO<sub>2</sub> and O<sub>2</sub> concentrations in the burrows of bank swallows (*Riparia riparia*). Physiol Zool 54:132–136
- Wicksten M (1994) Sytematics agenda 2000 (letters to the editor). Am Sci 82:205
- Widdicombe J (1997) Airway alveolar permeability and surface liquid thickness: theory. J Appl Physiol 183:3–12
- Widmer HR, Hoppeler H, Nevo A, Taylor CR, Weibel ER (1997) Working underground: respiratory adaptations in the blind mole rat. Proc Natl Acad Sci USA 94:2062–2067
- Wiebe AH (1933) The effect of high concentrations of dissolved oxygen on several species of pond fishes. Ohio J Sci 33:110-126
- Wiedersheim R (1879) Die Anatomie der Gymnophionen. Fisher, Jena
- Wiener F, Morkin E, Skalak R, Fishman P (1966) Wave propagation in the pulmonary circulation. Circ Res 19:834-850
- Wierenga PJ, Nielsen DR, Hagan RM (1969) Thermal properties of a soil based upon field and laboratory measurements. Soil Soc Am Proc 33:354–360
- Wigglesworth VB (1950) The principles of insect physiology. Methuen, London

Wigglesworth VB (1953) Surface forces in the tracheal system of insects. Q J Microsc Sci 94:507– 522

- Wigglesworth VB (1965) The principles of insect physiology, 6th edn. Methuen, London
- Wigglesworth VB (1972) The principles of insect physiology, 7th edn. Chapman and Hall, London
- Wigglesworth VB, Lee WM (1982) The supply of oxygen to the flight muscles of insects: a theory of tracheole physiology. Tissue Cell 14:501–518
- Wignall PB, Twitchett RJ (1996) Oceanic anoxia and the end-Permian mass extinction. Science 272:1155–1158
- Wilkie DR (1977) Metabolism and body size. In: Pedley TJ (ed) Scale effects in animal locomotion. Academic Press, New York, pp 23-36
- Wilkin PJ, Williams MH (1993) Comparison of the aerodynamic forces on a flying sphingid moth with those predicted by quasi-steady theory. Physiol Zool 66:1015–1044
- Williams DD, Rausch RL (1973) Seasonal carbon dioxide and oxygen concentrations in the dens of hibernating mammals (Sciuridae). Comp Biochem Physiol 44A:1227–1235
- Williams MH, Wesseldine S, Somma T, Schuster R (1981) The effect of induced erythrocythemia upon 5-mile treadmill run time. Med Sci Sports Exercise 13:169–175
- Williams RJP, Da Silva JJRF (1978) High redox potential chemicals in biological systems. In Williams RJP, Da Silva JJRF (eds) New trends in bio-inorganic chemistry. Academic Press, London, pp 121–171
- Willmer EN (1934) Some observations on the respiration of certain tropical fresh water fish. J Exp Biol 11:283-306
- Willmer EN (1970) Cytology and evolution, 2nd edn. Academic Press, London
- Wilson DE (1973) Bat faunas: a trophic comparison. Syst Zool 22:14-29
- Wilson EO (1992) The diversity of life. Belknap Press, Cambridge
- Wilson KJ, Kilgore DL (1978) The effects of location and design on the diffusion of respiratory gases in mammal burrows. J Theor Biol 71:73–101
- Wilson TA (1981) Relations among recoil pressure, surface area, and surface tension in the lung. J Appl Physiol 50:921–926
- Wilson TA, Bachofen H (1982) A model for mechanical structure of the alveolar duct. J Appl Physiol 52:1064–1070
- Wilson TA, Beck KC (1992) Contributions of ventilation and perfusion inhomogeneities to the Va/Q distribution. J Appl Physiol 72:2298–2304
- Wimsatt WA (1970) Biology of bats. Academic Press, London
- Winick M, Coscia A, Nobble A (1967) Cellular growth in human placenta. I. Normal cellular growth. Pediatrics 39:248–251
- Winker S, Woese CR (1991) A definition of the domains Archea, Bacteria and Eucarya in terms of small subunit ribosomal RNA characteristics. Syst Appl Microbiol 14:305–310
- Winterstein H (1908) Beiträge zur Kenntnis der Fischatmung. Pfluegers Arch Gesamte Pysiol Menschen Tiere 125:73–98
- Winterstein H (1925) Über die chemische Regulierung der Atmung bei den Cephalopoden. Z Vergl Physiol 2:315–328
- Wintrobe M (1934) Variations in size and haemoglobin content of erythrocytes in the blood of various vertebrates. Folia Hematol 51:32–47
- Wislocki GB, Belanger LF (1940) The lungs of the larger cetacea compared to those of smaller species. Ibid 78:289–297
- Withers PC (1992) Comparative animal physiology. Saunders, New York
- Withers PC, Casey TM, Casey KK (1979) Allometry of the respiratory and hematological parameters of Arctic mammals. Comp Biochem Physiol 64A:343-350
- Withers PC, Hillman SS, Simmons LA, Zygmut AC (1988) Cardiovascular adjustments to enforced activity in the anuran amphibian, *Bufo marinus*. Comp Biochem Physiol A 89:45– 49
- Wit F (1932) Über den Einfluss der Luftfeuchtigkeit auf die Grösse der Atemöffnung bei Landpulmonaten. Z Vergl Physiol 18:116–124
- Witt R, Lieckfeld CP (eds) (1991) Bionics: nature's patents. Pro Futura Verlag, Munich
- Wittenberg BA, Wittenberg JB (1985) Oxygen pressure gradients in isolated cardiac myocytes. J Biol Chem 260:6548–6554

- Wittenberg JB (1965) The secretion of oxygen into the swim bladder of fish. J Gen Physiol 44:521-526
- Wittenberg JB (1976) Facilitation of oxygen diffusion by intracellular haemoglobin: oxygen and physiological function. Professional Information Library, Dallas
- Wittenberg JB, Wittenberg BA (1962) Active secretion of  $\dot{O}_2$  in the eye of fish. Nature (Lond) 42:214–232
- Wittenberg JB, Wittenberg BA (1987) Myoglobin mediated oxygen delivery to mitochondria of isolated cardiac myocytes. Proc Natl Acad Sci USA 84:7503-7507
- Wittenberg JB, Wittenberg BA (1989) Transport of oxygen in muscle. Annu Rev Physiol 51:857– 878
- Wittmann J, Prechtl J (1991) Respiratory function of catecholamines during the late period of avian development. Respir Physiol 83:375–386
- Wolfenson D, Frei YF, Berman A (1982) Blood flow distribution during artificially induced respiratory hypocapnic alkalosis in the fowl. Respir Physiol 50:87-92
- Wolk E, Bogdanowicz W (1987) Hematology of the hibernating bat, *Myotis daubentoni*. Comp Biochem Physiol 88A:637-639
- Wollman H, Smith TC, Stephen GW, Colton ET, Gleaton HE, Alexander SC (1968) Effects of extremes of respiratory and metabolic alkalosis on cerebral blood flow in man. J Appl Physiol 24:60–65
- Wolvekämp HP (1955) Die physikalische Kieme der Wasserinsekten. Experientia 11:294-301
- Wolvekämp HP, Waterman TH (1960) Respiration. In: Waterman TH (ed) The physiology of Crustacea, vol I. Academic Press, New York, pp 35–100
- Wolvekämp HP, Baerends GP, Kok B, Mommaerts WFHM (1942) O<sub>2</sub> and CO<sub>2</sub> binding properties of the blood of the catfish (*Sepia officinalis*) and the common squid (*Loligo vulgaris*). Arch Neerl Physiol 26:203–218
- Wood CM, Cadwell FH (1978) Renal regulation of acid-base balance in a fresh water fish. J Exp Biol 205:301–317
- Wood CM, Randall DJ (1973) The influence of swimming activity on sodium balance in the rainbow trout (Salmo gairdnen). J Comp Physiol 82:207-233
- Wood CM, Randall DJ (1981) Oxygen and carbon dioxide exchange during exercise in the land crab (Cardisoma carnifex). J Exp Zool 218:7-22
- Wood CM, McMahon BR, McDonald DG (1970) Respiratory gas exchange in the resting starry flounder, *Platichthys stellatus*: a comparison with other teleosts. J Exp Biol 78:167–183
- Wood CM, Perry SF, Randall DJ, Wood CM, Bergman HL (1989) Ammonia and urea dynamics in the Lake Magadi tilapia, a ureotelic fish adapted to an extremely alkaline environment. Respir Physiol 77:1–20
- Wood CM, Bergman HL, Laurent P, Maina JN, Narahara A, Walsh PJ (1994) Urea production, acid base regulation and their interactions in the Lake Magadi tilapia, a unique teleost adapted to highly alkaline environment. J Exp Biol 189:13–36
- Wood LDH, Bryan AC (1971) Mechanical limitations of exercise ventilation at increased ambient pressure. In: Lambertsen CJ (ed) Underwater physiology. Academic Press, New York, pp 125–205
- Wood SC (1971) Effects of metamorphosis on blood respiratory properties and erythrocytes adenosine trophosphate level of the salamander, *Dicamptodon ensatus*. Respir Physiol 12:53-65
- Wood SC, Glass ML (1991) Respiration and thermoregulation of amphibians and reptiles. In:
   Woakes AJ, Grieshaber MK, Bridges CR (eds) Physiological strategies for gas exchange and
   metabolism. Cambridge University Press, Cambridge, pp 107–124
- Wood SC, Johansen K (1974) Respiratory adaptations to diving in the Nile monitor lizard, Varanus niloticus. J Comp Physiol 89:145-158
- Wood SC, Lenfant CMJ (1976) Physiology of fish lungs. In: Hughes GM (ed) Respiration of amphibious vertebrates. Academic Press, London, pp 257–270
- Wood SC, Lenfant CMJ (1979) Oxygen transport and oxygen delivery. In: Wood SC, Lenfant C (eds) Evolution of respiratory processes: a comparative approach. Lung biology in health and disease, vol 13. Marcel Dekker, New York, pp 193-223
- Wood SC, Moberly WR (1970) The influence of temperature on the respiratory properties of iguana blood. Respir Physiol 10:20–29

- Wood SC, Weber RE, Maloiy GMO, Johansen K (1975) Oxygne uptake and blood respiratory properties of the caecilian *Boulengerula taitanus*. Respir Physiol 24:355–363
- Wood SC, Johansen K, Gatz RN (1978) Pulmonary blood flow, ventilation-perfusion ratio, and oxygen transport in a varanid lizard. Am J Physiol 233:R89–R93
- Woodbury LA (1942) A sudden mortality of fishes accompanying a supersaturation of oxygen in Lake Waubesa, Wisconsin. Trans Am Fish Soc 71:112–117
- Woodson RD (1984) Hemoglobin concentration and exercise capacity. Am Rev Respir Dis 129:S72–S75
- Wourms JP (1993) Maximization of evolutionary trends for placental viviparity in the spadenose shark, *Scoliodon laticaudus*. Environ Biol Fish 38:269–294
- Wourms JP, Callad IP (eds) (1992) Evolution of viviparity in vertebrates. Am Zool 32:249-354
- Wourms JP, Groove BD, Lombardi J (1988) The maternal embryonic relationship in vivaparous fishes. In: Hoar WS, Randall DJ (eds) Fish physiology, II: The physiology of developing fish. Academic Press, New York, pp 1–134
- Wray GA, Levinton JS, Shapiro LH (1996) Molecular evidence for deep Precambrian divergences among metazoan phyla. Science 274:568–573
- Wu ER (1993) The development and evolution of a key morphological innovation: air breathing organs in the anabantoidei. Am Zool 33:14A
- Wu HW, Chang HW (1947) On the arterial system of the gills and the suprabranchial cavities in *Ophiocephalus argus*, with special reference to the correlation with bionomics of the fish. Sinensia 17:1–15
- Wüst G, Brogmus W, Noodt E (1954) Die zonale Verteilung von Salzgehalt, Niederschlag, Verdunstung, Temperatur und Dichte an der Oberfläche der Ozeane. Kiel Meeresforsch 10:137-161
- Xu L, Mortola JP (1989) Effects of hypoxia on the lung of the chick embryo. Can J Physiol Pharmacol 67:515–519
- Yager D, Butler JP, Bastacky J, Israel E, Smith G, Drazen JM (1989) Amplification of airway constriction due to liquid filling of airway interstices. J Appl Physiol 66:2873–2884
- Yalden DW, Morris PA (1975) The lives of bats. Quadrangle The New York Times Book Co, New York
- Yamaguchi KD, Nguyen-Phu D, Scheid P, Piiper J (1985) Kinetics of O<sub>2</sub> uptake and release by human erythrocytes studies by stopped-flow technique. J Appl Physiol 58:1215–1224
- Yamaguchi KD, Jürgens H, Bartels H, Scheid P, Piiper J (1988) Dependence of O<sub>2</sub> transfer conductance of red blood cells on cellular dimensions. In: Mochizuki M, Honig CR, Koyama T, Goldstick TK, Bruley DF (eds) Oxygen transport to tissue, vol. X. Plenum Press, New York, pp 571–578
- Yamao F, Muto A, Kawauchi Y, Iwami M, Iwagami S et al. (1985) UGA is read as trytophan in mycoplasma capricolum. Proc Natl Acad Sci USA 82:2306–2309
- Yang D, Oyaizu Y, Oyaizu H, Olsen GF, Woese CR (1985) Mitochondrial origins. Proc Natl Acad Sci USA 82:4443–4447
- Yeliseev AA, Krueger KE, Kaplan S (1997) A mammalian mitochondrial drug receptor functions as a bacterial "oxygen" sensor. Proc Natl Acad Sci USA 94:5101–5106
- Yen MRT (1989a) Elastic properties of pulmonary blood vessels. In: Chang HK, Paiva M (eds) Respiratory physiology: an analytical approach. Marcel Dekker, New York, pp 533–559
- Yen MRT (1989b) Elasticity of microvessels in postmortem human lungs. In: Lee JS, Skalak TC (eds) Microvascular mechanics. Springer, Berlin Heidelberg New York, pp 175–190
- Yoder MC, Checkley LL, Giger U, Hanson WL, Kirk RL, Capen RL, Wagner WW (1990) Pulmonary microcirculatory kinetics of neutrophils deficient in leukocyte adhesion-promoting glycoproteins. J Appl Physiol 69:207–213
- Yonge CM (1947) The pallial organs in the aspidobranch Gastropoda and their evolution throughout the Mollusca. Philos Trans R Soc Lond 232B:443–518
- Yonge CM (1952) The mantle cavity in *Siphonaria alternata* Say. Proc Malacol Soc Lond 29:190– 199
- Yonge CM (1958) Observations on the pulmonate limpet *Trimusculus (Gadinia) reticulatus* (Sowerby). Proc Malacol Soc Lond 33:31–37
- Youlson JH, Freeman PA (1976) Morphology of the gills of larval and parasitic adult sea lamprey, *Petromyzon marinus* L. J Morphol 149:73-104

Young BA (1992) Trachea diverticula in snakes: possible functions and evolution. J Zool (Lond) 227:567–583

- Young JS (1973) A marine kill in New Jersey coastal waters. Mar Pollut Bull 4:70P
- Young RE (1972) The physiological function of hemocyanin in some selected crabs. II. The characteristics of haemocyanin in relation to terrestrialness. J Exp Mar Biol Ecol 10:193–206
- Young RE (1978) Correlated activities in the cardioregulatory nerves and ventilatory system in the Norwegian lobster, *Nephrops norvegicus* (L). Comp Biochem Physiol 61A:387–394
- Young RE, Coyer PE (1979) Phase co-ordination in the cardiac and ventilatory rhythms of the lobster *Momarus americanus*. J Exp Biol 62:53-74
- Young FN, Zimmerman JR (1956) Variations in temperature in small aquatic situations. Ecology 37:609–611
- Youvan D, Marrs B (1987) Molecular mechanisms of photosynthesis. Sci Am 256:42-50
- Ysseling MA (1930) Über die Atmung der Weinbergschnecke (*Helix pomatia*). Z Vergl Physiol 13:1–60
- Zaccone G, Fasulo S, Ainis L (1995) Gross anatomy, histology and immunohistochemistry of respiratory organs of air-breathing and teleost fishes. In: Pastor LM (ed) Histology, ultrastructure and immunohistochemistry of respiratory organs in non-mammalian vertebrates. Servicio de Publicaciones de la Universidad de Murcia, Murcia, Spain, pp 15–33
- Zaccone G, Goniakowska-Witalinska L, Lauweryns JM, Fasulo S, Tagliafierro G (1989) Fine structure and serotonin immunohistochemistry of the neuroendocrine cells in the lungs of the bichirs *Polpterus delhezi* and *P. ornatipinnis*. Bas Appl Histochem 33:277–294
- Zadunaisky JA (1984) The chloride cell: the active transport of chloride and the paracellular pathways. In: Hoar WS, Randall DJ (eds) Fish physiology, vol XB. Academic Press, London, pp 129–176
- Zabzoule M, Marc-Vergnes JP (1986) A global mathematical model of the cerebral circulation in man. J Biomech 19:1015–1022
- Zander R, Schmid-Schörbein H (1973) Intracellular mechanisms of oxygen transport in flowing blood. Respir Physiol 19:279–289
- Zapol WM, Liggins GC, Schneider RC, Qvist J, Snider MT, Creasy RK, Hochachka PW (1979) Regional blood flow during simulated diving in the conscious Weddell seal. J Appl Physiol 47:968–973
- Zeuthen E (1953) Oxygen uptake as related to body size in organisms. Q Rev Biol 28:1-12
- Zeuthen E (1970) Rate of living as related to body size in organisms. Pol Arch Hydrobiol 17:21– 30
- Zhao-Xian W, Ning-Zhen S, Wei-Ping M, Jie-Ping C, Gong-Qing H (1991) The breathing pattern and heart rates of *Alligator sinensis*. Comp Biochem Physiol 98A:77–87
- Zhuang FY, Fung YC, Yen RT (1983) Analysis of blood flow in cat's lung with detailed anatomical and elasticity data. J Appl Physiol 55:1341–1348
- Ziebis W, Forster S, Huettel M, Jørgensen BB (1996) Complex burrows of the mud shrimp, *Callianassa truncata*, and their geochemical impact in the sea bed. Nature (Lond) 382:619– 622
- Zimmer G, Beyersdorf F, Fuchs J (1985) Decay of structure and function of heart mitochondria during hypoxia and related stress and its treatment. Mol Physiol 8:495–513
- Zinkler D (1966) Comparative metabolism of invertebrates. Z Vergl Physiol 52:99-144
- Zoond A (1931) Studies on the localization of respiratory exchange in invertebrates. III. The book lungs. J Exp Biol 8:263–266

## **Subject Index**

Abdominal cavity (see Cavity) 57, 289 Abdominal pumping 288 Acarina 285 Accessory respiratory organs 68, 102-103, 165, 183, 197, 222-224, 229, 231, 233 236, 238-244, 253-254, 261, 334 Acid-base balance 164-165, 183, 196, 228, 233-234, 244, 249, 273-275, 316, 340 Acid-base regulation 48, 236-237 Acid-base status 49-50, 187, 234, 239, 336, 357 Acidosis 50, 112, 127, 157, 197, 239, 274-275, 343-344, 354, 356 Acionyx jubatus 134 Acipenser transmontanus 86 Acrochordus javanicus 187 Acroloxus lacustris 97 Actinopterygian fish 20, 184, 255, 269 Active radicals (see Radicals) Active transport 207 Adaptation 3, 24, 35, 53, 57, 59, 61, 69, 71, 77, 81-82, 85, 95, 112, 122-123, 132 149, 151, 161, 164-165, 172-174, 182, 186, 202, 217, 219, 222, 224, 227 231, 238, 245, 249, 251-252, 261, 265, 274, 276, 299, 301, 318, 348 Adrenaline 198, 264, 338 Aerial gas exchange (see Gas exchange) Aerial hypoxia (see Hypoxia) Aerobic gas exchange 203 Aerobic glycolysis (see Glycolysis) Aerodynamic valving 119 Aerosphere 79, 152, 169, 181 Aestivation 237, 242 Agnatha 50, 160, 181, 204, 276 Agriolimax agrestis 248 Air breathing 20, 24-25, 38-39, 50, 55, 59-60, 87, 121, 156, 161, 181, 194, 196, 200 217, 219, 221-225, 227-229, 231, 233, 236-240, 242, 252-254, 258, 262-263, 265-266, 271, 273-175, 289, 299, 330, 343-345, 359 - amphibians 275, 277, 303 - fishes 100, 102, 109, 165, 182, 194, 222, 228-229, 233, 239-244, 253, 255 261, 267, 275, 277, 334 - crabs 97, 117, 249 - organs 24, 101, 143, 153, 156, 198, 241, 275-276 - snails 201, 245-246

- vertebrates 10, 20, 68, 96, 159, 253, 264, 289, 303, 309 Air capillaries 16, 127, 315-316, 319-320, 323, 334 - cells 58, 265, 302, 303-304, 332-333 - flow 22, 102, 119, 143, 147, 277, 312, 323 - gaping 224 – gills 150 - sacs 70, 86, 137, 143, 222, 250, 254, 277, 287-289, 304-305, 309, 314-316 318-321, 323, 348 Air-blood barrier 24, 135, 264 Alectura lathami 352 Alkaline lakes 181 Alkalosis 24, 50, 176, 316, 321 Allantois 350-351, 357 Alligator sinensis 117 Allosteric modulators 111 Alma emini 164, 167, 202 – nilotica 188 Alopex lagopus 123 Alveolar blood capillaries 66, 123, 319 - blood capillary barrier 83, 127, 178 - capillary blood 132, 178 - cells 341-342 - epithelium 123, 131, 179 - fluid 263, 332 - gas 13, 58, 123, 136, 274, 320 - lung 79, 176, 263, 304 - macrophage 328 - PCO<sub>2</sub> 49, 274, 277 - PO<sub>2</sub> 43, 54, 123, 132, 138, 277 - septa 58, 61, 83, 180, 228, 294 - surface 58, 83, 180, 263, 327, 338 - surface tension 59 Alveolus 54, 59 Alvinella pompejana 106, 167 Amazilia tzacatl 321 Amazonian basin 219, 224, 242, 254 Amblystoma tigranum 187 Ametabolic state 153 Amia calva 194, 225, 228, 237-238, 244, 267, 276, 332 Ammonia 8, 17, 99, 154, 160, 183, 217 Ammonia excretion 183 Amniotes 227 Amniotic fluid 264 Amphiaerobes 17, 33

Amphibians 20, 35, 38-39, 51, 60, 91, 95-96, 102, 109, 122, 125, 127-128, 149, 156 174-175, 184-191, 193, 204, 219, 225, 227, 233, 236, 239-243, 254, 262 271, 275-277, 289, 292-293, 298-304, 306, 309, 324, 329, 332-334, 349 Amphibious fish 219, 227-228, 261 Amphibolurus nuchalis 56, 306 Amphioxus 97, 184 Amphipnous cuchia 201, 222, 244, 253, 257 Amphipods 99 Amphiuma means 233 - tridactylum 271, 334 Amyda mutica 201 Anabas testudineus 102, 165, 197, 203, 228, 231, 243, 253, 257 Anaerobic fermentation 28 Anaerobic glycolysis (see Glycolysis) Anaerobic habitats 45, 153 - metabolism 12, 166, 241, 279, 285, 305, 308 - microorganisms 30 - state 84 Anal gills 188 Anas platyhynchos 176 Ancistrus anisitsi 253 Ancylus fluvialis 97 Andean goose 176 Angiosperms 35, 227 Angiotensin 17, 183, 338-340 Anguilla anguilla 51, 60, 197, 266-267, 269 - vulgaris 253 Annelids 92, 95, 98, 102, 106, 172, 185, 188, 200 Anomura 248-249 Anoxia 29, 46, 48, 116, 156, 165, 172-173, 198, 213, 303, 308, 343 Anser anser 176 - indicus 128, 176, 318 Antarctic icefish (see Icefish) Antioxidants 35 Antrozous pallidus 312 Anura 51, 115, 191, 193, 204, 300-303 Aorta 197, 242, 267, 269, 340 Aortic arch 69, 187 Aphrodite aculeata 200 Aplacophora 187 Aplysia californica 189, 198 - deplians 110 - fasciata 189 Apneustic breathing 240 Apoda 190, 300, 303, 327 Aquatic breathers 49, 109, 160, 219, 234, 271 - burrowers 172 - gas exchanger 127, 183, 223-224 - habitat 38, 155, 181, 227, 289, 341, 349 - hypoxia 159, 221, 223-224, 228 - life 35, 158, 193, 200, 252 - respiration 101, 115, 120, 151, 160, 165, 186, 203, 239, 251, 254, 261, 267 - respiratory organs 25 Arachnids 150, 278-279, 288

Arapaima gigas 236, 242, 267 Arboreal 2 Archilochus colubris 318 Arenicola cristata 187, 192 - marina 60, 99, 105, 115, 153, 167, 172, 334 Arhynchite pugettensis 103, 201 Aroid plants 273 Artemia 99, 105, 110 Arterial blood 13, 24, 49, 97, 112, 123, 125, 128, 138, 143, 176-177, 193, 196, 234 242-243, 248, 269, 275, 308, 316, 318-319, 340, 344, 356 Arterial PCO, 49, 128, 143, 159, 175-176, 191, 236-237, 239, 247, 321, 343-344, 357 Arterial PO2 48, 102, 104, 113, 138, 143, 191, 237, 242, 248, 250-251, 276, 279, 321 344, 357 Arterio-venous shunt 342 Arterio-venous anastomosis 124 Arthropods 2, 35, 66, 87, 91, 184, 188-189, 245, 265, 274, 276-277 Arum maculatum 273 Ascidians 97, 143, 184, 335 Ascites 80 Asphyxia 182, 274, 334 Aspidonotus spirifer 201 Aspirational breathing 104, 242 Astacus leptodactylus 99 Astylosternus robustus 92 Atelectasis 210, 332, 342 Athoracophoridae 247, 280 Atmosphere 2-3, 8-9, 26-27, 29-34, 36-40, 44-46, 54, 56, 87, 90, 99, 106, 121, 152-156, 165, 169, 170, 173-175, 227, 265-266, 318, 345-346, 348, 353 Atria 16, 66, 96, 208 Austropotamobius pallipes 116, 221 Autamorphism 53, 130 Avian pulmonary system 16, 86, 127, 143, 207, 304-305, 308-309, 314-316, 319, 320-323 Axolotl 193, 300 Bacteria 2-3, 30, 32, 40-41, 44-48, 130, 153, 166-167, 171 Balaena mysticetus 83 Bar-headed goose (see Anser indicus) Barometric pressure 9, 38, 156, 173-176, 353 Basommatophora 200 Bats 38, 56, 69, 71, 77, 79, 86, 101, 149, 309-314, 317, 320, 323, 349 Benthonic species 2 Bernoulli-Venturi effect 287, 348 Bicarbonate (HCO3<sup>-</sup>) 13, 48-50, 126, 155-156, 231, 233-234, 236, 266, 274-275 277, 357 Bichir 20, 164, 219, 222, 273, 332 Bifurcation 304, 306 Bifurcation points 87 Bimodal breathing 68, 102, 194, 217, 242-245, 250, 253-254, 275, 334 - respiration 254

- Biodiversity 1, 131
- Biomass 1, 40, 149, 164
- Biomphalaria sudanica 164, 201
- Biosphere 8, 20, 27, 29, 32, 35–38, 40, 87, 90, 149
- Birds 22-24, 38, 40, 49, 51, 70-71, 75-77, 79, 60, 82, 86, 97, 101, 122, 124-125, 127 139, 143-145, 149, 152, 159, 171, 176-177, 225, 236-238, 240-241, 244 251, 273, 277, 289, 293-294, 304-305, 308-324, 334, 336, 349-350, 352-353, 355
- Birgus latro 182, 231, 238, 249-251
- Bivalves 39, 95, 99, 111, 117
- Blennius pholis 229, 261
- Blood brain barrier 48
- capillaries 42, 58, 66, 77, 81, 83, 97, 123, 136, 143, 177, 187, 207, 255, 263 281, 293–294, 302, 319, 334
- circulation 14
- flow 56, 60, 64, 66, 77, 80, 83-84, 86, 94,
   96-97, 103, 110, 116, 124, 143 147, 175-176,
   178-179, 192, 197-198, 211, 213, 233, 238,
   240, 243-244 264, 266, 300, 308, 321, 341
- gas barrier 16, 54, 57-58, 71, 82-83, 123, 127, 133, 143, 151, 174, 207-208 244, 247-248, 250, 254-255, 271, 298, 303, 312, 316, 319-320, 322, 327 332, 354-355
  gills 188
- maternal (see Maternal blood)
- O<sub>2</sub> carrying capacity 76, 91, 104, 109, 111, 211, 248, 311, 355
- perfusion 59, 60, 71, 77–78, 87, 91–92, 99, 101, 115–118, 121, 127, 133–134 136, 138, 143, 145, 147, 171, 178, 207, 209, 211, 238, 240, 242–244, 261 276, 300, 303, 308, 320, 340
- pH 49, 50, 71, 92, 106, 109, 111–112, 154–156, 159, 163–167, 175–176, 181 192, 231, 233–234, 236–238, 241, 248, 266, 275, 277–278, 308, 344 357
- plasma 13, 49, 54, 66, 76-77, 104, 111-113, 126, 183, 196, 207-208 210, 233-234, 236, 264, 274-275, 340, 354-355, 357
- pressure 44, 72, 81, 83, 85, 93, 95, 97, 108, 125, 179–180, 198, 243, 249 264, 332, 340
- venous 13, 93, 97, 108, 134, 143, 179, 193-194, 222, 241-244, 249, 308, 319 321, 337
- vessels 69, 75, 83, 91–93, 101, 143, 174, 192, 211, 248, 293, 301
- viscosity (see Viscosity)
- volume 179, 196, 205, 294, 298, 301, 319–320, 355
- Blue-green algae 30, 32, 158
- Body mass 13, 16, 22, 24, 73, 80-82, 89, 91-92, 95-99, 102, 114, 116, 124-125 134, 201, 209, 247, 251, 276, 279, 287, 300-301, 305-306, 311, 313, 315 318, 321-322, 353
- Body size 12-13, 16, 36, 82-83, 87-88, 91, 95, 109, 177, 273, 286, 288, 322

Bohr (shift) effect 92, 105-106, 108-112, 122, 126, 165, 172, 266, 318, 321 Boleophthalmus boddarti 198 - pholis 231 Bombyx mori 286 Book lungs 66, 265, 278, 280, 323 Bothriolepis 20, 273 Boulengerula taitanus 301, 327 Bowfin (see Amia calva) Brachiopods 111, 188-189, 248 Brachvura 117, 172, 248 Bradycardia 95, 224 Bradykinin 17, 338-340 Brain 10, 16, 22, 38, 42, 45, 48, 56, 61, 70, 101, 103, 109, 118, 143, 175-176, 308 318, 321, 353 Branchial arch 69, 165, 196, 239, 240, 255 chambers 116-117, 183, 222, 249-251, 260 circulation 239 crown 184, 192, 335 vascular resistance 182, 240 Branchinella kugenumaensis 188 Branchioles 192 Branchiostegite 100, 231 Breath-holding 116 Breathing 116-118, 120-121, 127, 143, 150-151, 156, 159, 161, 164-165, 179, 181-182, 185-186, 190, 193-194, 196-198, 200-201, 217, 219, 221-225, 227-229, 231-234, 236-246, 248-255, 258, 261-265, 267, 271, 273-277, 280-289, 292, 299, 301, 303, 307-309, 311-312, 315, 330, 332, 334, 341-346 348, 357 control (see Control of breathing) 70 cost (see Cost of respiration) Bronchial tree 84-85 Bronchoalveolar lung 304 Brownian motion 124 Buccal cavity 24, 138, 222, 236, 253-254, 275, 289, 300-301 Buccal-force-pump 61, 69, 81, 101-102, 104, 115, 184, 241-242, 267, 289 Buccal-pharyngeal cavity 103, 185, 292, 300 Bufo bufo 187 Bufo marinus 51, 240, 302-303 Bullfrog 227, 275, 280 Buoyancy 83, 99, 229, 247, 267, 269, 274, 299 Burrow 12, 38, 98-100, 154-155, 166, 169-173, 192-193, 198, 200-201 Bythograea thermydron 167 Caecilians (see Apoda) Calcium carbonate (CaCO<sub>3</sub>) 69, 350 Callianassa californiensis 172 - truncata 98-99, 172 Callinectes sapidus 78, 182, 221

- Caloppa granulata 147, 151
- Camel 54
- Cancer magister 111, 117
- pagurus 99, 100
- productus 99, 111, 182

Capacitance 126, 165 - coefficient 106, 151, 170, 234 Capillary geometry 135 - loading 294, 320 - recruitment 124 - transit time 54, 57, 121, 124, 126 Carapace 100, 184, 237, 248, 250 Carassius auratus 51, 224 Carbon dioxide (CO<sub>2</sub>) 2, 9–10, 12–14, 16, 22, 24-28, 30, 32-40, 48-50, 53-54, 58 88, 91, 99, 104-106, 109, 114, 118, 121-124, 126, 128, 135, 143, 149, 151-159, 161, 163-166, 169-172, 174-176, 182, 187, 191, 193-194, 203, 208 219, 221, 223-225, 227, 231, 233-234, 236-241, 244, 246-249, 251, 253-255, 262, 265-266, 269, 271, 274-278, 293, 316, 318, 321, 343, 345-346 348-357 - monoxide 26, 29, 44, 110, 154, 164 Carbonic anhydrase 49, 126, 156, 231, 233, 240 Carboniferous period 21, 35-36, 39, 90, 222 Carcinus guanhumi 100 - maenas 100-101, 147, 182, 197, 251 Cardiac muscle 42, 106 - output 54, 82, 93, 95-97, 114, 117, 125-126, 133-134, 239-240, 242, 244 311, 319 Cardiopulmonary bypass 339 Cardisoma hirtipes 183 Carotid rete 143 Carrier pigments 14, 106, 112, 124 Catalyse 166, 233 Catecholamines 183, 341, 356 Caudata 190, 300-301 Caveolae cellulares 338 Cavity 24, 55, 57, 86, 94, 98-99, 101-103, 125-126, 138, 143, 183-185, 198, 200-201, 222-223, 225, 236, 245, 248-249, 251, 253-254, 265, 275, 289, 290-292, 300-301, 321, 334 Central nervous stimulus 318 - nervous system 3, 10 Cephalic rete 138 Cephalochordates (see Chordates) Cephalopods 78, 79-80, 95, 99, 102, 111, 116, 143, 185, 193, 269, 335 Cepola rubescens 98, 183 Cercopithecus aethiops 309 Cerebral artery 75, 84 - blood flow 143, 176, 318, 321 - cortex 56 - vasoconstriction 176, 321 Cerripedia 91 Cetaceans 121, 149 Chaenichthys rugosus 194 Chaenocephalus aceratus 113, 121, 194 Chaetopoda 91 Chaetopterus variopedatus 98, 172, 334-335 Chamsocecephalus esox 194 Channa argus 240, 244 - punctatus 244 - striata 197

Chaos (theory of) 11, 87 Character states 130 Chelicerates 190, 278 Chelonia 134, 170, 240, 274, 305, 308, 352 Chelonia mydas 240, 308 Chemical binding 105, 126 Chemoreceptors 48, 237-239 Chicken (see Gallus domesticus) Chilopoda 251, 285, 287 Chiromantis petersi 300, 303 - xerampelina 300, 303 Chironomus plumosus 335 Chiroptera 309-310 Chitin 182-183, 279, 285 Chloride cells 197-198, 244, 271 - shift 49 Chlorocruorin 111, 192 Chlorophyll 2 Chondrichthyces 160, 194 Chordates 69, 135, 184, 192, 273 Chorioallantois 350-351, 357 Choroid rete 123, 139 Chronobiology 3 Chryptochiton stelleri 110 Chrysemys (Pseudemys) scripta 51, 240-241 – picta 352 Chthonerpoton indistinctum 303 Ciliary movements 88, 98, 184-185, 190, 200 – currents 97 – feeding 335 – organ 336 – tufts 245 Circulation 14, 17, 21, 66, 72, 85-86, 91, 94-97, 157-158, 160, 178, 187, 200, 204 228, 239, 243, 255, 264, 308, 337-338, 340, 354 Circulatory system 16, 20, 49, 60, 76, 85, 90, 93-96, 104, 112, 137, 188, 193, 267 Cirripedia 188 Citric acid 47, 68 Clarias batrachus 257 – lazera 164 - mossambicus 68, 102, 164, 253, 257 Cleidoic egg 304, 349 Cleophaga melanoptera 176 Clinus superciliosus 349 Cloaca 102-103, 185, 200-201, 301 Closed circulation (see Circulation) Cockroach 287 Coelacanth 20-21, 86 Coelenterate 88-89 Coelomic cavity 86, 290-291, 312, 334 - fluid 91, 166-167, 187-188, 190, 200-201 Coenobita clypeatus 101 Coleoptera 164, 345 Colibri coruscans 319, 322 Colloidal osmotic pressure 111, 263 Columba livia 320 Compliance 302, 306-307, 316, 320 Concept organism 27

Conductance 58-59, 99, 210-211, 350, 352-353, 355, 357 Conger conger 233, 236 Control of breathing/respiration/ventilation 20, 38, 70, 152, 237-238, 242, 279, Conus arteriosus 243 Convection 99, 116, 123-124, 132, 135, 137, 151, 154, 156 Convective inertial force 119, - transport 14, 108, 151, 351 Cooperativity of O<sub>2</sub>-binding 105, 111, 192 Copepods 90 Coronary arteries 113, 194 Cortisol 197 Cossus 91, 286 Cost of respiration/breathing 104, 115, 120-121, 232 Costal suction pump 273 Cough mechanism 241 Counter-current gas exchange 100, 102, 126, 136, 138-139, 143, 147, 193, 213-214 266, 276, 319 Covalent bonds 11, 154 Crab 48, 78, 95, 99-101, 111, 116-117, 120, 147, 151, 167, 172-173, 178, 182-183 189, 197, 221, 223, 231, 236-238, 248-251, 271, 277 Cretaceous 3, 38-39, 82, 314 Crocidura flavescens 322 Crocodilia 204, 274 Crocodylus niloticus 304 Cross-current gas exchange system 136, 143, 147, 176, 316, 319 Crossopterygian fish 20, 228, 254 Crustacea 95-96, 99, 104, 111-112, 143, 172, 182-184, 190, 192-193, 231, 248 251-252, 276-277 Cryptic species 53 Cryptobiosis 10 Cryptobranchus alleganiensis 190 Cryptochaetum iceryae 193 Cryptozoic habitat 21, 90 Ctenidia 55, 184, 200, 223, 245, 265, 335 Ctenophorus nuchalis 333 Ctenopoma muriei 164 Cutaneous gas exchange 89, 95-96, 99, 127, 187, 190-192, 196, 233, 253, 264, 303 Cuticle 46, 184, 202, 265, 276, 346, 353 Cuticular gills 184 Cyanobacteria 30, 32, 46 Cyanocephalus volans 310 Cyclic - CO<sub>2</sub> - release 114 Cyclopterus lumpus 106 Cyclostomes 184, 194 Cygnus cygnus 318 - olor 319, 320-321 Cynea 89 Cyprinus carpio 106, 113 Cytochrome 24, 48, 108, 166, 206, 217 Cytotrophoblast 211

Danio dangila 56, 99, 196 DDT 352 Decompression sickness 342 Deformation of: - egg shell 352 - ervthrocytes 77 - gill lamellae 194 - lung parenchyma 178 Derelict waters 38, 163, 225, 252, 254, 257 Dermoptera 310 Deroceras reticulatum 94 Desert 3, 149, 249, 275, 287, 300 Design 2-4, 6, 8, 11-12, 16-17, 21-22, 25, 49, 53-57, 59, 61-62, 66, 68, 70-75, 78-79, 81, 83-86, 88, 94, 97-98, 104, 119, 131-133, 136-138, 177, 179, 181 184-185, 194, 204, 210, 215, 228, 242, 274-278, 280-281, 284, 286, 289, 303-306, 309, 311, 313, 315, 321-324, 336, 343, 351, 356 Desmognathus quadramaculatus 187 Devonian 20, 32-33, 35-36, 38-39, 161, 219, 221-222, 224, 274-276, 278, 298 Diaphragm 61, 91, 280, 290-291, 334 Dielectric constant 154-155 Diffusing capacity 43, 54, 57-59, 69, 73, 77, 79, 81-82, 112, 127, 131-134, 178-179 208-209, 211, 213, 234, 250, 254, 261, 263-264, 298, 305-306, 309, 319-320, 350, 354-355 Diffusion 14, 17, 24, 32, 42, 49, 60, 62, 77, 87-91, 99, 106, 108, 115-116, 121-124, 126-128, 131-132, 135-137, 143, 151, 156-157, 169-171, 174, 187, 190 205-209, 225, 247-248, 258, 264, 266-267, 273, 277-279, 286-287, 336 341, 344-346, 352-353, 355-357 - coefficient 288, 354-355 - distance 77 - gradient 208, 288 - limitation 123, 128, 138 - rate 122, 127 - resistance 112, 209 Dipnoi 20, 69, 96, 193-194, 219, 240, 243-244, 254, 257, 269, 298, 301-303, 309 Diptera 188, 193, 286 Dissociation curve 22, 123, 126, 355 Diversity 1-2, 5-6, 12, 21, 25, 34, 39, 53, 81-82, 87, 111, 119, 131-132, 157, 181 183, 215, 227, 245, 248, 253-254, 276, 299, 302, 304, 323, 340 Diving 103, 105-106, 109, 121, 159, 187, 240, 317, 345, 348-349 Diving women (see Korean ama) DNA 41, 42-44, 47, 209 Dogfish (see Scyliorhinus canicula) Dorsal aorta 197, 242, 267 Double capillary system 293-294, 306 - circulation (see Circulation)

Draco volans 310 Dromaius novaehollandiae 76, 321-322 Earth 1-3, 6-9, 11-12, 20-21, 25-27, 29-34, 36-40, 74, 79, 89, 91, 95, 105, 152-154, 160, 170, 174, 177, 181, 221, 274, 304, 309 Earthworms 21, 89, 91, 95, 105, 170, 257 Echinoderms 91, 98, 183, 188-189 Ecosystems 5, 13, 26, 28, 31, 149, 161, 164, 167, 181, 227, 274 Ectothermy 60, 87, 96-97, 114, 116, 125, 175, 228, 248, 277, 309, 324, 350 Eel 21, 51, 60, 95, 102, 110, 120, 151, 164, 172, 185, 187, 197, 201, 222, 236-237 244, 253-254, 257, 266-267, 269 Egg 54, 58-59, 69, 88, 90-91,122, 184, 193-194, 204-205, 212, 225, 300, 304, 348-356 - air space 357 - clutch 58, 350, 352 - incubation 58, 87, 350-357 - membrane 204 - permeability 354-356 - shell 58-59, 69, 204-205, 212, 349-356 - shell conductance 58, 350, 352-353 - size 354 Eichhornia crassipes 165 El Niño 221 Elasmobranchs 60, 80, 93, 169, 193-194, 240 Elastic recoil of the lung 241, 343 - of tissue 102, 289, 307, 327 Electrophorus electricus 249, 253-254 Elements 6-7, 21, 40, 45, 73, 121, 184,197, 239-240, 294, 307 Eleutherodactylus coqui 249, 253-254 Embiotoca lateralis 349 Embryo 45, 69, 88, 90-91, 95, 97-98, 165, 184, 204, 213, 294, 316, 350, 352-357 Emphysema 41, 180, 337 Emu (see Dromaius novaehollandiae) Endocrine function 39, 83, 204, 338 Endosymbiosis (see Symbiosis) Endothelial cells 10, 45, 91, 96, 126, 179, 197-198, 209, 248, 332, 334, 338, 340-341 Endotherm 24, 59, 72, 87, 90, 97, 109, 116, 124, 160, 271, 273, 281, 308-309, 314 323 Energy 3, 5, 7, 10-12, 16-17, 26-28, 30, 33, 41, 46, 49, 54-56, 50-60, 69, 71, 84-85 90, 99, 104, 110, 115-117, 121, 155, 160, 166, 175, 228, 241, 271, 273, 277 289, 305, 307, 310, 312-313, 317, 322, 324, 332, 343, 353 - budget 84, 104 - consumption 305, 317 - cost 85 - expenditure 12, 60, 138 - needs/requirements 241, 317, 343 - saving 69, 99, 115-116, 118, 121, 241, 277, 307, 310 Enicognathus ferrugineus 58, 350 Entomacrodus nigricans 228 Entropy 11

Environment 2-5, 8, 11-14, 16, 24, 28, 33, 40, 45-46, 49, 53, 55, 57-58, 61, 71-72 75-76, 80-81, 88, 90-91, 105, 108, 114, 119, 124, 127, 131-132, 134-135 141, 149, 151-153, 157, 160-161, 164-165, 167, 170, 173, 181, 191-192, 198 202, 217, 219, 221, 224-225, 228, 234, 245, 265, 271, 273, 276, 284, 292 301-302, 314, 320, 327, 329, 335, 344, 348, 350, 352, 355 Enzymes 6, 12, 14, 32, 42-47, 68, 114, 126, 166, 231 Ephemeroptera 200, 348 Epidermis 87, 187, 302 Episodic breathing/respiration 240-241, 277 Epithelial cells 116, 141, 145, 164, 167, 179, 197-198, 245, 306 Epitheliochorial placenta (see Placenta) Epithelium 123, 189, 191, 194, 196-197, 223, 234, 248, 261, 263-264, 266, 341 Epomophorus wahlbergi 310, 312 Eptatretus stoutii 93 Eremophilus mutisii 229 Erpetoichthys calabaricus 222 Erythrinus erythrinus 165, 224 Erythrocytes (see Red blood cells) Erythropoiesis 209, 355 Eukaryote 33, 47, 68, 87 Eunectes murinus 69 Eutheria 204, 308 Eutrophication 158 Evagination 25, 101, 193, 188, 254 Everglades (see swamps) Evolution 2, 4, 6-8, 10-12, 14, 17, 20-22, 24-27, 29, 31-33, 37, 39-40, 42, 44-48 53, 54-55, 61, 67-68, 70-72, 74-75, 78-79, 80, 85, 87-88, 96, 104, 106, 109 111, 149-150, 156, 169, 177, 184, 204, 215, 219, 222-223, 228-229, 231, 242 244, 250, 253, 265-266, 271, 273-275, 289, 299, 314, 317, 321, 349 Evolutionary adaptations/refinements 111, 274 - advances 45, 55, 70, 245, 291 blind alley 75 - chain 112 - changes 32, 68, 70 - dead end 183, 232, 303 - developments (see Evolutionary advances) - events 53 - expansions 74 - gaps 21 - innovations (see Evolutionary advances) - lineages 25, 181, 335 - novelties 217 - pathways 55, 61 - progression 289 - transformations 20 Exaptations 69 Excercise 13, 22, 41, 56, 60, 71, 78, 82, 86-87, 106, 115, 120, 123-124, 126, 133 143, 160, 174-175, 179-180, 209, 263, 301, 305-306,

External gills 98, 183-187, 190-191, 193-194, 225, 243, 276, 300 - respiration 14 Extinction 11, 13, 20, 38-39, 61, 80, 82, 219, 314 Extraction factor 102, 145, 147, 151, 185, 187, 192, 249, 252, 311-312, 320 Facultative air breather (see Air breathing) Fetal apnea 118 - birth weights 210 - blood 58, 205, 207-208 - breathing 118 - circulation 66, 205-206, 208-209 - erythropoiesis 209 - gills 188 - growth 40, 208-209 - haematocrit 207 - lung 80 - maternal gas exchange 205, 211 - maternal-placental barrier 205, 207-208 - membranes 213 metabolism 209 - nutrition 204 - O<sub>2</sub> consumption 118, 205, 213 - respiratory movements (see Fetal breathing) - villous system 205 Fetus 59, 204-205, 209-213, 215, 355 Fick's law 127 Fish 13, 16-17, 20-21, 50-51, 60, 62, 66, 68-69, 78-79, 86, 93-102, 104-106, 108-110, 113-118, 121, 124-126, 128, 133, 136, 138-139, 143, 145, 149, 151-152 155-156, 159-161, 164-165, 167, 172-173, 181-189, 193-194, 196-198, 202-203, 208, 217, 219, 221-231, 233-234, 236-244, 250, 252-255, 257-258, 260-263, 265-267, 269, 271, 274-277, 280, 298, 301-302, 306, 309-310, 330 332-334, 336, 340, 349 Flight 13, 16, 22, 56, 70-71, 76, 86, 90, 101, 114, 132, 143, 149, 174-177, 179, 273 281, 284, 286-288, 309-314, 316-320, 322-323, 336, 349 Flight muscles 16, 90, 114, 177, 273, 281, 284-285, 288, 314, 318-319, 322 Fluorocarbon 342 Force 4-5, 8, 11, 26, 38, 61, 69, 81, 86, 101-102, 104, 106, 115, 119, 123, 154-155 174, 177, 182, 184, 219, 222, 241 Form (see Life forms) 242, 263, 267, 286, 289, 294, 312, 330, 342, 346 Fossil 36-37, 71, 273, 310 Fossorial 2, 49, 159, 166, 169-172, 224, 301, 309 Fox 54, 57, 86, 123 Fractal geometry (see Geometry) Free radicals (see Radicals) Fresh water 60, 97, 99, 152, 155, 158-160, 188, 194, 197, 200, 217, 219, 221, 224 227, 245, 252, 254, 265, 280, 303, 310

Functional reserve 56-59, 86, 104, 118, 208-209, 211, 213, 308, 313, 320-321, 350 GABA ( $\gamma$ -aminobutyric acid) 3 Galago senegalensis 309 Gallus domesticus 51, 58, 76, 80, 247, 319, 322 Gammarus locusta 193, 238-240, 253, 332 Gar 20 4, 8-9, 12, 17, 25-27, 32-35, 37, 40, 44, Gases 59, 69, 77, 91, 121, 122, 124, 126-127, 131, 133, 136, 150-151, 153, 155-156, 158-159, 163-166, 170-172 174, 182, 217, 225, 348, 351, 353 Gas gill 200, 286, 345-346, 348 - bladder 68, 104-106, 116-117, 127, 222, 225 - bubbles 345-346, 348 - composition 170 concentrations 171 - density 120 - diffusion 169-170, 209-210, 315 - exchange components 316, 319-320, 323-324, 332, 342, 344 - exchange efficiency 178, 182, 309, 313, 320 exchange mechanisms 190 - exchange media 66, 119, 136, 139, 149 - exchange pathways 190, 193, 228, 288 - exchange surface 122, 176, 208, 320 - exchange tissue 143, 312, 319, 324-325 - exchange/transfer 17, 38, 49, 55, 58, 60, 64, 67, 71, 77-79, 82-83, 85-89 91-92, 95-96, 101, 112, 115, 120-124, 137-138, 143, 164, 177, 183-184 187-188, 191-192, 194, 196, 198, 200-202, 204-205, 207-208, 210, 214 227, 229, 231-232, 263, 267, 271, 287, 292, 319, 334, 348, 353 - exchangers 4, 11-12, 14, 16-18, 20-24, 46, 49-50, 53-57, 59, 61-62, 66 68-71, 73, 75-76, 78, 81-82, 85-87, 99, 101, 104, 112, 115, 118-119, 120-121, 126-127, 130-133, 135-136, 143, 150, 156, 160, 172, 183, 186, 188 204-205, 207, 212, 223-224, 233, 280, 288, 330, 334 - gland 106, 266 - requirements 169 - secretion organs 139 - spitting reflex 267 - turbulence 120, Gastrophilus intestinalis 348 Gastropod 55, 94-95, 105, 111-112, 150, 164, 187, 193, 200-201, 223, 225, 245-248, 277, 280 Gecarcoidea natalis 183 Genetic code 47 Geograpsus crinipes 183 – grayi 183 Geometry 58, 75, 84, 86, 119, 132, 135, 184, 170, 205, 354-355 Geomys bursarius 170-171 - pinetis 170

Geothermal springs 26, 167, 181, 352 Gestation 87, 205, 208-210, 212-213, 340, 350, 355 Gills 2, 21, 24-25, 49, 55, 60-62, 66, 68-69, 78-79, 86, 93, 95, 97-104, 106, 110-111, 113-118, 124, 126, 133, 136, 138, 143-145, 147, 150-151, 153, 155-156 160, 165, 167, 178, 181-194, 196-198, 200-201, 204, 219, 221-225, 231 233-234, 236-245, 248-251, 253-255, 260-263, 265, 267, 271, 274-276 286, 300, 330, 334-335, 340, 342, 344-345, 348 - arch 68-69, 184, 194, 196, 243-244, 261 – bar 193 - breathing 193 - lamellae (see Secondary lamellae) - ventillation 79, 102, 115, 117-118, 165, 221, 239, 241 Gillichthys mirabilis 181, 225, 241, 261, 267 Glaucomys volans 310 Global warming 37, 169, 221 Glossoscolex giganteus 92 Glucose 11, 28, 43, 92, 106, 109 metabolism (see Metabolism) Glutathione 46 Glycolysis 10, 203 Gobius cobitis 95 Goldfish 51, 95, 110, 121, 224 Goliathus goliathus 286 Gopher 170-171 Gourami 240, 334 Grasshopper 288 Gravity 4, 8-9, 76, 120, 156, 159, 177, 178–179, 182, 274, 322, 342, Greenhouse effect 9, 27, 34, 36-37, 154 - gases 34 Gulf of Mexico 156, 221, 318 Gymnarchus niloticus 164, 183, 267 Gymnophiona 69, 190, 300, 327 Gymnosperms 227 Gyps rueppellii 175 Habitat (see Environment) Haemoglobin 22, 24, 44, 49, 64, 76-77, 92, 104-106, 108-109, 110-114, 124, 126 164, 166-167, 177, 192, 194, 196, 200-201, 203, 210, 234, 276, 355 - concentration 71, 77, 82, 86, 106, 112, 123, 126, 134, 175, 209, 211, 301 311, 321 - content 76, 110, 322 - dissociation curve 123, 126, 176 - O<sub>2</sub> affinity 108-109, 121, 123-124, 177, 192, 208, 258, 303 Haemoglobinless fish 113-114, 121, 194 Haemolymph 89, 93, 98, 100, 104, 111, 117, 178, 183, 189, 231, 236, 249, 279 Hagfish 50, 93-94, 188, 194 Haldane effect 49, 126 Halichoerus grypus 349 Haliotes corrugata 110

Harmonic mean thickness 58, 83, 207, 211, 254-255, 322, 354 Hatching 184, 186, 194, 253, 265, 300, 350, 352-353, 355-356 Heart muscle 16, 42, 76, 106 - rate 49, 93-97, 101, 114, 117-118, 125, 191, 239, 308 Helicostoma 187 Hematin 166-167 Hematocrit 71, 76-77, 86, 110, 112, 123, 175, 207, 209–211, 301, 306, 311–312 Hemerythrins 111 Hemiptera 164, 345 Hemitripterus americanus 51, 95 Hemochorial placentae (see Placenta) Hemocyanin 246, 248, 279, 280 Hering-Breuer reflex 70 Heterocephalus glaber 134, 170, 172, 309 Heterocytes 46 Heteropneustes fossilis 225, 233, 243, 254 Heterothermy 13, 87, 134, 228 High altitude 22, 38, 40, 71, 76–77, 83, 126, 143, 173-176, 204, 210-211, 217, 309, 317-318, 353 Himalayan mountains 176 Hind gut 2, 102, 166-167, 200-201 Hirundo fuligula 322 Histamine 338–340 Holosteans 244, 253-254, 267, 273 Holothuria forksali 102, 200 – tubulosa 98 Holothurians 99, 195 Holthuisana transversa 100, 117–178, 249 Homarus vulgaris 59 Homeostasis 3, 11-12, 49, 236 Homeotherm 72, 87, 109, 160, 174, 271, 273, 308 Homo sapiens 70 Homology 61, 68-69, 113, 265-266, 269, 290, 355 Hoplerythrinus unitaeniatus 236 Hoplias lacerdae 221 – malabaricus 221, 224 Hoploerythrinus unitaeniatus 266-267 Hoplosternum thoracatum 257 Horizontal septum 289 Hormones 17, 183, 196-197, 204, 337, 340 Horse 54, 55, 76, 80-81, 101, 123, 178, 180 Hovering 322 Humidic habitat (see Cryptozoic habitat) Hummingbird 76, 125, 273, 318-319, 321-322 Hydrogen sulfide (H<sub>2</sub>S) 26, 154-156, 164, 166-167, 169, 172, 200-201, 217, 225 Hydrophobic hairs 346, 348 Hydrosphere 8, 32, 79, 152, 169 Hydrostatic pressure 3, 179, 322, 334, 345, 348 Hydrothermal vent 31, 106, 156, 167 Hydroxyl radical 41-44, 275

Hyla arborea 301, 303 – cricifer 92 – versicolor 92 Hymenoptera 288 Hyperbaria 120, 152 Hypercarpnia 48-49, 99-101, 110, 118, 153, 161, 166, 171–172, 191, 197–198, 219 221, 233, 236, 238, 240, 249, 274, 303, 352 Hyperoxia 22, 29, 46, 48-49, 118, 152-153, 157, 165, 222, 233, 239, 353, 355 Hypertension 66, 80, 125, 210 Hyperventilation 47, 120, 174-175, 210 Hypobaria 173 Hypocapnia 175-176, 318, 321, 353 Hypothermia 171, 277, 303, 352 Hypoxia 13, 22, 29, 39-40, 42, 46, 49, 56, 71-72, 76-80, 88, 90, 95, 98-99, 106, 110-112, 114-118, 123, 125-127, 143, 153, 157, 159, 161, 177, 185, 190-191 198, 207, 209-211, 219, 221-225, 227-228, 234, 237-241, 249, 254, 274 286, 288, 300, 303, 308, 318-319, 321, 335, 352-353, 355-356 Hypselosaurus priscus 350, 352 Icaronycteris index 310 Icefish 16, 106, 113–114 Icerya purchasi 193 Ichthyophis paucesulcus 303, 327 Ictalurus nebulosus 116 – punctatus 197 Incubation 58, 87, 350-357 Inert gases 124, 126, 266, 348 Infundibula 16, 208 Insects 1, 16, 38-39, 46, 70-71, 78-79, 86, 90-91, 95, 102, 110, 114, 117, 122, 131-132, 137, 149, 164, 171, 183–185, 189, 191, 193, 202, 204, 247, 251, 264-265 277, 280-281, 284-288, 292, 309-310, 314, 322-323, 330, 335-336, 344-346, 348 Interalveolar septa 208, 294 Intercostal muscle 237 Intermittent breathing 238, 241, 277, 308 - ventilatory pattern 116-117, 187 Internal gills 184, 191, 193-194, 300 - subdivisions 25, 293, 305, 316, 319, 323 Interstitial oedema 24, 179, 342 Intracardiac separation 243 Intramural blood pressure 198 - stress 179 Intrauterine hypoxia 211 Invagination 25, 61, 101, 131, 250, 271, 276, 278, 280-281 Invertebrates 16, 21, 46, 97, 105, 160, 169, 184, 187, 193, 245, 280 Ionic regulation 27, 60, 153, 155, 183, 186, 198, 231, 233, 236, 242, 245, 249, 254 271, 274 Ionocyte 196, 244, 271 Isopods 99, 150, 249, 278, 280 Jurassic Period 38

Kepler 8 Konia dikume 113 Korean ama 210 Labyrinthine organ 68, 102, 240, 253, 257 Lactic acid 56, 106, 308 Lakes 3, 109, 112, 152, 155-156, 158-161, 163-164, 174, 176, 181, 190, 254, 260 267, 302 Lamellae (see Secondary lamellae) Lampreys 102, 184-185, 188 Laplace's Law 59, 324 Lasionycteris noctivagans 310 Lasiurus borealis 310, 313 - cinereus 310 Latimeria chalumnae (see Coelacanth) Laws of planetary motion (see Kepler) - of thermodynamics 11 Leeches 96 Leiolopisma ladacense 174 Leipoa ocellata 352 Lepidosiren paradoxa 188-189, 193-194, 240, 243, 254-255, 298 Lepisosteus osseus 236, 253-254, 269, 332 Leptonychotes weddelli 13, 223 Leuciscus erythrophthalmus 223 Life forms 25, 33, 71, 177, 193, 252 Ligia oceanica 193 Limax maximus 246 Limnodrilus 200 Limulus polyphemus 190 Lineus longissimus 92 Lipids 17, 125, 269, 334 Liquid breathing 115, 273, 341, 343 Lithosphere 8, 26, 79, 169 Littorina rudis 245 Lizards 56, 68, 101, 149, 204, 247, 275, 289, 301, 304-306, 308, 310, 314, 333-334 349 Lobe-finned fish 20 Loligo pealei 78, 111 Lophius piscatorius 106 Lopholatilus chamaeleonticeps 98 Lugworm 60, 105, 109, 115, 153, 167, 192, 334 Lumpenus lampretaeformis 98, 172–173 Lung 13-14, 16-17, 20-21, 24-25, 38, 43, 45-48, 54-59, 61, 64, 66, 68-71, 73-87, 91 96-97, 99-102, 104, 108, 110, 112, 115, 117-119, 121, 123-124, 126-128, 133-134, 136, 138, 143, 145, 150-151, 156, 161, 165, 167, 171, 174, 176-182 184-185, 187-189, 191, 193-194, 196, 198, 200-204, 207-212, 219, 222-223, 225, 227, 229, 231, 233-234, 236-250, 252, 254-255, 257, 262-267, 269 271, 273-274, 281, 287-294, 298-309, 311-312, 314-316, 318-319, 320-325, 327, 329, 332-345, 349, 351, 357 Lung -air sac system 70-71, 86, 143, 304, 309, 314, 318, 320 Lungfish 20, 69, 96, 165, 184, 187-189, 194, 219, 229, 237-238, 240, 242-244, 254 269, 276-277, 280, 298, 301-302, 306, 309, 332

Lymnaea pereger 200-201 - stagnalis 245 Lymphatics 208, 263 Macrophage 41, 306, 328-329, 332, 341-342 Macrozoarces americanus 106 Magelona 111 Malacochersus tornieri 305 Mammals 12, 16, 22-24, 38, 40, 43-45, 47, 49, 51, 54, 57-59, 61, 64, 66-68, 70-71 73, 76-77, 79-86, 95, 97, 101, 106, 109, 110, 113, 116, 121, 124-127, 131 133-134, 136, 138, 143, 149, 159, 169, 171-172, 175, 178, 204-205, 207-208 211, 215, 225, 233, 236-238, 240-241, 244, 251, 263-264, 273, 277, 280-281, 289-290, 292-294, 304-306, 308-321, 323-324, 332, 334, 336, 343, 349, 355 Mantle cavity 55, 98-99, 102, 125-126, 183-185, 198, 200-201, 223, 225, 245, 248, 265, 278 Marginal channel 198 Marionina achaeta 89 Mars 8-9, 26-27, 32, 154 Maternal blood 59, 207, 209-210 - fetal barrier 207 - fetal PO<sub>2</sub> 211 Mean free path of  $O_2$  78, 288 Mechanoreceptors 238, 240 Medulla oblongata 238 Megaloprepes caeruleata 287 Megalops atlantica 222, 267 Megapodes 352 Megascolides australis 98 Meleagris gallopavo 350 Metabolic acidosis 354 - rate 6, 11-13, 43, 68, 78, 88, 90-91, 97, 99, 109-110, 112, 118, 124, 164 177, 187, 190-191, 225, 227, 237, 257, 269, 273, 279, 284, 286, 301, 303 305, 319-320, 322, 327, 352, 356 Metabolism 10, 12-13, 17, 33, 41, 47, 49, 55, 61, 79, 81-82, 88-89, 106, 114-116 120, 134, 151, 153-154, 166-167, 173, 196, 203, 209, 221-222, 225, 227-228, 233, 236, 241, 266-267, 271, 277, 279, 285-286, 289, 301, 303, 305 308, 324, 340, 350, 355 Metamorphosis 20, 109, 187, 225, 227, 236, 299 Metaphyta 1 Metazoa 1, 12, 17, 32, 54, 87 Methane (CH<sub>4</sub>) 1-2, 7-8, 26-27, 29, 34, 36, 155, 163, 343 Mezozoic 39, 254 Microridges 86, 196-197 Microsorex hoyi 59 Mictyris longicarpus 251 Misgurnus fossilis 254, 332 Mitochondria 10, 14, 16, 41-43, 47, 66-68, 72-73, 75, 89, 114, 123, 127, 134, 166 174, 196, 271, 279, 284-285, 306, 322, 336

Mitochondrial-rich cells (see Chloride cells) Mnierpes macrocephalus 227, 261 Models 36, 50, 57-58, 73, 84, 90, 110, 132-134, 136-137, 183, 192, 207, 219 225, 304, 313, 316 Mole rat 134, 170-172, 309 Mollusc 10, 79, 87, 95-98, 102, 110-111, 116, 183-184, 187-189, 224-225, 245-246, 269, 276, 335 Momarus americanus 111 Monopterus albus 228, 231 – cuchia 243 Morone saxatilis 117 Morphology 53, 108, 112, 181, 197, 215, 286, 299, 302, 323 Mt Everest 173, 176 Mucous cells 196 Mudskipper 183, 194, 198, 261 Multicameral 304, 307, 314, 323 Mus wagneri 309 Muscle 10, 13, 16, 42-45, 56, 66, 68, 73, 76, 90-91, 93, 98, 102, 106, 109-110, 113-115, 118, 120, 123, 133-135, 174-175, 177, 179, 185, 187, 190, 193, 198 200, 233, 236-237, 265, 267, 269, 271, 273, 278-279, 281, 284-286, 288-289, 291, 301-302, 305-307, 311-312, 314, 317-319, 322, 327, 336, 340, 343 Myocardium 84 Myoglobin 13, 16, 22, 24, 106, 110, 113, 283 Myotis daubentoni 312 - lucifugus 312 - velifer 310 Myriapoda 285, 287 Myxicola infundibulum 336 Myxine glutinosa 50, 93-94 Nasal cavity (see Cavity) Natrix rhombifera 305 Natural selection 2-4, 11, 23, 25, 53-56, 61, 68-70, 73-74, 76, 80-81 Nautilus 99, 102, 116, 125, 269, 335 Necturus maculosus 98, 185, 189-193, 271, 302-303 Nematoda 90, 245 Nemertines 92 Neoceratodus forsteri 238-240, 243, 254-255, 276 Neopulmo 316 Neoteny 20, 172 Nereis succinea 187 - virens 185 Nervous system 10, 277 Neurones 10, 17 Nitric oxide 44, 108-109 Nitrogen metabolism 196, 227-228, 271, 350 Niveoscincus metallicus 204 Noctilio albiventris 311, 313 Nocturnal 2, 249, 309, 314 Noradrenaline 198
Normoxia 49, 84, 97, 100, 102, 201 Notothermia gibberiforms 114 – neglecta 114 Numida meleagris 51, 322 Nyctalus noctula 310 Obligate air breather 104, 229, 236, 243, 253, 255 - water breather 243, 276 Oblique septum 289 Octopus vulgaris 99, 111, 185 Ocypode saratan 97, 101, 249 Oedema 24, 83, 126, 179-180, 210, 332, 342 Oligochaetes 89, 95, 99, 164, 167, 187-188, 200 Oncorhynchus kisutchi 221 - mykiss 48, 60, 116-117, 236 - tshawytscha 221 Ondatra zibethicus 349 Onycophora 285, 287 Open circulation 94, 96 Opercular breathing 186 - flap 101, 184 - pump 102, 115 Ophidia 69, 300, 305, 324 Ophisthobranchia 111 **Opisthocomus hoatzin** 314 Opthalmic rete 138, 143 Optimality 74 Optimization 54-55, 58, 71, 74-76, 78-79, 83, 105, 313, 350 Orconectes rusticus 101, 221 Ordovician 36, 38-39 Oreochromis alcalicus grahami 197, 267 Organic phosphate 22, 105-106, 109, 112 Orthoptera 289 Oryctolagus cuniculus 83 Osmoregulation 81, 112, 116, 183, 196, 227, 236, 244, 340 Osmotic pressure 54, 111, 197, 264 Osteichthyes 160, 265 Ostracoderms 35, 181 Ostrich 22, 24, 316, 350, 353 Otala lactea 246-247 Oviparity 204, 349-350 Oxidative metabolism 10, 79, 225 - radicals (see Radicals) Oxygen (O<sub>2</sub>) extraction factor 38, 102, 119, 123, 143, 145, 147, 151, 185, 187, 191-192, 249-250, 312-313, 319-320, 335 Paedomorphy 20, 172 Palaemon elegans 153 Paleopulmo 119, 316 Paleozoic 35, 38-39, 90, 152, 217, 254, 265, 269, 299, 314, 323 Pallial gills 182 Pangasius 68 Panting 24 Pantodon buchholzii 310

Pantopoda 91 Papio anubis 309 Parabronchial air flow 176, 316 gas exchange efficiency 308 - gas exchange tissue 136, 143, 319 - lumen 143 - lung 312, 321, 389, 304, 312, 321 Parabronchus 86, 119, 127, 136, 143, 319 Paramecium 90 Parapodia 184, 187–189, 198 Parenchyma 59, 178, 290, 304-305, 307, 312, 316, 319, 323 Partial pressure 33, 36, 45, 97, 126, 136, 155, 174-175, 187, 229 - gradient 18, 24, 60, 121, 132, 206, 269, 276-277, 345, 356 Passerines 316-318, 322 Passive ventilation 98, 102, 138, 300 Pasteur effect 28, 33 Patagona gigas 125 Patella granularis 182 Paucicameral 304-305 Pavement cells 196-197 Pelagic 2, 78, 117, 196, 266 Pelamis platurus 187 Pelomedusa subflava 240 Pelomyxa palustris 90 Pelvic gills 194 Perfusion 59-60, 71, 77-78, 87, 91-92, 101, 115-118, 121, 127, 133-134, 136, 138 143, 145, 147, 171, 178, 207, 209, 211, 238, 240, 242-244, 266, 276, 300 302, 308, 320, 340 Periodic breather 308 Periophthalmus cantonensis 172 - schlosseri 261 - vulgaris 261 Permian 35-36, 38-39, 90 Pesudacris t. macurata 92 Petromyzon 184 pH 49-50, 71, 92, 106, 109, 111-112, 154-156, 159, 163-165, 167, 175-176, 181 192, 231, 233-234, 236-238, 241, 248, 266, 275, 277-278, 308, 344, 357 Pharnacia serratipes 289 Pharyngeal cavity (see Cavity) Philodendron selloum 90, 273 Pholas dactylus 79, 201, 224, 335 Phospholipid 69, 269, 332 Photosynthesis 30, 32-33, 37, 39, 47, 152, 156-157, 159, 164 Phyllostomus hastatus 311-312, 317 Physa fontinalis 201 Pila ovata 201 Pillar cells 66, 194, 197-198 Pinnae 192 Pinnules 192 Pipa pipa 271, 302, 327 Pipistrellus pipistrellus 310 Pisces 51 Piston pump ventilation 115

Placenta 24, 57-59, 66, 123, 204-205, 207-215, 340-341, 350, 355 Placental barrier 205, 207-208 - diffusional pathway 207 - O<sub>2</sub> consumption 59, 207, 209 Planaria 21, 90 Planets 7-9, 27, 37-38, 40, 153-154 Planorbis corneus 105, 200-201, 245, 280 - ovata 202 Plasma HCO<sub>3</sub> 233-234, 236, 275 Plastron 286, 344, 346, 348-349 Platichthys fleus 116 Platyhelminthes 187 Plautus alle 76 Plecostomus plecostomus 253 Plecotus townsendii 313 Pleopods 150, 172, 190, 193 Plethodontidae 92, 99, 300-301 Pluvialis dominica 318 Pneumatic duct 267 Pneumocytes 208, 302, 306, 332 Pneumonate gastropod 55, 95, 105, 150, 223, 248, 277, 293, 345 Pneumostome 246, 267, 278, 280 Podilymbus podiceps 353 Podocnemys 201 Poecilia reticulata 71 Pogonophoran tube worm 167 Poikilothem 125, 160, 309 Polychaetes 89, 95, 97, 99, 106, 111, 143, 167, 172, 183-184, 188, 192, 200, 335 Polyodon spathula 117–118, 336 Polypterids 20, 193, 255, 269, 271 Polypterus bichir 164 Pomacea depressa 225 Pomatomus saltatrix 117, 196 Porcellio scaber 190, 279-280 Porosity 59, 69, 169-170, 205 Porosphere 169 PreCambrian 12, 30, 155 Pregnancy 210-211, 313, 340, 349 Priapulids 111 Primary bronchus 119 Principle of minimum work 74 Prokaryote 27-28, 30, 33, 40, 47, 68, 87 Prosobranchia 111 Proterozoic Era 31-32, 35 Protobiont 27 Protochordates (see Chordates) Protoplasmic streaming91Protopterus aethiopicus164, 187, 193, 238–240, 243, 254–255 - amphibius 193, 237, 242 Protozoa 16-18, 47, 62, 90-91, 245, 287 Pseudemys floridana 117, 121 - scripta 240, 254, 306 Pseudochaenichthys geogianus 113 Pseudopleuronectes americanus 95 Pseudothelphusa garhami 117, 236, 249 Pteranodon 315, 323

Pteropus edulis 314 – gouldii 317 Pterosaurs 39, 149, 310, 314 Pulmocutaneous blood flow 240, 300 Pulmonary air ways 75, 274, 320 - arteries 66, 69, 84-85, 178, 243, 269 - arterio-venous shunt (see Arterio-venous shunt) - blood capillaries 58, 81-83, 123-124, 143, 180, 255, 263, - blood flow 66, 83, 86, 116, 238, 308, 338 - blood pressure 264 - blood volume 179, 320 - capillary blood 293-294, 298, 306, 319 - capillary blood pressure 83, 179 - capillary transit time (see Transit time) - circulation 66, 85-86, 308, 338, 340 - circulatory resistance 66, 80, 338 - design 22, 75, 324 - diffusing capacity 81, 298, 319-320, 322 elastic tissue recoil (see Elastic recoil of the lung) endothelial cells (see Endothelial cells) gas exchange 341, 344 – haemorrhage 81 - hypertension 66, 80 - macrophage (see Macrophage) morphometric diffusing capacity (see Diffusing capacity) - morphometric parameters 80, 322 - oedema 83 - parenchyma (see Parenchyma) - pathology 45 - perfusion (see Perfusion) - pressure 83, 179 - respiratory surface area (see Surface area) - smooth muscle 289 - structural parameters 311 - surfactant (see Surfactant) - system 70 - vasculature 179, 240, 242 - vein 96, 143, 319 - ventilation (see Ventilation) - water loss 237 Pulmotracheates 280 Pyrrhocorax graculus 318 Quetzalcoatlus northropi 315 Rabbit (see Oryctolagus cunicula) Radicals 41, 44-47 Rainbow trout (see Oncorhynchus mykiss) Ram ventilation 117-118, 194, 336 Rana berlandieri 225 - catesbeiana 191, 227, 236, 242, 302 - pipiens 51, 104, 240, 303 – sylvatica 92 - temporaria 51, 88, 185, 187

Ray-finned fish 20, 273

Reactive radicals (see Radicals) Red blood cells 21-22, 43, 49, 54, 56, 66, 71, 76-77, 82-83, 91, 106, 108-113, 123 126, 175, 196, 198, 208-210, 221, 223, 234, 241, 255, 274, 301, 311, 322, 355 Redundancy 55, 75, 81-82, 320 Remora remora 185 Reptiles 35, 38, 49, 60, 81-82, 102, 109, 116, 122, 125, 127, 134, 149, 174, 178, 185 187, 204, 217, 227, 237, 240-241, 263-264, 274-275, 277, 289, 292-293 298, 304-306, 308-309, 314-315, 317, 319, 323-324, 334, 349, 352 Reserve capacity 56, 211, 213 Respiration (see Breathing) Respiratory acidosis (see Acidosis) - adaptation (see Adaptation) - air flow (see Air flow) - alkalosis (see Alkalosis) - control (see Control of breathing/respiration/ ventilation) - efficiency 53, 61, 69, 78, 115, 120, 134, 175, 242, 316 - gases 4, 17, 24, 48-49, 91, 94, 121, 124, 126, 134, 150-152, 157-158, 166 171, 181, 185, 187, 192, 205, 265, 277, 286, 351 - medium 14, 24-25, 66, 97, 99, 109, 111, 115, 121, 131, 150-151, 228, 245 276, 277, 341 - movements 98-99, 118, 185 - neural system 237 - quotient 99, 274 - surface area 22, 24, 59-61, 66, 71, 86, 98, 131, 183, 185, 194, 198, 203, 221 224, 247, 250, 254, 260-261, 280, 293-294, 303, 309, 312, 316, 320, 322 323-324, 327, 334 - system 22-23, 49, 59, 70, 72, 86-87, 105, 108, 133, 135, 178-179, 280, 288 317, 319-320, 323, 329, 334 - work 49, 101, 119, 121, 174, 196, 307, 332 Rete mirabile 106, 139, 266 Rete opthalmicum 138, 143 Rhacophorus dulitensis 310 Rhinodrilus fafner 89 Rhythmic breathing 98, 103, 200-201, 241 Rib cage 117, 289 Ribs 237, 242, 289, 316, 336 Riftia pachyptila 167 Rivulus marmoratus 166, 231 Rock pools 152 Root effect 106, 165, 266 Rotatoria 90 Sabellidae 192 Safety factor 55-57, 76, 83 Salamander 92, 96, 99, 164, 185, 187-190, 227, 271, 300–301, 333 Salamandra lacertina 164 - salamandra 265, 271

Salting out effect 266 Sandelia capensis 260 Sapharca inaequivalvis 105 Sarda sarda 196 Saturation time 124 Sauromalus obesus 275 Scandentia 310 Scaphoda 187 Scaphognathite 100-101, 172, 249, 251 Sceloporus aeneus 204 Schizobranchia insignis 336 Scomber scombrus 196 Scorpion 278, 280 Scotophilus viridis 310 Scyliorhinus stellaris 233 - canicula 51, 60 Sea 3, 37, 39, 98, 106, 121, 152 – anemone 18, 188 - cucumbers 102, 185, 198 - level 3, 8-9, 35-36, 38-40, 92, 133, 152-153, 155-158, 160-161, 167, 245, 248, 308-309, 343, 353 - raven 51, 95 - snake 187, 308 - turtles 170 - urchin 98, 335 - water 60, 78, 92, 102, 113, 115, 152, 155-157, 159, 173-176, 197-198, 200-201, 209-210, 227, 271 Sea women (see Korean ama) Secondary bronchi 119, 316 - gills 245 - lamellae 78, 86, 128, 138, 182, 194, 196-198, 243-244, 261 Sediment 2-3, 36-38, 89, 98, 100, 105, 153, 156, 166, 172, 192 Segmentum accelerans 119 Selasphorus platycerus 320 - rufus 318 Sepia officinalis 111 Septa 58, 61, 208, 289, 294, 301-303, 327 Serotonin 17, 198 Sheet-flow 62 Shell 8, 12, 57, 59, 69, 102, 184-185, 187, 201, 204-205, 212, 269, 349-357 Sphenomorphus quoyii 204, 349 Shrew 58, Shunts 59-60, 128, 165, 186-187, 191, 194, 197, 207, 209, 240, 242-244, 255, 261 308, 342 Sicyases sanguineus 229, 261 Silentia 190 Silurian 35-36, 221, 241, 274 Siphonaria capensis 245 – zelandica 187 Sipunculids 111 Siren intermedia 334 - lacertina 190, 233 Sirenians 294 Sirenids 164

Skeletal muscle 16, 66, 73, 106, 109, 133, 233, 236, 312, 317 Skin 18, 24, 61, 92, 97, 99, 102, 127-128, 131, 138, 151, 169, 174-175, 183, 185 187-188, 190-191, 193-194, 196, 200-201, 211, 223, 231-234, 236-237 246, 253-254, 267, 275, 280, 300-304, 335 Slug 94, 245-247, 278 Sminthilus limbatus 60 Smooth muscle 10, 44-45, 66, 198, 265, 289, 302, 306-307, 327, 340 Snake 24, 56, 68-69, 98, 149, 172, 187, 197, 204, 240, 253, 289, 300, 304-306, 308 310, 324 Soil 8, 35, 45, 79, 152, 169-171, 202, 217, 255, 352 Solar insolation 3, 26, 300 Solemya reidi 167 – velum 167 Solifugae 285 Solubility of: - CO<sub>2</sub> in water/blood 115, 126, 151, 156, 170, 274 - respiratory gases in tissue membranes 126 - O<sub>2</sub> in water/blood 124, 126, 150, 221, 227, 274 Sorex minutus 327 Spalax ehrenbergi 134, 171 Speciation 2, 32 Spermophilus lateralis 277 Spheciospongia vesparium 88 Spheniscus humboldti 76, 322 Sphenodontia 304 Sphenomorphus fragilis 204 Sphenomorphorus quoyii 204, 349 Spider 78, 265, 278-280, 288, 346 Spiracles 115, 279-280, 288, 346 Sponges 88, 143, 151, 335 Squalus acanthias 51, 204 - suckleyi 51, 145 Squamata 274, 305 Static recoil pressure 342-343 Sterna paradisea 317 Sternotherus minor 103 - odoralis 187 Stroke volume 99, 117, 125, 308 Strongylocentrotus droebachiensis 98 Structural-functional correlations (see Adaptation) Struthio camelus 24, 350 Sturnus vulgaris 312 Stygocapitella subterranea 89 Suctional breathing 38, 81, 101, 104, 242, 289, 292 Sun 7-9, 11, 33, 39, 104 Superoxide anion 41, 45 - dismutase 43, 46 Suprabranchial chamber 68, 102, 203, 253, 257, 260

Surface area 16, 22, 24-25, 43, 54, 58-61, 66, 71, 76-78, 82, 84, 86, 88, 91-91, 97-98, 111, 114, 121, 127, 131, 139, 143, 152, 175, 179, 182, 185-186, 192-194 196-198, 203, 205-208, 212, 221, 224, 247, 250, 254, 260-261, 264, 271 280, 286, 294, 303, 309, 312, 314-316, 319-320, 322-324, 327, 332, 334 336, 353 Surface skimming 224 Surface tension 59, 69, 154, 182, 312, 316, 324, 332-334, 346 Surface - to - volume ratio 54, 77, 88-89, 91, 187, 300, 304 Surfactant 17, 69, 263, 306-307, 324, 330, 332-334, 342-343 Swamps 161, 165, 181 Swamp worm 167, 202 Swim bladder 13, 69, 106, 123, 265, 266-267, 269, 332 Symbiosis 49 Symmorphosis 73-75, 79 Sympathetic nerves 269 Synapta 91 Synbranchus marmoratus 236-237, 276 Syndesmochorial placenta 208 Systemic arteries 308 - blood flow 240, 242-243, 255, 308 - vasculature 85, 179, 196, 242-244, 340 Tadarida brasiliensis 310, 313 Tadarida maxicanobrasiliensis 311 Tadpoles 109, 183, 189, 191, 193-194, 225, 227, 236, 275 Taenidia 285, 330 Tardigrada 91 Taxonomic diversity 1 Tectonic activity 32, 38 Teleosts 51, 60, 93, 95, 104–105, 113, 123, 183-184, 189, 193-194, 197, 204, 228, 240-241, 243, 244, 252-253, 265, 267, 269, 273, 349 Telmatobius culeus 174, 190, 302 Temperature 2-4, 8-10, 13, 26-27, 34, 37-40, 78, 90-92, 95, 98-100, 106, 109, 110 112-115, 117, 121, 124, 128, 151, 154-161, 165, 167, 169, 174-176, 181 190-192, 210, 219, 221-222, 225, 227, 236, 238, 241, 245-248, 258, 273-278, 300, 322, 340, 343, 346, 352-353 Tenebrio molitor 288 Terebellid worms 18, 184, 187 Terrapene ornata 275 Terrestrial 2-3, 8, 12, 24, 32-38, 49, 59, 74, 94-95, 97, 109, 116-117, 124, 149-150 160, 165-166, 183, 190, 194, 217, 219, 222-224, 227, 231-232, 236, 238-239, 242, 245-246, 248-253, 255, 257, 262, 264-265, 271, 273-274, 276-277, 287, 289, 292, 299-304, 308, 323, 344, 349 Tertiary bronchus (see Parabronchus)

Tertiary period 39, 105, 308 Testudo graeca 240, 306 – dendriculata 275 Tetrapods 20, 69, 74, 219, 227, 242, 254, 273-275, 298 Thamnophis sirtalis 204, 240, 306 Theraphosa blondi 279 Thermodynamics (see Laws of thermodynamics) Thickness of the blood-gas barrier (see Blood-gas barrier) Thomomys bottae 170 Thoracic cavity (see Cavity) Thoracic pumping 287 Thunnus thynnus (see Tuna) Tidal breathing 102, 185 Tilapia grahami 109 Tinca tinca 110, 221 Tissue barrier 14, 17, 24, 66, 110, 121, 126, 131, 135, 137, 174, 190, 201 - capillarity 72 - cells 14 16-17, 44, 62, 84, 88, 112, 118, 124, 280-281, 285-286, 303, 337 - elasticity 179 – fluid 12 - O<sub>2</sub> permeability 16 - oxidative capacity 306 - oxygenation 13 - protein turnover 12 - regeneration 46 Toad 51, 96, 240, 264, 302-303 Torpor 171, 273 Trabeculae 61, 301, 306 Trachea 16, 22-23, 70-71, 86, 90-91, 99, 131, 200, 247, 264, 277, 280-281, 284-288, 300, 309, 323, 330, 344-346, 348-349 Tracheal convolution 24 - dead space 22 - fluid 286 - gills 200, 348 - growth 288 - length 24 - respiration 280, 285 - respiratory surface area 24 - system 86, 91, 131, 277, 280, 286-288, 309, 323, 344, 348 - volume 22, 286 Tracheates 280, 284-285, 287 Tracheoles 91, 264, 281, 285-286, 288, 323 Tracheopulmonata 247 Trachurus mediterraneus 86 Transit time 54, 57, 121, 124, 126, 184, 313 Transitional breathers 228, 234, 242 Transpalcental O<sub>2</sub> transfer 206, 208 Triassic period 3, 38–39, 253 Trichoptera 200, 348 Trichotoxon copleyi 248 Trimodal gas exchange 190 Trionyx mucita 187 - spiniferous asperus 187

Triturus alpestris 301 – vulgaris<sup>°</sup> 332 Trophoblast 207, 209, 211-212 Tropical desert 3 - freshwater 160-161, 221, 224, 252, 276 - swamps 38, 156, 161, 163-164, 181, 194, 201-202, 224 Trout 48, 51, 60, 78, 113, 116–117, 143, 187, 236 Tubifex tubifex 200 Tubular myelin 332 Tuna 102, 106, 109, 113, 117, 160, 185, 194, 271 Tupinambis nigropunctus 247, 287, 304 Turkey (see Meleagris gallopavo) Tursiops truncatus 334 Typhlonectes compressicauda 188, 204, 300-301, 349 Umbilical artery 210-211 - blood flow rate 211 - cord 356 - venous/arterial blood circulation 211 - vessels 210-211 Umbra limi 267 Unicameral lungs 304 Uniramians 277 Unstirred water layer 128 Urea 237-238 - excretion 17 - metabolism 227 - toxicity 183 Urechis caupo 98, 102, 105, 166, 185, 201 Uric acid 227 Urodeles 98-99, 185-187, 190, 204, 271, 301, 303 Uterine vein 213 Uterus 350 van der Wall's force 155 Vapour pressure 150, 174, 224, 274, 353 Varanus exanthematicus 96, 304-305 Vascular resistance 66, 97, 120, 182, 197, 210, 240, 243, 302 - shunts 59, 186, 207, 209, 240, 243 Vasculosyncytial membrane 207, 209 Ventilation 48-49, 61, 68, 71, 78-79, 82, 87, 97-99, 100-103, 115-118, 120-121 133-134, 136, 138, 143, 145, 153, 157, 165, 173-178, 183-185, 189, 191 194, 200-201, 210, 221, 223, 233-234, 236-241, 244, 249, 255, 265, 275-277, 287, 289, 300, 305, 308, 319-320, 327, 335-336, 343 Ventilation-perfusion ratio 116, 133-134, 145, 178, 308, 320 Ventilatory work 115-116, 138 Vertebrates 1, 10, 12, 16-17, 21, 35, 48-49, 60-61, 69, 76, 92, 96, 101, 104-106 108, 110-113, 123-125, 134, 138, 143, 149, 156, 159, 204, 209, 219, 233-234, 236-241, 244, 249, 255, 265, 275-277, 287, 289, 300, 305, 308, 319-320, 327, 335-336, 343

Villi 205, 207, 211–212
Villous membrane 59, 207
Viscosity 196, 302, 311
Viscosity of:

air 318
blood 76–77, 110, 114, 124
blood plasma 111
red blood cells 112
water 115, 120, 150, 159, 265, 342–343

Vivaparity 204, 215, 349
Volcanic activity 3–4, 33, 112, 181, 267

Water breathing (see Breathing) – gill (see Gill) – hyacinth 164 loss 24, 38, 60, 97, 116, 131, 181–182, 222, 227, 237, 246, 271, 275–276 279, 300, 303, 350, 352–353
lung 150, 198, 200, 202–203, 245, 265
Water- blood barrier 60, 78, 127–128, 136, 138, 185, 190, 198
Water-air interface 217, 219
Whale 2, 13, 82–83
Wing beat – breathing ratio 69, 311–312
Wing beat kinematics 132
Xenopus laevis 239–240, 271, 302–303, 306
Xeric habitats 292, 303

Zoarces viviparous 204, 349

Xyelacyba myersi 185